18甘女g
$z$ INHISN
$Z_{z}$ NOILOLILSNI ${ }^{-}$NVINOSHLIWS

1 y $\forall \& \operatorname{cil}$


 INSTITUTION NOILRLILSNI＿N甘INOSHLIWS

S31ष甘प817






LIBRARIES

 INSTITUTION




SMITHSONIAN


INSTITL
 LIBRARIES SMITHSONIAN


NOILOLILSNI
 libraries s smithsonian NOILOLILSNI ${ }^{-1}$ NGINOSHLIWS



SヨIy TITUTION

$1 \quad y \forall 8817$


## $1 \forall \forall y 817$



NOUIISNI
STITUTION



I 바빅

## STITUTION





LIBRARIES
$\qquad$


Sヨ1y४y 17


SMITHSONIAN
 NOILOLILSNI NVINOSHLIWS SヨI甘甘
$\bar{z}$
-3
-1
-1
-1
0
2
 LIBRARIES SMITHSONIAN


INSTIT


INSTITI



NOILCLILSNI


46！ IBRARIES


SMITHSONIAN
 INSTITUTION


SヨIV


N甘INOSHLIWS

$\sum_{-1}^{\infty}$
SMITAN

站 ${ }^{\infty}$ SMITHSONIAN RARIES SMITHSONIAN INSTITUTION

## $\sum_{\infty}$

NOIIOIISNI NVINOSHIIWS
$\sum_{5}^{\sum} 518 \forall 8917^{2}=$ LIBRAI SBI

InIIISNI

N甘INOSHLIWS


SMITHSONIAN

## RARIES



RARIES


SMITHSONIAN
를
$\frac{1}{2}$
$\frac{t}{6}$
$\frac{n}{2}$

BRARIES ${ }^{\text {SMITHSONIAN }}$

|  |
| :---: |
|  |  |

IINLIISNI－${ }^{-1}$ N甘INOSHLIWS
$\frac{1}{2}$

BRARIES SMITHSONIAN


IIOLIISN
0
3
3
1
1
0
2
2
2
NSTITUTION

－NVINOSHLIWS
IInIILSN



NOI 1 ก1


NOILOLILSNI


SヨI \＆甘木g17 ${ }^{\text {z }}$ LIBRARIES


SMITHSONIAN

Sヨ1女甘y 17


LIBRA



NVINOSHLIWS


Sヨly甘yg17 SヨI甘甘yal7LIBRARIES


SヨIy甘y $17{ }^{-}$LIBRARIES



${ }^{2}$ SMITHSONIAN ${ }^{-1}$ INSTITUTION



Sま1 प甘女g 17

NOI」n．

LIBR

LIBRA
$\overline{2}$
$\frac{1}{-1}$
-1
-1
0
$z$

n
3
3
3
1
0
0
2
2
,


### 507.68

ANNALS OF THE
SOUTH AFRICAN MUSEUM
$\because$

VOLUME 59

# ANNALS <br> OF THE 

## SOUTH AFRICAN MUSEUM

VOLUME `59


PRINTED FOR THE
TRUSTEES OF THE SOUTH AFRICAN MUSEUM
1971-1972

## LIST OF CONTENTS

Page
Boonstra, L. D.
Discard the names Theriodontia and Anomodontia: a new classification of the Therapsida (published September 1972) ${ }^{\prime}$ ..... 315
Boonstra, L. D.
The early therapsids (published December 1971) ..... 17
Haigh, E. H.Development of Trachurus trachurus (Carangidae), the South African maasbanker(published May 1972)139
Haigh, E. H.
Larval development of three species of economically important South African fishes (published March 1972)47
Hendey, Q.B.
The evolution and dispersal of the Monachinae (Mammalia: Pinnipedia) (publishedMarch 1972)99
Hendey, Q. B. \& Repenning, C. A.
A Pliocene phocid from South Africa (published March 1972) ..... 71
Hendey, Q. B.
A Pliocene ursid from South Africa (published March 1972) ..... II 5
Hooijer, D. A.A Late Pliocene rhinoceros from Langebaanweg, Cape Province (published August1972) . . . . .. .. .. .. .. .. .. .. I5I
McKenzie, K. G.A new species of Paradoxostoma (Crustacea, Ostracoda) from South Africa (publishedMay 1972)${ }^{1} 33$
Maier, W.Two new skulls of Parapapio antiquus from Taung and a suggested phylogeneticarrangement of the genus Parapapio (published December 1971)I
Roeleveld, M. A.
A review of the Sepiidae (Cephalopoda) of southern Africa (published September 1972) .193

ANNALS OF THE SOUTH AFRICAN MUSEUM ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM

| Volume | 59 | Band |
| ---: | :---: | :--- |
| December | 1971 | Desember |
| Part | $\mathbf{1}$ | Deel |



TWO NEW SKULLLS OF PARAPAPIO ANTIQUUS FROM TAUNG AND A SUGGESTED PHYLOGENETIC ARRANGEMENT OF THE GENUS PARAPAPIO

By
WOLFGANG MAIER

## The ANNALS OF THE SOUTH AFRICAN MUSEUM

 are issued in parts at irregular intervals as material becomes availableObtainable from the South African Museum, P.O. Box 6i, Cape Town

Die ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM word uitgegee in dele op ongereelde tye na beskikbaarheid van stof

Verkrygbaar van die Suid-Afrikaanse Museum, Posbus 6r, Kaapstad
OUT OF PRINT/UIT DRUK
r, 2(1, 3, 5, 7-8), 3(i-2, 5, t.-p.i.), 5i-2, 5, 7-9),
$6(\mathrm{r}, \quad$ t.-p.i. $), \quad 7(\mathrm{I}-3), \quad 8, \quad 9(\mathrm{I}-2), \quad 10(\mathrm{I})$,
II (1-2, 5, 7, t.-p.i.), 21, 24(2), 27, 3I(1-3), 33, $3^{8}$

Price of this part/Prys van hierdie deel RI,8o

Trustees of the South African Museum (C) Trustees van die Suid-Afrikaanse Museum 1971

# TWO NEW SKULLS OF PARAPAPIO ANTIQUUS FROM TAUNG AND A SUGGESTED PHYLOGENETIC ARRANGEMENT OF THE GENUS PARAPAPIO 

By<br>Wolfgang Maier<br>Dr. Senckenbergische Anatomie der Universität Frankfurt a./M.<br>(With I plate, 4 figures, 3 tables)<br>[Ms. accepted 30 Fuly 1971]

## Contents



## Introduction

The travertine caves near Taung, Cape Province, are the type locality of Australopithecus africanus. In addition, some Pleistocene cercopithecoids and a considerable number of other fossil animals were found there. The fossil material, however, was not collected systematically and is not as well known as that of the australopithecine-bearing dolomite caves of the Transvaal. Peabody (1954) has compiled a faunal list for the Taung sites.

The fossil cercopithecoids from Taung were first mentioned by Haughton (1925), who proposed the name Papio antiquus for the material available at that time. Gear (1926), describing these and additional new specimens in more detail, distinguished another species, Papio izodi. Broom (1940) included both species in the genus Parapapio, which had been created by Jones (1937) for a primitive baboon-like form from Sterkfontein. Only Freedman (1957) recognized the clearcut differences between both taxa; he considered that only the first species belonged to Parapapio, P. antiquus Haughton, I925, while the latter was a primitive true baboon, Papio izodi Gear, 1926. In 1957 Freedman also described the colobid Cercopithecoides williamsi and the small Parapapio jonesi from Taung, and in 196I the same author described Papio wellsi, another true baboon found at this site.

Unfortunately most of the Taung caves were mined out by the early fifties and consequently have yielded no further fossil material. In 1952, however, Mr. James Kitching of the Bernard Price Institute for Palaeontological Research, Johannesburg, was able to rescue the last few primate specimens from the Taung dumps. The present author was kindly allowed to prepare this material, which appeared to consist of two fairly complete female skulls
of Parapapio antiquus. These two new specimens were embedded in a fine-grained pinkish breccia, which seems to be more calcified in M.3079 than in M.3078, the hardness of the former approximately corresponding to that of the pink cercopithecoid breccia (Brain's Upper Phase I) of the Makapansgat Limeworks. Both specimens are housed at the Bernard Price Institute for Palaeontological Research, Johannesburg.

## Description

The measurements of the two new specimens are incorporated in Tables I and 2, which at the same time provide comparative data. Unless otherwise stated, the technique of measuring is in accordance with the definitions of Freedman (1957), and most of the comparative data have been extracted from the publications of the same author.

## (1) Specimen M. 3078 (Pl. i)

This is a fairly complete and undistorted cranium with the third molars only newly erupted and not having been in occlusion. The muzzle is complete as is the left half of the braincase; the right half of both the upper face and the braincase together with most of the cranial base have been eroded away. The incisors, the canines and the right $\mathrm{P}^{3}$ were lost before fossilization, but their alveoli have been preserved. The remaining teeth are in relatively good condition. The morphology and size of both skull and teeth indicate that this was a young female of Parapapio antiquus.

The proportions of this cranium are similar to those of Tvl. 639 (Transvaal Museum, Pretoria; Freedman 1957: Fig. 48), the muzzle being short in relation to the braincase. The braincase itself is fairly flat in the frontal region, but drops relatively steeply in the parietal region. The nuchal plane is therefore situated deeply in the backward prolongation of the alveolar margins. As far as can be seen, the mastoid processes must have been well developed, whereas nuchal crests are absent. The nuchal line runs backward as a straight continuation of the jugal arch, the inion therefore being in a lowered position. The temporal crest shows the typical course met with in other specimens of Parapapio antiquus: it is well pronounced in its frontal part, exhibiting only a slight notch behind the orbit; hence, it overhangs the postorbital constriction of the lateral wall of the braincase (temporal fossa), resulting in a wide postorbital breadth when seen in dorsal view. On the parietal bone the faint temporal line very gradually converges toward the midline, but approaches to within only about 15 mm of it.

The anterior root of the jugal arch starts above the distal half of the second molar. The zygomatic part of the jugal arch is comparatively strong and broad, showing clearly the area for the insertion of the masseter muscle. The temporal part of the arch is narrower, but exhibits a strongly developed tubercle frontolateral to the articular fossa. This fossa is remarkably deep and distinctly concave transversely. Posteriorly it is bounded by a very small postglenoid process.

Medially, the glabellar region is very undeveloped, the nasal line running as a nearly straight continuation of the frontal outline. Laterally there exist shallow excavations between the supraorbital arcus and the cranial vault. The arcus are barely prominent, but possess distinct supraorbital notches. The left orbit does not seem to be disproportionately large and is fairly well rounded. The interorbital and nasal region show a straight contour, which is a typical feature of Parapapio antiquus as compared with female skulls of the other species of this genus.

The muzzle appears to be quite narrow and slender in this specimen, because the maxillary crests are not strongly developed. Hence, although the muzzle dorsum drops steeply towards the sides, the canine fossae are comparatively well excavated. There are 4 to 5 infraorbital foramina on each side, opening separately just at the posterior end of the maxillary crests. The premaxilla protrudes considerably, indicating a well-developed incisor row; the lateral wings of the premaxilla do not reach the nasal bone. The nasal aperture shows a typical ovoid outline.

The maximum breadth of the muzzle and of the ovoid tooth arch lies across the anterior half of the second molars. The palate seems to be short, the posterior margin lying between the last molars. The greater palatine foramina are slit-like and they are situated between the second and the third molars. The incisive fossa opens between the canines. The angle between the pharyngeal face of the base of the braincase and the palate is $135^{\circ}$; in two new female skulls of Parapapio broomi (M.3056 and M.3070) it is $127^{\circ}$ and $122^{\circ}$.

Due to the immaturity of the new specimen, the alveolar processes are quite undeveloped, resulting in a comparatively low facial height.

The alveoli of the (missing) medial incisors are about $5,5 \mathrm{~mm}$ in breadth, but those of the lateral incisors only about 4 mm , thus indicating the specialized broadening of the former ones. The alveoli of the canines measure about 6 by 7 mm ; in the male specimen T. 22 (Transvaal Museum, Pretoria) of Parapapio antiquus these dimensions are 9 by 9 mm , proving that the present skull is that of a female. Both premolars are well developed and comparatively elongated. In the last two upper molars, the distal pair of cusps, and particularly the disto-buccal cusps are conspicuously reduced in size.

## (2) Specimen M. 3079

This specimen is not as complete as the first one. It comprises only a fairly well-preserved facial skeleton and the frontal part of the calvaria. The few remaining teeth are very worn and although damaged to some degree, show that it was a very old animal. The front teeth were lost before fossilization, but their alveoli are still visible. The canine alveoli are comparatively small, thus indicating that the present cranial fragment is that of a female. The skeletal parts show some minor cracking, probably causing some slight distortion.

The muzzle of this specimen appears to be altogether heavier and stouter than that of the first specimen. The maxillary crests are more prominent, the
muzzle dorsum consequently being broader and more flattened, as is typical for the species (Freedman 1957). Most breadth measurements of this fragmentary cranium are distinctly greater than in M.3078, whereas the length measurements are very similar. The degree of excavation of the canine fossae is nearly identical in both specimens. In the present cranium the orbits seem to be more flattened and the supra-orbital arcus more developed, resulting in a more conspicuous ophryonic groove. These features are shared with specimen Tvl. 639, which also represents an old female. The temporal crests are very strong in M.3079, resulting in a very great intertemporal breadth. The zygomatic bone of the left side is partly damaged, but appears to have been very strong. The anterior root of the jugal arch, as in Tvl. 639, is also situated above the anterior part of the third molar; the tooth rows of these two old specimens were thus shifted relatively more forward than in the younger specimen. The same age differences are to be observed in the height of the face, the older specimens being distinctly higher. In living primates this downward and forward growth of the alveolar processes is well known to occur during adulthood (Scott 1967).

There is only one premolar left in the present cranium, and this has been partly damaged; it is fairly similar to those of M. 3078 and T. 17 (Transvaal Museum, Pretoria). The existing first molar is very worn and extruded and thus comparatively long. However, $\mathrm{M}^{2}$ and $\mathrm{M}^{3}$ are also very long, and they belong to the top of the known size range for this species. As far as can be seen in both of these posterior molars, there is considerable reduction in the breadth of the distal cusps, which is typical for Parapapio antiquus.

## Discussion

The two new skulls, described above, add in many respects to our knowledge of the fossil species Parapapio antiquus, which so far has been found only at Taung. These two specimens confirm that the peculiar shape of the muzzle, with its straight nasal and its well-developed maxillary crests and canine fossae, is very characteristic for this taxon. As the material comprises a young adult and a very old female skull, we can appreciate some of the morphological differences due to age. In the young specimen the muzzle is more slender and the face is narrower and less high than is the case in old specimens (Figs I and 2). Skull M. 3078 exhibits, for the first time, morphological details of the articular and infratemporal fossae of $P$. antiquus.

The lateral and oblique position of the temporal crests in $P$. antiquus indicates a backward-orientation of the temporal muscle, which may be correlated with some specialization of the masticatory function, i.e. a stressing of more anterior parts of the dentition. Interestingly, some of the tooth characters of $P$. antiquus seem to support this kind of functional interpretation (see below). This specific course of the temporal crests is already met with in the comparatively young animal M.3078, whereas the typical flattening of the muzzle dorsum is not yet evident.


## PARAPAPIO ANTIQUUS

- M. 3078

Fig. I. Pantographs of the young female specimen M. 3078 and of the old female M. 3079 are superimposed to show differences due to age. Both are shown in norma dorsalis, being orientated on the occlusal plane.


Fig. 2. Pantographs of four of the most complete skulls of Parapapio antiquus are superimposed in norma lateralis. Specimens Tvl. 639 and 56604 are reversed. The picture demonstrates an increasing degree of tooth row declination and facial height with increasing age; it shows also the small degree of sexual dimorphism in the known specimens.
Table i. Cranial measurements of Parapapio antiquus and P. broomi

|  | Parapapio antiquusfemales males |  |  |  |  |  |  |  |  |  | Parapapio broomi* females** males |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | M.3078 | M. 3079 | T.17 | T.ir | Tvl. 639 | 56 694* | n | X | n | X | n | X | n | X |
| Maximum length | 135 | - | - | (141) | (140) | - | 3 | 139 | 1 | (150) | 3 | 140 | 3 | 161 |
| Basal length . | (88) | - | - |  | (100) | - | 1 | 100 | - | - | 2 | 98 | 1 | 124 |
| Bizygomatic breadth . | (88) | - | - | - | - | - | 1 | (88) | 1 | 96 | 4 | 96 | 2 | 106 |
| Basion-Bregma. . | (56) | - | - | - | 62 | - | 2 | 59 | - | - | - | - | 1 | 66 |
| Basion - Glabella | (70) | - | - | - | 73 | - | 2 | 72 | - | - | 2 | 66 | 2 | 76 |
| Minimum interfrontal | (53) | (61) | - | 53 | 54 | 52 | 5 | 55 | 1 | 57 | 3 | 52 | 4 | 53 |
| Maximum temporal . | (70) | - | - | - | 72 | - | 2 | 71 | 1 | 73 | 2 | 74 | 2 | 78 |
| Inion-Glabella . . | 92 | - | - | (90) | $9{ }^{1}$ | - | 3 | 91 | 1 | 96 | 1 | 98 | 4 | 99 |
| Muzzle height ant. $\mathrm{P}^{3}$ | 17 | 16 | 18 | , | 19 | 15 | 5 | 17 | 2 | 23 | 1 | 15 | 2 | 26 |
| Muzzle breadth ant. $\mathrm{M}^{3}$ | 45 | 48 | 47 | - | - | 46 | 4 | 47 | 1 | 48 | 2 | 50 | 4 | 52 |
| Muzzle breadth ant. ${ }^{3}$ | 34 | 39 | - | - | 37 | 35 | 4 | 36 | 1 | 36 | 3 | 38 | 3 | 43 |
| Muzzle breadth dors. $\mathrm{M}^{2}$. | 30 | 37 | 35 | 30 | 35 | - | 4 | 33 | 1 | 38 | 2 | 33 | 3 | 40 |
| Nasion-Prosthion | 66 | (66) | - | (68) | (71) | 68 | 5 | 68 | 2 | 79 | 3 | 70 | 3 | 86 |
| Staphylion - Prosthion | 54 |  | (62) | ) | (64) | - | 4 | 59 | 1 | 66 | 2 | 60 | 2 | 66 |
| Nasion-Rhinion . | (30) | (30) | - | 33 | 32 | - | 4 | 31 | 1 | 41 | 3 | 35 | 1 | 43 |
| Interorbital breadth | 8 | (10) | - | 7 | 9 | 9 | 5 | 9 | 1 | 9 | 3 | 9 | 2 | 12 |
| External orbital breadth | (69) | (76) | - | 68 | 73 | 70 | 5 | 71 | 2 | 77 | 3 | 70 | 2 | 77 |
| Orbital height . . | 22 | 20 | - | 22 | 23 | 22 | 5 | 22 | 2 | 24 | 3 | 24 | 2 | 24 |
| Orbital breadth . . | 24 | 26 | - | 26 | 26 | 25 | 5 | 25 | 2 | 27 | 2 | 27 | 3 | 26 |
| Nasal aperture breadth | 15 | 16 | 17 | 16 | 17 | 19 | 6 | 17 | 1 | 17 | 3 | 15 | 2 | 18 |
| Nasal aperture height | 27 | (26) | 25 | 27 | 29 | - | 5 | 27 | 2 | 27 | 2 | 25 | 2 | 36 |
| Vertical height of face (Alveolar line-torus s.) | 51 | 53 | - | - | 62 | - | 3 | 55 | 2 | 62 | 2 | 59 | - | - |
| Alveolar 1. -infraorb. margin | 30 | 34 | 34 | - | 36 | - | 4 | 34 | 2 | 35 | 2 | 35 | - | - |

* After Freedman 1957.
** Including two new skulls from Makapansgat.

Although the morphology of the female skull of $P$. antiquus is fairly well known, metrical data are still poor, and the present knowledge of the male skull is very unsatisfactory (Table I). In overall size, female skulls of Parapapio antiquus are very similar to those of $P$. broomi, whereas male skulls of the latter species are considerably larger than those of the former, suggesting a lesser degree of sexual dimorphism for P. antiquis (Maier 1971). Figure 2 shows a craniogram of the most complete male skull known so far (University of California, Museum of Paleontology Specimen No. 56 604; unfortunately still partly embedded; see Figure 4 in Freedman 1965); the muzzle is only slightly longer and more declined than in the super-imposed female craniogram.

The relatively large numbers of teeth permit a statistical analysis to be made. This is, however, true only for premolars and molars, the front teeth still being virtually unknown. Table 2 provides comparative data for the other species of Parapapio. Although the length of the tooth row is very similar in P. antiquus and $P$. broomi, there seem to be some discrepancies in the dimensions of individual teeth. $\mathrm{P}^{4}-\mathrm{M}^{2}$ are distinctly longer in $P$. antiquus, whereas


Fig. 3. Breadth/length indices of the last upper premolar and the upper molars in the four species of the genus Parapapio. The values for $P$. broomi are considered to be 100; those of the other species are related to $P$. broomi. The diagrams demonstrate the aberrant tooth proportions of $P$. antiquus. For exact data see Table 2.
Table 2．Measurements in mm of upper teeth in females of various Parapapio species ${ }^{1}$

|  | Parapapio antiquus |  |  |  |  |  |  |  |  |  | P．broomi＊ |  |  | P．whitei＊ |  |  | P．jonesi＊ |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | M． 3078 | M． 3079 | T． 17 | T． 16 | T．25＊ | 56 608＊ | 56 694＊ | n | X | B／L 100 | n | X | B／L 100 | n | X | B／L 100 | $n$ | X | B／L 100 |
|  | 三 | 二 | 二 | － | － | 二 | 二 | － | 二 | － | $\begin{aligned} & 4 \\ & 2 \\ & 2 \end{aligned}$ | $\begin{array}{r} 7,3 \\ 6,9 \\ 11,0 \end{array}$ | 94，5 | ． | － | － | 2 1 1 | 6,6 6,7 11,0 | 101，5 |
| $\mathrm{I}^{2} \quad \begin{array}{lll}\text { a } \\ & \\ & \\ \\ & \mathrm{b} \\ \mathrm{h}\end{array} \quad$. | － | 二 | 二 | － | 二 | － | 二 | － | 二 | － | $\begin{aligned} & 4 \\ & 2 \\ & 3 \end{aligned}$ | 5,8 6,0 8,6 | 103，4 | － | 二 | － | $\begin{aligned} & 2 \\ & 1 \\ & 2 \end{aligned}$ | 4,9 6,1 7,0 | 124，4 |
| C $\quad \begin{array}{lll}\text { l } \\ & \text { b } \\ & \text { h }\end{array}$ | 二 | 二 | $(6,2)$ <br> $(6,3)$ | 二 | 二 | 二 | － | $\begin{array}{r}1 \\ 1 \\ \hline\end{array}$ | $(6,2)$ <br> $(6,3)$ | 101，6 | $\begin{aligned} & 4 \\ & 4 \\ & 2 \end{aligned}$ | $\begin{array}{r} 5,9 \\ 6,1 \\ 12,2 \end{array}$ | 103，4 | $\begin{aligned} & 1 \\ & 3 \\ & 1 \end{aligned}$ | $\begin{array}{r} 7,3 \\ 7,0 \\ 16,0 \end{array}$ | 95，9 | $\begin{aligned} & 2 \\ & 3 \\ & 3 \end{aligned}$ | 6,1 5,8 7,9 | 95，0 |
| $\mathrm{P}^{3} \quad{ }^{1(\mathrm{~h})}$ ． | 9,8 7,4 | － | $\overline{7,9}$ | － | － | － | － | $\begin{aligned} & 1 \\ & 2 \end{aligned}$ | $\begin{aligned} & 9,9 \\ & 7,7 \end{aligned}$ | $\overline{77,8}$ | $\begin{aligned} & 10 \\ & 10 \end{aligned}$ | $\begin{aligned} & 6,6 \\ & 7,4 \end{aligned}$ | 112，1 | $\begin{aligned} & 4 \\ & 3 \end{aligned}$ | $\begin{aligned} & 7,4 \\ & 7,9 \end{aligned}$ | 106，8 | $\begin{aligned} & 5 \\ & 6 \end{aligned}$ | 5,4 7,1 | 131，5 |
| P4 $\begin{array}{lll}\text { 4 } \\ & \text { b }\end{array}$ | 6,9 8,3 | $\begin{aligned} & (6,8) \\ & (8,0) \end{aligned}$ | $\begin{gathered} 7,1 \\ (8,0) \end{gathered}$ | － | 7，7 | 6,8 8,2 | 6，7 | $\begin{aligned} & 6 \\ & 4 \end{aligned}$ | $\begin{gathered} 7,0 \\ (8,1) \end{gathered}$ | $\begin{aligned} & (n=4) \\ & 117,7 \end{aligned}$ | $\begin{aligned} & 14 \\ & 12 \end{aligned}$ | 6,4 8,4 | $\begin{gathered} (n=23) \\ 131,5 \end{gathered}$ | $\begin{aligned} & 6 \\ & 6 \end{aligned}$ | $\begin{aligned} & 6,9 \\ & 8,7 \end{aligned}$ | $(n=7)$ 129,1 | $\begin{aligned} & 6 \\ & 5 \end{aligned}$ | 6,0 7,7 | $\begin{aligned} & (n=7) \\ & 129,8 \end{aligned}$ |
| $\mathbf{M}^{1} \begin{array}{r} 1 \\ \\ \\ \\ \mathrm{mb} \\ \mathrm{db} \end{array} .$ | 10,0 8,5 8,0 | 11,0 $(9,2)$ $(9,0)$ | $\begin{aligned} & (9,8) \\ & (9,5) \\ & (8,8) \end{aligned}$ | $\begin{aligned} & 10,9 \\ & 10,6 \\ & 10,0 \end{aligned}$ | 10,3 - | 9,2 9,1 7,7 | $\frac{(8,9)}{8,7}$ | $\begin{aligned} & 7 \\ & 5 \\ & 6 \end{aligned}$ | $\begin{array}{r} 10,0 \\ 9,4 \\ 8,7 \end{array}$ | $\begin{gathered} (\mathrm{n}=5) \\ 92,3 \\ 85,4 \end{gathered}$ | $\begin{aligned} & 14 \\ & 12 \\ & 10 \end{aligned}$ | 9,6 9,5 8,8 | $\begin{gathered} (\mathrm{n}=15) \\ 99,3 \\ 91,2 \end{gathered}$ | $\begin{aligned} & 6 \\ & 5 \\ & 5 \end{aligned}$ | $\begin{array}{r} 10,7 \\ 10,5 \\ 9,8 \end{array}$ | $\begin{gathered} (n=5) \\ 98,5 \\ 91,7 \end{gathered}$ | 6 5 5 | 8,7 8,8 8,0 | $\begin{gathered} (\mathrm{n}=6) \\ 105,1 \\ 96,5 \end{gathered}$ |
| $\mathbf{M}^{2} \begin{array}{r}\text { mb } \\ \\ \\ \\ \mathrm{db}\end{array}$. | 11,5 10,5 9,1 | 12,1 $(10,4)$ 9,5 | $(11,7)$ | 11,5 11,5 9,8 | $\begin{array}{r} 11,6 \\ 11,0 \\ 9,2 \end{array}$ | $\begin{array}{r} 11,0 \\ 10,5 \\ 9,1 \end{array}$ | 10,9 11,3 9,6 | $\begin{aligned} & 7 \\ & 6 \\ & 6 \end{aligned}$ | 11,5 10,9 9,4 | $\begin{gathered} (\mathrm{n}=6) \\ 95,2 \\ 82,1 \end{gathered}$ | $\begin{aligned} & 17 \\ & 12 \\ & 13 \end{aligned}$ | 11,1 11,1 10,1 | $\begin{gathered} (\mathrm{n}=22) \\ 101,7 \\ 93,1 \end{gathered}$ | 6 6 6 | 12,7 12,4 11,4 | $(n=7)$ 100,3 91,7 | 4 2 2 | 10,3 10,5 8,9 | $\begin{gathered} (\mathrm{n}=5) \\ 101,6 \\ 91,4 \end{gathered}$ |
| $\mathbf{M}^{3} \begin{array}{rr}1 \\ \mathrm{mb} \\ \mathrm{db}\end{array}$. | 10,3 10,2 7,6 | 10,9 $(10,7)$ 7,8 | $\begin{array}{r}10,4 \\ 11,1 \\ 8,3 \\ \hline\end{array}$ | 11,3 11,5 9,5 | $\begin{array}{r} 10,4 \\ 10,4 \\ 7,2 \end{array}$ | $\begin{array}{r} 9,6 \\ 10,4 \\ 7,2 \end{array}$ | 9,8 10,9 7,7 | $\begin{aligned} & 7 \\ & 7 \\ & 7 \end{aligned}$ | 10,4 10,7 7,9 | $(n=7)$ 103,6 76,0 | $\left.\begin{aligned} & 12 \\ & 11 \\ & 10 \end{aligned} \right\rvert\,$ | 11,0 10,7 8,8 | $\begin{gathered} (\mathrm{n}=21) \\ 99,3 \\ 82,8 \end{gathered}$ | 2 1 1 | 13,3 12,8 11,0 | $(n=2)$ 96,0 79,3 | 3 3 3 | 9,6 9,8 7,9 | $\begin{gathered} (n=8) \\ 102,7 \\ 82,1 \end{gathered}$ |
| $\begin{aligned} & \mathbf{P}^{4}-\mathbf{M}^{3} \\ & \mathbf{C}-\mathbf{M}^{3} \end{aligned}$ | 37,1 $(49)$ | $\begin{aligned} & (37,0) \\ & (48) \end{aligned}$ | $\begin{aligned} & 37,5 \\ & 49,6 \end{aligned}$ | － | 38，8 | 35，9 | $\stackrel{35,8}{-}$ | $\begin{aligned} & 6 \\ & 3 \end{aligned}$ | $\begin{gathered} 37,0 \\ (48,9) \end{gathered}$ | － | $\begin{aligned} & 7 \\ & 3 \end{aligned}$ | 36,3 48,6 | － | $\begin{aligned} & 2 \\ & 1 \end{aligned}$ | 39,6 51,7 | － | 3 1 | 33,3 43,1 | － |

${ }^{1}$ Indices comprise males and females including specimens from Maier 1971. $\mathrm{n}=$ number of specimens
$\mathbf{B}=$ average values
$\mathbf{B}=$ tooth breadth．
$\mathbf{L}=$ tooth length．

Table 3. Statistical analysis of the tooth lengths of $\mathrm{P}^{4}, \mathrm{M}^{1}, \mathrm{M}^{2}$ and $\mathrm{M}^{3}$ in the females of Parapapio antiquus and P.broomi

|  | Standard deviation $\mathrm{s}^{2 *}$ |  | Student's t-Test for the sample means* | Degrees of <br> freedom <br> df | Significance probabilities* |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Parapapio antiquus | P. broomi | t |  | t. 99 |  |
| $\mathrm{P}^{4}$ | 0,136 | 0,070 | 3,9 | 18 | 2,878 | O,OI |
| $\mathrm{M}^{1}$ | 0,743 | 0,366 | 1,4 | 19 | 2,86 I | 0,2-0,1 |
| $\mathrm{M}^{2}$ | o, 169 | 0,252 | 1,7 | 22 | 2,819 | O, 1 |
| $\mathrm{M}^{3}$ | 0,345 | 0,226 | 3,0 | 17 | 2,898 | O,OI |

* After Simpson, Roe \& Lewontin 1960.
$M^{3}$ seems to be reduced. The Student t-test proved these differences to be significant for $\mathrm{P}^{4}$ and $\mathrm{M}^{3}$ (Table 3).

The breadths of the premolars and molars being nearly identical in both taxa, the breadth/length index could be expected to express the different degrees of elongation (or reduction). The index also makes possible a comparison with the other species, $P$. jonesi being absolutely smaller and $P$. whitei larger than the previous two. Figure 3 shows the relations diagramatically, the indices of $P$. broomi being expressed as ioo. $P$. jonesi exhibits comparatively high values, especially for $\mathrm{M}^{1}$ and $\mathrm{M}^{3}$, this possibly being a primitive feature. $P$. whitei is very similar to $P$. broomi, showing a slight tendency to elongation, especially in $\mathrm{M}^{3}$. Again, P. antiquus appears to be very aberrant with its gradual increase of the index in mesio-distal direction. The indices for the distal breadths of the molars show that $P$. antiquus possesses the highest degree of reduction of the distal cusps in all molars. Absolutely this reduction is most pronounced in $\mathrm{M}^{3}$, whereas the relative value is lowest for $\mathrm{M}^{2}$.

Judging from tooth size and morphology, $P$. jonesi could tentatively be regarded as the most primitive of the fossil cercopithecids of South Africa, possibly being closely related to their common ancestor. New finds of male skulls, however, show clearly that this taxon was well advanced in some respects: its high degree of sexual dimorphism would exclude it from being a direct forrunner of $P$. antiquus and its well-pronounced maxillary crests and different cranial proportions from being a direct ancestor of both $P$. broomi and $P$. whitei (Maier 1971).

On its teeth alone, however, $P$. jonesi provides a model for understanding the evolutionary alterations within the genus Parapapio. Compared with $P$. jonesi, $P$. antiquus shows $\mathrm{P}^{4}$ and $\mathrm{M}^{1}$ very much elongated; $\mathrm{M}^{2}$ being still longer, while $\mathrm{M}^{3}$ is very similar in both taxa-apart from the conspicuous reduction of the distal cusps in $P$. antiquus, which cannot be understood simply as a consequence of small size.


Fig. 4. Phylogenetic diagram of the genus Parapapio and the hypothetical origin of the genus Papio. The diagram is based on the South African evidence only. P. jonesi occurs at all the mentioned sites; the forms on the left side occur only at Sterkfontein and Makapansgat, those on the right only at Taung, Swartkrans and Kromdraai. Less important sites have been disregarded.
$P$. broomi and $P$. whitei are very similar in tooth indices, underlining the close coincidences in their cranial morphology. Both these taxa are similar to $P$. jonesi in the proportions of $\mathrm{P}^{4}$ and $\mathrm{M}^{2}$, whereas $\mathrm{M}^{1}$ and $\mathrm{M}^{3}$ seem to be distinctly elongated. Summarizing, one can state that $P$. antiquus shows a progressive elongation of the ' $\mathrm{P}^{4} / \mathrm{M}^{2}$-field' of the tooth row, whereas in both $P$. broomi and P. whitei it is mainly the third molar which is increased. In some regards, these conclusions need to be confirmed by additional observations.

These peculiarities of the dentition could possibly be interpreted functionally: in $P$. antiquus, the centre of gravity of the chewing activity is shifted forward as compared with that of the related species. This would necessitate a more oblique direction of the temporal muscle, and would, in turn, explain not only the morphology of the temporal crests but possibly even the low position of the occipital region of the braincase as observed in $P$. antiquus.

## Classification of the genus Parapapio

Present mammalian systems are based mainly on the methods of comparative morphology and this is especially true for fossil forms. The classification of fossil cercopithecoids is fraught with many difficulties, and even that of the extant taxa has not yet been satisfactorily established. External and soft-part characters have proven most valuable for systematic purposes within this superfamily (Pocock 1925), the teeth and the skeletons being very uniform within the whole group (Remane 1960; Schultz 1970). As far as possible, ecological and functional aspects should be considered also and, in the case of fossils, it is important that there be an appreciation of the time factor as well. The present state of knowledge as to the classification of the Cercopithecoidea was discussed recently by the present author (Maier 1970).

All attempts to obtain absolute data on the ages of the South African australopithecine caves have so far been unsuccessful (Tobias \& Hughes 1969). However, recent finds have resulted in surprising changes of the chronology of the North and East African Pliocene and Pleistocene fossil sites. Olduvai Bed I has been dated at about $\mathrm{I}, 8 \mathrm{~m} . \mathrm{y}$., while the deposits at Omo, Kaiso, Kanapoi, Koobi Fora, Chemeron and Laetolil might reach back some 3 to 4 m.y. (Maglio 1970). These sites with their varied fossil faunas provide a good basis for comparison with the richly fossil-bearing cavern breccias in southern Africa, but not much work has so far been done in this field. Hendey (1970) has pointed out the major difficulties in comparing South African fossil faunas at the present stage of knowledge.

Whereas Kurtén (ig6o, ig68) suggested a Middle Pleistocene age for the South African ape-man cave deposits, we must now contemplate a Lower Pleistocene age for them. Ewer (1963:343) reviewed the then available evidence and concluded that 'Kromdraai and Swartkrans may correspond to the gap between [Olduvai] I and II, while Makapan and Sterkfontein belong to the period covered by the older deposits of Olduvai I and Omo'. This tentative
correlation was repeated by the same author in 1967. According to Cooke (pers. comm.) the pigs and elephants from Makapansgat compare well with forms from Kanapoi and Lower Omo beds, indicating an absolute age of some $2,5-3,5 \mathrm{~m} . \mathrm{y}$. for that site. Sterkfontein is estimated by Cooke to be about 2,5 m.y. and Swartkrans about 2,0 m.y. As in East Africa, the Pleistocene sites of South Africa will, most probably, have to be dated further back than was previously thought.

This evidence, so far based mainly on Suidae, Elephantidae and Carnivora, seems to be supported by the primate evidence. (Unfortunately, the rich East African cercopithecoid material has not yet been described comprehensively, but Mrs. Meave Leakey of the Kenya National Museums will shortly publish a monograph.) Arambourg (1947) recorded Dinopithecus brumpti from the Omo deposits, Butzer (197I) added Colobus sp., Cercopithecus sp., Parapapio sp., Papio sp. and Simopithecus sp. from this site. R. E. F. Leakey (1969) described Papio baringensis from the Chemeron Beds which shows much similarity with Papio robinsoni, and in 1970 the same author recorded from Koobi Fora ( $\pm 2,5$ m.y.) Cercopithecus sp., Papio sp. and Simopithecus sp. According to L. S. B. Leakey (1965), large forms of Papio and Simopithecus are known from Olduvai Beds I-IV. As far as I could see during a recent visit to the Kenya National Museums, Nairobi, the genus Parapapio occurs as well, both at Olduvai and Koobi Fora, the material most probably belonging to the species $P$. jonesi. A mandible from Kanapoi ( $\pm 4 \mathrm{~m} . \mathrm{y}$.) has recently been referred to this species as well (Patterson i968). A few small teeth from Lothagam would fit approximately some specimens from the 'grey breccia' of Makapansgat, being referred also to $P$. jonesi. Most of these better known sites seem to have three cercopithecid forms side by side: a small Parapapio, a large Papio (Dinopithecus and Gorgopithecus possibly being only synonyms) and a very large Simopithecus.

The South African cave deposits show a different arrangement, which may, however, be due partly to geographical separation and a different mode of deposition (Ewer 1967). The older sites at Makapansgat and Sterkfontein have so far yielded only various types of Parapapio and a comparatively small and primitive Simopithecus (Maier, in press), but no true baboon of the genus Papio whatsoever (Freedman 1957). Very small and primitive forms of baboons appear only at Taung, while, besides Parapapio and Simopithecus, Papio is abundantly represented in the younger sites of Swartkrans and Kromdraai. The fossil colobids of both East and South Africa are too different for useful comparisons to be made.

Pending more detailed information about the cercopithecoid material from the Lower Pleistocene of East Africa, the preliminary evidence seems to suggest rough contemporaneity of the more important South African faunas.

The faunal comparison of the South African sites is complicated by their geographical distance and by evident palaeo-ecological differences in the surroundings of the ancient deposits (Ewer 1956a). Thus, at Makapansgat the environment was probably more varied and less dry (Ewer 1956b; Wells 1967),
whereas Taung 'was distinctly more desert-like than . . . the other deposits' (Ewer 1957: 139). Zoogeographically Makapansgat shows more affinities with central Africa than the other sites. Considering these difficulties, Ewer (1957: 141) concluded: 'The probable time sequence of the deposits is Sterkfontein and Makapansgat close together, with the former very probably being the earlier; then Swartkrans and lastly Kromdraai, while the Taung deposit is most probably closest in time to Sterkfontein and Makapan.' From the morphological evidence of the cercopithecoids, Freedman (1957) considered the Taung deposit to be the oldest, followed in order by Sterkfontein, Makapansgat, Swartkrans and Kromdraai. Wells (1967) and Cooke (1967) seem to assume that Taung is slightly younger, the latter author giving a sequence Makapansgat, Sterkfontein and Taung for his 'Sterkfontein Faunal Span' (see his Table I).

Recently Wells (ig69) more clearly expressed his conviction that Makapansgat may be earlier than Sterkfontein, whereas Taung may be even closer to the Swartkrans-Kromdraai 'faunal span'. It seems to be very necessary that the newly prepared elephant material from Makapansgat be studied by experts who are well acquainted with the East African forms.

Freedman (1957: 248) stated that the more important South African fossil sites originate from 'a geologically short period just following the PlioPleistocene boundary', the time of depositing between the oldest (Taung) and the youngest (Kromdraai and Cooper's) breccias not being longer than about 250 ooo years. As, according to Simpson (1944), the minimum time span for the evolution of a new species amounts to about 0,5 m.y., Freedman (1957: 244) concluded: 'Therefore . . . it seems quite obvious that the faunal changes between the sites could not be due to in situ evolution' and that 'it would therefore seem that the most obvious and probable cause of the faunal replacements was successive migrations into and out of the areas as a result of local and/or distant environmental changes'. The recently suggested evidence of the very great age and long duration of these deposits would yield, however, a satisfactory temporal frame to explain the evolution and radiation of the numerous Pleistocene Cercopithecoidea in South Africa, without entirely discarding the possibility of some faunal shifting.

The existing classification of the genus Parapapio was elaborated in the studies of Broom (1940) and Freedman (1957). Based mainly on the occurrence of different-sized molars, these authors established four species: the small-sized and, as it appears now, widespread Parapapio jonesi, two medium-sized forms, $P$. antiquus and $P$. broomi, which 'are remarkably similar in tooth size but differ very considerably in skull shape' (Freedman 1957: 158), and finally the largesized $P$. whitei.

The small Parapapio jonesi could easily represent the generalized common ancestor of the Papionini sensu stricto. Its small teeth are unspecialized as compared with progressive features in the other species of Parapapio. Occurring in all of the australopithecine caves of South Africa, this taxon has now been recorded from various places in East Africa as well, probably covering some
${ }^{2-3}$ m.y. of the Pleistocene fossil record. New finds have shown, however, that the male skull is quite advanced, although retaining its primitive teeth (Maier 1971).

In cranial shape, tooth specialization and the small degree of sexual dimorphism, Parapapio antiquus seems to differ more from $P$. jonesi than does $P$. broomi, and might thus be an earlier offshoot, possibly being somehow adapted to the drier ecological conditions prevailing at Taung. Such an environment could also have stimulated the evolution of the small true baboons Papio wellsi and P. izodi: 'Parapapio antiquus is very similar in size and dental morphology to Papio izodi, and these two species may represent a morphological stage not far from the point at which the genera Parapapio and Papio started diverging from a common stem' (Freedman 1957: 245).

The nature of the molar specializations and the very similarly elongated male crania indicate a monophyly of both Parapapio broomi and P. whitei. The teeth of the latter species seem to be relatively larger than in the similarly sized P. broomi. Further material may close the existing size gap, but since both forms occur in the same blocks of the 'Upper Phase I' breccia of Makapansgat, they cannot form a chronocline. As we do not know their postcranial skeleton, it is not possible at the moment to assign different ecological niches to these apparently sympatric species. In Figure 4 an attempt is made to plot the evidence in the form of a phylogenetic diagram.

## Summary

Two new female skulls of the fossil cercopithecid Parapapio antiquus from Taung, Cape Province, South Africa, are described and discussed. The teeth of this species especially show some significant differences from the other three species of Parapapio. A phylogenetic arrangement of the genus is suggested.

## Acknowledgements

In the first place, I have to thank Mr. J. W. Kitching for placing his material at my disposal; I owe thanks to Miss J. Roets and Mr. B. Maguire for much help with the manuscript and to Mr. J. Henderson for help with the statistics.

## References

Arambourg, C. 1947. Contribution à l'étude géologique et paléontologique du bassin du Lac Rodolphe et de la Basse Vallée de l'Omo. $2^{\text {e }}$ partie: Paléontologie. Mission scient. Omo 1: 231-562.
Broom, R. 1940. The South African Pleistocene cercopithecid apes. Ann. Transv. Mus. 20:89-100. Butzer, K. W. 1971. The lower Omo Basin: geology, fauna and hominids of Plio-Pleistocene formations. Naturwissenschaften 58:7-16.
Cooke, H. B. S. 1967. The Pleistocene sequence in South Africa and problems of correlation. In bishop, w. W. \& clark, J. D., eds. Background to evolution in Africa: 175-184. Chicago; London: University of Chicago Press.

EWER, R. F. 1956a. The dating of the Australopithecinae: faunal evidence. S. Afr. archaeol. Bull. 11: $4^{1-45 .}$
Ewer, R. F. 1956b. The fossil carnivores of the Transvaal caves: two new viverrids, together with some general considerations. Proc. zool. Soc. Lond. 125: 259-274.
Ewer, R. F. 1957. Faunal evidence on the dating of the Australopithecinae. In Clark, J. D., ed. Third Pan-African Congress on Prehistory, Livingstone 1955: 135-142. London: Chatto \& Windus.
Ewer, R. F. 1963. The contribution made by studies of the associated mammalian faunas. S. Afr. F. Sci. 59: 340-347.

Ewer, R. F. 1967. The fossil hyaenids of Africa - a reappraisal. In bishop, w. w. \& clark, J. D., eds. Background to evolution in Africa: 109-123. Chicago; London: University of Chicago Press.
Freedman, L. 1957. The fossil Cercopithecoidea of South Africa. Ann. Transv. Mus. 23: 121-262.
Freedman, L. 1961. New cercopithecoid fossils, including a new species, from Taung, Cape Province, South Africa. Ann. S. Afr. Mus. 46: 1-14.
Freedman, L. 1965. Fossil and subfossil primates from the limestone deposits at Taung, Bolt's Farm and Witkrans, South Africa. Palaeont. afr. 9: 19-48.
Gear, J. H. S. 1926. A preliminary account of the baboon remains from Taungs. S. Afr. F. Sci. 23: 731-747.
Haughton, S. H. 1925. A note on the occurrence of a species of baboon in limestone deposits near Taungs. Trans. R. Soc. S. Afr. 12: lxviii.
Hendey, Q.B. 1970. A review of the geology and palaeontology of the Plio-Pleistocene deposits at Langebaanweg, Cape Province. Ann. S. Afr. Mus. 56: 75-117.
Jones, T. R. 1937. A new fossil primate from Sterkfontein, Krugersdorp, Transvaal. S. Afr. $\mathcal{F}$. Sci. 33: 709-728.
Kurtén, B. 196o. The age of the Australopithecinae. Stockh. Contr. Geol. 6: 9-22.
Kurtén, B. 1968. Dating the early stages of hominid evolution. In кurth, g., ed. Evolution und Hominisation: 75-81. Stuttgart: Fischer.
Leakey, L. S. B. 1965. Olduvai Gorge, 1951-61. Cambridge: University Press.
Leakey, R. E. F. 1969. New Cercopithecoidea from the Chemeron Beds of Lake Baringo, Kenya. Fossil Vert. Afr. 1: 53-69.
Leakey, R. E. F. i970. Fauna and artefacts from a new Plio-Pleistocene locality near Lake Rudolf in Kenya. Nature, Lond. 226: 223-224.
Maglio, V. J. 1970. Early Elephantidae of Africa and a tentative correlation of African PlioPleistocene deposits. Nature, Lond. 225: 328-332.
Maier, W. 1970. Neue Ergebnisse der Systematik und der Stammesgeschichte der Cercopithecoidea. Z. Säugetierk. 35: 193-214.
Maier, W. 1971. New fossil Cercopithecoidea from the Pleistocene cave deposits of the Makapansgat Limeworks, South Africa. Palaeont. afr. 13: 69-107.
Maier, W. The first complete skull of Simopithecus darti from Makapansgat, and its systematic position. 7. hum. Evolution. (In press.)
Patterson, B. 1968. The extinct baboon Parapapio jonesi in the early Pleistocene of northwestern Kenya. Breviora 282: 1-4.
Peabody, F. E. 1954. Travertines and cave deposits of the Kaap Escarpment of South Africa, and the type locality of Australopithecus africanus Dart. Bull. geol. Soc. Am. 65: 671-706.
Pocock, R. I. 1925. The external characters of the catarrhine monkeys and apes. Proc. zool. Soc. Lond. 1925: 1479-1579.
Remane, A. i96o. Zähne und Gebiss. In hofer, h., schultz, A. h. \& starck, A., eds. Primatologia. Handbook of primatology. 3: 637-846. Basel; New York: Karger.
Schultz, A. H. 1970. The comparative uniformity of the Cercopithecoidea. In napier, J. r. \& napier, p. h., eds. Old world monkeys: 39-51. New York; London: Academic Press.
Scott, J. H. 1967. Dento-facial development and growth. London; New York: Pergamon Press.
Simpson, G. G. 1944. Tempo and mode in evolution. New York: Columbia University Press.

Simpson, G. G., Roe, A. \& Lewontin, R. C. 196o. Quantitative zoology. Rev. ed. New York; Burlingame: Harcourt, Brace.
Tobias, P. V. \& Hughes, A. R. 1969. The new Witwatersrand University excavation at Sterkfontein. S. Afr. archaeol. Bull. 24: 158-169.
Wells, L. H. 1967. Antelopes in the Pleistocene of southern Africa. In bishop, w. w. \& clark, J. D. eds. Background to evolution in Africa: 99-107. Chicago; London: University of Chicago Press.
Wells, L. H. 1969. Faunal subdivision of the Quaternary in southern Africa. S. Afr. archaeol. Bull. 24: 93-95.


Parapapio antiquus M. 3078 (female) lateral and basal view. Note the straight contour of the muzzle dorsum, the low position of the occiput and the reduction of the third molar. Scale unit 10 mm .

# INSTRUCTIONS TO AUTHORS 

## Based on

CONFERENCE OF BIOLOGICAL EDITORS, COMMITTEE ON FORM AND STYLE. 1960.
Style manual for biological journals. Washington: American Institute of Biological Sciences.

## MANUSCRIPT

To be typewritten, double spaced, with good margins, arranged in the following order: (1) Heading, consisting of informative but brief title, name(s) of author(s), address(es) os author(s), number of illustrations (plates, figures, enumerated maps and tables) in the article. (2) Contents. (3) The main text, divided into principal divisions with major headings; subheadings to be used sparingly and enumeration of headings to be avoided. (4) Summary. (5) Acknowledgements. (6) References, as below. (7) Key to lettering of figures. (8) Explanation to plates.

## ILLUSTRATIONS

To be reducible to $12 \mathrm{~cm} \times 18 \mathrm{~cm}$ ( 19 cm including caption). A metric scale to appear with all photographs.

## REFERENCES

Harvard system (name and year) to be used: author's name and year of publication given in text; full references at the end of the article, arranged alphabetically by names, chronologically within each name, with suffixes $a, b$, etc. to the year for more than one paper by the same author in that year.
For books give title in italics, edition, volume number, place of publication, publisher.
For journal articles give title of article, title of journal in italics (abbreviated according to the World list of scientific periodicals. 4 th ed. London: Butterworths, 1963), series in parentheses, volume number, part number (only if independently paged) in parentheses, pagination.
Examples (note capitalization and punctuation)
Bullough, W. S. 196o. Practical invertebrate anatomy. 2nd ed. London: Macmillan.
Fischer, P.-H. i948. Données sur la résistance et de le vitalité des mollusques. 7. Conch., Paris 88: 100-140.
Fischer, P.-H., Duval, M. \& Raffy, A. i933. Etudes sur les échanges respiratoires des littorines. Archs Zool. exp. gén. 74: 627-634.
Kohn, A. J. 1960a. Ecological notes on Conus (Mollusca: Gastropoda) in the Trincomalee region of Ceylon. Ann. Mag. nat. Hist. (13) 2: 309-320.
Kohn, A. J. ig6ob. Spawning behaviour, egg masses and larval development in Conus from the Indian Ocean. Bull. Bingham oceanogr. Coll. 17 (4): 1 -51.
Thiele, J. i9io. Mollusca: B. Polyplacophora, Gastropoda marina, Bivalvia. In schultze, l. Zoologische und anthropologische Ergebnisse einer Forschungsreise im westlichen und zentralen SüdAfrika. 4: 269-270. Jena : Fischer. Denkschr. med-naturw. Ges. Jena 16: 269-270.

## ZOOLOGICAL NOMENCLATURE

To be governed by the rulings of the latest International code of zoological nomenclature issued by the International Trust for Zoological Nomenclature (particularly articles 22 and 51). The Harvard system of reference to be used in the synonymy lists, with the full references incorporated in the list at the end of the article, and not given in contracted form in the synonymy list.

## Example

Scalaria coronata Lamarck, 1816: pl. 451, figs $5 a, b$; Liste: 1 1. Turton, 1932: 80.

# ANNALS OF THE SOUTH AFRICAN MUSEUM ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM 

Volume 59 Band<br>December 1971 Desember<br>Part 2 Deel



## THE EARLY THERAPSIDS

By
L. D. BOONSTRA

# The ANNALS OF THE SOUTH AFRICAN MUSEUM <br> are issued in parts at irregular intervals as material becomes available 

Obtainable from the South African Museum, P.O. Box 61, Cape Town

## Die ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM

word uitgegee in dele op ongereelde tye na beskikbaarheid van stof

Verkrygbaar van die Suid-Afrikaanse Museum, Posbus 61, Kaapstad

## OUT OF PRINT/UIT DRUK

1, $2(1,3,5,7-8), 3(1-2,5$, t.-p.i. $), 51-2,5,7-9)$, $6(\mathrm{I}, \quad$ t.-p.i. $), \quad 7(\mathrm{I}-3), \quad 8, \quad 9(\mathrm{I}-2), \quad \mathbf{1 0}(\mathrm{I})$, II (1-2, 5, 7, t.-p.i.), 2I, 24(2), 27, $31(1-3), 33,38$

Price of this part/Prys van hierdie deel

$$
\mathrm{R}_{3,30}
$$

## Trustees of the South African Museum (C) Trustees van die Suid-Afrikaanse Museum 1971

Printed in South Africa by The Rustica Press, Pty., Ltd, Court Road, Wynberg, Cape

In Suid-Afrika gedruk deur
Die Rustica-pers, Edms., Bpk.
Courtweg, Wynberg, Kaap

# THE EARLY THERAPSIDS 

By<br>L. D. Boonstra<br>South African Museum, Cape Town<br>(With 3 figures)<br>[MS. accepted 8 September 1971]

## Contents



## Introduction

As early therapsids I consider those forms that have been recovered from Zones I and II of the Russian succession and from the Tapinocephalus zone of the Beaufort beds of the Karroo System. They thus range from the top of the Lower Permian to the end of the Middle Permian.

In this paper I am stressing, firstly, the great diversity of forms with which we are so suddenly confronted in one of the many explosive faunal developments that have so repeatedly occurred during the long history of animal life and apparently gainsaying the dictum natura non facit saltum.

Secondly, I shall attempt to arrange this assemblage into a number of morphological series, before attempting a phylogenetic arrangement, because I feel that this is a safer procedure, bearing in mind that this fauna with which we are confronted as a fait accompli consists of contemporaries and geologically speaking of the same age with no one the ancestor of any other.

Thirdly, I shall consider the derivation of these therapsids from the antecedent fauna of pelycosaurs with which in similar fashion we are confronted in Carboniferous-Permian times.

## Diversity

Of the pre-Upper Permian assemblage of therapsids we know at least 70 well-established genera which have on taxonomic criteria been brigaded into I9 families. This explosive radiation is much greater than the earlier Carboni-ferous-earlier Permian radiation of the pelycosaurs with its 8 families.

In the early therapsids the size variation is as much as that between a rat and a hippopotamus, with weights from about 500 g to two tons. In shape they vary from light and slender to massive and plump. Some are agile, others ponderous. There are long as well as short tailed forms, long snouted and extremely short snouted species lived side by side; locomotion varied from slinking, walking to running, with the body slung between the spread-eagled limbs or carried fairly high on more upright supports. A few were insectivorous, some carrion eaters, others predaceous carnivores and many herbivores; some feeding on soft marsh plants, whereas others, roaming on to higher ground, subsisted on more fibrous shrubs. In one group the teeth were largely replaced by horny sheaths analogous to those of tortoises.

This diverse assemblage has been classified into the following 19 families:
I. Eotitanosuchidae
2. Brithopidae
3. Anteosauridae
4. Titanosuchidae
5. Tapinocephalidae
6. Styracocephalidae
7. Estemmenosuchidae
8. Phthinosuchidae
9. Hipposauridae
io. Galesuchidae
i I. Otsheriidae
12. Venyukoviidae
13. Dromasauridae
14. Endothiodontidae
15. Dicynodontidae
16. Alopecodontidae
17. Pristerognathidae
18. Lycosuchidae
19. Scaloposauridae

The various authors who have established these 19 discrete families have done so on the basis of determined differences of a structural nature.

These differences, although often considerable, are also limited, and it is because of these limitations accompanied by certain basic similarities and trends that these families have been brigaded into one order-the Therapsida.

A profitable evaluative discussion can best be started by considering firstly those points of basic similarity and then the extent of the differences and variations.

Structural features common to all the known early therapsids are:
I. A single temporal fenestra lying below the posterior process of the postorbital, but the participation of this process and of the various other bones forming the temporal border varies.
2. The pterygoids tend to meet in the middle line behind the interpterygoid vacuity, which is thereby variously reduced, and applied to the basicranium with consequent loss of the primitive freely movable joint and the development of a longitudinal basicranial girder.
3. The jaw articulation never lies in a plane posterior to that of the occipital condyle and the anteriorly directed slope of the occiput is reduced, often becoming nearly vertical or even sloping backwards.
4. There is always a reflected lamina of the angular.
5. The septomaxilla, with a foramen, has a more or less well-developed lateral facial exposure but small in Otsheria and brithopids and phthinosuchids and not exposed laterally in dicynodonts.
6. There is no supratemporal and the anterior coronoid is always lost and sometimes both are absent.
7. The lacrimal never reaches the nostril.
8. The maxilla is always deep.
9. The squamosal flares out laterally and posteriorly to various degrees.
10. The quadrate ramus of the pterygoid reaches the quadrate.
iI. The vertebrae are amphicoelous and there are no dorsal intercentra.
12. The girdles and limbs are adapted for an early stage of a quadrupedal gait. The main adaptations are: loss of the supraglenoid buttress and foramen, loss of one central in the tarsus and one distal, number of phalanges reduced to varying degree, the glenoid is reduced in length and the humerus untwisted to varying degrees, the iliac blade is heightened, and an anterior process developed to varying degrees, the femur loses the Y system of ridges and develops a greater trochanter.

## Strugtural variations

## TEMPORAL FENESTRA

It is obvious that the origin of the m. capiti-mandibularis in the captorhinomorphs (and all other anapsids) could only have been from the inner or under surface of the temporal or cheek bones. Fox has reported that in a captorhinid examined by him the central part of the temporal covering is composed of very thin bone with a concomitant thickening peripherally of this weak area and infers that the attachment of the muscle was mainly on the thickened parts and that the thinning centrally was due to this area becoming non-functional and thus liable to fenestration.

Such fenestration has in fact taken place in the pelycosaurs. The area of bone-resorption is mainly situated at the junction of squamosal and jugal, but the fenestration in the pelycosaurs has been far from uniform. In fact in one species of Ophiacodon there is not a single but a double fenestra. Moreover, there
is considerable variation in the participation in the border of the fenestra by the bones of the cheek. The dorsal border or upper temporal arch is always formed by the postorbital and the squamosal. The lower border or zygomatic arch is mostly formed by the jugal and squamosal in varying proportions, but in all the three suborders there are forms in which the quadratojugal enters the lower border of the fenestra. Is this of sufficient importance to query the homology of the temporal fenestra?

The temporal fenestrae of the pelycosaurs lie mainly laterally in the cheeks and are separated from one another by a broad flat intertemporal skull table. In the pelycosaurs the original capiti-mandibularis has divided into a major medial mass, the temporal and a lateral mass, the masseter. The temporal originated from the inner face of the bones above the fenestra, viz. parietal, postorbital, postfrontal and squamosal. The masseter arose from the inner face of the zygomatic arch, i.e. from the jugal and squamosal lying below the fenestra. The function of the fenestra is undoubtedly to enlarge the adductor chamber for the bulging of the temporal muscle during contraction.

The temporal fenestra of the earliest therapsids, apparently homologous to that of the higher sphenacodonts, when first encountered already shows a number of modifications in divergent directions.

In Hipposaurus the fenestrae are still small and are separated by a wide intertemporal table, the posterodorsal flange of the postorbital, meeting the squamosal, lies in a horizontal plane with the temporal muscle originating, in part, from its ventral face.

In the early Galesuchidae the fenestra is larger both in length and width, but otherwise essentially as in the higher sphenacodonts.

In Phthinosuchus the fenestra is greatly enlarged both in height and length due to the outflaring of the squamosal laterally as well as posteriorly. (It extends forward into the jugal.) The postero-dorsal flange of the postorbital lying horizontally is, however, shortened and laterally flanked by a horizontally disposed lappet of the squamosal which on its ventral face provides a large area of the origin for the temporal muscle.

In Eotitanosuchus the fenestra is enlarged, extending forward into the jugal, and a groove on the outer edge of the postorbital indicates that the temporal in part arose from the lateral face of this bone.

In the Brithopidae, and even more so in the Anteosauridae, the fenestra is enlarged by a lateral as well as a posterior outflaring of the squamosal which greatly increases the size of the adductor chamber. The dorsal flange of the postorbital now shows a well-developed lateral face and the temporal muscle now arose in part from a ridge on the dorsal edge of the postorbital, confluent with a postero-lateral edge on the squamosal. Moreover, in the Brithopidae and Anteosauridae, the intertemporal skull table is much reduced in width and the original horizontally lying upper face of the postorbital now faces appreciably laterally and is practically excluded from the skull table.

In the Titanosuchidae the temporal fossa is only of moderate size. The
posterior flange of the postorbital, lying mainly vertical and applied to the outer face of the parietal, is greatly reduced and lying low down in the skull reaches the squamosal as a tapering splint.

The intertemporal skull table is reduced in width and the parietals form a fairly wide and fairly high sagittal crista.

The origin of the temporal muscle màss is mainly from the outer face of the posterior postorbital flange and extends up the lateral face of the parietal to the edge of the crista and posteriorly to the upper part of the squamosal, whose lateral edge is continued as a ridge on to the parietal.

The jugal is excluded from the fairly deep lower temporal arch, apparently due to the downgrowth of the strong postorbital.

In the Tapinocephalidae the greatly varying pachyostotic thickening of the skull bones affects the nature of the temporal fenestra, the adductor chamber and the degree of participation of the bones forming the borders of the fenestra.

As to position of the fenestra, the one extreme is seen in Riebeeckosaurus where the two fenestrae are separated by only a sharp parietal crista; in the other extreme the intertemporal width is so great in Criocephalus that the fenestra is not visible in dorsal view. In all the lower arch, formed solely by the squamosal, is very deep so that the fenestra is situated high up in the cheek. In general the strong postorbital bar makes the distance between orbit and fenestra great. As to shape, the fenestra is slitlike in some moscopines and tapinocephalines with the fore-aft diameter one-third of the dorso-ventral, whereas in Avenantia it is longer than high, with the struthiocephalines in an intermediate position.

The adductor chamber is roomy in Avenantia, moderately so in the struthiocephalines but antero-posteriorly compressed in the tapinocephalines and in Moschops and Criocephalus. As to the circum-fenestral bones, the Tapinocephalidae have one feature in common in that the jugal is wholly excluded from the lower arch, being pushed anteriorly by the thick postorbital bar and the forward growth of the squamosal due to the quadrate moving anteriorly.

In the tapinocephalids considerable variations occur in the upper temporal arch. In some of the struthiocephalines (where the pachyostosis is moderate) the dorso-posterior flange of the postorbital and the upper flange of the squamosal do not meet, being thus separated by the parietal. In the other tapinocephalids (where the pachyostosis is greatly developed) the junction of the postorbital and squamosal is pushed down to the lower half of the fenestra.

In the tapinocephalines (where the pachyostosis is great) abnormal overgrowth of both the frontal and postfrontal bones caused these bones to enter the dorso-anterior part of the rim of the fenestra.

In the moschopines (where the pachyostosis is in some respects even greater) only the postfrontal enters the dorso-anterior part of the rim of the fenestra. In all the tapinocephalids the main origin of the temporal muscle
mass is from the lateral surfaces of those parts of the parietal, postorbital and squamosal lying well within the upper part of the temporal fossa.

The rim of the fenestra thus lies lateral to the area of origin, i.e. the more fibrous part of the temporal muscle, and any bulging of the muscle mass on contraction could hardly have occurred through the fenestra, which is in any case small. The body or fleshy part of the muscle lies lower down and is covered by the deep lower arch (squamosal). The fenestra thus seems to have lost its primary function! The forward position of the lower jaw articulation with the concomitant great depth of the squamosal arch greatly lengthens the muscle mass and this increased length would compensate for a decreased ability to bulge locally.

In the Styracocephalidae the pachyostosis has caused a great reduction of the size of the temporal fenestrae, which are situated widely apart. Differential bone thickening has resulted in the rim of the fenestra being formed solely by the postorbital and squamosal.

A forward shift of the jaw articulation as in the tapinocephalids with the deep squamosal low arch has affected the working of the temporal muscles as described above for the tapinocephalids.

In the Estemmenosuchidae the fenestra is large particularly in length due to a forward extension into the jugal as well as a posterior outflaring of the squamosal. The dorsal flange of the postorbital lying in the skull table is shortened and fails to reach the squamosal. The intertemporal width is large. The temporal muscle thus in part originates from the lateral face of the parietal.

In the Otsheriidae the temporal fenestra is large, due to the outflaring of the squamosal both laterally and posteriorly as well as the reduction of the width of the intertemporal skull table. It is still primitively bounded by the three bones - postorbital, squamosal and jugal, but both the upper and the lower arch are modified.

In the upper arch the posterior flange of the postorbital only provides a narrow edge to bound the upper border of the fenestra and the temporal muscles arise in part from the latero-ventral edge of this splint-like flange.

The lower arch is fairly shallow but is deeper than broad, thus lying vertically, with a large contribution from the jugal. The postero-ventral corner of the squamosal is prolonged ventrally in the form of a pedicel to hold the quadrate in a position low down in the skull and also far posteriorly. The adductor chamber is roomy and the temporal muscles short but bulky.

In the Dromasauridae a single skull of Galeops from the Tapinocephalus zone is inadequately known.

The temporal fossa is short but deep and apparently bounded by the postorbital, squamosal and jugal.

The squamosal has a long ventrally directed pedicel similar to that of Otsheria.

In the Endothiodontidae and Dicynodontidae the oldest known forms from low down in the Tapinocephalus zone already have the temporal region,
which is so typically unique for all the Dicynodontia and basically retained throughout the long history of this group.

Of all the early therapsids the Endothiodontidae and Dicynodontidae show the greatest modification in the temporal region from the primitive pelycosaur condition.

The temporal fenestra is greatly enlargèd. The fore-aft diameter is uniquely lengthened due to the anterior position of the orbit accompanied by the slenderness of the postorbital bone and the posterior flaring of the squamosal. The medio-lateral diameter is enlarged due to the reduction of the intertemporal width of the skull table.

The upper postorbital-squamosal arch is fairly primitive except for the greatly lengthened dorsal flange of the postorbital which is somewhat bent down laterally from the horizontal. The temporal muscle arises in part from the lateral edge and under surface of the postorbital and the upper edge of the squamosal.

The greatest modification is seen in the structure of the squamosal.
In the zygoma the squamosal, originally lying in a vertical plane, is bent down laterally to lie in a horizontal plane with the original dorsal edge now forming the lateral edge of the bar. In addition the squamosal extends far anteriorly to terminate in a plane ventral to the orbit and the jugal is almost completely excluded from the lateral face of the zygoma.

The downward growth of the postero-ventral corner of the squamosal to form a pedicel, first seen in Otsheria, is also greatly modified. In Otsheria the face of this pedicel is lateral. In the Dicynodontia this face is now directed much anteriorly with the original posterior edge turned outwards to form a sharp lateral edge. To the lower part of this oblique face the quadratojugal is applied. Above the quadratojugal is the area of origin of the masseter mainly from fascia attached to the sharp lateral squamosal edges.

In all the four early therocephalian families (Pristerognathidae, Lycosuchidae, Alopecodontidae and Scaloposauridae) the temporal fenestra is large and faces more dorsally than laterally and the adductor chamber is very roomy. Here also the posterior flange of the postorbital is greatly reduced and lies as a small splint lying vertically and applied to the lateral face of the parietal, which now forms the greatest part of the upper border of the temporal fenestra.

The intertemporal width is greatly reduced and this part of the skull table is normally developed into a sagittal crista of varying width and height.

Here the temporal muscle had its main origin from the lateral face of the parietal.

## POSITION OF THE QUADRATE

The foregoing comparison of the temporal fenestra and its arches in the early therapsids drew our attention to the origin of muscles of the capitimandibularis mass.

The function of these adductors is related to the position of the jaw articulation and the insertion on the lower jaw. These two aspects will now be considered. In the pelycosaurs the quadrate is situated far posteriorly just posterior to the plane of the occipital condyle.

In the earliest Gorgonopsia the quadrate lies just anterior to the plane of the occipital condyle.

In the Eotitanosuchidae the quadrate apparently lies in the plane of the condyle.

In the Brithopidae the quadrate has shifted somewhat anteriorly to the plane of the condyle.

In the Anteosauridae the quadrate has shifted still further anteriorly and due to the backward tilt of the occiput lies very far anterior to the upper edge of the occiput.

In the early therocephalian families the quadrate still lies in the primitive posterior position.

In the Titanosuchidae, Tapinocephalidae, Styracocephalidae and Estemmenosuchidae the quadrate lies very far forward of the plane of the condyle and still more of the plane of the upper occipital edge.

In the Otsheriidae the quadrate would appear to have been situated somewhat anterior to the plane of the condyle.

This is also the position of the quadrate in the early Endothiodontidae and Dicynodontidae.

## INSERTIONS OF THE TEMPORAL MUSCLE

The primitive nature of the insertions of the m . capitimandibularis is still evident in the sphenacodonts and this condition is basically retained in the early therapsids.

The most significant change is seen in the Gorgonopsia and the Therocephalia, where the dentary developed a prominent free-standing dorsoposteriorly directed coronoid process for the reception of the temporalis.

No forms are known in which this development is incipient. Low down in the Tapinocephalus zone it is simply there fully developed in the oldest Gorgonopsia and Therocephalia.

As has already been mentioned above, the origin of the subdivided capitimandibularis is in the early Gorgonopsia still of primitive nature, but that in the earliest therocephalians it is already highly specialized in a mammalian direction. This very definite difference in origin of the muscles is remarkably not accompanied by any noteworthy change in the insertion.

The development of the coronoid process in these two groups thus appears to have been caused by a pull exerted by the adductors in a primitive way in the case of the gorgonopsians on the one hand and by an advanced mammallike way in in the therocephalians.

The presence of a coronoid in the Gorgonopsia can thus at most be considered as a parallel development and not one in a mammalian direction.

## THE REFLECTED LAMINA OF THE ANGULAR

This structure is a feature common to all the early therapsids and is concerned with the insertion of the anterior pterygoid and superficial masseter muscles.

This is also the condition in the higher sphenacodonts and held as strong evidence of their consanguinity with the therapsids.

In the early Dicynodontia the structure of the reflected lamina differs somewhat from that of the other early therapsids. This is probably associated with a difference in the origin of the anterior pterygoid muscle for we know that in the early Endothiodontidae and Dicynodontidae, but not in the Otsheriidae, the lateral pterygoidal flange is greatly reduced.

## THE MARGINAL TEETH

Of the oldest therapsid families, io have a carnivorous dentition and 7 are herbivorous, with the adaptations showing a quite remarkable diversity.

In the primitive pelycosaurs the tooth row is long and consists of simple pointed teeth. In the maxilla a pair of teeth well back in the row are enlarged as 'canines'. The replacement is distichial.

In the advanced sphenacodonts the enlarged 'canines' are situated near the front of the maxillary row. The functional replacement is by a member of the same tooth family but the upper canines are replaced alternately.

In the early therapsids the tooth row is reduced in all the carnivorous families, but is secondarily lengthened in the herbivorous Titanosuchidae, Tapinocephalidae and Styracocephalidae. In the Otsheriidae and Venyukoviidae the row is still fairly long but highly specialized. In the Endothiodontidae and Dicynodontidae development of horny sheaths radically reduces the marginal teeth.

In the early therapsids the upper canine when present is the first tooth in the maxilla in all the families, except the Scaloposauridae and Alopecodontidae.

A lower canine is present, except where secondary lost as in the herbivorous Endothiodontidae, Dicynodontia and Tapinocephalidae but persists in the herbivorous Titanosuchidae and Styracocephalidae. In the Lycosuchidae there are a pair of upper canines replaced alternately but functionally by a member of the same family.

In the pelycosaurs there appears to be no limit to the tooth replacement. This is also the case in the Titanosuchidae and probably also in the Tapinocephalidae. In the other families there is evidence of limited replacement in the Gorgonopsia and Therocephalia. The condition in the other early therapsid families is unknown.

In the early therapsids the upper teeth in occlusion lie lateral of the lower teeth, but in the Anteosauridae, Titanosuchidae, Tapinocephalidae and Styracocephalidae the incisors intermesh, so do the canines in the Titanosuchidae and the whole battery in the Tapinocephalidae.

In the sphenacodonts the teeth are simple and pointed.
In the early therapsids considerable variations have arisen.

In the Eotitanosuchidae the primitive condition is retained.
In the Brithopidae and Anteosauridae the incisors are progressively lengthened and in the latter the postcanines become bulbously spatulate.

In the carnivorous gorgonopsian and therocephalian families the distal edge of the incisors, canines and post-canines becomes serrated.

In the herbivorous Titanosuchidae, Tapinocephalidae and Styracocephalidae the incisors develop a talon and heel; in the Tapinocephalidae the canine and the postcanines develop a similar talon and heel, but in the Styracocephalidae only the postcanines. The canine remains fairly normal in the Titanosuchidae and Styracocephalidae. In the Titanosuchidae the long row of postcanines are spatulate with serrated edges.

In Otsheria and Venyukovia the teeth become bluntly conical.
In the Dicynodontia the incisors disappear, the upper canines present or absent and the lower canine always absent. There are no postcanines in the Dicynodontidae and in the Endothiodontidae they are reduced, and displaced medially from the jaw margin.

## PALATE

In the sphenacodonts the pterygoids do not meet in the median line posterior to the interpterygoid vacuity. The quadrate ramus is deep and strong. The transverse ramus is strong, prominent and dentigerous. The choana is long and situated anteriorly. There is no suborbital foramen or fenestra. The posterior end of the vomers is spatulate and the vomerine bar lies low down in the skull.

The early therapsids manifest considerable variations from the primitive pelycosaur palatal structure. They all have one advance in common, viz. that the basipterygoid joint is no longer freely movable. The quadrate ramus becomes weaker in the Gorgonopsia, Therocephalia and Dicynodontia. The anterior ramus is (generally) reduced; greatly so in the Dicynodontia. The transverse ramus is progressively weakened in the series Otsheriidae-Venyu-koviidae-Dicynodontia and becomes edentulous in practically all the therapsids.

The choana is somewhat shortened in the Brithopidae and Anteosauridae but in the dicynodontian families it is both shortened and pushed backwards by the enlarged palatal process of the premaxilla. Only in the therocephalian families is a well-developed suborbital fenestra present.

Only in the Eotitanosuchidae, Gorgonopsia and Dicynodontia is the vomer well raised above the general palatal level.

## THE LOCOMOTOR APPARATUS

In the ig known families of early therapsids the structure of the girdles and limbs is not adequately known in II of these families. Any comparative consideration must thus be tentative.

An overall picture of the locomotor apparatus in the other 8 families discloses considerable adaptive radiations, but in all there is an advance
beyond the crawling habit of the pelycosaurs to a slinking habit in the brithopids, anteosaurids and hipposaurids and a more upright walking gait in the endothiodontids, dicynodontids, the 3 therocephalian families and the titanosuchids and tapinocephalids.

We may commence by attempting to give a picture of the diversity exhibited in the structure of the girdles and limbs in these early therapsids.

In the hipposaurids the procoracoid has not been ousted from the glenoid; in all the others it has been ousted.

The procorocoid is enlarged in the hipposaurids and the Dinocephalia, small in the Dicynodontia and moderate in the other families.

Only in the Dicynodontia is an acromion process developed on the scapula. This feature, typical of the mammals, is however no evidence of affinity of the Dicynodontia to the mammals, but rather a case of parallelism as it is also found in the contemporary Pareiasauridae-a cotylosaur family with no affinity to the mammals. An ossified sternum is developed in the Dicynodontia and Gorgonopsia but in none of the other groups. In all the early therapsids the axial muscles have been forced off the outer face of the ilium and the iliac height is increased.

The anterior iliac process is incipient in the hipposaurids, moderate in the pristerognathids and brithopids and anteosaurids, well developed in titanosuchids and tapinocephalids and great in the endothiodontids and dicynodontids but undeveloped in the dromasaurids.

Only in the endothiodonts and dicynodontids has the acetabulum moved to the anterior pelvic border.

The pubo-ischiatic plate retains its great primitive length in hipposaurids, pristerognathids and anteosaurids. The pubic part is shortened in titanosuchids and tapinocephalids and greatly so in endothiodontids and dicynodontids, where a pubo-ischiatic fenestra replaces the pubic foramen present in all the other families.

The pelvic symphysis is strongly ossified in the hipposaurids, pristerognathids and anteosaurids but weak in all the other families.

## Humerus

All the early therapsids have lost the primitive strap-like caput of the humerus and there has been an untwisting of the proximal and distal ends relative to each other. These ends remain expanded to various degrees, but are greatly reduced in hipposaurids.

In hipposaurids no epicondylar foramina are present. In endothiodontids, dicynodontids, Tapinocephalidae and Anteosauridae there is no ectepicondylar foramen but it is present in brithopids, titanosuchids and Therocephalia.

## Femur

In all the early therapsids there has been a preaxial shift and a shortening of the caput femoris but to varying degrees in the various families. The distal condyles have shifted to lie in the same plane and this distally so that the knee
joint becomes a simple hinge well adapted to a more upright disposition of the limb.

Only in the Titanosuchidae and Tapinocephalidae has the femur become greatly broadened.

## Forefoot

The primitive phalangeal formula of $2,3,4,5,3$ has been reduced to $2,3,4,4,3$ in the hipposaurids and to $2,3,4,3,3$ in the anteosaurids and to $2,3,3,3,3$, in all the other families of the early therapsids.

## Hindfoot

In the tarsus the primitive medial central has been lost in all the early therapsids, where this structure is known, and the phalangeal formula reduced from the primitive $2,3,4,5,4$ to $2,3,4,4,3$ in the hipposaurids, $2,3,4,3,3$, in brithopids and anteosaurids and 2, 3, 3, 3, 3 in all the other early therapsids.

In the pristerognathids the astragalus tends to overlie the calcaneum and in the hipposaurids a sustentaculum tali is developed as well as a tuber calcis.

Now, what does this rather great diversity in the structure of the locomotor apparatus signify?

The main variations are apparently towards the acquisition of a greater degree of active movement than that of crawling - on the one hand that possible in a slinking habit and on the other in a more upright walking habit.

Is the improved locomotor ability correlated in any way with an improved masticatory ability?

In the herbivorous families the achievement of a walking gait would increase the area that can be grazed and the ability to reach higher ground would bring these reptiles into contact with hardier and more fibrous plants than those flourishing in more marshy terrain.

In the Otsheriidae and Venyukoviidae the bluntly conical teeth appear to be adapted to a coarser fare.

The horny jaw sheaths and plates of the Endothiodontidae and Dicynodontidae together with the fore-and-aft sliding of the lower jaw would greatly help in the shearing and milling of fibrous vegetable matter.

In the Titanosuchidae and Tapinocephalidae and Styracocephalidae the intermeshing talon-and-heel teeth showing considerable abrasion are obviously well adapted for piercing and crushing tough fibrous plants.

In the Eotitanosuchus-Anteosaurus series of carnivores the progressive development of a formidable battery of long pointed intermeshing incisors together with the strong canines would enable these reptiles to execute a strong piercing and jerking bite into the flesh of the larger herbivores. The progressive decrease in the role of the postcanines would accompany this method of biting.

A slinking habit of locomotion indicates that these carnivores did not run after their prey but rather lay in ambush and then pounced.

The early gorgonopsians with their moderate anterior teeth and reduced postcanine series and slinking but agile locomotory ability probably pounced on the small contemporary Dicynodontia or could have at times been carrion eaters.

The early therocephalians with their limbs well adapted to a more upright walking and running gait could pursue and overcome even some of the larger herbivores. With the postcanines greatly reduced in some genera, the front part of the jaws was mostly in action pulling and tearing out lumps of flesh.

The scaloposaurids with a long tooth row and small canines and with some cuspidate postcanines were better adapted as insectivores.

The variations in the adductor muscles in the above groups of early therapsids appear to be well correlated to both the varied dentitions and modes of locomotion.

## DIVERSE STRUCTURES

The quadratojugal has variable relations in the early therapsids. In all it is, however, much reduced in size and never enters the lower temporal arch as it does in some members of all three of the pelycosaurian suborders.

Primitively a surface bone of the postero-lateral corner of the skull, flanking the quadrate, it first tends to move medially in some of the higher sphenacodonts to rest on the quadrate above the lateral condyle as a bone of reduced size.

This process is seen continued in the Gorgonopsia, Therocephalia, Brithopidae and Anteosauridae. Whereas in the Dicynodontia the quadratojugal becomes a plate applied to the antero-lateral face of the everted squamosal, in the Titanosuchidae and Tapinocephalidae the quadratojugal, variable in size and shape, still forms part of the lateral skull surface. Does this indicate an origin from different pelycosaurian ancestors?

The lacrimal, primitively a long bone stretching from orbit to nostril, is reduced to an anterior circumorbital bone in all the therapsids. It is reduced in some sphenacodonts but also in the edaphosaurian, Mycterosaurus.

The supratemporal is absent in all therapsids and in all one coronoid is lost, but both coronoids are absent in the Dicynodontia.

In the early therapsids the preparietal is a new acquisition in only the Endothiodontidae and Dicynodontidae as well as the Hipposauridae and Galesuchidae. In the primitive dicynodontian family, Otsheriidae, there is however no preparietal. It is also absent in the possible gorgonopsian forerunner, Eotitanosuchus and Phthinosuchus.

In the early Gorgonopsia the preparietal lies anterior to the pineal foramen, but forms its anterior border in the early Dicynodontia which implies a different raison d'être.

The dorsal process of the premaxillaries varies in length in the pelycosaurs, being long in ophiacodonts, short to medium in sphenacodonts and edaphosaurs.

It is also variable in the early therapsids, being long in eotitanosuchids, brithopids and anteosaurids; very long in the Titansuchidae and Tapinocephalidae. In the dicynodontian families it is long in the primitive Otsheriidae and Venyukoviidae, but short in the more advanced early Endothiodontidae and Dicynodontidae.

Arranged into series as to length of premaxillary process we have:


Together with the development of a reflected lamina of the angular we see a reduction in the role of the posterior mandibular bones in the higher sphenacodonts and in all the early therapsids and this reduction is more pronounced in those therapsids where the dentary develops a coronoid process.

The braincase has its sidewall largely open in the Therocephalia and Dicynodontia, but much less so in the Anteosauridae, Titanosuchidae and Tapinocephalidae. In the Therocephalia there is no downward directed flange of the parietal whereas in all the other early therapsids, where known, it is present.

The sphenethmoidal complex is weakly ossified in the early Therocephalia but well developed in the Dinocephalia and in those early Dicynodontia where it has been studied. In the Dicynodontia it lies far anteriorly and has no contact with the prootic, whereas in the Dinocephalia contact is made above the lateral fenestra.

The fenestra ovalis lies low down in the skull in all the early therapsids, but there are considerable variations in the structure of the stapes. The dorsal process of the stapes is reduced in the Tapinocephalidae and absent in all the other early therapsids. A stapedial foramen usually present is absent in the hipposaurids, brithopids, anteosaurids and in all the early Dicynodontia where the stapes has been described.

The exoccipital apparently does not enter the floor of the braincase in Captorhinus. This is definitely the case in the early Endothiodontidae and Dicynodontidae and the early Gorgonopsia, whereas in Dimetrodon and all the Dinocephalia it forms the whole posterior part of the brain floor.

The prootics do not meet in the middle line in Captorhinus, but do meet in Dimetrodon. They meet in all the Dinocephalia, where known, but not in the Dicynodontia and just meet in the Therocephalia.

The dorsum sellae is very high in Captorhinus and high in Dimetrodon. In the former it is formed by the basisphenoid, whereas in the latter by the prootic.

In the Dinocephalia the upper part of the dorsum sellae is formed by the prootic, and in the Therocephalia the prootic just enters, whereas it is excluded in the early Dicynodontia. The sella turcica is deep in Captorhinus and Dimetrodon. This is also the case in all the Dinocephalia but is shallow in the Therocephalia and Dicynodontia.

The quadrate ramus of the pterygoid is strong in the sphenacodonts and is greatly strengthened in the Dinocephalia but weakened in Therocephalia, Gorgonopsia and Dicynodontia.

## Morphological series

In the foregoing the extent of the variations observed in the assemblage of early therapsids has been given in some detail. The result being that one cannot see the wood for the trees.

We must now consider whether these divergencies can be arranged in some orderly manner on a basis of possible consecutive ascending morphological stages.

The early therapsids form a fauna of discrete types of animals living together during a definite interval of time. They can thus, broadly speaking, be considered as contemporaries and thus some cannot be conceived as being ancestral to others.

What we can, however, attempt to do is to determine the possibility of arranging the animals exhibiting these various structural features in series, one derivable from the others in a morphological sense.

SERIES I
Captorhinidae $\rightarrow$ Pelycosauria $\rightarrow$ Eotitanosuchidae $\rightarrow$ Brithopidae $\rightarrow$ Anteosauridae.
Consecutive steps in the following features:

## (a) Temporal fenestra

Absent in captorhinids $\rightarrow$ small or double in pelycosaurs $\rightarrow$ large in eotitanosuchids $\rightarrow$ larger in brithopids $\rightarrow$ very large in anteosaurids. This progressive increase is mainly due to lateral and posterior outflaring of the squamosal.
(b) Intertemporal skull table

Wide in pelycosaurs $\rightarrow$ still wide in Eotitanosuchus $\rightarrow$ greatly reduced in brithopids $\rightarrow$ but less reduced in anteosaurids.

## (c) Posterior process of postorbital

Horizontal surface bone in pelycosaurs $\rightarrow$ just starting to tilt down in Eotitanosuchus $\rightarrow$ tilting progressively increased in brithopids and anteosaurids to culminate as a bone lying nearly vertically flanking the parietal inside the temporal fossa.

## (d) Area of origin of the temporal muscle on the postorbital

Under surface of postorbital in pelycosaurs $\rightarrow$ moving to lateral edge and dorsal face in Eotitanosuchus $\rightarrow$ on the morphological dorsal face in brithopids and anteosaurids which progressively becomes a functionally lateral face.

## (e) Insertion of temporal muscle

Notwithstanding the changes in the origin of the muscle in this series, the insertion on the mandible remains constant from sphenacodontid to anteosaurid.

## (f) Faw articulation

In captorhinids this lies posteriorly in a plane with the occipital condyle and level with the alveolar border $\rightarrow$ this is still the position in most pelycosaurs, but in the higher sphenacodonts it has shifted downwards $\rightarrow$ posterior and low in eotitanosuchids $\rightarrow$ shifted both anteriorly and ventrally in brithopids and anteosaurids.

## (g) Reflected lamina of the angular

Absent in captorhinids $\rightarrow$ still absent in most pelycosaurs, but developed in the higher sphenacodonts $\rightarrow$ progressively better developed in eotitanosuchids, brithopids and anteosaurids.

## (h) Marginal tooth row and 'canines'

In captorhinids the tooth row is long, without 'canines' $\rightarrow$ in pelycosaurs long to very long, canines absent or variously present in the three pelycosaur groups, but strong in Eothyris and most sphenacodontids $\rightarrow$ row reduced in eotitanosuchids, but strong definite canine present $\rightarrow$ progressive reduction of number of post-canines in brithopids and anteosaurids, canines very strong and incisors progressively lengthened to culminate in the very long intermeshing set of Anteosaurus.
(i) Septomaxilla, maxilla and lacrimal

In captorhinids the septomaxilla is intranarial, the lacrimal enters the narial border and the maxilla is low $\rightarrow$ these relations are retained in nearly all the pelycosaurs but in Mycterosaurus, Sphenacodon and Dimetrodon the lacrimal fails to reach the naris and the maxilla becomes high. This may be related to the greater development of 'canines' in these three genera, but other pelycosaurs have enlarged 'canines' without affecting the primitive relations of the maxilla $\rightarrow$ in eotitanosuchids, brithopids and anteosaurids the septomaxilla extending backwards becomes a bone of the lateral surface, the lacrimal fails to reach the nostril and the maxilla is high.

## (j) Dorsal process of the premaxilla

Short in captorhinids $\rightarrow$ moderately lengthened in some pelycosaurs but still fairly short in sphenacodonts $\rightarrow$ greatly lengthened in eotitanosuchids, brithopids and anteosaurids.
(k) Braincase

Insufficiently known in this series, but in both Dimetrodon and Anteosaurus the sphenoidal complex is well ossified and the exoccipital and the prootic enter the floor of the braincase and the prootics meet in the middle line in the dorsum sellae.

## (l) Locomotor apparatus

Insufficiently known, but in all brithopids and anteosaurids the femur has become a long, slender curved bone.

SERIES II

(a) Temporal fenestra

In brithopids large $\rightarrow$ slightly reduced in titanosuchids $\rightarrow$ moderately to very greatly reduced in tapinocephalids (except in Avenantia and Riebeeckosaurus).
(b) Intertemporal skull table

Moderately wide in brithopids $\rightarrow$ so also in titanosuchids $\rightarrow$ moderately to enormously widened in tapinocephalids (except in Avenantia and Riebeeckosaurus).
(c) Posterior process of the postorbital

In brithopids long and high with good contact with the squamosal $\rightarrow$ in titanosuchids reduced to a splint and just meeting the squamosal $\rightarrow$ in tapinocephalids shortened still further so that in some forms it fails to reach the squamosal.
(d) Area of origin of the temporal muscle on the postorbital

In brithopids from the fairly large tilted (dorsal) face $\rightarrow$ in titanosuchids this area is reduced and the origin transferred more on to the parietal $\rightarrow$ this is carried further in the tapinocephalids where the total area is small (except in Avenantia and Riebeeckosaurus).
(e) Insertion of temporal muscle

The primitive pelycosaurian position is retained throughout the series.
(f) Jaw articulation

In brithopids anterior to the plane of the occipital condyle $\rightarrow$ in titanosuchids still further anteriorly $\rightarrow$ in tapinocephalids very far anteriorly.

## (g) Marginal teeth

In brithopids moderately long pointed incisors, well-developed canines, fairly long postcanines row of bluntly conical teeth.

In titanosuchids we find a radical change to a herbivorous dentition; strong pointed canine is retained, the strong intermeshing incisors have developed a piercing talon and crushing heel and the very long postcanine row has cuspidate spatulate crowns.

In tapinocephalids this process is carried further in that the canine has disappeared as such and the very long series consists of isodont talon-and-heel teeth, all intermeshing but the anterior teeth are weaker than the incisors of the titanosuchids.

## (g) Dorsal process of the premaxilla

In brithopids this is of moderate length intercalated between the nasals, greatly lengthened in both titanosuchids and tapinocephalids and nearly reaching the frontal.

## (h) Braincase

Little known in the brithopids.
In both titanosuchids and tapinocephalids the lateral wall is well ossified and the sphenoidal complex strongly ossified; the exoccipital and prootic enter the floor of the braincase; the prootics meeting in the middle line form part of the dorsum sellae.

## (i) Locomotor apparatus

In brithopids the girdles and limbs are fairly lightly built whereas in both titanosuchids and tapinocephalids they are massive to very massive.

Styracocephalidae
With a persistent canine and the development of weak talon-and-heel incisors and postcanines and a secondary broadened intertemporal skull table and reduced temporal fossa. Styracocephalus can be derived from the titanosuchids as a branch somewhat divergent from the tapinocephalid branch.

Estemmenosuchidae
It is difficult to place Estemmenosuchus. The shagreen of palatal teeth is reminiscent of early pelycosaurs.

The broad intertemporal region with the upper part of the postorbital lying on the dorsal surface overhanging the temporal fenestra are eotitanosuchid features as is the large temporal fenestra. The incisors and canines are like those of the brithopids.

The great downward and forward shift of the quadrate and the long series of postcanines parallel features of both the titanosuchids and the tapinocephalids.

SERIES III
Pelycosauria $\rightarrow$ Phthinosuchidae $\rightarrow$ Hipposauridae $\rightarrow$ Galesuchidae.
(a) Temporal fenestra

This is small or double in the pelycosaurs $\rightarrow$ suddenly very large in Phthinosuchus $\rightarrow$ but only moderately enlarged in `Hipposaurus $\rightarrow$ then again large in the galesuchids. Clearly not a consecutive series.
(b) Intertemporal skull table

Wide in pelycosaurs $\rightarrow$ remains wide in Phthinosuchus $\rightarrow$ becomes very wide in Hipposaurus $\rightarrow$ but somewhat reduced in the galesuchids. Again not a consecutive series.
(c) Posterior process of the postorbital

In the whole series it persists as a horizontal surface bone overhanging the temporal fenestra. Long in pelycosaurs $\rightarrow$ short in Phthinosuchus $\rightarrow$ very long in Hipposaurus $\rightarrow$ long in galesuchids. In Phthinosuchus it is almost entirely excluded from the edge of the skull table due to the development of a lappet of the squamosal extending far anteriorly and lying laterally of the postorbital.
(d) Area of origin of the temporal muscle on the postorbital

In the whole series the origin remains on its under surface, but in Phthinosuchus also from the under surface of the squamosal lappet.
(e) Insertion of the temporal muscle

Partially inserted on the upper and outer face of the dentary in pelycosaurs $\rightarrow$ this primitive insertion retained in Phthinosuchus $\rightarrow$ but in hipposaurids and galesuchids mainly on the strongly developed coronoid process.

## (f) Jaw articulation

Posterior position in pelycosaurs $\rightarrow$ shifted anteriorly in Phthinosuchus, Hipposaurus and galesuchids. But situated far ventrally in Hipposaurus.
(g) Dorsal process of the premaxilla

Moderately long in pelycosaurs $\rightarrow$ unknown in Phthinosuchus $\rightarrow$ very short in Hipposaurus and the galesuchids.

## (h) Marginal teeth

Postcanines progressively reduced in the series. A single well developed canine present in Phthinosuchus, Hipposaurus and the galesuchids.
(i) Locomotor apparatus

Unknown in Phthinosuchus and the early galesuchids. In Hipposaurus the anterior iliac process remains weak as in pelycosaurs, an ossified sternum is developed;
the limbs have become long and slender as in anteosaurids. The tarsus is greatly specialized with the development of a tuber calcis and a sustentaculum tali; the phalangeal formula only slightly reduced, probably $2,3,4,4,3$.

## (j) Preparietal

Absent in pelycosaurs and in Phthinosuchus, suddenly present in Hipposaurus and all other gorgonopsians.

## (k) Vomer

Lying in general plane of palate with broad posterior end in pelycosaurs $\rightarrow$ raised or vaulted, with broad posterior end in Phthinosuchus $\rightarrow$ raised or vaulted in Hipposaurus and galesuchids, posterior end tapering and intercalated between palatines.
series iv
Pelycosauria $\rightarrow$ Otsheriidae $\rightarrow$ Endothiodontidae $\rightarrow$ Dicynodontidae.

## (a) Temporal fenestra

In pelycosaurs small or double, bounded by postorbital, squamosal and jugal in the higher sphenacodonts $\rightarrow$ in Otsheria large, bounded by postorbital, squamosal and jugal $\rightarrow$ in endothiodonts and dicynodonts very large mainly due to posterior outflaring of the squamosal, with jugal participation in its border reduced.

## (b) Intertemporal skull table

Wide in pelycosaurs $\rightarrow$ moderate in Otsheria and in endothiodonts and dicynodonts.

## (c) Posterior process of the postorbital

A horizontal surface bone in pelycosaurs and fairly long $\rightarrow$ in Otsheria showing only an edge as a surface bone and fairly long $\rightarrow$ in endothiodonts and dicynodonts slanting downwards laterally and very long.

## (d) Area of origin of the temporal muscle on the postorbital

In pelycosaurs from its undersurface $\rightarrow$ in Otsheria from its latero-ventral edge $\rightarrow$ in endothiodonts and dicynodonts from the dorsal surface now lying at a slant.

## (e) Squamosal

In most pelycosaurs the postero-lateral corner of the squamosal lies in the plane of the alveolar border, but in the higher sphenacodonts (and Edaphosaurus) it lies far ventrally and the lower temporal arch, in which the jugal plays a large part, lies in a vertical plane.

In Otsheria the process of the squamosal lies far ventrally and the lower arch, in which the jugal plays a large part, still lies in a vertical plane.

In endothiodonts and dicynodonts both these features are suddenly radically changed. The ventral process, still extending far ventrally, is uniquely everted to present a sharp lateral edge and an anterior face to which the quadratojugal is applied as a flat sheet of bone. The anterior process of the squamosal, now forming most of the lower temporal arch, is also everted and now lies in a nearly horizontal plane with its morphological dorsal edge facing laterally.

## (f) Temporal muscles

In sphenacodonts the origin of the temporal muscles is mainly from the under surface of the bones of the skull roof $\rightarrow$ in Otsheria it is partly shifted to the edge of the postorbital $\rightarrow$ in the early endothiodonts and dicynodonts the unique and radical changes in the nature of the squamosal is due to the radical changes in the areas of origin of both the temporal and masseter. Noteworthy is that the masseter lying medially of the zygomatic arch in the pelycosaurs now has its origin from the antero-lateral face of the squamosal below the zygomatic arch.

The insertion of these muscles has not changed much from the pelycosaur condition and no coronoid process is developed, but there is already an indication of a lateral flange on the dentary well developed in some later Dicynodontia.

## (g) Reflected lamina of the angular

In the higher sphenacodonts we have the first development of this structure $\rightarrow$ it is unknown in Otsheria, but in Venyukovia it is well developed $\rightarrow$ it is present in the early endothiodonts and dicynodonts but in nature differs considerably from that in the other contemporary therapsids indicating a difference in the insertion of the anterior pterygoid muscle. A fenestra in the lower jaw is known in Ophiacodon but in no other pelycosaur; it is also present in Venyukovia and the endothiodonts and dicynodonts.

In some pelycosaurs with 'canines' the maxilla is high $\rightarrow$ but in Otsheria without canines it is suddenly very high as it is in endothiodonts with or without canines. If the increase in maxillary height is due to the presence of canines, as has been maintained, then Otsheria must have inherited this feature from an ancester with canines.

## (h) Jaw articulation

This lies posteriorly in the series pelycosaur $\rightarrow$ Otsheria $\rightarrow$ endothiodonts and dicynodonts.

In pelycosaurs it lies high up (except in the higher sphenacodonts and Edaphosaurus) $\rightarrow$ very far ventrally in Otsheria and endothiodonts and dicynodonts.

## (i) Septomaxilla, lacrimal and maxilla

In pelycosaurs the septomaxilla lies internarially $\rightarrow$ in Otsheria there is a small lateral face $\rightarrow$ but in the endothiodonts and dicynodonts it is again internarial.

The lacrimal fails to reach the naris in some pelycosaurs (e.g. Dimetrodon
and Mycterosaurus) $\rightarrow$ in Otsheria it is greatly shortened (but still long in Venyukovia) $\rightarrow$ in endothiodonts and dicynodonts it has become a short bone of the anterior orbital border.

## (j) Snout

In nearly all pelycosaurs the snout is long with the orbit and nostril far apart $\rightarrow$ in Otsheria it is greatly shortened (but still fairly long in Venyukovia) $\rightarrow$ in the early endothiodonts and dicynodonts the snout is very greatly shortened with the naris very near the orbit.

## (k) Dorsal process of the premaxilla

Moderately long in pelycosaurs $\rightarrow$ long tapering intercalation between nasals in Otsheria (and Venyukovia) $\rightarrow$ but very short in endothiodonts and dicynodonts.

## (l) Palatal face of the premaxilla

Absent in pelycosaurs and choana extending far anteriorly $\rightarrow$ well developed in Otsheria and choana pushed posteriorly but not reaching the palatine (very well developed in Venyukovia, choana pushed back but still long, makes contact with the palatine) $\rightarrow$ well developed in endothiodonts and dicynodonts, choana pushed backwards and greatly shortened, it sometimes makes contact with the palatine.

## (m) Vomer

In pelycosaurs paired and lying in general plane of palate broadened posteriorly $\rightarrow$ unpaired in Otsheria lying low down broad posteriorly (paired in Venyukovia) $\rightarrow$ raised above (vaulted) general plane of palate, reaching interpterygoid vacuity in Dicynodontia.
(n) Lateral ramus of pterygoid

Strongly developed in pelycosaurs $\rightarrow$ quite strong in Otsheria (but weak in Venyukovia) $\rightarrow$ absent in endothiodonts and dicynodonts.
(o) Marginal teeth

In pelycosaurs the tooth row is long, with 'canines' in some forms, pointed in most $\rightarrow$ moderately long row in Otsheria, with incisors enlarged, no 'canine', postcanines spatulate (in Venyukovia 'canine' present, teeth bluntly conical, some with crushing face) $\rightarrow$ in early endothiodonts and dicynodonts the anterior part of the jaws is edentulous with development of horny sheaths, strong upper canines present or absent, reduced postcanines shifted away from jaw margin in endothiodonts but absent in dicynodonts.

## (p) Braincase

The exoccipital and prootic form floor of braincase in Dimetrodon and prootics meeting in middle line in the dorsum sellae $\rightarrow$ unknown in Otsheria $\rightarrow$ in early endothiodonts and dicynodonts the exoccipital and prootic do not enter the floor and the prootic does not enter the dorsum sellae.

The sphenoidal complex is well developed in Dimetrodon, situated far posteriorly but the lateral wall is widely open $\rightarrow$ unknown in Otsheria $\rightarrow$ very well developed in endothiodonts and dicynodonts but situated very far anteriorly with the result that the lateral wall is widely open because in addition the prootic has little anterior development.

## (q) Locomotor apparatus

Primitive in pelycosaurs $\rightarrow$ unknown in Otsheria $\rightarrow$ in the earliest endothiodonts and dicynodonts it is already highly specialized. The scapula has a welldeveloped acromion process; there is a strongly developed ossified sternum, the ilium has an enormous anterior iliac process; the pubis is greatly reduced; there is a large pubo-ischiatic fenestra; the phalangeal formula is reduced to 2, 3, 3, 3, 3 .

## Galeops

This very imperfectly known form from the Tapinocephalus zone shows a few features similar to those of the early Dicynodontia.

The temporal fenestra is short but high; the squamosal has a long downwardly directed process; in the lower jaw there is no coronoid process but a reflected lamina of primitive form is developed and a fenestra pierces the jaw; the jaws are edentulous.

With its large procoracoid and the absence of an acromion process Galeops is more primitive than the other early dicynodonts.

## Therocephalia

We, as yet, know no forms that could provide a morphological step intermediate between the pelycosaurs and the four earliest therocephalian families (Alopecodontidae, Lycosuchidae, Pristerognathidae and Scaloposauridae).

The big morphological gap will be evident if we, in summary, list the advances shown in these early therocephalians.

The temporal fenestra is immediately very large with outflaring squamosals, and the parietal is always intercalated between the postorbital and squamosal; the intertemporal skull table is narrow and developing a sagittal crista; the posterior process of the postorbital is reduced to a small splint applied to the lateral parietal face; the temporal muscle no longer arising from the under surface of the roof bones and is inserted on a strong coronoid process; one or two strong canines developed (except in the scaloposaurids); small teeth anterior to the large canine in alopecodonts but absent in the other families and the tooth row generally shortened sometimes radically; dorsal process of premaxilla always very short; jaw articulation shifted slightly forwards; but the exoccipital and prootic less prominent in the brain floor than in Dimetrodon and the sphenoidal complex less developed; a large suborbital fenestra present; the whole locomotor apparatus well developed in adaptation to a more upright walking gait and the phalangeal formula reduced to $2,3,3,3,3$.

## SUMMARY OF MORPHOLOGICAL SERIES

The series primitive sphenacodont (Haptodus?), eotitanosuchids, brithopids to anteosaurids, undoubtedly forms a morphological ladder with its bottom end resting further down among the captorhinomorphs with Anteosaurus on the highest rung.

The series primitive brithopid (Sydon?), titanosuchids to tapinocephalids, styracocephalids and estemmonosuchids, is clearly a line closely related to but diverging from the first series. That this series started from a primitive brithopid appears very probable but can be queried. The tapinocephalids are a very mixed lot but undoubtedly closely related and in various ways a rung up the ladder above the titanosuchids, but the picture is complicated by the developments seen in the other two related forms-Styracocephalus and Estemmenosuchus. This series also terminates at the top of the Tapinocephalus zone.

In the series primitive sphenacodont, Phthinosuchus, Hipposaurus to galesuchid gorgonopsians, the position of Phthinosuchus as intermediate between sphenacodonts and hipposaurids is very uncertain but a fairly close but less specialized form than Phthinosuchus would fit the bill. Moreover, Hipposaurus does not quite fit in as an antecedent stage to the galesuchids.

If those objections are valid then the phthinosuchids, hipposaurids and galesuchids form a triradiate branch arising from a sphenacodont group close to that from which the Dinocephalia is derived.

In the dicynodont series the morphological step from any known pelycosaur to Otsheria is very great and I find it difficult to visualise how such a step could have taken place; but the transition from Otsheria to endothiodonts and dicynodonts is small and obvious.

In the pelycosaur-therocephalian series no intermediate stages are known and the gap is very wide, but nothing that the discovery of some more primitive forms would not bridge.

In short, the Dinocephalia and Gorgonopsia can be derived from a sphenacodont near to Haptodus and the Dicynodontia and Therocephalia from two other as yet unknown primitive pelycosaurs.

The foregoing morphological analysis can also be presented in numerical form. For the various groups under consideration here I have tabulated the primitive reptilian characters determinable in each.

Arranging these in numerical order we get the following percentages:
Captorhinomorpha 100
Sphenacodontia 88
Eotitanosuchia 68
Dinocephalia 6o
Gorgonopsia $\quad 56$
Therocephalia 52
Otsheriidae 40
Dicynodontia 32

## Early therapsid history

From the foregoing there is no doubt that on purely morphological grounds the therapsids must be derived from the captorhinomorphs by way of the sphenacodont pelycosaurs.

Now, does this fit in with the known history of the early tetrapods?
We can begin the story with the primitive anthracosaurs which were the first tetrapods to successfully achieve an amphibious life. These are best known from the Lower Carboniferous of Scotland where the prevailing climate was warm and moist and eminently suitable for an existence partly in water and partly on land.

At the close of the Lower Carboniferous times the Scottish climate changed radically. The elevation caused by the Hercynian Foldings made the climate arid and thus unsuitable for these amphibians.

We now find the amphibian history continuing in central Europe and North America where during Upper Carboniferous times swampy conditions in a warmer climate prevailed.

Swampy conditions continued into Lower Permian times in central Europe and North America, but slowly changed to drier conditions and this sparked off the explosive development of the earliest cotylosaurs especially in North America, closely followed by the rise of the pelycoasurs also mainly in North America but with representatives in central Europe.

At the end of the Lower Permian the climate over North America became more and more arid and the cotylosaur-pelycosaur explosion came to an abrupt end.

In parts of Europe, however, the Lower Permian climate remained cool and favourable for the continued existence of the sphenacodonts and during the Middle Permian this cool climate continued in Cisuralian Russia where the first therapsids made their appearance in deltaic conditions and from there spread to southern Africa where the favourable flood plain conditions existed in a fairly cool to warm climate.

It would thus appear that for every major step in the phylogeny a change of scene was necessary.

This seeming capriciousness can, however, be reasonably accounted for.
During Carboniferous-Permian times the western part of the northern hemisphere formed a single continent-Laurentia-and, notwithstanding the upheavals caused by the Hercynian Foldings and the presence of Tethys, there would at this time have been fewer barriers for the transmigration of tetrapods than at the present time.

## Subsequent history

Arising in late Ecca (Lower Permian) times the therapsids formed a firmly established order of reptiles with four distinct suborders at the beginning of early Beaufort (Tapinocephalus zone-Middle Permian) times.

During the whole of the Tapinocephalus zone ( 2200 m of sediments) little further development took place.

The Dinocephalia were fully developed at the base of the zone-only Styracocephalus is first encountered above the lowest of the tripartite subdivisions of the zone. This greatly diversified suborder dominated the vertebrate life of the Middle Permian, consisting as it did of a family of large carnivores (Anteosauridae) and three families of large herbivores (Titanosuchidae, Tapinocephalidae and Styracocephalidae). Life during this time must have been easy with a cool moist climate in an area of low relief consisting of large expanses of fresh-water pools separated by low uplands with periodic floodings. But abruptly at the end of Tapinocephalus zone times the life span of the Dinocephalia was cut short, thus ending one of the first four developmental trends of the early therapsids.

This sudden extinction was apparently not caused by any radical change in the environmental conditions. The succeeding Endothiodon zone lies conformably on the Tapinocephalus zone without any radical change in lithological character - the only noteworthy feature being the increase of the number of purplish bands indicating more periods of somewhat drier conditions. There was also no sudden development of competitors or antagonists.

The only reason for the sudden extinction of the Dinocephalia I can advance is that they went to seed in too favourable living conditions, aggravated by the pathological pachyostosis induced by a pituitary hypertrophy.

The Gorgonopsia, represented by two families from low down in the Tapinocephalus zone, had by then already developed all the characters typical of this suborder and during the Middle Permian show no further development. They constituted a very minor element in the fauna of these times. This suborder of rather primitive therapsids is represented in the higher zones of the Beaufort beds to form a much more important element in the fauna.

The Hipposauridae, forming a very distinctive family, is not represented in the Endothiodon zone, but in the Cistecephalus zone we know three further genera. Thus they become extinct at the top of the Upper Permian.

The suborder is further represented in the Endothiodon zone by 13 genera and in the Cistephalus zone, by $4^{8}$ genera and these have been subdivided into as much as 17 discrete families, indicating that during the Upper Permian this suborder really went to town and during this period constituted an important element of carnivorous forms in the fauna.

Their span of life came to an abrupt end at the close of the Permian.
This second developmental trend of the early therapsids thus had a lifespan extending through the whole of the Middle and Upper Permian. During this period the gorgonopsians manifest but minor variations and retain such a uniform morphological pattern that the subdivision into separate families can at most be considered as of taxonomic convenience.

They form an interesting group of fairly long-lived primitive therapsids suddenly present at the beginning of the Middle Permian with their distinctive
cachet fully developed in one bound and wholly sterile.
It is of interest to note that the gorgonopsians, apparently of Cisuralian origin, have only three genera in the Russian Upper Permian. Inostrancevia is a giant gorgonopsian, whereas the aberrant Proburnetia is very similar to Burnetia of the Karoo.

The Dicynodontia, with two families in the Middle Permian, numerically rich in individuals, formed a significant element in the fauna as the sole assemblage of small herbivores preyed on by the smaller to medium sized carnivores of those times. With a fully developed morphological pattern from the base of the Middle Permian they waxed exceedingly until becoming extinct in the Middle Trias.

The basic pattern, fully developed at the beginning of the Middle Permian, remains unchanged during their long span of life. But in the Upper Permian a mass of small variations occur as witnessed by the fact that over 200 species have been named. During these times the dicynodonts were extremely abundant, by far outnumbering all their contemporary therapsids and constitute the bulk of the herbivores on which the gorgonopsians preyed.

In the Lower Trias the lystrosaurs were very nearly the only herbivores preyed on by some small cynodonts and the predaceous Chasmatosaurus.

In Cynognathus zone times we get the kannemeyerids forming the end of this line of development, which during its long span of life continued basically unchanged and sterile.

The Dicynodontia, with the Cisuralian form Otsheria as starting point, had but five descendant genera in Russia, but since the Upper Permian have spread to Scotland, China, Indo-China, North and South America, India and Antarctica.

Thus, notwithstanding their innate inability to escape from their confining basic structural pattern they were very adaptable herbivorous reptiles, well able to fit into all the vicissitudes of the invironmental changes encountered from the moist Middle Permian to deep into the arid Trias in six of our present continents.

The Therocephalia with four families and 18 genera in the Tapinocephalus zone form an important element of the contemporary fauna forming the small to medium sized insectivores and carnivorous predators of their time.

The Pristerognathidae continue into the Upper Permian with three genera in the Endothiodon zone and one in the Cistecephalus zone.

The Alopecodontidae have no representatives in the Endothiodon zone but there are two genera in the Cistephalus zone.

The Scaloposauridae continue into the Trias with two genera in the Endothiodon zone, 14 in the Cistecephalus zone and one in the Lystrosaurus zone.

In the Upper Permian five new families make their appearance, indicating a continued virile variability. In Euchambersia we have the first poisonous reptile with a poison gland and an appropriate fang.

In the specialized Whaitsiidae we find a tendency towards the develop-
ment of a secondary palate and the closure of the suborbital fenestra.
Of the Akidnognathidae and Ictidosuchidae there are in the Upper Permian nine genera of small advanced therocephalians.

In the Cynognathus zone there are nine genera of small to medium-sized Bauridae more advanced than the Scaloposaurids from which they arose.

In the Red Beds there follow the Tritylodontidae and in the Cave Sandstone we have the Diarthrognathidae.

The Cynodontia, appearing for the first time in the Cistecephalus zone, if not developed independently and directly from the pelycosaurs can only be derived from the Therocephalia.

In this versatile assemblage the later branches exhibit various trends towards the mammalian condition particularly in regard to the braincase, the dentition, the secondary palate, the reduction of the posterior mandibular bones and the establishment of an articulation of the dentary directly to the squamosal.

Of the four suborders of the therapsids, developed from pelycosaur ancestors before the beginning of the Middle Permian, the Dinocephalia, Gorgonopsia and Dicynodontia have been proved phylogenetically sterile.

Only the more plastic and versatile Therocephalia developed upper branches approaching structural levels very close to that of the first mammals and it seems reasonable to assume that one or more of these trends did actually culminate in the first mammals.

This phylogenetic success of the Therocephalia can be attributed to a number of factors. In the initial stages most of the primitive conservative structural patterns were bred out and a great amount of plasticity was retained by developing at a moderate tempo without any extravagant variations. In some of the later branches, as exemplified by Euchambersia and the whaitsiids, such sterile developments were however not avoided, but there was always a plastic core retained.

From the start excessive size was avoided, except in some pristerognathids and lycosuchids which soon petered out. Together with the small size there was the acquisition of an agile locomoter potential and this ability to live an active mobile life ensured their ultimate success.

## References

A full bibliography has been given in
Boonstra, L. D. 1969. The fauna of the Tapinocephalus zone (Beaufort beds of the Karoo). Ann. S. Afr. Mus. 56: 1-73.
Fox, R. C. 1964. The adductor muscles of the jaw in some primitive reptiles Univ. Kans. Publs. Mus. nat. Hist. 12: 657-68o.


Fig. i. The therapsid explosion in the Karoo.
This diagram very effectively illustrates the main features of the faunistic history of the therapsids during Permo-Triassic times.
The therapsids, arising in Cisuralian Russia in the Lower Permian, spreading southwards entered the Karoo Basin at the beginning of the Middle Permian as a well-established and diversified order of reptiles. In the cool, moist, equitable climate then prevailing in the Karoo they quickly established themselves as the dominant land vertebrates.

This initial burst slackened off towards the end of the Middle Permian.
But in the Upper Permian, with its more varied climate, in which periods of warmer drier conditions alternated with fairly cool and moist periods, in a further and greater expansive burst the therapsids attained their maximum faunistic development.
With the drastic ecological changes in the Lystrosaurus zone (lowest Triassic) with its heavy rains and marshes in a warmer climate, the therapsids all but petered out.
Later in the Triassic, in drier and sometimes arid conditions, only those therapsids far advanced in a mammal direction extended the life span of the therapsids, but now they were faunistically overshadowed by the sauropsid explosion which then came under way.


Fig. 2. The explosion of the therapsid suborders in the Karoo.
The Dinocephalia, arising in Russia in the late Lower Permian, entered the Karoo Basin as a diversified suborder and immediately became the dominant therapsids. But they quickly shot their bolt and from the middle of the Middle Permian declined rapidly to become extinct before the Upper Permian. Thus ended the first sterile therapsid trend towards an actively mobile life on firm land of some altitude.

The Gorgonopsia and Dicynodontia, entering the Karoo Basin fully fledged, but in a subordinate faunistic role, became the dominant therapsids in the Upper Permian. The predatory Gorgonopsians came to an abrupt end as the second sterile trend towards life on drier ground when the marshes of the Lystrosaurus zone made life for their herbivorous prey impossible on dry land, coupled with the predators' inability to pursue the surviving lystrosaurs into the marshes. The herbivorous Dicynodontia, adapted to upland life, found the swampy conditions of the Lystrosaurus zone all but impossible and only the lystrosaurs could adapt themselves, but this temporary success proved their final undoing and in the later dry to arid Triassic only the kannemeyerids could eke out an existence.
Thus ends the third therapsid attempt towards an active mobile life on firm land.
The Therocephalia entered the Karoo Basin as a diversified suborder with two families in a strong predaceous role and two in an insectivorous role. The predators flourished in the Middle Permian but were ousted from this role by the gorgonopsians in the Upper Permian. The more insectivorous families continued successfully into the later Triassic, but where overshadowed faunistically (except for their cynodont offshoot) by the upsurging sauropsids. This fourth trend of the therapsids towards upland active life proved to be genetically fertile in that they gave rise to the first mammals in the Upper Triassic.

diagramatic phylogeny of the early therapsids. phenacodontid represented by Haptodus ptitanosuchid represented by Eotitanosuchus ithopid represented by Syodon ateosaurid represented by Anteosaurus tanosuchid represented by Fonkeria yracocephalid represented by Styracocephalus apinocephalid represented by Tapinocephalus thinosuchid represented by Phthinosuchus pposaurid represented by Hipposaurus llesuchid composite of the early genera cosuchid represented by Trochosaurus isterognathid represented by Glanosuchus aloposaurid represented by Blattoidealestes sheriid represented by Otsheria cynodontid represented by Dicynodon adothiodontid composite of early genera omasaurid represented by Galeops.


## INSTRUCTIONS TO AUTHORS

## Based on

CONFERENCE OF BIOLOGICAL EDITORS, COMMITTEE ON FORM AND STYLE. 1960 . Style manual for biological journals. Washington: American Institute of Biological Sciences.

## MANUSCRIPT

To be typewritten, double spaced, with good margins, arranged in the following order: (1) Heading, consisting of informative but brief title, name(s) of author(s), address(es) of author(s), number of illustrations (plates, figures, enumerated maps and tables) in the article. (2) Contents. (3) The main text, divided into principal divisions with major headings; subheadings to be used sparingly and enumeration of headings to be avoided. (4) Summary. (5) Acknowledgements. (6) References, as below. (7) Key to lettering of figures. (8) Explanation to plates.

## ILLUSTRATIONS

To be reducible to $12 \mathrm{~cm} \times 18 \mathrm{~cm}$ (19 cm including caption). A metric scale to appear with all photographs.

## REFERENCES

Harvard system (name and year) to be used: author's name and year of publication given in text; full references at the end of the article, arranged alphabetically by names, chronologically within each name, with suffixes $a, b$, etc. to the year for more than one paper by the same author in that year.
For books give title in italics, edition, volume number, place of publication, publisher.
For journal articles give title of article, title of journal in italics (abbreviated according to the World list of scientific periodicals. 4th ed. London: Butterworths, 1963), series in parentheses, volume number, part number (only if independently paged) in parentheses, pagination.
Examples (note capitalization and punctuation)
Bullough, W. S. 1960. Practical invertebrate anatomy. 2nd ed. London: Macmillan.
Fischer, P.-H. i948. Données sur la résistance et de le vitalité des mollusques. F. Conch., Paris 88: 100-140.
Fischer, P.-H., Duval, M. \& Raffy, A. 1933. Etudes sur les échanges respiratoires des littorines. Archs Zool. exp. gén. 74: 627-634.
Kohn, A. J. 196oa. Ecological notes on Conus (Mollusca: Gastropoda) in the Trincomalee region of Ceylon. Ann. Mag. nat. Hist. (13) 2: 309-320.
Kонл, A. J. 196ob. Spawning behaviour, egg masses and larval development in Conus from the Indian Ocean. Bull. Bingham oceanogr. Coll. 17 (4): 1-51.
Thiele, J. igio. Mollusca: B. Polyplacophora, Gastropoda marina, Bivalvia. In schultze, l. Zoologische und anthropologische Ergebnisse einer Forschungsreise im westlichen und zentralen SüdAfrika. 4: 269-270. Jena: Fischer. Denkschr. med-naturw. Ges. Fena 16: 269-270.

## ZOOLOGICAL NOMENCLATURE

To be governed by the rulings of the latest International code of zoological nomenclature issued by the International Trust for Zoological Nomenclature (particularly articles 22 and 51). The Harvard system of reference to be used in the synonymy lists, with the full references incorporated in the list at the end of the article, and not given in contracted form in the synonymy list.

## Example

Scalaria coronata Lamarck, 18ı6: pl. 451, figs 5 a, b; Liste: 11. Turton, 1932: 80.
?

```
\(50 \geqslant 68\)
```

ANNALS OF THE SOUTH AFRICAN MUSEUM ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM

| Volume | $\mathbf{5 9}$ | Band |
| :---: | :---: | :--- |
| March | $\mathbf{1 9 7 2}$ | Mart |
| Part | $\mathbf{3}$ | Deal |



LARVAL DEVELOPMENT OF THREE SPECIES OF ECONOMICALLY IMPORTANT SOUTH AFRICAN FISHES

By<br>E. H. HAIGH

# The ANNALS OF THE SOUTH AFRICAN MUSEUM <br> are issued in parts at irregular intervals as material becomes available <br> Obtainable from the South African Museum, P.O. Box 61, Cape Town <br> Die ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM <br> word uitgegee in dele op ongereelde tye na beskikbaarheid van stof <br> Verkrygbaar van die Suid-Afrikaanse Museum, Posbus 6I, Kaapstad <br> <br> out of print/uit druk <br> <br> out of print/uit druk <br> <br> 1, 2(1, 3, 5, 7-8), 3(1-2, 5, t.-p.i.), 5(2, 5, 7-9), <br> <br> 1, 2(1, 3, 5, 7-8), 3(1-2, 5, t.-p.i.), 5(2, 5, 7-9), $6(\mathrm{I}, \quad$ t.-p.i.), $\quad 7(\mathrm{I}-3), \quad 8, \quad 9(\mathrm{I}-2), \quad 10(\mathrm{I})$, $6(\mathrm{I}, \quad$ t.-p.i.), $\quad 7(\mathrm{I}-3), \quad 8, \quad 9(\mathrm{I}-2), \quad 10(\mathrm{I})$, 1I(1-2, 5, 7, t.-p.i.), 21, 24(2), 27, 3 I(i-3), 33 

 1I(1-2, 5, 7, t.-p.i.), 21, 24(2), 27, 3 I(i-3), 33}

Price of this part/Prys van hierdie deel R2,20

Trustees of the South African Museum © Trustees van die Suid-Afrikaanse Museum 1972

In Suid-Afrika gedruk deur
Die Rustica-pers, Edms., Bpk.
Courtweg, Wynberg, Kaap

# LARVAL DEVELOPMENT OF THREE SPECIES OF ECONOMICALLY IMPORTANT SOUTH AFRICAN FISHES 

By<br>E. H. Haigh

South African Museum, Cape Town
(With II figures and 9 tables)
[MS. accepted I September 1971]
Contents

| Introduction |  |  |  |  | Page |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  | - | . | . | - | 47 |
| Material and methods |  | . | - | . | 49 |
| Description |  | - | - | - | 50 |
| Merluccius capensis |  | . | . | - | 50 |
| Thyrsites atun |  | - | - | - | 55 |
| Helicolenus dactylopterus |  | - | - | - | 60 |
| Distribution | - | . | - | - | 69 |
| Summary | - | - | - | . | 69 |
| Acknowledgements |  | - | - | . | 69 |
| References |  | - | - | - | 69 |

## Introduction

This paper is the first of a series of studies on larval fish development. Mainly economically important species will be described although other species of interest will also be included.

The species described in this paper are Merluccius capensis, the Cape stockfish, Thyrsites atun, the snoek, and Helicolenus dactylopterus, the jacopever.

The Cape stockfish or hake is of great economic importance in South Africa. The annual trawled catch has ranged from 68 oig 223 to 70686775 kilo over the five years from 196ı to 1965 (Ann. Rep. Div. Sea Fish. S. Afr. 33). At the end of 1970 the trawled landings were reported to be 64 million kilo. Irvin \& Johnson (1963) give an account of the economic importance and habitat of the adult. Although the adult is almost exclusively demersal, the larvae are caught in plankton nets, indicating a pelagic mode of life. However, the number of larvae, especially the older, larger forms, is relatively small in the samples.

The snoek is an important predator of the pilchard Sardinops ocellata and related pelagic fish. It is a large rapacious carnivore, with excellent, firm flesh and thus also an important seasonal food fish in South African waters. Annual catches vary between 7,2-9 million kilo per annum.

The jacopever is uncommon in fishmarkets around the coast, the larger number being caught in trawls and used in fishmeal manufacture. Between 196I and 1965 the landings ranged between 857427 and I 487624 kilo (Ann.

Rep. Div. Sea Fish. S. Afr. 33). Smith (1953) reports its flesh to be palatable. However it does not constitute a major portion of the fishing resources of the country.

Davies (1949) states that Helicolenus inhabits waters between 90 and 360 metres. It is a bottom-dwelling fish usually on the continental shelf. Superficially it is very like the Tristan da Cunha scorpaenid Sebastichthys capensis and several authors have reported both genera to be viviparous. Specimens between 3,5 and $4,2 \mathrm{~mm}$ standard length had well-developẻd jaws (indicating functionality) and several head spines. Davies (1949) suggests a November spawning season, Ahlstrom (1961) on the other hand suggests a winter and early spring release of young of Sebastodes spp. on the west coast of the United States. Moser (1967) states that there are two broods released in Sebastodes paucispinis, one in autumn and one in spring. The majority of samples in our collection were caught in late spring, October. These represent a complete range in sizes. As the collection is not very large, no conclusions can be drawn from this.

The three species described in this paper are classified as follows:

| Order: | Gadiformes | Perciformes | Scorpaeniformes |
| :--- | :--- | :--- | :--- |
| Suborder: | Gadoidei | Scombroidei | Scorpaenoidei |
| Family: | Merlucciidae | Gempylidae | Scorpaenidae |
| Genus: | Merlucius | Thyrsites | Helicolenus |
| Species: | capensis | atun | dactylopterus |
|  | Castelnau 1861 |  | Euphrasen 1791 | Delaroche 1809

The family Merlucciidae is distinguished by a separate caudal fin. According to Norman (1937, 1966) it has one genus comprising seven species, three in the northern and four in the southern temperate zones. Although Gilchrist (1921) and Barnard (1925) doubt the distinction made between the European species Merluccius merluccius and M. capensis, Norman (1937) confirms Regan's (1908) distinction. Ginsburg (1954) sheds more light on the American species of the Merlucciidae.

The Gempylidae is a small family comprising io genera each with only a small number of species. Its taxonomic history seems to have been untroubled.

The taxonomy of the family Scorpaenidae needs world-wide revision. Eschmeyer (1969) gives a good review of the Atlantic Scorpaenidae, synonymizing Helicolenus maculatus with H. dactylopterus and separating the Tristan da Cunha species from H. dactylopterus. However, he does state that the gradient in characters is rather disjointed and that a conclusive synonymy needs a more comprehensive study of material.

Eggs and larvae of Merluccius merluccius have been described by Ehrenbaum (1909) and D'Ancona (1933); of M. bilinearis by Kunz \& Radcliffe (1917); of M. productus by Ahlstrom \& Counts (1955). Marak (1967) describes the early pro-larvae of $M$. albidus and distinguishes them from the pro-larvae of $M$. bilinearis. Fischer (1959) describes eggs and pro-larvae of M. gayi from Chile
as do Santander \& Castillo (1969) from the coast of Peru. Hart \& Marshall (1951) report a larval Merluccius capensis between $19^{\circ}$ and $22^{\circ} \mathrm{S}$, extending the possible range further north than this present survey. Matthews \& De Jager (1951) described the development of the egg and pro-larva of $2,35 \mathrm{~mm}$ for M. capensis. Larvae of the genus Merluccius show a basic similarity in pigmentation, allowing for easy recognition.

Larvae of species of the family Gempylidae have been described: Gempylus serpens by Jones (1960); Nesiarchus nasatus and Gempylus spp. by Voss (1954) and Thyrsites atun by Regan (1914-16). De Jager (1955) described the development of artificially fertilized eggs and resultant larvae of Thyrsites atun up to the age of nine days and a length of $3,9 \mathrm{~mm}$. In the present paper the development from $4,6 \mathrm{~mm}$ to 25 mm is described, thus completing the description of Thyrsites atun development.

In the family Scorpaenidae larval development of two species of Sebastodes has been described by Ahlstrom (1963) and Sebastes marinus has been described by Bigelow \& Welch (1924). A paper on the distribution of Sebastodes spp. in Californian waters was published by Ahlstrom in 1961. In this paper he discussed briefly the distinguishing features of some scorpaenid larvae in the eastern North Pacific. However, Moser (1967) gives the complete development of Sebastodes paucispinis and gives illustrations and a list of characters of early stages for 14 further species.

## Material and methods

Specimens were obtained by research vessels of the Division of Sea Fisheries, Cape Town, using NiooB and NiooH plankton nets, from 1950 to 1967. Samples were fixed and stored in formalin which was replaced by $70 \%$ ethyl-alcohol. Specimens were stained, using the methods of Hollister (1934), Davies \& Gore (1935) and Moran (1956) but modified slightly by reducing the clearing time in KOH and reducing the concentration of the KOH used. This was done in order to preserve the pigment of the specimens. As pigments are inclined to fade, more than one larva in the size range was used in order to obtain the most characteristic pigmentation pattern. Stained specimens were preserved in glycerin and ethyl-alcohol.

Measurements were taken as follows:
standard length (s.l.): tip of lower jaw to caudal peduncle
snout:
eye diameter
head length:
trunk:
depth:
pelvic and first dorsal spine lengths for Thyrsites atun only

All proportions presented as percentage of standard length. All lengths cited in text are the standard lengths of the specimen.

Some specimens that should have been well ossified did not absorb stain properly. This was most probably due to decalcification of the bone by formalin. Vertebral counts include urostylar complex which is counted as two.

## Description

## Merluccius capensis

Merluccius capensis is characterized by 130-140 scales in longitudinal series, 13-14 gillrakers in lower part of anterior arch and a pectoral with 13-14 rays reaching to beyond the origin of the anal, while the pelvic extends nearly to the vent. Depth is $60 \%$ of length and headlength is $32-36 \%$ of length. The maxillary extends to below the posterior edge of the pupil or beyond and is less than half of head-length. $\mathrm{D}: 10-11 ; 35-40$. A:37-40 (Norman 1937).

## Pigmentation

The general pigmentation pattern of $M$. capensis is similar to that of M. merluccius as described by Ehrenbaum (1909) and D'Ancona (1933). The major pigmentation on the head consists of one or two large stellate melanophores situated at the postero-dorsal edge of the brain and anterior to the first dorsal fin-occipital spot. The dorsal peritoneal wall is always darkly pigmented with both stellate and closed chromatophores.

The tails of most larvae examined bear three areas of pigmentation. An anteropostanal spot situated latero-ventrally just behind the anus; a larger mediopostanal spot, covering the whole lateral surface of the tail, midway between the anus and the caudal fin; and one or two stellate melanophores comprising the caudal spot situated latero-ventrally on the caudal peduncle.

These pigmented areas are characteristic of the species and are to a greater or lesser extent augmented by smaller stellate and contracted melanophores at the dorsal aspect of the head and ventral aspect of the abdomen. These vary a great deal in intensity within a size group but generally increase in size with age during larval life.

In the early post-yolk-sac stage larvae (Fig. ra) the head bears a pigment spot on the anterior edge of the brain which becomes obscured as the larvae get larger and ossification commences. The cerebral area of the head is further dotted with a varying number of melanophores. With further development, melanophores may also appear on the jaw, around the eyes and on the opercular surface of the head (Figs Id, 2a).

Peritoneal pigmentation remains fairly constant throughout development. The melanophore anterior to the first dorsal fin becomes augmented by smaller stellate melanophores along the sides of the first dorsal and later the second dorsal fin.

The anteropostanal, mediopostanal and caudal spots remain relatively the same size throughout the larval life but in prejuvenile and juvenile stages,

b $5,5 \mathrm{~mm}$


Fig. 1. Merluccius capensis
Early larval stages showing position of major pigmentation areas and ossification development. Measurements indicate standard length.
they appear to become smaller and more discrete. They probably break up to form the smaller melanophores that abound on the dorsolateral sides of the juvenile (Fig. 2c).

Small, discrete pigment spots are present on the pectoral and pelvic fin-
buds of some of the larvae. As soon as ossification of the pelvic fins is complete, pigmentation appears on both fins. No caudal fin pigmentation was observed.

Pigment spots appeared on the dorsal fin late in the larval and early juvenile stages. Pigmentation was also present on the dorsal head and abdominal surfaces in juveniles.

## Ossification

The premaxilla, maxilla, mandible and cleithrum are ossified in larvae $3,6 \mathrm{~mm}$ long. The first traces of the gill-arches can be seen in slightly larger specimens. The branchiostegal rays then ossify progressively from dorsal to ventral (Fig. Ib). The premaxilla at $5,5 \mathrm{~mm}$ has between four to six teeth and the dentary four to six. The supracleithrum and posttemporal appear as slender rods. By $6,1 \mathrm{~mm}$ the epihyal and ceratohyal are well formed and bear six branchiostegal rays. The first traces of the preopercle, cranial and opercular ridges are ossified and there are eight mandibular teeth. By 7 mm the hyomandi-

Table 1
Ossification of skeletal elements of Merluccius capensis

| Size <br> in mm | $\underset{\text { Dorsal }}{\text { I }}$ | $\begin{gathered} \text { II } \\ \text { Dorsal } \end{gathered}$ | Anal | Caudal | Vertebrae | Neural spines | Haemal spines | Pectoral rays | Pelvic rays |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 4,5 | - | - | - | - | - | 3 | - | - |  |
| 5,4 | - | - | - | 6 | - | 4 | - | - | 2 |
| 6,0 | - | - | - | 8 | - | 6 | - | - | 3 |
| 6,6 | 2-3 | - | - | 12-14 | 8-13 | $8-10+2-5$ | 3-5 | - | 5 |
| 6,9 | 3 | - | - | 15 | $12+5$ | $8+5$ | 4 | - | 6 |
| 7,8 | 5-7 | $6-8+4-5$ | $\begin{gathered} 15-17+ \\ 5^{-7} \end{gathered}$ | 20-21 | $4^{1-42}$ | 50-54 | 24-25 | - | 7 |
| 9,0 | 6-7 | 26 | 30 | 19-21 | 47-49 | 54 | 24 | - | 9 |
| 10,2 | 8 | 34 | 33 | 28 | 57 | 54 | 27 | - | 9 |
| 12,0 | 9 | 35 | 35 | 30 | 57 | 54 | 29 | - | 9 |
| 14, I | 11 | 32 | 39 | 40 | 57 | 54 | 29 | 7 | 9 |

bular can be clearly distinguished and the quadrate has started ossifying in the articular region. The pterygoid has also started forming and the seventh branchiostegal is complete. Other bones have thickened and widened considerably. By $9,6 \mathrm{~mm}$ parietal and frontal bones are well formed, the nasals have started ossifying and so have the lacrymals. The first trace of the operculum is present, underlying the opercular ridges. The visible bones in the head have now been fully laid down and further development takes place to reach the juvenile stage between 14 and 16 mm (Fig. 2d).

The anterior neural spines ossify first, followed by the anterior centra. At $6,8 \mathrm{~mm}$ the first seven centra and five to eight haemal spines are formed. However, neural and haemal spines of some of the caudal vertebrae also show signs of ossification at this stage. During this growth-period the centra are rapidly laid down and at $9,6 \mathrm{~mm}$ all the vertebrae except the five preceding the last centrum have been fully formed. The last vertebra is ossified but the urostyle is still cartilaginous. At this stage the hypural and epural elements show some degree of ossification. Haemal spines are only found from the 26th


Fig. 2. Merluccius capensis
a-b. Late larval stages. c. Juvenile. d. Lateral view: 1. Lacrymal. 2. Maxilla. 3. Premaxilla. 4. Dentary. 5. Circumorbitals. 6. Pterygoid. 7. Quadrate. 8. Articular. 9. Preopercle. 10. Subopercle. 11. Branchiostegal rays. 12. Opercle. 13. Cleithrum. 14. Postcleithrum. 15. Pelvic girdle. 16. Pelvic rays. 17. Pectoral rays. 18. Anal rays. 19. Caudal rays. 20. Second dorsal fin. 21. First dorsal fin. 22. Supracleithrum. 23. Posttemporal. 24. Cranial bones. 25. Hyomandibular.
26. Frontal. 27. Nasal.
centrum at this stage, indicating the first 24 to 25 vertebrae to be abdominal. By $13,2 \mathrm{~mm}$ all the parapophyses of the abdominal vetebrae, except the first six, have enlarged somewhat. In the juvenile there are 18 abdominal vertebrae bearing parapophyses expanded lateroventrally (Fig. 2d).

Fin formation takes place in the following sequence: pectoral fin bud, caudal buds and pelvic bud between 3,0 and $5,0 \mathrm{~mm}$. In size range 5,0 to $6,0 \mathrm{~mm}$ the pelvic rays ossify, then the caudals start, followed by the first dorsals at between 5,5 and $6,8 \mathrm{~mm}$. The anal and second dorsal rays follow almost immediately. Pectoral rays only appear much later. By 6 mm the first five pelvic rays have appeared in the pelvic bud. The caudal buds appear at about $4,5 \mathrm{~mm}$. The ventral caudal rays appear first, followed at 6 mm by the dorsal caudal rays. Several caudal rays seem to ossify simultaneously. In some larvae of 6 mm the basal buds of the first four dorsal rays are also evident.

Although the anal fin starts ossifying only after the first dorsal, its development proceeds faster and by $6,8 \mathrm{~mm}$ between six and eight rays are ossified in the anal while only five dorsal rays can be distinguished. The pelvic now has six ossified rays and the caudal has 14 .

The pelvic attains its full complement of seven rays at approximately 9 mm . The caudal, dorsal and anal fins have an almost complete number of rays at $13,2 \mathrm{~mm}$ but as so few larvae over this size were obtained there is no certainty about the exact size when full ray number is attained. Ahlstrom \& Counts (1955) give 16 mm as the size where the full complement of both dorsal and anal fin rays is present in M. productus.

Seven pectoral fin rays were observed at $14,1 \mathrm{~mm}$. The juvenile of 37 mm has a full number of 14. M. productus larvae only develop pectoral fin rays at 24 mm and later (Ahlstrom \& Counts, 1955).
Changes in body form and growth rate
The eye diameter is larger than the snout length in the earlier stages but this difference is gradually diminished and by $6,0 \mathrm{~mm}$ they are approximately equal in length. The snout becomes longer than the eye diameter from $10,5 \mathrm{mms.l}$.

Table 2
Mean measurements in mm and proportions (\% of s.l.) of Merluccius capensis

| Size group | $\mathcal{N}$ o. | Snout l. |  | Eye diameter |  | Head l. |  | Trunk $l$. |  | Depth |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | \% |  | \% |  | \% |  | \% |  | \% |
| 2,5-3,49 | 2 | 0, 13 | 4,57 | 0,31 | 11, I2 | 0,64 | 22,44 | 1,71 | 63,33 | 0,98 | 33,05 |
| 3,5-4,49 | 9 | 0,25 | 6,57 | 0,32 | 8,10 | 1,07 | 25,70 | 1,73 | 43,35 | 1,06 | 26,60 |
| 4,5-5,49 | 21 | 0,48 | 9,91 | 0,53 | 11,07 | 1,43 | 28,39 | 2,86 | 46,69 | 1,40 | 29,55 |
| 5,5-6,49 | I I | o,62 | 10,32 | 0,62 | 10,32 | 1,89 | 31,08 | 2,80 | 48,65 | 1,69 | 30,60 |
| 6,5-7,49 | 18 | 0,65 | 8,65 | 0,65 | 8,65 | 1,96 | 27,10 | 3,23 | 46,54 | ı,66 | 26,09 |
| 7,5-8,49 | 13 | 0,71 | 9,67 | 0,73 | 9,36 | 2,30 | 30,24 | 3,66 | 46,61 | 2,03 | 25,81 |
| 8,5-9,49 | 1 | 0,90 | 10,00 | 0,90 | 10,00 | 2,40 | 26,66 | 4,20 | 46,66 | 2,10 | 23,33 |
| 9,5-10,49 | 8 | 0,97 | 8,86 | 0,90 | 9,22 | 2,92 | 30,01 | 4,60 | 47,55 | 2,40 | 25,02 |
| 10,5-11,49 | 6 | 1,05 | 9,73 | 0,90 | 8,33 | 3,20 | 30,09 | 4,90 | 45,35 | 2,40 | 22,71 |
| $\dagger$ - |  |  |  |  |  |  |  |  |  |  |  |
| 13,5-14,49 | 3 | 1,30 | 9,33 | 1,10 | 7,88 | 3,70 | 26,57 | 6,20 | 44,63 | 3,00 | 21,60 |
| * 20,40 | 1 | 1,80 | 8,82 | 1,50 | 7,35 | 6,60 | 32,35 | 9,60 | 47,05 | 4,20 | 20,58 |
| 26,60 | I | 2,10 | 7,89 | 1,80 | 6,76 | 7,20 | 27,06 | 11,40 | 42,85 | 5,10 | 19, 17 |
| 36,30 | 1 | 3,80 | 10,46 | 2,80 | 7,71 | 1 1,30 | 31,12 | 17,00 | 46,83 | 7,50 | 20,66 |
| 41,00 | I | 4,50 | 10,97 | 3,00 | 7,31 | 13,00 | 31,70 | 19,00 | 46,34 | 8,00 | 19,5 |
| 46,00 | I | 5,00 | 1 0,86 | 3,00 | 6,52 | 13,50 | 29,34 | 21,20 | 46,08 | 8,00 | 17,39 |

The head is $24 \%$ of the standard length in the smallest stages $(2,9$ to $4,4 \mathrm{~mm}$ ), increasing to $28 \%$ at 4,5 to $5,4 \mathrm{~mm}$ and remaining 25 to $32 \%$ of standard length for the rest of development from 5,5 to 46 mm . The head shows an average rate of increase of $0,36 \mathrm{~mm}$ for each millimetre increase in standard length.

The proportion of the trunk to the standard length varies between 42 and $48 \%$ but is usually about $45 \%$ throughout development and can be used as a taxonomic character (Ahlstrom \& Counts, 1955). The trunk shows an average increase of $0,47 \mathrm{~mm} / \mathrm{mm}$ increase of standard length. Up to a length of $9,75 \mathrm{~mm}$ the depth increases $0,23 \mathrm{~mm} / \mathrm{mm}$ increase in standard length, but between $9,75 \mathrm{~mm}$ and $13,2 \mathrm{~mm}$ the increase is only $0,127 \mathrm{~mm} / \mathrm{mm}$ increase in standard length. Thus the hake changes from a rather deep tadpole-like post-larva to a slender, evenly sloping juvenile (Fig. 2c).

## Thyrsites atun

The snoek is characterized by the maxilla reaching slightly beyond the anterior border of the eye, the long mandible projecting the upper jaw and reaching to the posterior third of the eye. Both jaws carry large canines. The caudal is deeply forked.
D: $18 \mathbf{- 2 1}+\mathbf{1 0 - 1 2 + 6 . A : 1 - 2 + 8 - 1 1}+6$. Pectoral: $2+1$ I. Pelvic: $\mathrm{I}+5$. Vertebrae: 34-35. Depth $\pm 7$ (Smith 1953; Fowler 1936; Beaufort \& Chapman 195I: 199).

## Pigmentation

Standard pigmentation in larvae of Thyrsites atun between 4,0 and 10,0 mm consists of a variable number of small stellate and closed melanophores over the snout and cerebral areas of the head. A dark area of pigmentation, standard in all larvae, is found on the antero-dorsal and lateral areas of the peritoneum. Small scattered spots are also to be found posteriorly, above the anus. The pigmentation is darker in smaller specimens of four to six millimetres, becoming more diffuse and evenly distributed as the fish grows larger. When the snoek larvae are II to 15 mm long, abdominal pigmentation consists of scattered stellate melanophores.

Most characteristic and stable are the two areas of tail pigmentation. On the ventral surface of the tail, midway between anus and anal fin, is a smaller spot consisting of only one melanophore. Also ventrally situated is the posterior pigment area between the anal and caudal fin. This spot consists of several stellate melanophores clustered together, and covers two to three times the area of the anterior pigment spot. However, these areas of tail pigmentation do not appear in the specimens figured by De Jager (1955). I have had occasion to examine the specimens of De Jager and his figures agree reasonably well with the specimens. It would appear that rapid migration of pigment takes place in early larval life. The pigment pattern described above is only evident in the last stage of De Jager's larvae at $3,9 \mathrm{~mm}$ but unfortunately is not obvious
in his Figure 18. The two tail pigment areas only disappear in juveniles over 16 mm and constitute a good diagnostic feature for the larvae of the species.

The characteristic black dorsal pigmentation of the adult snoek begins to show up in larvae of $6,0 \mathrm{~mm}$. On each side of the dorsal spines a thin line of black pigment appears, which becomes thicker and more obvious as the larvae grow. This line is characteristic of the Gempylidae. None of the fins shows pigmentation during development (Fig. 3).

## Ossification

At $4,6 \mathrm{~mm}$ the premaxilla with four teeth, the maxilla, and the mandible with two teeth are ossified as well as the cleithrum. Ossification of the preopercle has begun and two spines may be distinguished on the margin as well as one small spine originating in the middle of the preopercle in line with the


Fig. 3. Thyrsites atun
Early larval stages showing position of major pigmentation areas and ossification development. Measurements indicate standard length.
most dorsal spine. The first gill arches and five branchiostegals are formed.
Between 5 and 6 mm the premaxilla develops four more teeth and the anterior canines also start developing. The mandibular teeth increase to six. The palatine is clearly visible. Preopercular spines increase to four and the branchiostegals to six. Traces of the pterygoid, quadrate, symplectic, and opercle are ossified as well as the posttemporal and postcleithrum. The ceratohyal can also be faintly discerned. By 8 mm traces of the hyomandibular are laid down and the preopercular spines increase to five. By 10 mm palatine teeth develop. The cranial bones have by now become clearly ossified and traces of the subopercle are present. By ${ }_{11,5} \mathrm{~mm}$ traces of the nasals can be seen, the fangs are well developed (two on each side) and three palatine teeth are present. The subopercle is quite clear and the hyomandibular is well formed. By $20,0 \mathrm{~mm}$ the jaws are heavily ossified, the preopercle has assumed a more median position and the six preopercular spines are not as obvious as before. The quadrate is well ossified, obscuring part of the symplectic. Frontal and parietal bones are well formed but there are still wide gaps between them. The bones of the pectoral girdle have become much wider and flanges have developed on the posttemporal and cleithrum.

There are no traces of ossification in the vertebral column at $4,6 \mathrm{~mm}$, but between 5,4 and $6,0 \mathrm{~mm}$ the basic elements of the neural spines appear in some specimens. At $6,6 \mathrm{~mm}$ at least three, usually more, neural spines are ossified. Between 6,6 and 7,2 mm the first centra of the vertebrae become ossified and by $9,75 \mathrm{~mm}$ at least 18 vertebrae are ossified. By $8,15 \mathrm{~mm}$ the first haemal spines appear. The ossification of the vertebral spines exceeds that of the centra. The full complement of the haemal spines is ossified at $11,4 \mathrm{~mm}$. Of these, II are shorter abdominal and 14 to 15 are larger caudal spines. Only at 13 to 14 mm is the full complement of neural spines laid down and by 14 mm the vertebral centra are also fully ossified.

The vertebral centra ossify from the dorsal and ventral peripheries inward, except for the last three vertebrae where ossification proceeds dorsally from the ventral periphery. In these three vertebrae the neural spines also form later than in the other vertebrae. The snoek has 21 abdominal and 13 to 14 caudal vertebrae.

The urostyle begins to turn up at $7,2 \mathrm{~mm}$ and the first traces of urostylar and hypural ossification are evident at $9,0 \mathrm{~mm}$. At $11,0 \mathrm{~mm}$ the urostyle is fully ossified and bears two dorsal and three ventral hypural elements. The haemal spines of the ultimate and penultimate vertebrae have broadened to support the caudal fin.

The pectoral lobe is evident at $4,5 \mathrm{~mm}$ but the first fin ossification appears in the pelvic spine at $5,4 \mathrm{~mm}$. By $6,0 \mathrm{~mm}$ the initial ossification of the pelvic girdle has started and by $8,15 \mathrm{~mm}$ the pelvic spine has achieved its characteristic serrate appearance and proportional full length. The pelvic rays develop gradually until the full complement of five is attained at $14,0 \mathrm{~mm}$.

The first dorsal fin starts ossifying anteriorly between 5,5 and $6,0 \mathrm{~mm}$
and this proceeds rapidly posteriorly. At $7,2 \mathrm{~mm}$ there are 10 to 13 rays present, the most anterior having become hard and serrate. The second dorsal fin has traces of 10 to I 5 rays ossified at $8,15 \mathrm{~mm}$ and at $9,5 \mathrm{~mm}$ the distinction between the first and second dorsal is clear. There are 19 to 20 spines and 12 to 16 soft rays. However it is not until later in the juvenile stage that the finlets differentiate from the second dorsal. The only distinction that can be made, even at 20 mm , is that the last five rays are more widely separated than the rest.

Table 3
Table of skeletal elements of Thyrsites atun larvae at different stages

| $\begin{aligned} & \text { Size } \\ & \text { in } m m \end{aligned}$ | No. of specimens | Pectoral | Pelvic | Caudal | $\begin{aligned} & \text { Dorsal } \\ & I \text { \&尺 } I I \end{aligned}$ |  | Anal | Haemal spine | Neural spine | Vertebrae |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 5,1 | 2 | o | 0 | 0 | 0 | 0 | 0 | o | 0 | 0 |
| 5,4 | 6 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 6,0 | 5 | 0 | 1 | 0 | $4^{-8}$ | 0 | 0 | 0 | 0 | 0 |
| 6,6 | 6 | 0-4 | 1 | 0-4 | 9-13 | 0 | 0 | 0 | 0-31 | 0 |
| 7,2 | 6 | 4-6 | 1 | 5-8 | 10-13 | 0 | 3-6 | 0 | 7 | 6 |
| 8,0 | 6 | 7-9 | 1 | 10 | 15 | +10 | 7-10 | $5^{-10}$ | 17 | 15 |
| 9,5 | 6 | 12 | 1 | 17 | 19 | +14 | 1 1-12 | 16-19 | 20-27 | 18-20 |
| 11,4 | 5 | 13 | I +2 | $17+5$ | 19 | +17 | 13 | 23-25 | 28-32 | 25-28+2 |
| 14,0 | 1 | 13 | $\mathrm{I}+5$ | $17+12$ | 19-20 | +16-17 | $1-2+14$ | 23-25 | 34 | 34-36 |
| 20,0 | 1 | 14 | $\mathrm{I}+5$ | $17+10$ | 21 | $+16$ | $2+15$ | 26 | 35 | 36-37 |

The next fin-rays to ossify are those of the caudal and pelvic fins which do so almost simultaneously between $6,0 \mathrm{~mm}$ and $6,6 \mathrm{~mm}$. At $8,0 \mathrm{~mm}$ there are seven to nine pectoral rays and at $9,5 \mathrm{~mm}$ II to 13 , while the pectoral girdle has started to ossify. At II,5 mm the coracoid, scapula and radials have formed. In the caudal fin the ventral fin-rays are completed first, followed shortly after by the dorsal rays at $9,5 \mathrm{~mm}$. Between 10 mm and ${ }_{1} 5 \mathrm{~mm}$ the secondary caudal rays appear. By 14 mm the caudal fin is fully ossified except for some small secondary fin-rays. The caudal has eight ventral and nine dorsal primary rays.

The last fin to start ossifying is the anal fin. By $7,2 \mathrm{~mm}$ only traces of the first three to six rays are laid down and by $8,15 \mathrm{~mm}$ at least the first seven rays are ossified. The full complement of 16 rays is only laid down by 14 mm . In the specimens examined, the large majority had only one anal spine; a few had anal fins with two spines. As with the second dorsal fin, the anal finlets are not differentiated until late in the juvenile development.

The dorsal spines increase rapidly in actual and relative length between 6 and $7,5 \mathrm{~mm}$ and remain at 13 to $14 \%$ of standard length till 14 mm length is reached. After this the relative length decreases somewhat to about $8 \%$ of standard length by 21 mm .

## Changes in body form

The very young snoek larva has a large head and short abdomen with a fairly long tail (Fig. 4a). The head is $30,6 \%$ of standard length at $5,4 \mathrm{~mm}$ and increases to $35,2 \%$ by $8,5 \mathrm{~mm}$, more or less retaining that proportion till the juvenile stage is reached where it diminishes proportionately as standard length increases. The head increases by $0,42 \mathrm{~mm} / \mathrm{mm}$ increase in standard length


Fig. 4. Thyrsites atun
Larvae showing increase in size of area occupied by viscera.
between 4,08 and $14,25 \mathrm{~mm}$. After this size is reached, increase in head length appears to decrease and reach $0,225 \mathrm{~mm} / \mathrm{mm}$ increase in length.

The snout is $37 \%$ of the head length. Initially the snout is only $0,09 \mathrm{~mm}$ longer than the eye diameter, but this difference increases with age until at $21,0 \mathrm{~mm}$ the snout length is $\mathrm{I}, 05 \mathrm{~mm}$ greater than the eye diameter (Table 4).

Table 4
Mean proportions of Thyrsites atun larvae presented as \% of s.l.
Size range of
standard length
in mm
$4,08-5,49$
$5,5^{-6}, 49$
$6,5^{-7,49}$
$7,5^{-8}, 49$
$8,5^{-9}, 49$
$9,5^{-10,49}$
$10,5^{-11}, 49$
12,6
13,5
14,0
20,17
21,0

| No. of larvae | Head | Depth | Trunk | Ist Dorsal |
| :---: | :---: | :---: | :---: | :---: |
| - II | 30,6 | 21,4 | 42,0 | - |
| 7 | 32,6 | 21,2 | 45, I | 7,4 |
| 22 | 32,6 | 20,6 | 47,9 | 11,0 |
| 13 | 35,2 | 21,0 | 55,2 | 13,2 |
| 11 | 37,8 | 20,8 | 64,8 | 14,7 |
| 7 | 36,8 | 20,0 | 66,03 | 13,7 |
| 3 | 37,5 | 19,7 | 71,2 | 14,7 |
| I | 35,7 | 20,2 | 69,0 | 13,0 |
| 1 | 35,5 | 18,8 | 64,4 | 13,3 |
| 2 | 36,2 | 21,2 | 74,8 | 13,8 |
| 2 | 33,0 | 16,2 | 76,2 | 11, I |
| 1 | 31,4 | 16, I | 73,3 | 8,5 |

The ante-anal length or trunk length is initially $42,0 \%$ of the standard length. This, however, does not remain constant but increases as shown in Table 4. As far as can be judged from material available, the area occupied
by the intestine and viscera increases rapidly as the larva reaches 7 mm (Fig. 5 and Table 4). This rapid increase of an average of $0,98 \mathrm{~mm} / 0,95 \mathrm{~mm}$ increase in standard length continues till in mm s.l. is reached when the rate. of increase of the trunk slows down somewhat. At this stage (II mm s.l.) the anus has reached the origin of the anal fin, i.e. its juvenile position. It is possible that the rapid rate of visceral increase is linked to the change in the diet of the larvae. The larvae cease to feed on phytoplankton and become predatory on other fish. Head and eyes of larval fish have been found in the gut of snoek larvae as small as 8 mm .

Proportional depth of the snoek larvae remains fairly constant, between 18 and $21 \%$ of standard length, until $14,0 \mathrm{~mm}$ is reached, then it drops gradually to $16 \%$ of standard length at $21,0 \mathrm{~mm}$. Actual depth increase is of the order of $0,17 \mathrm{~mm}$ for each mm of standard length increase up to $13 \mathrm{~mm} \mathrm{s.l}$. After this, accuracy of calculation breaks down due to the small number of available specimens, but the rate appears to be less and in the order of $0,056 \mathrm{~mm} / \mathrm{mm}$ increase in s.l. Generally the larvae seem to become slimmer as they reach the juvenile stage.


Fig. 5. Thyrsites atun
Late larval and juvenile stages. Measurements indicate standard length.

## Helicolenus dactylopterus

The pectoral of Helicolenus dactylopterus is emarginate on the dorsal edge, with two unbranched rays followed by eight branched and nine unbranched
rays, the latter being free from the membrane for at least one-third of their length. Suborbital keel smooth with one spine small or absent, the mouth large with villiform teeth on the jaws, vomers and palatine. The maxilla reaches to below the hind margin of the eye. The spination of the adult head is as follows: i nasal, 2 supraorbital, 2 parietal, i pterotic, 2 posttemporal, 2 opercular, I small suborbital, 5 preopercular, second the longest (Fig. 6c). Soft dorsal higher than spinous dorsal. Pelvic reaches almost to vent. D: $12+{ }_{12-13}$ A: $3+5$. Pelvic: $\mathrm{I}+5$. Vertebrae: $\mathbf{2 4 - 2 5}^{-25}$.

Smaller specimens have black pigments near the end of the spinous dorsal. Pigment on body of juveniles in vertical bands (Eschmeyer 1969: 92-99).

## Pigmentation

All specimens from $3,5 \mathrm{~mm}$ have a clearly pigmented peritoneum with scattered melanophores on the posterior aspect of the head. This peritoneal colouring is still visible at 20 mm on the dorsal surface of the peritoneum. The pectoral fins in this species are unpigmented whereas some other scorpaenid larvae in the collection have variously pigmented pectoral fins.

## Ossification

Even the smallest larvae obtained had well ossified head spines and jaws as well as a cleithrum. By $4,5 \mathrm{~mm}$ (Fig. 7a) the premaxilla and maxilla are clearly defined, as is the lower jaw. The parietal, pterotic and posttemporal head spines are developed. The opercle is small and lightly ossified while the preopercle has three well-developed primary spines on the outer edge and two secondary spines on the median ridge. The middle of the primary spines is the longest. Four branchiostegal rays are present.

By $6,0 \mathrm{~mm}$ the frontal, parietal and pterotic bones are ossified but still easily distinguished. The hyomandibular has developed and the cleithrum is wider and better ossified than the $4,5 \mathrm{~mm}$ stage. A small subopercle is present.

Table 5
Mean measurements of Thyrsites atun larvae in mm

| Size range of standard length | No. of larvae | Standard length s.l. | Head | Snout | Eye diameter | Depth | Snout to anus | Length Pelvic spine | Longest dorsal spine |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 4,08-5,49 | 11 | 4,08-5,4 | 1,55 | 0,588 | 0,501 | 1,08 | 2,12 | - $(0,3)$ | $-(0,3)$ |
| 5,5-6,49 | 7 | 5,7-6,0 | 1,80 | o,681 | o,608 | 1,26 | 2,69 | 0,35 | 0,45 |
| 6,5-7,49 | 22 | 6,6-7,35 | 2,30 | o,86o | o,680 | 1,43 | 3,34 | o,77 | 0,76 |
| 7,5-8,49 | 13 | 7,5-8,42 | 2,88 | o,93 | o,87 | 1,70 | 4,52 | 1,24 | 1,09 |
| 8,5-9,49 | 11 | 8,55-9,45 | 3,42 | 1,32 | o,96 | 1,88 | 5,84 | 1,73 | 1,32 |
| 9,5-10,49 | 7 | 9,6-10,24 | 3,60 | 1,52 | 1,12 | 1,97 | 6,48 | 2,01 | 1,34 |
| 10,5-11,49 | 3 | 10,5-10,8 | 4,00 | 1,56 | 1,04 | 2,11 | 7,60 | 2,26 | 1,58 |
| 11,5-13,5 | 2 | 12,6 | 4,50 | 1,80 | 1,14 | 2,55 | 8,70 | 2,10 | 1,65 |
|  |  | 13,5 | 4,80 | 1,80 | 1,65 | 2,55 | 8,70 | 2,40 | 1,80 |
| ${ }^{13,5-15,5}$ | 2 | 13,8 | 4,8 | 2,10 | 1,26 | 3,00 | 10,20 | 2,40 | 1,80 |
|  |  | 14,25 | 5,4 | 1,95 | 1,56 | 2,95 | 10,80 | 3,00 | 2,10 |
| 15,5 \& over | 3 | 20,10 | 6,6 | 2,46 | 1,68 | 3,40 | 15,9 | 2,4 | 1,8 |
|  |  | 20,25 | 6,75 | 2,70 | 1,65 | 3,15 | 15,85 | 3,45 | 2,7 |
|  |  | 21,0 | 6,8o | 2,70 | 1,65 | 3,40 | 15,40 | 2,80 | 1,80 |



Fig. 6. Helicolenus dactylopterus
a. Larva at 10 mm standard length showing external features only. b. Dorsal view of head showing position of spines. c. Lateral view of head showing position of cranial bones and spines: I. Dentary. 2. Angular. 3. Articular. 4. Quadrate. 5. Branchiostegal rays. 6. Preopercle and five spines. 7. Subopercle. 8. Opercle. 9. Postcleithrum. 10. Posttemporal and spines. II. Hyomandibular. 12. Pterotic and spine. 13. Parietal and spines. 14. Frontal and supraorbital spine. 15. Premaxilla. 16. Maxilla. 17. Nasal and spine. 18. Lacrymal and suborbital spine. 19. Circumorbitals. 20. Prefrontal. 21. Pterygoid. 22. Cleithrum.

Gillrakers and gill-arches have started ossifying. A ceratohyal, quadrate and traces of the pterygoid are visible.

By $6,6 \mathrm{~mm}$ the lacrymal and endopterygoid are formed and the lacrymal bears a spine. Frontal and parietal are fused. Supratemporal and spine are evident. Six branchiostegal rays are ossified. By $7,35 \mathrm{~mm}$ the lacrymal is larger and bears two spines while the first and second suborbital are well ossified, obscuring the pterygoid to some extent. Thirteen gillrakers are present and the sutures between parietal, pterotic, posttemporal and hyomandibular have become indistinct. In the size range 6,6 to $7,35 \mathrm{~mm}$ the head spines present are: supraocular, parietal, pterotic, supratemporal, preorbital, suborbital, three primary and two secondary preopercular. Between 7,35 and $9,0 \mathrm{~mm}$ ossification of the head proceeds to near juvenile condition. New elements added include nasal spines, two extra primary preopercular spines and traces of a preocular


Fig. 7. Helicolenus dactylopterus
Early larval stages showing position and relative length of head spines. Measurements indicate standard length.
spine. By $10,2 \mathrm{~mm}$ the parietal spine has become bifid as in juveniles. Two opercular spines become evident at the posterior edge of the opercle as in adults. From late larval stage at $10,0 \mathrm{~mm}$ and juvenile stage between 15 and 20 mm the relative size of the head spines decreases and they become far less conspicuous. The secondary spines on the preopercular disappear and the


Fig. 8. Helicolenus dactylopterus
a. Larva. b. Early juvenile showing completed major ossification. Measurements indicate standard length.
second primary spine lengthens considerably to become as long as the third and eventually the longest spine, as in the adult condition. A small spine develops behind the supraorbital, also behind the supratemporal. Rows of villiform teeth develop on the upper and lower jaws in the late larval stage.

In the trunk and tail ossification of the neural spines commences between 5,0 and $5,4 \mathrm{~mm}$ while that of the centra starts between 5,5 and $6,0 \mathrm{~mm}$ and that of the haemal spines soon after. Ossification proceeds anteroposteriorly in sequence.

At $5,4 \mathrm{~mm}$ there are two neural spines and at $6,0 \mathrm{~mm} 5$ to 18 : Between 6,0 and $6,6 \mathrm{~mm}$ ossification proceeds rapidly and 17 to 20 neural spines and 14 to 16 haemal spines are formed, as well as 15 to 20 complete and 2 to 3 incomplete centra (see Table 6). Twenty vertebrae are fully ossified at $7,35 \mathrm{~mm}$ (Fig. 9d) and by $8,5 \mathrm{~mm} 24$ centra, with a full complement of 18 to 19 haemal and 23 neural spines ossified by standard length $9,5 \mathrm{~mm}$. Between ro,o mm and the juvenile stage the neural arches ossify completely and the spines become broader and stouter.

At $4,8 \mathrm{~mm}$ there are four ossified caudal rays, five or six at $6,0 \mathrm{~mm}$. The notocord becomes heterocercal at $6,6 \mathrm{~mm}$. The full complement of ${ }^{5} 5$ primary caudal rays is ossified by $7,35 \mathrm{~mm}$ as well as five to six secondary rays. The

Table 6
Range of skeletal elements in vertebral column of Helicolenus dactylopterus

| Length mm | Haemal spine | Neural spine | Vertebrae | No. of specimens |
| :---: | :---: | :---: | :---: | :---: |
| 5,4 | - | $0-3$ | - | 1 |
| 5,7 | $0-9$ | $0-15$ | $0-5+0-10\left(\frac{1}{2}\right)$ | 3 |
| 6,0 | $0-13$ | $5-18$ | $5-6+5-9\left(\frac{1}{2}\right)$ | 6 |
| 6,6 | $14-16$ | $19-23$ | $15-21+2\left(\frac{1}{2}\right)$ | 3 |
| 7,2 | $16-17$ | $19-23$ | $20-24+2\left(\frac{1}{2}\right)$ | 4 |
| 7,9 | $15-18$ | $19-23$ | $21-24+2\left(\frac{1}{2}\right)$ | 2 |
| 8,1 | $13-19$ | $18-23$ | $23-24$ | 2 |
| 9,0 | 19 | $23-24$ | $24-26$ | 2 |
| 10,5 | 19 | $24-25$ | $26-27$ | 4 |
| 15,0 | 19 | $24-25$ | $26-27$ | 5 |

( $\frac{1}{2}$ ) refers to partly ossified vertebrae.
Table 7
Average number of ossified elements in fins of Helicolenus dactylopterus larvae

| Size in $m m$ | Caudal | Dorsal <br> Spines Rays | Anal <br> Spines Rays | Pectoral <br> lobe | Pelvic |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 4,00 | 4 | - | - | - |  |
| 5,40 | 6 | - | - | 4 | - |
| 6,60 | $2+15+3$ | $\mathbf{8}+11 \frac{1}{2}$ | $\mathbf{1}+6$ | 12 | - |
| 7,60 | $3+15+3$ | $\mathbf{8}+3+12$ | $2+6$ | 17 | $1+3$ |
| 8,50 | $4+15+5$ | $\mathbf{5}+8+12$ | $2+6$ | 17 | $1+4$ |
| 9,50 | $6+15+6$ | $12+13$ | $3+5$ | 18 | $1+5$ |
| 10,50 | $8+15+9$ | $12+13$ | $3+5$ | 19 | $1+5$ |

(Bold face denotes present but unerupted spines)
urostyle and four hypural elements also ossify between 6,6 and $7,35 \mathrm{~mm}$. By $10,2 \mathrm{~mm}$ the caudal fin takes on a juvenile aspect and more secondary rays are present (Table 7).

The pectoral fin starts ossifying between 5,0 and $5,4 \mathrm{~mm}$. Ossification proceeds dorsoventrally and is completed by $10,2 \mathrm{~mm}$ when there are 19 pectoral rays. Differentiation into the characteristic Helicolenus pattern of two unbranched plus 8 to 9 branched plus 8 to 9 unbranched rays only takes place in late juvenile or early adult stage.

Dorsal and anal fins ossify between 6,0 and $6,6 \mathrm{~mm}$. At $6,6 \mathrm{~mm}$ II to 12 dorsal rays are lightly ossified and 6 to io dorsal spines visibly ossified but not erupted. The anal fin has six lightly ossified rays and one unerupted spine. Three posterior dorsal spines have erupted by $7,35 \mathrm{~mm}$ and by $10,0 \mathrm{~mm}$ the unpaired fins have become fully differentiated; all the dorsal spines have erupted between 9,0 and $9,5 \mathrm{~mm}$. The 12 dorsal spines are shorter and stouter than the 12 to 13 dorsal rays while the three anal spines are as long as the five rays except for the second spine which is longer. Between io,o mm and i5 to 17 mm the fin supports develop fully.

The pelvic fin is evident at $7,0 \mathrm{~mm}$ and the spine is first to ossify. One spine and three rays are complete at $7,35 \mathrm{~mm}$. One spine and five rays present the full complement and are present at $10,2 \mathrm{~mm}$. Ossification of skeletal parts seems to proceed at widely differing rates but as little is known about prevailing conditions under which growth took place, no deduction can be made.


Fig. 9
Distribution of Merluccius capensis from 1951 to 1966 .

Table 8
Average measurements of Helicolenus dactylopterus larvae in mm

| Size range <br> of s.l. | No. of <br> specimens | Standard <br> length | Head <br> length | Snout <br> length | Eye <br> diameter | Depth | Trunk <br> length |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | ---: |
| $3,50-4,49$ | 15 | 3,99 | 1,38 | 0,37 | 0,42 | 1,36 | 2,04 |
| $4,5-5,49$ | 27 | 4,82 | 1,81 | 0,57 | 0,60 | 1,68 | 2,51 |
| $5,50-6,49$ | 10 | 5,92 | 2,28 | 0,91 | 0,78 | 2,13 | 3,21 |
| $6,50-7,49$ | 14 | 6,92 | 2,70 | 0,94 | 0,92 | 2,48 | 3,70 |
| $7,50-8,49$ | 9 | 8,10 | 3,23 | 1,10 | 1,06 | 2,86 | 4,66 |
| $8,50-9,49$ | 4 | 8,92 | 3,20 | 1,12 | 1,20 | 3,60 | 5,30 |
| $9,50-10,49$ | 2 | 9,90 | 4,05 | 1,35 | 1,50 | 3,60 | 6,00 |
| $10,50-11,49$ | 4 | 11,10 | 4,27 | 1,42 | 1,18 | 3,82 | 6,75 |
| $11,50-12,45$ | 2 | 12,00 | 4,05 | 1,65 | 1,50 | 4,20 | 7,35 |
| $12,50-14,95$ | 1 | 12,60 | 4,20 | 1,20 | 1,50 | 4,20 | 7,20 |
| $16-17$ | 1 | 17,10 | 6,00 | 1,80 | 2,40 | 6,00 | 10 |
| $18-19$ | 2 | - | 6,90 | 1,95 | 2,40 | 6,30 | 11,25 |
| $20-21$ | 2 | - | 7,20 | 2,10 | 2,85 | 6,60 | 13,05 |

TAble 9
Body proportions of Helicolenus dactylopterus larvae in \% of standard length

| Size range <br> in mm | Snout <br> $\%$ | Eye <br> $\%$ | Head <br> $\%$ | Trunk <br> $\%$ | Depth <br> $\%$ | No. of <br> specimens |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $3,5^{-4,49}$ | 8,6 | 10,7 | 34,9 | 51,7 | 34,3 | 16 |
| $4,5^{-5}, 49$ | 14,2 | 12,8 | 36,4, | 50,5 | 33,9 | 23 |
| $5,5^{-6,49}$ | 13,6 | 13,1 | 38,6 | 54,2 | 36,0 | 11 |
| $6,5^{-7,49}$ | 13,6 | 13,3 | 39,1 | 53,6 | 35,9 | 14 |
| $7,5^{-8,49}$ | 11,9 | 13,1 | 40,0 | 57,8 | 35,0 | 9 |
| $8,5^{-9,49}$ | 12,6 | 13,4 | 36,1 | 59,6 | 40,3 | 4 |
| $9,5^{-10,49}$ | 13,6 | 15,2 | 40,9 | 60,3 | 36,4 | 3 |
| $10,5^{-11,49}$ | 12,8 | 12,8 | 38,5 | 60,8 | 34,5 | 2 |
| $11,5^{-12,49}$ | 13,7 | 12,5 | 33,7 | 61,0 | 40,0 | 4 |
| $12,5^{-13,49}$ | 9,5 | 11,9 | 33,3 | 57,1 | 33,3 | 1 |
| $16,5^{-17,5^{*}}$ | 10,5 | 14,0 | 35,1 | 63,1 | 35,1 | 5 |
| $18,5^{-19,5^{*}}$ | 8,2 | 11,8 | 32,5 | 61,8 | 30,9 |  |
| $20,5^{-21,6}$ | 10,0 | 13,5 | 34,2 | 62,1 | 31,4 |  |

* Size ranges not available in collection.


Fig. io
Distribution of Thyrsites atun from 195I to 1956 and 1960 to 1965.


Fig. II
Distribution of Helicolenus dactylopterus from 1962 to 1965 .

## Changes in body form

During development the body form changes from a rather deep anteriorly large to a more evenly proportioned shape with a depth approximately $30 \%$ of the standard length. The relative head length remains fairly constant during development, varying between 34 and $40 \%$ of the standard length. Head length increases $\pm 0,39 \mathrm{~mm}$ for each mm increase in standard length. The snout is 8 to $14 \%$ of standard length and eye diameter in to $15 \%$ of s.l., both remaining constant throughout development. The rapid increase in snout length during size range 3,4 to $5,49 \mathrm{~mm}$ could however indicate the growth and ossification of jaw elements. The trunk length increases $0,6 \mathrm{~mm}$ per mm increase in s.l., increasing to $0,9 \mathrm{~mm}$ per mm increase in s.l. at 9,0 to $10,0 \mathrm{~mm}$ s.l. Relative trunk length increases gradually during development from 50 to $5 \mathrm{I} \%$ of standard length between sizes of 4 and 5 mm , to 6 I to $62 \%$ of s.l. at 19 to 21 mm .

## Distribution

The area covered by the research vessels of the Division of Sea Fisheries on the pilchard research programme has varied since its inception. During I95I and 1952 the area worked lay between $32^{\circ}$ and $35^{\circ} 30^{\prime} \mathrm{S}$ and was delimited by the 200 fathom depthline to the west. Approximately the same area was worked between 1953 and 1957. In 1958 the eastern limit of the work area was extended round Cape Point to $19^{\circ} 30^{\prime} \mathrm{E}$. This area was worked until the end of 1960 , when the eastward delimitation was extended to $21^{\circ} \mathrm{E}$. During these years the westward delimitation extended to $16^{\circ} 3^{\prime} I^{\prime}$ E. From July 1963 to December 1965 the area covered by the research vessels was between $32^{\circ} 10^{\prime}$ to $36^{\circ} 10^{\prime} \mathrm{S}$ and $16^{\circ}$ to $21^{\circ} 30^{\prime} \mathrm{E}$. Stations lists are obtainable from the Annual Reports of the Division of Sea Fisheries for the relevant years.

## Summary

The larval stages of Merluccius capensis, Thyrsites atun and Helicolenus dactylopterus are described. All three species are economically important in South Africa. The taxonomy of each species is revised according to latest opinions. A brief description of distribution is included.

## Acknowledgements

The author wishes to thank the Division of Sea Fisheries's sea-going staff for the collection of the study material, Fisheries Development Corporation for financial assistance and Dr. N. A. H. Millard and Mrs S. Bruins for reading the manuscript and other assistance.

## References

Ahlstrom, E. H. 1961. Distribution and relative abundance of rockfish (Sebastodes spp.) larvae of California and Baja California. Rap. P.-V. Réun. Cons. perm. int. Explor. Mer 150: 169-176.
Ahlstrom, E. H. 1963. Kinds and abundance of fishes in the Californian Current region based on egg and larval surveys. Rep. Calif. coop. oceanic Fish. Invest. 10: 31-52.
Ahlstrom, E. H. \& Counts, R. C. 1955. Eggs and larvae of the Pacific hake Merluccius productus. Fishery Bull. Fish Wildl. Serv. U.S. 56: 295-31 1.
Barnard, K. H. 1925. A monograph of the marine fishes of South Africa. Ann. S. Afr. Mus. 21: I-4 18 .
Beaufort, L. F. De \& Chapman, W. M. 1951. The fishes of the Indo-Australian Archipelago. 9. Leiden: Brill.
Bigelow, H. B. \& Welch, W. W. 1924. Fishes of the Gulf of Maine. Bull. Bur. Fish., Wash. 40 (1): $1-567$.
D'Ancona, U. 1933. Uova larve e studi giovanili di Teleostei. Fauna Flora Golfo Napoli 38: 177-384.
Davies, D. H. 1949. Preliminary investigations on the foods of South African fishes. Investl Rep. Fish. mar. biol. Surv. Div. Un.S.Afr. 11: 1-36.
Davies, D. D. \& Gore, U. R. 1936. Cleaning and staining of skeletons of small vertebrates. Publs. Field Mus. nat. Hist. (Tech.) 4: 1-15.
De Jager, B. v. D. 1955. Development of the snoek (Thyrsites atun), a fish predator of the pilchard. Investl Rep. Div. Fish. Un.S.Afr. 19: 1-16.
Ehrenbaum, E. 1909. Eier und Larven von Fischen. 2. Teil. Nord. Plankt. 10: 217-413.
Eschmeyer, W. N. 1969. A systematic review of the scorpion fishes of the Atlantic Ocean (Pisces: Scorpaenidae). Occ. Pap. Calif. Acad. Sci. 79: 1-143.
Euphrasen, B. A. 1791. Scomber atun och Echeneis tropica beskrifna. K. svenska VetenskAkad. Nya Handl. 12: 315.

Fischer, W. 1959. Huevos, crias y prelarvas de la merluza (Merluccius gayi), Guichenot. Revta Biol. mar. 9: 229-249.
Fowler, H. W. 1936. The marine fishes of West Africa. Bull. Am. Mus. nat. Hist. 10: 630-631.
Gilchrist, J. D. F. 1921. Deep sea fishes procured by the S.S. 'Pickle' (Part 1). Rep. Fish. mar. biol. Surv. Un.S.Afr. 2 (Spec. Rep. 3): 41-79.
Ginsburg, I. 1954. Whitings on the coasts of the American continent. Fishery Bull. Fish Wildl. Serv. U.S. 56: 187-208.
Hart, T. J. \& Marshall, N. B. 195 I. Breeding ground of pilchards off the coast of South-West Africa. Nature, Lond. 168: 272-273.
Hollister, G. 1934. Cleaning and dyeing fish for bone study. Zoologica, N.T. 12: 89-10i.
Irvin \& Johnson Limited. 1963. South .African fish and fishing. Cape Town: Irvin \& Johnson.
Jones, S. 1960. On the snake mackerel Gempylus serpens Cuvier from the Laccadive Sea. J. mar. biol. Ass. India 2: 85-88.
Kunz, A. \& Radcliffe, L. 1917. Notes on embryology and larval development of twelve teleostean fishes. Bull. Bur. Fish., Wash. 35: 87-1 34 .
Marak, R. R. 1967 . Eggs and early larval stages of the off-shore hake, Merluccius albidus. Trans. Am. Fish. Soc. 96: 227-228.
Matthews, J. P. \& De Jager, B. v. D. 1951. The development of the Cape stock-fish Merluccius capensis. Investl Rep. Div. Fish. Un.S.Afr. 13: 1-10.
Moran, J. F. 1956. Differential staining of bone and cartilage in toto of fish. Proc. Indiana Acad. Sci. 65: 234-236.
Moser, H. G. 1967. Reproduction and development of Sebastodes paucispinis and comparison with other rockfishes off Southern California. Copeia 1967: 773-797.
Norman, J. R. 1935. Coast fishes. Part I. The South Atlantic (including the Cape Verde Islands, West Africa, South Africa, Ascension Island, Tristan da Cunha and Gough Island). 'Discovery' Rep. 12: 3-58.
Norman, J. R. 1937. Coast fishes. Part II. The Patagonian region (including the Straits of Magellan and the Falkland Islands). 'Discovery' Rep. 16: 3-150.
Norman, J. R. 1966. A draft synopsis of the orders, families and genera of recent fishes and fishlike vertebrates. London: British Museum (Natural History).
Regan, C. T. 1908. Descriptions of the new or little known fishes from the coast of Natal Ann. Natal Mus. 1: :-6.
Regan, C. T. 1916. Larval and post-larval fishes. Nat. Hist. Rep. Br. Antarct. Terra Nova Exped. (Zool.) 1: 125-156.
Santander, H. \& De Castillo, O. S. ig6g. Desarrollo y distribucion de huevos y larvas de merluza, Merluccius gayi (Guichenot) en la costa peruana. Boln Inst. Mar Peru 2: 79-107.
Smith, J. L. B. 1953. The sea fishes of southern Africa. 4th ed. Cape Town: Central News Agency.
Voss, N. A. 1954. The postlarval development of the fishes of the family Gempylidae from the Florida current. I. Neiiarchus Johnson and Gempylus Cuv. \& Val. Bull. mar. Sci. Gulf Caribb. 4: 120-157.

## INSTRUCTIONS TO AUTHORS

Based on<br>CONFERENGE OF BIOLOGICAL EDITORS, COMMITTEE ON FORM AND STYLE. 1960 .<br>Style manual for biological journals. Washington: American Institute of Biological Sciences.

## MANUSCRIPT

To be typewritten, double spaced, with good margins, arranged in the following order: (1) Heading, consisting of informative but brief title, name(s) of author(s), address(es) of author(s), number of illustrations (plates, figures, enumerated maps and tables) in the article. (2) Contents. (3) The main text, divided into principal divisions with major headings; subheadings to be used sparingly and enumeration of headings to be avoided. (4) Summary. (5) Acknowledgements. (6) References, as below. (7) Key to lettering of figures. (8) Explanation to plates.

## ILLUSTRATIONS

To be reducible to $12 \mathrm{~cm} \times 18 \mathrm{~cm}$ ( 19 cm including caption). A metric scale to appear with all photographs.

## REFERENCES

Harvard system (name and year) to be used: author's name and year of publication given in text; full references at the end of the article, arranged alphabetically by names, chronologically within each name, with suffixes $a, b$, etc. to the year for more than one paper by the same author in that year.
For books give title in italics, edition, volume number, place of publication, publisher.
For journal articles give title of article, title of journal in italics (abbreviated according to the World list of scientific periodicals. 4 th ed. London: Butterworths, 1963), series in parentheses, volume number, part number (only if independently paged) in parentheses, pagination.

## Examples (note capitalization and punctuation)

bullough, W. S. 1960. Practical invertebrate anatomy. and ed. London: Macmillan.
Fischer, P.-H. 1948. Données sur la résistance et de le vitalité des mollusques. F. Conch., Paris 88: $100-140$.
Fischer, P.-H., Duval, M. \& Raffy, A. i933. Etudes sur les échanges respiratoires des littorines. Archs Zool. exp. gén. 74: 627-634.
Kohn, A. J. i96oa. Ecological notes on Conus (Mollusca: Gastropoda) in the Trincomalee region of Ceylon. Ann. Mag. nat. Hist. (13) 2: 309-320.
Kohn, A. J. ig6ob. Spawning behaviour, egg masses and larval development in Conus from the Indian Ocean. Bull. Bingham oceanogr. Coll. 17 (4): 1-51.
Thiele, J. igio. Mollusca: B. Polyplacophora, Gastropoda marina, Bivalvia. In schultze. l, Zoologische und anthropologische Ergebnisse einer Forschungsreise im westlichen und zentralen SüdAfrika. 4: 269-270. Jena: Fischer. Denkschr. med.-naturw. Ges. Jena 16: 269-270.

## ZOOLOGICAL NOMENCLATURE

To be governed by the rulings of the latest International code of zoological nomenclature issued by the International Trust for Zoological Nomenclature (particularly articles 22 and 51). The Harvard system of reference to be used in the synonymy lists, with the full references incorporated in the list at the end of the article, and not given in contracted form in the synonymy list.

## Example

Scalaria coronata Lamarck, 1816: pl. 451, figs $5 a, b$; Liste: 11. Turton, 1932: 80.

ANNALS OF THE SOUTH AFRICAN MUSEUM ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM

| Volume | $\mathbf{5 9}$ | Band |
| ---: | :---: | :--- |
| March | $\mathbf{1 9 7 2}$ | Maart |
| Part | $\mathbf{4}$ | Deel |



# A PLIOCENE PHOGID FROM SOUTH AFRICA 

By
Q. B. HENDEY \& C. A. REPENNING

The ANNALS OF THE SOUTH AFRICAN MUSEUM are issued in parts at irregular intervals as material becomes available

Obtainable from the South African Museum, P.O. Box 61, Cape Town

Die ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM
word uitgegee in dele op ongereelde tye na beskikbaarheid van stof

Verkrygbaar van die Suid-Afrikaanse Museum, Posbus 61, Kaapstad

> OUT OF PRINT/UIT DRUK

I, $2(\mathrm{I}, 3,5,7-8), 3(\mathrm{I}-2,5$, t.-p.i. $), 5 \mathrm{I}-2,5,7-9)$,
$6(\mathrm{I}, \quad$ t.-p.i. $), \quad 7(\mathrm{I}-3), \quad 8, \quad 9(\mathrm{I}-2), \quad 10(\mathrm{I})$, II (I-2, 5, 7, t.-p.i.), 2I, 24(2), 27, 3 I (I-3), 33, 38

Price of this part/Prys van hierdie deel
$\mathrm{R}_{4,00}$.

Trustees of the South African Museum
(C) Trustees van die Suid-Afrikaanse Museum 1972

ISBN 0949940046

## A PLIOCENE PHOCID FROM SOUTH AFRICA

By<br>Q. B. Hendey<br>South African Museum, Cape Town

\&

Charles A. Repenning<br>U.S. Geological Survey, Menlo Park, California

(With plates 2-18, 2 figures and 7 tables)
[MS. accepted 9 September 1971]

Contents


## Introduction

Although Illiger recognized the basic features of the seals of the world in 18in by separating them from the sirenians and placing them in a separate order, the Pinnipedia, their classification is still in a state of flux. In 188o Allen divided the pinnipeds into two major groups: the 'walkers' and the 'wrigglers' which Smirnov (1908) subsequently named the superfamilies Otarioidea and the Phocoidea. The Phocoidea contains only one family, the Phocidae (Gray 1825, but defined with its present contents by Brookes, 1828), usually known as the 'earless seals' or 'true seals'. Subsequent to Kellogg's (1922) introduction of the subfamily Lobodoninae (respelled Lobodontinae by Hay, 1930) to include the Antarctic phocids, the family Phocidae was considered to include four subfamilies: Phocinae for the northern seals, Monachinae for the genus Monachus, Cystophorinae for the genera Cystophora and Mirounga, and Lobodontinae. Scheffer (1958) reduced the rank of the Antarctic phocids to that of a tribe, Lobodontini, within the family Monachinae. King (ig66), in possibly one of the most detailed explanations of a change in pinniped classification, abandoned the subfamily Cystophorinae, placing the genus Cystophora in the Phocinae and the genus Mirounga in the Monachinae. Most recently (at this writing) McKenna
(1969), in possibly one of the briefest explanations of a change in pinniped classification, stated that the formal taxon order Pinnipedia has been abandoned.

As here used, the order Pinnipedia contains three families: Odobenidae, the walruses; Otariidae, the sealions; and Phocidae, the true seals who must 'wriggle' on their bellies in terrestrial locomotion, their hind limbs permanently extended behind them. The family Phocidae contains two distinct subfamilies, the Phocinae, inhabiting temperate and arctic waters of the Northern Hemisphere, and the Monachinae, inhabiting parts of most oceans except the Arctic Ocean.

For the most part, the characters identified by King (1966) may be used to separate the living members of the two phocid subfamilies. Since it is assumed that the two subfamilies derive from a common ancestor, it is to be expected that fewer of these characters will be useful for familial identification in the fossil ancestors of the living true seals. Such is the case with the Pliocene phocid from South Africa.

Since 1958, when the first discoveries from the quarries of the African Metals Corporation were recorded by Singer \& Hooijer (1958), tens of thousands of vertebrate specimens have been recovered from these Pliocene phosphate deposits near Langebaanweg, Cape Province. At least 60 mammalian species, as well as a full complement of birds and cold-blooded vertebrates, have been recognized (Hendey, 1970a, 1970b). The fauna is both marine and terrestrial, apparently representing accumulation during a prolonged period of marine, estuarine, and terrestrial deposition in the area.

The dating of the Langebaanweg fauna is a major problem presently being investigated (Hendey, 1970b; Maglio \& Hendey, 1970). On the basis of an admittedly limited number of comparisons with faunal elements from betterdated localities in East Africa, we believe the Langebaanweg deposits bearing the main body of higher vertebrate remains to be perhaps 4 to 5 million years old. The most recent interpretation of the most reasonable definition of Pleistocene, and the approximation of its beginning between 2,6 to 3 million years ago (Savage \& Curtis, 1970), suggest that the Langebaanweg fauna should probably be called late Pliocene.

The Langebaanweg fauna includes a pinniped which was first reported by Boné \& Singer (1965). These authors tentatively referred it to the otariid genus Arctocephalus, but more recently discovered specimens show clearly that it is a monachine phocid which belongs to a previously unrecorded species of the extinct genus Prionodelphis, heretofore known only from Pliocene deposits in Argentina. In many respects this seal from Langebaanweg is similar to the extant Monachus and its ancestors, notably Pliophoca etrusca from the late Pliocene of Italy (Tavani, 1942). However, similarities to the extant phocids of the Antarctic seas are equally well marked.

In view of the small number of recorded specimens of Prionodelphis rovereti (Frenguelli, 1922, 1926), from Argentina, the Langebaanweg seal, represented
by a wide variety of specimens, is clearly important in that it provides the first good evidence of the antiquity and ancestry of the monachine seals in the Southern Hemisphere.

## Systematics

As here used, the subfamily Monachinae includes the same genera of living seals that were included in this subfamily by King (ig66). Except for the addition of the genus Mirounga, this agrees with the definition of the subfamily as originally defined by Trouessart (1897:373).

As noted by Scheffer (1958:1ri), when he reduced the Lobodontinae to tribal rank within the Monachinae, the major difference between these Antarctic seals and Monachus is one of geography. King (ig66) omitted reference to Scheffer's tribe Lobodontini when discussing the monachine relationship of Mirounga, with good reason as the genus does not conform to the geographic distinction mentioned by Scheffer. Recognition of any tribal subdivision of the Monachinae now seems pointless.

King (1966:397) noted that with some features otherwise typical of the monachine seals Monachus was an exception and regarded this genus as being not quite so advanced. Such exceptions to otherwise typical features of the Monachinae are even more evident in the Pliocene seal from Langebaanweg. At this stage in the evolution of the phocid seals the subfamily Monachinae can be distinguished from the subfamily Phocinae by the following characters of the skull.

## Subfamily Monachinae

Diagnosis. Seals having a mastoid bone without a prominent posterolaterally projecting rounded crest but, instead, having a posterolateral surface curving uniformly from the region of the parietal suture down to the region of the stylomastoid foramen; mandible with an extensive symphyseal surface that is elongate and smoothly oval in outline and that firmly articulates over the entire depth of the chin.

As will be shown in the following report, some postcranial bones of the Langebaanweg seal exhibit monachine features while other are simply phocid, with no subfamilial characteristics.

## Genus Prionodelphis

Type. Prionodelphis rovereti Frenguelli, 1922.
Known distribution. Pliocene of the South Atlantic Ocean.
Comment. The type species, Prionodelphis rovereti, was described as a squalodont cetacean from a few isolated teeth found in Pliocene deposits in Entre Rios in Argentina. A mandibular fragment bearing one tooth was later found at the same locality which led Cabrera (1926:390) to the conclusion that the animal was a pinniped, a conclusion supported by others (Frenguelli, 1926;

Kraglievich, 1934; Kellogg, 1942). We are aware of no additional material.
The material from South Africa now makes it possible to provide a better definition of the genus.

Diagnosis. A generalized monachine seal lacking the shortened rostrum and crowded teeth of Monachus monachus and the squared premaxillaries with aligned, upper incisors of M. tropicalis and M. schauinslandi; postcanine teeth low-cusped as in M. schauinslandi, M. tropicalis and Pliophoca etrusca and distinctly narrower in occlusal outline than those of M. monachus; upper fifth postcanine ${ }^{1}$ with recurved crown; ascending ramus of premaxilla strong, terminating against nasals and prominently visible in lateral view separating maxilla from nasal aperture; pre-orbital processes prominent; forehead broad in supra-orbital region; osseous nasal septum strongly developed; dental formula 2.1.5/2.1.5; tympanic bulla covers petrosum.

## The Langebaanweg Seal <br> Prionodelphis capensis n.sp.

Holotype. An incomplete skull with left canine and fourth postcanine, and right third postcanine (South African Museum No. L 15695).

Referred material. An incomplete skull (L 12695); temporal bone (L 15652 $^{2}$ ); mandible fragments (L 7556, L 12299); one lower and two upper incisors; three lower and four upper canines; and nine lower and ten upper postcanines.

Various elements of the postcranial skeleton have been recovered, of which the following have been selected for description: vertebrae ( $\mathrm{L}_{7563}, \mathrm{~L}_{15689}$, $L_{15849} A_{1} \& A_{2}, L_{15396} L_{15857}$ ); scapula (L 2160); humeri (L ${ }_{2157}$, L 4638); ulnae (L 2161, L 15682 ); radii (L 2935, L 12869 ), innominate (L i5849A), femur (L io131); tibiae (L 2138, Lioi28/9); calcaneum (Lioir8); astragali (L ioi30, L io993); navicular (L i585i); entocuneiform (L ior34); metapodial (L rog96); first phalanx (L iog99); second phalanx (L IO205).

All specimens are housed in the South African Museum, Cape Town. Except for the two incomplete skulls, which are too fragile to cast, casts of the more significant specimens are housed in the U.S. Geological Survey, Pacific Coast Center, Menlo Park, California.

Locality and horizon. The holotype and most of the referred material is from horizon 2, 'E' Quarry, Langebaanweg. Some postcranial elements are known from horizon I. These horizons are thought to be broadly contemporaneous (Hendey, 1970b). A few fragmentary remains from ' C ' and Baard's Quarries are excluded from this report, but they apparently represent the same species.

Comparative material. Skulls of all living phocid species except Pusa caspica have been available for comparison either in the South African Museum or in the Pacific Coast Center of the U.S. Geological Survey. Postcranial material has been somewhat less available, but comparisons were made with postcranial ${ }^{1}$ Called $\mathrm{P}_{1}$ by Frenguelli, 1922: 496.
elements of Monachus schauinslandi, Hydrurga leptonyx, Lobodon carcinophagus, Mirounga angustirostris, M. leonina, and all living phocine genera.

Diagnosis. Prionodelphis capensis differs from the type species in that it has only one anterior accessory cusp on the lower postcanines instead of two, and there is a greater reduction in size of the posterior root in the second to fourth upper postcanines. The cheek-teeth of the South African species approach more closely a three-cusped tooth pattern, and in addition, are slightly larger and more laterally compressed (Table 1).

Table I. Average length-width ratios for postcanines 2 to 4 of some phocids.

|  | Length | Width | W/L $\times 100$ | Upper <br> Lower |
| :---: | :---: | :---: | :---: | :---: |
| Hydrurga leptonyx |  |  |  |  |
| Upper ( $\mathrm{N}=6$ ) | 18, 0 mm | $8,8 \mathrm{~mm}$ | 49 | o,925 |
| Lower ( $\mathrm{N}=6$ ) | 17,6 | 9,3 | 53 |  |
| Proonodelphis capensis |  |  |  |  |
| Upper ( $\mathrm{N}=7$ ) | 13,8 | 7,5 | 54 | 1,200 |
| Lower ( $\mathrm{N}=8$ ) | 15,0 | 6,8 | 45 |  |
| Prionodelphis rovereti ${ }^{1}$ |  |  |  |  |
| Upper ( $\mathrm{N}=1$ ) | 12,25 | 8,0 | 65 | 1,204 |
| Lower ( $\mathrm{N}=2$ ) | 13,0 | 7,0 | 54 |  |
| Monachus schauinslandi |  |  |  |  |
| Upper ( $\mathrm{N}=6$ ) | 12,0 | 7,9 | 66 | 1,047 |
| Lower ( $\mathrm{N}=6$ ) | 12,1 | 7,6 | 63 |  |
| Monachus monachus |  |  |  |  |
| Upper ( $\mathrm{N}=6$ ) | 12,9 | ${ }_{8,}^{9,1}$ | 70 | 1,1II |
| Lower ( $\mathrm{N}=6$ ) | 12,9 | 8,0 | 63 |  |

## Description

The assessment of the fossil remains listed above is somewhat hampered by their fragmentary nature. The holotype is composed of about 60 individual pieces, including three teeth, found scattered over a wide area in the excavation No. LBW 1969/ı (South African Museum departmental records). Numerous small pieces could not be restored to the skull, although they undoubtedly belong, and others presumably remain in unexcavated parts of the deposit.

The second partial skull (L i2695) was similarly fragmented. Although the partially restored snout region is less complete than that of $\mathrm{L}_{15}{ }_{595}$, parts of the braincase of the second specimen were also recovered. Many of the individual pieces are extensively abraded, probably having suffered in a manner similar to that described for an alcelaphine skull recovered near by (Hendey, 1970a: 82).

As with much of the referred material, the temporal bone ( $\mathrm{Li}_{5} 65_{2}$ ) also came from the excavation LBW 1969/r, but from approximately 75 cm below the holotype. It probably belongs to another individual.

Most of the isolated teeth were recovered intact, and they vary from unworn to extremely worn.

Few elements of the postcranial skeleton were recovered intact, but in some cases sufficient numbers of a particular bone are known to enable a complete assessment of its characteristics.

## THE SKULL

The skull of the Langebaanweg pinniped is in many ways unique, and it exhibits a set of characteristics which makes it impossible to assign it to any previously known phocid species (Plates 2-8).

As a whole, the skull appears convincingly to be that of a monachine seal. The lack of swollen or crested mastoids, the broad and flat dorsal surface of the petrosal apex, and the deep and oval mandibular symphysis rule out any known phocine seal, while the incisor formula and cheek-tooth pattern strongly suggest a monachine seal. Although the postcranial bones in general also appear monachine, they possess some features that are characteristic of living phocine seals, such as an entepicondylar foramen on the humerus.

In general features, the skull most resembles those of Monachus and Hydrurga, although it is considerably more gracile and less elongated than the latter. Of all extant genera its teeth most resemble those of Monachus. However, except for the lack of great vertical exaggeration of the cusps, the teeth are also quite similar to those of Hydrurga. Of the extinct genera they are, as far as comparisons are possible, most similar to Prionodelphis rovereti and, less so, to Pliophoca etrusca. The basic pattern of the cheek-teeth appears to be that of earlier members of the family Phocidae.

Unlike the usual pattern in the Antarctic monachines, the premaxilla terminates against the nasal bone. Although it shares this characteristic with Monachus and several phocine seals, it differs markedly, as does Pliophoca etrusca, in the massiveness of the ascending ramus of the premaxilla. When the

Table 2. Dimensions of the skull of Prionodelphis capensis from Langebaanweg.

|  | Li5695 | Li 2695 | L7556 | Li 2299 |
| :---: | :---: | :---: | :---: | :---: |
| Rostral width | 48,0* | 61,0* | - | - |
| Distance between pre-orbital processes of maxilla | 94,0* | - | - | - |
| Minimum diameter of inter-orbital region | 36,o* | - | - | - |
| Distance between pre-maxillary projections and posterior limit of nasal aperture | 58,0* | 51,0* | - | - |
| Lengths of nasals . . . . | 62,0* | - | - | - |
| Distance between external alveolar margins of fifth postcanines. | 74,5* | 87,0* | - | - |
| Alveolar length of upper postcanine series | 72,5 | 72,5* | - | - |
| Alveolar length of lower postcanine series |  | 7,5 | 66,5 | - |
| Depth of mandible behind $\mathrm{M}_{1}$. . <br> * Estimated. | - | - | 27,9 | 31,6 |



FIg. i. Diagram of cranial measurements: 1. Rostral width. 2. Distance between pre-orbital processes of maxilla. 3. Minimum diameter of inter-orbital region. 4. Distance between premaxilary pranine series.
skull is viewed laterally the premaxilla is visible along its entire length rather than being partly hidden behind the maxilla, largely within the nasal opening, as in living monachines. The nasal opening is elongated antero-posteriorly, and of the southern phocids most resembles Hydrurga and Lobodon, rather than Leptonychotes and Ommatophoca in which there is foreshortening of the anterior part of the snout. In this respect it is also similar to Monachus and Pliophoca. The two halves of the premaxilla have fairly prominent projections at their most anterior point of contact, dorsal to the incisors. Of the extant monachines M. tropicalis, M. schauinslandi and Hydrurga have similar projections, but in Hydrurga inflation of the alveolar region of the incisors renders the projections less prominent, and in the two Monachus species the projections are quite widely separated. In the fossil species there is a marked step between the most anterior maxilla-premaxilla contact and the premaxillary projection. This is best seen in the specimen L 12695 .

Judging from the size of the alveoli, the lateral incisor is only slightly larger than the medial one. The latter is situated slightly anterior to the other. The relative size and position of the upper incisors are as in M. monachus and Hydrurga, and unlike the specialized condition found in Lobodon and Leptonychotes, in which genera the lateral incisor is much larger than the medial one and is situated in line with it.

Two upper incisors are known, both left medials judging from their size. The structure of the crown resembles that of the incisors of M. monachus and Pliophoca etrusca. The cross-section of the canines is only slightly elongated antero-posteriorly, and in this respect $P$. capensis differs markedly from the Antarctic phocids. The canines L ir686 (Plate 9B) and L 12695 have large bulbous roots, a feature also seen in some of the postcanines. This is a characteristic of old age in most or all pinnipeds.

There are five upper postcanines and at least one specimen of each dental category is known from the assemblage. The first, and smallest, is single rooted (Plate 9 C ). The remainder are all double rooted, with the posterior root being larger than the anterior one in the second, third, and fourth teeth, and the sizes reversed in the fifth. The latter is also the second smallest of the teeth and is set slightly separate from the others immediately below the infra-orbital foramen. The postcanine tooth rows curve outwards posteriorly as in Ommatophoca, and are not diverging straight lines as in Leptonychotes and Hydrurga; this pattern is rather close to that of $M$. schauinslandi as well as some other phocid species.

The postcanines resemble those of Pliophoca etrusca, although narrower and more gracile, and except for being much less massive, they are also similar to those of $M$. monachus (Plate 9D). They are quite distinct from the highly specialized teeth of extant Antarctic phocids; of this group the teeth of Leptonychotes are closest to the fossils, but they are nevertheless significantly different. The basic pattern of the first to fourth postcanines is similar to the corresponding teeth of $M$. monachus. There is a prominent central cusp with two smaller cusps situated anteriorly and posteriorly, with an additional small projection on the
most posterior part of the cingulum. The enamel is generally rugose, a condition which is found in all monachine seals except Mirounga, but which is also found in some phocine seals. The second, third and fourth upper postcanines have a marked inflation of the postero-internal cingular region, a condition also evident in some teeth of $P$. rovereti, and $P$. etrusca, and these teeth are broader posteriorly than they are anteriorly. The first upper postcanine has the internal cingulum inflated, and the maximum transverse diameter is at about the midpoint of the tooth. The upper postcanine of $P$. rovereti described by Frenguelli ( $1922: 493$ and figs. $\mathrm{Ib}, \mathrm{Ic}$ ) is shorter and relatively broader than those of $P$. capensis. It lacks the distinct anterior accessory cusp present in the latter species and has a larger posterior root.

The fifth postcanine of $P$. capensis lacks the posterior accessory cusp and cingular projection, and the anterior accessory cusp is much reduced, being barely discernible; the principal cusp is strongly recurved (Plate 9 E ). It resembles the cheek teeth of Leptonychotes in this respect. Both the fifth postcanines recovered to date are completely unworn, and it seems probable that this tooth did not occlude with the lower fifth postcanine. Leptonychotes also has a nonoccluding upper fifth postcanine. This tooth, in $P$. capensis, is very similar to one of the original $P$. rovereti specimens illustrated (as a $\mathrm{P}_{1}$ ) by Frenguelli (1922: fig. 2A). The principal differences are in the smaller size of the $P$. rovereti specimen, in that its anterior accessory cusp is situated higher up the crown, and also in that there is no cingulum on the buccal surface of the Langebaanweg specimens. The two $P$. capensis specimens differ from one another only in that one is slightly shorter and somewhat broader than the other. In the postcanines such differences distinguish upper from lower teeth, but judging from the size of alveoli in mandibles and maxillae known, these teeth can only be upper fifth postcanines.

Of the seven second, third and fourth upper postcanines known, two are unworn and five show wear angled from the principal cusp to the posterior cingulum.

The general features of the maxilla of $P$. capensis correspond most closely to those of Hydrurga. The fossil seal has very prominent preorbital processes. The one preserved in the holotype projects outwards and downwards as in Hydrurga, although in L 12695 it has an outwards and upwards inflection. The presence and form of the preorbital process is variable throughout the Pinnipedia, but within the monachine seals it is virtually absent in M. schauinslandi, M. tropicalis and Leptonychotes and present but variably developed in other extant monachine species.

In the Antarctic phocids the jugal terminates lateral to the infra-orbital foramen, whereas in P. capensis, P. etrusca, Monachus and several other seals it terminates above this foramen.

The infra-orbital foramen is oval shaped as in Hydrurga and Lobodon, but the orientation of the longitudinal axis of the foramen differs in that it is directed upwards and outwards in P. capensis, whereas the axis is upwards and inwards in Hydrurga and Lobodon.

The shape of the nasals is not perfectly known, but from a reconstruction of this region in the holotype (Plate 2), it appears that they do not correspond in shape to those of other monachines. Instead they are broader in the frontal region than between the maxillae.

The osseous nasal septum is a very stout bone which terminates at or near the most anterior limit of the nasals. Its proportions resemble those of $M$. schauinslandi, M. tropicalis, Hydrurga, Ommatophoca and Mirounga, but not M. monachus, Lobodon and Leptonychotes.

The supra- and post-orbital regions of the frontal bones are essentially similar to those of Hydrurga, Lobodon and Ommatophoca, and are not parallel-sided as in Monachus, Leptonychotes and Mirounga, nor as in P. etrusca. In the holotype there are the beginnings of a sagittal crest towards the posterior part of the frontals. There are two step-like projections on the frontals above and behind the preorbital processes of the maxilla. Similar features are present in Ommatophoca and are less distinctly represented in Hydrurga.

The second partial skull (L 12695) belongs to an aged individual and is somewhat more robust than that of the holotype (Plate 5). The difference in size of the two specimens may in part be due to the ages of the individuals concerned, but may also reflect sexual dimorphism in the species. Apart from size, the most striking difference between the two specimens is in the form of the nasal aperture. That of $\mathrm{L}_{12} 695$ is actually slightly shorter than that of $\mathrm{L}_{15695}$, although it is, as would be expected, wider and higher. A similar and probably related allometric feature is the correspondence in the length of the postcanine tooth rows.

The braincase of $P$. capensis is not known, although parts of the nuchal region of L 12695 were recovered. The nuchal crest is fairly well developed, being more similar to that of Hydrurga than other species, although, like the sagittal crest, it is considerably less prominent than that of Hydrurga. Unlike Hydrurga there is no marked concavity of the supra-occipital. As in the Antarctic seals, Monachus monachus, and Pliophoca etrusca, the nuchal crest extends anteriorly across the temporal, terminating near the external acoustic meatus, rather than uniting with an enlarged jugular process of the exoccipital as in M. schauinslandi and M. tropicalis.

The basi-cranium is largely unknown, but a number of specimens of the well-ossified temporal bone have been recovered. The most complete (L i5652) has the mastoid and most of the tympanic intact, and the post-glenoid process is still attached (Plate 6). That part of the ectotympanic which projects under the acoustic meatus has been broken off and lost.

Although the general appearance of the tympanic region is most reminiscent of Monachus because of the slight inflation of the bulla, some of its features strongly indicate a relationship between Prionodelphis capensis and the Antarctic phocids. These include the posterior extent of the bulla and the rounded apex of the petrosum.

As noted by King (1966:387), the posterior wall of the bulla in phocine
seals (except some individuals of Erignathus), and in species of Monachus, is located rather far forward so that the posterior part of the petrosum is exposed in ventral aspect (without recourse to peering through the posterior lacerate foramen), whereas in the Antarctic monachine seals, including Mirounga, the bulla covers the petrosum and essentially separates the mastoid from the posterior lacerate foramen by almost contacting the exoccipital. The latter condition is very evident in $P$. capensis, strongly suggesting an affinity with the Antarctic seals.

The dorsal (cerebellar) surface of the temporal resembles that in some living Antarctic seals (Plate 7). The apex of the petrosum is broad and rounded with low relief, as in Lobodon and Leptonychotes, and differs greatly from the pointed apex found in Monachus. It is not the globular structure typical of the phocine seals. However, the petrosal apex of $P$. capensis is smaller than in the living Antarctic seals, suggesting that this seal was less well adapted for directional underwater hearing, according to the interpretation of Repenning (in press). The cerebellar fossa is relatively large, as in Leptonychotes, and as in the latter, the squamosal extends medially to the edge of the cerebellar fossa and to the internal facial canal.

The ventral (external) side of the temporal most resembles that in Hydrurga. The external opening of the carotid canal is located well forward of the posterior limit of the bulla, as in Hydrurga, Leptonychotes and Lobodon, but in contrast to the more posterior location in Ommatophoca, Mirounga and Monachus. In P. capensis this foramen faces noticeably ventrally, as in Hydrurga, but the general outline of the bulla is more similar to that in Lobodon. The stylomastoid foramen is rather widely separated from the external cochlear foramen (Burns \& Fay, 1970:374). A similarly wide separation is found in Hydrurga.

Dissection of the middle ear was not undertaken.
The mandible is rather unspecialized and resembles that of Hydrurga and Monachus (Plate 8). It differs from that of Hydrurga in its smaller size and in having the symphyseal region relatively narrower transversely. In Hydrurga the two lower incisors are situated side by side, but in the fossil the medial incisor lies in an almost horizontal position above and behind the lateral incisor, in a manner comparable to that in most other species of phocid seals.

A single isolated lower medial incisor ( $\mathrm{L}_{15444 \mathrm{~A}}$ ) is known. It is similar in size to that of Leptonychotes but has a pronounced step on the lingual surface of the crown which, in lateral view, resembles that of Monachus.

The lower canines are similar to the uppers, but have a straighter root and are more rounded in cross section.

The five lower postcanine teeth are situated close to one another: the alveolar walls between the teeth are as narrow as, or narrower than, those between the two roots of one tooth. In the mandibular ramus L 7556, the teeth are positioned in much the same way as in the $P$. etrusca specimen described by Tavani (1943: fig. 6a). The first postcanine is single rooted, and the remainder, which are more or less equal in size, have two roots. These teeth apparently all
have a crown pattern similar to the first to fourth upper postcanines (Plate 9 G ). Nine isolated lower postcanines are known, one of which is identified as a first lower and two are thought to be second lowers. Both the latter are worn on their posterior surfaces. Of the remainder, three show most wear on their anterior surfaces, and four, including the first lower, show no perceptible wear at all. Judging from the wear on the upper and lower postcanines, it appears that they functioned as crushing agents, although two of the lower teeth show signs of a transverse shearing action.

The lower teeth are differentiated from the uppers by the fact that they are more slender, with little or no inflation of their internal cingula. The lower postcanines of $P$. rovereti (Frenguelli, 1922:499, figs. $2 \mathrm{~B}, 2 \mathrm{C}$ ) are shorter and relatively broader than those of $P$. capensis, and have a variably developed second anterior accessory cusp which is not present in the Langebaanweg species.

Table 3. Dimensions of the teeth of Prionodelphis capensis from Langebaanweg.

|  | No. | $\begin{gathered} \text { UPPER } \\ \text { A-P } \\ \text { dia- } \\ \text { meter } \end{gathered}$ | Transverse diameter |  | No. | $\begin{gathered} \text { LOWER } \\ \text { A-P } \\ \text { dia- } \\ \text { meter } \end{gathered}$ | Transverse diameter |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Incisors Med. Med. | $\begin{aligned} & L_{11689} \\ & L_{15381 B} \end{aligned}$ | 6,1 6,3 | 5,3 5,2 | Incisors Med. | L 15444A | 3,8 | 3,2 |
| Canines | Lir 686 <br> L 15241 <br> L i ${ }_{5} 630 \mathrm{~B} / 2$ <br> L 15695 | $\begin{array}{r} 10,1 \\ \text { c. } \begin{array}{r} 10,2 \\ \text { c. } 10,1 \\ 10,1 \end{array} ~ \end{array}$ | $\begin{array}{r} 7,5 \\ 7,5 \\ \text { c. } 8,0 \\ 7,4 \end{array}$ | Canines |  | $\begin{aligned} & 9,2 \\ & 9,0 \\ & 9,6 \end{aligned}$ | $\begin{aligned} & 7,4 \\ & 7,3 \\ & 7,8 \end{aligned}$ |
| Post- <br> canines ist 2nd | $\begin{aligned} & \mathrm{L}_{1}{ }_{168} 68 \\ & \mathrm{~L}_{5} 6 \mathrm{II} \end{aligned}$ | $\begin{array}{r} 9,0 \\ 13,9 \end{array}$ | $\begin{array}{r} 6,1 \\ c .7,0 \end{array}$ | Postcanines Ist 2nd ?2nd | L 15580 <br> L $15680 \mathrm{~B} / 2$ <br> L 15736 A | $\begin{array}{r} 9,7 \\ 14,5 \\ 14,2 \end{array}$ | $\begin{aligned} & 5,6 \\ & 6,4 \\ & 6,5 \end{aligned}$ |
| $\begin{aligned} & 3^{\text {rd }} \\ & 4^{\text {th }} \\ & 5 \text { th } \\ & 5 \text { th } \end{aligned}$ |  | 14,5 13,4 9,8 10,4 12 | $\begin{aligned} & 7,5 \\ & 7,4 \\ & 6,4 \\ & 5,5 \end{aligned}$ |  |  |  |  |
| ? | L 12557 | 12,9 | 8, 1 | ? | L $15429 / \mathrm{I}$ | c. 15,0 | 6,5 |
| ? | L 12556 | 13,7 | 7,8 | ? | L 15413 B | 15,4 | 6,4 |
| ? | $\mathrm{L}_{15630 B / 2}$ | 13,8 | 7,2 | ? | Lior6o | 14,8 | 7,2 |
| ? | $\mathrm{L}_{1} 5664$ | 14,1 | 7,8 | ? | $\mathrm{L}_{15444 \mathrm{~B}} \mathrm{~L}_{15771}$ | 15,2 15,0 | 7,0 |
|  |  |  |  | ? | L 12124 | 16,0 | 7,3 |

The proportions of the upper and lower cheek teeth to one another are similar in $P$. capensis and $P$. rovereti, and differ from some extant monachines (Table I).

The height of the mandibular corpus is fairly constant between the poste-
rior limit of the symphysis and the fifth postcanine (Plate 8). In the mandible fragment L 12299, the masseteric fossa begins about 20 mm behind the fifth postcanine, and the ascending ramus begins inclining at about this point. There is no corresponding upward inflection of the inferior margin of the corpus as there is in Leptonychotes, and the fossil resembles most other phocid species in this respect.

The symphysis of the mandible is short relative to that of some monachine seals, and terminates below the posterior root of the second postcanine. It is typically monachine, however, and has a strong, oval articular surface over the entire depth of the jaw. The mandibular condyle is not known.

There are multiple mental foramina towards the anterior part of the corpus.

## THE POSTGRANIAL SKELETON

While the skull characters of Prionodelphis capensis show its relationships to lie with the Monachinae, certain features of the postcranial skeleton are more commonly found among the Phocinae (see King, 1966). Presumably, the 'phocine' characteristics are inherited from the primitive ancestral stock, and are features which were lost by the Monachinae during their later development. Characteristics which are typically monachine are also evident.

## Vertebrae

Associated with the innominate to be described later ( $\mathrm{L}_{15849 \text { ), were a }}$ number of vertebrae, most of which were badly crushed and incomplete. However, two lumbar ( $\mathrm{L}_{15849 \mathrm{Ar}}$ and A2) and one caudal vertebra ( $\mathrm{L}_{15857}$ ) were reasonably well preserved although still incomplete (Plate ioC, D, F). Other vertebrae recorded are an axis ( $L_{7563}$ ), one other cervical ( $\mathrm{L}_{15689}$ ), and a sacral ( $\mathrm{L}_{15396 \text { ) (Plate ioB, E), all of which are damaged, and a number }}$ of other fragmentary specimens.

Only the centrum of the second cervical vertebra is preserved. The odontoid process is prominent, with a length of 16 mm and a maximum transverse diameter of $17,7 \mathrm{~mm}$. The total length of the centrum is 53 mm , and the transverse diameter of the anterior articular end is estimated to be about 55 mm . The other cervical vertebra, probably a fourth, consists of the centrum, parts of the left transverse processes enclosing the vertebrarterial canal, and part of the left half of the neural arch. The centrum is $47,6 \mathrm{~mm}$ long and the transverse diameter of the anterior epiphysis is $31,4 \mathrm{~mm}$. Both these specimens have the reduced transverse processes which characterize the Phocidae (King, 1964:98).

The two lumbar vertebrae, a second or third and a fifth, are similar in size to those of Pliophoca etrusca (see Ugolini, 1902, and Table 4). The transverse processes of the fifth lumbar vertebra are very prominent, which is characteristic of all Phocidae (King, 1964:99).

Although the sacral vertebra, a third, is from an adult individual it was not fused to the second. However, the anterior end of the preserved right transverse process is markedly rugose, suggesting that there was a strong cartilaginous

Table 4. Dimensions of lumbar vertebrae of Prionodelphis and Pliophoca.

attachment between it and the posterior end of the second sacral transverse process. This specimen is unusual in that it lacks the left transverse process, and the anterior end of the centrum and neural arch are, as a result, asymmetrical with the dorso-ventral median axis directed from right to left at a slight angle to the normal line. The length of the centrum is estimated to be 38 mm , while that of a Pliophoca etrusca specimen is 36 mm (Ugolini, 1902).

In the caudal vertebra, a first, much of the centrum is lost, apparently having been gnawed away. Crushing, punctures, and gnaw-marks resulting from grasping and chewing by carnivores are a not uncommon feature of the fossils from Langebaanweg. Both anterior and posterior zygapophyses of the caudal vertebra are well developed, and the distance between their anterior and posterior limits is 47 mm . The transverse diameter of the anterior epiphysis of the centrum is approximately 25 mm .

## Anterior Limb

## Scapula

A single incomplete pinniped scapula ( L 2160 ) is known from Langebaanweg (Plate ioA). This specimen consists of the articular end, neck and lower parts of the blade. The acromion and internal margin of the articulation are damaged.

The glenoid cavity is markedly elongated and concave; the concavity as well as the breadth/length ratio being comparable only to that of Lobodon and Monachus ( $67,9 \%$ for L 2160, 72,0\% for one Lobodon scapula, and 67,1 $\%$ for one Monachus schauinslandi scapula). Other monachine genera have shallower and more nearly equidimensional scapular glenoid fossae.

Also most comparable to Lobodon and particularly Monachus, the neck of the fossil scapula is extremely short and has an antero-posterior diameter of $45,8 \mathrm{~mm}$. The spine is strongly developed for a monachine seal, but might not exceed the development of that of Lobodon. Too little is preserved to be certain. The sharp scapular notch, where the anterior margin of the coracoid process turns into the inferior border of the supraspinous fossa, suggests that the anterior border of the scapula may have been straight and vertical as in the Antarctic monachines.

The medial surface of the subscapular fossa is divided into two parts by a prominent ridge most resembling that of Monachus in its prominence and location. The ridge is more prominent than in Monachus, however, and is better developed than in any living phocid. Although the scapula differs greatly in other respects, the prominence of this subscapular ridge is equalled only by the

Miocene Phoca vindobonensis Toula (1897: pl. 9, fig. 15a).

## Humerus

All fourteen humeri recovered to date in which the distal end is preserved have an entepicondylar foramen. The supinator ridge is well developed in all specimens (Plate ir). These features are characteristic of the extant Phocinae (King, 1966), but in at least one extinct monachine, Monotherium, an entepicondylar foramen was present (see Van Beneden, 1877). This is a characteristic of particular significance in the interpretation of the relationships of Prionodelphis capensis.

Because of the rather startling phocine appearance of the humerus of $P$. capensis, it is appropriate to examine this bone in greater detail. Plate 12 shows the right humeri of Monachus schauinslandi, P. capensis and Cystophora cristata. The humerus of Cystophora was selected because it, of all phocine genera, most resembles that of the fossil. The humerus of another phocine, Erignathus barbatus, is shown with those of $P$. capensis and M. schauinslandi in Figure 2. It is immediately evident that the $P$. capensis humerus exhibits not only phocine, but also monachine characteristics.

Apart from the two characters already mentioned, there is also a considerable difference between phocine and monachine humeri in the region of the deltoid crest. In all pinnipeds the pectoralis muscle is prominent, and its insertion on the humerus is strengthened. In the phocine seals this has been accomplished by an anteriorly directed enlargement of the medial edge of the deltoid crest toward the enlarged lesser tubercle, so that the intertubercular groove becomes circular in cross-section, coming to within $40^{\circ}$ of completely encircling the bicipital tendon in some species. In the otarioid seals, the pectoral insertion is similarly strengthened in this area, and the intertubercular groove becomes trenchant, although the lesser tubercle remains 'lesser'. Strengthening of the pectorial insertion on the phocine humerus does not take place by extending the insertional area distally along the shaft of the humerus but, rather, the insertional area terminates abruptly at a strong process on the distal end of the deltoid crest. Beyond this point the anterior margin of the phocine humerus shaft is concave as it curves to meet the distal articulation, and is devoid of muscle scars.

The transverse development of the deltoid crest of phocine humeri is also in evidence laterally, where a lip of bone overhangs the area of insertion of the deltoid muscle.

In the monachine seals anterior enlargement of the deltoid crest is minimal, the intertubercular groove remains widely open, and there is no overhanging of bone on the lateral edge of the crest. The pectoralis insertion on the humerus is strengthened by extending its area distally down the shaft toward the radial fossa (much reduced in monachine seals) and the distal articulation. The anterior margin of the monachine humerus is, therefore, straight or even convex, and muscle scars are prominent where the deltoid crest blends distally into the shaft. A similar elongation of the pectoral insertion is also present in the otarioid


Fig. 2. Anterior and medial views of the right humeri of Monachus schauinslandi (1), Prionodelphis capensis (2), and Erignathus barbatus (3). (D - deltoid crest; S - supinator ridge.)
seals, consistent with the greater development of the pectoral muscles in that group.

In the $P$. capensis humerus, the bicipital or intertubercular groove is widely open, the deltoid crest blends smoothly into the distal part of the shaft, and the muscle scars are prominent on the shaft below the deltoid crest. In these respects it is typically monachine. It does, however, have the lateral overhanging of the deltoid crest similar to that of the phocines. The deltoid crest is as a result, more prominently developed than in modern monachines.

It is concluded that the pattern of strengthening of the pectoral muscle insertion is a more useful character in classifying phocids than the two characters given by King (1966), which apply to modern species only. In the P. capensis humerus the loss of the entepicondylar foramen, a reduction in the size of the supinator ridge and reduction of the lateral development of the deltoid crest, would reduce it to an almost exact replica of that of $M$. schauinslandi.

The humerus of $P$. capensis is stoutly proportioned, and the most complete specimen known ( L 4638 ) has a total length of 138 mm between the head and median condyle. The transverse diameter of the distal end is $52,7 \mathrm{~mm}$ (mean of 8 specimens). It is stouter than that of living Monachus and Leptonychotes, and some fossil monachines such as Pliophoca etrusca and Monotherium aberratum; comparable in stoutness to the humerus of living Hydrurga and fossil Palaeophoca nystii; it is less stout than that of living Lobodon and Ommatophoca. Relatively shorter and stouter humeri suggest more pelagic adaptations in living phocids.

## Ulna

According to King (1966:390) there are no consistent differences between the ulnae of phocines and monachines. In our sample of ulnae from living species there is a suggestion that the tuberosity for insertion of the internal anconeal muscle (Howell, 1929:75) is much more produced and somewhat more posterior in location in the phocine seals. In the monachine seals, if any anconeal tuberosity can be said to exist, it is continous with the triceps insertion at the anterodorsal apex of the olecranon (ulnar orientation is here considered to be with the long axis vertical, as in fissiped carnivores). There also appears to be slight but persistent sigmoid flexure of the phocine ulnar shaft, in anterior or humeral aspect, because of a lateral curve distal to the radial notch, whereas the shaft of modern monachine seals is straight distal to the radial notch.

Of these two suggestive characters, the nature of the anconeal insertion is not preserved on the ulnae of $P$. capensis, but there appears to be a slight phocine outward curvature of the shaft in the vicinity of the interosseous crest, distal to the radial notch. It thus seems possible that the straight shaft of the living monachine seals has been recently acquired.

Except for the slight curvature of the shaft, the ulna of P. capensis (Plate 13) greatly resembles that of $M$. schauinslandi. The proportions are nearly equal and both are characterized by an extremely elongated posterior process of the olecranon, giving the bone a very hatchet-like appearance.

There is considerable generic variation, and no subfamilial differentiation, in the configuation and relative location of the humeral and proximal radial articulation on the phocid ulna. In these articular facets $P$. capensis also resembles living Monachus. The radial facet has minimal medial offset and distal separation from the facet for the humerus, and faces anteriorly rather than anterolaterally as in other phocids. The humeral facet has a distinct medial curvature, as does that of many other phocids. It also seems possible, therefore, that the lateral positioning of the head of the radius, as reflected in its articulation with the ulna, may be a rather recent development, at least in the monachine seals.

The total length of the ulna of $P$. capensis, estimated from two incomplete specimens, is about 170 mm .

## Radius

Reflecting the more anterior orientation of the radial articulation of the ulna, the radius of Prionodelphis capensis (Plate 14) was orientated more anteriorly from the ulna than is that of some living monachines and apparently all living phocines. As a result, the radial tuberosity lies distinctly on the medial side of the radius, as on the radius of living Monachus, and not on the posteromedial surface as is the case of the radius of living phocines.

King (1969: fig. 31) has pointed out that the distal articulations of the radii of Hydrurga and Ommatophoca have convex surfaces that curve on to the medial (flexor) side of the radius at or near the anterior (preaxial) limit of the articular surface. The radius of Halichoerus has this medially curving segment of the articulation about midway between the anterior and posterior limits of the articular surface. The condition in the latter produces a moderate indentation in the medial margin of the articular facet when viewed distally.

The pattern of distal articulation on the radius of Halichoerus seems to be a characteristic of all extant Phocinae. The radii of living monachine seals follow the patterns shown by King (1969) for Ommatophoca and Hydrurga. In Ommatophoca and Mirounga the distal articular surface is roughly rectangular, and the anteromedial quarter curves on to the medial surface. In Hydrurga, Lobodon and Monachus the anteromedial corner of the articulation is extended so that the part of the surface that curves on to the medial side of the radius is almost a separate articulation (see King, 1969: fig. 3ib).

In Prionodelphis capensis the distal articulation of the radius (L 2935) most resembles that of Mirounga. In general configuration the radius is markedly spatulate with a prominent anterior crest for insertion of supinator and pronator teres muscles, most closely resembling in this respect, Hydrurga and Ommatophoca of the living monachines.

The total length of the radius of $P$. capensis, estimated from two incomplete specimens, is about 145 mm with the greatest anteroposterior diameter of the shaft being $5 \mathrm{I}, 4 \mathrm{~mm}$. The dimensions of the proximal end are 30,8 by $23,4 \mathrm{~mm}$ (mean of seven specimens).

## Posterior Limb

## Innominate

The most complete innominate known ( $\mathrm{L}_{15849 \mathrm{~A}}$ ) lacks the most anterior part of the ilium, and the posterior parts of the ischium and pubis (Plate 15). There is, however, sufficient of this bone remaining to enable a fairly confident assessment of its characteristics.

The ilium is weakly everted, approximately to the extent of that of Erignathus and most monachine seals. Erignathus is atypical of the Phocinae, the remainder of which have a strongly everted ilium 'with a deep lateral excavation' (King, 1969:392); the latter character is absent in P. capensis.

Comparison of the post-acetabular proportions of the entire innominate is precluded by the incompleteness of the specimens. However, a comparison of the distance between the centre of the acetabulum and the apex of the ischiatic spine, to the width of the obturator foramen ventral to the ischiatic spine (not always maximum width), results in an equally distinct separation between monachine and phocine seals (Table 5). Interestingly, measurements of photographs and drawings published by $\operatorname{King}(1956,1966,1969)$, also conform quite well.

Table 5. Innominate proportions of some phocids.

${ }^{1}$ From photo in King, 1956.
${ }^{2}$ From drawing in either King, 1966, or King, 1969.
As with the living Antarctic monachines (but not Monachus schauinslandi) and some phocine seals, the innominate of $P$. capensis appears rather thick across the acetabulum. It appears similarly thick in available specimens of Pagophilus and Cystophora. This appearance is caused by a relatively small acetabulum.

## Femur

Only the distal end of the femur of $P$. capensis is known (L ior3i) (Plate 16A). The patellar facet is somewhat broader than tall, as in the monachine seals, while a fairly marked pit for the popliteus muscle on the lateral epicondyle shows resemblance to the phocine seals.

## Tibia

The tibia of $P$. capensis is remarkable for the development of pronounced fossae on the posterior and antero-lateral surfaces. In the specimen L 10128/9 (Plate 16B, C, D) the thickness of bone between these two fossae is as low as $0,75 \mathrm{~mm}$. This condition most resembles that in the tibia of Halichoerus. King (1966) states that the post-tibial fossa is more pronounced in the Phocinae than in the Monachinae.

Distally, the tibia of $P$. capensis is conspicuously broad and anteroposteriorly flattened (Plate i6E, F), a condition very similar to Pliophoca etrusca (Tavani, 1942: fig. 18). The fibular contact is sharply angled outward suggesting that the fibula was rather markedly bowed.

## Pes

A calcaneum (L ioir8), two astragali (L ioi30, L io993), one navicular (Li585I), one entocuneiform (Lio124), one metatarsal V (L io996) and two phalanges (L iog99, L io205) of Prionodelphis capensis are known (Plates i 7 \& 18). At first glance it seems obvious that $P$. capensis has long metatarsal bones relative to the size of the astragalus, and in fact, a sampling of the relative sizes of these two bones in seven living genera seems to bear this out (Table 6). The Langebaanweg seal appears to have relatively longer metatarsals than those of the compared living genera except Monachus.

Table 6. Tarsal-metatarsal comparison of some phocids.

| Genus |  | (1) <br> Greatest length <br> of MT V | (2) <br> Greatest length <br> of astragalus | (3) <br> Greatest length <br> of cuboid-MT <br> IV facet | $(2 / \mathrm{I})$ |
| :--- | :---: | :---: | :---: | :---: | :---: |

As the fossil astragali and metatarsal V were not found in association, they could represent different sized individuals, creating a false impression of relative metatarsal size. To check this possible error a similar comparison of the cuboid-metatarsal IV articular facet on metatarsal V , to the total length of the
the metatarsal was also made (Table 6). Comparable results were achieved, except for Mirounga and Pagophilus whose relative proportions were not consistent. It therefore appears probable that $P$. capensis did indeed have relatively elongated metatarsal bones and hence had relatively large hind flippers.

King (r966:393-394) has suggested that the more distal articular facet between the astragalus and the calcaneum is relatively long in the phocine seals and short in the monachines. Although there is no exception in the monachine specimens available, there seems to be great variation in the form of this facet on the astragalus of the phocine seals. Our specimen of Cystophora appears decidedly 'monachine' in this character: astragali of Erignathus appear to vary from distinctly 'monachine' to distinctly 'phocine'. This articular surface on $P$. capensis is distinctly elongated and hence 'phocine' to the extent that the character is valid.

In all respects other than their mutual lower articulation, the astragalus and calcaneum of $P$. capensis are extremely similar to those of $M$. schauinslandi. If the tibial articulation of the astragalus is arbitrarily taken to be dorsal, so that the fibular articulation is vertical, these surfaces are low relative to the body of the astragalus and the fibular articulation extends about to the most ventral limit of the bone. Among living monachines a similar condition is found in both Monachus and Mirounga, which differ greatly in this respect from Hydrurga. Wide variation is found also in the phocine seals: the astragalus of Pagophilus is perhaps most similar to that of $P$. capensis, while that of Phoca differs the most.

The tibial articulation is cylindrical, as in Mirounga, Monachus and Lobodon, rather than spherical as in Hydrurga. No angular boundary separates the distal articulation for the navicular from the adjacent articulation for the calcaneum. The astragalus is rather short-necked and has a short calcanear process.

The dimensions of the calcaneum L iori8 (Table 7) are remarkably similar to that of a M. schauinslandi specimen recorded by Robinette \& Stains (1970: table 1), while its porportions are clearly monachine rather than phocine (Robinette \& Stains, 1970: table 2). The metrical data presented in Table 7 confirms the observations on the similarities between the calcanea of P. capensis, Monachus (especially M. schauinslandi and M. tropicalis) and Mirounga, and also illustrates differences from those of Hydrurga, Leptonychotes and Ommatophoca.

The posterior ${ }^{1}$ astragalar articulation of the calcaneum of $P$. capensis is narrow relative to its length. This contrasts with that facet of the calcaneum of Hydrurga and Monachus schauinslandi, in which it is nearly as wide as it is long. Robinette and Stains (1970:535) state that the facet is narrower on the calcaneum of $M$. tropicalis than on that of $M$. schauinslandi.

The facet of the navicular, for articulation with the entocuneiform, is notably equidimensional and flat. The articular surface on the entocuneiform, for contact with metatarsal I, is notably elongated, suggesting a more slotted

[^0]Table 7. Dimensions of calcaneum of Prionodelphis capensis from Langebaanweg, compared with those of some modern monachines.

| Species |  | N | TL | W | DVH | W/TL | DVH/TL | DVH/W |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Monachus monachus $\dagger$ |  | 1 | 63,2 | 29,8 | 30,0 | 47 | 47 | 101 |
| Mirounga angustirostris $\dagger$ |  | 6 | 79,0* | 40,9* | 38,7* | 52 | 49 | 95 |
| Monachus schauinslandi $\dagger$ |  | 1 | 59, I | 31,9 | 30,3 | 54 | 51 | 95 |
| Monachus tropicalis $\dagger$ |  | 1 | 54,9 | 27,3 | 28,5 | 50 | 51 | 104 |
| Prionodelphis capensis |  | 1 | 58,4 | 30,7 | 30,4 | 53 | 52 | 99 |
| Hydrurga leptonyx $\dagger$. |  | 1 | 74,5 | 38,7 | 42,0 | 52 | 56 | 108 |
| Leptonychotes weddelli $\dagger$ |  | 1 | 71,5 | 38,2 | 41,9 | 53 | 58 | 110 |
| Ommatophoca rossi $\dagger$ |  | 1 | 56,o | 30,4 | 34,6 | 54 | 61 | 114 |
| $\dagger$ From Robinet $\mathrm{TL}=$ total leng |  |  |  |  | * Average figures.$\mathrm{W}=\text { width } .$ |  |  |  |

DVH $=$ dorsoventral height.
proximal articulation rather than the basined articulation on metatarsal I of most living seals. Although both the navicular and the entocuneiform appear large relative to the known astragali and calcaneum, they appear to be undiagnostic of subfamily affinities.

As has been mentioned, metatarsal V appears to be relatively elongated. The two known phalanges also appear to be conspicuously elongated and slender when compared with those of living phocids. Otherwise they seem to have no distinctive features.

## Discussion

In assigning the Langebaanweg phocid to the genus Prionodelphis, it is recognized that reassessment may be required when more material of $P$. rovereti is found. Generic identity is based upon the remarkable similarity of the few fragments from Argentina to the South African material and on the belief that the lack of greater knowledge is a stronger argument against the establishment of a new genus than it is against tentative assignment to the same genus.

The similarity of the cheek teeth of $P$. rovereti and $P$. capensis is very strong. The transverse narrowness of the cheek-teeth, more evident in the latter species, and a posterointernal shelf on the upper cingula only, are features found only in some of the Antarctic genera of the monachine seals. As mentioned, the tooth proportions and cingular shelf are most similar to the condition in Hydrurga. In addition, the greatly reduced and distinctly recurved last upper postcanine, of both species of Prionodelphis is singularly suggestive of a close relationship, and is not known in other monachine seals, although there is some resemblance to the more anterior cheek teeth of Leptonychotes, and to a lesser extent also Ommatophoca.

Apart from differences in the dimensions of the teeth of $P$. capensis and $P$. rovereti, some differences in morphology are also evident. Other than the upper
fifth, the only known upper postcanine of $P$. rovereti (Frenguelli, 1922: fig. 1), differs from the second to fourth upper postcanines of $P$. capensis in having a far larger posterior root, which is at least partially divided longitudinally. This feature of the Argentinian species, as well as its slightly broader cheek-teeth, can be interpreted as being less advanced characteristics. The anterior part of the crown of this tooth of $P$. rovereti lacks'a distinct accessory cusp, which is present in all the known postcanines of $P$. capensis. The anterior part of the $P$. rovereti tooth is markedly convex, and the lingual view (Frenguelli, 1922: fig. rc) shows a small step more or less where an accessory cusp might be expected. It is possible, therefore, that this specimen has the anterior accessory cusp masked by some individual variation, and that normally such a cusp was present. The lower postcanines of $P$. rovereti (Frenguelli, 1922: fig. 2B, C) are illustrated as having not one, but two anterior accessory cusps, with the anterior and posterior parts of the teeth being almost mirror images of one another. In none of the postcanines of $P$. capensis is a second anterior accessory cusp known, and its development in the Argentinian species could be a more advanced specialization.

The age of the Argentinian species is even more uncertain than that from Langebaanweg. The Entre Rios deposits, from which the specimens of $P$. rovereti came, appear to be of Pliocene age (Langston, 1965 : table 3) and would seem, therefore, to be roughly the same age as those from Langebaanweg. However, most often these deposits have been referred to as being late Miocene or early Pliocene, and the latter age is given by Romer (1966).

Discussion on the relationship of $P$. capensis to $P$. rovereti will be more meaningful when more specimens of the latter are known, but the observed differences between the two sets of specimens, the possibility of a temporal difference in the deposits from which they come, and their geographical separation suggest a distinction between the South African and Argentinian fossils at least at the species level.

The relatively poor fossil record of the Phocidae in general renders interpretation of the wider relationships of $P$. capensis equally problematical. Some features of the fossil seal from South Africa are found, among the living seals, only in the Phocinae. However, a number of features are clearly monachine and these suggest that the dichotomy from the primitive phocid into the two extant subfamilies was a result of two distinctly different patterns of specialization to better adapt to pelagic existence. These adaptations relate to, amongst other things, greater swimming ability and underwater hearing. As has been pointed out throughout the description, in all of these adaptations $P$. capensis has clearly followed the monachine pattern.

To judge from the comparisons between $P$. capensis and the living phocine and monachine seals, differences in adaptation toward greater swimming ability appear most evident in the proximal limb elements. Subfamilial differences in the structure of the humerus related to the strengthening of the pectoralis muscle have been outlined in the discussion of this bone. In the Phoci-
nae the pectoralis insertion has been strengthened by exaggeration of the deltoid crest, and in the Monachinae strengthening of this same muscle has been accompanied by a distally extended insertional area on the humerus shaft.

One might infer that the phocid ancestral to the living subfamilies had a humerus of relatively slender proportions showing a moderate development of both types of pectoralis insertion, such as seen on the humerus of 'Phoca' vindobonensis Toula (1897: pl. i, fig. 16) or Leptophoca lenis True (1906: pl. 75). Such fossil phocids as Monotherium aberratum Van Beneden (1877: pl. i7, figs. 1-4) appear to have the insertional area extended so far distally on the shaft that a monachine condition seems indisputable, while others such as Phocanella pumila Van Beneden (1877: pl. 14, figs. I-4), have clearly evolved the phocine condition by strengthening the deltoid crest and eliminating all pectoralis insertion on the shaft distal to the crest. In addition, the presence of an entepicondylar foramen on the humerus appears to be a primitive feature. It is present in all of these fossil seals, including Prionodelphis capensis, and is retained in the living phocine seals as well. Only at the stage of evolution evident in the living phocids does the presence of this foramen become diagnostic of subfamily affinity.

Consideration of the subfamilial differences in swimming adaptations which might be found in the pelvic limbs has been hampered in this study by the lack of a complete specimen of the femur of $P$. capensis. Nevertheless, King (1966:392) has pointed out that, except for the genus Erignathus, the phocine seals may be recognized by their extremely everted ilium. Both leverage and strength of the insertion of the massive muscles of the back, the iliocostalis system, are benefited by this structure, as well as are most of the gluteus group which directly transfers the forces of the back to the femur to produce the characteristic phocid swimming motion. The advantages of this structure seem so obvious that it is puzzling why none of the monachine seals have developed it, or why it developed so late in the history of the phocid seals. Few fossil seals in which the pelvis is known, exhibit the phocine everted ilium.

The interpretation of the functions of osteological characters in the phocine and monachine ear regions is subjective, but according to one interpretation (Repenning, in press), two of these differences relate to improved underwater hearing.

The presence of a more or less horizontal crest on the external surface of the mastoid bone in all phocine seals is correlated with a greater directional selectivity of sounds in water originating above or below the head; this crest is not present in the monachine seals, including $P$. capensis.

The development of an enlarged petrosal apex in all seals is correlated with a greater sensitivity to sound in water, and this development is conspicuously less in $P$. capensis than in the living Antarctic seals. The enlargement of the petrosal apex is in the form of a globular mass in the phocine seals, while in the monachine seals, with the exception of Mirounga, the apex is enlarged as a rather low and broad structure. In this respect $P$. capensis is clearly monachine.

It should also be noted that broadening is slight in Hydrurga, and that enlargement of the apex is partly accomplished by thickening; the structure does not appear globular as in the phocine seals, however. Furthermore, enlargement of a petrosal apex is minimal in Monachus, less than in P. capensis; in this respect Monachus might be expected to be most similar to the ancestral phocid from which the extant subfamilies evolved.

Monachus is the least specialized of the living monachine seals in the enlargement of the petrosal apex, strengthening of the humerus, distal broadening of the radius, enlargement of the ilium, strengthening of the femur and modification of the dentition. Except for the condition of the femur, which is unknown in the South African fossil, and possibly the enlargement of the ilium, which is incompletely preserved, Monachus is also less specialized in these features than P. capensis.

From the preceding consideration of Prionodelphis capensis and related seals, the following features appear most likely to be those that would characterize the ancestral protophocid from which the two modern subfamilies, the Phocinae and the Monachinae, evolved: dentition with primary cusp flanked by one accessory cusp anteriorly and one posteriorly, much the same as seen in Praepusa pannonica Kretzoi (1941: fig. 1), ear region much as in living Monachus, and postcranial skeleton unspecialized as in 'Phoca' vindobonensis Toula (1897).

The relationship of $P$. capensis to the Antarctic monachines is evident in a broad sense, but it is not clearly ancestral to any of the four living genera. The highly modified dentitions of the living genera differentiate them most strikingly from the Pliocene fossil. The great reduction of the last upper postcanine tooth of $P$. capensis seems to preclude the possibility of it being ancestral to Hydrurga or Lobodon, while the great simplification of the teeth of Leptonychotes and Ommatophoca leave little basis for interpretation. Hydrurga, Lobodon and Ommatophoca are all clearly better adapted to pelagic life in their postcranial specializations than was P. capensis. All of the Antarctic seals, and Leptonychotes in particular, have a greater development of the petrosal apex than does $P$. capensis, which indicates that the latter had less acute hearing underwater. In all these respects $P$. capensis is less advanced than the Antarctic seals, but more advanced than Monachus.

It seems probable that Prionodelphis, presently known only by the species $P$. rovereti and $P$. capensis, was not the only Pliocene monachine of the southern seas, and that some or all of the modern Antarctic genera derive from a related but unknown form.

All previously described fossil phocids are known from incomplete remains and a good many from a very few, or even one bone. The humeri and femora are the most commonly described because they are among the more durable bones of the body, and, presumably, their size lends them to discovery. Excluding mandibular fragments, Pliophoca etrusca Tavani (1942; a skull), Phoca pontica Eichwald (1853; a cranium) and Phoca pontica Alekseev (1924; a rostrum) are the only fossil phocids of which the skulls are even partially known. Despite its
fragmentary nature, Prionodelphis capensis is one of the most completely known fossil phocids, and since the systematic investigation of the Langebaanweg deposits is still in its early stages, it can be expected that much more material will become available for study in the future.

## Summary

Pinniped remains from the late Pliocene deposits at Langebaanweg in South Africa are described. The material is referred to Prionodelphis capensis n. sp. (family Phocidae, subfamily Monachinae). On the basis of this material, the genus Prionodelphis Frenguelli 1922 is defined. The relationships of the Langebaanweg species to extant and fossil monachines are discussed, and morphological characters, by which fossil Phocinae and Monachinae can be differentiated are suggested.

## Acrnowledgements

We wish to thank A. W. Mansfield and D. E. Sergeant of the Fisheries Research Board of Canada; C. W. Mack of the Harvard University Museum of Comparative Zoology; O. P. Pearson and R. L. Jones of the University of California Museum of Vertebrate Zoology; R. H. Mansville, K. W. Kenyon, and E. Kridler of the (U.S.) Bureau of Sport Fisheries and Wildlife; C. O. Handley, Jr. and G. E. Ray of the (U.S.) National Museum of Natural History; E. H. Bryan, Jr. of the Bernice P. Bishop Museum (Honolulu) ; F. H. Fay of the (U.S.) Arctic Health Research Center; and L. Giannelli of the Universita di Pisa Museo di Paleontologia for the loan or gift of material used in this study.

The current investigations at Langebaanweg are being supported by the South African Council for Scientific and Industrial Research, Chemfos Ltd. (a subsidiary of the African Metals Corporation) and Shell South Africa (Pty.) Ltd. The Wenner-Gren Foundation for Anthropological Research, New York, provided the vehicle used in the field work at Langebaanweg (Grant no. 2752-1834).

We are indebted to Mr. H. Krumm and Mr. G. Benfield of Langebaanweg, whose co-operation with the South African Museum led to the recovery of most of the material described in this paper.

Francis H. Fay and Clayton E. Ray kindly offered comments on the manuscript of this paper, and we are grateful to them for their observations and criticisms.

## References

Alekseev, A. K. 1924. [Seals in the Sarmatian deposits of southern Russia.] Zh. nauchno-issled. Kafedr Odesse 1: 26-34. [In Russian.]
Allen, J. A. i88o. History of North American pinnipeds: a monograph of the walruses, sealions, sea-bears and seals of North America. Misc. Publs U.S. geol. geogr. Surv. Territ. 12: $i-x v i, 1-785$.
Boné, E. L. \& Singer, R. 1965. Hipparion from Langebaanweg, Cape Province, and a revision of the genus in Africa. Ann.S. Afr. Mus. 48: 273-397.

Brookes, J. 1828. A catalogue of the anatomical and zoological museum of Joshua Brookes. London.
Burns, J. J. \& Fay, F. H. 1970. Comparative morphology of the skull of the Ribbon seal, Histriophoca fasciata, with remarks on systematics of Phocidae. F. Zool. 161: 363-394.
Cabrera, A. 1926. Cetáceos fósiles del Museo de La Plata. Revta Mus. La Plata. 29: 363-41 i.
Eichwald, C. E. von. 1853. Lethaea Rossica, ou Paleontologie de la Russie. 3: Stuttgart: Schweizerbart.
Frenguelli, J. 1922. Prionodelphis rovereti un representante de la familia 'Squalodontidae' en al Paranense Superior de Entre Rios. Boln Acad. nac. Cienc. Córdoba 25: 491-500.
Frenguelli, J. 1926. El Entrerriense de Golfo Nuevo en el Chubut. Boln Acad. nac. Cienc. Córdoba 29: 191-270.
Gray, J. E. 1825. An outline of an attempt at the disposition of Mammalia into tribes and families, with a list of the genera apparently appertaining to each tribe. Ann. Philosophy (n.s.) 10: 337-344.
Hay, O. P. 1930. Pinnipedia. In hay, o. p. Second bibliography and catalogue of the fossil Vertebrata of North America. 2: 555-565. Washington: Carnegie Institution. (Publs Carnegie Instn 390.)
Hendey, Q.B. 1970a. A review of the geology and palaeontology of the Plio/Pleistocene deposits at Langebaanweg, Cape Province. Ann. S. Afr. Mus. 56: 75-117.
Hendey, Q. B. 1970 $b$. The age of the fossiliferous deposits at Langebaanweg, Cape Province. Ann. S. Afr. Mus. 56: 119-131.
Howell, A. B. 1929. Contribution to the comparative anatomy of the eared and earless seals (genera Zalophus and Phoca). Proc. U.S. natn. Mus. 73: 1-142.
Illiger, C. 181 i. Prodromus systematis mammalium et avium. Berlin: Salfeld.
Kellogg, R. 1922. Pinnipeds from Miocene and Pleistocene deposits of California. Univ. Calif. Publs geol. Sci. 13: 23-132.
Kellogg, R. 1942. Tertiary, Quaternary, and Recent marine mammals of South America and the West Indies. Proc. 8th Am. Sci. Congr. (Washington) 3: 445-473.
King, J. E. 1956. The monk seals (genus Monachus). Bull. Br. Mus. nat. Hist. (Zool.) 3: 201-256.
King, J. E. r964. Seals of the world. London: British Museum (Natural History).
King, J. E. 1966. Relationships of the Hooded and Elephant seals (genera Cystophora and Mirounga). 7. Zool. 148: 385-398.
King, J. E. 1969. Some aspects of the anatomy of the Ross seal, Ommatophoca rossi (Pinnipedia: Phocidae). Scient. Rep. Br. Antarct. Surv. 63: 1-54.
Kraglievich, L. 1934. La antigüedad pliocena de las faunas de Monte Hermoso y Chapadmalal. Siglo ilustrado, Montevideo 938: $\mathbf{1 - 1} 36$.
Kretzor, M. 1941. Seehund-Reste aus dem Sarmat von Erd bei Budapest. Földt. Közl. 71: 350356.

Langston, W. 1965. Fossil crocodilians from Colombia and the Cenozoic history of the Crocodilia in South America. Univ. Calif. Publs geol. Sci. 52 : i-vii, 1-157.
Maglio, V. J. \& Hendey, Q.B. i970. New evidence relating to the supposed stegolophodont ancestry of the Elephantidae. S. Afr. archaeol. Bull. 25: 85-87.
Mckenna, M. 1969. The origin and early differentiation of therian mammals. Ann. N.T. Acad. Sci. 167: 217-240.
Repenning, C. A. Underwater hearing in seals: functional morphology. In Functional anatomy of marine mammals. London: Academic Press. (In press.)
Robinette, H. R. \& Stains, H. J. 1970. Comparative study of the calcanea of the Pinnipedia. 7. Mammal. 51: 527-541.

Romer, A. S. 1966. Vertebrate paleontology. 3rd ed. Chicago: University Press.
Savage, D. E. \& Curtis, G. H. 1970. The Villafranchian stage-age and its radiometric dating. Spec. Pap. geol. Soc. Am. 124: 207-231.
Scheffer, V. B. 1958. Seals, sea lions and walruses. Stanford: University Press.
Singer, R. \& Hooijer, D. A. 1958. A Stegolophodon from South Africa. Nature, Lond. 182: ioi-102.
Smirnov, N. A. 1908. [Review of Russian pinnipeds.] Mém. Acad. Sci. St. Petersb. (8) 23: 1-75. [In Russian].
Tavani, G. 1942. Revisione dei resti del pinnipede conservato nel Museo di Geologia di Pisa. Palaeontogr. ital. 40: 97-113.
Tavani, G. 1943. Revisione dei resti di pinnipedi conservati nel Museo geo-paleontologico di Firenze. Atti. Soc. tosc. Sci. nat. Memorie 6I: 34-42.
Toula, F. 1897. Phoca vindobonensis n. sp., von Nussdorf in Wien. Beitr. Paläont. Geol. Öst.-Ung. 11: 47-70.

Trouessart, E.-L. 1897. Catalogus mammalium tam viventium quam fossilium. Berlin: Friedländer. True, F. W. 1906. Description of a new genus and species of fossil seal from the Miocene of Maryland. Proc. U.S. natn. Mus. 30: 835-840.
Ugolini, R. 1902. Il Monachus albiventer Bodd. del Pliocene di Orciano. Palaeontogr. ital. 8: 1-20. Van Beneden, P. J. 1877. Description des ossements fossiles des environs d'Anvers. Annls Mus. r. Hist. nat. Belg. $\mathbf{1}$ : $1-88$.


> 乙

Dorsal view of skull L 15695 . Scale represents 5 cm .


Lateral view of skull Lis 569 . Scale represents 5 cm .


L

Ventral view of skull $\mathrm{L}_{15695}$. Scale represents 5 cm .


Right lateral view of rostrum $\mathrm{L}_{\mathrm{I} 2695}$.




D

A, B, C \& D. Dorsal, ventral, lateral and internal views of temporal $\mathrm{L}_{15}{ }_{5} 62$.



A, B \& C. Lateral, internal and dorsal views of mandible L 7556.

A. Lateral view of upper incisor Lir689.
B. Lateral view of upper canine $L_{1} 1686$ (aged individual).
C. Buccal and occlusal views of first upper postcanine L 11687.
D. Buccal and occlusal views of second, third of fourth upper postcanine $L_{1564}$.
E. Lingual and occlusal views of fifth upper postcanine $\mathrm{L}_{15429}$.
F. Lateral view of lower canine Li3152 (young individual).
G. Buccal and occlusal view of lower second, third, fourth or fifth postcanine L i5444B.

A. Lateral view of scapula $L_{2160}$.
B. Lateral and ventral views of second cervical vertebra $L 7563$.
C. Ventral view of a second or third lumbar vertebra L ${ }_{1} 849$ A2.
D. Ventral view of fifth lumbar vertebra Li5849 AI.
E. Anterior view of third sacral vertebra L 15396.
F. Anterior view of first caudal vertebra $\mathrm{L}_{15857}$.


A \& B. Anterior and posterior views of humerus L ${ }_{21} 57$.
C. Medial view of humerus L 4638 .


Anterior views of the right humeri of Monachus schauinslandi (A), Prionodelphis capensis (B) and Cystophora cristata (C). Scale approximately $\times \frac{1}{2}$.


A


B
A. Lateral view of left ulna $\mathrm{L}_{15682}$.
B. Medial view of right ulna L ${ }_{2161}$.


A


## B

A. Lateral view of right radius L 12869 .
B. View of distal articulation of right radius L 2935 (anterior parts of specimen are lost).



B


## E



D


F

A. Distal view of femur Lioi31.

B, C \& D. Posterior, anterior and proximal views of right tibia Lio128/9.
E. Posterior view of left tibia L 2138 .
F. View of distal articulation of right tibia $\mathrm{L}_{\text {Io }}$ 28/9.


A


A. Lateral, ventral and medial views of right calcaneum Lioir8.
B. Dorsal and ventral views of left astragalus L io993.


B
A. Dorsal, lateral and ventral views of left metatarsal V, L iog96.
B. Dorsal view of phalanges L 10205 and L rog99.

## INSTRUCTIONS TO AUTHORS

## Based on

CONFERENCE OF BIOLOGICAL EDITORS, COMMITTEE ON FORM AND STYLE. 1960.
Style manual for biological journals. Washington: American Institute of Biological Sciences.

## MANUSCRIPT

To be typewritten, double spaced, with good margins, arranged in the following order: (1) Heading, consisting of informative but brief title, name(s) of author(s), address(es) of author(s), number of illustrations (plates, figures, enumerated maps and tables) in the article. (2) Contents. (3) The main text, divided into principal divisions with major headings; subheadings to be used sparingly and enumeration of headings to be avoided. (4) Summary. (5) Acknowledgements. (6) References, as below. (7) Key to lettering of figures. (8) Explanation to plates.

## ILLUSTRATIONS

To be reducible to $12 \mathrm{~cm} \times 18 \mathrm{~cm}$ ( 19 cm including caption). A metric scale to appear with all photographs.

## REFERENCES

Harvard system (name and year) to be used: author's name and year of publication given in text; full references at the end of the article, arranged alphabetically by names, chronologically within each name, with suffixes $a, b$, etc. to the year for more than one paper by the same author in that year.
For books give title in italics, edition, volume number, place of publication, publisher.
For journal articles give title of article, title of journal in italics (abbreviated according to the World list of scientific periodicals. 4th ed. London: Butterworths, 1963), series in parentheses, volume number, part number (only if independently paged) in parentheses, pagination.
Examples (note capitalization and punctuation)
Bullough, W. S. 196o. Practical invertebrate anatomy. 2nd ed. London: Macmillan.
Fischer, P.-H. i948. Données sur la résistance et de le vitalité des mollusques. F. Conch., Paris 88: 100-140.
Fischer, P.-H., Duval, M. \& Raffy, A. 1933. Etudes sur les échanges respiratoires des littorines. Archs Zool. exp. gén. 74: 627-634.
Kohn, A. J. 1960a. Ecological notes on Conus (Mollusca: Gastropoda) in the Trincomalee region of Ceylon. Ann. Mag. nat. Hist. (13) 2: 309-320.
Koнn, A. J. ig6ob. Spawning behaviour, egg masses and larval development in Conus from the Indian Ocean. Bull. Bingham oceanogr. Coll. 17 (4): 1 -51.
Thiele, J. igio. Mollusca: B. Polyplacophora, Gastropoda marina, Bivalvia. In schultze, l. Zoologische und anthropologische Ergebnisse einer Forschungsreise im westlichen und zentralen SüdAfrika. 4: 269-270. Jena: Fischer. Denkschr. med-naturw. Ges. Jena 16: 269-270.

## ZOOLOGICAL NOMENCLATURE

To be governed by the rulings of the latest International code of zoological nomenclature issued by the International Trust for Zoological Nomenclature (particularly articles 22 and 51). The Harvard system of reference to be used in the synonymy lists, with the full references incorporated in the list at the end of the article, and not given in contracted form in the synonymy list.

## Example

Scalaria coronata Lamarck, 1816: pl. 451, figs 5a, $b$; Liste: I1. Turton, 1932: 80.
 ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM

| Volume | $\mathbf{5 9}$ | Band |
| :---: | :---: | :--- |
| March | $\mathbf{1 9 7 2}$ | Maart |
| Part | $\mathbf{5}$ | Deel |



# THE EVOLUTION AND DISPERSAL OF THE MONACHINAE (MAMMALIA: PINNIPEDIA) 

By<br>Q. B. HENDEY

# The ANNALS OF THE SOUTH AFRICAN MUSEUM are issued in parts at irregular intervals as material becomes available <br> Obtainable from the South African Museum, P.O. Box 61, Cape Town <br> Die ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM <br> word uitgegee in dele op ongereelde tye na beskikbaarheid van stof <br> Verkrygbaar van die Suid-Afrikaanse Museum, Posbus 61, Kaapstad <br> <br> out of print/uit druk <br> <br> out of print/uit druk <br> <br> 1, 2(1,3,5,7-8), 3(1-2,5, t.-p.i.), 5(2, 5, 7-9), <br> <br> 1, 2(1,3,5,7-8), 3(1-2,5, t.-p.i.), 5(2, 5, 7-9), 6(1, t.-p.i.), 7(1-3), 8, 9(I-2), 10(1), 6(1, t.-p.i.), 7(1-3), 8, 9(I-2), 10(1), II(I-2, 5, 7, t.-p.i.), 2I, 24(2), 27, 3 I(i-3), 33 

 II(I-2, 5, 7, t.-p.i.), 2I, 24(2), 27, 3 I(i-3), 33}

Price of this part/Prys van hierdie deel RI,40

Trustees of the South African Museum © Trustees van die Suid-Afrikaanse Museum 1972

In Suid-Afrika gedruk deur
Die Rustica-pers, Edms., Bpk.
Courtweg, Wynberg, Kaap

# THE EVOLUTION AND DISPERSAL OF THE MONACHINAE (MAMMALIA: PINNIPEDIA) 

By<br>Q.B. Hendey<br>South African Museum, Cape Town<br>(With 2 figures)<br>[Ms. accepted 15 November 1971]

## Contents



## Introduction

The recent description of a late Pliocene monachine seal from Langebaanweg in South Africa (Hendey \& Repenning 1972) included a discussion on the possible relationships of this species (Prionodelphis capensis) to other Monachinae. It is the purpose of the present paper to enlarge upon this topic, and also to comment on the phylogeny and zoogeography of the Monachinae.

The origin and evolution of the Pinnipedia have been the subject of numerous publications (e.g. McLaren 1960; Sarich r969 etc.) and of all the subfamilies of this order, the Monachinae is probably that with the poorest fossil record, and consequently the one whose evolution is least well understood.

A number of fossil monachines ranging in age from Miocene to Pleistocene have been recovered from localities in Europe, while there are Pleistocene records of monk seals from the south-eastern United States (King 1964). The Pliocene monachine from South Africa is only the second Southern Hemisphere record of its kind, the other being the poorly known Prionodelphis rovereti Frenguelli 1922 from the Pliocene of Argentina. There are later Pleistocene and Holocene records of Monachinae from the southern continents, but these are all of species still extant, and are from areas within, or near to, the present ranges of the species concerned. For example, remains of seals have been found preserved by icy conditions in Antarctica, although none of these specimens is more than a few thousand years old (Crane \& Griffen 1968). There are also late Pleistocene and Holocene records of Mirounga leonina from coastal human occupation sites in South Africa, an area where occasional stray individuals of this species are still found.

The sparse fossil record of the Monachinae in the southern continents is curious, since modern representatives of this subfamily are found mainly in the higher latitudes of the Southern Hemisphere. The only extant northern monachines are the comparatively small and widely scattered populations of monk seals (Monachus monachus, M. tropicalis and M. schauinslandi) and the northern elephant seal (Mirounga angustirostris).

Modern monachine distribution is one of the more remarkable features of this subfamily, and in speculating on phyletic relationships within the group it is convenient to consider the possibilities under three separate headings.

## The Monk Seals

Modern monk seals occur in the Mediterranean area and adjacent coast of north-west Africa (Monachus monachus), the Caribbean area (M. tropicalis, which may now be extinct), and in the Leeward chain of islands north-west of Hawaii (M. schauinslandi). On the basis of the admittedly poor fossil record, it seems probable that this group, and the subfamily as a whole, originated from a generalized phocid in the western Europe and Mediterranean areas.

The earliest record of a monachine seal anywhere is Monotherium aberratum Van Beneden 1877 from the late Miocene of Belgium. As far as can be seen from the fragmentary remains known, this species is little different from other generalized phocids of the Miocene. In the absence of any other monachines of comparable age, with the possible exception of $M$. maeoticum Nordmann i860 from southern Russia, it can conveniently be regarded as ancestral to the rest of the subfamily. Indeed, there is nothing in the morphology of known skeletal elements of $M$. aberratum which would preclude it from being ancestral to later Monachinae.

Other European fossil monachines include Pliophoca etrusca Tavani 1942 from the later Pliocene of Italy, and Palaeophoca nystii Van Beneden 1877 from the Pleistocene of Belgium. The status of the Pliocene Pristiphoca occitana Gervais \& Serres 1847 is uncertain, although Kellogg (1922: 78) states that it 'certainly belongs to the same genus, and possibly to the same species, as the fossil form from the Orciano in Italy' (i.e. Pliophoca etrusca). P. etrusca is very similar to the extant $M$. monachus, and it was in fact originally described as a fossil M. albiventer (=monachus) by Ugolini (1902). Whatever the correct nomenclature of the Italian fossils, they are clearly closely related to $M$. monachus, as is also Monotherium maeoticum (Kellogg, 1922). Several other species of Monotherium have been described (Kellogg 1922; King 1964).

This sparse and taxonomically confused fossil record clearly does not allow an unequivocable statement to be made on the phyletic relationships of the species concerned. However, the Monotherium aberratum - Monachus monachus lineage is here taken to include Pliophoca etrusca as an intermediate form, and Palaeophoca nystii as an offshoot.

Elsewhere in the areas of distribution of modern monk seals, the fossil record is extremely poor or non-existent. Nevertheless, some inferences can be
drawn from the few available records of fossil monk seals and related forms.
Although the South African Prionodelphis capensis does exhibit certain characteristics which are typical of modern Antarctic seals, in totality of characters there is a more marked resemblance between it and the monk seals and their ancestors. However, it differs from Pliophoca, Palaeophoca and Monachus in at least one important anatomical detail, and that is the presence of an entepicondylar foramen in the humerus. This feature is also present in the humerus of Monotherium aberratum. It is therefore concluded that the dichotomy of the European monachine lineage, and the lineage which includes $P$. capensis, occurred in the late Miocene or early Pliocene.

Similarly, in view of the late Pliocene date of the fossils from Langebaanweg, it is certain that the southern lineage was established in the south Atlantic sometime earlier in this epoch, or perhaps even during the late Miocene.

Having taken Europe as the centre for monachine evolution, the possible routes followed by early monachines into the South Atlantic need to be considered. When intercontinental migrations of seals took place, it is reasonable to suppose that these would be successful where distance between landfalls was least, and prevailing ocean currents favourable. The role of ocean currents in pinniped migrations may well be more complex than appears at first sight. For example, a factor directly related to ocean currents is that of water temperatures. King (1964) has noted a connection between pinniped distribution and water temperatures, and thus also ocean currents. It is to be expected that pinnipeds undertaking long migrations would follow prevailing ocean currents, not only because these would facilitate long-term directional movements, but also because water temperatures remain more equable within a single current system. Temperature is important in determining the ecosystem of any oceanic environment, and since pinnipeds are simply one element of such a system, their dispersal in the past is likely to have been influenced by sea temperature and ocean current patterns.

Pliocene to Recent records of monk seals in the Atlantic Ocean are as follows:
M. monachus and its ancestors-Europe and north-west Africa
M. tropicalis-Caribbean
$P$. rovereti-Argentina
P. capensis-South Africa

With Europe and north-west Africa taken as the centre from which the Monachinae dispersed, the other records can be accounted for in a number of ways.

However, bearing in mind the factors already mentioned, the pattern of dispersal outlined below is considered the most probable of the alternatives.

While east to west crossings of the Atlantic could conceivably have taken place in both hemispheres, it is simpler to suppose that some time in the late Miocene or early Pliocene a single such crossing took place, starting from the
north-west coast of Africa to the most easterly parts of South America. Even today M. monachus occurs as far south as Cape Blanc in Mauritania, and historic records indicate its presence even further south in Senegal (Van Wijngaarden 1962). Assuming a similar distribution for an early ancestor, an east to west crossing following the prevailing ocean currents by way of the Canary Islands at the shortest distance between Africa and South America would not have been difficult to achieve.

Once established in South America, these early monachines could have spread both north and south, the former to give rise eventually to M. tropicalis in the Caribbean, and the latter to give rise to $P$. rovereti in Argentina. All the monachines involved in these movements would have been species adapted to warm water conditions.

An early monachine population adapting itself to colder waters and moving into the far south of South America would then have been ideally situated for further dispersal into Antarctic regions, eventually to give rise to modern Antarctic monachines. This same group could also have undertaken another Atlantic crossing, this time from west to east, by way of islands near the Antarctic convergence and the prevailing ocean currents of the southern mid-latitudes, to South Africa. The presence of $P$. capensis at Langebaanweg is thus accounted for, and while Europe is regarded as the centre from which all later Monachinae were dispersed, the east coast of South America is here regarded as the centre from which the southern monachines arose (Fig. I).

Palaeontological evidence relating to the date of arrival of the ancestors of M. tropicalis in the Caribbean is inconclusive. While there are Pleistocene records of M. tropicalis in Florida and South Carolina (King 1964), early forms of this species must have been present in this area before the Pleistocene. This is a statement which is also based on inference rather than direct evidence.

Since the two records of Prionodelphis both date from the Pliocene, and only one east to west crossing of the Atlantic has been indicated, it suggests that ancestors of M. tropicalis were present in the Caribbean during the Pliocene as well.

There is further indirect evidence to substantiate this statement. It is well known that there is greater morphological correspondence between $M$. tropicalis and $M$. schauinslandi than exists between either of these species and M. monachus, which indicates that these two species have had a common ancestor more recently than the genus as a whole. In order that this common ancestor could give rise to the two modern species, it must have been present in both the Atlantic and Pacific Oceans.

Bearing in mind the existing distribution of M. tropicalis and M. schauinslandi, the most probable Atlantic-Pacific link penetrated by the common ancestor, was that which existed before the North and South American continents were linked by the isthmus of Panama. Once established on the Pacific coast of Central America, the dispersal of ancestors of $M$. schauinslandi to islands in the central Pacific would have been facilitated by the prevailing currents in the north equatorial region of this ocean (Fig. I).

Fig. I. Suggested dispersal routes of monachine seals.
Light arrows-Some of the principal surface currents of the oceans. 3. Spread of ancestors of Monachus schauinslandi to central Pacific. Southern Hemisphere dispersal:
I. Crossing to east coast of South America in late Miocene or early
2. Arrival of ancestors of modern southern monachines in Antarctic
egions, probably during the Plioce.
3. Crossing of South Atlantic of Prionodelphis during the Procene. America, probably during the late Pliocene.

The last time a crossing of the Central American region by pinnipeds was possible is not known for certain. Olsson (1932) has stated that the first evidence for a separation between marine invertebrates of northern Peru and the Caribbean was in the late Miocene. This suggests that the common ancestor of $M$. tropicalis and $M$. schauinslandi must have crossed somewhere near the present isthmus of Panama by this time. However, Simpson (1950) has referred to the Pliocene mammals migrating between the two New World continents as 'island hoppers', and indicates that the final North and South American land connection was achieved in the late Pliocene, a view supported by Whitmore \& Stewart (1965). A Pliocene Atlantic to Pacific crossing by early monachines would seem more reasonable in view of the late Miocene date for the earliest monachine in Europe. Even though the movement of marine invertebrates between the Atlantic and Pacific may not have been possible since the late Miocene, the movement of pinnipeds across the area between Central and South America during the Pliocene is probable. It is possibly the imprecise relative dating of the fossils and deposits referred to in the preceding discussion that causes the discrepancy in the evidence provided by invertebrate and vertebrate fossils. Gabunia \& Rubinstein (1968) give an indication of the difficulties encountered in the relative dating of deposits at the period of time which is critical in the present instance.

The preceding interpretation of the evolution and dispersal of the monk seals can be summed up as follows:
I. Monachus monachus evolved from a late Miocene monachine, largely in the area in which it is found today.
2. In the late Miocene or early Pliocene an early monachine crossed the Atlantic from the west coast of Africa to the east coast of South America.
3. One group of these immigrants moved southwards along the east coast of South America and gave rise to Prionodelphis rovereti.
4. This southward movement of monachines continued into Antarctic regions, while at least one population crossed the South Atlantic to South Africa and gave rise to Prionodelphis capensis.
5. A second group of the original immigrants to South America moved northwards into the Caribbean, and also across into the Pacific before the final linking of the North and South American continents. They eventually gave rise to Monachus tropicalis and M. schauinslandi, ancestors of the latter having moved from the Pacific coast of Central America to the central Pacific.

The monk seals are a remarkably conservative group. Although the three modern species have had a history independent of one another, perhaps dating back to the Miocene, and certainly dating back to the Pliocene, they are still morphologically very similar to one another. It seems that having once adapted to their environment they received little, if any, pressure to cause adaptive change. This conservative, low latitude group survives today, effectively isolated from other pinnipeds.

The Antarctic Monachines<br>(Excluding the southern elephant seal)

Before dealing with the evolution and dispersal of Antarctic monachines, it is necessary to consider in detail their possible phyletic relationships to the two species of Prionodelphis, based on considerations of comparative morphology.

A detailed study of the known remains of Prionodelphis capensis revealed that there is an undoubted connection between this species and the Antarctic monachines in general (Hendey \& Repenning 1972). However, it was also clear that no definite phyletic connection between the South African species and any one of the modern Antarctic seals could be demonstrated. In some respects $P$. capensis is morphologically intermediate between the genus Monachus and the Antarctic seals. Such features of $P$. capensis as the relatively narrow cheekteeth, the flat and broad petrosal apex of the tympanic, and the relatively stout humerus and spatulate radius, suggest a stage of development from the ancestral monachine condition (exhibited by the conservative Monachus) towards Hydrurga, Lobodon, Leptonychotes and Ommatophoca.

The highly modified dentitions of the Antarctic monachines are the features which differentiate them from $P$. capensis most strikingly. On the basis of their dentitions it is improbable that either Hydrurga or Lobodon could be derived from a $P$. capensis-like ancestor, since this would have necessitated the reversal of at least one evolutionary trend.

Both species of Prionodelphis have a reduced upper fifth postcanine, with only the vestiges of a single anterior accessory cusp remaining, this apparently having derived from the basic three-cusped pattern still evident in Monachus. Both Hydrurga and Lobodon have little reduction of this tooth, which in both cases has well-developed accessory cusps anteriorly and posteriorly. While it is not impossible that lost features can be redeveloped (Kurtén 1963), this is not usual, and the possibility of a phyletic connection between Prionodelphis and both Hydrurga and Lobodon becomes more improbable.

The multiplication of accessory cusps in Lobodon is yet another complicating factor. The lower postcanines of $P$. rovereti have two anterior and two posterior accessory cusps, but even this does not match the proliferation of posterior cusps in the second to fifth postcanines of Lobodon. Cusp development in this genus is variable, and additional accessory cusps appear to develop as buds from the principal cusp and as projections from the cingulum. There is no evidence to suggest that the archetype monachine cheektooth had an array of cusps such as is seen in Lobodon. Consequently, although deviation from the basic three-cusped pattern may not have been difficult to achieve, in the case of the Prionodelphis upper fifth postcanine it would require an even more complex reversal of the trend already mentioned.

In addition, Lobodon differs from other Antarctic monachines in having teeth which are relatively broad. A comparison of the cheekteeth of $P$. rovereti, P. capensis and Hydrurga shows a progressive narrowing of the teeth (Hendey
\& Repenning 1972: Table 1), and a similar transverse compression is also evident in the teeth of Leptonychotes and Ommatophoca. The teeth of Lobodon are more like the ancestral monachine condition in this respect.

Kellogg (1942: 453) has already suggested that $P$. rovereti might be ancestral to Leptonychotes. This suggestion is not affected by the one that $P$. capensis is a derivative of the Argentinian species. The phyletic connection between the two species of Prionodelphis is indicated partly by the inferred manner of dispersal of southern monachines, the earlier suggestion that in some respects P. capensis has more advanced characteristics (Hendey \& Repenning 1972: 93), and that the Entre Rios deposits are early Pliocene (Romer 1966), while those at Langebaanweg are late Pliocene (Hendey 1970).

Prionodelphis capensis could be regarded as intermediate between $P$. rovereti and Leptonychotes in view of the reduction in the size of the posterior root in its upper postcanines, the reduction in the number of cusps in its lower postcanines, and in the greater transverse compression of its cheekteeth. Much the same can be said in the case of Ommatophoca, but in this instance the differences between the cheekteeth of $P$. capensis and $O$. rossi are far greater, and changes would have needed to be at a greatly accelerated rate. Perhaps the feature most suggestive of a connection between Prionodelphis and Leptonychotes is the recurved crown and virtual lack of accessory cusps in the upper fifth postcanine of the former genus, characteristics which are matched in most of the cheekteeth of Leptonychotes. To a lesser extent these characteristics are evident also in the cheekteeth of Ommatophoca.

Thus on the basis of teeth alone, it appears that Prionodelphis is more likely to be ancestral to Leptonychotes and Ommatophoca, although $P$. capensis itself is probably not on a direct line to these species, but merely paralleled the trends which led to them. On the other hand Hydrurga and Lobodon are likely to be derived from an even more generalized monachine-a proto-Prionodelphis.

Looking once more at the distribution of the southern monachines, it is clear that, other factors aside, $P$. capensis was not geographically well situated to give rise to the Antarctic monachines, and Leptonychotes and Ommatophoca in particular.

In view of the suggestion that Hydrurga and Lobodon arose from a 'protoPrionodelphis', it is possible that early representatives of these genera reached Antarctica before the ancestors of Leptonychotes and Ommatophoca. Since the latter two genera have a more southerly distribution than Hydrurga and Lobodon (King 1964), it is likely that their ancestors entered Antarctica from the southern tip of South America $\left(55^{\circ} \mathrm{S}\right)$ via the South Shetland Islands to the Palmer Peninsula of Antarctica itself. The most likely route into Antarctica from South Africa ( $34^{\circ} \mathrm{S}$ ) would, because of prevailing ocean currents, have been by way of the south Indian Ocean, via islands near the Antarctic convergence and the pack ice to the Antarctic continent. In addition to being a more difficult route, it would have necessitated the crossing of an area (the outer fringes of the pack ice) which may already have been inhabited by the ancestors of Hydrurga and

Lobodon. While the four modern Antarctic species do not compete for food, this must become progressively less so further back in time when their respective ancestors were more generalized.

A final point concerning the possibility of a phyletic connection between $P$. capensis and one or more of the modern Antarctic monachines, concerns the age of $P$. capensis. Since $P$. capensis is considered to date back 4-5 million years (Hendey 1970), it would be required that the marked morphological differences between it and, say, Leptonychotes be developed in this period of time. If the early Pliocene date for $P$. rovereti is correct, the period of time for similar changes to occur might be two or three times as long. In itself this is meaningless, but clearly the greater the time involved, the greater the possibilities for radical morphological change.

In the absence of convincing evidence to the contrary, it is probable that differentiation and evolution of the Antarctic monachines took place in the high latitudes in which they occur today. The more extreme climate and climatic changes in high latitudes during the Pliocene and Pleistocene may well have provided the mechanism for accelerated changes in the Antarctic monachines, the nature of the environment acting as a stimulant to adaptive change. Owing to the lack of deposits of Pliocene and Pleistocene age in Antarctica, except for those beneath the Antarctic Ocean, it is very improbable that the nature of the development of Antarctic seals will ever be known from fossil remains. Thus with this group of pinnipeds, interpretations of their evolutionary history will probably remain speculative.

It is clear that there was a far more rapid evolution of specialized characters in the southern monachines than was the case with the monk seals, even though the independent history of the four southern species was probably no longer than that of the three monk seals. Thus the Antarctic seals contrast with the monk seals by being a high latitude and highly specialized group, as opposed to a low latitude and conservative group.

## The Elephant Seals

The genus Mirounga has long been the subject of controversy as to its relationships. It is now included in the Monachinae (King 1966) but anatomically and in its distribution it does still to some extent stand apart from other monachines.

While some conclusions on relationships can be drawn from comparisons between the cheekteeth of other monachines, the peg-like teeth of Mirounga are singularly uninformative. Burns \& Fay (1970:389), however, concluded that, 'craniologically Mirounga is more like other phocids of the Southern Hemisphere than is Monachus'. The mere fact that Mirounga is a highly specialized genus makes it probable that it would have closer connections to the diversified Antarctic monachines, rather than with the conservative monk seals. In other words, it is suggested that adaptive radiation in the Monachinae is confined to the southern high latitude group, to which Mirounga must therefore belong.

Furthermore, the distribution of the two modern species is more readily accounted for in assuming a southern origin for the genus. This is already suggested by the fact that $M$. angustirostris maintains a pattern of breeding (in the northern winter) which is typical of Southern Hemisphere phocids (King 1964).

The present distribution of the southern elephant seal includes the extreme southern end of South America, but within historic times it has been reported as far north as the Juan Fernandez Islands, which is in fact the type locality. A wider spread along the west coast of South America during the past, in a manner comparable to that of modern Arctocephalus australis, is conceivable owing to prevailing ocean currents and water temperatures of that coast. This would have brought the southern and northern ranges into far closer proximity, and indeed have allowed the southern species, or an ancestor, to reach the area in which the northern species occurs today. A split in this distribution, centred in low latitudes, of a single population of Mirounga, and a gradual retreat in its range in the south, would then account for the present pattern of elephant seal distribution. Mirounga angustirostris can thus be regarded as a relict species, surviving in isolation far from the origins of the genus.

Since there is an early Pleistocene record of Mirounga in California (C. A. Repenning, pers. comm.), it is probable that the genus arose during the Pliocene, and the present pattern of elephant seal distribution developed during the Pleistocene.

The suggested phyletic relationships of modern and some fossil Monachinae are illustrated in Figure 2.

## Discussion

The present pattern of monachine distribution is perhaps the most unusual of any of the pinniped subfamilies. The Otariinae and Arctocephalinae, which are taken by some to be a single subfamily (Mitchell 1968), have essentially similar distributions, with the eastern Pacific shorelines providing the closest links between the species of the two hemispheres. The Odobeninae and Phocinae are purely Northern Hemisphere groups, largely confined to high latitudes. The Monachinae alone can be subdivided into two geographically widely separated groups, while a third subdivision includes two equally widely separated species of a single genus.

King (1964) has discussed pinniped distributions as they relate to ocean currents and water temperatures, and has remarked upon the correlation with colder waters. The monk seals are one of the few exceptions to the general rule that, 'the $20^{\circ} \mathrm{C}$ summer isotherm in either hemisphere, where it approaches continental coasts forms a reasonable pointer to the limits of where one might expect to find seals' (King 1964:89). The critical temperature for the monk seals is higher. By contrast the Antarctic seals are limited to waters with a maximum temperature of $3-4^{\circ} \mathrm{C}$. The southern elephant seal also occurs in


the cold waters of the far south, while its northern relative conforms to the general rule quoted above.

It is reasonable to suppose that at one time the monachines had a less interrupted distribution, and the records of Prionodelphis give an indication that this was indeed the case. In order to explain the curious pattern of modern distributions, it is necessary to refer to the occurrence of other pinnipeds. It is found that the southern fur seals (genus Arctocephalus) and southern sea lion (genus Otaria) inhabit much of the intervening area which is suitable for occupation by seals. Bearing in mind the suggested manner of dispersal of early monachines, it is probable that much of their range during the Pliocene, and perhaps part of the Pleistocene as well, included the areas in which at least some of the species of Arctocephalus and also Otaria occur today. The indications are, therefore, that there has been replacement of monachines by otariids, especially in the Southern Hemisphere.

The earliest Southern Hemisphere records of the Otariidae are Otaria fischeri Gervais \& Ameghino 1880 from Argentina, a Phocarctos hookeri from New Zealand (Berry \& King 1970), and 'Arctocephalus' williamsi from Australia (King 1964). The two Australasian species are Pliocene in age, and while King (ig64) listed the Argentinian record as '? Miocene', it too probably dates from the Pliocene (Davies 1958). Excluding Pleistocene records, other recorded fossil otariids are from either the west coast of North America or Japan. Thus it is probable that while the Phocidae had the Atlantic Ocean as the centre for their evolution, the early history of the Otariidae was confined to the Pacific. The indications are, therefore, that the Otariidae only reached southern Africa after they had become established in Australasia and South America, i.e. the two southern continents bounding the Pacific Ocean. It must have been very late in the Pliocene, or perhaps only in the Pleistocene that the otariids became established in southern Africa. Although the fossil record is not conclusive on this point, it does seem likely that the extinction of Prionodelphis in South America and South Africa was more or less synchronous with the advent of otariids in these regions.

The present distribution of the two species of Mirounga discussed earlier, may at least be partly determined by the presence of Arctocephalus australis populations on the South American coast. King (1964) states that even though M. leorina and $A$. australis have a common diet they appear to co-exist without difficulty, but she also notes that when breeding season conflicts occur 'the fur seal usually wins' (King 1964: 25). The fact that M. leonina distribution has become considerably less in recent times suggests that the sympatric existence of these two species may be in a state of delicate balance, and, if upset, $A$. australis is in a position of dominance.

Unsuccessful competition by monachines with otariids cannot be proven, but the present distribution of these two groups suggests that there is a mutually exclusive inter-relationship between them.

The possible influence of climatic changes on monachine distribution
cannot, of course, be ignored. For example, $M$. leonina may be more sensitive to increasing temperatures than Arctocephalus, and the virtual disappearance of this species from the South African coast in the Holocene may be due to the general warming of conditions during this epoch. It is unlikely that prehistoric man played any significant role in the disappearance of the South African Mirounga, although this possibility cannot be entirely discounted. Late Pleistocene coastal hominid occupation sites are all below present sea-level, and the relative importance of Mirounga and Arctocephalus in diets of the hominids of the time is not known.

It is probable that the radiation of the otariids, coupled with changes in climate, jointly determined the nature of the present distribution of the Monachinae, which survive as isolated populations in low latitudes, and as firmly entrenched populations only in the southern high latitudes.

In reviewing the evolution of the Monachinae as outlined above, it is evident that this group does not conform exactly to some widely accepted principles. Crowson (1970: 133) lists certain 'rules' 'for determining the areas of origin of systematic groups from the patterns of distribution of their presentday members'. He states that it 'has been suggested that any group should be considered to have originated in that area where (a) it is represented by the greatest number of existing species . . or ( $d$ ) its most primitive living forms occur'. Crowson's conclusion that these 'rules' can be misleading is substantiated by the Monachinae, since the greatest number of living species (the Antarctic seals) are far removed from the most primitive living forms (Monachus). In this case 'rule' ( $d$ ) applies.

## Conclusion

There are undoubtedly alternatives to the speculations outlined in this report. However, since the facts which form its basis are so limited, their interpretation must to a large extent be subjective. This is inevitable when dealing with a group of mammals with a poor fossil record which is supplemented only at very infrequent intervals. The only two Southern Hemisphere records which throw any light on the early history of the southern Monachinae are Prionodelphis rovereti from Argentina and P. capensis from South Africa, the descriptions of which were published nearly 50 years apart.

Given such a situation it seems preferable to speculate now, rather than wait for further significant discoveries to be made.

If this paper serves to stimulate better reasoned interpretations of the limited fossil record of the Monachinae, it will have served a purpose. Whatever is finally accepted, it is clear that a reassessment of the taxonomy of the monk seals and all fossil Monachinae is necessary.

## Summary

The fossil record of the Monachinae is outlined, and it is concluded that this pinniped subfamily arose in the western Europe-Mediterranean area. It
is concluded that the monk seals, and the early ancestors of the Monachinae as a whole, are an essentially low-latitude and conservative group, from which arose the high-latitude and specialized 'southern' monachines, which include Mirounga angustirostris. It is suggested that the latter group underwent their differentiation in the high latitudes in which they are most commonly found today, having arisen from Prionodelphis and 'proto-Prionodelphis' populations which spread into Antarctic regions from South America. It is suggested that the decline of the Monachinae in southern mid-latitudes was due at least partly to the rise of the Otariidae in these regions.

## Acknowledgements

The present study was a direct outcome of an investigation of the Prionodelphis capensis fossils from Langebaanweg, which was undertaken jointly with Charles A. Repenning of the Pacific Coast Center of the U.S. Geological Survey. I am greatly indebted to him for the stimulation and assistance he provided in both these studies. The opinions expressed in the present paper do not necessarily reflect those of Mr. Repenning.

The current investigations at Langebaanweg are being supported by the South African Council for Scientific and Industrial Research, Chemfos Ltd. (a subsidiary of the African Metals Corporation) and Shell South Africa (Pty.) Ltd. The Wenner-Gren Foundation for Anthropological Research, New York, provided the vehicle used in the field work at Langebaanweg (Grant no. 2752-1834).

## References

Berry, J. A. \& King, J. E. 1970. The identity of the Pliocene seal from Cape Kidnappers, New Zealand, previously known as Arctocephalus caninus. Tuatara 18: 13-18.
Burns, J. J. \& Fay, F. H. 1970. Comparative morphology of the skull of the Ribbon seal, Histriophoca fasciata, with remarks on the systematics of the Phocidae. 7. Zool. 161: 363-394-
Crane, H. R. \& Griffen, J. B. ig68. University of Michigan radiocarbon dates XII. Radiocarbon 10: 61-114.
Crowson, R. A. 1970. Classification and biology. London: Heinemann.
Davies, J. L. 1958. The Pinnipedia: an essay in zoogeography. Geogr. Rev. 48: 474-493.
Frenguelli, J. 1922. Prionodelphis rovereti un representante de la familia 'Squalodontidae' en al Paranense Superior de Entre Rios. Boln Acad. nac. Cienc. Cordoba 25: 491-500.
Gabunia, L. \& Rubinstein, M. i968. On the correlation of the Cenozoic deposits of Eurasia and North America based on the fossil mammals and absolute age data. i3th Int. geol. Congr. 10: 9-17.
Gervais, H. \& Ameghino, F. 1880. Les mammifêres fossiles de l'Amerique du Sud. Buenos Aires; Paris: Savy.
Gervais, P. \& Serres, M. de. 1847. Nouvelles observations sur les mammifères dont on a trouvé les restes fossiles dans les sables marins de Montpellier. Annls Sci. nat. (3) 8: 224-226.
Hendey, Q. B. 1970. The age of the fossiliferous deposits at Langebaanweg, Cape Province. Ann. S. Afr. Mus. 56: 119-131.
Hendey, Q. B. \& Repenning, C. A. 1972. A Pliocene phocid from South Africa. Ann. S. Afr. Mus. 59: 71-98.
Kellogg, R. 1922. Pinnipeds from Miocene and Pleistocene deposits of California. Univ. Calif. Publs geol. Sci. 13: 23-132.

Kellogg, R. 1942. Tertiary, Quaternary, and Recent marine mammals of South America and the West Indies. Proc. 8th Am. Sci. Congr. (Washington) 3: 445-473.
King, J. E. 1964. Seals of the world. London: British Museum (Natural History).
King, J. E. 1966. Relationships of the Hooded and Elephant seals (genera Cystophora and Mirounga). 7. Zool. 148: 385-398.
Kurten, B. 1963. Return of a lost structure in the evolution of the felid dentition. Commentat. biol. 26: 1-12.
McLaren, I. A. 196o. Are the Pinnipedia biphyletic? Syst. Zool. 9: 18-28.
Mitchell, E. 1968. The Mio-Pliocene pinniped Imagotaria. 7. Fish. Res. Bd Can. 25: 1843-1900. Nordmann, A. 186o. Palaeontologie Südrusslands. Helsingfors: Friis.
Olsson, A. A. 1932. Contributions to the Tertiary paleontology of northern Peru: Part 5, The Peruvian Miocene. Bull. Am. Paleont. 19: 1-272.
Romer, A. S. 1966. Vertebrate paleontology. Chicago: University Press.
Sarich, V. M. 1969. Pinniped phylogeny. Syst. Zool. 18: 416-422.
Simpson, G. G. 1950. History of the fauna of Latin America. Am. Scient. 38: 361-389. (Reprinted in Simpson, G. G. 1965. The geography of evolution; collected essays: 165-208. New York: Capricorn Books.)
Tavani, G. 1942. Revisione dei resti del pinnipede conservato nel Museo di Geologia di Pisa. Palaeontogr. ital. 40: 97-1 13.
Ugolini, R. 1902. Il Monachus albiventer Bodd. del Pliocene di Orciano. Palaeontogr. ital. 8: i-20.
Van Beneden, P. J. 1877. Description des ossements fossiles des environs d'Anvers. Annls Mus. r. Hist. nat. Belg. 1 : 1-88.
Van Wijngaarden, A. ig62. The Mediterranean monk seal. Oryx 6: 270-273.
Whitmore, F. C. \& Stewart, R. H. 1965. Miocene mammals and Central American seaways. Science 148: 180-185.

## INSTRUCTIONS TO AUTHORS

## Based on

CONFERENGE OF BIOLOGICAL EDITORS, COMMITTEE ON FORM AND STYLE. 1960.
Style manual for biological journals. Washington: American Institute of Biological Sciences.

## MANUSCRIPT

To be typewritten, double spaced, with good margins, arranged in the following order: (1) Heading, consisting of informative but brief title, name(s) of author(s), address(es) of author(s), number of illustrations (plates, figures, enumerated maps and tables) in the article. (2) Contents. (3) The main text, divided into principal divisions with major headings; subheadings to be used sparingly and enumeration of headings to be avoided. (4) Summary. (5) Acknowledgements. (6) References, as below. (7) Key to lettering of figures. (8) Explanation to plates.

## ILLUSTRATIONS

To be reducible to $12 \mathrm{~cm} \times 18 \mathrm{~cm}$ ( 19 cm including caption). A metric scale to appear with all photographs.

## REFERENCES

Harvard system (name and year) to be used: author's name and year of publication given in text; full references at the end of the article, arranged alphabetically by names, chronologically within each name, with suffixes $a, b$, etc. to the year for more than one paper by the same author in that year.
For books give title in italics, edition, volume number, place of publication, publisher.
For journal articles give title of article, title of journal in italics (abbreviated according to the World list of scientific periodicals. $4^{\text {th }}$ ed. London: Butterworths, 1963), series in parentheses, volume number, part number (only if independently paged) in parentheses, pagination.

## Examples (note capitalization and punctuation)

bullough, W. S. 196o. Practical invertebrate anatomy. 2nd ed. London: Macmillan.
Fischer, P.-H. 1948. Données sur la résistance et de le vitalité des mollusques. 7. Conch., Paris 88: 100-140.
Fischer, P.-H., Duval, M. \& Raffy, A. 1933. Etudes sur les échanges respiratoires des littorines. Archs Zool. exp. gén. 74: 627-634.
Kohn, A. J. ig6oa. Ecological notes on Conus (Mollusca: Gastropoda) in the Trincomalee region of Ceylon. Ann. Mag. nat. Hist. (13) 2: 309-320.
Kohn, A. J. 1960b. Spawning behaviour, egg masses and larval development in Conus from the Indian Ocean. Bull. Bingham oceanogr. Coll. 17 (4): 1-51.
Thiele, J. igio. Mollusca: B. Polyplacophora, Gastropoda marina, Bivalvia. In schultze. l, Zoologische und anthropologische Ergebnisse einer Forschungsreise im westlichen und zentralen SüdAfrika. 4: 269-270. Jena: Fischer. Denkschr. med.-naturw. Ges. Jena 16: 269-270.

## ZOOLOGICAL NOMENCLATURE

To be governed by the rulings of the latest International code of zoological nomenclature issued by the International Trust for Zoological Nomenclature (particularly articles 22 and 51). The Harvard system of reference to be used in the synonymy lists, with the full references incorporated in the list at the end of the article, and not given in contracted form in the synonymy list.

## Example

Scalaria coronata Lamarck, 18ı6: pl. 45 I , figs $5 a, b$; Liste: in. Turton, 1932: 80.

# ANNALS OF THE SOUTH AFRICAN MUSEUM ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM 

| Volume | $\mathbf{5 9}$ | Band |
| ---: | :---: | :---: |
| March | $\mathbf{1 9 7 2}$ | Mart |
| Part | $\mathbf{6}$ | Peel |



# A PLIOCENE URSID FROM SOUTH AFRICA 

By<br>Q. B. HENDEY

# The ANNALS OF THE SOUTH AFRICAN MUSEUM <br> are issued in parts at irregular intervals as material becomes available <br> Obtainable from the South African Museum, P.O. Box 6i, Cape Town <br> Die ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM <br> word uitgegee in dele op ongereelde tye na beskikbaarheid van stof <br> Verkrygbaar van die Suid-Afrikaanse Museum, Posbus 6I, Kaapstad <br> OUT OF PRINT/UIT DRUK <br> I, $2(\mathrm{I}, 3,5,7-8), 3(\mathrm{I}-2,5$, t.-p.i. $), 5 \mathrm{I}-2,5,7-9)$, 6(1, t.-p.i.), 7(1-3), 8, 9(1-2), $10(1)$, II(1-2, 5, 7, t.-p.i.), 21, 24(2), 27, 31(1-3), 33 

Price of this part/Prys van hierdie deel

$$
\mathrm{R}_{2,00}
$$

## Trustees of the South African Museum (C) Trustees van die Suid-Afrikaanse Museum 1972

ISBN 0949940062

Printed in South Africa by
The Rustica Press, Pty., Ltd, Court Road, Wynberg, Cape

In Suid-Afrika gedruk deur Die Rustica-pers, Edms., Bpk.
Courtweg, Wynberg, Kaap

# A PLIOCENE URSID FROM SOUTH AFRICA 

By
Q.B. Hendey
South African Museum, Cape Town
(With plates 19-20, 2 figs and 3 tables)
[MS. accepted 16 February 1972]
Contents


## Introduction

There are few mammal-bearing deposits of Pliocene age known in Africa (Kurtén 197I: 134) and consequently occurrences which date from this epoch are of significance in that they may produce records of species which are of special phylogenetic and zoogeographic interest. The only Pliocene occurrences presently being investigated in southern Africa are those at Langebaanweg in the Cape Province (Hendey 1970a, 1970b), and one of the more remarkable records from this locality is that of an agriotheriine ursid (Hendey 1969). It was the first record of an agriotheriine in Africa, and is still the only ursid known from sub-Saharan Africa.

The Agriotheriinae, which in the most restricted sense may be taken to include the genera Agriotherium and Indarctos, are known from a number of late Tertiary and early Pleistocene occurrences in Eurasia and North America, and the South African record adds a new dimension to concepts of the evolution and dispersal of this group. In general, the recorded species are represented by rather fragmentary material and, unfortunately, this is also the case with the Langebaanweg form. However, the significance of the material is not diminished, although the description which follows might well require revision if more specimens are recovered in the future.

The terrestrial mammal fauna with which the Langebaanweg agriotheriine is associated includes a number of species which are unexpected in an African context. For example, one of the hyaenids is referred to Percrocuta, a genus which is otherwise known in Africa only from the Algerian Miocene (Thenius

## II5

Ann. S. Afr. Mus. 59 (6), 1972: $115-132,2$ pls, 2 figs, 3 tables.
1966). It was, however, widely distributed in Eurasia during the late Tertiary (Kurtén 1957a). A second unusual species is an as yet unnamed boselaphine antelope, which apparently derives from the Miocene Protragocerus labidotus of Kenya (Gentry 1970). The boselaphines were also common in Eurasia during the late Tertiary.

Other significant, although not unexpected, records from Langebaanweg include an early ancestor of Hyaena hyaena, a primitive form of Mammuthus subplanifrons which is one of the earliest of the true elephants (Maglio \& Hendey 1970), and an ancestor of the white rhinoceros, Ceratotherium simum (Hooijer, in press).

The Langebaanweg deposits are unique amongst the major late Cenozoic fossiliferous occurrences of sub-Saharan Africa in that a marine fauna is associated with the terrestrial vertebrate fossils. Vertebrates linked with the marine environment include the first recorded fossil penguin from Africa (Simpson 1971) and an unusual monachine seal, Prionodelphis capensis (Hendey \& Repenning 1972), which has shed some light on the origins of the Antarctic seals (Hendey 1972).

Viewed in relation to the fauna as a whole, the agriotheriine is but one of a series of important additions to the fossil record of the late Cenozoic of Africa.

In order that this species might be placed in taxonomic perspective, its description is preceded by a brief review of the Ursidae.

## The Family Ursidae

The Ursidae are a comparatively recently evolved mammalian group, with a relatively small number of constituent genera, and they have received a considerable amount of attention from palaeontologists and neontologists alike. Despite this they have proved an extremely controversial group and it is only recently that a measure of agreement has been reached on their phyletic and intra-familial relationships.

The most comprehensive account of the Ursidae is that of Erdbrink (1953), and this has provided an invaluable basis for more recent work on the family. Since the appearance of Erdbrink's monograph much attention has been focused upon the ursine bears (the genus Ursus and closely related forms). Their phylogeny is now one of the best known of all mammalian groups and this successful study has been largely due to the work of Kurtén (1957b, 1958, 1964, etc.) and Thenius (r959a). Much the same can be said of the tremarctine bears (the genus Tremarctos and its close relatives) (Kurtén 1966, 1967), although the origin of this group remains obscure.

The taxonomic position of the giant panda, Ailuropoda melanoleuca, has been a matter of controversy for more than a century owing to its combination of ursid and procyonid characteristics. Davis (1964) presented what is perhaps the definitive study of the anatomy of this animal, and he held the view that it is indeed a bear. While this conclusion is still not universally accepted, Ailuropoda is regarded as an ursid in the present study.

The Tertiary ursids, excluding the immediate ancestors of the ursine group, can conveniently be placed in two categories. The first includes those genera which bridge the evolutionary gap between the Canidae and 'true' bears. They are Cephalogale, Hemicyon and Dinocyon. Secondly, there is a more advanced group in which many specialized ursid characteristics were developed. This group includes Agriotherium and Indarctos. Not surprisingly these two groups may be broadly differentiated on temporal as well as morphological grounds, the former being essentially Miocene in age, and the latter dating largely from the Pliocene. The Tertiary ursids are not as well known as the Quaternary forms because they are less well represented in the fossil record. There are also many uncertainties regarding the relative ages of some members of the groups.

Frick (1926) separated these Tertiary forms from both the Canidae and Ursidae, placing them in a single unit which he called 'Hemicyoninae'. Subsequently Pilgrim (1932) referred them all to the Ursidae and this arrangement is generally followed today.

There have been many differences of opinion concerning formal subdivisions within the family. For example, Kraglievich (1926) proposed separation into three subfamilies, which Simpson (1945: 225) argued was 'of very doubtful theoretical validity and of little or no practical convenience'. Indeed, he even appeared to be in some doubt as to whether the bears merited family rank. Erdbrink (1953) was another not in favour of having subfamilies within the Ursidae. Today it has become fairly widely accepted that subfamilial grouping is both reasonable and desirable and the arrangement usually followed is that of Thenius (1959a), who recognized the Hemicyoninae, Agriotheriinae, Tremarctinae and Ursinae.

In spite of the formal system of zoological nomenclature, there is a strong element of personal opinion in the definition of many taxa. The inclusion or exclusion of the Hemicyoninae from the Ursidae appears to depend largely upon whether a 'vertical' or 'horizontal' system of classification (Simpson 1945) is favoured. An excellent example of this basic difference in approach is afforded by the controversy which has surrounded the taxonomy of early Pleistocene Hominidae. Reed (1967) has given a concise summary of the two points of view, and states that 'as gaps in the fossil record . . . have been filled, the tendency has been . . . to shift from a horizontal (grade) type of classification to a vertical (clade) type'.

While the general pattern of ursid evolution has long been appreciated, the intra-familial groupings have tended to emphasize morphological rather than phylogenetic aspects of the family. An attempt has been made here to classify the ursids according to their phylogeny, although this was hampered by the fact that there are still certain critical points in ursid evolution which are not satisfactorily resolved. The intra-familial classification to be given presently follows that of Thenius (1959a), but is modified on the basis of the tentative phylogeny presented in Figure 1. This phylogeny is based on the work of


Fig. r. Tentative phylogeny of the Ursidae.

Erdbrink, Kurtén and Thenius, and the morphological characters, temporal range and geographical distribution of each genus was taken into consideration.

The pattern which emerges is one of three distinct radiations within the family, each'successive radiation cutting out the one preceding temporally and/or geographically. In each case the stem genus is recorded in Europe, and dispersals were largely confined to the northern continents. The intrafamilial classification arrived at is as follows:

| Subfamily | Tribe |  |  | Genera included |
| :--- | :--- | :--- | :--- | :--- |
| Hemicyoninae | . | Hemicyonini | . | Cephalogale, Hemicyon, Dinocyon |
| Agriotheriinae | . | Agriotheriini | . | Ursavus, Indarctos, Agriotherium |
|  |  | Ailuropodini | .. | Ailuropoda |
|  |  |  | Tremarctini | . . | Arctodus, Tremarctos

The replacement of the Hemicyoninae by the Agriotheriinae, and the latter in turn by the Ursinae, can be accounted for in general by assuming that there was competition between better and lesser adapted groups. Such competition would, of course, have taken place at the species level, but would ultimately have been manifested in higher taxonomic categories as well.

Kurtén (1957c: 224) has concluded that the replacement of a species by an ecologically related species may occur in one of three ways:
(i) 'The extinction of the earlier form has no causal connection with the immigration of the later form. Both result from the action of other factors, for instance climatic.'
(2) 'The extinction of one species permits the subsequent immigration of another.'
(3) 'The immigrating form is adaptively superior to the local form, and ousts it through competition.'
While the third alternative is the one concluded to be the most generally applicable in the present instance, it was almost certainly not the sole factor involved in the extinction of ursids. Furthermore, competition was probably not confined only to members of the family, and some ursids probably became extinct as a result of competition with members of other families. This might well have been the case with the somewhat aberrant ursid Agriotherium.

## Intra-familial Categories Within The Ursidae

## Subfamily Hemicyoninae

Diagnosis (adapted from Pilgrim 193I). Ursidae with the upper profile of the skull almost rectilinear; snout relatively long and narrow; infra-orbital foramina rather remote from orbits; temporal fossae long and deep; occiput low; sagittal and lambdoidal crests prominent; zygomatic arches relatively narrow; $\mathrm{P}^{2}$ and $\mathrm{P}^{3}$ double-rooted; $\mathrm{P}^{4}$ situated behind infra-orbital foramen,
and antero-posterior diameter equal to or slightly exceeding that of $\mathrm{M}^{1} ; \mathrm{P}^{4}$ with prominent protocone situated towards the midpoint of the tooth, and parastyle absent; $\mathrm{M}^{1}$ larger than $\mathrm{M}^{2}$, with the transverse diameter of these teeth exceeding antero-posterior diameter, and with internal cusps crescentic inwards; $\mathrm{M}^{3}$ always absent; mandible with premasseteric fossa and full complement of teeth; $\mathrm{P}_{2}$ to $\mathrm{P}_{4}$ double-rooted; $\mathrm{M}_{1}$ large with talonid becoming prominent; $M_{2}$ smaller than $M_{1}$, double-rooted with antero-posterior diameter slightly greater than transverse diameter; $\mathrm{M}_{3}$ small, single-rooted and slightly elongated antero-posteriorly; postcranial skeleton a cursorial type, feet digitigrade; humerus with entepicondylar foramen.

Discussion. In general there was an increase in body size with time, and some later Hemicyoninae were as large as the biggest of modern bears. Although probably carnivorous on the whole, at least one genus (Cephalogale) apparently became progressively more omnivorous, and it is regarded as the ancestor of all ursids.

Early in their history the Hemicyoninae were confined to the Old World and only spread to North America at the peak of their radiation in the late Miocene. Although they were a restricted group generically, they were widespread and apparently very successful in the northern continents during the Miocene. They became extinct early in the Pliocene when the radiation of the Agriotheriinae was beginning.

## Subfamily Agriotheriinae <br> Tribe Agriotheriini

Diagnosis. Ursidae with the upper profile of the skull rather convex; snout fairly short and broad; infra-orbital foramina close to orbits; occiput moderately high; sagittal crest not prominent; zygomatic arches becoming broad; $\mathrm{P}^{4}$ situated below or slightly anterior to infra-orbital foramen, with anteroposterior diameter approximately equal to that of $\mathrm{M}^{1} ; \mathrm{P}^{4}$ protocone becoming progressively larger and sometimes with accessory cusps developed anterior to it; $\mathrm{P}^{4}$ parastyle develops and becomes progressively larger; $\mathrm{M}^{1}$ roughly square; $\mathrm{M}^{2}$ sometimes with talon; $\mathrm{M}^{3}$ always absent; mandible sometimes with premasseteric fossa; $\mathrm{M}_{1}$ usually large and sectorial; $\mathrm{M}_{2}$ elongated; $\mathrm{M}_{3}$ usually circular or very slightly elongated; postcranial skeleton progressively more heavily built; humerus (usually) with entepicondylar foramen.

## Tribe Ailuropodini

Diagnosis. Ursidae with upper profile of skull convex; snout short and broad; infra-orbital foramina close to orbits; sagittal crest not prominent; $\mathrm{P}^{1}$ vestigial or absent; $\mathrm{P}^{4}$ large with prominent protocone and antero-internal cusp (protocone lobe), and prominent parastyle; $\mathrm{P}^{4}$ situated below infraorbital foramen, with antero-posterior diameter slightly greater than that of $\mathrm{M}^{1} ; \mathrm{M}^{1}$ roughly square with prominent internal cingulum; $\mathrm{M}^{2}$ elongated with
prominent internal cingulum and talon on which multiple cusplets are developed; $\mathrm{M}^{3}$ always absent; mandible without premasseteric fossa; $\mathrm{P}_{1}$ vestigial; $M_{1}$ large, elongated and non-sectorial; $M_{2}$ large and slightly elongated with multiple cusplets on occlusal surface; postcranial skeleton robustly proportioned; non-cursorial; humerus with entepicondylar foramen.

## Tribe Tremarctini

Diagnosis. Ursidae with the upper profile of the skull rather convex; snout relatively short and broad; infra-orbital foramina close to orbits; sagittal crest sometimes prominent; zygomatic arches moderately broad; $\mathrm{P}^{4}$ situated anterior to infra-orbital foramen, with antero-posterior diameter usually less than that of $\mathrm{M}^{1} ; \mathrm{P}^{4}$ protocone prominent and parastyle absent; $\mathrm{M}^{1}$ roughly square or slightly elongated; $\mathrm{M}^{2}$ elongated with prominent talon; $\mathrm{M}^{3}$ always absent; mandible with premasseteric fossa; $\mathrm{M}_{1}$ large, elongated and non-sectorial; $\mathrm{M}_{2}$ elongated, antero-posterior diameter approximately equal to that of $\mathrm{M}_{1} ; \mathrm{M}_{3}$ moderately large and slightly elongated; postcranial skeleton robustly proportioned; non-cursorial; humerus with entepicondylar foramen.

Discussion. As here defined the Agriotheriinae are the most diverse of the ursid subfamilies. It includes the genus Ursavus which is the earliest member of the family which is unmistakably 'bear-like', and which is regarded as the stem genus of the ursids by those who include the Hemicyoninae in the Canidae (e.g. Kurtén 1966). It is known only in Europe.

An apparent off-shoot from Ursavus was Indarctos, a genus which is first recorded in the early Pliocene of Europe. It is uncertain which species of Indarctos is the earliest. One possibility is $I$. vireti Villalta \& Crusafont 1945 from Spain, while another is the agriotheriine from the lignites of Monte Bamboli in Italy (Erdbrink 1953). The transition from Ursavus to Indarctos appears to be principally a matter of an increase in size.

A number of other species of Indarctos have been recorded in the Pliocene of the northern continents. European species are I. arctoides Deperet 1895, I. atticus Dames 1883 and I. ponticus Kormos 1913, while I. lagrelli Zdansky 1924 is from China, I. punjabiensis Lydekker 1884 from India and I. oregonensis Merriam et al i9ı6 from North America. In general the recorded specimens of Indarctos are rather fragmentary, and consequently definitions of the species are often inadequate. Pilgrim (1931) noted the similarity between I. ponticus and I. lagrelli, and Kurtén (1957c) regarded them as conspecific. Probably the recovery of additional material and a review of the genus would result in further synonomies being recognized.

Much the same can be said of Agriotherium, except that this genus survived into the Villafranchian of Europe (Kurtén, r968), and as the new record from Langebaanweg shows (vide infra), it also became established in Africa. As far as could be determined there is no early Pliocene record of this genus. Species include A. insigne Gervais 1853 from Europe, A. maraghanus Mecquenen 1925
from Iran, A. palaeindicus Lydekker 1878 and A. sivalensis Falconer \& Cautley 1836 from India, and $A$. gregoryi Frick 1921 from North America.

Probably the best published account of the differences between Agriotherium and Indarctos is that of Pilgrim (1932). He was apparently the first person to conclude that Agriotherium is the more advanced of the two genera, although only in some respects and he stated that Indarctos was a development from Agriotherium. This is a traditional point of view which is still widely held, and Agriotherium has often been referred to as a link between the Canidae and Ursidae.

The present study led to the conclusion that the characteristics of Agriotherium are not primitive, but rather the result of the development of specializations. Agriotherium is regarded as a derivative of Indarctos in which there was a trend towards the development of more carnivorous habits. The alternative view that Indarctos was derived from Agriotherium is rendered a little unlikely by the known temporal ranges of the two genera, and the possibility that both are descended from a hypothetical common ancestor is an unnecessary theory.

The general trend in ursid evolution has been towards the development of characters suited to an omnivorous or herbivorous diet. There is at least one well-documented reversal of this trend. The polar bear, Ursus maritimus, is a purely carnivorous form which still retains many of the characters of $U$. arctos, the species from which it is derived (Kurtén 1964). This anomalous development may have come about during one of the Pleistocene glaciations when an $U$. arctos population adapted to life in a vegetation-less peri-glacial environment. Since the dichotomy of $U$. maritimus and $U$. arctos took place comparatively recently, the dentitions of the two species are still essentially similar. Given sufficient time the $U$. maritimus dentition would become increasingly modified, with the carnassials developing at the expense of the molars.

Agriotherium was probably just such an exception to the general rule in ursid evolution, but in this instance the carnivorous habits are reflected by the nature of the dentition. The numerous references to its 'primitive' and 'canidlike' characteristics imply 'carnivore-like', which is not necessarily primitive at all. In fact Erdbrink (1953:582) referred to the upper carnassial of Agriotherium as being 'very carnivorous in aspect'.

The most important characters which distinguish Agriotherium from Indarctos are to be found in the dentitions, and basically the differences are centred on the emphasis of the carnassials and the reduction of the other cheekteeth in Agriotherium (Pilgrim 1932: 42).

In Agriotherium the anterior premolars in both maxilla and mandible are reduced in size and sometimes number. This is not necessarily an indication of a carnivorous diet, since many ursines also have the anterior premolars reduced or lost, but in this group the molars are correspondingly enlarged and the reduction also affects the carnassials. This is not so in Agriotherium.

This genus differs from Indarctos in having the lingual margins of $\mathrm{M}^{1}$ and $\mathrm{M}^{2}$ shorter than buccal margins as a result of the paracones and metastyles
being more strongly developed than the protocones and hypocones. Since the buccal cusps are higher than lingual ones it is possible for them to act as shearing as well as crushing agents. The crushing function of the $\mathrm{M}^{2}$ of Indarctos is further indicated by the presence of a talon, and in this genus the $\mathrm{M}^{2}$ is always longer than $\mathrm{M}^{1}$. By contrast the $\mathrm{M}^{2}$ of Agriotherium is nearly always smaller than $\mathrm{M}^{1}$ and lacks the talon. Erdbrink's (1953:57I) conclusion that there is 'at best a beginning of a talon . . in A. insignis' is probably incorrect, and the talon in this species is regarded as vestigial.

The 'carnivorous aspect' of the upper carnassial of Agriotherium has already been mentioned, and this tooth is also the most important in so far as the inferences on the ancestry of the genus are concerned. It is characterized by the presence of a prominent parastyle, a cusp which is not found in any of the Canidae, Hemicyoninae or species of Ursavus. It is, however, present but small in some species of Indarctos (e.g. I. punjabiensis). This suggests a progressive development of a $\mathrm{P}^{4}$ parastyle in the Agriotheriinae as follows:

Ursavus (absent)-Indarctos (small)-Agriotherium (prominent).
In Ailuropoda, which is here regarded as another descendent of Indarctos, it is also prominent.

The lower carnassial of Agriotherium also exhibits 'carnivorous' characteristics. The talonid is reduced relative to the trigonid and the hypoconid is higher than the entoconid, which makes it a more efficient shearing tooth than that of Indarctos.

The reasons for the development of an apparently carnivorous lineage stemming from Indarctos are not known, but this might have been in response to competition with early Ursinae. By the late Pliocene when the ursine radiation was getting under way in Europe, Agriotherium was the only agriotheriine surviving in this area and it only became extinct in the early Pleistocene. It follows that if it was indeed a purely carnivorous form, its ultimate extinction in Europe and elsewhere cannot be explained in the same way as the extinction or limitation of other Agriotheriinae. In this instance competition with other fissiped carnivores may be the answer, although other undetermined factors might have been involved.

In the late Pliocene Indarctos was still present in Asia, an area in which the Ursinae had not yet become common, and it is only in Asia that Ailuropoda is recorded. Those authors who have accepted Ailuropoda as an ursid have invariably suggested its descent from the Agriotherium/Indarctos group (see Davis 1964), and Indarctos appears to be the only known fossil form from which Ailuropoda can be satisfactorily derived. Since Ailuropoda was well established early in the Pleistocene, and since it is a good deal more advanced than Indarctos, its differentiation must have taken place during the Pliocene, probably at about the time that Indarctos itself was nearing extinction.

Much of the controversy about the status of Ailuropoda seems to stem from the fact that it is almost always compared with modern ursine bears, from which it does indeed differ quite markedly. However, if it is taken into account
that their common ancestor was a Miocene form (Fig. I), the differences are hardly surprising. The differences between the European early Pleistocene Ursus minimus and Agriotherium insigne are as great, or even greater, than those between modern ursines and Ailuropoda, yet the referral of Agriotherium to the Ursidae is no longer questioned.

In order to illustrate that on dental evidence alone Indarctos could be ancestral to both Ailuropoda and Agriotherium, a list of some characters of the upper cheekteeth of these three genera is given in Table I. On the one hand the dentition is modified for a herbivorous diet (Ailuropoda), and on the other a carnivorous dentition is developed (Agriotherium).

Table r. Some characters of the upper dentitions of Ailuropoda, Indarctos and Agriotherium.

| Ailuropoda | Indarctos | Agriotherium |
| :---: | :---: | :---: |
| Herbivorous $\qquad$ lineage | Ancestral genus | $\longrightarrow$ Carnivorous lineage |
| Anterior premolars |  |  |
| $\mathrm{P}^{1}$ sometimes absent | $\mathrm{P}^{1}$ present | $\mathrm{P}^{1}$ present (?) |
| $\mathrm{P}^{\mathbf{2}} \& \mathrm{P}^{3}$ double-rooted | $\mathrm{P}^{2} \& \mathrm{P}^{3}$ double-rooted in early forms (?) | $\mathrm{P}^{2} \& \mathrm{P}^{3}$ single-rooted |
| Carnassial |  |  |
| $\mathrm{P}^{4}$ with prominent parastyle and antero-internal cusp | $\mathrm{P}^{4}$ parastyle absent or small, antero-internal cusp small | $\mathrm{P}^{4}$ with prominent parastyle, anterointernal cusp usually absent |
| Molars |  |  |
| $\mathrm{M}^{1}$ square with prominent lingual cingulum, four main cusps and smaller cusplets | $\mathrm{M}^{1}$ square, four main cusps only | $\mathrm{M}^{1}$ narrower lingually, four main cusps only |
| $\mathrm{M}^{2}$ elongated with prominent talon, four main cusps and many cusplets | $\mathrm{M}^{2}$ slightly elongated with small talon and four main cusps | $\mathrm{M}^{2}$ nearly square, talon vestigial or absent, usually four main cusps only |

The range of Ailuropoda diminished considerably during the Quaternary and it now survives in a natural state only in isolated areas in China. During the Pleistocene it was widely distributed in China, and is also recorded from Burma (Smith-Woodward 1915). It was during the Pleistocene that the Asiatic radiation of the Ursinae took place and this suggests that Ailuropoda may have been an unsuccessful competitor with this group. It is therefore another agriotheriine whose decline is attributed to the Ursinae.

The decision to include the tremarctines in the Agriotheriinae is not easily justified. Superficially at least, there are similarities between the extant Ailuropoda melanoleuca and Tremarctos ornatus, and both differ from Ursus. There are resemblances in general skull morphology, both being 'short-faced' forms, and Davis (1955: 29) states that, 'Except for the pre-masseteric fossa, the features that distinguish the skull of Tremarctos from the skull of Ursus, although much less exaggerated, are similar to the features that distinguish the skull of the
giant panda (Ailuropoda)'. Kurtén (1967) mentioned the similarity between the tremarctine Arctodus and Indarctos.

However, the teeth of the tremarctines are much closer to those of Ursus than any other agriotheriine. The $\mathrm{P}^{4}$ lacks a parastyle and the molars are similar to those of Ursus.

Its ancestry may lie with the agriotherine Ursavus, and the problematical (?) Ursavus pawniensis Frick 1926 from the North American Miocene may be the ancestral form. Erdbrink (1953) suggested that the tremarctines are not a homogeneous group, and he derived Tremarctos from ursine stock, but the 'arctotheres' (Arctodus) from Indarctos. However, Kurtén (ig66: 7) found that although the 'earlier history of Arctodus is poorly documented . . . there can be little doubt that it is a tremarctine'.

Another significant characteristic of tremarctines is the entepicondylar foramen of the humerus. This is a feature also present in the humerus of Indarctos (e.g. I. oregonensis), Ailuropoda and the Hemicyoninae from which the Agriotheriinae are derived. It is, however, not present in the humerus of the Ursinae. It is here regarded as a primitive characteristic retained in at least two agriotheriine lineages (Indarctos-Ailuropoda and ?Ursavus-Tremarctini), but lost in the Ursinae and perhaps also the Indarctos - Agriotherium lineage.

The conclusion reached here is that the tremarctines do belong in the Agriotheriinae, having stemmed from an Ursavus-like ancestor, and having paralleled the Ursinae in some respects.

As with Ailuropoda, the only surviving tremarctine, Tremarctos ornatus, occurs isolated from the Ursinae, in this instance in South America. Both Ailuropoda and the tremarctines co-existed with ursines for much of the Pleistocene, and in the case of the tremarctines for part of the Pliocene as well (Bjork 1970), so their inferred replacement by the ursines was a slow process. However, the fact remains that they were definitely in decline by the end of the Pleistocene, whereas the Ursinae were still remarkably successful. But for the advent of human civilization the Agriotheriinae might well have become extinct while the Ursinae might have remained a prominent part of the world's fauna.

## Subfamily Ursinae

## Diagnosis (see Pilgrim 1931).

Discussion. There is an extensive literature on modern and fossil ursids and a substantial proportion is devoted to the Ursinae. It is the best known and least controversial of the ursid subfamilies and only in the case of the sun bear, Helarctos, and the sloth bear, Melursus, are there any real doubts about ancestry. The subfamily apparently stems from the early Pliocene Protursus (Kurtén 1971), and the genera Ursus and Helarctos are first recorded in the late Pliocene, while Melursus is known only from the Quaternary.

Four categories may be distinguished within the subfamily. The first two are the Helarctos and Melursus groups, both of which are represented by a single
extant species, and in neither case is there a good fossil record. The genus Ursus can conveniently be divided into two groups. The first comprises $U$. americanus and $U$. thibetanus, the North American and Asiatic black bears, and the second is the brown bear group, $U$. arctos and related forms. The latter includes the polar and grizzly bears as well as a number of extinct species such as the giant $U$. spelaeus. They are an extremely successful group and at one time or another they have been distributed through much of the Northern Hemisphere, including the Arctic and North Africa.

## The Langebaanweg Bear <br> Agriotherium africanum n.sp.

Holotype. A left maxillary fragment with $\mathrm{P}^{4}$ (South African Museum No. L 2045).

Referred material. A part of an ulna (L 2154) and isolated teeth as follows:
L $1868 \mathrm{~A}-\mathrm{E}: \mathrm{I}_{3}, \mathrm{I}_{1}$, ? $\mathrm{P}^{3}$ and parts of $\mathrm{P}^{4}$ and $\mathrm{M}^{1}$.
L 12637: $\mathrm{M}^{2}$.
L i844 \& L 3I4I: $\mathrm{I}_{2}$ and $\mathrm{I}_{3}$.
L $1256 \mathrm{I}: \mathrm{M}_{3}$.
Locality. All the specimens are from 'E' Quarry, Langebaanweg.
Diagnosis. A species of Agriotherium of large size, in which the $\mathrm{P}^{4}$ has a prominent parastyle and a well-developed protocone lobe, the latter consisting of the protocone, an antero-internal cusp and a small intermediately situated cusp; the protocone lobe projects and is flattened posteriorly where it functions as a shearing surface additional to that of the paracone and metastyle. The $\mathbf{M}^{2}$ is smaller than $\mathrm{M}^{1}$ and is without a talon. The antero-buccal surface of $\mathrm{M}_{3}$ is inflated.

Etymology. The specific name is given in recognition of the fact that this is the first agriotheriine recorded from Africa.

## DESCRIPTION

Only two of the specimens, the ? $\mathrm{P}^{3}$ and $\mathrm{M}_{3}$ are complete in all respects, while the referred $\mathrm{P}^{4}, \mathrm{M}^{1}$ and $\mathrm{M}^{2}$ are so poorly preserved that not a single standard measurement could be taken on them. The latter are important, however, since they do give an indication of the morphology of the teeth concerned. In general the Agriotheriinae are not well represented in the fossil record, but the Langebaanweg species can be less adequately defined than most of the recorded species of the subfamily.

The ? $\mathrm{P}^{3}$ (L 1868C) (Plate 19 F, G) is referred to this species since it was found in association with the other L 1868 specimens, which unquestionably do belong to Agriotherium. In size ( $10,8 \times 7,8$ ) it is comparable to the $\mathrm{P}^{3}$ of an Indarctos atticus specimen described by Thenius (1959b), and it resembles this
tooth in being broadest anteriorly. It is a simple, low-crowned tooth with a barely perceptible principal cusp from which arise keels, one running posteriorly and the other antero-internally. The crown is supported by a single anteroposteriorly elongated root.

The carnassial fragment L 1868D is incomplete, but what remains matches corresponding parts of the holotype $\mathrm{P}^{4}$ (Plate $19 \mathrm{~A}-\mathrm{E}$ ). This tooth differs in some respects from those of previously described specimens of Agriotherium and Indarctos, although in size (Table 2) and general appearance it is similar to the $\mathrm{P}^{4}$ of these genera.

Table 2. Dimensions of the $\mathrm{P}^{4}$ of some species of Agriotherium

|  | A. insigne <br> France <br> (1) | A. sp. <br> Spain <br> (1) | A. palaeindicus <br> India <br> (2) | A. sivalensis <br> India <br> (2) |
| :--- | :---: | :---: | :---: | :---: | :---: |
| length . . . | $29, \mathrm{I}$ | 30,0 | 28,0 | 33,0 |
| breadth. | 21,0 | 23,0 | 21,0 | 19,8 |


|  | A. gregoryi <br> N. America <br> ( ) |  |  |  |  |
| :--- | :---: | :---: | :---: | :---: | :---: |
| length . | . | . | 23,3 | 35,4 | 36,5 |
| breadth | . | . | 21,7 | 25,8 | 25,0 |

(1) Frick (1926)
(2) Lydekker (1884)

The crown consists of a parastyle, which is damaged, paracone and metastyle, which are flanked lingually by a large protocone lobe made up of a protocone, antero-internal cusp and a small, intermediately situated cusp. There are two roots on the buccal side of the tooth and another supporting the protocone lobe. Although the parastyle is damaged, sufficient remains to indicate that it was large and made up about $25 \%$ of the total length of the tooth. In this respect it is typical of the $\mathrm{P}^{4}$ of Agriotherium in which the parastyle is always large, whereas in Indarctos it is usually not as well developed. The paracone and metastyle are approximately equal in length and make up the remaining $75 \%$ of the total length of the tooth. Shear facets have been worn on the lingual surfaces of these cusps.

The Langebaanweg $\mathrm{P}^{4}$ differs most markedly from previously described Agriotherium and Indarctos carnassials in the size and morphology of the protocone lobe. Its length ( $22,5 \mathrm{~mm}$ ) can be measured accurately since its anterior
and posterior limits are clearly defined. In most other Agriotheriinae this is not the case as the posterior limit of the protocone merges gradually with the lingual surface of the metastyle, but in any case they all have shorter protocone lobes. In addition, the protocone lobe of L 2045 differs from other species of Agriotherium in that it has a fairly prominent antero-internal cusp, although this cusp is present in Indarctos. It is small in I. punjabiensis,* but quite large in I. lagrelli (Zdansky 1924) and I. atticus (Thenius 1959b).

The protocone itself is unique in that instead of being conical, it has its apex elongated antero-posteriorly and compressed towards the paracone and metastyle. The elongation of the protocone lobe as a whole is largely due to the shape of the protocone. The functional advantage of this elongation is readily evident, since the posterior part of the protocone has developed on it a shear facet which is supplementary to that of the paracone and metastyle. Of all the Indarctos and Agriotherium upper carnassials presently known, that of the Langebaanweg species seems the best adapted to a shearing function.

Another unusual feature of the protocone lobe is the small cusp situated between the antero-internal cusp and the protocone. The cusp itself has been all but worn away, but its presence is marked by a circular patch of exposed dentine. It, and the most anterior part of the protocone have almost horizontal wear facets, indicating that the $\mathrm{P}^{4}$ served a crushing function as well.

Parts of the enamel of this tooth show the 'wrinkling' or rugosity said to be characteristic of Agriotherium (Erdbrink 1953).

In its general morphology the $\mathrm{P}^{4}$ of the Langebaanweg species is not dissimilar to that of Ailuropoda.

A small part of the maxilla of the holotype is preserved. The most anterior part of the alveolus of $\mathrm{M}^{1}$, and part of the antero-external root of this tooth are present. The $\mathrm{M}^{1}$ must have had a transverse diameter of at least 30 mm , which is in keeping with the size of this tooth in Agriotherium and Indarctos. The inferior margin of the infra-orbital foramen is also present and it is situated above and slightly posterior to the $\mathrm{P}^{4}$.

Little of the $M^{1}$ (L i868E) is preserved. Parts of the roots supporting the paracone and protocone are present, and that root beneath the protocone is large, antero-posteriorly elongated and inserted at an angle to the plane of the palate. The other preserved root is smaller, transversely elongated and inserted vertically into the maxilla. It presumably matched the now missing root which supported the metastyle. Most of the crown is lost and the only enamel preserved is near the paracone. Judging from the preserved parts of the crown and the roots, this tooth appears to have been narrower lingually. The transverse diameter is estimated to have been 30 mm , which is comparable to the figure inferred for the missing $\mathrm{M}^{1}$ of the holotype. The antero-posterior diameter must also have been about 30 mm .

[^1]The $\mathrm{M}^{2}$ ( L 12637) is an important specimen, since although it is incomplete, its morphology indicates that the affinities of the Langebaanweg agriotheriine lie with Agriotherium rather than Indarctos. It consists of a paracone and metastyle which are equal in size, situated parallel to a protocone and hypocone which are also similar in size. The latter cusps are lower than the paracone and metastyle. The enlarged and posteriorly elongated talon which characterizes the $\mathrm{M}^{2}$ of Indarctos is not in evidence. This tooth is appreciably smaller than the $\mathrm{M}^{1}$ and its dimensions are estimated to be $25 \times 25 \mathrm{~mm}$. It is thus smaller than the $\mathrm{M}^{2}$ of previously described species of Agriotherium (see Frick 1926: 81).

A reconstruction of the posterior upper dentition of the Langebaanweg agriotheriine is illustrated in Figure 2.


Fig. 2. A reconstruction of the posterior upper dentition of Agriotherium africanum based on the specimens L 1868C, L 2045, L 1868E and L 12637 (Natural size).

Little can be said of the lower incisors which are preserved (L i868A, B, L 1844, L 3141) (Plate 20 A ) other than that they are large and agree in all morphological respects with the corresponding teeth of Indarctos lagrelli (Zdansky 1924) and an Indarctos specimen from Samos (Helbing 1932). No descriptions or illustrations of the lower incisors of Agriotherium could be located, but presumably they are essentially the same as those of Indarctos.

The $\mathrm{M}_{3}\left(\mathrm{~L}_{12561}\right)$ (Plate 20 B ) is a single-rooted and low-crowned tooth with an almost circular and flat occlusal surface. The antero-buccal part of the crown is inflated and there is a wear facet in this region angled from the occlusal surface across the inflation towards the cingulum. This presumably results from occlusion with the lingual surface of the paracone or metastyle of $\mathrm{M}^{2}$. This is an indication that the post-carnassial teeth of this species functioned as shearing as well as crushing agents. The $M_{3}$ measures $16,5 \times 16,9 \mathrm{~mm}$.

The ulna (L 2154) (Plate $20 \mathrm{C}, \mathrm{D}$ ), which lacks the distal end, lower part of the shaft and anconeus process, is far too large to be confused with the ulna of any other carnivore species in the Langebaanweg assemblage. It compares closely in size (Table 3) and morphology with the ulna from Pikermi referred to Indarctos atticus by Pilgrim (1931).

Table 3. Dimensions of the Langebaanweg Agriotherium ulna, compared with that of an Indarctos (?) cf. atticus specimen from Pikermi (Pilgrim, 1931).

|  |  | L 2154. | Pikermi |
| :--- | :---: | :---: | :---: |
| Dorso-ventral diam. at coronoid process | . | 87,0 | 85,0 |
| Transverse diam. at coronoid process | . | 59,0 | 61,0 |
| Transverse diam. at proximal end | . | 50,0 | 52,0 |

DISGUSSION
The genera Agriotherium and Indarctos share many dental and osteological characteristics, but it is clear that the Langebaanweg agriotheriine has greater affinities to the former genus. It is regarded as a species distinct from those previously recorded since it exhibits certain apparently unique characteristics, and in addition it is the most geographically isolated record of the genus.

Agriotherium africanum differs from previously described species in the size of $\mathrm{M}^{2}$ and in the nature of its $\mathrm{P}^{4}$ and $\mathrm{M}_{3}$. All the species of Agriotherium are known from single, or perhaps a few individuals and it has therefore not been possible to assess the range of variation in any of them. However, R. H. Tedford (pers. comm.) has found 'considerable variation [in the $\mathrm{P}^{4}$ ] within and among populations of Hemphillian Agriotherium from the United States', although none of the North American specimens matched the $A$. africanum $\mathrm{P}^{4}$. If the Langebaanweg species is conspecific with a known species, then it is likely that it would be one of the Eurasian forms, which presumably also had variable upper carnassials.

In this connection the geographical location of $A$. africanum is probably significant. Of the 18 species of fissiped carnivores known from Langebaanweg, only four have affinities with contemporary Eurasian species. Much the same applies to the non-carnivorous mammals. Consequently it is probable that although there is a general similarity between the late Pliocene mammal faunas of Eurasia and Africa, each area was represented by its own lineages. For example, although the Langebaanweg Percrocuta is fairly similar to the Eurasian $P$. eximia, it is sufficiently different to warrant the status of a separate species. Similarly the boselaphine from Langebaanweg resembles Tragoportax salmontanus from the Siwaliks of India, but the two are clearly not conspecific.

Even if larger numbers of individuals of the Eurasian species of Agriotherium become available in the future, it seems unlikely material matching that from Langebaanweg will be recorded. It was on this basis that the decision was made to refer the Langebaanweg Agriotherium to a new species.

Probably it is just a matter of time before more agriotheriine remains are recovered elsewhere in Africa, especially in view of the attention presently being focused on Pliocene deposits in East Africa. It is also possible that more material of $A$. africanum will be found at Langebaanweg, since some of the deposits from which present specimens were derived remain unexcavated. Consequently more adequate definition of $A$. africanum might still be possible, and its phyletic relationships might yet be more accurately determined.

## Summary

An account of the family Ursidae (Mammalia: Carnivora) is given and a new ursid species, Agriotherium africanum, is described.

## Acknowledgements

The first draft of this manuscript was completed in 1967 and at that time I benefited greatly from correspondence with Dr. Björn Kurtén (University of Helsinki) and Dr. Richard H. Tedford (American Museum of Natural History). Both were very generous in sharing their knowledge of the Ursidae, and any merits which this paper may possess are due largely to them. Its shortcomings are, however, entirely of the author's own making.

The current investigations at Langebaanweg are being supported by the South African Council for Scientific and Industrial Research, Chemfos Ltd. (a subsidiary of the African Metals Corporation) and Shell South Africa (Pty.) Ltd. The Wenner-Gren Foundation for Anthropological Research, New York, provided the vehicle used in the field work at Langebaanweg (Grant no. 2752-1834).

I am indebted to the management of Chemfos Ltd, and also Mr. H. Krumm and Mr. G. Benfield for their unfailing assistance in the recovery of fossils from the quarries at Langebaanweg.

## References

Bjork, P. R. 1970. The Carnivora of the Hagerman Local Fauna (Late Pliocene) of southwestern Idaho. Trans. Am. phil. Soc. 60: 1-54.
Davis, D. D. 1955. Masticatory apparatus in the spectacled bear Tremarctos ornatus. Fieldiana, Zool. 37: 25-46.
Davis, D. D. 1964. The giant panda. A study of evolutionary mechanisms. Fieldiana, Zool. Mem. 3: 1-339.
Erdbrink, D. P. 1953. A review of fossil and recent bears of the Old World. Deventer: Jan de Lange.
Frick, C. 1926. The Hemicyoninae and an American Tertiary bear. Bull. Am. Mus. nat. Hist. 54: 1 -119.
Gentry, A. W. 1970. The Bovidae (Mammalia) of the Fort Ternan fossil fauna. Fossil vertebrates of Afr. 2: 243-323.
Helbing, H. 1932. Über einen Indarctos-schädel aus dem Pontien der Insel Samos. Abh. schweiz. paläont. Ges. 52: 1-18.
Hendey, Q. B. 1969. Quaternary vertebrate fossil sites in the south-western Cape Province. S. Afr. archaeol. Bull. 24: 96-105.

Hendey, Q.B. 1970a. A review of the geology and palaeontology of the Plio/Pleistocene deposits at Langebaanweg, Cape Province. Ann. S. Afr. Mus. 56: 75-117.
Hendey, Q. B. i970b. The age of the fossiliferous deposits at Langebaanweg, Cape Province. Ann. S. Afr. Mus. 56: 119-131.
Hendey, Q. B. 1972. The evolution and dispersal of the Monachinae (Mammalia: Pinnipedia). Ann. S. Afr. Mus. 59: 99-113.
Hendey, Q. B. \& Repenning, C. A. 1972. A Pliocene phocid from South Africa. Ann. S. Afr. Mus. 59: 71-98.
Hooijer, D. A. A late Pliocene rhinoceros from Langebaanweg, Cape Province. Ann. S. Afr. Mus. 59. (In press.)
Kraglievich, L. 1926. Los arctoterios norteamericanos (Tremarctotherium n. gen.) en relación con los de Sud América. An. Mus. nac. Hist. nat. B. Aires 33: 1-16.

Kurtén, B. 1957a. Percrocuta Kretzoi (Mammalia, Carnivora), a group of Neogene hyenas. Acta zool. cracov. 2: 375-404.
Kurtén, B. 1957b. The bears and hyenas of the Interglacials. Quaternaria 4: 1-13.
Kurtén, B. i957c. Mammal migrations, Cenozoic stratigraphy, and the age of Peking Man and the australopithecines. F. Paleont. 31: 215-227.
Kurtén, B. 1958. Life and death of the Pleistocene cave bear. Acta zool. fenn. 95: 1-59.
Kurtén, B. 1964. The evolution of the polar bear, Ursus maritimus Phipps. Acta zool. fenn. 108: $1-30$.
Kurtén, B. ig66. Pleistocene bears of North America. i. Genus Tremarctos, spectacled bears. Acta zool. fenn. 115: I-120.
Kurtén, B. 1967. Pleistocene bears of North America. 2. Genus Arctodus, short-faced bears. Acta zool. fenn. 117: 1-60.
Kurtén, B. ig68. Pleistocene mammals of Europe. London: Weidenfeld \& Nicolson.
Kurtén, B. 1971. The age of mammals. London: Weidenfeld \& Nicolson.
Lydekker, R. 1884. Indian Tertiary and post-Tertiary Vertebrata. Siwalik and Narbada Carnivora. Palaeont. indica (io) 2: 178-354.
Maglio, V. J. \& Hendey, Q. B. 1970. New evidence relating to the supposed stegolophodont ancestry of the Elephantidae. S. Afr. archaeol. Bull. 25: 85-87.
Matthew, W. D. 1929. Critical observations upon Siwalik mammals. Bull. Am. Mus. nat. Hist. 56: 437-56o.
Pilgrim, G. E. 1931. Catalogue of the Pontian Carnivora of Europe. London: British Museum (Natural History).
Pilgrim, G. E. 1932. The fossil Carnivora of India. Palaeont. indica (n.s.) 18: $\mathbf{1 - 2 3 2}$.
Reed, C. A. 1967. The generic allocation of the hominid species habilis as a problem in systematics. S. Afr. 7. Sci. 63: 3-5.
Simpson, G. G. 1945. The principles of classification and a classification of mammals. Bull. Am. Mus. nat. Hist. 85: 1-450.
Simpson, G. G. 1971. Fossil penguin from the Late Cenozoic of South Africa. Science 171: 1144-1145.
Smith-Woodward, A. 1915. On the skull of an extinct mammal related to Aeluropus from a cave in the ruby mines at Mogok, Burma. Proc. zool. Soc. Lond. 1915: 425-428.
Thenius, E. 1959a. Ursidenphylogenese und Biostratigraphie. Z. Säugetierk. 24: 78-84.
Thenius, E. r959b. Indarctos arctoides (Carnivora, Mammalia) aus dem Pliozän Osterreichs nebst einer Revision der Gattung. Neues fb. Geol. Paläont. Abh. 108: 270-295.
Thenius, E. rg66. Zur Stammesgeschichte der Hyänen (Carnivora, Mammalia). Z. Säugetierk. 31: 293-300.
Zdansky, O. 1924. Jungtertiäre Carnivoren Chinas. Paleont. sinica (C) 2: 1-149.


A-E Buccal, anterior, occlusal, lingual and oblique views of the Agriotherium africanum holotype L 2045.
F \& G Lingual and occlusal views of the $?^{3} \mathrm{P}^{3}$ of Agriotherium africanum L 1868 C .


A Lingual view of the $\mathrm{I}_{3}$ of Agriotherium africanum L 3141 and L 1868A.
B Occlusal and posterior views of the $\mathbf{M}_{3}$ of Agriotherium africanum Li256i.
C \& D Anterior and medial views of the ulna of Agriotherium africanum L 2154 .

## INSTRUCTIONS TO AUTHORS

## Based on

CONFERENCE OF BIOLOGICAL EDITORS, COMMITTEE ON FORM AND STYLE. 1960 .
Style manual for biological journals. Washington: American Institute of Biological Sciences.

## MANUSCRIPT

To be typewritten, double spaced, with good margins, arranged in the following order: ( I$)$ Heading, consisting of informative but brief title, name(s) of author(s), address(es) of author(s), number of illustrations (plates, figures, enumerated maps and tables) in the article. (2) Contents. (3) The main text, divided into principal divisions with major headings; subheadings to be used sparingly and enumeration of headings to be avoided. (4) Summary. (5) Acknowledgements. (6) References, as below. (7) Key to lettering of figures. (8) Explanation to plates.

## ILLUSTRATIONS

To be reducible to $12 \mathrm{~cm} \times 18 \mathrm{~cm}$ ( 19 cm including caption). A metric scale to appear with all photographs.

## REFERENGES

Harvard system (name and year) to be used: author's name and year of publication given in text; full references at the end of the article, arranged alphabetically by names, chronologically within each name, with suffixes $a, b$, etc. to the year for more than one paper by the same author in that year.
For books give title in italics, edition, volume number, place of publication, publisher.
For journal articles give title of article, title of journal in italics (abbreviated according to the World list of scientific periodicals. 4 th ed. London: Butterworths, 1963), series in parentheses, volume number, part number (only if independently paged) in parentheses, pagination.
Examples (note capitalization and punctuation)
Bullough, W. S. 196o. Practical invertebrate anatomy. 2nd ed. London: Macmillan.
Fischer, P.-H. i948. Données sur la résistance et de le vitalité des mollusques. 7. Conch., Paris 88: 100-140.
Fischer, P.-H., Duval, M. \& Raffy, A. 1933. Etudes sur les échanges respiratoires des littorines. Archs Zool. exp. gén. 74: 627-634.
Kohn, A. J. 1960a. Ecological notes on Conus (Mollusca: Gastropoda) in the Trincomalee region of Ceylon. Ann. Mag. nat. Hist. (13) 2: 309-320.
Kohn, A. J. ig6ob. Spawning behaviour, egg masses and larval development in Conus from the Indian Ocean. Bull. Bingham oceanogr. Coll. 17 (4): $1-51$.
Thiele, J. igio. Mollusca: B. Polyplacophora, Gastropoda marina, Bivalvia. In schultze, l. Zoologische und anthropologische Ergebnisse einer Forschungsreise im westlichen und zentralen SüdAfrika. 4: 269-270. Jena: Fischer. Denkschr. med-naturw. Ges. Jena 16: 269-270.

## ZOOLOGICAL NOMENCLATURE

To be governed by the rulings of the latest International code of zoological nomenclature issued by the International Trust for Zoological Nomenclature (particularly articles 22 and 51). The Harvard system of reference to be used in the synonymy lists, with the full references incorporated in the list at the end of the article, and not given in contracted form in the synonymy list.

## Example

Scalaria coronata Lamarck, 1816: pl. 451, figs $5 a, b$; Liste: II. Turton, 1932: 80.

ANNALS OF THE SOUTH AFRIGAN MUSEUM ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM

| Volume | $\mathbf{5 9}$ | Band |
| ---: | :---: | :--- |
| May | $\mathbf{1 9 7 2}$ | Mei |
| Part | $\mathbf{7}$ | Deel |



A NEW SPEGIES OF PARADOXOSTOMA (CRUSTACEA, OSTRACODA) FROM SOUTH AFRICA

## By

K. G. McKENZIE

Cape Town Kaapstad

## The ANNALS OF THE SOUTH AFRICAN MUSEUM

are issued in parts at irregular intervals as material becomes available

Obtainable from the South African Museum, P.O. Box 61, Cape Town

Die ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM word uitgegee in dele op ongereelde tye na beskikbaarheid van stof

Verkrygbaar van die Suid-Afrikaanse Museum, Posbus 6i, Kaapstad

> OUT OF PRINT/UIT DRUK
> I, $2(\mathrm{I}, 3,5,7-8), 3(\mathrm{I}-2,5$, t.-p.i. $) 5 \mathrm{I}-2,5,7-9)$, $6(\mathrm{I}, \mathrm{t} .-$ p.i. $), 7(\mathrm{I}-3), 8,8(\mathrm{I}-2), 10(\mathrm{I})$, II (i-2, 5, 7, t.-p.i.) $24(2), 27,3 \mathrm{I}(\mathrm{I}-3), 33$

Price of this part/Prys van hierdie deel
Ro,6o

## Trustees of the South African Museum (C) Trustees van die Suid-Afrikaanse Museum 1972

ISBN 0949940070

Printed in South Africa by The Rustica Press, Pty., Ltd, Court Road, Wynberg, Cape

In Suid-Afrika gedruk deur Die Rustica-pers, Edms., Bpk. Courtweg, Wynberg, Kaap

# A NEW SPECIES OF PARADOXOSTOMA (CRUSTACEA, OSTRACODA) FROM SOUTH AFRICA 

By<br>K. G. McKenzie<br>British Museum (Natural History), London

(With 12 figures)
[MS. accepted 30 November 1971]

## Contents

PAGE

## Introduction

During a working visit to the South African Museum in September 1970, I was shown a marine sample which had been collected at Sea Point, near Cape Town, by the late Dr. K. H. Barnard and which included a large population of a Paradoxostoma species. This species has proved to be new.

I am very grateful to the Director of the South African Museum, Dr. T. H. Barry, for making its facilities available; to Mr. B. F. Kensley, Curator of Crustacea, who drew my attention to the sample; to the National Institute for Water Research, Pretoria, and to my own Museum for financial support; and to Mr. D. Goode, the Transvaal Museum, who inked my original drawings.

Types are stored at the South African Museum under register number SAM A iloit and some paratypes are at the British Museum (Natural History) under register number $\mathrm{BM}(\mathrm{NH})$ 197.10.13.1-25.

## Systematics <br> Paradoxostoma kensleyin. sp.

Figures I-I2

## Derivation of name

For Mr. B. F. Kensley, who drew my attention to the sample and who has contributed several papers on the crustaceans of South Africa.

## Diagnosis

In lateral view, carapace ovate-subtriangular; of medium size (length up to about $\mathrm{o}, 65 \mathrm{~mm}$ ); smooth; without conspicuous colour patches in the specimen dissected (possibly, such coloration was present in life but disappeared


Paradoxostoma kensleyi n. sp., ovigerous ${ }^{\text {\& }}$, paratype
Fig. i. Internal view of right valve, $\times$ 300. Fig. 2. Antennule, $\times 750$. Fig. 3. Antenna with lobate antennal gland, $\times 750$. Fig. 4, Mandible coxale, $\times 750$.
following preservation) ; anterior margin subacuminate anteroventrally; dorsal margin strongly convex with a weak anterodorsal flexure in the right valve; posterior margin subacuminate posterodorsally, broadly rounded ventrally; ventral margin weakly inflexed in the vicinity of the oral cone; greatest height medial and about $60 \%$ of the length. In dorsal view, compressed; evenly elliptical. Internally, inner margin regular; line of concrescence also regular; vestibule continuous; radial pore canals few (about 7 anteriorly and 5 posteriorly) unbranched, short and straight; normal pore canals fairly numerous, scattered, simple; hinge adont or modified adont, with a weak terminal posterior projection in the right valve and a corresponding accommodation in the left valve; muscle scars comprising four adductors in a subvertical series, others not observed (Fig. I). Carapace sex dimorphism weak.

Antennule (Ai) 6-segmented; length ratios of the last four segments 18:27:13:3 (Fig. 2). Antenna (A2) 5-segmented, the penultimate segment appears to be sutured in its proximal half; length ratio of the terminal claws is about $3: 2$; the flagellum (Spinnborste) extends beyond the tips of the claws and is jointed at about $\frac{2}{3}$ its length from the proximal end; the gland to this Spinnborste is large, and lobate proximally (Fig. 3). Mandible with a styliform coxale (Fig. 4); palp two-segmented with 5 terminal bristles (Fig. 5). Oral cone present, with the characteristic suctorial modification (Fig. 6). Maxilla lacking a palp, trilobate (one lobe hidden in Fig. 7) ; epipod with about 13 Strahlen and with two downwards-pointing setae. First thoracic leg (Pi) pediform, four-segmented; protopod armed with a powerful dorsodistal clawlike spine (Fig. 8). Second and third thoracic legs ( $\mathrm{P}_{2}$ and $\mathrm{P}_{3}$ ) also pediform but with dorsodistal bristles instead of claw-like spines on their protopods (Figs 9, I I). None of the terminal claws on these legs are strongly spinose. Posterior of the body ( ( ) extended into a caudiform process with a terminal spine (Fig. 10). Hemipenis of male as illustrated (Fig. 12).

## Material

A very large population comprising numerous mature individuals and juveniles of both sexes.

## Locality

Sea Point, near Cape Town, Republic of South Africa.

## Collector and date collected

The late Dr. K. H. Barnard; March 1928.

## Dimensions

Holotype ( $0^{\top}$ ) Length $=0,52 \mathrm{~mm}$; Height $=0,31 \mathrm{~mm}$; Breadth $=0,18 \mathrm{~mm}$.
Allotype (ㅇ) Length $=0,56 \mathrm{~mm}$; Height $=0,34 \mathrm{~mm}$; Breadth $=0,20 \mathrm{~mm}$.

## Discussion

Of the previous workers on Recent South African marine Ostracoda (Brady 1880; Müller 1908; Klie ı940; Benson \& Maddocks 1964) only Klie


Paradoxostoma kensleyi n. sp., ovigerous ; paratype
(same specimen as in Figs 1-4)
Fig. 5. Mandible palp, $\times 750$. Fig. 6. Oral cone with suctorial disc, $\times 750$. Fig. 7. Maxilla, two lobes plus the downwards-directed setae, $\times 750$. Fig. 8. PI,$\times 750$. Fig. 9. P2,$\times 750$. Fig. 10. Posterior of body, $\times 750$. Fig. il. $\mathrm{P}_{3}, \times 750$.

Paradoxostoma kensleyi n. sp., mature đ̛, paratype
Fig. 12. Hemipenis, muscles only illustrated in the upper region, $\times 75^{\circ}$.
described any paradoxostomatids. He keyed six species in a Table (Klie 1940: 447) from which it appears that $P$. kensleyi is closest to the Klie species $P$. auritum and $P$. reflexum in characters based on the first four paired limbs (antennule, antenna, mandible, maxilla) and because it lacks pilosity on the ventral margin of the $\mathrm{P}_{3}$ third segment. But $P$. kensleyi differs from both these species in maximum size and has a different shape to that of P. reflexum. Apart from the maximum size difference, $P$. kensleyi has a different hemipenis to that of $P$. auritum and although similar in general carapace shape also appears to have a different line of concrescence (Klie 1940: 444). Another similar species is $P$. hypselum Müller 1908, which was described from the sub-Antarctic. I have recently determined a specimen which probably belongs to this species (USNM 137380) and the spines on the distal claws of the thoracic legs, for the $\mathrm{P}_{3}$ in particular, are distinctive, as pointed out by Müller (1908: 118, 119). Such distinct spines do not feature on the distal claws of the thoracic legs in kensleyi. Further, hypselum (length $\uparrow 0,72 \mathrm{~mm}, \widehat{0} 0,68 \mathrm{~mm}$ ) is a slightly larger species than kensleyi.

Summarizing, the known paradoxostomatid fauna of South Africa now comprises 7 species, namely: Paradoxostoma caeruleum Klie 1940, P. griseum Klie 1940, $P$. angustissimum Klie 1940, $P$. auritum Klie ı940, $P$. reflexum Klie 1940, $P$. semilunare Klie 1940 and $P$. kensleyi n.sp. It is likely that Dr. G. Hartmann, of the Zoologisches Museum und Staatinstitut, Hamburg, will describe further species when he monographs his large Recent South African collections.

## Summary

Paradoxostoma kensleyi, a new marine ostracode collected near Cape Town, South Africa, is described and compared with previously described South African paradoxostomatids.

## References

Benson, R. H. \& Maddocks, R. F. ig64. Recent ostracodes of Knysna Cape Province, Union of South Africa. Paleont. Contr. Univ. Kans. 34 (Arthropoda 5): 1-39.
Brady, G. S. i88o. Report on the Ostracoda dredged by H.M.S. Challenger during the years 1873-1876. Rep. Voy. Challenger 1873-76. x (Zoology 3): 1-184.
Klie, W. 1940. Beiträge zur Fauna des Eulitorals von Deutsch-Südwest-Afrika. II. Ostracoden von der Küste Deutsch-Südwest-Afrikas. Kieler Meeresforsch. 3: 404-448.
Müller, G. W. 1908. Die Ostracoden der Deutschen Südpolar-Expedition 1901-1903. Dt. Südpol.-Exped. 10: 52-182.

## INSTRUCTIONS TO AUTHORS

## Based on

CONFERENCE OF BIOLOGICAL EDITORS, COMMITTEE ON FORM AND STYLE. 1960 .
Style manual for biological journals. Washington: American Institute of Biological Sciences.

## MANUSCRIPT

To be typewritten, double spaced, with good margins, arranged in the following order: (i) Heading, consisting of informative but brief title, name(s) of author(s), address(es) of author(s), number of illustrations (plates, figures, enumerated maps and tables) in the article. (2) Contents. (3) The main text, divided into principal divisions with major headings; subheadings to be used sparingly and enumeration of headings to be avoided. (4) Summary. (5) Acknowledgements. (6) References, as below. (7) Key to lettering of figures. (8) Explanation to plates.

## ILLUSTRATIONS

To be reducible to $12 \mathrm{~cm} \times 18 \mathrm{~cm}$ ( 19 cm including caption). A metric scale to appear with all photographs.

## REFERENGES

Harvard system (name and year) to be used: author's name and year of publication given in text; full references at the end of the article, arranged alphabetically by names, chronologically within each name, with suffixes $a, b$, etc. to the year for more than one paper by the same author in that year.
For books give title in italics, edition, volume number, place of publication, publisher.
For journal articles give title of article, title of journal in italics (abbreviated according to the World list of scientific periodicals. 4th ed. London: Butterworths, 1963), series in parentheses, volume number, part number (only if independently paged) in parentheses, pagination.

## Examples (note capitalization and punctuation)

Bullough, W. S. 1960. Practical invertebrate anatomy. 2nd ed. London: Macmillan.
Fischer, P.-H. 1948. Données sur la résistance et de le vitalité des mollusques. F. Conch., Paris 88: 100-140.
Fischer, P.-H., Duval, M. \& Raffy, A. 1933. Etudes sur les échanges respiratoires des littorines. Archs Zool. exp. gén. 74: 627-634.
Конn, A. J. 196oa. Ecological notes on Conus (Mollusca: Gastropoda) in the Trincomalee region of Ceylon. Ann. Mag. nat. Hist. (13) 2: 309-320.
Kohn, A. J. 196ob. Spawning behaviour, egg masses and larval development in Conus from the Indian Ocean. Bull. Bingham oceanogr. Coll. 17 (4): 1-51.
Thiele, J. igio. Mollusca: B. Polyplacophora, Gastropoda marina, Bivalvia. In schultze, l. Zoologische und anthropologische Ergebnisse einer Forschungsreise im westlichen und zentralen SüdAfrika. 4: 269-270. Jena: Fischer. Denkschr. med-naturw. Ges. Jena 16: 269-270.

## ZOOLOGICAL NOMENCLATURE

To be governed by the rulings of the latest International code of zoological nomenclature issued by the International Trust for Zoological Nomenclature (particularly articles 22 and 51). The Harvard system of reference to be used in the synonymy lists, with the full references incorporated in the list at the end of the article, and not given in contracted form in the synonymy list.

## Example

Scalaria coronata Lamarck, 1816: pl. 451, figs $5 a, b$; Liste: 11. Turton, 1932 : 80 .

ANNALS OF THE SOUTH AFRICAN MUSEUM ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM

| Volume | $\mathbf{5 9}$ | Band |
| ---: | :---: | :--- |
| May | $\mathbf{1 9 7 2}$ | Mei |
| Part | $\mathbf{8}$ | Deel |



DEVELOPMENT OF TRACHURUS TRACHURUS (CARANGIDAE), THE SOUTH AFRICAN MAASBANKER

By<br>E. H. HAIGH

# The ANNALS OF THE SOUTH AFRICAN MUSEUM 

are issued in parts at irregular intervals as material becomes available

Obtainable from the South African Museum, P.O. Box 6i, Cape Town

## Die ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM

word uitgegee in dele op ongereelde tye na beskikbaarheid van stof

Verkrygbaar van die Suid-Afrikaanse Museum, Posbus 6I, Kaapstad

> OUT OF PRINT/UIT DRUK
> 1, 2(1, 3, 5, 7-8), 3(1-2, 5, t.-p.i.), 5(2, 5, 7-9), $6(\mathrm{I}, \quad$ t.-p.i.), $\quad 7(\mathrm{I}-3), \quad 8, \quad 9(\mathrm{I}-2), \quad 10(\mathrm{I})$, II(1-2,5,7, t.-p.i.) $24(2), 27,31(1-3), 33$

Price of this part/Prys van hierdie deel RI,35
$\begin{array}{ll}\text { Trustees of the South African Museum (C) Trustees van die Suid-Afrikaanse Museum } \\ & 1972\end{array}$

ISBN 0949940089

Printed in South Africa by
The Rustica Press, Pty., Ltd.
Court Road, Wynberg, Cape

In Suid-Afrika gedruk deur
Die Rustica-pers, Edms., Bpk.
Courtweg, Wynberg, Kaap

# DEVELOPMENT OF TRACHURUS TRACHURUS (CARANGIDAE), THE SOUTH AFRICAN MAASBANKER 

\author{
By <br> E. H. Haigh <br> South African Museum, Cape Town <br> (With 4 figures and 3 tables) <br> [MS. accepted 29 November 1971] <br> \section*{Contents} <br> 

## Introduation

The systematics of the genus Trachurus of the family Carangidae seem to be rather confused and need revision, based on an adequate world-wide collection. Berry (personal communication) is of the opinion that there are two species in South African waters. However, his review paper on the genus Trachurus is only to appear in three or four months. The present larval fish collection is not geographically wide enough for accurate comparison of the two possible species and as the distinguishing features are essentially adult characters, the designation Trachurus trachurus for these specimens is felt to be the most accurate, at the same time indicating the similarity of these larvae to European Trachurus trachurus larvae.

Several papers describing the larval development of specimens bearing either the generic or specific name Trachurus have appeared since late in the nineteenth century and this seems an opportune time to review what is known about larvae of this genus.

The description of Caranx trachurus by Holt (1898) does not give details and the illustrations in Annales du Musée d'Histoire Naturelle de Marseille (5, 1899: 27-32, figs 53-63) to which he refers were not available. However, Ehrenbaum (1909: 27-30) reviews Holt's publications as do Heincke \& Ehrenbaum (igoo).

All authors seem to be satisfied as to the identity of the egg of Trachurus trachurus and describe it as having a diameter of 0,7 to $1,09 \mathrm{~mm}$, a totally segmented yolk and an oil globule of 0,19 to $0,28 \mathrm{~mm}$ diameter with yellowish to brownish pigment around it which remain in the anterior part of the yolk
until resorbed. South African maasbanker eggs that have been measured are seldom larger than I mm in diameter-usually about $0,9 \mathrm{~mm}$ with an oil globule of $0,2 \mathrm{~mm}$ diameter. This falls within the range mentioned above.

Ehrenbaum (1909) gives a short description and six rather inadequate figures of Trachurus trachurus, but these do serve to confirm the basic similarity to the larvae described by Schnakenbeck (193I). Unfortunately Schnakenbeck fails to provide adequate dimensional data. By modern standards Schnakenbeck's size groups are rather large, but comparative reworking of measurements in the present paper produces values which approximate these.

Delsman (1926) described eggs and early larval stages from the Pacific round the Indonesian islands under the name Caranx kurra. However, both eggs and larvae are so similar to descriptions by Ehrenbaum (1909) that they could easily be larvae of the genus Trachurus, probably maccullochi, which Nichols (1940) put as a race of Trachurus trachurus.

The description of Trachurus symmetricus larvae from the Pacific coast of America by Ahlstrom \& Ball (r954) again shows the small differences between species of Trachurus. Trachurus symmetricus larvae can perhaps be separated from Trachurus trachurus larvae by the very slight difference in degree of pigmentation, the former being less pigmented. However, degree of pigmentation hinges to a very large extent on the length and method of preservation, and perhaps also on the time of capture, be it daylight or after dark. Pigmentation is thus an unreliable characteristic for the distinction of species so closely related.

Aleev (1957) described a good developmental series of Trachurus mediterraneus ponticus Aleev. Unfortunately his diagrams do not show ossification details or pigmentation patterns and can thus not be successfully compared with the figures published by Dechnik \& Seniokova in 1964 of Trachurus mediterraneus (?).

In 1969 Zhudova published three figures of Trachurus larvae described as T. trachurus and a distribution map giving distribution of Trachurus larvae between $5^{\circ} \mathrm{N}$ to $12^{\circ} \mathrm{S}$ and $10^{\circ} \mathrm{E}$ to $14^{\circ} \mathrm{W}$ in the Gulf of Guinea and in 1970 Kiliachenkova published several good figures of eggs and larvae up to $8,9 \mathrm{~mm}$ in length caught along the West African west coast between $24^{\circ}$ and $15^{\circ} \mathrm{N}$ and $19^{\circ} 50^{\prime}$ and $17^{\circ} 50^{\prime} \mathrm{W}$.

The adult maasbanker is found in most parts of the Atlantic down to a depth of 400 metres. It occurs abundantly on the west coast of southern Africa, where, with Sardinops ocellata (Pappe, 1853), the South African pilchard, it forms the basis of the fishmeal industry. Like the pilchard, it is a plankton feeder, its diet consisting mainly of zooplankton including amphipods, euphausids and fish larvae.

Eggs and larvae are usually found in deep seas (see section on distribution), while juveniles are found mostly in sheltered bays close inland. Several authors report the clustering of small and juvenile Trachurus under jellyfish (Ehrenbaum 1909) and floating debris and seaweed (Delsman 1926). Apparently this habit is common to the Carangidae.

## Materials and Methods

Specimens were obtained by research vessels of the Division of Sea Fisheries, Cape Town, using NiooB and NiooH plankton nets, from 1950 to 1967. Samples were fixed and stored in formalin which was replaced by $70 \%$ ethyl alcohol. Specimens were stained, using methods of Hollister (1934) but modified slightly by reducing the clearing time in KOH and reducing the concentration of the KOH used. This was done in order to preserve pigment in specimens. As pigment is inclined to fade with time, more than one larva in the size range was used in order to obtain the most characteristic pigment pattern. Stained specimens were preserved in glycerin.

Measurements were taken as follows:
Standard length (s.l.) : tip of lower jaw to end of caudal peduncle
snout: tip of lower jaw to anterior margin of eye
eye diameter:
head length:
trunk length:
depth:
the eye being essentially round, could be measured in any direction
tip of snout to cleithrum
tip of snout to posterior end of anus, measured along the midline with verticle to snout and anus taken at posterior edge of head

All proportions are presented as percentage of standard length.
All lengths cited in text are the standard length.

## Description

The adults and juveniles of Trachurus are distinguished from other genera of the family Carangidae by the absence of separate anal and dorsal finlet, the presence of laterally expanded scutes over the full length of the lateral line and a procumbent spine before the spinous dorsal fin. As far as the distinctive South African species is concerned, Smith (1953) says that Trachurus trachurus has between 70 and 90 scutes on the lateral line, a depth of $45 \%$ and more than 30 dorsal rays.

Among later larval stages, dorsal fin-count of $8+3 \mathrm{I}-33$ and anal fin-count of $3+27-29$ combined with distinctive pigmentation pattern will serve to separate Trachurus larvae from Decapterus larvae which have a darkly pigmented area on the head.

## Ossification

The smallest specimens show slight ossification of the premaxilla, dentary and cleithrum. The three horizontal, one corner and one vertically situated preopercular spines are lightly ossified (Fig. iA). Between $3,0 \mathrm{~mm}$ and $4,5 \mathrm{~mm}$ ossification takes place rapidly. The premaxilla with four fine teeth is well formed. The maxilla is lightly ossified. The dentary and angular can be clearly distinguished and there are six ossified branchiostegal rays. The preopercle now


Fig. I. Trachurus trachurus. Early larval stages showing pigmentation, pattern and position and degree of bone development. Measurements indicate standard length.
A. $3,30 \mathrm{~mm}$.
B. $4,50 \mathrm{~mm}$.
C. $5,8 \mathrm{omm}$.
D. $7,2 \mathrm{~mm}$.
bears four spines on the horizontal arm, the corner spine has become very long and there are two spines on the vertical arm. The number of spines laid down between this stage and $5,4 \mathrm{~mm}$ is usually the number that remains for the rest of the development. There is a spiny ridge on the outer anterior edge of the preopercle with six to seven small spines on it.

The cleithrum broadens very gradually and by $5,4 \mathrm{~mm}$ the supracleithrum has appeared as have the first traces of the opercle, the quadrate and the pterygoid. There are now seven branchiostegals. Between $5,4 \mathrm{~mm}$ and $6,6 \mathrm{~mm}$ further ossification takes place, all above-mentioned bones becoming heavier, darker and broader. New bones make their appearance, i.e. the frontal, sphenotic and parietal as well as the supra-occipital crest over the brain region. Laterally from this is the first indication of the pterotic and behind the pterotic
what appear to be the first traces of the exoccipital. The hyomandibular and symplectic can be clearly distinguished as can the opercle, the subopercle and interopercle. The epihyal and ceratohyal show the first traces of ossification. The articular and dentary have become fairly well amalgamated. By $7,8 \mathrm{~mm}$ the pterygoid has become fairly extensive (Fig. 2A) and traces of the lacrimal and nasal can be seen. The postcleithral bones which first become evident at about 6 mm are now joined by traces of the scapula and the pelvic girdle is also evident. Ceratohyal and epihyal can be easily seen.

The cranial ossification proceeds rapidly after $7,8 \mathrm{~mm}$. The whole brain region becomes ossified and bones join up. The post-temporal appears above the supracleithrum and there are several centres of ossification in the supratemporal region. The circumorbital bones are evident but the nasal region is still fairly unossified. The maxilla has now become the major bone in the upper jaw, almost completely obscuring the dentary, when the mouth is closed. The spines on the preopercle are less prominent. Ceratohyal and epihyal are well ossified but still separate. The pectoral girdle elements present are scapula and three pterygials while the pelvic girdle is well ossified.

Table I
Average meristic counts during development of Trachurus trachurus.

| Average s.l. | Dorsal spines rays | Anal <br> spines rays | Vertebrae <br> trunk tail | Neural spines | Haemal spines | Pectoral fin | Pelvic <br> fin | Caudal. fin |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 4,25 |  | - - | $\mathrm{O}-10+\mathrm{O}-\mathrm{I}$ | 3-16 | 4-6 | 0-3 | - | 1-5+1- |
| 6,03 | 0-4 | 0-5 | $\mathrm{O}-10+0-4$ | 0-18 | o-8 | --5 | - | $3-5+3-5$ |
| 6,82 | 0-6+ 0-17 | $0-1+0-14$ | $0-10+0-12$ | 2-20 | 3-10 | 3-8 | - | 1-9+2-8 |
| 8,34 | $0-7+3-24$ | $1-2+7-22$ | $10+10-14$ | 20-22 | 1 I-I3 | 6-14 | I-3 | $8-9+8$ |
| 9,16 | $6-8+13-25$ | $1-3+12-19$ | $10+11-14$ | 20-22 | II-I3 | 8-15 | 3-4 | $9+8$ |
| 9,8o | $5-8+20-30$ | $3+16-24$ | $10+13-14$ | 22 | 12-13 | 9-17 | 3-5 | $9+8$ |
| I 1,6I | $8+\mathrm{I}+28-33$ | $2+1+26-28$ | $10+14$ | 22 | 13 | ı6-19 | $1+4-5$ | $9+8$ |
| 13,51 | 8-9+1+25-33 | $2+1+23-28$ | $10+14$ | 22 | 13 | 7-18 | $1+5$ | $9+8$ |
| 15,37 | $8+\mathrm{I}+3 \mathrm{I}-32$ | $2+1+26-27$ | $10+14$ | 22 | 13 | 18-21 | I +5 | $9+8$ |
| 26,00 | $8+1+30$ | $2+1+26$ | $10+14$ | 22 | 13 | $1+21$ | $1+5$ | $9+8$ |
| 29,25 | $8+\mathrm{I}+33$ | $2+1+29$ | $10+14$ | 22 | 13 | $1+20$ | $\mathrm{I}+5$ | $9+8$ |
| 34,54 | $8+\mathrm{I}+34$ | $2+1+30$ | $10+14$ | 22 | 13 | $\mathrm{I}+2 \mathrm{I}$ | $\mathrm{I}+5$ | $9+8$ |
| 48,50 | $8+\mathrm{I}+33$ | $2+1+30$ | $10+14$ | 22 | 13 | $\mathrm{I}+2 \mathrm{I}$ | $\mathrm{I}+5$ | $9+8$ |

In the postcranial region ossification starts at about $4,5 \mathrm{~mm}$ in some specimens while others show only the first traces of the haemal and neural spines at about $6,0 \mathrm{~mm}$. The ossification of the vertebral column appears to start anteriorly with two to three neural spines, then some haemal and neural spines ossify medially, and after this the neural spines over the intestinal sac ossify, followed by the posterior haemal and neural spines. Between $5,4 \mathrm{~mm}$ and $6,6 \mathrm{~mm}$ the vertebral centra ossify rapidly, and apparently several at once, from the anterior, with the result that by $6,6 \mathrm{~mm}$ there can be as many as
ten trunk centra and up to twelve tail centra partly or fully ossified. The urostyle ossifies before the penultimate vertebra.

By $7,8 \mathrm{~mm}$ ten tail centra are ossified, and by $9,9 \mathrm{~mm}$ the full complement of ten trunk and fourteen tail centra, including the urostyle, is ossified. The neural and haemal spines have broadened by now, especially in the caudal plate where three haemal and two neural spines support the caudal fin rays. Schnackenbeck (193I) gives an extensive account of the caudal ossification of European species and ossification in the specimens described follows the same pattern.

The median fin rays are first evident between 5,8 and $6,6 \mathrm{~mm}$ and those in the dorsal and anal fins are formed simultaneously. Between $6,5 \mathrm{~mm}$ and $7,8 \mathrm{~mm}$ the dorsal and anal spines appear. The third anal spine is associated with the soft anal and only becomes thickened at about 9, Imm . The middle dorsal spines appear first then the most posterior and the seventh and eighth spines last of all, usually between 9,0 and $10,0 \mathrm{~mm}$. The small procumbent dorsal spine is formed only during juvenile stage. The spine of the soft dorsal becomes thickened only at about $8,5-9,0 \mathrm{~mm}$. The rays of the anterior and midsections of both fins are formed first and ossification proceeds posteriorly gradually until both fins are fully ossified at about ir,o mm s.l. Between 9 mm and 12 mm the fin supports of both fins are formed.

The pectoral lobe is present even in smaller specimens and ossification of the rays starts as early as $5,2 \mathrm{~mm}$ in a few specimens but in the majority of specimens examined general pectoral fin ray ossification commences between 6,0 and $6,50 \mathrm{~mm}$ at the most dorsal aspect of the fin and proceeds round the periphery ventrally in sequence. Ossification takes place rapidly and by $10,0 \mathrm{~mm}$ there are usually about 17 rays ossified. The full complement of 22 rays with the most anterior ray considerably thickened can be seen only in juveniles of over $20,0 \mathrm{~mm}$.

There are no evident pelvic lobes and the first ray appears only at $7,5 \mathrm{~mm}$ but by $9,8 \mathrm{~mm}$ there can be as many as five rays ossified and by $10,0 \mathrm{~mm}$ the outermost ray has become noticeably thickened. The full number of one spine and five rays is present at $11,0 \mathrm{~mm}$.

The caudal fin ossification starts very early on in development, as soon as jaws and cleithrum have ossified, and well before the urostyle has turned up. Rays ossify from the middle outward and by $6,5 \mathrm{~mm}$ some specimens show the full complement of nine dorsal and eight ventral primary caudal rays. Secondary caudal rays start ossifying almost immediately after this size and proceed gradually until well into the juvenile stage. The urostyle turns up gradually at about $6,0 \mathrm{~mm}$ s.1.

Juvenile ossification takes place mainly in terms of consolidation of bones although there is of course a tremendous amount of growth taking place. The juvenile stage is reached between 10,0 and $12,0 \mathrm{~mm}$ although the lateral line scutes, a major distinguishing feature of the species, appear only between 18 and 19 mm .

Ossification of Trachurus trachurus larvae does not take place at the same rate in all specimens and commencement also varies considerably. If position of capture, time of year and water temperature are considered, it appears that larvae of similar sizes caught in the same latitudes will show different degrees of ossification, depending upon whether they were caught early or late in spring or summer. Larvae of $6,5 \mathrm{~mm}$ were caught in water temperatures of $15,79^{\circ}$ and $22,18^{\circ} \mathrm{C}$. Those caught in $15,79^{\circ} \mathrm{C}$ (August) showed far less vertebral ossification than those caught at $22,18^{\circ} \mathrm{C}$ (January).

Pigmentation


Fig. 2. Trachurus trachurus. Late larval stages with ossification nearing completion. A. $8,4 \mathrm{~mm}$ standard length. B. $10,5 \mathrm{~mm}$ standard length showing ossification. sop. subopercle. iop. interopercle. C. Same specimen showing pigmentation pattern.

The pigmentation pattern in Trachurus species from the southern African region is much the same as that described for European specimens by Ehrenbaum (1909) and Schnakenbeck (193I) as well as that of Trachurus symmetricus, described by Ahlstrom \& Ball (1954).

In the early larvae (Fig. iA-B) the pigment is distributed in three main areas, dorsally along the edge of the body and over the brain area, mediolaterally along the peritoneal wall and over the medioposterior region of the chorda and ventrally along the edge of the trunk and tail with a few scattered melanophores on the upper and lower jaws and in the region of the cleithrum. The dorsal pigmentation is soon augmented (Fig. ${ }_{\mathrm{I}} \mathrm{C}$ ) by melanophores appearing on the dorsolateral sides of the body musculature. The median row of elongated contracted melanophores in the lateral line region which is so characteristic of many carangid larvae, becomes darker and more distinct. A second row of pigment appears directly over the notochord. On the peritoneum pigment darkens dorsally and scattered melanophores appear laterally. Chromatophores are still present on the nose, the jaws and underneath the branchiostegal rays. By $6,6 \mathrm{~mm}$ (Fig. ID) a second pigmentation on the caudal plate itself has become more clearly defined. Between 7 mm and 10 mm the body wall thickens and fins are formed obscuring much of the deeper-lying pigmentation. The balance of pigmentation changes rapidly, the dorsal aspect of the fish becoming far more heavily pigmented than the ventral (Fig. 2C). There are large and small melanophores on the dorsolateral aspect. The more dorsally situated ones seem to be far larger than those on the lateral side.

The dorsal aspect of the head has also become pigmented to uniformity with the body. There are still chromatophores on the jaws and nose but none is visible on the trunk region. The ventral aspect of the tail still bears widely scattered melanophores but the caudal plate and unpaired fins have become pigmented. Both spiny sections of the median fins are pigmented, while the rayed parts are clear. The lateral line row of melanophores is still very distinct and remains so until the juvenile stage is reached.

Changes in body-form
The earliest stages of Trachurus sp. present in these collections have the yolk-sac absorbed and intestine with one fold developed at $2,45 \mathrm{~mm}$. The eye has become pigmented but the snout is shorter than eye diameter ( $7,7 \%$ v.s. $10,0 \%$ of s.l.). During the next millimetre increase in length the head increases proportionately rapidly from $25 \%$ to $33,5 \%$ of standard length and the depth remains constant at about $30 \%$. However, this depth is measured at the region where the head joins the trunk and it is noticeable (Table 2) that this original proportion is slightly but definitely decreased in the larger stages. This is no doubt due to rapid increase in the head during early development and the increase in tail size in later development. If larvae shown in Figures 1 and 2 are compared, it will be noticed that the younger stages are proportionately much deeper anteriorly than posteriorly while the later stages are of more even proportions.

The larval development of Trachurus sp. is thus characterized by a smooth and gradual development. Fins form in adult positions and body proportions change gradually and slightly.

Table 2
Mean measurements of Trachurus trachurus in mm.

| Size range | Average size | No. | Snout | Eye | Head | Trunk | Depth |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 2,5-3,5 | 3,17 | 5 | 0,25 | 0,33 | 0,90 | 1,70 | 0,95 |
| 3,5-4,5 | 4,04 | 29 | 0,45 | 0,40 | 1,43 | 2,40 | 1,30 |
| 4,5-5,5 | 4,93 | 45 | 0,53 | 0,50 | 1,65 | 2,80 | 1,55 |
| 5,5-6,5 | 5,84 | 32 | 0,70 | o,6o | 2,00 | 3,40 | 1,80 |
| 6,5-7,5 | 6,92 | 32 | 0,75 | 0,75 | 2,50 | 4,00 | 2,30 |
| 7,5-8,5 | 8, 15 | 29 | 0,95 | 0,95 | 3,00 | 4,90 | 2,60 |
| 8,5-9,5 | 9,04 | 16 | 1,05 | 1,05 | 3,20 | 5,30 | 2,80 |
| 9,5-10,5 | 10,01 | 13 | 1,15 | 1,10 | 3,40 | 5,80 | 3,05 |
| 10,5-11,5 | 10,89 | 6 | 1,20 | 1,25 | 3,70 | 6,20 | 3,20 |
| 11,5-12,5 | 11,28 | 17 | 1,40 | 1,40 | 4,10 | 6,8o | 3,65 |
| 12,5-13,5 | 12,99 | 3 | 1,60 | 1,40 | 4,6o | 7,20 | 3,90 |
| 13,5-14,5 | 13,90 | 7 | 1,50 | 1,50 | 4,8o | 7,70 | 4,00 |
| 14,5-15,5 | 15,31 | 6 | 1,50 | 1,70 | 5,20 | 8,40 | 4,50 |
| 1 5,6-16,6 | - | o | - | - | - | - | - |
| 16,6-1 7,6 | 16,90 | 3 | 2,00 | 1,80 | 5,80 | 9,70 | 4,8o |
| 1 7,6-18,6 | 18,20 | 1 | 1,80 | 2,00 | 5,90 | 9,8o | 5,50 |
| 18,6-19,6 | 18,88 | 4 | 2,10 | 1,90 | 6,40 | 9,8o | 5,20 |
| 19,6-20,6 | 20,15 | 2 | 2,20 | 2,00 | 6,50 | 10,50 | 6,10 |
| 20,6-21,6 | 21,01 | 4 | 2,10 | 2,20 | 6,70 | 11,10 | 6,00 |
| 23,67 | 23,67 |  | 2,27 | 2,27 | 7,80 | 12,02 | 5,85 |
| 24,05 | 24,05 | I | 2,27 | 2,60 | 7,80 | I 1,30 | 6,20 |
| 25,67 | 25,67 | 1 | 2,60 | 2,92 | 8,45 | 13,65 | 6,82 |
| 29,00 | 29,00 | I | 3,00 | 3,00 | 9,50 | 14,60 | 7,50 |

Table 3
Mean body proportions of Trachurus trachurus larvae as \% of standard length.

| Size range <br> in mm | Average <br> size <br> in mm | No. | Snout | Eye | Head | Trunk | Depth |
| :---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: |
| $2,5-3,5$ | 3,17 | 5 | 7,5 | 10,0 | 25,0 | 54,0 | 30,0 |
| $3,5-4,5$ | 4,04 | 29 | 10,5 | 9,5 | 33,5 | 56,0 | 30,5 |
| $4,5-5,5$ | 4,93 | 45 | 11,0 | 10,5 | 32,0 | 57,0 | 31,5 |
| $5,5-6,5$ | 5,84 | 32 | 11,5 | 10,5 | 35,0 | 53,0 | 31,0 |
| $6,5-7,5$ | 6,92 | 32 | 11,0 | 11,0 | 36,0 | 59,5 | 32,0 |
| $7,5-8,5$ | 8,15 | 29 | 13,0 | 11,5 | 36,0 | 59,5 | 31,5 |
| $8,5-9,5$ | 9,04 | 16 | 12,0 | 11,5 | 35,0 | 58,0 | 31,0 |
| $9,5-10,5$ | 10,01 | 13 | 11,5 | 11,0 | 34,0 | 58,0 | 31,0 |
| $10,5-11,5$ | 10,89 | 6 | 10,5 | 11,5 | 34,0 | 57,0 | 29,0 |
| $11,5-12,5$ | 11,28 | 17 | 11,5 | 11,5 | 34,5 | 57,0 | 30,5 |
| $12,5-13,5$ | 12,99 | 3 | 12,0 | 10,5 | 35,0 | 55,5 | 30,0 |
| $13,5-14,5$ | 13,90 | 7 | 10,5 | 11,0 | 34,5 | 55,5 | 29,0 |
| $14,5-15,5$ | 15,31 | 6 | 9,6 | 11,0 | 34,0 | 54,5 | 29,0 |
| $16,6-17,6$ | 16,90 | 3 | 11,0 | 11,0 | 34,0 | 57,0 | 28,0 |
| $17,6-18,6$ | 18,20 | 1 | 10,0 | 10,5 | 32,0 | 53,5 | 30,5 |
| $18,6-19,6$ | 18,88 | 4 | 11,0 | 10,0 | 33,5 | 52,0 | 27,5 |
| $19,6-20,0$ | 20,15 | 2 | 11,0 | 10,5 | 32,5 | 53,0 | 30,0 |
| $20,0-21,0$ | 21,01 | 4 | 10,0 | 10,0 | 32,0 | 52,5 | 28,5 |
| 23,67 | 23,67 | I | 9,5 | 9,5 | 33,0 | 51,0 | 24,5 |
| 24,05 | 24,05 | I | 9,5 | 11,5 | 32,5 | 47,0 | 25,5 |
| 25,67 | 25,67 | 1 | 10,0 | 11,5 | 33,0 | 53,0 | 26,5 |
| 29,00 | 29,00 | 1 | 10,5 | 10,5 | 32,5 | 50,5 | 26,0 |

The air-bladder starts as a small clear patch behind the cleithrum and extends rapidly posteriorly until it occupies about $50 \%$ of the dorsal longitudinal distance of the peritoneum.


Fig. 3. Trachurus trachurus juvenile

| ang. | angular | hyp. | hypurals | ptm. | post-temporal |
| :--- | :--- | :--- | :--- | :--- | :--- |
| art. | articular | la. | lacrimal | pto. | pterotic |
| brstg. | branchiostegal rays | mx. | maxilla | ptr. | pterygoid |
| clt. cleithrum | na. | nasal | ptryg. | pterygials |  |
| cor. coracoid | pa. | parietal | quad. | quadrate |  |
| c. orb. circumorbitals | pclt. i \& 2 | postcleithra | scap. | scapula |  |
| deth. dermethmoid | pelv. | pelvic girdle | soc. | supraoccipital |  |
| dn. dentary | pf. | prefrontal | supcl. | supracleithrum |  |
| f. | frontal | pmx. | premaxilla | sym. | symplectic |
| hyom. hyomandibular | pop. | preopercle | u. | urostyle |  |

## Distribution

The area covered by the research vessels of the Division of Sea Fisheries on the pilchard research programme has varied since the inception of the programme. During 195I and 1952 the area worked lay between $32^{\circ}$ and $35^{\circ} 30^{\prime} \mathrm{S}$ and was delimited by the 200 fathom depthline to the west. Approximately the same area was worked between 1953 and 1957. In 1958 the eastern limit of the work area was extended round Cape Point to $19^{\circ} 30^{\prime}$ E. This area was worked until the end of 1960 when the eastward delimitation was extended to $21^{\circ}$ E. During these years the westward delimitation extended to $16^{\circ} 31^{\prime} \mathrm{E}$. From July 1963 to December 1965 the area covered by the ships was between $32^{\circ}$ 1о' to $36^{\circ}$ ro'S and $16^{\circ}$ to $21^{\circ} 30^{\prime} \mathrm{E}$. Station lists are obtainable from the Annual Reports of the Division of Sea Fisheries, for the relevant years. Figure 4 shows positions where larvae and juveniles were captured from 195I to 1965 . The most productive year was 1964 when Trachurus sp. were caught at 81 stations visited, followed by 1965 when 68 stations yielded Trachurus sp.

Trachurus larvae were caught throughout the year, the smallest number of stations yielding larvae in July and the largest number during October, that is late spring in the Southern Hemisphere. The spring months, August, September and October, yielded the largest collection of larvae while the three other seasons yielded far fewer and were not markedly different.


Fig. 4. Distribution of Trachurus trachurus larvae in the research area between 1951 and 1965 .

## Summary

The development of the South African larvae of Trachurus trachurus is described and a general map of the distribution of the larvae provided.

## Acknowledgements

The author wishes to thank the Division of Sea Fisheries' sea-going staff for the collection of study material, the Fisheries Development Corporation for financial assistance, and the South African Museum, Cape Town, for housing. Dr Naomi Millard read and criticized the manuscript.

## References

Ahlstrom, E. H. \& Ball, O. P. 1954. Description of eggs and larvae of jack mackerel (Trachurus symmetricus) and distribution and abundance of larvae in 1950 and 1951. Fishery Bull. Fish Wildl. Serv. U.S. 56: 209-254.

Aleev, Y. G. 1957. [On ten species of Trachurus in the U.S.S.R. seas.] Trudy sevastopol'. biol. Sta. 9: 167-242. (In Russian.)
Dechnik, T. V. \& Seniokova, V. I. 1964. [Distribution of pelagic fish eggs and larvae in the Mediterranean Sea.] Trudy sevastopol'. biol. Sta. 15: 77-1 15. (In Russian.)
Delsman, H. C. 1926. Fish eggs and larvae from the Java Sea (i). Treubia 8: 199-239.
Ehrenbaum, E. 1909. Eier und Larven von Fischen. i.Teil. Nord. Plankt. Lief. 4: 1-216.
Heincke, F. \& Ehrenbaum, E. 1900. Eier und Larven von Fischen der deutschen Bucht. Wiss. Meeresunters. (Abt. Helgoland) 3: 277-279.
Hollister, G. 1934. Clearing and dyeing fish for bone study. Zoologica, N.N. 12: 89-10i.
Holt, E. W. L. i898. Notes on the reproduction of teleostean fishes in the south-western district. 7. mar. biol. Assoc. U.K. 5: 107-1 55.
Kiliaghenkova, V. A. i970. Development and distribution of eggs and larvae of Trachurus trachurus L. Rapp. P.-v. Réun. Cons. perm. int. Explor. Mer 159: 194-198.
Nichols, J. T. 1940. Notes on carangin fishes. V. Young Trachurus in the Gulf of Mexico. Am. Mus. Novit. 1067: 1-4.
Schnakenbeck, W. 1931. Carangidae. Rep. Dan. oceanogr. Exped. Mediterr. 2(A 14): 3-13.
Smith, J. L. B. 1953. The sea fishes of southern Africa. 4th ed. Cape Town: Central News Agency.
Zhudova, A. M. ig69. [Materials and study of the eggs and larvae of some species of fish from the Gulf of Guinea and the adjacent waters of the open ocean.] Trudy AtlantNIRO Inst. ryb. Khoz. Okeanogr. 22: 135-163. (In Russian.)

## INSTRUCTIONS TO AUTHORS

## Based on

CONFERENCE OF BIOLOGICAL EDITORS, COMMITTEE ON FORM AND STYLE. I 960.
Style manual for biological journals. Washington: American Institute of Biological Sciences.

## MANUSCRIPT

To be typewritten, double spaced, with good margins, arranged in the following order: (1) Heading, consisting of informative but brief title, name(s) of author(s), address(es) of author(s), number of illustrations (plates, figures, enumerated maps and tables) in the article.
(2) Contents. (3) The main text, divided into principal divisions with major headings; subheadings to be used sparingly and enumeration of headings to be avoided. (4) Summary. (5) Acknowledgements. (6) References, as below. (7) Key to lettering of figures. (8) Explanation to plates.

## ILLUSTRATIONS

To be reducible to $12 \mathrm{~cm} \times 18 \mathrm{~cm}$ ( 19 cm including caption). A metric scale to appear with all photographs.

## REFERENCES

Harvard system (name and year) to be used: author's name and year of publication given in text; full references at the end of the article, arranged alphabetically by names, chronologically within each name, with suffixes $a, b$, etc. to the year for more than one paper by the same author in that year.
For books give title in italics, edition, volume number, place of publication, publisher.
For journal articles give title of article, title of journal in italics (abbreviated according to the World list of scientific periodicals. 4th ed. London: Butterworths, 1963), series in parentheses, volume number, part number (only if independently paged) in parentheses, pagination.

## Examples (note capitalization and punctuation)

bullough, W. S. 1960. Practical invertebrate anatomy. 2nd ed. London: Macmillan.
Fischer, P.-H. 1948. Données sur la résistance et de le vitalité des mollusques. F. Conch., Paris 88: $100-140$.
Fischer, P.-H., Duval, M. \& Raffy, A. i933. Etudes sur les échanges respiratoires des littorines. Archs Zool. exp. gén. 74: 627-634.
Kohn, A. J. 196oa. Ecological notes on Conus (Mollusca: Gastropoda) in the Trincomalee region of Ceylon. Ann. Mag. nat. Hist. (13) 2: 309-320.
Kонл, А. J. 196ob. Spawning behaviour, egg masses and larval development in Conus from the Indian Ocean. Bull. Bingham oceanogr. Coll. 17 (4): 1-51.
Thiele, J. igio. Mollusca: B. Polyplacophora, Gastropoda marina, Bivalvia. In schultze. L, Zoologische und anthropologische Ergebnisse einer Forschungsreise im westlichen und zentralen SuidAfrika. 4: 269-270. Jena: Fischer. Denkschr. med.-naturw. Ges. Fena 16: 269-270.

## ZOOLOGICAL NOMENCLATURE

To be governed by the rulings of the latest International code of zoological nomenclature issued by the International Trust for Zoological Nomenclature (particularly articles 22 and 51). The Harvard system of reference to be used in the synonymy lists, with the full references incorporated in the list at the end of the article, and not given in contracted form in the synonymy list.

## Example

Scalaria coronata Lamarck, 1816: pl. 451, figs $5 a, b$; Liste: 1 1. Turton, 1932: 80.

### 507.68

ANNALS OF THE SOUTH AFRICAN MUSEUM anNale van die SUid-afrikaanse museum

| Volume | $\mathbf{5 9}$ | Band |
| ---: | :---: | :--- |
| August | $\mathbf{1 9 7 2}$ | Augustus |
| Part | $\mathbf{9}$ | Deel |



## A LATE PLIOCENE RHINOCEROS FROM LANGEBAANWEG, CAPE PROVINCE

## By

D. A. HOOIJER

The ANNALS OF THE SOUTH AFRICAN MUSEUM are issued in parts at irregular intervals as material becomes available

Obtainable from the South African Museum, P.O. Box 61, Cape Town

Die ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM
word uitgegee in dele op ongereelde tye na beskikbaarheid van stof

Verkrygbaar van die Suid-Afrikaanse Museum, Posbus 61, Kaapstad
out of print/uit druk
1, 2(1, 3, 5, 7-8), 3(1-2, 5, t.-p.i.), 5(2, 5, 7-9), $6(\mathrm{I}, \quad \mathrm{t} .-\mathrm{p} . \mathrm{i}), \quad 7(\mathrm{I}-3), \quad 8, \quad 9(\mathrm{I}-2), \quad 1 \mathrm{I}(\mathrm{I})$, 1I(1-2, 5, 7, t.-p.i.), 24(2), 27, $31(1-3), 33$

Price of this part/Prys van hierdie deel $\mathrm{R}_{5,30}$

Trustees of the South African Museum © Trustees van die Suid-Afrikaanse Museum 1972

ISBN 0949940127

Court Road, Wynberg, Cape

In Suid-Afrika gedruk deur
Die Rustica-pers, Edms., Bpk.
Courtweg, Wynberg, Kaap

# A LATE PLIOCENE RHINOCEROS FROM LANGEBAANWEG, CAPE PROVINCE 

By

D. A. Hooìjer<br>Rijksmuseum van Natuurlijke Historie, Leiden

(With plates 21-34 and 5I tables)
[MS. accepted 14 February 1972]

## Contents



## Introduction

The rhinoceros remains described in the present paper are from the 'E' Quarry at Langebaanweg, situated approximately $32^{\circ} 5^{\circ}{ }^{\prime} \mathrm{S}, 18^{\circ} 9^{\prime} \mathrm{E}$ in the Sandveld region of the south-western Cape Province, some 105 km N.N.W. of Cape Town. They are more abundant than those of any other large mammal in the Langebaanweg fauna. The literature on the geology and palaeontology of the Langebaanweg deposits is reviewed in Hendey (1970a); the geological age is discussed in Hendey (1970b) and Maglio \& Hendey (1970). The 'E' Quarry rhinoceros has been cited as Diceros aff.bicornis, but I found it to be a very early Ceratotherium, the same as that from Kanapoi, Ekora and Lothagam-I in N.W. Kenya described as Ceratotherium praecox Hooijer \& Patterson (1972). This species is still very close to a Diceros like D. bicornis (L.) in some dental characters which take the eye even at a cursory look, such as the transversely placed proto- and metaloph, absence of medifossettes, well-developed paracone style, and angular antero-internal crown corners. In these as well as other characters the fossil teeth from Langebaanweg and those of $D$. bicornis differ from those of Ceratotherium simum (Burchell), which has obliquely placed proto- and metalophs, medifossettes, no paracone style, and rounded antero-internal crown angles. We believe that Ceratotherium praecox is directly ancestral to the living C. simum, and its occurrence at the Kenya sites, near the 4 million year level (Maglio 1970; Cooke \& Maglio 197r; Bishop 1971a: 511) is perfectly in accordance with the Late Pliocene age that is now becoming accepted for the Langebaanweg deposits.

Ann. S. Afr. Mus. 59 (9), 1972: 151-191, 14 pls, 51 tables.

Abbreviations used in this paper are:
K.N.M. Kenya National Museum
L.M. Leiden Museum
M.C.Z. Museum of Comparative Zoology, Harvard University
S.A.M. South African Museum

## Ceratotherium praecox Hooijer \& Patterson

Ceratotherium praecox Hooijer \& Patterson 1972: 19.
The present species was based on three incomplete skulls and mandibles with teeth, some fragments without teeth, an upper molar and an imperfect humerus from Late Pliocene sites in north-western Kenya. The Langebaanweg rhino collection comprises four upper dentitions, parts of three skulls and ten mandibles (mostly with teeth), ioo isolated upper and 50 isolated lower cheek teeth, 3 upper incisors, 20 deciduous cheek teeth, and 650 postcranial bones. The cranial and dental characters of the Langebaanweg rhinoceros are the same as those of the Kenya collection already described, but the Langebaanweg collection adds to our knowledge of the species information on the upper incisors and milk teeth which were unknown before, and the postcranial characters which were virtually unknown until the Langebaanweg material became available. The data provided in the present paper show the amount of individual variation within a single species of Pliocene rhinoceros. It is not saying too much now that $C$. praecox odontologically and osteologically is better known than its extant descendant, although, of course, its external characters are for ever lost to us.

The cranial characters of the present species are as follows: dorsal surface more concave, posterior portion less extended behind, occiput less posteriorly inclined, nuchal crest less thickened than in Ceratotherium simum. The premaxillae bear two incisors each, about 10 mm in diameter. The symphysial part of the mandible is narrower than in C. simum, and similar to $D$. bicornis. The premolars and molars (upper as well as lower) are more hypsodont than those in D. bicornis, but decidedly less so than in C. simum. The flattened ectolophs, marked protocone folds in the molars, strong internal cingula in the premolars, angular anterointernal corners of the crowns in premolars and molars alike, the posterior bulging of the protocones, which make up three-fifths of the internal crown faces, the medifossettes that rarely occur (mostly in $\mathrm{P}^{2-3}$ and $\mathrm{M}^{3}$, if at all), and the medisinus and postsinus depths being very nearly equal, all these are characters shared by the Kanapoi and the Langebaanweg C. praecox.

The species in question is rather Diceros-like in skull and dentition, the teeth differing in their relatively higher crowns, with a flattened ectoloph on which the paracone style is almost completely suppressed, the postsinus being very nearly as deep as the medisinus, and the posterior protocone bulge slightly more marked. In these points the Kanapoi and Langebaanweg rhinoceros is evolving toward the Quaternary Ceratotherium simum, in which the crown height
is still greater, the paracone style completely suppressed and the parastyle raised, forming a concave area on the ectoloph where the paracone style had been, medifossettes are common, formed by the union of crochet and crista, postsinus and medisinus are equal in depth, the protocone bulge is more marked, the protoloph is obliquely placed and the antero-internal crown angles are rounded. In the early subspecies C. simum germanoafricanum (Hilzheimer), which is indistinguishable from the extant $C$. simum simum cranially, and which occurs at Laetolil, the basal Olduvai Beds, and Chemeron Formation locality J.M. 90 ( $=9 \mathrm{I}$ ), the crowns are not quite so hypsodont and the metaloph is still transverse in its course rather than oblique as in the modern form, although the rounded antero-internal crown angles and the medifossettes of C. simum germanoafricanum are as in C. simum simum. In my earlier paper on Pleistocene East African rhinoceroses (Hooijer 1969), published at a time when I had not yet studied the material from Kanapoi, Lothagam-I and Ekora, I referred specimens from the Chemeron Formation, locality J.M.507, and from the Mursi Formation of the Omo Basin (=lower level of the Omo collection made by Mr. R. E. F. Leakey in 1967) to C. simum germanoafricanum which I now recognize as belonging to Ceratotherium praecox instead; this will be dealt with in the final section of this paper. The discovery of Ceratotherium praecox vindicates the view of Thenius (1955) that Ceratotherium split off from the Diceros stock sometime in the Pliocene.

## Dentition and skull

The individually youngest upper dentition, $\mathrm{Lr}_{3} 3035$, comprises $\mathrm{P}^{2}-\mathrm{M}^{3}$ from the right side as well as $\mathrm{M}^{1-3} \sin$. (Pl. 21). The crowns of $\mathrm{P}^{3}$ and $\mathrm{M}^{2}$ dext. only are virtually complete. There was a $\mathrm{DM}^{1}$ or $\mathrm{P}^{1}$ as there is an anterior facet on $\mathrm{P}^{2}$.
$\mathrm{P}^{2}$ is worn down to a height of 28 mm externally. The external enamel layer is missing for the most part; only the metastyle portions remain. There is a very marked internal cingulum, rising on the protocone and the hypocone from its lowest point at the medisinus entrance. The internal portions of protoloph and metaloph are connected at their bases by a small ridge; there is a small pit between it and the internal cingulum. The medisinus is slightly deeper than the postsinus, and there is a very small crochet, hardly more than a point.
$\mathbf{P}^{3}$, worn to 45 mm from the base externally, has a very prominent internal cingulum, reaching its lowest point at the entrance to the medisinus, which is narrow and V-shaped. There is a very weak crochet, and no crista or antecrochet. Medisinus and postsinus are equal in depth. The ectoloph is flattened, with a weak cingulum; there is no parastyle fold or paracone style. The protoloph is hardly indented anteriorly, but there is a vertical groove in the metaloph marking off the hypocone.
$\mathrm{P}^{4}$, with an external height as worn of 60 mm , has the flattened ectoloph detached from the remainder of the crown, which shows the narrow, V-shaped
medisinus entrance, the heavy internal cingulum, weak crochet, and the medisinus depth equal to that of the postsinus, as in $\mathrm{P}^{3}$. An internal view of $\mathrm{P}^{\mathbf{2 - 4}}$ dext. of $\mathrm{L}_{13035}$ is given in Plate 25, top.
$\mathrm{M}^{1}$, the right of which lacks most of ectoloph and protoloph, and the left of which is entire but for the antero-external angle, is worn to a height of 52 mm externally. The lingual entrance to the medisinus is V -shaped, and there is an anterior fold in the metaloph marking off the hypocone. This molar, in contrast to the premolars, has a deep fold anteriorly in the protoloph marking off the protocone (the protocone fold), a strong crochet extending all across the medisinus, not receding near the base, and the internal cingulum hardly marked except along the protocone and for a tubercle at the entrance to the medisinus. The inner portion of the protoloph is recurved backward, forming three-fifths of the internal surface. The ectoloph is flattened, without styles, and medisinus and postsinus are of the same depth.
$\mathrm{M}^{2}$, nearly entire on both sides, is worn to 75 mm from the external base. This is clearly a hypsodont tooth, the anteroposterior diameter of the crown being 62 mm externally. A weak paracone style is seen in the upper part of the crown only, to 60 mm from the base, flattening out further rootward. There is no groove marking off the hypocone, but the description of $\mathrm{M}^{1}$ would otherwise fit the $\mathrm{M}^{2}$.
$\mathrm{M}^{3}$ of dentition $\mathrm{Li}_{3} 035$, both incomplete behind, are 90 mm high as worn and the length of the outer surface is about 75 mm . The marked protocone fold, internal protocone cingulum, and strong, even bifid crochet, are as in the other molars of this individual. The paracone style is weak but discernible, reaching from the top of the crown to approximately 50 mm from the crown base.

Another upper dentition, $\mathrm{L}_{2} 5^{1}$ 9, likewise consists of isolated teeth, which are $\mathrm{P}^{2}-\mathrm{M}^{3}$ dext. and $\mathrm{P}^{3}-\mathrm{M}^{1} \sin$. (Pl. 22). They are rather well preserved although a number of crown angles are missing.
$\mathrm{P}^{2}$ is just 20 mm high as worn externally. The medisinus is still open internally. It shows a crochet united with a small crista so that a medifossette is formed. The same feature is seen in both $\mathrm{P}^{3}$, which are worn down externally to 35 mm from the crown base. The protocone fold, which is preserved only in $\mathrm{P}^{3}$ dext., is more marked than that in $\mathrm{P}^{3}$ of dentition Li3035. The postsinus is almost as deep as the medisinus; the ectoloph is just as flattened, with a weak cingulum, and the internal cingulum is just as prominent as that in $\mathrm{P}^{3}$ of Li3035. $\mathrm{P}^{4}$, present on both sides in L2519, is 45 mm high as worn externally, and the right specimen has an imperfectly formed medifossette, while the left has a bifid crochet and a small crista that do not join. There is no difference in depth between the postsinus and the medisinus, and the ectoloph and the internal cingulum are as in $\mathrm{P}^{3}$.
$\mathrm{M}^{1}$, on both sides, has a particularly powerful crochet, nearly twice as thick as that in $\mathrm{M}^{1}$ of $\mathrm{L}_{13035}$, but no crista. The external crown height is 35 mm , as worn. In addition to the anterior protocone fold there is an internal indenta-
tion in the protocone as seen in $\mathrm{M}^{1} \sin$. The posterior bulging of the protocone is such that it forms three-fifths of the internal crown face. There is a distinct hypocone fold, visible in both the right and the left molar. The internal cingulum is not preserved in these molars but externally there is a very weak cingulum, mainly posteriorly, as in all the molars. The ectoloph is flattened, and the narrow postsinus appears to be slightly less deep than the medisinus.

The $\mathrm{M}^{2}$ dext. of L2519, 50 mm high as worn, has a crochet that is not thickened; it is recurved outward at the apex but the crista is just barely indicated and no medifossette is formed. The posterior bulging of the protocone is such that it occupies 27 out of the 45 mm long internal basal anteroposterior diameter. The lingual entrance to the medisinus is V -shaped, and the protocone is indented lingually. The portion of the crown that would have shown the protocone fold is missing; the hypocone fold is weakly developed, and so is the internal cingulum; the ectoloph is flattened, without styles. The two sinuses are equally deep.
$\mathrm{M}^{3}$ dext. of L2519 lacks most of the outer surface (ecto-metaloph) so that no measurements can be given. It has a crown height as worn of about 60 mm . There is a very marked protocone fold, and a weak cingulum on the depressed internal surface of the protocone. The crochet is well-developed and there is a crista, too. These projections, however, remain separate down to the bottom of the medisinus. On this rather worn molar there is no trace of a paracone style such as we see on less worn specimens; the paracone style is no longer visible in the basal $50-60 \mathrm{~mm}$ of the crown.

The next upper dentition to be described is Li3747 (Pl. 23). Of this set of teeth the small anterior premolar $\mathrm{P}^{1}$, or a persisting $\mathrm{DM}^{1}$, is preserved on the right side, as the teeth are still in situ in the maxillary. It is about 23 mm anteroposteriorly, and about 20 mm transversely; nothing can be said about its structure as the crown is worn flat.
$\mathrm{P}^{2}$ is nearly entire on both sides. Although the worn crown height is the same as that in $\mathrm{P}^{2}$ of L25 $19(20 \mathrm{~mm})$ the valley between protocone and hypocone is closed as wear has reached the bottom of the sinus in between. There is a rather strong internal cingulum, and a pit is formed between it and the joint bases of proto- and hypocone, as in $\mathrm{P}^{2}$ of $\mathrm{Li}_{3} \mathbf{0} 35$. Postsinus and medisinus are of equal depth. There is a crochet but no crista.
$\mathrm{P}^{3}$ lacks the entire outer surface on the left side, and has only the anteroexternal angle on the right. The protocone fold is weakly developed. The internal cingulum is very marked, continuous with that on the anterior surface, and it carries a series of tubercles. It extends all along the protocone, reaching its lowest point at the narrow medisinus entrance, and rises along the hypocone, i.e. the same development that we noticed in the premolars of the two upper dentitions dealt with above. There is only a crochet, which is not very prominent, making the central portion of the medisinus rather wide. The depth of this portion of the medisinus is the same as that of the postsinus.
$P^{4}$, the worn ectoloph height of which is 40 mm , is rather damaged on the left but well preserved on the right side. There is a paracone style, which is rather more developed than that in the less worn dentitions Li 3035 and L2519; in these teeth there is no trace left of the paracone style, but in $\mathrm{LI}_{3747}$ it continues to about 20 mm from the crown base. There is a weak external cingulum along the posterior moiety of the ectoloph. The posterior bulging of the protocone occupies three-fifths of the internal surface of the crown. The internal cingulum is less developed, and the crochet more prominent than that in $\mathrm{P}^{3}$. The protocone fold is hardly shown. The medisinus is as deep as the postsinus and has a narrow, V-shaped entrance.
$\mathrm{M}^{1}$, which is between 20 and 25 mm in worn ectoloph height, has the protocone fold well marked. The protocone takes up 30 out of the 50 mm of internal anteroposterior crown diameter, and is slightly indented internally. The bottom of the narrow internal medisinus entrance is almost reached by wear, but its central portion is still about 15 mm deep, which is also the depth of the postsinus. The very thick crochet is free from the ectoloph at the level of wear. In $\mathrm{M}^{1}$ dext. it would have closed off a medifossette with the small crista if wear had proceeded some 5 mm more, but in $\mathrm{M}^{1} \mathrm{sin}$. no medifossette would have been formed in this way. In this advanced stage of wear no trace remains of the paracone style; the internal cingulum is so weak as to be practically absent.
$\mathrm{M}^{2}$ is 40 mm high at the worn ectoloph. $\mathrm{M}^{2}$ dext. has a vertical fracture in the ectoloph, but the external surface of $\mathrm{M}^{2}$ sin. is undamaged although detached from the remainder of the crown. The paracone style is shown as a weak bulge only along the worn edge of the crown. The protocone fold is very distinct, and the internal indentation of the protocone shows just as it does in the $\mathrm{M}^{1}$. The protocone occupies 40 out of the 65 mm of internal anteroposterior crown diameter. The crochet is narrower than that in $\mathrm{M}^{1}$ and does remain free at its apex so that no medifossette is formed. There is hardly any trace of an internal cingulum.
$\mathrm{M}^{3}$, incomplete on both sides, has the external surface worn down to 50 mm ; the paracone style is shown only in the apical 15 mm . The anterior fold, internal indentation, and posterior bulging of the protocone are as in $\mathrm{M}^{2}$. The crochet extends all across the medisinus but does not close off a medifossette.

Whereas the two dentitions first described (Li3035 and L2519) are rather similar in dimensions (see Table I) dentition Li 3747 is larger, but there are no significant differences in structure. The only point worth making is that the paracone style is slightly more marked in these large teeth than in those earlier described.

A crushed skull, L6658, has a good portion of the palate with $\mathrm{P}^{4}-\mathrm{M}^{2}$ dext. and $\mathrm{P}^{2}-\mathrm{M}^{2}$ sin., and the two last molars detached (Pl. 24). The dental dimensions are more or less intermediate between those of L2519 and Li3747
(Table i). There is a small portion of the anteriormost premolar, on the left side. $\mathrm{P}^{2} \sin$. is incomplete internally and much worn down: the ectoloph height is reduced to 15 mm , and the medisinus is cut off from the lingual border. There is a tiny medifossette, which would have disappeared with a little more wear. $\mathrm{P}^{3} \sin$. has a medifossette too; its external height as worn is almost 30 mm , and no paracone style is shown. The medisinus is just closed off lingually. The internal cingulum, with its lowest point at the junction of protocone and hypocone, is well developed. It is slightly less marked in $\mathrm{P}^{4}$, present on both sides, with a worn ectoloph height of 40 mm . The crochet is bifid in $\mathrm{P}^{4}$ dext., and single in $\mathrm{P}^{4} \sin$. The premolars $\mathrm{P}^{3}$ and $\mathrm{P}^{4}$ agree in the postsinus being as deep as the medisinus, the posterior bulging of the protocone forming three-fifths of the internal surface (21 out of 35 mm in $\mathrm{P}^{3}$, and 27 out of 45 mm in $\mathrm{P}^{4}$ ), and in their weak protocone folds.
$\mathrm{M}^{\mathbf{1}}$, lacking the antero-external angle on both sides, is some 25 mm high as worn externally. The protocone fold is very marked, and there is an internal indentation in the protocone, which occupies three-fifths of the internal border. The lingual cingulum is weakly developed, the lingual medisinus entrance very narrow, and the crochet is rather thick, as usual in first molars. $\mathrm{M}^{2}$, the right of which is partially embedded in the bone, has a worn ectoloph height of 45 mm , and does not show the paracone style any more. The characters are those of $\mathrm{M}^{1}$; only the crochet is more slender. The $\mathrm{M}^{3}$, of which the left is virtually complete, is 60 mm high as worn externally. The paracone style can be traced in the apical I5-20 mm only. The crochet extends all across the medisinus, and joins the posterior wall of the protoloph, thus cutting off the external portion of the medisinus. The protocone fold is strongly marked, the internal cingulum very weak.

Table i
Measurements of upper teeth of Ceratotherium praecox from Langebaanweg (mm)

| No. of specimen | Li 3035 | L25 19 | Li3747 | L6658 |
| :---: | :---: | :---: | :---: | :---: |
| P2, ant.post. | c. 35 | 33 | 36 | 32 |
| ant.transv. | - | 40 | 44 | 37 |
| post.transv. | c. 45 | - | 50 | $40+$ |
| $\mathrm{P}^{3}$, ant.post. | 45 | 46 | - | 43 |
| ant.transv. | 58 | 57 | 66 | 62 |
| post.transv. | 54 | - | - | 58 |
| $\mathrm{P}^{4}$, ant.post. | 48 | 51 | 53 | 50 |
| ant.transv. | - | 65 | 75 | 67 |
| post.transv. | - | 60 | 73 | 63 |
| $\mathrm{M}^{1}$, ant.post. | - | 58 | c. 6 o | 57 |
| ant.transv. | 70 | 70 | 80 | 73 |
| post.transv. | 64 | c. 64 | 72 |  |
| $\mathrm{M}^{2}$, ant.post. | 62 | 64 | 68 | 64 |
| ant.transv. | 72 | 72 | 82 | 74 |
| post.transv. | 67 | 65 | c. 70 | 69 |
| $\mathrm{M}^{3}$, ant.post.(int.) | 65 | - | 67 | 66 |
| ant.transv. | 72 | - | 78 | 73 |
| length outer surface | c. 75 | - | 79 | 83 |
| Length $\mathrm{P}^{2}-\mathrm{M}^{3}$ | c. 300 | c. 300 | 330 | 305 |
| Length $\mathrm{P}^{2}-\mathrm{P}^{4}$ | c. 135 | 130 | 135 | 125 |
| Length $\mathrm{P}^{4}-\mathrm{M}^{3}$ | c. 230 | c. 225 | 245 | 235 |

To dentition $\mathrm{LI}_{3} 3035$ belongs a skull portion, giving a zygomatic width of 390 mm , very near to the maximum, observed by Heller (1913) in modern Ceratotherium simum, viz., 384 mm . The length from $\mathrm{M}^{3}$ to the back of the postglenoid process is $c .220 \mathrm{~mm}$, slightly less than the length $\mathrm{P}^{4}-\mathrm{M}^{3}(c .230 \mathrm{~mm})$. In subadult skulls of $C$. simum in which $\mathrm{M}^{3}$ has not erupted yet the length $\mathrm{P}^{4}-\mathrm{M}^{3}$ exceeds the postdental length from $\mathrm{M}^{3}$ to the back of the postglenoid process (e.g., S.A.M. $2138 \mathrm{I}: \mathrm{P}^{4}-\mathrm{M}^{3} c .225 \mathrm{~mm}$; postdental length c. 190 mm ). In skulls with $\mathrm{M}^{3}$ slightly worn the two lengths are subequal (S.A.M. 21382: $\mathrm{P}^{4}-\mathrm{M}^{3} 215 \mathrm{~mm}$; postdental length 200 mm ; S.A.M. 21379: $\mathrm{P}^{4}-\mathrm{M}^{3} 225 \mathrm{~mm}$; postdental length 210 mm ). In fully adult $C$. simum skulls with $\mathrm{M}^{3}$ well worn down the length $\mathrm{P}^{4}-\mathrm{M}^{3}$ is exceeded by the postdental length (M.C.Z., Dept Mamm. 24917 and $34850: \mathrm{P}^{4}-\mathrm{M}^{3}$ 190-205 mm; postdental length $270-275 \mathrm{~mm}$ ). In the holotype skull of Ceratotherium praecox from Kanapoi, which is quite adult, the postdental length is the larger of the two, though not to the extent seen in the recent species (Kanapoi $\mathrm{P}^{4}-\mathrm{M}^{3} 205 \mathrm{~mm}$; postdental length $230-250 \mathrm{~mm}$ ).

The premaxillaries of $\mathrm{Lr}_{3} 3747$ are preserved, and they show two alveoli on each side, one behind the other. The anterior alveolus holds a tooth crown that is unerupted, about 12 mm anteroposteriorly and 9 mm transversely. The posterior alveolus is of the same size but empty; its depth is only 7 mm . The specimens are shown in Plate 28, top. The occurrence of rudimentary upper incisors in C. praecox is interesting, as the recent species of Ceratotherium no longer shows them. An isolated $\mathrm{I}^{1}$ has a rounded crown and a strong, posteriorly recurved root. The height of the crown and root combined is 37 mm , while the crown diameter is II mm ( Pl .28 , top right).

The nasal horn boss of L6658 is crushed, but its width is about 180 mm . The nasal portion of another skull, L2520, is 180 mm wide at the horn boss; this width is $170-208 \mathrm{~mm}$ in adult males, and $146-173 \mathrm{~mm}$ in adult females of recent Ceratotherium simum (Heller 1913). The frontal region of the skull L2520 shows the second horn boss, on the frontals, but the upper borders of the orbits are not preserved. Dorsal views of skulls L2520 and L6658 are given in Plate 27.

Skull $\mathrm{L}_{3} 347$ is broken in many pieces; the right half of the top of the skull has been reassembled (Pl. 26). Although the angle between the dorsal and the occipital planes cannot be exactly measured it is approximately $60^{\circ}$. This is $65^{\circ}$ in skull K.N.M. KP3o from Kanapoi, against $65-80^{\circ}$ in Diceros bicornis, and $45-50^{\circ}$ in Ceratotherium simum. These figures tend to show that in the fossil C. praecox the occiput is less posteriorly inclined relative to the dorsal surface than in C. simum, and rather resembles D. bicornis in this respect. In keeping with the less marked posterior inclination of the occiput, the nuchal crest in C. praecox is not as thickened as it is in modern C. simum, in which it is quite massive, overhanging the occipital condyles.

The mandible Li3035 is nearly entire, lacking only part of the ventral border of the left horizontal ramus and the right coronoid process (PI. 30).
$P_{3}-M_{3}$ dext. and $P_{2}-M_{3} \sin$. are in situ; an internal view of the right ramus is given in Plate 3I. Mandible Lir 849 has the symphysis as well as $\mathrm{P}_{2}-\mathrm{M}_{3}$ dext., somewhat more worn than $\mathrm{Li}_{3} 035$ (Pl. 32, right). There is further a symphysial portion of the mandible, L6o58, with the alveoli for $\mathrm{P}_{2}$ (Pl. 33). There appear to be small alveoli for incisors in the symphyses examined, but none of these elements has been found. The premolars and molars of the Langebaanweg Ceratotherium do not show the tendency toward obliqueness of the lophids, or that toward fossettid formation seen in Ceratotherium simum.

The length of the mandible, $\mathrm{Li}_{13035}$, is 570 mm ; this measurement is $565-635 \mathrm{~mm}$ in adult males, and $550-588 \mathrm{~mm}$ in adult females of recent C. simum (Heller 1913). The length of the symphysis is 125 mm in $\mathrm{Li}_{3} 3035$ and
 females of $C$. simum. The fossil specimens agree with the recent in both length measurements. However, the width at the symphysis is 60 mm in $\mathrm{LiI}_{1849}$ and 65 mm in L6058, which is decidedly less than that in recent males ( $96-\mathrm{r} 25 \mathrm{~mm}$ ) and females ( $9 \mathrm{I}-1 \mathrm{II} \mathrm{mm}$ ) of $C$. simum (Heller 1913). It follows from this that in C. praecox the symphysis is relatively (and absolutely) narrower than in C. simum. It is in $D$. bicornis that we find such a narrow symphysis: S.A.M. 35658 has a length of symphysis of 105 mm by a width at symphysis of only 45 mm .

In the height at $\mathrm{M}_{1}, 125 \mathrm{~mm}$, the fossil mandible Li 3035 equals C. simum (S.A.M. 21 1379), whereas in D. bicornis (S.A.M. 35658) this height is only 85 mm . The distance from the dental foramen to the base of the posteromedial articular surface is 160 mm in Li 3035, against 230 mm in C. simum and 135 mm in D. bicornis; the jaw orientation in the fossil was evidently nearer to that in D. bicornis than to that in C. simum. The condyles in Li 3035 are not entire, but the condylar area appears to be more massive, and wider below the condyle than in C. simum. The medial surface below the condyle is more hollowed than in either of the two living species. These are also the characters of the Kanapoi C. praecox.

Table 2
Measurements of lower teeth of Ceratotherium praecox (mm)

| No. of specimen | $\mathrm{Li}_{3} \mathrm{O} 35$ | Lir 849 |  | Li3035 | Lir 849 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| $\mathrm{P}_{2}$, ant.post. | 30 | - | $\mathrm{M}_{1}$, ant.post. | (54) | - |
| ant.transv. | 15 | 17 | ant.transv. | 32 | 35 |
| post.transv. | 17 | 17 | post.transv. | 32 | 37 |
| $\mathrm{P}_{3}$, ant.post. | - | - | $\mathrm{M}_{2}$, ant.post. | 63 | 56 |
| ant.transv. | - | 24 | ant.transv. | 36 | 37 |
| post.transv. | 27 | - | post.transv. | 34 | 37 |
| $\mathrm{P}_{4}$, ant.post. | 48 | 45 | $\mathrm{M}_{3}$, ant.post. | c. 64 | 60 |
| ant.transv. | 30 | 31 | ant.transv. | 35 | 35 |
| post.transv. | 32 | 32 | post.transv. | 32 | 34 |
|  |  |  | Length $\mathrm{P}_{2}-\mathrm{M}_{3}$ | 290 | 290 |
|  |  |  | Length $\mathbf{M}_{\mathbf{1}}-\mathbf{M}_{3}$ | 175 | 170 |

Dental measurements of the two mandibles are given in Table 2. Isolated lower teeth to be recorded further on considerably expand the variation ranges in size.

Among the isolated teeth from Langebaanweg there are a few unworn or very slightly worn crowns showing the degree of hypsodonty; these will be mentioned in the following pages.

An unworn P ${ }^{4} \sin$., Li 3760 (Pl. 25, bottom left) has an ectoloph height of 90 mm by a greatest anteroposterior length of the ectoloph, in the apical third of the crown, of 55 mm , which gives a height/length index of 164 . An unworn recent $\mathrm{P}^{4}$ of Diceros bicornis (Leiden Museum, cat.ost.e) has the same greatest ectoloph length by an ectoloph height of 80 mm , giving a height/length index of 145 . On the other hand, an unworn $\mathrm{P}^{4}$ of recent Ceratotherium simum (S.A.M. 21382 ) has an ectoloph height of 103 mm by a greatest anteroposterior ectoloph length of 46 mm , giving a height/length index of 224 .

Among the last upper molars in particular there are several nearly unworn crowns, as follows: an $M^{3}$ dext., L6696 (Pl. 25, middle), an $\mathrm{M}^{3} \sin$. , L71o6, of the same individual; an $\mathrm{M}^{3}$ dext., L6291 (Pl. 25, middle), an $\mathrm{M}^{3} \sin ., \mathrm{L}^{2} 46 \mathrm{I}$, of the same individual as L6291; an unworn $\mathrm{M}^{3}$ dext., L6638, incomplete basally and a very slightly worn $\mathrm{M}^{2}$ sin., L6636. In L6696 the total height of the outer surface is 94 mm by a length of the outer surface of 78 mm , giving a height/length index of 121 . The paracone style is a narrow ridge, which fades away in the basal 35 mm of the ectoloph. L6291 has a height of the outer surface of 85 mm ; the length of the outer surface is 70 mm , giving a height/length index of 121. Finally, L6638 has a height of the outer surface of 96 mm by a length of this surface of approximately 80 mm ; height/length index c.120. This is just about the height/length index of $\mathrm{M}^{3}$ in modern Diceros bicornis (outer surface height 64 mm , outer surface length 54 mm , height/length index 119: Hooijer 1969:87), but the Pleistocene Diceros bicornis from the Omo Beds is lower-crowned than the living form (two specimens of $\mathrm{M}^{3}$, height of unworn outer surface $56-59 \mathrm{~mm}$, length of outer surface $55-58 \mathrm{~mm}$, height/length index 102: Hooijer 1969:87). In modern Ceratotherium simum $\mathbf{M}^{3}$ is $120-130 \mathrm{~mm}$ high (Dietrich 1945:59), and a slightly worn recent M ${ }^{3}$ (S.A.M. 21379) is Ioo mm high at the outer surface, while an unworn recent $\mathrm{M}^{3}$ (S.A.M. 21382), the outer surface of which is not quite fully calcified at base, is just over 100 mm high at the incompletely formed external surface. In these recent $\mathrm{M}^{3}$ there is no paracone style but a depression behind the parastyle instead.

The $\mathrm{M}^{2}$ from Langebaanweg, L6636, slightly worn, has an ectoloph height at the metaloph origin of 98 mm by a greatest anteroposterior ectoloph length of 73 mm ; its height/length index is 134 . The hypsodonty of Ceratotherium praecox $\mathrm{M}^{2}$ has already been demonstrated in a slightly worn $\mathrm{M}^{2}$ from Lothagam-I (K.N.M. LT89 in Hooijer \& Patterson 1972) that has an ectoloph height at the metaloph origin of 74 mm by a greatest anteroposterior ectoloph length of 63 mm , giving a height/length index of 117 . In modern Diceros bicornis $\mathrm{M}^{2}$ (two specimens) the ectoloph is not so much higher than wide, although the difference is small: M.C.Z., Dept Mamm., no. 51479, height at metaloph origin 56 mm , length 54 mm , height/length index 104, and Leiden Museum, cat.ost.b, height 74 mm , length 68 mm , height/length index 109.

Since the Omo $\mathrm{M}^{3}$ of Diceros bicornis (Pleistocene) is less hypsodont than the modern $\mathrm{M}^{3}$, the same doubtless holds for the $\mathrm{M}^{2}$. In the fossil $\mathrm{M}^{2}$ from Langebaanweg the paracone style is present on the apical half of the crown only.

There are two very slightly worn $\mathrm{P}_{4}$ in the Langebaanweg collection, L5356 and L6693, both from the right side. They are rather similar in dimensions (Table 3), and intermediate in height/length indices between recent Diceros bicornis (first column) and recent Ceratotherium simum (last column of Table 3). The discrepancy in height/length indices is the same as that found in $\mathrm{P}^{4}$.

| Table 3 |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| Measurements of $\mathrm{P}_{4}$ in Diceros and Ceratotherium (mm) |  |  |  |  |
|  | D. bicornis | Ceratot | praecox | C. simum |
| No. of specimen | L.M.cat.e | $\mathrm{L}_{535} 6$ | L6693 | S.A.M. 21382 |
| Greatest length of outer surface | 44 | 49 | 48 | 47 |
| Height of metalophid | 63 | 74 | 70 | 94 |
| Height of hypolophid (b) | 55 | 68 | 65 | 88 |
| Height/length index (a) | 143 | 151 | 146 | 200 |
| Height/length index (b) | 125 | 139 | 135 | 187 |

Four isolated lower molars, either $\mathrm{M}_{1}$ or $\mathrm{M}_{2}$, are unworn or very slightly worn. These are L6667 and L 2526 , from the right side, and L6664 and L668o, from the left. The height of the anterior (meta-) lophid, taken from the external base of the crown, varies from 70 to 80 mm ; the hypolophid height varies between the same limits. Unworn $\mathrm{M}_{1-2}$ of recent D. bicornis are $c .55$ to 65 mm high, and those of $C$. simum $c .80$ to 100 mm .

The isolated upper premolars and molars from Langebaanweg are enumerated in the tables that follow.

Of $\mathrm{P}^{2}$ we have nine specimens (Table 4) the first three of which are from the right side, the others from the left. There is a double crochet in L6649, a medifossette in L6751, L4750, and L6648, while a bifid crochet is shown in L6623.

$\mathrm{P}^{3}$ is represented by nineteen specimens (Table 5) the first eight of which are from the right side, the others (starting with $\mathrm{LII}_{180}$ ) from the left. In L6630 there is seen a slender crista extending to the tip of the crochet; the internal cingulum is rather weak in this specimen as well as in L6627. L6625 and $\mathrm{L}_{5} 665$ have a bifid crochet, $\mathrm{L}_{5} 665$ has in addition a very small crista.

| Table 5 |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Measurements of $\mathrm{P}^{3}$ of Ceratotherium praecox (mm) |  |  |  |  |  |  |  |
| No. of specimen | L6629 | L6630 | L663I | L5444 | L6625 | L6295 | L6646 |
| Ant.post. | 48 | c. 50 | 47 | 40 | 43 | 40 | c. 45 |
| Ant.transv. | 65 | c. 67 | 64 | 59 | 60 | 60 | 62 |
| Post.transv. | 58 | - | 56 | 52 | 55 | 57 | 58 |
| No. of specimen | $\mathrm{Li}_{3}{ }_{7} 65$ | Lir8oi | L6639 | Lir 1996 | L6627 | L5695 |  |
| Ant.post. | 45 | 45 | 43 | - | 47 | 44 |  |
| Ant.transv. | 58 | 64 | 62 | 60 | 68 | 61 |  |
| Post.transv. | 51 | 57 | 58 | 56 | 61 | 58 |  |
| No. of specimen | $\mathrm{L}_{5671}$ | L9114 | L545 ${ }^{\text {I }}$ | L6640 | $\mathrm{L}_{5} 665$ | Li3099 | Aterir |
| Ant.post. | 45 | - | 50 | 46 | 48 | 44 | 45 |
| Ant.transv. | 60 | 64 | 66 | 63 | 65 | 61 | 58 |
| Post.transv. | 54 | 58 | 63 | 57 | 59 | 56 | 54 |

There are twenty specimens of $\mathrm{P}^{4}$ (Table 6) the first eleven of which are from the right side, the others (from Liris2 onward) from the left. L2525 has a crista joining the crochet. Lirin2 belongs to the same individual as Lirini, has a double crochet the lateral part of which is joined to a crista, thus forming a medifossette (Pl. 29, bottom). L6655, slightly worn, shows a double crochet and a crista (Pl. 29, bottom). Li376o shows the full height of the ectoloph, with a height/length index of 164 (Pl. 25, bottom left). Medifossette formation is very rare in $\mathrm{P}^{4}$ and $\mathrm{M}^{1-2}$, one in twenty or three in forty Langebaanweg teeth.

Table 6
Measurements of $\mathrm{P}^{4}$ of Ceratotherium praecox (mm)

| No. of specimen | L6717 | L2525 | L6652 | L6299 | Lir67 | L66ı9 | L6635 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ant.post. | 51 | 54 | 49 | 49 | 51 | 50 | 50 |
| Ant. transv. | 70 | 74 | 70 | 70 | 75 | 71 | 76 |
| Post.transv. | 63 | 69 | 61 | 61 | 67 | 61 | 70 |
| No. of specimen | L6739 | L5696 | L3454 | Lilitir | LiH ${ }^{2} 2$ | L66ı8 |  |
| Ant.post. | c. 57 | 51 | 47 | 48 | $4^{8}$ | c. 52 |  |
| Ant.transv. | 75 | 69 | 67 | 68 | 68 | 71 |  |
| Post.transv. | 64 | 63 | 56 | 62 | 62 | 66 |  |
| No. of specimen | L663 ${ }_{2}$ | L6296 | L4612 | L6622 | Li3760 | Li3099 |  |
| Ant.post. | - | 48 | 50 | c. 50 | 47 | 48 |  |
| Ant.transv. | 69 | 70 | 71 | 68 | 68 | 64 |  |
| Post.transv. | 63 | 64 | 68 | 63 | 62 | 60 |  |

Table 7
Measurements of $\mathrm{M}^{1}$ of Ceratotherium praecox (mm)

| No. of specimen | L6626 | L6624 | L6703 | L5445 | L9113 | L6628 | LII 798 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ant.post. | 58 | 55 | - | 57 | 60 | 52 | 55 |
| Ant.transv. | 73 | 72 | 73 | 72 | 74 | 71 | 72 |
| Post.transv. | 68 | 67 | 66 | 65 | 65 | 63 | 67 |
| No. of specimen | $\mathrm{L}_{5912}$ | L6293 | Li 2039 | L53II | L6647 | L5418 | L662 I |
| Ant.post. | 55 | - | c. 52 | 56 | 56 | 61 | - |
| Ant.transv. | 68 | 67 | 66 | 69 | 73 | 72 | 75 |
| Post.transv. | 65 | 60 | 61 | 65 | 68 | 68 | - |
| No. of specimen | L6465 | L4749 | L5919 |  |  |  |  |
| Ant.post. | c. 53 | 54 | 58 |  |  |  |  |
| Ant.transv. | 69 | 70 | 70 |  |  |  |  |
| Post.transv. | 66 | 61 | 65 |  |  |  |  |

$\mathrm{M}^{1}$ is represented by seventeen specimens (Table 7) the first eight of which are from the right side, the left specimens beginning with L6293. Medifossettes are not formed; L6626 has a double crochet (Pl. 28, bottom right).

There are twenty-four specimens of $\mathrm{M}^{2}$ (Table 8) the first ten of which are from the right side, the remaining specimens (starting with L6636) from the left. A true medifossette, formed by the union of crochet and crista, is shown only in L91 $16, L_{591} 6$ (external surface broken off: Pl. 28), and Liog83. A small crista is seen in $\mathrm{L}_{5917} \mathrm{~F}^{2} \mathrm{~L}^{2} \mathrm{I}_{7} 7$ (in which the crochet makes a contact with a small projection on the posterior face of the ectoloph: Pl. 28), L6746, L6654, L664i, L6644A, and Li2360. L91 18 consists of ectoloph and crochet only; the crista is in contact with the crochet apically (Pl. 29). L6636 is the specimen with the ectoloph slightly worn, and a height/length index of 134 , already referred to above.

Table 8
Measurements of $\mathrm{M}^{2}$ of Ceratotherium praecox (mm)

| No. of specimen | $\mathrm{L}_{5917}$ | L6617 | L6694 | L9116 | L6631 | L6634 | L5911 | L6746 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ant.post. | c. 50 | 67 | c. 62 | 62 | c. 60 | 61 | 63 | c. 64 |
| Ant.transv. | 75 | 82 | 82 | 75 | 79 | 77 | 79 | 74 |
| Post.transv. | 68 | 75 | 69 | 73 | 71 | 72 |  |  |
| No. of specimen | L6637 | L6643 | L6636 | L5916 | L9118 | L6654 | L6645 | Liog83 |
| Ant.post. |  | - | c. 60 | - | - | c. 55 | 56 | 55 |
| Ant.transv. | 80 | 80 | 73 | - | - | 76 | 78 | 71 |
| Post.transv. | 72 | 73 | 66 | - | - | 68 | 67 | 67 |
| No. of specimen | L6644 B | L6654 | L664 ${ }^{1}$ | L6644 A | Lir 898 | Li2360 | L6653 | L9115 |
| Ant.post. | c. 62 | 63 | 67 | 65 | 65 | c. 55 | c. 55 | - |
| Ant.transv. | 74 | 70 | 75 | 79 | 77 | 73 | 71 | 77 |
| Post.transv. | 67 | 65 | 67 | 72 | 68 | 70 | 65 | 68 |

We have seventeen specimens of $\mathrm{M}^{3}$ (Table 9) the first ten of which are from the right side, the remaining (to begin with L71o6) from the left. L6696 is a slightly worn specimen with a height/length index of 121 (Pl. 25); L6291 is a somewhat smaller specimen likewise slightly worn and with the same index (Pl. 25). The left $\mathrm{M}^{3}$ L7io6 belongs to the same individual as L6696, and the left M ${ }^{3}$ L646I belongs to the same individual as L629I. The specimen L6638 is unworn but incomplete at the base of the crown; its height/length index is $c .120$.

| Table 9 |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| No. of specimen | Measurements of $\mathbf{M}^{3}$ of Ceratotherium praecox (mm) |  |  |  |  |  |  |
|  | L6696 | L6291 | L6638 | L6294 | L6620 | L5666 | Liog84 |
| Ant.post. (int.) | 66 | 58 | c. 65 | 56 | 58 | 61 | 61 |
| Ant.transv. | 71 | 65 | 75 | 68 | 69 | 74 | 66 |
| Length outer surf. | 78 | 70 | c. 80 | 78 | 75 | 75 | 70 |
| No. of specimen | L6641 | L6290 | Lir997 | L7106 | L646ı | Li3614 |  |
| Ant.post. (int.) | 64 | 6 I | 72 | - | 58 | 60 |  |
| Ant.transv. | 71 | 75 | 75 | - | 65 | 73 |  |
| Length outer surf. | 76 | 81 | 80 | 78 | c. 70 | 72 |  |
| No. of specimen | L6289 | L6642 | Lirogi | L6466 |  |  |  |
| Ant.post. (int.) | 60 | 57 | 58 | 66 |  |  |  |
| Ant.transv. | 67 | 69 | 69 | 72 |  |  |  |
| Length outer surf. | 75 | 73 | 68 | 74 |  |  |  |

These specimens have already been referred to above. L6294 has a crochet extending all across the medisinus; L664i and Lirg97 have a very large crochet, and Liog84 has a small crista and an internal projection at the base of the crochet.

Some of the remaining lower cheek teeth are in situ in incomplete mandibles, as follows: $\mathrm{L}^{6} 6_{15}$, a right mandibular ramus, has the posterior portion of $\mathrm{P}_{4}$ and the three molars; the lengths are reduced as a result of interproximal wear, their transverse diameters slightly exceed those in Li $\mathrm{Li}_{3} 035$ and Lir 849 (Table 2), and the height at $M_{1}$ is $130 \mathrm{~mm} . \mathrm{P}_{2}-\mathrm{M}_{2} \sin$. and $\mathrm{P}_{2}-\mathrm{P}_{4}$ dext. of one and the same individual, L6659, are narrower-crowned, as are those recorded in Table 2. A right and a left mandibular ramus with the much worn $\mathrm{M}_{1}-\mathrm{M}_{3}$ on either side (L6612, L6614), give an $\mathrm{M}_{1}-\mathrm{M}_{3}$ length shorter than that in the less worn dentitions. The height of the ramus at $\mathrm{M}_{1}$ is 125 mm . L6793 is a right mandibular ramus with $\mathrm{P}_{2}-\mathrm{M}_{2} ; \mathrm{Lirl}_{1989}$ is a right ramus fragment with $\mathrm{M}_{1}, \mathrm{M}_{2}$, and part of $\mathrm{M}_{3}$. Two parts of right rami, $\mathrm{L}_{1} 3759$ and $\mathrm{L}_{1} 8805$, have $\mathrm{M}_{2}$, and $\mathrm{M}_{3}$, respectively, in situ. The measurements of these teeth are given in Table 1 o.

Table 10

| No. of specimen | L6615 | L6659 | L6612 | L6793 | Lir989 | Li3759 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\mathrm{P}_{2}$, ant.post. | - | - | - | - | - | , |
| ant.transv. | - | 18 | - | - |  | - |
| post.transv. | - | 20 | - | - |  |  |
| $\mathrm{P}_{3}$, ant.post. | - | (47) | - | - | - | - |
| ant.transv. | - | 24 | - | - | - |  |
| post.transv. |  | 28 | - | - |  |  |
| $\mathrm{P}_{4}$, ant.post. | - | 52 | - | (43) | - |  |
| ant.transv. | - | 29 | - | ${ }^{31}$ | - |  |
| post.transv. |  | 31 | - |  | - |  |
| $M_{1}$, ant.post. | (40) | 57 | (44) | - | 55 | - |
| ant.transv. | 37 | 32 | 37 |  | 35 |  |
| most.transv. $\mathrm{M}_{2}$, ant.post. | (51) | 34 <br> 65 | 36 <br> $(52)$ | (54) |  |  |
| ant.transv. | 38 | - | 37 | 37 | 37 | 36 |
| post.transv. |  | - | 39 | 37 | $3^{8}$ | 37 |
| $\mathrm{M}_{3}$, ant.post. | 59 | - | 58 |  | - | 63 |
| ant.transv. | $3^{8}$ | - | - | - | - | 37 |
| post.transv. | 38 | - | 35 | - | - | $3^{8}$ |
| Length $\mathrm{M}_{1}-\mathrm{M}_{3}$ | 160 | - | 150 | - | - | - |

There are seven isolated specimens of $\mathrm{P}_{2}$ (Table in) the first four of which are from the right side. The first and the last specimen are decidedly larger than the $\mathrm{P}_{2}$ in the two mandibles of Table 2.

Table il
Measurements of $\mathbf{P}_{\mathbf{2}}$ of Ceratotherium praecox (mm)

| No. of specimen | Lir8i2 | L6684 | L6676 | Li 1999 | Li 1959 | L6665 | LII815 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ant.post. | 33 | (30) | - | (27) | 30 | 32 | 34 |
| Ant.transv. | 20 | 18 | 17 | 17 | 18 | 17 | 21 |
| Post.transv. | 23 | 23 | 21 | 18 | 20 | 18 | 23 |

There are ten isolated specimens of $\mathrm{P}_{3}$ (Table 12) the first three of which are from the right side.

Table 12
Measurements of $\mathrm{P}_{3}$ of Ceratotherium praecox (mm)

| No. of specimen | $\mathrm{Li}_{210} 7$ | Lir8io | L6671 | L668 ${ }_{\text {I }}$ | L6669 | Lil8if |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ant.post. | (40) | (44) | (41) | 48 | (45) | (38) |
| Ant.transv. | 25 | 27 | 24 | 25 | 28 | 24 |
| Post.transv. | 28 | 30 | 31 | 26 | 28 | 26 |
| No. of specimen | L2525 | L5698 | Lir809 | L5697 |  |  |
| Ant. post. | (42) | (42) | (43) | 50 |  |  |
| Ant.transv. | 25 | 24 | 27 | 27 |  |  |
| Post.transv. | 30 | 28 | 30 | 28 |  |  |

$P_{4}$ is represented by eight specimens (Table 13) the first two of which are from the right side. $\mathrm{L} 6687, \mathrm{Li}_{12} 108$, and $\mathrm{LII}_{18} 04$ are larger, especially wider, than their homologues in the dentitions of Tables 2 and 10.

Table I3
Measurements of $\mathrm{P}_{4}$ of Ceratotherium praecox (mm)

| No. of specimen | L6687 | Li2 $\mathrm{I}_{10} 8$ | LII804 | Lif8ı6 | L6762 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Ant.post. | 49 | 53 | 51 | (45) | 50 |
| Ant.transv. | 33 | 33 | 33 | 30 | 29 |
| Post.transv. | 34 | 34 | 37 | 31 | 31 |
| No. of specimen | L6670 | L475 ${ }^{\text {I }}$ | L6662 |  |  |
| Ant.post. | (47) | 55 | (49) |  |  |
| Ant.transv. | 28 | - | 31 |  |  |
| Post.transv. | 35 | 3 I | 33 |  |  |

Fourteen isolated lower molars represent either $M_{1}$ or $M_{2}$ (Table 14); the first eight are from the right side, the remaining six (beginning with L6672) from the left.

Table I4
Measurements of $\mathbf{M}_{1}$ and $\mathbf{M}_{2}$ of Ceratotherium praecox (mm)

| No. of specimen | L6678 | L6679 | L2525 | L6690 | L6302 | L5690 | Lir894 |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ant.post. | $(56)$ | 58 | 60 | $(57)$ | 60 | 65 | $(50)$ |
| Ant.transv. | 34 | 30 | 31 | 34 | 35 | 34 | 37 |
| Post.transv. | 35 | 33 | 33 | 35 | 34 | 34 | 37 |
| No. of specimen | L6677 | L6672 | L4752 | L9126 | L5669 | L6689 | Li3390 |
| Ant.post. | 62 | 60 | - | 60 | $(55)$ | $(49)$ | 65 |
| Ant.transv. | 38 | 37 | 38 | 38 | 38 | 35 | 38 |
| Post.transv. | 36 | 39 | 35 | 37 | 37 | 37 | 36 |

The last lower molar, $M_{3}$, is easily distinguishable from $M_{1}$ or $M_{2}$ by its reduced posterior cingulum; in well-worn specimens the absence of a posterior pressure scar of course is characteristic for $M_{3}$. There are eight isolated $M_{3}$ (Table 15) the first two of which are from the right side.

Table 15
Measurements of $\mathbf{M}_{3}$ of Ceratotherium praecox (mm)

| No. of specimen | L5667 | Li 1802 | L6613 | Lif989 | L9609 | L9125 | L9iso | L9120 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ant.post. | 68 | 66 | (55) | 65 | 67 | 65 | 65 | (57) |
| Ant.transv. | 38 | 40 | 37 | 42 | 41 | 39 | 36 | 36 |
| Post.transv. | 34 | 37 | 36 | 37 | 35 | 34 | 34 | 34 |

In Tables 10 to ${ }^{5} 5$, inclusive, the anteroposterior diameter is in parentheses when it is much reduced because of interproximal wear.

There are a number of teeth belonging to the milk dentition of Ceratotherium praecox. Deciduous teeth were not present among the material of this species described from Kanapoi, Lothagam-i and Ekora (Hooijer \& Patterson 1972). Therefore, the Langebaanweg milk teeth are compared below with those of the two living African species. The differential characters of the milk molars of Diceros bicornis and Ceratotherium simum are recorded in Hooijer (1959). In a collection from Late Pleistocene sites near Swartklip, Cape Province, reported upon by Hendey \& Hendey (1968), there are milk molars of $C$. simum, which have been used for comparison.

The maxillary milk dentition of Ceratotherium praecox comprises two isolated and much worn DM ${ }^{1}$, both from the left side, L6674 and L6675, measuring 22 mm anteroposteriorly and 23 mm transversely. In $C$. simum $\mathrm{DM}^{1}$ is more elongated anteroposteriorly than in $D$. bicornis because of the greater forward projection of the parastyle in the former, but this character is lost in much worn specimens like those from Langebaanweg and a distinction cannot be made at this stage of wear.

Of $\mathrm{DM}^{2}$ there are two specimens in the Langebaanweg collection, $\mathrm{L}_{4} 6 \mathrm{o} 8$ (Pl. 29) and $\mathrm{L}_{5} 664$, both from the left side. $\mathrm{DM}^{3}$ is represented only by a single specimen, L9io5B (Pl. 29), from the left side and lacking most of the ectoloph. Finally, of the last upper milk molar, $\mathrm{DM}^{4}$, we have three specimens, one right lacking the outer surface, L 6727 , one entire left $\mathrm{DM}^{4}, \mathrm{~L}_{13} 8 \mathrm{r} 8$, and another left specimen, much worn down, L665I (Pl. 29). The upper milk molars in $C$. simum are distinguished from those in $D$. bicornis by the more prominent parastyle, suppression of paracone style, greater crown height, absence of inner cingula, stronger crista joining the crochet and forming a medifossette, and the postsinus being approximately as deep as the medisinus instead of shallower. The inner portion of the protoloph is more distinctly curved backward in C. simum than in $D$. bicornis, but this difference is more marked in the posterior milk molars than in $\mathrm{DM}^{2}$, in which it is not or hardly evident. Upper milk dentitions of $C$. simum and of $D$. bicornis have been described from the Early Pleistocene Makapansgat caves (Hooijer 1959); they tend to be on the large side but otherwise indistinguishable from their recent homologues. Variation ranges in dimensions of the milk teeth of the recent species are presented in Table 16 along with the measurements of the Langebaanweg specimens and those from Swartklip in the South African Museum; the Swartklip specimens conform to those of $C$. simum in every respect (they bear catalogue numbers preceded by ZW).

The $\mathrm{DM}^{2}$ of Ceratotherium praecox, L 4608 , has a prominent parastyle as in C. simum but has an internal cingulum along the protocone, as in D. bicornis. There is a tubercle at the medisinus entrance that is absent in $\mathrm{L}_{5} 664$; both specimens have a well-developed crista joining the crochet and forming a
medifossette. The postsinus is almost as deep as the medisinus; these are, again, C. simum characters.

| Table 16 |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Measurements of upper milk molars of C. praecox and recent species (mm) |  |  |  |  |  |  |
| $\mathrm{DM}^{2}$, no. of specimen | L46o8 | $\mathrm{L}_{5} 664$ | D. bicornis | C. simum | ZWi92 | ZW26ıo |
| Greatest length ectoloph | 42 | - | 38-41 | 41-51 | 41 | 42 |
| Antero-transverse | 37 | $35+$ | 33-39 | $36-4{ }^{1}$ | 36 | 34 |
| Postero-transverse | 42 | $38+$ | 35-40 | 35-43 | - | 33 |
| $\mathrm{DM}^{3}$, no. of specimen | L9105 ${ }^{\text {B }}$ | D. bicornis | C. simum | ZWi842 |  |  |
| Greatest length ectoloph | - | 45-52 | 53-61 | 53 |  |  |
| Antero-transverse | - | 40-50 | 46-48 | 46 |  |  |
| Postero-transverse | - | 39-47 | 44-46 | 45 |  |  |
| DM, ${ }^{4}$ no. of specimen | Li38ı8 | $\mathrm{L}_{6651}$ | D. bicornis | C. simum |  |  |
| Greatest length ectoloph | 60 | $54+$ | 50-55 | 66-68 |  |  |
| Antero-transverse | 56 | 53 | 45-53 | c. 54-55 |  |  |
| Postero-transverse | 53 | 52 | 40-51 | 52-60 |  |  |

$\mathrm{L}_{5} 664$ is incomplete internally, but the minimal transverse diameters can be given. The Langebaanweg $\mathrm{DM}^{2}$ tally well in size with those of $C$. simum. The two Swartklip specimens of $\mathrm{DM}^{2}$, ZWig2 and ZW26ro, both from the right side, lack the internal cingulum, display well-formed medifossettes, and have the postsinus as deep as the medisinus, as in C. simum to which they belong. The same holds good for the Swartklip $\mathrm{DM}^{3}$, ZWi842, which is from the left side. In the Langebaanweg collection there is but one $\mathrm{DM}^{3}, \mathrm{~L} 9105 \mathrm{~B}$, wanting most of the ectoloph. There is a slender crista, not joining the crochet, hardly any trace of an inner cingulum (except at the medisinus entrance), but the postsinus is less deep than the medisinus, as in D. bicornis. No measurements can be given. Of $\mathrm{DM}^{4}$ we have three Langebaanweg specimens, one right, lacking the outer surface, and two from the left side, as listed above. The entire specimens show the absence of the paracone style, the formation of a medifossette, and the absence of an inner cingulum, as in C. simum, although the postsinus is decidedly less deep than the medisinus, as in $D$. bicornis. Thus, the C. praecox milk molars combine characters found in C. simum and D. bicornis, whereas in size they are intermediate between the two.

Of the mandibular milk dentition there are the following specimens: L6686, $\mathrm{DM}_{2}$ dext., slightly worn; L9105C, $\mathrm{DM}_{3}$ dext., unworn (Pl. 32), metalophid height 41 mm , and hypolophid height 38 mm ; L630r, $\mathrm{DM}_{3}$ dext., slightly worn; L9105A, $\mathrm{DM}_{4}$ dext., unworn, metalophid height 50 mm , and hypolophid height 46 mm ; L6689, $\mathrm{DM}_{4}$ dext., much worn down; L6795, left ramus with incomplete $\mathrm{DM}_{3-4} ; \mathrm{L} 6660, \mathrm{DM}_{4}$ dext. in ramus fragment, slightly worn (Pl. 32), crown not fully erupted, anteroposterior diameter 54 mm , as in $\mathrm{L}_{9} 105 \mathrm{~A}$; $\mathrm{L} 2524, \mathrm{DM}_{4}$ sin. in ramus fragment, crown edge broken, lingual base not exposed; Li2870 and L6757, both $\mathrm{DM}_{4}$ sin., slightly worn.

As shown in Table 17, the Langebaanweg lower milk molars are larger than those in D. bicornis, as were the upper milk molars, but they correspond rather well with those from Swartklip, which represent C. simum. These Swartklip specimens are: ZWi837, $\mathrm{DM}_{2-4}$ dext. in ramus fragment; ZW2036,
$\mathrm{DM}_{2-3}$ dext.; ZWı867, $\mathrm{DM}_{2}$ dext.; ZWı876, $\mathrm{DM}_{3}$ dext., and ZWig66, $\mathrm{DM}_{3}$ sin., unworn, metalophid height 45 mm , hypolophid height 42 mm . The $\mathrm{DM}_{3}$ of $C$. praecox that is unworn, L 9105 C , has the anteroposterior diameter

Table 17
Measurements of lower milk molars of C. praecox and recent species (mm)

| $\mathrm{DM}_{2}$, no. of specimen | L6686 |  |  |  | $\begin{aligned} & \text { ZW } \\ & 1837 \end{aligned}$ | $\begin{gathered} \text { ZW } \\ 2036 \end{gathered}$ | $\begin{aligned} & \text { ZW } \\ & 1867 \end{aligned}$ | D. bicornis |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Ant.post. | 41 |  |  |  | 40 | 40 | 39 | 27-33 |
| Ant.transv. | 16 |  |  |  | 16 | - |  | 13-15 |
| Post.transv. | 18 |  |  |  | 20 | - | - | 15-18 |
| $\mathrm{DM}_{3}$, no. of specimen | L9105 | L6301 | L6795 |  | $\begin{aligned} & \text { ZW } \\ & 1837 \end{aligned}$ | $\begin{gathered} \text { ZW } \\ 1876 \end{gathered}$ | $\begin{gathered} \text { ZW } \\ 1966 \end{gathered}$ | D. bicornis |
| Ant.post. | 48 | $4^{6}$ | c. 47 |  | 46 | 45 | 44 | 38-41 |
| Ant.transv. | 20 | - | c. 23 |  | 22 | - | 22 | 19-20 |
| Post.transv. | 22 | 21 | 25 |  | 23 | - | 23 | 20-22 |
| $\mathrm{DM}_{4}$, no. of specimen | L9105 | L6689 | Li2870 | L6757 | $\begin{aligned} & \text { ZW } \\ & 1837 \end{aligned}$ |  |  | D. bicornis |
| Ant.post. | 54 | (46) | 51 | 51 | 48 |  |  | 41-45 |
| Ant.transv. | 23 | 23 | 25 | 25 | - |  |  | 22-23 |
| Post.transv. | 26 | 25 | 27 | $23+$ | - |  |  | 23-25 |

longer than that in the unworn $\mathrm{DM}_{3}$ of C. simum, ZWig66 (48 against 44 mm ), whereas both in metalophid height and in hypolophid height L9io5C is less than is ZWig66 ( 4 I and 38 mm against 45 and 42 mm ). It follows from this comparison that the milk tooth of C. praecox is less hypsodont than that of C. simum; we got the same result from the unworn permanent premolars and molars.

## Postcranial Skeleton

The postcranial material, which is very abundant at the Langebaanweg 'E' Quarry, is listed in the tables of measurements that follow ( 18 through $5^{\circ}$ ). Measurements of the bones of D. bicornis and C. simum have been given in previous papers (Hooijer \& Singer 1960; Hooijer 1969) from skeletons in the South African Museum, Cape Town, and in the Osteology Department, National Museum Centre for Prehistory and Palaeontology, Nairobi, respectively. In both cases the C. simum skeleton is larger than that of D. bicornis, with more massive metapodials (higher width/length ratios), but other than that no skeletal differences between the two extant species are apparent. Most of the Langebaanweg bones are larger than their homologues even in C. simum.

Eleven proximal portions of scapulae are in the Langebaanweg collection (Table 18) the first five of which are from the right side.

Table 18

| Measurements of scapula (mm) |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| No. of specimen |  |  |  | L8244 | $\mathrm{LII}_{1773}$ | LII524 | L8245 |
| 1. Ant.post. diameter of collum scapulae |  |  |  | 135 |  | - | c. 130 |
| 2. Ant.post. diameter over tuber scapulae and glenoid cavity |  |  |  | c. 170 | 155 | - | c. 170 |
| 3. Ant.post. diameter of glenoid cavity |  |  | , | c. 110 | c. 100 | c. 110 | c. 105 |
| 4. Transverse diameter of idem |  |  |  | c. 95 | c. 90 | 95 | 95 |
| 5. Transverse diameter of tuber scapulae |  |  |  | - | 55 |  | 60 |
| No. of specimen | L8306 | Li 3857 | L8288 | L8287 | L8290 | L8266 | Li3779 |
| 1. | - | 125 | 120 | 130 | 125 | - | 135 |
| 2. | - | 165 | c. 155 | 165 | 165 | 165 | 175 |
| 3. | c. 105 | 105 | c. 100 | 105 | 105 | 105 |  |
| 4. | 95 | c. 90 | c. 90 | 100 | - | - | - |
| 5. | - | 65 | 65 | c. 60 | c. 60 | c. 70 | - |
| No. of specimen | D. bicorni | C. simum |  |  |  |  |  |
| 1. | 100 | 130 |  |  |  |  |  |
| 2. | 130 | 160 |  |  |  |  |  |
| 3. | 85 | 105 |  |  |  |  |  |
| 4. | 80 | 100 |  |  |  |  |  |
| 5. | 45 | 60 |  |  |  |  |  |

The mid-portion of the shaft of a right humerus, L342I, has a width at the deltoid tuberosity of 170 mm , and a least width of 85 mm , as in C. simum (Hooijer 1969: 91). There are further only distal portions of the humerus, nine in all (Table 19) the first six of which are from the right side.

Table 19
Measurements of humerus (mm)

| No. of specimen | L6886 | L6977 | Li 3559 | L6947 | L6878 | L6899 |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: |
| I. Least width of shaft | 90 | 80 | c. 90 | 85 | - | 90 |
| 2. Greatest distal width | - | 190 | - | - | c. 180 | c. 190 |
| 3. Width of trochlea | 135 | 130 | 130 | c. 130 | 125 | 125 |
| No. of specimen | L3443 | L6965 | Li3463 | D. bicornis | C. simum |  |
| I. | 80 | - | 80 | 60 | $70-85$ |  |
| 2. | 185 | - | - | $150-155$ | 180 |  |
| 3. | 120 | 125 | - | 100 | 120 |  |

A radio-ulna dext., $\mathrm{L}_{12818 \text {, is slightly damaged proximo-medially; the }}$ radius is longer than any of the fossil radii, four of which are nearly entire (Table 20); only the last specimen in this table, $\mathrm{L}_{4967}$, is from the left side.

Table 20
Measurements of radius (mm)

| No. of specimen | Li28ı8 | L7997 | L6375 | L8ı 14 |
| :---: | :---: | :---: | :---: | :---: |
| 1. Median length | 400 | 375 | c. 370 | 385 |
| 2. Proximal width | - | 125 | 130 | 130 |
| 3. Proximal ant.post. diameter (medial side) | - | 80 | 85 | 85 |
| 4. Least width of shaft | 75 | 75 | 70 | 70 |
| 5. Greatest distal width | 120 | 120 | 115 | - |
| 6. Width distal articular surface | 105 | 100 | 100 | 95 |
| No. of specimen | L4967 | D. bicornis | C. simum |  |
| 1. | 390 | 345-350 | 365-380 |  |
| 2. | 135 | 100 | 120-125 |  |
| 3. | 90 | 60 | 75 |  |
| 4. | 75 | 45-55 | 65-70 |  |
| 5. | - | 95 | 120 |  |
| 6. | 105 | 80 | 100 |  |

There are twenty-one proximal portions of the radius (Table 21); the first seven are from the right side.

Table 21
Proximal measurements of radius ( mm )

| No. of specimen |  |  | Li3175 | L637 ${ }^{1}$ | L6370 | L3425 | L998ı | L7983 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 2. Proximal width |  |  | 135 | 130 | 125 | 125 | 120 | c. 130 |
| 3. Proximal ant.post. diameter (medial side) |  |  | 90 | c. 80 | 80 | 85 | c. 85 | 90 |
| 4. Least width of shaft |  |  | - | 65 | 65 | - | - |  |
| No. of specimen | L7934 | L7968 | L8or 7 | L8007 | L4205 | Li3845 | Li 2888 |  |
| 2. | 130 |  | 125 | c. 125 | 115 | 125 | 130 |  |
| 3. | 90 | c. 95 | 80 | - | 85 | 85 | 90 |  |
| 4. |  |  |  | 65 | 70 | 65 | 70 |  |
| No. of specimen | L7986 | L4959 | L9978 | L2229 | L8015 | L2289 | L7958 | L9988 |
| 2. | 125 | 130 | 135 | c. 125 | 125 | 125 | 125 | c. 120 |
| 3. | 80 | 85 | c. 90 | 85 | 80 | c. 80 | c. 85 | 80 |
| 4. | 75 | 70 | 80 | 70 | - | - |  | - |

Distal radius portions number thirty-three (Table 22), fifteen from the right and eighteen from the left side.

Table 22
Distal measurements of radius (mm)

| No. of specimen | L6369 | L6367 | L7973 | L6177 | L4202 | $\mathrm{Lr}_{3} 843$ | L5234 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 4. Least width of shaft | 65 | 70 | 70 | - |  | 65 |  |
| 5. Greatest distal width | 120 | 120 | 120 | 120 | c. 115 | c. 110 | 110 |
| 6. Width distal articular surface | 100 | 105 | 105 | 110 | 95 | 90 | 95 |
| No. of specimen | L9985 | L7911 | L8oio | L796ı | L6362 | L6ı 73 | L3065 |
| 4. | - | - | - | - | - | - | - |
| 5. | 120 | 120 | c. 115 | 120 | 120 | c. 120 | 120 |
| 6. | 100 | 105 | 105 | 105 | 100 | 105 | 100 |
| No. of specimen | $\mathrm{L}_{51} 74$ | L8ol2 | Li3842 | L2290 | L2293 | L2291 | L9733 |
| 4. | - | 70 | 70 | 70 | 65 | 65 | 65 |
| 5. | c. 120 | 110 | c. 120 | 115 | - | 120 | - |
| 6. | 95 | 90 | 105 | 105 | 100 | 100 | 90 |
| No. of specimen | L9986 | $\mathrm{L}_{4} \mathrm{O}_{3}$ | L9730 | L6170 | L4194 | L6372 | L3067 |
| 4. | - | - | - | - | - | - | - |
| 5. | 115 | 115 | c. 115 | 110 | c. 115 | 120 | 110 |
| 6. | 105 | 100 | 100 | 95 | 100 | 100 | 100 |
| No. of specimen | L7924 | L8oo6 | L4957 | L7920 | L4200 |  |  |
| 4. | - | - |  | - | - |  |  |
| 5. | 120 | c. 105 | 110 | 110 | 115 |  |  |
| 6. | 95 | 100 | 90 | 90 | 100 |  |  |

The ulna of the radio-ulna dext., $\operatorname{Li2818\text {,istheonlyentireulnainthe}}$ Langebaanweg collection; it has a maximum length of 530 mm (D. bicornis 450 mm ; C. simum 510 mm ), and a length from the processus anconaeus (beak) to the extremity of the olecranon of 175 mm (D. bicornis 140 mm ; C. simum $\left.{ }_{1} 65 \mathrm{~mm}\right)$. Further measurements are given in Table 23. In this table, twenty proximal and distal ulna portions are listed; the first twelve are from the right side, the remaining eight (beginning with L8o52) from the left.

Table 23
Measurements of ulna (mm)

| No. of specimen |  | L12818 | L806o | Li3836 | L8038 | L8071 | L8055 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| I. Width at semilunar notch |  | - | - | c. 95 | 95 | c. 105 | 105 |
| 2. Greatest distal diameter |  | 90 | - | - | - | - | - |
| 3. Ant.post. diameter distal articular surface |  | 60 | - | - | - | - | - |
| No. of specimen | L4210 | L7984 | L7959 | L8029 | L8025 | L7985 | L804 ${ }^{\text {I }}$ |
| 1. | - | - | - | - | - | - | - |
| 2. | 85 | c. 80 | c. 80 | c. 80 | c. 70 | 85 | - |
| 3. | 60 | 55 | c. 50 | 55 | 55 | 55 | 55 |
| No. of specimen | L8052 | Li3833 | L9994 | L625 ${ }^{\text {I }}$ | Li3839 | Li2891 | L3554 |
| 1. | 105 | 110 | 105 | 105 | - | - | - |
| 2. | - | - | - | - | 80 | 85 | 90 |
| 3. | - | - | - | - | 60 | 60 | 60 |
| No. of specimen | L7927 | D. bicornis | C. sim |  |  |  |  |
| 1. | - | 90 | 110 |  |  |  |  |
| 2. | 80 | 75 | 90 |  |  |  |  |
| 3. | 55 | 60 | 65 |  |  |  |  |

There are twenty-six scaphoids (Table 24) the first ten of which are from the right side.

Table 24
Measurements of scaphoid (mm)

| No. of specimen |  |  | L6oio | L6ol 2 | L6oo3 | L6oo9 | L9477 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| I. Posterior heig |  |  | 63 | 60 | 58 | 57 | 60 |
| 2. Anterior heigh |  |  | 58 | 60 | 59 | 59 | 61 |
| 3. Proximal width |  |  | 55 | 56 | 54 | 55 | 54 |
| 4. Proximal ant. | eter |  | 75 | 73 | 74 | 70 | 78 |
| 5. Maximum dia | tal facets |  | 70 | 70 | 69 | 68 | 71 |
| No. of specimen | Lir 767 | L7850 | Lir 768 | L5284 | L9483 | Li3472 | L6218 |
| 1. | 63 | 60 | 65 | 62 | 60 | 66 | 62 |
| 2. | 62 | 60 | 63 | 61 | 58 | 64 | 59 |
| 3. | 55 | 61 | 57 | 60 | 53 | 55 | 61 |
| 4. | 74 | 76 | 79 | 77 | 75 | 77 | 77 |
| 5. | 75 | 77 | 75 | 79 | 75 | 73 | 75 |
| No. of specimen | L7809 | L6or 4 | L6008 | L7849 | L5986 | L3569 | L7803 |
| 1. | 60 | 67 | 60 | 63 | 67 | 63 | 65 |
| 2. | 59 | 60 | 63 | 63 | 64 | 64 | 59 |
| 3. | 55 | 60 | 57 | 58 | 60 | 55 | 57 |
| 4. | 75 | 87 | 73 | 84 | 80 | 74 | 78 |
| 5. | 75 | 78 | 72 | 75 | 77 | 74 | 76 |
| No. of specimen | L7735 | L7826 | L4290 | $\mathrm{Li}_{3} 616$ | L786I | L7738 | $\mathrm{L}_{5282}$ |
| 1. | 66 | 65 | 61 | 64 | 64 | 67 | 57 |
| 2. | 63 | 65 | 59 | 60 | 63 | 68 | 58 |
| 3. | 60 | 60 | 56 | 55 | 59 | 59 | 54 |
| 4. | 76 | 79 | 73 | 8 I | 82 | 77 | 68 |
| 5. | 73 | 75 | 69 | 75 | 80 | 77 | 68 |
| No. of specimen | D. bicornis | C. simum |  |  |  |  |  |
| I. | 50 | 62 |  |  |  |  |  |
| 2. | 54-60 | 58-65 |  |  |  |  |  |
| 3. | 55 | 60 |  |  |  |  |  |
| 4. | 63 | 75 |  |  |  |  |  |
| 5. | 62-70 | 73-78 |  |  |  |  |  |

Of the lunar there are thirty-six specimens (Table 25), and the first eighteen are from the right side.

Table 25
Measurements of lunar (mm)

| No. of specimen |  | L4253 | Li 2379 | L6oo6 | L7853 | L7829 | L7882 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| I. Anterior height |  | 64 | 61 | 65 | 69 | 61 | 61 |
| 2. Proximal width |  | 67 | 61 | 64 | 68 | 63 | 62 |
| 3. Greatest ant.post. diameter |  | 79 | 83 | 81 | 87 | 78 | 80 |
| No. of specimen | $\mathrm{L}_{5281}$ | L4270 | L3049 | L4787A | L9475 | L4287 | $\mathrm{LI}_{3} 828$ |
| 1. | 65 | 57 | 58 | 60 | 59 | 60 | 63 |
| 2. | 68 | 58 | 66 | 58 | 61 | 60 | 60 |
| 3. | $75+$ | 72 | 76 | 78 | 78 | 77 | 77 |
| No. of specimen | L5290 | Li3823 | L7737 | L7755 | L5975 | Li 3824 | LI 12187 |
| 1. | 62 | 62 | 58 | 64 | 59 | 66 | 63 |
| 2. | 64 | 60 | 59 | 62 | 56 | 66 | 66 |
| 3. | 79 | 78 | 76 | 80 | 75 | 78 | 82 |
| No. of specimen | Lir 596 | L7885 | L7822 | L7771 | Li3727 | L9184 | L 9183 |
| 1. | 66 | 58 | 61 | 60 | 58 | 63 | 62 |
| 2. | 65 | 60 | 63 | 60 | 66 | 64 | 63 |
| 3. | 86 | 76 | 76 | 78 | 76 | 83 | 81 |
| No. of specimen | L5293 | Lif598 | L7896 | L4286 | L5972 | L7890 | L9457 |
| 1. | 60 | 58 | 64 | 63 | 61 | 65 | 66 |
| 2. | 60 | 63 | 65 | 63 | 63 | 65 | 63 |
| 3. | 75 | 75 | 80 | 81 | 79 | 78 | 82 |
| No. of specimen | L3806 | L7774 | D. bicornis | s C. sim |  |  |  |
| 1. | 59 | 64 | $44-48$ | $54-6$ |  |  |  |
| 2. | 58 | 65 | 48 | 58-6 |  |  |  |
| 3. | 77 | 80 | 64-68 | 75 |  |  |  |

Fifteen specimens of the cuneiform are in the Langebaanweg collection (Table 26), ten right and five left; L9465 is presented in anterior view in Pl. 33 (bottom).

Table 26
Measurements of cuneiform (mm)

| No. of specimen |  |  | $\mathrm{Li}_{27} \mathrm{C}_{5}$ | L3405 | $\mathrm{L}_{5218}$ | L4265 | L5286 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1. Anterior height |  |  | 57 | 51 | 52 | 47 | 61 |
| 2. Distal width |  |  | 56 | 48 | 46 | 49 | 51 |
| 3. Proximal ant.post. diameter |  |  | 51 | 47 | 44 | 43 | 46 |
| 4. Greatest horizontal diameter |  |  | 68 | 63 | 58 | 58 | 61 |
| No. of specimen | L7808 | L7833 | L7869 | L7898 | $\mathrm{Li}_{3} 82 \mathrm{I}$ | L9254 | L7749 |
| 1. | 55 | 53 | 56 | 57 | 56 | 45 | 59 |
| 2. | - | 44 | 50 | 51 | c. 47 | 43 | 50 |
| 3. | $4^{8}$ | 41 | 43 | 47 | 45 | 43 | 49 |
| 4. | 61 | 56 | 61 | 64 | 60 | 57 | 64 |
| No. of specimen | L3566 | L947 ${ }^{1}$ | L9465 | D. bicorni | is C. sim |  |  |
| 1. | 59 | 52 | 53 | 50 | 56-5 |  |  |
| 2. | 52 | 50 | 53 | 38-40 | 45- |  |  |
| 3. | 51 | 43 | 48 | $3^{8-40}$ | $4^{8-5}$ |  |  |
| 4. | 67 | 57 | 62 | 53 | 66 |  |  |

Three pisiforms, one right, L6oo4, and two left, L7854 and L7892, are in the Langebaanweg collection (Table 27); L7892 is presented in anterior view in Plate 33 (bottom). The bones have the two facets, for ulna and cuneiform.

Table 27
Measurements of pisiform (mm)

| No. of specimen | L6004 | L7854 | L7892 | D. bicornis | C. simum |
| :--- | :---: | :---: | :---: | :---: | :---: |
| I. Length | 71 | 72 | 67 | 6 I | 60 |
| 2. Distal height | 51 | 49 | 43 | 35 | 36 |

An exceptional bone is L 7823 , a cuneiform sin. with the pisiform completely ankylosed to it. The part representing the cuneiform is normal in shape, but it forms a solid mass with the pisiform, and the ulnar facets of the two bones are confluent ( Pl . 33, middle). The greatest horizontal diameter of the anomalous bone is just over 110 mm (the distal extremity of the pisiform is incomplete). For comparison a cuneiform and a pisiform are figured along with the cuneipisiform (Pl. 33, bottom).

The trapezium, the radial of the distal row of carpal bones, with facets for the scaphoid and the trapezoid, is represented in the Langebaanweg collection by a single specimen, L 3497 ; it is from the right side. In Table 28 the fossil bone is shown to be larger than its homologue in $C$. simum, as is usual for Langebaanweg bones.

Table 28
Measurements of trapezium (mm)
No. of specimen

1. Height
2. Proximal diameters

The trapezoid is represented in the Langebaanweg collection by five specimens, three from the right and two from the left side (Table 29).

## Table 29

Measurements of trapezoid (mm)

| No. of specimen | L7798 | Lir88i | Li3999 | L4263 | L4267 | D. bicornis | C. simum |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| I. Anterior width | 35 | 37 | 37 | 35 | 38 | 30 | 35 |
| 2. Anterior height | 38 | 45 | 34 | 39 | 38 | 31 | 32 |
| 3. Posterior height | 35 | 50 | 33 | 36 | 37 | 29 | 36 |
| 4. Ant.post. diameter | 51 | 55 | 49 | 48 | 52 | 41 | 49 |

Of the magnum we have twenty-one specimens (Table 30) the first ten of which are from the right side.

Table 30
Measurements of magnum (mm)

| No. of specimen |  |  | L7793 | L4244 | $\mathrm{LI}_{12824}$ | $\mathrm{L}_{4} \mathrm{O}^{8} 8$ | L5568 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| I. Anterior width |  |  | 59 | 59 | 65 | 58 | 56 |
| 2. Anterior height |  |  | 40 | 40 | 38 | 34 | 42 |
| 3. Proximal ant.post. diameter |  |  | 83 | 80 | 83 | 68 | 76 |
| 4. Greatest diameter |  |  | 103 | 104 | 115 | c. 85 | 96 |
| No. of specimen | L9473 | L9460 | L9459 | L6or3 | L4264 | $\mathrm{L}_{5184}$ | L6217 |
| 1. | 62 | 61 | 57 | 52 | 56 | 56 | 56 |
| 2. | 38 | 45 | $4{ }^{1}$ | 38 | 40 | 40 | $4{ }^{1}$ |
| 3. | 80 | 85 | 84 | 74 | 77 | 82 | 78 |
| 4. | - | - | - | - | - | 105 | 107 |
| No. of specimen L7876 | L7876 | L5259 | L7743 | Lif592 | L9476 | L4283 | L7745 |
| 1. | 59 | c. 60 | 57 | 60 | c. 55 | 54 | 57 |
| 2. | 38 | 42 | 45 | 43 | 42 | 40 | c. 40 |
| 3. | 77 | 86 | 88 | - | 80 | 77 | 78 |
| 4. <br> No. of specimen | 94 | 105 | 112 | 101 | 101 | - |  |
|  | L7797 | L7759 | D. bicornis | S C.sim |  |  |  |
| $\begin{aligned} & \text { I. } \\ & 2 . \\ & 3 . \end{aligned}$ | 51 | c. 55 | 44-49 | 57-5 |  |  |  |
|  | 34 | 40 | 32 | 38 |  |  |  |
|  | 77 | 82 | 63-67 | 70 |  |  |  |
| 4. |  | - | 77-85 | 84 |  |  |  |

There are forty specimens of the unciform in the Langebaanweg collection (Table 31), twenty from the right, and the same number from the left side.

| No. of specimen |  | L7762 | L7812 | Li2824 | L9193 | LI2766 | L5262 | L7747 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| I. Anterior height |  | 58 | 64 | 65 | 57 | 62 | 56 | 61 |
| 2. Anterior width |  | 81 | 87 | 83 | 77 | 8ı | 75 | 83 |
| 3. Greatest diameter |  | 103 | 112 | 107 | ı00 | 108 | 98 | 105 |
| No. of specimen | Lif590 | L6or6 | L4240 | L946r | L9201 | L6005 | L7879 | Lir 597 |
| 1. | 62 | 62 | 55 | 60 | 56 | 55 | 63 | 61 |
| 2. | 85 | 81 | 76 | 78 | 72 | 80 | 88 | 85 |
| 3. | 111 | 105 | 102 | 100 | 98 | 96 | c. 115 | 103 |
| No. of specimen | L9456 | L7855 | L526o | L7870 | L9184 | L4076 | Liro97 | L7837 |
| 1. | 56 | 64 | 55 | 60 | 57 | 58 | 56 | 62 |
| 2. | 72 | 87 | 73 | 77 | 75 | 83 | 75 | 83 |
| 3. | 98 | 108 | 98 | 102 | 95 | 109 | 97 | 105 |
| No. of specimen |  | L4285 | Li 2826 | L9468 | L5973 | $\mathrm{Li}_{3} 829$ | Lif591 | L7840 |
| 1. |  | 57 | 61 | 60 | 54 | 58 | 63 | 58 |
| 2. |  | 76 | 79 | 81 | 76 | 83 | $9{ }^{1}$ | 85 |
| 3. |  | 100 | 105 | 102 | 96 | 108 | 112 | 110 |
| No. of specimen |  | L4789A | L4256 | Lif6oo | L5263 | L5258 | L774 ${ }^{2}$ | L9469 |
| 1. |  | 55 | 63 | 53 | 55 | 65 | 60 | 57 |
| 2. |  | 74 | 85 | 78 | 77 | 82 | 80 | 80 |
| 3. |  | 104 | 107 | 95 | 99 | 106 | 102 | 108 |
| No. of specimen |  | L5257 | L9466 | L9464 | D. bicornis | S C. sim | um |  |
| 1. |  | 57 | 60 | 57 | 49-51 | 51-5 |  |  |
|  |  | 74 | 84 | 79 | 63-65 | 74- |  |  |
| 3. |  | 100 | 106 | 105 | 84-90 | 99-1 |  |  |

There are twenty entire second metacarpals (Table 32) the first five of which are from the right side. The ratio middle width/median length in the fossil series varies from 0,21 to 0,28 , which includes the observations on the recent Mc.II (taken from Hooijer \& Singer 1960, and Hooijer 1969).

> Table $32^{`}$
> Measurements of second metacarpal (mm)

| No. of specimen | $\mathrm{L}_{3066}$ | $\mathrm{~L}_{4890}$ | $\mathrm{~L}_{5934}$ | $\mathrm{~L}_{12819}$ | $\mathrm{~L}_{5988}$ |
| :--- | :---: | :---: | :---: | :---: | :---: |
| 1. Median length | 173 | 177 | 167 | I $_{72}$ | 160 |
| 2. Proximal width | 42 | 39 | 43 | 44 | 38 |
| 3. Proximal ant.post. diameter | 52 | c. 50 | 55 | 54 | 50 |
| 4. Middle width | 43 | 40 | 42 | - | 38 |
| 5. Middle ant.post. diameter | 24 | 23 | 23 | - | 21 |
| 6. Greatest distal width | 50 | 49 | 56 | 56 | 50 |
| 7. Width distal trochlea | 44 | 43 | 45 | 46 | 44 |
| 8. Distal ant.post. diameter | 5 I | 49 | 49 | 50 | 48 |
| 9. Ratio middle width/length | 0,25 | 0,23 | 0,25 | - | 0,24 |


| No. of specimen | L9225 | L7111 | L7129 | L7iog | L7072 | L9395 | L7083 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1. | 175 | 158 | 172 | 167 | 159 | 172 | 176 |
| 2. | 45 | 37 | 44 | 40 | 37 | 4 I | 41 |
| 3. | 55 | 55 | 53 | 51 | 47 | 58 | 51 |
| 4. | 45 | 37 | 42 | 36 | 35 | 4 I | 37 |
| 5. | 25 | 20 | 23 | 23 | 21 | 25 | 24 |
| 6. | 53 | 52 | 49 | 46 | 47 | 52 | 56 |
| 7. | 46 | 44 | 45 | 42 | 42 | 45 | 47 |
| 8. | 48 | 48 | 48 | 47 | 45 | 51 | 45 |
| 9. | 0,26 | 0,23 | 0,24 | 0,22 | 0,22 | 0,24 | 0,21 |
| No. of specimen | L7093 | L7090 | L4104 | L7064 | L7154 | L413 ${ }^{2}$ | L6064 |
| 1. | 162 | 176 | 176 | 158 | 183 | 157 | 163 |
| 2. | 42 | 40 | 42 | 40 | 41 | 40 | 38 |
| 3. | c. 50 | 52 | 57 | 49 | 53 | 54 | $5{ }^{1}$ |
| 4. | 41 | 39 | 41 | 35 | 45 | 39 | 35 |
| 5. | 19 | 26 | 29 | 22 | 23 | 24 | 24 |
| 6. | 50 | 53 | 53 | 47 | 57 | 52 | 50 |
| 7. | 42 | 46 | 42 | 42 | 46 | 45 | 45 |
| 8. | 45 | 47 | 48 | 43 | 51 | 48 | 45 |
| 9. | 0,25 | 0,22 | 0,23 | 0,22 | 0,25 | 0,25 | 0,21 |
| No. of specimen | L7071 | D. bicornis |  | C. simum |  | $\begin{aligned} & \text { Chemeron } \\ & { }_{1} 65 \end{aligned}$ |  |
| 1. | 166 | 147 | 148 | 160 | 160 |  |  |
| 2. | - | 32 | 40 | 44 | 45 | c. 45 |  |
| 3. | 58 | 46 | 36 | 44 | 49 | c. 50 |  |
| 4. | 47 | 33 | 31 | 40 | 40 | 42 |  |
| 5. | 28 | 19 | 18 | 20 | 24 | 23 |  |
| 6. | 57 | 39 | 37 | 45 | 50 |  |  |
| 7. | 48 | 33 |  |  | 40 | - |  |
| 8. | 50 | 4 I | 38 | 43 | 45 | - |  |
| 9. | 0,28 | 0,22 | 0,21 | 0,25 | 0,25 | 0,25 |  |

The third metacarpal is represented in the Langebaanweg collection by twenty-two entire specimens (Table 33), eleven right and eleven left. The bone L3070 is a diseased specimen, somewhat like the second metatarsal of Dicerorhinus leakeyi Hooijer (1966, pl. 15) from Rusinga Island.

Table 33
Measurements of third metacarpal (mm)

| No. of specimen |  |  | L5962 | Lir ${ }_{3}{ }^{6}$ | L7086 | L708ı | L7100 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| I. Median lengt |  |  | 186 | 206 | 192 | 198 | 185 |
| 2. Proximal wid |  |  | 67 | 78 | 65 | 76 | 71 |
| 3. Proximal ant. |  |  | 58 | 67 | 55 | 65 | 63 |
| 4. Middle width |  |  | 57 | 66 | 56 | 63 | 57 |
| 5. Middle ant.po |  |  | 27 | 31 | 27 | 28 | 26 |
| 6. Greatest dista |  |  | 74 | 78 | 70 | 78 | 73 |
| 7. Width distal |  |  | 63 | 72 | 62 | 69 | 65 |
| 8. Distal ant.pos |  |  | 51 | 57 | 52 | 58 | 51 |
| 9. Ratio middle |  |  | 0,31 | 0,32 | 0,29 | 0,32 | 0,31 |
| No. of specimen | L7080 | Li3750 | L3070 | L6215 | L2275 | L6045 | L2276 |
| 1. | 193 | 186 | 195 | 187 | 203 | 194 |  |
| 2. | 70 | 64 | 69 | 65 | 72 | 67 | 74 |
| 3. | 56 | 57 | - | 57 | - | 58 | 57 |
| 4. | 58 | 54 | c. 60 | 58 | 61 | 59 | 61 |
| 5. | 30 | 24 | - | 27 | 27 | 26 | 31 |
| 6. | 75 | 67 | - | 69 | 80 | $70+$ | 78 |
| 7. | 63 | 58 | 65 | 62 | 67 | 62 | 68 |
| 8. | 51 | 50 | - | 54 | 55 | 50 | 54 |
| 9. | 0,30 | 0,30 | - | 0,31 | 0,30 | 0,30 | 0,31 |
| No. of specimen | L7001 | L12822 | L5937 | L593 I | L4149 | Li3756 | L938 ${ }^{\text {I }}$ |
| 1. | 183 | 195 | 182 | 188 | 188 | 206 | 192 |
| 2. | 72 | 77 | 70 | 72 | 78 | 69 | 71 |
| 3. | 57 | 61 | 57 | 56 | 58 | 58 | 54 |
| 4. | 63 | 66 | 55 | 58 | 64 | 63 | 57 |
| 5. | 25 | 28 | 24 | 27 | 26 | 24 | 24 |
| 6. | 73 | - | 73 | 71 | 75 | 71 | 73 |
| 7. | 61 | 66 | 61 | 58 | 64 | 60 | 62 |
| 8. | - | 56 | 53 | 50 | 54 | 54 | 51 |
| 9. | 0,34 | 0,34 | 0,30 | 0,31 | 0,34 | 0,31 | 0,30 |
| No. of specimen | L9408 | L7085 | Li 3580 | D. bi | ornis | C. sim |  |
| 1. | 186 | 183 | 192 | 162 | 166 | 173 | 176 |
| 2. | 72 | 78 | 77 | 59 | 60 | 70 | 68 |
| 3. | 58 | 61 | 60 | 48 | 51 | 55 | 52 |
| 4. | 64 | 64 | 58 | 46 | 45 | ${ }_{56}$ | 58 |
| 5. | 27 | 30 | 30 | 22 | 22 | 24 | 28 |
| 6. | - | 82 | 74 | 61 | 52 | 66 | 71 |
| 7. | 69 | 64 | 60 | 51 | - | - | 59 |
| 8. | - | 52 | 53 | 44 | 41 | 48 | 48 |
| 9. | o,34 | 0,35 | 0,30 | 0,28 | 0,27 | o,32 | o,33 |

The variation range in width/length ratio in the Langebaanweg Mc.III, 0,29 to 0,35 , is such that it includes the observations of $C$. simum but the two D. bicornis metapodials are relatively more slender than the fossil specimens, although the difference is small.

There are sixteen entire fourth metacarpals in the Langebaanweg collection (Table 34), the first five of which are from the right side. In this metacarpal, only one of the two $D$. bicornis is below the variation range in width/length ratio in the fossil specimens.

Table 34
Measurements of fourth metacarpal (mm)

| No. of specimen |  | L663 ${ }^{1}$ | L7084 | L5936 | L5949 | L7098 | L12820 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1. Median length |  | 148 | 147 | 155 | 151 | 149 | 157 |
| 2. Proximal width |  | 53 | 50 | 54 | 55 | 53 | 58 |
| 3. Proximal ant.post. diameter |  | 47 | 52 | 51 | $4^{8}$ | 50 | 50 |
| 4. Middle width |  | 39 | 42 | 40 | 37 | 40 | 44 |
| 5. Middle ant.post. diameter |  | 24 | 24 | 24 | 23 | 27 | 25 |
| 6. Greatest distal width |  | 50 | 58 | - | 48 | 52 | 55 |
| 7. Width distal trochlea |  | 46 | 49 | $4^{6}$ | 42 | 42 | 45 |
| 8. Distal ant.post. diameter |  | 46 | 45 | - | 42 | 44 | 46 |
| 9. Ratio middle width/length |  | 0,26 | 0,29 | o,26 | 0,25 | 0,27 | 0,28 |
| No. of specimen | L2285 | L94 11 | L7078 | L4131 | L9246 | L7089 | L7102 |
| 1. | 150 | 153 | 147 | 160 | 161 | 163 | 155 |
| 2. | 54 | 50 | 51 | 57 | 57 | 58 | 51 |
| 3. | 50 | - | 49 | 53 | 52 | 55 | 51 |
| 4. | 39 | 39 | 40 | 36 | 38 | 43 | 37 |
| 5. | 23 | 21 | 24 | 22 | 25 | 24 | 26 |
| 6. | 54 | 48 | 50 | 51 | 51 | 52 | 51 |
| 7. | 46 | 43 | 42 | 43 | 45 | $4{ }^{2}$ | 42 |
| 8. | 44 | 41 | 42 | 43 | 46 | 44 | 45 |
| 9. | 0,26 | 0,25 | 0,27 | 0,23 | 0,24 | 0,26 | 0,24 |
| No. of specimen | L7095 | L940 ${ }^{\text {I }}$ | L7ioi | D. bi | ornis | C. sim |  |
| 1. | 156 | 145 | 157 | 136 | 135 | 145 | 143 |
| 2. | 64 | 57 | 59 | 43 | $3^{8}$ | 55 | 54 |
| 3. | 53 | 49 | 51 | 43 | 44 | 51 | 50 |
| 4. | 42 | 38 | 38 | 33 | 30 | 40 | 41 |
| 5. | 25 | 25 | 26 | 18 | 19 | 23 | 23 |
| 6. | 58 | 47 | 52 | 43 | 35 | 48 | 52 |
| 7. | 47 | 46 | 46 | 37 | - | - | $4{ }^{2}$ |
| 8. | 48 | 43 | 44 | 38 | 34 | 43 | 45 |
| 9. | 0,27 | o,26 | o,24 | 0,24 | 0,22 | 0,28 | 0,29 |

The fifth metacarpal of Ceratotherium praecox is reduced, mammiform, as it is in the recent species. There is one specimen in the Langebaanweg collection, $\mathrm{L}_{1}$ 606, with the two facets for the unciform and Mc.IV. It is 46 mm in length, and 35 by 29 mm in proximal diameters. In D. bicornis these diameters are 35 mm , and 27 by 26 mm ; in C. simum the bone is larger, as usual, viz., length 45 mm , and 33 by 26 mm proximally.

Of the femora in the Langebaanweg collection there is only one that is nearly entire, Li2292, from the left side, lacking portions of the caput and of the trochanter major, and most of the medial part of the trochlea (first column in Table 35). In length from caput to medial condyle it exceeds the femur of C. simum, but in diameter of the caput it is just as large as the larger of the two C. simum femora. There are two isolated femur heads, Li2632 and Li2676, with the same diameter as Li2292. The width across the third trochanter, 175 mm , is also found in a mid-shaft portion of a left femur, Li3254. There are several juvenile shaft portions showing the third trochanter, viz., $\mathrm{Li}_{1381}, \mathrm{LI}_{3} 867$ 13869, and L3409. Three distal portions of femora, L8ir 8 and Li268ı from the right side, and $\mathrm{Lir}_{175} 8$ from the left, complete the list of femora in the Langebaanweg collection (Table 35).

Table 35
Measurements of femur (mm)

No. of specimen
I. Greatest length
2. Diameter of caput
3. Width across third trochanter
4. Greatest distal width
5. Distal ant.post. diameter, medial side
6. Distal ant.post. diameter, lateral side

No. of specimen
4. Greatest distal width
5. Distal ant.post. diameter, medial side
6. Distal ant.post. diameter, lateral side

| Li2292 | D. bicornis |  | C. simum |  |
| :---: | :---: | :---: | :---: | :---: |
| 590 | 440 | 460 | 510 | 530 |
| 110 | 80 | 85 | 110 | 95 |
| 175 | - | 140 | 155 | - |
| c. 175 | 120 | 125 | 155 | 150 |
| - | 160 | 165 | 190 | 190 |
| 155 | - | 125 | 155 |  |
| L8ı18 | Li 2681 | $\mathrm{LII}_{175}{ }^{\text {d }}$ |  |  |
| 175 | 180 | 165 |  |  |
| 210 | 225 | c. 200 |  |  |
| c. 160 | c. 175 | c. 145 |  |  |

Twenty-one entire patellae are in the Langebaanweg collection (Table 36), nine from the right, and eleven from the left side. All of them are larger than the recent bones even of $C$. simum.

Table 36 Measurements of patella (mm)

| No. of specimen | $\mathrm{LI}_{4} 035$ | Li3725 | Lif589 | L92io | L4250 | $\mathrm{LII}_{1} 887$ | L3069 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| I. Length | 130 | 115 | 105 | 110 | 115 | 130 | c. 135 |
| 2. Width | 120 | 110 | 105 | 105 | 105 | 120 | 125 |
| No. of specimen | L6226 | L606o | L4268 | L7766 | L7895 | L7739 | L406I |
| 1. | 130 | 115 | 120 | 125 | 120 | 115 | 130 |
| 2. | I 10 | IIO | 110 | I 10 | 105 | 105 | 110 |
| No. of specimen | $\mathrm{L}_{128} 8_{33}$ | Li 3968 | L7787 | L4246 | $\mathrm{L}_{5} \mathrm{SI}_{7} 7$ | L5927 |  |
| 1. | 135 | 120 | 120 | 115 | I 10 | 115 |  |
| 2. | 125 | 110 | 115 | 105 | 105 | 100 |  |
| No. of specimen | L5926 | D. bic | ornis | C. sim |  |  |  |
| 1. | 115 | 95 | 100 | 105 | 105 |  |  |
| 2. | 105 | 85 | 90 | 90 | 95 |  |  |

Of the tibia there are no entire specimens in the Langebaanweg collection; the most complete specimen, Li 805 , has only the medial portion of the proximal articular surface, and distally the lateral portion is damaged. The length, measured along the medial surface, is 355 mm , and the greatest length was probably 380 mm (D. bicornis 335 mm, C. simum $350-380 \mathrm{~mm}$ ). There are five proximal portions of the tibia (Table 37), the first three of which are from the right side.

Table 37
Proximal measurements of tibia ( mm )

| No. of specimen |  | L9702 | Li2619 | L7934 | $\mathrm{Li}_{31} 74$ | L7944 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1. Proximal width |  | Ler | 150 | 140 | 150 | 155 |
| 2. Proximal ant.post. diameter |  | 115 | 145 | $120+$ | - | c. 135 |
| 3. Least width of shaft |  | 70 | - | - | 75 | 80 |
| No. of specimen | D. $b i$ | cornis | C. si |  |  |  |
| 1. | 110 | 115 | 135 | 140 |  |  |
| 2. | - | 120 | - | 145 |  |  |
| 3. | - | 55 | - | 65 |  |  |

There are no less than forty-one distal portions of the tibia (Table 38); the first nineteen from the right side, and the remaining twenty-two (starting with Li3477) from the left.

|  |  | TAbl |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Distal m | asuremen | ts of tib | (mm) |  |  |  |
| No. of specimen |  | L7947 | L7908 | L6ı71 | L7953 | Li806 | L4968 |
| 3. Least width of shaft |  | 80 | 75 | 70 | 55 | 70 | 65 |
| 4. Distal width |  | 105 | 100 | 95 | 90 | 95 | 100 |
| 5. Distal ant.post. diameter |  | 100 | 90 | 85 | 85 | 80 | 90 |
| No. of specimen | LII 770 | L7909 | L4963 | L4742 | L9980 | L3073 | L6ı67 |
| 3. | - | - | - | - | - | - |  |
| 4. | 95 | 100 | 105 | 100 | c. 95 | 95 | 100 |
| 5. | 90 | 95 | 95 | 90 | 85 | 85 | 95 |
| No. of specimen | L7910 | L7930 | L6I 74 | L6373 | L6374 | L7948 | Li 3477 |
| 3. | - | - | - | - | - | - | 75 |
| 4. | 90 | 100 | 90 | 100 | 105 | 100 | 100 |
| 5. | 85 | 95 | $80+$ | 90 | 95 | - | 95 |
| No. of specimen | L4965 | Li3858 | L794 ${ }^{1}$ | L7940 | L4969 | L2262 | L6ı65 |
| 3. | 65 | - | 70 | 85 | 75 | 70 | - |
| 4. | 95 | 95 | 95 | 100 | 95 | 95 | 100 |
| 5 | 90 | 90 | 85 | 100 | - | 85 | 95 |
| No. of specimen | L7946 | L7914 | L7950 | L7931 | L7947 | L795 ${ }^{\text {I }}$ | L7912 |
| 3. | 75 | - | - | - | - | - | - |
| 4. | 90 | 95 | 100 | 105 | 100 | 95 | 100 |
| 5. | 90 | 90 | 95 | 100 | 85 | 85 | 90 |
| No. of specimen | L2264 | L7926 | L6366 | LII 529 | L4186 | $\mathrm{L}_{4} 187$ |  |
| 3. | - | - | - | - | - | - |  |
| 4. | 100 | 95 | 90 | 95 | 100 | 90 |  |
| 5. | 90 | 95 | 85 | 90 | - | - |  |
| No. of specimen | L7921 | D. bic | rnis | C. sim |  |  |  |
| 3. | - | - | 55 | - | 65 |  |  |
| 4. | 90 | 85 | 95 | 95 | 115 |  |  |
| 5. | - | 70 | 95 | 80 | 85 |  |  |

The astragalus is represented in the Langebaanweg collection by sixtyseven entire specimens (Table 39), thirty-six from the right side, and thirty-one (beginning with $\mathrm{L}_{4} \mathrm{I} 66$ ) from the left. The astragalus is the numerically best represented bone in the Langebaanweg collection, to which its solid build undoubtedly contributed.

Like the other bones from Langebaanweg, the astragali are on the large side when compared with their homologues in the living African species. Twenty-six out of the sixty-seven Langebaanweg astragali exceed the larger of the two $C$. simum astragali in all dimensions taken. The ratio medial height/total width varies between much wider limits in the Langebaanweg series (0,74-0,9I) than it does in the few recent bones of $D$. bicornis and $C$. simum, as may be expected. However, the variation range in this ratio in the Ceratotherium praecox series does not overlap with that in the Miocene brachypotheres of Africa and Europe (Brachypotherium heinzelini and B. brachypus: o,64-0,73; cf. Hooijer 1966: 148). In nearly all of the Langebaanweg astragali the trochlea width is

| Table 39 |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| No. of specimen |  | L5886 | L5891 | L5929 | L4169 | L5888 | $\mathrm{L}_{4}{ }^{173}$ |
| I. Lateral height |  | 93 | 96 | 89 | 91 | 94 | 94 |
| 2. Medial height |  | 90 | 94 | 88 | 81 | 85 | 90 |
| 3. Total width |  | 110 | 110 | 104 | 101 | 110 | 116 |
| 4. Ratio medial height/to | width | o,82 | o,85 | o,85 | o,8o | o,84 | o,78 |
| 5. Trochlea width |  | 103 | 95 | 93 | 95 | 98 | 105 |
| 6. Width of distal facets |  | 86 | 90 | 82 | 90 | 93 | 92 |
| No. of specimen | L5928 | L7222 | L7230 | L7226 | L7209 | L7207 | L7204 |
| 1. | 85 | 88 | 84 | 85 | 96 | 90 | 90 |
| 2. | 88 | 85 | 83 | 80 | 94 | 87 | 87 |
| 3. | 109 | 100 | 103 | 99 | 111 | ı06 | 115 |
| 4. | o,81 | o,85 | o,81 | o,8ı | o,85 | o,82 | o,76 |
| 5. | 102 | 96 | 93 | 92 | 103 | 96 | 100 |
| 6. | 82 | 86 | 83 | 82 | 90 | 78 | 95 |
| No. of specimen | L7211 | L7212 | L7195 | L4865 | Li803 | L7197 | L7198 |
| 1. | 95 | 90 | 94 | 80 | 83 | 92 | 94 |
| 2. | 92 | 89 | 94 | 80 | 8 I | 85 | 85 |
| 3. | 110 | 108 | 119 | 104 | 100 | 115 | 111 |
| 4. | o,84 | o,82 | o,79 | 0,77 | o,81 | 0,74 | 0,77 |
| 5. | 103 | 99 | 103 | 89 | 90 | 102 | 96 |
| 6. | 90 | 84 | 95 | 83 | 78 | 80 | 92 |
| No. of specimen | L7200 | Li 1577 | Lif903 | L6349 | Li3822 | L4161 | L6209 |
| 1. | 100 | 88 | $9{ }^{1}$ | 95 | $9{ }^{1}$ | 88 | 94 |
| 2. | 89 | 92 | 94 | 96 | 92 | 88 | 90 |
| 3. | 120 | 116 | 112 | 116 | 111 | 115 | 112 |
| 4. | o,74 | o,79 | o,84 | o,83 | o,83 | o,77 | o,8o |
| 5. | 104 | 95 | 98 | 98 | 102 | 97 | 97 |
| 6. | 98 | 94 | 93 | 94 | 92 | 93 | 88 |
| No. of specimen | Lir58i | L5427 | L4874 | L2267 | L4868 | L4864 | L7196 |
| 1. | 93 | 91 | 87 | 91 | 105 | 89 | 92 |
| 2. | 94 | 84 | 84 | 89 | 98 | 84 | 88 |
| 3. | 117 | 113 | 107 | 111 | 122 | 113 | 112 |
| 4. | o,8o | o,74 | o,79 | o,8o | o,80 | o,74 | o,79 |
| 5. | 103 | 99 | 96 | 97 | 105 | 96 | 102 |
| 6. | 88 | 86 | 85 | 81 | 103 | 87 | 87 |
| No. of specimen | L9489 | $\mathrm{Li}_{2515}$ | L4166 | L4162 | L4168 | L6065 | L5890 |
| 1. | 88 | 88 | 88 | 93 | 85 | $77+$ | 86 |
| 2. | 87 | 88 | 89 | 95 | 84 | 87 | 89 |
| 3. | 105 | 103 | 109 | 112 | 103 | 107 | 115 |
| 4. | o,83 | o,85 | o,82 | o,85 | o,82 | o,8ı | o,77 |
| 5. | 93 | 90 | 102 | 100 | 90 | 94 | 105 |
| 6. | 82 | 79 | 93 | 90 | 77 | 83 | 87 |
| No. of specimen | $\mathrm{L}_{4} 167$ | L7213 | L7225 | L7215 | L7216 | L5930 | L9492 |
| 1. | 87 | 86 | 94 | 88 | 91 | 88 | 90 |
| 2. | 86 | 91 | 91 | 87 | 92 | 94 | 88 |
| 3. | 116 | 115 | 120 | 110 | 107 | 115 | 111 |
| 4. | o,74 | 0,79 | o,76 | o,79 | o,86 | o,82 | o,79 |
| 5. | 99 | 105 | 105 | 96 | 98 | 108 | 98 |
| 6. | 83 | 96 | 95 | 87 | 82 | 88 | 95 |
| No. of specimen | Li 2655 | L7488 | L9495 | L7219 | L5717 | L7208 |  |
| 1. | 85 | 87 | 80 | 86 | 83 | 85 |  |
| 2. | 92 | 82 | 81 | 85 | 80 | 88 |  |
| 3. | 104 | 105 | 98 | 107 | 105 | 108 |  |
| 4. | o,88 | o,78 | -,83 | o,79 | o,76 | o,8ı |  |
| 5. | 90 | 90 | 87 | 95 | 94 | 92 |  |
| 6. | 83 | 83 | 80 | 84 | 83 | 85 |  |


| No. of specimen | L7210 | L7203 | L7206 | Li 1578 | Li 1583 | $\mathrm{L}_{4}{ }^{1} 64$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1. | 91 | 88 | 85 | 87 | 92 | 84 |
| 2. | 87 | 91 | 87 | 93 | 92 | 92 |
| 3. | 107 | 100 | 110 | 120 | 106 | 103 |
| 4. | 0,8I | 0,91 | 0,79 | 0,78 | o,87 | o,89 |
| 5. | 92 | 91 | 98 | 105 | 100 | 92 |
| 6. | 86 | 79 | 86 | 101 | 86 | 84 |
| No. of specimen | $\mathrm{LII}_{5} 82$ | $\mathrm{L}_{5} 889$ | L5869 | L4870 | L9491 | L4863 |
| 1. | 85 | 94 | 87 | 87 | 85 | 83 |
| 2. | 93 | 97 | 93 | 90 | 82 | 84 |
| 3. | 114 | 108 | 108 | 104 | 106 | 100 |
| 4. | 0,82 | o,89 | 0,86 | o,87 | 0,77 | o,84 |
| 5. | 94 | 97 | 96 | 93 | 90 | 92 |
| 6. | 91 | 88 | 85 | 83 | 84 | 86 |
| No. of specimen | L9486 | D. bicornis |  | C. simum |  |  |
| I . | 92 | 65 | 71 | 74 | 76 |  |
| 2. | 94 | 68 | 70 | 75 | 84 |  |
| 3. | 114 | 86 | 83 | 95 | 104 |  |
| 4. | 0,82 | 0,79 | 0,84 | 0,79 | 0,8ı |  |
| 5. | 99 | - 78 | 78 | 83 | 87 |  |
| 6. | 87 | 73 | 72 | 85 | 88 |  |

greater than the medial height, although in some by a narrow margin only; in three specimens ( $\mathrm{L}_{7203}, \mathrm{~L}_{4} \mathbf{1 6 4}_{4}$, and $\mathrm{L}_{5} 889$ ) the trochlea width equals the medial height, and in one ( $\operatorname{Li2655)~the~trochlea~width~is~just~a~little~less~than~}$ the medial height. This evidently exceptional condition in C. praecox is the rule in Aceratherium and Dicerorhinus (Hooijer 1966: 173); in Brachypotherium trochlea width exceeds medial height, as it does also in Paradiceros (Hooijer 1968:89) and Chilotheridium (Hooijer 197I : 377).

The calcaneum is represented in the Langebaanweg collection by fifty specimens, twenty-four right and twenty-six left (Table 40). In greatest height all of these exceed the recent bones used for comparison; in anteroposterior diameter thirty-six fossil calcanea exceed the recent.

| Table 40 |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Measurements of calcaneum (mm) |  |  |  |  |  |  |  |
| No. of specimen |  | LII584 | Lir 771 | $\mathrm{L}_{5} 867$ | L4174 | L7186 | L598o |
| 1. Greatest height |  | 149 | 153 | 143 | 140 | 141 | 145 |
| 2. Greatest width |  | 85 | 90 | - | - | - | - |
| 3. Ant.post. diameter |  | - | 83 | 76 | 72 | 73 | 76 |
| No. of specimen | L5893 | $\mathrm{L}_{5} 855$ | $L_{5982}$ | $L_{5981}$ | L4177 | L5851 | L3052 |
| 1. | 142 | 144 | 134 | 146 | 143 | 148 | 140 |
| 2. | 83 | 81 | 81 |  |  | 82 |  |
| 3. | 70 | 78 | 69 | 75 | 76 | 76 | 73 |
| No. of specimen | L3052 | L488ı | L7180 | L7169 | L7184 | L5856 | L7190 |
| 1. | 140 | 143 | 146 | 142 | 149 | 152 | 146 |
| 2. | - | - | - | - | 95 | 97 | 88 |
| 3. | 73 | 74 | 76 | 73 | 77 | 88 | 79 |
| No. of specimen | L7182 | L7198 | L7i8ı | L7191 | L7166 | L6348 | $\mathrm{Lr}_{38} \mathrm{O}_{4}$ |
| 1. | 145 | 157 | ${ }^{1} 53$ | 150 | 152 | 146 | 148 |
| 2. | 90 | 95 | - | 94 | - | - | - |
| 3. | 77 | 8 I | 77 | 81 | 80 | 76 | 76 |


| No. of specimen | L5892 | L3536 | L3149 | L7194 | L6055 | L5853 | L9503 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1. | 144 | 145 | 140 | 156 | 153 | 151 | 145 |
| 2. | 83 | - | 87 | - |  | 84 | 83 |
| 3. | 78 | 76 | 70 | 84 | 80 | 79 | 72 |
| No. of specimen | L4175 | L5461 | L7175 | L3790 | L8654 | L7188 | L7187 |
| 1. | 150 | 145 | 152 | 142 | 143 | 141 | 147 |
| 2. |  |  | - | 90 |  | - | 85 |
| 3. | 73 | 77 | 79 | c. 75 | 80 | 80 | 77 |
| No. of specimen | L7177 | L7171 | L7172 | L4179 | $\mathrm{LIH}_{15} 8_{5}$ | L6054 | Li802 |
| 1. | 143 | 145 | 140 | 146 | 161 | 141 | 141 |
| 2. | - | 79 | - | - | c. 95 | 85 | 83 |
| 3. | 75 | 70 | 73 | 77 | 83 | 72 | 74 |
| No. of specimen | L9501 | L7192 | $\mathrm{LI}_{3} 825$ | D. bicornis |  | C. simum |  |
| 1. | 152 | 153 | 138 | 110 | 110 | 125 | 125 |
| 2. | 82 | 83 | - | 65 | 70 | 80 | 82 |
| 3. | 75 | 82 | 79 | 60 | 65 | 75 | 66 |

The naviculars in the Langebaanweg collection number twenty-seven (Table 41), the first fifteen of which are from the right side.

Table 41
Measurements of navicular (mm)

| No. of specimen |  | L9516 | L7775 | L5567 | L3675 | L95 ${ }^{5}$ | L95 12 | L9181 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1. Anterior height |  | 35 | 32 | 34 | 37 | 31 | 31 | 34 |
| 2. Total width |  | 56 | 62 | 58 | 54 | 57 | 63 | 60 |
| 3. Ant.post. diameter |  | 71 | 72 | 82 | 75 | 72 | 76 | 80 |
| No. of specimen | L7852 | L7854 | L7888 | Lif623 | L6064 | L4242A | L6067 | L4251 |
| 1. | 32 | 31 | 32 | 30 | 33 | 33 | 34 | 33 |
| 2. | 56 | 57 | 61 | 53 | 60 | 55 | 59 | 54 |
| 3. | 76 | 72 | 73 | 69 | 76 | 69 | 72 | 74 |
| No. of specimen | L7757 | L9507 | L6o65 | L9510 | L5241 | L4 $4_{42}{ }^{\text {B }}$ | L4257 | $\mathrm{L}_{5} 6_{4}$ |
| 1. | 33 | 32 | 32 | 30 | 37 | 30 | 32 | 34 |
| 2. | 66 | 58 | 58 | 56 | 60 | 52 | 53 | 54 |
| 3. | 70 | 72 | 69 | 72 | 78 | 70 | 76 | 75 |
| No. of specimen | Li 2627 | L7889 | L7841 | L6o66 | D. bi | rnis | C. sim |  |
| 1. | 33 | 33 | 33 | 33 |  |  | 29 |  |
| 2. | 60 | 58 | 55 | 63 | 4 |  | 55 |  |
| 3. | 73 | 76 | 66 | 78 | 5 |  | 62 |  |

There are twenty-nine cuboids (Table 42), the first eleven of which are from the right side.

Table 42
Measurements of cuboid (mm)

| No. of specimen | L622 ${ }^{\text {I }}$ | $\mathrm{L}_{3} 804$ | L4262 | L3676 | L3496 | L9482 | Li2823 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1. Anterior height | 53 | 49 | 49 | 48 | 51 | 50 | 54 |
| 2. Anterior width | 53 | 48 | 54 | 49 | 47 | 49 | 58 |
| 3. Greatest ant.post. diameter | 77 | 75 | 76 | 69 | 75 | 73 | 86 |
| No. of specimen | L4269 | L7796 | Lir 750 | L4069 | L6223 | L426o | L7871 |
| 1. | 51 | 55 | 51 | 51 | 53 | 49 | 46 |
| 2. | 49 | 54 | 50 | 49 | 53 | 44 | 48 |
| 3. | 76 | 79 | 73 | 78 | 82 | 73 | 70 |

A LATE PLIOGENE RHINOGEROS FROM LANGEBAANWEG, CAPE PROVINGE 183

| No. of specimen | L945 8 | L9474 | L6620 | L4068 | $\mathrm{L}_{77} 85$ | Li 2008 | L7803 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1. | 48 | 52 | 52 | 52 | 46 | 50 | 53 |
| 2. | 51 | - | 5 I | 52 | 49 | 54 | 54 |
| 3. | 72 | 71 | 81 | 83 | 76 | 78 | 87 |
| No. of specimen | L9472 | L7770 | L5273 | $\mathrm{L}_{4289}$ | L5287 | L5288 | L5280 |
| 1. | 49 | 52 | 55 | 44 | 50 | 55 | 47 |
| 2. | 44 | 50 | 48 | 43 | 44 | 47 | 53 |
| 3. | 77 | 79 | 78 | 73 | 71 | 76 | 78 |
| No. of specimen | $L_{5294}$ | D. bicornis | C. sim |  |  |  |  |
| 1. | 48 | 37 | 43 |  |  |  |  |
| 2. | 46 | 44 | 52 |  |  |  |  |
| 3. | 73 | 65 | 80 |  |  |  |  |

The cuboid of Ceratotherium praecox is higher than wide anteriorly in sixteen specimens, and wider than high in nine. We find the same variation in Aceratherium and Dicerorhinus (Hooijer 1966: ı76); it is in Brachypotherium and Chilotherium that the width is distinctly greater than the height, and this is true to a lesser extent in Chilotheridium (Hooijer 1971: 380).

Eight ectocuneiforms, six right and two left (Table 43), have the anterior width about two times the anterior height, as in the recent African species, Aceratherium and Dicerorhinus, and Chilotheridium; in Chilotherium the width is three times the height (Hooijer 1966: 177 ; 1971:380-381).

Table 43
Measurements of ectocuneiform (mm)

| No. of specimen | L4075 | L9514 | L9517 | L7820 | L4053 | L4070 |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: |
| I. Anterior height | 30 | 32 | 28 | 30 | 27 | 27 |
| 2. Anterior width | 60 | 56 | 54 | 57 | 53 | 55 |
| 3. Ant.post. diameter | 71 | 59 | 57 | 57 | 51 | 56 |
| No. of specimen | L7749 | L7773 | D. bicornis | C. simum |  |  |
| I. | 33 | 33 | 24 | 27 |  |  |
| 2. | 57 | 58 | 45 | 57 |  |  |
| 3. | 58 | 59 | 53 | 54 |  |  |

One mesocuneiform, from the left side, is the remaining tarsal bone in the collection (Table 44).

Table 44
Measurements of mesocuneiform (mm)

| No. of specimen | $\mathrm{LI}_{1} 6663$ | D. bicornis | C. simum |
| :--- | :---: | :---: | :---: |
| 1. Height | 24 | 14 | 19 |
| 2. Width | 24 | 24 | 22 |
| 3. Ant.post. diameter | 45 | 34 | 43 |

A set of right metatarsals, Li3548-i 3550 , belong to one and the same individual ( Pl .34 ). Their measurements are given in the first columns of Tables 45-47.

Of the second metatarsal there are fifteen entire specimens (Table 45), the first seven of which are from the right side. The variation range in width/
length ratio is rather small, o,18-0,22 only, and one of the D. bicornis metapodials remains below these limits, that is, it is more slender in build.

Table 45
Measurements of second metatarsal (mm)

| No. of specimen |  |  | Li3550 | L2279 | L4II8 | L4886 | $\mathrm{LI}_{3} 802$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| I. Median length |  |  | 176 | 166 | 161 | 164 | 162 |
| 2. Proximal width |  |  | 41 | 35 | 37 | 35 | 37 |
| 3. Proximal ant.post. diameter |  |  | - | 49 | 55 | 51 | 54 |
| 4. Middle width |  |  | 32 | 32 | 30 | 33 | 29 |
| 5. Middle ant.post. diameter |  |  | 32 | 30 | 29 | 26 | 29 |
| 6. Greatest distal width |  |  | 49 | 45 | - | 44 | - |
| 7. Width distal trochlea |  |  | 44 | $4{ }^{1}$ | 39 | 39 | - |
| 8. Distal ant.post. diameter |  |  | 52 | 43 | 44 | 45 | 43 |
| 9. Ratio middle width/length |  |  | 0,18 | 0,19 | 0,19 | 0,20 | 0,18 |
| No. of specimen | L5943 | L7075 | Lil ${ }_{7} 72$ | L605 ${ }^{2}$ | L4109 | L4127 | L9380 |
| 1. | 174 | 158 | 162 | 153 | 160 | 168 | 162 |
| 2. | 43 | 33 | 36 | 33 | 33 | 33 | 39 |
| 3. | 56 | 51 | 58 | 50 | 50 | 54 | 56 |
| 4. | 35 | 29 | 30 | 28 | 32 | 30 | 35 |
| 5. | 33 | 27 | 32 | 24 | 27 | 30 | 30 |
| 6. | 48 | 43 | 49 | 41 | 41 | 44 | 48 |
| 7. | 43 | 38 | 42 | 39 | 38 | 38 | 44 |
| 8. | 48 | 43 | 45 | 42 | 44 | 48 | 45 |
| 9. | 0,20 | 0,18 | 0,19 | 0,18 | 0,20 | 0,18 | 0,22 |
| No. of specimen I. | L7097 | Lir904 | $\begin{gathered} \mathrm{L}_{4}{ }^{1} 4^{2} \\ 160 \end{gathered}$ | D. bicornis |  | C. simum |  |
|  | 157 | 153 |  | 129 | 135 | 148 | 151 |
| 2. | 38 | 35 | 39 | 25 | 24 | 38 | 34 |
| 3. | 56 | 5 I | 55 | 42 | 33 | 49 | 47 |
| 4. | 32 | 33 | 32 | 25 | 22 | 30 | 28 |
| 5. | 30 | 30 | 31 | 19 | 20 | 22 | 24 |
| 6. | 46 | 45 | 44 | 33 | 31 | 40 | 39 |
| 7. | 45 | 40 | 41 | 29 | - | - | 36 |
| 8. | 47 | 44 | 48 | 36 | 35 | 42 | 40 |
| 9. | 0,20 | 0,22 | 0,20 | 0,20 | 0,16 | 0,20 | 0, 19 |

There are twenty entire third metatarsals (Table 46), ten from the right and ten from the left side. The range of variation in width/length ratio of the fossil bones $(0,26-0,33)$ is very nearly the same as that in the four recent bones.

Table 46
Measurements of third metatarsal (mm)

No. of specimen
I. Median length
2. Proximal width
3. Proximal ant.post. diameter
4. Middle width
5. Middle ant.post. diameter
6. Greatest distal width
7. Width distal trochlea
8. Distal ant.post. diameter
9. Ratio middle width/length

| Li3548 | L604 8 | L4138 | Li3752 | LI3754 |
| :---: | :---: | :---: | :---: | :---: |
| 198 | 192 | 179 | 181 | 171 |
| 70 | 62 | 62 | 61 | 55 |
| - | 57 | 57 | 56 | 53 |
| 60 | 53 | 55 | 54 | 47 |
| 35 | 31 | 30 | 30 | 25 |
| 82 | 68 | - | 71 | 58 |
| 69 | 57 | 54 | 58 | 54 |
| 55 | 50 | 50 |  | 49 |
| 0,30 | 0,28 | 0,31 | 0,30 | 0,27 |

## A LATE PLIOGENE RHINOGEROS FROM LANGEBAANWEG, CAPE PROVINGE

| No. of specimen | L7068 | L7065 | L7062 | Li2615 | LI 1855 | L5960 | L6043 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1. | 191 | 187 | 182 | 180 | 188 | 198 | 186 |
| 2. | 63 | 64 | 57 | 61 | 62 | 67 | 58 |
| 3. | 61 | 55 | - | 57 | 55 | c. 60 | 51 |
| 4. | 54 | 52 | 53 | 52 | 57 | 62 | 49 |
| 5. | 29 | 29 | 26 | 31 | 27 | 33 | 28 |
| 6. | 67 | 67 |  | 68 | 70 | 80 | - |
| 7. | 59 | 56 | 56 | 56 | 60 | 66 | 53 |
| 8. | 52 | 51 |  | 51 | 51 | 58 | 49 |
| 9. | 0,28 | 0,28 | 0,29 | 0,29 | 0,30 | 0,31 | 0,26 |
| No. of specimen | L7000 | L5932 | L4148 | Li380ı | Li3749 | L9379 | L7152 |
| 1. | 180 | 171 | 178 | 190 | 183 | 177 | 182 |
| 2. | 59 | 58 | 58 | 59 | 59 | 61 | 60 |
| 3. | 53 | 52 | 55 | - | 54 | 51 | 53 |
| 4. | 48 | 53 | 49 | 55 | 53 | 54 | 60 |
| 5. | 26 | 29 | 28 | 29 | 31 | 31 | 29 |
| 6. | 66 | 64 | 66 | 71 | - | 68 | 74 |
| 7. | 55 | 54 | 55 | 59 | 55 | 59 | 62 |
| 8. | 50 | $4^{6}$ | 48 | 54 | 51 | 49 | 51 |
| 9. | 0,27 | 0,31 | 0,28 | 0,29 | 0,29 | 0,31 | 0,33 |
| No. of specimen | L7092 | D. bi | ornis | C. sim |  | Aterir |  |
| 1. | 183 | 148 | 152 | 160 | 169 | 180 |  |
| 2. | 59 | 48 | 50 | 59 | 55 | 58 |  |
| 3. |  | 48 | 45 | 47 | 49 | 50 |  |
| 4. | 50 | 40 | 40 | 51 | 48 | 49 |  |
| 5. | 28 | 21 | 19 | 22 | 25 | 24 |  |
| 6. | 65 | 54 | 45 | 56 | 66 | 68 |  |
| 7. | 57 | 47 | - | - | 51 | - |  |
| 8. | 50 | 42 | 40 | 46 | 47 | 45 |  |
| 9. | 0,27 | 0,27 | 0,26 | 0,32 | 0,28 | 0,27 |  |

There are sixteen entire fourth metatarsals in the Langebaanweg collection (Table 47), eight right and eight left. The width/length ratio does not vary a

Table 47
Measurements of fourth metatarsal (mm)


| No. of specimen | L5942 | L9390 | L924 ${ }^{1}$ | L7099 | D. bicornis |  | C. simum |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| I. | 152 | 157 | 155 | 154 | 125 | 127 | 138 | 146 |
| 2. | 49 | 57 | 45 | 49 | 42 | 39 | 44 | 49 |
| 3. | 46 | 55 | 56 | 46 | 40 | 40 | 47 | 45 |
| 4. | 29 | 30 | 30 | 27 | 26 | 26 | 35 | 29 |
| 5. | 35 | 40 | 34 | 33 | 24 | 23 | 26 | 28 |
| 6. | $3^{8}$ | 42 | 40 | $4{ }^{1}$ | 36 | 31 | 44 | 39 |
| 7. | 34 | 36 | 36 | 38 | 33 | - |  | 37 |
| 8. | 39 | 39 | 42 | 40 | 38 | 34 | 41 | 41 |
| 9. | 0,19 | o,19 | 0,19 | 0,18 | 0,21 | 0,20 | 0,25 | 0,20 |

great deal ( $0,17-0,22$ ). One of the recent bones (the first under the head C. simum) is not within these limits but above them; it is more massively built than the other recent, and the fossil fourth metatarsals.

There are nineteen first phalanges of median digits (Table 48), whether from the manus or from the pes I am unable to tell.

| Table 48 |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Measurements of phalanx I, median digit (mm) |  |  |  |  |  |  |  |
| No. of specimen |  | L3046 | L8418 | L6og9 | L8416 | L4214 | L9250 |
| 1. Median length |  | 38 | 41 | 40 | 43 | 38 | 42 |
| 2. Proximal width |  | 66 | 67 | 67 | 66 | 64 | 60 |
| No. of specimen | L8417 | $\mathrm{L}_{5276}$ | L9520 | L8415 | L5326 | L8420 | L6216 |
| 1. | 43 | 44 | 37 | 37 | 37 | 40 | $4{ }^{2}$ |
| 2. | 61 | 64 | 58 | 61 | - | 60 | 57 |
| No. of specimen | L8419 | L925 ${ }^{1}$ | L5993 | $\mathrm{Li}_{376}{ }^{\text {7 }}$ | L5275 | L7252 |  |
| 1. | 39 | 37 | 40 | 44 | 39 | - |  |
| 2. | 65 | 57 | 61 | 64 | 57 | 71 |  |
| No. of specimen | D. bicornis |  | C. simum |  |  |  |  |
|  | manus | pes | manus | pes |  |  |  |
| I. | 31 | 33 | 42 | 41 |  |  |  |
| 2. | 51 | 49 | 58 | 63 |  |  |  |

Four second phalanges of median digits are available (Table 49).
Table 49
Measurements of phalanx II, median digit (mm)

|  |  |  | D. bicornis |  |  |  | C. simum |  |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| No. of specimen | L 8426 | L 9253 | L9518 | $\mathrm{L}_{11607}$ | manus | pes | manus | pes |
| I. Median length | 30 | 33 | 34 | 26 | 26 | 28 | 30 | 30 |
| 2. Proximal width | 64 | 62 | 53 | 64 | 55 | 56 | 65 | 73 |

There is one third phalanx of a median digit (Table 50).
Table 50
Measurements of phalanx III, median digit (mm)
No. of specimen
r. Median length
2. Greatest width

|  | D. bicornis |  | C. simum |  |
| :---: | :---: | :---: | :---: | :---: |
| L8427 | manus | pes | manus | pes |
| 32 | 26 | 28 | - | 34 |
| 93 | 84 | 80 | - | 107 |

Five bones (L8421, L8422, L9256, L9519 and Lir879) represent first phalanges of lateral digits; they vary in median length from 35 to 37 mm , and in proximal width from 42 to 50 mm . A third phalanx of a lateral digit, L9257, has a median length of $c .30 \mathrm{~mm}$, and a greatest diameter of ${ }_{51} \mathrm{~mm}$. Two proximal sesamoids remain to be recorded; the larger bone, L7364, length 41 mm , width ${ }_{21} \mathrm{~mm}$, presumably belonged to a median digit, while the smaller, L4074, length 28 mm , width 17 mm , may have belonged to a lateral digit.

## Other C. praEcox sites in East and South Africa

We have evidence of the occurrence of Ceratotherium praecox at sites other than Kanapoi, Ekora and Lothagam- I in Kenya, and Langebaanweg in the Cape Province. Fragmentary teeth from the Mursi Formation of the Omo Basin in southern Ethiopia and from the Chemeron Formation in Kenya, previously referred to Ceratotherium simum germanoafricanum (Hooijer 1969: 86, 77), in the light of the discovery of Ceratotherium praecox at Kanapoi and Langebaanweg, should be identified as $C$. praecox. The teeth from the 'lower level' (Mursi Formation), which had been collected by R. Leakey in 1967, were re-examined by me in July 1971 at the Centre for Prehistory and Palaeontology, National Museum, Nairobi. There are a $\mathrm{P}^{4} \sin$. and a $\mathrm{M}^{2-3} \sin$. in palatal portions (Hooijer 1969, pl. 5, figs 4-5) displaying, as far as preserved, an angular antero-internal corner. In $\mathrm{M}^{3}$ there is a true medifossette, whereas in $\mathrm{P}^{4}$ and $\mathrm{M}^{2}$ the crochet extends across the medisinus without uniting with a crista to form a medifossette. $\mathrm{P}^{4}$ shows the internal indentation of the protocone also seen in $\mathrm{M}^{2}$. The internal face of $\mathrm{M}^{2}$ is 50 mm anteroposteriorly, and 30 mm of this are taken up by the protocone. Although all the teeth are incomplete externally the basal external crown outline is preserved, and the transverse diameters can be approximately given (Table $5^{1}$ ). They are within the limits of their homologues in the Langebaanweg collection. Although the ectolophs of the Mursi Formation specimens cannot be studied, in all observable characters these teeth agree with those of Ceratotherium praecox; the medifossette is not normally formed in this species, and its presence in the Mursi $\mathrm{M}^{3}$ is exceptional. The Chemeron maxilla with $\mathrm{M}^{1-3}$ (Hooijer 1969, pl. 2, fig. 1 ), from locality J.M.507, do not have medifossettes, and $\mathrm{M}^{2}$ has a distinct antero-internal crown angle. The teeth are very much worn down, and $\mathrm{M}^{1}$ and $\mathrm{M}^{3}$ are so fragmentary that the width cannot be determined, but those of $\mathrm{M}^{2}$ are approximately the same as those in the Mursi specimen (Table 51). The skull from J.M.91, Chemeron Formation (Hooijer 1969: 76, pl. I) is more advanced in its dentition and shows the rounded antero-internal crown angles, the medifossettes, and the posterior extension of the protocone characteristic of the modern species; this specimen moreover has the backwardly inclined occiput, extending beyond the occipital condyles, characteristic of C. simum germanoafricanum, and as such it was identified in my earlier paper. The presence of both Ceratotherium praecox and Ceratotherium simum germanoafricanum in the Chemeron Formation is puzzling,
for the mammalian fossils in the Chemeron Formation were found so closely together (Dr. W. W. Bishop, pers. comm.) as to make it unlikely that they were not of the same age. The Chemeron locality J.M.go (=J.M.91) is placed by Cooke \& Maglio (i97ı, fig. 2) at the 2 million year level, whereas the remainder of the Chemeron Formation is left at the 4 million year level. This arrangement is in accordance with the evidence provided by the rhinoceroses. Bishop (1971b), with a faunal list, gives the age of the Chemeron Formation as greater than 2,0 m.y. and less than 5,4 m.y.

A metapodial of a rhinoceros from the Chemeron Formation, locality J.M.5II, is a left second metacarpal. Whether it represents $C$. praecox or C. simum I am unable to tell; the measurements have been added to Table 32 and agree with those of either of the two species.

From locality J.M.5II of the Chemeron Formation there is a $\mathrm{P}^{4}$ dext. of a large chalicothere, a new element to the Chemeron Formation fauna (cf. Bishop 197Ib). It was collected on 5 August 1967; I found it in the Chemeron collection of the Department of Geology at Bedford College, London, on 18 November 1971, and it was given to me for study by Dr. W. W. Bishop. The specimen is of considerable interest as it adds to the younger elements of the Chemeron Formation fauna, and chalicothere teeth are rare anyway. The specimen is referable to Ancylotherium hennigi (Dietrich), a species recorded before from Laetolil and Bed I at Olduvai (Dietrich 1942: 105; Butler 1965: 226). It is very well preserved and not much worn; the lingual cusp is only just touched by wear, and the height of the worn ectoloph is 33 mm . The crown measures 28 mm anteroposteriorly and 3I mm transversely, and has all the characters of Ancylotherium (Thenius 1953:98 and fig. ı). The Olduvai material consists of a few carpals, metacarpals and phalanges only, but among the Laetolil collection there is an $\mathrm{M}^{2}$ (Dietrich 1942, pl. IV, fig. 37; pl. XII, fig. 79), measuring $55,0 \mathrm{~mm}$ anteroposteriorly and $40,0 \mathrm{~mm}$ transversely. The newly found $\mathrm{P}^{4}$ and the Laetolil $\mathrm{M}^{2}$, when compared with their homologues in an upper dentition of Ancylotherium pentelicum (Gaudry \& Lartet) as figured by Thenius, prove to be on a par for size. In the $A$. pentelicum dentition $\mathrm{P}^{4}$ measures 33,3 by $37,5 \mathrm{~mm}$, and $\mathrm{M}^{2} 67,2$ by $50,5 \mathrm{~mm}$ (Thenius 1953: 105); the Chemeron $\mathrm{P}^{4}$ and the Laetolil $\mathrm{M}^{2}$ are both one-sixth smaller in dimensions than the corresponding teeth in A. pentelicum. Laetolil and Olduvai Bed I are around the 2 million year level (Maglio 1970; Cooke \& Maglio 1971), and that is where part of the Chemeron Formation (locality J.M.go and 91) was placed by Cooke \& Maglio. However, as stated above, in the opinion of geologist Dr. Bishop, the geological evidence does not support a time gap of 2 million years between some Chemeron sites (J.M.90, 9I) and others. The tooth of Ancylotherium hennigi (locality J.M.5II) as well as the skull of Ceratotherium simum germanoafricanum (locality J.M.9I) and stage 2 or 3 of Elephas recki (Cooke \& Maglio 1971) suggest an age for the Chemeron Formation closer to 2 million years than to 4 million years. On the other hand we have elements like the maxillary of Ceratotherium praecox (locality J.M.507) in addition to Loxodonta adaurora Maglio, Mammuthus
subplanifrons (Osborn), Anancus cf. kenyensis (Cooke \& Maglio 1971) or Anancus sp. (Bishop 1971b), and Nyanzachoerus species 'A' of Cooke \& Ewer, which are suggestive of an age around 4 million years. If the Chemeron Formation fauna is really unified as to age, it may tentatively be placed around the 3 million year level, as suggested to me by Dr. W. W. Bishop. However this may be, further faunal studies are needed, and the record of Ancylotherium hennigi from locality J.M. 5 II of the Chemeron Formation is here given as a contribution for that end.

| Table 5I |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| Measurements of upper teeth of Ceratotherium praecox (mm) |  |  |  |  |
|  | Langebaanweg | Mursi Fm. | $\begin{aligned} & \text { Chemeron } \\ & \text { Fm. } \\ & \text { J.M. } 507 \end{aligned}$ | Swartlintjes Farm, Namaqualand |
| $\mathrm{P}^{4}$, ant.post. | 47-c. 57 | - | - | - |
| ant.transv. | 65-76 | c. 68 | - | - |
| post.transv. | 56-73 | c. 60 | - | - |
| $\mathrm{M}^{2}$, ant.post. | c. 50-68 | - | - | - |
| ant.transv. | 70-82 | c. 75 | c. 75 | 74 |
| post.transv. | 65-75 | c. 65 | c. 65 | 70 |
| $\mathrm{M}^{3}$, ant.post. (int.) | 56-72 | c. 60 | - | - |
| ant.transv. | 65-78 | c. 67 | - | - |
| length outer surface | 68-83 | - | - | - |

The Aterir Beds in the Baringo area, Kenya, which are placed by Maglio (1970), Cooke \& Maglio (1971) and Bishop (1971b) near the 4 million year level (as is the Mursi Formation = Yellow Sands), contain material of C. praecox. The inner portion of an upper right molar, marked $5 / \mathrm{B}_{4} / 6$, has the marked protocone fold, internal indentation of the protocone, slight internal cingulum, a crochet but no medifossette, and the marked antero-internal crown corner characteristic of the present species. No measurements can be given, but in its internal anteroposterior diameter, nearly 50 mm (of which 28 mm for the protocone), and the massive crochet it is nearest to $\mathrm{M}^{1}$. Another Aterir specimen, an upper left premolar, marked $5 / \mathrm{BI}$, again with the angular antero-internal corner, no medifossette, and a very weak paracone style, has the dimensions of $\mathrm{P}^{3}$ in dentition $\mathrm{Li}_{3} \mathbf{0 3 5}$. Its measurements have been added to Table 5 . There is further in the Aterir collection a right third metatarsal (marked I/I8 and I/23) indistinguishable from its Langebaanweg homologue; its measurements have been added to Table 46: Finally, there is a proximal sesamoid, presumably of a median digit, marked $5 / \mathrm{B}_{1}$. This Aterir specimen is 41 mm long and 20 mm wide, just about as large as the Langebaanweg sesamoid L7364.

An isolated, rolled $\mathrm{M}^{2}$ dext., lacking the antero-external and posteroexternal angles, and originating from Swartlintjes Farm, Hondeklipbaai, Namaqualand, C.P. (about 160 km north of Langebaanweg), represents the same species of Ceratotherium as that from Langebaanweg. According to the geologist who presented the specimen to the South African Museum, Mr. A. J. Carrington, the fossil molar came from ill-sorted angular felspathic fluviatile gravels at an elevation of $c .18 \mathrm{~m}$. The gravels overlie what are taken
as Lower Pleistocene marine sands, and would be Upper Pleistocene. However, it is difficult to reconcile this view with the characters of the rhinoceros molar, which are those of the Late Pliocene Ceratotherium praecox. Its rolled condition suggests that it was derived from an earlier deposit. The specimen is figured in Plate 25 (bottom right), and bears the South African Museum number Qi771. The ectoloph is 77 mm high as worn. There is a well-marked protocone fold and internal indentation of the protocone, an angular antero-internal crown corner, and further there are a weak cingulum internally at the protocone, a strong but relatively slender crochet, no crista, and a postsinus very nearly as deep as the medisinus. The antero-transverse diameter is 74 mm , the posterotransverse 70 mm , very close to those in L6658.

The Namaqualand site is the only one in the Cape Province other than Langebaanweg from which Ceratotherium praecox is recorded, and this species is further known only from north-western Kenya and southern Ethiopia. It is already proving useful in African correlations, and may become more so if and when found in other parts of Africa.

## Summary

Numerous remains of an extinct species of rhinoceros have been obtained by parties of the South African Museum at the ' $E$ ' Quarry of the Langebaanweg site, 104 km north-northwest of Cape Town, C.P. They are more abundant than those of any other large mammal in the Langebaanweg fauna; there are remains of seven skulls, ten mandibles (most of them with teeth in situ), ${ }_{1} 70$ isolated teeth, and 650 postcranial bones. This material is referred to Ceratotherium praecox Hooijer \& Patterson described from the Late Pliocene of Lothagam-1, Kanapoi, and Ekora in north-western Kenya. Ceratotherium praecox is little removed from the point of divergence of the genus Ceratotherium and the genus Diceros, and is held to represent the immediate ancestor of the modern white rhinoceros, Ceratotherium simum. The species is further recorded in the Cape Province from Swartlintjes Farm, Hondeklipbaai, Namaqualand (approximately 160 km north of Langebaanweg). It is also known from the Mursi Formation in southern Ethiopia, and the Chemeron Formation and the Aterir Beds in the Baringo area, Kenya, all deposits dated around the 4 million year level. The discovery of this species is proving most useful in inter-African correlation and adds to the evidence already available that the 'E' Quarry Langebaanweg site is Late Pliocene in age.

## Agknowledgements

It is a great pleasure to thank Dr. T. H. Barry, Director, Mr. and Mrs. Q. B. Hendey, and Mrs. D. Hirschon, Palaeomammalogy Department, South African Museum, for facilitating my work at the museum in May and June 1971, and for courtesies extended. I am indebted to Dr. W. W. Bishop for permission to include C. praecox remains from other East African sites, to Mr. R. E. F. Leakey who let me study Mursi Formation, Ethiopia, and

East Rudolf, Kenya, rhinoceroses, and to Mr. Neville Eden who took the photographs. My journey to South Africa has been made possible by a grant from the Wenner-Gren Foundation for Anthropological Research, Inc., New York.

## References

Bishop, W. W. $1971 a$. The Late Cenozoic history of East Africa in relation to hominoid evolution. In Turekian, K. K., ed. Late Cenozoic glacial ages: 493-527. Cambridge (Mass.) : Yale University Press.
Bishop, W. W. 197ı $b$. Stratigraphic succession 'versus' calibration in East Africa. In bishop, w. w. \& miller, J. A., eds. Calibration of hominoid evolution. Edinburgh: Scottish Academic Press.
Butler, P. M. 1965. East African Miocene and Pleistocene chalicotheres. Bull. Br. Mus. nat. Hist. (Geol.) 10: 163-237.
Cooke, H. B. S. \& Maglio, V. J. 1971. Plio-Pleistocene stratigraphy in East Africa in relation to proboscidean and suid evolution. In bishop, w. w. \& miller, J. A., eds. Calibration of hominoid evolution. Edinburgh: Scottish Academic Press.
Dietrich, W. O. i942. Ältestquartäre Säugetiere aus der südlichen Serengeti; DeutschOstafrika. Palaeontographica 94(A): 43-133.
Dietrich, W. O. 1945. Nashornreste aus dem Quartär Deutsch-Ostafrikas. Palaeontographica 96(A) : 46-90.
Heller, E. 1913. The White Rhinoceros. Smithson. misc. Collns 6r: i-77.
Hendey, Q. B. 1969. Quaternary vertebrate fossil sites in the south-western Cape Province. S. Afr. archaeol. Bull. 24: 96-105.

Hendey, Q.B. i970a. A review of the geology and palaeontology of the Plio/Pleistocene deposits at Langebaanweg, Cape Province. With an Appendix: The Langebaanweg Bovidae by A. W. Gentry. Ann. S. Afr. Mus. 56: 75-117.

Hendey, Q.B. r97ob. The age of the fossiliferous deposits at Langebaanweg, Cape Province. Ann. S. Afr. Mus. 56: i19-131.
Hendey, Q.B. \& Hendey, H. 1968. New Quaternary fossil sites near Swartklip, Cape Province. Ann. S. Afr. Mus. 52: 43-73.
Hooijer, D. A. i959. Fossil rhinoceroses from the Limeworks Cave, Makapansgat. Palaeont. afr. 6: 1 -I3.
Hooijer, D. A. 1966. Miocene rhinoceroses of East Africa. Bull. Br. Mus. nat. Hist. (Geol.) 13: II7-190.
Hooijer, D. A. i968. A rhinoceros from the Late Miocene of Fort Ternan, Kenya. Zool. Meded., Leiden 43: 77-92.
Hooijer, D. A. 1969. Pleistocene East African rhinoceroses. Fossil Vertebr. Afr. 1: 71-98.
Hooijer, D. A. i971. A new rhinoceros from the Late Miocene of Loperot, Turkana District, Kenya. Bull. Mus. comp. Zool. Harv. 142: 339-392.
Hooijer, D. A. \& Patterson, B. i972. Rhinoceroses from the Pliocene of north-western Kenya. Bull. Mus. comp. Zool. Harv. 144: 1-26.
Hooijer, D. A. \& Singer, R. ig6o. Fossil rhinoceroses from Hopefield, South Africa. Zool. Meded., Leiden 37: $113-128$.
Maglio, V. J. 1970. Early Elephantidae of Africa and a tentative correlation of African PlioPleistocene deposits. Nature, Lond. 225: 328-332.
Maglio, V. J. \& Hendey, Q.B. 1970. New evidence relating to the supposed stegolophodont ancestry of the Elephantidae. S. Afr. archaeol. Bull. 25: 85-87.
Thenius, E. 1953. Studien über fossile Vertebraten Griechenlands. III. Das Maxillargebiss von Ancylotherium pentelicum Gaudry und Lartet. Annls géol. Pays hell. 5: 97-106.
Thenius, E. 1955. Zur Kenntniss der unterpliozänen Diceros-Arten (Mammalia, Rhinocerotidae). Annln naturh. Mus. Wien 6o: 202-21 I.

## Explanation of the plates

Note. All specimens are Ceratotherium praecox Hooijer \& Patterson from Langebaanweg, except Plate 25, bottom right, which is from Swartlintjes Farm, Namaqualand.


Upper dentition, Li3035, crown view, $\times$ o,44.


Upper dentition, L2519, crown view, $\times$ o,52.


Upper dentition, Li3747, crown view, $\times \mathbf{0 , 3 5}$.


Skull, L6658, palatal view, $\times \mathbf{0}, 37$.


Top, $\mathrm{P}^{2-4}$ dext., Li3035, internal view, $\times 0,78$.
Middle, outer surfaces of $\mathrm{M}^{3}$ dext., L6291 and L66g6, external views, $\times 0,57$.
Bottom left, ectoloph of $\mathrm{P}^{4}$ sin., Li376o, external view, $\times 0,56$.
Bottom right, $\mathbf{M}^{2}$ dext., Swartlintjes Farm, Namaqualand, S.A.M. Qi77I, crown view, $\times$ o,82.


 views, $\times 0,52$.

Top row, from left to right, $\mathrm{DM}^{3} \sin ., \mathrm{L}_{1} 1_{5} \mathrm{~B}, \mathrm{DM}^{2} \sin ., \mathrm{L} 4608$, and $\mathrm{DM}^{4} \sin$., L 6651 , crown
views, $\times \mathrm{o}, 78$.
Bottom row, from left to right, $\mathrm{P}^{4} \sin ., \mathrm{LIII}_{1} 2, \mathrm{P}^{4} \sin$., L6655, and ectoloph of $\mathrm{M}^{2} \sin$., L 9118 ,


Right mandibular ramus, $\mathrm{L}_{13035, ~ i n t e r n a l ~ v i e w, ~} \times 0,27$.



Top, symphysis of the mandible, L6o58, top view, $\times$ o, 67 .
Middle, ankylosed cuneiform and pisiform sin., L7823, anterior view, $\times 0,6 \mathrm{r}$.
Bottom, cuneiform sin., L9465, and pisiform sin., L7892, as they articulate, anterior views, $\times 0,6 \mathrm{r}$.


Metatarsals II, III and IV dext., Li $354^{8-1} 355^{\circ}$, articulated, front view, $\times 0,6$ I.

## INSTRUCTIONS TO AUTHORS

## Based on

CONFERENCE OF BIOLOGICAL EDITORS, COMMITTEE ON FORM AND STYLE. I 960.
Style manual for biological journals. Washington: American Institute of Biological Sciences.

## MANUSCRIPT

To be typewritten, double spaced, with good margins, arranged in the following order:
(1) Heading, consisting of informative but brief title, name(s) of author(s), address(es) of author(s), number of illustrations (plates, figures, enumerated maps and tables) in the article.
(2) Contents. (3) The main text, divided into principal divisions with major headings; subheadings to be used sparingly and enumeration of headings to be avoided. (4) Summary. (5) Acknowledgements. (6) References, as below. (7) Key to lettering of figures. (8) Explanation to plates.

## ILLUSTRATIONS

To be reducible to $12 \mathrm{~cm} \times 18 \mathrm{~cm}$ ( 19 cm including caption). A metric scale to appear with all photographs.

## REFERENCES

Harvard system (name and year) to be used: author's name and year of publication given in text; full references at the end of the article, arranged alphabetically by names, chronologically within each name, with suffixes $a, b$, etc. to the year for more than one paper by the same author in that year.
For books give title in italics, edition, volume number, place of publication, publisher.
For journal articles give title of article, title of journal in italics (abbreviated according to the World list of scientific periodicals. 4th ed. London: Butterworths, 1963), series in parentheses, volume number, part number (only if independently paged) in parentheses, pagination.

Examples (note capitalization and punctuation)
bullough, W. S. 1960. Practical invertebrate anatomy. 2nd ed. London: Macmillan.
Fischer, P.-H. 1948. Données sur la résistance et de le vitalité des mollusques. F. Conch., Paris 88: 100-140.
Fischer, P.-H., Duval, M. \& Raffy, A. 1933. Etudes sur les échanges respiratoires des littorines. Archs Zool. exp. gén. 74: 627-634.
Конл, A. J. 196oa. Ecological notes on Conus (Mollusca: Gastropoda) in the Trincomalee region of Ceylon. Ann. Mag. nat. Hist. (13) 2: 309-320.
Koнn, A. J. 196ob. Spawning behaviour, egg masses and larval development in Conus from the Indian Ocean. Bull. Bingham oceanogr. Coll. 17 (4): 1-51.
Thiele, J. 19io. Mollusca: B. Polyplacophora, Gastropoda marina, Bivalvia. In schultze. l, Zoologische und anthropologische Ergebnisse einer Forschungsreise im westlichen und zentralen SüdAfrika. 4: 269-270. Jena: Fischer. Denkschr. med.-naturw. Ges. Jena 16: 269-270.

## ZOOLOGICAL NOMENCLATURE

To be governed by the rulings of the latest International code of zoological nomenclature issued by the International Trust for Zoological Nomenclature (particularly articles 22 and 51). The Harvard system of reference to be used in the synonymy lists, with the full references incorporated in the list at the end of the article, and not given in contracted form in the synonymy list.

[^2]

ANNALS OF THE SOUTH AFRICAN MUSEUM ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM

| Volume | $\mathbf{5 9}$ | Band |
| ---: | :---: | :--- |
| September | $\mathbf{1 9 7 2}$ | September |
| Part | 10 | Deel |



# A REVIEW OF THE SEPIIDAE (CEPHALOPODA) OF SOUTHERN AFRICA 

By<br>MARTINA A. ROELEVELD<br>Cape Town Kaapstad

# The ANNALS OF THE SOUTH AFRICAN MUSEUM 

are issued in parts at irregular intervals as material becomes available

Obtainable from the South African Museum, P.O. Box 6I, Cape Town

# Die ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM <br> word uitgegee in dele op ongereelde tye na beskikbaarheid van stof 

Verkrygbaar van die Suid-Afrikaanse Museum, Posbus 61, Kaapstad

> out of print/uit druk
> 1, 2(1, 3, 5, 7-8), 3(i-2, 5, t.-p.i.), 51-2, 5, 7-9), $6(\mathrm{I}, \quad$ t.-p.i. $), \quad 7(\mathrm{I}-3), \quad 8, \quad 9(\mathrm{I}-2), \quad 10(\mathrm{I})$, 1i(1-2, 5, 7, t.-p.i.), 24(2), 27, 3i(i-3), 33

Price of this part/Prys van hierdie deel
Rio,8o

## Trustees of the South African Museum <br> (C) Trustees van die Suid-Afrikaanse Museum 1972

Printed in South Africa by The Rustica Press, Pty., Ltd, Court Road, Wynberg, Cape

In Suid-Afrika gedruk deur
Die Rustica-pers, Edms., Bpk. Courtweg, Wynberg, Kaap

# A REVIEW OF THE SEPIIDAE (CEPHALOPODA) OF SOUTHERN AFRICA 

By
Martina A. Roeleveld
South African Museum, Cape Town
(With plates 35-45, 20 figures and 53 tables)
[MS. accepted I March 1972]
Contents
Introduction . . . . . . . . . 194
Methods ..... 194
Abbreviations and glossary ..... 196
Key to the Sepiidae of southern Africa ..... 197
Key to shells only ..... 199
Systematic discussion
Diagnoses of family and genera ..... 200
Description of southern African species
Sepia zanzibarica Pfeffer, 1884. ..... 202
Sepia officinalis vermiculata Quoy \& Gaimard, 1832 ..... 204
Sepia acuminata Smith, 1916 ..... 208
Sepia confusa Smith, 1916 ..... 210
Sepia incerta Smith, 1916 ..... 213
Sepia burnupi Hoyle, 1904 ..... 217
Sepia joubini Massy, 1927 ..... 221
Sepia adami n. sp. ..... 224
Sepia australis Quoy \& Gaimard, 1832 ..... 227
Sepia tuberculata Lamarck, 1798 ..... 231
Sepia papillata Quoy \& Gaimard, 1832 ..... 235
Sepia simoniana Thiele, 1920 ..... 240
Sepia angulata n. sp. ..... 242
Sepia hieronis (Robson, 1924) ..... 243
Sepia insignis Smith, 1916 ..... 248
Sepia robsoni (Massy, 1927) ..... 250
Sepia faurei n. sp. ..... 251
Sepia (Hemisepius) typica (Steenstrup, 1875) ..... 257
Sepia (Hemisepius) dubia Adam \& Rees, 1966 ..... 264
Sepiella cyanea Robson, 1924 ..... 266
Discussion
Relationships ..... 268
Geographical distribution ..... 271
Vertical distribution ..... 276
Growth ..... 278
Summary ..... 280
Acknowledgements ..... 280
References ..... 280
Appendix: Tables 9-53 ..... 283

## Introduction

The aim of this paper is to collate and extend, where possible, the knowledge of the Sepiidae of southern Africa (defined by Day 1967: vii, as Africa south of the twentieth parallel of latitude). Hitherto, the sepiids have been described in various papers dealing with South African cephalopods (e.g. Robson 1924a, $b$; Massy 1925, 1927, 1928; Voss 1962b, 1967) and in some detail by Adam \& Rees (1966) in their excellent review of the family. The collection in the South African Museum, however, comprises some 664 animals and 4II shells, belonging to 18 species, and these specimens provide a number of interesting additions to our knowledge of the southern African Sepiidae. Three new species and the female animal of Sepia insignis (previously known only by its shell) are described.

All but two of the species have been redescribed from the specimens available and compared with previous descriptions. Unfortunately the collection is poor in species occurring off the Natal coast (called here the 'doratosepion' group). This gap has been filled to some extent by the donation of a few Sepiidae caught off the Moçambique coast by the Oceanographic Research Institute, Durban, and by the loan of some Sepiidae from the Natal Museum, Pietermaritzburg. Most of the latter specimens have previously been described by Miss A. L. Massy (1925, 1928). No specimens were available for Sepia robsoni and S. (Hemisepius) dubia. Only one specimen, the type, is known of each of these two species; these types are deposited in the British Museum (Natural History), and are not sent out on loan.

Before his death Dr. K. H. Barnard compiled notes on the South African Sepiidae and constructed a rough key to the species. But since several specimens have been added to the collection in the interim, and the notes include a number of errors, the present work was not based on his notes, but was started afresh. Some points of interest found in Barnard's notes are discussed under the relevant species.

This paper was submitted in partial fulfilment of the requirements of the Degree of Master of Science in Zoology at the University of Stellenbosch.

## Methods

All animals in reasonable condition were measured with dividers and millimetre rule. Each dimension was calculated as a percentage of dorsal mantle length for animals and of shell length for shells. For animals of mantle length less than 25 mm , only mantle dimensions were recorded, as the animals are too small to handle without damaging them. Measurements of southern African Sepiidae previously published by other authors were included where possible to increase the numbers to a significant level. Tables of all relative dimensions are given at the end of the paper. For some of the more significant dimensions (mantle width, head width, fin width, length of tentacular club and shell dimensions) ranges and arithmetic means were calculated. Calculations
for male and female animals of each species were carried out separately, but shell dimensions of both sexes were combined, since in the case of a shell found on the beach, the sex of the animal by which it was secreted cannot be determined (except perhaps in the case of Sepiella cyanea).

The dimensions of preserved specimens are affected by a number of factors. The relative mantle width is generally greater in dead animals than in live ones, as the mantle collapses and flattens out. However, since all animals measured were dead and fixed in formalin, one may assume the error to be fairly constant. Relative arm lengths are not considered to be of much significance, since they vary considerably, depending on the extent of contraction. However their ranges and means were calculated for comparative purposes, where only large differences may be considered significant. The length of the tentacles is similarly affected, but more so, as the tentacles seem to be more contractile than the arms. In addition, some animals were preserved with the tentacles retracted, and these tentacles were sometimes difficult to straighten out for measuring. Similarly the presence or absence of keels on the arms frequently appears to depend on the condition of the animal at the time of preservation.

In most cases coloration is an unreliable guide within the Sepiidae, as these animals are masters of the art of camouflage, and can show a wide range of colour patterns. Generally the background is a pale cream, overlaid by brownblack and orange-red chromatophores. By progressive expansion of these chromatophores, the animal can assume a range of colours from black, through purple to reddish-brown, over all or part of the body. With complete contraction of the chromatophores only the pale background is seen.

Five of the species described below, namely Sepia confusa, S. incerta, S. burnupi, S. joubini and S. adami, have been called the 'doratosepion' group for convenience, since these species seem to be closely related and are frequently discussed as a group or compared with each other in the text. Most of these species have been included at some time in Rochebrune's genus Doratosepion. This genus was created for Sepiidae with an elongated body, short arms with biserial suckers, short tentacular clubs with unequal suckers, a very elongated shell with two posterior wings and a spine. Whilst Rochebrune's classification of the Sepiidae has been rejected by Adam (1944) and Adam \& Rees (ig66) with good reason, 'doratosepion' is used here as a collective name for the abovementioned five species. Its use, however, in no way implies the retention of the genus Doratosepion.

Distribution ranges have been constructed from localities of specimens in the South African Museum collection and from published records, and have in some cases been extended by locality records from the University of Cape Town Ecological Survey. In the distribution figure (Fig. 18) only localities of animals are recorded. Shell localities cannot be considered to extend the distribution range of a species, since sepiid shells are known to drift over long distances, and their place of origin is thus unknown.

Finally, it should be noted that the Natal Museum has listed the shells of
animals in the collection under separate numbers, e.g.:
N.M.956: S. incerta, $10^{\hat{1}}$ from Natal coast, in stomach of Ground Shark.
N.M.957: S. incerta, I of from the same locality.
N.M.958: shells of the above two specimens (and one shell of S. burnupi, presumably added later).

## Abbreviations and Glossary

MLd -dorsal mantle length along midline
MLv -ventral mantle length along midline
MW -maximum mantle width (excluding fins)
HL -head length dorsally (from anterior tip of nuchal cartilage to edge of dorsal interbrachial membrane)
HW -maximum head width (usually across the eyes)
FL -length of single fin along curve of mantle
FW -width of single fin, from lateral edge of mantle to free edge of fin
AL I-IV -arm length measured from inner base of most proximal sucker to tip of arm
TL -length of tentacle, from point of emergence from tentacular sac to tip of club
$\mathrm{Tcl} \quad$-length of tentacular club, from basal sucker to tip of club
Shell:
L -total length of shell, excluding posterior spine (where present)
W -maximum width of shell
Th -maximum thickness of shell, along midline
Str z -length of striated zone
km -kilometres
m -metres
mm -millimetres
$\mathrm{N} \quad$ - number of specimens measured
N.M. - Natal Museum
O.R.I. - Oceanographic Research Institute
P.F. - Cape Fisheries survey vessel Pieter Faure
S.A.M. - South African Museum
U.C.T. - University of Cape Town Ecological Survey
acuminate -forming an acute angle
arm length formula - comparative lengths of the arms in decreasing order
attenuated -suddenly becomes very slender distally (usually referring to arm tips)
'doratosepion' group-includes Sepia confusa, S. incerta, S. burnupi, S. joubini and S. adami. See page 195 for definition.
emarginate -with a broad semicircular or rectangular notch (usually referring to anterior mantle margin ventrally)

## Key to the Sepidae of Southern Africa

Figures I and 2 illustrate the external morphology of Sepia and its shell, and most of the terms used in the keys. Sepia angulata is not included in the first key as this species is known only by its shell.
I Posterior gland present, opening via a pore situated between the posterior extremities of the fins (genus Sepiella) Sepiella cyanea
Posterior gland absent (genus Sepia) ..... 2
2 Tentacular club with numerous subequal suckers (Figs 14c, 17b) ..... 3
Tentacular club with a few median suckers enlarged (Fig. II) ..... 11
3 Mantle produced dorsally ..... 4
Mantle convex dorsally ..... 7
4 Mantle ventrally entire ..... 5
Mantle ventrally emarginate ..... S. insignis
5 Dorsal arms with biserial suckers S. hieronis
Dorsal arms with quadriserial suckers ..... 6
6 Buccal membrane with a few small suckers S. zanzibarica
Buccal membrane without suckers S. acuminata
7 Mantle ventrally entire S. simoniana
Mantle ventrally emarginate ..... 8
8 Tips of dorsal arms finger-like, devoid of suckers (Fig. 16a) ..... 9
Tips of dorsal arms normal, with suckers to the tips ..... 10
9 Few or no papillae dorsally along outline of shell and on head ..... S. robsoni
Densely papillose dorsally (Fig. 15a) S. faurei
10 About 12 pairs of pores in the ventral mantle surface (Fig. 17b) S. (Hemisepius) typica No pores in the ventral mantle surface S. (Hemisepius) dubia
${ }_{11}$ Skin tuberculate dorsally; ventrally with two large wrinkled patches on the mantle (Fig. 12) ..... 12
Skin smooth dorsally; no ventral wrinkled patches ..... 13
12 Diameter of large tentacular suckers approximately equal to width of club (Fig. IIb) S. papillata
Diameter of large tentacular suckers less than width of club (Fig. I ia). S. tuberculata
13 Shell ovate, width $32-46 \%$ length; inner cone well developed, reflexed and completely fused to outer cone (Pls 35c, d, 36 a, b) S. officinalis vermiculata
Shell elongate; inner cone reduced, with narrow limbs ..... 14
14 Shell broadly elongate, width $29-37 \%$ length; no posterior wings on outer cone (Pl. 39a, b). 'Light organ' in mantle cavity S. australisShell narrow elongate, width $14-26 \%$ length; outer cone with posterior wings (Fig. 2).No 'light organ' in mantle cavity. ('doratosepion' group)15
15 Males (male of $S$. adami not known) ..... 16
Females. The females of the 'doratosepion' group are difficult to separate, and this part of the key is very tentative ..... 19
16 Dorsal arms normal ..... 17
Dorsal arms modified (Figs 6c, 8e) ..... 18
17 Fins rounded posteriorly ..... S. joubini
Fins extended posteriorly to form 'tail' (Fig. 4) ..... S. confusa
18 Ventral arms with hectocotylized region and distal cirri (Fig. 8c, d); fins extended into points posteriorly
Ventral arms without hectocotylized region or cirri; fins rounded posteriorly ..... S. incerta


Fig. 1. General morphology of the Sepiidae, illustrated by Sepia australis. a. External features; b. the mantle cavity (the 'light organ' is characteristic of S. australis and does not occur in other Sepiidae); c. dorsal arm, to show the quadriserial arrangement of the suckers. Longitudinal series numbered $1-4$. The suckers of series 1 and 4 are obscured distally by the protective membranes.
19 Lateral arms attenuated distally20
Lateral arms not attenuated distally
S. adami

20 Lateral arms attenuated over their distal half; distal suckers biserial 21 Lateral arms attenuated over less than half the arm length; distal suckers quadriserial . 22

21 Protective membranes on distal part of dorsal arms expanded . S. joubini Protective membranes on distal part of dorsal arms not expanded - S. confusa

22 Shell with inner cone raised posteriorly; striated zone convex; striae convex (Fig. 6d)
S. incerta

Shell with inner cone low posteriorly; striated zone $M$-shaped; striae angular
(Fig. 8a, Pl. 4d) . . . . . . . . . . . S. burnupi


Fig. 2. Ventral view of the shell of Sepia joubini (A30141) showing some of the features mentioned in the descriptions.

Length 37 mm .

## Key to Shells Only

Sepia robsoni is not included in this key, as its shell is insufficiently known.
I Posterior spine present
2 Ventral part of inner cone well developed . . . . . . . . 3
Ventral part of inner cone reduced . . . . . . . . . 4
3 Ventral part of inner cone not reflexed (Pl. 35b) S. zanzibarica
Ventral part of inner cone reflexed and completely fused to outer cone (Pls $35 \mathrm{~d}, 36 \mathrm{~b}$ )
S. officinalis vermiculata
4 Outer cone with posterior wings (Fig. 2) ..... 5
Outer cone without posterior wings ..... 8
5 Inner cone raised posteriorly, forming a deep pocket over the end of the phragmocone (Pls 37d, 39b) ..... 6
Inner cone low posteriorly (Pl. 38 d ) ..... 7
6 Posterior part of inner cone with a deep longitudinal groove (Fig. 6d) ..... S. incerta
Posterior part of inner cone rounded, without longitudinal groove (Fig. 5) S. confusa
7 Limbs of inner cone raised, lying on the phragmocone; striae $\Lambda$-shaped (Pl. 38d, Fig. 8a) S. burnupi
Limbs of inner cone not raised, lying at sides of phragmocone; striae convex
(Figs 2, 10a) *S. joubini, S. adami
8 Shell rhomboidal, posterior spine not keeled (Pl. 37a, b) S. acuminata
Shell elongate oval, posterior spine keeled (Pl. 39a, b) S. australis
9 Phragmocone considerably shorter than dorsal shield ..... 10
Phragmocone almost as long as dorsal shield ..... 12
io Anterior margin of phragmocone transverse, not parallel with corresponding margin of dorsal lamella (Fig. i7d) ..... II
Anterior margin of phragmocone convex, more or less parallel with corresponding margin of dorsal lamella (Fig. 16c) S. faurei
I I Shell dorsally chitinous, inner cone indistinct (Fig. I7c, d) S. (Hemisepius) typica
Shell dorsally calcified, inner cone distinct S. (Hemisepius) dubia
12 Outer cone expanded posteriorly, inner cone reduced (Pl. 42d) (genus Sepiella) Sepiella cyanea Outer cone narrow posteriorly, inner cone well developed and completely reflexed ..... 13
13 Shell rounded anteriorly ..... 14
Shell acuminate anteriorly ..... 17
14 Ventral surface flat or concave (Pls 39d, 4od) S. tuberculata
Ventral surface convex ..... 15
I5 Limbs of inner cone fairly broad, narrowing gradually anteriorly ..... 16
Limbs of inner cone broad posteriorly, narrowing suddenly anteriorly ( Pl .42 b ) S. simoniana
16 Shell very broad (width 50-60\% length) (Pl. 45); angle between striated zone andsmooth zone pronounced in lateral view (Pl. IOd) . . . . . S. angulataShell elongate oval (width $37-55 \%$ length) (Pl. 41); no pronounced angle betweenstriated zone and smooth zone in lateral view
${ }_{17} 7$ Limbs of inner cone lie at sides of phragmocone (Pl. 43) ..... - S. hieronis
Limbs of inner cone raised, lying on phragmocone - S. insignis

## Systematic Discussion

## DIAGNOSES OF FAMILY AND GENERA

## Family Sepiidae

Mantle short, oval or rounded, dorso-ventrally flattened; fins lateral, occupying almost entire lateral margin of mantle; eye covered by a continuous membranous lid; arm suckers usually quadriserial, occasionally biserial; left

[^3]ventral arm of male usually hectocotylized; tentacles completely retractable, tentacular club distinct from stalk, with subequal or unequal suckers. Shell internal, usually calcareous; phragmocone retained; conotheca reduced ventrally, represented by inner cone; posterior spine present or absent.

Genus SEPIA Linnaeus, $775^{8}$
Diagnosis as for family. Posterior gland absent; tentacular club with subequal or unequal suckers; locking apparatus simple, oval (Fig. 3a). Outer cone of shell not expanded posteriorly.


Fig. 3. Comparison of the mantle locking apparatus of a. Sepia (S. australis, ㅇ, A30154) and b. Sepiella (S. cyanea, ${ }^{\top}$, A 6526 ). Left, mantle component; right, funnel component. Scale $=1 \mathrm{~mm}$.

Subgenus Sepia s.s.
Mantle slender to moderately broad; arm suckers biserial or quadriserial; tentacular suckers subequal or unequal. Shell slender to broadly oval, length approximately equal to dorsal mantle length; phragmocone occupies almost all dorsal shield; posterior spine present or absent.

Subgenus Hemisepius Steenstrup, 1875
Mantle very broad; arm suckers biserial; tentacular suckers subequal. Shell very thin, phragmocone shorter than dorsal shield; posterior spine absent.

Subgenus Metasepia Hoyle, 1885
Mantle broadly oval; arm suckers quadriserial; tentacular club with few unequal suckers. Shell rhomboidal, much shorter than mantle; posterior spine absent. (No southern African representatives.)

Genus SEPIELLA Gray, 1849
Posterior gland present; tentacular club with numerous subequal suckers; locking apparatus with tubercle on mantle component and corresponding depression in funnel component (Fig. 3b). Shell without posterior spine; outer cone expanded posteriorly.

DESGRIPTION OF SOUTHERN AFRICAN SPECIES
Sepia zanzibarica Pfeffer, 1884
(Pl. 35a, b. Tables 9, io)
Sepia zanzibarica Pfeffer, 1884: 9, fig. in, ıı. Hoyle, 1886: 22, 217. Smith, 1916: 21. Tomlin, 1923: 40. Massy, 1925: 2 II. Voss, 1962b: 248. Adam \& Rees, 1966: 7, pl. 2, figs 9-11, pl. 41, fig. 247.

## Type locality <br> Zanzibar.

## Distribution

Animals: Zanzibar (Pfeffer 1884: 9), German East Africa (Massy 1925: 21 I). Depth not known.
Shells: Mombasa (Adam \& Rees 1966: 7) to Tongaat, Natal (Smith igı6: 21); also Malagasy (Adam \& Rees $1966: 7$ ).

## Material

N.M. 959, 960, German East Africa (det. A. L. Massy); i đ
S.A.M. A2141, Chinde, mouth of Zambezi River; 2 shells

Locality unknown; i shell
Description
Male (N.M.959) originally described by Massy (1925:211), now in rather poor condition.

Mantle broadly oval, anterior mantle margin produced dorsally, ventrally entire. Fins narrow, rounded, separate posteriorly. 'The buccal membrane has a single sucker on five of its tips' (Adam \& Rees 1966: 8). These not clearly visible in male, due to distorted state of buccal membrane.

Skin smooth, except for few tiny papillae mid-dorsally on mantle. Colour dark purple dorsally, with paler fins. Narrow dark purple line along fin bases. Colour ventrally lighter purple laterally on mantle, fading to mottled buffpurple mid-ventrally.

Arms unequal in length; shortest dorsally, longest ventrally, of formula 4.3.2.I. In female, arms I to III subequal, arms IV considerably longer (Pfeffer 1884 : 9). Arms joined by shallow interbrachial web, except between ventral pair. All arms keeled, provided with well-developed protective membranes folding over inner surface; arm tips attenuated.

Suckers on all arms quadriserially arranged to tips. Chitinous rings of suckers smooth.

Left ventral arm hectocotylized. Basal two-thirds of arm bearing quadriserial suckers; suckers minute over six rows on distal third (as described by Massy 1925: 21 I). Adam \& Rees (1966: 8) were mistaken in presuming that the hectocotylus was situated on the proximal third of the arm. Three dorsal series of minute suckers in normal position, ventral series displaced, leaving naked, grooved region on arm; tip of arm bearing normal quadriserial suckers.

Tentacular club long, bearing numerous subequal suckers in rows of about six, 'but probably form oblique transverse rows of eight' (Adam \& Rees 1966: 8). Of these, three median series of suckers a little larger than others. In addition, three larger distal suckers partially concealed by reflexed tip of club. Chitinous rings of tentacular suckers finely dentate. Protective membranes well developed, remaining separate proximally and continuing along tentacular stalk for some distance. Natatory membrane a little longer than club.

Shell (N.M.96o) of male specimen badly damaged. Three other shells (Pl. 35a, b) in fairly good condition, but somewhat worn.

Shell broadly oval, tapering anteriorly and posteriorly. Dorsal surface roughly granular posteriorly, more finely so anteriorly. Median ridge ill-defined. Two fairly well defined dorsal grooves diverging from posterior end correspond with position of ventral limbs of inner cone. Posterior spine short but strong, directed dorsally and coloured blue; spine not keeled. Striated zone long ventrally, occupying two-thirds to three-quarters shell length. Striae broady $\Lambda$-shaped, becoming somewhat more rounded anteriorly. Median groove broad, with phragmocone raised on either side. Inner cone very well developed posteriorly, free, curving over posterior part of striated zone to form a pocket. Limbs of inner cone broad, curving over lateral edges of phragmocone. Outer cone of present shells damaged, but according to Pfeffer's figure (fig. ira) it broadens somewhat posteriorly without actually forming wings. Shell thickest on either side of midline, near anterior end of striated zone.

## Remarks

It is not certain if this may be considered a southern African species, since only shells have been found on our coasts.*

Tomlin (1923: 40) mentions a specimen, presumably a shell, from Isipingo, but gives no details.

## Sepia officinalis vermiculata Quoy \& Gaimard, 1832

$$
\text { (Pls } 35 \mathrm{c}, \mathrm{~d}, 36 \mathrm{a}, \mathrm{~b} \text {. Tables } \mathrm{I}, 2, \mathrm{I} \mathrm{I}-\mathrm{I} 3)
$$

Sepia vermiculata Quoy \& Gaimard, 1832 : 64 , pl. I, figs I-5. Férussac \& d'Orbigny, 1835-1848: 279, pl. IIIbis. Gibbons, 1888: 202. Bartsch, 1915: 250. Smith, 1916: 20. Robson, 1924a: 12. Massy, 1925: 209; 1928: 91 .
Acanthosepion vermiculatum Rochebrune, 1884: 113. Adam, 1944: 234.
Acanthosepion vermiculata: Robson, 1924b: 639. Massy, 1927: 156.
Sepia officinalis vermiculata Adam, 1940: 130; 1941: 99, 102, 106, pl. IV, fig. 1; 1962: 1 1. Voss, 1962b: 248, 249. Adam \& Rees, 1966: 30, pl. 10, figs 55, 56, pl. 45, fig. 27 I .
? Sepia jousseaumi Rochebrune, 1884: 117. Smith, 1916: 22. Adam, 1941: 108, pl. IV, fig. 3; 1944: 235.
? Sepia jousseaumei: Bartsch, 1915: 250.
? Sepia hierredda (non Rang) Turton, 1932: 2.

## Type localities

Cape of Good Hope (S. vermiculata and $S$. jousseaumi).

## Distribution

Animals: $30^{\circ} 42^{\prime} \mathrm{S}, 15^{\circ} 59^{\prime} \mathrm{E}$ (Voss 1962b: 250) to Delagoa Bay, Moçambique (Massy 1927: 156; Adam 1962: in). Depth o-248 m.
Shells: Saldanha Bay to Chinde (S.A.M.).

## Material

S.A.M. A2 I43, Chinde, mouth of Zambezi River; I shell

A2 144, Durban; i shell
A3or25, $\mathrm{S}_{2}{ }^{\circ} \mathrm{W}$ of Cape Point, $23 \mathrm{~km}, \mathrm{I} 56 \mathrm{~m} ; 4$ juveniles
A3oi 28, SE of Cape St. Blaize, $9 \mathrm{~km}, 62 \mathrm{~m}$; i juvenile
A30129, Swartkops; i juvenile
A3or30, Algoa Bay fishing grounds; I $\widehat{0}$
A3or3r, Table Bay; i 9
A30182, locality unknown; I đ
A30183, locality unknown; i q
A30483, Still Bay; I shell
A30487, locality unknown (det. A. L. Massy); i shell
A30496, locality unknown; i shell
A31238, off Hartenbos, near Mossel Bay, 18-24 m; 5 우
A31292, $33^{\circ}$ oi'S, $17^{\circ} 5^{\prime} \mathrm{E}$ (Saldanha Bay); i $q$

[^4]Locality unknown; i $q$
Knysna estuary; 30 ô, 18 ㅇ
Durban Bay; I ô, 6 ㅇ
Ysterfontein beach; i shell
Nature's Valley; i shell, discarded
Breede River mouth; i shell
Krom River mouth, Cape St. Francis; 12 shells
Saldanha Bay; 2 shells

## Description

Mantle broadly oval, anterior margin somewhat produced dorsally, ventrally entire. Head short and broad; fins broad and rounded but separate posteriorly. Mantle slightly more slender and fins slightly wider in males than in females.

Skin smooth, except in one male (A30I30), which is sparsely papillose dorsally, mainly on head. No indication in any specimens of long ridge-shaped tubercles near fins, as mentioned by Massy (1925: 210), although some have pale pink round spots in this region. Colour dark dorsally, pale ventrally and some specimens show the well-known transverse zebra-like stripes dorsally on mantle and on fourth arms, or at least at fin bases. Three specimens (males A30130 and A3or82 and female, locality unknown) with pale stripes on dark background, but one female ( $\mathrm{A}_{3} \mathrm{OI} 83$ ) with dark stripes on pale background.

Arms longest ventrally, shortest dorsally, with arm length formula 4.3.2.I. Arms joined by shallow web; arms III and IV keeled, arms II sometimes keeled, arms I usually not-depending on manner of preservation. Arm tips somewhat attenuated. Suckers quadriserially arranged on all arms to tips, decreasing regularly in size from arm base. All sucker rings finely dentate, distal teeth being longer than proximal ones. Protective membranes well developed.

Left ventral arm of male hectocotylized basally. About six normal suckers at base of arm followed by 9-12 rows of modified suckers. The latter much smaller and separated by transverse ridges on arm. Arm normal distally.

Tentacular club bearing small distal suckers in oblique rows of eight. Suckers on proximal part of club variously enlarged: from ventral side, first series of suckers of normal size, second series $1,5^{-2}$ times as big as first series, third series 2,5-3 times as big as first series, fourth series $\mathrm{I}-\mathrm{I}, 5$ times as big as first series, fifth series same size as first series. Rings of large club suckers smooth, those of smaller suckers toothed distally. One sucker at tip of club concealed by reflexed tip; immediately below this, two suckers, about twice as big as their proximal neighbours, partly concealed by tip. Protective membranes of club not meeting proximally; natatory membrane a little shorter than club.

Shell (Pls 35c, d, 36a, b) broadly oval, tapering somewhat anteriorly and posteriorly; posterior spine present. Posterior end of shell and base of spine covered with horny covering. Dorsal surface of shell tuberculate, with fairly broad chitinous margin. No marked dorsal ridge. Ventral striated zone fairly
long (about half total length), with median longitudinal ridge. Anterior border of striated zone convex on either side of median ridge. Inner cone broad, reflexed and fused to broad outer cone.

Seven shells from Durban Bay have two deep lateral grooves in ventral surface ( Pl .35 d ). These grooves do not occur in any other shells, and it is strange that all seven shells should have them, as these animals, though from the same locality, were not all caught at the same time (two were caught in February 1970 and the other five in April 1970).

## Remarks

Adam (1941: 104) has shown that Sepia officinalis in the eastern Atlantic Ocean is represented by four geographic races:
S. o. officinalis Linnaeus, 1758: from the Atlantic coast of France to Rio de Oro (Cap Blanc).
S. o. filliouxi Lafont, 1868 : from the Atlantic coast of France to the southern coast of Scandinavia.
S. o. hierredda Rang, 1837: from south of the Baie du Lévrier (Mauritania) to the coast of Angola.
S. o. vermiculata Quoy \& Gaimard, 1832: southern Africa.

In addition a fifth race, S. o. mediterranea Ninni, 1884, lives in the Mediterranean Sea (Adam \& Rees 1966: 32).

Table i. A comparison of the relative dimensions (as \% MLd) of the animals of the races Sepia officinalis vermiculata and Sepia officinalis hierredda. Only animals with dorsal mantle length greater than 100 mm are included. The figures for $S$. officinalis hierredda were calculated from relative dimensions given by Adam (1941).

|  |  | $\begin{gathered} \text { Fen } \\ \text { Range } \end{gathered}$ | Mean | Males <br> Range | Mean |
| :---: | :---: | :---: | :---: | :---: | :---: |
| MW | S. o. vermiculata <br> S. o. hierredda | $\begin{gathered} 43,8-63, \mathrm{I} \\ 4^{\mathrm{I}-5 \mathrm{I}} \end{gathered}$ | $\begin{aligned} & 55,0 \\ & 46,0 \end{aligned}$ | $\begin{gathered} 38,4-57,4 \\ 40-51 \end{gathered}$ | $\begin{aligned} & 49,9 \\ & 44, \mathrm{I} \end{aligned}$ |
| HW | S. o. vermiculata <br> S. o. hierredda | $\begin{array}{r} 34,9-50,0 \\ 3 \mathrm{I}-4^{2,5} \end{array}$ | $\begin{aligned} & 43,4 \\ & 35,3 \end{aligned}$ | $\begin{gathered} 36,9-46,3 \\ 28-36 \end{gathered}$ | $\begin{aligned} & 4 \mathrm{I}, 5 \\ & 3 \mathrm{I}, 9 \end{aligned}$ |
| AL I | S. o. vermiculata <br> S. o. hierredda | $\begin{aligned} & 27,7-55,0 \\ & 26,5-33,5 \end{aligned}$ | $\begin{aligned} & 38,9 \\ & 30,2 \end{aligned}$ | $\begin{aligned} & 28,8-46,3 \\ & 26,5-40 \end{aligned}$ | $\begin{aligned} & 37,3 \\ & 34, \mathrm{I} \end{aligned}$ |
| AL II | S. o. vermiculata <br> S. o. hierredda | $\begin{array}{r} 29,8-57,9 \\ 28-36,5 \end{array}$ | $\begin{aligned} & 42,7 \\ & 31,3 \end{aligned}$ | $\begin{gathered} 34, \mathrm{I}-49,3 \\ 30-40 \end{gathered}$ | $\begin{aligned} & 39,4 \\ & 36,1 \end{aligned}$ |
| AL III | S. o. vermiculata <br> S. o. hierredda | $\begin{aligned} & 31,9-60,3 \\ & 28,5-37,5 \end{aligned}$ | $\begin{aligned} & 44,0 \\ & 33,8 \end{aligned}$ | $\begin{aligned} & 35,7-54 \\ & 33,5-43 \end{aligned}$ | $\begin{aligned} & 43,3 \\ & 39,2 \end{aligned}$ |
| AL IV | S. o. vermiculata <br> S. o. hierredda | $\begin{aligned} & 37,2-79,3 \\ & 32,5-4^{8} \end{aligned}$ | $\begin{aligned} & 53,6 \\ & 39,5 \end{aligned}$ | $\begin{gathered} 40,5-63,2 \\ 40-63 \end{gathered}$ | $\begin{aligned} & 48,9 \\ & 50,2 \end{aligned}$ |
| Tcl | S. o. vermiculata <br> S. o. hierredda | $\begin{aligned} & 25,7-38,2 \\ & 20,5-24,5 \end{aligned}$ | $\begin{aligned} & 30,4 \\ & 23,0 \end{aligned}$ | $\begin{array}{r} 23,8-34 \\ 19-23 \end{array}$ | $\begin{aligned} & 28,2 \\ & 22,2 \end{aligned}$ |

Adam (1941: 103) was able to show some differences in the shells of the Atlantic races, but could find no marked differences in the forms of the animals. He observed that $S$. o. vermiculata differs from $S$. o. hierredda in that the shell is relatively wider and thicker and is more strongly tuberculate dorsally. Adam \& Rees (1966: 32) added that the posterior part of the shell of $S$. o. vermiculata seems to be more broadly rounded than that of $S$. o. hierredda.

A comparison of the measurements of the $S$. o. vermiculata specimens described above, with those given for $S$. o. hierredda by Adam (1941: 157-159), shows that there are indeed some differences, but only in specimens having a dorsal mantle length exceeding 100 mm . The most marked differences between the two races is observable in the tentacular clubs (Table i). The clubs of S. o. vermiculata are relatively longer than those of S. o. hierredda, and there is apparently no overlap in the ranges of this dimension for the two races. The differences in the other relative dimensions are less marked, but $S$. o. vermiculata has a relatively wider mantle and head, and longer arms (except the fourth arm in the males) (Table 1). The relative dimensions of the shells also show some differences between the two races (Table 2). The shells of S. o. vermiculata are relatively wider and thicker and have a slightly shorter striated zone. These differences become more marked, the larger the shells.

Table 2. Comparison of relative shell dimensions (as \% L) of S. officinalis vermiculata and $S$. o. hierredda. Only shells of length greater than 100 mm are considered. The figures for $S$. o. hierredda were calculated from relative dimensions given by Adam (1941).

|  |  | Range | Mean |
| :---: | :---: | :---: | :---: |
| Width | S. o. vermiculata <br> S. o. hierredda | $\begin{array}{r} 32,8-41,7 \\ 30-36,5 \end{array}$ | $\begin{aligned} & 38,5 \\ & 34,0 \end{aligned}$ |
| Thickness . | S. o. vermiculata <br> S. o. hierredda | $\begin{aligned} & 9,1-15,2 \\ & 9,4^{-12,6} \end{aligned}$ | $\begin{aligned} & 12,5 \\ & 11,1 \end{aligned}$ |
| Length of striated zone | S. o. vermiculata <br> S. o. hierredda | $\begin{aligned} & 40,5-74,8 \\ & 43,5-70,5 \end{aligned}$ | $\begin{aligned} & 50,3 \\ & 57,5 \end{aligned}$ |

The males of $S$. o. vermiculata and S. o. hierredda cannot be distinguished by their hectocotyli. The hectocotylus of $S$. o. hierredda has 8-13 transverse rows of modified suckers (Adam 1941: io6). In the present specimens of S. o. vermiculata the hectocotylus has $9-12$ rows of modified suckers. The extent of sucker modification seems to increase with the size of the animal. Massy (1925: 210) reported a large male of $S$. o. vermiculata (MLd $6 \frac{1}{2}$ inches $=165 \mathrm{~mm}$ ) having 17 rows of modified suckers on the hectocotylus.

The southernmost record of $S$. officinalis hierredda is the Baia dos Tigres, Angola (about $15^{\circ} 20^{\prime} \mathrm{S}$ ); S. officinalis vermiculata is known to occur as far north as $30^{\circ} 42^{\prime} \mathrm{S}$ (north of the Olifants River) off the west coast of southern Africa
(Fig. 18). Dr. M.-L. Penrith (personal communication) points out that no sepiid shells were found on any but the northernmost beaches of South West Africa, although much other debris was washed up. In Angola, where S. o. hierredda is known to occur, sepiid shells were present on the beaches. The apparent paucity of Sepiidae off the South West African coast suggests that there may be no region of overlap between $S$. o. hierredda and S. o. vermiculata.

On the east coast, animals of $S$. o. vermiculata have been found as far north as Delagoa Bay (Massy 1927: 156; Adam 1962: 11). Barnard collected a shell (A2 I43) from Chinde, near the mouth of the Zambezi River; but this cannot be considered a reliable locality, since sepiid shells have been known to drift over long distances.

It is of interest to note that Sepia officinalis is the only species of Sepia known to occur in estuaries in southern Africa. This, its wide geographic distribution and its division into several geographic races, indicate that it is a very adaptable species.

## Sepia acuminata Smith, 1916

(Pl. 37a, b. Tables $14-\mathrm{I} 6$ )
Sepia acuminata Smith, 1916: 21, pl. II, fig. 3 (partim). Tomlin, 1923: 40. Robson, 1924a: 12 Massy, 1928: 91, pl. VIII, figs 1-7. Turton, 1932: 1. Voss, 1962b: 248. Adam \& Rees 1966: 53, pl. 16, figs 91, 92, pl. 43, fig. 26i.
non Sepia acuminata Smith, 1916: non pl. II, fig. 4 ( $=$ S. hieronis).
Rhombosepion acuminata Robson, 1924b: 643 .
Sepia sp. Adam, 1941: 121, pl. IV, fig. 6.

## Type localities

Port Elizabeth; Tongaat beach, Natal (shells only).

## Distribution

Animals: $29^{\circ} 54^{\prime} \mathrm{S}, 3 \mathrm{I}^{\circ} \mathrm{I} 5^{\prime} \mathrm{E}$ (Robson $1924 b: 643$, Sta. 103) to Punta Zavora, Moçambique (S.A.M.). Depth 64-369 m.
Shells: Cape St. Francis (S.A.M.) to ? Mombasa (Adam \& Rees 1966: 53).

## Material

S.A.M. A30147, locality unknown; 1 đ

A3ı398, Corner, Moçambique, $25^{\circ} \mathrm{I} 5^{\prime} \mathrm{S}, 35^{\circ}$ ıo'E, $\pm 266 \mathrm{~m}$; i đ̂, i $q$
A3r399, Corner, Moçambique, $25^{\circ}$ I $5^{\prime} \mathrm{S}, 35^{\circ}$ Io'E, 248 m ; i đ
A31400, Corner, Moçambique, $25^{\circ}$ I $5^{\prime} \mathrm{S}, 35^{\circ}$ ro'E, 257 m ; i juvenile
A3ı40i, Punta Zavora, Moçambique, $24^{\circ} 35^{\prime} \mathrm{S}, 35^{\circ} 25^{\prime} \mathrm{E}, 220-257 \mathrm{~m}$; 1
 Krom River mouth, Cape St. Francis; i shell (broken)
Umngazana River mouth, west Pondoland; i shell

## Description

Mantle broadly oval, anterior margin produced to fairly sharp point dorsally, halfway along eyes or more; ventrally entire. Fins narrow, rounded and separate posteriorly.

Skin smooth. Coloration (described from recently caught specimens from Moçambique; other specimens considerably faded): dense concentration of purple chromatophores dorsally on mantle, head and arms; fins pale at edges. Chromatophores less dense ventrally, resulting in paler purple colour near fins and brownish colour medially on mantle. Ventral surface of head and arms almost white.

Arms short (less than half MLd), but somewhat longer in females than in males. Arms I to III subequal in length, arms IV somewhat longer. Shallow interbrachial web present, except between arms IV. All arms keeled to some extent and bear moderately well developed protective membranes.

Arm suckers globose, quadriserially arranged to tips of arms. Chitinous rings of suckers finely toothed distally, with nodular surface; adjacent skin of suckers wrinkled.

Left ventral arm of male hectocotylized distally. Six or seven rows of normal suckers on basal third of arm, followed by nine to ten modified rows, in which all suckers greatly reduced in size. Two dorsal series of modified suckers in normal position, but the two ventral series have moved close together on ventral edge of sucker-bearing surface, almost forming single line. Thus two dorsal series widely separated from two ventral series by naked, wrinkled region. Distal third of arm with normal suckers, decreasing in size to arm tip.

Tentacular club recurved, bearing numerous small subequal suckers in transverse rows of eight. Reflexed tip of club obscures two somewhat larger suckers. Sucker rings toothed distally; protective membranes separate proximally; natatory membrane somewhat longer than club.

Shell (Pl. 37a, b) broadly elongate, sharply pointed anteriorly, rounded posteriorly, bearing spine with tip directed dorsally; spine not keeled. Dorsal surface of shell usually pink in colour, with broad chitinous margins laterally and with median ridge sometimes sunken below level of rest of dorsal surface. Striated zone long ventrally (about two-thirds total length); median groove very faint or absent; striae regularly convex. Inner cone not much thickened posteriorly, with narrow limbs curving over lateral edge of striated zone. Outer cone only slightly calcified and mostly chitinous posteriorly, where it curves sharply in ventral direction. Shell thickest at anterior end of striated zone.

## Remarks

In Smith's (1916: 21) original description, he mentioned and figured (pl. II, fig. 4) one specimen from Tongaat Beach which differed from the other examples of $S$. acuminata. From the figure, this specimen is clearly $S$. hieronis (cf.).

The shell from the 'Ph. Dautzenberg' collection, illustrated by Adam (1941, pl. IV, fig. 6) looks very much like that of S. acuminata.*

Adam \& Rees (1966:53) reported some broken shells, almost certainly pertaining to $S$. acuminata, from Mombasa. This identification is the more probable since $S$. acuminata is now known to occur as far north as Moçambique, and is apparently a subtropical species. The male specimen reported by Adam \& Rees from the U.C.T. Ecological Survey, AFR 105 I K, comes from $29^{\circ} 54^{\prime}$ S, $31^{\circ}{ }^{1} 3^{\prime}$ E, 369 m (ex U.C.T. catalogue).

Sepia confusa Smith, 1916
(Pl. 37 c , d. Figs 4, 5. Tables 17-19)
Sepia burnupi Hoyle, 1904: 27, pl. I, fig. 192 (partim).
Sepia confusa Smith, 1916: 24, pl. II, figs 7, 8. Tomlin, 1923: 41. Robson, 1924a: 12. Turton, 1932: 1. Voss, 1962b: 248. Adam \& Rees, 1966: 65, pl. 18, figs i12, 1 13, pl. 42, fig. $24^{8 .}$
Doratosepion confusa Massy \& Robson, 1923: 435, figs 1-3. Carleton \& Robson, 1924: 259, pl. 3, figs $1-6$.
Doratosepion confusum: Robson, 1924b: 647.
Sepia (Doratosepion) confusa Massy, 1925: 221, pl. XIII, figs 20, 21, 24-28, pl. XIV, fig. 38; 1928: 93.

Type localities
Port Elizabeth; Tongaat beach, Natal (shells only).

## Distribution

Animals: $29^{\circ} 52^{\prime} \mathrm{S}$, $31^{\circ}{ }^{\circ} 7^{\prime} \mathrm{E}$ (off Durban) (Robson 1924a: 12, 1924b: 647, Sta. 95) to $5^{\circ} 39^{\prime} \mathrm{S}, 39^{\circ}$ 16'E (Zanzibar area) (Adam \& Rees 1966: 65). Depth 64-352 m.
Shells: Port Elizabeth (Hoyle 1904: 27) to Chinde, mouth of Zambezi River (S.A.M.).

## Material

S.A.M. A2 i40, Chinde, mouth of Zambezi River; i shell

A6516, off Tugela River, in6-1 ${ }^{2} 4 \mathrm{~m}$; I ${ }^{\text {a }}$
A30292, 35 km S of Tugela River, $116-134 \mathrm{~m} ; 2{ }^{\boldsymbol{\delta}}$, in poor condition A31402, Corner, Moçambique, $25^{\circ}{ }^{1} 5^{\prime} \mathrm{S}, 35^{\circ}{ }^{\circ} 0^{\prime} \mathrm{E}, \pm 266 \mathrm{~m} ; 4$ o $^{\circ}$ A31403, Corner, Moçambique, $25^{\circ}{ }^{1} 5^{\prime} \mathrm{S}, 35^{\circ}{ }^{\circ}$ ıo'E, 257 m ; i ㅇ $\mathrm{A}_{3}$ I404, Punta Zavora, Moçambique, $24^{\circ} 35^{\prime} \mathrm{S}, 35^{\circ}{ }^{2} 5^{\prime} \mathrm{E}, 220-257 \mathrm{~m}$; 2 ठ
N.M. 96ı, 962, 48 km off Tugela River, 64 m (det. A. L. Massy); 2 of
N.M. 963 , Natal coast (det. A. L. Massy); 2 shells

## Description

Mantle elongate, anterior margin strongly produced dorsally, ventrally slightly emarginate. Fins of male very broad, especially towards posterior end, and extended to form 'tail' beyond posterior end of mantle (Fig. 4). 'Tail'

[^5]

Fig. 4. Ventral view of Sepia confusa male, to show 'tail'-like extension of fins. Modified after Massy \& Robson (1923).
length approximately equal to or greater than MLd. Fins of female rounded and separate posteriorly, not forming 'tail'.

Skin smooth, except in one male (A6516) which has few small papillae mid-dorsally on mantle. Colour dark purple mid-dorsally on mantle and above eyes. Rest of body paler pinkish-brown, where chromatophores less expanded. Ventral surface of head almost white.

Arms relatively short (not more than half MLd). Dorsal arms of males slightly longer than others, which are subequal in length. Arms of female subequal. Interbrachial web deepest between arms III and IV, absent between fourth pair. Arms I to III slightly keeled, arms IV more strongly so. Protective membranes fairly well developed, especially along attenuated part of arms, where membrane curves over, partially covering suckers. Arms attenuated at tips in males, over about half arm length in female. No sign of hectocotylization in males.

Arm suckers globose, with smooth rings; quadriserially arranged to tips of arms in males. Proximally, two median series of suckers much larger than lateral series, but on attenuated part of arm all suckers minute and of same size. In female, suckers as those of males proximally, but becoming biserial and widely spaced over flattened attenuated part of arm.

Tentacular club slightly recurved, with distal suckers in oblique rows of eight, and with nine median suckers variously enlarged. Of these, five much larger, other four grading to normal size. Three slightly larger suckers partially concealed by reflexed tip of club. Sucker rings broad and nodular, with finely dentate inner edge. Protective membranes of club moderately well developed, separate proximally. Natatory membrane a little longer than club.

Shell (Pl. 37c, d, Fig. 5) very narrow, pointed anteriorly, with posterior spine directed dorsally. In male specimens from Moçambique, delicate keel runs from base of spine and for some distance along dorsal face of shell; dry shells show no indication of keel on spine. Dorsal surface of shell with narrow, heavily calcified region medially, coloured pink; shell chitinous laterally. Distinct broad median ridge, narrowing posteriorly, limited by lateral grooves. Whole dorsal surface shows concentric striae, $\wedge$-shaped. Ventral striated zone just over half total length. Striae wavy, anterior border of striated zone angular on either side of midline ( $M$-shaped). Median longitudinal groove distinct. Inner cone forms free ledge posteriorly (Fig. 5) and has long narrow limbs. Outer cone broad and deep posteriorly, forming chitinous wings. Shell thick, ventral surface strongly convex; maximum thickness occurring immediately anterior to striated zone.

## Remarks

Detailed structure and possible function of the 'tail' are discussed by Massy \& Robson (1923) and by Carleton \& Robson (1924).

The male of $S$. confusa is distinguished from the other 'doratosepion' species by its 'tail'-like extension of the fins. The female may be separated from all
except $S$. joubini by the biserial sucker arrangement on the attenuated part of the arms. It is distinct from the latter species in that the protective membranes on the tips of the dorsal arms are not expanded as they are in S. joubini.


Fig. 5. Sepia confusa male (A31402). Detail of posterior end of shell, ventral view.

Sepia incerta Smith, I916
(Pls 38a, b, 39a, b. Fig. 6. Tables 20-22)
Sepia burnupi Hoyle, 1904: 27, pl. I, figs 190, 191 (partim).
Sepia incerta Smith, 1916: 23, pl. II, fig. 6. Tomlin, 1923: 41. Voss, 1962b: 248. Adam \& Rees,

Sepia (Doratosepion) incerta Massy, 1925: 219, pl. XIII, figs 22, 23, 29-36, pl. XIV, figs 40, 43.
Sepia (Doratosepion) burnupi (non Hoyle) Massy, 1925: 215, pl. XII, figs 12-19, pl. XIV, figs 39, 41, 42; 1928: 94. Barnard, 1962: 252, fig. 4.
? Sepia incerta: Turton, 1932: 1.
Type localities
Port Elizabeth; Tongaat beach, Natal (shells only).

## Distribution

Animals: $33^{\circ} 07^{\prime} \mathrm{S}, 27^{\circ} 56^{\prime} \mathrm{E}^{*}$ (East London area) (Barnard 1962: 252) to Durban (Adam \& Rees 1966: 67). Depth 70-79 m.

[^6]Shells: Port Elizabeth (Hoyle 1904: 27) to Tongaat beach, Natal (Smith 1916: 23).

## Material


A30480, locality unknown; 2 shells
S.A.M. Si, Punta Zavora, Moçambique; 9 shells
N.M. $956,957,958$ A, B, Natal coast, in stomach of Ground Shark (det. A. L. Massy); $2{ }^{t}, 2$ shells
N.M.969, 970 , Cape Henderson (det. A. L. Massy); 1 , 3 shells

## Description

Mature males: mantle elongate, about three times as long as wide. Anterior mantle margin produced dorsally, emarginate ventrally. Fins narrow, fused posteriorly over tip of mantle.

Chromatophores densest mid-dorsally, fewer towards fins and sides of head; ventrally sparse, forming spots on mantle and, in one large specimen, larger spots on funnel. Series of large spots present along dorsal midline of each fin.

Arm lengths unequal (formula 1.4.2 $=3$ ). Dorsal arms longest, modified (Fig. 6c): proximal quarter bearing io-18 normal suckers basally, then 8-10 suckers on thickened transverse ridges on arm. Above this, protective membranes expanded over about half arm length, supported by transverse thickenings in the membrane. Membranes joined over inner surface, distal to sucker bearing portion of arm; ventral membrane more expanded than dorsal one and forming lamella with maximum width of $\mathbf{1 0}-\mathbf{2 0} \mathrm{mm}$, about three-fifths from arm base. Distal quarter of arm attenuated, with protective membranes rapidly reduced towards arm tip.

Lateral and ventral arms bearing suckers quadriserially arranged almost to tips; median suckers larger than lateral suckers, situated on protective membranes. Suckers on these arms decrease gradually in size towards tipnone enlarged. No sign of hectocotylization on either ventral arm in any males.

Suckers globose; rings very finely serrated, almost smooth. Skin wrinkled immediately adjacent to chitinous rings of suckers.

Arms joined by shallow web, deepest between arms III and IV; no web between ventral pair.

Tentacular club recurved, bearing numerous small suckers distally in obliquely transverse rows of eight. Four or five suckers at proximal end of one of the median rows much enlarged. Large club suckers with finely serrated rings, small ones with toothed rings. Natatory membrane extends a little beyond proximal limit of club; protective membranes separate distally.

Young male (A30143, MLd 93 mm ): dorsal arms shorter than ventral arms, but modified to some extent. Protective membranes well developed,

[^7]

Fig. 6. Sepia incerta. a. Ventral and b. dorsal views of female (A30143). c. Right dorsal arm of male; d. detail of posterior end of shell ( ${ }^{\top}$, A3or43), ventral view. c. Modified after Barnard (1962) ; a. b. and d. original.
increasing gradually in width towards distal end, but not forming marked lamellate expansions found in mature males. Protective membranes only 3 mm wide at maximum width, but already fused over inner surface of arm. On proximal two-fifths, 14 normal suckers and io-12 suckers on transverse ridges. Ventral arms relatively shorter than in mature males.

Females (Fig. 6a, b) : small in comparison with large males, but mature. Ovary well developed, containing numerous oval eggs. Well developed nidamental glands present. Females differ from males in that dorsal arms not modified; arm formula variable. Lateral arm pairs not equal in length, unlike those of males. Dorsal arms of female with well developed protective membranes, but these not expanded into lamellae. All arms of female bear quadriserial suckers. Arm tips (less than half arm length) attenuated, bearing quadriserial suckers and having well-developed protective membranes folding over inner surface of arm.

Shell (Pls 38a, b, 39a, b) narrow, elongate, acuminate anteriorly and posteriorly. Dorsally with wide chitinous margins and three median longitudinal ridges, pink in colour, separated by two longitudinal grooves. Median ridge broad anteriorly, narrowing posteriorly; two lateral ridges narrow and indistinct anteriorly, but broader and more marked posteriorly. Ventrally with shallow longitudinal median groove. Striated zone long and anteriorly convex. Striae $\wedge$-shaped posteriorly, becoming more convex anteriorly. Inner cone with long narrow limbs lying at sides of striated zone; inner cone raised posteriorly to form ledge over end of striated zone. Centre of this posterior ledge bisected by deep longitudinal groove (Fig. 6d). Outer cone forming posterior, chitinous wings, and thickened over base of spine. Spine not keeled; directed dorsally.

## Remarks

The soft parts of Sepia incerta were first described by Massy (1925: 215, 219). She described two males as $S$. burnupi and two females as $S$. incerta, but a re-examination of the shells of these specimens has indicated that in fact all four specimens are referable to $S$. incerta. Barnard (1962:252) described another six specimens of $S$. incerta under the name $S$. burnupi.
$S$. incerta is distinguished from the other species of the 'doratosepion' group as follows: The male of $S$. incerta is characterized by its modified dorsal and unmodified ventral arms. The female is distinct from those of $S$. confusa, $S . j o u b i n i$ and $S$. adami in that less than half the arm length is attenuated distally, and the suckers on the attenuated part are quadriserial. From the female of S. burnupi it can only be separated by the differences in the shells (and the larger size of the animals).

The shell of $S$. incerta differs from those of S. burnupi, S. joubini and S. adami in that the inner cone is raised posteriorly, forming a deep pocket over the end of the striated zone, as in S. confusa. It differs from the latter species in having a deep longitudinal groove in the centre of the raised posterior portion of the inner cone.

Sepia burnupi Hoyle, 1904

## (Pl. 38c, d. Figs 7, 8. Tables 23, 24)

Sepia burnupi Hoyle, 1904: 27, pl. I, figs 188, 189 (partim). Bartsch, 1915: 250. Smith, 1916: 23, pl. II, fig. 5. Voss, 1962b: 248. Adam \& Rees, 1966: 81, pl. 20, figs 127, 128.
non Sepia burnupi Hoyle, 1904: non pl. I, figs 190, 191 ( $=$ S. incerta), non pl. I, fig. 192 ( $=$ S. confusa). Sepia exsignata Barnard, 1962: 250, fig. 3.
non Sepia burnupi: Barnard, 1962: 252, fig. 4 ( $=$ S. incerta).
? Sepia burnupi: Turton, 1932: 1 .
Type localities
Umkomaas, Natal (S. burnupi, shells only); off Umhlanga River, Natal, 40-48 m (S. exsignata).

## Distribution

Animals: Off Umhlanga River, Natal (Barnard ig62: 252). Depth 40-48 m. Shells: ? Port Alfred (Turton 1932: 1) to Tongaat beach, Natal (Smith 1916: 23).
Material
S.A.M. A2 147, Scottburgh, Natal; 3 shells

A6525, off Umhlanga River, Natal, $40-48 \mathrm{~m}$; I $\overline{\text { on }}$, 1 \& (holotype and allotype of S. exsignata)
N.M. 958 C, ? Natal coast; i shell
N.M.4073, off Umhlanga River, Natal, $40-48 \mathrm{~m}$; I ${ }^{1}$ (paratype of S. exsignata)

## Description

Mantle elongate oval, anterior margin somewhat produced dorsally, emarginate ventrally. Mantle pointed posteriorly. Fins beginning at anterior mantle edge, fairly wide. Fins of male drawn out into overlapping points posteriorly (Fig. 7b); rounded ventrally and meeting in midline in female (Fig. 7a).

Skin sparsely papillose dorsally over head and mantle, with a series of elongate tubercles along fin bases dorsally. Skin smooth ventrally, except for longitudinal dermal fold on each side of mantle, between midline and fin bases.

Colour dark dorsally, particularly between eyes and on that part of mantle covering shell; fins pale. Ventral surface pale, but with scattered chromatophores between dermal folds and fins. In addition, female has four oval silveryblue spots dorsally between eyes.

Arms of female subequal in length, except ventral pair, which a little longer. Arm tips somewhat attenuated; arms III and IV keeled. Suckers on all arms quadriserial, decreasing in size towards tips. Median series of suckers somewhat larger than lateral series.

In male, dorsal and ventral arms modified. Dorsal arms (Fig. 8e) bearing long cirriform processes laterally, and shorter ones on dorsal margin, supporting broad protective membranes. Basally, suckers normal and quadriserial, but rapidly decreasing in size distally, where cirriform processes extend further


Fig. 7. Sepia burnupi, A6525. a. Female (allotype of S. exsignata), b. male (holotype of $S$. exsignata). Left: dorsal and right: ventral views.
towards midline of arm, until at distal end they alternate medially with minute, biserial suckers. Tips of lateral cirri swollen. Distal tip of arm (about 2 mm ) bare, without suckers or cirri.

Right ventral arm of male with normal quadriserial suckers over most of its length, decreasing markedly in size distally, followed by short bare portion of the arm. Tip bearing short cirri, of which dorsal cirri better developed than ventral ones (Fig. 8d). There are a few small suckers between the cirri. Left ventral arm (Fig. 8c), the hectocotylus, basically like right ventral arm; but in second quarter of arm, ventral protective membrane very well developed, thrown into folds, and ventral series of suckers absent.

Lateral arms like those of female, with quadriserial suckers decreasing in size towards attenuated arm tips. In both sexes, sucker rings without teeth, nodular on upper surface.

Tentacular club of both sexes slightly recurved (Fig. 8b), bearing small suckers in oblique rows of eight, and five suckers greatly enlarged. Two suckers at tip larger than their neighbours, partly covered by reflexed tip of club. All tentacular sucker rings finely dentate. Protective membranes well developed, approximating very closely at base of club, without fusing. Natatory membrane only a little longer than club.

Shell (Pl. 38c, d, Fig. 8a) narrowly elongate, pointed anteriorly and posteriorly. Dorsally, shell calcified only along narrow median strip; laterally with broad chitinous margins. Calcified region finely granular, coloured pink, with fairly well defined median ridge. Posterior spine directed dorsally, not keeled. Deep median ventral groove present over whole length of shell. In region of striated zone, ventral surface highest at limbs of inner cone, shelving to lowest point at median groove, giving ventral surface $M$-shape in crosssection. Striated zone long, with acuminate anterior border. Striae $\wedge$-shaped. Inner cone has curved around lateral edges of striated zone, covering its sides with very thin layer (through which striae can be seen) and having, like S. insignis, its limbs lying on the phragmocone and not at its sides. Inner cone only very slightly raised posteriorly, not forming ledge. Outer cone fairly narrow, but forming wings posteriorly. Shell thickest just anterior to striated zone.

## Remarks

Hoyle (1904: 27) originally described S. burnupi on the basis of five shells. Smith (1916: 23), on re-examining these shells, and with additional material, separated them into three species, $S$. burnupi, $S$. incerta and $S$. confusa.

The first animals referred to $S$. burnupi were two male specimens described by Massy (1925: 215), but these in fact pertain to S. incerta. Barnard (1962:250) described two males and one female (S.A.M. A6525, N.M. 4073) as a new species, Sepia exsignata. Adam \& Rees (1966: 83), going only by Barnard's description, suggested that $S$. exsignata is probably synonymous with $S$. burnupi. The decalcified shell of $S$. exsignata was examined and, despite its rather poor


Fig. 8. Sepia burnupi, A6525. a. Ventral view of shell of female; b. left tentacular club of male; c. hectocotylized left ventral arm of male; d. tip of right ventral arm of male; e. modified dorsal arm of male. c. d. e. Modified after Barnard (1962); a. and b. original.
condition, was found to be identical with that of $S$. burnupi. The two species are here formally synonymized.

The shell of $S$. burnupi differs from those of $S$. confusa and $S$. incerta in that the inner cone is low posteriorly and does not form a deep pocket over the end of the phragmocone. It differs from $S$. joubini and $S$. adami in the $M$-shape of the striated zone, with the raised limbs of the inner cone lying near the peaks of the $M$ and by the angular shape of the striae.

The male of $S$. burnupi is distinguishable from those of the other species of the 'doratosepion' group by the modified tips of the ventral arms. The female is distinct from $S$. confusa, $S$. joubini and $S$. adami in that the arms are attenuated over less than half their length, and the suckers on the attenuated portion of the arms are quadriserial; from $S$. incerta it can be separated only by the shell.

## Sepia joubini Massy, 1927

(Figs 2, 9. Tables 3, 25, 26)
Sepia (Doratosepion) joubini Massy, 1927: 161, pl. XVIII, figs 1-10.
Sepia joubini: Voss, 1962 b: 248 . Adam \& Rees, $1966: 70$, pl. 43 , fig. 257.

## Type localities

Tugela River mouth, NW by N $\frac{3}{4} \mathrm{~N}, 25 \mathrm{~km}, 66-77 \mathrm{~m}$; Cape Natal, W by N, $10,5 \mathrm{~km}, 99 \mathrm{~m}$.

## Distribution

Animals: Off Tugela River mouth to Cape Natal (Massy 1927: 16i, and S.A.M.). Depth 66-I 34 m.

Material

$$
\begin{aligned}
& \text { A30142, } 35 \mathrm{~km} \text { S of Tugela River mouth, in } 6-134 \mathrm{~m} \text {; I dt, } 2 \text { 아 }
\end{aligned}
$$

$$
\begin{aligned}
& \text { A31393, off Cape Natal, S } 79^{\circ} \mathrm{E} \text { io, } 5 \mathrm{~km}, 99 \mathrm{~m} \text {; in ơ }
\end{aligned}
$$

## Description

Animals small. Mantle elongate, anterior margin produced dorsally, slightly emarginate ventrally. Posterior end pointed. Fins narrow, widening somewhat posteriorly, where rounded and separate.

Skin smooth in most specimens, but some (A30172) have a few dark papillae dorsally. Colour pale, with sparse chromatophores. Dark area dorsally over shell, and oval orange tubercles at fin bases. Pale silvery-blue areas present dorsally in front of and behind eyes, and sometimes also between eyes. These silvery regions not visible in all specimens. Ventral mantle surface pale, with somewhat darker region between midline and fin base. Also a few tubercles present on lateral margin of this area. Head translucent under and on either side of funnel, but beige at arm bases. Males with distinct red spot on lateral side of each dorsal arm, near base. Smaller red spot present on lateral side of
ventral arm, and sometimes small spots also present on other arms. These red spots absent in females.

Arms short (about one-third MLd or less). In males arms subequal in length, attenuated only at tips. Suckers on dorsal arms arranged in about three pairs basally, then quadriserial to arm tip. Suckers on lateral arms arranged in oblique quadriserial rows, except for few irregularly arranged suckers basally. Minute suckers on extreme tip of arm biserial for a few rows. Right ventral arm bearing large irregularly arranged suckers on basal half; suckers much smaller distally, arranged quadriserially. Left ventral arm hectocotylized, somewhat longer than its right counterpart. Suckers large basally, variously arranged. Distal hectocotylized region obscure and not always recognizable as such. Protective membranes expanded and small suckers arranged quadriserially but widely spaced over about eight rows. Distal tip of arm bearing minute quadriserial suckers.

In female, lateral arms longer than dorsal and ventral ones. Dorsal arms not markedly attenuated, bearing quadriserial suckers separated distally by deep median longitudinal groove. Protective membranes very well developed near arm tip and folded over inner surface of arm. Lateral arms attenuated over about half their length. Proximal part of arm bearing suckers arranged quadriserially, except a few irregular ones basally. Distally, suckers become minute and biserial, separated by longitudinal groove. Ventral arms not attenuated, bearing quadriserial suckers to tips.

Suckers globose in both sexes, having smooth rings. Dorsal and ventral


Fig. 9. Sepia joubini, A30172. Detail of posterior part of shell, ventral view. See also Figure 2.
arms keeled; interbrachial web highest between lateral arms, absent between fourth pair.

Tentacular club with many small subequal suckers distally in obliquely transverse rows of eight, and four median basal suckers greatly enlarged. All tentacular suckers with finely toothed rings. Tip of club reflexed, partly concealing two somewhat larger suckers. Natatory membrane extending beyond club.

Shells all decalcified, and damaged when removed. Shape of shell (Fig. 2) narrow elongate, pointed anteriorly. Indication of median longitudinal ridge dorsally; posterior spine present, not keeled, directed dorsally (Fig. 9). Median longitudinal groove present ventrally. Striated zone long (about two-thirds shell length), convex on either side of median groove, becoming flattened posteriorly. Striae convex and wavy, becoming more angular anteriorly. Inner cone with long narrow limbs, forming low ledge posteriorly. Outer cone forming posterior wings. Shell thickest in region of anterior end of striated zone. Not known if peculiar structure at base of posterior spine (Fig. 9) occurs in all specimens.

## Remarks

The specimens A3oi4i are topotypes, since they were from the same haul as Massy's syntypes (P.F.io715). These topotypes were not seen by Massy.

Barnard stated in his notes: 'the whip-like tips (of the arms, in the female)

Table 3. Sepia joubini females: increase in arm length and extent of attenuation of the dorsolateral arm with increase in size.

|  | MLd (mm) | Arm II <br> Length (mm) |  |  |
| :---: | :---: | :---: | :---: | :---: |
| A30141 | 26 | 8 | 3 | 37,5 |
| A30141 | 28 | 8 | 3 | 37,5 |
| A30141 | 29 | 8 | 2 | 25,0 |
| A30141 | 30 | II | 5 | 45,5 |
| A30141 | 32 | 11 | 4 | 36,4 |
| A30142 | 32 | 13 | 7 | 53,8 |
| A3014 1 | 34 | 10 | 4 | 40,0 |
| A30142 | 35 | 14 | 7 | 50,0 |
| A30141 | 35 | 13 | 7 | 53,8 |
| A30141 | 36 | 13 | 7 | 53,8 |
| A30141 | 36 | 15 | 10 | 66,7 |
| A30141 | 37 | 14 | 8 | 57, I |
| A30141 | 37 | 17 | 9 | 52,9 |
| A30172 | 38 | 20 | 12 | 60,0 |
| A30141 | 39 | 16 | 9 | 56,3 |
| A30141 | 39 | 17 | 9 | 52,9 |
| A30141 | 40 | 13 | 7 | 53,8 |
| A3014 ${ }^{1}$ | 43 | 17 | 9 | 52,9 |

begin to develop from 30 mm mantle length; below this length the whip is not clearly demarcated from the proximal portion and specimens may be difficult to separate from australis. The latter in fact occurred in the same haul. The whip becomes longer relatively to the proximal portion, and may become a little longer than the latter.'

Table 3 shows the extent of attenuation of the dorsolateral arm in females of 26 to 43 mm dorsal mantle length. In the smallest females, the tips of the lateral arms already show some degree of attenuation, but the suckers on the attenuated part are quadriserial. The degree of attenuation of the arm increases with growth of the animal, to a mantle length of about 36 mm , then levels off, although the increase in arm length with growth appears to be linear. In the larger specimens the suckers on the attenuated part of the arm are arranged biserially, although this probably corresponds to a crowding of the quadriserial condition.

The 'australis' specimens caught in the same haul as S. joubini (A30141) are in fact $S$. adami (A30149, A31394). In any case, young S. joubini without attenuated lateral arms may be distinguished from $S$. australis by the absence of a 'light organ' in the mantle cavity, and by the differences in the shells. S. australis shells are less narrow and elongate; they have a more marked dorsal rib and have a spine with a strong dorsal keel, whereas $S$. joubini shells have an unkeeled spine. The outer cone forms posterior wings in $S$. joubini but not in S. australis.

The male of S. joubini may be distinguished from the other 'doratosepion' species by the red spots on the arms, the unmodified dorsal arms and rounded fins. The female is characterized by the expanded protective membranes distally on the dorsal arms.

Sepia adami n. sp.
(Fig. 10. Table 27)

## Type locality

S $79^{\circ} \mathrm{E}$ of Cape Natal $10,5 \mathrm{~km}, 99 \mathrm{~m}$.

## Material

S.A.M. A31394, S $79^{\circ} \mathrm{E}$ of Cape Natal $10,5 \mathrm{~km}$, 99 m (P.F.10717, 14 December 1900); I ㅇ (holotype)

A30i49, same locality as holotype; 5 아 (paratypes)

## Description

Only females of this species known.
Mantle elongate oval, sharply pointed posteriorly (Fig. ioc, d). Anterior mantle margin produced dorsally to about anterior level of eyes in large specimens (a little less in smaller ones); ventrally entire or slightly emarginate. Head short and broad. Fins beginning a few mm behind mantle margin, narrow and rounded posteriorly, but fused over posterior tip of mantle.


Fig. io. Sepia adami female, A31394 (holotype). a. Ventral view of shell; b. dorsal view of anterior part of shell; c. ventral view of holotype; d. dorsal view of holotype; e. right tentacular club.

Skin sparsely papillose dorsally on head and mantle. Raised lunate tubercles along fin bases dorsally. Colour (of preserved specimens) pale cream, with somewhat darker region mid-dorsally over shell and over eyes. Scattered reddish-brown spots present on arms. Ventral surface pale. Sometimes also a line of pale lunate tubercles in this region (not in type specimen).

Arms subequal in length, joined by shallow web except between ventral arms. All arms keeled. Suckers globose, quadriserial on all arms from base to tip. The two median series of suckers much larger than the lateral ones; but on tips of arms all suckers the same size and markedly smaller than basal suckers. Sucker rings smooth.

Tentacular club (Fig. Ioe) short, slightly recurved, with about five median suckers enlarged and smaller distal suckers in five to six longitudinal series. Two suckers at tip of club slightly larger than their neighbours and partially concealed by reflexed tip of club. Rings of large club suckers finely toothed distally, those of small suckers toothed right round. Natatory membrane broad, extending a little beyond club proximally. Protective membranes separate at base of club.

Shells of all specimens very soft and difficult to remove undamaged. Only shell of holotype extracted in reasonably complete state (shell length 59 mm , including spine). Unfortunately, this shell is somewhat abnormal, having apparently been damaged and repaired, since outgrowth present mid-dorsally, corresponding with deep cleft in mid-ventral region. These do not occur in other shell examined by dissection.

Shell (Fig. ioa, b) narrow, elongate, broadest in anterior third, tapering sharply towards posterior end. Shell calcified only in median region dorsally; calcified part consisting of flat median ridge, broadest anteriorly, narrowing posteriorly, and two lateral ridges, narrow and indistinct anteriorly, broadening posteriorly to about twice width of median ridge. Posterior part of shell somewhat damaged dorsally, but ridges apparently fuse and become less distinct in this region. Calcified part of shell pale pink in colour, partially rugose. Rest of shell chitinous dorsally. Posterior spine directed dorsally, not keeled.

Striated zone occupies just over half ventral surface of shell. Anterior border of striated zone convex, with invagination at median groove. But this apparently due to repair outgrowth mentioned above, since anterior border $\Lambda$-shaped in other shell. Inner cone forming shallow pocket posteriorly over striated zone, but not markedly raised. Limbs of inner cone long and narrow, but it is not certain exactly how far they extend. Outer cone widens posteriorly, forming small wings. Shell thickest anterior to striated zone.

## Remarks

These specimens were found in the collection of the South African Museum, labelled Sepia australis, and indeed they do superficially resemble this species. S. adami differs from $S$. australis in the following ways:

1. The shell is markedly different in that it is more elongate, thicker, has posterior wings and a spine without keels.
2. There is no 'light organ' in the mantle cavity.
3. The arm tips are more slender than in S. australis.
4. The colour is much paler than that of $S$. australis and there is no indication of an orange band in the region of the fin bases.
With the six female specimens of $S$. adami described above were caught eleven males which almost certainly pertain to $S$. joubini, and have been given a separate catalogue number (A31393). The females of S. adami certainly differ from those of $S$. joubini. In $S$. adami the suckers on the dorsal arms are clearly quadriserial to the tips, whereas in $S$. joubini females the suckers on the dorsal arms appear to be biserial, although in fact they are in very oblique rows of quadriserial suckers. Neither are the tips of the dorsal arms provided with expanded protective membranes, as in S. joubini. In the latter species the lateral arms are very attenuated over at least half their length, and the suckers on the attenuated portion of these arms are biserial. In $S$. adami the lateral arms are not attenuated and bear quadriserial suckers to the tips of the arms.

## Distinctive characters

I. Tentacular club with unequal suckers
2. Shell with unkeeled spine and posterior wings
3. Arms subequal, with quadriserial suckers

This species apparently falls into the 'doratosepion' group, and is most closely related to the other species of this group, viz. S. confusa, S. incerta, S. burnupi and S. joubini. It differs from the females of all these species in that the arms are not attenuated distally.

Sepia adami has been named after Professor Dr. W. Adam of the Institut Royal des Sciences Naturelles de Belgique, in recognition of his extensive work on the Sepiidae.

## Sepia australis Quoy \& Gaimard, 1832

(Pl. 40a, b. Figs 1, 3a, 21. Tables 28-30)
Sepia australis Quoy \& Gaimard, 1832: 70, pl. 5, figs 3-7. Hoyle, 1912: 281, fig. 8. Bartsch, 1915: 250. Smith, 1916: 24, pl. II, fig. 9. Tomlin, 1923: 41. Robson, 1924a: 11. Turton, 1932: 2. Adam, 1941: 117, pl. IV, fig. 5; 1942: 10; 1959: 149, fig. 10. Voss, 1962b: 248, 252; 1967: 64. Adam \& Rees, 1966: 89, pl. 21, figs 138-142, pl. 45, fig. 270.
Sepia capensis d'Orbigny, 1845a: 283. Férussac \& d'Orbigny, 1835-1848: 278, pl. VII, figs 1-3, pl. XII, figs $7-11$, pl. XVII, figs 18, 19. Tryon, 1879: 198, pl. 94, figs 440-442. Hoyle, 1886: 23, 217. Gibbons, 1888: 202. Bartsch, 1915: 250. Thiele, 1920: 438, pl. LII, fig. 14, pl. LIII, figs $\mathrm{I}-5$.
Sepia sinope Gray, 1849: 106.
Rhombosepion australe Rochebrune, 1884: 85. Adam, 1944: 223.
Rhombosepion capense Rochebrune, 1884: 85. Robson, 1924b: 641, fig. 24. Adam, 1944: 222.
Sepia (Doratosepion) australis Massy, 1925: 214.
Rhombosepion australis: Massy, 1927: 156.
non Sepia australis: d'Orbigny, 1845a: 294. Férussac \& d'Orbigny, 1848: 285, pl. VII, fig. 4. Hoyle, 1886: 22, 220. (=S. novaehollandiae).
non Sepia capensis d'Orbigny, 1826: Gray, 1849: 1 10 ( $=$ S. cultrata).

## Type localities

Agulhas Bank ( $S$. australis), Cape of Good Hope ( $S$. capensis), ? China (S. sinope).

## Distribution

Animals: South African coast, from $31^{\circ} 43^{\prime} \mathrm{S}, 16^{\circ} \mathrm{I} 3^{\prime} \mathrm{E}$ (off the Olifants River) (Adam \& Rees 1966: 89) to $\mathrm{S} 73^{\circ} \mathrm{E}$ of Rame Head, 5 km (near Port St. Johns) (S.A.M.).
Red Sea (Rochebrune 1884: 85; Adam 1942: 10, 1944: 222, 1959: 149).
Depth 2-459 m.
Shells: South African coast, from Braak River (Namaqualand coast) (S.A.M.) to Port Alfred (Turton 1932: 2).
? China (Gray 1849: io6).

## Material

S.A.M. 2727 , Kalk Bay; 17 shells

A8g82, Hout Bay, $17-37 \mathrm{~m}$ (det. A. L. Massy); $2{ }^{\wedge}, 3$, 8 , in poor condition

A8986, south of Knysna Heads $16 \mathrm{~km}, 95 \mathrm{~m} ; 25$ ť, 34 ¢ +51 juveniles, all in poor condition
A8g88, $\mathrm{N} 48^{\circ} \mathrm{W}$ of Lions Head $80 \mathrm{~km}, 422 \mathrm{~m} ; 4$ specimens in poor condition
A8992, $\mathrm{S}_{42}{ }^{\circ} \mathrm{W}$ of Cape St. Blaize $17,5 \mathrm{~km}, 2-6 \mathrm{~m}$ (det. A. L. Massy); I + in poor condition
A8994, $\mathrm{N} 87^{\circ}$ E of Cape Point lighthouse $15 \mathrm{~km}, 59 \mathrm{~m}$ (det. A. L. Massy); II specimens in poor condition
A8997, mouth of Hout Bay, 73-92 m (det. A. L. Massy); i $甲$ in poor condition

A29734, west of Slangkop, ioo m (det. G. L. Voss) ; 2 ㅇ
A2g8or, False Bay, $59 \mathrm{~m} ;$ i +
A29802, False Bay, 59 m ; I ${ }^{\text {or }}$
A30148, S $57^{\circ} \mathrm{W}$ of Cape Point lighthouse in $\mathrm{km}, 147 \mathrm{~m}$; $10{ }^{\boldsymbol{t}}$, 9 ㅇ
$\mathrm{A}_{3} 0150, \mathrm{~S} 54^{\circ} \mathrm{W}$ of Gericke Point $21,5 \mathrm{~km}, 79 \mathrm{~m} ; 7$ t ${ }^{2}, 35$ 아
A3015I, S ${ }_{11}{ }^{\circ} \mathrm{W}$ of Great Fish Point lighthouse $5 \mathrm{~km}, 73 \mathrm{~m} ; 4 \mathrm{~d}^{\boldsymbol{0}}, 6$ ㅇ

A3O153, $33^{\circ}{ }^{\circ} 4^{\prime} \mathrm{S}, 27^{\circ} 54^{\prime} \mathrm{E}, 50 \mathrm{~m} ; 4$ ㅇ, 2 ? ?
A30154, $34^{\circ} 19^{\prime} \mathrm{S}, 18^{\circ}{ }^{32^{\prime} \mathrm{E}}$ (Buffels Bay), $59-64 \mathrm{~m} ; 6$ d', 12 아
$\mathrm{A}_{3015}{ }^{5}$, SE of Cape St. Blaize $9 \mathrm{~km}, 62 \mathrm{~m}$; I 우
A30156, Mossel Bay, $34^{\circ}$ 14'S, $22^{\circ}{ }^{\circ} 3^{\prime} \mathrm{E}, 66 \mathrm{~m}$ (det. A. L. Massy); 1 ơ A30157, S $37^{\circ} \mathrm{W}$ of Cape Hangklip 45,5 km, $183 \mathrm{~m} ; \mathrm{t}^{\boldsymbol{\delta}}, 4$ ㅇ
A30158, S $70^{\circ} \mathrm{W}$ of Cape Infanta $8 \mathrm{~km}, 64 \mathrm{~m}$ (det. A. L. Massy); 1 우 A30159, S of Knysna Heads $16 \mathrm{~km}, 95 \mathrm{~m}$; 3 dै, I ㅇ

A3oi6o, S $79^{\circ} \mathrm{W}$ of Table Mt. $64 \mathrm{~km}, 459 \mathrm{~m}$; I ${ }^{7}, 6$ 우
Ajoi6i, S $16^{\circ} \mathrm{W}$ of Cape Point light $16 \mathrm{~km}, 156 \mathrm{~m} ; 2{ }^{\text {th}}, 5$ ㅇ
A30162, S $34^{\circ} \mathrm{W}$ of Cape Infanta $30 \mathrm{~km}, 84 \mathrm{~m} ; 1$ ? ${ }^{\wedge}$, 1 ? ?, 2 juveniles, all in poor condition
A30163, S $73^{\circ}$ E of Rame Head $5 \mathrm{~km}, 79 \mathrm{~m} ; 2$ すै
A30164, S $62^{\circ}$ E of Bird Island light $13,5 \mathrm{~km}, 72 \mathrm{~m} ; 7$ th, 3 ㅇ
A30165, S $70^{\circ} \mathrm{W}$ of Cape Infanta $8 \mathrm{~km}, 64 \mathrm{~m}$ (det. A. L. Massy); I ${ }^{\text {d }}, 2$ 아
A30166, S $20^{\circ}$ E of Sebastian Bluff ${ }^{5} 5,5 \mathrm{~km}, 73 \mathrm{~m}$; i 우
A3or67, $\mathrm{S} 48^{\circ} \mathrm{W}$ of Cape St. Blaize $43 \mathrm{~km}, 8 \mathrm{r}-84 \mathrm{~m} ; 2$ す
A30168, off Buffels Bay (False Bay), $55 \mathrm{~m} ; 2$ ô, 4 ㅇ
A3or69, S of Cape St. Blaize $57,5 \mathrm{~km}, 99 \mathrm{~m}$ (det. A. L. Massy); 5 specimens in poor condition
A30170, $\mathrm{S} 37^{\circ} \mathrm{W}$ of Cape Infanta $7 \mathrm{~km}, 68 \mathrm{~m}$; I ${ }^{\mathbf{~}}$, I 우
A30171, S $14{ }^{\circ} \mathrm{W}$ of Gericke Point $8 \mathrm{~km}, 64 \mathrm{~m} ; 3$ ㅇ
A30173, S $6^{\circ} \mathrm{E}$ of Cape Infanta $21,5 \mathrm{~km}, 77 \mathrm{~m} ; 2{ }^{7}$, 1 ? \& in poor condition
A30175, S $42^{\circ}$ E of Cape St. Blaize 9,5 km, $66 \mathrm{~m} ; 2$ ta, 2 우
A3o190, Salt River power station (Cape Town); i +
A30263, N $87^{\circ}$ E of Cape Point lighthouse, 59 m (det. A. L. Massy); 5 specimens in poor condition
A30264, S $14{ }^{\circ} \mathrm{W}$ of Gericke Point $8 \mathrm{~km}, 64 \mathrm{~m}$ (det. A. L. Massy); I ${ }^{\wedge}$, I + , in poor condition
A $_{3} 0266, \mathrm{~S}_{59}{ }^{\circ} \mathrm{E}$ of Sebastian Bluff ${ }_{1} 3,5 \mathrm{~km}, 62 \mathrm{~m}$ (det. A. L. Massy); I ${ }^{\text {º }}$
A30267, S $34^{\circ} \mathrm{W}$ of Cape Infanta $30 \mathrm{~km}, 84 \mathrm{~m}$ (det. A. L. Massy);
I ${ }^{t}$, I + , in poor condition

A30504, Bloubergstrand; I shell
A30555, Bloubergstrand; 2 shells
A30564, Between Strandfontein and Muizenberg; I $\widehat{1}$
A30604, $32^{\circ} 24^{\prime} \mathrm{S}, 17^{\circ} 28^{\prime} \mathrm{E}, 193 \mathrm{~m}$; $13 \mathrm{~J}^{\top}, 53$ of, all in poor condition Ysterfontein beach; i shell
Mossel Bay (south of Slangkop on Cape Peninsula); 22 shells
Olifantsbosbaai (Cape Point Reserve); 7 shells
Millers Point, Simonstown; 6 shells
48 km N of Olifants River; 2 shells
Strandfontein (False Bay); I shell
Bloubergstrand; I shell
Between Strandfontein and Muizenberg; in shells
Namaqualand coast, between Sout and Braak Rivers; I shell
Simonstown; 2 shells
Arniston; 2 shells
Still Bay; 40 shells
Krom River mouth, Cape St. Francis; i shell (broken)

## Description

Mantle elongate oval, anterior margin dorsally produced, ventrally entire or slightly emarginate. Head short and broad, fins narrow and rounded but separate posteriorly.

Skin smooth. Colour dark purple dorsally on head and mantle, with slightly paler area medially over shell, and narrow reddish-brown to orange line at bases of fins. This line broadens posteriorly, but not meeting in the midline. Fins pale yellow, with small purple spots. Mantle as dark ventrally as dorsally, with purple chromatophores more concentrated near fin bases and posteriorly on mantle. Fins pale ventrally, without chromatophores; head pale ventrally except for some chromatophores on keeled edges of ventral arms.

Arms subequal in length, joined by shallow interbrachial web, deepest between arms II and III, absent between ventral pair. Protective membranes fairly well developed, especially at arm bases. Arms III moderately, arms IV well keeled.

Suckers globose, quadriserially arranged, with the two median series much larger than the lateral ones. Enlargement of median sucker series more marked in male than in female. Arm tips relatively blunt (definitely not attenuated), bearing minute suckers still arranged quadriserially. In male, distal suckers with long teeth on distal part of rings; rings on proximal suckers smooth on arms I and II, finely dentate proximally on arms III and IV. In female, rings finely dentate in distal suckers, smooth in proximal suckers. Sucker rings nodular on upper surface in both sexes, and adjacent skin of suckers wrinkled.

Left ventral arm of male hectocotylized over proximal two-thirds: five or six normal suckers at base, followed by six to seven rows of minute suckers. The two dorsal series of modified suckers arranged in normal positions, but the two ventral series merged to form a single series of 12 -14 minute suckers situated on edge of ventral protective membrane. Distal third of arm normal. Modified suckers described by Adam \& Rees (1966: 90) as consisting of ' 13 slightly alternate pairs of smaller ones'. This in fact corresponds to $6-7$ quadriserial rows of suckers, of which two ventral series have merged to form a single series, as mentioned above.

Tentacular club somewhat recurved, with small distal suckers in rows of about five. Four median suckers at base enlarged, and two suckers at tip a little larger than their neighbours, partially concealed by reflexed tip of club. Large suckers with smooth rings, smaller ones finely dentate. All rings broad and nodular. Natatory membrane very broad, a little longer than club. Protective membranes not meeting proximally.

Shell (Pl. 40a, b) broadly elongate, somewhat pointed anteriorly, more sharply so posteriorly. Posterior spine with dorsal keel, continuing along dorsal surface of shell for some distance. Shell with broad median rib dorsally, limited on either side by lateral groove. Striated zone long ventrally (about threequarters shell length); striae wavy. Anterior border of striated zone also wavy and may be sharply pointed or rounded medially. Ventral surface raised in
middle to form broad ridge on either side of distinct median longitudinal groove. Lateral to these ridges, shell concave. Inner cone not well developed posteriorly but raised to form deep pocket; limbs of inner cone narrow. Outer cone narrow, not forming posterior wings. Shell thickest and widest near anterior border of striated zone.

## Remarks

In the past there has been considerable confusion in the naming of this species. Sepia australis was first described from a specimen from the Cape of Good Hope by Quoy \& Gaimard in 1832. Later d'Orbigny claimed that he had described the same species as Sepia capensis in 1826, and used the name S. australis for an Australian species. Hoyle (1909: 266) could find no confirmation of the use of $S$. capensis prior to 1832, and upheld Quoy \& Gaimard's name S. australis, renaming $S$. australis d'Orbigny as Sepia novaehollandiae.

Regarding Sepia sinope, Smith (1916: 24) remarked: 'The name S. sinope was substituted by Gray for the $S$. australis, Q. \& G. (non d'Orb.), and he quoted a single imperfect shell in the British Museum collection which was labelled "China". No information concerning its acquirement is attached to the specimen, and consequently in all probability the locality cannot be relied upon. It certainly belongs to the present species' (i.e. S. australis).
S. australis is one of the commonest Sepiidae along the coast of the western Cape and its shells are found in abundance on the beaches. Turton (1932: 2) mentioned that this species is rare at Port Alfred, and indeed not many records are known east of Port Elizabeth.

The University of Cape Town Ecological Survey has a record of S. australis from Durban. Unfortunately the animals were discarded and their identity cannot be checked. Since $S$. australis has been confused with $S$. joubini and $S$. adami in the past, this record remains doubtful.

Adam \& Rees (1966: 89) report a shell from Grahamstown, Natal. But the only Grahamstown in South Africa known to the author is in the eastern Cape, and is 64 km from the sea.

Voss ( $1962 b: 252,1967: 64$ ) is mistaken in stating that $S$. australis is known only from southern Africa, since Rochebrune (1884:85) and Adam (1942: 10 , 1944: 222, 1959: 149) have reported this species from the Red Sea. The occurrence of $S$. australis in China (Gray 1849: 106) is doubtful (see above).

## Sepia tuberculata Lamarck, 1798

## (Pls 39c, d, 40c, d. Fig. ina. Tables 4, 31-33)

Sepia tuberculata Lamarck, 1798: 130; 1799: 9, pl. I, figs IA-B; 1822: 668; 1845: 372. Bosc, 1802: 45. Montfort, 1805: 274, pl. vii, figs $1-6$. Blainville, 1825 : 368, pl. I, fig. 2; 1827a: figs 2-6; 1827b: pl. ı, figs 2-6. Deshayes, 1832: 945. Férussac \& d'Orbigny, 1835-1848: 277, pl. VI, figs i-4. d'Orbigny, 1845a: 28ı; 1845[-47] b: pl. 3, fig. 11; 1845[-47]c: pl. 3, fig. ir. Hoyle, 1886: 24, 217. Gibbons, 1888: 202. Smith, 1903: 356. Adam, 194 I : in 3, pl. III, fig. 8. Voss, $1962 b$ : 248. Adam \& Rees, 1966 : 106 , pl. 26, figs 169 , 170 , pl. 27 , figs 171 , 172, pl. 28, figs 173, $^{174}$, pl. 44, figs $265,268$.
? Sepia mammilata Leach, MS. Férussac \& d'Orbigny, 1835-1848: 277 (S. mamillata), pl. IVbis (S. mammilata).

Spathidosepion tuberculatum Rochebrune, 1884: 93, pl. IV, fig. 3. Adam, 1944: 226.
Hemisepius (?) tuberculatus Smith, 1916: 25.
non Sepia tuberculata: Férussac \& d'Orbigny, 1835-1848: non pl. XVII, figs 13-15. Steenstrup, 1875: IV, pl. I, figs 20, 21, pl. II, fig. 6. Hoyle, 1910: 265 , figs 9,10 , pl. Va, figs $4^{-1} 4$. ( $=$ S. papillata) .

Type locality
?
Distribution
Animals: Melkbosstrand (S.A.M.) to Knysna (U.C.T.). Depth o-3 m.
Shells: Kommetjie, Cape Peninsula to Nature's Valley (S.A.M.) and Malagasy (Adam, 194I: 114).
Material
S.A.M. A2978i, Mossel Bay; i

A29867, Strandfontein; i $¢$
A30121, David's Kraal (near Cape Hangklip); $\mathbf{1}$ đ
A30123, Simonstown Harbour, 3 m ; I +
A30139, locality unknown; 19
A30180, Melkbosstrand; i 아
A30279, locality unknown; I $\widehat{0}$
A30485, Simonstown; i shell (of juvenile)
A30493, Simonstown; I shell
A30500, Gordon's Bay; 2 shells
A305II, Dalebrook; 2 S, I juvenile ?
A30559, Dalebrook; i +
A30600, Millers Point, Simonstown; 2 §
A31235, locality unknown; I ${ }^{\text {® }}$
Nature's Valley; 6 shells, discarded
Between Strandfontein and Muizenberg; 3 shells
Millers Point, Simonstown; 3 shells
Betty's Bay; I shell
Strandfontein (False Bay) ; I shell
Kommetjie; I shell
Die Kelders; I shell, discarded
? Cape Agulhas; 4 shells
? Milnerton; 2 shells
? Pearly Beach, S of Gansbaai; 4 shells
? Die Kelders; I shell
? II km NW of Cape Agulhas; 7 shells

## Description

Mantle short and broad, anterior mantle margin convex dorsally (not produced), ventrally entire; mantle rounded posteriorly. Fins wide, beginning a few mm behind anterior mantle margin, rounded but separate posteriorly.

Skin densely tuberculate on dorsal surface of head, arms, mantle and fin bases. Ventral surface smooth except for large oval wrinkled area on each side of mantle and on ventral surfaces of fourth arms. Colour dark purple dorsally, pale buff ventrally.

Arms subequal in length, about half dorsal mantle length. Tips of all arms except ventral pair attenuated. Interbrachial membrane present, except between ventral arms. Arms III and IV keeled.

Suckers not globose; with finely dentate horny rings. Protective membranes well developed. Suckers quadriserial on all arms to tips.

Left ventral arm of male hectocotylized basally. Dorsal series of suckers normal, but the two ventral series are widely separated from these by a broad naked area with transverse ridges. Distal half of arm normal.

Tentacular club long, slightly recurved, bearing small distal suckers arranged in very oblique rows of eight. Four or five median suckers enlarged. Suckers at tip of club no larger than their neighbours. Horny rings of large suckers smooth, those of small suckers dentate. Protective membranes meeting proximally. Natatory membrane extending a little beyond club.

Shell (Pl. 4oc, d) oval but somewhat angular anteriorly. Dorsal surface finely granular. Median ridge faint or absent. No posterior spine or knob, but only a small hump present in this position. No distinct median groove ventrally. Striated zone long, striae wavy with overall convex shape. Inner cone completely reflexed and fused to outer cone, but not well developed posteriorly;


Fig. in. Comparison of tentacular clubs of a. Sepia tuberculata (A30139) and b. S. papillata (A30124).
limbs of inner cone long and narrow. Outer cone broad laterally, narrowing sharply posteriorly so that inner cone almost reaches posterior margin of shell. Shell generally very thin, ventral surface flat or slightly concave.

Several shells (indicated with query in list of material) (Pl. 39c, d), found on beaches, with markedly longer striated zone, and sometimes also with somewhat wider outer cone.

## Remarks

This species was synonymized with $S$. papillata by Férussac \& d’Orbigny (1835-1848: 277), but the two were rightly separated by Rochebrune (1884: 93-95). Hoyle (1910: 267), following Férussac \& d'Orbigny's synonymy, described a specimen of $S$. papillata under the name $S$. tuberculata. In fact the differences between the two species are quite clear (Table 4), the most marked being the relative sizes of the large tentacular suckers. The two specimens mentioned by Steenstrup (1875: IV) almost certainly also pertain to S. papillata (cf.).

Adam (194I: 114) has mentioned a specimen of $S$. tuberculata from Port Dorey, New Guinea. The correctness of this locality is doubtful (Adam, personal communication), and has not been included in the distribution lists.

The wrinkled areas on the ventral surfaces on the mantle and fourth arms are apparently used to hold on to a hard substratum such as rocks. S. tuberculata kept in aquaria have been observed to use these wrinkled areas to cling to the glass walls.

Table 4. Comparison of Sepia tuberculata with S. papillata.

|  | S. tuberculata | S. papillata |
| :--- | :--- | :--- |
| Dorsal mantle margin | Convex | Attenuated |
| Tips of arms I-III | Diameter less than width <br> of tentacular club | Not attenuated <br> Tentacular <br> club <br> to width of tentacular clubLarge <br> suckers |
|  | Protective <br> membranes | Joined proximally |
| Dorsal <br> surface | No median ridge; no <br> posterior knob | Separate proximally <br> Median ridge and posterior <br> knob present |
| Ventral <br> surface | Flat or concave; no dis- <br> tinct median groove | Convex; median groove <br> present |
| Inner <br> cone | Not well developed pos- <br> teriorly; limbs narrow | Well developed posteriorly; <br> limbs usually broad (but see <br> description) |
| Thickness | Mean 6,6\% shell length | Mean 10,1\% shell length |

The shells with an exceptionally long striated zone ( $70,0-88,5 \%$ shell length) show no other differences from those of $S$. tuberculata (striated zone $54,1-73,9 \%$ shell length) and may constitute an extension of the known range of striated zone length for this species, or may be distinct. No decision can be made on their status until the soft parts can be studied.

Sepia papillata Quoy \& Gaimard, 1832
(Pl. 4ra-d. Figs irb, 12. Tables 4, 34-37)
Sepia papillata Quoy \& Gaimard, 1832 : 61, pl. I, figs 6-14. Férussac \& d'Orbigny, 1835-1848:
 1928: 92. Turton, 1932: 1. Adam, 1939a: 55, pl. III, fig. 6; 1941: 112. Voss, 1962b: 248, 25 1. Adam \& Rees, 1966: 108 , pl. 28, figs ${ }^{175-1} 78$.
Spathidosepion papillatum Rochebrune, 1884: 94. Adam, 1944: 226.
Sepia tuberculata (non Lamarck) Férussac \& d’Orbigny, 1835-1848: pl. XVII, figs 13-15. Steenstrup, 1875: IV, pl. I, figs 20, 21, pl. II, fig. 6. Gibbons, 1888: 202. Hoyle, 1910: 265 , figs 9 , ro, pl. Va, figs $4^{-14}$.
non Sepia papillata: Smith, 1916: 22, pl. II, figs $\mathbf{1}, 2$ ( $=$ S. simoniana) .

## Type locality

Cape of Good Hope.

## Distribution

Animals: Lüderitzbucht (Angra Pequena) (Hoyle 1910: 265) to Natal coast, off Tugela River and Umvoti River (Massy 1928: 92). Depth 26-127 m.
Shells: Orange River mouth (S.A.M.) to ? Tongaat, Natal (Tomlin 1923: 40).
Material
S.A.M. A30118, Woodstock power station, Cape Town; $2{ }^{\circ}$ (one shell missing) A30119, S 62-79 ${ }^{\circ}$ E of Bird Island lighthouse $15 \mathrm{~km}, 73-48 \mathrm{~m}$; 1 ㅇ (shell missing)
A30120, Woodstock power station, Cape Town; i $\begin{gathered}\text { त }\end{gathered}$
A30124, locality unknown; I + (shell missing)
A3or 36, S $87^{\circ}$ E of Cape St. Blaize $9 \mathrm{~km}, 5 \mathrm{Im}$; i +
A30137, Hout Bay, 37-73 m; I ơ
A30138, N $3^{\circ}$ E of Green Point lighthouse $4 \mathrm{~km}, 40 \mathrm{~m}$; I ${ }^{1}$

A30476, Bloubergstrand; I shell
A30482, $\mathrm{S} 62-79^{\circ}$ E of Bird Island lighthouse $15 \mathrm{~km}, 73-48 \mathrm{~m}$; I shell A30497, locality unknown; I shell
A30507, Elandsbaai beach, after red tide; 2 § 2 , 아
A30509, Elandsbaai beach, after red tide; I $\widehat{0}$
A30553, Orange River mouth; 2 shells
A30554, Bloubergstrand; i shell
A31250, Castle Rock, False Bay; I ${ }^{\text {ot }}$
Ysterfontein; I shell

Nature's Valley; i shell, discarded
Between Strandfontein and Muizenberg; 14 shells
Mossel Bay (S of Slangkop on Cape Peninsula); i shell
Olifantsbosbaai (Cape Point Reserve); I shell
Millers Point, Simonstown; 2 shells
48 km N of Olifants River; 4 shells
Strandfontein (False Bay); 6 shells
Betty's Bay; i shell
Umngazana River mouth, west Pondoland; 2 shells
Muizenberg; 2 shells
Kommetjie; 4 shells
Milnerton beach, Cape Town; 2 shells
Namaqualand coast, between Sout River and Braak River; i shell
Still Bay; 8 shells
Simonstown; 2 shells
Arniston; 2 shells
Die Kelders; 3 shells, discarded
Krom River mouth, Cape St. Francis; 32 shells

## Description

Mantle broadly oval, anterior margin somewhat produced dorsally, slightly emarginate ventrally. Fins broad, beginning a few mm behind mantle margin; posteriorly rounded and separate.

Skin tuberculate dorsally and laterally on head, arms and mantle, but smooth ventrally except for two large wrinkled patches on mantle, and smaller patches along ventral surfaces of fourth arms, as in S. tuberculata. Colour dark reddish-brown to purple dorsally on tuberculate surfaces, pale ventrally with scattered chromatophores, which are somewhat more dense on fins.

Arms subequal in length, about $50 \%$ MLd; arms I to III attenuated over about distal quarter. All arms keeled, and joined by fairly deep interbrachial web, which may attain half arm length.

Suckers not globose; with finely toothed chitinous rings. In the female, suckers quadriserially arranged on all arms. Suckers on attenuated arm tips minute, but still quadriserial. In the males, suckers quadriserial over most of arm, but attenuated tip broader and flatter than in female, and minute suckers arranged in eight series. Ventral arms not much attenuated at tips, but suckers also show multiplicity, although less extensively than on other arms.

Left ventral arm of male hectocotylized proximally. Basally about 13 suckers arranged normally, followed by modified region, consisting of eight quadriserial rows of suckers. In this region the two dorsal series are separated from the two ventral series by a naked region with transverse ridges. Modified suckers described by Massy ( $1925: 212$ ) as ' 15 rows of diminutive suckers placed 2 in a row, in zigzag order, with shallow grooves between them'. This in fact corresponds with a quadriserial condition in which two series come close


Fig. 12. Sepia papillata male, A30ı37. Ventral view to show wrinkled areas on mantle and ventral arms.
together (on either side of median grooved section of arm) until apparently forming single longitudinal zigzag series on each side. Suckers normally quadriserial distally, but at arm tip (which is slightly attenuated), minute suckers arranged in oblique rows of eight.

Tentacular club long, bearing small distal suckers in oblique rows of eight. Four suckers enlarged medially; of these, the middle two extremely large, with diameters equal to width of sucker-bearing surface of club. Chitinous rings of large suckers smooth, those of small suckers dentate. Protective and natatory membranes very well developed; protective membranes not meeting basally, natatory membrane extending a little beyond club.

Barnard (unpublished notes) discovered that there are two forms of shells
belonging to $S$. papillata. Both shell forms (Pl. 4oa-d) broadly oval, tapering somewhat anteriorly. Dorsal surface rugose, with faint, broad median ridge, broadening anteriorly. Usually no spine posteriorly, but a broad rounded knob present. In a few shells, however, this knob continues as small spine lying close to shell surface and thus directed ventrally. Spine not exceeding posterior margin of shell. Striated zone long ventrally, with narrow smooth area on either side of it anteriorly. Striae wavy, with overall $\wedge$-shape. Median longitudinal groove distinct. Shape of inner cone different in the two forms, but completely reflexed and fused to outer cone in both. Outer cone broad laterally. Inner cone frequently (but more often in form A than B) drawn out posteriorly, almost reaching edge of shell, as in $S$. simoniana.

Shell form A (Pl. 40a, b) with broad inner cone; difficult to distinguish from $S$. simoniana in extreme cases. Phragmocone of this form generally thicker at anterior region of striated zone, on either side of midline. Shell form B (Pl. 4oc, d) differs in that inner cone narrow and thicker, with limbs forming narrow raised ridges, as in $S$. tuberculata. Phragmocone generally thinner than in form A, but always convex to some extent.

## Remarks

As mentioned above, the animals of this species have frequently been confused with those of $S$. tuberculata, but they are in fact distinct (see Table 4).

Of the two shell forms, form A (having a broad inner cone) markedly resembles the shell of $S$. simoniana, but the animals differ primarily in that S. simoniana has a tentacular club with numerous subequal suckers, and has a smooth skin, whereas $S$. papillata has a tentacular club with very unequal suckers, and has a tuberculate skin. Shell form B of S. papillata closely resembles the shell of $S$. tuberculata, but the differences between the two species are as listed in Table 4. A comparison of the relative dimensions of shell forms A and B of $S$. papillata revealed no statistically significant differences. The present collection includes six male and four female animals having shells of form A, and only two males with shells of form B. No differences could be found between the animals having either shell form, and all undoubtedly belonged to $S$. papillata. The significance of these shell forms is at present unknown, but they may reflect a relationship with $S$. simoniana on the one hand and $S$. tuberculata on the other.

Steenstrup (1875: IV) mentioned two specimens (male and female) of Sepia which he thought to be S. tuberculata. He illustrated the buccal view of the female (pl. II, fig. 6), suckers (pl. I, fig. 2 I) and an arm with minute suckers at the tip arranged in eight series (pl. I, fig. 20). As $S$. tuberculata and $S$. papillata have so often been confused, we may assume that Steenstrup's specimens could belong to either species, since neither tentacular club nor shell were illustrated. But of these two species only the male of $S$. papillata has the minute suckers at the tips arranged in eight series. Hence the male specimen was almost certainly $S$. papillata, and presumably the female belonged to the same species.

As mentioned above, Hoyle's (1910: 265) three female specimens, described as $S$. tuberculata, were in fact $S$. papillata. This can clearly be seen from his figures (pl. IVa, figs 4-6). In the description, Hoyle said that the arms are about as long as the mantle, but according to his measurements, they are only half as long, as also in the present specimens.

The description of $S$. papillata shells given by Smith (1916:22) could cover both $S$. papillata and $S$. simoniana, and indeed he stated that 'It should be noted also that the limbs of the inner cones are rather variable. Sometimes, as in the Astrolabe figure, they do not expand much posteriorly. On the contrary, in some specimens they spread considerably, and become rather pointed posteriorly.' The shell in Smith's figure (pl. II, fig. i) is that of S. simoniana. Unfortunately, he does not give a list or the number of specimens examined, but mentions only that there were shells from Port Elizabeth (Ponsonby, Spencer) and Tongaat beach, Natal (Burnup). One of the specimens presented to the British Museum by Spencer was 134 mm long and 60 mm wide.

Adam (194I: in3) lists one of Smith's specimens (length 134 mm , width $45 \%$ * length) from Port Elizabeth in a table of measurements of $S$. papillata shells, but does not mention a shell from Port Elizabeth in his list of material examined.

Adam \& Rees (1966) do not include Smith (igi6) in the synonymy for S. papillata, but included in the list of material for $S$. simoniana are three shells from Port Elizabeth (Ponsonby) and three shells (of which one is doubtful) from Port Elizabeth (Spencer). These are presumably the same shells as originally examined by Smith. Of the specimens donated to the British Museum by Spencer, one shell is recorded by Adam \& Rees as being 137 mm long, with a width of $44 \%$ of the length. This must be the shell measured by Smith. Presumably Adam reconsidered his opinion of 1941 of Smith's specimens, and now considers them all to pertain to $S$. simoniana. Adam \& Rees (1966: 109) in fact state that the shell figured by Smith (1916, pl. II, figs i, 2) as S. papillata belongs to $S$. simoniana.

Tomlin's (1923: 40) localities listed for S. papillata were obviously quoted from Smith (1916), and thus refer to $S$. simoniana (with the exception of Quoy \& Gaimard's locality of the type of $S$. papillata).

Massy (1925:211) is the only author to have previously described a male of S. papillata, but she does not mention the presence of multiplicity of the suckers on the arm tips. Since this condition occurs in all the present male specimens, Massy probably overlooked it in hers.

Voss (1962b: 25I) was mistaken in saying that $S$. papillata 'was not reported from South Africa by either Massy or Robson and may be an uncommon species'. S. papillata was described by Massy in 1925, and mentioned again in 1928. In fact the species appears to be very common, at least around the western Cape, since the shells are commonly washed up on the beaches.

[^8]Sepia simoniana Thiele, 1920
(Pl. 42a, b. Tables 38-40)
Sepia simoniana Thiele, 1920: 436, pl. LII, figs 5-13. Odhner, 1923: 7. Tomlin, 1926: 285.
Voss, 1962b: 248, 250. Adam \& Rees, 1966: 109, pl. 29, figs 179-182, pl. 42, fig. 254.
Sepia natalensis Massy, 1925: 212, pl. XI, figs 1-11, pl. XIV, fig. 37.
Sepia tuberculata (non Lamarck) Gray, 1849: 101, 102.
Sepia papillata (non Quoy \& Gaimard) Smith, 1916: 22, pl. II, figs 1, 2. Tomlin, 1923: 40 (partim).
Type localities
Simons Bay (S. simoniana); 25,5 km NE of Bird Island (S. natalensis).

## Distribution

Animals: Simons Bay (Thiele 1920: 436) to off Tugela River (S.A.M.). Depth 14-134 m.
Shells: $\quad 48 \mathrm{~km} \mathrm{~N}$ of Olifants River mouth (S.A.M.) to Tongaat beach, Natal (Smith 1916: 22).
Material
S.A.M. A30127, locality unknown; 1 §

A30132, $34^{\circ}$ 12'S, $18^{\circ} 3^{\prime} \mathrm{E}, 40 \mathrm{~m}$; 1 ㅇ
A3or33, Simons Bay; i $q$
A30134, locality unknown; i 9
A30135, 35 km S of Tugela River mouth, in6-134 m; i 9
A3or 79, Millers Point, Simonstown; I juvenile
A30498, Table Bay; i shell
A30499, Still Bay; I shell
A31239, Sunny Cove, False Bay, 14 m; 2 juveniles
A31251, Castle Rock, False Bay; 2 〇
Between Strandfontein and Muizenberg; 4 shells
Simonstown; 2 shells
48 km N of Olifants River mouth; i shell
Betty's Bay; 2 shells
Strandfontein (False Bay); 2 shells
Umngazana River mouth, west Pondoland; 4 shells
Still Bay; I5 shells
Arniston; 4 shells
Die Kelders; i shell, discarded
Krom River mouth, Cape St. Francis; 46 shells

## Description

Mantle broadly oval, anterior mantle margin somewhat produced dorsally, entire ventrally. Fins fairly narrow, beginning a few mm behind anterior mantle margin, rounded and separate posteriorly.

Skin very finely papillose dorsally on head and mantle. Skin somewhat wrinkled ventrally on mantle in some specimens, but this is possibly due to preservation. One juvenile (A3OI79), however, has distinct wrinkled oval
patches ventrally on mantle, as in S. papillata and S. tuberculata. Wrinkled areas usually also present on ventral surfaces of fourth arms. Colour pinkish-brown dorsally, pale ventrally, with sparse chromatophores, except for two slightly darker regions on either side of midline, beginning at anterior mantle margin and fading towards posterior.

Arms subequal in female, about half dorsal mantle length, with third arms a little longer than others. In males, arms unequal in length, of formula 3.2.1.4. All arms except ventral pair somewhat attenuated over about distal quarter. Depth of interbrachial web approximately half arm length, except between ventral pair, where web absent. Third and fourth arms keeled.

Suckers on all arms quadriserial to tips; suckers largest about one-third from arm base, then decrease in size to tips, where they suddenly become minute. On fourth arms, although tips not attenuated, distal suckers also minute. Rings of large suckers smooth, those of smaller ones very finely dentate. Suckers of male more globose than those of female.

Left ventral arm of male hectocotylized proximally. About five to nine normal suckers at base of arm, followed by modified region, where two dorsal series of suckers smaller than usual and separated from the two ventral series by naked region with transverse ridges. Suckers in the two ventral series minute. Distal half of arm normal. According to Thiele's (1920: 438) description of the hectocotylus, second longitudinal series of suckers almost lacking in modified region, although in one specimen this absence was less extensive. But in the present specimens, second series of suckers complete, though smaller, and it seems probable that in Thiele's specimens some suckers were lost after capture.

Tentacular club very long, occupying about half length of tentacle. Club bears very many minute subequal suckers, and four (not two, as stated by Voss 1962b: 250) somewhat larger suckers, partly concealed by reflexed tip of club. Protective membranes very well developed, also bearing suckers, except at outer edge. Rings of small suckers with well defined teeth; those of four larger suckers at tip of club smooth. Protective membranes joined basally. Natatory membrane well developed, somewhat shorter than club.

Shell (Pl. 42 a, b) broadly oval, tapering a little anteriorly. Dorsal surface finely granular. At most a faint indication of median longitudinal ridge dorsally, broadening anteriorly. Longitudinal grooves on either side of median ridge ill defined. Spine absent posteriorly, but broad knob present, sometimes tapering into small point, but not attaining posterior margin of shell. Striated zone long, about two-thirds shell length. A narrow smooth area present on either side of striated zone. Striae wavy, with overall $\wedge$-shape. Deep median longitudinal groove present ventrally, with convex striated zone on either side. Striated zone most strongly convex near its anterior end. Inner cone well developed, reflexed and fused to outer cone. Limbs of inner cone very broad, usually narrowing suddenly anteriorly, giving characteristic shape. Inner cone drawn out posteriorly, almost reaching posterior margin of shell. In some shells,
however, anterior narrowing of limbs not as sharp, and these shells are difficult to separate from those of $S$. papillata form A.

## Remarks

As has been pointed out above, shell form A of S. papillata is very similar to that of $S$. simoniana, but the animals differ markedly.

A shell from the Cape of Good Hope, identified by Gray (1849: 102) as S. tuberculata, belongs in fact to $S$. simoniana, according to Adam \& Rees (ı96: 109), who re-examined the shell.

As mentioned above, the specimens described by Smith (1916: 22) as S. papillata are probably $S$. simoniana.
S. simoniana Thiele and $S$. natalensis Massy are indubitably synonymous, as has already been remarked by Voss (1962b: 25I) and by Adam \& Rees (ı966: iog).

Sepia angulata n. sp.
(Pls 44d, 45a-d. Table 4i)

## Type locality

Bloubergstrand (shells only).

## Distribution

Shells: Bloubergstrand to Still Bay (S.A.M.).
Material
S.A.M. A3i3i7, Bloubergstrand (coll. Roeleveld, 9 November 1969); i shell (holotype)
A313i8, Milnerton beach (coll. Kensley, 29 January i969); 3 shells
A3i319, Still Bay (coll. Du Preez, April 1969); 2 shells
A31320, Still Bay (coll. Du Preez, October 1969); 3 shells
A3i395, Bloubergstrand (coll. Roeleveld, 9 November 1969); 3 shells

## Description

Only shells of this species known.
Shell (Pls 44d, $45^{\mathrm{a}-\mathrm{d}) ~ b r o a d l y ~ o v a l, ~ r o u n d e d ~ a n t e r i o r l y ~ a n d ~ p o s t e r i o r l y . ~}$ Dorsally, part of dorsal shield covering phragmocone clearly distinguishable from outer cone: the former brown in colour; the outer cone white. Dorsal surface finely granular, becoming somewhat more coarsely so posteriorly; entire dorsal surface with iridescent sheen. Faint indication of dorsal longitudinal rib, widening anteriorly. No posterior spine or knob.

Ventrally, striated zone fairly short, flat from side to side,* except at extreme lateral edges, which are slightly rounded. Striated zone rises rapidly from posterior to anterior. Then smooth zone, also flat, descends sharply towards anterior, forming distinct obtuse angle between striated and smooth zones (Pl. 44d). A number of shallow grooves radiate over striated zone from

[^9]posterior end in holotype; median longitudinal groove no more distinct than others. Striae wavy over the grooves, with overall convex shape. In some shells, however, radiating grooves less distinct than median groove, and in these shells, striae more regularly convex.

Inner cone completely reflexed, fused to outer cone; posteriorly well developed, forming transverse ridge. Limbs of inner cone broad, as in S. papillata form A. Outer cone very broad and deep laterally and posteriorly, giving shell its characteristic tubby shape. Slight fold present in outer cone on each side, near posterior end, from inner cone to margin.

## Remarks

This species somewhat resembles $S$. papillata and $S$. simoniana, but the shell of $S$. angulata is somewhat shorter and broader, differing also in that the median ventral groove is indistinct, and the striated zone and last loculus are remarkably flat and form an angle at the point of meeting, about halfway along the shell. $S$. angulata differs from $S$. tuberculata in that it is relatively broader, the phragmocone is much thicker, particularly at the anterior end of the striated zone, and the outer cone is much broader posteriorly. The inner cone is also more strongly developed in $S$. angulata.
S. angulata has been so named after the angle formed between the striated and smooth zones in lateral view (angulatus, L.-angular).

## Distinctive characters

I. Shell very broad ( $50-60 \%$ length $)$
2. Obtuse angle between striated and smooth zones, in side view
3. Both striated zone and smooth zone very flat from side to side

> Sepia hieronis (Robson, 1924)
> (Pl. 43a-d. Fig. 13. Tables 5, $4^{2-44)}$

Sepia acuminata Smith, 1916: 21, pl. II, fig. 4 (partim).
Sepia sp. A Robson, 1924a: 13.
Rhombosepion hieronis Robson, 1924b: 645, pl. II, figs 9, II. Massy, 1927: 158.
Sepia hieronis Voss, 1962b: 248, 251 ; 1967: 64. Adam \& Rees, 1966: 112, pl. 30, figs 187, 188, pl. 43 , fig. 262.
Type localitues
S.S. Pickle, Sta. 2: $33^{\circ} 03^{\prime} \mathrm{S}, 17^{\circ} 4^{\prime}$ E, 215 m

Sta. 7: $32^{\circ} 32^{\prime} \mathrm{S}, 17^{\circ} 42^{\prime} \mathrm{E}, 206 \mathrm{~m}$
Sta. 8: $32^{\circ} 33^{\prime} \mathrm{S}, 17^{\circ} 29^{\prime} \mathrm{E}$, 23 Im
Sta. 33 : $32^{\circ} 54^{\prime} \mathrm{S}, 17^{\circ} 25^{\prime} \mathrm{E}, 275 \mathrm{~m}$

## Distribution

Animals: West coast: $30^{\circ}$ 13'S, $15^{\circ}$ 18 $\mathbf{E}^{*}$ (Adam \& Rees 1966: 112, g) to west of Slangkop (Voss 1967: 64). Depth 43-457 m. East coast: Monte Belo, Moçambique (S.A.M.). Depth 43I-459 m.
Shells: Bloubergstrand (S.A.M.) to Tongaat beach, Natal (Smith 1916:21).

[^10]
## Material

S.A.M. Agooo, $80 \mathrm{~km} \mathrm{~N} 42^{\circ} \mathrm{W}$ of Lions Head, 422 m (det. A. L. Massy); I $\widehat{\delta}$, I juvenile, in poor condition
A29728, west of Slangkop, 250 m (det. G. L. Voss); i ${ }^{\text {® }}$
A30145, locality unknown; i
A30146, S $76^{\circ} \mathrm{W}$ of Lions Head $45 \mathrm{~km}, 257 \mathrm{~m}$; 1 §
A30260, N $48^{\circ} \mathrm{W}$ of Lions Head 8 km , 422 m (det. A. L. Massy); 2 specimens in poor condition
A30268, N $5 \mathrm{I}^{\circ} \mathrm{W}$ of Lions Head 75 km , 32 I m (det. A. L. Massy); $1{ }^{\wedge}$ in poor condition
A30563, $34^{\circ} \mathrm{o4}^{\prime} \mathrm{S}, 17^{\circ} 45^{\prime} \mathrm{E}, 275 \mathrm{~m}$; 1 o
A31243, $32^{\circ} \mathrm{I} 5^{\prime} \mathrm{S}, 17^{\circ}$ o8'E, 283 m ; 1 ơ
A3ı405, Monte Belo, Moçambique, $25^{\circ} 35^{\prime} \mathrm{S}$, $33^{\circ} 30^{\prime} \mathrm{E}, 43^{\mathrm{I}-455 \mathrm{~m} \text {; i đ }}$ A3ı406, Monte Belo, Moçambique, $25^{\circ} 35^{\prime} \mathrm{S}$, $33^{\circ} 30^{\prime} \mathrm{E}, 440-459 \mathrm{~m}$; i đ A31407, Monte Belo, Moçambique, $25^{\circ} 35^{\prime} \mathrm{S}, 33^{\circ} 30^{\prime} \mathrm{E}, 459 \mathrm{~m}$; 1 o
Mossel Bay (S of Slangkop on Cape Peninsula) ; 19 shells, broken
Olifantsbosbaai (Cape Point Reserve); i shell, broken
Millers Point, Simonstown; i shell, broken
Bloubergstrand; 2 shells, slightly damaged

## Description

The specimens previously described by Massy (1927: 158) now very shrivelled and cannot be measured or properly examined. Remaining specimens (seven males and one small female) in good condition.

Mantle ovate, anterior mantle margin produced to fairly sharp point dorsally between eyes in males, less strongly produced in small female (about half as much as in males). Ventrally, anterior mantle margin entire in males, emarginate in female. Fins narrow, beginning a few mm behind mantle margin, rounded and separate posteriorly.

Colour generally mottled reddish-brown dorsally (but in two males and in female chromatophores contracted to small spots on pale background). Mantle ventrally pale in middle, somewhat darker laterally towards fin bases, where chromatophores more dense. Skin sparsely papillose dorsally on head and mantle in female; smooth in males.

Arms generally shortest dorsally, longest ventrally, having formula 4.3.2.1, but varying somewhat in different specimens (female measured by Adam \& Rees 1966: II3, had subequal arms). Interbrachial web present, except between ventral arm pair, and is deepest between arms II and III.

In males, dorsal arms attenuated over about distal third, bearing biserial suckers gradually diminishing in size towards tips, where suckers minute. Sucker arrangement on lateral arms very characteristic. On dorsolateral arms, suckers of basal two-thirds biserial, sometimes becoming somewhat irregularly arranged or even quadriserial for a few rows on middle third of arm, then three to five pairs of greatly enlarged suckers. Of these, middle ones have diameter


Fig. 13. Sepia hieronis male, A30563. a. Ventral view; b. dorsal view; c. hectocotylus; d. sucker from the base of a lateral arm, diameter about I mm ; e. one of the suckers distal to the enlarged suckers on arms II to IV, diameter about I mm; f. one of the enlarged suckers on the distal part of the hectocotylus, diameter about 1 mm ; g . enlarged sucker from one of the lateral arms, diameter about 2 mm ; h . oral view of sucker from the tentacular club, diameter about $0,2 \mathrm{~mm}$.
about twice that of basal suckers. The more distal enlarged suckers of peculiar shape (Fig. ine), being elongated and attached by long stalks. Distal suckers minute, arranged quadriserially. Arrangement of suckers on ventrolateral arms generally the same as that on dorsolateral arms, but a number of quadriserial rows of suckers present on middle third of arm. Enlarged suckers not as large (relative to those at arm base) as on dorsolateral arms. On right ventral arm, suckers on basal third arranged as on lateral arms. Quadriserial suckers present on middle third, followed by three to eight biserial rows; then about five biserial rows of suckers larger than those immediately proximal to them, but not as large as basal suckers. Minute suckers at tip of arm also biserial, unlike those of lateral arms.

Left ventral arm of male hectocotylized. One to three normal suckers at base followed by modified region, extending approximately half way along arm, which is transversely wrinkled and bears minute suckers laterally: seven pairs of
minute suckers on dorsal border; ventrally, the two series of suckers have moved together to form a single series of 14 suckers situated on extreme ventral edge of arm. Suckers biserial distally; the more proximal of these suckers (9-10 pairs) large, those on the tip minute.

In the female, all arms attenuated distally. Skin wrinkled basally on arms, next to sucker-bearing surface. This apparently not due merely to preservation, and has not been observed in any other Sepia species. Suckers on all arms biserial, and diminish gradually in size from base to tip. No enlargement of suckers on middle third of arms as found in males (Adam \& Rees 1966: 113, however, found that females may also have enlarged suckers on the arms. See below).

Tentacular club small and recurved, with numerous subequal suckers in transverse rows of about eight. Rings of suckers without teeth, but covered with numerous small knobs. Protective membranes separate proximally. Natatory membrane extends along tentacular stalk for a distance a little less than half club length.

Shell (Pl. 43a-d) ovate, acuminate anteriorly, narrowing and rounded posteriorly. Dorsal surface usually pink in colour, finely granular posteriorly. Chitinous margins broad in smaller shells. Distinct rounded median rib present dorsally, broadening somewhat anteriorly. Rib limited by distinct lateral grooves. Spine absent posteriorly, but pronounced rounded knob present. Behind this, shell bends sharply towards ventral. Striated zone long, with narrow concave region laterally, raised and flattened medially. Striae convex to angular ( $\wedge$-shaped), but with notch on either side, corresponding with point of meeting of raised middle region of striated zone with concave lateral areas. Median groove usually very faint, sometimes almost indistinguishable, but in a few shells is quite marked. Outer cone broad and deep.

Table 5. Sepia hieronis: a comparison of the relative dimensions (as $\%$ shell length) of the shells from the east and west coasts of southern Africa.

|  | West coast |  |  | East coast |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | N | Mean | Range | N | Mean | Range |
| Width | 4 | 39, 1 | 38,3-40,4 | 3 | 42,4 | 40,7-45,7 |
| Thickness | 4 | 11,0 | 9,6-11,7 | 3 | 14,3 | 13,0-15,2 |
| Striated zone | 4 | 72,2 | 70,0-73, 1 | 3 | 65,6 | $64,8-66,7$ |

Shells from east and west coasts show some marked differences, although having an overall similar appearance. In shells from west coast (Pl. 43a, b), inner cone not as well developed posteriorly, and with narrow limbs. In eastern shells ( $\mathrm{Pl} .43 \mathrm{c}, \mathrm{d}$ ), on the contrary, inner cone well developed posteriorly, reflexed and fused to outer cone, and with broad limbs. Eastern shells wider and thicker, have shorter striated zone, and median region of striated zone is more markedly raised than in western shells (Table 5). Yet differences between the
two shell forms are those of degree, and do not suggest different species (except perhaps differences of inner cones). No differences could be found between animals from the two coasts. Differences in relative dimensions of shells from east and west coasts possibly due to size, since shells from east coast generally smaller than those from west coast. Alternatively, differences may be due to contrasting environmental conditions on east and west coasts.

## Remarks

Smith (1916: 21), in the original description of $S$. acuminata, mentioned and figured (pl. II, fig. 4) a shell from Tongaat beach, Natal, which differs from the other examples of $S$. acuminata. From the description and figure, this specimen clearly belongs to $S$. hieronis, and differs from S. acuminata in that there is no spine, but only a posterior knob (just visible in Smith's figure), the outer cone is well developed posteriorly, the striated zone is more raised, and is divided into three distinct regions: one flat median and two concave lateral areas.

The arrangement of the suckers seems to vary considerably in S. hieronis, since every author describes them somewhat differently. But all are agreed that there are a number of pairs of enlarged suckers on the lateral arms of the males.

Robson (1924b: 646) described the hectocotylus as having one series of suckers ventrally in the modified region. This in fact corresponds to two series which have moved together to form a single series, as mentioned above. In one of the present specimens the hectocotylized arm is contracted, and the ventral series has become a zig-zag line, illustrating its origin from two longitudinal series of suckers.

According to Massy (1927: 159) the distal suckers on the lateral arms are biserial, but in the present male specimens they are quadriserial.

Adam \& Rees (1966: 113), in the description of the male, mention that the suckers of all the arms are proximally quadriserial, and imply that those of the dorsal arms are quadriserial throughout. In the present males, the suckers of the dorsal arms are biserial from base to tip, and on the lateral arms are biserial over at least part of the proximal half, although these conditions are sometimes rendered less clear due to contraction of the arms.

In one female (MLd 61 mm) described by Adam \& Rees (1966: 113 ) the dorsolateral arms have four pairs of enlarged suckers in the middle, minute suckers quadriserially arranged distally; 'the tips of all the arms, except the dorsal ones, seem to have the quadriserial arrangement of the suckers, but we are not sure that this is not due to contraction and that, in fact, all the suckers are biserial'. The female syntype has no enlarged suckers on the dorsolateral arms. In another female the enlarged suckers are well developed on the lateral and even on the ventral arms. In the present female (A30145, MLd 33 mm ) all the arm suckers are biserial, and none is enlarged. It is possible that the sucker enlargement only develops as the animals grow to maturity.

Previously, animals of $S$. hieronis were known only from the west coast of

South Africa, from $30^{\circ} 13^{\prime} \mathrm{S}, 15^{\circ}{ }^{1} 6^{\prime} \mathrm{E}$ to west of Slangkop (Robson $1924 a$ : 13 , 1924b: 645; Massy 1927: 158, 159; Voss 1962b: 251, 1967: 64; Adam \& Rees 1966: i12). Adam \& Rees (1966: 53, 112) found one shell of $S$. hieronis with specimens of S. acuminata from Stations 95 and 103 (Robson 1924a: 12, 13, 1924b: 643). These localities are approximately $30^{\circ} \mathrm{S}, 31^{\circ} \mathrm{E}$, which is off the Natal coast, near Durban. Smith's (1916:21) shell from Tongaat beach, Natal, was the only other record of $S$. hieronis from the east coast.

The present specimens from Monte Belo, Moçambique, are the first records of $S$. hieronis animals from the Indian Ocean. These specimens undoubtedly pertain to $S$. hieronis, although the shells show some differences from those found off the west coast. It is strange that neither the shell nor the animal of $S$. hieronis has so far been recorded between Cape Point and Durban.

Sepia insignis Smith, 1916
(Pl. 44a-c. Fig. I4. Tables 45, 46)
Sepia insignis Smith, 1916: 25, pl. II, fig. ıo. Tomlin, 1923: 41. Voss, 1962b: 248. Adam \& Rees, 1966: i14, pl. 3ı, figs 189-19ı.

## Type locality

Tongaat beach, Natal (shells only).

## Distribution

Animal: $34^{\circ} 15^{\prime} \mathrm{S}, 18^{\circ} 47^{\prime} \mathrm{E}$ (False Bay) (S.A.M.). Depth $4^{2} \mathrm{~m}$.
Shells: Bloubergstrand (S.A.M.) to Tongaat beach, Natal (Smith 1916: 25).

## Material

S.A.M. A30486, locality unknown; i shell

A31236, Kommetjie; i shell
A31241, Simonstown; 3 shells (r broken)
$\mathrm{A}_{3} 1247,34^{\circ}{ }^{\circ} 5^{\prime} \mathrm{S}, 18^{\circ} 47^{\prime} \mathrm{E}, 42 \mathrm{~m}$; I +
A $_{3}$ I248, between Strandfontein and Muizenberg; i shell
Still Bay; I shell
Bloubergstrand; i shell, broken
Description
Female specimen rather badly preserved, with mantle compressed laterally.

Mantle elongate oval, anterior margin produced dorsally, emarginate ventrally. Head short and broad. Fins rather wide, beginning a few mm behind anterior margin of mantle, rounded and separate posteriorly.

Skin sparsely papillose dorsally with concentric chromatophores on head and mantle. Mantle very wrinkled ventrally, but this may be due to preservation. Colour dark purple dorsally on head and mantle; chromatophores on mantle concentrated mid-dorsally, less dense laterally towards fins. Fins pale, with a few scattered chromatophores both dorsally and ventrally. Mantle pale mid-ventrally, with band of chromatophores on each side along bases of fins.


Fig. 14. Sepia insignis female, A31247. a. Dorsal view; b. ventral view; c. right tentacular club.
Arms I and II subequal in length, about one-third MLd, arms III a little shorter, arms IV longer, very well developed, and wrinkled ventrally. Shallow interbrachial web present, except between ventral arms. Dorsal arms attenuated distally over about half arm length. Arms III and IV strongly keeled. Protective membranes well developed on all arms, and wide enough to meet over inner surface of arm.

Arms unfortunately rather distorted, and some suckers lost, especially from delicate dorsal arms. Suckers on dorsal arms biserially arranged from base to tip. On arm II, five pairs of suckers basally followed by very oblique quadriserial rows to arm tip. On arm III, two pairs of suckers basally, then one or two to fill in gap before obliquely quadriserial suckers, which continue to tip. Suckers on ventral arms very obliquely quadriserial, with one row of three suckers basally. Quadriserial rows so oblique that suckers appear to be almost biserial. None of the suckers enlarged; size of suckers decreases gradually from base to tip on all arms. Suckers globose, with nodular rings which have irregular but not toothed edges.

Tentacular club fairly small, bearing a number of subequal suckers in transverse rows of about eight. Suckers have large rings with nodular surface and smooth edge. Natatory membrane well developed, continuing a little beyond club. Protective membranes well developed; ventral membrane curves around base of club, but not meeting dorsal membrane, which ends just before base of club. Upper surface of club with a number of transverse rows of chromatophores.

Shell (Pl. 44a-c) elongate, fairly narrow, sharply acuminate anteriorly, then about same width over approximately two-thirds of its length; rounded
posteriorly. Posterior spine absent. Shell a pale pink colour dorsally. Mid-dorsal longitudinal ridge present, but not limited by lateral grooves. Ridge ends posteriorly in blunt knob, whereafter shell curves sharply to ventral. Striated zone long, narrowing markedly posteriorly; striae $\wedge$-shaped between limbs of inner cone. Well defined median longitudinal groove present over striated zone, continuing along last loculus, where it is less distinct. Inner cone well developed posteriorly, completely reflexed and fused to outer cone. Inner cone curved over sides of striated zone laterally, as in S. burnupi, and limbs of inner cone lie on phragmocone. In smallest shell, limbs of inner cone much nearer edges of phragmocone than in larger shells. It would appear that limbs of inner cone move towards middle of shell in older shells. Outer cone broad laterally.

## Remarks

Until now only shells of $S$. insignis were known. The present female was found in the collection of the University of Cape Town Ecological Survey, and is now lodged in the collection of the South African Museum.

The shell from Bloubergstrand (Pl. 44c) must have measured about 55 mm when whole, and is the largest shell known of $S$. insignis.

This species seems to be related to $S$. hieronis. The shells of the two species show some resemblance, and the animals are similar in that both species have subequal tentacular suckers and biserial suckers on the dorsal arms.

Sepia robsoni (Massy, 1927)
(Tables 6, 50)
Rhombosepion robsoni Massy, 1927: 159, pl. XVII, figs $1-8$.
Sepia robsoni Voss, 1962b: 248. Adam \& Rees, 1966: 120, pl. 46, fig. 279.

## Type locality

Hout Bay, $17-37 \mathrm{~m}$.

## Description

Only one specimen (male) ever caught, and was not available for examination. Following description after Massy (1927: 159) and Adam \& Rees (1966: 120).

Animal small. Mantle broadly oval, anterior mantle margin dorsally slightly convex (Adam \& Rees 1966: 120), very slightly produced in the centre (Massy 1927: 160), ventrally deeply emarginate. Fins wide, beginning three mm behind mantle margin, separate posteriorly.

A few tubercles present dorsally along outline of shell and on head. Fleshy ridge present on either side on mantle ventrally, near fin bases, as in S. typica, but pores absent. Flesh-colour to pale brown, with minute dark chromatophores dorsally, extending partly on to fins. Ventrally with a few chromatophores along fin base.

Arms subequal in length, with ventral arms slightly longer than dorsal ones. Interbrachial web well developed between arms I to III, attaining half
arm length, lower between arms III and IV, absent between ventral pair. Suckers globose, with smooth rings; skin adjacent to rings grooved; suckers biserially arranged, none enlarged.

Dorsal arms with finger-like tips, devoid of suckers, and with eight pairs of suckers proximal to naked tips. Dorsolateral arms with eight pairs of large subequal suckers and a few small suckers distally. Ventrolateral arms with nine pairs of large subequal suckers. Right ventral arm with seven pairs of large suckers, of which suckers of ventral series larger than those of dorsal series, and much larger than those of other arms. Left ventral arm hectocotylized over basal three-quarters of its length, bearing ten pairs of minute suckers on modified region.

Tentacular club crescent shaped, bearing about 53 more or less subequal suckers, in transverse rows of four to six, median suckers being slightly larger than others. Rings of tentacular suckers papillose (? nodular), dentate on at least part of the ring (Massy 1927: 160). Natatory membrane very broad, extending beyond base of club for a distance equal to about half club length; dorsal protective membrane very broad at base of club.

Shell completely chitinized (? decalcified), in poor condition. Anterior part strongly acuminate, resembling that of $S$. hieronis.

## Remarks

According to Massy (1927: 159), Sepia robsoni resembles S. hieronis, but differs from the latter in having grooved suckers (smooth rings) biserially arranged on all the arms, and none is enlarged in the male. 'The shell somewhat resembles that of $S$. insignis, but the latter is even more acuminate anteriorly and seems to be much wider in its posterior part' (Adam \& Rees 1966: 121).
$S$. robsoni also resembles $S$. dubia (for comparison, see Table 6) in that all the arm suckers are biserially arranged, the mantle is very broad, and the ventral mantle surface has fleshy keels without pores. The hectocotylus is like that of $S$. typica, and the interbrachial web is well developed. The tentacular club is exactly like that of S. typica (Adam \& Rees 1966: 121). Sepia robsoni differs, however, from both $S$. typica and $S$. dubia in its shell, which presumably has the phragmocone covering almost the entire dorsal lamella (since it is compared with those of $S$. hieronis and $S$. insignis) and in the dorsal arms with the tips devoid of suckers. In this latter feature $S$. robsoni resembles $S$. faurei.

## Sepia faurei n . sp.

(Figs 15, 16. Tables 6, 50)

## Type locality

S $14{ }^{\circ} \mathrm{E}$ of Cape Seal 88 km , 168 m.

## Material

S.A.M. A30144, S $14{ }^{\circ} \mathrm{E}$ of Cape Seal $88 \mathrm{~km}, 168 \mathrm{~m}$; I ㅇ (holotype)

## Description

Animal small. Mantle broadly oval, anterior mantle margin convex dorsally (not markedly produced), emarginate ventrally. Fins narrow, beginning a few mm behind mantle margin, separate posteriorly.

Colour uniform brown on head and mantle dorsally, with small darker spots on arms. Skin densely papillose dorsally. Fleshy ridge present on either side of mantle ventrally, near fin bases, as in S. typica, but pores absent.


Fig. 15. Sepia faurei female, A30144 (holotype). a. Dorsal view and b. ventral view.

Arms subequal in length; interbrachial web high dorsally, attaining about half arm length, but low between ventrolateral and ventral arms, and absent between ventral pair. Suckers globose, without teeth on chitinous rings, but these covered with numerous raised knobs. Suckers on all arms biserially arranged. Distally all arms except ventral pair attenuated over about half arm length; suckers on attenuated portion (abòut six pairs on arm I, if pairs on arm II, and i6 pairs on arm III) minute, and protective membranes very well developed. Tips of attenuated distal half of dorsal arms finger-like, devoid of suckers and protective membranes (Fig. 16a). Six to nine pairs of subequal suckers proximally (nine pairs on arm I, six pairs on arm II, and seven pairs on arm III), of which last pair somewhat smaller. Ventral arms not markedly attenuated distally, bearing eight pairs of subequal suckers proximally, and about 12 pairs of minute suckers on tips.


Fig. 16. Sepia faurei female, A30144 (holotype). a. Detail of distal part of dorsal arm. Position of missing suckers indicated by dotted rings. b. Right tentacular club. c. Ventral view of shell (incomplete).

Tentacular club broad, slightly recurved, bearing 33 subequal suckers in transverse rows of four to six (Fig. 16b). Median suckers a little larger than lateral suckers. Sucker rings not toothed, with nodular surface. Dorsal protective membrane well developed, separate from ventral membrane proximally. Natatory membrane broad, continuing along tentacular stalk for a distance equal to club length.

Shell (Fig. 16c) not calcified, very thin, as in S. typica, broadly ovate,
Table 6. Comparison of the main characters of Sepia robsoni (Massy), S. dubia Adam \& Rees and S. faurei n. sp.

|  | S. robsoni | S. dubia | S. faurei |
| :---: | :---: | :---: | :---: |
| Sex . | male | female | female |
| MLd | 17 mm | 17 mm | 21 mm |
| Anterior mantle margin | Dorsally slightly convex, ventrally deeply emarginate | Dorsally slightly convex, ventrally deeply emarginate | Dorsally convex, ventrally emarginate |
| Dorsal papillae | A few tubercles along the outline of the shell and on the head | Dorsal surface of mantle, head and arms covered with well-spaced round papillae, creating a very rugose appearance. Mid-dorsally on the mantle there are two oval patches of contracted papillae, one on each side of the median line, and a third one anteriorly near the mantle margin | Skin dorsally on head, mantle and arms densely covered with small papillae |
| Ventral mantle surface | Much like that of Hemisepius typicus with the fleshy ridge, but there is no trace of any pores | Smooth, with a thick fleshy keel parallel to the outer margin, as in H. typicus, but without pores | A fleshy ridge on either side near the fin bases, but no pores |
| Arm lengths | Arms subequal in length | Arms subequal in length | Arms subequal in length |
| Interbrachial web | The web reaches halfway up the arms between arms I, II and III; between arms III and IV it is much lower and is completely lacking between arms IV | The web is very high between arms I, II and III, reaching halfway up the arms; it is much lower between arms III and IV and non-existent between arms IV | Web does not attain half arm length between arms I, II and III; it is even lower between arms III and IV, and is absent between arms IV |
| Arm suckers | Suckers globose; biserially arranged on all the arms | Rather small and biserial | Globose; biserial on all arms |


| The suckers on the dorsal arms | The tips of the attenuated distal half |
| :--- | :--- | of the dorsal arms are finger-like and devoid of suckers and protective membranes


| Dorsal arms | 'The distal portions are thickened and end in rounded knobs without suckers. When observed laterally the surface is seen to be folded as if suckers might be in the course of formation. The appearance may therefore be due to accident' (Massy 1927). But in Massy's figures (pl. XVII, figs 1 , 2) the dorsal arms appear slender at the tips. According to Adam \& Rees (1966), who re-examined the specimen, 'the dorsal arms have finger-like tips, devoid of suckers'. They do not mention any thickening | The suckers on the dorsal arms continue to within $0,4 \mathrm{~mm}$ of the tips (Taylor 1968, personal communication) |
| :---: | :---: | :---: |
| Tentacular club | About 53 suckers arranged in transverse rows of $4-6$. About 3 median rows have larger suckers than the rest. Dorsal protective membrane very broad at base of club. Natatory membrane very broad and extending beyond base of club for about half the length of the club | Minute subequal suckers arranged in 4 or 5 longitudinal series (Adam \& Rees 1966). There are in fact 54 suckers, of which the median ones are slightly larger (Taylor i968, personal communication). Dorsal protective membrane wide and separated from the ventral one at base of club. Natatory membrane well developed, extending beyond base of club for about half its length |

Table 6 (continued)

|  | S. robsoni | S. dubia | S. faurei |
| :---: | :---: | :---: | :---: |
| Shell . | Completely chitinized (? decalcified); in poor condition. Its anterior part is strongly acuminate and resembles that of $S$. hieronis | Shell broadly oval, acuminate anteriorly, but less so than in H. typicus and is broadly rounded behind. It is a low dome-shape in the middle but is flatter near the margins. Nearly the whole dorsal surface is calcareous, with a reticulate pattern. No spine. Ventral surface strongly concave and spoon-shaped. The central part has a reversed conical shape and is slightly higher than the surrounding part; it occupies a little more than half the shell length, as in H. typicus. Last loculus trapezoid, widest at anterior margin. This last loculus represents about half the length of the striated area. Behind the last loculus, the striated area occupies only the central third portion of the whole inner zone and is flanked by two broad, smooth, marginal areas. The striae, about ${ }^{1} 5$ in number, are widely spaced, transverse, nearly straight, being only slightly wavy. There is a distinct inner cone, brownish in colour in alcohol, with rather broad limbs, that are completely fused to the outer cone; its posterior part surrounds a very shallow depression. The broad outer cone surrounds completely the inner cone | Shell completely chitinous. It is broadly oval, somewhat acuminate anteriorly. The ventral phragmocone roughly triangular in shape, and does not cover the anterior part of the dorsal lamella. Last loculus, which has a convex anterior border, probably covered about half the length of the phragmocone. The striated zone occupies the central region of the phragmocone. On either side of the striated zone there is a smooth marginal area. The striae are transverse, and slightly rounded. Unfortunately the posterior part of the shell has been lost, and no indication of the inner cone remains |

somewhat acuminate anteriorly. Posterior end damaged. As in S. typica, phragmocone not covering anterior part of dorsal lamella. Anterior border of last loculus convex. Striae fewer and last loculus longer than in S. typica. Striated zone occupying central region of phragmocone; smooth marginal area present on either side of striated zone. Striae transverse, slightly convex in shape. No indication of inner cone remains.'

## Remarks

Sepia faurei resembles $S$. robsoni in the absence of suckers distally on the dorsal arms. It resembles both $S$. robsoni and $S$. dubia in that the mantle is very broad and has fleshy keels, without pores ventrally, and the arm suckers are biserially arranged. The three species are compared in Table 6.

The shell of $S$. faurei is similar to those of $S$. typica and $S$. dubia, being very thin, with the phragmocone not covering the entire dorsal lamella. In S. typica and $S$. dubia, however, the anterior border of the phragmocone is not parallel to the corresponding sides of the dorsal lamella, whereas in $S$. faurei it is more nearly so, and the phragmocone is somewhat longer in the latter species.

Sepia faurei, showing relationships with both S. robsoni and S. dubia, seems to represent an intermediate link in the transition from Sepia to Hemisepius. On the one hand it is related (by virtue of the dorsal arms) to S. robsoni, which apparently has a Sepia-like shell with the phragmocone covering almost the entire dorsal lamella anteriorly, and on the other hand $S$. faurei is related to $S$. (Hemisepius) dubia and S. (Hemisepius) typica with a Hemisepius shell, in which the phragmocone is much shorter than the dorsal lamella.

## Distinctive characters

I. Tips of dorsal arms finger-like, devoid of suckers and protective membranes
2. Suckers biserially arranged on all arms
3. Mantle very broad, with fleshy keels ventrally
4. Skin densely papillose dorsally on head, mantle and arms
5. Shell with phragmocone considerably shorter than dorsal lamella, but with anterior margin of phragmocone convex in shape
The holotype of Sepia faurei was collected during one of the cruises of the Cape Fisheries survey vessel Pieter Faure (P.F.14290, ig February 1902), after which this species has been named.

Sepia (Hemisepius) typica (Steenstrup, 1875)
(Fig. 17. Tables 7, 47-49)

[^11]
## Type localities

Table Bay; St. Francis Bay (var. chuni).

## Distribution

Animals: Saldanha Bay (Hoyle, 1912: 281) to Cape Natal, W by N, 10 km (Massy 1927: 161). Depth 2-156 m.
Material
S.A.M. A8989, locality unknown; I $\begin{gathered}\text { t in poor condition }\end{gathered}$

A2g6o8, south side of Schaapen Island, Saldanha Bay, 4 m; 3 ㅇ
A29717, SSE of Ystervarkpunt, 92 m (det. G. L. Voss); 7 d $\boldsymbol{\text { d }}$, 6 ㅇ
A29783, Saldanha Bay, 7 m ; 15 d', 31 아
A30176, S $34^{\circ} \mathrm{W}$ of Cape Infanta $30 \mathrm{~km}, 84 \mathrm{~m} ; 5 \delta^{\circ}$ (of which two in poor condition), 2 아
A30177, $34^{\circ} 14^{\prime} \mathrm{S}, 22^{\circ} 23^{\prime} \mathrm{E}$ (Mossel Bay), $60 \mathrm{~m} ; 5$ ot, 4 아
A30269, $\mathrm{S}_{1} 6^{\circ} \mathrm{W}$ of Cape Point lighthouse $16 \mathrm{~km}, \mathrm{I}_{5} 6 \mathrm{~m}$; $\mathrm{I} \mathrm{o}^{\wedge}, 4$ f, all in poor condition
A30484, locality unknown; i shell in poor condition

## Description

Animals small; largest specimen in present collection being a female of MLd 25 mm ; largest male has MLd 21 mm .

Mantle very broadly oval, almost as wide as long. Anterior mantle margin convex dorsally, emarginate ventrally. Ventral surface of mantle bearing two fleshy ridges near fin bases; each ridge with a number of pores anteriorly (most commonly $10-12$ on each side, but the number may vary between five and 15 . Number of pores not always the same on both sides). Ridges becoming less distinctive posteriorly. No evidence of longitudinal groove linking pores, as described by Steenstrup (1875: II).

Head short and broad. Fins narrow, beginning a few mm behind anterior mantle margin, fused posteriorly.

Colour dark reddish-purple dorsally on head, arms and mantle, with darker diamond-shaped region mid-dorsally over shell. Two pale round tubercles with concentric chromatophores present in middle of darker region. Fins pale dorsally with scattered dark chromatophores, most dense near fin bases. In some cases short transverse orange bands present on fins. Colour pale ventrally, except fór darker colour of ridges bearing pores, and sparse chromatophores between ridges and fin bases; a few chromatophores present laterally on funnel.

Skin very sparsely papillose dorsally, a few papillae around eyes being the most marked. Mid-dorsal tubercles on mantle very flattened.

Arms subequal in length, fairly short (about $40-50 \%$ MLd). Interbrachial web deep, attaining half arm length, but absent between ventral arms. All arms triangular in cross section, but only well developed ventral arms keeled.

Suckers on all arms biserial, globose, not flattened and disc-shaped, as described by Steenstrup (1875: II). Chitinous rings smooth edged, with nodular
surface. In female, size of suckers decreases gradually from base, but suckers suddenly become much smaller at tips of arms; about six to eight pairs of minute biserial suckers present. In male, one to four pairs of suckers near tips of arms I to III enlarged. No enlargement occurs on ventral arms, but in some cases suckers of dorsal series of right ventral arm larger than those of ventral series. Thore (1945:52) found, however, that suckers of 12 th to 18 th rows on right ventral arm were enlarged in his specimens from Table Bay and Oukraal (Oude Kraal). One male from Saldanha Bay (A29783) shows abnormal arrangement of suckers on third arms, perhaps due to contraction. Middle of right arm III bears three rows of suckers in irregular quadriserial arrangement, and one row of three suckers. Left arm III has one row of three suckers, four very oblique quadriserial rows, then a single dorsal sucker medially. These arms normal distally.


Fig. 17. Sepia (Hemisepius) typica. a. Dorsal and b. ventral view of female, A29783. c. Dorsal and d. ventral view of shell of female, A2g6o8.

Left ventral arm of male hectocotylized. Basal half or more of arm modified, bearing nine to 13 pairs of minute suckers arranged in two widely spaced series separated by fleshy transverse ridges on arm. Distal half of arm normal, bearing five to six pairs of normal suckers, then about eight pairs of minute suckers on arm tip. A few basal modified suckers sometimes larger than the rest.

Tentacular club small and straight, bearing numerous subequal suckers in oblique transverse rows of six. Rings of suckers broad and nodular, without teeth. Protective membranes separate proximally. Natatory membrane very
well developed, continuing along stalk for a distance from one-half to once club length.

Shell (Fig. 17c, d) not calcified, very thin and fragile. One shell, successfully dissected out (A29608), has dimensions: length $19,5 \mathrm{~mm}$, width ${ }_{10}, 5 \mathrm{~mm}$, length of striated zone 10 mm . It is broad, pointed anteriorly, rounded posteriorly. No posterior spine or knob, and no median dorsal ridge present. Phragmocone very short, triangular in shape, with last loculus constituting base of triangle anteriorly. Last loculus not covering anterior part of dorsal shield. No median longitudinal groove ventrally. Striated zone occupies most of phragmocone; striae wavy. Inner cone barely discernible, completely fused with outer cone; forming, together with its limbs, a circle about posterior point of striated zone. Outer cone broad.

## Remarks

When first describing this species, Steenstrup (1875:468) created a new genus for it, on the basis of the following characters:
I. The ventral mantle surface has deep pores, which in Hemisepius typicus are arranged in two lines of 12 pores each.
2. The shell is poorly developed, with very rudimentary, calcareous loculi not covering the anterior part of the dorsal lamella, and their anterior border is not parallel to the corresponding sides of the extremely thin lamella.
3. All arms with biserial suckers, which are very flattened, almost smooth, disc-like.
Until recently, these characters clearly separated H. typicus from all the remaining Sepiidae (although the number of pores was found to vary somewhat, and the suckers are not flattened, but globose) and the genus was valid. But a recently discovered species, Sepia dubia, first described by Adam \& Rees (1966: II 19 ), shows a number of characters which bridge the gap between the genera Hemisepius and Sepia, and Adam \& Rees relegated Hemisepius to subgeneric status.

Chun (1915:412) first described the male of $S$. typica, and illustrated the hectocotylus. He found that most of the suckers in the ventral series of the modified region were absent, but these had probably been lost, since all the males in the present collection have a complete ventral series on the hectocotylus.

Smith (1916: 26) suggested the possibility that S. tuberculata be identical with S. typica. It is difficult to understand how Smith came to this supposition, since both the shells and the animals of these two species are very different. The only possible similarities in the shells are the general outline and the thinness (although the shell of S. typica is much broader and very much thinner than that of $S$. tuberculata). As remarked by Adam (1941: i16), the suggestion is clearly untenable.

Thore (1945:50) found that his specimens of S. typica from Table Bay and Oukraal (MLd $22-27 \mathrm{~mm}$ ) were larger than that described by Chun (1915:412)
(MLd $\mathrm{I}_{7} \mathrm{~mm}$ ) from a more easterly locality, St. Francis Bay, but comparable with those of Steenstrup (from Table Bay) and of Massy (1927: 164) (from False Bay and Hout Bay). On the basis of this, he stated: 'I think we have to postulate a constant difference in size between the eastern and western form of Hemisepius, the latter being the largest' (Thore 1945:50), and proposed that the eastern forms be named $H$. typicus var. chuni.

Thore found that his western male specimens also differed from that described by Chun in the number and size of the suckers on the ventral arms. The hectocotylus of Thore's specimens differed from that figured by Chun in that the dorsal row of suckers was separated from the edge of the arm by a distinct longitudinal groove; the proximal suckers in the ventral row were not enlarged; the second-last sucker of the dorsal row (in the modified region) was not enlarged; the enlarged suckers distal to the modified region were about 12 in number (five in Chun's specimen) and there were about 20 minute suckers at the tip of the arm ( 13 in Chun's specimen).

In all, only three specimens of S. typica are known from localities east of $24^{\circ}$ E: one male (MLd 17 mm ) from St. Francis Bay (Chun 1915: 412), one female (MLd 18 mm ) from east of Port Elizabeth (Voss 1962b: 252) and one male (MLd 13 mm ) from Cape Natal (Rhombosepion sp. A, Massy 1927: 16r, determined as $S$. typica by Adam \& Rees 1966: 117). In the male from Cape Natal, the suckers distal to the modified region of the hectocotylus were absent, presumably lost. This leaves one eastern male with which to compare the western specimens. A comparison of the specimens of Thore and Chun is given in Table 7, together with the specimens in the collection of the South African Museum.

From observations on the specimens and the data in the table, the following points become apparent:
I. Chun's specimen from St. Francis Bay is not unusually small, as it falls into the size ranges of mature specimens from Saldanha Bay and Mossel Bay, and is larger than those from Cape Infanta and Ystervarkpunt.
2. The number and position of enlarged suckers on arms I to III apparently varies randomly; there is no correlation with mantle length or the locality of the specimens. The distal suckers on arms III are not always markedly enlarged.
3. Right arm IV: the total number of suckers increases with the size of the animal, but the specimens from Ystervarkpunt have fewer suckers than those of the same size from other localities; Chun's specimen has an unusually low number of suckers for its size-a specimen of MLd 17 mm from Mossel Bay has 2I pairs of suckers on the fourth right arm. The position of the distal enlarged suckers is not closely related to the total number of suckers, but some enlarged distal suckers are almost always present; in this, Chun's specimen again differs. There are usually three pairs of enlarged suckers basally, but occasionally two or

|  |  | Saldanha Bay (A29783) | Table Bay (Thore 1945) | Off Cape Infanta $\left(\right.$ A $\left._{3017}\right)$ |
| :---: | :---: | :---: | :---: | :---: |
| Dorsal mantle length |  | 11-19 mm | 22-27 mm | $1 \mathrm{i}-16 \mathrm{~mm}$ |
| Enlarged suckers | Arm I | Variable enlargement of suckers from 8-12th rows, usually including those of 9-roth rows | Enlargement of ioth sucker pair, and variable enlargement of 2-3 pairs of distal suckers, or of some suckers of one series | Variable enlargement of suckers from 8-11 th rows, usually including those of gth row |
|  | Arm II | Variable enlargement of suckers from 8-12th rows, usually including those of ro-isth rows | As arm I | Variable enlargement of suckers from 7-I ith rows, usually including those of 9th row |
|  | Arm III | Variable enlargement of suckers from ${ }^{12-16 t h}$ rows, usually including those of 13-14th rows | Distal enlargement of suckers (which rows not specified) | Variable enlargement of suckers from ro-12th rows, usually including those of ro-rith rows |
| Suckers on right arm IV | Total number | 19-27 pairs | About 30 pairs | 21-22 pairs |
|  | Enlarged distally | Variable enlargement from 11-17th rows | 12th to 18th pairs | 12th to 14th pairs |
|  | Enlarged proximally | Basal 3 or 4 pairs | Basal 3 pairs | Basal 2 to 3 pairs |
| Suckers on hectocotylus (left arm IV) | Number minute distally | 10-20 | About 20 | 12-19 |
|  | Number enlarged between modified region and minute distal suckers | 8-10 | 12 | 8-9 |

Table 7 (continued)

|  |  | Off Ystervarkpunt (A29717) | Mossel Bay (A30177) | St. Francis Bay (Chun 1915) |
| :---: | :---: | :---: | :---: | :---: |
| Dorsal mantle length |  | 10-15 mm | $14^{-21 ~ m m}$ | 17 mm |
| Enlarged suckers | Arm I | Variable enlargement of suckers from 6-9th rows, usually including those of 7-8th rows | Variable enlargement of suckers from 7-I I th rows, usually including those of 9th row | 1oth pair enlarged |
|  | Arm II | Variable enlargement of suckers from 6-1oth rows, usually including those of 8th row | Variable enlargement of suckers from 8-13 th rows, usually including those of 9-1oth rows | roth pair enlarged |
|  | Arm III | Variable enlargement of suckers from $10-12$ th rows, usually including those of ro-i ith rows | Variable enlargement of suckers from $9^{-1} 4^{\text {th }}$ rows | No enlargement reported |
| Suckers on right arm IV | Total number | 16-21 pairs | 21-26 pairs | 13 pairs, gradually decreasing in size from base to tip |
|  | Enlarged distally | 9-1 $4^{\text {th }}$ pairs | 1 1-16th pairs |  |
|  | Enlarged proximally | Basal 2-3 pairs | Basal 3-4 pairs |  |
| Suckers on hectocotylus (left arm IV) | Number minute distally | 8-12 | 16-20 | 13 |
|  | Number enlarged between modified region and minute distal suckers | 6-10 | 8-12 | 5 (or 8?) |

four pairs; Chun's description does not preclude the possibility that these were also present in the specimen from St. Francis Bay.
4. Hectocotylus: the number of minute distal suckers of Chun's specimen is within the normal range for its size. The number of enlarged distal suckers in Chun's specimen is unusually small; from the figure (Chun 1915, fig. 34), Thore deduced that the penultimate sucker of the dorsal series on the modified region is enlarged, and that there are five enlarged suckers distal to the modified region of the hectocotylus. An alternative interpretation is that this single enlarged dorsal sucker is part of the group of distal enlarged suckers, of which the second pair has failed to become enlarged. A similar case was observed in a specimen (MLd 16 mm ) from Cape Infanta, in which one sucker of the second pair of enlarged suckers has remained minute. The longitudinal groove separating the dorsal series of suckers from the edge of the modified surface, as reported by Thore, was observed in some specimens but not in others, and apparently depends on the state of preservation of the specimens. The three enlarged proximal suckers in the ventral row on the modified region, as illustrated by Chun, are absent in Thore's specimens. In the present specimens the proximal suckers of the dorsal and/or ventral row were sometimes found to be somewhat larger than the more distal suckers on the modified region of the hectocotylus. But the proximal enlargement is not marked, and the size of the suckers of the modified region gradually diminishes distally. The number of large suckers, and their position, apparently varies randomly.
Thus Chun's specimen differs from those from more westerly localities only in that it has fewer suckers on the right arm IV, of which none are enlarged distally. Whether or not this constitutes a valid character for separating eastern and western forms of S. typica cannot be decided on the basis of the presently known specimens, and a decision must await the collection of further specimens from the eastern coast of South Africa. In any case, the 'variety' is no longer recognized as a valid taxon within the system of nomenclature, and should be replaced by the term subspecies, provided the eastern and western forms of S. typica are found to differ sufficiently.

Sepia (Hemisepius) dubia Adam \& Rees, 1966
(Tables 6, 50)
Sepia (Hemisepius) dubia Adam \& Rees, 1966: 119, pl. 34, figs 198-201, pl. 46, fig. 272, text fig. I.

## Type locality

False Bay, $34^{\circ}$ II'S, $18^{\circ} 27^{\prime} \mathrm{E}, 25 \mathrm{~m}$.

## Description

The only known specimen (female) was not available for examination. Following description after Adam \& Rees (1966: ir9) and Taylor (personal
communication), who kindly supplied some additional information about the type specimen.

Animal small. Mantle broadly oval, anterior mantle margin slightly convex dorsally, deeply emarginate ventrally.

Dorsal surface of mantle, head and arms covered with well spaced round papillae; in addition, three oval patches of contracted papillae present medially: one on either side of median line, and third one anteriorly near mantle margin. Ventral mantle surface smooth, with thick fleshy keel, parallel to outer margin, as in S. typica, but without pores.

Arms subequal in length, laterally compressed, keeled on outer sides; protective membranes narrow. Web very high between dorsal and lateral arms, attaining half arm length; lower between arms III and IV, absent between ventral pair. Arm suckers rather small; biserially arranged.

Tentacular club small, crescent shaped, bearing 54 minute subequal suckers arranged in four or five longitudinal series (according to Taylor, personal communication, the median suckers are slightly larger than the others). Natatory membrane well developed, extending beyond base of club for a distance equal to about half club length. Dorsal protective membrane wide, separated from ventral membrane at base of club.

Shell broadly oval, somewhat acuminate anteriorly, broadly rounded posteriorly. Almost whole dorsal surface calcareous, with reticulate pattern. Posterior spine absent. Ventral surface strongly concave, spoon shaped. Phragmocone has reversed conical shape, occupying a little more than half shell length, as in S. typica. Last loculus trapezoid, widest at anterior margin. Striated zone about twice as long as last loculus, but occupying only central third of width of phragmocone; broad smooth marginal area present on either side of striated zone. About 15 widely spaced, transverse, slightly wavy striae present. Inner cone distinct, brownish in colour, with rather broad limbs, completely fused to outer cone. Posterior part of inner cone surrounds shallow depression. Outer cone broad, completely surrounding inner cone.

## Remarks

Sepia dubia resembles $S$. typica in that it has biserial suckers on the arms, the tentacular clubs are the same, and the shell is very similar, with a very short phragmocone, whose anterior border is not parallel to the corresponding sides of the thin dorsal lamella. The shell differs however, in that it has a calcareous covering of the dorsal surface, and a distinct inner cone. Sepia dubia has fleshy keels on the ventral mantle surface, but there are no pores. It also differs from S. typica in that the skin is not smooth, but is covered with well spaced papillae dorsally.

Whilst $S$. dubia is obviously closely related to $S$. typica, the differences between the two species include a character (pores in the ventral mantle surface) listed by Steenstrup (1875: II) as defining the genus Hemisepius, and led Adam \&

Rees (1966: 143) to state that 'the genus or subgenus Hemisepius may be maintained for H. typicus and H. dubius, but its separation from other Sepiidae has become less distinct'. In the taxonomic section of their review (Adam \& Rees 1966: 117, 119) they have relegated Hemisepius to subgeneric status for S. typica and $S$. dubia.

Sepiella cyanea Robson, 1924
(Pl. 42c, d. Fig. 3b. Tables 5i-53)
Sepiella cyanea Robson, 1924a: 13; 1924b: 648, figs $25-27$, pl. II, fig. 6. Adam, $1939 b$ : 109 , figs $\mathrm{I}_{4} \mathrm{~A}-\mathrm{B}$, pl. IV, figs 3,4 . Voss, $1962 b: 24^{8}$. Adam \& Rees, 1966: 12 I , pl. 36, figs 208-215. Sepia sp. a Voss, 1962a: 3.
? Sepiella obtusata (non Pfeffer) Massy, 1928: 95.
Type locality
S.S. Pickle, Sta. 476: $29^{\circ} 17^{\prime} \mathrm{S}, 31^{\circ} 33^{\prime} \mathrm{E}$, $5^{1} \mathrm{~m}$ (lectotype, designated Adam \& Rees 1966: 121).

## Distribution

Animals: Port Elizabeth (Adam \& Rees 1966: 121) to $29^{\circ}{ }^{\circ} 7^{\prime} \mathrm{S}$, $31^{\circ} 33^{\prime} \mathrm{E}$ (off Tugela River) (Robson 1924a: 14, Sta. 476) and Nosy N'Tangam (Malagasy) (Adam \& Rees 1966: 121). Depth 51-73 m.
Shells: Port Elizabeth (Adam \& Rees 1966: 121) to ? Tongaat (Massy 1928: 95) and Ambavanibé, Malagasy (Adam 1939b: ro9).

## Material

S.A.M. A6526, Port Alfred; I ô, 5 shells

## Description

Mantle elongate oval, anterior mantle margin produced dorsally to eye level, emarginate ventrally. Mantle bluntly rounded posteriorly and posterior gland (characteristic of genus) opens via pore situated between and just below posterior extremities of fins. Fins fairly wide, beginning a few mm from mantle margin, rounded and closely approximated posteriorly. Posterior region of fins somewhat damaged in present specimen, but according to Adam \& Rees (1966: 122), fins fused at base.

Skin smooth; colour of head and mantle dark blue-purple mid-dorsally, paler towards fins, on each of which a series of dark wedge-shaped patches present in male. Ventral surface pale, with sparse chromatophores medially, more concentrated near fins.

Arms fairly short, with arm length formula 4.3.1.2, and joined by low interbrachial web, also present between ventral arms (but Adam \& Rees 1966: 123, report that web absent between ventral arms). All arms keeled. Arm tips attenuated. Suckers quadriserially arranged on all arms; chitinous rings of suckers toothed distally with about 12 long teeth.

Left ventral arm hectocotylized over basal half. Minute suckers in modified region arranged in one dorsal, one medio-dorsal and two ventral series. Arm surface between dorsal and ventral series is transversely ridged, and suckers of medio-dorsal and dorsal series are situated on the ridges. Middle and distal parts of arm somewhat mutilated, but distal part apparently normal, with quadriserial suckers.

Tentacles of present specimen missing. According to Adam \& Rees (1966: 123) 'the tentacular stem is triangular in cross-section with a rounded keel on the outer side and a flat, transversely-striated, inner surface, which is limited by two membraneous ridges, these being the continuation of the protective membranes of the club. The swimming-membrane of the latter is not very broad and barely reaches the base of the club. The protective membranes are narrow. The minute, subequal suckers are arranged in about 12 longitudinal series in both sexes (pl. 36, fig. 215). Their chitinous rings are each armed with a few, blunt, spaced teeth.' Robson ( $1924 b: 648$ ) also observed two enlarged suckers at the extremity of the club.

Shell (Pl. 42c, d) elongate oval, somewhat narrower anteriorly, but not sharply acuminate; posteriorly rather broadly rounded. Dorsal surface calcareous, with median longitudinal ridge and lateral grooves. Chitinous margin fairly broad. No posterior spine, but slight hump present over posterior extremity of striated zone. Broad shallow groove present, running from hump to posterior margin. Striated zone long, fairly broad posteriorly. Anterior margin of striated zone broadly convex, somewhat wavy. Last loculus continues along sides of striated zone to meet limbs of inner cone. Anterior part of smooth zone shows some compression in most shells. Faint indication of median longitudinal groove over striated zone only. Inner cone forms knob posteriorly, and has very short narrow limbs. Outer cone very broad.

Females differ from males in that arms are relatively much shorter, protective membranes on arms better developed, covering distal suckers (Adam \& Rees 1966: 123) and sucker rings almost smooth (Robson 1924b:648). Shell of female broader, ventral surface thinner, inner cone more developed and outer cone broader (Adam \& Rees 1966: 123) and striated zone less pointed (Robson 1924 $b$ : 649).

## Remarks

The genus Sepiella is represented by only one known species, $S$. cyanea, in southern African waters. Together with $S$. ornata and $S$. weberi, it differs from the other species of Sepiella in the number of sucker series (10-14) on the tentacular club ( $S$. melwardi from Australia is known only by its shell).

Sepiella ornata (from West Africa) differs from S. cyanea in that the shell is narrower, the striated zone is shorter, and the posterior part of the shell is less broad; in S. weberi (from Timor and Soemba) the posterior part of the striated area is less acuminate than in S. cyanea (Adam \& Rees 1966: 123).

## Discussion

## RELATIONSHIPS

The genus Sepia was created by Linnaeus (1758) to include all cephalopods without an external shell. Of these, the only true Sepia included in the genus was S. officinalis. The genus was restricted by Lamarck (i799: 4) to include only those cephalopods with an internal calcareous shell. At that time, the only known species were $S$. officinalis and $S$. tuberculata.

Gray (1849: 106) first used the name Sepiella for a group of shells which are 'oblong, posterior end expanded, produced, cartilaginous, not beaked, convex beneath'. Steenstrup ( $1875: 468$ ) created the genus Hemisepius, with H. typicus as the type species. This author (1880: 347) redescribed the group Sepiella and on the basis of the characters of both the shell and the soft parts raised Sepiella to generic status.

With the discovery of numerous species of Sepia in the course of time, several attempts were made to divide the genus into subgeneric groups, mainly on the basis of sucker arrangement on the sessile arms and tentacular clubs, and on the structure of the shell (d'Orbigny $1845 a$ : 26ı-298; Gray 1849: 96-1 12). Attempts have also been made to split up the genus Sepia into several genera, mostly without success. Rochebrune (1884: 74) divided the Sepiidae into ten genera, mainly on shell characters. This classification has been shown to be extremely contradictory and unnatural (Adam 1944). More recently, Iredale (1954: 81) divided the Australian Sepiidae into three families, four subfamilies and 13 genera! Criticisms of this classification are given by Adam (1964: 265) and Adam \& Rees (1966: 132). These authors have suggested a more reasonable classification,* retaining only the genera Sepia and Sepiella, which is outlined below in slightly modified form, together with the southern African representatives of each group:
I. Shell with well-developed posterior spine; ventral part of inner cone strongly developed. Tentacular suckers subequal, in 8-20 series. Sepia zanzibarica
II. Shell with well-developed posterior spine; inner cone well developed, with wide limbs, but completely reflexed on to and fused with the outer cone. Tentacular suckers unequal, usually in eight longitudinal series. Sepia officinalis vermiculata
III. Shell with posterior spine which is generally not keeled, but may be keeled dorsally, ventrally, or on both sides; inner cone more reduced, with narrow limbs; in most of these species the outer cone has two posterior wings which form, in the narrower shells, a typical cup-like expansion. Tentacular suckers nearly always arranged in eight longitudinal series.

[^12]a. Outer cone without wings; posterior spine with or without keels. Tentacular suckers minute and subequal or slightly unequal. Sepia acuminata
b. Outer cone with wings; posterior spine not keeled. Tentacular suckers subequal. No southern African representatives
c. Outer cone with wings; posterior spine without keels. Tentacular suckers unequal. In some species the ventral part of the inner cone forms a short, rounded ledge. Sepia confusa, S. incerta, S. burnupi, S. joubini and S. adami
d. Outer cone with or without wings; posterior spine keeled. Tentacular suckers unequal or subequal. Sepia australis
$e$. Outer cone with wings; posterior spine absent. Tentacular suckers subequal or unequal. No southern African representatives
IV. Shell relatively broad, with a more or less developed inner cone and without posterior wings at the outer cone; posterior spine absent. Tentacular suckers unequal or subequal. Sepia tuberculata, S. papillata, S. simoniana, S. angulata, S. hieronis and S. insignis
V. Form of shell not well known. Tentacular suckers subequal; dorsal arms with finger-like tips, devoid of suckers. Sepia robsoni and S. faurei
VI. Subgenus Hemisepius: Shell very thin, without posterior spine; phragmocone considerably shorter than dorsal shield; inner cone reduced. Sepia typica and S. dubia
VII. Subgenus Metasepia: Shell rhomboidal, much shorter than the mantle, with a completely chitinous dorsal surface. Tentacular suckers very few in number, unequal. Inner cone very narrow; posterior spine absent. No southern African representatives
VIII. Genus Sepiella: Mantle with a posterior gland and characteristic locking apparatus (Fig. 3b). Tentacular suckers subequal, in 8-32 longitudinal series. Shell with outer cone expanded, inner cone reduced; posterior spine absent. Sepiella cyanea
The degree of affinity within the different groups varies. The southern African species falling into group IIIc, the 'doratosepion' group (Sepia confusa, S. incerta, S. burnupi, S. joubini and S. adami) show close interrelationships and a marked resemblance in the characters mentioned above (p. 195) as defining Rochebrune's genus. Not all the suckers on the sessile arms are biserial, but in most of these species there is a biserial arrangement of suckers on some part of the arms in one or both sexes. These species are also remarkable in that nearly all show sexual dimorphism, such as the 'tail' in males of $S$. confusa, the modified dorsal arms in the male of $S$. incerta, and the modified dorsal and ventral arms in the male of $S$. burnupi. The males of $S$. joubini are less remarkable, being distinguished from the females (apart from the hectocotylization of the left ventral arm) mainly by the red spots on the arms. The male of $S$. adami is not known.

The southern African 'doratosepion' species fall into two orders of size. $S$. confusa and $S$. incerta attain dorsal mantle lengths of $85-90 \mathrm{~mm}$ in the females and about 150 mm in the males. $S$. burnupi, S. joubini and $S$. adami attain dorsal mantle lengths of $3^{6-59 ~ m m}$ in the females and $4^{1}-45 \mathrm{~mm}$ in the males (the male of $S$. adami is not known). All these species occur off the east coast of southern Africa.

Sepia australis (group IIId) shows a superficial resemblance to the smaller 'doratosepion' species, and particularly to $S$. joubini and S. adami. But the shell is wider, has a keeled spine and no posterior wings on the outer cone. Its distribution also differs, and this species has been assigned to the Cape faunistic province (see below).

Sepia acuminata (group III $a$ ) differs markedly from the other species of group III. The animal is generally broader and the shell is not narrow elongate but broad, almost rhomboidal, with no posterior wings on the outer cone.

The affinities between the southern African representatives of group IV (Sepia tuberculata, S. papillata, S. simoniana, S. angulata, S. hieronis and S. insignis) are much less clear than those of group III. Whilst there are similarities between some of these species, the only characters common to all are the absence of the posterior spine and wings on the outer cone of the shell, and the well developed inner cone which is completely reflexed and fused to the outer cone. The shell of $S$. papillata ( $\mathrm{Pl} .4 \mathrm{ra-d}$ ) is similar to that of $S$. tuberculata (Pls 39c, d, $4^{\circ} \mathrm{c}, \mathrm{d}$ ) on the one hand, and on the other hand to that of S. simoniana (Pl. 42a, b), from which it is sometimes almost indistinguishable, and to that of S. angulata ( $\mathrm{Pls} 44 \mathrm{~d}, 45^{\mathrm{a}-\mathrm{d}}$ ). The shells of $S$. hieronis ( $\mathrm{Pl} .43^{\mathrm{a}-\mathrm{d} \text { ) and } S \text {. insignis }}$
 consideration of the tentacular suckers, however, divides these species differently: $S$. tuberculata and $S$. papillata have unequal tentacular suckers, whereas in $S$. simoniana, $S$. hieronis and $S$. insignis they are subequal. The soft parts of $S$. angulata are not known. The distribution of these species also varies (Fig. i8, Table 8), but all occur in the Cape-South West African province.

Only two species, S. typica and S. dubia, fall into group VI, and both are southern African. These small animals show a close affinity in general appearance and in the shells. S. typica has previously been separated from the other Sepiidae in the genus Hemisepius, but the discovery of S. dubia has rendered the separation of Hemisepius from Sepia less clear. The distribution of these species, and those of group V, falls under the Cape-South West African province.

The systematic position of Sepia robsoni and S. faurei (group V) seems to be intermediate between those of groups IV and VI. Both species show a relationship with the species of group VI in general shape (animal very small, mantle very broad, with fleshy keels ventrally); other similarities between the species of groups V and VI include the similarity of hectocotylus of S. robsoni and S. typica (those of $S$. faurei and $S$. dubia not known), the biserial arrangement of the suckers on all the arms, and the similarities of the tentacular clubs, bearing
subequal suckers. The shell of $S$. robsoni, of which little is known, apparently resembles those of $S$. hieronis and $S$. insignis (group IV). The shell of $S$. faurei, on the other hand, approaches the Hemisepius-like shell: it is very thin and chitinous, and the phragmocone is considerably shorter than the dorsal lamella, but is not as short as those of $S$. typica and $S$. dubia. The shell of $S$. faurei differs from those of group VI in that the anterior margin is more nearly parallel to the corresponding margins of the dorsal lamella.

An evolutionary series leading to the Hemisepius condition can be traced as follows: Sepia robsoni (shell Sepia-like, dorsal arms with bare tips, fleshy keels on mantle ventrally, pores absent) to $S$. faurei (shell approaching Hemisepius condition, dorsal arms with bare tips, fleshy keels on mantle ventrally, pores absent) to S. dubia (Hemisepius-like shell, dorsal arms normal, fleshy keels on mantle, no pores) to S. typica (Hemisepius-like shell, dorsal arms normal, fleshy keels with pores).

It is impossible at this stage to draw any further conclusions regarding the phylogenetic relationships between the sepiids. Adam (1939a: 92) concluded that the shell and tentacular club present the best features for distinguishing the species, and Adam (1964: 268) and Adam \& Rees (1966: 135) have suggested that the following characters are probably primitive: a well-developed ventral part of the inner cone; the presence of a posterior spine; arm suckers of equal size and quadriserially arranged; subequal tentacular suckers, arranged in eight longitudinal series; and the presence of minute suckers on the buccal membrane (of the species described here, only S. zanzibarica has buccal suckers). But an arrangement of species according to shell structure does not agree with an arrangement of a series according to the structure of the tentacular club, e.g. a comparison between $S$. papillata and S. simoniana, whose shells are very similar, but whose tentacular clubs are very different.

## GEOGRAPHIGAL DISTRIBUTION

Sepiids are cephalopods inhabiting the continental shelf and slope, and in some cases the intertidal zone. Although they are capable of active swimming they apparently spend most of their time on the bottom and do not move over very wide areas or go far beyond the continental shelf (sepiids are not generally found deeper than about 500 metres*). This is borne out by the distribution of the individual species; it is found that although sepiids occur around the coasts of Europe, Africa, Asia, the Indo-Pacific islands and Australia, there are no known cosmopolitan species. On the other hand, the number of endemic species is high.

The earliest fossil record of sepiids is that of Voltzia palmeri Schevill, from Upper Jurassic deposits. Apart from this, sepiids are known almost exclusively through the Tertiary to the present. Five genera have been recorded from the Eocene (Bülow-Trummer 1920; Roger 1952): Archaeosepia, Belosepia, Pseudosepia,

[^13]Sepia and Stenosepia, but of these only Sepia has been recorded from later Tertiary deposits (Oligocene, Miocene, Pliocene) and still occurs in recent times. Thus it would appear that Voltzia became extinct in the Jurassic, and Archaeosepia, Belosepia, Pseudosepia and Stenosepia in the Eocene. All recent sepiids were presumably derived from the fossil genus Sepia.

All but three of the fossil sepiids were found in European deposits. The three exceptions are Voltzia palmeri Schevill from Cuba (Upper Jurassic), Belosepia incurvata Cossmann \& Pissaro from West Pakistan (Sind Region; Eocene) and Belosepia ungula Gabb from North America (Texas, Missouri, Alabama; Eocene).

The absence of sepiids from New Zealand and both coasts of the American continent today is apparently due to the separation of these land masses by extensive, deep oceans and/or very cold water in the regions where migration could otherwise occur. The ocean between Australia (where numerous species of Sepiidae occur) and New Zealand is wide and deep, and the currents are adverse to a crossing in this direction.

Table 8. Distributional categories of the Sepiidae of southern Africa (in the same order as in figure 18).

| Moçambique-Malagasy province | Natal province | Cape-South West African province |
| :---: | :---: | :---: |
| Sepia confusa | Sepia acuminata <br> Sepia joubini <br> Sepia burnupi <br> Sepia adami <br> Sepia incerta <br> Sepiella cyanea | Sepia faurei <br> Sepia simoniana <br> Sepia insignis <br> Sepia dubia <br> Sepia robsoni <br> Sepia tuberculata <br> Sepia typica <br> Sepia officinalis vermiculata <br> Sepia papillata <br> Sepia australis <br> Sepia hieronis <br> ? Sepia angulata |

At least two species of fossil sepiids have been recorded from America, the most recent from the Eocene. The absence of later fossil evidence suggests that these species became extinct, and that recolonization was prevented, perhaps by the low temperatures prevailing in the only relatively shallow areas via which Sepiidae could migrate from Asia or Europe, viz. the Bering Straits or via the Faeroe Islands, Iceland and Greenland.

In the waters around southern Africa, i9 species of Sepiidae are known to occur. Of these, 16 are endemic, one is tropical, one is an Atlantic species and the remaining species has an interrupted distribution.

The tropical species, Sepia confusa, enters southern African waters at the southern end of its range. It is known to occur from Zanzibar to Durban, and is


Fig. 18. Distribution ranges of the Sepiidae of southern Africa.
related to other sepiids of the 'doratosepion' group occurring off the east coast of southern Africa.

One Atlantic species, Sepia officinalis, enters southern African waters. It occurs from Scandinavia to Delagoa Bay (Moçambique), but the subspecies S. officinalis vermiculata is endemic to southern Africa, occurring from off the Groene River mouth on the west coast to Delagoa Bay on the east coast.

Sepia australis has an interrupted distribution, occurring in southern African waters from the Olifants River on the west coast to Rame Head, near Port St. Johns, on the east coast, but is also found in the Red Sea.

The endemic species are divided into two groups: the subtropical species restricted to the east coast of southern Africa, and the temperate species, occurring from the west coast round to the south coast and gradually diminishing along the east coast.

Stephenson (1948: 228), on the basis of extensive surveys of the intertidal fauna, divided southern Africa into three faunistic provinces, namely the subtropical population of Natal, the warm temperate fauna of the south coast and the cold temperate fauna of the west coast. Day (1967: in) found, however, that these divisions do not apply to shelf fauna, since the change in temperature at, say, 100 m is not as marked as it is in the intertidal zone. Thus the surface temperature of the south coast (Bashee River to Cape Point) ranges from $15^{\circ}$ to $20^{\circ} \mathrm{C}$, whereas on the west coast (Cape Point to South West Africa) the surface temperature range is about $12^{\circ}$ to $15^{\circ} \mathrm{C}$. At 100 m , however, the bottom temperature is far more constant, the range being about $12^{\circ}$ to $14^{\circ} \mathrm{C}$ from Port Elizabeth to Lüderitzbucht (Day 1967: 12).

Day (1967: 12) suggests the following faunistic provinces to include both intertidal and shelf fauna:
I. The Moçambique-Malagasy province, reaching Delagoa Bay; dominated by tropical species.
2. The Natal province, from Delagoa Bay to Bashee River; many tropical species, but also fair numbers of endemics and Atlantic species.
3. The Cape-South West African province, from Bashee River to about Cape Frio; dominated by endemics but with a few tropical species and several other components. The intertidal fauna of this province differs on the Indian and Atlantic coasts, i.e. are separated into warm temperate and cold temperate forms.
The distribution of the Sepiidae of southern Africa agrees with Day's faunistic provinces, except that the boundary between the Natal and the Cape-South West African species seems to lie a little further south, between Port Elizabeth and East London (Fig. 18). The species of Sepiidae arbitrarily assigned to the various provinces are listed in Table 8.

1. The Moçambique-Malagasy province: only one tropical sepiid species, Sepia confusa, is known to occur here. It also occurs further south, as far as Durban. Sepia hieronis has been caught in this region, but as it has also been recorded off
the west coast, this is not a tropical species but an endemic one with a peculiar interrupted distribution. Sepia acuminata, a subtropical species, enters this province at the northern end of its range.
2. The Natal province: all the species of this category are endemic, and are restricted to the east coast of southern Africa; they do not extend further south than Port Elizabeth. The main component of this group of subtropical species is the 'doratosepion' group, including Sepia incerta, S. burnupi, S. joubini and S. adami. These species are very similar anatomically and are obviously closely related. S. confusa (see above) also belongs to the 'doratosepion' group, but it is not endemic to southern Africa, as it has been recorded as far north as Zanzibar. Since, however, the waters off the east African coast have not been extensively sampled, further collection may well show that the sepiids of the Natal province occur further north than is known at present.

Two other species, Sepiella cyanea and Sepia acuminata, are found in the Natal province. The latter species extends into the Moçambique province, and has been recorded as far north as Zavora. Sepiella cyanea has also been recorded from Nosy N'Tangam, Malagasy (Adam \& Rees 1966: 121).

Several species from the Cape-South West African province extend into the Natal province (see below).
3. The Cape-South West African province: Sepiids do not seem to enter South West African waters to any marked degree. Only one Cape species, S. papillata, has been recorded from South West Africa. The apparent absence of Cape Sepiidae from South West African waters does not seem to be due to temperature, since this varies little along the west coast of southern Africa. It may however be due to the difference in coastal conditions, since the South West African shores consist largely of long sandy beaches, with pounding surf and very little kelp. Rocky outcrops are few and far apart. According to Penrith \& Penrith (1969: 100), fishes of the genus Lithognathus show a similar break in distribution off the South West African coast. L. lithognathus occurs from Natal to the Orange River mouth, but has not been recorded off South West Africa, where the species is replaced by $L$. aureti and $L$. olivieri off the northern half of South West Africa. No species of Lithognathus have been recorded off the southern half of South West Africa, between Sandwich Harbour (on the tropic of Capricorn) and the Orange River mouth.

Of the Cape species of Sepiidae, Sepia tuberculata occurs from Melkbosstrand to Knysna. The relatively short range of this species may be linked with its shallow-water habitat (depth range o-3 m).
S. papillata and S. typica have wider ranges of distribution, extending into the Natal province (to Durban). On the west coast, S. papillata has been recorded as far north as Lüderitzbucht-one of the few rocky areas on the South West African shore; S. typica does not seem to occur north of Saldanha Bay.

As mentioned above, $S$. officinalis vermiculata is endemic, and occurs in the

Cape faunistic province. It has been recorded from Groene River mouth to Delagoa Bay, the latter being the northernmost record for temperate Cape species of Sepiidae along the east coast (except perhaps S. hieronis).

Sepia simoniana is the only Cape species which does not occur off the west coast. It has been recorded from False Bay to the Tugela River mouth.

Other endemic species of the Cape province are Sepia insignis, S. robsoni, $S$. faurei and $S$. dubia, each known from only one locality, and S. angulata. The soft parts of the latter species are as yet unknown; it is not included in Figure 18. $S$. hieronis has provisionally been allocated to the Cape species, although it has not so far been recorded from the south coast; it is known to occur off the west coast (from Hondeklip Bay to Slangkop) and off Monte Belo, Moçambique.

## VERTICAL DISTRIBUTION

The vertical distributions of the southern African sepiids are not well known. Many records give no indication of the depths at which the specimens were caught, and others, where open trawls were used, are unreliable. In the latter case the depth at which the trawl was hauled is given, but the specimens could as easily have been caught while the trawl was raised or lowered. However, since sepiids tend to live on or near the bottom, the depth over which the trawl was fished probably gives a fairly good indication of the depth at which the sepiids were living. The depth records available (Fig. 19) show some interesting features.

Off the south coast, where the continental shelf is very wide (about 220 km at the widest point), few sepiids have been recorded below 100 m . Off the east and west coasts the continental shelf is much narrower (maximum about $40-50 \mathrm{~km}$ ) and sepiids have been recorded as deep as 460 m , that is, some way out beyond the edge of the shelf.

Sepia officinalis vermiculata is unusual in that it is found in shallow water in estuaries and sheltered bays (Saldanha Bay, Breede River mouth, Knysna lagoon, Bushmans River mouth and Durban Bay). Strangely it has not so far been found in False Bay. This subspecies is not limited to shallow water, however, and has also been recorded from depths to 249 m off the Natal coast (Massy 1925: 209).

Sepia tuberculata is apparently more closely restricted to the inshore waters, and has been collected mainly from rock pools. There is also one record from Simonstown harbour, at a depth of 3 m . Many of the records do not give any reference to depth, and this species is not included in Figure 19.

Sepia typica is common in fairly shallow water in Saldanha Bay, but elsewhere it is found somewhat deeper. In Table Bay and False Bay this species is found below ${ }^{1} 7-18 \mathrm{~m}$, off the south coast below 40 m , and off Durban at 99 m . Similarly the upper depth limit of $S$. simoniana is deeper off the east coast ( $116-134 \mathrm{~m}$ ) than off the south coast (below about 10 m in False Bay). S. papillata and S. australis show a similar though less marked trend at the eastern end of their distribution ranges.


Fig. 19. Vertical distribution of the Sepiidae of southern Africa. Linked symbols indicate continuous hauls between the indicated depths.

The east coast sepiids also show a tendency to occupy deeper water further northwards, as the climate becomes hotter, though the absence of records from shallow water off the Moçambique coast may well be due to insufficient sampling. The east coast species are generally caught below 60 m off the Natal coast and below 200 m off the Moçambique coast. Two exceptions are $S$. burnupi, caught at $40-48 \mathrm{~m}$ off the Umhlanga River, and Sepiella cyanea, recorded from $5^{1}$ and 73 m off the Tugela and Umvoti Rivers. For both these species, these are the only available depth records.

## GROWTH

For each species the relative dimensions were calculated for as many specimens as possible, and the ranges and means were calculated for those dimensions thought to be of some significance. Tests for correlation between various relative dimensions and the standard length (MLd or shell L) were performed for species where ten or more specimens had been measured (males and females being considered separately), to test if there is any change in relative body (or shell) dimensions with growth. In general the results of the correlation tests were disappointing, due to the great variation in measurements.

The only species giving significant correlations for most of the body measurements was Sepia australis, and even here the scatter about the regression lines is wide, though 59 males and 77 females were measured. Figure 20 shows the scatter diagrams for most of the dimensions of $S$. australis males and shells. Where correlation was found to be statistically significant, the regression lines were calculated by the least squares method. These lines are included in Figure 20 for interest, but they clearly do not fit the scatter diagrams well, since the variation is very wide. Trends can, however, be observed. With growth, the mantle becomes much narrower relative to its length; the head becomes relatively shorter and narrower, and the fins also become relatively narrower. The change in relative length of the tentacular club is small, but the arms become relatively longer. In fact there seems to be a general trend to elongation with growth, producing a more streamlined body, perhaps enabling faster locomotion.

Growth of the $S$. australis females is very similar to that of the males, except that the arms are relatively shorter than those of the males, although also becoming longer in larger animals.

The shell dimensions also show a wide scatter, despite the fact that the shell is a rigid structure and is not subject of contractility as are the soft parts. The shell is however secreted by the animal and the variation in its dimensions suggests that the wide scatter of the relative dimensions of the soft parts is not solely due to their contractility. The shell becomes relatively narrower and slightly thinner with growth, and the striated zone becomes relatively longer.





Fig. 20. Sepia australis males and shells. Change in dimensions (expressed as \% MLd or \% shell length) with growth (increase in MLd or shell length). The straight lines through the scatter diagrams indicate the calculated regression lines, where a statistically significant correlation was found to obtain.

## Summary

A complete synonymy, distribution and depth ranges, and descriptions of external morphology are given for each of 20 species. Three species, Sepia adami, S. angulata and $S$. faurei, are new. The soft parts of $S$. insignis (previously known only by its shell) are described for the first time. The animals and shells described by Massy (1925) as $S$. incerta and $S$. burnupi have been re-examined, and all are found to pertain to $S$. incerta. The first known animals of $S$. burnupi (formerly the types of $S$. exsignata) have been redescribed, and the first recorded specimens of $S$. hieronis from the east coast are described. Keys to the soft parts and to the shells are provided. The relationships of the southern African Sepiidae, their geographical and vertical distribution, and their growth, are discussed.

## Acrnowledgements

I am grateful to Professor W. Adam of the Institut Royal des Sciences Naturelles de Belgique, Dr. D. van Z. Engelbrecht of the University of Stellenbosch, Dr. N. A. H. Millard of the South African Museum, Dr. P. B. Best of the Division of Sea Fisheries, Cape Town and Dr. M.-L. Penrith, formerly of the South African Museum, for valuable advice on the manuscript. Also to Mr. S. X. Kannemeyer, for the photography.

For the donation and/or loan of specimens, I should like to thank Mr. J. Bass of the Oceanographic Research Institute, Durban, Dr. J. A. Pringle of the Natal Museum, Dr. J. D. Taylor of the British Museum (Natural History), Dr. J. G. Field of the University of Cape Town, and the Division of Sea Fisheries. I would also like to thank the numerous individuals who have brought me sepiid shells.

## References

Adam, W. 1939a. Cephalopoda. II-Révision des espèces Indo-Malaises du genre Sepia Linn 1758. Siboga Exped. Monogr. 55b: 35-92.

Adam, W. 1939b. Cephalopoda. III-Révision du genre Sepiella (Gray) Steenstrup, 1880. Siboga Exped. Monogr. 55b: 93-122.
Adam, W. 1940. Les races de la seiche commune (Sepia officinalis Linné). Bull. Soc. zool. Fr. 65: 125-131.
Адам, W. 1941. Résultats scientifiques des croisières du navire-école belge 'Mercator'. III (4). Cephalopoda. Mém. Mus. r. Hist. nat. Belg. (2) 21: 83-161.
Adam, W. 1942. Les céphalopodes de la mer Rouge. Bull. Inst. océanogr. Monaco 822: 1-20.
Adam, W. 1944. Révision de l' 'Étude monographique de la famille des Sepiadae' d'A. T. de Rochebrune (1884). Mém. Mus. natn. Hist. nat., Paris (n.s.) 18: 219-242.
Adam, W. 1959. Les céphalopodes de la mer Rouge. In Mission Robert Ph. Dollfus en Egypte (Décembre 1927-Mars 1929). S.S. 'Al Sayad'. Résultats scientifiques 3 (28): 125-193. Paris: Centre National de la Recherche Scientifique.
Adam, W. 1962. Céphalopodes de l'Archipel du Cap-Vert, de l'Angola et du Mozambique. Mems fta Invest. Ultramar (2) 33: 7-64.
Адам, W. 1964. Considérations sur la systématique des Sepiidae (Cephalopoda). Zoöl. Meded., Leiden 39: 263-278.

Adam, W. \& Rees, W. J. 1966. A review of the cephalopod family Sepiidae. Scient. Rep. John Murray Exped. 11 (1): $\mathbf{1}-165$.
Barnard, K. H. 1962. New species and records of South African marine Mollusca from Natal, Zululand and Moçambique. Ann. Natal Mus. 15: 247-254.
Bartsch, P. 1915. Report on the Turton collection of South African marine mollusks, with additional notes on other South African shells contained in the United States National Museum. Bull. U.S. natn. Mus. 9x : 1-305.
Blainville, H. M. D. de. 1825. Manuel de malacologie et conchologie. Paris: Levrault.
Blainville, H. M. D. de. 1827 a. In cuvier, f., ed. Dictionnaire des sciences naturelles. Paris.
Blainville, H. M. D. de. 1827 b. Atlas to Manuel de malacologie et conchologie. Paris: Levrault.
Bosc, L. A. G. 1802. Histoire naturelle des vers. 1. In buffon, g. L. L. de. Histoire naturelle de Buffon. Nouv. éd. (Suite). Paris.
Bülow-Trummer, E. v. 1920. Cephalopoda dibranchiata. Fossilium Cat. ( 1 : Animalia) II: 1-313.
Carleton, H. M. \& Robson, G. C. 1924. On the histology and function of certain secondary sexual organs in the cuttlefish Doratosepion confusa. Proc. R. Soc. (B) 96: 259-271.
Chun, C. 1915. Die Cephalopoden. II. Teil: Myopsida, Octopoda. Wiss. Ergebn. dt. TiefseeExped. 'Valdivia' 18: 405-552.
Day, J. H. 1967. A monograph on the Polychaeta of southern Africa. Part I. Errantia. London: British Museum (Natural History).
Deshayes, G. P. 1830-32. Histoire naturelle des vers. Par M. Brugière (continuée par M. G. P. Deshayes). 2-3. In Encyclopédie méthodique. Paris.
Férussac, A. de \& Orbigny, A. d'. 1835-48. Histoire naturelle générale et particulière des céphalopodes acétabulifères vivants et fossiles. Paris: Bertrand.
Gibbons, J. S. 1888. Partial list of the South African Mollusca. Trans. S. Afr. phil. Soc. 4: 201-219.
Gray, J. E. 1849. Catalogue of the Mollusca in the collection of the British Museum. 1. Cephalopoda Antepedia. London: British Museum (Natural History).
Hoyle, W. E. 1885 . Preliminary report on the Cephalopoda collected during the cruise of H.M.S. 'Challenger'. 2. The Decapoda. Proc. R. phys. Soc. Edinb. 13: 281-310.

Hoyle, W. E. 1886. Report on the Cephalopoda collected by H.M.S. Challenger during the years 1873-1876. Rep. Voy. Challenger 1873-76 16: 1-245.
Hoyle, W. E. 1904. Sepia burnupi n. sp. from Natal. 7. Conch., Lond. 11: 27-28.
Hoyle, W. E. 1909. A catalogue of recent Cephalopoda. Second supplement, 1897-1906. Proc. R. phys. Soc. Edinb. 17: 254-299.
Hoyle, W. E. 1910. Mollusca: Cephalopoda. In schultze, l. Zoologische und anthropologische Ergebnisse einer Forschungsreise im westlichen und zentralen Südafrika 4: 261-268. Jena: Fischer. Denkschr. med-naturw. Ges. Jena 16: 261-268.
Hoyle, W. E. 1912. The Cephalopoda of the Scottish National Antarctic Expedition. Trans. R. Soc. Edinb. 48: 273-283.

Iredale, T. 1954. Cuttle-fish 'bones' again. Aust. Zool. 12: 63-82.
Lamarck, J. B. i 798 . Extrait d'un mémoire sur le genre de la sèche, du calmar et du poulpe, vulgairement nommés polypes de mer. Bull. Soc. philomath. Paris 2: 129-131.
Lamarck, J. B. i 799 . Sur les genres de la sèche, du calmar et du poulpe, vulgairement nommés polypes de mer. Mém. Soc. Hist. nat. Paris 1: 1-24.
Lamarck, J. B. 1822. Histoire naturelle des animaux sans vertèbres. Ist ed. 7. Paris: Baillière.
Lamarck, J. B. 1835-45. Histoire naturelle des animaux sans vertèbres. 2nd ed. 11. Paris.
Leach. [Unpublished manuscript, referred to in Férussac \& d'Orbigny, 1835-48.]
Linnaeus, C. 1758. Systema naturae. roth ed. r. Holmiae: Laurentii Salvii.
Massy, A. L. 1925. On the Cephalopoda of the Natal Museum. Ann. Natal Mus. 5: 201-229.
Massy, A. L. 1927. The Cephalopoda of the South African Museum. Ann. S. Afr. Mus. 25: 151-167.
Massy, A. L. 1928. On the Cephalopoda of the Natal Museum. Part II. Ann. Natal Mus. 6: 89-96.
Massy, A. L. \& Robson, G. C. 1923. On a remarkable case of sex-dimorphism in the genus Sepia. Ann. Mag. nat. Hist. (9) 12: 435-442.
Montrort, P. D. de. i805. Histoire naturelle des mollusques. In buffon, g. l. l. de. Histoire naturelle. Nouv. éd. (suite). Paris.
Odhner, N. Hj. 1923. Contribution to the marine molluscan faunas of South and West Africa. Göteborgs K. Vetensk.-o. VitterhSamh. Handl. (4) 26 (7): 1-40.

Orbigny, A. D. d'. i826. Tableau méthodique de la classe des céphalopodes. Annls Sci. nat. 7: 96-169.
Orbigny, A. D. d'. 1845[-47]a. Mollusques vivants et fossiles. 1: $1-605$, with Atlas of 36 plates. Paris.
Orbigny, A. D. d'. 1845[-47]b. Paléontologie universelle des coquilles et des mollusques: 1-392, plates II-104. Paris.
Orbigny, A. D. d'. $1845[-47]$ c. Paléontologie des coquilles et des mollusques étrangères à la France. Plates 9-60. Paris.
Penrith, M. J. \& Penrith, M.-L. i969. A new species of Lithognathus (Pisces: Sparidae) from the northern coast of South West Africa. Cimbebasia (A) 1: 99-1 I 1 .
Pfeffer, G. 1884. Die Cephalopoden des Hamburger Naturhistorischen Museums. Abh. naturw. Ver. Hamburg 8: 63-90.
Quoy, J. R. \& Gaimard, J. P. ı832. Mollusques. In Zoologie du voyage de l'Astrolabe, pendant les années 1826-1829. 2. Paris: Tastu.
Robson, G. C. 1924a. Preliminary report on the Cephalopoda (Decapoda) procured by the S. S. "Pickle". Rep. Fish. mar. biol. Surv. Un. S. Afr. 3 (Spec. Rep. 9): 1-14.

Robson, G. C. $1924 b$. On the Cephalopoda obtained in South African waters by Dr. J. D. F. Gilchrist in 1920-21. Proc. zool. Soc. Lond. 1924: 589-686.
Rochebrune, A. T. de. 1884. Étude monographique de la famille des Sepiadae. Bull. Soc. philomath. Paris (7) 8: 74-122.
Roger, J. 1952. Sous-classe des Dibranchiata Owen 1836 (Coleoidea Waagen, Endocochlia Schwartz). In piveteau, J., ed. Traité de Paléontologie. 2: 689-755. Paris: Masson.
Smith, E. A. 1903. A list of species of Mollusca from South Africa, forming an appendix to G. B. Sowerby's 'Marine shells of South Africa". Proc. malac. Soc. Lond. 5: 354-402.

Smith, E. A. 1916. On the shells of the South African species of Sepiidae. Proc. malac. Soc. Lond. 12: 20-26.
Steenstrup, J. 1875. Hemisepius, en ny slaegt af Sepia-blaeksprutternes familie med bemaerkninger om Sepia-formerne i almindelighed. K. danske Vidensk. Selsk. Skr. (5) 10: 465-482.
Steenstrup, J. i88o. Sepiella Gray. Stp. Vidensk. Meddr dansk naturh. Foren. 1879/80: 347-356.
Stephenson, T. A. 1948. The constitution of the intertidal fauna and flora of South Africa. Part III. Ann. Natal Mus. 11: 207-324.
Thiele, J. 1920. Die Cephalopoden der deutschen Südpolar-Expedition 1901-1905. Dt.Südpol.Exped. 16: 433-465.
Thore, S. 1945. On the Cephalopoda of Prof. O. Carlgren's expedition to South Africa in 1935. K. fysiogr. Sällsk. Lund Förh. 15: 49-57.

Tomlin, J. R. le B. 1923. On South African marine Mollusca with descriptions of several new species. 7. Conch., Lond. 17: 40-52.
Tomlin, J. R. le B. 1926. On the South African marine Mollusca, with descriptions of new species. Ann. Natal Mus. 5: 283-301.
Tryon, G. W. 1879. Cephalopoda. Man. Conch. 1: 1-316.
Turton, W. H. 1932. The marine shells of Port Alfred, S. Africa. London: Oxford University Press.
Voss, G. L. 1962a. List of the types and species of cephalopods in the collections of the Academy of Natural Sciences of Philadelphia. Notul. Nat. 356: 1-7.
Voss, G. L. 1962b. South African cephalopods. Trans. R. Soc. S. Afr. 36: 245-272.
Voss, G. L. 1967. Some bathypelagic cephalopods from South African waters. Ann. S. Afr. Mus. 50: 6ı-88.

## APPENDIX

Table 9. Sepia zanzibarica. Relative dimensions as \% MLd.


Table 10. Sepia zanzibarica shells.
Relative dimensions as \% shell length.

Table in. Sepia officinalis vermiculata males. Relative dimensions as \% MLd.

|  |  | A30130 | A30182 | Knysna Estuary |  |  |  |  |  |  |  | Durban | Adam | $\begin{aligned} & \text { Adam } \\ & \& \\ & \text { Rees } \end{aligned}$ | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm |  | 220 | 198 | 126 | 111 | 111 | 111 | 102 | 94 | 80 | 67 | 136 | 124 | 148 |  |  |  |
| MLv | . . | 90,9 | 87,4 | 90,5 | 91,0 | 89,2 | 89,2 | 92,2 | 91,5 | 90,0 | 92,5 | 89,7 | 87 | 78 | 13 | 89,2 | 78 -92,5 |
| MW | . . | 50,9 | 38,4 | 52,4 | 51,4 | 49,5 | 50,5 | 48,o | 53,2 | 53,8 | 61,2 | 57,4 | 51 | 49 | 13 | 51,3 | 38,4-61,2 |
| HL | - . | 26,8 | 22,7 | 26,2 | 28,8 | 27,0 | 27,9 | 22,5 | 24,5 | 27,5 | 26,9 | 33,1 | 28 | 26 | 13 | 26,8 | 22,5-33,1 |
| HW | - . | 40,0 | 36,9 | 41,3 | 45,0 | 41,4 | 41,4 | 41,2 | 42,6 | 50,0 | 52,2 | 46,3 | 40 | 41 | 13 | 43,0 | 36,9-52,2 |
| FL. | - . | 106,8 | 106, 1 | 107,1 | 109,9 | 103, 6 | 101,8 | 105,9 | 111,7 | 103,8 | 113,4 | 110,3 | 96 | 100 | 13 | 105,9 | $96-113,4$ |
| FW | . . | ${ }^{1} 5,9$ | 15,7 | 15,9 | 16,2 | 13,5 | 11,7 | 16,7 | 18,1 | 13,8 | 17,9 | 14,7 | 13 | 15 | 13 | 15,2 | 11,7-18,1 |
| AL I | . | 36,4 | 38,9 | 31,7 | 33,3 | 32,4 | 28,8 | 37,3 | 38,3 | 37,5 | 40,3 | 46,3 | 44 | 44 | 13 | 37,6 | 28,8-46,3 |
| AL II |  | 34,5 | 39,9 | 34, 1 | 36,9 | 34,2 | 35,1 | 36,3 | 39,4 | 40,0 | 38,8 | 49,3 | 47 | 47 | 13 | 39,4 | 34, r-49,3 |
| AL III |  | 38,6 |  | 35,7 | 39,6 | 36,9 | 40,5 | 41,2 | 41,5 | 36,3 | 41,8 | 52,9 | 50 | 54 | 12 | 42,4 | 35,7-54 |
| AL IV |  | - | 54,0 | 43,7 | 43,2 | 40,5 | 40,5 | 45, 1 | 47,9 | 53,8 | 49,3 | 63,2 | 53 | 57 | 12 | 49,3 | 40,5-63,2 |
| TL ${ }^{\text {Rt. }}$ |  | 95,5 | 106, 1 |  |  |  | - | - |  |  |  | ${ }^{11} 5,4$ | 161 |  | 8 |  |  |
| ${ }^{\text {LL }} \mathrm{Lt}$. |  | 93,2 | III, 1 | - | - | - |  | - |  | - | - | 117,6 | 153 | - | 8 | 119,1 | 93,2-161 |
| Tcl |  | 29,5 | 27,8 | 23,8 | 30,6 | 26,1 | 24,3 | 28,4 | 26,6 | 35,0 | 34,3 | 29,4 | 28 | 34 | 13 | 29, I | 23,8-35,0 |

Table 12. Sepia officinalis vermiculata females and juveniles. Relative dimensions as $\%$ MLd.

|  | A30131 | A30183 | A31292 | Locality unknown | Knysna Estuary |  |  |  |  |  | Durban Bay |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm . | 208 | 287 | 188 | 242 | 103 | 105 | 100 | 102 | 98 | 96 | 156 | 172 |
| MLv . | 88,5 | 87,5 | 88,8 | 86,4 | 99,0 | 93,3 | 90,0 | 87,3 | 92,9 | 89,6 | 91,7 | 92,4 |
| MW . | 43,8 | 50,9 | 44,7 | 55,8 | 63,1 | 50,5 | 50,0 | 56,3 | 53,1 | 54,2 | 59,6 | 59,9 |
| HL | , | 24,4 | 23,9 | 34,3 | 25,2 | 20,0 | 27,0 | 25,5 | 29,6 | 26,0 | 33,3 | 33,1 |
| HW | - | 37,6 | 38,8 | 39,3 | 47,6 | 43,8 | 42,0 | 44,1 | 44,9 | 47,9 | 48,7 | 50,0 |
| FL | 101,0 | 107,3 | 109,0 | 110,7 | 119,4 | 106,7 | 104,0 | - | 105,1 | 110,4 | 110,3 | 113,4 |
| FW | 12,0 | 12,2 | 16,0 | 16,5 | 19,4 | 12,4 | 16,0 | $\bar{\square}$ | 12,2 | 17,7 | 13,5 | 16,3 |
| ALI | 31,7 | 29,6 | 27,7 | 55,0 | 40,8 | 34,3 | 32,0 | 42,2 | 40,8 | 35,4 | 40,4 | 42,4 |
| ALII | 33,7 | 32,8 | 29,8 | 57,9 | 40,8 | 34,3 | 31,0 | 43,1 | 40,8 | 38,5 | 50,0 | 47,7 |
| AL III | 33,7 | 36,6 | 31,9 | 60,3 | 42,7 | 36,2 | 36,0 | 44,1 | 38,8 | 42,7 | 44,2 | 48,8 |
| AL IV | - | 44,3 | 37,2 | 79,3 | 53,4 | 44,8 | 42,0 | 52,0 | 52,0 | 51,0 | 56,4 | 50,0 |
| TL Rt | 101,0 | 158,5 | 85,1 | 203,3 | - | - |  |  |  |  | 105,8 | 116,3 |
| 1 L L | 117,8 | 163,1 | 74,5 | 159,1 | - | 257 | - | - | 34.7 | 30 | 115,4 | 104,7 |
| Tcl. | 26,4 | 27,9 | 26,6 | 36,0 | 29,1 | 25,7 | 31,0 | 38,2 | 34,7 | 30,2 | 31,4 | 32,0 |


|  |  | Durban Bay (cont.) |  |  |  | Adam 1962 | Adam \& Rees 1966 | $\mathrm{N}_{0}$ | Mean | Range ${ }_{\text {O }}$ | A30128 juvenile | A30129 juvenile |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm |  | 162 | 149 | 172 | 168 | 147 | 120 |  |  |  | 50 | 39 |
| MLv | . | 91,4 | 89,9 | 92,4 | 89,9 | 90 | 85 | 18 | 90,3 | $85-99,0$ | 90,0 | 87,2 |
| MW | . | 58,0 | 62,4 | 56,4 | 58,3 | 52 | 54 | 18 | 54,6 | 43,8-63,1 | 52,0 | 61,5 |
| HL . | - | 31,5 | 32,9 | 29,7 | 32,1 | 31 | 33 | 17 | 29,0 | 20,0-34,3 | 28,0 | 30,8 |
| HW | - | 49,4 | 48,3 | 34,9 | 36,9 | 46 | 42 | 17 | 43,7 | 34,9-50,0 | 48,0 | 48,7 |
| FL | . | 113,6 | 117,4 | 104,7 | 114,3 | 100 | 98 | 17 | 108,5 | 98-119,4 | 106,0 | 97,4 |
| FW | - | 14,8 | 15,4 | 17,4 | 14,9 | 10 | 13,5 | 17 | 14,7 | $10-19,4$ | 12,0 | 10,3 |
| AL I |  | 38,9 | 41,6 | 39,5 | 44,0 | 42 | 33 | 18 | 38,4 | 27,7-55,0 | 22,0 | 23,1 |
| AL II | . | 46,3 | 52,3 | 46,5 | 49,4 | - | 33 | 17 | 41,6 | 29,8-57,9 | 28,0 | 30,8 |
| AL III | - | 48,1 | 49,7 | 47,1 | 47,2 | 46 | 33 | 17 | 43,1 | 31,9-60,3 | 26,0 | 30,8 |
| AL IV | . | 56,8 | 59,7 | 54,7 | 58,3 | 58 | 46 | 17 | 52,7 | 37,2-79,3 | 30,0 | 38,5 |
| TL Rt | - | 104,9 | 117,4 | 173,3 | 157,7 | - | 150 | 21 | 129,3 |  | 100,0 | 117,9 |
| $\mathrm{Tcl}^{\text {Lt }}$ | . | 112,3 | 107,4 | 148,8 | 138,1 | - | 150 | 17 | 129,3 | 74,5-203,3 | 100,0 | 125,6 |
| Tcl | - | 28,4 | 28,9 | 29,1 | 33,3 | - | 32 | 17 | 30,6 | 25,7-38,2 | 20,0 | 20,5 |

Table 13. Sepia officinalis vermiculata shells. Relative dimensions as \% shell length.

|  | $\stackrel{\mathrm{L}}{\text { in } \mathrm{mm}}$ | W | Th | Str z |
| :---: | :---: | :---: | :---: | :---: |
| A30129 | 37 | 45,9 | 8, I | 43,2 |
| A30130 | 214 | 36,9 | 13, 1 | 49,5 |
| A30182 | 193 | 33,2 | 10,9 | 63,2 |
| A30183 | 278 | 38,5 | 12,6 | 69, I |
| Breede River | 45 | 44,4 | 10,0 | 46,7 |
| Knysna | 120 | 38,3 | 11,7 | 50,0 |
| Estuary | 95 | 37,9 | 12,6 | 49,5 |
|  | 126 | 38,1 | 12,3 | 47,6 |
|  | 98 | 38,8 | 11,2 | 44,9 |
|  | 94 | 39,4 | 11,7 | 47,9 |
|  | 92 | 39, 1 | II,4 | 47,8 |
|  | 56 | 41,1 | 11,6 | 50,0 |
|  | 127 | 33, 1 | 9,8 | 43,3 |
|  | 125 | 32,8 | 10,0 | 41,6 |
|  | 102 | 41,2 | 12,7 | 47, 1 |
|  | 97 | 38,1 | 11,3 | 46,4 |
|  | 61 | 41,0 | 13,1 | 52,5 |
|  | 112 | 40,2 | 12,5 | 47,3 |
|  | 110 | 39,1 | 11,7 | 47,3 |
|  | 109 | 38,5 | 11,5 | 48,6 |
|  | 105 | 39,0 | 12,4 | 48,6 |
|  | 105 | 41,0 | 12,9 | 48,6 |
|  | 101 | 41,6 | 12,9 | 46,5 |
|  | 103 | 37,9 | 11,7 | 50,5 |
|  | 102 | 39,2 | 12,7 | 48,0 |
|  | 100 | 40,0 | 12,5 | 46,0 |
|  |  | 40,0 | 12, I | 50,5 |
|  | 96 | 39,6 | 11,5 | 49,0 |
|  | 96 | 40,6 | 12,5 | 47,9 |
|  | 93 | 40,9 | 11,8 | 47,3 |
|  | 89 | 40,4 | 12,9 | 50,6 |
|  | 91 | 38,5 | 11,5 | 49,5 |
|  | 85 | 38,8 | 12,4 | 48,2 |


|  | $\underset{\text { in } \mathrm{mm}}{\mathrm{~L}}$ | W | Th | Str z |
| :---: | :---: | :---: | :---: | :---: |
| Knysna <br> Estuary (cont.) | 86 | 38,4 | 11,0 | 51,2 |
|  | 81 | 39,5 | 11,7 | 53, 1 |
|  | 79 | 40,5 | 12,7 | 49,4 |
|  | 77 | 40,3 | 12,3 | 49,4 |
|  | 78 | 39,7 | 12,2 | 50,0 |
|  | 74 | 40,5 | 12,2 | 48,6 |
|  | 72 | 40,3 | 12,5 | 48,6 |
|  | 69 | 40,6 | 1 1,6 | 49,3 |
|  | 68 | 42,6 | 13,2 | 52,9 |
|  | 63 | 41,3 | 12,7 | 47,6 |
|  | 63 | 41,3 | 11,9 | 52,4 |
|  | 62 | 41,9 | 12,9 | 51,6 |
|  | 63 | 41,3 | 1 1,9 | 49,2 |
|  | 58 | 39,7 | 12, 1 | 51,7 |
|  | 56 | 41, I | 12,5 | 50,0 |
|  | 54 | 42,6 | 13,0 | 53,7 |
|  | 50 | 42,0 | 12,0 | 50,0 |
|  | 45 | 42,2 | 10,0 | 48,9 |
| Saldanha | 139 | 41,7 | 10, 1 | 74,8 |
| Bay | 165 | 34,5 | 9,1 | 69, I |
| Krom River | I 57 | 39,5 | 14,0 | 47,8 |
| Durban <br> Bay | 148 | 39,2 | 14,2 | 40,5 |
|  | 164 | 39,0 | 15,2 | 41,5 |
|  | 157 | 39,5 | 14,0 | 48,4 |
|  | 138 | - | 15,2 | 43,5 |
|  | 127 | 40,2 | 15,0 | 45,7 |
|  | 169 | 39,6 | 14,2 | 49, I |
|  | 168 | 40,5 | 13,7 | 51,2 |
| N |  | 60 | 6I | 61 |
| Mean |  | 39,7 | 12,2 | 49,7 |
| Range |  | $\begin{aligned} & 32,8- \\ & 45,9 \end{aligned}$ | $\begin{gathered} 8,1- \\ 15,2 \end{gathered}$ | $\begin{aligned} & 40,5- \\ & 74,8 \end{aligned}$ |

Table 14. Sepia acuminata males. Relative dimensions as \% MLd.

|  | A30147 | A31398 | A31399 | N.M. 964 | Massy <br> 1928 | Adam \& Rees 1966 | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd | 89 | 67 | 89 | 78 | 80 | 67 |  |  |  |
| MLv | 83, 1 | 8o,6 | 80,9 | 82,1 | 80,0 | 78 | 6 | 8o,8 | $78-83,1$ |
| MW | 44,9 | 68,7 | 60,7 | 64,1 |  | 49 | 5 | 57,5 | 44,9-68,7 |
| HL | 27,0 | 34,3 | 36,0 | 37,2 |  | 25 | 5 | 31,9 | $25-37,2$ |
| HW | 40,4 | 52,2 | 48,3 | 42,3 | 41,3 | 36 | 6 | 43,4 | $36-52,2$ |
| FL | 95,5 | 86,6 | 82,0 | 94,9 |  | 87 | 5 | 89,2 | 82,0-95,5 |
| FW | 11,2 | 6,0 | 7,9 | 9,0 |  | 9 | 5 | 8,6 | 6,0-11,2 |
| AL I | 25,8 | 38,8 | 39,3 | 33,3 | 42,5 | 25 | 6 | 34, 1 | $25-42,5$ |
| AL II | 24,7 | 35,8 | 36,0 | 29,5 | 41,3 | 27 | 6 | 32,4 | 24,7-41,3 |
| AL III | 24,7 | 38,8 | 42,7 | 35,9 | 40,0 | 28 | 6 | 35,0 | 24,7-42,7 |
| AL IV | 27,0 | 50,7 | 44,9 | 43,6 | 51,3 | 33 | 6 | 41,8 | 27,0-51,3 |
| $\mathrm{TL}_{\text {Rt }}^{\text {Rt }}$ | 44,9 60,7 | 182,1 | - | 135,9 | 145,0 | 105 | 6 | 112,3 | 44,9-182, 1 |
| Tcl | 11,2 | 22,4 | - | I 5,4 | 15,0 | 13 | 5 | 15,4 | 11,2-22,4 |

Table I5. Sepia acuminata females and juveniles. Relative dimensions as \% MLd.

|  | $\begin{gathered} \mathrm{A}_{3} 1398 \\ \circ \\ \text { ㅇ } \end{gathered}$ | $\begin{gathered} \text { N.M. } 964 \\ \text { 아 } \end{gathered}$ | $\begin{gathered} \text { Adam \& } \\ \text { Rees } \\ \text { 1966 } \\ \dot{q} \end{gathered}$ | $\mathrm{N}_{\text {우 }}$ | Mean 우 | Range ㅇ | $\mathrm{A}_{3} 1400$ juv. |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 92 | 92 | 79 |  |  |  | 39 |
| MLv . . | 73,9 | 82,6 | 83 | 3 | 79,8 | 73,9-83 | 87,2 |
| MW | 65,2 | 54,3 | 53 | 3 | 57,5 | $53-65,2$ | 59,0 |
| HL . | 38,0 | 31,5 | 29 | 3 | 32,8 | $29-38,0$ | 38,5 |
| HW | 51,1 | 40,2 | 40 | 3 | 43,8 | $40-51,1$ | 48,7 |
| FL | 76, 1 | 90,2 | 89 | 3 | 85, 1 | 76,1-90,2 | 82,1 |
| FW . | 6,5 | 12,0 | 11 | 3 | 9,8 | 6,5-12,0 | 7,7 |
| AL I | 40,2 | 29,3 | 32 | 3 | 33,8 | 29,3-40,2 | 35,9 |
| AL II | 44,6 | 38,0 | 32 | 3 | 38,2 | $32-44,6$ | 35,9 |
| AL III | 40,2 | 37,0 | 32 | 3 | 36,4 | $32-40,2$ | 33,3 |
| AL IV | 50,0 | 41,3 | 35 | 3 | 42, I | $35-50,0$ | 48,7 |
| TL Rt . | - | 137,0 | 95 | 4 |  | 95-155,4 | 202,6 |
| ${ }^{\text {L }}$ Lt . | 155,4 | 105,4 | 95 | 4 | 123,2 | $95-155,4$ | 202,6 |
| Tcl. | 22,8 | 18,5 | 13 | 3 | 18, 1 | $13-22,8$ | 23, I |

Table 16. Sepia acuminata shells. Relative dimensions as \% shell length.

|  | A31398 | A31399 | A31400 | West Pondoland | N.M. 965 |  | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| L in mm | 72 | 92 | $\pm 39$ | $\pm 108$ | 101 | $\pm 77$ |  |  |  |
| W . | 38,9 | 38,0 | $\pm 48,7$ | $\pm 43,5$ | $\pm 35,6$ | $\pm 39$ | 6 | 40,6 | 35,6-48,7 |
| Th. | 9,7 | 9,2 | $\pm 10,3$ | $\pm 10,2$ |  | $\pm 10,4$ | 6 | 10,0 | 9,2-10,4 |
| Str z | 59,7 | 68,5 | $\pm 64, \mathrm{I}$ | $\pm 64,8$ | 64,4 | $\pm 6 \mathrm{I}$ | 6 | 63,8 | 59,7-68,5 |

## See opposite for Table 17

Table 18. Sepia confusa females. Relative dimensions as \% MLd.

|  | $\mathrm{A}_{3} 1403$ | Adam \& Rees 1966 |  |  |  | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | $\pm 53$ | 77 | 85 | 80 | 84 |  |  |  |
| MLv | 73,6 | 80 | 79 | 75 | 8ı |  |  |  |
| MW | 49, I | 36 | 33 | 35 | 33 | 5 | 37,2 | $33-49,1$ |
| HL | 26,4 | 22 | 23 | 19 | 18 |  |  |  |
| HW | 32,1 | 31 | 33 | 30 | 33 | 5 | 31,8 | 30-33 |
| FL . | 79,2 | 87 | 79 | 81 | 80 |  |  |  |
| FW | 5,7 | 13 | 13 | 12,5 | 8,5 | 5 | 10,5 | 5,7-13 |
| AL I | 45,3 | 38 | 33 | 34 | 30 | 5 | 36, I | 30-45,3 |
| AL II | 45,3 | 34 | 32 | 35 | 32 | 5 | 35,7 | $32-45,3$ |
| AL III | 49, I | 34 | 32 | 32 | 32 | 5 | 35,8 | $32-49,1$ |
| AL IV | 50,9 | 36 | 33 | 35 | 34 | 5 | 37,8 | $33-50,9$ |
| TL | - |  | 82 |  | - |  |  |  |
| Tcl | - | 13,5 | 14 | - | - | 2 | 13,8 | ${ }^{1} 3,5{ }^{-1} 4$ |

Table 19. Sepia confusa shells. Relative dimensions as \% shell length.

|  | A2140 | N.M. 961 | A31402 |  |  |  | A31403 | $\begin{gathered} \text { Adam } \\ \& \\ \text { Rees } \\ 1966 \end{gathered}$ | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| L in mm | 88 | $\pm 141$ | $106 \pm$ | $\pm 105$ | $\pm 93$ | $\pm 104$ | $\pm 53$ |  |  |  |  |
| Th : | 19,3 9,1 | 15,6 6,4 | 17,9 8,5 | 18,1 7 7, | 19,4 8,6 | 19,2 | 24,5 11,3 | 19 | 8 | 19,1 8,5 | $15,6-24,5$ $6,4-11,3$ |
| Str z | 55,7 | 65,2 | 54,7 | 58,1 | 59,1 |  |  | $\pm 62$ | 6 | 59,1 | 54,7-65,2 |

Table 17. Sepia confusa males. Relative dimensions as \% MLd.

|  | A65i6 | N.M.96I | N.M.962 | $A_{31402}$ |  |  |  | Adam \& Rees 1966 |  |  | Massy 1925 | $\begin{gathered} \text { Massy } \\ 1928 \end{gathered}$ | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 140 | 141 | 133 | 106 | 105 | 93 | 104 | 90 | $\pm 105$ | $\pm 100$ | 135 | 148140 |  |  |  |
| MLv | 77,9 | 75,2 | 75,9 | 65, I | 70,5 | 72,0 | 69,2 | 72 | 71 | 70 | 68,9 | 68,9 72,9 |  |  |  |
| MW | 32, I | 38,3 | 39, 1 | 38,7 | 42,9 | 48,4 | 49,0 | 38 | 36 | 35 | 40,0 | - - | II | 39,8 | 32, 1-49,0 |
| HL . | 20,0 | 21,3 | 29,3 | 24,5 | 23,8 | 31,2 | 21,2 | 22 | 24 | 25 |  |  |  |  |  |
| HW | 25,7 | 24,8 | 21, 1 | 33,0 | 34,3 | 34,4 | 36,5 | 32 | 27 | 28 | 25,2 | 20,9 | 12 | 28,6 | 20,9-36,5 |
| FL | 85,7 | 90, I | 86,5 | 75,5 | 8r,o | 80,6 | 81,7 | - |  | - |  |  |  |  |  |
| FW . . | 17,9 | 12,8 | I 1,3 | 7,6 | 8,6 | 9,7 | 13,5 | 14,5 | 12,5 | 13 |  |  | 10 | 12,1 | 7,6-17,9 |
| AL I | 35,7 | $4 \mathrm{I}, \mathrm{I}$ | 46,6 | 49, I | 43,8 | - | 52,9 | 33 | 35 | 35 | 48,9 | 37,2 47,9 | 12 | 42,2 | $33-52,9$ |
| AL II | 26,4 | 36,9 | 35,3 | 40,6 | 32,4 | 43,0 | 48, I | 28 | 29 | 30 | 38,5 | 42,6 38,6 | 13 | 36, I | 26,4-48, 1 |
| AL III | 27, 1 | 32,6 | 38,3 | 40,6 | 42,9 | 43,0 | 38,5 | 28 | 29 | 28 | 34,8 | 38,5 38,6 | 13 | 35,4 | 27, 1-43,0 |
| AL IV | 30,7 | 41, I | 37,6 | 45,3 | 42,9 | 48,4 | 45,2 | 28 | 31 | 30 | 40,0 | 45,3 40,0 | 13 | 38,9 | $28-48,4$ |
| TL ${ }_{\text {Rt }}$ | 67, 1 | 123,4 | - |  | - | - |  | - | 85 | 90 |  | 125,0 100,0 |  |  |  |
| ${ }^{\text {Lt }}$ | 65,7 | - | 105,3 | 166,0 |  | - | 146,2 |  | 8 |  |  |  |  |  |  |
| Tcl ; | 10,7 | 9,9 | 10,5 | 15,1 | - | - | 16,3 | - | 13 | 14 |  | 10,1 10,0 | 9 | 12,2 | 9,9-16,3 |
| 'Tail' | 95,0 | 125,5 | 38,3+ | 77,4+ | 82,9 | 8o,6+ | - |  |  |  | 96,3 | 135, $\mathrm{I}+$ - |  |  |  |

Table 20. Sepia incerta males. Relative dimensions as \% MLd.

|  | N.M. 956 | N.M. 957 | $A_{30143}$ |  |  |  |  | Massy 1925 |  | Massy 1928 |  | $\begin{gathered} \text { Adam \& } \\ \text { Rees } \\ \text { Ig66 } \end{gathered}$ | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 148 | 146 | 120 | 127 | 127 | 142 | 93 | ${ }^{1} 36$ | 125 | 150 | 147 | 135140 |  |  |  |
| MLv | 84,5 | 82,2 | 85,0 | 83,5 | 81, 1 | 78,9 | 83,9 |  |  | 85,3 | 83,7 | $67 \quad 74$ |  |  |  |
| MW | 34,4 | 34,3 | 35,0 | 30,7 | 35,4 | 31,7 | 35,6 |  |  |  |  | $37 \quad 34$ | 9 | 34,2 | 30,7-37 |
| HL | 22,3 | 23,3 | 19,2 | 18,9 | 16,5 | 21,8 | 18,3 |  |  |  |  | $27 \quad 21$ |  |  |  |
| HW | 23,0 | 23,3 | 21,7 | 25,2 | 28,4 | 23,2 | 24,8 |  |  | 22,7 | 20,4 | $24 \quad 25$ | 11 | 23,8 | 20,4-28,4 |
| FL | 95,4 | 93,8 | 95, | 93,7 | 91,3 | 83, I | 94,7 |  |  |  |  |  |  |  |  |
| FW | 9,5 | 6,2 | 8,3 | 10,2 | 10,2 | 9,2 | 9,7 |  |  |  |  | II II | 9 | 9,5 | 6,2-11 |
| AL I | 85,8 | 93,8 | 75,0 | 76,4 | 68,5 | 88,0 | 32,3 | 127,9 | 107,2 | 92,7 | 91,8 | 81 84 | $12^{*}$ | 89,3* | 68,5-127,9* |
| AL II | 38,5 | 37,0 | 40,8 | 38,6 | 39,4 | 35,2 | 30, I |  |  | 42,7 | $4 \mathrm{I}, 5$ | $\begin{array}{ll}37 & 39\end{array}$ | $12^{*}$ | 41,6* | 35,2-61, ${ }^{\text {* }}$ |
| AL III | 37,2 | 36,3 | 40,0 | 39,4 | 37,8 | 37,3 | 33,4 | 62,5 | 45,6 | 49,3 | 40, 1 | $\begin{array}{ll}38 & 39\end{array}$ | 12* | 41,9* | 36,3-62,5* |
| AL IV | 66,9 | 66,5 | 63,3 | 60,6 | 66, I | 63,4 | 38,7 | 102, 2 |  | 74, ${ }^{\text {o }}$ | 74, 1 | 59 <br> 6 | 12* | 69,5* | 59-102,2* |
| $\mathrm{TL}^{\text {Rt. }}$ | 66,9 |  | 8ı,6 | 107,9 | - | 96,5 | 73, 1 |  | 96,8 | 67,3 | 74, r |  |  |  |  |
| ${ }^{\text {TL }} \mathrm{Lt}$. | $6 \mathrm{r}, 5$ | 82,2 | - | 83,5 | - |  |  | ${ }^{130,1}$ | 138,4 | 65,3 | 74,8 |  |  |  |  |
| Tcl | 12,8 | 14,4 | 15,8 | 15,8 | - | 13,4 | 15, I | 22,1 | 19,2 | 12,7 | 12,2 | $15 \quad 14$ | 12 | 15,2 | 12,2-22,1 |

* Young male (MLd 93 mm ) not included.

Table 21. Sepia incerta females. Relative dimensions as \% MLd.

|  | A30143 | Massy 1925 |  | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 82 | 90 | 80 |  |  |  |
| MLv | 85,4 |  |  |  |  |  |
| MW | 36,6 | 48,9 | 53,8 | 3 | 46,4 | 36,6-53,8 |
| HL . | 18,3 |  |  |  |  |  |
| HW | 26,8 | 33,3 | 30,0 | 3 | 30,0 | 26,8-33,3 |
| FL . | 92,7 |  |  |  |  |  |
| FW . | 8,5 |  |  |  |  |  |
| AL I | 26,8 |  | 45,0 | 3 | 42, I | 26,8-54,4 |
| AL II . | 22,0 | 57,8 | 61,3 | 3 | 47,0 | 22,0-61,3 |
| AL III . | 31,7 | 64,4 | 56,3 | 3 | 50,8 | 31,7-64,4 |
| AL IV . | 35,4 | 60,0 | 62,5 | 3 | 52,6 | 35,4-62,5 |
| TL ${ }_{\text {Rt }}$. | 87,8 |  | - |  |  |  |
| Tcl ${ }^{\text {Lt }}$. | 67, 1 | $16,7$ | - |  |  |  |
| Tcl . . | 14,6 |  | - | 2 | 15,7 | 14,6-16,7 |

Table 22. Sepia incerta shells. Relative dimensions (approximate) as \% shell length.
Many of the shells are broken.

|  | $\begin{array}{cc} \text { Massy } 1925 \\ \text { No. } 8 & \text { No. } 16 \end{array}$ |  | $\underset{\text { A. }}{\substack{\text { N.M. } 958 \\ B}}$ |  | $\stackrel{\mathbf{B}}{\mathrm{N} . \mathrm{M} .970}{ }_{\mathbf{C}}$ |  | Punta Zavora |  |  |  | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\underline{L}$ in mm | 135 | 125* | 148* | 146* | 81 | 65 | 106 | 77 | 66 | 49 |  |  |  |
| W . | 17 | 16,8 | 14,9 | 13,7 | 19,8 | 20,0 | 17,9 | 20,8 | 19,7 | 22,4 | 10 | 18,3 | 13,7-22,4 |
| Th |  | 8 | 6,1 | 5,5 | 7,4 | 7,7 | 7,5 | 7,8 | 7,6 | 8,2 | 9 | 7,3 | 5,5-8,2 |
| Str z | ? 70 | ? 66 | 57,4† | 47,9† | 67,9 | 56,9 | 69,8 | 62,3 | 56,1 | 63,3 | 8 | 64,0 | 56,1-? 70 |

[^14]Table 23. Sepia burnupi. Relative dimensions as \% MLd.


Table 24. Sepia burnupi shells. Relative dimensions (approximate) as \% shell length.

|  | A2147 |  |  | $\underset{\mathrm{C}}{\mathrm{~N} . \mathrm{M}^{2} 98}$ | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| L in mm | 47* | $4^{1}$ * | 55* | 50* |  |  |  |
| W | 25,5 | 24,4 | 23,6 | 24 | 4 | 24,4 | 23,6-25,5 |
| Th | 7,4 | 6,1 | 6,4 | 7 | 4 | 6,7 | 6,1-7,4 |
| Str z | 85,1 | 78,0 | 81,8 | 74 | 4 | 79,7 | $74-85, \mathrm{I}$ |

* Anterior tip of shell missing.

Table 25. Sepia joubini males. Relative dimensions as \% MLd.

|  | A3014 ${ }^{1}$ | A30142 | A30172 |  |  |  |  | Massy 1927 |  | A31393 |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 29 | 34 | 36 | 36 | $4^{1}$ | 39 | 39 | 40 | 33 | 35 |  | $4{ }^{1}$ |
| MLv | 79,3 | 85,3 | 75,0 | 80,6 | 80,5 | 79,5 | 84,6 |  |  | 77, 1 | 82,9 | 75,6 |
| MW | 37,9 | 41,2 | 38,9 | 41,7 | 31,7 | 38,5 | 38,5 |  |  | 40,0 | 36,6 | 36,6 |
| HL | 24, I | 26,5 | 22,2 | 22,2 | 24,4 | 20,5 | 15,4 |  |  | 22,9 | 19,5 | 22,0 |
| HW | 34,5 | 35,3 | 33,3 | 33,3 | 31,7 | 28,2 | 30,8 | 30,0 | 36,4 | 34,3 | 31,7 | 31,7 |
| FL | 79,3 | 82,4 | 83,3 | 86,1 | 85,4 | 87,2 | 84,6 |  |  | 82,9 | 80,5 | 80,5 |
| FW | 5,2 | 8,8 | 5,6 | 8,3 | 9,8 | 7,7 | 7,7 |  |  | 8,6 | 7,3 | 7,3 |
| AL I | 24, 1 | 32,4 | 22,2 | 30,6 | 24,4 | 25,6 | 28,2 | 40,0 | - | 25,7 | 24,4 | 24,4 |
| AL II | 24, 1 | 29,4 | 22,2 | 30,6 | 24,4 | 23,1 | 30,8 | 37,5 | - | 22,9 | 22,0 | 22,0 |
| AL III | 20,7 | 29,4 | 25,0 | 27,8 | 19,5 | 23,1 | 25,6 | 40,0 | 39,4 | 22,9 | 22,0 | 22,0 |
| AL IV | 17,2 | 32,4 | 22,2 | 25,0 | 22,0 | 23,1 | 25,6 | 45,0 | 45,5 | 25,7 | 24,4 | 22,0 |
| $\mathrm{TL}^{\mathrm{Rt}}$. | 110,3 | - | - | 91,7 | - | 117,9 | 125,6 |  |  | 91,4 | 100,0 | 78,0 |
| ${ }^{\text {L }}$ Lt. | 89,7 | - | - | - | 73,2 | 102,6 | - | - | 121,2 | 94,3 | 104,9 | 82,9 |
| Tcl | 10,3 | - | - | 13,9 | 12,2 | 10,3 | 7,7 | - | 12,1 | 10,0 | 9,8 | 9,8 |


|  | A31393 (cont.) |  |  |  |  |  |  |  | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 39 | 36 | 34 | 33 | 26 | 30 | 21 | 20 |  |  |  |
| MLv | 74,4 | 77,8 | 76,5 | 75,8 | 76,9 | 76,7 | 76,2 | 70,0 |  |  |  |
| MW | 35,9 | 38,9 | 38,2 | 42,4 | 42,3 | 40,0 | 47,6 | 35,0 | 18 | 39,0 | 31,7-47,6 |
| HL | 17,9 | 22,2 | 20,6 | 21,2 | 23, 1 | 20,0 |  |  |  |  |  |
| HW | 30,8 | 30,6 | 32,4 | 36,4 | 42,3 | 36,7 |  |  | 18 | 33,4 | 28,2-42,3 |
| FL. | 82,1 | 77,8 | 82,4 | 78,8 | 80,8 | 80,0 |  |  |  |  |  |
| FW | 5,1 | 5,6 | 5,9 | 6,1 | 11,5 | 6,7 |  |  | 16 | 7,3 | 5, 1-1 1,5 |
| AL I | 25,6 | 27,8 | 20,6 | 21,2 | 26,9 | 26,7 |  |  | 17 | 26,5 | 20,6-40,0 |
| AL II | 20,5 | 25,0 | 20,6 | 21,2 | 26,9 | 23,3 |  |  | 17 | 25, 1 | 20,5-37,5 |
| AL III | 20,5 | 25,0 | 20,6 | 21,2 | 30,8 | 23,3 |  |  | 18 | 25,5 | 19,5-40,0 |
| AL IV | 23,1 | 27,8 | 23,5 | 24,2 | 34,6 | 23,3 |  |  | 18 | 27,0 | ${ }_{1} 7,2-45,5$ |
| $\mathrm{TL}^{\text {Rt.}}$ | 94,9 | 100,0 | 105,9 | 75,8 | - | 86,7 |  |  |  |  |  |
| ${ }^{\text {TL }}$ Lt. | 102,6 | 102,8 | 111,8 | - | - 1 | 106,7 |  |  |  |  |  |
| Tcl | 10,3 | 13,9 | 11,8 | 12,1 | - | 13,3 |  |  | 14 | 11,3 | 7,7-13,9 |

Table 26. Sepia joubini females. Relative dimensions as \% MLd.

|  | $\mathrm{A}_{3}{ }^{1} 41$ |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 40 | 27 | 29 | 28 | 31 | 35 | 33 | 35 | 35 | $3^{8}$ | $3^{8}$ | 39 |
| MLv | 77,5 | 81,5 | 82,8 | 82, 1 | 8o,6 | 82,9 | 81,8 | 80,0 | 82,9 | 8ı,6 | 81,6 | 82, I |
| MW | 35,0 | 40,7 | 41,4 | 42,9 | 41,9 | 40,0 | 39,4 | 40,0 | 37,1 | 36,8 | 36,8 | 35,9 |
| HL | 22,5 | 25,9 | 20,7 | 25,0 | 25,8 | 25,7 | 24,2 | 22,9 | 25,7 | 23,7 | 21,1 | 20,5 |
| HW | 32,5 | 40,7 | 37,9 | 39,3 | 38,7 | 34,3 | 36,4 | 34,3 | 34,3 | 28,9 | 31,6 | 33,3 |
| FL | 82,5 | 81,5 | 79,3 | 82, I | 83,9 | 82,9 | 84,8 | 77, 1 | 82,9 | 81,6 | 78,9 | 76,9 |
| FW | 5,0 | 7,4 | 10,3 | 7, 1 | 6,5 | 7,1 | 9, I | 5,7 | 5,7 | - | 5,3 | 6,4 |
| AL I | 25,0 | 33,3 | 27,6 | 28,6 | 29,0 | 31,4 | 27,3 | 28,6 | 31,4 | 31,6 | 28,9 | 33,3 |
| AL II | 35,0 | 25,9 | 31,0 | 28,6 | 25,8 | 34,3 | 30,3 | 31,4 | 45,7 | 36,8 | 39,5 | 41,0 |
| AL III | 37,5 | 25,9 | 31,0 | 28,6 | 29,0 | 37, I | 30,3 | 28,6 | 42,9 | 39,5 | 39,5 | 43,6 |
| AL IV | 30,0 | 29,6 | 31,0 | 32, 1 | 29,0 | 31,4 | 33,3 | 28,6 | 34,3 | 31,6 | 34,2 | 30,8 |
| TL ${ }_{\text {Rt }}$ | 90,0 | 140,7 | - | 107, 1 | 1 12,9 | 94,3 | 87,9 | 114,3 | 100,0 | - | 78,9 | - |
| ${ }^{\text {L }} \mathrm{Lt}$ | 92,5 | 107,4 | 96,6 | 110,7 | - | 122,9 | 87,9 | 105,7 | 100,0 | 84,2 | 84,2 | $100,0$ |
| Tcl | 10,0 | 14,8 | 13,8 | 14,3 | 12,9 | 14,3 | 12,1 | 14,3 | 12,9 | 10,5 | 13,2 | 10,3 |


|  | A3014I (cont.) |  |  | A30142 |  | A30172 | Massy 1927 <br> (Adam <br> \& Rees <br> 1966) <br> PFili74i PFio7ı |  | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 40 | 40 | 43 | 36 |  | 39 | 47 | 36 |  |  |  |
| MLv | 82,5 | 80,0 | 76,7 | 86, I |  | 82, I | 83 |  |  |  |  |
| MW | 35,0 | 32,5 | 32,6 | 38,9 | 40,6 | 38,5 | 32 |  | 19 | 37,8 | $32-42,9$ |
| HL | 20,0 | 22,5 | 20,9 | 30,6 | 40,6 | 20,5 | 25 |  |  |  |  |
| HW | 30,0 | 27,5 | - | 33,3 | 34,4 | 33,3 | 30 | 30,6 | 19 | 33,8 | 27,5-40,7 |
| FL | 80,0 | 80,0 | 76,7 | 80,6 | 90,7 | 87,2 | 85 |  |  |  |  |
| FW | 7,5 | 7,5 | 9,3 | 5,6 | 6,3 | 7,7 | 8,5 |  | 18 | 7,1 | 5,0-10,3 |
| AL I | 30,0 | 30,0 | 27,9 | 27,8 | 43,8 | 30,8 | 30 | 38,9 | 20 | 30,8 | 25,0-43,8 |
| AL II | 37,5 | 37,5 | 37,2 | 38,9 | 43,8 | 43,6 | 38 | 47,2 | 20 | 36,5 | 25,8-47,2 |
| AL III | 37,5 | 37,5 | 37,2 | 38,9 | 31,3 | 43,6 | 38 | 47,2 | 20 | 36,2 | 25,9-47,2 |
| AL IV | 32,5 | 30,0 | 27,9 | 30,6 | 43,8 | 28,2 | - | 38,9 | 19 | 32,0 | 27,9-43,8 |
| TL Rt | 90,0 | 95,0 | 81,4 |  |  | 94,9 |  | 127,8 |  |  |  |
| ${ }^{\text {L }}$ Lt | 97,5 | 107,5 | 74,4 |  | 93,8 | 92,3 |  |  |  |  |  |
| Tcl | 15,0 | 15,0 | 11,6 |  | 15,6 | 12,8 | 9,5 | 13,9 | 19 | 13,0 | 9,5-1 5,6 |

Table 27. Sepia adami females. Relative dimensions as \% MLd.

|  | A31394 | A30149 |  |  |  |  | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 59 | 47 | 44 | 28 | 24 | 25 |  |  |  |
| MLv | 76,3 | 74,5 | 72,7 | 78,6 | 75, 0 | 80,0 |  |  |  |
| MW | 40,7 | 38,3 | 38,6 | 39,3 | 45,8 | 44,0 | 6 | 4 1 , I | 38,3-45,8 |
| HL . | 22,0 | 23,4 | 20,5 | 25,0 |  |  |  |  |  |
| HW | 35,6 | 36,2 | 36,4 | 39,3 |  |  | 4 | 36,9 | 35,6-39,3 |
| FL | 86,4 | 80,9 | 77,3 | 82, 1 |  |  |  |  |  |
| FW | 6,8 | 6,4 | 6,8 | 7,1 |  |  | 4 | 6,8 | 6,4-7, 1 |
| AL I | 32,2 | 29,8 | 27,3 | 25,0 |  |  | 4 | 28,6 | 25,0-32,2 |
| AL II | 30,5 | 25,5 | 25,0 | 21,4 |  |  | 4 | 25,6 | 21,4-30,5 |
| AL III | 28,8 | 29,8 | 25,0 | 25,0 |  |  | 4 | 27,2 | 25,0-29,8 |
| AL IV | 32,2 | 29,8 | 31,8 | 25,0 |  |  | 4 | 29,7 | 25,0-32,2 |
| TL Rt | 98,3 | 63,8 | - | 85,7 |  |  |  |  |  |
| ${ }^{\text {L }}$ Lt | 78,0 | 78,7 | 88,6 | 92,9 |  |  |  |  |  |
| Tcl . | 1,3,6 | 12,8 | 12,5 | 14,3 |  |  | 4 | 13,3 | 12,5-14,3 |

Table 28．Sepia australis males．Relative dimensions as \％MLd．

|  |  | A30148 |  |  |  |  |  |  |  |  |  | $\mathrm{A}_{3} \mathrm{O}_{50}$ |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | －．．．． | 20 | 27 | 28 | 25 | 39 | 47 | 52 | 42 | 50 | 49 | 39 | 40 | 27 | 27 |
| MLv | ．．．．． | 70，0 | $8 \mathrm{I}, 5$ | 82，1 | 84，0 | 84，6 | 85,1 | 82，7 | 83，3 | 88，o | 87，8 | 82，1 | 85，0 | 77，8 | 81，5 |
| MW | ．．．．． | 60，0 | 48， 1 | 46，4 | 52，0 | 43，6 | 42，6 | 38，5 | 40，5 | 42，0 | 42，9 | 43，6 | 42，5 | 48，1 | 44，4 |
| HL | ．．．．． |  | 25，9 | 21，4 |  | 23，1 | 21，3 | 19，2 | 19，0 | 24，0 | 22，4 | 20，5 | 22，5 | 25，9 | 18，5 |
| HW | －．．． |  | 48，1 | 46，4 |  | 43，6 | 42，6 | 38，5 | 40，5 | 46，0 | 42，9 | 41，0 | 42，5 | 44，4 | 44，4 |
| FL | ．．．．． |  | 77，8 | 71，4 |  | 100，0 | 85,1 | 73， 1 | 81，0 | 82，0 | 83，7 | 84，6 | 80，0 | 77，8 | 81，5 |
| FW | ．．．．． |  | 14，8 | 14，3 |  | 10，3 | 8，5 | 7，7 | 9，5 | 10，0 | 10，2 | 12，8 | 10，0 | I 1， 1 | II，${ }^{\text {I }}$ |
| AL I | ．．．．． |  | 29，6 | 28，6 |  | 25，6 | 31，9 | 30，8 | 31，0 | － | 32，7 | 25，6 | 32，5 | 25，9 | 22，2 |
| AL II | ．．．．． |  | 29，6 | 28，6 |  | 30，8 | 29，8 | 30，8 | 28，6 | － | 32，7 | 25，6 | 30，0 | 22，2 | 22，2 |
| AL III | ．．．．． |  | 29，6 | 28，6 |  | 30，8 | 31，9 | 32，7 | 28，6 | － | 34，7 | 28，2 | 32，5 | 22，2 | 22，2 |
| AL IV | ．．．．． |  | 29，6 | 32，1 |  | 33，3 | 34，0 | 32，7 | $3 \mathrm{r}, \mathrm{o}$ | － | 36，7 | 33，3 | 30，0 | 25，9 | 29，6 |
| $\mathrm{TL}^{\mathrm{Rt}}$ | －．．． |  | 111， 1 | － |  | 102，6 | 97，9 | 98，1 | 102，4 | － | 75，5 | 92，3 | 102，5 | H11， | 100，0 |
| $\mathrm{TL}_{\text {Lt }}$ | ．．．．． |  | 114，8 | － |  | 107，7 | 112，8 | 103，8 | 100，0 | － | 89，8 | 102，6 | － | HII， 1 | 107，4 |
| Tcl | ．．．．． |  | 18，5 | － |  | 17，9 | 14，9 | ${ }^{15} 54$ | 14，3 | － | 14，3 | 12，8 | 12，5 | I 1， 1 | 14,8 |


| $\begin{aligned} & \text { in } \\ & 0 \\ & 0 \\ & \text { en } \end{aligned}$ |  <br>  |
| :---: | :---: |
| $\begin{aligned} & \text { H } \\ & \text { N } \\ & \text { O } \end{aligned}$ |  <br>  <br>  <br>  <br>  <br> a <br> む瓦会 <br>  |
| N |  |
| 合 | men <br>  <br>  <br>  |
|  |  |
|  |  |

Table 28 （continued）

|  | A30157 |  | A30159 |  |  | A30160 | $A_{30161}$ |  |  |  |  | A $_{3} \mathrm{O}_{1} 63$ |  | A30164 |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 45 | 51 | 31 | 17 | 24 | 32 | 54 | 48 | 46 | 23 | 24 | 34 | 22 | 36 | 36 | 36 |
| MLv | 84，4 | 82，4 | 80，6 | 82，4 | 79，2 | 81，3 | 81，5 | 85，4 | 87，0 | 87，0 | 79，2 | 88，2 | 81，8 | 83，3 | 80，6 | 77，8 |
| MW | 40，0 | 37，3 | 41，9 | 52，9 | 45，8 | 43，8 | 37，0 | 39，6 | 39，I | 56，5 | 50，0 | 50，0 | 54，5 | 41，7 | 44，4 | 38，9 |
| HL． | 20，0 | 21，6 | 25，8 |  |  | 21，9 | 18，5 | 20，8 | 23，9 | 30，4 | 25，0 | 29，4 | 31，8 | 25，0 | 16，6 | 25，0 |
| HW | 35，6 | 37，3 | 38，7 |  |  | 37，5 | 37，0 | 37，5 | 37，0 | 47，8 | 45，8 | 47， 1 | 50，0 | 38，9 | 41，7 | 33，3 |
| FL | 75，6 | 66，7 | 77，4 |  |  | 81，3 | 79，6 | 77， 1 | 78，3 | 78，3 | 75，0 | 88，2 | 81，8 | 91，7 | 83，3 | 77，8 |
| FW | 8，9 | 9，8 | 9，7 |  |  | 9，4 | 7，4 | 8，3 | 6，5 | 8，7 | 8，3 | 11，8 | 9，1 | 11，1 | 11，1 | 11，1 |
| AL I | 33，3 | 27，5 | 29，0 |  |  | 28，1 | 27，8 | 29，2 | 28，3 | 26，1 | 25，0 | 29，4 | 27，3 | 27，8 | 33，3 | 27，8 |
| AL II | 31，1 | 23，5 | 29，0 |  |  | 28，1 | 29，6 | 29，2 | 26，1 | 30，4 | 29，2 | 29，4 | 27，3 | 30，6 | 30，6 | 30，6 |
| AL III | 31,1 | 27，5 | 32，3 |  |  | 31，3 | 29，6 | 27，1 | 28，3 | 30，4 | 29，2 | 29，4 | 22，7 | 30，6 | 33，3 | 33，3 |
| AL IV | 31，1 | 31，4 | 32，3 |  |  | 31，3 | 33，3 | 31，3 | 32，6 | 30，4 | 33，3 | 26，5 | 27，3 | 30，6 | 33，3 | 30，6 |
| TL Rt | 111， 1 | ， | 87， 1 |  |  | 93，8 | 83，3 | 100，0 | 97，8 | 126，1 | 91，7 | 79，4 | 90，9 | 88，9 | 75，0 | ， |
| ${ }^{12} \mathrm{Lt}$ | 122，2 | 103，9 | 90，3 |  |  | 90，6 | 103，7 | 104，2 | 115，2 | 104，3 | 112，5 | 76，5 | 90，9 | 80，6 | 77，8 | 83，3 |
| Tcl ． | 13，3 | 13，7 | 12，9 |  |  | 12，5 | 13，0 | 14，6 | 13，0 | ${ }^{17} 74$ | 20，8 | 11，8 | 13，6 | 13，9 | ${ }^{13,9}$ | 13，9 |


|  |  |
| :---: | :---: |
| 范 |  |
| z |  |
| $\begin{aligned} & \hline \hline \text { H } \\ & \text { o } \\ & \text { < } \end{aligned}$ |  <br>  |
| $\begin{aligned} & \text { O} \\ & 0 \\ & 0 \\ & \text { < } \end{aligned}$ |  <br>  |
| $\begin{aligned} & \text { n } \\ & 0 \\ & 0 \\ & \hline \end{aligned}$ |  <br>  |
|  | かのにNのMNMNのN みが |
| － |  <br>  |
|  |  |
| 6 0 0 0 4 |  <br>  |
|  |  <br>  <br>  <br>  |
|  |  |

Table 29. Sepia australis females. Relative dimensions as \% MLd.


| $\begin{aligned} & \text { dy } \\ & \text { d } \\ & \text { on } \end{aligned}$ |  <br>  |
| :---: | :---: |
| $\begin{aligned} & 0 \\ & \stackrel{0}{0} \\ & \stackrel{\infty}{6} \end{aligned}$ |  <br>  <br>  <br>  <br>  <br>  <br>  |
| $\begin{aligned} & \text { in } \\ & \text { in } \\ & \text { on } \end{aligned}$ |  <br>  <br>  <br>  <br>  <br>  <br>  <br>  |
| 产 |  ส f <br>  |
|  |  |

Table 29 （continued）

|  | A30154（cont．） |  |  |  |  |  |  |  |  |  | A30155 | A30157 |  |  |  | $\mathrm{A}_{3}{ }^{1} 59$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 39 | 36 | $3^{8}$ | 39 | 31 | 30 | 29 | 22 | 19 | 17 | 38 | 49 | $4^{2}$ | 30 | 33 | 46 |
| MLv | 87，2 | 86，I | 84，2 | 87，2 | 83，9 | 86，7 | 82，8 | 90，9 | 89，5 | 82，4 | 84，2 | 79，6 | 83，3 | 83，3 | 81，8 | 84，8 |
| MW | 43，6 | 47，2 | 44，7 | 46，2 | 48，4 | 50，0 | 51，7 | 54，5 | 52，6 | 47， 1 | 44，7 | 42，9 | 42，9 | 46，7 | 45，5 | 43，5 |
| HL | 23，1 | 19，4 | 18，4 | 20，5 | 22，6 | 26，7 | 27，6 |  |  |  | 21， 1 | 20，4 | 23，8 | 20，0 | 24，2 | 19，6 |
| HW | 38，5 | 38，9 | 36，8 | 41，0 | 4 I .9 | 40，0 | $4^{1,4}$ |  |  |  | 39，5 | 34，7 | 38，I | 40，0 | 39，4 | 34，8 |
| FL． | 89，7 | 94，4 | 89，5 | 92，3 | 90，3 | 93，3 | 89，7 |  |  |  | 89，5 | 67，3 | 71，4 | 66，7 | 69，7 | 84，8 |
| FW | 12，8 | 13，9 | 13，2 | 12，8 | 12，9 | 13，3 | 13，8 |  |  |  | 5，3 | 8，2 | 11，9 | 6，7 | 9， 1 | 8，7 |
| AL I ． | 25，6 | 25，0 | 28，9 | 25，6 | 25，8 | 26，7 | 27，6 |  |  |  | 23，7 | 28，6 | 26，2 | 20，0 | 24，2 | 23，9 |
| AL II | 25，6 | 27，8 | 28，9 | 23， 1 | 29，0 | 26，7 | 27，6 |  |  |  | 26，3 | 28，6 | 26，2 | 23，3 | 24，2 | 26，I |
| AL III | 25，6 | 27，8 | 28，9 | 25，6 | 29，0 | 26，7 | 27，6 |  |  |  | 23，7 | 28，6 | 23，8 | 23，3 | 27，3 | 26， 1 |
| AL IV | 25，6 | 27，8 | 28，9 | 28，2 | 29，0 | 30，0 | 27，6 |  |  |  | 26，3 | 30，6 | 28，6 | 26，7 | 27，3 | 23，9 |
| TL ${ }_{\text {Rt }}$ ． | 87，2 | $6 \mathrm{I}, \mathrm{I}$ | 68，4 | 82， 1 | 106，5 | 93，3 | 93，r |  |  |  | － | 100，0 | 109，5 | I 10，0 | ， | 80，4 |
| ${ }^{T}$ Lt ． | 84，6 | 52，8 | 65，8 | 84，6 | 100，0 | 100，0 | 86，2 |  |  |  | － | 108，2 | 128，6 | 1 10，0 | 100，0 | 84，8 |
| Tcl | ${ }^{1} 5,4$ | 13，9 | ${ }^{1} 5,8$ | 15，4 | 16，I | 16，7 | 13,8 |  |  |  | － | 12，2 | 1 1，9 | 13，3 | 18，2 | 10，9 |


| $\begin{aligned} & \text { n } \\ & \stackrel{0}{0} \\ & 0 \\ & 0 \end{aligned}$ |  か が <br> व <br>  |
| :---: | :---: |
| $\begin{aligned} & \text { +1 } \\ & \underset{\sim}{m} \\ & \end{aligned}$ |  <br> 0001120120120 <br>  |
| $$ | $\simeq \stackrel{0}{1}$ <br> 00000000000000 <br>  <br>  <br>  <br> NONDOM N O N O ONN <br>  <br>  |
| $\begin{aligned} & 8 \\ & \stackrel{0}{0} \\ & \underset{\sim}{e} \end{aligned}$ |  |
|  |  |

Table 29 (continued)

|  | A30166 | A30168 |  |  |  | $\mathrm{A}_{30170}$ | $A_{30171}$ |  |  | A30175 |  | A30190 | A30334 | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 43 | 43 | 23 | 25 | 22 | 49 | 45 | 43 | 43 | $4^{8}$ | 36 | 54 | 58 |  |  |  |
| MLv | $8 \mathrm{r}, 4$ | 81,4 | 82,6 | 80,0 | 77,3 | 83,7 | 86,7 | 83,7 | 83,7 | 77,1 | 83,3 | 88,9 | 86,2 |  |  |  |
| MW | 41,9 | 44,2 | 47,8 | 48,0 | 54,5 | 40,8 | 42,2 | 44,2 | 44,2 | 39,6 | 44,4 | 40,7 | 39,7 | 77 | 46,0 | 37,5-66,7 |
| HL | 25,6 | 16,3 |  |  |  | 20,4 | 17,8 | 20,9 | 18,6 | 18,8 | 22,2 | 24,1 | 19,0 |  |  |  |
| HW | 39,5 | 34,9 |  |  |  | 36,7 | 37,8 | 37,2 | 39,5 | 35,4 | 41,7 | 37,0 | 37,9 | 66 | 39,5 | 24,1-48,1 |
| FL | 88,4 | 88,4 |  |  |  | 81,6 | 91,1 | 86,0 | 86,o | 72,9 | 77,8 | 77,8 | 84,5 |  |  |  |
| FW | 11,6 | 1 1,6 |  |  |  | 10,2 | [1, 1 | 9,3 | 9,3 | 6,3 | 5,6 | 9,3 | 12,1 | 66 | 10,7 | 5,3-15,0 |
| AL I | 27,9 | 25,6 |  |  |  | 24,5 | 26,7 | 23,3 | 25,6 | 25,0 | 25,0 | 29,6 | 29,3 | 66 | 26,6 | 19,6-34,0 |
| AL II | 25,6 | 25,6 |  |  |  | 26,5 | 26,7 | 27,9 | 25,6 | 22,9 | 25,0 | 25,9 | 29,3 | 66 | 26,7 | 16,0-34,5 |
| AL III | 25,6 | 25,6 |  |  |  | 28,6 | 28,9 | 30,2 | 25,6 | 20,8 | 22,2 | 24, 1 | 27,6 | 66 | 26,9 | 20,0-35,4 |
| AL IV | 27,9 | 27,9 |  |  |  | 26,5 | 26,7 | 25,6 | 23,3 | 22,9 | 25,0 | 25,9 | 27,6 | 66 | 27,5 | 19,0-37,9 |
| TL Rt | 51,2 | 58,1 |  |  |  | 83,7 | 82,2 | 79,1 | 76,7 | 91,7 | 77,8 | 101,9 | 69,0 |  |  |  |
| ${ }^{12}$ Lt | 69,8 | 72,1 |  |  |  | 77,6 | 93,3 | 76,7 | 67,4 | 87,5 | 55,6 | 101,9 | - |  |  |  |
| Tcl . | 16,3 | 14,0 |  |  |  | 12,2 | 1 1, I | 1 1,6 | 1 1,6 | 12,5 | 11,1 | 13,0 | 12,1 | 64 | 14, 1 | 9,5-19,2 |

## A REVIEW OF THE SEPIIDAE (CEPHALOPODA) OF SOUTHERN AFRICA <br> 299

Table 30. Sepia australis shells. Relative dimensions as \% shell length.

|  | L in mm | W | Th | Str z |
| :---: | :---: | :---: | :---: | :---: |
| 2727 - . . . . . . . | 33 | 36,4 | 9,1 | 69,7 |
|  | 47 | 34, 0 | 8,5 | 72,3 |
|  | 48 | 35,4 | 8,3 | 75,0 |
|  | 47 | $34, \mathrm{o}$ | 7,4 | 72,3 |
|  | 46 | 34,8 | 8,7 | 71,7 |
|  | 49 | 32,7 | 8,2 | 71,4 |
|  | 51 | 33,3 | 7,8 | 72,5 |
|  | 43 | 32,6 | 8, 1 | 69,8 |
|  | 48 | 33,3 | 9,4 | 70,8 |
|  | 51 | - | 7,8 | 78,4 |
|  | 54 | 35,2 | 8,3 | 72,2 |
| A30504 . . . . . . | 41 | 36,6 | 9,8 | 70,7 |
| Mossel Bay (Cape Peninsula) | 52 | 32,7 | 8,7 | 75,0 |
|  | 58 | 29,3 | 7,8 | 72,4 |
| North of Olifants River | 57 | 33,3 | 8,8 | 71,9 |
|  | 55 | 29, 1 | 7,3 | 74,5 |
| Strandfontein to Muizenberg | 45 | 33,3 | 8,9 | 66,7 |
|  | 46 | 32,6 | 8,7 | 76, 1 |
| Millers Point | 55 | 30,9 | 9,1 | 70,9 |
|  | 52 | 32,7 | 8,7 | 71,2 |
|  | 43 | 34,9 | 9,3 | 72,1 |
| Namaqualand coast | 59 | 32,2 | 9,3 | 72,9 |
| Simonstown | 40 | 32,5 | 8,8 | 72,5 |
| Arniston . |  | 32,1 | 7,5 | 73,6 |
|  | 56 | 32,1 | 7,1 | 75,0 |
| N. |  | 24 | 25 | 25 |
| Mean |  | 33,2 | 8,5 | 72,5 |
| Range . . . . . . |  | 29, 1-36,6 | 7,1-9,8 | 66,7-78,4 |

Table 31. Sepia tuberculata males. Relative dimensions as \% MLd.

|  | A30121 | A30279 | A30600 |  | A31235 | Adam <br>  <br> Rees <br> 1966 | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 25 | 42 | 27 | 33 | 51 | 47 |  |  |  |
| MLv | 92,0 | 85,8 | 96,3 | 90,9 | 90,2 | 92 |  |  |  |
| MW | 64,0 | 69,0 | 63,0 | 66,7 | 58,8 | 66 | 6 | 64,6 | 58,8-69 |
| HL | 48,0 | 50,0 | 59,3 | 57,6 | 37,3 | 60 |  |  |  |
| HW | 60,0 | 52,4 | 70,4 | 63,6 | 47, 1 | 53 | 6 | 57,8 | 47,1-70,4 |
| FL | 104,0 | 85,8 | 96,3 | 103,0 | 103,9 | 96 |  |  |  |
| FW . | 16,0 | 16,7 | 18,5 | 15,2 | 19,6 | ${ }_{15}+$ | 5 | 17,2 | 15,2-19,6 |
| AL I | 52,0 |  | 70,4 | 72,7 | 47, 1 | 57 | 5 | 59,8 | 47,1-72,7 |
| AL II | 48,o | - | 70,4 | 66,7 | 43, 1 | 60 | 5 | 57,6 | 43,1-70,4 |
| AL III | 56,o | 45,3 | 66,7 | 66,7 | 49,0 | 57 | 6 | 56,8 | 45,3-66,7 |
| ALIV | 52,0 | 52,4 | 55,6 | 54,5 | 47,1 | 57 | 6 | 53,1 | 47,1-57 |
| TL | 76,0 |  | ${ }^{140,7}$ | 127,3 | 49,0 | 106 |  |  |  |
| Tcl . | 32,0 | - | 37,0 | 30,3 | 21,6 | 23,5 | 5 | 28,9 | 21,6-37,0 |

## See page $\mathbf{3 0 2}$ for Table 32

Table 33. Sepia tuberculata shells. Relative dimensions as \% shell length.


Table 33 (continued)

|  |  |  |  |  |  |  |  |  |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |

Table 32. Sepia tuberculata females. Relative dimensions as \% MLd.

|  | A29781 | A29867 | A30123 | A30139 | A30 80 |  |  | A30559 |  | $\operatorname{lam} 19$ |  | Adam \& Rees 1966 | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm . | 30 | 75 | 71 | 82 | 70 | 48 | 58 | 36 | 56 |  | 49 | 64 |  |  |  |
| MLv . | 80,0 | 89,4 | 97,2 | 89,0 | 90,0 | 96,o | 98,4 | 97,2 | 84 | 96 | 88 | 92 |  |  |  |
| MW | 80,0 | 66,7 | 77,5 | 73,2 | 68,6 | 66,7 | 67,3 | 63,9 | 62,5 | 70 | 61 | 70 | 12 | 69,0 | 6ı -80,0 |
| HL | 46,7 | 45,4 | 46,5 | 45,0 | 53,0 | 50,0 | 46,6 | 52,8 | 46,5 | 56,5 | 53 | 47 |  |  |  |
| HW . | 60,0 | 49,4 | 53,6 | 41,5 | 53,0 | 58,4 | 55,2 | 61,1 | 44,5 | 47 | 51 | 55 | 12 | 52,5 | 41,5-61,1 |
| FL | 113,3 | 112,0 | 121,1 | 109,8 | 1 18,6 | H12,5 | 108,7 | 105,6 | 89 |  | 92 | 102 |  |  |  |
| FW | 16,7 | 17,3 | 18,3 | 23,2 | 17,2 | 20,8 | 20,7 | 19,4 | 10,7 | - | 8,2 | 19 | 11 | 17,4 | 8,2-23,2 |
| AL I | 43,3 | 48,0 | 67,6 | 55,0 | 55,8 | 46,0 | 51,8 | $6 \mathrm{I}, \mathrm{I}$ | 52 | 45,5 | 51 | 53 | 12 | 52,5 | 43,3-67,6 |
| AL II | 50,0 | 58,7 | 66,2 | 52,5 | 63,0 | 64,6 | 57,0 | 58,3 | 50 | 39,5 | 53 | 52 | 12 | 55,4 | 39,5-66,2 |
| AL III | 46,7 | 57,3 | 67,6 | 50,0 | 57,2 | 64,6 | 53,5 | 58,3 | - | 45,5 | 47 | 52 | 11 | 54,5 | 45,5-67,6 |
| AL IV | 50,0 | 46,7 | 63,4 | 46,4 | 53,0 | 58,4 | 51,8 | 52,8 | 46,5 | 43,5 | 45 | 59 | 12 | 51,4 | 43,5-63,4 |
| TL | 76,7 | 106,7 | 107,0 | 89,0 | 190,0 | 114,5 | 136,0 | 108,3 |  | 104 | - | 117 |  |  |  |
| Tcl | 33,3 | 30,7 | 43,7 | 30,5 | 37,2 | 33,4 | 46,6 | 36,1 |  | 30 | 28,5 | 3 r | 12 | 33,8 | $25-46,6$ |

Table 34. Sepia papillata males. Relative dimensions as \% MLd.

|  | A30118 |  | A30120 | A30137 | A30138 | A30507 |  | A30509 | A31250 | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLdin mm | 100 | 115 | 112 | 100 | 110 | 115 | 105 | 105 | 113 |  |  |  |
| MLV | 95,0 | 87,0 | $\pm 69,6$ | 87,0 | 86,4 | 78,3 | 81,0 | 90,5 | 87,6 |  |  |  |
| MW | 68,0 50 | 60,9 38 | 62,5 | 60,0 | 59,1 | 59,1 | 76,2 | 64,8 | 58,4 | 9 | 63,2 | 58,4-76,2 |
| HL | 50,0 | 38,3 | 35,7 | 44,0 | 40,9 | 26,1 | 41,0 | 33,3 | 46,9 |  |  |  |
| FL | 1130 | ${ }_{1174}$ | 42,9 89 | 10,0 | 100,0 | 34, | 104, | 43,8 | 46,0 | 9 | 44,8 | 34,8-52,4 |
| FW | 20,0 | 26,1 | 14,3 | 15,0 | 13,6 | 13,0 | 28,6 | $\pm$ - | 17,7 | 8 | 18,5 | 13,0-28,6 |
| AL I | 53,0 | 62,6 | 42,0 | 54,0 | 52,7 | 47,8 | 56,2 | - | 64,6 | 8 | 54,1 | 42,0-64,6 |
| AL II | 57,0 | 56,5 | 42,0 | 55,0 | 57,3 | 47,8 | 61,9 | - | 64,6 | 8 | 55,3 | 42,0-64,6 |
| AL III | 64,0 | 61,7 | 42,9 | 55,0 | 65,5 | 47,8 | 64,8 |  | 58,4 | 8 | 57,5 | 42,9-65,5 |
| ALIV | 65,0 | 55,7 | 56,3 | 48,0 | 50,9 | 43,5 | 54,3 | 57,1 | 52,2 |  | 53,7 | 43,5-65,0 |
| TL | 105,0 | 64,3 | 160,7 | 75,0 | 77,3 |  |  |  | 167,3 |  |  |  |
| Tcl | 38,0 | 30,4 | 30,4 | 30,0 | 32,7 | - | - | - | 30,1 | 6 | 31,9 | 30,0-38,0 |

Table 35. Sepia papillata females. Relative dimensions as \% MLd.

|  | A30119 | A30124 | A30136 | A30140 |  |  | Hoyle 1910 | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm . | 130 | I I 5 | 95 | 140 | 120 | 135 | I 15 |  |  |  |
| MLv . | 92,3 | 91,3 | 88,4 | 82, I | 91,7 | 88,9 |  |  |  |  |
| MW . | 63,8 | 82,6 | $6 \mathrm{I}, \mathrm{I}$ | 60,7 | 75,0 | 66,7 | 62,6 | 7 | 67,5 | 60,7-82,6 |
| HL | 42,3 | 60,0 | 46,3 | 44,3 | 58,3 | 44,4 |  |  |  |  |
| HW | 42,3 | 56,5 | 44,2 | 37,9 | 43,3 | 44,4 | 46, 1 | 7 | 45,0 | 37,9-56,5 |
| FL | I I 5,4 | 124,3 | 106,3 | 103,6 | I 16,7 | 88,9 |  |  |  |  |
| FW | 15,4 | 21,7 | 13,7 | 17,9 | 20,8 | 18,5 | I 1, 3 | 7 | 17,0 | 11,3-21,7 |
| AL I | 50,0 | 63,5 | 54,7 | 42,9 | 58,3 | 40,7 | 47,8 | 7 | 51, I | 40,7-63,5 |
| AL II | 50,0 | 64,3 | 58,9 | 46,4 | 58,3 | 44,4 | 48,7 | 7 | 53,0 | 44,4-64,3 |
| AL III | 55,4 | 65,2 | 57,9 | 46,4 | 66,7 | 48, I | 56,5 | 7 | 56,6 | 46,4-66,7 |
| AL IV | 50,0 | 68,7 | 49,5 | 45,0 | 52,5 | 45,9 | 47,8 | 7 | 51,3 | 45,0-68,7 |
| TL | 161,5 | 234,8 | 194,7 | 107, 1 | 140,8 | 139,3 | 113,0 |  |  |  |
| Tcl | 44,6 | 55,7 | 42, I | 38,6 | 29,2 | 25,9 |  | 6 | 39,4 | 25,9-55,7 |

Table 36. Sepia papillata shells (form A). Relative dimensions as \% shell length.


Table 37. Sepia papillata shells (form B). Relative dimensions as \% shell length.


Table 38. Sepia simoniana males. Relative dimensions as \% MLd.

|  | A30127 | $\mathrm{A}_{31251}$ |  | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm. | 116 | 141 | 144 |  |  |  |
| MLv | 92,2 | 91,5 | 87,5 |  |  |  |
| MW | 62,9 | 63, I | 57,6 | 3 | 61,2 | 57,6-63, |
| HL | 49, 1 | 43,3 | 44,4 |  |  |  |
| HW | 37, 1 | 49,6 | 42,4 | 3 | 43,0 | 37,1-49,6 |
| FL | 94,8 | ${ }_{11} 13,5$ | 109,0 |  |  |  |
| FW | - | 15,6 | 16,0 | 2 | ${ }^{15} 5$ | ${ }^{15,6-16,0}$ |
| AL I | 62,9 | 59,6 | 55,6 | 3 | 59,4 | 55,6-62,9 |
| AL II | 63,8 | 66,o | 63,2 | 3 | 64,3 | 63,2-66,0 |
| AL III | 73,3 | 70,9 | 68,8 | 3 | 71,0 | 68,8-73,3 |
| AL IV | 60,3 | 53,2 | 54,9 | 3 | 56,1 | 53,2-60,3 |
| TL ${ }^{\text {Rt }}$ | - | 177,3 | 172,2 |  |  |  |
| $\mathrm{TL}_{\text {Lt }}$ | - | 173,8 | 160,4 |  |  |  |
| Tcl | - | 70,2 | 63,2 | 2 | 66,7 | 63,2-70,2 |

Table 39. Sepia simoniana females. Relative dimensions as \% MLd.

|  | A30132 | A30133 | A30134 | A30135 | Adam \& Rees 1966 | $\begin{gathered} \text { Massy } \\ 1925 \end{gathered}$ | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLdin mm | 174 | 172 | 139 | III | $\begin{array}{llllll}180 & 165 & 162140\end{array}$ | 147 |  |  |  |
| MLv | 87,4 | 92,4 | 87,8 | 87,4 | $\begin{array}{llll}83 & 90 & 87 & 84\end{array}$ |  |  |  |  |
| MW | 53,4 | 57,6 | 57,6 | 60,4 | $\begin{array}{lllll}53 & 58 & 59 & 56\end{array}$ |  | 8 | 56,9 | $53-60,4$ |
| HL | 46,o | 43,6 | 40,3 | 37,8 | $\begin{array}{lllll}42 & 51 & 44 & 49\end{array}$ |  |  |  |  |
| HW | 40,2 | 47,7 | 41,7 | 45,0 |     <br> 1 39 $4^{1}$ $4^{\text {I }}$ | 39,5 | 9 | $4^{1,8}$ | $39-47,7$ |
| FL. | 100,0 | 116,3 | 105,8 | 100,0 | $\begin{array}{llll}94 & 97 & 99 & \pm 93\end{array}$ |  |  |  |  |
| FW | 10,9 | ${ }^{15}$, I | 10,8 | 9,0 | $11-12 \pm 8,5$ |  | 6 | 11,5 | 9,0-15, 1 |
| AL I | 43,7 | 52,3 | 39,6 | 38,7 | 47 - 4039 | 45,6 | 8 | 43,2 | 38,7-52,3 |
| AL II | 43, I | 53,5 | 41,0 | 41,4 | 44 - 4043 | 40,8 | 8 | 43,4 | $40-53,5$ |
| AL III | 44,3 | 55,8 | 45,3 | 46,8 | $50-434^{6}$ | 54,4 | 8 | 48,2 | $43-55,8$ |
| AL IV | 41,4 | 54,1 | 43,2 | 41,4 | 47 - 4343 | 51,0 | 8 | 45,5 | 41,4-54,1 |
|  | 103,4 | 142,4 | 89,9 | 101,8 | 122 - 99 - | 187, I |  |  |  |
| ${ }^{\text {Lt }}$. | 100,6 | 127,9 | 73,4 | 111,7 | $122-99-$ |  |  |  |  |
| Tcl | 48,9 | 55,2 | 51,8 | 51,4 | $49-56-$ | 60,5 | 7 | 53,3 | 48,9-60,5 |

## See opposite for Table 40

Table 41. Sepia angulata shells. Relative dimensions as \% shell length.

|  | L in mm | W | Th | Str z |
| :---: | :---: | :---: | :---: | :---: |
| $\mathrm{A}_{31317}$ (Holotype) . | 60 | 56,7 | 13,3 | 51,7 |
| A31395 | 57 | 57,9 | 14,0 | 47,4 |
|  | 61 | 50,8 | 13,1 | 47,5 |
|  | $\pm 4^{6}$ | 56,5 | 13,0 | 52,2 |
| A31318 | 66 | 53,0 | 15,2 | 50,0 |
|  | $\pm 75$ | 56,o | 14,0 | 50,5 |
|  | 25 | 60,0 | 16,0 | 60,0 |
| $\mathrm{A}_{31319}$ | 63 | 57, 1 | 11,9 | 60,3 |
|  | 67 | 52,2 | 11,9 | 61,2 |
| A31320 | $4^{1}$ | 56, 1 | 12,2 | 58,5 |
|  | 35 | 60,0 | 14,3 | 54,3 |
|  | 34 | 52,9 | 11,8 | 58,8 |
| N . |  | 12 | 12 | 12 |
| Mean |  | 55,8 | 13,4 | 54,4 |
| Range |  | 50,8-60,0 | 11,8-16,0 | 47,4-61,2 |

Table 40. Sepia simoniana shells. Relative dimensions as \% shell length.


Table 42. Sepia hieronis males. Relative dimensions as \% MLd.

|  | A29728 | $A_{301} 6$ | A30563 | A31243 | $\mathrm{A}_{3}{ }^{4} \mathrm{O} 5$ | $\mathrm{A}_{3}{ }_{4}{ }^{06}$ | $A_{31407}$ | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm . | 59 | 54 | 62 | 61 | 47 | 56 | 48 |  |  |  |
| MLv . | 69,5 | 70,4 | 75,8 | 82,0 | 83,0 | 76,8 | 93,8 |  |  |  |
| MW | 49,2 | 55,6 | 56,5 | 65,6 | 70,2 | 55,4 | 62,5 | 7 | 59,3 | 49,2-70,2 |
| HL | 37,3 | 37,0 | 37, 1 | 50,8 | 44,7 | 44,6 | 45,8 |  |  |  |
| HW | 44, I | 42,6 | 43,5 | 50,8 | 46,8 | 37,5 | 54,2 | 7 | 45,6 | 37,5-54,2 |
| FL | 78,0 | 79,6 | 82,3 | 90,2 | 78,7 | 80,4 | 83,3 | 7 |  | 37,5 54, |
| FW | 10,2 | 5,6 | 6,5 | 9,8 | 8,5 | 7, I | 8,3 | 7 | 8,0 | 5,6-10,2 |
| AL I | 30,5 | 26,0 | 38,7 | 55,7 | 34,0 | 39,3 | 41,7 | 7 | 38,0 | 26,0-55,7 |
| AL II | 33,9 | 26,0 | 35,5 | 62,3 | 42,6 | 37,5 | 47,9 | 7 | 40,8 | 26,0-62,3 |
| AL III | 37,3 | 31,5 | 38,7 | 60,7 | 46,8 | 41, I | 58,3 | 7 | 44,9 | 31,5-60,7 |
| AL IV | 49,2 | 46,3 | 45,2 | 68,9 | 48,9 | 35,7 | 52, I | 7 | 49,5 | 35,7-68,9 |
| TL | 127, I | 101,9 | 90,3 | I 1 4, 8 | 142,6 | 121,4 | 185,4 |  |  |  |
| Tcl | 1 1,9 | 14,8 | 14,5 | 13, 1 | 12,8 | 14,3 | 14,6 | 7 | 13,7 | 11,9-14,8 |

Table 43. Sepia hieronis females. Relative dimensions as \% MLd.

|  | $\mathrm{A}_{30145}$ |  <br> Rees 1966 | N | Mean |
| :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 33 | 61 |  |  |
| MLv | 78,8 | 74 |  |  |
| MW | 60,6 | 54 | 2 | 57,3 |
| HL . | 48,5 | 38 |  |  |
| HW | 51,5 | 51 | 2 | 51,3 |
| FL | 84,8 | 85 |  |  |
| FW | 9, I | 8 | 2 | 8,6 |
| AL I | 18,2 | 38 | 2 | 28, I |
| AL II | 24,2 | 38 | 2 | $3 \mathrm{I}, 1$ |
| AL III | 30,3 | 38 | 2 | 34,2 |
| AL IV | 45,5 | 38 | 2 | 41,8 |
| TL | 157,6 | - |  |  |
| Tcl | 12, I | 10 | 2 | I I, I |

Table 44. Sepia hieronis shells. Relative dimensions as \% shell length.

|  |  | L in mm | W | Th | Str z |
| :---: | :---: | :---: | :---: | :---: | :---: |
| West coast | A29728 | 60 | 38,3 | 11,7 | 70,0 |
|  | A30146 | 52 | $4 \mathrm{C}, 4$ | 9,6 | 73,1 |
|  | A30563 | 62 | 38,7 | 11,3 | $\pm 72,6$ |
|  | A31243 | 67 | 38,8 | 11,2 | 73,1 |
| East coast | $\mathrm{A}_{31405}$ | 46 | 45,7 | 15,2 | 65,2 |
|  | $\mathrm{A}_{31406}$ | 54 | 40,7 | 14,8 | 66,7 |
|  | $\mathrm{A}_{314}{ }^{2} 7$ | 54 | 40,7 | 13,0 | 64,8 |
| N |  |  | 7 | 7 | 7 |
| Mean |  |  | $4^{0,5}$ | 12,4 | 69,4 |
| Range |  |  | 38,3-45,7 | 9,6-15,2 | 64,8-73, 1 |

Table 45. Sepia insignis female. Relative dimensions as \% MLd.

|  | A31247 |
| :---: | :---: |
| MLd in mm . | 44 |
| MLv . | 90,9 |
| MW . | $\pm 52,3$ |
| HL | 27,3 |
| HW | 45,5 |
| FL | 95,5 |
| FW | 6,8 |
| AL I | 36,4 |
| AL II | 36,4 |
| AL III | 29,5 |
| AL IV | 45,5 |
| TL ${ }^{\text {Rt }}$ | 68,2 |
| ${ }^{\text {L }}$ L | 75,0 |
| Tcl | 13,6 |

Table 46. Sepia insignis shells. Relative dimensions as \% shell length.

|  | L in mm | W | Th | Str z |
| :---: | :---: | :---: | :---: | :---: |
| A30486 | 29 | 34,5 | 10,3 | 55,2 |
| A31236 | 29 | 37,9 | 10,3 | 55,2 |
| A31241 | $\begin{aligned} & 34 \\ & 17,5 \end{aligned}$ | $\begin{aligned} & 35,3 \\ & 37, \mathbf{I} \end{aligned}$ | 8,8 | $\begin{aligned} & 61,8 \\ & 60,0 \end{aligned}$ |
| A $_{312} 247$ | 44 | 31,8 | 11,4 | 63,6 |
| Still Bay . | $\pm 32$ | 31,3 | 10,9 | 65,6 |
| Adam \& Rees 1966 . | 26 | 33 | 16,5 | 65 |
| N . |  | 7 | 6 | 7 |
| Mean |  | 34,4 | 11,4 | 60,9 |
| Range |  | 31,3-37,9 | 8,8-16,5 | 55,2-65,6 |

Table 47. Sepia typica males. Relative dimensions as \% MLd.


Table 48. Sepia typica females. Relative dimensions as \% MLd.

|  | A29608 |  |  | A29717 |  |  |  |  |  | A29783 |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLdin mm | 22 | 21 | 18 | 9 | 10 | 12 | 8 | 10 | 11 | 23 | 25 | 19 |
| MLv | 93,2 | 100,0 | 94,4 | 88,9 | 90,0 | 83,3 | 75,0 | 80,o | 72,7 | 91,3 | 92,0 | 89,5 |
| MW | 72,7 | 76,2 | 77,8 | 88,9 | 80,0 | 75,0 | 75,0 | 70,0 | 72,7 | 82,6 | 64,0 | 89,5 |
| HL | 63,6 | 81,o | 61, 1 | 66,7 | 70,0 | 66,7 | , | 60,0 | 63,6 | 56,5 | 48,0 | 57,9 |
| HW | 54,5 | 57, I | 55,6 | 66,7 | 70,0 | 66,7 | - | 60,0 | 63,6 | 52,2 | 48,0 | 57,9 |
| FL. | 113,6 | 109,5 | 105,6 | 100,0 | 100,0 | 100,0 | - | 110,0 | 90,9 | 113,0 | 104,0 | 110,5 |
| FW | 9, 1 | 9,5 | I I, I | - | - | 12,5 | - | - | - | 10,9 | 12,0 | 10,5 |



|  | A29783 (cont.) |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 14 | 15 | 15 | 15 | 16 | 15 | 16 | 18 | 17 | 14 | 18 | 19 |
| MLv | 85,7 | 86,7 | 93,3 | 86,7 | 87,5 | 93,3 | 93,8 | 88,9 | 88,2 | 92,9 | 83,3 | 94,7 |
| MW | 78,6 | 73,3 | 80,0 | 86,7 | 81,3 | 80,0 | 81,3 | 77,8 | 82,4 | 85,7 | 77,8 | 84,2 |
| HL | 57, I | 53,3 | 60,0 | 66,7 | 62,5 | 66,7 | 62,5 | 66,7 | 64,7 | 64,3 | 66,7 | 68,4 |
| HW | 57, I | 60,0 | 60,0 | 60,0 | 56,3 | 60,0 | 56,3 | 50,0 | 58,8 | 57, 1 | 55,6 | 52,6 |
| FL | 107, 1 | 93,3 | 106,7 | 106,7 | 1 12,5 | 100,0 | 106,3 | 100,0 | III, 8 | 114,3 | 105,6 | 100,0 |
| FW | 7, 1 |  | - | - | - | - | 18,8 | 8,3 | I 1, 8 | 7, I | 13,9 | - |


|  | A29783 (cont.) |  | $A_{30176}$ |  | A30177 |  |  |  | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 18 | 17 | 18 | 15 | 19 | 20 | 15 | 15 |  |  |  |
| MLv | 88,9 | 94, I | 94,4 | 86,7 | 100,0 | 78,9 | 93,3 | 93,3 |  |  |  |
| MW | 77,8 | 82,4 | 72,2 | 73,3 | 78,9 | 75,0 | 80,0 | 86,7 | 44 | 79,3 | 64,0-90,0 |
| HL . | 61, 1 | 64,7 | 55,6 | 66,7 | 63,2 | 55,0 | 66,7 | 66,7 |  |  |  |
| HW | 55,6 | 58,8 | 55,6 | 60,0 | 52,6 | 60,0 | 60,0 | 66,7 | 43 | 59,0 | 48,0-70,0 |
| FL. | 105,6 | 105,9 | 100,0 | 66,7 | 110,5 | 110,0 | 113,3 | - |  |  |  |
| FW . |  | 5,9 | - | 10,0 | - | - |  | - | 17 | 10,9 | 5,9-18,8 |

Table 49. Sepia typica shells. Relative dimensions as \% shell length.

|  | L in mm | W | Th | Str z |
| :--- | :---: | :---: | :---: | :---: |
| A29608 . . . . | 19,5 <br> A1 | 53,8 <br> 57,1 | - | 1, <br> Adam \& Rees 1966 . |
| Mean . . . . |  | 55,5 |  | - |

Table 50. Sepia robsoni, S. faurei and S. dubia. Relative dimensions as \% MLd (or shell length).


* Taylor, personal communication.

Table 5I. Sepiella cyanea males. Relative dimensions as \% MLd.

|  | A6526 | Adam \& Rees 1966 |  |  |  |  |  | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MLd in mm | 62 | $74 \quad 75$ | 56 | 55 | 53,5 | 50 | 48 |  |  |  |
| MLv | 75,8 | 74 81 | 88 | 87 | 86 | 86 | 86 |  |  |  |
| MW | 40,3 | $45 \quad 41$ | 57 | 56 | 60 | 58 | 60 | 8 | 52,2 | 40,3-60 |
| HL. | 19,4 | $23 \quad 24$ | 25 | 25 | 26 | 26 | 27 |  |  |  |
| HW | 41,9 | 4139 | 52 | 49 | 49 | 52 | 50 | 8 | 46,7 | 39-52 |
| FL | 80,6 | 9385 | 100 | 98 | 98 | 98 | 92 |  |  |  |
| FW | 8, I | I5 9,5 | 19,5 | 18 | 20 | 20 | 12,5 | 8 | 15,3 | 8,1-20 |
| AL I | 35,5 | $4^{1} \quad 41$ | 36 | 35 | 36 | 38 | 40 | 8 | 37,8 | $35-41$ |
| AL II | 33,9 | $41{ }^{1}$ | 36 | 36 | 36 | 38 | 40 | 8 | 37,7 | 33,9-41 |
| AL III | 41,9 | $47 \quad 45$ | 46 | 44 | 43 | $4^{8}$ | 50 | 8 | 45,6 | 41,9-50 |
| AL IV | 46,8 | 6 I 55 | 54 |  | 50 | 54 | 56 | 8 | 53,2 | 46,8-61 |
| TL. |  | 8ı 106 |  |  |  |  | - |  |  |  |
| Tcl | - | $19 \quad 17,5$ | 23 | 22 | 20 | 24 | 21 | 7 | 20,9 | 17,5-24 |

Table 52. Sepiella cyanea females. Relative dimensions as \% MLd.

| MLd in mm . | Adam \& Rees 1966 |  |  |  |  |  |  | N | Mean | Range |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 75 | 71 | 69 | 37 | 37 | 37 | 29 |  |  |  |
| MLv . | 80 | 84 | 86 | 87 | 87 | 84 | 89 |  |  |  |
| MW | 53 | 45 | 52 | 65 | 59 | 65 | 64 | 7 | 57,6 | $45-65$ |
| HL | 24 | 27 | 25 | 27 | 27 | 27 | 31 |  |  |  |
| HW | 40 | 42 | 49 | 51 | 51 | 51 | 55 | 7 | 48,4 | $40-55$ |
| FL | 87 | 95 | 100 | 100 | 100 | 95 | 91 |  |  |  |
| FW | 13,5 | 14 | 19 | 19 | 19 | 16 | 12 | 7 | 16, I | $12-19$ |
| AL I . | 36 | 38 | 29 | 27 | 27 | 27 | 28 | 7 | 30,3 | $27-38$ |
| AL II | 33 | 39 | 29 | 27 | 27 | 27 | 28 | 7 | 30,0 | $27-39$ |
| AL III | 36 | 39 | 30 | 27 | 27 | 27 | 31 | 7 | 31,0 | $27-39$ |
| AL IV | 47 | 48 | $3^{8}$ | 35 | $3^{8}$ | 38 | 35 | 7 | 39,9 | $35-48$ |
| TL | 146 | - | - | - | - | - | - |  |  |  |
| Tcl | 22,5 | 22,5 | 24,5 | - | 24 | - | - | 4 | 23,4 | 22,5-24,5 |

Table 53. Sepiella cyanea shells. Relative dimensions as \% shell length.

|  | Sex | L in mm | W | Th | Str z |
| :---: | :---: | :---: | :---: | :---: | :---: |
| A6526 . . . | ठ | 61 | 31, I | 1 1,5 | 50,8 |
| Adam \& Rees 1966 | ${ }^{\circ}$ | 69 | 29 | 10 | 58 |
|  | ${ }^{*}$ | 54 | 33 | 14 | 56 |
|  | 아 | 74 | 30,5 | 11 | 51 |
|  | 아 | 70 | 31,5 | 12 | 53 |
|  | 아 | 69 | 35,5 | 13 | 58 |
| A6526 . . . |  | 77 | 33,8 | 11,7 | 54,5 |
|  | ? | 76 | 32,9 | 11,8 | 52,6 |
|  | ? | 62 | 32,2 | 11,3 | 54,8 |
|  | ? | $\pm 75$ | 30,7 | 12,0 | 61,3 |
| Adam \& Rees 1966 | ? | 80 | 32,5 | 12 | 60 |
|  | ? | 76 | 31,5 | 12,5 | 58 |
| $\mathrm{N}_{\text {total }}$.$\mathrm{Mean}_{\text {total }}$$\mathrm{Range}_{\text {total }}$. |  |  | 12 | 12 | 12 |
|  |  |  | 32,0 | I 1,9 | 55,7 |
|  |  |  | $29-35,5$ | $10-14$ | 50,8-61,3 |
|  |  |  |  |  |  |
|  |  |  | 31,0 | 11,8 | $54,9$ |
|  |  |  |  | 10-14 | 50,8-58 |
| $\begin{array}{ll}\mathrm{N}_{¢} \\ \mathrm{Mean}_{¢} \\ \mathrm{Range}_{+} & \cdot \\ \\ \end{array}$ |  |  | 3 | 3 | 3 |
|  |  |  | $32,5$ | 12 | $54$ |
|  |  |  | $30,5-35,5$ | II -13 | $5^{1} \quad-58$ |



Sepia zanzibarica Pfeffer, A2141: a. Dorsal and b. ventral views of shell.
Sepia officinalis vermiculata Quoy \& Gaimard, $\%$ from Durban: c. Dorsal and d. ventral views of shell.
Scale $=20 \mathrm{~mm}$.


Sepia officinalis vermiculata Quoy \& Gaimard,, , A30183: a. Dorsal and b. ventral views of shell (posterior spine broken).
Sepia officinalis hierredda Rang, ơ, A31291, from Baia de Cabo Negro, Angola: c. Dorsal and d. ventral views of shell of comparable size to the above.

Scale $=100 \mathrm{~mm}$.


Sepia acuminata Smith, ${ }^{\text {on }}$, A31398: a. Dorsal and b. ventral views of shell. Sepia confusa Smith, A2140: c. Dorsal and d. ventral views of shell.

$$
\text { Scale }=10 \mathrm{~mm} .
$$



Sepia incerta Smith, N.M.970: a. Ventral view of shells described by Massy (1925: 219) as $S$. incerta.
Sepia incerta Smith, N.M.958: b. Ventral view of shells described by Massy (1925: 215) as S. burnupi.

Sepia burnupi Hoyle, A2147: c. Dorsal and d. ventral views of shell (anterior tip broken).
Scale $=10 \mathrm{~mm}$.


Sepia incerta Smith, from Punta Zavora: a. Dorsal and b. ventral views of shell.
Sepia tuberculata Lamarck, from Cape Agulhas: c. Dorsal and d. ventral views of shell with exceptionally long striated zone.

Scale $=10 \mathrm{~mm}$.


Sepia australis Quoy \& Gaimard, from Still Bay: a. Dorsal and b. ventral views of shell. Sepia tuberculata Lamarck, $9, \mathrm{~A}_{3} \mathrm{O}_{5} 11$ : c. Dorsal and d. ventral views of shell with normal striated zone.
Scale $=10 \mathrm{~mm}$.


Sepia papillata Quoy \& Gaimard, shell form A, from Milnerton beach: a. Dorsal and b. ventral views of shell.
Sepia papillata Quoy \& Gaimard, shell form B, đ̛, A30120: c. Dorsal and d. ventral views of shell. Scale $=10 \mathrm{~mm}$.


Sepia simoniana Thiele, from Still Bay: a. Dorsal and b. ventral views of shell. Sepiella cyanea Robson, ${ }^{7}$, A6526: c. Dorsal and d. ventral views of shell.

Scale $=10 \mathrm{~mm}$.


Sepia hieronis (Robson), ${ }^{\text {tr }}$, A29728 (west coast): a. Dorsal and b. ventral views of shell. Sepia hieronis (Robson), ơ, A31407 (east coast): c. Dorsal and d. ventral views of shell.


Sepia insignis Smith, A3124I: a. Dorsal and b. ventral views of shell. c. Ventral view of part of large shell from Bloubergstrand.
Sepia angulata f. sp.: d. Median view of half shell, cut longitudinally to show angle between striated zone and smooth zone.

Scale $=10 \mathrm{~mm}$.


Sepia angulata n. sp., holotype, A31317: a. Dorsal and b. ventral views of shell. Sepia angulata n. sp., paratype, A31320: c. Dorsal and d. ventral views of shell.

[^15]
## INSTRUCTIONS TO AUTHORS

## Based on

CONFERENCE OF BIOLOGICAL EDITORS, COMMITTEE ON FORM AND STYLE. I960.
Style manual for biological journals. Washington: American Institute of Biological Sciences.

MANUSCRIPT
To be typewritten, double spaced, with good margins, arranged in the following order: (1) Heading, consisting of informative but brief title, name(s) of author(s), address(es) of author(s), number of illustrations (plates, figures, enumerated maps and tables) in the article. (2) Contents. (3) The main text, divided into principal divisions with major headings; subheadings to be used sparingly and enumeration of headings to be avoided. (4) Summary. (5) Acknowledgements. (6) References, as below. (7) Key to lettering of figures. (8) Explanation to plates.

## ILLUSTRATIONS

To be reducible to $12 \mathrm{~cm} \times 18 \mathrm{~cm}$ ( 19 cm including caption). A metric scale to appear with all photographs.

## REFERENGES

Harvard system (name and year) to be used: author's name and year of publication given in text; full references at the end of the article, arranged alphabetically by names, chronologically within each name, with suffixes $a, b$, etc. to the year for more than one paper by the same author in that year.
For books give title in italics, edition, volume number, place of publication, publisher.
For journal articles give title of article, title of journal in italics (abbreviated according to the World list of scientific periodicals. 4th ed. London: Butterworths, 1963), series in parentheses, volume number, part number (only if independently paged) in parentheses, pagination.

## Examples (note capitalization and punctuation)

Bullough, W. S. 196o. Practical invertebrate anatomy. 2nd ed. London: Macmillan.
Fischer, P.-H. 1948. Données sur la résistance et de le vitalité des mollusques. 7. Conch., Paris 88: 100-140.
Fischer, P.-H., Duval, M. \& Raffy, A. 1933. Etudes sur les échanges respiratoires des littorines. Archs Zool. exp. gén. 74: 627-634.
Kohn, A. J. 1960a. Ecological notes on Conus (Mollusca: Gastropoda) in the Trincomalee region of Ceylon. Ann. Mag. nat. Hist. (13) 2: 309-320.
Kонл, A. J. 196ob. Spawning behaviour, egg masses and larval development in Conus from the Indian Ocean. Bull. Bingham oceanogr. Coll. 17 (4): 1 -51.
Thiele, J. igio. Mollusca: B. Polyplacophora, Gastropoda marina, Bivalvia. In schultze, l. Zoologische und anthropologische Ergebnisse einer Forschungsreise im westlichen und zentralen SüdAfrika. 4: 269-270. Jena: Fischer. Denkschr. med-naturw. Ges. Jena 16: 269-270.

## zoological nomenclature

To be governed by the rulings of the latest International code of zoological nomenclature issued by the International Trust for Zoological Nomenclature (particularly articles 22 and 51). The Harvard system of reference to be used in the synonymy lists, with the full references incorporated in the list at the end of the article, and not given in contracted form in the synonymy list.

## Example

Scalaria coronata Lamarck, 1816: pl. 451, figs 5a, b; Liste: II. Turton, 1932: 80.

ANNALS OF THE SOUTH AFRICAN MUSEUM ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM

| Volume | $\mathbf{5 9}$ | Band |
| ---: | :---: | :--- |
| September | $\mathbf{1 9 7 2}$ | September |
| Part | II | Deel |



## DISCARD THE NAMES

THERIODONTIA AND ANOMODONTIA:
A NEW CLASSIFICATION OF THE THERAPSIDA

By<br>L. D. BOONSTRA

## The ANNALS OF THE SOUTH AFRICAN MUSEUM

are issued in parts at irregular intervals as material becomes available

Obtainable from the South African Museum, P.O. Box 6I, Cape Town

## Die ANNALE VAN DIE SUID-AFRIKAANSE MUSEUM

word uitgegee in dele op ongereelde tye na beskikbaarheid van stof

Verkrygbaar van die Suid-Afrikaanse Museum, Posbus 6i, Kaapstad

> OUT OF PRINT/UIT DRUK
> 1, $2(1,3,5,7-8), 3(1-2,5$, t.-p.i. $), 51-2,5,7-9)$, 6(1, t.-p.i.), 7(1-3), 8, 9(1-2), 10(1), II(1-2, 5, 7, t.-p.i.), 24(2), 27, 31(1-3), 33

Price of this part/Prys van hierdie deel
R2,10

Trustees of the South African Museum (C) Trustees van die Suid-Afrikaanse Museum 1972

Printed in South Africa by The Rustica Press, Pty., Ltd, Court Road, Wynberg, Cape

In Suid-Afrika gedruk deur
Die Rustica-pers, Edms., Bpk.
Courtweg, Wynberg, Kaap

# DISCARD THE NAMES THERIODONTIA AND ANOMODONTIA: A NEW CLASSIFICATION OF THE THERAPSIDA 

By<br>L. D. Boonstra<br>South African Museum, Cape Town

(With 2 figures)
[MS. accepted 25 Fanuary 1972]

## Contents



## Introduction

Since the discovery of certain Permian reptiles in Russia and South Africa in the early thirties of the 19th century showing some characters of a mammalian nature and their first descriptions by Kutorga in 1838 and Owen in 1844 numerous attempts have been made by various authors to fit them into the taxonomic system in such a way as to indicate in what manner these reptiles could be considered related and ancestral to the mammals.

I have found the historical study of the various classifications proposed by authors most interesting and illuminating and was tempted to publish a detailed historical review, but on second thoughts have decided that confining myself to the essentials of the ever increasing precision of the phylogenetic views would be more important and valuable and satisfying.

At the present time we know over 300 recorded genera of 'reptiles' which possess to varying degrees characters indicating a development in mosaic pattern in a general mammalian direction.

For this assemblage of 'reptiles' we have the name Therapsida coined by Broom in 1905.

In the present state of our knowledge this group as a whole appears to have evolved from captorhinomorph and sphenacodontid precursors.

The oldest known therapsids, from low down in the Permian, consist of an assemblage in which discrete lines of development are clearly evident.

For these lines of development we have the following denominations available:
I. Anomodontia (Owen 1859)
2. Dicynodontia (Owen 1860)
3. Cynodontia (Owen 1860)
4. Theriodontia (Owen 1876)
5. Dinocephalia (Seeley 1895)
6. Gorgonopsia (Seeley 1895)
7. Therocephalia (Broom 1903)
8. Scaloposauria (Boonstra 1953)
9. Phthinosuchia (Romer 1961)
10. Eotitanosuchia (Boonstra 1963)

There exists strong evidence that the Dinocephalia and Gorgonopsia evolved through the Eotitanosuchia from common sphenacodontid ancestors of the morphological habit such as that of the genus Haptodus. Taxonomically there may thus be some reason to coin a higher denomination to include the Eotitanosuchia, Gorgonopsia and Dinocephalia in order to indicate their consanguinity. Both these sublines of development, each showing the development of certain mammalian characters, became extinct-the Dinocephalia at the end of the Middle Permian and the Gorgonopsia at the end of the Upper Permian.

The dinocephalian line shows certain stages of development. The most primitive stage is represented by a group of animals for which the name Eotitanosuchidae has been used by Tchudinov ( 1960 ).

Ascending directly from the Eotitanosuchidae is the group Brithopodidae (Efremov 1954). From the Brithopodidae two higher groups arose dichotomously, viz. the Anteosauridae (Boonstra 1954) and the Titanosuchidae (Broom 1903). From the Titanosuchidae three higher stages diverged, viz. the Tapinocephalidae (Owen 1876), the Styracocephalidae (Haughton 1929) and the Estemmenosuchidae (Tchudinov 1960).

The Gorgonopsia line shows the following stages of development represented by groups with the following denominations:

Phthinosuchidae (Efremov 1954)
Hipposauridae (Watson \& Romer 1956)
Gorgonopsidae (Lydekker 1890)
Burnetiidae (Broom 1923)
The dicynodontian line arising from unknown but probably sphenacodont ancestors has as its oldest known representative the genus Otsheria from which I have proposed the group name Otsheriidae (Boonstra 1963). Diverging from this base there are the short-lived groups Venyukoviidae (Efremov 1940) and Galeopsidae (Broom 1912) and the longer lived but also sterile line of the Dicynodontia (Owen 1860) which split up into the Endothiodontidae (Lydekker 1890) and Dicynodontidae (Owen 1876) from which arose the Kistecephalidae (Seeley 1895), Lystrosauridae (Broom 1903) and the Kannemeyeriidae (Von Huene 1948).

The third line, first encountered in the Middle Permian is that of the Therocephalia (Broom 1903). Arising from as yet unknown but probably
sphenacodont ancestors they formed an important group of carnivores during Tapinocephalus zone times, when they had already developed diverging branches with the following denominations:

Pristerognathidae (Broom 1906)
Lycosuchidae (Broom 1910)
Alopecodontidae (Broom 1932)
This line, strongly developed in the Middle Permian, became extinct at the end of the Upper Permian with the last off-shoots represented by the Whaitsiidae (Haughton 1918) and the Euchambersiidae (Broom 1931).

The last line, also beginning in the Middle Permian, with a few inadequately known forms, is that of the Scaloposauria (Boonstra 1953). This line may have arisen from the therocephalian line during the Lower Permian and is certainly closely related. Both the therocephalian and scaloposaurian lines apparently arose from some earlier sphenacodonts, but we have no certain indication of this as we have in the first line where a Haptodus-like form is indicated.

Commencing in the Cistecephalus zone and continuing to the top of the Cynognathus zone we find the Bauriamorpha (Watson 1917) which are generally considered to have arisen from the earlier ictidosuchian Scaloposauria.

Also commencing in the Cistecephalus zone and continuing into the Red Beds we have a final branch-the Cynodontia (Owen 1860) culminating in the near-mammals - the Tritylodontia (Simpson 1925). If the Cynodontia are not a parallel branch to the Scaloposauria, both to be derived from primitive therocephalians, then one must postulate a direct and separate derivation from some earlier sphenacodont.

From the foregoing it is clear that I think that we have three main branches of Therapsida.

An older view was that there were two main branches which have been labelled:

Anomodontia (Owen 1859) and
Theriodontia (Owen 186o)

## Anomodontia

Although Owen initially in 1859 - 60 clearly intended the terms Anomodontia and Dicynodontia to have as type, Dicynodon, he later included some theriodonts.

Since then the term Anomodontia has had a chequered career, being used by authors to include a variety of other forms manifestly un-Dicynodon-like. The term Anomodontia being thus misused it would be pragmatic to drop it altogether and rather retain the name Dicynodontia solely for those forms showing a Dicynodon-like structure as originally intended by Owen.

## Theriodontia

In 186o Owen coined the term Cynodontia with Galesaurus as the type and included it as a 'family' of his Anomodontia. In 1876 Owen introduced the
term Theriodontia for the same genera included in his former Cynodontia, apparently to supersede the latter name. As Owen included genera now considered as Gorgonopsia and others as Cynodontia in his Theriodontia, and Watson recently also the Titanosuchia, which three groups are now known to lie on different lines of development, the term Theriodontia unites incompatible groups and should be dropped.

Deciding to discard the names Anomodontia and Theriodontia because they each bracket together lines of development which, as I have indicated, are not nearly related, it appears necessary to coin three new names for the three main branches of the Therapsida, and to include as subdivisions of each of these three new denominations those groups which are in fact closely related.

For the first of these branches I propose the name Alphatherapsida to include the subdivisions Eotitanosuchia, Dinocephalia and Gorgonopsia.

For the second branch-Betatherapsida-to include only those forms related to Dicynodon.

For the third branch the name-Gammatherapsida-to include those fertile groups directly related and finally leading to the first mammals, viz. Therocephalia, Scaloposauria and Cynodontia.

## Diagnoses

## ALPHATHERAPSIDA

Dentition primitively carnivorous with pointed incisors, canines and postcanines adapted for snatching and tearing out flesh without cutting or chewing. Short-lived side branches with dentition transformed to herbivorous talon and heel teeth adapted for piercing and crushing without cutting and chewing.

Choanae anteriorly situated with air passage without bony ventral floor to partition it off during feeding.

The temporalis primitively originating from under surface of skull roof and inserted in the adductor fossa and on the upper edge of the dentary.

In the gorgonopsian branch the insertion was improved by the precocious development of a coronoid process on the dentary. Here we thus have a primitive origin coupled with an advanced insertion.

In the dinocephalian branch the origin of the temporalis shifts away from the under surface of the skull roof to the lateral surface of the intertemporal bones, but no coronoid process is developed. Here we thus have an advanced origin coupled with a primitive insertion. The later pachyostosis bedevils this aspect. The postdentary bones of the mandible are persistently well developed due to their retaining the primitive insertion of the adductors which exerted a greater horizontal than vertical pull, with the joint a simple hinge. But in the herbivorous forms a fore-and-aft motion allows for a crushing bite.

Concomitant with the primitive carnivorous jaw-mechanism the locomotory apparatus is of a crawling habit with sprawling limbs and little upraising of the body and only slightly reduced digital segments.

In the braincase the sphenoidal complex is very well ossified, but the prootic is feebly ossified, thus leaving a wide gap in the lateral wall with a loose standing unwidened epipterygoid. In the pachyostotic Dinocephalia the gap in the lateral wall is greatly reduced, but the narrow epipterygoid remains uninvolved.

In the Alphatherapsida the gorgonopsian branch has its characteristic structures developed early and these are retained, with only insignificant variations, throughout its span of life, notwithstanding that they survived to the end of the Upper Permian.

The dinocephalian branch commencing as a primitive carnivorous group early in its history, develops herbivorous twigs but the whole branch is short lived and unprogressive and is soon cut short by the pathological pachyostosis.

## BETATHERAPSIDA

Dentition herbivorous, primitively with a series of marginal teeth, later with marginal teeth in part or wholly replaced by horny sheaths. Choanae shifted moderately posteriorly, with part of air passage with bony partition, separating it from buccal cavity, formed by plates of the premaxilla.

The jaw adductors highly specialized, particularly in their origins, with concomitant great lengthening of the temporal fossa and the development of a unique triradiate squamosal, and everted zygoma accompanied by a lengthened sliding articular facet allowing fore-and-aft sectorial movement of the jaw when feeding, insertions tending to shift on to the outer face of the dentary.

The postdentary bones unreduced.
Feeding on upland plants (except lystrosaurs and Kistecephalidae) the locomotor apparatus is adapted for a more upright walking gait, with an acromion process, greatly enlarged anterior iliac process, obturator foramen and reduced digital segments. In the braincase the sphenoidal complex is very well developed and situated far anteriorly; the prootic short, thus leaving a very long gap in the lateral wall, with a loose standing slender epipterygoid.

Notwithstanding its long span of life this group remained stationary on its early achieved developmental niveau. What variations arose were quite insignificant, initiating nothing phylogenetically fertile.

## GAMMATHERAPSIDA

Dentition primitively carnivorous, but variations commence quite early, viz. reduction of postcanines (in lycosuchids and whaitsiids); development of additional precanines (in alopecodontids and Scaloposauria); tricuspid sectorial postcanines arose (in some Scaloposauria and some Cynodontia); grinding surfaces developed on postcanines (in some Scaloposauria and Cynodontia); differentiation into 'premolars' and 'molars' (in Cynodontia). Thus the primitive snatching and tearing dentition became adapted to cutting and grinding with a process of chewing.

Primitively the choanae were anteriorly situated, but concomitant with
the developing of a chewing habit the choanae shifted backwards with the development of a secondary bony palate partitioning off the air passage from the buccal cavity during the process of mastication.

The temporalis originated from the lateral face of the intertemporal bones and inserted on the coronoid process.

Primitively, where the posterior mandibular bones are still well developed, the superficial masseter inserted on the reflected lamina of the angular and the internal pterygoid wrapped round the ventral edge of the angular. But in the cynodonts part of the superficial masseter inserted on the postero-ventral corner of the dentary and the internal pterygoid also partly moved on to the inner face of the corner of the dentary, which resulted in a reduction of the function and thus the size of the angular.

With the pull of the jaw muscles having strong horizontal components in the early forms the posterior mandibular bones remained strong to withstand the strain on the jaw joint. But with the jaw-closing muscles developing less horizontal and greater vertical pull the strain on the jaw joint decreased with a resulting decrease in size of the posterior mandibular bones.

With the increased strain on the dentary, due to its capturing some of the muscle insertions from the posterior bones, it developed a very large coronoid process and a prominent angle and, extending further and further posteriorly, in the final stages made contact with the squamosal to form a double jaw joint.

In the locomotor apparatus a more upright walking gait is developed with a reduction of the phalanges to $2,3,3,3,3$ in all but the early cynodonts where the $4^{\text {th }}$ and $5^{\text {th }}$ digits have 4 segments.

Primitively with pubic foramen, advanced with obturator foramen. Primitively without, advanced forms with, infra-spinatus fossa.

In the braincase the sphenoidal complex is primitively not well ossified, but moderately so in some advanced forms. Primitively the prootic is feebly developed, but in advanced forms extends anteriorly to meet the epipterygoid.

Primitively the epipterygoid is usually slender but broadened in the primitive lycosuchids. In some advanced forms it is greatly broadened and meeting the prootic enters into the sidewall of the braincase (cynodonts).

Postorbital bar primitively well developed as also in some advanced forms but in some others it becomes weak and even incomplete.

Occipital condyle primitively single (tripartite), in advanced forms double and formed by the exoccipitals. Primitively with suborbital fenestra, but sometimes reduced, large in Scaloposauria, absent in Cynodontia.

In contrast to the Alphatherapsida and Betatherapsida the Gammatherapsida were a very versatile group in which developments, besides leading into a number of early as well as later blind alleys, produced very progressive parallel branches all in a general mammalian direction, with the procynosuchid -galesaurid-tritylodont branch most probably including the actual ancestors of the first mammals.

## SYNAPSIDA

The term Synapsida (Osborn 1903) has by all recent students been used with the taxonomic rank of Subclass to include those vertebrates popularly known as the mammal-like reptiles. These animals possess a mosaic of characters some of which pertain to the Class Mammalia and others to the Class Reptilia.

They are thus neither true mammals nor true reptiles and do thus not fit into the Class Mammalia or into the Class Reptilia.

I thus support those recent authors who have proposed that the name Synapsida should have the rank of a separate Class.

The Class Synapsida would then include the two Subclasses Pelycosauria and Therapsida.

For the Subclass Therapsida I propose the following classification:

| Subclass | Superorder | Order | Suborder | Family |
| :---: | :---: | :---: | :---: | :---: |
| Therapsida | Alphatherapsida | Eotitanosuchia <br> Dinocephalia |  | Eotitanosuchidae Phthinosuchidae Rubidginidae |
|  |  |  | Brithopia | Brithopodidae Anteosauridae |
|  |  |  | Titanosuchia | Titanosuchidae <br> Tapinocephalidae <br> Styracocephalidae <br> Estemmenosuchidae |
|  |  | Gorgonopsia |  | Hipposauridae Gorgonopsidae Burnetiidae |
|  | Betatherapsida | Venyukovioidea |  | Otsheriidae <br> Venyukoviidae <br> ? Dromasauridae |
|  |  | Dicynodontia |  | Endothiodontidae <br> Dicynodontidae <br> Kistecephalidae <br> Lystrosauridae <br> Kannemeyeriidae |
|  | Gammatherapsida | Therocephalia |  | Pristerognathidae <br> Lycosuchidae <br> Whaitsidae |
|  |  | Scaloposauria | Ictidosuchia | Alopecodontidae Ictidosuchidae Scaloposauridae |


| Subclass | Superorder | Order | Suborder | Family |
| :--- | :---: | :---: | :--- | :--- |
| Therapsida <br> (continued) | Gammatherapsida <br> (continued) | Scaloposauria <br> (continued) <br> Cynodontia | Bauriamorpha | Procynosuchia |
|  |  | Bauriidae <br> Ericiolacertidae <br> Procynosuchidae <br> Galesauridae <br> ? Silphedestidae |  |  |
|  |  | Tritylodontia | Cynognathidae <br> Diademodontidae <br> Chiniquodontidae <br> Traversodontidae <br> Tritylodontidae |  |
| Trithelodontidae |  |  |  |  |
| Diarthrognathidae |  |  |  |  |

## Diagnoses of the Higher Therapsid Taxa

## SUBGLASS THERAPSIDA

Advanced synapsids of the Permian and Triassic. There is strong evidence that one therapsid superorder, at least, was directly derived from sphenacodont pelycosaurs, but the derivation of the other two superorders from sphenacodonts, although very probable, is less certain. The therapsids include the direct ancestors of the mammals.

Further advanced than the pelycosaurs in that: the pterygo-basicranial joint is no longer freely movable; a longitudinal girder is developed, the interpterygoid vacuity is never widely open but partly or completely closed; the squamosal is outflaring with a posterior face; there is no supratemporal; the lacrimal never reaches the nostril and the maxilla is deep.

At the beginning of the Middle Permian the therapsids had already developed in diverse directions each showing a lesser or greater acquisition of certain mammalian characters.

Of the three main branches one became successfully adapted and dominated the scene during the Middle and Upper Permian, comprising herbivores and their predatory carnivores, but proved sterile; a second branch of herbivores became adapted to their special niche, waxed exceedingly and very successfully maintained themselves to near the end of the Triassic when they died out without issue; the third branch, already well established at the beginning of the Middle Permian, firstly as predators and later developing herbivorous side branches, developed more and more in the mammalian direction, with one or more twigs producing the first mammals late in the Triassic.

## SUPERORDER ALPHATHERAPSIDA

Permian therapsids a stage further developed than the early Permian sphenacodonts from which they arose, not leading to mammals.

The intertemporal skull table is primitively broad and flat, with the posterior flange of the postorbital lying horizontally in the dorsal skull roof (but modified in some Dinocephalia) and reaching the squamosal; the postorbital bar is always complete.

The lower jaw primitively without a prominent coronoid process (but present in Gorgonopsia), the dentary always strong, but without a definite postero-ventral angle; the postdentary bones always well developed.

The quadrate is primitively robust with the quadrate ramus of the pterygoid strong (except in Gorgonopsia).

Primitively with simple conical incisors, canine and postcanines (but modified in some Dinocephalia) and palatal teeth on the pterygoid and palatine.

There is no secondary palate and no suborbital fenestra.
The epipterygoid is slender and does not enter the sidewall of the braincase; the prootic is weakly developed with a free anterior edge; the sphenoidal complex is well ossified.

The postfrontal is well developed; the dorsal premaxillary process is long (but short in Gorgonopsia).

A pineal foramen is always present.
The occipital condyle is single.

## ORDER EOTITANOSUCHIA

The most primitive therapsids descending from sphenacodonts, with all the primitive characters listed in the diagnosis of the superorder Alphatherapsida of which they are the morphological ancestors.

## Family Eotitanosuchidae

Eotitanosuchidae with the squamosal not extending into the intertemporal skull roof and the primary palate closed.

## Family Phthinosuchidae

Eotitanosuchians with the squamosal developing a lappet entering the intertemporal skull roof and the primary palate with a median cleft.

## Family Rubidginidae

Younger relict eotitanosuchians with a short series of serrated postcanines and a small temporal fenestra.

## ORDER GORGONOPSIA

Middle and Upper Permian alphatherapsids descending from eotitanosuchians, which have developed a prominent coronoid process; the quadrate posteriorly situated is reduced in size and the quadrate ramus of the pterygoid is lightly built; the dorsal process of the premaxillary is shortened; a preparietal is developed. Extinct at the end of the Permian leaving no descendants.

## Family Hipposauridae

Primitive gorgonopsians with a very broad intertemporal skull roof; small temporal fenestra, deep suspensorium, fairly long postcanine series, with the dorsal skull contour strongly curved.

## Family Gorgonopsidae

Intertemporal skull roof somewhat reduced in both earlier and later forms, but in the latter sometimes secondarily greatly widened; large temporal fenestra, fairly shallow suspensorium, postcanine series reduced, gape of jaws in some Upper Permian forms enormous with very strong canines.

## Family Burnetiidae

Later aberrant gorgonopsians with very wide intertemporal region, reduced temporal fenestra, dentition reduced with weak teeth; with pachyostotic thickening of roofbones of skull in the form of bosses and ridges.

## ORDER DINOGEPHALIA

Early alphatherapsids derivable from an eotitanosuchian niveau and die out without issue at the end of the Middle Permian; with basically primitive structure obscured in some branches by pachyostosis and some abortive specializations.

Width of intertemporal skull table reduced, sometimes greatly so, but secondarily greatly widened where the pachyostosis is great, temporal fenestra moderate to large except where secondarily reduced by the pachyostosis.

No coronoid process on the dentary; primitively with a carnivorous dentition of simple conical teeth, later specialized carnivorous with an early development of a herbivorous branch and intermeshing of some or all the upper and lower batteries.

The quadrate robust, as also the quadrate ramus of the pterygoid; quadrate shifting anteriorly.

Dorsal process of the premaxillary long to very long.

## SUBORDER BRITHOPIA

Primitive dinocephalians linked to the eotitanosuchians and morphologically ancestral to the other dinocephalian groups.

The intertemporal skull roof reduced in width with the posterior process of the postorbital lying at a slant down from the horizontal; the temporal fenestra large.

The dentition carnivorous, with the incisors tending to lengthen and the postcanines becoming reduced; the lower and upper incisors and canines intermesh.

Palatal teeth primitively well developed, later practically confined to the palatine.

Quadratojugal never a surface bone.
With no or little general pachyostosis.
Dorsal process of the premaxillary moderately long.

## Family Brithopodidae

Primitive brithopians with a fairly long postcanine series; incisors not greatly lengthened; postfrontal not bulbously swollen and no other pachyostosis; quadrate with little anterior shift; moderate outflaring of squamosals. Palatal teeth well developed.

## Family Anteosauridae

A stage further advanced than the brithopids. Postcanine series reduced, incisors greatly lengthened. Postfrontals becoming greatly bulbously swollen and the skull roof moderately pachyostosed. Strong outflaring of squamosals, especially posteriorly.

Palatal teeth reduced, practically confined to the palatine.

## SUBORDER TITANOSUCHIA

Advanced dinocephalians derived from a brithopid niveau.
The intertemporal region reduced in width, sometimes to a narrow sagittal crest, but secondarily greatly to enormously widened where the pachyostosis is great; the temporal fenestra large to very large but secondarily greatly reduced.

The dentition is herbivorous, initially with a large conical canine and with only the incisors developing a talon and heel; later the canine is not distinguishable as such and all the marginal teeth develop a talon and heel; the postcanine series always very long. Palatal teeth practically absent.

Quadratojugal sometimes a surface bone.
The pachyostosis is moderate to enormous.

## Family Titanosuchidae

Primitive titanosuchians developed from a brithopid level and indicating the morphological level from which the tapinocephalids arose.

The intertemporal width reduced with a low thick sagittal crest, posterior process of the postorbital reduced, temporal fenestra fairly large, but squamosals not outflaring.

Strong incisors with piercing talon and crushing heel and large conical canines, a long series of spatulate postcanines, which do not intermesh as do the incisors and canines of the two jaws. No palatal teeth.

Moderate pachyostosis.

## Family Tapinocephalidae

Specialized titanosuchians derived from a titanosuchid level.
The intertemporal region very variable, mostly of moderate width, sometimes with a sharp sagittal crest, in one subfamily enormously broadened as a result of the excessive pachyostosis; temporal fenestra large to greatly reduced.

All the marginal teeth with talon and heel, upper and lower battery intermeshing.

Pachyostosis light to great.

## Family Styracocephalidae

Middle Permian aberrant titanosuchians. Intertemporal region very
broad, but temporal fossa roomy with posteriorly flaring squamosal; pachyostosis in the form of 'horns' and bosses.

Weak conical incisors and canine and a long series of postcanines; palatal teeth very well developed, even on the vomer.

## Family Estemmenosuchidae

The 'horns' situated on the frontals, and directed dorsally, whereas in the Styracocephalidae the 'horns' are formed by the tabular and directed posteriorly. Otherwise with features very similar to those of the Styracocephalidae.

## SUPERORDER BETATHERAPSIDA

Permian and Triassic therapsids on a developmental niveau far above that of the early Permian sphenacodonts, not leading to mammals.

The intertemporal skull table is primitively reduced in width, but flat, becoming narrow and later developing a sagittal crest, but secondarily widened in the Kistecephalidae; the posterior process of the postorbital inclined downwards from the horizontal and reaching the squamosal, later reduced; the postorbital bar is always complete.

The lower jaw without a coronoid process of the dentary and without a coronoid bone; the dentary always strong without a postero-ventral angle; the postdentary bones well developed. The quadrate is robust lying low down on a pedicel of the uniquely triradiate squamosal, and the quadrate ramus of the pterygoid is weak. Primitively with a modified set of marginal teeth in both jaws, which very early are radically reduced and sometimes wholly lost and replaced by horny sheaths; there are no palatal teeth.

The premaxillaries, primitively paired but later fused, develop plates to form a unique type of secondary palate and the choanae are shifted posteriorly; primitively the maxilla and palatine have no inward palatal growth but later extend palatally but never meet below the air passage. The epipterygoid is slender and does not enter the sidewall of the braincase; the prootic is weakly developed with a free anterior edge; the sphenoidal complex is well ossified and lies far anteriorly.

The postfrontal is primitively well developed but reduced later; the dorsal premaxillary processes are primitively paired and long, later fused and short.

A pineal foramen present; preparietal primitively absent, later present. The occipital condyle is single. There is a fenestra in the mandible between the dentary and angular.

## ORDER VENYUKOVIOIDEA

Primitive betatherapsids not directly linked to the sphenacodonts, leading to the higher Dicynodontia.

The width of the intertemporal region is reduced, without sagittal crest and the temporal fenestra is short, the jugal has a large entry into the zygoma which is not strongly everted.

The dentition consists of a well-developed series of bluntly conical marginal teeth on the premaxilla, maxilla and dentary.

No inward growth of palatine and maxilla and the posterior part of the palate is thus primitive, except that the lateral ramus of the pterygoid is somewhat or much reduced.

The premaxillaries are not fused and have a long dorsal process; the septomaxilla is largely superficial, the postfrontal is well developed and there is no preparietal.

## Family Otsheriidae

The incisors are enlarged, the choana is short, the palatine does not meet the premaxilla; the lateral ramus of the pterygoid is still prominent, the lacrimal is short and there is no pachyostosis.

## Family Venyukoviidae

The incisors are enlarged, the choana is long; the palatine meets the premaxilla; the lateral ramus of the pterygoid is much reduced; the lacrimal is long; there is some pachyostosis.

## Family Dromasauridae

The dentition consists of a series of isodont marginal teeth or the jaws are edentulous; the temporal fenestra is very short and deep.

## ORDER DICYNODONTIA

Advanced betatherapsids morphologially derivable from Otsheria; a long-lived order, rich in species varying in minor characters with a single main theme and phylogenetically sterile.

The intertemporal region reduced in width, sometimes very much so, but is secondarily widened in one aberrant family; the sagittal crest feeble to very high or wholly absent. The temporal fenestra is very long and the jugal is practically ousted from the zygoma by the squamosal which is uniquely everted.

The marginal teeth are greatly modified, there are never any incisors; an upper conical canine present or absent; postcanines present or absent and when present shifted medially and variously disposed.

Palatal flanges of the palatine and maxilla tending to grow inwards to form a variable open trough for the air passage; the lateral ramus of the pterygoid is lost.

The premaxillaries are fused and the dorsal process is short; the septomaxilla tending to shift interiorally; the postfrontal is primitively present but is lost in later forms; a preparietal is developed.

## Family Endothiodontidae

With postcanine teeth, number and disposition very variable, canines present or absent, with a postfrontal.

## Family Dicynodontidae

Without postcanine teeth, canines present or absent, the postfrontal is frequently absent.

## Family Kistecephalidae

The intertemporal region is secondarily greatly widened, without sagittal crest, edentulous, without canines, without pre- and postfrontals and preparietal.

## Family Lystrosauridae

Without postcanines and canines usually present, postfrontal present, nares shifted posteriorly and premaxilla lengthened.

## Family Kannemeyeriidae

Very high sagittal crest; depression leading into pineal foramen.

## SUPERORDER GAMMATHERAPSIDA

Permian and Triassic therapsids probably derived from Early Permian sphenacodonts and including the immediate ancestors of the mammals.

The intertemporal skull table narrow, usually with a sagittal crest, but secondarily widened in one late family; the postorbital is reduced and never reaches the squamosal; the postorbital bar primitively and usually complete, but incomplete and even wholly absent in some advanced forms.

The dentary primitively and usually with a prominant coronoid process, strong and finally greatly enlarged with a strong postero-ventral angle and making contact with the squamosal; the postdentary bones primitively well developed but greatly reduced in some advanced groups.

The quadrate small, with weak to incomplete quadrate ramus of the pterygoid.

Primitively with carnivorous dentition of simple conical teeth, later very variable, often with accessory cusps in the postcanines and in some advanced forms with highly elaborated crowns of a mammalian nature.

Primitively without secondary palate, in later groups incipient in various ways, to incomplete, and finally fully developed in mammalian fashion.

The epipterygoid is primitively slender and remains so in many forms, but is widened in some early forms and in advanced forms very broad and incorporated into the sidewall of the braincase; the prootic is primitively weakly developed, but later growing forwards meets the epipterygoid (alisphenoid) suturally; the sphenoidal complex is usually not well ossified.

The postfrontal small or absent.
The occipital condyle single in earlier forms, later becomes notched and finally with double condyles formed by the exoccipitals.

## ORDER THEROGEPHALIA

Middle to Upper Permian gammatherapsids; initially primitive but with a wide gap between them and their sphenacodont precursors; with a degenerate family in the Upper Permian; relation to higher gammatherapsids is uncertain.

The intertemporal region is narrow with a reduced postorbital; the postorbital bar always complete; pineal foramen always present; dentary with a well-developed coronoid process and the postdentary bones well developed; no prominent postero-ventral angle to dentary.

Primitively without secondary palate but aberrantly incipient in the Upper Permian family.

Epipterygoid slender or widened but never incorporated in the sidewall of the braincase.

Dentition primitively carnivorous with a long postcanine series, later greatly reduced to lost, always uncusped. Postfrontal small or absent, suborbital fenestra large in earlier forms but reduced to absent in later forms.

The occipital condyle is always single.

## Family Pristerognathidae

Middle Permian therocephalians with a well-developed carnivorous dentition with a single canine; no secondary palate, epipterygoid slender, postfrontal small, suborbital fenestra large.

## Family Lycosuchidae

Middle Permian therocephalians with a well-developed carnivorous dentition, with double canines; no secondary palate, epipterygoid widened; postfrontal small, suborbital fenestra large.
Family Whaitsiidae (including Lycedeopsidae and Euchambersiidae as subfamilies)

Upper Permian therocephalians with reduced dentition, postcanines feeble, few or wholly absent, lower incisors sometimes absent, aberrant development of an incipient secondary palate, epipterygoid widened, suborbital fenestra large to small or absent, dentary scimitar-shaped with postdentary bones not robust.

## ORDER SCALOPOSAURIA

Middle Permian to Lower Triassic gammatherapsids, probably independently derived from sphenacodonts with a considerable gap; fairly primitive in the Middle Permian, but advanced in the Upper Permian and Lower Triassic; relations to cynodonts uncertain.

The intertemporal region is usually narrow with a sagittal crest, but widened in one family with loss of crest; the postorbital bar is slender and complete or incomplete; the postorbital is sometimes greatly reduced or even absent; the jugal spur of the postorbital bar is usually present but absent in a few forms; the pineal foramen is sometimes absent.

The coronoid process of the dentary is strong, feeble or absent; there is no prominant postero-ventral angle to the dentary; the postdentary bones are well developed or weakened. A secondary palate is primitively absent, later incipient to well developed. The epipterygoid is mostly slender but sometimes widened and partially included in the sidewall of the braincase in one form.

A prominent canine is usually retained but sometimes not recognizable as such, accessory small canines are usually present, maxillary teeth usually numerous and the postcanines variable, being simple, cusped or with transversely widened crowns.

The suborbital fenestrae are always well developed, the postfrontal is reduced or absent.

The occipital condyle is initially single but later sometimes incipiently double.

## SUborder ictidosuchia

Mostly Permian scaloposaurians just extending into the Triassic; the intertemporal region is usually narrow but later widened in one family, the postdentary bones are weakened and the dentary lightly built; the secondary palate is primitively absent, sometimes incipient but never complete, the epipterygoid is slender but in one case partially enters the sidewall of the braincase.

## Family Alopecodontidae

Primitive Middle Permian ictidosuchians close to the contemporary therocephalians.

The intertemporal region is narrow with a sagittal crest; the postorbital bar is complete, the coronoid process is strong, the dentary robust and the postdentary bones well developed; there is no secondary palate.

Dentition carnivorous, always with a prominent canine and two small accessories, the postcanines are simple conical teeth, the postfrontal is small and the occipital condyle single.

## Family Ictidosuchidae

Upper Permian ictidosuchians linked to the alopecodontids.
The intertemporal region is narrow, usually with a sagittal crest and a pineal foramen present; primitively with a complete postorbital bar, but later incomplete; the coronoid process is prominent, the secondary palate absent, incipient to weakly developed; with an enlarged canine behind smaller accessories and the postcanines simple conical teeth, a single occipital condyle.

## Family Scaloposauridae

Permian to Lower Triassic ictidosuchians.
Primitively with a narrow intertemporal region, but in some later forms this is widened and the pineal foramen is often absent.

Primitively with a complete postorbital bar, but later incomplete and sometimes even without a jugal spur.

The coronoid is weak or absent.
The secondary palate is incipient to weakly developed.
Primitively with an enlarged main canine, but in advanced forms not distinguishable, the postcanines are primitively conical but sometimes cusped.

## SUBORDER BAURIAMORPHA

Triassic scaloposaurians further advanced than the ictidosuchians; the intertemporal region is narrow, usually with a sagittal crest and pineal foramen.

Postorbital bar, complete or incomplete.
Dentary with a prominent coronoid process; weak or robust with postdentary bones well developed or weak; with a well-developed closed secondary palate; epipterygoid moderately widened, lying lateral to braincase.

Incisors and canine conical or peglike, postcanines with cusps and transversely expanded. Occipital condyle notched or double.

## Family Bauriidae

Intertemporal region narrow with sagittal crest; pineal foramen absent or present; the postorbital bar complete or incomplete, sometimes without jugal spur.

Dentary strong with well-developed postdentary bones. The vomer does not enter the secondary palate. Incisors and prominent canine conical and postcanines expanded.

## Family Ericiolacertidae

Intertemporal region broadened, without sagittal crest, no pineal foramen; postorbital bar incomplete, without a jugal spur.

Dentary and postdentary bones lightly built.
The vomer enters the secondary palate.
No outstanding canine, incisors modified and postcanines peglike with cusps and expanded transversely.

## ORDER CYNODONTIA

Advanced Upper Permian and Triassic gammatherapsids, derived from sphenacodonts probably through an intermediate stage at a morphological level near that of the Middle Permian therocephalians and scaloposaurians; including the ancestors of the mammals.

The intertemporal region is narrow with a sagittal crest, pineal foramen primitively present but later lost; postorbital bar complete in earlier forms but later incomplete. Dentary with weak to very strong coronoid process, the postero-ventral angle to the dentary is primitively weakly developed but very prominent in later forms; primitively without a posterior process but this is developed in later forms and in some forms reaches the squamosal to form an accessory articulation; the postdentary bones well developed in earlier forms but later much reduced. Primitively with a cleft secondary palate, but later closed.

The epipterygoid is widened and enters the sidewall of the braincase and becomes suturally joined to the prootic.

The dentition primitively with conical incisors and canines sometimes with accessory small canines; the postcanines developing cusps and later with widened variously elaborated crowns; primitively polyphyodont later diphyo-
dont with 'premolars' and 'molars' distinguishable.
Postfrontal lost; no suborbital fenestra; occipital condyle notched and later double.

## SUBORDER PROCYNOSUGHIA

Primitive Upper Permian and Triassic cynodonts, related to the two older gammatherapsid orders and linked to the first mammals.

Pineal foramen present and postorbital bar complete, zygoma lightly built.
In the dentary the postero-ventral angle is absent or only moderately developed; a masseteric fossa on the coronoid process is incipient to fairly well developed but there is still no masseteric process on the jugal; the posterior process of the dentary is still undeveloped; the postdentary bones are still well developed, but the reflected lamina of the angular is reduced. Initially the secondary palate is still cleft but is later closed.

The incisors and canine conical with accessory small canines sometimes present; accessory cusps on the postcanines.

## Family Procynosuchidae

Upper Permian primitive procynosuchians still with a cleft palate, fairly weak coronoid process, with accessory precanine maxillary teeth; the occipital condyle is incipiently double.

## Family Galesauridae

Upper Permian and Lower Triassic procynosuchians with a closed secondary palate, strong coronoid process without precanine maxillary teeth, and a double occipital condyle.

## SUBORDER GYNOGNATHIA

Advanced specialized Triassic cynodonts, with a carnivorous and herbivorous branch, derived from Upper Permian procynosuchians, becoming extinct in the Upper Triassic.

Pineal foramen present, postorbital bar complete and a very strong zygoma.

Dentary greatly enlarged, prominent and strong coronoid process with masseteric fossa fairly to very well developed; strong to very strong posteroventral angle; posterior process of dentary moderately to well developed and in some advanced forms making contact with the squamosal in an accessory articulation; masseteric process on jugal present or absent with a step between maxilla and jugal; postdentary bones greatly reduced with all but loss of reflected lamina of the angular. Secondary palate well developed and closed. Incisors and canines conical, without accessory anterior canines, postcanines with fore and aft accessory cusps or with crowns transversely expanded and further elaborated.

## Family Cynognathidae

Earlier carnivorous cynognathians with the dentary not making contact
with the squamosal, and with small angular process and jugal process; maxillary teeth divided into premolars with crenulated crowns and molars with a series of sectorial cusps in a longitudinal row.

## Family Chiniquodontidae

Later carnivorous cynognathians with the dentary making contact with the squamosal in some advanced forms, usually no angular process to the dentary.

## Family Diademodontidae

Earlier herbivorous cynognathians with very strong masseteric process on the jugal; long series of maxillary teeth with peg-like premolars and transversely widened crushing molars.

## Family Traversodontidae

Later herbivorous cynognathians with a step between the maxilla and jugal and no masseteric process on the jugal.

## SUBORDER TRITYLODONTIA

Advanced Upper Triassic cynodonts, derived from Upper Permian procynosuchians.

Without pineal foramen, postorbital bar incomplete without postorbital and postfrontal; zygoma very strong or fairly weak.

Dentary greatly enlarged with strong coronoid process and well developed masseteric fossa; postero-ventral angle very prominent, posterior process of dentary well developed and making contact with the squamosal in advanced forms; postdentary bones greatly reduced; no jugal process.

Secondary palate closed but greatly reduced in width with median shift of postcanines. Transverse ramus of pterygoid reduced.

Incisors primitively conical or specialized and recumbent, conical canine present or absent, postcanines cusped and further elaborated with crushing crowns.

Mononarial or binarial.

## Family Trithelodontidae

Zygoma moderately strong; posterior process of the dentary not reaching the squamosal; incisors primitively conical, long diastema, long series of widened molars; mononarial.

## Family Tritylodontidae

Zygoma strong; posterior process of the dentary not meeting the squamosal; incisors reduced, one enlarged, recumbent in dentary, no canine, long diastema, molars quadrangular with elaborate crushing crowns; mononarial.

## Family Diarthrognathidae

Zygoma fairly weak; posterior process of the dentary making contact with the squamosal in an accessory articulation; incisors and canine primitively conical, no diastema, molars transversely widened with cusps; binarial.

## Summary

The classification of the Therapsida is re-evaluated and the older view of two main branches, Anomodontia and Theriodontia, discarded in favour of three main branches for which the names Alphatherapsida, Betatherapsida and Gammatherapsida are proposed.

## References

Boonstra, L. D. 1953. A new scaloposaurian genus. Ann. Mag. nat. Hist. (12) 6: 6or-6o5.
Boonstra, L. D. 1954. The cranial structure of the titanosuchian: Anteosaurus. Ann. S. Afr. Mus. 42: 108-148.
Boonstra, L. D. 1963. Early dichotomies in the therapsids. S. Afr. 7. Sci. 59: 176-195.
Brink, A. S. 1963. The taxonomic position of the Synapsida. S. Afr. 7. Sci. 59: 153-159.
Вroom, R. 1903. On the classification of the theriodonts and their allies. Rep. S. Afr. Ass. Advmt Sci. 1: 286-294.
Вroom, R. 1905. On the use of the term Anomodontia. Rec. Albany Mus. 1: 266-269.
Broom, R. 1912. On some new fossil reptiles from the Permian and Triassic beds of South Africa. Proc. zool. Soc. Lond. 1912: 859-876.
Broom, R. 1923. On the structure of the skull in the carnivorous dinocephalian reptiles. Proc. zool. Soc. Lond. 1923: 661-684.
Broom, R. 1931. Notices of some new genera and species of Karroo fossil reptiles. Rec. Albany $M u s$ 4: 161-166.
Broom, R. 1932. The mammal-like reptiles of South Africa and the origin of mammals. London: Witherby.
Efremov, I. A. 1940. Preliminary description of the new Permian and Triassic Tetrapoda from USSR. Trudy paleont. Inst. 10 (2): 1-140.
Efremov, I. A. 1954. [A fauna of terrestrial vertebrates in the Permian cupriferous sandstones of the western Ural region.] Trudy paleont. Inst. 54: 1-416. (In Russian.)
Haughton, S. H. 1918. Investigations in South African fossil reptiles and Amphibia. (Part ir.) Some new carnivorous therapsids, with notes upon the brain-case in certain species. Ann. S. Afr. Mus. 12: 175-216.

Haughton, S. H. 1924. A bibliographic list of pre-Stormberg Karroo fossils. Trans. R. Soc. S. Afr. 12: 51-104.
Haughton, S. H. 1929. On some new therapsid genera. Ann. S. Afr. Mus. 28: 55-78.
Huene, F. von. 1948. Short review of the lower tetrapods. In royal society of south africa. Robert Broom commemorative volume: 65-106. Cape Town: Royal Society of South Africa. (Special publication.)
Kutorga, S. 1838. Beitrag zur Kenntniss der organischen Ueberreste des Kupfersandsteins am westlichen Abhange des Urals. St. Petersburg: Eggers.
Lydekrer, R. 1890. Catalogue of the fossil Reptilia and Amphibia in the British Museum. Part IV. London: British Museum.
Osborn, H. F. 1903. The reptilian subclasses Diapsida and Synapsida and the early history of the Diaptosauria. Mem. Am. Mus. nat. Hist. 1: 449-507.
Owen, R. 1844. Description of certain fossil crania, discovered by A. G. Bain, Esq., in sandstone rocks at the south-eastern extremity of Africa, referable to different species of an extinct genus of Reptilia (Dicynodon) and indicative of a new tribe or sub-order of Sauria. Proc. geol. Soc. Lond. 4: 500-504.
OWEN, 1859 (1860). On the orders of fossil and recent Reptilia, and their distribution in time. Rep. Br. Ass. Advmt Sci. (Aberdeen, 1859) 29: 153-166.
OWEN, R. 1860. Palaeontology; or, A systematic summary of extinct animals and their relations. Edinburgh: Longman.

Owen, R. 1876. Descriptive and illustrated catalogue of the fossil Reptilia of South Africa in the collection of the British Museum. London: British Museum.
Reed, C. 1960. Polyphyletic or monophyletic ancestry of mammals, or: what is a class? Evolution 14: 314-322.
Romer, A. S. 196i. Synapsid evolution and dentition. In International colloquium on the evolution of lower and non specialized mammals . . . September 1960. 1: 9-56. Brussels: Koninklijke Vlaamse Academie voor Wetenschappen, Letteren en Schone Kunsten van Belgie.
Seeley, H. G. 1895. Researches on the structure, organization, and classification of the fossil Reptilia. Part IX, section 1. On the Therosuchia. Phil. Trans. R. Soc. (B) 183: 311-370.
Simpson, G. G. 1925. A Mesozoic mammal skull from Mongolia. Am. Mus. Novit. 201: 1 -1 1.
Tchudinov, P. K. 196o. [Upper Permian therapsids of the Ezhovo locality.] Paleont. Zh. 1960 (4): 8r-94. (In Russian.)

Van Valen, L. ig6o. Therapsids as mammals. Evolution 14: 304-313.
Watson, D. M. S. 1917. Sketch classification of the pre-Jurassic tetrapod vertebrates. Proc. zool. Soc. Lond. 1917: 167-186.
Watson, D. M. S. \& Romer, A. S. 1956. A classification of therapsid reptiles. Bull. Mus. comp. Zool. Harv. 114: 37-89.


Fig. I. Schematic representation of the evolutionary story of the mammals. The diagram is based on the number of described genera.

We commence the story nearly 400 million years ago.
At that time (the Devonian) there lived a group of freshwater fish, known as the Rhipidistia, whose paired fins had become adapted to propelling the body forwards with a purchase on the muddy floor of shallow pools.

These rhipidistians were succeeded (during the Carboniferous) by a group of amphibians, known as the Embolomeri, with two pairs of extremities capable of ungainly locomotion on dry land and with the ability of utilizing atmospheric oxygen but returning to the water for reproduction.

Just before the next period (the Permian) we encounter two groups of reptiles that had become completely adapted to life on land. These were the primitive Captorhinidae, with a simple adductor muscular mass for closing the jaw when feeding and the Sphenacodontia, where the adductor muscles had developed into a more efficient mechanism for feeding and able to raise the body from the ground for better locomotion.

Arising from these early reptiles (during the later part of the Permian) we have the Therapsida, which evolved in various directions, but with this in common viz. a great improvement in the jaw mechanism and locomotor ability. The majority of the therapsids, developing along differing but nearly parallel lines, became extinct, some in the Permian and others successfully competing to the end of the Trias.

Of the more successful therapsids we indicate in the scheme a group consisting of the familiar Scaloposauridae, Bauriidae, Tritylodontidae and Diarthrognathidae, brigading them
together as the Premammalian Therapsida. In this group the locomotor ability is greatly improved and the jaw mechanism adapted more and more for chewing their food. For the latter the lower jaw became more and more dominated by a single bone - the dentary - and the teeth developed shearing, cutting and crushing cusps and the respiration during the chewing process was facilitated by the development of a bony secondary palate, separating the air passage from the buccal cavity.

Up to the end of the Triassic period all the vertebrates were poikilothermic or 'cold-blooded', i.e. they had no built in mechanism for temperature control. About this time certain of the higher therapsids, with their higher rate of metabolism, made possible by the improved locomotor and masticatory ability, developed mechanisms to dissipate excess body heat or to conserve it. For the former a skin with glands for sweating and a diaphragm for panting became imperative. To conserve heat the development of an insulating cover of hair or fur took place. These features, together with the dentary-squamosal jaw hinge made these small rat-like creatures mammals.

From their beginning late in the Triassic (about 150 million years ago) these first mammals were small rat-like animals forming a very inconspicuous part of the vertebrate fauna. This continued throughout the Jurassic and Cretaceous, when vertebrate life was dominated by the sauropsid reptiles which included the dinosaurs during their heyday.

But from the Tertiary the mammals waxed exceedingly to fill every possible ecological niche including besides terrestrial conditions varying from arctic to tropical climates, excursions into fresh and salt water and into the air. The culminating event, less than a million years ago, is the emergence of Man.

Since then this single genus has attained a dominant position in the living world, which it is ravaging at an alarming rate.


Fig. 2. Schematic phylogeny of the Therapsida.

## INSTRUCTIONS TO AUTHORS

## Based on

CONFERENCE OF BIOLOGICAL EDITORS, COMMITTEE ON FORM AND STYLE. 1960 .
Style manual for biological journals. Washington: American Institute of Biological Sciences.

## MANUSCRIPT

To be typewritten, double spaced, with good margins, arranged in the following order: (1) Heading, consisting of informative but brief title, name(s) of author(s), address(es) of author(s), number of illustrations (plates, figures, enumerated maps and tables) in the article. (2) Contents. (3) The main text, divided into principal divisions with major headings; subheadings to be used sparingly and enumeration of headings to be avoided. (4) Summary. (5) Acknowledgements. (6) References, as below. (7) Key to lettering of figures. (8) Explanation to plates.

## ILLUSTRATIONS

To be reducible to $12 \mathrm{~cm} \times 18 \mathrm{~cm}$ ( 19 cm including caption). A metric scale to appear with all photographs.

## REFERENCES

Harvard system (name and year) to be used: author's name and year of publication given in text; full references at the end of the article, arranged alphabetically by names, chronologically within each name, with suffixes $a, b$, etc. to the year for more than one paper by the same author in that year.
For books give title in italics, edition, volume number, place of publication, publisher.
For journal articles give title of article, title of journal in italics (abbreviated according to the World list of scientific periodicals. 4 th ed. London: Butterworths, 1963), series in parentheses, volume number, part number (only if independently paged) in parentheses, pagination.
Examples (note capitalization and punctuation)
Bullough, W. S. 196o. Practical invertebrate anatomy. 2nd ed. London: Macmillan.
Fischer, P.-H. 1948. Données sur la résistance et de le vitalité des mollusques. 7. Conch., Paris 88: 100-140.
Fischer, P.-H., Duval, M. \& Raffy, A. i933. Etudes sur les échanges respiratoires des littorines. Archs Zool. exp. gén. 74: 627-634.
Kohn, A. J. rg6oa. Ecological notes on Conus (Mollusca: Gastropoda) in the Trincomalee region of Ceylon. Ann. Mag. nat. Hist. (13) 2: 309-320.
Kohn, A. J. ig6ob. Spawning behaviour, egg masses and larval development in Conus from the Indian Ocean. Bull. Bingham oceanogr. Coll. 17 (4): 1 -51.
Thiele, J. 19ı. Mollusca: B. Polyplacophora, Gastropoda marina, Bivalvia. In schultze, L. Zoologische und anthropologische Ergebnisse einer Forschungsreise im westlichen und zentralen SüdAfrika. 4: 269-270. Jena: Fischer. Denkschr. med-naturw. Ges. Jena 16: 269-270.

## ZOOLOGIGAL NOMENCLATURE

To be governed by the rulings of the latest International code of zoological nomenclature issued by the International Trust for Zoological Nomenclature (particularly articles 22 and 51). The Harvard system of reference to be used in the synonymy lists, with the full references incorporated in the list at the end of the article, and not given in contracted form in the synonymy list.

## Example

Scalaria coronata Lamarck, 1816: pl. 451, figs $5 a, b$; Liste: 11. Turton, 1932: 80.
-


DTION

|  |
| :---: |
|  |  |
|  |  |
|  |  |
|  |  |
|  |  |
|  |  |
|  |  |
|  |  |

## ITUTION





LIBRARIES SMITHSONIAN


2
2
-1
-1
-1
-1
2
2 LIBRARIES


SMITHSONIAN


Sヨ1y甘y 17

institution

institution



SMITHSONIAN INSTITUTION


LI


## y甘ygl



NOILOLILSNI

LIBRARIES SMITHSONIAN



[^0]:    ${ }^{1}$ As used by Robinette \& Stains (1970: fig. 1); this is the anterior articulation of King (1966: 393).

[^1]:    * This cusp was shown in Lydekker's (1884) illustration, but others (e.g. Matthew 1929) apparently overlooked it.

[^2]:    Example
    Scalaria coronata Lamarck, 1816: pl. 451, figs $5 a, b$; Liste: 1 1. Turton, 1932: 80.

[^3]:    * On the available specimens no constant differences could be found between the shells of these two species.

[^4]:    * Voss (personal communication) has two females caught off Beira in 1964 by the U.S. Indian Ocean Expedition. The localities are $19^{\circ} 51^{\prime} \mathrm{S}, 36^{\circ} 21^{\prime} \mathrm{E}, 62 \mathrm{~m}$, and $20^{\circ} 30^{\prime} \mathrm{S}, 35^{\circ} 49^{\prime} \mathrm{E}, 32 \mathrm{~m}$. The latter locality is within the limits of southern Africa as defined on page 194.

[^5]:    * This identification has been confirmed by Adam (personal communication).

[^6]:    * Not $33^{\circ} 04^{\prime} \mathrm{S}, 27^{\circ} 54^{\prime} \mathrm{E}, 27 \mathrm{fm}$., as stated by Barnard (1962:252).

[^7]:    * See footnote on p. 213.

[^8]:    * Not $57 \%$ (Adam, personal communication).

[^9]:    * In one specimen (A31319, length 63 mm ) the striated zone is concave.

[^10]:    * Not $30^{\circ} 09^{\prime} \mathrm{S}, 19^{\circ} 02^{\prime} \mathrm{E}$, as stated by Adam \& Rees.

[^11]:    Hemisepius typicus Steenstrup, 1875: 468, pl. I, figs 1-10, pl. II, fig. 1. Hoyle, 1886: 26, 217 ; 1912: 281. Gibbons, 1888: 202. Smith, 1903: 356; 1916: 25 . Chun, 1915: $4^{11}$, figs 33, 34. Massy, 1927: 164. Thore, 1945: 50, fig. I. Voss, 1962b: 248, 252; 1967: 64.
    Hemisepion typicum: Rochebrune, 1884: 78, pl. 3, fig. I.
    Rhombosepion sp. A Massy, 1927: 161.
    Hemisepius typicus var. chuni Thore, 1945: 50.
    Sepia (Hemisepius) typica Adam \& Rees, 1966: 117 , pl. 32, figs 192-195, pl. 33, figs 196, 197.

[^12]:    * Adam (pers. comm.) says that he and Rees do not attach any systematic value to these groups. Where, however, a genus contains as many species as does Sepia, the use of some such grouping of similar species greatly facilitates the study of interspecific relationships.

[^13]:    * Three known exceptions are Sepia elliptica, S. hedleyi and S. pharaonis, which have been collected from depths to 1000 metres.

[^14]:    * Length of shell if complete
    $\dagger$ Part of striated zone missing; percentage should be higher. These dimensions excluded from the calculation of the mean relative length of striated zone for this reason.

[^15]:    Scale $=10 \mathrm{~mm}$.

