

THE DANISH
INGOLF-EXPEDITION

VOL. V D

PUBLISHED AT THE COST OF THE GOVERNMENT
BY
THE DIRECTION OF THE ZOOLOGICAL MUSEUM OF THE UNIVERSITY



COPENHAGEN
PRINTED BY BJANCO LUNO BØGH
1911-1919



Contents of Vol. V D

- XII. O. CARLSEN: Actinaria, II, p. 1-92 (9 plates) 1942
- XIII. F. JENSENIUS MADSEN: Octocorallia, p. 1-95 (11 plates) 1944
- XIV. P. L. KRAMP: Medusa, III, Frachylina and Scyphozoa, p. 1-99 (8 plates) 1944

THE DANISH INGOLF-EXPEDITION

VOLUME V

12

ACTINULARIA

PART II

BY

OSKAR CARLGREN

WITH 6 PLATES AND 95 FIGURES IN THE TEXT



COPENHAGEN

PRINTED BY BIANCO LUNDA

1942

Preface.

It is now about twenty years since the first part of the report of The Ingolf-Actiniaria was published. Several circumstances have retarded the publication of the second part, I think, however, that this delay has not been altogether a loss. On the contrary, our knowledge of the forms provided with acontia has increased during the interval, especially since the systematization of the nematocysts has been carried out. I have here followed the classification proposed by WEILL (1935), but somewhat modified by me (1940), of the nematocysts. As I have previously pointed out, it is sometimes difficult to distinguish especially the microbasic *p*-mastigophors from the microbasic amastigophors in preserved material, but I think that, in the main, my determination of

the types is correct. In this connection I will note that the breadth of the nematocysts was given as somewhat too small in part I of the Ingolf-Actiniaria, because I then used too low a magnification. In this part I have described several species new to science, some of them not belonging to the Acontinaria. The latter have been added in an Appendix. Several species only very shortly characterized by me (1928, 1940) have here been given a more detailed description.

I have written this paper according to nearly same plan I have indicated in 1921 in the first report of the Actiniaria of the Ingolf expedition.

I. Description of the species.

Subtribus Acontinaria.

Nynantheae the pedal disc of which is provided with basilar muscles and acontia or organs like acontia. Sphincter usually mesogloal.

Family Bathypheilliidae.

Acontinaria, the acontia of which carry only basitrichs. Mesenteries differentiated into macro- and microcnemes. Sphincter mesogloal.

I proposed this family in 1932 for the genera *Bathypheilia*, *Acraspedanthus* and *Phelliogoton*.

Genus *Bathypheilia* Carlgr.

Bathypheilliidae with broad pedal disc. Column divisible into scapus and scapulus, the former with cuticle and tenaculi. Sphincter strong, mesogloal. Tentacles hexamerously arranged, fewer than the mesenteries. Longitudinal muscles of tentacles and radial muscles of oral disc ectodermal. Two siphonoglyphs. Mesenteries hexamerously arranged, those of the last cycle growing from the aboral end upwards. 6 pairs perfect, 2 pairs of directives. Only the perfect pairs fertile and provided with filaments and acontia. Retractors of the perfect mesenteries concentrated, also present but weak on the stronger imperfect mesenteries. Nematocysts of the acontia only basitrichs. Cnidom: spirocysts, basitrichs, microbasic *p*-mastigophors.

Type and single known species, *B. margaritacea*.

The genus is easily distinguished from *Acraspedanthus* Carlgr., which has a smooth column and more mesenteries in the distal than in the proximal part, and from *Phelliogoton* Carlgr., the indistinct scapus of which lacks cuticle and tenaculi. It cannot be identical either with *Paraphellia* Hadd., which also is devoid of

a cuticle and has the mesenteries of the second and third order fertile, or with the imperfectly known *Eaphyllia* Pax, which according to PAX is provided with cinclides in longitudinal rows, or with *Cataphellia* Stephenson, which also has cinclides and comes near *Hornothia*, *Phollia*, *Isophellia* Carlgr. and probably *Decaphellia* Bourne belong to the Sagartiidae and Isophelliidae inasmuch as the acontia contain microbasic amastigophors as well as basitrichs.

Bathypheilia margaritacea (Dan.).

Pl. III fig. 10

Phellia margaritacea n. sp. DANIELSEN 1890 p. 54, Pl. 3 fig. 7.

Pl. 12 figs. 6-12. CARLGRÉN 1928 p. 258, 300.

Bathypheilia margaritacea (Dan.) CARLGRÉN 1932 p. 292.

Diagnosis: Body cylindrical, in expanded state somewhat higher than broad, in contracted state of about same length and breadth. Column divisible into scapus and scapulus, without cinclides but with tenaculi. Sphincter very strong, broad, alveolar, not stratified, perfectly separated from the endodermal muscles of the column, its muscle meshes narrow, in its uppermost part arched. Tentacles cylindrical, rather short, in number between 24 and 36, hexamerously arranged, the last cycle incomplete. Longitudinal muscles of tentacles and radial muscles of oral disc ectodermal. Pairs of mesenteries arranged in 2 or 3 cycles, the third cycle incomplete. Only the first 6 pairs perfect, fertile and provided with filaments and acontia. Retractors of the perfect mesenteries

17-19 μ about 2 μ in size. Moreover I have, in the maceration preparations, found nematocysts and spirocysts of same size as those of the tentacles but these capsules belonged probably not to the scapulus but were stuck on the drawn in tentacles.

The ectoderm of the tentacles is high and provided with rather numerous nematocysts and very numerous spirocysts. The ectodermal, longitudinal muscles of the tentacles are weak at the outside of their bases, at the inside considerably stronger. Here the folds of the muscle lamella reach about the same thickness as the mesogloea. In the upper part of the tentacles the muscles are about equally developed on both sides. The radial muscles of the oral disc are ectodermal, weak in the inner part of the disc as also in the radial furrows, somewhat stronger between the

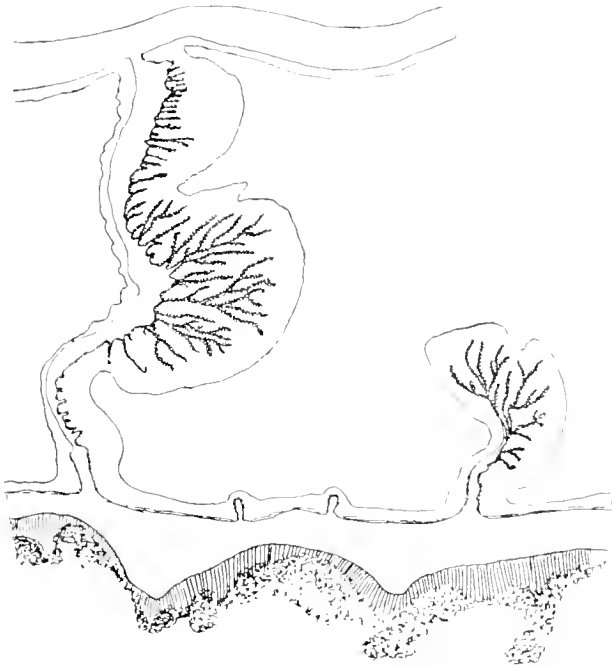


Fig. 3. *Bathypheilia margaritacea*. Cross-section of a directive mesentery and of one mesentery of the second and two of the third order in the lower part of the actinopharynx.

furrows. The ectoderm of the actinopharynx is considerably higher than that of the oral disc. The actinopharynx is devoid of longitudinal muscles, its mesogloea is thinner than its ectoderm, in the 2 siphonoglyphs thickened.

The mesenteries are hexamerously arranged in 2 or 3 cycles, of which the third is incomplete, as a rule usually consisting of a few pairs some of which reach the distal end of the body. In one specimen with 26 tentacles I have counted, however, about 48 mesenteries at the base. The exocoels, in which the mesenteries of the third order arise, do not correspond in the specimens. The 6 first pairs of mesenteries are perfect, the others imperfect. The longitudinal muscles of the perfect mesenteries form distinct retractors. In the uppermost part of the mesenteries they are concentrated and have high folds, in the aboral part of the actinopharynx and below it the pennons are more diffuse with the highest folds in the outer part of the mesenteries. Also the mesenteries of the second order, and rarely the strongest mesenteries of the third, are provided with rounded but weak retractors as seen in cross-sections. Usually the mesenteries of the third order are very weak and project only a little beyond the surface of the endoderm. In textfig. 3 I have drawn one directive, one mesentery of the second order and 2 mesenteries of the third at the level of the lower part of the actinopharynx. The parietobasilar and basilar muscles are weak. Only the mesenteries of the first cycle have filaments, acontia and reproductive organs. The ciliated tracts of the filaments are well developed, as also the acontia.

Genus *Daontesia* nov. gen.

Bathypheiliidae divisible into scapus and acontia. Scapus with tenaculi and with cinclides at the limbus. Siphonoglyphs. Tentacles not numerous, hexamerously arranged in number of the mesenteries. Two siphonoglyphs and 2 pairs of directives. Perfect pairs of mesenteries 12, provided with filaments. At least the mesenteries of the first cycle fertile. Retractors diffuse, strong. Cnidomorph spirocysts, microbasal *p*-mastigophors, basitrichs.

I have previously referred the species described below to the genus *Sagartiogeton*. It recalls in some respects also *Kabusactis* but several maceration preparations, as well as sections of isolated parts of the acontia, have shown that only basitrichs were present in these organs, wherefore the genus must be joined either with the family Hormathiidae or the family Bathypheiliidae. Because the mesenteries of the third cycle are well developed (as it seems, lack filaments and acontia, I refer the genus to the latter family though it shows a certain relation to *Chelobathypheilia*, a Hormathiid. In fact the genus forms a link between the typical forms provided with macro- and microcnemes and those the mesenteries of which are not so differentiated. Such transitional forms exist also between the families Isophelliidae and Sagartiogetonidae.

Daontesia praelonga (Carlgr.)

Pl. III fig. 1, Pl. V fig. 1.

Sagartiogeton praelongus n. sp. CARLGRÉN 1928, p. 259, 302.

Diagnosis: Body elongated. Tenaculi of the scapus very close set, very numerous, especially in the upper part of the scapus. Their cuticle multi-stratified, giving the scapus a shaggy appearance. Cinclides at the limbus probably arranged in longitudinal rows. Sphincter strong, alveolar. Tentacles and mesenteries about 48, hexamerously arranged. Two siphonoglyphs and 2 pairs of directives. Perfect pairs of mesenteries 12. Inner parts of the mesenteries curved towards the exocoels, those of the directives towards the endocoels. Retractors of the mesenteries diffuse, strong, especially those of the perfect pairs, with numerous high folds. Parietobasilar muscles weak. Only the perfect pairs provided with filaments. Dioecious. Nematocysts of the scapulus 17.5-22 μ (about 2.8 μ), often a little curved, basitrichs, those of the tentacles 26-34 μ (about 2.8 μ), those of the actinopharynx partly 24-34 μ (2-5.5 μ), microbasal *p*-mastigophors, partly 24-31 μ (3.5-5.5 μ), basitrichs, those of the filaments partly 24-31 (34) μ (3.5-5.5 μ), microbasal *p*-mastigophors (length of exploded shaft that of the capsule or a little shorter) partly 24-24 (28) μ (about 2.8 μ), partly 12-17 μ (1.5-2.5 μ), both basitrichs, rare, those of the acontia 45-58 μ (3.5 μ), in very small specimens 41-48 μ , basitrichs. Spirocysts of tentacles about 24 μ (2.5-60 μ), 5.5 μ .

Colour in alcohol: Scapus dirty grey.

Dimensions in contracted state, 1) length of the body 2.2 cm., breadth 1.3 cm., inner tentacles about 0.6 cm. long (RYDER leg.); 2) length of the scapus 4.6 cm., breadth of basal disc 4 cm., that of the upper part of the scapus 0.8 cm. (Ingolf exp. St. 64), 3) length 0.8, breadth 0.4 cm. (smallest specimen).

Occurrence: Danmark Strait 65°30' N, 28°25' W, 1041 m. Ryder 1888, 1 specimen, S. of Iceland 62°06' N, 19°00' W, 1960 m. Bottomtemp. 3.1 (Ingolf expel. St. 64) 1 specimen, E. of Iceland 65°31' N, 7°31' W, 1435 m. Bottomtemp. 0.8 (Ingolf expel. St. 105) 1 specimen, basal part lacking, W. of Faroe Islands 61°32' N, 11°36' W, 1356 m. Bottomtemp. 2.40 (Ingolf expel. St. 4) 1 specimen, basal part lacking.

The distribution of the species is given in fig. 94, p. 76.

Exterior aspect. The pedal disc of the largest specimen (no. 1) to a great part torn up, that of a smaller specimen well expanded and provided with a cuticle to which mud is attached. The colour

the ciliated tract, the lower portion is divided into two parts, the former is part with thin but dense setae invested with cuticle. The latter part of the sectioned mesentery is longitudinal in nature below, but owing to the nature of the part I have not been able to determine. The other part of the scapulus is of ordinary length and

is like the scapulus the raised portion of the sectioned specimens. I counted 50 mesenteries at 4, 50 mesenteries at 5, 50 or so or what fewer at 6, 50 or so, rather broad at their distal end, at least in the contracted state, rather wide, irregularly sulcated, in the largest specimen, very



FIG. 4. The two filiform mesenteries from the actinopharynx.

FIG. 5. Section of two mesenteries of the second and third cycles in the region of the ciliated tracts.

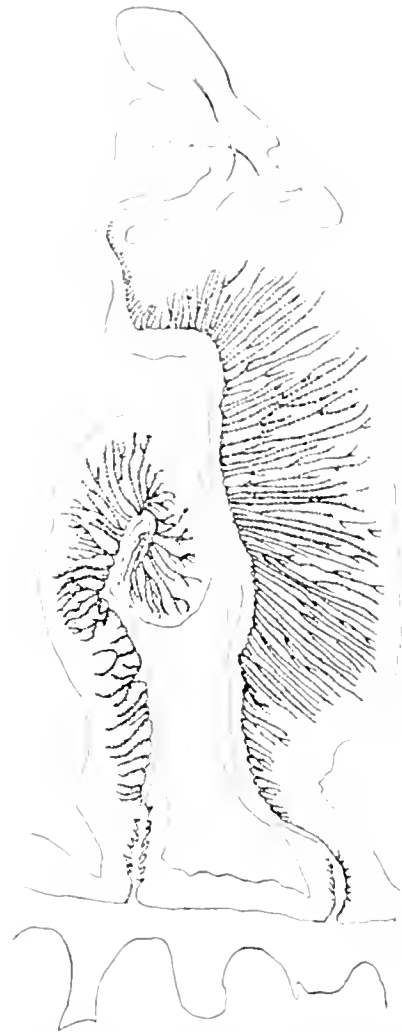


FIG. 5. *Danobesia praelonga*. Section of two mesenteries of the second and third cycles in the region of the ciliated tracts.

numerous strata. The greater part of the papillae is formed by the cuticle, whereby the papillae come to have a very characteristic appearance. In the figure 1 PL. V, I have given a longitudinal section of some papillae with contiguous parts of the column-wall. Between the chitinized ectoderm cells gland cells are probably intercalated, though they were torn up during sectioning. The mesogloea of the column is of ordinary thickness and extended into the papillae. The cinclides seem to be exclusively invaginations of the ectoderm. The ectoderm of the scapulus is about as thick as that of the scapus, its nematoecysts numerous. The circular muscles of the column are weak, the sphincter on the other hand very strong, long and broad. It occupies the whole, strongly thickened mesogloea of the scapulus and the uppermost part of the scapus, and is wholly separated from the endodermal circular muscles of the column (fig. 4). Its muscles are small in the lowermost part, in other parts it consists of larger and smaller muscle meshes and its structure is alveolar. They are of about the same width but diminish rather rapidly proximally.

The ectoderm of the tentacles is high, its nematoecysts few, its spirocysts numerous. The longitudinal muscles of the tentacles are ectodermal and developed to the usual extent. The radial muscles of the oral disc agree with the longitudinal muscles of the tentacles, and are about equally developed at the insertions of

the mesenteries as between them. The ectoderm and the mesogloea of the actinopharynx is commonly of about same thickness, in the ridges, and especially in the 2 siphonoglyphs, the mesogloea is thicker. All nematocysts of the actinopharynx are broader in their basal end.

The arrangement of the mesenteries is, in the largest specimen, $6 + 6 + 12 + 1 = 25$ pairs. The extra pair is situated next to the one directive pair. In a second specimen there are 24 pairs, 12 pairs are perfect in the whole tract of the actinopharynx, the others imperfect. The retractors form strong, diffuse pennons distributed over the greater part of all mesenteries, though they are considerably weaker in the mesenteries of the third cycle than in those of the others but here also fairly strong

(fig. 5). The folds of the muscle lamella are very high and close set but not much branched. The inner parts of the mesenteries are curved towards the exocoels in the non-directives, towards the endocoels in the directives. The parietobasilar muscles are not strong, and form only short, rather few folds. In the larger individuals the two first cycles of mesenteries are certainly provided with filaments. On the mesenteries of the third cycle I have seen neither filaments nor acontia, though these mesenteries reach the margin. The ovaries were lost from the mesenteries in the larger specimens, in a smaller one at least the mesenteries of the first cycle were fertile.

Remarks see the genus.

Family Sagartiidae.

Acontiaria with mesogloea sphincter. Mesenteries not differentiated into macro- and microcnemes. Their acontia provided with microbasal amastigophors and basitrichs.

Genus Sagartia.

Sagartiidae with well developed pedal disc. Column with scattered suckers (constructed as the warts but without any endodermal evaginations), capable of attaching foreign bodies. Cinclides present. Sphincter mesogloea, fairly well developed. Tentacles fairly numerous. Longitudinal muscles of tentacles and radial muscles of oral disc ectodermal. Siphonoglyph one or two, distinct. Number of mesenteries about the same at the base as at the margin. More than 6 pairs of mesenteries perfect. No differentiation of the mesenteries into macro- and microcnemes. Retractors of the mesenteries diffuse or somewhat concentrated, never circumscribed. Gonads present on the mesenteries of first cycle onwards. Sometimes asexual reproduction. Acontia well developed. Unidom: spirocysts, microbasal amastigophors, microbasal *p*-mastigophors, basitrichs.

Sagartia troglodytes (Price).

Pl. VI fig. 4.

Literature and synonyms see STEPHENSON 1935 p. 324-325, also *Sagartia troglodytes* PRICE CARLGRÉN 1930 p. 4; 1939 p. 8; 1940 p. 43 figs. XIV 6-8; PAX 1936 p. 113.

Diagnosis: Pedal disc wide. Column in expanded state pillar-like, in full extension many times higher than broad, in the upper two-thirds provided with strong, adhesive suckers. Cinclides scattered, evaginations of the endoderm as well as invaginations of the ectoderm, generally the evaginations are the stronger. Sphincter strong, separated from the endodermal muscles of the column by a thin lamella, gradually diminishing downwards, in its upper part filling up almost the whole mesogloea and here reticular, in its lower part more alveolar. Tentacles conical, relatively short, hexamerously arranged, up to 192. Two distinct siphonoglyphs. Mesenteries hexamerously arranged, up to 192, 2 pairs of directives. At least 12 pairs perfect. Retractors of the mesenteries diffuse, bandlike with comparatively low folds, directive retractors concentrated. Parietobasilar muscles fairly well developed, basilar muscles weak but distinct. Acontia very slender, emitted only on strong stimulation, with small nematocysts. Never asexual reproduction. Nematocysts of the column partly H 12 : 2 μ , basitrichs, partly H 12-17 : 3.5-4 μ microbasal amastigophors, those of the tentacles partly H 10-22 : 2-2.5 μ , basitrichs, partly H 17 : 2.5-3 μ , few, microbasal amastigophors, those of the actinopharynx partly 22-24 : 1-1.5 μ , microbasal

p-mastigophors partly 22-27 : about 2.5 μ , basitrichs, those of the filaments partly 12-24 : 1.5-5 μ , microbasal *p*-mastigophors, partly 17-20 : 2-2.5 μ , basitrichs, those of the acontia partly 17-29 : 3.5- about 4 μ , microbasal amastigophors, partly 12-17 : 1.5-2 μ , basitrichs. Spirocysts of tentacles up to about 22.6 : 3.5 μ .

Colour very variable (see GOSSE (1860) and STEPHENSON (1935)). The individuals from Bohuslän are usually coloured as follows. Column pale salmon red, in its upper part greyish or greenish with pale suckers and white longitudinal bands most conspicuous at the base, where longer bands alternate with shorter ones. Cinclides without coloured annulus. Tentacles seldom opaque white, greyish, sometimes with a longitudinal, brownish or olive-green band on each side and with two or three cross-bands of yellowish-white. At the base of the stronger tentacles a distinct B-mark of more or less deep-black colour; at the base of the outer tentacles only a black spot. Oral disc greyish to greyish-black with thin ochre coloured, longitudinal lines and white cross-lines. In the middle of each radius of the two or three oldest cycles a white patch. The colour-variety *melanoleuca* of Gosse is not rare on the Swedish west coast.

Size: height 1 cm or more, diameter of base 3.4 cm, of disc 2 cm (STEPHENSON 1935). The individuals from the Scandinavian waters seem not to reach the size of those from Great Britain.

Occurrence: Iceland: Helmaey, Vestmannaeyar, Grindavik, Kollavik.

Faroe Islands: Trangisvaag (Fjeldstrup).

Norway: Sklinna; Bessahorne; Vällersund, Lersvik, low water (Dons); Svalbardvagen, Titran, low-water (Dons); Trondheim-fjord; Brisund among sea-weed, low-water (E. Pettersson); Rognevaldsvaag low-water (Dons); Herdla, low-water (Carlgrén).

Sweden: Bohuslän, Gullmaren; N.A. of Flatholmen (Bock); Gasvik; Strommarne, a few metres (Carlgrén and others), between Kallsö and Vargo H 12 m (Währberg and Eliasson).

Denmark: Länfjord; Livo Bredning (Peterson); west of Fur (Carlgrén); N. of Haustholm 57-24' N, 38° E, 33 m, (Dana), Frederikshavn 8 m (Kramp); N. of Borrebjerg bank 8 m, Marens bank (Kramp); Hirtsholms north-west bank 2-3 m (Kramp); Hellebæk (Mortensen 1910); (N. of Feno 30 m (Carlgrén 1929); mouth of Koldingfjord 44 m (Carlgrén 1929); the occurrence of the specimens in the two latter localities is somewhat dubious as I cannot find any proof of them.)

Further distribution: North Sea, coast of Great Britain and Ireland, Atlantic coast of France. Probably also the Mediterranean.

Exterior aspect: The pedal disc is wide, not undulated, the body conical, in strong extension pillar like and then considerably higher than broad. The column is, in its upper part, provided with

and the oral disc is very prominent and, outside the longitudinal ridges, the surface of the mesenteries is covered with numerous small suckers, and the oral disc is covered with numerous small suckers. The margins are very strongly invaginated. The suckers are relatively short in comparison with those of *S. elegans* (Dall) and hexamerously arranged in the oral disc (see also GOSSE). The oral disc is covered with numerous longitudinal ridges and with numerous longitudinal and radial siphonoglyphs.

Structure of the column.—The ectoderm of the column is covered with numerous small suckers. The structure of the column is very similar to that of the vermiform *Tealia*, *Banohutis* and *Agardhiopsis* but the oral disc here takes no part in the contraction of the column (PAX 1936, fig. 4). The cinclides are partly invaginations of the ectoderm, partly invaginations of the ectoderm and partly covers from the canal to about same extent. The lower part of the canal is covered by the ectoderm. The oral circular muscles of the column are very weak. The subdermis is strong, separated from the ectoderm by a thin lamella of the mesogloea. The oral disc is not pulled up almost the whole mesogloea, but it is pulled up gradually. Its structure is reticular, especially in the upper part, sometimes the lower part is more alveolar. The oral and radial muscles of the oral disc are strong.

The mesenteries are hexamerously arranged with two pairs of retractors (8 individuals examined). At least 12 pairs are perfect. The retractors of the stronger mesenteries are band-like and diffuse, those of the primary height and often a little branched, sometimes the retractors of the directives are a little concentrated. The basilar muscular folds show few folds, but are, at least in larger individuals, situated on a shelf of the mesogloea. The basilar muscles are weak but distinct. All stronger mesenteries are fertile. The acontia are very thin, but long.

I have never observed asexual reproduction in this species. The individuals from the Swedish coast seem to be oviparous. In an individual from Titter (Norway) there were, however, embryos in the coelenteric cavity.

The species is easily recognised by the scattered adhesive suckers and above all by the thin acontia, the nematocysts of which are very small—the microbasic amastigophors never longer than 3μ (12 individuals examined). As to the appearance of the nematocysts see CARLGRÉN 1940 p. 43. For further details of exterior and colour see STEPHENSON 1935.

Remarks.—This species is often confounded with other *Sagartia* species and with *Cereus polyanthatus* (*Hebertes bellus*) and vice versa. It is at present impossible to state if it occurs in the Mediterranean. VOX HEIDERS *S. troglodytes* from Trieste is a *Cereus*. LINNARVA's short description 1889 of his *troglodytes* from Marseilles is a *Cereus* to certain information that he has examined this species.

***Sagartia elegans* (Dall)**

Part of the literature.—see STEPHENSON 1935 p. 306, also STEPHENSON 1936 p. 10, CARLGRÉN 1924 p. 27, STEPHENSON 1935 p. 10, CARLGRÉN 1939 p. 8, PAX 1936 p. 112 fig. 68.

Description.—Pencil-like. Column with scattered suckers and a few small suckers at upper part. Cinclides present in the upper part of the column, whereas at the limbus. Upper cinclides principally invaginations of the ectoderm, lower ones invaginations of the mesogloea. Mesogloea at base a little undulated and overhanging, becoming more regular orally, in smaller individuals the oral disc is pulled up, separated from the endodermal surface by a thin lamella of the mesogloea, pushing downwards, in its upper part it is pulled up the whole breadth of the mesogloea. The oral circular muscles are very weak, up to about 192 hexamerously arranged in the oral disc. The oral radial muscles of tentacles

and radial muscles of oral disc ectodermal. Actinopharynx with longitudinal ridges. One siphonoglyph or two. Three cycles of mesenteries perfect in regular specimens. One pair of directives or two. Retractors of mesenteries diffuse, forming a band about in the middle of the mesenteries, provided with fairly low folds on the non-directives. The three first cycles of mesenteries fertile. Asexual reproduction by fragmentation (bearing). Acontia thick and very numerous, after stimulation rapidly ejected from the mouth and the cinclides. Nematocysts of the column partly $18.3-24.5-37.5-4.5\mu$, often curved, microbasic amastigophors, partly $18.3-21.5-2.8-3\mu$, basitrichs, those of the tentacles partly $24-26-$ about 4.2μ , microbasic amastigophors, partly (17) $24-28-2.8\mu$, basitrichs, those of the actinopharynx partly $19-25-4.2-5\mu$ microbasic *p*-mastigophors, partly $24-35-3\mu$, basitrichs, partly $14.5-18.3-4.2-5.6\mu$ microbasic amastigophors sparse, those of the filaments partly $20-25.4-4.2\mu$ microbasic *p*-mastigophors, partly $8-11-1.5$ basitrichs, rare, those of the acontia, partly $(39-11-77(80)-(4.5)-5-7\mu)$, microbasic amastigophors, partly $(21)-24-11-2.8-4.2\mu$, basitrichs. Spirocysts of the tentacles $19-2-$ about $28-4.5\mu$.

Colour of an individual from Bohuslän: Column brick-red with white suckers, at the base white longitudinal lines. Outer tentacles at the base orange coloured, otherwise blackish violet with one white cross-band and white to yellowish-white apex, inner tentacles brownish-grey with two white bands and yellowish-white apex, at the base a rather distinct B mark, inwards on the oral disc a white zone and over the stronger endocoels a ring of white spots about in the middle of the disc. Actinopharynx shading in orange. Another specimen from Bohuslän had the column brownish-yellow but at the limbus flesh-coloured, the suckers were uncoloured, the tentacles coloured as the former specimen, but the orange colour very indistinct on the outer tentacles. An individual from Ulvesund had the column salmon red, in stronger contraction shading into orange with light suckers, the uppermost part of the column was violet-grey. The outer tentacles had at their base a small, indistinct yellowish-white zone (sometimes not present), the most part of them was orange coloured, the apex was greyish shading into violet. The inner tentacles were light grey shading into violet, in more contracted state blackish violet with 3 yellowish cross-band. At the base of the tentacles there was a violet-black, triangular spot, more distinct in contracted state (no distinct B mark present). The oral disc was flesh-coloured powdered with violet-grey to violet-black, sometimes with irregularly arranged small, white spots. The actinopharynx was flesh-coloured. The colour of this species is very variable, see GOSSE (1860), NAFILYAN (1912) and STEPHENSON (1935).

Dimensions at least up to 3.8 cm at the base and 1.5 cm at the margin (STEPHENSON 1935). All Scandinavian individuals were smaller.

Occurrence: Iceland: Vestmannaeyar at the beach, Rockall 185 m (*Sagartia rockalliensis*). Norway: Hjeltefjord; Ulvesund and Ösund, a few metres, on Laminaria (Carlgrén); Gulløy, Solsvik, littoral (Dons); Skagerrak: mouth of Gullmarfjord, Smedjan 30 m (Carlgrén); Kattegat: S.E. of Store Middelgrund 30 m "Akka" St. 153, N.W. of Hallands Väderö 30 m ("Akka" St. 164); S.W. of Hesselö 22 m ("Akka" St. 159); N.W. of Kullen 30 m ("Akka" St. 160); off Aalsgaard (Kramp); off Hellebæk, 24 fms. (Mortensen); Helgoland (Carlgrén).

Distribution: Kattegat, Skagerrak, the North Sea, the south west coast of Iceland, British Isles, Atlantic coast of France, the Mediterranean.

Exterior aspect: The column is conical or more cylindrical in connection with different state of contraction. The cinclides seem to vary from 4 up to 5 in the compartments, the former

from the weakest endocoels, the latter from the strongest. The tentacles are shorter than those of *Sagartiogeton viduatus*, *undatus* and *anguicomus*. The oral disc is wide. There is often a single siphonoglyph, sometimes two. Moreover see the diagnosis.

Anatomical description: The anatomy of this species is previously described by F. DIXON (1888) and by STEPHENSON (1920), wherefore I have not much to add. The suckers of the high ectoderm seem to be of the same structure as those of *S. troglodytes*, though they are weaker and seldom used for adherence. The sphincter is reticular in older individuals.

The mesenteries are in regular specimens arranged hexamerously and up to 192 in number. Often their arrangement is irregular. Characteristic of this species is that often only a single directive pair is present. As the asexual reproduction takes place here in the same manner as in *Metridium* viz. by "tearing" (STEPHENSON) of pieces from the limbus, the presence of a single directive pair is probably due to the persistence of old mesenteries in the lacerated pieces (see CARLGRÉN 1901, 1909). F. DIXON has examined in all 10 individuals, of which 8 were provided with a single siphonoglyph. Of 6 examples dissected by myself, 2 had two pairs of directives the others only one pair. An individual (from Ulvesund) was a double animal with more than 200 tentacles and with two mouths but only one oral disc showing uncommonly strong undulation for this species. The one mouth and the corresponding throat were smaller than the others. Only one directive pair joined the smaller throat, while 2 directive pairs were attached to the larger one. The perfect pairs vary in number, in regular individuals usually 3 cycles of mesenteries are perfect, in irregular ones there can be more or less than 24 perfect pairs. The retractors are band-like, diffuse, in the directives somewhat more concentrated, with richly branched folds. The parietal muscles are weak and not folded, the basilar muscles fairly well developed. The acontia are thick. I have examined the nematocysts of the acontia in 31 (33) individuals from different localities (Iceland, Helgoland, Norway, Sweden, and Denmark). The length of the microbasal amastigophors varied in 31 specimens between 47 and 77 μ , the average size of all was 57.54. STEPHENSON has examined 12 acontia, he found the average length of the amastigophors to be 53.93 μ . The basitrichs varied in 33 specimens between 26 and 41 μ , the average length of all was 33.3, in STEPHENSON's specimens 33.4, thus a good agreement.

Remarks: I think that *Sagartia rockaliensis* described by me 1924 is identical with *S. elegans*. I have (1935 p. 6) also described *elegans* from the Mediterranean (the gulf of Genoa). Possibly it belongs to another *Sagartia*-species though there was good agreement as to the size of the nematocysts of the acontia with those of *S. elegans*.

Genus *Kadosactis* Dan.

Diagnosis: Pedal disc well developed, with a cuticle. Column divisible into scapus and scapulus, the former with a cuticle, usually easily deciduous, and with tenaculi rarely very weak, usually strong with a stratified cuticle. Column with cinchides formed by the ecto- as well as by the endoderm and situated in various places, sometimes in reduction. Margin more or less distinct. Sphincter strong, mesogloecal. Tentacles of variable length, conical, hexamerously or irregularly arranged. Outer tentacles with or without basal thickenings on the outside. Number of tentacles usually fewer than that of the mesenteries. Longitudinal muscles of the tentacles ectodermal, at the base stronger on the inside than on the outside. Radial muscles of oral disc ectodermal. 1 or 2 up to 4 siphonoglyphs, 1-4 pairs of directives. Arrangement of the mesenteries hexamerous or irregular. At least 6 pairs of mesenteries, usually 12 or more perfect. Mesenteries growing from the limbus towards the margin. Perfect mesenteries and sometimes the strongest imperfect with strong diffuse retractors, extended over the whole surface and never concentrated, nor circumscrip-

and provided with filaments and reproductive organs. Parietobasilar muscles not strong. The stronger mesenteries, not all directives, fertile. Acontia well developed, their nematocysts microbasal amastigophors and basitrichs, the former often very large. Cnidom: spirocysts, microbasal *p*-mastigophors, microbasal amastigophors, basitrichs.

As I have previously pointed out it is not necessary to set up a distinct family for this genus (compare STEPHENSON 1920 p. 191 and CARLGRÉN 1933 p. 24). The genus is related to *Phellia* but separated from it among other features by the presence of cinchides and a different structure of the tenaculi, which sometimes are very small and not rarely visible to the naked eye as in *K. (Sagartiogeton) antarctica*. As to the subgenus *Kadosanthos*, proposed by me 1934 for *K. subeatus*, differing from the typical *Kadosactis*-species in the presence of large tubercle-like formations, it is questionable, if the tenaculi are wholly reduced. As, however, the ectoderm of the column was wholly lost and the species was taken in very deep water, it is probable, that the rubbing of the clay against the animal during the dredging had removed every trace of the tenaculi, which at any rate may have been very small.

STEPHENSON (1935 p. 358) thinks that it may be necessary to unite the genera *Kadosactis* and *Actinothoe* because some species referred to *Actinothoe* are provided with tenaculi in conformity with *Kadosactis*. I think it is impossible to refer, for inst., the type of *Actinothoe*, *A. sphyrodacta*, a very delicate species, to the robust species of *Kadosactis* here named. Already the type of *Kadosactis*, *K. rosea*, is against such an opposition. For inst., the thin tentacles in *Actinothoe* are very different from the thick tentacles of *Kadosactis rosea*, which moreover are provided with thick mesogloecal swellings on their outside. The retractors of the mesenteries have also another appearance in the two genera. True, *Actinothoe undata* has more mesenteries at the base than at the margin in conformity with *Kadosactis* but there is, however, a real difference. In *Kadosactis* the mesenteries grow from the base towards the distal part of the column, in *undata* from the limbus as well from the margin, though the mesenteries originate earlier at the base than at the margin (see *Sagartiogeton undatus*). Therefore I think that these two species must be referred to two different genera. On the other hand it seems to me possible to unite *undata* with the type of genus *Sagartiogeton* proposed by me 1924 for *S. robustus* (see *Sagartiogeton*).

To the genus *Kadosactis* I refer the following species: *K. rosea* Dan., *K. abyssicola* Kor. & Dan., *K. (Sagartiogeton) antarctica* Carlgr., *K. spitzbergensis* (Dan.) and *K. (Sagartiogeton) subeata* Carlgr., the last species being the type for a sub-genus, *Kadosanthos*.

Kadosactis rosea Dan.

- Kadosactis rosea*, n. sp. DANIELSSEN 1890 p. 8, Pl. I fig. 2, Pl. 7 fig. 11, Pl. 9 fig. 8, CARLGRÉN 1932 p. 254, 1933 p. 20 fig. 11.
Phellia bathybia n. sp. DANIELSSEN 1890 p. 61, Pl. I figs. 1-4, Pl. 13 figs. 7-9.
Phellia violacea n. sp. DANIELSSEN 1890 p. 70 Pl. I fig. 7, Pl. 13 fig. 10, Pl. 14 figs. 9-10.
Hormathia mosenlosa n. sp. GRAVIER 1918 p. 15, 1922 p. 69 Pl. 6 figs. 58, 59, Pl. 10 fig. 99.

Diagnosis: Tenaculi rather small, cuticle of the scapus easily deciduous. An annulus of cinchides at the limit between the scapus and scapulus, no cinchides at the limbus. Sphincter strong, reticular to somewhat alveolar, with large muscle meshes. Number of tentacles in large specimens 31-44. Outer tentacles with bulbous thickenings at the outside of the base, at least as long as the inner tentacles, 2 siphonoglyphs and 2 pairs of directives. Pairs of mesenteries normally 24 (6 + 6 + 12), of which not all reach the distal body-end, 12 pairs of perfect mesenteries with strong, diffuse retractors often forming humps. Stronger mesenteries of the third order without filaments and reproductive organs, but

Oral disc directed towards the scapus. Outer tentacles 12 + 21, about 2.5 *u*, 12 + 21, probably micro-*u*. Inner tentacles 20 + 20, tentacles 26 + 36 + 36 + 36, pharynx partly 11 + 65 + 65 + 65, scapus partly 38 + 45, about 3 *u*, 42 + 50, partly 42 + 50, 42 + 55 *u* micro-*u*, 42 + 45 + 2 + 2.5 *u* basitrichs, those of the pharynx partly 39 + 118 + 55 + 7 *u*, 39 + 118 + 55 + 7 *u*, basitrichs, 39 + 118 + 55 + 7 *u*, resp. 36 + 48 + 55 + 7 *u*, resp. 2 + 60 + 6 + 7 *u*.

Colour: Scapus at first white, with a faint rosy tinge, later rosy. Outer tentacles coloured as the column, the inner ones, the pharynx and the oral disc brown coloured, the oral disc and the pharynx rosy-brown with a darker annulus. *Kalosoactis rosea* teste DANIELSEN. Scapus 100 *u*. Scapus yellow-red. Inner tentacles and oral disc yellowish. Outer tentacles at the base and on the sides yellowish, coloured as the scapulus but otherwise quite white. *Phellia scapula* teste DANIELSEN. Scapulus light violet, the rest of the scapus colour with a white annulus round the oral disc. Outer tentacles white. Oral disc pale-violet with yellowish-brown. A siphonophore chestnut brown (*Phellia violacea* teste DANIELSEN). The body is colourless in preserved state, the oral disc and the oral disc are sometimes dark brown, the siphonophore brown, the siphonoglyphs yellowish.

Diameter of preserved state 1.5 cm high and broad (or 2 cm of a 1.5 cm high).

Occurrence: Bahá Bay, 69° 50' N, 61° 37' W, 1880 m. Bottom temp. 1.04. (Godthaab exp. St. 51) 25 specimens; 67° 48' N,

7° 06' W, 2165 m. Bottom temp. 1.1. (Ingolf exp. St. 113) several specimens; 69° 13' N, 8° 23' W, 1889 m. Bottom temp. 1.1. (Ingolf exp. St. 117) several specimens; 70° 51' N, 13° 03' E, 2354 m. Bottom temp. 1.2. 69° 02' N, 11° 26' W, 1836 m. Bottom temp. 1.1. (Norw. N. Atl. exp. 205, St. 210, *Phellia violacea*); 68° 27' N, 8° 20' W, 1996 m. Bottom temp. 1.1. (Ingolf exp. St. 118) 2 specimens; 67° 53' N, 10° 13' W, 1902 m. Bottom temp. 1.1. (Ingolf exp. St. 119) 1 specimen; 67° 29' N, 11° 32' W, 1666 m. Bottom temp. 1.1. (Ingolf exp. St. 120) 1 specimen; 66° 23' N, 7° 25' W, 1802 m. Bottom temp. 1.1.1. (Ingolf exp. St. 101) 15 specimens; 67° 11' N, 8° 18' W, 1619 m. Bottom temp. 1.0.9. (Ingolf exp. St. 111) 3 specimens; 65° 53' N, 7° 18' W, 2127 m. Bottom temp. 1.1.1. (Norw. N. Atl. exp. St. 51, *Phellia bathybia*); 63° 53' N, 6° 22' W, 1960 m. Bottom temp. 1.1. (Michael Sars exp. 1900) numerous specimens; 63° 22' N, 5° 29' W, 2222 m. Bottom temp. 1.2. (Norw. N. Atl. exp. St. 10, *Kalosoactis rosea*).

Exterior aspect: The wide pedal disc is commonly attached to the mud, and then the investment of the pedal disc agrees with that of the scapus. Often the investment is lost, so that the insertions of the mesenteries are clearly visible. 2 specimens (from the Ingolf expedition St. 101) were attached to dead shells of *Pecten frigidus* and also in a bottle from the Norwegian Atlantic expedition a shell of this species was present together with the holotype of *Kalosoactis*. Often the limbus is not distinct in the specimens attached to the mud. The column is usually cylindrical and more or less high according to the stronger or weaker contraction (compare DANIELSEN's figures 1-1 Pl. I of *Phellia bathybia*, which probably are good). If low, the diameter of the pedal disc is considerably larger than the height of the column, especially in the specimens fastened to shells. The scapus is provided with a rather weak cuticle, often richly incrustated with a thick layer of mud. The cuticle is easily deciduous. If



Fig. 9. *Kalosoactis rosea*, Splanchnon.

Occurrence: 69° 00' N, 9° 04' E. (Godthaab exp. St. 162) 1 specimen; 69° 00' N, 9° 04' E, 1865 m. (Monaco exp. W. 1899) 1 specimen; 70° 36' N, 12° 10' E, 1750 m. (Ingolf exp. St. 110) 1 specimen; 70° 51' N, 13° 03' E, 2354 m. Bottom temp. 1.2. (Norw. N. Atl. exp. 205, *Phellia violacea*); 69° 31' N,

removed, the surface of the scapus often shows irregularly arranged, low papillae of various appearance. A nearer examination of their investment indicates, that they are weak tenaculi (compare below). The scapulus is not high, smooth and longitudinally sulcated, at least in contracted specimens. The species is provided with ctenidies. In a small, well preserved specimen from the

Godthaab expedition the cinclides were distinct and visible under a lens. They were situated at the limit between scapus and scapulus and placed in an annulus round the body. To judge from a section of a piece containing three pairs of mesenteries, it seems to be a cinclis from each endocoel in the upper part of the body. On the other hand I have not found any cinclides in a sectioned piece with 5 mesenteries of the limb. The margin is not distinct.

The number of the tentacles is always fewer than that of the mesenteries. 7 specimens examined had 27, 28, 31, 36, 38, 43, and 44 tentacles. DANIELSSEN stated, that *Phellia bathybia* had 18 + 18 and *Kadosactis rosea* 36 + 36 tentacles. As to the latter I have not been able to count all tentacles of the type-specimen,

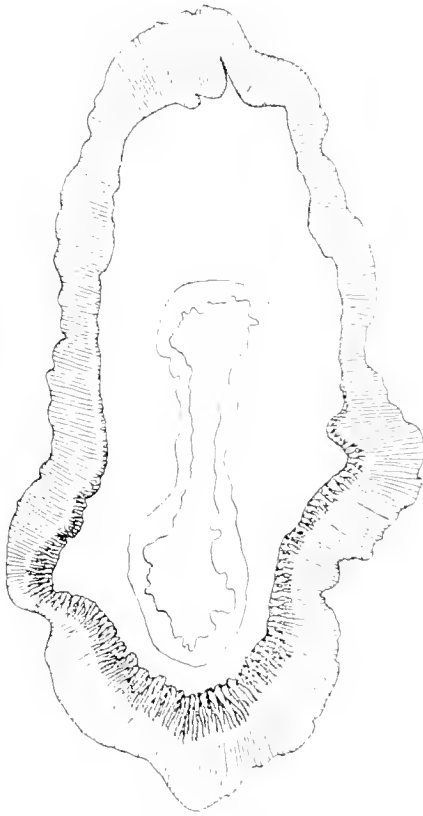


Fig. 7. *Kadosactis rosea*. Transverse section of tentacle.

but judging from the remaining tentacles, there were certainly considerably fewer tentacles than DANIELSSEN states. Also DANIELSSEN's statement of the number of mesenteries in *Kadosactis* is certainly incorrect. Also the tentacle cycles are more than 2 and probably 3. Moreover, it is difficult to decide, whether the tentacles are strictly hexamerously arranged, probably not, as small displacements seem to have taken place in connection with the development of the strong basal swellings on the outer tentacles and the fact that not all mesenteries reach the margin. At the outside of the base the outer tentacles are provided with strong mesogloea thickenings in the form of bridges connected with the mesogloea of the column. In sections of the inner tentacles the mesogloea was somewhat thickened on the corresponding places but these thickenings were visible only in sections. The distal part of the outermost tentacles was sometimes filamentously drawn out and according to DANIELSSEN in the living state almost as long as the column and varying. Also the middle tentacle cycle is often of the same appearance as the third as also sometimes some tentacles of the inner cycle. The tentacles of the latter cycle are, however, commonly shorter and not so much extended. Thus, the outer tentacles are longer or at least as long as the inner ones. The oral disc is wide, in the contracted state of the animal deeply excavated and provided with radial furrows corresponding to the insertions of the mesenteries. About the inner half of the

oral disc is devoid of tentacles. The actinopharynx is rather long, and on each side provided with 10 longitudinal ridges corresponding to the insertions of the perfect mesenteries. The siphonoglyphs are broad and distinct.

Anatomical description: For the anatomical examination I have used 2 type-specimens of *Phellia bathybia* and 4 other specimens and also sectioned parts of several others as also of *Phellia bathybia*, *colacca* and *Kadosactis rosea*. The ectoderm of the scapus is rather high, in the tenaculi lower. The cells, probably chitinized, forming the connection between the mesogloea and the cuticle in the tenaculi, are rather few and separated from each other by rather larger parts of common cells; whether gland cells are present or not, I cannot decide, as the cells were badly preserved. Probably the fact that the cuticle is easily deciduous is in relation to the fact that these connections are few and weak. The sphincter (fig. 6) is strong and mostly reticular with tendency to become alveolar (especially in the type-specimen of *Kadosactis rosea*) and the muscle meshes are usually large especially in the upper half. It is separated from the endoderm by a rather thick mesogloea lamella. In one sectioned specimen the distal part of the sphincter was situated close to the endoderm in a part of the column, in another part close to the ectoderm.

The ectoderm of the tentacles is rather high. Their longitudinal muscles are ectodermal, and at the base of the abaxial side very weak or absent (fig. 7), on the alaxial side very strong, especially on the ridges i. e. the tentacles are here longitudinally furrowed. Here the muscle lamella forms densely packed, high folds. In the distal part of the tentacles the longitudinal muscles are weaker and of the same strength on the outside as on the inside. The mesogloea is strongly thickened at the base of the abaxial side in the outer tentacles. Sometimes the mesogloea of the inner tentacles is somewhat thickened at the same place. The ectoderm of the tentacles, oral disc and actinopharynx is pigmented. The radial muscles of oral disc are ectodermal, weak in the vicinity of the actinopharynx, strong with high ramified folds in the outer parts. Only in the radial furrows the muscles are weak.

The ectoderm of the actinopharynx is rather high and provided with nematocysts of different size. In a large specimen they were partly 58-65 \times 5-5.5 μ (numerous), partly 41-55 \times 2.5 μ basitrichs, partly 13-17 \times 1.5-2 μ basitrichs, in a smaller specimen 41-50 \times 1.5 μ , 38-41 \times 2.5 μ , 41-42 \times 1.5 μ . In the types of *Phellia colacca* and *Kadosactis rosea* the large nematocysts were 43-53 \times 4.5-5 μ resp. 43-50 \times 1.5 μ . The larger capsules of the actinopharynx and filaments were often exploded, the length of the shaft and that of the capsule was about the same. They look like amastigophors but because I have seen several ones with a very short thread I think they are *p*-mastigophors the thread of which has usually been broken off. It may be possible that also amastigophors were present, but in such a case they are, as to their size, not different from the *p*-mastigophors. The mesogloea of the actinopharynx is strong in the ridges, in the furrows weak, in the siphonoglyphs thickened.

The mesenteries are generally arranged hexamerously in 3 cycles (6 \times 6 = 12 = 24 pairs) 2 pairs of which are directives. Of 7 specimens examined in detail 5 were normally developed with the 24 pairs regularly arranged. Of these 12 pairs were perfect, the other 12 imperfect, though the strongest of the latter almost reach the actinopharynx. The pairs of the last cycle were not equally developed, some pairs were very weak and do not reach the distal body-end, others are considerably stronger and provided with developed retractors, especially in the distal part. In one specimen the weaker and the stronger pairs alternate, in the other specimens the arrangement was irregular. The different development of the mesenteries of the third cycle was connected with the above named, various number of tentacles. One specimen showed a somewhat irregular arrangement of the mesenteries in



FIG. 1. *Phellia bathybia*. A, detail of the distal body-end showing the mouthparts and the bases of the tentacles; B, detail of a filament showing its structure and the bases of the tentacles; C, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; D, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; E, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; F, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; G, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; H, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; I, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; J, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; K, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; L, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; M, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; N, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; O, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; P, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; Q, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; R, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; S, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; T, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; U, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; V, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; W, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; X, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; Y, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles; Z, detail of the mouthparts showing the upper and lower lips and the bases of the tentacles.



FIG. 2. Mesenteries in the lowest part of the scapus of *Phellia bathybia*.

of the margin. The longitudinal tentacles are very well developed with many high folds (fig. 8) and often branched. The third cycle are provided with long retractions in their distal parts, which are directed towards the exocoels. The tentacles, however, lack pennons

and reach only a little over the inner side of the column. The nematobasilar muscles as well as the basilar muscles are weak. All perfect mesenteries are provided with filaments and reproductive organs, which are lacking in the mesenteries of the third cycle. The size of the nematocysts of the acontia of 8 specimens was as follows:

	Amastigophors	Basitrichs
196-118	5.5-6 μ	53-65 : 3.5-4 (41.5 μ)
101-113	5.5-6	53-60 : 3.5-4.5
91-106	6-6.5	53-61 : 3.5-4.5
101-110	about 6.5	36-53 : 3.5-4.5
88-101	5.5-6	46-55 : 4-4.5
86-98	about 6	36-48 : 3.5-4
86-103	6-7	41-53 : 3.5-4.5
98-110	about 6-7	48-55 : about 4.5

The size of the basitrichs of the acontia of the specimens 1-7 is dubious as the macerated preparations of the filaments certainly contained also nematocysts of the filaments. As I have pointed out (1933 p. 21), I could not previously isolate perfectly the acontia from the filaments. In a well preserved specimen from the Godthaab expedition on the other hand the acontia were not stuck together with the filaments. I think that the smallest size of the basitrichs of the acontia may be about 48 μ . The ciliated tracts of the filaments are well developed and supported by large thickenings of the mesogloea. The intermediate streak seems to be structured as in *Halicampa arctica* (CARLSEN 1921 p. 121-122 Pl. 2 figs. 11, 15). The species is dioecious.

Remarks: *Phellia bathybia* and *violacea* (DANIELSEN 1890) are undoubtedly nothing but *Kadosactis rosca*. The outer tentacles are in all three species provided with thickenings on the outside of their bases. Also their inner organization agrees very well. Identical with *K. rosca* is also GRAVIER's *Hornathia? musculus* (CARLSEN 1931 p. 11). The species is a pronounced arctic deep-sea form always living in 1-2 temperatures.

Kadosactis abyssicola (Dan. & Koren.).

Pl. III fig. 4. Pl. IV fig. 3.

Phellia abyssicola n. sp. DANIELSEN and KOREN 1879 p. 78 Pl. 9 figs. 3-4. APPELLÖF 1891 p. 11. GRIEG 1897 p. 5, 11, 1913 p. 113. APPELLÖF 1905 p. 68.
non *Sapartia (Phellia) abyssicola* D & K. DANIELSEN 1890 p. 30.
non *Sapartia abyssicola* n. sp. VERRILL 1882 p. 311.

Diagnosis: Tenaculi weak, numerous. Cinclides situated only close below the sphincter. Sphincter strong, reticular to somewhat alveolar, not stratified. Tentacles up to about 60, without distinct basal thickenings on their outside, considerably fewer than the mesenteries, the outer tentacles considerably shorter than the inner ones. Usually one siphonoglyph and one pair of directives, sometimes more than 2. More than 12 pairs of perfect mesenteries. Retractors of the mesenteries weaker than in *K. rosca* and their muscles more delicate. Reproduction also by fragmentation. Nematocysts of the column partly 43-58 : 5-6.5 μ , microbasal amastigophors, shaft with about 25 rows of barbs, partly 9.9-17.5 : 2-3 μ , basitrichs, those of the tentacles partly 52-70.5 : 7.5 μ , microbasal amastigophors, partly 21-31 : about 2.5-3 μ , basitrichs, common, those of the actinopharynx partly 41-63.5 : about 1.5-5 μ , microbasal amastigophors?, very numerous, partly 28-36 : 2.5-3 μ , basitrichs, those of the filaments partly 32.5-36 : about 1.2 μ microbasal amastigophors, partly 10.5-13.5 : 2 μ , basitrichs, those of the acontia partly 120-156 : 7-10.5 μ , microbasal amastigophors, partly 71-91 : 5-6(6.5) μ , basitrichs. Spinozooids of tentacles 17 : 1.5-13 : 6.5 μ .

Colour: Cuticle of the scapus greyish-brown, speckled with darker brown spots. When the cuticle is removed the scapus is

pale flesh-coloured. Outer tentacles pale yellowish red, inner tentacles on their lower half and on all the inner surface brownish red, on the outer side considerably paler. Actinopharynx and oral disc dark brown-red, the latter with paler stripes proceeding from the mouth towards the inner tentacles (teste DANIELSSEN and KOREN). A preserved specimen from Hjeltefjord was olive-brown, its actinopharynx brown-red. The cuticle of another specimen from the same locality was dirty yellow, the inner tentacles on their inside and the oral disc brownish-red, other parts uncoloured.

Dimensions of the best preserved specimens from Hjeltefjord: 1) Height 1 cm, largest breadth 1.5 cm, length of the inner tentacles 0.5-0.6 cm. 2) Height 1.2 cm, largest breadth about 2 cm. Of a specimen from Trondheimfjord: Largest breadth 1.1 cm, largest height 1.2 cm, length of the inner extended tentacles about 1.4 cm and of the outer tentacles about 0.7-0.9 cm. A specimen from Hvidingsø was 2.7 cm broad and 1.5 cm high.

Occurrence: Norway: Trondheimfjord 70-150 m, clay ("Gunnerus" 1922), 2 specimens; Røberg 130-180 m ("Gunnerus" 1933) 1 spec.; Nordfjord: Vaagsfjord, Slaaken, and Stegene 188-377 m; off Skavöskallen, 188-226 m (teste GREGG); Bergen: Hjeltefjord, deep water (APPELLÖF), 2 specimens; Korsfjord 171 m (teste KOREN and DANIELSSEN); Hardangerfjord 188-282 m (ASBJØRNSEN); Hardangerfjord (teste GREGG); Digerns Sound (ASBJØRNSEN); Hvidingsøya 151-188 m (G. O. SÆRS); Osløfjord, Braendtangen S. of Hvidsten 151 m, 1 specimen; Norway without distinct locality, 3 specimens.

Exterior aspect: The pedal disc is well developed, wide, rather thick and provided with fragments of a cuticle. The column



Fig. 9. *Kadosactis abyssicola*, Sphincter.

is somewhat conical, wider in the proximal than in the distal part. The scapus is invested with a wrinkled, rather thick but easily deciduous cuticle "interwoven with fine sand and clay" (DANIELSSEN). When the cuticle is removed, the scapus is very rugose, giving it an appearance as if it were provided with larger

and smaller irregular tubercles, which, however, sometimes—in specimens from Trondheimfjord, were inconspicuous to the naked eye. A closer examination of these imperforate tubercles, invested with cuticle, shows that we have to do with tenaculi,



Fig. 10. *Kadosactis abyssicola*, Section of three perfect mesenteries of the upper part of the body.

In the specimen from Røberg the tenaculi, forming irregular elevations, were present (fig. 3 Pl. 4), fragments of such occurred also in the worst preserved specimens. Cinclides are also present here, they are, however, not situated on the tubercles, as DANIELSSEN and KOREN state, but situated close below the sphincter and seem to be at least 2 in certain compartments. In a sectioned piece with 3 pairs of mesenteries of a rather well preserved specimen I observed 2 in each endocoel of the outer pairs but no cinclides in the endocoel of the middle pair. The cinclides seem to be of the same structure as in *Saprotroglocha rapana* (compare this species) but are very narrow in their middle parts, so that they can be easily overlooked in sections. Manifestly they are here in reduction, so that it is not probable that the acontia could be ejected through them. The margin was, in the best preserved specimens, crenulated. At the insertions of the mesenteries the uppermost part of the column, the scapus, was longitudinally sulcated (Pl. III fig. 4).

The tentacles are conical, in the contracted state rather thick, when extended considerably longer, the inner tentacles about twice as long as the outer ones and sometimes longitudinally and transversally furrowed. The number of tentacles is considerably fewer than that of the mesenteries. Four specimens had 40, 44, 54, and 58 tentacles, while the mesenteries were 72, 72, 80, and 90 at the limbus. The tentacles are devoid of basal thickenings. The oral disc is rather wide and its inner part without tentacles, it is radially sulcated. The actinopharynx is rather short and longitudinally

18 arranged hexamerously, the last cycle only in the proximal part of the body. 6 (12?) pairs of perfect mesenteries. Longitudinal muscles of the mesenteries forming retractors only on the mesenteries of the first and second cycles. Retractors rather strong but their folds not, or only in their uppermost part sometimes a little

smooth and provided with longitudinal furrows. The margin is distinct.

The tentacles are short, conical, the outer tentacles considerably smaller than the inner ones. The number of tentacles was 18 (6 + 6 + 12 + 24) in both specimens. DANIELSSEN stated incorrectly that 3 cycles of tentacles, 24 in each, were present. The tentacles are not thickened on the outside of the base. The actinopharynx was transversally furrowed, probably there were 2 siphonoglyphs.

Anatomical description: The ectoderm of the scapus was of ordinary thickness but considerably thinner than the mesogloea, the sphincter (fig. 12) was very strong and broad, occupying a great part of the here thickened mesogloea and wholly separated from the endodermal muscles of the column. The muscle meshes are of ordinary size, often drawn out in a radial direction and showing a tendency to vertical stratification. The longitudinal muscles of the tentacles and radial muscles of the oral disc were ordinarily developed and ectodermal.

I have examined the mesenteries of the same specimen, which DANIELSSEN has dissected. They were hexamerously arranged and formed 48 pairs, the last cycle of 24 pairs only present in the lowest part of the body. As far I can see only 6 perfect pairs were present, while DANIELSSEN says that there were 12 pairs. It must, however, be remarked that the section figured in Pl. 15 fig. 3 has not cut through the actinopharynx but the oral disc, which I have discovered from the remains of the dissected specimen. The longitudinal muscles of the 2 first cycles form rather strong diffuse retractors (fig. 13) sometimes a little concentrated in their uppermost parts. In the main the retractors agree in their appearance fairly well with those of the species described above of *Kadosactis* but not



Fig. 12. *Kadosactis spitzbergensis*. Sphincter.

concentrated. Parietobasilar muscles long but forming few folds. Nematocysts of the scapulus 17-19 \times 2.5-3 μ , those of the tentacles 22.6-29.6 \times (2.4) 2.8 μ , those of the actinopharynx partly 22.6-28.2 \times 3-3.5 μ , partly 11-12 \times 1.5 μ , the latter basitrichs, those of the acontia partly 38-47 \times 5.5-6.5 μ !, microbasal amastigophors, partly 28.2-38 \times 3.5-4.2 μ , basitrichs. Spirocysts of tentacles 21.2-36.6 μ .

Colour: Cuticle of the scapus greenish with a faint violet-coloured substratum, capitulum blue-violet, margin paler, tentacles intense blue-violet with reddish extremities. Oral disc blue-violet with lighter-coloured rays and a lighter-coloured annulus round the mouth (DANIELSSEN).

Dimensions in contracted state: Spec. 1) breadth of the pedal disc 1.1 \times 1 cm, length of the body about 0.7 cm. Spec. 2) breadth of the pedal disc 1 \times 0.8 cm, length of the body 0.3 cm.

Occurrence: 80°03' N, 8°28' E, 475 m. Temperature at the bottom 1.1° (Norw. N. Atl. Exped.) 2 specimens.

Exterior aspect: The pedal disc is well developed and covered with a cuticle. In the contracted state the body forms a low conus. The column is divisible into scapus and scapulus, the former provided with a cuticle, to which foreign bodies are attached. The scapus is probably provided with weak tenacula, because the ectoderm was very low in several places. As the ectoderm was more incrustated here than in other parts, it is probable that the species has tenacula, so much the more as the cells seem to be chitinized in the lower parts of the ectoderm. I have not observed any cirrillides in my sections which, however, does not prove that such may be present. In the contracted state the scapulus is



Fig. 13. *Kadosactis spitzbergensis*. Section of a perfect mesentery at the upper part of the body.

with those of *Sapartiaopton*. They are somewhat weaker on the mesenteries of the second cycle than on those of the first. The longitudinal muscles of the other mesenteries are weak. The parietobasilar muscles form no or some very short folds but are extended rather far towards the distal body end. The basilar muscles are weak but distinct; the filaments of typical structure. The acontia

conical. The scapus is rugose and provided with numerous tenaculi of the same structure as in *D. praelonga* but shorter. At the limbus



Fig. 14. *Sagartiogeton robustus*. Section of a cinclis in the lowest part of the body. On the right a part of another cinclis is visible.

the scapus seems to lack the papillae, at any rate they are very weak. Here cinclides, close situated and arranged in longitudinal rows, are present. The exact arrangement of the cinclides is dif-



Fig. 15. *Sagartiogeton robustus*. Sphincter.

ficult to determine but I have observed between 6 and 2 (seldom 1 cinclis) in each row. I have sectioned a piece just above the upper end of the scapus but not found any cinclides here; it is therefore

probable that the cinclides occur only at the limbus of this species. The scapus is devoid of a cuticle and irregularly wrinkled, the margin is rather distinct.

The tentacles were 170-180 in the uncontracted specimen. They are conical to more cylindrical in the contracted state and rather long, the inner tentacles twice as long as the outer ones. They are close set and arranged in several cycles, but the arrangement is probably irregular as only one or two siphonoglyphs, the latter asymmetrically arranged, were present. More than half of



Fig. 16. *Sagartiogeton robustus*. Section of a perfect mesentery in the region of lowest part of the actinopharynx.

the oral disc is occupied by tentacles. The oral disc is radially sulcated, the furrows correspond to the insertions of the mesenteries. The actinopharynx is longitudinally ridged. The sectioned specimen was provided with a well developed siphonoglyph.

Anatomical description: The ectoderm of the scapus is of ordinary thickness. The cinclides are partly invaginations from the ectoderm, partly evaginations from the endoderm, mostly the first (fig. 14). The tenaculi recall as to their structure those of *D. praelonga*, but the stratified cuticle forms not so many strata as those of these species (PLA fig. 3). The mesogloea of the column is very thick at least in the contracted state of the body and contains numerous cells poor in protoplasm. The circular muscles of the column are ordinarily developed as also is the sphincter. It is broad and extended over somewhat more than the half of the scapulus, ends rather abruptly and is wholly differentiated from the circular muscles of the column. Towards the tentacles it is reticular, in

yard and Delaware Bay in many localities 76-610 fms; 39° 59' 46" N, 70° 18' 30" W, 260 fms; 38° 21' 50" N, 73° 32' W, 193 fms. (Blake-exped.)

Exterior aspect: The pedal disc is broad, attached to pebbles or shells, often clasping the tubes of *Hyalinocia artifex*. The column is elongated in extended state, the middle part narrower than the proximal and distal ones, in the contracted state it forms a broad, low cone, sometimes it is nearly flat. The scapus is provided with a cuticle and especially in the lower part of the body provided with tenacula, which are very distinct when the cuticle is removed. According to VERRILL the scapus of some specimens can be nearly naked, probably the cuticle of such specimens is lost. Cinclides are present below the sphincter and in the proximal part of the scapus. The scapulus is naked and longitudinally sulcated in introverted state of the distal body-end.

The tentacles are slender, acute, the inner considerably longer than the outer ones, in number up to about 96, in the "Ingolf"-specimens fewer and hexamerously arranged. More than half the oral disc is occupied by tentacles. The 2 siphonoglyphs are distinct.

Anatomical description: The ectoderm and mesogloea of the scapus are of ordinary thickness. The cinclides seem to be principally excavations of the endoderm. The endodermal muscles of the column are weak, the sphincter rather strong, not stratified, reticular in its upper part, more alveolar in its lower, long and

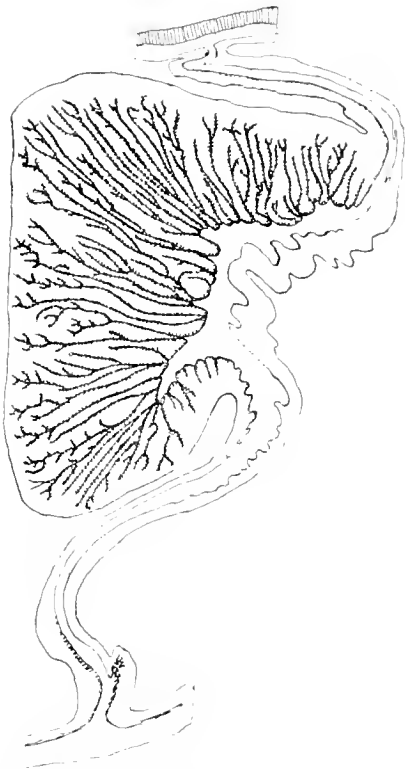


Fig. 18. *Sagartiogeton verrilli*. Section of a perfect mesentery of the first order about in the middle of actinopharynx (U.S. Fish Com.)

usually not occupying half the breadth of the mesogloea and wholly separated from the endodermal muscles of the column. The sections of the sphincter of two specimens, the one taken by the U. S. Fish Commission, the other by the Ingolf expedition (fig. 17), show good agreement.

The longitudinal muscles of the tentacles are ectodermal, at the base about equally developed on the outside as on the inside, the folds like a palisade, are of about same height as the main part of the mesogloea. The radial muscles of the oral disc agree in their appearance with the longitudinal muscles of the tentacles, but are weaker at the insertions of the mesenteries. The ectoderm

of the actinopharynx is rather high and contains numerous nematocysts, the mesogloea is of ordinary thickness, in expanded state of the actinopharynx thin, in the siphonoglyphs thickened.

A sectioned specimen showed the following arrangement of the mesenteries: 12 pairs of the first and second order and 6 of the third were perfect, the other 6 of the third order and those of the fourth imperfect. Counted from the one directive pair the pairs of mesenteries 1, 2 and 4 of the third order were imperfect in both halves of the body. The fourth cycle of mesenteries was almost complete the specimen had 92 tentacles. In another specimen there were also more than 12 perfect pairs. In a third

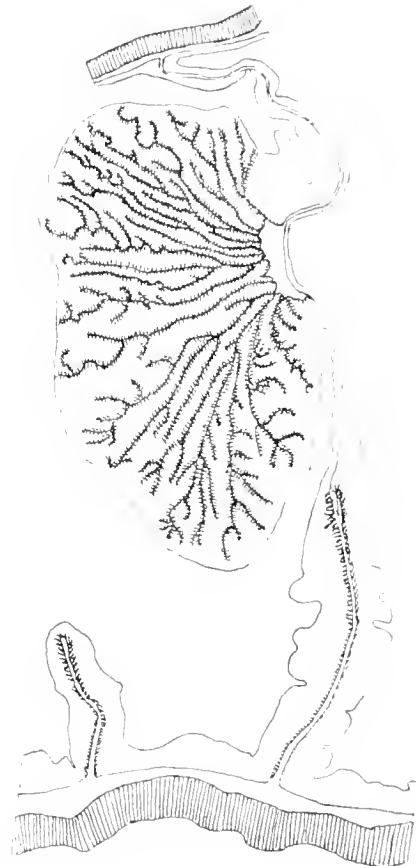


Fig. 19. *Sagartiogeton verrilli*. Section of a directive mesentery and of a mesentery of the third order at the lowest part of the actinopharynx (Ingolf St. 95).

individual I counted 74 tentacles but 124 mesenteries at the base. The longitudinal muscles of the mesenteries form retractors on the mesenteries of the first, second and third order. The retractors are strong in the mesenteries of the first and second order, especially in the former, and kidney-like in their upper part. Also in the middle of the mesenteries the folds of the retractors are high, in the lower part the retractors disappear. I have figured (figs. 18 and 19) the retractors of two specimens. The parietobasilar muscles reach to the upper part of the mesenteries and form a distinct offset on the stronger pairs, but their folds are few and low (figs. 18 and 19). The basilar muscles are weak, oral and marginal stomata present, acontia broad and well developed. All (or almost all?) mesenteries are provided with filaments and reproductive organs. The mesenteries of the last cycle are weak and have filaments and reproductive organs in their lowest part. The species is dioecious.

Sagartiogeton ingolfi Carlgr.

Sagartiogeton ingolfi n. sp. CARLGREN 1928 p. 259, 301.

Diagnosis: Scapus with a cuticle, probably weak and easily deciduous, with scattered weak papillae (tenacula). Cinclides in

with a sphincter. The column is about 80 in number. Mesenteries are about 10. The mesenteries are concentrated in the middle part of the body, and are weak. Paratentacles are weak. Paratentacles muscles are about 14-19.7-3.5 μ . Paratentacles are about 14-19.7-3.5 μ . Paratentacles are about 14-19.7-3.5 μ .



Sphincter.

... of the tentacles partly ... anastigmophors, partly 22-24 ... of the actinopharynx partly ... anastigmophors, partly 25-28 ... anastigmophors, few, partly 24-38 ... 13-17, 2-2.5 μ basitrichs, ... 2.0-2.7, about 1.2 μ micro ... about 3 μ basitrichs, ... anastigmophors, partly 16-2 ... partly 60-88, 8-7 ... 15-22, 3.5-4.2 μ basitrichs, 14-15, 5-5.5 μ .

... 1 expanded, length ... 2 ... 1 ... about 0.7 cm long ... 14-11 N, 32-52 W, 1838 μ ... 18-92, 7 specimens.

Exterior aspect. The examples were very badly preserved and strongly contracted so that I cannot give any good description of the species. The column, certainly divisible into scapus and scapulus as I have observed fragments of a cuticle incrustated with foreign bodies in a specimen, is according to the state of contraction conical or cylindrical. In the latter case, when the body is more expanded, the basal and distal ends are broader than the middle. Moreover, the column is irregularly wrinkled and forms small papillae, certainly tenacula, below the sphincter. On sections I have found enclides in the proximal part of the body as well as below the sphincter. The margin is rather distinct.

The tentacles are about 80 in number (2 specimens examined). Especially the outer tentacles are thin in the expanded state and not provided with thickenings at the base. The inner tentacles are more than twice as long as the outer ones. The oral disc is rather wide and radially sulcated, about half the disc is devoid of tentacles. The actinopharynx is well developed and sulcated. 2 well developed siphonoglyphs, running out in aboral prolongations, are present.

Anatomical description: The ectoderm of the column is high, the endodermal circular muscles are rather well developed. The sphincter is rather strong and not stratified. It occupies a great part of the mesogloea. The muscle meshes are fairly large and uniformly distributed and separated from each other by thin mesogloea-balks (fig. 20).

The anastigmophors of the tentacles are few. The longitudinal muscles of the tentacles are ectodermal, fairly strong and arranged like a palissade. The radial muscles of the oral disc are stronger



Fig. 21. *Saprot-optima imidi*. Section of a digestive mesentery in the uppermost part of the body.

and also ectodermal. Their folds are high between the insertions of the mesenteries and often once branched, at the insertions of the mesenteries weak. The ectoderm of the actinopharynx is rather high and thicker than the mesogloea, also in the ridges that, only in the siphonoglyphs the mesogloea is somewhat thickened.

The mesenteries were irregularly developed in the sectioned specimen. In the distal part 38 pairs of mesenteries were present, in the middle region of the body only 25 pairs and in the lower

part 37 pairs (the mesenteries at the limbus, however, not examined). On the one side of the two pairs of directives there were 1 perfect pairs, on the other side 3¹₂, all with strong muscle penons. Marking the perfect mesenteries with P, the other mesenteries provided with retractors with M, the weaker with W and the weakest with w the arrangement of the mesenteries was as follows (dm: directive mesenteries).

^{dm}
PP, ww, WW, ww, PP, *w*w, WM, *w*w, PP, *w*w, WW, *w*w, PP,
ww, PP, ww, WW, ww, PP, ^{dm}ww, PM, ww, WW, ww, PP, ww,
MM, ww, WM, ww, PP, ww, MM, ww, PP, ww, WW, ww.



Fig. 22. *Sagartiogeton ingolfi*. Section of a directive mesentery and of some other mesenteries in the lower part of the body.

In the middle of the body all ww, except those marked with spaced-out figures, seem to be absent. Also the mesenteries in one and the same pair are often not equally developed. Another individual had about 88 mesenteries at the limbus. The longitudinal muscles of the mesenteries are, especially in the perfect mesenteries, very strong and form concentrated retractors showing in their distal part a characteristic appearance with a large fold directed outwards and separated from the main lamella of the mesentery. In textfigures 21 and 22, I have figured mesenteries sectioned through the distal and proximal parts of the body. The longitudinal muscles are weak inside and outside the retractors as also in the mesenteries marked with W and w. These latter mesenteries are fertile, seemingly devoid of filaments and reach only a short way into the coelenteron. The parietobasilar muscles are distinct, situated on a fold of the mesogloea in the proximal part of the body, in the middle they begin to disappear. Oral and aboral stomata are present. All stronger mesenteries, inclusive the directives, are fertile, in the sectioned specimen with ovaries.

Several nematocysts were exploded but the exploded shaft broken into pieces so that it was difficult to clear up the types of nematocysts. I think, however, that I am right when I interpret the broader and longer nematocysts, having a long shaft, as amastigophors, the other broader with a short shaft as *p*-mastigophors. The amastigophors as well as the basitrichs of the acontia varied. In an individual the former were $71.7 \pm 8.7 \cdot 7.5 \mu$, the latter $55 \cdot 62 \cdot 3.7 \cdot 1.2 \mu$ in two individuals $60 (63) \cdot 66 \cdot 7 \mu$ resp. $48 \cdot 53.6 \cdot 3.5 \cdot 1.2 \mu$.

Sagartiogeton flexibilis (Dm.).

Phellia flexibilis n. sp. DANIELSSEN 1890 p. 51, Pl. 3 figs. 5, 6, Pl. 12 figs. 1, 5.

Diagnosis: Pedal disc wide. Scapus with a cuticle incrustated and well developed, provided with tenaculi. Scapulus with cinclides. Sphincter very strong, alveolar-reticular with tendency to stratification and occupying about two thirds of the breadth of the mesogloea. Tentacles about 36. Pairs of mesenteries 21 ($6 + 6 = 12$). Perfect pairs of mesenteries probably 6, with very strong, strongly concentrated, sometimes kidney-shaped retractors. Folds of the retractors few but rather much branched. Longitudinal muscles of the other mesenteries weak. Parietobasilar muscles weak. Filaments on the mesenteries of the first and second orders.

The nematocysts of the scapus were $42.7 \cdot 45.5 \cdot 2.8 \cdot 3.5 \mu$, often a little curved, probably microbasic amastigophors, those of the tentacles partly $26 \cdot 29 \cdot 3.5 \cdot 1 \mu$, microbasic amastigophors, partly $17 \cdot 24 \cdot 2.8 \mu$, basitrichs, those of the actinopharynx partly $26.8 \cdot 28.2 \cdot 3.5 \cdot 1 \mu$ microbasic *p*-mastigophors?, partly $26 \cdot 27 \cdot 2.5 \cdot 2.8 \mu$, basitrichs, those of the acontia partly $(42?) \cdot 53 \cdot 66 \cdot 5.6 \cdot 7 \mu$ microbasic amastigophors, partly $34 \cdot 41 \cdot 2.8 \cdot 3.5 \mu$, basitrichs. Spirocysts of tentacles up to about $27 \cdot 4.5 \mu$.

Colour: Scapus yellowish-brown, scapulus either almost white or pale rose-red. Outer tentacles pale red, inner tentacles brown, at their bases dark chestnut brown. Oral disc strong brown with a white annulus round the mouth, from which white stripes issue to the tentacles (teste DANIELSSEN).

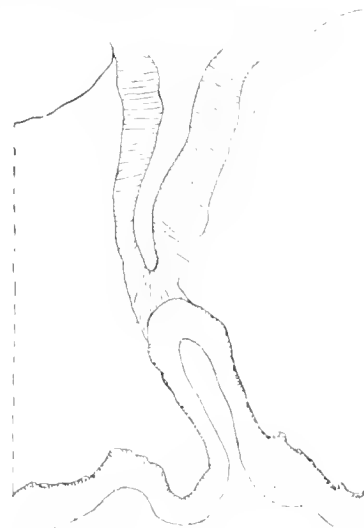


Fig. 23. *Sagartiogeton flexibilis*. Cinclides.

Dimensions of the specimen taken at station 8 in contracted state: Largest breadth of the pedal disc 0.7 cm, height of the column 0.4 cm. — In expanded state 2 cm high (DANIELSSEN).

Occurrence: Sognefjord, 61° 43' N., 6° 36' E., 1189 m. Temperature at the bottom 6.6° (Norw. N. Atl. Exped. St. 4) 1 specimen. 61° 00' N., 4° 19' E., 366 m. Temperature at the bottom 6.6° (Norw. N. Atl. Exped. St. 8) 1 specimen.

XXI. 1952. Type
 Pl. 12, fig. 2 in
 1952. The pedal



Fig. 24. *Saprotrophus flexibilis*. Splincter.

of the oral disc (20-22) at distal body end. The scapus is a well-developed cuticle, incrustated with foreign particles, which were distinctly conspicuous only when the oral disc had been removed. Then they appeared as small granules. The scapulus is rather high and proximal part of it is narrower in section as the splincter I have described above.

The tentacles are about 36 in number as far I can see, but in other specimens were 48 tentacles but this number is not constant. The tentacles are arranged in more than 20 pairs as reported by DAVIESSEN, probably 6-16 pairs in each cycle. The oral disc is not broad, but rather narrow and gradually sulcated. Proboscis is not developed.

The outer, thick, ectoderm of the actinopharynx is very thin. The mesogloea of the actinopharynx is thicker than the ectoderm. The ectoderm is very thin, it is 1 and contains nematocysts. The ectoderm is formed by the formation of enclides which are very thin, they participate to the same layer as the mesogloea, very strong with numerous filaments, forming about two-thirds of the thickness of the ectoderm (Fig. 24). The cellular part is very thin. The longitudinal muscles of the

tentacles are ectodermal. The ectoderm of the actinopharynx is higher than the mesogloea and pigmented.

The perfect pairs of mesenteries were probably 6, as far I can see (in conformity with DAVIESSEN). In a section of about 1/8 of the middle part of the body I counted 3 perfect mesenteries and between those, one pair and a single mesentery, 1 pair of the second and 2 pairs of the third order. Of the latter one pair seems to be weaker than the other and probably does not reach the distal body end. The 6 perfect pairs are provided with very strong, high and often kidney shaped retractors, the folds of which are rather much branched (fig. 25). It is probable that also some of the mesenteries of the second order form distinct retractors although weaker than those of the perfect pairs. In another sectioned part (see above), containing one pair of directives, one pair of the second and two pairs of the third order, the one mesentery of the second cycle was provided with weak but distinct retractors of the same appearance as those of the first cycle. As also filaments and reproductive organs are present in the mesenteries of the first as well as of the second order, there is practically no distinct difference between these mesenteries here. The muscles of mesenteries of the third order are longitudinal and weak, the parieto-basilar and basilar muscles weak. DAVIESSEN stated that the 6 perfect pairs of mesenteries were sterile, I cannot confirm this observation, the perfect mesenteries inclusive the directives were undoubtedly provided with testes, which, however, were little developed. At least certain mesenteries of the second order were also fertile. The specimen was apparently not full-grown.

The nematocysts were usually not well preserved, only in the acontia they were good. Especially it was difficult to distinguish



Fig. 25. *Saprotrophus flexibilis*. Part of a directive mesentery in the fertile region.

the microbasic *p*-mastigophors from the amastigophors. In maceration preparations of the ectoderm of the actinopharynx as well as of the filaments with adjacent parts of the mesenteries I have found oval nematocysts recalling holotrichs and a very large spirocysts 52-7 μ undoubtedly foreign capsules, residues of the food taken (compare CARLIGREN 1940 p. 50).

Sagartiogeton abyssorum n. sp.

Diagnosis: Pedal disc wide. Column smooth, possibly without cinclides. Margin distinct. Sphincter in its upper part reticular, in its lower alveolar, occupying only half of the thickness of the mesogloea. Tentacles between 90 and 100, conical, fairly long, fewer than the mesenteries at the base, their longitudinal muscles ectodermal. Actinopharynx longitudinally sulcated. More than 12 pairs of perfect mesenteries, 2 pairs of directives. Retractors of the mesenteries band-like but often more or less separated from

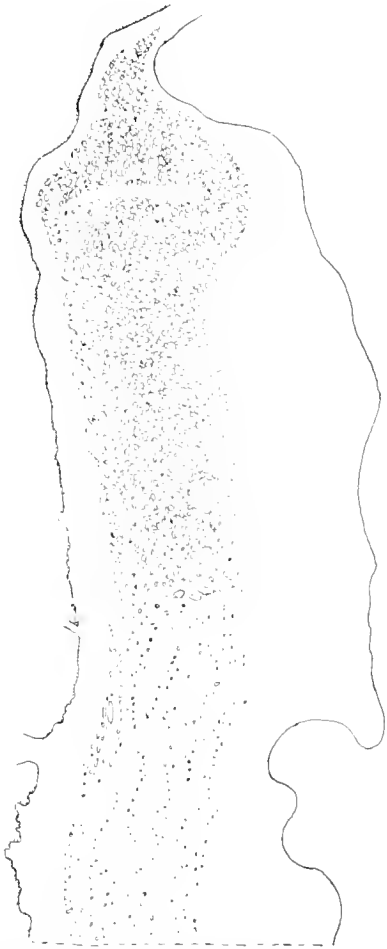


Fig. 26. *Sagartiogeton abyssorum*. Sphincter.

the mesenteries, in extreme cases forming a band-like transversal circumscribed formation. Parietobasilar muscles weak but forming a little shelf. Nematocysts of the column partly 30-36 · 5 μ , microbasal amastigophors, partly 19.7-26.8 · 2.8 over 3 μ , basitrichs, those of the tentacles partly 41-50 · about 5.5 μ , often a little curved, microbasal amastigophors, partly 22-26 · 2.8 μ , basitrichs, those of the actinopharynx partly 35-41 · 5.5 (6) μ , microbasal *p*-mastigophors, partly 31-38 · about 3.5 (1) μ , common, basitrichs, those of the filaments partly 28.2-41 · 4.2-5 μ , microbasal *p*-mastigophors, partly 11.3-15.5 · 1.5-2 μ , basitrichs, those of the acontia partly 72-86 · 6.5-7 μ , microbasal amastigophors, partly 31-55 · 3-3.5 μ , partly 15.5-19.7 · 2 μ , both basitrichs. Spirocysts of tentacles up to about 16 · 5 μ .

Colour unknown.

Size of the best preserved specimen: breadth of pedal disc 1.5-1.3 cm, height of the body about 4 cm.

Occurrence: 62° 06' N, 19° 00' W, 1960 m, Bottom temp, 3.4 (Ingolf exped. St. 64) 2 specimens.

Exterior aspect: Both specimens were strongly contracted, the one very damaged in its upper part. The pedal disc was broad and irregularly folded, that of the damaged specimen drawn out in two portions, the one considerably longer than the other. The column was irregularly wrinkled. I cannot decide if cinclides were present, considering the thickness of the mesogloea they were possibly absent.

Anatomical description: The textfigure 26 shows the structure of the sphincter. The mesenteries were more numerous at the base than at the margin. Owing to the strong and irregular contraction of the pedal disc it was very difficult to decide the exact number of the mesenteries at the base without demolishing the specimen. In about three fourth of the limbus I counted, however, about 100 mesenteries, wherefore the number of them may have been at least 130. The best preserved specimen had 2 pairs of directives, on the one side of the directive pairs there were 8 perfect pairs, on the other 7 $\frac{1}{2}$; thus in all 17 pairs and a single mesentery perfect. The retractors of the perfect mesenteries were very strong and band-like, their folds high and close set and partly richly branched. The muscles issued now directly from the mesenteries (textfig. 27 a), now only the inner parts of the retractor were fastened at the mesenteries while the outer, larger part formed a special lobe, now the retractor were joined with the

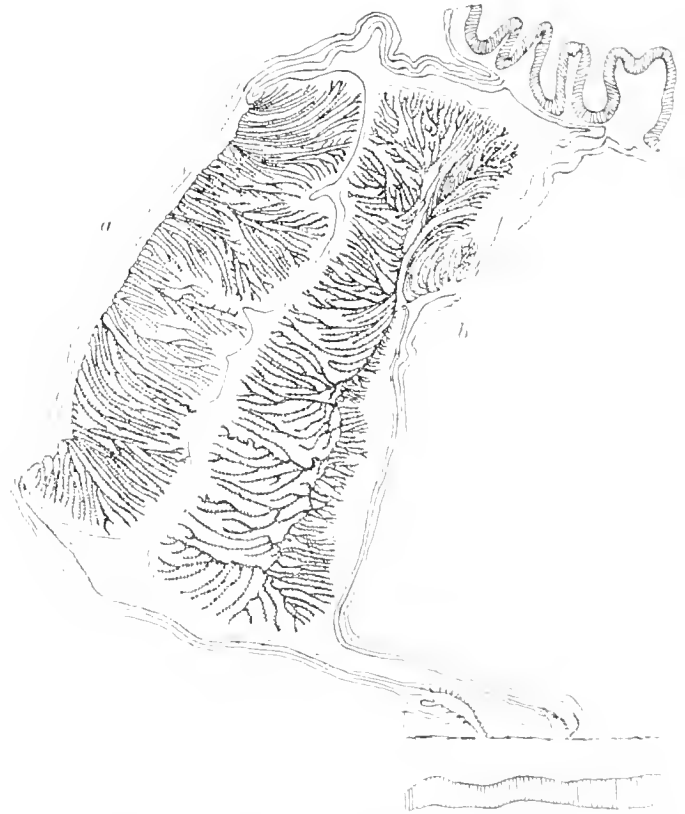


Fig. 27. *Sagartiogeton abyssorum*. Transverse section of a perfect mesenterial pair, the third from the one directive, in the region of the actinopharynx.

mesentery only with a mesogloecal lamella giving the retractor appearance of a circumscribed sphincter drawn out in transversal direction (textfig. 27 b).

Remarks: The species is certainly nearly related to *S. angulif* but may be a distinct species because the retractors and the sphincter show different appearance, and the sizes of the nematocysts are different.

Sagartiogeton (Actinothoe) *undatus* (O. F. Mull.)

(Cuvier)

- Actinothoe undata* O. F. MÜLLER, 1788, II, p. 59, Pl. 63, figs. 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 151, 152, 153, 154, 155, 156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 190, 191, 192, 193, 194, 195, 196, 197, 198, 199, 200, 201, 202, 203, 204, 205, 206, 207, 208, 209, 210, 211, 212, 213, 214, 215, 216, 217, 218, 219, 220, 221, 222, 223, 224, 225, 226, 227, 228, 229, 230, 231, 232, 233, 234, 235, 236, 237, 238, 239, 240, 241, 242, 243, 244, 245, 246, 247, 248, 249, 250, 251, 252, 253, 254, 255, 256, 257, 258, 259, 260, 261, 262, 263, 264, 265, 266, 267, 268, 269, 270, 271, 272, 273, 274, 275, 276, 277, 278, 279, 280, 281, 282, 283, 284, 285, 286, 287, 288, 289, 290, 291, 292, 293, 294, 295, 296, 297, 298, 299, 300, 301, 302, 303, 304, 305, 306, 307, 308, 309, 310, 311, 312, 313, 314, 315, 316, 317, 318, 319, 320, 321, 322, 323, 324, 325, 326, 327, 328, 329, 330, 331, 332, 333, 334, 335, 336, 337, 338, 339, 340, 341, 342, 343, 344, 345, 346, 347, 348, 349, 350, 351, 352, 353, 354, 355, 356, 357, 358, 359, 360, 361, 362, 363, 364, 365, 366, 367, 368, 369, 370, 371, 372, 373, 374, 375, 376, 377, 378, 379, 380, 381, 382, 383, 384, 385, 386, 387, 388, 389, 390, 391, 392, 393, 394, 395, 396, 397, 398, 399, 400, 401, 402, 403, 404, 405, 406, 407, 408, 409, 410, 411, 412, 413, 414, 415, 416, 417, 418, 419, 420, 421, 422, 423, 424, 425, 426, 427, 428, 429, 430, 431, 432, 433, 434, 435, 436, 437, 438, 439, 440, 441, 442, 443, 444, 445, 446, 447, 448, 449, 450, 451, 452, 453, 454, 455, 456, 457, 458, 459, 460, 461, 462, 463, 464, 465, 466, 467, 468, 469, 470, 471, 472, 473, 474, 475, 476, 477, 478, 479, 480, 481, 482, 483, 484, 485, 486, 487, 488, 489, 490, 491, 492, 493, 494, 495, 496, 497, 498, 499, 500, 501, 502, 503, 504, 505, 506, 507, 508, 509, 510, 511, 512, 513, 514, 515, 516, 517, 518, 519, 520, 521, 522, 523, 524, 525, 526, 527, 528, 529, 530, 531, 532, 533, 534, 535, 536, 537, 538, 539, 540, 541, 542, 543, 544, 545, 546, 547, 548, 549, 550, 551, 552, 553, 554, 555, 556, 557, 558, 559, 560, 561, 562, 563, 564, 565, 566, 567, 568, 569, 570, 571, 572, 573, 574, 575, 576, 577, 578, 579, 580, 581, 582, 583, 584, 585, 586, 587, 588, 589, 590, 591, 592, 593, 594, 595, 596, 597, 598, 599, 600, 601, 602, 603, 604, 605, 606, 607, 608, 609, 610, 611, 612, 613, 614, 615, 616, 617, 618, 619, 620, 621, 622, 623, 624, 625, 626, 627, 628, 629, 630, 631, 632, 633, 634, 635, 636, 637, 638, 639, 640, 641, 642, 643, 644, 645, 646, 647, 648, 649, 650, 651, 652, 653, 654, 655, 656, 657, 658, 659, 660, 661, 662, 663, 664, 665, 666, 667, 668, 669, 670, 671, 672, 673, 674, 675, 676, 677, 678, 679, 680, 681, 682, 683, 684, 685, 686, 687, 688, 689, 690, 691, 692, 693, 694, 695, 696, 697, 698, 699, 700, 701, 702, 703, 704, 705, 706, 707, 708, 709, 710, 711, 712, 713, 714, 715, 716, 717, 718, 719, 720, 721, 722, 723, 724, 725, 726, 727, 728, 729, 730, 731, 732, 733, 734, 735, 736, 737, 738, 739, 740, 741, 742, 743, 744, 745, 746, 747, 748, 749, 750, 751, 752, 753, 754, 755, 756, 757, 758, 759, 760, 761, 762, 763, 764, 765, 766, 767, 768, 769, 770, 771, 772, 773, 774, 775, 776, 777, 778, 779, 780, 781, 782, 783, 784, 785, 786, 787, 788, 789, 790, 791, 792, 793, 794, 795, 796, 797, 798, 799, 800, 801, 802, 803, 804, 805, 806, 807, 808, 809, 810, 811, 812, 813, 814, 815, 816, 817, 818, 819, 820, 821, 822, 823, 824, 825, 826, 827, 828, 829, 830, 831, 832, 833, 834, 835, 836, 837, 838, 839, 840, 841, 842, 843, 844, 845, 846, 847, 848, 849, 850, 851, 852, 853, 854, 855, 856, 857, 858, 859, 860, 861, 862, 863, 864, 865, 866, 867, 868, 869, 870, 871, 872, 873, 874, 875, 876, 877, 878, 879, 880, 881, 882, 883, 884, 885, 886, 887, 888, 889, 890, 891, 892, 893, 894, 895, 896, 897, 898, 899, 900, 901, 902, 903, 904, 905, 906, 907, 908, 909, 910, 911, 912, 913, 914, 915, 916, 917, 918, 919, 920, 921, 922, 923, 924, 925, 926, 927, 928, 929, 930, 931, 932, 933, 934, 935, 936, 937, 938, 939, 940, 941, 942, 943, 944, 945, 946, 947, 948, 949, 950, 951, 952, 953, 954, 955, 956, 957, 958, 959, 960, 961, 962, 963, 964, 965, 966, 967, 968, 969, 970, 971, 972, 973, 974, 975, 976, 977, 978, 979, 980, 981, 982, 983, 984, 985, 986, 987, 988, 989, 990, 991, 992, 993, 994, 995, 996, 997, 998, 999, 1000.

Depth 100-150 m. Pedicel disc wide. Column provided with cinclides throughout, not part as well as at the base, and usually (always?) 8-10 pairs, starting from a region below the margin. Margin with 8-10 rather strong alveolar with a little tendency to be complete, some places, separated from the endodermal muscles by a narrow, narrow band of the mesogloea. Tentacles long, 100-150, 150-192, at least originally hexamerously arranged. Longitudinal muscles of tentacles and radial muscles of column well developed. Actinopharynx with longitudinal ridges 100-150, corresponding to the number of perfect mesenteries. Esophagus 2-3, 2-3, rarely 1 or 3, aborally a little prolonged. Mesenteries hexamerously arranged, but the arrangement in the region of the actinopharynx usually disturbed by retarding of certain mesenteries in their growth. As a rule 2 pairs of directives, rarely 3 or rarely 1. Perfect pairs in varying number up to at least 18, some single mesenteries. More mesenteries at the base than at the margin. Retractors of the mesenteries band-like, more or less contractile, rarely circumscribed, those of the directives strongly concentrated, rarely circumscribed. Parietobasilar muscles well developed, not forming a sheet, basilar muscles well developed. A stronger mesenteries incl. the directives fertile. Dioecious, unarmoured. Nematocysts of the column partly 11-22 \times 3.5-5 μ , of the tentacles partly 8.5-17 \times 1.5-3 μ basitrichs, those of the tentacles partly 19-27.5 \times 3.5-4.5 μ microbasic amastigophors, partly 8.5-17 \times 1.5-2.5 μ basitrichs, those of the actinopharynx partly 21-29 \times 3.5-4.2 μ , microbasic *p*-mastigophors, partly 19.7-24 \times 2.5-2.8 μ , basitrichs, those of the filaments partly 12-11.5 \times 4.2-5.5-6 μ , partly 18.3-29 \times 4.5 μ , both microbasic *p*-mastigophors, partly 9.5-17 \times 1.5-3 μ basitrichs, those of the tentacles partly 34-58 \times 5-6.5-7 μ , microbasic amastigophors, partly 22-31 \times 2-2.5-2.8 μ , basitrichs. Spirocyts of tentacles partly 21 \times 4.2 μ .

Coloration. Pedicel disc flesh coloured. Column flesh- or salmon-colored, not greenish when contracted. The parts which are adjacent to the endocoels are colored by red- or olive-brown longitudinal stripes forming longitudinal bands. Upper cinclides reddish-brown, annulus at the limbus light olive-brown. Lower cinclides as in *colantus*. Tentacles light grey to yellowish, annulus at each side provided with an olive green, rarely yellowish, annulus, thickened at the base of the tentacles and forming a distinct marker mark. Inside those on the disc at the margin of the tentacles 2 darker marks which, together with the annulus, form a distinct \otimes mark when the tentacles are contracted. Oral disc black-brown, annulus at the margin orange-colored. The parts of the oral disc which are adjacent to the stronger endocoels are often provided with a yellowish, rarely yellowish white, almost square spots situated at the margin of the disc. Dissections of the mesenteries on the margin of the disc yellow or orange. Sometimes the directive mesenteries are more reddish on the other part of the disc and sometimes the directive mesenteries, the oral disc is crossed by a black longitudinal line, which, together with the directive plane (see *Actinopharynx* of *Actinopharynx* 1935, Pl. XXI fig. 2).

Sizes of two well preserved, large specimens 1) length 2.5 cm, breadth at the base about 2 cm, 2) length 1.5 cm, breadth at the base 3 cm. The longest individual seen by myself was, when living and well expanded, 4 cm long, at the limbus and margin 3 cm broad, in the middle of the body 2 cm; the inner tentacles were about 3 cm long (CARLQVIST 1893).

Occurrence: Norway: Trondheimsfjord, Bergenfjord 32 m; Oslofjord, Hallandspollen, Skagerak: Bohuslän, Kosterfjord between Runno och Storsholm 4.7 m. (Oldevig & Eliasson); between Salto bank and Gulskår (scapus incrustated with sand) 10 m (Wahrberg & Eliasson); between the mainland and Tjerno (Wahrberg & Eliasson); Kornö Brandskären 30 m; Gullnaren-Bjorkholm 26 m, Strommarne, Skatholmen 7 m (Carlqvis).

Kattegat, Stigfjord 12-16 m (Carlqvis, Gustafson, Molander); Koon, Marstrand 38-20 m (Lagerberg); mouth of Göta river (Wahrberg & Oldevig); Frederikshavn (Feddersen, Carlqvis); E. S. E. of Frederikshavn 23-26 m; N. of Borrelberg bank 8 m (Kramp); 3 miles E. to N. of Sahy 11 m (Thor expd.); Limfjord, Livo Bredning N. of Fur (Späreck); Kungsbackafjord 15 m ("Akka" St. 3); off Kungsbackafjord 16 m ("Akka" St. 37); S. S. W. of Nordsto ("Akka" St. 17); W. of Middlegrund 25 m ("Akka" St. 166); S. of Anholt 26 m ("Akka" St. 163); N. W. of Hallands Väderö ("Akka" St. 153); Väderö-Bastad 13.5 fms; Väderö, Sandhamn 10 fms (Lönnberg); Lysegrund (Mortensen); N. N. W. of Lysegrund ("Akka" St. 164); S. W. of Hesselö ("Akka" St. 159); off Aalgaard 10-15 fms (Kramp); Hellebæk; the Sound, Snekkersten 17-21 m; Samsö (Jacobsen); Store Belt (Winther); Fæno, Middelfart (Lütken); Koldingfjord 22 m (Carlqvis); [Bay of Kiel (Möbius)? I have not seen any specimens from this locality.]

Exterior aspect: As far as I can remember all individuals from Bohuslän I have seen in living state were provided with tenaculi. Thus, the column was divisible into scapus and scapulus. Many preserved specimens from different localities had also tenaculi, only a few, badly preserved individuals seemed to lack them, but sections of some of them showed that there were at least traces of such formations present. Therefore I think that the presence of tenaculi is characteristic of the typical *undata*. Usually the scapus is covered with mud, if the mud is partly removed the tenaculi show an appearance of rounded spots covered with a cuticle (figs. 15, 16, Pl. III). When the scapus is wholly deprived of its investment the tenaculi project in form of papillae on the scapus. Rarely other objects than mud are stuck to the scapus. I have, however, seen two individuals, the investment of which consisted of only small grains of sand. Sections of the tenaculi show another appearance than that in other parts of the scapus ectoderm (Pl. VI fig. 6). Two tenaculi are visible in the figure, their ectoderm is probably at least partly chitinized as seen more distinctly in sections of other individuals. In the figured ectoderm that of the tenaculi is fairly high, in sections of tenaculi of other individuals considerably thinner than the other ectoderm. In the ectoderm of the scapus parasites similar to those mentioned in *S. viduatus* (see below) have often been found. The cinclides seem to be sometimes ectodermal but more usually endodermal. Of 8 cinclides examined in three specimens 6 were evaginations of the endoderm, in one cinclis the ectoderm and endoderm took part to about same extent, the 8th cinclis was almost wholly lined by ectoderm.

The number of tentacles seem never (or rarely?) to reach 192. One large specimen had 170 tentacles but 192 mesenteries at the base, another 122 tentacles and 96 pairs of mesenteries at the limbus, 96 mesenteries were present in the whole animal but as the tentacles were 122 it is clear that the younger mesenteries grow from the margin as well as from the limbus, though the latter develop earlier than the former. The mesenteries are certainly arranged after a hexamerous plan, but as of the youngest, stronger mesenteries the one mesentery is often perfect, its partner imperfect and some of them are retarded in their development

the arrangement seems irregular. Usually there are 2 pairs of directives, rarely 1 or 3. Of 27 examined individuals from Stigfjord and Gullmaren 25 had 2 pairs of directives and 2 only 1 pair. Of 6 individuals of a collection attached to a shell of *Ostrea*, 3 had 2 pairs of directives, 2 1 pair and 1 individual 3 pairs of directives. The number of perfect mesenteries varies. I have dissected 6 specimens with 2 pairs of directives. The number and arrangement of the perfect mesenteries were in 1) 18 pairs and 2 single mesenteries - d (directive pair) $8\frac{1}{2}$ in d $8\frac{1}{2}$ - 2) 18 pairs and 1 single mesentery - d 8 d $8\frac{1}{2}$ - in 3) 15 pairs and 1 single mesentery - d 7 d $6\frac{1}{2}$ - in 4) 13 pairs and 2 single mesenteries - d $6\frac{1}{2}$ d $5\frac{1}{2}$ - in 5) 16 pairs perfect - d 7 d 7 - in 6) 13 pairs and 2 single mesenteries - d $6\frac{1}{2}$ d $5\frac{1}{2}$. A specimen from Gullmaren with 1 pair of directives had 16 pairs and 2 single mesenteries perfect. Three specimens from the shell of *Ostrea* had the following number of perfect mesenteries 1) 15 pairs and 2 single mesenteries - d $3\frac{1}{2}$ d $3\frac{1}{2}$ d 6 - 2) 6 pairs and 1 single mesentery - d $2\frac{2}{2}$ d $2\frac{2}{2}$ - 3) 6 pairs and 3 single mesenteries d $5\frac{3}{2}$. The specimens 2 and 3 were small and, however, not sectioned in the uppermost part of the actinopharynx wherefore the statement of the number of perfect mesenteries is somewhat uncertain.

I have examined the nematocysts of the acontia in 46 specimens almost all from Stigfjord in Bohuslän, in 15 specimens the length of some amastigophors were more than 50μ , in a single acontia up to 58μ , the average length of all was 46.38μ . The basitrichs of the acontia have been measured in 32 specimens, the average length was 25.9μ .

var. *anguicomus* (Price).

For literature and synonymy see STEPHENSON 1935 p. 355. Also *Actinothoe anguicomus* Price PAX 1936 p. 111 fig. 139.

STEPHENSON has (1935) given a summary of this form which he regards as a distinct species, however admitting that it may be identical with *undatus*. Of course it is very difficult to decide if the one is a variety of the other or whether both are distinct species. I have seen living *anguicomus* in Helgoland and in Denmark (S. of Esbjerg and Hirtshals) and have also examined a specimen from Plymouth. The small differences between the two forms are to my mind 1) *undatus* is provided with tenaculi on the column, *anguicomus* has a smooth column 2) *anguicomus* grows larger than *undatus* 3) the colour of *anguicomus* varies more than that of *undatus*. I have seen *anguicomus* in Helgoland the colour of which recalls that of *undatus*. On the other hand both forms have sometimes the same black transverse band on the disc. The acontia too seem to have same colour in both. 4) *undatus* has, though rarely, 3 directive pairs or only 1. In *anguicomus* I have found only 2 directive pairs. Too few specimens of *anguicomus*, however, have been examined to decide whether this is a real difference. Of these differences only the first seems to me important. But as I have shown that the usually smooth *laccratus* sometimes has tenaculi, and the anatomy of *undatus* and *anguicomus* agree well, it is to my mind best to regard *anguicomus* as a variety of *undatus* or vice versa but as the latter is the older name the species must be called so.

I have examined some individuals as to their anatomy which is imperfectly known. The sphincter of two large specimens sectioned (from Helgoland and Plymouth) showed same appearance. It was principally reticular and separated from the endodermal muscles of the column by a thin stripe of the mesogloea. 10 specimens (7 from Helgoland, 1 from Plymouth and 2 from Hirtshals) had 2 pairs of directives. The number of perfect pairs of mesenteries seems always to be more than 12. One individual from Plymouth had 18 pairs and 2 single mesenteries perfect - d (directives) $9\frac{1}{2}$ d $7\frac{1}{2}$. Four specimens from Helgoland had the following number of perfect pairs 1) 20 pairs and 2 single mesenteries d $9\frac{1}{2}$ d $9\frac{1}{2}$ - 2) 20 pairs - d 9 d 9 - 3) 18 pairs and 3 single mesenteries - d $9\frac{1}{2}$ d $7\frac{2}{2}$ - 4) 11 pairs and 1 single mesentery - d 6 d $6\frac{1}{2}$, this specimen was small and not fertile. All stronger

mesenteries incl. the directives are fertile. The retractors and the parietobasilar muscles showed the same appearance as those of the typical *undatus*. The nematocysts of the column were partly 17-19 - 3.5μ , microbasal amastigophors, partly 8.5-15.5 - 1.5-2.8 μ , basitrichs, those of the tentacles partly 17-22.6 - about 3.5-4 μ , microbasal amastigophors, partly 12.7-17 - 1.5-2.8 μ , basitrichs, those of the actinopharynx partly 22.6-25.5 - 1.2-1.5 μ , microbasal *p*-mastigophors, partly 19.7-22.6 - 2.5-2.8 μ , basitrichs, those of the filaments partly 12-14 - 3.5-5 μ , partly 22.6-26.8 - 3.5-4.2 μ , both microbasal *p*-mastigophors, partly 11.3-13 - 2 μ , few, basitrichs, those of the acontia partly 39.5-50.8 - 5-5.5 μ , microbasal amastigophors, partly 22-26.8 - 2.5-2.8 μ , basitrichs (all information of the nematocysts from a large specimen from Plymouth, the nematocysts of the tentacles, however, measured in a large specimen from Helgoland). The microbasal amastigophors of the acontia varied between 34 and 54 μ (average size of all 40.65μ) in 18 specimens, the basitrichs between 24 and 29 μ (average size of all 24.6μ) in 16 specimens. The average size of the nematocysts of the acontia was somewhat smaller than in the typical *undatus*. As is to be noted, however, that a large proportion of the *anguicomus* examined were small.

Sagartiogeton (Actinothoe) viduatus (O. F. Mull.).

Actinia viduata n. sp. O. F. MULLER 1776 p. 231, 1788 p. 31 Pl. 63 figs. 6-8; RATHKE 1813 p. 146; SARS 1857 p. 141. (Laurker 1861 p. 191.

A. Isacmaca viduata EHRENBURG 1831 p. 31. (Örsted 1814 p. 74.

Paractis? viduata MULL. p. p. MILNE-EDWARDS 1857 p. 259.

Cylista viduata MULL. p. p. ANDREES 1883 p. 359.

Sagartia viduata O. F. MULL. PETERSEN 1892 p. 172, 176; CARL-GREX 1893 p. 88 Pl. 4 fig. 2. Pl. 6 figs. 7, 8, textfig. 26; GRIEG 1913 p. 143; BLEGGVAD 1914 p. 86 p. p.; BROCH 1927 p. 6.

Actinothoe viduata MULL. STEPHENSON 1935 p. 312. PAX 1936 p. 115 fig. 138; CARL-GREX 1910 p. 41 fig. XIII 6-11

A. filiformis n. sp. RAPP 1829 p. 57 Pl. III figs. 2, 3. SARS 1835 p. 3.

Diagnosis: Pedal disc wide. Column smooth, always without tenaculi, with cinclides in its upper part as well as at the limbus. Upper cinclides (ectodermal invaginations) irregularly arranged in number 0-3 from each endocoel of the three oldest cycles. At the limbus 1-3 cinclides from the oldest endocoels. Sphincter ordinarily developed, extended, filling up about half of the mesogloea, reticular to alveolar, wholly separated from the endodermal muscles of the column. Tentacles conical, thin, long, in extended state considerably longer than the column, regularly hexamerously arranged in number up to 192. Actinopharynx with numerous longitudinal ridges. Always 2 siphonoglyphs without aboral prolongations. Pairs of mesenteries up to 192, hexamerously arranged, 2-3 cycles perfect, 2 pairs of directives. The youngest cycle only in the distal and proximal parts. Retractors of the mesenteries diffuse with ordinary high folds, in the region of the actinopharynx situated in the middle of the mesenteries. Retractors of the directives stronger than those of the other mesenteries and situated especially in their upper part close to the actinopharynx. Parietobasilar muscles fairly weak, basilar muscles distinct. At least the three first cycles of mesenteries with filaments, acontia, and gonads. Acontia thick. Never asexual reproduction. Nematocysts of the column partly (22) 24-27 - 4-4.5 μ , microbasal amastigophors, partly 10-19 - 1.5 over 2.5 μ , basitrichs, those of the tentacles partly (20) 22-26 (20) - about 3.5 μ , microbasal amastigophors, partly 15.5-19 - about 2.8 μ , those of the actinopharynx partly 22-24 - 4-4.5 μ , microbasal *p*-mastigophors, partly 14-26 - 2-2.5 μ , basitrichs, those of the filaments partly 17-20 - 4 μ , partly 24-27 - 4.5 μ , both microbasal *p*-mastigophors, partly 13-15 - almost 2 μ , basitrichs, those of the acontia partly 34-62 - 5.5-6.5 μ microbasal amastigophors, partly 22-29 - about 2.5 μ , basitrichs. Spirocysts of tentacles up to about 23 μ .

...s. at the distal end of the column, ...to the ...of the ...by an olive ...site provided ...somewhat broadened ...with the ...coloured. Within the base ...spot of same colour as the ...of 12-21 opaque grey, ...to the endocoels ...tertiles and the actino- ...stronger coloured than ...X 260, fish or yellowish white. Gonads

...column 5 cm long and 1.5 cm ...Fertacles at least as long as

- (1) ...Narvik (Tredøherred, Eidsboth) on *Zostera*;
- (2) ...Bergen (Flor) Manger (teste Sars); Hjelte-
- (3) ...Havregerthord on *Zostera* (teste Gregg);
- (4) ...Kjøbenhavn (Zostera) (teste Broch); Drøbak on
- (5) ...Sweden; Gullmarön, Väderöar on
- (6) ...Kosterfjord, Rasso on *Zostera*
- (7) ...Denmark; Lundfjord; Nykolung; Liva
- (8) ...off Glyvigore 23 m (Kramp); Hol-
- (9) ...Kattegat without distinct locality
- (10) ...Mortensen. The species lives usually on
- (11) ...in the vicinity
- (12) ...on floating *Zostera*

The exterior of the species has been de- ...1893. The always smooth column is usually ...*Zostera*, rarely pillarlike. As to the ...principally invaginations of the ecto- ...In a large specimen I have ...of the enci- ...0, 2, 2, 2, 3, 1, 2, 3, 3, 2, 3, 1, 1, 2, 2, 0, 2, 1, 3, ...At the base one enci- ...the older endocoels.

As to the description, see CARLGRÉN 1893. As to the ...principally to be invagina-



Fig. 28. Enci- ...at the limbus.

...from the endoderm ...I cannot decide with ...the endodermal ...textfig. 28).

At any rate we have to do at least with praeformed enci- ...In these toterm of the column sometimes encysted parasites, probably gregarines, occur (textfig. 29). They were situated about in the middle of the ectoderm or somewhat approached to the mesogloea. As in other species of *Sagartiogeton* the younger mesenteries grow from the base upwards as well as from the margin downward, but the mesenteries at the base seem to arise a little earlier than those

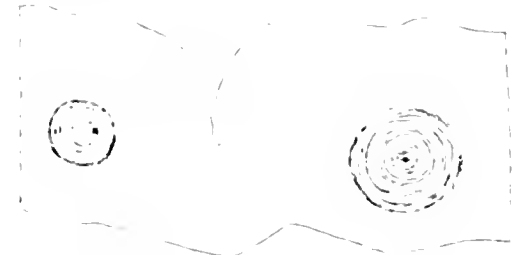


Fig. 29. *Sagartiogeton rubratubus*. Encysted Gregarines in the ectoderm of the column.

at the margin. The retractors of the non-directives are band-like, in the directives concentrated and in the region of the actinopharynx situated close to it. The acontia are thick. The species is dioecious, the male and female gonads are ripe at the end of June in Bohuslan. The species never propagates asexually, but pieces of the basal part regenerate easily (see CARLGRÉN 1901, 1909).

Remarks: The species is not identical with *Sagartia rubrata* of GOSSE (see also STEPHENSON 1935 p. 357). Also the species mentioned by SCHULZE (1875), HARTLACH (1890) and PAX (1928) from Helgoland is not our species but *amphicemus*. The *rubrata* too from Bulk (MÖHR'S 1873) and from other coasts of Germany is probably not this species. This is also the case with the species mentioned by ØRSTED 1814 and LITCKEN 1861 from the Sound. It is, however, possible that *rubratubus* at these times was present there because we must always take into account that the fauna has changed from time to time in such an area as the Sound. Also the individuals enumerated by LEVINSSEN 1893 from Danish waters were not *rubratubus*. It is likely that the species shortly described by RAPP 1929 as *A. filiformis* n. sp. and noted by SÆRS 1835, is this species although the figures are not good; but the species undoubtedly had enci- ...

Sagartiogeton (Actinothoë) laceratus (Dal.).

Pl. III fig. 13.

For literature and synonyms see STEPHENSON 1935 p. 347. Also *Sagartia unilata* (O. F. M.) LÖNNBERG, 1898 p. 55, 1902 p. 59 p. p. *Actinothoë lacerata* Dal. PAX 1935 p. 124 fig. 118; CARLGRÉN 1910 p. II fig. XIV 1-5.

Diagnosis. A relatively small species. Pedal disc broad and undulated in connection with the asexual reproduction. Column usually smooth but sometimes with weak tenaculi distributed over the most part of the column. Enci- ...absent at the limbus, principally endodermal evaginations. Splinter strong, alveolar to reticular, separated from the endodermal muscles of the column by a thin band of the mesogloea, at its proximal end not pointed. Tentacles fairly long but shorter than those of *rubratubus* and *unilatus*, conical, in number up to about 140-150, usually considerably fewer, hexamerously, sometimes octomerously or decamerously arranged, 2, rarely 3 siphonoglyphs. Perfect pairs of mesenteries typically 12 with concentrated diffuse retractors. Parietobasilar muscles weak, basilar muscles distinct. Asexual reproduction by laceration (constriction). Development of the mesenteries in the lacerated pieces basal. Acontia well developed, thick. Nematocysts of the column partly 19-25 - about 3.5 μ , often curved, microbasal anastigophores, partly 11-18 - (2) 2.5 μ , basitrichs, those of the

tentacles partly 22-29 \times 1-1.5 (5) μ , microbasal amastigophors, partly (14) 19-22 (28) \times 2.5 μ , basitrichs, those of the actinopharynx partly 26-29 \times 3.5-1.5 μ microbasal *p*-mastigophors, partly (22) 23-27 \times 2.5-3 μ , basitrichs, those of the filaments partly 10-13 \times 4-1.5 μ , partly 22-26 \times 1-1.5 μ , both microbasal *p*-mastigophors, partly 11-15 \times 1.5 μ , basitrichs, those of the acontia partly 36-60 \times 1-5.4 μ microbasal amastigophors, partly 21-31 \times 2-2.6 μ basitrichs. Spirocysts of tentacles up to about 21 \times 1 μ .

Colour: I have noted the following colours in some specimens: Column white with pale brown-red longitudinal bands. Cinclides white. Tentacles in expanded state greyish with 5 grey to yellowish-

("Akka" St. 74); W. of Morup 16 m ("Akka" St. 103); S.W. of Galtabaek 30 m ("Akka" St. 131); S.W. of Store Middelgrund 33 m ("Akka" St. 178); S.W. of Lysegrund 35 fms (Mortensen), Hellebæk (Kramp), The Sound: S.W. of Sofiero 26-32 m (Lomborg), Raa 16 fms (Lomborg).

Exterior aspect: The pedal disc is wide and typically strongly undulated (fig. 43 Pl. III) in connection with the asexual reproduction by constriction. MARY STEPHENSON (1925 p. 891) characterises the species (*vaccina*) "among other things by its lacinate basal outline and constant habit of asexual reproduction by basal fragmentation". Whether this habit is constant is question-

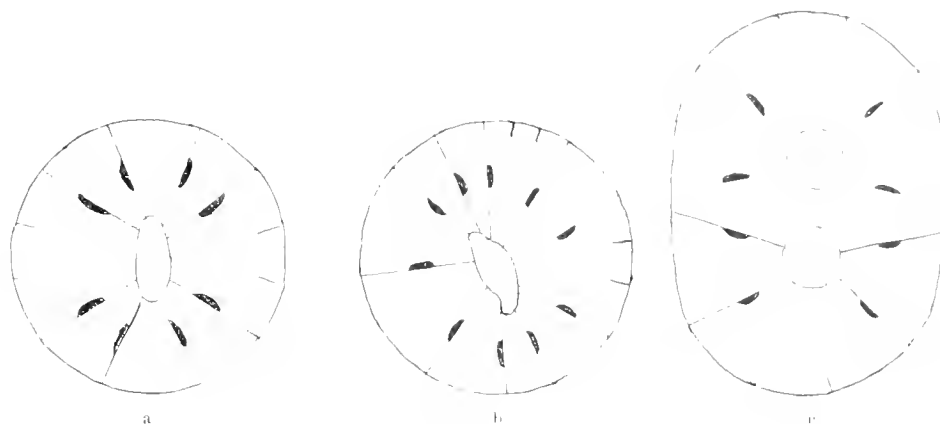


Fig. 30. *Sagaractipton lacustris*. Schematic arrangement of mesenteries in lacinated pieces.

white cross-bands. On the inside of the tentacles a brown longitudinal line and a more or less distinct dark brown B spot at the base. Oral disc dark brown with yellowish radial lines or yellowish-white with dark brown radial stripes. One individual with a brown annulus around the mouth (specimens from Rotvold bay, Trondheimfjord).—Column pale salmon red with brick-red longitudinal band sometimes appearing as spots, tentacles greyish with 2 or 3 white or yellowish-white transverse bands, between these an area of lilac especially distinct in contracted state. B spots distinct at the base of the inner tentacles, brown, on the outer tentacles at same place a brown band. Oral disc paler or darker with yellowish, in the outer parts orange-coloured insertions of the mesenteries. Often 6 more or less marked, orange-coloured radial bands. Actinopharynx pale, little shading in orange (specimens from Gullmaren). See further Gosse (1860 *Sagaritia vaccina*) and STEPHENSON (1935).

Dimensions: Diameter of base up to 1.5 cm. Largest well preserved specimen length 2.2 cm, oral disc about 2 cm. According to STEPHENSON the disc and the tentacles can reach 3 cm, and the height of expanded animal 2.2 cm or more. The Scandinavian individuals are usually smaller.

Occurrence: Norway: Foldenfjord 9-27 m (Sot-Ryen); Trondheimfjord, Rotvold Bay 100-10 m (Carlgren); He Bay 50 m (Petersen); Elandwaagen 6-15 m; Fosenwaagen 25-30 m; Sodenæs 100 m (Mortensen); Bjugnbotn 5-10 m; Rissen 15-13 m (Östergren). Dröbak, Skibkalla 30 m (Broch). Sweden: Bohuslän Strömstallfjord 8-15 m; N.W. of Nord Koster 30 m; Kosterfjord between Flatskär and Danielknimmel 25-50 m; $\frac{1}{2}$ mile S.W. of Klövskär, 35 m (Wahrberg & Eliasson); between Gaveskär and Knippelholmen 11 m (Wahrberg & Oldevig); Styro 35 fms. Andöholm (Olsson); Kornö Brandskär 30 m; Gullmaren off Langegap 35 m; off Lysekil 60 m; Björkholmen (Carlgren); Bonden Tova; Stigfjord (Carlgren & Gustafsson, Molander); Kattegat: Frederikshavn S.E. of Hirtsholm 11 fms (Carlgren); 6 miles E.S.E. of Frederikshavn 23 m (Kramp); west coast of Læso Channel 36 m (Kramp); Læso Channel 35-15 m (Carlgren); Kungsbackafjord 27 m, 25 m ("Akka" St. 6, 7); W. off Traslov 54 m

able though as a rule it is so. The column is as a rule smooth, sometimes, however, provided with weak tenaculi in most part of it. I have seen several individuals from different localities in Bohuslän and at Frederikshavn having such formations. STEPHENSON (1929 p. 128 and 1935) has referred *Phellia pecta* Gosse to *A. lacustris*. According to Gosse (1860) this species was provided "with a very thin membranous epidermis" extending about half-way up the column. Perhaps it was a *lacustris* furnished with tenaculi. STEPHENSON (1935) mentions, however, that he never has observed any tenaculi in the British individuals of *lacustris*.

In the upper part of the column there are few cinclides present, usually only one, sometimes two communicating with the older endocoels. At the base cinclides seem to be absent. The tentacles are usually hexamerous, sometimes octomerous, decamerous or irregularly arranged. Their number varies considerably but seems never to reach 192. In a very large individual I counted 75 tentacles in about the half of the animal. Usually they are fewer than 100. They are considerably shorter than those of *calvatus* and *undulatus* also in extended state. The actinopharynx is longitudinally ridged, the siphonoglyphs, usually 2 rarely 3, are well developed. The size of the species is smaller than that of *calvatus* and *undulatus*.

Anatomical description: The arrangement of the mesenteries is as a rule hexamerous but sometimes octomerous or decamerous. The arrangement varies so much that it seems as if never a single directive pair should be present in this species. Of 26 specimens examined from different localities 21 had 2 pairs of directives symmetrically or almost symmetrically situated, 2 individuals were also provided with 2 such pairs but separated from each other only by a perfect common pair, 3 specimens had 3 pairs of directives, in 2 of these two pairs of directives were situated as in the latter specimens, the third pair was about opposite the two others, in the third specimen the three directive pairs were more symmetrically arranged. The difference in this respect between *lacustris* and *Sagaritia elegans*, which latter species often has only a single directive pair, is certainly correlated with the different mode of asexual reproduction in the two species. In *elegans* new

was long and occupied the most part of the scapulus. It seems to be somewhat reticular in its upper part (the preservation of the sphincter was bad here), in its lower part, where the mesogloea

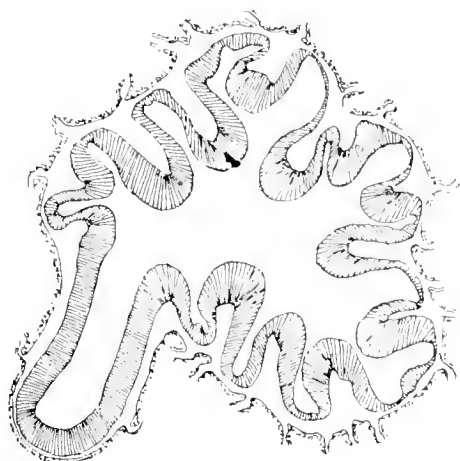


Fig. 32. *Sagartiogeton tubicolus*. Transverse section of the actinopharynx.

was thick, it were alveolar (textfig. 31). The ectodermal muscles of the tentacles show coarse folds, the radial muscle folds of the oral disc were closer set. The actinopharynx was of the usual struc-



Fig. 33. *Sagartiogeton tubicolus*. Section of a perfect mesentery at the end of the actinopharynx.

ture, the longitudinal ridges of the ectoderm pass over into the endoglandular tract of the filaments. The figure 32 shows a section of the actinopharynx with the single siphonoglyph.

The mesenteries were irregularly arranged in correspondence

with the presence of only one pair of directives. There were 23 mesenteries perfect and several imperfect in almost the whole region of the actinopharynx, I have, however, not examined the uppermost part of the body, possibly some more mesenteries were perfect there. Of the perfect mesenteries 10 formed pairs, three mesenteries were single. At the limbus I counted 120-130 mesenteries, at the margin the mesenteries may have been 72 as the specimen had 72 tentacles. In the middle part of the body the mesenteries seemed to be fewer. Thus, the mesenteries grow from the margin as well as from the limbus but certainly more rapidly from the latter. The retractors of the perfect mesenteries were concentrated in their upper part and provided with high folds (fig. 33). The mesogloea of the mesenteries was thickened, especially in the lower part of the specimen, and the parietobasilar muscles situated on a distinct fold of the mesogloea (fig. 33). The nematocysts of



Fig. 34. *Sagartiogeton tubicolus*. Section of the column with a tenaculum (specimen from Stjernevandsund).

the scapulus were partly 25.4 29 - 4.5 5.5 μ , microb. amastig., partly about 11 - 2.8 μ , basitrichs, rare, those of the tentacles partly 28.2 32.1 - 4 5.6 μ , microb. amastig., partly 19.7 22.6 - 2.8 μ , basitrichs, those of the actinopharynx partly 31 33.8 - 4.2 5 μ microb. *p*-mastig., partly 25.4 28.2 - 2.8 μ , basitrichs, partly 11.3 14 - 2 μ , basitrichs, those of the filaments partly 21 29.6 - 4.2 5.6 μ , microb. *p*-mastig., partly 11.3 14 - 4.2 μ , microb. *p*-mastig., partly 11.3 13.1 - (1.5) 2, basitrichs, those of the acontia partly 11 60 - 5.6 μ microb. amastig., partly 19.2 - 15.1 - 4.2 μ , basitrichs. Spirocysts up to about 33 - 5 μ .

Since I have examined the specimen described above I have had the opportunity to examine several specimens taken at two localities, Stjernevandsund and Rognsvandsund, situated not far from each other in the Finmark. The specimens from Stjernevandsund agree in their exterior and anatomy and also in the size of the nematocysts fairly well with those taken in Hjeltvedfjord, so that they may belong to one and the same species. Also the specimens from Rognsvandsund resemble the named specimens so much that I am inclined to place them together but as the microbasal amastigophors and *p*-mastigophors were considerably larger here, it may be possible that it is a distinct species. I have, however, at present referred them together, the specimens from Rognsvandsund as a variety *minor*.

The exterior of the Stjernevandsund and Rognsvandsund specimens agrees. The pedal disc was very wide, the column in contracted specimens conical or in more expanded state more or less cylindrical, though the base was considerably broader than the distal part. The column seemed to be quite smooth also by examination with a lens, but microscopic examination of one Stjernevandsund specimen sectioned completely showed that there were small tenaculi here (fig. 34). Probably there had been a very thin cuticle also in other specimens though it had been torn off during the dredging. The tentacles were in 3 specimens from Stjernevandsund 66, 80 and 118, in 4 specimens from Rognsvandsund 60, 62, 84 and 96, the mesenteries at the base 180, 156 and 220 resp. 92, 128 and 96

of the specimens from Hjeltefjord. The retractors were not observed with several specimens. The specimens from Stjerneøysund had a single siphonoglyph. Rognsund likewise, had a single siphonoglyph situated. The specimens from Hjeltefjord I have sectioned the mesenteries at each locality; it seems that the specimens from Hjeltefjord. The retractors were not observed in the section of the specimen from Hjeltefjord. The retractors were not observed in the section of the specimens,



FIG. 1. Section of a perfect mesentery at the mesenteric plane in specimen from Stjerneøysund.

Remarks: In the specimens from Hjeltefjord the folds of the mesogloea were more numerous. Figure 1 shows a section of a perfect mesentery of a specimen from Stjerneøysund. The stronger mesenteries, incl. the retractors.

As in Figure 1, the comparison of the size of the nematocysts from the specimens from Stjerneøysund and Rognsund. The nematocysts from the specimens from the Stjerneøysund specimen were partly 14.2-17.9 μ , the largest was anastig., partly 18.3-22.6 = 3(3.5) μ (1 spec. exam.), those of the tentacles partly 24-33 = 4.5 μ , (1 spec. exam.), those of the filaments partly 22.6-24 = 2.5-2.8 μ (1 spec. exam.), those of the actinopharynx partly 24-31 = 4-5.6 μ , (1 spec. exam.), those of the siphonoglyph partly 19-21 = 4.2-5 μ , microb. basitrichs partly 22-29.6 = 2.5-2.8, basitrichs partly 26-32 = 4.5-5 μ , microb. amastig. partly 12.7-14.1 = 3.5-4.2, microb. amastig. those of the acontia partly 38-48 = 3-4.5 μ , microb. amastig. partly 38-48 = 3-4.5 μ .

The specimens from Rognsund were: those of the filaments partly 11-12 = 1.6-1.9 μ , microb. amastig. (1 spec. exam.), those of the tentacles partly 31-41 = 4.5-5 μ , microb. amastig. (1 spec. exam.), those of the filaments partly 19.7-24 = 2.8 μ (1 spec. exam.), those of the actinopharynx partly 33-42 = 5-5.6 μ , microb.

p-mastig. (1 spec. exam.), partly 24-27 = 4.2-5 μ , microb. *p*-mastig. (1 spec. exam.), partly 24-32 = 2.5-2.8 μ , basitrichs (1 spec. exam.), those of the filaments partly 34-43 = 5 μ , microb. *p*-mastig. (3 spec. exam.), partly 14.4-19.7 = 3.5-4.2 (5) μ , microb. *p*-mastig. (2 spec. exam.), partly 11.3-11 = 1.5 μ (1 spec. exam.) those of the acontia partly 62-82 = 5.6 almost 7 μ , microb. amastig. (6 spec. exam.), partly 11-53 = 3.5-4.2 μ basitrichs (6 spec. exam.). As we see the microbasal amastigophors and the microbasal *p*-mastigophors were considerably longer in the specimens from Rognsund than those from the other localities as the following comparison of the average length shows:

	Acontia micr. am.	Filament mpm.	Tentacles micr. am.	Actinopharynx mp·m?
Hjeltefjord	59.5 μ	26.3 μ	39.3 μ	32.9 μ
Stjerneøysund	57.5 μ	28.4 μ	28.8 μ	28.5 μ
Rognsund	71.8 μ	38.3 μ	37.75 μ	38.4 μ

The average size of the basitrichs of the acontia was 42.8, 42.97 and 46.75 thus, a fairly little difference. It is to be noted that the smallest specimen from Rognsund (see above: size) had larger nematocysts than those of the largest specimen from Stjerneøysund.

The occurrence of a single siphonoglyph in so many specimens indicates that asexual reproduction, probably by laceration, is prevalent in this species.

Remarks: I have with some hesitation identified the specimens with *Phellia tabacola*, Kor. & Dan.

Genus *Phellia* Gosse.

Sagartiid with broad pedal disc. Column without cinclides, divisible into scapus and scapulis, the former with a cuticle and with somewhat modified tenacula forming distinct wart-like papillae covered with a very thick cuticle. Sphincter fairly well developed, long, mesogloea, wholly separated from the endodermal muscles of the column. Tentacles at least in younger individuals fewer than the mesenteries, the inner tentacles longer than the outer ones. Longitudinal muscles of tentacles and radial muscles of oral disc ectodermal or meso-ectodermal. Siphonoglyphs varying, as also the pairs of directives (1-4). Younger mesenteries growing from the base upwards. Arrangement of the mesenteries often irregular owing to asexual reproduction (laceration or longitudinal fission?). Perfect pairs of mesenteries usually more than 6, sometimes fewer, with strongly concentrated to almost circumscribed retractors and well developed parietobasilar muscles forming a distinct strong fold. The stronger imperfect mesenteries with or (in younger individuals) without retractors. Basilar muscles distinct. Filaments, acontia and gonads from the first cycle of mesenteries (incl. the directives) onwards. Acontia well developed provided with microbasal amastigophors and basitrichs. Cnidom: spirocysts, basitrichs, microbasal amastigophors and microbasal *p*-mastigophors.

The recovery of Gosse's *Phellia gausapata* by STEPHENSON and the examination of this species by STEPHENSON as well as by myself — I have examined a specimen from Sklinna, Norway and lately an individual from Iceland — have disturbed our previous idea about the genus *Phellia*. The anatomy of this species, which STEPHENSON (after stating that the first named *Phellia*, *P. noronhaiata*, is identical with *Sagartia troglodytes*) regards as type of the genus, agrees more with the genus *Sagartia* than one previously has suggested, so that it is not necessary to retain the family Phellidae. Numerous species placed in the genus *Phellia* have been described but belong to the genus *Telmatactis* or to several other genera. In my paper on the Actinaria of the German Deep-Sea expedition (p. 236, note) I have stated that *Sagophellia* and *Phellia* are synonyms. In fact, at present we can refer only *Sagophellia aucklandica* Carlg., *Hormathia arlex* McMurr., and

possibly *Symphellia dubia* Carlgr. to the genus *Phellia*. All these species live in northern boreal or subantarctic waters.

The genus, I think, is nearly related to *Kadosactis*. Both genera are characterized by irregularity in the mesenteries and the number of siphonoglyphs, though *Phellia* is more aberrant in these respects, with sometimes 4 pairs of directives and 4 siphonoglyphs. There are, however, some differences, the tenaculi are of different structure in the two genera and *Kadosactis* is provided with cinclides which are lacking in *Phellia*.

***Phellia gausapata* Gosse.**

Pl. V fig. 2.

Literature and synonyms see STEPHENSON 1935 p. 371 Pl. XXI fig. 5, Pl. XXII fig. 1, textfigs. 75, 76, 99 D, 106. Also CARLGRÉN 1930 p. 7; PAX 1936 p. 111.

Diagnosis: Pedal disc fairly broad, at least in younger individuals broader than the oral disc. Tenaculi with a very strong cuticle. Sphincter thin but long, at least half of it situated in the scapus. Tentacles in variable number, often irregularly arranged. Actinopharynx with 1-3 not strong siphonoglyphs, 1-3 pairs of directives. Perfect pairs of mesenteries varying from 4 to 8, sometimes one mesentery perfect, its partner imperfect. Mesenteries, at least in younger examples, more numerous than the tentacles, arranged in 4 or 5 (6) cycles. Retractors of the stronger mesenteries varying from concentrated diffuse to more circumscribed. In older individuals also the mesenteries of second and third orders, are provided with retractors, in younger ones not. Filaments at least on all older mesenteries. Gonads from the oldest mesenteries onwards, in younger individuals on the first and second cycles. Acontia well developed. Their basitrichs somewhat larger than the amastigophors. Nematocysts of the scapus partly $11.3-20 \times 3.5-4 \mu$, microbasic amastigophors, partly $11-17.6 \times 3.2-4.2$, those of the scapulus $14-18 \times 3.5-4 \mu$, those of the tentacles partly $11-18 \times 3.2-4 \mu$, microbasic amastigophors, partly $16.6-22.6 \times 2-2.5 \mu$, basitrichs, those of the actinopharynx partly $21.2-25 \times 3.8-4.6 \mu$, microbasic amastigophors?, partly $21.6-29.2 \times 2-2.6 \mu$, basitrichs, those of the filaments partly $10-14 \times 2.8-3.5 \mu$, microbasic *p*-mastigophors, partly $11.3-17.6 \times 1.5-2 (2.5) \mu$, basitrichs, those of the acontia partly $27.5-37 \times 3-3.5 \mu$, microbasic amastigophors, partly $30-43 \times 3.5-4.6 \mu$, basitrichs. Spirocysts of tentacles up to $31 \times 4 \mu$.

Colour: "Lowest part of scapus cream coloured, highest part greenish grey. Scapulus pale flesh-colour tinged with purple, longitudinally streaked with dull purple, the streaks irregular. Tentacles translucent, grey, marked with opaque cream and dull brick-red. Primary tentacles with a transverse cream bar halfway up, interrupted in the middle and with red areas above and below it, secondary tentacles similar to the primaries but with an additional cream bar at the base. Tertiary tentacles with two cream bars. Oral disc purplish grey with a soft pattern. A pale zone round the mouth imperfectly outlined by a scalloped ring of opaque cream. Primary and secondary radii dark, the latter with a pale orange triangle, tertiary radii pale orange with a little V at the tentacle base. Actinopharynx cream." (STEPHENSON 1935).

Size: According to STEPHENSON up to 2-3 cm in length and 1.5-2 cm in breadth at the oral disc, when expanded. The individual marked with (1) was in contracted state 1.2 cm long and 1.5-0.7 cm broad at the base, 0.3 cm at the distal part. The size of the example marked with (2) was: length 1.6 cm, breadth at the base 0.7 cm, at the margin 0.3 cm.

Occurrence: Norway: Sklinna, low-water (DONS 1927), 4 specimen (1). Iceland: Vestmannaeyar low-water (SEYMUNDSON 1899), 4 specimen (2).

Further distribution: North-east coast of Scotland, Wick (Gosse, Stephenson).

Exterior aspect. The single individual from Norway was contracted, conical with broad base, narrow in its upper part. The scapus was provided with a firmly adherent cuticle, very thick in the strong, wart-like tenaculi. The scapulus was smooth,



Fig. 36. *Phellia gausapata*. Section of a perfect mesentery in the region of the actinopharynx.

the margin tentaculate. The tentacles were 32 in number and considerably fewer than the mesenteries at the base, the inner tentacles considerably larger than the outer ones. The actinopharynx



Fig. 37. *Phellia gausapata*. Section of a mesentery of the second order in the region of the ciliated tracts.

had few but high longitudinal ridges. There were 2 siphonoglyphs symmetrically situated. STEPHENSON has noted 1-3 siphonoglyphs.

Anatomical description. The ectoderm of the scapus was fairly low, in the tenaculi thinner, on the other hand the cuticle of the tenaculi was very thick (fig. 2 Pl. V), in other parts fairly thin. In distinction from the cuticle of the tenaculi in *Kadosactis* it shows a very compact structure, at any rate it is not so lamellar as in this genus. The structure of the ectoderm in the tenaculi seems



Fig. 38. *Chondrophellia coronata*. Lower half of the sphincter.

Habitat.	scapulus n.	tentacles		actinopharynx n.	acoutia n.
		n.	sp.		
St. 78	24-27	19 · 1.5-43 · 7 μ	22-29	31-41 · about 2.8 μ
St. 90	18-23 · 2.5 μ	26-32	19 · 1.5-50 · 7.5 μ
St. 11	26-31	22 · 2-50 · 7 μ	26-34	38-53 · 2.8-4 μ
St. 93	(20) 22-26	19 · 1.5-46 · 7.5 μ	33-41 · 2.8 μ
St. 32	30-42 · 2.8-3 μ
St. 166 (small spec.)	30-36 · 2.8 μ
St. 2791 (Albatross exp.)	26-34	34-42 · 2.8-4 μ

W. 2148 m. Bottom temp. 1.6 (Ingolf exp. St. 11) 2 specimens; 62°57' N. 19°58' W. 957 m. (Thor exp. 1903. St. 166) 1 specimen; 61°33' N. 19° W. 2051 m. Bottom temp. 3' (Ingolf exp. St. 65) 1 specimen; 60°37' N. 27°52' W. 1505 m. Bottom temp. 4.5' (Ingolf exp. St. 78) 1 specimen; 19°25' N. 12°20' W. 1275-1180 m. (Thor exp. 1905 St. 93) 3 specimens.

Figure 95 shows the distribution of the species.

Further distribution: East coast of U. States 300-980 fms, Marthas Vineyard 41°29'15" N. 65°47'10" W. 980 fms (Blake exp. St. 307 teste VERRILL). East Atlantic 38°26' N. 26°30'45" W. 1165 m. 31°43'30" N. 10°46'45" W. 2165 m (Monaco exp.). Coast of Chile 38°08' S. 75°53' W. 677 fms. (Albatross exp. St. 2791).

Exterior aspect: The pedal disc is thin, commonly not or slightly broader than the cylindrical column and provided with a thin cuticle. The column is firm and its scapus provided with an adherent cuticle, which is thick and hard especially in the tuberculated area. The most part of the scapus shows longitudinal and transversal furrows giving the scapus a checkered appearance. Distally the squares have a more tuberculated facies. In the summit of the scapus there are 12 tuberculated ridges drawn out longitudinally, each divisible by shallow incisures into 2 or mostly 3 tubercles giving the ridges a characteristic, denticulate aspect (Pl. I fig. 10). The scapulus is short and longitudinally sulcated. The tentacles in the contracted state are rather thick and conical, the inner tentacles of ordinary length, the outer ones about half as long as the inner ones. They are hexamerously arranged in 5 cycles, but the last cycle was incomplete in the specimens

examined (from St. 78). I counted here 39 tentacles in the one half, possibly there were a few more as some tentacles were very small. About half of the oral disc is devoid of tentacles. The oral disc is radially furrowed at the insertions of the mesenteries, the actinopharynx is of ordinary length and provided with 2 distinct siphonoglyphs symmetrically situated and devoid of aboral prolongations.

Anatomical description: The ectoderm of the pedal disc and column is rather thin, that of the scapulus contains numerous nematocysts. The mesogloea of the column is thick and provided with rather numerous cells poor in protoplasm. The circular muscles of the column are weak, on the other hand the mesogloea sphincter (fig. 38) is very well developed and alveolar, in the middle region with a little tendency to be reticular. The muscle meshes are large, especially in the upper part of the sphincter.

The ectoderm of the tentacles is higher than the mesogloea and contains numerous spirocysts but more sparse nematocysts, especially in the specimens from St. 78 and 90. In the specimens from St. 11 they were more numerous in the apex of the tentacles. The longitudinal muscles of the tentacles are ectodermal with high folds arranged like a palisade and often somewhat ramified. The radial muscles of the oral disc are also ectodermal, sometimes, but not always, they are weaker at the insertions of the mesenteries. The ectoderm of the actinopharynx is somewhat thicker than that of the scapus and provided with numerous nematocysts. The mesogloea of the actinopharynx is of ordinary thickness. The sizes of the nematocysts (n) and spirocysts in the different regions of the body were the following:

The pairs of mesenteries were 18 in the sectioned specimens from St. 78, of these only 21 were present in the most part of the body, i. e. the fourth cycle was developed in the uppermost and lowermost parts of the column. At the hubus there were 24 pairs present, at the margin fewer. Only 6 pairs of mesenteries are perfect. The mesenteries of the second order are also well developed, those of the third order weaker and those of the fourth very weak. In the region of the actinopharynx the perfect mesenteries are thin in the inner half, in the outer thickened; the mesenteries of the second and third orders weaker here than below the actinopharynx. The mesenteries of the 3 first orders form diffuse retractors the folds of which are high, especially in the middle part of the retractors, and often rather much branched. Textfigure 39 shows a transverse section of 4 mesenteries of the 3 first cycles below the actinopharynx. The inner parts of the non-directives are here curved towards the exocoels, those of the directives towards the endocoels. The retractors do not reach the basal disc but are continued in a weaker muscle lamella in the proximal parts of the mesenteries (textfig. 40). The parietobasilar muscles are rather weak and form few and low folds, in the mesenteries of the third order they are in contact with the retractors. The basilar muscles are weak, with few but distinct folds. The mesenteries of the first and second order incl. the directives are provided with filaments and reproductive organs, either ovaries or testes. The acoutia are very well developed.

Remarks: The anatomy of this species is described by McMurrian (1893 p. 188), in several points, however, the description is incomplete. In contradistinction to McMurrian I have found

retractors are fertile in 2 specimens from the M. M. specimens. Thus, it is



FIG. 1. Section of the scapus with a directive mesentery and of the second and third order.

1925) does not belong to the genus *Actinurup* but to a new genus, for which I have already proposed the name *Chondrophellia*. Concerning the species M. M. has substituted the name *coronata* for *tastigata*



FIG. 2. A directive mesentery in *Actinurup*.

in order to obviate confusion with *Hormathia (Chitonactis) coronata*. As the species must be placed in another genus than *H. coronata* it is not necessary to give the species a new name. The name *tastigata* must be dropped.

As far as I can see the Albatross specimens, dredged off the coast of Chile, belong to the same species as the Atlantic specimens: of the former I have examined one specimen, considerably larger than the Atlantic specimens. Also *Hormathia elongata* described by GRAVIER (1918, 1922) is certainly this species or at least a species belonging to the genus *Chondrophellia*. The figures of this species agree very well with *Chondrophellia* as also GRAVIER'S description of its exterior and of its imperfectly described anatomy. GRAVIER'S statement that the parietobasilar muscles are well developed, while the retractors are weak, is certainly due to a confusion of these muscles. It is opposite to remark that the retractors of *Chondrophellia* are situated close to the column.

Genus Allantactis Dan.

Hormathidae with smooth, fairly thick column which is devoid of tubercles, cuticle and cinctides. Margin distinct. Sphincter very strong, mesogloea. Tentacles fairly short, the inner longer than the outer ones, without basal bulbs, 96 in number, hexamerously arranged. Longitudinal muscles of the tentacles ectodermal, radial muscles of oral disc meso-ectodermal. 2 well developed, broad siphonoglyphs, 6 pairs of perfect and sterile mesenteries, 2 pairs of directives. Not more mesenteries than tentacles. Retractors of the mesenteries diffuse. Parietobasilar and basilar muscles fairly weak. Cnidom: spirocysts, basitrichs, microbasic *p*-mastigophors.

Allantactis parasitica Dan.

PL. III fig. 14, PL. IV fig. 9.

Allantactis parasitica n. gen. n. sp. DANIELSSEN 1890 p. 20 Pl. 2 fig. 3, Pl. 9 figs. 1-4.

Allantactis parasitica DAN. KWIETNIEWSKI 1898 Pl. 14 figs. 1-3; CARLGRÉN 1902 p. 50, 1912 p. 5; PAX 1915, 1922 p. 250; CARLGRÉN 1928 p. 263, 1932 p. 263, 1939 p. 11; CARLGRÉN in KRAMP 1933 p. 17; CARLGRÉN in BROCH 1936 p. 1.

Calluactis krogeri n. sp. DANIELSSEN 1890 p. 36 Pl. 2 fig. 2, Pl. 8 figs. 6, 13, 14.

Pedal disc fairly wide embracing shells of *Sipho*, *Neptunus* or other gastropods. Column smooth. Sphincter very strong separated from the endodermal muscles of the column by a rather thin stripe of the mesogloea, mesogloea, distinctly transversally stratified. Tentacles, at least in preserved state, more or less longitudinally furrowed, their longitudinal muscles well developed. Radial muscles of oral disc principally ectodermal, strong in the outer part, weaker in the inner one. Actinopharynx with longitudinal ridges. Siphonoglyphs with well developed gonidial tubercles and aboral prolongations. Retractors of the perfect mesenteries band-like, in the region of the actinopharynx forming fairly low folds, in the directives stronger, below the actinopharynx the folds of the retractors are strongest inwards. Parietobasilar muscles forming no shelf. Mesenteries of the second cycle may sometimes be sterile. Nematocysts of the column 23.5-39 (31) · 3.5-4.2 μ , basitrichs, those of the tentacles (25.5) 28-39.5 · 3.5-4.2, basitrichs, those of the actinopharynx (26) 28-42 · 3-4.2 (4.9), basitrichs, those of the filaments partly (24) 24-33.8 · 4.2 (4.9) μ , microbasic *p*-mastigophors, partly 12.7-17 · 1.5-2 μ basitrichs, those of the acontia partly 31-43.7 · 3-3.5 μ , partly 15.5-22.6 · almost 2 μ , both basitrichs. Spirocysts up to between 50-60 · 7-7.5 μ .

Colour: Column yellow, shading a little to brown, distal part near the tentacles rose-red. Tentacles pale violet. Oral disc rose red, around the mouth a purple-red oblong annulus (*Allantactis parasitica*, DANIELSSEN); Column carmine-red with light-red

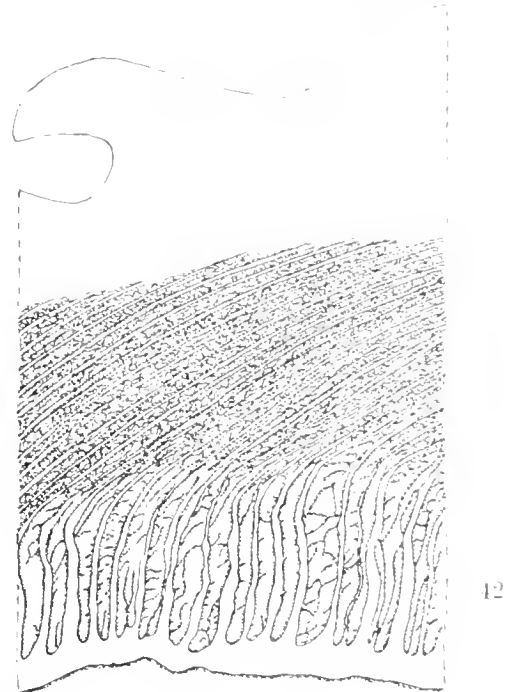
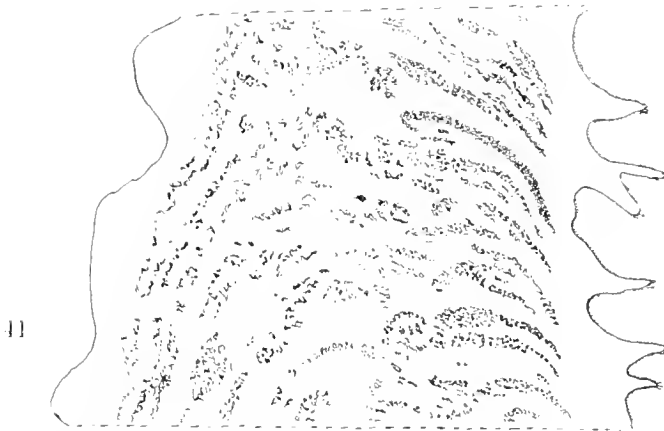
longitudinal stripes, tentacles and oral disc pale buff colour. The colour varies, however, somewhat in the different individuals, from scarlet to carmine (*Calliactis kroyeri*, DANIELSSEN); reddish with deeper coloured stripes, distal part of the body reddish (specimen from Hurry Inlet); colour white to brown-yellow upwards reddish (specimen from Franz Joseph fjord); yellow-red with yellow-grey tentacles (specimen from Fame Islands).

Sizes in preserved state up to 10 cm long and 4 cm broad.

Occurrence: Jones Sound, Gaasefjord 76°18' N, 88°40' W, (Fram Exp. 1900) 75°51' N, 81°01' W, 610 m. Bottom temp. $\pm 0.6^\circ$.

West Greenland: Baffin Bay and Davis Strait: 77°28.5' N, 68°16' W, 875 m. Bottom temp. $\pm 0.4^\circ$, 77°17' N, 69°59' W, 930 m.

Joseph fjord 12-35 m, entrance of Franz Joseph fjord 200-300 m. Inner part of Moskusoxefjord 100 m, outer part 220 m (Kolthoff Exp. 1900). Between Bontekoe and Cape Bennett 290 m, Franz Joseph fjord 502 m. Temp. 1.3, Dusen fjord 185-75 m. Bottom temp. ± 1.2 (Norwegian Exp. 1930, 1931), Geologfjord near head 650-625 m. Bottom temp. ± 1.61 , Grantaffjord 82 m. Bottom temp. ± 1.56 (Norwegian Exp. 1932), 73°20' N, 21°30' W, 70 m, 73°15' N, 25°12' W, 760 m, 72°15' N, 22°56' W, 35-60 m, 72°13' N, 26°38' W, 35-60 m (Sw. Greenland Exp. 1899), Sofia Sound 200 m, entrance of Kempe fjord 100-55 m, Vega sound 250 m, 30 m, 250-190 m, Alpe fjord 70 m, Antarctic harbour (Norwegian Exp. 1930), Forsblad fjord 11-3 fms. (Danish E. Greenland Exp. 1900), Scoresbysund, Fame Isl. 70°50' N, 22°33' W, 23-25 m, Hurry Inlet 70°13' N, 22°29' W, 70 m. (Sw. Greenland Exp. 1899), 7°0



Figs. 41, 42. *Allantactis parasitica*. Section of 2 sphincters in their middle part. The outer part of the sphincter figured in 42 is somewhat schematic.

Bottom temp. $\pm 0.4^\circ$, 77°05.5' N, 71°13' W, 790 m. Bottom temp. $\pm 0.4^\circ$, 76°36' N, 68°51' W, 180-80 m. Bottom temp. ± 1.3 , 75°35' N, 65°41' W, 490 m. Bottom temp. 0.7° , 75°26' N, 62°26' W, 820 m. Bottom temp. 0.7° (Godthaab Exp.), 75°26' N, 67°27' W, 260 fms. (Sofia Exp.), 74°52.5' N, 62°12' W, 150 m. Bottom temp. 0.7° , 73°12' N, 58°08' W, 850 m. Bottom temp. 0.5° (Godthaab Exp.), Upernavik (Drosvad 1929), 72°08' W, 74°20' W, 30-80 fms. (Nilsson), 70°53' N, 51°03' W, about 700 m. Bottom temp. 1.05° , 70°51' N, 52°01' W, 733 m. Bottom temp. 1.1° (Godthaab Exp.), Umanak 260 fms. (Tjalfe Exp. 1908), 250 fms. (Hansen), Ikerasak, inner part of Umanakfjord 70°36' N, 51° W, 80-120 fms. (Oirik), 69°16' N, 51°35' W, (Tjalfe Exp. 1908 St. 146), 69°41' N, 51°38' W, 350 fms. (Sofia Exp.), North part of Disco fjord 171-227 fms., 69°17' N, 52°50' W, 225 fms., 69°17' N, 52°11' W, 227-231 fms., 69°15' N, 53°18' W, 114-161 fms. (Tjalfe Exp. 1908 St. 115, 117, 18, 122, 182), 68°17' N, 58°14' W, 410 m. Bottom temp. 2.55° , Totness Road, Exeter Sound 75-200 m (Godthaab Exp.), Tunugdliarfik 280-300 m. Bottom temp. 2.1° (Rink Exp. 1912 St. 139). Greenland without distinct locality (Oirik).

East Greenland: Tirolerfjord near the head 122-126 m. Bottom temp. $\pm 1.10^\circ$, near Young Sound 320 m. Bottom temp. ± 1.73 (Norwegian Exp. 1932), Clavering fjord 338-100 m. Between Jackson Isl. and Cape Mary 250-230 m. Bottom temp. $\pm 1.05^\circ$, 1/4 quarter-miles S. of Hold with Hope 310-260 m. Bottom temp. 0.3 (Norwegian Exp. 1931) Mackenzie Bay N. of Franz

fus. (Danish E. Greenland Exp. 1900), Kangerdlugssuak 175 m (2nd East Greenland Exp. 1932).

N. E. and E. of Iceland: 67°10' N, 15°10' W, 195 fms. Bottom temp. ± 0.6 (Ingolf St. 121) 66°14' N, 11°33' W, 781 fms. Bottom temp. ± 0.8 (Ingolf Exp. St. 110), 66°32' N, 18°50' W, 192 m (Heegaard), 61°53' N, 10°0' W, 630 m. Temp. 600 m ± 0.69 (Michael Sars Exp. 1900), 65°21' N, 10°42' W, 650 m (Monaco Exp.) 63°05' N, 3°00' E, 960 m. Bottom temp. ± 1.1 (Norw. N. Atl. Exp.), 63°06' N, 2°46' E, 915 m. Temp. at 910 m ± 1.07 (Michael Sars Exp. 1900), 63°07' N, 1°38' E, 1150 m, 62°58' N, 1°56' E, 1100 m, 62°13' N, 1°26' E, 775 m (Michael Sars Exp. 1902, all stations from the cold area).

West Spitzbergen: 80°01' N, 10°51' E, 130 m (Monaco Exp.) Kings Bay 60-160 fms. (Sw. Spitzb. Exp. 1861) Prince Charles Forland 78°57' N, 11°80' E, 115 m (Olga Exp.), 78°18' N, 8°47' E, 199 m. Bottom temp. 1.1 (Norw. N. Atl. Exp.) 78°22' N, 17°10' E, 102 m (Monaco Exp.), Safe Bay 118-127 m. Temp. at 108 m 0.95 , entrance Tundra Bay 71-68 m. Bottom temp. ± 0.93 , off Tundra Bay 117-111 m. Temp. at 110 m ± 0.62 , off Cape Erdmann 263-256 m. Bottom temp. $2.2.6$, off Alkhorn 106-395 m. Temp. at 382 m 2.61 , Advent Bay 70-12 m. Temp. at 41 m 1.85 , entrance East fjord 199-226 m. Temp. at 210 m 1.27 , Temple Bay, Bona Harbour 30 m. Bottom temp. 3.78 , Green Bay 90-80 m, Billen Bay 37-35 m. Bottom temp. 1.5 , 150-110 m ± 1.67 , Northfjord

richardi. The column is higher than broad and divisible into scapus and scapulus. The former is in its lowermost part almost smooth, in its upper part provided with more or less numerous, but always small tubercles arranged in vertical rows. In the contracted state the column is transversally furrowed, and shows in the tuberculated area a checkered appearance. There are no distinct coronial tubercles. As the ectoderm of the scapus is lost, I cannot confirm, whether the scapus is provided with a cuticle. At any rate the

and ridges, in the contracted state also transversally furrowed. The 2 siphonoglyphs, symmetrically situated, are broad and aborally prolonged.

Anatomical description: The mesogloea of the column is thick and contains cells poor in protoplasm, the sphincter very strong and separated from the weak endodermal muscles by a considerable mesogloea-layer. The muscle meshes are arranged in groups, elongated transversally and separated from each other by thicker balks of mesogloea. Therefore the sphincter is distinctly transversally stratified. In its lowest part the muscle meshes are more scattered and separated by thicker mesogloea-balks and curved (fig. 13).

The ectoderm of the tentacles is thick apart from that of the basal swellings. The longitudinal muscles of the tentacles are ectodermal, in their proximal parts weaker on the outside than on the inside, in the basal swellings probably absent (fig. 14). In the outer parts of the swellings I have found some muscle meshes in the mesogloea in transverse sections, but probably these muscle meshes belong to the sphincter. The mesogloea of the tentacles is thick in their basal parts especially in the swellings. The radial muscles of the oral disc are meso-ectodermal and well developed, at the insertions of the mesenteries not weaker than between the mesenteries.

The mesenteries are typically arranged in 4 cycles (6 + 6 + 12 + 24 = 48 pairs). Only the mesenteries of the first cycle are perfect. The longitudinal muscles form weak diffuse retractors in the stronger mesenteries (fig. 15). In the region of the actinopharynx the retractors are situated in the middle of the mesenteries, or somewhat more outwards, and show large folds seen with the naked eye. The parietobasilar muscles are weak, as are also, probably, the not well preserved basilar muscles. The mesenteries of the first cycle are sterile, the other fertile. In maceration preparations of the filaments with adjacent parts of the mesenteries there were fairly numerous nematocysts of different size and appearance, one part certainly holotrichs, the other atrichs or holotrichs all evidently residues of the food taken (see CARLQVIST 1910 note 2 p. 509).

Remarks: HADDOX (1889 p. 315) has referred the species to the genus *Chitonactis*, and HADDOX (1898 p. 459) and STEPHENSON (1929 p. 535) to the genus *Hormathia*. As the genus *Actinauge* may be maintained, the species *longicauda* undoubtedly belongs to this genus. It agrees as well in appearance as in organisation much with *Actinauge verrilli* and *richardi*.

Actinauge richardi (Marion)

Pl. III fig. 7, Pl. IV fig. 1.

For literature and synonyms see STEPHENSON 1935, moreover

Chitonactis richardi (Mar.) FISCHER 1889 p. 269 Pl. 6 figs. 3, 4.
Actinauge richardi (Mar.) CARLQVIST 1934 p. 16; PAN 1936 p. 111.

Diagnosis: Pedal disc thin, usually deeply excavated, forming a cup-shaped concavity filled with mud or sand which serves as an anchor for the animal, sometimes, however, attached to other objects and then flattened. Column divisible into scapus and scapulus, the former covered with a cuticle which sometimes is thin, sometimes strong and provided with tubercles showing a tendency to be arranged in longitudinal and vertical rows. The tubercles are of very variable appearance, now low, now large, rounded or more acuminate. At the uppermost part of the scapus usually 12 stronger tubercles. Scapulus with ridges which sometimes are broken up by transverse interruptions into tubercles, and which communicate with the swellings of the tentacles of at least the fourth cycle. Sphincter separated from the very weak endodermal muscles of the column by a fairly thick stripe of the mesogloea, fairly long, not much broader in its upper part than in its lower, usually alveolar with a tendency to longitudinal and transverse stratification or forming a net of coarse meshes in which smaller



Figs. 44, 45. *Actinauge longicauda*. Transverse sections of a tentacle (fig. 44) and a perfect mesentery (fig. 45) in the region of the actinopharynx. Inner part of the mesentery not figured.

cuticle, if present, is weak. In the contracted state of the animal the scapulus is furnished with numerous longitudinal furrows and alternately broader and smaller ridges, the number of the ridges seems to correspond to the number of tentacles, as each ridge ends at the base of a tentacle. Thus the margin is not distinct. The tentacles are acute, and in the expanded state long (teste VERRILL), in the contracted state of ordinary length, the inner are longer and broader than the outer ones. The capitular ridges, especially the larger ones, end with thickenings at the base of the tentacles, the latter are bulbous swollen at the outside of the base. These swellings are most distinct in the inner tentacles. The number of tentacles seems to correspond to that of the mesenteries, and is 96. In VERRILL's figure I Pl. 5 (1883) the last cycle is not indicated. They are probably hexamerously arranged. The inner half of the oral disc is radially furrowed and lacks tentacles. According to VERRILL the oral disc "can expand very broadly". The actinopharynx is long and provided with several longitudinal furrows

Scapus 10-15 cm high, up to 10 cm broad, up to 12 cm high. Oral disc 1.5-2 cm in diameter, with 5-6 cycles, each cycle with 5-6 ridges, the ridges of the 5th cycle, the most numerous, with swellings on their outside. Retractors of the oral disc fairly weak on the meso-ectoderm. Radial muscles of oral disc meso-ectodermal. Actinopharynx 1.5-2 cm wide, with about 24 longitudinal filaments, up to 10 cm partly prolonged aborally. Mesenteries 96 or rarely a few more, hexamerously arranged, 6 pairs of perfect and sterile mesenteries. Retractors of the mesenteries 10-15 cm long, with a little concentrated, forming a sheath, 1.5-2 cm wide, not forming a sheath, a little concentrated, 1.5-2 cm long. Nematocysts of the scapus 10-15 cm, 2.5-2.8 μ , partly about 11.3-15.2 μ , basitrichs, those of the actinopharynx partly 22-36-28 μ , basitrichs, partly about 19.7-3 μ , very rare, microbasal p -mastigophors, those of the filaments partly 10-11.3-15.2 μ , basitrichs, partly 16.2-20-3-4.2 μ , microbasal p -mastigophors, those of the tentacles partly 24-34-3-3.5 μ , partly 8.5-10-12 μ , very rare, basitrichs. Spirocysts of tentacles up to 60 μ , 7 μ .

Colour: Scapus partly white tinged with green, tubercles dull red, scapus and base of tentacles whitish. Tentacles white or orange-brown with brown longitudinal bands, or blood-red to chocolate-brown with longitudinal bands of red and white. Oral disc usually white, rarely streaked with brown, muddy brown, or chocolate-brown colour (HADDON, DÉRÈGNÉ).

Size: up to 8 cm broad and 12 cm high.

Occurrence: Scotland 60°36' N, 0°37' W, 115 m (Jäger-ke, 1910); Hebrides 58°31' N, 8°03' E, 300 m (Jägerskiöld); 61°40' N, 3°11' E, 490 m. Temperature at the bottom 6.31. Melville Sars exp. St. 51 (1902); 61°44' N, 2°43' E, 115 m. Temperature at the bottom 6.78. Michael Sars exp., St. 50 (1902); Farøe group (teste STEPHENSON), great fishing banks N.W. of S.W. of Bergen 130-180 fms., N.W. of Egersund, Jæderen, Gjøra, rock (Swedish fishermen), North Atlantic; 59°22' N, 11°44' W, 184 m, 49°27' N, 8°36' W, 116 m. Temperature at 157 m, 6.56, 6.58. Michael Sars exp., 1910; 36°07' N, 8°03' W, 110 m. Michael exp., St. 1096 determined by GRAVIER as *Chondactinia* (Hadd.) 35°32' N, 7°07' W, 1215 m, temperature at 157 m, 16.10, 35°31' N, 6°37' W, 735 m, temperature at 157 m, 17.2. Michael Sars exp., 1910; 26°55' N, 15°05' W, 189 m, temperature at the bottom 11.2 (German Deep Sea exp.), locality? (teste VERRILL, 1922) determined by MARIOT as *Chondactinia* (Hadd.) 15. Mediterranean, Naples, Palanghesari, Bochicella.

Further distribution: Several localities S.W. of Ireland (Hadd.), St. Vincent, Bay of Biscay, coast of Portugal, west coast of Africa as far as Senegal (teste FISCHER) according to MARIOT (1922), from 70 to 2000 m.

The genus *Chondactinia* species has been described by HADDON (1888), Sars (1902, 1918), and VERRILL (1928). Finally STEPHENSON (1928) has given a good summary of the exterior and the interior of the species. The illustrations made by him in other respects are excellent. The number of nematocysts is summed up from 10 specimens, but the number of the filaments only from 3.

Actinange verrilli (Hadd.) McMurr.

Actinange (Hadd.) McMurr. CAPRIORI, 1928, p. 292 and 1933, p. 103.

Colour: Scapus and base of tentacles of *T. richardi*. Column divisible into 5-6 cycles, each cycle with a tubercle rarely deciduous. Tubercles of the scapus with tubercles arranged in longitudinal rows. Tubercles very variable in their shape, some are small, round, low large and more flattened

and formed by growing together of smaller tubercles. Lower part of the scapus not tuberculated or with indistinct tubercles. Scapulus agreeing with that of *richardi*, the ridges however more numerous. Sphincter elongated, occupying only a part of the thick mesogloea, only in its upper part somewhat below its apex stronger, alveolar with distinct tendency to transverse stratification. Tentacles 96, rarely a few more, short, especially in comparison with the length of the animal, conical, at least in the preserved state longitudinally furrowed, at their outside provided with sometimes very strong, sometimes weaker swellings of the mesogloea. The longitudinal muscles of tentacles ectodermal, not strong, very weak in the bulbs. Radial muscles of the oral disc ectodermal or meso-ectodermal, well developed. Actinopharynx long, longitudinally furrowed. Two distinct but not very broad siphonoglyphs aborally prolonged. Mesenteries 96 or rarely a few more, hexamerously arranged, 6 pairs of perfect and sterile mesenteries. The mesenteries of the second order also seem to be sterile. Retractors of mesenteries diffuse with low folds. Nematocysts of the scapus 10-15 cm, 13.4-2 μ , basitrichs, those of the scapulus partly 11.3-18.3-2-2.8 μ , basitrichs, partly 21-28.2-1.2 μ , microbasal p -mastigophors, those of the tentacles 22.6-36-2.8-3 (3.5) μ , basitrichs, those of the actinopharynx partly 26-12.2-2.8-3.5 μ , partly 14-19- about 2 μ both basitrichs, partly 21-36-1-4.5 μ , microbasal p -mastigophors, those of the filaments partly 12.7-18.3-2 μ , basitrichs, partly 21-31-1.2 (1.5) μ , microbasal p -mastigophors, those of the acontia partly 31-13-3-1 μ , partly 11-21.5-2 μ , both basitrichs. Spirocysts of tentacles up to 60 μ , 7 μ .

Colour: Scapus dull pale red, flesh colour or salmon, warts pink or whitish. Scapulus red, orange or chocolate-brown often present in stripes of darker or lighter tints. Tentacles usually dark pink, salmon, orange or orange-brown varying to dull red and chocolate-brown. Oral disc usually orange or reddish brown or chocolate, with lighter and darker radii (teste VERRILL 1922 p. 6, 95).

Size: up to 10-15 high and 8-10 cm broad, larger tentacles up to 4.5-2 cm long. (VERRILL 1922).

Occurrence: Baffin Bay, Davis Strait and West Greenland; 77°28' N, 68°46' W, 875 m, bottom temp. \pm 0.1; 77°05' N, 71°73' W, 790 m, bottom temp. \pm 0.1; Melville Bay 75°26' N, 62°26' W, 820 m, bottom temp. 0.7; 71°52' N, 62°12' W, 450 m, bottom temp. 0.7; off Lancaster Sound 71°12' N, 77°00' W, 680 m, bottom temp. 0.1; 73°12' N, 58°08' W, 850 m, bottom temp. 0.5 (Godthaab exp. 1928); Upernivik (Drosvad); 70°53' N, 51°03' W., about 100 m, bottom temp. 4.95; 70°51' N, 52°01' W, 733 m (Godthaab exp. 1928); Umanakfjord 260 fms. (Tjalfe exp. 1908), Umanak 397 fms. (Ingegerd and Gladan exp. 1871); 250 fms. (Amundsen); Discøfjord 130 fms. (Holm); Discøfjord, Nipisite harbour 10-20 fms. (Ingegerd and Gladan exp. 1871); 68°24' N, 51°05' W, 220-280 fms. (Tjalfe exp. 1908); 66°49' N, 56°28' W, 235 fms. (Wandel 1889); 66°44' N, 56°08' W, 175 fms. (Tjalfe exp. 1908); 66°42' N, 56°12' W. (Tjalfe exp. 1909); 66°35' N, 56°38' W, 318 fms, bottom temp. 3.9 (Ingolf exp. St. 32) 65°36' N, 56°24' W, 349 fms. (Wandel); 65°34' N, 51°31' W, 68 fms., bottom temp. 0.2 (Ingolf exp. St. 29); 65°30' N, 55°26' W, 289 fms. (Wandel 1889); 65°14' N, 55°42' W, 120 fms., bottom temp. 3.5 (Ingolf exp. St. 28); 63°30' N, 54°25' W, 582 fms., bottom temp. 3.3 (Ingolf exp. St. 25); Kyaneffjord 120 m, 300-500 m, 200-410 m (Rink exp. St. 5, 11, 23).

N. of New Foundland 51°20' N, 52°25' W, 232 fms. (Ingegerd and Gladan exp. 1871); St. Lawrence estuary 50-330 m (Préfontaine); 80 miles S. of Marthas Vineyard (U. S. Fish. Com.); 40°03' N, 70°28' W, 135-160 fms. (U. S. Fish Commission).

Iceland, Halli of Isafjardardjup 170-235 m; Reyðarfjörður 226 m.

Further distribution: East coast of N. America from Chesapeake Bay to west of New Foundland 50-1098 fms. (teste

Verrill 1922). West coasts of N. and S. America? (teste McMURRIE; probably, as he remarks, another species).

Exterior aspect: The exterior of this species varies considerably in the distribution and size of the warts of the column. In the specimen from U. S. Fish Commission (Pl. II fig. 9) the warts are small and rounded and the arrangement in rows very distinct, the specimens from the Godthaab expedition (Pl. II figs. 6, 8, 10) agree with the former, but the warts are fewer and sometimes very indistinct. In an individual from the Tjalle ex-

into scapus and scapulus, the former provided with a more or less developed cuticle. Tentacles always more than 96, not so numerous as the mesenteries, usually with distinct swellings at the base of their abaxial side, arranged at the margin in 2 (or more?) close cycles. Longitudinal muscles of the tentacles ectodermal. Oral disc wide, sometimes asymmetric, bilobed. Radial muscles of oral larger disc ectodermal or meso-ectodermal rather strongly folded, in the parts corresponding to the endocoels, however, unfolded. Two well developed siphonoglyphs. Pairs of perfect mesenteries commonly 6, sometimes 7-8 or 6-7 + some extra single mesen-

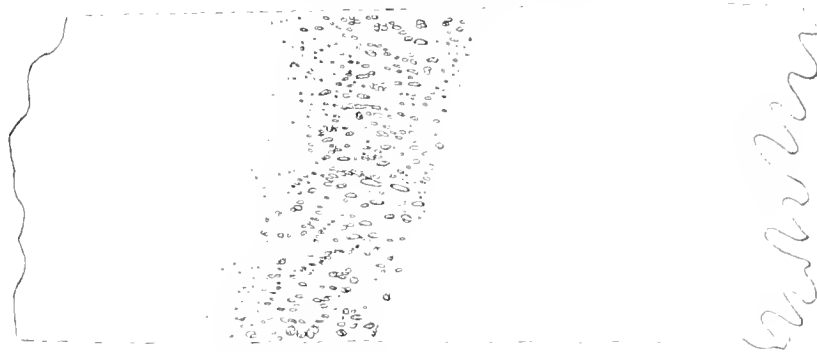


Fig. 46. *Actinauge richardi*. Section of the sphincter in its middle part.

pedition (Pl. II fig. 7) the upper warts have begun to grow together in the upper part of the scapus at the same time as they have become larger. Much larger warts are met with in a specimen from the Ingolf expedition (Pl. II fig. 4). Three other individuals, all from same locality, show a different appearance. The specimen figured in Pl. II fig. 2 has the warts of about ordinary size but the arrangement is more irregular. Figs. 1 and 3 Pl. II show individuals provided in their upper parts with very large warts (the lower part of the scapus is not visible) evidently arisen from growing together of smaller warts. When I first examined these specimens I suggested that they possibly belonged to *Phelliactis*, but a closer examination of the oral disc of the specimens figured in Pl. II figs. 2, 3 and 4 showed that the characteristic arrangement of the muscles in this genus were absent here, wherefore these specimens must be referred to *Actinauge verrilli*, all the more as all other characters, also the size of the nematocysts, agreed with this species. The ridges of the scapulus and the coronal tubercles, often not distinct, seem to vary in number.

Anatomical description: McMURRIE has 1893 given a good description of the anatomy of this species. The sphincter is, in comparison to the size of the specimens, fairly weak, strongest in the upper part apart from the uppermost. It shows a tendency to transverse stratification though not so distinctly as in *A. longicornis*. A portion of the sphincter has been drawn in figure 46. The basitrichs of the acontia have been measured in 25 specimens, their average length was 36.2μ resp. 17.6μ , those of the tentacles in 18 individuals, their average length was 28.2μ . The nematocysts of the actinopharynx were examined in 15 specimens, those of the filaments in 5. The average length of the microbasal *p*-mastigophors in the former was 28.48μ , in the latter 28.1μ . The average length of the basitrichs in the actinopharynx was 35.97μ resp. 16μ , in the filaments 15.58μ . The nematocysts of the scapus and scapulus have been examined in only 1 specimen, because the ectoderm usually was disturbed or incrustated with mud and other objects.

Genus *Phelliactis* Simon.

Hornathiidae with well developed pedal disc. Column in its upper part sometimes more or less asymmetric, usually provided with large tubercles, especially in the distal region. Column divisible

teries, 2 pairs of directives. Mesenteries more numerous at the limbus than at the margin, last cycle of mesenteries as a rule developing earlier at the mesenteries of the first and second order than at the third. Retractors of the mesenteries in the lower parts fairly weak, stronger in their uppermost part. Parietobasilar muscles weak, not forming a shelf. At least 6 perfect pairs sterile. Acontia well developed only with basitrichs. Unidom: spirocysts, basitrichs, microbasal *p*-mastigophors.

As I have shown (1928a p. 207) the genus *Chondrodactis* is synonymic with *Phelliactis*.

Phelliactis hertwigi Simon.

Pl. IV fig. 11.

Phelliactis hertwigi n. sp. SIMON 1892 p. 75; CARLSEN 1928 p. 289; 1928a p. 207, 1931 p. 11 textfig. 9.

Chondrodactis duplicata n. sp. STEPHENSON 1918 p. 112 Pl. 14 fig. 6, Pl. 15 fig. 3, Pl. 18 figs. 19-21, Pl. 19 figs. 2-9; 1920 p. 491, 513, 539 fig. 25.

Diagnosis: Pedal disc well developed. Column with large, but not high, tubercles especially in the upper part of the scapus. No sharp difference between scapus and scapulus because the cuticle of the former is thin and easily deciduous (perhaps sometimes not present?). Uppermost part of the column distinctly asymmetric, bilobed. Sphincter well developed, broad distally, gradually diminishing downwards, alveolar with tendency to transverse and longitudinal stratification, separated from the endoderm by a rather broad mesogloea-lamella. Tentacles up to about 160, fewer than the mesenteries with distinct bulbs at the base of their abaxial sides. Longitudinal muscles of tentacles ectodermal, well developed at the adaxial side. Radial muscles of the bilobed oral disc, as also the siphonoglyphs, as in other *Phelliactis*-species. Mesenteries arranged in 5 cycles, the last cycle more or less incomplete. Always more than 6 perfect pairs; the extra perfect mesenteries forming now pairs, now consisting of one or more mesenteries, the partner of which is imperfect, generally symmetrically arranged on both sides of the one directive pair. Nematocysts of the tentacles $32.4-46$ + about 2.8 (3.5) μ , basitrichs, those of the actinopharynx partly $35.2-48$ + 2.8μ , basitrichs, partly $31-39.5$ + 1.2 (1.5) μ , microbasal *p*-mastigophors, those of the filaments partly $32.1-38$ + 2.8μ , basitrichs, partly $26.8-31$ + about 3.5μ , microbasal *p*-mastigophors, those of the

contraction of the specimens. The distal part of the body is more or less asymmetric, at least in older specimens, but not so distinctly as in *P. hertwigii*. The column is divisible into scapus and scapulus, the former of which is covered by a cuticle, usually lost or only present between the tubercles. In some specimens, however, the cuticle is better preserved and sometimes, as in the specimens from the stations 27 and 10 (Pl. I figs. 9, 12) and from the Sofia expedition (Pl. IV fig. 8), the tubercles are provided with a thick cuticle. The lower part of the scapus has smaller tubercles, the upper part commonly very large ones. Most specimens show an appearance like that reproduced in Pl. I fig. 12. The scapulus is provided with high ridges which are tuberculated when the bulbous swellings of the tentacles are large.

The short tentacles are situated at the margin. Their number varies, but the tentacles are always fewer than the mesenteries at the limbus. One specimen examined had 192 mesenteries at the base but only about 160 tentacles, another 156 mesenteries at the lowest part of the actinopharynx but about 130 tentacles. At the base of the abaxial side the mesogloea forms more or less large bulbs, continuations of the ridges of the scapulus; sometimes the bulbs are small and rather indistinct but always present at least on several tentacles. The oral disc is wide with radial ridges,



Fig. 47. *Phelliactis robusta*. Sphincter of a small specimen.

the actinopharynx long with few longitudinal ridges, the two siphonoglyphs are broad but aborally hardly prolonged.

Anatomical description: The ectoderm of the scapus is low, the mesogloea very thick, the endodermal circular muscles of the column are weak. The mesogloea sphincter is, in comparison with the size of the specimens, not strong, especially on the higher side; that is to say, there is usually a little difference in the development

of the sphincter in the lower and higher sides of the contracted animal. It is wholly separated from the endodermal circular muscles and rather broad in its upper part but gradually diminishing downwards. Its aboral end, forming only a thin musculamella, is situated nearer the endoderm than the ectoderm, but



Fig. 48. *Phelliactis robusta*. Uppermost part of the sphincter in a larger specimen.

separated from the former by a broad mesogloea. Its structure is alveolar (Fig. 47) showing a tendency to horizontal stratification in the middle part; sometimes the upper part of the sphincter is more reticular but with thick mesogloea-meshes (fig. 48).

The ectoderm of the tentacles is high, the longitudinal muscles are at the base considerably stronger at the adaxial side than at the abaxial. The meso-ectodermal radial muscles of the outer part of the oral disc agree with those in other *Phelliactis*-species viz. are weaker in the regions corresponding to the stronger endocoels than in other places, sometimes the difference is not so strongly marked as in *Phelliactis hertwigii*. In the endoderm there were small often exploded basitrichs.

The mesenteries are hexamerously arranged, at the limbus more numerous than at the margin. There are 5 cycles of mesenteries, usually those of the last cycle are not developed in all exocoels. I have dissected several specimens but in no case I have observed more than 6 perfect pairs, in contrast to the case of *hertwigii*, in which there always are more than 6 perfect pairs even in small individuals. As most specimens are considerably larger than the small specimen of *hertwigii* figured by CARLOREX (1931 b fig. 9), there is no reason to suppose that in *robusta* more mesenteries than 6 pairs would later grow together with the actinopharynx. In one specimen the mesenteries were 192 at the base, another specimen had 156 mesenteries at the end of the actinopharynx, a third about 131 mesenteries in the middle of the animal, at the limbus about 160. The number of mesenteries in the six sextants was more or less different. As in other *Phelliactis*-species the mesenteries of the last cycle show a tendency to develop earlier in the exocoels nearer the mesenteries of the first and second order than those of the third. The muscles of the mesenteries agree with those in other *Phelliactis*-species. The retractors are considerably stronger in the upper part of the body than in the lower part though weaker than in *Paraphelliactis*.

I have measured the nematocysts of the acontia in 10 specimens. They show good agreement in all. The small nematocysts of the acontia are rare and perhaps sometimes absent. The dimensions of the cuidae in other parts of the body are summed up from 3 specimens. The measures of the nematocysts of the filaments is, however, from a single specimen. In the smallest, not fertile specimen, the nematocysts were somewhat smaller. In a maceration preparation of the filaments - parts of the mesenteries I found several large holotrichs, evidently residues of the food taken.

Genus *Paraphelliactis* Carlgr.

Hormathiidae with well developed pedal disc. Column cylindrical, divisible into scapus and scapulus, the former strongly

lamella on the outside, while they were folded on the inside (fig. 51). I have made several maceration-preparations of the tentacular ectoderm from the more closely examined specimen but have not observed any nematocysts. In the other large specimen some small tentacles were visible; maceration preparation from these showed the presence of several nematocysts $29.6-36.7 \times 2.5 \mu$, but as acontia were ejected close to the tentacles it is possible that the capsules belonged to the acontia and only were stuck to the



Fig. 52. *Paraphelliactis spinosa*. Part of a mesentery in the region of the endo-glandular tract.

tentacles. On the other hand the spirocysts were extraordinarily numerous. The radial muscles of the oral disc were meso-ectodermal. The arrangement of these muscles agrees more with that in *Actinauge* than that in *Phelliactis*. True, the height of the muscle-lamella can be somewhat different in certain parts (fig. 50) but there was no such regular diminishing of the muscles in the middle of the parts corresponding to the larger endocoels.

The pairs of mesenteries were arranged in 5 cycles ($6 + 6 + 12 + 21 + 48$) in the specimen examined, but the last cycle was incomplete and also the mesenteries of this cycle present were very weak, especially in the lower part of the animal. As I counted 168 tentacles, the mesenteries at the margin may have been 168. I have observed 164, of which about 10 pairs were small. In the lower part of the body the mesenteries seemed to be fewer. Owing to the irregular contraction of the pedal disc I could not, however, count with any certainty, as the limbus was damaged, but I think that there were fewer mesenteries there. Somewhat higher up at most 150 mesenteries seemed to be present, but the mesenteries of the last cycle were also there very small. Only the mesenteries of the first cycle were perfect with 2 pairs of directives symmetrically situated. In the distal part the retractors of the mesenteries formed very strong but diffuse pennons on the stronger mesenteries (fig. 50), in the lower part they were weak (fig. 52) only the muscles of the mesenteries of the first order were here rather well developed. The parietobasilar muscles were weak, the ciliated tracts of the filaments strong. The mesenteries of the first cycle were sterile, the stronger of the other mesenteries fertile. The specimen examined was a female with small ova.

Remarks. Because remains of the ectoderm of the tentacles were stuck to the actinopharynx it was difficult to clear up the nematocysts there. I think, however, that the present information about them is more correct than that I gave in 1928. The size given there of the largest nematocysts was incorrect, caused probably by an error in measuring. In the maceration preparations of filaments with part of the mesenteries also large nematocysts similar to those of the acontia were present, probably belonging to these. The inner parts of the body, apart from the retractors, were not well preserved.

Genus *Adamsia* Forbes.

Hormathiidae with very wide base secreting a cuticle which may project beyond the mouth of the shell to which it is fastened. Column smooth, provided with cinclides sometimes situated on slight elevations. Sphincter well developed, mesogloal. Tentacles in several cycles, never long. Two siphonoglyphs and 2 pairs of directives. 12 pairs of perfect and fertile mesenteries. Mesenteries more numerous at the margin than at the limbus. The species of this genus live in symbiosis with hermit crabs, in such a way that their mouths are always situated beneath that of the hermits. Cnidom: spirocysts, basitrichs, and microbasie *p*-mastigophors.

Adamsia palliata (Bohadsch).

Actinia palliata Boh. STRÖM 1762-1766 p. 164.

Medusa palliata Boh. STRÖM 1788 p. 250 figs. 1-5.

Adamsia palliata Boh. SARS 1853 p. 20, DANIELSEN and KÖREN 1856 p. 87; APPELLÖF 1891 a p. 7; ARNDT 1912 p. 121; GRIEG 1913 p. 111; PAX 1936 p. 110.

For further literature see ANDRES 1883 and STEPHENSON 1935.

Diagnosis: Cinclides endocoelic in the middle and lower part of the column, usually not more than one in each endocoel. Margin distinct. Sphincter well developed with fairly large muscle-meshes showing at least a tendency to transverse stratification. Tentacles hexamerously arranged in up to 7-8 cycles, in number between 100-500, fairly short. Retractors of the mesenteries weak, diffuse. The perfect mesenteries, and some of the older imperfect, sterile. Occurs in symbiosis with *Eupagurus prodeauri* Leach. Nematocysts of the column $17-19.7 \times 2.2-2.5 \mu$, basitrichs, those of the tentacles $19.7-21 \times 2.5-2.8 \mu$, basitrichs, those of the actinopharynx $21-26.8 \times 2.8 \mu$, basitrichs, those of the filaments partly $19-22 \times 2.8-3(3.5) \mu$, microbasie *p*-mastigophors, partly $15.5-16 \times 2-2.5 \mu$, basitrichs, very rare, those of the acontia $26.8-33.2 \times 2.1-3.5 \mu$, basitrichs. Spirocysts of tentacles up to about $32.5 \times 3.5 \mu$.

Colour: Column brownish, light orange or reddish-brown with more or less vivid magenta spots of variable size, the parts beneath the hermit crab white or pale. Oral disc and tentacles usually white or cream-coloured. Acontia usually rose or lilac, rarely white.

Size: Longitudinal diameter of the discs up to about 5-7 cm.

Occurrence: Norway: Trondheimsfjord (teste ARNDT), the fjords at Bergen (teste APPELLÖF) Søndmor (teste STRÖM), Herdalfjord 20-30 m (Carlgren), Florø, Manger, Hardangerfjord, Bergenfjord (Sars), Flekkefjord (Esmark).

Further distribution: North Sea, British Isles, Atlantic coast of France, The Mediterranean.

Genus *Hormathia* Gosse.

Hormathiidae with well developed pedal disc often adherent to shells. Column divisible into scapus and scapulus. Scapus provided with tubercles showing at least a tendency to be arranged in rows,

of the scapus (100-200) (fig. 1) and only a ring of 12 coronal tubercles (100-200) (fig. 2) on the uppermost part of the scapus. The coronal tubercles are often small or absent, especially in the smaller specimens. Well developed cuticle, scapulus whitish, scapular siphon mesoglochal, strong. Tentacles 96, long, conical, reddish-yellow, with their outside, not more than 96 microns in diameter. Tentacles and mesenteries in same plane. Siphon 100-200 microns long. Siphon and radial muscles of the scapus, and the latter, sometimes meso-ectodermal, with a siphonoglyphis, 6 pairs of perfect and sterile siphonoglyphs, 6 pairs of perfect and sterile siphonoglyphs. Retractors diffuse. Oviducts, spermatocytes, basitrichs, and actinopharynx.

Hormathia digitata (O. F. Mull.)

- Hormathia digitata* (O. F. Mull.) GRAVEY 1857, p. 11, Pl. III figs. 9, 11, 12.
Hormathia digitata O. F. M. GRAY 1913, p. 144, 1923, 24, p. 29; 1926, p. 29.
Hormathia digitata O. F. M. CARLGRÉN 1930, p. 3, 1932, p. 263, 267, 1933, p. 27, 92, 13, 1939, p. 9, fig. 2; 1940, p. 44, fig. XIV, 9, 11, PAX 1956, p. 119.
Hormathia digitata n. sp. GRAVEY 1857, p. 11, 1922, p. 60, Pl. IV, figs. 42, 43, Pl. IX, fig. 97; CARLGRÉN 1934, p. 13.
 For further references see CARLGRÉN 1893, p. 110, 1928, p. 296 and SUNDINSSON 1935, p. 259.

Diagnosis. Pedal disc wide. Scapus usually provided with tubercles, the uppermost of which, forming 12 coronal tubercles, are stronger than the other and sometimes alone present. In younger individuals the scapus may be not tuberculated or the tubercles are small. Scapulus sometimes with ridges issuing from the coronal tubercles. Sphincter very strong, wholly separated from the ectodermal circular muscles of the column showing at first a tendency to transverse stratification, alveolar to more reticulate, especially in its upper part. Tentacles 96, long, conical. Actinopharynx of moderate length. Siphonoglyphis broad, aborally enlarged. Mesenteries 96, hexamerously arranged. Retractors of the scapus mesenteries well developed, diffuse, parietobasilar muscles weak, circular muscles fairly well developed. Viviparous. Nematocysts of the scapus partly 17-22 \times about 2.5 μ , basitrichs, partly 7-12 \times probably basitrichs, those of the scapulus 17-29 \times about 2.5 μ , basitrichs, those of the tentacles 23-34 \times 2.5-3.5 μ , basitrichs, those of the actinopharynx partly 26-36 \times about 4 μ , microbasitrichs, p-mastigophors, partly 26-31 \times 3.5-4 (4.5) μ , actinopharynx, those of the filaments partly 19-24 \times 4.5 μ , microbasitrichs, those of the filaments partly small basitrichs, sparse, those of the scapus 26-38-40 \times 3.5-4 (4.5) μ , partly about 11 μ , both basitrichs. Siphonoglyphs of tentacles up to about 43 \times 7 μ .

Coloration. Scapus more or less whitish, flesh-coloured, reddish or red-orange. Scapulus often of same colour as the tentacles, but more except the ridges. Tentacles reddish-yellow, dull or bright red or purple-brown, sometimes almost flesh-coloured. Oviducts reddish or coppery, pinkish-white sometimes with a pattern. Siphon and siphonoglyphs pinkish buff or with alternate stripes of reddish and white. SUNDINSSON (1935, p. 296) notes that the oral siphon is reddish or grayed in dark pigment on a paler ground and that the siphonoglyphs have pale pigment on a darker ground as *Hormathia digitata*, *Polydora cincta*, *Cataphylla*. As far I can remember *Siphonoglyphis* *digitata* has a pattern. See further CARLGRÉN 1893, p. 110, 1928, p. 296, 1945.

Size. Body length up to about 8 cm broad.

Localities. Iceland: 62° 05' N, 52° 49' W, 161 fms. (Ingegjerd exp., 1874).

Localities. Iceland: Upphæfils-straeti, Reyder 1857), 68° 20' N, 22° 30' W, 210 m. (Ingolf exp., 1908, S. 117). Egedesund (60° 50' N, 25° 17' N, 58° 44' W, 410 m. Bottom temp. 2.55). Ingegjerd (62° 05' N, 52° 49' W, 175 fms. (Tjalfe exp., 1925, St. 27). Skogv. Hólfjörðubanke, 66° 44' N, 56° 08' W.,

330 m (Tjalfe exp., St. 100); 66° 37' N, 56° 37' W., 450 m (Dana exp., 1925); 66° 35' N, 55° 54' W., 88 fms., bottom temp. 1.6 (Ingolf exp., St. 31); 65° 34' N, 4° 31' W., 68 fms., bottom temp. 0.2 (Ingolf exp., St. 29); 63° 56' N, 53° 12' W., 130 fms. (Wandel 1889); Fiskenas, Bredefjord 220-310 m, 245-410 m, 230 m, 250-280 m, 110-180 m, 115 m (Rink exp., 1962, St. 66, 77, 87, 97, 91, 111); Kvanefjord 115 m (Rink exp., 1912, St. 25); Tunngliartik 125-177 m (Rink exp., 1912, St. 110); Skovfjord 70-110 m (Rink exp., 1912, St. 156); Juhanehaab 1-10 fms. (Amundsen); Davis Strait 100 fms. (Holm), Greenland without distinct locality (Holm).

Iceland (see the chart fig. 2 CARLGRÉN 1939): Dýrafjörður 24 m; 66° 18' N, 25° 59' W, 621 m; Isafjörður off Hnífsdalur 50 m; Isafjörðardjúp 52 m; Jokulliröðir 63 m; Steingrimsfjörður 50-100 m, 188 m; Skagaströnd 56 m, 75-113 m; Hofsó's depth unknown; Eyjafjörður S. of Hrisey 32-38 m; Akureyri on a harbour pier; Skjálfaði 170 m; Axarfjörður 38 m; Þistillfjörður, Grenjanes 77 m; Mjólfjörður 60 m; Reyðarfjörður 130 m; 226 m, Berufjörður; 10-15 miles, S.S.W. of Eystrahorn, depth not noted; Mýrabúgur 63° 52' N, 16° 18' W, 109 m; 63° 12.5' N, 20° 06' W, 510 m; Vestmannaeyjar, 2 localities, one of them 75 m, 63° 12' 5' N, 21° 29' W, 151 m; 63° 35' N, 21° 29' W, 151 m; Keflavík 30 m; 64° 05' N, 22° 25' W, 38 m; Faxalói 64° 44' N, 23° 24' W, 65 m, Breiðfjörður 65° 11' N, 21° 21' N, 85 m, Iceland without distinct locality on the carapax of *Hyps aratus*.

West Spitzbergen, Spitzbergen Norway 79° 47' N, 11° 15' E., 100 fms. (Swedish Spitzb. exp., 1872-73). Prince Charles Forland 78° 41' N, 16° 08' E., 115 m (Olga expel., St. 27); Icefjord 221-254 m, bottom temp. 2.55 (Swedish Spitzb. exp., 1908, St. 40); 76° 43' N, 13° 40' E., 160 m; 25 miles N. of Bear Is., 179 m; 75° 23' N, 17° 45' E., 110-110 m; 74° 39' N, 18° 07' E., 110-155 m; 74° 15' N, 17° 36' E., 180 m; 73° 52' N, 19° 55' E., 130-200 m (Olga exp. Sts. 10, 16, 54, 51, 49, 44); 75° 31' N, 17° 50' E., 225 m. Bottom temp. 1.6 (Norwegian North Atl. exp., St. 326); 79° 20' N, 8° 17' E., 188-236 m, temp. at 180 m 2.17; 78° 56' 59" N, 10° 20' E., 40-237 m, temp. at 30 m 2.09, at 220 m 1.59; 78° 45' 78° 16' N, 15° 27' 15° 30' E., 87-100 m, temp. at 100 m 1.7; Advent Bay 78° 40' N, 13° 46' 5' E., 112 m; 77° 48' N, 11° 07' E., 120 m; 77° 47' N, 11° 22' E., 132-162 m; 77° 45' N, 11° 56' E., 117-160 m, temp. at 100 m 3.8; 77° 45' N, 11° 07' E.; 125-175 m; 77° 44' N, 11° 45' E., 185-228 m; 77° 44' N, 10° 40' E., 284-285 m; Bell Sound 241 m, bottom temp. 0.01; 120-190 m; 76° 50' N, 12° 50' E., 212-301 m; 76° 34' N, 76° 37' N, 28° 40' 28° 46' E., 155-170 m; 76° 16' N, 28° 43' E., 164 m, temp. at 150 m 1.9; 76° 08' N, 27° 30' E., 184-203 m; 75° 58' N, 28° 28' E., 197 m; 75° 58' N, 26° 52' E., 201 m; 75° 39' 8' N, 26° 47' E., 180-191 m, temp. at 170 m 0.78; 74° 29' N, 21° 42' E., 200 m; 74° 16' N, 20° 35' E., 115-125 m; 74° 15' N, 20° 30' E., 119-128 m; 74° 15' N, 21° 15' E., 135-120 m; 74° 12' N, 21° 52' E., 223-248 m, temp. at 210 m 1.89; 74° 04' N, 19° 40' E., 126-152 m, bottom temp. 1.9; 74° 74° 7' N, 20° 30' 20° 38' E., 175-245 m. (All localities teste GRAY); 78° 08' 30" N, 13° 44' E., 393 m; 76° 45' N, 23° 20' N, 23° 20' E., 186 m; 76° 30' N, 25° 27' E., 48 m; 72° 37' N, 20° 00' E., 177 m; 72° 37' N, 20° 00' E., 394 m (all 5 localities teste GRAVEY).

East Spitzbergen: King Charles Land 79° 00' N, 25° 55' E., 195 m.; Between Kamin Peninsula and Swatvi Noss, 78° 62 m; about 70° N, 40° E., 160 m (teste PAX); Kolafjord (Averinzeff, Derjugin); 10-160 fms. (teste PAX); 70° 49' N, 33° 46' E., 215 m, temp. at 200 m 1.6 (Michael Sars expel., 1901); Motovskifjord 69° 35' N, 32° 40' E., 53 m (Andr. Perwoswanny exp.); Chevanna 30 fms. (Sandeberg 1877); Semiostrova 50-55 fms. (Sandeberg 1877); N. off Ribatschi Peninsula 138 m (teste PAX); 70° 09' N, 31° 00' E., bottom temp. 1.05 (Michael Sars exp., 1901); Orafjord; Varangerfjord, E. off Ekero (Michael Sars exp., 1901); Vadsö 20-30 fms. (Klinekowsstrom); 71° 36' N, 25° 45' E., 300 m, bottom temp. 3.9 (Michael Sars exp., 1900, St. 57). North Cape, Magerö (Verkrusen); Porsangerfjord (Sars) 200 m, bottom temp. 0.2 (Nordgaard); Kistrand 20 fms.; Hammarfest (Lovén); Vargsund 250 m, Stjernov 200-350 m, Skaryskjar, Stjernovsund 300-350 m,

Hella, Öksfjord 200-25 m (Dons); Kvaenangen, Jokelfjord 1000 m, bottom temp. 1° (Nordgaard), Kvaenangen (Aurivillius); Lyngen fjord, Hornesgrunden 15-20 fms. (Bidenkap); Karlo 30-40 fms. (Malmgren); Ulvsfjord 25-30 fms. (Goës and Malmgren); Kjosen, Ulvsfjord (Norw. North Atlantic exp.); Kalfjord 30 fms.; Ramfjord 80-100 fms. (Goës and Malmgren); Grötsund 70 fms. (Goës and Malmgren); Kvalsund 20 fms.; Tromsø (Kier, Sars); 20-30 fms. (Wulff); Balsfjord, Storsteinnes, 70 fms. (Kier); Malangen, Stønnesbottu, 10-80 m (Nordgaard); Andenes (Kier); Bjarkøy 10-15 m; 30-50 m, 35-55 m, 50-60 m, 110-150 m; Gibstad 40 m, 50 m; Evenskiær 20-30 m, 35 m, 15-55 m; Ögsfjord, 100 m, bottom temp. 2.1° (Nordgaard); Skovna 200-300 fms. (Sars); Vestfjord (Bodöm); Föddenfjord 13 m, 15-60 m (Soot Ryen), 530 m, temp. at 500 m 6.55° (Nordgaard); 66° 12' N, 13° 13' E, 177 m (teste GRAYIER); Tjøtta, Nordland (Collin); Trondheimfjord: Tautra about 200 m (Mortensen); 30-50 m (Carlgren), Røberg about 300 m (Carlgren); Storfosen 200 m, Garten 250 m, off Södenes, about 100 m (Mortensen); Gjeiteneset 100-200 m (Dons); N.W. of Siggen in the archipelago of Bergen, 100 fms. (Swedish fishermen); fishing banks W.N.W. of Bergen, 80-170 fms. (Lambert); fishing banks N.W. of Bergen 60-200 fms. (Swedish fishermen); Glesvær (Lönnberg and Jägerskiöld); 15-21 miles S.E. of Haugesund and Bergen 100-170 fms. (Swedish fishermen); Hardangerfjord (teste GRÆG); Jäderen (Swedish fishermen); N.W. of Egersund 100 fms. (Swedish fishermen); Jutland banks 50-150 fms. (Swedish fishermen), Oslofjord 50 fms.; Björnen Ranö 200 fms.; Skagerrak 370 fms. (Gunnild exp. St. 7); 11 miles N.W. to N. of Höjen 58° 16' N, 9° 35' E., 660 m (Thor exp. 1908 St. 285); 36 miles N.W. to N. of Höjen 58° 12' N, 9° 53' E., 535 m (Thor exp. St. 223); Skagerrak, 220 m (Thor exp. 1903 St. 20); off Grisdarane, 85-98 m; Skagerrak (Petersen, *Urticina crassicornis*!); Kosterfjord 20-60 fms. (Aurivillius); Ramsö (Carlgren), Southern part of Kosterfjord, 220-230 m (Sandberg); off Hallö, 211-33 m (Thor exp. St. 212); Väderöar (B. Fries, Carlgren); W.S.W. $\frac{1}{2}$ W. of Maseskiær, 120 m (Thor exp. St. 1091); Gullnaren 25-80 m (Carlgren and others); W. of Winga, 27-29 fms. (Thor exp.); Kattegat (Lovén, Olsson, Petersen); Nidingeknölen 30 m ("Akka" St. 70); about 56° 51' N, 12° 05' E., 16 m ("Akka" St. 103); about 56° 51' N, 12° 11' E., 56 m ("Akka" St. 81); 10 miles N.W. to W. of Kullen, 32 m (Thor exp. St. 309); N.E. to E. of Torokow, 11 fms. (Lönnberg); between Kullen and Vinga skär at Hallands Väderö, 14.5 fms. (Lönnberg); Krapperrup (Gyllenstierna); Danmark, Hellebæk (Lütken); the Sound (Lütken, Winther and others); N. of Hven, 14 fms. (Kramp).

Faroe Isl. 12 miles S. of Akraleiti, Sudero 260 m; Kvivig, Stromo (teste FABRICIUS and LANDI) 61° 03' N, 2° 13' E., 130 m, temp. at 125 m 6.78° (M. Sars Exp. 1902 St. 49), 61° 28' N, 1° 15' E., 100 fms. (Mich. Sars Exp. 1902 St. 28), 61° 16' N, 1° 18' E., 150 m (Swed. Greenl. Exp. 1899), 55° 30' N, 1° E., 40 fms., Shetland Isl. Outer Skerries Haaf (teste NORMAN).

Further distribution: North Sea.

Exterior aspect. The exterior of this species varies considerably and in the same manner as *Hormathia nodosa*. In younger individuals the scapulus is smooth or almost so, or provided with small tubercles, older individuals have more or less strong tubercles, especially in the upper part of the scapulus (Pl. I figs. 5, 8), sometimes the tubercles are small apart from the coronal tubercles, (Pl. I fig. 2) sometimes only the coronal tubercles are present. I have noted (1933 p. 26) that most specimens from West Greenland have this appearance (Pl. I fig. 11) but also in the Skagerrak apparently similar ones occur. Pl. I fig. 6 shows an individual from this water, here the coronal tubercles were very small. Rarely the species lives in accidental commensalism with the crayfish *Hyas* (Pl. I fig. 3). Small individuals can easily be confounded with *H. nodosa* but the species can be distinguished from each other by the size of the nematocysts. *H. nodosa* seems always to be more elongated than *digitata* also in the contracted state (see the figures 2, 3 Pl. III and 6, 10 Pl. IV).

Anatomical description see CARLGRÉN 1893. The nematocysts of the tentacles have been measured in 15 individuals, those of the actinopharynx in 10, those of the acontia in 20 specimens. The breadth noted by me in STEPHENSON, 1935, of the nematocysts is too narrow.

In a small, strongly contracted specimen from Kvanefjord, Greenland, curious parasites occur in the mesogloea below the sphincter which here showed a more reticular appearance than usually in *digitata*. These parasites were now scattered in the meso-



Fig. 53. *Hormathia digitata* var. *vegae*. a, sphincter of a specimen. b, part of sphincter in another specimen (both from Stolbowed-Büschmi).

gloea, now more and commonly accumulated in smaller or larger groups which sometimes were of considerable size (Pl. V fig. 4). Evidently these parasites are identical or nearly related to those I have noted (1934 p. 15 and 17, fig. 11) in the ectoderm of oral disc of *Paraphellactis michaelsarsi* and in the mesogloea of the scapus of *Actinotage abyssorum*, though they are considerably more numerous here. In the figs. 2 and 3 Pl. VI I have drawn some of these parasites in high magnification. Several of the parasites were free and their form was more or less fusiform or more irregular, sometimes groups of them were encysted (Pl. VI fig. 2). Probably we have to do with Gregatines, probably of another kind than those I have noted in *Sagittotrocton calvatus* and *andatus* (p. 26 text fig. 29).

In the collection of the Actiniaria from the Vega expedition there were 4 specimens which I with some hesitation refer to *Hormathia digitata* but as a special variety, *vegae*. One individual was taken east of Taymyr peninsula either at 76° 52' N, 116° E., in a depth of 36 fms. (bottom temp. = 1.4°) or at 76° 40' N, 115° 30' E., in a depth of 35 fms. (bottom temp. = 1.6°) (the collections from both localities which, according to SUMMERS (1880), had same annual life, seem to have been, at least partly, put together), the three others were dredged between the islands Stolbowed and Büschmi at 73° 53' N, and 138° E., in a depth of 12 fms. (bottom temp. = 0.4°). The size of the largest specimen was: largest breadth of the pedal disc 3 cm, height of the column

scapus 17-20 μ , the smallest specimen 17 μ . I have drawn the exterior of the scapus in figure 12 Pl. III. As we see the setae are very closely set, but not so closely as in *H. doptata*. The types and size of the setae are in general in agreement with those in *H. doptata*. The projections of the filaments seem to be of the typical form I have, however, drawn in figure 13 Pl. III. The spinneter (fig. 53) recalls that of *H. doptata* (fig. 42) and is provided with 11-13 specimens of setae. The setae as shown is showed a different appearance from those of *H. doptata*. Pl. III fig. 12 Pl. III. The scapus is covered with a developed cuticle, in the tubercles somewhat thicker. It has several outshoots from the mesogloea here and there (fig. 8). Sometimes these outshoots were shorter, sometimes longer, and cast to the cuticle.

When the tentacles are running out from these projections the connection with certainty decide, but at any rate the connection is a definite connection between them and the cuticle. The tentacles being red in mucous cells which were absent, and the connections. The whole organization recalls that of *H. doptata*.

Polysyllus taken from the Danish Dymplina expedition there were 4 specimens taken in the Kara Sea and attached to tubes of *H. doptata*. I have drawn one of them, the largest, in figure 6 Pl. III. The three others were smaller and had only about 48 tentacles. All were not sexually ripe. Especially one of the smallest specimens was provided with papillae recalling tenaculi. Sections were made but we have to do with such formations. Here as far as I could see the cells connecting the mesogloea with the cuticle were not present. Sections of the column of the largest specimens were in general in agreement with the individuals from the Vega expedition. The size of the nematocysts was the same as in *H. doptata* (2 individuals examined). Moreover the whole organization recalls that of this species. There is, I think, no doubt that the specimens from the Dymplina expedition were identical with those from the Vega expedition but younger.

Remarks.—Owing to presence of more or less developed tentacles with some hesitation referred specimens described above to the genus of *H. doptata*. Nothing is known about such formations in *H. doptata* but it is possible that very young specimens are provided with tenaculi in some form or other. As far as I can see the specimens of *H. doptata* have sometimes an appearance of the smaller individuals from the Dymplina expedition. I have, however, at present no opportunity to examine the specimens. In case that tenaculi never occur during the life of *H. doptata* I think it is necessary to erect a new genus for the specimens from the Dymplina and Vega expeditions. I name this genus *Hormathia nodosa*.

Hormathia nodosa (Fabr).

Cuvier, *Atlas*, p. 14, Pl. IV, figs. 6, 19.

Forsk., *Gen. Mus.*, 1823, 21, p. 29, 1926, p. 26.

H. nodosa (Fabr.)—Forsk., *Gen. Mus.*, 1823, 21, p. 29, 1926, p. 26; *Atlas*, 1795, 14, 1963, p. 352, 1939, p. 9.

Forsk., *Gen. Mus.*, 1823, 21, p. 12, Pl. III, fig. 4.

H. nodosa (Fabr.)—Forsk., *Gen. Mus.*, 1823, 21, p. 115, 1928, p. 291; *Atlas*, 1795, 14, 1963, p. 352, 1939, p. 9.

Body cylindrical, with column elongated, often flattened. Scapus and scapulus, scapulus with setae more sparsely set, sometimes with a few more slender, rarely absent or weakly developed. Filaments on scapulus rarely absent or weakly developed. Nematocysts present in the uppermost part of the scapus and oral tubercles. Cuticle of the

scapus more or less strong, scapulus smooth. Spinneter situated nearer the ectoderm than the endoderm, reticular in its uppermost part, for the rest alveolar with a distinct tendency to transverse stratification, in young individuals reticular (always 4) but the muscle meshes somewhat larger. Tentacles 96, in contracted state often longitudinally sulcated. Longitudinal muscles of tentacles strong, ectodermal, radial muscles of oral disc meso-ectodermal. Actinopharynx very long, siphonoglyphs very broad. Retractors of the mesenteries as those of *H. doptata*. Parietobasilar and basilar very weak. No marginal stomata. Nematocysts of the scapus partly 17-23 \times 2.5 μ , partly 19-12 \times 1.5 μ both basitrichs, those of the tentacles partly 26 \times 2.5-11 (13) \times 3.5 μ , partly 14-19 \times 1.5 μ rare, both basitrichs, those of the actinopharynx partly 26-37 \times 2.8-3.5 μ , partly 11-19 \times 1.5 μ , both basitrichs, partly 26-32 \times 2.5 μ - microbasic *p*-mastigophors, those of the filaments partly 12.7-15 \times 1.5-2 μ , basitrichs, partly 21-29 \times 1-1.5 μ microbasic *p*-mastigophors, those of the acontia (13 specimens examined) partly 31-18 \times 3.5-1 μ , partly 11-18 \times 1.5 μ , few, both basitrichs. Spirocysts of tentacles up to 58 \times 5-6 μ .

Colour: Scapus light brown, flesh coloured or pale red, covered with brown or mud-coloured cuticle. Scapulus pink or red. Inner tentacles brown-red, intermediate tentacles paler, outermost pale whitish. Round the mouth is a pale yellow-white narrow annulus from which pale rays issue to the inner tentacles (Danielsen 1891, Verrill 1922).

Size up to 8 cm in length and 5 cm broad in the contracted state. The largest tubercles up to 1 cm in diameter.

Occurrence: Hudson Bay, Richmond Gulf, 25 fms.; Jones Sound, Havnefjord between 76 25' N. and 76 40' N. and 80 20' W.-81 15' W. (Fram Exped. 1898-1902); Baffin Land, Entrance to Furry and Hekla Straits, 29-30 fms. (Norcross-Bartlett exp. 1933); Labrador, Saglek Bay (Bartlett).

North Greenland without distinct localities (Kjeldsen, Ryder, Møller).

West Greenland: 76 40' N., 76 20' W., 580 m, bottomtemp. \pm 1.1 (Godthaab exp.); Cape Dudley Digges 76 01' N., 68 28' W., 30-15 m (Nilsson); Melville Bay 75 26' N., 67 27' W., 260 fms. (Sofia exp. 1883); Upernavik (Ryder); Upernaviks district, Prøven 16-100 fms. (Torell, Amundsen, Olrik, Bollbroek; Umanak (Amundsen, Torell); N. of Hare Isl. 70 20' (McClain); Disco Isl. 69 31' N., 56 01' W., 100 fms. (Valorous exp.); Jakobshavn (Traustedt); Christianshaab, 15-30 fms. (Öberg); Egedesminde (Traustedt); Ikamiut (Lohmann); North Stromfjord St. 10 A, 22-31 (Nordmann); Davis Strait, 100 fms. (Holm); Kangamiut, 130 fms. (Nielsen); 65 36' N., 56 21' W. (Wandel); 65 31' N., 54 31' W., 68 fms., bottomtemp. 0.2 (Ingolf exp. St. 29); 65 17' N., 54 17' W., 55 fms. (Ingolf exp. St. 31); 63 56' N., 53 12' W., 130 fms. (Wandel); Fiskernes (Öberg); Ivigtut (Munster); Bredefjord, 170-180 m (Rink exp.); 60 22' N., 47 27' W., 120 m, bottomtemp. 5.8 (Godthaab exp. St. 188); Hua (Lundholm); Greenland from the stomach of dog-fishes.

East Greenland: Stormbugt, 8-10 fms., 20 fms.; Danmarks Havn, 10-15 fms.; "Oresund", 10-15 fms.; sound between Maatten and Renskiar, 25-30 fms. (all 4 localities from the Danmark exp.); 73 55' N., 19 20' W., 150 m (Swedish Polar exp. 1900); 73 30' N., 28 18' W., Cape Broer Rys, 25-27 m (Swedish Greenland exp. 1899); Clavering Isl., 10-35 m (Bartlett); King Oscarfjord at the entrance to Kempfjord, 100-55 m (Norwegian exp. 1930); Heklas Havn, 3-6 fms. (Ryder 1894); Tasiusak, 25-30 fms. (East Greenland exp. 1899); Kangerdlugssuak, 11-15 m (E. Mikkelsen 1932); Jan Mayen (Austrian Polar exp.).

Iceland: 2 miles off Seyðisfjörður, 113 m; Reyðarfjardardjúp, 226 m; N.E. of Iceland 66° N., 14 41' W., 280 m (Thor exp. 1903); 64 53' N., 10 W., 630 m, temp. at 100 m \pm 0.69 (Michael Sars exp. 1900); 62 38' N., 14 0' E., 650 m, bottomtemp. \pm 0.45; 62 35' N., 1 04' E., 620-640 m, temp. at 620 m \pm 0.03 (Michael Sars exp. 1902).

Faroe Island: Thorshavn (Müller); 250 fms. on fish-line (Michael Sars exp. 1902).

N. of Spitzbergen: Ross Isl. 80 18' N. 20 23' E., 85 m (Römer and Schaudinn exp. 1898 St. 13); Low Isl., 16 fms. (Swedish Spitzberg exp. 1861).

West Spitzbergen: Mossel Bay, 9-10 fms.; N.W. off Mossel Bay, 15-20 fms.; Gray Hook, 60 fms.; Norwegian Isl. (all 4 localities from Swedish Spitzberg exp. 1861); 79° N. 11° E., 36-140 m; 78°44' N. 10° 8' E. 115 m (Olga exp.); 79° 10' N. 11° E., W. of Cape Mitra, 100 m (Swedish Polar exp. 1900); Cross Bay, 40-60 fms. (Swedish Spitzb. exp. 1861).

Icefjord: Cape Boheman, 40-50 m (Swed. Spitzb. exp. 1898); entrance of Save Bay, 125-150 m, temp. at 125 m, 0.87°; 141 m 1.23°, entrance of Dickson Bay, 14-14 m, 29-27 m, bottom temp. about 2°; Klas Billen Bay, 32-40 m, bottom temp. 1.5-2°; Sassen Bay, 19-28 m, bottom temp. 2-3° (all localities from Swedish Spitzberg exp. 1908); Save Harbour, 30-40 fms. (Malmgren 1864); Bel Sound 39-40 fms. (Torell); Duyns Point (Swedish Spitzberg, exp. 1872-73); off Horn Sound 210 m (Swed. Spitzberg, exp. 1898); 76° 13' N. 13 10' E., 160 m; 76° 23' N. 15 7' E., 145 m (Olga exp.) - teste GRIGG: 79 20' N. 8 17' E., 188-236 m; 78 56' 59" N. 10 20' E., 40-237 m, temp. at 220 m, 1.59°; 78 15' N. 9 22' E. 282-297 m, temp. at 275 m, 2.85°; 78 10' N. 13 46' E., 412 m; 77 13' N. 12 07' E., 130 m; 77 11' N. 10 10' E., 281-285 m; 77 41' N. 11 50' E. 113-150 m; 76 58' 5 N. 12 15' E., 220-253 m; 76 56' N. 12 50' E., 212-301 m.

East Spitzbergen: 80° N. 19 05' E., 40 fms.; 79 57' N. 17 05' E., 25 fms.; Treurenberg Bay, 6-30 fms. (Swedish Spitzberg, exp. 1861); Treurenberg Bay, 22 m (Monaco exp. St. 1074); 79 40' N. 19 5' E. Foster Isl. 10 fms.; Waigat Isl. 30 fms. (Swedish Spitzberg, exp. 1861); Hinlopen Strait, Behm Isl. 79 20' N. 20 55' E., 80 m; Bismarck Strait 78 58' 5" N. 20 35' E., 35 m; Greatfjord close by Chanking Point 78 25' N. 20 E., 105-110 m; Cape Blanck 77 49' N. 20 3' E., 65 m (Römer and Schaudinn); W. Thymen Strait, 39-46 fms. (Malmgren 1864); 78 14' N. 21 45' E., 38 m; Ryk Yse Isl. 77 49' N. 25 12' E., 60-80 m; 78 5' N. 26 40' E., 290 m (Römer and Schaudinn); King Charles Land, Bremer Sound, 100-110 m (Swedish Spitzberg, exp. 1898); between Jena and Abel Isl., 40 m; Swedish Forland, 2 miles from Cape Arnesen, 85 m (Römer and Schaudinn). East Spitzbergen without distinct locality (Kükenthal).

Spitzbergen-Bear Island Norway: 75 49' N. 24 25' E., 80 m (Swedish Spitzberg, exp. 1898); 75 10' N. 17 30' E., 179 m; 71 31' N. 17 E., 110-110 m; 75 09' N. 17 17' E., 138-191 m; 71 39' N. 18 07' E., 140-155 m; 74 31' N. 17 E., 165 m; 71 17' N. 17 35' E., 156 m; 73 52' N. 19 55' E., 130-200 m (Olga exp.); 22 quarter-miles S.E. of Bear Isl., 130 m, bottom temp. 0.05°; Bear Island, 130-140 m (Michael Sars exp. 1901); 72 27' N. 20 51' E., 349 m, bottom temp. 3.5° (Norwegian North Atl. exped. St. 290); -teste GRIGG: 76 16' N. 28 13' E., 161 m, temp. at 150 m, 1.9°; 76 15' N. 27 29' E., 136 m, temp. at 125 m, 0.75°; 76 08' N. 27 30' E., 181-203 m; 75 39' 8 N. 26 17' E., 180-191 m, temp. at 170 m, 0.78°; 75 28' N. 26 13' E., 207 m; 75 23' N. 26 33' E., 192 m; 71 71 7' N. 20 30' 20 38' E., 175-215 m; 74 57' N. 26 08' E., 197-202 m, temp. at 190 m, 0.95°; 71 15' N. 20 30' E., 119-128 m; 71 15' N. 21 15' E., 135-220 m; 73 19' N. 32 07' E., 330 m, temp. at 315 m, 0.9°; 73 17' N. 48 20' E., 210-243 m, temp. at 200 m, 1.98°.

Frantz Joseph Land: 10 miles E. of Victoria Isl., 70 m; 4-2 miles off Cape Flora, 50-60 m; S.W. of Camp. Ziegler, 60-80 m (Norwegian exp. 1930); northwest coast of McClintock Isl. 80 22' (Baldwin-Ziegler exp. 1902).

Barents Sea: 70 49' N. 33 46' E., 245 m, temp. at 200 m, 1.6°; 70 9' N. 31 E., 200 m, bottom temp. about 1.5° (Michael Sars exp. 1901); close by the entrance of Motowski fjord, 272 m (Andrej Perwosvanni exp.) 13 miles N. of Sem Isl., 174-184 m (teste PAX), Murman Coast, several localities (teste DERJUGAN, BREITFUSS, PAX); Olenja Guba (teste Pax).

Norway: Varangerfjord E. of Ekero; Kiberg 200 m; Kongsfjord 100 m (Michael Sars exp. 1901); Hammarfest (Lovén); Porsangerfjord (Nordgaard).

W. of Kara Strait, 70 25' N. 57 56' E., 66 m (Duc d'Orléans exp.). Arctic Sea of Siberia 76 52' N. 116 E., 36 fms., bottom temp. 1.4°; 76 40' N. 115 30' E., 36 fms., bottom temp. 1.6° (Vega exp.).

Exterior aspect: The species varies very much in its exterior (see the diagnosis). In the figures 2 and 3 Pl. III and 6 and 10 Pl. IV I have given some examples of this variation. In the specimen drawn in fig. 2 only the coronal tubercles are developed, fig. 3 shows an individual with fairly closely set, rounded tubercles, in the specimen photographed in fig. 6 the tubercles are fairly sparse. The largest specimen I have seen (fig. 10) had very large, often a little acuminate tubercles scattered over the whole surface of the scapus. Usually the species is considerably longer than *H. digitata*. Where both species occur together it is possible that they hybridize (Carlgren 1902 p. 52).

Anatomical description see CARLGRÉN 1893 p. 115.

Genus *Stephanauge* Verr.

Hormathiidae with the pedal disc well developed, often elongated in the transverse plane and often clasping octocorals, sponges or tubes of worms. Column rather low, indistinctly divisible into scapus and scapulus, the former smooth, the latter, at least in the contracted state, longitudinally furrowed and sometimes provided with tubercles more or less regularly situated. Cinchides few, only in the directive compartments. Sphincter mesogloae, well developed, perfectly separated from the endodermal circular muscles of the column. Tentacles of ordinary length or longer, with or without mesogloae thickenings at the outside of the base, probably more numerous than the mesenteries at the limbus. Longitudinal muscles of the tentacles ectodermal, radial muscles of the oral disc ectodermal or ecto-mesogloae. Two siphonoglyphs. Perfect pairs of mesenteries 6 or 12, 2 pairs of directives. Mesenteries hexamerously arranged. Retractors of the mesenteries rather weak, parietobasilar muscles weak. All stronger mesenteries, the directives sometimes excepted, fertile. Acontia few (sometimes absent?). Cnidom: spirocysts, microbasic *p*-mastigophors, basitrichs.

The difference between the genera *Stephanauge* and *Amphiatthus* I have pointed out in 1925 and 1928. As we see below I have not observed any acontia in the type species, *S. neaxis*, and as to *S. acanellae* (*abyssicola* Mos.) the presence of acontia is somewhat doubtful, I think, however, that an examination on good material will show the presence of acontia also here as in *S. barbosa*; the gonads of the specimens examined of *neaxis* and *acanellae* were very strong and filled up almost the whole coelenteric cavity rendering the examination more difficult. VERRILL mentions, however, the presence of few acontia in *neaxis*.

Stephanauge neaxis (Verr.).

Actinauge neaxis n. sp. VERRILL 1883 85 p. 55 Pl. 16 figs. 1, 5, 1883 b p. 511, 531 Pl. 7 figs. 22, 22 a; 1899 p. 145.

Stephanauge neaxis (Verr.) VERRILL 1922 p. 99 G., Pl. 22 figs. 5, 6, Pl. 28 figs. 1-4, Pl. 30 fig. 3.

Stephanauge abyssicola p. p. VERRILL 1899 p. 217 fig. 34.

Diagnosis: Pedal disc prolonged, clasping the denuded axis of *Balticina* and other Pennatulids. Column rather low, with thick mesogloae, smooth in its proximal part, in the distal part, at least in the contracted state, with longitudinal ridges terminating sometimes at the margin in irregular tubercles. A circular, distinct wall below the furrows in certain states of contraction. Cinchides 1-4 or 1-2 rarely 3-2. Sphincter strong but not filling up

stomata were well developed as also the marginal stomata in the uppermost part of the stronger mesenteries. The gonads were present in all or almost all mesenteries, the directives can be fertile



Fig. 55. *Stephanauge nexilis*. Transverse section of tentacle.

or sterile. Because of the strong development of the gonads, filling up almost the whole cavity of the body, I have not been able to find any acontia, in the maceration preparations of the



Fig. 56. *Stephanauge nexilis*. Section of a directive mesentery in the lower part of the actinopharynx.

base of the mesenteries I have observed large nematocysts of very different length (13.2) 59.2 93.1 \cdot 1.2 (5) μ , which perhaps belonged to acontia as there were no such nematocysts in the

filaments. VERRILL 1922 p. 99 mentions, however, that acontia are present, but apparently few. He has drawn a figure (6 Pl. 22) of this species with 1 ejected acontia. As far as I can see the figure represents this species. As all specimens examined by me concerning the arrangement of the mesenteries had two pairs of directives symmetrically arranged, the species probably does not propagate asexually. The nematocysts were usually rather opaque but I think that the nematocysts marked as microbasic p mastigophors are such.

Remarks: VERRILL in 1922 described the species in detail but in several points incorrectly. The tentacles are about twice as many than VERRILL states. The cinclides are present only in the directive chambers, not scattered. The figure 6 Pl. 22, certainly a magnification of fig. 5 Pl. 6 in the paper of 1883 (Blake-exped.), showing threads representing ejected acontia, is probably incorrect, as it gives an idea of scattered cinclides. The perfect pairs of mesenteries are only 6, not 12 or more as VERRILL states. His figured sections Pl. 28 figs. 3, 4, Pl. 30 fig. 3, 1922, are sections through the oral disc, not through the actinopharynx. Also the first cycle of mesenteries are fertile, in opposition to VERRILL'S statement.

Stephanauge acanellae (Verr.)

Sagartia acanellae n. sp. VERRILL 1883 85 p. 16 Pl. 6 figs. 2, 2 a; 1883 b p. 511, 534 Pl. 11 fig. 25.

! *Actinia abyssicola* n. sp. MOSELEY 1877 p. 297 Pl. 4, 5 fig. 5.

! *Stephanaectis abyssicola* (Mos.) R. HERTWIG 1882 p. 79 Pl. 2 fig. 13.

! *Stephanauge abyssicola* (Mos.) VERRILL 1899 p. 115; p. p. CARL-GREN 1928 p. 300.

! *Raphactis abyssicola* (Mos.) VERRILL 1922 p. 101.

Diagnosis: Pedal disc prolonged in the transverse plane of the body and clasping the stalk of *Acanella* or *Mopsea* so that the lateral lobes come into contact with each other. Column either, and usually, smooth, even in large specimens, or provided with tubercles in the uppermost part. In the latter case the tubercles are sometimes numerous and closely set, sometimes few, scattered or with a tendency to an arrangement in longitudinal rows. Sometimes there are 12 larger tubercles on the circular wall, which is more or less distinct according to the stronger or weaker contraction of the column. Cinclides distinct, often 2 from each directive compartment. Splincter strong, alveolar, of about equal breadth, filling up most of the breadth of the mesogloea. Tentacles numerous, up to about 130-140, hexamerously arranged in 5 or 6 cycles, thin, conical, not thickened at the outside of the base, the inner comparatively long, the outer shorter and considerably thinner. Longitudinal muscles of the tentacles stronger at the inside than at the base of the outside. Radial muscles of the oral disc rather well developed. Two distinct siphonoglyphs with strong aloral prolongations and situated about in the transverse plane of the prolonged basal disc. Mesenteries in 5 cycles, hexamerously arranged, the last cycle incomplete. Mesenteries at the limbus somewhat fewer than at the margin, 6 pairs of perfect mesenteries as a rule; the mesenteries of the second cycle, so far as can be seen, are rarely perfect in the uppermost part of the actinopharynx. Longitudinal muscles of the mesenteries weak, directive mesenteries, however, with distinct retractors. Parietobasilar and basilar muscles weak. Gonads in all the stronger mesenteries. Nematocysts of the column very numerous, partly 17 24 \cdot 3.5 1.2 μ microbasic p -mastigophors, partly 12.7 11.1 \cdot 3.5 1.2 μ probably basitrichs; those of the tentacles partly 19.7 21.1 \cdot 1.2 (1.5) μ microbasic p -mastigophors, partly 12 15.5 \cdot 3.5 1.9 μ basitrichs; those of the actinopharynx partly 18.3 22.6 \cdot 5 5.6 microbasic p mastigophors?, partly 16.2 19.7 \cdot 1.2 μ basitrichs; those of the filaments partly 19.7 22.6 \cdot 3.5 1.5 μ , partly 10.2 15.5 \cdot almost 2 2 (2.5) μ , basitrichs, partly 16.9 21.1 \cdot (3.5) 5 (5.6) μ , probably microbasic p -mastigophors, spirocysts of the tentacles up to about 36 (11) \cdot 6.3 μ .

Color of the oral disc, pedoderm, and column, colorless, or brownish (1 specimen); anal disc larger than the body, often orange or reddish brown, or yellowish green, but frequently dark purple (1 specimen). Color of the tentacles usually similar to the tentacles usually present in the column with brown (VERRILL) (*abyssicola*, 1 specimen), or white with few radial streaks of slightly darker color (1 specimen); coronar wall pale pinkish yellow, oral disc pink and disc dark pink (MOSELEY).

Length of the body larger diameter of the base in larger specimens 1.5-2.5 cm, transverse diameter up to 1.5 cm, height of column up to 1.5 cm, length of the tentacles (VERRILL). In the specimen taken in Davis Strait the length of the base was 2.5 cm, length of the body 1 cm.

Localities: Davis Strait 66° 49' N, 56° 28' W, 113 m (Wandel 880 fms. depth, S. of Iceland 62° N, 21° 36' W, 1591 m, Bottom 333 fms., Ingot exped., St. 491, 42° 59' N, 51° 15' W, 1100 m, Temperature at 914 m, 3.70° C. Michael Sars' exped., 1910, St. 70) Georges Bank, E. of Georges Bank 677 fms., St. 2528-1885, 2 specimens; off Nantucket Shoals 497 fms., (U. S. Fish Com., St. "Albatross" St. 2046) 1 specimen; off Martha's Vineyard 372 fms., 365 fms., Fish Hawk St. 893, 894 7, 1 specimen; 40° 02' N, 68° 50' W, 547 fms., (U. S. Fish Com., St. "Albatross" St. 2048) 13 specimens (see VERRILL). Several localities between 39° 50' 15" and 41° 32' N, and between 65° 35' 30" and 70° 18' W, 260-1212 fms. (Blake exped.), Nova Scotia, Grand Bank; (*abyssicola* 40° 17' N, 66° 48' W, 1350 fms., Challenger Exped.).

Exterior aspect. The pedal disc was, in all specimens examined by me, prolonged about in the transverse plane of the gonads and usually clasping branches of *Acanella*. The column was usually smooth, some specimens had small tubercles, showing a tendency to be arranged in longitudinal rows in the uppermost part of the column. In the specimen taken in Davis Strait, one side was provided with numerous small tubercles distally and an annulus of low but broader tubercles on the indistinct coronar wall. Smooth specimens as well as such having tubercles have been taken in the same locality. The circular wall is nothing more than a product of a certain contraction of the body, often not visible or only visible in a part of the periphery. The number of the cinclides varies in the directive compartments but seems usually to be 2 × 2 or 2 × 1 more seldom fewer or more numerous. Of 22 closely examined specimens 9 had 2 × 2 cinclides, 7 2 × 1, 3 1 × 1, 2 3 × 2 and 1 1 × 1 cinclides. The cinclides were usually situated perpendicularly to the longitudinal axis of the pedal disc although sometimes small displacements in their arrangement occurred.

The tentacles are rather numerous, more than 100, in larger specimens up to 130-140. They are comparatively long, conical, and thicker at the outside of the base and often, also in preserved specimens, with long filiform tips. The inner tentacles are larger than the outer and somewhat longer than the often filiform outer ones. The tentacles seem to be somewhat more numerous than the tentacles at the limbus, though the difference is small. In one specimen I counted 149 tentacles and 101 mesenteries at the limbus, in another 190 resp. 101, in a third 114 resp. somewhat more than 100 tentacles and 120 resp. 120. Thus, the last cycle of mesenteries is rather small, the larger and grow downwards or at least upwards, and are greater at the base than at the limbus. At any rate the number of tentacles between 9 × specimens and 8 × *orbis* on the one hand and the 20 × *Ampeliontha* on the other, inasmuch as the latter has a considerably more mesenteries at the limbus than the former. The oral disc is very wide and the mouth opening is very small. The actinopharynx is provided with 2 broad bands of muscle, one above and one below the oral disc.

Actinopharynx and tentacles. The ectoderm of the column is contracted in the contracted state of the animals rather than in the extended state. The ectoderm is thickened between the

insertions of the directive mesenteries. The sphincter is alveolar, perfectly differentiated from the circular, endodermal muscles of the column, in all places of almost same breadth and filling up most of the breadth of the mesogloea. Its muscle meshes are small, in the uppermost part somewhat larger (fig. 57). The ectoderm of the tentacles is very high, the mesogloea about equally developed on the abaxial as on the adaxial side. The longitudinal muscles of the tentacles are ectodermal and weak, especially on the abaxial side. Also the radial muscles of the oral disc are weak. The ecto-



Fig. 57. *Stephananthe acanella*, Sphincter.

derm and the mesogloea of the actinopharynx are rather thick. The ectoderm of the whole actinopharynx is pigmented, as also that of the column, tentacles, and oral disc.

The mesenteries are hexamerously arranged in 4 cycles, to which sometimes are added some mesenteries of a fifth cycle. There are two pairs of directives (always 4). As a rule only the 6 first pairs of mesenteries are perfect, in one specimen it seems as though the mesenteries of the second order reach the uppermost part of the actinopharynx. The longitudinal muscles of the mesenteries form weak retractors, somewhat stronger pennons are present on the directives. The parietobasilar and basilar muscles are weak. The gonads are present at least in all the stronger mesenteries. Also in this species I have not been able to ascertain the presence of acontia because of the very strong development of the gonads and the bad preservation of the interior of the animals. The type of the nematocysts were very difficult to decide. Apart from the other nematocysts of the column and tentacles there were strongly opaque nematocysts present. They were in the column 19.7-25.1 × 5-5.6 (6.3) μ , in the tentacles (44) 20.4-25.4 μ . Probably they were developmental stages. The broadest nematocysts of the filaments showed a strongly granular appearance, so that it was impossible to distinguish the type, but in another example they seem to be microbasic *p* mastigophors. In this example I found also stenothels

and holotrichs, certainly foreign bodies, in the maceration preparations of the filaments and mesenteries.

Remarks: Although it may be possible that MOSELEY's *Actina abyssicola* (= *Stephanactis abyssicola* R. Hertwig) and *Sagartia acanellae* Verr. are identical species, I have here, in opposition to what I wrote in 1928 (p. 300-301), preferred to call the above species *Stephanauge acanellae* and put up with a query *A. abyssicola* as synonymous with *acanellae*. If the statement of MOSELEY and R. HERTWIG is correct as to the number of mesenteries and the tentacles—MOSELEY mentions about 60 tentacles and HERTWIG 90-100 mesenteries at the limbus in *abyssicola*—we should have to do with two different species, *Amphianthus abyssicola* and *Stephanauge acanellae*. But there were 2 specimens in the collection and it is doubtful whether the statements refer to the same specimen. However that may be, *Sagartia acanellae* is nearly related to *Stephanauge uerilis* and may go to the same genus. As to *A. abyssicola* Mc MERRICH (1893 p. 193) and VERRILL (1899 p. 115) have suggested, that this species is identical with *Stephanauge uerilis*, but later VERRILL (1922 p. 101) refers it to his genus *Raphactis* (= *Amphianthus* CARLGRÉN 1931 p. 10).

Genus *Amphianthus* R. Hertw.

Syn. *Korenia*, *Gephyra*, *Gephyropsis*, *Raphactis*, *Stelidactis*, *Sycaanthus* (p. p.), *Stephanactis* (p. p.).

Hormathiidae with the pedal disc often prolonged in the transverse plane and clasping branches of octocorals, sponges or other objects. Column rather low, mostly thick, sometimes smooth, sometimes provided with small tubercles, commonly arranged in more or less distinct longitudinal rows. Cinclides as a rule only from the directive compartment, sometimes singular, sometimes 2-4 in each compartment, in the latter case one above the other, probably always principally endodermal evaginations (cinclides in young specimens indistinct or absent?). Column in contracted state often forming a circular wall in the lowest part of the sphincter which is mesogloea, rather strong to strong, and wholly separated from the circular muscles of the column. Margin distinct. Tentacles cylindrical or conical, sometimes more or less thickened at the outside of the base, hexamerously or irregularly arranged. Inner tentacles considerably stronger than the outer ones. Number of tentacles fewer than that of the mesenteries. Longitudinal muscles of tentacles and radial muscles of oral disc ectodermal. Actinopharynx with few longitudinal ridges. Siphonoglyphs 1-3, often situated perpendicularly to the longitudinal axis of the pedal disc. Mesenteries hexamerously or irregularly arranged, 1-3 pairs of directives. At least 6 pairs of mesenteries perfect, often more, sometimes there are pairs of mesenteries, one mesentery of which is perfect, its partner imperfect. Retractors of the mesenteries comparatively weak, apart from those of the directives. Mesenteries of the first order and at least the stronger of the other fertile, the directives usually excepted. Acontia mostly short but broad, their nematocysts basitrichs. Oidum: spirocysts, basitrichs, microbasal *p*-mastigophors.

In my papers (1925, 1928 a) I have discussed the Amphianthids, so that there is no need to repeat this discussion in all details here. I have pointed out that the genus *Stephanauge* (type *uerilis*) probably is a distinct genus separated from *Amphianthus* probably by a richer development of mesenteries at the margin than at the limbus. The examination of *uerilis* supports such an organisation though the difference between the number of mesenteries at the distal end and that of the proximal is not so conspicuous as in *Stephanauge bulbosa* (CARLGRÉN 1928 a p. 222). On the contrary the mesenteries at the limbus are more numerous than those of the margin in *Amphianthus* or, in other words, the mesenteries, at least those of the younger cycles, grow from the base upwards in *Amphianthus*, from the distal end downwards or possibly

from both ends in *Stephanauge*. This difference is sufficient to my mind to separate the two genera though undoubtedly near related to each other.

The near relationship of *Gephyra* (*Gephyropsis*) with the Amphianthids HERTWIG (1882 p. 76) and ANDRES (1883 p. 382, 576) have already suggested. In fact the genus *Gephyra* is identical with *Amphianthus* and *Korenia* (CARLGRÉN 1928). Also the genus *Raphactis* is nothing but *Amphianthus*—I have examined the type *R. nitida* Verr. and also observed acontia here (CARLGRÉN 1931). As to the genus *Sycaanthus* it is more difficult to decide its identity with *Amphianthus* as the first description of the type *S. mirabilis* Verr. is very imperfect. ANDRES (1883 p. 584) suggested, probably rightly, it may be a Zoanthid. VERRILL's *mirabilis* of 1883 belongs to the genus *Isozoanthus* (CARLGRÉN 1921 p. 187), while his specimens of 1899 and 1922 probably are *Amphianthus* or possibly *Stephanauge*. Also the description of *Bunodes nitida* R. Hertw. suggests that we have to do with an *Amphianthus*. As a matter of fact, before reading the description I have stated the near relationship of this species to *Amphianthus* during my visit to London in 1897.

Amphianthus margaritaceus (Dan.).

Korenia margaritacea n. sp. DANIELSSEN 1890 p. 1 Pl. 1 fig. 1. Pl. 7 figs. 1-5.

Amphianthus (*Korenia*) *margaritaceus* (Dan.) CARLGRÉN 1928 p. 258, 300; 1932 p. 261; 1933 p. 32; CARLGRÉN in Broch 1936 p. 5; 1939 p. 11, 22.

Diagnosis: Pedal disc wide, with a cuticle. Column in the contracted state with or without a distinct circular wall, smooth in young specimens, in older provided with rounded mesogloea-papillae being small but numerous next to the tentacles and here, at least in contracted specimens, not distinctly arranged in rows; below these papillae a small smooth or somewhat tuberculated area, at the circular wall often large tubercles and further down smaller tubercles arranged in 15-18 longitudinal rows. The lowest part of the column smooth. Cinclides 1-1, sometimes indistinct or absent (?) in young specimens. Sphincter strong, broad, extended over almost the whole breadth of the mesogloea, in its upper and inner part reticular with rather large meshes, in its outer part alveolar. Tentacles about 60-70, conical, rather short, often somewhat thickened at the base of their abaxial side, inner tentacles about twice as long as the outer ones. Two siphonoglyphs, 6 pairs of perfect mesenteries; moreover, the mesenteries of the second cycle are often perfect in the uppermost part of the actinopharynx. Retractors fairly well developed on the strongest mesenteries, especially on the directives, parietobasilar and basilar muscles weak. All stronger mesenteries, except the directives, fertile. Nematocysts of the column partly about 14-28.2 × 3.5-4.2 μ , basitrichs, partly 17-18.3 × 4.2-5 μ , microbasal *p*-mastigophors, those of the tentacles 14.1-28.2 × 3.5-4.2 (5) μ , basitrichs (on the outer side of their base 18.3-24 × 4.2-49 μ , microbasal *p*-mastigophors, sometimes absent), those of the actinopharynx partly 28.2-36 × 3.5-4.2 μ , basitrichs, partly 27.5-31 × 3.5-5 μ , microbasal *p*-mastigophors, those of the filaments partly 18.3-24 × 2.8-4 μ , basitrichs, 22.6-29 (34) × 3.5-4.2 μ microbasal *p*-mastigophors, those of the acontia 31-16.5 × 4.2-5 (5.5) μ , basitrichs (13.4-15.5 × 2 μ , rare, basitrichs), spirocysts of the tentacles up to 55 × 7 μ .

Colour: Column and pedal disc pale rose red to almost white with mother-of-pearl lustre. Oral disc a darker rose red with a slightly darker annulus around the mouth. Tentacles somewhat darker rose-red than the oral disc (DANIELSSEN). The ectoderm of the column contains, in preserved state, a yellow brown pigment.

Dimensions: Column about 1 cm high, at the margin 1 cm, at the limbus 2.2-4 × 0.5 cm broad. Largest specimen (St. 26 Swed. exp.) in contracted preserved state: Largest breadth 2.2 × 1 cm, height 1 cm.

it supports my suggestion that the basitrichs of the Anthozoa have arisen from hoplotelic microbasic *b*-mastigophors (see CARL-GREN 1910 p. 54, 55).

Remarks: Possibly *Sagartia splendens* DAN. is identical with this species but, as the type of *S. splendens* is lost, it is difficult to decide. Therefore I prefer to regard the specimens above described as a new species.

Amphianthus mopseae (Dan.).

Stelidiactis mopseae n. sp. DANIELSSEN 1890 p. 17 Pl. 2 figs. 4, 5, Pl. 8 figs. 7-11.

? *Stelidiactis tubulariae* n. sp. DANIELSSEN 1890 p. 19 Pl. 2 figs. 6-7, Pl. 8 fig. 12.

Diagnosis: Pedal disc prolonged in the directive plane (always?) clasping the stalk of *Mopsea* or *Tubularia*. Column provided with 24 ridges (teste DANIELSSEN) without tubercles. Cinclides at least 2 from the directive compartments. Sphincter rather strong, alveolar, not stratified, not occupying the whole breadth of the mesogloea. Tentacles up to about 48, not thickened on their outside, short, conical. More than 6 pairs of mesenteries perfect. Two siphonoglyphs (always?). Mesenteries at the limbus more numerous than those at the margin. Retractors of the mesenteries, parietobasilar, and basilar muscles weak. All stronger mesenteries, including the directives, fertile. Nematocysts of the tentacles about $14 \times 1.5 \mu$ basitrichs, those of the actinopharynx 19-21 ($24 \times 2.8-3.5 \mu$) those of the filaments 18.3 $22.6 \times 2.8-3.5 \mu$, those of the acontia 30-36 $\times 4.2-1.5 (5) \mu$, basitrichs. Spirocysts of the tentacles up to 31 $\times 4.5 \mu$.

Colour: Column mother-of-pearl coloured, tinged faintly with rose red. Oral disc salmon red with somewhat darker rays issuing from the mouth. Tentacles a little lighter in colour than the oral disc (DANIELSSEN, *Stelidiactis mopseae*). Column as the former. Tentacles somewhat darker red with a bright red annulus round the base (DANIELSSEN *Stelidiactis tubulariae*).

Dimensions in contracted state: height 0.5-0.4 cm, breadth of the base about 0.9 \times 0.15, in expanded state: length of the pedal disc 3.4 cm, height of the column and breadth of the oral disc 1.2 cm (DANIELSSEN *S. mopseae*). *Stelidiactis tubulariae* in expanded state: Pedal disc 2.5, height of the column and breadth of the oral disc 2 cm (DANIELSSEN).

Occurrence: Norway: Westfjord 68 12' N, 15 10' E, 621 m. Bottom temp. 6.5° (Norw. N. Atl. Exped. St. 255 *S. mopseae*) 1 specimen: 64 18' N, 6 32' E, 283 m. Bottom temp. 6.9° (Norw. N. Atl. Exped. St. 79 *S. tubulariae*).

Exterior aspect: I have examined the single, not well preserved specimen of *S. mopseae*. The pedal disc was oblong and clasped a branch of *Mopsea borealis*. It is drawn out in the directive plane, provided with a cuticle and radially folded. According to DANIELSSEN the comparatively low column is provided with 24 ridges and between these deep furrows and in the furrows cinclides here and there but without any regularity. In the preserved state of the specimen there is hardly anything to see of these formations, only traces of ridges. In sections through a part of the column enclosing 5 perfect mesenteries I found at least 2, probably 3 cinclides arranged in a longitudinal row in the directive compartment—a slide with sections was a little damaged during the manipulations with the sections, so I cannot decide with certainty, if 2 or 3 cinclides were present. Whether there were cinclides in other compartments than in the directives is very doubtful. In order to confirm it, it would have been necessary to cut the most of the specimen, which I have given up, as only a single specimen was present. Probably no other cinclides than those issuing from the directive compartments were present as in other *Amphianthus*-species. It must be noted that

DANIELSSEN falsely stated that in *A. (Koreoia) motpuritaceus* the papillae were provided with a minute round opening (loop-hole) in the extremity (compare this species). The figure 8 Pl. 8 of *S. mopseae* in the work of DANIELSSEN is certainly very schematic, the supposed cinclides are e. g. here regularly arranged contrary to the statement in the text. The furrows of the lower part of the body do not correspond to the insertions of the mesenteries (see fig. 9 Pl. VIII in DANIELSSEN's work).

According to DANIELSSEN the number of tentacles was 24 in *A. mopseae*, 48 in *A. tubulariae*. Meanwhile I think that the tenta-

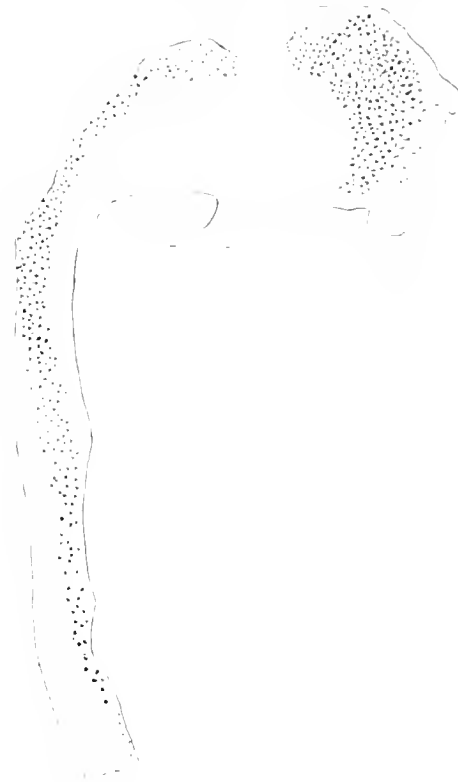


Fig. 63. *Amphianthus mopseae*. Sphincter.

cles of the former were more than 24, as in a small segment of the specimen I found three tentacles invaginated into three adjacent endocoels. The tentacles are short (conically acuminate and thick at the base, DANIELSSEN) and lack basal thickenings on the outside. The oral disc is round, strongly arcuate (DANIELSSEN), also in preserved state the mouth is situated on a cone. The actinopharynx is rather long and longitudinally sulcated. The two siphonoglyphs are distinct, situated in the longitudinal axis of the animal and provided with distinct aboral prolongations.

Anatomical description: The ectoderm of the column is of ordinary height to judge from small remaining fragments of it, the mesogloea very thick, especially in the lower part of the column, and contains cells poor in protoplasm. The cinclides, principally evaginations from the endoderm, are wide, conical towards the endoderm, the outer aperture considerably smaller. The sphincter is rather strong, mesogloea, not stratified, alveolar with small muscle meshes, perfectly differentiated from the endodermal muscles of the column, occupying only a part of the mesogloea, in its upper part situated nearer to the ectoderm than to the endoderm (fig. 63).

The ectoderm of the tentacles is very high, to judge from the invaginated tentacles. In maceration preparations I observed only some small nematocysts $11-17 \times 1.5-2 \mu$ in size, and spirocysts up to about $31 \times 1.5 \mu$. Moreover there were also some spool-like opaque capsules $11.3-11 \times 2.8-3.5 \mu$. The statements of the size of the stinging capsules are, however, uncertain as only fragments

of the tentacles. As the capsules were rather opaque or granular, I could not see their type, but I think that they were of the same type, as well as basitrichs present on the acontia. The former probably somewhat shorter than the latter. The longitudinal muscles of the oral disc were very weak. The ordinarily high ectoderm of the oral disc and of the longitudinal muscles, the mesogloea is thin and transparent.

Remarks: I cannot determine the exact number of the mesenteries, but they were probably about 48 pairs at the base. In almost a quarter of the specimens I counted 23 mesenteries at the limbus.

At the base of the mesenteries are considerably fewer and probably 18-24 pairs. DANIELSSON stated that only 6 pairs of mesenteries were perfect. In fact there were certainly also a second cycle of mesenteries attached to the actinopharynx, but whether all mesenteries of this cycle were perfect I cannot decide. The mesogloea of the mesenteries is rather thick, especially in their lowest part. The longitudinal muscles are weak forming distinct but weak retractor only in the reproductive region, the parietobasilar and basilar muscles weak. Oral and well developed marginal stomata are present. The acontia were packed together close by the directives in the sectioned piece. Also other mesenteries than the retractors seem to be provided with acontia. The nematocysts, basitrichs, of the acontia were very numerous. All stronger mesenteries and the directives were fertile, containing testes. DANIELSSON has drawn a figure (cf. PL. VIII) of the reproduction organs. The sign 2 shows distinctly a testis, though DANIELSSON has mistaken it. If the figure is correct, it seems as also an ovum were present. DANIELSSON described the reproduction organs as ovaries.

Remarks: I think that *A. tabulariae* is the same species as *A. islandicus*. The colour and the description of the species agree very well, but as no specimens of *tabulariae* were preserved, it is not possible to decide with certainty.

Amphianthus islandicus n. sp.

Description: Pedal disc wide. Column smooth, without tubercles. Cinclides 2 from the directive endocoel. Sphincter very strong consisting of very fine meshes (or single fibrillae), wholly separated from the general muscles of the column, occupying in its upper part almost the breadth of the mesogloea, diminishing downwards. Tentacles conical, up to about 100 in number, without longitudinal thickenings. Siphonoglyph broad, almost without longitudinal furrows. Mesenteries irregularly arranged, at the base about 100-170. Perfect mesenteries more than 6 pairs. One or two directives present. Retractors of the mesenteries very weak. Parietobasilar muscles weak. Acontia well developed. Nematocysts of the acontia: part 17-22 μ , 4-2.5 μ microbasal *p*-mastigophors?, part 8.5-11 μ , 1.5-2 μ , basitrichs, those of the acontia: part 19.5-19.7 μ , 3.5-5.5 μ microbasal *p*-mastigophors?, part 10-14 μ , 1.9-2.8 μ , basitrichs very sparse, those of the acontia: part 17-21 μ , 3.5-4 μ microbasal *p*-mastigophors, part 10-17 μ , 2-2.8 μ few basitrichs, those of the filamentous acontia: part 14.2-19.7 μ , 3.5-4.2 μ microbasal *p*-mastigophors, part 10-17 μ , 2-2.8 μ basitrichs, those of the acontia 23-28 μ , 3.5-4.2 μ . Siphonoglyph of tentacles up to 41 μ , 7 μ , 10 μ long.

Material: Type specimen, breadth of pedal disc 1.3-1.2 cm, length of siphonoglyph of column that of the smallest, breadth of column 1.2-1.1 cm.

Locality: 10° 15' N, 22° 20' W, 326-216 m (Thor. exped., 1952).

Remarks: Both specimens were contracted and the siphonoglyph was almost circular, the column

smooth, in its upper part provided with indistinct longitudinal furrows possibly caused by contraction. In the larger specimens there were 2 cinclides close to each other in the part corresponding to the single directive endocoel, the smaller specimen was possibly devoid of cinclides, at least they were not visible under a lens. The tentacles of the larger individual occupied about half of the oral disc and were about 100 in number. The actinopharynx was

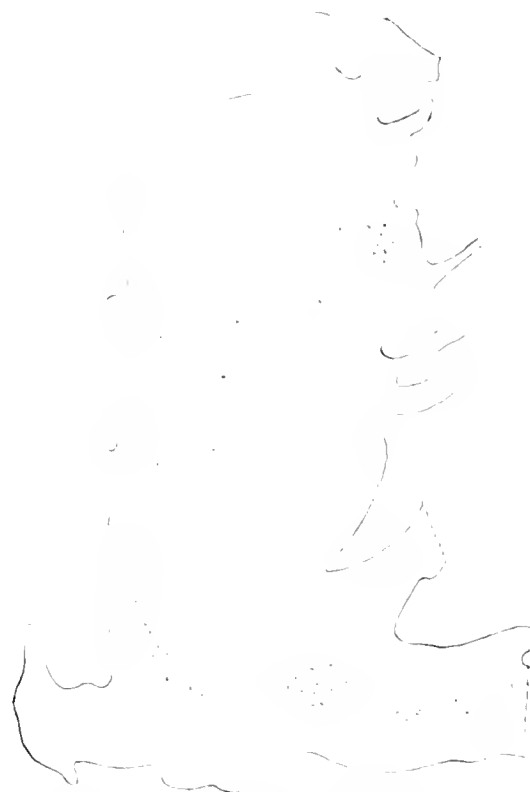


Fig. 61. *Amphianthus islandicus*, Sphincter.

provided with 16 longitudinal furrows in the larger specimen which had only one siphonoglyph. The smaller had probably two.

Anatomical description: The anatomy of this species agrees with other species of the genus *Amphianthus*. Fig. 61 shows a section of the sphincter. The muscle meshes are considerably finer than in *Amphianthus dohrnii*. The longer individual had 10 pairs and 3 single mesenteries perfect, among them a single directive pair. The perfect mesenteries of the smaller individual were 9¹/₂. Two directives were probably present here, I have, however, only dissected this individual. The longitudinal muscles of the mesenteries formed very weak retractors, the parietobasilar muscles too were also weak, the acontia thick.

The basitrichs were, apart from those of the acontia, few, so that the above named size of them is somewhat uncertain. Some of the larger nematocysts were exploded, their shaft about of some length as the capsule; the shaft was not drawn out in a thread, thus the nematocysts may have been amastigophors, but possibly the threads were broken off. However, an examination of living material is necessary to ascertain if amastigophors are present in the genus *Amphianthus*.

Remarks: The species is certainly nearly related to *A. dohrnii* and it may be possible that they belong to same species. *Dohrnii* has, according to ANDRÉ, up to 96 tentacles. I have found up to about 70 in specimens from Naples. The cinclides were commonly apparently 1-1 but 2 cinclides have been found in the part corresponding to the directives. The sphincter of *dohrnii* has, however, coarser meshes than *islandicus*, and the basitrichs of the acontia are somewhat larger in the former than in the latter.

***Amphianthus ingolfi* n. sp.**

Diagnosis: Pedal disc wide. Column smooth. 1 cinclis from each directive endocoel. Sphincter alveolar, strong with fine meshes which are situated far from each other in the strongly thickened uppermost part of the column. Tentacles about 48, considerably fewer than the mesenteries, not thickened on the aboral side.



Fig. 65. *Amphianthus ingolfi*. Sphincter.

1 siphonoglyph and 1 pair of directives (in the larger specimen) Mesenteries at the base about 116. Retractors of the mesenteries weak. Asexual reproduction by fragmentation (tearing). Nematocysts of the tentacles partly $15.5-26.8 \times 3-1.2 \mu$, microbasic p -mastigophors, partly $11.3-22.6 \times 2-3 \mu$, basitrichs, those of the actinopharynx partly $17.6-21 \times 3-3.5 \mu$, microbasic mastigophors, partly $21-25.1 \times 2.8-3 \mu$, basitrichs, those of the filaments partly $19.7-21 \times 2.8-3 \mu$, microbasic p -mastigophors, partly $10-16.9 \times 1.5-2 \mu$, basitrichs, those of the acontia $35.2-16.5 \times 1.2-5.5 \mu$, basitrichs. Spirocysts of tentacles up to $48 \times 7 \mu$.

Colour unknown.

Size of the larger specimen in the contracted state: breadth of pedal disc 0.7×0.5 cm, height of the column $0.15-0.5$ cm, that of the smaller: breadth of pedal disc 0.7×0.6 cm, height of column 0.1 cm, breadth of distal part 0.35 cm.

Occurrence: $60^{\circ} 37' N$, $27^{\circ} 52' W$, 1595 m, bottom temp. 1.5° . Ingolf exped. St. 78, 2 specimens.

Exterior aspect: Both examples were of same appearance. The smaller example had a very wide pedal disc, the body of the larger example was of about same breadth. The larger specimen had 1 cinclis, on the smaller one I could not find any cinclis under the lens. The tentacles were about 48 in the larger individual. The larger specimen was provided with a bud-like swelling at the base indicating reproduction by fragmentation.

Anatomical description: The interior of the smaller individual was not well preserved so that the description has been made from the larger specimen. A section of the sphincter is drawn in fig. 65. There were about 48 mesenteries in the upper part of the body, at the base about 116. Probably only 1 directive pair was present, 7 pairs and 2 single mesenteries were perfect. The retractors were weak. I have measured the basitrichs of the acontia in both specimens. In the smaller example the length varied between 35.2 and 16.5μ , in the larger one between 36.7 and 15μ . The average size of 52 measured nematocysts, 26 of each specimen, was 11.22μ .

***Amphianthus verruculatus* n. sp.**

Diagnosis: Pedal disc wide, often embracing Hydroids or skeletons of Octocorals. Column smooth in its lower part, in its uppermost provided with more or less distinct rows of very small tubercles (sometimes absent?). Probably only one cinclis from the directive compartment. Sphincter very strong, alveolar-reticular to reticular with rather large muscle meshes, occupying in its upper part almost the whole breadth of the mesogloea, diminishing downward. Tentacles between 40 and 50, without aboral swellings. Number of siphonoglyphs and pairs of directives one or two. Considerably more mesenteries than tentacles. Nematocysts of the column $17-21 \times$ about 3.5μ , microbasic p -mastigophors, those of the tentacles partly $(14) 17-21 \times 3.5 \mu$, microbasic p -mastigophors, partly $11.3-18.3 \times 3.5-4 \mu$, basitrichs, those of the actinopharynx partly $19.7-21 \times 3.5 \mu$, microbasic p -mastigophors, partly $15.5-21 \times$ about 3μ , basitrichs, those of the filaments partly $15.5-21 \times 3-4 \mu$, microbasic p -mastigophors partly $12-22 \times 2.5-3 \mu$, basitrichs, those of the acontia $29.6-13.7 \times 1.2-5 \mu$, basitrichs. Spirocysts of tentacles up to $37-50 \times 7$ about 8μ .

Colour unknown.

Size of the largest specimen from St. 35: the very expanded basal disc about 1 cm broad, height of the body 0.6 cm. All other specimens were very small: largest length of pedal disc $0.4-0.6$ cm, height of the body $0.3-0.15$ cm.

Occurrence: $66^{\circ} 35' N$, $56^{\circ} 38' W$, 599 m, bottom temp. 3.9° . Ingolf exped. St. 32, 1 specimen; $66^{\circ} 22' N$, $57^{\circ} 16' W$, 686 m, Tjalle exped. 1909, 1 specimen; $65^{\circ} 16' N$, $55^{\circ} 05' W$, 682 m, bottom temp. 3.6° . Ingolf exped. St. 35, 2 specimens; $64^{\circ} 54' N$, $55^{\circ} 10' W$, 710 m, bottom temp. 3.8° . Ingolf exped. St. 27, 1 specimen; $62^{\circ} 06' N$, $22^{\circ} 30' W$, 1587 m, bottom temp. 3.1° . Ingolf exped. St. 68, 1 specimen.

$70^{\circ} 53' N$, $54^{\circ} 03' W$, about 700 m. Bottom temp. 1.5° . Godthaab exped. St. 113

Exterior aspect: The column was smooth, apart from its uppermost part which had more or less distinct, longitudinal rows of very small tubercles only visible under a strong lens. The rows of tubercles were most distinct in the smaller example from St. 35 and in that taken by the "Tjalle". In the larger example dredged at St. 35 no tubercles were visible, but, as the uppermost part of it was very contracted it is possible that this specimen also has had them. As the sphincter was of same appearance as those of the other specimens, and the size of the nematocysts

of the tentacles were two or a little larger than those of the same species. The diameter of the mesenteries at the base of the tentacles was 40-50 μ . Owing to the smallness of the specimens the structure of the inner organs I cannot

They varied between 31 and 36.7, the nematocysts of the largest individual from 35 to 43.7.

Remarks.—I have with some hesitation referred the specimen from station 143 to *erruculatus*. The nematocysts agreed fairly well with this species but were somewhat longer in the column, tentacles and actinopharynx, but the specimen was also consider-



Fig. 66.

Anchithus erruculatus. Sphincter, that in 67 figured from the Godthaab station 143.



Fig. 67.

ably larger (the column was 1 cm high and the base up to 1.7 cm broad). Also the sphincter (fig. 65) showed same appearance but was more extended, probably in correspondence with a different state of contraction. The nematocysts of the column were (15) 18-23 \cdot 3.5-4 μ , those of the tentacles 19-25 \cdot about 2.8 μ , those of the actinopharynx partly 25-29 \cdot 1.5 μ , microbasic *p*-mastigophors, partly 25-29 \cdot about 3 μ , basitrichs, those of the acontia 33-37 \cdot about 4 μ , basitrichs.

Also the largest specimen from station 143, that I have examined the structure of the tentacles from, that taken at station 27 (Fig. 65, text, 66) I have drawn the sphincter (fig. 65) from station 32 (fig. 66). The lowermost part of the tentacles is microalveolar. In other specimens the structure is reticular but the appearance of it was similar to that of the nematocysts of the acontia (fig. 65, text, 66). The structure of the sphincter in the specimen from St. 32,

Family Metridiidae.

Acontia the acontia of which are provided with microbasal *b*-mastigophors and microbasal amastigophors, the latter sometimes very rare or absent. Sphincter mesogloal. Mesenteries not divided into macro- and microemes.

The separation of the genus *Metridium* from the family Hormathiidae is undoubtedly necessary. The genus has very few characters of important ones only the mesogloal sphincter which, however, has another situation in *Metridium*, and the usually 6 perfect pairs of mesenteries—in common with this family. Already the types of the nematocysts are very different. PAX (1936 p. 103) places *Metridium* together with *Hormathia* and other Hormathiids and writes "Wenn aber die Metridiidae als eigene Familie aufrecht erhalten werden sollen, so muss sich doch ein einziges Merkmal angeben lassen durch das man sie mit Sicherheit von den Hormathiidae unterscheiden kann. Das ist tatsächlich nicht der Fall." It is to be regretted that PAX had so little knowledge of the organization of the Hormathiidae. But it is too much to expect of an author who has determined specimens of *Hormathia* as *Paranthous* (see CARLGRÉN 1927 p. 88, 89).

Genus *Metridium* Oken.

Metridiidae with wide pedal disc. Column smooth, divisible into scapus and capitulum, the former ending distally in a collar. Cinclides scattered on the scapus. Sphincter mesogloal, not situated close to the tentacles but at the uppermost part of the scapus. Tentacles short and delicate, extraordinarily numerous in large individuals. Longitudinal muscles of tentacles ectodermal, radial muscles of oral disc ectodermal or meso-ectodermal. Oral disc lobed in large races. Siphonoglyphs and directive pairs up to 3. Typically 6 perfect and sterile pairs of mesenteries, but especially in small races, and in individuals arisen from asexual reproduction, there are more, though not many, perfect pairs and some of the 6 primaries may be fertile. More mesenteries at the margin than at the limbus. Retractors of the mesenteries diffuse to more concentrated. Cnidom: spirocysts, microbasal *b*-mastigophors, microbasal *p*-mastigophors, microbasal amastigophors, basitrichs, Spirocysts small.

Metridium senile (L.).

Metridium dianthus (ELL.) APPELLÖF 1905 p. 63, 83, 86, 105; GRIEG 1913 p. 143; CARLGRÉN 1930 p. 3; BROCH 1927 p. 6.

Metridium senile (L.) CARLGRÉN 1933 p. 281; 1939 p. 8; RAWLINSON 1934 p. 901; STEPHENSON 1935 p. 261 Pl. XV fig. 1, Pl. XVI figs. 1-2, Pl. XXV figs. 1-2, Pl. XXVI figs. 1-2, Pl. XXVII figs. 1-2, textfigs. 7 (13) 34 (A-C) 74, 82; PAX 1936 p. 108 figs. 66, 67.

Metridium senile var. *finbriatum* (VERT.) CARLGRÉN 1934 p. 353; 1936 p. 23. For further references see CARLGRÉN 1893 p. 102, 1928 p. 298, ANDRES 1883 p. 345 and STEPHENSON 1935 p. 267.

Pedal disc wide. Column usually pillarlike. Sphincter usually alveolar at least with tendency to longitudinal stratification. Tentacles up to many hundred, usually short, sometimes longer. Typically 6 pairs of perfect mesenteries but in some races up to 15. Reproduction by fragmentation (tearing) from the limbus, possibly by longitudinal fission too. Nematocysts of the column partly 9.5-30 × 3.5 (1) μ , basitrichs, partly 11-35 × 3-5 μ , microbasal amastigophors, those of the tentacles partly 15.5-36 × 3.5-5 μ , microbasal amastigophors, partly 12-31 × about 2 μ , basitrichs, those of the actinopharynx partly 17-16 (53) × 3-5 μ , microbasal *p*-mastigophors, partly 19.7-13 × 2.5-3 μ , basitrichs, those of the filaments partly 10-14 × 3-3.5 μ , partly 21-48 × 4-5.5 (6) μ , both microbasal *p*-mastigophors, partly 15.5-24 × 2 μ , very rare, basitrichs, those of the acontia partly 33-80 × 3-5.5 μ , microbasal *b*-masti-

gophors, partly 32-89 × 3-6.5 (8) μ , microbasal amastigophors, the latter sometimes absent or almost so. Spirocysts up to about 29 × 3.5 μ .

Colour very variable: Column red-brown, flesh-coloured, olive-brown, salmon-coloured, pale orange-coloured, cream or pure white. Generally the disc and the tentacles are of same colour as the column, the latter often provided with a transverse bar some way above their base. The lips are white or cream, often reddish or orange. The pale orange and especially the pure white forms occur in Bohuslän principally in deeper waters.

Size up to 19 cm long and 10-11 cm broad in well preserved state (var. *finbriatum*).

Occurrence: West Greenland: Ikamiut (Lohmann); Egedesminde (Traustedt); Holsteinsborg 20 fms. (Holm), 22-35 m (Godthaab exp.), (Bernberg, Traustedt); Holsteinsborg harbour (Ingolf exp.); the district of Holsteinsborg (Bernberg); Ikertokfjord; head of Kapisigdlík fjord W. of Laxeely 10-0 m (Dana exp.).

Arctic coast of America to Cape Cod (teste PARKER); from Long Island Sound and Northern New Jersey to Bay of Fundy, Gulf of St. Lawrence (teste VERRILL); New Haven, Newport; New York; Chaleur Bay (Johansen); New Foundland (Verkrüzen); Grand Manan (U. States Fish Com.).

Iceland: Dýrafjörður, 3 localities, one of them 15 m. (Ingolf exp.) (Lundbeck), 8 m (Koch); Isafjörður (Caroe, Ingolf exp.); Isafjardardjup 9-28 m; Skötufjörður 9-28 m; Steingrimsfjörður; Skagaströnd 70-100 m; Eyjafjörður 22-38 m (Möller); S. of Hrísey; Eyjafjörður 17-20 fms. (Beskytteren exp. 1903); Akureyri, littoral; Berufjörður (Torrell); S. of Eystra Horn, 61° 17' N, 14° 11' W, 75 m (Michael Sars exp. 1900) 10-15 $\frac{1}{2}$ miles S.S.W. of Eystra Horn; Heimaey, Vestmannaeyar from the ventricle of a cod-fish; 61° 05' N, 22° 22' W, 45 m; Hafnarfjörður 7.5-13 m; Reykjavík; Hvalmúsvík; Hvalfjörður 0-20 m.

Faroe Islands (Møller), east side of Klaksvig (Bordo), Vestmannaasund S. of Egilsnes (the small individuals not determined (CARLGRÉN 1930 p. 11) were certainly *M. senile*); Orkney and Shetland Isls. (teste STEPHENSON and NORMAN), North Sea: 55° 13' N, 3° 15' E, 30 fms. (Dana exp. 1922); 54° 15' N, (Dana exp. 1922); Helgoland (Carlgrén); Büsum (teste PAX); coast of Schleswig-Holstein (teste MÖBIUS); west coast of Jutland: 36 miles N. to W. $\frac{3}{4}$ W. of Horns Rev lightship, 10 m (Thor exp.); Jutland Bank, 60-130 fms. (Uddstrom); Hauvrighuk, Ringkjöbing fjord (Johansen); west of the British Isles; English Channel (teste Gosse and STEPHENSON); north west coast of France to Rade du Croisic, Plerac (teste FISCHER); the Mediterranean (temporary?, teste PAX).

Murman Coast, W. off Kamin Peninsula (teste PAX), Kildin Isl., relict sea of Mogilnoje (teste PAX); Kolafjord (Avernzell, Derjugin), Norway: Öksfjord (midertidal); Hasvik; Soroy; Grotøy; Rost; Terningen, Verrafjord; Titran; Mastad, Veroy, Flatvadsund, Florø (Dons); Ulfsfjord, Kjosén; Kvenangen, 90 m (Nordgaard), Tromsø, 20-30 fms. (teste Sars), 50 m; Balsfjord, 70 fms. (Kjaer); Foldenfjord, Namdalen (Collett); Trondheimfjord (Nordgaard, Dons), Hsvika, Beian (Kjaer), Vaagsfjord (teste GRIEG); Bergen (teste Sars); Solsvig, Manger, 20 fms., Florø (Sars), Godosund, Espevaer (Bidenkap).

Skagerak: Lindesnes, Korshamn, littoral, Kjøpsø, 20-0 m (teste Broch); Oslofjord (Esmareck); Drøbak (Broch and others); Haunholm (Carlgrén); N.W. of Hirtshals, 16 fms. (Thor exp.); Bohuslän on many localities.

Linnfjord (Mortensen, Carlgrén and others); Tisted Brodning, 11-12 m (Biol. Station).

Kattegat: Frederikshavn (Carlgrén, Kramp and others);

1) *M. (M.)* *puberulum* (Göteborg). Rarities found in the Scania region (Göteborg), extent about 57° 03' N., 13° 12' E. (Linné), 56° 48' N., 12° 20' E., (Müller), 56° 45' N., 13° 15' E. (Ström), 57° 02', 89', 79', 43' 5" N., 14° W. (Aronson), 27° 15' (Thorén), 57° 15' N., 14° W. (Lindström), 23 m. S. of Helsingør (H. Jørgensen), 21 m. Lönneberg (S. of Helsingør), 25 Adsgårde, Krampø, Landskrona (Lindström), 20 m. S. of Knäbek (Lönneberg), 57° 15' N., 14° W. (Krampø), 17 miles S.W. of Venø (S. of Århus), 60 miles S. of Ukkasharpen, S. of

1) *M. (M.)* *puberulum* (Mortenson, Lurken), between Ström and Malmø (Lindström), S. E. of Flenø (Petersen), 57° 15' N., 14° E. (Forsberg), Bering Sea (Michaelson), 57° N. (Northrop), North Sea (low water) (E.W. Nelson), Alaska (C. H. Hart), 1932, Sitka, Whale Isl. near Sitka, St. Michael, Yakutat, Koonak, (to St. Thomas), Puget Sound, Nanaimo, Metchikan, Fort Vancouver, Kincaid, Victoria, British Columbia; San Francisco, Oakland, harbour, Kamchatka, Petropaulowsk, Avacha Bay, Avanton Bay (Swedish Kamchatka exp. 1920-21); Japan, No. 196, Hokkaido, (see WASSILIEFF).

There are several varieties or races of *Metridium scabrum*. At present we can distinguish 4 or 5, between some of them intermediate forms have been noted.

1) var. *puberulum* and *puberulum*, I have described the form and the locality of this form in 1896. It is characterized by the large tentacles having a strongly lobed oral disc and no retractors, rare anastigophors in their acontia. In small tentacles, however, the disc is not lobed and the anastigophors are present in the acontia. The nematocysts in larger individuals are larger in size. The sphincter is alveolar and shows at least a tendency to longitudinal stratification. The nematocysts of a small

individual about 1 cm long and 0.5-0.6 cm broad from Bohuslan were in the scapus partly 10-15.5-15-3 μ , basitrichs, partly 17-19.7-3.5-4.2 μ , microbasic anastigophors, those of the tentacles partly 15.5-25-3.5-4.2 μ , microbasic anastigophors, partly 15.5-18.3-2.5-2.8 μ , basitrichs, those of the actinopharynx partly 19.7-25.4-4-4.5 (5) μ , microbasic *p*-mastigophors, partly 19.7-28.8-2.5-2.8 μ , those of the filaments partly 21-24-4.5-5.5 μ , partly 11.3-11-3-3.5 μ , both microbasic *p*-mastigophors, those of the acontia partly 39.5-48-3-3.5 μ , microbasic *b*-mastigophors, partly 38-46.5-4.2-5.6 μ , microbasic anastigophors. The spirocysts of tentacles were up to 19.7-3.5-4.2 μ .

2) var. *puberulum*. It agrees in its exterior with *dianthus* but grows considerably larger than this variety and its acontia are always provided with anastigophors. The nematocysts are larger than those of *dianthus* and reach the maximum of size in *puberulum*.

3) var. *pallidum*, a smaller variety than *dianthus*, usually not lobed. The sphincter is reticular and the cinclides seem to be predominantly endocoelic, in contradistinction to those of *dianthus* in which they are generally exocoelic. The tentacles are larger than those of *dianthus*. The size of nematocysts of this variety is unknown. Intermediate forms between *pallidum* and *dianthus* occur (STEPHENSON 1935).

4) a dwarf race described by Miss RAWLINSON 1931 from Dingle. A small but fertile form. The oral disc is rarely distinctly lobed. The acontia are provided with anastigophors. The nematocysts agree fairly well with those of small young specimens of *dianthus*.

5) a small form not described in detail by PAX (1911) from the relict sea Magilnoje. The variety *pallidum* is known from the British Isles and France, and Helgoland, the Dingle race from England, the variety *puberulum* from Bering Sea and the Northern Pacific. Probably a closer examination of the species on the Scandinavian coast will show that different forms exist also there. See also STEPHENSON'S description of *Metridium* 1935.

Appendix.

Family Edwardsiidae.

Edwardsia norvegica n. sp.

Diameter 2 cm. Physa well developed, capable of involution. Column with 10-12 conical goral. Nemathybones small, numer-



Fig. 1. Cross-section of the nemathybones.

ous in the lower part of the scapus, fewer in the upper. Periderm thick in the lower part of the scapus, distally thinner. Tentacles 16. Retractors of the macrocemes concentrated in the upper part of the gonad region with at most 20 folds, as a rule richly ramified. Parietal muscles well developed, now elongated, now more concentrated, with high, somewhat branched folds. The outer lamellar part of the macrocemes attached close to the outside of the retractors. The expansion of the parietal muscles on the column of ordinary type. Nematocysts of the nemathybones 47-58-34- about 2.5 μ (numerous) basitrichs, those of the tentacles 29-34- about 2.5 μ (numerous) basitrichs, those of the actinopharynx partly 11-53- about 3 μ , microbasic *p*-mastigophors, partly 22-24-2.5 μ , basitrichs, spirocysts of the tentacles 13-2-25-3.5 μ .

Colour unknown.

Dimensions in somewhat contracted state: length 2 cm, largest breadth 0.4 cm.

Occurrence. Norway, Trondheimfjord, Galgenes 250-300 m ("Gunnerus" 1, 1925) 1 specimen.

Exterior aspect. The physa was well developed and completely drawn in, so that it is evidently capable of involution. The uppermost part of the column was a little invaginated, the distal ends of some tentacles were, however, visible. The column was somewhat polygonal, especially conspicuously in the contracted basal

and distal ends. The nemathybomes were small, in the lower part of the scapus numerous and scattered over the whole surface (fig. 68, 69), in the upper part fewer and mostly placed in the middle-line between the insertions of the mesenteries, sometimes, however, situated nearer the insertions. Small fragments of foreign bodies were stuck to the cuticle of the scapus.

Isoedwardsia nidarosiensis n. sp.

Diagnosis: Ectoderm and cuticle of the scapus rather thin, the latter very incrustated. Nemathybomes few, small, scattered. Tentacles 16. Retractors of the macrocnemes rather strong, in the upper part of the reproductive region with about 15-20 folds

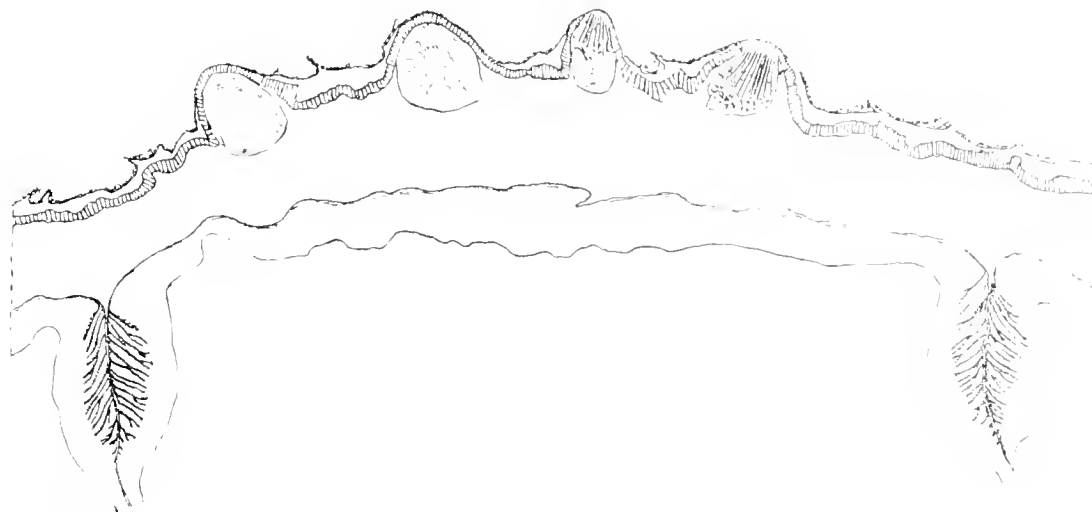


Fig. 69. *Edwardsia norvegica*. Section of the column between two mesenteries in the lower part of the body. 4 nemathybomes are visible.

The tentacles were 16 and of typical appearance as also was the actinopharynx.

Anatomical description: The ectoderm of the physa was high, that of the scapus thinner. The cuticle was thin in the upper part of the scapus and outside the nemathybomes, in the lower part thick, its outer parts were easily deciduous. The ectoderm of the tentacles was of the usual appearance.

The retractors of the macrocnemes are not strong, their folds high and richly branched, but few in number. In the upper part of

ramified especially in their outer parts. Lamellar part of the macrocnemes issuing not far from the middle of the retractors. Parietal muscles very strong in the upper part of the gonad region, their expansion on the column of ordinary type. Nematocysts of the nemathybomes about $62-67 \cdot 2.5-3 \mu$, microbasal *b*-mastigophors (thinner type), those of the tentacles $31-45 \cdot$ about 2.8μ , basitrichs, common, those of the actinopharynx partly $60-71.5 \cdot 3.5 \mu$, basitrichs, very numerous, partly $26-31 \cdot 2.5-2.8 \mu$, basitrichs, sparse, partly $41-49.5 \cdot 5.6 \mu$, microbasal *p*-mastigophors, very sparse, those of the filaments partly $28-45 \cdot 1.5-6.5 \mu$, microbasal *p*-mastigophors, partly $20-29.5 \cdot 2.5-2.8 \mu$, partly $27-42.5 \cdot 1.2 \mu$, both basitrichs. Spirocysts of the tentacles $11-24 \cdot 2-3 \mu$.

Cnidom: spirocysts, basitrichs, microbasal *b*-mastigophors, microbasal *p*-mastigophors.

Colour in formalin two days after preservation: Scapus dirty gray, capitulum, tentacles, oral disc and actinopharynx orange-coloured.

Dimensions in strongly contracted state: length 1 cm, largest breadth of the proximal part of the scapus 0.5 cm, that of the capitular region 0.3 cm, length of the tentacles about 0.2 cm.

Occurrence: Norway, Trondhømfjord, Hestdalen, Selvikstrand 125-150 m. ("Gunnerns" 1921) 1 specimen.

Exterior aspect. No distinct physa. Although in a very small area in the middle of the proximal end of the body the mesogloea was considerably thinner than in the other parts of the scapus, I think we are not dealing with a real physa here. The ectoderm has a structure like that of the other parts of the scapus to judge from remaining fragments of the ectoderm provided with a cuticle. Because there were nemathybomes present in the proximal end, though still more sparse than in other parts of the scapus, the species may be an *Isoedwardsia*. The proximal end was rounded, the proximal part bulbous swollen, the upper part of the scapus and the capitulum somewhat polygonal. Owing to contraction the scapus was strongly and irregularly wrinkled, giving it a granular appearance. The cuticle was rather thin but strongly incrustated with foreign bodies, spicula, sand grains and detritus. The nemathybomes were small, few and scattered and only visible in sections.

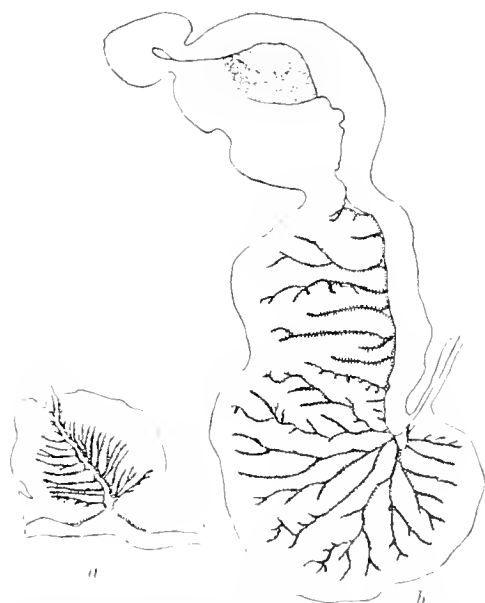


Fig. 70. *Edwardsia norvegica*. Parietal muscle (a) and retractor (b) of mesentery in the upper part of the fertile region.

the gonad region there were about 20 folds (fig. 70 b) in the lower part fewer. As to the parietal muscles (fig. 70 a) compare the diagnosis.

Remarks: I have not been able to identify this species with any before known.

Fig. 71. *Andwakiia* sp. n. Retractor (fig. 71) and parietal muscle of a mesentery in the upper part of the scapus (fig. 72).

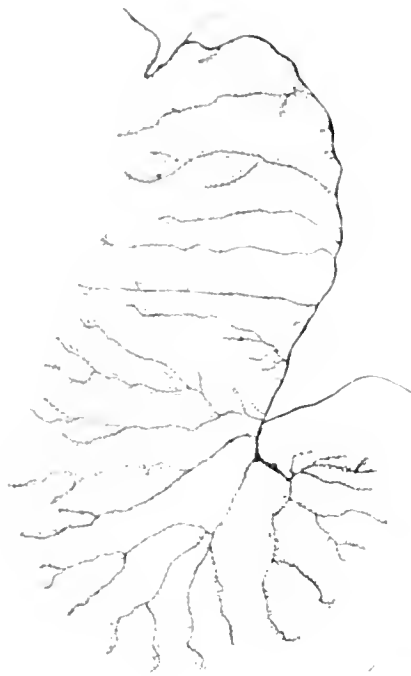


Fig. 71.

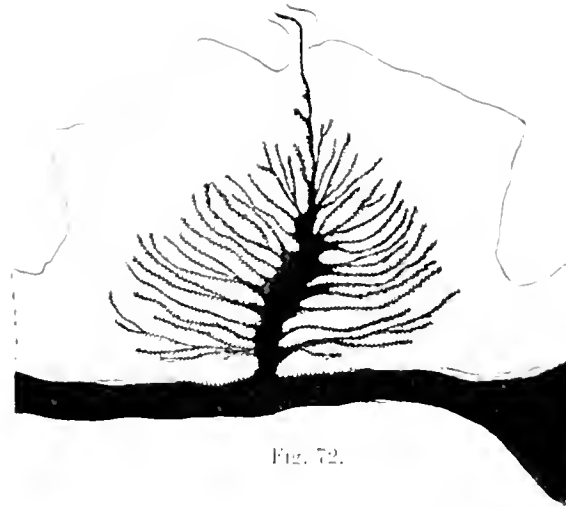


Fig. 72.

Fig. 71. *Andwakiia* sp. n. Retractor (fig. 71) and parietal muscle of a mesentery in the upper part of the scapus (fig. 72).

Andwakiia sp. n. For the anatomical examination I used a part of the proximal end and a piece of the scapus, tentacles, oral disc and actinopharynx and the upper part of the scapus.

The oral disc of the scapus was low also in the proximal part and provided with a cuticle. The nemathyboes were numerous, which were broader in the basal than in the apical part. The mesoglea was stratified owing to contractile elements in its outer parts and contained rather large nematocysts. The endodermal circular muscles were provided with high folds, somewhat weaker in the capsule. The ectoderm of the scapulus was more than three times thicker than that of the scapus, its nematocysts rather numerous, partly $12-15 \times 3.5-4 \mu$ and $17-22 \times$ about 2μ in size.

The high ectoderm of the apex of the tentacles had extraordinarily numerous nematocysts (size compare diagnosis). The ectodermal longitudinal muscles of the tentacles were rather strong.

In the high ridges of the actinopharynx there were extraordinarily numerous large nematocysts and sparse smaller ones.

The retractors of the macroemes were rather strong, forming about 15-20 primary folds in the upper part of the scapus. Especially in the outer part of the retractors the folds were ramified (fig. 71). The parietal muscles were strong, especially in comparison with the pennons, and provided with numerous folds (fig. 72). In transverse sections they were mostly triangular in the upper part of the scapus, sometimes more elongated in a radial direction. The expansion of the parietal muscles on the column was as usual. The outer lamellar part of the macroemes was attached not far from the middle of the pennons. The specimen was a male with well developed testes.

Family Andwakiidae.

Andwakiia parva Carlgren

Pl. IV fig. 7, Pl. VI fig. 7.

Andwakiia parva n. sp. Carlgren 1910, p. 28, figs. VI 6-8.

Andwakiia sp. n. Scapus and scapulus. The scapus attached to the substratum, scapulus provided with a thick cuticle. Sphincter muscles and muscle meshes. Tentacles 12-15 \times 24-30 μ , short. Longitudinal muscles of the tentacles fan-like arranged. One pair of tentacles regenerate. Mesenteries usually 12-15 \times 3.5-4 μ in the proximal part and 17-22 \times about 2μ in the distal part. Mesenteries not branched. Macroemes, mastigophors, partly 12-15 \times 3.5-4 μ , partly 17-22 \times about 2μ , both basitrichs, those of the filaments partly 11-15 \times 4-4.5 μ , microbasics μ -mastigophors, partly 8-12 \times 1-1.5 μ , basi-

provided with retractors, ciliated tracts of the filaments (and reproductive organs?). Retractors of the stronger mesenteries concentrated, with few high, somewhat branched folds. Parietal muscles well developed, in their upper part fan like, in their lower ones more radially elongated. Nematocysts of the scapulus partly 12-15 \times 3.5-4 μ , microbasics amastigophors, partly 12-13 \times 3-4 μ , almost 3.5 μ , partly 9-10 \times 2 μ , both basitrichs, those of the tentacles partly 11-17 \times over 2.5 μ , microbasics amastigophors, partly 16-20 \times about 2.5 μ , basitrichs, those of the actinopharynx 18-22 \times about 1 μ , probably microbasics μ -mastigophors, partly 17-22 \times about 2.5 μ , basitrichs, those of the filaments partly 11-15 \times 4-4.5 μ , microbasics μ -mastigophors, partly 8-12 \times 1-1.5 μ , basi-

trichs, those of the acontia partly $24-31 \times 3.5 \mu$, microbasic amastigophors, partly $22-23 \times 2.5 \mu$, basitrichs. Owing to the irregularities of the arrangement of the mesenteries and siphonoglyphs the species probably propagates itself asexually.

Colour: Scapus grayish brown or grayish, scapulus orange-coloured, tentacles at the base orange, two white spots more or



Fig. 73. *Andwakia parva*. Section of the scapus showing a tenaculum.

less transversally situated, sometimes joined at about the middle of the tentacles or a little nearer their bases, tentacles for the rest uncoloured, opaque, sometimes a little powdered with white, in contracted state their endoderm seems darker. Oral disc orange-coloured, especially at the insertions of the mesenteries. Actinopharynx pale orange. In smaller specimens the colour is often pale and the spots on the tentacles sometimes absent.

Dimensions: A small specimen, not exceeding 1 cm in length and 0.3 cm in breadth.

Occurrence: Sweden: Bohuslän, Väderöarne in the vicinity of the *Lophohelia*-reef at a depth of 60-70 m together with *Octineon succicum* on shells and small stones, numerous specimens (CARL-GREN and GUSTAFSSON 1932-30).

Exterior aspect: The species is cylindrical in the expanded state, when contracted hemisphaeric (Pl. IV fig. 7, Pl. VI fig. 7). The column is divisible into physa, scapus and scapulus. The physa is, however, seldom visible, as it is closely attached to the substratum on which the species lives, but when the animal releases itself from the substratum the basal part of the body forms a distinct physa which it uses for locomotion. The scapus is provided with a cuticle and numerous tenaculi of same structure as those of *Phellia gausapata*. To the tenaculi grains of sand are often fastened. The scapulus is rather short, the tentacles thin and rather short, in number never more than 24 (in 5 specimens I counted 19-22 (I:19, 3:20, I:22), and arranged in two cycles. The oral disc is inconsiderable, the actinopharynx longitudinally sulcated. The siphonoglyphs are very little differentiated, in number two or one.

Anatomical description: The ectoderm of the scapus is rather high with a cuticle, which is very thick in the tenaculi. The tenaculi seem to be of the same appearance as in *Phellia gausapata*, although sometimes the chitinized cells, connecting the mesogloea with the cuticle, are few (fig. 73). The mesogloea of the tenaculi is thickened. The ectoderm of the scapulus is higher than in the scapus, the mesogloea sphincter strong, mostly consisting of large muscle-meshes and wholly separated from the endodermal muscles of the column: more than half of the sphincter (fig. 74) is situated in the scapus. The ectodermal muscles of the tentacles are arranged like a palisade and very little ramified. The ectoderm of the actinopharynx is high, that of the indistinct siphonoglyphs has somewhat longer cilia than in other parts of the actinopharynx.

The mesenteries are more or less irregularly arranged, 6 specimens sectioned showed the following arrangement of the mesenteries: d: directive pairs, p: perfect mesenteries, i: imperfect mesenteries:



Fig. 74. *Andwakia parva*. Sphincter.

1.	^{dd} pp, u, ^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pp, ii	= 20	mesenteries: 2 pairs of directives.
2.	^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pp, ii, ii	= 22	: 2
3.	^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pp, ii	= 20	: 1 pair -
4.	^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pp, ii, ii	= 24	: 1 -
5.	^{dd} pp, ii, ^{dd} pp, ii, ^{dd} ip, ii, ^{dd} pp, ii, ^{dd} pi, ii	= 20	: 1 -
6.	^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pp, ii, ^{dd} pi, ii, ^{dd} ip, ii	= 20	: 1 -

the distal part of the retractors, *pp* formed the parietal muscle (the same side), *p* in the middle of the mesentery, *p* was



FIG. 76. *Andreakia parva*. Transverse section of a perfect mesentery in the region of the endoglandular tract.

FIG. 77. Retractor, perhaps not perfect but provided with acentia.

The arrangement of mesenteries agrees with that of the tentacles. *Andreakia parva* does not develop mesenteries earlier in the life cycle than the distal part. The retractors, present only on the perfect mesenteries, were concentrated with rather few, somewhat



FIG. 77. *Andreakia parva*. Transverse section of a directive mesentery in its lower part.

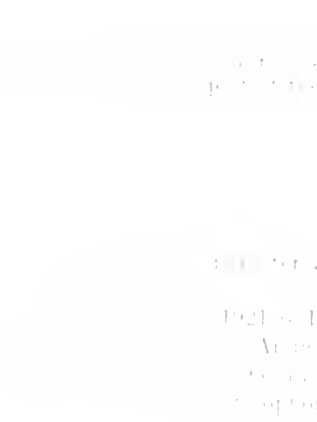


FIG. 78. *Andreakia parva*. Transverse section of an imperfect mesentery proximal to the oesopharynx.

ramified folds (fig. 75). The parietal muscles of the perfect as well as of the imperfect mesenteries (fig. 76) are in their upper part more or less fan-like, in their lower part elongated in a radial direction, the muscles are here somewhat weaker on the side where the retractor is situated than on the opposite (fig. 77). The ciliated tracts of the filaments are well developed on the perfect mesenteries, the endoglandular tract is present also at least on several imperfect mesenteries which, however, had not ciliated tracts.



FIG. 77. *Andreakia parva*. Transverse section of a directive mesentery in its lower part.

The perfect and some of the imperfect mesenteries are provided with acentia. No sectioned specimen was fertile, but owing to the smallness of the imperfect mesenteries it is not likely that they develop generative organs. The species propagates probably asexually, as there are so many irregularities in the arrangement of the mesenteries. The specimens occur often together on shell-fragments or on small stones.

The nematocysts of the acentia I have examined in 4 specimens, their size showed good agreement. The microbasic amastigophors were 24-32, 27-31, 26-31 and 24-31 μ , the basitrichs 24-29, 26-29, 24-30 and 22-23 μ .

Family Octineonidae.

1903, 79 and acentia, the latter (1903, 80) was referred to the family Amastigophoridae, with which it was placed. It is suggested, that in the present case, as well as basitrichs

in the acentia as I could at that time examine the acentia only in sections. As the following species had no amastigophors in the acentia I have now made a renewed examination of these organs in *O. loadable* on material presented me 1934 by the British Museum. The maceration preparations of the acentia showed clearly that in this species also only basitrichs are present.

Genus *Octineon* Fowl.

Pedal disc flat, broad, body conical, the upper part often raised as a cylinder from the lower one, divisible into scapus and scapulus, the former with a cuticle, sometimes incrustated with grains of sand as also is the pedal disc. Ectoderm of the scapus showing tendency to reduction in certain places. Scapulus without spirocysts and cuticle. Sphincter mesogloea. Tentacles few, hexamerously arranged, rather weak, considerably fewer than the mesenteries, capable of invagination. Siphonoglyphs indistinct. Only the 8 "Edwardsia"-mesenteries perfect, fertile, with filaments and acontia and circumscript retractors, the 5th and 6th couples, forming pairs with the lateral "Edwardsia"-mesenteries, stronger than those of the second cycle but of same appearance. Parieto-basilar muscles broad but weak, their innermost part forming a shelf (always?) on the "Edwardsia"-mesenteries. Considerably more mesenteries in the proximal than in the distal part. Chidom: spirocysts, basitrichs microbasic *p*-mastigophors.

Octineon sueticum Carlg.

Octineon sueticum n. sp. CARLGRÉN 1910 p. 28, 59 figs. IX 14-16.

Diagnosis: Cuticle of the scapus strong, neither the scapus nor the pedal disc incrustated with sand. Sphincter mesogloea with small muscle meshes, wholly separated from the endodermal circular muscles of the column. Tentacles small, up to 21. Pennons of the "Edwardsia"-mesenteries and the muscles of the 5th and 6th couples stronger than in *O. lindahli*. Relation between the number of mesenteries and that of the tentacles less than in this species (about 1:1). Nematocysts of the scapus 12-13 - about 3.5 μ , basitrichs, those of the tentacles 11-19 - almost 2-2 μ , often curved, basitrichs, those of the actinopharynx partly 17-23 - about 3.5 μ , microbasic *p*-mastigophors, partly 14-24 - almost 3 μ , basitrichs, those of the filaments partly 11-14 - 3-3.5 μ , microbasic *p*-mastigophors, partly 17-19 - (3.5) 4 μ , microbasic *p*-mastigophors, partly (8) 10-12.5 μ , basitrichs, those of the acontia partly 48-65 - 1.5-5.5 μ , partly 11-20 - 1.5-almost 2 μ , very sparse, both basitrichs. Spirocysts of tentacles about 12 - 2-24 - 2.5 μ .

Colour: Cuticle in older specimens brownish or dirty grey, the upper part of the scapulus provided with 6 opaque white, elongated spots, alternate with them on the lowest part of the scapulus three opaque-white longitudinal lines, especially visible when the animal is wholly expanded, sometimes more indistinctly arranged. Oral disc colourless or reddish-brown, tentacles uncoloured, sometimes with an indistinct cross-band in the middle, their base white or yellowish-white, actinopharynx brownish, shading into orange.

Dimensions: height up to 0.8 cm in half contracted state, pedal disc 0.9 - 0.6 cm.

Occurrence: Sweden, Bohuslän Väderöar in the vicinity of the Lophohelia-reef 60-70 m; on small stones or shells together with *Ambakia parva*.

Exterior aspect: The pedal disc is flattened and very wide. When wholly contracted the animal is almost disc-like, half contracted it forms a low conus, wholly or almost wholly expanded, the upper part rises as a cylinder from the strongly flattened, lower part. The body is divisible into scapus and scapulus, the former provided with a cuticle, which in older specimens is thick and in contracted state of the animal very wrinkled. The tentacles are thin and delicate and capable of invagination as those of *O. lindahli* and *Halcampoides*. In sections of contracted specimens some tentacles are often invaginated. Their number are about 21, at least in older specimens. The actinopharynx is of ordinary length, the siphonoglyphs are indistinct.

Anatomical description: The organization of this species agrees very well with that of *O. lindahli* (compare FOWLER 1894, CARLGRÉN 1921 p. 132-133, 1931 p. 10). The ectoderm of the



Fig. 78. *Octineon sueticum*. Section of a part of the scapus showing the gland-cells in the transformed ectoderm.

scapus is mostly thin, but certainly present and provided with a cuticle. In certain places it is thicker than the mesogloea, and the nuclei of the cells and small nematocysts are clearly visible, in

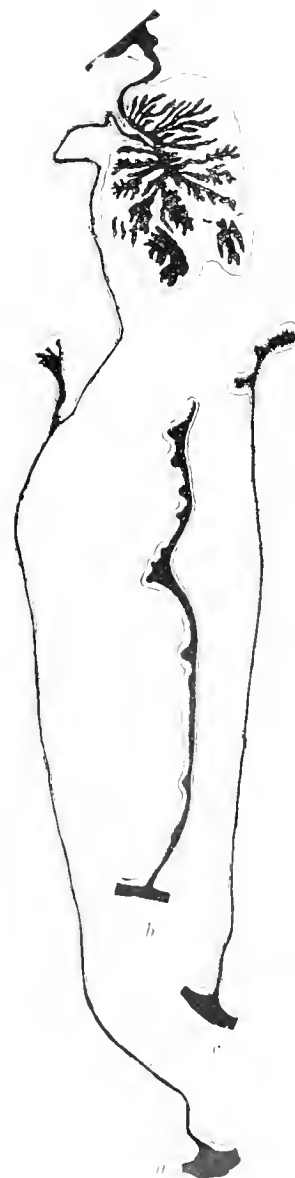


Fig. 79. *Octineon sueticum*, a: Perfect mesentery in the lower part of the actinopharynx, b: mesentery of the second order, c: the same in its lower part; the exocoelomic side on the left. Cross sections.

other places, also in the same section, not so distinct, as there seems to be a subcuticle above the reduced and possibly partly chitinized ectoderm cells. Possibly we have to do with an organization as in the *tenaculi*, but as the reduction of the ectoderm is distributed

of the mesogloea, the numerous papillae, it is questionable whether the mesogloea is provided with distinct tenacules. The mesogloea of *O. lundabli* as the ectoderm is often not provided with tenacules, but only by the gland cells, which are present in the inner layer (2-78). Such gland cells are present in *O. succicum* and especially in the endoderm of the acontia, where they form an almost continuous layer at the surface (2-78). Also in the endoderm of the mesenteries the gland cells are present in a similar manner. Probably we have to do with the same glands as the gland cells of the mesogloea sometimes present in the ectoderm. FOWLER, having not seen any ectoderm of *O. succicum* (in opposition to myself), suggested that the "tenacules" of mesogloea for the adhesion of the sand" (1910, p. 105) were effected by wandering cells from the endoderm, (fig. 100) "it is reasonable to doubt their presence". Whether there are any wandering cells in *lundabli* I cannot confirm owing to the poor preservation of the specimens, but in *succicum* such wandering cells may be present.

The sphincter is ring-shaped and rather strong, its muscle meshes are well developed, separated from the circular muscles of the pharynx. Whether the sphincter in the expanded state of the animal is in a state of extension as in *lundabli*, I have not been able to determine, as the mesogloea in the region of the sphincter was in a state of retraction owing to strong contraction. The ectoderm of the pharynx, tentacles and oral disc is of usual appearance, that of the actinopharynx, apart from that in the ridges, considerably stronger than the thin mesogloea.

The mesenteries are more numerous in the proximal than in



Fig. 80. Cross-section of a retractor of a perfect mesentery of *O. succicum*, close to the actinopharynx.

the distal. The relation is about 4:1. The number of mesenteries at the oral disc in one specimen of *O. succicum* is 16. The tentacles are macrotenacles, and the acontia are small and unproductive organs, all of which are present in pairs. The folds of the retractors are

more robust and more numerous, at least in older specimens (fig. 79 a) than those of *O. lundabli*, sometimes the pennons showed a tendency to be divided into two parts at the upper part of the actinopharynx, a tendency more conspicuous in other retrac-



Fig. 81. *Octopus succicum*, inner part of the sixth couple of mesenteries.

tors than in that figured (fig. 80). Apart from the retractors the longitudinal muscles of the "Edwardsia"-mesenteries are missing or, if present, at any rate very weak; on the opposite side of the retractors the muscles are clearly visible, forming a distinct layer and terminating, at least sometimes, as a fold running out from the main lamella of the mesenteries. The description of FOWLER indicates that the muscles form folds in a similar manner in *O. lundabli*, but whether this fold is so deep, that it forms a distinct lamella is questionable, as the figures 14-17 Pl. 30 are difficult to interpret; an offshoot in the mesentery drawn in fig. 14 may possibly be a similar fold. A closer examination is desirable. The imperfect mesenteries of the 5th and 6th couples, forming pairs with the lateral "Edwardsia"-mesenteries, have no distinct retractors but their free borders have rather numerous longitudinal muscle folds, stronger on the 5th couple than on the 6th (fig. 81). On the endocoel sides the muscles are missing or, at any rate, if present, very weak, on the opposite sides, however, clearly visible. Thus, the muscles corresponding to the parietobasilar muscles are here developed. The mesenteries of the second cycle (figs. 79 b, c) agree with those of the 5th and 6th couples but are weaker and show the same arrangement of the here weaker muscles. The ciliated tracts of the filaments are well developed. One of the specimens examined was fertile and had ovaries, enclosing small ova, in the perfect mesenteries.

I have measured the numerous basitrichs in the acontia in 4 specimens. They were $48.60 \pm 5\mu$, $50.60 \pm 5\mu$, $53.58 \pm 5\mu$ and $52.65 \pm 5\mu$. Moreover there occur in the acontia also small but few nematocysts about $14.20 \pm 1.5\mu$, basitrichs. The cnidae of the tentacles and actinopharynx I have examined in 2 specimens.

The size of the cnidae in *O. lundabli* agrees very well with those in *succicum*. In maceration preparations of *lundabli* I have found the following measures of the cnidae: tentacles: basitrichs $14.48 \pm 1.5 \pm 2\mu$; acontia: basitrichs unexploded $50.57 \pm 5\mu$, exploded $48.58 \pm 5\mu$, moreover smaller basitrichs $17.49 \pm 2\mu$, very sparse; spirocysts of the tentacles about $12.25-21 \pm 5\mu$.

Family Actiniidae.

Genus *Anemonia*.

Actiniidae with well developed pedal disc. Column smooth, provided with marginal sphaerules on the parapet (not in the fossa as in *Actinia*), sometimes mis-carried or absent in young individuals. Sphincter endodermal, concentrated or extended, diffuse. Tentacles long, sometimes not perfectly retractile, considerably fewer than the mesenteries at the base; longitudinal muscles of tentacles ectodermal. Siphonoglyphs in varying number. Perfect mesenteries numerous. Gonads appearing from the mesenteries of the first cycle onwards. Unidom: Spirocysts, atrichs, basitrichs, microbasal *p*-mastigophors (and sometimes holotrichs?).

Anemonia sulcata (Pennant).

Literature and synonyms see ANDRES 1883 and STEPHENSON 1935, also: *Anemonia sulcata* PENN. WEILL 1931 p. 567 figs. 375, 376.

Diagnosis: Tentacles long, almost of same length as the column when expanded, in number up to about 200 in large individuals, often irregularly arranged owing to asexual reproduction by longitudinal fission. Marginal sphaerules, when present, in varying number. Sphincter concentrated or more extended, diffuse. Arrangement of mesenteries often irregular. Number of mesenteries at the base up to about 250 (or more?). Retractors of mesenteries extended, diffuse, parietobasilar muscles weak. Endoderm provided with zooxanthellae. Nematocysts of the column partly 14-17 \times 2.5-3 μ , basitrichs, partly 24-27 \times about 5 μ , holotrichs?, sparse, those of the marginal sphaerules partly 29.6-39 \times 3-4.2 μ (45-55 \times 4.5 μ), those of the tentacles 17-34 \times (40) \times 2-3 (5) μ , basitrichs, those of the actinopharynx partly 27-32.5 \times 3-3.5 μ (up to 70 \times 10 μ ! WEILL 1931) basitrichs, partly 21-22.6 \times about 4.2 μ few, microbasal *p*-mastigophors, those of the filaments partly 26-35.2 \times 4.2-almost 5 μ , partly 18.3-24 \times 2 μ , both probably basitrichs, partly 18.3-24 \times 3.5-4.5 (5) μ , microbasal *p*-mastigophors. Spirocysts of tentacles up to about 39 \times 3 μ .

Colour of the present specimens according to DOXS: Column greyish brown, tentacles emerald green.

Size of one of the largest individuals: breadth of the very contracted pedal disc 0.7-1.2 cm, that of the expanded oral disc 1.3 cm, length of inner tentacles about 1 cm.

Occurrence: Norway, Espeland in the vicinity of Bergen on algae (DOXS 1936) several specimens.

Further distribution: South and west coasts of England, west coast of Scotland, Ireland, France, Spain, the Mediterranean, Madeira, Canaries.

The information of the size of the nematocyst and spirocysts in the diagnosis is based on an examination of an individual of medium size from the Canary Islands. In brackets I have given the measures which WEILL (1931) has noted and which more essentially differ from mine. I am very sceptical that the basitrichs can reach a size of 70 \times 10 μ in the actinopharynx. This may be a mistake of WEILL who did not note the large basitrichs in the filaments. WEILL (1931) has drawn a figure (376 p. 568) of a nematocyst in size about 27.5 μ . Although he has not seen this nematocyst in an exploded state he thinks that it may be an atrich. I have found several such in my individuals, but the thread of the capsules showed sometimes a more regularly arrangement about as in the text-figure 1 d of *Arachnanthus australae* (CARLQVIST 1937) though the spirals were fewer (9 in one specimen). To my mind it is very questionable if these nematocysts are atrichs. They recall very much some holotrichs in the Cerantharia and the holotrichs in *Diodon* (CARLQVIST 1940 p. 38 fig. 17). The wall of the capsules seem to be very thin, the thread thick and, as far I can see, unfortunately no capsules were exploded - armed or at any rate not smooth. Possibly one could suggest that the thread was very compressed and does not become its definite length ere it explodes. Such a presumption is, however, hardly plausible, as I have never found atrichs the thread of which is compressed in such a way in the capsules, and the thread of the atrichs is thin. For my part I must at present characterize these nematocysts as holotrichs, though an examination of the thread in an exploded state is desirable in order to definitively decide the nature of these capsules. In the diagnosis of the genus and species I have used the term concentrated-diffuse instead of circumscribed-diffuse because this term seems to me better to answer to the appearance (see STEPHENSON 1928 fig. 1 c). When we speak of a circumscribed-diffuse sphincter a part of it may be circumscribed.

The pedal disc and the lower part of the body were fairly strongly contracted in the present specimens, the upper part of the body was expanded, with longitudinal furrows corresponding to the insertions of the mesenteries. No marginal sphaerules were developed. The sphincter was, in one specimen examined, concentrated-diffuse. The largest specimen had 81 tentacles but between 130-140 mesenteries at the base. Of 4 specimens examined 2 had 2 siphonoglyphs, 2 only one. There were many perfect mesenteries. The nematocysts agree rather well in their size with those of the specimen from the Canary Islands. The basitrichs of the column were 12.7-15.5 \times 2.5-2.8 μ , the holotrichs? 24-27 \times about 5 μ , the basitrichs of the tentacles 21-30.5 \times about 3 μ , the basitrichs of the actinopharynx 27-34 \times 3.5-4.2 μ , the microbasal *p*-mastigophors 21-24 \times 4.2 μ , the larger basitrichs of the filaments 28.2-35.2 \times 4.2-5 μ , the microbasal *p*-mastigophors 18.3-22.6 \times 4.2-5 μ .

For further details see SIMON (1892), SCHNEIDER (1902, 1908), PAX (1907) and STEPHENSON (1935).

Family Actinostolidae.

Parasicyonis ingolfi n. sp.

Diagnosis: Body low. Sphincter very strong, reticular to alveolar in the outer and upper parts, with traces of stratification. Tentacles 72, probably arranged 6 + 12 + 18 + 36. About half of the oral disc devoid of tentacles. Arrangement of the mesenteries probably 6 + 12 + 18 + 36 \times 72 pairs, 2 pairs of directives. The 36 first pairs with rather well developed diffuse retractors in the upper part of the body. Parietobasilar muscles weak. Nematocysts of the column 17-21 \times 2.5-2.8 μ , basitrichs, those of the tentacles 22-34 \times 2.5-2.8 μ , basitrichs, those of the actinopharynx partly 25.8-28.2 \times 4.2 μ , microbasal *p*-mastigophors, partly 23.3-

28.2 \times 2.8-3.2 μ , basitrichs, those of the filaments partly (18.3) 24-28.2 \times (4.2) 4.5-5 μ , microbasal *p*-mastigophors, partly 12.7-21.8 \times 2.1-2.8 μ , basitrichs. Spirocysts of tentacles up to 48 \times about 4 μ .

Colour in alcohol: yellowish with large dark brown spots of indeciduous ectoderm. Tentacles and actinopharynx paler than the column but pigmented.

Dimensions: Breadth 2.8 cm, length of the column about 1.8 cm, height of the strongly contracted specimen about 0.5 cm.

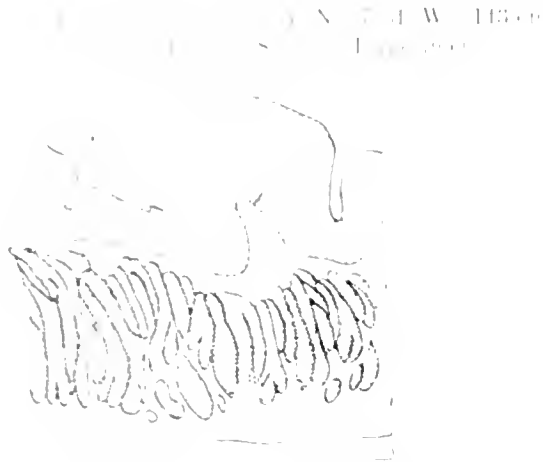


Fig. 82. Section of a part of a tentacle.

The specimen was very contracted, the distal part of the actinopharynx and the tentacles were conspicuous. In this stage

the tentacles were deeply and provided with a cuticle, the distal part distinct tubercles but with irregular thickening between these flattened thickenings, the distal, revolved part is long. Because the margin is not distinct, owing to the contraction, it is difficult to decide the position of the tentacles. As a rule, the ridges of the uppermost part of the outer tentacles without definite thickening of the basal outer sides of the tentacles are broad, irregularly wrinkled, and they meet together from the sides and not



the tentacles meet together from their distal ends. The outer tentacles are arranged as in the figure, but it seems as

if there were only two cycles of tentacles. About the inner half of the oral disc is devoid of tentacles. The actinopharynx is of ordinary length and provided with 2 siphonophylphs.

Anatomical description. Owing to the strong contraction of the specimen and the bad preservation I cannot give a detailed description of the anatomy.

The ectoderm of the pedal disc is of ordinary thickness and pigmented, its cuticle strong. The mesogloea is thin in comparison with that of the column, but thicker than the ectoderm, and fibril-



Fig. 84. *Pteris equans rapalli*. Section of a mesentery in the region of the endoglandular tract.

lar. The ectoderm of the column is pigmented, without a cuticle and rather high but many times thinner than the mesogloea. The mesogloea is thick, especially in the sphincter region, the fibrillae are extended in the inner parts of the mesogloea, in the outer they cross each other, so that a basket work of fibrillae arises. The endodermal circular muscles of the column are weak, on the other hand the sphincter is strong and very broad, but diminishing rather abruptly, mainly reticular with the meshes drawn out in a radial direction. In its upper and outer parts the sphincter shows a tendency to be alveolar and stratified (fig. 83).

The ectoderm of the tentacles is high and very folded, as also the mesogloea. The longitudinal muscles are strong and mesogloea, the large muscle meshes often drawn out in a radial direction (fig. 82). The radial muscles of the oral disc agree with the longitudinal muscles of the tentacles.

The parts of mesenteries are probably 72 (6 · 12 · 18 · 36 · 72), the last 36 of which are present only in the proximal part of the body, the other mesenteries reach the distal end. In about

the half of the animal there were 18 perfect mesenteries. Thus, I think that $6 + 12 = 18$ pairs are perfect in the whole specimen. Of these two pairs are directives. Possibly irregularities of the arrangement of the youngest mesenteries may occur, but owing to the bad preservation of the mesenteries it is very difficult to decide it. The longitudinal muscles of the 36 first pairs are rather well developed and form broad, diffuse retractors which, in some places, especially in the upper part of the body, are rather high (fig. 8f). The muscles of the 36 other pairs are weak. The parieto-

basilar muscles are weak, at least in their upper part, and not situated on a fold of the mesogloea. In the lower part of some mesenteries there are muscle meshes enclosed in the mesogloea. Possibly these enclosures indicate parts of the parietobasilar muscles, but as they lie closer to the longitudinal muscles I am more inclined to suppose that they belong to these. The basilar muscles are distinct but not strong. Only the 36 youngest pairs of mesenteries are fertile and provided with large ova. All mesenteries are provided with filaments.

II. Distribution of the species.

In some previous papers I have given a survey of the Actiniaria from West Greenland and adjacent areas (1933), from East Greenland (1928), from Iceland (1939) and from the Faroe Islands (1930). These reports are here brought near to completion. I now account for all the material of the Actiniaria as well as of the Ceriantharia and Zootharia (the two latter in brackets) which I have examined from the Ingolf expedition and from northern and arctic regions. We begin with the enumeration of the species from the westernmost waters.

West and Northwest Greenland, Davis Strait and Baffin Bay: *Actinernus nobilis*, *Edwardsia andresi*, *Halcampoides purpurea*, *Aethelmis intestinalis*, *Peachia parasitica*, *Halcampa arctica*, *Cactosoma abyssorum*, *Haliactis arctica*, *Liponema multicornis*, *Bolocera tardia*, *Bolocera maxima*, *Banodactis stella*, *Banodactis spetsbergensis*, *Cribrinopsis similis*, *Tealia felina crassicornis*, *Epiactis nordmanni*, *Anthosactis jan mayeni*, *Tealidium jungermani*, *Pycnanthus laevis*, *Sicyonis tuberculata*, *Sicyonis ingolfi* (S. of Greenland), *Parasicyonis groenlandica*, *Actinostola callosa*, *Actinostola groenlandica*, *Actinostola spetsbergensis*, *Stomphia coccinea*, *Kadosactis rosea*, *Chondrophellia coronata*, *Phelliactis hertwigi*, *Phelliactis robusta*, *Actinauge cerrilli*, *Allantactis parasitica*, *Hornathia nodosa*, *Hornathia digitata*, *Metridium senile dianthus*, *Stephanauge acanellae*, *Amphianthus margaritaceus*, *Amphianthus verruculatus*, (*Siphonactinopsis laevis*?); in all 38 (39!) species.

To these is to be added the unidentified *Actinia*? *spectabilis* Fabr.

Cerianthus sp.? (*lloyli*? see below) *Isozoanthus bulbosus*, *Isozoanthus davisi*, *Isozoanthus ingolfi*, *Epizoanthus lindahli*, *Epizoanthus erdmanni*, *Epizoanthus danielsenii*, *Epizoanthus glacialis*; in all 8 species.)

As to the distribution of the species in the different areas see CARLGRÉN (1933).

East Greenland and Danmark Strait: East Greenland, *Edwardsia citrea*, *Edwardsia arctica*, *Edwardsia incerta*, *Nematostella* (*Milne-Edwardsia*) *polaris*, *Nematostella nathorsti*, *Halcampoides purpurea*, *Banodactis stella*, *Tealia felina crassicornis*, *Anthosactis jan mayeni*, *Hornathia nodosa*, *Allantactis parasitica*, Danmark Strait, northern, arctic part: *Parasicyonis ingolfi*, southern atlantic part: *Tealidium jungermani*, *Sicyonis tuberculata*, *Danotesia praelonga*, *Sagartiogeton cerrilli*, *Sagartiogeton ingolfi*, *Chondrophellia coronata*, *Phelliactis robusta*, *Phelliactis hertwigi*, *Paraphelliactis spinosa*, in all 21 species. Probably some more species are present in the waters of the not closely examined south-east coast.

(*Cerianthus* sp.?— *Isozoanthus bulbosus*, *Isozoanthus arborescens*, *Isozoanthus magnisulcosus*, *Isozoanthus islandicus*, *Epizoanthus lindahli*, *Epizoanthus erdmanni*; in all 7 species.)

Iceland, coastal area: *Ptychodactis patala*, *Edwardsia tuberculata*, *Edwardsia andresi*, *Edwardsia islandica*, *Nematostella polaris*, *Limnactinia laevis*, *Halcampoides purpurea*, *Halcampa arctica*, *Bolocera tardia*, *Banodactis stella*, *Cribrinopsis similis*, *Tealia felina crassicornis*, *Parasicyonis sarsi*, *Actinostola spetsbergensis*, *Stomphia coccinea*, *Sagartia troglodytes*, *Sagartia elegans*, *Phellia gonsapata*, *Hornathia digitata*, *Hornathia nodosa*, *Allantactis parasitica*, *Actinauge cerrilli*, *Amphianthus islandicus*, *Metridium senile dianthus*; in all 21 species.

(*Isozoanthus multisulcosus*, *Isozoanthus altisulcatus*, *Parazoanthus anguicomus*, *Epizoanthus erdmanni*, *Epizoanthus incrustatus*; in all 5 species.)

Atlantic abyssal area S.W. to S.E. of Iceland: *Isoboloesia ingolfi*, *Bolocera tardia*, *Epiactis dubia*, *Sicyonis variabilis*, *Actinostola callosa*, *Danotesia praelonga*, *Sagartiogeton abyssorum*, *Chondrophellia coronata*, *Phelliactis hertwigi*, *Phelliactis robusta*, *Stephanauge acanellae*, *Amphianthus ingolfi*, *Amphianthus verruculatus*; in all 13 species.

(*Isozoanthus arborescens*, *Epizoanthus papuciphilus*; in all 2 species.)

The area between Iceland and the Faroe Islands: *Halcampa arctica*, *Cribrinopsis similis*, *Hornathia digitata*, *Hornathia nodosa*; in all 4 species.

The Faroes, coastal area: *Edwardsia tuberculata*, *Edwardsia danica*, *Actinia equina*, *Bolocera tardia*, *Cribrinopsis similis*, *Tealia felina crassicornis*, *Stomphia coccinea*, *Sagartia troglodytes*, *Hornathia digitata*, *Hornathia nodosa*, *Metridium senile dianthus*; in all 11 species.

(*Parazoanthus haldoni*, *Epizoanthus incrustatus*, in all 2 species.)

Cold deep basin of the Norwegian Sea: *Bathypollia margaritacea*, *Kadosactis rosea*, *Amphianthus margaritaceus*. To this area probably belong also *Anthosactis ingolfi*, *Parasicyonis ingolfi* and *Kolidades pedunculata*¹ taken at depths between about 1370 and 2000 m. each in a single locality. In all 3 species.

The boundary at the area between 0 and - temperatures in the Norwegian Sea: *Edwardsia citrea*, *Halcampoides purpurea*, *Cactosoma abyssorum*, *Banodactis spetsbergensis*, *Cribrinopsis similis*, *Epiactis arctica*, *Pycnanthus densus*, *Actinostola spetsbergensis*; in all 8 species. All localities, apart from that of *Cactosoma*, which was taken at 0 as well as - the temperatures were below zero (°C).

(*Cerianthus cogli* (new locality: 63°26' N, 7°56' W, 171 fms. bottom temp. -0.69°, Ingolf exp. St. 138). *Epizoanthus roseus*, *Epizoanthus glacialis*, *Epizoanthus koreni* () in all 1 species.)

West Spitzbergen: *Edwardsia citrea*, *Nematostella polaris*, *Halcampa arctica*, *Banodactis stella*, *Cribrinopsis similis*, *Tealia felina crassicornis*, *Actinostola spetsbergensis*, *Stomphia coccinea*, *Stomphia polaris*, *Kadosactis* (?) *spetsbergensis*, *Allantactis parasitica*, *Hornathia digitata*, *Hornathia nodosa*; in all 13 species.

(*Cerianthus* sp.? (*roulei*) *Epizoanthus erdmanni*; in all 2 species.)

East Spitzbergen: *Edwardsia citrea*, *Nematostella polaris*, *Nematostella nathorsti*, *Halcampa arctica*, *Haliactis arctica*, *Banodactis stella*, *Banodactis spetsbergensis*, *Cribrinopsis similis*, *Tealia felina crassicornis*, *Actinostola spetsbergensis*, *Stomphia coccinea*, *Stomphia polaris*, *Allantactis parasitica*, *Hornathia digitata*?, *Hornathia nodosa*; in all 11 (15!) species.

(*Cerianthus* sp.?; *Epizoanthus erdmanni*; in all 2 species.)

The area between Spitzbergen and Norway: *Edwardsia*

¹ I have examined the type, but as it was badly preserved and only part of it remains, it was impossible to clear up its systematic place. The sphincter was mesogloea and very strong, occupying almost the whole breadth of the mesogloea, its muscle meshes very large.

100° W. to 105° W., 60° N. to 65° N.: *Haliactis arctica*,
Parasactis fuscus, *Actinostola callosa*, *Actinostola*
spetsbergensis, *Stomphia coccinea*, *Hornathia*
digitata, 12 species.
 100° W. to 105° W., 65° N. to 70° N.: *Edwardsia*
arctica, *Halocampa duodecimcirrata*, *Tealia felina*
coriacea, *Stomphia coccinea*, *Sagartia troglodytes*,
Sagartiopecton undatus, *Sagartiopecton lacustris*,
Hornathia digitata, *Metridium senile danthous*, in all 12 species.
 100° W. to 105° W., 70° N. to 75° N.: *Edwardsia*
arctica, *Halocampa duodecimcirrata*, *Tealia felina*
coriacea, *Sagartia troglodytes*, *Sagartiopecton undatus*,
Hornathia digitata, *Metridium senile danthous*, in all 6 species. Probably some more
 species may be present here.
 Little Belt: *Edwardsia danica*, *Halocampa duodecimcirrata*,
Tealia felina coriacea, *Sagartia troglodytes*?, *Sagartiopecton undatus*,
Metridium senile danthous; in all 5 (6?) species.
 Kiel Bay: *Edwardsia danica*, *Halocampa duodecimcirrata*,
Tealia felina coriacea, *Metridium senile danthous* and probably
Sagartiopecton undatus in all 4 (5) species.
 Lübeck Bay: *Edwardsia danica*, *Halocampa duodecimcirrata*,
Tealia felina; in all 3 species.
 Mecklenburger Bay: *Edwardsia danica*, *Halocampa duodecim-*
cirrata; in all 2 species.
 Arcoria Basin and Bornholm to about 16° E.: *Halocampa*
duodecimcirrata, (*Tealia felina lufotensis*?)¹; S. of Bornholm: 1 (2?)
 species.
 Coast of Murman: *Edwardsia fusca*, *Actinia equina*, *Banodac-*
tis stella, *Cerbrinopsis similis*, *Tealia felina crassicornis*, *Actinostola*
spetsbergensis, *Stomphia coccinea*, *Allantactis parasitica*, *Hornathia*
digitata, *Hornathia nodosa*, *Metridium senile danthous* and var.
naqilnopsensis; in all 11 species.
 (*Ceranthus*, probably *loggii*, *Archiactis albida*, *Epizoa*
edmanni; in all 3 species).
 Franz Joseph Land: *Edwardsia arctica*, *Halocampa arctica*,
Banodactis spetsbergensis, *Tealia felina crassicornis*, *Stomphia*
coccinea, *Hornathia nodosa*; in all 6 species.
 The White Sea and Barents Sea: *Nematostella polaris* (new
 localities: 65° 36' N., 19° 02' 18" E., 65° 13' 12" N., 37° 15' E.), *Hal-*
ocampa arctica, *Haliactis arctica*, *Actinostola spetsbergensis*, *Allantactis*
parasitica, *Hornathia digitata*, *Hornathia nodosa*; in all 7 species.
 (*Epizoa* *lindahli*.)
 Kara Sea: *Edwardsia arctica*, *Banodactis stella*, *Tealia felina*
crassicornis, *Anthosactis jan mayeni*, *Actinostola spetsbergensis*,
Allantactis parasitica, *Hornathia digitata* var. *regae*?, in all 7
 species.
 Siberian Sea, Cape Tscheljuskin to Cape Onnan: *Haliactis*
arctica, *Banodactis stella*, *Epiactis marsupialis*, *Epiactis incerta*,
Anthosactis jan mayeni, *Actinostola spetsbergensis*, *Allantactis*
parasitica, *Hornathia digitata* var. *regae*, *Hornathia nodosa*; in
 all 9 species.
 Bering Strait, Cape Lisburn Cape Onnan to Cape Chaplin-
 Norton Sound: *Ptychodactis patula*, *Edwardsia regae*, *Haliactis*
arctica, *Banodactis stella*, *Banodactis spetsbergensis*, *Tealia felina*
crassicornis, *Epiactis marsupialis*, *Epiactis levisi*, *Epiactis arctica*
 (N. of Bering Str.), *Actinostola spetsbergensis*, *Stomphia coccinea*;
 in all 11 species.
 Bering Sea, Bering Strait to Aleutian Isl.—Bering Isl.: *Hal-*
ocampoides purpurea, *Prachia parasitica*, *Halocampa*? *regae*, *Charisca*
sarcoida, *Liponema multicornis*, *Banodactis stella*, *Cnidopus ritteri*,
Anthopleura xanthogrammica, *Cerbrinopsis similis*, *Tealia felina*
crassicornis and var. *coriacea*, *Actinostola spetsbergensis*, *Stomphia*
coccinea, *Metridium senile pubriatum*; in all 13 species.
 Since the Actinaria dealt with here include not only those of
 the Ingolf expedition but also those taken by several other ex-
 peditions from arctic, boreal and North Atlantic regions it is clear
 that their zoogeographical character is very different. I have
 below made an attempt to classify them though, of course, the
 zoogeographical classification is dubious for several species.

thous danicus (only in Långfjord. MORTENSEN notes (1897) also
*Zoothous*¹ *salicatus* from Långfjord, possibly identical with *danicus*.)
 In all 3 species.)

The Sound from Gillsberg Hoved Kullen: *Edwardsia longi-*
cornis, *Edwardsia danica*, *Halocampa duodecimcirrata*, *Banodactis*
stella, *Tealia felina coriacea*, *Stomphia coccinea*, *Sagartia troglody-*
tes, *Sagartiopecton undatus*, *Sagartiopecton lacustris*,
Hornathia digitata, *Metridium senile danthous*, in all 12 species.

Great Belt: *Edwardsia danica*, *Halocampa duodecimcirrata*,
Tealia felina coriacea, *Sagartiopecton undatus*, *Stomphia coccinea*,
Metridium senile danthous, in all 6 species. Probably some more
 species may be present here.

Little Belt: *Edwardsia danica*, *Halocampa duodecimcirrata*,
Tealia felina coriacea, *Sagartia troglodytes*?, *Sagartiopecton undatus*,
Metridium senile danthous; in all 5 (6?) species.

Kiel Bay: *Edwardsia danica*, *Halocampa duodecimcirrata*,
Tealia felina coriacea, *Metridium senile danthous* and probably
Sagartiopecton undatus in all 4 (5) species.

Lübeck Bay: *Edwardsia danica*, *Halocampa duodecimcirrata*,
Tealia felina; in all 3 species.

Mecklenburger Bay: *Edwardsia danica*, *Halocampa duodecim-*
cirrata; in all 2 species.

Arcoria Basin and Bornholm to about 16° E.: *Halocampa*
duodecimcirrata, (*Tealia felina lufotensis*?)¹; S. of Bornholm: 1 (2?)
 species.

Coast of Murman: *Edwardsia fusca*, *Actinia equina*, *Banodac-*
tis stella, *Cerbrinopsis similis*, *Tealia felina crassicornis*, *Actinostola*
spetsbergensis, *Stomphia coccinea*, *Allantactis parasitica*, *Hornathia*
digitata, *Hornathia nodosa*, *Metridium senile danthous* and var.
naqilnopsensis; in all 11 species.

(*Ceranthus*, probably *loggii*, *Archiactis albida*, *Epizoa*
edmanni; in all 3 species).

Franz Joseph Land: *Edwardsia arctica*, *Halocampa arctica*,
Banodactis spetsbergensis, *Tealia felina crassicornis*, *Stomphia*
coccinea, *Hornathia nodosa*; in all 6 species.

The White Sea and Barents Sea: *Nematostella polaris* (new
 localities: 65° 36' N., 19° 02' 18" E., 65° 13' 12" N., 37° 15' E.), *Hal-*
ocampa arctica, *Haliactis arctica*, *Actinostola spetsbergensis*, *Allantactis*
parasitica, *Hornathia digitata*, *Hornathia nodosa*; in all 7 species.
 (*Epizoa* *lindahli*.)

Kara Sea: *Edwardsia arctica*, *Banodactis stella*, *Tealia felina*
crassicornis, *Anthosactis jan mayeni*, *Actinostola spetsbergensis*,
Allantactis parasitica, *Hornathia digitata* var. *regae*?, in all 7
 species.

Siberian Sea, Cape Tscheljuskin to Cape Onnan: *Haliactis*
arctica, *Banodactis stella*, *Epiactis marsupialis*, *Epiactis incerta*,
Anthosactis jan mayeni, *Actinostola spetsbergensis*, *Allantactis*
parasitica, *Hornathia digitata* var. *regae*, *Hornathia nodosa*; in
 all 9 species.

Bering Strait, Cape Lisburn Cape Onnan to Cape Chaplin-
 Norton Sound: *Ptychodactis patula*, *Edwardsia regae*, *Haliactis*
arctica, *Banodactis stella*, *Banodactis spetsbergensis*, *Tealia felina*
crassicornis, *Epiactis marsupialis*, *Epiactis levisi*, *Epiactis arctica*
 (N. of Bering Str.), *Actinostola spetsbergensis*, *Stomphia coccinea*;
 in all 11 species.

Bering Sea, Bering Strait to Aleutian Isl.—Bering Isl.: *Hal-*
ocampoides purpurea, *Prachia parasitica*, *Halocampa*? *regae*, *Charisca*
sarcoida, *Liponema multicornis*, *Banodactis stella*, *Cnidopus ritteri*,
Anthopleura xanthogrammica, *Cerbrinopsis similis*, *Tealia felina*
crassicornis and var. *coriacea*, *Actinostola spetsbergensis*, *Stomphia*
coccinea, *Metridium senile pubriatum*; in all 13 species.

Since the Actinaria dealt with here include not only those of
 the Ingolf expedition but also those taken by several other ex-
 peditions from arctic, boreal and North Atlantic regions it is clear
 that their zoogeographical character is very different. I have
 below made an attempt to classify them though, of course, the
 zoogeographical classification is dubious for several species.

¹ The locality seems to me somewhat dubious. Possibly a confound-
 ing of labels has taken place.

Especially this is the case when a species has been taken in a single locality or in two localities one of which showed a negative temperature the other a positive one. In brackets I have also included the Ceriantharia and Zoantharia as I have previously given a zoogeographical survey of only a few species of these orders.

Atlantic abyssal species: *Actinocrinus nobilis*, *Isoedwardsia ingolfsi*, *Bolocera marina*, *Tealidium jungermani*, *Epiparactis dubia*, *Pycnanthus laevis*, *Parasicyonis groenlandica*, *Sicyonis tuberculata*, *Sicyonis ingolfsi*, *Sicyonis caribialis*, *Davantesia praelongata*, *Sagartiogeton abyssorum*, *Sagartiogeton ingolfsi*, *Chondrophellia coronata*, *Phelliactis hertwigi*, *Phelliactis robusta*, *Paraphelliactis spinosa*, *Stephanauge axinellae*, *Amphianthus ingolfsi*, *Amphianthus verruculatus*.

(*Isozoanthus islandicus*, *Isozoanthus dubius*, *Isozoanthus multinsulosus*, *Epizoanthus abyssorum*, *Epizoanthus paguriphilus*.)

Boreal species: *Protanthea simplex*, *Edwardsia tuberculata*, *Edwardsia longicornis*, *Edwardsia pallida*, *Edwardsia danica*, *Edwardsia islandica*, *Edwardsia finmarchica*, *Edwardsia norvegica*, *Edwardsia koreni*?, *Edwardsia costata*, *Isoedwardsia nidarosienensis*, *Fagesia* (*Milne-Edwardsia*) *locni*, *Fagesia carnea*, *Paraedwardsia arenaria*, *Paraedwardsia sarsi*, *Linnactinia laevis*, *Prachia boeki*, *Halcampa duodecimcirrata*, *Antrakia mirabilis*, *Antrakia parva*, *Octineon succicium*, *Tealia felina* varieties *coriacea*, *tuberculata* and *lofolensis*, *Parasicyonis sarsi*, *Actinostola abyssorum*, *Kadosactis abyssicola*, *Sagartiogeton robustus*, *Sagartiogeton flexibilis*, *Sagartiogeton undatus* with its variety *anguicomus*, *Sagartiogeton lacertatus*, *Sagartiogeton viduatus*, *Sagartiogeton tubicolus*, *Phellia gausapata* (*Stephanauge acrilis*, *Actinauge longicornis*?) *Amphianthus norvegicus*, *Amphianthus mopsuec*, *Amphianthus islandicus*.

(*Cerianthus floppii*, *Pachycerianthus multiplicatus*, *Arachnanthus sarsi*, *Botryanthifer norvegicus*, *Isozoanthus altisulcatus*, *Isozoanthus danicus*, *Isozoanthus multinsulosus*, *Isozoanthus arborescens*, *Parazoanthus anguicomus*, *Parazoanthus haddoni*, *Epizoanthus norvegicus*, *Epizoanthus koreni*?).

Boreal-atlantic species: *Bolocera tuedae*, *Actinostola callosa*, *Actinauge richardi* (east-atlantic species).

Boreal-west-atlantic species: *Sagartiogeton verrilli*.

Boreal-lusitanic-mediterranean species (not abyssal): *Gonactinia prolifera*, *Prachia hastata*, *Anemonactis* (*Eloactis*) *muzelii*, *Actinia equina*, *Anemona sulcata*, *Sagartia troglodytes*, *Sagartia elegans*, *Adamsia palliata*.

(*Epizoanthus incrastatus*.)

Low-arctic-boreal species: *Edwardsia andresi*, *Metridium senile*.

Arctic-boreal species: *Ptychodactis patula*, *Sideractis glacialis*, *Actinauge verrilli*, *Hormathia nodosa*, *Hormathia digitata*.

(*Cerianthus floppii*? possibly only boreal; the specimens from arctic regions were so badly preserved that an examination of better material is necessary in order to decide their identity with *floppii*; *Isozoanthus ingolfsi*?, *Epizoanthus glacialis*.)

Low-arctic species: *Cactosoma abyssorum*, *Liponema multicornis*, *Pycnanthus densus*?

(*Epizoanthus erdmanni*, *Epizoanthus daniclseni*.)

Arctic, strongly marked abyssal species: *Anthosactis ingolfsi*, *Parasicyonis ingolfsi*, *Bathypheilia margaritacea*, *Kadosactis rosea*, *Amphianthus margaritaceus*¹⁾, *Kodioides pedunculata*, (*Cerianthus rogti*, *Cerianthus rothi*? - *Isozoanthus bulbosus*²⁾, *Epizoanthus roseus*).

Arctic, not, or usually not, abyssal species: *Edwardsia arctica* (also abyssal see below), *Edwardsia regae*, *Edwardsia citrea*, *Nematostella* (*Milne-Edwardsia*) *polaris*, *Nematostella nathorsti* (taken also in the abyssal region), *Arthelias intestinalis*, *Prachia*

parasitica, *Halcampa arctica*, *Halcampa? regae*, *Epactis arctica*, *Epactis nordmanni*, *Epactis marsupialis*, *Epactis incerta*, *Epactis lewesi*, *Actinostola groenlandica*, *Stomphia polaris*, *Kadosactis spetsbergensis*.

(*Cerianthus daniclseni* Roule, *Epizoanthus lovabli*.)

Panarctic species: *Haliactis arctica*, *Banodactis stelba*, *Tealia felina crassicornis*, *Cribriopsis similis*, *Banodactis spetsbergensis*, *Anthosactis jan mayeni*, *Actinostola spetsbergensis*, *Stomphia coccoinea*, *Allanactis parasitica*.

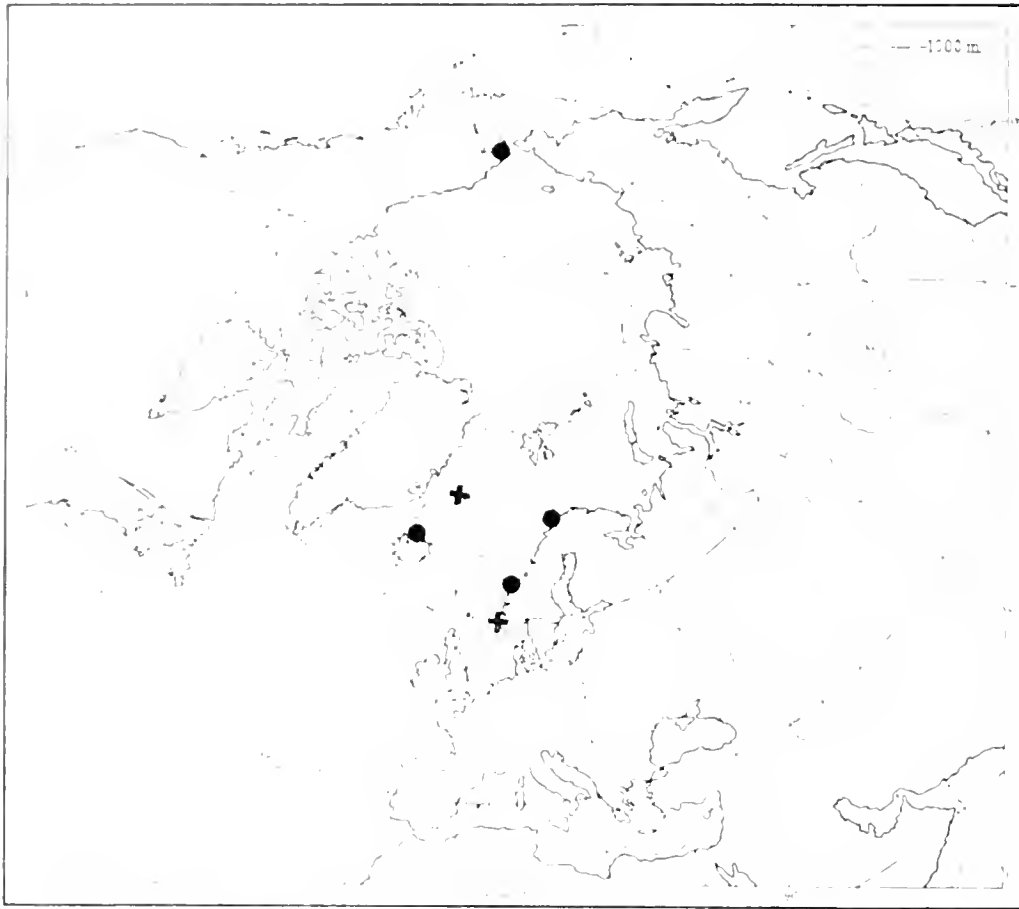
Cosmopolitan species: *Halcampoides purpurea*.

It is not my intention to discuss in detail the distribution of each species here. In the maps given by me in 1933 and in this paper (figs. 85-95) of the distribution of several species we find species showing a westerly distribution as *Actinauge cerebella* as well as an easterly one as *Hormathia digitata*. Of some interest are, however, some of the panarctic species as those living in Bering Strait and southwards. Although the Actinian fauna of Bering Strait and the northern Pacific Ocean is slightly explored I think, however, that we can draw some conclusions when comparing this fauna with that of the North Atlantic. Of the species occurring in Bering Strait or in Bering Sea or in both, some, *Cnidopus ritteri*, *Charisea saricola* and *Anthopleura xanthogrammica*, belong to the North Pacific fauna, some others are in common with those in the North Atlantic, one, *Metridium senile puberulum*, has its corresponding variety in the boreal atlantic *Metridium senile dianthus*, three species, two of them *Epactis lewesi* and *Edwardsia regae*, dredged in Bering Strait, the third, *Halcampa? regae*, taken in Bering Sea are known each from only a single locality. The panarctic species, *Haliactis arctica*, *Tealia felina crassicornis*, *Actinostola spetsbergensis* and *Stomphia coccoinea*, occur in Bering Strait as well as in Bering Sea, *Banodactis spetsbergensis* only in the former, *Cribriopsis similis* only in the latter (it is, however, possible that the specimens, being very badly preserved, belonged to the nearly related north-american *C. williamsi*), *Anthosactis jan mayeni* and *Allanactis parasitica* in their distribution (see CARLGRÉN 1933 figs. 7, 16) have not reached Bering Strait, though it is possible that they occur northward of it in colder water. To the panarctic species seem to belong also *Ptychodactis patula* and *Liponema multicornis*, the former taken in Bering Strait, the latter in Bering Sea, though they are dredged in few localities. The nature of *Epactis arctica* found only in two localities, the one north of Bering Strait, the other in the North Atlantic but in + temperature, is somewhat dubious as to its zoogeographical character. It is interesting that the boreal *Tealia felina coriacea* (or possibly *tuberculata*) occurs in Bering Sea giving support to my opinion that *crassicornis* is the mother form from which the varieties of *Tealia* provided with verrucae have originated. Very interesting also is the fact that the high-arctic abyssal *Amphianthus margaritaceus* and the high-arctic *Edwardsia arctica* occur in the North Pacific, the former in the Sea of Okhotsk at about a depth of 3500 m and at a bottom temperature of -1.52° (locality 46 11'5" N, 147 28' E.), the latter in the Sea of Japan at depth of 2300 m and at a bottom temperature of -0.18° (locality 41 38'5" N, 132 08' E.). How do we explain this distribution? As far I can see there is no other possibility than that the named species are relicts from the period in the earth development when the waters between N America and Asia were considerably deeper than now. The presence of *Ptychodactis*, *Liponema* and *Tealia felina coriacea* in the Bering Strait or in the Bering Sea indicate that these waters do not have a high arctic character. But more investigations of the waters from Bering Strait southward are necessary before we can hope to solve all zoogeographical problems concerning these waters. It is not excluded that we shall meet more surprises than those Broen found (Oktokoralen des nordhellen Pacifischen Ocean, Norsk Vid. Akad. Oslo 1935) when he found several boreal species of Octocorallia in the Sea of Okhotsk.

At the end I give figures (figs. 85-95) of the distribution of some species. As to the distribution of some other species see my paper of 1933.

¹⁾ See below.

²⁾ As to the distribution of this species and the comments on it see CARLGRÉN 1933 fig. 7 and p. 14 and 16. Because *Davantesia* has apparently transgressed the banks between Iceland and Faeroe Islands it occurs on the atlantic side in + temperatures as well as on the border to the Norwegian Sea in - temperature it is possible that the distribution of *Isozoanthus bulbosus* can be explained in the same way.



● *Vitinaria palustris* + *Vitinaria subcostalis* (taken also in Trondheimfjorden).

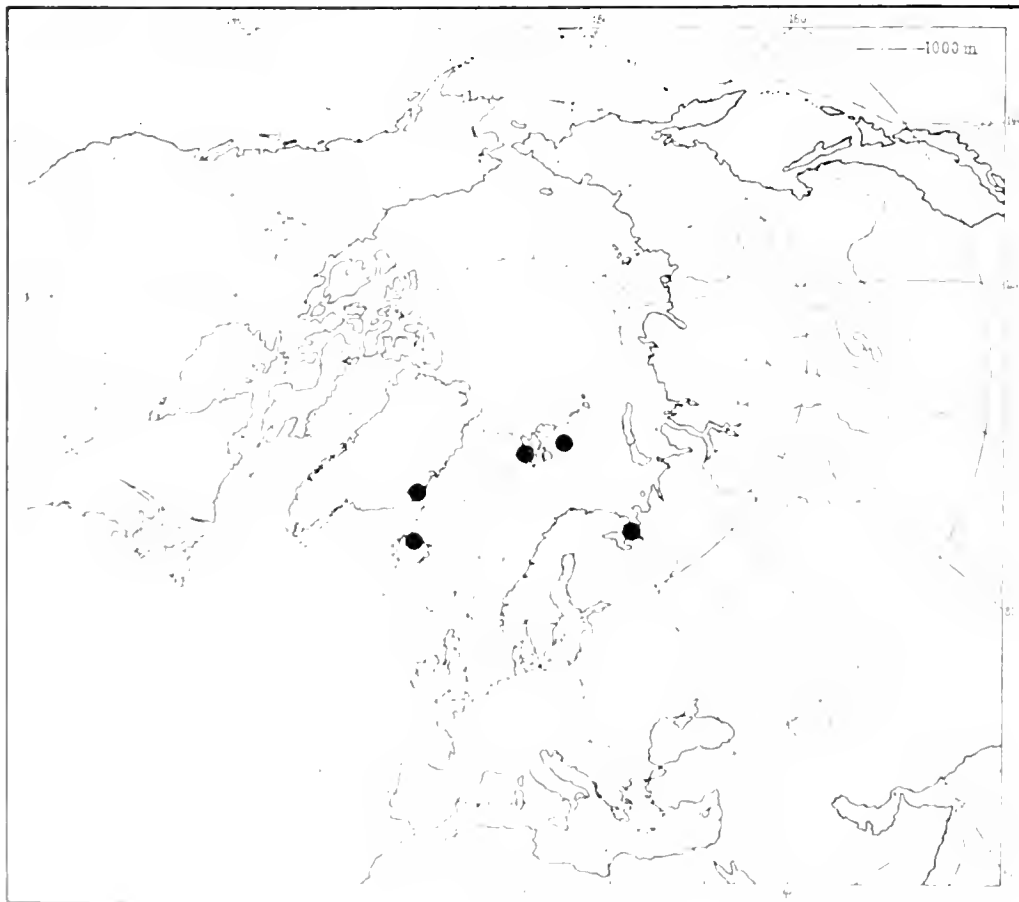


Fig. 50. *Vitinaria palustris*.

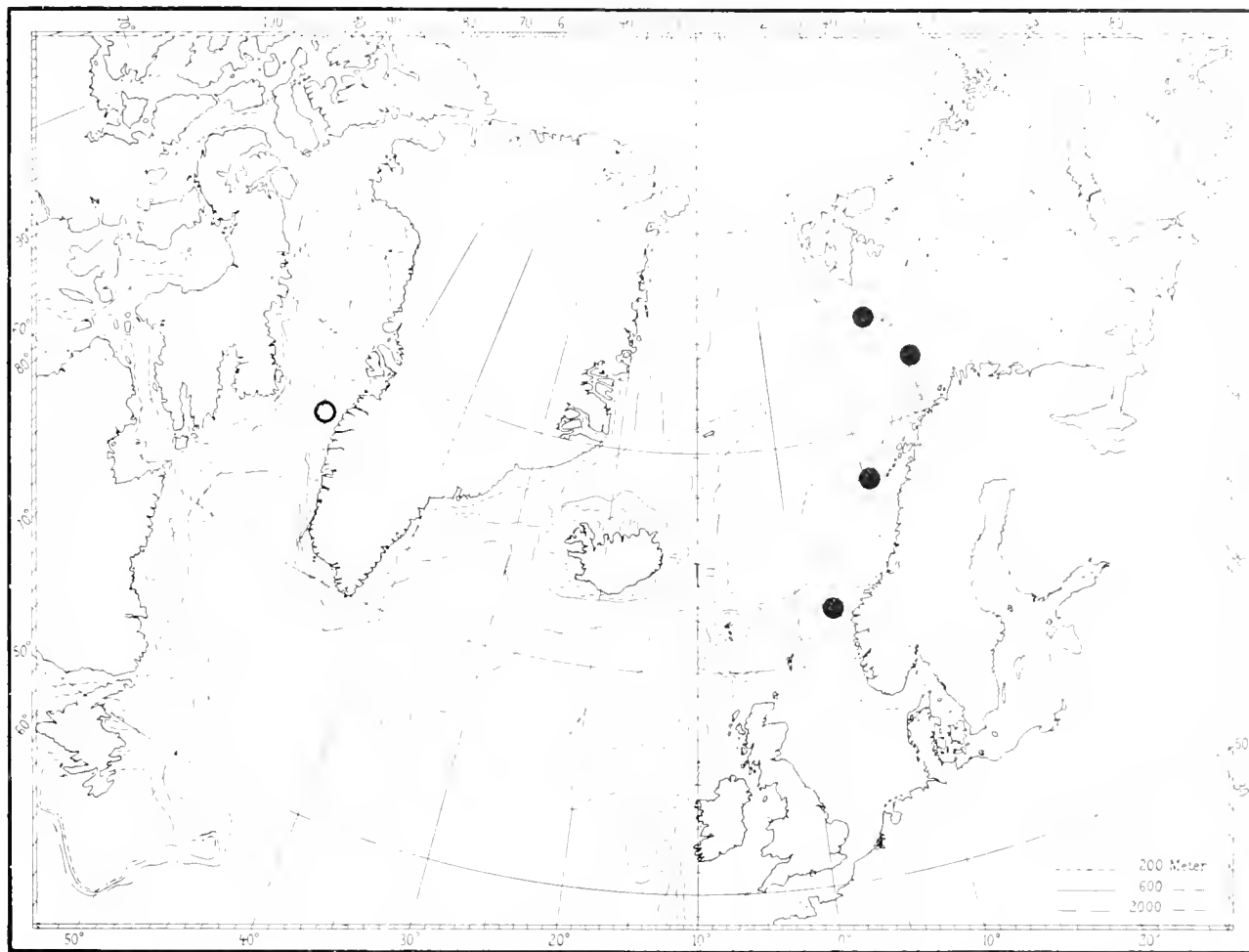


Fig. 87. *Cactosoma abyssorum*. ○ exact locality unknown.

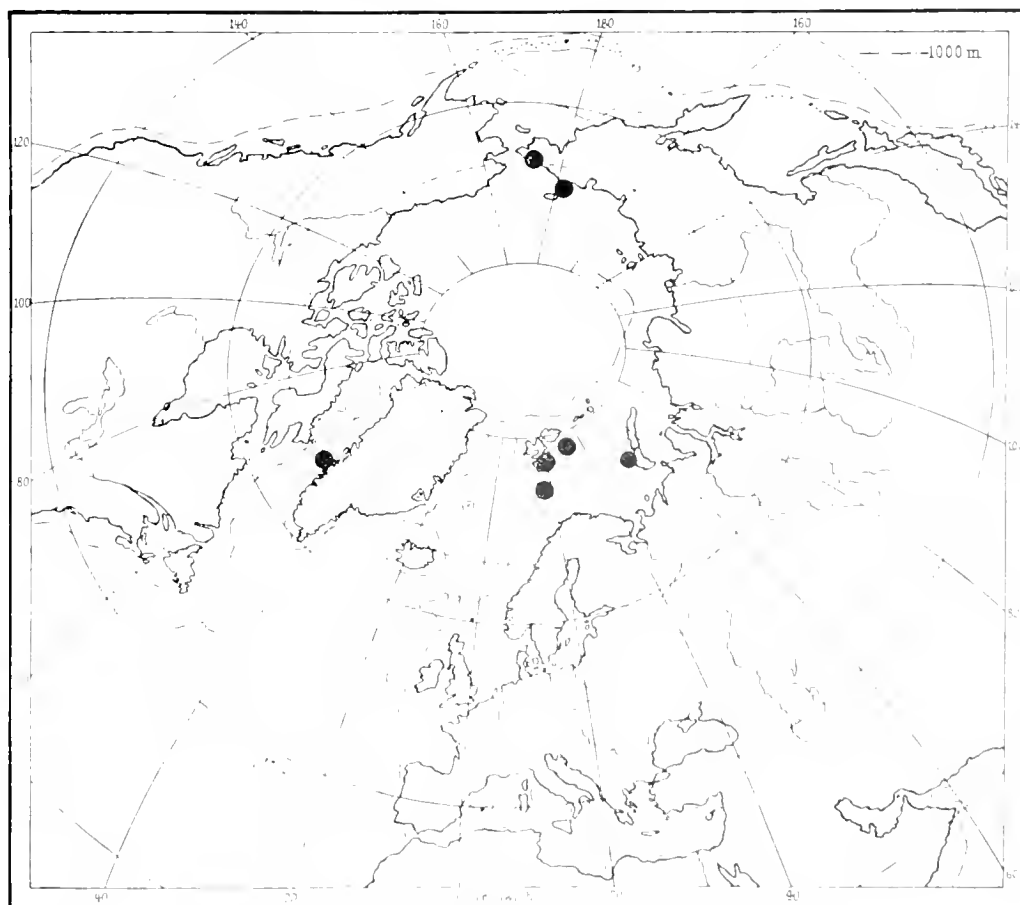


Fig. 88. *Halodis arctica*.

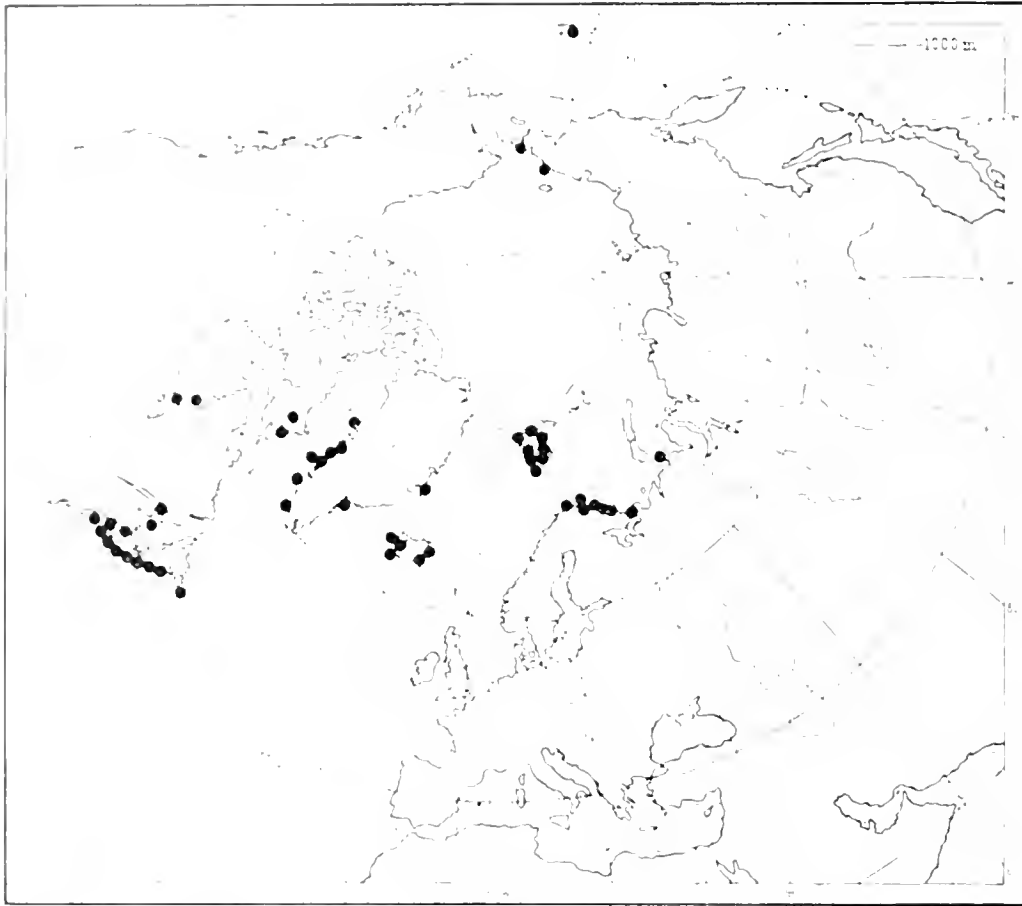


Fig. 89. *Bunodactis stella*.

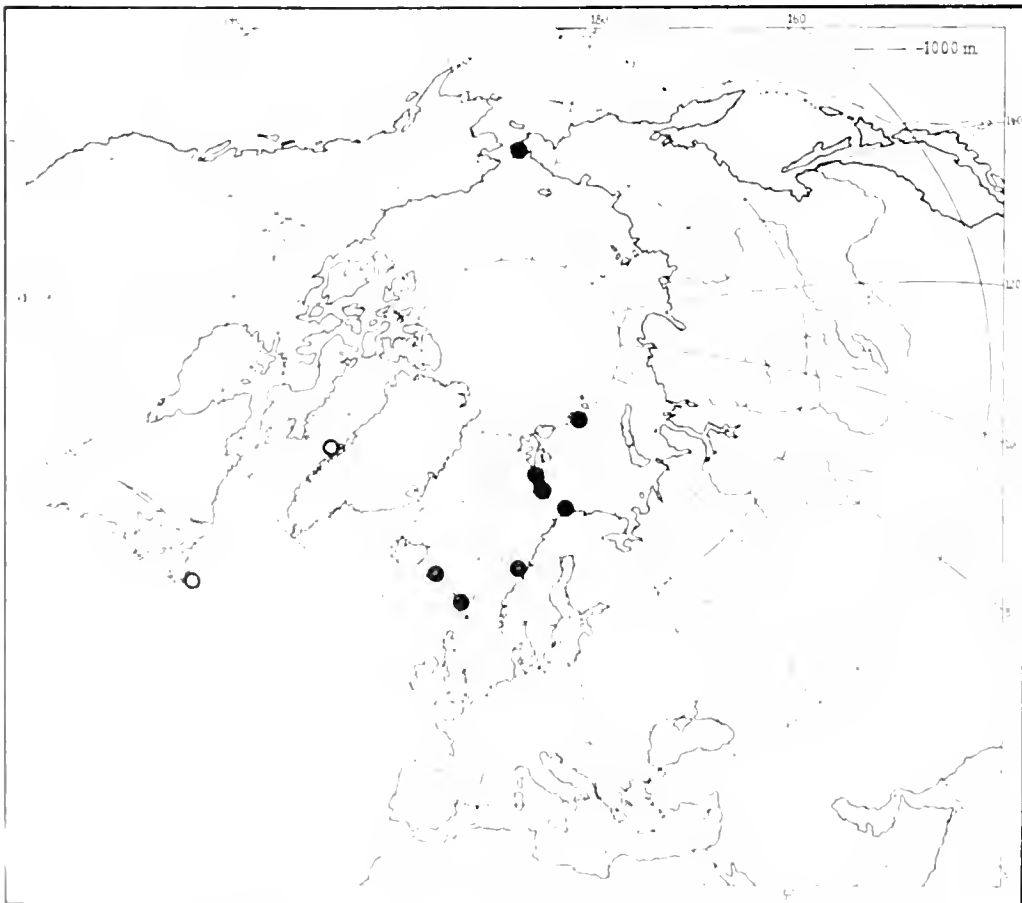


Fig. 90. *Bunodactis* sp. (●) — exact locality unknown.

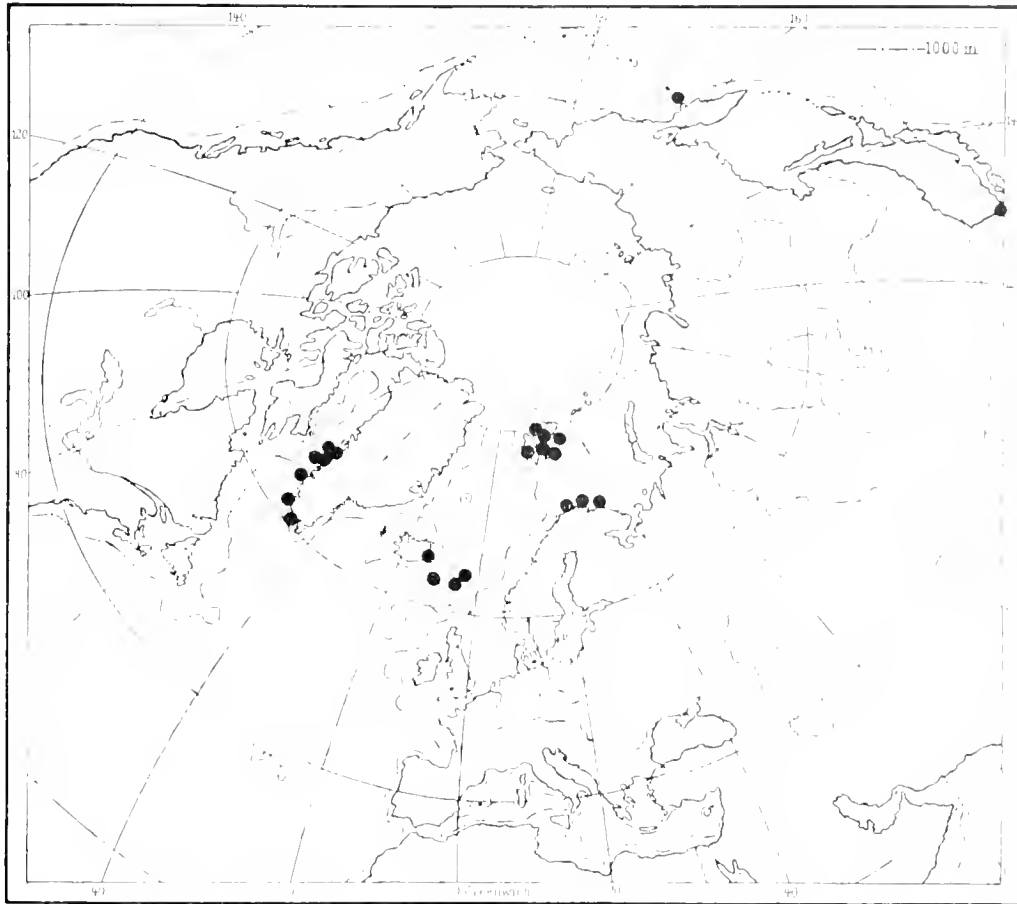


Fig. 91. *Cribrinopsis similis*.

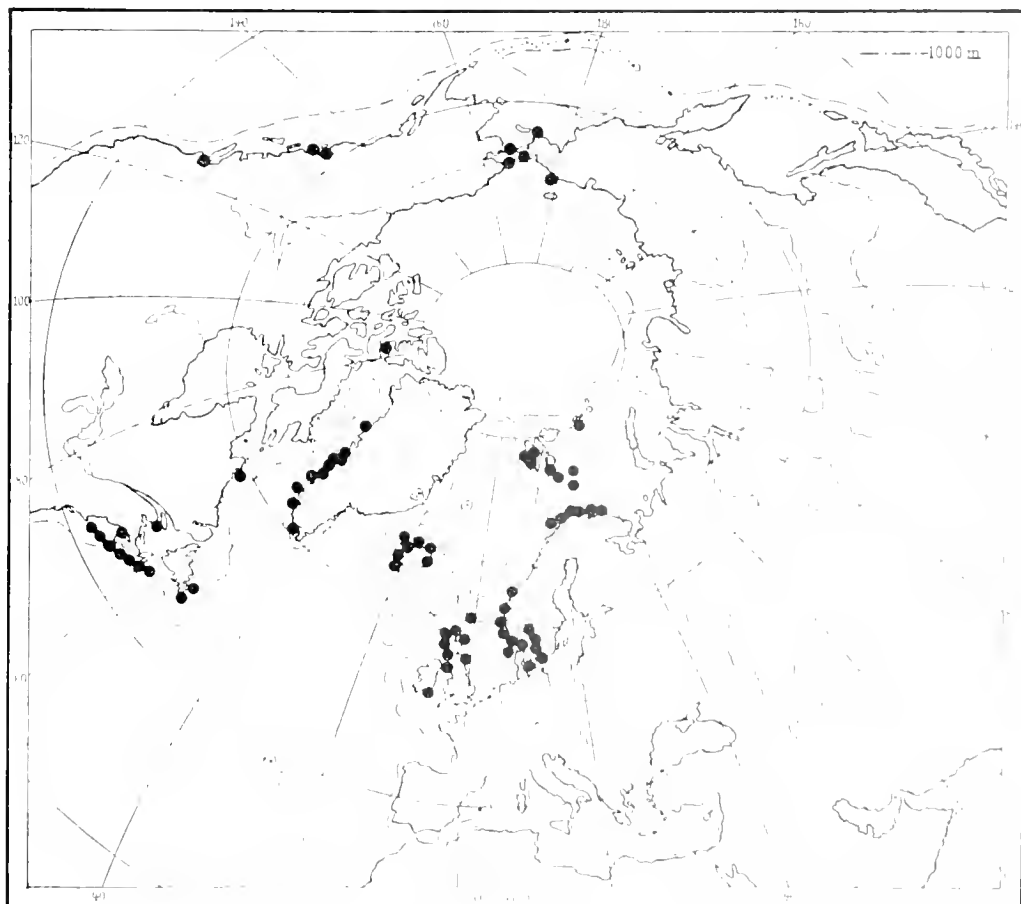


Fig. 92. *Stomphia coccinea*.

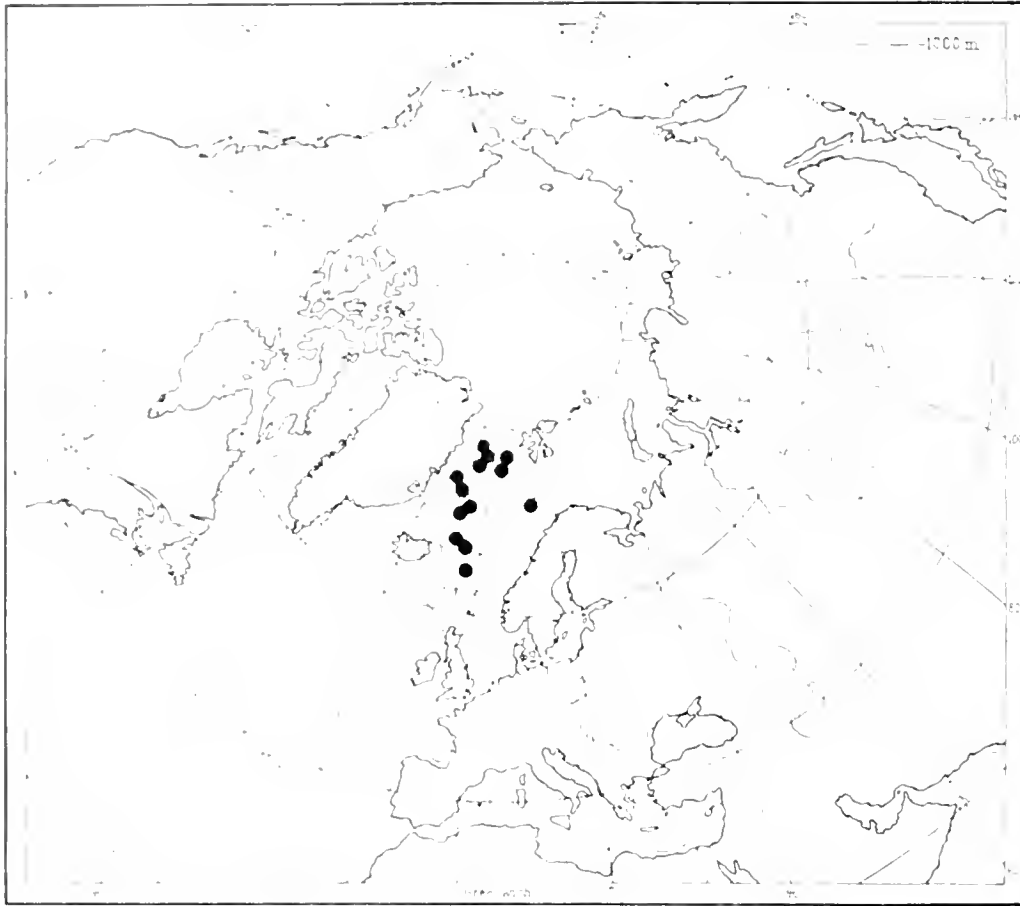


Fig. 93. *Bathypellia margaritacea*.

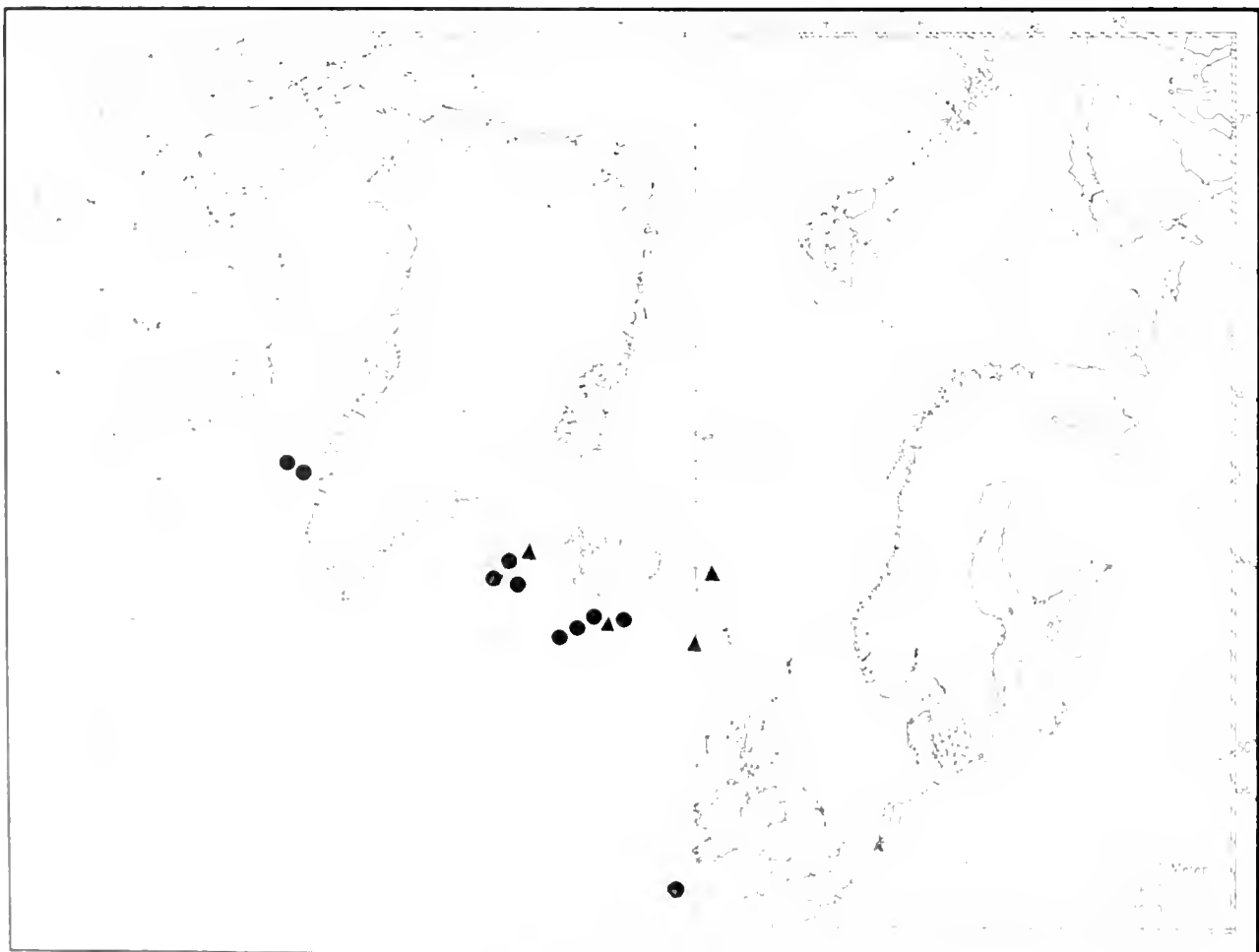


Fig. 94. ▲ *Pholichthys robusta*, ● *Pholichthys robusta*.

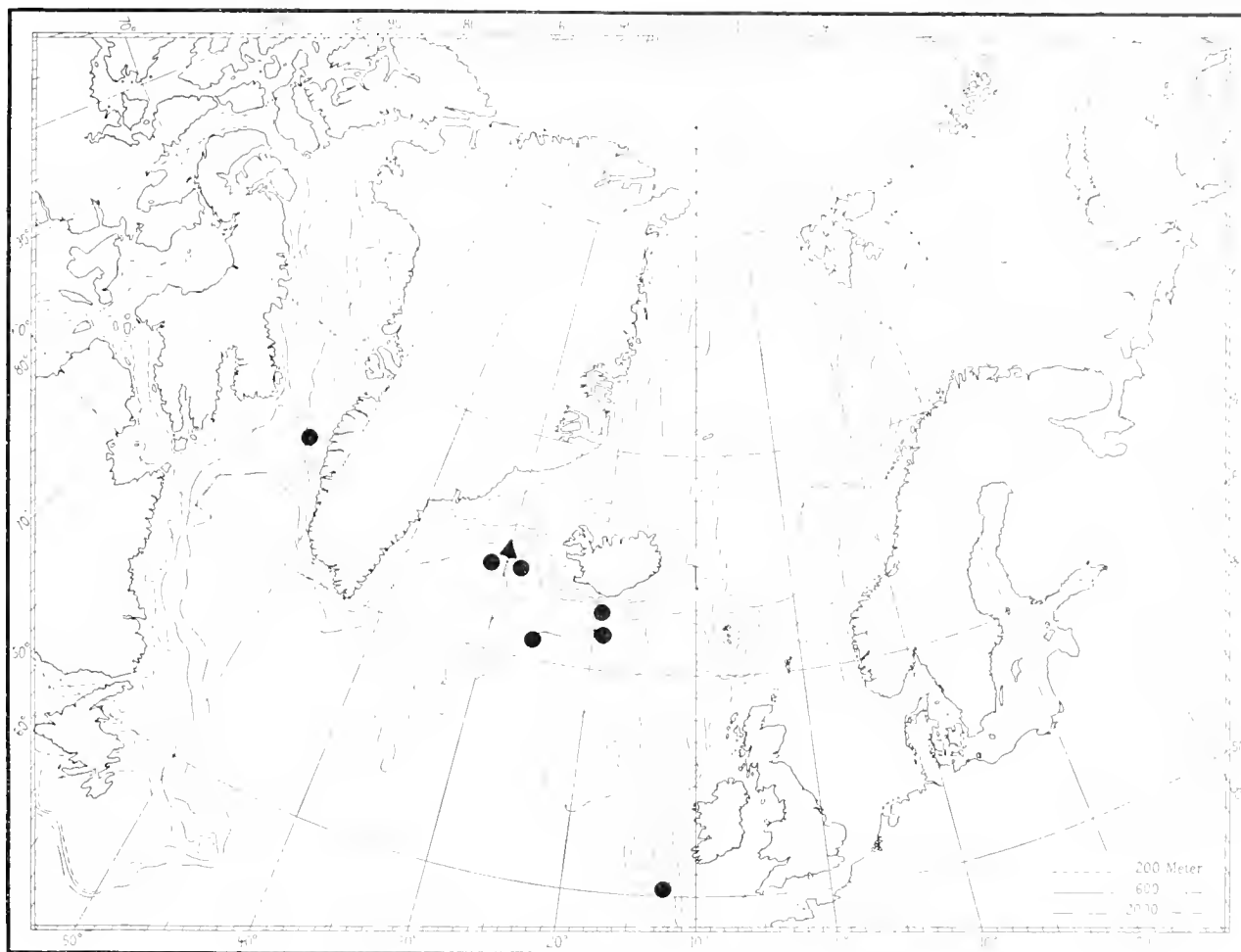


Fig. 95. ▲ *Paraphelliactis spinosa*, ● *Chondrophellia coronata*.

III. Contributions to the Anatomy, Genealogy, and Classification of the Actiniaria.

The Actiniaria belong to the groups of animals the classification of which is based on their anatomy. Since R. HERTWIG (1882, 1888) published his report on the Actiniaria of the Challenger expedition our knowledge of their organization has been considerably augmented. A short survey of the anatomy is to be found in the works of Pax (1914 and 1925) — the latter work is, however, unreliable in several points — and in those of STEPHENSON (1920-22 and 1928). Both authors have made an attempt to classify the Actiniaria. STEPHENSON has also to a certain extent dealt with their genealogy. By his classification of the Actinaria STEPHENSON (1920 p. 168) used a "new" method based on simple summation of anatomical characters. Such a method is principally usable. I had already (in 1900) pointed out that we must take into account all anatomical characters in classification — provided that we give some characters a greater value than the others, i. e. we must make a balance not only between more important and other less important characters, but also between the important characters themselves. This also seems to have been in STEPHENSON'S mind (1920 p. 470, and 1921 p. 508), when he speaks of the systematical place of *Boloceraoides*. The particulars on which STEPHENSON (1920-22) grounded his classification, are, in several points, open to criticism, for which reason I wrote some years later the following unpublished survey of the anatomy and its use in classification, a survey, however, completed here by the results of later investigations. Since then I have in some points modified my opinion as to the

use of certain anatomical details for classification, in several others STEPHENSON now agrees with me, so that our classification in the main is now the same. Although it might seem superfluous to publish this survey, I think, that it is necessary in order to make clear my own opinions as to the importance of certain anatomical details for classification, and because many anatomical facts have become known since STEPHENSON wrote his survey in 1928. A forecast of the ideas here proposed by me can be found in several of my previous papers.

The proximal part of the body is either rounded, sometimes more or less distinctly separated from the other parts of the column and forming a physa, or it is flattened, disc like, sometimes passing over into the column without any distinct boundary, and not provided with basilar muscles, which, however, are present in the more differentiated genera, of which the proximal part has formed a pedal disc, distinctly marked off from the column, in order to attach the body more firmly to the substratum.

The rounded proximal part of the body is often provided with apertures, (e. g. in *Edwardsia*, *Halcampoides*, *Halcampa* and *Pachia*) mostly arranged in one or several cycles around a central porus and surrounded by a sphincter. Sometimes, in some species of *Edwardsia*, the endoderm at the apertures forms a mobile stop (CARLIGREN 1921 pp. 50, 53). In the fertile Actinaria the ectoderm of the proximal end of the body has no radial muscles, except as it seems in *Subantus* (CARLIGREN 1921 p. 9), no, or very seldom,

columnar muscles, and in the pedal disc. The presence of a nerve layer (often provided with spirocysts) in the columnar muscle layer (see STEPHENSON 1935) is a conservative character, as well as a primitive character in the aboral end of the column. A basal disc may also have had no muscles (see STEPHENSON 1935). Actinaria is thus a primitive form. STEPHENSON (1921 p. 74) however, seemed to be of the opinion that the presence or absence of a definite basal disc, or the presence of basilar muscles, sometimes provided a key to the division of Actinaria without basilar muscles together with the presence of a basal disc (STEPHENSON 1935 p. 15). The base can vary considerably in form, from a distinct basal disc, to a basal disc-like process. Forms with a distinct basal disc are usually rounded, or flattened to a physalid-like shape, on the other hand forms with a basal disc-like process, or a rounded basal end can flatten it out to a disc-like appearance. The appearance of the basal end is sometimes not a conservative character, in opposition to the presence or absence of a basal disc. For my part I cannot place species with and without basal muscles in one family, and families having or lacking basal muscles near each other. Since the forms having basal muscles also show modes of life differing from that of the forms without basal muscles, it seems to me, that the presence or absence of basal muscles is an important character of fairly high classificatory value. I think, however, that STEPHENSON now agrees with my point of view (see STEPHENSON 1935).

The structure of the column agrees in several species more or less with that of the tentacles, inasmuch as longitudinal muscles, and a nerve layer with nerve cells are present in both, and are more or less lost in most Actinaria. As to the longitudinal muscles of the column, which are always ectodermal, they are present in genera extended over the whole column, now fairly long (*Psittichia*, *Gonactinia*, *Isocorallium* and *Boloceroidea*, *Cyathoceras*, *Psychodactylus*, *Dactyloanthus*, *Saberactis*, *Corallium*, *Cyathactis*, *Banodactopsis*). Other genera are provided with such muscles only in the uppermost part of the column. This is the case with *Thamactis*, they are well developed, and in *Necadua*, *Phylloactinia*, *Leptactinia*, *Aiptasia*, *Bartholomea*, *Caribbeacolla*, *Heteractis*, *Halicampyon* and possibly *Linnæactinia* (I am not sure, that we really have to do with such muscles).

McMURDO (1893 p. 113) has also observed ectodermal columnar muscles in *Halicurians palatus*, and FOWLER (1888) in *Leptactinia*. In the case of the former species the statement of McMURDO reports ventilation, as neither STEPHENSON nor I have observed such muscles in other species of *Halicurians* and in other related genera. Concerning *Thamactis* nothing is known concerning the structure, but I think that the ectodermal muscles here are to be regarded as the somewhat swollen bases of the ectodermal tentacles (see O'DONNELL and HERRING 1879 p. 15 Pl. III figs. 8, 9). To the columnar muscles described in several stichodactylid genera (see STEPHENSON 1900) may be placed (CARLGRÉN 1921 p. 116) and (STEPHENSON 1935) I have not found such muscles in the columnar muscles of the stichodactylids examined by DE LINDEN. The above named genera with ectodermal columnar muscles belong to the families Boloceroideidae, Psychodactylidae, Cyathocerasidae, Nematocystidae, Alciidae, Aiptasiidae, Halicampyonidae and Leptactinidae.

As to the evolutionary character of the presence of ectodermal columnar muscles, it is of the opinion, that the ancestor of the Actiniaria had such muscles. As to the occurrence of these muscles in the Actiniaria, the evolutionary workers regard it as a resurrection character, which character remained in the primitive genera of the Actiniaria, and was lost, while I in 1893, when I tentatively proposed the division of the Actiniaria, at that time only few species of Actiniaria were known, and the columnar muscles were known only in the primitive genera. The columnar muscles had genetic value, and I think they retain my original view in its

entirety. I am of opinion that the presence of these columnar muscles is a factor of fairly high classificatory value, but not to be taken alone, but in connection with other primitive characters, as already pointed out by me (1900). Also I think that the use of this factor in classification does not lead to unnatural results. Considering the families having ectodermal muscles in the whole column we find that they all belong to the tribus Protanthecae (in my original view) and to Boloceroidea, a group without basilar muscles, and, to my mind, derived from the family Gonactinidae (CARLGRÉN 1921b p. 19). As to the other genera with ectodermal muscles only in the uppermost part of the column, the question is, whether these muscles are new formations or not. Concerning the family Alciidae it may not be the case, as it is a descendant of the family Boloceroideidae, and perhaps it is the same in the Aiptasiidae, which possibly also are descended from a Boloceroideid-like form (CARLGRÉN 1921b p. 19). As to *Halicampa* the weak columnar muscles might be a new formation, but I have already (CARLGRÉN 1921 p. 117) indicated, that they are possibly prolongations of the muscles of the tentacles and of the oral disc, which is here small. I think that some transformations have taken place here, as the distal part of the small sphincter has entered into the base of the tentacles. It seems to me not impossible to suppose that the small part containing the capitular muscles originally belonged, to an earlier, somewhat wider, oral disc.

The presence of spirocysts in the body wall of many larvae (CARLGRÉN 1921a, 1931b) indicates that it is an ancestral character in the Actiniaria. Often the spirocysts occur along with the ectodermal columnar muscles, but also in genera devoid of these muscles. The spirocysts are certainly more conservative than these muscles, and are sometimes present in fairly highly organized genera, such as *Actinopsis*, and in the family Aurelianiidae. Solitary or scattered spirocysts one can find in other genera such as *Bolocera*, and in the marginal sphaerules, but usually they are lost in the column.

The column is often smooth without any differentiation, sometimes, however, divisible into a longer proximal part, scapus, and a shorter, distal part, scapulus, the former provided with a cuticle. To these an uppermost thin-walled capitulum can be added (STEPHENSON in CARLGRÉN and STEPHENSON 1928 p. 16) occurring also without the presence of a cuticularised scapus. The column of the more differentiated genera often has outgrowths of a different appearance. Their use is either as stinging organs or more passive means of protection.

Of stinging organs we can distinguish several sorts:

1) Nematophores: Batteries of nematocysts containing microbasal *b*-mastigophors, for the greater part enclosed in the mesogloea of the scapus (in several Edwardsiidae).

2) Marginal sphaerules in the most distal part of the body. They always contain atrichs, usually very numerous, but also basitrichs and spirocysts (in the family Actiniidae incl. Banodactiidae and Phyllactiidae and in the genus *Nemactis*? (Verrill 1869: 74 p. 187)).

3) Marginal pseudospherules, simple or compound, often perforated vesicula-like outgrowths, forming more or less strong nematocyst-batteries in the most distal part of the body and recalling the marginal sphaerules, but never provided with atrichous nematocysts (in several Actiniidae).

4) Hollow, simple vesicula-like outgrowths, not situated at the margin, and containing same nematocysts as in other parts of the column (in *Haloclaca* and *Dactyloanthus*).

5) Hollow, compound outgrowths provided with macrobasal amastigophors (as in the family Alciidae) or with microbasal amastigophors (as in *Banodactopsis*).

6) Spots of nematocysts, sometimes also with spirocysts, not situated in special protuberances, or forming small, not hollow, protuberances (in *Fopsis* (*Milne Edwardsia*), *Halicurians*, *Syphacurians*, *Mona*, *Pelocates*, *Halicampypton* and in the capitulum of *Milca*).

The passive protective organs of the column are:

1) Hollow verrucae, evaginations of the gastral cavity containing all germ-layers. Their ectoderm consists, in their apex especially, of supporting cells and of fewer gland cells swollen at their base but provided with a very thin efferent duct (in several Actiniidae). The verrucae as well as the formations mentioned below under 2 and 3 serve as attachment for foreign bodies.

2) Spots or small protuberances of the same structure as the verrucae, but not in connection with evaginations of the column (in the species of *Sagartia*).

3) Tenaculi ("*Halcampa*-papillae"), spots or shorter or longer elevations, in which the supporting ectoderm-cells for the most part are chitinized and connected with the cuticle. Between the supporting cells are gland cells (in *Halcampa* and other Athenaria and in several Acontaria).

4) More or less strong tubercles covered with a weaker or stronger cuticle, as in several genera provided with acontia.

All these formations on the column are adaptations to a certain mode of life and are usually of lesser classificatory value. As to the stinging organs STEPHENSON (1921 pp. 170-71) has, however, pointed out the absence of such organs in the species having mesogloal sphincter and acontia, and has, among others, taken that as an instance of a relationship between the forms with mesogloal sphincter and those with acontia. In *Pelocotes*, a species with acontia, however, stinging warts occur, but they are of simple structure, and according to information—whether correct or not I will leave unsaid—given by VERBIL (1869 71 p. 487) the Sagartiid *Nemactis* has marginal sphaerules. Be this as it may, there are other factors against the idea that the species having acontia, and those having a mesogloal sphincter but no acontia, make a genetic whole.

The column of the Actinaria often has apertures, cinclides, corresponding to the apertures of the physa in some genera (p. 78) and provided with a weak sphincter. The cinclides, in some cases principally ectodermal invaginations, in others endodermal evaginations, either only of one sort in a species or of both, are safety-valves against rupture of the column at strong contraction of the body, and widely distributed in the Nyantheae and here present both in several Actinians provided with acontia and in the family Actiniidae (in a wider sense)—the marginal sphaerules and the pseudospherules are often provided with a cinclis—but also in some Athenaria as in *Hareuactis* and *Eloactis*. In the Thenaria with mesogloal sphincter but without acontia, they seem to be lacking. As they are present both in those species having as well as in those lacking acontia, it is evident that they stand in no near relation to the acontia (STEPHENSON). Generally they occur in thin-walled species, sometimes also in thick-walled as in *Stephanauge* and *Amphianthus*, but here reduced in number and at least as a rule only present in the directive compartments, more sparingly in species the column of which has a cuticle (as in *Kadosactis*). That a reduction of cinclides takes place in thick-walled deep-sea species having acontia is very probable—in one species, *Kadosactis* (*Kadosanthus*) *sulcata*, taken by the "Michael Sars" expedition at a depth of 1215 m, the cinclides are arranged in an annulus at the limit between the scapus and scapulus, but their duct is irregular in its course and probably undergoing reduction; on the other hand the cinclides of *Kadosactis* (*Sagartiopecton*) *antarctica* taken at a depth of 1636 m show no traces of reduction—a difference possibly connected with the fact that the mesogloea of *K. sulcata* is considerably thicker than in *K. antarctica*, but whether all thick-walled deep-sea species having acontia formerly had cinclides, which have later been lost, is very questionable. However this may be, the presence or absence of cinclides is of little classificatory importance.

The circular muscles of the uppermost part of the column are often differentiated into an endodermal or mesogloal sphincter. The endodermal sphincter, transformed into a mesogloal when passing through the mesenteries—the term endodermal sphincter

referring only to the parts of it situated between the mesenteries—is sometimes weak, usually well developed, either diffuse or circumscript, in the latter case either palmate having several or no main-lamellae, or pinnate provided with only one main-lamella. Especially the more strongly circumscript sphincter shows a tendency to be mesogloal next to the mesenteries, inasmuch as the muscle-folds sometimes fuse together in the interior of the sphincter, and also the diffuse sphincter shows a similar tendency when forming a so-called aggregate sphincter (McMURRIEN 1893 p. 152). For these reasons, and from a theoretical point of view, there is nothing to hinder the supposition that an endodermal sphincter can be transformed into a mesogloal one, especially in a species where the mesenteries in their uppermost part are thickened, as in the genus *Oractis* (CARLGRÉN 1931 p. 45). On the other hand a reduction of a mesogloal sphincter to an endodermal one, or to no sphincter at all, has never been stated. PAX'S (1936 p. 86) assertion that the sphincter of *Dadumene* is reduced is only a vague presumption. The endodermal sphincter occurs in several Corallimorphiae (Protostichodactylinae), among the Athenaria in *Oractis* and *Andresia*, and in many families of the Thenaria. As such a sphincter is present in the Corallimorphidae and Discosomidae—a group which was certainly differentiated very early from the other Actinaria—and occurs also in certain Zoantharia such as the Madreporaria, the suborders nearest related to the Actinaria, and having a common ancestor with 8 mesenteries, arranged as the perfect mesenteries of *Edwardsia*, there is no doubt that the endodermal sphincter in the Actinaria also has at least a diphyletic, but probably a polyphyletic, origin. At least it ought to be remarked that the endodermal sphincter is sometimes so weak that it may be questionable whether it really exists, or is only undifferentiated circular muscles, that is to say no sphincter at all.

The mesogloal sphincter is generally alveolar, with scattered muscle-meshes and fibrillae, or it may be reticular. Rarely it is very weak, and consists of a single row of meshes formed, as it seems, by endodermal folds growing together and situated immediately outside the endodermal circular muscle, as in *Aiptasia couchii*, sometimes it is fairly weak, but it is usually strong. Often it is separated from the endodermal muscles of the column by a thinner or thicker band of mesogloea, sometimes the sphincter enters without distinct boundary into the circular muscles of the column. Sometimes it shows a more or less distinct tendency to transverse or longitudinal stratification. In rare cases there are two sphincters, either situated not far from each other, as in *Haliathanella*, or the lower sphincter has been developed at a distance from the upper one, as in *Mena*. The mesogloal sphincter never occurs in the Protantheae, s. str., nor in the Corallimorphiae, Ptychodacteae, and Endocoelelantheae, but in some Athenaria, in the families Halcampidae, Octineonidae, and Andwakidae and in the Mesomyaria and most Acontaria.

There is no doubt, I think, that the mesogloal sphincter arises partly from an undifferentiated circular muscle layer of the column, partly from a more or less differentiated endodermal sphincter. The sphincter of *Aiptasia couchii* e. g. is certainly developed in the former manner, and possibly this is the case in many species showing a distinct longitudinal stratification of the mesogloal sphincter. Here we must anticipate that the formation of the longitudinal bands of muscle meshes has taken place independently at different times, each time through a growing together of the folds of the endodermal muscles followed by an increase of the mesogloea in the area of the development. I have pointed out above that the endodermal sphincter sometimes shows a tendency to be somewhat mesogloal. In the section of the aggregate sphincter of *Myonanthus ambiguus* (McMURRIEN 1893 p. 152 PL. XXII fig. 23), if not taken close to a mesentery but more or less in the middle between the mesenteries, we have already here a mesogloal sphincter in being. At least some of the mesogloal sphincters with distinct transverse stratification have certainly arisen from an endodermal sphincter as in *Isosagamis alba* (CARLGRÉN 1927

1928a, p. 172, fig. 26). On the other hand, as I have mentioned above, and as I have pointed out elsewhere, in connection with the fact that in the Zoanidae the mesogloea is provided with either double or single mesogloea, and in the Actiniaria on the other side with the origin of mesogloea in the ectoderm of the tentacles and in the mesogloea of the oral disc (Actiniaria (see below p. 80), we must not forget that the genera provided with a mesogloea sphincter (see below p. 82), wide in other words the mesogloea sphincter, are provided with it in different stages during the evolution of the Actiniaria. As far as I can conclude STEPHENSON (1921, p. 44) supposed at least a diphyletic origin of the mesogloea sphincter, when he placed together *Haliuapoa*, with its mesogloea sphincter, and *Haliampoles* with no sphincter. From a phylogenetic point of view I think that STEPHENSON'S division (1921, pp. 341-342) of the Thelamnia into Endomyaria and Mesomyaria is unfounded, although for the present, the division can be used to a certain extent for classification (see below p. 85-86). Still, it is as in the case with PAX'S division (1925, p. 181-820, 1926) of the Actiniaria, apart from the Endocoelactaria and Stichodactylina into Endomyaria and Mesomyaria—it is only a simple scheme of examination without any traces of an attempt at a division founded on the phylogeny of the Actiniaria.

Almost all Actiniaria are provided with tentacles, only the genus *Limnactinia* lacks them, probably in connection with its mode of living. In some Corallimorphinae, especially the genus *Discosoma*, the tentacles are much reduced and raise themselves hardly above the surface of the oral disc. All other Actiniaria described as lacking tentacles certainly possessed them, but either they have been thrown off or been invaginated into the coelenteric cavity.

The development of the first 12 tentacles takes place mostly after a bilateral type, seldom after a biradial as in *Peachia* and some other Athemaria (CARLQVIST 1904, p. 544). Usually a single tentacle stays in communication with each endocoel and exocoel; in most Corallimorphinae (Protostichodactylinae) and in the so-called Stichodactylinae, however, the tentacles are arranged in radial rows, at least in the parts corresponding to the endocoels. The arrangement with a single tentacle from each endocoel and exocoel is more primitive than the arrangement in radial rows. In the primitive Corallimorphinae, *Sideractis* and *Nectactis*, there is also only one tentacle to each endo- and exocoel, and in the genus *Aethapsos*, among the Stichodactylinae, we meet a species of the stichodactylina type of which is but slightly indicated. The origin of the stichodactylina arrangement of the tentacles is, in the Actiniaria at least, diphyletic, as the Corallimorphinae (Protostichodactylinae) and the Stichodactylinae are so different in their organization, that we must suppose that the stichodactylina arrangement is a parallelism. Such an arrangement of the tentacles is not met, however, in the Ceramtharia.

The tentacles are simple, smooth, more seldom dendritic or branched (with bifurcated or pinnate knob-like elevations or transverse ridges). A special form of tentacle is the capitate. Usually the tentacles are specially differentiated stinging batteries, but they may be of other kind, of course, than those in the other parts of the body. So, for example, in *Thalassianthus* and *Heterodactyla*, the tentacles differ in such a way, as to form hemisphaeric stinging batteries, carrying the marginal sphaerules in their appearance and in their stinging apparatus. Also in the simple tentacles the tentacles may be collected in spots, e.g., in *Banodeopsis* (see below p. 80) and in *Actinia*. Usually the nematocysts are most numerous in the distal part of the tentacle. Sometimes the stinging capsules are of a special kind, in the catch tentacles of *Dendrocygus*, lobed and atrichs, in the ordinary tentacles of the genera *Microbase*, *Amastigophors*, in the tentacles of the genus *Strophodon*, *microbase*, *Amastigophors* (see below p. 89). In a few genera, *Aethapsos*, *Tealulana* and *Haliuapoa*, the tentacles, at their bases, are provided

with a stinging battery with specially large microbasic hemastigophors. Sometimes the mesogloea of the tentacles is more or less thickened aborally at the base as in *Actinotage*, in many Actinostolidae, and in some species of the genera *Kubosactis*, *Banodeactis*, *Phelliactis*, *Amphianthus* and so on. All these differentiations usually have only lesser classificatory value, but in certain cases, and in connection with other characters, show a near relationship between certain genera. The ectoderm of the tentacles usually contains numerous spirocysts. In the genera *Eloactis* and *Haliuclara* the spirocysts seem wholly, in *Discosomidae* almost, reduced.

The longitudinal muscles of the tentacles vary much as to their place in the tissues. Often they are purely ectodermal, sometimes more or less mesogloea viz. meso-ectodermal or ecto-mesogloea, sometimes exclusively mesogloea. Of two closely related genera one may be provided with ectodermal, the other with more or less mesogloea tentacle muscles. Exceptionally, as in *Tealia filina*, the muscles vary from ectodermal to almost mesogloea. The more or less mesogloea muscles of the tentacles occur in different families having an endodermal, a mesogloea or no sphincter, as in Actiniidae (in the wide sense), in Actinostolidae and in Condylanthidae. From the above mentioned facts it is clear that the mesogloea muscles of the tentacles have arisen here and there in the more highly differentiated Actiniaria, and that forms having them do not all have a common ancestor, in other words the origin of the mesogloea muscles of the tentacles has run parallel in several groups of the higher Actiniaria.

The oral disc of the Actiniaria shows only little differentiation. Exceptionally it is provided with small papillae representing small nematocyst batteries as in *Dofleinia*. In the genus *Limnactinia* the ectoderm of the oral disc is strongly thickened. The place of the radial muscles varies in a manner similar to that of the longitudinal muscles of the tentacles. These more or less mesogloea radial muscles of the oral disc occur in the more differentiated Nyantheae, in species with or without acontia, as well as in species with an endodermal or a mesogloea sphincter. Thus there is no reason to anticipate that the species provided with radial muscles more or less enclosed in the mesogloea are descendants of a common ancestor, having a tendency to develop such muscles. The oral disc generally has radial furrows corresponding to the insertions of the mesenteries. Here these muscles are usually weaker than in other parts of the oral disc, but in the genus *Phelliactis* the weakest muscles are over the endocoels. A singular case it met with in *Bathylactylus*, in which the ectodermal radial muscles at the insertions of the mesenteries are enclosed in the mesogloea (CARLQVIST 1928a, p. 172, fig. 26).

The actinopharynx usually agrees with the column in its structure. A weak longitudinal muscle layer is not seldom present, and more usual than in the column. Commonly the actinopharynx has two siphonoglyphs combined with two pairs of directives. In several genera, however, there is only one siphonoglyph, apparently always a ventral one if the specimen has arisen by sexual reproduction. Sometimes it is very weak as in *Edwardsia*, sometimes very strong as in *Peachia*, *Eloactis*, *Oractis* and some other Athemaria. Several genera have no distinct siphonoglyphs. Rarely, as in *Thalassianthus* and *Reetia*, there are several siphonoglyphs not joined to directives. In case of asexual reproduction of a specimen the number of siphonoglyphs varies. If there is only a single siphonoglyph present, as far as we know it is a dorsal one. The siphonoglyphs are often aborally prolonged, and reach farther down than the other parts of the actinopharynx. In *Peachia* the single siphonoglyph is orally prolonged in a more or less lobed conchula.

Since in the primitive Actiniaria the siphonoglyphs are indistinct or weakly developed it is to be supposed that the ancestor of the Actiniaria had no siphonoglyphs (or possibly a very weak ventral one).

The development of the first 8 mesenteries usually takes place more or less simultaneously, the ventrolateral mesenteries, however, apparently, always appear earlier than the others. In the

Edwardsia-stage, provided with only 8 mesenteries, the longitudinal muscles of which are characteristically arranged, no recent Actinian remains, as all species belonging to the family Edwardsiidae have, in addition to their 8 perfect "*Edwardsia*"-mesenteries, at least 4 very weak mesenteries in the uppermost part of the column. These 4 mesenteries, arising in the lateral and ventrolateral compartments as couples, grow, reach in most Actiniaria the actinopharynx, and form pairs with the 4 lateral "*Edwardsia*"-mesenteries. From this stage, the so-called *Halcampula*-stage (VAN BENEDEK), *Halcampoiles*-stage (STEPHENSON), characterized by the presence of two pairs of directives and 4 common pairs of mesenteries, or perhaps in a small set somewhat earlier, before the lateral mesenteries have been arranged in pairs, the origin of new mesenteries differs. In this group, the Endocoelelantheae, the new pairs, the muscles of which are orientated as in the directives, arise in the much enlarged endocoels, either cyclically or bilaterally, in numerous or few developmental zones (CARL GREX 1918). Judging from the arrangement of the mesenteries and their muscles in the family Minyadidae (CARL GREX 1894 p. 22) the development of the mesenteries has probably taken place in a similar manner. All other Actiniaria establish the new pairs in the exocoels as common pairs with their longitudinal muscles facing inwards, as a rule cyclically, but in the genus *Polysiphonia* bilaterally in a few, 12 or somewhat more, developmental zones, but not until 12 or a few more pairs have originated (CARL GREX 1918 p. 13). Because of the manner of the development of the tentacles, both mesenteries of a pair probably do not usually (never?) appear quite simultaneously in the distal part of the body; this is especially conspicuous in such forms as the Endocoelelantheae, *Polysiphonia* and some other forms, in which the two mesenteries of a pair differ in size. Also in forms with a richer development of mesenteries in the basal part than in the distal part, the mesenteries of a pair may arise at different times, as in *Stomphia*, *Actinostola*, *Sicyonis* and others. In the Actiniaria the mesenteries grow either from the oral side towards the base, or the reverse, or from the oral as well as from the aboral end more or less simultaneously. Through arrest and differential growth of the mesenteries a pentamerous or octomerous arrangement instead of a hexamerous comes about in some species. Rarely, as in *Thalassianthus* and *Rivettia*, the directives are absent. As the development of the mesenteries in these genera is unknown, we cannot at present clear up the origin of the radial symmetry. It is hardly to be supposed that this symmetry has arisen here by asexual reproduction, which has, in many cases, caused very different symmetries and irregularities of the mesenterial arrangement, symmetries in many cases identical with those arising sexually, but being of no importance for classification and for the study of the phylogeny.

All the species of Actiniaria, the ontogenetic development of which is known, pass through an *Edwardsia*-stage, and all species having only these mesenteries perfect have been considered as more or less primitive forms. In fact these species often show other fairly primitive characters, only a few forms such as *Halcampa*, *Octineon*, and *Aiptasia* are provided with a mesogloal sphincter, the two latter also with acontia, indicating a more advanced stage. Although many forms, having only the "*Edwardsia*"-mesenteries perfect, are without doubt primitive, it may be possible that in some of them this stage has arisen secondarily. I am especially thinking of such a species as *Halcampa duodecimcirrata*, which seems at least sometimes to be sexually ripe at a stage with 8 perfect mesenteries, but which later reaches the *Halcampula*-stage with 6 perfect pairs. Either this is the beginning of a reduction of the mesenteries¹⁾ or the reproductive period has been advanced to an earlier stage in the life of the animal (CARL GREX

1903 p. 23). However this may be, the Actiniaria having only the *Edwardsia* mesenteries perfect, form no genetic whole. The Gonactiniidae, and the above mentioned species belonging to different families of the Athenaria, and an *Aiptasia*¹⁾, which remain in the *Edwardsia*-stage, show in other characters little or no relation to each other. Thus such a tribus as the Protactiniinae (McMURRIE), based on the presence of only 8 perfect mesenteries, is invalid.

As to the reason why in one case the development of mesenteries takes place in the endocoels, in the other in the exocoels, we know nothing. Moreover we cannot make clear the real cause of the origin of the octomerous or decamerous arrangement, though the transformation of a hexamerous type into a decamerous one is known in a few cases. Sometimes a great increase of certain tentacles seems to influence the arrangement of the mesenteries. Most conspicuous is this in *Neradna*, where a strong growth of the third or fourth order of tentacles seems to cause a doubling of the mesenteries of the third or fourth order (CARL GREX 1925a, PANIKKAR 1937). Also the bilateral development of the mesenteries in *Polysiphonia* and *Actinornis* is probably related to irregular growth of the tentacles (CARL GREX 1918 p. 14, 15). A similar correlation between the origin of the mesenteries and that of the tentacles we find in laceration pieces, where a great increase in size of a directive tentacle may give rise to geminates (double-animals) (CARL GREX 1901a).

The longitudinal mesenterial muscles situated on one side of the mesenteries are sometimes weak, usually more or less strong and forming diffuse, concentrated or circumscribed²⁾ retractors or pennons. As I have previously pointed out (CARL GREX 1911 p. 15), the strength of the retractors is mostly correlated with the size of the animal. Very elongated Actiniaria as a rule have few mesenteries and strong retractors, usually limited to the macrocnemes, broader species are provided with more numerous mesenteries and mostly weaker retractors. But also in some genera with broad bases, as in *Aureliana*, *Condylanthus*, very strong retractors may appear. A weak longitudinal muscle layer in the mesenteries is more primitive than the presence of a distinct retractor. As I have explained already (1894a p. 89), the ectodermal muscles, present in the column of the most primitive Actiniaria, must be supposed to have been reduced in connection with the appearance of a stronger muscularity in the mesenteries. Nothing contradicts such a suggestion, as almost all species provided with longitudinal muscles in the column have weak longitudinal muscles in the mesenteries. In the few exceptions from the rule the longitudinal columnar muscles are reduced in most part of the column and are only present in the uppermost part of it. Regarding the mesenteries and their longitudinal muscles we can distinguish two types (STEPHENSON 1920 p. 156), one, in which the mesenteries are differentiated into macrocnemes and microcnemes, and another without any such differentiation. In the first type only the macrocnemes, the perfect mesenteries, are provided with retractors, filaments and reproductive organs, in the second there is no distinct difference between the mesenteries, at least as far as the presence of retractors and filaments is concerned. Sometimes, however, transitions between the two types occur. Although STEPHENSON (1920) admits, that the mesenteries of the Actiniaria originally had a weak musculature, he suggests that in the more highly

¹⁾ The small oral disc with the few, broad, and densely packed tentacles, makes it possible that a reduction of the size of the oral disc has taken place here. In spite of the fact that the mesenteries of the second cycle are present in the whole length of the body, there are never any tentacles corresponding to this cycle (CARL GREX 1911 p. 23).

²⁾ I have previously (1911 p. 21) suggested that the *T. ovalis*-stage sometimes occurring in sexually ripe *T. ovata* could be a result of asexual reproduction. Although such a supposition is not quite to be excluded, it is, however, hardly probable, because a binodal development of the mesenteries has been described in the case of laceration-pieces of *Aiptasia* (CARL GREX 1904).

³⁾ A real circumscribed retractor (resp. sphincter) is, to my mind, present only when either the more or less numerous folds of the mesogloa issue from a single main-lamella, as in *Octineon* and *Aureliana*, or the outer and inner lamellar part of the mesenteries are attached to the retractor very close to each other; in other cases, when the retractor in cross-section has a more or less kidney-like appearance, we may rather speak of a concentrated retractor.

1920, 1921, 1922, 1923, 1924, 1925) stage, and a professor (STEPHENSON, 1920, 1921, 1922, 1923, 1924, 1925) with the increase of the complexity of the structure of the mesenteries is taken into account. It is not surprising (STEPHENSON, 1922) and that there is a transformation of the circumscript retractors to become diffuse retractors (STEPHENSON, 1920, 1921, 1922, 1923, 1924, 1925). The further development on the part of the mesenteries, the loss of these special retractors and the development of a larger number of diffuse retractors (STEPHENSON, 1920, 1921, 1922, 1923, 1924, 1925). As I have previously stated (1911, p. 15, 1924b, p. 19), there is a special type of retractors (see also a theory). It must be remembered that among the more or less primitive Actiniaria, the Protantheae, the Corallimorphae and the Ptychodacteae, as well as among the more advanced, the Bologrodaria, no species, except those of the genus *Halicampus*, has stronger retractors, on the contrary, the longitudinal muscles of the mesenteries form no or very weak retractors. Concerning the genera having the strongest proper retractors, such as *Octocorva*, *Cordylanthus*, *Paraphyllia*, *Leclercia*, and *Actinoporus*, the first I have a wide basal part of a considerable number mesenteries in the proximal than in the distal part of the body. *Octocorva* has 8 mesenteries perfect, *Cordylanthus* 12, *Paraphyllia* 12-24, and *Actinoporus* a considerable number. That *Octocorva* and *Cordylanthus*, in spite of their numerous mesenteries, have retained the strong retractors on the few perfect mesenteries is easily understood, as only the perfect mesenteries and the mesenteries attached to the inner part of the oral disc can move the distal part, but why in *Paraphyllia* the circumscribed retractors of the 12-24 perfect mesenteries have been retained, and why in *Leclercia*, and still more in *Actinoporus*, which latter has all mesenteries perfect, these retractors have not been transformed into diffuse retractors, is difficult to understand. STEPHENSON (1920, p. 541) explains the retention of the circumscribed retractors in *Paraphyllia* in the following manner: "*Paraphyllia* generally lives on a sandy bottom, and it may need the powerful retractors to help the sphincter to withdraw the tentacular crown very suddenly to avoid its being bitten off by hunting fish in search of such things, or molluscs emerging foot-foremost from the sand. Thus, or some similar reason, would account for retention or redevelopment of the circumscribed muscles". I think this explanation rather unsatisfactory. Moreover it must be remarked that the genus *Paraphyllia* also lives on other objects than sand. I have examined a species of this genus, possibly identical with *P. papillata*, taken on Gorgonids, *Dendrophyllia* and other objects in Morocco. Can it be probable that the circumscribed retractors have regenerated when new mesenteries in a greater number are present in a species? There is, I think, nothing in favour of such a supposition. On the contrary there is much to be said for the supposition that the evolution of the longitudinal muscles of the mesenteries has gone from weaker to stronger muscles in connection with the form of the body and the mode of life.

The parietobasilar muscles, in the genera lacking basilar muscles, have been developed in a different way. In the not elongated forms, with a broad base, Protantheae, Corallimorphae and Ptychodacteae, as well as in some Athenaria, they are very weak, in the elongated genera of the Athenaria strong, forming, with the longitudinal part of the longitudinal mesenterial muscles, a more or less strong sphenoid muscle, the function of which is to contract the body, the contraction of which cannot be performed by the other muscles, which are separated from the parietal muscles by the oral disc, and from the proximal part of the mesentery. Sometimes, in the elongated forms, the parietal muscles are more or less prolonged into the oral disc. In the broad Actiniaria the parietobasilar muscles are very weak, in the elongated forms often forming a distinct projection, which is not so much, as they grow over the transverse part of the oral disc. In such cases parts of the transverse part of the parietobasilar muscles, are enclosed in the oral disc. In the elongated forms of the parietobasilar muscles, the relation to the form of the body and the mode of life is not so clear as that of attachment for the body. As

the parietobasilar muscles are very weak in the Zoantharia and Madreporaria and in the primitive Actiniaria, I think a slightly developed parietobasilar muscle to be a primitive character.

As the basilar muscles appear only in connection with the development of a proper pedal disc, the species having basilar muscles represent a secondary stage of evolution.

The filaments of the mesenteries are usually divisible into two portions, a distal one consisting of a middle tract and two ciliated tracts, separated from each other by an endodermal part, the intermediate tract, and a proximal, the endoglandular tract, a continuation of the middle tract. In some groups, Protantheae, Corallimorphae and Ptychodacteae, the ciliated tracts are missing, and the filaments consist of only a single tract. We meet a special differentiation of the filaments in the Ptychodacteae, where the single tract of the imperfect mesenteries is drawn out orally into two lobes, forming together a half-funnel, possibly with the same function as the ciliated tracts. In some genera, *Halicampa* and *Bologrodaria*, a closer examination will probably show a similar structure in some other filaments. The ciliated tracts are a little prolonged into the furrows of the intermediate tracts, a structure recalling the filaments of the Zoantharia, though in this group the prolongation of the ciliated tracts is considerably more pronounced. Rarely, in *Isochordaria*, *Limaetia*, and *Scytophorus*, the ciliated tracts are discontinuous, exceptionally the filament itself is interrupted (in the imperfect mesenteries of *Calliactis parasitica*). Whether the presence of a simple tract in the Madreporaria and in the above named Actiniaria is primitive or not, authors somewhat disagree. Almost all authors who have discussed this problem are of opinion that the simple filaments represent an earlier type in phylogeny than those provided with ciliated tracts, only STEPHENSON (1921, pp. 556-558) suggested that the stage with a simple filament has possibly arisen secondarily. Although S. allows "that the ancestor of all Anthozoa was without ciliated streaks", he proposes (p. 557) "that it is likelier that the ancestor of all three groups of Zoantharia, the Edwardsids, the Zoanthids, and the Dodecaactiniaria (Actiniaria and Madreporaria) had ciliated streaks perhaps only in a slightly differentiated form, and it is a simpler way of putting things to think of some forms losing them than of three groups joining them". Certainly if there were three groups of Anthozoa with identical ciliated streaks, STEPHENSON'S way of thinking could be taken into consideration. Now it is not so. The Edwardsids are no special group comparable with the Actiniaria and Zoantharia, but must be regarded as a relatively highly differentiated family among the Actiniaria, and the ciliated streaks (tracts) of the Actiniaria and Zoantharia are not identical, though resembling each other. Small Zoantharia also have very weak ciliated tracts, according to DIERCKX (1898, p. 461, 1907) none, but I think this statement needs verification. There are ciliated tracts also in Ceriantharia, though different from those of the other groups. Moreover, as the ciliated tracts develop later than the single tract, and the Madreporaria, Protantheae, Corallimorphae and Ptychodacteae in more than one respect must be regarded as primitive, there is hardly any reason to suggest that the ciliated tracts are reduced in these groups. And how do we explain the origin of the half-funnel-like filaments in the Ptychodacteae according to STEPHENSON'S opinion? The ancestor should have had no ciliated tracts, then such organs were to appear, later to be reduced, and at last the half-funnel was to have arisen, I find such a supposition hardly acceptable. It is easier to accept the development of the half-funnels from a simple filament. Moreover, STEPHENSON'S supposition that the ciliated tracts have been reduced in the Madreporaria on account of the intercalation of septa keeping the mesenteries apart, and so making the function of the ciliated tracts unnecessary, can just as well be interpreted in the sense that the ciliated tracts, being unnecessary, have never been developed. Also S.'s remarks, that the forms without ciliated tracts have weak or no sphincter in opposition to forms with ciliated tracts having mostly strong sphincters, hardly holds good, as there is a great

number of the Actinaria having no or a weak sphincter, but well developed ciliated tracts. Also the Ceriantaria, which are not provided with a sphincter have, as also S. remarks, well developed ciliated tracts.

Summing up all this there is to my mind nothing in favour of the assumption that a reduction of the ciliated streaks has taken place in the Anthozoa, at least in the groups previously mentioned. Possibly, however, there is a certain reduction of the ciliated tracts in the above mentioned few genera, where the ciliated tracts have been interrupted. This question is on the whole connected with the origin of the ciliated tracts, which is not yet well enough known.

As a special differentiation of the filaments the acontia may be considered, which, at least in many cases, appear so early as in the larvae with only 8 mesenteries (CARLGRÉN 1934). In all Actinaria having acontia these organs are of a practically uniform structure. The statement of O. and R. HERWIG (1879) that the muscles in *Calliaetis* (*Sagartia*) *parasitica* are situated on the same side as the nematocysts is incorrect, in fact the muscles occupy also in this species the opposite side. Curious organs like acontia, but with few nematocysts, are present in the genus *Nemanthous* (CARLGRÉN 1940b). The nematocysts of the acontia are, however, different in structure and size in the different families. According to some authors the acontia are rudimentary in some species. Although it is possible that such statements are correct in a few cases, I think, however, that most of them are false, and due to bad preservation of the material examined, and that an examination of better preserved specimens would give other results. The occurrence of rudimentary acontia gave STEPHENSON (1920 p. 498, 504) the idea, already put forward by VERRILL (1899 p. 143) that all Thesaria having a mesogloecal sphincter originate in forms provided with acontia and cinclides. STEPHENSON thus suggested (l. c. p. 505) that the loss of activity and slower metabolism of deep-water forms may cause a degeneration of the cinclides and acontia in these forms. As to the cinclides it is probable that a degeneration has sometimes taken place (see above p. 79), but, considering the variability in the appearance of the cinclides—of genera closely related to each other one may be provided with cinclides, another not—it is not likely that the ancestor of all genera provided with a mesogloecal sphincter have had cinclides, the more so as pores occur in so different places, such as in the physa, in the body-wall, in the marginal sphaerules and pseudo-sphaerules, and in many of the genera having an ectodermal or no sphincter. Now as to the reduction of the acontia, STEPHENSON, as above mentioned, suggested that the acontia have been reduced in deep-sea forms. Before discussing this question we may make clear the primitive function of the acontia. Are they in the first place developed as defensive arms, and to be used outside the animal, or is their primitive function to kill the booty, which has been put into the coelenteron alive? I think the latter function the more primitive. If not it seems to me difficult to understand why the acontia are not reduced in forms lacking cinclides, for an ejection of the acontia from the mouth certainly usually has only a slight effect as defensive arms. Accepting my interpretation as to the primitive function of the acontia, a reduction of these organs in species having once obtained them is not easily cleared up. It is untrue that many (most?) deep-sea Actinaria having a mesogloecal sphincter are devoid of acontia, on the contrary there are several forms living the deep-sea, which have well developed acontia, for instance *Phelliaetis*, *Kadosactis* (*Kadosanthus*), *Chondrophellia*, and other Hornathiidae. Moreover there are many genera living in shallow water and provided with a mesogloecal sphincter but no acontia. A positive instance of degeneration of acontia in deep-sea forms is as yet unknown. The specimens of *Liliella* (STEPHENSON 1918a p. 33) were badly preserved, so that a renewed examination of this species is desirable¹⁾. The matter

stands somewhat differently, if *Stephanauge* (*S. acialis* and *acuellae*) should prove to lack acontia²⁾, this genus is certainly related to *Amphianthus*, which has acontia. But also with this single instance of reduction of the acontia, it is not permissible to generalize so as to say that all genera now devoid of acontia, but provided with a mesogloecal sphincter, have their origin in a form with such organs, the more so as it is certain (see p. 80) that the sea-anemones provided with a mesogloecal sphincter are not descendants from a common ancestor. So I cannot endorse STEPHENSON'S opinion that the family Actinostolidae (the former family Paractinidae) has arisen from a form having acontia. Moreover it is questionable if in fact, all sea-anemones with acontia have a common ancestor. The presence of acontia even in the genera lacking basilar muscles, shows that the first appearance of these organs is to be found in a rather primitive stage of the Nymphaeae. Among the Athenaria there are 3 families, Haleampectidae, Andwakiidae and Octoneonidae, all having acontia, the first family without a sphincter, the other two with a mesogloecal one. The family Haleampectidae is the most primitive, and from it the two other families can be derived, and from a pre-*Haleampectis*-stage, with weak retractors perhaps, several families of the Actinaria with basilar muscles have arisen. But, as I have suggested, (1924b p. 19), there is possibly another line going from forms lacking basilar muscles to forms having these, that is to say from the Boboceroidaria, or forms allied with this group, to such genera as *Aheia* (from here to perhaps the Diadumenidae) which has retained several primitive characters. But the question whether the sea-anemones provided with acontia have a monophyletic or a di-(poly-)phyletic origin is in reality very difficult to clear up. It seems to me, however, that the acontia-like organs developed in the family Nemanthidae (CARLGRÉN 1940b) indicate a di- or polyphyletic origin of the acontia. The presence of different types of nematocysts in the acontia (see CARLGRÉN 1940 p. 50 note 4, in which a lapsus penae is to be corrected—Acontiferidae instead of Acontiphoridae)—need, however, not render necessary a polyphyletic origin for many of the Acontinaria, if they originate from a form having only microbasic b-mastigophors in its acontia (see schedule p. 55 CARLGRÉN 1940), as is the case with *Metridium*, which has sometimes only these nematocysts in its acontia, while other varieties of it are provided also with microbasic anastigophors. It is, however, to be suspected that the anastigophors have been reduced in the acontia of some forms of *Metridium*. Finally we have to consider the cnidae. The distribution of the spirocysts has been dealt with above. According to WEILL (1935) we have to deal with 6 categories of nematocysts in the Anthozoa, viz. atrichs, holotrichs, microbasic mastigophors, microbasic and macrobasic anastigophors, and basitrichs. Because of the different appearance and distribution of the microbasic mastigophors in the Anthozoa, I have divided them into *b*- and *p*-mastigophors, thus, to my mind, 7 categories occur. There is, so far as we at present know, no genus provided with all types of nematocysts. In the genus *Dialanassa* s. str., however, all types, apart from the microbasic *b*-mastigophors, are present. Usually only two types, microbasic *p*-mastigophors and basitrichs, occur in the Actinaria, sometimes there are 3, rarely one or four (see WEILL 1935 and CARLGRÉN 1940). I have above remarked that there are certain types of nematocysts in the marginal sphaerules, in the tentacles of some genera, and in the acontia of different families, all certainly of systematic value. Moreover there is no doubt that the knowledge of the cnidom, i. e. the types of cnidae occurring in a species or genus, is generally valuable in finding out its relationship to other genera, though an agreement in the cnidom between two species does not indicate an absolute relationship, because the development of the types of nematocysts from more primitive types to more advanced ones has evidently ceased in some types, or in other words it is a parallelism of development (see CARLGRÉN 1940). For instance

¹⁾ The presence of acontia has been stated here (CARLGRÉN & STEPHENSON, Actinaria, Australasian Antarctic expedition 1911-1914, Ser. C. 9, P. 2, 1929).

²⁾ According to VERRILL (1922 p. 99 Pl. XXI fig. 68) *Stephanauge* has acontia, but apparently few.

8) The endium of the Actiniaria probably had the same endium, but the types of nematocysts and their distribution may be different. It is not clear, however, that it may be possible to determine their relationship to each other. The distribution of nematocysts is very different, that some genera or species have a certain type of nematocysts and at the same time other types. For instance the family *Stolidobranchia* (e.g. *Stolidia*, *Asapha*) have atrichs and *Stolidia* has also *Stolidia* spirocysts and *p*-mastigophors, *Asapha* has *Asapha* spirocysts as above mentioned, all types of nematocysts are also mentioned by CAMERON (1940). The relationship of the endium of the genus is however undoubtedly valuable for the determination of its relationship, though one must be careful not to draw a conclusion from an agreement of the endium of two genera that they are related to each other.

9) The evolution of most of the organs of the Actiniaria have, in the course of time, gone in the following directions:

1) The basal part of the body was primitively a basal plate, provided with radial impressions, without basilar muscles, and with a columnar structure as the column. This plate has either developed into a broad physa or been enlarged and eventually provided with a broad physa. Perhaps also the physa can form a distinct column by its flattening and by getting basilar muscles.

2) The column was primitively provided with ectodermal circular muscles and spirocysts, which, in primitive sea-anemones are more or less retained, but in more differentiated genera is fully, wholly reduced. The spirocysts are more conservative than the muscles, and sometimes present in highly differentiated genera. A resurrection of the longitudinal muscles, if once reduced, does not, or exceptionally, in *Lamactinia* (?) take place.

3) The ancestor of the Actiniaria had no sphincter. The circular muscle tract arises, by folding into the distal part of the column, into a ring of an endodermal or mesogloea sphincter. This latter has arisen either directly from the circular muscles or passed through an endodermal sphincter stage. There is no reason to suppose a reduction of ancestor on one side for the forms having a diffuse or circumscript sphincter and, on the other, for those provided with a mesogloea sphincter. A reduction of a mesogloea sphincter has not been stated.

4) The longitudinal muscles of the tentacles and the radial muscles of the oral disc were, in the primitive stage, ectodermal. Radial and circular enclosing, meso-endodermal, endo-mesogloea and at least mesogloea muscles arise in different species which often are related.

5) The ancestor of the Actiniaria probably had no siphonous tentacles, or a weak ventral one. A single, ventral siphonous tentacle, well developed, is present in some genera; whether this is primitive or not is questionable. In many other species siphonous tentacles occurs, but always in connection with asexual reproduction caused by regeneration. Usually a dorsal siphonous tentacle is developed.

6) The ancestor of the Actiniaria was probably provided with 8 pairs of tentacles, arranged as the macrocnemes in *Edwardsia* (see fig. 8, tentacles). No recent Actinarian remains at this stage. In the course of time 4 couples of mesenteries have arisen forming a dorsal, ventral, lateral and ventrolateral complex. From this complex the mesenteries, before the mesenteries have been differentiated, probably of development have originated, one pair of the mesenteries forming in the endoderm new pairs of tentacles with their longitudinal muscles orientated towards the anterior, the other forming new pairs in the endoderm with their longitudinal muscles facing inwards.

7) The ancestor of the Actiniaria probably had 12 mesenteries, new mesenteries grow in the course of time, especially in forms with a cylindrical body usually only form relatively few mesenteries and strongly concentrated or circumscript retractors on the macrocnemes, broader and lower species usually numerous mesenteries, not differentiated into macrocnemes and microcnemes, and usually diffuse retractors. No reduction of the strong retractors of the macrocnemes takes place in connection with the increase in the number of mesenteries.

8) The filaments of the Actiniaria originally consist of a single tract. To this later come two ciliated tracts in the upper part of the filaments, or the single tract may, but rarely, form curious half-funnel-like organs in its upper part. A total reduction of the ciliated tracts, a regression to a stage with a single tract, can hardly be imagined, though the ciliated tracts or the whole filament may sometimes be reduced in certain places and thus become discontinuous.

9) The acontia are a differentiation of the filaments, originating, as far as is yet known, before the ciliated tracts of the filaments arise. Their origin is probably not monophyletic, because, among others, organs like acontia but with few nematocysts occur. The acontia are provided with nematocysts, which are of different types in certain families, but probably can be derived from microbasic *b*-mastigophors, though these nematocysts are rarely present in the acontia.

10) The knowledge of the endium of a genus is of importance in order to clear up its relationships. If two species have the same endium, it is, however, not certain that they are related to each other, because the types of nematocysts have changed more or less parallel with the development of other organs during evolution. The ancestor of the Actiniaria had at least atrichs, probably also holotrichs and microbasic *b*-mastigophors, perhaps also *p*-mastigophors, in its endium.

We now proceed to the classification of the Actiniaria.

It is relatively easy to arrange the genera in families, considerably more difficult, however, to arrange the families in larger groups. The cause is that the Actiniaria are relatively simply constituted, and that their development has certainly been parallel in several cases, as already announced by myself (1941 p. 28). There are, however, some groups, which are distinctly separated from the other Actiniaria, viz. the Psychodacteae (Psychodactylinae). The Corallimorphiae (Protostichodactylinae) and the Endocoolanthae. Some authors, e. g. STEPHENSON and WELLS, consider the Corallimorphiae (Asclerocorallia) members of the Madreporaria. In some papers (1924c p. 180 and 1940 pp. 57-59) I have pointed out my view about the place of this group. I cannot find any real argument for placing it in the Madreporaria, in which, as far as we know at present, it would be wholly isolated. Evidently the presence of numerous holotrichs in the Corallimorphiae as well as in the Madreporaria has been of great importance in the theory of their relationship. But as holotrichs have now been found in *Dadaman*, a form provided with acontia, and in a larva the ripe form of which is unknown, there is a difference in quantity but not in quality as to the holotrichs between the Madreporaria and the Actiniaria proper. But specially regarding the presence of certain types of nematocysts and removing the Corallimorphiae from the other Actiniaria, we must for even more reasons also remove the Psychodacteae, the endium of which consists of only atrichs and few spirocysts, and which moreover in their organisation considerably deviate from the typical Actiniaria. For this reason I, in 1940, proposed to give the Psychodacteae and Corallimorphiae a higher rank, and coordinate them with the Actiniaria and Madreporaria, as I find such an arrangement acceptable. The Psychodactaria are certainly a group which has separated early from the other Actiniaria and gone their own way, and the Corallimorpharia have run through a development fairly parallel with the Madreporaria. As to the Endocoolanthae they belong to the Actiniaria proper, but are a very homogenous group, characterized, among other features, by the curious origin of their mesenteries. If we go further to the primitive Actiniaria proper, we first have

to consider the Protantheae, a group to which I, in earlier papers, reckoned the above mentioned groups to belong. I now agree with STEPHENSON and include in the Protantheae only the family Gonactiniidae, characterized by the presence of longitudinal columnar muscles, by the absence of ciliated tracts of the filaments, and of basilar muscles. Undoubtedly this family is very primitive, only the presence of microbasal amastigophors suggests a more advanced stage. As coordinated with Protantheae and Endocoel-antheae I have proposed the group Nynantheae, provided with ciliated tracts of the filaments in agreement with the Endocoel-antheae. Among the Nynantheae we can distinguish three groups, the two first, the Athenaria or Abasilaria, and the Boloceroïdaria without basilar muscles, the third, the Thenaria or Basilaria with basilar muscles. The Athenaria are usually provided with a physa and commonly have a very elongated body, and their mesenteries, as a rule, are divisible into macro- and microcunemes; some of the Athenaria also have acontia. The Boloceroïdaria have a different exterior and agree more with the common Actiniaria. They are certainly derived from the Protantheae (CARLGRÉN 1924b p. 19, 1940 p. 30, PAXIKKAR 1937a p. 87) and separated from them only by the presence of ciliated tracts of the filaments and the absence of atrichs; in all other characters such a form as *Boloceroïdes* agrees with *Protanthea*. As to the forms with basilar muscles, the Thenaria, STEPHENSON (1920-22) has divided my group at once into Endomyaria and Mesomyaria, the latter group including also forms with acontia, a subdivision, however, which is not successful, as he counted in with the Mesomyaria also some forms having acontia but no sphincter. It is also very difficult to classify the Actiniaria which are provided with basilar muscles, and possibly it would be most suitable, at present, not to divide them into larger groups. It is not certain that all forms with no sphincter, or with an endodermal sphincter, genetically belong together. The same holds good also of the forms with a mesogloal sphincter. It is true that the family Actiniidae and the stichodactyline families show a near relationship, but the family Aliciidae, having no sphincter but provided with macrobasal amastigophors, is an aberrant family not related to the family Actiniidae. In fact, I think the origin of this family is to be sought not far from the Boloceroïdidae, the microbasal mastigophors of which (CARLGRÉN 1940 figs. 1, 3, 5, 7) seem to me to be a prestadium of the macrobasal amastigophors. Moreover the basilar muscles of the Aliciidae show all transitions from very weak to strong ones, in the genus *Alicia* they are only indicated, in *Lebrania* they are very strong. As to the forms with acontia there are several difficulties concerning their classification, some of them have no basilar muscles, a few have no sphincter, and most have a mesogloal one. The genera placed here below with the Athenaria have the same endom in the acontia as two groups provided with basilar muscles. Very different from the others is the family Diadumenidae (the three species of the genus *Diadumene*), which has no sphincter, but is provided with all types of nematocysts apart from the microbasal *b*-mastigophors, including atrichs, holotrichs, and macrobasal amastigophors. It is not probable that the family has been developed from the same line as the other forms provided with acontia, but from a line not far from that of *Alicia*, for it must be remarked that only the families Aliciidae and Diadumenidae have macrobasal amastigophors—as far as we at present know. Possibly the family Aiptasiidae has its origin not far from the Diadumenidae. If we retain for the present STEPHENSON'S groups Endomyaria and Mesomyaria, it is, however, necessary to separate the forms with acontia from the Mesomyaria (see above). So I have proposed a new sub-tribus, the Acontiarina, coordinated with the Endomyaria and Mesomyaria (see STEPHENSON 1935 p. 181). The erection of these groups, however, has to my mind only a temporary value.

STEPHENSON has (1920-21) made an attempt to give an evolutionary history of the families of the Mesomyaria and Endomyaria. Issuing from an *Eosagartia* provided with acontia, he thinks,

among other theories, that the families now lacking acontia previously had such organs, but later lost them. I find nothing to support such a supposition (see p. 83), and STEPHENSON'S scheme of the evolution of the Mesomyaria is to my mind invalid. Nor can I accept his scheme for the evolution of the Endomyaria. When going into details there is much more to be discussed about STEPHENSON'S classification of 1920-22, but as space is limited in this paper I cannot enter into a closer discussion here.

The genera can, to my mind, at present be arranged in the following manner. Several genera, the anatomy of which is quite unknown, have not been included here. The names of a few others are somewhat dubious, as their synonymy is not satisfactorily cleared up.

Sub-order Ptychodactiaria.

Fam. Ptychodactiidae. Genera: *Ptychodactis* App., *Dactyloanthus* Carlgr.

Sub-order Corallimorpharia (Asclerocorallia).

Fam. Sideractiidae. Genera: *Sideractis* Dan., *Nectactis* Gray.

Fam. Corallimorphidae. Genera: *Corallimorphus* Mosell., *Isocorallion* Carlgr., *Corpaectis* Allm.

Fam. Discosomidae. Genera: *Discosoma* Leuck., *Paraliscosoma* Carlgr., *Oriolia* Duch. & Mich., *Rhodactis* Milne-Edw. & Haime, *Actinotrypa* Duch. & Mich., *Ricoidea* Duch. & Mich.

Sub-order Actiniaria (proper).

Tribus Protantheae.

Fam. Gonactiniidae. Genera: *Protanthea* Carlgr., *Gonactinota* Sars.

Tribus Endocoelanthaeae.

Fam. Haleuriidae. Genera: *Haleurias* McMurr., *Carlpenia* Steph.

Fam. Actinernidae. Genera: *Actinernus* Verr., *Syphalaeurias* Carlgr., *Syphaeternus* Carlgr., *Isaeternus* Carlgr.

Tribus Nynantheae.

Sub-tribus Boloceroïdaria.

Fam. Boloceroïdidae. Genera: *Boloceroïdes* Carlgr., *Bolocraetis* Panik., *Banodropsis* Andr.

Fam. Nevadaeidae. Genus: *Nevadae* Steph.

Sub-tribus Athenaria or Abasilaria.

Fam. Edwardsiidae. Genera: *Edwardsia* Quatr., *Isosedwardsia* Carlgr., *Fagesia* Delphy (= *Milne-Edwardsia* Carlgr.), *Parasedwardsia* Carlgr., *Nematostilla* Steph.

Fam. Linnactiniidae. Genus: *Linnactinia* Carlgr.

Fam. Haleanpoididae. Genera: *Haleanpoides* Dan., *Haleanpella* (Andr.) R. Hertwig, *Syphalcampella* Carlgr., *Pentactina* Carlgr., *Aethalms* Lurk., *Scytophorus* R. Hertw., *Haleanpogoton* Carlgr.

Fam. Haloclavidae. Genera: *Haloclava* Verr., *Anemonactis* Andr. (= *Eloactis* Andr.), *Peuchia* Gosse, *Mesamaca* Andr., *Oractis* McMurr., *Harenactis* Torr., *Siphonactinopsis* Carlgr.

Fam. Halecampidae. Genera: *Halecampa* Gosse, *Parahalecampa* Carlgr., *Halecampaster* Carlgr., *Cactosoma* Dan., *Haliandrella* Kwietn., *Mena* Steph.

Fam. Octineonidae. Genus: *Octineon* Fowl.

Fam. Andwakiidae. Genus: *Andwakia* Dan.

Fam. Halecampaetidae (perhaps two families). Genera: *Halecampaetis* Farq., *Phlocactes* Ann., *Phylocactes* Ann., *Phylocactopsis* Panik., *Stephensonactis* Panik., *Haliactis* Carlgr.

Sub-tribus Endomyaria.

Fam. Aliciidae. Genera: *Alicia* Johns., *Phyllodiscus* Kwietn., *Lebrania* Duch. & Mich.

Fam. Complanthidae. Genera: *Complanthus* Carlgr., *Charisca* Torr., *Macrocnema* Carlgr.

1939. BROCH, H. Anthozoa, mainly from Pacific waters collected by U.S.S.R. Expeditions 1930-1932. Exploration de Mers de l'URSS Fasc. 33 (text also Russian).
1792. BROCQUÉ, M. Histoire naturelle de Vers 1. Encycl. méthodique. Paris 1792.
1890. BRUNCHONST, J. Die biologische Meeresstation in Bergen. Bergens Mus. Aarsb. for 1890.
1891. CARLQVIST, O. Beiträge zur Kenntnis der Actinien-Gattung *Bolocera*. Övers. K. Svenska Vet.-Akad. Forh. 1891 N: 1. Stockholm.
- 1891a. *Protanthea simplex* nov. gen. n. sp. eine eigentümliche Actinie *ibm.* No. 2 1891.
1892. Beiträge zur Kenntnis der Edwardsien. *ibm.* No. 1 1892.
- 1893a. Studien über Nordische Actinien I. K. Svenska. Vet.-Akad. Handl. 25 No. 10. Stockholm.
- 1893b. — Über das Vorkommen von Braträumen bei Actinien. Övers. K. Vet.-Akad. Forh. 1893 No. 1. Stockholm.
1894. — Zur Kenntnis der Minyaden *ibm.* 1894 No. 1.
1898. — Zoantharien. Hamburger Magelhaensische Sammelreise 4. Hamburg 1899.
1899. — Über abschürfbare Tentakel bei den Actiniarien. Zool. Anzeig. 22. No. 578.
- 1900a. — Ostafrikanische Actinien. Jahrb. Hamburg wiss. Anstalt 17. Beiheft 2, 1900.
- 1900b. — Über *Pentactinia californica* n. gen. n. sp. Övers. K. Svenska Vet.-Akad. Forhand. 57.
1901. — Die Brutpflege der Actiniarien. Biol. Centralbl. 21 No. 15.
1902. — Die Actiniarien, in Zoologische Ergebnisse einer Untersuchungs-fahrt im Sommer 1898 auf S.M.S. "Olga". Wiss. Meeresuntersuch. etc. N.F. 5. Abt. Helgoland No. 1.
- 1903-1908. Anthozoa. Bronns Klassen und Ordnungen. 2. Abt. 2 Coelenterata. Leipzig.
1901. — Kurze Mitteilungen über Anthozoen I 3. Zool. Anzeig. No. 16, 17. 1901.
- 1901a. — Studien über Regenerations- und Regulationserscheinungen P. I. K. Svenska Vet.-Akad. Handl. 37 No. 8 1901.
1905. — Kurze Mitteilungen über Anthozoen I. Zool. Anzeig. 28 No. 11-15 1905.
1905. — in NORDGAARD, see NORDGAARD.
1906. — Die Actinien-Larven. Nordisches Plankton 11. 1906. Kiel u. Leipzig.
1909. — Studien über Regenerations- und Regulationserscheinungen 2. K. Svenska Vet.-Akad. Handl. 43 No. 9. Stockholm.
1911. — Über *Daetylanthus (Cystiactis) antarcticus* (Clubb.). Wiss. Ergebn. Schwed. Südpolar Exped.
1912. — in BROCH, see BROCH.
1912. — in STEPHENS, see STEPHENS.
1913. — Actiniaria. Report Sec. Norwegian Exped. in the Fram 1898-1902 No. 31. Kristiania 1913.
1914. — On the Genus *Porponia* and related Genera. Scottish Nat. Antarctic Exped. Trans. R. Soc. Edinburgh 50 P. 1 No. 1.
1916. — Actiniaria and Zoantharia of the "Danmark" Expedition. Medd. om Grønland 13. 1916. Kjøbenhavn.
1918. — Die Mesenterienordnung der Halcurriden. Lunds Univ. Arskr. N. F. Avd. 2. Bd. 11. No. 29. 1918. Lund.
1921. — Actiniaria P. I. The Danish Ingolf Expedition 5. 9. Copenhagen.
1921. — Actiniaria in Dollfus. Contribution à la faune des Invertébrés du banc de Rockall. Bull. l'Inst. Océanogr. No. 138. Monaco 1921.
- 1921a. — Die Larven der Ceriantbarien, Zoantharien und Actiniarien etc. Wiss. Ergebn. Deutschen Tiefsee-Expedition (Valdivia) 19 H. 8. Jena.
- 1921b. CARLQVIST, O. On *Boloceroidea*, *Bunobopsis* and their supposed allied genera. Ark. for Zool. 17 A No. 1. Stockholm.
- 1911c. Actiniaria from New Zealand and its Subantarctic Islands. Vid. Medd. fra Dansk naturh. Foren. 77.
1925. On the Actinarian family Amphianthidae. Ark. för Zool. 17 B No. 1. 1925.
- 1925a. A revision of the Actiniaria of the Chilka Lake. *ibm.* 17 A No. 21. 1925.
1927. Actiniaria and Zoantharia. Further zoological Results of the Swedish Antarctic Expedition 2. No. 3 1927. Stockholm.
1928. Ceriantbarier, Zoantharier och Actinarianer in *Conspectus Faunae Groenlandicae*. Med. om Grønland 23. Suppl. Kjøbenhavn.
- 1928a. Actiniaria der Deutschen Tiefsee-Expedition. Wiss. Ergebn. Deutschen Tiefsee Exp. 22 H. 1. Jena.
1930. Actiniaria and Zoantharia in Zoology of the Faroes VIII. Copenhagen.
1931. Zur Kenntnis der Actiniaria *Abasilaria*. Ark. for Zool. 23 A No. 3. Stockholm.
1932. Die Ceriantbarien, Zoantharien und Actinarianer des arktischen Gebietes. Fauna arctica 6. 3. Jena.
1933. Zoantharia and Actiniaria in The Godthaab Expedition. Medd. om Grønland 79 No. 8. Kjøbenhavn.
1933. in KRAMP, see KRAMP.
1934. Zur Revision der Actiniarien. Ark. f. Zool. 26 A No. 18. Stockholm.
- 1934a. — Some Actiniaria from Bering Sea and arctic waters. Journ. Washington Acad. Sci. 24 No. 8. 1934.
- 1934b. Ceriantbaria, Zoantharia and Actiniaria. Report Michael Sars North Atlantic Deep-Sea Exped. 1910. Bergen.
1935. Di alcune Attinie e Zoantbari raccolti nel Golfo di Genova. Bull. Mus. Lab. Zool. Anat. Università Genova 15 No. 83.
1936. in BROCH, see BROCH.
1937. Ceriantbaria and Zoantharia. Sci. Reports Great Barrier Reef Expedition 5 No. 5. British Museum. London.
1939. Actiniaria, Zoantharia and Madreporaria. The Zoology of Iceland 2. P. 8. Copenhagen and Reykjavik.
1939. in BROCH, see BROCH.
1940. A contribution to the knowledge of the structure and distribution of the Cnidaria in the Anthozoa. Lunds Univ. Arssk. (N.F.) Avd. 2. 36 No. 3.
- 1940a. Actiniaria from Alaska and Arctic waters. Journ. Washington Acad. Sci. Jan. 1940.
- 1940b. Actiniaria from the Gulf of California. Zoologica. New York Zool. Soc. 25 P. 2.
1928. O. and STEPHENSON, The British Edwardsiae. Journ. Marine Biol. Assoc. (N.S.) XV. No. 1.
1851. COCKS, W. P. Actiniae procured at Falmouth. Ann. Report R. Cornwall Polytechn.
1890. CUNNINGHAM, J. F. *Tedia tuberculata* (Cocks) a Study in Synonymy. Journ. Mar. Biol. Assoc. 1 (N.S.) P. 2.
1856. DANIELSSEN, D. C. & KOREN, J. *Næ Actiner* in *Fauna littoralis Norvegiae* 2. 1856. Bergen.
1877. *ibm.* 3. 1877. Bergen.
1890. DANIELSSEN, D. C. Actinida. The Norwegian North Atl. Exped. Zool. 19.
1858. DAWSON, J. W. On Sea Anemones and Hydroid Polyps from the Gulf of St. Lawrence. Canadian Nat. and Geol. 3.
1901. DELAGE, Y. & HÉROUARD, E. Traité de Zoologie concrète 2. Les Coelentérés. Paris 1901.
1888. DIXON, A. F. On the arrangement of the mesenteries in the Genus *Sagartha* Gosse. Sci. Proc. R. Dublin Soc. (N. S.) 6. Dublin.

1758. LINNÉ (LINNÆUS) C. VON. Systema Naturae Edit. 10. Holmiae 1758.
1761. -- Fauna Suecica Stockholmiæ 1761.
- 1766-68. Systema naturae Edit. 12. Holmiae 1766-68.
1861. LÜTKEN, C. Nogle Bemærkninger om de ved de danske Kyster iagttagne Arter af Aktiniernes Gruppe. Vid. Medd. naturh. Foren. Kjøbenhavn 1860.
1875. -- A revised Catalogue of the Anthozoa and Calycozoa of Greenland. Manual and Instructions for the Arctic expedition. London 1875.
1898. LÖNNBERG, E. Undersökningar rörande Öresunds djurlif. Medd. K. Landbruksstyrelsen No. 1. Upsala 1898.
1902. -- Undersökningar rörande Skeldervikens och angränsande Kattegat-områdes djurlif. ibm. No. 2. 1902.
1878. MARENZELLER, E. VON. Die Coelenteraten, Echinodermen und Würmer der K. K. österreichisch-ungarischen Nordpol-expedition. Denks. K. Akad. Wissensch. Wien. Math.-naturw. Classe 35. 1878.
1884. MACKINTOSH, H. W. Report on Irish Zoophytes P. 1. On some rare sea anemones taken at Greystones co. Wicklow. Proc. R. Irish Acad. (2) 4. Dublin.
1889. McMURRIEN, J. P. The Actiniaria of the Bahama Islands. Journ. of Morphol. 3 No. 1. Boston.
1891. -- Contribution to the Morphology of the Actinozoa III. ibm. 5 No. 1. Boston.
1893. Report on the Actiniae collected by the United States Fish Commission Steamer "Albatross" during the winter of 1887-1888. Proc. U.S. Nat. Mus. 16. Washington.
1898. -- Report on the Actiniaria collected by the Bahama Expedition of the State University of Iowa 1893. Bull. Lab. Nat. Hist. Iowa. 1. Iowa 1898.
1901. -- Report on the Hexactiniae of the Columbia University Expedition to Puget Sound etc. Ann. N.Y. Acad. Sci. 11. New York.
1901. --- The Actiniae of the Plate collection. Zool. Jahrb. Suppl. 6. Jena.
1910. -- The Actiniaria of Passamaquoddy Bay with a discussion of their Synonymy. Trans. R. Soc. Canada (3) 4 sect. 4. Montreal.
1913. -- On two new Actinians from the coast of British Columbia. Proc. Zool. Soc. London 1913.
1863. MEYER, A. and MÖBIUS, K. Beschreibung der Edwardsia duodecimerrata aus der Kieler Bucht. Wiegmanns Arch. f. Naturg. 29. 1. Berlin.
1857. MILNE-EDWARDS, H. Histoire naturelle des Coralliaires ou polyps proprement dits. Tom. 1. Atlas. Paris 8.
1897. MORTENSEN, TH. Smaa biologiske og faunistiske Iagttagelser. Vid. Medd. Dansk Naturh. Forening. Kjøbenhavn (p. 316).
1877. MOSELEY, H. N. On new forms of Actiniaria dredged in the Deep-Sea. Trans. Linnæan Soc. (2) 1.
1776. MÜLLER, O. F. Zoologia Danicae Prodronus seu Animalium Danicae et Norvegiae. Havniæ 1776 8.
- 1788-1806. Zoologia Danica seu animalium Danicae et Norvegiae descriptiones et historia. Havniæ 1788-1806. P. 1. 1788. P. 3. 1789 P. 4 1806.
1873. MÖBIUS, K. Die wirbellosen Thiere der Ostsee. Berichte Exped. Pommerania. Kiel.
1893. Über die Thiere der schleswig-holsteinischen Austernbanke, ihre physikalischen und biologischen Lebensverhältnisse.
1912. NAFILYAN, Z. G. Matériaux pour la fauna des Actinies des côte de France. Les Actinies de Roscoff. Mém. Soc. zool. France 25. Paris.
1905. NORDGAARD, O. Hydrographical and Biological Investigations in Norwegian Fjords. Bergens Museum. Bergen 1905.
1868. NORMAN, A. M. Shetland final dredging Report P. II on the Crustaceans etc. Report British Ass. f. Adv. Sci. 1868.
1865. PACKARD, A. S. View of the recent Invertebrate Fauna of Labrador. Mem. Soc. Nat. Hist. Boston 1 P. 2. Boston 1867.
1936. PANIKKAR, N. K. The structure, bionomics and systematic position of two new brackish-water Actiniaria from Madras. Proc. Zool. Soc. London P. 1. 1936.
1937. Notes on *Neradic glauca*. Records Indian Museum 39 P. 1. Calcutta.
- 1937a. The Morphologic and Systematic Relationship of a new Boloceroïdarian etc. Proc. Indian Acad. Sc. A No. 2.
1900. PARKER, G. H. Synopsis of North American Invertebrates 13. The Actiniaria. Amer. Naturalist 31 No. 195. 1900.
1907. PAX, F. Vorarbeiten zu einer Revision der Familie Actiniidae. Inaug. Diss. Breslau.
1908. Die Actinienfauna Westafrikas. Denks. med.-naturw. Ges. Jena. 30.
1909. Aktinienstudien I-III. Jena Zeits. Naturw. 15. Jena.
1912. Révision des types des Actinies décrites par Quoy et Gaimard. Ann. Sci. Natur. (9) zool. 16.
1911. Die Aktinien. Ergebn. Fortsch. Zool. 1. H. 3 4. Jena.
1915. Hexacorallien der Murmanküste. Trav. Sci. Imp. Natur. Petrograd 114.
1920. Die Aktinienfauna von Büsum. Schr. Zool. Stat. Büsum 5. H. 6.
1922. Zoantharien und Actiniarien in Untersuchungsfahrt des Reichsforschungsdampfers "Poseidon" in das Barentsmeer etc. Wiss. Meeresunt. Neue Folge Abteil. Helgoland 13. 2.
1925. Coelenterata. Unterklasse Hexacorallia in Handbuch der Zoologie P. 1. 1923-25. Berlin und Leipzig.
1928. Die Tierwelt Deutschlands T. 4. 1928. Jena.
1936. Anthozoa. Die Tierwelt der Nord- und Ostsee XXX. 3 Leipzig.
1885. PENNINGTON, A. S. British Zoophytes. London 1885 8.
1892. PETERSEN, C. G. J. Beretning til Indenrigsministeriet fra den danske biologiske Station 1. 1890-1891. Sertryk af Fiskeriberetningen. Kjøbenhavn 1892.
1914. POUME, F. Das System der Coelenteraten. Arch. f. Naturgeschichte 80 Ab. A.
1891. PROUHO, H. Observations sur la Gonactinia prolifera. Arch. Zool. Exp. Gén. (2) 9 No. 2. Paris.
1829. RAPP, W. Über die Polypen im Allgemeinen und die Actinien insbesondere. Weimar.
1813. RATKE, H. Beiträge zur Fauna Norwegens. Nova Act. Acad. Caes. Leop. Carol. Nat. Cur. 29. Viaticslaviae & Boniae 1813.
1931. RAWLINSON, R. A comparative Study of *Metrobium scoble* (L.) var. *diathanus* (Ellis) and a Dwarf Variety of this Species etc. Journ. Mar. Biol. Assoc. U. K. 19. No. 2.
1913. REES, O. M. On Elnectis Mazeli. Journ. Mar. Biol. Assoc. U. K. 10. Plymouth.
1829. SARS, M. Bidrag till Soedyrenes Naturhistorie. Bergen 1829 8.
1833. Beitrag zur Naturgeschichte der See-thiere. Isis 1833.
1835. Beskrivelser og Iagttagelser over nogle mærkelige eller nye i havet ved den Bergenske kyst levende Dyr af Polypernes Classer. Bergen 1835 8.
1851. Beretning om en i Sommeren 1849 foretagen Reise i Lofoten og Finnmarken. Nvt. Mag. Naturv. 6. Christiania 1851.
1853. Bemærkninger over det Adriatiske Havs Fauna sammenlignet med Nordhavets. ibm. 7. 1853.
1857. Bidrag til Kendskabet om Middelhavets Littoral fauna. ibm. 9. 1857.
1861. Beretning om en i sommeren 1859 foretagen Reise ved Kysten af Romsdals Amt. ibm. 11. 1861.

Contents.

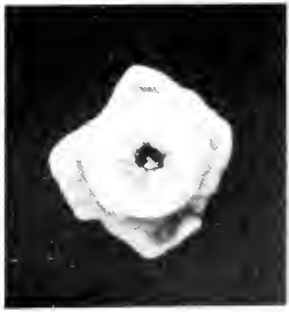
	Page		Page
ACTINIARIA. PART I (1921).			
Section I. Description of the species	1	Genus <i>Cybrina</i> s. <i>Ranodactylus</i> : <i>Cybrina bella</i> (Verr.)	148
Subtribus <i>Protactinaria</i> , Family <i>Gonactinidae</i>	1	<i>Cybrina spetsbergensis</i> (n. sp.)	151
Genus <i>Protanthea</i> , <i>Protanthea simplex</i> Carlg.	2	Genus <i>Cybrinopsis</i>	155
Genus <i>Gonactinia</i>	3	<i>Cybrinopsis similis</i> n. sp.	156
<i>Gonactinia prolifera</i> (M. Sars)	4	Genus <i>Urticina</i>	160
Genus <i>Sideractis</i>	5	<i>Urticina jellina</i> (L.)	161
<i>Sideractis glaucalis</i> Dan.	6	— <i>curata</i> & <i>tuberculata</i>	163
Fam. <i>Ptychodactylidae</i> , Genus <i>Ptychodactylus</i> , <i>Ptychodactylus patula</i> App.	11	— <i>lapolensis</i>	168
Genus <i>Actinernus</i> , <i>Actinernus nobilis</i> Verr.	14	— <i>crassicornis</i>	170
Subtribus <i>Xyactinaria</i> , <i>Athecaria</i> s. <i>Athecaria</i>	15	— <i>crassicornis</i> = <i>Cybrinopsis</i> (of <i>Cybrina</i>)	174
Fam. <i>Edwardsiidae</i> , Genus <i>Edwardsia</i>	22	Genus <i>Epactis</i>	174
Subfamily <i>Edwardsiinae</i> , Genus <i>Edwardsia</i>	27	<i>Epactis noronhaiensis</i> Carlg.	175
<i>Edwardsia tuberculata</i> Dub. & Kor.	29	<i>arctica</i> (Verr.)	177
— <i>longicornis</i> Carlg. (n. sp.)	31	<i>nordmanni</i> n. sp.	180
— <i>pallida</i> Carlg. (n. sp.)	35	<i>uvula</i> n. sp.	181
— <i>dantea</i> n. sp.	37	Fam. <i>Paractinidae</i>	183
— <i>arctica</i> n. sp.	39	Genus <i>Anthosactis</i>	190
— <i>jasea</i> Dan.	42	<i>Anthosactis pinnaequi</i> Dan.	191
— <i>andresi</i> Dan.	43	— <i>ingolfs</i> n. sp.	194
— <i>islandica</i> n. sp.	47	Genus <i>Tcalidium</i>	196
— <i>virida</i> (Dan.)	49	<i>Tcalidium jungermanni</i> n. sp.	197
— <i>roque</i> n. sp.	53	Genus <i>Epiparactis</i>	198
— <i>finmarchica</i> n. sp.	54	<i>Epiparactis dubia</i> n. sp.	199
Genus <i>Isocardiaria</i> , <i>Isocardiaria ingolfs</i> n. sp.	56	Genus <i>Pycnanthus</i>	200
Genus <i>Milne-Edwardsia</i>	59	<i>Pycnanthus boreis</i> n. sp.	202
<i>Milne-Edwardsia borei</i> Carlg.	60	— <i>diversus</i> n. sp.	204
— <i>carina</i> (Gosse)	62	Genus <i>Parasacquinis</i> , <i>Parasacquinis sarotti</i> n. sp.	208
— <i>polaris</i> n. sp.	65	Genus <i>Sicquinis</i>	211
— <i>norholsia</i> n. sp.	67	<i>Sicquinis tuberculata</i> n. sp.	212
Genus <i>Paracardiaria</i> , <i>Paracardiaria arenaria</i> Carlg.	69	— <i>ingolfs</i> n. sp.	217
<i>Paracardiaria sarsii</i> (Dub. & Kor.)	71	— <i>caribialis</i> n. sp.	218
Fam. <i>Linnactinidae</i> , Genus <i>Linnactina</i>	75	Genus <i>Actinostola</i>	221
<i>Linnactina laevis</i> n. sp.	75	<i>Actinostola spetsbergensis</i> Carlg.	222
Fam. <i>Halecampoidae</i> , Genus <i>Halecampoides</i>	80	— <i>callosa</i> (Verr.)	227
Genus <i>Halecampoides</i> , <i>Halecampoides purpurea</i> Stud.	82	— <i>abyssorum</i> (Dan.)	229
Genus <i>Aethelmus</i> , <i>Aethelmus ulcristatus</i> (Fabr.)	92	— <i>quadrilobata</i> Carlg.	230
<i>Aethelmus schaudinnii</i> n. sp.	95	Genus <i>Stomphia</i>	232
Genus <i>Peachia</i> , <i>Peachia parasitica</i> (L. Agass.)	97	<i>Stomphia coccarda</i> (O. F. M.)	234
<i>Peachia hastata</i> Gosse	100	— <i>polaris</i> (Dan.)	239
— <i>beckii</i> (Dan. & Kor.)	105	ACTINIARIA. PART II (1922)	
Genus <i>Halochara</i>	106	Preface	1
<i>Halochara producta</i> (Stimps.)	107	I. Description of the species, subtribus <i>Leptactinaria</i>	1
Genus <i>Elaeactis</i>	110	Fam. <i>Bathyphectidae</i> , Genus <i>Bathyphectes</i> , <i>Bathyphectes tuberosa</i> (Dan.)	1
<i>Elaeactis mazilli</i> (Aom.)	111	Genus <i>Dianthesia</i> n. gen., <i>Dianthesia parviflora</i> (Carlg.)	1
Genus <i>Siphonactinopsis</i> n. gen., <i>Siphonactinopsis laevis</i> n. sp.	115	Fam. <i>Sagartidae</i> , Genus <i>Sagartia</i> , <i>Sagartia laevis</i> (Carlg.)	1
Fam. <i>Halecampidae</i> , Genus <i>Halecampa</i>	117	<i>Sagartia chopini</i> (Dal.)	8
<i>Halecampa duodecimcristata</i> (M. Sars)	119	Genus <i>Kallosactis</i> , <i>Kallosactis rosea</i> (Dan.)	1
— <i>arctica</i> Carlg.	120	<i>Kallosactis abyscotti</i> (Dan. & Kor.)	12
— <i>roque</i> (?) n. sp.	123	— <i>abyssorum</i> (Dan.)	14
Genus <i>Cactosama</i> , <i>Cactosama abyssorum</i> Dan.	124	Genus <i>Sagartoplecton</i> , <i>Sagartoplecton laevis</i> Carlg.	16
Fam. <i>Halecampactidae</i> , Genus <i>Haliactis</i> , <i>Haliactis arctica</i> n. sp.	128	<i>Sagartoplecton verrilli</i> nov. nom. (Dan. & Kor.)	18
Fam. <i>Andrakidae</i>	131	— <i>ingolfs</i> Carlg.	19
Genus <i>Andrakia</i>	133	— <i>flavialis</i> (Dan.)	21
<i>Andrakia mirabilis</i> Dan.	134	— <i>abyssorum</i> n. sp.	23
— <i>Thunaria</i> s. <i>Basidaria</i> , Fam. <i>Actinidae</i>	137	— <i>actinothoe</i> <i>andriata</i> (O. F. M.)	24
Genus <i>Actinia</i> , <i>Actinia capina</i> L.	134	— <i>var. tricolor</i> (Price)	25
Fam. <i>Boloceridae</i>	139	— <i>rudialis</i> (O. F. M.)	25
Genus <i>Bolocera</i> , <i>Bolocera tuediae</i> (Johnst.)	140	— <i>laevitatis</i> (Dal.)	26
<i>Bolocera multicornis</i> Verr.	143	— <i>tuberosus</i> (Dan. & Kor.)	28
Fam. <i>Cybrinidae</i> s. <i>Ranodactylidae</i>	145		

Page		Page
50	Genus <i>Actinella</i> , <i>Actinella rubra</i> <i>actinellus</i> (Dan.)	51
51	<i>Actinella rubra</i> (Dan.) n. sp.	53
52	<i>Actinella</i> <i>actinella</i> (Dan.)	55
52	<i>Actinella</i> <i>actinella</i> n. sp.	56
54	<i>Actinella</i> n. sp.	57
56	<i>Actinella</i> n. sp.	57
57	Fam. <i>Metridiidae</i> , <i>Metridium senhale</i> (L.)	59
58	Appendix Fam. <i>Lobosoridae</i> , <i>Lobosoria mottiqua</i> n. sp.	60
59	<i>Lobosoria mottiqua</i> n. sp.	61
59	Fam. <i>Androsolidae</i> , <i>Androsoma pinn</i> Carlzt.	62
61	Fam. <i>Octonoidae</i> ,	64
62	Genus <i>Octonion</i> , <i>Octonion maccum</i> Carlzt.	65
63	Fam. <i>Actinoidae</i> , <i>Actinonia salcata</i> (Penna)	67
63	Fam. <i>Actinostolidae</i> , <i>Parasiparis impile</i> n. sp.	67
64	II. Distribution of the species	69
66	III. Contributions to the Anatomy, Genealogy, and Classification	
67	of the Actiniaria	77
69	Literature	86

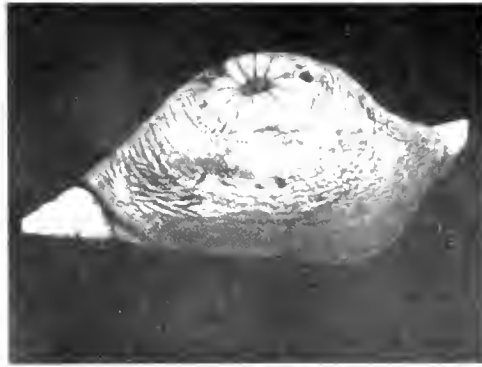
PLATES

Plate I.

- Fig. 1. *Phelliactis coronata*, $\frac{1}{2}$, Trondheimsfjord.
2. *Hormathia digitata*, $\frac{1}{4}$, N.W. to W. of Højen, Skagerrak, 5369.
3. *Hormathia digitata* upon *Hets*, about $\frac{1}{2}$, Iceland.
4. *Paraphelliactis spinosa*, $\frac{1}{4}$, Ingolf exp. St. 95.
5. *Hormathia digitata*, about $\frac{1}{4}$, Kattegat, 10 $\frac{1}{4}$ miles N.W. to W. of Kullen.
6. *Hormathia digitata*, $\frac{1}{4}$, Skagerrak, 44 miles N.W. to W. of Højen, 6990.
7. *Paraphelliactis spinosa*, almost nat. size, Ingolf exp. St. 95.
8. *Hormathia digitata*, about nat. size, Norway, Finnmarken, Rognsund.
9. *Phelliactis robusta*, almost nat. size, Ingolf exp. St. 10.
10. *Chondrophellia coronata*, a little magnified, Ingolf exp. St. 18.
11. *Hormathia digitata*, $\frac{1}{4}$, West Greenland, Ingolf exp. St. 29.
12. *Phelliactis robusta*, $\frac{1}{4}$, Ingolf exp. St. 32.



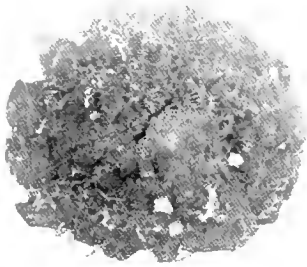
1



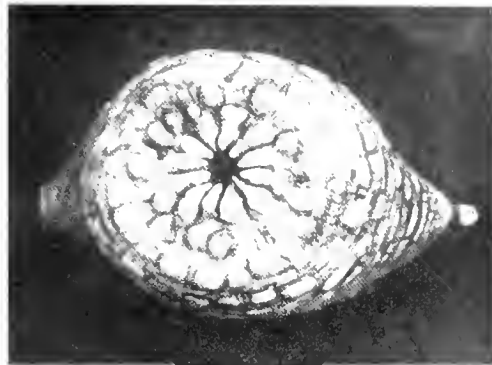
2



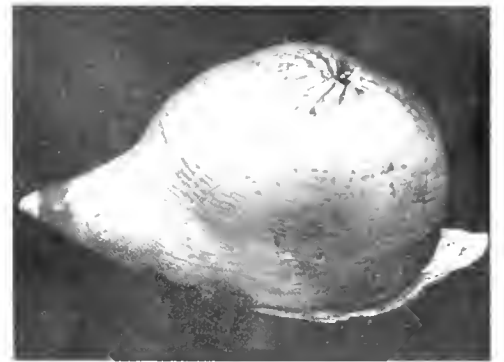
3



4



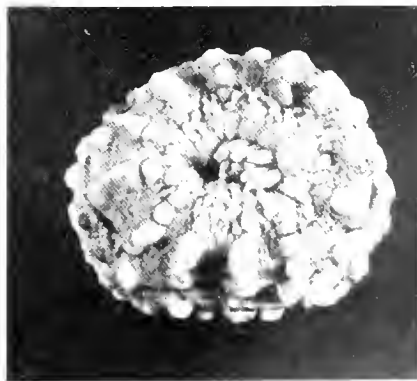
5



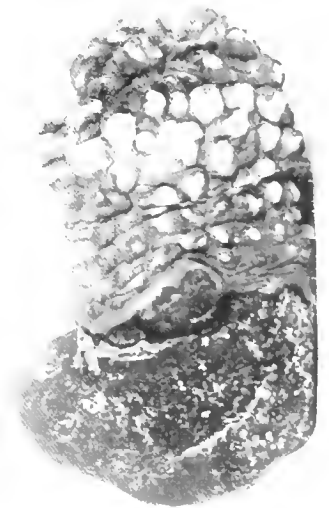
6



7



8



9



10



11



12

Plate II.

Actinopterygii.

- Fig. 6. A little reduced, Godthaab exp. St. 61.
- 7. ¹/₄, Tjalfe exp. St. 100.
- 8. A little reduced, Godthaab exp. St. 77.
- 9. A little reduced, U.S. Fish Com. 49 03' N. 70 28' W.
- 10. A little reduced, Godthaab exp. St. 77.

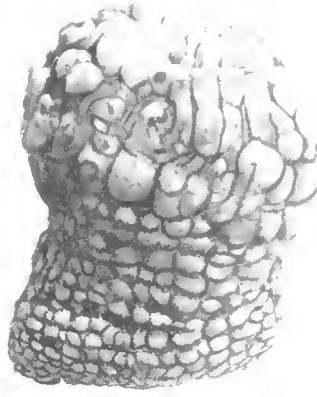
Fig. 6. A little reduced, Godthaab exp. St. 61.
7. ¹/₄, Tjalfe exp. St. 100.
8. A little reduced, Godthaab exp. St. 77.
9. A little reduced, U.S. Fish Com. 49 03' N. 70 28' W.
10. A little reduced, Godthaab exp. St. 77.



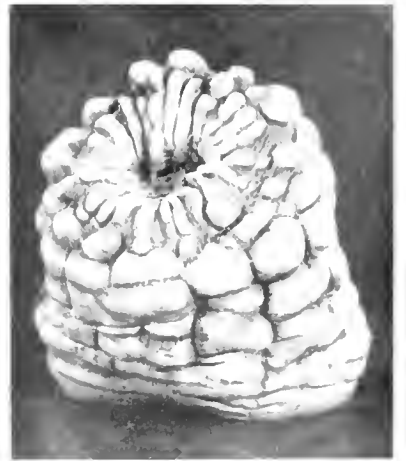
1



2



3



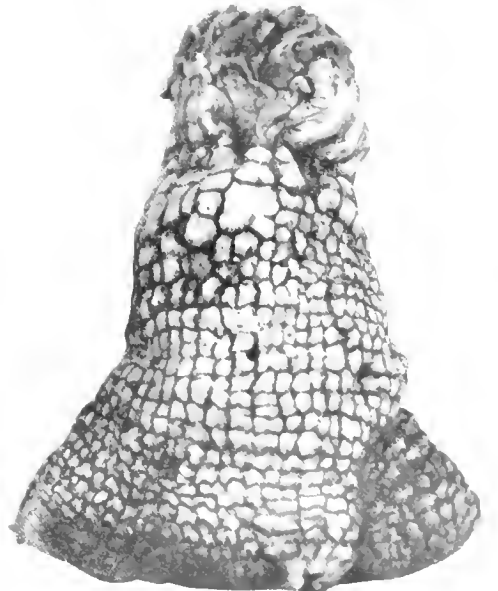
4



5



6



7



8



9



10

Plate III.

- 1. *Dactylopusia* sp. $\times 100$, $\frac{1}{1}$, River leg.
- 2. *Hornothia* sp. $\times 100$, $\frac{1}{1}$, Ost.
- 3. *Hornothia* sp. $\times 100$, $\frac{1}{1}$.
- 4. *Kryptothoa* sp. $\times 100$, about $\frac{1}{1}$, Norway, Hjeltefjord.
- 5. *Leptothoa* sp. $\times 100$, $\frac{1}{1}$, Jæderen, Norway.
- 6. *Leptothoa* sp. $\times 100$, $\frac{1}{2}$, Trondheimsfjord.
- 7. *Leptothoa* sp. $\times 100$, $\frac{1}{1}$, Øst, Bergen, Norway.
- 8. *Sagittiopton* *undulatus*, $\frac{1}{2}$, Rockall.
- 9. *Hornothia* sp. *digitata* var. *cegae*, about $\frac{1}{1}$, Kara Sea.

- Fig. 10. *Bathyphebia* *margaritacea*, $\frac{2}{1}$.
- 11. *Hornothia* *digitata* var. *cegae*, $\frac{1}{1}$, E. of Tajmyr Peninsula, Siberia.
- 12. *Hornothia* *digitata* var. *cegae*, $\frac{1}{1}$, Blischmi, Siberia.
- 13. *Sagittiopton* *laccatus*, $\frac{1}{1}$, The Sound.
- 14. *Allantactis* *parasitica*, $\frac{1}{1}$, E. of Tajmyr Peninsula, Siberia.
- 15. *Sagittiopton* *undulatus*, $\frac{1}{1}$, The Sound.
- 16. *Sagittiopton* *undulatus*, $\frac{2}{1}$, Marstrand.

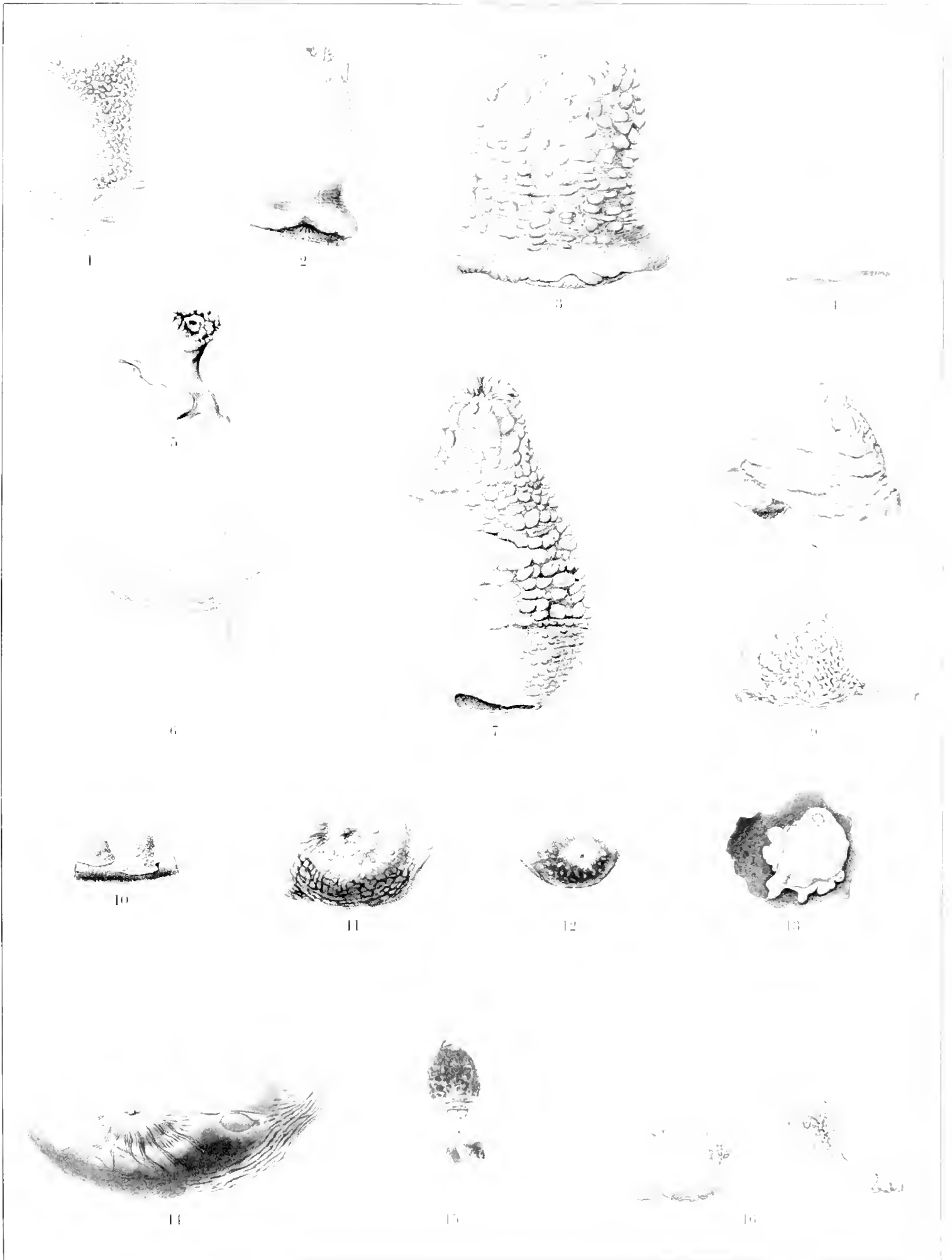
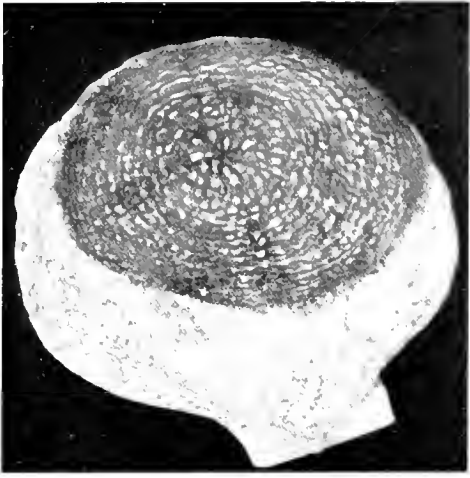


Plate IV.

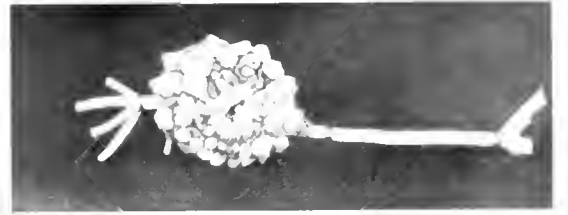
- Fig. 1. *Phallactis robusta*. A little magnified. Michael Sars exp. No. 10. St. 14.
 2. *Syllis ruficornis*. $\frac{1}{4}$. Helgefjord. Brodholmen, Norway.
 3. *Alciostoma* sp. A little magnified. Trondhemfjord.
 4. *Phallactis robusta*. $\frac{1}{4}$. Ingolf exp. St. 40.
 5. *Syllis ruficornis* var. *naana*, about $\frac{1}{4}$. Finnmarken, Russia.
 Fig. 6. *Hornathia nodosa*. $\frac{1}{4}$. Faroe Isl.
 7. *Apudrakia parca*. Very much magnified.
 8. *Phallactis robusta*. About $\frac{1}{4}$. Sofia exp. 1883.
 9. *Allantactis parasitica*. About $\frac{3}{4}$. East Greenland.
 10. *Hornathia nodosa*. $\frac{1}{4}$. Hammarfest.
 11. *Phallactis bertwigi*. $\frac{2}{3}$. Ingolf exp. St. 61.



1



2



3



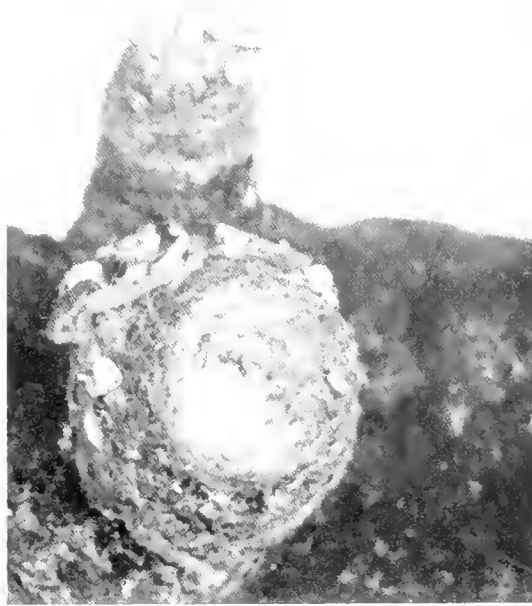
3



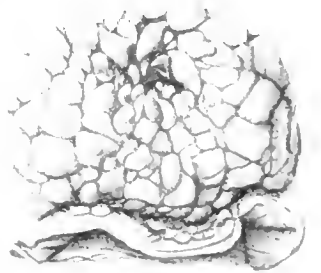
7



6



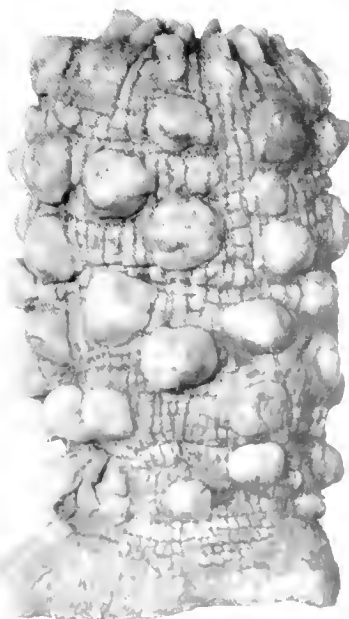
7



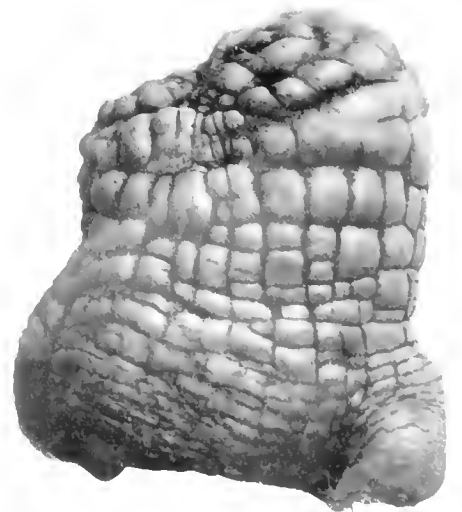
8



9



10



11

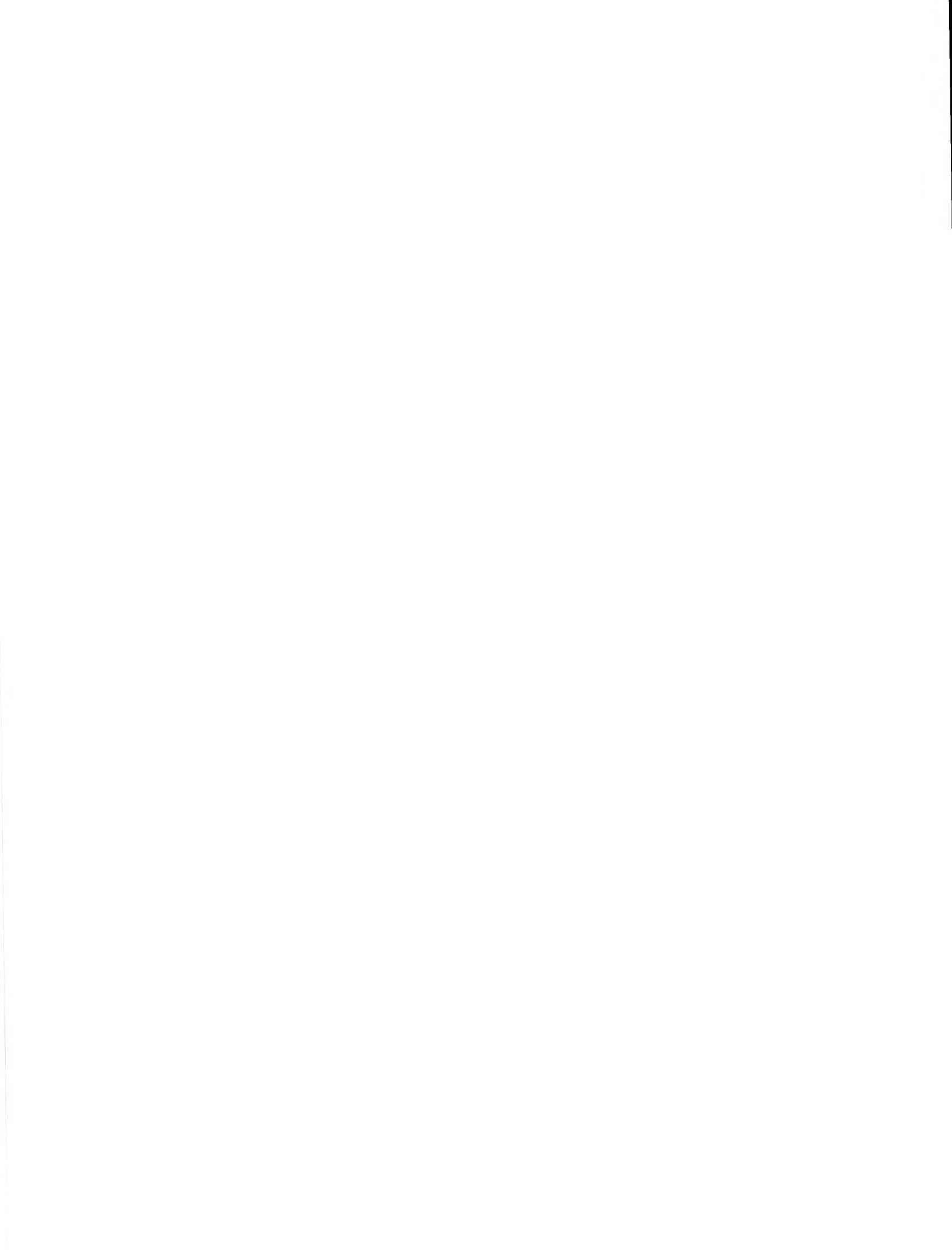


Plate V.

- Fig. 1. *Prorhynchus glacialis*. Section of the scapus showing tenaculi.
2. *Prorhynchus glacialis*. Section of the scapus with tenaculum.
3. *Stomatopoda glacialis*. Section of the scapus with a tenaculum.

- Fig. 4. *Hormathia digitata*, Kvanefjord, W. Greenland. Section of scapus showing islets of parasites in the mesogloea.



1



2



3



4

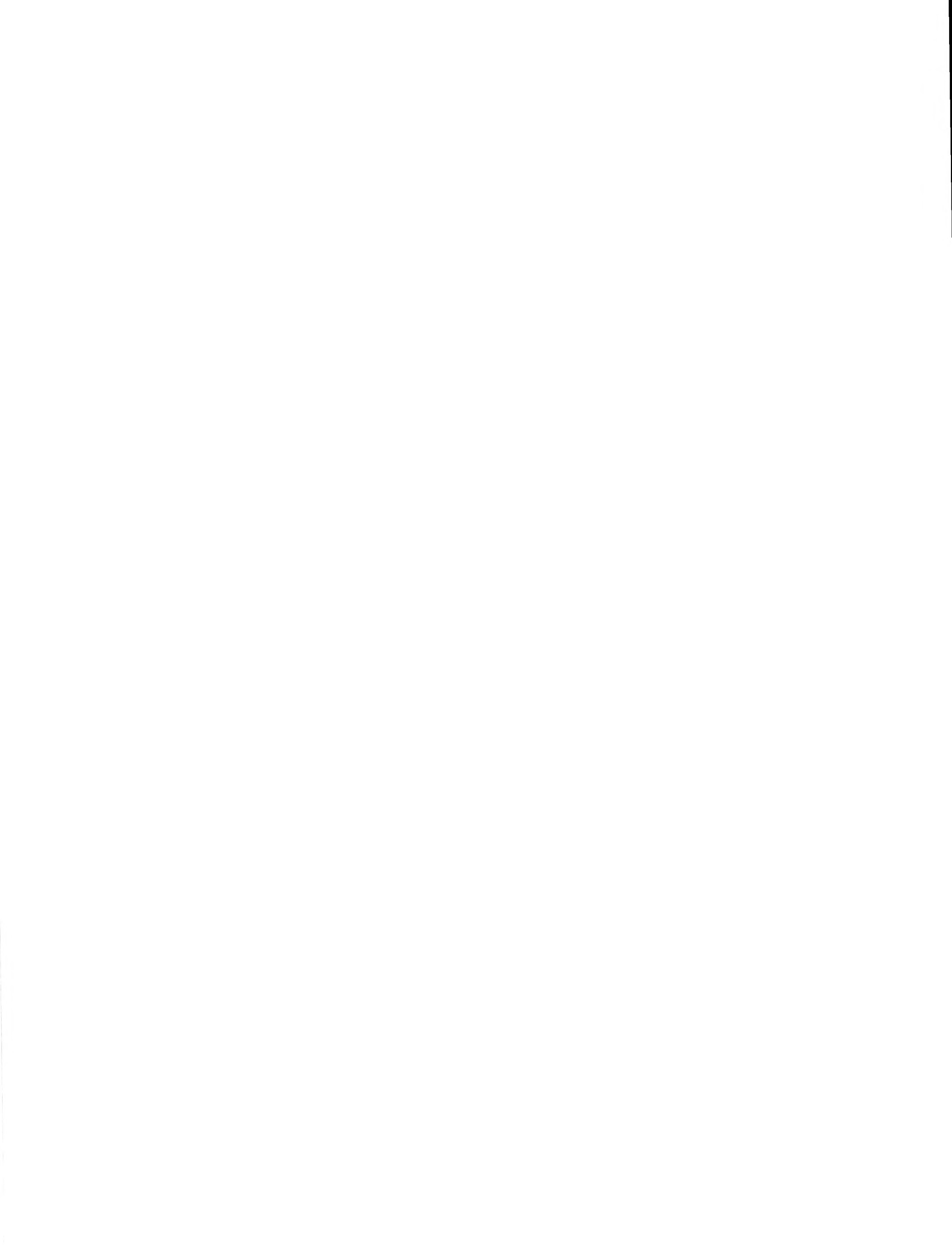


Plate VI.

1. Section of the scapus showing a parasite, *Hornathia* sp. (H.).
2. Parasites in the nosogloea of *Hornathia* sp. (H.).
3. Section of the column showing a parasite, *Saprotrocton* sp. (S.).

Fig. 5. *Hornathia diptata* var. *vequa*. Section of the scapus.
6. *Saprotrocton andatus*. Section of the column showing two tenaculi (t.).
7. *Aulacitia parca*. About $\frac{1}{4}$.



1



2



3



4



5



6



7

THE INGOLF-EXPEDITION

1895-1896

THE LOCALITIES, DEPTHS, AND BOTTOMTEMPERATURES OF THE STATIONS

Station Nr.	Date	Lat. N.	Long W.	Depth in m.	Bot- tom- temp.	Sta- tion Nr.	Date	Lat. N.	Long W.	Depth in m.	Bot- tom- temp.	Sta- tion Nr.	Date	Lat. N.	Long W.	Depth in m.	Bot- tom- temp.
1895						1896											
1	11-V	62° 30'	8° 21'	249	7.2	24	25-VI	63° 06'	56° 00'	2258	2.4	45	11-V	61° 32'	9° 43'	1211	4.17
2	12 -	63° 04'	9° 22'	493	5.3	25	26 -	63° 30'	54° 25'	1096	3.3	46	- -	61° 32'	11° 36'	1356	2.40
3	- -	63° 35'	10° 24'	512	0.5			63° 51'	53° 03'	256		47	12 -	61° 32'	13° 40'	1789	3.23
4	13 -	64° 07'	11° 12'	416	2.5	26	- -	63° 57'	52° 41'	64	0.6	48	- -	61° 32'	15° 11'	2165	3.17
5	- -	64° 40'	12° 09'	292				64° 37'	51° 24'	205		49	13 -	62° 07'	15° 07'	2109	2.91
6	16 -	63° 43'	14° 34'	170	7.0	27	1-VII	64° 54'	55° 10'	740	3.8	50	- -	62° 43'	15° 07'	1921	3.13
7	17 -	63° 13'	15° 41'	1130	4.5	28	- -	65° 14'	55° 42'	791	3.5	51	15 -	64° 15'	11° 22'	128	7.32
8	19 -	63° 56'	24° 40'	256	6.0	29	5 -	65° 31'	54° 31'	128	0.2	52	- -	63° 57'	13° 32'	791	7.87
9	20 -	64° 18'	27° 00'	555	5.8	30	10 -	66° 50'	54° 28'	41	4.05	53	16 -	63° 15'	15° 07'	1497	3.08
10	- -	64° 24'	28° 50'	1484	3.5	31	11 -	66° 35'	55° 54'	166	4.6	54	18 -	63° 08'	15° 40'	1301	3.9
11	21 -	64° 34'	31° 12'	2448	1.6	32	11 -	66° 35'	56° 38'	599	3.9	55	19 -	63° 33'	15° 02'	595	5.9
12	22 -	64° 38'	32° 37'	1958	0.3	33	12 -	67° 57'	55° 30'	66	0.8	56	- -	64° 00'	15° 09'	128	7.57
13	- -	64° 47'	34° 33'	1171	3.0	34	18 -	65° 17'	54° 17'	104		57	20 -	63° 37'	13° 02'	659	3.4
14	- -	64° 45'	35° 05'	331	4.4	35	- -	65° 16'	55° 05'	682	3.6	58	- -	64° 25'	12° 09'	397	0.8
15	4-VI	66° 18'	25° 59'	621	0.75	36	28 -	61° 50'	56° 21'	2702	1.5	59	- -	65° 00'	14° 16'	584	0.4
16	5 -	65° 13'	26° 58'	471	6.1	37	29 -	60° 17'	54° 05'	3229	4.4	60	21 -	65° 09'	12° 27'	234	0.9
17	16 -	62° 49'	26° 55'	1403	3.4	38	30 -	59° 12'	51° 05'	3521	4.3	61	- -	65° 03'	13° 06'	104	0.4
18	17 -	61° 41'	30° 29'	2137	3.0	39	9-VIII	62° 00'	22° 38'	1629	2.9	62	31 -	63° 48'	19° 12'	436	7.92
19	18 -	60° 29'	31° 14'	2949	2.4	40	- -	62° 00'	24° 36'	1591	3.3	63	1-VI	62° 40'	19° 05'	1596	4.0
20	20 -	58° 20'	40° 48'	3192	1.5	41	12 -	61° 39'	17° 10'	2345	2.0	64	- -	62° 06'	19° 00'	1960	3.4
21	21 -	58° 01'	44° 45'	2595	2.4	42	14 -	61° 41'	19° 17'	1177	0.4	65	2 -	61° 33'	19° 00'	2051	3.0
22	22 -	58° 10'	48° 25'	3474	1.4	43	- -	61° 42'	19° 41'	1215	0.05	66	- -	61° 33'	20° 43'	2124	3.5
23	24 -	60° 43'	56° 00'	<small>not made</small>		44	- -	61° 42'	9° 36'	1026	4.8	67	3 -	61° 30'	22° 30'	1836	3.0

THE DANISH INGOLF-EXPEDITION

VOLUME V

13

OCTOCORALLIA

(STOLONIFERA TELESTACEA NENIDEA ALCYONACEA GORGONACEA)

BY

F. JENSENIUS MADSEN

WITH 1 PLATE AND 53 FIGURES IN THE TEXT



COPENHAGEN

PRINTED BY BIANCO LUNO A/S

1944

Contents.

	Page		Page
Order <i>Chalcididae</i> (Hymenoptera)	3	Genus <i>Parapopra</i> Milne-Edwards	32
<i>Chalcididae</i> (Hymenoptera)	3	<i>arborea</i> (Linne)	32
<i>Chalcididae</i> (Hymenoptera)	4	Family: <i>Scapellatoidea</i> Amyvilibus	32
<i>Chalcididae</i> (Hymenoptera)	5	Genus: <i>Anthobola</i> Verrill	32
<i>Chalcididae</i> (Hymenoptera)	5	<i>grandiflora</i> (M. Saito)	32
<i>Chalcididae</i> (Hymenoptera)	6	Suborder: <i>Habitania</i> Studer	33
<i>Chalcididae</i> (Hymenoptera)	6	Family: <i>Acanthopopridae</i> Kükenthal	33
<i>Chalcididae</i> (Hymenoptera)	8	Genus: <i>Acanthopopra</i> Gray	33
<i>Chalcididae</i> (Hymenoptera)	9	<i>armita</i> Verrill	33
<i>Chalcididae</i> (Hymenoptera)	10	Family: <i>Mantoidae</i> Gray	36
<i>Chalcididae</i> (Hymenoptera)	10	Genus: <i>Paramantia</i> Kolliker	36
<i>Chalcididae</i> (Hymenoptera)	11	<i>platanus</i> (Linne)	36
<i>Chalcididae</i> (Hymenoptera)	11	Genus: <i>Trachepanicea</i> Deichmann	37
<i>Chalcididae</i> (Hymenoptera)	11	<i>kalethali</i> (Broch)	37
<i>Chalcididae</i> (Hymenoptera)	11	Family: <i>Primaoidae</i> Verrill	38
<i>Chalcididae</i> (Hymenoptera)	11	Subfamily: <i>Primaoidae</i> Vershuys	39
<i>Chalcididae</i> (Hymenoptera)	12	Genus: <i>Primaon</i> Lamontoux	39
<i>Chalcididae</i> (Hymenoptera)	15	<i>residuipanis</i> (Günther)	39
<i>Chalcididae</i> (Hymenoptera)	15	Subfamily: <i>Primaoidae</i> Vershuys	39
<i>Chalcididae</i> (Hymenoptera)	16	Genus: <i>Primaella</i> Gray	39
<i>Chalcididae</i> (Hymenoptera)	16	<i>janqisani</i> n. sp.	39
<i>Chalcididae</i> (Hymenoptera)	20	Family: <i>Gonopidae</i> Dana	43
<i>Chalcididae</i> (Hymenoptera)	20	Genus: <i>Stenopopra</i> Verrill	43
<i>Chalcididae</i> (Hymenoptera)	20	<i>haralis</i> Kramp	43
<i>Chalcididae</i> (Hymenoptera)	20	<i>rosa</i> Grzeg.	43
<i>Chalcididae</i> (Hymenoptera)	24	Family: <i>Chrysopopridae</i> Verrill	44
<i>Chalcididae</i> (Hymenoptera)	23	Subfamily: <i>Leptopopridae</i>	44
<i>Chalcididae</i> (Hymenoptera)	24	Genus: <i>Ridicopus</i> Stearns	44
<i>Chalcididae</i> (Hymenoptera)	24	<i>challapici</i> (Wright & Studer)	44
<i>Chalcididae</i> (Hymenoptera)	24	<i>quacis</i> (Verrill)	46
<i>Chalcididae</i> (Hymenoptera)	24	Subfamily: <i>Chrysopopridae</i> Studer	49
<i>Chalcididae</i> (Hymenoptera)	24	Genus: <i>Chrysopopra</i> Duchassaing & Michelotti	49
<i>Chalcididae</i> (Hymenoptera)	24	<i>apissata</i> (Verrill)	49
<i>Chalcididae</i> (Hymenoptera)	25	<i>campanula</i> n. sp.	54
<i>Chalcididae</i> (Hymenoptera)	26	Family: <i>Isidellae</i> Studer	56
<i>Chalcididae</i> (Hymenoptera)	28	Genus: <i>Isidella</i> Gray	56
<i>Chalcididae</i> (Hymenoptera)	29	<i>arbuscula</i> (Johnson)	56
<i>Chalcididae</i> (Hymenoptera)	29	Genus: <i>Isidella</i> Gray	57
<i>Chalcididae</i> (Hymenoptera)	29	<i>lapidaria</i> (M. Saito)	57
<i>Chalcididae</i> (Hymenoptera)	30	Zoogeographical and ecological remarks	58
<i>Chalcididae</i> (Hymenoptera)	30	List of synonyms	60
<i>Chalcididae</i> (Hymenoptera)	32	List of literature	63
<i>Chalcididae</i> (Hymenoptera)	32		

Introduction.

The present paper is a systematic-geographical treatment of the recent Octocorals, apart from the Pennatulacea, from the North-Atlantic Ocean north of about 50° latitude and adjacent seas. It should, however, be noted that species which apparently have their northernmost limit in the Lusitanian area, at the latitudes of Ireland and England, are not discussed here. (A list of these species: *Ceratosis grayi*, *Chelidonisis aurantiaca*, *Corallium johnsoni*, *Stachyodes verlugsi* etc. is to be found in STEPHENS' paper 1909).

The material of northern Octocorals in the Zoological Museum of Copenhagen which forms the basis of this paper was mainly collected by the Danish "Ingolf" Expedition, but amongst other also the Danish research steamers "Thor" and "Dana" collected several specimens. The Pennatulacea of the "Ingolf" Expedition were described in 1901 by the late Professor, Dr. H. F. E. JUNGERSEN, who also in various papers from 1915-1917 several times announced the report of the other Octocorals from the "Ingolf" Expedition as being in preparation. JUNGERSEN died, however, in 1917 before this work was finished, leaving no written notes apart from a manuscript on *Anthomastus*, on which species he in 1916 at the sixteenth Scandinavian Naturalists' meeting had read a small paper, published in the Danish language in 1918. The larger posthumous manuscript was published in 1927 under edition of HJ. BROCH. In three other papers from 1915-1917 JUNGERSEN dealt with the Octocorals from the waters around Greenland and the Octocorals taken in the Norwegian Sea by the Norwegian research steamer "Michael Sars". These papers include many zoogeographical remarks and very extensive synonym-lists, but they contain neither closer discussion of the systematics nor descriptions of the material, and the only new species mentioned, *Stenogorgia borealis*, was but a nomen nudum. *Stenogorgia borealis* was, however, described in 1930 by KRAMP who in various papers has dealt with the Octocorals of the Faroic- and Icelandic-shelf and from the Davis Strait. BROCH (1928) treated the northern Nephthylidae, but since JUNGERSEN no authors have dealt more thoroughly with the northern deep-sea Octocorals. And though more than 25 years have elapsed since the death of JUNGERSEN, the great material still is of the same importance as in 1916 and the three species, which JUNGERSEN on the labels considered new ones, are still undescribed.

JUNGERSEN had before his death sorted out and provisionally labelled most of the large collection of Octocorals preserved in the Zoological Museum at his time; only the material of small stoloniferous forms was mostly marked with queries or not sorted out at all. The three species which JUNGERSEN designated as new were *Chrysogorgia campanula*, *Primoella jungersei* (named in this way as the name *gracilis* proposed by JUNGERSEN for this delicate little species had later been used by MOLANDER for another species of *Primoella*), and the Stoloniifera *Charularia greggii*. Moreover, the material of small stoloniferous specimens included four other species new to science, two of them of the greatest interest as the one evidently represents a new genus, and the other belongs to a very little known genus which without doubt should be ranged in a new family, though at present it is impossible with certainty to state to which order this family belongs.

There are only some few of JUNGERSEN's determinations which the author has not been able to confirm. The *Radicipes* specimens which were secured by the "Ingolf" were by JUNGERSEN iden-

tified with *Strophogorgia challengeri* Wright & Studer, to which species JUNGERSEN with a query also referred *Lepidogorgia gracilis* Verrill (JUNGERSEN, 1915, p. 1183). This, however, is clearly contradicted by the material which includes two different species, the one, only represented by one specimen, identical with *R. challengeri*, the other, represented by several specimens, identical with *R. gracilis*. Beyond doubt, however, JUNGERSEN himself would have discovered and corrected this lapsus during the closer examination and preparation for the intended manuscript. JUNGERSEN also (1917) united *Paramuricea plumosus* (L.) and *P. kükenthalii* Broch, but in some of the material of the latter species he had on the labels written *P. plumosus* f. *kükenthalii*. Finally, the present author disagrees with JUNGERSEN in the view upon the northern Nephthylidae with retractile anthocodiae, as he arrived at the conclusion that among these only one species can be distinguished, whereas JUNGERSEN in his papers (1915-1917) maintained two species in the group.

Among the authors who have contributed to our knowledge of the northern deep-sea Octocorals may be mentioned M. SARS, DANIELSEN, GRIEG and of more modern students besides JUNGERSEN, MOLANDER and BROCH. A general account of the northern Octocorals has not yet been given. JUNGERSEN, as mentioned, has in 1901 treated the Pennatulacea monographically, and MOLANDER (1915) gave a survey of the Aleyonacea s.lat. Though in this paper stress is laid upon the deep sea species, short references have been made to the other species at present recognized in the area, and thus the reports of the "Ingolf" Expedition give a synopsis of the hitherto known northern Octocorals.

As to the taxonomic arrangement the author has followed a system such as proposed by HICKSON (1930), however, with the emendation that the family Xenidae is regarded as a distinct order (cf. BOCK 1938). Thus, as also the Tubiporidae should most conveniently be regarded as a special order (cf. BOCK 1938), the following eight orders¹⁾ are recognized within the Octocorallia:

- Stoloniifera,
- Telestacea
- Xenidea,
- Aleyonacea,
- Tubiporidae,
- Heliporidae,
- Gorgonacea { Scleraxonia,
- } Holaxonia,
- Pennatulacea.

Of these orders only the Tubiporidae and the Heliporidae have not hitherto been found in the northern seas.

As to the terminology, the author hopes to have used it in such a way that confusion may not arise as often has been the case in the literature on Octocorals. Explanations of most of the terms used may be found in HICKSON'S paper (1930) or in the list of "Technical Terms" in DEICHMANN'S paper (1936 p. 28-29). The signification of other terms used is thought to appear directly from the use.

¹⁾ The six first-mentioned orders are by Bock regarded as sub-orders of the order Aleyonacea; it seems however more convenient to regard them as orders coordinate with the Gorgonacea and Pennatulacea.

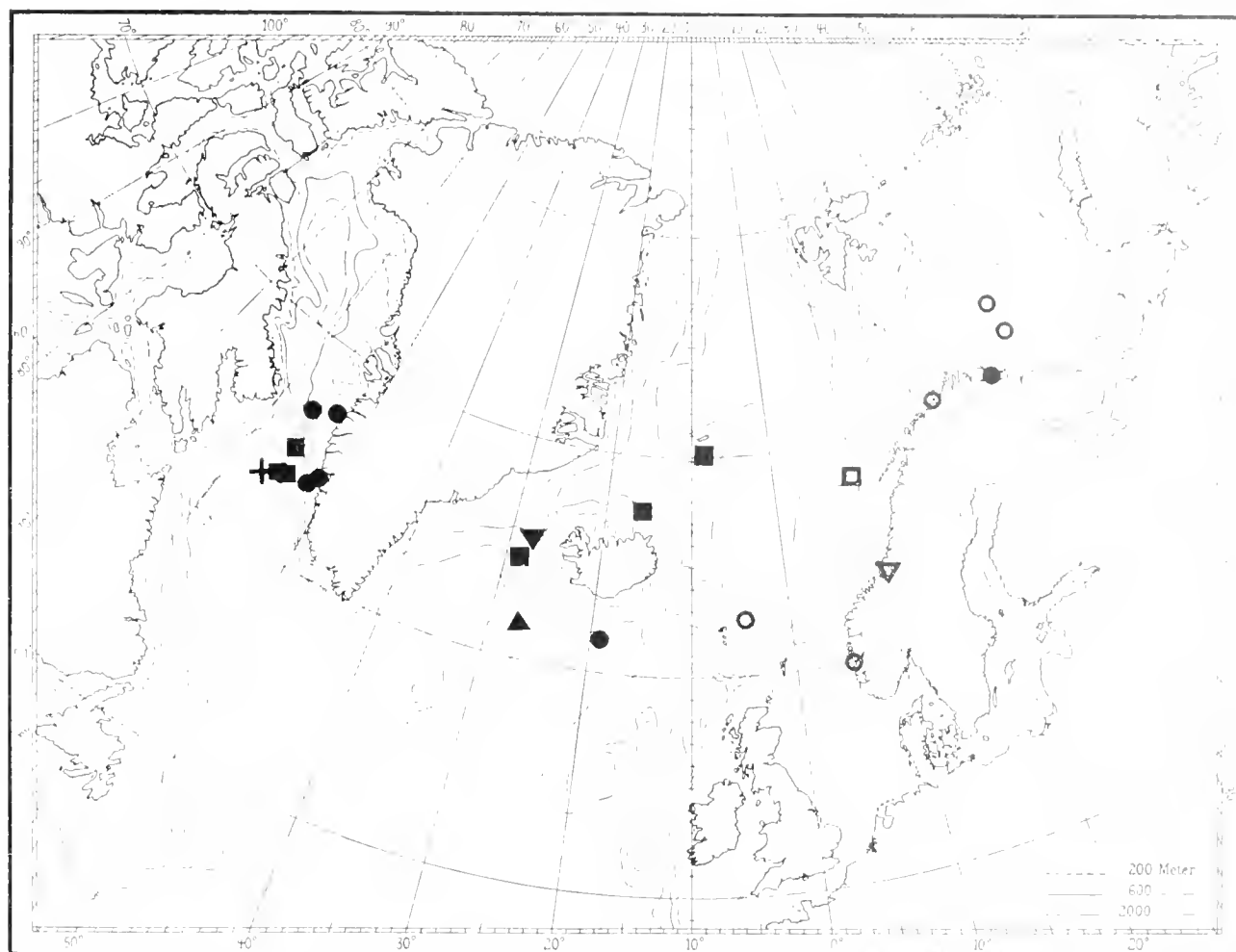


Fig. 1. ● *Clavularia arctica* (M. Sars), ■ *Clavularia alba* (Grieg), ▲ *Clavularia greppi* n. sp., + *Clavularia leddeni* n. sp. & *Clavularia* n. sp., ▼ *Anthelia borealis* (Køren & Danielssen). Black figures: localities represented in the Zoological Museum of Copenhagen. Open figures: localities recorded in the literature.

If thus DEICHMANN's identification of the specimen is correct, it may be said that VERRILL's description of the species only gives a poor idea of it, and the generic name would then have to be retained for such species which have a 16-pointed anthocodial crown as described by BROCH and then can not be used for the northern *Clavularia arctica* as MOLANDER does in 1929.

It may be noted that all the species mentioned in the Clavulariidae in this paper have pinnate tentacles, which, apart from the unpaired tip, have only one single row of pinnulae on each side.

Genus *Clavularia* Quoy & Gaimard 1830.

Clavularia arctica (M. Sars 1860).

- Rhizoxema arctica* M. Sars, 1860, p. 2.
- Clavularia arctica* Køren & Danielssen, 1883, p. 12, pl. 3 figs. 25-35.
- Clavularia arctica* Broch, 1912 *c*, p. 3-8, figs. 1-3.
- Jungersen, 1915, p. 1461.
- 1917, p. 1-5.
- Rylov, 1926 *b*, p. 237-239, 243-241, fig. 5.
- J. A. Thomson, 1927, p. 7.
- Cornulariella* Molander, 1929, p. 23.
- Clavularia* Kraup, 1932, p. 16, 19.

For further references vide BROCH 1912 *c* and JUNGENSEN 1917.

Description: Vide BROCH 1912 *c*.

Material:

- Vadso (Varangerfjord, northern Norway) 95-110 m. Colony of *Clavularia arctica* Køren & Danielssen.
- Store Hellefiskebanke (N. of Holstenborg, W. Greenland) 90 m. "Fylla" 1881, 1 colony.
- Davis Strait 188 m. "Fylla" 1881, Several colonies.
- 63° 56' N. 53° 12' W. 215 m. Wandel 1889, 1 colony.
- 66° 19' N. 56° 28' W. 150 m. Wandel 1889, 1 colony.
- 61° 33' N. 49° 00' W. 2051 m. 3 C. "Ingolf" St. 65, 1 colony.
- Godthaabsfjord (W. Greenland) "Tulle" St. 52, 1 colony.

Remarks: The material consists of ribbon-like, so-called anastomosing stolons, creeping upon stones, hydroids, sponges, barnacles, polyzoan colonies, or mollusc shells.

In most of the specimens the zooids are placed at fairly regular intervals of a few millimetres. In one colony (the "Tulle" St. 52) the more than half a hundred zooids are, however, so densely crowded that there are no spaces between them. Most zooids are more or less contracted and nearly all have the anthocodium retracted into the anthostele. The largest zooids, which are those from the type locality Vadso, have an anthostele which measures about 43 mm in height, having a diameter of about 3 mm. In the other colonies the zooids are somewhat smaller, the largest anthostele measures in some of the colonies from the Davis Strait about 10 mm in height, whereas in other specimens from the same locality it measures but 5 mm. BROCH states for the largest extended zooids in his material a height of 16 mm (broken) by 2.5-3 mm in breadth.

The spiculation agrees in all the specimens, apart from that

of the species. The description of *Clavularia* given by GREG (1888) is not complete, the type locality being given as "the island of St. Olaf" (p. 7). However, it is not clear what locality is meant in this connection. The type locality is given as "the island of St. Olaf" (p. 7). However, it is not clear what locality is meant in this connection. The type locality is given as "the island of St. Olaf" (p. 7). However, it is not clear what locality is meant in this connection.

- 1. GREG, 1888, p. 7, fig. 15, pl. 1, figs. 1-34.
- 2. HEDLEY, 1894, p. 376.
- 3. MOLLER, 1900, p. 383.
- 4. LANGESEN, 1915, p. 1162.
- 5. ANDERSEN, 1916, p. 28.
- 6. KIRKENIDAL, 1919, p. 99.
- 7. MOLANDER, 1919, p. 37.
- 8. MOLANDER, 1929, p. 36-37, pl. 3, fig. 9.
- 9. KRISTENSEN, 1932, p. 4.
- 10. MOLANDER, 1936, p. 37.
- 11. MOLANDER, 1938, p. 37.
- 12. MOLANDER, 1949, p. 99.
- 13. MOLANDER, 1949, p. 99.
- 14. MOLANDER, 1949, p. 99.
- 15. MOLANDER, 1949, p. 99.
- 16. MOLANDER, 1949, p. 99.
- 17. MOLANDER, 1949, p. 99.
- 18. MOLANDER, 1949, p. 99.
- 19. MOLANDER, 1949, p. 99.
- 20. MOLANDER, 1949, p. 99.

Clavularia alba Greg 1888

- 1. GREG, 1888, p. 7, 15, pl. 1, figs. 1-34.
- 2. HEDLEY, 1894, p. 376.
- 3. MOLLER, 1900, p. 383.
- 4. LANGESEN, 1915, p. 1162.
- 5. ANDERSEN, 1916, p. 28.
- 6. KIRKENIDAL, 1919, p. 99.
- 7. MOLANDER, 1919, p. 37.
- 8. MOLANDER, 1929, p. 36-37, pl. 3, fig. 9.
- 9. KRISTENSEN, 1932, p. 4.
- 10. MOLANDER, 1936, p. 37.
- 11. MOLANDER, 1938, p. 37.
- 12. MOLANDER, 1949, p. 99.
- 13. MOLANDER, 1949, p. 99.
- 14. MOLANDER, 1949, p. 99.
- 15. MOLANDER, 1949, p. 99.
- 16. MOLANDER, 1949, p. 99.
- 17. MOLANDER, 1949, p. 99.
- 18. MOLANDER, 1949, p. 99.
- 19. MOLANDER, 1949, p. 99.
- 20. MOLANDER, 1949, p. 99.

The material consists of some small fragments of stolon which are attached to polyzoan colonies and a dead fragment of *Lophalia*. Altogether fifteen zooids are preserved. The material consists of some small fragments of stolon which are attached to polyzoan colonies and a dead fragment of *Lophalia*. Altogether fifteen zooids are preserved.

The material consists of some small fragments of stolon which are attached to polyzoan colonies and a dead fragment of *Lophalia*. Altogether fifteen zooids are preserved.

synonymous with *Anthothela grandiflora* (M. Sars), by a re-examination in 1918 he, however, changed his view and now regarded the species as a membranous form of *Gaisemua futuosa* (M. Sars). There is, however, no doubt that *Rhaserania alba* is a distinct species which is neither identical with *Anthothela grandiflora* nor with *Gaisemua futuosa*, and to this conclusion MOLANDER also arrived in his paper of 1929 where on p. 36-37 he withdraws his previous statements, holding that the species should be regarded as a membranous species of the genus *Anthothela*, though not identical with *A. grandiflora*.

When DELIUMANN in 1936 enumerated *R. alba* in the synonymy list of *Anthothela grandiflora* it was on the basis of MOLANDER's statement, since no specimens were examined, and KIRKENIDAL, who in 1919 mentions *R. alba* among the synonyms to *Anthothela grandiflora*, in 1921 omitted it here.

Distribution. Besides the above-mentioned localities the species is only known from the type locality. Its distribution comprises thus the Norwegian Sea, Danmark Strait, and Davis Strait in depths from 552-1181 m. Chart of distribution, fig. 1.

Clavularia griegii n. sp.

Material:

- 61° 41' N, 27° 00' W, 913 m, 64 C, "Ingolf" St. 81.

Diagnosis: Membranous Octocorals with zooids distinctly divided into a rigid, thick-walled anthostele and an entirely retractile anthocodia. Size of verrucae up to 2 mm in height by 3.5 mm in diameter at the base. Sclerites of stolon warty, pointed spindles of two types, the most common, slender ones measuring about 0.4 - 0.63 mm, and scattered among these some larger ones measuring about 0.6 - 0.075 mm. Sclerites of anthostele warty and pointed spindles measuring up to 1.1 mm in length, and besides a few which have an upper more irregularly formed end, sometimes giving the sclerite a "club" shaped appearance. Sclerites of the anthocodia in the lower part scattered, transversally arranged, straight, slender spindles, in the upper part somewhat larger and often club-shaped ones, measuring up to about 0.8 mm, arranged on chevron and continuing uninterruptedly in the sclerite rows of the tentacle dorsa. In the lower part of tentacle dorsum club-shaped sclerites arranged in a double row with the blunt ends pointing outwards. In the outer part small, flat, transversally arranged rods. In pinnulae numerous small, flat sclerites.

Type. In the Zoological Museum of Copenhagen.

Description. The material consists of some small fragments of stolon which are attached to polyzoan colonies and a dead fragment of *Lophalia*. Altogether fifteen zooids are preserved.

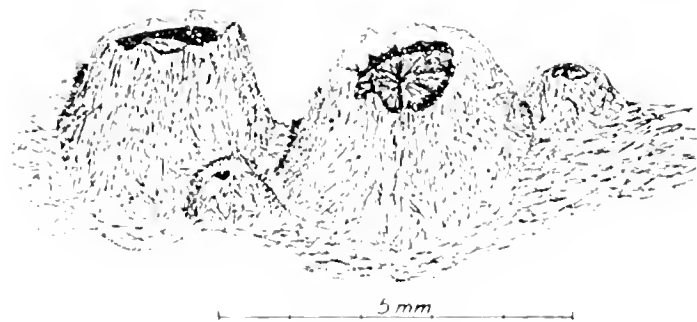


Fig. 2. *Clavularia griegii* n. sp. Sketch of a fragmentary colony showing the club-shaped verrucae which are formed when the anthocodiae are retracted into the anthostelae.

The largest fragment of a stolon is 43 mm in length measuring, where it is broadest, about 4 mm in width. The stolon is branched, the side branch measuring about 5 mm in length, and it is provided with eight rather closely placed zooids. The largest interval between two zooids is about 4 mm. Another fragment of a stolon

(fig. 2) measures about 9.5 mm in length and 3 mm in width, being provided with four zooids, which all have the anthocodia retracted into the anthostele. The two largest anthostelae measure 2 mm in height and 3 and 3.5 mm respectively in diameter at the base. The interval between them is scarcely 0.3 mm. The smaller zooids which are placed close at each other have anthostelae about 0.75 mm high.

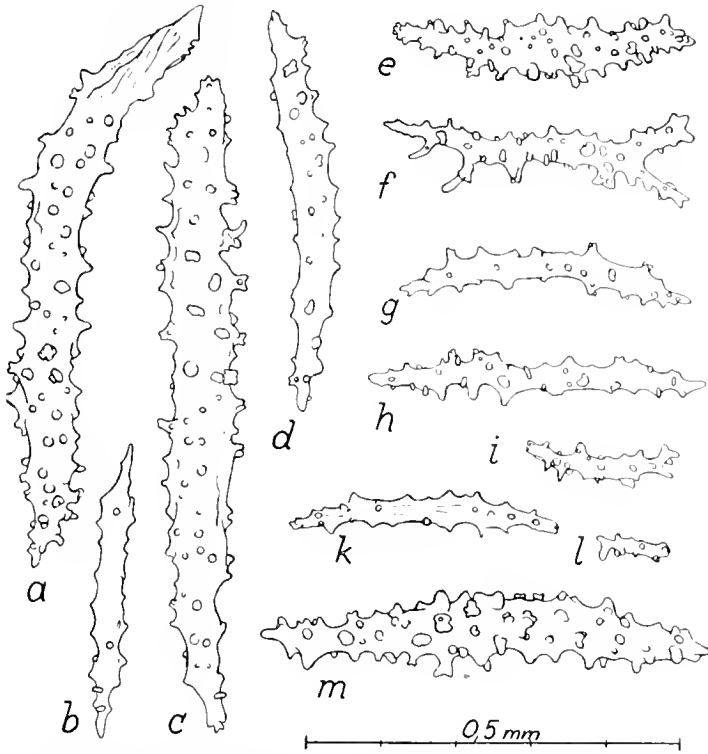


Fig. 3. *Clavularia greigii* n. sp. a-d, sclerites of anthostele, e-m, sclerites of stolon, g-k, of the usual slender type, e, m, of the larger and more seldom type, m, of the usual size, l, a branched sclerite from the stolon.

In none of the stolon-fragments the zooids are placed in a single row, and as moreover the stolons are somewhat extended here and there, it is most probable that the stolon in larger specimens is membranous. The verrucae measure from 0.5-2 mm in height.

When alive the specimens were stated to be straw-coloured and they are now in alcohol greyish white.

The sclerites in the stolon are warty spindles crosswise placed, though mostly in a longitudinal direction (fig. 3 e m). The great majority of them are slender, straight or somewhat curved, generally measuring 0.35-0.15 - about 0.03 mm (fig. 3 g l); scattered among these are, however, a few considerably stouter and also more strongly warty ones in average measuring 0.6 - 0.075 mm (fig. 3 m). A few slightly branched spindles may be found (fig. 3 f), and in a small fragment of a stolon which is supposed to be of the same species, though most unfortunately all the zooids have been worn off, most of the spindles are of this type, several of them even much more branched.

The zooids have entirely retractile anthocodia and are provided with a distinct, thick-walled anthostele in which the sclerites are mostly closely and irregularly placed in a longitudinal direction though often indistinctly arranged in eight converging double rows. At the base the sclerites are arranged more transversally.

The sclerites of the anthostele are warty, straight or curved spindles with both ends pointed (fig. 3 b d). The largest measured is 1.1 mm in length, but else their length only seldom exceeds about 0.8 mm. Numerous smaller sclerites occur among the larger ones, and in the younger zooids all the sclerites are small

and of about the same size. Besides the pointed spindles there may be, in each of the eight double rows, a few, usually curved, sclerites which have the upper end more irregularly formed, without warts, provided with ridges, often flattened and blunt and sometimes slightly protruding (fig. 3 a). Sometimes these sclerites may have the appearance of clubs.

The sclerites in the lower part of the anthocodia are slender and usually straight spindles, which are transversally and fairly scatteredly arranged and generally measure less than 0.5 mm in length (fig. 4 m o). The sclerites in the upper part of the anthocodia are arranged in chevron in eight double rows (fig. 4 p). They are curved, warty spindles which measure up to 0.2 mm. Those lowermost in the anthocodial crown have both ends pointed whereas the uppermost have a blunt and irregularly formed upper end (fig. 4 f, k) and resemble the corresponding type of the anthostele. The points of the anthocodial crown continue uninterrupted in the rows of sclerites in the tentacle dorsa. In the tentacle base the sclerites (fig. 4 h) resemble those of the anthocodial crown, but else the sclerites in the lower part of the tentacle dorsum are about 0.1 mm long, curved clubs which are arranged in a double row with their blunt ends pointing outwards (fig. 4 j, g). The upper part of the tentacle dorsum is provided with

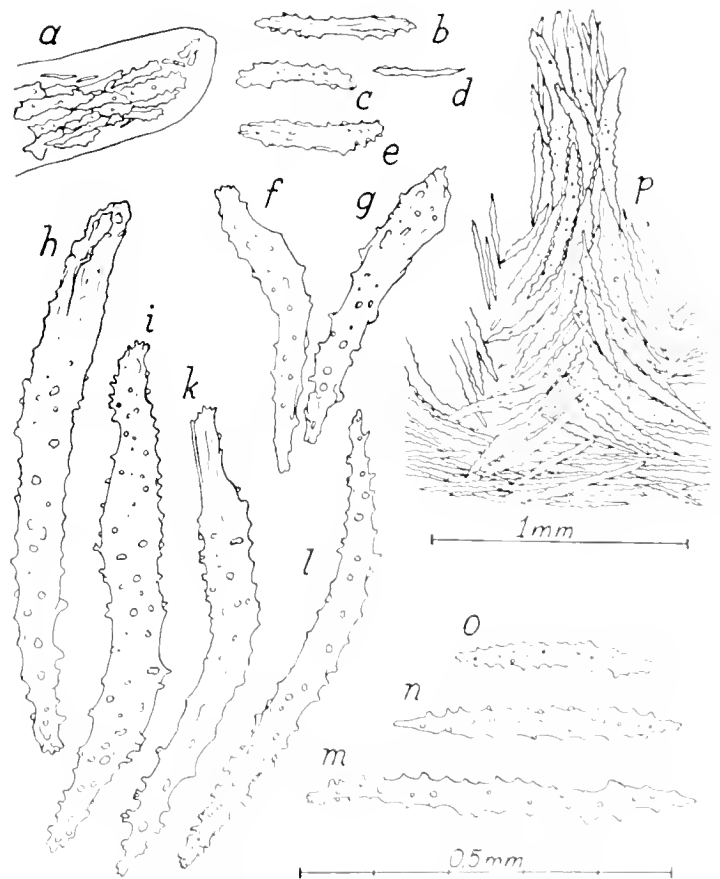


Fig. 4. *Clavularia greigii* n. sp. a, a pinnule with sclerites, b, sclerites of upper part of tentacle, c, d, sclerites of lower part of tentacle, e, sclerite from tentacle base, f, g, sclerites of anthocodial crown, h, o, sclerites of lower part of anthocodia, p, upper part of anthocodia showing the sclerites in natural position in one of the eight points of the anthocodial crown and the tentacle base.

irregularly transversally arranged, generally flattened rods measuring about 0.2 mm (fig. 4 b c), and also the pinnulae which number about 8-10 pairs are strongly armed with numerous small, flat sclerites (fig. 4 a).

The anthocodia retracts into the anthostele in a similar manner as that described below for *Clavularia levidensis*. The very rigid anthostele may, however, not be drawn together above the re-

of the zooids. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern.

The zooids are arranged in a regular pattern. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern.

The zooids are arranged in a regular pattern. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern.

The zooids are arranged in a regular pattern. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern.

The zooids are arranged in a regular pattern. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern.

Clavularia levidensis n. sp.

Author: [Name], 1922, p. 24, fig. 24. About 500

The zooids are arranged in a regular pattern. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern.

The zooids are arranged in a regular pattern. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern. The zooids are 1.5-2.0 mm high and 1.0-1.5 mm wide. The zooids are arranged in a regular pattern.

only 7 mm high. Most zooids are somewhat smaller than those of which the measures are given here.

The colour of the specimens in alcohol is greyish or light brown.

The sclerites of the species are almost needle-like and slightly spinous. They are in the lower part of the zooid body typically

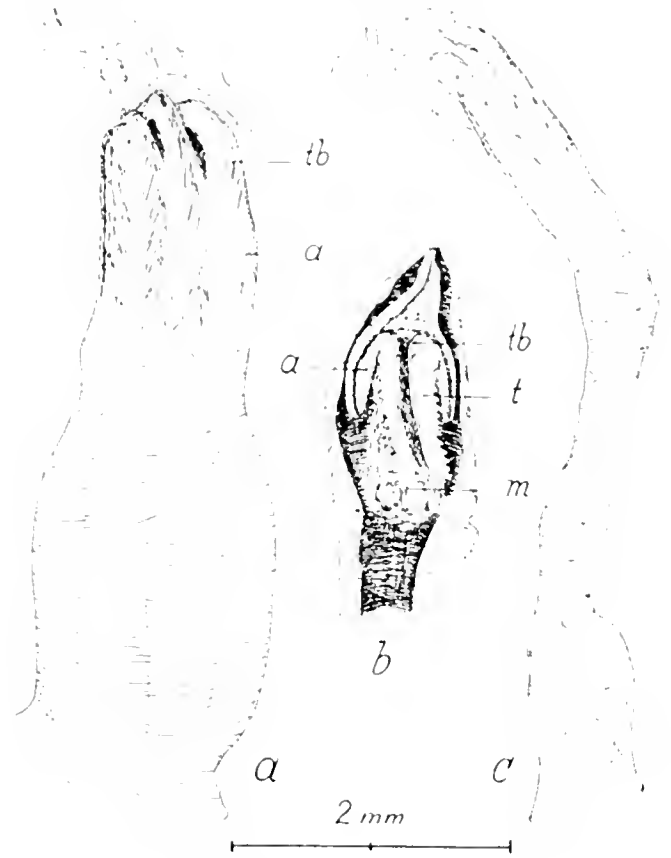


Fig. 5. *Clavularia levidensis* n. sp. a, c, sketches of two zooids, c, younger than a (compare the size of the anthocodial crowns but more extended. b, zooid with retracted anthocodia shown in longitudinal section. m, mesenteric filaments, only indicated, t, tentacles, tb, tentacle base, a, anthocodial crown.

horizontally placed (fig. 5 a) become, however, in the more extended zooids more irregularly arranged, often in fan-shaped groups (fig. 5 c). In most zooids the sclerites measure generally 0.2-0.3 mm (0.015 mm) (fig. 6 a). The sclerites in some specimens may, however, have a somewhat larger average size, the maximum size of them being about 0.6 mm (0.03 mm), and whereas the sclerites generally are almost straight, with blunt ends and about the same diameter in their whole length, those of the larger size are more pointed and often slightly curved (fig. 6 b).

In the upper part of the zooid body the sclerites are arranged in chevron in eight intermesenterial double rows. The number of sclerites in the rows increases considerably with the age of the specimens. These sclerites are slightly curved and measure in average about 0.3 (0.01-0.015 mm) (fig. 6 c), and also in those specimens with larger sclerites in the lower part of zooid body their maximum size does not seem to exceed about 0.4 mm.

The eight rows of sclerites continue uninterruptedly in those of the tentacle dorsa. At the tentacle base the sclerites are likewise arranged in chevron whereas in the lower part of the tentacles they are longitudinally arranged. They here reach a length of about 0.2 mm and are fairly strongly warty (fig. 6 d). In the outer half of tentacles the sclerites decrease in size and are scattered and more transversally arranged (fig. 6 e, f).

The pinnulae number about 12 pairs in each tentacle and are devoid of sclerites.

The specimens are probably all fairly young ones as generally there is no trace of a stolon. However, the zooids are mostly not juvenile as in some of them fairly large eggs were found. In a

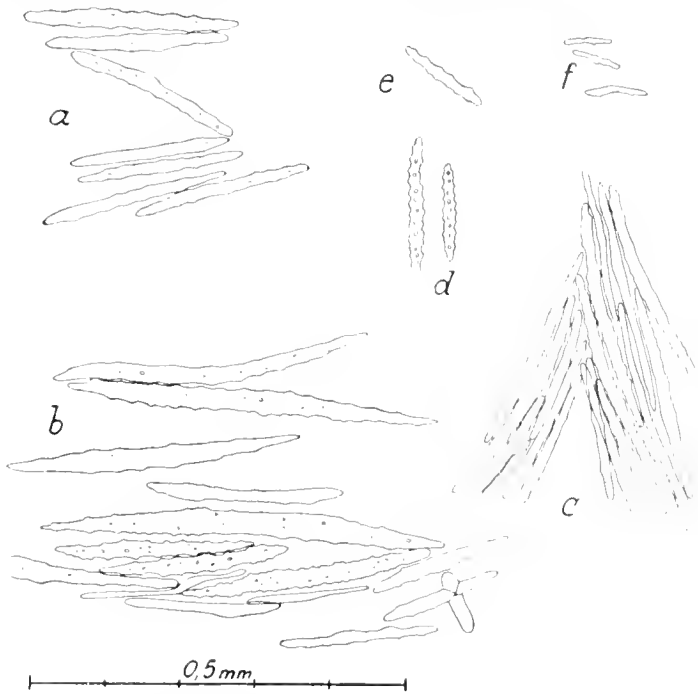


Fig. 6. *Clavularia luridensis* n. sp. a, b, sclerites from lower part of body wall (anthostele) in natural position. a, sclerites of the usual size. b, sclerites of the largest type. c, sclerites of anthocodial crown in natural position. d, longitudinally arranged sclerites from lower part of tentacle. e, sclerite from outer part of tentacle. f, sclerites from outermost part of tentacle in natural position.

few zooids the base is somewhat extended and flattened down upon the foraminiferans. In these rudiments of stolon the sclerites are of the same type as in the zooid body.

The zooids are thin-walled in their whole length, and thus it is not possible to distinguish clearly between an anthostele and an anthocodia; and if the upper part in one of the fifty specimens had not been retracted the species should probably erroneously have been described as non-retractile. The dissection of the only retracted specimen shows (fig. 5 b) that, by retraction, the upper part of the zooid with the crown of sclerites is simply withdrawn into the coelenteric cavity, whereas the upper part of that portion of the zooid body which is armed with transversally arranged sclerites (i. e. the lower part of the anthocodia) closes about and above it. The tentacles are not inverted when retracted, but simply put down into the coelenteric cavity.

Discussion: See under the following species.

Distribution: Davis Strait, 2258 m. Chart of distribution, fig. 1.

***Clavularia venustella* n. sp.**

Material:

63° 06' N, 56° 00' W, 2258 m, 24°C. "Ingolf" St. 21. About 100 zooids.

Diagnosis: Zooids small, less than 10 mm high, with retractile upper part, but thin-walled in their whole length. Stolon ribbon-like, branched at varying angles and sometimes anastomosing. Sclerites of stolon longitudinally arranged, warty spindles about 0.3-0.4 mm in length. Sclerites of lower part of zooid body transversally arranged, warty, often somewhat flattened rods, size generally 0.2-0.3 mm. Sclerites of upper part of zooid body warty spindles, maximum size about 0.1-0.03 mm, arranged in chevron in eight double rows which continue uninterruptedly in

the rows of sclerites in the tentacle dorsa. The lower part of tentacle dorsum provided with longitudinally and closely placed fairly short, warty spindles. The outer part provided with small and flattened, often transversally arranged sclerites.

Type: In the Zoological Museum of Copenhagen.

Description: The material consists of about a hundred zooids distributed on several colonies, many of these consisting of only one zooid and a small stolon.

The stolon is flat and creeps on sand incusted foraminiferans and small worm tubes. It has a width varying from 0.7-1.2 mm and may be somewhat branched at varying, sometimes right angles. Also anastomoses may occur. From the stolons the zooids arise at varying intervals, generally 1.5 mm, but the interval between them may be only about 0.5 mm. The largest observed interval between two zooids was about 7 mm, none of the preserved stolons or fragments of stolon are, however, now more than about 14 mm long.

The largest zooid measures from stolon to base of tentacles about 8 mm and has a diameter of about 1.5 mm. Most of the zooids are, however, considerably smaller, their average size being about 4-4 mm. The largest zooids have often a diameter of about 2 mm and are generally placed at the bifurcations of the stolon. The shape of the zooids are somewhat varying, many

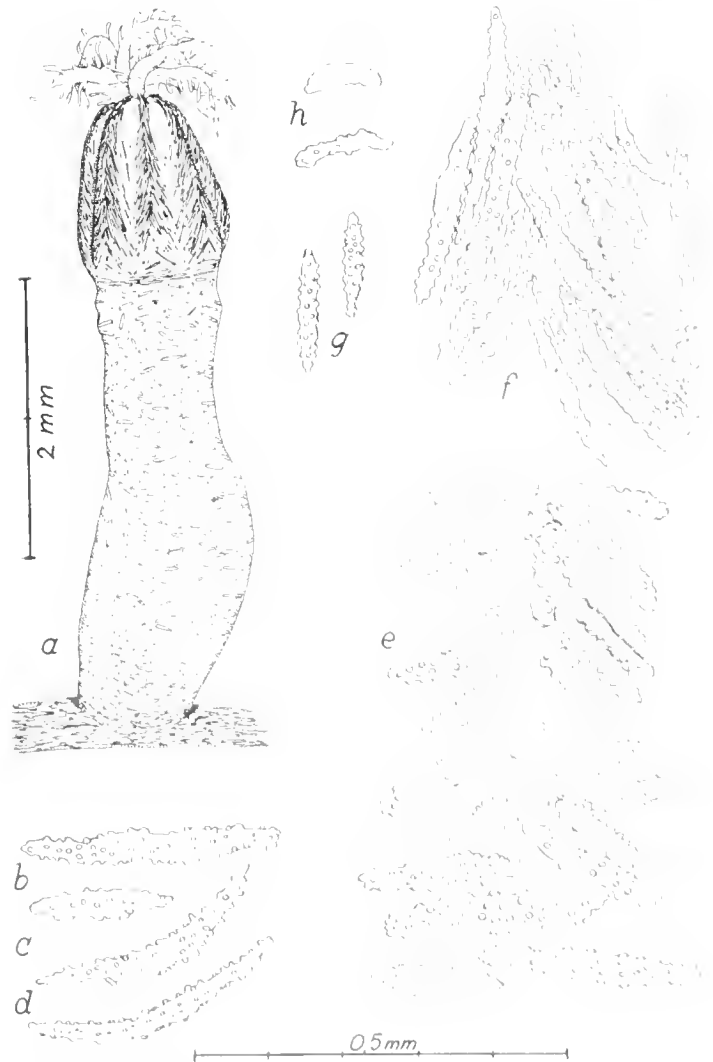


Fig. 7. *Clavularia venustella* n. sp. a, sketch of a zooid with the tentacle halfway withdrawn. b, d, sclerites of stolon. c, sclerites of body wall in natural position, the dotted lines indicate the attachments of the mesenteries. f, sclerites from uppermost part of zooid body (anthocodial crown) in natural position. e, longitudinally arranged sclerites from lower part of tentacle. h, sclerites from outer part of tentacle. g, sclerites from lower part of tentacle.

The sclerites in the basal part of the tentacles are longitudinally arranged straight spindles (fig. 8*c*). The sclerites in the outer part of the tentacles are flattened and transversally arranged. Due to the scarcity of material the spiculation of the pinnulae was not studied.

Discussion: The specimen described above has been referred to *Anthelia borealis* as in all essentials it is well in accordance with Broch's description of this species. In any case it is impossible to point out any distinguishing character of greater value. It may be admitted that the sclerites of the zooid body, judging from Broch's figure 2, may be stouter in the specimens from the Trondheim Fjord than they are in that taken by the "Ingolf" Expedition, but this may be due to individual variation. The size of the specimen described here is considerably shorter than that stated for the largest zooids of the specimens from the Trondheim Fjord, but it must be remembered that the described specimen is juvenile.

The specimen taken by the "Ingolf" Expedition resembles also *Anthelia armata* J. A. Thomson 1927 (p. 9) from off Madeira in 2380 m depth, which species apparently has a similar arrangement of the sclerites in the zooid body, although J. A. Thomson writes about the sclerites: "Ceux de la partie inférieure du polype sont plutôt disposés horizontalement que longitudinalement, mais avec une tendance à former 8 crêtes longitudinales." In J. A. Thomson's species the largest stated size of sclerites is about 0.5 mm.

Anthelia borealis from the Trondheim Fjord, the specimen from the "Ingolf" St. 21, and J. A. Thomson's *A. armata* are apparently very closely related and may prove to belong to the same species. J. A. Thomson's description is, however, not very thorough and no figures are given of the sclerites.

Distribution: *Anthelia borealis* is previously known from Trondheim Fjord in depths of 200-100 m. If the identification of the specimen described above is correct, and there is no evidence that it is not, the species is also found in the Danmark Strait west of Iceland at 817 m, which also favours the supposition that it has been found at Madeira in 2380 m depth under the name *A. armata*. Chart of distribution, fig. 1.

Anthelia fallax Broch 1912.

Anthelia fallax Broch, 1912*d*, p. 13-16, figs. 5-8.

Anthothela grandiflora var. *fallax* Molander, 1918*b*, p. 10-11, fig. 3.

Anthothela grandiflora var. *fallax* Molander, 1929, p. 36.

Diagnosis: Vide Broch 1912*d*.

Family: Cyathopodidae n. fam.

Genus Cyathopodium Verrill 1868.

This genus was proposed by VERRILL (1868*b*, p. 115) for a small stoloniferous coral from the Pamotu Archipelago which DANA in 1846 described and referred to the extinct genus *Aulopora*.

The author has not had access to DANA's original large paper, but has seen the original atlas and the synopsis of the species from the original paper which DANA published in 1859. DANA writes (p. 126) about the family IV—Tubiporidae (of the suborder II Aleyonaria) the following: "Coralligenous Aleyonaria: coralla tubular, calcareous or semicalcareous; tubes not striate within. Genus I. *Aulopora*. Non-aerogenous Tubiporidae, budding from a creeping filiform stolon. Coralla calcareous, tubular, encrusting, often reticulately anastomosing; cells subremote, sometimes a little prominent. *Aulopora tenuis* (Dana). Red, tubular corallum filiform, reticulate, mostly $\frac{1}{3}$ of a line broad, cells circular not prominent."

Material:

Fragment of the type specimen from Trondheim Fjord

Remarks. It seems worth noting that the spiculation of the zooid body in this species much resembles that of an *Acanthogorgia*, especially in the arrangement of the peculiarly bent sclerites (cf. fig. 23*b*); however, also in the spiculation below and at the tentacle base, where the sclerites are a little protruding although in *A. fallax* they do not have a distinctly protruding, smooth ray. No doubt MOLANDER is wrong in referring *A. fallax* to *Anthothela grandiflora* (see further Broch 1935 p. 5), and if the species should be removed from the Stoloniifera and placed among the Gorgonaceans it may undoubtedly be in the vicinity of the family Acanthogorgiidae.

Distribution: At present only known from Trondheim Fjord in depths of 300-500 m.

Genus Sarcodictyon Forbes 1847.

Sarcodictyon catenatum Forbes 1847

Sarcodictyon catenata Forbes, in Johnston 1847, p. 179, pl. 33 figs. 4-7.

Rhizoxema filiformis M. Sars, 1856, p. 65-67, pl. 19, figs. 13-17.

Sarcodictyon catenata Herdman, 1883, p. 31-51, pl. I-3.

Rhizoxenia filiformis Appelf., 1895, p. 11.

Sarcodictyon catenata Stephens, 1909, p. 1.

Evagora catenata Anrivillius, 1926, p. 114.

Sarcodictyon catenatum J. A. Thomson, 1927, p. 8-9.
Carlgren, MS.

Description: Vide HERDMAN 1883

Material:

Hjeltefjord (Norwegian coast near Bergen), 55 m, 1 colony.

Distribution: This insignificant little species is not actually known from the area investigated by the "Ingolf" Expedition, but it is very likely that it sooner or later will prove to occur here.

The species has been recorded from the Irish and Scottish Atlantic coasts in depths down to about 900 m, and under the name *Rhizoxenia filiformis* from Scandinavia from Mangerfjord and Herlofjord near Bergen, in the vicinity of which the above mentioned specimen was taken, it has further been mentioned from the Skagerrak from Gullmarfjord by ANRIVILLIUS (1926) and from Kosterfjord and Seghsfjord by CARLGREN (MS). The species is also recorded from the English Channel and the Mediterranean. Probably it has a very wide distribution.

Bathymetrical distribution: About (20) 50-850 m.

VERRILL (1868*b*, p. 115) refers the species to a new genus *Cyathopodium* with the following words: "This new genus is proposed for the remarkable little polyp, rightly described by Prof. DANA as closely allied to *Tubipora*. It is in fact a *Tubipora*-like polyp, with short cup-shaped tubes, connected by narrow calcareous stolons which correspond to the transverse plates of *Tubipora* and from which new buds arise."

Neither DANA's nor VERRILL's description are thus very distinct and the genus was therefore in many years by students of the Stoloniifera group considered a very doubtful one. DEICHMANN (1936 p. 38) describes, however, a new species from the West Indies and refers to the authority of VERRILL to the genus *Cyathopodium*. DEICHMANN further tells that VERRILL in his not published notes had placed the genus *Cyathopodium* in the family Telestidae and thus may, as it will follow below, be an important information.

DEICHMANN refers the genus *Cyathopodium* to the Tubiporidae.

by slight magnification, rather indistinct. Microscopic examination shows, however, that in the spiculation there is a very strong distinction between the lower part of the anthocodia and the upper part of the anthostele. In the latter the sclerites are considerably larger and have a somewhat different appearance.

The sclerites in the upper part of the anthostele are closely placed in a few layers. They are either fairly smooth and generally richly branched simple rods, measuring 0.2-0.3 mm in length, or quadruplet formations of the same. Uppermost in the anthostele most of the sclerites are free (fig. 11 *a, d*), several of them, however, fuse together (fig. 11 *c*), and the main anthostele is in this manner developed into a very solid, calcareous tube, the walls of which are formed by a close calcareous meshwork (fig. 12) built of fused simple rods and quadruplets arranged in a layer about two to three sclerites thick.

The outer walls of the stolons are built in the same manner. They are, however, usually somewhat stouter than the figured section of an anthostele, formed by somewhat thicker sclerites. The inner part of the stolons is for the most part filled with free sclerites, which are considerably smaller than those which build the outer wall, and among which sixtuplets with a diameter of

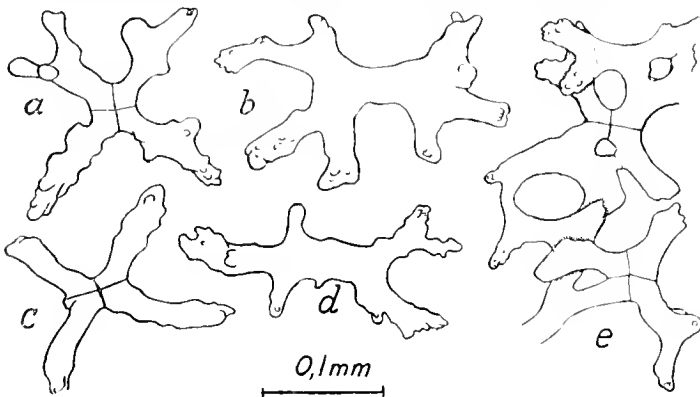


Fig. 11. *Cyathopodium vulguli* n. sp. Solitary simple sclerites (*b, d*) and quadruplets (*a, c*) from the upper part of the anthostele, compare fig. 13 *b*, zone *as₁*, *e*, some of the fused sclerites from the same part of the zooid.

about 0.1 mm are rather frequent. The outer calcareous meshwork in the stolon is separated from the inner skeleton by a delicate, foliated and perforated horny or chitinous membrane.

Due to the scarcity of the material it has not been possible to ascertain whether in the body wall of the zooid an inner horny meshwork is present, the occurrence of which DEICHMANN (1936) uses as a diagnostic character for the genus *Cyathopodium*; as, however, such an inner horny layer exists in the stolons it is almost certain that a similar layer may be found in the zooids.

The zooids are provided with a, sometimes fairly thick, horny or chitinous cuticula.

The fragmentary basal part of a zooid, which is figured in fig. 11 *f*, shows a very interesting feature, viz. that a calcareous skeleton divides the coelenteric cavity into eight main longitudinal canals separated by tiny calcareous walls, forming in the centre of the zooid a more irregular meshwork with smaller canals, the structure of this inner calcareous skeleton is the same as in the outer walls of the anthostele. The height of this fragmentary zooid is about 1.5 mm, and the diameter is about 1 mm, i. e. of about the same size as in the small zooid figured in fig. 11 *a, c* and considerably smaller than in the above described zooid (fig. 13) in which no trace of such an inner skeletal structure was seen.

An intrusion in the basal part of the coelenteric cavity of a very loose calcareous meshwork was, however, observed in a very damaged zooid about 3 mm long in contracted state and scarcely 1 mm in diameter, but due to the bad state of preservation it

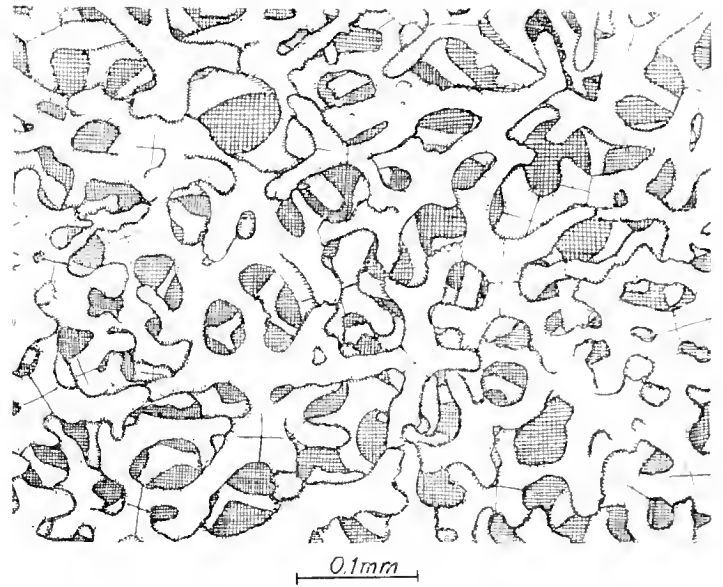


Fig. 12. *Cyathopodium vulguli* n. sp. Section of the calcareous skeleton in the main part of the anthostele, compare fig. 13 *b*, zone *as₂*.

was not possible to state anything definite about this calcareous meshwork, e. g. whether it was situated in the mesenteries, which probably has been the case, which a comparison with the Telestacean species *Pseudochladorhynchus jacksoni* Versluys 1907 might indicate, or not. See further the discussion on the following species. It is the comparison with VERSLUYS' description of *Pseudochladorhynchus* which justifies the description of the fragment in fig. 11 *f* as an anthostele. As it is seen from fig. 11 *c* a similar anthostele may be found also in smaller zooids.

The basal part of the specimen in fig. 13 (on the figure marked *s₁*) has, on the contrary, been described as a stolon, and it is

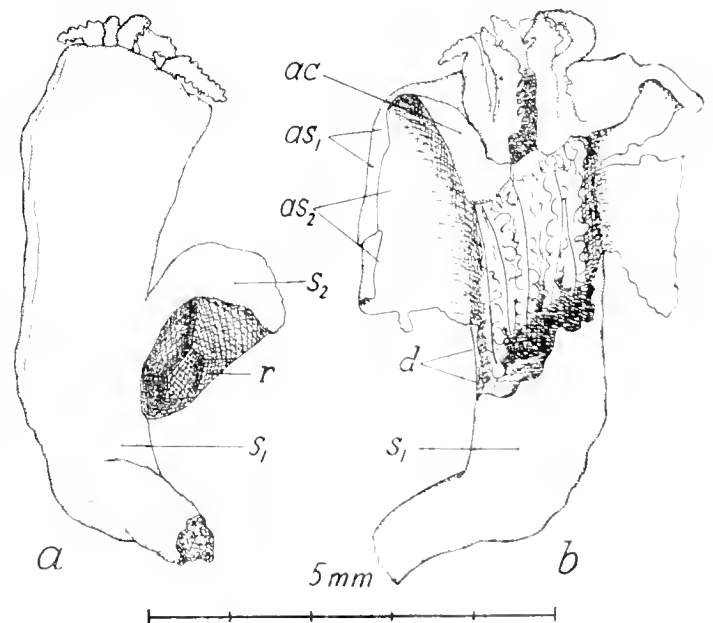


Fig. 13. *Cyathopodium vulguli* n. sp. *a*, small fragmentary, partly free stolon with a single zooid; *b*, the same specimen from the opposite side; the small piece of rock *r* with the stolon attached to it *s₂* has been removed and the zooid opened. The mesenteries with the mesenteric filaments are only indicated, the dorsal ones are shown at the left. There is no development of a skeleton in the mesenteries and no distinct canals issue from the bottom of the coelenteric cavity. *c*, a small piece of rock, *s₁* free stolon, *s₂* creeping stolon, *d*, the scar where the creeping stolon has been broken off, *ac*, the anthocodia with free sclerites fig. 10, *as₁*, the upper part of the anthostele with free sclerites and rigid calcareous skeleton fig. 11, *as₂*, the main part of the anthostele with the calcareous skeleton fig. 12.

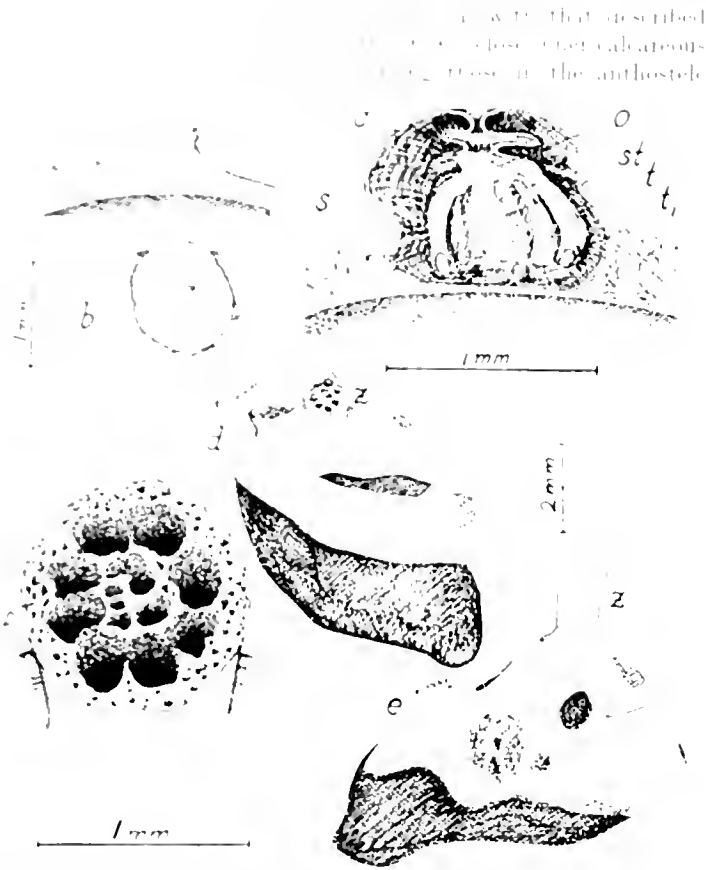


FIG. 1. *Cyathopodium* sp. n. (a) small contracted zooid shown from above and from above; (b) the same zooid shown in section, showing the inverted tentacle t_i , the not inverted tip of tentacle t , and the part of an inverted tentacle; (c) the stomodaeum, showing the part of an inverted tentacle; (d) a fragmentary colony on a stone drawn to show the fragmentary zooid from above and from the side respectively; (e) more extended zooid of the zooid seen from above, showing the thick coarsened work in the outer body wall and the eight distinct canals cut by the calcareous septae. This zooid was covered with a cracked horny or chitinous colourless cuticula.

is a stolon, and when it has given off the zooid it continues as the part of the figure marked s_1 which creeps on a small stone. Of course part s_1 might have been described as an anthostele, but s_2 is a distal stolon given off from the zooid, but as it is not seen from the above mentioned this would have been erroneous. From the present material it is also in some cases seen how

a flat stolon, which creeps upon a small stone, leaves this, then rounds off and for a time grows as a free stolon, round in section, before it again becomes attached to a small stone and again flat in section. The whole material of this species is also supposed to have belonged to one single colony, and the connections between the fragments have without doubt partly been constituted by such free stolons.

Discussion. It is for the present impossible to state anything definite about the systematic position of the family Cyathopodidae, as the structure of the zooids is not sufficiently cleared up. Some remarks may, however, be made. DANA in 1846 placed the genus in the Tubiporidae, and the same is done also by VERRILL (1868 *b*) and DEICHMANN (1936). In fact, however, there seems to be no closer relation between the Cyathopodidae and the Tubiporidae. In the period between VERRILL's paper of 1868 *b* and DEICHMANN's of 1936 *Cyathopodium* has, on account of DANA's original description, been regarded as a very doubtful species in the Clavularidae. In the present paper it is also provisionally placed in the order Stolonifera, partly as it is most convenient to refer a stoloniferous species to this group, and partly because the order Stolonifera already before hand is a sort of limber-room. It is, however, most probable that the real systematic place of the Cyathopodidae is in the order Telestacea, which point of view is supported by the above described anthostele. It may also be supposed that similar conditions in the *Cyathopodium elegans* Deichmann induced VERRILL, in his not published manuscript to place the genus among the Telestidae. As, however, secondary zooids have not yet been observed in any of the species referred to the genus *Cyathopodium* it has not been found convenient for the present to refer the Cyathopodidae to the order Telestacea, in which precisely the principal character is the occurrence of secondary zooids.

Cyathopodium ingolfi is in the colourless sclerites distinguished from *C. tenuis* which probably has light red sclerites, and *C. elegans* which has lemon coloured or yellow sclerites. The structure of the zooid in *C. tenuis* is not known. The zooid of *C. elegans* is in contrast to *C. ingolfi* provided with an operculum, and DEICHMANN also states the occurrence of a collarlet below the tentacles. In *C. ingolfi* nothing corresponding is found. The shape of the sclerites and the abundant occurrence of quadruplets in *C. ingolfi* yield also useful distinguishing characters, and it may be doubtful whether *C. elegans* and *C. ingolfi* in reality should be placed in the same genus.

Distribution. South of Iceland at about 1300 m. Chart of distribution, fig. 15.

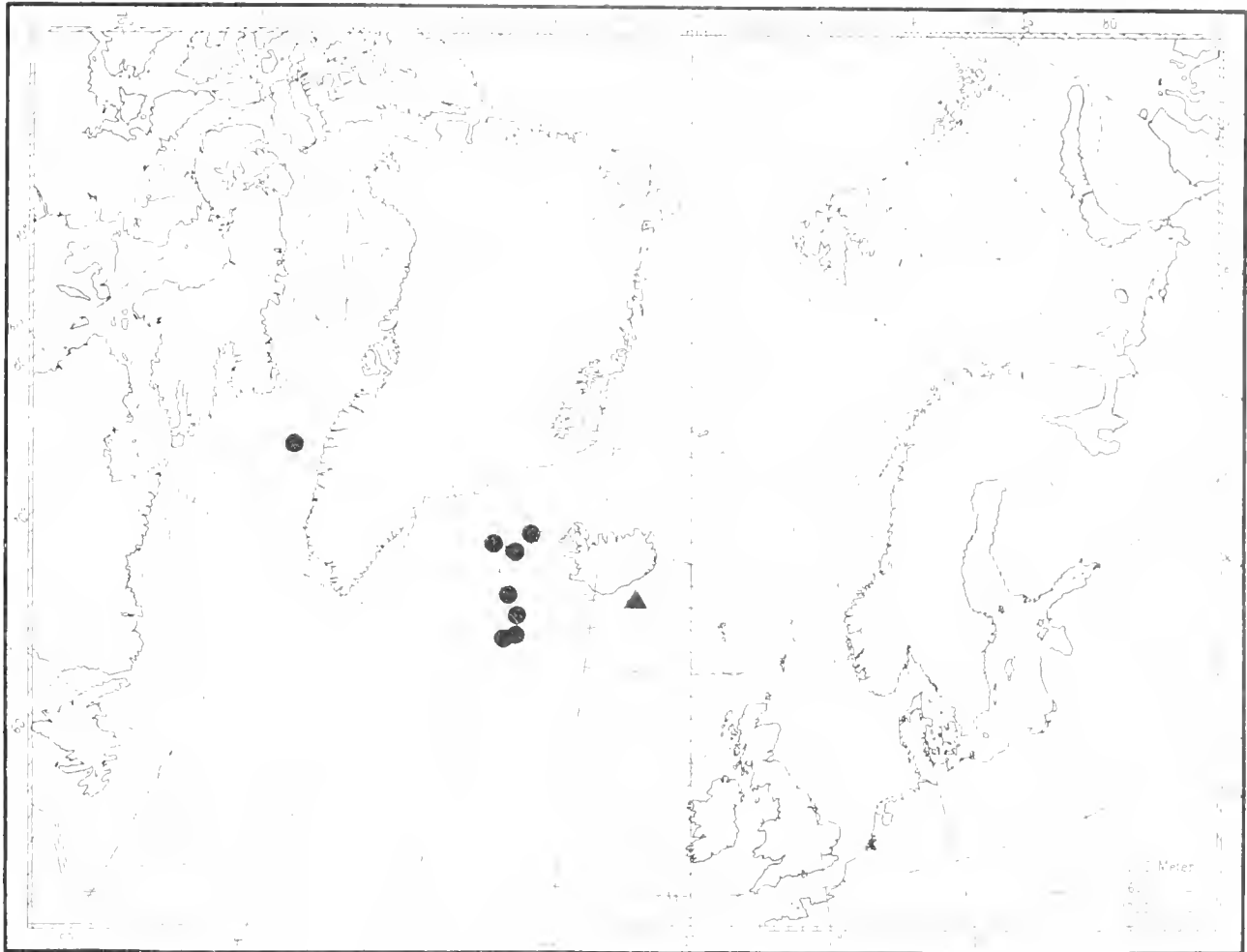


Fig. 15. ▲ *Cyathopodium angolli* n. sp.
● *Telestaba septentrionalis* n. sp.

Ordo: Telestacea Hickson 1930.

Diagnosis. Stoloniiferous, branching Octocorals, where in the monopodial species stem and branches and in the sympodial species stem- and branch-sections each is formed by one single zooid. The secondary zooids only at their base connected with the primary ones, their coelenteric cavities connected with that

of the primary zooid through canals in the body wall of this latter. Sclerites numerous and sometimes fused into a calcareous meshwork. In some genera the lower part of the coelenteric cavity partially filled out with a mesogloea tissue containing numerous often fused sclerites.

Family: Telestidae Milne Edwards 1857

Diagnosis. The same as for the order.

Remarks: In the order Telestacea only one family, viz. Telestidae, is recognized at present. This family comprises, with the diagnosis above, both the monopodial branching genera of the usual *Telesta* type, viz. *Telesta* Lamouroux, *Coclogorgia* Milne Edwards, and also *Pseudogorgia* Kolliker, and the sympodial branching and differing *Pseudocladohoris* Versluys and the new genus *Telestaba* described below, which occupies an intermediate position between the usual *Telesta* type and *Pseudocladohoris*.

Some authors have also in the Telestidae included *Scleranthella* Studer, which genus STREPER, however, placed among the Clavularidae, which point of view seems to be the most con-

venient, though MOLANDER (1929 p. 31) by a re-examination of the genus arrives at the conclusion that *Scleranthella* belongs to the genus *Parisis*.

Also another genus has been reckoned to the *Telestidae*, viz. *Chalazaria* sensu Molander 1929. In MOLANDER'S much amended sense *Chalazaria* (diagnosis: MOLANDER 1929 p. 19) comprises only the following species: *C. crades* Quoy & Gaimard, *C. violacea* Quoy & Gaimard, *C. capitata* Schenk and *C. coronata* Burchard. MOLANDER'S conclusion that these species should be included in the *Telestaba* is based partly on observations of young zooid buds directly upon elder zooids in some of the species and partly on some observations on the anatomy of *C. crades*. HICKSON (1930) however, dissents emphatically from this conclusion and

surface, often also with scattered larger warts and usually with a rather irregular outline. The largest may measure about $0.35 \cdot 0.15$ mm, most of them are, however, about $0.25 \cdot 0.07$ mm. These sclerites are longitudinally placed and form a very dense

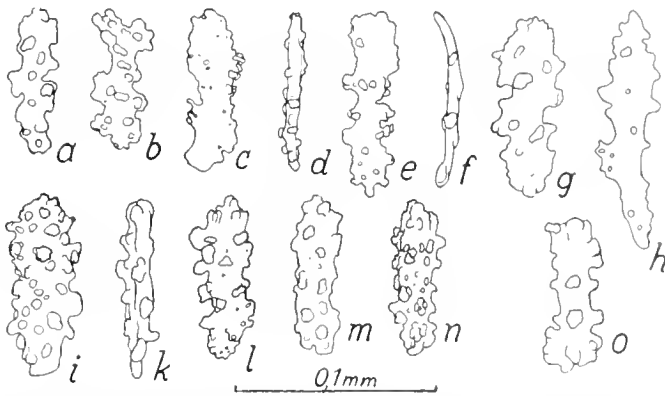


Fig. 17. *Telesstaba septentrionalis* n. g., n. sp. a-h, sclerites of tentacles, a-g, of the usual type, h, an extraordinarily large one, d, f, shown from the side, i-n, sclerites of anthocodia, k, shown from the side, n, a not flattened one from the lower part of anthocodia, o, sclerite from the stomodaecium.

covering of the stolon. Below this sclerite mail, in the inner and lowest part of the stolon, there is a loose skeleton formed by the other, somewhat different, type of sclerites (fig. 18 k-m), which are rod- or spindle-like with large warts and often branched. They are smaller than those of the other type, generally less than 0.15 mm, and have a less granulated surface. Sometimes some are fused together.

The sclerites in the body wall of the anthostele resemble those of the outer part of the stolon, but are more warty and generally more regularly shaped than these. Two sclerites of the typical shape are shown in fig. 18 o, p from above and from the side respectively. Their average size is generally about $0.35 \cdot 0.13$ mm, but several smaller ones also occur. Besides some larger sclerites may be found, in the smaller zooids and the upper part of the more lengthened ones measuring about $0.15 \cdot 0.25$ mm, in the lower part of the longer zooids, however, some may measure up to $0.7 \cdot 0.33$ mm. The sclerites are longitudinally and very closely arranged, placed in the furrows of the zooid in a layer, mainly only one sclerite thick, in the ridges of the zooid, however, somewhat more crowded (compare the transverse section pl. I fig. 3). The largest and most irregularly formed sclerites are found in the ridges.

Twins and quadruplets may be rather common both in the anthostele and the stolon.

The sclerites in the anthocodial part of the body wall are much smaller, warty, rod-like, and flattened ones, all of about equal size, i.e. about $0.08 \cdot 0.025$ mm. It seems, however, as if those of the uppermost part of the anthocodia are generally more flattened than those from the lower part. They are mostly irregularly obliquely or transversely placed, probably all becoming longitudinally arranged in the entirely expanded anthocodia, and mainly restricted to eight intermesenterial ridges which form a continuation of the ridges of the anthostele. As only very few sclerites occur outside these rather narrow rows, an almost complete contraction of the lower part of the by retraction inverted anthocodia is made possible (compare pl. I fig. 1).

The transition from the anthostele to the anthocodia is fairly smooth. In the anthocodia the rows of sclerites are, however, interrupted a little below the tentacles.

The tentacles, which in full-grown zooids measure $1.6 \cdot 1.8$ mm in contracted state, are strongly armed on all sides with a dense layer of transversely arranged sclerites (fig. 17 a-b) which resemble those of the anthocodia though being somewhat more

flattened (fig. 17 d, f), generally less warty and also a little more varying in shape and size. Average size about 0.08 mm, maximum size about 0.12 mm. Also the pinnulae, which number about 10-12 pairs and in the contracted state are very short, are well provided with small sclerites (compare pl. I fig. 3).

The stomodaecium, which in comparison with the mesenteries is fairly long, is provided with rather numerous and transversally arranged sclerites of the same type as those from the tentacles (fig. 17 o).

Most zooids in the present material measure less than $7 \cdot 8$ mm in height in contracted state and in these the dorsal mesenteries extend down to the stolon. In the larger zooids, however, the basal part of the coelenteric cavity is filled with a loose skeleton of warty and often branched sclerites (fig. 18 s-q) resembling those from the inner part of the stolon but of a larger size, measuring up to about 0.3 mm in length. By the intrusion of these sclerites into the coelenteric cavity the lower part of the longer zooids is developed into a sort of stem which may constitute the greater part of the zooid.

The following measures may be given. In a zooid, which measures 8 mm with tentacles extended, no sclerites in the coelenteric cavity were noticed. In a 10.5 mm long contracted zooid the "stem" constitutes 4 mm; and the "stem" is 18 mm long in the zooid, fig. 16 b, which in contracted state measures about 24 mm. In this last-mentioned zooid the diameter at the stolon is about 4 mm, whereas at the tip of the anthostele it is about 1.4 mm. The anthocodia without tentacles measures about 2.5 mm

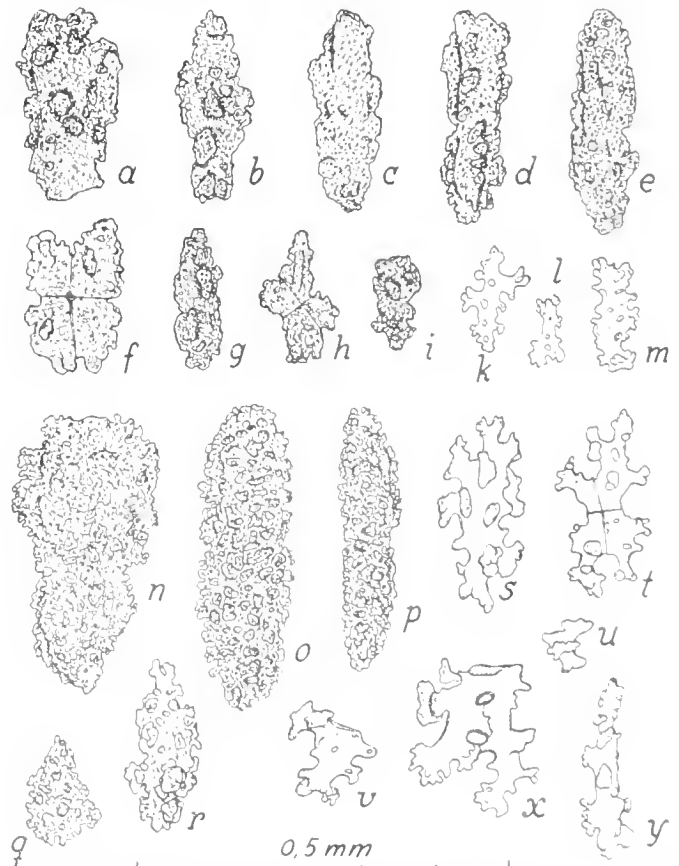


Fig. 18. *Telesstaba septentrionalis* n. g., n. sp. a-e, sclerites of stolon, a, c, from the outer wall of the stolon, b, d, from the inner part of the stolon, a, a, sclerites of zooid, a, sclerites of body wall, p, shown from the side, s, q, sclerites from the inner part of "stem", q, r, quadruplets of fused sclerites.

and is not completely retracted. The tentacles measure about 1.8 mm. Several single zooids are found in contracted state measuring more than 20 mm in height.

The development of the "stem", in the lower part of which

the "stem" is a spiracled ridge (about 1/200th of the zoid's length). It is clear from the above that in the "stem" eight longitudinal canals may be distinguished which represent the "stem" canals. Generally, these canals are arranged in a spiral, but often a few of them may be arranged in a straight part of the "stem" and thus only eight "stem" canals may be distinguished. The "stem" canals also give off branches, and in the upper part of the "stem" small endodermic canals are seen between the sclerites in the middle. The sclerites in the inner part of the "stem" are arranged in spiral and sections through this part of the "stem" are, what is without canals and has been observed in a zigzag and fused sclerites, and in a circle around this part of the "stem" canals and other endodermic canals. In this part of the "stem" canals are fairly small, whereas in the other endodermic canals are comparatively large and they cannot be distinguished from the "stem" canals. In the "stem" several mesogloal cell canals besides the "stem" canals are found, and such are also present in the lower part of the zooids, compare pl. 4 fig. 2. In six zooids, i.e. about 10% of the maternal, secondary zooids arise from the "stem" of the primary ones (fig. 16 b). A zooid, averaged, 35 mm long zoid with a diameter of 2.12 mm, in its whole length may probably have had two lateral zooids, but otherwise only one secondary zooid is found in the colonies.

The secondary zooids resemble the primary ones, and when they are 270 µm long about 6 mm they have their lower part developed into a "stem" similar to that of the primary zooids.

Discussion.—The above described species should be placed in the genus *Telastata* on account of the occurrence of secondary zooids, however scarce they are. If, however, the lateral zooids are not included these zooids with secondary ones, the genus *Telastata* probably have been given a doubtful position in the genus *Stichometra*. It is also probable that some of the hitherto described species of *Carabalia* should be referred to *Telastata*, the type species are identical with *Telastata septentrionalis*. It may be mentioned here that there is no closer relationship between *Telastata* and *Carabalia* sensu Molander, the latter having distal to the lateral zooids and a quite differing spiculation.

On the other points *Telastata* resembles the genus *Telista* very much, the distinct longitudinal furrows and the horny or chitinous plates are common to both genera and also the sclerites in the "stem" are of about the same type as in *Telista*.

Telastata septentrionalis differs from *Telista* in that lateral zooids are not generally occurring. As mentioned above the secondary zooids are only found in about 10% of the zooids in the colonies. In most of the eight stations from which the material was taken the species branching zooids are found only in station 81, 83 and 197. Another species, which without doubt should be referred to the genus *Telastata*, viz. *Pseudotelastata* Thomson & Dean (1931, p. 216, pl. 15 figs. 5, 9, pl. 27 fig. 7 (9 00'3" S, 126 24'5" E, 112 m)) shows, however, externally the same habitus as *Telastata*. See further below.

The characters which distinguishes *Telastata* from *Telista* is the occurrence of the "stem" and the creation of the new genus is the result of the discovery of a mesogloal, sclerite in the lower part of the coelenteric cavity of the older zooids. In the younger zooids of the much larger *Telista* the sclerites are fused to the stolon (LAVACKMANN 1916, p. 107, fig. 12). LAVACKMANN, it is true, in his description of the species of *Telastata* found an internal sclerite in the lower part of the coelenteric cavity, divided into two parts (LAVACKMANN 1916, p. 96, pl. 7 fig. 10). The sclerites in the lower part of other colonies of *Telastata* are not fused to the stolon, but exceptionally may be

found there is still a primary difference, as the "stem" in *Telastata* is a normal formation which starts its development as soon as the anthostele of the zooid reaches a length of about 6 mm, and also are found in the secondary zooids when they exceed the same size.

The "stem" might also have been described as a vertical stolon, but such a point of view is contradicted by comparison with the long primary zooid in *Telista* and also by comparison with the development of the "stem" in the Pacific genus *Pseudotelastata*, excellently described by VERSLUYS in 1907. In *Pseudotelastata* the sclerites in the zooid, apart from the anthostele, are fused together in such a manner that a solid, perforated, calcareous tube is formed. The calcareous skeleton of the zooid extends also in the basalmost part of the mesenteries, and VERSLUYS' preparations showed that the mesenteries in the lower part of the coelenteric cavity meet and fuse in the centre, in which way eight "stem" canals and the mesogloal filling in the "stem" are formed (VERSLUYS 1907, pls. 2, 3). The "stem" canals are, however, in *Pseudotelastata* considerably larger in comparison with the mesogloal tissue than in *Telastata*, and the conditions as they are figured for *Pseudotelastata* by VERSLUYS (pl. 3 fig. 13) much resemble those found in the anthostele of *Cyathopodium equifolii* (compare fig. 11 f).

The branching of the zooids in *Pseudotelastata* differs principally from that in *Telastata*, as the lateral zooids issue from the body wall of the others, above the "stem" (VERSLUYS 1907, pl. 13 fig. 11), whereas in *Telastata* they issue from the "stem" (fig. 16 b). In *Pseudotelastata* the branching is sympodial, all the zooids being equal, whereas in *Telastata* it is monopodial with a long primary zooid and shorter secondary ones. In *Pseudotelastata* the lower part of all the zooids is developed into a "stem".

VERSLUYS' figures 2, 3 and pl. 3 fig. 11 show that the tentacles in *Pseudotelastata*, by retraction of the anthostele, are bent down, whereas in the specimens examined of *Telastata* they point upwards also in retracted state (compare fig. 16 b), probably, however, this is of no systematic value.

It may be remarked that J. A. THOMSON & DEAN (1931) have referred two new species to the genus *Pseudotelastata*, which, however, apparently differ too much from the type described by VERSLUYS to be placed in the same genus as this. Due to the scarcity of their material THOMSON & DEAN have not been able to give a fully exhaustive description of the structure of the colonies. There seems, however, to be no doubt that one of the species, viz. *P. mosaiica* Thomson & Dean (1931, p. 216, pl. 15 figs. 5, 9, pl. 27 fig. 7 (9 00'3" S, 126 24'5" E, 112 m)) belongs to the genus *Telastata* and is very closely related to *T. septentrionalis*.

Telastata mosaiica (Thomson & Dean) agrees with *T. septentrionalis* as regards the spiculation of the anthostele, and the sclerites in the two species are much alike, though it seems as if those of the inner part of the "stem" in *T. mosaiica* are longer and more slender than the corresponding from *T. septentrionalis*. At present the occurrence of numerous tertiary zooids in *T. mosaiica* probably yields the best distinguishing character between the two species.

The other species *P. carlsbergi* Thomson & Dean (1931, p. 217, pl. 2 fig. 7, pl. 25 figs. 9 a, 9 b) is after the description difficult to place in the system. In outer habitus the species differs considerably from *P. hicksoni* Versluys, it is much more robustly built, it does not show the zigzag arrangement of the zooids, which all are arranged close together about the end of the primary one, and the sclerites are not fused. All this favours the supposition that *P. carlsbergi* Thomson & Dean is far more closely related to the genus *Telastata* than to *Pseudotelastata*, and if it should not be placed in a genus of its own, which seems probable, it may most conveniently be ranged in the genus *Telastata*. It would be very interesting to know whether the secondary zooids bud from the "stem" of the primary one as in *T. septentrionalis* or from the body wall above the "stem" as in *P. hicksoni*.

The genus *Telestata* described here bears resemblance both to the usual *Telesta* type and to *Pseudocladochonus*; it is, however, without doubt most closely related to the former. When the previous family Telestidae in agreement with HICKSON (1939) is raised to the rank of an order, it is also the most convenient to remove *Pseudocladochonus* from the Telestidae sensu stricto and place it in a family of its own, which may be named

Pseudocladochenidae n. fam.

comprising but one genus with only one species. VERSTUYL'S excellent description includes all the facts which speak for in-

cluding the species in the order *Telestacea* and for excluding it from the family Telestidae s. str. from which it differs principally in the equal zooids and the sympodial branching.

Distribution. *Telestata septentrionalis* was taken by the Ingolf Expedition in the North Atlantic Ocean southwest and west of Iceland, in Denmark Strait and Davis Strait, at depths from 740-2418 m. Probably it is a common species in the deeper parts of the North Atlantic Ocean. Chart of distribution, fig. 15.

Ordo: Xeniidea Bock 1938.

The order consists of soft-bodied Octocorals which have comparatively little contractility and mainly by their cylindrical anthocodiae are distinguished from the other orders of Octocorals. In the order only one family is

Remarks. It may be noted that KUKENTHAL (1902, 1906) and HICKSON (1931), as a diagnostic character for this group, use the feature that the zooids only possess one pair—the dorsal pair—of mesenteric filaments. In *Ceratocaulon wandeli*, however, mesenteric filaments are present on all the mesenteries.

Family Xeniidae Verrill 1866

Members of this group are for the most part shallow water forms, but there are two deep water forms which are known at present to be occurring in the North Atlantic Ocean, being

is fairly well extended and measures 45 mm in height, and of this the stalk makes out 25 mm. The smallest specimen measures about 20 mm, and of this the stalk is about 12 mm. The diameter of the cylindrical stalks are 8 mm and 5 mm respectively. The largest specimen has about 25 zooids, the smallest about 14 zooids.

Genus *Ceratocaulon* Jungersen 1892.

Polyps with Xeniidae with monomorphic zooids and a distinct two-tiered anthocodiae and surrounded by a conspicuous cuticle. The anthocodiae set in a fairly distinct row and are retractile.

The cylindrical stalks are often somewhat narrowed just above the base which is slightly expanded and has been attached to stones or similar solid objects. The stalk is surrounded by a very conspicuous, fairly thick, firm and smooth, brown, horny or chitinous cuticula, which, towards the cluster of anthocodiae, gradually becomes thinner and a little below these is not discernible any more. In the largest and most extended specimen the cuticula disappears about 5 mm below the anthocodiae. In the type specimen, which measures about 25 mm, it disappears about 1 mm below the anthocodiae.

Ceratocaulon wandeli Jungersen 1892.

The specimens all have a single stalk; JUNGERSEN (1917, p. 8) says, however, about a specimen from the museum of Bergen

- Ceratocaulon wandeli* Jungersen 1892, p. 231-232, figs. 1-4.
Xeniidae Kükenthal, 1906a, p. 21-22.
 Møller, 1915, p. 39.
Ceratocaulon wandeli Jungersen 1917, p. 8-9.
Xeniidae Hickson 1931, p. 154.
 Møller, 1932, p. 10.
 1. N. 22. W. 540. H. 0.24. Wandel. The type specimen.
 2. N. 22. W. 621. H. 0.75. Ingolf. St. 15. 1 colony.
 3. N. 7. W. 751. H. 0.10. Ingolf. St. 143. 1 colony.
 4. N. 142. W. 790. H. 0.78. "Michael Sars". St. 96.
 5. N. 24. W. 810. H. 0.50-0.79. Thor. St. 255 (14).

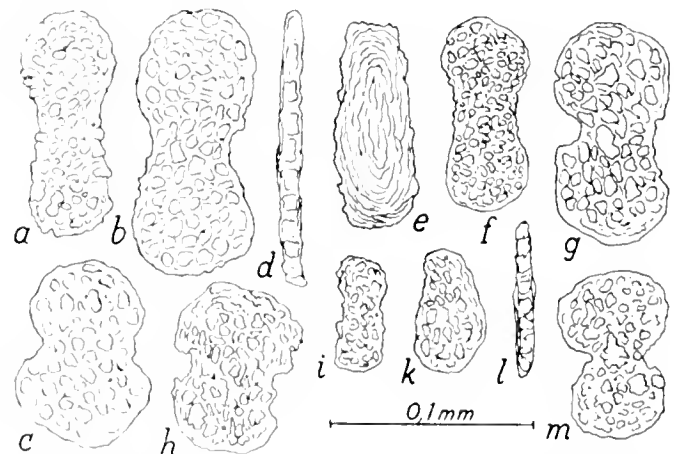


FIG. 19. *Ceratocaulon wandeli* Jungersen. a, sclerite from tentacle dorsum, b, d, sclerites from anthocodia, d, shown from the side, c, m, sclerites from stalk, l, shown from the side.

tentacles bear a single row of pinnulae laterally. Number of pinnulae about 20 pairs. The pinnulae are of the ordinary growth. The upper part of the pinnulae retract into the cuticular sheath, leaving a rough surface, and a sclerite, about 0.1 mm, present in a pinnula. The sclerite of the stalk, anthocodial and tentacle sclerites. Mesenteric filaments present.

Measurements of zooids of the species are given in the table. Tentacles with two rows of pinnulae in each row.

The colony of one of the specimens is fairly well preserved in alcohol. The largest specimen

that "the thick "horny" stem divides into two equal, short and clumsy, branches, each covered with its cuticular sheath and terminating with its cluster of polyps."

The colour of one of the specimens when alive was for the stalk stated as brown, horn coloured, a colour still retained in the cuticula of the preserved specimen. The cluster of antho-

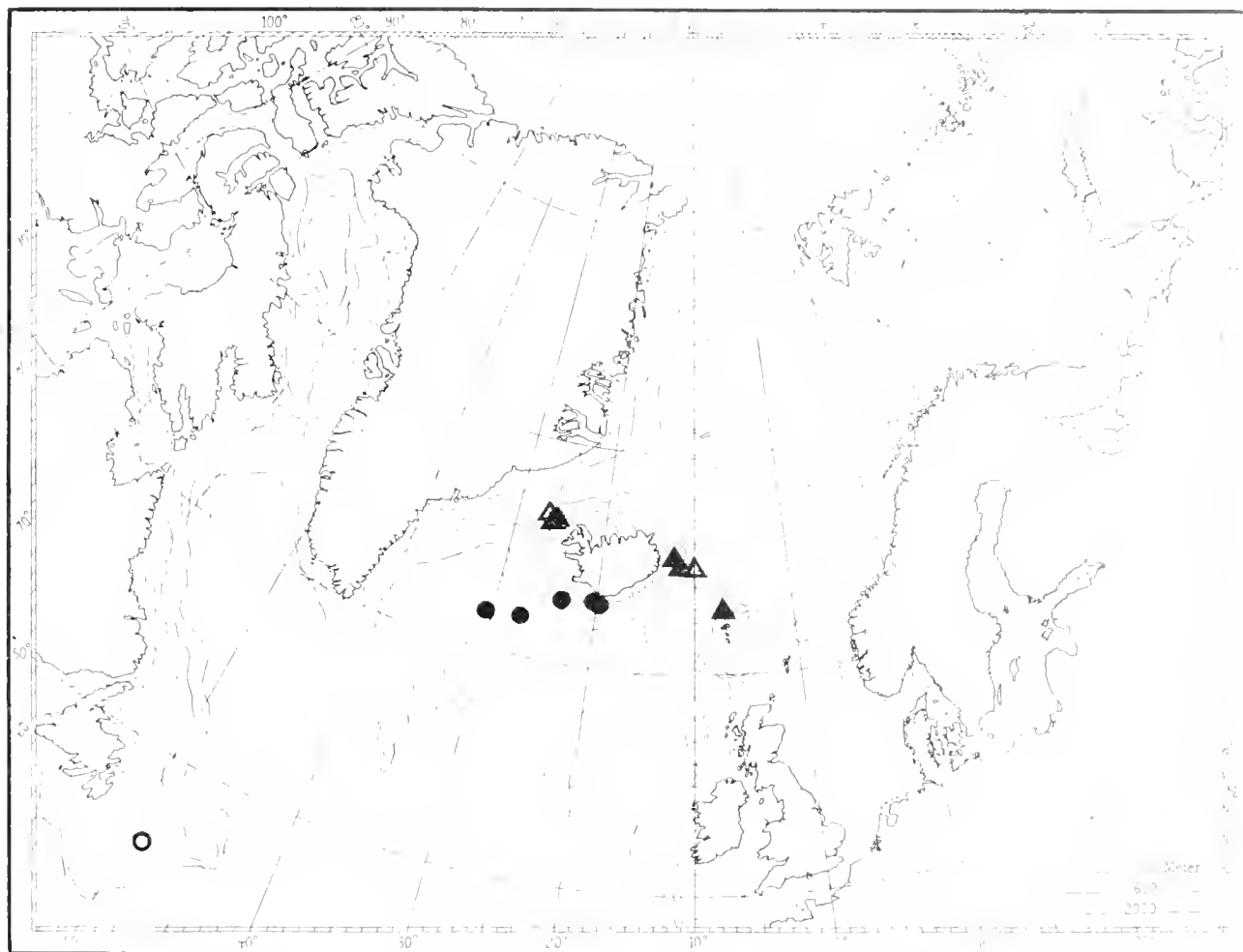


Fig. 20. ▲ *Ceratocaulon wandeli* Jungersen.
● *Acanthopora armata* Verrill.

codiae was described as distinctly pale-red, their colour in the specimens in alcohol is now greyish or light brown.

In contracted specimens the anthocodiae are apparently closely set in equal height. In more extended specimens, however, the stalk is lengthened, especially in the centre of the colony, and the anthocodiae are then placed more scattered. The lowest and outermost placed anthocodiae are moreover also the shortest, and thus in the most extended specimens the tentacle crowns become quite free of each other.

The largest anthocodiae measure about 14 mm in height, the largest tentacles a little more than 3 mm. Latero-ventral on each side of the tentacle there are about twenty slender pinnulae in a single row.

The anthocodiae of the Xenidae are usually described as non-retractile, a statement which often also is used in the diagnosis of the family. HICKSON (1931 p. 138) states, however, about the anthocodiae that "there can be no doubt that they all possess considerable powers of contraction." This species shows power of complete retraction of the anthocodiae and also a considerable power of contraction of the entire colony. In one of the present colonies thus most of the anthocodiae are completely retracted, and JUNGENSEN (1917 p. 9) writes about the specimens from the Bergen Museum that "in two cases the upper naked part of the stem with all its polyps was found retracted into and concealed in the "horny" part," a very interesting statement which has been quite overlooked by the later students of the Xenidea. Probably the retraction of the anthocodiae in *C. wandeli* is a very slow process only taking place when exposed to very

strong influences. By retraction the anthocodiae are entirely inverted.

The stalk of the species is composed of the anthostelae of the zooids which reach down nearly to the base of the colony. The zooids are arranged with their dorsal sides towards the axis of the colony and the mesogloea tissue is but poorly developed.

The sclerites are, as is usual in the Xenidea, of a very uniform type. They are minute, flat, oblong disks with a rugged surface, rounded ends and generally a constricted middle. The largest measure 0.13 - 0.06 mm, most are, however, somewhat less than 0.1 mm (fig. 19).

The spiculation of the specimens is very varying, thus one of the specimens is apparently entirely devoid of sclerites, this may of course be due to bad preservation, but in another specimen where sclerites are found in the stalk such are almost quite absent in the anthocodiae, and a specimen with fairly richly spiculated anthocodiae has a stalk only poorly provided with sclerites. In the most spiculated specimen the sclerites of stem are closely crowded in a rather thick layer and in the anthocodiae they are closely and crosswise arranged in eight broad intermesenterial rows which also continue uninterruptedly in the dorsum of the tentacles. In the tentacles the sclerites are longitudinally arranged. The row of sclerites is at the tentacle base about three to four sclerites broad, becomes, however, in the upper part of the tentacle, rapidly only one sclerite broad. Besides the interseptal rows of sclerites a few sclerites are found along the attachments of the mesenteries in the uppermost part of the zooid body. Sclerites may also be found in the stalk in the walls between the colonies.

the anterior part with 4 observed setae, the posterior part with 30 setae. The 10 pairs of legs of the head are provided with two dorsal mesonotal setae and one ventral seta, whereas the 10 pairs of legs of the abdomen have a length of 1.5-2.0 times the width. The type specimen is deposited in the Museum of Natural Sciences, Stockholm, Sweden, respectively.

The genus *C. (C.)* was not by KUKEN (1922) regarded as a distinct genus, but only as a subgenus of *Xenodermatobia* (comprising also *Xenodermatobia antarctica* BOGALTTA, 1922, pp. 1-2, pl. 5, figs. 22-24). The genus *C. (C.)* as *C. (C.)* was, however, not followed

here, as, in reality, the horny cuticula, the comparatively large sclerites, the presence of eight mesenteric filaments, and the whole primitive character of *C. (C.)* speaks for placing it in a distinct genus. To this genus should also be reckoned the only other known deep water species of Xenidea, viz. *X. antarctica* Kükenthal from the Bouvet Island at 157 m depth.

C. (C.) has, as distinct from *C. (C.)*, the oral side of the tentacles provided with numerous small warts. *C. (C.)* has further the horny cuticula incrustated with foreign bodies. The absence of sclerites in *C. (C.)* is, however, of no distinguishing value as such may apparently be absent also in *C. (C.)*.

Distribution: The species is found in the Norwegian Sea in several localities just north of the Wyville Thomson Ridge from the Faroes to the Danmark Strait at depths from 510-850 m. Chart of distribution, fig. 20.

Ordo: **Alcyonacea** Verrill 1866 emended.

Family: **Alcyoniidae** Verrill 1865.

Subfamily: **Alcyoniinae** Kükenthal 1906.

Diagnosis: Vide KÜKENTHAL 1906 *a*, p. 28.

Genus: **Alcyonium** Linné 1758 emended.

Diagnosis: Vide KÜKENTHAL 1906 *a*, p. 28.

Alcyonium digitatum Linné 1758.

Alcyonium digitatum Broch, 1912 *d*, p. 27-31, figs. 18-23.

norvegicum Broch, 1912 *d*, p. 21-26, figs. 11-17.

digitatum Molander, 1915, p. 36-42, fig. 11, pl. 2
figs. 21-26.

Jungersen, 1917, p. 5-8.

(?) Deichmann, 1936, p. 18-19.

- *norvegicum*, Pax 1936, p. 223-225, figs.
117-119.

Carlgren, MS.

For further references vide JUNGENSEN 1917.

Description: Vide BROCH 1912 *d* and MOLANDER 1915.

Remarks: Of modern authors BROCH (1912 *d*) has tried to distinguish the northern *Alcyonium* into two different species, i. e. besides the typical *A. digitatum* L., in a form which always has membranous growth and is identical with *Sympodium norvegicum* Koren & Daniélsen 1883. MOLANDER in 1915 recognized only one species, however divided into two varieties, viz. *A. digitatum* var. *norvegicum* and *A. digitatum* var. *digitatum*, both varieties typically of the usual clumsy, lobular shape, but sometimes also found in membranous forms.

It seems, however, impossible to maintain that the two forms are specifically different and also JUNGENSEN (1917) regarded them as synonymous.

Distribution: The species is found on the European coast as far north as Lofoten and as far south as the Bay of Biscay. It is further found at the Faroes and off South Iceland and probably also on the American coast off New England and Nova Scotia.

Bathymetrical distribution: 1-789 m, generally, however, at depths from 20-150 m.

Vide further JUNGENSEN 1917 and the author's paper in 1913.

Genus: **Anthomastus** Verrill 1878.

Diagnosis: Vide KÜKENTHAL 1906 *a*, p. 29.

Anthomastus grandiflorus Verrill 1878.

Anthomastus grandiflorus Jungersen, 1927, p. 5-11, textfigs.
1-4, pl. I.

Remarks: The very large collection which the Ingolf-Expedition secured on 13 different stations has previously been worked up by JUNGENSEN in "The Danish Ingolf Expedition", Vol. V, pt. II.

Distribution: The species is found on the Norwegian coast from Bergen to Trondheim and in the southern part of Davis Strait. It is further found in the North Atlantic Ocean south of the Wyville Thomson Ridge (s. lat.), and in the eastern part of the Atlantic Ocean it is found as far south as the Canaries, in the western part as far south as the Caribbean Sea.

Bathymetrical distribution: About 110-2875 m.

Family: **Nephtyidae** Gray 1859

Diagnosis: Vide KÜKENTHAL 1903 p. 4-5.

Remarks: The family Nephtyidae is very difficult to work up because so many genera and species have been established, most of which have proved not to hold true. In this family were thus described several genera from the northern seas with all together more than half a hundred species, though probably all these species are merely synonyms of a few of one or two genera.

KÜKENTHAL (1906, 1907) was the first to reduce the great number of species, those recognized by him mostly belonging to the genus *Euaephthya* sensu Kükenthal, and some years later

MOLANDER and JUNGENSEN, both in 1915, published the results of their studies on the group. Both the last mentioned authors agree with KÜKENTHAL that most of the previously described forms are invalid, both being of the opinion that still some of the species recognized by KÜKENTHAL are but synonyms of others. Otherwise, however, the two authors do not agree.

JUNGENSEN, after having examined more than a thousand specimens from the northern seas, arrived at the result that all the northern Nephtyidae belonged to only four species of one single genus (*Euaephthya* sensu Jungersen), in his papers (1915, 1917) he published very extensive lists of synonyms to these

of the genus *Eunephthya* with state-
ments which would be set
aside. As a result of the Ingolt Expedi-
tion, however, in the introduction, JUNG-
ERSEN (1916) writes even before he had time
to see the material:

I have not had time to publish the results of his studies
on the genus *Eunephthya*. In 1928 published a treatment of
the genus *Eunephthya* and p. 4 in this paper Broch
has given the opportunity during several discourses
to state my own points of view completely.

As BROCH (1936) recognizes among the northern
Nephthiidae the four species mentioned by JUNG-
ERSEN, and as he has pointed out and discussed below, some
of the views of BROCH and DEICHMANN's descriptions
are incorrect.

The author of this paper deeply regrets that JUNG-
ERSEN, the founder of the northern Nephthiidae, did not
publish a more extended report, as many points are still rather
unclear. Some of JUNG-ERSEN's determinations of the specimens
in the collection of the Zoological Museum of Copenhagen need
to be corrected, that the material directly yields the examiner.
The modern students of the northern Nephthiidae all
agree in the division of these corals into two main groups
according to their capability of retraction of the anthocodiae.

The first group, with entirely retractile anthocodiae, com-
prises according to JUNG-ERSEN, BROCH, and DEICHMANN the two
species *Eunephthya rubiformis* (Ehrenberg) and *E. fruticosa* (M.
Sars), according to MOLANDER the group comprises the genus
Gersemia Molander, in which he, however, recognizes several
species. MOLANDER (1917), and VERRILL (1922) agrees quite with
MOLANDER's point of view.

The second group, with non-retractile anthocodiae, com-
prises according to JUNG-ERSEN, BROCH, and DEICHMANN, the two
species *L. gracilata* Verrill and *L. flauda* (Rathke); according to
MOLANDER the group comprises the genus *Eunephthya* with
three species recognized, distributed over two groups, viz.
E. gracilata gracilata and *L. andalbatia*. The first group cor-
responds to *L. gracilata* sensu Jungersen, the second group to
L. andalbatia sensu Jungersen. VERRILL (1922) agrees practically with
MOLANDER except that he retains the generic name
Eunephthya for one other species and for the first of MOLANDER'S
forms under the generic name *Dipla* Danielssen 1887, for the
second form under *Dipla* Korten & Danielssen 1883.

In the second group, with retractile anthocodiae, it
is generally supposed that the three authors JUNG-ERSEN,
BROCH, and DEICHMANN were in accordance in their view on
the species which they recognize in the group. This is, how-
ever, not the case. In a foot note by DEICHMANN (1936)
it is stated that the material is to be found, "Broch's figure 5,
is not correctly and correctly wrongly labelled. Both the shape
and the colour of the spicules indicate *L. rubiformis*, likewise
the colour of the spicules of them are deep red. I have never seen
any other species of red spicules in *E. fruticosa*,
and the figure 4, labelled *rubiformis*, resemble much
more *E. fruticosa*. DEICHMANN is, however, not right in
this. BROCH and DEICHMANN have been confused. The
figures 4 and 5 of BROCH's key of determination
of *Eunephthya* and correspond also to the
figures 4 and 5 of BROCH's paper (1912 b). BROCH (1935, p. 19)
states that he is not certain whether the colour of the
spicules is of red or not, that "Durch
Vergleichen der *Eunephthya fruticosa* im
Zoologischen Museum mit dem Stücker und starker zur
Eunephthya rubiformis".

It is clear that BROCH's *E. rubiformis* does not
belong to the same species as DEICHMANN'S *E. fru-*
ticosa. The two species are identical. Likewise there

is no absolute accordance between BROCH'S and JUNG-
ERSEN'S conceptions of *E. rubiformis* and *E. fruticosa*. As appears from the
above, red sclerites may possibly be found in BROCH'S *E. fruticosa*
whereas such do not occur in JUNG-ERSEN'S *E. fruticosa*. JUNG-
ERSEN (1916, p. 189) writes about *E. rubiformis* that "The colour
in this species is due to red spicules" and he further states that
all other arctic species of *Eunephthya* possess colourless sclerites.
Nor are JUNG-ERSEN'S and DEICHMANN'S *E. rubiformis* quite
identical as appears from JUNG-ERSEN'S papers where, among
the synonyms to *E. rubiformis*, is included *E. rubiformis* Broch
(1912 b) which apparently according to DEICHMANN should be an
E. fruticosa.

E. rubiformis sensu Deichmann comprises among other forms
E. rubiformis sensu Molander to which also VERRILL'S *E. rubi-*
formis corresponds. Also JUNG-ERSEN (1917) includes in his syno-
nym list to *E. rubiformis*, *E. rubiformis* sensu Molander. BROCH
(1928, p. 15) states however "JUNG-ERSEN (1916), probably by a
slip of the pen, ranged MOLANDER'S "*Eunephthya rubiformis*"
among the synonyms of *Eunephthya rubiformis* (Ehrenberg).
MOLANDER'S figures and description, however, leave no doubt
that these have been based upon typical specimens of *E. fruti-*
cosa (M. Sars)." It is evident that MOLANDER'S *E. rubiformis*
belongs to BROCH'S *E. fruticosa*; it is, however, not quite certain
that JUNG-ERSEN referred MOLANDER'S *E. rubiformis* among the
synonyms to *E. rubiformis* in his sense owing to "a slip of the
pen"; it may be due to a conception of *E. rubiformis* which does
not quite correspond to BROCH'S view on the same species.

The great confusion which exists as to the various students'
conception of the northern Nephthiidae with retractile antho-
codiae is only indicated in the above-mentioned. Below, under
the discussion of the genus *Gersemia*, the author revert to the
problem in more detail.

In the second group of northern Nephthiidae, those with
non-retractile anthocodiae, there are better accordance between
the modern students' conceptions of the species, though the two
species, which JUNG-ERSEN, BROCH, and DEICHMANN recognize,
according to MOLANDER (and VERRILL) each is a group of species.

During the preparation of this paper the author has had
access only to the material in the collection of the Zoological
Museum of Copenhagen. However, in the 25 years which have
elapsed since JUNG-ERSEN'S death, this material has grown to such
an extent that it now comprises more than 1300 specimens from
very widely spread localities in the North-Atlantic Ocean and
adjacent seas. Thus the material in the museum of Copenhagen
is now about one fourth larger than the whole material to which
JUNG-ERSEN had access in the Scandinavian museums, and con-
sequently allows a thorough examination of the northern Neph-
thiidae.

At first the author was convinced that JUNG-ERSEN had gone
far too wide in his synonym lists and that a point of view, such
as taken by MOLANDER, would be the most convenient. However,
by repeated, careful studies of the great material at disposal the
author arrived at the conclusion that in the group with retractile
anthocodiae only one single species might be distinguished, that
thus *E. rubiformis* and *E. fruticosa* are synonymous, and that in
the other group, with non-retractile anthocodiae, it was possible
to distinguish between the same two species as JUNG-ERSEN did.
The one of these species, however, comprising three forms which
possibly may be regarded as valid sub-species (one of the forms
perhaps a valid species) and which correspond to MOLANDER'S
species in the group *Eunephthya dicaricatae glomerata*. It is to
be noted that the material of one of these forms, identical with
MOLANDER'S *E. quoclandua*, only recently has been included in
the collections of the Zoological Museum of Copenhagen and,
though JUNG-ERSEN knew the form from MOLANDER'S paper, he
had no personal knowledge of it at the time when he wrote his
papers on the northern Octocorals.

The present author believes that it is correct to maintain a

generic difference between the two main groups of northern Nephthyidae, as done by MOLANDER (1915, 1918) and by BROCH (1939), and this course, which also seems the most convenient, is consequently followed below. Which generic names, however, should be used for these groups may be discussed.

JUNGERSSEN ranged, as mentioned above, all the northern Nephthyidae into one single genus, viz. *Eunephtya*, a genus which was established by VERRILL (1869 p. 284) and in which VERRILL as type designated a previously described species, "*Nephtya thyrsoidea* Verrill 1865". Besides, VERRILL in the genus described a new species, *Eunephtya glomerata*, the type specimen of which was from Greenland and sent to VERRILL by LÜTKEN under the MS. name *Alcyonium glomeratum*.

The original description of *E. glomeratum* is short but distinct and VERRILL in 1922, more than fifty years later, redescribed the type specimen and gave several figures. The original description of *Nephtya thyrsoidea* is, on the contrary, very deficient, and KÜKENTHAL (1907 p. 377) doubts whether that species actually belongs to the genus *Eunephtya* in his sense. VERRILL (1922) shares this doubt, and redescribes on p. 29 *Eunephtya thyrsoidea* on a cotype, as the original specimen was destroyed by the great Chicago fire, and states that his species is identical with *Capnella rugosa* Kükenthal 1901. VERRILL is of the opinion that KÜKENTHAL has made a wrong use of the generic name *Capnella* (in his diagnosis of the new genus *Capnella* erected for *Alcyonium imbricatum* Quoy & Gaimard GRAY (1869 p. 129) e. g. states "Polypes retractile"), and VERRILL therefore retains the generic name *Eunephtya* for those species which KÜKENTHAL ranges into the genus *Capnella*, and instead he uses for *Eunephtya glomerata* the generic name *Drifa*, the genus then having as type *Drifa hyalina* Daniélsen 1887, a species which is identical with VERRILL's original *E. glomerata*. In those forms which VERRILL (1922) ranges in *Drifa glomerata* are, however, included both forms which belong to *E. glomerata* sensu Molander and such forms which belong to *E. flavescens* sensu Molander. *Drifa hyalina* belongs to the latter.

BROCH in 1928 with a query places *Eunephtya thyrsoidea* Verrill 1922 among the synonyms of *E. glomerata*, and DEICHMANN (1936) does the same without a query. The present author can not agree with this; VERRILL's description in 1922 distinctly shows that *E. thyrsoidea* is different from *E. glomerata* and, as VERRILL states, most probably identical with *Capnella rugosa* Kükenthal. If this is correct the species should, however, still be named *Capnella rugosa* Kükenthal 1902, as not until VERRILL's redescription in 1922 was an identification possible. It is in any case not correct when VERRILL rejects the generic name *Capnella*. This name is prior to *Eunephtya* and that GRAY describes the zooid in *C. imbricata* (Q. & G.) as retractile should, if not a misprint, probably only allude to the fact that the tentacles may be withdrawn and the body contracted; QUOY & GAIMARD themselves (1833 p. 281) describe the zooids in *Alcyonium imbricatum* as "non retractilibus", and KÜKENTHAL's descriptions (1903) of the species of *Capnella* leaves no doubt as to the generic accordance between *C. imbricata* and *C. rugosa*.

There exists, however, much accordance between *Eunephtya glomerata* and the genus *Capnella* and maybe it is impossible to maintain a generic difference between the species of Nephthyidae now reckoned to *Capnella* and the North-Atlantic Nephthyidae with non-retractile anthocodiae. BROCH (1939 p. 11), in any case, arrives at the result that there is no generic difference between the two groups and that consequently *E. glomerata* and *E. florida* should be ranged into the genus *Capnella* whereas the generic name *Eunephtya* should be retained for the other northern Nephthyidae, i. e. those with retractile anthocodiae.

If actually *E. glomerata* Verrill belongs to the same genus as *Capnella imbricata* (Quoy & Gaimard), and much favours this supposition, then BROCH is right that the generic name *Capnella* Gray 1869 should be used for the northern Nephthyidae with

non-retractile anthocodiae, and also in the present paper that name is adopted for the group. It will, on the other hand, be evident from the above that the generic name *Eunephtya* should not be used for the northern Nephthyidae with retractile anthocodiae if this group is referred to a genus different from that into which *E. glomerata* is ranged, and that BROCH (1939) made a lapse in doing so. The correct generic name for the group would have been *Gersemia* Marenzeller 1878 as used both by MOLANDER (1915, 1918) and by VERRILL (1922). The name *Gersemia* was by MARENZELLER (1878 p. 375) first used for some specimens which he identified with RATHKE's *Gorgonia florida*, a species belonging to the group with non-retractile anthocodiae. MARENZELLER showed, however, in 1886 that the identification was not correct and renamed the specimens *Gersemia danieleseni*, and KÜKENTHAL (1906) after a re-examination identified the specimens with his *E. frutesca*. On account of this and of the fact that also the other species described by MARENZELLER 1878 in the genus *Gersemia* (*G. longata*) clearly belongs to the group with retractile anthocodiae it is thought that no objection can be made against the use of the generic name *Gersemia* for that group.

Genus: *Gersemia* Marenzeller 1878.

Remarks: In this group of northern Nephthyidae, easily recognized by their entirely retractile anthocodiae, a considerable number of species have, as mentioned above, previously been described. Some of the modern students of the group recognize, however, only two species, though, as shown above, without agreeing in their conception of these.

BROCH (1928) uses as the principal distinguishing character the shape of the sclerites in the bark of the branch tips of the colony. In *E. rubiformis* sensu Broch these sclerites should be "broad, fusiform or rod-like ones, up to 0.35 mm in length and richly armed all over with irregularly arranged compound wharts" (BROCH 1928 p. 9).

In *E. frutesca* sensu Broch these sclerites should mainly be "double stars up to 0.2 mm in length and with transversally arranged compound wharts generally in whorls" (BROCH 1928 p. 9).

DEICHMANN (1936) uses another character to distinguish the group into two species. *E. rubiformis* sensu Deichmann comprises those specimens which have "Zooids small, only few mm in length, and retractile into the common cushion of tissue which forms the end of the branches" (DEICHMANN 1936 p. 60).

E. frutesca sensu Deichmann comprises those specimens which have "Zooids tall, up to 8 mm in length, and retractile singly into the tubes which form the ends of the branches" (DEICHMANN 1936 p. 60).

Explained in another way *E. rubiformis* sensu Deichmann has small anthocodiae set well apart on the branches, and thus when the anthocodiae are expanded there are distinct spaces of coenenchyma between them. In *E. frutesca* sensu Deichmann, however, the larger anthocodiae are more closely set, and when expanded no distinct spaces of coenenchyma are left between them.

The collection in the Zoological Museum of Copenhagen includes nearly a thousand specimens which may be ranged in the genus *Gersemia*, many of them determined by JUNGERSSEN. A study of the comparatively few specimens which JUNGERSSEN labelled *rubiformis* showed that all these specimens possessed anthocodiae set well apart and of a size less than about 3 mm, and thus these specimens also belong to *E. rubiformis* sensu Deichmann. Only some of them (colonies from the Kara Sea, Spitzbergen and East Greenland) had, however, in the branch tips, sclerites corresponding to BROCH's diagnosis of *E. rubiformis*, whereas the other colonies from about the same localities possessed sclerites by which they could just as well or better have been referred to *E. frutesca* BROCH, and the same may be said about

NO. 1. *E. rubiformis*. A colony from off Cape Cod, U.S.A., in the Smithsonian Institution (No. 1000) is a typical *E. rubiformis* sensu Deichmann. A colony from Alaska, a name not included in the description, is also a typical *E. rubiformis* sensu Deichmann, as determined by DEICHMANN (1916, p. 10). A colony from the U.S. Nat. Mus., No. 1000, is also a typical *E. rubiformis*. All these colonies were taken in 1870, and are possibly of the same lot. The material from off Cape Cod agree also perfectly with the description of *E. rubiformis* given by DEICHMANN (1916, p. 10). It appears from the above that *E. rubiformis* sensu Deichmann and *E. rubiformis* sensu Jungersen was the same, this is also shown as is shown by a study of the material by DEICHMANN (1916, p. 10). In fact more than half of the specimens regarded as *E. fruticosa*, correspond with *E. rubiformis* sensu Deichmann, and both in the material of DEICHMANN (1916, p. 10) corresponding to *E. rubiformis* sensu Deichmann and that corresponding to *E. fruticosa* sensu Deichmann the specimens are found with sclerites by which they correspond to *E. rubiformis* sensu Broch.

Other colonies from West Greenland (Jakobshavn, Egedesboe) are typical *E. fruticosa* corresponding to *E. fruticosa* sensu Deichmann. The colour of the stem from base to top of the colonies was quite uniform, sclerites, such ones only occurring in the anthocodial part of the root body.

Apart from the colour of sclerites it has been found quite possible to find characters by which JUNGENSEN may have regarded his *E. rubiformis* from his *E. fruticosa*. It seems as if JUNGENSEN has regarded specimens of *Gersemia* with more or less sclerites as *E. rubiformis*, and the few colonies, by JUNGENSEN regarded *E. rubiformis*, in which the sclerites now are missing have probably lost their colour since the time when JUNGENSEN determined the specimens. From his papers (1887, 1916, 1917) it also appears that *E. rubiformis* sensu Jungersen correspond sclerites, and in the paper (1916, p. 10) JUNGENSEN states the correct identification of a specimen which is thought to be a typical *E. rubiformis* though it "does not possess the usual sclerites, but is only showing a faint pink colour."

By a careful study of the large collection at disposal the author has reached the conclusion that whereas it perhaps might have been possible to separate a smaller collection of northern Nephthya with retractile anthocodiae into various "species" as done by MOLANDER (1915, 1918*a*) this becomes quite impossible in the case of the present large collection. In spite of many attempts the author has been quite unable to find any distinguishing character of any value.

Other than the colour of the sclerites, as apparently used by DEICHMANN (1916, p. 10) as the principal distinguishing character, is worth mentioning the stem, which apart from the colour of sclerites has been found to become distributed over two different groups.

The most striking character used by Broch (the shape of the stem) and the shape of the branch tips, is of equally small value. In a study of the material it is perhaps to be seen a clear difference, in a larger number of specimens, and all possible intermediate stages are found between the typical *E. fruticosa* sclerites sensu Broch and the typical *E. rubiformis* sclerites sensu Broch.

Other than the colour of the sclerites, about the character used by DEICHMANN (1916, p. 10) as the principal distinguishing character, a comparison with the coenenchyma has been made, but as all possible intermediate stages are found here between colonies of typical *E. fruticosa* sensu Deichmann and such of typical *E. rubiformis* sensu Deichmann, the character is of little value as any of the other characters mentioned above.

Material from the Zoological Museum of Copenhagen (No. 1000) is a typical *E. fruticosa* M. Sars, from Vadso (No. 1000) is a typical *E. fruticosa* in the sense of DEICH-

MANN, after the sclerites it may also come nearest to *E. fruticosa* sensu Broch.

It has thus been quite impossible to find any character or combination of characters which might be used to separate the northern Nephthya with retractile anthocodiae (= *Gersemia*) into more species or well-defined groups, as all the various forms in which *Gersemia* has been found pass by every structural degree insensibly into each other, consequently the author has regarded them all as belonging to one species which should be named *Gersemia rubiformis* (Ehrenberg 1831). And by this, maybe somewhat drastic, but the author thinks absolutely justifiable course, all the various problems, all the confusion which exists in connection with the synonymies of the northern Nephthya with retractile anthocodiae has been done away with.

It may be noted that also BROCH in 1935 doubts whether it is in reality possible to distinguish clearly between *E. rubiformis* and *E. fruticosa*. Broch writes on p. 19 about some specimens referred to *E. fruticosa* that: "man kann bei solchen Exemplaren auch gelegentlich in Zweifel geraten, ob eine scharfe Grenze gegenüber der vorhergehenden Art (*E. rubiformis*) vorhanden sei."

In a form such as *Gersemia rubiformis* (Ehrenberg) s. l. it is evident that the conception of species is more comprehensive than e. g. the conception of the species in the family Chrysogorgiidae, where only little variation is allowed or found in the various forms. *Gersemia rubiformis* may perhaps be regarded either as a group of very closely related races which have about the same geographical distribution and quite freely hybridize, or as a species which in nature mutates backwards and forwards just as *Diosiphila* in the laboratory.

The author thinks that the best of the proposed characters for distinguishing between two groups of the forms of *Gersemia fruticosa* is that proposed by DEICHMANN, though it may be regarded as of no specific value. In the enumeration below of the material collected by the "Ingolf" and other material kept in the Zoological Museum of Copenhagen it is therefore noted whether the specimens in question had small anthocodiae set well apart on the branch tips (*E. rubiformis* sensu Deichmann) or somewhat larger anthocodiae set more closely and thus when expanded without distinct spaces of coenenchyma between them (*E. fruticosa* sensu Deichmann). For part of the material it has, however, been impossible to state to which group the specimens in question should be reckoned.

It may be repeated that the rather few specimens with red sclerites (i. e. *E. rubiformis* sensu Jungersen) all belong to *E. rubiformis* sensu Deichmann. It may further be noted that no colony with red sclerites has been found with a distinct stem below the cluster of branches and that the largest colony with red sclerites measures only about 6 cm in height in contracted state. The reason why these specimens, representing *E. rubiformis* sensu Jungersen, in some way makes up a group of their own is without doubt that the presence of red sclerites is a rather rare phenomenon.

Gersemia rubiformis (Ehrenberg 1831).

- Lobularia rubiformis* Ehrenberg, 1831, p. 58, 59.
Eumphthya fruticosa = *E. rubiformis* Jungersen, 1915, p. 1171-1181.
Gersemia rubiformis = *G. uvaeformis* = *G. clavata* = *G. fruticosa* = *G. mirabilis* Molander, 1915, p. 18-70, fig. 12, pl. 1, figs. 1-13.
Eumphthya rubiformis = *E. fruticosa* Jungersen, 1916, p. 187-193.
 Jungersen, 1917, p. 9-11.
Gersemia rubiformis = *G. uvaeformis* = *G. clavata* = *G. fruticosa* = *G. mirabilis* Molander, 1918*a*, p. 1-11.
Gersemia rubiformis = *G. canadensis* = *G. carnea* = *G. fruticosa* = *G. mirabilis* = *G. clavata* = *G. studeri* = ? *G. longiflora* Verrill, 1922, p. 1-8, 20-28, 48-49, textfig. 4, 13, pl. 1, pl. 2, pl. 3, figs. 5-8, pl. 4, figs. 1-3, 8, pl. 5, figs. 3-5, pl. 11, pl. 11, figs. 3-3*a*, pl. 17*a*, fig. 1.

Gersemia rubiformis + *G. uvaeformis* + *G. clavata* = *G. fruticosa* Rylov, 1926 *a*, p. 65-70, figs. 3-10.
Gersemia rubiformis + *G. clavata* + *G. fruticosa* Rylov, 1926 *b*, p. 239-244.
Gersemia danielsseni + *Eumephtya fruticosa* J. A. Thomson, 1927, p. 44, 45-46, pl. 1, fig. 9.
Eumephtya rubiformis = *E. fruticosa* Broch, 1928, p. 3-10, 44-49, figs. 4, 5, 8, 9.
fruticosa = *E. rubiformis* Kramp, 1932, p. 10, figs. 3, 4.
rubiformis + *E. fruticosa* Broch, 1935, p. 17-19.
rubiformis + *E. fruticosa* Deichmann, 1936, p. 63-66, pl. 1, fig. 6, pl. 4, figs. 14-25.

Eumephtya fruticosa Pax, 1936, p. 227.
fruticosa = *E. rubiformis* Kramp, 1939, p. 3-4.
fruticosa Carlgren, MS.

For further references vide JUNGENSEN's papers and the list of synonyms at the end of this paper.

Description. References may be made to the papers by BROCH and MOLANDER, and to DANIELSSEN (1887). It should only be remembered that the differences which these authors use to separate species or even genera of the northern Nephthyidae are altogether useless, as the various forms pass insensibly into each other.

Anthocodiae larger, more closely set and when expanded without distinct spaces of coenenchyma between them

Anthocodiae fairly small and set well apart on the branches

Material:

63° 35' N, 10° 24' W, 512 m, 0.5 C, "Ingolf" St. 3	3
64° 07' N, 11° 12' W, 416 m, 2.5 C, ..	4
63° 06' N, 56° 00' W, 2258 m, 2.4 C, ..	24
63° 30' N, 54° 25' W, 1096 m, 3.3 C, ..	25
65° 14' N, 55° 42' W, 791 m, 3.5 C, ..	28
65° 34' N, 54° 31' W, 428 m, 0.2 C, ..	29
66° 35' N, 55° 54' W, 166 m, 1.6 C, ..	31
65° 17' N, 54° 17' W, 404 m, ..	34
64° 50' N, 56° 21' W, 2702 m, 4.5 C, ..	36
65° 00' N, 11° 16' W, 581 m, 1.0 C, ..	59
65° 38' N, 26° 27' W, 260 m, 5.9 C, ..	98
66° 23' N, 7° 25' W, 1802 m, 1.1 C, ..	104
65° 34' N, 7° 34' W, 1435 m, 1.0 C, ..	105
67° 14' N, 8° 48' W, 1619 m, 1.0 C, ..	111
67° 57' N, 6° 44' W, 2386 m, 1.1 C, ..	112
69° 34' N, 7° 06' W, 2465 m, 1.0 C, ..	113
69° 43' N, 8° 23' W, 4889 m, 1.0 C, ..	117
67° 29' N, 14° 32' W, 1666 m, 1.0 C, ..	120
67° 40' N, 15° 40' W, 932 m, 1.0 C, ..	124
67° 19' N, 15° 52' W, 552 m, 1.0 C, ..	126
63° 26' N, 7° 56' W, 887 m, 1.0 C, ..	138
63° 29' N, 6° 57' W, 1169 m, 1.0 C, ..	140
62° 58' N, 7° 09' W, 731 m, 1.0 C, ..	143
62° 53' N, 4° 14' E, 850 m, "Michael Sars" St. 51	51
62° 58' N, 1° 56' E, 1120 m, "Michael Sars" St. 35	35
62° 35' N, 4° 04' W, 650 m, 1.0 C, "Michael Sars" St. 67	67
64° 58' N, 14° 12' W, 565 m, 1.0 C, "Michael Sars" St. 96	96
66° 02' N, 11° 05' W, 1010-900 m, "Thor" St. 51 (03)	51
66° 20' N, 12° 40' W, 690 m, "Thor" St. 80 (08)	80
Godthaabsfjord, "Tjalfe" St. 52	52
66° 14' N, 56° 08' W, 325 m, "Tjalfe" St. 100	100
68° 20' N, 54° 03' W, 400-500 m, ..	107
69° 17' N, 52° 14' W, 430 m, ..	123
69° 46' N, 51° 35' W, ..	116
69° 46' N, 54° 22' W, ..	155
66° 22' N, 57° 16' W, 686 m, ..	366
65° 09' N, 53° 33' W, 55 m, ..	419
65° 06' N, 54° 49' W, 83 m, ..	422
55° 00' N, 56° 34' W, 314 m, 2.8 C, "Godthaab" St. 44	44
69° 14' N, 57° 22' W, 245 m, 1.20 C, (200 m) "Godthaab" St. 50	50
69° 50' N, 64° 36' W, 1880 m, 1.0 C, (1860 m)	54
71° 52'.5 N, 62° 42' W, 450 m, 0.72 C, (430 m)	73
75° 26' N, 62° 26' W, 820 m, 0.69 C, (790 m)	77
76° 36' N, 68° 54' W, 480-80 m, 1.3 C, ..	86
76° 25' N, 69° 38' W, 465 m, 1.19 C, (445 m)	107

Capnella glomerata (Verrill 1869) sensu Jungersen.

- Eumephytha glomerata* Jungersen, 1915, p. 1164-1169.
 - Jungersen, 1916, p. 193-195.
 - Jungersen, 1917, p. 11-16.
 - Molander, 1918*a*, p. 11-12.
Drifa glomerata + *D. racemosa* Verrill, 1922, p. 31-34, pl. 5 fig. 2 pl. 11 figs. 2, 4, pl. 15 figs. 4-5, pl. 17*a* figs. 2-3 textfig. 5.
Eumephytha glomerata Rylow, 1926*a*.
 - Broch, 1928, p. 3-10, 10-12, figs. 1, 2, 6.
 - Kramp, 1932, p. 5, fig. 1.
 - Deichmann, 1936, p. 61-62.
 - Pax, 1936, p. 227, fig. 153, 151.
 - Kramp, 1939, p. 2, 3.
Capnella glomerata Broch, 1939, p. 14.
 - Carlgren, MS.

For further references vide JUNGENSEN's papers.

Remarks: As mentioned above JUNGENSEN's and BROCH's *Eumephytha glomerata* corresponds to MOLANDER's group *E. divaricatae-glomeratae*, in which group MOLANDER (1915) recognizes three main species viz. *E. glomerata* Verrill sensu Molander, *E. flavescens* (Dan.) sensu Molander, and *E. groenlandica* Molander.

The collection of the Zoological Museum of Copenhagen includes about 200 specimens of *Capnella* referable to *C. glomerata* sensu lato and in this material all the forms recognized by MOLANDER are represented. A study of the material gave the result that the form, by MOLANDER named *Eumephytha groenlandica*, apparently is so well distinguished from the other forms of *C. glomerata* that it possibly should be regarded as a valid species. However, the great variation known in the northern Nephthyidae and the fact that in the material the form is only represented from two localities, are in favour of at present regarding *C. groenlandica* (Molander) as but a distinct form of the species *C. glomerata* (Verrill) s.l. The two other forms recognized by MOLANDER are generally also fairly well distinguished in the material; there are, however, some few colonies which occupy an intermediate position and thus can not with certainty be referred to any of MOLANDER's forms.

Capnella glomerata (Verrill) f. **lütkeni** (Marenzeller).

The form named in this way corresponds to MOLANDER's *E. glomerata*. It has the following synonyms.

- Ammothea arctica* Lutken, 1875, p. 186 (nomen nudum).
 - Luetkeni Marenzeller, 1877, p. 372-374, pl. 3 fig. 1.
Drifa islandica + *Nephthya polaris* Danielssen, 1887, p. 65-68, 92-98, pl. 6 figs. 30-71, pl. 13 figs. 2-15.
Eumephytha glomerata Broch, 1912*b*, p. 12-11, figs. 11-13.
 - Molander, 1915, p. 72-71, fig. 13*a*, pl. 2 fig. 19.
 - Molander, 1918*a*, p. 11-12.

Description: Vide MOLANDER 1915.

Material:

(Specimens which are not well distinguished from *C. glomerata* f. *flavescens* are marked with a query.)

- 63 35' N, 10 24' W, 512 m, 0.5 C, "Angolf" St. 3.
 (?) 63 13' N, 11 34' W, 170 m, 7 C, - 6.
 63 13' N, 15 41' W, 1130 m, 4.5 C, - 7.
 61 54' N, 55 10' W, 710 m, 3.8 C, - 27.
 65 34' N, 51 31' W, 128 m, 0.2 C, - 29.
 65 17' N, 54 17' W, 101 m, - 34.
 63 15' N, 15 07' W, 1197 m, 3.08 C, - 53.
 65 00' N, 11 16' W, 581 m, 0.1 C, - 59.
 64 56' N, 36 19' W, 381 m, 4.4 C, - 91.
 62 35' N, 4 01' W, 512 m, 0.09 C, "Michael Sars" St. 67.

- 65 12' N, 13 57' W, 413 m, "Thor" St. 189 (63).
 66 41' N, 56 08' W, 330 m, "Tjalfe" St. 100.
 69 08' N, 53 12' W, 261 m, - 113*a*.
 69 16' N, 54 35' W, - St. 116.
 65 09' N, 53 33' W, 55 m, - 119.
 65 06' N, 51 19' W, 83 m, - 122.
 71 52.5' N, 62 42' W, 450 m, 0.72 C, (130 m) "Godthaab" St. 73.
 (?) 77 05.5' N, 71 13' W, 790 m, 0.12 C, (725 m) "Godthaab" St. 87.
 76 25' N, 69 38' W, 165 m, 1.1-1.9 C, (115 m) "Godth." St. 107.
 76 10' N, 76 20' W, 85 m, 1.1-1.2 C, (75 m) "Godthaab" St. 111.
 76 08' N, 80 53' W, 80 m, 1.1-0.5 C, (70 m) "Godthaab" St. 116.
 Exeter Sound, Baffin Land 100 m, "Godthaab" St. 196*c*.
 63 06' N, 40 40' W, 437 m, 3.03 C, (130 m) "Dana" St. 6004, Iceland (Bakkefjord, Midfjord).
 Greenland¹⁾, numerous localities both off West and East Greenland; off East Greenland found so far north as Danmarks-Havn.

Remarks: The anthocodiae of this form have in contracted state generally a length of from 2 to 3 mm. The upper part (head) of the anthocodia is strongly spiculated on the abaxial side of the contracted zooid, whereas the adaxial side and the lower part (shaft) of the anthocodia are more poorly spiculated and sometimes quite devoid of sclerites. The sclerites of the abaxial side of the anthocodia are fairly clumsy clubs (fig. 22*c-g*) generally of a length of about 0.22 mm and with a maximum size of about 0.35-0.45 mm. They are arranged in a single layer with their heads pointing upwards and outwards and give the anthocodiae of this form of *E. glomeratum* a very characteristic appearance which has been very well figured by Broch (1912*b*). The figure has also been reproduced by Pax 1936, fig. 153.

Distribution: See under the following form.

Capnella glomerata (Verrill) f. **flavescens** (Danielssen).

To this form the following synonyms belong.

- Eumephytha glomerata* Verrill, 1869, p. 284.
Gersemia candida Koren & Danielssen, 1883, p. 9-10, pl. 5 figs. 1-15.
Drifa hyalina + *Nephthya flavescens* + *N. rosea* Danielssen, 1887, p. 59-61, 81-91, pl. 7 figs. 1-11, pl. 11, pl. 12, pl. 13 fig. 1.
Eumephytha racemosa Studer, 1901, p. 33-34, pl. 4 figs. 1-2.
 - flavescens Molander, 1915, p. 74-78, fig. 13*b*, pl. 2 figs. 15, 16.

Description: Vide MOLANDER 1915.

Material:

(Specimens which are not well distinguished from *C. glomerata* f. *lütkeni* are marked with a query.)

- 63 43' N, 15 44' W, 1430 m, 4.5 C, "Angolf" St. 7.
 64 21' N, 28 50' W, 1184 m, 3.5 C, - 10.
 (?) 66 18' N, 25 59' W, 621 m, 0.75 C, - 15.
 (?) 65 43' N, 26 58' W, 474 m, 6.1 C, - 16.
 64 54' N, 55 10' W, 710 m, 3.8 C, - 27.
 (?) 61 12' N, 9 36' W, 1026 m, 4.8 C, - 41.
 63 08' N, 15 40' W, 1304 m, 3.9 C, - 51.
 63 37' N, 43 02' W, 659 m, 3.4 C, - 57.
 62 58' N, 7 09' W, 731 m, 0.4 C, - 113.
 (?) 65 27' N, 51 45' W, 126 m, Wandel.
 (?) 66 49' N, 56 28' W, 112 m, -
 (?) 63 45' N, 9 35' W, 540 m, -
 5 sml. E. of Soudisfjord (Iceland), Wandel.
 69 46' N, 51 22' W, 471 m, "Tjalfe" St. 155.

¹⁾ Further particulars on the Greenlandic finds of Octocorals will be set forth in "The Zoology of East Greenland".

W	141° 57'		
W	140° 00'	Coast	St. 57
W	140° 50'	Dana	St. 2341
W	140° 00'	Dana	St. 2361
W	140° 00'		St. 2835
W	140° 00'		St. 2840
W	141° 00'	Dana	St. 6001
W	141° 00'		St. 6005

specimen of *Eumphtha glomerata* Verrill (cf. VERRILL, 1922) and of *Aciphothya flavescens* Danielsson are forms of *E. flavescens* which show likeness to specimens of *E. lütkeni*.

Distribution. According to MOLANDER'S statements (MOLANDER 1915, 1918*a*) and the material kept in the Zoological Museum of Copenhagen the two forms have an almost identical distribution comprising the Barents Sea, the Norwegian Sea, the Baffin Bay, the Davis Strait, the North Atlantic Ocean just south of the Waville-Thomson Ridge, and the West Atlantic Ocean off New-Foundland and Nova Scotia.

Also the bathymetrical distribution is identical for the two forms, *C. glomerata* f. *lütkeni* has been met with in depths from 14-1497 m, and *C. glomerata* f. *flavescens* has been found in depths from about 60-1184 m. (According to VERRILL *C. glomerata* in N. America is found at about 2700 m).

Though the distributions of these two forms of *C. glomerata* comprise the same areas, it seems that in the Norwegian Sea *C. glomerata* f. *flavescens* is by far the most common in the eastern part, off the coast of Norway, whereas in the western part, off the coast of East Greenland, *C. glomerata* f. *lütkeni* is the more common.

***Capnella glomerata* (Verrill) f. *groenlandica* (Molander).**

Eumphtha groenlandica Molander, 1915, p. 78-79, fig. 13 c, pl. 2 fig. 18.

Description: Vide MOLANDER 1915.

Material

North of northern Iceland.
62° 19' N, 8° 51' W, 501 m, 1-21°C. (190 m). "Dana" St. 6005.

Remarks. The colonies are low, less than 6 cm in height, and of a beautiful light-red colour (in alcohol). The anthocodiae are very large, in contracted state measuring up to 8 mm in length by 3.5 mm in width, and distinctly ridged and very powerfully armed with densely crowded sclerites. That which most clearly distinguishes this form from *C. glomerata* f. *flavescens* is, besides the somewhat larger and somewhat differently shaped clubs of the anthocodial head (fig. 22 s, a-x), the occurrence of numerous very long and slender spindles in the anthocodia (fig. 22 x, t). These spindles measure up to 0.6 mm or more i. e. nearly twice the length of the longest spindles in f. *flavescens*.

E. glomerata f. *groenlandica* is at present well distinguished from the other northern forms of *E. glomerata* and it might perhaps be regarded as a valid species.

VERRILL (1922 p. 34) supposed that *Eumphtha racemosa* Studer might be identical with *E. groenlandica* Molander. However, judging from the description and the measurements of sclerites given by STUDER (1901), this may not be the case. *E. racemosa* belongs undoubtedly to *C. glomerata* f. *flavescens*.

Distribution. Besides in the above mentioned localities, north of Iceland and a little west of the Faroes, the species is only known from an uncharted locality off East Greenland.

***Capnella florida* (Rathke 1806).**

- Gorgonia florida* Rathke, 1806, vol. 4, p. 20-21, pl. 137.
- Eumphtha florida* Broch, 1912*c*, p. 39-43, figs. 30, 31.
- Jungersen, 1915, p. 1169-1171.
- rosea* = *E. florida* = *E. spitzbergensis* Molander, 1915, p. 79-81, fig. 13*d, e*, pl. 2 figs. 14, 16, 20, 21, 23.
- Eumphtha florida* Jungersen, 1916, p. 195-197.
- Jungersen, 1917, p. 16-18.
- Diva multiflora* Verrill, 1922, p. 35-36, pl. 1 fig. 7, pl. 15 figs. 6, 7, textfig. 6, 7.

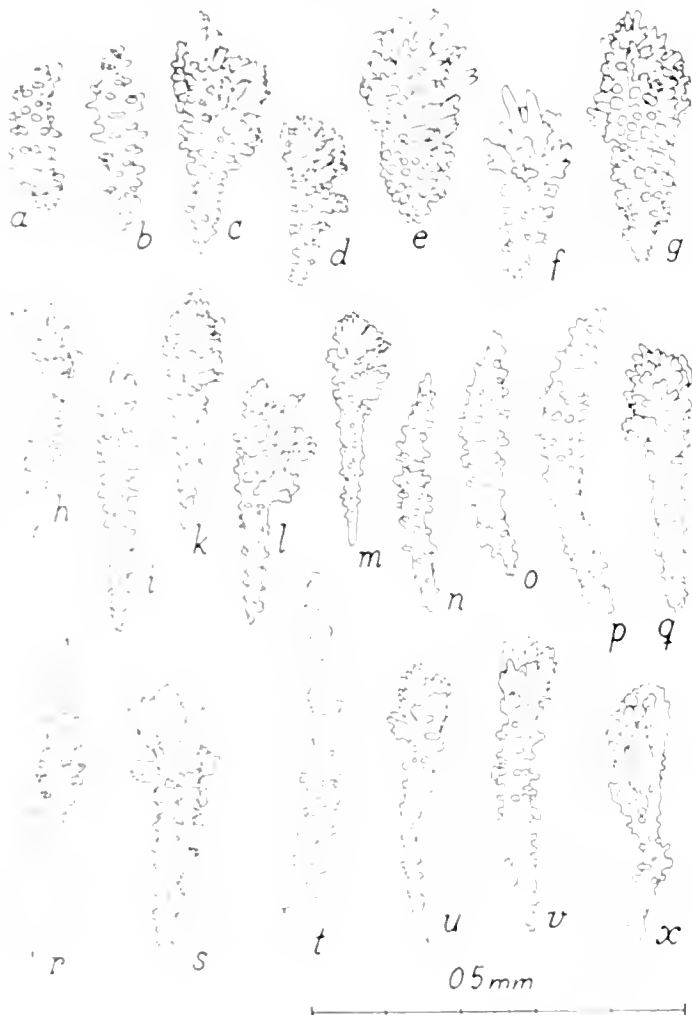


FIG. 22. a-x. Anthocodial sclerites, a-x, of *C. glomerata* f. *groenlandica* (Molander); a, c, of the specimen from the locality of the "Dana" St. 6005 (Danielsson); b, d, e, of the specimen from the locality of the "Dana" St. 2341 (Molander). The sclerites are drawn more or less in side view, a, b, c, d, e, f, g, from behind.

1. The length of the anthocodia in this form is somewhat greater than in f. *flavescens*, and also the anthocodial head is larger. The head is provided with eight distinct, rather long spines, which are more dense than in the other form. The shaft of the anthocodia is provided with a head tube at the mesenterial end (fig. 22 a, b), which may attain a length of 1.5 mm or more. Generally measure less than 0.5 mm in length, the sclerites are mainly cylindrical, but some are short shaft and attain a length of 0.6 mm or more. In *C. glomerata* f. *groenlandica* and *C. glomerata* f. *lütkeni* the sclerites are generally shorter, and the occurrence of long slender spindles may be mentioned. It may be maintained that the specimens from the locality of the "Dana" St. 6005 are *E. groenlandica*, both the type

Eumephithya rosea J. A. Thomson, 1927, p. 15.
florida Broch, 1928, p. 3, 10, 12-11, figs. 3, 7.
 Kramp, 1930, p. 1-5.
 Kramp, 1932, p. 6, fig. 7.
 Kramp, 1933, p. 17.
 Deichmann, 1936, p. 62-63.
 Pax, 1936, p. 227, fig. 155.
 Kramp, 1939, p. 3.

Capnella florida Broch, 1939, p. 14.
 For further references vide JUNGERSSEN 1917.

Description: Vide Broch 1912c.

Material:

62°30' N. 8°21' W.	249 m.	7°2 C.	"Ingolf" St.	1.
63°01' N. 9°22' W.	493 m.	5.3 C.		2.
63°35' N. 10°24' W.	512 m.	0.5 C.		3.
63°13' N. 15°11' W.	1130 m.	1.5 C.		7.
66°18' N. 25°59' W.	621 m.	0.75 C.		15.
63°30' N. 54°25' W.	1096 m.	3.3 C.		25.
66°35' N. 55°54' W.	166 m.	1.6 C.		31.
63°33' N. 15°02' W.	595 m.	5.9 C.		55.
63°37' N. 13°02' W.	659 m.	3.1 C.		57.
60°37' N. 27°52' W.	1505 m.	4.5 C.		78.
64°45' N. 27°20' W.	584 m.	8.1 C.		89.
65°38' N. 26°27' W.	260 m.	5.9 C.		98.
66°33' N. 20°05' W.	83 m.	5.6 C.		127.
62°58' N. 7°09' W.	731 m.	±0.1 C.		143.
67°03' N. 27°08' W.	375 m.		WANDEL.	
61°42' N. 27°13' W.				
65°30' N. 55°26' W.	525 m.			
65°22' N. 54°02' W.				
61°23' N. 5°01' W.	180 m.			
66°20' N. 25°12' W.	180 m.			
5 sml. E. of Seidisfjord (Iceland).			WANDEL.	
61°16' N. 11°15' W.				
61°40' N. 7°10' W.			"Diana".	
66°40' N. 11°05' W.	900-1040 m.		"Thor" St. 51 (03).	
63°15' N. 22°23' W.	216-326 m.			171 (03).
65°50' N. 26°51' W.	392 m.	5.83 C.		150 (01).
60°19' N. 2°50' W.	525 m.		"Michael Sars" St. 9.	
E. of the Faroes.	365-110 m.		"Michael Sars" St. 9.	
62°35' N. 4°01' W.	620-610 m.	0.09 C.	"Michael Sars" St. 67.	
62°53' N. 9°06' W.	160 m.	3.86 C.	"Michael Sars" St. 85.	
70°12' N. 51°28' W.	478 m.		"Tjalfe" St. 177.	
68°28' N. 51°17' W.	317-461 m.			199.
66°22' N. 57°16' W.	686 m.			366.
63°57' N. 53°18' W.	680 m.	4.35 C. (500 m)	"Godthaab" St. 32.	
68°31' N. 53°57' W.	520 m.	4.79 C. (500 m)		158.
60°22' N. 17°27' W.	120 m.	5.76 C. (110 m)		188.
61°25' N. 23°05' W.	121 m.	7.46 C. (118 m).	"Dana" St. 3200.	
62°27' N. 4°54' W.	150 m.	4.66 C. (125 m).		5866.
61°30' N. 8°52' W.	515 m.	8.13 C. (500 m)		5950.
			Trondheim Fjord.	
			East Greenland (Turner Sound, Cape Tobin, Miki's Fjord).	6-130 m.
			West Greenland (Bredefjord).	

Remarks: The material consists of about 150 specimens and comprises both colonies with anthocodiae almost devoid of sclerites, colonies with strongly spiculated anthocodiae, and all possible transitional forms. The material confirms thus Broch's and JUNGERSSEN's view when they united into one all the species described under the generic name *Dana* by KOREN & DANIELSSEN (1883) and DANIELSSEN (1887). Attention may also be called to the somewhat diverging form described above p. 28, which has unusually large anthocodial sclerites (fig. 21 *g, h*).

Distribution: In the Norwegian Sea the species has been found along the Scandinavian coast from the latitude of Bergen as far as Kola Fjord, at Spitzbergen, and in East Greenland as far north as Scoresby-Sound. It is very common on the Wyville Thomson Ridge s. l. and is also known from several localities in Baffin Bay and Davis Strait. In the eastern part of the North-Atlantic Ocean the species has been found as far south as off the Irish coast, in the western part as far south as about 41° N.

The bathymetrical distribution of the species ranges from 6 m (at East Greenland) to 1505 m, southwest of Iceland; it is, however, mainly found at depths greater than about 200 m.

Vide further JUNGERSSEN (1915, 1916, 1917) and Broch (1928), chart of distribution by Broch (1928 p. 13, fig. 7).

Ordo: **Gorgonacea** Verrill 1865.

Subordo: **SCLERAXONIA** Studer 1887

Diagnosis: Vide KIKKENDALL 1921, p. 8.

Family: **Paragorgiidae** Aurivillius 1931

Diagnosis: Vide AURIVILLIUS 1931, p. 10.

Genus: **Paragorgia** Milne Edwards 1857.

Paragorgia arborea (Linné 1758).

- A. *arborea* (Linné 1758), p. 809.
Verrill 1865, Broch 1912 c, p. 10-13, figs. 4-10.
Jagerskiöld, 1917, p. 19-22.
Kjellmån, 1919, p. 77-80, fig. 31.
Verrill 1922, p. 17-18, textfig. 1, pl. 13, figs. 1-4.
Kjellmån, 1924, p. 28-29.
Norgren, 1930, p. 1-2, fig. 1.
Broch 1935, p. 20.
Deichmann, 1936, p. 81-82.
Pax, 1936, p. 254-256, figs. 177, 178.
Stasny, 1937, p. 71-78, textfig. Z, pl. 1, figs. 31-33.
Verseveldt, 1939, p. 20-32, figs. 5-10.
Carlgren, MS.
For further references vide Broch (1912 c), JAGERSKJÖLD 1917, and VERSEVELDT 1939.
- B. *arborea* Verrill 1865.

Material:

The Norwegian coast (Finnmarken, Lofoten, Trondheim Fjord) 90-200 m. Several colonies.
60° 51' N., 14° 02' E., 125 m., 5 F.C. "Michael Sars" St. 16, 3 colonies.
61° 29' N., 6° 22' W., 282 m., "Dana" St. 5702, 1 colony.

Remarks: This species was not taken by the "Ingolf" Expedition, and in the eastern part of the North-Atlantic Ocean and adjacent seas it has not previously been recorded far away from the Norwegian coast. The above-mentioned find made by the "Dana" in 1938 at a locality in the Faroe Sea somewhat east of Sudero represents in this area the hitherto westernmost find of *Paragorgia arborea* and adds the species to the fauna of the Faroes.

Distribution: The species is at present known to occur along the Scandinavian coast from Hardangerfjord to Finnmarken and as far west as the Faroes, off the Portuguese coast (one locality), in the Barents Sea, off the American North-Atlantic coast on the banks of New-Foundland and Nova Scotia, and further in the North Pacific.

Vide further JAGERSKJÖLD 1917 and BROCH 1935 (chart of distribution, Broch 1935, p. 10-11).

Bathymetrical distribution: About 75-800 m.

Family: **Semperinidae** Aurivillius 1931.

Diagnosis: Vide AURIVILLIUS 1931, p. 10.

Genus: **Anthothela** Verrill 1879.

Anthothela grandiflora (Milne Edwards 1857).

Anthothela grandiflora (Milne Edwards 1857).

- Milne Edwards 1857, p. 63-75, pl. 10, figs. 10-12.
Verrill 1865, Broch 1912 c, p. 5-9, figs. 1-3.
Jagerskiöld 1918, p. 6-8, fig. 1.
Verrill 1922, p. 13-14, fig. 17.
Verrill 1922 c, p. 18-19, textfig. 1, pl. 6, figs. 1-4.
Kjellmån 1924, p. 15-16, figs. 13-11.
Verrill 1927, p. 16-18, pl. 1, fig. 29.
Kjellmån 1929, p. 16, fig. 28.

Anthothela grandiflora Pax, 1936, p. 254, figs. 175-176.

Deichmann, 1936, p. 78-79.

Stasny, 1937, p. 20-23, textfigs. F1, F2, pl. 1, figs. 6, 7.

Jagerskiöld, 1939, p. 21.

Verseveldt, 1939, p. 37-47, figs. 13-15.

Carlgren, MS.

For further references vide Broch 1912 c.

Description: Vide BROCH 1912 c, VERSEVELDT 1939.

Material:

Finnmarken (northern Norway). (Cotype ?) Fragment.
Trondheim Fjord (Roberg, Skarnsund, Brettingnes), 150-300 m, several colonies.

63°15' N, 15°07' W, 1197 m, 3 08°C, "Ingolf" St. 53, 1(2) colonies.
61°11' N, 27°00' W, 913 m, 6 1°C, "Ingolf" St. 81, Fragments.
63°12' N, 20°06' W, 510 m, "Thor" St. 168 (03), 1 fragment.

Remarks: The material consists of upright branched colonies, of which one from the Trondheim Fjord measures 22 cm, and of some membranous specimens from the same locality.

The material from the "Ingolf" St. 81 differs somewhat from the usual type. It consists of four tiny fragments measuring from about 1.2 cm. The fragments are slightly branched and have a diameter of about 0.75-1 mm. The colony to which they have belonged may have had a much slender habitus than the usual colonies of *Anthothela grandiflora*, even more slender than in Sars' original specimens from Finnmarken. The verrucae measure about 1.2 mm in height. The anthocodial part of the zooid body in the largest expanded zooids measures about 5 mm. The sclerites of the specimen are of the same type as in those from the Trondheim fjord (vide Broch 1912 c) only they are not so crowded in the coenenchyma.

Distribution: The species is known from the Scandinavian

coast from Finnmarken in the north to the Skagerrak in the south, where it has been recorded by JÄGERSKIÖLD (1939) from 58°31' N, 10°05' E, 510 m. Also MORIUS (1873, p. 119) mentions *Braconium grandiflorum* from the Skagerrak (off Arendal 75 m), but this record is somewhat doubtful as the *B. grandiflorum*, which MORIUS (1874, p. 260) records from East Greenland, by a re-examination by JUNGEBSEN (1915, p. 1156, 1916, p. 195) was recognized as a specimen of *Eumphylla (Capnella) glomerata*. JUNGEBSEN, however, did not make any reference to *E. glomerata* from the Skagerrak, and it therefore is possible that MORIUS' *B. grandiflorum* from the Skagerrak actually was correctly identified. In Scandinavian waters *Anthothela grandiflora* is found at depths from about (75)150-600 m. It has further been found in the waters south and southwest of Iceland in depths from about 500-1500 m, and along the American coast from off Martha's Vineyard in the south to off Nova Scotia in the north, and J. A. THOMSON in 1927 records the species from off Madeira, the Azores and the Cape Verde in depths from about 1100-1700 m. Chart of distribution, fig. 32.

Bathymetrical distribution: About (75)150-1700 m.

Subordo: **HOLAXONIA** Studer 1887

Diagnosis: Vide KÜENTHAL 1924, p. 237.

Family: **Acanthogorgiidae** Kükenthal & Gorzawsky 1908.

Diagnosis: Vide ACRIVILLIUS 1931, p. 38-39.

Genus: **Acanthogorgia** Gray 1857.

Diagnosis: Vide KÜENTHAL 1929, p. 239.

Acanthogorgia armata Verrill 1878.

- Acanthogorgia armata* Verrill, 1878, p. 376.
Verrill, 1882, p. 361.
Verrill, 1883, p. 31-33, pl. 3 figs. 1, 1 a, 1 b, 2, 2 a, 2 b.
Verrill, 1884, p. 220.
verrilli Studer, 1901, p. 14-15, pl. 7 figs. 1-6.
? *armata* Hickson, 1905, p. 225.
? *armata* Nutting, 1910, p. 13-14, pl. 19 fig. 1.
? J. St. Thomson, 1911, p. 880.
? Kükenthal, 1924, p. 249-250.
? J. A. Thomson, 1927, p. 37-38, pl. 1 fig. 12.
? Deichmann, 1936, p. 119-150, pl. 16 figs. 1-4.
? Stiasny, 1939, p. 137-138, pl. 4 fig. 7, pl. 5 fig. 10.
? 1940, p. 21-22.
? 1942, p. 37-38.
? 1943, p. 129.

Material:

- Off Georges Bank or a neighbouring locality, in great depth, 1 colony. (This specimen was given to the Zoological Museum of Copenhagen 1880 by the Danish Lieutenant Commander C. F. Wandel who during the summer took part in the cruises of the "Blake", in charge of A. Agassiz.)
61°14' N, 30°29' W, 2137 m, 3 0°C, "Ingolf" St. 18, 3 fragments of axis.
62°58' N, 23°28' W, 915 m, 5 5°C, "Ingolf" St. 73, Axis-fragment.

61°11' N, 27°00' W, 913 m, 6 1°C, "Ingolf" St. 81, 3 colonies.
62°57' N, 18°58' W, 957 m, "Thor" St. 166 (03), 6 colonies.
63°05' N, 20°07' W, 557 m, "Thor" St. 167 (03), 3 colonies.

Diagnosis: Colony richly branched, shrub-like or bush-like, often in one plane, with brown, horny basal disk and axis. Zooids cylindrical, slightly widened at the corona, size of verrucae generally 6-7 × 1-1.2 mm, placed at right angles and distributed on all sides all over the colony, usually fairly closely set and much crowded at the branch tips. In zooid body eight interseptal, double rows of crossing, peculiarly bent, spinous spindles, arranged on chevron, average length 0.7-0.8 mm, maximum length about 1.5 mm. Neck region of zooid not very distinct. Coronal sclerites arranged on chevron, slender, angularly bent, with a short, much warty ray and a long, projecting, smooth spine, average size about 1.6 mm, maximum size about 2.4 mm. In the lower part of tentacle curved spindles arranged on chevron, in the upper part small, flattened rods, transversally arranged. Sclerites of coenenchyma much varying, mostly bi- or quadrangulate, decreasing in size from the smaller branches towards the base where small, lumpy sclerites may be the most common.

Description: The specimens vary in height from 6 to 17 cm and are often somewhat broader than high. They are mostly richly branched, several of them in only one plane, but most of them more irregular. Often there is no distinct main stem, and from the base more stems may rise. The branching is nearly right-angled, the branches, however, soon curving and becoming parallel to the stem or branch from which they issue.

The colonies have been attached to solid objects. The diameter of the stem at the base is in a 8 cm high colony about 1.2 mm, in a 15 cm high colony 3 mm, and in the largest specimen, a 17 cm high one, about 4 mm.

The bare axis fragments from the "Ingolf" St. 18 and St. 73 may have been of colonies about 10 cm high.

width of 2 to 187 micradial. The coronal sclerites are a bright yellowish color (fig. 23). The sides of the stems and the tentacles are much crowded, the spines between them. In a single



Fig. 23. *Acanthopagurus armatus* Verrill. *c-g*, coronal sclerites, *c*, of about average size, *b*, sclerites of zoid body in natural position, arrow indicates the attachment of the mesenteries.

zoid the zooids are likewise crowded from the base to the tip of the colony.

The stems are cylindrical, slightly widened at the corona, and the contracted tentacles generally 6-7 mm in length (0.25-0.3 mm width, their appearance then as shown in VERRILL (1883, p. 32, figs. 1, 1a). They may measure up to 10 mm in length in one colony, all the zooids are somewhat shorter than the stems, measuring about 3-4-1.5 mm, and in appearance resembling the zooids figured by VERRILL in pl. 3

of his paper. In the stems the sclerites are long, slender, spinous, and curved, and appear, which typically are placed crosswise to the stem, that their straight part is placed in a longitudinal position pointing towards the apex of the zooid. The straight part crosses over and ends in the nearest interseptal row (fig. 23*h*). The sclerites in the stems are much closer and more transversally placed than the warty one, the differing forms of sclerites being due to the different degrees of contraction. The stems are on an average 0.7-0.8 mm in diameter, the coronal sclerites at the middle, and coronal sclerites of the Verrill (1883, p. 33). The largest measured about 1.5 mm, and the smallest, in a distinct neck region, measuring about 0.2 mm. The coronal sclerites may be some times transversally arranged

The corona is formed by some long, projecting coronal sclerites in each interseptal row below the tentacles, arranged in chevron as those of the main zooid body and generally bent in a similar manner with the one ray strongly warty and the other, much longer and projecting one, the so-called spine, generally smooth in its whole length (fig. 23*e-g*). The lower half or more of the spine of some of the coronal sclerites may be granulated (fig. 23*d*), but is never coarsely warty as the short basal ray. The largest measured coronal sclerites had a length of about 2.1 mm, the maximum length is, however, seldom more than about 1.9 mm, and in most zooids the largest coronal sclerites are about 1.6 mm which corresponds to the largest size stated by VERRILL (1883, p. 32). In DEICHMANN'S statement (1936, p. 150) that "The marginal spicules may be as much as 1.1 mm in length" 1.1 mm is probably a misprint for 2.1 mm. The long coronal sclerites generally number four to six in each interseptal row and generally a few much shorter and often straight ones may be found besides (fig. 23*a-c*), and similar ones occur also at the base of the tentacles.

The tentacles are rather short with only half a dozen pairs of pinnulae and by contraction bent down over the peristome. Their dorsum is strongly armed, in the lower part with curved, spinous spindles (fig. 24*a, b*) arranged in chevron, maximum size about 0.4 mm, and in the upper part with numerous, mostly transversally placed, flattened rods (fig. 24*g-i*) measuring from 0.1-0.02 to 0.3-0.05 mm.

Sclerites were noticed neither in the pinnulae nor in the peristome or stomodaenum.

The sclerites of the coenenchyma are of several types and vary much both in the different parts of the colony and also in the different specimens. The largest sclerites are found in the smallest branches and in the branch tips and, generally speaking, the sclerites gradually decrease in size towards the base where the smallest sclerites of the colony are to be found.

In the smaller branches the sclerites are more or less bent spindles resembling those of the zooid body and of the same maximum length, but often of a somewhat larger average size (about 1 mm) (fig. 25*a*). Besides, in some colonies as e. g. that from off Georges Bank, triradiate sclerites of nearly corresponding size are just as common (fig. 25*b*). In the same colony, which is about 9 cm high and 12 cm broad, the most common sclerites in the main branches are tri- or quadriradiate, measuring up to 0.4 mm in diameter (fig. 25*c, d*). Straight or curved spindles are, however, also fairly numerous, and these may measure up to 0.8-0.05 mm and are either scarcely warty in their whole length or have a strongly warty, and a smooth part, and thus much resemble the coronal sclerites though being of a considerably smaller size and also having the warty ray much more pointed (fig. 25*f, g*). Besides the said sclerites also some coarse and blunt rods of fairly

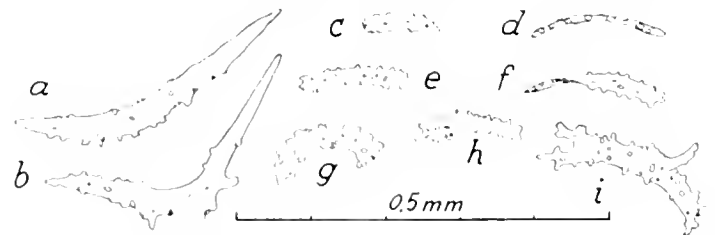


Fig. 24. *Acanthopagurus armatus* Verrill. *a, b*, sclerites of tentacle base, *c, d*, sclerites of outer part of tentacle, *d*, shown from the side.

large size (0.8-0.11 mm) may occur (fig. 25*e*). In the coenenchyma of the base of this specimen tri- and quadriradiate sclerites are likewise the most common, measuring about 0.25 mm in diameter, but here also small lumpy sclerites are found in large number.

The sclerites of the coenenchyma from corresponding parts

of the other specimens seem, generally speaking, to be somewhat smaller than in the above-mentioned colony, and are in some specimens more variable.

The sclerites of the branches in a colony from the "Thor" St. 167, 13 cm high, are mostly slender tri- or quadriradiate ones

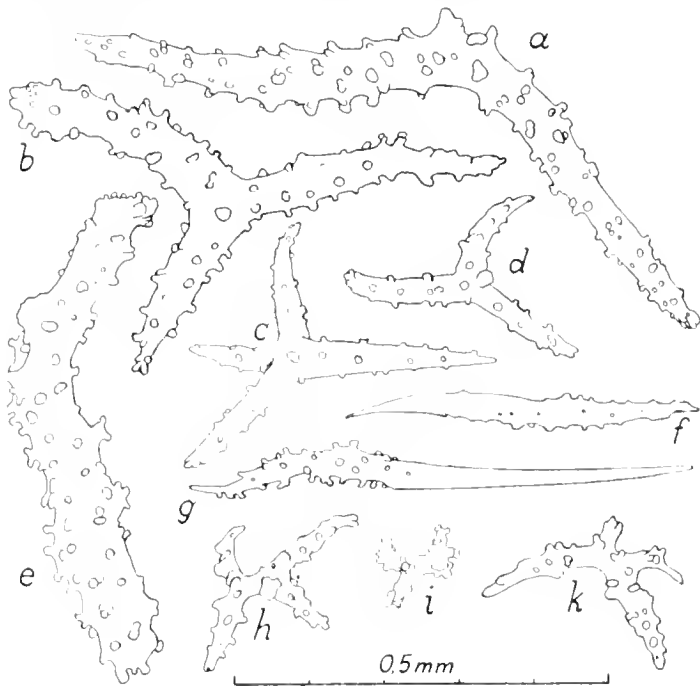


Fig. 25. *Acanthogorgia armata* Verrill. Specimen taken by the "Blake" Expedition off Georges Bank or a neighbouring locality. *a, b, d*- and tri-radiate sclerites from one of the smaller branches, both types about equally common. *c-g*, sclerites from one of the main branches (stems) in the colony. *c, d*, tri- and tetra-radiate sclerites which are the most common. *e*, coarse spindle, a rather rare form. *f, g*, spindles occurring in fairly large number. *h-k*, sclerites from the base.

(fig. 26 *a, b*), but besides several larger and more coarsely built ones (fig. 26 *c*) occur. In the same specimen the sclerites of the base are mostly some lumpy ones (fig. 26 *m, q*) less than 0.2 mm in size, whereas only rather few and small tri- and quadriradiate sclerites are found, some of which, however, are almost six-radiate due to strongly projecting processes, which are nearly equal the rays in length (fig. 26 *l*). Also spindles of about 0.3 mm in length occur, but they are only few.

Spindles occur everywhere in the coenenchyma, though in a very varying number. They are e. g. fairly common in the stem of a 6 cm high colony from the "Ingolf" St. 81 and here they are either long, very slender ones (fig. 26 *d*) measuring up to 1.2 · 0.05 mm, and often being angularly bent, or smaller crescent-shaped ones (fig. 26 *h, k*) measuring up to 0.1 · 0.06 mm. In the same stem the triradiate sclerites were only small ones, less than 0.2 mm in diameter and often irregularly shaped (fig. 26 *e, q*), and some very coarsely built, not branched sclerites of varying shape were also found rather common (fig. 26 *i*).

Discussion: There can be no doubt that the above-described specimens, in spite of some variation in the sclerites of the coenenchyma, belong to one species, viz. *Acanthogorgia armata* Verrill, to which species, as shown by DEICHMANN in 1936, STÜDER's *Acanthogorgia verrilli* is also referable.

Under the name *A. armata* some specimens have also been

recorded from the Cape of Good Hope by HICKSON (1905) and from the coast of Sumatra by NETTING (1910) respectively, the identities of which, however, are doubted by KÜRENTHAL (1924). Specimens of the first form (South Africa, 300-575 m) have been examined by STIASNY, who (1910), with a query, records them as *A. armata*. STIASNY has further (1912), likewise with a query, recorded *A. armata* from the Mediterranean, the bay of Capri and (1939 *b*), without a query, from the coast of Morocco, 20-160 m. Unfortunately, however, the author has had access neither to the paper by HICKSON nor to those by STIASNY (1939 *b*, 1910) and thus is not able to discuss the statements made in these papers.

As regards NETTING's specimen it is difficult from the description to ascertain whether he has dealt with a previously known species or a new one. The present author found it possible that NETTING's material would prove to belong to *A. doylei* Kükenthal & Gorzawsky 1908. STIASNY, however, announces in his paper of 1913 that in his revision of the Acanthogorgiidae of the Siboga-Expedition he will describe the specimen as a new species under the name *Acanthogorgia paramata*, and in the same paper STIASNY designates *A. doylei* as an uncertain species.

Distribution: *Acanthogorgia armata* is known from American waters from off New Foundland in the north to off Martha's Vineyard in the south, in depths from 275 to 1267 m. The species is also by J. A. THOMSON (1927) recorded from off the Azores and from the Portuguese coast (off Cape Mondega) in depths from 550 to 1250 m, and by STIASNY (1939 *b*) from off the Moroccan coast in depths from 20 to 160 m. The finds made by the "Ingolf" and the "Thor" were made south and south-west of Iceland in depths from 557 to 957 (2137) m. The latter depth parenthetically since here only a dead colony was secured. Chart of distribution, fig. 20.

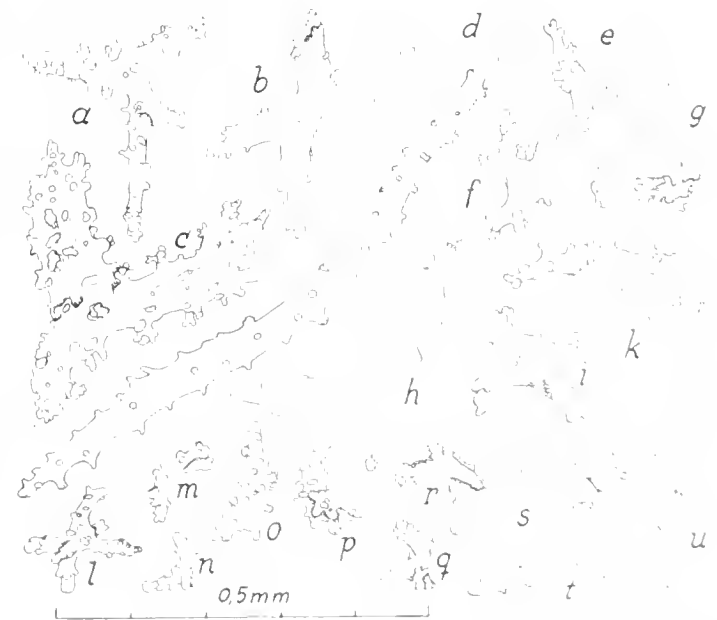


Fig. 26. *Acanthogorgia armata* Verrill. *a, c*, sclerites from a branch of a colony taken by the "Thor", St. 167, *a, b*, of the usual type, *c*, the more rare form, *d, k*, sclerites from one of the main branches (stems) in a colony taken by the "Ingolf" St. 81, *e, q*, the most common type, *d*, one of the larger spindles, *h, k*, crescent-shaped spindles which are fairly common, *i*, a lumpy sclerite, *m, o*, sclerites of the base in the same colony, *m, p*, representing the most common type, *l*, one of the rather rare spindles.

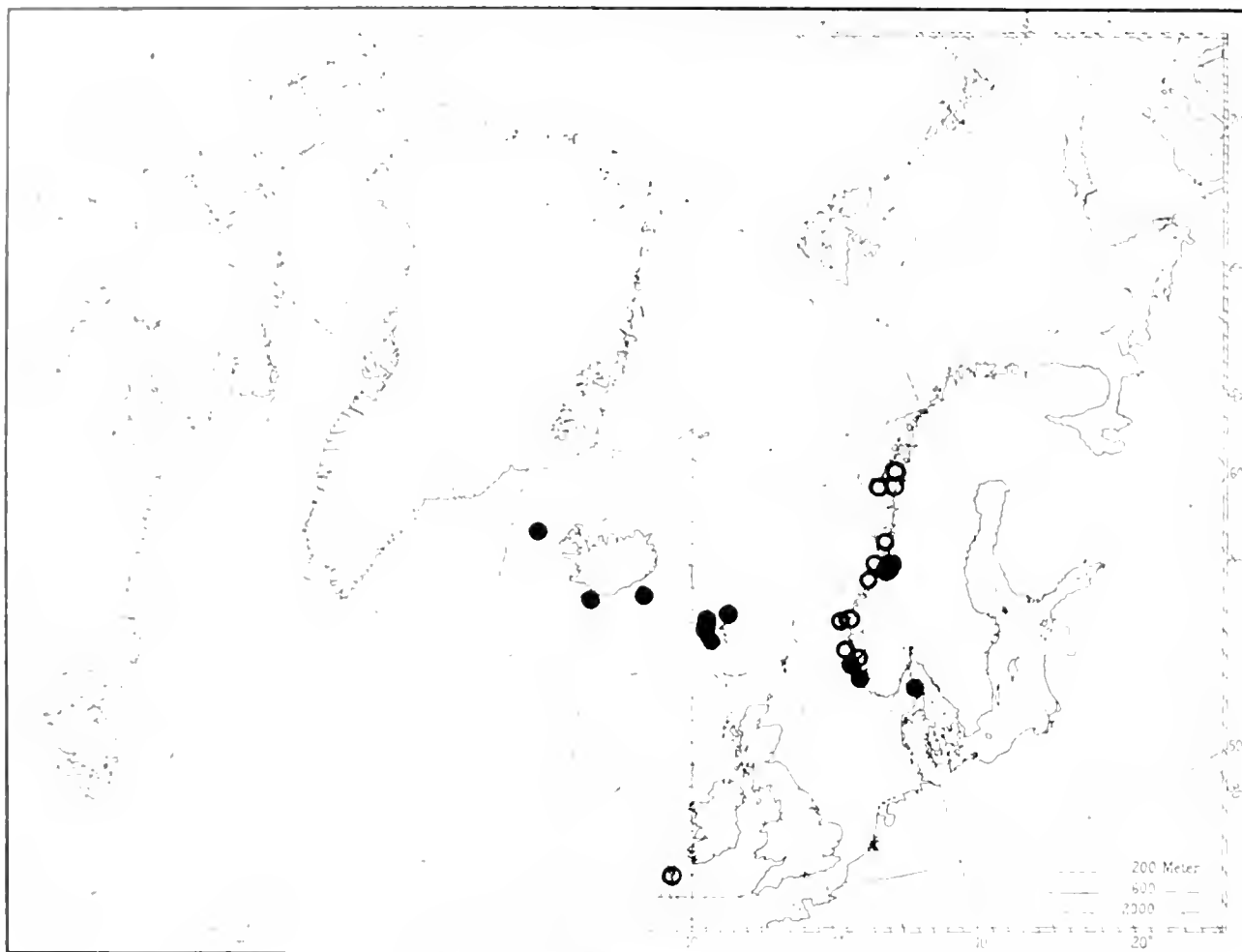


FIG. 20. *Paramuricea placomus* (Lac.)

Family Muriceidae Gray 1859 emended

Diagnosis. Vide ABRVILLIUS 1931, p. 94.

Genus. Paramuricea Kolliker 1865,

Opusc. Ichth. Scand., vol. 1, p. 456.

Paramuricea placomus (Lacépede 1758)

Lacépede, 1758, p. 892.

Kolliker, *Walden's Reise*, 1853, p. 13, pl. 9.

Brady, 1912, p. 17, 20, figs. 9, 10.

Brady, 1912, p. 21, 26, figs. 11, 11.

Brady, *Atl. Zoog.*, 1917, p. 27, 31.

F. A. Davis, 1927, p. 39, pl. 1, fig. 2.

Nordenskiöld, 1910, p. 2.

Arvillius, 1931, p. 166, 170, fig. 32.

Brady, 1936, p. 135, 136, pl. 11, figs. 11, 12.

Brady, 1936, p. 257, 12, 180.

Brady, *Kolliker*, 1939, p. 1, 5.

Brady, MS.

Brady, *Abrvillius*, 1931.

Brady, 1932, 300, *Arvillius*, 1931.

61 32 N, 7 55 W, 352 m, "St. 1," 1 fragmentary

colony.

63 33 N, 15 02 W, 595 m, 5 9 C, "Ingolf" St. 55, 10 colonies.

63 12.5 N, 20 06 W, 510 m, "Thor" St. 168 (03), 1 colony.

Skagerrak (29 miles N, $\frac{5}{8}$ W, of the Skagen light-ship) 317 m, "Thor" St. 297 (01), 1 colony.

62 44 N, 6 06 W, 330 m, "Dana" St. 58-10, 1 fragmentary colony.

62 07 N, 8 35 W, 375 m, 6 58 C, (325 m) "Dana" St. 6007, 2 fragmentary colonies.

61 39 N, 7 55 W, 352 m, "Dana St. 6008, 1 fragmentary colony.

Besides, the collection of the Zoological Museum of Copenhagen includes numerous colonies from Norway (Hvidingsu, Bergen, several localities in the Trondheim Fjord and many without exact locality).

Remarks. The specimens of *P. placomus* kept in the Zoological Museum of Copenhagen are generally branched mainly in one plane, but there are some which are one-sided bush-like branched. Anastomoses between the branches are rather frequent, especially in the large colonies.

The specimens taken in the North Atlantic are only small, several of the specimens from the Norwegian coast, however, measure 50 cm or more in height, and one specimen, from very old time kept in the zoological collections of Copenhagen, measures about 90 cm in height and about 110 cm in width.

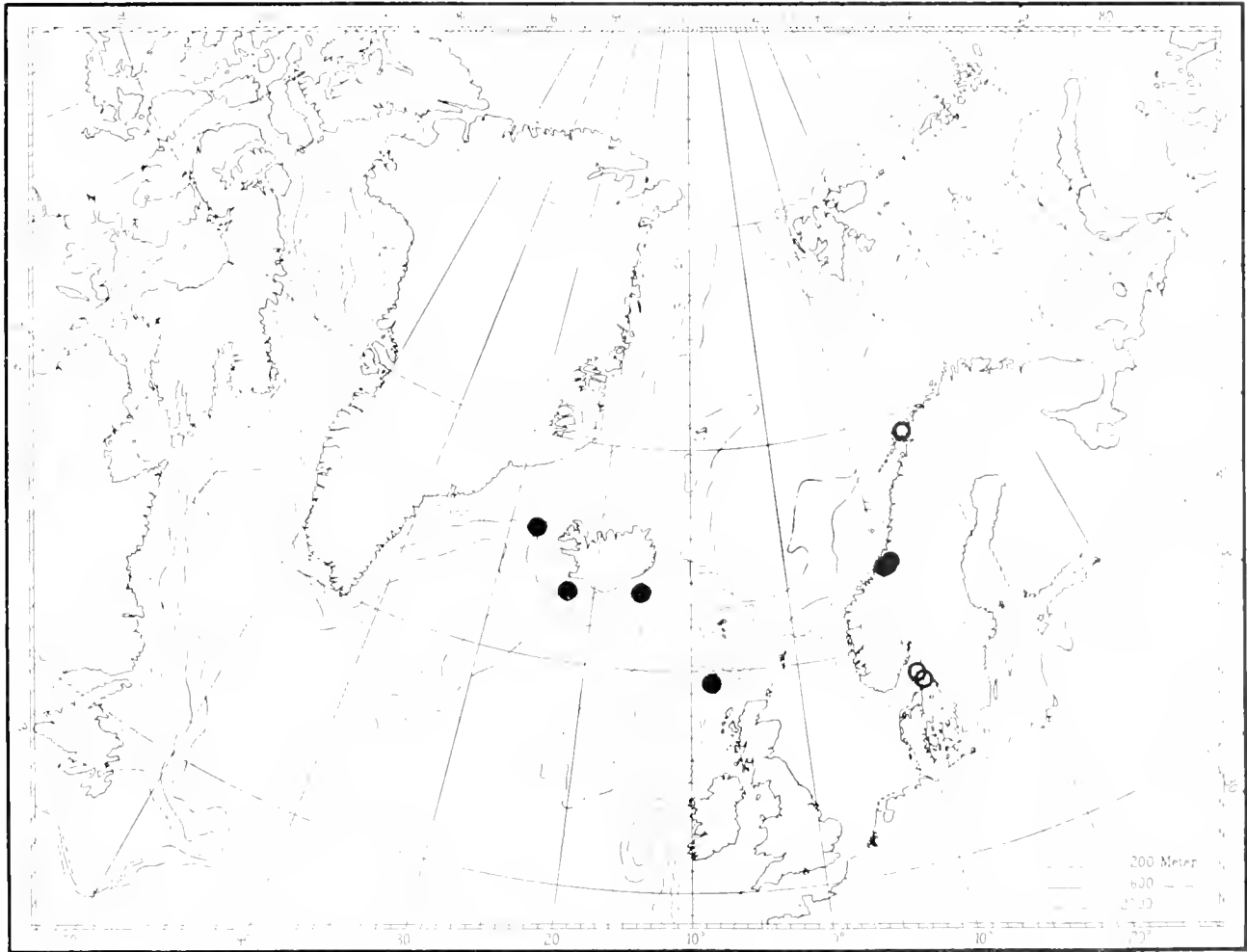


Fig. 28. *Trachymuricea kükenthali* (Broch).

On the authority of BROCH (1935 p. 11, footnote), *Cl. stormi* KOREN & DANIELSSON is included among the synonyms of *P. placomus*, and it appears from the original description and BROCH's redescription of this form that BROCH's point of view in 1935 without doubt is correct. A quite analogous membranous form of *Trachymuricea kükenthali* will be described below.

JUNGERSEN (1917 p. 29) records *P. placomus* from the "Michael Sars" St. 76 a, b, c and St. 85. However, three of the specimens from the "Michael Sars" St. 76 kept in the Zoological Museum of Copenhagen have proved to belong to *Trachymuricea kükenthali* (Broch) and were also by JUNGERSEN labelled as *P. placomus* f. *kükenthali*. On the other hand, the specimen which JUNGERSEN mentions from the Skagerrak ("Thor" St. 297) is a distinct *P. placomus*.

Distribution: The species is with certainty known to occur along the Scandinavian coast from Lofoten in the north (Bodo) to the Skagerrak in the south. It occurs further in the North-Atlantic Ocean on the Wyville Thomson ridge from the Faroes, south of Iceland to Denmark Strait and on the eastern coast of North America (off New England). The species is also recorded from south of Ireland, the west coast of France, the Cape Verde, and the Mediterranean. It seems, however, to be open to doubt whether the last-mentioned specimens actually belong to the North-Atlantic *P. placomus* (cf. AUVILLIUS 1931 p. 170, and STIASNY 1912 p. 26-27). In the northern part of the Atlantic Ocean and along the Norwegian coast *P. placomus*, however, is a common species. Chart of distribution, fig. 27.

Vide further JUNGERSEN 1917.

Bathymetrical distribution: From about 150 m (75 m) to about 1600 m.

Genus: *Trachymuricea* Deichmann 1936.

Diagnosis. Vide DEICHMANN 1936, p. 132.

***Trachymuricea kükenthali* (Broch 1912)**

Paramuricea kükenthali Broch, 1912 c, p. 26-31, figs. 15-20.

Arndt, 1912, p. 125.

Broch, 1913 a, p. 181.

placomus (pars) Jungersten, 1917, p. 27-31.

Muriceoides kükenthali Auvillius, 1931, p. 175-180, fig. 34.

Paramuricea kükenthali Dons, 1932, p. 11.

Jagerskiöld, 1935, p. 15.

1936, p. 18.

Trachymuricea Deichmann, 1936, p. 131.

Muriceoides Pax, 1936, p. 257.

Paramuricea Jagerskiöld, 1937, p. 21.

placomus (pars) Kramp, 1939, p. 1-5.

Muriceoides kükenthali Carlgren, MS.

Description: Vide BROCH 1912 c and AUVILLIUS 1931.

Material.

65° 13' N, 26° 58' W, 171 m, 6-1°C, "Ingolf" St. 16, 1 fragment.

63° 33' N, 15° 02' W, 595 m, 5-9°C, "Ingolf" St. 55, 1 fragment.

Wright & DeKay (1845, p. 80), St. 76, 3 colonies, measuring 0.17-0.20 as *P. placans*, with a colony from St. 171 (1956) 2 colonies, measuring 0.18-0.20. BROEN (1913, p. 159) 300 m.

Specimens were first described by BROEN (1912) on the basis of material from the Trondheim Fjord, but were by JENSEN (1935) referred to *Placans* with *Paramuricea placans* (L.), 200 fathoms, 2 specimens. ACRIVILLIUS (1931) likewise on specimens from the Trondheim Fjord referred to the genus *Muriceodes* Wright & DeKay, and in 1936 established a new genus to include among others *T. kükenthalii* (Broch).

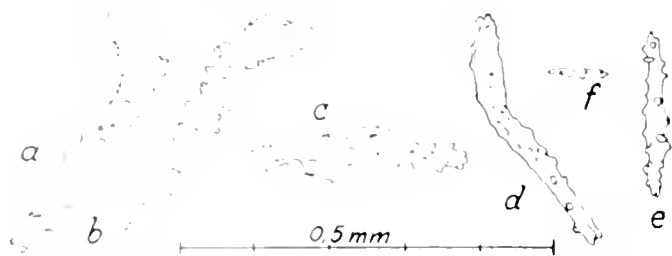


FIG. 29. *Trachyphela kükenthalii* (Broch). a, b, sclerites from anthodia neck, lower part of anthodia neck of zooid; c, sclerite from distal end of tentacle; d, sclerite from outer part of tentacle; e, pinnula sclerite.

As a result of the authors' studies on the several specimens in the Zoological Museum of Copenhagen which may be referred to *Trachyphela*, agrees in all essentials with ACRIVILLIUS' exhaustive treatment of the species (ACRIVILLIUS 1931), and a closer examination of the material has therefore not been found worth

presenting in this paper (1917), without any explanation, to *T. kükenthalii*, among the synonyms to *P. placans*; on the other hand, most of the specimens he had, however, noted *P. placans* as *T. kükenthalii*. That JENSEN united the two species (1935) is, though apparent, he did not intend entirely to support *T. kükenthalii*, was no doubt on account of the surprising similarity in appearance and spiculation which is met with in *P. placans*, and another fact which also may account for his view on the species is that the two forms in the material he had, found together, thus both species are common in the Trondheim Fjord, and of the three stations (St. 171, 1956) where *P. placans* two also gave *T. kükenthalii*.

Specimens taken by the "Ingolf" and "Thor" are small fragments, the former only fragments. The material from the "Michael Sars" on the other hand, consists of three specimens, fragmentary colonies, measuring from about 0.17-0.20 mm.

Material from the type locality (the Trondheim Fjord) consists of rather well developed, branched and not always

distinctly in one plane, those taken by the "Michael Sars" are only very sparsely branched, consisting of an upright stem (up to 4 mm in diameter) with a very few (e. g. four) side-branches distinctly arranged in one plane and likewise only slightly or not at all branched. Anastomoses were found in one colony. The specimens thus differ somewhat in their main habitus from the typical, it may also be noted that their coenenchyma is fairly thick and that the verrucae are only very slightly protruding.

MOLANDER (1929, p. 3) supposes that *Trachyphela rudis* Verrill 1922 is a *Paramuricea*, probably synonymous with *P. kükenthalii*, but an identity with the latter species is, in spite of some similarities, not present (see further the discussion on *Clavularia gracilis* n. sp.).

The Zoological Museum of Copenhagen has, however, a small, quite membranous colony from the Trondheim Fjord which without doubt is a young specimen of *T. kükenthalii*.

The colony in question consists of a membranous stolon expanded on a fragment of *Lophelia* and measures about 15 x 6 mm. From the base about 18 closely placed zooids arise.

The spiculation of the specimen agrees with that of typical upright and branched specimens. The sclerites of the anthostele (verrucae in ACRIVILLIUS) are mainly longitudinally arranged, warty spindles, most of them provided with a distinctly out-turned upper end (fig. 29 a, b) and on average measuring about 0.4 mm.

In the lower part of the anthocodia (polypal neck in ACRIVILLIUS) the sclerites are scattered, transversally arranged, warty spindles with blunt ends, measuring about 0.35 mm (fig. 29 c). In the upper part of the anthocodia the sclerites are arranged en chevron in eight rows which continue uninterruptedly in the rows of sclerites in the tentacle dorsa.

The sclerites in the anthocodial crown and in the lower part of the tentacles are bent, somewhat club-shaped spindles (fig. 29 d) with the upper and shorter ray generally provided with ridges. The largest of these sclerites measure about 0.5 mm, and the average size is about 0.35 mm. The upper ray of these sclerites are longitudinally arranged in the tentacles along the middle of the dorsum as distinct from the arrangement of the sclerites in the tentacles of the very similar form described in this paper as *Clavularia gracilis* n. sp.

The outer part of the tentacles are provided with a broad row of longitudinally arranged, warty rods about 0.2 mm in length (fig. 29 e). In each pinnula there are a few small sclerites (fig. 29 f).

Distribution: Besides from the type locality, the Trondheim Fjord, the species is recorded from a locality just north of Lofoten (Broen 1913) and from some places in the Skagerrak at about 200 m depth (JENSEN 1935, 1936, 1937). The localities of the specimens kept in the Zoological Museum of Copenhagen are all just south of the Wyville Thomson Ridge, from the Faroes to Denmark Strait, and probably the species may prove to be common in the whole North-Atlantic Ocean. Chart of distribution, fig. 28.

Bathymetrical distribution: From about 150 m in the Trondheim Fjord to about 1300 m south of the Faroe Bank.

Family Primnoidae Verrill 1866, emended 1883

COLEMAN, 1919, p. 309.

The family Primnoidae (L. SÄNDER in 1887) was first defined by VERRILL (1866) in the Challenger Report, and was later followed by KÜKENBAL (1890) and ACRIVILLIUS (1906), who in 1931 proposed a new subfamily classification, and in 1936 SÄNDER'S classification,

on p. 156-158 however proposed a new subfamily classification, which with the modifications given by ACRIVILLIUS (1931, p. 238) seems to give the best picture of the relationship of the different genera, and which accordingly is followed below.

Only two of the five subfamilies are represented in the North-Atlantic Ocean, each with one genus and one species.

Subfamily: **Primnoinae** Versluys 1906, emended.

Diagnosis: Zooids with operculum. Opercular scales overlapping each other in adaxial direction. Adaxial side of zooids devoid of body scales or only with reduced ones. In the abaxial longitudinal rows 2-7 body scales. Zooid body scales having their outside plentifully set with small and pointed warts, which, however, do not form actual ridges.

Genus: Primnoa Lamouroux 1812.

Diagnosis: Vide KÜKENTHAL 1919, p. 358.

Primnoae resedaeformis (Gunnerus 1763).

Gorgonia resedaeformis Gunnerus, 1763, p. 321-329, pl. 9.

Primnoa resedaeformis Broch, 1912 *e*, p. 32-37, figs. 21-25.

Jungersen, 1917, p. 25-27.

— Kükenthal, 1919, p. 360-362.

— Nordgaard, 1930, p. 2.

Aurivillius, 1931, p. 293-296, fig. 58.

— Broch, 1935, p. 29-33, figs. 17-18.

— Deichmann, 1936, p. 157.

— Pax, 1936, p. 257, figs. 181, 182.

— Kramp, 1939, p. 1.

— Stiasny, 1939 *a*, p. 35.

— Carlgren, MS.

For further references vide JUNGENSEN 1917, KÜKENTHAL 1919 and DEICHMANN 1936.

Description: Vide BROCH 1912 *e* and AURIVILLIUS 1931.

Material:

Off the Scandinavian coast in several localities from the Skagerrak in the south to Bodø in the north. 150-170 m. Many colonies.

S. W. Greenland (Grødefjord, Frederikshaab, Bredefjord, Julianehaab, Hua). 225-510 m. Some colonies.

S. E. Iceland. 225 m. 1 colony.

42° 06' N, 63° 15' W. 1 colony.

62° 01' N, 1° 00' E. 373 m. "Michael Sars" St. 53. 1 colony.

62° 19' N, 8° 51' W. 501 m. 1-21 C. (190 m). "Dana" St. 6005. 1 colony.

62° 07' N, 8° 35' W. 375 m. 6-58 C. (325 m). "Dana" St. 6007. 1 colony.

Pacific coast of Alaska. 1 colony.

Remarks: The author did not succeed in finding any characters in the Pacific specimen of *P. resedaeformis* (without the broken-off base 65 cm in height) which could distinguish this specimen from an Atlantic one; this also corresponds with the view of AURIVILLIUS (1931) and BROCH (1935) that the Pacific specimens of *P. resedaeformis* only on geographical reasons can be separated from the Atlantic as a distinct variety.

Distribution: In the North-Atlantic Ocean the species occurs along the Scandinavian coast from the Skagerrak to Fmmerken (according to BRATTSTRÖM also found in the Varangerfjord, CARLGRÉN MS.), along the coast of North America from George's Bank to Fundy Bay, in the southern part of Davis Strait, and further it has been found in the Barents Sea. In all these localities it has only been taken at moderate depths (95-565 m). In the eastern part of the Atlantic Ocean it has, however, also been recorded from off the Portuguese coast and here at fairly great depths, about 1000 m.

In the northern Pacific Ocean the species is known from several localities both along the American and the Asiatic coast, and is here mainly found at somewhat greater depth than in the Atlantic (335-832 m).

Vide further JUNGENSEN (1917) and BROCH (1935) with chart of distribution p. 40-41.

Subfamily: **Primnoellinae** Versluys 1906, emended.

Diagnosis: Zooids with operculum, opercular scales overlapping each other in adaxial direction. Body scales of adaxial side of zooid sometimes wanting. Generally more than 7 (and not less than 1) zooid body scales in the abaxial longitudinal rows. Outside of body scales either smooth or provided with ridge formations.

According to VERSLUYS the subfamily comprised only the genus *Primnoella*. With the above-mentioned diagnosis the subfamily, however, also includes the genus *Calogorgia* Gray, which was referred to it by AURIVILLIUS in 1931 on account of the close relationship which seems to exist between that genus and *Primnoella*. It may thus be difficult in some cases to ascertain to which genus some of the species should be referred (cf. KÜKENTHAL 1919 p. 183).

Genus: Primnoella Gray 1857.

Diagnosis: Colonies generally unbranched or in some species scarcely branched. The zooids, which usually are placed in whorls, are more or less bilateral-symmetrical and are, besides the operculum, provided with a circum-operculum. Zooid body scales fairly smooth on the outside, warty on the inside. In the stem a layer of scale-like sclerites under which an inner layer of lumpy ones may be found.

Primnoella jungerseni n. sp.

Material:

61° 31' N, 31° 12' W. 2148 m. 1-6 C. "Ingolf" St. 11. 8 colonies.

61° 11' N, 30° 29' W. 2137 m. 3-0 C. "Ingolf" St. 18. 2 colonies.

Diagnosis: Colony unbranched, flexible, flaccid, with disk-like attachment. Zooids bilateral-symmetrical, ascending and placed in whorls, two or three zooids in each whorl. The intervals between the whorls measuring about 2-5 mm. Size of verrucae generally 1.6-1.8 mm. Opercular sclerites and circum-opercular sclerites of about the same type, the former being somewhat smaller than the latter which measure about 0.5 mm. Zooid body scales placed in eight longitudinal rows with about 6-7 scales in the abaxial rows and about 4 scales in the adaxial rows. From the abaxial side of zooids, besides the two abaxial, also the two abaxial lateral rows of sclerites visible. In the stem a single layer of scales (no inner layer of differently formed sclerites).

Type: In the Zoological Museum of Copenhagen.

Description: The largest of the colonies was collected at the "Ingolf" St. 18 and has been selected as the type. It is an entirely intact specimen which measures 91 mm in height and is attached to a small stone by a basal disk of about 1.1-2 mm in diameter. The stem which is very flaccid and highly flexible is in the lower part of the colony about 0.1 (0.5) mm in diameter and is only slightly more slender towards its end, being about

the uppermost whorl. The axis is covered with scales, which are 0.18-0.2 mm in diameter. The intervals are broadest about 3 mm from the base. At regular intervals, the diameter of the intervals is about 0.5 mm. These are those between the

their mouth. The verrucae measure about 1.6-1.8 (2) mm in length by 0.4-0.5 mm in diameter. In the uppermost whorl of zooids the verrucae measure, however, only 1.4 mm in length. The number of zooid body scales in the abaxial rows is usually 6-7, rarely 8.

The other colony from the "Ingolf" St. 18 consists of two fragments which are supposed to be of one set, viz. a 7 mm large, bare axis attached to a stone by a basal disk, 0.8 mm in diameter, and a 49 mm large, fragmentary colony without base. All the 13 remaining whorls of zooids consist of but two zooids. The intervals between the whorls vary between 1.8-3 mm and the size of the verrucae vary between 1.5-1.9 mm. The zooid body scales number 6-7 per abaxial row.

Most of the colonies from the "Ingolf" St. 11 are more or less damaged. The largest specimen is 53 mm in height; its zooids number two or three per whorl, and their verrucae measure about 1.5-1.65 mm. In a section of 28 mm there are 11 whorls, the largest interval between two whorls being 1.5 mm.

One of the other colonies is 49 mm high and has 13 whorls of zooids, the lowermost whorl being placed 8 mm above the base and the intervals between the rest varying from 5-2.2 mm. The number of zooids in the four lowest whorls is 2, in the fifth 3, in the sixth 2 and in the rest of the whorls 3 zooids. The upper zooid whorl is somewhat damaged, the point of axis being bare. The zooid body scales number 6-7-8 and exceptionally 9 per abaxial row.

Two colonies have been attached on a small stone. Of the one only the basal disk is left, of the other 28 mm of the stem with 5 whorls of zooids is left, the lowermost whorl being placed 5.5 mm above the base and the intervals between the others being 5.1, 3.1, 5 and 3.6 mm respectively. The number of zooids is two in the two lowest whorls, three in the rest. The size of the verrucae varies between 1.8-2 mm in height.

A fragmentary colony (38 mm high without base and lower stem) is, besides the type specimen, the only one in which the tip of the colony is intact. The specimen has 13 whorls of zooids left, one of which (the sixth from the top) consists of three zooids whereas in all the other whorls there are but two zooids. The intervals between the whorls vary from 2-2.7 mm; the interval between the uppermost of the whorls but two and the uppermost but one being, however, 1.7 mm and the interval between this whorl and the uppermost being only 1.4 mm. The size of the verrucae varies between 1.55 and 1.9 mm, in the zooids of the uppermost whorl it is, however, only 1.3 mm and in the zooids of the uppermost whorl but one the verrucae measure 1.3 and 0.8 mm respectively.

Of the rest of the colonies from the "Ingolf" St. 11 two measure 44 and 45 mm respectively, whereas the others consist of smaller fragments.

All the colonies are pure white (in alcohol), the colour being due to the sclerites.

The colour of the axis, which in some colonies is laid bare in large sections, is, in the lower part, slightly yellow brown, towards the upper part of the colony it becomes lighter and is here light blue green iridescent. The terete and very flexible axis has in the lower part a width of about 0.12-0.2 mm and tapers gradually towards the tip of the colony. Where in the basal disk it is extended to form the attachment of the colony, it is in one colony pure white, i.e. strongly calcified, whereas in other colonies no such strong calcification has taken place. The width of the stem varies from about 0.5 to 0.25 mm in diameter, and the basal disk which is always roundish, measures from 0.6-1.2 mm in diameter.

At the "Ingolf" St. 18, however, a fairly lobated basal disk, which measures about 1 mm in diameter and about 1 mm in height and which may possibly have belonged to an older colony of *P. purpurina*, was found attached to a stone.

The coenenchyma of the stem and basal disk is fairly thin. The basal disk is covered with a close layer of scale-like and

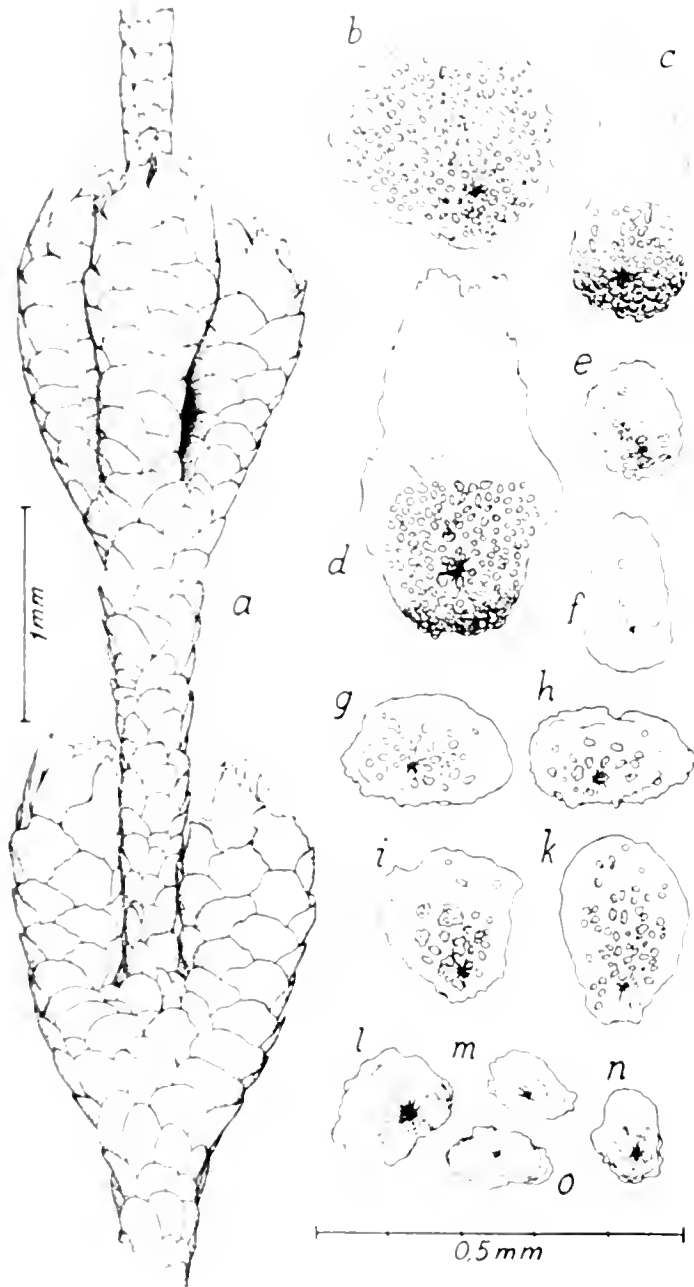


FIG. 1. a) Lateral view of a colony. b) Transverse section of the uppermost whorl. c) Transverse section of the uppermost whorl showing lateral sclerites. d) Transverse section of a whorl showing the arrangement of zooids. e) Transverse section of a whorl. f) Transverse section of a whorl. g) Transverse section of a whorl. h) Transverse section of a whorl. i) Transverse section of a whorl. k) Transverse section of a whorl. l) Transverse section of a whorl. m) Transverse section of a whorl. n) Transverse section of a whorl. o) Transverse section of a whorl.

intervals between the whorls, are about 0.5 mm. The intervals between the uppermost and the next whorl are usually 1.5-2 mm. The whorls are usually placed close to each other, only the uppermost whorl consists of three zooids. The zooids are attached against the stem, the mouth being directed outwards. The coenenchyma is closed over

overlapping sclerites, oval or more rounded in shape (fig. 30 *l*). These scales are usually small, about 0.1–0.15 mm in diameter, the largest having a diameter of about 0.2 mm. They are not warty, but have on their outside more or less marked, concentric ridges which are the impressions of the overlapping scales. Both these and all the other sclerites of the colony are delicately striped from the sclerite-centre ("nucleus") outwards towards the edges.

In the coenenchyma of the stem, one layer of scale-like sclerites is found imbricately covering each other from the bottom upwards. These scales (fig. 30 *e–k*) are thin, with a smooth outside and an inside which is more or less warty, especially in the basal portion, with sometimes fairly large, rugged warts (fig. 30 *i*). The sclerites are usually oval in the longitudinal direction of the stem and with an eccentric placed sclerite-centre (fig. 30 *i, k*). They may, however, also be oval in a direction across of the stem (fig. 30 *g, h*) and have then a more centrally placed sclerite-centre. Their outlines are usually fairly regular; only in their basal part, where they are attached into the coenenchyma, they may be somewhat irregular. Their size usually varies by about 0.2 mm (0.12–0.35 mm).

The zooid whorls consist in some of the colonies of but two ascending zooids placed opposite, usually, however, there are in the colony both whorls with two and three zooids (cf. the above-stated descriptions of the habitus of some of the colonies). The intervals between the whorls (the "internodes") vary somewhat, but are always a little longer than the length of the verrucae (= the contracted zooid). All the zooids are contracted and closely pressed against the stem (fig. 30 *a*). They (the verrucae) measure in length about 1.5–2 mm, usually 1.6–1.8 mm, by 0.4–0.5 mm in width.

The zooids are imbricately covered with eight rows of scales, which are placed with the sclerite-centre off the mesenteries of the zooid. The uppermost, the opercular sclerites, which may be laid down so as to cover the mouth of the zooid, are of a rectangular or pointed triangular shape (fig. 30 *c*, fig. 31 *5*). Their basal part with the sclerite-centre is closely warty on the inside and the basalmost margin is somewhat thicker than the rest of the sclerite; the upper and larger part of the sclerite is smooth also on the inside. The opercular sclerites measure about 0.3–0.15 mm in length, those of the adaxial side of the zooid being the smallest.

The operculum is covered by the eight large marginal sclerites which form a distinctly circum-operculum. The circum-opercular sclerites (fig. 30 *d*, fig. 31 *2, 4, 8*) are of about the same type as the opercular sclerites. Their size varies from about 0.5–0.6 (0.65) mm, those of the adaxial rows usually being the smallest, in any case the most narrow, whereas those from the lateral and abaxial side of the zooid may be fairly broad and with a much more irregular outline (fig. 30 *a*).

When the operculum and circum-operculum are closed above the mouth of the zooid, a pointed cone in which the adaxial sclerites are covered by the abaxial is formed.

The zooid body scales (fig. 30 *b*, fig. 31 *1, 4*) are of a rather uniform appearance, they are roundish with a smooth outside and have nearly their whole inside covered with small warts, only a narrow border along the upper part of the scale being smooth. They measure up to about 0.1 mm, the uppermost scales of the lateral rows usually being the larger. Those of the abaxial rows decrease only little in size from the top of the zooid towards the base, whereas those of the adaxial and adaxial-lateral rows decrease considerably in size; the basalmost scales measuring only about 0.1–0.13 mm.

The zooid body scales usually number 6–7 in the abaxial rows, sometimes, however, there may be 8, and in one zooid 9 scales were counted in the abaxial rows. From the abaxial to the adaxial side of the zooid the number of scales in the longitudinal rows gradually decreases so that in the adaxial rows there are usually about 4 zooid body scales.

The strongly contracted zooids are usually somewhat concave on their adaxial side. In some zooids so concave that a distinct deep furrow is formed in which the zooid body scales of the two adaxial rows are placed with their outside against each other. The zooid which is figured in fig. 31 is one of the least contracted and is only slightly concave on its adaxial side, nor are the circum-operculum and operculum here fully closed above the mouth of the zooid.

There are no deposits in the tentacles. In some of the whorls one of the zooids is considerably smaller (i. e. younger) than the others, measuring but 0.8–1.3 mm in contracted state. In these young zooids the zooid body scales in the abaxial rows number 6–7; and from the full-grown zooids the young ones in habitus differ mainly by their less developed circum-opercular sclerites.

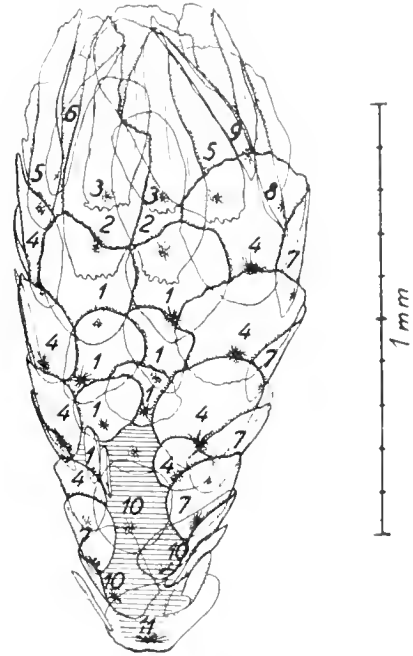


Fig. 31. *Primnoella junipiscus* n. sp.

A zooid made transparent and shown from the adaxial side. The nuclei of the sclerites, but not the warts on the inside, are indicated. The zooid is cut from the stem, by which operation some of the stem scales at the base of the zooid are cut through and the basalmost pair of adaxial scales removed somewhat from their natural position. The hatched area is the place where the cut has been performed.

- | | | |
|-----|--|-----------------------------|
| 1. | zooid body | scales of the adaxial rows. |
| 2. | circum-opercular | - - - |
| 3. | opercular | - - - |
| 4. | zooid body | - - - adaxial-lateral rows. |
| 5. | circum-opercular | - - - |
| 6. | opercular | - - - |
| 7. | zooid body | - - - abaxial-lateral - |
| 8. | circum-opercular | - - - |
| 9. | opercular | - - - |
| 10. | zooid body scales of the abaxial rows. | |
| 11. | the large stem scale direct below the basalmost pair of abaxial body scales. | |

In the two colonies where the tip of the colony is intact the stem is flattened between the uppermost pair of zooids (the uppermost whorls in these colonies consisting of only two zooids placed opposite each other). The stem ends somewhat below the top of the verrucae, and the point of the axis ends a little further below, at about the base of the adaxial side of the zooids. In both these colonies the uppermost zooids are somewhat smaller than the other zooids (cf. the above-stated description). Thus a tip-growth is indicated besides the intercalary growth.

Discussion: The species is referred to the genus *Primnoella* on account of the shape of its scale-like sclerites, its unbranched colony and its whorls of zooids. There are, however, some differ-

of the stem (i.e. the lumpy spicules) (*P. polita* and *P. jamgerseni*) and the absence of lumpy spicules (missing, the stem smooth) (*P. distans*). WRIGHT & STUDER (1889), VERSELYS (1909) and DEICHMANN (1936) regard this as a generic character, but it is not clear from their text whether they believe that all the previously mentioned species have an inner layer of differentiated cells or whether the absence of such a statement is due to an actual absence of that layer. The lumpy spicules in description within the relation to the whorls with and species without an inner layer of the stem are also known.

P. distans differs from most other *Primoella* species in having a low number of zooids in the whorls. No other *Primoella* species has a lower number in which the most common number of zooids in the whorls has been two and the largest number of zooids in the whorls has been three.

In the case of species of zooids in the whorls, the comparatively low number of abaxial zooid body scales, and the absence of lumpy spicules in the stem are all characters which indicate a relationship between *P. jamgerseni* and some of the other described species of *Calopogon*.

Of the two of the other species of *Primoella* may perhaps be compared with *P. jamgerseni*, viz. *P. distans* Studer 1878 and *P. polita* Deichmann 1936. *P. distans* Wright & Studer 1889). The first of these species was described on a specimen, 23 cm high (colony base collected in the South Pacific (22° 21' S, 151° 7.7' E) at a depth of about 1000 m, 550 Fathm, and WRIGHT & STUDER (1889) failed to have recaptured some specimens of this species from the West Atlantic Ocean (off Pernambuco, Brazil, 220-730 m, and the West Indies, about 825 m). KÜKENTHAL (1919) (1941) has not found it, whether this is true, and DEICHMANN (1936) described a new species, *P. polita*, which is identical with *P. distans* Wright & Studer 1889, on colonies from off Frederiksted, West Indies, at a depth of about 920 m.

The two species, *P. distans* Studer 1878 (p. 644 pl. 1 fig. 9), *P. polita* Deichmann 1936 (p. 162-163 pl. 26 fig. 12) (*P. distans* Wright & Studer 1889) (p. 87-89 pl. 17 fig. 1) have the low number of zooids in the whorls in common. STUDER states for *P. distans* that the lowermost whorl has two zooids and that the number of zooids per whorl soon increases to four. WRIGHT & STUDER state for their material (i.e. the species *P. polita*) a number of whorls which in the lowermost whorl is two, in the next whorl four, and then four, but in some whorls five. DEICHMANN (1936) has a number of zooids in the whorls as usually three, rarely four. Both these species have thus a somewhat higher number of zooids in the whorls than *P. jamgerseni*.

Distances between whorls. It is not pointed out any differences between *P. distans* and *P. polita*. KÜKENTHAL, however, who as mentioned above has pointed out that Studer's specimen from 1878 and WRIGHT & STUDER's specimens from 1889 are identical, gives as basis for the measurements the different appearance of the figured specimens. It is very difficult to explain, that the intervals between whorls of *P. distans* are larger than those in WRIGHT & STUDER's specimens. The statements in the texts are, however, that the intervals between whorls are 2.5-3.0 mm, the intervals as 3-4 mm, and the intervals between the whorls as 3-4 mm. In the case of the intervals between the whorls in species as e.g. in *P. jamgerseni*, the intervals are measured from the attachment of the calyx to the calyx. Thus in one specimen the intervals between whorls measured 2.7 mm, whereas in another specimen the intervals measured 5.4 mm, and in a third specimen the intervals measured 3.4 mm. It is to be noted that whereas the intervals between whorls of *P. jamgerseni* are measured from the attachment of the calyx to the calyx, the intervals between whorls of *P. distans* and *P. polita* are measured from the base of the zooids.

STUDER states for the verrucae (Zellen) of *P. distans* a size of 2 mm, and WRIGHT & STUDER state for the verrucae (calyces) in their material of *P. polita* a size of 1.5-2 mm. DEICHMANN gives 1.5-1.75 mm. Thus the size of the verrucae in the three species discussed here is nearly equal and then cannot yield any distinguishing character. Such may, however, be found in their shape and scale covering. The verrucae of *P. distans* are, as pointed out by KÜKENTHAL in 1919, curved adaxially towards the stem, whereas the verrucae in WRIGHT & STUDER's specimens issue from the stem at an acute angle and are straight, as is also the case in DEICHMANN's specimen. STUDER's figures show further (if they are correct) that the verrucae of *P. distans* are more slender than those of *P. polita* and *P. jamgerseni* and that in the abaxial rows of zooid body scales the number of scales is considerably higher (about 10-15). It also appears from the figures that in the zooids of *P. distans* just as in *P. polita* there are only two abaxial rows of scales visible from the abaxial side of the zooid.

STUDER'S *P. distans* is somewhat larger than both *P. polita* and *P. jamgerseni* as it measures 23 cm without the lacking basal part, whereas the largest *P. polita* (that figured by WRIGHT & STUDER 1889 pl. 17) including the base of the colony measures about 22 cm and the largest *P. jamgerseni* only measures 9.1 cm. *P. distans* differs from the two other species in having a very considerable part of the stem without zooid whorls; STUDER states 11 cm between the scar of the broken off base and the lowermost whorl.

P. distans may in the slender verrucae with more than 10 zooid body scales in the abaxial rows be sufficiently distinguished from the two other species of *Primoella* with a low number of zooids in the whorls; i.e. *P. jamgerseni* and *P. polita*; and these may easily be distinguished from each other, since in the verrucae of *P. jamgerseni* four rows of scales are visible from the abaxial side, whereas in *P. polita* only two rows are visible, as so nicely shown in WRIGHT & STUDER's figures.

In *P. jamgerseni* the verrucae are also chubbier. The base consists in *P. jamgerseni* of an only small basal disk, whereas in *P. polita* it consists of a lobed or root-like portion which may be of a rather large extent. Nor for *P. polita* is described an inner layer of lumpy spicules in the stem, and such a layer may probably be missing.

P. jamgerseni belongs to the group Convexae Wright & Studer 1889 (Diagnosis: Vide KÜKENTHAL 1919 p. 385). To this group also *P. distans* and *P. polita* have been reckoned though with the same right they might have been referred to the other group, Carinatae Wright & Studer 1889 (= Compressae KÜKENTHAL 1919). The shape of their zooids is as described for the Convexae, but the number of scale rows visible from the abaxial side of their verrucae is only two as in the Carinatae.

Distribution. *Primoella jamgerseni* is known from two localities, viz. in the Danmark Strait (south of the Wyville Thomson Ridge) and in the North-Atlantic Ocean somewhat south of the first locality, Chart of distribution, fig. 32, the depths being 2418 m and 2137 m respectively. It is to be noted that this species is the first *Primoella* which has been found in the North-Atlantic Ocean, whereas hitherto all the about 20 species described of this genus have been found in the southern hemisphere and only one of them, the above-mentioned *P. polita*, also has been taken north of the equator, at the West Indies.

Besides in the geographical distribution *P. jamgerseni* also differs somewhat from the other species of *Primoella* in the bathymetrical distribution as hitherto no other *Primoella* has been found in depths greater than about 1100 m.

Primoella jamgerseni may be expected to have a very wide distribution as it is a deep-sea species of a genus which otherwise is nearly exclusively restricted to the southern hemisphere.

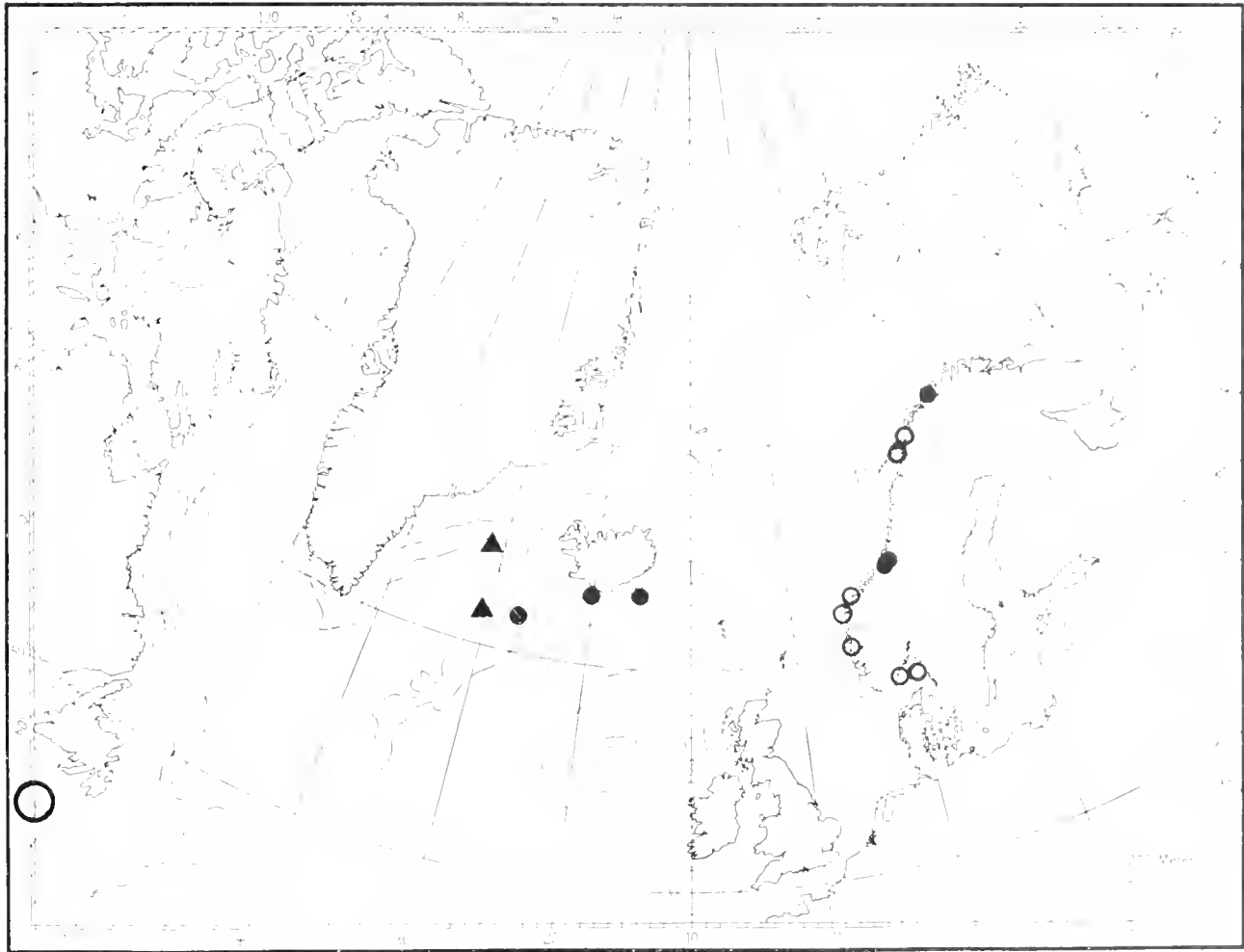


Fig. 32. ● *Anthothela grandiflora* (M. Sars). ▲ *Primonella pumescens* n. sp.
The large open figure indicates that the species is recorded from the area, though without exact locality.

Family: **Gorgonidae** Dana 1846 emended.

Diagnosis: Vide Kükenthal 1924, p. 324.

Genus: **Stenogorgia** Verrill 1883.

Diagnosis: Kükenthal 1924, p. 347.

Stenogorgia borealis (Jungersen) Kramp 1930.

Stenogorgia borealis Jungersen, 1915, p. 1186, nomen nudum.
Kramp, 1930, p. 641, figs. 2-5.

Description: Vide Kramp 1930.

Material:

65° 13' N, 26° 58' W, 171 m, 64°C, "Ingolf" St. 16,
62° 00' N, 22° 38' W, 1630 m, 2.9°C, - 39,
61° 11' N, 27° 00' W, 913 m, 6.1°C, - 81,
65° 38' N, 26° 27' W, 260 m, 5.9°C, - 98,
66° 33' N, 20° 05' W, 83 m, 5.6°C, - 127
The Faroes,
S. W. Greenland, Kvanefjord, 565 m, "Tjalle",
S. W. Greenland, Bredefjord, 700 m, "Rink" St. 125,
S. E. Greenland, Lindenowfjord, 300 m,
63° 50' N, 10° 55' W, 330 m, "Dana" St. 6000,
63° 33' N, 11° 25' W, 317 m, 4.31°C, (315 m), "Dana St. 6001

Remarks: The colony from the "Ingolf" St. 98 was with-
out exception stated to be grey with a pale blue touch.

Distribution: The species is found in several localities on
the Wyville Thomson Ridge from the Faroes to Greenland, it
is further found in two localities somewhat south west of Ice-
land and in some of the southernmost fjords of Greenland both
on the east and west coast. The list above includes all the pre-
viously known localities which apart from the three last mentioned
all are marked on a chart in Kramp's paper (1930 p. 10).

Stenogorgia rosea Græb 1887

Stenogorgia rosea Græb, 1887, p. 158, pl. 1, figs. 1-11, pl. 2,
figs. 1-9.
Bütschowsky, 1929, p. 189-190, fig. 31,
Kükenthal, 1924, p. 349,
Pax, 1936, p. 258,
Carlgren, MS.

For further references vide Kükenthal 1924.

Description: Vide Græb 1887 and Bütschowsky 1929.

1929

1929. — *Chryso-gorgiidae* (subfamily) in *Chryso-gorgiidae* (subfamily) of the Kattegat, to the south of the island of Bornholm. — *Baltische Fauna* (1929) 1929, p. 121. (To be a mistake for *Chryso-gorgiidae* was not dealt with in the

author's paper. The Danish Octocorals from the Waters inside Skagen. — Professor CARSTEN states, however, in his manuscript on the Danish Anthozoa (= Koraller, Danmarks Fauna²) that the species actually is taken in the northern Kattegat, at Tistlarna, by JÄGERSKOLD, the locality being situated somewhat south west of Göteborg.

Bathymetrical distribution: About 20–100 m.

Family Chryso-gorgiidae Verrill 1883

Diagnosis: Vide KÜKENHAL 1919, p. 189–190.

Subfamily Lepidogorgiinae Verhulst 1902

Genus Radicipes Stearns 1883

1883. — *Ann. Kükenthal* 1919, p. 541.

All the members of the species of this genus are described by Verhulst (1902), and of these species two were recaptured by J. A. Thomson (1927). Except for J. A. Thomson (1927), it is true, referred to the genus of the "Ingolf" to only one species, viz. *Radicipes* (Wright & Studer) but this was, as will be evident from the descriptions below, a lapse, as, in reality, only one of the species is referable to that species, whereas the other un-*Radicipes* species, viz. *R. quaedam* (Verrill), was referred to *Verhulstia* with a query was made a synonym of *Radicipes* (Wright & Studer) by KÜKENHAL (1919) though doubtful, and in fact the Verrill's description, though on some points different, especially gives the main characteristic features of the genus, and comparison with the other species known in

Radicipes challengerii Wright & Studer 1883

1883. — *Ingolf* (Wright) 1883, p. 691.

1883. — *Ingolf* (Wright & Studer) 1883, p. 3–4, pl. 1, figs. 1, 1a, pl. 5, a, fig. 2.

1902. — Verhulst 1902, p. 14–15, figs. 18–19.

1927. — J. A. Thomson 1927, p. 26, fig. 2, *quaedam* (partim) Jungermann, 1915, p. 151.

1919. — Kükenthal 1919, p. 543–544.

1924. — Verhulst 1924, p. 411–412.

1927. — J. A. Thomson 1927, p. 20, pl. 1, fig. 17, *quaedam* (partim) Krøyer, 1932, p. 19.

1927. — Wright & Studer 1883, fig. 2, 1 colony.

The colony is composed of an open spiral. Stem slender, cylindrical, with the zooids attached to the stem in a regular spiral. The zooids are situated at a distance from the tentacles having a diameter of about 1.5 mm, the tentacles reaching a length of about 4 mm. The intervals between the tentacles are 2–3 mm. Intervals between the zooids are 4–5 mm. Intervals between the tentacle crowns are of a few mm number. In stem diameter at 8–10 cm. (type specimen) size about 0.5–0.6 mm. In diameter at 20–30 cm. size about 0.15–0.05 mm. In diameter at 40–50 cm. size about 8-shaped 0.1–0.15 mm (0.17–0.05 mm) or 0.1–0.15 mm (0.17–0.05 mm) or 0.1–0.15 mm (0.17–0.05 mm). In the diameter at 60–70 cm. size about 0.25–0.03 mm. In the diameter at 80–90 cm. size about 0.25–0.02 mm. In the diameter at 100–120 cm. size about 0.12–0.03 mm. In the diameter at 140–160 cm. size about 0.12–0.03 mm.

Description: The material consists of a single colony which is broken into several fragments and lacks the basal part. The colony is unbranched, highly flexible and somewhat elastic and may, when alive, have measured about 70 cm or more in length. It has been coiled into an open spiral with several windings and has had the zooids placed on the inner side of these.

The colour of the specimen (in alcohol) is white.

The round axis is white, slightly light-blue iridescent, measuring in the lower part of the colony about 0.5 mm in diameter. The diameter of the stem is about 0.75 mm.

The zooids are placed obliquely on the stem in such a manner that a rather long part of the zooid body is parallel to the axis (fig. 33). They are arranged in a unilateral row, usually quite close to each other, but the successive intervals may also be up to 4 mm. The intervals seem, however, larger on account of the long part of the zooid body which lays parallel to the axis. The intervals between the tentacle crowns vary from 2 to 12 mm. Occasionally the zooids may be placed so close together that a part of one zooid overlaps a part of another; in one place in the colony four zooids (tentacle crowns) are thus crowded on only 6 mm.

The zooid body reaches a length of generally 7–8 mm, measured parallel to the axis, and has, perpendicularly on the axis, a height which reaches about 2 mm, being generally 1.2–1.3 mm, and to this come the tentacles which may have a length of about 4 mm.

The heights of the zooid bodies are thus somewhat greater in this specimen than in the type specimen examined by VERHULST, who in 1902 (p. 15) for the corresponding measure states "höchstens 4 mm, meist 0.9 oder 0.8 mm und weniger." VERHULST's statement that the intervals between the zooids usually are 4–5 mm and in the upper part of the colony about 7 mm, refers probably to the intervals between the tentacle crowns.

The zooids, seen from above, may reach a width of about 1.75 mm and are thus somewhat protruding from the stem. Below the tentacles they have a diameter of about 1.5 mm. Several of the zooids are filled with and expanded by ripe or nearly ripe eggs, and only in these the lower part of the zooid body is easily distinguishable from the stem (compare fig. 33).

The tentacles are of unequal size. The adaxial one is always small, often quite rudimentary, usually of a size of about 1 mm or less. The other tentacles usually measure up to about 4 mm. Some (two or three) of the abaxial are, however, often distinctly smaller than the other. The tentacles are not retractile, but by contraction curved in over the peristome. The pinnulae may number up to about 20–25 pairs.

Among the full grown zooids a less developed one may occasionally be found.

The tip of the colony which has been broken off is regenerating, and the stem measures here, below the scar, about 1.3 mm in

diameter, but above the scar, in the regenerating part, it measures only about 0.1 mm in diameter. The old axis is not continuing into the regenerating tip, but several millimetres below the scar a new axis is developed lying parallel to and very close up to the old axis and continuing into the new tip of the colony. This

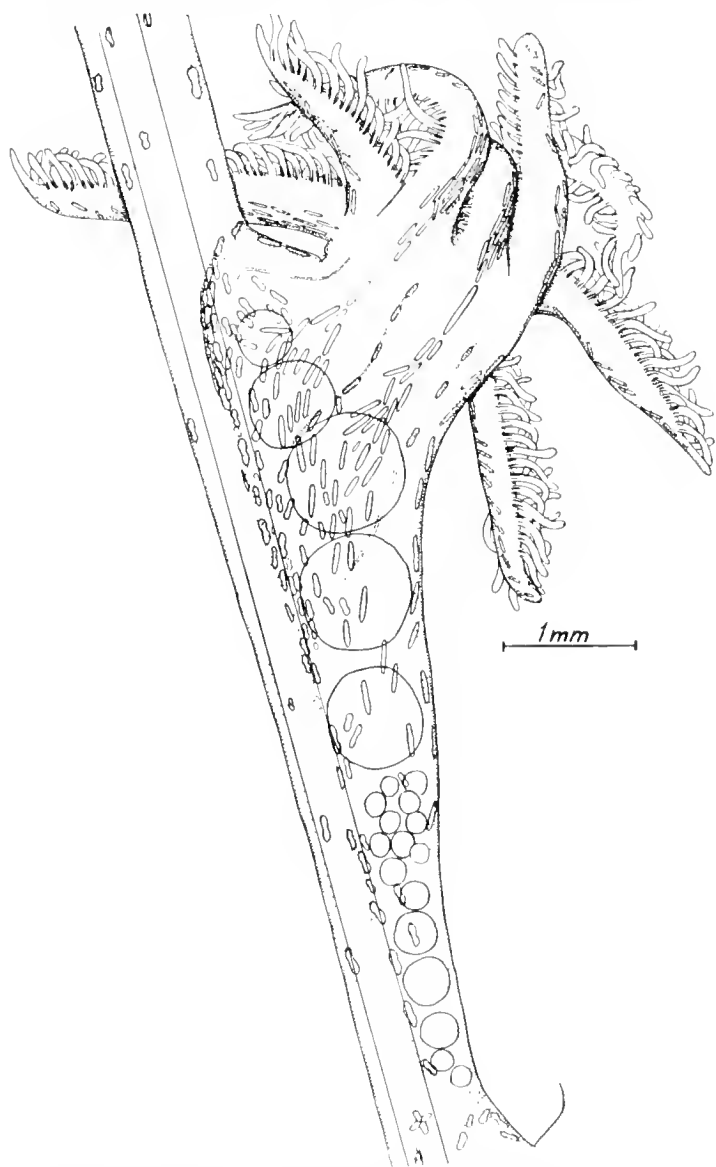


Fig. 33. *Radicipes chalonperi* (Wright & Studer). The zooid has been made transparent and the axis and the eggs are indicated. It may be noted that both somewhat more and somewhat less spiculated zooids are found.

tip bears two zooids which have not yet developed an oral opening or tentacles but nevertheless contain large eggs.

The coenenchyma of the stem is thin and provided with extremely few sclerites, which are placed so scattered that there may often be about 2 mm between them; a few sclerites may, however, sometimes be found rather close to each other (fig. 31 *k*). The stem sclerites are scale-like and placed in the longitudinal direction of the stem. The most common type is the oblong form with rounded ends and narrowed middle (i. e. nearly 8-shaped) which is figured fig. 31 *a, b*; the largest of this type measures about 0.25 · 0.07 mm, but besides these some somewhat smaller sclerites with a less smooth surface (fig. 31 *c, g*) or a more irregular outline (fig. 31 *h, i*) may be found. The latter form, the largest of which measures about 0.15 · 0.05 mm, is by VERSLUYS (1902 p. 11) considered the most common form in the specimen examined by him.

In the zooids the sclerites are likewise only few in number. In the basal part of the zooid body they are for the most part transversally placed, in the middle and upper part of the zooid body they are, however, longitudinally arranged and principally restricted to eight intermesenterial rows (fig. 33). These rows of sclerites continue sometimes without interruption into the rows of sclerites in the tentacle dorsa, often, however, the sclerites are quite absent in the upper part of the zooid body. Besides the intermesenterial rows of sclerites a few sclerites may be found along the attachments of the mesenteries.

The sclerites of the zooid body are scale-like and rather smooth, yet with some very small teeth at the ends. The transversally placed ones at the base of the zooids are mainly of the same type as the most common in the stem. The largest of these 8-shaped sclerites are about 0.25 mm in length, but generally they are less than 0.2 mm (fig. 35 *d, e, o*). The longitudinally placed sclerites in the main zooid body are generally of a somewhat longer and narrower type without constricted middle (fig. 35 *c*). They usually measure about 0.25 · 0.03 mm and not more than 0.3 mm in length. Only the single sclerite, which in the upper part of the zooid body often fills out the space between the rows of sclerites in the zooid body and the tentacle dorsum, may be somewhat longer and is often somewhat narrower (e. g. 0.1 · 0.02 mm). Twins, quadruplets (fig. 35 *g, h*) and some small differently shaped sclerites (fig. 35 *f*) may be found between the other sclerites, and 8-shaped sclerites may be found in the longitudinal rows of sclerites, as also in the basal part of the zooid body some of the other type occur.

The sclerites of the tentacles are in the tentacle dorsum rather long, narrow scales or flattened spindles (fig. 35 *p, s*) the largest of which generally measure about 0.25 · 0.02 mm. In the lower part of the tentacle they are placed longitudinally in a row which may be four (sometimes six) sclerites broad, in the middle and

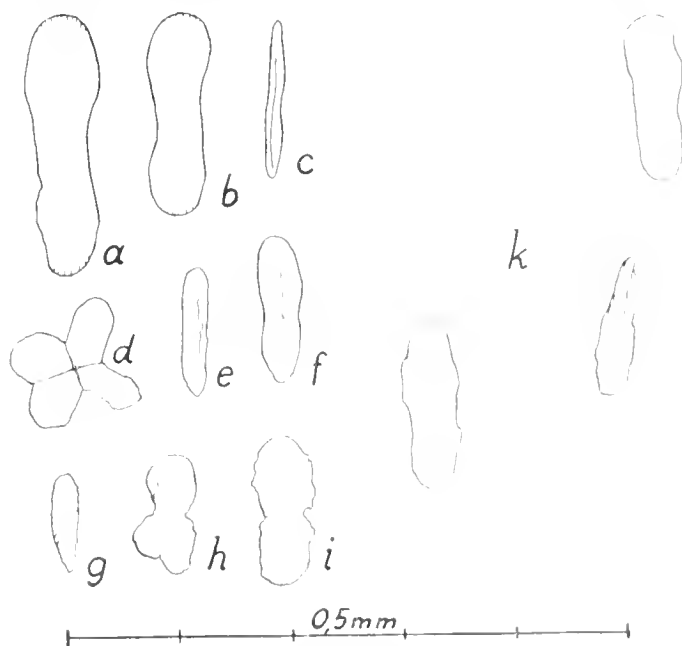


Fig. 34. *Radicipes chalonperi* (Wright & Studer). *a, b*, sclerites from the stem, *a, b*, of the most common type, *c*, shown from the side, *k*, a group of three sclerites from the stem in natural position.

upper part of the tentacles they decrease in size and are more scattered and usually more transversally placed (in the fully outstretched tentacles they become probably all longitudinally arranged).

In each pinnula there are two sclerites, one on each side. They are scale-like and measure generally 0.12 (· 0.2) · 0.05 mm

with length. They are, however, arranged in such a manner that one can see them at the base of the pinnula. The base of the pinnula with its basal part. Thus that picture is seen in Verrill's (1902, p. 15) description of *Radicipes*.

Wright & Studer (1902) describe *Radicipes* with the following figures:

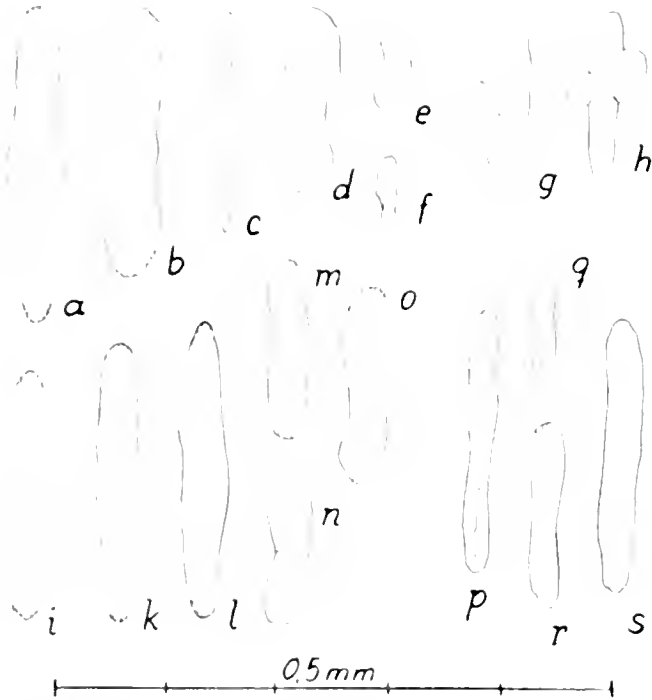


Fig. 1. *Radicipes gracilis* (Wright & Studer): a, h, selerites from zooid body; b, d, of the most common type, c, shown from the dorsal border; e, selerites from upper part of zooid body; f, g, of the dorsal type; p, selerites from tentacle dorsum.

Description. The description above differs on several points from that given by Wright & Studer's original description of *Radicipes gracilis* (1902). Verrill (1902) gave, however, after having examined one of the type specimens, a redescription of the species with correction of Wright & Studer's errors, and his description, by the "Ingolf" agrees in all essentials with that given above, which only in some few points was not quite correct.

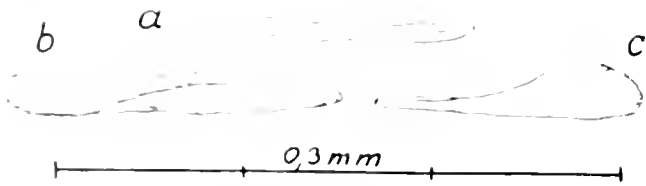


Fig. 2. Wright & Studer: Selerite from pinnula of *Radicipes gracilis*, shown from the side.

Radicipes gracilis is hitherto only known from the coast of the island of Gibraltar, 1080 m, and a locality in the Atlantic Ocean, 8° 10' N, 15° 40' W, 750 m (recorded by the "Ingolf"). The latter locality is situated some 100 miles from the coast. It is probably the species mentioned by Verrill (1902) as occurring in the deeper parts of the Atlantic Ocean (his illustration, fig. 37, is identical with that given here, fig. 1, 7, 8, 1717).

***Radicipes gracilis* (Verrill 1884).**

Lepidogorgia gracilis Verrill, 1884, p. 220.
 1885, p. 512, pl. 2, figs. 10, 10a.
 Verrill, 1902, p. 15.
Radicipes challengerii (= *gracilis* (partim)) Jungersen, 1915, p. 1183-1184.
gracilis Kukenthal, 1919, p. 518.
 Verrill, 1922, p. 12, figs. 10, 10a.
Lepidogorgia verrilli J. V. Thomson, 1927, p. 20-21, pl. 3, fig. 18, pl. 5, fig. 20.
Radicipes gracilis Kukenthal, 1924, p. 112.
challengerii Kramp, 1932, p. 10.
gracilis Deichmann, 1936, p. 227.

Material

- 61 50' N, 56 21' W, 2702 m, 15°C, "Ingolf" St. 36, 13 juvenile colonies - some fragments.
- 62 00' N, 21 36' W, 1591 m, 3-3°C, "Ingolf" St. 10, 11 colonies.
- 62 06' N, 22 30' W, 1587 m, 3-1°C, "Ingolf" St. 68, 1 colony.
- 62 57' N, 19 58' W, 957 m, "Thor" St. 166 (03), 1 juvenile colony.
- 62 19' N, 56 00' W, 2550 m, 1-91°C, (2525 m), "Godthaab" St. 21, 2 colonies.

Diagnosis: Colony erect, slightly spirally twisted. Stem slender, rigid, only towards the tip flexible and elastic. Calcareous base root-like, much branched. Zooids fairly cylindrical and obliquely seated at intervals of generally 5-10 mm. Zooid body when fully grown measuring nearly 2 mm in length by about 1 mm in diameter. Tentacles of about equal size; when contracted about 3 mm in length. Size of verrucae usually 3-4 mm. Sclerites of coenenchyma scale-like, oblong with rounded ends and constricted middle, longitudinally arranged in a generally fairly close layer, average size about 0.13-0.035 mm, maximum size about 0.3-0.05 mm. Zooids strongly armed, the interseptal sclerites somewhat obliquely placed and of the same type as the stem sclerites, the septal ones, and those of the lower half of the tentacle dorsum, longitudinally placed larger scales usually without constricted middle, average size 0.3-0.05 mm, maximum size about 0.5-0.06 mm. In some zooids, usually in the tentacle bases, also some few more or less flattened spindles measuring up to 0.7-0.05 mm. Sclerites of outer half of tentacle dorsum obliquely placed scales 0.1-0.02 mm 0.2-0.03 mm. Sclerites of pinnulae slender, scale-like, somewhat twisted, measuring up to 2 mm in length; often a few in each pinnula.

Description. The material includes colonies of a much varying age.

The smallest specimen is the juvenile one from the "Thor" St. 166 which only measures about 1.4 cm in height and has not yet developed more than the primary zooid, nor has the specimen developed a calcareous base, but the tiny stem, which measures only 0.1-0.2 mm in diameter, passes below over in a small bag formed extension into which also the axis of the specimen extends. The single zooid measures, with incurved tentacles, scarcely 2 mm in length, having a width of scarcely 1 mm.

Also the collection from the "Ingolf" St. 36 consists of juvenile specimens which, however, all have developed the characteristic features of the species. They vary in height from about 2.5 cm to 4.5 cm and bear three to four zooids placed unilaterally. The root-like calcareous bases, which anchored the specimens into the mud, are richly branched and in comparison with the stems fairly large. None of the bases bear now more than one stem, but from some of the bases it seems that more stems have risen when the specimens were alive. The diameter of the stem is scarcely 0.2 mm. The interval between the base and the lowermost zooid varies from 15-25 mm and the intervals between the zooids may vary from about 1 to about 5 mm. The largest zooids measure, with the tentacles slightly incurved, about 5 mm in

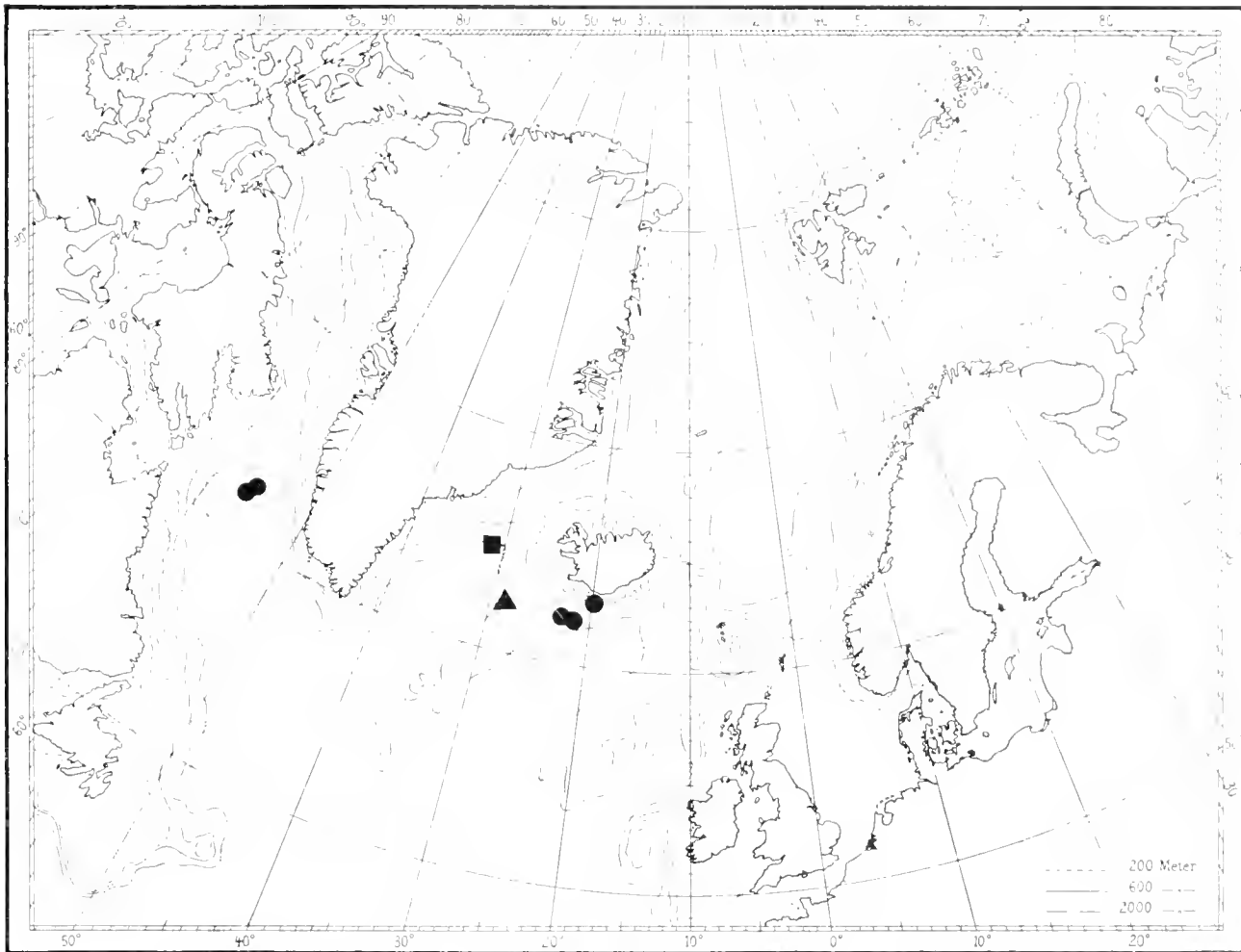


Fig. 37. ▲ *Radicipes challengeri* (Wright & Studer), ● *Radicipes gracilis* (Verrill), ■ *Chrysopogon campanula* n. sp.

length by about 1 mm in width. The uppermost zooid is always full-grown, but else in the colonies every second of the zooids is generally a young one.

The material from the "Godthaab" St. 21 consists of a single calcareous base from which, however, two stems arise. One of these is 7.2 cm in length and has five zooids, the interval between the base and the lowermost zooid being 32 mm, and the intervals between the zooids varying from 7-10 mm. The other stem measures about 8.5 cm in length and bears full-grown zooids. The interval between the base and the lowermost full-grown zooid is 30 mm, and the intervals between the zooids vary from about 5 to 9 mm. The stem has in the lower part a diameter of about 0.75 mm, having about 25 mm above the base two small zooid-buds, less than 0.5 mm high, placed 2 mm apart. The verrucae measure about 3-4 mm.

The colonies from the "Ingolf" St. 10 are of much varying size, the smallest measures about 7.5 cm and the largest may, when alive, have measured more than 50 cm in length. The interval between the base and the lowermost zooid is about 60 mm in a 15 cm high colony, in a 39 cm high colony about 80 mm, in a 25 cm high colony about 75 mm, and in a 10 cm high colony about 15 mm. The largest interval between the base and the lowermost zooid is found in the colony 34 cm high, from "Ingolf" St. 68, and measures about 95 mm. The intervals between the zooids are generally 5-8 mm, but may vary from 1 to 11 mm. The zooids seem to vary somewhat in size. In some of the larger colonies the largest verrucae are scarcely 3 mm in height. In other colonies the verrucae are 1.5 mm high by 1.1-

1.2 mm in width. In more of the colonies the zooids are distinctly of unequal age, every second being considerably smaller than the others.

The diameter of the stem is generally about 1 mm in the lower part of these colonies which are from 10 cm to 50 cm high, but in one colony, 15 cm high, the stem is considerably thicker, measuring below about 2 mm in diameter and in nearly its whole length being distinctly thicker than the zooids, which however are fairly small in this colony. In this specimen also the spiculation of the stem, as will be mentioned below, differs somewhat from the typical.

The colonies are erect, slightly spirally turned, and the larger specimens are rigid, only towards the tip somewhat flexible. The axis is roundish, having the surface provided with short longitudinal scratches. In the smaller colonies the colour of axis is whitish and light-blue iridescent, in the lower part of the medium-sized colonies more yellow and in the larger colonies slightly light-brown.

The colour of the colonies is in alcohol white. The juvenile colonies from the "Ingolf" St. 36 were noted as colourless when alive and the large colony, 36 cm high, from the "Ingolf" St. 68 was noted to have the axis shining whitish through the coenenchyma and very pale pink zooids.

The zooids are fairly cylindrical in shape and placed somewhat obliquely upon the stem which is slightly swollen at the places where they are attached. The zooids are strongly spiculated and by contraction somewhat curved towards the stem. The tentacles are of about equal size and not retractile. The zooid

of the stem is about 2 mm) whereas the sclerites are generally of a length of 0.3-0.5 mm. Typically the sclerites are arranged longitudinally in the stem, but in some colonies they are arranged transversely. The sclerites are regularly placed and in the lower part of the stem they are arranged at the middle of the zooids. The sclerites of the stem are scale-like, with a constricted middle. Their average length is 0.3-0.5 mm, their maximum size about 0.6 mm. They are arranged, generally in a regular pattern. In the lowest part of the stem the sclerites are scattered whereas they are

arranged (fig. 38). The interseptal sclerites of the zooid body are generally of a slender scale-like form with more or less rounded ends and usually without constricted middle. Their maximum



Fig. 38. *Verillia*: *a*, a zooid from one of the juvenile colonies from the "Ingolf" St. 36; *b*, a zooid of the colony from the Godthardsholmen St. 24.



Fig. 39. *Radicipes gracilis* (Verill). *a*, sclerites in natural position in the stem of a juvenile colony from the "Ingolf" St. 36, between the uppermost and next uppermost zooid; *b*, a section of coenenchyma with the sclerites in natural position, from the middle part of stem in a colony 32 cm high from the "Ingolf" St. 49; the diameter of the stem in the lowest part of this colony scarcely 1 mm.

size is about 0.5 - 0.06 mm, and generally they measure about 0.3 mm in length (fig. 41 *a, b, m, q*).

In the tentacle base and nearly the lower half of the tentacle dorsum similar sclerites are found arranged longitudinally in a row generally 5-8 sclerites broad. The sclerites decrease gradually in length from the tentacle base, where the largest sclerites of the zooid are usually found, until they become about 0.2 mm in length at the outer half of the tentacles. The sclerites in the outer half of the tentacle dorsum are typically obliquely placed in two rows, one on either side of the middle of the tentacle. Their

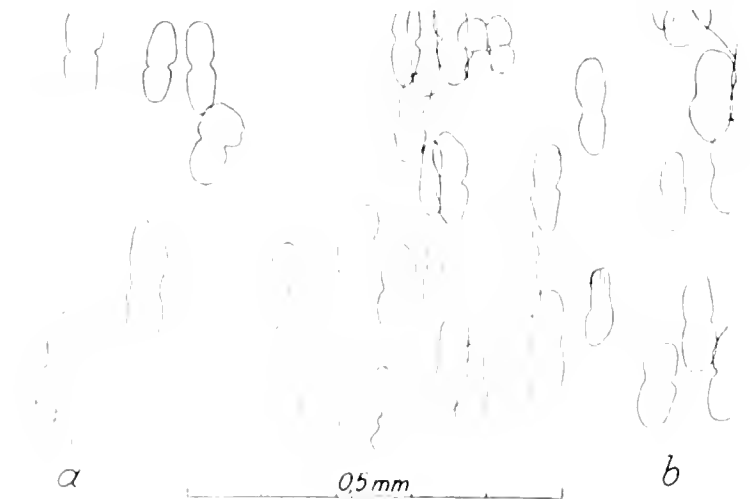


Fig. 40. *Radicipes gracilis* (Verill). Sections of coenenchyma of stem in the colony, 45 cm high, from the "Ingolf" St. 49, which in the lower part has a diameter of about 2 mm. *a*, from the lowest part of the stem below the zooids; *b*, from the upper part of the stem between two zooids.

tentacle base layer near the zooids (comp. fig. 38). In the figures 39 *a, b* that the sclerites of the stem of colonies from the "Ingolf" St. 36 are somewhat smaller than those from the larger colonies, and, as can be seen, there is a corresponding difference in the diameter of the stems. In the somewhat differing colony from St. 49, in the unusually thick stem the sclerites are arranged, generally, as in fig. 40, but also in this colony the sclerites are arranged at the places of the zooids. The sclerites of the stem from the "Thor" St. 496 sclerites are arranged in the lower part of the stem and in the zooid. The sclerites of the tentacle base is often devoid of sclerites, but they occur and are then of the same type as those of the stem, also longitudinally.

The sclerites of the generally scale-like sclerites of the stem are of the same type as the sclerites of the stem, generally parallel to the stem. The sclerites in the zooid body are placed in the same type as those from the stem, but they are not obliquely arranged in the tentacle dorsum (fig. 38 *b, 41 b*). The sclerites of the stem are typically longitudinally

length varies from 0.2–0.1 mm and their width from 0.03–0.02 mm (fig. 11 *r* *u*).

The pinnulae number from about 15 to 20 pairs. Especially the outermost ones may be devoid of sclerites, but else the pinnulae are generally provided with two or a few slender, scale-like, somewhat twisted sclerites (fig. 11 *v* *a*) the largest of which measure about 0.2 mm.

In some zooids, in the lower half of the tentacles or in the upper part of the zooid body, a few sclerites may be found which differ from the typical ones in being more slender and spindle-shaped, though usually more or less flattened (fig. 11 *f* *h*). In the juvenile colonies, where, as already mentioned, the sclerites distinguish themselves by their comparatively long and slender form, also some few of the interseptal sclerites which extend into the tentacle base are exceptionally large and of a very slender, flattened spindle-shaped type measuring up to about 0.7 mm in length by about 0.05 mm in width (fig. 11 *v*, *k*).

All the sclerites are fairly smooth, with rather smooth edges, only the ends having a fine-toothed appearance.

No sclerites were noticed in the peristome.

Discussion: There may be no doubt that the specimens described here belong to VERRILL's species, *Lepidopogona gracilis*. Though this species was somewhat insufficiently described, VERRILL's original description includes all its most conspicuous characters. Thus VERRILL (1881 p. 222) described the axis as "tall, slender, terete, tapering to a flexible tip" and just this axis, which in the larger colonies is rigid and only is flexible towards the tip, seems to be a characteristic feature in this species in comparison with the other species of *Radiocipes*. Likewise VERRILL's description of the general appearance of the colony and his statement of the intervals, and the diameter of the zooids and stem corresponds well with the material of the "Ingolf" Expedition.

VERRILL states for the colonies a size of up to 90 cm and for the living colonies an orange or salmon colour.

From the descriptions above of *R. challengerii* and *R. gracilis* it will be evident that these two species are quite distinct, and that JUNGERSSEN in 1915 made a lapse when he, though with a query, united them. By the closer examination of the specimens, which JUNGERSSEN, if he had lived, would have had to make before he was able to publish the rest of his intended reports on the northern Octocorals, he himself would certainly have discovered and corrected the error.

R. gracilis is easily distinguished from all other, previously described, species of *Radiocipes* especially by its rigid, erect stem, as the stem in all other species of *Radiocipes* is coiled into a spiral and very flexible. That species, which most resembles *R. gracilis*, is *R. aureus* Kükenthal (1919 p. 514) from the Indian Ocean which likewise has fairly distinctly placed zooids with tentacles of about equal size. In this species the stem sclerites seem, however, typically to be without constricted middle, and the sclerites of the

zooid body attain a considerably larger size, measuring up to 1.3 mm, and seem also, after KÜKENTHAL's figures, to be much more irregularly placed than in *R. gracilis*. The species has also, as distinct from *R. gracilis*, the usual spirally coiled stem.

It seems most probable that the *Lepidopogona verrilli* which

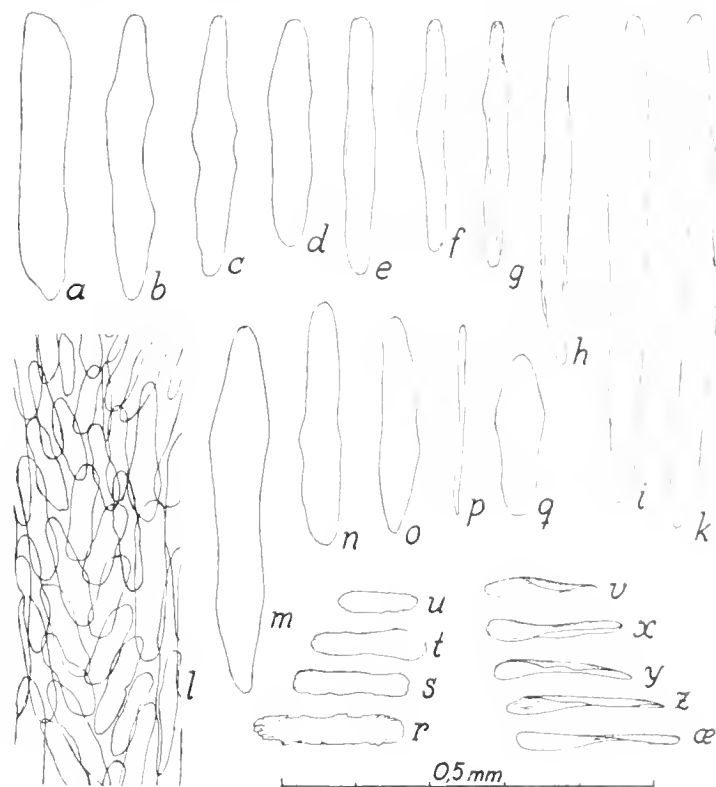


Fig. 41. *Radiocipes gracilis* (Verrill). Sclerites of zooid, *a*, *b*, *m*, *q*, interseptal scale-like sclerites from zooid body, *p*, shown from the side *h*, *k*, exceptional large and rather spindle-shaped sclerites from upper part of zooid body and tentacle base, *e*, *g*, sclerites from lower half of tentacle dorsum, *c*, *d*, scale-like, *e*, *f*, flattened spindle-like, *a*, spindle-shaped one about round in section, *l*, section of zooid body showing the septal sclerites in natural position, *r*, *u*, sclerites from outer half of tentacle dorsum, *v*, *a*, sclerites from pinnulae.

J. A. Thomson in 1927 described from 42° 40' N, 62° 19' 30" W, 1450 m, is actually a *R. gracilis*, both the locality and Thomson's figures support this supposition. *R. verrilli* (Wright) is moreover a Pacific species.

Distribution: The species is found in the Davis Strait and south of Iceland and has further been recorded from the western part of the Atlantic ocean from off Georges Bank in the north to off Delaware Bay in the south. Chart of distribution, fig. 37.

Bathymetrical Distribution: about 957–3173 m.

Subfamily: *Chrysogorgiinae* Studer 1887, emended Versluys 1902

Genus: *Chrysogorgia* Duchassaing & Michelotti 1866.

Diagnosis: Vide KÜKENTHAL 1919, p. 506.

The genus comprises several species of which, however, only a few belong to the Atlantic fauna. Most of the species are found in the tropical parts of the oceans at moderate depths whereas the two North-Atlantic species are found at fairly great depths.

Chrysogorgia agassizii (Verrill 1883).

Dasygorgia agassizii Verrill, 1883, p. 22–23, pl. 2, figs. 1, 1a, 1b, 1881, p. 220.

1885, p. 511–512, pl. 9, fig. 199.

Roule, 1896, p. 304.

Chrysogorgia agassizii Versluys, 1902, p. 60.

nec. Nutting, 1912, p. 55.

Chrysogorgia agassizii Kükenthal 1919, p. 530, fig. 233.

1921, p. 103.

C. pentasticha J. A. Thomson, 1927, p. 22.

agassizii Deichmann, 1936, p. 233–234, pl. 23, figs. 31–40, pl. 24, figs. 1–2.

Material.

61° 31' N, 31° 12' W 2418 m, 1 6 C "Ingolf" St. 11, 12 more or less

fragmentary colonies, some axis fragments and 16 basal disks.

62° 06' N, 19° 00' W, 1960 m, 3 1 C "Ingolf" St. 64, 9 more or less fragmentary colonies and some axis fragments.

62° 58' N, 25° 21' W 1492 m, 1 5 C "Ingolf" St. 81, 2 axis fragments.

about a right angle from the first stem, but soon bending and becoming parallel to this. It is noteworthy that the new stems are not developed at the issue of a branch, but outside the spiral of branch-building coenenchyma.

One of the larger colonies is bent at right angles on two places, 50 mm apart. The colony has apparently twice in its life been

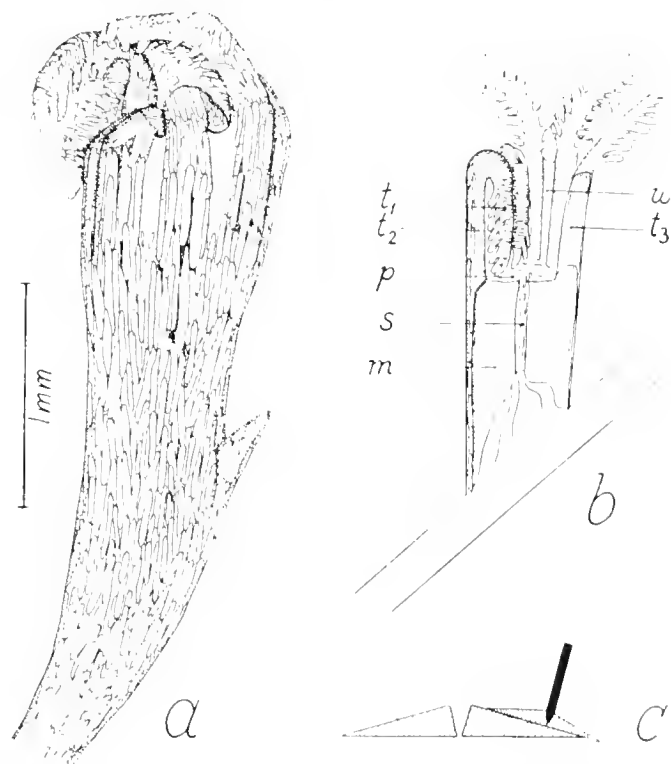


Fig. 12. *Chrysogorgia agassizii* (Verrill). a, a tip-zooid. b, scheme of zooid. t_1 , outer free, pinnae-bearing part of tentacle. t_2 , lower half of tentacle. u , the web which connects the lower halves of tentacles. t_3 , the hollow tentacle. p , the peristome. s , the stomodaeum. m , mesenteries. c, two basal disks in section, showing the conical attachments of the horny axis into the calcareous base.

overturned and has reacted on the injuries by in the first case developing a new stem perpendicular to the old one and in the other case by bending the stem at a right angle.

In one colony an anastomosis was noticed between two different branches.

The intervals between successive stem-branches are generally about 2.5 mm, varying between 2 and 3.5 mm, yet in some colonies a few of the lowermost intervals may be considerably larger, up to 13 mm. The orthostiche intervals (intervals between successive branches of the same vertical plane) vary from 10 to 15 mm and are generally 11-12 mm.

The branching of stem is, as mentioned above, typically 2_5 turn dextrorse. This regular mode of branching is, however, occasionally somewhat disturbed. The branches are e. g. in sections of a few of the colonies placed in such a manner that about every fourth or third stand in the same vertical plane whereas the branching otherwise is the usual 2_5 turn dextrorse. In some colonies a few extra branches issue from outside the typical spiral of branch-building coenenchyma generally being much smaller and slender than the normally placed ones. And one colony is distinguished by having two series of branches: the normal ones placed in a spiral 2_5 turn dextrorse and with orthostiche intervals of about 12 mm, and besides a series of smaller branches which are placed more scattered in an 1_5 spiral turning towards the left and with orthostiche intervals of about 25 mm.

The branches bifurcate generally only once or twice, some are, however, bifurcating three times, and branches may be found where bifurcating has taken place up to six times. The internodes

of first order measure generally 3-9 mm and the largest measure 16 mm. Zooids are found only exceptionally on these internodes, still there were in one case two zooids on the first internode, the one, however, placed at the bifurcation. The other internodes, apart from the outermost, have typically one zooid which, however, may be missing. If exceptionally there are two zooids, then the one is always placed at the point of bifurcation. The outermost internodes may reach a considerable size, the largest being about 75 mm, and are usually provided with several zooids. The largest number noticed was 19 zooids on a 73 mm long internode. The largest branches are about 80 mm and are found in colonies which probably may have measured 25-30 cm in height, often these branches bifurcate but once with the inner internode measuring about 5 mm and the outer about 75 mm. The angle of bifurcation is generally about 60-70° and may narrow down to about 50°.

The zooids measure in contracted state (verrucae) about 2.5 (2-3) mm in height having a width of 0.5-0.75 mm. Those of the uppermost part of the colonies are often all young ones but such are else only found on the outermost internodes inside the full-grown tip zooids.

It is noteworthy that the zooid in this species has about the lower, strongly spiculated half of its tentacles connected by a web, so that above the peristome a tube is formed which by contraction of the zooid protects the upper free pinnae-bearing parts of the tentacles (fig. 12).

The species may probably live in dense growths on the sea bottom, and one of the colonies shows very distinctly that it has been placed in the outer part of such a growth. The stem is bent, showing that the specimen during its growth has been pushed away by growing neighbour colonies, and the branches on that side of the specimen which has turned towards the growth are less than 30 mm in length and generally do not bifurcate, whereas those of the outer side of the colony measure up to 55 mm and bifurcate two or three times.

The sclerites in the coenenchyma of the base, stem and branches are thin and scale-like and of an oblong shape with rounded ends and usually a narrowed middle part. In the outer parts of the base they are placed fairly scattered (fig. 13), near

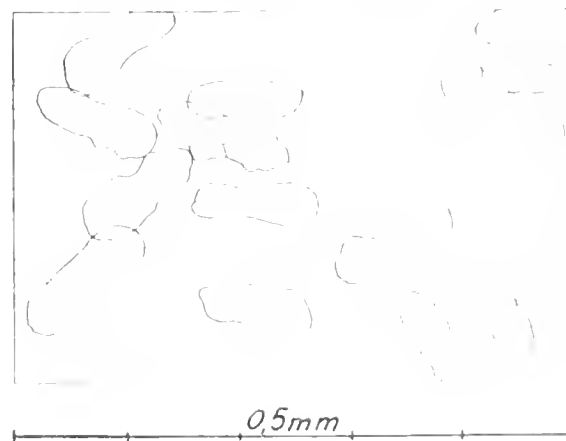


Fig. 13. *Chrysogorgia agassizii* (Verrill). Sclerites in natural position from outer part of basal disk.

the stem, however, in a rather close layer and here arranged with their longitudinal axis pointing towards the stem. The sclerites in stem and branches are longitudinally arranged in a rather close layer about one sclerite thick (fig. 14 a, b). The sclerite layer is, however, neither in stem nor in the branches continuous, but interrupted by scattered sclerite free spaces.

The largest scales are found in the base and the lower part of the stem and measure about 0.2 mm in length by about 0.06 mm in width at the ends and 0.1-0.5 mm at the narrowed middle

of the same length, some of the larger sclerites of this type are found in the lower part of the tentacle. In the lower part of the tentacle the largest measuring about 0.25 mm, but often narrower, measuring



FIG. 16. *Verrellia* sp. (a) section of coenenchyma of the lower part of the colony, about 15 cm high, showing the position of sclerites; (b-h) sclerites from the lower part of the tentacle.

about 0.15 mm. The sclerites are at the base more or less perpendicular to the longitudinal axis of the tentacle, but in the lower part of the tentacle they are in a very close layer. The sclerites are of two different types. The most common type is a spinous, slender, spindle-shaped sclerite with rounded ends and a maximum length of about 0.25 mm (fig. 16*b, c*). The other type of sclerites from the coenenchyma is a rod-like form (fig. 16*a, e*), which is found in the branches and stem of the colony. In the lower part of the tentacle the rod-like sclerites occur in a very close layer in the lower part of the tentacle nearly up to the attachment of the rod-like sclerites.

They occur in a very varying number, and whereas in some zooid bodies about half the sclerites are of the scale-like type, this type seems in other zooids to be almost absent. Intermediate stages between the two types of zooid body sclerites may be found.

The lower parts of the tentacles, i.e. the parts connected with each others by a web, are strongly armed with a row, up to 6-8 sclerites broad, of spinous spindles, resembling those from the zooid body, but usually being round and having pointed ends. One of the typical form is figured in fig. 15*c*, but some may be found which are thicker and of a more irregular shape and often with blunt ends. Their maximum size is about 0.5-0.08 mm. The sclerites decrease in size towards the free pinnulae-bearing part of the tentacle (about 0.2 mm), but are still round spindles (fig. 15*f, g*). In the lowermost free part of the tentacle the sclerites are of about the same type (fig. 15*b*) and also longitudinally directed, but else the tentacle dorsum is covered with a close layer of transversally arranged, flat and somewhat curved sclerites with a rugged surface (fig. 17*a-c*) the largest measuring about 0.3-0.07 mm, but most being much smaller.

The pinnulae generally number 6-12 pairs. They may sometimes be devoid of sclerites but are generally provided with a bundle of a few sclerites which usually are less than 0.25 mm in length and of a very varying shape. They have a rugged surface and are sometimes nearly rod-like, sometimes with flattened, often extended ends, and sometimes flat, resembling those of the tentacle dorsum (fig. 17*d-h*).

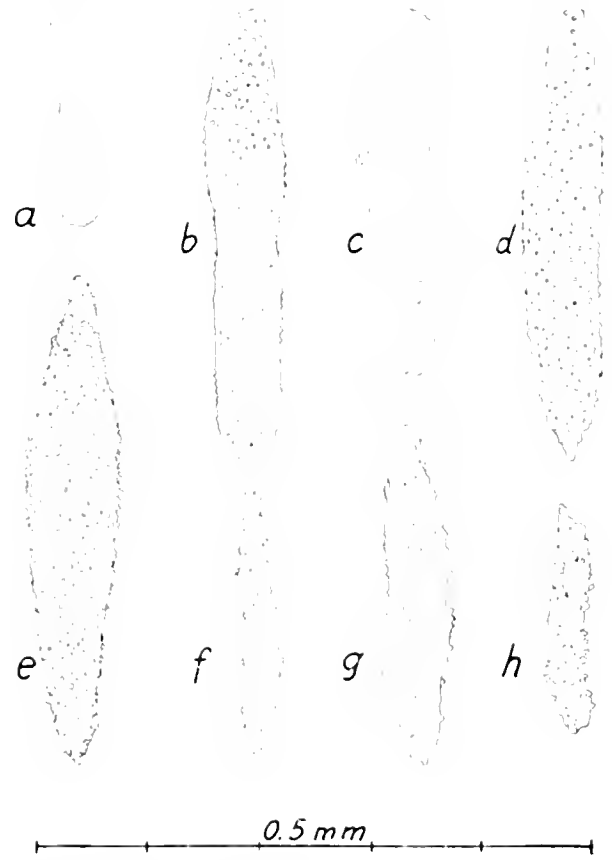


FIG. 17. *Verrellia* sp. (a) *Verrellia* sp. (a-d) sclerites of zooid body, (e-h) sclerites from tentacle dorsum of the lower half of tentacle, (e-h) sclerite from the lower part of pinnulae-bearing part of tentacle.

The peristome is provided with small, somewhat flattened sclerites measuring about 0.15-0.02 mm (fig. 17*q-l*). Generally there are only a few sclerites, placed very scattered, but sometimes a rather close layer of these sclerites may be found. In some zooids a few sclerites of the same type occur also in the upper part of the stomodaeum, mainly along the attachments of the mesenteries.

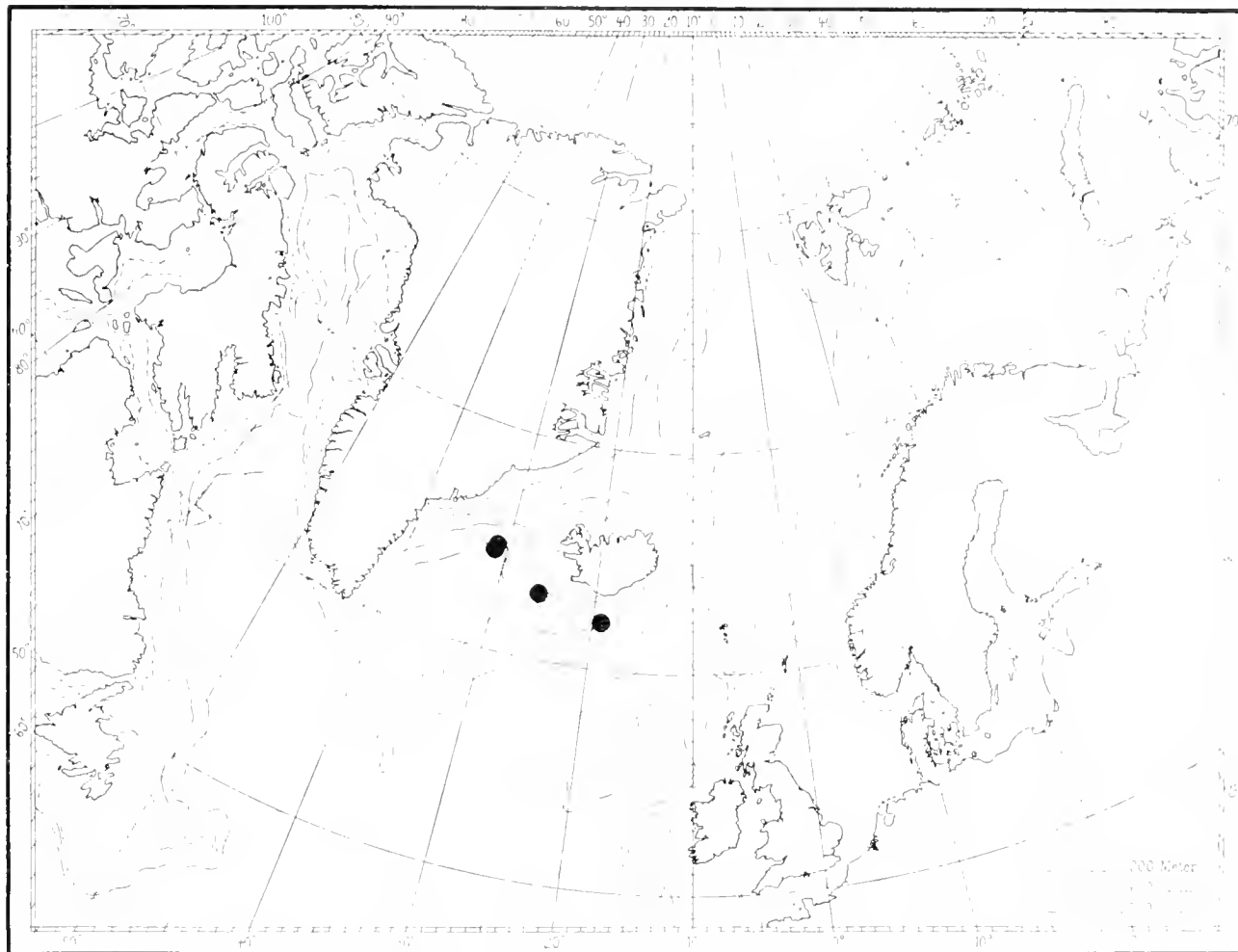


Fig. 46. *Chrysopaqa agassizi* (Verrill).

Discussion: The species, to which this collection of the "Ingolf" Expedition is referred, was described by VERRILL in 1883 on two specimens from 11 21'45" N, 65 35'30" W, in 2171 m, and apart from one point the material of the "Ingolf" agrees well with VERRILL's description and figures both of the general appearance of the colony and of the sclerites. VERRILL thus mentions both the two types of sclerites from the zooid body. He states, however, that the scale-like type is the most common whereas the opposite proved to be the case in the "Ingolf" collection; but this is a point to which no importance may be attached as DEICHMANN in 1936 (p. 231) not at all mentions this scale-like type from the zooids though she re-examined VERRILL's specimen. In the specimens taken by the "Ingolf" the scale-like sclerites also occurred in a very varying number. The measures, which VERRILL states for the sclerites, agree also well with those of the "Ingolf" collection. It is true that the greatest length, which he mentions for the scale-like type from the zooid body, is only 0.18 mm; on pl. 2 fig. 1 b, c a sclerite is, however, figured which after the magnification scale may have been 0.3 mm in length.

When VERRILL (p. 22) states "The tentacles are not entirely retractile" this may allude to the fact that their lower halves are coalesced.

DEICHMANN (p. 231) states that the zooids are "with few spicules", an information which does not agree with the conditions in the "Ingolf" specimens, and, also seems to be in contrast with VERRILL's figure pl. 2 fig. 1 a.

The specimen, upon which VERRILL's description was mainly based, lacks the base, but VERRILL (1883 p. 22) describes the base in the other specimen from the same locality as divided "into several long, divergent, irregular, palmate, flattened pro-

cesses" and he writes (1885 p. 511) about some other specimens from the same area in about 2162-2191 m, that "its root-like base is divided into short, flat, irregular branches." Also the two photos of *C. agassizi* reproduced by DEICHMANN (1936 pl. 34) show beautiful stoloniferous bases.

In regard to the shape of the bases there is thus a distinct difference between the specimens from off Georges Bank in the West Atlantic and those with basal disks from the North Atlantic south and west of Iceland. How much importance should be attached to this difference is, however, difficult to ascertain. In

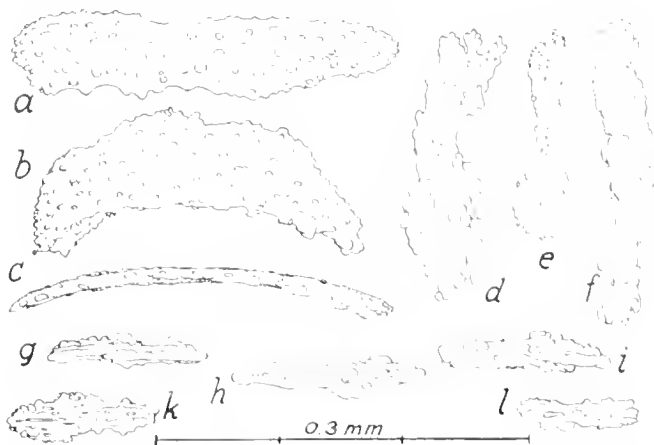


Fig. 17. *Chrysopaqa agassizi* (Verrill). a, c, sclerites from outer part of tentacle dorsum, c, shown from the side. d, f, sclerites of pinnular. g, i, sclerites from peristome and stomodaenum.

of the stem is found with the base broken off and has been anchored into the substrate. The stem is found with a tendency to curve downwards from the same colonies and is attached to a stoloniferous base.

It is interesting to note, however, that the base is not attached to the stem by a horny joint (as in *C. apassizi*) apparently originates from VERMILION (1927) and if it proves to correspond to the stem of *C. apassizi* of Georges Bank, then the differences between the specimens from the West Atlantic and those from the North Atlantic may not be greater than stated above and probably represent a subspecific distinction as in the latter species. On the contrary, it is sink into the calcareous base. KUKENTHAL (1919, p. 530) already doubted the identity of the specimens from the Pacific which NEUMANN (1912, p. 55) refers to as *C. apassizi*. KUKENTHAL points out that the coenenchyma in North Atlantic specimens is thick, with few sclerites, whereas in North Pacific specimens is thin and with numerous sclerites, and to this it may be added that NEUMANN describes the sclerites as "small, rounded" whereas in the coenenchyma of *C. apassizi* they are "large, smooth" and that the spiculation of the tentacles is quite different. In NEUMANN'S specimen "Double rows of thin, pointed spines extend along the basal parts of the dorsal surfaces of the tentacles, whereas there are spinous spindles in the basal parts of the tentacles of *C. apassizi*."

The *C. apassizi* which J. A. THOMSON (1927, p. 22) reports from the Azores off the Azores in 1425-1968 m. as DEICHMANN (1926, p. 222) supposes, most probably identical with *C. apassizi* is probably also J. A. THOMSON'S *C. flexilis* from off Cape Verde in 1477 m. and off the Azores in 3948 m. and his *C. orientalis* from off Cape Verde in 1311 m. may prove to belong to the same species. *C. apassizi* belongs to the group Spiculosa Wright & Stead, and since it has both scales and spindles in the zooid it seems to be most nearly related to the species of the *squarrosus*-group (see KUKENTHAL, 1919, p. 519) from which it, however, can be easily distinguished by its regular $\frac{2}{3}$ turn dextrorse branching of the stem and in generally having the branches bifurcated one, two or three times. The sclerites of the zooid in *C. apassizi* are moreover of a quite different type and are also considerably larger than those of *C. orientalis* and *C. morbi*. From the other species, which have the same $\frac{2}{3}$ turn dextrorse branching pattern, *C. apassizi* is easily distinguished partly by its spiculation of tentacles partly by its few internodes, which apart from the basal internode only bear one zooid each.

Description. The species is previously known from off Georges Bank in 2271-2391 m. and from the Bay of Biscay in 1757-1710 m. recorded by ROGER in 1896. The "Ingolf" localities are situated somewhat south and west of Iceland in 1192-2118 m. The specimens from the Azores and Cape Verde probably also found off the Azores in 1311-3948 m. Chart of distribution, fig. 46. Height of stem: 1425-2491 (3948) m.

Chrysogorgia campanula n. sp.

Fig. 47. 12 W. 2448-2460 (Ingolf), St. II, 1 colony.

Stem: 12 W. 2448-2460 (Ingolf), St. II, 1 colony. Stem somewhat purple. Branches bifurcated, pseudo-dichotomously branched with intervals between successive branches usually about 2 mm. Internodes 2.5-9 mm. Zooids standing at right angles to the branches (though also contracted) and one zooid on each internode, with the exception that where two zooids may occur. Size of zooids: 2.5-3.2 mm in height by 1.5-2.5 mm in width. Scales of stem measuring up

to about 3.5-4 mm. Scales of branches somewhat narrower, measuring up to about 3.5-4 mm. Scales of zooid body typically measuring about 3.5 mm in length having a width of 0.1 mm at the narrowed middle. Scales of tentacle dorsum typically about H-shaped, curved, transversely arranged and reaching a length of about 3.5 mm. No coidal papillae.

Type. In the Zoological Museum of Copenhagen.

Description. The single colony is somewhat damaged, it is without base and tip and measures in this condition about 85 mm in height, and it may be supposed that the entire colony has not been very much larger as 1) the interval between the scar of the broken off base and the lowermost scar of a branch measures about 1 mm, whereas the successive intervals between the nearest eight branches do not exceed 2 mm, and as 2) the diameter of the uppermost left part of the stem is only slightly thicker than the diameter of the proximal internodes of the upper branches.

The stem is slightly bent in zigzag fashion and gives rise to a branch at each bend. The branches are slightly ascending, forming an angle on about 60-70° with the longitudinal axis of the colony, and placed along the stem in a shrew-line which turns towards the left from the bottom upwards, every sixth branch being placed nearly exactly in the same vertical plane and the shrew-line turning twice round the stem between them. This regular mode of branching is, however, interrupted in one place in the lower part of the colony, where diametrically opposite a branch an extra very small one with only one zooid issues. The colony has altogether 26 branches, many of which, however, are broken off. The intervals between successive branches vary from 1.8-1.5 mm but are usually 3-3.5 mm. The shortest intervals are found in the lower part of the colony.

The branches themselves may be pseudo-dichotomously divided as many as six times, still they (the preserved ones) are usually bifurcated only two or three times and this applies especially to the lowermost. The most bifurcated branch is the eighth reckoned from the broken tip of the colony. Usually the branching takes place in such a manner that the first bifurcation and thus the intervals of second order lie in a nearly horizontal plane, whereas the third bifurcation and thus the intervals of fourth order lie in a nearly vertical plane. The angle of bifurcation is generally about 90° (varying from 80-100°) and also the angle between the branches and the stem is usually about 90°.

The size of the internodes may vary somewhat, e.g. the size of the internodes of first order (the inner) varies from about 3.5 to 8 mm and is usually about 5 mm. The size of the internodes of second order varies between 2.5 and 9 mm, and most frequently they are of about the same size as the corresponding inner internode. The outermost internode may be as long as 9 mm but is general much shorter. The shortest intervals are found in the lowermost preserved branch which is the sixth reckoned from the base. Though the pseudo-dichotomous branching is often symmetrical, this is not always the case. In one branch, e.g. the internodes of second order are 2.5 and 8.5 mm respectively, and whereas the one part of the branch bifurcates six times the other bifurcates only three times.

The zooids stand at right angles to the branches and are placed in a number of one on each internode. Only on the outermost internode there may be another, sometimes young, zooid besides the tip-zooid, whereas the inner internode may be quite without zooids, or, if there is one, this is usually placed at the point of bifurcation. Apart from the tip zooids, which are placed lateral to the ends of the branches so that the branch tips project beyond them, the zooids are very clearly distinguished from the branches. Generally speaking, the zooids are placed on the branches in such a manner that their mouths point upwards, but some are found directed downwards. Most of them are rather contracted and have their tentacles curved in over their mouth,

and in this condition the largest measure about 3.2 mm in height; the most common size is, however, 2.5-2.7 mm and some are also found which measure only 2 mm. The tip zooids are generally the largest (highest and broadest) having a diameter of 1.5-1.7 mm

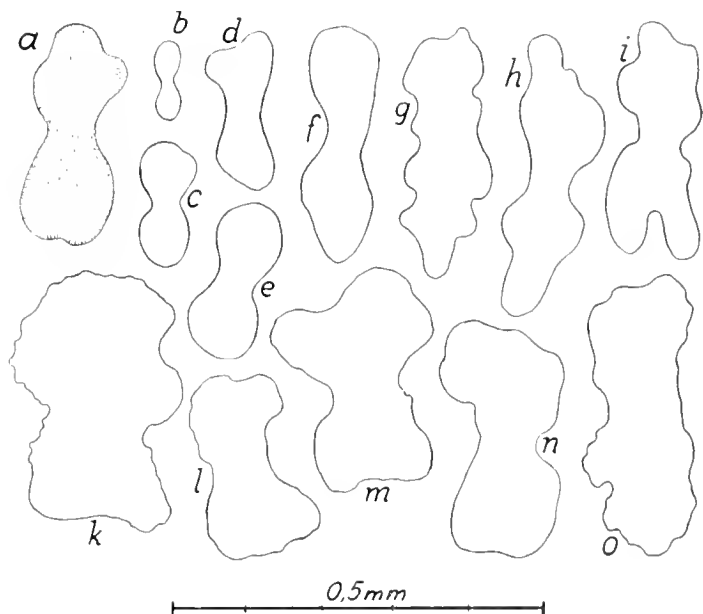


Fig. 48. *Chrysosporga campanula* n. sp. a-i, sclerites of branches, a-f, of the typical shape, a, with the finely toothed surface indicated, g-i, of the largest size, k-o, sclerites from lower part of stem.

(fig. 50 b). The other zooids are usually somewhat more slender, having a diameter of about 1 mm, most of them being contracted about the middle and thus becoming bell-shaped (fig. 50 a).

On most of the stem the coenenchyma is rubbed off. In the upper third of the colony, however, where it is partly preserved, there is a single zooid placed on the stem itself opposite a branch.

The basal part of the stem is about 1 mm in diameter. The coenenchyma is fairly thin and the round axis measures here about 0.7 mm in diameter. The diameter of the stem in the top of the colony is about 0.15 mm, and the diameter of the internodes of first order may be about 0.6 mm; it is, however, usually only 0.5-0.4 mm and sometimes even less. The outer internodes are

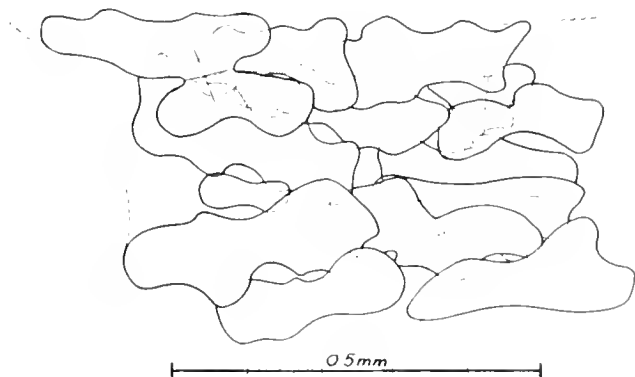


Fig. 49. *Chrysosporga campanula* n. sp. Sclerites from a branch in natural position.

much thinner, only about 0.2 mm in diameter. The axis of the branches is tapering, being hairfine at the point. In some of the preparations it was observed that the medulla runs quite eccentrically in the axis where a new branch issues.

No cnidal papillae were noticed.

The colour of the specimen in alcohol is white, and the colour of the axis is light yellow-brown and light blue-green iridescent.

The sclerites are all thin and scale-like and provided with numerous small spines which, especially at the ends, give the edges a finely denticulate appearance (as indicated on fig. 48 a), and they are all of about the same type, derived from an oblong form, with rounded ends and constricted middle, as shown in fig. 48 b, c.

In the coenenchyma of the stem and branches the scales form a close overlapping layer which in the lower part of the stem may be three sclerites thick, but in the branches is mainly only one sclerite thick. The scales from the stem may reach a length of 0.4 mm and a width of 0.25 mm; besides these, however, also scales of the same type as those from the branches are found which, though they may reach the same length as the largest from the stem, are always much narrower. These sclerites of

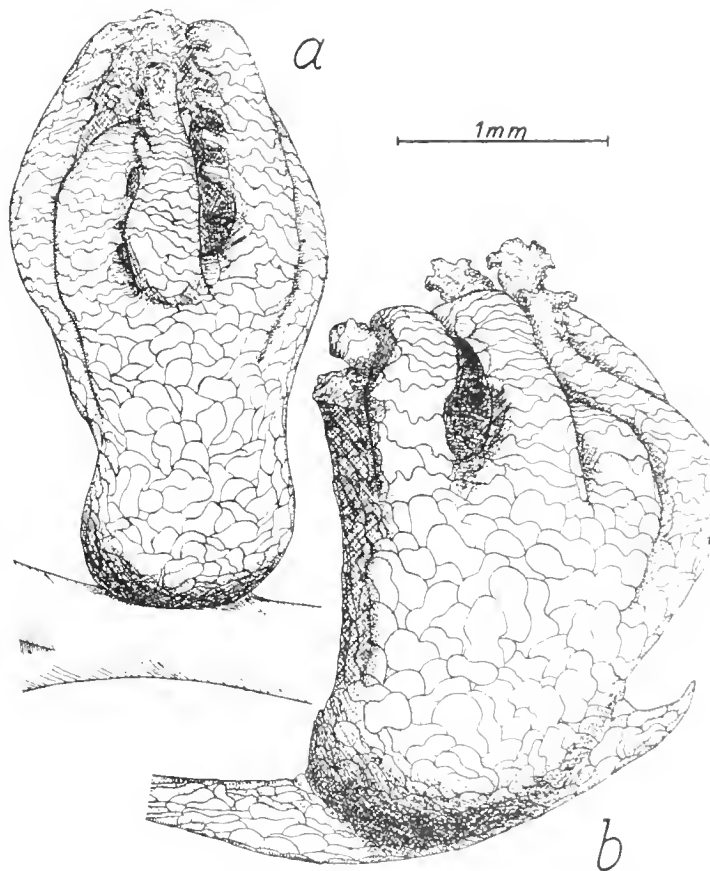


Fig. 50. *Chrysosporga campanula* n. sp. Two zooids, a, a tip-zooid. Some zooids are, though of the same height, somewhat slimmer than that figured fig. 2, a.

branches are typically beautifully S-shaped (fig. 48 a, b) and measure generally about 0.35 mm in length by 0.08-0.1 mm in width at the extended oval ends, and about 0.5 mm in width at the narrowed middle. The broadest of the sclerites have with a length of about 0.4 mm, a width of about 0.13 mm at the ends, and somewhat less at the middle. Several of the scales are, however, rather irregular, with forked ends or with more narrowed parts, but even in the most diverging sclerites the S-shaped type is always discernible. Scales of fairly small size may be found sporadically between the others, and both in stem and branches twin- and quadruplets-formations are common. The scales are all placed in the longitudinal direction of the stem or the branches (fig. 49).

The scales of the zooid body overlap each other mainly in a direction from the bottom upwards. They are at the base of the zooid placed transversally (i. e. lengthwise to the stem), become in the upper portion of the base more obliquely directed, and are in the main zooid body arranged quite irregularly (fig. 50 and 51). The zooid scales are beautifully S-shaped and fairly broad with nearly circular ends. They measure generally about 0.33 mm in length by about 0.16 mm in width at the ends and



FIG. 51. *Chirogasterella pinnata* n. sp. Sclerites of zooid body in natural position. Some of the forked ends possibly represent developmental stages of the sclerites, not found in the zooid body.

only two or three fully developed ones (fig. 51). It is thought that many of the eight other full grown sclerites are in a young one, about 0.8 mm in length, which has just hatched out. In the young zooid it was, however, the forked ends from the zooid body were somewhat shorter than from the full grown zooids, only measuring 0.4 mm in length.

In the zooids, the sclerites are transversely arranged, as in figure 51. The scales in the above mentioned young zooid are arranged about the middle, also however in a transverse shape. Those of the full grown zooids are arranged in a transverse shape, typically broad x shaped with the forked ends reaching down on the sides (fig. 51). Figure 52 shows some of the sclerites of the tentacle

dorsum reach, however, only down on the one side of the tentacle and have correspondingly only the one end extended and forked. The tentacles are thus laterodorsal covered with the ends of the dorsal sclerites, but besides sclerites may be found arranged lengthwise to the tentacle. The lateroventral part of the tentacle is bare or has small transversely arranged sclerites.

The tentacles have six to ten pairs of pinnulae which are provided with sclerites of a very varying size. These sclerites have typically a forked end which reaches into the tentacle among the lateral sclerites, and a rather long, "gutter"-formed part which reaches into the pinnule with the open part of the "gutter" pointing downwards (fig. 52 *b*). They may reach a length of about 0.5 mm but are usually much smaller. In each pinnule there may be a few sclerites; generally, however, only one of these is of a larger size, and very often all the pinnulae sclerites are rather small. Many of them, especially the smaller ones, are not "gutter" shaped.

The peristome is devoid of sclerites.

Discussion. The species belongs to the group *Squamosea tapuan* Versluys and seems to be most nearly related to *C. acanthella* (Wright & Studer) from which it may, however, be distinguished by its larger zooids, the absence of cnidal papillae, and its much closer layer of scales in the coenenchyma.

Distribution. Only known from one locality, in the Danmark Strait south of the Wyville Thomson Ridge, Depth 2418 m. Chart of distribution, fig. 37.

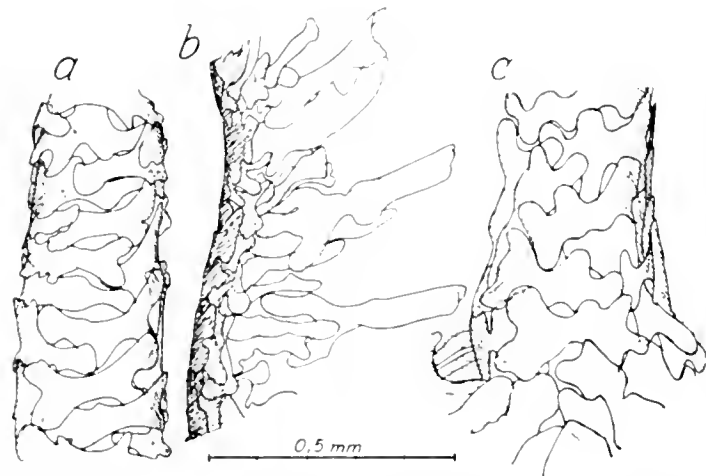


FIG. 52. *Chirogasterella pinnata* n. sp. Sclerites in natural position in tentacles. *a*, tentacle dorsum; *b*, part of tentacle from the side showing the sclerites in the pinnulae; *c*, tentacle base.

Family Isididae Sinder 1878

Diagnosis. Vide KUKENTHAL 1919, p. 556

Genus Acanella Gray 1870.

Acanella Gray 1870, p. 573

***Acanella arbuscula* (Linn.) Sinder 1878**

- 1878, p. 573, fig. 216, pl. 31, fig. 1, 1a.
- 1878, p. 573, fig. 216, pl. 31, fig. 1, 1a.
- 1878, p. 573, fig. 216, pl. 31, fig. 1, 1a.
- 1878, p. 573, fig. 216, pl. 31, fig. 1, 1a.
- 1878, p. 573, fig. 216, pl. 31, fig. 1, 1a.
- 1878, p. 573, fig. 216, pl. 31, fig. 1, 1a.
- 1878, p. 573, fig. 216, pl. 31, fig. 1, 1a.
- 1878, p. 573, fig. 216, pl. 31, fig. 1, 1a.
- 1878, p. 573, fig. 216, pl. 31, fig. 1, 1a.
- 1878, p. 573, fig. 216, pl. 31, fig. 1, 1a.

Description. Vide VERRILL 1883 and KUKENTHAL 1919.

Material

- Off. Marthas Vineyard, 375-1130 m. U. S. F. Comm.
- 66 19 N. 56 28 W. 410 m. Wandel.
- 64 21 N. 28 50 W. 1181 m. 3 5 C. Ingolf St. 10.
- 63 06 N. 56 00 W. 2258 m. 2 10. 21
- 63 30 N. 54 25 W. 1096 m. 3 30. 25.
- 65 11 N. 55 12 W. 791 m. 3 50. 28.
- 62 00 N. 22 38 W. 1629 m. 2 90. 39.
- 62 00 N. 21 36 W. 1591 m. 3 30. 49.
- 63 33 N. 15 02 W. 595 m. 5 90. 55.
- 62 40 N. 19 05 W. 1596 m. 4 00. 63.
- 62 06 N. 19 00 W. 1960 m. 3 10. 61.
- 61 30 N. 22 30 W. 1836 m. 3 00. 67.
- 62 06 N. 22 30 W. 1587 m. 3 10. 68.

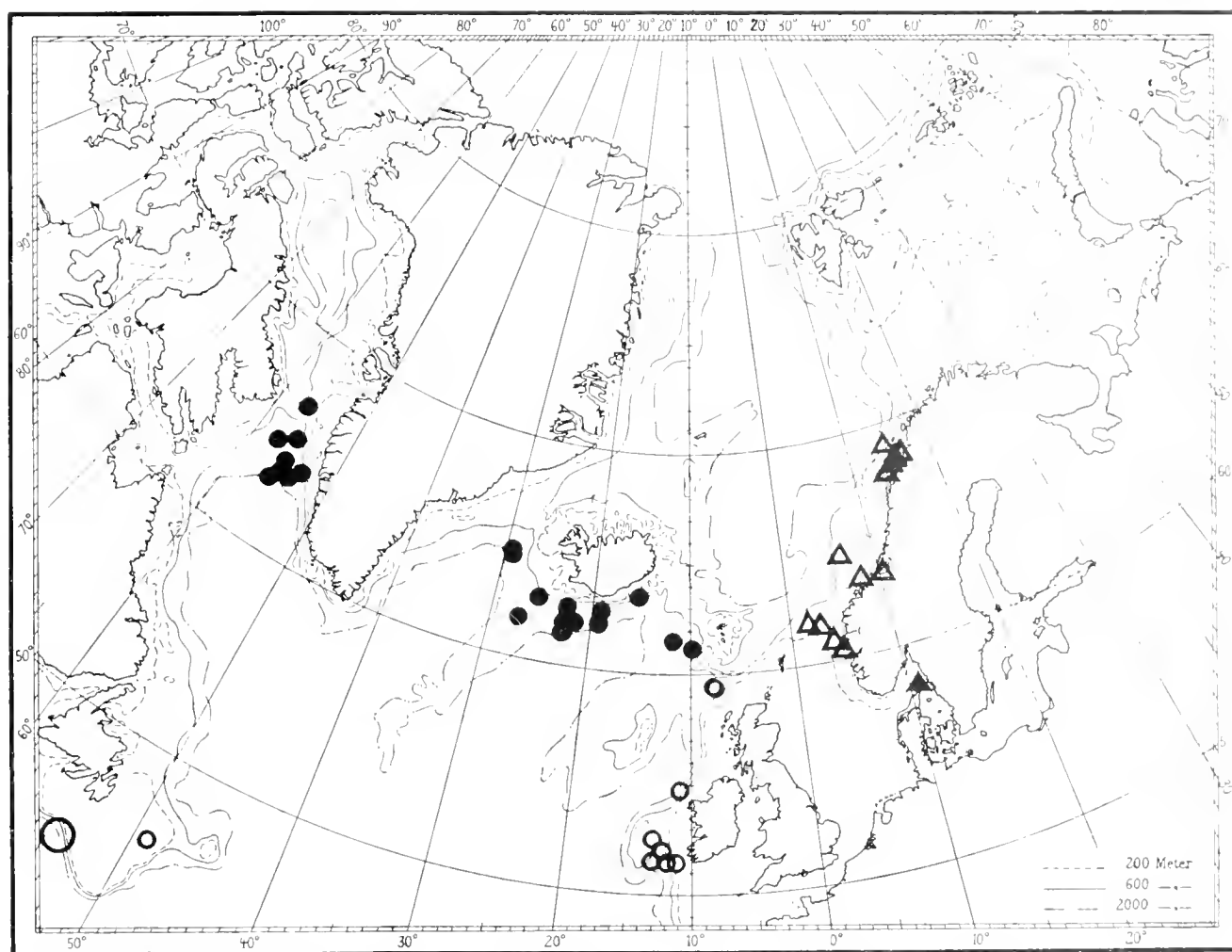


Fig. 53. ● *Acanella arbuscula* (Johnson),
▲ *Isidella lofotensis* M. Sars.

62 40' N, 22 17' W, 1109 m, 3.9 C, "Ingolf" St. 69.
61 14' N, 27 00' W, 913 m, 6.1 C, - 81.
62 58' N, 25 24' W, 1192 m, 1.8 C, - 81.
61 45' N, 29 06' W, 1070 m, 4.4 C, - 90.
61 08' N, 9 16' W, 847 m, "Michael Sars" St. 79.
61 05' N, 55 20' W, 1100 m, "Tjalfe" St. 337.
64 10' N, 56 37' W, 750 m, - 102.
63 54' N, 53 15' W, 988 m, - 428.
63 51' N, 53 15' W, 988-1100 m, "Tjalfe" St. 429.
63 36' N, 55 45' W, 1200 m, 3.28 C, (1150 m) "Godthaab" St. 479.

Apart from the Nephthyidae *Acanella arbuscula* is by far the most common of the North-Atlantic Octocorals. At the stations where it was secured by the "Ingolf" it usually occurred in fairly great quantities.

JUNGERSEN (1915 p. 1186), besides the above-mentioned material, also mentions specimens taken by the Danish research vessel "Thor" south-west and south of Iceland, and south-west of Ireland at a depth of 2100 m. This material is, however, not preserved.

Distribution: The species is very common in the North-Atlantic Ocean north of 0° latitude and south of the submarine ridges which connect Scotland, Iceland, Greenland and Baffin Land. Undoubtedly the species will prove to occur in the deeper part of the whole Atlantic Ocean. Chart of distribution, fig. 53.

Vide further JUNGERSEN 1915 and 1917.

Bathymetrical distribution: about 375-3173 m.

Genus: *Isidella* Gray 1870.

Diagnosis: Vide KÜKENTHAL 1919, p. 561.

***Isidella lofotensis* M. Sars 1868.**

Isis hippuris (pars) Gummerus, 1768, p. 70-71, fig. 8.

Isidella lofotensis M. Sars, 1868, p. 22-23.

Mopsea borealis M. Sars, 1869, p. 250.

Mopsea borealis G. O. Sars, 1872, p. 50-57, pl. 5 figs. 1-23.

Isidella hippuris Grieg, 1891, p. 3-11, figs. 1-22.

Acanella hippuris Broch, 1912 c, p. 39-45, figs. 26-29.

Isidella hippuris Jungersten, 1917, p. 24-25.

lofotensis Kükenthal, 1924, p. 415-416.

Pax, 1936, p. 259.

Carlgren, MS.

For further references vide JUNGERSEN 1917 and KÜKENTHAL 1924.

Description: Vide Broch 1912 c.

Material:

Lofoten, 560 m, Cotype to *Mopsea borealis* M. Sars, 1 colony.
58 06' N, 9 00' E, 660-120 m, "Thor" St. 1571, 1 colony.

Distribution: The species is at present only known from off the Scandinavian coast from Lofoten in the north to the Skagerrak in the south (the above mentioned localities represent the northernmost and the southernmost finds). The westernmost find hitherto made is in 61 52 30' N, 4 12 06' E. Chart of distribution, fig. 53.

Bathymetrical distribution: About 200-700 m.

Zoogeographical and Ecological Remarks.

On the zoogeography and ecology of these animals we have to base our remarks on the study of the collection of the Gorgonacea in the Davis Strait. The following remarks may be made:

1. The zoogeography of Octocorals is not known with certainty. It is well stated after experiments in aquaria, that *Anthothela grandiflora* was entirely carnivorous, and JENKINS (1904) found specimens of the Pennatulid *Virgata* from the Davis Strait (between Denmark and Sweden) feeding on the remains of *Radiocipes quacalis* (JENKINS, 1904, p. 159). In specimens of *Radiocipes quacalis* taken from the dorsal cavities of several zooids, but also from the cavities of zooids examined in other traces of food were not detected. In many zooids are found stuffed with minute Foraminiferans especially Globiferina. A considerable amount is found not only in the stomachs but also in the stomodaeum, and in the space between the zooids. Without doubt, it may be supposed that it is the zooids adjacent to the colony in question was caught by the disturbance of the material, and on this disturbance the Octocoral may retract its zooids. We may guess that the Octocorals mainly feed upon living prey and that the zooids, on the fact that the single zooids only in the cavities of zooids have access to food, are the reasons why the remains of prey are found in the zooids, but we cannot say anything exact about the question.

It is also possible that detritus falling down from above may be the main access to part of the food, and though the zooids in the bottom may not be supposed to be able to catch detritus, such might no doubt be a valuable source of food at great depths where living prey may be scarce. WILSON (1889, p. 141) also pointed out that the zoogeographical deep-sea species of Octocorals are "supposed to eat all the zooids have an 'equal' opportunity of catching what falls from above". It may, therefore, be supposed that in *Chrysosporus aquassizi* many zooids may have access to food, that their mouth is directed downwards.

2. The zoogeography of the reproduction of the Octocorals. Of the species treated in this paper *Anthothela grandiflora*, *Capnella glomacata*, *Prumnella pumescens*, *Radiocipes challoperii*, *R. quacalis* and *Acanthopora armata* have been found with larvae in the Davis Strait. *Anthothela grandiflora* occurs referably in the Davis Strait together with larvae in the Davis Strait.

3. The zoogeography of the attachment. It has been shown that *Anthothela grandiflora*, *Anthothela* species, *Capnella glomacata* and *Prumnella pumescens* are attached to irregular objects, and that *Chrysosporus aquassizi* and *Acanthopora armata* are attached to regular objects. *Anthothela* species, like *Anthothela grandiflora*, are attached to regular expansion of the stem, and *Acanthopora armata* anchors the

Among the Gorgonacea there are both species which are attached to solid subjects 1) by an expansion of the base of the stem, e. g. *Anthothela grandiflora*, 2) by a basal disk formed by extension of the horny axis, e. g. *Acanthopora armata* and 3) by a calcareous basal disk, e. g. *Prumnella pumescens*, and species which are anchored into the mud by a calcareous root-like base as e. g. *Acanthopora armata*. It is noteworthy that also one of the Gorgonaceans provided with a calcareous base, viz. *Chrysosporus aquassizi*, is rather independent on the condition of the bottom in a similar way as the Nephthyidae. Its base may be developed either as a calcareous disk attached to a solid object or as a hollow calcareous formation which comprises a lump of clay and in this manner anchors the colony into the bottom, and moreover the West Atlantic form of *Chrysosporus aquassizi* is found with a root like calcareous base such as in *Acanthopora armata*.

A few concluding remarks may be made about the zoogeographical position of the deep-sea species dealt with in this paper. Undoubtedly, these species are extremely stenothermic and therefore quite unable to live in lesser depths where the temperature is subject to any significant variations during the year. However, the Nephthyidae form an exception from the general rule as these forms occasionally, although only in arctic areas, are met with in rather shallow water.

As mentioned in the introduction several species of Octocorals not dealt with in this paper have been found in deep waters off the coast of Ireland. Apart from *Acanthopora armata* and *Anthothela grandiflora* (if identical with *Gymnosata bathybius* Kent) none of the species found in this area have been taken by the "Ingolf" or other research vessels investigating the North Atlantic Ocean south of the Faroes and Iceland, and the whole distribution supports the supposition that most of the forms found off the coast of Ireland require temperatures not below about 8° C. Thus their distribution in the eastern part of the Atlantic Ocean is, generally speaking, limited in the north at the course of the 8° isotherm in 1000 m depth.

Several of the species dealt with in this paper have only been found in the North Atlantic Ocean south of the ridges in the Davis Strait, the Danmark Strait and between Iceland and the Faroes, viz. *Clavularia quacalis*, *Cl. levidensis*, *Cl. canastella*, *Capthopodaria capelli*, *Felastula septentrionalis*, *Acanthopora armata*, *Prumnella pumescens*, *Chrysosporus aquassizi*, *C. campanula*, *Radiocipes challoperii*, *R. quacalis*, and *Acanthopora armata*. These species have mainly been met with at depths greater than about 1000 m. *Acanthopora armata* and *Acanthopora armata* have, however, also been taken in depths of only a few hundred metres and the first-mentioned species possibly even in only 20 m depth. Several of the species mentioned here have a rather wide distribution in the North Atlantic Ocean and probably several of them may prove to occur in the whole abyssal Atlantic, at any rate down to about 3000 m. They have mainly lived at temperatures about 3° C. and though most of them have also been found at somewhat lower temperatures, none have been found at temperatures below zero. The temperatures at which they have been taken vary from 1.5-6.1°C.

South of and on the submarine ridges in the North-Atlantic Ocean several species are also found which besides are distributed in the warm area of the Norwegian Sea, i. e. along the Norwegian coast between the Skagerrak in the south and Lofoten in the north. These species are *Anthelia borealis*, *Anthomastus grandiflorus*, *Anthothela grandiflora*, *Paramuricea placomus*, and *Trachymuricea kükenthali*, and in this group probably also the following boreal Atlantic species should be ranged, viz. *Stenogorgia borealis* which, however, has not hitherto been found east of the Faroes, and *Stenogorgia rosca* and *Isidella lofotensis* which, however, have not hitherto been found far away from the Scandinavian coast and always at moderate depths (down to about 100 m and 700 m respectively), and probably also *Anthelia fallax* which, however, is only known from the Trondheim Fjord. The temperatures which are required by this group of species seem on an average to be slightly higher than those required by the preceding group, i. e. between about 3° and 8° C. The Danish expeditions in the North Atlantic have taken them at temperatures between 2.1° C. and 7.2° C.

Also *Clavularia arctica* belongs to the abyssal Atlantic + boreal Atlantic species. This species is, however, also found north of Lofoten in the Barents Sea, here, however, only at depths less than 120 m (55-120 m), whereby the cold water of the Norwegian Sea has been avoided. South of Lofoten the bathymetrical distribution of the species is between 180 and 300 m.

Two species, viz. *Paragorgia arborea* and *Primonot resedaeformis*, are found besides in the boreal Atlantic also in the boreal Pacific but only at moderate depths (down to about 850 m). The temperatures required by these species seem mostly to vary between 5° and 8° C.

The only one of the Octocorals of the groups dealt with in this paper which for the present may be said to be true arctic is *Ceratocaulon wuellerstorfi* which is known from some localities on the northern side of the Wyville Thomson Ridge mainly at temperatures a little below zero (-0.75° C.), though in one locality at 0° C.

Also the three Nephthyidae and *Clavularia alba* have been found at temperatures below zero, and at present *Caprella glomerata*, *Gersonia rubiformis* and *Cl. alba* are the only ones which have been taken in the main part of the Norwegian Sea. These species have a fairly wide distribution, *Gersonia rubiformis*, which also has been found in the northern Pacific, is arctic + subarctic and circumpolar, *Caprella glomerata* is arctic + arctic boreal, and *Cl. florida* is arctic boreal, *Cl. alba* is mainly arctic. They are usually met with at temperatures between about -1° C. and -6° C. However, *Cl. florida* is mainly found in the temperature interval between 3° and 6° C. (also found at temperatures up to 8.1° C.), whereas the other species seem to prefer somewhat lower temperatures, mainly between -1° and -4° C.

Of the species dealt with in this paper only *Sarcodictyon catenatum*, *Aegonium digitatum* and *Stenogorgia rosca* are normally met with in the littoral zone. The zoogeographical position of *Ale. digitatum* has been discussed by the author in a small paper of 1913, and probably similar conditions apply to *Sarcodictyon catenatum*. *Stenogorgia rosca* seems to be an endemic Scandinavian species.

During the "Ingolf" Expedition Octocorals were secured at altogether 66 stations, and at 62 of the stations species of the orders dealt with in this paper were taken, yet larger forms, i. e. Gorgonaceans, were only taken at 21 of the stations.

- elegans, Paramuricea, Grieg 1887 — Paramuricea placomus (L.).
 fallax, Anthelia, Broch 1912.
 Anthothela grandiflora var., Molander 1918 — Anthelia fallax Broch.
- filiformis, Clavularia, Hickson 1891 — Sarcodictyon catenatum Forbes.
 — Rhizoxenia, M. Sars 1856 — Sarcodictyon catenatum Forbes.
- flabelliformis, Gorgonia, Gummerus 1765 — Paramuricea placomus (L.).
- flava, Duya, Danielssen 1887 — Capnella florida (Rthk.).
 — Paraspongodes, May 1900 — Capnella florida (Rthk.).
- flavescens, Eumephtya glomerata var., Kükenthal 1906 — Capnella glomerata (Verr.) f. flavescens.
 Eumephtya, Molander 1915 — Capnella glomerata (Verr.) f. flavescens.
 Nephthya, Danielssen 1887 — Capnella glomerata (Verr.) f. flavescens.
 — Paraspongodes, May 1899 — Capnella glomerata (Verr.) f. flavescens.
- florida, Capnella, Broch 1939 — Capnella florida (Rthk.).
 — Duya, Koren & Danielssen 1883 — Capnella florida (Rthk.).
 Eumephtya, Kükenthal 1906 — Capnella florida (Rthk.).
 Gersenia, Marenzeller 1878 — Gersenia rubiformis (Ehrbg.).
 Gorgonia, Rathke 1806 — Capnella florida (Rthk.).
 Paraspongodes, May 1900 — Capnella florida (Rthk.).
- fricci, Briareum, Koren & Danielssen 1853 — Paragorgia arborea (L.).
- frigida, Clavularia, Danielssen 1887 — Gersenia rubiformis (Ehrbg.).
 Duya, Danielssen 1887 — Capnella florida (Rthk.).
 Paraspongodes, May 1900 — Capnella florida (Rthk.).
- fruticosa, Eumephtya, Kükenthal 1906 — Gersenia rubiformis (Ehrbg.).
 Gersenia, Molander 1915 — Gersenia rubiformis (Ehrbg.).
 — Paraspongodes, May 1898 — Gersenia rubiformis (Ehrbg.).
 — Voeringia, Jungersen 1887 — Gersenia rubiformis (Ehrbg.).
- fruticosum, Aleyonium, M. Sars 1860 — Gersenia rubiformis (Ehrbg.).
- glacialis, Duya, Danielssen 1887 — Capnella florida (Rthk.).
 Paraspongodes, May 1900 — Capnella florida (Rthk.).
 Paraspongodes, May 1898 — Gersenia rubiformis (Ehrbg.).
- globosa, Paraspongodes, May 1900 — Gersenia rubiformis (Ehrbg.).
- glomerata, Ammothea, Carter 1880 — Capnella glomerata (Verr.).
 — Capnella, Broch 1939 — Capnella glomerata (Verr.).
 Drifa, Verrill 1922 — Capnella glomerata (Verr.) f. lütkeni + f. flavescens.
 Eumephtya, Verrill 1869.
- gracilis, Lepidogorgia, Verrill 1881 — Radicipes gracilis (Verr.).
 — Radicipes (?) Jungersen 1916 — Radicipes gracilis (Verr.).
- grandiflora, Anthothela, Verrill 1879 — Anthothela grandiflora (M. Sars).
 Anthothela, Molander 1915 — Anthothela grandiflora (M. Sars).
- grandiflorum, Briareum, M. Sars 1856 — Anthothela grandiflora (M. Sars).
- grandiflorus, Anthomastus, Verrill 1883.
- griegii, Clavularia, F. J. Madsen 1911.
- griegi, Paraspongodes, May 1900 — Gersenia rubiformis (Ehrbg.).
- groenlandica, Eumephtya, Molander 1915 — Capnella glomerata (Verr.) f. groenlandica.
- huppuris, Acanella, Broch 1913 — Isidella lofotensis M. Sars.
 Isidella, Grieg 1891 — Isidella lofotensis M. Sars.
 Isis, Gummerus 1788 — Isidella lofotensis M. Sars.
- hyalina, Drifa, Danielssen 1887 — Capnella glomerata (Verr.) f. flavescens.
 Eumephtya, Kükenthal 1906 — Capnella glomerata (Verr.) f. flavescens.
 Eumephtya flavescens var., Molander 1915 — Capnella glomerata (Verr.) f. flavescens.
- hyalina, Haimea, Koren & Danielssen 1883 — Aleyonium digitatum (L.).
 Paraspongodes, May 1899 — Capnella glomerata (Verr.) f. flavescens.
- hyalinum, Sympodium, Grieg 1887 — Gersenia rubiformis (Ehrbg.).
- ingolfi, Cyathopodium, F. J. Madsen 1911.
- irramosa, Danielssenia, Grieg 1887 — Stenogorgia rosea Grieg.
 Leptogorgia, Molander 1919 — Stenogorgia rosea Grieg.
- islandica, Drifa, Danielssen 1887 — Capnella glomerata (Verr.) f. lütkeni.
 Eumephtya glomerata var., Kükenthal 1906 — Capnella glomerata (Verr.) f. lütkeni.
 Paraspongodes, May 1899 — Capnella glomerata (Verr.) f. lütkeni.
- jan-mayeni, Voeringia, Danielssen 1887 — Gersenia rubiformis (Ehrbg.).
- jungersenii, Primmocella, F. J. Madsen 1911.
- kükenthali, Muriceoides, Aurivillius 1931 — Trachymuricea kükenthali (Broch).
 Paramuricea, Broch 1912 — Trachymuricea kükenthali (Broch).
 Trachymuricea, Deichmann 1936 — Trachymuricea kükenthali (Broch).
- lepadifera, Gorgonia, Linné 1767 — Primmocella resedaeformis (Gunn.).
 Primmocella, Lamouroux 1816 — Primmocella resedaeformis (Gunn.).
- levidensis, Clavularia, F. J. Madsen 1911.
- lofotensis, Isidella, M. Sars 1868.
- loricata, Gersenia, Marenzeller 1878 — Gersenia rubiformis (Ehrbg.).
 Gersenia fruticosa forma Molander 1915 — Gersenia rubiformis (Ehrbg.).
 Paraspongodes, May 1900 — Gersenia rubiformis (Ehrbg.).
- lütkeni, Ammothea, Marenzeller 1878 — Capnella glomerata (Verr.) f. lütkeni.
 — Paraspongodes, May 1899 — Capnella glomerata (Verr.) f. lütkeni.
- lütkeni, Aleyonium, Verrill 1879 — Capnella glomerata (Verr.) f. flavescens (?).
 Eumephtya, Verrill 1883 — Capnella glomerata (Verr.) (= Gersenia rubiformis (Ehrbg.)).
- margaritaceum, Sympodium, Grieg 1887 — Aleyonium digitatum (L.).
 Clavularia, May 1900 — Aleyonium digitatum (L.).
- massa, Sympodium, Ehrenberg 1831 — Aleyonium digitatum (L.).
- membranaea, Gersenia fruticosa forma, Molander 1915 — Gersenia rubiformis (Ehrbg.).
- mirabilis, Eumephtya, Kükenthal 1906 — Gersenia rubiformis (Ehrbg.).
 Gersenia, Molander 1915 — Gersenia rubiformis (Ehrbg.).
 Paraspongodes, May 1900 — Gersenia rubiformis (Ehrbg.).
 Voeringia, Danielssen 1887 — Gersenia rubiformis (Ehrbg.).
- multiflora, Duya, Verrill 1922 — Capnella florida (Rthk.).
- multiflorum, Aleyonium, Verrill 1879 — Capnella florida (Rthk.).
- nodosa, Paragorgia, Koren & Danielssen 1883 — Paragorgia arborea (L.).
- nordenskiöldi, Organidus, Danielssen 1887 — Gersenia rubiformis (Ehrbg.).
- normani, Acanella, Verrill 1878 — Acanella arbuscula (Johns.).
- norvegicum, Aleyonium, Broch 1912 — Aleyonium digitatum (L.).
 Erythropodium, Broch 1912 — Aleyonium digitatum (L.).
 Sympodium, Koren & Danielssen — Aleyonium digitatum (L.).
- pacifica, Primmocella, Kimoshita 1907 — Primmocella resedaeformis (Gunn.).
- pallida, Gersenia fruticosa var., Molander 1915 — Gersenia rubiformis (Ehrbg.).
- palmata, Paraspongodes, May 1900 — Gersenia rubiformis (Ehrbg.).

List of Literature.

The list includes only the more important works and the works quoted. References to the rest of the literature on the northern Ocean are to be found in the works referred to in the text.

- APPELLÖF, A. 1895. Faunistiske Undersøgelser i Herlofjorden. Bergens Mus. Aarbog 1894-95.
- ASHWORTH, J. H. 1898. The Stomodaeum, mesenterial filaments and endoderm in *Xenia*. Proc. Royal Soc. London, vol. 53.
1899. The Structure of *Xenia hicksoni* n. sp. with some observations on *Heteroxenia elisabethae* Kolliker. Quart. Journ. Micr. Sci., vol. 12, n. s.
- AURIVILLIUS, M. 1926. Några fynd av sällsynta havsevertebrater. Fauna och Flora, vol. 21.
1931. The Gorgonarians from Dr. Sixten Bock's Expedition to Japan and Bonin Islands 1914. Kungl. Svenska Vetensk. Akad. Handl., ser. 3, vol. 9, no. 1.
- BIELSCHOWSKY, EVA. 1918. Eine Revision der Familie Gorgoniidae. (Dissertation) Breslau.
1929. Die Gorgonarien Westindiens. Kap. 6. Die Familie Gorgoniidae, zugleich eine Revision. Zool. Jahrb. Abt. Syst., Suppl. 16, Heft 1.
- BOCK, SIXTEN. 1938. The Aleyonarian Genus *Bathyalcyon*. Kungl. Svenska Vetensk. Akad. Handl., ser. 3, vol. 16, no. 5.
- BOURNE, G. C. 1900 a. Coelenterata, in Lankester's Treatise on Zoology, pt. II.
- 1900 b. On the genus *Lemnalia* Gray with an account of the branching-systems of the order Aleyonacea. Trans. Linn. Soc. London, vol. 7.
- BROCH, HJ. 1912 a. Coelenterés du fond. Duc. D'Orléans: Campagne arctique de 1907.
- 1912 b. Die Aleyonaceen des Kolafjordens. Travaux Soc. Imp. Nat. St. Pétersbourg, vol. 11.
- 1912 c. Bemerkungen über *Clavularia arctica* (M. Sars). Det Kgl. Norske Vid. Selsk. Skr. 1911.
- 1912 d. Die Aleyonarien des Trondhjemsfjordes. I. Aleyonacea. Det Kgl. Norske Vid. Selsk. Skr. 1911.
- 1912 e. Die Aleyonarien des Trondhjemsfjordes. II. Gorgonacea. Det Kgl. Norske Vid. Selsk. Skr. 1912.
- 1913 a. Arktiske Aleyonarier i Tromsø Museum. Tromsø Mus. Aarshefter, vol. 34.
- 1913 b. Die Aleyonarien des Trondhjemsfjordes. III. Pennatulacea und IV. Biogeographische Übersicht. Det Kgl. Norske Vid. Selsk. Skr. 1912.
1916. Aleyonaria. Results of Dr. E. Mjöberg's Swedish Scientific Expedition to Australia 1910-1913, No. XI. Kungl. Svenska Vetensk. Akad. Handl., vol. 52, no. 11.
1928. Aleyonarians, with a Systematic-Biogeographical Discussion of the Northern Eumephytha-Species. The Norweg. North Polar Exp. "Maud" 1918-1925, Ser. Res., vol. 5, no. 7.
1929. Die Oktokorallen der Arktis. Fauna Arctica, vol. 5.
1932. Über einige geographisch interessante Fundstellen von Aleyonarien und Hydrokorallen im nördlichen Stillen Ocean. Explor. des mers de l'URSS, fasc. 17, 1933.
1935. Oktokorallen des Nordlichsten Pazifischen Ozeans und ihre Beziehungen zur Atlantischen Fauna. Avhandl. utgitt av Norske Vidensk. Akad. i Oslo, I. Mat. Nat. Kl. 1935, no. 1.
- BROCH, HJ. 1939. Some South African Shallow Water Octocorals. Kgl. Fysiograf. Sällsk. i Lund. Forhandl., vol. 9, no. 6.
1940. Anthozoa, mainly from Pacific Waters, collected by USSR Exped. 1930-1932. Explor. des mers de l'URSS, vol. 23, 1940.
- CARLQVIST, O. MS. Koraldyr. Danmarks Fauna.
- DANA, J. D. 1846. Zoophytes. United States Exploring Expedition during the Years 1838-1842.
1859. Synopsis of the Report on Zoophytes of the U.S. Exploring Expedition under C. Wilkes 1838-1842.
- DANIELSEN, D. C. 1887. Aleyonida. The Norwegian North Atlantic Exped. 1876-1878. XVII.
- DEICHMANN, ELISABETH. 1936. The Aleyonaria of the Western Part of the Atlantic Ocean. Mem. Mus. Comp. Zool. Harvard, vol. 53.
- DOXS, C. 1914. Et og andet fra mine ekskursioner. Tromsø Mus. Aarsberetning for 1913.
1932. Om Nord-Norges korallsamfund. Det Kgl. Norske Vid. Selsk. Forhandl., vol. 5, no. 4.
- EUREMBERG, C. G. 1834. Die Coralleniere des rothen Meeres. Kgl. Akad. Wiss. Berlin.
- FORBES, E. & GOODE, F. 1853. On some remarkable marine Invertebrata new to the British Seas. Trans. Royal Soc. Edinburgh, vol. 20.
- GISLÉN, T. 1930. Epibioses of the Gullmar Fjord II. Kristinebergs Zoologiska Station 1877-1927.
- GRAY, J. E. 1869. Notes on the fleshy Aleyonoid Corals. Ann. Mag. nat. Hist., ser. 4, vol. 3.
- GRUO, I. A. 1887. Bidrag til de norske Aleyonarier. Bergens Mus. Aarsberetning 1886.
1888. To nye Corullarier fra den norske Kyst. Bergens Mus. Aarsberetning 1887.
1891. Tre nordiske Aleyonarier. Bergens Mus. Aarsberetning 1890, no. 2.
1891. Bidrag til Kjendskaben om de nordiske Aleyonarier. Bergens Mus. Aarbog 1893, no. 2.
1907. Invertébrés du Fond. Duc. D'Orléans: Campagne arctique de 1907. Belgique 1905.
- GRONVIG, I. E. 1763. Om en Søvext, alvegne, som findes med Froehuse, *Gorgonia reschotensis*. Det Trondhjemske Selsk. Skr. Anden Deel. Kbhvn.
1768. Om nogle Norske Coraller. Det Kgl. Norske Vidensk. Selsk. Skr. Fjerde Deel. Kbhvn.
- HERDMAN, W. A. 1883. On the Structure of *Sarcodictyon*. Proc. Roy. Phys. Soc. Edinburgh, vol. 8.
- HICKSON, S. J. 1891. A Revision of the Genera of the Aleyonaria (Stolonifera), with a Description of one new Genus and several new Species. Trans. Zool. Soc. London, vol. 13, pt. 13.

1861. Gorgonacea. *Explor. Voy. U.S. Mail Steamer "Albatross" Good Hope, 1849-50*. U.S. Geol. Surv. Geol. Surv. Atlantic, vol. 3.
1862. Gorgonacea. *Explor. Voy. U.S. Mail Steamer "Albatross" Good Hope, 1849-50*. U.S. Geol. Surv. Geol. Surv. Atlantic, Proc. Zool. Acad., vol. 1, p. 10.
1863. A new Genus of the Family Nereidae, with a Revision of the Genus *Squilla*. *U.S. Geol. Surv. Geol. Surv. Rept. Exped. 1828-29*, vol. 1, p. 10.
1864. Description of some New Corals from Madeira. *Zool. Surv. London*, 1862.
1862. Description of Two Corals from Madeira, belonging to the Genus *Porolithothamnium*. *Proc. Zool. Soc. London*, 1862.
1867. *1847. A History of the British Zoophytes.*
1868. *1847. H. F. L. 1887. Korallthierier. Aeyonidae, Dijnuphidae, Pterothulidae, Tubarien.*
1862. Corals from Wrangell, in the arktisk Aeyonide. *Vid. Medd. Dansk. For. Kobenhavn*, 1891.
1864. Pterothulida. *The Danish "Ingolf" Exped.*, vol. 5, no. 1.
1865. Aeyonaria, Antipatharia og Madreporaria. *Consp. Faun. Gronland. Medd. om Gronland*, vol. 23.
1866. The Aeyonaria of East Greenland. *Medd. om Gronland*, vol. 24.
1867. Aeyonaria and Madreporarian Corals in the Museum of Bergen, collected by Fram Expedition 1898-1900 and by the Michael Sars, 1900-1906. *Bergens Mus. Aarbok* 1915-16, 1918. Om Korallthierier, Anthomastus, og Udviklingen af dens Korallform. *Forhandl.*, vol. 16, skand. naturforsker-selsk., 1916.
1867. Anthomastus. *The Danish "Ingolf" Exped.*, vol. 5, no. 11.
1868. *1863. Koraller i Svenska Hav. "Jorden Rigtig"*
1865. Götterborgs Mus. Arstryck 1865.
1866. Götterborgs Mus. Arstryck 1866.
1867. Götterborgs Mus. Arstryck 1867.
1868. Nya stotag och gamla problem. *Göteborgs Handels- och Sjöfartstidning*, no. 200, 221, 233.
1869. Götterborgs Mus. Arstryck 1869.
1869. W. Sars. 1870. On two New Genera of Aeyonoid Corals taken in the recent Expedition of the Yacht *Norma* to the Coast of Spain and Portugal. *Quart. Journ. Micr. Sci.*, no. 10.
1869. J. & P. Dyer. 1857. Beskrivelse over nogle nye Gortterater. *Fauna Litt. Norvege*, Helt. 5.
1884. New Aeyonidae, Gorgonidae og Pennatulidae tilhørende Norge. *Fauna Bergens Mus.*
1869. P. L. 1860. Aeyonaria. *Zoology of the Faroes*.
1862. Aeyonaria, Antipatharia and Madreporaria. *The Geol. Surv. Rept. 1828. Medd. om Gronland*, vol. 79.
1866. Coelenterata, Ctenophora and Chaetognatha. *Medd. om Gronland*, vol. 194.
1869. *1869. The Zoology of Iceland*, vol. 2, pt. 7.
1869. W. 1892. Versuch einer Revision der Aeyonaceen. *1869. Fauna der Nordsee. Zool. Jahrb. Abt. Syst.*, vol. 15.
1869. Versuch einer Revision der Aeyonaceen. II. Die Familie *Nereidae*. *1869. Zool. Jahrb. Abt. Syst.*, vol. 19.
1869. *1869. Wiss. Ergeb. Deutsche Tiefsee Exped.*, vol. 1.
1869. *1869. U.S. Geol. Surv. Rept. Exped. (1898). Wiss. Geol. Surv. Rept. Exped. (1898). Vol. 8. Abt. Helgoland.*
1869. *1869. Revisión de Aeyonariae. II. Die Familie Nereidae. 1869. Zool. Jahrb. Abt. Syst.*
1869. *1869. Die Gattungen Eunephthya, Vermetophylla, Murexella. Zool. Jahrb. Abt. Syst.*
1869. *1869. Aeyonariae des Sibirischen Eismeer. 1869. Zool. Jahrb. ser. 8, vol. 18.*
1869. *1869. Zool. Jahrb. der Isididae. Zool.*
1869. *1869. System und Stammesgeschichte der Primnoidae. Zool. Anz.*, vol. 16.
1869. *1869. Die Gorgonarien Westindiens. Zool. Jahrb. Suppl.*, 11.
1869. *1869. Gorgonaria. Wiss. Ergeb. Deutsche Tiefsee Exped. 1898-1899*, vol. 30.
1869. *1869. Gorgonaria. Das Tierreich*, Lief. 17.
1869. *1869. Beiträge zur Kenntnis der Polypen. Verhandl. phys. med. Gesellsch. Würzburg, N. Folge*, vol. 2.
1869. *1869. Zur Kenntnis der Aeyonarien Gattung Telesto Linn. Zool. Jahrb. Suppl.*, 11.
1869. *1869. Systema naturae*, Ed. 10.
1869. *1869. A Revised Catalogue of the Anthozoa and Calycozoa of Greenland. T. Rupert Jones. Manual of the Nat. Hist. Greenland*
1869. *1869. The Danish Octocorals from the Waters inside Skagen. Zoogeographical and Ecological Remarks. Vid. Medd. Dansk. nat. For.*, vol. 107.
1869. *1869. Die Coelenteraten, Echinodermen und Würmer d. K. K. Österreichisch-Ungarischen Nordpol-Expedition. Denkschr. Kais. Akad. Wissch. mat. nat. Cl.*, vol. 35.
1869. *1869. Poriferen, Anthozoen, Ctenophoren und Würmer von Jan Mayen. Die Österreich. Polart. Jan Mayen*, vol. 3.
1869. *1869. Ueber die Sarcophytum benannten Aeyoniden. Zool. Jahrb.*, vol. 1.
1869. *1869. Aeyonaceen von Ost-Spitzbergen, nach der Ansicht der Herren Prof. Dr. W. Kükenthal und Dr. A. Walther im Jahre 1889. Zool. Jahrb. Abt. Syst.*, vol. 11.
1869. *1869. Beiträge zur Systematik und Chorologie der Aeyonaceen. Jenaische. Ztschr. Nat. wiss.*, vol. 33, 1900.
1869. *1869. Die arktische, subarktische und subantarktische Aeyonaceenfauna. Fauna Arctica*, vol. 1.
1869. *1869. Histoire naturelle des Coralliaires ou Polypes prominent dits. Vol. 1.*
1869. *1869. Northern and Arctic Invertebrates in the Collection of the Swedish State Museum. VII. Aeyonacea. Kgl. Svenska Vetensk. Akad. Handl.*, vol. 51, no. 11.
1869. *1869a. Die Aeyonaceen des Eisfjords. Kgl. Svenska Vetensk. Akad. Handl.*, vol. 51, no. 11.
1869. *1869b. Membranöse Ausbildung der Kolomen bei Gorgonacea nebst einigen Bemerkungen über Rhizoxenna alba Grieg. Arkiv f. Zool.*, vol. 11, no. 21.
1869. *1869c. Der Kelch als systematischer Character bei den Aeyonaceen. Arkiv f. Zool.*, vol. 11, no. 22.
1869. *1869. Bemerkungen über Leptogorgia irramosa (Grieg). Arkiv f. Zool.*, vol. 12, no. 5.
1869. *1869. Die Octactmarien. Further Zool. Res. Swedish Antaret. Exped. 1901-1903.*
1869. *1869. Annual communities on soft bottom areas in the Gullnar fjord. Kristinebergs Zoologiska Station 1877-1927.*
1869. *1869. Coelenterata, Echinodermata, Bryozoa. Ber. d. Comm. z. wiss. Untersuch. d. deut. Meere, Jahrg. 1.*
1869. *1869. Mollusken, Würmer, Echinodermen und Coelenteraten. Die zweite deutsche Nordpolart. vol. 2.*
1869. *1869. Hydrographical and Biological Investigations in Norwegian Fiords. Bergens Mus.*
1869. *1869. Mofjordens Naturforhold. Det Kgl. Norske Vidensk. Selsk. Skr.*, 1906.
1869. *1869. Havstrømmene og den norske marine Fauna. Det Kgl. Norske Vidensk. Selsk. Skr.*, 1915.
1869. *1869. Faunistic Notes on Marine Evertbrates VII. On the Distribution of some Aeyonarian Corals in Northern Norway. Det Kgl. Norske Vidensk. Selsk. Forhandl.*, vol. 3.
1869. *1869. Descriptions of the Aeyonaria collected by the U.S. Bureau of Fisheries Steamer Albatross in the Vicinity of the Hawaiian Islands in 1902. Proc. U.S. Nat. Mus.*, vol. 31.
1869. *1869. The Murexidae. Siboga Exped.*, vol. 13 b.
1869. *1869. Anthozoa. Tierwelt d. Nord- und Ostsee. Lief. 30.*

- PREXANT, M. 1927. Faune marine sessile Roscoff. Trav. Stat. Biol. Roscoff. 1927.
- QUOY & GAIMARD. 1833. Voyage de découvertes de l'Astrolabe, vol. 4.
- ROULE, L. 1896. Résultats scient. de la Campagne du "Candan" dans les Golfe de Gascogne. Ann. Univ. de Lyon, vol. 26.
- ROXAS, A. H. 1933. Philippine Aleyonaria. The Families Cornulariidae and Xeniidae. Philipp. Journ. Sci., vol. 50.
- RYLOV, V. 1916 a. Aleyonacea, gesammelt von der Expedition des Schwimmenden Wissenschaftlichen Meeresinstituts im Jahre 1921. Ber. Wiss. Meeresinst., vol. 2, Moskva.
- 1926 b. Hydroidea et Aleyonaria, collectionnés dans la mer de Barentz par la Station Biologique du Mourman pendant les croisières le long du méridien de Kola en 1921. Ann. Mus. Zool. Acad. Sc. de F.U.R.S.S.
- SARS, G. O. 1872. On Some Remarkable Forms of Animal Life from the Great Depths off the Norwegian Coast.
- SARS, M. 1851. Beretning om en i Sommeren 1849 foretagen zoologisk Reise i Lofoten og Finnmarken. Nyt Mag. for Naturvidensk., vol. 6.
- 1856. Nye Polyper. Fauna litt. Norvegiae. Heft. 2.
- 1860. Bemærkninger over norske Coelenterater. Videnskabselsk. Forhandl. Christiania 1860.
- 1868. Om Echinodermter og Coelenterater fundne ved Lofoten. Videnskabselsk. Forhandl. Christiania 1867.
- STEPHENS, JANE. 1909. Aleyonarian and Madreporarian Corals of the Irish Coasts. Fisheries, Ireland, Sci. Invest., 1907, V.
- STIASNY, G. 1937. Revision der Scleraxonia mit Ausschluss der Melitodidae und Coralliidae. Die Gorgonacea der Siboga-Expedition. Suppl. II. Siboga Exped., vol. 13 b, 8.
- 1939 a. Gorgonaria von Portugal. Arch. Mus. Bocage, vol. 10.
- 1939 b. Gorgonaires du Maroc (Cote Atlantique). Bull. Soc. sc. nat. du Maroc, Rabat, vol. 19.
- 1939 c. System der Octocorallia. Zool. Medd., vol. 21.
- 1940. Aleyonaria und Gorgonaria von Südafrika. Aus der Sammlung des South African Museum Capetown. Verh. Nederl. Akad. Wet., Ser. 2, vol. 39.
- 1942. Aleyonaria und Gorgonaria aus dem Golf von Neapel. Pubbl. Staz. Zool. Napoli, vol. 19.
- 1943. Die Gorgonarien-Familie Acanthogorgiidae Kükenth. & Gorz. mit besondere Berücksichtigung des Material der Siboga-Expedition. Zool. Anz., vol. 141.
- STUDER, TH. 1878. Übersicht der Anthozoa Aleyonaria, welche während der Reise S. M. S. Gazelle um die Erde gesammelt wurden. Mtsber. Kgl. Preussisch. Akad. Wiss. Berlin 1878.
- 1901. Aleyonaires provenant des campagnes de l'Hirondelle. Rés. camp. scient. Albert I Prince d. Monaco, vol. 20.
- THEEL, HD. 1907. Om utvecklingen af Sveriges zoologiska hafstation Kristineberg och om djurlivet i angränsande haf och fjordar. Arkiv f. Zool., vol. 1, no. 5.
- THOMSON, J. A. 1910. Note on Eunephthya glomerata, Verrill, from the Faeroe Channel. Proc. Roy. Phys. Soc. Edinburgh, vol. 18.
- 1927. Aleyonaires provenant des Campagnes scientifiques du Prince Albert I de Monaco, vol. 73.
- THOMSON, J. A. & DEAN, LAURA. 1931. The Aleyonacea of the Siboga Expedition with an Addendum to the Gorgonacea. Siboga Exped., vol. 13 d.
- THOMSON, J. STUART. 1911. The Aleyonaria of the Cape of Good Hope and Natal. Proc. Zool. Soc. London, 1911.
- VERRILL, A. E. 1861. Revision of the Polyps of the Eastern Coast of the United States. Mem. Boston Soc. Nat. Hist., vol. 1.
- 1865. Synopsis of the Polyps and Corals of the North Pacific Exploring Expedition. . . . With Descriptions of some additional Species from the West Coast of North America. Proc. Essex Inst., vol. 1.
- 1866. On the Polyps and Echinoderms of New England, with Descriptions of new species. Proc. Boston Soc. Nat. Hist., vol. 10.
- 1868 a. Notes on the Radiata in the Museum of Yale College, with Descriptions of new Genera and Species. Trans. Connecticut Acad. Arts and Sci., vol. 4.
- 1868 b. Critical remarks on the Halyeonid Polyps in the Museum of Yale College, with Descriptions of new Genera. Amer. Journ. Sc. and Arts, ser. 2, vol. 15.
- 1869. Critical Remarks on Halyeonid Polyps. Amer. Journ. Sc. and Arts, ser. 2, vol. 17.
- 1878. Notice on recent additions to the Marine Fauna of the eastern coast of North America. Amer. Journ. Sc. and Arts, ser. 3, vol. 16.
- 1879. Notice of recent additions to the marine Invertebrata, of the northeastern coast of America, with descriptions of new genera and species and critical remarks on others. Proc. U. S. Nat. Mus., vol. 2.
- 1882. Notice of the remarkable Marine Fauna occupying the outer banks of the Southern coast of New England. Amer. Journ. Sc., ser. 3, vol. 23.
- 1883. Report on the Anthozoa, and on some additional Species dredged by the "Blake" in 1877-1879, and by the U. S. Fish Commission Steamer "Fish Hawk" in 1880-1882. Bull. Mus. Comp. Zool. Harvard, vol. 11.
- 1884. Notice on the remarkable Marine Fauna occupying the outer banks off the Southern Coast of New England. Amer. Journ. Sc., ser. 3, vol. 27.
- 1885. Results of the explorations made by the Steamer "Albatross", off the northern coast of the United States, in 1883. U. S. Comm. of Fish. Fisheries, vol. 11.
- 1922. Aleyonaria and Actinaria. Rep. Canadian Arctic Exped. 1913-18, vol. 3, pt. G.
- VERSEVELDE, J. 1910. Studies on Octocorallia of the Families Briareidae, Paragorgonidae and Anthothelidae. Temminckia, vol. 5.
- VERSLUYS, J. 1902. Die Gorgoniden der Siboga Expedition. I. Die Chrysogorgonidae. Siboga Exped., vol. 13.
- 1906. Die Gorgoniden der Siboga Expedition. II. Die Prionoidae. Siboga Exped., vol. 13 a.
- 1907. Pseudochladochonus hicksoni n. g. n. sp. Siboga Exped., vol. 13 c.
- WEDDIN, E. P. & STUDER, TH. 1889. Report on the Aleyonaria collected by H. M. S. Challenger. Rep. Sc. Res. Challenger, vol. 31.

Plate I.

Telestula septentrionalis n. g., n. sp.

- Fig. 1 Schematic sketch of a retracted zooid cut longitudinally in half (on plate textfig. 6b), I, II and III showing where the three sections drawn on the plate have been cut.
- Fig. 2 Transverse section through the distal part of the contracted zooid showing how completely the contracted lower part of the anthocodia closes for the opening to the inverted anthocodia and the coelenteric cavity.
- Fig. 3 Transverse section through the zooid a little above the peristome of the inverted anthocodia showing the tentacles and the arrangement of papillae and sclerites.
- Fig. 4 Transverse section through the "stem" showing the eight "stem" canals.

cc. ectoderm

ec. endoderm

mes. mesogloia

sc. mesenteries

t. tentacles

p. papillae

sc. cavities left by sclerites

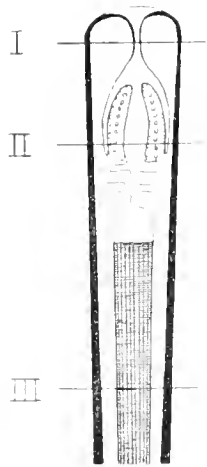
after decalcification.

st. c. "stem" canals.

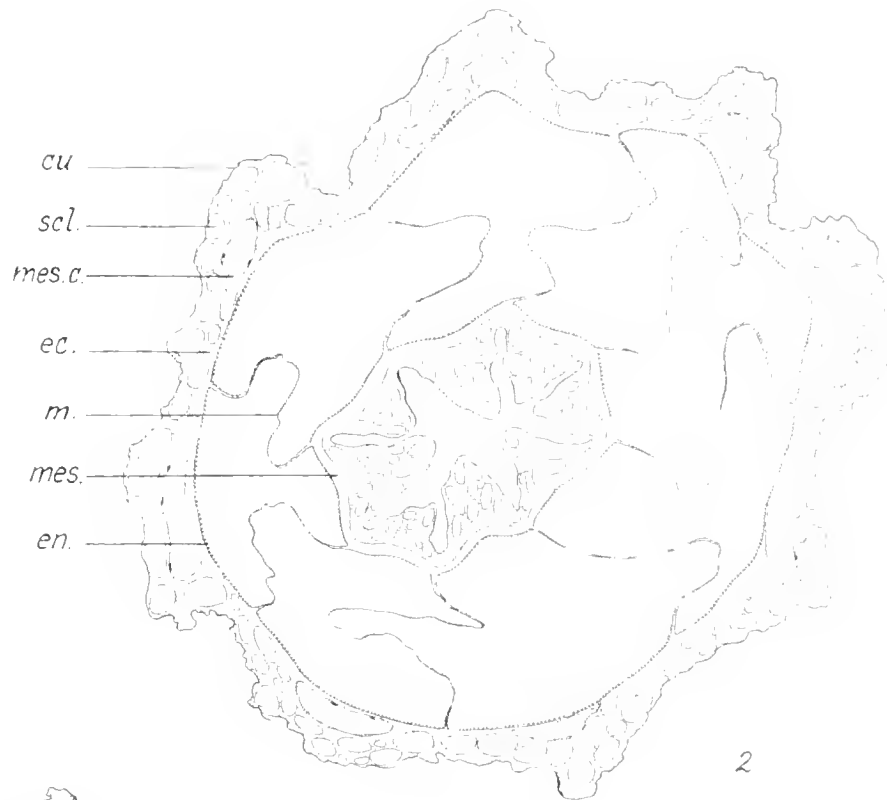
ec. c. other endodermal canals.

mes. c. mesogloial cell canals.

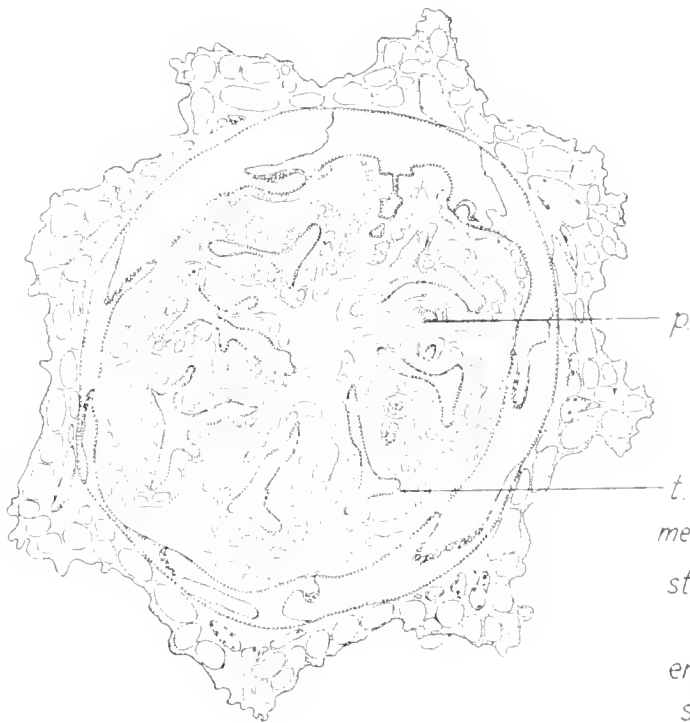
ca. cuticula



1



2



3



4

THE INGOLF-EXPEDITION

1895-1896

THE LOCALITIES, DEPTHS, AND BOTTOMTEMPERATURES OF THE STATIONS

Station Nr.	Date	Lat. N.	Long W.	Depth in m.	Bottom-temp.	Station Nr.	Date	Lat. N.	Long W.	Depth in m.	Bottom-temp.	Station Nr.	Date	Lat. N.	Long W.	Depth in m.	Bottom-temp.
1895						1896											
1	11-V	62° 30'	8° 21'	249	7.2	24	25-VI	63° 06'	56° 00'	2258	2.4	45	11-V	61° 32'	9° 43'	1211	4.17
2	12 -	63° 04'	9° 22'	493	5.3	25	26 -	63° 30'	54° 25'	1096	3.3	46	- -	61° 32'	11° 36'	1356	2.40
3	- -	63° 35'	10° 24'	512	0.5			63° 51'	53° 03'	256		47	12 -	61° 32'	13° 40'	1789	5.25
4	13 -	64° 07'	11° 12'	446	2.5	26	- -	63° 57'	52° 41'	64	0.6	48	- -	61° 32'	15° 11'	2165	3.17
5	- -	64° 40'	12° 09'	292				64° 37'	54° 24'	205		49	13 -	62° 07'	15° 07'	2109	2.91
6	16 -	63° 43'	14° 34'	170	7.0	27	1-VII	64° 54'	55° 10'	740	3.8	50	- -	62° 43'	15° 07'	1921	5.13
7	17 -	63° 13'	15° 11'	1139	4.5	28	- -	65° 11'	55° 42'	791	3.5	51	15 -	64° 15'	14° 22'	128	7.32
8	19 -	63° 56'	24° 40'	256	6.0	29	5 -	65° 34'	54° 31'	428	0.2	52	- -	63° 57'	13° 32'	791	7.87
9	20 -	64° 18'	27° 00'	555	5.8	30	10 -	66° 50'	54° 28'	41	1.05	53	16 -	63° 15'	15° 07'	1497	4.08
10	- -	64° 24'	28° 50'	1484	3.5	31	11 -	66° 35'	55° 54'	166	1.6	54	18 -	63° 08'	15° 40'	1391	5.9
11	21 -	64° 34'	31° 12'	2448	1.6	32	11 -	66° 35'	56° 38'	599	3.9	55	19 -	63° 33'	15° 02'	595	5.9
12	22 -	64° 38'	32° 37'	1958	0.3	33	12 -	67° 57'	55° 30'	66	0.8	56	- -	64° 00'	15° 09'	128	7.57
13	- -	64° 47'	34° 33'	1171	3.0	34	18 -	65° 17'	54° 17'	194		57	20 -	63° 37'	13° 02'	669	3.4
14	- -	64° 45'	35° 05'	331	4.4	35	- -	65° 16'	55° 05'	682	3.6	58	- -	64° 25'	12° 09'	397	5.8
15	4-VI	66° 18'	25° 59'	621	0.75	36	28 -	61° 50'	56° 21'	2702	1.5	59	- -	63° 00'	11° 16'	84	5.1
16	5 -	65° 43'	26° 58'	471	6.1	37	29 -	60° 17'	54° 05'	3229	1.4	60	21 -	63° 00'	12° 27'	34	5.9
17	16 -	62° 49'	26° 55'	1103	3.4	38	30 -	59° 12'	51° 05'	3524	1.3	61	- -	63° 05'	13° 06'	104	5.4
18	17 -	61° 44'	30° 29'	2137	3.0	39	9-VIII	62° 00'	22° 38'	1629	2.9	62	31 -	61° 48'	10° 41'	156	7.9
19	18 -	60° 29'	34° 14'	2949	2.4	40	- -	62° 00'	21° 36'	1591	3.3	63	1-VII	62° 00'	13° 00'	139	4.0
20	20 -	58° 20'	40° 48'	3192	1.5	41	12 -	61° 39'	17° 10'	2345	2.9	64	- -	62° 06'	13° 00'	155	4.6
21	21 -	58° 01'	44° 45'	2505	2.4	42	14 -	61° 41'	10° 17'	1175	0.4	65	2 -	61° 00'	13° 00'	397	5.9
22	22 -	58° 10'	48° 25'	3474	1.4	43	- -	61° 42'	10° 11'	1215	0.5	66	- -	61° 33'	10° 00'	254	5.4
23	24 -	60° 43'	56° 00'	1737	0.5	44	- -	61° 42'	9° 36'	1026	4.8	67	- -	61° 33'	12° 00'	1836	5.0

Station	Date	Time	Loc		Dist m	Dist to p	Sta No.	Date	Lat. N	Lon. W	Depth in m	Bot- tom- temp					
			Lat. N	Lon. W													
100	4	22	64	44	3	32	1838	44	118	24-VII	68	27'	8	20'	1996	1.0	
101	9	26	64	24	3	34	1414	44	119	25	-	67	53'	10	49'	1992	1.0
102	4	24	64	56	36	49	384	44	120	-	-	67	29'	11	32'	1666	1.0
82			66	31	33	45	491		124	-	-	66	59'	13	41'	996	0.7
103	27	64	65	44	39	39	1416	24	122	26	-	66	42'	13	44'	217	1.8
104	9	28	65	24	29	00	1384	42	123	28	-	66	52'	15	40'	273	2.0
105	9	42	65	28	27	39	847	55	124	-	-	67	40'	15	40'	932	0.6
106	4	28	65	38	26	27	260	59	125	29	-	68	08'	16	02'	1373	0.8
107	31	29	66	43	25	54	352	64	126	-	-	67	49'	15	52'	552	0.5
108	4	29	66	25	14	02	111	64	127	2-VIII	66	35'	20	05'	83	5.6	
109	18	44	66	23	12	05	1911	67	128	-	-	66	59'	20	02'	365	0.6
110	19	22	66	23	19	26	1412	69	129	3	-	66	35'	23	47'	229	6.5
111	5	45	66	24	8	52	1990	66	130	8	-	63	00'	20	40'	636	6.55
112	2	44	66	24	7	25	1892	44	131	-	-	63	00'	19	09'	1314	4.7
113	31	40	65	34	7	31	1435	68	132	-	-	63	00'	17	04'	1497	4.6
114	6	44	65	34	8	54	842	66	133	9	-	63	14'	11	24'	433	2.2
115	2	44	65	29	8	46	878		134	-	-	62	34'	10	26'	563	4.1
116	31	46	65	33	19	28	926	63	135	10	-	62	48'	9	48'	598	0.4
117	8	48	65	30	12	07	183	44	136	-	-	63	04'	9	44'	482	4.8
118	7	49	65	29	13	25	74	45	137	-	-	63	14'	8	31'	559	0.6
119	11	48	66	44	11	33	1414	68	138	-	-	63	26'	7	56'	887	0.6
120	12	49	67	44	8	48	1619	69	139	-	-	63	36'	7	30'	1322	0.6
121	13	42	67	35	6	44	2686	44	140	11	-	63	20'	6	57'	1469	0.9
122	23	44	69	31	7	06	2465	40	141	-	-	63	22'	6	58'	1279	0.6
123	6	43	67	29	7	29	1456	40	142	-	-	63	07'	7	05'	1105	0.6
124	8	43	67	30	8	29	162	64	143	-	-	62	58'	7	09'	731	0.4
125	14	46	67	30	8	26	990	64	144	-	-	62	49'	7	12'	529	1.6
126	15	44	69	30	8	24	1889	40									

THE DANISH INGOLF-EXPEDITION

VOLUME V

14

MEDUSÆ

PART III.

TRACHIYLINA AND SCYPHOZOA

WITH ZOOGEOGRAPHICAL REMARKS ON ALL THE MEDUSÆ
OF THE NORTHERN ATLANTIC

BY

P. L. KRAMP

WITH 6 PLATES AND 26 FIGURES IN THE TEXT



COPENHAGEN

PRINTED BY BIANCO LUNO

1947

Introduction.

Many years have elapsed, since I published the first two parts of the medusæ of the northern Atlantic in the "Ingolf" publications (Leptomedusæ 1919, Anthomedusæ 1926). For various reasons the treatment of the Trachylina and the Scyphomedusæ was repeatedly postponed; other work, particularly on animals from Greenland waters, occupied much of my time available for scientific work; moreover I wanted the final treatment of the North-Atlantic medusæ to be as complete as possible by including new collections (mainly from the "Dana" expeditions), and also to be better acquainted with the distribution of the oceanic species in other parts of the Atlantic Ocean. In this respect the working out of the extensive collections of Hydromedusæ from the "Dana" expeditions in 1921-22 and 1930 and from the "Discovery" expeditions has given valuable information.

In the meantime it was decided to restrict the "Ingolf" papers still awaiting publication to comprise deep-sea animals only, hoping at last to come to an end with this series. The majority of the Trachylina and some of the Scyphomedusæ are decidedly bathypelagic, and they might therefore be treated to about the same extent as the Leptolma, but in accordance with the new plan the few neritic forms will only be briefly mentioned.

To conclude the present volume a short survey is given of all the medusæ occurring in the areas under consideration, i.e. the Atlantic Ocean north of about 50° N. and adjacent waters, paying regard to new information on the Leptolma.

One new species is described, *Haliscera bipolara* n. sp.

If nothing else is stated, the material dealt with in the present paper belongs to the Zoological Museum of Copenhagen.

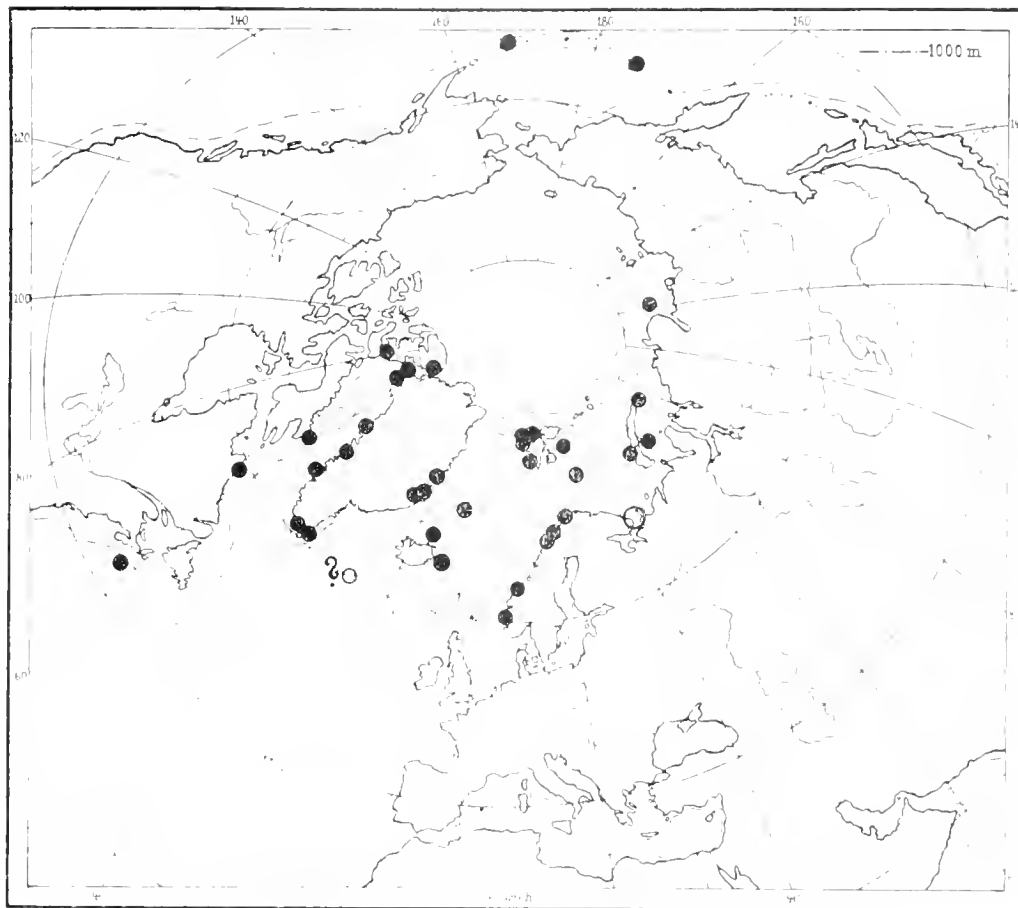


Fig. 1. Distribution of *Ptychogastris polaris* — exact locality unknown.

16 clusters; in all other respects these young specimens resemble the adult, and the gonads are already visible on the eight stomachal lobes which are all of equal size.

Colour: According to sketches, made on board the "Ingolf", Sr. 126, the colour of the subumbrella is a light pink, the gonads are scarlet, the stomach a deep red (Pl. VI figs. 1-2).

Comparison with other species. I quite agree with VANHÖFFEN (1912a p. 386) that *Ptychogastris antarctica* (Haeckel) differs too much from *P. polaris* to be placed in the same genus, or even in the same family. The genus is, however, really represented in the Antarctic by *P. opposita* Vanhöffen (1902b), a species greatly resembling the arctic *P. polaris*, *P. asteroides* (Haeckel) from the Adriatic Sea and the Straits of Gibraltar likewise seems to be a true *Ptychogastris*, though there is one feature which (provided that the description is correct) presents a serious obstacle against its affinities to this genus: the adhesive tentacles are said to be hollow, and in HAECKEL'S figure of a transverse section (1881, Pl. 7 fig. 4) the central canal is seen to be surrounded by an epithelium of endoderm cells with cilia waving into the hollow space, and the ectoderm is remarkably thick; these tentacles are, accordingly, fundamentally different from those of *P. polaris*, and from the tentacles of any other Trachymedusa as well.

Systematic position. As far as the systematic position of *Ptychogastris* is concerned, I absolutely disagree with the supposition put forth by MAAS (1906a p. 481) and provisionally followed by BIGELOW (1909b p. 311) that *Ptychogastris* should be associated with *Crossota* on account of "the arrangement of the several rows of tentacles". In *Ptychogastris* the tentacles are placed in distinct clusters, whereas in *Crossota* they make an uninter-

rupted whorl; and as previously pointed out by me (1942 p. 80) the tentacles of *Crossota* are not arranged in several rows or series. Thus the only point of connection supposed to exist between these two genera does not hold good, and in all other respects they are highly different. The species of *Ptychogastris* constitute a distinct family with an isolated position within the Trachymedusa.

Occurrence:

Almost all the specimens in our collections were collected at the bottom of the sea, with dredge or trawl, at very different depths, varying from about 10 m (in East Greenland) to 330 m (Inglefield Bay, West Greenland); most of the records in the literature are from depths of less than 200 m. HAECKEL (1881) records it from off Halifax, Nova Scotia, at a depth of 2200 m, but the specimen in question may possibly have been caught pelagically in a higher level. In all parts of the area of distribution the medusa has most frequently been collected at the bottom, where it most likely attaches itself to solid objects by means of the adhesive disks of its tentacles. It appears to BROWN (1903 p. 29) that "a medusa, which has exceptionally powerful circular muscles lining the whole subumbrella and a strong muscular velum of exceptional width, would be a strong and active swimmer and not likely to spend much of its time crawling about the bottom of the sea". As a matter of fact, the vast majority of the specimens hitherto observed have actually been stationed on or near the bottom, but occasionally the medusa is met with swimming in the upper strata, sometimes even very near the surface of the water. It seems most likely, therefore, that the habit of the medusa is to spend some of its time attached to the bottom, usually at rather considerable depths, but occasionally to ascend to higher levels, and for such a habit of living the possession of strong muscles is highly required. Very likely MAAS (1906a p. 480)

of the species *Halicreas* with that of *Halicorella* and *Halicorella* to great depths (1000-2000 fathoms) includes in order to

FIGURE 1. *Halicreas* (from map, text fig. 1). 1. *Halicreas* *conicum*, 2. *Halicreas* *albium*, 3. *Halicreas* *papillosum*, 4. *Halicreas* *glabrum*, 5. *Halicreas* *rotundatum*. The locality in deep water south of the Cape of Good Hope on the map seems to me to be the locality where MAAS (1906, p. 20) was based upon his description, and I feel sure that the identification is correct.

Fam. Halicreasidæ Fewkes 1896.

The family is a group of radial canals, with numerous tentacles in each octant, all structurally alike and arranged in a regular pattern, each divisible into a soft flexible proximal part and a distal region, with free sensory clubs, with numerous long proboscis. (Bigelow 1919, p. 321, slightly modified.)

FIGURE 2. (cf. Bigelow) that the medusa which agree with the above constitute a separate family, and also that the number of tentacles in each octant is the most characteristic feature of the group. It is to be deplored, therefore, that another expert author (Cundy 1928, p. 80) not only reduces the present family to a subfamily (Halicreasinae), but will have the genus *Homonocentia* in which a proper manner of tentacles is present.

Various suggestions have been advanced concerning the number of tentacles in the limitation of its genera and species, but I am afraid to arise on account of too vague descriptions of the species. I hope before long to be able to enter into a more detailed discussion of these problems, based upon examination of extensive collections from various parts of the world, but in the meantime must, however, be forwarded on the subject as follows.

On account of the varying conceptions of MAAS of the genus *Halicreas* created by himself in 1896, I shall not discuss the question of the rate of that genus in the present paper, but will only remark that it seems to me highly probable that the *Halicreas* he has described and figured by him (figs. 21-24) are the same as *Homonocentia platygonum* Fewkes, 1896, the type species, which is the type species of the family Halicreasidæ, as distinctly provided with four lips, and is not related to the Halicreasidæ. Some further remarks on the genus *Halicreas* will be found below under the discussion of the genus *Halicorella*.

On account of the limitation of genera and species of the family Halicreasidæ, the incompleteness of the description of the genus *Halicreas* of the *Willarr* medusa is not surprising, and is not beyond doubt.

The genus *Halicorella* (the first genus, *Halicorella* (1896), has been derived from the genus *Halicreas* (1896) by the species *Halicreas nana* (1896) described thoroughly by Bigelow (1919), and named by Bigelow *Halicorella nana*. A complete review of the genus *Halicorella* is not possible, but I will mention a few of the more important specimens. The specimen of *Halicorella* (1896) is a female, and is identical with the specimen of *Halicorella* (1896) of Brown (1902), and is very like *Halicorella* (1896) of Bigelow (1904), and *Halicorella* (1896) of Vanhoffen (1902), and is very like *Halicorella* (1896) of Maas (1906), and

which I have seen a few specimens, is certainly a distinct species, probably of *Halicorella*.

The specific value of VANHOFFEN'S several species has been doubted by various authors, but none of them have come to definite conclusions. THOM (1936) certainly went much too far in reducing the number of species to only two: 1) *Halicreas albium* Vanhoffen (with only one kind of tentacles) also comprising *H. conicum* (Vanhoffen) and *H. macrotza* (Maas), 2) *Halicreas papillosum* Vanhoffen (with eight long and numerous short tentacles), also comprising *Halicreas glabrum* Vanhoffen and *Halicreas rotundatum* Vanhoffen. RANSON (1936) maintains the genera *Halicreas* (with more than 15 tentacles in each octant) and *Halicorella* (with 12 or less tentacles in each octant), and he also observed a characteristic difference in the conformation of the umbrella in these two genera; he retains all VANHOFFEN'S species of *Halicreas*, and refers *Homonocentia platygonum* Maas to *Halicorella*. According to BIGELOW (1938, p. 121) it is obvious that "the five named representatives of the smooth-belled subdivision of the genus (*conicum*, *albium*, *glabrum* and *rotundatum* Vanhoffen 1902, *macrotza* Maas, 1906) stand in need of drastic reduction, successive studies (Bigelow 1909, Ranson 1936) having proved that the features on which Vanhoffen (1902) laid chief stress (degree of doming of the exumbrella, and precise location of the gonads on the radial canals), are too variable to be of taxonomic significance."

I am however not quite sure that the number of species may be much reduced, my own studies have convinced me that a number of distinct species exist, but I find it difficult to bring them into concordance with VANHOFFEN'S descriptions. As a matter of fact, no proper descriptions were given, except of *H. papillosum*, but only scattered remarks on certain distinguishing features, and the beautiful coloured figures are more artistic than elucidating. After I wrote my paper on the medusa in the waters west of Greenland (KRYMR 1912) I have seen numerous specimens of *Botryocentia brucei* Browne from almost every part of the Atlantic Ocean, which further confirmed my supposition that *Halicreas glabrum* Vanhoffen belongs to the same genus, and probably even to the same species; the peculiar structure and position of the tentacles thus entirely escaped the attention of VANHOFFEN, in the uncoloured figure of *Halicreas papillosum*, seen from the exumbrella (Pl. IX fig. 8), the difference in size of the eight periradial and the other tentacles is clearly shown, but in the lateral view (fig. 7) nothing of the kind is indicated. One might be inclined to think that the artist has taken similar liberties in his delineation of some of the other figures.

Halicreas rotundatum is expressly stated to have 20 tentacles in each octant when 18 mm in diameter, it also has an evenly rounded, dome shaped umbrella. I have not seen any medusa corresponding to this species; none of the dome-shaped specimens which I have seen had more than about 12 tentacles in each octant, nor has any other author, (cf. *Halicorella hutchinsii*, p. 9).

The most characteristic feature of *Halicorella conica* is the conical shape of the umbrella, it was taken in one or two localities, south of Africa and in the Indian Ocean, one of the specimens was 18 mm in diameter, and according to fig. 33 on Pl. II it had 9 tentacles and two statocysts in each octant (fig. 6 on Pl. 9 shows 11 tentacles). Various authors have doubted the specific value of the shape of the umbrella, it should be pointed out, however, that the jelly of the Halicreasidæ is rather firm and resistant and apparently not very liable to alter its shape by preservation. As a matter of fact, all the numerous specimens of *Halicorella* which I have seen in the "Discovery" collections from the southern Atlantic and the Antarctic Ocean have the conical shape characteristic of *H. conica* and agree very well with that species, with the exception that the number of tentacles rarely exceeds 8 in each octant. It is the same species which occurs in the Mediterranean.

On the other hand, all the numerous specimens of *Halicorella*

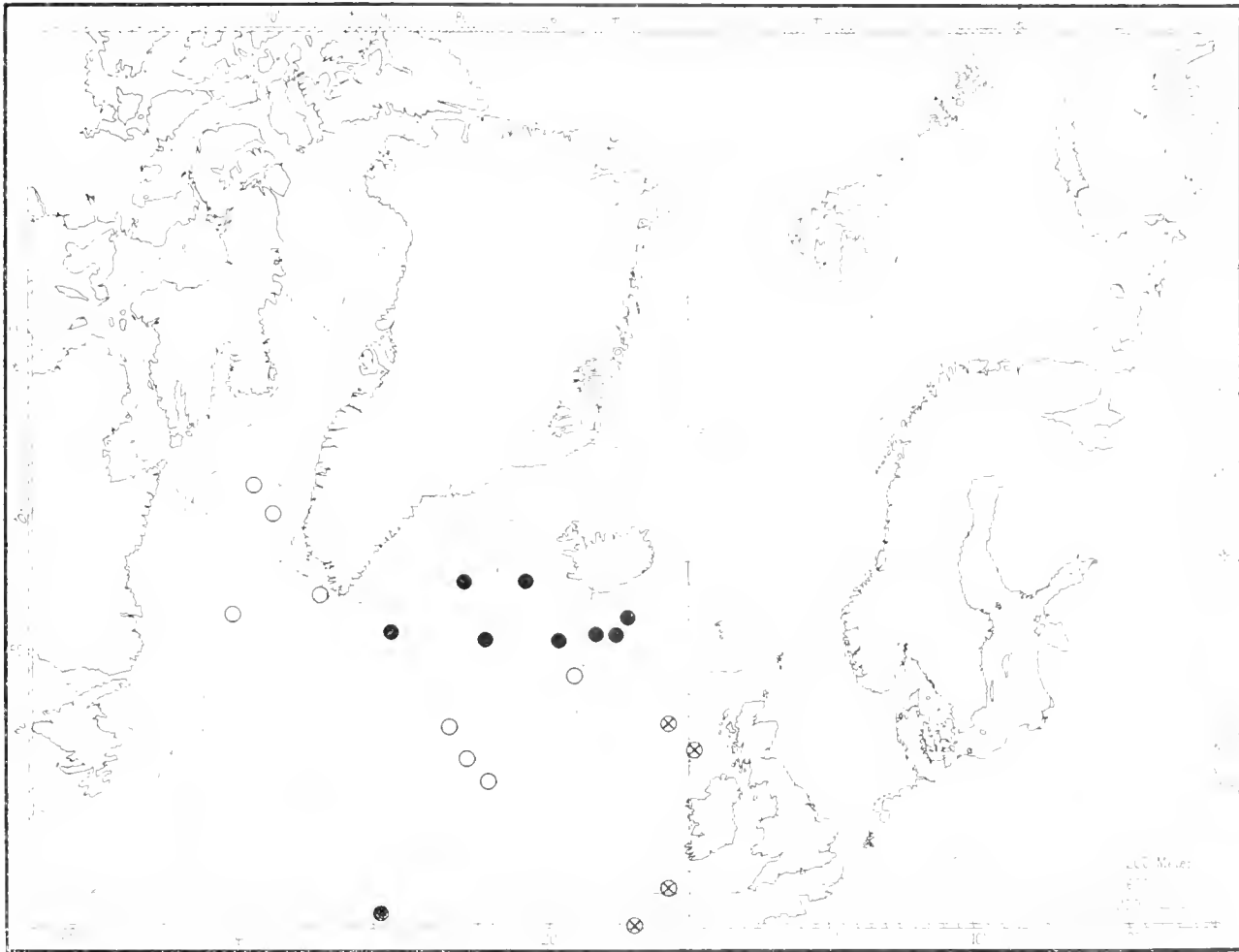


Fig. 2. Distribution in the northern Atlantic of *Halicreas minimum* ● new records ○ previous records, ⊗ records not yet published.

from the northern Atlantic examined by me, and further mentioned below, have an evenly rounded, dome-shaped umbrella with a thick jelly. They are about 17 mm in diameter, with about 12 tentacles in each octant. They agree fairly well with the specimens from the eastern tropical Pacific described and figured by BIGELOW (1909 a p. 142) under the name of *Homococyma alba* (Vanhöffen); but it seems to me doubtful to refer them to that species. VANHÖFFEN'S description of *Halicrecia alba* was based on one specimen from the South Atlantic, a large medusa 35 mm in diameter, yet with only 8-10 tentacles in each octant. Moreover its gonads are described and figured as lance-shaped, occupying a rather considerable portion of the radial canals, whereas in the Pacific as well as in the North-Atlantic specimens the gonads are short and broadly oval or nearly circular in outline. I prefer, therefore, to describe this form as a new species and name it in honour of professor H. B. BIGELOW, *Halicrecia bigelowi* n. sp.

Genus Halicreas Fewkes 1882

Halicreasidae with eight radial canals; with a continuous row of tentacles; with periradial, gelatinous papillae on the exumbrella.

***Halicreas minimum* Fewkes,**

Pl. VI fig. 3.

Principal references.

Halicreas minimum Fewkes 1882 p. 306.
papillosum Vanhöffen 1902 b p. 68, Pl. IX figs. 7-8, Pl. XI fig. 30.
Halicreas papillosum Bigelow 1909 a p. 138, Pl. 3 fig. 3, Pl. 33 figs. 8, 9, Pl. 34 figs. 1-3, 5, 8, 10, 11
Halicreas minimum Bigelow 1938 p. 122.

North-Atlantic records:

Halicreas minimum Fewkes 1882 p. 306, 1886 p. 953
papillosum Kramp 1920 p. 5
 Bigelow 1926 p. 67.
 Ranson 1936 p. 161
minimum Kramp 1942 p. 70.

Material (see the map, textfig. 2)

61° 34' N, 19° 05' W.,	¹⁹ 1904, "Thor" St. 180,	2 specimens
61° 30' N, 17° 08' W.,	²¹ 1904,	183-6
63° 19' N, 26° 50' W.,	²¹ 1928, "Godthaab" St. 1,	1000 m wire 2 spec.
59° 30' N, 15° 23' W.,	²⁹ 1928,	5, 3000 m 1
56° 56' N, 51° 17' W.,	³ 1928,	10, 3000 m 7
62° 19' N, 56° 00' W.,	¹¹ 1928,	21, 1000 m 6
		21, 2500 m 1
60° 59' N, 22° 29' W.,	²⁹ 1925, "Dana" St.	2306, 3000 m wire 3
60° 20' N, 29° 21' W.,	³¹ 1925,	2307, 1500 m 1
59° 21' N, 37° 56' W.,	¹² 1925,	2308, 3000 m 115
		2308, 1000 m 58
61° 17' N, 52° 35' W.,	⁹ 1925,	2101, 1900 m 2
62° 35' N, 32° 53' W.,	²⁷ 1925,	2137, 1900 m 1
17° 02' N, 31° 15' W.,	²⁷ 1931,	1201, 2000 m 7
		1201, 3000 m 70
		1201, 1000 m 10
		1201, 5000 m 33
		1201, 6000 m 25
62° 23' N, 16° 05' W.,	²⁹ 1932,	1102, 3000 m 13
		1102, 1000 m 17
62° 36' N, 32° 18' W.,	¹⁶ 1933,	1687, 2000 m 7

the same localities in which Michael Sars (North Atlantic) took these specimens, but were not recorded. They were taken in 1910.

For the description of *Haliscera* VANHÖFFEN (1902 b), more material is necessary. The excellent figures illustrating the anatomy of the medusa (Pl. IX, figs. 1-5) are the majority of the number of specimens examined, and are in a poor state of preservation. The figures of the tentacles, however, with some of the tentacles themselves retained, they fully agree with the description of VANHÖFFEN. A discussion on the individual specimens and their localities is better postponed for a later date, in connection with the extensive collections from the deep-sea of the North Atlantic which have been placed at my disposal.

The coloration of the medusa are not quite as given by VANHÖFFEN (1902, Pl. IX, fig. 7); the gonads are a pure scarlet, the radial canals pink, the tentacle stumps, carmine (Pl. VI, fig. 2), brown, coloured sketch made by me on board the "Thor" (1908).

Haliscera is a bathypelagic medusa with a world-wide distribution. The map, textfig. 2, clearly shows that its geographical range in the northern Atlantic is limited by the submarine ridges and the submarine ridges connecting Scotland, Iceland, Greenland, and Baffin Land. With one exception ("Godthaab" St. 12, North Atlantic localities are all outside the limits of the Greenland Sea. St. 5 was near the south point of Greenland, latitude 60° 29' N, and one single specimen was taken in 1920, 200 m below the surface in water with a salinity of 34.91 ‰. It was taken in the deep strata outside the shelf, where the medusa occurs. It is also the only instance of the medusa occurring in the arctic region of this geographical region; all other North Atlantic specimens were taken in hauls with at least 1000 m wire out (2000-6000 m below the surface). The species was nowhere taken in series of less than about 2000 m below the surface, and in most of the hauls taken with 3000-6000 m wire out (2000-4000 m below the surface) brought forward a large number of specimens. The vertical distribution is particularly well illustrated in the catches of "Dana" St. 1201, the ring trawl with a 200 m opening and with an opening diameter of 150 cm, towed for 2 hours at a depth of 1500 m, and with 2000 m wire out, caught 7 specimens, whereas a considerable number were taken in hauls made with a 100 m net towed simultaneously with 3000 m wire (10) and 5000 m wire (33); the deepest hauls made with a larger net, diameter 300 cm, and with a 200 m opening net, it took 25 specimens during the haul. The species was taken in still greater numbers in a haul made with 100 m net, "Dana" St. 2308, where 115 specimens were taken in 2 hours towed during a haul of 2 hours' duration, and 18 specimens in the haul of 14 hours' duration. The following were taken in the hauls made

THOR (1935, p. 46) found an increase of the average size of the specimens from the upper strata downwards, and he concluded that the eggs or young larvae rise towards the surface, whereas the medusa, while growing larger, gradually sink back into the deep-sea (the specimens at his disposal were all young ones, less than 10 mm in diameter). A tendency in the same

m wire out	Diameter of specimens, mm
1000	22-29
2000	9-26
3000	9-35
4000	12-35
5000	20-30
6000	22-30

direction is indicated by the North-Atlantic collections examined by me, as seen from the accompanying table showing the size limits of the individuals taken at different depths. I would, however, not attach too much importance to these figures which may prove to be entirely casual; but I shall return to the question, when my studies on the collections from the central and southern parts of the Atlantic Ocean are finished.

Geographical distribution: *Haliscera minimum* occurs in the deep parts of all the oceans, except in the arctic seas. There are regions of the Indian and Pacific Oceans, from which it has not yet been recorded, but there is every reason to believe that its distribution in these oceans really comprises the entire deep-sea areas. In the Atlantic Ocean it is generally distributed in the deep basins from the submarine ridges in the North Atlantic to the shelf of the Antarctic Continent. In the Pacific its area of distribution comprises the Bering Sea, the deep-sea of that water is in direct connection with the deep-sea of the Pacific Ocean and forms a continuous part of it, in the same manner as the deep-sea areas west and east of the southern part of Greenland are continuous portions of the Atlantic deep-sea basin.

Genus **Haliscera** Vanhöffen 1902.

Halisceridae with eight radial canals; with a continuous row of tentacles; without exumbrel papillae.

Haliscera bigelowi n. sp.
Pl. I figs. 5-8, Pl. II figs. 1-2.

Homocoenia alba Bigelow 1909 a p. 112, Pl. 3 figs. 1, 2, Pl. 33 figs. 6, 11, Pl. 34 fig. 9.

Material (see the map, textfig. 3).

57° 03' N, 41° 20' W., 28 1/2 1908, "Thor" St. 12, 300 m wire, 2 specim.	
63° 19' N, 26° 50' W., 23 1/2 1928, "Godthaab" St. 1, 1000 m wire, 1 spec.	
63° 32' N, 26° 23' W., 28 1/2 1925, "Dana" St. 2138, 1500 m wire, 75	
62° 23' N, 16° 05' W., 29 1/2 1932,	1102, 3000 m 6
62° 23' N, 16° 05' W., 25 1/2 1932,	1102, 3000 m 6
	1102, 4000 m 4
62° 36' N, 32° 48' W., 19 1933	4987, 4000 m 5
62° 45' N, 16° 01' W., 7 1/2 1934	5083, 1000 m 1
	5083, 1500 m 30
	5083, 2000 m 20

Moreover taken by the "Michael Sars" in the following localities.

47° 31' N, 43° 41' W., 11 1/2 1910,	"Michael Sars" St. 80, 950-525 m
48° 02' N, 39° 55' W., 12 1/2 1910	81, 2000 m wire
46° 48' N, 27° 46' W., 17 1/2 1910	87, 1900-0 m
45° 26' N, 25° 15' W., 18 1/2 1910	88, 1000 m wire
48° 29' N, 13° 55' W., 23 1/2 1910	92, 600 m
50° 13' N, 41° 23' W., 29 1/2 1910	91, 600 m
56° 33' N, 9° 30' W., 7 1/2 1910	98, 1000 m
	98, 1500 m
57° 41' N, 41° 48' W., 27 1/2 1910	101, 1000 m
	101, 2000 m

in the central and southern Atlantic Ocean the occurrence of the medusa is more restricted to the deep strata. The vertical distribution in the North Atlantic is shown in the accompanying map, about 7 cm to about 800 m below the surface, and between 50 and 100 m below the surface. The species occurs to be more abundant in the deep-sea of the North Atlantic. *Haliscera papillosa* (Vanhöffen) and *Haliscera minima* (Vanhöffen) are also recorded from the North Atlantic. *Haliscera minima* (Vanhöffen) has been recorded from the deep-sea of the North Atlantic, but it occurs right up to the surface in the central and southern Atlantic Ocean. It has been recorded from the deep-sea

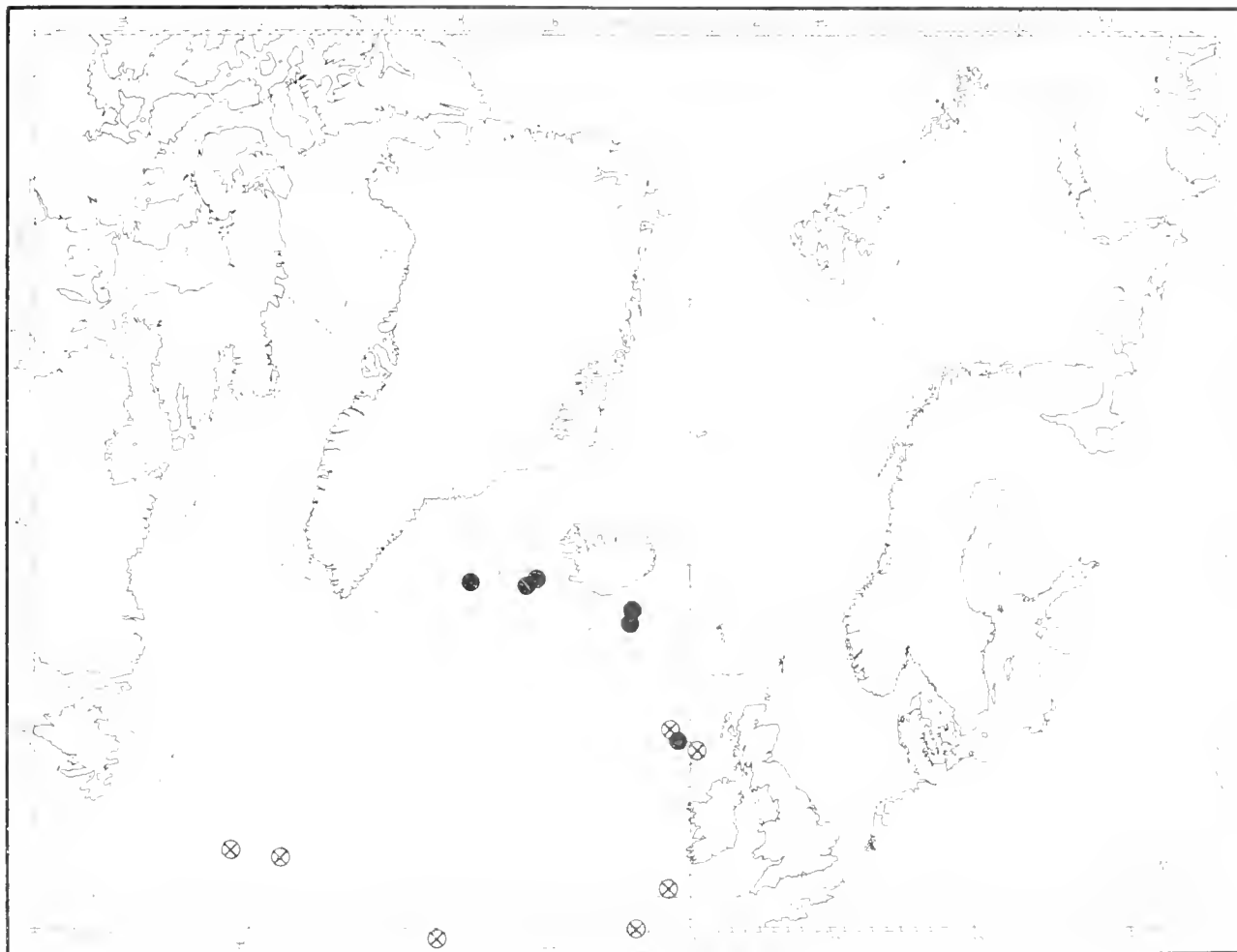


Fig. 3. Distribution in the northern Atlantic of *Halisera bigloui*. ● new records; ⊗ records not yet published.

Description: (Pl. I fig. 5) Umbrella almost hemispherical, its central portion consisting of a thick, hemispherical mass of jelly, by a slight constriction separated from the marginal portion which has fairly thin walls, the apical gelatinous portion comprises nearly two thirds of the entire height of the medusa.

The stomach (Pl. I fig. 6) is very broad and flat, slightly conical in shape, it has a circular outline, and the mouth is a circular opening varying in width according to the state of contraction. There are eight broad radial canals and a broad circular vessel. The gonads (Pl. I fig. 6) are broadly oval, comprising about two fifths of the length of the radial canals and placed somewhat nearer to the stomach than to the circular vessel; in female specimens each gonad contains about seven large eggs in one layer. The gonad is usually rather flat, but in certain states of contraction it is doubled up after a longitudinal line and thereby attains a laterally compressed appearance.

In the specimens examined almost all the tentacles are broken off, leaving only the basal stumps (Pl. I fig. 7); a few small tentacles are retained, showing that they have the same structure as in other species of *Haliereasidae* (Pl. I fig. 8). The number of tentacles is usually about 96 (12 in each octant); in some of the octants there may be 11 or 13; the full number of tentacles, as attained at an early stage of development, in the smallest specimen examined, which is 5 mm in diameter, there are about 80 tentacles; another small specimen, 6 mm wide, has about 96, and in the largest specimen, 19 mm in diameter, the number does not exceed 100. The tentacles are all of the same structure, but varying in size; the eight periradial tentacles are not distinctly larger than the others. The distribution of large and small tentacles between

the periradials is rather irregular, two small ones or two or only large ones are frequently placed side by side.

The statocysts (Pl. II figs. 1-2) are elongated club-shaped, like other species of *Haliereasidae*; the endoderm core consists of about 10 cells each with a large nucleus except the two or three distal cells, in which no nuclei are seen; as all the specimens are preserved in formalin, the statoliths have completely disappeared. The ectoderm consists of two different kinds of cells; in the distal half or two thirds of the club there are two lateral rows of rectangular cells, rather high and rich in protoplasm, they have most probably carried sensory hairs; in the proximal portion of the club and on the abaxial and adaxial sides of the distal portion, the epithelium consists of large, flattened cells with irregular outlines, usually somewhat elongated in the longitudinal direction of the club.

There are three statocysts in each octant, and their position among the tentacles is very characteristic and rather constant (Pl. I fig. 7) when the number of tentacles in an octant is 12 we shall almost always find the statocysts placed as follows: (1) the medusa seen from the exumbrellar side, the bell margin lower way, to the right hand side of a periradial tentacle we first see an outer tentacle and then a statocyst; (2) following 1 by 3 tentacles, one statocyst, 3 more tentacles, and again one statocyst; but between the third statocyst and the first one in the next octant there are 6 tentacles! A statocyst in the middle of this group of six tentacles would result in a completely symmetrical and regular position of statocysts round the umbrella margin, with three tentacles between two successive pairs of statocysts; but I have never seen a statocyst on the left hand side of a periradial tentacle or in the

the apical knob is frequently broken and the radial canals are often irregular. They are also frequently broken in a few specimens. The shape of the apical knob and the variations in the number of tentacles in an octant as well as the shape of the perradial tentacles are thus asymmetrically distributed, but are repeated in the same manner.

Female. Umbrella 100 mm. or more, but with a weak muscular contraction. Bell margin 15-17 mm. height 10-12 mm.

Female gonads. The female seen alive ("Goolthaab") is a very young specimen. St. 1 is chosen as the **type specimen** because it is an individual, fairly well preserved, with the apical knob and bell margin as damaged. The above description of the female gonads is based on other specimens. (M. J. S. 1938, 80-81.)

Comparison with other species. The North Atlantic species here described agrees in almost all respects with the species described by BIRLOW (1909a) of the medusa from the Pacific, referred to him by HALPERN as *Haliscera alba* Vanhoffen; the only difference is that I feel almost sure they belong to the same species. According to BIRLOW the female gonads contain 12-22 eggs in the largest number observed, whereas in the North Atlantic species there are rarely more than seven eggs in each of the largest number observed. BIRLOW describes the female gonads as deep, and not flat, as stated above; the female gonads there are sometimes flat, sometimes laterally flattened. The only reasons of doubt against uniting the Pacific and Atlantic forms into one species, is the relative size of the female gonads according to BIRLOW "the radial ones are 1.5-2 times larger than those between the canals," whereas in the North Atlantic form there is a difference, it is not distinctly proportioned, and in the same specimen the eight perradial tentacles are of one or more of somewhat varying size, and the female gonads in the Pacific specimens, as seen from the figure (1909a, fig. 22) of the bell margin, Pl. 33 fig. 11, in which the perradial tentacles are of very different size, and one is much larger than some of those between the two canals. It may be remarked that apparently the tentacles are broken off exactly at their point of issue from the umbrella, and that we cannot be sure that the size of the perradial tentacles at the end of the endodermal core gives a true impression of the size of the tentacle which in the living specimen is broken off. The difference in the descriptions by BIRLOW and myself therefore does not seem to me to be of sufficient importance to indicate a specific difference between the two species. As mentioned above (p. 7) HALPERN has referred to *Haliscera alba* Vanhoffen specimens collected from *Haliscera caeca*, which has 64 tentacles and 16 statocysts, and from *Haliscera alba* Vanhoffen, which has 160

localities in the eastern tropical Pacific between about 0° and 25° S., where it was taken in vertical hauls from 300 fathoms to the surface. If the medusa found in these two widely separated areas really belong to one species, it is to be expected that this species will prove in future to have an extensive distribution in the deep-sea areas of the oceans.

Genus *Botrynema* Browne 1908

Halisceridae with eight radial canals, with sixteen groups of tentacles (two groups containing many tentacles in a single row in each octant) and eight solitary perradial tentacles.

In a previous paper (KRAMÉ 1942, p. 73) I have discussed this genus and its species. Numerous specimens had been collected in the deep-sea areas west of Greenland, and I pointed out that all specimens from the Baffin Bay had an evenly rounded umbrella, whereas in all specimens taken in the Atlantic water south of the submarine ridge across Davis Strait the apical jelly had an enormous thickness and was provided with a very distinct and sharply defined apical knob (Pl. VI fig. 4). Though the two forms were alike in all other respects, I concluded that they belonged to two different species, the northern form, from the arctic basin of the Baffin Bay, certainly belongs to *Botrynema allmanni* (Hartlaub), the southern form agrees perfectly with *B. braucei* Browne, originally described from the Antarctic Ocean. Later on I have examined extensive collections of *Botrynema braucei* from almost all parts of the Atlantic Ocean, and all these numerous specimens, except very young ones, possess the remarkably thick jelly and the distinct apical knob, which I had observed in the specimens from Davis Strait. They confirm, therefore, my supposition that the arctic *B. allmanni* and the Atlantic *B. braucei* are two distinct species, though they differ from each other only in colour and in the shape of the umbrella, this difference however being remarkably constant.

In the paper quoted above I also called attention to the possibility that *Haliscera glabrum* Vanhoffen might be identical with *Botrynema braucei* Browne. The name *Haliscera glabrum* was given by VANHOFFEN (1902 b) to a medusa taken by the "Valdivia" expedition in the tropical and southern parts of the Atlantic; the species was not properly described, and if the figure (Pl. 9 fig. 3) really has been drawn after a specimen of *Botrynema*, it gives an altogether wrong conception of the bell margin. I am therefore still of the opinion, expressed in my former paper (1942, p. 77) that "the name of the cosmopolitan species must remain *Botrynema braucei* Browne and should not be replaced by *glabrum*, because the description of that species is really beyond recognition."

The identity of the genera *Botrynema* Browne 1908 and *Allomaniota* Hartlaub 1909 was pointed out by BIRLOW (1913, p. 52); he examined some specimens taken in the northern Pacific and referred them to *B. allmanni* (Hartlaub), but they most certainly belong to *B. braucei* (see KRAMÉ 1942, p. 76).

Both species are now so well known that I have very little to say about their morphology. I am able to state, however, that the tentacles have the same structure as in other species of Halisceridae, in all specimens hitherto observed the tentacles are all broken off near their base, but in some of the specimens at my disposal I have found a few very young tentacles fully preserved (Pl. I fig. 9). The statocysts are likewise very similar to those of *Haliscera* and *Haliscera* (Pl. II fig. 3). They are placed in the 16 spaces between the groups of tentacles, in both species there are usually 3 statocysts in each of the interradial spaces and one or two on either side of the perradial tentacles, but in a few specimens of *B. braucei* I have found a perradial tentacle flanked by 3 statocysts on either side.

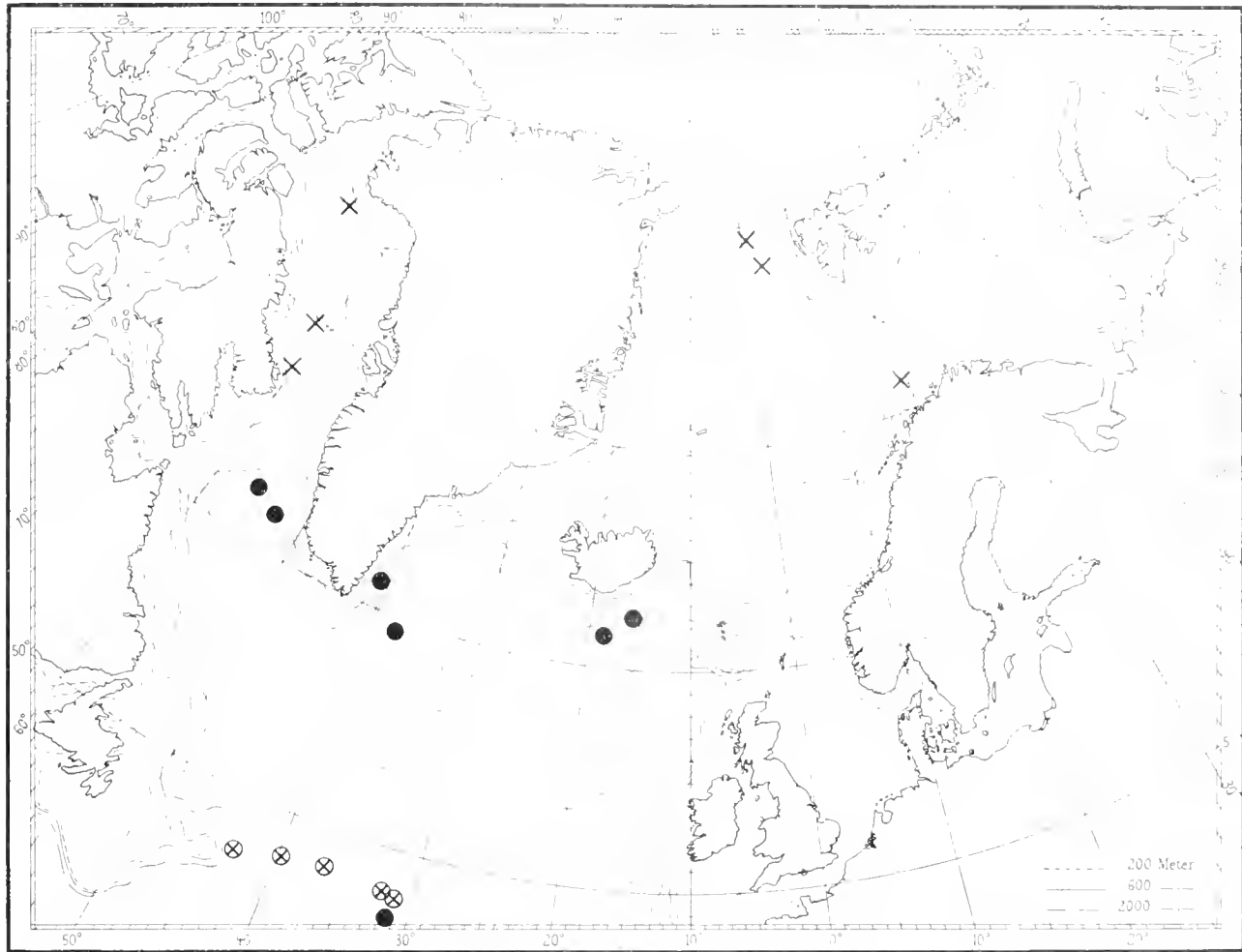


Fig. 4. Distribution in the northern Atlantic of: ● *Botrynema brucei*, new records; ○ do., records not yet published; X *Botrynema ellinorae*.

***Botrynema ellinorae* (Hartlaub).**

Alloionema ellinorae Hartlaub 1909 p. 8, Pl. 76 figs. 3, 4, 6.
Botrynema ellinorae Kramp 1912 p. 77.

I have seen no other specimens of this species than those taken by the "Godthaab" expedition 1928 and mentioned by me in 1942; they were taken in three localities in the deep basin of Baffin Bay, about 1000-1800 m below the surface at temperatures between 0° and -0.4°.

Further distribution (see the map, textfig. 4): Between East Greenland and Spitzbergen and north of Norway, in deep water (HARTLAUB).

***Botrynema brucei* Browne.**

Pl. I fig. 9, Pl. II fig. 3, Pl. VI fig. 1.

✓ *Haliereas glabrum* Vanhoffen 1902b p. 70, Pl. 9 fig. 3
Botrynema brucei Browne 1908 p. 239, Pl. I figs. 8-9, Pl. 2 fig. 1, Vanhoffen 1912a p. 382, textfigs. 18-19, Pl. 25 fig. 5.
Botrynema ellinorae Bigelow 1913 p. 53, Pl. I figs. 1-4.
 ✓ *Haliereas glabrum* Ranson 1936 p. 167, Bigelow 1938 p. 124.
Botrynema brucei Kramp 1912 p. 77.

North-Atlantic record:

Botrynema brucei Kramp 1912 p. 77.

Material (see the map, textfig. 4):

61 31°N, 19 05°W., ¹⁰ 1901, "Thor" St. 180, 1 specimen.		
62 19°N, 56 00°W., ¹¹ 1928, "Godthaab" St. 21, 1000a, 2500 m wire		
59 21°N, 37 56°W., ¹² 1925, "Dana" St. 2308, 3000 m wire, 70 spec.	2308, 4000 m	24
61 17°N, 52 55°W., ⁹ 1925	2401, 1900 m	8
61 13°N, 40 57°W., ²⁵ 1925.	2136, 1700 m	1
17 02°N, 31 15°W., ²⁷⁻²⁸ 1931	1201, 1000 m	85
	1201, 5000 m	235
	1201, 6000 m	12
62 23°N, 16 05°W., ²⁵ 1932	1102, 1000 m	20

In the map, textfig. 4, are included the localities in which this species was collected by the "Michael Sars" North Atlantic Deep-Sea Expedition 1910.

This species is easily recognizable, even if badly preserved. The gelatinous substance is very resistant, and the medusa is, therefore, at once conspicuous by the characteristic shape of the umbrella. Moreover, in all the numerous specimens I have seen, traces of the sixteen groups of tentacles are always distinguishable, even in specimens in a very poor condition. The female gonads have never been properly described, and unfortunately none of the female specimens at my disposal are sufficiently well preserved to allow a closer examination of their gonads. I am, therefore, unable to state whether they have contained many small eggs or few large ones. The male gonads are shield shaped, with the apex pointing towards the bell margin, and rather flat.

Color: The stomach and the radial canals are bright scarlet, the circular vessel somewhat lighter, the tentacle stumps brown.



Fig. 5. Distribution in the northern Atlantic of: ● *Rhopalomena velatum* — *Rhopalomena punctatum*.

Rhopalomena velatum Gegenbaur.

North-Atlantic records:

- Rhopalomena coeruleum* Browne 1906 p. 172.
- velatum* Vanhöffen 1912a p. 371.
- Bigelow 1911 p. 20.
- 1918 pp. 388, 433.
- 1926 p. 54.
- Ranson 1936 p. 137.

Material (see the map, textfig. 5):

- 57° 17' N, 11° 13' W., ⁷/₆ 1905, "Thor" St. 71, 1 specimen
- 49° 01' N, 14° 52' W., ⁶/₆ 1906, 65, 300 m wire, 1 spec.
- 56° 56' N, 9° 01' W., ²⁸/₃ 1908, 11, 65 m 1
- 47° 02' N, 31° 15' W., ²⁷/₂₈ ²⁸/₃₁ "Dana" St. 1201, 50-6000 m wire
- 49° 19' N, 30° 22' W., ³⁰/₆ 1931, 1203, 50-600 m

In the two last-mentioned localities the species was taken at different depths, from very near the surface to about 1000 m down, though it is possible that some of the specimens taken in the deepest hauls were actually captured during the hauling in of the nets. At both stations the four uppermost hauls, with 50-600 m wire out, were made with the 2 m ringtrawl (S 200) with bag of stramin; at stat. 1201 four deeper hauls, with 2000-5000 m wire out, were made with a somewhat smaller net, 1½ m in opening diameter; in the deepest haul at each station the appliance (E 3000) was much larger, 3 m in diameter, but with a bag of coarser net, not

suitable to catch this small medusa. The duration of the hauls was also different. For the sake of comparison between the catches it is necessary, therefore, to re-calculate the number of specimens to a certain standard. In the table below are given: the appliance used at each haul, the duration of the haul, the actual number of specimens caught, and the number of specimens reduced to number per one hour's haul with S 200.

Stat.	m wire out	Appliance used	Duration of haul	Actual number of specimens	Reduced number	Diameter of specimens		
1201	50	S 200	60 min	115	115	4-8 mm		
	100			500	500	4-8		
	300			20	20			
	600			10	10	4-8		
	2000			S 1500	240 min	35	15	6-8
	3000					5	15	5-8
	4000					60	25	5-8
1203	5000			60	25	4-9		
	6000	E 3000		1	15	7		
	50	S 200	120 min	8	4	6-8		
	100			9	15	6-9		
	300			13	6.5	6-8		
1000	600			10	5	4-9		
	1000	E 3000		0	0			

of the specimens were taken in the upper strata of the water column. Its distribution in the western Atlantic is similar to the areas of the eastern tropical Pacific. The "Data" show that the species *Rhopalonema* it mainly occurs in the upper strata. It is true that the medusa was very abundant at St. 4291, the centre of the Gulf Stream, and at St. 4292, at the edge of St. 4293 which was near the edge of the Gulf Stream, at St. 4295, still further west, and at St. 4296, still further west. At St. 4297, 48° N. 66° W. it was not taken at all.

Rhopalonema species mainly belongs to the upper strata of the water column, but is rather common to partly found in deeper water layers, especially very far below the surface. At "Data" St. 4291 a vertical haul was made in the haul with 100 m wire out to a depth of 100 m below the surface, but several specimens were also taken in the upper strata, and it is not likely that all of the specimens taken in the upper strata during the hauling in of the haul were taken in the upper strata.

The vertical parts of the specimens do not show any remarkable vertical distribution of the various size classes.

Geographical distribution. *Rhopalonema velatum* is a cosmopolitan species in the warmer parts of all the oceans, including the Mediterranean. In the western Atlantic its northern limit of distribution is off Georges Bank, about 40° N. On the European coast it is known from the Bay of Biscay, and, as appears from the present collection, it follows the Gulf Stream towards the western coasts of the British Isles, where it has been found as far north as 57° 47' N. west of Scotland.

Rhopalonema funerarium Vanhoffen.

(Pl. II figs. 4-5)

Diatrypastrum Quoy & Gaimard 1827

Rhopalonema funerarium Vanhoffen 1902 b, p. 61, Pl. 9, fig. 2, Pl. 10, fig. 17, Pl. 11, fig. 34

Rhopalonema funerarium Maas 1904, p. 28.

Rhopalonema coeruleum Maas 1905, p. 51, Pl. 10, figs. 67, 68.

funerarium Bigelow 1909 a, p. 132

coeruleum Mayer 1910, p. 380

funerarium Vanhoffen 1912 a, p. 372.

Bigelow 1917, p. 306

Kramp 1924, p. 22

Broch 1929, p. 196

Ranson 1936, p. 111

Clarke, *Transl.* 1936, p. 10

N. S. P. Atlantic record 4

Rhopalonema funerarium Kramp 1924, p. 22

Maas 1904, p. 28, fig. 5 (text fig. 5)

St. 4291, 48° N. 66° W., 1905, 1 spec. St. 4295, 2 specimens.

St. 4297, 48° N. 66° W., 1906, 1 spec. 184996, 1800 m wire, 7 spec.

Rhopalonema funerarium (Thom.) St. 181° S. W. of Ireland (see also the present paper) (KRAMP 1924). It is true that the present collection of opinion that the specimens preserved from *R. velatum* were taken in the upper strata, but later it was discovered by formalin, but later it was discovered by formalin, they are not in very good condition, and they can only be referred to *Rhopalonema funerarium* (Vanhoffen 1902 b) as *Rhopalonema coeruleum* (Maas 1904) or *Rhopalonema coeruleum* (Mayer 1910) or *Rhopalonema coeruleum* (Bigelow 1909 a) or *Rhopalonema coeruleum* (Vanhoffen 1912 a) or *Rhopalonema coeruleum* (Bigelow 1917) or *Rhopalonema coeruleum* (Kramp 1924) or *Rhopalonema coeruleum* (Broch 1929) or *Rhopalonema coeruleum* (Ranson 1936) or *Rhopalonema coeruleum* (Clarke, *Transl.* 1936).

Bigelow 1909 a, p. 132
Bigelow 1917, p. 306
Kramp 1924, p. 22
Broch 1929, p. 196
Ranson 1936, p. 111
Clarke, *Transl.* 1936, p. 10

which nearly reach to the circular vessel, and by the configuration of the subumbrellar musculature in its upper parts around the manubrium, forming a star-shaped figure, as neatly represented in VANHOFFEN'S figures and similar to that found in *Colobonema*. In four of the specimens examined by me the manubrium is very short, only about one fourth as long as the height of the subumbrella cavity, but in one specimen it is more than half the height of the bell cavity, and the distal half part of the manubrium is narrowed and terminates in four distinct mouth lips (Pl. II fig. 4). The eight perradial tentacles have a broad and bulbous base, but are all broken off short. Most of the adradial cirri have disappeared leaving only a tiny protuberance on the bell margin, but some of the interradial cirri are present (Pl. II fig. 5); they are club-shaped with an almost spherical distal knob and a thin pedicel, in which the endoderm consists of one row of elongated, cylindrical cells surrounded by an ectodermal epithelium of large, flat cells. Several of the statocysts are preserved, and they are placed in the middle of the spaces between the other marginal organs (tentacles and cirri); they are not in such a condition that I can give a description of their minor structure, but I can state that they are enclosed in the same manner as in *R. velatum*. The diameter of the specimens is 9-17 mm, the height 7-12 mm (see KRAMP 1924).

This species clearly differs from *R. velatum* by the much greater length of the gonads, by the greater number of statocysts, and particularly by the position of these latter; the 8 or 16 statocysts in *R. velatum* are placed close by the base of the perradial tentacles and interradial cirri, whereas the 32 statocysts in *R. funerarium* alternate with the tentacles and cirri and are situated in the middle of the spaces between them. In the apex of the umbrella the mesogloea may be somewhat thickened in *R. funerarium*, but it never has a distinct apical knob as in *R. velatum*. *R. funerarium* also differs from the various species of *Rhopalonema* described by HAECKEL (*clarigerum*, *caeruleum*, and *polydactylum*) all of which are most probably identic with *R. velatum* Gegenbaur. It also seems to me improbable that, as indicated by BIGELOW (1909 a), *R. funerarium* should be identic with *R. striatum* Maas 1905; in *R. striatum* the gonads are broadly oval and situated in the proximal parts of the radial canals.

I cannot agree with THIEL (1936) in uniting all species of *Rhopalonema*, including *R. funerarium*, into one species, *velatum*, and when THIEL remarks (p. 15): "Nur KRAMP (1921) hat noch einmal *Rh. funerarium* als gute Art erwähnt." I may refer to BIGELOW (1909 and 1917) and BROCH (1929) who likewise retain *R. funerarium* as a distinct species.

Geographical distribution: *Rhopalonema funerarium* occurs in the deep and intermediate strata, and it seems to have its principal distribution in the tropical parts of the oceans. It is recorded from the eastern tropical Pacific (BIGELOW 1909 a), the Malayan Archipelago (MAAS 1905), the warm parts of the Indian Ocean (VANHOFFEN 1902 b), the tropical Atlantic from the Cape Verde Islands southwards (VANHOFFEN 1902 b and 1912 a), near Monaco in the Mediterranean (RANSON 1936), the surroundings of the Azores (RANSON 1936); in the western Atlantic it has been taken in one locality, off Georges Bank, 40° 06' N., 68° 06' W., at a comparatively high level, *viz.* in a vertical haul from 300 m (BIGELOW 1917). The localities south-west of Ireland mentioned above are the northernmost localities in which this species has been taken up to now.

Genus *Homoeonema* Maas 1893; Browne 1903.

The genus *Homoeonema*, erected by O. MAAS in 1893, has a very sad history, which is rather difficult to unravel, and the confusion is partly due to MAAS himself who, in some subsequent papers (1897 and, especially, in 1906 b) referred a number of quite

different species to the same genus and thoroughly altered its definition. Several authors have attempted to solve the intricate taxonomic problems which have arisen around this genus, and their opinions are very different. As the genotype, *Homococconeia platygonon* Maas, belongs to the fauna of the North-Atlantic area, I take this opportunity to make a new attempt to settle the question as to what generic names ought to be applied to the several species which from time to time have been provided with the name of *Homococconeia*. I am quite aware that I cannot reach a final result, because some of the species are still unknown to me, so that I must rely on the descriptions in the literature. Nevertheless, the attempt must be made, and I hope that the following considerations may serve to elucidate the matter.

In MAAS' original description (1893) *Homococconeia* was a genus of the family Trachynemidae, and his definition is as follows: "Tentakel alle unter sich gleich, aber nicht in bestimmter, an die Zahl der Radiärkanäle gebundener Anzahl (8 oder 16) vorhanden, sondern zahlreich, 32 bis 64 und mehr."

The actual number of tentacles is usually not a character of generic value, but in connection with other features it is certainly of considerable importance whether the number is fixed or varying. The above definition excludes the genera *Rhopalococconeia* and *Santathoa* Gegenbaur 1856 and *Pantachogon* Maas 1893, as also the subsequently described *Colobococconeia* Vanhoffen 1902. On the other hand, if *Haliereas* and *Aphantha* and their allies are reckoned among the Trachynemidae, as actually done by various authors, these are all covered by the definition of *Homococconeia*.

The genus originally comprised three species: *platygonon* nov., *militare* nov., and *elongatum* which was proposed as a new name for *Rhopalococconeia polydactylum* Haeckel; this latter is, however, most certainly a *Rhopalococconeia*, probably identical with *R. elatum*. *H. militare* is evidently a Trachynemid, as it has a distinct prismatic manubrium with four mouth-lips; it has broad, oval gonads in the distal half of the radial canals which, in the description, are said to be broad, though in the figure (Pl. I fig. 1) they appear to be fairly narrow. VAXHÖFFEN (1902 b p. 56), BIGELOW (1913 p. 14), and THIEL (1936 p. 21) refer this species to *Pantachogon*, because they regard this genus in a wide sense as comprising all Trachynemidae with numerous tentacles, in one row, and all alike, without a stomacal peduncle (BIGELOW 1913). MAAS (1905 p. 51) is opposed to the transmission of *H. militare* to *Pantachogon*, because, in his opinion, *Pantachogon* is mainly characterized by discontinuous gonads which, however, is not correct (see below, p. 19). Considering that the number of tentacles in *H. militare*, according to the figure, is 48 as in *P. haeckeli*, I am inclined to think that the said authors are right in transferring it to *Pantachogon*, but I would not go so far as THIEL and identify it with the species *P. haeckeli*.

Homococconeia platygonon was a small medusa with broad gonads on the proximal portions of the radial canals, which likewise are remarkably broad; the species therefore bears a great resemblance to a Haliereasid, and several authors, including MAAS himself in some of his later papers, really refer it to the Haliereasidae.

A small Norwegian medusa was described by E. T. BROWNE (1903 p. 21, Pl. 2 figs. 2, 3) and by him referred to *H. platygonon* Maas; one of the difficulties in the forthcoming discussions is due to the uncertainty of this identification. The specimens were taken in Skjerstadvjord and Byfjord in Norway; I myself have seen 19 specimens from Herlofjord exactly agreeing with BROWNE'S description (KRAMP & DAMAS 1925 p. 318); two of them are now in the Zoological Museum of Copenhagen, and I have been able to re-examine them. This Norwegian medusa certainly belongs to the Trachynemidae, because it possesses a prismatic manubrium and a mouth opening provided with four short lips, whereas the Haliereasidae all have a large, circular stomach and a wide, circular mouth opening without indication of lips. But it differs from all other Trachynemidae in the shape of the gonads, which are remarkably broad, occupying the proximal halves of the radial canals

and connected with each other around the base of the stomach, it therefore cannot be included in any of the known genera of Trachynemidae, except perhaps in *Homococconeia* sensu Maas 1893.

The stomach and its surroundings are not seen in MAAS' figure of *H. platygonon*, but in the text he expressly states that the stomach is "gleich dem der typischen Formen," i. e. the species of *Rhopalococconeia* described on the preceding pages. It seems to me, therefore, that in spite of the deficiency of the figure we must regard *Homococconeia platygonon* Maas as belonging to the Trachynemidae sensu strictu. When further comparing it with BROWNE'S Norwegian medusa we will find that both of them have a small but distinct apical knob, numerous tentacles, and only four statocysts which, in both of them, are described as vesicular. The only difference is that the circular canal and the distal halves of the radial canals, beyond the broad gonads, in BROWNE'S medusa "are not so broad and conspicuous as those figured by MAAS." If MAAS' medusa were a Haliereasid, it would be the only member of that family with vesicular statocysts; it is also much smaller than any known species of Haliereasidae, and owing to the advanced development of the gonads it cannot be a young individual.

It therefore seems to me very likely that the medusa described by MAAS and by BROWNE really belong to the same species, *Homococconeia platygonon*; and if this is agreed upon, it becomes the only species which can bear the generic name *Homococconeia*, because no other known medusa can be congeneric with it such as we know it from BROWNE'S description. Their identity, if it is true, cannot be decided with certainty; but even if the supposed identity of the two medusae be denied, it seems to me that the generic name ought never to be attached to any other species either of Trachynemidae or of Haliereasidae (see below). In such case *Homococconeia platygonon* Maas must be regarded as an obsolete species, and, if the rules of nomenclature were to be strictly followed, the medusa described by BROWNE should be provided with a new name; but as the Norwegian medusa, which was well described by BROWNE, has ever since been known under the name of *Homococconeia platygonon* and has repeatedly been recorded from new localities (KRAMP & DAMAS 1925, RUNNSTRÖM 1932, BERNSTEIN 1934, JASCHNOV 1939), it seems wise to retain this name and designate BROWNE as its author, adding *Homococconeia platygonon* Maas as a doubtful synonym.

In his paper on the "Albatross" medusa, MAAS (1897 p. 22) described a new species, *Homococconeia typicum*, he referred it to *Homococconeia* because it was a "Trachynemide mit zahlreichen und gleichartigen kurzen Tentakeln," but it is entirely different from *H. platygonon*, and in 1905 MAAS himself transferred it to *Colobococconeia* Vanhoffen 1902 together with a number of specimens taken by the "Siboga" and presumably belonging to the same species; this view has been accepted by the majority of subsequent authors, who only differ as to the question of whether the two species, *typicum* Maas and *sericum* Vanhoffen, are identical or not.

In his "Valdivia" paper (1902 b) VAXHÖFFEN gives a new definition of *Homococconeia*: "Trachynemide mit 8 perkanalen und 72 oder mehr interkanalen Tentakeln, mit am proximalen Ende der Radiärkanäle dem Magen anliegenden Gonaden," in contradistinction to *Pantachogon*, in which the number of intercal tentacles does not exceed 56, and the oval or spindle shaped gonads are placed along the radial canals. He transfers *Homococconeia militare* Maas to *Pantachogon*, and describes two new species of *Homococconeia*—*H. aphantha* and *H. macropaster*—both of them have club-shaped statocysts and numerous (more than 80) tentacles, and the gonads are almost spherical swellings on the eight radial canals close by the stomach. They differ so considerably from *Homococconeia platygonon* (sensu Maas as well as sensu Browne) that they cannot belong to the same genus, and it is also impossible to place them within any other of the genera known by them. It was fully justified, therefore, when MAAS (1906 b) erected a new genus, *Isococconeia* (by VAXSON 1936 altered to *Arctapoda* etc.

non *Homocconema platygonon* Browne 1906.
non *Halicercera platygonon* Ranson 1936.

***Pantachogon militare* (Maas 1893).**

Homocconema militare Maas 1893.
Pantachogon militare Vanhöffen 1902 b.

***Colobonema typicum* (Maas 1897).**

Homocconema typicum Maas 1897.
Colobonema typicum Maas 1905.

***Arctapodema amplum* (Vanhöffen 1902). Genotype.**

Homocconema amplum Vanhöffen 1902 b.
Isonema amplum Maas 1906 b.
Isonema amplum Vanhöffen 1912 a.
Pantachogon amplum Bigelow 1913.
Arctapodema amplum Ranson 1936.

***Arctapodema macrogaster* (Vanhöffen 1902).**

Homocconema macrogaster Vanhöffen 1902 b.
Isonema amplum Maas 1906 b.
Pantachogon macrogaster Bigelow 1913.
Arctapodema? amplum Ranson 1936.

Isonema antarcticum, australe, and tetragonium Vanhöffen 1912 a are Trachynemide of doubtful systematic position.

Species belonging to the family Halicreasidæ.

***Halicercera alba* Vanhöffen 1902. Genotype.**

Halicercera alba Vanhöffen 1902 b.
Homocconema album Maas 1906 a.
Halicercera alba Mayer 1910.
Halicercera alba Vanhöffen 1912 a.

non *Homocconema album* Bigelow 1909 a.

***Halicercera racovitzæ* (Maas 1906).**

Homocconema (*Halicercera*) *racovitzæ* Maas 1906 b.
Homocconema racovitzæ Bigelow 1909 a.
Halicercera racovitzæ Mayer 1910.
Halicercera racovitzæ Vanhöffen 1912 a.

***Halicercera bigelowi* Kramp 1916.**

Homocconema album Bigelow 1909 a.
Halicercera bigelowi Kramp 1917.

? *Homocconema platygonon* Maas 1906 a.
 ? *Homocconema platygonon* Browne 1906.
 ? *Halicercera platygonon* Ranson 1936.

The genus *Homocconema* may now be defined as follows: Trachynemide with eight radial canals; with numerous tentacles structurally alike; with gonads forming a continuous band around the base of the stomach extending outwards along the radial canals, without a stomachal peduncle. Only species: *platygonon* Browne 1903.

***Homocconema platygonon* Browne.**

Pl. II fig. 6

North-Atlantic records:
 ? *Homocconema platygonon* Maas 1893 p. 15, Pl. I fig. 8.
Homocconema platygonon Browne 1903 p. 21, Pl. 2 figs. 2, 3.
 Kramp & Damas 1925 p. 318.
 Runnström 1932 p. 30.

Homocconema platygonon Bernstein 1931 pp. 26, 53.
 Jaschnov 1939 p. 111.

Material:

Herlofjord, Norway, October 1908, 300 m.

Herlofjord is in the neighbourhood of Bergen, where professor D. DAMAS collected 19 small specimens (1-2 mm in diameter) of this medusa. I saw these specimens, when I stayed with prof. DAMAS at Liège in 1920; later on they were sent to Bergens Museum, except two specimens which I secured for our museum at Copenhagen. The shape of the umbrella is not quite as high as shown in the figure by BROWNE, somewhat more like MAAS' figure. The gonads (Pl. II fig. 6) have a somewhat wavy outline and are attached to the subumbrella along fairly narrow lines; on the subumbrella side of the radial canals the gonads are continuous, as also seen in BROWNE'S figure, and the gonads are likewise confluent in the interradia. As a matter of fact, therefore, the gonads form one uninterrupted band around the base of the stomach with outgrowths along the eight radial canals, about halfway to the bell margin. In the specimens examined by me there are about 10-11 tentacles in each octant, thus altogether 80 or more.

The specimens described by BROWNE were taken in Skjerstadfjord (about 67° 4' N.) in April and in Byfjord (near Bergen) in November; the present specimens were found in Herlofjord (= Herdalfjord) in October; RUNNSTRÖM (1932) also found it in the neighbouring Hjeltefjord, and he gives valuable information on its occurrence in Herdalfjord, where he has taken it throughout the year, with maximal occurrence in the autumn and minimum in spring; the largest specimens (2-3 mm) occur in the autumn, but in March only very small specimens (about 1/2 mm) are found. RUNNSTRÖM further states that it belongs to the deep strata in the fjord, mainly between 200 and 400 m, but during the autumn young specimens may be found at higher levels, 50-100 m below the surface. This species thus is quite common in the fjords in the surroundings of Bergen.

It is very interesting that *Homocconema platygonon* also has been found in some localities in the northern part of the Kara Sea, partly in fairly deep water (155-100 m), partly in the upper strata, 17-0 and 10-0 m (BERNSTEIN 1931). This Russian author takes it for granted that the specimen taken by the German "Plankton-Expedition" between Iceland and the south point of Greenland (MAAS 1893) belongs to the same species as the Norwegian specimens, and he discusses its geographical distribution: "The zoogeographical character of this form remains uncertain. Our knowledge is not sufficient to explain its appearance in the Kara Sea. It is possible that this small medusa . . . has escaped the attention of the planktonists, and that it has a greater distribution in the arctic seas. If between Iceland and Greenland it lives in a region of cold currents, it is even possible that *Homocconema platygonon* is an arctic medusa. Future investigations may probably bring this information. So far it thus seems probable that the connection between the western distribution of this medusa and its occurrence in the Kara Sea may be accomplished as well through the Barents Sea as through the Polar Basin."

In the intermediate and deep strata in Hjeltefjord, Herdalfjord, and Byfjord in Norway the temperature of the water is about 7° C, with only slight variations during the year (rarely below 6° and rarely above 8°), and the plankton consists of a mixture of boreal and arctic species. The zoogeographical character of *Homocconema platygonon*, therefore, is not determined by its occurrence in these fjords; its occurrence in the Kara Sea, however, seems to indicate that it is an arctic species. Its presence in the Atlantic between Iceland and the south point of Greenland is not very elucidating, partly because we are not sure of the identity of the medusa found there, partly because the locality is in a mixed area with water masses of different origin. BERNSTEIN is right, therefore, that the zoogeographical character of this species

62° 35' N, 32° 53' W., ²⁷ / ₅ 1925, "Dana" St. 2437, 1900m wire, 1 spec.	
47° 02' N, 31° 45' W., ²⁷⁻²⁸ / ₆ 31.	1201, 2000m 15
	1201, 1000m 7
	1201, 5000m 2
62° 45' N, 16° 01' W., ⁷ / ₅ 1931.	5083, 2000m 6
63° 38' N, 11° 13' W., ¹⁸ / ₅ 1931.	5113, 2000m 4

The morphology of *Colobomena sericeum* is comparatively well known, with only a few uncertain points still awaiting elucidation. BIGELOW (1938, p. 118) is of the opinion that the stump-like appearance of the tentacles is normal, and not the result of mutilation; I do not think this view is correct; in several specimens the tips of the tentacle stumps show distinct marks of being broken, so that evidently the tentacles have been longer. An account of the rate of development of the tentacles and the gonads in relation to size of specimens will be postponed to another occasion, when I have finished the examination of the extensive collections from other geographical areas.

As a rule the manubrium of the preserved specimens is strongly contracted and very short; but sometimes the manubrium has been preserved in a more or less extended condition, and such specimens perfectly resemble that figured by MAAS (1905, Pl. X fig. 62) from the "Siboga" collection. We can therefore state with certainty that the Siboga specimens of *Colobomena* belong to *C. sericeum*. But I must still maintain my former opinion (KRAMP 1924, p. 28) that the "Albatross" specimen, as it is described and figured by MAAS (1897, *Homonocnema typicum*) cannot be referred to the same species, partly on account of the much greater number of tentacles, and also because the gonads are much shorter. RANSON (1936, p. 153) has ably expressed the same opinion of *H. typicum* in the following way: "elle est différente de *Colobomena sericeum* on la description . . . est tellement défectueuse que nous ne devons pas en tenir compte." There is no reason, therefore, to introduce the specific name of *typicum* for the well described and well known medusa *Colobomena sericeum* Vanhoffen.

Remarks on the vertical distribution: In the northern Atlantic as well as in its other areas of distribution, *Colobomena sericeum* is a distinctly bathypelagic medusa. At the "Dana" St. 1201 the hauls containing specimens of this species were all made with the 1½ m ringtrawl with bag of stannum ("S. 150") hauled horizontally during four hours; the greatest number of specimens were taken in the haul with 2000 m wire out, and several of them were young ones (height of bell 4-27 mm), unfortunately, they are all in a bad state of preservation, unsuitable for closer examination; the specimens caught with 1000 and 5000 m wire out were 21-36 and 25-33 mm respectively. This indicates that the young individuals preferably occur at a higher level than the fully developed ones. The other North-Atlantic catches, from which the depth of capture is known, seem to confirm this (e.g. the only specimen taken with 1000 m wire out, "Thor" St. 165 (1905), was only 11 mm high); but further investigations must decide, whether this is the general rule.

Horizontal distribution: The three specimens from "Thor" St. 181 (1906) S. W. of Ireland, were mentioned by me in a previous paper (KRAMP 1921); moreover one specimen is recorded from a locality, 51° 05' N, 26° 08' W., taken by the Norwegian ship "Armauer Hansen" (KRAMP 1920). On the map, textfig. 6, are also included some localities in which this species was taken by the "Michael Sars" North-Atlantic Expedition in 1910, not yet published. BROWNIE (1906) records a specimen from the Bay of Biscay.

Colobomena sericeum is apparently generally distributed in the deep parts of all the great oceans; it has never been taken in the Mediterranean, and its distribution northwards in the Atlantic is evidently limited by the continental shelves and the submarine ridges between Scotland, Iceland, and Greenland. It has not yet

been found in the southern part of Davis Strait, where several others of the bathypelagic medusae from the Atlantic Ocean are known to occur.

Genus *Pantachogon* Maas 1893

Trachynemide with 64 or more tentacles all of one kind; with gonads extending along the radial canals, with free, club-shaped statocysts; without a stomacal peduncle; with the apical outlines of the subumbrella muscular fields forming an entire circle.

In the original definition of this genus, MAAS (1893, p. 17), emphasized the supposed discontinuity of the gonads as the most characteristic feature, and in his "Siboga" paper (1905, p. 51) he maintains that this is not due to the state of preservation; but it must now be regarded as an established fact that it really is so (see KRAMP 1912, p. 78). We can also state that *Pantachogon rubrum* Vanhoffen (1902 b) is identical with the genotype, *P. haeckeli* Maas (1893). Adult specimens of this species always have 64 tentacles; only two more species are known, *P. scottii* Brownie (1910) with about 120 tentacles and *P. militare* (Maas) (formerly *Homonocnema militare*, see above, p. 15). The relation between *Pantachogon* and *Colobomena* has been discussed above (p. 18).

Pantachogon haeckeli Maas.

Pl. II figs. 7-8

North-Atlantic records.

<i>Pantachogon haeckeli</i> Maas 1893, p. 17, Pl. I fig. 2.	
" - " 1901, p. 29.	
<i>rubrum</i> Kramp 1913 a, p. 271.	
" 1914, p. 133.	
" 1920, p. 5.	
" 1921, p. 22.	
<i>haeckeli</i> Kramp 1912, p. 78.	

Material (see the map, textfig. 7).

61° 15' N, 9° 35' W., ²¹ / ₅ 1901, "Thor" St. 93, 1 specimen	
65° 00' N, 28° 10' W., ¹⁹ / ₆ 1901.	152, 5 specimens
61° 30' N, 17° 08' W., ¹¹ / ₇ 1904.	183, 7
57° 17' N, 11° 33' W., ⁷ / ₆ 1905.	71, 3
60° 00' N, 10° 35' W., ²⁹ / ₅ 1905.	165, 1000m wire, 1 spec.
57° 16' N, 9° 55' W., ¹ / ₉ 1905.	167, 1500m " 5
60° 59' N, 22° 29' W., ²⁹ / ₅ 1925, "Dana" St. 2306, 2000m wire, 125 sp.	
60° 20' N, 29° 21' W., ³⁰ / ₅ 1925.	2307, 1500m " 65
59° 21' N, 37° 56' W., ^{1, 2} / ₆ 1925.	2308, 1000m " 2
	2308, 3000m " 1200
62° 35' N, 32° 53' W., ²⁷ / ₅ 1925.	2437, 1900m " 1
63° 19' N, 26° 50' W., ²¹ / ₅ 1928, "Godthaab" St. 1, 1000m	20
47° 02' N, 31° 45' W., ²⁷⁻²⁸ / ₆ 31 "Dana" St.	1201, 2000m 270
	1201, 3000m 650
	1201, 1000m 180
	1201, 5000m 180
62° 23' N, 16° 05' W., ¹⁵ / ₆ 1932.	1102, 2000m 235
	1102, 3000m 250
	1102, 1000m 500
61° 52' N, 35° 30' W., ¹⁵ / ₅ 1933.	1686, 6000m " 1
62° 36' N, 32° 48' W., ¹⁶ / ₅ 1933.	1687, 2000m 250
62° 45' N, 16° 01' W., ⁷ / ₅ 1931.	5083, 2000m " 33
63° 38' N, 11° 13' W., ¹⁸ / ₅ 1931.	5113, 1800m 165
	5113, 2000m 200

Among these numerous specimens very few are in such condition that a closer examination of their morphology can be carried out; in a few cases, however, I have seen tentacles which are somewhat more than stump-like, i. e. they are broken at some distance from their base, and I have no doubt that they have been of greater length.

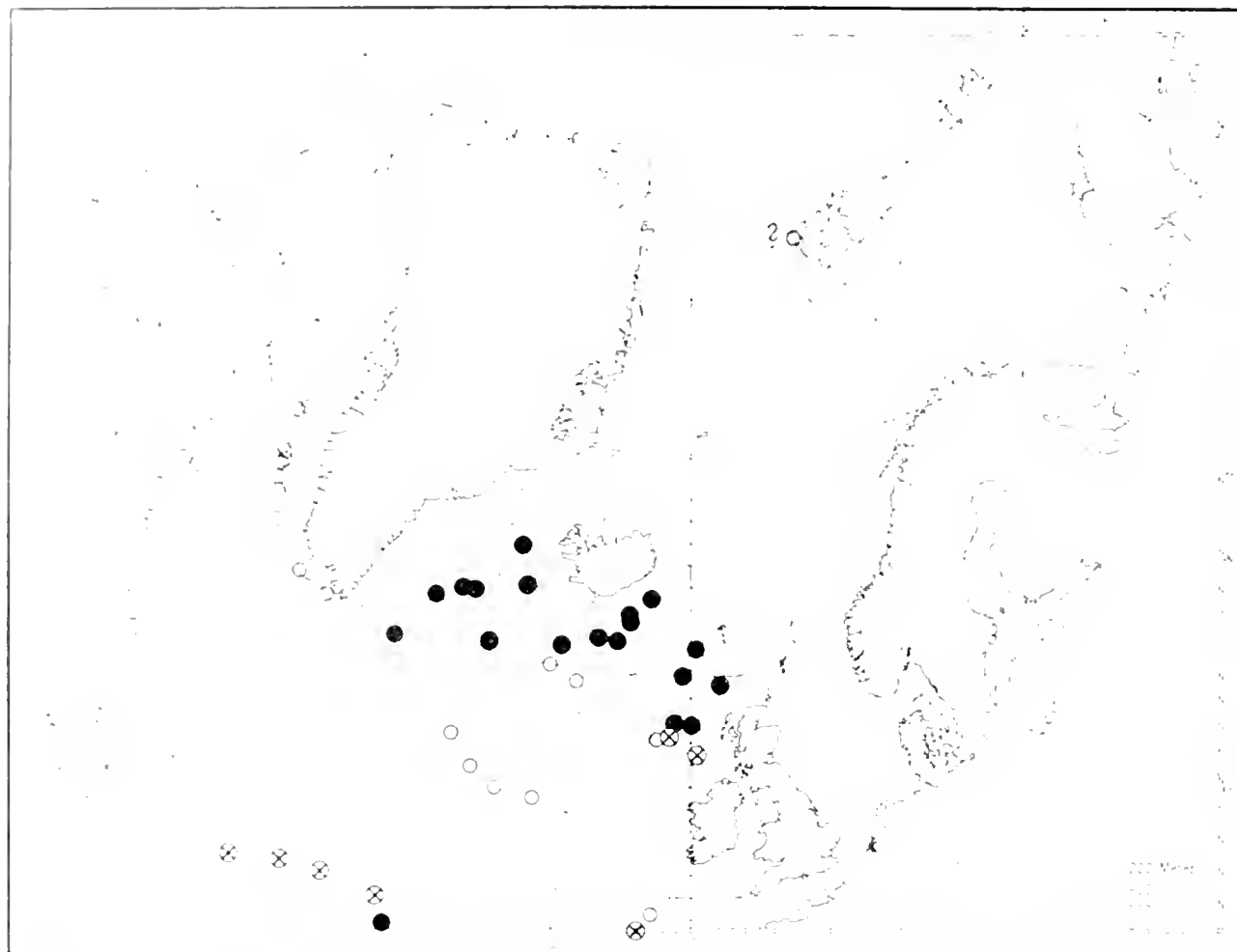


FIG. 7.—The North Atlantic. 1. *Pantchoon hatchelli*. ● new records; ○ previous records; × records not yet published.

of the primary canal, each of them containing one primary canal (Fig. 7, 8).

I have not looked for young specimens which might show the development of the tentacles, but they are in a very good state of preservation. I shall comment on the observations which I was able to make.

In specimens 2 mm high there are 2 tentacles and one canal between every successive pair of radial canals. In specimens 3 mm high there is one, interradial, canal between every pair of tentacles. In a few specimens 3 mm high there are two canals between the radial canals, but it cannot be seen which is the longest. When the height of the specimens is 4 mm there are three tentacles between the canals and one canal between every successive pair of tentacles. In specimens 5 mm high there are four tentacles between the canals and two canals between every successive pair of tentacles. In specimens 6 mm high there are five tentacles between the canals and three canals between every successive pair of tentacles. In specimens 7 mm high there are six tentacles between the canals and four canals between every successive pair of tentacles. In specimens 8 mm high there are seven tentacles between the canals and five canals between every successive pair of tentacles. In specimens 9 mm high there are eight tentacles between the canals and six canals between every successive pair of tentacles. In specimens 10 mm high there are nine tentacles between the canals and seven canals between every successive pair of tentacles. In specimens 11 mm high there are ten tentacles between the canals and eight canals between every successive pair of tentacles. In specimens 12 mm high there are eleven tentacles between the canals and nine canals between every successive pair of tentacles. In specimens 13 mm high there are twelve tentacles between the canals and ten canals between every successive pair of tentacles. In specimens 14 mm high there are thirteen tentacles between the canals and eleven canals between every successive pair of tentacles. In specimens 15 mm high there are fourteen tentacles between the canals and twelve canals between every successive pair of tentacles. In specimens 16 mm high there are fifteen tentacles between the canals and thirteen canals between every successive pair of tentacles. In specimens 17 mm high there are sixteen tentacles between the canals and fourteen canals between every successive pair of tentacles. In specimens 18 mm high there are seventeen tentacles between the canals and fifteen canals between every successive pair of tentacles. In specimens 19 mm high there are eighteen tentacles between the canals and sixteen canals between every successive pair of tentacles. In specimens 20 mm high there are nineteen tentacles between the canals and seventeen canals between every successive pair of tentacles. In specimens 21 mm high there are twenty tentacles between the canals and eighteen canals between every successive pair of tentacles. In specimens 22 mm high there are twenty-one tentacles between the canals and nineteen canals between every successive pair of tentacles. In specimens 23 mm high there are twenty-two tentacles between the canals and twenty canals between every successive pair of tentacles. In specimens 24 mm high there are twenty-three tentacles between the canals and twenty-one canals between every successive pair of tentacles. In specimens 25 mm high there are twenty-four tentacles between the canals and twenty-two canals between every successive pair of tentacles. In specimens 26 mm high there are twenty-five tentacles between the canals and twenty-three canals between every successive pair of tentacles. In specimens 27 mm high there are twenty-six tentacles between the canals and twenty-four canals between every successive pair of tentacles. In specimens 28 mm high there are twenty-seven tentacles between the canals and twenty-five canals between every successive pair of tentacles. In specimens 29 mm high there are twenty-eight tentacles between the canals and twenty-six canals between every successive pair of tentacles. In specimens 30 mm high there are twenty-nine tentacles between the canals and twenty-seven canals between every successive pair of tentacles. In specimens 31 mm high there are thirty tentacles between the canals and twenty-eight canals between every successive pair of tentacles. In specimens 32 mm high there are thirty-one tentacles between the canals and twenty-nine canals between every successive pair of tentacles. In specimens 33 mm high there are thirty-two tentacles between the canals and thirty canals between every successive pair of tentacles. In specimens 34 mm high there are thirty-three tentacles between the canals and thirty-one canals between every successive pair of tentacles. In specimens 35 mm high there are thirty-four tentacles between the canals and thirty-two canals between every successive pair of tentacles. In specimens 36 mm high there are thirty-five tentacles between the canals and thirty-three canals between every successive pair of tentacles. In specimens 37 mm high there are thirty-six tentacles between the canals and thirty-four canals between every successive pair of tentacles. In specimens 38 mm high there are thirty-seven tentacles between the canals and thirty-five canals between every successive pair of tentacles. In specimens 39 mm high there are thirty-eight tentacles between the canals and thirty-six canals between every successive pair of tentacles. In specimens 40 mm high there are thirty-nine tentacles between the canals and thirty-seven canals between every successive pair of tentacles. In specimens 41 mm high there are forty tentacles between the canals and thirty-eight canals between every successive pair of tentacles. In specimens 42 mm high there are forty-one tentacles between the canals and thirty-nine canals between every successive pair of tentacles. In specimens 43 mm high there are forty-two tentacles between the canals and forty canals between every successive pair of tentacles. In specimens 44 mm high there are forty-three tentacles between the canals and forty-one canals between every successive pair of tentacles. In specimens 45 mm high there are forty-four tentacles between the canals and forty-two canals between every successive pair of tentacles. In specimens 46 mm high there are forty-five tentacles between the canals and forty-three canals between every successive pair of tentacles. In specimens 47 mm high there are forty-six tentacles between the canals and forty-four canals between every successive pair of tentacles. In specimens 48 mm high there are forty-seven tentacles between the canals and forty-five canals between every successive pair of tentacles. In specimens 49 mm high there are forty-eight tentacles between the canals and forty-six canals between every successive pair of tentacles. In specimens 50 mm high there are forty-nine tentacles between the canals and forty-seven canals between every successive pair of tentacles. In specimens 51 mm high there are fifty tentacles between the canals and forty-eight canals between every successive pair of tentacles. In specimens 52 mm high there are fifty-one tentacles between the canals and forty-nine canals between every successive pair of tentacles. In specimens 53 mm high there are fifty-two tentacles between the canals and fifty canals between every successive pair of tentacles. In specimens 54 mm high there are fifty-three tentacles between the canals and fifty-one canals between every successive pair of tentacles. In specimens 55 mm high there are fifty-four tentacles between the canals and fifty-two canals between every successive pair of tentacles. In specimens 56 mm high there are fifty-five tentacles between the canals and fifty-three canals between every successive pair of tentacles. In specimens 57 mm high there are fifty-six tentacles between the canals and fifty-four canals between every successive pair of tentacles. In specimens 58 mm high there are fifty-seven tentacles between the canals and fifty-five canals between every successive pair of tentacles. In specimens 59 mm high there are fifty-eight tentacles between the canals and fifty-six canals between every successive pair of tentacles. In specimens 60 mm high there are fifty-nine tentacles between the canals and fifty-seven canals between every successive pair of tentacles. In specimens 61 mm high there are sixty tentacles between the canals and fifty-eight canals between every successive pair of tentacles. In specimens 62 mm high there are sixty-one tentacles between the canals and fifty-nine canals between every successive pair of tentacles. In specimens 63 mm high there are sixty-two tentacles between the canals and sixty canals between every successive pair of tentacles. In specimens 64 mm high there are sixty-three tentacles between the canals and sixty-one canals between every successive pair of tentacles. In specimens 65 mm high there are sixty-four tentacles between the canals and sixty-two canals between every successive pair of tentacles. In specimens 66 mm high there are sixty-five tentacles between the canals and sixty-three canals between every successive pair of tentacles. In specimens 67 mm high there are sixty-six tentacles between the canals and sixty-four canals between every successive pair of tentacles. In specimens 68 mm high there are sixty-seven tentacles between the canals and sixty-five canals between every successive pair of tentacles. In specimens 69 mm high there are sixty-eight tentacles between the canals and sixty-six canals between every successive pair of tentacles. In specimens 70 mm high there are sixty-nine tentacles between the canals and sixty-seven canals between every successive pair of tentacles. In specimens 71 mm high there are seventy tentacles between the canals and sixty-eight canals between every successive pair of tentacles. In specimens 72 mm high there are seventy-one tentacles between the canals and sixty-nine canals between every successive pair of tentacles. In specimens 73 mm high there are seventy-two tentacles between the canals and seventy canals between every successive pair of tentacles. In specimens 74 mm high there are seventy-three tentacles between the canals and seventy-one canals between every successive pair of tentacles. In specimens 75 mm high there are seventy-four tentacles between the canals and seventy-two canals between every successive pair of tentacles. In specimens 76 mm high there are seventy-five tentacles between the canals and seventy-three canals between every successive pair of tentacles. In specimens 77 mm high there are seventy-six tentacles between the canals and seventy-four canals between every successive pair of tentacles. In specimens 78 mm high there are seventy-seven tentacles between the canals and seventy-five canals between every successive pair of tentacles. In specimens 79 mm high there are seventy-eight tentacles between the canals and seventy-six canals between every successive pair of tentacles. In specimens 80 mm high there are seventy-nine tentacles between the canals and seventy-seven canals between every successive pair of tentacles. In specimens 81 mm high there are eighty tentacles between the canals and seventy-eight canals between every successive pair of tentacles. In specimens 82 mm high there are eighty-one tentacles between the canals and seventy-nine canals between every successive pair of tentacles. In specimens 83 mm high there are eighty-two tentacles between the canals and eighty canals between every successive pair of tentacles. In specimens 84 mm high there are eighty-three tentacles between the canals and eighty-one canals between every successive pair of tentacles. In specimens 85 mm high there are eighty-four tentacles between the canals and eighty-two canals between every successive pair of tentacles. In specimens 86 mm high there are eighty-five tentacles between the canals and eighty-three canals between every successive pair of tentacles. In specimens 87 mm high there are eighty-six tentacles between the canals and eighty-four canals between every successive pair of tentacles. In specimens 88 mm high there are eighty-seven tentacles between the canals and eighty-five canals between every successive pair of tentacles. In specimens 89 mm high there are eighty-eight tentacles between the canals and eighty-six canals between every successive pair of tentacles. In specimens 90 mm high there are eighty-nine tentacles between the canals and eighty-seven canals between every successive pair of tentacles. In specimens 91 mm high there are ninety tentacles between the canals and eighty-eight canals between every successive pair of tentacles. In specimens 92 mm high there are ninety-one tentacles between the canals and eighty-nine canals between every successive pair of tentacles. In specimens 93 mm high there are ninety-two tentacles between the canals and ninety canals between every successive pair of tentacles. In specimens 94 mm high there are ninety-three tentacles between the canals and ninety-one canals between every successive pair of tentacles. In specimens 95 mm high there are ninety-four tentacles between the canals and ninety-two canals between every successive pair of tentacles. In specimens 96 mm high there are ninety-five tentacles between the canals and ninety-three canals between every successive pair of tentacles. In specimens 97 mm high there are ninety-six tentacles between the canals and ninety-four canals between every successive pair of tentacles. In specimens 98 mm high there are ninety-seven tentacles between the canals and ninety-five canals between every successive pair of tentacles. In specimens 99 mm high there are ninety-eight tentacles between the canals and ninety-six canals between every successive pair of tentacles. In specimens 100 mm high there are ninety-nine tentacles between the canals and ninety-seven canals between every successive pair of tentacles.

When the height of the tentacles does not exceed 2 mm, it appears that in specimens 2 mm high there is one, interradial, canal between every successive pair of tentacles. I have not seen any specimens of the form that there are two canals between every successive pair of tentacles. In specimens 3 mm high, exceptionally, there are two canals between every successive pair of tentacles. In specimens 4 mm high there are three canals between every successive pair of tentacles. In specimens 5 mm high there are four canals between every successive pair of tentacles. In specimens 6 mm high there are five canals between every successive pair of tentacles. In specimens 7 mm high there are six canals between every successive pair of tentacles. In specimens 8 mm high there are seven canals between every successive pair of tentacles. In specimens 9 mm high there are eight canals between every successive pair of tentacles. In specimens 10 mm high there are nine canals between every successive pair of tentacles. In specimens 11 mm high there are ten canals between every successive pair of tentacles. In specimens 12 mm high there are eleven canals between every successive pair of tentacles. In specimens 13 mm high there are twelve canals between every successive pair of tentacles. In specimens 14 mm high there are thirteen canals between every successive pair of tentacles. In specimens 15 mm high there are fourteen canals between every successive pair of tentacles. In specimens 16 mm high there are fifteen canals between every successive pair of tentacles. In specimens 17 mm high there are sixteen canals between every successive pair of tentacles. In specimens 18 mm high there are seventeen canals between every successive pair of tentacles. In specimens 19 mm high there are eighteen canals between every successive pair of tentacles. In specimens 20 mm high there are nineteen canals between every successive pair of tentacles. In specimens 21 mm high there are twenty canals between every successive pair of tentacles. In specimens 22 mm high there are twenty-one canals between every successive pair of tentacles. In specimens 23 mm high there are twenty-two canals between every successive pair of tentacles. In specimens 24 mm high there are twenty-three canals between every successive pair of tentacles. In specimens 25 mm high there are twenty-four canals between every successive pair of tentacles. In specimens 26 mm high there are twenty-five canals between every successive pair of tentacles. In specimens 27 mm high there are twenty-six canals between every successive pair of tentacles. In specimens 28 mm high there are twenty-seven canals between every successive pair of tentacles. In specimens 29 mm high there are twenty-eight canals between every successive pair of tentacles. In specimens 30 mm high there are twenty-nine canals between every successive pair of tentacles. In specimens 31 mm high there are thirty canals between every successive pair of tentacles. In specimens 32 mm high there are thirty-one canals between every successive pair of tentacles. In specimens 33 mm high there are thirty-two canals between every successive pair of tentacles. In specimens 34 mm high there are thirty-three canals between every successive pair of tentacles. In specimens 35 mm high there are thirty-four canals between every successive pair of tentacles. In specimens 36 mm high there are thirty-five canals between every successive pair of tentacles. In specimens 37 mm high there are thirty-six canals between every successive pair of tentacles. In specimens 38 mm high there are thirty-seven canals between every successive pair of tentacles. In specimens 39 mm high there are thirty-eight canals between every successive pair of tentacles. In specimens 40 mm high there are thirty-nine canals between every successive pair of tentacles. In specimens 41 mm high there are forty canals between every successive pair of tentacles. In specimens 42 mm high there are forty-one canals between every successive pair of tentacles. In specimens 43 mm high there are forty-two canals between every successive pair of tentacles. In specimens 44 mm high there are forty-three canals between every successive pair of tentacles. In specimens 45 mm high there are forty-four canals between every successive pair of tentacles. In specimens 46 mm high there are forty-five canals between every successive pair of tentacles. In specimens 47 mm high there are forty-six canals between every successive pair of tentacles. In specimens 48 mm high there are forty-seven canals between every successive pair of tentacles. In specimens 49 mm high there are forty-eight canals between every successive pair of tentacles. In specimens 50 mm high there are forty-nine canals between every successive pair of tentacles. In specimens 51 mm high there are fifty canals between every successive pair of tentacles. In specimens 52 mm high there are fifty-one canals between every successive pair of tentacles. In specimens 53 mm high there are fifty-two canals between every successive pair of tentacles. In specimens 54 mm high there are fifty-three canals between every successive pair of tentacles. In specimens 55 mm high there are fifty-four canals between every successive pair of tentacles. In specimens 56 mm high there are fifty-five canals between every successive pair of tentacles. In specimens 57 mm high there are fifty-six canals between every successive pair of tentacles. In specimens 58 mm high there are fifty-seven canals between every successive pair of tentacles. In specimens 59 mm high there are fifty-eight canals between every successive pair of tentacles. In specimens 60 mm high there are fifty-nine canals between every successive pair of tentacles. In specimens 61 mm high there are sixty canals between every successive pair of tentacles. In specimens 62 mm high there are sixty-one canals between every successive pair of tentacles. In specimens 63 mm high there are sixty-two canals between every successive pair of tentacles. In specimens 64 mm high there are sixty-three canals between every successive pair of tentacles. In specimens 65 mm high there are sixty-four canals between every successive pair of tentacles. In specimens 66 mm high there are sixty-five canals between every successive pair of tentacles. In specimens 67 mm high there are sixty-six canals between every successive pair of tentacles. In specimens 68 mm high there are sixty-seven canals between every successive pair of tentacles. In specimens 69 mm high there are sixty-eight canals between every successive pair of tentacles. In specimens 70 mm high there are sixty-nine canals between every successive pair of tentacles. In specimens 71 mm high there are seventy canals between every successive pair of tentacles. In specimens 72 mm high there are seventy-one canals between every successive pair of tentacles. In specimens 73 mm high there are seventy-two canals between every successive pair of tentacles. In specimens 74 mm high there are seventy-three canals between every successive pair of tentacles. In specimens 75 mm high there are seventy-four canals between every successive pair of tentacles. In specimens 76 mm high there are seventy-five canals between every successive pair of tentacles. In specimens 77 mm high there are seventy-six canals between every successive pair of tentacles. In specimens 78 mm high there are seventy-seven canals between every successive pair of tentacles. In specimens 79 mm high there are seventy-eight canals between every successive pair of tentacles. In specimens 80 mm high there are seventy-nine canals between every successive pair of tentacles. In specimens 81 mm high there are eighty canals between every successive pair of tentacles. In specimens 82 mm high there are eighty-one canals between every successive pair of tentacles. In specimens 83 mm high there are eighty-two canals between every successive pair of tentacles. In specimens 84 mm high there are eighty-three canals between every successive pair of tentacles. In specimens 85 mm high there are eighty-four canals between every successive pair of tentacles. In specimens 86 mm high there are eighty-five canals between every successive pair of tentacles. In specimens 87 mm high there are eighty-six canals between every successive pair of tentacles. In specimens 88 mm high there are eighty-seven canals between every successive pair of tentacles. In specimens 89 mm high there are eighty-eight canals between every successive pair of tentacles. In specimens 90 mm high there are eighty-nine canals between every successive pair of tentacles. In specimens 91 mm high there are ninety canals between every successive pair of tentacles. In specimens 92 mm high there are ninety-one canals between every successive pair of tentacles. In specimens 93 mm high there are ninety-two canals between every successive pair of tentacles. In specimens 94 mm high there are ninety-three canals between every successive pair of tentacles. In specimens 95 mm high there are ninety-four canals between every successive pair of tentacles. In specimens 96 mm high there are ninety-five canals between every successive pair of tentacles. In specimens 97 mm high there are ninety-six canals between every successive pair of tentacles. In specimens 98 mm high there are ninety-seven canals between every successive pair of tentacles. In specimens 99 mm high there are ninety-eight canals between every successive pair of tentacles. In specimens 100 mm high there are ninety-nine canals between every successive pair of tentacles.

between the four mouth lips, whereas there is always a tentacle exactly opposite the "primary" canals which agree in position to the lips; as, however, the cases observed are very few, I am not sure that it is the general rule.

Occurrence in the North Atlantic.—MAYNARD (1893) originally recorded *Pantchoon hatchelli* from a locality south of Iceland, Lat. 60° N., and in 1901 he referred, with some doubt, a mutilated specimen from Spitzbergen to the same species. In three papers (KRAMP 1913, 1911, and 1912) I have dealt with its occurrence in the waters west of Greenland, and moreover I have seen a number of specimens collected by the "Armauer Hansen" in six localities in deep water west and east of the Rockall plateau (KRAMP 1920). Some few specimens, taken by the "Thor" south west of Ireland in 1906 and in the Bay of Biscay in 1909, were recorded by me in 1921. On the map, textfig. 7, are also given the localities, at which it was taken by the "Michael Sars" North-Atlantic Expedition in 1910 within the area dealt with here.

The species seems to be generally distributed over the deep-sea areas of the northern Atlantic, in the north-eastern part the distribution is limited by the continental shelves and the submarine ridges Scotland-Iceland-Greenland (apart from the doubtful record from Spitzbergen, MAYNARD 1901); the finds in this area are all outside the 600 m line. In the waters west of Greenland, on the other hand, it has occasionally been taken in more shallow water. It has its main occurrence in the deep strata, but is not strictly limited to the deep-sea. Within the area under consideration it has been taken in altogether 75 hauls from which the depth of the haul is known; an enumeration of the catches made by the "Tjalfe", "Godthaab", "Thor", "Dana", "Armauer Hansen",

and "Michael Sars" will show that 65 catches were made with 1000-5000 m wire out (about 650-3300 m below the surface); west of Greenland the "Tjalle" took numerous specimens in two hauls with 800-900 m wire out, and on five occasions it has been taken with 600 m wire out. Exceptionally it has even been found at still higher levels: 100 m wire ("Godthaab" St. 18, 1 specimen), 300 m wire ("Godthaab" St. 5, 1 specimen), and 100 m wire ("Michael Sars" St. 81, east of the Newfoundland Bank, 9 specimens). In hauls with less than 800 m wire the number of specimens caught has always been very small, but from about 500 m below the surface downwards the species is frequently taken in great abundance: at the "Dana" St. 2308, east of the south point of Greenland, no less than about 1200 specimens were taken in a haul of 2 hours' duration with the 2 m ringtrawl with 3000 m wire out. *Pantachogon haeckeli* is thus a predominantly bathypelagic medusa, though occasionally it may ascend into higher strata, especially in colder regions: in certain areas, e.g. in the waters west of Greenland, it may therefore be carried with the currents into the coastal waters: thus it was taken by the "Godthaab" expedition near the south point of Greenland and off the entrance to Hudson Strait, and it is also able to cross the submarine ridge in Davis Strait and penetrate into the deep basin of Baffin Bay (for details, see KRAMP 1912 pp. 78-79).

In the hauls from the "Dana" the specimens caught in the stramin-nets varied in size from 3 or 5 mm to 10 or 11 mm in diameter; specimens more than 11 mm wide were found on two occasions only: St. 1201, 5000 m wire (12 mm), and St. 2308, 3000 m wire (13 mm). (In Baffin Bay, "Godthaab" St. 51, one specimen 11 mm wide was taken in a haul with 3000 m wire out). The measurements of the specimens present no evidence of a difference in the vertical distribution of the various size classes of individuals.

Geographical distribution: *Pantachogon haeckeli* is generally distributed over the deep parts of all the oceans from the slopes of the antarctic continent to the Bering Sea and to the submarine ridges separating the North-Atlantic deep-sea area from the arctic basins.

Genus *Crossota* Vanhöffen 1902.

Trachynemida: with 8 or more radial canals; with numerous densely crowded tentacles all of one kind; with pendent, sausage-shaped gonads on the radial canals; with free, club-shaped statocysts; with or without a short stomachal peduncle.

Since this genus was established by VANHÖFFEN (1902 b p. 72) the supposed multiserial position of the tentacles has been emphasized by all subsequent authors as the most characteristic feature of the genus. In my paper on the "Godthaab" medusa (KRAMP 1912 p. 80) I expressed a different opinion and doubted the correctness of one of VANHÖFFEN's figures (Pl. 12 fig. 17), and now, having examined a large number of well-preserved specimens of the genotype, *Crossota brunnea* Vanhöffen, I can state that I was entirely right in my view: the tentacles of *Crossota* are not placed in several rows; they all make their appearance on the bell margin proper in close connection with the ring canal, but during growth their basal portion becomes adnate to the lower margin of the exumbrella; in older tentacles, therefore, the point of issue of the free portion is gradually displaced a little outwards on the exumbrella. If this should be called an arrangement in several rows, it is at least effected in quite another way than that illustrated by VANHÖFFEN in his figure 17, in which the youngest tentacles are the farthest removed from the ring canal, implanted in the jelly of the exumbrella without the slightest connection with the ring canal. In the other species which I have

seen, the arrangement is the same as in *C. brunnea*, but the displacement of the free portion is less pronounced (see below).

VANHÖFFEN gave no definition of the genus but a comparatively thorough description of the species *Crossota brunnea*, which was found to be widely distributed in the deep-sea of the Atlantic and Pacific Oceans between the equator and about Lat. 60° S. He also mentioned another new species, *C. norvegica*, from the Norwegian Sea, but he gave no description of it, beyond stating that it was characterized by its cherry red colour and smaller size as compared with *C. brunnea*.

BIGELOW (1909 a p. 434) showed that *C. brunnea* also occurs in the tropical Pacific; he considered *C. norvegica* (which he only knew from VANHÖFFEN's insufficient description) a synonym of *C. brunnea* and placed the genus in the family Pectylidae, whereas MAYER (1910 p. 395) placed it near *Botryocoma*, *Halocreas*, and *Homonocoma*.

In 1913 a (pp. 17 ff.) BIGELOW found that *Crossota* was not related to *Ptychopastera* but to *Aglantha* and *Aglaura*, and he described two new species from the north-western Pacific, *C. alba* and *C. pedunculata*, the latter characterized by the presence of a short, solid, gelatinous peduncle. He also found numerous specimens of a *Crossota* which he called *C. brunnea* var. *norvegica*. I have seen two of these specimens, kindly sent to me by professor BIGELOW; they are very similar to the North-Atlantic species *C. rubrobrunnea*, which I described in the same year (KRAMP 1913 a p. 273), and undoubtedly belong to the same species (see below).

When I described this species from the deep strata in the Davis Strait, I saw that it resembled an *Aglantha* in the pendent, sausage-shaped gonads, the ribbon-like radial canals, and the shape of the stomach, and I even thought I saw a trace of a stomachal peduncle; it also had a great resemblance to "*Melicerium proboscifer*" MAAS (1897 p. 19, Pl. II figs. 5-7), which was provided with a long gelatinous peduncle and undoubtedly was closely allied to *Aglantha*; I therefore also referred my new species to *Aglantha*. Later on I was very sorry that I had made such a mistake, but now I am aware that I was not altogether wrong: *Crossota* is undoubtedly closely allied to *Aglantha*. Shortly after my paper had been published, I received a letter from professor VANHÖFFEN who was inclined to think that my "*Aglantha rubrobrunnea*" was identical with his *Crossota norvegica*, and I, being young and inexperienced, naturally complied with such an authority and in a subsequent paper (KRAMP 1920 p. 5) referred some other North-Atlantic specimens, collected by the Norwegian vessel "Armauer Hansen", to *C. norvegica*. At about the same time I found in the collections of the Zoological Museum of Copenhagen some specimens of a larger *Crossota* with a cherry red colour and with 10-11 radial canals, collected by the "Ingolf" Expedition in deep water in the Norwegian Sea, north-east of Iceland; they were labelled by G. M. R. LEVINSEN *Crossota* n. sp. How I wished to know what VANHÖFFEN'S *C. norvegica* was really like! My curiosity was soon disposed of, for during my stay with professor D. DAMAS at Liège in Belgium in the autumn of 1920 I saw VANHÖFFEN'S original specimens and found that they had 10-12 radial canals (a fact overlooked by VANHÖFFEN) and in every respect were similar to the specimens which I had seen in our own museum. This discovery was published in KRAMP & DAMAS (1925 p. 317) with an addition that *Crossota rubrobrunnea* Kramp was a distinct species occurring in the northern Atlantic, but not in the Norwegian Sea nor in the Polar Sea. Unfortunately, the specimens of *C. rubrobrunnea* from the "Armauer Hansen" (KRAMP 1920) are still mentioned under *C. norvegica* in the "Nordisches Plankton" by BROWN (1929 p. 597).

As mentioned above (p. 12), UCHIDA (1928 p. 80) refers *Crossota* to the subfamily Aglaurinae.

RANSON (1936 pp. 160 ff.) describes a specimen of *C. rubrobrunnea* from the Bay of Biscay and discusses the genus, which he places near *Actapodema*; he will not admit the presence of a gelatinous peduncle in *C. pedunculata* Bigelow, and he is cor-

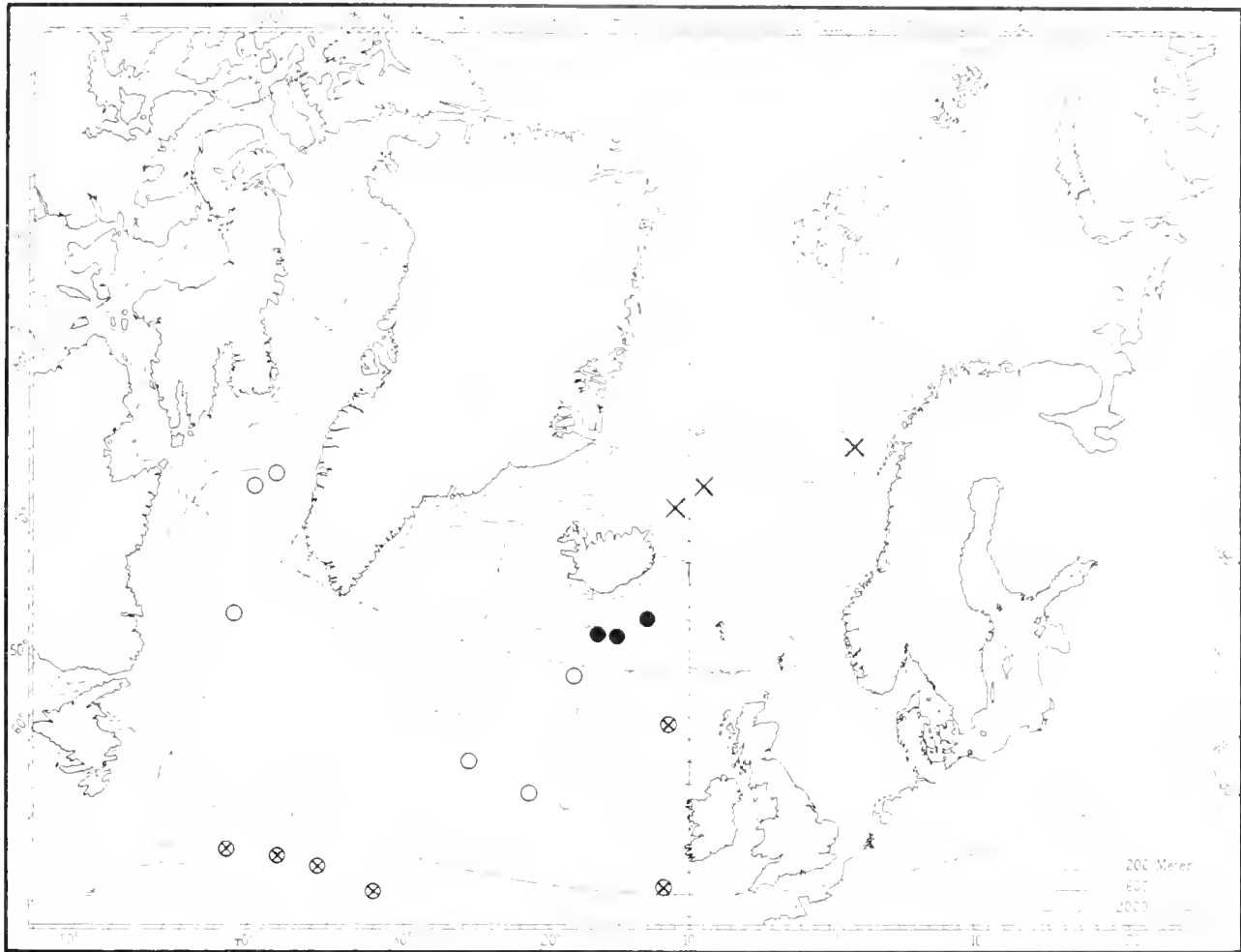


Fig. 8. Distribution in the northern Atlantic of: ● *Crossota rufobrunnea*, new records; ○ *Crossota rufobrunnea*, previous records; × *Crossota rufobrunnea*, records not yet published; ⊗ *Crossota unceyfica*.

Crossota brunnea Bigelow 1938 p. 119.
rufobrunnea Kramp 1942 p. 79.

North-Atlantic records:
 Kramp 1913, 1920, 1912; Ranson 1936.

Material (see the map, textfig. 8):

61 31°N, 19°05'W., ¹⁹7/1901. "Thor" St. 180. 1800m wire. 2 spec.
 61 30°N, 17°08'W., ¹¹7/1901. " " 183. 1 specimens
 62 23°N, 16°05'W., ²⁵8/1932. "Dana" St. 1102. 3000m wire. 35 spec.
 " " " " 4102. 1000m " " 220

Description: Umbrella (Pl. VI fig. 5), when moderately contracted, somewhat higher than a hemisphere, up to 15 mm in diameter and 10 mm in height, evenly rounded; gelatinous substance fairly thin, about 1.5 mm at the apex, gradually tapering in thickness towards the bell margin. Exumbrella with about 20 meridional grooves in each octant, running from the bell margin upwards and all of equal length, terminating at about the level of the base of the manubrium, leaving the top of the umbrella smooth. The circular musculature of the subumbrella is weak. Velum broad, but thin and with weak muscles.

Manubrium (Pl. III figs. 1-2) bottle-shaped, its length about one-third of the height of the subumbrella cavity, varying according to state of contraction. The basal part, the stomach proper, is rather broad, the mouth-tube somewhat narrower and shorter than the stomach, mouth with four short but well-developed, out-

turned lips. In transverse section the mouth-tube is quadrangular with thin walls, in each corner is a string of large, vacuolated endoderm cells covered by a thin ectoderm and visible on the external side as a prominent edge, on the internal side marked by a sharp groove (Pl. III figs. 1, 2, 3); these four strings and the corresponding grooves are continued to the points of the lips and upwards to the top of the stomach. The attachment of the stomach to the subumbrella is star shaped, corresponding to the eight radial canals. The walls of the stomach are rather thin and provided with two whorls of deep folds. Those of the upper whorl are eight small, narrow lappets hanging down into the cavity of the stomach (Pl. III fig. 2); their inner edges almost meet in the centre (Pl. III fig. 4), they are hollow, and their cavities communicate with the bell cavity through eight small, radiating fissures in the apical wall of the stomach between the radial canals. The lower whorl consists of eight large elongated pouches, placed immediately below those of the upper whorl, sharply marked off from them, and gradually tapering downwards towards the distal portion of the stomach (Pl. III fig. 2). On the external side of the stomach they are seen as eight deep, open grooves (Pl. III fig. 1); when the stomach is strongly contracted in circular direction, these grooves may become so narrow as to be termed fissures, and the spaces between them may then protrude like eight longitudinal ridges. The large pouches of the lower whorl are not quite equidistant, the four spaces with the above mentioned four prominent edges being somewhat broader than the four other spaces (Pl. III fig. 4). The endoderm of the pouches is slightly thickened and has a papillose surface.

Externally no trace of a stomacheal peduncle is observed,

the upper pouches, as seen in PL. III, fig. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100. The longitudinal sections, PL. III, figs. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100. The transverse sections, PL. III, figs. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100. The sections through the upper pouches, PL. III, figs. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100. The sections through the lower pouches, PL. III, figs. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100.

PL. III, fig. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100.

PL. III, fig. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100.

PL. III, fig. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100.

PL. III, fig. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100.

continuous epithelium, but in the lateral bands this epithelium is gradually dissolved into a meshwork of branched cells winding themselves in the spaces between the flattened, pigmented cells of the endoderm lamella, which, outside the border of the "lateral band" becomes directly adnate to the thin supporting lamella separating it from the muscular layer of the subumbrella. The "lateral bands" of the radial canals are the portions where two layers of endoderm cells are present without being separated by a cavity. Moreover the bands are made conspicuous by a difference in configuration of the cells of the endoderm lamella, the exumbrellar, pigmented epithelium of the canal consists of small cells, fairly high and with large, round vacuoles; in the lateral bands the pigmented cells are flattened, large, roundish, and separated by rather broad spaces partly filled by the branching non-pigmented cells, outside the bands the cells are somewhat smaller, with wavy outlines and closely set. Sometimes the lateral bands of the radial canals are very sharply marked out by a clear line on each side, this is undoubtedly due to artificial rupture of the tissues caused by strong contraction at the moment of fixation. In the material available no specially developed musculature may be observed in the lateral bands of the radial canals.

The ring canal is fairly broad; its subumbrellar epithelium is unpigmented and similar to that of the radial canals.

The eight sausage-shaped gonads are attached to the eight radial canals near the stomach (PL. III, fig. 2); they are hollow, their internal cavity communicating with the radial canals. In fully developed specimens the gonads are somewhat shorter than the manubrium, in young individuals they are small globular swellings.

The length of the tentacles cannot be decided, as in all the specimens available they are broken off at some distance from their base, but from what remains they seem to have been rather long. The abaxial side of their basal portion is adnate to the lower margin of the exumbrella (PL. IV, figs. 1, 2), and the point of issue of their free portion is therefore somewhat removed from the ring-canal, mainly in the oldest tentacles, but the displacement is less pronounced in this species than in *C. brunnia*. The tentacles are provided with a large and broad endodermal spur protruding into the gelatinous substance of the umbrella in front of the ring-canal. Tentacles in different stages of development are always present simultaneously, but they are not regularly alternating. The tentacles are usually separated from each other by spaces similar to their own breadth.

The total number of tentacles in fully developed individuals is usually about 250. It is rarely possible to count the tentacles within every single octant, partly because some of the radial canals in badly preserved specimens often cannot be exactly located right out to the bell margin. The radial canals are not always quite equidistant, and the number of tentacles per octant varies accordingly; in a specimen 10 mm high with altogether 240 tentacles the number per octant varied from 16 to 37; in some other specimens of similar size I have counted the following numbers:

27	22	23	26	28	25	28	26	205
36	34	35	26	34	34	33	25	251

The statocysts are evidently readily lost, even among the best preserved specimens I have seen only a few statocysts; they agree with those found by BIERVOR (1913, p. 19 and 50, PL. 9, fig. 12) in the Pacific specimens and in *C. alba*; two of them are seen in PL. IV, figs. 3, 4.

Colour (PL. VI, fig. 5). Fresh or well preserved specimens are so densely pigmented that they are almost completely opaque; the colour is a deep reddish brown and, as mentioned above, it is due to dense accumulations of minute pigment granules in the cells. The pigmentation comprises the following tissues: the

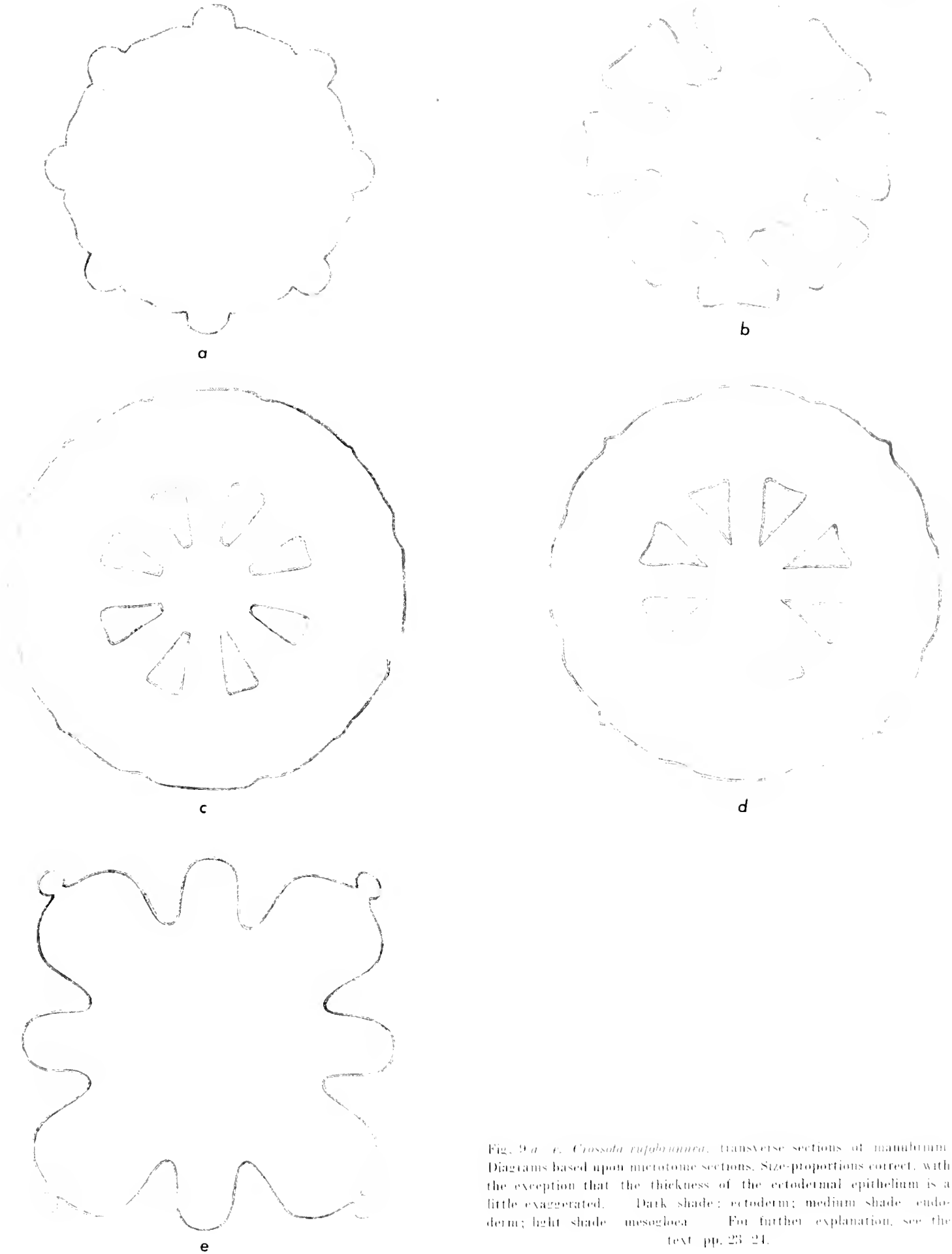


Fig. 9a-e. *Crossota rajabiannea*, transverse sections of manubrium. Diagrams based upon microtome sections. Size-proportions correct, with the exception that the thickness of the ectodermal epithelium is a little exaggerated. Dark shade: ectoderm; medium shade: endoderm; light shade: mesogloea. For further explanation, see the text pp. 23-24.

of the tentacles, and the epithelium of the oral groove from the periphery to the centre, which the octopus-like animals do not do; the tentacles, especially the older ones, in poor condition and the oral groove is frequently rubbed off. The colour is as in my original description (textfig. 196), but that of the distal part of the tentacles of the specimens white with four red spots, due to abrasion; the pigmented epithelium of the oral groove, except on the four prominent edges, is also rubbed off. The best preserved specimens are 196, reddish-brown.

Diagnosis.—*Crossota* (cap. textfig. 8). Davis Strait and north-western part of Greenland, in the deep strata (Arctic Ocean), with temperatures between 3.1 and 3.8°, partly between 3.0 and 3.5° (Egede, 1909 and Godthaab, 1928), South Atlantic, 2000–3000 m deep, with 1800–4000 m wire out ("Thor" stat. 118; De Wit, 1932; West of the Rockall Bank, in hauls with 1500–3000 m wire out; Armand Hansen, 1913). Also collected in the waters of the Norwegian expedition with the "Michael Sars" (1910, not published), in the channel east of Rockall Stat. 200–8 W of Isora 1–8, 391, four localities east of the New Zealand, Bay of Sars, 80, 81, 82, and 84; these localities are included in textfig. 8, further in the surroundings of the Azores, 87, 87, 86, and between the Azores and Bermuda 87, 61, 3, 11, N. 17, 52, W. the specimens collected by the "Michael Sars" were taken in hauls with 1500–3000 m wire out. Material of this species has been found in the Bay of Biscay, off Azores, 46, 29, 15, N. 5, 19, 30, W. in a vertical haul 2650.0 m (Paris, 1912), and it was undoubtedly also this same species which was taken in some localities near the Bermuda Islands, in hauls between 1097.0 and 1829.0 m, and mentioned by Bigelow (1938) as *C. hannaea*. The distribution of *Crossota norvegica* covers the entire deep-sea area of the northern Atlantic, from as far south as about 30° N. It is a well-marked bathypelagic form and it never crosses the submarine ridges separating the Arctic basin from the Arctic deep-sea basins (the Norwegian Sea and Baffin Bay). In the northern Pacific Ocean, the species *Crossota americana* (Borrow, 1913), it was mainly collected in hauls from 300 fms. (459 m) to the surface, on the continental shelf (De Wit, 1932, p. 207).

***Crossota norvegica* Vahlbøen**
Pl. IV, fig. 5, 6

- 1. *Crossota norvegica* Vahlbøen, 1902, p. 75.
- 2. *Crossota norvegica* Krøyer & De Wit, 1925, p. 317.
- 3. *Crossota norvegica* De Wit, 1929, p. 307.
- 4. *Crossota norvegica* De Wit, 1932a, p. 153.
- 5. *Crossota norvegica* Bigelow, 1933, p. 48.
- 6. *Crossota norvegica* Krøyer, 1929, p. 7.
- 7. *Crossota norvegica* Krøyer, textfig. 8.
- 8. *Crossota norvegica* De Wit, 1937, St. 118, 3 specimens.
- 9. *Crossota norvegica* Vahlbøen, 1902, p. 120, 2.

Material. — In poor condition, the others in good condition; details are well preserved. *Diagnosis.*—Umbrella, dome-shaped, 18–20 mm in diameter and 18 mm high; oral groove, substance fairly thin; manubrium, 16–18 mm long, broad, about 16–18 mm wide; velum fairly broad, 10–11 mm long, 10–11. In two of

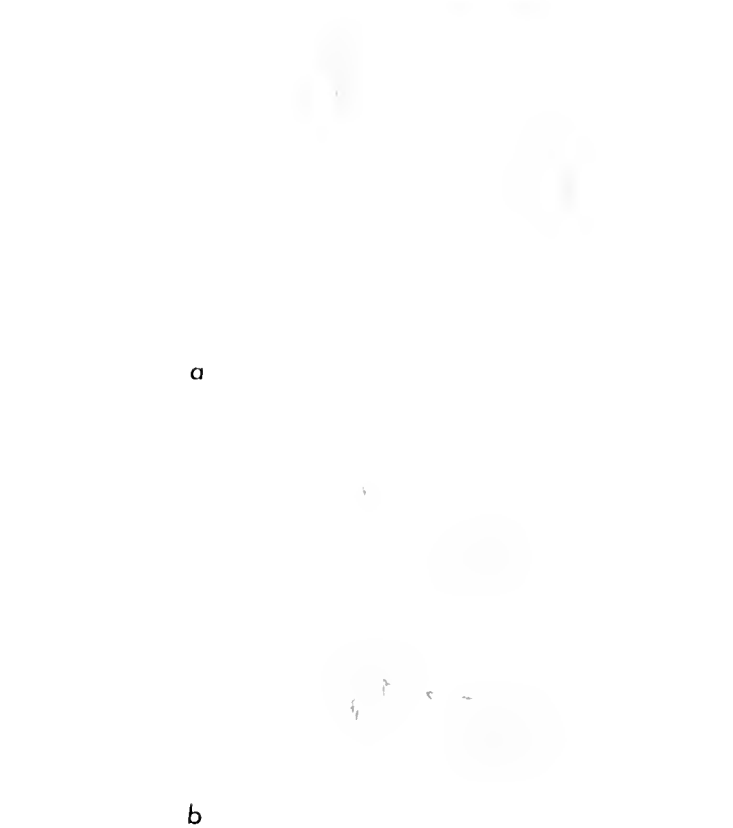


Fig. 10. *a, b*, *Crossota norvegica*. Mode of issue of the radial canals in two specimens: *a* with 11 radial canals and one blind canal ("Ingolf" stat. 118); outline of stomach indicated; *b* with 11 radial canals ("Ingolf" stat. 120).

the specimens, with 10 and 11 radial canals; they all issue separately from the centre of the subumbrella; in two other specimens (textfig. 10*a* and *b*) one or more of the canals are seen branching off from the others at a short distance from their proximal ends. One of these specimens (textfig. 10*a*) has 11 complete radial canals reaching the ring-canal, and also one short blind canal. The terminal points of the radial canals, where they join the ring-canal, are not quite equidistant.

The manubrium (Pl. IV fig. 6) is bottle-shaped, about one-third to two-fifths as long as the height of the bell cavity. The mouth has 5–7 outturned lips (the number of lips being about half the number of radial canals); from each of the lips a prominent ridge proceeds upwards over the mouth tube; in the proximal portion of the stomach proper some of these ridges become more or less irregularly divided. The stomach is provided with similar invaginations as those described above in *C. rubrorannea* and also seen in *C. hannaea*, but in *C. norvegica* they are variable in number and irregular in position.

The gonads are sausage-shaped, about half as long as the manubrium, attached to the radial canals very near the stomach; the number of gonads is equal to the number of radial canals.

The tentacles could not be counted exactly; in the best preserved specimen there are about 275. The displacement of the point of issue of the older tentacles is not very pronounced. The basal endodermal spur is comparatively long, more elongated than in *C. rubrorannea*. Statocysts are not observed.

Colour. This medusa has about the same deep reddish-brown colour as that found in *C. rubrorannea*.

Variation. The seven specimens of this species observed up to now have the following dimensions:

	VANHÖFFEN'S original specimens		"Ingolf" St.				
	118	118	118	120	120	120	
Diameter, mm.	20	20	15	17	19	18	20
Number of radial canals. . .	10	12	?	11	10	11	11

Distribution: The two localities, where *C. norvegica* was taken by the "Ingolf" expedition, are north-east of Iceland in the deep basin of the Norwegian Sea; at St. 120, nearest to Iceland, the depth of the bottom was 1666 m, at St. 118 it was 1996 m. The labels do not tell us how far below the surface of the water the specimens were captured, but according to the journal of the expedition they must have been caught in the trawl, either near the bottom of the sea or at some higher level during the hauling in of the trawl. VANHÖFFEN'S original specimens were found in the eastern part of the Norwegian Sea, 69°13' N., 10°40' E., west of the Lofoten, at a depth of about 1000 m. We may suppose, therefore, that the "Ingolf" specimens were likewise taken in the deep strata where the temperature of the water is very low; at the "Ingolf" St. 118 and 120 the temperature was below 0° from about 500 metres downwards, being ± 1°0 at the bottom. *C. norvegica* may thus be designated as an arctic deep-sea medusa.

Genus *Aglantha* Haeckel 1879.

Trachynemide with a well-developed, gelatinous stomachal peduncle; with eight radial canals; with numerous tentacles all alike; with free, club-shaped statocysts; with eight pendent, sausage-shaped gonads, equally developed and attached to the subumbrella.

By this diagnosis *Aglantha* is distinguished from *Aglaura* in which the eight gonads are upon the peduncle, and from *Anphogoua*, in which the eight gonads are not equally developed; moreover *Aglantha krampii* Ranson, with linear gonads on the peduncle, is removed from the genus *Aglantha* (see above, p. 12).

In two papers (1932 and 1936) RANSON has dealt with the history of the Aglaurinae and discussed the various genera and species referred to this division of the Trachynemide, and in most respects I can agree with him. As far as the species which have been referred to the genus *Aglantha* are concerned, we may safely state as follows: *A. globulifera* (Haeckel) belongs to *Aglaura*; *A. ignea* Vanhöffen should be transferred to the Rhopalomeninae; *A. cautchatica* (Brandt), *A. rosea* (Forbes), *A. occidentalis* Maas, and *A. conica* Hargitt are forms (races or varieties) of *A. digitale*. *A. elata* (Haeckel) is probably identic with *A. elongata* (Lesson), and RANSON is inclined to identify this form with *A. digitale*; considering its occurrence off the African coasts, far removed from the area of distribution of *A. digitale*, it seems to me that, at least at present, we must retain it as a proper species (the specimens from the Bay of Biscay, identified by MAAS (1904 p. 30) as *Agliscia elata*, certainly belong to *A. digitale* as demonstrated by RANSON, who has re-examined the specimens). *Aglantha digitale* var. *intermedia* Bigelow (1909 a p. 122, Pl. 29 figs. 1-10), taken by the "Albatross" in the eastern tropical Pacific, differs from *A. digitale* in the shape of the manubrium and in the peculiar S-like course of the canals upon the peduncle; therefore, and also for geographical reasons, it seems to me that this form should better be regarded as a distinct species. *Melocertum proboscifer* Maas is most probably an *Aglantha*, distinguished by the considerable breadth of the umbrella in proportion to the height (see above, p. 22). As mentioned above (p. 12) *Aglantha krampii* Ranson seems to me to represent a proper genus, *Ransonia* n. g.

Only one species of *Aglantha* occurs in the northern portions of the Atlantic area, viz. *A. digitale* (O. F. Müller); different races may be more or less distinctly recognized, but on the present occasion I shall not enter into this matter because only scattered portions of the material at my disposal are in a condition which

make them suitable for this kind of investigations. I shall only give a general survey of the North-Atlantic occurrence of the species as a whole, particularly of the bathymetrical occurrence in the different regions within the area of distribution.

As, from several points of view, *Aglantha digitale* is one of the most important species of medusæ in northern seas, it might be desirable for once to give a list of the references in the literature, as complete as possible. As to the literature previous to 1910 I refer to MAYER, Medusæ of the World (1910 p. 492), with addition of LEVINSEN 1893 p. 116 containing a list of Greenland localities. — In the following list the North-Atlantic records are marked by an *n*.

Aglantha digitale (O. F. Müller).

- Aglantha digitale* Mayer 1910 p. 492 (list of previous literature).
n — *digitalis* Le Danois 1913a p. 188, and 1913b p. 27, figs. 11-17 (description of development; localities S.W. of Ireland, near Jan Mayen, and between Jan Mayen and Iceland).
n — *digitalis* Le Danois 1911 p. 311 (Bay of Biscay, Faroes, Jan Mayen).
n — *digitalis* Linko 1913 (Kara Sea).
n — *digitale* Bigelow 1913 p. 43 (N.W. Pacific; discussion of species).
n — *digitalis* Kramp 1913 a p. 269 (W. Greenland).
n — — 1913b p. 527 (N.W. Europe).
n — — 1914 p. 128 (W. and E. Greenland).
n — *digitale* Bigelow 1914 (New England).
n — *digitalis* Kramp 1915 (Great Belt and Kattegat).
n — *digitale* Bigelow 1915 (discussion of species; between Nova Scotia and Cape May).
n — *digitale* Bigelow 1917 pp. 303, 301, 305 (Cape Cod—Halifax).
n — *digitale* Bigelow 1918 p. 388 (near Chesapeake Bay).
n — *digitalis* Kramp 1920 p. 5 (N. Atlantic).
n — *digitale* Bigelow 1920 p. 10 (Alaska) and p. 17 (Labrador).
n *Aglaura hemistoma* var. "laterna" Sverdrup 1921 p. 26, Pl. 3 fig. 14 (Kristianiafjord).
n *Aglantha digitalis* Sverdrup 1921 p. 27, Pl. 4 fig. 16 (Kristianiafjord).
n — *digitale* Bigelow 1922 p. 131 (Gulf of Maine).
n — — Lebour 1922 p. 664 (food).
n — — Peacock 1923 p. 95 (Northumberland coast).
n — *digitalis* Jespersen 1923 p. 109 (N. W. Greenland).
n — *digitale* var. *rosea* Coy 1924 p. 56 (Northumberland coast).
n — *digitale* var. *rosea* Peacock 1924 p. 60 (Northumberland coast, remarks on var.).
n — *digitale* Kramp 1924 p. 29, fig. 24 (Bay of Biscay, S.W. of Ireland).
n — *digitale* Kramp & Damas 1925 p. 318 (Norway).
n — *rosea* Russell 1925 p. 786 (Plymouth).
n — *digitale* Uchida 1925 p. 97 (Japan).
n — — Bigelow 1926 pp. 38, 40, 48, 50, 352 (Gulf of Maine).
n — *digitale* Uchida 1927 p. 225 (Japan).
n — — Jaschnov 1927 p. 7 (Kara Sea).
n — *rosea* Russell 1927 p. 573 (Plymouth).
n — *digitale* Kramp 1927 (Denmark; races in Danish waters).
n — *digitalis* Uchida 1928 p. 79 (Japan).
n — *digitale* Broch 1929 p. 512 figs. 22, 23 (northern seas).
n — *rosea* Sanderson 1930 p. 229 (Northumbrian coast).
n — — Watson 1930 p. 236 (Northumbrian coast).
n — *digitalis* Uchida 1930 p. 335 (Japan).
n — *rosea* Plymouth marine Fauna 1931 p. 83 (Plymouth).

18. Sars 1897, p. 21, figs. 70 (east coast of Iceland; 1902, p. 41, Norway).
19. Sars 1902, p. 41 (Cape Spake Bay).
20. Sars 1902, p. 104 and 1902b (distribution; B. 1902, p. 76, Plymouth).
21. Sars 1903, p. 132, fig. 8 (Kamchatka).
22. Kramp 1903, p. 16 (E. Greenland).
23. Bernstein 1931, pp. 27-29 (Kara Sea).
24. Russell 1935a, pp. 311, 315, 318 (Channel).
25. Russell 1935a, p. 27 (biology).
26. Kunze 1935, p. 95 (Baltic).
27. Russell 1936, p. 177, Pl. 2, figs. 18, 20 (temperate Atlantic; discussion of species and varieties).
28. *digitalis rosea*, Kunze 1937a, p. 6 (Baltic).
29. Russell 1937b, pp. 139, 147, 151 (North Sea).
30. Frost 1937, p. 26 (Newfoundland).
31. *digitalis* Hardy 1936 (N. of Spitzbergen).
32. Kramp 1937, p. 130, fig. 59 (Denmark).
33. Uchida 1938a, p. 51 and 1938b, p. 43 (Japan).
34. Russell 1938, pp. 113, 116, 117, 119, 133, 137 (Plymouth).
35. *digitalis* Bigelow & Sears 1939, p. 362 (Cape Cod; Cape Spake Bay).
36. *digitalis* Kramp 1939, p. 16 (Iceland).
37. Jaschnov 1939, pp. 112, 111 (Kara Sea, Beringotski Sea).
38. *digitalis* var. *rosea* Russell 1940, p. 517 (metacysts).
39. Uchida 1940, p. 292 (Japan).
40. De Greef 1942, p. 74 (around Baltic Land).
41. Kramp 1942, p. 81 (W. Greenland; discussion of species; biology).
42. *digitalis* Kramp 1943, p. 7 (E. Greenland).

The 1800 (or Zoologist O. F. Müller (1776, p. 233) was the first to give a brief diagnosis of "*Medusa digitale*", for which, according to him, was introduced to O. F. FAHNEBUS. In "Fama Gronoviana" (1780, p. 396, FAHNEBUS gave the same diagnosis, and gave a somewhat more detailed description of the specimens which he had collected in Greenland. Though this species is one of the most common, restricted and common medusae in the northern hemisphere, it was not reported before it was found in other regions. *Medusa antichthona* was collected by MERLINUS in the Pacific and first described by BRANDI (1838, p. 354, Pl. Figs. 1-5), and the first record from European seas (the Shetland Islands) was by FROST (1848, p. 34, Pl. Fig. 2, *Cera rosea*). From the coast of North America it was recorded for the first time by V. A. Sars (1893, p. 57, figs. 81-86, *Trachymedusa digitale*). The name "*Trachymedusa digitale*" is introduced by HAYAKUMI; his new designation is based upon the specimens in our museum in Copenhagen, collected at several localities off the west coast of Greenland (and in the northern Atlantic), mainly from the vessels "Danmark" and "Greenland", most of these specimens were collected in the Greenland localities, and some of them were first published by LINDSEY (1893, p. 146); the specimens were first published in details, are as follows: 66° 23' N., 8° 25' W., and 68° 27' N., 8° 26' W. (MORSE, 1898, N. 16, W. 17, and 17° 58' N., 30° 59' W. (OURD, 1898, p. 75, W. Bay, 1898).

As far as is known, the medusa "*Aequorea digitale*" is common in the northern hemisphere from the Polar Sea to the Mediterranean, and to the coast of Manx in the West. It is common in the northern hemisphere, both sides of the Atlantic, and is common in the Mediterranean to give a complete list of the localities where it occurs in our collections.

In some previous papers I have dealt with the occurrence in the waters round Denmark, Norway, Iceland, and Greenland, and several other authors have given equally thorough accounts of the occurrence in other coastal areas. It may be worth while, however, to examine the distribution in the large, open oceanic basins, from which the records in the literature are somewhat scattered and from which rather considerable collections are available in our museum. Though *Aequorea digitale* is a holoplanktonic medusa, we cannot beforehand be sure that it is equally abundant everywhere within its extensive area of distribution.

For the following discussion of the occurrence I have divided the area under consideration into six sections, as seen from the map, textfig. II, and I am going to deal with them separately, leaving out the records from the coastal regions.

I. The Kara Sea: Material: Kara Sea, "Dijmphna" 1882-83. Previous records: LINKO 1913, JASCHNOV 1927, BERNSTEIN 1931, JASCHNOV 1939.

Unfortunately, we have no information of the exact localities at which the specimens were taken by the "Dijmphna" expedition, and no records of depth or date. The expedition comprised the southern portion of the Kara Sea, south of 72° N., and west of 96° E. LINKO was the first to mention *Aequorea digitale* from the Kara Sea, but his paper has not been accessible to me. JASCHNOV (1927, p. 7) has dealt with its occurrence at some length; according to this author the medusa was never taken east of 66° E., where the salinity of the water is considerably lowered by the influence of the great Siberian rivers; adult specimens were mainly taken in the deeper strata at very low temperatures, whereas very young specimens were found almost exclusively in the surface layers. The largest specimens measured by him were 15 mm in height, thus considerably smaller than those found in other arctic regions. According to BERNSTEIN this species may occur in the Kara Sea in great abundance.

II. The Barents Sea: No new material. Records in the literature: LINKO 1901a, p. 15 (preliminary list of species) and 1901b, p. 249; KRAMP 1913b. *Aequorea digitale* may be extremely abundant in this region, but the quantity seems to be variable from one year to another. LINKO has given valuable information of the seasonal occurrence, and he concludes that the propagation predominantly takes place during the winter in the neighbourhood of the coasts.

III. The Norwegian Sea and the Polar Sea east of Greenland:

66° 23' N., 8° 52' W.,	¹⁰ / ₅ 1896,	Ingolf St., 103, 188-0 m, 2 spec.
66° 23' N., 7° 25' W.,	¹¹ / ₅ 1896,	101, 188-0 m, 3
68° 27' N., 8° 26' W.,	²¹ / ₅ 1896,	118, 1 specimen
62° 58' N., 7° 09' W.,	¹¹ / ₆ 1896,	113, 1
abt. 62 ¹ / ₂ ° N., 1° E.,	²⁰ / ₆ 1900,	E. Greenland, Exp. vert. hauls, 3 + 2 spec.
abt. 63° N., 3 ¹ / ₂ ° E.,	²⁰ / ₆ 1900,	3 + 2 + 1
abt. 63 ¹ / ₂ ° N., 0°	²¹ / ₆ 1900,	1 spec.
abt. 61° N., 1 ¹ / ₂ ° W.,	²¹ / ₆ 1900,	7
61° 05' N., 9° 38' W.,	⁵ / ₅ 1901,	Thor. St., 63, 3 specimens
63° 36' N., 6° 20' W.,	¹¹ / ₅ 1903,	12, 600m wire, 22 spec.
61° 04' N., 4° 33' W.,	²¹ / ₆ 1905,	121, 3 specimens
66° 49' N., 24° 59' W.,	⁷ / ₅ 1927,	Dana, St. 3221, 800m wire, 200 sp.
66° 22' N., 6° 26' W.,	²³ / ₅ 1931,	5112, 600m
65° 14' N., 6° 06' W.,	²³ / ₅ 1931,	5112, 900m
		5113, 1100m
		5113, 1900m
		5113, 2400m

Previous records, LE DAVOIS 1913a and b, 1911 records this species from the neighbourhood of Jan Mayen; KRAMP 1913b, in which the collections by the International Plankton

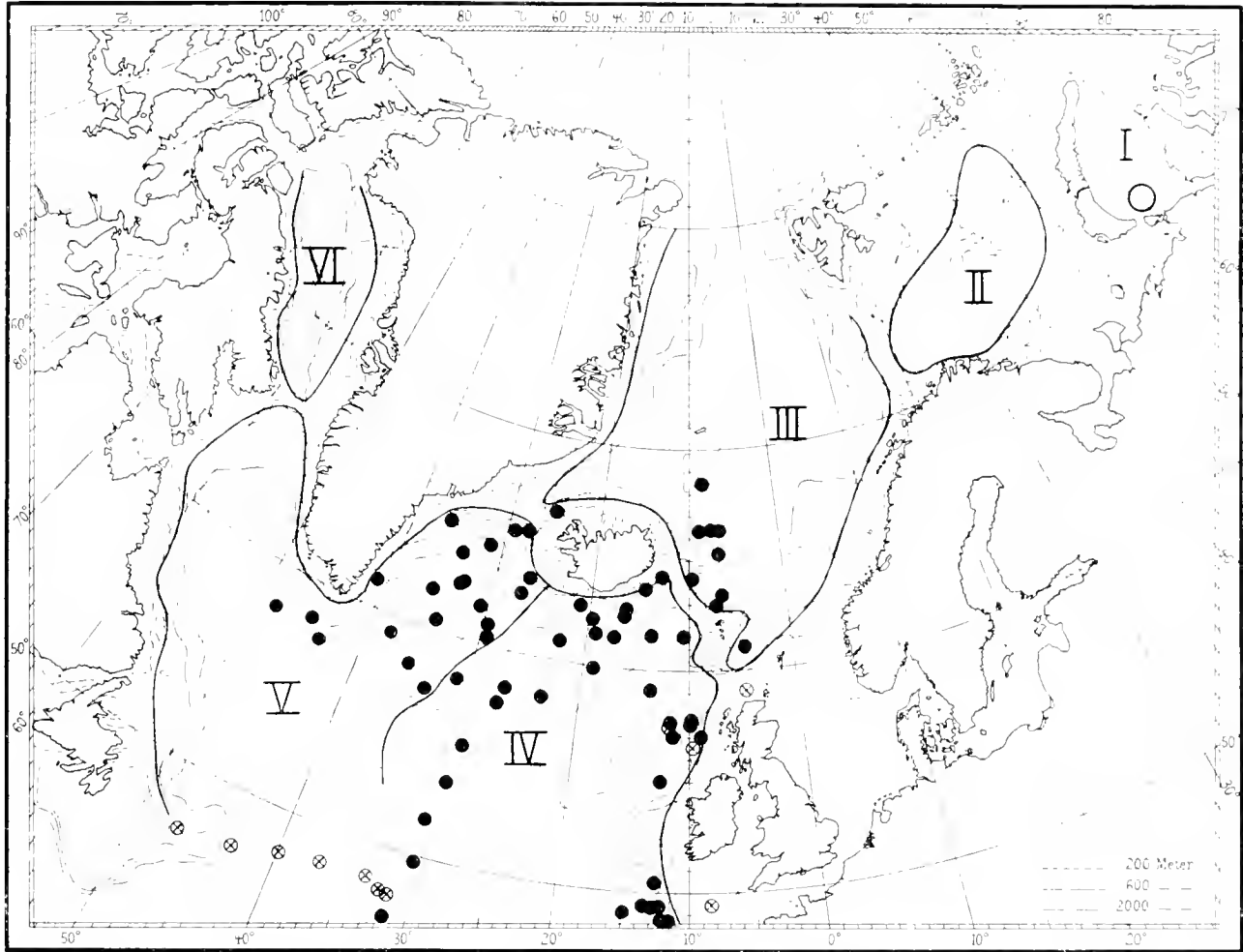


Fig. 11. *Aglantha digitale*. ● new records outside the coastal waters; ○, records yet not published; ⊗ exact localities not stated. The strongly drawn lines indicate the boundaries of the regions mentioned in the text.

Investigations 1902–1908 are summarized and illustrated on a map: *Aglantha digitale* was taken in several localities between Norway and Iceland, usually in small numbers, but in May 1903 it was taken in very great abundance in three localities about 67° N.

The few specimens collected by the “Ingolf” Expedition were taken in hauls through the upper strata with fairly small nets; the specimens from St. 104 were small, 2 mm, the others varied in height between 14 and 18 mm. The specimens taken by the East-Greenland Expedition 1900 in vertical hauls with small nets were all very small, 2–4 mm. The specimens taken by the “Thor” at two stations east of the Wyville Thomson Ridge in May 1903 and 1904 were of considerable size, 19–24 mm high. “Thor” St. 124, 1905, was in the Faroe-Shetland Channel; the three specimens taken there were 10, 17, and 21 mm high. The most interesting material available from the Norwegian Sea is that collected by the “Dana” in May 1931, St. 5112 and 5113, above deep water east of Iceland; the hauls were made with the 2 m ringtrawl with bag of stramin, and at St. 5113 the duration of the hauls was two hours; as seen from the list above great numbers of *Aglantha* were taken at all depths between about 600 and 1600 metres below the surface, especially in the haul with 1900 m wire out (about 1300 m below the surface); there were no remarkable differences in the size of the specimens at these different depths, almost all the specimens were large, up to 20 mm in height, with a slight admixture of small ones. The haul with 600 m wire out at the neighbouring St. 5112 caught about 800 specimens during one hour; the size limits were about the same as in the deeper hauls at St. 5113, 5–19 mm, but very few of them were adult,

the vast majority being young ones, about 6 mm in height. There can be no doubt, therefore, that the numerous large specimens brought up by the deep hauls were really caught in the deep strata, where the temperature of the water was below 0°, whereas the vast majority of the young ones occurred at higher levels with positive temperatures.

IV. North-East Atlantic, east of the Mid-Atlantic ridge running southward from south-western Iceland (the Reykjanes Ridge).

Material.

57° 13' N, 27° 03' W.,	17	1868, BANG.	
58° 17' N, 30° 59' W.,	22	1859, OLERIK	
58° 27' N, 26° 43' W.,		1857, MOBERG	
59° 07' N, 13° 32' W.,			
59° 00' N, 19° 02' W.,		1883, RYDER	
58° 17' N, 23° 19' W.,	22	1898, OSTENFELD	
55° 25' N, 29° 05' W.,	18	1890, LUNDBECK	
61° 41' N, 10° 17' W.,	11	1895, “Ingolf” St. 42,	188–0 m, 18 spec.
61° 40' N, 13° 33' W.,	2	1903, “Thor” St. 87,	5 specimen
62° 10' 5" N, 19° 36' W.,	12	1903	161, 20 specimens
62° 41' N, 20° 41' W.,	9	1901	179, 50 m wire, 1 spec.
61° 31' N, 19° 05' W.,	10	1904	180, 100 m — 10 —
61° 30' N, 17° 08' W.,	11	1904	183, 12 specimens
57° 17' N, 11° 33' W.,	7	1905	71, 3
57° 52' N, 9° 33' W.,	8	1905	72, 600 m wire, 8 spec.
57° 46' N, 9° 55' W.,	31	1905	167, 22 specimens
48° 43' N, 12° 05' W.,	21	1906	52, 300 m wire, 78 spec.
50° 25' N, 12° 44' W.,	5	1906	62, 1500 m — 16

61° 52' N., 35° 30' W.,	¹⁵ 1933, "Dana" St. 1686,	600 m wire	none sp.
62° 36' N., 32° 48' W.,	¹⁶ 1933, — St. 1687,	600 m	300
—	— 1687,	1000 m	500
—	— 1687,	2000 m	500

Previous records: The occurrence of *Aglantha digitale* in the coastal waters of the north-western Atlantic has been dealt with by several authors; summaries are given by BIGELOW 1926 p. 352 (east coast of North America, with special reference to regional and seasonal occurrence in the Gulf of Maine); KRAMP 1911, 1912, and 1913 (the coasts of Greenland); KRAMP 1939 (Iceland). Special records from Newfoundland and Labrador are given by BIGELOW 1909b p. 312 and 1920 p. 17 and by FROST 1937 p. 26. From the off-shore areas of this section very few records are found in the literature. The Plankton-Expedition in 1889 did not take the typical form west of 30° W., but MAAS (1893 p. 24) described a new species or variety, *occidentalis*, from a series of localities across the Newfoundland Bank. The "Tjalff" expedition, on its voyage to the west coast of Greenland in 1908, collected numerous specimens of *Aglantha* in a series of localities about 58° N. and 31° 53' to 39° 24' W. (KRAMP 1913a p. 239). The occurrence in the waters between the southern part of the west coast of Greenland and the eastern coasts of Labrador and Baffin Land is dealt with by KRAMP 1913, 1914, and 1912.

Besides the material listed above and the extensive collections from the waters between Greenland and Labrador (KRAMP 1912) I have seen numerous specimens collected by the "Michael Sars" expedition in 1910 in a series of localities east of the Newfoundland Bank, and we may state that the species is generally distributed and very abundant in this part of the Atlantic. In the Davis Strait it is particularly numerous above the coastal banks on both sides, off the Greenland coast as well as off the coast of Labrador, but less abundant in the middle parts of this region; in the area east of the southern part of Greenland, however, it may also be taken in great quantities above the greatest depths. At most of the "Dana" stations the greatest number of specimens were taken in hauls with 600 m wire out, but considerable numbers were also sometimes taken in deeper strata. The appliance used by the "Dana" was almost always the 2 m ringtrawl with bag of stramin, and the numbers caught in different depths may therefore be directly compared. Only at St. 2136, 2138, and 4235 the deepest haul was made with a coarser net with more open meshes, which may account for the small number of specimens taken in these hauls. The deepest haul, with 1000 m wire out, at St. 2308 was, however, made with the 2 m ringtrawl and only brought up 9 specimens of *Aglantha*, which may even have been caught at higher levels during the hauling in of the net; undoubtedly, therefore, the species is really rare in the very deep strata. At St. 1687, in the middle of the deep basin east of southern Greenland, the hauls with 1000 and 2000 m wire contained more specimens than the haul with only 600 m wire out.

At all depths and at any time between May and August, from which material is available, small as well as large specimens were taken. The largest specimens, 21-22 mm in height, were found as well in the neighbourhood of the east coast of Greenland as above the Reykjanes Ridge and in the deepest parts of the basin. In the Davis Strait the specimens frequently attain a still larger size, especially in the cold marginal zones off the coasts of Greenland and Labrador, where they grow to a size of about 28 mm in height.

VI. The Baffin Bay. — No new material. — A general account of the occurrence of *Aglantha* in this deep and cold basin is given in my paper on the medusa of the "Godthaab" expedition (KRAMP 1912). In certain parts of this cold area the specimens attain a very considerable size, 31 mm or more in height.

The statements above confirm the previous conception of *Aglantha digitale* as a very common medusa throughout the northern part of the Atlantic and adjacent waters, frequently occurring in great numbers. It is rarely taken in the surface water, and as a rule it is likewise rare in the very deep strata, though sometimes it may be met with in considerable numbers as deep down as 3000 metres or more below the surface. We know from the literature that in the southern part of its area of distribution it only occurs in the intermediate and deep strata.

Aglantha digitale has a circumpolar distribution; in the Pacific it penetrates southwards as far as Misaki in Japan (about 31° N.) and Vancouver on the west coast of North America (about 50° N.). In the western Atlantic it is common in the Gulf of Maine and occasionally found as far south as Chesapeake Bay (about 37° N.). In the eastern Atlantic it is frequently mentioned from the Bay of Biscay (BROWNE 1906, LE DANOIS 1911, KRAMP 1921, RANSON 1936); according to BROWNE it is even the most common of all medusæ in the Bay of Biscay. It is also recorded from a locality west of Cape Finisterre (13° N., RANSON 1936). Re-examination of the specimens taken by the "Thor" in 1909 and 1910 in two localities off the Straits of Gibraltar and formerly (1921) referred by me to *Aglantha digitale*, has proved that they belong to the species which later on (1932) was described by RANSON as *Aglantha krampi*. According to RANSON *Aglantha digitale* is found east and south of the Azores (southernmost locality 36° 17' N., 28° 53' W.) and between the Azores and the Bermudas. In the collections of the "Dana" expedition in 1921-1922 (which I have worked up but not yet published) *Aglantha* was completely lacking, also from the northernmost stations in the Atlantic about 35°-36° N. The southern limit of distribution thus seems to be at about 36° N. across the Atlantic Ocean.

Pl. VI fig. 6 presents a typical specimen of the red form of *Aglantha* as found in the cold areas in the waters west of Greenland. The figure was drawn by me on board the "Godthaab" in 1928.

Fam. Geryonidæ Eschscholtz 1829.

Trachymedusa with 4 or 6 radial canals, with blindly ending centripetal canals; with well-developed manubrium mounted upon a peduncle; with leaf-like gonads upon the radial canals.

Gen. *Liriope* Lesson 1843

1-rayed Geryonidæ.

Every author of recent time, who has dealt with this genus, emphasizes the difficulty, or impossibility, of finding reliable characters for distinction and limitation of the species. RANSON (1936 p. 188) still retains at least three Atlantic species, *virgata*, *curybia*, and *tetraphylla*. THIEL (1936 p. 15) unites all the numerous species into one, *L. tetraphylla* (Chamisso & Eysenhardt), and BIGELOW (1938 p. 126) is inclined to follow him in this respect. They may be right, but I prefer to leave the question open until I have examined the extensive collections at my disposal.

None of the various forms of *Liriope* are constant inhabitants of the areas here dealt with. Shoals of them are sometimes carried into the western part of the British Channel, where they are usually identified as *L. virgata*, and some few specimens were taken at two of the stations of the "Dana". They agree perfectly with the descriptions of *L. curybia* as given by HAECKEL and subsequent authors.

Larropia eurybia H.

H.	1864	20-302	P. 12	11-25
	1879	201		
S.	1883	P. 11	12-25	
M.	1910	120		

M.

1°N	3°W	100-1000	1201-1000	2 spec
			1201-1000	1
1°N	2°W	100	1203-500	2
			1203-3000	1

These localities are near the northern boundary of the Gulf Stream, where the temperature of the water in the upper strata was about 13-14° C. In the colder water a little further north the species was lacking.

Geographical distribution. *Larropia eurybia* is abundant in the Mediterranean and is also common in the warm parts of the Atlantic; it is recorded by MAYS (1893) from the Northern Equatorial Current and the Florida Current, and by RANSON (1936) from the Bay of Biscay.

II. Narcomedusæ.

Fam. Solmaridæ.

Genus *Pegantha* Hæckel.

Pegantha clara R. P. Bigelow.

Pegantha clara R. P. Bigelow 1909 p. 80, 2 figs.
 — Mayer 1910 p. 445, fig. 298A.
 H. B. Bigelow 1918 p. 397.
 1938 p. 131.

Material:

17° 02' N, 31° 15' W., 27-28 g. 31.	"Dana" St. 1201.	50 m wire.	1 spec.
—	1201.	100 m	1
—	1201.	300 m	1
—	1201.	5000 m	1
49° 49' N, 30° 22' W., 30 g. 1931	1203.	100 m	2
	1203.	300 m	2
	1203.	600 m	1

The original description of this species was based upon one large, well-preserved specimen, taken near the borders of the Gulf Stream, off Woods Hole; MAYER (1910) examined the same specimen and gave new figures of it. H. B. BIGELOW (1918 and 1938) has seen several small specimens, most probably belonging to the same species, taken in the surroundings of the Bermudas and the Bahama Islands. The type specimen was 53 mm in diameter with 14 long tentacles alternating with the same number of smaller size. The specimens examined by H. B. BIGELOW varied in diameter between about 10 and 20 mm, with about 20 to 24 tentacles; BIGELOW does not state, whether the tentacles are of different sizes.

I have no doubt that the present specimens, from the northern boundary area of the Gulf Stream, belong to the same species; they are in rather poor condition, but in some places the otocorpa and the stomach sacculi may be traced. They vary in size between 7 and 17 mm; the tentacles, 14-21 in number, are of different sizes, but large and small ones are usually not regularly alternating, as will appear from the adjacent diagrams (textfig. 12).

Geographical distribution: H. B. BIGELOW (1938) is inclined to think that the medusa identified by VANUÖFFEN (1912a and b) as *Polycalpa forskalei* Hæckel, and also *Pegantha smaragdina* Bigelow, belong to *Pegantha clara*; in such case this species has a very wide distribution in the oceans, being found in the tropical Pacific between the Hawaiian and Caroline Islands and off Chile and Peru, in the tropical Atlantic off the Cape Verde Islands, and in the localities mentioned above in the north-western Atlantic. *P. smaragdina* (found off Peru) is a large medusa, 50-75 mm wide, with 28-31 tentacles; these are of almost equal size, in contradistinction to the alternately large and small tentacles of the large original specimen of *P. clara* as described by R. P. BIGELOW. Most records are from the upper strata, but, as stated by BIGELOW, the species has also been taken in "open net-hauls from considerable depths." The specimen from "Dana"

St. 1201, 5000 m wire, may have been taken in the upper strata during the hauling in of the net.

The distribution of *Pegantha clara* in the North Atlantic seems to be extended still farther north than hitherto supposed, if the parasitic larvae, which shall now be described, really belong to that species.

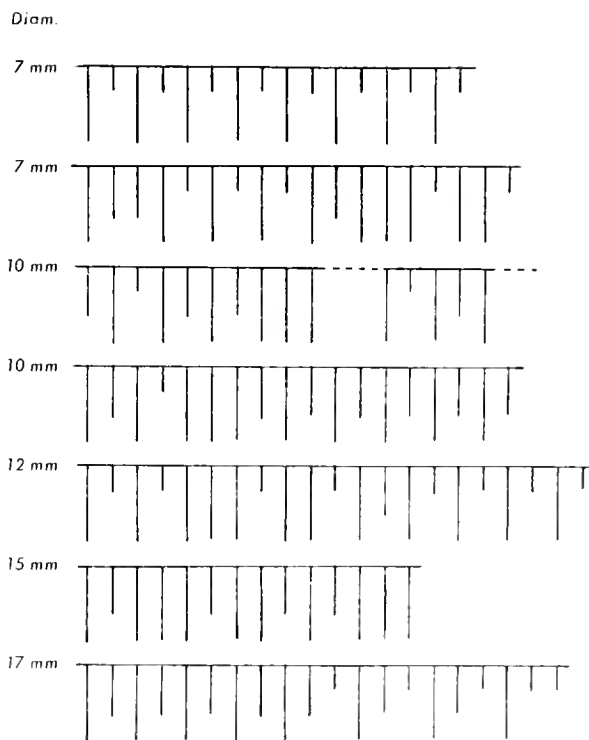


Fig. 12. *Pegantha clara*. Diagram showing relative length of tentacle in seven specimens from "Dana" St. 1201 and 1203.

Larvæ of *Pegantha clara* in Periphylla.

Pl. IV, fig. 7, Pl. V, figs. 1-10.

Several Narcomedusa larvæ in different stages of development were found in the stomach cavity of a young *Periphylla periphylla* from "Godthaab" St. 4, south west of Iceland, 63° 49' N, 26° 50' W., 21 g., 1928, 1000 m wire haul. The larvæ were lying free in the gastric cavity of the host, most of them in the stomach proper, but a few young stages were seen in the distal part of one of the marginal lappets. Pl. IV, fig. 7 presents the *Periphylla* with the larvæ *in situ*. Two of them are medusoids ready for liberation.

the actinulae propagate by budding from the oral region of the parent medusa (as in *Narcomedusa* and *Actinostole* Metchnikoff), and in other cases (as in *Cannothutha* Muller) the young medusae (in the present species the budding takes place in the oral region of other species of *Cannothutha*, *Reptanthus*, *Uthusa*, *Canna parvula*) are egg-shaped larva probably of *Canna* type, attached to the mouth region of the parent medusa and propagate a large number of medusae. A special case is described by D. DAVIES (1910) in the case of *Narcomedusa* larva, presumably *N. tenuis* (Gegenbaur), were found as parasites in the parthenia of the polychaete worm *Taeniocera* which propagate by budding and increase in number, causing a complete castration of the host. The young medusae are also known as *Cannothutha* Muller, *Canna parvula* Bigelow, and *Paputha* Bigelow, but their final development is unknown. No other medusa living free in the gastric cavity of another medusa has been described hitherto for the first time.

The youngest stages observed are two complexes of larva, each consisting of three or four individuals (Pl. V figs. 1 and 2). One of the two complexes differs considerably from the other in that it consists of a minute body with two very large tentacles provided with a distal knob of nematocysts. I would consider this as a primary polyp (no. I in the following) and the others are developed by budding. One of the two medusae (no. I) is an actinula with a large egg-shaped body and two tentacles placed opposite each other, one of them considerably larger than the other and both provided with a distal knob of nematocysts. The next bud (III) is still very small, and the tentacles just visible as a small protuberance with a knob of nematocysts. In fig. 2 a still younger knob may be distinguished to the left of no. III. When a bud has attained a certain size it detaches itself from the primary polyp and becomes a free actinula with two tentacles. Pl. V fig. 3 shows a chain of three actinulae, no. I is an actinula with its two tentacles of unequal size, no. II is at the other end of the chain and smaller than no. I and its tentacles are both quite small. Between nos. I and II are two swellings evidently representing the young medusae in their first stages of development (III and IV). In this chain there is no individual like the primary polyp with the two large tentacles as seen in figs. 1 and 2. It is probable therefore that an actinula after its liberation from the parent medusa is sometimes able to produce other actinulae (fig. 3).

The youngest actinula seen in Pl. V figs. 1 and 2 is represented in fig. 4. Two more tentacles have been developed, but they are still much smaller, and are placed between the two larger ones. It will also be seen that the body of the actinula is considerably smaller in size after its liberation from the parent medusa.

The transition of the young actinula into a medusa is accompanied by the formation of a circular collar of tentacles which extends outside the tentacles, and the difference in size between the two larger tentacles here becomes almost alike. It is probable therefore that the tentacles are provided with a basal cushion-like pad, and that the radiating grooves are already provided with a basal cushion-like pad, and from

each statocyst an otopore is seen running some distance towards the central part of the aboral surface, the middle one in each octant being the longest and the others decreasing in length towards both sides. The peronia and otoporopae are also seen in fig. 7, which shows a section of the aboral surface of a larva similar to those figured in figs. 5 and 6. The oral portion of the larva, below the collar, is still egg-shaped, and there is no indication of a mouth opening.

I have seen no further developmental stages between the actinula just described and the two medusae shown in Pl. V figs. 8-10. The medusae are about 2 mm in diameter. There are still only four tentacles, with a solid core of cylindrical endoderm cells, and with the nematocysts still clustered in the tip of the tentacle. The umbrella is watchglass-shaped, its mesogloea moderately thick. The umbrella margin is turned inwards, and it is slightly notched outside the four peronia. The statocysts are well developed, club-shaped with a basal cushion-like pad; their number is unaltered, five in each quadrant. The most remarkable feature is the extraordinary length of the otoporopae, the four interradial otoporopae are nearly concurrent at the apex of the exumbrella, and some of the others are only slightly shorter, their length however decreasing towards the tentacular radii. The velum is well developed.

The egg-shaped body of the actinula has now been transformed into the manubrium of the medusa, it has decreased in size, but it is still hanging down like the tongue of a bell; in one of the two specimens the manubrium is even somewhat constricted at its base. In this developmental stage of the medusa the first trace of a mouth is observed as a minute depression in the distal end of the manubrium (Pl. V fig. 10), not yet quite perforating the tissue to connect the stomach cavity with the outside. Until now the nutrition of the larva must evidently have been accomplished by osmosis. Probably the young medusa will leave the host as soon as the opening of the mouth is fulfilled, enabling the medusa to catch and swallow its own food.

The specific affinity of the larva here described seem to me beyond doubt. Very few species of *Narcomedusa* are known from the northern Atlantic, and the possession of well-developed otoporopae in the young medusa at once separates it from *Salmaris corona*, *Salmassus caesa*, *Aequi citrea*, *Aeginaria grimaldi*, and *Aeginopsis laeucata*. The only remaining species is *Paputha clara*, which is characterized by the possession of 2-5, usually 3, long, slender, linear otoporopae on each of the marginal lappets. The fully developed medusa may have as many as 28 tentacles and marginal lappets, but as mentioned above the number is increasing with age; in a specimen 7 mm in diameter I counted 8 large tentacles and as many very small ones, and there can hardly be any objection against the supposition that the young medusa begins its free-swimming life with only four tentacles as in the specimens found in the stomach of *Periphalia*. It seems to me highly probable, therefore, that the parasitic larvae described above belong to *Paputha clara* R. P. Bigelow, a species which seems to be widely distributed in the northern Atlantic.

In the closely related species *Paputha smaragdina* H. B. Bigelow, which occurs in the tropical Pacific, the development of the larva takes place in the stomach cavity of the parent medusa. MAVER (1910) and also Bigelow himself (1938) are inclined to think that the two species are identical. This supposition seems, however, to be contradicted by the development of the larvae. In both species the larvae, while in the actinula stage, propagate by aboral budding, and the proceeding of the budding process is very similar in both species. But during the further development the larva of *P. smaragdina* differs from that of *P. clara* in several respects. It has a greater number of tentacles, 6-8 in the older actinula stages, and 10-12 in the young medusa before its liberation. The actinula soon attains a broadly flattened shape, very different from the elongate shape of the actinula of *P. clara*. A mouth opening is pierced in very young actinula stages of

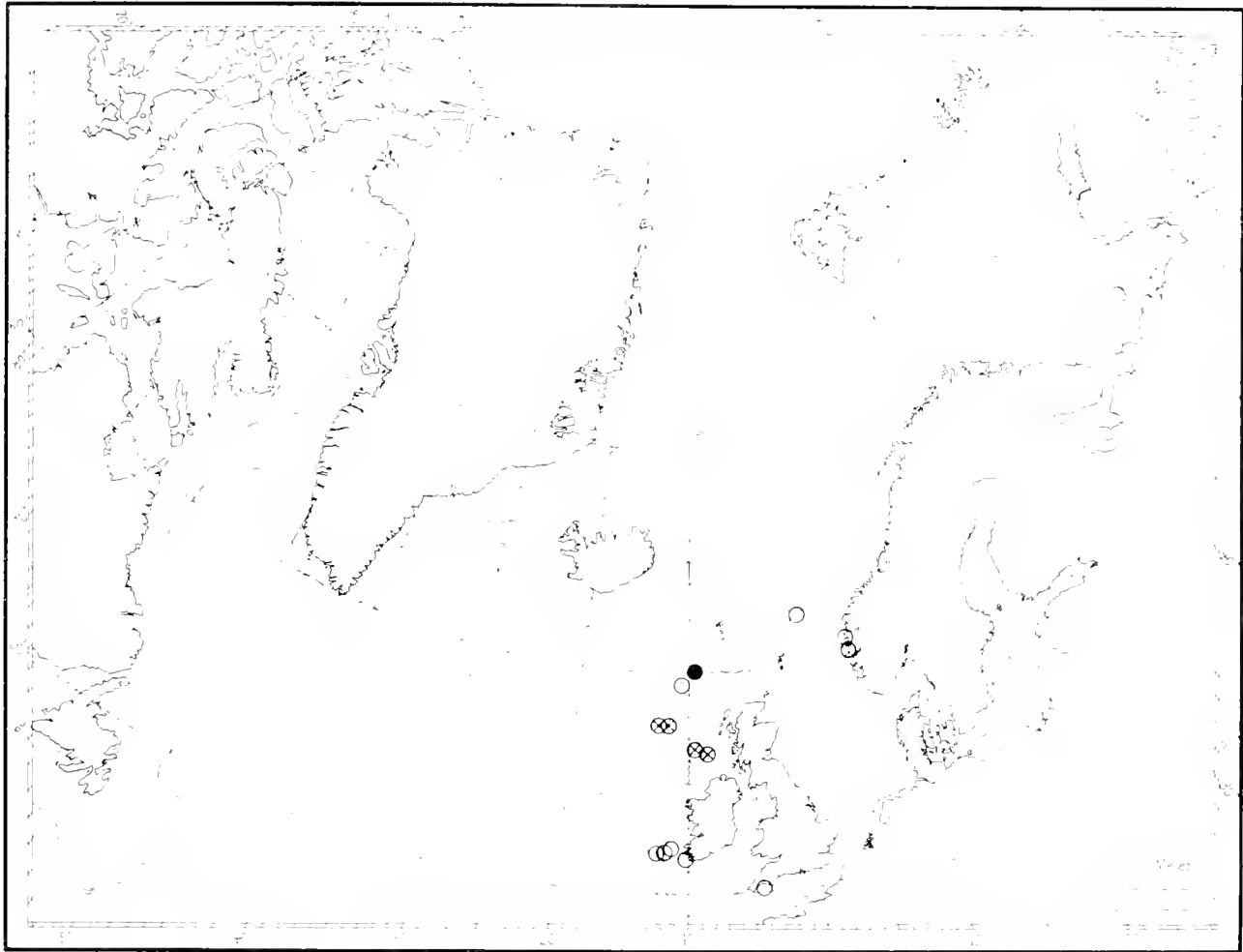


Fig. 13. Distribution in the northern Atlantic of: ● *Solmaris corona*, new record; ○ *Solmaris corona*, previous records; ⊗ *Solmaris corona*, records not yet published; ⊙ *Solmaris multilobata*.

P. smaragdina, whereas in *P. clara* the mouth does not appear until immediately before the liberation of the medusa. The statocysts of *P. smaragdina* make their first appearance in much later developmental stages of the larva, and even in the latest state of the medusa before its liberation there are only one or two statocysts between each successive pair of tentacles. Moreover otoporpoæ are completely lacking in the larvæ of *P. smaragdina*.

Metagenesis. — A very important difference between the larvæ of the two species is the absence of what I have called "primary polyp" in *P. smaragdina*. BIGELOW expressly states (p. 96) that larvæ produced by budding are indistinguishable from those developed directly from eggs. The larvæ being all alike, they probably all develop into medusæ, and consequently the developmental circle of this species is a hypogenesis and not a metagenesis. In *P. clara*, on the other hand, a metagenesis most probably takes place. It is very improbable that the larva described above with its two very large tentacles, much larger than the tentacles in even the most advanced actinula stages, might be transformed into a typical actinula; this would only be possible by a reduction in size of its tentacles, which hardly seems to be possible. This polyp should evidently be regarded as an oozoid, directly derived from an egg; it propagates asexually by budding, producing a number of larvæ which are transformed into medusæ, but it never becomes a medusa itself; it represents an asexual generation, and its offspring, the medusæ, constitute the sexual generation. Thus the cycle of development in *Pequathia clara* is a typical metagenesis, secondarily arisen in connection with the parasitic habit of the larva.

How the *Periphylla* becomes infected with the parasitic *P.*

gautha-larvæ, can only be guessed; no stages younger than the primary larvæ with two large tentacles were observed. Probably these primary larvæ have entered the stomach cavity of the host while in a ciliated planula stage.

The larvæ of *Cymina laticentris*, parasitic in *Tomopteris* (DAMAS 1936) apparently enter the body cavity of the host in very early stages, as DAMAS found morula stages in the parapodia of the polychæte. They increase enormously in number by asexual propagation, first by polyembryonic fission of the morula, in later stages by aboral budding, which is continued after the larvæ have attained the medusoid shape. The budding takes place in almost the same manner as in the species of *Pequathia* and, as in *P. smaragdina* but unlike *P. clara*, the larvæ derived directly from the eggs are similar to those developed by budding, so that probably no metagenesis takes place.

It is possible that a metagenetic development will be discovered in some of the species of Narcomedusæ, in which the cycle of development is not yet fully elucidated; until now, however, the classical example of *Canna proboscidea rubiginosa* was the only undoubted case of metagenesis known among the Narcomedusæ. In *Pequathia clara* the parasitic habit has brought about a metagenesis of quite another type.

Genus *Solmaris* Haeckel

Solmaris corona (Kieferstein & Ehlers)

Egmeta corona Kieferstein & Ehlers 1861, p. 94, Pl. 11, figs. 7-9.
Solmaris corona Haeckel 1879, p. 358.

coronantha Haeckel 1879, p. 359, Pl. 20, figs. 7-10.

1889, p. 122, Pl. 1, fig. 10.
 1890, p. 122, Pl. 1, fig. 10.

Agina citrea (Eschscholtz)

1829, p. 113, Pl. 11, fig. 4.
 1829, p. 115, Pl. 10, fig. 3.
 1859, p. 338, Pl. 20, figs. 14-15.
 1931, p. 37, Pl. 4, figs. 7-13.

1896, p. 13, Pl. 4, figs. 7-13.

1896

N. 64° 42' W., 1900 East Greenland Exped. 1 spec.

The specimens collected S.W. of the Faroe Bank, moreover from the Rockall Channel, specimens collected by the "Michael Sars" North Atlantic expedition between Scotland and Rockall in August 1910.

The specimens from the East Greenland expedition are 9 mm in diameter, with a crown of 36 tentacles and marginal lappets.

The Mediterranean specimens described by KEFERSTEIN & FRIEDLÄNDER (1861) were 11-20 mm diameter with 27-30 tentacles and 12-15 lappets; the specimens from the Canary Islands, described by HAECKEL as *S. canariensis*, were 10-15 mm wide with 21-32 tentacles. Specimens from north-western Europe generally have a somewhat larger number of tentacles and lappets, but it is not clear, as pointed out by BROWNE (1906), that they belong to the same species. BROWNE (1906), who examined a number of specimens from the south coast of Ireland, found that very small specimens, 2-4 mm wide, had only 12-16 tentacles, but 29 were found in 6 specimens 10 mm wide, and in specimens 12 mm in diameter the number of tentacles varied between 25 and 35; the greatest number recorded by him was 36, in a specimen 10 mm wide. In 1906, 10 specimens, 10-15 mm wide, from the Norwegian coast (BROWNE 1906; KRAMER & DAVES 1925) 31-42 tentacles have been counted, and specimens from the "Michael Sars" the number of tentacles varied between 33 and 44, regardless of the size of the individuals, according to FRIEDLÄNDER. I have no doubt, therefore, that the specimens from the Hebrides, examined by VAXHOFFEN (1908), belong to the same species; they were 5-10 mm in diameter with 34-45 tentacles.

VAXHOFFEN related his specimens to *S. multilobata* MAYER (1866), a matter of fact it seems probable that this species is identical with *S. canaria* or *S. multilobata*, as described by MAYER (1866), and with a larger number of tentacles and lappets than *S. citrea*, more than 64 in specimens 12-18 mm in diameter; the specimens from west of the Hebrides, in almost the same locality, examined by VAXHOFFEN. Later on the species has frequently been taken in great abundance in the waters off the south of the British Isles and on the Norwegian coast, and *S. multilobata* has never been observed since it was first described.

More than 200 specimens of this species were taken on one haul with the plankton net in the Rockall Channel, and the number of tentacles and lappets makes the only record of the species in two hauls; and it is certainly very astonishing that *S. multilobata* was abundant in 1889 in the waters off the coast of the British Isles, at a period when several investigations during several years had failed to record the other form, *canaria*. The specimens were taken in 1889 in that particular summer, and it is probable that the entire population of *Sabulites* in the area had just completed its annual development, and that the normal number of tentacles and lappets had not yet attained the size of the specimens taken in 1889.

The species is recorded from 20° N., and very common in the waters off the coast of the British Isles; it certainly belongs to the northern boundary area of the Gulf Stream (August to November) in

is more or less regularly carried by the Gulf Stream to the southern part of the west coast of Norway, and also into the Channel, at least as far as Plymouth.

Further distribution: Mediterranean, the Canary Islands.

Fam. Æginidæ.

Ægina citrea Eschscholtz.

Ægina citrea Eschscholtz 1829, p. 113, Pl. 11, fig. 4.
rosæ Eschscholtz 1829, p. 115, Pl. 10, fig. 3.
rhodina Haeckel 1859, p. 338, Pl. 20, figs. 14-15.

North-Atlantic record.

Ægina rhodina Kramp 1924, p. 37.

Material (see the map, textfig. 14)

57° 52' N., 9° 53' W., 8 ₆ 1905	"Thor" St. 72	600 m wire	1 spec.
49° 22' N., 12° 52' W., 4 ₆ 1906	181	1800 m	1
47° 02' N., 31° 45' W., 27 ₂₈ 1931	"Dana" St. 4201	50 m wire	1 spec.
		4000 m	3
62° 23' N., 16° 05' W., 25 ₆ 1932	1102	2000 m	1
63° 38' N., 14° 43' W., 18 ₅ 1934	5113	1800 m	3
		5113, 2000 m	3

The specimens are fairly small, 5-15 mm in diameter; they all have four tentacles, and, as far as can be seen from their state of preservation, the eight stomachal lobes have an entire margin with no indication of a secondary notch; the specimens belong, accordingly, to the *rosæ* type.

Geographical distribution: The various forms of *Ægina*, all of which probably belong to one species, *Æ. citrea*, are widely distributed in the tropical and subtropical parts of all the oceans, including the Mediterranean. In the Pacific it penetrates northwards into the Bering Sea. In the Atlantic it is quite common east of Florida and near the Bahamas and the Bermuda Islands; in the eastern part of the Atlantic it is recorded from the Azores and off the coast of Portugal (RAXSOX 1936). As demonstrated by the present collections the area of distribution in the Atlantic extends considerably farther north; on a previous occasion (KRAMER 1924) I have recorded the species from a locality S.W. of Ireland ("Thor" St. 181, 1906). It was also taken by the "Thor" (St. 72, 1905) west of Scotland and by the "Michael Sars" (St. 101, 1910) in the Rockall Channel. The "Dana" has taken it in the northern boundary area of the Gulf Stream about midway between Newfoundland and the British Isles (St. 1201) and also in two localities off the south coast of Ireland, as far north as 62° 23' and 63° 38' N. (St. 1102 and 5113).

The vertical range is very considerable. THOMAS (1936, p. 77) designates this species as a typical deep-sea medusa, but as a matter of fact, it occurs at all depths, being frequently taken in the surface waters. On the "Dana" St. 4201 one specimen was taken near the surface (with 50 m wire out), others in a deep haul, 1000 m wire. The other specimens in the present collections were all taken at rather considerable depths, most of them in hauls with 1800-2000 m wire out.

We may state, accordingly, that *Ægina citrea* is a widely distributed oceanic medusa, its northwards distribution in the Atlantic being limited by the submarine ridges separating the Atlantic proper from the adjacent arctic basins, occurring at all depths in the warmer regions, but restricted to the deep strata in the northern boundary areas.

Æginopsis laurentii Brandt.

North-Atlantic records.

Æginopsis laurentii Wagner 1885, p. 70.

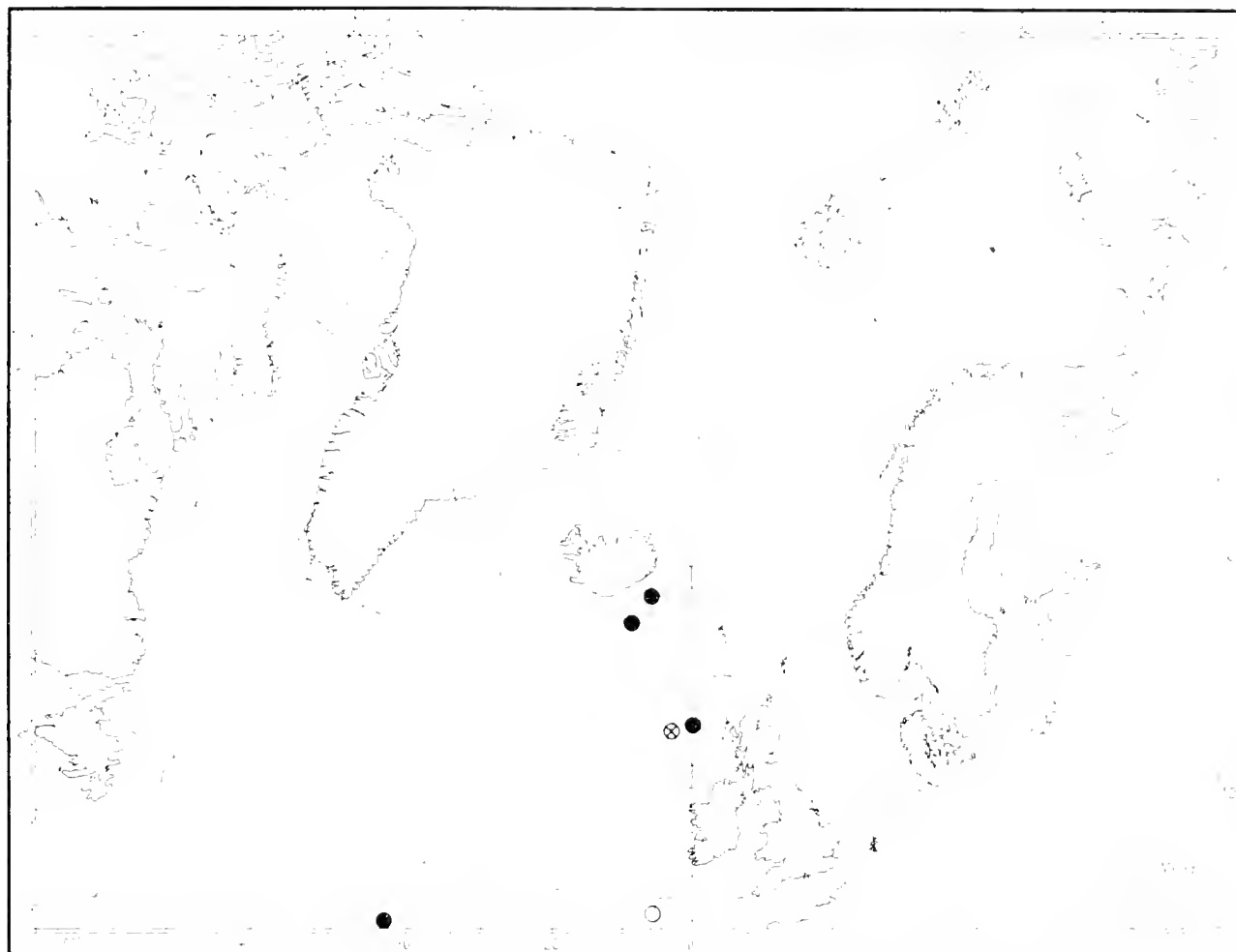


Fig. 14. Distribution in the northern Atlantic of *Equisetum*. ● new records; ○ previous records; --- record not yet published.

Solmundus sp. Fewkes 1888.

Equisetopsis laurentii Schläger 1891 p. 312.

— — — Birula 1896 p. 347.

— — — Vanhoffen 1897 p. 273.

Solmundus glacialis Grönberg 1898 p. 466, Pl. 27 figs. 7, 8.

— — — Aurivillius 1899

Equisetopsis laurentii Linko 1904b p. 219.

— — — Maas 1906a p. 185.

Solmundus glacialis Broch 1905 p. 8.

— — — Broch 1907 p. 8.

Equisetopsis laurentii Hartlaub 1909 p. 172, Pl. 76 figs. 7, 8.

— — — Bigelow 1909b p. 314, Pl. 32 figs. 2, 6.

— — — Kramp 1911 p. 431.

— — — Kramp & Damas 1925 p. 320.

— — — Thiel 1932b.

— — — Kramp 1933c p. 16.

— — — Bernstein 1931 pp. 9, 26.

— — — Ranson 1936 p. 212.

— — — Jaschnov 1939 pp. 112, 111.

— — — Dunbar 1912 p. 71.

— — — Kramp 1912 p. 97.

— — — 1913 p. 8.

Material: The numerous specimens in our collections are all from the Greenland waters and have been dealt with by me in the papers cited above. The species was well described and figured by BIGELOW (1909b), and I have nothing to add to his description.

MAYER (1910 p. 472) includes *Solmaris tetraonema* Hargitt (1902 p. 17 and 1901 p. 58) among the synonyms of *Equisetum laurentii*. An identification of these two medusae seems to me rather ques-

tionable. *Solmaris tetraonema* was deficiently described from a single specimen, somewhat damaged, so that "accurate determination was not practicable." The bell was flat, discoid, and between the four primary tentacles were "what appeared to be 4 undeveloped or rudimentary tentacles." It was found near the Gulf Stream off Woods Hole, thus quite outside the area of distribution of *Equisetopsis laurentii*. For geographical reasons it also seems to me rather objectionable to refer *Equisetopsis mertensii* Haeckel to *E. laurentii*. This latter has never been observed in the Pacific south of the Bering Strait, whence it was described by BRANDT (1838). *E. mertensii* was taken in the Sea of Japan, and HAECKEL expressly states that the only specimen was "ein sehr unvollkommen erhaltenes Spiritus-Exemplar." It should at present be regarded as an obsolete species, until some medusa be found in Japanese waters which may correspond to HAECKEL'S description.

Geographical distribution. *Equisetopsis laurentii* is an arctic, circumpolar species. THIEL (1932b p. 166) designates it as arctic-boreal, but its occurrence outside the true arctic regions is evidently restricted to areas, where cold currents prevail. It is common in the waters west of Greenland at least as far north as Smith Sound and is also found on the north coast of Ellesmere Land.¹ It follows the cold Labrador Current southwards along the coast of Labrador to Fogo Island, Newfoundland (BIGELOW 1909b). It also occurs along the entire east coast of Greenland. It has been found in deep water in a locality between Iceland and Norway in the cold bottom water of the Norwegian Sea. From Vardo in

¹ Recently recorded from several localities around Baffin Land (DUNBAR 1912).

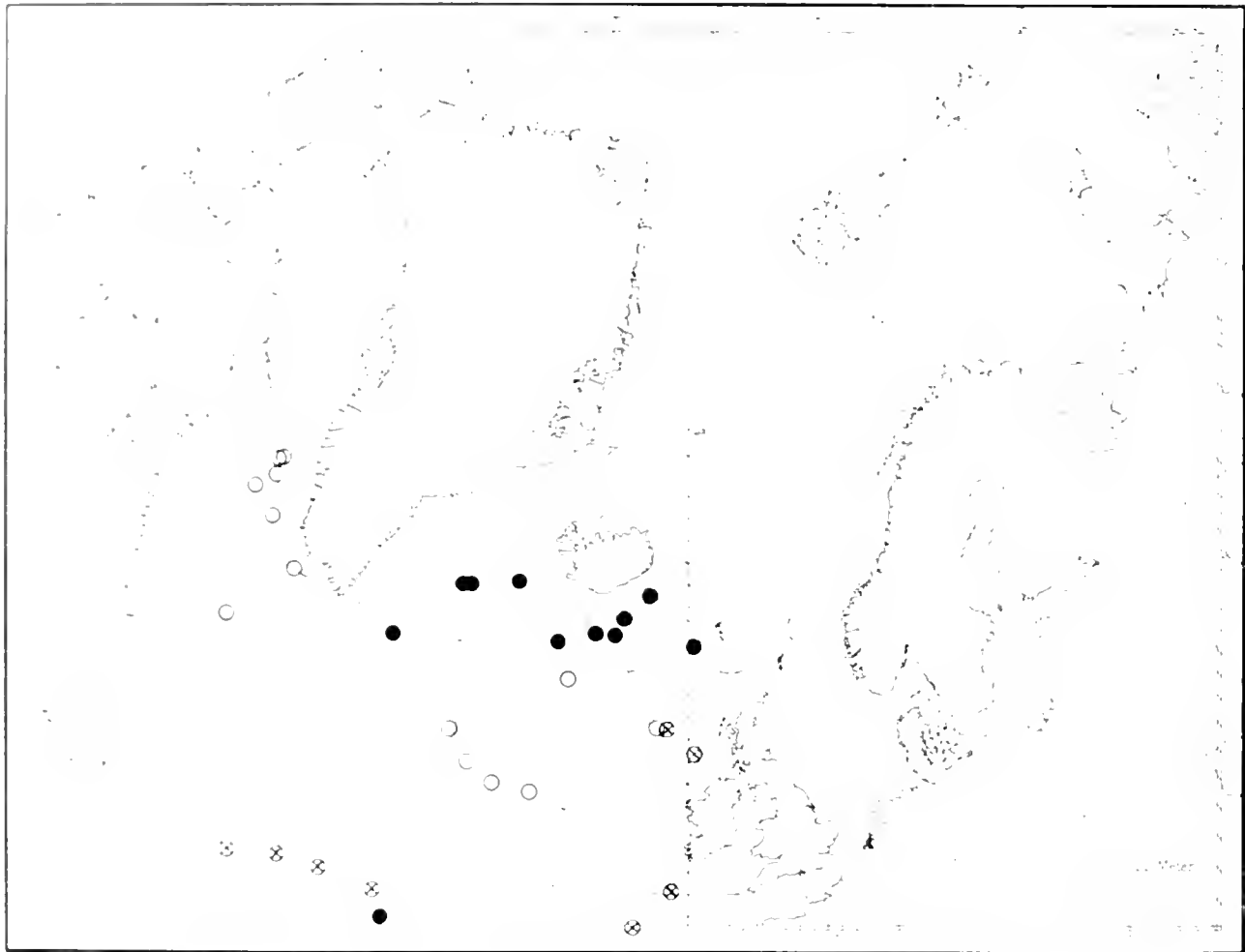


FIG. 15.—Distribution of Atlantic of *Eupima grimaldii*. ● new records, ○ previous records, ⊗ records not yet published.

It is distributed northwards to the north coast of Russia and Alaska (Bragg 1950) to the north coast of Alaska (Bragg 1950) from Laurent Bay in Bering Strait. It does not penetrate further southward into the Davis Strait. Its vertical occurrence ranges from the surface to 1000 m. In the Davis Strait it avoids the Atlantic water of the deep-sea area; in the North Atlantic it inhabits the upper strata which are influenced

62° 35' N 32° 53' W, 27, 1925	"Dana" St. 2137	1900 m wire	40 spec.
63° 19' N 26° 50' W, 24, 1928	"Godthaab" St. 1	1000 m	8 "
47° 02' N 31° 45' W, 27, 28, 31, 1928	"Dana" St. 4201	5000 m	12 "
62° 23' N 16° 05' W, 25, 1932		1402, 2000 m	500 "
		1402, 3000 m	650 "
		1102, 1000 m	125 "
62° 36' N 32° 48' W, 16, 1933		1687, 2000 m	600 "
63° 38' N 14° 13' W, 18, 1934		5113, 1800 m	75 "
		5113, 2000 m	130 "

Genus *Eginura* Haeckel

Eginura grimaldii Ma.

62° 35' N 32° 53' W, 27, 1925	40 spec.
63° 19' N 26° 50' W, 24, 1928	8 "
47° 02' N 31° 45' W, 27, 28, 31, 1928	12 "
62° 23' N 16° 05' W, 25, 1932	500 "
	650 "
	125 "
62° 36' N 32° 48' W, 16, 1933	600 "
63° 38' N 14° 13' W, 18, 1934	75 "
	130 "
62° 35' N 32° 53' W, 27, 1925	40 spec.
63° 19' N 26° 50' W, 24, 1928	8 "
47° 02' N 31° 45' W, 27, 28, 31, 1928	12 "
62° 23' N 16° 05' W, 25, 1932	500 "
	650 "
	125 "
62° 36' N 32° 48' W, 16, 1933	600 "
63° 38' N 14° 13' W, 18, 1934	75 "
	130 "

On the map, textfig. 15, are included eight localities in which this species was taken by the "Michael Sars" in 1910 m hauls with 1000-3000 m wire out.

Eupima grimaldii was well described by MAAS (1901 p. 38, Pl. III figs. 19-28) from a locality outside the Bay of Biscay, 47° 13' N, 17° 10' W, where one single specimen was taken by the Prince of Monaco at a depth of 781 m. The original specimen was only 42 mm in diameter, but the medusa may attain a much larger size; the largest specimens at my disposal are 31 mm wide.

The specific name has been discussed by THIEL (1935 p. 37 and 1936 p. 86), BRAGG (1938 p. 132), and KRYMP (1942 p. 100).

THIEL has found that in the tropical parts of the Atlantic young specimens may occur in the upper strata; in the northern Atlantic, however, this species is truly bathypelagic. In one locality, south of Iceland ("Dana" St. 2306) a specimen was taken in a haul with only 600 m wire out, at the same time as hauls with 2000 and 3000 m wire captured a large number of specimens, and in no other North Atlantic localities was this medusa taken in hauls with less than 1000 m wire out, 600 metres or more below the surface.

Very large specimens, more than 25 mm in diameter, were taken only on one occasion ("Dana" St. 2308, 3000 m wire out), otherwise there is no indication of a difference in the bathymetric occurrence of specimens of different sizes; young as well as adult ones, 1-25 mm wide, were taken at all depths.

Geographical distribution: The map, textfig. 15, clearly shows that the horizontal distribution of this species in the North Atlantic is strictly limited by the continental shelves and the submarine ridges connecting Scotland, Iceland, Greenland, and Baffin Land; it is generally distributed and very common throughout the deep-sea area of the northern Atlantic. It is also common in the areas around the Bermudas and the Azores, and is frequently met with in the deep-sea west of southern Europe and north-western Africa, but it seems to be fairly rare in the southern Atlantic. Some young specimens were taken by the "Meteor" in a few localities (TUNEL 1936), and it is recorded from the Guinea Current and the Benguela Current off the west coast of Africa, where it was taken by the "Valdivia" (VAXHOFFEN 1908); in the vast collections of the "Dana" expedition in 1930, from Cape of Good Hope northwards, this species was altogether lacking south

of the Cape Verde Islands. It is unknown in the Mediterranean, but it is recorded from several places in the Indian and Pacific Oceans; in the Pacific it occurs as far north as in the Bering Sea. *Equorea grimaldi* may thus be designated as a bathypelagic medusa with a world wide distribution, comprising all the great oceans, except the Mediterranean and the arctic seas, though apparently it is not equally abundant everywhere.

Narcomedusæ indeterminate.

Fragments of medusæ, probably Narcomedusæ, which I am unable to determine, were taken in the following localities:

60° 59' N, 22° 29' W, 1 st / ₅	Dana, St. 2306, 2000 m wire
59° 21' N, 37° 56' W, 1 st / ₆	2308, 3000 m
61° 13' N, 40° 57' W, 2 nd / ₇	2136, 1700 m
62° 35' N, 32° 53' W, 2 nd / ₇	2437, 1900 m
47° 02' N, 31° 45' W, 2 nd / ₈	4201, 5000 and 6000 m wires
62° 23' N, 16° 05' W, 2 nd / ₆	4402, 2000 m wire
62° 36' N, 32° 48' W, 1 st / ₆	4687, 2000 m
63° 38' N, 14° 13' W, 1 st / ₅	5113, 1800 and 2000 m wire

by HYGOM 1857; it must, accordingly, have been written between 1851 and 1857. It appears from these notes that STEENSTRUP himself has seen *Periphylla* between the Faeroes and Iceland during his voyage to Iceland in 1839, and also that on his arrival the medusa was named to him as "merchant-cap". The possibility, therefore, exists that FABER's medusa was really a *Periphylla*, and if so FABER becomes the author of the specific name *hyacinthina*.

In 1877 our whole collection of medusæ was sent to HÆCKEL in Jena together with a complete hand-written list of the specimens, written by LÜTKEN with provisional identifications of the species; the list was returned in 1880 with the final identifications in HÆCKEL's hand-writing. The several specimens of *Periphylla* are by LÜTKEN designated as "*Charybdea hyacinthina* Faber" or "*Periphylla*, s: *Charybdea hyacinthina*", and to all of them HÆCKEL has added the name *Periphylla hyacinthina* Steenstr. - Two of the first numbers in the list are dated 1837 and 1842 (specimens from Greenland), and this is probably what

induced HÆCKEL to quote the "Acta Musei Hafniensis" 1837 and 1842.

If FABER's medusa from Iceland was really a *Periphylla*, the specific name *hyacinthina* takes precedence over *dubia* Brandt 1838 as well as *bicolor* Quoy & Gaimard 1833. But, as mentioned above, there is no doubt that the same species was formerly described by PÉRON & LESNEUR (1809) under the name of *Charybdea Periphylla*. The description was not very clear, and the atlas, which was intended to follow the text, was never issued as a whole; but the figure of *Charybdea periphylla* was reproduced in 1834 by BLAINVILLE and in 1839 by MILNE-EDWARDS and leaves no doubt of the identity. The correct specific name of the only existing species of this genus thus becomes *periphylla* Péron & Lesueur.

In order to determine the generic name we must look up all the names which have been applied to the medusa since it was first described; they will appear from the following synopsis, in which the four so-called species are placed in four separate columns.

Péron & Lesueur 1809		Carybdea Periphylla		
Faber 1829	Medusa (Melitea) hyacinthina			
Quoy & Gaimard 1833		Carybdea bicolor		
Brandt 1837		Carybdea (Cyclophylla) bicolor	Carybdea (Cyclophylla) periphylla	
Brandt 1838				Chrysaora (Dodecabostrycha?) dubia
Lesson 1843		Carybdea bicolor	Carybdea periphylla	Cassiopa (Dodecabostrycha) dubia
Mörsch 1857	Dodecabostrycha Umataursak Groenlandorum			
L. Agassiz 1862		Quoyia bicolor	Charybdea periphylla	Dodecabostrycha dubia
Lütken 1875	Charybdea hyacinthina			
Hæckel 1880	Periphylla hyacinthina	Periphylla bicolor	Periphylla peroni	Periphylla dodecabostrycha

Carybdea Péron & Lesueur 1809 comprised two species, *periphylla* nov. and *marsupialis* Linné; this latter is generally adopted as the genotype of *Carybdea*, and it would cause a most disastrous confusion if that generic name were applied to *periphylla*. AGASSIZ (1862 pp. 172-174), it is true, adopted the division introduced by LESSON, in which *Carybdea* was retained as the generic name of *periphylla*, whereas the name of *Carybdea marsupialis* was changed to *Marsupialis planci*. BRANDT (1837 p. 187), on the other hand, divided the genus *Carybdea* P. & L. into three subgenera, one of which was *Carybdea genuinae* comprising *C. marsupialis* P. & L. and *alata* Reynaud.

FABER (1829 p. 197) referred his *Medusa hyacinthina* to the genus *Melitea* Péron & Lesueur. *Melitea purpurea* P. & L. (1809 p. 343) is however a Rhizostomid.

The name *Dodecabostrycha* is due to BRANDT and has been adopted as generic or sub-generic name by LESSON, MÖRSCH, and AGASSIZ, and as specific name by HÆCKEL; but all these and several subsequent authors have misunderstood the combinations of names in BRANDT's well-known paper of 1838, because they have paid no regard to his previous paper (BRANDT 1837 pp. 187 and 189¹⁾, in which the genus *Carybdea* is divided into three subgenera, one of which is *Cyclophylla* (comprising *C. periphylla* P. & L. and *C. bicolor* Q. & G.), and the genus *Chrysaora* is likewise divided into three subgenera, one of which is *Dodecabostrycha* with the species *quadrifurcatus* Lesson (= *Desmonema quadrifurcatus* of the Rhizostomeæ). In 1838, when BRANDT described

the new species *dubia*, he was in doubt of its affinities and thought it possible, but not certain, that it might belong to the sub-genus *Dodecabostrycha*. His doubt also appears from the various applications of queries. In the text (p. 387) the species is called ?*Chrysaora* (*Dodecabostrycha*!) *Dubia*; in the explanation of the plates and also on one of the plates (Tab. 29) is written ?*Chrysaora* (*Dodecabostrycha*) *dubia*, and on the other plate (Tab. 30) the same without any query. Accordingly *Dodecabostrycha* cannot be applied to any of the forms of *Periphylla*, because it was originally established for a Rhizostomid.

Cyclophylla, on the other hand, was introduced by BRANDT (1837 p. 187) as the name of a new sub-genus comprising the two species *C. periphylla* P. & L. and *bicolor* Q. & G. If we were to decide a change of the generic name *Periphylla*, we must accordingly take into consideration the possibility of replacing it with *Cyclophylla* Brandt.

This name, at any rate, takes precedence over *Quoyia*, introduced by AGASSIZ (1862 p. 173) for *Carybdea bicolor* Quoy & Gaimard; it would certainly be a most inconvenient name, especially if the name of the family, to which the genus belongs, should be derived from it (*Quoyiadae*!).

In the printed literature *Periphylla* is used for the first time as a generic name in HÆCKEL's monograph (1880 p. 418). A strict application of the law of priority would lead us to cancel that name in favour of the older name *Cyclophylla* Brandt 1837. It seems to me, however, most deplorable if *Periphylla* should be replaced by a name, which has appeared only once in the literature and never revived, whereas this very common and

¹⁾ HÆCKEL, in his fifth addition (p. 659) regarded almost all the names in that paper as obsolete.

of the genus *Periphylla* employed in this paper. I have followed most authorities in regarding *Periphylla* as a monogeneric genus. I most unfortunately retained the name *Periphylla* because I did not know at the time that this name had already been used by HAECKEL, nor was it invented by me. The name *Periphylla* from which HAECKEL adopted the generic name, was first used in the literature in 1829 by PÉRON & LESNEUR for two species which, in their classification, were placed in the genera *Carybdea*, *periphylla* and *marsupialis*, which were very far from each other, in two different families. Since then I saw that this medusa from Iceland and Greenland was placed by PÉRON & LESNEUR as *Carybdea periphylla* P. & L., and as a new generic name was required, he elevated PÉRON'S specific name to the rank of generic name, a procedure not uncommon in those days. HAECKEL adopted the idea and introduced the generic name *Periphylla* into the zoological literature, where it will be wise to retain it.

We may state that the name of the only species of this genus is given by

Periphylla periphylla (Péron & Lesneur).

Periphylla periphylla (Péron & Lesneur).

North-Atlantic records

- Medusa* (*Melita*) *hyacinthina* Faber 1829 p. 197.
Dactylostricha *Umataursak* *Greenlandorum* Mørch 1857 p. 95.
Carybdea *hyacinthina* Lutken 1875 p. 188.
Periphylla *hyacinthina* Haeckel 1880 p. 119, Pl. 21 figs. 11-16.
Dactylostricha *dubia* Fewkes 1881 p. 138.
Periphylla *hyacinthina* Fewkes 1886 p. 939.
Umata Fewkes 1886 p. 931.
hyacinthina Leysen 1893 p. 118.
Umata Vanhoffen 1892 p. 6, Pl. I figs. 1-10, Pl. II figs. 3-8, Pl. III figs. 1-7.
Nordgaard 1900 p. 18.
Browne 1903 p. 30.
Kramp 1913 p. 277.
Kramp 1914 p. 159.
Bjelow 1914 p. 27.
Broch 1914 p. 1, textfig. 1.
regina Broch 1914 p. 8.
hyacinthina Kramp 1920 p. 7.
Bjelow 1922 p. 138.
Hayno 1926 p. 286.
Runnstrom 1932 p. 31.
Kramp 1939 p. 29.
Straszny 1940 p. 6.
Kramp 1942 p. 106.

Material from the map, textfig. 16)

62° 21' N, 16° 56' W	15 ₆ 1895	E. Byst. 2 specimens
61° 43' N, 26° 57' W	15 ₆ 1925	Togoli 181, 17, 2 specimens
63° 32' N, 26° 23' W	15 ₆ 1926	76 1
62° 35' N, 27° 12' W	15 ₆ 1925	Thor. St. 153 4
62° 37' N, 19° 57' W	15 ₆ 1924	180 1
62° 38' N, 17° 58' W	15 ₆ 1924	183 1
61° 52' N, 35° 30' W	15 ₆ 1933	61 2
62° 36' N, 32° 48' W	15 ₆ 1933	71 3
63° 51' N, 33° 51' W	19 ₅ 1931	72 4
62° 23' N, 16° 05' W	19 ₅ 1932	165 5
61° 52' N, 35° 30' W	19 ₅ 1933	167 2
63° 38' N, 14° 43' W	18 ₅ 1931	Dana. St. 2308, 20,000m wire, 75 spec.
61° 43' N, 40° 57' W	18 ₅ 1925	2436, 6000m 14
		2436, 10000m 11
		2436, 17000m 21
62° 35' N, 32° 53' W	18 ₅ 1925	2437, 6000m 28
		2437, 10000m 26
		2437, 19000m 32
63° 32' N, 26° 23' W	18 ₅ 1925	2438, 6000m 1
		2438, 15000m 8
51° 48' N, 30° 30' W	1 ₇ 1931	1205, 3000m 1
		1205, 6000m 1
53° 38' N, 29° 41' W	1 ₇ 1931	1206, 1000m 2
		1206, 3000m 2
		1206, 6000m 38
		1206, 10000m 1
63° 51' N, 33° 51' W	19 ₅ 1931	1235, 6000m 2
62° 23' N, 16° 05' W	19 ₅ 1932	1402, 2000m 22
		1402, 3000m 6
		1402, 4000m 6
61° 52' N, 35° 30' W	15 ₆ 1933	1686, 6000m 2
62° 36' N, 32° 48' W	15 ₆ 1933	1687, 600m 7
		1687, 1000m 9
		1687, 1500m 140
		1687, 2000m 100
63° 38' N, 14° 43' W	18 ₅ 1931	5113, 1800m 12
		5113, 2000m 19

In our collections are numerous specimens from other localities in the North-Atlantic area, which have previously been recorded in the literature; they are indicated on the map, textfig. 16, by a special signature.

Periphylla periphylla is very abundant in the Atlantic Ocean south of the submarine ridges between Scotland, the Faroes, Iceland, Greenland, and Baffin Land. It has its main occurrence in the deep and intermediate strata, but it may sometimes ascend towards the surface and may then cross the ridges and be carried considerably farther northwards, but it evidently avoids areas where cold currents prevail. I have previously dealt with its occurrence in the waters west of Greenland (KRAMP 1912), where stray specimens are sometimes seen in the coastal area even as far north as Godhavn, about 70° N. From the deep-sea south of Iceland it may penetrate as far as the north-west point of the island, but we have no evidence of its occurrence off the north coast, unless the "*Melita hyacinthina*" Faber, recorded from Eyjafjord, be really a *Periphylla*. By the Gulf Stream it is frequently carried to the west coast of Norway; it is recorded from the Sognefjord in May and the fjords in the neighbourhood of Bergen in January, March, and July, and according to HAYNO (1926) it is a regular visitor to the Lofoten region, about 68° N. In Sognefjord it was taken in great numbers, partly small individuals, which might indicate that the medusa is indigenous in this deep fjord; it is also possible, however, that the fjord, being provided with a submarine barrier at the mouth, acts as a trap to the medusae which happen to be carried in across the barrier (Broch 1914 p. 8). *Periphylla* is also said to occur at Spitzbergen (MAYER 1910 p. 546), but I have been unable to find any precise records from this region in the literature.

As far as the horizontal distribution is concerned the present material only adds a number of localities within the same area, from which the species was previously known; but the numerous representative samples from different depths taken by the "Dana" provide us with valuable information of the vertical distribution of the various developmental stages of the medusa. In my paper on the West Greenland medusae (KRAMP 1912 p. 108) I have mentioned the disagreement between the bathymetrical occurrence, especially of the young individuals, in the Greenland waters as observed by me, and the corresponding observations from other

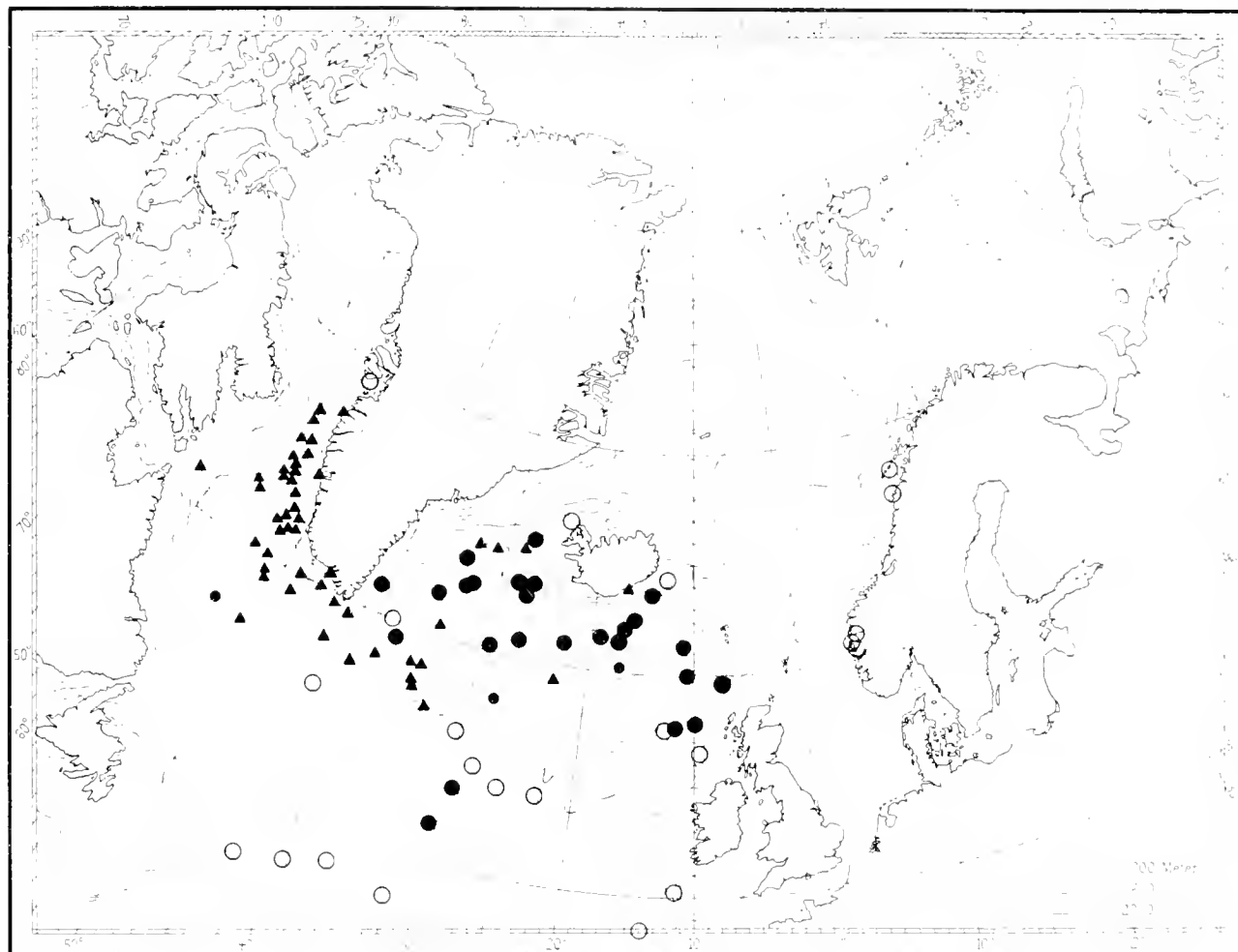


Fig. 16. Distribution in the northern Atlantic of *Periphylla periphylla*. ● new records; ▲ previous records, specimens in Zoological Museum, Copenhagen; ○ other previous records.

regions. In a previous paper (1913a) I pointed out that in Davis Strait the young specimens are particularly abundant in the deep strata, whereas STIASNY (1931 p. 363) is of the opinion that the small individuals mainly occur at higher levels than the large ones, and this opinion was based partly upon his own examination of the "Discovery" collections, partly on the statements by BROEN in the "Michael Sars" report (1914).

I have measured the diameter of all the specimens taken by the "Dana"; in the collections from the older expeditions the specimens were picked out and preserved more or less at random, and these collections are therefore not representative and are not considered in the following calculations. The results of the measurements are given in Tables V-VII, and for comparison I have summarized the results from other geographical areas as obtained by STIASNY, BROEN, and myself in Tables I-IV. The results are not directly comparable, because the diameter of the medusa has been measured in different ways by the various authors. The diameter of the "Dana" specimens is measured to the base of the marginal lappets; BROEN preferred to measure the diameter of the central disk above the annular furrow. None of these methods are quite reliable, because the shape of the medusa is very variable according to the state of contraction, and this applies not merely to the bell margin, but to the gelatinous central disk as well. Nevertheless, in spite of the unavoidable incorrectness of the measurements, they give an impression of the bathymetrical distribution of small, middle-sized, and large specimens in the various regions, sufficient for the purpose.

In the southern part of the Atlantic (Table I, STIASNY 1931, "Discovery" collections) the largest specimens (forma *regina*)

evidently have their principal occurrence between about 1250 and 2000 m below the surface, whereas middle-sized (forma *leptocynthia*) and small specimens (forma *dodecabostrigata*) are mainly caught between about 750 and 1000 m below the surface, and several specimens were even taken at depths of less than 250 m, whereas these stages were almost entirely lacking in the deepest strata, from about 1250 m downwards.

In the Mediterranean (Table II, KRUMH 1921, "Thor" collections) this species never seems to attain any considerable size, the largest specimens observed being only 35 mm wide. In the collections by the "Thor" specimens more than 25 mm in diameter were restricted to the deepest strata, about 1100-1700 m below the surface (1600-2600 m wire out; in my paper of 1921 I have calculated the depth as about one half the length of wire out, whereas two thirds probably comes nearer to the truth), whereas middle-sized and small specimens occur at all depths, "the young individuals seem normally to ascend into the intermediate, occasionally even into the upper strata, but sink to the deep water once more before breeding."

Table III is a contraction of the table, fig. 3, given by BROEN (1914, "Michael Sars" collections). In this portion of the North Atlantic, predominantly between 25° and 50° N., small and middle-sized specimens likewise occur at all depths, the majority being found rather high up in the water, about 500-600 m below the surface, occasionally even quite near to the surface, whereas the large individuals evidently prefer the deep strata.

Table IV illustrates the facts as observed in the Davis Strait by the "Trafalgar" expedition (KRUMH 1913); in the corresponding table in that paper are included some observations from the

Table I. Vertical distribution of *Periphylla* in the South Atlantic, from SRIJANJ 1931

Depth (m)	Diameter of specimens, mm		
	5-15	15-25	25-35
0-100	3	4	
100-200	2	1	1
200-300	2		1
300-400	24	10	
400-500	8	1	
500-600	1		14
600-700			5
700-800			1

Table II. Vertical distribution of *Periphylla* in the Mediterranean, from KRAMP 1924

Atmospheric depth (m)	Diameter of specimens, mm		
	5-15	15-25	25-35
0-100	2	2	
100-200	26	18	
200-300	4		
300-400	15	5	
400-500	1	2	2
500-600	17	34	11
600-700	3	2	2

Table III. Vertical distribution of *Periphylla* in the North Atlantic, between about 25° N and 50° N, from BROOK 1911.

Atmospheric depth (m)	Diameter of specimens, mm			
	5-15	15-25	25-40	40
0-100	6			
100-200	8	1		
200-300	5	3		
300-400	30	4	2	
400-500	13	4	2	
500-600	10	5	2	1
600-700	5	2		
700-800	4	6	2	

Table IV. Vertical distribution of *Periphylla* in the Davis Strait, from KRAMP 1923

Atmospheric depth (m)	Small	Medium-sized	Large
0-100		1	
100-200	7		
200-300		1	
300-400			1
400-500			4
500-600			1
600-700		1	
700-800			4
800-900			1

Irvinger Sea; they are left out here. In this region specimens of all sizes may be met with at all depths; all of them predominate in the deep strata, and young specimens are very rarely taken in the upper layers, where, on the other hand, large specimens have repeatedly been observed, not merely by the "Tjalfe" expedition, but also on other occasions.

The material collected by the "Dana" in the North Atlantic between about 52° and 61° N, gives the following results (Tables V-VII). Table V presents the actual number of individuals taken in hauls of two hours' duration with the 2 m ringtrawl (S 200), the numbers obtained by hauls of only one hour's duration being multiplied by 2. This table is directly comparable with the tables presented above from other geographical regions, and it shows that the vast majority of the smallest individuals, 1-10 mm in diameter, are densely crowded in the deeper strata, only very few being taken in the upper layers, whereas the middle-sized and large specimens seem to be somewhat more dispersed. The number of hauls made at each depth must, however, be taken into consideration, and this is done in Table VI, giving the number of specimens per one haul of two hours' duration with the S 200, (only positive hauls are included in the table). It confirms the impression of a dense accumulation of small specimens, 1-15 mm wide, at depths between about 1000 and 1350 m, the number decreasing gradually downwards and very suddenly upwards; specimens 15-25 mm in diameter are far less numerous than the small ones, and they are about equally common from 100 to about 1350 m below the surface; the same apparently applies to the larger specimens, 25-40 mm wide. Specimens more than 40 mm in diameter were altogether scarce, but were taken at all depths, even in the uppermost strata, and it may be noted that the specimen taken nearest to the surface (with only 50 m wire out, "Dana" St. 2306, south of Iceland) was the largest of the specimens brought home, being about 250 mm in diameter. Table VII, giving the percentage number of specimens of each size-class at the various depths, shows that at depths between about 100 and 650 m below the surface the stock of *Periphylla* mainly consists of middle-sized individuals, whereas in the deeper strata, from 1000 m downwards, the vast majority are quite young ones.

In this portion of the North Atlantic, accordingly, the vertical distribution of the various developmental stages of *Periphylla periphylla* is very nearly the same as in the Davis Strait, which was also to be expected, as the southern, deep part of Davis Strait is a direct continuation of the deep basin of the Atlantic Ocean. In both areas the majority of the specimens were taken in strata which had a temperature of about 3-4°.

SRIJANJ (1931, p. 361) is inclined to think that "the small stages of *Periphylla* have their origin on the continental slope or near the coast . . . and that they are driven into the open sea and sink slowly to greater depths," and also (p. 363) that the occasional occurrence of young stages in the superficial layers in the tropical regions "is perhaps to be explained by cold vertical currents which carry deep-sea animals to the surface." - To this may be remarked: In the North Atlantic there is no indication of small specimens of *Periphylla* being particularly plentiful in the neighbourhood of the continental slopes; the proportionate number of small and larger individuals seems to be independent of the position of the localities, and we may conclude, therefore, that breeding takes place everywhere in the deep-sea areas. The development of *Periphylla* is unknown, but it is generally supposed that, like other Scyphomedusae, the specimens pass through a fixed, benthonic polyp stage. The remarkable fact, pointed out above, that the majority of the young specimens are accumulated in certain intermediate strata (different in different regions) and almost lacking deeper down, regardless of the distance to the bottom, makes it probable that the development is independent of the sea bottom, so that no fixed bottom stage comes into the cycle of development.

Moreover it should be remarked that the horizontal currents

Tables V-VII. Vertical distribution of *Periphylla* collected by the "Dana" in the North Atlantic between about 52° N. and 61° N.

Table V. Actual number of specimens taken in hauls of two hours' duration with the 2 m ringtrawl.

Length of wire out, m	Approximate depth, m	Diameter of specimens mm							Total number of specimens	Number of hauls made	Number of hauls containing <i>Periphylla</i>
		4-10	10-15	15-20	20-25	25-30	30-40	40			
50	25	1	1	9	1
100	60	2	2	..	4	10	2
300	200	2	1	..	3	10	2
600	400	20	31	49	26	10	4	2	142	12	10
1000	650	2	13	19	17	1	2	3	57	7	4
1500	1000	302	28	42	6	1	349	3	3
1800-2000	1350	175	143	33	19	5	2	1	378	5	5
3000	2000	15	16	6	2	..	39	2	2
4000	2700	4	2	2	3	2	13	2	2

Table VI. Number of specimens per one haul with the 2 m ringtrawl in two hours.

Table VII. Percentage number of specimens of each size-class in the various depths.

Approximate depth, m	Diameter of specimens, mm				Total number per one haul	Approximate depth, m	Diameter of specimens, mm			
	4-15	15-25	25-40	40			4-15	15-25	25-40	40
25	0.1	0.1	25	0	0	0	100
60	0.2	..	0.2	..	0.4	60	50	0	0	50
200	0.2	0.1	0.3	200	67	0	0	33
400	4.2	6.2	1.2	0.2	11.8	400	36	53	10	1
650	2.1	5.1	0.4	0.4	8.1	650	26	63	5	5
1000	110.0	9.0	..	0.3	119.3	1000	95	5	0	0.3
1350	63.5	10.4	1.4	0.2	75.5	1350	86	11	2	0.3
2000	15.5	3.0	1.0	..	19.5	2000	79.5	15.5	5	0
2700	3.0	2.5	1.0	..	6.5	2700	46	39	15	0

in the intermediate strata of the Atlantic Ocean are slow, their velocity being somewhat more or less than 1 cm/sec., which means that it takes the water with its contents of plankton organisms about a year to pass through a distance of 300 kilometres; floating animals, bred on the continental slopes, must therefore have attained a rather advanced age before they can reach the central parts of the oceanic basins. E. g. at the "Dana" St. 4206, 53°38' N. 29°41' W., several small specimens of *Periphylla*, 5-10 mm in diameter, were taken in a haul with 600 m wire out, nearly 900 kilometres from the nearest continental shelf (the Rockall Bank); it should have taken them about three years to cover that distance providing they followed a straight line, and still more by a curved route. It is highly improbable, therefore, that these specimens have arisen from benthonic polyps growing somewhere on the continental slopes surrounding the northern basin of the Atlantic Ocean.

The difference in the vertical distribution of the various stages of development in different geographical areas must be due to differences in the hydrographical conditions. I quite agree with BIGELOW (1938 p. 158) that "it seems likely that the upper boundary for *Periphylla* is set by light combined with temperature." As pointed out by the same author in a previous paper (BIGELOW 1928 p. 496), "*Periphylla* never normally swims upward to the zone of strong illumination in Tropic seas, contrasting with its not exceptional occurrence right at the surface in arctic and sub-

arctic latitudes." In the various geographical regions the majority of the specimens are accumulated in strata with very different temperatures, about 13-15° in the Mediterranean, about 7-12° in the subtropical belt of the North Atlantic, about 2-5° in the southern Atlantic, about 3-4° in the northern part of the Atlantic north of 50° N. and in the Davis Strait. 15° C. probably marks the upper limit of temperature endurable to the species. The young medusæ are most probably bred in the intermediate strata above the great depths; the fact that in some areas the young individuals are inclined to ascend towards the surface, while in other areas, as in the northernmost portions of the Atlantic, the young specimens are almost entirely restricted to the deeper strata and only large specimens occasionally occur in the surface layers, is most probably due to differences in the vertical circulation of the water masses, and it seems likely that the large specimens are more capable of resistance against the movements of the water. Upwelling currents will carry young specimens towards the surface, but in the subarctic portions of the Atlantic the vertical movement of the water is predominantly downwards, and thus will keep the small medusæ in the deeper layers, while the large ones have the capability to force their way upwards towards the surface.

In the above discussion of the vertical distribution of *Periphylla* I have only dealt with the specimens collected by the "Dana" in the 2 m ringtrawl with bag of stramm (the so called

in his account of the "Michael Sars" Scyphomedusae (1914), both are identical with *N. rubra* Vanhöffen, and he may be right as far as *atlantica* is concerned. It seems to me, on the other hand, that *N. globifera* differs so much from the two other species that it is unsafe to unite them, unless further studies of a larger number of well preserved specimens perhaps might lead to the conclusion that the differences were only due to individual variation. The specimens of *N. rubra* from the eastern tropical Pacific, examined by BIGELOW himself (1909a p. 36), agree very well with the original description by VANHÖFFEN (1902a p. 29), in which it is expressly stated that the pedalia are prominent, the tentacular spaces between the marginal lappets larger than the rhopalar, and the marginal lappets narrow and pointed; in all the specimens examined by me (in so far as they are sufficiently well preserved to show the outline of the margin) the lappets are equidistant and broad and rounded, exactly as in BROCH's original specimens. Moreover the gonads of *N. rubra* are separated from each other by large and equal spaces, which is in striking contrast to the characteristic position of the gonads in *N. globifera* as observed by BROCH as well as by me.

Geographical distribution: *Nausithoe globifera* occurs in the deep and intermediate strata of the north-eastern Atlantic; two of the new localities recorded in the present paper are considerably farther north than the localities from which the species was previously known; the distribution is evidently limited by the continental slopes of Iceland and the British Isles.

Fam. Atollidæ.

Genus *Atolla* Haeckel.

Atolla wyvillei Haeckel.

North-Atlantic records:

- Atolla bairdii* Fewkes 1886 p. 936, Pl. 1-3.
- *verrillii* Fewkes 1886 p. 939, Pl. 4-5.
- *bairdii* + *verrillii* Fewkes 1889 p. 530.
- *wyvillei* Roule 1896 p. 302.
- *bairdii* Browne 1906 p. 179.
- - Vanhöffen 1906 p. 44, textfigs. 4, 5.
- *tenella* Hartlaub 1909 p. 477, Pl. 77 figs. 1, 2.
- *bairdii* Kramp 1913a p. 279.
- - Kramp 1911 p. 453.
- *wyvillei* + *bairdii* Broch 1914 p. 14.
- *bairdii* Kramp 1920 p. 7.
- *wyvillei* Bigelow 1928 p. 508.
- - Stiasny 1910 p. 14.
- - Kramp 1912 p. 109.

Material (see the map, textfig. 18):

- 61° 38'N, 32° 37'W., ²²/₅1895. "Ingolf" St. 12, 1958 m. 1 specimen
- 62° 00'N, 21° 36'W., ⁹/₈1895. - 40, 1591 m. 1 -
- 62° 58'N, 23° 28'W., ⁸/₆1896. - 73, 911 m. 1 -
- 62° 25'N, 28° 30'W., ¹⁴/₆1896. - 83, 1717 m. 1 -
- 64° 11'N, 31° 00'W., ²⁵/₆1896. - 91, 2328 m. fragments
- 66° 23'N, 7° 25'W., ¹¹/₇1896. - 101, 1802 m. 1 specimen
- 67° 57'N, 6° 44'W., ²⁰/₇1896. - 112, 2386 m. 3 -
- 68° 27'N, 8° 20'W., ²⁴/₇1896. - 118, 1996 m. 1 -
- 67° 29'N, 11° 32'W., ²⁵/₇1896. - 120, 1666 m. 1 -
- 65° 00'N, 28° 10'W., ¹⁹/₆1901. "Thor" St. 152, 800 or 1000 m wire, 2 specimens
- 61° 31'N, 19° 05'W., ¹⁰/₇1901. "Thor" St. 180, prob. 1800 m wire, 1 sp.
- 61° 30'N, 17° 08'W., ¹¹/₇1901. - 183, 1800 m. 1 -
- 57° 52'N, 9° 53'W., ⁸/₆1905. - 72, 1500 m. 1 -
- 48° 19'N, 13° 53'W., ³/₉1906. - 180, 1800 m wire, 3 spec.
- 60° 59'N, 22° 29'W., ²⁹/₅1925. "Dana" St. 2306, 2000 m wire, 1 spec.

- 47° 02'N, 31° 45'W., ²⁷/₂₈1931. "Dana" St. 1201, 3000 m wire, 1 spec.
- 1201, 1000 m. fragm.
- 1201, 5000 m. 7 spec.
- 1201, 6000 m. 13 -
- 49° 19'N, 30° 22'W., ³⁰/₆1931. - 1203, 1000 m. 2 -
- 53° 38'N, 29° 11'W., ¹/₇1931. - 1206, 600 m. 1 -
- 1206, 1000 m. fragm.
- 62° 23'N, 16° 05'W., ²⁵/₆1932. - 1402, 2000 m. 7 spec.
- 62° 36'N, 32° 48'W., ¹⁶/₆1933. - 1687, 2000 m. 7 -
- 63° 38'N, 44° 13'W., ¹⁸/₅1931. - 5113, 1800 m. 1 -
- 65° 14'N, 6° 06'W., ²¹/₅1931. - 5113, 2400 m. 1 -

FEWKES described this species (under the names of *Atolla bairdii* and *verrillii*) from the Gulf-Stream area off the east coast of North America between about 35 and 43° N. ROULE (1896) and BROWNE (1906) recorded it from the Bay of Biscay. According to VANHÖFFEN (1906, Nordisches Plankton) it has been taken near the Faroes, "ferner von der Ingolf-Expedition unter 64° N. Br., endlich vom "Michael Sars" in einem Fange aus 1900 m Tiefe noch höher im Norden..." but no precise localities are given by this author. It was taken at several stations by the "Michael Sars" North Atlantic Deep-Sea Expedition in 1910 (BROCH 1914) and by the Norwegian vessel "Armauer Hansen" in the north-eastern basin of the Atlantic in 1913 (KRAMP 1920). Also found in several localities in the deep parts of Davis Strait (KRAMP 1913a, 1911, and 1912). BIGELOW records it from the area off the east coast of North America as far north as off New York, about 40° N.

Atolla has about the same world-wide distribution as *Periphylla* in the deep portions of the oceans, but it seems to be more strictly confined to the deep strata, especially in northern waters. In the North-Atlantic area it is not, like *Periphylla*, known to ascend into the upper strata and to be carried across the submarine ridges by the currents along the west coasts of Greenland and Norway. It is rather astonishing, therefore, that it was taken by the "Ingolf" expedition at four stations (St. 101, 112, 118, and 120) and also by the "Dana" (St. 5113) in the deep, cold basin of the Norwegian Sea. *Atolla* was never taken by the "Ingolf" by implements for pelagic fishery, but always in the trawl. If this had happened once or twice, one might suppose that the medusa was captured on higher levels during the hauling in of the trawl, but the fact that all the specimens, more than ten, in all the nine localities, were taken in benthonic hauls, leads us to the conclusion that they really lived near the bottom. (The specimens recorded by ROULE from the Bay of Biscay were likewise taken in the trawl). In the five localities south of the ridges the depth varied between 914 and 2328 m, and the bottom temperature between 0.3 and 5.5; at the four stations in the Norwegian Sea the bottom temperature was very low, 1.1; 1.0 at depths between 1802 and 2386 m; by the "Dana" a specimen was taken in a haul with 2400 m wire out, i. e. about 1600 m below the surface, at a similar low temperature, about 0.8. We may conclude, therefore, that *Atolla* is indigenous in the deep strata of the Norwegian Sea, and this confirms the supposition that the two small medusae described by HARRLEMAN (1909) as *Atolla tenella* were young specimens of *Atolla verrillii*; they were collected by the "Belgica" in a locality between N. E. Greenland and Spitzbergen, 79° 34' N, 2° 37' W., at depths between 1200 and 1800 m. The area of distribution of this species accordingly comprises the high arctic regions.

The material available is too small to serve as base for a discussion of the vertical distribution of the various stages of development; some of the specimens are quite small, 10-20 mm in diameter.

Geographical distribution: *Atolla verrillii* is a true cosmopolitan deep-sea medusa, occurring in all the oceans from the

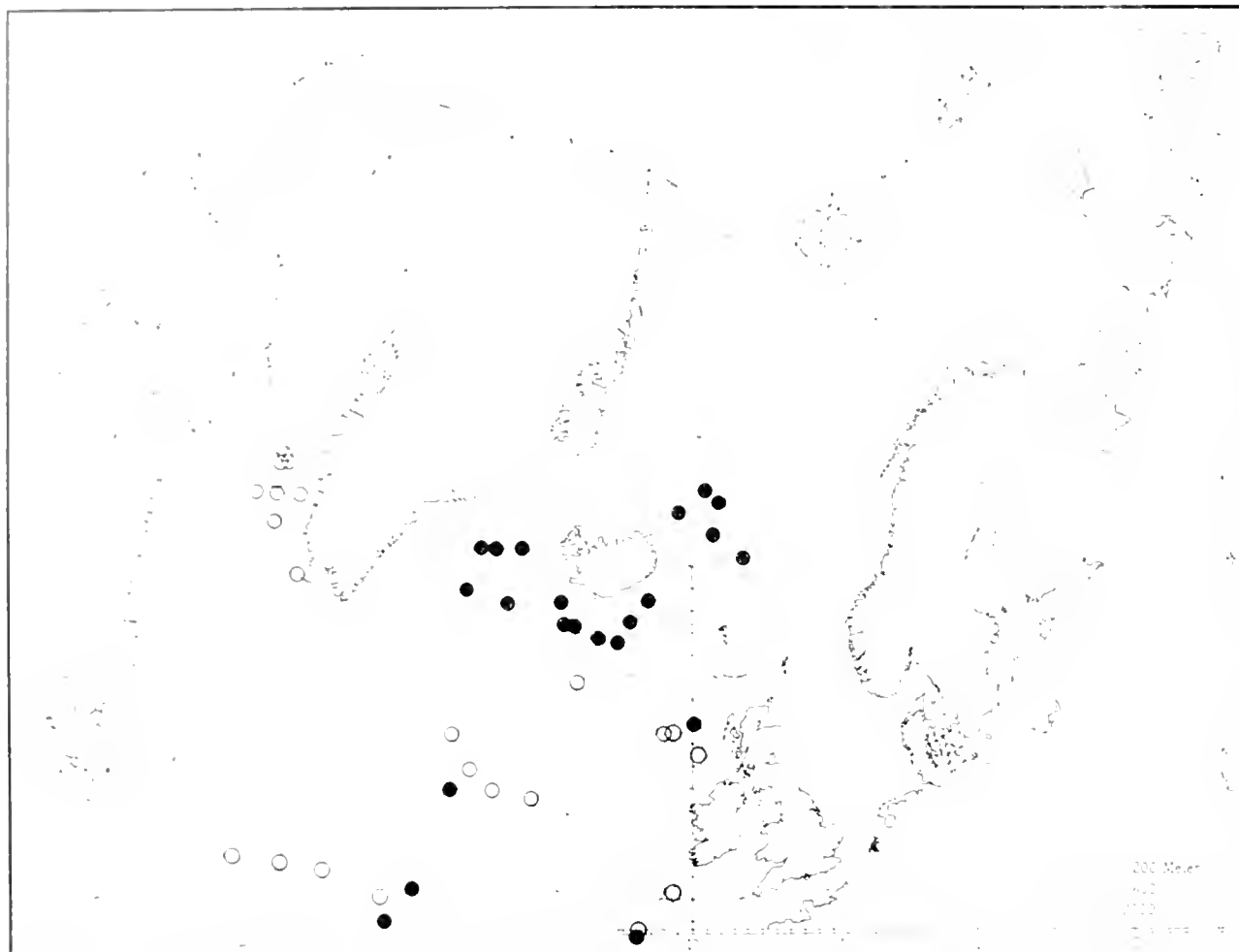


FIG. 18. Distribution in the northern Atlantic of *Aurelia aurilla*. ● new records; ○ previous records.

At the present time in the Pacific it is found as far north as 42° N. In the Atlantic area it penetrates into high-arctic

Order Semæostomæ.

Very few North Atlantic forms belonging to this order are known to be pelagic, and only occasionally are taken in the open ocean. They will, therefore, be briefly mentioned in this paper.

Pelagia noctiluca Forskahl

Pelagia noctiluca Forskahl

- W. J. E. Shaw, *J. Biol.*, 1900, p. 103.
 A. J. Clark, *J. U. S. Geol. Surv.*, 1901, p. 71.
 Clark, *ibid.*, 1901, p. 71.
 Clark, *ibid.*, 1901, p. 71.
 Clark, *ibid.*, 1901, p. 71.

Clark, *ibid.*, 1901, p. 71. (Clark, 1899, in *Trans. U. S. Geol. Surv.*, 1901, p. 71.)
 Clark, *ibid.*, 1901, p. 71. (Clark, 1899, in *Trans. U. S. Geol. Surv.*, 1901, p. 71.)
 Clark, *ibid.*, 1901, p. 71. (Clark, 1899, in *Trans. U. S. Geol. Surv.*, 1901, p. 71.)
 Clark, *ibid.*, 1901, p. 71. (Clark, 1899, in *Trans. U. S. Geol. Surv.*, 1901, p. 71.)
 Clark, *ibid.*, 1901, p. 71. (Clark, 1899, in *Trans. U. S. Geol. Surv.*, 1901, p. 71.)

medusa was found in several localities across the Atlantic between the edge of the Newfoundland Bank and the mouth of the Channel, a little south of Lat. 50° N., and the two stations, where it was taken by the "Dana", are within the same range. It is very abundant in the Bay of Biscay, whence it is frequently carried into the mouth of the Channel and northwards along the western coasts of Ireland and Scotland, sometimes even right to the Norwegian coast. According to VAXHOFFEN (1906, p. 15) a big shoal has been observed west of the Rockall Bank, about 57°-58° N., 22°-28° W. The specimen mentioned above, taken by E. BAST, shows that it may even occur still farther north, at 62° N., not very far from the south coast of Iceland.

Aurelia aurita (L.).

Syn. *Aurelia flavulata* L. Agassiz 1862, non Péron & Lesueur 1809.

- Material:
 62° 30' N. - 8° 21' W. NW. of the Faroes, 11₃ 1895, "Ingolf" St. 1
 10 specimens.
 Thorshavn on the Faroes, 2₃ 1901, "Dana", 2 specimens.
 Lerwick on the Shetland Islands, 2₆ 1905, "Thor", 1 specimen.
 Espvikpollen, Hardangerfjord, Norway, 2₆ 1927, R. SEJREK,
 5 specimens.

Moreover our collections contain numerous specimens from Iceland and West Greenland, previously mentioned by me (KRAMP 1932 and 1942).

In my paper on the West-Greenland medusa (1942, p. 126) I expressed the opinion that *Aurelia aurita* had probably invaded

the Greenland seas in recent time, because all the specimens collected previously proved to belong to *A. limbata*. Among the old drawings of Greenland meduse made by H. P. C. MÖLLER I have however found one, undoubtedly representing *A. aurita*; the locality is not stated, but the drawing was probably made somewhere in West-Greenland.

The localities mentioned above add nothing to the known area of distribution of this almost cosmopolitan species. It occurs along the entire west coast of Norway, round Iceland, and on the west coast of Greenland as far north as about 70° N., but has never been found on the East-Greenland coast.

Cyanea capillata (L.).

Syn. *Cyanea arctica* Péron & Lesueur.

Material:

58° 21' N., 0° 20' W., ¹²/₆ 1891. HARTZ. (West of the Hebrides).
 48° 09' N., 8° 30' W., ²⁰/₆ 1905. "Thor" St. 88. (Off the mouth of the Channel).
 57° 36' N., 7° 05' W., ²⁷/₅ 1908. "Thor" St. 8. (At the Hebrides).
 64° 15' N., 12° 10' W., ¹⁰/₇ 1927. "Dana" St. 3079. 700 m wire. (S. E. of Iceland).
 Tyrolerfjord, ²³/₆ 1929. and Nordfjord, Strindberg's peninsula, ³⁰/₆ 1929. The Danish East-Greenland Expedition 1929. (E. Greenland).

The occurrence round Iceland and on the coasts of Greenland is dealt with in some of my previous papers (1939, 1942, 1943). The specimens from the two East-Greenland localities mentioned above were not available to me, when I wrote my paper on the meduse in "The Zoology of East Greenland" (1943); these localities are in the Franz Joseph Fjord area, about 71° N.

Cyanea capillata is widely distributed in the coastal areas of temperate and arctic seas. In the waters west of Greenland it occurs at least as far north as in Jones Sound between Ellesmere Land and North Devon, about 76° N.; it is very common along the west coast of Greenland from Cape Farewell to Upernavik. On the east coast it has been found in several scattered localities as far north as Danmarks Havn, about 76½° N. It also occurs round Iceland, Jan Mayen, and Spitzbergen, and along the European coasts from France to northern Russia.

Order Stauromedusæ.

Besides the specimens from Greenland and Iceland of *Hali-mocynthus lagenæ*, *Halielystus octoradiatus*, *Lucernaria quadricornis*, and *Lucernaria haeckeli* our collections in Copenhagen contain some Lucernarians from a few other localities in the North-Atlantic area, mainly from the Faroes. The collections from Greenland and Iceland are dealt with in some of my previous

papers (KRAMP 1939, 1942, and 1943), the others will be briefly mentioned here.

Halielystus octoradiatus (Lamarek).

Syn. *Halielystus auricula* Clark.

Material:

The Faroes (without further statement). 2 specimens collected by STEENSTRUP.

Bordøvig, the Faroes, ²²/₅ 1899. TH. MORTENSEN. Numerous young specimens.

This species occurs on the east coast of North America north of Cape Cod; the west coast of Greenland as far north as Thule, about 76½° N.; the south coast of Iceland; the European coasts from France to northern Norway; Spitzbergen. Moreover known from Alaska and Japan.

Lucernaria quadricornis O. F. Müller

Material:

The Faroes. MÜLLER. 3 specimens (identified by STEENSTRUP).
 Bergen in Norway. KOREN. 1 specimen.
 69° 31' N., 7° 06' W., ²¹/₇ 1896. depth 2465 m. "Ingolf" St. 113. 1 spec.
 Svino, the Faroes, ¹/₅ 1897. H. JOHNSON. 1 specimen.
 Kvannesund, the Faroes, 1898. H. JOHNSON. 1 specimen.
 Bordøvig, the Faroes, ¹⁰/₅ 1902. 13-28 m. "Diana". A. DITLEVSEN. 1 specimen.

One of these localities, "Ingolf" St. 113, is of particular interest being situated in the Norwegian Sea, north-east of Iceland, at the considerable depth of 2465 m, which is unusual for *Lucernaria quadricornis*. It was originally referred to that species by G. M. R. LEVINSSEN, and a careful re-examination has convinced me of the correctness of the identification. The total height of the specimen is 22 mm, the pedicel is 9 mm in length and distinctly marked off from the calyx; the perradial notches between the arms are not much broader than the interradial. Thus it is quite different from *Lucernaria bathyphila* Haeckel, which was described from deep water in the southern part of the Norwegian Sea, between the Faroes and the Shetland Islands. The specimen is mentioned in the journal of the Ingolf Expedition, so that the possibility of a confusion of labels is excluded. The capture of a large specimen in Inglefield Bay, N.W. Greenland, at a depth of 930 m, shows that also in other arctic regions this species may occasionally occur in the abyssal region.

Lucernaria quadricornis occurs on the east coast of North America north of Cape Cod and along the west coast of Greenland as far north as Inglefield Bay, 77° 17' N.; also taken in some localities on the east coast of Greenland between about 65 and 71° N. Also recorded from Spitzbergen, but never seen at the coasts of Iceland. Common along the European coasts from southern England to the White Sea.

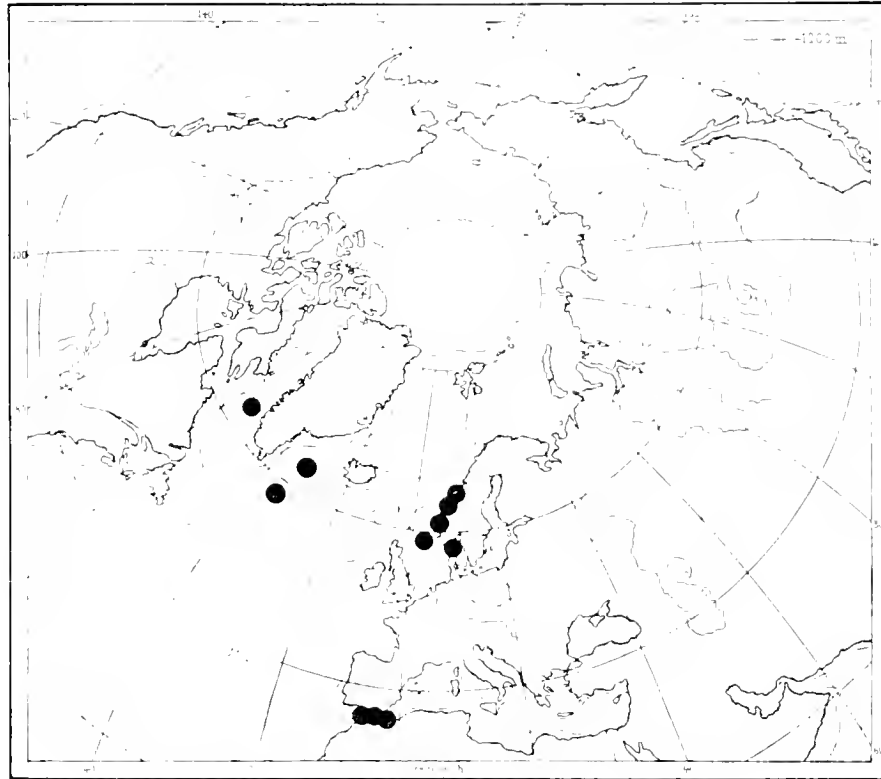


Fig. 19. Distribution of *Tiarranna rotunda*.

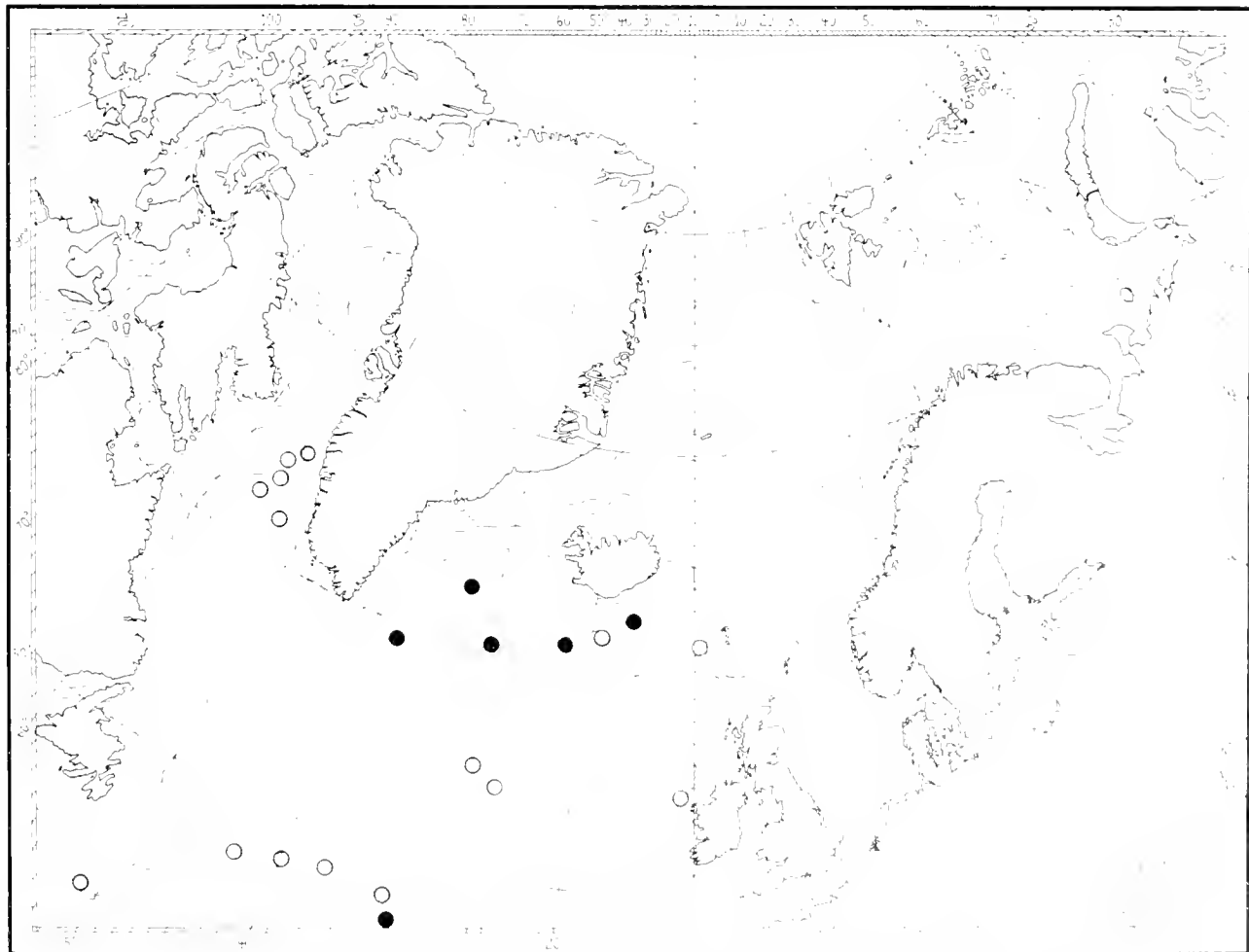


Fig. 20. Distribution in the northern Atlantic of *Chromalmonema rubrum*. ● new records; ○ previous records

61 22° 30' N, 17° 45' W, 1100 m; 2 specimens, men-
 62 23° 15' N, 16° 05' W, 1100 m; 2 specimens, the eastern
 edge.

		E. V&G	
N	W	1100	2400-2000 m wire.
N	W	1102	2307-1500 m.
N	W	1102	2308-3000 m.
N	W	1101	4201-3000, 4000, and 6000 m wire.
N	W	1102	4402-2000 m wire.
N	148° W	1100	4687-2000 m.

It is interesting to note the conception of this species as a bathy-
 thermophilic, generally distributed in the deep basins of the
 Atlantic, with the southern slopes of the submarine
 ridges of the Iceland, Greenland, and Baffin land.

The taxonomic position of *Chromatommata* has been much
 confused. It is a member of the medusae of West Greenland (KRAMP
 1942, p. 4). It is related to my original view (of 1919) that it belongs
 to the Anthomedusae related to the Laedidae among the Lepto-
 medusae, and to the Pandidae among the Panderidae among the
 Anthomedusae. Since I have become acquainted with RUSSELL'S
 classification of the medusae of Hydromedusae, III* (RUSSELL
 1951, p. 518), owing to the war this paper was inaccessible to me,
 I have looked up the Swedish library in 1945). I agree with him
 that it is related to the Anthomedusae and placed with
Leptomedusa in a special family, the Tiarannidae.

Chromatommata eburnea (Kramp): As previously mentioned
 (p. 142) and the colour of living specimens, as I saw them
 (p. 142, 1928), differs rather considerably from the
 yellowish-brown colour in preserved specimens. Pl. VI fig. 7
 is a reproduction of a coloured sketch made by me on board
 the "Thor" (p. 142).

Lausilia undulata (Forb. & Goods.):

- 63 26° N, 22° 28' W., ¹ 1927, "Dana" St. 3162, 600 m wire.
 62 23° N, 16° 05' W., ² 1932, 4402, 600, 1000, and
3000 m wire.

Ptychopoda crocea Kramp & Damas

Mangerfjord near Bergen, Norway, July 1932. TH. MORTENSEN.
 11 specimens, taken near the bottom, about 300 m.

Stauraphora mertensi Brandt:

- 65 27° N, 18° 17' W., ³ 1924, "Dana" St. 2193, 700 m wire.

Mitrocomella polydromata (Romanes):

- 57 21° N, 3° 20' E., ⁴ 1927, "Dana" St. 3006.

Cyrtocanna arctica Bigelow:

Mangerfjord near Bergen, Norway, July 1932. TH. MORTENSEN.
 15 specimens, taken near the bottom, about 300 m.

Phialella quadrata (Forbes):

- 57 30° N, 1° 58' W., ⁵ 1927, "Dana" St. 3009, 65 m wire.

KUSSE (1937a, p. 6) records "*Encopium quadratum*" from the
 Baltic, *publ. MORIN*, but this is erroneous.

Phialidium hemisphaericum (L.):

- 60 35° N, 3° 15' W., ⁶ 1926, "Dana" St. 2998, 600 m wire.

- 64 15° N, 12° 10' W., ¹⁰ 1927, 3079, 700 m.

- 63 26° N, 22° 28' W., ¹ 1927, 3162, 600 m.

Phialidium islandicum Kramp:

- 64 15° N, 12° 10' W., ¹⁰ 1927, "Dana" St. 3079, 700 m wire.

Octocanna fuscata (Q. & G.):

- 57 16° N, 9° 55' W., ¹ 1905, "Thor" St. 167, 1500 m wire, 1 spec.

Outside the Mediterranean this species was hitherto only known
 from some of the deep fjords on the west coast of Norway; the
 locality mentioned above is in the Atlantic between the west
 coast of Scotland and the Rockall Bank.

Zoogeographical Remarks

on the Medusæ of the northern Atlantic and adjacent Waters.

In Table VIII is given a general survey of the distribution of the North-Atlantic free-swimming medusæ, and on the following pages the composition of the fauna in each of the different geographical areas will be briefly discussed. In the Table I have included the coastal area on the east coast of North America between Cape Cod and the south-east point of Newfoundland (about 42–17° N.), because its pelagic fauna bears a considerable likeness to that in higher latitudes in north-western Europe.

The first three columns in the table comprise the three deep-sea areas: the Atlantic basin (including the deep, southern part of Davis Strait), the Baffin Bay, and the Norwegian Sea. The occurrence of the bathypelagic species is mainly restricted to these areas, but some of them may also be found in the neighbourhood of the coasts, either as stray visitors, or as constant inhabitants of deep fjords, e. g. on the west coast of Norway. On the other hand, neritic species with a somewhat prolonged pelagic life-time may sometimes be carried far out into the open sea, where they may be found in the upper strata above deep water; they are marked with an S in the table. The medusæ of the genus *Obelia* cannot be specifically separated; in the table the distribution of the corresponding hydroids is given, marked H. In some other species the hydroid has been found in areas from which the free medusa has not yet been recorded; in such cases the occurrence of the species is likewise indicated by an H.

The number of species of medusæ which at present are known from the areas here dealt with (north of about 50° N.) amounts to 142; but in Table VIII are also included 14 species taken off the American coast between Cape Cod and Newfoundland, but never recorded further north.

Among the 142 species 61 are Anthomedusæ, 38 Leptomedusæ, 1 Linnomedusæ, 16 Trachymedusæ, 6 Narcomedusæ, and 14 Scyphomedusæ (see Table IX). The majority of the species are meroplanktonic and neritic. It is true that the developmental cycle is unknown in several species, but probably all of the Leptolina (Antho-, Lepto- and Linnomedusæ) have a fixed polyp stage, and all of the Trachylina (Trachy- and Narcomedusæ) are presumably holoplanktonic. Among the Scyphomedusæ 8 species are known or supposed to be meroplanktonic, and these are also neritic; but we know that *Pelagia noctiluca* is holoplanktonic, and the vertical distribution of the oceanic species *Periphylla periphylla*, *Nausithoe globifera*, *Atolla wyvillei*, and *Poralia rufescens* gives reason to believe that they are likewise destitute of a fixed bottom stage. Among the species with doubtful development is also reckoned the high-arctic *Nausithoe limpida*, which is only known from the north east coast of Greenland.

Almost all the species of Trachylina are true oceanic forms, independent of the sea-bottom; the only exception among the northern species is *Ptychogasteria polaris*, which seems to spend

part of its time attached to the bottom of the sea, whence it occasionally swims towards the surface. The vast majority of the Leptolina are neritic forms, derived from polyp-stages attached to the bottom in the coastal areas, but some few of them occur in deep water. Most of these medusæ are however only taken at rather short distances outside the continental shelves or in deep fjords, which indicates that their fixed polyps live on the continental slopes and not in true oceanic basins. *Paraploea bathybia* is only known from the southern, deep part of Davis Strait; *Annularia affinis* and *Pandea rubra* occur in the eastern parts of the Atlantic Ocean; *Bythotrephes norragi* is likewise found in the eastern Atlantic and also in the Norwegian Channel and in some of the Norwegian fjords; *Calyropsis simplex* is only known from one of the fjords on the west coast of Norway; *Tiaranna rotunda* occurs at Gibraltar, in Norwegian fjords, and in the Skagerrak, but it is also known from the Irminger Sea and Davis Strait. *Ptychogena hyperborea* is only known from Smith Sound between Greenland and Ellesmere Land and is presumably an arctic deep-sea medusa; *Ptychogena crocea* and *Cyclocanna walshii* have been taken in some of the Norwegian fjords, the latter species also in the Skagerrak and off the east coast of North America; *Octocanna funeraria* is common in the Mediterranean and in some of the fjords on the west coast of Norway, and in the present paper it is recorded from the channel between Scotland and the Rockall Bank. In contradistinction to all these species *Chromatonema rubrum* should be designated as a true oceanic species, being generally distributed over the entire North-Atlantic deep-sea area, apparently quite independent of the continental slopes (see above, the map textfig. 20).

As far as the Trachylina are concerned, it is difficult to distinguish between bathypelagic species and species belonging to the upper strata. Most of the Trachymedusæ are well marked bathypelagic forms, but *Aglaura hemistoma* and *Licopi crocea*, which are stray visitors to the European coasts, belong to the upper strata, and *Aglantha digitale* occurs almost everywhere, though as a rule it is rare at great depths as well as near the surface; in most places it has its principal occurrence some hundreds of metres below the surface; in Table IX it is listed among the species of the upper strata. The Narcomedusæ are poorly represented in the northern seas. *Solmaris corona*, *Pogonantha clara*, and *Egina vitrea* have their main occurrence in the upper strata in warmer seas, but in these northern latitudes they show a marked tendency to seek deeper water. *Egina grimaldii* is a well marked deep-sea species with a cosmopolitan distribution; *Eginopsis laurentii*, on the other hand, is an arctic species, which decidedly prefers cold water; it is therefore mainly taken in the upper strata in the waters round Greenland and in the deeper strata in the Norwegian Sea.

Table VIII.

Area	Depth categories		Coastal areas	
	Atlantic Ocean	Baffin Bay	North Sea and Skagerrak	Other
Atlantic Ocean	II			
Baffin Bay		II		
North Sea and Skagerrak			II	
Other				II

Table VIII (continued)

	Deep areas		Coastal area	
	Atlantic Ocean	Baltic Bay	Northwestern Sea	
			Cape Cod - Newfoundland	
			Newfoundland - Ellesmere Land	
			West Greenland	
			East Greenland	
			S. and W. Iceland	
			N. and E. Iceland	
			Faroe	
			British Isles - West	
			Channel	
			North Sea and Skagerrak	
			Kattegat and Baltic	
			Norway S. of Lofoten	
			Norway N. of Lofoten	
			Spitzbergen	
			Barents Sea and Kara Sea	

Table VIII (continued).

	Deep-sea areas			Coastal areas															
	Atlantic Ocean	Baffin Bay	Norwegian Sea	Cape Cod - Newfoundland	Newfoundland - Ellesmere Land	West Greenland	East Greenland	S. and W. Iceland	N. and E. Iceland	Faroes	British Isles West	Channel	North Sea and Skagerrak	Kattegat and Baltic	Norway S. of Lofoten	Norway N. of Lofoten	Spitzbergen	Barents Sea and Kara Sea	
<i>Nansithoe globulera</i> Broch																			
<i>Atolla wyvillei</i> Haeckel, .	✓																		
<i>Pelagia noctiluca</i> (Forsk.)																			
<i>Chrysaora lysoscella</i> (L.)																			
<i>Cyanea capillata</i> (L.)		✓																	
<i>lamareki</i> Pér. & Les.																			
<i>Discomedusa lobata</i> .																			
<i>Phacelophora ornata</i> (Verrill) .																			
<i>Poralia rufescens</i> Vanh.																			
<i>Amelia aurita</i> (L.) .																			
<i>limbata</i> (Brandt)																			
<i>Rhizostoma pulmo</i> (L.)																			
Number of species . . .	32	6	6	39	19	32	11	29	19	24	68	68	73	39	59	24	15	26	

Table IX. Systematic and biological survey of the pelagic Medusa in the Atlantic Ocean and adjacent waters north of about 50° N.

	AXILIO- MEDUSAE	LEPTO- MEDUSAE	LEPTO- MEDUSAE	LEPTO- MEDUSAE	TRACHY- MEDUSAE	NARCO- MEDUSAE	SCYPHO- MEDUSAE	Total number
Meroplanktonic species	61	38	4				8	111
Holoplanktonic species					16	6	1	23
Development doubtful							5	5
Nentic species	57	34	4	1			9	105
Oceanic species	7	4		15	6	5		37
Mainly in upper strata	57	34	4	1		5	11	113
Mainly in deep strata	7	4		12	1	3		27
Total number of species	64	38	4	16	6	14		142

The Fauna in the Different Geographical Areas.

I. Coastal Areas.

1. The coastal waters between Cape Cod and Newfoundland. The plankton fauna, and especially the medusa, of this area has been thoroughly dealt with in several papers by H. B. DUNBAR, the most important being the large volume: Plankton of the Coastal Waters of the Gulf of Maine, 1926. The area comprises the Gulf of Maine, the Bay of Fundy, the southern coasts of Nova Scotia and Newfoundland, and the Gulf of St. Lawrence. Among the 40 species recorded from the area at least 20 are probably native to the species indigenous in the coastal waters; the other 20 species are possibly also indigenous, but as they are recorded only once with on very rare occasions, we cannot be sure of this (e.g., *Periphylla bairdella*, *Bougainvillia bairdiana*, *Lobelia bairdiana*, *Diphylosomma japonica*, and *Phacellophora* sp.). The other five were found in the Gulf of Maine, *Deplanura* sp., *Phacellophora* sp., *Phacellophora* sp., off Newfoundland. The holoplanktonic fauna of the area is likewise indigenous in the coastal area, but the water of the gulf is mainly due to influx from the North Atlantic. The warm water of the Gulf Stream carries with it a great number of species of southern origin: *Stomatopoda pteropoda*, *Tropidocyclops*, *Lucicutia borealis*, *Eurytemora alba*, *Eurytemora affinis*, *Alpheidae*, *Aphanaeus borealis*, *Aphanaeus borealis*, *Leucophaea tetraodon*, *Leucophaea tetraodon*, *Leucophaea tetraodon*. The Gulf Stream water is a surface stratum of warm and salt water, generally lying close outside the coast of the gulf, but when some of the 9 species mentioned above are recorded in the gulf, they serve as indicators of an influx of water from the off-shore banks of this warm water of the North Atlantic. A continuation of the ice-cold Labrador Current flows along the northern coasts of Newfoundland and Nova Scotia, and occasionally enters the gulf, carrying with it species of northern origin. When the following four species of medusa are recorded in the coastal area here concerned, they must be regarded as introduced: *Sarsia praeceps* (found as far south as the southern coast of Newfoundland), *Cataglyphis rosaceum* (found off Halifax), *Physophora polaris* (off Halifax), *Physophora polaris* (off Halifax). A great number of indigenous species also occur in the coastal waters of northern Europe; the only exceptions are *Physophora praeceps*, *Physophora praeceps*, and *Physophora praeceps* which are only known from the American coast. The fauna of the gulf is, with the only exception of the oceanic species *Physophora polaris* and the northern forms mentioned above, a mixture of species of the Gulf Stream and of those occurring in the

imperfectly known; up to now it comprises 19 species of medusa; the hydroid of *Obelia quincalata* occurs on the east coast of Labrador; the other species are recorded from scattered localities between Hudson Strait and Smith Sound (for details, see DUNBAR 1912¹ and KRAMP 1912); eight of them are decidedly arctic species, *Aglantha digitale* and *Cyanea capillata* are widely distributed in arctic as well as in boreal regions.

3. The west coast of Greenland. The fauna of medusa in the waters west of Greenland was thoroughly dealt with by me in a recent paper (KRAMP 1912) to which I refer. In the table above (Table VIII) 32 species are listed as occurring in the West-Greenland coastal area; three of them are however only represented by their hydroids (*Leuckartaria abyssii*, *Obelia quincalata*, and *Obelia longissima*). The species are all indigenous in the area, except *Periphylla peripholla* which belongs to the deep-sea outside the southern part of the coast; it sometimes ascends to the surface layers and may then be carried by the currents into the coastal area. The great majority of the species (28) are meroplanktonic and neritic. The fauna is of a mixed character corresponding to the mixed composition of the waters; among the 28 meroplanktonic species 7 are predominantly arctic, 6 arctic-boreal, 9 northern-boreal, 2 boreal, 1 southern-boreal, and 3 cosmopolitan. The 3 holoplanktonic species are *Aglantha digitale* and the two arctic forms *Equisops laureata* and *Physogastrin polaris*.

Almost all the West-Greenland medusa are also known from the coasts of northern Europe; the only exceptions are: *Halithobus pauper*, which occurs at Iceland but not on the coasts of the European continent (also recorded from the northern Pacific); *Cataglyphis multiseptata* and *Aurelia limbata* which are arctic-boreal forms in the Pacific; *Euchilota centriolaris* which is an American medusa distributed from Florida to Vineyard Sound and once taken in the southern part of the Greenland coast. The distribution of the various species along the west coast of Greenland depends on the hydrographical conditions in the various sections and is discussed by me in the paper quoted above.

4. The east coast of Greenland. The ice-cold water of the East Greenland Polar Current moves southwards all along the east coast of Greenland, decreasing in thickness towards the south; below it is a stratum of Atlantic water with temperatures above 0°; in the southernmost section this comparatively warm layer partly consists of water from the Iminger Current which turns westwards from Iceland, and it is possible that medusa are sometimes carried into the coastal area of East Greenland by this current. Up to now, however, we know only 9 species of medusa from East Greenland besides two species which are only represented

¹ DUNBAR, H. B. 1912. Plankton of the Coastal Waters of the Gulf of Maine. U.S. Fish and Game Commission, Washington, D.C.

² KRAMP, G. 1912. Die Fauna der Westgrönlandküste. I. Die Medusen. Denkschriften der Naturforschenden Gesellschaft in Zürich, 41, 1-100.

³ Owing to the war DUNBAR'S paper has only quite recently been accessible to me; four species, not previously known from this area, are recorded by him from the coasts of Baffin Land, and three others were taken farther north than known before.

by their hydroids (*Leuckartiara abyssii* and *Obelia longissima*). There are the usual three arctic holoplanktonic forms, and 8 meroplanktonic, neritic species, all of which must be considered indigenous in the area. *Nausithoe limpida* was taken off North East Greenland, and its further distribution is unknown. *Cyanea capillata* is generally distributed along the coast being fairly common; the other neritic species are found in some few scattered localities. All of the East-Greenland species also occur in West Greenland and in North-European seas.

5. Iceland. — 33 species of pelagic medusae are known to occur in the coastal waters round Iceland, including *Podocoryne carnea*, *Bougainvillia ramosa*, and the three species of *Obelia*, the presence of which is only stated by the occurrence of their hydroid polyps. The medusae of Iceland have previously been dealt with by me in a special paper (KRAMP 1939); in the table above (Table VIII) are included two species (*Paratiara digitalis* and *Ptychogasteria polaris*), which had been left out in the paper quoted, because they were only taken at some distance from the coast; they are, however, considered as belonging to the fauna of the Icelandic coastal waters.

Only two of the Icelandic species of medusae are holoplanktonic: *Aequorea digitale*, which is common everywhere, and *Ptychogasteria polaris*, which has been taken in two localities north and east of the island at depths of somewhat more than 500 m. The bathypelagic *Periphylla periphylla* is rather frequently carried towards the south and west coast from the Atlantic deep-sea area. The remaining 30 species are meroplanktonic and neritic, and most of them are indigenous in the Icelandic coastal areas, but rather few are generally distributed around the island.

The northern branch of the Gulf Stream moves towards the south coast of Iceland turning westwards (the Irminger Current) and continues in a clockwise direction along the west and north coast, but it is considerably diminished in extent after turning round the north-west point of the island under the cooling influence of the Polar Current coming from the North; the main body of the Polar Current, however, follows the east coast, which therefore is the coldest part of the Icelandic coasts. Off the south-east coast there is usually a fairly sharp limit between the cold water of the Polar Current and the warm water derived from the Gulf Stream. The combined effect of the currents is, accordingly, a marked decrease of the temperature of the water from the south-east round the whole island in a clockwise direction. Species belonging to southern and boreal regions, therefore, are mainly found on the south and west coasts, occasionally carried round the corner to the north coast, whereas arctic species are only taken on the north and east coasts. The species which have a wide distribution round the island are mainly those belonging to arctic-boreal and northern-boreal tracts (for details, see KRAMP 1939, especially Table III, p. 31). In the table above (Table VIII) the medusae occurring on the south and west coast (29 species) and those taken on the north and east coast (19 species) are separated in two columns.

Specimens of meroplanktonic medusae, especially of the larger forms with a fairly long pelagic period, may undoubtedly sometimes be carried to the southern coasts of Iceland from distant areas to mix with the indigenous population, and some decidedly southern forms, such as *Bougainvillia ramosa* (the hydroid with medusa buds taken once on the south coast), *Lizzia blondina* (taken once near Cape Horn), *Leuckartiara octona* (on the south coast, twice), and *Cyanea lamarchi* are probably not constant inhabitants of the Icelandic coastal waters, but may occasionally settle there for some time.

The Icelandic fauna of medusae has a predominantly boreal character, and the majority of the species occur in the western as well as in the eastern parts of the North-Atlantic area. Only one species has a decidedly western distribution: the arctic medusa *Halitholus pauper*, known from the northern Pacific and from

both sides of Greenland; in Iceland it has only been taken off the north-western part of the coast. On the other hand, the following seven species have never been recorded from the western Atlantic: *Lizzia blondina*, *Podocoryne arcuolata*, *Paratiara digitalis*, *Neotarris pileata*, *Phialidium islandicum*, *Eutonina indicans*, and *Cyanea lamarchi*. They are all inhabitants of the coastal waters in north-western Europe (*Neotarris* and *Eutonina* also known from the Pacific), and they have a predominantly boreal or southern distribution. *Phialidium islandicum* occurs all round Iceland, the others have only been taken on the southern and western coasts.

6. The Faroes. — The fauna of medusae at the Faroes is imperfectly known and has never been the subject of special treatment. The number of species recorded up to now and listed in Table VIII amounts to 20, but a closer examination of the neritic fauna around and between these small islands would certainly add several species to the list, especially such which are common to Iceland and the British Isles.

With the exception of *Aequorea digitale* the medusae known from the immediate neighbourhood of the Faroes are all meroplanktonic and neritic, and most probably all of them are indigenous in the area, though in this respect *Bougainvillia saeperciliaris* and *B. principis* may perhaps be a little doubtful. Besides the three species of *Obelia*, *Phialella quadrata* is included in the list, because the corresponding hydroid has been taken at the islands, whereas the free medusa has not yet been observed.

The Faroes are washed by the Gulf Stream, and in accordance herewith their marine fauna is mainly of a boreal character. Under normal conditions a number of species with a predominantly southern distribution might also be able to occur round the islands; the cold water of the East-Iceland Polar Current, which is moving southwards along the north-eastern edge of the Wyville Thomson Ridge below the Gulf Stream, may however occasionally rise towards the shallow-water area of the Faroe plateau, and the temperature of the water around the islands is therefore subject to considerable variations, which may prevent a constant settling down of the southern species. Only two of the medusae hitherto recorded from the Faroes (*Mitrocomella polydiademata* and *Phialella quadrata*) do not occur in the Icelandic waters, whereas all of the Faroese species are known from the British coasts and (with the exception of *Phialella quadrata*) from the southern part of the west coast of Norway.

7. Atlantic coasts of the British Isles. — The medusae occurring off the coasts of the British Isles have invoked the interest of several British zoologists, particularly E. FORBES, E. T. BROWNE, and in recent years F. S. RUSSELL. As most of their investigations have been carried out from the various marine laboratories, the different portions of the coasts have not been equally well examined. We may however suppose that almost all the species actually occurring there have really been observed. In Table VIII the British medusae are enumerated under three different columns, because the conditions on the Atlantic coasts, in the Channel, and in the North Sea present rather considerable dissimilarities.

68 species of pelagic medusae are recorded from the coastal areas west of the British Isles; the two bathypelagic species *Bythotrephes murrayi* and *Otococconeum junceum* were however only taken in a few localities on the continental edge; they are included here, because, as mentioned above (p. 53), they are supposed to pass their fixed bottom stage on the continental slopes and not in the true oceanic basins. *Aequorea digitale* is very common, and it is mainly the comparatively small, southern form, forma *rosea*, which occurs on the British coasts. *Solmaris corona* is indigenous in the waters west of Scotland and Ireland and is mainly taken in the upper strata. The third holoplanktonic medusa is *Pelagia noctiluca*; it is probably not indigenous in British waters, but is frequently carried by the currents from more southerly

Bythotrephes murrayi, *Tiaranna rotunda*, and *Cyclocanna welshi* are bathypelagic forms only found in the deep strata of the Skagerrak.

Tima bairdi probably is indigenous only on the slopes of the Norwegian Channel; during the period of its pelagic life, which lasts about a year, it spreads far around in the surrounding waters, into the Kattegat as well as towards the south and west in the North Sea, though never into the southernmost portions.

The following species are only known from a few localities, where they however seem to be indigenous: *Margelopsis haeckeli* (Belgium and Heligoland), *Eleutheria dichotoma* (Gullmarfjord on the Swedish coast of the Skagerrak), *Chalumena radiatum* (Belgium, the Limfjord in Jutland, Gullmarfjord), *Bougainvillia nordguardi* (once observed in Oslofjord, else known only from the surroundings of Bergen), *Bougainvillia marborivua* (in the Heligoland Bight, probably transported by ships from the Antarctic), *Nemopsis bachei* (Zuider Sea), *Agastrea mira* (Dover and Heligoland), *Eutima elephas* (Heligoland), *Willia stellata* (east coast of Scotland), *Pachella polymena* (one locality south of the Shetland Islands), *Gamionemus marbachi* (Oslofjord and Gullmarfjord).

A comparatively large number of the medusae (13 species) occurring in the North Sea and Skagerrak have never been recorded from the south and west coast of the British Isles: The two deep-sea species *Tiaranna rotunda* and *Cyclocanna welshi*; the decidedly northern species *Bougainvillia superciliaris*, *Stauropora mertensi*, and *Phialidium islandicum*; *Leuckartiara abyssii* and *Tima bairdi* which have their proper home in the Norwegian Channel; *Eutima indicans*, common in the North Sea except in the southern part; and the following species with a narrow distribution (see above): *Margelopsis haeckeli*, *Bougainvillia nordguardi*, *B. macbratnae*, *Nemopsis bachei*, and *Eutima elephas*.

10. The Kattegat and the Baltic. - The fauna of Hydro-medusae in the Danish waters inside the Skaw was thoroughly dealt with by me in the paper quoted above (KRAMP 1927) and also in the series "Danmarks Fauna" (1937) comprising also the Scyphomedusae (as well as the Siphonophora and the Ctenophora). To the lists given in these papers should only be added *Stauridium productum* and *Leuckartiara abyssii*, the hydroids of which have been found in the Kattegat.

The Kattegat is an interesting sea, being a transition area between the North Sea and the Baltic; the salt water of the North Sea (the Jutland Current) penetrates into the deep channels of the Kattegat as an undercurrent, still traced through the Belts into the western part of the Baltic; it is overlaid by the brackish water of the Baltic running northwards to the Skagerrak, where it is forced towards the coasts of Sweden and Norway and finally reaching the North Sea off the Norwegian coast.

39 species of pelagic medusae occur in the Kattegat, 26 of which are indigenous there. The following 13 species are more or less regular visitors from the Skagerrak, most of them derived from the North Sea: *Bougainvillia britannica* (sometimes penetrating into the Great Belt which constitutes the principal connection between the Kattegat and the Baltic), *Leuckartiara nobilis* (found only once, in the Great Belt in 1923), *Laobucca undulata*, *Stauropora mertensi* (only under exceptional conditions), *Melicertum octocostatum*, *Mitrocomella polydiademata*, *Cosmetira pilosella*, *Eucheilota maculata*, *Saphenia gracilis*, *Eutima insignis*, *Chrysaora isosecles*, *Cyanea lamarecki*, and *Rhizostoma pulmo*. Under exceptional conditions (as in 1936) *Cyanea lamarecki* may be carried as far south as into the Belt Sea. *Chrysaora* and *Rhizostoma* are very rarely seen in the Kattegat, but in 1933 *Chrysaora* appeared in the northern part of the Kattegat, and in the same year *Rhizostoma* even occurred at the northern entrance of the Great Belt.¹

¹ In October and November 1946 *Rhizostoma* was observed in the southern Kattegat and in the Sound, even as far south as in Køge Bay south of Copenhagen.

In the Baltic 16 species have been taken, 9 of which are indigenous in the western part, whereas only 1 species (*Sarsia tubulosa*, *Halitholus circatus*, *Cyanea capillata*, and *Aurelia aurita*) are indigenous in the Baltic proper east of the Gdser-Düsserort threshold. Of special interest are the two arctic medusae *Euphysa tentaculata* and *Halitholus circatus*. The former occurs in the southern Kattegat, the Belts, and the western Baltic, and besides it is only known from the Barents Sea and West Greenland; *Halitholus circatus* is very abundant in the deep, cold basins of the Baltic proper, less common in the Belt Sea and the Kattegat, and its further distribution is purely arctic (see Table VIII); it must be designated as an arctic survivor in the Baltic.

The only holoplanktonic medusa in this area is a particularly small form of *Aequorea digitata*, indigenous in the deeper parts of the Kattegat, occasionally carried into the Baltic.

11. The west coast of Norway. A special treatment of the Hydromedusae occurring along the west coast of Norway is given by KRAMP and DAMAS (1925); some few species are added to the list by RUNNSTRÖM (1932), KRAMP (1933b), and REES (1938 and 1941).

A branch of the Gulf Stream approaches the Norwegian coast, mainly through the Faroe-Shetland Channel; it is particularly powerful in late summer and autumn. As a rule it is separated from the coast by a belt of coastal water of lower salinity, partly derived from the Baltic Current. The Gulf Stream follows the Norwegian coast northwards and is still traced in the Barents Sea, but north of the Lofoten it is considerably cooled by the influence of the polar water, which accounts for the great difference in number of species found in the two portions of the Norwegian coastal region (59 in the southern section against 23 in the northern section). The majority of the species are meroplanktonic forms indigenous in the coastal areas. The few holoplanktonic forms are: *Ptychopasteria polaris* (taken in several localities in the northern section, rarely seen in the southern), *Homococconeia platygonon* (in some of the fjords in the surroundings of Bergen), *Aequorea digitata* (generally distributed), *Solmaris coronata* (rather frequently carried to the Norwegian coast by the Gulf Stream), and *Pelagia noctiluca* (a rare visitor from the Atlantic). *Periphylla periphylla*, which possibly also is holoplanktonic, is frequently carried to the Norwegian coast, and the possibility cannot be excluded that it is also indigenous in some of the deep fjords.

The following meroplanktonic species are constant inhabitants of the deep strata in some of the fjords of the southern section: *Bythotrephes murrayi*, *Calyceopsis simplex*, *Tiaranna rotunda*, *Ptychogona crocea*, *Cyclocanna welshi*, and *Ocyropsis pinnata*.

Besides the above-mentioned holoplanktonic species *Cosmetira pilosella* and *Phialidium islandicum* are probably visitors brought to the southern part of the Norwegian coast by the Gulf Stream.

When the following species occasionally are met with in the coastal water of southern Norway, they are most probably derived from the Jutland Current, which sometimes crosses the mouth of the Skagerrak: *Eutima elephas*, *Epyroca citrina*, *Chrysaora lysosocella*, *Cyanea lamarecki*, and *Rhizostoma pulmo*.

Apparently very few of the numerous species which are indigenous in the southern section only are able to pass the boundary at Lofoten; *Laobucca undulata* and *Melicertum octocostatum* may probably be regarded as visitors from the southern section, when they occur north of the Lofoten; all the other species found in the northern section are probably indigenous there.

12. Spitzbergen. The medusa fauna of Spitzbergen is imperfectly known. As the Gulf Stream is still traced at least on the western coasts, one might expect to find occasional visitors from southern regions, but the 13 species hitherto observed are commonly occurring also in other arctic regions. Three of the species are holoplanktonic, *Aequorea digitata* and the two decidedly arctic forms *Ptychopasteria polaris* and *Eugnopsis laurentii*.

of the Gulf Stream (WILSON, 1931; KILGUS, 1931). The fauna of the northern part of the Gulf Stream is characterized by LINSKO (1931) and JASCHKOV (1932). Twenty species of the northern part of the Gulf Stream have been recorded by SARGENT (*Uppister*, *S. barbata*, and *S. setacea*) in the planktonic zone (see Table VIII). The fauna of the southern part of the Gulf Stream (*Heterostoma*, *Chelapoda*) are known from the Gulf Stream as well. The fauna is however not purely northern, but the Gulf Stream is still remarkable for the presence of a good accordance net with the fauna of the northern part of the Gulf Stream and northern boreal species. The fauna of the Gulf Stream is characterized by BERNSTEIN (1934), as the fauna of the northern part of the Gulf Stream and the northern boreal species. *Uppister*, *S. barbata*, and the small Trachymedusa *Homonotus* (*Uppister*) are recorded (p. 17), both of which demonstrate their origin from the Barents Sea.

II. The Deep-Sea Areas.

A. The Atlantic basin north of about 50° N. The metaplanktonic, neritic medusae. The great majority of the metaplanktonic medusae are neritic, and the duration of their pelagic life is usually too short to enable them to be drifted away from the coastal areas, from which they are derived. Some few of the larger species may, however, sometimes be met with at considerable distances from their place of origin. The Anthomedusae *Leulastania robusta* and *Neotauris pilenta*, and the Leptomedusae *Leulastania data* and *Halopsis ocellata*, all of which are reported from the coastal areas of the north-eastern Atlantic, have also been taken above deep water west of the British Isles and south-west of Iceland, following the circulations of the Gulf Stream system. The Leptomedusa *Phialopsis diapensis*, which is indigenous to the eastern tropical Atlantic, has also occasionally been taken south-west of Iceland and in the Irminger Sea. The Scyphomedusa *Yusistha punctata*, indigenous in the warm portions of the ocean, is recorded by VANHOUTEN (1902, p. 29) from the Gulf Stream north-west of Scotland.

Very peculiar is the occurrence of three East Atlantic neritic species in the central portion of the North Atlantic between 30° and 40° W. *Phialopsis diapensis* was taken in July 1910 by the "Michael Sars" east of the Newfoundland Bank (St. 81), 48° 02' N, 29° 55' W, in a haul with 200 m. wire out. Some specimens of *Leulastania robusta* were taken in June 1931 by the "Dana" (St. 1201) 19° 10' N, 17° 02' W, and 17° 02' N, 31° 15' W (St. 1201) and 19° 19' N, 31° 22' W (St. 1203) in hauls with 100 and 50 m. wire out; at St. 1203 a few specimens of *Neotauris pilenta* were also found (in hauls with 100 m. wire out, possibly caught on a higher level than the 100 m. net). Apart from one record of *L. robusta* at New York (p. 12, Fines, 1937, p. 26) these three species were only recorded from the coastal areas in the eastern parts of the Atlantic basin and the water south and south-west of Iceland. We know from the investigations of the surface water currents in the northern part of the Atlantic that the Gulf Stream has a strong influence on the habitual circulation of the surface water, and one might conclude that the occurrence of these species in these western tracts is due to their being carried out of surface water from the eastern part of the Atlantic basin (to the south-west). In all three localities the water was evidently pure Gulf Stream water (temperature 14-15° C.) together with such species as *Phialopsis scottiana*, *Rhopalumma* (*Rhopalumma*) *relatum*, etc. The only possible explanation seems to be that the species *Leulastania robusta*, and *Neotauris pilenta* are carried from the American coastal waters, and that the species *Phialopsis diapensis* has its only record of *L. robusta* at New York, and that it has never been observed there.

upper strata above the northern part of the Atlantic deep-sea basin, is *Aequorea duplicata*. The distribution of this species is dealt with above (pp. 27-9); it occurs throughout the area, being particularly common some hundreds of metres below the surface, less frequent in immediate neighbourhood of the surface as well as in the very deep strata. *Aequorea homostoma* and *Solmaris incisa* are rare visitors from the south. The Narcomedusa *Solmaris corona* has only been found within the coastal areas of the British Isles.

Pequathia clara and *Egusa vitrea* belong to the upper strata of warmer seas; they are occasionally met with in the northern waters, where they seem to prefer the deeper strata. *Pequathia clara* is a predominantly West Atlantic form, and the free medusa has not been met with east of 30° W. in the northern section, whereas parasitic larvae, probably belonging to this species, were taken south-west of Iceland (see p. 33). *Egusa vitrea* occurs in the entire tropical and subtropical belt of the Atlantic Ocean and penetrates far towards the north in the north-eastern Atlantic, being recorded from deep water in localities not far from the south coast of Iceland (see p. 36).

Four species of *Larope* enter the northern seas as visitors from the warm portions of the Atlantic, but three of them have only been taken in the coastal areas: *L. tetraphylla* and *scutigera* along the American coast into the Gulf of Maine, *L. exigua* along the European coasts reaching as far as the English Channel as an occasional visitor. Only one species of this genus, *L. curybia*, has been found above deep water in the northern Atlantic; it is distributed right across the tropical Atlantic, and some few specimens were taken by the "Dana" (St. 1201 and 1203) in the Gulf-Stream water about midway between Newfoundland and Ireland (see p. 32). *Pelagia noctiluca*, which is widely distributed in the warm portions of the Atlantic, is frequently met with along the northern border of the Gulf Stream between the edge of the Newfoundland Bank and the British Isles, and in the north-eastern Atlantic it may even on rare occasions be carried as far north as 62°, not far from the south coast of Iceland, following the northernmost branch of the Gulf Stream (see p. 18).

Rhopalumma relatum has a similar distribution in the North Atlantic as *Pelagia noctiluca*, following the Gulf Stream towards the northern parts of the British Isles, but in the central portion of the North-Atlantic basin it is not restricted to the surface water, several specimens being taken even in the deepest hauls (with 1000-5000 m. wire out) at "Dana" St. 1201 (see p. 13).

3. Bathypelagic medusae. The following two species should be excluded from the discussion, because their distribution is imperfectly known: The Anthomedusa *Paraploca bathybia* was taken in deep water in the southern part of Davis Strait by the "Godthaab" expedition (KRUMH 1912)¹. The Scyphomedusa *Paralia rubescens* is known from deep water in a few scattered localities in the Pacific, Indian, and Atlantic Oceans, and one specimen was taken in 1910 by the "Michael Sars" about midway between Newfoundland and the British Channel.

The metaplanktonic medusae *Annatarea affinis*, *Pandea rubra*, *Bythotrephes mirruga*, *Taranna rotunda*, and *Otocanna funeraria* only seem to occur in the neighbourhood of the continental shelves, and probably their fixed hydroid stages live on the continental slopes, whence the free medusae are not likely to be carried much farther out into the deep-sea basins, the currents being slow in the deep and intermediate strata, where these species occur.

Most of the other deep-sea medusae of the northern Atlantic are known or supposed to be holoplanktonic (see p. 53), and the majority of them are generally distributed in the North-Atlantic deep-sea basin until the submarine ridges Scotland-Iceland-Greenland-Baffin Land, a few of them penetrating more or less north of the ridges.

3. I have found a very similar medusa, apparently the same species, in a sample of medusae taken by the "Dana" expedition 1930 near Cape of Good Hope in South Africa!

Two species have only been taken in the eastern portion of the area: the Scyphomedusa *Nausithoe globifera* occurs in the deep-sea basin east of the Mid-Atlantic ridge from about 45° N. until the channels west of Scotland, and in the present paper it is also recorded from deep water south of Iceland. The Trachymedusa *Rhopalonema funerarium* is widely distributed in the Tropics, and in the eastern Atlantic it occurs as far north as off the south-west coast of Ireland, where it has been taken in deep water; in the western part of the North Atlantic it was met with in a locality outside the Gulf of Maine, where it occurred in the Gulf-Stream water less than 300 m from the surface, but it has never been observed in the North-West Atlantic deep-sea.

Among the bathypelagic medusae, which are distributed throughout the deep-sea basins of the North Atlantic, the following species have a world-wide distribution in the great oceans: the Trachymedusae *Halireas minimum*, *Botrycnema brucei*, *Colobonema sericeum*, and *Pantachogon haeckeli*; the Narcomedusa *Eginaura grimaldii* (which however is rare in the South Atlantic), and the Scyphomedusae *Periphylla periphylla* and *Atolla wyvillei*. All these are found in the entire deep-sea basins in the northern Atlantic, with the exception of *Colobonema sericeum*, which has not been observed in the Labrador Sea and Davis Strait.

Three of these species may also be met with north of the submarine ridges. *Periphylla periphylla* rather frequently ascends towards the upper strata in colder seas and may therefore be carried across the ridges by the currents; but it avoids the very cold bodies of water and has never been taken in the deep, cold strata of the Baffin Bay and the Norwegian Sea; its distribution north of the ridges follows the Gulf Stream and its off-shoots to the west coast of Norway and the southern and western coasts of Iceland, and it is likewise carried northwards along the west coast of Greenland, but not very far, mainly following the comparatively warm water moving northwards outside the edges of the off-shore banks. *Pantachogon haeckeli* is more strictly confined to the deep and intermediate bodies of true Atlantic water, but occasionally it ascends into higher levels; the record of a specimen from Spitzbergen (MAAS 1904) is doubtful. West of Greenland some few specimens were taken by the "Godthaab" in deep water, about 800 and 1750 m, in Baffin Bay, about 70° N., at temperatures between 0.3 and : 0.4, probably carried directly across the ridge (which in its central part rises to about 700 m below the surface), afterwards sinking into the deep strata of Baffin Bay (KRAMP 1942 pp. 78 and 140). — *Atolla wyvillei* is a well marked bathypelagic species, at any rate in the northern seas. It has been found in five localities in the Norwegian Sea, preferably near the bottom at depths between 1600 and 2100 m and at temperatures about : 1°, and some young specimens, probably belonging to the same species, are recorded from deep water between north-east Greenland and Spitzbergen. The occurrence of this species in the deep-water of the Norwegian Sea can hardly be due to transportation by the currents across the Wyville Thomson Ridge; it must be indigenous in the Norwegian Sea. It has never been observed in Baffin Bay.

Whereas all these species have an almost cosmopolitan distribution in the deep parts of the great oceans, there are three bathypelagic species which are confined to the northern parts of

the Atlantic Ocean (partly found also in the Pacific). All of them are generally distributed in the deep-sea on both sides of the Mid-Atlantic ridge, their southern limit of distribution being at 30 or 40° N. Towards the north they all reach the continental slope south of Iceland and the submarine ridges between Scotland, Iceland, and East Greenland. *Haliscera hyslopae* has been found east of the Newfoundland Bank, but not in the Labrador Sea and Davis Strait; *Chromatonema cabeani* and *Crossota rafabrunnea* also occur in the deep part of Davis Strait south of the ridge between West Greenland and Baffin Land. The occurrence of these North-Atlantic species in the northern basins of the Atlantic is thus quite similar to that of the cosmopolitan species mentioned above. They are equally adapted to the conditions in these northern waters, and the reasons why the three last-mentioned species do not penetrate into the southern seas should be considered in connection with the question of the evolution of species within the various genera. Some genera, as e. g. *Atolla*, *Periphylla*, *Eginaura*, *Colobonema*, and *Halireas*, comprise only one species each, all of which are cosmopolitan; others, like *Pantachogon* and *Botrycnema*, contain one cosmopolitan species and one or more species of restricted distribution; *Haliscera*, *Crossota*, and *Chromatonema* are split up in a number of species, each with its particular area of distribution; *Haliscera hyslopae* and *Crossota rafabrunnea*, which are common in the northern Atlantic, also occur in the Pacific.

B. The arctic basins.

Very few medusae are indigenous in the deep basins of Baffin Bay and the Norwegian Sea. Some of the neritic species may occasionally drift out above the deep-sea areas; they will not be considered here. The Narcomedusa *Egimopsis laurenti* is a well-marked arctic species which prefers the coldest water; in Baffin Bay it is mainly found in the upper strata in the neighbourhood of the coasts, but in the Norwegian Sea it avoids the comparatively warm surface water and sinks into the cold, deep strata. *Aequorea digitata*, which occurs over the entire areas, is rare in the deep strata in Baffin Bay, whereas in the Norwegian Sea it may be taken in great abundance at least as far down as 1600 m below the surface (see p. 29). The Leptomedusa *Ptychopora hyperborea* has only been taken in Smith Sound; it is probably a bathypelagic species. *Sminthea arctica* is a doubtful species, taken in deep water west of Spitzbergen (HARTLAUB 1909). *Pantachogon haeckeli* is a stray Atlantic visitor in Baffin Bay (see above). The cosmopolitan deep-sea medusa *Atolla wyvillei* is indigenous in the deep, cold strata of the Norwegian Sea, but has not been found in Baffin Bay (see above).

There are, however, two decidedly arctic deep-sea medusae; one of them, *Botrycnema ellmanii*, is very abundant in Baffin Bay about 1000–1800 m below the surface at temperatures between 0° and : 0.4 and has also been taken in deep water between Spitzbergen and Greenland and north of Norway. Among the medusae this is the only species which confirms the supposed resemblance between the deep-sea faunas of Baffin Bay and the Norwegian Sea. In the deep, cold strata of the Norwegian Sea, 1000–2000 m below the surface, we also find *Crossota norvegica*, which has not been observed anywhere else.

1881. FEWKES, J. F. Studies of the Jelly-fishes of Narragansett Bay. Bull. Mus. Comp. Zool. Harvard Coll. Vol. VIII, no. 8.
1882. FEWKES, J. F. On the Acalophae of the East Coast of New England. Bull. Mus. Comp. Zool. Harvard Coll. Vol. IX.
1886. Report on the Medusae collected by the U.S.F.C. steamer Albatross in the region of the Gulf Stream, in 1883-84. Rep. U. S. Fish. Comm. for 1884.
1888. Medusae. The Lady Franklin Bay Expedition. Vol. 2, Appendix 132.
1889. Report on the Medusae collected by the U.S.F.C. steamer Albatross in the region of the Gulf Stream, in 1885-86. Rep. U. S. Fish. Comm. for 1886.
1848. FORBES, E. A Monograph of the British Naked-eyed Medusae. Ray Society.
1937. FROST, N. Further Plankton Investigations. Ann. Rep. Fishery Research Lab., Newfoundland, 1936-37.
1856. GEGENBAUR, C. Versuch eines Systemes der Medusen. . . Zeitschr. wiss. Zool. Bd. 8.
1898. GROENBERG, G. Die Hydromedusen des arktischen Gebiets. Zool. Jahrb. Abt. Syst. Bd. XI.
1903. GUNTHER, R. T. Report on the Coelenterata from the intermediate waters of the N. Atlantic. . . "Oceana" 1898. Ann. Mag. nat. Hist. Ser. 7, vol. XI.
1864. HAECKEL, E. Beschreibung neuer craspedoter Medusen aus dem Golfe von Nizza. Jenaische Zeitschr. für Naturwiss. Bd. I.
- 1879-80. Das System der Medusen.
1881. Report on the deep-sea Medusae. . . Rep. Sci. Res. H. M. S. Challenger. Zool. Vol. I.
1936. HARDY, A. C. The Arctic Plankton collected by the Nautilus Expedition 1931. I. General Account. Journ. Linn. Soc. London, Vol. 39, Zool.
1902. HARGETT, C. W. Notes on a few Medusae new to Woods Hole. Biol. Bull. Vol. I.
1904. The Medusae of the Woods Hole Region. Bull. Bureau of Fisheries, Vol. 24.
1909. HARTLAUB, C. Méduses. Duc d'Orléans, Croisière Océanogr., "Belgica" dans la Mer du Groenland 1906.
1926. HAYNO, E. J. Periphylla hyacinthina. Naturen. Bd. 50.
1927. JASCHNOV, W. A. Das Zooplankton des Kaspischen Meeres. Ber. wiss. Meeresinst. Moscow, T. 2.
1939. Hydroméduses de l'Océan Glacial. Bull. Soc. des naturalistes de Moscou, Sec. Biol., N. S. Tome 48.
1923. JESPERSEN, P. Dr. Thorild Wulff's Plankton-Collections in the waters west of Greenland. II. Thule Exped. Meddel. om Grønland, Bd. 61.
1861. KEFERSTEIN, W. & E. EHLERS. Zoologische Beiträge gesammelt im Winter 1859-1860 in Neapel und Messina. Leipzig.
- 1913a. KRAMP, P. L. Medusae collected by the "Tjalff" Expedition. Vidensk. Meddel. dansk naturhist. Foren. Bd. 65.
- 1913b. Coelenterata. Résumé Planktonique III. Bull. trimestriel etc., Bureau du Conseil internat. pour l'exploration de la mer.
1914. Meduser og Siphonophorer. Conspectus Faunae Groenlandicae. Meddel. om Grønland, Bd. 23.
1915. Medusae, Ctenophora, and Chaetognathi from the Great Belt and the Kattegat 1909. Meddel. Komm. f. Havundersog. Ser. Plankton. Bd. I, No. 12.
1919. Medusae I. Leptomedusae. The Danish Ingolf Exped. Vol. V, Part 8.
1920. List of Medusae collected by the M.S. "Armauer Hansen" in the North Atlantic in 1913. Bergens Museums Aarbok 1917-18, 2. Hefte, nat. R. 8.
1921. Medusae. Rep. Danish Oceanogr. Exped., 1908-1910 to the Mediterranean and adjacent Seas. Vol. II, H. I.
- 1926a. Medusae II. Anthomedusae. The Danish Ingolf Exped. Vol. V, Part 10.
- 1926b. KRAMP, P. L. Occasional Note on Coelenterata. I. Vidensk. Meddel. dansk naturhist. Foren. Bd. 82.
1927. The Hydromedusae of the Danish Waters. Kgl. danske Vidensk. Selsk. Skrifter, nat. nat. Afd. R. S. Bd. XII, I.
1930. Hydromedusae collected in the south-western part of the North Sea and in the eastern part of the Channel in 1903-1911. Mem. no. 45 du Musée R. d'hist. nat. de Belgique.
- 1933a. Occasional Notes on Coelenterata. II. Vidensk. Meddel. dansk naturhist. Foren. Bd. 94.
- 1933b. Leptomedusae. Nordisches Plankton. Bd. XII, Teil 3.
- 1933c. Coelenterata, Ctenophora, and Chaetognatha. The Scoresby Sound Comm. 2nd East Greenland Exped. in 1932. Meddel. om Grønland, Bd. 104.
1936. On the Leptomedusae of the Genera Eirene Eschsch. and Heliciritha Hartl. Vidensk. Meddel. dansk naturhist. Foren. Bd. 99.
1937. Polypdyr II. Gopler. Danmarks Fauna, Bd. 43.
1939. Medusae, Siphonophora, and Ctenophora. The Zoology of Iceland, Vol. II, 5 b.
1912. Medusae. The "Godthaab" Expedition 1928. Meddel. om Grønland, Bd. 81.
1913. Medusae, Siphonophora, and Ctenophora. Zoology of East Greenland. Meddel. om Grønland, Bd. 121.
1925. KRAMP, P. L. & D. DAMAS. Les Méduses de la Norvège. Introduction et Partie spéciale I. Vidensk. Meddel. dansk naturhist. Foren. Bd. 80.
1935. KUNNE, C. DE W. MUELER & C. KUNNE. Fischbrut- und Plankton-Untersuchungen auf dem . . . "Poseidon" in der Ostsee, Mai-Juni 1931. Wiss. Meeresunters. Abt. Helgoland, N. F. Bd. 19.
- 1937a. Über als "Freundlinge" zu bezeichnende Grossplanktonen in der Ostsee. Rapports et Procès-Verbaux . . . Conseil internat. . . Vol. 102.
- 1937b. Über die Verbreitung der Leitformen des Grossplanktons in der südlichen Nordsee im Winter. Ber. d. Deutschen Wiss. Komm. für Meeresforsch. N. F. Bd. 8.
1922. LEROUX, M. V. The Food of Plankton Organisms I. Journ. Mar. Biol. Assoc. Plymouth, Vol. XII.
1843. LESSON, R. Histoire naturelle des Zoophytes, Acalophes.
1893. LEVINSSEN, G. M. R. Meduser, Ctenophorer og Hydroder fra Grønlands Vestkyst. Vidensk. Meddel. naturhist. Foren. i Kjøbenhavn for 1892.
1900. LIXKO, A. Bericht über Medusen und Ctenophoren aus den Eismeer. Travaux Soc. Imp. des Naturalistes de St. Petersbourg. T. 31.
- 1904a. Planktonliste des Barents Meeres. Exped. für wissenschaftl. prakt. Unters. an der Armauer Küste. Comite für Unterstützung der Küsten-Bevölkerung des Russ. Nordens.
- 1904b. Zoologische Studien im Barents-Meer. Zool. Anzeiger. Bd. 28.
1913. Zooplankton de la Mer Glaciale de Sibirie d'après les récoltes de l'Exped. Polaire Russe en 1900-1903. Mem. Acad. sci. St. Petersb. T. 29, 1-3.
1875. LAUREN, C. A revised list of the Acalophae and Hydrozoa of Greenland. Arctic Manual and Instructions. Copenhagen.
1893. MAVAS, O. Die craspedoten Meduser der Plankton Expedition. Ergebnisse d. Plankton Exped. Bd. II, K.
1897. Die Medusen. Rep. Explor. West Coast of Mexico. Albatross. Mem. Mus. Comp. Zool. Harvard Coll. Vol. 23, No. 1.
1901. Meduses provenant de campagnes de Yacht "Hirondelle" et "Princesse Alice". Resultats de campagnes scientifiques. Prince de Monaco. Fasc. 28.

PLATES

Plate 1.

Figs. 1-4 *Photomastax palaris* Allman.

- Fig. 1. Tentacles with adhesive disk. — Figs. 1-2. $\times 45$, fig. 3. $\times 120$.
Fig. 4. Optical section of filiform tentacle. — $\times 70$.

Figs. 5-8 *Halosoma baylouni* n. sp.

- Fig. 5. Male individual. — $\times 6$.
Fig. 6. Stomach, showing the circular mouth opening; one of the radial canals with female gonad.
Fig. 7. Part of umbrella margin, *c, c.* circular vessel, *n, n.* nerve ring, *r, c.* radial canal, *v.* velum. — $\times 49$.
Fig. 8. Distal end of a juvenile tentacle. — $\times 300$.

Fig. 9 *Balanus beana* Browne.

- Fig. 9. Young tentacle: *a* near base, *b* middle portion, *c* distal part. — $\times 137$.

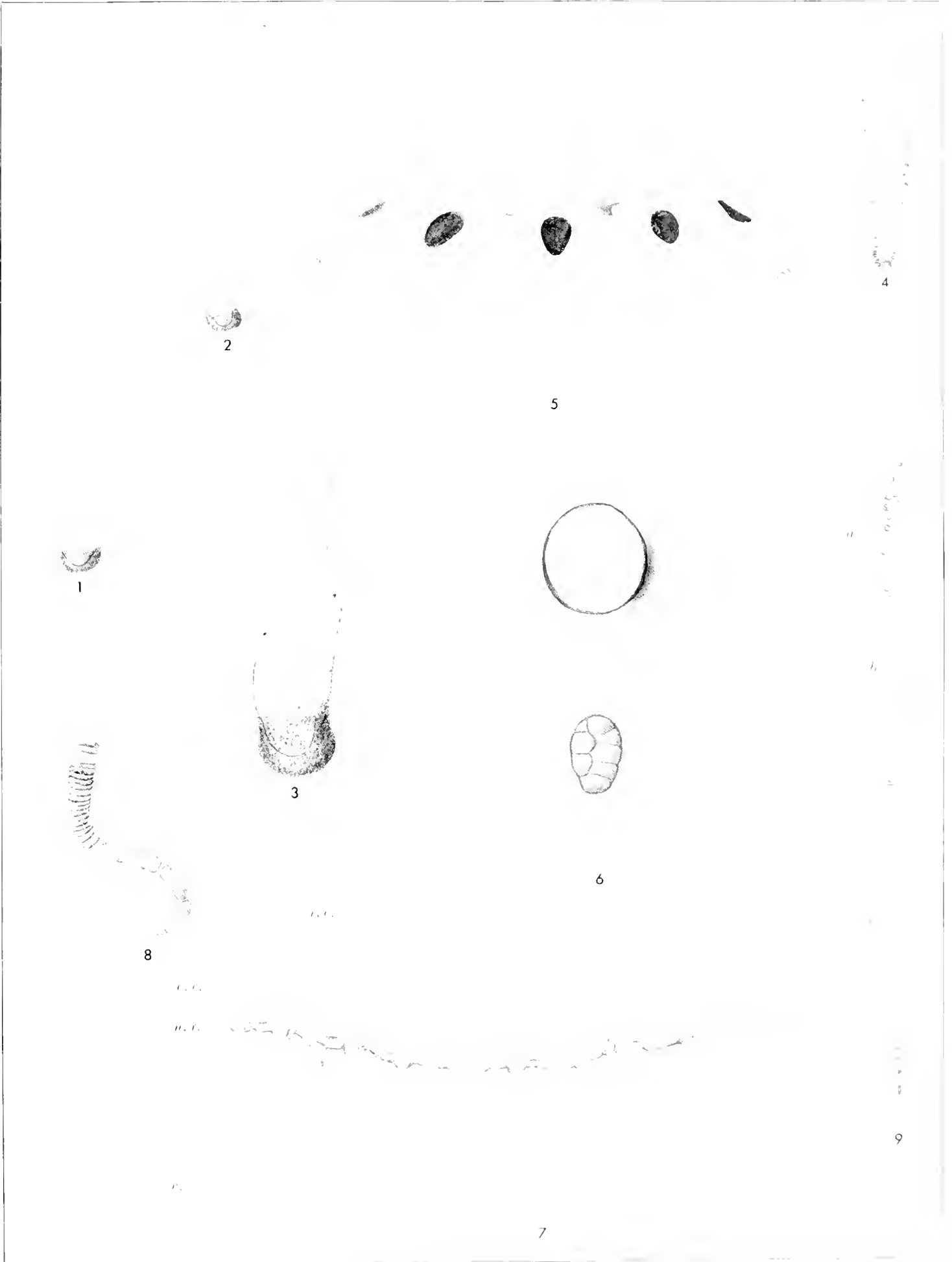


Plate II.

- Figs. 1-2. Statoocysts of *Halioreca haphanta* n. sp. 180
- Fig. 3. Statoocyst of *Balanocystis brucei* Browne. 180
- Figs. 4-5. *Rhopilema tantulum* Vauhoffen.
- Fig. 4. Manubrium, Specimen from "Thor" stat. 181(99).
- Fig. 5. Interradial currus. 225
- Fig. 6. *Hymenocystis platygonum* Browne, stomach and male gonads, seen from the exumbrella. 70
- Figs. 7-8. Statoocysts of *Pantachocopa hutchinsii* Maas. 675
- Figs. 9-10. *Crossota rotunda* Kramp.
- Fig. 9. Transverse section of radial canal, *ex.* exumbrella, *end.* endoderm lamella, *mc.* circular muscle fibres, *sub.* subumbrella. 70
- Fig. 10. Part of radial canal with its surroundings. On the right-hand side part of the ectodermal muscular epithelium is removed, disclosing the cells of the endoderm lamella; through an opening in the subumbrella wall of the radial canal the pigmented cells of its exumbrella wall are seen. For further explanation, see the text, p. 24. 135



P
K
L
C
1

2
3
4
5
6
7
8
9
10

11
12
13
14
15
16
17
18
19
20



end. l.
m.
sub.



Plate III.

Cassida catabannica Kramp

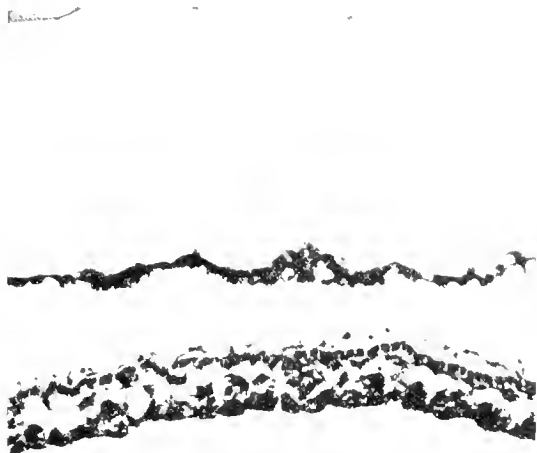
- Fig. 1. Manubrium, external view.
- Fig. 2. Manubrium, internal view, showing the two whorls of invaginated pouches and one of the longitudinal grooves; *g*, gonad.
- Fig. 3. Transverse section of one corner of the mouth tube, showing the string of large, vacuolated endoderm cells.
- Fig. 4. Transverse section of proximal part of the stomach, through the upper whorl of invaginated pouches (cf. textfig. 9, p. 25).
- Figs. 5-6. Longitudinal sections of stomach; for further explanation, see p. 24.
- Fig. 7. Longitudinal section of radial canal; *ex*, exumbrella, *mc*, circular muscle fibres, *p*, pigmented endodermal epithelium, *np*, non-pigmented endodermal epithelium of radial canal, *sub*, subumbrella.
- Fig. 8. Meridional section of umbrella; *end*, *l*, endoderm lamella, *mc*, layer of circular muscle fibres, *sub*, subumbrella.



1



2



7



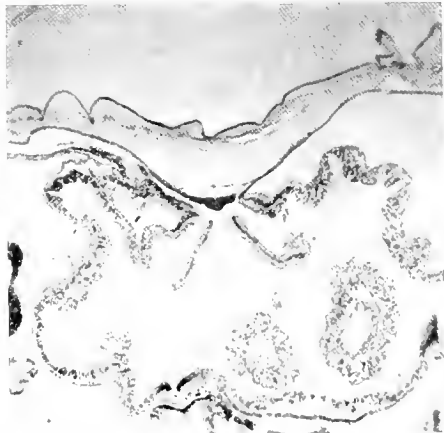
3



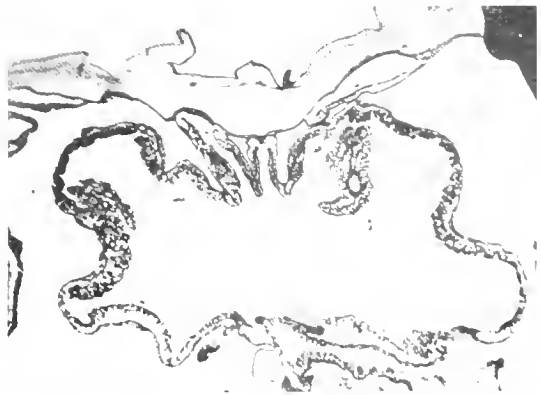
8



4



5



6

ca.

ps.

m. p.

m.

sub.

end. l.

m.

sub.

Plate IV.

Figs. 1-4. *Cassata catobannata* Kraup.

- Fig. 1. Umbrella margin, abaxial aspect.
Fig. 2. Umbrella margin, adaxial aspect (velum removed): *c. v.*, circular vessel; *n. r.*, nerve ring; *sub.*, subumbrella.
Figs. 3-4. Statocysts, longitudinal sections. $\times 500$.

Figs. 5-6. *Cassata papuana* Vanhoffen.

- Fig. 5. Specimen from "Ingolf" stat. 129, $\times 4$.
Fig. 6. Mammum of a specimen with 11 radial canals, "Ingolf" stat. 129.

Fig. 7. *Papudha elata* R. P. Bigelow.

- Fig. 7. Larva in the gastric cavity of *Periphalia periphalia*, "Godthaab" stat. 1, $\times 4$.



3



4



5



6

sub.

c. l.

n. l.



1

2



7

Plate V.

*Larvae of *Pogonotha clara* R. P. Bigelow*

- FIGS. 1-2. Primary polyps (I) with actinula buds of successive stages of development (II and III).
- FIG. 3. Chain of actinula buds in successive stages (I-IV).
- FIG. 4. Young actinula with two well-developed and two small tentacles; without statocysts, and without aboral collar.
- FIGS. 5-6. Two actinulae, each with four tentacles almost equally developed, and with 20 statocysts on the rim of the aboral collar.
- FIG. 7. Part of aboral surface of actinula, showing otoporpe and peronai tentacles cut off at their bases.
- FIGS. 8-9. Aboral view of medusa, showing the four tentacles and the long otoporpe.
- FIG. 10. Oral view of medusa, showing four peronai, twenty statocysts, and the first trace of mouth opening.

All figures $\times 40$.



Plate VI.

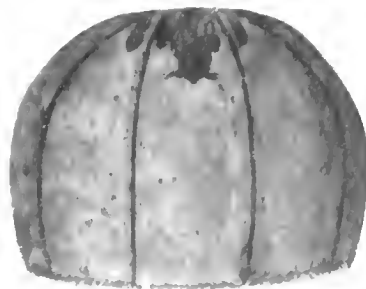
- Figs. 1-2. *Prorhynchus pectus* Allman. Coloured sketches made on board the "Ingolf" stat. 126, by C. E. WARDL.
- Figs. 3. *Heterocis acuminata* Fewkes.
- Figs. 4. *Balanocystis hawaii* Brown.
- Figs. 5. *Cassida californiana* Klamp.
- Figs. 6. *Aphanta digitata* (O. F. Muller).
- Figs. 7. *Chromatomyia rubra* Fewkes.
- Figs. 3-7 are reproductions of coloured sketches drawn from life by the author on board the "Godthaab" in Davis Strait 1928.



1



2



5

4



7

6

THE INGOLF-EXPEDITION

1895-1896

THE LOCALITIES, DEPTHS, AND BOTTOMTEMPERATURES OF THE STATIONS

Sta- tion Nr.	Date	Lat. N.	Long W.	Depth in m	Bot- tom- temp.	Sta- tion Nr.	Date	Lat. N.	Long W.	Depth in m	Bot- tom- temp.	Sta- tion Nr.	Date	Lat. N.	Long W.	Depth in m	Bot- tom- temp.
1895						1896											
1	11-V	62° 30'	8° 21'	249	7.2	24	25-VI	63° 06'	56° 00'	2258	2.4	45	11-V	61° 32'	9° 43'	1211	1.15
2	12 -	63° 04'	9° 22'	493	5.3	25	26 -	63° 30'	54° 25'	1096	3.3	46	- -	61° 32'	11° 36'	1356	2.49
3	- -	63° 35'	10° 21'	512	0.5			63° 51'	53° 03'	256		47	12 -	61° 32'	13° 40'	1789	3.23
4	13 -	61° 07'	11° 12'	446	2.5	26	- -	63° 57'	52° 41'	61	0.6	48	- -	61° 32'	15° 11'	2165	3.17
5	- -	61° 40'	12° 09'	292				64° 37'	51° 21'	205		49	13 -	62° 07'	15° 07'	2109	2.91
6	16 -	63° 43'	14° 34'	170	7.0	27	1-VII	64° 54'	55° 10'	740	3.8	50	- -	62° 43'	15° 07'	1921	3.13
7	17 -	63° 13'	15° 11'	1130	4.5	28	- -	65° 14'	55° 42'	791	3.5	51	15 -	61° 15'	14° 22'	128	7.32
8	19 -	63° 56'	24° 40'	256	6.0	29	5 -	65° 34'	54° 34'	128	0.2	52	- -	63° 57'	13° 32'	791	7.87
9	20 -	61° 18'	27° 00'	555	5.8	30	10 -	66° 50'	51° 28'	11	1.05	53	16 -	63° 15'	15° 07'	1497	3.08
10	- -	64° 24'	28° 50'	1484	3.5	31	11 -	66° 35'	55° 54'	166	1.6	54	18 -	63° 08'	15° 10'	1301	3.9
11	21 -	64° 34'	31° 12'	2448	1.6	32	11 -	66° 35'	56° 38'	599	3.9	55	19 -	63° 33'	15° 02'	595	5.9
12	22 -	64° 38'	32° 37'	1958	0.3	33	12 -	67° 57'	55° 30'	66	0.8	56	- -	64° 00'	15° 09'	128	7.57
13	- -	64° 17'	34° 33'	1171	3.0	34	18 -	65° 17'	54° 17'	101		57	20 -	63° 37'	13° 02'	659	3.4
14	- -	64° 15'	35° 05'	331	1.4	35	- -	65° 16'	55° 05'	682	3.6	58	- -	64° 25'	12° 09'	397	0.8
15	4-VI	66° 18'	25° 59'	621	0.75	36	28 -	64° 50'	56° 21'	2702	1.5	59	- -	65° 00'	11° 16'	581	0.1
16	5 -	65° 43'	26° 58'	171	6.1	37	29 -	60° 17'	54° 05'	3229	1.1	60	21 -	65° 09'	12° 24'	744	0.9
17	16 -	62° 19'	26° 55'	1403	3.4	38	30 -	59° 12'	54° 05'	3521	4.3	61	- -	65° 03'	13° 06'	194	0.4
18	17 -	61° 41'	30° 29'	2437	3.0	39	9-VIII	62° 00'	22° 38'	1629	2.9	62	31 -	63° 48'	19° 12'	136	1.92
19	18 -	60° 29'	34° 11'	2949	2.4	40	- -	62° 00'	21° 36'	1591	3.3	63	1-VI	62° 40'	19° 05'	1506	4.0
20	20 -	58° 20'	40° 48'	3192	1.5	41	12 -	61° 39'	17° 10'	2315	2.0	64	- -	62° 06'	19° 00'	1960	3.1
21	21 -	58° 01'	44° 45'	2505	2.4	42	11 -	61° 41'	19° 17'	1477	0.4	65	2 -	61° 33'	19° 00'	564	5.9
22	22 -	58° 10'	48° 25'	3471	1.1	43	- -	61° 42'	19° 11'	1215	0.05	66		61° 34'	20° 45'	944	3.3
23	24 -	60° 43'	56° 00'	2021		44	- -	61° 42'	9° 36'	1026	1.8	67	3 -	61° 39'	21° 33'	846	3.0

Stn. No.	Lat. N	Long. W	Depth in m.	Bottom temp.	Station No.	Date	Lat. N	Long. W	Depth in m.	Bottom temp.
187	64 41	32 52	1838	1.4	118	24-VII	68 27'	8 20'	1996	1.0
188	64 41	33 14	1444	1.6	119	25 -	67 53'	10 49'	1902	1.0
189	64 53	36 19'	384	4.1	120	- -	67 29'	14 32'	1636	1.0
190	64 46	36 45'	401	-	121	- -	66 59'	13 41'	996	0.7
191	64 42	36 34	1416	2.1	122	26 -	66 42'	14 44'	217	1.8
192	64 58	36 28	1384	1.2	123	28 -	66 52'	15 40'	273	2.0
193	64 45	34 36	847	5.5	124	- -	67 40'	15 40'	932	0.6
194	64 57	26 55	260	5.9	125	29 -	68 08'	16 02'	1373	0.8
195	64 28	25 00'	352	6.1	126	- -	67 19'	15 52'	552	0.5
196	64 28	26 25	111	0.4	127	2-VIII	66 33'	20 05'	83	5.6
197	66 50	26 50'	1011	0.7	128	- -	66 50'	20 02'	365	0.6
198	66 46	26 59	1412	0.9	129	3 -	66 35'	23 47'	220	6.5
199	66 57	27 52'	1090	0.6	130	8 -	63 00'	20 40'	636	6.55
200	66 52	28 58	1802	1.1	131	- -	63 00'	19 09'	1314	4.7
201	64 02	29 32'	1435	0.8	132	- -	63 00'	17 04'	1407	1.6
202	64 44	27 00'	842	0.6	133	9 -	63 14'	14 24'	433	2.2
203	64 55	27 28'	878	-	134	- -	62 34'	10 26'	563	4.1
204	64 25	28 50'	926	0.3	135	10 -	62 48'	9 48'	508	0.4
205	62 56	26 01'	183	1.1	136	- -	63 01'	9 11'	482	4.8
206	62 56	25 30'	72	1.5	137	- -	63 14'	8 31'	559	0.6
207	62 58	25 24'	1471	0.8	138	- -	63 26'	7 56'	887	0.6
208	64 21	25 21'	1619	0.9	139	- -	63 36'	7 30'	1322	0.6
209	64 04.6	24 47.6	2386	1.1	140	11 -	63 29'	6 57'	1469	0.9
210	64 32.4	24 56.2	2465	1.0	141	- -	63 22'	6 58'	1279	0.6
211	64 58	24 25'	1456	1.0	142	- -	63 07'	7 05'	1105	0.6
212	64 47	27 26	162	0.4	143	- -	62 58'	7 09'	731	0.4
213	64 45	29 06	699	0.4	144	- -	62 49'	7 42'	520	1.6
214	64 44	31 09	1889	1.0	-	-	-	-	-	-

DL Danish Ingolf-Expedition,
5 1895-1896
D3 The Danish Ingolf-
v.5D expedition
nt.12-14

50M4d

PLEASE DO NOT REMOVE
CARDS OR SLIPS FROM THIS POCKET

UNIVERSITY OF TORONTO LIBRARY

