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CORNELL UNIVERSITY AGRICULTURAL EXPERIMENT STATION

THE LEPIDOPTERA OF NEW YORK AND NEIGHBORING STATES

Primitive Forms Microlepidoptera Pyraloids Bombyces

WILLIAM T. M. FORBES

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PREFACE

The work which follows is intended primarily to make easier the recognition of the Lepidoptera of the State of New York, and to assemble so far as possible the known data on their life histories, whether published or in the form of notes associated with the various collections. The work is, in a sense, a compilation, but largely of matter never before published. The keys, in particular, are for the most part a new venture, since for many groups of American Lepidoptera none have ever before been worked out; even the published keys to European forms are in need of revision. I cannot hope that my keys are perfect, but rather that, as a contribution in a new field, they may serve as a basis on which to build in the search for further knowledge.

The present contribution includes about half — the more primitive half — of the Lepidoptera. I have accumulated some data on the remainder, but it may be long before my notes are ready to publish.

My indebtedness is so general as to make a detailed acknowledgment impossible. Every one I have approached has helped me, and this means almost every Lepidopterist and the authorities of every considerable museum in the eastern United States. My first and greatest debt has been to the late Dr. John B. Smith, who, at Rutgers, encouraged me to begin such a piece of work, and gave me a great deal of valuable counsel. The signed contributions of Dr. Annette F. Braun and that of Mr. Carl Heinrich speak for themselves. No one else could have worked up the Nepticulidæ, the Coleophoridæ, and Lithocolletis. I have also been in almost continual consultation with them concerning many other groups where their names do not appear. I have consulted Mr. August Busck on every single group of the Tineids, and in most cases have followed his advice. I made the final draft of this memoir and prepared the drawings in the laboratories of Cornell University in 1919. I have incorporated a considerable part of the data published since that date, and have completely rewritten the section dealing with the Olethreutinæ and the Pterophoridæ; but it has not been possible to make the additions complete, nor to verify the New York records and adjust them to the new work. In the most important cases these have been noted in the text.

WILLIAM T. M. FORBES

Ithaca, New York June, 1923.

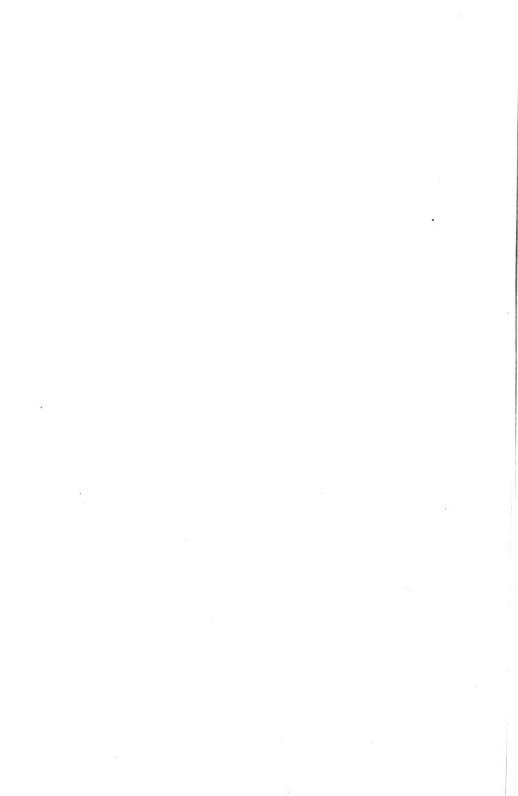


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LEPIDOPTERA OF NEW YORK AND NEIGHBORING STATES WILLIAM T. M. FORBES

The primary purpose of this memoir is to describe the kinds of Lepidoptera, or butterflies and moths, which occur in New York, and those which, though they have not as yet been taken in the State, belong to the same fauna, as many of these will sooner or later be found within And a second purpose, of nearly equal if not of greater importance, it. is to indicate, as briefly as practicable, the main facts in the life history of each. Though monographs of certain isolated groups are available, there is no work dealing comprehensively with the American forms of this interesting and economically most important order.

As a measure of economy, colored figures, bibliography, and ref-These appear unnecessary since W. J. Holland, erences are omitted. in The Moth Book¹, provides colored figures of a large proportion of our more conspicuous species; while Barnes and McDunnough have a full bibliography nearly ready for the press, and Dyar's $List^2$, already published, furnishes a convenient index to the literature.

Every part of this memoir has been made as concise as possible without sacrifice of clearness. For this reason two types of keys are used. The one in which the categories are indicated by successive indentation is most convenient for quick reference, and is employed wherever practicable. In the case of very bulky keys, however, such as the keys to the families, and the one to the species of Tortricidæ resembling Eucosma, a more compact arrangement has proved necessary. Descriptions of genera and species have been reduced to skeletons, in the manner customary in zoological publications. Dimensions are given in millimeters: this is the more convenient unit in the case of the small forms; in the larger it is easily remembered that 25 millimeters equals 1 inch. Dates and distributions are approximate only, but as accurate as can be given with the material at hand.

As to nomenclature, the author has followed the tradition of usage more closely than any particular code, though such rules of usage as are almost universally accepted have been followed even to the upsetting of a few familiar names. In any disputed case the more familiar name is used. Other names which have gained currency are noted in parentheses. Names of varieties and races, so far as they appear to indicate a real difference, are noted, the more important ones in separate paragraphs, though without a number, the less important by casual phrases in the description of the species to which they belong.

¹ Holland, W. J. The Moth Book, p. 1-479. 1903. ² Dyar, Harrison G. A list of North American Lepidoptera and key to the literature of this order of insects. United States National Museum. Bul. 52: 1-723. 1902.

TAXONOMY

This is not the place for a monograph on the laws of nature, but the following outline will make clearer the writer's point of view as to the status of the forms discussed.

A species is considered to be: A group of individuals separated from all others by tangible characters, breeding freely among themselves, but not with other individuals. Fifty years ago this would have been sufficient. Now that the continuity of evolution has become obvious, we must recall that at some time any given species was coming into being, was being set off from its relatives, and was acquiring its characteristic property of not interbreeding with them. During such a period, which may conceivably have been long or short, the species would be imperfectly defined; individuals would interbreed with their cousins, but more and more rarely, and with imperfect fertility; and the various strains would be acquiring adaptations to new environments or climates, which would reduce the probability of internating. And along with all this, differences of structure would be developing, distinguishing them to the human eye. In fact, many groups are now in this intermediate state, as witness the asters and the violets in the plant kingdom, and the Apanteses and the Euxoas among the Lepidoptera. Further, it is often, and in the Lepidoptera usually, unknown, to what point this isolation has reached; so we must use our best judgment in deciding what is a species and what a mere variety in any given case.

In order to make clearer the relationships of animals, species are grouped in a series of successively larger categories: genus, family, order, class, and phylum (subkingdom), within the animal kingdom. Besides these, in groups as large and as complexly related as the insects, intermediate groupings are employed: subgenus for a group of species within a genus; subfamily, suborder, and subclass similarly. Superfamily is used to designate a group of families³, and tribe for a group of closely related genera. The values of these categories are really arbitrary, and there has been a continual tendency to split groups; in fact, the superfamily of the present day is smaller than the genus of Linnæus (1758). As to the genus, however, we have an imperfect criterion. Member of a single genus (on the average) produce hybrids, but sterile or of low fertility; members of different genera do not. Even here we find Nature draws no sharp lines, for, in isolated favorable cases, hybrids have been produced between widely different animals, though such always seem destined to an early death, in an embryonic stage.

In the course of the splitting of groups just mentioned, it is the practice to preserve the original name for one of the sections into

Some early authors call this a tribe.

which an earlier category is divided. In the case of a tribe or larger group, whose name is derived from that of a genus, the subdivision in which that genus remains holds its name. When, however, a genus is divided, a species typical of the genus is taken (the type), which shall always be included in the genus retaining the old name. In many cases the proposer of a genus has designated the type; in others, the selection has been left to a later worker, and there is a good deal of confusion as to its selection. This confusion has been the basis of most of the changes of name of familiar insects with which the worker on Lepidoptera is cursed.

In the present memoir we are dealing with part of a single order, the Lepidoptera, belonging to the class Insecta. The suborders, superfamilies, families, subfamilies, tribes, genera, subgenera, and species are all systematically arranged in the text, and need no further explanation.

VARIATION

A species is, next to an individual, the most fundamental unit of living nature; but species themselves are not homogeneous. For intercrossing, which tends to bring things to an average, is offset by variation, which is continually producing new forms.

Although not as a rule striking, and often ill-defined, the most fundamental subdivision of a species is the race (also called subspecies, or variety, as that word is used by many German and English workers); these are forms isolated by difference of locality or habitat. Races are more or less clearly defined according to the amount of intercrossing between them; and this, in its turn, is a function of both the distances involved and the mobility of individuals. Some stocks, also, appear to be inherently more variable than others. So an active species, like Vanessa cardui, has few races, though its distribution is almost worldwide; while in the case of some Lycenas and Haploas, each patch of ground has its local form — a little race — distinct from those around Such minute races are known as strains. In general, races are it. distinguished from each other by a large number of minute characters, none of which are wholly constant, so that occasionally there will be found, in any region, forms that belong rather to some other race. Races breed true, at least so long as their environment remains constant.

Another type of variation is *seasonal*. Many species have more than one generation a year; and in that case specimens from one season are often recognizably, and sometimes strikingly, different from those of another. This variation is usually a response to the weather, wholly or in part, so that it is not very rare for a form normal, perhaps, in the spring, to appear in a cold summer, or in a cold breeding cage, or even erratically if the individual larva has grown up and trans-

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formed in abnormal surroundings. In the period of change from larva to pupa, specimens have been found to be most sensitive to external conditions, but the effect may be perceptible throughout larval life.

Thirdly come *food-varieties*, which are, perhaps, sometimes of the same nature as strains, sometimes as seasonal forms, but associated with differences in larval food. Aside from the minor cases of stunting from unfavorable food, and of exuberant growth with favorable, there is at least one case, that of *Halysidota*⁴ tessellaris, where definite forms are associated with different food plants.

Mendelian forms, like races, breed true so long as pure. They do not need to be segregated locally, but act as units in inheritance. Some are dominant, their characteristics appearing in every individual that earries the factor that causes them; others are recessive, only appearing in those individuals that happen to inherit the proper factor from both parents. The latter, of course, may be covered up for a series of generations, and then suddenly appear as the result of a favorable mating. The details of this type of inheritance are exceedingly complicated, and are explained in several recent textbooks on heredity. Striking cases of inheritance of this type exist in the genus Apantesis. In A. nais it involved the black costal edge, the black-spotted collar, the black face, the red hind wings, and some other characters, each inherited independently of the others. Such Mendelian characters are supposed to have arisen by an abrupt change, or *mutation*, in the germ of some individual. Many cases have been studied in the Lepidoptera. Mendelian forms, as well as minor races, are often called strains on account of their tendency to breed true.

Many variations, lastly, are produced wholly or largely by external causes; and we are ignorant of the true nature of many. These are generally known as *varieties* ("aberrations" of Staudinger, Jordan, and their followers), or, if rare and widely different from the normal form, as *aberrations*.

Monstrosities are deviations in structure from the norm of a species. They may be slight, or so radical as to be incapable of continued life; and may be a result either of an inherited factor, or of some external influence in an early stage. Mere mutilations are not considered monstrosities.

It is customary to use as the name of a species its genus and species names. The former is a noun in the singular, and of Latin form; the latter, when an adjective, is made to agree with it in gender. All names of genera are written with a capital, and in entomology it is customary nowadays to write those of all species with a small letter. To

[&]quot;This is Hübner's original spelling of the name, justified by its derivation from the Greek.

this combination the name of the author who first described or figured the species is often added. For a *race*, its name (with that of the author if desired) is added after the species name; and for a seasonal form, or named variety, or aberration, or Mendelian form, its name is added, preferably prefixing the notation variety (abbreviated var. or v.), *aberration* (*ab.*), or *form* (*f.*), to distinguish it from a race.

As an example, to give fully the name of the early spring form of the Azure Blue we write Lycana argiolus pseudargiolus f. lucia Kirby.

There is often uncertainty as to whether a given form is a species or a race. I have listed such forms according to what appears to me their most probable status, sometimes noting the uncertainty.

The following striking cases of variation are listed from the Lepidoptera of our fauna:

Races (Local forms):

Argynnis aphrodite (Atlantic), alcestis (Mississippi Valley), and cipris (Rocky Mts.).

Basilarchia astyanax (Atlantic), arthemis (Northeastern), rubrofasciata (Northern Rockies), and arizonensis (Arizona).

Cercyonis pegala (Gulf strip), alope (Eastern), nephele (Northeastern), and olympus (Mississippi Valley).

Lycana argiolus (Europe), pseudargiolus (general eastward), nigra (Appalachians), etc.

Lycana couperi (Northern), and lygdamas (Appalachians).

Pholus satellitia (Antilles), and pandorus (Northeastern).

Hemaris thysbe (Northern), and floridensis (Southern).

Zale lunifera (Atlantic), and lineosa (Mississippi Valley).

Automeris io (Northern), and lilith (Florida).

Eacles imperialis (Northern), and nobilis (Texas).

Seasonal forms:

Papilio marcellus (early spring), telamonides (late spring) and lecontei (summer).

Phyciodes that (summer), and marcia (spring).

Polygonia comma (winter), and dryas (summer).

Polygonia interrogationis (winter), and umbrosa (summer): these two imperfectly seasonal.

Hemaris diffinis (summer), and tenuis (spring).

Lycana pseudargiolus lucia (partly local), marginata, violacea, pseudargiolus.

Eutrapela kentaria (spring), and glaucaria (summer).

Eutrapela alciphearia (spring), and ornata (summer).

Peronea minuta (summer), and cinderella (winter).

Sexual forms (a few striking examples):

Argynnis diana (male red, female blue).

Argynnis idalia (male with red marginal spots, female with white).

Chrysophanus thoë (male purple, female red and black).

Lycana pseudargiolus (female only with black border).

Lycana comyntas and scudderi (male blue, female blaek).

Pieris rapæ (female with an additional spot).

Eurymus philodice and eurytheme (female with spots in border).

Pamphila brettus and zabulon (male tawny, female black).

Callosamia promethea (male black and diurnal, female red and noeturnal).

Automeris io (male yellow, female dark).

Eacles imperialis (male with red patches, absent in female).

Estigmene acraa (male with yellow hind wings, female white).

Apantesis phalerata (male with pale hind wings, female with red).

Alypia langtonii (eastern male with two white spots on hind wing, female always with one yellow).

Tarache terminimacula (pattern different in sexes).

Biston quernaria (male dark and short-winged, female light and longwinged).

Erannis tiliaria (male yellow and brown, female black and white, and wingless).

Nymphula maculalis (male black and white, female normally gray). Nymphula semincalis (male with white submarginal band).

Acrolophus mora (male blackish, female brown).

Prionoxystus robiniæ (male with yellow hind wing, female gray).

Prionoxystus macmurtrei (female many times larger than male).

Dimorphism not wholly sexual (so far as known, Mendelian):

Eurymus species (white and yellow females).

Eurymus eurytheme (both sexes yellow or orange).

Pieris rapæ (white), and novaangliæ (yellow male).

Papilio turnus (yellow), and glaucus (southern black female).

Pamphila hobomok (tawny), and pocahontas (black female).

Pamphila massasoit (yellow below), and suffusa (dark below).

- Apantesis nais (red and yellow females); also many less striking cases in the genus.
- Catocala palæogama and other species of Catocala (black-banded forms).

Deilinea liberaria (tawny), and helena (brown-banded).

Aplodes brunnearia and bistriaria (green and brown forms).

Nymphula icciusalis (yellow or brown ground).

Nymphula maculalis (dark or spotted male; white or gray female).

The so-called species of *Haploa*, except *clymene*, should perhaps be put in this category.

In the west we have also the white and yellow males of *Estigmene* acræa, and the yellow and black forms of *Papilio polyxenes*. There are many cases of dimorphic or polymorphic larvæ, where the moth varies but little; the following are conspicuous cases, the alternative color being usually green and brown or black.

Dimorphic or polymorphic larvæ

Papilio polyxenes (light or dark). Vanessa cardui (white to black). Deilephila lineata (green, checkered, or black). Lapara harrisii (green and white, or brown). Sphecodina abbotii (green spots on brown, or checkered). Herse cingulata (green or brown). Sphinx chersis (green or pink). Several other Sphingidæ (green or brown). Tropæa luna (green or reddish head). Eacles imperialis (green, tawny and black, or black). Citheronia regalis (green, blue, brown, or rose). Acronycta (many species). Noctua c-nigrum (brown; in summer often green). Leucania unipuncta (brown or black). Cosymbia lumenaria (green and striped, or brown and checkered). Lygris diversilineata, and many other geometers (green or brown). Cucullia asteroides (green, or yellow and white). Moths of high variability

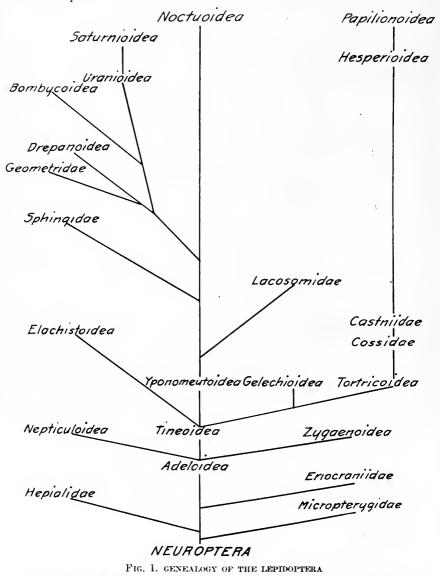
Telea polyphemus (tawny, yellow, brown, or olive). Automeris io (female). Feralia jocosa (green, cream, or brown). Brotolomia iris (amount of green and purple). Lithacodia muscosula (green or gray). Metrocampa prægrandaria (green or gray, both fading to yellow).

The instability of green and purple pigments is notable. Most species with these colors fade rapidly while still alive, sometimes even before emergence from the pupa.

RELATIONSHIPS

It is impossible to represent the true relationships of the Lepidoptera in a linear arrangement. In general, the main branches of the order have been placed according to their degree of specialization as a whole; but where development has been in such varied directions this is a

more or less arbitrary process. Thus the butterflies (Rhopalocera) have been placed at the end, as furthest developed of all, but the relationships of the base of their stock are clearly with the Cossidæ —



At the time of preparing this chart I was unable to place the Pyralidoidea. I now believe they should be derived from the Yponomeutoidea

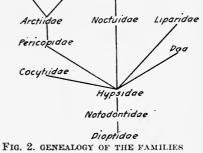
an undoubtedly early type; and the lowest butterflies are but little removed from the highest of the Cossid type (the tropical American and Australian Castniidæ). In certain ways (as in the thoracic sclerites) even the highest butterflies show their primitive character. For such reasons some recent authors have inserted the butterflies next to the Cossidæ, and have put the Euchromiidæ or Noctuidæ at the pinnacle of the order. It should be remembered, in any case, that each is the terminal of a long line of separate descent; and a true arrangement would put them side by side rather than one above the other.

The diagram (fig. 1) indicates the writer's impression of the probable relationship of the families. The Jugatæ are undoubtedly an early type and approaching extinction. The Frenatæ can scarcely have descended from either of the three Jugate families, but must have come from a stock with the haustellum, ovipositor, and tibial spurs of the Eriocraniidæ, and the larva of the Hepialidæ. The earliest type of Frenate must have been much like Incurvaria, but doubtless larger, and with characters that have been lost in all the aculeate genera, but survive in the lower Tineidæ, such as Acrolophus and Scardia. From such a type the Tineine superfamilies have arisen as separate branches, the Tortricoidea perhaps as another branch, or in association with the Yponomeutoidea, and the Pyraloids and higher Frenatæ doubtless from a common stem with the Yponomeutoids, but diverging very early.

In a similar way, the individual superfamilies of the Macrofrenatæ must have separated very early; but the Drepanoidea, Uranioidea, Saturnioidea, and Geometroidea may have hung together a short time after the Sphingidæ and the Noctuoidea had become distinct.

As to the internal evolution of the superfamilies, little is clear in most cases, save that the author has Novidge Amoridae Againstidae

aimed to arrange the families in order, from the more generalized to the specialized. In the noctuid group the relationship is a little clearer (fig. 2), the connection between Notodontidæ, Liparidæ, and the remaining families being through the exotic families Hypsidæ and Pericopidæ. The Hypsidæ have the primitive venation of the Liparidæ but have preserved the ocelli and the tongue. The Dioptidæ have the trifid venation and the free **Sc** of the



RELATED TO THE NOCTUDÆ

Notodontidæ, but the tympanum is of a primitive type from which the others may be derived.

In the butterflies, the line of descent through Hesperiidæ and Papili-

onidæ to the Pieridæ is clear, and the Nymphalidæ are certainly a direct offshoot of the Pieridæ; but the attachment of the Lycænidæ is less certain.

DISTRIBUTION

The life zones of New York and the adjacent States are:

1. Arctic-alpine

4. Alleghanian or Transition

- 2. Hudsonian
- 5. Carolinian or Upper Austral
- 3. Canadian
- 6. Austroriparian or Lower Austral

In northeastern North America, the Arctic-alpine zone is confined to the true Arctic region and the tops of the White Mountains of New Hampshire. The Green Mountains, the Adirondacks, Mt. Katahdin, the Catskills, and even some lesser peaks have stray alpine species, but south of the Catskills there are none. The bleak coast of Labrador belongs to this zone, but the inland shows the wooded character of the Hudsonian zone.

The Hudsonian zone is marked by an average midsummer temperature of about 50° to 57° F., and by the dominance of fir, spruce, and tamarack. It covers the continent from the Laurentians north, and shows on the higher levels of all our mountains, even south to North Carolina. Characteristic of its northern portion are *Brenthis* montinus, Polygonia gracilis, and Alypia langtonii. South of the Mohawk Valley its Lepidoptera have not been recognized as distinct from the Canadian forms.

The Canadian zone has an average temperature of 57° to $64^{\circ}F$, and also is a great area of conifer forest, but one readily giving place to many deciduous trees. While characteristic of Canada and northern Maine, the most fully cultivated parts of Canada, including the sheltered inner half of Nova Scotia and all of Quebec and Ontario south of the Saguenay and Laurentians district, belong to a warmer area. South of Canada, most of the mountains of New England and New York, as well as all the cooler summits of the Appalachians, belong to this zone. The peat bogs and swamps at lower levels often show the fauna of this zone, especially where tamarack and the heaths (Labrador tea, etc.) are dominant, rather than the southern white cedar. Characteristic Lepidoptera are Basilarchia arthemis, (which runs into the Transition), Chrysophanus epixanthe, Pieris oleracea (formerly wide spread in the Transition also), Eurymus interior, Polygonia faunus, and others.

The Transition, or Alleghanian, zone is typical of New York, and the data of seasons and broods are designed especially to apply to this district. In this zone deciduous trees are quite as common as conifers, and the northern beech and southern chestnut overlap. It can hardly be said to have a special flora and fauna, but only a special combination of forms. It covers the area with a summer temperature of 64° to 72° , or a total growing heat (sum of daily excesses over 43°) of $10,000^{\circ}$ to $11,500^{\circ}$ F.

The Upper Austral, or Carolinian, zone has a summer temperature range of 72° to 79°, or a total growing heat of 11,500° to 18,000°F. It is the most northern area for many characteristic species: *Papilio* marcellus (and its food, the papaw), cresphontes, philenor, Pieris protodice, Eurema nicippe, Euptoieta claudia, Chlorippe clyton and celtis (with Celtis, their food) and others, besides a large number of skippers.

The most striking of the characteristic moths are *Herse cingulata*, *Phlegethontius sexta*, and *Citheronia regalis*. Besides a part of the Mississippi Valley, it embraces the extreme southern part of Ontario and the Great Lakes strip of New York; but on the Atlantic Coast it is more restricted, including the major part of Virginia and Maryland and sending long arms along the coast and up the rivers. Distinctively Austral forms even reach Albany on the Hudson, Springfield on the Connecticut, and Boston along the coast. Part of the Austral species named are strong flyers, and also appear sporadically far out of their range, where they rarely or never breed.

The lower Austral zone, or Austroriparian, scarcely enters the region under discussion, but can be considered to include the coast of Virginia, and possibly isolated points in Maryland, and Cape May, New Jersey. Its summer temperature is over 79°F., and its total heat is 18,000°. Most probably in this case the total heat is the controlling factor.

Only a few lower Austral species are recorded in this monograph, either those that are particularly striking or those that have been often reported as strays in the North. In fact, our part of the area has scarcely been studied.

South of this there is a series of tropical zones, which do not concern us.

The control of humidity is, in our region, less striking, as it limits itself mainly to the setting off of the very damp off-shore islands, where ground vegetation is rank and trees are often gnarled and stunted. This strip has been so changed by man—largely through fires and sheep pasturing—that it is no longer possible to say what its natural stock was; but it is particularly marked by a wide northward range of species otherwise sub-tropical, like the *Prenes* and *Prionapteryx nebulifera*, mixing side by side with the boreal forms. This is perhaps because the blanket of fog masks the severity of the winters; an effect that reaches an extreme on Nantucket, whose indigenous Lepidoptera have now largely disappeared because of sheep pasturing. There is a marked difference between the Mississippi Valley forms and those of the Atlantic Slope, but this may be due as much to the barrier of the Appalachians as to any difference in climate. This subdivision shows clearly only in the Transition and Upper Austral zones. West of our territory there is an arid subdivision, which invades Illinois in a few isolated stations. The sand region of the Coast and in the vicinity of Albany and Peru, New York, forms a sort of artificial arid subdivision, where live a few characteristic western species. The Synedas may be examples; also Plagiomimicus and its relatives.

The control of physical barriers is best marked by the confining of the colder-zone insects in New York to certain of the higher peaks, and by the wider barriers of the St. Lawrence Plain (which stops $C\alpha n$ onynpha inornata, for instance) and the Mohawk Valley, which is perhaps the barrier for typical Pieris oleraca, and marks a varietal difference in certain geometers. These barriers, like that of humidity, are much less effective with us than in the Western States.

Distribution may also be viewed on a much smaller scale. Each spot, or station, where a colony of a species occurs, has its own peculiar characteristics of moisture and average temperature, and its own surrounding barriers, more or less effective: of these factors moisture is the most obvious variable, but fog or sunlight, close or easily drained soil, high or low water-table, each has an effect at least on the available food-plants and on the possibilities of pupation of the insect. Standing water has its own characteristic types (largely Nymphulinæ), and even running water has its peculiar species (Elophila fulicalis). Characteristic of the swamps are many Noctuids, a large part of them recognizable by their striated wings, simulating dead grass or reeds; as, for example, Leucania pallens and the Borolia group, the Nonagrias and their kin, Senta, Ommatostola, and Euchalcia venusta, as well as the species of Prenes, Chrysophanus epixanthe, Darapsa versicolor (which can only transform in wet moss), the Raphipteras, Epimartyria, and many others. Dry, open fields are relatively barren of Lepidoptera, but even they are the principal home of the Crambinæ.

It is a general rule that species in their most favorable local conditions will range far beyond their principal zone, and at optimum conditions of temperature will invade abnormal types of environment. This doubtless explains the curious mixture of northern and southern types, for instance, in the peat-bogs, where such arctic genera as Œneis find their only chance in the Canadian zone, and where the typically southern Exyras invade New England. So also the Nymphulinæ, which are richly developed in the tropics, find protection from the frost only in our ponds and streams, where they winter below the ice.

STRUCTURE

Imago

The *head* (fig. 3) bears a pair of antennæ, normally long and conspicuous, a large, minutely facetted compound eye on each side. in many cases with a minute simple eye, or ocellus, just above it; and the mouth parts, which include the labrum, a pair of mandibles, a pair of maxillæ, and the labium.

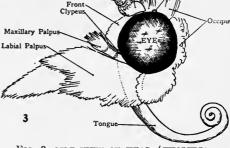
The antenna is composed of a large number of segments, most often from twenty to fifty. The

Antenna - shaft

basal segment, the scape (fig. 3), is much larger than the others; the second, or pedicel. contains a peculiar sense organ; and the remaining segments comprise the flagellum. Frequently the distal segments of the flagellum are thicker and shorter, forming a club (figs. 5 to 7). In this case the slender part of the antenna is the *shaft*. The club is usually without scales on the under side, even when the shaft is fully scaled, and varies in form and structure.

In the primitive forms the club is fusiform (fig. 4) gradually thickening and as gradually tapering to a point. It may be bent in the middle, or hooked, as in the skippers (fig. 5), and the hook may vary in stoutness and length, and in the number of segments involved. The higher butterflies, which have no hook, often show, by the asymmetry of the terminal segments, how the hook has become vestigial. In some forms the club is not sharply set off from the shaft (as in Feniseca), and in a few Satyrids its past existence is indicated only by an abrupt change in the color of the flagellum. Antennæ which are not clubbed usually taper to a point; those which have lost a club usually end bluntly.

The normal antenna is partly clothed with scales, which typically form two rings about each segment or two bands on the dorsal side (fig. 8), but always leave more or less extensive sensory areas covered with minute hairs, which by contrast appear naked. In many lower forms these bare areas are inconspicuous, but as a rule the under half, more or less, of each segment is naked. Lepidoptera with clubbed



dicel

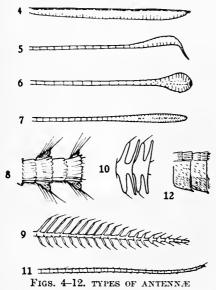
Scape

Vertex

Ocellus

FIG. 3. SIDE VIEW OF HEAD (THOLERIA REVERSALIS)

antennæ usually have the shaft completely scaled, and the sensory area confined to the club, which it may completely cover. In this case there are often special areas marked by pits or grooves separated by longitudinal ridges, and containing special structures. When each



4, Fusiform; 5, clubbed and hooked; 6, 7, clubbed; 8, serrate and fasciculate, dorsal view; 9, bipectinate; 10, doubly bipectinate; 11, simple and ciliate; 12, laminate or prismatic, side view

segment has a quadrangular extension on the under side covered with these sensory hairs, the antenna is called *laminate* or *prismatic* (fig. 12); when the segments are extended sideways in little teeth, the antenna is serrate (fig. 8); and when there are long branches, it is pectinate, or, in extreme cases, plumose (feathered). If there is but one series of such branches, it is unipectinate (a rare condition); if there are two, it is bipectinate (fig. 9), and when each segment bears two pectinations or branches in each row, it is doubly bipectinate, as in the Luna moth and its kin, (fig. 10). In doubly bipectinate antennæ the pectinations are often alternately long and short, or thick and thin. Where there is a long, strong pair of bristles on each segment, the antenna is *ciliate* or bristled (fig. 11); if the bristles come in distinct tufts, it is fasciculate, but if they are numerous and scattered evenly it is pubescent (fig. 87). In many sphinx moths, especially

males, the bristles are in two vertical rows on each side of each segment, whose tips curve and meet so as to seem to form loops. The scales on the antenna usually form two transverse rows on each segment (fig. 46); sometimes they are scattered (fig. 48); and in the Saturniidæ and many butterflies they are either wholly absent, or fugitive, except on the two basal joints.

The eyes vary in size and shape. Typically they are circular in side view (fig. 3), and about as wide as the distance between the two eyes as seen in a front view. Often the eyes are much narrower, and much higher than wide (elliptical, fig. 271) or even concave on the posterior margin (reniform). In the Lycznidz they appear as if cut

off behind, and there is a slight notch in the upper front side, where the antenna arises. Often there are minute straight hairs arising

EYD

Labial

Palpus

FRONT

Clypeus

Maxillae

Tongue

between the facets, in which case the eye is called *hairy* (fig. 14). A *lashed eye* is one in which bristles arise around the edge and curve over it (fig. 15), especially in front below the antennæ, and behind. The facets are usually finer and more numerous in nocturnal forms.

The simple eyes, or ocelli (fig. 3), when present, lie

FIG. 13. FRONTAL VIEW OF LOWER PART OF HEAD, behind the root of the an-DIAGRAMMATIC tennæ. They vary in size

and in distance from the eye, and are often absent, as in all butterflies. The portion of the head between the eyes in front is the *front*, called *clypeus* or *clypeus posterior* by some entomologists (figs. 3, 13), and

Maxillary



EVE

Pilifer

Epipharynx

13

Gena

FIG. 14. HAIRY EYE (HIGH MAGNIFICA-TION)

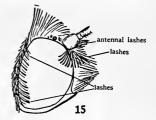


FIG. 15. LASHED EYE (MODER-ATE MAGNIFICATION)

the sharp projecting ridge along its lower boundary is the true *clypeus*. The head between the eyes above is the *vertex*; and the back of the head, behind the eyes and ocelli, is the *occiput* (fig. 3).

The *labrum* of the imago (fig. 13) is a three-lobed structure. The middle lobe merely closes the space between the bases of the maxillæ; the two lateral lobes are the *pilifers*, and project across the base of the tongue, each bearing a tuft or row of bristles, which provide important characters in the classification of the Sphingidæ.

The mandibles are almost always rudimentary or absent, difficult to find, and unimportant (fig. 49). In Epimartyria alone there are regular biting mandibles; but in a dried specimen they are usually difficult to see without dissection.

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Each maxilla (save in Epimartyria) consists mainly of a slender, eoiled portion, the galea (figs. 37, 49). The base of the maxilla in many eases bears a two- to four-jointed sensory organ on the outer side, the maxillary palpus. The two maxille are grooved on their inner sides and hooked together to form a hollow sucking tube. Together they are known as the tongue.

The tongue may vary in size and stiffness. When it is shorter than the head in slender moths, or shorter than the thorax in heavy and strong ones, it is considered rudimentary in systematic work, as it is in the Pyralids if it is not large enough to show between the palpi when coiled up. It has sensory bristles at the tip and often bears scales at the base, but as a rule the base is naked. In certain noctuids, as *Alabama argillacea*, the bristles at the tip form a rasping organ with which the rinds of fruits can be pierced. A few moths lack the tongue. The maxillary palpi are conspicuous and five-jointed in the lowest moths, and are folded at rest, being more or less movable. As one goes up the scale they become smaller, till in our Noctuidæ they are mere microscopic, scaly tufts, and in the butterflies and the Geometridæ they are unrecognizable.

The *labial palpi*, often called merely the palpi, arise on each side behind the base of the tongue (in the lower forms from a small labium,

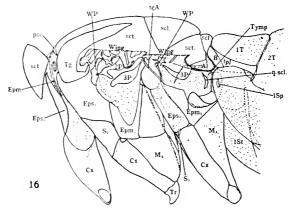


FIG. 16. LATERAL VIEW OF THORAX, DENUDED, WITH WINGS AND LEGS REMOVED

Al. alula of hind wing; B. entrance to tympanic bulla; Cx, coxa; Epm, epimeron; Eps, episternum: M, meron; n. scl. nodular sclerite of tympanum; 2P, 3P, paraptera (3P is the "subalar sclerite"); psc, prescutum; scl, scutellum; sct, scutum (sct, is the patagium); 1Sp, spiracle of first segment of abdomen] S, sternum (thorax); 1St, sternite of first segment of abdomen; Tg, tegula; tgA, tegular arm; tpl, tergopleural groove; Tymp, membrane of tympanum; WP, wing process of pleurites; (The subscript numbers, 1, 2, 3, indicate the segments of the thorax)

in the higher ones directly from the surface of the head), and extend forward on each side of the base of the tongue. They are composed of three joints, and are densely clothed with scales, which are usually thicker on the upper and especially the lower side than on the lateral faces, giving them a more or less blade-like form. If the palpi extend nearly straight forward, they are called *porrect*; if the scaling on the end of the second joint ends abruptly with a slight tuft, they are *clavate* (fig. 244). In the Tineidæ they usually have strong bristles besides the scales and soft hair; but this is rarely the case in higher families of moths.

The thorax (fig. 16) is composed of three segments, prothorax, mesothorax, and metathorax, each provided with a pair of legs, and the last two each with a pair of wings. The legs (fig. 17) are segmented, the segments being, named in order beginning at the base: coxa, trochanter, femur, tibia, and five tarsal segments. The coxe of the middle and hind legs are partly fused with the body and divided into an anterior part, the true coxa, and the posterior part, the meron, by a suture. The trochanter is minute. The femur is rarely modified but often bears long hairs, even when the rest of the leg is scaled. In the males of the Catocala group, there is a minute spine, the gonyodon, on the apex of the fore femur.

The fore tibia bears a leaf-like structure on its inner side, the *epiphysis*; this is clothed with stiff bristles, and serves mainly to clean

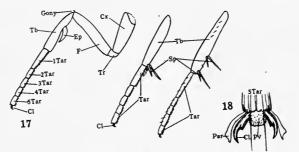


FIG. 17, 18. LEGS AND FOOT

Legs: Cl, claw; Cx, coxa; Ep, epiphysis; F, femur; Gony, gonyodon; Sp, spurs; Tar, tarsus; Tb, tibia; Tr, trochanter

Foot: Cl, claw; Par, paronychium; Pv, pulvillus; 5 Tar, body of fifth segment of tarsus

the tongue and antennæ. The higher butterflies and a very few moths have lost the epiphysis. The tibia also sometimes bears one or more enlarged spines or claws at the tip. The mid and hind tibiæ have a pair of *spurs* at the tip — scaled, spine-like structures articulating with

the tibia. The hind tibiæ have a second pair, usually near or below the middle.

In many butterflies and Noetuidæ the tibiæ bear spines similar to those on the tarsus, but less regularly arranged. Males frequently have tufts of sex-hairs, and in many cases the tibia is grooved to protect them. On the hind legs of many Ennomid geometers, the tibia is largely hollow and the tuft almost completely enclosed, though its point of attachment is regularly on the exterior.

The last five segments constitute the *tarsus*, or foot, the most basal being the *metatarsus*. This segment is usually much larger than the other four, and occasionally bears special tufts or spines. The final segment (fig. 18) ends in two articulated and curved *claws*, and an adhesive pad between them, the *pulvillus*. On the inner side of each claw, toward the pulvillus, there is often a sensory membrane, the *paronychium*. These are large and conspicuous in the Sphingidae, and are often forked. The pulvillus is rarely minute or absent.

The tarsal segments are also armed with smaller spines, which usually form three or four rows on the ventral surface, but sometimes are dorsal also, or gathered into tufts at the tips of the segments. They are sometimes concealed in the scaling, though rarely absent. Often they are wanting on the base of the metatarsus and are differently arranged on the last segment. In the family Nymphalidæ, the fore legs are reduced and not used for walking. They have no claws and the segments are more or less fused, but the general appearance of a leg is retained. The male carries the reduction slightly further than the female. The Erycinidæ and Libytheinæ show less reduction in the male and none in the female: in their males the tarsus is very small and without claws, and the tibia extends beyond its insertion as a sort of claw. In the males of the Lycanida the fore legs are still less reduced; while the leg has lost its claws, it is functional, ending either in a single claw-like spine or a group of spines. In the Herminiinse the leg is more curiously modified in the male, although perfectly normal in the female. The coxa may be very long and movable (it is never as closely fused with the body in the fore leg as in the middle and hind ones). The trochanter is sometimes lengthened. The femur tends to be weak, the tibia and first joint of the tarsus, either or both, may be hollowed out to a mere shell, enclosing enormous tufts of hair; and the tarsus is sometimes minute. The leg as a whole may be enlarged or reduced. Many moths, especially geometers, bear a tuft of fine hair at the junction of the hind tibia and femur, or on the tibia near the base; this is also often contained in a hollow in the tibia, and in some cases is lost at copulation. In the Acidaliinæ the tibia may be much enlarged to enclose it, or may be reduced, even to an extreme, while

the tarsus in these cases is usually reduced. In several exotic Acidaliinæ one of the spurs bears a comb which seems to be used in handling this tuft. Some species of Ptychopoda (Eois) have almost completely lost the hind legs. These modifications are confined to males, or are carried much further in males than in females. Besides these features spurs are often reduced or lost, and occasionally (Leucania, Nematocampa) modified in form. In the lower Tineina the tibial spines are represented by long, slender, but stiff bristles which may be either in regular rows or irregular. In some cases, as in Nepticula and Acrocercops, tibial spines are a conspicuous feature, but in the Œcophoridæ and others they seem to intergrade with the ordinary hairs. In some Sphingidæ and Plusiæ the spines at the base of the tarsus may form a distinct *comb* (fig. 17, middle leg).

The wings (fig. 19) are usually more or less triangular, the three

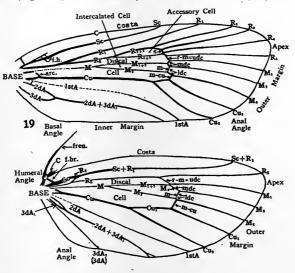


FIG. 19. WINGS (ACROLOPHUS POPEANELLUS d)

C, costa; Sc, subcosta; R, radius; R₁, R₂, R₂, R₃, R₆, branches of radius; R_s radial sector (usually merely labelled R in the hind wing); M, media; M₁, M₂, M₃, branches of media; Cu, cubitus; Cu₂, Cu₂, branches of cubitus; 1stA, 2dA, first and second anal veins; 3dA, third anal vein; 3dA₁, 3dA₂, branches of third anal vein (3dA₂ of the hind wing is usually merely labelled 3dA); f.br., frenulum-brace (humeral); arc., arculus; S, sectorial cross-vein; r-m, radio-medial cross-vein; m, medial cross-vein; m-cu, medio-cubital cross-vein; udc, upper discocellular vein (in this case the same as r-m); mdc, middle discocellular vein (in this case M and a portion of M₃); fren., frenulum; f.h., frenulum hook

sides, costa, outer (or hind), and inner margin, and the three angles, base, apex, and anal angle, being indicated in the figure. Sometimes the inner margin of the fore wing is extended out near the base, forming a basal angle, or the costa of the hind wing may be similarly extended, forming a humeral angle. The wing is stiffened with a regular pattern of hollow rods, the *veins*, which are important when the wing is expanding, besides serving to stiffen the mature wing. These veins have a definite arrangement, based on that of the pupal tracheæ around which they form (see the figures). From the base of the wing there run out costal, subcostal, radial, medial, cubital, and some anal main Of these the costa is simple and forms the front edge of the stems. wing; the subcosta may be simple, but in a few low forms it forks once at the tip. The radius forks normally into five branches, primitively as shown in figure 40 but in various ways in the higher forms. Often part of these branches are lost, and in a few of the lowest species the first branch forks again (fig. 36). Media is three-branched, but usually the base of it is lost, and the branches are variously attached to the stems of radius and cubitus; when one is attached to cubitus, the latter is called trifid (fig. 425), when two, quadrifid (fig. 427), the upper median being always free or attached to radius. The middle branch of media, sometimes called the independent, is often weak or lost. *Cubitus* is two-branched, and is a very constant feature. The anals are somewhat uncertain in origin, but usually appear in low Frenatæ as three radiating veins, the first running along the principal concave fold of the wing. The first anal disappears in higher forms, leaving only the fold; the second is persistent; and the third tends to grow short in the hind wing, and to join its tip to the second, or even to disappear, in the fore wing. In the Jugatæ (figs. 31, 36, 40) the arrangement is more complex and not fully understood. This description, intended mainly for the fore wing, applies also to the hind wing of the Jugata. In the hind wing of the others the radius is only two-branched, and the upper branch fuses more or less, often almost completely, with subcosta, as indicated by the lettering of the earlier figures. Commonly $Sc+R_1$ is merely marked Sc, and R_s , merely R. Besides these veins, which are based on tracheæ, there are certain crossveins; the humeral (\mathbf{h}) at the base of the costa, often pushed to the base of the wing and so lost; the sectorial (s.), running across between the third and fourth branches of radius, and so enclosing the accessory cell, often lost by the fusion of the veins at that point; and the discocellular, running more or less irregularly from radius aeross media to cubitus, enclosing the discal cell (" cell ") between it and the base of the wing, and supplying an attachment for the branches of media when the base of media is lost. The arculus (arc.) connects the media and the cubitus at the base of the wing, but shows clearly only in the

fore wing of the Jugatæ; and there is occasionally a cross-vein in the anal region. The discocellular is divided into four parts by the three branches of media: upper, middle, and lower discocellulars (udc., mdc., ldc.), and the false base of media-three (m-cu.). In the hind wing the humeral is usually replaced by a short spur from the base of the subcosta, which may not reach the margin.

The Jugatæ have a special lobe on the inner margin of the fore wing, the jugum (fig. 40). In the Hepialidæ this slips under the hind wing in flight and helps the wings to work together.⁵ In the others it overlies the hind wing and is sometimes called a *fibula*. In most cases the wings are attached to each other by a bristle or group of bristles, the frenulum, which is attached to the base or the humeral angle of the hind wing. The frenulum either runs through a hook, the frenulum hook, attached to Sc near the base of the fore wing; or under a tuft of scales, the retinaculum, spreading up fanlike from near the base of Cu. The male almost invariably has a single frenulum running through a hook; the female has usually several bristles (the approximate number characteristic of the family) held merely by a retinaculum. In several groups (some Pyralididæ, some Pterophoridæ, Ægeriidæ, Euteliinæ) the frenulum of the female is also simple, but the hook is represented at most by a second tuft of scales. In many forms there is no attachment between the wings, which merely overlap; in a very few (Ægeriidæ, Pterophoridæ) the inner margin of the fore wing and the costa of the hind wing bear series of recurved spines, which interlock (fig. 223), the hamuli.

In the males of many species the wings bear scent glands. Sometimes these are merely scattered, associated with special scales, the androconia, which are designed to spread the scent, (as in Pieris and Eurymus). Frequently there are structures developed for their protection; as, for instance, the pocket on Cu in Danaus, on the inner margin in Carsia, the folded inner margin of many Olethreutes (fig. 266), of Lobophora and Heterophleps, and so on. The costal edge is particularly apt to be folded over, forming the costal fold (fig. 265, 306), and this may contain a large tuft or mass of androconia, as in a large part of our Hesperiinæ and Tortricidæ. In some instances the secreting area is clothed with special scales, forming a stigma (fig. 249). This is usually easy to see, but in such forms as Cercyonis and a few Pamphilas it is easiest to moisten the wings with chloroform or benzine and hold the butterfly to the light. When not specially noted, the location of a stigma is the upper side of the fore wing below the cell, in the Satyrinæ and Hesperiidæ, or at the end of the cell as in Thecla.

 $^{^{\}rm 5}A$ somewhat similar lobe occurs on the hind wing of some male Geometridæ and Tortricidæ (fig. 267), but probably with the function of a scent organ.

Two other structures on the thorax are occasionally of importance in some species. The dorsal part of the prothorax may be extended in a pair of mushroom-shaped bodies, the *patagia*, or, taken together, the collar. Sometimes they are called tegulæ (fig. 16, sct_1). They are only conspicuous in the Noctuoidea, and even there are so buried in vestiture as to appear merely part of the general body surface. The true tegulæ (sometimes called patagia) are attached at the base of the fore wing and loop about it, covering the base of the costa and the whole articulation (fig. 16, tg.). The part of the mesothorax exposed between the tegulæ is the disc. The metathorax is narrow above, widening a little to support the wings, and bears the basal hair. The general surface of the thorax is divided into a considerable number of sclerites, as in other insects; but as they are little used in classification, the figure is sufficiently explanatory.

To study the surface it must be denuded of scales and hairs. The back of the insect, above the level of the wings, is the *dorsum* or *tergum*, and its sclerites are *tergites* (sct, scl); the sides from wings to legs, are *pleura*, composed of *pleurites* (eps, epm, etc.), and the lower side, between the legs, is the *venter*, made up of *sternites* (s).

The abdomen is normally composed of ten segments, several of which are usually concealed, and it bears at its end the sexual organs, the most conspicuous of which, in the male, are the *valves*, a pair of claspers for holding the female; and, in the female, the *ovipositor*, or instrument for placing the eggs.

The abdomen may be divided like the thorax into dorsum, pleura, and venter, though in the absence of legs and wings the boundaries may seem a bit arbitrary. The membrane on the sides is considered to represent the pleura. On the first segment there is however a deep groove, the *tergo-pleural groove* (fig. 16, **tpl**.), and a small pleurite below it. Scent-tufts occur on the abdomen in many males, especially near the base in the Noctuidæ and near the apex in the Geometridæ. They are usually retractile in pockets when not in use, and are rarely seen expanded. Those near the tip of the abdomen (on the seventh segment) are called *coremata*.

In the males of most species there are eight visible segments. The body of the ninth (fig. 20) is reduced to a ring, the tergite of which is the *tegumen*, and its sternite the *saccus* or *vinculum*. Attached to the junction of tegumen and saccus is the *valve* (harpé), the name of which indicates a common form. The valve may be composed of a thickened dorsal edge, or *costa*, a central part, the *valvula*, or valve proper, and a lower part which is strongly curved and spoon-like, the sacculus. Lying in the middle line between the valves is the penis, or

adæagus, which is retractile, and often works through a ring, the *juxta*. Its reversible lining is often armed with spines, known as *cornuti*. The *furca*, a curious fork or simple spine in some Ennomids, is formed from the lower part of the juxta and may resemble an additional ventral valve. Its two halves are often separate. Besides these parts there

is a great variety of secondary structures, of which the most important are the *clasper*, a hook developed on the inner face of the valve, and the *peniculus*, a hairy prominence arising from the edge of the tegumen above the valve.

The end of the valve, especially in the Noctuidæ, often bears a row or mass of recurved spines, the corona (fig. 21). A finger-like process on the dorsal edge of the valve is a digitus, one on the ventral edge a pollex, and one near the base on the inner face (generally hairy) is the editum (dorsally) or clavus (attached to the sacculus). When the elagone is divided as in Accorvate

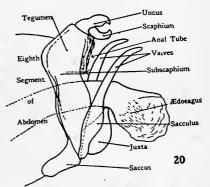
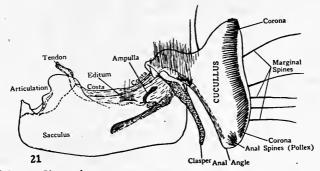


FIG. 20. TYPICAL MALE GENITALIA, SIDE VIEW (EUBAPHE .OSTENTA)

clasper is divided, as in Acronycta, its dorsal arm is the ampulla.

The tenth segment is further reduced. Its dorsal part is the uncus, which is most often a strong hook, but may be divided or reduced to

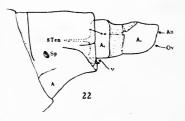


to see without dissection.

FIG. 21. VALVE OF MALE GENITALIA (HADENA DEVASTATBIX)

a plate, which in some forms is indistinguishably fused with the tegumen or scaphium; on each side it frequently bears a small appendage, the socius, which is conspicuous in the Thyatiridæ and the Hemitheinæ. The ventral part of the segment, when it appears at all, forms a plate between the anal tube and the penis, known as the *transatilla*, in many cases articulated with the base of the valves.

The end of the intestine is commonly a long tube, in many species strengthened by the *scaphium* above and the *subscaphium* below. Whether these structures represent the eleventh segment is an open question. They may be simple bands or plates, or they may bear spines or be of striking forms. The *gnathos* is a partially free subscaphium articulating directly with the uncus.





In most females the abdomen has seven visible segments (fig. 22). The eighth and ninth segments are retracted within the seventh and are, for the major part, composed of membrane. On the ventral surface of the eighth segment is the vagina, with a chitinous plate below it. The remainder of the segment is, in most cases, a short cylinder. The ninth segment is divided into two lobes between which lie the anus and the opening of the oviduct. The usual condition is shown in figure 22.

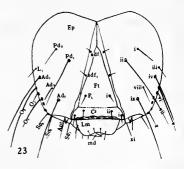


FIG. 23. HEAD OF LARVA WITH SET.#. NUMBERED BY DYAR'S SYS-TEM (RIGHT) AND HEINRICH'S SYSTEM (LEFT)

Ad₁, Ad₂, Ad₃, Anterodorsal setæ of epicranium; adf₁, adf₂, adfrontal setæ; Ant, antenna; Cl, clypeus; Ep, epicranium; Ft, front; F₁, its seta; L₁, lateral seta of epicranium; Lm, labrum; md, mandible; O₁, O₂, O₃, ocellar setæ of epicranium; Pd₁, Pd₂, posterodorsal setæ of epicranium; So₁, So₂, So₃, subocellar setæ of epicranium

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The four most primitive families show a different structure. In the case of the Micropterygidæ there are eight fully developed segments, and the ninth and tenth are invaginated within the eighth, the tenth segment being fairly well developed (fig. 34). The Eriocraniidæ and the Incurvariidæ have a complex piercing ovipositor, whose structure is shown in the figure of Eriocrania (fig. 38). The Hepialidæ have nine segments, but the structure is complex and not fully understood.

Larva

The caterpillars show the same essential structures and appendages as do the imagoes, but they are very different in development. As a rule, only the head is chitinized, the skin of the body being thin and flexible. The major part of the head (fig. 23) is composed of a pair of sclerites, the epicrania. Between these, on the face, lies the front, which is in most instances triangular and rarely reaches the top of the head. Between the front and epicrania there are two very narrow sclerites, the adfrontals, in some works called the paraclypeals. Below the front is a third narrow sclerite, the clypeus. The lower half of this, more or less, is composed of membrane. On the under side of the head there are two small triangular sclerites, the postgenæ.

The antennæ are very short and small, and lie immediately above the mandibles, which are the principal structures associated with the mouth. In front of the mandibles there is a flap, the labrum (fig. 24), which serves as an upper lip. Behind the mandibles there is a second,

somewhat thicker flap which functions as a lower lip. It is composed of the fused maxillæ and labium. It is complex in structure and but little used in classification. The eyes are represented by six small, separate ommatidia, located in a group low down on the side of the head, as shown in figure 232. There are a considerable number of setæ which are constant in number and position in any given NUMBERED ACCORDING TO FORBES'S species. A typical arrangement of these system (BIGHT) AND HEINRICH'S is shown in figure 23. There are two two L_{y} L_{z} , L_{z} , L_{z} , lateral setæ; M_{1} , M_{2} , methods of designating the setæ. The M_2 , median setæ right side of the figure is labeled accord-

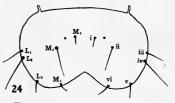
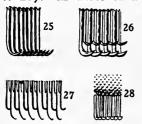


FIG. 24. LABRUM, THE SETÆ

ing to the method most often used in the past, while the left side bears the system recently proposed by Heinrich.

The body is composed of thirteen distinct segments, three belonging to the thorax, and ten to the abdomen. The ninth segment of the abdomen is, in most cases, much smaller than the others. The prothorax bears a sclerite on its dorsal surface known as the *cervical shield* (shown in the diagrams at the head of each family). The setæ on the side are arranged in two groups, called the prespiracular, and the subventral, warts. Each of the segments of the thorax bears a small leg which shows the usual segments, coxa, femur, tibia, and tarsus; but all are very small, and the tarsus is composed of a single segment and ends in a single claw. The abdomen is without true legs, but the third to sixth and last segments bear membranous prolegs, which may have, at the tip, a series of hooks, or *crotchets*. There are *spiracles* on the side of the first segment of the thorax and the first eight of the abdomen. In many caterpillars, especially aquatic forms, the last spiracle lies on the dorsum and faces backward, as shown in the figure of Carposina (fig. 293). The body, as a rule, bears a regular arrangement of seta on each segment. These seta are designated by numbers, as shown in the figures, and are known as primaries; when there are a small number of additional setæ, they are known as subprimaries (tigs. 434, 439); a large number, indefinite in position, are called secondaries (fig. 405). In certain families, in place of single set in the primary positions, the tubercles from which the hairs arise are grouped in warts, each bearing a tuft of setæ (fig. 406). In this case the caterpillar is spoken of as having tufted hair.

The hooks on the prolegs also show a definite arrangement (figs. 25 to 28). If those of a series are all of the same length, or regularly



FIGS, 25-28. HOOKS OF PROLEGS, TYPICAL ABRANGE-MENTS

dinal; 28, multiserial

graded in length, they are called uniordinal; when alternately of two lengths, biordinal; and when of three lengths, triordinal. If there are one or more rows of rudimentary hooks at the base of the functional ones, the arrangement is called multiserial. The series of hooks may be arranged either in one or two transverse bands, a longitudinal band, or an ellipse which is usually broken at one or two points. In very rare cases, when there is a longitudinal band on the inner face of the proleg, there is a weaker band, or some scattered hooks, on the 26, biordinal; 27, trior outer face. Fracker calls this arrangement a pseudocircle.

Pupa

The pupa, or chrysalis (fig. 29), also shows all the structures characteristic of an insect. All the appendages, however, are folded back on the body and may be soldered to it. The most conspicuous appendages are the wings, which lie on either side of the venter, at the front end of the body. The hind wings are almost completely covered by the fore wings, but a small portion of their posterior edges may show. On the midventral line lies the tongue, and between the tongue and the

wings are the first two pairs of legs and the antennæ. The mesothorax and metathorax are easily recognized by their attach-

ment to the wings. The prothorax lies immediately in front of them, and the remainder of the front of the body belongs to the head. In many species this is divided, by transverse sutures, into two or three sclerites. The mouth is surrounded by the *labrum* in front and the tongue behind. On the sides the mandibles are located when they are developed, but they are absent in most species. In the more primitive forms the maxillary palpi (or "eve-collar") appear immediately behind the eyes as small oblong or triangular sclerites. The abdomen is composed of a regular series of segments, the first three or four of which are, in most cases, immovably soldered together. Then there come one or more movable incisures, and the terminal segments are again fused. An in*complete* pupa is one in which there are four or more movable incisures, one more in the male than in the female, and in which the body is provided with spines to enable the pupa to work out of its burrow or cocoon or out of the soil. In an obtect pupa there are three or less movable incisures in both sexes, and the pupa does not leave the cocoon. At the end of the body there may be an extension bearing a tuft or group of setæ, which are hooked in most instances. This is the cremaster. In some pupze the male may be distinguished by its wider antennæ, and in some

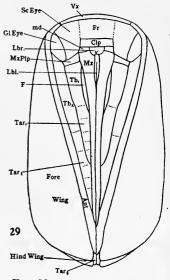


FIG. 29. VENTRAL VIEW OF AN-TERIOR PART OF PUPA, DIAGRAM-ATIC, SHOWING STRUCTURES

Ant, antenna; Clp, clypeus; F, fore femur; FT, front; Gl Eye, glazed eye; Lbi, labium, with palpi; Lbr, labrum, with pilifers; md, mandible; Mx, maxilla; Mx Plp, maxillary palpus (eyecollar); Sc Eye, sculptured portion of eye; Tar,, fore tarsus; Tar₂, mid-tarsus; Tar₈, tip of hind tarsus; Tb₁, fore tibia; Tb₂, mid-tibia; Vx, vertex

the two separate reproductive openings of the female can be recognized. 2

SYNOPSIS OF THE FAMILIES OF LEPIDOPTERA

Suborder Jugatæ. Moths with fore and hind wings similar in form and venation, and with at least four radials in the hind wing. Base of cubitus fused with 1st A, a short portion of it appearing like a cross vein between 1st A and the apparent base, which is in fact the arculus. Wing membrane spinulated (fig. 30). Frenulum rudimentary or absent. Fore wing with a separate lobe, or jugum, at base of inner margin. Larvæ with dorsal setæ similarly arranged on thorax and abdomen.

Family 1. Micropterygidæ. Minute moths, with functional mandibles, and maxillæ of primitive type formed of galea, lacinia, and palpus; feeding on pollen. Fore wing with subcosta forked near its middle. Female with ten abdominal segments preserved and without ovipositor. Larvæ with modified clubbed setæ, feeding on wet moss. Pupæ with large mandibles.

Family 2. Eriocraniidæ. Minute moths, with rudimentary mandibles, and maxillæ possessing a short colled tongue. Lacinia absent. Subcosta of fore wing forking near its apex. Female with complex piercing ovipositor, laying its eggs in the tissue of leaves. Larvæ without hooks on the prolegs, which are rudimentary; with simple setæ, mining in leaves. Pupæ with large mandibles.

Family 3. Hepialidæ. Large, or very large, moths, with rudimentary, nonfunctional mouth parts, save for the labial palpi. Subcosta of the fore wing forking near its middle or simple. Female without piercing ovipositor. Larvæ of normal caterpillar form, boring in stems and roots; with a circle of hooks on the prolegs. Pupæ with small obscure mandibles, and rudimentary, divergent maxillæ.

Suborder Frenatæ. Moths with hind wing much smaller, or shorter and broader, than fore wing, with at most two free branches of radius. Cubitus straight to base, no portion of it appearing like a crossvein. Wing membrane rarely spinulated. No jugum. Hind wing usually with a frenulum. Antennæ rarely clubbed, and clubbed in none of our forms which lack a frenulum. Pupa with trachea \mathbf{R}_{4+5} arising from the stem of radius beyond that of \mathbf{R}_1 . Larvæ almost never with dorsal setæ similarly arranged on thorax and abdomen.

*Wings with acule over the general surface.

Superfamily Incurvatioidea. Venation fairly complete; antennæ without eye-cap; female with piercing ovipositor (so far as known): larvæ usually case-bearers when mature.

Family 4. Incurvariidæ. Characters of the superfamily.

Superfamily Nepticuloidea. Venation much reduced; cell very small or absent; antenna with large eye-cap; female without ovipositor,

the eggs laid exposed. Larvæ leaf-miners or bast-miners when mature.

Family 5. Nepticulidæ. Characters of the superfamily.

**Aculeæ confined to small areas or absent.

[†]Larvæ with warts i and ii, iv and v united.

Superfamily Zygænoidea. Moth with primitive venation, large wings, and minute maxillary palpi or none; Sc and R of hind wing usually uniting along cell. Larvæ short and slug-like; typically with diffuse hair-tufts; pupa primitive, incomplete, with all segments free; first abdominal spiracle uncovered, maxillæ usually extended and toothed laterally, but without maxillary palpus.

Family 6. Megalopygidæ. Moths with tongue obsolete; wings heavily and loosely clothed with soft scales, often mixed with curly hair; hind wings with Sc and R fused for most of length of cell. Larvæ with 14 prolegs, 10 of them with an angulate or broken band of hooks.

Family 7. Eucleidæ. Moths with tongue obsolete; wings usually heavy and loosely scaled; hind wings with Sc and R shortly fused near base. Larvæ with prolegs abcont, replaced by a system of sucking discs; hairy, spiny, or naked.

Family 8. Pyromorphidæ. Moths with tongue strongly developed. Wings translucent; hind wings with Sc and R fused toward outer end of cell, the basal part of R also fused with Sc or obsolete. Larvæ with tufted hair; with 10 prolegs, provided with normal hooks.

ttLarvæ with warts i and ii separate, iv and v often united.

 \ddagger ''Micros.'' Larvæ with three setæ on prespiracular wart, usually concealed feeders; moths with soft-scaled wings with broad fringes, broadest at anal angle, the wings often lanceolate or linear; \mathbf{R}_5 often running to costa or apex; accessory cell, when distinct, with its broad side resting on discal cell, often fusing with it; hind wing with Sc and R never fusing beyond cell, and very rarely at any point, the base of R often obsolete; 1st A rarely lost unless wing is very narrow; tongue very often scaled at base.

Superfamily **Tineoidea**. Larvæ with setæ iv and v separate; pupa normally incomplete; moth normally with rough head, often with folded maxillary palpi; tongue scaled when present; fore wing usually with \mathbf{R}_5 running to costa, hind wing with **R** and \mathbf{M}_1 separate, **Sc** and **R** typically approximate at base.

Family 9. Tineidæ. Larvæ with anal prolegs unlike the ventrals; most often scavengers; moths winged in both sexes, without a heavy anal tuft in the female; usually with rough head, folded maxillary palpi, and bristles on the labial palpi, or with the first joint of the labial palpi enlarged.

Family 10. Psychidæ. Larvæ with anal prolegs similar to the ventrals, each with a single ellipse of hooks, broken on inner side; always living in a case. Female moth with a tuft of heavy hair-scales at the end of the abdomen, which

are mixed with the eggs; almost always wingless, and often maggot-like with all appendages rudimentary. Male with rudimentary mouth parts.

Family 11. Tischeriidæ. Larvæ leaf-miners; the thoracic legs absent, and each ventral proleg with two transverse rows of hooks. Moths with a loose tuft of scales on vertex, a small scape, short porrect palpi without bristles, small maxillary palpi, and very hairy hind tibiæ; fore wing with Cu running through the center of the wing, with all radials running to costa.

Family 12. Lyonetiidæ. Larvæ leaf-miners; structurally much like the Tineidæ, flattened, with setæ iv and v more widely separated. Moths with head smooth, at least on front, with a well-marked eye-cap, the hind wing with R running through the axis of the wing, which is often linear.

Family 13. Opostegidæ. Larvæ very slender, cylindrieal, without legs; bastminers; moths with folded maxillary palpi; with very large eye-caps; fore wing with three or four unbranched veins, hind wing linear. (Position of family uncertain.)

Family 14. Gracilariidæ. Larvæ when young very much flattened, with bladelike mandibles, and rudimentary maxillæ and labium; living as leaf-miners and bast-miners and eating the sap only; when grown usually with normal mouthparts, eating the parenchyma, and often living as leaf rollers. No prolegs on sixth segment of abdomen; the other legs variable. Moth with maxillary palpi porrect or obsolete, our species without eye-cap; fore wing lanceolate, normal or with somewhat reduced venation; hind wing often linear, with principal vein through the center of the wing; sometimes with \mathbf{R}_1 free. (Position of family uncertain.)

Family 15. Coleophoridæ. Larvæ with iv and v eloser than usual; with very strong true legs, and prolegs reduced, each with two transverse bands of hooks or none; usually leaf-miners when young, or feeding within seeds; almost always becoming case-bearers when grown. Moths with smooth head; palpi moderate, usually tufted, with the third joint often set on at an angle; fore wing with cell set obliquely, the cubitals extremely short and running directly to inner margin. Epiphysis rudimentary, at apex of fore tibia, or absent. Antennæ turned forward in repose. (Position of family uncertain.)

Superfamily **Cycnodioidea**. Larvæ and pupæ not well known; apparently the larvæ with setæ iv and v close together; leaf-miners when young and case-bearers when grown. Pupæ without maxillary palpi, with nearly all segments of abdomen free, or, in Elachista, which pupates exposed, with all soldered. Moth with \mathbf{R}_{2+3} of hind wing preserved, running to costa, maxillary palpi minute and straight or lost, and with characteristic genitalia; \mathbf{R}_5 running to costa, lost in all our species; **Sc** and **R** of hind wing widely separated, in our species with **R** running through middle of wing.

Family 16. Cycnodiidæ. Palpi moderate, upturned; hind wing lanceolate, with well-formed cell, and nearly complete venation. Larva sixteen-legged, a blotchminer; pupa suspended, exposed, immobile.

Family 17. Douglasiidæ. Palpi short, drooping; ocelli very large; hind wing without cell; with R_{2+3} separating off from radial stem before M_1 . Larvæ leafminers, hardly known. (Position of family uncertain.)

Family 18. Heliozelidæ. Palpi short, drooping; hind wing without cell; one or two medials arising from R-stem, which forks into R_{2+3} and R_{4+5} near apex. Larvæ legless, with two pairs of ocelli; blotch-miners; cutting out a case just before pupation.

Superfamily Gelechioidea. Larva with setæ iv and v closely approximated or on the same tubercle, usually with developed prolegs; pupa obtect, maxillary palpi usually present, but pilifers not marked, and front femora and labial palpi usually connected. Moth with all radial branches running to costa as a rule, and usually all present; \mathbf{R}_4 and \mathbf{R}_5 stalked except in Stenominæ. Hind wing with Sc and R closely approximate at base, separating before end of cell (shortly fused in Blastobasidæ). Head more or less smooth; palpi long and upturned, maxillary palpi minute, folded over base of tongue, which is scaled.

Family 19. Cecophoridæ. Fore wing without stigma; R_2 and Cu_2 well back from end of cell; hind wing at least half as broad as fore wing, with apex of membrane bluntly rounded; R and M_1 well separated at origin; 1st A preserved in both wings.

Family 20. Xylorictidæ. Fore wing without stigma; R_2 and Cu_2 well back from end of cell, R_4 and R_5 often separate, when stalked forking over apex; hind wing broad, not excavated below apex, with R and M_1 stalked.

Family 21. Gelechiidæ. Fore wing without stigma, R_1 and Cu_2 well back from end of cell, R_4 and R_5 stalked or united, and both running to costa; hind wing with R and M_1 closely approximated or stalked, or with wing strongly excavated below apex; 1st A lost in both wings.

Family 22. Blastobasidæ. Fore wing with a stigma between R_1 and R_2 , which are widely separated, veins R_2 to Cu_2 all closely crowded around end of cell. Hind wing with Sc and R very shortly fused near base; lanceolate, and rather narrower than fore wing; R and M_1 well separated at origin.

Family 23. Lavernidæ. Fore wing without stigma, lanceolate, with 1st A variable, sometimes anastomosing with 2d A. Hind wing much narrower than fore wing, narrow-lanceolate to linear, pointed, with R and M₁ strongly approximate at origin, or stalked, and M₂ and Cu₁ usually widely separated.

Superfamily **Yponomeutoidea**. Larva with setæ iv and v variable, pupa obtect in forms with iv and v separate, with maxillary palpi; normally with labial palpi and femora exposed; pilifer not marked. Moth with \mathbf{R}_5 when present usually running to outer margin; maxillary palpi usually small or minute, and porrect; hind wing with **R** and \mathbf{M}_1 variable, **Sc** and **R** approximate at base, and usually connected by a cross vein. Head usually smooth.

Family 24. Yponomeutidæ. Ocelli small or absent; legs typically smoothscaled, venation but little reduced: fore wing usually with all veins separate; hind wing with R and M_1 well separated except in the Cerostoma group, M_1 and M_2 often stalked, 1st A distinct in both wings. Egg of flat type so far as known. Larvæ with beta lower than alpha on prothorax, prespiracular wart of three setæ and iv and v widely separate on abdomen. Pupa obtect, with maxillary palpi exposed, pilifers represented by distinct lobes only in Atteva, and fore femora exposed except in Scythris. (To this family are attached various isolated genera of uncertain position.)

Family 25. Glyphipterygidæ. Ocelli large and conspicuous, maxillary palpi minute, tongue scaled, labial palpi upturned to middle of front or beyond, often beyond vertex; wings exceptionally broad, macro-like in shape, the fringe relatively narrow, and but little widened at anal angle. R_{\star} and R_{\star} usually separate,

Cu, arising close to angle of cell; R and M_1 usually separate in hind wing; 2d A very strongly forked at base. Egg of upright type; larva with front acute, reaching well toward, but not to, vertex; alpha of prothorax farther from middorsal line than beta, abdomen with i nearer the middle line than ii on eighth segment, iv and v close together, legs with a single circle of hooks. Pupa incomplete, with only anterior rows of fine spines on the segments, cremaster rudimentary; maxillary palpi, labial palpi and fore femora exposed; mesothorax extending back in a lobe, nearly cutting the metathorax in two.

Family 26. Heliodinidæ. Palpi very short, drooping (medium sized in Euclemensia); maxillary palpi minute, porrect; tongue strong; tarsi with more or less distinct whorls of bristles, the tibiæ also often with stiff bristles, the hind legs displayed at rest. Larvæ various, not well known, pupæ hardly known.

Family 27. Ægeriidæ. Antennæ often fusiform, normally with a tuft of bristles at apex; wings strong, very narrow, usually more or less transparent. Fore wing with \mathbf{R}_s running to outer margin, anal region much reduced, hind wing with costa bearing a backward-directed keel, with a row of spines interlocking with a row on inner margin of fore wing. Larvæ borers, with high front, hooks of prolegs uniordinal in two transverse rows, iv and v close together, and last spiracle higher than the others; pupa incomplete.

Superfamily Tortricoidea. Larva with set \mathbf{iv} and \mathbf{v} approximate, pupa incomplete, typical, spined dorsally for progression; moth normally with \mathbf{R}_5 running to outer margin, free in lower forms; stalked with \mathbf{R}_4 in higher forms, hind wing with Sc and R as in the preceding superfamily, R and M₁ usually approximate; head and body usually with rough vestiture, that on the head shorter than in the Tineidæ; palpi rough and more or less triangular or porrect; maxillary palpi minute.

Family 28. Tortricidæ. Fore wing with 1st A preserved, Cu_2 arising less than three-fourths way out on cell; larva with biordinal or triordinal hooks; abdomen of pupa with a distinct cremaster, or with setæ on the anal rise.

Family 29. Phaloniidæ. Both wings without 1st A; Cu_2 arising more than three-fourths way out in cell; hind wing with M_1 preserved, usually stalked with R; palpi alike in both sexes. Larva with uniordinal hooks; pupa with end of abdomen obliquely truncate, with irregular spines, without cremaster.

Family 30. Carposinidæ. Both wings without 1st A; Cu_2 arising more than three-fourths way out; hind wing with M_1 and M_3 completely lost; palpi much longer in female than in male. Larva a fruit borer with a complete circle of uniordinal hooks and last spiracles dorsal.

Family 31. Cossidæ. Large, stout, heavy moths, with small palpi, rudimentary tongue, partly hairy vestiture, and short tibial spurs. Wing veins heavy, including R_{4+5} and the dividing veins in the cell, 1st A strong and tubular, from near base. Frenulum sometimes obsolete. Egg sometimes upright; larva stout and strong, with mandibles heavy and turned forward, usually with multiordinal hooks in a complete circle; setæ ii as far apart on ninth abdominal as on other segments. Pupa very heavy, with extremely short maxillæ which meet on middle line; no maxillary palpi, and no cremaster.

‡‡"Pyraloids." Larvæ with only two setæ on prespiracular wart, with iv and v approximated, rarely with secondary or tufted hair (in which case the spiracles are round); pupa

obtect but almost always with distinct maxillary palpi; practically never progressing from the cocoon. Moth normally with firmly and finely scaled wings, with ample anal region in hind wing; usually with three anals in hind wing, but with 1st A almost always lost in fore wing; maxillary palpi of porrect type; tongue scaled at base; no accessory cell.

Superfamily **Pyralidoidea**. Characters as just stated for the Pyraloids.

Family 32. Thyrididæ. Wings heavy and close-scaled, 1st A absent in both wings, all radials separate (variously stalked in exotic species), \mathbf{R}_s to outer margin; hind wing with Sc and R closely parallel beyond end of cell; maxillary palpi obsolete, pilifers large. Larva with two setæ on vii of mesothorax, and i higher than ii on ninth abdominal segment.

Family 33. Pyralididæ. Wings entire, without special scaling along the veins beneath; fore wing with 1st A usually lost. Hind wing with Sc and R very closely approximate, or more often fused, beyond end of cell. Larva with primary sets only, with normal prolegs, usually bearing biordinal or triordinal hooks, pupa with seventh abdominal segment fixed in both sexes, usually smooth; with a deep furrow between ninth and tenth abdominal segments in the few species without maxillary palpi.

Family 34. Pterophoridæ. A series of specialized spatulate scales along Cu and its forks in hind wing; our species with fore wing divided into two, and hind wing into three, feathers; 1st A preserved. Larva usually with secondary or tufted hair, with very long, stem-like prolegs with expanded plantæ. Pupa usually exposed, suspended by the tail, often very hairy and angular; seventh abdominal segment movable in male; without maxillary palpi or groove between ninth and tenth abdominal segments. Cremaster supplemented by a second tuft of spines on venter.

Family 35. Orneodidæ. Both wings deeply cleft into six feathers. Only one anal preserved (1st A?). Larva with uniordinal hooks on prolegs and a single seta vii on mesothorax. Pupa with maxillary palpi concealed; seventh segment fixed. Cremaster simple.

‡‡‡"'Macros." Larvæ with only two setæ on prespiracular wart, with iv and v separate except when there is dense secondary hair; with oval spiracles, and often with dense tufted or secondary hair. Usually external feeders. Pupa obtect, progressing only in the Citheroniidæ, without distinct maxillary palpi or pilifers; moth broad-winged, with 1st A rudimentary or absent in all wings, base of M lost, maxillary palpi of porrect type and almost always rudimentary or lost, tongue not scaled; wings usually firmly scaled, with narrow fringes; accessory cell when present separated by a fully developed vein from discal.

Superfamily Uranioidea. Larva with simple hair; set x i and ii separate, iv and v both well below spiracle, and tending to approximate

each other; hooks on prolegs biordinal, in an elliptical band; one or more additional setæ on prolegs. Pupæ hardly known. Egg of flat type. Moth with a slight chitinization subventrally on first segment of abdomen, representing tympanum; fore wing typically with \mathbf{R}_5 and \mathbf{M}_1 approximate or stalked, in Lacosomidæ with \mathbf{R}_4 and \mathbf{R}_5 stalked and widely separated from \mathbf{R}_3 ; Cu apparently three-branched (trifid). Hind wing with Sc and R sharply diverging from close to base of wing.

Family 36. Epiplemidæ. Larva living practically exposed; with circle of hooks on prolegs broadly interrupted; moth with \mathbf{R}_s stalked with \mathbf{M}_1 , free from \mathbf{R}_s ; moth resting with hind wings rolled about body. Frenulum normal in our species.

Family 37. Lacosomidæ. Larva with hooks of prolegs in a complete ellipse; living in a case, open at both ends; thin-skinned, with chitinized thorax and posterior callosity. Moth with \mathbf{R}_4 and \mathbf{R}_5 widely separated from \mathbf{R}_3 ; frenulum rudimentary in our species; resting position normal. (Position of family doubtful.)

Superfamily **Saturnioidea**. Larva always with fine, usually rudimentary secondary hair; primaries on warts or spines which also bear secondary setæ; iv and v united, i of eighth segment of abdomen usually united in middle line; prolegs with a straight band of biordinal hooks. Egg of flat type. Moth without tympanum, the metathorax not modified; tongue rudimentary in our species; fore wing always with trifid venation; \mathbf{M}_2 closely associated with radial stem, one radial always lost (\mathbf{R}_5 ?), \mathbf{R}_2 - \mathbf{R}_4 stalked and much crowded; hind wing as in the family Lacosomidæ; frenulum lost.

Family 38. Citheroniidæ. First-stage larva usually with primary hair; ninth segment with a mid-dorsal spine, anal plate tuberculate or spined; body spines more or less horn-like, never with long spinules, and strongly unequal in our species. Pupa formed in the ground, active, hard and rough, with flanged segments. Cremaster bifurcate, without hooks. Moth with male antennæ doubly bipectinate halfway to apex; fore wing with M_1 more or less stalked with R_2 -R₄ parallel to M_2 ; hind wing with two anals.

Family 39. Saturniidæ. Larva with dense bristly spines in earlier stages, never with primitive first stage; warts i of eighth segment of abdomen fused into a caudal horn in all our species, ii rarely fused, ii of ninth segment fused only in the Hemileucinæ, which have subequal bristly spines and a smooth anal plate. Pupa almost always in a cocoon, not spinulose; cremaster simple or represented by spines only; abdominal segments often without flanges, and telescoping when dried. Moth with antennæ plumose to apex in male, in all our species, \mathbf{M}_{u} in our species, free from R, typically closely associated with \mathbf{M}_{2} ; 3d A usually rudimentary.

Superfamily Bombycoidea. Larva always with much secondary hair, even on head; with warts, often obscured after first stage; with an additional subdorsal wart; legs as in the Saturnioidea; larva often tending to be flattened. Egg of flat type. Pupa normally with secondary hair, with visible labial palpi. Moth various, without

tympanum, ocelli, or maxillary palpi, with pectinate antennæ; tongue very rudimentary or lost. Hind wing with Sc and R closely parallel from base, diverging before end of cell; R, usually distinct.

Family Bombycidæ. Larva with rudimentary hair; with caudal horn. Moth with Cu apparently 3-branched; \mathbf{R}_4 and \mathbf{R}_5 stalked farthest; no humeral veins; and rudimentary frenulum. Traces of 1st A preserved. (Cultivated only.)

Family 40. Eupterotidæ. Larva with fine dense hair, usually mixed with some spatulate scales, with dorsal hair-pencils. Pupa not hairy; in the ground. Moth with R_2 and R_8 , R_4 and R_8 stalked; Cu apparently 3-branched, and frenulum in our species normal. No humeral veins nor traces of 1st A.

Family 41. Lasiocampidæ. Larva with fine secondary hair, in some exotic species mixed with scales; without slender pencils. Pupa hairy; in a cocoon. Moth with apparently 4-branched cubitus; R_s stalked with M_1 , and R_2 with R_s ; no trace of 1st A. Hind wing with frenulum lost, with two or more humeral veins, and expanded humeral angle, which is exposed in the resting position.

Superfamily Drepanoidea. Larva with some subprimaries but without secondary hair; tubercle iv somewhat higher than v and well separated; prolegs with a band of biordinal hooks, and usually a few outer hooks also, uniordinal in Eudeilinea. Anal legs slightly weakened or absent. Egg flat; pupa thin-shelled, in a cocoon. Imago with tympanum formed of a large, double, subventral chitinization on abdomen, not opening to exterior unless through the pleural suture; thin-winged; **Cu** quadrified in the American species, at least in the hind wing; hind wing with **S**c and **R** separate to beyond end of cell, then sometimes fusing.

Family 42. Thyatiridæ. Larva with all legs. Cu trifid in fore wing; vestiture deep, making the body appear stout. Frenulum knobbed at tip.

Family 43. Drepanidæ. Larva with anal prolegs lost (except in the Indian genus Euchera). Moth with Cu quadrifid in both wings, slender, often with hooked wings. Frenulum weak and normal or lost; humeral angle expanded.

Superfamily Geometroidea. Larva very rarely with secondary hair, but always with ventral subprimaries (at least one on sixth segment of abdomen), setæ iv and v remote, iv higher; prolegs with a band of biordinal hooks, often interrupted by a sucker. Egg of flat type except in Alsophila. Pupa normal without exposed maxillary palpi or labium. Imago with a large tympanic hood at base of abdomen, opening subventrally, below spiracle. No ocelli or maxillary palpi; wings usually thin, hind wing with **Sc** and **R** closely parallel or fused part of length of cell, separated at base and beyond cell, strongly curved or angled at base, usually sending a brace across to base of frenulum.

Family 44. Geometridæ. Characters of the superfamily; Cu trifid, except in Operophtera.

Superfamily Sphingoidea. Larva with many minute secondary hairs, obscuring the primaries, which are single; in first stage with seta v higher than iv; i of eighth segment of abdomen united on middorsal line, usually on a horn. Egg of flat type, usually spherical. Imago heavy-bodied, with very strong wings; no tympanum; fore wing with 3d A a strong tubular vein, running into 2d A; hind wing with Sc and R closely parallel, connected by \mathbf{R}_1 , which is as strong as the other veins. Abdomen almost always spined along posterior edge of segments.

Family 45. Sphingidæ. Characters of the superfamily.

Superfamily Noctuoidea. Larvæ with variable vestiture, often with tufted hair; seta (or wart) iv widely separated from v, higher, sometimes behind spiracle; hooks or prolegs *uniordinal* in a single band. Egg *upright*. Imago with a stretched membrane in metepimeron which is more or less broken up into several sclerites; hood on base of abdomen, either above level of spiracle or enclosing it, occasionally lost; ocelli often present; maxillary palpi present and scaled, but almost always minute; 3d A of fore wing weak, hind wing with Sc and R never divergent from base, rarely connected by a brace-vein to frenulum, never closer together beyond cell than along cell, usually fused for part of length of cell or connected by a eross vein. Cu quadrifid, except in the Notodontidæ, and the western family Dioptidæ, which also always lack the hood.

Family 46. Notodontidæ. Cu trifid; Sc and R of hind wing independent, or connected by a weak cross vein; larva with anal prolegs more or less reduced or modified; almost invariably raised in resting position.

Family 47. Liparidæ. Cu quadrifid, as in the following families; palpi short; tongue rudimentary, ocelli absent; antennæ plumose in male; hood above spiracle. Sc and R of hind wing connected by a cross vein, or touching at a point, more than a third way out on cell. Larva with tufted hair, with two bright-colored dorsal glands on abdomen.

Family 48. Noctuidæ. Palpi various; tongue normally functional; ocelli present (except in Menopsimus), hood usually enclosing spiracle, less often above spiracle or obsolete. Sc and R of hind wing touching at a point, less than a third way out on cell, or shortly fused; larva with either simple or tufted hair; in the latter case with secondary hair also, or with wart iv much lower on seventh abdominal segment than the others, and often obsolete.

Family 49. Agaristidæ. Like the Noctuidæ; antennæ clubbed, hood absent.

Family 50. Arctiidæ. Tongue often weak; ocelli always present; Sc and R fusing for at least a fifth, usually a half of length of cell, but not beyond end of cell; hood above spiracle. Larva with tufted and without secondary hair, the tufts rarely lost in last stage; wart iv of seventh abdominal segment not lower than on others. Two subdorsal warts on mesothorax and metathorax.

Family 51. Lithosiidæ. Similar to the family Arctiidæ, with the ocelli lost; wings smooth-scaled; hood sometimes lost. Larvæ usually with tufted hair much reduced in last stage; but present, at least when young; when well developed

in last stage, with the subdorsal warts of mesothorax and metathorax longitudinally placed.

Family 52. Nolidæ. Ocelli lost; tongue present, weak; fore wings tufted; hind wings with Sc and R fused more than half of length of cell; hood above spiracle. Larva with only 14 legs, with tufted hair; wart iv obsolete.

Family 53. Euchromiidæ. Ocelli present; tongue strong; palpi strong; hind wings with free part of Sc lost, the first developed vein being \mathbf{R} . Hood very large, the abdomen constricted behind it in many exotic species. Diurnal. Larva with tufted hair, with only a single subdorsal wart on mesothorax and metathorax.

Suborder **Rhopalocera**. Butterflies with hind wing much shorter and broader than fore wing; with only a single free radial; fore wing with cubitus straight to base, sometimes with a rudiment of **1st A** arising from it near base; the rest of **1st A** lost. \mathbf{R}_{4+5} of pupa given off from radial stem before the origin of \mathbf{R}_1 , obsolete in imago but often with a trace showing as a short spur or a fold. No jugum or frenulum; humeral angle of hind wing enlarged, usually with a humeral vein. Antennæ more or less obviously clubbed, with the scaleless sensory area often covering the whole club, and rarely extending on the shaft (Feniseca). Tongue and labial palpi always strong; ocelli and maxillary palpi always absent. Egg upright, larva with tubercles **iv** and **v** well separated and both low in first stage, usually obscured in later stages; prolegs typically with triordinal hooks. Our butterflies are all diurnal, and all except Thanaos sleep with the wings raised over the body or outspread.

Superfamily Hesperioidea. (Skippers). Head very broad; front twice as wide as high; antennæ widely separated at base (two to four times their own width), usually with a strong but slender tuft of lashes in front of eye; fore legs with epiphysis, hind tibiæ usually with all spurs; fore wing with all veins present and arising separately from cell; hind wing with humeral vein usually running across from tip of basal thickening (costa) to bend in Sc. Larva with prothorax much smaller (in our species) than head or following segment; head capsule closed ventrally behind base of mouth parts by a small sclerite (gula); prolegs with a complete circle of hooks. Larva always a concealed feeder. Pupa rounded, suspended by a Y-shaped girth in a more or less perfect cocoon; with maxillæ extending out at base to reach eyes.

Family 54. Hesperiidæ. Frenulum and frenulum-hook absent.

Superfamily **Papilionoidea**. (True butterflies). Head narrower, the antennæ separated at their base by about their width or less; front less than twice as wide as high; eyes not lashed; fore legs without epiphysis (except in the Papilionidæ), hind legs with end-spurs only; humeral vein, when present, extending free from **Sc** toward costa, often forked at tip, but not reaching edge of wing. Larva with prothorax not noticeably narrowed, without gula; prolegs with a single band of hooks, or if with a second outer band it is much reduced; suspension girdle of pupa a simple loop or absent; the tongue, in the pupa, not touching eyes.

Family 55. Papilionidæ. Fore leg with epiphysis; head broad; fore wing, in our species, with all radials; M_2 associated with Cu stem (quadrifid), 3d A free, turning down toward inner margin; hind wing with only one anal (in our species). Egg spherical. Larva very stout, with tufted hair in first stage; with osmeteria. Pupa girt, but loosely, the anterior end with two points.

Family 56. Pieridæ. Fore leg fully developed in both sexes, with claws and pulvillus, but without epiphysis, head narrower, but with the antennal sockets not encroaching on the eyes; fore wing with one or two radials lost in our species, except in Zegris, M_3 stalked with R-stem, M_2 associated with R-stem in both wings, 3d A of fore wing rudimentary, running up into 2d A; hard wing with two anals. Larva with fine secondary hair, slender and normal in form. Pupa girt loosely, angular, ending in a single spine.

Family 57. Lycænidæ. Fore leg nearly fully developed in male, with spinules on tarsus, but without normal terminal claws and pulvilli, wholly normal in female; head very narrow, the antennal sockets encroaching more or less on the eyes; front not depressed. Fore wing with one or two radials lost in our species, \mathbf{M}_1 usually free; \mathbf{M}_2 arising from the cross vein halfway between \mathbf{M}_1 and \mathbf{M}_3 , often weak, the cross veins both weak. \mathbf{M}_2 of hind wing as in fore wing; humeral lost in our species; anals as in the Pieridæ. Egg flat. Laiva slug-like with retractile head, and fine secondary hair. Pupa short and rounded, closely girt.

Family 58. Erycinidæ. Fore leg quite reduced and brushlike in male; without claws or spines, with tibia expanded in a spine beyond articulation of tarsus; normal in female; head as in the Lycænidæ; venation as in the Lycænidæ, but with humeral vein preserved and our species with costa thickened out to humeral angle. Egg and larva and pupa of our species as in the Lycænidæ.

Family 59. Nymphalidæ. Fore leg reduced and nonfunctional in both sexes (except female of Libythea); head as in Pieridæ, front depressed; fore wing in our species with all radials, M_1 free, M_2 associated with R-stem in both wings; hind wing usually with humeral vein; anals as in the Pieridæ. Egg as high as wide, vertically ribbed; larva slender, with spines or fleshy filaments, or forked tail; rarely as in Pieridæ; pupa suspended by the tail only, or (in a few Satyrinæ) in a slight cocoon.

ARTIFICIAL KEY TO THE FAMILIES OF LEPIDOPTERA6

Imago



FIG. 30. PORTION OF WING MEMBRANE SHOWING SOCKETS OF SCALES AND WING SPINULES (ACULEÆ).HIGHLY MAGNIFIED

1.	Winged
1.	Wingless or with rudimentary wings
2.	Hind wings with four or five radials, with at least ten veins besides anals
	(figs. 31, 40), wing-membrane spinulated (fig. 30)
2.	Hind wings with only one free radial (two in the otherwise much reduced
	Douglasia group, page 224); with at most six (or, with Sc, seven) veins
	from cell
3.	Wings hardly wider than their fringe, expanse about one-half inch4
3.	Wings ample, fringe narrow, expanse over one inchHepialidæ (p. 66)
4.	Middle tibiæ with a spur; mouth parts formed for sucking; Sc of fore wing
	forking near apex
4.	forking near apexEriocraniidæ (p. 64) Middle tibiæ unarmed, mouth parts formed for biting; Sc forking near mid- dle
	dleMicropterygidæ (p. 62)
5.	Each wing deeply cleft into six narrow stripsOrneodidæ (p. 652)
5.	Fore wing moderately cleft into two, and hind wing deeply, into three, feathers
	(figs. 400–403)Pterophoridæ (p. 639)
5.	Wings entire, or one pair only, moderately cleft
6.	Inner margin of fore wing and costal margin of hind wing with series of
	recurved spines and interlocking; fore wing at least four times as long as wide,
	and base, at least, of hind wing transparent (figs. 224-229) Ægeriidæ (p. 360)
6.	Wings not interlocking at middle of margin, very rarely transparent, and, if
	so, with broader fore wings7
7.	Hind wing lanceolate, without marked anal angle, or notched below apex and
	trapezoidal, or cleft; the fringe almost as wide as wing, or wider. (Micros
_	in part)
7.	Hind wings much broader than their fringe, never lanceolate, and rarely
~	trapezoidal with produced apex8
8.	A double series of enlarged and divergent scales along Cu of hind wing below;
0	wings, body, and legs, very longPterophoridæ (Agdistinæ)
ð.	No such specialized scales
9.	Fore wing with two anal veins well developed at middle of wing or at outer
•	margin, the first a tubular vein at middle of wing in broad-winged forms,
	sometimes tubular only at the margin in narrow ones

•In the case of extra-limital families, no page is indicated; in those not included in this memoir, page references are made to the brief family definitions in the synopsis, pages 41 to 44.

9.	Fore wing with only one anal reaching margin, 1st A rudimentary, or repre-
10	sented by a fold; $3d A$ at most by a short spur
10.	Castniidæ
10.	Antennæ tapering regularly, or very slightly fusiform
11.	Sc separating from cell shortly before the apex of the cell (figs $62, 72)$ 16
11.	Sc arising separate from R, running closely parallel to it to well beyond end
	of cell, or (in our species) fused with it beyond end of cell; the base of R
	in that case either complete or showing as a short spur (fig. 298) Pyralididæ (Schænobiinæ) (p. 525)
11.	Sc anastomosing with cell for short distance or not at all; not closely parallel
	to R beyond end of cell
12.	Accessory cell well marked
12.	Accessory cell absent
13.	wings ample (fore wing not half longer than wide), body short and slender;
13.	mouth parts rudimentary
	parts rudimentary
13.	Wings more or less oblong, usually twice as long as wide, and lightly veined;
	body small and slender; mouth parts usually developed, with scaled
14	tongue
14.	wing (fig. 101)
14.	Fore wing with 1st A and 2d A independent beyond extreme base of wings15
	Fore wing with \mathbf{R}_{s} running to outer margin, base M distinct, running through
	center of cell; Sc and R of hind wing connected by a strong cross vein or
15	anastomosing; tongue absentEucleidæ (p. 102) Sc and R of hind wing independent; or connected by a cross vein, when R_s runs
19.	to costa (Ethmia); or anastomosing when base of media of fore wing
	runs near lower edge of cell or is completely lost (Tortricidæ); tongue usually
	runs near lower edge of cell or is completely lost (Tortricidæ); tongue usually present
16.	\mathbf{R}_{s} long-stalked (fig. 72), colors light, the northern species with crinkly hair
16	on fore wing; tongue absent
10.	Pyromorphidæ (p. 113)
	Hind wing with three anals, the first often fading out toward base (fig. 19)18
17.	Hind wing with two anals or less; at most with a short spur of 1st A at
18	margin in broad-winged forms (figs. 409-432)
10.	Sc and R of hind wing closely parallel or fused beyond end of cell (figs. 312- 399Pyralididæ in part (p. 523)
18.	Sc and R strongly divergent from before end of cellMicros in part
19.	Antennæ distinctly swollen toward tip, (figs. 5-7), and frenulum wanting
10	(butterflics)
10.	wings with a strong frenulum
20.	Fore wing with all veins present, from cell, eyes strongly lashed in front;
	antennæ separated at base by a distance greater than half width of
00	eyes
20.	closer together 91
21.	Fore wing with 3d A free at tip, hind wing with only one anal, save in one
	closer together
21.	Hind wing with two well-developed anals; 3d A of fore wing running into
	2d A or lost
	ern species, with ten or eleven veins

22.	M_2 distinctly associated with radial stem, in one, and usually in both wings; lower discocellular vcin often obsolete; with at least a trace of a humeral
ດາ	A humeral vein in hind wing
Z3.	A numeral vent in find wing
23.	No humeral vein
24.	Butterfly walking on four legs (except female of Hypatus), radius hve-
	branched: M. from cell
24.	Butterfly using all its legs for walking; radius usually four-branched, M,
	stalked with itPieridæ (p. 44)
25.	lorth American species very stout, and with wings 55 mm. (two inches) or more in expanse; the hind wings rarely reaching beyond middle of
	or more in expanse: the hind wings rarely reaching beyond middle of
	abdomen; Sc and R of hind wing connected at the middle of the cell or rather
	before by a vein (\mathbf{R}_1) which is as strong as any; and then closely parallel
	to end of cell or beyond
95	Wings proportionately larger; Sc and R rarely connected by a strong cross
40.	vein, and if so, strongly divergent beyond it (fig. 426)
96	veni, and it so, strongly divergent beyond it (ng. 420)
Z 0.	Sc and R separate, but connected by a more or less distinct cross vein; acces-
	sory cell fused with discal cell, but with the line of separation (\mathbf{R}_{i+5}) indi-
	cated by a slight thickening starting from an angulation in the stem of
~ ~	R; species under 30 mm. in expanse (fig. 155) A few Micros59
26.	Accessory cell separated by a full-sized vein, or completely absent27
27.	Cu of fore wing apparently three-branched (in a couple of Lithosians two-
	branched)
27.	branched)
28.	Frenulum normal
28.	Frenulum normal
	absent
29.	Sc and R fused from base of hind wing beyond middle, then rapidly diverging;
	swollen at the base; slender mothsLithosiidæ in part (p. 42)
29.	Sc and R separate at extreme base; then closely approximate or fused a
	greater or less distance
29.	greater or less distance
30.	Hind wing with Sc angled near base, connected by a strong cross vein to
	humeral angle
30.	Sc of hind wing moderately curved or straight at base
31.	Hind wing with Cu apparently 4-branched, M. being much nearer M. than M.:
	Sc and R closely parallel beyond end of cell; vestiture deep (fig. 432).
	Thyatiridæ (p. 686)
31.	Hind wing with Cu apparently 3-branched, M_2 being as near to M_1 as to M_3 ;
	Sc and R separating before end of cell (fig. 425)
32.	Stout species; no tympanic hood; 1st A lost
32.	Usually slender species: when rather stout, with a well-marked lateral bood
	on first segment of abdomen
33.	Tongue absent; fore wing with R_2 and R_3 , R_4 and R_5 stalked together (fig. 425);
	northern species with hyaline dots on fore wing
	Eupterotidæ (Apatelodes) (p. 678)
33	Tongue present, often weak; fore wing fully scaled; usually with accessory
00.	cell, or R ₃ and R ₄ long-stalked togetherNotodontidæ (p. 42, 678)
34	Subcosta straight to base and but little swollen; no trace of a tympanic hood.
04.	Dioptidæ.
34	Subcosta strongly sinuous and much swollen at base; a lateral hood at base
94. •	of abdomen
35	Sc and \mathbf{R} of hind wing fused for a very short distance, then sharply divergent,
55.	separate from base, or connected by a weak cross vein (figs. 413-422);
	tympanic hood absent
	VIIIDanic mood abschu

	Sc strongly divergent from R at extreme base, then sharply bent and touching, fusing or closely parallel to it, or connected by a strong cross vein; tympanic hood conspicuous, lateral
20.	Antenna lacely as led as upon side (for 0 10)
30.	Antennæ closely scaled on upper side (figs. 9, 10)
	M. stalked with R in both wings or neither; hind wing usually with one anal (figs. 413-416); male antennæ pectinate to apexSaturniidæ (p. 668)
37.	M_1 stalked with R in fore wing but not in hind wing; hind wing with two anals
	(figs. 419, 422); male antennæ pectinate halfway to apex.
	Citheroniidæ (p. 664)
38.	Sc of hind wing sharply divergent from R from close to base (fig. 411)39
38.	Sc and R parallel at base, connected by a weak crossvein (fig. 426)41
39.	R_{4+5} widely separated from R_s all the way from cell to margin (fig. 411). Lacosomidæ (p. 656)
39.	\mathbf{R}_{s} arising from cell closely associated with \mathbf{R}_{s}
40.	\mathbf{R}_{s} and \mathbf{M}_{s} stalked or closely approximate at base, and separate from \mathbf{R}_{s} (like
	fig. 409)Uraniidæ
40.	\mathbf{R}_{5} separate from \mathbf{M}_{1} (like fig. 414)Lonomiidæ
41.	R_s separate from M_1 (like fig. 414) Lonomiidæ Frenulum about one-sixteenth length of hind wing; four radial veins in fore
	wingÉupterotidæ (Eupterotinæ) Frenulum obsolescent, not exceeding l meral angle, or absent, 5 radials.
41.	Frenulum obsolescent, not exceeding lumeral angle, or absent, 5 radials.
	Bombycidæ (p. 679)
42.	Cu, of fore wing arising from cell about a third way out from base, or even
	nearer base; \mathbf{R}_s stalked with \mathbf{M}_1 ; with humeral veins and without frenulum
10	in the North American species (fig. 427)Lasiocampidæ (p. 679)
42.	Cu ₂ of fore wing arising well beyond middle of cell; hind wing usually with
12	frenulum
40.	cubitals arising separately, or with R_2 and R_3 shortly stalked (fig. 195).
	Thyrididæ (p. 521)
43.	R ₃ and R ₄ , or R ₄ and R ₅ long-stalked, or with some veins absent44
44.	Sc and R of hind wing parallel to beyond separation of R from cell, and then
	approaching very close or fusing a short distance (figs. 306, 435)
44.	Sc fusing to upper side of cell or wholly independent46
45.	First segment of abdomen with chitinized subventral bullæ, and a subdorsal
	cavity opening backward; northeastern species expanding over 25 mm.
45	Drepanidæ (p 688)
40.	First segment of abdomen not modified; northeastern species expanding less than 20mm
48	is superant we have a second at extreme base with P
10.	Euchromiidæ (p. 43)
46.	Sc and R separating before end of cell
47.	Antennæ swollen toward tip
47.	Shaft of antennæ regularly tapering
48.	Ocelli present (fig. 3)
48.	Ocelli absent
49.	Sc and R of hind wing fused to middle of cell or beyond. Most Arctifice (p. 42)
49.	Sc and R fused for more than a fifth length of cell, but the fusion not reaching middle
49.	Sc and R fused for less than a fifth length of cell, the fusion sometimes imper-
50	fect
50.	Hind tarsus ordinarily not more than eight times as long as thick, tibia often with reduced spure. M reduced only in Eulephania in the rest of full strength
	with reduced spurs; M, reduced only in Eubaphe; in the rest of full strength and associated with cubital stem; moths often stont; Sc very much swollen
	at base; hood above spiracle
	and above sphare sphare (p. 42)

⁷Also most tropica lThyrididæ.

5	i0.	Hind tarsus ordinarily much more slender, the tibia with long spurs; M_2 usually well separated from cubital stem, though nearer it than radial, and often weaker than the other veins; Sc not more than twice as thick as R in their basal portion; usually slender moths; hood surrounding spiracle (fig. 16), except in some slender speciesSome Noctuidæ (p. 42)
		Tympanic hoods enlarged dorsally, showing from dorsal side as two rounded bosses on the first segment of the abdomen, separated by a third of the width of the abdomen; brilliantly marked speciesPericopidæ
5	51.	Tympanic hoods less conspicuous dorsally; separated by half the width of the abdomen
5	52.	White or yellow species with palpi not reaching the middle of the smooth- scaled front, and four-branched Cu in both wings; hood above spiracle, con- spicuous
5	52.	Species with longer palpi, three-branched Cu in hind wings or gray ground color; and lateral hood
5	53.	Fore wing with raised scale-tufts; small species with Sc and R ordinarily fused to near middle of cell but free at base
5	3.	Fore wings smoothly scaled
5	5 4 .	Fore wings smoothly scaled
5	54.	Sc and R fused from base to middle of cell. Most Lithosiidæ and Menopsimus (Noctuidæ) (p. 42)
5	54.	Sc sharply divergent from R at base, then angulate and becoming closely
ŧ	55.	approximate or fusing with itA few Geometridæ (p. 41) Legs lost, moth never leaving cocoonPsychidæ (?s in part) (p. 140)
5	55.	With normal legs
Ē	00.	Cocoon seedlike, with a valve at one end (being formed of the larval case), the moth normally not leaving it; moth less than 6 mm. long.
£	56.	Psychidæ ($\hat{\mathbb{Q}}$ s in part) (p. 140) Cocoon normally felted of the larval hair, or rudimentary and underground;
E	57.	moth more than 6 mm. long
5	57.	Ocelli absent
5	58.	Ocelli absent
ŧ	58.	Abdomen smoothly clothed with fine light woolly hair; moth not normally leaving the cocoon, which is composed of the larval hair.
		Liparidæ (a few \Im s) (p. 42)
5	59.	Maxillary palpi conspicuous, folded in resting position (fig. 49)60
5	59.	Maxillary palpi straight and porrect (fig. 3), or rudimentary
6	30.	Antenna with large eye-cap, larger than eye
6	31.	Fore wing with branched veins, normally with a small, four-sided cell (figs. 52-57)
6	81	Fore wing with three or four simple veins only (fig. 113). Opostegidæ (p. 16)
		Head entirely smooth, \mathbf{R}_{5} , when distinct, running to costa (fig. 114). Oinophilidæ
(62.	Head with a few erect hairs; R ₅ present and running to outer margin (fig. 202)
	62	Head tufted at least on vertex: R. running to costa
(63.	Head tufted, at least on vertex; R_5 running to costa
		Most Incurvariidæ (p. 72)

	Wing membrane not aculeate; \mathbf{R}_i rarely as strong as the other veins, and when distinct separated from the base of the wing by several times its length
	(fig 73)
64.	margin (figs. 123, 175)
65.	long and longitudinal; rarely obsolete (figs. 129, 185)
65.	Hind wing linear, R and M, connate or stalked
66.	Fore tibia exceptionally slender, with epiphysis rudimentary, at its apex, or
66.	absent; antenna turned forward in reposeColeophoridæ (p. 202) Fore tibia usually stout, the epiphysis conspicuous and situated at its middle;
67.	antennæ folded back in repose
	duced apex; rarely bifid (figs. 156, 160, etc.)
01.	Hind wing rounded at apex or trapezoidal, often broader than its fringe; anal region developed (fig. 138 etc.) 68
67.	region developed (fig. 138, etc.)
68	fringe (fig. 180)
00.	Carposinidæ (p. 513)
68.	Carposinidæ (p. 513) Hind wing with at most one vein lost
69.	usually stout; venation complete, with base of media preserved
	Vestiture of thorax and fore and middle tibiæ mostly of normal scales; body
	usually slender
70.	Palpi upturned beyond middle of front, usually beyond vertex, or long and porrect; with very long first joint; eyes usually hairy.
	Tineidæ (Acrolophinæ) (p. 116)
	Palpi small, not reaching middle of front; eyes nakedCossidæ (p. 516)
11.	Cu_2 arising less than three-fourths way out on cell (fig. 234); palpus beak-like with second joint rough and usually porrect or oblique, and third small (figs. 241 to 243, and 271 to 283, inclusive)Tortricidæ (p. 376)
71	Cu ₂ arising more than three-fourths way out on cell or (rarely) palpus smooth
	and upturned beyond vertex (fig. 147)
72.	Palpus rough-scaled and beak-like, with second segment roughly scaled on
	upper side, and third porrect, inconspicuous; 1st \mathbf{A} lost; \mathbf{R} and \mathbf{M}_1 closely
72.	approximate or stalked (fig. 288)
	mentary; female (so far as known) with abdomen heavily tufted at apex;
	R and M_1 normally separate but strongly divergent (figs. 103, 104)73
	Palpus bristled on outer side of second segment, sometimes with a mass of bristles, third segment ovoid, well set off, sometimes porrect; R and M_1 nor-
	mally separate
	usually tapering
73.	Male antennæ simple or bristled in American species, female with abdominal
73	tuftPsychidæ (p. 140) Male antennæ plumose; female unknownEpipyropidæ
74.	Wing membrane not spinulated; antennæ shorter than fore wing.
	Tineidæ (p. 116)
74.	Wing membrane spinulated; antennæ of male much larger than fore wing; ovipositor chitinized, piercingIncurvariidæ (p. 72)

75. R and \mathbf{M}_{i} of hind wing closely approximated or stalked; tongue usually present (for 159)
(fig. 152)
1st A usually preserved at margin
rate, R _s running to outer margin, 1st A lost; maxillary palpi absent.
Thyrididæ (p. 521)
76. Body and wings light, and soft-scaled; maxillary palpi perceptible, folded77
77. R, and R ₅ stalked or united, both running to costa (fig. 155); 1st A lost.
Gelechiidæ (p. 255) 77. \mathbf{R}_{s} and \mathbf{R}_{s} separate, or forking over apex, all veins present; 1st A strong
78. Fore wing blunt. maxillary palpi of folded typeXylorictidæ (p. 250)
78. Fore wing falcate, maxillary palpi minute, but porrect.
Yponomeutoidea (Cerostoma) (p. 341)
79. R ₅ running to costa (fig. 141)
from cell
from cell
Ecophoridæ (p. 230) 80. \mathbf{R}_{s} and \mathbf{R}_{s} separate (fig. 200), or ocelli very largeYponomeutoidea (p. 335)
80. \mathbf{R}_4 and \mathbf{R}_5 separate (ng. 200), of ocean very narge
81. Antennæ with rudimentary eye-cap or none
81. Antennæ with rudimentary eye-cap or none
82. Palpi reaching beyond middle of frontA few Gracilariidæ (p. 161) 83. Head with long, bristly vestiture covering vertex and face; tongue and maxil-
lary palpi absent
83. Face at least smooth-scaled
84. Hind wing with radial stem central, sending off a branch to costa near middle
(\mathbf{R}_{2+3}) and one to dorsal margin nearer apex (fig. 131). Douglasiidæ (p. 224)
84. R-stem not sending a branch to costa, or with the branch close to apex (fig
181, etc.)
85. Hind wing lanceolate, at least a sixth as wide as long, with the R -stem running through its center (fig. 134)
85. Hind wing with \mathbf{R} -stem closely associated with Sc toward base, or lost; or wing
linear with veins crowded (figs. 180, 186)
86. Maxillary palpi distinct, porrect
86. Maxillary palpi obsolete
palpi usually upturned beyond middle of frontCycnodiidæ (p. 218)
87. Cu-stem of hind wing simple, free; palpi small, drooping (fig. 134).
Heliozelidæ (p. 225) 88. Fore wing linear, with three or four simple veins only (fig. 221).
Heliodinidæ (Cycloplasis) (p. 356)
88. Fore wing lanceolate with branched veins but no cell (fig. 133).
Heliozelidæ (Coptodisca) (p. 225) 88. Fore wing with formed cell
89. Fore wing with only four veins running to costa, and five or six to inner margin
stalked with M_2 Some Yponomeutoidea (p. 335)
89. Fore wing with five veins running to costa or only three or four to inner margin R and M ₁ approximate or stalked to hind wing90
90. Vertex rough or with a rough anterior crest
90. Entire head smooth-scaled Lavernidæ (p. 318)
91. Accessory cell small or more often absent (figs. 115-120) hind tibiæ less hairy often smooth or bristledGracilariidæ (p. 161)
or on one of or

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Larva⁸

1.	Thoracic legs wanting or reduced to fleshy swellings, without chitinized seg-
	ments
1.	Thoracic legs developed, with chitinous segments
	from the vertex by the epicrania)Incurvariidæ (Prodoxus) (p. 72)
2.	Body cylindrical or flattened; when somewhat fusiform with front reaching
	vertex
3.	Head with six small ocelli on each side
3.	Head with two ocelli on each side, front not reaching vertex. Heliozelidæ (p. 225)
3.	Head with a single large ocellus on each side, or none
4.	Abdomen with rudimentary ventral prolegs on third to sixth segments, bear-
	ing hooks
4.	Abdomen without prolegs on sixth segmentGracilariidæ (p. 161)
b.	Ocellus frontal, front triangularEriocraniidæ (p. 64)
0. 6	Ocellus lateral, front quadrangular
•••	fifth segments of abdomen when presentGracilariidæ (p. 161)
6.	Front widest at anterior end, body cylindrical; rudimentary prolegs on second
-	to seventh segments of abdomen or none7
7.	Body normal, five times as long as thick, normally with prolegs.
7	Nepticulidæ (p. 79) Body very slender, ten times as long as thick, without prolegs.
••	Opostegidæ (p. 160)
8.	Setæ replaced by large, ovate scales, arranged in pairsMicropterygidæ (p. 62)
8.	Setæ normal9
9.	Crochets wanting, prolegs rudimentary or absent
	Prolegs at least represented by rudimentary crochets
10.	Front not extending to vertex (except when vertex is represented by a very
	narrow slit)
11.	Head completely retracted, body frequently with spines or secondary hair, the primary setæ obsolete; body with obscure incisures, usually with con-
	the primary setæ obsolete; body with obscure incisures, usually with con-
11	spicuous pitsEucleidæ (p. 102) Head normally exposed; body with primary setæ only, which are usually dis-
	tinct: with strong incisures
12.	tinct; with strong incisures
	(ng, 51) $(ngeneration for a first f$
12.	Setæ iv and v adjacent (fig 174); prolegs absentA few Gelechiidæ (p. 255)
10.	Body without secondary or tufted setæ, tubercle vi single; vii of three setæ at most, unless the proleg has a multiserial circle of hooks, when it may
	have four setæ; sometimes with a couple of dorsal subprimary setæ14
13.	Body with tufted or secondary hair; at least two setse on tubercle vi on sixth
	segment of abdomen, or with additional set x on proleg
14.	Sixth segment of abdomen without a prolegGracilariidæ (p. 161)
14.	Sixth segment of abdomen with a proleg, the more anterior segments some-
15.	times without
	transverse bands

^{*}Modified from the key in The Classification of Lepidopterous Larva, by S. B. Fracker Illinois Biological Monographs, volume 2, number 1.

15.	Hooks of prolegs in a single band, sometimes with a few rudimentary outer
	nooks besides
16.	hooks besides
16.	Prespiracular wart of prothorax with three set (fig. 174)
17.	Hooks uniordinal (fig. 25), body cylindrical, vii of mesothorax with a single
	seta
17.	Hooks biordinal or triordinal (figs. 26, 27), or uniordinal in larve with stout
	fusiform body and bisetose tubercle vii of mesothorax Pyralididæ (p. 523)
10	Hooks of prolegs arranged in two transverse bands (fig. 107)
10.	House in a since or alling a construct branch intermeted (for 40, 022) of
10.	Hooks in a circle or ellipse, sometimes broadly interrupted (figs. 42, 233)24
19.	Prolegs with two simple series of nooks
19.	Prolegs with a single series of hooks, or with two bands formed of several
	Prolegs with two simple series of hooks
20.	Setæ iv and v of abdomen remoteLyonetiidæ (Bucculatrix) (p. 155)
20.	Setæ iv and v of abdomen adjacent (fig. 319)
21.	Setæ iv and v of abdomen adjacent (fig. 319)
21.	Hooks of anal prolegs in a single series
22	Front extending about one-third way to vertex Cossidæ (Cossula) (p. 5]6)
20	Front extending at least two thirds way to vertex
<u>.</u>	Front extending at least two-thirds way to vertex
20.	spiracies empirical, normal in size, those of eighth segment of abdomen ingher
-	than the others
23.	Spiracles very small, circular, the last pair about in line. Coleophoridæ (p. 202)
24.	Setæ iv and v of abdomen remote, or, in a few minute species, v absent25
24.	Setæ iv and v adjacent, often on a common tubercle, no small hooks at base
	of principal series on prolegs
25.	Hooks arranged in a single complete ellipse
25.	Hooks arranged in a single complete ellipse
	at the base of the normal ones (fig. 28) 28
26.	Prothorax with setæ of prespiracular group about as far from spiracle as
	from each other; seta i on abdomen higher than iiLyonetiidæ (p. 148)
26.	Prothorax with setæ of prespiracular group about twice as far from spiracle
H 0.	as from each other
97	Sata i of abdoman much lower than ii (fig 02) Tinsida (n 116)
07	Seta i of abdomen, much lower than if (ig. 52)
41.	as from each other
28.	Mesothorax and metathorax with seta ia in front of ib and well separated,
~~	abdomen with iv above level of spiracle (fig. 42)
28.	Mesothorax and metathorax with setæ ia and ib closely associated; abdomen
	with iv below level of spiracle
29.	with iv below level of spiracle
	Tineidæ (Acrolophinæ) (p. 116)
29.	Prothorax with seta beta lower than alpha
30.	Last pair of spiracles dorsal, closer together on middle line than setze i of
	anterior abdominal segments (fig. 293)Carposinidæ (p. 513)
30.	Last pair of spiracles nearly normal
31	Mesothorax with two setse vii (above base of leg)
31	Mesotheray with a single seta vii ninth addominal segment with set
	higher then i
20	higher than i
04.	actor i bishar than ii
90	seta i higher than ii
02.	Fromozene spirache with long axis norizontal
33.	Setæ ii of ninth abdominal segment closer together than on any other, fre-
	quently on the same plate (fig. 290)
33.	Seta 11 of ninth abdominal segment as far apart as on the other segments,
	very rarely (Leuzera) on the same plate
34.	Abdomen with setse iv and v practically horizontally placed; hooks of pro-
	very rarely (Zeuzera) on the same plate

34.	Abdomen with setae iv and v in a diagonal or vertical line; hooks usually multiordinal (fig. 27)
35.	multiordinal (fig. 27)Tortricidæ (p. 376) Coxæ of metathoracic legs twice as far apart as wide; prolegs small; small
~ "	species (fig. 194)Lavernidæ (p. 318) Coxæ of metathoracic legs closer together
35.	Coxæ of metathoracic legs closer together
36.	Setæ i and ii adjacent on abdominal segments.
	Heliodinidæ (Schreckensteinia) (p. 356)
36.	Setæ i and ii widely separated
37.	Front reaching less than half way to vertex (about half way in some very
	large species with horizontal head and triordinal hooks)
37.	Front reaching two-thirds way to vertex or a little shorter, and ending in an
	attenuate point; small species with uniordinal or biordinal hooks39
38.	Borers: abdomen with set x is and y on separate tubercles on the ninth seg-
00.	Borers; abdomen with setæ iv and v on separate tubercles on the ninth seg- ment (fig. 296)Cossidæ (p. 516) Leaf feeders; abdomen with setæ iv and v on the same tubercle on the ninth
38	Leaf feeders: abdomen with set x is and y on the same tubercle on the ninth
50.	segment
20	Hooks of prolegs biordinal
90. 90	Hooks of prolegs uniordinal
40	Second, third, and fourth ocelli grouped together, more widely separated from
40.	Second, tinru, and tourth ocent grouped together, more where separated from
	first and lower (fig. 149) Ecophoridæ (p. 230)
40.	Ocelli evenly spaced ⁹ Gelechiidæ (p. 255) Abdomen with iii farther back than spiracle on eighth segment ⁹ .
41.	Abdomen with in farther back than spiracle on eighth segment.
	Blastobasidæ (p. 308)
41.	Abdomen with seta iii not farther caudad than spiracle.
	Glyphipterygidæ (p. 350)
42.	Prespiracular wart on prothorax with three setæ (figs. 150, 151)43
42.	Prespiracular wart on prothorax with two setæ
43.	Set a iv and v of abdomen remote: or, if approximate, set a beta much closer
	together than set alpha on prothorax (like fig. 210) and prolegs long and
	slender
43.	slender
	usually short (p. 244)
44.	Tubercle vii on mesothorax and metathorax with two setæ
44.	Tubercle vii on mesothorax and metathorax with a single seta47
45.	Setæ minute, tubercles reduced to obscure rings, head unusually wide; and
	prolegs reducedThyatiridæ (p. 686)
45.	Setæ minute, tubercles reduced to obscure rings, head unusually wide; and prolegs reduced
46.	Tubicle in of abdomen with two setæLithosiidæ (p. 42)
46.	Seta iii of abdomen singleArctiidæ (Utetheisa) (p. 42)
47.	North American species with enlarged, contrasting tubercles: and a hump
	on eighth segment of abdomen; contrastingly striped transversely (or spotted) with black
	spotted) with black
47.	Caterpillars of other types. Noctuidæ (p. 42)
48.	Caterpillars of other types
	first pair of ventrals much reduced 49
48.	first pair of ventrals much reduced
	reduced
48	reduced
40	Hair tufted; hooks of prolegs uniordinal; 14 legs
40	With a few subprimery being only (in come systic applied with the state of the stat
чΰ.	. With a few subprimary hairs only (in some exotic species with fine secondary
	hair), sometimes with only a single subventral subprimary on sixth segment of abdomen; usually 10 legs
50	And prolong wholly lost
50.	Anal prolegs wholly lost
50.	nar profess represented by a pair of large tubercles, or flagella at least,
	normally fully developed

[&]quot;These characters are said to be inconstant but none are better known to me.

52. 52.	Hooks of prolegs uniordinal (fig. 25)
53.	Anal plate bifurcated, head roughly papillose; third ocellus very large. Nymphalidæ (Satyrinæ) (p. 44)
54.	Anal plate simple; head smoother; third ocellus rarely much enlarged54 Larva parasitic, with hemispherical body and a complete circle of uniordinal hooks
54.55.	Larva more normal in shape, not parasitic
55.	expanded plantaPterophoridæ (p. 639) Spiracles elliptical, larger; ventral prolegs short56
56.	Secondary setæ dense
56.	Secondary setæ dense
57.	few subprimaries
57.	Notch acute, with convergent sides; anal prolegs much reduced and not used;
58.	warts rudimentary and dominated by a single hair (Melalopha) or absent (Datana)
	Lipandæ (Doa) Tubercle iv much lower on seventh than on other segments; anal prolegs more
50	or less reduced or modified
50	Skin shagtened
99. 60	Skin not shagreened
60.	No eversible dorsal glands on two segments of abdomenLipatidae (p. 42)
61.	No eversible dorsal glands
61.	Spiracles elliptical, normal in size
62.	Ventral prolegs short, with a straight band of heavy hooks.
62.	Ventral prolegs slender, with an expanded planta sometimes bearing a circle of hooksPterophoridæ (p. 639) Mesothorax with only a single large wart above level of spiracles.
63.	Mesothorax with only a single large wart above level of spiracles. Euchromiidæ (p. 43)
63	Mesothorax with two or three warts above level of spiracles
64.	Wart (or seta) iv at about the same level on seventh abdominal segment
64.	as on the sixth and eighthArctiidæ (p. 42) Wart (or seta) iv much lower on seventh segment or absent (perhaps fused
65.	Wart (or seta) iv much lower on seventh segment or absent (perhaps fused with v)
	row \dots A few Noctuldae (p. 42)
66.	Hooks of prolegs abruptly decreasing in size near each endPericopidæ Body without general secondary hair, often with a few subprimaries; with
66.	not more than eight hairs on prolegs
	always well developed
67. 67.	Hooks in a complete circle (fig. 412)
•	much weaker band on outer side70
05.	Subdorsal setæ of abdomen represented by warts. Yponomeutidæ (Scythris) (p. 349)

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68. 69.	Subdorsal setæ of abdomen simple
69.	Head smooth, body widest at first segment of abdomen. Xylorictidæ (Ptochoryctis) (p. 250)
70	Prespiracular wart of prothorax with three setæ; setæ iv and v of abdomen
10.	respiratural wait of protocax with three sets, sets it and to abduce a province (for 151) (p. 244)
70	approximate (fig. 151) Geophoridæ (Ethmia) (p. 244) Prespiracular wart of prothorax with two setæ; setæ iv and v of abdomen
10.	respiratural wait of protociax with two sets, sets it and t of abdomin
71	usually distant
71	Spiracles subequal
	Setæ very irregular in length, some ten times as long as others; with obscure
12.	warts, at least in younger stages, sometimes with spatulate scale-hairs73
72	Setæ subequal or supplemented by prominent warts or by spines
73	Labrum notched two-thirds of its depth, or with the notch somewhat shallower
10.	and continued as a groove to the base of the labrum; North American
	species with small dorsal hair pencilsEupterotidæ (Apatelodes) (p. 678)
73.	Labrum less deeply notched, the notch not continued by a groove; no dorsal
	hair pencilsLasiocampidæ (p. 679)
74.	hair pencilsLasiocampidæ (p. 679) Eighth segment of abdomen with a mid-dorsal horn, plate, or tubercle75
74.	Eighth segment of abdomen not armed in mid-dorsal line
	Body with numerous branching spines or enlarged tubercles
	Body with at most two pairs of small spines on thorax
	Head angulated or spined dorsally, or abdomen with several mid-dorsal spines;
	hooks of prolegs usually triordinal (fig. 27)Nymphalidæ (p. 44)
76.	Head evenly rounded; hooks biordinal (fig. 26)77
77.	A mid-dorsal spine on ninth segment of abdomen; spines of body segments
	strongly unequal, and armed with short nodules or spiracles (fig. 423).
	Citheroniidæ (p. 664)
77.	No mid-dorsal spine on ninth segment, or body spines subequal and armed
	densely with long poison-spinules (fig. 418)Saturniidæ (p. 668)
78.	Segments with six or eight annulets, prolegs normal in position.
=0	Sphingidæ (p. 42; 360)
18.	Segments with two or three obscure annulets; prolegs unusually widely
70	Segments with two or three obscure annulets; prolegs unusually widely separated
79.	Head high, triangular
80	Head not triangular
00.	An inconspictous ind-dorsal spine on innen abdominal segment.
63	Citheroniidæ (Anisota) (p. 664) No mid-dorsal spines
81.	Hooks in an ellipse at most narrowly interrunted Hesperidæ (p. 43)
81.	Hooks in one band, occasionally interrupted, rarely in two widely separated
	bands
82.	Band of hooks reduced or interrupted at middle
82.	Principal band of hooks continuous
83.	Head half diameter of body; secondary hair relatively prominent.
83.	Head rarely more than a third as wide as body; secondary hair less prom-
	inent
84.	Head rarely more than a third as wide as body; secondary hair less prom- inentLycænidæ (p. 44) A forked, eversible dorsal gland just behind head (osmeterium).
	Papinonidæ (p. 44)
84.	No osmeterium
85.	Body with branching spines, high hairy tubercles, fleshy filaments, bifur-
	cated anal plate or angulated or spined headNymphalidæ ¹⁰ (p. 44)

,

¹⁰ The western Saturniid genera, Agapema and Saturnia, will run here. They have regular, branching spines, strong prolegs, and a small, smooth head, unlike any Nymphalidæ known to me.

Pieridæ (p. 44)

Pupa¹¹

	-
1.	Mandibles large, movable, crossing in front of the face. Micropterygidæ (p. 62); Eriocraniidæ (p. 64)
1	Mandibles small fixed or obsolete
	Mandibles small, fixed, or obsolete
2.	line
3.	Maxillary palpi present, separated by a suture from maxillæ4
3.	Maxillary palpi absent or represented merely by lateral extensions of the maxillæ
4.	Dorsum of abdomen with a covering of fine spines, not arranged in rows; dorsal head-piece longer than prothorax on mid-dorsal line
	Several abdominal segments each with a row of spines near the anterior edge, sometimes with a second posterior row, but with scattered spines rudimen- tary or absent
5.	Maxillary palpi distinct, extending as a band along posterior margin of eye6
5.	Maxillary palpi minute, not extending along posterior margin of eye.
6.	A few Gracianidæ (p. 161) Abdominal segments with irregular areas of diffuse spinules only; first segment with spiracle exposed
6.	Abdominal segments each with an anterior row of spines, more prominent than the area of fine spinules; first segment with spiracles covered by wings. Incurvariidæ (p. 72)
7	Abdomen with two rows of spines on each of the middle segments
	Abdomen with the anterior rows of spines only
8.	Cremaster represented by a tuft of spines or absent, no spines on the anal rise; wings narrow and pointed; large spines present on venter of tenth abdom- inal segment
8.	Cremaster specialized, forming a definite process, or spines present on anal rise; wings broad, not pointed; no large spines on venter of tenth segment
9.	Last abdominal segment with a group of angular nodules, overshadowing the
9.	setæ. Maxillary palpi going with tongue on dehiscence. Phaloniidæ (p. 499) Last abdominal segment with prominent setæ. Maxillary palpi separating from
10.	tongue on dehiscence
10.	than a quarter as longGlyphipterygidæ (p. 350) Mesonotum not produced in a lobe; metathorax rarely less than half as long.
	Tineidæ (p. 116) Dorsal head-piece much longer than prothorax on middle line, often twice as long; dorsum of abdomen with scattered fine spines, rarely dominated by a stronger anterior row
11.	stronger anterior row
12	the other sclerites)16 Antennæ extending half the length of the wings; labrum very long and lobe- like, covering a fourth of labial palpi
	(P. Mas)

¹¹ Based on the keys in A Classification of the Lepidoptera, based on Characters of the Pupa, by Dr. Edna Mosher, in the bulletin of the Illinois State Laboratory of Natural History vol. 12, article 2.

12.	Antennæ extending at least three-fourths the length of the wings; labrum
13.	normal
13.	equal in length, and seldom extending beyond tip of wings
14.	ing beyond tip of wings
14.	mate on mid-dorsal line on middle abdominal segments Tischeriidæ (p. 145) Labial palpi invisible; last segment with two lateral spines; setæ i normal.
15.	well separated Lyonetiidæ (Bucculatriæ) (p. 148) Abdominal segments 3 to 7 each with two deep punctures at the anterior margin near the mid-dorsal line; segments 8 to 10 together shorter than segment 7, not separated by distinct incisures.
15.	Lyonetiidæ (Phyllocnistis) (p. 148) No such pits on dorsum; abdominal segments 8 to 10 relatively longer. Gracilariidæ (p. 161)
16.	Cremaster developed with a distinct stem, longer than thick. Tortricidæ (Peronea) (p. 376)
17.	Cremaster not developed, its hooks attached directly to body
17.	first abdominal spiracles invisible
18.	abdominal segment
	Megalopygidæ (p. 101) Abdominal segments without areas of fine setæ; no tubercles behind the spiracles
19.	spiracles
19.	wings
20.	Mesothorax less than twice as long as metathorax; maxillæ quadrangular. widely separated, divergent
20.	Mesothorax more than twice as long as metathorax; maxillæ longitudinal, their tips often meeting in midventral line; or all appendages obsolete
21.	Third abdominal segment movable on second; segments with an anterior row of spines and a posterior row of setæPsychidæ (p. 140)
21.	Third abdominal segment fixed; segments with anterior and posterior rows
22.	of spines
22.	Pilifers absent, labrum simple or obscurely bilobed 31
23.	suture (fig. 29)
23.	Maxillary palpi absent25
24.	Epicranial suture absent, fronto-elypeal about half visible; eighth segment of
-1.	abdomen free from the seventh in male; no deep dorsal furrow between ninth
	and tenth segments of abdomen
24.	Epicranial suture distinct at sides; sometimes running into suture between
	head and thorax at middle (except in the Phycitinæ which have a deep groove
	between ninth and tenth segments of abdomen); fronto-clypeal suture
	absent: eighth segment fixed
25.	A deep groove between ninth and tenth abdominal segments dorsally; fore
	temora exposed
25.	No deep groove between ninth and tenth segments

20	the second second solution is a second in a between energy and antenna.
26.	Antennæ not swollen, fore and middle legs extending between eyes and antennæ; fore femur normally exposed
26	fore femur normally exposedPterophoridæ (p. 639) Antennæ swollen outwardly; fore legs abutting squarely on eyes, middle legs
20.	sometimes extending between eyes and antennæ, fore femur concealed (butter-
	flies)
97	Outer angles of maxillæ in contact with eyes; tongue often projecting beyond
41.	tips of wings, pung normally in a cocoon Hesperiide (n 43)
97	tips of wings; pupa normally in a cocoon
	normally exposed
28	normally exposed
20.	rarely in a cocoon
28.	Middle legs reaching forward to eyes, and often between eyes and antennæ;
	pupa suspended by tail only, or rarely in a cocoonNymphalidæ (p. 44)
29.	Tongue falling short of tip of wings, pupa roundedLycænidæ (p. 44)
29.	Tongue almost always reaching tip of wings
30.	Anterior end of pupa with two pointsPapilionidæ (p. 44)
30.	Anterior end of pupa with a single point
31.	Fore wings extending far beyond the posterior edge of the fourth abdominal
	segment in the midventral line; fore femora and almost always labial and
	maxillary palpi exposed; fronto-clypeal suture obsolete; prothorax notice-
	ably narrower on mid-dorsal line than at sides; antennæ very rarely in con-
	tact on midventral line
31.	tact on midventral line
	segment; or fore femora and labial palpi concealed, or fronto-clypeal suture
90	distinct; prothorax usually subquadrate; maxillary palpi often covered34
32.	Pupa incapable of motion; maxillary and labial palpi concealed.
32	Lyonetiidæ (p. 148) Pupa with several free segments; labial palpi exposed
33.	Maxillary palpi present; caudal end of body without lateral projections ending
	in spines
33.	in spines
	spines
34.	Fore wings extending well beyond fourth abdominal segment, or abdomen
	depressed, with incisures much deeper dorsally and ventrally than on sides,
	and capable mainly of dorso-ventral motion; antennæ in contact on middle
	line; first four segments of abdomen usually longer than the remainder;
	epicranial suture present, maxillary palpi usually, and femora and labial
	paipi rarely, present
34.	Fore wings not extending beyond fourth segment, or maxillary palpi absent;
	abdomen not depressed and specialized for dorso-ventral motion; first four abdominal segments rarely longer than the others; epicranial suture rarely
	visible
35	Maxillary palpi absent; no movable segments; a specialized cremaster present;
	pupa suspended and girt like that of a butterfly; tongue as long as fore
	legs
35.	legsCycnodiidæ (p. 218) Maxillary palpi present, and body capable of dorso-ventral motion, except in
	a few genera which have no cremaster; tongue longer than middle legs,
	reaching antennæ, except in Ethmia
36	Antennæ four-fifths as long as fore wings, meeting only at their apex; labial
	palpi large, maxillaries minute, fronto-clypeal suture complete, strongly
	sinuate of middle; most of setæ hookedYponomeutidæ (Scythris) (p. 337)
30	Antennæ reaching almost or quite to tip of wings, coming in contact at two-
	thirds length of wings, and often diverging again at apex; labial palpi con-
37	cealed unless fore femora are exposed
37	Antennæ diverging at apex; three flexible incisures on abdomen

38.	Tongue shorter than fore legs Ecophoridæ (Ethmia) (p. 244)
38.	Tongue longer than fore legs
39.	Maxillary palpi large; labial palpi obsolete; fronto-clypeal suture complete;
90	Temora concealed
39.	Maxiliary paipi minute and ironto-civical sutine interrupted; or maxiliary
	palpi lost, or large in primitive species with fore femora exposed. Lavernidæ (p. 318)
40	Fronto-clypeal suture complete, maxillary palpi large, in contact with both
	fore and middle legs (n. 255)
40.	Fronto-clypeal suture obsolete in middle
41.	Maxillary palpi large, no hooked setæ on venter of ninth abdominal segment.
	Ecophoridæ (p. 230)
41.	Maxillary palpi minute, distant from maxillæ; with hooked setæ on venter of
	ninth abdominal segmentXylorictidæ (p. 250)
42.	Labial palpi exposed, lanceolate
42.	Labial palpi reduced to a minute area behind mouth, or lost
40.	Body with more or less dense secondary setæ (sometimes very minute) not arranged around the larval wartsLasiocampidæ (p. 679)
43	Body with primaries only, or with setæ arranged around the larval warts44
44.	Fore femora exposed
44.	Fore femora concealed
45.	Fore femora concealed
	reaching to maxillary palpus or eye; or a special ridge developed on fifth
	reaching to maxillary palpus or eye; or a special ridge developed on fifth segment of abdomenNoctuidæ (p. 42) Abdomen ending in a group of pyramidal points, the setæ obscure, middle legs
45.	Abdomen ending in a group of pyramidal points, the setæ obscure, middle legs
	touching maxillary palpi; no special ridge on fifth segment. Pyralididæ (Diatræa) (p. 590)
46	Body setæ not arranged in circles about the scars of the larval warts.
10.	Agaristidæ (p. 42); Noctuidæ (p. 42)
46.	Body setæ arranged in circles about the scars of the warts
47.	Tongue less than half as long as fore wingsLiparidæ (p. 42) Arctiidæ (Halysidota) (p. 42); Euchromiidæ (p. 43)
	Arctiidæ (Halysidota) (p. 42); Euchromiidæ (p. 43)
47.	Tongue less than half as long as fore wings Liparidæ (p. 42)
48.	Maxillary palpi preserved, close to antennæ; a dorsal crest on thorax and base
10	of abdomen; no specialized cremasterPyralididæ (Galleriinæ) (p. 532) Maxillary palpi lost; rarely with a mid-dorsal keel; usually with a cre-
40.	maxinary paper lost, rately with a ind-dorsal keer; usually with a cre- master
49.	Antennæ distinctly swollen toward tip; cremaster obsolete.
	Nymphalidæ (Œneis) (p. 44)
49.	Antennæ not swollen toward tip; often pectinate; or cremaster well devel-
	oped
50.	Setæ arranged in circles around the larval warts
50.	Setæ not arranged in circles around the larval warts
51. 51	Antennæ more than half as long as fore wings
51. 52	A long cremaster, as long as ninth and tenth segments of abdomen together,
02.	with hooked setæ, abdomen without flanged plates.
	Noctuidæ (Pantheinæ) (p. 42)
52.	Cremaster rudimentary or without hooked setæ, in the latter case with flanged
	plates on abdomenArctiidæ (p. 42)
53.	Cremaster rudimentary or without hooked setæ, in the latter case with flanged plates on abdomen
	$\mathbf{B}_{\mathbf{T}} \mathbf{V}_{\mathbf{T}} = \mathbf{B}_{\mathbf{T}} \mathbf{V}_{\mathbf{T}} \mathbf{U}_{\mathbf{T}} $
53.	Body covered with fine soft secondary setæ; cremaster strong, with recurved hooks
59	Body with set x simple obscure or mostly lost

- 56. Antennæ usually considerably broader near the base, their greatest width usually greater than that of the prothoracic legs; antennæ usually more than three-fourths the length of the wings; if not, then the epicranial suture is present, or the cremaster is wanting, or if present, bifurcate at the distal end or bearing hooked setæ; dorsum of the abdomen usually with a deep furrow between the ninth and tenth segments; scar of a caudal horn never present on the dorsum of the eighth abdominal segment; labial palpi sometimes visible as small triangular or polygonal areas caudad of the labrum..57
- 57. Maxillæ usually more than three-fifths the length of the wings; if not, then the caudal end of the body with hooked setæ, or the spiracles of the third abdominal segment concealed by the wings and those of the sixth segment farther ventrad than those of the other segments; prothoracic femora often exposed; a deep furrow usually present on the dorsum of the abdomen between the ninth and tenth segments; posterior margin of mesothorax never with a row of deep pits with smooth tubercle-like areas between.

Geometridæ (p. 41)

Saturniidæ (Hemileucinæ) (p. 669)

Order LEPIDOPTERA

Suborder JUGATÆ

Body and wings covered with minute spinules or aculeæ. Head (fig. 32), with ocelli, when developed, separated by a wide, unscaled space from the eyes; ocelli in the Hepialidæ rudimentary (Sthenopis) or absent. Antennæ without sense-cones, with aculeæ on their surface; in the North American species proportionately small and simple, but pectinate in exotic species. Clypeus separated by well-marked sutures from both front and labrum. Thorax loosely constructed, the metathorax fully half as large as the mesothorax and similar in structure; the halves of the metascutum usually meeting on the middle line. Fore wing (figs. 31, 36, 40) with subcosta more or less distinctly forked; humeral vein present; base of media preserved; jugum present. supported by the rudiments of a fourth and a fifth anal vein; with an oblique vein from **M** to 2d **A** near base, which appears like a cross vein, but whose upper half is in fact a portion of cubitus (fig. 41). Hind wing with four or five branches of radius preserved, the venational plan about as in the fore wing, but with the oblique vein in the base of the anal region obscure. Jugum usually rudimentary or absent; frenulum composed of a series of weak bristles or absent.

Caterpillar with dorsal setæ placed similarly on mesothorax and metathorax, and on abdomen (figs. 39, 42). Eyes, when six, in an irregular group, when single, on face. Pupa with four or more movable segments, with mandible set off by a suture, or articulated.

The three families seem relics of an earlier age, and are not closely related.

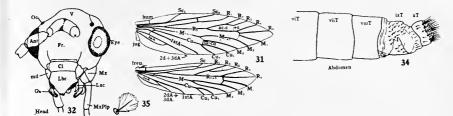
Family 1. MICROPTERYGIDÆ

(Eriocephalidæ)

Head broad (fig. 32); the ocelli widely separated by a bare space from the small eyes; all sutures preserved; loosely hairy, with a naked space above eyes. Antennæ moniliform, with whorls of stiff bristles, the scape and pedicel scaled. Mandibles large, functional; maxillæ with a large, folded, five-jointed palpus, without a tongue, but with a small, stiff lacinia which serves in handling food, as well as the twojointed galea (fig. 33); labium with a well-developed basal segment, supporting the two palpi; the mouth parts as a whole exactly as in ordinary biting insects; middle tibia with a tuft of hair at tip, but without spurs; hind tibia with four spurs. Abdomen of female ending in two retractile segments, without a trace of an ovipositor (fig. 34).

Fore wing (fig. 31) bluntly lanceolate, \mathbf{R}_{5} running to costa. Sc

forked shortly beyond middle, \mathbf{R}_1 forked or connected by a cross vein to Sc, accessory cell present. 2d A with large fork at base, con-



FIGS. 31, 32, 34, AND 35. MICROPTERYGIDÆ

31, Epimartyria, venation

32, Micropteryx, head: Ant, antenna; Cl, clypeus; fr, front; Ga, galea; Lac, lacinia; Lbr, labrum; md, mandible; Mx, maxilla; MxPlp, maxillary palpus; Oc. ocellus; ∇ , vertex

34, Micropteryx, end of abdomen; viT to xT, inclusive, terga of sixth to tenth segments of abdomen

35, One of the scales representing sets on the larva of Microptervx (after Chapman). $\times 120$

nected across 1st A to Cu by an apparent cross vein, no distinct vein below the lower fork of 2d A; jugum overlying hind wing. Hind wing with similar venation but with Sc_1 lost, anal system reduced, and no jugum.

Egg spherical, apparently of the flat type; studded with blunttipped spines, white or yellowish; the eggs laid in small clusters in, and under, wet moss. Caterpillar of a slug-caterpillar type, polygonal in cross section, exceedingly thin-skinned, and soon shriveling if allowed to dry; with two double rows of socketed scales on each side (fig. 35), irregular on prothorax and last segments; ninth abdominal segment well developed. Prolegs represented by conical processes on first eight segments of abdomen, with a large sucker on the ninth and tenth. True legs normal. Head with mouth turned forward, eye apparently single and dorsal. Antennæ longer than head, the second segment the longest. Food wet moss, the known species feeding on Hypnum and liverworts. Pupa in a dense, parchment-like cocoon, with large, crossed mandibles; not well known. Hibernation probably as larva in the cocoon.

Distribution probably world-wide; about 35 known species.

The imagoes feed freely on the pollen of various flowers, using both mandibles and maxillæ.



FIG. 33. MICROPTERYX. MAXILLA Abbreviations as in figure 32; art, articulation; st, stipes

1. MICROPTERYX Hübner

(Eriocephala Curtis)

Characters of the family. Fore wing with \mathbf{R}_i simple. Our species belongs to the subgenus Epimartyria Walsingham, with \mathbf{R}_i and R₅ stalked.

1. M. auricrinella Walsingham. Purple. Head with golden gloss; base of fore wing with golden scaling; under side and hind wing mouse gray with purple iridescence. 9 mm.

The moth may be found in partly shaded, wet places in May, and is extremely local. It is commoner northward. It is nearly invisible in flight and is most often picked up in sweeping for Diptera. Its life history is unknown. This may be a synonym of M. Inteiceps Walker, described from Nova Scotia as a slightly larger form (6 lines = 12 mm.).

Distribution general in the northeastern States and Canada. New York: Mt. Marcy (4000 feet).

Family 2 ERIOCRANIIDAE

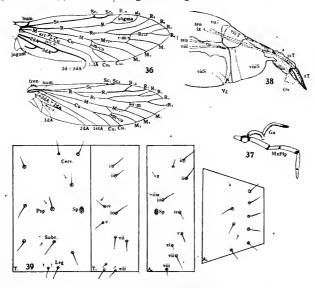
(Micropterygidæ, in part)

Head dorsally like that of the Micropterygidæ, antenna filiform, with sparse scaling on shaft; mandibles rudimentary, conical; maxillæ with long folded palpus, but with a short, spirally coiled tongue, and no lacinia (fig. 37); basal joint of labium preserved. Fore wing (fig. 36) with Sc shortly forked near tip, \mathbf{R}_1 forked, accessory cell variable, \mathbf{R}_5 running to apex, \mathbf{R}_{4} stalked, one radial sometimes absent. Anals similar to those of the Micropterygidæ, but with an additional vein below the fork of 2d A (fig. 36). Jugum as in the Micropterygidæ. Hind wing similar to fore wing; Sc_1 rudimentary or absent, anal system quite variable (the most complex arrangement known to me is shown in figure 36). Frenulum rudimentary. Hind tibia normal, hairy; middle tibia with a single spur. Abdomen of female terminating in a horny, piercing ovipositor, with powerful muscles (fig. 38).

Egg ellipsoidal, soft, laid in the tissues of the leaf. Caterpillar of leaf-miner type (fig. 39) with large head; flattened; body-setæ much reduced, but apparently with i and ii vertically placed on abdomen, and ia and ib similarly on mesothorax and metathorax. Prolegs rudimentary, apparently present on first eight segments of abdomen; true legs absent. Ninth segment well developed. Head with mouth pointing forward, with a single ocellus on dorsal surface.

The caterpillar makes a roomy blotch mine, starting from a short; linear one, in the hardly expanded young leaves of Amentiferæ, which are sometimes distorted in their further growth. It scatters its frass loosely. It feeds quickly, in the spring, and then spins a tough cocoon in the ground and summers and winters there, pupating in the spring.

Pupa with enormous mandibles, crossed in repose, which it uses to cut its way out of the cocoon and to dig up to the surface. All parts loose (pupa libera), there being some power of motion even between



FIGS. 36-39, ERIOCRANIIDÆ

36, Mnemonica, venation; fore wing: c.v., crossvein, 1st A to 2d A; hind wing: c.v., crossvein, 2d A to 3d A. 37, Eriocrania, maxilla: Ga, galea MxPlp, maxillary

palpus

38, Eriochania, end of abdomen: viiT to ixT, inclusive, seventh to ninth tergites of abdomen; xT, tenth tergite or ovipositor; viiS, viiiS, seventh and eighth sternites; ten viii, ten ix, tendons of eighth and ninth segments; Ov, oviduct; Vg, vagina

39, Mnemonica auricyanea: seta map (after Busck)

the thoracic segments; skin extremely thin, shrivelling on emergence. All sutures distinct; labrum large, free, with six pairs of setæ. First seven abdominal segments free.

The imagoes are short-lived, and usually do not feed. The family is apparently world-wide in distribution, with about 20 known species.

1. ERIOCRANIA Zeller

(*Micropteryx*, in part; *Eriocephala* Dyar, laps. cal.)

Characters of the family. Wings lanceolate. Our species belong to the subgenus Mnemonica Meyrick; with all veins present, accessory cell absent in both wings, and R of hind wing 4-branched.

3

h

22

D

1. E. auricyanea Walsingham. Head and thorax with long, gray hair, with slight iridescence. Fore wing golden with scattered, small, purple spots, each of several scales, the spots denser toward base and inner margin. Larger blotches along margin toward apex, and at anal angle, and a streak running up from inner margin near base. 12 nm.

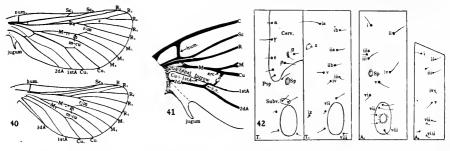
Moth in March and April. Larva in a large blotch mine on oak, chestnut, and chinquapin, in April and May; extremely local, sometimes confined to a single tree. District of Columbia. New York: Karner.
2. E. griseocapitella Walsingham. Similar to the preceding species but without

2. E. griseocapitella Walsingham. Similar to the preceding species but without any purple blotches, the spots being all small and rounded; perhaps not distinct. Described from the District of Columbia.

Family 3. HEPIALIDAE

(Swifts)

Head loosely hairy over whole upper surface, concealing the rudimentary ocelli when they are present; mouth parts rudimentary, mandibles and maxillæ recognizable under the microscope, basal joint of labium present, bearing the small, hairy palpi; antennæ, in our species, about as long as width of thorax, somewhat moniliform, pedicel large



FIGS. 40-42. HEPIALIDÆ

40, Hepialus sylvinus (Europe), venation; 41, same, detail of base of fore wing; 42, Sthenopis humuli (Europe), seta map (after Fracker)

and sealed, but flagellum scaled at base only. Legs weak, without spurs; the hind legs of many species with a large tuft of hair in the male. Fore wings (fig. 40) with humeral vein, with **Sc** more or less distinctly forked but little beyond its middle, \mathbf{R}_1 simple, \mathbf{R}_2 and \mathbf{R}_3 stalked, the apex below \mathbf{R}_3 ; base of **M** fully preserved, forming a large intercalated cell. Oblique anal eross vein present, as in the Mieropterygidæ (fig. 41), but anal system more or less reduced, often with only **2d A** fully developed. Jugum slender, with a long, free tip; underlying hind wing in flying position. Hind wing with practically identical venation, but anal region more reduced, with simple veins. Thorax very loosely organized, leaving a distinct space between the

bases of the fore and hind wings. Abdomen long and elumsy, without any modifications; in the female ending in complicated structures not yet fully understood.

Eggs of flat type, rounded, usually scattered broadcast in the neighborhood of the food. Larva (fig. 42) a borer; slender, cylindrical, with a rather long head. Ocelli six, the four anterior not in a crescent, but in an oblong group, the other two close to them. Maxillæ of a peculiar type, the palpus with three free segments. Mesothorax and metathorax with setæ ia in front of ib, as on abdomen, v higher than iii and iv. Abdomen with lateral setæ high, iv higher than spiracle, and the three arranged in an oblique row; iiia and \mathbf{x} well-developed setæ, but ix minute; ninth segment of abdomen large, with setæ normally arranged. Prolegs normal, with multiserial hooks in a complete ellipse. the upper hooks rudimentary and grading into the skin granulations. The caterpillars normally bore in roots and root-stocks, and those of Sthenopis are practically aquatic; they are very active.

Pupa slender, fitting the burrow; mandibles rudimentary, but sharply defined; all sutures of head preserved except the elypeo-labral, even the gena being distinct; antennæ short, maxillæ very short and widely divergent, quadrangular, not covering the labium, the parts not differentiated; abdomen with two series of spines on each segment, the end of the abdomen roughly spined, without a eremaster; setæ iv not as high as in the larva. Abdominal segments 2 to 7 of male and 2 to 6 of female free. Pupa leaving burrow on emergence.

The moths are mostly dusk-fliers, although H. hyperboreus flies in the daytime. The family is world-wide and largely developed in Australasia, with nearly 200 species.

Key to the genera

1. STHENOPIS Packard

(*Hepialus*, in part)

The larvæ, so far as known, bore in the roots of trees and shrubs growing partially submerged, and usually work below the water level. The moths have the habit of swarming like midges, the males having a wavy, zigzag flight, and the females apparently entering the swarm of males one by one. They are very short-lived and do not come to light or sugar. The larvæ feed at least two seasons, pupating in early summer. The pupa has a peculiar comb of short spines on the venter of the seventh segment of the abdomen, which also shows weakly in Phassus. The genus is doubtfully distinct from Phassus.

WILLIAM T. M. FORBES

Key to species

1. Pale yellow (fading to dirty	white)
1. Brown with bronzy iridescence.	4. auratus.
1. Ochre yellow, or brown and gra	
2 Hind wing salmon brighter t	than the vellowish fore wing

2. Hind wing salmon, brighter than the yellowish fore wing.

3. quadriguttatus. 2. Hind wing straw yellow, shading into ochreous toward border. 1. argenteomaculatus ♀.

- 2. Hind wing mouse gray.
 - 3. Median dark band not noticeably invading cell \mathbf{M}_s ; postmedial dark band with a clearly marked outer defining line from \mathbf{M}_1 to below \mathbf{Cu}_2 at least; median and apical dark areas not suffused with yellowbrown; hind wing normally shaded with ochreous at apex; lower silvery spot usually enlarged, triangular....l. argentcomaculatus β .

1. S. argenteomaculatus Harris. Grayish, with darker brown, pale-edged, confused bands, the two principal bands converging from base and apex of costa toward middle of inner margin, with two shorter ones between these, and a dark marginal band. Hind wing mouse gray in male, yellowish in female, 65-100mm (H 41:14).

Midsummer. Larva in rootstocks of alder.

Massachusetts to Pennsylvania and Minnesota; probably wider spread, but confused with the next two forms. New York: Catskills, Glendale, L. I.

2. S. purpurascens Packard. Similar to S. argentcomaculatus except as noted in synopsis. 75-100 mm.

Var. los Strecker has a single silver spot near base and one or two at end of cell; var. perdita Dyar is without silver (H 41:13).

This species is confused with the last in records. I have seen it from northern Ontario and Canada.

3. S. quadriguttatus Grote. Fore wing ochreous, hind wing salmon, markings exactly as in *purpurascens*, sometimes suffused. Hardly distinct from the last.

New England to northern Ontario and western New York. New York: Lancaster and Buffalo (VanDuzee), Albany.

4. S. auratus Grote. Grayish with a rosy tinge; bands yellow-brown, pale-margined, the submarginal and marginal often more or less completely fused, and the basal markings confused and largely fused. Angular, brassy-yellow spots at base, a discal spot, and a series of about three, subterminally. Hind wing mouse gray, tawny at margin. 50 mm.

July northward; end of June in North Carolina. Very tew specimens known.

Quebec; North Carolina. New York: Fentons (Lewis County), Lancaster, Ithaca, McLean, Catskills (Summit).

5. S. thule Strecker. Pale yellow with obscure markings and a brown patch from base to beyond middle of costa; a couple of silvery points.

July. Sometimes common where found at all. Larva in roots of willow. Wisconsin; Hudson Bay; Montreal; New York. New York: Waddington.

2. HEPIALUS Fabricius

Similar to Sthenopis; antenna full as long as thorax and less moniliform. The moths occasionally come to light. The larvæ bore in herbaceous plants.

Key to species

1 ellow-pro	wm	 	 	hyperboreus.
Brownish	gray	 	 	2. gracilis.

l. H. hyperboreus Möschler. Yellow-brown; male with an irregular, silver V-mark extending across the wing or nearly so, and sometimes with silvery terminal spots. Female similar to male or with an obscure blackish V only. 50 mm.

The species is hardly distinct from H. ganna of the Alpine regions of the Old World, and the forms in this country are not well understood. The form with silvery terminal spots flies by day, and is typical hyperboreus (Labrador, Alberta); the one without is macglashani Henry Edwards (H 41:15) and is a dusk-flier (Ontario, California).

Hymers, Ontario; Arctic America, and westward.

2. H. gracilis Grote. Fore wing brownish gray, mottled, the most distinct marking being an irregular band from base to middle of inner margin, and one from apex nearly to the same point, both with a gray central shade, and often with a blackish shade where they meet. \mathcal{J} over 30 mm.; \mathcal{Q} over 40 mm. The larva is likely to bore in ferns.

Maine to Massachusetts; Colorado. Var. mustelinus Packard. is a more northern form of the species dominant in Nova Scotia and Quebec, and also ranging to Colorado. It is smaller (β under 30, 9 under 40 mm.) more decidedly brown, and more contrastingly marked than the type, the female being as contrastive as the male of the typical form. Intergrades are not rare.

WILLIAM T. M. FORBES

Suborder FRENATÆ

(Heterocera, in part)

Aculea present only in a few of the most primitive forms. Ocelli, when present, close to the upper margin of the eye, placed directly behind the antenna, but often absent. Antenna with sense-cones except in a few primitive forms, the second segment, or pedicel, usually proportionately small. Vertex separated by sutures in the lower forms; in the higher ones with practically all the head-sutures obliterated. Mouth parts never of the mandibulate type, when not rudimentary with the labrum three-lobed, the central lobe filling the gap between the bases of the two maxillæ, the lateral lobes forming the pilifers. Maxillæ with basal structures reduced; the lacinia absent, and the palpi in the majority of forms reduced or absent. Labium usually rudimentary or absent, except for its three-jointed palpi. Thorax with the third segment smaller than in the Jugate, often reduced dorsally to a narrow strip, but always developed ventrally for the attachment of the legs; the two halves of the scutum usually widely separated. Abdomen in primitive forms (Adelidæ) with a horny, piercing ovipositor (fig. 50), which becomes lost in the higher types, though the two pairs of strong tendons and muscles are preserved. In many of these higher forms the ovipositor ends in two lobes.

Fore wing with humeral vein rudimentary or absent; Sc always simple; R₁ always simple, base of media and 1st A often lost, 2d A always preserved, and usually apparently forked at the base, as a result of the attachment to it of the tip of the upper fork of 3d A; lower fork of **3d A** usually lost; (sometimes the lower fork is free, or **3d A** wholly lost). Jugum never present, the inner-marginal cord always running directly into the scutellum. (In the nonaculeate forms, there is a small patch of minute spines on the inner margin near the base, which may be a persisting remnant of the acule. Hind wing with the radius much reduced, typically with only one free branch (**Rs**, usually spoken of merely as **R**); **Rs** once forked in the lower Elachistidæ, and in our genus Tinagma, in which latter genus the dorsal venation is much reduced; \mathbf{R}_1 crossing over to \mathbf{Sc} and fused with it from its junction to the apex, except in a few Gracilariidæ with very narrow wings; if Sc and Rs remain separate, R_1 looking like a crossvein, usually located near the base of the wing in those forms where **Sc** and \mathbf{R}_{s} remain independent, but often obliterated by the fusion of Sc and R at the point where R_1 should cross. Hind wing usually with three apparent anals, or two by the loss of 1st A; for the upper fork of 3d A has become almost completely fused with 2d A, in the Tortricide and broad-winged Tineids (in the broad sense), showing as a distinct basal fork of 2d A, as on the fore wing. Cross-

veins between the anals very rarely present but obvious in a few Cossidæ and Psychidæ between 1st A and 2d A of the fore wing, and visible at the base of the wing in a few low Tineids. Secondary veins rare, present on the inner margin of the fore wing of the Megalopygidæ, and near the base of the costa of the hind wing of most Lasiocampidæ (fig. 428), where they are known as humerals, one of them, perhaps, being the true humeral. Humeral vein often present; when the frenulum is present usually running across to its base. Costal edge thickened out to the point of origin of the frenulum, being a rudiment of vein **C**. Frenulum lost in several of the higher families, either in part or all of the genera; when present, it is practically always single, in the male, running through a membranous retinaculum attached just below the costal edge of the fore wing; in the female, however, usually multiple, formed of two to many bristles, and held in place by stiff hair-scales projecting down from Sc and R, and up from below **Cu** of the fore wing. Frenulum occasionally single in the female also.

Caterpillar with the setæ ia and ib of the thorax vertically placed, on the abdomen with i anterior to ii; thorax normally with \mathbf{v} lower than iii and iv; abdomen with iv level with the spiracle or lower, and iv, v, and vi not lying in an oblique line; iiia and x minute; prolegs rarely either modified or absent; body often with tufted or secondary setæ. Maxillary palpus with only two free segments; four anterior ocelli, when present, arranged in a semicircle. When only a single ocellus occurs, it is on the side of the face.

In boring larvæ, the structures are usually normal; but leaf-miners may be exceptions to most of the characters given in this definition.

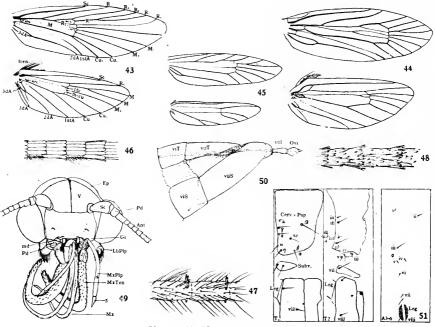
Pupa with head-sclerites more or less completely fused; metathorax smaller than in Jugatæ and thoracic segments never free, but in other particulars varying from forms with almost the structure of the Jugatæ, to forms with all the parts soldered together.

The Frenate have been derived from a Jugate type intermediate between the three surviving families; the Rhopalocera are derived from primitive Frenate in the neighborhood of the Cosside, as is indicated by a series of intermediate exotic forms.

WILLIAM T. M. FORBES

SUPERFAMILY INCURVARIOIDEA Family 4. INCURVARIIDÆ (Adelidæ)

Vertex very rough-haired, front either rough or scaled; with a distinct naked area above eye behind antennæ, except when the eyes are very large. Vestiture lightly attached. Eyes extremely variable



FIGS. 43-51. INCURVARIDÆ

- 43, Adela ridingsella, venation
- 44, Prodoxus decipiens, venation
- 45, Eudarcia simulatricclla, venation
- 46, Adela ridingsella, 9; section of antenna, ventrolateral view
- 47, Incurvaria, antenna
- 48, Paraclemensia, antenna

49, Tegeticula guccasella, Q, head: Ant, antenna; Ep, epieranium; Ge, gena; LbPlp, labial palpus; Mx, maxilla (tongue); MxPlp, maxillary palpus; MxTen, maxillary tentaele; Pd, pedicel of antenna; Pil, pilifer; Sc, scape of antenna; V, vertex; (the segments of the maxillary palpus are numbered 1 to 5)

50, Tegeticula guccasella, Q, end of abdomen: abbreviations as in figure 38 **51**, Adela cuprella (Europe), seta map

in size; in some western and exotic species nearly meeting on vertex, with two sizes of facets in male. Antennæ in some genera short, in Adela the longest known in the Lepidoptera, several times as long as fore wings; often shorter in female, and more densely scaled; commonly with two equal whorls to a segment, or irregularly scaled and bristled (figs. 46-48); maxillary palpi long and folded, short and porrect, or rudimentary,- in Prodoxinæ the longest known in the Lepidoptera; tongue scaled at base. Palpi and hind tibiæ often densely hairy, especially in male. Fore wing with complete venation or a single vein lost, cell large, accessory cell and base of M usually distinct, 1st A distinct at margin, 2d A forked at base. Membrane aculeate over the whole surface of the wing. Hind wing with Sc swollen at base, \mathbf{R}_1 normally forming a heavy basal fork of it, though completely fused in Adela; base of **Rs** obsolete, **R** and \mathbf{M}_1 stalked only in Nemotois, 3d A_1 occasionally free at tip (fig. 43); scaling often hair-like, and iridescent or metallic in many species. Female, so far as known, with last segments heavily chitinized, with strong, piercing ovipositor; laying its eggs in the tissues of the food-plant.

Caterpillars, so far as known in the Adelinæ, case-bearers; the case typically lenticular and formed of a piece of leaf; borers in seeds and stems of Yucca and related Liliaceæ, in the Prodoxinæ. Head normal in form, the adfrontals reaching the vertex, the front about two-thirds way, setæ as a rule obscure; iv and v rather close together, and iv sometimes migrating far up behind spiracle, as in the Hepialidæ. Prolegs normally with two areas of granulations bearing rudimentary hooks, with a single line on sixth segment of abdomen, and no anal prolegs; prolegs reduced to one or two rows of minute hooks in Incurvaria, and all lost in the Prodoxinæ; true legs also lost in Prodoxus.

Pupa incomplete, chitinized, with a frontal beak; thorax, appendages and terminal segments only lightly soldered; third segment of abdomen free; dorsal spines in a patch or pair of patches on each segment, sometimes supplemented by a row of spines; maxillary palpi distinct, extending from antennæ to base of maxillæ; prothorax very narrow and more or less depressed; dorsal head piece conspicuously widened (the "Adelid prothorax"). A few Incurvarias (of group Lampronia) have only the series of spines on abdomen, and many genera are still unknown in the early stages.

This is a small and ancient world-wide family containing two or three hundred known species. The genera Triptodema, Mea, and Pelates have not been examined for aculeæ or ovipositor, and so are tentatively allowed to remain in the Tineidæ; in several other genera

the ovipositor has not been examined. The genera of this family are repeated in the key to the Tineidæ for convenience in identification.

Key to the genera

1. Antennæ at least a fourth longer than fore wings.
2. R and \mathbf{M}_1 of hind wing stalked
1. Antennæ hardly if at all longer than fore wings.
2. Maxillary palpi very short and porrect, or absent.
3. Tongue strong, labial palpi moderate
3. Tongue obsolete, labial palpi minute
2. Maxillary palpi well developed, folded, usually covering base of tongue.
3. Folded part of maxillary palpus two-thirds as long as width of head.
4. Females (with exserted ovipositor).
5. Maxilla with a long, thick, naked, coiled tentaole besides palpus and
tongue2. Tegeticula.
5. Maxilla with a sharp angle on second joint of maxillary palpus only.
1. Prodoxus.
4. Males, (with large claspers).
5. Valve one-fourth as long as fore wingl. Prodoxus.
5. Valve less than one-sixth as long as fore wing2. Tegeticula.
3. Folded part of maxillary palpus half as long as width of head.
4. Venation complete
4. Fore wing with a vein lost.
5 Hind wing with 7 yours lanceolate with open cell 6 Fudarcia

5. Hind wing with 7 veins. lanceolate, with open cell......6. Eudarcia. 5. Hind wing broader, with complete, normal venation...5. Paraclemensia.

1. PRODOXUS Riley

Similar to Tegetienla, male with larger valves; female with only a rudiment of the maxillary tentacle. Caterpillars boring in Yucca, our species in Y. filamentosa; much like Promba but without thoracic legs or projections representing the abdominal legs. Pupa fairly smooth, with spine-patches, and an anterior, toothed ridge on each segment. Pupa transforming in a silk cocoon at the mouth of its burrow (fig. 44).

1. P. quinquepunctella Chambers. Cream white, typically with about five black points, but in var. decipiens Riley immaculate. 15-20 mm. (H. p. 438 f. 255-259.) Common. The caterpillar bores in the flesh of the fruit and fruit-stalk

of Y. filamentosa, emerging a little earlier than Tegeticula yuccascila.

Central New Jersey to Missouri and south.

2. TEGETICULA Zeller

(Pronuba Riley 1872, not Thomson 1860; Valentinia Coolidge 1909, not Walsingham 1909)

Antennæ smoothly scaled, the scaling not regularly formed of two whorls to a segment; scales easily lost, uncovering the fine, chitinous spinules; palpi rather small and slender, upturned, with scales and bristles, the basal segment decidedly the longest; maxillary palpi folded, five-jointed, the first joint very long; vertex roughly hairy. The female with a long, roughly spinulose, coiled tentacle growing out from the junction of the first and second joints of the maxillary palpus (fig. 49). Fore wing with normal primitive venation, with base of **M** and \mathbf{R}_{4+5} preserved, veins all arising separately from the cell, or one or two shortly stalked.

Egg laid in the tissues of the ovary of a Yucca flower, larva feeding on the developing seeds, but only damaging part of those in the capsule. Head not depressed, small; front triangular, separated from the vertex by the adfrontals; body stout, somewhat fusiform, strongly tapering behind, not flattened; with small thoracic legs, the prolegs on the third to sixth abdominal segments represented by fleshy humps, but without crochets. Pupa with a strong frontal spine, with a heavy, serrated ridge on each segment of abdomen, with very stout, subdorsal hooks on eighth segment of abdomen pointing forward, but no cremaster.

The moth uses its tentacles to gather and carry a ball of pollen. After laying each egg the moth inserts one of the tentacles into the ovary of the flower through the stigma, and fertilizes the ovules with it.

1. T. yuccasella Riley, (Yucca borer). White, immaculate, usually with gray hind wings. 20-25 mm. (H. p. 442, f. 262-263.)

The moth is common where Yucca is found, emerging during the flowering period in May and June. Larva in Y. filamentosa.

New York to Ohio, Missouri, and south. New York: Long Island.

3. INCURVARIA Haworth

(Lampronia Stephens; Tinea, in part)

Similar to Prodoxus, but with smaller maxillary palpi, and, in our species, black, or dark metallic blue in color. Typically with whole head rough-hairy, but in group Lampronia with the face smooth and head sometimes not very rough. Male antennæ pectinate in some exotic species. The characters are superficially much like Tinea, but the scaling of the antennæ is smooth, and less regularly arranged in two rows to a segment (fig. 47); if the scales are rubbed off, the underlying spinules show, the antenna in either case appearing smooth and velvety to a low-power lens, and commonly bristly under higher power.

Larvæ sometimes borers in stens, like Prodoxus, sometimes leaf-miners when young, and later cutting out a flat, lenticular case. Head as in Tegeticula, body smoothe", prolegs with one or two transverse rows of minute hooks, setæ iv and v rather close together, below and behind spiracle. Pupa apparently like Prodoxus, with dorso-anal spines.

Key to the species

1. Straight, antemedial fascia, no costo-apical spot.....l. russatella.

- 1. An irregular, pale spot on costa before apex, besides the one three-fourths way out.

 - 2. Head pale straw yellow, antemedial fascia angulate, or not reaching costa. 2. taylorella.

1. I. russatella Clemens. Deep brown; head ochreous, with rough face; antennæ annulate, purple and gold; a pale golden fascia a fourth way out on fore wing, slightly wider at inner margin, not always reaching costa; costal and dorsal spots at middle and traces of a pale costal subterminal spot. Apical fringe white-tipped. 13 mm.

Apparently not common. May; July.

Both wings are normally scaled.

Canada to Pennsylvania and District of Columbia. New York: Ithaca.

2. I. taylorella Kearfott. Head pale straw yellow, darker above; antennæ annulate, pale straw and brown; fore wing with fascia extending up from inner

margin, typically not reaching costa, but if so, narrow and angulate; spot beyond middle of costa very large, and rather beyond the corresponding dorsal spot; subterminal spot sending a spur down in dorsal fringe almost to anal angle. 18 mm.

May; July. The structure is as in russatella, and the species is near capitella of Europe, which bores in the stem of Ribes. It is doubtfully distinct from I. russatella and occurs with it at New Brighton, Pennsylvania.

Mt. Washington, New Hampshire; Ottawa, Ontario; British Columbia. New York: Newcomb. 3. I. aureovirens Dietz. Hind wing narrower than fore wing, ovate-lanceolate,

with hair-scaling. Markings about like I. taylorella. 6. mm. Pennsylvania. This species is unknown to me.

1. labradorella Clemens, with fuscous head and markings much like russatella, belongs here doubtfully; the type only is known, from Labrador.

4. CHALCEOPLA Braun

(Cyanauges Braun, not Gorham; Incurvaria; Tineola, in part)

Head very hairy, labial palpi minute, drooping, not extending beyond the front; tongue and maxillary palpi obsolete. Venation as in Incurvaria; fore wing (in the eastern species) overlaid with golden hair-scales.

Key to the species

Ground purplel. dietziella.

1. C. dietziella Kearfott. Face very rough; M_1 and M_2 stalked in hind wing. Fore wing when fresh with golden hair-scales mixed with the normal scales. Head old gold; body and antennæ fuscous; fore wing with the normal scales purple, with golden bases, which are exposed in rubbed specimens. Hind wing slightly brower. 9 mm. (Incurvaria, Tineola.)

June. The larva possibly on Cornus.

Essex Co., New Jersey.

2. C. cyanella Busck. M_1 and M_2 free. Head brilliant orange-ochre, the antennæ contrasting, blackish; body and fore wings deep metallic green, the hair-scales golden; hind wings purple. (Incurvaria Busck.)

Alleghany Co., Pennsylvania; Cincinnati, Ohio; doubtfully from Maryland.

5. PARACLEMENSIA Dyar

(Brackenridgia Busek 1903, not Aldrich 1902; Incurvaria, in part)

Similar to Incurvaria, with narrow sparse scaling on the lanceolate hind wing; \mathbf{M}_2 of fore wing lost, \mathbf{M}_1 and \mathbf{M}_2 of hind wing short-stalked. Antennæ irregularly scaled (fig. 48). Larva at first a leaf-miner, later in a lenticular case, cating crescentic paths in the tissue of the leaves. (Fig. 48.)

1. P. acerifoliella Fitch. Under-scaling dcep purple-blue, densely overlaid with peacock green; hind wing pale, translucent. Head orange, antennæ black. 9 mm. (iridella Chambers; Ornix Fitch).

Larva (Maple case-bearer) on maple (normally only on rock maple); and more rarely birch, oak, beech, and huckleberry (Braun); occasionally in injurious numbers.

Massachusetts to British Columbia. New York: St. Lawrence County, generally; Black Brook (Clinton Co.), Ithaca, McLean, Albany, Bolton (Felt), Deposit.

6. EUDARCIA Clemens

(Meessia Spuler)

Face exceptionally rough; eyes extremely small, but prominent and visible from above, behind the antennæ, which are longer than fore wing. Fore wing (fig. 45) with one medial lost; closing vein of accessory cell weak and arising before end of discal cell, making \mathbf{R}_2 and \mathbf{R}_3 appear stalked together; hind wing two-thirds as wide as fore wing, with broad fringe; one medial lost, cell open above \mathbf{Cu} and base of media preserved.

The larva lives in a flat, ovoid, rough case, on lichens. E. simulatricella is closely related to E. vinculella of Europe.

1. E. simulatricella Clemens. Blackish; head ochreous. Fore wing with a fascia a third way out, costal and dorsal spots opposite each other at middle, an apical spot, not reaching either margin, and extreme tip of fringe white. 9 mm. (*bipunc-tella* Walsingham).

Pennsylvania.

2. E. cæmitariella Chambers. Similar to E. simulatricella, no white in fringe, markings generally much narrower and more oblique. 9 mm.

In the type specimen the fascia is broken; there are three median spots, the middle one farther out and obscure, and two white spots beyond, on costa.

This species has been confused with the similarly marked Ornix guttea, but the type is undoubtedly Adelid. I have seen no other specimens.

Kentucky; Gulf States.

7. ADELA Latreille

Antennæ (fig. 46) at least as long as fore wings, almost always several times as long in male, but relatively short and roughly scaled toward base in most females; with large basal joint. Eyes varying from very small to very large, larger in male; tongue strong, scaled at base; maxillary palpi minute, porrect; labial palpi bristly; vertex with long, bristly hair, front variable. Hind tibiæ in several species heavily bristled. Ovipositor of female strong, simpler than in Tegeticula. Fore wings (fig. 43) with all ten veins arising from the cell, prcserved, \mathbf{R}_s running to costa; all veins free, or \mathbf{R}_s and \mathbf{R}_4 shortly stalked; hind wings with all veins free, or \mathbf{M}_1 and \mathbf{M}_2 connate or stalked; Sc much swollen at base, \mathbf{R}_1 not recognizable.

The larvæ (fig. 51), when young, live, as a rule, in flowers or seeds; when older, they form a lenticular case of two, flat, oval pieces of leaf, in which they pupate. Their prolegs are extremely primitive, each consisting of two areas of hooks, grading off into the normal skin-granulations, and separated by a narrow fold of membrane. The leg on the sixth segment of the abdomen is reduced, and there is none on the anal segment. The pupa is normal.

Key to the species

A patch of black spots near anal angle (speculum).....l. ridingsella. No speculum.

1. A. ridingsella Clemens. Antennæ relatively short (in male twice as long as fore wing, and one and one-fourth times in female); face hairy, palpi moderate, eyes small; fore wing with \mathbf{R}_{a} and \mathbf{R}_{4} stalked, hind wing with \mathbf{M}_{1} and \mathbf{M}_{2} stalked. Head dark, antennæ annulated with white, fore wing yellow-brown, with a pale grayish patch beyond middle, covered with rows of black dots; with a silvery

fascia before its middle, and shorter ones beyond it near margins; speeulum of about 6 black patches with silver scales between them. 13 mm. (schlægeri Zeller; Dicte coruseofasciella Chambers).

June. July.

Maine to mountains of North Carolina; northern Pennsylvania. New York: Rock City (Cattarangus Co.), Ithaca.

2. A. purpura Walker. Antennie in male three times as long as fore wing; face hairy, palpi very bushy, eyes of moderate size; all veins widely separate in fore wing, $\dot{\mathbf{M}}_1$ and \mathbf{M}_2 stalked in hind wing. Greenish to purplish bronzy with a white medial fascia, and a costal bar beyond the fascia, both edged with black. 14 mm. (biriella Zeller).

The moth is found on willow bloom in April.

Nova Scotia to northern New Jersey and Manitoba. New York: Cranberry Creek.

3. A. bella Chambers. Antennæ four times as long as fore wing in male; one and one-half times as long and thickened with scales toward base in female; face smoothly scaled, strongly oblique inward to mouth; eyes small, palpi small and sakly bristled; hind tibia smooth. Venation as in A. purpura. Purple-green with lines of brighter, blue-purple iridescence toward apex. Antennæ black, with outer half white. 14 mm. (chalybeis Zeller; oichroa Zeller).

This species may be the same as A. earuleella Walker.

Southern New Jersey, and southern Ohio to Texas. New York: Crugers.

8. NEMOTOIS Hübner

(Adela, in part)

Similar to Adela in all stages; eyes typically very large in male, though small

in our species; **R** and \mathbf{M}_1 of hind wing stalked. **1. N. bellella** Walker. Male antennæ over twice as long as fore wings; female short and thickened with black scales on the basal half. Fore wing dull, old gold, striate with purple on vens, the inner margin and apex solidly purple; a broad. golden fascia two-thirds way out, and a streak in base of fold. 16 mm.

Canada; Colorado.

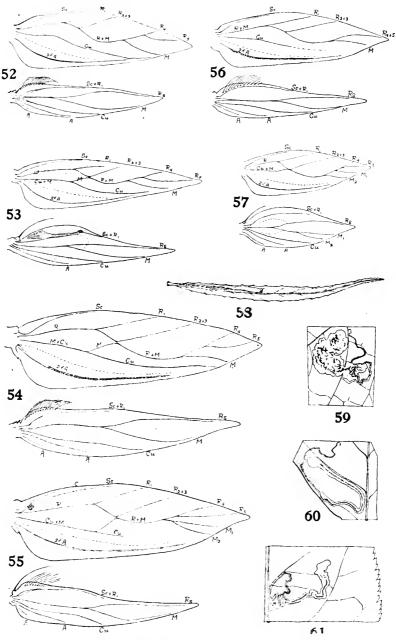
SUPERFAMILY **NEPTICULOIDEA** Family 5. **NEPTICULIDÆ**

Annette F. Braun

Head and face tufted. Antennæ not exceeding three-fourths of wing length, rather thick, with basal segment enlarged and concave beneath to form an eye-cap. Labial palpi short, porrect or drooping. Maxillary palpi long, filiform, folded. Tongue rudimentary. Posterior tibiae with bristles above. Wing membrane aculeate. Fore wings (fig. 52) with media coalescing with radius from base to beyond middle of wing, so that all the branches of radius and media appear to arise from one stem; or coalescing with cubitus for a short distance from base, then, either passing obliquely outward to radius just beyond \mathbf{R}_{2+3} , and anastomosing with radius to beyond middle of wing, as before, or remaining separate from radius, in which case (Trifurcula) \mathbf{R}_{4+5} is absent. \mathbf{R}_{2+3} coincident. \mathbf{R}_{4+5} separating beyond \mathbf{M} , or coalescing to apex. Cubitus unbranched, sometimes coincident with M or becoming obsolete beyond its point of separation from M. Second anal vein very prominent. Crossveins absent. A fibula (jugum) present in females of the more primitive genera. Hind wings (fig. 52) with subcosta and R, coincident; Rs and M coalescing to about the middle of the wing. Media one- or two-branched. Cubitus unbranched. No crossveins. Frenulum of male consisting of a single strong spine; of female, rudimentary, of several minute spines. The function of the frenulum is performed, in the female, by a series of curved spines along base of costa. Hind wing one-half to almost as broad as the fore wing.

The moths, because of their minute size and retired habits, and very rapid and irregular flight, are not frequently seen. Early in the spring, some species may be collected resting in the crevices of bark. Later, moths may sometimes be found on leaves, usually those of their food plants. Occasionally, because of the peculiarity of all the individuals of a single generation maturing and emerging at the same time, great numbers of moths may be seen on leaves of the food plant and neighboring plants. To secure an adequate representation of the group, however, rearing of the moths from larvæ is necessary.

With the exception of several gall-making species of Ectedemia, the larvæ of all species of which the life history is known, are miners within the tissues of leaves or in bark (rarely in fruits). They show a preference for trees and shrubs, but not a few mine leaves of herbaceous plants. When full grown, the larva, with few exceptions, leaves the mine, and, dropping to the ground, spins a dense, flattened cocoon amongst the rubbish or in the loose surface soil.



FIGS. 52-61. NEPTICULIDÆ (Annette F. Braun)

52, Wings of Nepticula nyssacfoliella, female; 53, wings of Nepticula nyssacfoliella, male; 54, wings of Ectoedemia heinrichi, female; 55, wings of Obrussa ochrefasciella, male; 56, wings of Nepticula terminella, female; 57, wings of Glaucolepis saccharella, male; 58, mine of Nepticula pallida; 59, mine of Nepticula nyssacfoliella; 60, mine of Nepticula saginella; 61, mine of Nepticula pomivorella

The egg is a minute oval body attached to the surface of the leaf or bark by a minute, glistening speck of cement, which renders its location visible even to the naked eve.

The larva of Nepticula upon hatching eats directly into the leaf, and makes a very narrow, linear mine (figs. 58 to 61), which is at first often difficult to discern, since in its early stages, the larva consumes but a small part of the leaf tissue. This mine may continue as a linear mine, gradually broadening throughout its course, or it may at some period abruptly enlarge into a blotch. In the latter parts the mine is semitransparent and easily visible. The mine of any one species is very constant and characteristic in appearance, and, in most instances, serves for immediate identification of the species. The species of Ectædemia are gall-producers or bark-miners in forest trees.

The larva is slightly flattened, with the head deeply retracted into the prothorax, due to the lengthening of the dorsal side of the head. Locomotor organs are represented by mere roughened protuberances; such rudimentary feet are present on segments three and four, on segments six to eleven, inclusive, and sometimes on the last segment in Nepticula; in Ectodemia there are sometimes one or two additional pairs of rudimentary processes.

The cocoon is spun of dense brown or yellowish silk, flattened oval in general outline, but usually broader at its anterior end, around which a fissure extends, guarded by the smooth projecting edges of the two halves of the cocoon. Through this fissure the pupa is thrust at emergence. In some species, the flat, projecting edges form a rim extending entirely around the cocoon.

The pupa is flattened ovate; all the appendages are free and segmented; and segments one to seven inclusive of the abdomen are free. The pupa shows in some respects a resemblance to that of the primitive Eriocraniidae.

The moth is active almost immediately after emergence, running rapidly up and down the sides of the breeding jar, and in an incredibly short time has the full use of its wings. When at rest, the wings lie almost horizontal, meeting in a line down the middle of the back.

Key to the genera

(European genera in brackets)

A. \mathbf{R}_{s} of fore wing present. B. Media of fore wing with three branches (Scoliaula).

BB. Media of fore wing with one or two branches.

CC. Media of fore wing single-branched.

D. Middle spurs of posterior tibiæ in or above the middle....4. Nepticula. DD. Middle spurs of posterior tibiæ below the middle.....3. Ectædemia.

WILLIAM T. M. FORBES

1. GLAUCOLEPIS Braun

Eye-cap large. Middle spurs of posterior tibiæ in the middle. Fore wings elongate ovate; hind wings nearly equaling the fore wings in breadth, in the male; three-fourths of the width of the fore wing in the female. Fore wings (fig. 57); cubitus coincident with media, which anastomoses with radius from \mathbf{R}_{2*3} to beyond middle of wing; \mathbf{M}_3 absent; \mathbf{M}_2 arising before separation of \mathbf{M} and \mathbf{R} . Hind wings: media two-branched.

1. G. saccharella Braun. Tuft brownish ocherous, eye-caps bluish white. Thorax and basal fourth of fore wing blue or purple metallic; remainder of wing black with a broad, bluish silvery fascia just beyond middle; eilia pale bluish. Hind wings of male with oval, yellow patch of androconia. 4 mm.

Very long serpentine mines in leaves of maples.

2. OBRUSSA Braun

Eye-cap large. Labial palpi well developed. Middle spurs of the posterior tibiæ above the middle. Fore wings elongate ovate, with fibula in the female; hind wings a little over one-half the breath of the fore wings. Fore wings (fig. 55); media coalescing with cubitus at base, then passing obliquely to radius beyond \mathbf{R}_{2-37} and anastomosing with radius to beyond middle of wing. \mathbf{R}_4 separate. \mathbf{M}_1 and \mathbf{M}_2 coalescing for a short distance beyond separation of \mathbf{M} and \mathbf{R} . \mathbf{M}_3 absent. Cubitus becoming obsolete beyond its separation from \mathbf{M} . Hind wings: media single-branched.

Represented by a single species whose early stages are entirely unknown.

1. 0. ochrefasciella Chambers. Tuft ochraceous; eye-caps buff. Fore wing blackish brown with a pale ocherous fascia at basal third; scattered ocherous scales at two-thirds, forming indistinct transverse line in female. Last row of scales at apex and cilia pale ocherous. Underside of wing of male with androconia. 6.5-8 mm.

3. ECTEDEMIA Busek

Basal segment of antennae enlarged and concave beneath to form an eye-cap. Labial palpi somewhat longer than in Nepticula. Middle spurs of posterior tibiae below the middle. Fore wings elongate ovate, pointed; fibula present in the female; hind wings two-thirds to three-fourths as wide as fore wings; nearly as long as the fore wings. Fore wings (fig. 54); media coalescing with cubitus at base, then passing obliquely to radius beyond \mathbf{R}_{243} , and anastomosing with radius to beyond middle of wing. \mathbf{R}_4 separate. Media single-branched. Cubitus reaching margin. Hind wings: media single-branched.

The position of the middle spurs on the posterior tibiae, the relatively smaller eye-caps, and the broader wings will distinguish this genus from those species of Nepticula which have identical venation.

The larvæ of the species whose life history is known form galls on twigs or petioles, or are miners in the bark of twigs. The egg is somewhat more circular in outline than that of Nepticula. There is but a single generation of the moths a year, as would be expected from the peculiarities of the life history.

In all but the unicolorous *E. populella*, the fore wings are mottled with fuscous scales, or with dark-tipped scales. Ill-defined markings are formed by the grouping of these dark scales in patches. The markings differ from those species of Nepticula which resemble Ectordemia most in structural characters.

Key to species

a. Fore wings unicolorousl.	populeUa.
aa. Fore wings mottled.	
b. Dark-tipped scales evenly distributed2.	castaneæ.
bh. Dark-tipped scales more or less collected into patches.	
c. Base of fore wing with scattered dusting, except near costa5.	obrutella.
cc. Dusting dense near base of wing.	
d. A poorly defined, pale fascia at basal third4.	heinrichi.

l. E. populella Busck. Tuft reddish ocherous, eye-caps pale yellowish. Fore wings shining coppery brown, with green and violet iridescence. 7-8.5 mm.

The larvæ form almost globular galls, of the size of a pea, on the petioles of leaves of poplar. The larva is full-grown in October. The moth appears in May. 2. E. castaneæ Busck. Tuft black above; eye-caps creamy-white. Fore wing clothed with bluish white scales, which are mostly deeply tipped with blackish brown, so that the wing is almost uniformily densely dusted. 7.5–8 mm.

The larvae form cylindrical galls encircling young twigs of chestnut. 3. E. phleophaga Busck. Tuft ocherous. Thorax and basal half of fore wing dark bluish fuscous, outer half paler, bluish with dark-tipped scales; an ill-defined, ocherous costal and an opposite dorsal patch at apical third. 9-10 mm. Serpentine mines in bark of chestnut; larva full grown in April and May;

imago in September.

4. E. henricht Busck. Tuft black, eye-caps creamy-white. Fore wing pale ocher-ous, densely dusted with blackish fuscous scales, which tend to form patches. The dark dusting is usually absent or scattered at the extreme base of wing Axcept along costa and on two poorly defined, transverse fasciae, one at basal third, the other at apical third; the second fascia sometimes almost obliterated by dusting. 9-10 mm.

The larva forms a characteristic, flattened-oval, spiral mine in the bark of young branches of pin oak (*Quercus palustris*). The larvæ are full-grown in October and early November, producing moths in May and June of the following year.

5. E. obrutella Zeller. Differs from the two preceding species chiefly by the scattered dusting of the basal half of wing. Food plant unknown, thus far recorded only from Texas.

4. NEPTICULA von Heyden

Basal segment of antennæ dilated and concave beneath to form a large eye-cap. Middle spurs of posterior tibiæ in or above the middle. Fore wings elongate ovate, pointed; hind wings one-half to two-thirds as wide as fore wings. Fore wings (figs. 52, 53, 56); media coalescing with radius from base to beyond middle of wing, or coalescing with cubitus at base and passing obliquely to radius beyond \mathbf{R}_{2+3} and anastomosing with radius to beyond middle of wing as before. \mathbf{R}_{i} sometimes coincident with \mathbf{R}_{i} . Media single-branched. Cubitus usually reach-

ing nearly to margin. Hind wings: media single-branched. As far as is known, the larvæ of all of the North American species are miners within the tissues of leaves. The egg is placed on either the upper or under surface of the leaf, often along the side of a vein, and the larva passes directly into the interior of the leaf. The larva usually mines just beneath the upper epidermis, consuming the palisade layer of cells, and in later stages, some of the spongy parenchyma cells. In thin leaves, the mine seems more transparent, because of the originally smaller number of these cells and the looseness of their arrangement. Where the upper or lower surface is mined indiscriminately, as is done by N. populetorum in the leaves of poplar, the cross section of the leaf shows palisade

cells on either side. Some species mine different sides of the leaf at different periods of larval life. The mine may be a linear tract, gradually increasing in breadth to its end, or it may at some point suddenly enlarge into a blotch. A change in the character of the mine usually indicates the beginning of a new instar. There are four larval instars. The mine formed during the first instar is very short, rarely exceeding a few millimeters in length. The large, conspicuous part is made during the last larval instar, in the few days preceding the escape of the larva from the mine. The larva leaves the mine by a semi-circular slit in the upper or the lower epidermis and spins the characteristic cocoon, usually brownish, but occasionally yellowish or whitish, in the surface soil or amongst rubbish, often near the base of the tree; cecasionally it spins on the twigs or branches. Pupation does not take place immediately; in the summer generations it cecurs a few days before emergence, in the overwintering generation it may be delayed until spring.

A few species have but one generation a year; most species have two or three; a few of the oak-feeding species may have as many as four generations. The length of the life cycle is approximately six weeks, except in the single-generation species, where several months may elapse between oviposition and the attainment of full growth by the larva. The moths from the over-wintering generation of harva emerge in May and June, a few species, however, appearing in April.

Key to the species

a. Fore wings with pale spots or fasciæ.
b. A silvery or pale golden metallic spot at, or very near, the base of the
wing.
c. A median fascia.
d. A costal and a dorsal spot at three-fourths
dd. No such spots at three-fourths.
e. Thorax and extreme base of wing purplel. argentifaseiella.
ce. Thorax and extreme base of wing golden
cc. No median fascia
bh. Without such a spot.
c. Basal third of wing buff9. eerea.
cc. Basal third of wing not buff.
d. A pale costal spot at one-third5. trinotata.
dd. Without a pale costal spot at one-third.
e. Two pale fasciæ.
f. Thorax creamy buff
ff. Thorax dark.
g. Head black
gg. Head ochraceous
ce. One pale fascia.
f. Fascia more or less interrupted.
g. Male with a chitinous plate from base to near middle of costa
of hind wing.
h. Fascia silvery
hh. Fascia not silvery; male with long hair-pencil from base
of costa of hind wing
gg. No such chitinous plate in male
ff. Fascia complete.
g. Fascia before the middle; wing lusterless.
h. Whitish costal and dorsal spots at three-fourths.
35. thoracealbella.
hh. No such spots at three-fourths

gg. Fascia in or beyond middle of wing. hh. Apex not white (sometimes with white apical cilia). i. Fore wing almost lusterless; fascia not more shining than remainder of wing. ii. Fore wing almost lustreless; fascia shining white, silvery, or golden. jj. Collar not conspicuously paler than the head. k. Apical cilia white; marginal line defined. kk. Apical cilia not white; marginal line not defined. iii. Fore wing with a metallic luster; fascia silvery or golden. j. Fascia preceded by a purple or deep golden brown band. 15. purpuratella. jj. No such band. k. Fascia the only pale marking. 1. Basal halt of wing metallic golden or bronzy; fascia ill-defined internally. m. Entire apical area deep purple....14. unifasciella. mm. Costal half of apical area purple. 13. resplendensella. ll. Fascia well-defined internally, contrasting with ground color. m. Wing purple before the fascia, brown beyond. 19. altella mm. Wing not as above. n. Fascia noticeably beyond the middle. o. Fascia indistinct.....16. obscurella. oo. Fascia distinct. p. Entire tuft ochraceous....17. ostryæfoliella. pp. Tuft ochraceous behind only.. 18. paludicola. nn. Fascia at or near the middle. oo. Tuft reddish or ccherous. p. Ground color bronzy. q. Collar pale yellowish (usually). 21. opulifoliella. pp. Ground color purplish black. 23. juglandifoliella. kk. With additional silvery or golden markings. l. A semi-elliptical, golden, metallic spot on dorsum proximal to fascia.....10. rhoifoliella. 11. Metallic markings along termen or at apex. m. Termen margined with silvery scales from dorsum11. terminella. to apex mm. Apex golden metallic, concolorous with fascia. 12. villosella.

aa. Fore wings without pale spots or fascile.

b. Ground color pale ocherous or yellowish.

e. A purplish fuscous band across apex of fore wing.

1. Nepticula argentifasciella Braun. Tuft black behind, ochraceous in front; eye-caps silvery white. Thorax and base of fore wings dark purple. Fore wings dark brown, with metallic reflections. At the basal fifth of the wing is a brilliant silvery fascia, sometimes broadening so considerably on dorsum as almost to reach the base of the wing. At the middle of the wing a second fascia; at extreme apex a silvery patch of scales of variable extent. 4 to 4.5 mm.

Larva in leaves of basswood (*Tilia americana*). Mine narrow serpentine, expanding into a blotch. Cocoon reddish.

There are two or three generations a year. The larvæ become full-grown toward the end of June, in August, and in the latter part of September.

2. Nepticula scintillans Braun. Tuft and collar black; eye-caps silvery white. Thorax and base of fore wing golden. Fore wing, except at the base, very dark purple with a silvery fascia across the middle, broadest on the dorsal margin. A second silvery fascia across the apex of the wing. The eilia at the extreme apex dark brown, elsewhere silvery grav. 3 mm. Ohio. Narrow serpentine mines in leaves of haw apple (*Cratacgus mollis*); two generations.

3. Nepticula pteliaeella Chambers. Tuft dark brown; eye-caps white. Thorax and base of fore wing at the dorsum silvery; a silvery faseia before the middle, a costal and opposite dorsal spot at three-fourths, on a dark brown ground eolor. Cilia silvery around the apex, becoming brown toward the dorsum. Hind wings dark brown. 4 to 4.5 mm. Kentucky; Ohio.

The larva is a miner in the leaves of the hop tree (*Ptelea trifoliata*); the mine (fig. 12), which is everywhere much contorted, is at first very indistinet, and sometimes blotch-like; later it becomes more distinct.

There are two generations a year. The larvæ may be collected in July and in August and September.

4. Nepticula quadrinotata Braun. Head dark brown; eye-caps silvery white. Thorax and fore wings dark brown; markings silvery white, consisting of an elongate spot at base of dorsum, a small spot on the costa before the middle, a larger triangular spot at the tornus, and a similar spot on the costa nearer the apex. 4 to 5 mm. Ohio; Kentucky.

The larva mines leaves of hornbean (*Carpinus caroliniana*) and hazel (*Corylus americana*). The mine is at first linear, usually closely following the midrib or one of the lateral veins; later doubling on itself for a short distance before it expands into an irregular, pale brownish blotch.

There are two generations a year: The larva mines in July and from late August to the middle of October, but is never common.

5. Nepticula trinotata Braun. Tuft ocherous, eye-caps whitish. Fore wings with deep blue reflections in the basal third, velvety back beyond, and somewhat irrorated in the apical third, the scales here having pale bluish iridescent bases. At the basal third on the costa is a white spot of variable size, faintly tinted with lilac in some lights. At the apical third there is a costal and an opposite dorsal spot, each shining white and larger than the spot at the basal third. 4.5 to 5 mm. Ohio.

The larvæ form blotch mines on *Carya cordiformis* and cccasionally on *C. ovata* The mine is at first an extremely narrow, linear tract, expanding into a broader tract, which, in turn, becomes a blotch.

There are two generations a year, the mines of the first appearing during the early part of July and those of the second generation, at the beginning of September.

 $\hat{\mathbf{0}}$. Nepticula bifasciella Clemens. Tuft ochraceous; eye-caps shining, cream color. Thorax and base of fore wings to the first fascia dark purple; beyond the first fascia, wings dark brown with bronzy reflections; the fasciæ silvery or golden according to the light, the first fascia at one-third, the second at two-thirds of the wing length. 4 to 4.5 mm.

The larva is a miner in leaves of wild cherry (*Prunus serotina*), and occasionally on wild plum (*Prunus americana*). The mine is much contorted, especially at first, often, by confluence, forming a blotch; later distinct. The leaf of wild cherry is discolored and reddish around the mine.

This species is one of the earliest to appear in the spring, the larvæ become full-grown by the middle of May; later generations appear in June and July and in September.

7. Nepticula intermedia Braun. Head black; eye-caps silvery white. Thorax bronzy, base of fore wing plum-purple, followed by a shining silvery or golden fascia. Beyond this fascia wing dark brown, with but faint bronze reflections. A second silvery or golden fascia crossing the wing at two-thirds. 3 to 3.5 mm. Ohio; Kentucky.

This species makes serpentine mines on leaves of sumac (Rhus spp.). Usually there are but two generations a year, the larvæ maturing in July usually over-wintering, but occasionally a third generation appears.

8. Inepticula rhamnicola Braun. Tuft ocherous in the summer generation, black in the overwintering generation. Thorax creamy buff, patagia dark brown. Fore wings brown, the tips of the scales blackish. At the basal third of the wing a cream-colored fascia with its edges often indented by dark scales. At twothirds of the wing length a more shining silvery fascia. 4.5 to 5.5 mm. Ohio.

The larvæ are found in the leaves of *Rhamnus lanceolata*; the mine is at first linear, contorted, and on lower surface; later it crosses to the upper side where it finally becomes a blotch. There are three generations; the mines are most abundant in October.

9. Nepticula cerea Braun. Head buff; eye-caps a little paler. Thorax and base of fore wings to just beyond one-third, creamy buff. From the base a few fuscous scales extending along the costa to the middle of the pale area, where they join a small, triangular, fuscous spot which is sometimes faintly connected with the dorsum by a few scattered fuscous scales. Following the pale basal area, a broad dark-brown band across the wing, succeeded by a narrower, silvery white fascia. The apical third of the wing dark brown, except for the pure white cilia at the apex. 3.5 mm.

Ohio; Pennsylvania.

10. Nepticula rhoifoliella Braun. Head black; eye-caps silvery white. Thorax blackish purple. Fore wings very lustrous, base of the costa plum-purple. A large, semi-elliptical patch of scales just beyond the base of the wing and resting on the dorsum, but not reaching to the extreme costa, of deep, brilliant golden.

shading along its edges into reddish bronze. Beyond this, wing deep purple with bronze reflections; wing crossed at three-fifths its length by a straight, shining, silvery or pale golden fascia. 3.5 mm.

Ohio; Kentucky; Missouri.

The larvæ make contorted, serpentine mines on the upper side of leaves of poison ivy (*Rhus toxicodendron*).

There are three generations; mature larvæ may be found in June, toward the end of July, and in September.

11. Nepticula terminella Braun. Tuít on the face dull brownish, on the vertex and head black; collar and eye-caps shining white, with a very faint yellow tinge. Thorax bronzy. Costal half of the fore wing to the fascia, blue-purple, the blue predominating at the extreme edge; below the costa the wing shading into a deep, brilliant, golden color, becoming more bronzy as it nears the fascia. Fascia situated just beyond the middle of the wing, almost straight, and with a brilliant, silvery luster. Apical third of the wing blue-purple, blue predominating. Just below the apex a double row of silvery seales margining the termen, becoming a single row toward the dorsum, and sometimes connected with the fascia. 5 to 5.5 mm.

Ohio; Kentueky; Pennsylvania.

The mine is seen on various species of oak, though most commonly on red oak (Quercus rubra) and pin oak (Q. palustris), it is a pale greenish, gradually broadening, linear tract, 3.5 mm. wide at the end. The larva is yellow, even when very young. Thus this mine can early be distinguished from the other linear mines on oak.

There are three generations a year, and in favorable seasons, a fourth.

12. Nepticula villosella Clemens. Tuft orange-ochraceous; eye-caps pale golden. Thorax and fore wings to the fascia brilliant, metallic bronzy, somewhat purple at the base of costa; fascia at two-thirds golden. Wing beyond the fascia bluepurple, with a large spot at the apex and the apical cilia golden, concolorous with the fascia. 4.5 mm.

The larva is a miner in leaves of blackberry (*Rubus* spp.) and occasionally wild raspberry (*Rubus occidentalis*). The mine is a tortuous, brown, linear tract scarcely broader than the pale brownish larva within.

There are three generations.

This is distinguished from all other species by the metallic golden apex.

13. Nepticula resplendensella Chambers. Palpi whitish, tuft pale reddish saffron. Fore wings, including cilia, with a brilliant metallic luster, golden or silvery, except the basal half of the costal margin, and a large spot extending along the base of the eostal eilia nearly to the tip and more than half way across the wing, which is deep purple. Tips of tarsi pale yellowish. 6 mm.

Kentucky.

14. Nepticula unifasciella Chambers. Head orange-ochraeeous; eye-caps silvery white. Upper surface of thorax and basal two-thirds of fore wing brilliant metallic bronzy or golden, except toward the costal margin where the color shades into purple, so that a silvery or golden fascia at the apical third is scarcely defined internally. Behind the fascia, wings deep purple. Cilia purple, golden at their tips. 4.5 mm.

Kentueky; Texas; Ohio.

15. Nepticula purpuratella Braun. Tuft ochraceous or orange; eye-caps silvery white. Thorax deep bronzy or golden. Extreme base of the fore wing concolorous with the thorax, shading outwardly to a paler, lustrous, golden color, and this, at the outer limits of the basal third, followed by a deep, bronzy band with purple and reddish reflections varying in intensity, and occupying approximately the middle of the wing. This followed by a brilliant, silvery fascia. Apical area beyond the fascia deep bronzy, usually suffused with brilliant purple. The purple

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reflections sometimes almost entirely absent, so that the dark band preceding the fascia and the apical area are deep bronzy-golden. 4.5 to 4.8 mm.

Pennsylvania.

16. Nepticula obscurella Braun. Tuft ocherous, eye-caps whitish. Fore wings shining golden brown, tinged with bronze along the extreme costa and in the apex. Just beyond two-thirds of the wing-length, an indistinct, narrow, whitish fascia, broadest in the middle of the wing and fading out toward the ends. Viewed from some angles, this fascia scarcely visible. Cilia of the general hue, their tips around the apex paler. however, and concolorous with the fascia. 3.5 mm.

New Jersev; New York.

The mine is a narrow serpentine track on the upper side of bayberry (Myrica carolinensis). There are two generations a year.

17. Nepticula ostryaefoliella Clemens. Tuft ochraceous; eye-caps and collar shining cream-colored. Thorax and fore wings shining brown with faint bronzy and purple reflections, which become deeper toward the predominantly purple apex. At two-thirds of the wing length is a shining silvery fascia. Cilia tipped with white around the apex. 4 mm. Pennsylvania; Ohio; Kentucky; North Carolina; British Columbia.

The mine is a rather broad, serpentine track, gradually increasing in breadth to the end, where it measures about 2 millimeters across; it occurs on various species of birch (*Betula* spp.) and on hop hornbeam (Ostrya). 18. Nepticula paludicola Brann. Distinguished from the preceding species by the following characters: tuft clay-colored or fuscous on the face, shading to buff

or cchraceous on the head; fascia slightly nearer the base. 3.5 to 4.5 mm.

New Jersey; Ohio.

The mine is a serpentine track on leaves of cranberry (*Oxycoccus macrocarpus*). 19. Nepticula altella Brann. Tuft orange-ochraceous in front, becoming pale behind; eye-caps creamy white. Thorax dark purplish brown. Fore wings before the fascia purple-brown, beyond it brown with purple reflections; general color to the naked eye deep purple before the fascia and brown beyond it. A silvery fascia crosses the wing at three fifths, and is usually a little broader on the margins of the wings. 6.5 to 7 mm.

Southwestern Ohio, locally in pin-oak forests.

The species has but one generation a year, the moths appearing in May. The mines are found only on the first leaves of the pin oak (*Quercus palustris*) that appear in the spring, never on the leaves that come later. The mine, which may best be regarded as a lower-side mine, is at first much contorted, winding and twisting within a small area, and causing a brownish discoloration of the surrounding leaf. This part of the mine seems to be formed early in the season, and the leaf around it is always dead when, in October, further feeding is resumed. At this time the larva starts out to mine into the fresh, green part of the leaf, where the mine is more distinct, due to the larva's partial eating of the leaf substance. The larva then becomes full-fed in a week or ten days and leaves the mine to spin a dark brown cocoon. The mine is extremely long but measures only 1 to 1.5 mm. in width at its end. The larva is yellow, with a row of dark brown dashes along the mid-ventral line.

This species may be separated from all other species by the fact that the wing is purple before the fascia and brown beyond, the reverse being true in all other cases where there is a difference in color before and behind the fascia.

20. Nepticula corylifoliella Clemens. Tuft ocherous to orange-ochraceons. Eyecaps silvery white, sometimes shading to fuscous outwardly. Thorax and fore wings bronzy brown, with blue-purple reflections toward the costa and in the entire apical part of the wing beyond the fascia, the color being there predominantly purple. The purple reflections sometimes entirely lacking proximal to the fascia. Fascia situated just beyond middle of wing, rather broad, narrowing toward the costa. Cilia silvery-tipped at the apex. 3.5 mm.

The food plants of N, corylifolicilla include hazel (Corylus americana), hop hornheam (Ostrya virginiana), hornbeam (Carpinus carolinina), and black birch (Betula lenta). The mine is a long, very narrow, winding tract, searcely broader than the larva. The larvae may be found in June and early July, and from late Angust until October.

21. Nepticula opulifoliella Brann. Tuft ochraceous; collar usually pale yellowish; eye-caps pale, shining buff, sometimes fuscous ontwardly. Thorax dark fuscous, with purple and blue reflections. Fore wings with pronounced purple and blue reflections toward the costa and beyond the fascia, shading to bronzy green below the fold. At three-fifths of the wing length a broad, very shiny, silvery fascia with faint, golden luster. 3.5 to 4 mm.

Ohio; North Carolina.

The larvæ form brownish, much contorted, serpentine mines in leaves of Opulaster (*Physocarpus opulifolius*). There are two generations, the larvæ appearing in July and September,

This species is very close to N, corylifoliella, from which it is difficult to distinguish it. The pale collar, when present, is a reliable character. The more yellowish eye-caps, and the less lustrons wing, with the absence of reddish tints, will aid in distinguishing this species from N, corylifoliella.

22. Nepticula quercipulchella Chambers. Head black: collar and eye-caps yellowish white, silvery; thorax and fore wings deep blue-black, bronzed, and with purple and violet reflections; the fascia behind the middle, silvery white and a little the widest on the dorsal margin; wing behind the fascia darker than before it, but cilia paler and less lustrous than wing. Under surface of wing, abdomen, and legs cupreous black. 4 mm.

Kentneky.

The above description is essentially that given by Chambers. According to him "the larva is bright green, with a deeper green line of contents; it makes a long, narrow, winding, and gradually widening track, similar to that of *N. quereicasta-nella* Chambers in leaves of *Quereus alba*." Chambers asserts that the larva from which he bred the type specimen formed a new mine when nearly grown, a fact, if true, at variance with all observations on this group.

23. Nepticula juglandifoliella Clemens. Tuft ochraceons, eye-caps and collar shining creamy white. Thorax and fore wings deep purplish black, uniformly purple beyond the fascia which is situated just beyond the middle; silvery white and broadest on the dorsum. 3.5 to 3.8 mm.

The mine is a serpentine track, usually whitish, and is found on various species of hickory and on walnut and butternut. The larva is pale green (almost white when feeding on walnut).

There are three generations. Full-grown larvæ may be found in the middle of June, the latter part of July, and in late August and early September.

The purple fore wings and white collar distinguish this species from its nearest allies.

24. Nepticula apicialbella Chambers. Tuft ochraceous, collar ereamy white, eye-caps white. Thorax dark purplish brown. Fore wings dark brown, with a faint, purple luster. Beyond the middle of the wing is a narrow, oblique white fascia, convex outwardly and reaching the margin farther from the base on the dorsum. The scales at extreme tip of wing white, forming, with the whitish apical cilia, a very distinct pale patch. 4 mm.

Kentucky; Ohio.

The larvæ make upper-side, serpentine mines on elm. The mine is brownish in color, with a conspicuous line of frass through the middle.

There are three generations. The full-grown larvæ are found in mid-June, late July, and in August and September.

The oblique fascia and white apex distinguish this species from all others.

25. Nepticula tiliella Braun. Tuft ochraceous, collar white. Antennæ black, eye-caps shining white. Fore wings almost black, with a very faint, purple luster. At the middle of wing a shining, pure white fascia, slightly oblique and a little convex outwardly. Cilia gray on the dorsum, shining white from the tornus to the costa. 3.5 mm.

Ohio; Kentucky.

The larvæ make serpentine mines on the upper side of leaves of basswood (*Tilia americana*). The mine is characterized by the tendency toward a spiral form, with either the early or the later part inside, and by the frequency of angular turns.

There are two generations, the full-grown larvæ appearing in early July and late August.

The pure white fascia and the white cilia, against which the outermost row of black scales is sharply defined, distinguish this species.

26. Nepticula rubifoliella Clemens. Head ocherous; eye-caps silvery white. Thorax and fore wings almost black, with a shining silvery, or faintly golden, fascia at the middle of the wing; fascia convex outwardly and somewhat nar rowed at its middle, sometimes almost interrupted. Cilia whitish, so that the marginal line of scales is defined. 4 mm.

The larvæ mine leaves of blackberry, forming at first very narrow, linear mines, which closely follow a vein or the margin of the leaf before enlarging into an irregular blotch.

Mines containing the larvæ may be collected in July and September.

27. Nepticula nyssæfoliella Chambers. Tuft ochraceous; eye-caps shining white. Thorax and fore wings black with very faint, purple reflections. In the middle of the wing a shining silvery, or pale golden, fascia, slightly convex outwardly. Cilia around the apex white, with marginal line of scales defined. Hind wings pale gray, with an oval patch of androconia in the male. 4.5 to 6 mm.

The larvæ mine in the leaves of sour gum (Nyssa sylvatica), forming narrow, linear mines (fig. 59) which abruptly enlarge into blotches measuring 2 cm. or more in length, with an average width of 5 or 6 mm. There are two or three generations a year, the larvæ of the first generation becoming full-grown in June.

This is one of the most abundant species; moths may often be collected in great numbers in the vicinity of the food plant.

From *N. rubifoliella*, its nearest ally, it is distinguished by its larger size and the equal breadth of fascia throughout.

28. Nepticula slingerlandella Kearfott. Tuft ochraceous, becoming paler behind, where it, merges into the pale ocherous or whitish collar. Eye-caps white. Thorax and fore wings black with a faint bronzy luster, somewhat irrorated beyond the shining white fascia situated just beyond the middle of the wing. Cilia pale gray. 3.5 to 5 mm.

New York; Ohio.

The larvæ mine leaves of cultivated plums and prunes, wild plum (*Prunus americana*), and occasionally sweet cherry, forming narrow, linear mines which abruptly enlarge into irregular blotches. This species attains economic importance in the plum orchards of northern New York, where its ravages have been the subject of a bulletin by C. R. Crosby, in which are given further details of its life history, together with numerous figures. There is a single generation of moths in New York, and the larvæ which are full grown in July do not produce moths until the following year. Farther south, a second brood of larvæ may usually be collected in September.

29. Nepticula rosæfoliella Clemens. Tuft ochraceous; eye-caps shining creamy white. Fore wings almost black, with a very faint, dark blue and bronzy luster. Just beyond the middle of the wing is a rather broad, straight, silvery, or very pale, golden taseia. Cilia of the general hne, searcely paler tipped opposite the apex. 4.5 mm.

The larva mines the leaves of various species of rose. The mine is serpentine, usually much contorted, and frequently closely follows the edge of the leaf in its early course. A broad line of frass is visible. The mine at its end measures 1.5 to 2 mm. across. There are three generations, the larva being full-grown in June and early July, in August, and in October. Mines containing larvae may, however, he found at almost any time during the summer and fall up to November.

This species may be distinguished from N. slingerlandella by the darker head, dark cilia, and somewhat broader wings.

30. Nepticula fuscotibiella Clemens. Tuft ochraceous; eye-caps whitish. Thorax and fore wings fuscous, faintly purple, with the scales before the fascia paler at their bases, so that this part of the wing is somewhat irrorated and paler than that beyond the fascia where the wing is dark fuscous purple. Fascia just beyond the middle, dull white, sometimes a little convex and broadening on the dorsum. Cilia gray, pale gray around the apex. 4 to 4.5 mm.

The larvæ mine leaves of various species of willow. The mine is a gradually broadening, linear tract, sometimes straight, but often bent back on itself toward the end. Occasionally (on *Salix discolor*) its latter part is a more or less spiral blotch. There are at least three generations a year. The larvæ may be collected from June until the end of October.

31. Nepticula ulmella Braun. Tuft ocherous on face, tinged with red above, and sometimes with a few dark brown scales behind. Antenne creamy white, broadly banded above with dark brown, so that only a narrow line of the pale color appears between the annulations. Eye-caps creamy white. Thorax brownish, somewhat peppered. Scales of the fore wing creamy white, shading to dark brown at their tips, except where they form a creamy white, oblique fascia at the middle of the wing. The general color of the fore wing is thus a somewhat mottled, dark brown. Fascia, from the middle of the wing on the costa, extending to dorsum somewhat behind the middle, and sometimes broken with a few, dark-tipped scales. Cilia creamy white. 4 to 5 mm.

The larvæ are miners in leaves of red elm and cork elm (Ulmus fulva and U. racemosa). The mine starts as a very fine brown, or, rarely, whitish, line, abruptly enlarging to a breadth of 1 mm., then increasing gradually in width, until it attains a breadth slightly in excess of 2 mm. The broad portion of the mine is usually so much contorted that it is not possible to trace the course of the mine, the whole having the appearance of an irregular blotch.

There are two generations, the mature larvæ being found in July and in September.

The creamy white fascia and cilia, together with the pale bases of the scales, distinguish this species.

32. Nepticula platanella Clemens. Tuft pale ocherous to ochraceous; eye-caps silvery white. Thorax and fore wings dark brown with a bluish luster. At middle of costal margin a small oblique silvery streak, and opposite it on the dorsal margin is a similar streak, usually larger than the costal streak and broader on the margin. Rarely both spots are very minute. Occasionally these two streaks meet, forming a more or less interrupted concave fascia. Last row of scales around the apex pale yellowish at their bases, thus forming a dark line in the whitish eilia. Hind wings yellowish fuscous, in the male with a swordshaped, yellowish, chitinous plate on the upper side from base to near middle of costa, margined along the costa with bristly black scales. Beyond this costa excised. 5.5 to 7 mm.

The mines are abundant on leaves of sycamore (*Platanus occidentalis*) and begin as slender, linear tracks usually filled with frass. Several days before pupation, the mine is abruptly enlarged into a large, usually almost circular blotch, which in many instances covers the linear part of the mine.

There are three generations of larvæ, the earliest being found during June.

There is considerable variation in the size of the white spots and it is but rarely that they form a fascia. Females may be distinguished from specimens of N. clemensella by their larger size, and the males from that species and all others except N. similella, by the peculiar chitinous plate along the costa of the hind wing.

33. Nepticula clemensella Chambers. Tuft ochraceous; eye-caps silvery white. Fore wings bluish black. A narrow, oblique, silvery streak on the middle of the costa (rarely inconspicuous) and an opposite dorsal streak, usually meeting in the male to form a narrow, oblique fascia. Cilia silvery with a brown line formed by the dark tips of the terminal row of scales around the apex. Hind wings vellowish fuscous, similar in both sexes. 4.5 to 5.2 mm. The larva mines in the leaves of sycamore (*Platanus occidentalis*), forming a

linear mine which gradually increases in breadth. Its terminal portion expands into a small blotch, three or four times the diameter of the end of the linear mine. There are three generations a year.

This species is much less common than N. platanella and uniformly smaller, the

largest specimens scarcely attaining the expanse of the smallest N. platanella. 34. Nepticula similella Braun. Tuft ocherous to ochraceous; eye-caps silvery white, occasionally tinged with ocher. Thorax and fore wings deep bluish black, the extreme bases of the scales more or less iridescent blue, especially in the apical half of the wing. At the middle of the wing an oblique narrow, costal streak, shining white but not silvery, usually meets the apex of a shorter, broader, dorsal streak. Tips only of the last row of scales around apex dark, thus forming a dark line in the white cilia. Hind wings gray; in the male with a narrow, chitinous plate from base to one-third of costa, with a long, yellowish, costal hair-pencil lying along it. 5 to 6 mm.

Ohio; Kentucky.

The larva makes a characteristic mine in the leaves of pin oak (Quercus palustris) and occasionally of chestnut. The early part of the mine is very narrow, completely filled with frass, and bent several times in close, S-shaped curves. The larva next mines just above the lower epidermis, forming a blotch scarcely visible above except for occasional spots here and there toward the edges of the blotch, where the leaf substance is more fully consumed. During the last stage, a conspicuous large blotch is formed, where the mine is transparent and whitish, with the frass accumulated toward the beginning of the blotch.

Females of this species can only be distinguished from those of N. platanella by their less shining costal and dorsal spots; the males differ in the yellowish costal hair-pencil of their hind wings.

35. Nepticula thoracealbella Chambers. Tuft ochraceous on the face, becoming reddish brown on the vertex; eye-caps white. Thorax white, with a few, scattered, dark brown scales in occasional specimens. Fore wings dark brown, slightly irrorated; a creamy white, irregular fascia just before the middle, concave toward the base and usually wider on the dorsal margin; at the apical fourth a distinct, creamy-white, costal spot, and an opposite dorsal spot, whose apices occasionally touch; cilia creamy white, sometimes grayish on the dorsum. 4 to 5 mm.

Kentucky; Ohio; Pennsylvnia.

36. Nepticula pomivorella Packard. Tuft orange-ochraceous; eye-caps and collar shining pale buff. Thorax and fore wings shining bronzy, with strong purple and blue reflections increasing toward the apex. 5 mm.

The larvæ make long, narrow, serpentine tracks (fig. 61) in the leaves of apple, gradually widening the mine to 2 or 2.5 mm. at the end.

37. Nepticula chalybeia Braun. Tuft ocherous, sometimes shading to reddish brown above; collar yellowish white; antennæ fuscous; eye-caps yellowish white. Thorax steel gray. Fore wings very narrow, steel gray with faint, greenish golden reflections. 3.5 to 4 mm.

Ohio.

The larvae mine leaves of wild pear (*Pyrus communis*) and cultivated pear, making rather short, serpentine tracks, often not exceeding 2 cm, but sometimes reaching 3 cm, in length, and broadening to 1.5 to 2 mm, across at the end. There are three generations a year. Mined leaves may be collected in early June, in July, and during the last part of August.

Its paler color, with the absence of purple, the narrow wings, and its smaller size distinguish this species from *Nepticula pomirorella*.

38. Nepticula flavipedella Brann. Tuft usually dark brown, collar creamy white; rarely tuft reddish ocherons on the face, and brown on the vertex; eye-caps creamy white. Thorax dark purplish brown. Fore wings dark brown, with dark blue and purple reflections; cilia with silvery tips. Fore legs, except the femora, dark brown; middle legs pale silvery, tarsi pale ocherous; hind legs silvery, tibiz dark brown, tarsi pale ocherous, 3.5 to 4.5 mm.

Ohio; Kentucky.

The mine is a very characteristic linear tract, found most commonly on pin and swamp white oak, but occasionally on other species of oak. The larva, for the first few millimeters, mines near the upper surface, making a very narrow indistinet mine. Then the mine is slightly but abruptly enlarged and for a length of 8 or 9 mm., the leaf substance is entirely consumed and the mine rendered transparent. Then follows another enlargement, and the mine, often much contorted, increases very gradually in breadth to the end where it measures 2 mm. across. The latter part of the mine is not transparent, but the mine is distinctly visible.

There are three generations a year. Mined leaves may be collected during early June, the latter half of July, and the early part of September.

Though very distinct in larval work, this species in the imaginal state is almost indistinguishable in appearance from *N. castaneafolicila*. The yellowish middle and hind tarsi and the deeper purple suffusion of the fore wings are, however, constant differences.

39. Nepticula castaneæfoliella Chambers. Tuit black; collar, eye-caps, and palpi creamy white. Thorax and fore wing dark brown, with slight bronzy and purple reflections; tips of scales somewhat darker, so that under a lens the wing is slightly irrorated, especially toward the apex. Cilia silvery at the tips. Posterior tible and fore legs, except the femora, dark brown; legs otherwise whitish. 4 to 4.5 mm.

Kentucky; Ohio; Virginia.

The larvæ form very long, much contorted, linear mines in the leaves of chestnut (*Castanea dentata*) and oak. They measure but little over 1 mm. in width at the end, and have a fine, central line of frass.

40. Nepticula latifasciella Chambers. Tuft on the face ocherous, dark brown on the vertex; collar and eye-caps creamy. Thorax and extreme base of fore wings creamy-buff. Remainder of fore wing deep purple-brown, with a very broad, creamy-buff fascia just before the middle; two or three creamy-buff scales at the extreme apex, forming with the creamy-white cilia around the apex, a conspicuous pale spot. Cilia clsewhere gray. 4 to 4.5 mm. The larvæ mine leaves of oaks, and probably chestnut. Although the mine

The larve mine leaves of oaks, and probably chestnut. Although the mine varies in length from 3 to 5 cm. on different species of oak, with a breadth of about 1.5 mm. at its end it has in general the same appearance. The frass is at first deposited in a broad, blackish line through the center, later dispersed across the entire breadth, and, toward the end, collected into a broad band.

This is one of the carliest species to appear in the spring; the moths may be found resting on tree trunks during the latter part of April. Mines may be found in June, the latter part of July, in September, and often during late October.

41. Nepticula cratægifoliella Clemens. Tuft ocherous, faintly tinged with red above. Antennæ ocherous, partly suffused with fuseous; eye-caps ocherous. Thorax

and fore wings ocherous, the extreme edge of the costa near the base purplish fuscous, and a bread, purplish fuscous band at the apex of the wing. The cilia, beyond this band, pale ocherous, giving the appearance of an ocherous apex preceded by a dark band. Cilia opposite the ends of the band concolorons with it. 3.5 to 4.5 mm.

Pennsylvania; Ohio; Kentucky.

The mines occur on several species of hawapple. The mine is comparatively short, rapidly increasing in diameter and measuring about 2 mm. in width in the later part of its course.

There are two generations a year. Larvæ may be collected in early July and at the beginning of September.

This species is distinguished from all others with yellowish ground color by its entire absence of dusting. The much broader mine and bright green larva distinguish it, in its early stages, from *N. scintillans*.

42. Nepticula nigriverticella Chambers. Face ochraceous, tuft above dark brown; collar and eye-caps pale ocherous, antennal stalk fuscous. Thorax and fore wings pale ocherous, dusted with purplish black scales. At the base of the dorsal margin a purplish black spot extending halfway across the wing, and occasionally, as a narrow line, reaching the costa, which is often dark brown near the base. At the beginning of the cilia a broad, purplish-black fascia; beyond it cilia pale ocherous. 5 mm.

Kentucky; Ohio; Texas.

43. Nepticula populetorum Frey and Boll. Tuft ocherous, becoming dark brown behind. Collar and eye-caps pale creamy. Thorax and fore wings buff or pale ocherous, more or less densely dusted with purplish fuscous scales. These scales form a purplish fuscous fascia at the beginning of the cilia. Cilia pale gray, whitish around apex. 5 mm.

Texas; Ohio; Kentucky; California.

The larvæ mine the leaves of several species of poplar, commonly the leaves of cottonwood (*Populus deltoides*). The mine is indiscriminately placed on the upper or the lower side of the leaf. It is whitish, gradually broadening, linear track, 2 to 2.5 mm in width at its extremity.

This species differs from the preceding in the absence of the dark spot at the base of the dorsum.

44. Nepticula saginella Clemens. Face ocherous or pale buff, head, above, dark brown; collar and eve-caps pale ocherous. Thorax and fore wings pale ocherous, buffish, or even whitish, and dusted with fuscous scales, often more densely dusted toward the outer half of the wing, where the dark scales are either evenly distributed or collected into spots, but never form a band. Cilia pale ocherous. 4 to 5.5 mm.

The mine (fig. 60) is a whitish, linear tract on various species of oaks and on chestnut; it varies in length and width, but is usually about 1.5 to 2 mm. wide at its extremity. The larvæ are found from June to October.

45. Nepticula pallida Braun. Tuft ocherous; the scales on the vertex tipped with orange; antennæ pale ocherous, eve-caps whitish. Fore wings very pale buff, evenly dusted with purplish gray, a little more densely toward the apex of the wing. Cilia very pale buff. 4 mm.

Cedar Point, Ohio.

The food plant of N. pallida is willow, Salix sp. The mine (fig. 58) is made on the lower side of the leaf and is extremely narrow at first, extending along the midrib, later doubling on itself once or twice, and gradually and evenly increasing in breadth to its end, where it measures a scant 1.5 mm. across. The entire length of the mine is approximately 4.5 cm.

The pale head distinguishes this species from N. sagincila.

In addition to the above species, there are several species, namely N. amelanchierella, N. anguinella and N. platea, which are still known only in the larval state. Their mines are described in the Synopsis of species by food plants. Synopsis of species by food plants

Salix spp., willow:

(1) N. pallida; mine linear, narrow at the end.

(2) N. fuscotibiclla: mine linear, gradually broadening, club shaped at the end, sometimes blotch-like toward the end.

Populus grandidentata, poplar:

(1) E. populella; globular swelling of petiole close to leaf.

Populus deltoides, cottonwood:

(1) N. populctorum; whitish mine, gradually increasing to a breadth of 2 to 2.5 mm. at end.

Myrica carolinensis, bayberry:

(1) N. obscurella; mine serpentine, very narrow.

Juglans cinerea, butternut:

(1) N. juglandifoliella.

Juglans nigra, walnut:

(1) N. juglandifoliclla; serpentine mine, very gradually increasing in breadth. Carya spp., hickory:

N. juglandifoliella; serpentine mine, very gradually increasing in breadth.
 N. tringtata; linear track, expanding into a blotch.

Corylus americana, hazel:

(1) N. corylifoliella; very narrow, serpentine mine, searcely broader than the larva.

(2) N. quadrinotata; narrow linear mine, expanding into an irregular blotch. Ostrya virginiana, hop hornbeam:

- (1) N. corylifoliella; very narrow, scrpentine mine, scarcely broader than the larva.
- (2) N. ostructoliella; linear track, gradually reaching a breadth of 2 mm., at the end.

Carpinus caroliniana, hornbeam:

(1) N. corylifoliella; very narrow, serpentine mine, scarcely broader than the larva.

(2) N. quadrinotata; narrow linear mine, expanding into an irregular blotch. Betula spp., birch:

- (1) \hat{N} . corylifoliella; very narrow, screentine mine, scarcely broader than the larva.
- (2) N. ostryafoliella; linear track, gradually reaching a breadth of 2 mm. at the end.

Castanea dentata, chestnut:

- (1) N. castaneæfolicila; long, contorted mine, with central line of frass; larva green.
- (2) N. saginella; shorter, whitish mine, with frass in a central line or dispersed; larva green.
- (3) N. latifasciella; serpentine minc; frass at first in a broad line, later, dispersed, and toward end gathered into a band; larva green.
- (4) N. similella; see under Quereus (8).

(5) E. castanea; larva makes a gall encircling twig.

(6) E. phleophaga; larva makes a serpentine track in the bark.

Quercus spp., oaks:

- (1) \overline{N} , terminella; mine a broadening, linear tract, 3.5 mm, wide at the end; larva yellow.
- (2) N. saginella; whitish, linear mine with frass in a central line or dispersed; larva green.
- (3) N. latifasciella; serpentine mine; frass at first in a broad line, later, dispersed, and toward the end gathered into a band; larva green.
- (4) N. flavipedella; linear mine; a short, indistinct part followed by a transparent area, and then a gradually broadening, serpentine track, not transparent, but easily visible above; larva green.

- (5) N. anguinella Clemens, Proc. Ent. Soc. Phila., vol. 1, p. 85, 1861; Tin. No. Am., 175, 1872. May be found in the leaves of oaks early in October and in the latter part of June. The mine is a very narrow serpentine tract, which is filled or discolored throughout its length by blackish excrement. The larva fits the mine closely, in color lemon-yellow, with ten square dark brown or blackish spots on the ventral surface."
 - It should be possible to recognize this species, when found, from Clemens' description of the larva.
- (6) N. platea Clemens, Proc. Ent. Soc. Ph.a., vol. 1, p. 85, 1861; Tin. No. Am., 175, 1872. "Mines oaks early in October. The mine is a moderately broad, winding tract, with a broad line of dispersed grains of excrement. The larva is purplish, with a pale green vascular line and a row of reddish-brown dorsal dashes. The mine is much broader than that of the preceding miner."
 - The statement that the larva is purple cannot be regarded as conclusive, since such color is often produced in larvæ feeding on leaves with autumnal coloration.

Quercus alba, white oak:

(7) N. quercipulchella; long, narrow, winding and gradually widening track; larva bright green.

Quercus palustris, pin oak:

(8) N. similella; linear mine, expanding into an underside blotch, followed by a large, conspicuous, nearly transparent blotch; larva pale green.

(9) N. altella; lower-side, very long, serpentine mine, not distinctly visible; larva yellow with row of dark brown dashes.

(10) E. heinrichi; flattened-oval, spiral mine in bark of young branches. Ulmus spp., elms:

(1) N. apicialbella; narrow, serpentine mine.

(2) N. ulmella; narrow, linear track, enlarging into an irregular blotch.

Platanus occidentalis, sycamore:

- (1) N. platanella; narrow, linear mine, abruptly enlarging into a large blotch.
- (2) N. clemensella; linear mine, gradually broadening with terminal portion expanded into a small blotch.

Physocarpus opulifolius, Opulaster:

(1) N. opulifoliella; brownish, contorted, serpentine mine.

Pyrus communis, pear:

(1) N. chalybeia; serpentine mine.

Pyrus malus, apple:

(1) N. pomicorella; serpentine mine, usually broadening to 2 or 2.5 mm. Amelanchier canadensis, service berry:

(1) N. amelanchierella Clemens, Proc. Ent. Scc. Phila., vol. 1, p. 84, 1861; Tin. No. Am., 174, 1872. "In the leaves of service-berry or Juneberry, Amelanchier canadensis, in June and July. The mine rather a broad tract, sometimes much contorted, with rather irregular edges, placed most often towards the base of the leaf and having a rather broad "frass" line of a dark brown color."

This species seems to be distributed wherever its food plant occurs; mines observed in Ohio, Kentucky, North Carolina.

Cratægus spp., hawapple:

- (1) N. cratægifoliella; serpentine mine, with a breadth of about 2 mm. at the end.
- (2) N. scintillans: contorted, serpentine mine, scarcely exceeding 1 mm. in breadth.

Rubus spp., blackberry, raspberry:

(1) N. villosella; narrow, serpentine mine.

(2) N. rubifoliella; short, narrow, linear mine, expanding into a blotch. Rosa spp., rose:

(1) N. rosæfoliella; serpentine mine.

Prunus serotina, wild black cherry:

(1) N. bifasciella; narrow, serpentine mine.

Prunus americana, wild plum:

(1) N. bifasciella; narrow, serpentine mine.

(2) N. slingerlandella; narrow, linear mine, abruptly enlarging into an irregular blotch (also on cultivated plums and prunes, and sweet cherry). Ptelea trifoliata, hop tree:

(1) N. pteliaella; very long, much contorted, narrow, serpentine mine. Rhus toxicodendron, poison ivy:

(1) N. rhoifoliella; narrow, contorted, serpentine mine.

Rhus spp., sumac: (1) N. intermedia; narrow, contorted, serpentine mine. Acer saccharum, sugar maple:

(1) Glaucolepis saccharella; very long, linear mine. Acer rabrum, red maple:

(1) Glaucolepis saccharella; very long, linear mine. Rhamnus lanceolata, buckthorn:

(1) N. rhamnicola; linear mine, expanding into an irregular blotch. Tilia americana, basswood:

 N. tilliella; serpentine mine.
 N. argentifasciella; indistinct, linear mine, expanding into a blotch. Nyssa sylvatica, sour gum or pepperidge:

(1) N. nyssæfoliella; linear mine, abruptly expanding into a blotch.

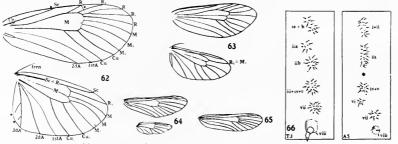
Oxycoccus macrocarpus, cranberry:

(1) N. paludicola; serpentine track, in part following the margin of the leaf.

SUPERFAMILY ZYGÆNOIDEA

(Slug caterpillars)

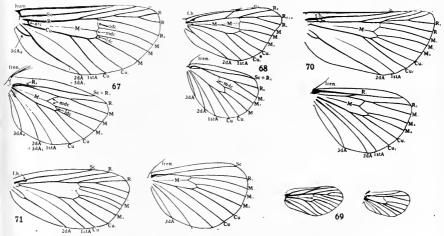
This series, which seems to be derived from a stock intermediate between the Hepialidæ and the lowest known Tineidæ, apparently became adapted very early, in the larva, to external feeding. The



FIGS. 62-66. ZYGÆNOIDEA: PYROMORPHIDÆ

62, Pyromorpha dimidiata, Q, venation. (The asterisks indicate accessory veins, the other abbreviations as in figure 19.); 63, Acoloithus falsarius, venation; 64, Harrisina texana, venation; 65, Harrisina americana, venation of fore wing; 66, Zygæna trifolii (Zygænidæ, Europe), seta map

families Eucleidæ and Megalopygidæ, with their extralimital relatives, the Dalceridæ and Aididæ, are closely related; the Zygænidæ and Pyro-



FIGS. 67-71. ZYGÆNOIDEA: EUCLEIDÆ

67, Phobetron pithecium, \mathcal{Q} , venation; 68, Adoneta spinuloides, venation; 69, Euclea delphinii, venation; 70, Prolimacodes scapha, venation; 71, Packardia geminata, venation

morpidæ form a second group, but the western and exotic family Epipyropidæ is not closely related to either, and in fact seems more closely connected with the Tincoidea.

The group is distinguished primarily by the larva (fig. 66) which has tubereles i and ii united into a single wart and likewise iv and v; but in most Eucleidæ these warts have been lost, and the body may be smooth. In the adult the venation is quite complete, including the

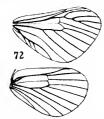


FIG. 72. ZYGÆNOIDEA: MEGALOPYGIDÆ Lagoa crispata, venation

base of media (not always forked) and the entire length of **1st A** in both wings. The maxillary palpi are rudimentary, and usually the tongue as well. In the Zygænidæ and Pyromorphidæ the tongue is strong. The wings are ample, of the macro type, with short fringes; the scaling sometimes soft, but fairly firm in the otherwise rather Yponomeutid-like Zygænidæ. The frenulum of the female is a tuft of many bristles.

The egg is always of the flat type, and is very thin and wafer-like in the Eucleidæ (except the primitive genus Monoleuca). The larva

is always more or less shortened and slug-like, with a retractile but large head. The hooks of the prolegs, when present, are in a single, uniordinal band. The two epicrania only unite for a very short distance above the front, leaving a large, triangular, or more narrow, slit-like space, filled with cervical membrane. Typically, the epicrania extend internally into the neck, as in leaf-miners. The pupa is of a low incomplete type, with thin skin and more or less movable appendages. Abdominal segments 3 to 6 are free, and 7 also in the male. The first abdominal spiracle is uncovered (as also in the Nepticulidæ). The maxillæ have a strong, lateral extension at the base, sometimes reaching the antennæ, but lack a separate maxillary palpus. In Lagoa the maxillæ are reduced and divergent, much like those of the Hepialidæ. The abdominal segments are armed with scattered spines. The cocoon is of silk, usually rather parchment-like, with a more or less specialized trap-door at the anterior end for the emergence of the pupa. The larva hibernates in the cocoon, pupating in the spring.

4

Family 6. **MEGALOPYGIDÆ**

(Lagoidæ; Liparidæ, in part)

(The flannel moths)

Closely similar to the Eucleidæ. Male, and sometimes female, antennæ pectinate to the tip; fore wing (fig. 72) with vestiture often crinkly, of more or less hairlike scales. Hind wing in our species with **Sc** and **R** fused most of the length of the cell, but free at base, unlike the North American Pyromorphidæ, in which also \mathbf{R}_4 and \mathbf{R}_5 of the fore wing are stalked. Mouth parts much reduced, and buried in dense hair.

Egg ellipsoidal. Larva with stinging hair mixed with dense, soft hair, arranged in diffuse tufts, ventrally with a few subprimaries only. Prolegs, without hooks present on abdominal segments 2 and 7, beside the normal ones on segments 3 and 6 and the last segment, which each have a uniordinal row of hooks, sharply angulated in the middle, or interrupted by a more or less distinct sucker. Pupa with all abdominal segments free (except the usual terminal ones). Appendages free. Maxillæ rudimentary, divergent, quadrangular as in the Hepialidæ, leaving the labial palpi wholly uncovered. Abdominal segments 2 to 6, inclusive, with conical tubercles behind the spiracles. Abdomen with fine, spinulated bands on anterior part of segments and a tuft of bristles on each side of dorsum behind them. Eyes and first spiracles as in the Eucleidæ.

This small family is almost wholly American.

Key to the genera

1. MEGALOPYGE Hübner

(Lagoa Harris)

Fore wings with discolored, crinkly hair toward costa. Frenulum rudimentary. Caterpillar with hair from tufts rather long and spreading so as nearly to conceal the body.

Key to the species

1. Wholly orange ochrepyxidifera.
1. Crinkly hair in part contrasting, brown and black.
2. Thorax ochre yellow to brown
2. Thorax cream colorcrispata.
I. Male antennae half as long as fore wing, of female about a third as long
as fore wing; cocoon with two transverse flanges on upper side.
(Megalopyge).
1. M. opercularis Smith and Abbot. Thorax and basal tuft of abdomen ochre;

crinkly hair, a mixture of light, chocolate brown and white, blackish on costa; outer margin yellow; fore legs blackish with white at apex of tibia. 25-35 mm. (H 38:25.)

Caterpillar with little, curled tufts on each side of tail.

The moth occurs normally from Maryland south. A stray has been found at Ithaca, New York.

II. Male antennæ two-thirds as long as fore wing, female over one-third; cocoon oval; larva without curled tufts of hair at tail. (Lagoa).

2. M. pyxidifera Smith and Abbot. Wholly ochre yellow. 25-35 mm. H 38:41. Caterpillar grayish white when young, gray in last stage; on oak. Southern States.

3. M. crispata Packard. Cream color; crinkly hair black and brown, most of it along the costa. Fore legs pale with black tips. 25-35 mm. (H 38:23).

Caterpillar cream white when young, dull red shading into smoky gray in front in the last stage; on various shrubs and trees.

In aberration grisea Barnes and McDunnough, the ground color is mouse gray with a contrasting pale fringe.

Massachusetts and southward. New York: Onteora Mountain, New Baltimore. Bethlehem, Karner, Albany, Rhinebeck, Poughkeepsie, New Windsor, Katonah, Staten Island; Brooklyn, Woodhaven, and elsewhere, Long Island.

2. NORAPE Walker

(Carama Walker; Lagoa, subgenus Ulosota Grote)

Frenulum well developed.

1. N. ovina Sepp. Pure white, with a little crinkly hair. .30 mm. (cretata Grote, pura Butler). (H38:22.)

Larva with sparse tufts of hair and contrastingly spotted body, on Celtis. District of Columbia and southward.

Family 7. EUCLEIDÆ

(Cochlidiidæ, Limacodidæ)

(The slug caterpillars)

Normally small, stumpy-winged moths with deep, often woolly vestiture. Head small and retracted, palpi fairly well developed, tongue and maxillary palpi much reduced. Thorax of low type, with metascutum narrowly continuous. Fore wing with two, and hind wing with three, completely preserved anals, base of media preserved, sometimes forked. Venation complete or nearly so; **Cu** 4-branched, \mathbf{R}_3 , \mathbf{R}_4 , and \mathbf{R}_5 stalked or united in fore wings; **Sc** and **R** in contact or fused for a short distance at or before middle of cell in hind wing, rarely merely connected by the apparent crossvein \mathbf{R}_1 .

Egg of the flat type, very thin and waferlike except in Monoleuca. Larva slug-like, with the prolegs replaced by suckers. Head large but permanently concealed in the thorax, save for a slit in the under side of the prothorax to allow feeding; head lightly chitinized, the vertex cleft almost to the front. Labrum with ii much higher and smaller than i. Larvæ in primitive forms with long, hairy, more or less stiff lappets,

or with strong spines provided with stinging hair, in the specialized forms with smooth bodies and practically all the hair lost. Thoracic legs present but small. Pupa of a low incomplete type, apparently with first abdominal segment free, as well as all appendages; diffuse spinulation all over dorsum of abdomen; collar projecting over head in a pair of subdorsal lobes, and mesoscutum extending back across metathorax to abdomen. Maxillæ with tongue-case extending along labial palpi, but not to their end, with a lateral projection at base, reaching in Prolimacodes under the eye to the antenna. Eye strongly sculptured, and forming a movable cap, covering end protecting the first spiracle, which is ventral, not dorsal as in most families.

The larva hibernates in the cocoon. The cocoon is oval, simple, and has a hinged lid for emergence.

There are about 400 species, the Phobetron group being American, the spined group, largely tropical in both hemispheres, and the group with smooth larvæ, north temperate.

Key to the genera (males)

1. Antennæ pectinate at base, with serrate or simple outer half. 2. Fore wings lacking one vein (fig. 68). 3. Inner margin of fore wings evenly convex. 1. Antennæ pectinate nearly or quite to apex. 2. Palpi not reaching vertex. 3. Head strongly retracted, male smoky with translucent wings, wholly unlike female.....l. Phobetron 3. Head fairly prominent, sexcs similar. 4. Hind tibiæ with terminal spurs only......9. Sisyrosea 4. Hind tibiæ with four spurs......10. Natada 1. Antennæ serrate or simple. 2. Hind tibiæ with terminal spurs only.....14. Heterogenea 2. Hind tibiæ with four spurs. 3. Palpi reaching beyond vertex. 4. Palpi extremely long, with third joint more than half as long as second. 3. Cnidocampa 4. Palpi reaching barely above vertex, third joint shorter..15. Lithacodes 3. Palpi reaching barely to vertex, or shorter. 4. Robust, with servate antennæ.....11. Prolimacodes 4. More slender, with simple or laminate antennæ. 5. Fore wing with rounded apex, but oblique outer margin. 13. Tortricidia 5. Fore wing with square apex and nearly straight costa. 12. Cochlidion 5. Fore wing broad and bluntly rounded.....16. Packardia

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	c · ·	7		
Arti	ncial_	keu	to	species

	There's group
1.	Thorax green
1.	Thorax brown or yellow
2.	Hind wing half vellow and half brownP. chloris (p. 108)
0	Hind wing nearly evenly colored, or with narrow brown border.
	This wing hearly evenly colored, of with harrow brown bound.
	<i>P. indetermina</i> (p. 107) Fore wing marked with green
- 3.	Fore wing marked with green
3.	Fore wings without green
- 91	/ Fore wings with all using presented over bluntly remaded
0,	2. Fore wings with an veries preserved, apex biantity rounded
	E. delphinii (p. 107)
31	4. Fore wings with 11 veins, apex subfalcate. A. spinuloides (rarely) (p. 108)
- <i>A</i> '	Small; hind wings triangular, transparent except on inner margin.
4.	Small; find wings trangular, transparent except on inner margin.
	P. pithecium (p. 105)
4.	Hind wings more rounded, heavily scaled.
5	A large, subtriangular, dark brown area, resting on costaP. scapha (p. 110)
5	No much sortal with
- i).	No such costal patch
6.	With ocellate spots at anal angle
6.	No such spots
7	Spots at anal angle black, pale ringedP. clegans (p. 112)
<u>_</u> .	Gods and had been had
(.	Spots pale, dark ringed
-8.	Hind wings paler than fore wingsP. geminata (p. 112)
8.	Hind wings blackishP. albipunctata (p. 113)
0	Marked with pure white
<i>a</i> .	marked with pure white the life of second second band 12
9.	No pure white, rarely a straight, dinuse, whitish, median band
10.	An irregular, transverse, median bandL. fasciola (p. 112)
10	Such a band on inner half of wing only11
10	A minute streak on middle of inner margin and on costa before apex12
10.	A minute streak on might of miler margin and on costa before a pex
10.	Two subapical dots and a dot below cell only
11.	Band extending into cell
11.	Band sapping abruptly at Cu
10	Cround light cohrours
14.	Ground nght demedias
	Ground red-brown
13.	Base of fore wing lemon yellow, contrasting with the buff ground
	C. flavescens (p. 106)
19	Page of function and contractional vallow
19.	Base of fore wing not contrastingly yellow14
14.	Fore wing straw yellow, contrasting with the dark brown hind wing; immaeu-
	late
14	Fore wing not contrastingly pale, or with complex markings15
15	The table set of the string of the templex markings
19.	Hind tibiæ with end-spurs only16
15.	Hind tibiæ with 4 spurs17
16.	Expanse under 15 mm.; hind wing blackish
16	Expanse over 15 mm.; hind wing chocolate brownS. textula (p. 109)
17	Destruction 1 in the state of the second of
14.	Postmedial line distinct, wavy
17.	Postmedial line even, or marks obsolescent19
18.	Palpi hardly exceeding frontP. pithecium (p. 105) Palpi extending above vertexI. beutenmulleri (p. 106)
18	Palpi avtanding above vertex
10.	A the second magnetic second s
19.	A transverse band formed of brown, longitudinal striations on the veins.
	<i>T. testacea</i> (p. 111)
19.	No longitudinal striations on veins 20
20	<i>T. testacea</i> (p. 111) No longitudinal striations on veins
20.	Subterminal line nearly straight, ending on inner margin; antemedial line
	similar, more obliqueN. nasoni (p. 109)
20	Subterminal line areuate, running across apex, or markings obsolete
51	Subterminal line whitted, and autors of a lock harm prior 1 at 1
.1.	Subterminal line whitish, and cutting off a dark brown apical patch.
	$C. \ biguttata$ (p. 110)
21.	Subterminal line dark, the wing not dark beyond it

22.	Antemedial line pale-shaded beyond; no median line parallel to it
22.	Ground color even, ochre, a median line from M_3 to inner margin, parallel to
	the antemidial line
23.	Hind wing blackish
23.	Hind wing straw yellow
24.	Clay color, markings obsolescentT. pallida (p. 111)
24.	Fore wing orange, hind wing pale yellow, markings obsolescent.
	T. flavula (p. 111)
24.	Yellow with well-marked, curved subterminal line25
25.	Hind wing nearly black, contrasting with the fore wingT. fiskeana $(p. 111)$
25.	Hind wing concolorous with fore wing, or slightly paler26
26.	Subterminal line filled below with brown, and whole median area brown in
	many specimens
26.	With the slightly diffuse lines only T , flexuosa (p. 111)

Tribe Phobetrini

(Tropic hairy type)

Larvæ hairy; subventral space somewhat reduced; with more or less deciduous, setose horns, the subdorsal one strong and laterals reduced; spiracle of first abdominal segment higher than the others, the tubercle above it absent; three tubercles on mesothorax. First stage singlehaired, with tubercles i and ii united. Pupa not seen. Moth with antennæ pectinate to tip; \mathbf{R}_{0} , stalked beyond the origin of \mathbf{R}_{1} .

1. PHOBETRON Hübner

(Economidea Westwood; Thyridopteryx, in part, Packard; Limacodes, in part)

Body very stout, especially in male, and woolly; male antennæ with about 8 simple joints at apex. Fore wing of male triangular, the outer edge nearly straight, and fully as long as inner; hind wing with apex acute, outer margin (fig. 67) wings normal, hind wing proportionately small. Hind tibiæ short, stout, and tufted like the Euclea group, with the upper spurs weak.

The larva has an irregular and variable set of curved, detachable lateral processes, which are covered with fine stinging hair. These processes are detached before pupation and arc woven into the cocoon. A few additional species occur in South America.

1. P. pithecium Smith and Abbot. Male translucent, smoky; veins and borders, 1. P. Interetim Sinth and Abbot. Mate transmeent, sindsy; tends and borders, especially inner margin of hind wing, smoky gray; with vague, darker, trans-verse bands. Female with wings opaque, the fore wing mixed light wood-brown, fuscous, and dull straw yellow, with a few blue scales, the border dark; post-medial one sinuous and scalloped; markings quite clean cut. Hind wing dark, \mathcal{J} 20 mm. Q 25 mm. (abbotana Hübner, T. nigricans Packard, hyalinus and tetradactylus Walsh). (H 47:6 \mathcal{J} , 7 Q.) The large is a general feeder on shrubs. (H 1:4)

The larva is a general feeder on shrubs. (H. 1:4).

Montreal, Quebec, and south. New York: Bolton (larvæ), Niagara Falls, Oswego, Lancaster, Bath, Hudson Valley, from Port Ewen south; Long Island.

2. ISOCH_ETES Dyar

(Semyra, in part)

Body relatively smaller; palpi much longer and smooth; hind tibiæ with end spurs only Venation as in female Phobetron. Caterpillar very fluffy, with all the subdorsal spines long.

1. I. beutenmulleri 11. Edwards. Similar to female pithecium; markings less clean-cut, postmedial line not distinctly wavy; outer margin pale yellow; blue scales more prominent. 20 mm. (H. 47:17.)

The larva is green.

Staten Island, New York, to Florida.

Tribe Eucleini

(Tropic spined type)

Larvæ spiny, with subventral space reduced; tubercles tending to be horn-like, not deciduous, poisonous in several species; 2 tubercles on thoracic segments; first abdominal segment as before; first stage with several hairs to a tubercle. Antennæ of male moth normally pectinate with simple tip; hind tibiæ usually with end spurs only, palpi moderate, \mathbf{R}_5 stalked, \mathbf{R}_2 usually free. Pupa with lateral processes of maxillæ reaching to base of antennæ.

3. CNIDOCAMPA Dyar

(Miresa)

Palpi stout, very long, with second and third joints equal; male antennæ simple, laminate. Larva similar to Sibine but with rudimentary subdorsal spines on the middle segments, besides the long ones, and with a small, blue rectangle in place of the saddle-marking; feeding on Japanese Ivy (Ampelopsis). 1. C. flavescens Walker. Fore wing with outer half light fawn, with two oblique, brown lines; base lemon yellow, much more broadly on costa. 30 mm. Booton Massedweatte, introduced from Asia Saldem in jurious.

Boston, Massachusetts, introduced from Asia. Seldom injurious.

4. SIBINE Herrich-Schæffer

(*Empretia* Clemens)

Similar to Euclea, except for the sinuous inner margin. \mathbf{R}_{s} sometimes free but closely connate with $\mathbf{R}_{3:4}$. Larva with strong anterior and posterior subdorsal spines, but in middle segments rudimentary ones only, or, in our species, none. There are several other species in tropical South America. 1. S. stimulea Clemens (Saddle-back caterpillar). Shining, deep seal brown, shaded with blackish; with three white points. Hind wing paler. 30 mm.

(H 47:9.)

Larva with characteristic green saddle, and with brown central patch; feeding on various trees. (H 1:6.)

Massachusetts to southern Illinois and southward. New York: Poughkeepsie, Staten Island; Brooklyn, and Woodhaven, Long Island.

5. EUCLEA Hübner

(*Limacodes*, in part)

Antennæ of male closely pectinate about two-fifths way out, then strongly serrate for a short distance, but with simple spical half; antennæ of female nearly simple. Palpi moderate, curved, and a little rough. Wings broad, all veins present. Fore wing (fig. 69) with outer and basal part of inner margin strongly convex. \mathbf{R}_5 strongly, and \mathbf{R}_2 often shortly, stalked.

Larva stout and prismatic, not much smaller in front, subdorsal and lateral spines irregularly but about equally developed; small pointed clusters of detachable spines between the terminal horns, the so-called caltrop spines.

The genus is not really distinct from Parasa, and P. indetermina is intermediate. 1. E. delphinii Boisduval. Brown, tending to shade into purplish brown and tawny, more or less marked with green, the boundary often edged with white. 25-30 mm. (cippus, quercicola, tardigrada, etc.) The larva feeds on various trees and shrubs. The moth flies in June. Montreal, Quebec to Illinois and southward. New York: Plattsburg, Peru.

The typical form, with varieties interjecta Dyar and viridiclava Walker, is rather common southward. Variety elliotii Pearson is recorded from Big Indian Valley, Coney Island, and Glendale, Long Island, and variety panulata Clemens is reported from New York State by Packard.

The following varieties are recognized, one or two of which may prove, on breeding, to be distinct species.

Key to the varieties

1. Dominantly brown, the cell brown. One large irregular green patch.....riridiclava Walker (H 47:23). Two green spots connected by a row of dots.....interjecta Dyar. Two green spots only. Lower spot, below base of cell, small and triangular; upper spot some delabilities (H 47.24) times broken into separate dots.....delphinii (H 47:24). 3. Lower spot large and forked, or running out below cell.

querceti Herrich-Schæffer.

1. Dominantly green, cell green. 2. A large, brown, discal spot.....elliottii Pearsall. 2. No discal spot.....panulata Clemens (H 47:5).

6. PARASA Moore

(Euclea, in part; Callochlora Packard)

Similar to Euclea, \mathbf{R}_2 less frequently stalked; thorax green in our species.

Larva without caltrop spines. 1. P. indetermina Boisduval. Male antennæ merely serrate at middle. Vertex and thorax green. underside brown; fore wing green, with a brown patch at base, and a brown, slightly irregular border, about one-fifth as broad as length of wing, normally with a darker shade at middle of outer margin. 25 mm. (viridis Reakirt, vernata Packara) (H 47:10 3, 15 9 as chloris.)

This species and *chloris* have been much confused and even interchanged, making published records uncertain. The caterpillar is not humped in front, and has strong spines on the second, fifth, and last segments of abdomen. The caterpillar

is red or yellow, purple, and white. It feeds on apple and many other trees and shrubs.

New York to Illinois and southward. New York: Staten Island, Woodhaven, etc., Long Island.

2. P. chloris Herrich-Schaffer. Antennæ of male broadly pectinate rather beyond middle. Similar to *P. indetermina*, smaller; the outer border generally broader and more even in outline, with the veins across it generally darker; no darker 20-25 mm. (fraterna Paekard, Newra Herrich-Scheffer.) patch at middle. (H 47:26 ♂ 29 Q.)

The larva is strongly humped anteriorly, with much-reduced spines and a

pointed tail. It is dead-leaf brown with a red, posterior patch. Southern New York; and New Jersey to Western Pennsylvania. New York: New Windsor, Staten Island, Forest Park, Brooklyn,

7. MONOLEUCA Grote and Robinson

(*Limacodes*, in part)

Antennæ of male pectinate beyond middle, palpi like Euclea; fore wings evenly rounded, with \mathbf{R}_{1} and \mathbf{R}_{4} completely fused; \mathbf{R}_{2} stalked. Built slighter than Euclea. Larva with spines short and subequal, the lateral ones on first segment of abdomen missing, replaced by the spiracle, a pair of spines at each end of body somewhat longer than the others.

1. M. semifascia Walker. Fore wing red-brown with a narrow, somewhat wavy, white fascia from about the middle of cell to inner margin. 20 mm.

The moth flies in July. The larva is vermilion red with two pairs of vermilion stripes edged with vellow, running along the rows of spines. There are also dorsal and lateral bands, each composed of three blue lines, and a subventral band of two lines.

M. subdentosa is probably found only south of our area.

8. ADONETA Clemens

Similar to Euclea. Fore wing (fig. 68) with outer margin slightly con-cave below apex, \mathbf{R}_{a} and \mathbf{R}_{i} united, stalked with \mathbf{R}_{5} ; \mathbf{R}_{2} free. Larva with short spines, without detachable caltrop spines between the terminal ones, spines on abdominal segments 2, 5, and 8 longer than those between, but all very small.

1. A. spinuloides Herrich-Schæffer. Fore wing brown, of various shades, tending to be darker, and generally also dark-shaded on the veins toward the margin. An irregular and broken dark postmedial line edged with white toward costa and inner margin, or (in a single specimen seen) with small green patches like E. delphinii, 20 mm. (H. 47:3).

The larva eats varions shrubs. It is green, with a broad, irregular, purple dorsal band, and yellow subdorsals, the terminal spines short. The moth flies in July and August.

Montreal to Illinois, south to North Carolina. New York: Albany, New Windsor, Staten Island, Pinelawn, Yaphank, Long Island.

A. leucosigma Packard is probably a variety of this species in which the dark marginal shading is absent. It is reported from New York to Texas. I have

seen no specimens from a definite locality in New York. 2. A. bicaudata Dyar. Ground color light yellow, the border diffusely and rather narrowly cinnamon brown. Markings as in A. spinuloides, but often without the dark discal bar. Larva with a pair of long spines at posterior end.

This form is known from the District of Columbia to North Carolina.

9. SISYROSEA Grote

(Isa Packard)

Male antennæ broadly pectinate to apex; antennæ of female simple. Palpi in male obliquely truncate, reacning middle of front, in female smaller. Hind tibiæ with end spurs only. Moth quite stout. Fore wing with outer margin decidedly oblique, bent at middle, and slightly concave above, \mathbf{R}_2 given off from radial stem at a sharp angle, beyond the origin of \mathbf{R}_3 . Surface of wing when perfect, showing wavy scaling in transverse striæ, much as in the Megalopygidæ.

The larva is strongly flattened. Its subdorsal spines are irregularly reduced, and the lateral spines are very long, even, and feathery, and are held appressed to the surface on which the larva rests.

Sisyrosea is a primitive genus, in some ways transitional to the Phobetrini.

I. S. textula Herrich-Schæffer. Fore wing shaded pale gray, brownish, and light flesh color. 20 mm. (*Limacodes inornata* Grote and Robinson.) (H. 47:14.) Larva green; feeding on wild cherry and many other shrubs.

Massachusetts to Pennsylvania. New York: General from Plattsburg southward. Larva more often seen than moth.

10. NATADA Walker

(Sisyrosea, in part)

Similar to the last genus; \mathbf{R}_2 free, wings rounded, with marked apex; scaling normal; tibiæ with four spurs. Larva high, prismatic, with short, subequal horns, and no caltrop spines.

1. N. nasoni Grote. Clay color, somewhat dusted with fuscous; with two oblique dark lines. 25 mm.

Larva green with yellow lines, and red tubercles; on oak and other trees.

The moth, though commoner in the southern States, seems rare. New York: Ronkonkoma and Yaphank, Long Island.

Tribe Cochlidiini

(Smooth Eucleids)

Larva without tubercles or spines after first stage, spiraele of first abdominal segment normal, and its lateral tubercle present. Pupa with maxilla not extending out to antennæ. Moth with \mathbf{R}_5 free, usually arising about half way between \mathbf{R}_{3+4} and \mathbf{M}_1 ; usually with all spurs on hind tibia.

11. PROLIMACODES Schaus

(Limacodes; Eulimacodes Dyar, not Möschler)

Stouter than the following genera; male antennæ serrate; **R** and \mathbf{M}_i of hind wing normally stalked (fig. 70). Larva with broad dorsal and subventral areas meeting in a sharp ridge, but without lateral area; in first stage with 2-haired warts.

An aberrant genus representing a typically South American group, the "tropic smooth type" of Dyar.

1. P. scapha Harris. Fore wing with a large triangular, deep brown area extending from a quarter way out on costa to apex, and down to A, with a narrow extension along costa to base; rest of wing pale brown, becoming silvery toward the dark brown patch. 25 mm. (H 47:8.)

The moth appears in July and August. Florida and Arizona specimens represent very distinct races (argentimacula Barnes and McDunnough and trigona H. Edwards). The larva is green, brown, and yellow, and lives on various trees and shrubs.

New York to Florida and Arizona. New York: Geneva, Ithaca, Florida, Rhinebeck, Staten Island; Brooklyn, East New York, Newtown, etc., Long Island. Judging from the larvæ seen, the species will prove much wider-spread.

12. COCHLIDION Hübner

(Limacodes Latreille)

In this and the following genera, the male antennæ are simple and laminate. **R** and \mathbf{M}_1 of the hind wing are normally free. In the larva, the dorsal and lateral areas are equally broad, and the subventral area reduced. The first stage has simple setæ. They are the "palearctic smooth type" of Dyar.

simple sets. They are the "palsarctic smooth type" of Dyar. Moth of Cochlidion with palpi upturned to about the middle of the front, longest in *C. biguttata*. Vestiture composed of hair and narrow spatulate hair. Hind tibiæ with all spurs, less heavily fringed than in Prolimacodes, but more than in some of the later genera. Fore wing with apex rectangular; outer part of costa and upper part of outer margin straight; all veins present; \mathbf{R}_2 and \mathbf{R}_5 both free, but approximate. Larva with small, but sharp, depressed spaces; with posterior end not extended. Skin spinulated. In the first stage setæ i and ii are very unequal, on the same wart.

1. C. biguttata Packard. Very pale gray, base and anal and apical spots wood brown, pale-edged, postmedial line nearly straight. 25-30 mm. (*Limacodes* Packard.)

June and July.

The larva has been found on oak only. It is pale whitish green, with a white sub-dorsal line, edged with dark, and no transverse yellow line on anterior margin of thorax.

Montreal, Quebec to New York and Illinois. New York: Plattsburg, Ithaca, Ronkonkoma and Yaphank, Long Island.

2. C. rectilinea Grote and Robinson. Yellowish or reddish brown, the median area rather grayer, with base darker; antemedial line white, preceded by a dark line; lines across apex and at anal angle straighter, and not cutting off well-marked dark patches. Hind wing rather darker gray-brown. Over 20 mm.

The moth occurs in the South, perhaps not ranging as far north as Virginia, northern records being based on *C. latomia*.

New York: reported in the vicinity of New York (Beutenmuller, Grote, and Robinson).

3. C. latomia Harvey. Similar to C. rectilinea, but with paler fore wing and pale, straw-yellow hind wing, no distinct spot at anal angle, and only a slight dark shade before the antemedial line. (H 47:27.)

New York to Texas; commoner southward.

4. C. y-inversa Packard. Dull light ocherous, with pale or dark shades; lines dark, even, sharply defined; antemedial from middle of costa to inner margin at a third way out; a second line, parallel to it, below middle of wing; a line across apex, and dark fringe; hind wing yellow. 25 mm.

Larva on hickory and blue beech; similar to C. biguttata, but with a yellow line across mesothorax.

Florida specimens belong to the variety parallela. Moth in July.

Gaspé, Quebee, to Illinois and Florida. New York: Batavia, Lewiston, Buffalo, Ithaca, Big Indian Valley, Poughkeepsie, New Windsor, New York City, Staten Island, Long Island.

13. TORTRICIDIA Packard

(*Heterogenea*, in part, etc.)

Palpi obliquely upturned to well beyond middle of front; fore wing with outer margin strongly oblique, apex rounded, costa more arched, and inner margin less sinuous than in Cochlidion. Larva with depressed spaces large and sharp; skin granulated but not spinulated; marked with red dorsally. In first stage, with setæ i and ii equal.

Kronæa minuta probably belongs to this genus or to Heterogenea.

1. T. fiskeana Dyar. Fore wing tawny yellow, with a straight, oblique median line, sharply defined on its inner side, and a strongly curved one across the apex, the space between them sometimes more or less filled with brown. Hind wing nearly black. 14-15 mm.

This form is perhaps a variety of minuta. The lines are less diffuse than in Heterogenea shurtleffi.

Durham, New Hampshire; Iowa.

2. T. flexuosa Grote. Similar to T. fiskeana, both wings pale yellow, the lines not suffused. 15-25 mm.

In variety cæsonia Grote (H 47:12) the medial area is suffused with brown. Larva green with yellow subdorsal lines; the red dorsal markings, when extended laterally in cross form, not reaching more than halfway to the lateral margins; sometimes reduced to one or two spots.

Quebec to New Jersey and western Pennsylvania. New York: Ithaca, De Bruce, Big Indian Valley, Onteora Mountain, Ilion, Sharon Springs, Rhinebeck, Poughkeepsie, Windsor, Bronx Park, New York City, Long Island. The dark form cæsonia Grote, is known from Big Indian Valley, Coeymans, and

Poughkeepsie.

3. T. pallida Herrich-Schæffer. Fore wing light pinkish brown; hind wing straw yellow, typically immaculate, in var. flavula Herrich-Schæffer showing traces of lines. 10-18 mm.

The larva occurs on willow oak, and other shrubs. It is green, with the usual two subdorsal yellow lines, with a large, red, cross-shaped mark extending from end to end of the body, and from side to side at the middle, the anterior arm of the cross narrow, but the posterior widening into a patch. The moth flies in July.

Southern Maine to southern New York; probably more widespread but overlooked. New York: Plattsburg, Keene Valley, Ithaca, Staten Island; Brooklyn, and Bellport, Long Island.

4. T. testacea Packard. Dull, light brown with a broad, dark, median shade, darkest along the veins. Hind wings paler. 15 mm. (H 47:11.)

Larva with the dorsal red patch as long and wide as the body, but symmetrical

from front to back; feeding on oak, birch, wild cherry, and so forth. Nova Scotia to northern New Jersey and Illinois. New York: Plattsburg, Saranac Lake, Red Lake, Ithaca, McLean, Big Indian Valley, Staten Island; Marriek Long Leand Merrick, Long Island.

5. T? (Kronæa) minuta. Fore wings straw yellow, hind wings blackish, immaculate 10 mm.

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Larva with yellow line across the front of the thorax and a larger red patch than in H, shurtleffi.

Atlantic States. The type is lost, and no other specimens are known to exist.

14. HETEROGENEA Knoch

Similar to Tortricidia, but with upper spurs of hind tibia lost, and palpi weaker, hardly reaching the middle of the front.

1. H. shurtleffi Packard. Base of wings rusty, outer part brown, obscuring the markings, which are brown, and much as in C. y-inversa. 12 mm. (H 47:20.)

This species is apparently rare. The larva is similar to the Tortricidias, but has a transverse yellow line across the mesothorax, and only a small red dot in the middle of the dorsum.

Massaehusetts to New Jersey. New York: Vicinity of New York City (Eliot and Angus).

15. LITHACODES Packard

Similar to Packardia; palpi upturned beyond vertex. or, when obliquely held. projecting strongly, and reaching level of antenna. Larva similar to that of Cochlidion, but with a mixture of granules and spinules on the skin.

1. L. fasciola Herrich-Schæffer. Ochre vellow with a white, irregular, transverse, median band, usually followed by a deep brown shade; rarely all brown except the terminal area, and with the white bands strongly contrasting. 20 mm. (H 47:2).

Larva with a yellow subdorsal line, but no dark one, and no red markings. Moth in July.

Common generally; Quebec to Texas. New York: Buffalo, Geneva, Ithaca, Scheneetady, and the southern Catskills.

16. PACKARDIA Grote and Robinson

(Cyrtosia Packard, not Perris)

Slender moths with sealy vestiture. Antennæ simple; palpi upturned beyond middle of front, acute; tibiæ with all spurs. Fore wing (fig. 71) with arehed costa, bluntly rounded apex and outer margin, \mathbf{R}_5 arising from end of cell nearly midway between \mathbf{R}_{3+4} and \mathbf{M}_1 , \mathbf{R}_2 arising from cell.

Larva with depressed spaces small; end of body produced in a pointed tail; skin granular and not spinose nor crested at any stage; no dorsal markings. First stage with tubercles i and ii represented by simple, short spines. 1. P. elegans Packard. Typically dark bronze-gray (in var. fusca Packard

1. P. elegans Packard. Typically dark bronze-grav (in var. fusca Packard light buff) with a white band across end of wing to just above anal angle, and two lesser pale bands to middle of inner margin; ground more or less dark-shaded. (The two black dots at the anal angle replaced by a vague dark shade in var. fusca.) 20 mm. (H 47:16.)

Moth in June and July. Larva translucent yellowish green, with a broken, yellow subdorsal line and scattered patches of whitish green pigment. It feeds on the thin, shaded leaves of trees in dense woods.

Southern Quebec to northern New York. New York: Goat Island, Ithaca, Newburgh, New Windsor, Katonah, New York City, Long Island. 2. P. geminata Packard. Light straw or cream color, shaded with brown, except

2. P. geminata Packard. Light straw or cream color, shaded with brown, except at base and beyond the outer line, which runs as in *P. clegans*. Antemedial line oblique and far out. Hind wings nearly concolorous. 25 mm. (H 47:1.)

The moth occurs in June. The caterpillar is densely pigmented, whitish green with a nearly white, subdorsal line. It is found on very small shrubs only a few inches high, especially on wild cherry. The female moth, unlike most Eucleids, will often wait a few days for fertilization before flying.

Southern Quebec to New York and Wisconsin. New York: Forest Park. Brooklyn.

3. P. albipunctata Packard. Similar to P. geminata but with blackish hind wings. Fore wings typically much shaded with brown and ochre, but in var. ocellata Grote as in geminata. 25 mm.

Larva unknown.

The moth occurs in New York (Karner, Dutchess County) but its general distribution is unknown; it is probably confused with P. geminata, of which it may be a Mendelian form.

Family 8. PYROMORPHIDÆ

(Zygænidæ, Anthroceridæ in part; Sphingidæ, in part, of early workers)

Moderately small moths, with smoky, translucent wings, in a few species with metallic coloring. Male antennæ generally thickened and shortly pectinate; head prominent, with small palpi and small or rudimentary maxillary palpi, but a strong, naked tongue. Ocelli present, well separated from eyes, sometimes very small. Fore wing (figs. 62 to 65) rounded with two fully developed anals, traces of base of media preserved, but not forked; no accessory cell; all veins except \mathbf{R}_2 to \mathbf{R}_4 inclusive arising separately from the cell. Hind wings with Sc and R fused beyond middle of cell, the basal part of **R** lost, or represented by a short spur, so that Sc seems to spring from the outer part of the cell; Sc much swollen or forked at base. (In the true Zygænidæ or Anthroceridæ, Sc and R are not fused, but are connected by a cross vein.) Hind wing with 1st A preserved, but 3d A lost in a couple of genera.

Egg of flat type. Labrum of larva with seta ii higher than i but not reduced; body with somewhat diffuse tufts representing warts i+ii, iii, iv+v and vi, with two tufts representing vii, but with viii simple; hairs serrate, but not feathery as in some Arctiids.' Prolegs with hooks in a straight, uniordinal band, without any additional prolegs or suckers. Pupa incomplete but without motion between appendages; third to seventh abdominal segments free, and eighth in male also free from seventh. Tongue of macro type, reaching well toward tips of wings, and covering labial palpi, tongue laterally extended at base, but without a tooth to represent the maxillary palpi. Abdomen diffusely spinulated above.

A small and wholly American family, of about 50 species.

Key to the genera

1.	Fore	wing	with	nine	veins	from	cell,	all	separate			1. Aco	loithus.
1.	Fore	wing	with	R _a an	d R, 1	stalked	l toge	ether	, sometim	es wit	h ·10	veins	arising
	from	n cell			-		0						

2. hind wing with about same area as fore wing, with 3 anals 2. Pyromorpha.

2. Hind wing very small, with 2 anals only

1. ACOLOITHUS Clemens

(Harrisina, in part)

Wings bluntly rounded and somewhat translucent, hardly reaching end of abdomen. Fore wing (fig. 63) with 4 radials, all arising separately from cell, hind wing with base of \mathbf{R} represented by a spur in cell; \mathbf{M}_1 lost; 3 anals; anal angle slightly lobed. Caterpillar with inconspicuous hair, with a broad dark lateral line; found on grape and ampelopsis.

1. A. falsarius Clemens. Black with orange collar. 16-18 mm. (sanborni Packard) (H 16:14).

The moth may be found on flowers in May and June. New York to Louisiana, Florida, and Texas. New York: Syrac Rhinebeck, Crugers; Centerport and Lake Roukonkoma, Long Island. Svracuse, Ithaca.

2. PYROMORPHA Herrich-Schæffer

(Malthaca Clemens)

Wings (fig. 62) bluntly rounded, hind wing larger than in Acoloithus, usually extending well beyond end of abdomen; female with wings very slightly reduced. Fore wing with all veins preserved, \mathbf{R}_3 and \mathbf{R}_4 stalked; hind wing with all veins; base of \mathbf{R} lost. Wings very thin, with hair-scales. Larva with inconspicuous hair, dark, checkered with pale patches; on dead oak leaves. **1. P. dimidiata** Herrich-Schæffer. Smoky; basal half of fore wings except on inner margin, and costal edge of hind wings, pale tawny, the tawny less extensive in female. 25 mm. (*Malthaca perlucidula* Clemens). (**H** 47:33 \mathcal{J} .) The moth flies from the end of May to July

The moth flies from the end of May to July.

New York to Florida; also reported from the Western States. New York: Coy Glen (Ithaca), Staten Island, Brooklyn.

3. HARRISINA Packard

(Aglaope; Procris)

Fore wings four times as long as wide, outer and inner margins equal; \mathbf{R}_2 stalked, but \mathbf{R}_s arising from cell, and widely separated from \mathbf{R}_{2+3+4} . Hind wing of less than half the area of the fore wing, with M_1 and 3d A lost. Wings more heavily scaled than in other genera.

Caterpillar with black tufts contrasting with the pale ground; social on grape. 1. H. americana Guérin. Black, slightly greenish iridescent, with orange collar.

Fore wing with vein R_s lost (fig. 65). 18-25 mm. (H. p. 372 f. 210.) The larva is nearly white, with the black markings confined to the tufts. The moth flies from May to August. Perhaps the species has a wider distribution, but

it has always been confused with H. texana.

Massachusetts to Virginia. New York: Troy, Poughkeepsie, Staten Island.

2. H. texana Stretch. Similar to H. americana, but with all veins preserved, R2, R3 and R, stalked (fig. 64). 20-28 mm. (H. 43:34 as americana.)

Larva with black transverse bands, sometimes broken, and a reddish lateral band.

New York to Texas. New York: Poughkeepsie; Woodhaven and Yaphank, Long Island.

The southwestern variety australis Stretch, which has been taken in Florida and Georgia, has orange on the base of the tegulæ, as well as on the collar.

SUPERFAMILY TINEOIDEA (restricted)

The Tineoidea, as represented by the typical Tineidæ, are hardly distinguished from the Adeloidea, save by the general loss of aculeæ on the wing surface. In a few genera, as in Solenobia and Opostega, a few aculeæ are still to be found about the base of the cell and along the veins. I think I have also seen a few in Tischeria and Setomorpha, but they are not present on the general wing surface. Another rather general character for the genera of this series, which are otherwise closest to the Incurvarioidea, is that the antenna is regularly provided with two whorls of scales to a segment, one whorl being of sharply outstanding scales, thus making the antenna rough and annulate (figs. 87 to 89). This character holds in Tinea and the genera reduced from it, but fails in most of the aberrant forms, in which the scaling is smooth and sometimes in more than two rows (Acrolophus, Psychidæ). The more primitive forms of Tineoidea are often case-bearers, but the lenticular case typical of the Adeloids is rare.

The moths of the Tineoidea have the vestiture of the vertex variable, though always rough in the lower forms; and there is no naked area above the eye. The eyes are often small. The ocelli are variable. and, when present, are as a rule near the edge of the eye. The tongue, when present, is scaled at the base. The labial palpi typically are long and folded, but often reduced, or lost. In the Gracilariidæ, which may not really belong of this superfamily, they are porrect. The primitive forms have a complete primitive venation (fig. 19), the base of 1st A fading out; in higher forms, it is much reduced. In the hind wing, **R**, as a rule, is closely parallel to **Sc** at the base, and they are connected by a cross vein, (\mathbf{R}_1) a short distance from the base. The cross yein and the base of R often faded out together, but the cross vein is never preserved when **Rs** is weak, as in the Incurvariidæ. The Lyonetiidæ are an exception. The venation is sometimes extremely reduced, but 1st A is preserved as long as 2d A. The female has no piercing ovipositor.

The eggs are of the flat type. The pupa is incomplete. In the larva, setæ iv and v are usually widely separated (figs. 91, 92) as in the Noctuidæ, but there are three setæ on the prespiracular wart.

The Tineoidea and the next four superfamilies have been somewhat tentatively delimited and are not sharply defined, but the groupings here made will roughly indicate the relationships of the families. This superfamily, with the next three and the Incurvatioidea, forms the Tineid series as usually understood.

The Psychidæ have been widely separated by most workers, but there is little besides the egg-laying habit to distinguish them from the true Tineidæ; some have winged females (as in the case of Kearfottia, in

WILLIAM T. M. FORBES

the United States), but in some the female is more reduced than in any other known form of Lepidoptera, and never leaves the cocoon.

Family 9. **TINEIDÆ** (somewhat restricted)

Head usually with high, rough, bristling vestiture; the vestiture finer and more macro-like in Acrolophus, and short in a few genera near Amydria which have a strongly bristled palpus. Front often rough and palpus often bristled on side, though but weakly in reduced forms and not at all in Aerolophus. Maxillary palpus typically five-jointed, folded, sometimes reduced or absent; tongue sealed, or rarely absent; antenna never much longer than wing, but usually more than half as long; often with a peeten, but without an eye-eap; typically with two whorls of seales to a joint, one of them of divergent seales (in Aerolophus the antenna is smooth-scaled, with several rows to a segment); antenna in our species simple, but peetinate in a few exotie Aerolophinæ. Fore wing with all veins, or with one vein absent; accessory cell usually present, separated from cell by a long, weak vein; base of media present in lower forms; 2d A frequently forked at base; wing always rather narrow and sometimes laneeolate; hind wing varying from ample (Aerolophus) to linear (fig. 81); when broad, with \mathbf{R} and \mathbf{M}_{i} , well separated, but in some genera \mathbf{M}_{i} and \mathbf{M}_{s} stalked; when narrow, often with an open cell; with most veins present, but frequently extremely weak. No aculeæ. Hind tibia usually with a mixture of bristles and hair, never smooth-scaled.

Egg of flat type, oval. Larva with set \mathbf{iv} and \mathbf{v} well separated, otherwise differing in the two subfamilies. Pupa incomplete, with well-marked maxillary palpus; with Adelid prothorax; abdomen with segments 4 to 7 inclusive, free, and 8 in male; but segment 2 fixed; each segment with a single anterior row of spines; tongue shorter than labial palpus.

The Tineidæ appear to furnish the point of origin of each of the higher tineoid superfamilies, and possibly of the macros as well, but the macros seem a little closer to the Yponomeutoids. The two subfamilies are not closely related, and could well be treated as families, as they often are.

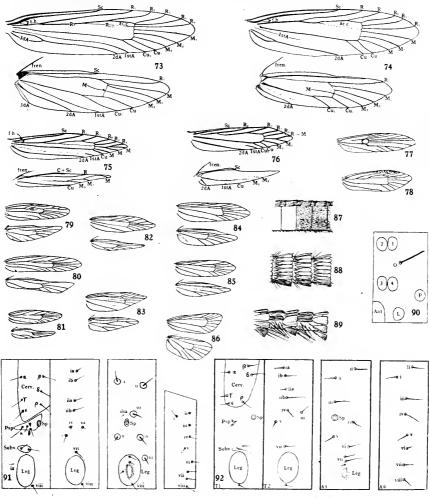
Key to the genera of Tineida and Adelida

1. Vestiture deep and spatulate on thorax, with well-marked anterior and posterior tufts; palpus also with deep vestiture, not bristled (Acrolophina)

1. Acrolophus.

Vestiture mainly scaly, except on head; palpus small, usually bristled.
 Only three short veins from cell to dorsal margin (fig. 106).

Tischeria (Tischeriidæ) (p. 145).



FIGS. 73-92. TINEID.E

73, Tinea pellionella, venation; 74, T. granella, venation; 75, Œnoë hybromella, venation; 76, Homostinea curvilineella, venation; 77, Monopis rusticella, venation of fore wing; 78, Triehophaga tapetzella, venation; 77, Monopis rusticella, venation; of fore wing; 79, Setomorpha insectella Q, venation; 80, Homosetia costisignella, venation; 81, Leucomele miriamella, venation; 82, Diachorisia relatella, venation; 83, Hybroma chrysocomella, venation; 84, Isocorypha mediostriatella, venation; 85, Hybroma servulella, venation; 86, Choropleca vesaliella, venation; 87, Acrolophus arcanellus, male antenna, side view; 88, Tinea, first group (Amydria and the rest are similar); 89, Cyanc visaliella, male antenna; 90, Tinea granella, ocellar group of larva (typical of the subfamily); 91, Acrolophus arcanellus, seta map (after Fracker); 92, Seardia fiskeella, seta map (after Fracker)

4. Second joint of palpus with a bushy mass of scales below, more or less rounded in side view and very large; third joint long.

5. Antenna without a pecten; tuft nearly spherical; maxillary palpus porrect; vertex nearly smooth......19. Amydria,

- 5. Antenna with a pecten.
 - 6. Tuft less developed; maxillary palpus small but distinctly folded. 7. Tuft compact and triangular; wings with all veins arising
 - 7. Tuft of loose spatulate hair.
 - 8. Hind wing with all veins free (figs. 73, 74)..4. A few Tinea
 - 6. Tuft and maxillary palpus like Amydria; vertex with high, bristling vestiture. Fore wing with small, raised scale tufts.

18. Xylesthia.

- 4. Second joint merely rough-scaled; third joint often short; palpus often drooping.
 - 5. Maxillary palpus, and usually tongue, obsolete.

6. Fore wing with ten veins arising from cell.

7. Antenna longer than fore wing.

(Adela and Nemotois-Adelidæ) (p. 77) 7. Antenna shorter than fore wing......7. Tineola. 6. Fore wing with a radial lost; wings lanceolate.....8. Tenaga.¹² 5. Maxillary palpus developed, folded.

6. Fore wing with complete venation (ten veins arising from cell).

7. M_3 and Cu_1 of fore wing stalked, sometimes Cu_2 also (fig. 77).

6. Monopis.

- 7. M_1 and M_2 arising out of R_514. Homostinea.
- 7. With only \mathbf{R}_4 and \mathbf{R}_5 stalked, or all veins free. 8. Tips of R_1 to R_3 fusing to form a stigma, with a thickened
 - 8. No stigma.
 - 9. Cell of hind wing open above Cu; venation incomplete (fig. 85).

10. Antenna nearly as long as fore wing....17. Triptodema. 10. Antenna two-thirds as long as fore wing.

- 11. Hind wing broad-lanceolate, with convex costa and separate medial stem.....12. Hybroma.
- 11. Hind wing narrow, with sinuous costa, and M associated with R-stem (fig. 81).....13. Leucomele.
- 9. Hind wing normal, with all veins preserved (six arising from cell).

- 10. A of fore wing simple at base (fig. 82). 11. Fore wing smooth-scaled; \mathbf{Cu}_2 of hind wing more than half as long as width of wing (fig. 84) .. 9. Isocorypha.
 - 11. Fore wing more or less distinctly tufted; Cu of hind wing very short (fig. 82).....10. Diachorisia.
- 10. A of fore wing forked at base.
 - 11. Each segment of antenna with a whorl of raised scales (fig. 88); fore wing with \mathbf{R}_2 and \mathbf{Cu}_2 well back from end of cell......4. Tinea.

¹² Sclenobia, with broad, bluntly rounded wings, likely to run out here (Psychidæ p. 143).

11. Antenna smoothly scaled or hairy; fore wing with R_2 and Cu_2 nearer end of cell (Prodoxus, Tegeticula, Incurvaria; — Adelidæ).

6. Fore wing with one vein lost.

7. \mathbf{M}_1 and \mathbf{M}_2 stalked with \mathbf{R}_6 (fig. 75).....15. Ence.

7. M_2 stalked with R_5 ; M_1 lost (fig. 76).....14. Homostinea.

7. M_1 and M_2 stalked with R_5 (fig. 75)......15. *Encë*.

M₂ stalked with R₅; M₁ lost (fig. 76).....14. Homostinea.
 8. Hind wing with 7 veins (counting the anals as one, as usual);

cell ope nabove Cu (fig. 45)..... (Eudarcia, Adelidæ) (p. 77) 8. Hind wing with normal venation.

9. A of fore wing forked at base

(Paraclemensia, Adelidæ) (p. 76) 9. A of fore wing simple at base.

10. Hind wing linear, with sinuate costa.

12. Hybroma in part. 10. Hind wing linear, with sinuate costa. 10. Diachorisia in part.

Subfamily ACROLOPHINÆ

(Anaphorinæ; Psychidæ, in part, of Kirby and Tutt)

Head retracted; eyes medium-sized, usually hairy; ocelli absent; palpus large, with the first joint large, upturned to the middle of the front, and, in the western genus Eulepiste, as long as the other two together; the palpus as a whole often upturned to vertex, or even far beyond, especially in the male; in the female, typically porrect and shorter. Maxilla obsolete. Antenna with somewhat globular scape; the shaft smoothly scaled above, sometimes in more than two rows, and finely pubescent with sense hairs below; in male, laminate, rarely pectinate. Vestiture of head, including palpi, thorax and femora, rough, but fine and very dense, largely of spatulate hair; abdomen also rather hairy; \mathbf{R}_s of fore wing running to outer margin. Hind tibia hairy rather than bristled.

Larva (fig. 91) with front reaching only halfway to vertex, the adfrontals very wide and reaching vertex; ocelli six, but not regularly arranged, the fourth and lower being much closer together than the second and third are; head ventrally chitinized behind labium. Leg with trochanter one-third as wide as femur; prothoracic legs separated by a distinct, chitinized sternum; thorax with setæ on large shields; tubercles iii to v apparently taking the place of pleural sclerites; cervical shield extending the whole width of the prothorax and enclosing the spiracle; prolegs with one complete ellipse of hooks, preceded by several (3-6)rows of rudimentary ones; the anal proleg with a curved band.

The larva of one species is known to live in a tube near the surface of the ground, feeding on grass and the like, and pupating in the ground.

Pupa heavily chitinized, suited for coming up through the earth; antenna shorter than fore wing; fore wing bluntly rounded.

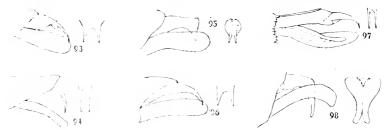
This group is entirely American, and predominantly tropical. Besides Acrolophus there are a couple of other well-defined genera.

1. ACROLOPHUS Poey

Including Hypoclopus Walsingham, Anaphora Clemens, Pseudanaphora Walsingham, Eutheca Grote, and other genera)

Characters of the subfamily.

The genus has been divided into various genera, but Walsingham and Durrant conclude that there are no satisfactory distinguishing characters and therefore reunite them.



FIGS, 93-98. MALE GENITALIA OF SPECIES OF ACROLOPHUS

In each case the lateral view is shown, as seen after removing the scales. The dorsal view of the uncus is also shown. **93**, *A. arcanellus*; **94**, *A. mora*; **95**, *A. popeanellus*; **96**, *A. texanellus*; **97**, *A. plumifrontellus*; **98**, *A. mortipennellus*

Key to the species (males)

- 2. Valve apparently simple in side view.
 - - Dorsal edge of uncus sharply bent at a right angle near middle (fig. 95).
 Valve two-thirds as wide as length of vertical part of uncus (fig. 95).

3. popeanellus.

- 5. Valve fully as wide as length of vertical part of uneus...4. confusellus. 4. Dorsal edge of uncus evenly curved.
 - 5. Spines of uncus fully one-third as long as valve, slender, separated by one-third their length; eyes naked (fig. 94).....l. mora.
 - 5. Spines of uneus less than one-third as long as valve or widely separated. 6. Body stout; palpus upturned, reaching just above the vertex
 - (fig. 93)2. arcanellus.
 - - 7. Valve narrow and slightly spatulate (fig. 96).....6. texanellus.
 - 7. Valve slender and regularly tapering toward apex....7. hulstellus.

I. Body rather slender; palpus of male extending back over middle of thorax or beyond; in female short and porrect; antenna of male as in Pseudanaphora or narrower. and subservate; fore wing more pointed, that of male being about like that of female arcanella. Eyes naked. (Sapinella Kirby, Eutheca Grote, not Keisenw).

1. A. mora Grote. Male blackish; female usually dull ochre yellow, or, rarely. dull brown and obscurely mottled; a dark bar at middle of fold, and a slight dark spot at end of cell. Male (fig. 94) with branches of uncus long, slender. and parallel, not tapering to the bluntly rounded tip; valves also slender and of even width. 25-30 mm. (arcanella of authors).

The moth flies in October and November. The larva has been bred from birch by Pearsall. The egg is long, narrow, and strongly ridged.

Concord, Massachusetts to North Carolina, and western Pennsylvania.

II. Body stout, and, with vestiture, more than one-third as wide as length of fore wing; palpus upturned to just beyond vertex, but not extending back over thorax in male; palpus of female short and porrect; antenna smooth, and, in male, with laminations one-half wider than length of segments; eyes hairy. (Pseudanaphora Walsingham).

2. A. arcanellus Clemens. Pattern and colors same as those of A. popeanellus; rough, black, raised striations on fore wing; spatulate vestiture on thorax much coarser than that of A. popeanellus; wing of female rather longer and more pointed than that of A. popeanellus. (Fig. 93.) 25-28 mm.

The larva and pupa are described under the genus. The larva has been bred on corn. The moth flies in late June and July.

I have seen the moth from New York, New Jersey, western Pennsylvania, and Illinois.

III. Male antenna with laminations as long as, or even longer than, wide; palpus as in Sapinella, or even longer, extending well beyond thorax in male, short in female; wings blunt; vestiture of fine, flattened hair; eyes hairy, the hair short and easily lost in A. tenuis. (Anaphora Clemens).

3. A. popeanellus Clemens. Brownish luteous or dull reddish brown, rarely more purplish; distinctly paler along fold and inner margin. Thorax often pale, especially in female. Fore wing obscurely and irregularly striate with darker brown; scales not noticeably raised; blackish patches at end of cell and at middle of fold, and, normally, a dark antemedial bar across fold. (Fig. 95.) 25-30 mm. (scardina Zeller, agrotipennella Grote, confusellus Beutenmüller).

New Jersey to Missouri, Florida, and Texas.

A. busckella Haimbach, found at Brown's Mills, New Jersey, appears to be a black

 A. ous defined in the spectral and the state of the spectral state of the spectres state of the spectral state of the spectres state of the s A. tenuis. Slightly smaller than A. popeanellus, on the average. Male genitalia about as in A. popeanellus, but with the uncus shorter and weaker and the valve markedly broader.

The moth occurs in July, in the northern part of its range.

Plummer's Island, Maryland, to Georgia and Texas.

5. A. tenuis Walsingham. Nearly uniform purplish, with obscure dark markings; uncus curved over evenly, not angulated, and with more widely separated points than in A. popeanellus; valve spoon-shaped, distinctly narrowed at base, and with small supplementary process. 20-25 mm. (violaccellus Bentenmüller).

This is a more southern species than A. popeanellus. It flies in June.

Florida; North Carolina; central Illinois; Iowa; and elsewhere. 6. A. texanellus Chambers. Moth smaller and rather slenderer than A. tenuis, with more or less distinct traces of an oblique shade crossing the apex, as in

A. hulstellus. Uneus (fig. 96) with points separate, down-curved; valves slender, concave, uniform in width.

There seems to be some confusion as to the identification of this species. In the Barnes collection, the name is represented by a form with scattered white scales on the fore wing, and with valves as in A. hulstellus. North Carolina, Texas.

7. A. hulstellus Beutenmüller. Similar to A. texanellus but larger and coarser, with a strong, oblique, dark shade across the wing two-thirds of the way to the apex. Plummer's Island, Maryland, to Florida.

8. A. mortipennellus Grote. Pale, dull brown, usually patchily marked with dark brown; the discal spot most persistent. Female longer-winged and more evenly yellowish brown. Uncus of male (fig. 98) much as in A. popeanellus, but with the points somewhat enlarged and bluntly rounded; valves very slender, constant in width, and evenly down-curved. R_3 and R_4 or R_4 and R_5 are often stalked. 25-30 mm. (Hypoclopus Dyar).

The distribution is uncertain. The material before me is from Missouri.

IV. Palpus of male as in group III, but with somewhat heavier vestiture; palpus of female long, rather drooping, beaklike; thoracic vestiture as in group III; fore wing with all veins free; male antenna with laminations hardly twice as high as the length of a segment. Male with long supplementary process beside the value. Eyes hairy. (Acrolophus).

9. A. plumifrontellus Clemens. Usually light reddish gray, with irregular, dark striation, gathering into patches at end and near middle of cell, and also, basad of middle and near middle of fold; these patches sometimes connected to form a zigzag mark, but sometimes partly or wholly absent, especially in the female. Rarely with but little or no reddish tint. (Fig. 97.) 35 mm.

The moth flies in June and July. The southern race cervinus Walsingham is somewhat smaller and more brilliant.

Southern New York to western Pennsylvania and Florida.

V. Fore wing with \mathbf{R}_{a} and \mathbf{R}_{4} invariably long-stalked; uncus simple. Eyes naked. (Hypoclopus).

A. griseus Walsingham. Typically ashy white, often ranging from light blue-gray to fuscous, with confused blackish markings. Hind wing brown. Valve broad, oblique, narrow at base, squarely truncate. 22-25 mm. July. The eastern record for this species is based on a related but quite dis-

tinct, undescribed species, differing in structure as well as in its darker color. Texas; Arizona.

Subfamily TINEINÆ.

(Tineidæ, in strict sense)

Head more prominent, though often with small eyes; eyes naked; palpus porrect. or somewhat upturned in life, not large, clothed with scales and bristles, which are regularly arranged in most cases; tongue usually present; maxillary palpus usually folded, porrect in Amydria, absent when tongue is absent. Antenna typically with the basal whorl of scales on each segment reduced or replaced by a whorl of bristles, often entirely concealed; the outer whorl often raised at a sharp angle, and when flat covering the whole surface of the segment, whereas the two whorls (when there are only two) of the Adelidæ and Acrolophinæ are equally developed and usually of contrasting colors. Outer whorl rarely somewhat interrupted below, but antenna never laminate as in Acrolophinæ. Head with rough, bristly vestiture on both vertex and front, though comparatively smooth in the Amydria group. Hind tibia often bristled, but not with a regular row of bristles. Fore wing with R₅ running to costa or apex.

Larvæ with front not reaching vertex; prespiracular setæ of prothora, (fig. 92)

closely grouped; setæ i of abdomen farther apart than ii; iv and v widely separated, on separate tubercles. Prolegs with uniordinal hooks, in a complete ellipse or shortly interrupted on inner posterior side; the anal prolegs with a short band.

The larvæ are generally case-bearers, the case usually ending in a three-cornered valve, as in the lower Coleophoridæ and Psychidæ; occasionally it is lenticular. The larvæ are often scavengers.

Pupa only lightly soldered together; wing lanceolate; antenna extending to wing tip; in Tinea, at least, with two prominent spines at the end of the abdomen. Cocoon of silk, usually in the larval case.

2. SCARDIA Treitschke

Antenna moderate, with pecten and with raised scale-whorls. Maxillary palpus small but folded; tongue strong and naked. Palpus long, porrect, with a large tuft on under side of second joint and bristles on outer side; third joint long, upturned. Head rough. Wings heavy, hardly lanceolate. Fore wing sometimes with \mathbf{R}_3 and \mathbf{R}_4 stalked, hind wing with \mathbf{M}_1 and \mathbf{M}_2 connate; with fringe one-half as wide as membrane. Hind tibia with long hair.

The larvæ bore in fungi.

Key to the species

1. Blackish, with contrasting, pale inner and outer margins.

2. Fore wing broader6. anatomella.

2. Fore wing narrower: with distinct, paired, pale costal striæ.....5. fiskeella. 1. Ground paler, often heavily overlaid with blackish; inner and outer margins

not pale. 2. A black discal bar (inconspicuous when the region about it is blackish).

3. Fore wing with a dark postmedial fascia, angled on outer, and erect on inner,

- 2. No black discal bar.
 - 3. A contrasting, pale antemedial fascia between the dark basal and medial

1. S. fúscofasciella Chambers. Clay color, dusted and mottled with brown, with more or less paired, dark costal strik; the dark postmedial fascia quite obscure and concave below the angulation. 28 mm. (Euplocamus Chambers).

This species was described from Kentucky.

2. S. errandella Busck. Similar to *S. fuscofasciella*, the ground more evenly luteous, the markings dull fuscous, with the usual purplish iridescence more sharply contrasting. The paired costal striæ distinctly pale; the darker post-medial band as in *S. fuscofasciella*, containing the slightly darker discal bar. 18 mm. (tessulatella auct., not Zeller).

July and August in western Pennsylvania.

3. S. approximatella Dietz. Luteous, dusted and mottled with fuscous; the dark fascia somewhat anterior to the middle; irregular on basal side, and often running in below the cell to join an oblique fascia from base of costa, thus forming a sort of W which is nearly erect on outer side. A subterminal dark streak

running to the costa before the apex. Antennal cilia long (2). 15 mm. The moth occurs in July. The larva has been found in a rotting sycamore log. Massachusetts to western Pennsylvania and Georgia. New York: Ithaca. 4. S. pravatella Busck. Similar to S. fuscofasciella, with basal third heavily dusted and contrastingly dark except on inner margin. Wing narrow. 23 mm.

Western Pennsylvania.

5. S. fiskeella Busek. This species is distinguished from *fuscofasciella* and *prava*tella by the lack of transverse bands, from anatomella by the narrow wings and the tendency for the pale outer margin to widen more opposite the cell. 25 mm.

North Carolina.

6. S. anatomella Grote. Dull black, reticulate and mottled with intense black; discal bars light. Slight pale spots on costa; outer and inner margins very irregularly edged with a contrastingly pale, clay-colored area, dusted with dark fuscous, and sometimes broken into spots. 25 to over 30 mm.

The moth occurs in June and July in western Pennsylvania and is also known from western Ontario. Dietz records it from New York.

3. ELATOBLA Herrich-Schæffer

(Abacobia Dietz, not Lacordaire; Dietzia Busck)

Similar to Tinea: \mathbf{M}_1 and \mathbf{M}_2 of hind wing stalked: tongue and maxillary palpus smaller than usual but not absent. Antenna half as long as fore wing, with pecten. Hind wing as wide as fore wing. Palpus with second joint clavate, third with some bristles. Female ovipositor membranous. Larva dirty white, with reddish head and anal plate and blackish cervical shield

and tubercles; boring in the bark of pine in early spring.

1. E. fuliginosella Zeller. Dark fuscous brown; hairs of head and palpi white-tipped; discal spots darker. Hind wing paler. 14-19 mm. (martinella Walker, carbonella Dietz).

The moth occurs in July.

Hazleton, Pennsylvania; Hudson Bay district; Colorado; Europe.

4. TINEA Linnaus

Small moths, the largest hardly an inch in expanse. Palpus smallish, smoothscaled, and more or less distinctly bristled on outer side, occasionally with some spatulate scales. Head, including front, with rough, bristling hair, often con-trastingly colored. Maxillary palpus and tongue developed; antenna with whorls of raised scales and pecten. Hind tibia with spurs above middle. All veins free. Base of **M** more or less distinct; \mathbf{M}_2 and \mathbf{M}_3 of hind wing stalked in *T*. rileyi. Hind wing lanceolate, about as wide as fore wing.

Argyresthia may be distinguished by the smoother face, and by Rs running to outer margin; Acrolepia, by the much smoother head. In the following key I have only been able to include the species known to me

personally, and a few others of striking character. Incre are a good many other described forms, some or which may not be valid.

Key to the species

- 1. Hind wing (fig. 73) as broad as fore wing; subcosta extending three-fourths its length or more, and cubital branches long, except in a few species with yellowish head and simply marked fore wings; costa not sinuous.
 - 2. Head dull ochre to fuscous. Fore wing usually nearly concolorous, or if blackish, strongly mottled, and with dark discal dots (obscure in the dark form of \overline{T} . carnariclla).

3. Eyes narrower than the distance between them below.

- 4. Eyes very small, half as wide as the distance between them; fore wing obscurely mottled in two shades of fuscous......2. obscurostrigella.
- 4. Eyes two-thirds as wide as the space between them; mottling contrasting, blackish on luteous.....l. misella.
- 3. Eyes wider than the space between them below.

4. Ground even, shining; each scale dark-tipped; discal dots distinct.

10. pellionella.

- 4. Fore wing mottled, with blackish scales in groups, or with scattered, contrasting, dark scales, at least toward apex.
 - 5. A blackish longitudinal streak from base to apex.....4. apicimaculella.

- 5. No longitudinal streak from base to apex.
 - 6. Head ochreous; scape of antenna with an ochreous dot.
 - 7. Fore wing dominantly blackish, with contrasting yellow inner margin6. bimaculella.
 - 7. Fore wing dominantly yellowish or fuscous.
 - 8. Hind wing much narrower than fore wing, with a short Cu₂. 9. Expanse about 12 mm.; fore wing with a single (claviform)
 - 8. Hind wing fully as wide as fore wing, with long Cu₂.
 - 9. Fore wing with a conspicuous series of blackish bars in base of fringe9. grumella. 9. Fore wing without contrasting dark bars in fringe.

8. carnariella.

- 6. Head fuscous; fore wing dominantly fuscous; scape of antenna all fuscous.
 - 7. Ground dark purplish fuscous, either plain or mottled with blackish; tegulæ unicolorous......11. grisseella. 7. Ground luteous, mottled with fuscous or brown; tegulæ only half
- 2. Ground blackish, without discal dots, or with pale ones; head frequently vellow and strongly contrasting.
 - 3. Fore wing without pale marginal spots.
 - 4. Thorax with a central, yellow stripe.....14. thoracestrigella.
 - 3. Fore wing with a series of contrasting, cream-white marginal spots.

15. ophrionella.

- 1. Hind wing (fig. 74) almost without exception decidedly narrower than fore wing; the fringe wider than the membrane; Sc reaching margin three-fourths of the way out; costa a little emarginate near the middle; discal spots generally obsolete; head white, as a rule, and. in broad-winged species, never ochre yellow.
 - 2. Fore wing with longitudinal stripes.

3. Fore wings white, markings dark; head white.

4. Two longitudinal stripes through the disc16. roburella.
4. Numerous longitudinal lines17. multistriatella.
3. Head and fore wings dark brown, streaked with white
3. Head brown and white, for wwing dark-striped on gray $\dots 17\frac{1}{2}$. atriftua.
For wing without strongly wayked longitudinal strings

- Fore wing without strongly marked longitudinal stripes.
 - 3. Scape of antenna blackish.
 - 4. Fore wing with considerable areas of brown, often forming brown shades; median fascia often confused.
 - 5. Two oblique, dark, median fasciæ, the outer half of the wing half blackish24, marmorella. 5. A single angulate, or broken, black median fascia.

 - 6. Head white, slightly gray behind antennæ.
 - 4. Fore wing lightly dusted with gray (the scale-tips dark); a square, black patch at middle of costa alone contrasting 20. auropulvella. 3. Scape white; fore wing white.
 - 4. Ground with a few gray-tipped scales; fascia of powdery gray.

21. acapnopennella; 21½, tylodes.

4. Ground white, contrastingly marked with black.

- - 5. Median black fascia not reaching inner margin; with but little black at base of costa.

 - 6. The strongest marking a broad median fascia crossing the fold.

23. angulifasciella.

See also: 12 misceella, 25. fuscomaculella, 29. nigratomella, and 28. maculabella.

1. T. misella Zeller. Eyes small, elliptical, separated by nearly twice their width. Palpus rather longer than usual, with considerable hair on the second segment. Cu_2 of hind wing three-fourths as long as width of wing.

Clay color; shoulder fuseous; fore wing heavily mottled with fuseous strike, tending to gather into quadrangular spots, among these the inconspicuous discal spots. Costal fringe barred. 12-23 mm.

The Pennsylvania specimens before me are darker, and appear to have less loose hair on the palpi than the European ones.

June to August.

Europe; Minnesota; Pennsylvania; Texas; Florida.

2. T. obscurostrigella Chambers. Similar to T. miscila. Eyes rather smaller, fore wings more evenly and less contrastingly mottled (sometimes nearly even, except for the discal dots). 12-18 mm.

May, July, and October.

Hazelton, Pennsylvania; St. Louis, Missouri.

3. T. fuscipunctella Haworth. Pale fuscous, tending to clay color, with a somewhat reddish head. Antennæ blackish. Palpi and antennæ nearly concolorons. A transverse, dark band across front of thorax and basal half of tegulæ, extending along the costa of the fore wings a tenth of the way out. Fore wing more finely and less extensively mottled with fuseous than in *T. obscurostrigella*; costal and dorsal fringes barred. Dorsal spots large, diffuse, contrasting, sometimes joined by a dark shade extending to base of wing. Hind wing paler, hardly iridescent. 11-17 nm. (*nubilipennella* Clemens, *Œcophora frigidella* Packard).

The black bar at the base of the costa seems to be distinctive. The eyes are normal in this and the remaining species of Tinea. This moth appears to be our commonest Tinea, and flies in May and June, and again in August. The larva breeds in the trash in pigeons' nests and similar refuse.

Labrador to Pennsylvania; Europe.

4. T. apicimaculella Chambers. Dull straw yellow or elay color. Outer side of palpi blackish. Thorax dark with some clay color on the tegulæ. Fore wing with the costal region shaded heavily with fuscous, the shade extending to the blackish streak on the outer part of the wing, and beyond it to the fold, toward the base. A blackish streak from the base almost to the apex, interrupted around the black outer discal dot. Reniform and elaviform dots strong, blackish.

Basal half of fringe barred with blackish. 11-15 mm.

The moth has been taken in June northward, and from April to July in the south.

New Jersey to Florida, Kansas, and Louisiana.

5. T. straminiella Chambers. Straw yellow. Sides of palpi and of thorax brown. Fore wing dusted with brown toward apex, with a row of dark spots along the fold and one at the end of cell. 8 mm.

This species was taken in June. No authentic material seems to be known: I believe it is a light variety of *T. bimaculclla*.

Kentucky.

6. T. bimaculella Chambers. Head pale yellowish, antennæ fuscous; tegulæ dark brown. Fore wing narrow, lanceolate, light yellow, heavily dusted with fuscous. or solidly purplish, most of the inner margin contrasting, clear light yellow. Outer discal dot large, blackish when not lost in the dark ground. Three blackish spots in fold, near base, and before and beyond middle. Hind wing narrow as in the granella group. 9 mm. (mandarinella Dietz).

I have seen the types of both names, and they are the same species. The species determined by Kearfott as bimaculella is not a Tinea.

June to August.

New Jersey to Ohio and Kentucky. 7. T. _____. Straw yellow. yellow, three-fourths as wide as the fore wing. Cu, short. 12 mm. (trimaculella Chambers?)

July. St. Louis, Missouri; (District of Columbia; Kentucky). New York: Otto. 8. **T. carnariella** Clemens. Similar to the preceding species. Deeper ochre yellow, the fore wing sometimes suffused with fuscous, leaving a contrasting yellow inner margin. Head without a black spot behind antennæ; palpi yellower on inner face; disc of thorax mostly yellowish. Fore wing more or less dusted with blackish scales, costal edge less contrastingly blackish, discal spots larger, diffuse, the one in the fold obscure. Hind wing dirty white, broad. 15 mm.

St. Louis, Missouri; Pennsylvania.

9. T. grumella Zeller. Head ochreous, darker than in T. pellionella. Yellowish, shaded with fuscous, especially toward costa and beyond the discal dot, but leaving the fold contrastingly pale. Usual spots dark, the one in the fold short. A series of blackish terminal bars. Hind wing somewhat narrower than fore wing. 16-18 mm. (?)

The length of the fore wing is given as four lines, which is too large for anything I have seen determined as grumella. The present description is condensed from Zeller's.

July.

Massachusetts.

10. T. pellionella Linnæus. (The old-fashioned clothes-moth.) Pale fuscous gray, a little shining. Head yellower than ground, but not decidedly yellow as in the preceding species. Antennæ, including whole of scape, and palpi, blackish. Fore wing with a good many scales darker toward apex, but with no definite dusting; discal spots distinct, but blurred; no terminal bars. Hind wings light gray. 10-15 mni. (H. p. 473, f. 253.)

The larva is white with a brown head and collar, and lives in a parchment-like case of white silk. It is one of our clothes moths, feeding on woollen goods, furs, and other dry substances of animal origin, but appears to be much less common under American conditions than is Tineola bisselliella.

The distribution is said to be general; I have seen the species from Michigan and Missouri, and, I think, from Alabama.

New York: Reported from Louisville, Canandaigua, and Alfred Center. 11. T. grisseella Chambers. Fuscous, head somewhat yellower than fore wing, with ocherous and fuscous palpi. Fore wing heavily striolate on a clay-colored ground. Outer discal dot distinct, and also two dark bars in the fold. 8 mm.

I have seen no authentic material of this species. The original description is strongly suggestive of T. pellionella.

Kentucky.

12. T. misceella Chambers. Head and palpi pale yellowish; antennæ pale fuscous; fore wings dusted with fuscous and saffron yellow in about equal proportions; two discal spots, one two-fifths of the way out, in the fold, and a smaller one at end of cell. 8 mm.

This species is unknown to me; it should be recognizable by the light palpi, and the absence of the first diseal dot, or its fusion with the elaviform.

Kentucky.

13. T. croceoverticella Chambers. Head bright ochre; antennæ blackish, with some ochroous scales on scape; palpi dull ochroous, gravish on outer side, with rougher, longer vestiture than usual, twice as long as the width of the joint, but not spatulate as in Elatobia. Thorax dark mouse gray, the apical half of the tegulæ clay color, contrasting. Fore wing blackish, with slightly darker shades in place of the discal dots, with obscure, pale marginal bars toward the apex, and a pale streak at the base. Two dark lines in fringe. Hind wing dark. 10-15 mm.

The larva is a case-bearer and feeds externally on the flat, white bracket fungi on beech, usually on small bits. The case tapers to a neck at each end and then has a flaring mouth. It is covered externally with sawdust and grass. It has been found in June and the moths emerge about the first of July.

Maryland to Kentucky, Kansas, and Saskatchewan (Regina) Canada.

14. T. thoracestrigella Chambers. Similar to T. croceoverticella, brighter, not bronzed, without lines in the fringe. 10 mm. smaller,

This species is definitely known only from Texas, but is doubtfully credited to Kentucky in Dyar's list.

15. T. ophrionella Dietz. Deep purple-brown. Head yellowish white with some black hairs behind. Scape silvery, inner side of palpi white. Fore wing with a series of six or seven white marginal bars, with a wider space between the two middle ones, and several on the inner margin lying mostly in the fringe; a single, white discal dot and several white patches below the fold. 11-14 mm. June.

Parry Sound, Ontario. New York: Uphill Brook (Mt. Marcy).

16. T. roburella Dietz. Head white, a fuscous spot on vertex; thorax and fore wing white, dusted and shaded with pale brown. A large, seal-brown spot from middle of costa to middle of wing, crossed by two longitudinal, dark streaks. Inner margin also narrowly dark on outer part of wing. Margin and fringe barred with brown, the latter with a dark central line. 13 mm.

Essex County, New Jersey.

17. T. multistriatella Dietz. White, lightly dusted and finely streaked with fuscous; with five or six partly confluent streaks, sometimes broken up into dashes and dots, the costal spots smaller. 17 mm.

Bred from fungi on beech stumps.

Toronto, Ontario; Maryland. 17½. T. atriflua Meyrick. Head white, shaded with fuscous on the sides; thorax dark, with apex of tegulæ whitish. Fore wing dark-dusted on a grayish white ground, the margins narrowly white, also with dark scales. Two blackdusted streaks, one from base of costa to apex, widening outwardly and occupying nearly half the width of the wing at the outer margin; the other below the fold; both edged with white. Some small bars on onter margin, and powdery dark dots in the white fringe. Hind wing pale gray with paler fringe. 13 mm. (Unknown to me).

June.

Toronto, Ontario.

18. T. rileyi Dietz. Scal-brown, head nearly black, including outer side of palpi and scape; tegulæ white-dusted. Fore wing with about five fine, white lines and white costal and dorsal edges. Fringe checkered, largely white.

The larvæ feed on rungi. The moths emerge in June.

Pennsylvania; District of Columbia; Florida.

In the remaining species of Tinea, the hind wing is about threefourths as wide as the fore wing, with the costa noticeably sinuate and \mathbf{Cu}_2 no longer than \mathbf{Cu}_1 ; the head and palpus are white, the palpus

being black on the outer side of the second segment and extreme base of the third, except as noted.

19. T. arcella Fabricius. Head all white, tegulæ with a black spot at base. palpus occasionally all black on outer side, normally typical; scape white. Thorax and fore wing white, fore wing with a little yellow shading and an angulate, black, median band, sometimes broken; a spot covering the basal fifth of the costa, and smaller blackish marks outwardly. 12 mm.

American specimens are not typical and may represent a new species. The larva as known in Europe is yellowish white, with a bright brown head, and has been bred on fungi under the bark of dead alder twigs.

July.

Europe; Connecticut to District of Columbia and Indiana; California (?), New York: Ithaca (Dietz).

20. T. auropulvella Chambers. Head white; scape partly dark gray; second segment of palpus outwardly all black, third segment all white. Fore wing white, mottled with a little light golden brown, with a square, black spot on the middle of the costa, continued by a narrow streak across the cell in dark specimens; a black antemedial spot and sometimes a basal spot on the costa; some other small black spots present, mostly on the margins.]1-16 mm.

June and July.

New Hampshire to North Carolina and Kentucky, westward to British Columbia. New York: Ithaca.

21. T. acapnopennella Clemens. Head with some gray scales, thorax and fore wing in some specimens considerably dusted with black-tipped scales, scape white, palpi typical. Pattern of fore wing about as in *T. auropulvella*, but less contrasting; the spots at the base of the costa rarely conspicuous. 10-16 mm. (minutipulvella Chambers).

June to August. Larva on Polyporus tulipiferus; whitish, with dark head and cervical shield.

Canada to Louisiana. New York: Ithaca, Albany. 21¹/₂. T. tylodes White. Antenna light gray, palpus with a dark streak on outer side of second joint; shoulder with a gray spot. Fore wing with some irregular yellowish scaling, and gray dusting, especially toward outer margin; markings blackish gray; some strike on margins; a streak along basal fifth of costa; a semioval spot on middle of costa with a rhomboidal spot before it on inner margin; an irregular outer spot on costa. Apical fringe with two more or less distinct gray shades. Hind wing light gray with paler fringe. 14 mm. Meyrick does not compare this species, which is unknown to me, with any

other. It may be the same as acapnopennella.

July and August.

Toronto and Muskoka Lake, Ontario. 22. T. apicisignatella Dietz. Head and scape white, shaft of antenna light brown; palpi typical. Fore wing dull whitish, spotted with black; a black square at middle of costa, extending well into the cell, and an equally large, black, subterminal spot separated by a narrow white band from the black apex; fringe barred. 12-13 mm.

This species is very close to T. auropulvella but appears distinct.

New Hampshire; Pennsylvania.

23. T. angulifasciella Dietz. Head white with a gray posterior tuft, half of third segment of palpus black; scape white; shaft of antenna pale; shoulders black; fore wing white; markings all black; heavy basal line reaching the fold, the base of the costal edge black; a small antemedial dot, a heavy angulate median fascia, extending to below the fold and sometimes weakly even to the inner margins; outer part of wing strongly mottled with black, forming a large, central dark patch. \mathbf{R}_4 and \mathbf{R}_5 stalked.

Plummer's Island, Maryland.

24. T. marmorella Chambers. Head and palpi yellowish white; second segment and half of third segment of palpi brown on outer side; antenna dark; scape blackish. Fore wing white, spotted with black; basal fifth of costa black; a little black toward inner margin; an antemedial, outwardly oblique, black fascia crossing the cell and fold (apparently interrupted in the specimen before me); a similar postmedial fascia covering the end of the cell; some other black markings on dise, and margins barred with black. 10 mm. (not "three-sixteenths inch"!)

Toronto, Ontario, May 25.

25. **T. fuscomaculella** Chambers. Closely similar to T. marmorella; the basal black streak on the costa running into an oblique fascia as in T. marmorella and granella, but the markings on the outer part of the wing less confluent. 12 mm.

I am not sure that this form is distinct from T. marmorella; the Canadian specimen of marmorella before me seems intermediate.

Kentucky.

26. **T. granella** Linnæus. Head rarely yellowish, with some gray behind the antenna. Scape black; palpi with only the tip of the third segment white outwardly. Fore wing white, more or less mottled with dark, bronzy brown, and spotted with black. Disc of thorax fuscous, tegulæ black with white tips. Fore wing with a black basal bar, antemedial spot, median bar extending into the cell, with a separate dot in the fold, and smaller outer bars, the two outermost sometimes as in T. apicisignatella. The markings often leaving clear a vague, white discal dot. Fringe with dark center-line, crossed by two white bars. 12 mm. (variatella Clemens).

The caterpillar is yellowish white, with a red-brown head and two curved, brown stripes on the neck. It feeds in grain, dried fruits, bracket fungi, and similar substances, breeding continuously when the temperature permits.

Europe. Distribution apparently general in the eastern United States. New York: Albany, Batavia.

T. cloacella Haworth is very similar, but with a yellow head, more dark behind the antennæ, and sparser and paler brown mottling, contrasting with the black spots. It is something of a pest in Europe but has not been authentically reported from America.

27. T. fulvisuffusella Dietz. Very near T. granella, but smaller, with lighter brown mottling, and less extensive black markings. $111\frac{1}{2}$ mm.

Hampton, New Hampshire.

28. **T**. maculabella Chambers. White. Antennæ sordid yellowish white. Thorax with three brown spots in a triangle. Fore wing white, lightly dusted with dark brown, and marked with dark brown; a brown spot at base of costa, an antemedial spot and an oblique irregular streak from before the middle of the costa to beyond the fold, interrupted on the fold; a small spot on middle of costa; and a postmedial streak running into a large, longitudinal patch in the middle of outer part of wing; a couple of subapical, longitudinal streaks. All the markings more or less mottled and defined with reddish yellow. A series of dark terminal spots. \mathbf{R}_{i} and \mathbf{R}_{5} stalked. 14-22 mm.

This species is unknown to me, but its large size appears distinctive, as well as the longitudinal apical dashes.

Pennsylvania; Kentucky.

29. T. nigratomella Dietz. Palpi, head, and antennæ sordid yellowish white; palpi dusted with brown, antennæ faintly annulate. Fore wing white, heavily and almost evenly dusted with brown, gathering in two vague transverse bands, the first erect and the other oblique. Costa at apex barred with alternate dark brown and white spots. $8\frac{1}{2}$ mm.

This species is unknown to me and may be a Diachorisia, like "Tinea" marginimaoulella and "T." fuscopulvella, with which it is compared.

Montclair, New Jersey.

I have seen a specimen from Pennsylvania labelled *Tinea defectella* Zeller. Its fore wing is white, patchily mottled with fuscous, tending to form oblique fasciæ, and the head is fuscous and white. 8 mm. I am not sure it is rightly determined.

T. uterella Walsingham, a blackish tropical species a good deal like T. misella, but lacking the pecten, is likely to occur in the warmer States. The larva is Tinea-like but with heavy thoracic sclerites, and forms a flat case like a melon seed. The species is usually put in the genus Tincola, but the tongue and maxillary palpi are preserved.

5. TRICHOPHAGA Ragonot

(*Tinea*, in part)

Tongue obsolete; but maxillary palpi preserved; costal veins as noted in key (fig. 78). Otherwise like the first group of Tinea. Antennæ rather more smoothly scaled than in Tinea, with only a narrow naked area at the base of each joint.

1. T. tapetzella Linnæus (Case-bearing clothes moth). Blackish, outer half of fore wing contrastingly whitish. 12-24 mm. (H p. 434 f. 254.) This species is apparently nearly world wide. The larva is a clothes moth and general scavenger, often breeding in owl-pellets; it is pale, with a black head

and a dark brown shield.

Distribution general south of New York City. New York: New Dorp, Staten Island.

6. MONOPIS Hübner

Blabophanes Herrich-Schæffer

Body and head like Tinea. Antennæ usually with rather strong sense bristles. Fore wing (fig. 77) with a small fovea near middle; \mathbf{M}_3 and \mathbf{Cu}_1 stalked, \mathbf{R}_4 and \mathbf{R}_5 sometimes stalked, \mathbf{Cu}_2 often stalked with \mathbf{M}_3 and \mathbf{Cu}_1 . Hind wing with \mathbf{M}_1 and M₂ rarely stalked.

Spuler makes a separate family, Monopidæ, for this genus; but the characters are not unlike those of Tinea, and the larval habits are the same.

Key to the species

1. Disc of thorax and inner margin of fore wing contrasting yellow.

2. Thorax and inner margin bright yellow.

3. Fore wing blackish with violet iridescence 3. ferruginella. 3. Fore wing blackish fuscous, flecked with pale ochreous.

4. croeieapitella.

- 2. Thorax and inner margin yellowish white2. dorsistrigella.
- 1. Disc of thorax concolorous, and inner margin slightly paler, or with concolorous inner margin
 - 2. A pale spot on middle or outer part of costa.
 - 3. Inner margin solidly dark or with a small dot at anal angle
 - 5. monachella. 3. Inner margin paler or with a pale patch, at least half as large as the costal one.
 - 4. Costa without dark spots before the middle.....6. biflavimaculella.
 - 4. Costa with alternate clay-colored and dark striæ

	5	. Tips of	tegulæ j	ale		 		. irrorella.
	ð.	. Tips of	tegulæ	concoloro	us	 	.7. margi	nistrigella.
2.	No p	ale areas	s on any	part of	wing.	 	8.	rusticella.

I. Cell half as long as fore wings: \mathbf{R}_{4} and \mathbf{R}_{5} separate, as a rule; \mathbf{Cu}_{2} free, \mathbf{M}_{1} and \mathbf{M}_2 of hind wing separate. Male values with straight ends and sharp angles. Cell more than half the length of fore wing (Blabophanes).

1. M. irrorella Dietz. Equally mixed with whitish and dark brown; head saf-

fron yellow; anal angle dark brown, inner margin dominantly pale, except at anal angle; costa also paler, especially toward middle. Thorax blackish; tips of tegulæ luteous. 11-14.5 mm.

June.

Pennsylvania; New York: Otto.

2. M. dorsistrigella Clemens. (Cu. rarely stalked). Head, disc of thorax, and inner margin of fore wing white, sometimes tinted with yellow; streak on inner margin widened before and beyond the middle; a triangular white patch in middle of costa, reaching down to the fovea; a few white flecks at the apex. Tegulae and ground color of fore wing dark purplish brown. 12-14 mm. (subjunctella Walker).

June and July.

Massachusetts and North Carolina to southern Ontario, Missouri, and New Mex-

ico; probably general. New York: Ithaca, Sea Cliff, Long Island.
3. M. ferruginella Hübner. Black with violet iridescence, Dorsal streak light cream-color, slightly widened before and beyond the middle; costal patch white, usually reduced to a few scattered scales. Head bright ochre. A few silvery tlecks at apex. Hind wing purplish fuscous in male, darker in female. 11-15 mm.

The larva is a clothes moth and a general scavenger.

This species is definitely known only in Europe, but is to be looked for, mixed with M. crocicapitella.

4. M. crocicapitella Clemens. Similar to M. ferruginella; the ground color somewhat purple-brown, paler and flecked with yellow along the costa and outer third of the wing; the triangle not distinct; hind wing and fringes pale gray. 9-17.5 mm. (hyalinella Staudinger, lombardica Hering, heringi Richardson, ferruginella Dyar, not Hübner).

July to October.

The larva has been bred from seeds of absinth and refuse, and the moth has been taken in a cave, associated with bats.

New York to California, probably general; also in Europe and Hawaii. New York: Morrisania (New York City); Clove Valley, Staten Island.

II. Fore wing with R_4 stalked with R_5 , Cu_2 with Cu_1 and M_3 ; force small. Cell about half as long as wing. Hind wing with M₁ and M₂ separate.

5. M. monachella Hübner. Mottled fuscous gray. Head and thorax white; a large, trapezoidal, costal white patch, reaching the fovea. \mathbf{R}_4 and \mathbf{R}_5 stalked, \mathbf{Cu}_2 short-stalked. 11-17 mm.

Europe.

This species seems to have been introduced in Manitoba, and at St. Hilaire, Quebec. It flies in June and July.

6. M. biflavimaculella Clemens. Head pale luteous. Thorax dark brown with a pale spot in front. Fore wings motiled umber brown tinged with violet; a strongly contrasting, triangular, cream-white patch on middle of costa, and a smaller one farther out on inner margin. 15 mm. (insignisella Walker).

The moth occurs in June to September. I have seen a specimen with M1 and \mathbf{M}_2 of the hind wing stalked, and one with \mathbf{R}_4 and \mathbf{R}_5 of the fore wing completely united.

Labrador to Texas and Washington. New York: Peru, Wilmington, Ithaca, Big Indian Valley.

7. M. marginistrigella Chambers. Dark purplish brown mixed with white; head contrasting yellowish white. Pale spots as in the last species, the one on hind margin more narrowly continued toward base. Apex largely pale. 9 mm.

June and September.

Kentucky; southern Ohio; western Pennsylvania.

III. Fore wing with end of cell and forea well before middle of wing; \mathbf{R}_{\star} and $\mathbf{R}_{\mathbf{S}}$ and \mathbf{Cu}_2 long-stalked, \mathbf{M}_1 and \mathbf{M}_2 of hind wing long-stalked; values of male rounded oval (Monopis).

8. M. rusticella Hübner. Almost evenly mottled with purplish fuscous; fovea pale. Head whitish yellow. 14-20 mm.

The moth has apparently been introduced in the New World, and occurs only sporadically in our territory. It flies from May to midsummer, and again in autumn, having two broods.

Eurasia, Truro, Nova Scotia; Toronto; Hampton, New Hampshire. New York: Ithaca (probably introduced).

7. TINEOLA Herrich-Schæffer

(*Tinea*, in part)

Moth similar to Tinea but with rudimentary tongue and maxillary palpi. Pecten present. Eyes of male very large.

There is a rudiment of the stigma of Trichophaga, the genus to which this seems most nearly related. It comes from the old-world tropics.

1. T. bisselliella Hummel (the clothes moth). Pale straw yellow, head more rusty. Fore wing occasionally with dark outer discal dot, 12-15 mm. (lanariella Clemens). (H. p. 432 f. 252.)

The larva of this species is the commonest of the clothes moths, and is white with a brown head. It forms slight silken galleries, but lives nearly free. It pupates in a translucent cocoon. It will eat wool, fur, insect specimens and occasionally other dried animal substances, but avoids cotton.

Common and general in distribution. New York: common everywhere.

8. TENAGA Clemens

Eyes very small with a naked space all around. Antennæ nearly as long as fore wing; palpi nearly concealed in the rough facial hair. No maxillary palpi Fore wing ovate-lanceolate; cell narrow, below middle of wing; one or tongue. radial lost; \mathbf{R}_4 and \mathbf{R}_5 stalked, \mathbf{M}_1 and \mathbf{M}_2 stalked, but free from \mathbf{R}_4 and \mathbf{R}_5 ; accessory and discal cells weakly closed outwardly. Hind wing lanceolate without wellmarked cell. Sc ending about two-thirds way out, but costa not sinuate; M, and \mathbf{M}_2 stalked; \mathbf{M}_3 free; \mathbf{Cu}_1 angulate where the discocellular vein usually is attached; fringe wider than the membrane; anals reduced.

This genus apparently is related to Tineola. 1. T. pomiliella, Clemens. Yellowish ochreous, with three, irregular, broken, brown fasciæ and scattered spots. 10 mm.

June, August, and September.

Southern Ohio; Pennsylvania.

9. ISOCORYPHA Dietz

(Incurvaria Clemens, in part)

Similar to Diachorisia. Antennæ smooth-scaled, with a single whorl to a segment. Fore wing (fig. 84) smooth scaled and slightly shining; \mathbf{R}_4 and \mathbf{R}_5 short-stalked, and \mathbf{A} simple. Hind wing three-fifths as wide, with \mathbf{Cu}_2 two-thirds as long as its width; and costa not sinuate. All veins separate; anals traceable.

I. mediostriatella Clemens. Antenna nearly as long as fore wing. Head yellow. Shining brown; a large golden yellow patch two thirds way out on costa; a broad streak through middle of wing, almost or quite connecting with it, and half as wide as the wing; and a patch at beginning of dorsal fringe, usually joining the streak. Hind wing dark. Costa slightly sinuous beyond middle. 8 mm.

July to September.

Superficially, this form is extremely close to Hybroma chrysocomella, but it has a brown thorax and paler antennæ, besides the venational difference.

10. DIACHORISIA Clemens

(Homosetia Clemens; Pitys Chambers; Calostinea and Stenoptinea Dietz; *Pelates* Dietz?)

Similar to the last group of Tinea but with some raised scaling on the fore wings; wings often heavily tufted; or more reduced, the fore wing (fig. 82) being lanceolate and the hind wing sometimes linear, with cell nearly marginal. A of fore wing, in the latter case, not forked at base, and wing-scaling sometimes nearly smooth. Occasionally with metallic markings. Antennæ usually smoother than in Tinea, the outer whorl of scales covering nearly the whole of each segment.

The species have not been fully studied and may not all be congeneric. Besides the names listed in the synonymy above, Infurcitinea seems also to be a synonym of this genus, being related to group Calostinea. Some of the species listed below may even belong to the Adelidæ.

Key to the species

- 1. With accessory cell; no definite scale-tufts; hind wing broader, with Cu_2 nearly as long as width of cell, and with anals developed.
 - 2. Male antenna distinctly public public public products of the stalked; costa of hind wing sinuate (Diachorisia, fig. 82)...1. velatella.
 - 2. Male antennæ smooth-scaled; fore wing with metallic markings; costa of hind wing hardly sinuate, \mathbf{R}_4 and \mathbf{R}_5 free (Calostinea).
- the margin.
 - 2. Both wings with all veins free; antennæ long, normal; costa of hind wing sinuate; fore wing with scale-tufts (Homosetia, fig. 80).
 - 3. Fore wing dark brown or blackish brown.
 - 4. Face white, vertex black.
 - 5. A large, well-defined silver spot near the base.
 - 6. No white terminal dots.....4. cristatella. 6. Distinct white terminal dots on costa and outer margin.

3. argentinotella.

- 3. Fore wing suffused with tawny or golden brown.
 - 4. White costal spots.
- 5. Spots toward the apex of the fore wing, perpendicular to the costa, 3. Fore wing dominantly grayish fuscous on a white ground, with yellowish spots, if there is any yellow.
 - 4. Fore wing with golden dots.

5. Fore wing wider; subcostal tuft of median facia large.

		1. miscecristatella.
	5. Fore wing narrower, the tuft small	10. tricingulatella.
	4. No yellow or golden14	. marginimaculella
2 .	. Hind wing with \mathbf{M}_1 and \mathbf{M}_2 stalked, front with a conical t	tuft (Stenoptinea).
	3. Apex of fore wing and fringe buff-brown	13. auriferella.
	3. Apex of fore wing and fringe not pale	12. ornatella.

1. D. velatella Clemens. (Fig. 82.) Head mixed brown and gray; palpi dark outwardly. Fore wing white, dusted with fuscous; with four or five unequal spots along costa and the usual three discals, which are rather large; the apex with additional confused spots. 9 mm. Antennæ with outer whorls of scales on each segment nearly complete, but no longer than the inner whorls. Ventral surface with strong pubescence, as long as a segment in male; shorter but of the same character in female. Hind wing with cubitus and Cu_2 running straight across to inner margin, with a quarter of the width of the wing below it. Cu_2 half as long as width of wing. (In the rest of the genus Cu is closely parallel to the inner margin, and the free part of Cu_2 is very short.)

July.

Maryland; Pennsylvania.

2. D. argentistrigella Chambers. Rich, iridescent, maroon brown; face and antennæ below, silvery white; antennæ annulate above. An irregular, silvery fascia before middle, the wing beyond it dusted with white; six costal streaks toward apex, the next to last being the longest, and two dorsal streaks. 12 mm. (Semele Chambers.)

I have not seen this form.

Kentucky.

3. D. argentinotella Chambers. Similar to *D. argentistrigella*; the fascia on the fore wing absent, or replaced by a costal patch, which typically reaches the cell. 10-17 mm. (Semele Chambers.)

The cubitals in the hind wing are very short. In Florida and Texas the moth flies in June.

Western Pennsylvania to Florida and Texas.

4. D. cristatella Chambers. Face white, sometimes suffused with fuscous; vertex black; thorax black, with more or less broken silvery or golden basal, antemedial, postmedial, and subterminal bands, defined with raised scales; a heavy tuft beyond the postmedial band on the costa, and a lesser one near the inner margin. Costal fringe dark, dorsal white. 13 mm.

June.

In Illinois they have bred what I believe to be this species from fungus. Kentucky,

5. D. obscurella Dietz. Similar to D. cristatella, the silvery markings weaker, and absent at base of costa. 11 mm.

Probably this is a variety of the last species. I have only seen one type, which was in poor condition.

July.

Nicholson, Pennsylvania; Kansas.

6. D. chrysoadspersella Dietz. Deep brown; head orange; vertex darker. Fore wing dusted with silvery gray and sprinkled with golden scales; with irregular silvery fasciae about like *D. cristatella*, but the fringe heavily barred with white at hase. Tufts forming heavy transverse bars at a third and two-thirds way out. 8.5 mm.

Plummer's Island, Maryland.

7. D. costisignella Clemens. Head white; antennæ dull tawny; palpi tawny yellow. Fore wing tawny; costa marked with about eight pale bars alternating with dark brown and white, the markings extending into the base of the fringe; one small scale tuft at end of cell. 12 mm.

The moth occurs in damp woods in July.

New York; New Jersey; Pennsylvania. New York: Ithaca (Dietz).

8. D. fasciella Chambers. Golden bronze; tufts placed on two contrasting dark fasciæ, but themselves small, well separated, and easily overlooked; without the brown, black, and white mottling of the last species 12 mm. (*Pitys* Chambers).

New Hampshire; Pennsylvania; New Jersey; Kentucky.

9. D. maculatella Dietz. Head bright ochre yellow; antennæ three-quarters as long as fore wing, fuscous above, and silvery below; thorax powdery; tegulæ dark brown anteriorly. Fore wing dull, powdery gray, tinted with yellow, with irregular dark costal spots and broken fasciæ; tufts small, \mathbf{R}_{*} and \mathbf{R}_{5} stalked. 10-12 mm.

I have never seen this form.

Pennsylvania; California.

10. D. tricingulatella Clemens. Head blackish; face whitish. Antennæ gravish; palpi grav on outer side. Fore wings over six times as long as wide, grav, slightly dusted with fuscous; a dark spot at base of costa; a band a third way out, hardly reaching inner margin; a broad, irregular, brassy-brown band in middle of wing, besides some brassy areas nearer base, and other irregular dark bands outwardly. Fringe checkered black and white, pale outwardly. 10 mm.

This species and the next are easily recognized by their pattern, which is formed of gravish white, blackish, and golden in about equal proportions.

Northern New Jersey and Pennsylvania.

11. D. miscecristatella Chambers. Differs from *D. tricingulatella* only as noted in the key. Fore wing only five times as long as wide; yellow spots normally less developed (*fuscocristatella* Chambers).

Var. auricristatella Chambers is pale, with the head wholly whitish.

I have taken the moth in July.

Kentucky; Worcester, Massachusetts.

12. D. ornatella Dietz. Face yellowish white; vertex and thorax dark brown; antennæ silver gray. Fore wing dark brown with irregular, silvery and golden patches and oblique bars; with raised, brown tufts. Hind wing more gray. 6.5 mm.

This is said to be the smallest Tincid with folded maxillary palpi. It occurs in July.

District of Columbia.

13. D. auriferella Dietz. Similar, the golden and silvery markings on the fore wing less distinct. 8 mm.

District of Columbia.

14. D. marginimaculella Chambers. Fuscous ash-gray; head lighter, dull clay color; antennæ gray. Fore wing neatly and regularly spotted with black; rough looking, without any yellow or golden scales. Costa heavily black-marked at base, with four large, black bars toward the middle and three small ones toward apex. Fold with dark spots at middle and four-fifths way out, a dorsal spot toward apex, and spots a quarter, half, and three-fourths way out on the cell, the latter, and the outer one in the fold, larger. Hind wing gray. 10 mm. (maculimarginella Chambers; Tinea Chambers).

I have seen the type and believe it belongs in Diachorisia. It is certainly no Tinea. It eame from Kentucky.

15. D. (?) heteropalpella Dietz. Silvery white, reticulate with golden brown; with more or less alternating, dark costal and dorsal spots. Head whitish, mixed with brown on vertex; thorax white; front of tegulæ brown. Maxillary palpi long, folded, the first joint as long as the remainder. Eyes small, round; antennæ three-fourths as long as fore wings, with whorls of loose scales like Tinea; fore wing apparently with \mathbf{R}_s and \mathbf{M}_1 stalked; hind wing narrow, costa sinuate. 8 mm.

This species may belong to the Adelidæ (subgenus Bathroxena Meyrick; Pelates Dietz, preoccupied).

Plummer's Island, Maryland.

16. D. (?) afflictella Walker is a plain blackish species, 25 mm. in expanse, which very likely does not belong here (*Tinea* Walker).

11. CHOROPLECA Walsingham and Durrant

(Cyane Chambers, not Felder)

Similar to Tinea (fig. 89, antenna). Fore wing ample (fig. 86), with Cu arising out on the cell not unlike Incurvaria; all veins separate. Hind wing trapezoidal, broad; one vein wanting, all veins separate. 1. C. vesaliella Chambers. Whitish, dusted, striate and mottled with blackish.

1. C. vesaliella Chambers. Whitish, dusted, striate and mottled with blackish. Head dirty white, mixed with brown behind. Antennæ annulate, with groups of darker segments near base, half, three-fourths, and seven-eighths way out. Fore wing with costal area and base dark brown; black in patches, the dark area extending halfway across the wing at its middle as a large spot, and cut by several white bars. Inner margin with a couple of brown spots, and smaller ones on the disc. Apical fringe with white bars, dorsal fringe pale. 8.5-11 mm.

This species looks rather like a Tinea. It flies in June.

Parry Sound, Ontario; south to Florida and Louisiana. "New York" (Dietz).

12. HYBROMA Clemens

(With *Isocorypha* Dietz, in part)

Antennæ rather more than half as long as fore wing; fore wing normal, typically with all veins free (fig. 85) in *H*. (?) chrysocomella (fig. 83) with \mathbf{R}_4 and \mathbf{R}_5 stalked and \mathbf{Cu}_2 lost. (*H. servulella* shows a trace of the basal fork of A.) Hind wing with costa arched, Sc extending well beyond middle, R straight, running to costa, **M** primitive, forking well before middle of wing, \mathbf{M}_{1+2} connected by a transverse vein to R but \mathbf{M}_5 free; \mathbf{M}_1 running to apex. Cu forked, free. In placing chrysocomella in this genus rather than in Isocorypha, I have put

In placing *chrysocomella* in this genus rather than in Isocorypha, I have put more weight on the unique venation of the hind wing than on the fore wing, which agrees with that of *mediostriatella*.

1. H. servulella Clemens. Bright light yellow; antennæ fuscous. Fore wing with costal edge brown from base, widening into a patch beyond middle, which may rarely extend across the wing; a second patch two-thirds way out; basal half of inner margin with an irregular brown patch. Yellow ground when least extended, appearing as four, partly confluent, rounded spots, besides the yellow apex. 11 mm.

June and July.

Cohasset, Massachusetts, to western Pennsylvania, northern New Jersey, and Missouri. New York: Ithaca.

2. H. (?) chrysocomella Dietz. Closely similar to *H. servulella*, but with brown markings more extensive, usually cutting off a rounded yellow spot at the anal angle. Ground rather darker, the markings more purplish. 7 mm.

Kansas and westward.

13. LEUCOMELE Dietz

Similar to typical Hybroma; wings (fig. 81) much narrower, hind wing with sinuate costa, Sc extending less than half the length of the wing, with all three branches of \mathbf{M} connected to \mathbf{R} by a short cross-vein, and base of \mathbf{M} lost. Ovipositor extensile, membranous,

1. L. miriamella Dietz. White; vertex and thorax dark purplish brown; fore wing dusted with blackish, gathering to form streaks along costa and base of inner margin; a couple of bars in fold, and several lesser spots. Apex darker. Fringe checkered at base, outer part whitish with a dark dividing line. 9.5-14 mm.

The types were taken on the trunk of a cherry tree in June. The apical third of the fore wing may be suffused with black except for the barred fringes; and the bars in the cell may fuse into a heavy oblique fascia.

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14. HOMOSTINEA Dietz

Antenna two-thirds as long as fore wings; fore wings (fig. 76) lanceolate, R₆, M₁, and M₂ stalked; M₁ sometimes lost, accessory cell present. Hind wings narrower than fore wings, narrow-lanceolate, cell open below media; costa sinuate; Sc only two-fifths as long as fore wing; one medial lost, and radius and M_2 often barely traceable.

1. H. curvilineella Dietz. Pale yellow dusted with blackish; antennæ blackish; fuscous dusting variable, and sometimes very slight, gathering at base of costa and forming obscure spots and streaks. First discal dot normally represented by a black spot, and the outer one by a curved, oblique streak. 9-12 mm.

The moth flies in the Gulf Strip in June, and northward in July. District of Columbia to Missouri, Kansas, and Texas. New York (?)

15. *ŒNOE* Chambers

Head normal; antennæ three-fourths as long as fore wing. Fore wing lanceolate (fig. 75); R₅, M₁, and M₂ stalked; Cu₂ lost; hind wing linear, with short Sc and simultate costa; M represented by a free, forked or trifid vein; Cu obscurely forked at margin or simple.

1. CE. hybromella Chambers. Head yellowish white, dusted with brown; thorax and basal third of fore wing maroon brown, ending at an erect silvery fascia. Outer part pale green, easily fading to pale yellow, somewhat irregularly dusted with brown. 8-9 mm.

Northward, the moth flies in July, and in the Gulf Strip in June.

Maryland; Pennsylvania; southern Ohio; Mississippi.

16. MEA Busck

(Progona Dietz, not Berg)

Palpi with second joint bushy, clothed with spreading, hair-like scales on its apical half; leaving the upper edge free, third joint rough and blunt. Antennæ rough-scaled, two-thirds as long, or as long, as fore wings. Fore wing lanceolate, Cu_1 lost, M_2 and M_3 stalked, R_5 and M_1 stalked; R_2 arising well back from angle of cell; hind wing with Sc ending two-fifths way out on costa; costa sinuate; cell closed, M_2 and M_3 stalked. Fringe nearly twice as wide as membrane.

The venation is Tineid, but the sexually dimorphic antennæ suggest the Adelidæ. I have had no opportunity to examine the structure fully. 1. M. skinnerella Dietz. Vertex white; palpi and face black. Antennæ fuseous

brown, white toward the base. Thorax white; fore wing white, dusted with brown, shaded with orange toward apex and on fringe; costa irregularly edged with dark brown well toward apex, and with some smaller, ill-defined, brown spots. 7.5 mm. July.

Northern New Jersey.

2. M. bipunctella Dietz. White; shoulders black; dark costal shading forming only two projecting waves.

Florida; Ohio.

17. TRIPTODEMA Dietz

Fore wing with normal venation, all veins free; hind wing lanceolate, the costa not sinuate, with M primitive, three-forked, with no cross-vein between the forks, but one present between M_s and Cu. Fringe nearly twice as wide as membrane. Ovipositor membranous.

This genus apparently is related to Hybroma. 1. T. sepulchrella Dietz. Dark brown, dusted with yellow, the yellow dominant on the thorax. Head russet. Fore wing with yellowish streaks in cell and fold, cut by the dark brown discal spots. A large dark brown spot near base. 8 mm. August.

Plummer's Island, Maryland.

18. XYLESTHIA Clemens

Head wholly rough, palpi with third joint very thin, half as long as second; second joint with a large brush of hair as in Amydria. Antennæ two-thirds as long as fore wings, as in Tinea, with a very heavy pecten. Wings as in Amydria, narrower, with rough scaling and scale tufts.

1. X. pruniramiella Clemens. Dull pale brown, usually with paired paler striæ and the scales largely black-tipped; sometimes nearly immaculate or with a darker shade through the middle. Fringe barred in var. kearfottella Dietz. 12-15 mm. (congeminatella Zeller, clemensella Chambers).

The moth is locally common from May to August. The larva is dirty white, with a brown head and cervical shield. It mines in woody excressences on plum. The cocoon is made of frass and is placed near the mouth of the burrow. April and May.

New Hampshire to Georgia, Kansas and Texas. New York: Ithaca, West Farms.

19. AMYDRIA Clemens

(Myrmecozela Meyrick, in part)

Head rough, though less so than usual in the Tineidæ, with short flattened hair. Antennæ whorled with scales, half as long as fore wings, pecten absent. Tongue and maxillary palpi very small; the latter porrect; labial palpi upturned to vertex with a spherical tuft on the second segment; the third segment about as long, and slender. Venation like that of Tinea.

Almost all the moths are identical in pattern, and are light straw yellow with brownish markings and reticulation; but they differ widely in the male genitalia. The figures show the outline of the valve as shown by denuding.

I. Head rougher, third segment of palpi compressed laterally (Amydria).

1. A. effrenatella Clemens. Fuscous shading usually forming a discal patch extending obliquely down and basad into the fold, a shade in the fold near the



FIGS. 99-100. RIGHT VALVES OF MALE GENITALIA OF SPECIES OF AMYDRIA

99, A. effrenatella; **100**, A. brevipennella (from co-type in the United States National Museum)

base, spots about the outer margin, and a shade along the inner margin. 15-25 mm. The male uncus ends in two long spines, the valve has a moderate spine on the ventral edge (fig. 99).

June and August.

Massachusetts and Parry Sound, Ontario to New Jersey, Missouri and Pennsylvania. New York: Ithaca.

2. A. brevipennella Dietz. Tuft on palpus smaller than in A. effrenatclla; moth usually paler with light reticulation, leaving the marginal spots and discal patch strongly contrasting; sometimes differing from small specimens of A. effrenatella only in structure. 14-15 mm.

The male is easily distinguished by the absence of the spine on the lower edge of the valve. The uncus is only slightly cleft (fig. 100). A blackish variety of this species is known.

Maryland; District of Columbia; Virginia.

II. Head smoother; vestiture of front less prominent; third segment of palpus flattened from front to back (Dysmasia Herrich-Schaffer).

3. A. dyarella Dietz. Gray to dark fuscous brown, inner margin with distincter

markings, fringe with nine dark bars. Dark markings contrasting and clean-cut

on a nearly evenly colored ground, tending to be transverse. 13-15 mm. I have not seen this form, and nothing has been published as to its structure. It was taken at Hazelton, Pennsylvania. It is darker than any of a large group of related southern species.

Pennsylvania; Maryland.

20. SETOMORPHA Zeller

(Semiota, Apotomia Dietz)

Head smoothly scaled, palpi moderate, ascending, flattened, second joint thickened with scales, and with lateral setæ; tongue and maxillary palpi obsolete. Wings rather narrow, ovate-lanceolate. Fore wing (fig. 79) with a more or less hyaline fovea in the accessory cell, which is not wholly separated from the discal cell, the forea distorting the venation in both sexes; \mathbf{R}_a to \mathbf{R}_5 stalked, \mathbf{Cu}_1 and \mathbf{Cu}_2 united in male, stalked in female. Hind wing with costa sinuate, Sc short, \mathbf{M}_1 and \mathbf{M}_2 stalked, M_3 and Cu_1 united in male, separate in female. M_3 and Cu_1 of fore wing stalked in some specimens, in others, free. Fovea with fine spinules on the wing-membrane, which may represent aculeæ.

1. S. insectella Fabricius. Dull luteous, dotted with light gray-brown. 8-20 mm. (operosella, inamœnella, ruderella Zeller; multimaculella Chambers, majorella, sigmoidella, transversestrigella, fractilineella Dietz).

If the species occurs at all in the northeastern States, it is a stray. It is an important pest of stored food in the tropics.

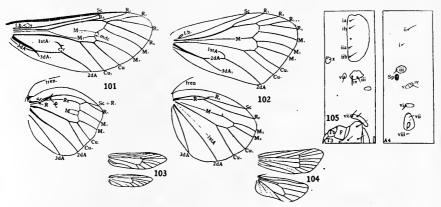
Family 10. **PSYCHIDÆ**

(With Tineidæ, in part)

Mouth parts typically all rudimentary, the palpi usually reduced to hairy tubercles, and the rest lost; a little more developed in Kearfottia. Head with loose, hairy vestiture, that on body variable; ocelli absent, with rare exceptions. Eyes small and retracted, often nearly buried in vestiture; antennæ moderate to very short, in higher forms broadly pectinated, with scaling irregular and confined to the upper side; in the lower forms with the normal two whorls and some bristles.

Abdomen of female ending in a bushy tuft, whose hairs are mixed with the eggs. Legs short, and in higher forms, hairy, the spurs tending to disappear. Typically with more or less translucent or transparent wings. Fore wing with base of **M** quite distinctly preserved and often forked; **Cu** apparently 2- or 3-branched; accessory cell separated from cell by a fine vein, or fused with it as in other Tineoids, \mathbf{R}_{s} always running to outer margin, near apex. At least the middle por-tion of 1st A preserved, often connected to 2d A by a crossvein or running into 2d A; 3d A free at base and quite strong; usually with the tip very distinctly forked, and the upper fork joining 2d A. Hind wing typically with \mathbf{R}_1 full developed, running across to \mathbf{Sc} , often with tip of Sc free, and often with an anastomosis between R and M, farther out; two or three anals, none distinctly forked at

base. Very often a few veins are lost, or there may be secondary veins developed, especially in the costal region of both wings, making the homologies uncertain. The foregoing description applies fully only



FIGS, 101-105. PSYCHIDÆ

101, Thyridopteryx ephemeraformis, male venation; 102, Eurycyttarus confederata, male venation; 103, Solenobia walshella, male venation; 104, Kearfottia albafasciella, venation; 105, Thyridopteryx ephemeraformis, seta map of larva

to the males of the more specialized genera; in the lower ones, the venation is as in the Acrolophid Tineidæ, from which only the tufted abdomen of the female, and small first segment of palpi seem to differentiate them in the adult. The females in the lowest forms are much like the males, except for the abdominal tuft; but in the higher, they gradually become reduced till in the most specialized genera the female is maggot-like, without recognizable structures, and with only the persistent abdominal tuft to show that it belongs to the Lepidoptera. These last forms never leave the pupa shell, but are fertilized within the cocoon by means of the long extensible abdomen of the male, and lay their eggs within the empty pupa shell, shrinking as they lay them. Even the lower forms that have preserved their legs and other appendages usually lay their eggs in or on the cocoon.

Eggs thin, of flat type, laid in a mass mixed with hairs. Larva (fig. 105) with the true legs strong, and the prolegs reduced practically to their hooks, the prolegs all similar, with a single horse-shoe of uniordinal hooks, open on the inner posterior side, even the anals being practically like the others. Head with adfrontals massive, typically not reaching much above top of front, the front never reaching the vertex. Cervical shield large, surrounding the spiracle, which is lengthened horizontally; meso- and metathorax also with dorsal plates. Body with setae i and ii unusually variable in position, normal in lower forms, in higher with ii nearer the mid-dorsal line than i, or even directly above it; iv and v adjacent. All the known larvæ live in a movable case, with a large anterior opening for feeding, and a smaller posterior one through which the excrement is passed, through which the pupa emerges, and, in the higher forms, through which the extensible abdomen of the male is passed to fertilize the female. In the lowest forms the case is flattened and ellipsoidal (very likely so in Kearfottia), in Solenobia tending to be triangular, with a three-valved posterior opening, like that of many Coleophoridæ; in the Psychinæ fusiform, tapering strongly to each end and covered on the outside with bits of foreign matter.

Pupa with second segment of abdomen more or less movable, dorsal headpiece narrow and prothorax unusually wide for a Tineoid; antennæ short, broad in those species whose imagoes have pectinate antennæ; maxillæ rather quadrangular, not covering the labial palpi at all, exactly as in the Hepialidæ; the mandibles forming a distinct separate selerite. Abdomen with each segment armed with an anterior row of spines and a posterior one of bristles; female pupæ of higher forms larviform, intermediate between the larva and grub-like adult in appearance, but brown and chitinized.

The family characters as given above are from the more typical specialized forms; the lower ones have not been fully studied, and show an intergradation to the lowest Tineoids, the pupæ having diffuse areas of spines dorsally. In spite of the wide variation, the European genera, which are many, form almost a perfect series from the lowest to the highest. The position of Kearfottia is doubtful, in the lack of knowledge of the early stages, but it seems related to the Lypusinæ, rather than to the Tineidæ or Yponomeutidæ. In many of its characters it is an exception to the definition of the Psychidæ, but the female has the characteristic, bristly anal tuft.

Subfamily Lypusinæ. Female winged. Palpi distinct; vertex in our species moderately roughened; antennæ practically simple. Fore wing (fig. 104) with \mathbf{R}_s running to outer margin, with all veins preserved, in our species with all free; accessory cell distinct, separated from discal by a fine vein; base of \mathbf{M} in our species not forked. Cubitals rather short, and running squarely across to inner margin; 1st \mathbf{A} free, weak. Hind wing with \mathbf{R}_i appearing merely as a crossvein from Sc to R, cell much shorter on anterior side, the area above the base of \mathbf{M} in our species quite small; with complete venation, or (in Kearfottia) with \mathbf{Cu}_i lost. Anal region a little reduced, though fully veined. Larva in a usually flattened case; normally a scavenger or carnivorous; pupa with cremastral spines dorsal. Kearfottia.

Subfamily **Talaeporiinæ**. Female wingless, but with normal eyes, antennæ, legs, and other appendages; leaving the cocoon for fertilization but laying its eggs within it. Male structure as in *Lypusinæ*, antennæ bristled; the fore wing in Solenobia with a vein lost (fig. 103). Case somewhat flattened, but with triangular valve; the larva feeding on lichens. **Solenobia**.

Subfamily Psychinæ. Female wingless, legless, and naked except for the terminal tuft; never leaving the pupal shell, or only after the eggs are laid; male antennæ pectinate; abdomen extremely extensible, conical when retracted; fore wing with all radials preserved or, rarely, one lost, often with a medial lost (by union of M_a and Cu_1 ?), often with extra veins. Accessory cell not distinct, 1st A con-nected to 2d A by cross vein, or running into 2d A. Hind wing with tip of Sc free, as a spur, the veins toward the margin variable and anastomosing, often with additional spurs, normal in Chalia; M_3 stalked with M_2 or lost. Larva of northeastern species in a fusiform case; with true legs shorter and stouter than usual, with ii higher than i on abdomen. Pupa with cremastral spines subventral (save in Chalia rilevi).

Male with transparent wings, hind wing subtriangular and small; fore wing more than twice as wide (fig. 101); abdomen conical. Larva with tubercle ii directly over i (fig. 105). Thyridopteryx. Male with opaque, smoky wings, broad and ample, the abdomen hardly exceeding them (fig. 102); larva with tubercle ii on the annulet behind i.

Eurvevttarus.

Male with translucent smoky, hairy wings, intermediate in width, the hind wing not lobed and relatively small. Fore wing with two veins lost, the rest free; 1st A curving down into 2d A; hind wing with seven veins, all free; the cell short in front. Chalia.

1. KEARFOTTIA Fernald

Front smooth, vertex with more or less rough scaling, leaning forward; much as in the Ecophoridæ; palpi oblique, reaching middle of front, second segment with long, loose hair-scales, third porrect; tongue minute; maxillary palpi scaly; no ocelli; antennæ rather less than half as long as fore wing, heavily ciliate, the cilia as long as the segments. Fore wing (fig. 104) elliptical, three times as long as wide; with complete venation, the veins from \mathbf{R}_2 to \mathbf{Cu}_1 nearly equidistant, Cu_2 a little more widely spaced, R_1 arising a third of the way out. Hind wing about half as wide, nearly semicircular, Cu, arising two-thirds way out on the cell, anals all present, 2d Å forked at base, sinuate, Cu_1 missing, mdcv unusually long and oblique, the simple base of M continued as M_{1+2} ; udcv short. transverse. Female similar with a heavy body and large terminal tuft. Larva unknown.

The genus would fit almost as well in the Tineidæ (between the two subfamilies) or in the Yponomeutidæ (in the broad sense) as here, but the resemblance to Narvcia and Diplodoma suggest a position near the foot of the Psychid series.

1. K. albafasciella Fernald. Head, thorax, and basal and apical thirds of fore wing deep brown; middle third cream white, usually with four dark spots along costal, and three along dorsal edge. 38 mm., 9 12 mm.

July.

Maryland; southern Ohio; Missouri (?).

2. SOLENOBIA Zeller

Characters of the subfamily. No ocelli, \mathbf{M}_1 (?) lost (fig. 103), \mathbf{M}_2 and \mathbf{M}_3 stalked or connate; hind wing with \mathbf{M}_2 and \mathbf{M}_3 stalked. A small area of aculeæ near the base of cell. Female with a minute lanceolate rudiment of a wing.

The pupa is similar to that of Tinea, but the rows of spines are triple, rather than single; and the maxillæ are very short and widely separated, but have distinct maxillary palpi, separated by a suture. The antennæ are also shorter than in Tinea.

1. S. walshella Clemens. Male smoky; fore wing more or less contrastingly mottled and dusted with whitish; translucent. 12 mm. Female smoky black, rough, with sparse hair-scales.

The larva feeds on lichens on trunks of trees. The moth is not rare but is easily overlooked.

New York and south. New York: Ithaca, Bronxville (Woodruff).

3. CHALIA Moore

Antennæ short, broadly pectinate; abdomen conical and hairy, with large genitalia exposed at the end. Fore wing triangular, with costa rather straight toward base; apex rounded over, and inner and outer margins straight. Two anterior radials lost; \mathbf{R}_i and \mathbf{R}_b connate or shortly stalked, forking over the apex, M2 and M3 connate, Cu1 and Cu2 both arising from lower margin of cell. 3d A running into 2d A. Hind wing two-thirds as long as fore wing, but of only half the area. Sc free; cell narrow in front; all the veins widely spaced, and one medial lost.

The pupa has widely separated, quadrangular maxillæ and distinct mandibles. as in the Hepialidæ. The cremastral spines are nearly terminal but above the anus; the wings of the male less than half the length of the body, obsolete in the female. The abdominal segments have two toothed ridges. The case is nearly cylindrical but somewhat fusiform. It is 11 mm. long, and is covered with fine, dust-like material (lichens ?).

This description is drawn up from C. rileyi, which may prove to be distinct from the genus Chalia. It is more primitive than our other Psychinæ. 1. C. rileyi Heylaerts. Translucent smoky, without markings. 10-12 mm. The larva is flesh color; its thorax yellow, striate with dark brown, and shin-

ing. The moth emerges in September.

Missouri; Jefferson County, West Virginia.

4. EURYCYTTARUS Hampson

(*Psyche*, in part)

Male antennæ very broadly plumose; tibial epiphysis quite small; one medial

lost in both wings (fig. 102).
l. E. confederata Grote and Robinson. Smoky, the wings thinly but evenly scaled. 15 mm. (H 41:8; 1:16, larval case.)

The larvæ feed on low plants, elimbing up on the trunks of trees to pupate. in the early spring. The case is fusiform; less than 25 mm. long. It is covered with pieces of leaf, and so forth, running lengthwise, and, usually more than half as long as the case. New York to Colorado and south. New York: Ithaca, New Baltimore, Staten

Island; Newtown, Long Island.

5. THYRIDOPTERYX Stephens

Male antennæ broadly but stiffly pectinate, with simple, serrate apex. Hind wing rounded-triangular (fig. 101), extended at anal angle; wings transparent, except costæ and inner margin of hind wing.

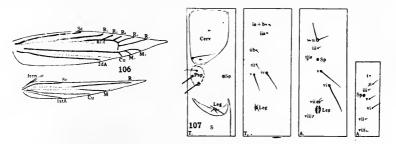
1. T. ephemeræformis Haworth. Transparent and smoky. 25 mm. (H 41:12.) The larva feeds on various trees and shrubs, even arbor vitæ and other evergreens, and is sometimes injurious. The moths emerge in late fall, and the eggs hibernate. The case of the full-grown female may be nearly 50 mm. long; that of the male is always much smaller.

Connecticut to Florida. New York: Geneva (cases), Fort Edward, New Balti-more, Mt. Vernon, New York City, Staten Island; general on Long Island. Not hardy at Ithaca, but a pest on Long Island.

Family 11. TISCHERIIDÆ

(Tineidæ, in part)

Vertex with a rather smooth, spreading, flat tuft covering base of antennæ; front smooth-scaled, high, tapering strongly below. Ocelli absent; maxillary palpi obsolete; tongue weak, scaled; labial palpi smallish to minute, somewhat roughly scaled, but without bristles, not upturned. Antennæ with scape smallish, a third the width of the eye, with a small, modified, scaly pecten. Segments of shaft slender, and eiliate below, with outer whorl of scales perceptibly raised. Eyes large and naked. Hind tibiæ hairy, the upper spurs well toward base. Fore wing lanceolate (fig. 106) more or less caudate, with large discal and accessory cells and all veins arising from them short. All radials present; \mathbf{R}_1 arising before the middle of the cell; \mathbf{R}_5 running to costa; one or more dorsal veins lost, and those present very short; the last cubital running directly across to inner margin. Hind wing normally



FIGS. 106-107. TISCHERIIDÆ 106, Tischeria, venation; 107, Tischeria malifoliella, seta map of larva

hardly half as wide as fore wing, in a few males about as wide, pointed; female frenulum of 2 bristles. Venation reduced and cell open.

Larva (fig. 107) less specialized than the Lyonetiidæ and Opostegidæ, strongly flattened; epicrania with an obscure lateral keel, with the normal setæ preserved, the epicrania extending a third their width behind the vertex. Hypostoma well developed, with divergent lateral sutures. Labrum with well developed pilifers; front reaching vertex, but sometimes narrowed to a point behind. Ocelli preserved, equal in size. Mouth parts small, normal. Mandibles of the biting type, the larva eating the parenchyma. Thoracic legs wanting. Abdominal segments slightly lobed laterally; prolegs with transverse bands of rudimentary hooks.

The larva is a leaf-miner, usually making a large blotch. Sometimes the feeding is all on one side, leaving the beginning of the mine

as a projecting point (trumpet-mine). The frass is ejected through a small slit in the leaf.

Pupa incomplete, of a low type, with segment 3, and possibly segment 2 free; fairly well chitinized. Setæ sometimes very heavy, i and ii approximate on the movable segments. Dorsal spines fine, in one patch, or in two patches of spines of unequal size; sometimes with a row of larger posterior spines (oak species). Cremaster of two heavy, upeurved hooks. Head with large dorsal piece; prothorax small; maxil-læ long, nearly covering palpi; no maxillary palpi.

There appears to be only a single genus north of Florida. It is rather like the Tineidæ but the early stages are specialized for leafmining, and at the same time have some primitive characters.

1. TISCHERIA Zeller

(With *Coptotriche* Walsingham)

Characters of the family. In many species, the under side of the fore wing is clothed with many fine spinules that may represent aculeæ, but they are too coarse, and are more likely to be modified scales. Various sexual modifications of the wing are common, the most extreme form being that of T. zelleriella, in which the costa of the hind wing is notched, and the usual fringe replaced at the notch by fine, short bristles.

Key to the species

1. Nearly even dark grav.	
2. Scales evenly colored and quite purplish	nea.
2. Scales merely shining fuscous, with paler tips; a smaller species.	
1. malifoliella, 3. rosetic	cola.
1. Fore wing powdered with blackish or with bands of powdering.	
2. Scattered small areas of black dusting	ella.
2. Two oblique transverse fasciæ	
2. Heavily dusted all over	
1. Fore wing yellow, often broadly shaded with darker yellow, or with a	verv
little brown dusting; larvæ on oak and chestnut.	•
2. Hind wing of male as wide as fore wing.	
3. Male with apical fringe notched16. zelleri	ella.
3. Male with fringe normal.	
4. Hind wing regularly lanceolate	irea.
4. Hind wing widened at base, abruptly attenuate just beyond middle.	
14. clemens	ella.
2. Hind wing of male half as wide as fore wing.	
3. Male with a fuscous patch near base of fore wing below, and less man	rked
ones on both sides of hind wing	ella.
3. Male without such patches.	
4. A patch of fuscous scales at anal angle of fore wing above.	
5. Vertex deep brown; fore wing pale lemon yellow, apex reddish	or
brown, sometimes dusted with black11. badii	
5. Vertex concolorous; fore wing dull ochreous10. tinctori	ella.
4. No such patch.	
5. Abdomen densely dusted beneath with fuscous brown on a pale ye	llow
ground \dots 9. castaneæ	ella.
5. Abdomen not dark dusted.	

6. Fore wing dull ochre.....8. concolor. 6. Fore wing reddish yellow, margined with purplish fuscous.

7. fuscomarginella. 6. Fore wing pale straw yellow, with dark-dusted spots.

13. albostraminea.

1. T. malifoliella Clemens. Mouse gray, a little shining, the scale-tips showing some golden iridescence. 6 mm.

Larva on apple, starting in a linear mine, which is gradually widened into a large blotch; but is not tentiform until the formation of the cocoon, when a fold is made, to contain the pupa. Larva in August; moth in May.

This moth, the "trumpet-miner", is generally distributed. New York: Menands (Albany Co.), Schenectady, East Greenbush. The larval work is seen throughout the State, but I have seen no records.

2. T. aënea Frey and Boll. Similar to T. malifoliella, rather larger, distinctly brighter and more purplish, with strong bronzy iridescence and purple apex; the

hind wing rather more nearly concolorous. 7 mm. The larva occurs on Rubus. The mine is generally straight, and runs to the edge of the leaf, usually between two veins. The mine is crumpled in parallel folds, much like that of the oak-feeding species and Lithocolletis.

Massachusetts; Ohio; Pennsylvania; Texas. 3. **T. roseticola** Frey and Boll. Indistinguishable from *malifoliella* in the adult stage, but usually a little smaller, duller, with broader wings, slightly more roughly scaled toward the apex, and with a yellower face.

The larva is a trumpet miner on rose.

4. T. ambrosiæella Chambers. Head powdery dull brown; fore wing luteous, heavily dusted with blackish, showing traces of the transverse bands of T. heliopsisella, the dark markings leaving two or three lightly dusted and more distinctly yellow areas toward the outer margin. Fringe mouse gray with some black-tipped scales. Hind wing mouse gray. 6 mm.

The larva mines Ambrosia trifida and does not form a nidus.
Kentucky; Ohio; Missouri.
T. heliopsisella Chambers. Deep ochre yellow. Fore wing with two oblique gray fascia from a third way out on costa to middle of inner margin and from beyond middle of costa to three-fourths way out on inner margin; a costal streak from the latter to the apex, and a streak in base of fold; all the black dusting a white ground. 8 mm. (nolckenii Frey and Boll).

The larva occurs on Heliopsis and Ambrosia, in August. It spins a circular white nidus for shelter within the mine, and pupates in it.

Southern Ohio; Kentucky; California.

6. T. solidaginifoliella Clemens. Cream color or light straw yellow; shaded with bright yellow, becoming yellow-brown in the costal fringe. Head solid yellow; fore wing with scattered spots of black dusting. 7 mm.

August. Larva in a flat, white blotch on upper side of leaves of Solidago. Pennsylvania to Texas. New York: Albany. 7. T. fuscomarginella Chambers. The larva is a miner on the under side of oak leaves.

Kentucky.

8. T. concolor Zeller. This species is known only from Texas. The mine is at the edge of a leaf of oak.

9. T. castaneæella Chambers. Pale yellow, all the margins of the fore wing strongly shaded and dusted with yellow brown; the under side of the abdomen contrastingly dusted with dark brown except at the apex. 8 mm.

The larva forms a narrow blotch mine along the edge of a leaf of chestnut. I have never seen a male.

Virginia; Kentucky.

10. T. tinctoriella Chambers. Dull ochre, the margins noticeably dusted with dark-brown, darker than badiiclla. Fore wing of male gray-scaled below, except at the apex and below the fold. (quercitella Clemens).

Larva in a blotch-mine on the upper side of an oak leaf, the mine extending out into lobes and marked with zigzag purple lines. Kentucky; Missouri. New York: Albany (New York State Museum).

11. T. badiiella Chambers. Light straw yellow, with contrasting, dark-brown vertex; apex of fore wing usually dusted with black (var. pruinosella Chambers) typically with the two dots only; fore wing more or less tawny-shaded from anal angle to apex. A small dark spot two-thirds way out on costa as well as the one on the inner margin. (citrinipennella Walsingham, not Clemens; purinosella, pruinosella Chambers).

The larval mine is a crumpled blotch on oak like that of T. zelleriella. The moth occurs in August and in March.

Quebec to Texas.

12. T. citrinipennella Clemens. Yellow; head and thorax yellow-brown; costa and apical third shaded with yellow-brown; fringes much paler and duller. Hind wing whitish, becoming umber brown at base. Abdomen dusted with brownish yellow below, but much paler than in T. castaneeella. 8 mm. (quercivorella Chambers; fuscomarginella Walsingham, not Chambers.)

Mine on oak; crumpled like that of T. badiiella. Moth in August.

Pennsylvania; Ohio; Missouri. New York: Crosby (Yates Co.), Ithaca.

13. T. albostraminea Walsingham. Pale straw color, not shaded with ochre; with blackish-dusted spots two-thirds way out on costa, five-sixths way out on inner margin, and over the apex, the latter the strongest. Apical fringe deep ochre. Underside pale with deep-ochre apical fringe. 5 mm.

The larva forms a small blotch on white oak. The moth emerges in August. It is our smallest species.

District of Columbia; Kentucky. New York: (type).

14. T. clemensella Chambers. Yellow, costal edge and some dusting toward apex below browner. Basal half of fore wing, below, with yellow sex-scaling (bicolor Frey and Boll).

Texas.

15. T. sulphurea Frey and Boll. Texas. This moth is unknown to me. 16. T. zelleriella Clemens. Light ochre, with a deep yellow shade in fringe over apex, costal fringe of hind wing brown, dorsal pale. Hind wing whitish in the male, gray in the female. Under side of fore wing with long, stiff hair, extending obliquely up from near inner margin, nearly across to costa, and from base threefifths length of wing; cell clothed with dense fine scales. 9 mm. (Coptotriche Walsingham; complanoides Frey and Boll; latipennella Chambers).

The larva makes a trumpet-mine on oak, and is unique in Tischeria in leaving its frass within the mine in a series of curved lines. The moth occurs generally in March and April.

Distribution general.

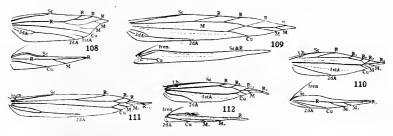
New York. (Henry Edwards.)

Family 12. LYONETIIDÆ

(Tineidæ, in part)

Vertex either hairy or scaled; face smoothly scaled, strongly oblique; tongue weak and naked or obsolete; maxillary palpi usually obsolete; more rarely quite short, straight, and drooping; labial palpi short and scaled, drooping or obsolete, best developed in Phyllocnistis, which only

doubtfully belongs to this family. Ocelli absent. Antennæ with scape enlarged and flattened; with eye-cap except in a couple of species of Phyllocnistis. Shaft with two nearly equal whorls of scales to a segment. Hind tibiæ with long bristly hair above, becoming a row of strong bristles in Phyllocnistis; upper spurs attached well above the middle. Fore wing lanceolate, often caudate (with a slender prolongation of the apical membrane), \mathbf{R}_5 running to costa; radials often reduced to three; \mathbf{R}_1 leaving cell well toward base of wing, except in



FIGS. 108-112. LYONETIIDÆ

108, Paraleucoptera albella, venation (after Heinrich); 109, Bedellia somnulentella, venation; 110, Bucculatrix, venation; 111, Lyonetia, venation of fore wing; 112, Phyllocnistis, venation

Phyllocnistis, or lost. Not more than four dorsal veins. Vein 1st A weak, free; 2d A variable, even within the genus Cemiostoma. Hind wing linear or lanceolate, its fringe much wider than itself, the membrane abruptly narrowed near or before middle of costa. Sc short, running close to costal edge, or even fused with it; sharply divergent from the R-stem from the base as in the Elachistoidea; R-stem running to apex, with one or two medials given off from its lower side; Cu separate, simple except in Philonome; anal region extremely reduced, often without distinct veins.

Egg flat. Larva with both true legs and prolegs present; 16 legs, except in Phyllocnistis. Body nearly cylindrical. Head much flattened in the normal series, with strong lateral keels and reduced mouth parts. Ocelli, six, in two groups; some setæ lost. Front quadrangular; hypostoma as in the Tischeriidæ. Setæ iv and v on abdomen remote, i nearer the middle line than ii; setæ of prothoracic wart widely separated; setæ of prolegs uniordinal in a complete ellipse; rarely with traces of a second row.

Pupa with all appendages soldered together and no free segments; labial and maxillary palpi not visible; labrum a separate sclerite; appendages nearly as long as the body; antennæ longer than wings; prothorax narrow, especially on the middle line, the dorsal head piece wide. Spiracles small, circular and slightly projecting. Dorsum of abdomen not spinulated.

The larvæ and pupæ of Bucculatrix and Phyllocnistis are aberrant and are discussed under their genera.

The family is somewhat heterogeneous and has been divided by some into four or five, in fact one for each really well-marked genus. Bucculatrix is quite isolated, but its image agrees with the characters of the family, while Phylloenistis is practically half way between the Lyonetiidæ and the Graeilariidæ, showing a mixture of the characters. of both in all stages. European workers tend to separate a Lyonetia group from a Cemiostoma (Proleucoptera) group, but there are North American genera which connect the two so well that subfamilies can hardly be made.

Key to the genera

1. Vertex rough, bristly, contrasting with lower face.

2. Eye-cap edged below with a row of strong bristles; cell entirely above middle of wing; wing with a scale-tuft at middle of dorsal margin.

3. Front running down far below eyes in a blunt point......7. Bucculatrix.

3. Four veins running from cell to inner margin.

4. Hind wing with **R**, **M**, and **Cu** represented by 5 veins....1. Corythophora. 4. Hind wing with **R**, **M**, and **Cu** represented by four veins..2. Proleucoptera.

3. Three veins from cell to inner margin (figs. 109, 111). 4. Vertex with a small, fine tuft.....4. Lyonetia.¹³

1. Entire head smooth.

2. Palpi moderate; tongue obsolete; eye-cap tending to disappear; fore wing with lanceolate discal cell and no accessory cell (fig. 112)...6. Phyllocnistis. 2. Palpi minute; tongue present in our species; accessory cell present, but open

outward so that \mathbf{R}_2 and \mathbf{R}_3 are stalked together; hind wing broader, lance-

1. CORYTHOPHORA Braun

Face smooth, with an erect tuft between antennæ; antennæ four-fifths as long as fore wing; eye-cap pointed anteriorly below; palpi moderate, smooth, drooping; maxillary palpi rudimentary. Fore metatarsi thickened with scales; hind this hairy. Fore wing sublance later, \mathbf{R}_1 absent, \mathbf{R}_2 and \mathbf{R}_3 as in Cenniostoma, cell with four posterior veins; tip of 1st A distinct, 2d A simple; hind wing lanceolate, two-thirds as wide as fore wing; veins obsolescent at base; cell open both above and below M; Cu forked; anal region reduced.

1. C. aurea Braun. Golden yellow; head and appendages very pale, projecting point of eye-cap rather darker; thorax nearly white, with golden tegulæ; fore wing becoming deep orange at apex; middle of costal and dorsal margins fading to white; fore tibiæ and tarsi dark brown externally. 9-9.5 mm.

July.

Southern Ohio; Balsam, North Carolina.

¹⁸ The Gracilariid genus Leucanthiza is likely to run to Lyonetia but is distinguished by its less perfec t eye-cap, moderately developed palpi. and orange coloring.

2. PROLEUCOPTERA Busek

(Leucoptera; Cemiostoma, in part)

Face smooth; vertex with a small fine tuft; antennæ four-fifths as long as fore wing; eye-cap good-sized, rounded, with regularly imbricated scaling, less perfect than in Opostega. Palpi small, drooping; tongue weak but distinct. Fore wing broad, with lanceolate, caudate membrane, with 10 veins, all separate, 2d A forked at base, M_3 and R_4 absent; hind wing narrow, without cell, with M_s and Cu_1 absent; fringe four times as wide as membrane.

This genus is very closely related to Leucoptera (Cemiostoma) but is slightly 'more primitive.

1. P. smilaciella Busck. Similar to C. albella; larger; first fascia much narrower, three times as long as wide, strongly oblique, and starting nearer the base; the second fascia a mixture of yellow and white, ending outwardly in four dark lines that converge on the apex; lead-colored spot smaller than in C. albella, and completely surrounded before and above with the yellow band.

The moth has been obtained from June to September. The mine is a large, dirty upper-side blotch on Smilax, beginning as a line. The cocoon is formed on the leaf, under two bands of silk.

Maryland; District of Columbia; southern Ohio; Pennsylvania.

3. LEUCOPTERA Hübner

(*Cemiostoma* Zeller)

Head smooth, with the usual large, vertical scales on occiput only. Eye-cap well developed, apparently continuous with the head vestiture, when closed completely covering eyes; mouth parts obsolete. Hind tibiæ with bristly hair; fore wing oblong-lanceolate, caudate; \mathbf{R}_1 weak or absent. Only three veins running from cell to hind margin, \mathbf{M}_{1+2} , $\mathbf{M}_{s}+\mathbf{Cu}_1$, and \mathbf{Cu}_2 ; hind wing linear-lanceolate.

In the American species (subgenus Paraleucoptera Heinrich,) \mathbf{R}_2 and \mathbf{R}_3 are stalked, \mathbf{R}_5 stalked with \mathbf{M} , but very short, and $\mathbf{2d} \ \mathbf{A}$ is forked at the base in the fore wing; the hind wing has only one medial. Typically, \mathbf{R}_3 and \mathbf{R}_5 are lost, and $\mathbf{2d} \ \mathbf{A}$ is simple, but there are two medials in the hind wing.

1. L. albefla Chambers. Sometimes with a few hairs to represent vertical tuft. Snow white; antennæ pale fuscous with white eye-cap and apex; fore wing with a nearly square golden fascia from costa beyond middle, pointing toward anal angle; a larger spot on costa beyond it, reaching apex, sometimes broken into fasciæ, both of which are edged with brown; and a silvery gray spot near anal angle, preceded and followed with black spots; with a yellow line before it, preceded by fuscous-tipped scales. Fringe fuscous at apex and anal angle, paler between. 6 mm.

Larva flat, lobed at sides, with pro- and mesothorax widest, then a couple of relatively narrow segments, and abdomen wider again, with minute but normal legs.

It lives, often socially, in a very dirty frass-filled mine, on poplar and willow. The cocoon is like that of *P. smilaciella*.

Kentucky; Colorado and west.

WILLIAM T. M. FORBES

4. LYONETIA Hübner

Vertex with bristling hair, intermediate between Proleucoptera and Bedellia, and Bucculatrix. Antennæ with a strongly developed eye-cap, about as long as fore wings; palpi longer than eye, drooping, divergent; maxillary palpi minute; tongue obsolete. Hind tibiæ with long, sparse hair. Fore wing narrow-hanceolate (fig. 111), caudate, with \mathbf{R}_1 arising from near middle of cell, cell narrow, without accessory cell; with 4 to 6 veins arising from near its tip; 2d A strongly forked at base. Hind wing linear, with \mathbf{M}_1 and \mathbf{M}_2 arising out of \mathbf{R}, \mathbf{M}_3 lost, and Cu simple. The larva normally makes a linear mine. Spuler puts Lyonetia as a separate

family from the preceding and following genera; but its characters do not scem very important. The imago hibernates under bark.

Key to the species

1. Light brown with longitudinal white streak	saliciella.
1. Ground white.	
2. No distinct yellow area at apex	l. speculella.

1. L. speculella Clemens. Shining white; antennæ dark brown above, more or less annulate, paler below; palpi fuscous outwardly; hind wings and fringe, also a narrow band on hind edge of fore wings, mouse gray, bronzed with purple. A brown streak from base along costal edge to beginning of fringe, typically very weak; the streak then running obliquely to inner margin, enclosing a white spot at inner margin. Another longitudinal bent streak in the middle of wing, meeting the transverse part of the first; beyond this an orange-yellow spot. A black apical dot. Fringe with three bars on costal side and one below, or sometimes with five costal and three dorsal streaks. Typically with blackish bars at middle and three-quarters way out, below fold. The type form is fully marked, but var. gracilella Chambers more lightly. 7 mm. (gracilella, apicistrigella Chambers, midificansella Packard).

The moth hibernates under bark. The larva occurs on grape, and, when common, on many other trees and shrubs. The mine at first is linear, with the frass left in it; then it abruptly changes to a blotch, and the frass is then ejected through a hole. The cocoon is slung like a hammock on the back of a leaf. I have seen a specimen of the moth suffused with gray.

The distribution is probably general; I have seen it from Pennsylvania to British-Columbia.

2. L. latistrigella Walsingham. Similar; fore wing with a faseia running from middle of inner margin to three-fourths way out on costa, followed by a ferruginous patch; without the costal streak of L. speculella; apical black dot preceded by white scales. 9 mm.

Larva in soft young leaves of *Rhododendron maximum*; mine at first linear, 3 cm. long and black, then abruptly widening but still linear for 3 cm., then a brownish blotch about 5 mm. by 4 mm. Pupa naked, suspended by a few silk threads to a bent leaf.

Atlantic States (?) (locality not stated).

L. saliciella is known definitely only from the west but is to be expected in our territory; its larva feeds on willow.

5. BEDELLIA Stainton

Vertex and upper part of face with a large tuft of hair, smooth below; no ocelli; antenna with an extremely large, massive pecten, practically an eye-cap; tongue naked, weak; palpi small, hanging, appearing as if socketed in the face, as in the Heliodines group; maxillary palpi absent. Hind tibiæ with bristly hair. Fore wing narrow-lanecolate (fig. 109), not decidedly caudate, cell narrow, in middle of wing; four veins running from cell to costa and three (two in the Floridian B. minor) to inner margin; A forked at base. Hind wing nearly linear, with reduced obscure venation, and with very broad fringe.

The larva lives in a light web on the under side of the leaf of Ipomea. I have not seen an adequate account of its structure, but it is less specialized than in the Proleucoptera group.

1. B. somnulentella Zeller. Dull light gray, sparsely and irregularly dusted with fuseous. 11 mm. (staintonella Clemens).

The moth has two broods, one cecurring in August, and the second from mid-

September to spring. The larva may be found from July to September. This species is of general distribution in the northeast, and also occurs in Europe. New York: New York City (Lintner).

6. PHYLLOCNISTIS Zeller

Eve-cap variable in size, absent in P. insignis; no pecten; eves small. Head smooth, palpi rather well developed; hind tibiæ with regular rows of long bristles. Fore wing caudate (fig. 112), with smallish cell and no accessory cell; $\mathbf{\hat{R}}_1$ arising well beyond middle of wing, R5 running to apex; in some species all radials present; A short, not forked.

Feeding larva of the flat, Gracilariid type, with similar, very thin, blade-like mandibles and with hardly recognizable free portions of labium and maxillæ, but entirely without legs. Last-stage larva cylindrical; with rudiments of legs; with head practically reduced to a spinneret; not feeding. Mine serpentine, white or pale green, often shining (snail-track mine) with or without central frass-line; parenchyma not eaten, but only the sap. Pupa within the mine, in a partly folded edge of the mine; of Gracilariid, rather than Lyonetid type; rather heavily chitinized; antennæ and hind legs extending well beyond end of wings; segments 4 to 6 of abdomen movable and 7 in male, 3 fixed; segments 3 to 7 with dorsal pits near the anterior margin, and heavily chitinized setæ; segments 8 to 10 combined, shorter than 7.

The moth occurs in May, July, September and October.

The genus is as near the Gracilariidæ as the Lyonetiidæ, but is a little less out of place in this already rather heterogeneous family. Several entomologists make it the type of a separate family, Phyllocnistidæ. Smilacisella, once put in this genus, is a Marmara.

Key to the species

1. Ground color orange, larva on Compositæ1. insignis.
1. Ground color white.
2. A longitudinal golden streak from base to middle, connecting with the first
dorsal fascia; larva on Magnoliaceæ.
3. Streak partly edged with black
3. Streak not at all edged with black
2. No such streak.
3. Larva on sweet gum
3. Larva on poplar4. populiella.

3. Larva on grape or Ampelopsis.

- 4. Middle of apical region white or golden.

1. P. insignis Frey and Boll. Head and thorax lead-colored; fore wing orange; inner margin gray to middle, the gray area almost reaching costa at base, costal edge, black, expanding into a black-edged, lead-colored triangle at middle; similar costal and dorsal lead-gray spots beyond middle; fringe black-barred. Sometimes with a little white in the costal black streak.

Larva on Erechtites. Moth in July and September.

Southern Ohio; Missouri; Kentucky.

2. P. liriodendrella Clemens. Similar to populiella. Apex broadly yellow; with a broad, yellow, longitudinal streak through middle of wing, joined by the tip of the oblique, postmedial streak; all markings finely edged with brown. 6 mm.

The larva makes a convoluted tract-mine, with a central frass-line, in the smaller terminal leaves of the tulip tree, in July.

3. P. magnoliella Chambers. Similar to P. liriodendrella; the brown edgings weaker, and absent from the basal streak.

Larva like that of the last species on various Magnolias.

4. P. populiella Chambers. White. Antennæ pale yellowish; eye-cap white; fore wing somewhat silvery; a dark gray, fine and broken, excurved postmedial fascia. with three slight dark bars in the fringe between it and apex. A strong black apical dot, from which extend two or three dark lines, in apical fringe; also an oblique streak before it in dorsal fringe; a slight, yellow shade before first fascia. and at apex. 6 mm.

The larva forms a broad mine of the shining, snail-track type, but with distinct central lines of frass. It feeds on aspen and other poplars.

Apparently common and general in distribution, but rare in collections. New York: Hemlock Lake (bred).

5. P. vitifoliella Chambers. Similar to P. populiella; generally with black markings rather more distinct, the second fascia usually extending onto the wing membrane, and yellow areas rather less distinct. 41/2 mm.

Mine on grape, like that of P. populiella, with quite distinct frass line.

Generally distributed; the commonest of the grape Phyllocnistises. New York (Bentenmuller).

6. P. vitigenella Clemens. Imago often indistinguishable from the other white species, but usually with a gray, antemedial shade on inner margin and with heavier and more extensive black markings; the second fascia, however, in the fringe only.

The mine is a narrow, almost invisible snail-track, without a frass-line; sometimes it is formed on Ampelopsis, and then is much contorted. 7. P. ampelopsiella Chambers. Similar to P. populiella and ritigenella; with

bars heavier, and the postmedial one distinctly double; apical region with a goodsized gray area on dorsal margin, in which the costal dark streaks often end; and, in some specimens, with a distinct, longitudinal, gray streak on basal half. and gray antemedial patch. Yellow shading distinct.

Mine a narrow and much contorted tract, often fusing into a blotch; ou Ampelopsis.

8. P. liquidambarisella Chambers. Moth like *populiella*; slightly variable: the markings typically reduced to a minimum, with hardly any yellow. Mine of snail-track type without central frass-line; on sweet-gum.

7. BUCCULATRIX Zeller

(The ribbed-cocoon makers)

Head rough on vertex; front smooth, extending far below the eyes in a point. Antennæ not long, with a well-developed eye-eap, fringed with stiff hairs; labial palpi small, hanging; maxillary palpi minute; tongue very weak, naked, distinctest in the *nixeella* group, which also spin atypical cocoons. Fore wing lanceolate (fig. 110) with cell'mostly in upper half of wing. Cu running nearly through the eenter of the wing; with all radials; \mathbf{R}_1 arising near base, oblique; the others running directly aeross to costa; A not forked. Hind wing also lanceolate, with broad fringe; with veins more distinct than in Bedellia, **R** running to apex; two medials preserved; Cu simple.

The larva at first forms a serpentine mine with a central frass-line, much like that of Nepticula; then leaves the mine, and skeletonizes the leaves, leaving one epidermis uneaten; it then lives exposed on the leaf.

Larva with head normal, the front extending about two-thirds way to vertex. Body cylindrical, stout, green; set normal, iv and v distant; i farther from middle line than ii, even on segments 8 and 9 of abdomen; prolegs strong, with two transverse hands of hooks (the anal pair as usual with a single row). True legs normal. Pupa in a silk coccon, as a rule attached longitudinally to a twig; abdomen dorsally spinulated, with third segment free; a stronger row of spines on anterior edge of each segment. Labial palpi covered, hind legs much longer than antennæ; abdomen ending in a pair of laterally directed angles or spines.

The species are closely similar, and numerous, and may not always run correctly in the key; I have been obliged to omit a couple from lack of material. The cocoon is almost always spun in a characteristic way, so as to form longitudinal ridges with the silk threads in a regular pattern between them; in two or three species it is simply oval, with irregularly arranged silk. Bucculatrix is an aberrant genus, apparently nearest to the Lyonetiidæ, but with the larva modified for external life.

Key to the species

- 1. Fore wing, head, and eye-cap dominantly white or cream yellow.
 - 2. Cream yellow, with vague shaded markings; no black; very small, 4 mm.

1. luteella.

- 3. With more extensive markings.

 - 4. Thorax and ground color of wings pure white (the thorax rarely
 - tinted with straw yellow). 5. Expanse 8-12 mm., markings defined, pale yellow.
 - 6. Three costal streaks besides the brown apical shading...4. montana.
 - 6. Two costal streaks besides the brown apical shading.
 - 7. First costal streak most distinctly connected to anal angle.

2. magnella. 7. First costal streak turning longitudinally in middle of wing,

6. Markings clean-cut, gray-brown; apical marks broken up.

- 7. Seales in outer transverse band all more or less bicolored;
 - 7. Scales in outer band black-tipped toward costa but evenly
- colored toward inner margin.....10. capitcalbella.

1. Fore wing dominantly brown or ochre yellow.

- 2. A white or silver basal dash.
 - 3. Dash extending across thorax; ground brilliant yellow-brown.
 - (Philonome elemensella). 3. Dash not extending across thorax; ground often dark brown.
 - 21. erescentella.
 - 2. No defined basal dash; rarely with a pale longitudinal shade. 3. Light yellow-brown dominant.
 - - 4. Eye-caps mostly or wholly white; antennæ annulate; markings blurred; fore wing with basal third of the paler (lemon-yellow) color.

13. packardella.

- 4. Eye-caps and white bands of fore wing with white, brown-tipped scales. 12. litigiosella.
- 4. Eye-caps usually yellow; white bands clear. Larger. Antennie even yellow-brown; markings of fore wing defined and more or less silvery, basal third of the darker yellow-brown.
 - 5. Tuft an isolated black spot.....14. coronatella. 5. Tuft followed by an equally large, chocolate brown area.

15. quinquenotella.

3. Fuscous-brown dominant.

4. Light; markings mainly of oblique streaks, and largely cream white.

17. ambrosicefoliella.

- 4. With an oval spot over and beyond tuft, and a similar one opposite it on costa; base mainly pale.....16. pomifoliella.
- 4. Shining deep brown with yellowish-silver spots.
- 5. Only four spots......19. locuples. 4. Suffused powdery umber brown, the dorsal oval spot alone distinct,
- and outlined with white; or with scattered white patches.

20. ainsliella.

Not in key: species C, copeuta, cuneigera, angustata.

1. B. luteella Chambers. Cream white, most of head and eye-caps pure white, shaft of antenna and vertex pale yellow; thorax and fore wing shaded with pale yellow, especially at middle and apex, contrasting with the white fringe. A very small tuft at middle of inner margin; no apical spot. 4 mm. This diagnosis has been drawn up from the type at Washington.

March.

Kentucky.

2. B. magnella Chambers. White. A luteous band across eye-cap; fore wing with a luteous stripe from middle of costa to lower half of apex, sometimes extending farther toward base; another oblique costal streak meeting the first at outer end; and a streak at middle of inner margin, with some raised black scales at its inner end; scales toward apex with black tips. Costal fringe white; apical fringe brown, dorsal pale yellow toward apex, with two black lines of scale-tips. 8 mm.

The tongue and palpi are concealed by the overhanging face in this series.

June. Larva on Solidago; cocoon smooth.

Connecticut to Kentucky.

3. B. fusicola Braun. Near B. magnella; head with a few fuscous scales in middle of tuft; outer part, only, of antenna fuscous. Fore wing with longitudinal streak from base strong in female, weak in male; first costal streak more oblique

than second, its outer part longitudinal A line of black subterminal scales and a black line in fringe. 12 mm. (magnella in part of collections.)

Larva in a spindle-shaped gall on Helianthus tracheliiformis, located in the upper part of the stem; about 2 cm. long and 5 mm. wide. Cocoon grayish brown, smooth. and strongly flattened. Larva in September; moth from end of May to early July. Cincinnati, Ohio.

4. B. montana Braun. Similar to B. magnella and B. fusicola. Vertex with some fuscous scales; shaft of antenna all gray. Fore wing with *three* oblique streaks from the costa, parallel and equidistant, besides the dark apical shading; the first two streaks confluent below. 11 mm.

June.

Mountain Lake, Virginia.

5. B. niveella Chambers. White; slightly yellowish at tip of wing and vertex; fore wing with some black scales in costal fringe, and two lines in dorsal fringe only.

Magnella is probably a fully marked variety of this; in fact I have seen inter-mediate specimens, bred from Solidago. Kentucky. New York: Rock City (Cattaraugus County).

6. B. errans Braun. Head and thorax white, brown in center; shaft of antennæ brown. Fore wing dark brown, with longitudinal white streaks at base above and below middle, leaving a brown longitudinal streak between them; two oblique white streaks on costa, the more basal heavier, and a white costo-apical spot in fringe; a streak, followed by two white dots, on dorsum. A dark line in fringe. Sometimes with the white streaks so enlarged as to dominate over the brown ground, and more or less fused. 10 mm.

ground, and more or less tused. 10 mm.
Food Aster shortii. Larva forming a contorted linear mine in the fall, ending in an enlargement in which is spun a silken wintering cocoon. Larva in the spring boring in the tip of a growing shoot, killing it. Pupal cocoon whitish, with faint longitudinal ridges. Moth in May. Cincinnati, Ohio; Okefinoke Swamp, Georgia.
7. B. species C. Head dirty white; eye-caps white; antennæ annulate, light dull brown and dirty white; fore wing whitish and dull light brown, with a quadrate, blackish patch in middle of fold; a short oblique brown shade before middle, extending narrowly along costa to have:

qualate, oneckish patch in mindle of fold; a short oblique brown shade before middle, extending narrowly along costa to base; another from three-fifths way out on costa, to outer margin above apex, ending in a black dot; and a subtriangular, costal subterminal patch; the bands broader than the distance between them; a contrasting, black, apical hook; tuft black, followed by a small brown area. 6-8 mm. (*ambrosizefoliella* auct., apparently not of Chambers). Doublefully distinct from the work two species.

Doubtfully distinct from the next two species. Larva on Ambrosia; cocoon ribbed.

Kentucky; western Pennsylvania; Missouri; and elsewhere.

8. B. agnella Clemens. Bands powdery black on light brown, not strongly contrasting; anal dot reduced to a few scales. Head mostly white. 6 mm.

Kentucky; District of Columbia.

9. B. copeuta Meyrick belongs to this group. It was described from Ontario.

10. B. capitealbella Chambers. Antennæ annulate with yellow-brown; ground color pure white and markings brighter yellow-brown; no black scales at anal angle; first fascia not continued along costa to base. Otherwise like *B. agnella*.

I have seen the type.

11. B. albicapitella Chambers. Cream color, very sparsely dusted with fuscous, no brown; center of tuft and somewhat blurred bands on wing yellow. 51/2 mm. This is *luteella* of collections, but not of Chambers.

Kentucky.

12. B. litigiosella Zeller. Pale straw yellow; the scales of the ground tipped with contrasting dark brown; little or no brown on face; the brown more distinct on eye-caps and very strong on the pale bands of the wing. Antennæ annulate in yellow and two shades of brown; vertex yellow-brown. Fore wing with costa before middle, and a transverse median band yellow-brown, and some brown farther out; fringe with black-tipped scales in basal half, without lines. 6 mm.

Early spring. Larva on white oak.

Pennsylvania.

13. B. packardella Chambers. Head and eye-caps white; tip of vertical tuft mixed pale golden and brownish; antennæ pale yellow, brown-dotted; thorax white, brown-dusted; basal half of fore wing white, fleeked with brown; a chrome-orange streak on fold and one on costa, spreading into the chrome-orange outer half of the wing, which is more or less brown-powdered on the costa; a faint white streak from middle of wing to anal angle and one across apex beyond it; fringe yellow, with two lines. 6 mm. (*trifasciella* Clemens, *obscurofasciella* Clambers).

Larva on chestnut, oak, and beech; moth in April.

The name *trifasciella* will have priority if it really represents this form.

Distribution general, extending west to California.

14. B. coronatella Clemens. Head with tuft pale ochreous, face yellowish white; eye-caps mainly pale yellow, more orange hehind. Fore wings pale orange chrome with a whitish patch near the base over fold, one nearly opposite and joined to it on inner margin, and one near middle of costa; near tip, a whitish, transverse band to middle of dorsal fringe; extreme tip whitish; the tuft and a black dot at apex and line in fringe being the only black marks. 6 mm.

Larva on black birch.

Pennsylvania to District of Columbia, and vicinity.

15. B. quinquenotella Chambers. Pale straw color, the eye-caps palest, and wings darkest. Vertex brown-tipped; thorax yellow-brown, with straw-yellow edges and tegulæ; fore wing straw color, the ground nearly covered by a broad, antemedial, yellow-brown fascia, and confluent outer bands, leaving pale spots, much as in *canadensisella*. Tuft with blackish scales followed by a chocolate brown area; an oblique, pale gold subterminal streak from costa, preceded by chocolate brown; all the markings but the last and the tuft, very diffuse. 7 mm. This species has been bred from a ribbed cocoon on Ampelopsis.

16. B. pomifoliella Clemens. Head and eye-cap cream white, the tuft centered with brownish; antennæ pale ochreous, dotted with dark fuscous; fore wings cream white, dusted and shaded with brown; base with brown streaks on costa. fold, and inner margin; patch on middle of margin large, oval; a streak from middle

of costa to anal angle, ending in a black dot, broadest on costa; a dark brown apical spot, and a dark line in fringe across apex. 7 mm. (pomonella Packard, curvilineatella Packard.) (H. p. 432 f. 251.) Larva on apple in September; dark yellowish green with brown head, with distinct dark hairs, with the usual habits, cocoon ribbed on the twice. The moth

tinct, dark hairs; with the usual habits; cocoon ribbed, on the twigs. The moth flies mostly in June and is generally distributed; common everywhere in New York.

17. B. ambrosiæfoliella Chambers. Head white, with discolored tuft; antennæ annulated, dark brown and white; thorax ochreous yellow, faintly sprinkled with brown (or with three yellow lines on a white ground); fore wings shaded, ochreous and white; an oval, brown, scaled area on inner margin containing the tuft, and edged with white; a short, brown, antemedial costal bar; a postmedial bar edged with white at the costa, extending across the wing to the anal angle, and along the outer margin to the apex; one slanting black line in dorsal fringe; and some scales in dorsal fringe. 7 mm.

Kentueky.

I think I have recognized the species mixed with *B. pomifoliella*. It cats Ambrosia, but is not the better-known Composite-feeder (*agnella*?).

18. B. canadensisella Chambers. Similar to B. pomifoliella, but darker; head white; tuft centered with yellow brown; thorax brown with a complete white

margin; fore wing brown, white at base; - the oblique, brown, antemedial area followed by a white fascia, interrupted at middle of wing and erect below, with the black tuft on its edge; white spots at middle of costa and anal angle; tending to join a streak from costa. Much like augustata, but wholly lacking the silver basal dash. A velvety brown apical spot, followed by a curved line in the fringe. (H. p. 431 f. 250.)

The larva feeds late in the autumn, in the usual manner on white birch, which it may disfigure badly; but it does little real damage because the leaves are almost ready to fall when it appears. The moth emerges in the spring.

This species is of general distribution south to Pennsylvania. New York.

19. B. locuples Meyrick. Blackish brown; head pale bronzy, with darker tuft. Fore wing with triangular golden spots at middle and near apex of costa; a larger transverse spot at middle of inner margin and a smaller spot at tornus. Hind wings dark; fringes gray. 7 mm. (Unknown to me.) July.

Toronto.

20. B. ainsliella Murtfeldt. Blackish, heavily dusted on a dirty white base; vertex with fuscous tuft; eve-caps whitish, antennæ annulate. Fore wing with ground color very largely blackish; the pale parts outlining an oval, blacker patch on inner margin, parallel to whose upper, outer side there is a pale streak running down from the costa (variabilis Braun). The larva normally feeds on black oak, and has the habits and cocoon usual

in the genus; in an epidemic it attacks many trees.

B. llecella Busck, a similar, holly feeding species described from Texas, is to be expected in the range of its food plant.

21. B. crescentella Braun. Head whitish; some dark hair in tuft; fore wing ochreous or darker brown; basal dash white, faint in lighter specimens, extending to middle of wing, with a dark shade below it; a costal streak at middle of wing, oblique and concave outwardly; a less oblique streak three-fourths way out; with a darker space between the two; a white streak over an irregular black spot at apex; dorsal spot dark brown, edged with white. 7-9 mm.

The larva makes a trumpet mine, with a central frass-line, on aster, Solidago, and Erigeron, a single larva making several mines. But it never lives externally. The cocoon is normal, white. The moth flies in July.

Ohio to Toronto, Ontario, and New Hampshire. New York: Otto, Florida. 22. B. cuneigera Meyrick appears to be similar to crescentella but with the disc of thorax white. It was described from Ontario.

23. B. angustata Frey and Boll, of which *crescentella* may be a variety, is similar, with a white head and brown thorax, and a dark brown fore wing, with white markings. The thorax is typically white in B. crescentella.

8. PHILONOME Chambers

Hardly distinct from Bucculatrix. Face shorter, truncate below, exposing the drooping labial, and minute, folded maxillary palpi. Only one species known.

1. P. clemensella Chambers. Palpi, face, and eye-caps white, the latter with orange upper edge; antennæ reddish orange at base, the rest nearly white; thorax white, with broad pale orange area behind, and with orange spots on shoulders; fore wing reddish orange, with broad white streaks below costa and on inner margin, the latter extending to a small tuft of brown scales, then turning obliquely up and meeting the end of the other streak; an oblique streak from costa, two-thirds way out, extended along costa toward base. Apex dusted with dark brown; a brown hook in apical fringe, and two converging streaks in dorsal fringe. 8 mm.

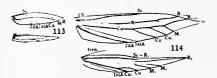
Larva on hickory and linden. Moth in July.

Northern New Jersey to Kentucky. New York (Beutenmuller).

Family 13. OPOSTEGIDÆ

(Tineidæ, in part)

Head covered with large, smooth seales, but with a strong tuft of long hair between the antennæ; palpi small, without bristles; maxillary palpi small, but quite distinctly folded, tongue very small; antennæ short, stout; with a large eye-cap, which is covered by regularly imbricated scales, and ribbed on the inner side. Body small; middle and hind tibiæ with long, stiff bristles. Shaft of antennæ with a single whorl of scales to a segment. Fore wing in the American species with no branched veins (fig. 113); with four or five simple ones tending to join at base, only the one through the middle of the wing distinctly tubular. Aculeæ present but abnormal, pointing costad; arranged in regular rows, and confined to a small area about the base of wing. Hind wing half as wide as fore wing, lanceolate; Sc and Cu simple,



FIGS. 113-114.

113, OPOSTEGIDÆ, Opostega salaciella (Europe), venation; 114, OINOPHILIDÆ, Opogona aurisquamosa (Hawaii), venation with a single, three-branched vein between them; frenulum a diffuse tuft of bristles, somewhat concentrated in male, of a lower type than is elsewhere known in the Frenatæ.

Larva extremely slender, cylindrical, with the setæ apparently arranged in regular circles about the body; legs wholly absent. Head flattened, with thickened lateral keels on the epicrania; setæ re-

duced. Front wider at back. Ocellus single, obsolete. Mouth parts small and reduced; labrum modified and retracted into a notch in the clypeus, mandible thin, but of biting type, with a membranous process. Dorsal part of head not extending far into thorax, but extended by a couple of heavy tendons; ventral side largely membranous, hypostoma rudimentary.

The larvæ mine in bast; the only known European species in flowerstalks of Caltha, O. albogalleriella in Ribes. The pupa has not been studied.

The family is small, and almost entirely Oriental, where a couple of other genera occur, as well as Opostega. The relationships are quite obscure, but the group seems to represent, as near as anything, the point of origin of the Lyonetiidæ from the common Tineoid stem; as it shows characters that appear also in the Tineidæ, Nepticulidæ, Psychidæ and Lyonetiidæ. The family Oinophilidæ is hardly distinct, and some genera of the latter show a nearly complete, normal venation; the genus Opogona (fig. 114) is about halfway between the two families.

OPOSTEGA Zeller

Eye-cap very large, covering the base of the wings, as well as the whole side of the head, in repose; fore wing caudate, without forked veins. Larvæ bast-miners.

1. O. albogalleriella Clemens. Silvery white, with a minute black dot at tip of membrane of fore wing; apex of costa somewhat yellowish; two fine, dark lines in costal and dorsal fringes (nonstrigella Chambers, accessoriella Frey and Boll).

Var. quadristrigella Chambers has a gray shade near middle of inner margin.

The larva is a bast-miner on gooseberry and red and black currant. The mine often starts in a two-year-old stem and usually ends in it, running up and down the canes, and leaving a scar which is usually visible before the larva leaves the cane, late in June. The cocoon is seed-like and brown. It is spun in the soil, like that of Nepticula.

Massachusetts to Texas. New York: Otto, Geneva.

It is possible that quadristrigella is a distinct species; it may feed on Rancunculaceæ.

2. O. cretea Meyrick. White; three lines in costal and two in dorsal fringe; a minute black apical spot; an oblique gray dorsal spot at middle of wing. 8-9 mm. Lake Muskoka, Ontario; July and August.

This species is unknown to me and indistinguishable from its description from O. quadristrigella.

3. O. scioterma Meyrick. Fore wing similar to O. quadristrigella, with an additional gray antemedial band nearly meeting the dorsal one, and a gray border. 9 mm. (Unknown to me.)

June.

Toronto, Ontario.

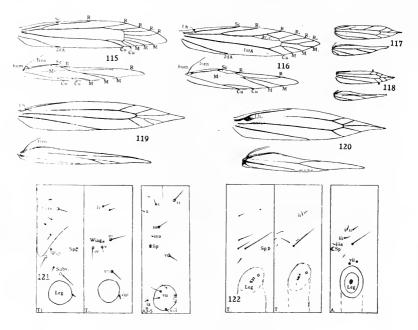
Family 14. **GRACILARIIDÆ**

(Tineidæ, in part)

Vertex either rough or smooth; face of the northern species smooth, at least below; ocelli usually absent; tongue well-developed; labial palpi moderate or long, often upturned, but usually with the third joint set on at a slight angle to the second, and the individual joints not noticeably curved; third joint normally blunt; no bristles. Maxillary palpi of porrect type, never folded across the front of the tongue, moderate to rudimentary, sometimes practically absent. Hind tibiæ normally smooth-scaled, often with a series of bristles above; the midtibiæ also bristled in some exotic species. Antennæ relatively long, often as long as fore wing, simple, with two full whorls of scales to a segment; eye-cap absent in our species.

Wings without aculeæ; fore wing lanceolate or linear-lanceolate, with lanceolate cell; the accessory cell rarely well-marked; **Sc** rather short, \mathbf{R}_1 arising before middle of cell, or absent, \mathbf{R}_5 running to costa (when recognizable); when stalked it is stalked farther with \mathbf{M}_1 than with \mathbf{R}_4 except in Cremastobombycia. The other veins short, and usually running sharply across to margin, part of them lost in the majority of forms. Apex of membrane very often drawn out in a long point, which rarely projects beyond the costal and dorsal fringe (caudate). 1st A a well-marked fold, 2d A simple, often strongly sin-

uous, without a trace of basal fork. Hind wing lanceolate (fig. 115) or linear, usually abruptly narrowing before middle; often with strongly sinuous costa; the fringe much broader than the wing. Frenulum of female with two bristles. Venation of the hind wing usually more or less obscure, the veins being mere thickenings and



FIGS. 115-122. GRACILARIIDÆ

115, Gracilaria alchimiella (Europe), venation; 116, Parornix anglicella (Europe), venation; 117, Marmara salictella, venation; 118, Leucanthiza amphicarpexfoliella, venation (the asterisks mark inconstant veins); 119, Cremastobombycia solidaginis, venation; 120, Lithocolletis emberizxpennella \mathcal{Q} (Europe), venation; 121, Acrocercops strigifinitella, larva, seta map (after Heinrich); 122, Lithocolletis species (from maple), seta map

difficult to trace either in denuded dry wings or in stained ones. Sc and **R** closely parallel when the venation is best developed, and connected by what appears to be an oblique crossvein about twofifths way out, which is much farther from base at its lower end; Sc either stopping at that point or running well toward apex; **R** reaching apex; \mathbf{M}_1 and \mathbf{M}_2 arising from radial stem, or connected to it by a short cross-vein; \mathbf{M}_3 from **Cu**-stem; cell always open between

 \mathbf{M}_2 and \mathbf{M}_3 , \mathbf{Cu}_2 and either \mathbf{M}_3 or \mathbf{Cu}_1 apparently always preserved. The free tip of \mathbf{R}_1 is preserved in primitive forms, apparently arising from the cross vein connecting **Sc** and **R**. Anal region obsolete.

The caterpillars (figs. 121, 122) always leaf (or bast) miners, at least when young, often changing their habits when part grown and forming nests in a folded leaf. Young caterpillar unique, specialized for feeding on the sap of the cells, which it opens with its blade-like mandibles; much flattened; lobed at the sides; widest near the anterior end. Head extremely flat; front widening toward the posterior end; adfrontals not normally recognizable, perhaps obsolete; maxillæ and labium very narrow as a rule, strap-shaped, and almost wholly incorporated in the head capsule, the free part of the maxillæ and labium minute. Ocelli reduced in size or number (frequently to a single pair). True legs sometimes absent; body-setæ much reduced, frequently unrecognizable, iv and v well separated; prolegs more or less reduced, often absent when the true legs are absent; hooks, when present, in one or two transverse or curved series, uniordinal, but sometimes with one of the series doubled. Prolegs always absent on sixth segment of abdomen (characteristic of the family). The caterpillar in this stage is always a miner, forming a more or less opaque mine (because the parenchyma is not eaten), which is often invisible from one surface of the leaf.

Full-grown larva strikingly different. Head but little flattened, with normal mandibles and maxillæ, and labium with spinneret. The bases of the maxillæ and labium long and slender. Prolegs always well developed on third to fifth and last segments of abdomen, but wholly absent on the sixth, the hooks as in the flat stage, but always more or less developed. Body cylindrical, with minute but recognizable setæ.

This type of caterpillar feeds on the parenchyma, either in an inflated mine, more or less lined and puckered with silk, or externally (skeletonizing the leaf) in a shelter which, in the typical forms takes the form of a cone, with the larger end closed by folding over the end of the leaf. In some species the cylindrical larva does not feed, but immediately spins the cocoon. The cocoon is usually of white silk, and is not woven in regular meshes. The mining species spin, as a rule, within the mine. The external cocoons of Marmara and some species of Acrocercops are ornamented with four groups of white bubbles, which are ejected as a froth through slits cut in the cocoon after it is spun; and soon set hard.

The change from one type of larva to the other takes place at different stages in different genera, and even in members of the same genus. In a couple of species of Acrocercops (**Neurobathra Ely**) the change is gradual, covering two or three molts.

Pupa incomplete, but specialized; derived from an early or pretineid type. Vertex very large, prothorax wide at sides, much narrowed in middle; front often with a serrate cocoon-cutter; maxillary palpi minute or absent; labrum well marked; antennæ and tongue long, the latter as long as the hind legs and extending free beyond the tip of the wings. Abdominal segments with fine, diffuse spining dorsally, without a specialized structure at anterior edge of middle segments dorsally, fourth segment of abdomen *fixed*, but eighth movable in male. Hibernation in imago or pupa, (in the latter case, with the imago well advanced inside).

The genus Phyllocnistis (fig. 112) might well be transferred to this family, but is markedly aberrant, especially in having preserved the legs of the sixth abdominal segment and in having lost the mouth parts in the evlindrical larva. The pupa is distinctly Gracilariid, but has abdominal segments 8, 9, and 10 much reduced, and peculiar dorsal structures on segments 3 to 7. The family as a whole is closely cognate with the Lyonetiidæ, indicating its Tineine derivation; but in its porrect maxillary palpi it shows relationship to the Yponomeutoidea, and it is superficially convergent with the Lavernidæ.

Key to the genera

Moth

- 1. Hind tibiæ with a series of bristles above.
- 2. Bristles fully formed; maxillary palpi (in our species) distinct.

3. Acrocercops.

- 2. Bristles imperfect, scale-like; maxillary palpi rudimentary. 4. Apophthisis. 2. Bristles very strong, maxillary palpi rudimentary; antennæ often with(Lyonetiidæ-Phyllocnistis).
- eye-cap
- 1. Hind tibiæ without series of bristles. 2. Vertex with rough bristly hair.
 - 3. Fore wing with R, absent, first radial arising near tip of cell; maxillary palpi obsolete (figs. 119, 120).
 - 4. Fore wing with three veins running from cell to inner margin (fig. 119). 8. Cremastobombycia.
 - 4. Two veins running from cell to inner margin. M₃ absent (fig. 120). 9. Lithocolletis.
 - 3. Fore wing with \mathbf{R}_1 present, arising from middle of cell (fig. 116); maxillary palpi conspicuous.....l. Parornix.
 - 2. Head all smooth scaled (the scales somewhat erectile).
 - 3. Fore wing with \mathbf{R}_1 absent (fig. 117).
 - 4. Maxillary palpi moderate6. Marmara.
 - 3. Fore wing with R, preserved (fig. 115).
 - 4. Middle tibiæ thickened irregularly with rough scales 2. Gracilaria.
 - 4. Middle tibiæ slightly thickened at tip, or smooth; often with a little hair.
 - 5. Hind tibiæ smooth; fore wings with a definite pattern. 7. Parectopa. 5. Hind tibiæ rough above; fore wings without markings.

4. Apophthisis.

Pupa

- 1. Prothorax crested, and four times as long on sides as on middle line.
 - 2. Dorsum of abdominal segments with only one size of spines; the segments not crested behind.
 - 3. Spined areas the whole length of the segments.

 - distinct 5. Leucanthiza.
 - 3. Spined areas a fourth as wide as the length of the segments.

 - 4. Head with a prominent crest (cocoon cutter); tongue extending less
 - 2. Spines of two sizes; the posterior edge of the segments usually crested. 9. Lithocolletis.
- 1. Prothorax depressed and neck-like, not more than twice as wide on side as on mid-dorsal line.
 - 2. Spines very coarse, sometimes with fine ones intermixed.
 - 3. Head with a ventral cutting-plate, which is usually serrate; maxillæ as

1. PARORNIX Spuler

(Ornix Treitschke, in part)

Vertex strongly roughened; front smooth; antennæ as long as fore wing, without pecten; palpi moderate, smooth; third segment somewhat shorter than second; maxillary palpi fairly large, similar to labials. Hind tibiæ smooth. Fore wing lanceolate (fig. 116); five veins running to costa, and four to inner margin; \mathbf{R}_i arising a third way out on cell; accessory cell usually preserved and \mathbf{R}_s and \mathbf{M}_i stalked. Hind wings narrow-lanceolate; tip of \mathbf{R}_i free but Sc coincident for a considerable distance with costal edge; venation nearly complete. M-stem usually forked; fringe three times as wide as membrane.

Larva of cylindrical type with normal legs (except when very young) with two curved bands of hooks, enclosing a straight row.

The larvæ leave their mine when well grown and form a nest by folding over the edge of a leaf, usually flatly. The pupa is formed in a similar nest. Most species feed on Rosaceæ and Amentiferæ. The moths rest with head raised, and fore and middle legs displayed. They fly freely in the afternoon. The species resemble each other closely, and are indeterminable if at all rubbed. The pupa hibernates.

Key to the species

- 1. Ground evenly brown, shining, with white or yellow marginal spots.
 - 2. A complete fascia; head concolorous.
 - 3. Five costal spots, outer half of apical fringe gray......3. preciosella.
 - 3. Five costal spots followed by a white bar in apical fringe; longer fringescales at apex pale, and black-tipped 2. kalmiella.

 No complete fascia; head yellow
 5. cratugifoliella. 7. Palpus with third segment bearing, at most, smaller fuscous spots. 4. Costal striæ regular
 6. A slight yellow tint on head and base of fringe of hind wing. 16. inusitatumella. 6. No yellow tint
 Costal striæ much reduced, mere spots
l. P. guttea Haworth. First costal and dorsal spots opposite each other. Head and collar pale. Fore wing dark brown, with pinkish gloss; typically with four or more costal spots, but the outer ones sometimes minute, and two dorsal spots.

Variety solitariella Dietz has the fourth costal spot large; three dorsal spots and a white sector in the fringe. 10-13 mm.

One brood, flying in May. Larva translucent, yellow, with black cervical marks only, as usual in Parornix; narrowly turning under the edge of a leaf; on apple. New Hampshire to District of Columbia; westward to the Paeifie.

2. P. kalmiella Dietz. Golden brown; the three dorsal spots farther out than the corresponding costal ones, and the one in fringe larger. 8 mm.

¹⁴Maxillary palpi whitish; antennæ brown above, annulate, paler below; fore wing with numerous striæ, the costal strlæ often more obscure toward base, and more oblique than the outer ones; last stria curved, usually crossing both fringes and enclosing a blackish apical spot; ground color toward costa usually dark; usually two antemedial and medial dark patches (the plical spots) in the fold; inner margin whitish, mostly white at base; three dark lines in apical fringe (rarely four). Fore and middle legs blackish fuscous, displayed; femora white, spotted; tips of tibiæ white; tarsi annulate; hind legs whitish, the joints annulate with darker. There are two broods, the moths flying in May and in August.

Moth in May. Larva pale green, in a pale orange blotch on the upper side of leaves of Kalmia angustifolia; in the autumn.

Connecticut; Pennsylvania.

3. P. preciosella Dietz. Dark bronzy-brown; spots violet-silver, the first dorsal joining the second costal spot, and the fourth dorsal minute but normally joining the fifth costal. 8 mm.

May. Larva on swamp huckleberry. (V. corymbosum) in autumn.

Pennsylvania; Connecticut.

4. P. anglicella Stainton. Dark gray, the inner margin not noticeably pale, the third line in fringe ending abruptly at the lower of the two white rays. 10 mm. (fragariæ Stainton.)

The larva forms a slender, conical roll of the edge of a leaf; on thorn and strawberry. Europe; New England.

P. innotata Walsingham was not recognizably described, nor associated with a definite food-plant or locality.

United States.

P. trepidella Clemens. Palpi annulated with dark brown near tip.

This species is unknown to me.

Pennsylvania.

5. P. cratægifoliella Clemens. Head intermixed dark brown and gray; antennæ faintly annulate with whitish. Fore wings dark brown with a purplish hue, dusted heavily with whitish along inner margin; a dark brown streak in base of fold, and a blotch near middle which almost reaches the inner margin. Whitish costal streaks distinct near apex, the last pair enclosing the dark apex. Fringe with two dark streaks and with white tips. 8 mm.

The larva is a miner on thorn; it is greenish white with reddish brown dorsum and brown head. It pupates under the folded edge of a leaf.

The moth flies in may. Pennsylvania; District of Columbia.

6. P. dubitella Dietz. Inner margin whitish. contrasting with ground; with the blackish plical spots, and two or three smaller ones on inner margin. 10 mm.

The larva forms a tract-mine on an undetermined shrub.

Pennsylvania.

7. P. conspicuella Dietz. Inner margin white on basal two-fifths only. Ground otherwise rather even. Striæ white, contrasting; about eight striæ on costa and five or more on inner margin, the outermost ones crossing fringe and cutting off the line in fringe, as in anglicella, but distinctly above the apex. Occipital tuft pale. 8 mm.

Caterpillar on black birch.

Pennsylvania.

8. P. arbitrella Dietz. No contrasts; costal striæ rather distinct, dorsal ones obsolete; a white discal bar. 8 mm.

Larva on Vaccinium.

Pennsylvania.

9. P. melanotella Dietz. Brown, with about nine equally strong, white streaks on costa; two white spots on fold, on a blackish ground; basal half of inner margin pale. 7 mm.

Larva on Cratægus.

Pennsylvania.

10. P. geminatella Packard. Fore wing dark brown and powdery gray, with obscure costal striæ and very obscure plical spots. 8 mm. (prunivorella Chambers ?)

Larva in a tentiform mine on apple and quince. Distribution general.

11. P. quadripunctella Clemens. Face tuscous; tuft dark brown; fore wing with inner margin contrastingly whitish. 8 mm.

Larva on apple and chokecherry.

Eastern States to Kansas.

12. P. vicinella Dietz. Dark brown with a purplish tint, with dark and obscure markings. 7 mm.

Larva on yellow birch.

Pennsylvania.

13. P. sorbivorella Dietz. Grayish brown; occiput and thorax creamy white; markings mostly weak; plical spots black, contracting; a white discal bar. 10 mm. (strobirorella Dietz, misprint.)

Larva in a cone like Gracilaria, on mountain ash.

Pennsylvania.

14. P. arbutifoliella Dietz. Gray with brown costa; inner margin mostly white, with triangular black spots in fold. Head and thorax white. 9 mm.

Larva on Pyrus arbutifolia (chokeberry).

Pennsylvania.

15. P. obliterella Dietz. Purple-brown; fourth and fifth striæ widely separate; inner third of wing half white and half black. Head and thorax with much white. 8 mm.

Larva on Betula nigra.

Pennsvlvania.

16. P. inusitatumella Chambers. Ground color uniform; three entire lines in fringe; moth similar to *prunivorelia*, but smaller, with more distinct striæ, and no semierect scales on inner margin. $7\frac{1}{2}-8$ mm.

Larva in an upper-side mine on *Cratægus tomentosus* and *C. mollis*, forming a nearly circular, whitish blotch with scattered frass, later made tentiform. Cocoon yellowish brown, spun on upper side of mine, outside.

Kentucky; Ohio.

P. trepidella and festinella Clemens are unknown to me; for "Ornix" quercifoliella Chambers and boreasella Clemens see Acrocercops.

2. GRACILARIA Haworth

(With Coriscium Zeller; Euspilapteryx Stephens, etc.)

Similar to Ornix; head smooth; scape sometimes with the rudiment of an eyecap; palpi smooth, rough, or with a triangular tuft on second segment. Fore wing with all veins (fig. 115) or with one dorsal vein lost; \mathbf{R}_1 arising close to base, easily mistaken for Sc, which is inconspicuous. Hind wing lanceolate or linear; venation weak and somewhat variable; typically with the tip of \mathbf{R}_1 free as in Parornix.

The moth rests with head raised, and the two rough-scaled anterior pairs of legs displayed. Larva, after an early stage, cylindrical; at first forming a blotch-mine, later rolling a leaf into a conical nest. A few species are miners till mature.

The species of this genus are difficult to identify without knowledge of the food plants. as they are closely similar and often highly variable. The key will be a partial guide only.

Key to the species

- 1. Fore wing with a subtriangular, or larger, irregular, golden patch, normally contrasting with the darker ground; well-defined, at least on its basal side, and extending a third way across the wing, or more.
 - 2. Upper part, at least, of face, and palpi dark. 19. purpuriella, 20. stigmatella.
 - 2. Upper part of face and palpi not dark.

- Palpus with bands broad and fuscous, the ground more or less dusted with fuscous so that about half the total scales are dark; patch normally diffuse; ground of wing predominantly golden......21. negundella.
 Palpus white or partly yellow with less than half of the third segment contrasting black, and sometimes a narrow black ring on the second; rarely dark, in which case the fore wing also is dark.
 A Bace on inner margin accelerates the whole ground dark purplish.
- - 4. Base on inner margin concolorous, the whole ground dark purplish, always with two well-marked golden spots. 5. Costa solid dark brown, not barred before apex; ground umber brown,
 - with a violet-purple iridescence. Third segment of palpus with more black.
 - 6. Brilliant royal purple; antenna pale; spots normally separated by half their width, the first crescentic and the second long and

 - yellow-brown, with brilliant crimson-purple iridescence; palpus with a small black spot near apex of third segment; spots large and nearly

 - 5. Palpi strongly yellowish, markings usually suffused, the outer spot often nearly obliterated......23. ostryæella.
 - 5. Palpi white, markings clean-cut......15. vacciniella. 6. First spot large and triangular, and second one nearly in contact with it, or with the spots fused into a large, irregular patch.

16. belfrageella.

- 4. Fore wing dark purplish, with middle of base contrastingly golden; costal patch large and irregular.....25. blandella, 26. juglandivorella.
- 4. Fore wing crimson, the base of the inner margin more or less contrastingly paler.
 - 5. A single, narrow, triangular, costal, golden patch, ending about at middle of costa.

- 6. Head and thorax light golden yellow.....18. packardella. 5. A large, irregular patch extending well toward apex, often diffuse
 - except at base; or with two patches, the outer one suffused. 6. Vertex light golden, much paler than the ground of the wings; or 6. Vertex concolorous.

7. Thorax wholly golden; head predominantly golden.

24. superbifrontella.

7. Tegulæ dark, vertex more extensively dark.

8. Vertex and tegulæ rusty, with crimson iridescence.

24. alchimiella.

8. Vertex and tegulæ duller grayish purple; patch diffuse. 22. azaleæ.

5. Two well-separated and sharply defined patches......17. coroniella.

1. No golden area with sharply defined, oblique, basal boundary, on costa of fore wing.

2. Palpus with a small, triangular tuft on second segment. 3. Ground all pale luteous and white, with very fine, black seale-tips. 31. paradoxa. 3. Ground darker, with considerable brown or gray. 4. Fore and middle tibiæ and under side of wings red-brown, tarsi whitish. 32. quercinigrella. 4. Fore and middle legs powdery grav, with narrow white bands; underside gray. 5. Ground dominantly white, with complete, blackish fasciæ; scales dark-tipped 5. White ground reduced to scattered spots, the fore wing almost wholly powdery gray and brown; fasciæ broken or obscure; scales white-2. Palpus smoothly scaled or nearly so. 3. Reddish, with costal third suffused with yellow. 4. Darker ground suffused with crimson. 5. Larger; darker; costal edge with two larger dark dots, hind tarsi contrastingly darkened at tips of joints......4. sassafrasella. 5. Smaller; paler; several black points on costa; hind tarsi evenly silver gray10. violacella. 3. Reddish, with costal edge very narrowly, or not at all, yellow. 6. elongella. 3. Not reddish; gray or brown. 4. Middle half of costal edge narrowly pale with dark dots. 5. Pale edge sharply defined below; ground darker and richer brown. 2. aceriella, 3. juglandiella. 5. Pale edge shading into the fuscous ground......5. rhoifoliella. 5. Fuseous gray, more or less violet-iridescent, the costal third faintly paler and costal edge striate at middle.....8. minimella. 4. With traces of the two golden triangles; costal edge with dark dots 23. ostruæella. 4. Costal edge concolorous, or nearly so. 5. Dark brown, spotted with yellow-brown. 6. Spots bright ochre and conspicuous, two on eosta and two or three 5. Dark brown, powdered on a yellow ground, less heavily on costa. 6. clongella. 5. Dark brown, powdered on a yellow ground; base darker. 28. atomosella. 5. Dusted with blackish on a white ground, more or less heavily. 6. elongella. 5. Blackish without noticeable markings.....l. strictella. Synopsis by foods Willow and poplar (moths with a single golden triangle): Light red20. stigmatella. Dark purple19. purpuriella. Myrica: Rusty, with yellower costa......9. flavella. Walnut:

Ironwood (Carpinus and Ostrya):
Markings strongly variable
Birch:
Two costal patches; ground yellow and crimson
Alder:
More or less distinct costal triangles
No costal triangles
Oak:
Powdery gray
Crimson, with large golden patch
Sassafras:
Dark crimson, with costal yellow shade bearing two dark dots4. sassafraselia.
Witchazel:
Crimson, with large patch and golden thorax24. superbifrontello.
Apple: Powdery gray
Cherry:
Black and silver
Desmodium:
Crimson, with yellow costal shade and minute dots10. violacella.
Sumach:
Fuseous, with pale costa and dark points
Maple:
Suffused, with golden costal patch often diffuse (on A. Negundo only).
21. negundella.
Purple; two large triangles
Crimson; one large triangle
Purple; four small spots14. Species A.
Deep brown; a series of costal points2. aceriella.
Cornus (all species purple with two spots):
Dorsal margin golden
Spots approximate; first one crescentic
Spots distant; first one quadrangular12. cornusella.
Azalea: Light crimson and golden
Vaccinium:
Two small patches15. vaeciniella.
Privet:
Gray and white
Ash:
Powdery gray
Unknown:
1 strictella 7 flarimaculcila, 8. minimella, 28. atomosella, and 31. paradoxa.

I. Palpus smooth or nearly so (Gracilaria).

1. G. strictella Walker. Fore wings dark gray; with diffuse, blackish patches, and about five inconspicuous black dots, two of them costal. 18 mm. (adaptella Walker,)

This species is unknown to me; Ely associates it with G. alnirorella Chambers. This type was caught west of Hudson Bay.

2. G. aceriella Chambers. Similar to *G. juglandiella*, but slightly smaller; tip of palpus more distinctly white; fringe divided by two white lines. This species was apparently discovered at Amherst, Massachusetts.

It is unknown to me. It was bred from maple.

3. G. juglandiella Chambers. Deep purple-brown. Face whitish below; palpus pale, with a narrow black terminal ring on second segment, and a narrow basal, and very broad subapical, ring on third. Fore wing obscurely striate with blackish along inner margin; middle half of costa eream-white, cut into about eight, squarish spots by black hars. Apex and fringe with more or less distinctly paletipped scales. 9 mm. (juglandisnigracella Chambers.)

Moth in September. Larva in August on walnut, at first in a short, linear mine, which is later converted into a blotch. Later folding down the edge of a leaflet and feeding outside. Pupation in the nest.

South Ohio; Kentucky; Missouri (?).

4. G. sassafrasella Chambers. Ochre yellow; the dorsal two-thirds of the wing heavily suffused with deep, dull rese. Head and thorax more brownish; antennæ black-annulate except at base. Fore wing with somewhat diffuse, blackish dots at middle and just below apex, and some scattered black scales. Apex brown, with three blackish lines in fringe. 12 mm.

July. Larva first forms a serpentine mine on sassafras, which is later converted into a tentiform one. Then it forms a clumsy case by rolling down the tip of a leaf, usually a young leaf. The pupa is formed in a yellow cocoon on a leaf. The larva is slender, lemon yellow, and with a light brown head. The larva occurs in June.

Connecticut to Missouri. New York; Ithaca.

5. G. rhoifoliella Chambers. Fuscous. Front white; inner side of palpus white with blackish tips to second and terminal joints. Fore wing darker toward apex and base of costa, with purple iridescence (much duller than sassafrasella), costa with hasal and apical sixths wholly blackish, the rest shaded with creamcolor and cut with dark striæ, which are more numerous than in juglandiella, and thinner, with a single heavy one at the middle. Inconspicuous coarser striæ on inner margin, and sometimes on fold. Apical fringe with a couple of pale bars. 12 mm.

Larva with the same habits as that of G. sassafrasella, feeding on Rhus toxicodendron and R. copallina. Moth in July and August.

New Jersey to Missouri and Minnesota. New York: East Aurora, Ithaca. 6. G. elongella Linnæus. An extremely variable species, some of its forms determinable only by breeding from its food, alder. Typically crimson, unmarked, with red-brown fore and middle tibiæ, and dirty whitish hind tibiæ tinted with brownish. Forms occur with narrow, contrasting, yellow costal edge (inconstans Stainton), or with more extensive yellow costal markings, which are sometimes extended into a vague likeness of the costal triangle of the normal group of Gracilaria; and there may be also similar markings on the inner margin. In var. punctella (signipennella Hübner) there are a couple of fuscous dots, and the middle tibiæ are blackish. Form inconstans and most of the mottled forms have white hind tarsi; var. signipennis has the white tarsi, and three blackish dots on the fold; and the immaculate forms also occur with white tarsi. For American forms (which some consider a distinct species, calling it alnivorella Chambers) alnicolella Chambers represents the uniform red-brown form; var. sanguinella Beutenmuller is the red and yellow one; pulchella Clemens, the brickred one; fuscoochrella Beutenmuller is suffused generally with fuscous; nigristrigella and ruptostrigella Beutenmuller are mottled, brown, yellow, and black varieties, differing in slight details. Alnivorella Chambers is the pepper-and-salt form and is the oldest American name; and shastaëlla Beutenmuller is the greenishwhite form with sparser black dotting. All the forms and a variety of inter-mediates have been bred from alder in this country or Europe, and seem to represent a single species. Many of the forms were described from definite localities, but any are likely to occur anywhere in our area. The larva makes the usual cone on alder.

The species is evidently widespread but appears common only northward. New Wilmington, Mt. Whiteface, McLean. York:

7. G. flavimaculella Ely. Brown; face overlaid, and vertex more or less scaled, with pale yellow-brown; antennæ annulate with dark brown and yellow-brown; palpi straw color, the second segment heavily shaded with brown outwardly, and the third brown except at base and apex. Fore wings with rather diffuse mark-ings of pale yellow-brown, especially along the fold, dorsal margin, and costal margin beyond the middle, forming two conspicuous yellow splashes on costa. Front and middle tibiæ dark brown; tarsi straw color, annulate with dark brown. 10 mm.

Moth from July to September. Larva unknown.

Southern Connecticut.

8. G. minimella Ely. Fore and hind wings each with a dorsal vein lost. Fore wings dark mouse-gray with perceptibly paler costal third, cut by very faint costal striations, especially on middle half of costa. Palpus on inner side white, with a broad blackish band on third segment; second segment on outer side with a blackish tip, third mostly blackish with a white tip. Unlike *rhoifoliella* in the lack of a whitish costa and different palpi. 9-91/2 mm.

July and August. Larva unknown.

Southern Connecticut.

9. G. flavella Ely. Yellowish tan; face and palpi pale yellowish; vertex with purplish iridescence. Costa bordered with bright straw color; hind wings and fringes yellow-gray, the apical fringe straw-color, intermixed with tan. Legs as usual. 10 mm.

July. Larva on Myrica cerifera in June. Mine at first linear, then a small blotch (up to 2 by 4 mm.). Cone formed by rolling the tip of a leaf downward. Southern Connecticut.

10. G. violacella Clemens. Head and antennæ purplish brown; face white; palpi pale, with second and third segments each heavily dark-tipped. Fore wing with costal third light yellow except at apex, the rest light brown; a dozen dark points on costal edge. All the dark portions heavily shot with violet. 8 mm. (desmodifoliella Clemens). There is also a darker phase with the yellow costa extending only two-thirds way to the apex, and the third tarsi shaded with gray. The larva feeds on Desmodium, at first in a small linear mine, then in a tenti-form one; later rolling down the edge of a leaf from the tip. The moth emerges in August 1 t is generally distributed

in August. It is generally distributed.

The remaining species of this group all show more or less clearly a triangular golden patch on the costa, before the middle, or a larger and irregular patch. **11. G. burgessiella** Zeller. Fore and middle tibiæ dull black, contrasting with the whitish tarsi and whitish hind legs, as in all this series. Face silver white to level of antennæ, vertex brown. Thorax brown; palpi white with a short black tip. Fore wing umber brown, with purple iridescence, the more basal golden such as the straight or conceave side outward. spot comma-shaped or semicircular, with the straight or concave side outward; marked with dark dots on costa. Postmedial spot nearly quadrate, twice as long as wide. No subterminal bars on costa. Fringe fuscous with faint lines in apex. 11-14 mm.

Moth at the end of July and in August; apparently rare. Caterpillar on Cornus. in July; at first in a linear mine, then a blotch, then a cone formed by rolling the tip of the leaf downward.

Cocoon formed outside the mine. This name has been frequently applied to a variety of dark purple Gracilarias without a pale base to the inner margin, but seems unquestionably to belong to the species described above. Massachusetts; Connecticut. New York: Ithaca.

12.G cornusella Ely. Similar to G. burgessiclia, smaller; ground blackish, with faint iridescence; first golden spot normally quadrate, extending to fold, not large; second spot minute, separated from the first by twice its diameter. No subapical costal dots. Fringe and apex immaculate dark brown. Antennæ darker than G. burgessiella. 10 mm. (burgessiella of authors).

The caterpillar when young forms the mine usual in the genus; then lives in a cylindrical nest made by rolling down one side of a leaf. The cocoon is spun in the roll. The moth has been obtained in April and in August.

Connecticut to Missouri, New York: Ithaca,

13. G. bimaculatella Ely. Light brown with crimson iridescence, the antennæ yellow-brown. Head and thorax dark brown, darker than the wings; the face silvery. Palpi with a little black at the tip only. Fore wing with first golden spot large, triangular, practically reaching inner margin; and second nearly, or quite, in contact with it, less perfectly triangular; or roughly semicircular, with a few, obscure, dark spots between it and the apex. Apical fringe yellow-gray with darker lines. 11 mm. (burgessiella of collections; amphidelta Meyrick?).

The caterpillar lives in a cone on red and soft maple. The species seems fairly common but is usually taken for something else.

Ontario to Connecticut; New York; Missonri. New York: Ithaca.

14. G. species A. Similar, much smaller; first golden spot narrow, barely reaching fold, crescentic, followed by four successively smaller, golden dots on costa, the last practically apical, and each defined on the outer side with a black line fringe mostly blackish, with the usual darker lines. 7-8 mm.

Moth in September and October. Larva on maple.

New York; Missouri.

15. G. vacciniella Ely. Dark purplish, including vertex and upper part of face; palpi with brown tips; antennæ annulate, as usual in the genus; fore wing with two spots, the first triangular, with its basal margin rounded and its outer margin perpendicular, and reaching to fold; outer spot separated from the first by about its width. Base of inner margin yellow. Fringe smoky gray, dark at apex and with obscure lines in it. 11 mm.

The moth flies at the end of July. The larva feeds on Vaccinium. Pennsylvania.

G. anthobaphes Meyrick is similar, but with a small additional golden spot at anal angle. It occurs at Lake Muskoka, Ontario, in July and August.

16. G. belfrageella Chambers. Purple. Face and palpi white; a dark dot on tip of second segment of palpi; costal triangle typically pale golden, truncated on the fold, and extended as a wide band along the costa to the beginning of the fringe; base of inner margin dark. 11 mm.

Rarely the costal patch is cut into two separate spots.

The larva feeds on *Cornus asperifolia*, at first in a linear winding tract on the under side of a leaf, then in a blotch, which is erinkled later; finally in the usual cone. It pupates in a fold of the leaf. Other specimens, which roll the whole leaf in a long cylinder, appear to make the same moth, but more often with separate spots. This name is often misapplied to one or another of the maple species.

Southern Ohio; Texas.

17. G. coroniella Clemens. Dark yellowish, overlaid with crimson; head dark yellowish; palpi with third segment dark on outer side. Fore wing with a triangular patch at middle extending only to fold, and not running out on costa; a couple of dark points on costa within it, and a small pale spot on costa beyond it; apex pale; fringe yellowish, tipped with black. 10 mm.

Larva on birch.

Illinois.

18. G. packardella Chambers. Light orange with strong crimson iridescence, strongly variable in brilliance; antennæ ringed with golden and fuscous as usual in the group; occiput golden, face silver white; fore wing with a nearly equilateral, golden triangle, reaching nearly to the inner margin; and inner margin at base shaded with pale golden. 12 mm. (*inornatella* Chambers, in part; *elegantella* Frey.)

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Larva on sugar maple. G. packardella has been confused with G. alchimiella, which has an entirely differently shaped patch.

Generally distributed. New York: Ithaca. 19. G. purpuriella Chambers. Dull grayish brown with purple iridescence, the face typically white below, but often only slightly paler; labial palpi blackish on outer side, except at extreme apex, dirty white within. Fore wing with base of inner margin concolorous, the triangle white, often scaled with fuscous, and variable in size, but normally narrow with concave sides, ending in a point near or above the fold; costa outwardly with faint pale and dark striæ. Apical fringe gray, barred with black. Tarsi with more dark scaling than usual. 12 mm.

This form is general in distribution; and is common in August, and from October to April. The caterpillar feeds on willow and poplar, rolling the leaf down from the tip, and sometimes using a whole leaf. The pupa is usually formed in the roll, rarely in a web outside. Ely considers this form conspecific with G. stigmella, but they appear distinct to me.

New York: Ithaca.

20. G. stigmatella Fabricius. Closely similar, but with paler reddish ground. Face on the average more extensively pale; triangle with very little dark scaling, and relatively larger, not pointed on the fold (upupæpennella). Larva on willow. Moth in July.

Wales, Maine; Framingham, Massachusetts; Cincinnati, Ohio; New Brighton, Pennsylvania; Europe. Possibly introduced in America. 21. G. negundella Chambers. Light tawny brown, suffused with crimson and

greenish; the costa patch not at all contrasting in light specimens. Palpi dirty yellowish white, with tips of joints blackish; antennæ brown, obscurely annulate. Head and thorax dull, and rather greenish on the whole; face pale below. Fore wing with costal patch large and irregular, extending nearly to apex, as in typical belfrageella; apex with obscure yellow spots; fringe with three blackish stripes. 13 mm.

The palest specimens show brownish spots in the ground, especially toward the Eastern specimens are darker than the types (from Colorado) and show apex. black dusting on the palpi, and the basal half of the costal patch has some black

edging. 13 mm. July and August; October to early spring. Caterpillar at first in a narrow. linear mine on under side; then erossing to upper side; then forming a large, whitish blotch; and finally, the usual cone: on Acer negundo.

Ohio and west (probably also eastward where the food occurs.) 22. G. azaleæ Busck. Head, thorax, and base of fore wing, except on inner margin, dark purple; costal half of fore wing golden almost to apex, dorsal margin tawny with crimson iridescence; the boundary usually diffuse. 10 mm.

This species is a green-house pest on evergreen azaleas, and has apparently been introduced from Japan via Holland. It has been confused with *zachrysa* Meyrick, but appears distinct. The latter is an Indian species feeding on apple.

Massachusetts. New York: Rochester, Yonkers. 23. G. ostryæella Chambers. Autumn form: Antennæ gravish, annulate with dark brown; palpi yellowish white, the second and third joints with broad black apical bands; maxillary palpi similar. Face golden below, more or less scaled with brown and sometimes with a brown cross-line. Fore wing pinkish brown. speckled with patches of blackish scales, with a more or less distinct elongate patch near anal angle, leaving a few golden scales on the margin. Pale costal triangle dark-margined and separated from the patch beyond the triangle; golden. suffused with dark except at its boundaries on costa and inner margin; outer spot more or less suffused. 10 mm.

spot more or less suffused. 10 mm. The larva feeds on Ostrya, in a linear, whitish mine on the upper side of a leaf; then in a whitish, digitate blotch over a vein, which it eats out transparent before

deserting it, leaving a network of brown veins. Finally the larva forms the usual cone.

Southern Ohio; Kentucky.

Supposed summer form: Antennæ ochreous, broadly annulate with dark; darker toward the apex; palpi yellowish white; third segment annulate with dark just before the apex; maxillary palpi yellowish white. Face, head, and thorax pale golden; vertex bronzy; fore wing suffused with purplish bronze; a pale golden patch at base; triangle pale golden, large, truncated on fold, extended out on costa, with very slight, dark points on the costal edge. Hind wing fuscous with

reddish fringe. Hind tibiæ and tarsi mostly yellowish white. 10 mm. The larva lives in a mine on the *under* side of Carpinus and Ostrya, the blotch when finished becoming similar to that of the autumin form, as the parenchyma is eaten out.

24. G. superbifrontella Clemens. Rose-violet, iridescent on a base of tawny yellow; palpi yellow with brownish tip; antennæ dull yellow, vertex shaded with reddish violet; thorax wholly golden. Hind wing very dark. Fore wing with large, irregular, costal patch, and base of inner margin suffused with golden. 11 mm. (alchimiella of authors).

June to August. Larva in a conical roll on Hamamelis; pale green with pale brown cervical shield.

Distribution general. New York: Crosby (Yates County); Ithaca. G. alchimiella Scopoli, a European oak species, has been reported from Essex County, New Jersey. The golden spot on the inner margin is more sharply defined; the head is lighter crimson rose; and the tegulæ and a spot on the disc of the thorax are tawny. Packardella has sometimes been determined as this species.

25. G. blandella Clemens. Dark purple; face yellowish; vertex with purpletipped scales; antennæ strongly annulate; palpi yellowish with a brown spot on outer side of second joint, and on tip of third. Fore wing with usual patch very large and irregular; the wing strongly golden-iridescent at base, sometimes forming a central, diffuse, basal spot. Fringe dark with a pale line in the middle; hind wings pale fuscous.

This species is unknown to me.

Virginia.

26. G. juglandivorella Chambers. Closely similar to G. blandella; joints of maxillary palpi purple-tipped; vertex largely purple; thorax with three lemonyellow stripes; yellow patch perhaps less extensive. 8 mm.

Larva on black walnut; at first in a linear mine, reaching an inch long, then under a series of small flaps, eating the parenchyma, and pupating under the last flap. The larva changes to the cylindrical form on leaving the mine.

I have not seen this form. The preceding species and this probably identical; and are so considered by Ely.

Virginia.

27. G. glutinella Ely. Summer form. Reddish bronze, face yellow; palpi straw color, shaded with dark brown just before the apex, darker outwardly; antennæ Fore and middle femora and tibiæ reddish bronze; hind tibiæ darkannulate. shaded at apex. Fore wing with some straw-yellow scales, especially toward apex of costa; triangle shining golden, truncated shortly on fold; no outer spot. Fringe gray, with two dark lines around the apex. 12-13 mm.

July-August. Larva in a roll on Alnus glutinosa, in July.

Southern Connecticut.

Supposed winter form: Ground dark purplish, intermixed with straw-colored scales; face pale yellowish, edged with brown on sides; vertex darker and more straw colored; palpi shaded throughout with dark purple, third segment with a heavy black ring at the tip; maxillary palpi also shaded with purple. Antennæ

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darker. Fore wing with costal triangle pale straw yellow, almost obliterated with dark purple suffusion; fringe gray, almost concolorous at apex with two dark lines. 13 mm.

The moth is distinguished from coroniella by the darker coloring. It flies in September; doubtless wintering, and reappearing in April.

The larva occurs on alder in August, at first on the under side of a leaf, in a linear mine; which ends in a small blotch, which is usually near the margin; then later in one or two shelters formed by turning down the edge of a leaf; and finally, in a cone at the apex of the leaf. The cocoon is formed outside the mine.

28. G. atomosella Zeller. Thorax and fore wing heavily dusted and shaded with blackish, on a pale ochreous ground; fore wing with pale striæ toward costa and inner margin; one of these, before the middle of the costa, being the most distinct marking, and extending obliquely out almost to the inner margin. Base darker. Head white, somewhat tufted behind. Palpi black with two white rings on second joint, and base of third joint white. Maxillary palpi with white dots. 14 mm. (atmosella Elv — misprint).

This species is unknown to me.

Texas; "Atlantic States" (Ely).

II. Palpus with a triangular tuft on under side of second segment (Coriscium).

29. G. cuculipennella Hübner. White, with oblique fasciæ formed of blackbarred scales, and mixed with brown scales, which concentrate at the edges of the fasciæ, forming brown lines; base powdery gray; basal half of wing with two fasciæ, extending obliquely outward, the first heaviest on inner margin, the second stronger, and usually ending just above the inner margin; a similar medial fascia. Outer half of wing with diffuse spots and short vertical fasciæ; fringe with dark-tipped scales, gathering in three or four lines. Head, thorax, palpi, and tibiæ powdery gray, barred with white, the tarsi hardly lighter. Inner side of palpi white. 10-14 mm.

Larva on privet.

Europe.

American records probably apply to fraxinella, which may be only a race of this species.

30. G. fraxinella Ely. Similar. Powdery gray areas more diffuse, the scales mostly finely white-tipped; usually with the fasciæ broken up, and obscure; rarely with fasciæ contrasting and black, but broken into spots. Fringes more suffused with gray than in G. cuculipennella; head and thorax darker. The white ground color is reduced to a large number of spots, many of them rounded. 12-14 mm.

Moth in July and May, the latter doubtless specimens which had emerged the preceding fall. Caterpillar on ash and apple, in the usual cone, in June. This may be a synonym of G. paradoxa Frey and Boll.

Connecticut. New York: Ithaca. 31. G. paradoxa Frey and Boll. White, heavily and rather evenly dusted with inely black-tipped, clay-colored scales, leaving a white antemedian fascia across the wing, and obscure, alternately light and dark mottlings along the margins outwardly. Head rather more lightly dusted, and palpi more heavily, except the tip of the second segment and base of the third. 12–15 mm. (Parectopa.)

Moth in April and early May. Larva unknown. Boston, Massachusetts; New Brighton, Pennsylvania. 32. G. quercinigrella Ely. Similar to G. fraxinella, powdery gray. Palpi only lightly dusted; fore and middle tible red-brown; tarsi white, ringed with redbrown; hind legs yellowish; the femora and coxæ shaded with red-brown. Forc wing with markings diffuse; base much paler below fold than above; anteme-dial fascia oblique outward, pale gray; outer part of wing with obscure, concuve fasciae. Under side red-brown. Hind wing pale-gray, partly red-brown below. 10 mm.

The moth flies in September. The cone is formed by bending down and under a lobe of the food leaf (red oak, etc.). The leaf is eaten through in a series of small holes where the edge of the flap is attached. The larva is whitish with brownish incisures, and occurs early in July. Apparently there is only one brood.

Southern Connecticut.

33. G. serotinella Ely. Palpi with a very small pointed tuft. Ground blackbrown; basal half of third segment of palpi yellow; tarsi pale. Fore wing with numerous, greenish-silvery spots, irregularly scattered and more or less fused, especially so as to form an outwardly oblique sub-basal fascia and a V-shaped antemedial spot resting on costa (enclosing a spot of the ground color). Apical fringe cut with four yellow bars. Dorsal fringe and hind wing mouse gray. 15 nm.

The moth occurs in July and August. The larva forms the tip of a leaf of *Prunus serotina* into a cone. The ecocon is boat shaped, flat on top, and is spun in a partly folded leaf.

Dublin, New Hampshire; East River, Connecticut. New York: Ithaca.

3. ACROCERCOPS Wallengren

(Gracilaria, Coriscium, in part)

Similar to Gracilaria; hind tibiæ of the northeastern species with a couple of rows of stiff straight bristles above, about a quarter as long as the length of the tibiæ, or somewhat less. Antennæ fully as long as the fore wings, palpi typically with a slight tuft on second segment, large and triangular in A. quinquestrigella; maxillary palpi minute in some exotic species. Wings about as in Graeilaria; in the fore wing with all veins present, or one dorsal (apparently Cu) lost, as in A. *onosmodiclla*. Hind wing often a little more reduced than in Graeilaria, \mathbf{R}_1 regularly being lost, and \mathbf{M}_3 also in A. *strigifinitella*. Resting position with the fore and middle legs displayed as usual, but sometimes with the head appressed to the object on which the moth is resting.

The caterpillar is of various types, but in all cases it feeds in the cylindrical stage (fig. 121). The transformation to the cylindrical stage is sometimes gradual. The larva are usually blotch miners, the cocoou, typically, being formed in the mine. Our species belong to various groups of the genus, which is very large in the Orient.

Key to the species

- 1. Inner margin with silvery spots or fascia only.

 - Costal and dorsal series at right angles to each other.....l. strigifinitell.
 Stripes running across the wing, farther cut on inner margin....3. strigosa.
 Moderate white fascile and no black ones.

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Maxillary palpi very small, less than a fifth as long as labials; palpi smoothly scaled; spining on hind tibiæ moderate. M₃ of hind wing lost. Larva changing gradually from flat to cylindrical type (Neurobathra Ely).

1. A. strigifinitella Clemens. Palpi yellowish white, barred with brown; head and antennæ dull brown; fore wings gravish brown, striate obliquely from both margins with fine white striæ, edged with black, alternating with heavier solid black ones. An apical ocellus. 8 mm. (*Dialectica* Walsingham; Ornix quercifoliella, Gracilaria duodecimlinella Chambers.)

The larva feeds on chestnut, chinquapin, oak, and beech. In the first two stages it is of the flat type, and makes a linear mine; the third stage larva is cylindrical, with an essentially normal head, but with rudimentary legs and the head flatter than it is later. It bores in a lateral vein. The fourth and later stages are of the normal cylindrical type, and bore in the midrib. If the supply of food in the midrib fails, the late larva forms a large, putfy blotch-mine, with scattered frass. There is no non-feeding stage. Pupation is outside the mine, in a white cocoon decorated with four separate masses of froth. The species breeds continuously all summer.

New York to Missouri. The species is in the Robinson collection, presumably from this State.

11. Maxillary palpi a fourth as long as the labials, which have a more or less distinct triangular tuft on the second segment; larra (so far as known) changing abruptly to final stage; tibial bristles variable. Fore wing with traces, at least, of all dorsal veins (Acrocercops).

2. A. astericola Frey and Boll. Lutecus. more or less dusted with brown; head, with its appendages, white. Fore wing with a broad, oblique, more or less triangular, white fascia two-fifths way out, edged, especially inward, with black; not reaching inner margin; two others, half as wide, beyond it, which have a heavier black edging. Apex white, preceded with black and followed by a double bar in the fringe; the outer fascia reaching the inner edge of the membrane. A short antemedial costal fascia, and two on inner margin. 9 mm. (*Gracilaria, Parectopa*.)

May, August.

The larva forms a large, tentiform blotch mine on Aster corymbosus etc.; the pupa is outside the mine, in a firm cocoon.

Massachusetts to western Pennsylvania. New York: Ithaca.

3. A. strigosa Braun. Dull brown, with seven slender, parallel, outwardly oblique, white fasciae across the wing, more or less edged with black, and somewhat broken. 10 mm.

The moth occurs in June. The larva is found on *Quercus prinus*, at first in a narrow white gallery, then in a large, whitish blotch, from which the parenchyma is gradually eaten out. The cocoon is dense and brownish, with the usual globules. The larva is full grown at the end of May.

Tryon, North Carolina; Powell County, Kentucky.

4. A. (?) boreasella Clemens. Fuscous, with nearly round, white, postmedial and subterminal spets on costa, and medial and subterminal ones on inner margin, the last extending into the fuscous fringe. 9 mm. (Ornix ? Clemens, Gracilaria ? of authors.)

Labrador.

5. A. onosmodiella Busck. Golden brown; head, therax, and appendages for the most part silvery; antennæ dark with silver line; fore wing with markings heavily black-edged; a large, antemedial crescent on costa, followed by three triangles; two silver triangles on outer part of inner margin, alternating with two costal ones; base of inner margin and apex silver white. Palpi wholly white. 9 mm. (pnosmodiella Busck, lapsus calami).

July. The larva a blotch miner on Onosmodium, of the family Boraginaceæ. District of Columbia to California.

6. A. quinquestrigella Chambers. Palpi white, second segment with a large, triangular tuft, like that of Coriscium; two dark rings on the third. Fore wing fuscous brown; inner margin narrowly and evenly white, with some dark scales, running out at anal angle; with paired, whitish streaks running obliquely out from costa and from upper edge of the dorsal streak, the outermost ones in the base of the fringe, and meeting at a right angle over the apex; these streaks are slightly edged with darker brown, and have dark streaks between them. 5 mm. (*rhombiferellum* Frey and Boll).

Kentucky; Texas.

7. A. albinotella Chambers. Head, palpi, antennæ, and fore wing silvery white; antennæ annulate with brown; palpus with second joint tufted, brown-gray, with white tip. Fore wing shining, light olive brown; thorax with a silvery, central band continued as a band on the inner margin of the wing, gradually narrowing to the anal angle, and interrupted by projections of the brown area; three white, outwardly oblique, costal fasciæ, the first usually broken, and joining the white dorsal area; a white apical spot covering the costal fringe, and edged below with a black crescent; apical fringe brown, vaguely banded; dorsal fringe shaded with white. 9 mm.

Moth in April and June, and again in August. Larva in a large, tentiform mine on under side of leaves of oak; at first in a linear mine. (Coriscium albinatella, albonotella Chambers.)

New York to Maryland, Ohio, and Missouri. "New York" (Beutenmuller).

III. Fore wing with one dorsal vein completely lost; R₄ and R₅ stalked (Leucospilapteryx Spuler).

S. A. venustella Clemens. Head and eye-cap silvery white; palpus with two blackish spots; antennæ dark brownish. Fore wing dark, purplish ash gray, with a white streak along inner margin from base half way to apex; a small, white, antemedial, costal spot, and three somewhat convergent white streaks beyond, equidistant at costa, and extending obliquely outward. Apical spot white, with dark margin, and containing a black dot. Fringe mouse gray, mixed with white. 8 mm. (eupatoriclla Chambers).

The moth occurs from late July to September. The larva makes several, successive tentiform blotch-mines on Eupatorium. Apparently the same species occurs on Ambrosia artemisiæfolia but I have seen only a rubbed specimen. Possibly it is A. astericola.

Cincinnati, Ohio, and District of Columbia to Pennsylvania, Kentucky, and Missouri.

4. APOPHTHISIS Braun

Similar to Acrocercops; antennæ rather shorter than fore wing, with a pecten; maxillary palpi rudimentary; bristles on hind tibia rudimentary, rather like stiff scales. Fore wing with \mathbf{R}_1 weak, \mathbf{R}_5 and \mathbf{M}_1 stalked, forking over the apex; \mathbf{M}_2 and \mathbf{M}_3 absent; hind wing as in Acrocercops.

The resting position is about as in Ornix.

1. A. pullata Braun. Dirty white; scales fuscous-tipped; head and palpi gray, antennæ lightly annulate. Fore wing with darker streaks near base and beyond middle of fold; a faint, darker line in fringe. $5\frac{1}{2}-6$ mm.

middle of fold; a faint, darker line in fringe. $5\frac{1}{2}-6$ mm. The moth has been taken late in July and in May. The larva is a miner in buckthorn, in July and October; the mine at first is obscure and linear, and later becomes a large blotch, 5-8 mm. wide. Pupation takes place outside the mine, in a flat, oval, yellow cocoon.

Cincinnati, Ohio.

5. LEUCANTHIZA Clemens

Front and vertex smooth, a small, loose tuft on occiput; antennæ as long as fore wings, basal joint small, not modified; palpi short, straight, drooping; maxillary palpi rudimentary; tongue short, apparently naked, hind tibiæ with long bristly hair (?). Fore wing lanceolate (fig. 118), with four traceable veins running to costa, \mathbf{R}_1 being lost; two or three veins from cell to inner margin. Hind wing with **R** and **M**₁ closely parallel, but without forked veins.

R and **M**, closely parallel, but without forked veins. Caterpillar of Gracilariid type, forming a large blotch-mine on upper side of a leaf. Cocoon of white silk, outside the mine. At rest the moth holds the antennæ extended laterally, as do other members of the family. This is an aberrant genus, which has not been fully studied, and may be misplaced in the keys. The short palpi may cause it to be sought among the Lyonetiidæ; and there may be a real kinship to Phyllocnistis.

Key to the species

Base of inner margin brown......amphicarpexfolicIla. Base of inner margin golden.....dircella.

1. L. amphicarpeæfoliella Clemens. Head lead color; antennæ golden brown, tips silvery white. Fore wings deep orange, shading to golden brown toward the apex; base deep brown, bordered by a strongly excurved, lead-gray line from costa to basal angle; a large, irregular, triangular, golden patch on middle of inner margin, extending up nearly to middle of wing; a golden streak at beginning of costal fringe, with two dots before it; fringe golden brown; hind wings gray. 6 mm.

May. Larva in an upper surface mine, on Amphicarpa monoica; green with brown dorsal, and darker ventral, spots, after the last molt becoming wholly green. Connecticut; Maryland; Pennsylvania.

2. L. dircella Braun. Orange yellow; head golden with darker hairs behind; antennæ dark brown, tips silver white; thorax golden, dark brown in front. Fore wing with base dark brown, except toward inner margin; apical area dark brown; antemedial golden fasciæ straight. Base of inner margin orange; an oblique, curved golden streak from antemedial region below costa to middle of costa; a small, costal streak at beginning of fringe; a short, golden, dorsal streak margined toward base by an oblique line, dark brown scales; a longer oblique streak at anal angle, and a faint, golden, marginal line. Fringe and hind wing dark brown. Legs golden with dark brown tarsi. 5-6 mm. July. Larva in late June and in September, in a large, sometimes digitate, upper-

July. Larva in late June and in September, in a large, sometimes digitate, upperside blotch, on *Dirca palustris*; often social. Cccoon outside the mine.

Clermont County, Ôhio.

6. MARMARA Clemens.

(*Æsyle* Chambers; *Gracilaria*, in part)

Head smooth; antennæ with pecten scaly and suggesting an eye-cap, or normal; palpi moderate, smooth-scaled, ascending in life but usually porrect when dry; maxillary palpi moderate, porrect; tongue naked. Tibiæ smooth-scaled. Fore wing (fig. 117) with only five veins arising from the cell, three running to the costa and two to the inner margin, all free. Fringe as usual in the family. Hind wing linear-lanceolate, with fringe four times as wide as membrane; Sc and R obscure; a forked stem in middle of wing, free from Rs; Cu simple.

The larvæ are of the flattened type in all the feeding stages, becoming cylindrical just before pupation. They are extremely broad at the anterior end and lobed at the sides. They are leaf miners, or miners in the bast of twigs, usually forming a tract. The cocoon is spun outside the mine and is covered with more or less confluent masses of pearl-like, white bubbles. The genus is only definitely known from North America, but has been generally confused with Gracilaria. Besides the

species mentioned in this work, there is an undescribed species of the light group, on elm; and one of the dark group, on Hibiscus.

Key to the species

1. Dominantly pale.

2. A large, basal, blackish patch on costa.....l. fulgidella.

1. Dominantly dark.

- 2. Second fascia erect, complete, as wide as the dark space before it...4. fasciclla.
- 2. Second fascia outwardly oblique, bent, and half as wide as the dark space before it, or broken.
 - 3. First fascia twice as wide on inner margin; the second offset its width

3. First fascia even in width or broken.

- 4. Brilliant, golden brown, with silver markings.....9. auratella. 4. Dull fuscous brown, hardly shining.
 - 5. Ground powdery, the pale scale-bases showing; palpus with heavy
 - 5. Ground even, palpi with third joint pale.

6. Ground nearly black.

- 7. Fasciæ complete or nearly so.
- 8. Head silver white......3. salietella.

6. Markings edged with black, on a lighter brown ground.

4. quinquenotella.

1. M. fulgidella Clemens. Head and antennæ vellowish white. Maxillary palpi dark. Fore wing silvery white with a dark brown blotch at base, not extending across the wing: a dark brown band rather below middle, sharply angulated just above the inner margin, and darkest along its outer edge; outer part with two broad, costal fasciæ confluent in the middle of the wing, with a white costal spot between them, and a white spot on inner margin opposite. Apex dark, with a white costal streak before it. Hind wing dark fuscous. 71/2 mm.

Larva on Quereus prinus; with the habits of scrotinella.

Pennsylvania.

2. M. elotella Busck. Paler than M. fulgidella; moth mostly white: antenne yellow. darker toward the tip; palpi with black bars; tibiæ mostly white. Fore wing white, with yellow-brown bands edged ontwardly with black; the first costal spot slight and tending to break up, the two outer fasciæ markedly separate, diverging in the dorsal fringe, and followed by an apical dot. Hind wing clay color.

Larva under bark of apple twigs, leaving a winding scar. South Connecticut. There is a closely similar, possibly the same, species on elm in Virginia.

3. M. salictella Clemens. Antennæ gravish fuseons; head and palpi white; occiput (as in the following species) fuscous. Maxillary palpi dark; second joint of labials touched with fuscous. Fore wing dark gray-brown, with silvery-white markings; a fascia at basal third; and a somewhat oblique one at middle, slightly excurved and sometimes broken at middle: with two costal spots and one or more on inner margin beyond it; the outermost spot sometimes forming a bar across the apical fringe. Fringes paler, whitish at apex; hind wing gravish fuscous. 6 mm. June, July.

The caterpillar forms a long, linear mine under the bark of willow twigs in the

fall, which at that time is inconspicuous. In the spring it is revealed by the scarring of the bark over it. The larva in early spring is pale vellow; later developing

transverse red stripes and two dark cervical spots. Pupation is outside the mine. Occasionally this species is injurious by girdling the twigs and small branches.

Connecticut to Virginia.

4. M. fasciella Chambers. Similar to M. salictella; ground lighter, and distinctive brown; each fascia black-edged on the inner side, the first two fascia equal, and as wide as the space between them; third fascia sometimes broken; second tascia straight; maxiliary palpi pale. (*Esyle* Chambers, *Acrocercops* of authors.)

Quinquenotella Chambers is generally considered an aberration of this species. In it the second fascia is broken into squarish costal and dorsal spots, the dorsal one lying farther out than the costal.

5. M. opuntiella Busck. Paler and more mottled than salicitla; distinctly blackpowdered on a whitish ground; first fascia obscure, second only nearly complete; palpus with a black band on third segment; maxillary palpus pale yellowish.

This species was described from Texas, but probably occurs as far north as the prickly pear, its food plant. The larva makes a long, irregular, winding mine under the epidermis of Opuntia.

6. M. smilacisella Chambers. Similar to M. salictella, but with the silvery markings more restricted, even the antemedial fascia being sometimes reduced to a bar on inner margin. (Phyllocnistis Chambers.)

The larva makes a very complexly winding and anastomosing linear mine with a central frass-line on leaves of Smilax hispida and glabra. The mine is about 2 mm. wide.

Cincinnati, Ohio; southern Kentucky.

7. M. serotinella Busck. Ground dark brown, slightly shiuing, not powdery. Similar to M. salictella, but with the first fascia nearly as wide as long, and much widened on the inner margin, with a sharply bent outer boundary; twice as wide as second fascia. Fasciæ not black-edged.

Larva on Prunus serotina (type only seen). M. pomonella, a dark species with small alternating spots only, similar to opuntiella, is a western species feeding on apple; and is to be looked for in the north of our territory.

8. M. apocynella Braun. Head whitish on face, more yellow above, fuscous, as usual, on vertex; palpi whitish, second joint tipped with fuscous; antennæ grav. Fore wings black; antemedial fascia straight; an oblique fascia at middle, slightly angulate above inner margin; a larger costal and a minute dorsal one two-thirds way out; a white, costal streak before the apex; tip of fringe whitish. Legs black, annulate with white; four outer segments of tarsi mostly white.

Larva in a long, whitish, serpentine mine on Apocynum cannabinum; cocoon typical, with bubbles. Larva in early July, the moth later in the month.

The moth is smaller than salictella, with a yellower head and more oblique median fascia.

Southern Ohio.

9. M. auratella Braun. Golden bronze; face metallie lead-gray; including antennæ; tips of both palpi whitish; fore wing with brilliant, silvery markings; antemedial fascia a fourth way out, wider below fold; an oblique costal streak halfway out, and a more erect one just beyond it on inner margin; triangular costal and dorsal streaks two-thirds way out, nearly meeting; and a fine streak across wing near apex. Apical fringe white. 61/2 mm.

There are two broods, in early June and in early August. The larva is a bastminer in Rudbeckia laciniata (so far as known, only in the cultivated golden glow) and in Dahlia. The mine is long, serpentine, usually tending downward, in the lower part of the stem. The cocoon is spun in a flap loosened from the cover of the end of the mine, opening not directly outward but by a silk-lined tunnel through the tissues of the flap; it is without pearl globules. This species possibly was introduced, with its food, from Mexico.

Southern Ohio.

7. PARECTOPA Clemens

(Gracilaria, Coriscium, in part)

Similar to Gracilaria; the vestiture of the vertex often raised, but without a definite tuft; middle tibiæ with at most a small tuft at apex; hind tibiæ smooth-scaled. Venation of fore wing slightly variable, one or two veins sometimes lost. Hind wing with \mathbf{M}_{a} lost; \mathbf{M}_{1} and \mathbf{M}_{2} and \mathbf{Cu}_{2} stalked. Larva, when mature, of the cylindrical type; the northeastern species making a digitate blotch-mine, but some of the primitive species in a cone, like Gracilaria.

Key to the species

- - 2. Less black in apical fringe, otherwise marked; face white. 3. Most of vertex behind antennæ light brown, white at edges; ground of fore

2. robiniella, 3. lespedezæfoliella.

I. Hind wing with traces of \mathbf{M}_3 preserved; fore wing with $\hat{\mathbf{R}}_1$ arising from near base of cell; \mathbf{R}_5 and \mathbf{M}_1 stalked, and all veins preserved. (Micrurapteryx Spuler).

1. P. salicifoliella Chambers. Apical fringe caudate, but with the tuft not contrastingly colored. Head and thorax white, with some gray scaling on head, especially on the two fanlike tufts over the bases of the antennæ; palpi with second joint mostly blackish; third joint with black rings at base, middle, and apex; maxillary palpi blackish. Legs black and white barred. Tegulæ and fore wing dull dark fuscous, lighter toward the margins. Two strongly oblique, white streaks at middle of costa, and three short, strongly convergent ones toward apex. Apex black, with a blackish bar around it in the fringe. Inner margin with a somewhat irregular white fascia, edged above with black, and sometimes containing a black bar near its upper edge, two-thirds way out, and scattered black scales. 10 mm.

The species occurs in the latter half of the year, apparently breeding continuously. The larva is a blotch-miner on willow, sometimes eating out the entire leaf, and is usually confined to a single mine. The pupal stage is passed under a silken carpet, on a leaf, and lasts two weeks.

Kentucky; southern Ohio; Missouri.

II. Hind wing with \mathbf{M}_{3} lost; fore wing with \mathbf{R}_{1} arising a third way out on cell; \mathbf{M}_{1} free and one or two dorsal veins lost. (Parectopa.)

2. P. robiniella Clemens. Dark brown, somewhat shining, but less so than *penn-sylvaniella*. Head white, occiput black, the black area covering the whole width of the head. Palpi blackish, third segment white; antennæ with scape white, with a fine black line in front, shaft blackish, and feebly annulate. Legs banded, black and white. Fore wings with three white fasciæ strongly outwardly oblique from costa toward middle, the first one shorter; an antemedial, white, dorsal spot or fascia, not always reaching inner margin and nearer the base than the first costal; a short oblique fascia before the second costal, and one nearly opposite

the third; a fine, vertical, white stripe across the apex, usually interrupted in middle. Fringes white, the costal and dorsal each with two black lines, which converge to the apex. Apex black, but not caudate; first dorsal line interrupted just below the apex.

Larva in a digitate mine on locust. *

Pennsylvania to Missouri; New York: Ithaca (mines not rare).

3. P. lespedezæfoliella Clemens. Similar; first costal fascia absent; dorsal fasciæ reduced to spots (mirabilis Frey and Boll).

Larva on Lespedeza and perhaps Desmodium, marking several small, digitate mines; the frass thrown outside through a hole on the under side. Cocoon white, outside the mine, often on the ground.

Pennsylvania.

4. P. pennsylvaniella Engel. Rich golden brown; head white; vertex light brown, white above eyes; palpus white, with most of second segment and a spot on third brown; apex of second segment white; maxillary palpi dark, with terminal joint white. Scape of antenna brown above, white below, without fine stripes. Fore wing with three strong silvery fasciæ on costa, outwardly oblique; the first fascia hardly if at all shorter than the other two; and a short, triangular, white spot before the apex. Dorsal margin with a strong, nearly longitudinal dash from near base to fold a quarter way out; two oblique fasciæ before and beyond middle, and a white bar in fringe opposite the last costal spot. All fasciæ edged with black. A black and blue ccellate spot just below and before apex. Fringe with a strong black hook in the apex, with a white area below it, preceded by a black line across the extreme apex of the membrane; dorsal fringe below the white patch gray, not barred. 9 mm.

May, August, October. Larva on Aster cordifolia; the mine large and inflated when mature.

Connecticut to western Pennsylvania.

5. P. plantaginisella Chambers. Golden brown, lighter than P. pennsylvaniella, the markings more heavily black-edged. Head bronze-brown, with a little white above the mouth and along the eyes; palpi bronze on outer side, mostly white within; thorax as in the proceeding species. Fore wing with the three costal fasciæ almost half as wide as the space between them; a subapical silver dot, with rascize almost half as wide as the space between them; a subapical silver dot, with a white bar in the fringe opposite it; inner margin with basal dash in fold; a short silver bar at middle, and a longer one three-fourths way to apex, with its tip almost touching the third costal one. A strong silver subterminal spot, opposite the tip of, the third costal, continued by a white bar in fringe. Apical ocellus with a large silver center. Fringe black, strongly caudate, with a white triangle below the apex. 7 mm. (erigeronella Chambers, geiella Chambers). September to November. Larva in a yellowish mine, which is at first slender and frass-filled, but later becomes a large inflated blotch; on Erigeron and Plan-targo.

tago. Pupa on the ground. Kentucky; Missouri.

8. CREMASTOBOMBYCIA Braun

Annette F. Braun

Face smooth, vertex rough-tufted. Labial palpi moderate, porrected or drooping, pointed. Maxillary palpi minute. Antennæ about as long as the wings; basal segment thickened; with slight pecten. Fore wings (fig. 119) lanceolate; \mathbf{R}_1 and \mathbf{R}_n . beginning unkerhed, with sight peterin. For which single the mass $(\mathbf{m}_1, \mathbf{m}_2)$ matching \mathbf{M}_1 absent, \mathbf{M}_1 absent, \mathbf{M}_2 and \mathbf{M}_3 short stalked; Cu single-branched; 2d A simple. Hind wings about one-half as wide, linear-lanceolate; \mathbf{M}_1 and \mathbf{M}_2 stalked; the base of **M** sometimes distinct; \mathbf{M}_3 absent; Cu single-branched; discal vein absent between Cu and \mathbf{M}_2 . Posterior tibiæ with loosely appressed hairs. The larvæ are similar to those of the cylindrical larval group of Lithocolletis. All of our species mine leaves of Compositæ, forming wrinkled, tentiform mines. They pupate in an elongate, dense, white cocoon sometimes ornamented with longitudinal ridges. This cocoon is suspended inside the mine by two slightly diverging, silken threads at the posterior end, and by either one or two threads at the anterior end.

The markings of the fore wings consist of a more or less distinctly developed basal streak, four white costal streaks, and two or three dorsal streaks. There may be one or two more or less distinctly angulated fasciæ, formed by the confluence of opposite streaks.

Key to the species

A. Basal streak distinct, reaching almost to, or beyond, the basal fourth.

B. Basal streak narrow, pointed, unmargined......l. solidaginia. BB. Basal streak dilated posteriorly......2. ambrosiella. AA. Basal streak minute or wanting; markings silvery......3. ignota.

1. C. solidaginis Frey and Boll. Fore wings reddish ocherous; a narrow, whitish basal streak below fold; costal, oblique streak at one-third, oblique costal and dorsal streaks at one-half, usually meeting in middle of wing; two posterior costal, and one posterior dorsal, streak, all margined outwardly with blackish scales. 7–9 mm. Underside miner in leaves of golden rod, *Solidago* sp.

Distributed over the entire United States.

2. C. ambrosiella Chambers. Fore wings reddish orange; a short, posteriorly dilated, white basal streak; an oblique costal streak at one-fourth, a more or less angulated median fascia; two posterior, costal, and one posterior dorsal, streak, all dark margined externally. 5.5-6.5 mm. Mine on underside of Ambrosia and Actinomeris; not projecting conspicuously on upper surface.

3. C. ignota Frey and Boll. Distinguished from the preceding by the silvery luster of the markings and the very short, indistinct basal streak. 6.5-7.5 mm. Tentiform mine on under surface of leaves of Actinomeris, Helianthus, and other Compositæ. Conspicuous as a tubercular swelling on upper side.

9. LITHOCOLLETIS Hübner

Annette F. Braun

Face smooth, crown rough-tufted. Labial palpi porrected or drooping, moderately long, pointed. Maxillary palpi rudimentary. Tongue moderate. Antennæ nearly attaining the wing length; simple in male; basal joint thickened; bearing a peeten (except in *desmodiella*). Fore wings (fig. 120) lanceolate; \mathbf{R}_1 and \mathbf{R}_4 absent, \mathbf{M}_1 absent, \mathbf{M}_2 and \mathbf{M}_3 coincident; \mathbf{Cu} single-branched; 2d A simple. Hind wings about one-half as wide, linear lanceolate; \mathbf{M}_1 and \mathbf{M}_2 coincident, \mathbf{M}_3 absent; \mathbf{Cu} single-branched; discal vein absent between \mathbf{Cu} and \mathbf{M}_{1+2} . Posterior tibiæ with loosely appressed hairs.

Our representatives fall naturally into two distinct and well defined groups, identical, however, in the structural details of the imago. The first group, comprising those species having a cylindrical larva, agrees closely with the European species in the early stages and in the type of markings of the imago. The second group includes all those species having a flattened larva, and is characterized by a definite type of wing marking.

The following characters will separate the two groups:

Larva evlindrical; white streaks and fasciæ dark margined internally....Group I. Larva flattened; white streaks and fasciæ dark-margined externally

(p. 195).....Group II.

Group I

The larva of the first group is cylindrical or subcylindrical in form, and has, beside the thoracic legs, four pairs of prolegs, on segments 7, 8, 9, and 13. It is usually of a pale greenish or yellowish color.

The mine may be placed on either the upper or lower surface; but in either case, the loosened epidermis is lined with silk, causing it to contract, and thus producing a roomy, tent-like mine. The mine is at first narrow and somewhat winding, but soon spreads out into a blotch, which sometimes includes the earlier, winding part. The outline of the loosened epidermis represents the final size of the mine, no further increase taking place. The mine may be oval or circular, or, in rare instances, nearly rectangular in shape, sometimes limited by two veins. Usually the larva feeds from the circumference inwardly; sometimes it begins at one end and sometimes it feeds irregularly in spots.

With the exception of ostensackenella, which leaves the mine to pupate, the pupa is formed within the mine, and may or may not be enclosed in a cocoon. When not in a cocoon, it is usually suspended in the mine by a thin meshwork of silken threads. When a cocoon is present, several varieties may be distinguished. The cocoon may be rather small, ovoid, formed of frass and silk; or large, loosely woven, semitransparent, occupying sometimes nearly one-half the mine; or an oval ring with ontlines formed of frass.

The fore wings of the imagoes are usually of some shade of yellow or brown, sometimes, however, with the basal two-thirds almost pure shining white. Upon this yellowish ground color, the more or less white markings appear. The transverse markings consist of costal and dorsal streaks, usually curved and oblique, and slightly curved or angulated fasciae, or combinations of these. These streaks, or fasciae, are usually margined with darker scales toward the base; the internal margins of some of the streaks are sometimes lacking. In a few species, some or all of the streaks, or fasciae, may be more or less margined externally also; but in no case is the external margin heavier than the internal one, or is an external margin present, when the internal one is absent. Longitudinal markings, when present, consist of a median streak from the base, sometimes accompanied by streaks along the costal and dorsal margins. The apex sometimes contains a well-defined, regularly shaped, black dot, formed of closely overlapping scales. In other species, these scales are more or less scattered. The hind wings are unicolorous.

Key to the species

A. Ground color of the fore wings pure white, marked with fuscous, irrorated

B. Fore wings dusted with fuscous scales.

C. Costal and dorsal streaks large, conspicuous, and curved backward

26. salicifoliella.

CC. Costal and dorsal streaks narrow, straight, forming angulated fasciæ 44. celtifoliella.

BB. Fore wings not dusted.

C. With an apical dot (indistinct in obsoleta).

D. Without a complete median fascia.

E. One white dorsal streak before the tornus; basal half of wing mostly white.

F. Three white costal streaks.

FF. Four white costal streaks.

G. Basal half of fore wings pure white.....5. clemensella. GG. A golden basai streak from base to near middle.

11. Basal streak margined with dark brown on its lower edge

argent		

F. With a median pale basal streak.

G. Basal streak extended across the fold to the dorsum....9. hageni. GG. basal streak not extended to the dorsum.

II. First dorsal streak short and broad, produced to base

8. albanotella. HH. First dorsal streak extending more than halfway across the wing.

I. A white costal streak from base to one-third.....17. *rileyella*. II. No such streak.

J. Basal streak dark margined toward costa.

KK. Apex of first dorsal streak opposite that of first costal.

L. Head and thorax white......15. olivaeformis. LL. Head and thorax reddish suffron.....18. kearfottella. JJ. Basal streak unmargined.

K. Thorax and basal portion of fore wings shining white 16. caryaealbella.

KK. Thorax and basal portion of fore wings not white 14. ostryaefoliella.

FF. Without a median pale basal streak.

G. Apex of first dorsal streak opposite the apex of second costal.

H. First dorsal streak very large.....l. fitchella. HH. First dorsal streak not larger than opposite costal streak

32. robiniella

- GG. Apex of first dorsal streak opposite the apex of first costal. H. Pale markings very indistinct.....12. obsoleta.
 - HH. Pale markings well defined.
 - I. Apex of second dorsal streak opposite the space between the second and third costal streaks; margins of opposite streaks never unitingll sexnotella.

11. Apex of second dorsal streak opposite second costal; mar-

gins of opposite streaks uniting.....10. *ariferella*. DD. With a complete median fascia.

- F. Dorsal half of the wing below the fold dark brown.

GG. Fascia at one-third; basal streak very short.....36. diversella. CC. Without an apical dot.

D. Oblique costal and dorsal streaks, rarely meeting; no straight fascia; sometimes one acutely angulated fascia.

FF. Fascia, if present, formed by the meeting of opposite streaks. G. With three, long, dorsal streaks before the tornus.

HH. Two costal streaks, first opposite second dorsal streak

28. occitanica.

GG. With two dorsal streaks before the tornus.

H. Basal streak unmargined or margined toward its apex only.

I. Basal streak confluent with the upper edge of the first dorsal streak.

II. Basal streak not confluent with first dorsal streak.

J. First pair of streaks very oblique and extended along the

JJ. First pair of streaks not extended to base..22. scudderella.

HH. Basal streak dark margined toward the costa.

I. Costal margin white from base to near one-third. .25. populiella. II. Costal margin not white from base.

JJ. Four costal streaks.

K. First dorsal streak beginning much nearer the base

than first costal; expanse 8-9 mm. 19. propinquinella. KK. First dorsal streak beginning nearly opposite first

costal; expanse 6.5-7 mm......20. cratægella. DD. Usually two complete fasciæ; if but one, median and nearly straight.

E. A median fascia; two posterior costal streaks.

FF. Head and thorax not white.

G. Two posterior costal streaks.

H. Fasciæ nearly straight.

I. Ground color ocherous..... II. Ground color of entire wing dark, reddish brown

40. desmodiella

III. Head and base of wings dark brown 41. ostensackenella. HH. Fasciæ distinctly bent outward near the middle..43. mariæella.

1. L. fitchella Clemens. Head and thorax white. Fore wings reddish saffron with five, white, costal streaks, the first produced along costa to base; three dorsal streaks, the first before the middle, very large; the second opposite the third costal streak; a black apical dot. 7.5-8 mm.

Tentiform mine on under side of leaves of oak, preferably Quercus macrocarpa and Q. bicolor.

2. L. bataviella Braun. White, marked with angulated fasciæ composed of gray-tipped, golden-fuscous scales; a fascia at basal fourth produced along margins to the base; an angulated fascia at middle of wing, followed by a Y-shaped mark. with its arms on costa, and its outer margin emitting a line of scales connecting with a curved fascia near apex. Apex dark dusted. 7-7.5 mm.15

Ohio, local.

3. L. trinotella Braun. Head, thorax, and basal two-thirds of wing white; apical

¹⁵ Larva on oak (Murtfeldt, in Cornell collection) W. T. M. F.

third pale golden. A pale golden basal streak parallel and close to costa for one-fourth, thence deflexed and passing into golden apical part. In apical part three white costal streaks and two dorsal streaks, the first at tornus, opposite first costal. Black apical dot. 5-6 mm.

Ohio; New Jersey.

An underside miner in leaves of red and silver maple.

4. L. quercialbella Fitch. Distinguished from the preceding by the straight basal streak parallel to fold, darker margins to white streaks, and slightly darker color and larger size. 7 mm.

An underside miner in leaves of oak.

5. L. clemensella Chambers. White; apical half of wings golden, with four costal and three dorsal, dark-margined white streaks, the dark margin of the first costal streak oblique and produced along costa toward base; first dorsal at tornus opposite second costal. Black apical spot. 6-6.5 mm.

Underside miner in leaves of sugar maple.

6. L. argentifimbriella Clemens. Costal and dorsal streaks as in L. clemensella; in addition, a long, golden, basal streak, margined with dark brown along its lower edge. 6.5-7 mm.

An underside miner in leaves of oak, especially white oak.

7. L. lucidicostella Clemens. Differs from the preceding by the pale, unmargined, golden, basal streak and the partial suffusion of the wing with gold below the fold. 6.5-7 mm.

An underside miner in leaves of sugar maple.

8. L. albanotella Chambers. Head white, with a few brownish scales; thorax white. Fore wings pale golden brown with a broad, white. dark-margined, basal streak; four white costal streaks, the first very oblique; opposite it, a broad white dorsal streak, dark-margined on its upper edge, is continued as a broad band along dorsal margin to base, where it is confluent with the basal streak; two posterior dorsal streaks; a black apical spot. 6-7.5 mm.

Ohio; Kentucky; Texas.

Rather small, tentiform mines on underside of leaves of oaks; usually at the edge of the leaf; with the loosened epidermis in numerous parallel ridges.

9. L. hageni Frey and Boll. Head and thorax white. Fore wings saffron brown four, small, white costal streaks, dark-margined on both sides; a large, dorso-basal, white patch, outwardly concave, occupying the basal fifth of wing except along costa (this patch is rarely extended to costa); a large curved white spot at middle of dorsum, and a triangular spot at tornus, both dark-margined on both sides. Apical spot usually present. 7.5-10 mm.

An underside miner in leaves of oaks, especially Q. bicolor.

10. L. æriferella Clemens. Tuft on head dark brown; wings reddish saffron, with four white costal streaks, the first, in the middle of the wing, oblique: three dorsal streaks, the first large and oblique, margined inwardly and around the tip; margins of the second and third dorsal streaks, in the middle of the wing, where they are dull leaden-colored, unite with the margins of the corresponding costal streaks; margin of second dorsal streak usually heavy and conspicuous. A black apical spot preceded by bluish scales. 7-8.5 mm.

An underside miner in leaves of oaks, particularly chestnut oaks and shingle cak. (Quercus imbricaria); distinguished by the dense, ovoid cocoon of frass and silk.

11. L. sexnotella Chambers. Differs from the preceding in the following respects: pale golden ground color; purer white costal and dorsal streaks, more posterior position of the second dorsal streak, whose apex is opposite the space between the second and third costal streaks; the margins of opposite streaks not uniting. 7-7.5 mm.

Kentucky; Pennsylvania.

12. L. obsoleta Frey and Boll. Wings ocherous, with markings as in the preceding species, but not clearly defined, and sometimes almost entirely obscured by suffusion with ground color. 8 mm.

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Massachusetts.

13. L. obscuricostella Clemens. Hade white; forewings pale golden, with a white basal streak dark margined above; four, dark-margined, costal streaks, the first very oblique; three dorsal streaks, the first very long, reaching apex of second costal; second dorsal obliquely opposite third costal; apical spot black. 6-6.5 mm.

Underside, slightly wrinkled mines between veins of leaves of hop hornbeam (Ostrya virginiana).

14. L. ostryaefoliella Clemens. Fore wings pale golden, with an unmargined, median, basal streak, and a white streak along base of mmer margin; four costal, and three dorsal, white streaks, the first pair oblique with apices opposite. 6-6.5 mm.

Large wrinkled mines on underside of leaves of hop hornbeam; dense cocoon of trass and silk.

15. L. olivaeformis Braun. Head and thorax white; fore wings pale brownish otherous; a median, white, dark-margined, basal streak; base of dorsum narrowly white; four costal and three dorsal streaks, first pair oblique with apices opposite; margin of first dorsal bent backward on fold; small black spical spot. 6.5-7 mm. Ohio.

An underside miner in leaves of hickory and pecan (Carya spp.); cocoon of trass and silk.

16. L. caryaealbella Chambers. Head, thorax, and base of fore wings, before the costal and dorsal streaks, white, except for a wide, golden streak along costal margin to first costal streak and a partial suffusion below fold with golden; four costal, and three dorsal, white streaks, on a pale-golden ground color; black apical spot. 6.5 mm.

Kentucky; Wisconsin.

Tent mine on underside of hickory leaves; cocoon of frass and silk.

17. L. rileyella Chambers. Wings golden; a white, unmargined, basal streak; five costal white streaks, the first, before the middle, produced along the costa to the base; three dorsal streaks, first large, with its apex opposite apex of second costal; apical spot small. 7-8 mm.

Missouri; Texas; Ohio.

Tentiform mines on underside of oak; in Ohio, on Quercus imbricaria.

18. L. kearfottella Braun. Head, thorax, and fore wings reddish saffron; a darkmargined, white, basal streak; first costal at one-third, produced along costa to basal fourth; first dorsal large; apical dot large. 7 mm.

Elongate mines on lower side of leaves of chestnut.

19. $\dot{\mathbf{L}}$. propinquinella Braun. Tuft dark brown; thorax and fore wings brownish golden; fore wings darkened with brownish scales in the middle, from second pair of streaks to apex, and in dorsal half of wing; basal streak long, dark-margined above and around its apex; a short, dorso-basal, white streak; four costal, and three dorsal, white streaks, the first pair very oblique, the dorsal one of the pair Leginning much nearer the base than the costal. 8–9 mm.

An underside miner on wild cherry, Prunus serotina.

20. L. crataegella Clemens. Similar to the preceding in color and markings, but differing in the smaller size and less oblique first pair of streaks, of which the dorsal streak begins nearly opposite the costal. 6.5-7 mm.

Small, underside mines on apple, Crataegus and Prunus.

21. L. malimalifoliella Braun. Color as in L. crataegella, from which it differs by the three, equally spaced, costal streaks. 5.5-6 mm.

Underside miner on apple and quince; mine wrinkled, speckled above.

22. L. scudderella Frey and Boll. Head, thorax, and fore wings pale yellowish brown. Basal streak long, margined around apex only; four costal and three dorsal white streaks, first pair very oblique, blackish internal margining often indenting first dorsal on fold; when this accumulation of dark scales is absent, the first dorsal streak appears less oblique. A streak of black scales in apex, margined above and before with white. 7.5-9 mm.

Underside miner on willow, Salix spp.

23. L. salicivorella Braun. Color pale grayish, with four costal, and three dorsal, white streaks, and a white basal streak confluent with the upper edge of the first dorsal streak. 7 mm.

New Jersey.

Underside miner on willow.

24. L. deceptusella Chambers. Color golden brown; four costal, and three dorsal, white streaks, and a white basal streak confluent with upper edge of first dorsal; an elongate patch of brown scales in apex. 6 nm.

Kentucky.

25. L. populiella Chambers. Head white; wings pale golden; three white basal streaks, costo-basal, median, and dorso-basal, respectively; four costal, and three dorsal, white streaks, large, broad, and but little oblique. 6-7 mm.

Ohio; Kentucky.

An underside miner in leaves of silver-leaved poplar, Populus alba.

.26. L. salicifoliella Clemens. Head white; tuft often with brown scales. Fore wings golden or brownish-yellow with white streaks; both streaks and ground color typically dusted with black scales (in some forms, dusting entirely absent). A short, median, basal streak and dorso-basal streak uniting with a dorsal streak at the basal fourth, enclosing a small patch of ground color; two oblique costal streaks, curving backward along the middle of the wing, and three, posterior, less-oblique, costal streaks; first dorsal streak oblique, curving backward, and usually confluent with the first two costal streaks; a triangular spot at tornus, followed by a curved streak usually uniting with the opposite, costal streak. 7-8 mm.

An underside miner in leaves of poplar and willow. Moths appearing in August usually lack the black dusting.

27. L. argentinotella Clemens. Wings golden yellow with a white basal streak to one-fourth, five costal streaks, three dorsal streaks before the tornus and one beyond. 6.5-8 mm.

An underside miner in leaves of elm.

28. L. occitanica Frey and Boll. Differs from the preceding only by the absence of the first and last two costal streaks of that species and may not be distinct. Reported only from Texas. An underside miner on elm.

29. L. auronitens Frey and Boll. Wings brownish golden; a very short basal streak; a long oblique costal streak at one-fourth and an opposite dorsal spot; a pair of triangular streaks in middle of wing whose apices meet; two posterior costal, and two dorsal, streaks. 6.5-8 mm.

Massachusetts; North Carolina.

An underside, rather large, much-wrinkled mine on alder. Alnus spp.

30. L. morrisella Fitch. Tuft dark brown; thorax and fore wing, below the fold, dark brown; remainder of wing golden brown; markings silvery white. A silvery basal streak uniting at one-third with a white dorsal streak; costal streak at one-third; a convex fascia at middle; beyond this, two perpendicular costal streaks, and opposite the first of these a dorsal streak. In the fold between this streak and the fascia, a velvety black streak. Apical spot large, velvety black. 6-7 mm.

Underside whitish mines on hog peanut, Amphicarpa monoica.

31. L. uhlerella Fitch. Differs from morrisella by the absence of the silvery basal streak and the less brilliant luster of the white markings.

Whitish, rather flat mines on underside of leaves of Amorpha fruticosa.

32. L. robiniella Clemens. Agrees with the two preceding in color and general type of markings; no basal streak; first dorsal streak but faintly indicated; median fascia replaced by opposite oblique streaks. 6-6.5 mm.

Under or upper side mines on locust, Robinia pseudo-acacia; white silken cocoon within the mine.

33. L. celtisella Chambers. Fore wing ocherous, with a long, white, basal streak, dark-margined above; an angulated, median, white fascia; a straight fascia at three-fourths, formed by confluent streaks; both faseiæ margined internally with dense dusting; apex white, densely dusted with dark brown. 6-7 mm.

Ohio; Kentucky.

Mine on leaves of blackberry, Celtis occidentalis; at first linear on lower side,

while on leaves of blackberry, *Ceits occulatitis*; at first linear on lower side, then expanding to a blotch on upper side, made tentiform by a longitudinal ridge. 34. L. basistrigella Clemens. Fore wings golden yellow; a long, narrow, median, basal streak; a pair of very oblique, linear, costal and dorsal streaks of equal width throughout and produced along the margins to the base; three posterior costal, and one dorsal, white streak. 8 mm. An underside miner on leaves of oaks; mine scarcely wrinkled; cocoon flat,

outlined by a characteristic oval ring of frass. 35. L. gemmea Frey and Boll. Fore wings reddish saffron; basal streak to

one-third, dark-margined above and below, near middle a nearly straight fascia; two posterior costal, and two dorsal, white streaks; a large brown spot in apex. 7-7.5 mm.

Massachusetts.

An upperside miner on leaves of locust.

36. L. diversella Braun. Thorax and base of wing sometimes deep, metallic golden, wing elsewhere golden or reddish brown; markings silvery or white. A very short basal streak; an almost straight fascia at one-third; two posterior costal, and two dorsal, white spots; an irregular, dark-brown, apical spot, preceded by white scales. 5-7 mm.

Ohio; Kentucky;

Underside mines on huckleberry, Gaylussacia, and on sorrel tree, Oxydendrum arboreum.

37. L. martiella Braun. Fore wing reddish saffron; a white basal streak to one-third; a slightly curved, white fascia just before middle; three posterior costal, and two dorsal, white streaks; a black apical spot. 7 mm.

North Carolina; British Columbia.

Narrow, tentiform mines on under side of leaves of birch, Betula spp.

38. L. symphoricarpella Chambers. Fore wings brownish golden; a short, indistinct, basal streak; a slightly curved fascia just before middle; at beginning of cilia a second fascia, sometimes divided into opposite costal and dorsal streaks; an indistinct costal streak preceding dusted apex. 5.5-6 mm.

Ohio; Kentucky; Texas.

Small, tentiform mines on underside of leaves of Symphoricarpos orbiculatus.

39. L. lucetiella Clemens. Head, thorax, and basal half of fore wings white; apical part suffused with golden; a golden costal streak from base, not reaching middle. At about the middle, a white fascia, bordered with golden inwardly; and a black spot on costa inwardly; at beginning of cilia a pair of white streaks, black-margined inwardly; an unmargined white streak encircling apex. 6-7 mm.

Rectangular transparent mines on underside of leaves of basswood, Tilia americana; pupa contained in an oval cocoon. 40. L. desmodiella Clemens. Tuft dark brown; thorax and base of fore wing

ruby-tinted; ferruginous brown beyond, with two silvery fasciæ, dark-margined on both sides; two posterior costal, and one dorsal white spot. 3-5 mm.

Tentiform mines on underside of leaves of various leguminous plants, such as Desmodium, Lespedeza and Phaseolus. Larvæ often gregarious, in large, inflated mines

41. L. ostensackenella Fitch. Head, thorax, and base of wing dark brown; remainder of wing brilliant golden brown. Two silvery, almost straight fasciæ, at one-fourth and one-half, respectively, shading outwardly into the ground color. Two posterior costal, and two dorsal streaks. 5.5-6 mm. 21

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Mine a yellow blotch on upper or under side of leaves of locust; cocoon flat, oval, yellow, spun outside the mine.

42. L. tritænianella Chambers. Thorax and fore wings pale reddish ocherous; two straight white fascix, a third, sometimes almost divided into opposite streaks, and slightly angulated; an indistinct, whitish, costal streak near the apex. 7-8 mm.

Tentitorin mines on upper side of leaves of hop hornbeam; mine at first blotchlike and speckled, later wrinkled.

43. L. mariæella Chambers. Thorax and fore wings golden brown; two white fasciæ, one at basal fourth, the other in the middle of the wing; both bent outward near the median line and margined inwardly with dark brown; at threefourths, a curved, white, costal streak meeting a similar dorsal streak; a white streak encloses apex. 8-8.5 mm.

Missouri.

Tentiform mines on lower side of leaves of Symphoricarpos orbiculatus.

44. L. celtifoliella Chambers. Wings reddish saffron, with three, acutely angled, white faseiæ, densely dusted internally with dark-brown, which often obscures the white; each of the first two fascize sending a white streak from its angle almost to the next fascia; the third running into a dusted costo-apical patch; a dorso-apical patch of dark dusting on a white ground; a median basal streak, densely dusted dorsally, this dusting often confluent with a dusted, dorso-basal, white patch. 6 mm.

Ohio; Kentucky; West Virginia.

Tentiform mines on underside of leaves of hackberry. 45. L. fragilella Frey and Boll. Fore wings brownish or deep reddish saffron, with two, outwardly angulated, narrow, indistinct, but complete fasciæ at basal fifth and at two-fifths, respectively; a third nearly straight at three-fifths interrupted in the middle by a projection from its internal margin; two posterior costal streaks, and a dorsal streak indicated by its margin. The fasciæ and streaks margined internally with dark dusting, the margins becoming broad on the costa. Apex dusted. 8.5-9 mm.

Tentiform mines on underside of leaves of honeysuckle; Lonicera spp.

46. L. tiliacella Chambers. Head, thorax, and base of wings, except along costa, pure white; middle part of wing to beyond beginning of cilia pale golden; a few golden scales in apex; three white fascia, the first two straight and darkmargined internally, the third slightly bent and its dark margin interrupted in the middle. 6-7 mm.

Circular tent mines on upper side of leaves of basswood; mine white, speckled with brown.

47. L. lysimachiæella Chambers. Named from mine on underside of leaves of Lysimachia lanceolata.

Group II

The larva of the flat group is very much depressed, almost flat, with the sides of the segments projecting, thus giving the entire larva a beaded appearance. The head is flat, somewhat triangular in shape, usually of a shining, reddish brown color, with the mouth parts projecting in front. The first three segments of the body are broader than the others and the body tapers toward the posterior end. While the legs are of the same number and occupy the same position as in the cylindrical larva, all are very rudimentary, appearing as small, tubercular projections. On the upper and lower side of most, and sometimes all, of the body segments, are distinctly outlined, shining, darker spots — the maculae. These vary in shape on the different segments, being elliptical or trapezoidal, but are constant for a given species in each stage. After the seventh, or last, moult, the larva assumes a more cylindrical shape; the legs are better developed; and the dark maculae gradually disappear.

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All of the species are miners on the upper side of leaves, where they make a flat, sometimes irregularly shaped blotch, or a rather broad, linear tract. The larva feeds from the centre outwardly, thus gradually increasing the extent of the mined portion of the leaf. The mine retains its flat, blotch-like character until after the seventh moult. The larva then lines the loosened epidermis with silk and by contraction produces from one to three narrow folds or ridges. Beneath this folded portion, the floor of the mine is thinly covered with silk. Then the larva, lying on its back, spins a flat, semi-transparent sheet of silk, oval or nearly circular in shape, and attached around its edges to the floor of the mine. Beneath this, along its long axis, the pupa is formed. The pupa, which is protruded from the mine in emergence, is thrust through a transverse slit near one end of the flat cocoon. Such a cocoon is formed in all but a few of the species of which the life history is known.

In almost all of the species, however, this cocoon is only made in the brood of which the imagoes are to appear in the same summer. In a later brood, the hibernating period, with one or two exceptions, is passed in the larval state. In a number of species, hibernation takes place beneath the folded epidermis. In most cases, however, an especially prepared, silken-lined chamber is formed. After the floor of the mine is loosely covered with silk, the upper epidermis is fastened down in a circular or oval outline, and the whole cavity is then lined with silk. Ample space is provided by a characteristic oval, or hemispherical, projection upon the under side of the leaf. The change to pupa occurs in the spring. Except for the absence of cremaster, the pupa is identical with that of the cylindrical-larval group.

The moths of this group may easily be recognized by the fact that the white markings of the fore wings are always externally dark-margined, and often densely dusted with black scales behind. In some cases there is, in addition, a slight internal margin, consisting of a few dark scales near the costa. The markings consist of white costal and dorsal streaks, usually oblique. Opposite streaks may unite to form a fascia, either outwardly angulated or straight.

Key to the species

A. Oblique costal and dorsal streaks; fasciæ, if present, distinctly angulated; never two straight fasciæ.

C. An oblique white streak or patch at the base of the dorsal margin, rarely indistinct (*lentella*, *caryaefoliella*) usually two, angulated fasciæ.

D. No costal streak before the middle; a median fascia. 51. saccharella. DD. A fascia at one-fourth and one at one-half.

- E. White streak at base of dorsum indistinct, or indicated by external, dark scales only.

EE. White streak at base of dorsum large and distinct.

D. Dorsal margin white from base to beyond middle.

E. Dorso-basal streak extending to oblique streak above the cilia.

F. Antennæ annulate with brown for their whole length.

54. conglomeratella.

DD. Dorsal margin not white.

E. With two costal streaks or spots before the white streak or spot forming the anterior edge of the apical dusting.

F. Ground color of wings pale, with a coppery lustre toward the

EE. With at most one costal streak before the white streak or spot

forming the anterior edge of the apical dusting. F. Median fascia but slightly angulated; other white marks small or

absent.

FF. Median fascia sharply angulated; white marks conspicuous.

G. First dorsal streak long, oblique, curved 59. fletcherella. GG. First dorsal streak perpendicular.

HH. Marks shining white, strongly margined.

I. External dusting at angle of fascia produced backward; apex densely dusted.

II. External margin of fascia not produced.....60. arcuella.

AA. Two straight, or nearly straight, fasciæ, nearer the base on the dorsal margin. BB. A costal and a dorsal streak at three-fourths.

C. Without a paler streak at the base of the dorsal margin.

D. Apical dusting, black on a whitish ground, and extended to the

CC. With a white streak or paler shade from inner angle to fold.

D. First fascia and its dark margin broken near the costa...68. æsculisella. DD. First fascia complete.

E. Color deep reddish......69. hamameliella. EE. Color paler ocherous.

F. Dorsal streak at tornus very oblique and pointing toward apex.

48. L. hamadryadella Clemens. Head, thorax, and fore wings white. Wing markings as follows: a small patch of black scales at base of costa; a slightly angulated band of black dusting on an ocherous ground, followed by an angulated line of black dusting; an angulated ocherous fascia at one-third, broadly dusted internally with black scales; a second, similar fascia just beyond the middle; the space between these two fasciæ marked by a line of black scales; a similar line of scales between the second fascia and costal and dorsal patches, which are separated from one another in the middle of the wing hy a patch of dark dusting; apex ocherous. dusted with black. 6.5-8.5 mm.

Irregular, whitish blotch mine on upper side of leaves of oak, especially Quercus alba, rarely (perhaps accidentally) on *Magnolia* and *Ostrya*. 49. L. cincinnatiella Chambers. Fore wings shining ocherous; a curved white

streak from base of inner margin to fold, dusted outwardly with black scales; two angulated fasciæ, at one-third and one-half, respectively, dusted externally with black scales, the dusting produced at the angle; at two-thirds a small, dusted. costal spot, beyond which is a white streak; an oblique dorsal streak from just before tornus sometimes extended to form inner margin of dusted apex. 6.5-7.5 mm. Larvæ gregarious, in brownish-yellow, blotch mines on leaves of oaks, principally

white oak.

50. L. macrocarpella Frey and Boll. Close to cincinnatiella; differs by the following characters: a small tuft of brown scales in apical cilia (wanting in cincinnatiella); larger size (8.5-9 mm.), darker, less shining ground color, more oblique, and slightly curved, dorsal arms of fascia.

Upper side blotch on oak; each mine with a single larva.

Eastern, southern, and western United States, chiefly near coast.

51. L. saccharella Braun. Fore wings ocherous; an oblique, curved, white streak at base of inner margin, usually uniting with an oblique dorsal streak at basal fourth; an acutely angled, slightly interrupted fascia at middle, a white costal spot beyond, and a long, oblique, dorsal streak opposite; apex densely dusted. 5-7 mm.

Small, irregular blotches on upper side of leaves of sugar maple, and occasionally on other maples.

52. L. caryaefoliella Clemens. Fore wings reddish ocherous; an indistinct, oblique, curved streak from base of dorsum; two angulated fasciæ, placed as in L. cincinnatiella; a third angulated fascia formed by the meeting of two, opposite, slightly oblique streaks; a small, white-dusted, costal spot beyond; apex black-dusted on a white ground. 6-7 mm.

Irregular blotches on upper side of leaves of hickory and, occasionally, of butternut and walnut.

53. L. lentella Braun. Markings at base and two angulated fasciæ as in preceding species; beyond second fascia, on costa, a broad white spot, margined on each side and below with black scales; opposite it a long, oblique, dorsal streak, opposite whose apex a second small, white dusted, costal spot; apex of wing dusted with black on a white ground, 6.5-7 mm.

United States west to Arizona.

Blotch mines on birch and hop hornbeam, often containing several larvæ. Epidermis much wrinkled, bending leaf into a fold.

54. L. conglomeratella Zellcr. Fore wings reddish saffron; two oblique costal streaks, at one-third and one-half, respectively; a costal spot at two-thirds; a narrow, white streak from base along dorsal margin nearly to tornus, then deflexed and passing obliquely toward apex, forming the inner margin of the apical dusting and sometimes partially obscured by the latter; a tuft of brown scales in the apical cilia. Antennæ annulate with brown. 7.5-9 mm.

New Jersey to Ohio; south and west to Texas and California. Blotch mines on oak, especially live oak (Quercus virginiana). 55. L. ulmella Chambers. Differs from L. conglomeratella by the more oblique, costal streaks, the absence of tuft of scales in apical cilia, and the white basal third of antennæ. 6.5-7 mm.

Blotch mines on upper side of leaves of elm.

56. L. quercivorella Chambers. Distinguished from both ulmella and conglomeratella by dorso-basal, white streak extending scarcely beyond the middle and not connecting with the oblique, dorsal streak before tornus. 6.5-7 mm.

A small blotch mine on upper side of leaves of oak, especially red oak (Q. rubra), and usually placed near the tip of a lobe.

57. L. platanoidiella Braun. Fore wings reddish ocherous; an oblique, costal streak at one-third; angulated fascia at middle, its external dusting being prolonged backwards at angle; a pair of white streaks at apical third; a small, white spot on costa between the costal streak of the pair and the white streak forming the inner margin of the apical dusting. 6.5-8 mm.

Blotch mines on upper side of leaves of oak, especially on $Quercus \ alba, Q.$ macrocarpa, and Q. bicolor.

58. L. cervina Walsingham. Distinguished from the preceding by the pale ground color of wings, their coppery luster toward apex, and the indistinct markings. 6 mm. New York.

59. L. fletcherella Brann. Fore wings reddish ocherous; a pair of costal and dorsal streaks at one-third, the dorsal one long, oblique, eurved, with its pointed apex directed toward apex of costal streak; a median, angulated fascia; a pair of opposite streaks at two-thirds; between these and a white streak before apex, in middle of wing, a white spot. 8.5-9 mm.

Ottawa, Canada.

Blotch mines on white oak.

60. L. arcuella Braun. Fore wing reddish orange with markings shining white; first costal streak at one-third broader than long; first dorsal streak nearly perpendicular, rectangular; median angulated fascia, with dark margin not produced at angle; costal streak at two-thirds strongly curved; an opposite dorsal streak; a curved costal streak enclosing apex; apex dusted with blackish-brown scales. 10 mm.

Virginia.

61. L. betulivora Walsingham. Wings pale reddish ocherous and markings dull white, with faint margins; markings as in preceding, but streaks at two-thirds very indistinct, and dusted apex not enclosed by a white streak. 7 mm.

North Carolina.

Small, round, blotch mines on leaves of birch.

62. L. bethunella Chambers. Fore wing reddish orange; markings as in two preceding species, except that first pair of streaks are longer and the costal one of the pair is decidedly oblique; the external dark dusting of the fascia produced at the angle to the space between the streaks, at two-thirds; apical dusting preceded by a white spot. Tarsi white. 6.5-7.5 mm.

Blotch mines on leaves of various species of oak.

63. L. picturatella Braun. Distinguished from the preceding by the conspicuous, black spots near the ends of the tarsal segments. 6.5-7 mm.

Connecticut; New York; New Jersey.

Brownish blotch mines on upper side of leaves of bayberry, Myrica carolinensis. 64. L. fasciella Walsingham. Fore wings reddish orange; a median, very

obtusely angulated fascia, its external, dark dusting being produced backward into the apical dusting. No costal or dorsal streaks. 6-7 mm.

Ohio and Kentucky.

Oval blotch mines on leaves of various species of oaks.

65. L. castaneæella Chambers. Fore wings reddish orange; a small, white, costal spot at one-third; a median fascia very obtusely angled, its dustings extending backward to the costal streak at two-thirds; opposite this streak, a faint dorsal streak; a small, white streak before dusted apex; dusting behind fascia and in apex sometimes almost entirely lacking. 6-7.5 mm.

Oval, blotch mine on chestnut and oak.

66. L. guttifinitella Clemens. Reddish orange; two straight, or nearly straight, fasciæ, at one-third and one-half, respectively; a costal spot at two-thirds and opposite it, a rather oblique, dorsal streak; apical dusting black on a white ground and extended to tornus; a white streak along upper edge of dusting near apex. 7 mm.

Mine a broad, tortuous tract, on poison ivy, Rhus toxicodendron.

67. L. obstrictella Clemens. Reddish brown; two, nearly straight, silvery fasciæ, the second nearly perpendicular, opposite costal and dorsal streaks at two-thirds,

often uniting to form a third, nearly straight fascia. Apex velvety black, preceded by a few silvery scales. 7-8 mm.

Mine a broad, branched tract, usually Y-shaped, often crossing midrib, on leaves of oaks, most commonly chestnut oak. Passes the winter in pupal state, contrary to the usual habit of species of this group.

68. L. aesculisella Chambers. Reddish ocherous; a faint, whitish streak from base of inner margin to fold; two fasciæ; the first fascia and its dark margin broken near costa, slightly angulated, and its dorsal arm more oblique than the second fascia, the latter being nearly straight. At two-thirds, a costal spot and an opposite, longer, dorsal streak. 8-9 mm.

Central United States.

Mine a broad, linear tract, on leaves of buckeye, Aesculus glabra and A. octandra; often containing several larvæ.

69. L. hamameliella Busck. Deep reddish orange; an oblique white streak from base of inner margin to fold; two straight, oblique, silvery fasciæ; a silvery spot at two-thirds; a long, very oblique, opposite, dorsal streak, parallel to termen, uniting in apex of wing with a short apical streak. Marginal line in cilia dis-tinct. 7 mm.

Mine a whitish blotch on leaves of witch-hazel, Hamamelis virginiana.

70. L. aceriella Clemens. Fore wings reddish ocherous; markings as in preceding species, except that dorsal streak at tornus is usually less oblique and marginal line in cilia not defined. 7-9 mm.

Mine a flat, broad tract, in leaves of maple, most commonly red maple. 71. L. ostryarella Chambers. Markings of the general character of the three preceding species, but distinguished as follows: first fascia concave outwardly on fold, more oblique than second; dorsal streak at tornus oblique; apex dusted

with dark brown. 6-7 mm. Irregular blotch mines on hop hornbeam (Ostrya) and ironwood (Carpinus); larvæ sometimes gregarious; hibernating chamber of winter outlined by a raised, circular ridge.

72. L. corylisella Chambers. Distinguished from the preceding species by the erect, dorsal streak at tornus. 6.5-7 mm.

Blotch mines on upper side of leaves of hazel, Corylus americana.

73. L. tubiferella Clemens. Head white; wings deep ocherous, with two straight, slightly oblique, white fasciæ; no costal or dorsal streaks; sometimes a minute, white spot in the apex. 7.5-8 mm.

Mine a very characteristic tract, gradually increasing in breadth, on leaves of oaks. Larva placed transversely in the mine.

Synopsis of species of Lithocolletis by food plants

Salix spp., willow:

L. scudderella. Tentiform mine on underside of leaf. L. salicivorella. Tentiform mine on underside of leaf. L. salicifoliella. Tentiform mine on underside of leaf.

Populus alba, silver-leaved poplar:

L. populiella. Small, tentiform, underside mine.

Populus spp., poplar:

L. salicifoliella. Rather large, tentiform, underside mine.

Myrica carolinensis, bayberry:

L. picturatella. Upper side, blotch mine.

Juglans cinerea, butternut:

L. caryaefoliella.

Juglans nigra, walnut:

L. caryaefoliella. Upper side, irregular, blotch mine.

Carya spp., hickory:

L. caryacalbella. Tentiform, underside mine; dense, ovoid cocoon of frass and silk.

L. olivacformis. Tentiform, underside mine; dense, ovoid cocoon of frass and silk.

L. caryacfoliella. Upper side irregular, blotch mines, often confluent. Corylus americana, hazelnut:

L. coryliscila. Upper side, irregular blotch mine. Ostrya virginiana, hop hornbeam:

L. obscuricostella. Small, slightly wrinkled, tentiform, underside mine; pupa in a slight, silken web.

L. ostryacfoliella. Underside mine, larger than that of preceding species and more wrinkled; ovoid eocoon of frass and silk.

L. tritanianella. Upper side, blotch mine; epidermis of leaf white, speckled, and later, much wrinkled and leaf folded.

L. lentella. Brownish, upper-side, blotch mines, often containing several larvæ; epidermis of leaf in numerous longitudinal ridges; leaf folded.

L. ostryarella. Irregular, flat, upper-side, blotch mines, usually containing several larvæ; hibernating chamber outlined by a circular ridge.

Carpinus caroliniana, ironwood:

L. ostryarella. (Described under Ostrya.)

Betula spp., birch:

L. martiella. Narrow, tentiform, underside mine.

L. lentella. (Described under Ostrya.)

L. betulivora. Small, nearly circular, upper-side, blotch mine.

Alnus spp., alder:

L. auronitens. Underside, rather large, tentiform mine.

Castanea dentata, chestnut:

L. kearfottella. Rather small, narrow, tentiform, underside mine, between veins.

Many upper-side miners on oak are also often found on chestnut. These mines are described under Quercus.

Quercus spp., oaks:

Underside, tentiform mines:

L. fitchella. Large, tentiform mine; epidermis of leaf slightly and finely wrinkled; most commonly on Q. macrocarpa and Q. bicolor.

L. hageni. Mine similar to that of fitchella.

L. rileyella.

L. quercialbella.

L. argentifimbriella. Especially on white oak, Q. alba.

L. albanotella. Small mine, usually near margin; epidermis of leaf with numerous, distinct, longitudinal ridges; especially on Q. bicolor and Q. macrocarpa.

Tentiform, wrinkled mine; ovoid cocoon of frass and silk; L. *æriferella*. especially on chestnut oak and shingle oak (Q. imbricaria).

L. basistrigella. Rectangular, transparent, scarcely wrinkled mine; flat cocoon outlined by ring of frass.

Upper-side blotches or flat broad tracts.

L. hamadryadella. Whitish, irregular, blotch mine.

L. cincinnatiella. Large, brownish blotches; larvæ usually gregarious; priucipally on white oak.

L. macrocarpella. Blotch mine.

L. conglomeratella. Blotch mine, especially on live oak (Q. virginiana).

I. quercivorella. Small, blotch mine, usually near the tip of a lobe, especially on red oak (Q. rubra).

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L. platanoidiella. Blotch mine.

L. fletcherella. Blotch mine on white oak.

L. bethunella. Small blotch mine.

L. castaneælla. Small blotch mine.

L. fasciella. Small blotch mine.

L. obstrictella. Mine a broad, branched tract, usually Y-shaped; most commonly on chestnut oaks.

L. tubiferella. Mine a broad, linear tract, gradually increasing in breadth and occasionally branched; larva placed transversely in mine.

Ulmus spp., elms:

L. argentinotella. Large, tentiform, underside mine.

L. occitanica. Tentiform, underside mine.

L. ulmella. Upper-side, blotch mine.

Celtis occidentalis, hackberry:

Underside, linear mine, expanding into an upper-side blotch, L. celtisella. with a longitudinal ridge.

L. celtifoliella. Underside, tentiform mine.

Hamamelis virginiana, witch-hazel: L. hamameliella. Whitish, upper-side, blotch mine.

Pyrus malus, apple:

L. malimalifoliella. Small, tentiform, underside mine; also on quince.

L. cratægella. Small, tentiform, underside mine.

Cratægus spp., hawapple:

L. cratagella. Small, tentiform, underside mine.

Prunus serotina, wild black cherry:

L. propinquinella. Large, tentiform, underside mine.

L. cratægella. Tentiform mine, smaller than that of L. propinquinella. Amorpha fruticosa:

L. uhlcrella. Whitish, rather flat mines on underside of leaves.

Robinia pseudoacacia, locust:

L. robiniella. Under- or upper-side, white mines. L. gemmea. Upper-side miner.

L. ostensackenella. Yellow, blotch mine, on upper or lower side. Larva leaves the mine to pupate.

Amphicarpa monoica, hog peanut: L. morrisella. Large, white, tentiform, underside mine, sometimes containing several larvæ.

Desmodium spp., Lespedeza spp., and Phaseolus:

L. desmodiella. Tentiform, underside mine, sometimes inflated and containing several larvæ.

Rhus toxicodendron, poison ivy:

L. guttifinitella. A broad, tortuous, irregular tract on upper side of leaf. Acer spp., maples:

L. clemensella. Underside, tentiform mine, on sugar maple.

L. lucidicostella. Underside, tentiform mine, on sugar maple.

L. trinotella. Small, underside, tentiform mine, on red and silver maple.

L. saccharella. Irregular, upper-side, blotch mine, chiefly on sugar maple.

L. aceriella. A flat, broad, tract on upper side; most commonly on red maple. *Esculus* spp., horse-chestnnt, buckeye:

A broad, linear, upper-side tract, sometimes containing five L. asculisella. or six larvæ.

Tilia americana, basswood:

L. lucetiella. Transparent, unwrinkled, underside mine; cocoon oval.

L. tiliacella. Circular, whitish, speckled, tent mine on upper side.

Gaylussacia spp., huckleberry:

L. diversella.

Oxydendrum arboreum, sorrel tree:

L. diversella. Tentiform, underside mine.

Steironema (Lysimachia) lanceolatum:

L. lysimachiæella. Small, tentiform, underside mine.

Lonieera spp., honeysuekle;

L. fragilella. Rather large, tentiform, underside mine.

Symphoricarpos orbiculatus, coral-berry:

L. symphoricarpella. Very small, underside, tent mines; half the mine partitioned off to form a pupal chamber. L. mariæella. Tent mines, larger than those of L. symphoricarpella; an ovoid,

L. marixella. Tent mines, larger than those of L. symphoriearpella; an ovoid, silken cocoon.

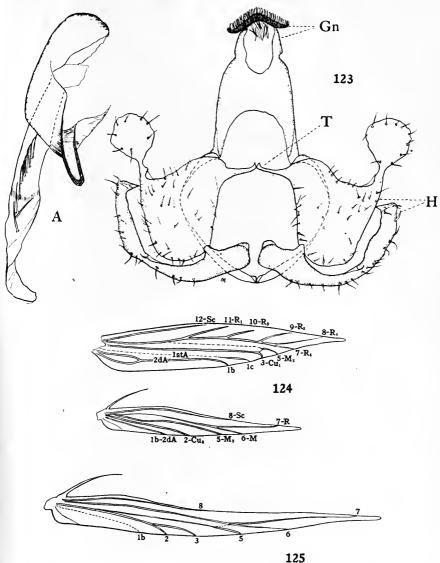
Family 15. COLEOPHORIDÆ

Carl Heinrich¹⁶

Head smooth. Antennæ $\frac{2}{3}$ to 1; thickened with scales toward base; basal joint with appressed scales, rough scaled, or with projecting scale tuft. Labial palpi moderate or long; ascending. Maxillary palpi absent. Posterior tibiæ rough haired above or smooth. Fore wings (fig. 124) elongate, narrow; never more than eleven veins; 1b (2d A) furcate; 2 (Cu_2) sometimes absent; 4 (M_3) sometimes absent or united with 3 (\mathbf{Cu}_1) ; 6 (\mathbf{M}_1) absent; 7 (\mathbf{R}_5) to termen; 7 and 8 $(\mathbf{R}_4 \text{ and } \mathbf{R}_5)$ approximate, connate, or stalked; pattern limited to irrorations of darker or lighter scales, longitudinal lines, or dustings, or one or more stigmata: wings often unicolorous, never with transverse markings or fasciæ. Hind wings (figs. 124, 125) linear-lanceolate; narrower than fore wings; 6 to 8 veins; crossveins between 5 and 6 (\mathbf{M}_1 and \mathbf{M}_2) weak or absent. 3 (Cu_1) sometimes absent; 4 (M_3) sometimes absent; 5 and 6 ($\mathbf{M}_1, \mathbf{M}_2$) separate; 6 and 7 (\mathbf{R}, \mathbf{M}_1 closely approximate, connate, or stalked. Male genitalia (fig. 123, A) with harpe partly divided; costal arm of harpe free for half its length, weakly chitinized; arms of gnathos fusing and forming a complete ring about anus, terminating in a more or less scobinate knob; gnathos occupying place of, and apparently functioning as, an uncus; uncus absent. Ædœagus rather stout, short or moderately long, hinged to vinculum.

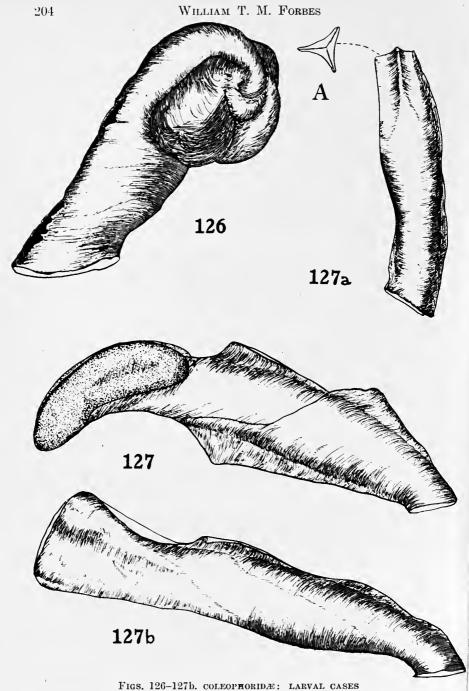
Larvæ inhabiting portable cases (figs. 126, 127, 127a, b) feeding on leaves, flowers, fruits, and seeds of various plants; external feeders or miners; never boring into stems of plants or folding or rolling leaves; with 3 setæ on prespiracular shield of prothorax. II caudolaterad of I on abdominal segments 1 to 8, IV and V approximate on abdominal segments 3 to 8; prothoracic spiracle vertically placed.

¹⁶ Of the Bureau of Entomology, United States Department of Agriculture.



FIGS. 123-125. COLEOPHORIDÆ

123, Male genitalia of Coleophora atlantica Heinrich: H, harpe; Gn, gnathos;
T, transtilla; A, Aedœagus and penis (lateral view)
124, Venation, fore and hind wings of Coleophora laricella Hübner
125, Venation of hind wing of Coleophora vernoniæella Chambers
(Drawn by Ada F. Kneale, of the Bureau of Entomology of the United States Department of Agriculture)



126, Larval case of Coleophora atlantica Heinrich (typical "pistol case") 127, Larval case of Coleophora leucochrysella Clemens

· 127a, Larval case of Coleophora fletcherella Fernald: A, triangular compression of posterior end

137b, Larval case of Coleophora laticornella Clemens (typical "cigar case") (Drawn by Ada F. Kneale, of the Bureau of Entomology of the United States Department of Agriculture)

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The family as here characterized includes, besides the typical Coleophora Hübner, two other small European genera, Goniodoma Zeller and Metriotes Hübner (Asychna Stainton). Of the exotic genera, which I have not seen, two of Meyrick's (the Australian Corythangela and the African Platybathra) should probably also be included. Venational and genital characters, namely the obsolescence of the fork in 1b (2d A) of fore wing, the stalking or fusing of 5 and 6 (M_1 and M_2) in the hind wing, and the presence of both guathos and a large functioning uncus, exclude Batrachedra Stainton which is usually associated with Coleophora.

In North America all our species belong to the genus *Coleophora* Hübner, which may be defined as follows:

1. COLEOPHORA Hübner

Antennæ $\frac{4}{5}$; porrected in repose. Labial palpi smooth or with slight tuft on under side of second joint toward apex. Posterior tibiæ rough haired above; anterior spurs of hind tibiæ from beyond middle. Fore wing with 7 and 8 (\mathbf{R}_4 and \mathbf{R}_6) connate or stalked. Hind wing $\frac{2}{3}$; 6 to 8 veins; 3 (\mathbf{Cu}_1) rarely absent; 4 sometimes absent; crossvein between 5 and 6 (\mathbf{M}_1 and \mathbf{M}_2) nearly obsolete.

Larva with head elongate ovoid, longer than wide; frons extending nearly to incision of dorsal hind margin; anterior and lateral setæ and punctures crowded forward on head, nearly in a straight line approximate to ocelli; ultra-posterior portion of epicranium large. Body setæ much reduced; thoracic tubercles enlarged, usually fusing on dorsum of meso- and metathorax to form secondary shields. Crochets uniordinal, in a flattened ellipse opening inwardly, or in two transverse bands; frequently reduced in number; sometimes altogether absent.

Pupa incomplete; with appendages soldered together, but not to body, extending nearly to or beyond caudal margin of body; caudal end of body with lateral extensions ending in sharp spines; femora of prothoracic legs defined; maxillary palpi absent; epicranial suture present; prothorax very short on meson, long on lateral margin, forming a double triangle; abdomen not spined above; cremaster absent. (This description probably applies to the entire family, but inasmuch as only pupal representatives of *Colcophora* were seen, the description is given under the genus.)

Coleophora is a very large genus, numbering between four and five hundred described species, largely confined to the northern hemisphere, and reaching its greatest development in middle and southern Europe and the United States. A couple of species have been described from South America, several from South Africa and India, and a few from Eastern Asia. In the United States we have some ninety odd described species, of which over half are found in the region covered by this hand book. Several attempts have been made to divide this unwieldy genus, but so far all have failed. The following table and arrangement of species, while arbitrary, attempts to follow natural lines, as far as is consistent with convenience of identification, and to bring together related species. Group VIII appears to be a natural one and on purely larval characters and habits should have generic rank; but so far, no consistent adult characters have been found to separate it from other *Coleophora*. The antennal character given (projecting hair tufts on basal joint) holds for the described North American species; but in Europe is also found on some species outside the free feeding group.

For the student of Lepidoptera, this family offers a fascinating study. There are still many new life histories to be worked out and many species yet to be

described. The group as a whole, however, is such a difficult one and so many of the species are similar, that accurate determinations can be made only by one who has a large collection available and has had considerable experience with the group. As it is, even he can not always be sure of his species unless he has both larval case and food plant records as well as the adult. No one should think of describing new *Coleophora* except from bred specimens carefully determined. To describe from collected adult material alone is nothing short of a crime, as it can only add to the inconvenience and confusion of future workers.

Key to the species

1.	Antenna with basal joint simple, rough scaled, or with only slight tuft, never
	with large projecting tuft; fore wing never pure white unmarked by longi-
	tudinal lines
	Antenna with large projecting tuft on basal joint; or, if tuft is absent or much
0	reduced, fore wing snow white unmarked by longitudinal lines
z.	Fore wing metallic or gray brown, unicolorous, sometimes tinged with ochreous
	or darker shades at apex but never striped or otherwise marked
2	Fore wing dark gray to grayish brown, semi-lustrous4
э.	Fore wing with shining, irridescent, metallic luster
1	Apex of fore wing distinctly tinged with grayish ochreous
4.	(1) cerasivorella. (2) zelleriella.
	Apex of fore wing not distinctly tinged with grayish ochreous
5	Head with decided ochreous tinge
0.	Head gray or grayish brown
6.	Entire insect shining steel gray
	Entire insect gray-brown or drab
7.	Fore wing somewhat paler than hind wing towards apex(8) concolorella.
• •	Fore wing concolorous with, or somewhat darker than, hind wing towards
	apex
8.	Legs and underside of moth with pale dustings grayish and largely obscured by
	dark scaling
	Legs and underside of moth with pale dustings white and conspicuously pre-
	dominating over darker scaling9
9.	Fore and hind wing a uniform, dark, smoky gray-brown(7) pruniella.
•	Fore and hind wing rather pale gray brown(9) cinerella.
10.	Antenna simple, thickened at base only
• •	Antenna clothed with rough scales for $\frac{1}{2}$ or more
11.	Antenna with rough scaling to $\frac{1}{2}$; annulated beyond(11) coruscipennella. Antenna with rough scaling to $\frac{4}{5}$; apical $\frac{1}{5}$ silver white(12) apicialbella.
19	Fore wing yellow to dark brown, more or less streaked with white
	Fore wing white streaked with yellowish white (or yellowish white streaked
	with white)
13.	Fore wing golden to dark brown; costa white; otherwise unmarked14
	Fore wing yellow unicolorous; or yellow with white costal and dorsal margins;
	or ochreous fuscous with the veins outlined in white; or brownish with two
	or more white stripes
14.	Fore wing dull blackish brown15
	Fore wing drab brown to golden brown16
15.	Fore wing drab brown to golden brown
	Antenna not annulated
16.	Fore wing very dark (blackish brown) toward apex
	Fore wing nearly uniform golden brown, scarcely darker towards apex18

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17.	Head and scaling on basal joint of antenna grayish ochreous with little admix-
	ture of white(15) carpinella.
	ture of white
	ture of white(16) umbratica.
18.	Dorsal cilia on fore wing dull grayish fuscous
10	Lorved one emotion for the second state of the
19.	Larval case smooth
20.	Alar expanse more than 10 mm (19) limosinennella
1 0.	Alar expanse less than 10 mm
21.	Alar expanse less than 10 mm
	slightly curved edge
	Larval case smooth; mouth deflected to 45 degrees; posterior end flattened to a
~ ~	straight edge
22.	Ground color of fore wing yellow
<u></u>	Costa of fore wing dark
40.	Costa of fore wing pale
24.	Costal and dorsal margins of fore wing white
	Costal and dorsal margins of fore wing white
	of wing
25.	Ground color of fore wing pale cream-yellow(25) cretaticostella.
	Ground color of fore wing dark yellow
26.	Ground color of fore wing drab brown
97	Ground color of fore wing grayish brown
	Fore wing with four white longitudinal streaks (29) infuscatella
28.	Fore wing with no fuscous or dark ochreous dustings on white areas
	Fore wing with no fuscous or dark ochreous dustings on white areas
	ous
29.	Alar expanse 10 mm. or over
20	Alar expanse less than 10 mm
	(31) borea. Fore wing without such
31.	Longitudinal white and ochreous markings on fore wing sharply contrasted32
	Longitudinal white and ochreous markings on fore wing not sharply con-
	trasted
32.	White median shade along upper margin of cell sending only two branches to
	costa
	to costa
33.	Only three otherous lines on fore wing (33) coenosinennella
	More than three ochreous lines on fore wing
34.	'three ochreous lines above fold in fore wing distinctly separated by white
	lines (34) cratipennella.
95	Three ochreous lines above fold tending to fuse
50.	Antenna distinctly annulated for basal half (37) ericodes.
36.	Antennæ not distinctly annulated for basal half
	few scattered scales toward apex
	Head and greater part of fore wing othrous
37.	Outer margin of fore wing mostly white
20	Outer margin of fore wing ochreous
30.	Hind tarsi annulated with fuscous

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39.	Head whitish ochreous; white on fore wing distinct
	Head dark grayish ochreous; white on fore wing almost completely obscured
	by ochreous or fuscous scaling
40.	by ochreous or fuscous scaling
	spots or dashes
	Fuscous dustings pronounced on white areas, crowded to form a blackish-brown
	dash at end of cell
41.	Antennæ distinctly annulated
	Antennæ not distinctly annulated
42.	Antenna simple or white; only slight basal tuft
	Basal tuft on antenna well developed
43.	Fore wing pure white, unmarked
	Fore wing pure white except apex which is strongly tinged with pale golden
	yellow
44.	Fore wing white at least at base
	Fore wing without any white; ochreous at base
45.	White on fore wing limited to basal half of wing
	White on fore wing not limited to basal half of wing
.46.	Fore wing shading from white at base to straw color; coppery at apex.
	(47) viburnella.
	Fore wing brown or clouded with brownish from middle to apex
47.	Cilia of fore wing and hind wing ochreous fuscous, lighter than wings.
	(48) nigralineella.
	Cilia of fore wing and hind wing dark smoky fuscous, concolorous with darkest parts of wings
	parts of wings
48.	Head, thorax, and autennal tuft pure white
	Head, thorax, and antennal tuft white sprinkled with grayish fuscous.
	(51) malivorella.
49.	Fore wing faintly striped and dusted toward apex with ochreous and fuscous.
	(52) atromarginata.
	Fore wing more or less dusted with ochreous fuscous but not striped50
50.	Dark dustings abundant and well scattered over fore wing (53) elæagnisella.
	Dark dustings sparse and limited to apical half of fore wing
51.	Apex of fore wing lightly dusted with blackish fuscous scales (54) querciella.
	Apex of fore wing lightly dusted with ochreous or golden scales. (55) atlantica.
	. Moths with fore wings gray brown, unicolorous, sometimes tinged with ochre-
	or darker shades but not striped or otherwise marked. Larval cases with
	terior ends flatly or triangularly compressed. Larvæ leaf-miners or seed
tee	ders.

(1) C. cerasivorella Packard.

Alar expanse: 9-9.5 mm. Localities: Massachusetts; Wisconsin. (June). Food plants: Cherry; apple. Larval case: Unknown.

Moths in the United States National Museum. are without the larval case. They were reared from apple in Bellingham, Wisconsin. Packard reared the moths from larvæ taken feeding on leaves of cherry, at Salem, Massachusetts, but he gives no description of the larval case.

(2) C. zelleriella Chambers.

Alar expanse: 9 nm.

Locality: Kentucky.

Food plant: Unknown.

Larval case: Ochreous; slender; cylindrical; tapering slightly to each end; posterior end triangularly compressed; 12 mm. long (From Chambers' description).

This species is known only from Chambers' meager description. It is probably a synonym of cerasivorella Packard.

(3) C. occidentis Zeller.

Alar expanse: 10 mm.

Locality: Massachusetts.

Food plant: Prunus serotina.

Larval case: Brown, cylindrical; upper edge serrate; mouth deflected to

45 degrees; posterior end triangularly compressed. This species is known only from Zeller's description. It is placed in our specific key with fletcherella only tentatively. If the two are one species, as is quite possible, Zeller's name will take precedence over the better known fletcherella. In our lists occidentis is wrongly listed as a synonym of pruniella Clemens.

(4) C. fletcherella Fernald.

Alar expanse: 10-12 mm.

Locality: Northern United States and Canada (June).

Food plants: Apple, pear, cherry, hawthorn.

Larval case (fig. 127a): Brown; cylindrical; smooth; mouth deflected from 45 degrees to 60 degrees; posterior end triangu-

This is the economically important "cigar case-bearer" on apple, and is easily distinguished from other gray-brown, unicolorous species by its yellowish head. It is probably a synonym of occidentis Zeller, but this can only be decided by further and more extensive rearings. Zeller describes the larval case of occidentis as serrate above. All cases of fletcherella in the United States National Museum have the upper edge smooth.

(5) C. laricella Hübner. (Fig. 124).

Alar expanse: 9-10 mm.

Locality: northeastern United States.

Food plant: Larch.

Larval case: Gray, with a yellowish or brownish patch on the outer side extending from the mouth and formed by a part of the leaf fiber woven into the case; mouth deflected from 45 degrees to 60 degrees; posterior end triangularly com-

This species is commonly known as the "larch case-bearer," and often does considerable damage to the trees.

(6) C. unicolorella Chambers.

Alar expanse: 8-9 mm.

Locality: Eastern United States (June).

Food plant: Seeds of Juncus.

Larval case: Dirty gray, ornamented with many small frass pellets; stout; cylindrical; mouth deflected to 90 degrees; posterior end broad, pyramidal, triangularly compressed; 4.5-5 mm. long.

(7) C. pruniella Clemens.

Alar expanse: 12.5-13 mm.

Localities: Pennsylvania; Ohio. (June).

Food plant: Prunus serotina.

Larval case: A double affair, composed of a small curved, silken first case, inserted into a later one cut from the serrate edge of the mined leaf, thus resembling a pistol butt projecting from a holster (in the spring often losing the silken pos-terior part); mouth slightly deflected; posterior end rounded and flatly compressed; 5-6 mm. long. (8) C. concolorella Clemens. Alar expanse: 10 mm. Locality: Pennsylvania (June). Food plant: Unknown. Larval case: Unknown.

A pale mauve species near cinerella Chambers but apparently distinct.

(9) C. cinerella Chambers. Alar expanse: 10 mm. Localities: Kentucky; District of Columbia. (End of May to July). Food plants: Birch and alder. Larval case: Similar to that of pruniella Clemens; dark rusty brown; roughly cylindrical with a couple of servate projections

on outer side; mouth deflected to 45 degrees; posterior end rounded and flatly compressed.

II. Entire insect of a shining metallic luster; unicolorous, larval case with apex triangularly compressed. Larvæ leaf miners on low plants.

(10) C. aenusella Chambers.

Alar expanse: 11 mm.

Locality: Kentucky.

Food plant: Unknown. Larval case: Unknown.

Known only by Chambers' description. Easily recognized, however, being the only metallic bronzy species in the eastern United States with simple antenna thickened at basal joint only.

(11) C. coruscipennella Clemens.

Alar expanse: 11-12 mm. Locality: United States (May-August).

Food plant: Unknown.

Larval case: Unknown.

A very common species, found nearly everywhere in the United States. Has been frequently identified as the European spissicornis Haw. (fabriciella Vill.). C. auropurpurella Chambers is a synonym.

(12) C. apicialbella Braun (apicella Braun).

Alar expanse: 9.5-11 mm.

Locality: Cincinnati, Ohio. (late May and early June).

Food plant: Silene virginica.

Larval case: Gray, darker on dorsal surface; eylindrical; mouth slightly deflected; 7-9 mm. long.

Easily distinguished from other species of this group by the antennæ. These are clothed with rough, purplish scales for basal four-fifths, the apical fifth being silvery white without annulations but with a black dot on underside of each segment.

III. Moths with fore wings golden to dark brown; costa white, otherwise unmarked; antenna thickened with scales near base, with very slight tuft on basal joint. Larval cases cylindrical and with apices flatly compressed (The socalled "cigar-case" type fig. 127b). Larva leaf miners, mostly on trees and shrubs.

(13) C. rosacella Clemens.

Alar expanse: 10.5 mm.

Locality: Pennsylvania (late May and early June).

Food plants: Rose and sweet briar.

Larval case: Brown; smooth except for one or more servations on upper edge; mouth deflected (from Clemens' description).

The darkest of the brown Coleophoras with white costal markings on the fore wing.

(14) C. albiantennaella Wild.

Alar expanse: 12-13 mm.

Locality: Buffalo, New York (late July).

Food plant: Cornus.

Larval case: Gray-brown; smooth; upper edge not serrate; mouth deflected to 45 degrees; posterior end as broad as, or broader than, middle of case; 11 mm. long.

This species easily distinguished from others of this group by the lack of annulations on the antenna.

(15) C. carpinella Heinrich.

Alar expanse: 7-7.5 mm. Locality: Maryland (June-July).

Food plant: Carpinus caroliniana.

Larval case: Yellowish brown; slender; smooth; mouth deflected to 45 degrees; posterior end as wide as middle of case, flattened to a straight edge; 6-7 mm. long.

(16) C. umbratica Braun.

Alar expanse: 8-9 mm. Localities: Ohio; Virginia (middle to end of June).

Food plant: Prunus americana. Larval case: Brown; rather slender; smooth except upper edge which is strongly serrate; mouth deflected from 45 degrees to 60 degrees; posterior end as wide as middle of case, flattened to a straight edge; 5-6 mm. long.

This species and *carpinella* are superficially very close. They may be dis-tinguished from each other by the characters given in the table, the larval cases, and the food plants; and from the other golden brown species of this group (caryaefoliella and allies), by the distinctly darker shading of the fore wing towards the apex.

(17) C. caryæfoliella Clemens.

Alar expanse: 9-10 mm.

Locality: Eastern United States (May-June in the South; June-July in the North).

Food plants: Hickory and pecan.

Larval case: Brown; smooth; without serration on upper edge; mouth deflected to 45 degrees; posterior end as wide as middle of case, flattened to a straight edge; 6-6.5 mm. long.

This very common species is found nearly everywhere that its food plants occur. It is a rather important enemy of the hickory in the north and of the pecan in the south. In economic literature it is referred to as the "hickory cigar case-bearer" or the "pecan cigar case-bearer." In spite of its importance, comparatively little is known about it, and it should be more intensively studied. There is quite possibly more than one species attacking the hickory, which would account for the wide range of variation in the specimens now included under this name. (18) C. ostryæ Clemens.

Alar expanse: 7.5-8 mm.

Localities: Pennsylvania; Ohio (June).

Food plant: Ostrya virginica.

Larval case: Pale reddish brown; rather wide; smooth except for upper edge which normally has a slight notch near posterior end; mouth slightly deflected; posterior end as wide as middle of case, flattened to a straight edge.

Miss Braun states that she has also reared this species from hickory and considers it a synonym of caryaefoliella Clemens. Until the latter is better known, however, it would seem better to retain the two as separate species.

(19) C. limosipennella Duponchel (laticornella Clemens). Alar expanse: 10.5-12.5 mm. Locality: Northeastern United States (June-July). Food plant: Elm. Larval case: (fig. 127b): Brown; rather broad; smooth except for upper

edge which is more or less servate and slightly sinuate; mouth deflected to 60 degrees; posterior end a trifle wider than middle of case, flattened to a slightly curved edge; 10-11 mm. long.

Locally important in this country as a pest of the elm. In Europe also recorded from alder and birch.

(20) C. corylifoliella Clemens.

Alar expanse: 8-9 mm.

Localities: Pennsylvania; Maryland; Virginia (June-July).

Food plant: Corylus americana.

Larval case: Dark brown; rather broad; rough and fibrous; upper edge serrate; mouth deflected to 90 degrees; posterior end narrower than middle of case, flattened to a slightly curved edge; 6-6.5 mm. long.

(21) C. alniella Heinrich.

Alar expanse: 8-9 mm.

Localities: Maryland; Virginia (June and early July).

Food plant: Alnus.

Larval case: As in corylifoliella Clemens.

The only appreciable difference between reared specimens of this species and corvifoliella is in the somewhat darker and more distinctly marked brown annulations on the antenna of the latter. It is quite probable that it will eventually prove to be a synonym of Clemens' species.

(22) C. lentella Heinrich.

'Alar expanse: 8 mm.

Locality: Long Island, New York (June).

Food plant: Betula lentu.

Larval case: Gray brown; broad, smooth except for upper edge which is markedly serrate; mouth deflected to 45 degrees; posterior end narrower than middle of case, flattened to a straight edge; 4.5-5 mm. long.

1V. Moths with fore wings yellow, unicolorous or golden yellow, more or less striped with white; not dusted or spotted. Antennæ with basal joints thickened or slightly tufted. Larval cases with posterior ends flatly compressed.

(23) C. ochrella Chambers.

Alar expanse: 12 mm. Locality: Kentucky (June).

Food plant: Unknown. Larval case: Unknown.

A dull dark yellow, unicolorous species with basal third of costa a trifle darker than ground color of fore wing. The only described species of this kind from North America.

(24) C. gaylussaciella Heinrich.

Alar expanse: 10 mm.

Locality: Falls Church, Virginia (June).

Food plant: Gaylussacia baccata.

Larval case: Brown; cylindrical; widest just beyond middle; tapering slightly to both ends; mouth deflected to 90 degrees; posterior end rounded; 6-6.5 mm. long.

Close to cretaticostella Clemens but distinguished by characters given in the table.

(25) C. cretaticostella Clemens.

Alar expanse: 12.5 mm.

Localities: Pennsylvania; Canada; Ohio; Massachusetts; Maryland; (May, June).

Food plant: Blackberry.

Larval case: Dark brown; of the *pruniella* type; composed of silk and leaf; slightly irregular in outline but not appreciably curved; posterior end rather abruptly tapering; mouth deflected to 45 degrees; 7-7.5 mm. long.

A very pretty species distinguished by the white costal and dorsal margins and basal part of fore wing.

(26) C. kearfottella Barnes and Busck.

Alar expanse: 11-12 mm.

Locality: Essex County, New Jersey.

Food plant: Salix.

Larval case: Formed of silk and bud scales; very irregular; as broad as long; mouth slightly deflected; 5-6 mm. long.

Close to *cretaticostella* Clemens but distinguished by the much darker yellow ground color of the fore wings.

V. Moths with fore wings dark grayish brown or drab, faintly striped with white. Antennæ with basal joints thickened but not appreciably tufted. Larval cases flatly or triangularly compressed.

(27) C. polemoniella Braun.

Alar expanse: 13-14 mm.

Locality: Cincinnati, Ohio (May and early June).

Food plant: Polemonium reptans.

Larval case: White with a gravish patch on dorsal surface adjacent to mouth; rather slender, cylindrical; mouth deflected to 45 degrees; posterior end flatly compressed and flaring, considerably broader than middle of case; 9-10 mm. long. An easily recognized species. In some specimens the white markings nearly

An easily recognized species. In some specimens the white markings nearly obsolete except on costa.

(28) C. astericola Heinrich.

Alar expanse: 13 mm.

Locality: Boston, Massachusetts (September).

Food plant: Aster multiflorus.

Larval case: Gravish white; slender; cylindrical; mouth deflected to 45 degrees; posterior end rounded and flatly compressed, narrower than middle of case; 13 mm. long.

 (29) C. infuscatella Clemens. Alar expanse: 11 mm. Locality: Pennsylvania. Food plant: Unknown. Larval case: Unknown.

VI. Moths with fore wings white, longitudinally streaked with yellowish white or whitish-yellow streaked with white, with or without fuscous or ochreous dustings on white areas. Antennæ with basal joints thickened but never appreciably tufted. Larval cases with posterior ends triangularly compressed. Larvæ seed feeders or leaf miners on low plants.

 (30) C. quadrilineella Chambers. Alar expanse: 7 mm. Locality: Eastern United States (June through August). Food plant: Seeds of *Juncus*. Larval case: Gray; covered with particles of frass on posterior fourth; along upper anterior portion of three-fourths of its length a shining shield formed from part of the seed capsule of the food plant: cylindrical; slender; mouth deflected to 90 degrees; 4-4.8 mm. long.

- (31) C. borea Braun. Alar expanse: 15-16 mm. Locality: Cincinnati, Ohio. Food plant: Polygonum scandens. Larval case: Blackish brown; stout; cylindrical; month slightly deflected; 8-9 mm. long (From Miss Braun's description).
- (32) C. biforis Braun.
 Alar expanse: 11.5-12.5 mm.
 Locality: Cincinnati, Ohio.
 Food plant: Lazula campestris.
 Larval expanse: "isoladar oulindria
 - Larval case: "slender, cylindrical, tapering to the three-valved apex. The sides of the obtuse apical angles of the valves are very short; from the point of union of adjacent valves, a gradually lowering ridge runs about halfway down the case"; S-8.5 mm. long. (From Miss Braun's description).
- (33) C. coenosipennella Clemens. Alar expanse: 11.5-12 mm. Locality: Eastern United States. Food plant: Stellaria pubera. Larval case: Grayish; "decorated with numerous dark reddish granules;"
- (34) C. cratipennella Clemens.
 (34) Alar expanse: 14.5 mm.
 Locality: Eastern United States (May and early June).
 Food plant: Seeds of *Juncus*.
 Larval case: Grayish ochreous, shading to reddish brown at apex; cylin
 - drical; smooth; unornamented; mouth deflected to 45 degrees; 8 mm. long.
- (35) C. shaleriella Chambers.

Alar expanse: 13.5-15 mm.

Localities: Kentucky; Ohio (August to September).

Food plant: Seeds of Polygonum punctatum.

Larval case: Pale straw color; elongate; slender; tapering gradually to pointed apex; decorated from mouth with four or five narrow strips of leaf extending backward four-fifths the length of the case, and by seven or eight distinct lines of frass extending backward about ¹/₄; 10-11 mm. long.

(36) C. caespititiella Zeller. Alar expanse: 11-12 mm. Localities: Maryland; Virginia (May-June). Food plant: Seeds of Juncus. Larval case: Pale straw color to pale gray brown; cylindrical; smooth; undecorated; mouth deflected to 45 degrees; 6-6.5 mm. long.

(37) C. ericodes Braun.
 Alar expanse: 12 mm.
 Locality: Cincinnati, Ohio (September).
 Food plant: Seeds of Aster ericodes.
 Larval case: Pale straw color to ashy brown; rather densely covered with pappus from flowers of food plant which extend beyond the apex; mouth deflected; 4.5-5 mm. long.

(38) C. vernoniælla Chambers (fig. 125) (veroniæella of Dyar's list). Alar expanse: 11-13.5 mm. Localities: Kentucky; Missouri; Ohio; Virginia (late June to mid-July). Food plants: Vernonia, Helianthus. Dark gray brown; slender elongate; tapering; needle-like; Larval case: mouth deflected from 20 degrees to 45 degrees; 13-20 mm. long. (39) C. duplicis Braun. Alar expanse: 11.5-14.5 mm. Localities: Ohio; North Carolina. Food plants: Aster shortii, Aster cordifolina, Solidago cæsia, S. latifolia. Larval case: Irregular, formed from bits of seeds and flowers attached to a cylinder of silk; mouth deflected. (40) C. granifera Braun. Alar expanse: 12.5-14 mm. Locality: Cincinnati, Ohio (June-July). Food plant: Leaves of Aster shortii. Larval case: Brownish red, paler towards apex, with a dark brown, granular spot on upper side near mouth; cylindrical; mouth slightly deflected; 10 mm. long. (41) C. fagicosticella Chambers (synonym, C. lineapulvella Chambers). Alar expanse: 8-10 mm. Locality: Eastern United States (end of May, through June). Food plant: Seeds of Juncus. Larval case: Composed of silk and entire seed pod of food plant, bractlets forming an outer scalloped enveloped for silk portion of case protruding beyond; mouth deflected to 90 degrees; 5 mm. long. A very common species. (42) C. amaranthella Braun. Alar expanse: 12-14 mm. Localities: Kentucky; Ohio (late July-August). Food plant: Seeds of Amaranthus hybridus. Larval case: Brownish, decorated with numerous, small, buff-gray particles; mouth deflected; apex pyramidal. 6 mm. long (43) C. vagans Walsingham. Alar expanse: 12 mm. Locality: New York City (August). Food plant: Grass. Stone gray with brown patch on upper side near mouth Larval case: (similar to quadrilineella Chambers); cylindrical, slightly bulged in center; mouth deflected to 90 degrees; 7-7.5 mm. long. Distinguished from other grass- and Juncus-feeding species by the heavy fuscous dustings nearly obscuring the whitish lines on fore wing. (44) C. lapidicornis Walsingham. Alar expanse: 11.5 mm.

Locality: Ohio (August).

Food plant: Peach.

Larval case: Dark, dusty gray brown; smooth; cylindrical; rather stout; scarcely tapering; mouth deflected to 90 degrees; 7 mm. long.

VII. Moths with fore wings white, unmarked, or shading to yellowish toward apex, not longitudinally striped or spotted; antennæ simple or with slight tufts. Larval cases formed of silk and leaf; posterior ends flatly compressed. Larva leaf miners in trees and shrubs.

(45) C. argentialbella Chambers. Alar expanse: 10-11 mm. Locality: Kentucky. Food plant: Unknown. Larval case: "Long and slender" (Chambers). Easily recognized, being the only pure white species with simple antenna described from the eastern United States.

(46) C leucochrysella Clemens Alar expanse: 14-15 mm. Locality: Eastern United States (June). Food plant: Castanea dentata. Larval case (fig. 127): Irregular; of the pruniella type, with silken first

case inserted in later one cut from mined portion of leaf; silken part grayish tinged with black; leaf part yellow or brownish; mouth deflected to 45 degrees; posterior end curved; 10-11 mm. long. An easily recognized species; fairly common wherever its food plant grows.

(47) C. viburnella Clemens.

Alar expanse: 11.5 mm.

Localities: Pennsylvania; Ohio (end of May).

Food plant: Viburnum prunifolium.

Larval case: Of the pruniella type; upper edge with flattened, serrate, winglike projection rising about 2.5 mm. above the barrel of the case; reddish brown; mouth deflected; posterior end slightly curved; 10 mm. long.

The basal joint of the antenna is somewhat more tufted than that of the other species in this group. The moth, however, is easily recognized by the characters given in the table.

VIII. Moths with large projecting tuft on basal joint of antenna. Larval cases of the "pistol" type (fig. 126), made of silk and frass; posterior ends never triangularly compressed. Larvæ external feeders on leaves or flowers of trees and shrubs; never miners.

(48) C. nigralineella Chambers.

Alar expanse: 12-13 mm.

Locality: Kentucky (July).

Unknown. Larval case found attached to stem of Juglans Food plant: nigra.

Larval case: Yellow; pistol handle brown on upper side, white underneath; on upper side, near mouth, a small triangular projection used for attachment of case during pupation (from Chambers' description).

(49) C. tiliæfoliella Clemens.

Alar expanse: 14-14.5 mm.

Pennsylvania; New York; Ontario, Canada (June-July). Localities: Food plant: Tilia (basswood).

Larval case: Uniform black; pistol handle turned down abruptly (at right angle with upper edge); small toothed projections about middle of under edge; upper edge straight; flaps present, small, appressed; mouth deflected to 45 degrees; 6.5-7 mm. long.

(50) C. rosæfoliella Clemens.

Alar expanse: 12.5 mm.

Locality: Pennsylvania (end of May).

Food plant: Buds of common hundred-leaf rose.

Larval case: Of silk, covered with granulations; brown varied with gray; posterior end (pistol handle) turned down like a hook; mouth slightly deflected (from Clemens' description).

I have never seen the case, and the species is therefore only tentatively placed in this group. (C. H.)

(51) C. malivorella Riley.

Alar expanse: 12.5-14 mm.

Locality: Atlantic States (early July).

Food plant: Apple.

Black with large, whitish fibrous patch along upper edge; Larval case: handle short and evenly rounded; small, toothed projec-tion on underside close to handle; flaps present, scalloped, small, and closely appressed within angle of handle; mouth deflected to 45 degrees; 8-9 mm. long. This is the well-known "pistol case-bearer" on apple. The larvæ eat the buds,

leaves, and young fruit.

(52) C. atromarginata Braun (currucipennella Walsingham nec Zeller).

Alar expanse: 10.5-14.5 mm.

Locality: Eastern United States (June-August).

Food plants: Quercus platanoides and Quercus rubra. Larval case:

Black, with faint white V marking on underside of barrel; handle turned down abruptly; flaps present, moderately large, not closely appressed; mouth deflected to nearly 45 degrees; 5.5-8 mm. long.

(53) C. elæagnisella Kearfott.

Alar expanse: 15-21 mm.

Locality: Ottawa, Canada (July).

Elæagnus argentea; Hippophoe rhamoides; Shepherdia Food plants: argentea.

Larval case: Grayish brown; elongate; without flaps; pistol handle reduced. giving a sort of scimitar shape to the case; upper edge smooth; a slight projection from near middle of lower edge; mouth deflected to 45 degrees; 10-12 mm. long.

(54) C. querciella Clemens.

Alar expanse: 12 mm.

Localities: Pennsylvania; Virginia; Long Island, New York. (June). Food plants: Quercus prinus and Q. alba.

Larval case: Gravish, with conspicuous black, somewhat lumped patch on back near posterior end; without flaps; handle much reduced, turned down at 45 degrees angle with upper edge; a slight projection back of middle on lower edge; mouth deflected to 45 degrees; 9-10 mm. long.

(55) C. atlantica Heinrich (fig. 123).

Alar expanse: 11-15 mm. Locality: Eastern United States.

Food plant: Prunus serotina.

Larval case (fig. 126): Black; handle evenly curved; flaps present, rather small and appressed; mouth deflected to about 45 degrees; 8-9 mm. long.

This is the pistol case species wrongly identified by Kearfott as C. pruniella and so listed in the New Jersey list.

SUPERFAMILY CYCNODIOIDEA

(Tineina, in part)

The Cycnodioidea are a small group, composed of isolated remnants of a primitive type which appears to have preserved \mathbf{R}_{2+3} in the hind wings, as an independent vein. In the African genus Cycnodia, (fig. 128) the vein is fully preserved, as well as all the normal veins of the wing; in Aphelosetia, which is otherwise extremely close to Cycnodia, it appears to be lost; in Tinagma (fig. 131) it is preserved, but several dorsal veins are lost; while in the Heliozelidæ the venation is so reduced that the point cannot be settled. A distinctive character of these reduced forms, not so well shown in Cycnodia, is the unusually wide space between **Sc** and **R**, so that **R** forms the axis of the wing, instead of **M**, which takes that place in some other narrow-winged forms.

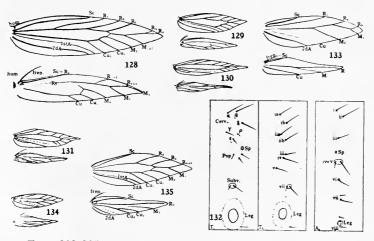
Head smooth; first joint of antennæ quite small; palpi moderate and slightly upturned or small, not bristled; maxillary palpi minute; hind tibiæ hairy. Fore wing with \mathbf{R}_5 running to costa when recognizable, but generally lost; with some dorsal veins usually lost also. Venation extremely reduced in *Coptodisca*. **1st A** free, **2d A** usually simple, but forked in Tinagma and some Aphelosetias. Hind wing narrower than fore wing; **Sc** and **R** strong, \mathbf{R}_1 usually not developed, but when present located close to base of wing; stem of **R** running nearly through axis of wing, in the narrow-winged forms with the outer veins branching from it. Male genitalia complex and characteristic.

Eggs of flat type as far as known; larvæ leaf-miners, at least when young — the Heliozelidæ making a flat case before pupation exactly as in many Adelidæ. Pupæ various, never obtect.

Family 16. CYCNODIIDÆ

(Elachistidæ)

Head smooth or slightly ruffled on vertex; palpi slender and smoothly sealed, somewhat upturned or porrect, often strongly divergent, third segment well-developed; no ocelli; no maxillary palpi. Antenna with scape small, with pecten, shaft scaled more or less roughly all around, with two equal whorls to a segment. Tongue weak, scaled at base. Eyes small. Fore wing broad-lanceolate, the cell located in the middle, and cut off obliquely at its outer end (fig. 129) \mathbf{R}_1 arising from **R**-stem at middle of cell, \mathbf{R}_4 and \mathbf{R}_5 stalked or united, \mathbf{M}_1 stalked with them, sometimes with \mathbf{R}_5 beyond the point of separation of \mathbf{R}_4 , \mathbf{R}_3 sometimes also stalked; \mathbf{M}_3 and \mathbf{Cu}_1 completely united, sometimes lost (A. praematurella). Hind wing narrower, lanceolate, with \mathbf{R}_1 strong, but



FIGS. 128-135. CYCNODIIDÆ, DOUGLASHIDÆ, AND HELIOZELIDÆ

128, Cyenodia cygnipennella (North Africa), venation; 129, Aphelosetia argentella (Europe), venation; 130, Douglasia ocnerostomella (Europe), venation; 131, Tinagma dryadis (Europe), venation; 132, Douglasia balteolella (Europe), seta map of larva; 133, Coptodisca species (on grape), venation; 134, Antispila treitschkiella (Europe), venation; 135, Heliozela stannella (Europe), venation (after Spuler)

erowded to extreme base of wing; **Sc** and \mathbf{R}_s widely separated, **Sc** in Aphelosetia ending at middle of costa; \mathbf{R}_{2+3} lost in Aphelosetia, present as a short spur in Cycnodia; \mathbf{R}_{4+5} running to costa near apex, \mathbf{M}_1 (in our species) stalked with it; \mathbf{M}_2 lost in Aphelosetia; \mathbf{M}_3 associated with **Cu**-stem, the cell usually weakly closed. Anal region reduced.

This description is based on Aphelosetia, the only genus in castern North America; in Cycnodia all the veins are present, and \mathbf{R}_s of the hind wing is less widely separated from **Sc**. Superficially the moths are like Cosmopterygidæ; but can be distinguished by the radically different hind wing; the moderately long, divergent palpi are also rare in the Cosmopterygidæ.

The larvæ of Aphelosetia are leaf-miners on various grasses and sedges; they are flattened, with normal mouth-parts and sixteen legs. The mines vary in character, and the frass, unlike that of Cosmopteryx, is left in the mine. The pupa is formed exposed or in a mesh-work cocoon, and is attached by a band around the middle. In several species there are three longitudinal raised dark stripes. All its parts are fused, the antennæ nearly as long as the wings, not separated by the legs or tongue; labial palpi eovered and maxillary palpi absent. The headsclerites are of geleehiid type; cremaster without spines or hooks; and frontoelypeal suture absent. The body is roughened with rounded tubereles.

APHELOSETIA Stephens

(Elachista Treitsehke, in part; Cosmiotes Clemens; Eurynome Chambers, not Leach; Buschia Dyar, Phigalia Chambers, not Duponehel; Aphigalia Dvar)

The characters are given in the description of the family. There is some variation in structure, especially in venation, but it is not large, and as there is some confusion in names no attempt is made here to divide the genus. Elachista concolorella Chambers is an Eriphia, but is not the Eriphia concolorella described by him on the same page. E. orichalcella, Dr. Braun informs me, is also a Cosmopterygid.

Key to the species (Braun)

1. Ground color dark brown or gray, with white or silvery spots and fasciæ. 3. Veins Cu_2 , $M_3 + Cu_1$, and M_2 arising opposite the space between R_2 and R_3 in fore wing. 4. A silvery spot at tip of wing membrane. 5. Antennæ silvery white at tip.....4. madarella. 4. Costal streak at two-thirds way to apex, eurving outward in middle 6. Head silvery gray, apex of antennæ white......9. sylvestris.
6. A white patch on face, antennæ fuscous, annulate..10. leucofrons.
2. No white or metallic patch at base of fore wing.

- 3. No apical white spot.
 - 4. A silvery costal streak in fringe beyond the pair of costal and dorsal streaks12. illectella.
 - 4. A costal streak only, beyond the fascia.....13. maculosella.
 - 4. Fascia followed by a costal and an opposite dorsal streak; no other markings.
 - 5. Fascia and streaks metallic white.....14. unifasciella.
 - 5. Fascia and streaks dull white......15. irrorata.

1. A. orestella Busek. White, slightly vellowish; antennæ smoky on outer half, fore wing with a slight vellow tinge in fold, with black dots at middle of wing and at end of cell; abdomen and legs somewhat smoky. 11 mm.

Larva gravish or green, with two more or less prominent dark dorsal stripes on prothorax, ending behind in a pair of spots. Mine on Hystrix, beneath the

upper epidermis (which is turned down by a twist in the leaf), 3 or 4 cm. long, gray for the most part, but with a wrinkled central stripe which remains green. Larva wintering in the mine, sometimes moving to a new leaf in the spring. Pupa in a very slight cocoon, stout, ovate, with four rather prominent mesothoracic tubercles; smooth and shining. Moth in May to July and September.

New York to Ohio. New York: Peru, Perry, Ithaca. 2. A. brachyelytrifoliella Clemens. Dark grayish brown, apical half of fore wing black-brown, base light; oblique white streaks from costa a third way, and which show the second almost reaching one which rises up from the dorsal margin. A white stripe along base of costa, joining the first transverse streak; two white streaks edged with black in costal fringe; a black apical spot, and a white dot in dorsal fringe. Head in front, and palpi, silver white. 6 mm. Larva mining a grass, *Brachyelytrum aristatum*, early in July; mine at first linear, then a blotch taking up most of the leaf. Pupa external, on a slight web linear.

web. Imago emerging in late July.

Easton, Pennsylvania.

3. A. enitescens Braun. Head and thorax metallic-leaden, the head nearly black; antennæ gravish black throughout; rest of fore wing dark brown. A silvery or and the gray is black throughout, less of hole wing dark blown. A silvery of golden fascia a third way out, oblique inward on costal half, broader and erect on dorsal half; two spots opposite each other at two-thirds, and a spot at apex of membrane; base of inner margin sometimes with silvery or golden scales. Metallic markings with some reddish or purple luster. Fringes and upper side of abdomen dark gray; hind wing dark brown. $7-7\frac{1}{2}$ mm.

Larva on Scirpus atrovirens (bulrush), forming a long transparent mine in a basal leaf, extending from the base upward; wintering in the mine, and moving to a second leaf after some feeding in the spring. Larva yellow with a pair of ill-defined dark patches on posterior part of prothorax; Pupa in a light flat eccoon of rhomboidal meshes. Pupa stout, with dorsum of abdomen flattened, and with a subdorsal series of short blunt spines; thorax with three or four prom-inent lateral tubercles. Moths in May and early June.

The larva feeds only at night, retiring by day to the base of the leaf, sometimes beneath the water level.

Cincinnati, Ohio.

4. A. madarella Clemens. Similar to A. enitescens; head paler, tips of antennæ white, base of fore wing more golden and legs paler. 8 mm. (Cosmiotes Clemens).

Larva on Carex, especially C. pubescens and cristata, and on Scirpus; mines similar to those of A. enitescens, several being made by a single larva, the larva with the same habit of retiring to the base of the mine by day. Larva pale green or whitish, with a pair of prominent, dark brown spots, sometimes L-shaped, on the prothorax. Cocoon and pupa similar to A. enitescens, pupa broader with rougher thorax, and a beaded ridge with a broad sinus at its middle, across the vertex. Moth in late May and June, rarely flying into July.

Pennsylvania and Ohio; abundant in Ohio.

5. A. argentosa Braun. Thorax deep golden brown, shading into metallic gray behind; head also metallic gray, antennæ wholly black-brown. Fore wing nearly black, with a fascia and costal and dorsal streaks as in A. enitescens; but bluishsilvery, the fascia produced outward a little on the inner margin, and the costal streak curving out at its lower end to the apex of the membrane, where it may be broadened, and sometimes meets the dorsal streak. Legs silvery gray, middle tibiæ and all tarsi dark brown, with tips of segments silvery. 7-71/2 mm.

Larva in a narrow-leaved Carex, mine extending down from the tip of a leaf, lying nearer the upper epidermis, the larva consuming most of the parenchyma. Epidermis slightly wrinkled at point of exit of larva, which is not always at the extreme base of the mine. Pupa in a slight cocoon of very irregular meshes; stout, ovate, shining, and chitinous, with rows of minute tubercles on dorsum

and prominent tubercles on sides of mesothorax, faint lines of tubercles on wings, and prominent tubereles on front of head. Moth in June.

The costal streak of this species corresponds to the fused costal streak and apical spot of A. madarella.

Clermont County and Cincinnati, Ohio.

6. A. prælineata Braun. Black, face lead-colored, palpi fuscons, paler above, with two bands on third segment; tips of tegulæ and a few scales on disc of thorax white; fore wing with white base, a slightly curved erect fascia two-fifthsway out, and costal and dorsal spots nearly opposite each other, four-fifths-way

out; apical fringe white. 7 mm. Mine on *Hystrix patula*, at first a line, then a blotch 4 to 5 mm. wide, usually on the under side of the leaf. Mine in early July; moth in August. Pupa stout and shining, heavily tuberculate.

Ohio

7. A. cucullata Braun. Black; palpi white, second joint brown outwardly, third sometimes with fuscous shading outwardly; antennæ black; face and head white; fore wing beyond base slightly brownish; a silvery fascia almost at base, broader on dorsum, a second fascia just beyond middle, bent at middle of wing, and not quite reaching inner margin; a silvery subapical costal streak, and a dorsal triangle a little before it. Fringe and hind wing dark. 8-9 mm.

Larva in Carex Jamesii; mine, in the fall, a narrow tract, which is widened into a broad, inflated tract covering most of the width of the leaf, in the spring. Larva red, with browner head; thorax with pinkish mid-dorsal line, and abdomen with lateral pinkish lines also. Pupa elongate, tapering, with prominent dorsal and lateral ridges, the former bifurcated in front and ending on a pointed hood projecting over the face; pupa suspended by a girth and a few strands of silk. Moth emerging from middle of May to early June.

Cincinnati, Ohio. New York: Ithaca.

8. A. albicapitella Engel. Dark brown with purple iridescence. Antennæ dark brown, tips gray; face silvery; under side of second segment of palpi gray; lower edge of tegulæ and back of thorax white. Fore wing with base and first fascia as in A. prælineata, and with costal and dorsal streaks four-fifths way to apex, sometimes meeting; fringe powdery dark gray. 8 mm.

Larva in overwintering lower leaves of Poa sylvestris; the early part of the mine with parenchyma partly eaten, the later part somewhat inflated, and with almost all the parenchyma eaten. Larva pale yellowish, with two dark dorsal stripes on prothorax, widening and sometimes becoming confluent behind; mid-dorsal line whitish. Pupa stout, ovate, with prominent tubercles on sides of mesothorax, in a slight cocoon of transverse threads. Larvæ leaving mine at end of March, moths emerging in May or June (Ohio).

Pennsylvania and southern Ohio; Wisconsin. 9. A. sylvestris Braun. Blackish brown, with a faint golden brown luster. Face and front of head silvery gray; palpi with outer side of second joint and a little of apex of third blackish; apical fifth of antennæ whitish; tips of tegulæ and of mesothorax silvery. Fore wing with a silvery basal patch, a nearly vertical, slightly irregular fascia just before middle, and large subterminal dorsal and costal spots, the latter farther toward apex.

Larva in stem-leaves of Poa sylvestris. Mine at first linear, at the margin of a leaf, inconspicuous; later mine in a second leaf, white, extending the entire width of the leaf and about 8 cm. long. Larva wholly pale yellow; pupa slender, elongate, dull except toward the head, with irregular tubercles on mesothorax; without cocoon. Mines in May; moths emerging in early June.

Cincinnati, Ohio.

10. A. leucofrons Braun. Blackish, slightly irrorated; face below antennæ with a white patch; palpi black below, with base of third segment white, white above with tip of third segment black; antennæ annulate with gray and fuscous; tegulæ

with white tips; fore wing with irroration sometimes forming faint longitudinal lines. Base of fore wing white, fascia narrow and irregular, oblique on costal and erect on dorsal half; a subterminal spot on inner margin, and a spot further out on costa, rarely obsolescent. Fringe with a line of black dots in base. Tibiæ and tarsi blackish, tips of segments and basal half of hind tibia white. 9-10 mm.

Mine on Hystrix and especially Elymus; whitish, on upper side of leaf, and usually extending its whole width, the under side of the leaf remaining green. Larva pale greenish or grayish, with narrow dorsal and broad lateral whitish lines, and a curved brown mark at rear of prothorax. Pupa attached flat to the leaf, head up; slender, elongate, not shining; with low rounded tubercles on sides of mesothorax and small ones on head; yellowish gray. Larva overwintering, the moths emerging in May.

Cincinnati, Ohio.

11. A. præmaturella Clemens. Palpi white above, fuscous below. Fore wing dull purplish black, dusted with white; head, thorax, and base of fore wing grayish fuscous, somewhat contrasting; antennæ darker. Fore wing with a white fascia before middle, and costal and dorsal spots toward apex, almost meeting. Extreme apex white, a row of dark scales in the fuscous fringe. $6-7\frac{1}{2}$ mm. (albapalpella and cristatella Chambers).

Larva on nodding wild rye (*Elymus canadensis*); mine linear, becoming a blotch as wide as the leaf, 10-11 cm. long. Moth in April to June and August. Late specimens tend to be suffused.

Pennsylvania, Ohio.

12. A. illectella Clemens. Fuscous, dusted with dark brown. Head and palpi yellowish, antennæ fuscous. Fore wings with the fascia located before the middle, and spots nearly meeting four-fifths way out; a silver streak in costal fringe, edged with black. (Cosmiotes Clemens.)

Pennsylvania.

13. A. maculosella Clemens. Blackish, suffused with dark golden brown; head dark brown, palpi dull yellowish, antennæ fuscous. Fore wing with fascia at middle silvery; a spot on costa near apex; apex blackish, fringe grayish brown. 7 mm.

I have seen only the type, which is too poor to recognize.

Pennsylvania.

14. A. unifasciella Chambers. Brownish, iridescent with reddish purple. Palpi white. Fore wing with fascia just before the middle oblique, white, the ground much deeper beyond it; a small white spot just before the dorsal fringe begins, with a narrow white streak nearly crossing the wing just beyond it. Legs spotted with yellowish white. 6 mm.

Canada.

15. A. irrorata Braun. Dark gray, heavily dusted with black; palpi black beneath, gray above; antennæ black. Fore wing with a narrow, irregularly indented fascia just beyond one-third the wing-length; and costal and dorsal white dots just beyond two-thirds. Fringe with a series of black dots in base. Hind wing concolorous. Pale under side of abdomen less contrasting than usual. 8 to 11 mm.

Larva in leaf of *Glyceria nervata* (as a stray on *Agrostis*) in wet places. Mine linear, very narrow, yellowish green; starting at the base and terminating near the apex of the leaf, the larva retiring to the base by day, and feeding at night; more rarely in a short, detached mine at apex of leaf, which is deserted by day. Larva yellow when young, glaucous above when grown. Pupa attached to the upper side of a leaf near its base, head downward, broader and flatter than *A. leucothorax*, with more tubereles on mesothorax, and stronger lateral ridges. Larva in early spring; moth in late May and early June.

Ohio; common.

Family 17. DOUGLASIIDÆ

(Elachistidæ; Glyphipterygidæ, in part)

Head about as in the other two families of the Elachistoidea; palpi intermediate in size, stout and drooping, the lower part of the face rather more smoothly scaled. Ocelli very large. Fore wing lanceolate; \mathbf{R}_{1} , when present, free from \mathbf{R}_{4} , but stalked with \mathbf{M}_{1} , running to costa; sometimes a radial absent; number of dorsal veins varying, but apparently all present in our species; 1st A free, but weak; 2d A strongly forked at base. Hind wing narrow-lanceolate, Sc ending about at middle of costa; R-stem running through middle of wing, bearing \mathbf{R}_{2+3} on its anterior side two-thirds way out; one medial arising before, and one beyond **R**₂₊₃. **Cu** simple, free. Larva a leaf-miner on Rosaceæ and related plants. Larva short

(fig. 132), fusiform, with normal head and eyes; front extending twothirds way to vertex and adfrontals reaching vertex. Cervical shield with six setæ, arranged in a hexagon; prespiracular wart with three large setæ, and subventrals of all three thoracic segments with two; ia and ib obliquely placed, much like i and ii of the abdomen. Abdomen with i and ii, iiia and iii obliquely placed, similar, approximate, iv and v equal, approximate, and on a level; two upper setæ of vii widely separated from the lower one, which is on the leg base; prolegs small, rather near midventral line, apparently without hooks; proleg of seventh segment smaller than the others; ninth segment with setæ iv lower than ii, nearest to iii. Spiracles circular, pupa not studied.

This larva appears as aberrant as the moth, and helps little in placing the family, which is only tentatively associated with the Cvenodiidæ.

TINAGMA Zeller

(Douglasia, in part)

Palpi rough-scaled below; wing scaling not shining. Antennal segments of normal length, with single whorls of very slender scales (fig. 131). Larva on Potentilla, Geum, and related Rosaceæ.

1. T. obscurofasciella Chambers. Blackish, dusted heavily on a dirty white ground, the effect mouse-gray, the bands yellowish brown, lightly dusted and more or less defined with black. A broad band, with slightly excurved outer boundary at middle of wing, and extending almost to base, lightly edged with white; and a more irregular diffuse fascia near apex. not reaching costa; apex vellower. 7 mm. (crenulellum Engel; Douglasia Chambers.) Moth probably generally distributed but overlooked; flying in May. Ontario to southern Ohio. New York: Black Brook (Clinton County), Rock

City (Cattaraugus County).

LEPIDOPTERA OF NEW YORK AND NEIGHBORING STATES

Family 18. **HELIOZELIDÆ**

(Elachistidæ, in part)

Head smoothly scaled, broad, with small eyes, like the Elachistidæ, but with the palpi shorter and drooping. Antennæ short, with thick joints, scape small, shaft with scaling as in Elachistidæ. Tongue short, with some scaling at base, the base covered over with a rough tuft of scales between the palpi. Maxillary palpi absent. Hind tibiae with stiff hair. Fore wings lanceolate, with four veins running to costa and four to inner margin from the central cell; or venation more reduced, without cell, \mathbf{R}_5 and \mathbf{M}_1 stalked, forking over the apex. Anal region rather broad, the anals free and simple. Accessory cell never indieated. Hind wing with \mathbf{R} widely separated from \mathbf{Sc} , the three medials arising from it separately, or venation reduced, with a single free medial. **Cu** separate; forked, or simple; anal region reduced.

Larva forming a blotch mine, which is small, apparently indicating that the larva feeds largely on sap in spite of its mandibles being of the biting type; frass voluminous, sometimes nearly filling the mine. Larva at pupation cutting out an oval piece of the mined leaf, of which it forms a lenticular case much like that of the Adelidæ. Legs wanting; front reaching vertex (?); abdomen with setae **iv** and **v** rather close together. Larva strongly flattened, as usual in leafminers. Pupa with all appendages free and separate; very lightly chitinized, with spines on dorsum in obscure patches, hardly developed; abdomen with third to seventh segments free, and eighth in male. No maxillary palpi; labial palpi expessed; antennæ half as long as wings. Labrum forming a well developed free lobe projecting over base of labial palpi.

In habits and pupa the family is distinctly primitive, but in larval and imaginal structure, more specialized than *Aphelosetia*.

Key to the genera

- 1. Fore wing with lanceolate cell; hind wing lanceolate.
 - - 2. **R**₁ absent; find wing with a single medial, free from **R**. (Fig. 135). Heliozela.
- 1. Fore wing without cell; hind wing linear. (Fig. 133)..... Coptodisca.

ANTISPILA Hübner

Fore wing (fig. 134) with \mathbf{R}_{3+4} and \mathbf{R}_{5} sometimes stalked with \mathbf{M}_{1} . Hind wing with \mathbf{M}_{1} occasionally lost, **R** running to apex or just above.

The species are dark brown, with more shining silvery face, the fore wing with a fascia a third way out, and usually spots at two-thirds, the dorsal spot rather nearer the base. The key may not apply invariably and the species are best indentified by breeding.

8

Key to the species

1. A white apical spot.

2. Outer joints of antennæ white.....l. hydrangiæella.

2. Only the minute last two joints of antennæ white......2. ampelopsiella. 1. No apical spot.

- 2. Costal spot moderate.
 - 3. Fascia curved, turning obliquely in to inner margin at basal angle; narrow, half as wide on inner margin as the dark areas before and beyond it.
 - 3. Fascia straight, rather beyond basal angle, at inner margin nearer to silver spot than to base, and as wide as the dark areas before and beyond it.

4. Antennæ with white apical segment; ground usually coppery.

4. cornifoliella.

1. A. hydrangiæella Chambers. Similar to ampelopsiella, the apical spot bright silver and as large as the pre-apical ones. 5 mm.

Mine and case like those of A. viticordifoliella on Hydrangea nivea. Moth in August.

Kentucky; southern Ohio.

2. A. ampelopsiella Chambers. A small dot of silver scales at apex. 5 mm. Larva on Ampelopsis. Moth in August. Kentucky; Missouri; Ohio; New York: Fort Edward.

3. A. major Kearfott. Middle third of antennæ silvery white; fore wing with base broadly metallic; fascia very narrowly interrupted at middle; dorsal dash narrow, extending up and out, about half in the fringe; a fascia across the wing just before the apex, not extending into the fringe, nearly broken at the middle, and zig zag like a thick S. 9 mm.

Moth in early June.

Black Mountains, North Carolina.

4. A. cornifoliella Clemens. Antennæ dark brown, scape somewhat ochreous. Fore wing not very bright, with a coppery hue; fascia rather narrow, not constricted on the fold, less distinct, and suffused with copper toward costa. Ground bright coppery toward apex, with the usual two golden spots. 71/2 mm.

The larva mines Cornus florida in September. It is white, with dark brown head and neck, and series of dorsal and ventral dots.

I have seen material from Cineinnati, Ohio.

5. A. isabella Clemens. Almost like A. viticordifoliella; sometimes with the greenish iridescence of nyssafoliella. 6 mm. The moth has been found in May. The larva mines grape, cutting out a cir-

cular case; it is yellowish white, with brown dots at the middle of the body dorsally and a single ventral one.

Pennsylvania; Missouri.

6. A. viticordifolielia Clemens. Moth brown with a brillant coppery tinge and silver fascia and spots; differing from ampelopsiella only in the lack of the minute silver apex, and probably not a distinct species.

Larva yellowish green, without dorsal or ventral spots; cutting out a small oval case for pupation.

Pennsylvania.

7. A. nyssæfoliella Clemens. Antenna dark brown with the scape yellow, as usual; green tint of ground variable, and not strong; golden markings broad, the faseia much broader at inner margin than at costa; outer spots more obliquely placed than in the grape species. $7\frac{1}{2}$ mm. Larva with dorsal spots rather fine, and a couple of ventral ones forming a

Larva with dorsal spots rather fine, and a couple of ventral ones forming a dark line; mining on Nyssa multiflora; case oval.

The moth emerges in May and ean be certainly identified only by breeding.

Pennsylvania; New York; New Jersey; District of Columbia. New York: Mohonk Lake (larvæ); Glens Falls and New York City (Lintner).

There is also a species on sweet fern, which has not been distinguished from A. isabella.

HELIOZELA Herrich-Schæffer

Fore wing (fig. 135) with only three veins running to costa; hind wing with only one medial. Our species has not been examined structurally, and may be an Antispila, but the markings are rather as in Heliozela.

1. H. æsella Chambers. Deep purple-brown, with two fasciæ, starting from • inner margin near base and at middle of wing, tapering above, and only reaching halfway to costa; the basal fascia broader and blunter. 6 mm. Larva in a flattened gall on leaves of grape; the gall lying on both sides of a

Larva in a flattened gall on leaves of grape; the gall lying on both sides of a vein, and almost completely eaten out before the maturity of the larva. Pupal case cut from the epidermis of the gall, at first oval, but made over into a spindle shape. Larva mature the middle of June; moth emerging about the first of the following May.

District of Columbia to Ohio.

COPTODISCA Walsingham

(Aspidisca Clemens, not Ehrlich)

Head and body characters like Antispila; eyes very small and not visible from above. Fore wing (fig. 133) without cell, the R-stem giving off \mathbf{R}_1 from its anterior side near base, and two veins to inner margin near apex; the main stem terminating in costa near apex (probably \mathbf{R}_{1+5}). Rest of M and Cu obsolete; A simple. Hind wing narrower, linear-lanceolate, Sc simple, short; R-stem simple, in middle of wing, forking over apex, with a branch from lower side at middle; Cu long, simple.

Larva and pupa essentially like Antispila. Mine relatively smaller, often almost entirely filled with frass, except the part cut out to form the ease, which is kept clean.

The various nominal species are described from specimens bred from different food plants, but the imagoes are nearly indistinguishable; very possibly several are merely slight food-varieties of a single species.

1. C. splendoriferella Clemens. Head bronzy; antennæ fuscous, tinged with golden; basal half, or rather more, of fore wings lead-gray, slightly metallic; from middle to tip golden, with a broad, nearly straight, triangular, silvery streak running from costa three-fourths way (near tip of broad part of wing) to center of wing, dark-margined on both sides; nearly joined by a dorsal streak opposite it, with converging dark margins, behind which is a dark brown blotch. In costo-apical fringe a silver streak edged on both sides with black. A black apical spot with bluish-silver center. Dorsal fringe brown with a black basal line interrupted by the silvery streak. Hind wing leaden gray, fringe yellowish brown. $4-4\frac{1}{2}$ mm. (Aspidisca Clemens; pruniella Clemens; Lyonetia saccatella Packard). Mine at first linear, then a small transparent blotch, the cocoon using almost

all of the clean part; on Cratægus, wild cherry, and apple.

The species is general. New York, Ithaca, East Greenbush, Albany, Fishkill.

Some of the following species may be valid, or all may be food-varieties of *splendoriferella*. Besides these species, larvæ of the genus have been found on several other food plants, but not distinctively named.

2. C. lucifluella Clemens. Larva on hickory. Moth slightly grayer than *splen-doriferella*, the golden area toward apex forming a broad band on costal third only; rest of ground of outer half solidly dark brown; the gray basal portion somewhat less extensive, and the markings generally not quite so far out.

Cocoons may be found on the trunk under loose bark.

3. C. ostryæfoliella Clemens. Larva on iron-wood (Ostrya); the mine larger than usual in the genus, and the case comparatively small.

The moth looks like a somewhat more richly colored *splendoriferella*, with somewhat more extended golden area.

4. C. saliciella Clemens. Larva on willow. Practically indentical with *splen-doriferella* but with the brown patch less sharply set off from the yellow; with two full rows of yellow scales between it and the black apical spot.

Missouri; Washington.

5. C. diospyriella Chambers. Larva on persimmon. Moth markedly variable, being either more or less yellow than *splendoriferella*.

The moth flies in June in Ohio.

6. C. ella Chambers, from hickory, is probably a strict synonym of *lucifluella*.

7. C. juglandiella Chambers. Markings of moth like those of *splendoriferella*, but larva feeding on black walnut; the case smaller; and the moth appearing earlier.

8. C. magnella Braun. Larva on Gaylussaeia in August, in the usual mine. Moth with the blackish area beyond the dorsal spot reduced, not extending beyond it toward costa, and sometimes practically confined to the fringe.

9. C. negligens Braun. Head, thorax and basal half of fore wings pale leadenmetallic. Antennæ blackish. Apical half of fore wing bright orange-yellow, more orange than any other species of the genus. Black and silvery apical markings essentially as in *C. splendoriferella*.

Larva mining cranberry in May; working in the overwintering leaves. Moth in June. One brood; apparently hibernating in the egg stage.

Buckeye Lake, Ohio.

Another rather distinct form occurs on white oak.

SUPERFAMILY **GELECHIOIDEA**

Vestiture usually scaly even on the head (figs. 147, 148) which never shows the high bristling vestiture of the normal Tineidæ; ocelli small or absent; antennæ normally with dorsal surface scaled, with two rows to a segment; the outer row much longer than the other and often encircling the antenna. Ventral surface bristled, the bristles very long in some Blastobasidæ and Œcophoridæ. Antennæ pectinate in some exotic forms (see Ptochoryctis). Palpi almost always upturned beyond middle of front, the third segment long and pointed, regularly tapering for most of its length (unlike the fusiform segment of most Tineoidea and Tortricoidea); second joint frequently with a tuft, which is usually less ragged than in the Tortricide; never bristled; third segment rarely tufted. Palpi rarely reduced (Pigritia group; fig. 179). Tongue usually moderate, scaled at base; maxillary palpi, when present, characteristic, minute, but of folded type, and curving over base of tongue; absent in forms with much reduced mouth parts. Epiphysis Palpi and tarsi never bristled or spined; hind tibiæ rarely large. bristled, but almost always hairy.

The legs are not displayed as in the Heliodinidæ, and habits of display are less common than in that family and the Glyphipterygidæ, but are shown, for instance, by *Stomopteryx agrimoniella*.

Wings highly variable in form and venation; but the fore wing always with an ample cell, frequently with a well-marked accessory cell imperfectly set off from it. \mathbf{R}_5 often running to the outer margin, but only in primitive forms with broad wings, \mathbf{R}_4 also terminating in the outer margin in a few exotic *Xylorictidæ*; **1st A** commonly lost (in the whole large family Gelechiidæ); **2d A** more or less forked at base. Hind wing variable, normally oval, but also trapezoidal, or even bifurcated in the Gelechiidæ, narrowing in other forms till it is linear in some Cosmopterygidæ; **1st A** varying as in the fore wing, and **3d A** also often vanishing with it; **2d A** forked as usual. Costa often sinuate and often bearing a tuft of bristles two-fifths way out. \mathbf{R}_1 frequently distinct, becoming free in a couple of the most reduced genera. Base of **M** obsolete in both wings. Female frenulum of few (normally two or three) bristles.

The egg is of the flat type and is laid externally: so far as I know, always singly. The larva (fig. 150) always has three setse on the prespiracular wart, and a single subventral on the meso- and metathorax, and the prothoracic spiracle normal. On the abdomen, setse iv and v are closely approximated, i and ii separated and usually at nearly the same level; on the ninth abdominal segment, ii is not much nearer the mid-dorsal line than the other subdorsals, and is usually nearer to i than to its mate. Normally the prolegs have their hooks arranged in a complete or broken ellipse; they are always uniserial but often triordinal. Secondary hair occurs in Blastodacna, a few subprimaries in Ethmia. The habits are various.

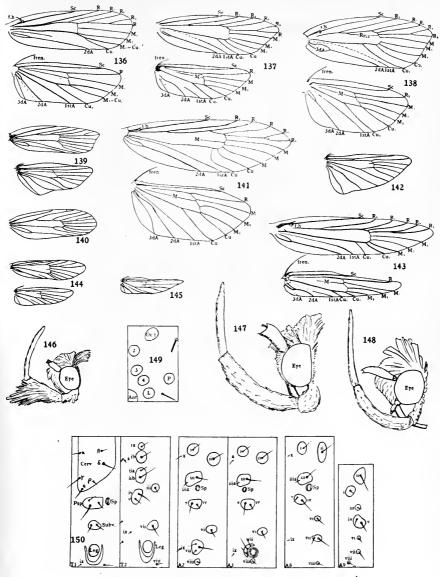
The pupa is also characteristic and is obtect; usually the abdomen is capable of dorso-ventral motion only, at three incisures. The body is depressed. The epicranial suture is present. The prothorax is hardly narrower on the mid-dorsal line than at the sides. The first four abdominal segments are usually longer than the others. The antennæ are usually adjacent for some distance on the midventral line.

The Gelechioidea form the most homogeneous of the subordinate groups of Microlepidoptera, and the largest of those groups. The Thyrididæ show some points of resemblance but will better be treated with the Pyralidoidea. Besides these only the genera Scythris and Euclemensia might perhaps be placed in this superfamily.

Family 19. **CCOPHORIDÆ**

(Depressariidæ; Gelechiidæ, in part)

Moderately small moths, rather larger than the average size of the Tineoidea. Head, typically, smoothly scaled, often somewhat rough. Labial palpi long, upturned, reaching or surpassing the vertex, the third joint nearly as long as the second, which is normally closely scaled, or with a longitudinal divided tuft. Tongue developed, scaled. Antennæ most often without pecten, never with eve-cap; both rows of scales on the segments broadly interrupted in Semioscopis, Ethmia, and a few other forms, as in the Maerolepidoptera. Hind tibia with long loose hair, sometimes rather bristly, the bristles gathering into tufts at the spurs. Tarsi nearly smooth, the spinules covered with scales. Fore wing with \mathbf{R}_{4} and \mathbf{R}_{5} stalked, rarely united; the other veins most often free; never with more than one vein lost. Cu_2 arising well out toward the end of the cell, which is normal. 1st A preserved toward margin, 2d A forked at base. 1st A very rarely appearing like a fork near tip of 2d A, as in many Cosmopterygidæ. Hind wing with **R** separate from **Sc** but closely parallel on the basal part of the cell, often connected to it by an oblique cross vein (\mathbf{R}_1) , which rarely is as strong as the other veins (Endrosis; Ethmia, in part). R and M, well separated at origin and parallel halfway to margin, then divergent, but rarely more than twice as far apart at margin as at origin; apex more or less rounded; the outer margin not distinctly concave below the tip of \mathbf{R} . \mathbf{M}_3 usually connate or stalked with Cu_1 , M_2 usually from below middle of end of cell, stalked with Cu in Triclonella, arising from cell nearer \mathbf{M}_1 , than \mathbf{M}_3 in Ethmia



FIGS. 136-150. ŒCOPHORIDÆ

136, Martyringa latipennis, venation; 137, Eumeyrickia trimaculella, venation; 138, Cryptolechia tentoriferella, venation; 139, Agonopteryx costosa (Europe), venation; 140, Depressaria heydenii (Europe), venation of fore wing; 141, Semioscopis aveilana (Europe), venation; 142, Ethmia pusiella (Europe), venation of hind wing; 143, Borkhausenia species, venation; 144. Dasycera imitatrix (Europe), venation; 145, Endrosis lacteella, venation of hind wing; 146, Eumeyrickia trimaculella, side view of head; 147, Cryptolechia tentoriferella, head; 148, Psilocorsis species, head; 149, Depressaria heracliana, arrangement of larval ocelli; 150, D. heracliana, seta map of larva

and Eumeyrickia. Hind wing variable in width, typically sublanceolate, with the anal region a little reduced, but more commonly ample, and fully as broad as fore wing; **2d A** more or less distinctly forked at base.

The family is close to the Blastobasidæ, but the pecten is never as heavy, there is no stigma at the termination of \mathbf{R}_1 , the veins do not show any distinct tendency to be grouped at the angles of the cell, and **Sc** is free from **R** in the hind wing.

Larva (fig. 150) typical of the superfamily: — head normal in form; all legs present and normal; normally with a biordinal ellipse

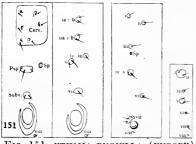


FIG. 151. ETHMIA PUSIELLA (EUROPE) SETA MAP.

of hooks, broken on the outer side; typically without secondary setæ; iii of abdomen in front of spiracle and slightly higher, single; head with adfrontals reaching membrane of vertex; front reaching two-thirds way to vertex; fourth occllus much closer to third than to lower, second much closer to third than to first. Larvæ more or less concealed feeders; usually web or rolled leaf; Endrosis a scavenger.

Pupa with epicranial suture pres-

ent, frontoclypeal imperfect or absent; maxillary palpi large, usually in contact with maxillæ, but labials and front femora concealed; antennæ in contact for some distance on the middle line, then normally diverging and exposing the hind tarsi. No hooked setæ on venter of ninth segment of abdomen.

The genus Ethmia (figs. 142, 151) is strongly aberrant in the early stages and slightly so in the imago, and is sometimes made the type of a distinct family, or transferred to the Yponomeutide. \mathbf{M}_2 arises from the middle of the end of the cell, much nearer to \mathbf{M}_1 than to \mathbf{M}_3 . \mathbf{R}_1 when distinct, is near the middle of cell or beyond (one-fifth way out normally) and the genitalia are distinctive. The larva has the hooks of the prolegs in a single band; its front is shorter, ending abruptly, and its adfrontals do not reach the vertex; sometimes there are additional hairs in group vii of the abdomen. The pupa has not been studied. The imago of Eumeyrickia shows essentially the same characters.

The family is large, and primitive for the superfamily, being much better developed in Australia than elsewhere. The Cryptolechia group appears primitive, Depressaria leads to the Gelechiidæ, which are very closely related, and Endrosis, to the Blastobasidæ. Most of the Cosmopterygidæ appear to be reduced Œcophoridæ.

LEPIDOPTERA OF NEW YORK AND NEIGHBORING STATES

Key to the genera

1. R_a and R₄ stalked with R₅, M₂ lost (fig. 136).....1. Martyringa. 1. R₃ free.

2. R₅ running distinctly to outer margin; all veins present (fig. 138).

3. Palpi with a triangular tuft on under side of second joint (fig. 146).

2. Eumeyrickia.

- 3. Palpi smooth-scaled, slender (fig. 147).
 - 4. Cu_1 and Cu_2 remote at origin.
 - 5. Hind wing with R and M1 divergent, palpi not reaching vertex, pecten
 - 5. Hind wing with \mathbf{R} and \mathbf{M}_1 parallel (fig. 138), palpi far beyond vertex, no pecten4. Cryptolechia.
- 2. \mathbf{R}_5 to costa or apex (figs. 139, 140).
 - 3. Abdomen strongly flattened, palpi with divided tuft on second joint, pecten present.

 - 4. Cu_1 and Cu_2 of fore wing separate (fig. 140).....7. Depressaria.
 - 3. Abdomen cylindrical; second joint of palpi without longitudinally divided tuft. The species with broad hind wings, which run here, lack the pecten.
 - 4. Hind wing with a median missing; M_2 stalked with Cu_1 (fig. 145). 17. Endrosis.
 - 4. Hind wing with all veins preserved.

 - 5. **M**₃ and **Cu**₁ of hind wing separate at origin. 6. Fore wing with a radial lost.....12. Decantha. 6. All veins present; costa concave (fig. 219, p. 356).
 - (Euclemensia: Heliodinidæ.) 5. \mathbf{M}_3 and \mathbf{Cu}_1 connate or stalked.

6. Fore wing with one radial missing (\mathbf{R}_4 and \mathbf{R}_5 completely fused).

- 6. All veins present; \mathbf{R}_4 and \mathbf{R}_5 stalked.
 - 7. Antennæ thickened with scales on shaft, toward base..14. Ecophora.
 - 7. Antennæ not specially thickened toward base; the segments about twice as long as wide.
 - 8. M₂ free (fig. 143).
 - 9. Antenna with pecten.....16. Borkhausenia. 9. No pecten.

10. Hind wing obtuse, as wide as fore wing.

10. Hind wing lanceolate, three-fourths as wide as fore wing.

- 11. Schiffermuelleria.
- 11. Hind wing with M_2 nearest M_1 (fig. 142).....10. Ethmia.
- 11. Hind wing with M_2 rather nearer M_3 , rarely central (fig. 141).
 - 12. Fore wing with Cu_1 and Cu_2 approximate or stalked; hind wing ample.....9. Semioscopis.

12. Fore wing with $\hat{C}u_1$ and Cu_2 widely separate at base.

8. Inga.

13. Fabiola.

1. MARTYRINGA Busek

(*Egoconia* Walsingham)

Antenna heavy, flattened, strongly serrate; no pecten; no ocelli, palpus upturned beyond vertex, second segment a little thickened, third about as long as second. Fore wing (fig. 136) more than three times as long as wide, with rounded outer margin; \mathbf{M}_1 arising well below angle of cell, \mathbf{M}_2 lost, \mathbf{M}_3 approximate to \mathbf{Cu}_1 and \mathbf{Cu}_2 , which are stalked; 1st A apparently lost; \mathbf{R}_5 running to apex or costa. Hind wing ample, \mathbf{M}_3 lost, \mathbf{M}_2 connate with \mathbf{Cu}_1 . \mathbf{M}_1 from a third way down end of cell.

A curious form, looking like a Depressaria, but with some Gelechiid characters. Early stages unknown.

1. M. latipennis Walsingham. Clay color, dusted with blackish, and with slightly blurred blackish marks; middle half of costa dark-shaded; base also more broadly shaded with dark. Orbicular and claviform spots oval, black, rarely fusing, reniform the larger, and squarish, the region beyond it darker almost to the margin, defining a pale, irregularly sinuous postmedial line. Terminal line gray, narrow. Hind wing paler and grayer. 15 mm.

July to August.

Known from southern Connecticut to North Carolina and Missouri; New York: Ithaca.

2. EUMEYRICKIA Busek

Palpi with triangularly tufted second joint and much longer, very slender third joint (fig. 146); antennæ slender, normal, without pecten; no ocelli. Fore wing (fig. 137) more triangular than usual, with subfalcate apex and arched costa. \mathbf{R}_4 and \mathbf{R}_6 forking over apex, the other veins separate, \mathbf{Cu}_2 straight. Hind wing ample, with veins \mathbf{R} to \mathbf{M}_3 nearly evenly spaced, and \mathbf{M}_3 almost connate with \mathbf{Cu}_1 .

Busck suggests that this genus is related to Ethmia. The moth favors hollow trees, and struts about with wings elevated like some Glyphipterygidæ.

1. E. trimaculella Fitch. Third joint of palpus blackish with two longitudinal white lines, not quite reaching base; second joint white-tipped and with white patches on inner side. Fore wing fuscous, dusted heavily on a luteous ground, leaving two pale costal spots toward apex and other faint markings; fringe dark with a double series of pale bars. Hind wings dark. 13 mm.

The moth flies in June southward, and in early July in the North.

Parry Sound, Ontario, and Megantic, Quebec, to North Carolina and Ohio. New York: Upper Ausable Lake.

Eido albapalpella Chambers. (Venilia Chambers, not Duponchel) has never been identified and may be the same species as the preceding; but as described the palpus is paler, the third joint is white, except for the blackish base, and the second white on its inner side.

Kentucky.

3. GERDANA Busek

Palpi somewhat loosely held, upturned nearly to vertex, with slightly rough and thickened second joint, and somewhat shorter third; pecten present; abdomen slightly flattened. Fore wing elongate, ovate, blunt; veins M_2 to Cu_1 approximate at lower angle of cell; R_s reaching margin just below apex; Cu_2 remote. Hind wing with costa sinuate, R and M₁ distant but divergent. M_s and Cu_1 connate or stalked. The habitus of this genus is rather like that of Depressaria.

1. G. caritella Busck. Straw to ochre yellow, suffused with darker brownish yellow; palpi brown; face, head, and thorax lighter; fore wing with basal half of

costal edge somewhat dusted with fuscous; two obliquely placed dark dots at one-third, and a series of three at end of cell, the discal largest; sometimes the three fused into an oblique band. A strongly bent subterminal band parallel to tip of costa and outer margin. Apex infuscated. Markings tending to disappear in rubbed specimens. Hind wing whitish, abdomen and fringes straw. 13 mm.

The moth occurs in July and August.

Southern Massachusetts to Ohio, Maryland, and Texas. New York: Ithaca.

4. CRYPTOLECHIA Zeller

(In part; Machimia Clemens)

Similar to Gerdana; palpi much longer, second joint reaching vertex; wings in our species more ample. This genus also occurs in Africa and Australia (figs. 138, 147).

1. C. tentoriferella Clemens. Light ochreous; when fresh, more or less shaded with reddish and dusted with black; two contrasting discal dots, and a blackish spot in fold halfway between them; a broken punctiform postmedial line parallel to outer margin and a series of black terminal dots. Head pale, palpi nearly white, with basal half contrasting blackish. Hind wing lightly infuscated. 22 mm. (fernaldella Chambers).

General in distribution; common in late August to October on trunks of trees. Caterpillar in August on oak, cherry, maple (?), chestnut, etc., in a flat tent on under side of leaf at one side of midrib, bending the leaf; feeding outside the tent. Caterpillar dark green. Cocoon under a folded edge of the leaf; of dense silk except at the point where the moth emerges. New York: Potsdam, Ithaca, Big Indian Valley, Schenectady, Karner, Rhinebeck,

New Windsor, New York City; Moravian Cemetery, Staten Island.

5. *PSILOCORSIS* Clemens

(Cryptolechia, in part)

No pecten. Palpus very long and slender; third joint nearly as long as second (fig. 148). Fore wing with squarish apex and more vertical outer margin than in Cryptolechia; \mathbf{R}_s rarely running to the apex, as in some Depressarias, from which the genus is distinguished by the lack of pecten, and by its smoother palpi. Ground color luteous to light brown. Second joint of palpus with a black ventral line edged on both sides with white; third joint white with three black lines, the middle one continuous with the line on the second joint. Scape of antennæ with two black lines with a white one between them; shaft on upper surface normally with two longitudinal black lines toward base, with white between them, becoming transversely barred with brown outwardly; the under side evenly pale. Fore wing transversely strigose with darker brown or black, usually with a black terminal line and sometimes with a dark fringe, but without other sharply defined markings.

All the species are very similar and are practically impossible to determine without a knowledge of the larva.

The larva lives in a slight web between two leaves of the food plant, skeletonizing the leaves, which turn brown. It has rather distinct tubereles; on the eighth segment of the abdomen, iii is directly in front of the spiracle, and iv and v, below

it. Pupation takes place in the web. 1. P. quercicella Clemens. Luteous, strigose with brown. Terminal line black, broken, contrasting, variable in length; fringe lead gray. A dark shade across

wings beyond middle, or a patch in fold, which is rarely obseure. Fore tibia solid brown, or with a complete, broad, longitudinal stripe. 12-15 mm.

Larva ou oak and chestnut in September; pale yellowish or greenish; head and entire thorax black. Moth in March and April; and in July. Generally distributed. New York: New Windsor (Morton). 2. P. obsoletella Zeller. Closely similar to P. quercicella, but usually, slightly

larger (16 mm.), and more evenly strigose; usually without the postmedial shade across the wing. Larva also on oak and chestnut, but with only prothorax black; mesothorax with some red on sides, and metathorax pale, like the abdomen.

Ohio. New York: Ithaea.

P. dubitatella Zeller appears to be a synonym of P. obsoletella, but without knowledge of the larva this must be a little uncertain.

3. P. faginella Chambers. Mouse-gray, with the strige less contrasting, deep brown; the terminal line reduced to about four dots at apex, inconspicuous. Longitudinal striping of antennæ unusually long, obscure; white lines on palpi very slender, broken. Fore tibia with a brown patch on anterior side. 13 mm. (Hagno Chambers).

Larva on beech; whitish with ferruginous head, some ferruginous on prothorax and a pinkish patch on each side of mesothorax.

4. P. cryptolechiella Chambers. Very similar to P. faginella, rather evenly colored, with slender, inconspicuous, broken terminal line and light fringe. Head, thorax, and base of fore wing reddish orange. 15 mm.

This species is unknown to me. It was bred from holly. In the United States National Museum it is identified with a species from Vaccinium, known from Massachusetts to North Carolina.

5. P. reflexella Clemens. Fore wings dull straw color, strigose with fuscous, appearing dull fuseous brown. Terminal dots obscure; fore tibia slightly darkened. 20 mm. (quercicella Zeller, not Clemens; cressonella Chambers).

Larva on oak. Moth in June.

Massachusetts to Virginia. New York: Ithaca (determined by Chambers).

6. P. ferruginosa Zeller. Ground color and fringe, light ochreous, markings as in P. reflexella, with distinct black discal dot. Fore tibia light. 18 mm. (confertella Walker).

Maine to Ohio. New York: Otto.

7. P. fletcherella Gibson. Antennæ black, annulate with light ochreous; fore wing pale golden vellow, rather heavily dusted with pale brown, with purplish reflections; discal dot black, conspicuous; fringe ochreous, shaded with brown. Legs bright pale ochreous; fore leg sometimes dark; tarsal joints dark. 19 mm. Larva on aspen poplar; yellowish green, head jet black, cervical shield blackish,

brown in middle, tubereles inconspicuous, a few blotches of crimson on eighth segment of abdomen, above spiracles; anal plate black.

Ottawa, Ontario.

Members of the genus have also been bred from Ambrosia, Amelanchier, and Carpinus, but not named.

6. AGONOPTERYX Hübner

(Depressaria, in part; Agnopteryx)

Similar to Depressaria, but with Cu_1 and Cu_2 stalked in the fore wing (fig. 139); hind wing less strongly lobed, as a rule.

The imagoes of this genus and of Depressaria emerge in the fall, often as early as August, and overwinter. They are often found under bark or flying by day in the early spring.

LEPIDOPTERA OF NEW YORK AND NEIGHBORING STATES

Key to the species

Ι.	Thorax	and	extreme	base	of	fore	wing	black,	contrasting	 atrodorsella	ı.
۱.	Base of	fore	e wing no	t bla	ck.		0		U		

2. Ground of fore wing pure white.

3. Ground heavily dusted with black, appearing ash gray......20. allenella. 2. Ground of fore wing not pure white.

- 3. Discal dot \mathbf{d} with white or whitish scales in center much paler than
 - ground.17
 - 4. Discal dot b wholly or part white, or replaced by a larger irregular white dot; a and c often white also.

 - 6. No trace of a pale or dark postmedial fascia; white scaling of discal
 - dots a and b often fused into a patch. 7. Black dots a and b also fused into a crescentic mark; discal dot d
 - even; third segment of palpus smooth, black-tipped.

6. walsinghamella.

6. A faint angulate postmedial line; discal dots all minute.

5. A contrasting curved black line on disc.

- 6. Ground mouse-gray14. hyperella. 6. Ground pale gray, tinted with red in fresh specimens.. 13. curvilineella.
- 5. No such mark; fore wing often with a vague squarish dark spot on disc. 6. Ground reddish, discal dot d often with a single white scale or a
 - little transverse streak.....12. fulva.
 - 6. Ground clay-color to ochre, discal dot d with half a dozen white scales in a rounded group. 7. A black patch in disc before discal dot **d**, usually obliterating **c**.

8. Patch larger; ground mottled on a bright ochre under-color.

3. pulvipennella.

- 8. Patch smaller; ground light, even clay-color; hind wing, below, very pale.....4. argillacea.
- 7. No black patch in disc; with raised scaling. 8. Palpi strongly tufted on outer half of second joint; hind wing, below, dusted with blackish.....8. plummerella.
- 6. Ground dull fuscous gray, with contrasting ochreous base.

11. nigrinotella.

- 3. Discal dot \mathbf{d} without any white scales, or any scales noticeably paler than the ground color.
 - 4. With a continuous dark terminal line toward apex, and a curved black spot on disc.....16. lythrella.
 - 4. A series of terminal dots. 5. Fore wing straw-yellow......17. flavicomella.

¹⁷ In this genus there are typically four dots in the cell. The first two are obliquely placed and represent the orbicular, the outer two longitudinal, representing the reniform. I have indicated these from the base outward with the letters a, b, c, d. Some authors speak of b as the inner and d as the outer diseal dot.

5. Fore wing bright yellow, mottled with red.

6. A triangular darker shade at end of disc.....18. robiniella. 6. No such shade.....19. lecontella. 5. Fore wing dull red-brown, practically immaculate.....12. fulva.

- 5. Fore wing dull yellowish or grayish. 6. Thorax pale; fore wing mottled and scaled with white, base contrastingly paler; fuscons patch in cell faint; fresh specimens with
 - slight pink suffusion 15. senecionella. 6. Brownish ochreous; thorax darker, with a little black behind.

9½. pteleæ.

- 6. Anterior half of disc of thorax gray; wings evenly clay-color, the base hardly paler; patch in cell contrasting, small, nearly black,
- 6. Wings powdery pale luteous, but without white; with fuscous, and raised black scales.....9. scabella and 8. plummerella (see above).

1. A. atrodorsella Clemens. Cream color; most of thorax and costal half of base of fore wing black; six or eight black costal dots; a rosy shade over end of cell, running into streaks on the veins; a dark costal-apical patch; black terminal dots, and rosy fringe. Upper part of face blackish, vertex rufous. 20-23 mm.

September to April. Caterpillar green, with darker dorsal and subdorsal stripes. blackish tubercles and spiracles; yellow-brown head, and a light cervical shield with a black dot at each side; feeding on Bidens, folding the leaf lengthwise. New Hampshire to District of Columbia, west to Wisconsin. New York: Fen-tons (Lewis County), Crosby (Lake Keuka), Ithaca, Delmar, Schenectady, New

Windsor, Staten Island.

2. A. canella Busck. White; palpi mottled with light brown, vertex brown; fore wings lightly dusted with black and brown scales except toward the base. Middle of costa with a black and brown shade. Discal dots a and b usually distinct, black or fused into a bar. Hind wings light fuscous with pale fringes. 20 mm.

July to September. The related European species, D. alstræmeriana, feeds on Conium.

Southern New Hampshire; Connecticut; New York; Washington State. New York: Wilmington, Ithaca, Catskills.

3. A. pulvipennella Clemens. Clay color, more or less shaded with reddish, and heavily dusted and shaded with black-brown, the whole effect powdery wood-color; base paler; costa with short black bars. Discal dots slightly raised in perfect specimens; a and b black, d with a white center, c obliterated, claviform indicated by a black dot; fringe reddish. Face and inner side of palpi whitish; vertex rose-brown, 22 mm. (*eupatoriella* Chambers, *solidaginis* Walsingham).

September to May; July; the summer brood probably dropping out northward. Very common. Caterpillar green, sometimes with darker dorsal and lateral lines. Tubercles, head, and cervical shield nearly concolorous; head marked with brown; cervical shield with lateral brown dots, anal plate brown-edged. Caterpillar folding the leaves of Eupatorium and Solidago lengthwise.

There is a rare coal-black aberration.

Massachusetts to Virginia, Illinois, and Missouri. New York: Fentons (Lewis

County), Ilion, Crosby (Lake Keuka), Ithaca, Schenectady. 4. A. argillacea Walsingham. Pale gray, hardly yellowish, *lightly* sprinkled with black; a small blackish patch near end of cell; base of inner margin pale, followed by a blackish shade. Terminal dots weak, outer discal dots inconspicuous, the white centers dull and not contrasting. 20 mm.

This species is the American representative of A. yeatiana, which eats Umbelliferae. It has been taken in March and in September.

Connecticut; New Hampshire; central Missouri. New York: Ithaca.

5. A. applana Fabricius, race clemensella Chambers. Rose-brown, dusted and mottled with fuscous and sometimes luteous. Head, thorax, and base of wings more yellowish; the four discal dots white, black-ringed, the first with most black. Postmedial line dark, parallel to the outer margin, angulated below costa, obscure, and very easily overlooked. Costal edge heavily spotted with black and luteous white, almost completely crowding out the ground, even at the base of the wing. 18 mm.

August to April. Caterpillar on parsnip and wild parsnip (Heracleum). The European larva is green, with darker dorsal and subdorsal lines, black tubercles, and grav-green head.

Connecticut to District of Columbia. New York: Ithaca, New Windsor.

A. ciliella Stainton, a slightly larger and smoother-looking species with yellowheaded larva is to be expected, but not definitely recorded in the northeastern States.

6. A. walsinghamella Busck. Crimson red, brighter than A. applana and A. ciliella; middle half of wing toward costa sprinkled with whitish and black, leaving the base of the costa red, contrasting with the cream white base of the inner margin; discal dots a, b, and c, with their white portions usually fusing into a spot, their black portions distinct. 20 mm. (Depressaria fernaldella Walsingham, not Chambers, hilarella of authors, not Zeller).

Caterpillar on Myrica.

Orono, Maine, to Connecticut and Wisconsin. New York: Albany.

7. A. nebulosa Zeller. Powdery fuscous, a little yellower than A. argillacea, with inconspicuous raised black points, as usual in the group. The four ordinary discal dots distinct, their white scaling tending to be suffused and to unite them in pairs, (a and b being generally united) but dull and hardly paler than the ground color. Under side of hind wing striolate toward tip. Second segment of palpus with funnel-shaped tuft toward tip. 18 mm.

Apparently general in distribution. 8. A. plummerella Busck. Closely similar to A. nebulosa. Discal dots very inconspicuous, apparently separate; base of hind wing a little more whitish. 24 mm.

Plummer's Island, Maryland; Ohio. New York: Rock City (Cattaraugus County).

9. A. scabella Zeller. Second joint of palpus nearly smooth. Markings as in the last two species, the raised tufts, especially the one in the fold below discal dot a, more distinct; no well-marked white scaling. Under side of hind wing with scattered black dots toward apex, and a fine dark terminal line.

Ohio.

9½. A. pteleze Barnes and Busck. Brush on second segment of palpus widest at middle, tapering to both ends. Fore wing rough with raised scaling. Thorax dark ochreous mottled with brown and black scales, the extreme posterior tip black; head redder. Fore wing ochreous with a large nearly round blackish shade over end of cell. None of the discal dots scaled with white. A series of ill-defined dark spots along costa, and obscure terminal dots. Hind wing light ochreous fuscous, the base and inner margin pale. Under side of abdomen with two series of black dots. 20-22 mm.

Larva on hop tree.

Decatur, Illinois.

- Clay-color. Palpus with two moderate dark rings on third 10. A. segment. Anterior half of disc of thorax powdery gray. Base of fore wing paler, defined with a little blackish shading toward inner margin. Base of costal edge black. Most of wing with scattered black scales, the costa more distinctly barred. All discal dots black, a small black patch on disc. Under side of fore wing with scattered light fuscous fleeks, and a broken black terminal line. 19 mm.

The only specimen before me was taken in April.

New York: Ithaea.

11. A. nigrinotella Busek. Palpi light yellow-brown, the base black-shaded and extreme tip black. Head light red-brown, thorax yellow-brown, — the front, back, and tegulæ darker. Base and basal half of costa yellow-brown, the rest of the wing brownish fuscous, with scattered black scales. Discal dot \mathbf{d} only, white-scaled. Hind wing shining light yellowish fuscous. 22 mm.

Larva on hop tree, with A. pteleæ.

Ohio; Illinois.

12. A. fulva Walsingham. Dull red-brown without yellow tint, with the usual paler base followed by a dark shade. Costa interrupted with black and yellowish. 22 mm.

This form is easily distinguished from the other red species by the lack of white diseal dots. The locality was not reported, and the species is very probably western.

13. A. curvilineella Beutenmüller. Head white, the fore wing grayish white, shaded and mottled with pale pearl gray, with a more or less distinct pink tint, leaving the base and costa pale, the wing lightly dusted with black scales, which gather into larger black spots along the costa; first three discal dots fused into a black crescent, concave upward, the fourth one white, ringed with black. Hind wing also pale, with white fringe. 18 mm.

October to April.

New York to Missouri. New York: Ithaca, New York City.

14. A. hyperella Ely. Similar to A. curvilineella; most of the wing-surface nearly evenly mouse-gray. Vertex dark gray, unlike even the darkest specimens of A. curvilineella. 16 mm.

End of May. Larva on *Hypericum prolificum* in April. Possibly a seasonal form of the preceding species, which however appears to be single-brooded in the north.

Great Falls, Virginia.

15. A. senecionella Busck. Luteous, slightly shaded with pink, and strongly and irregularly with dull gray, leaving the base pale and sharply defined except at costa. Paler areas also over the discal dots **a** and **b** and beyond **d**, and **a** vague, paler postmedial line. Third discal dot not distinct. Ground sometimes more evenly gray. Hind wing pale gray, the veins darkest. 17 mm.

Larva in March; imago in May.

Plummer's Island, Maryland.

A. canadensis Busck, which is slightly more dusted with black, and has the base of the hind wing paler, is likely to occur in the west of our area.

16. A. lythrella Walsingham. Tawny reddish, heavily shaded with mouse gray and dusted with black and pale cinercous toward costa; a distinct dark spot in the dorsal part of the whitish base, normally nearly covering the pale area; a thick, black, oblique crescent, edged with the reddish ground and followed by a group of whitish-cinercous scales. Discal dot **d** represented by a vertical white bar. Terminal line continuous, fading out at apex. Fringe purplish gray. Hind wings mouse gray. 16 mm.

Caterpillar on Lythrum alatum. Dull whitish green, immaculate; webbing the leaves in June. Moth in July.

Illinois; North Carolina.

17. A. flavicomella Engel. Straw yellow, shaded with red-brown and flecked and obscurely striate with blackish or umber brown. Outer discal dot black, in a brown spot or shade; inner (b) a black point, or obscure; more or less distinct traces of a postmedial fascia. Black bars on costa and terminal dots. Hind wing whitish. Blackish shade near base and shade over the discal dots typically strongly contrasting. 16 mm.

June and July.

Connecticut to North Carolina; Illinois. New York: Ithaca.

18. A. robiniella Packard. Yellow, heavily shaded and dusted with dull erimson red and a little fuscous, the latter forming a series of terminal dots. A vague triangular reddish shade over middle of wing, with a circular pale area before it containing discal dot a, and with the small pale discal dot d on its outer edge. A darker subterminal shade. Hind wing fuscous. Head and thorax concolorous Caterpillar on Robinia in June. Moth in July. General in distribution. New York: Schenectady, Bridgetown, Rhinebeck,

Crugers, Katonah. 19. A. lecontella Clemens. Head dull ochreous, with some fuscous on second, and two dark rings on third joint of palpus. Antennæ fuscous; thorax dull ochreous with two black-brown dots in front. Fore wings slightly darker, nearly evenly dotted with dark, and somewhat washed with rufous; a slight dark shade only on disc. Discal dots a and d distinct. Hind wing gray. 22 mm.

Caterpillar possibly on Sanieula and Pimpinella. Vermont to Maryland and Ohio. New York: Ithaca, Schenectady, New Windsor; Sea Cliff, Long Island.

20. A. allenella Walsingham. Fore wings broader with more arched costa, with Cu_1 and Cu_2 sometimes only very closely approximate. Abdomen only slightly flattened. Pecten weak, palpi practically smooth. Fore wing with rough scales and a discal tuft. Pale gray; palpi powdery; fore wing minutely mottled and dusted with blackish, with somewhat more distinct black points on costa, especially toward base; a fuscous subterminal shade, more distinct on costa, and a slight streak in base of fold. Two black discal dots at end of cell. Hind wing of the paler gray ground-color. 18 mm. (Semioscopis).

June. Larva on oak.

Maine to Virginia and western Pennsylvania. New York: McLean, Rhinebeck.

7. DEPRESSARIA Haworth

(In part; Schistodepressaria Spuler)

Head somewhat rough-scaled, with small ocelli and maxillary palpi; labial palpi with a furrowed brush on under side of second joint, third joint long and smooth; antennæ with pecten. Abdomen markedly flattened, and wings folded flat over the back at rest. Fore wing more or less oblong (fig. 140) with blunt or rounded apex, \mathbf{R}_s running to costa or apex, \mathbf{Cu}_1 strongly curved at base, \mathbf{Cu}_2 well separated from it, often nearly straight. Hind wing broader, usually strongly lobed at anal angle, with all veins present; \mathbf{R} and \mathbf{M}_1 parallel, \mathbf{M}_3 and \mathbf{Cu}_1 connate or shortstalked, M_2 arising near M_3 .

The fore wings tend much more to longitudinal marking than in Agonopteryx. The discal dots a and b usually fuse into a streak, and there is almost always a blackish bar along the base of the inner margin, in place of the pale base of most Agonopteryxes. The moths emerge in July and go immediately into winter quarters, coming out and laying their eggs the following March or April. The caterpillars (figs. 149, 150) live in webs on various plants, especially Umbelliferæ. The generic name Depressaria was formerly used to include a great variety of Ecophoridæ and even Gelechiidæ, mostly species with similar tufted palpi.

WILLIAM T. M. FORBES

Key to the species

1. Base and costal edge whitish...5. cinercocostella. 1. Base and costal edge not whitish. 2. With a pale fourth discal dot. 3. Fourth discal dot white, conspicuous, and preceded by a long white line. 7. emeritella.

3. Fourth discal dot less conspicuous, preceded by a heavy black bar.

3. maculatella, 4. symmochlota, 3. Fourth discal dot less conspicuous and not preceded by a longitudinal bar.

4. Hind wing whitish, much paler than fore wing.....(1.) togata.

4. Hind wing light gray, paler at base.....l. betulella. 2. No pale fourth discal dot.

1. D. betulella Busck. Fuscous, obscurely mottled, with the pale base of inner margin most conspicuously contrasting; terminal dots present; black discal dots obscure; head yellowish. 23 mm.

Caterpillar in a silken tube between leaves of black birch; pale green with black tubercles. Moth emerging in June.

Vermont to Pennsylvania.

D. togata Walsingham, described from the Rocky Mountain district, is closely similar but with rather darker fore wings and paler hind wings. It may enter the northern part of our territory northward, and has been reported from Vermont, though apparently in error for the preceding species.

2. D. grotella Robinson. Dull straw color, second joint of palpi blackish below; fore wing nearly evenly colored, a little redder on the disc, with vague longitudinal streaks of fuscous, the one in the outer part of the cell hardly more contrasting than the rest, but present, the most distinct streaks being a postmedial series between the veins. Fourth discal dot dark, terminal dots distinct; hind wing pearl gray. 20 mm.

Caterpillar green, darkest dorsally; cervical shield green, uumarked; head green with a black dot on each side above the jaws; on Corylus.

This species is unknown to me.

North Atlantic States. New York (Robinson).

3. D. maculatella Busck. Similar, crisply black-dusted on a chalk-white ground, appearing light gray. Fore wing with fuscous streaks between the veins, more or less interrupted by a faint, pale, dentate postmedial line; two pale discal dots, with a heavy black bar in the cell between them, touching the first. 22 mm.

New Hampshire and Connecticut to Ontario and western Pennsylvania.

4. D. symmochlota Meyrick. Similar to D. maculatella, but ground mouse gray or fuscous, with some pale shading and scaling, especially indicating the post-medial line. Sparse black scaling on the dark portions. Bar in cell as in D. maculatella, conspicuous. 20 mm.

Late July to early August. Perhaps the same as D. grotella. Sebec Lake, Maine; Parry Sound, Ontario; Manitoba.

5. D. cinereocostella Clemens. Fore wing reddish-brown, marked with numerous short black dashes; costa, head, and thorax normally much paler, whitish gray; hind wing grayish fuscous. 18 mm. Occasionally the costa is practically concolorous with the rest of the wing, but

such specimens are recognizable by their pinkish tint, distinct pale postmedial line of whitish scaling, minute or wanting fourth discal dot, and absence of any blackish shading in the cell.

Fresh moths occur in July The caterpillar is bright sulphur yellow with a

more ochreous cervical shield, and a dark ochre head, with black dots at the eyes. The tubercles and hairs are black. It folds and ties together leaves of water-parsnip (Sium) and caraway.

Massachusetts to Virginia. "New York" (Busck). 6. D. heracliana DeGeer. Luteous shaded with brown, forming longitudinal stripes or shades. Longitudinal stripes in cell, fold, and near base of inner margin; the stripes in the outer part of the wing lying between the veins, and interrupted by the angulate pale postmedial line; a dark discal dot. Palpus with two dark rings on third joint. 25 mm. (umbellana Fabricius, pastinacella Duponchel, ontariella Bethune.)

Caterpillar webbing together the flower heads and seeds, more rarely also the leaves of wild parsnip (Heracleum) and other Umbelliferæ, in June and July; teases of white parsmip (meracleum) and other Umbelliferæ, in June and July; usually entering the hollow stems to pupate. Green-gray, paler and yellower below, with head, plates, true legs, and tubercles black. Moth emerging in July and immediately retiring till the following spring. Of general distribution and decidedly common. New York: Crosby (Yates County), Ithaca, Albany, Rhine-beck, New Windsor, West Farms.

7. D. emeritella Stainton, race alienella Busck. Fore wing red-brown, dusted with white, with shaded, angulate, pale postmedial line, and white discal dot. Hind wing with red fringe.

The typical species occurs in Europe.

Caterpillar on Artemisia and presumably other aromatic Compositæ. Oregon. New York: Ilion.

8. INGA Busek

(Cryptolechia, Anesychia, in part)

Closely similar to Depressaria; no pecten; abdomen cylindrical. Fore wing with Cu₁ and Cu₂ subparallel and strongly curved at base. No ocelli or maxillary palpi; labials long, upturned, and smooth. The black and white coloring is shared by Depressaria canella. 1. I. sparsiciliella Clemens. White; a black dot at base of costa and bar near

base of inner margin, a squarish black spot near middle of costa, two black discal dots; black subterminal dots tending to run together. Hind wings and under side dark gray, head white with some black. 16 mm. (contrariella Walker, atropicta Zeller).

Southward this species flies in May and June.

New York to Texas. New York: West Farms.

9. SEMIOSCOPIS Hübner

(Enicostoma, in part; Epigraphia Stephens)

With ocelli. No distinct maxillary palpi. Labial palpi upturned in front of face, the second joint with indications of a bifurcated tuft toward the tip, the third shorter than in Depressaria and slender; tongue somewhat weak; no pecten. Fore wing with marked apex and oblique outer margin, Cu, more strongly curved than Cu. (The two are stalked in *S. inornata* [subgenus Epigraphia]). Of our species *S. megamicrella* also may be an Epigraphia, but I have no full notes concerning it.

The caterpillars are found on deciduous trees in the fall; the moths flying in the spring from April to June. Agonopteryx allenella, with but slightly flattened abdomen, forms the transition to this genus, but may be distinguished by its pecten and blunter wings.

Key to the species

1. Black longitudinal mark reaching base of wing.

2. Line heavy, the basal part strongly curved, and running from extreme base

2. Line more slender, running straight out in fold......4. aurorella. 1. Black mark, if present, not reaching base.

2. No definite black marking.

3. With conspicuous black terminal dots, and dots on disc.

2. megamicrella. 3. Evenly powdery gray; terminal dots absent, and dots on dise diffuse.

1. inornata.

1. S. inornata Walsingham. Powdery ash gray; outer half of fringe white with gray-stained tips; usual marks indicated by a slight irregularity of the dusting. Hind wing of the lighter ground color, the base of the fringe distinctly powdered. This species is easily confused with the next, but is apparently the only one

which normally has Cu₁ and Cu₂ stalked.

Ottawa, Ontario; Pennsylvania; Manitoba. New York: Ithaca, Karner.

2. S. megamicrella Dyar. Whitish, powdered with light brown; a double antemedial black dot on cell and a distinct black lunated discal bar, in some specimens broken in the middle; a rather distinct series of dark subterminal dashes running parallel to outer margin. Hind wing nearly white, with a faint continuous pale gray line in base of fringe. Cu_1 and Cu_2 of fore wing may be stalked. 18-25 mm.

April and May.

Massachusetts to Ohio. New York: Ithaca; East New York, Long Island. 3. S. merriccella Dyar. Closely similar to *S. packardella*, but rather more dis-tinctly striate, the black dash not reaching the base of the wing. 25-30 mm. (H 48:38.)

May.

New Hampshire to western Pennsylvania. New York: Big Indian Valley, Albany.

4. S. aurorella Dyar. Light gray, dusted somewhat irregularly with dull brown. Black basal dash fine, nearly straight, starting at base of costa, but running almost wholly in fold; its outer end attached to a curved line running up into cell, and often connected with the crescentic discal bar. Obseure dark dashes in the subterminal region, and a postmedial dash in fold. Terminal dots strong. Hind wing pale gray. 25-30 mm.

New York to Manitoba. New York: Ithaca, Big Indian Valley.

5. S. packardella Clemens. Gray, slightly streaked and dusted with brown. A heavy black bar running from base of costa out along base of \mathbf{R} , then eurying down into fold, and then up across cell to middle of costa. A slender branch running toward base of wing in fold, and a thicker one from where it crosses R to lower angle of cell; the latter bar convex outward. Hind wing pale gray. 25 mm.

June.

Northern Atlantic States. New York: Ithaca, Big Indian Valley, Albany.

10. ETHMIA Hübner

(Psecadia Hübner; Anesychia)

Moth similar to Semioscopis. Sensory area on antennæ more extensive, covering half the antennæ, even at the base. Palpus without any tufts on second joint. Fore wing (fig. 142) narrower, with Cu_1 and Cu_2 widely. separated at origin and nearly straight. Hind wing with **R** often preserved, M_2 rather nearer

LEPIDOPTERA OF NEW YORK AND NEIGHBORING STATES

to \mathbf{M}_1 than to \mathbf{M}_3 and nearly straight. Caterpillar with a single band of biordinal to M_1 that to M_2 and hearly straight. Catchphar with a single blind of blordmar hooks on prolegs; sometimes with additional setw in group vii of abdomen; otherwise normal; alpha of prothorax nearer mid-dorsal line than beta. Pupa obtect, with 5–6 and 6–7 capable of dorso-ventral motion; flattened and rounded, without cremaster. Maxillary palpi large; labials minute; epicranial suture present, but fronto-clypeal absent; tongue not nearly as long as fore legs, middle legs separated on midventral line by fore legs; hind legs concealed by the antennæ, which lie in government. which lie in contact for their posterior third. Wing cases hardly extending beyond fourth segment of abdomen. End of body unarmed, rounded over. Anal prolegs preserved, provided with hooks, and taking the place of a cremaster; located on each side of the genital opening; apparently corresponding to the anterior group of hooks in Stenoma.

The peculiar larva of this genus (fig. 151), which is most closely matched in the Choreutis group, has caused some workers to place it in a special family, Ethmiidæ, of the Yponomeutoidea. The moth and pupa, however, are very close to the more primitive Œcophoridæ. The typical group feed on Boraginaceæ, enclosed in a light web.

Key to the species

1. Basal segment of abdomen blackish; the rest yellow, contrasting.

1. fuscipedella.

1. Abdomen all gray.

2. Middle of wing broadly black from base to apex.

3. Expanse less than 20 mm., fore wing not streaked between veins. 2. trifurcella

3. Expanse 25 mm. or over; fore wing gray-streaked between veins.

3. macelhosiclla.

2. No dark longitudinal stripe.

3. One short black dash in cell, and dot at lower angle....4. longimaculella. 3. Two dashes in cell, sometimes connected into a slender streak, and no

1. E. fuscipedella Walsingham. Mouse gray; head, thorax. except tegulæ, and base of abdomen, concolorous gray; the rest of abdomen and hind tibix, yellow, immaculate. Two black dots on thorax. A black dot in middle of cell, one or two at end, and antemedial dot in fold. Usually with strong black terminal dots. Hind wing concolorous, immaculate. 25 mm.

The larva should be sought for on Thalietrum.

North Carolina; Iowa; Nebraska; Manitoba. 2. E. trifurcella Chambers. White, thorax with a black central stripe and a dot on each side of it. Palpi annulate and tipped with black. Fore wing with a black median stripe from base to apex, either trifurcate at apex or much nar-rower and flanked by two black bars. With black terminal dots and two black dots near inner margin (at ¼ and ¾ way out on wing). Antennæ dark brown. 17 mm.

No authentic material is known of this species.

Kentucky.

3. E. macelhosiella Busck. White, lightly dusted with black; the black tending 5. E. maceinosiella Busck. White, lightly dusted with black; the black tending to form streaks between the veins, especially toward costa, ending in heavier terminal dots. A broad, irregular, and slightly diffuse black band from base to apex, crossing apical fringe, and nearly interrupted by a white bar at end of cell. Hind wing gray. Fringe paler. Thorax black and white. 25-28 mm. Larva on *Phacelia* (Hydrophyllaceæ) at the beginning of May. With a tuft of hairs on wart vii of ninth segment of abdomen only, the rest of the body with the usual primary hairs. Head, tubercles, setæ, cervical and anal plates, true

legs, and plates on prolegs black. Dorsum of body dull black, with a white mid-dorsal stripe, extending onto the cervical and anal plates; a broad white lateral band, more or less suffused with pinkish and sometimes with yellow; under side gravish white. Pupa formed in a gallery in dead wood or bark, the entire summer passed in the pupal stage. Pupa stout, somewhat flattened, dark brown; with the anal prolegs preserving their hooks and taking the place of a eremaster; located on the ventral surface well before the end of the body. Moth emerging in September to November; and apparently hibernating.

Maryland; Missouri.

4. E. longimaculella Chambers. White, often slightly grayish. A black dot on vertex and front of thorax, and two on disc of thorax; two black annulations on third segments of palpus. Fore wing with a rounded black spot at end of cell and one below it and slightly farther out. Base of costal edge black; a strong black dash on outer half of cell below middle, and shorter one on base of \mathbf{R} ; also three at middle of wing below costa, one beyond cell, and one before middle of fold; rarely with the second dash in the cell, shown by the next species, trace-able. Hind wing light gray. 22 mm. (Hyponomeuta Chambers).

Caterpillar in a very flimsy web on stoneseed (*Lithospermum*). Head dull black. Body purplish black, cervical shield large, bright orange, bisected by a yellow line; tubercles concolorons. Neck white; white or yellowish transverse bands behind second and third segments of thorax and fifth and sixth segments of abdomen, the anterior ones slender dorsally and expanded below. Feet all ringed with black. Moth in May and July; larva in July.

New York to Kentucky. New York: Plattsburg. Ithaca.

5. E. zelleriella Chambers. Closely similar to *E. longimaculella*. A heavy black bar on base of costa, one toward base, and one in outer part of cell, obliterating the discal dot; one in subterminal region opposite the last, larger than in *longimaculella*; and several streaks toward costa. 18 mm.

Caterpillar on *Phacelia dubia*, living almost completely exposed. Head black, rounded, with a gray spot on upper part of front, and a rounded spot on each side, separated from the frontal spot by the black adfrontal sclerite. Tubercles of body, except vii, in large rounded velvety black spots; thoracic feet black, prolegs white. Body white with diffuse yellow dorsal, and even less definite lateral bands. Pro- and mesothorax and first segment of abdomen banded in front with black; smoky ventrally. Abdomen with rounded black dorsal spots on segments and smaller ones on incisures; with broad subdorsal gray shades, defined on lower side, and joined by lateral oblique ones. Anal plate sooty. Caterpillar found in May; moth emerging the following April.

Montreal, Quebec to Wisconsin, and south to Texas; apparently rarer north-ward.

11. SCHIFFERMUELLERIA Hübner

(*Ecophora*, in part; *Callima* Clemens, not *Kallima* Westwood; *Epicallima* Dyar)

Palpi very long, recurved, smooth; second segment thickened, third slender, shorter. Antenna without pecten; tongne developed. Fore wing rather narrow, sublanceolate, with oblique outer margin, with all veins preserved, \mathbf{R}_4 and \mathbf{R}_5 stalked, both running to costa, the rest free, and \mathbf{Cu}_2 nearly straight. Hind wings narrower, lanceolate but rounded at apex, with all veins preserved, \mathbf{M}_1 rather nearer to \mathbf{M}_3 than to \mathbf{M}_1 ; \mathbf{M}_3 and \mathbf{Cu}_1 connate. Brilliantly marked species.

Key to the species

1. Dark brown with yellow patches.

2. Three or four rounded or irregular spots......4. dimidiella.

2. A comma-shaped, or semilunate spot two-thirds way out on inner margin. 5. coloradella.

- 1. Tricolored.
 - 2. Yerlow with red-brown markings.....l. argenticinctella.
 - 2. Mainly tawny yellow and black.

* Eyes large. Cilia on antennæ half as long as segments (Schiffermuelleria).

1. S. argenticinctella Clemens. Head pale; thorax orange; palpi brown and white; antennæ black and white. Fore wing orange and yellow; base of inner margin with a silvery band edged with black; an antemedial oblique bar, not reaching costa; a bar along costa, well out, and a semicircular one opposite it, on inner margin; all silvery and edged more or less fully with black. Base, apex, and area enclosed by the line, deep orange or chocolate brown. A brown spot at apex and some dark subterminal suffusion. Fringe pale. Hind wing fuscous. 12 mm.

End of June to August. Caterpillar under bark of elder.

Of general distribution and not very rare. New York: Ithaca, Rhinebeck, New York City.

** Eyes small. Cilia on antennæ at least as long as segments.

2. S. edithella Busck. Dark bronze brown, with golden and silvery markings, face white; palpi somewhat golden; antennæ bronzy black; vertex and thorax deep bronze; fore wing with a transverse, erect, silvery fascia at a fourth way out, widening somewhat on inner margin; and an apical fascia two-thirds way out, reaching the costa and interrupted at its middle by a golden streak; a fine. silvery, subcostal streak in median area, and two close together in middle of wing: all dark-edged. A longitudinal golden shade on and beyond cell, interrupted by the silvery markings. Fringe, abdomen, and hind wings dark bronzy brown, with silvery anal tuft. 9-10 mm.

Center Harbor, New Hampshire; Delaware Water Gap, Pennsylvania.

3. S. lucidella Busck. Palpi golden yellow. Antennæ black with silvery white tips. Head and thorax bronzy. Fore wing black on margins toward base, along dorsum, and around apical edge; the rest a yellow area, covering costal half of wing and sending a long process toward apex; a narrow, black-edged blue fascia a third way out, crossing the golden area and ending in a pale yellow discal spot. Two pairs of black-edged longitudinal streaks in cell; a small pale yellow spot at middle of costa; and a dash two-thirds way out. Fringe iridescent blackish. Hind wing, abdomen, and legs black; tips of tarsi and spurs silver white. 12 mm. Early June.

Western Pennsylvania.

4. S. dimidiella Walsingham. Dull brown, spotted with straw yellow. Face white, antennæ black and white, with white scape. Fore wing with a very large yellow triangle resting on inner margin near base, and smaller spots on costa at middle and before apex, the latter spot continued as a curved yellow post-medial line to inner margin. 12 mm.

This moth can be superficially distinguished from *Ecophora borkhausenii* by its fringe and thorax being mostly black; from *Symmoca novimundi*, by its much more extensively pale head; and from *Triclonella determinatella*, by the basal spot not reaching the costa. It flies from the end of April till June. Toronto, Ontario, and west to British Columbia and California.

5. S. coloradella Walsingham (variety). Smoky gray-brown, somewhat powdery and dull. Some yellow on vertex and thorax; a half-lunate spot on inner margin extending a third way across the hind wing. (In the northern form there is a narrow yellow dorsal edge extending from this spot to the base of the wing, and continued as a pair of streaks on the thorax.) 15 mm.

The typical form of this species occurs in Colorado and lacks the yellow dorsal streak. It is very close to *sulphurella*, the type of Œcophora; in fact all the species here listed, except *argenticinctella*, might well be transferred to that genus. My only date for this species is July 28.

Parry Sound, Ontario; British Columbia.

12. DECANTHA Busek

(*Ecophora*, in part)

Similar to Schiffermuelleria, but palpi not so long; antennæ with strong pecten; fore wing with \mathbf{R}_4 and \mathbf{R}_5 completely fused; hind wing with \mathbf{M}_3 and \mathbf{Cu}_1 separate.

1. D. borkhausenii Zeller. Blackish with yellow thorax, rounded patches near base and middle of fore wing; a less regular transverse subterminal bar, and a smaller postmedial spot on inner margin; the spots more or less white-edged. Fringe yellow, hind wing brown. 15 mm. (boreasella Chambers).

Caterpillar translucent white, with yellow-brown head, cervical shield, etc.; under bark of pine, in June; the moth emerging in June and July.

Europe. Cohasset, Massachusetts; District of Columbia; Arizona; British Columbia. New York: Ithaca.

American specimens (boreasella) are aberrant, and may represent a distinct species.

13. FABIOLA Busek

(*Ecophora*; Borkhausenia, in part)

Similar to Borkhausenia, but lacking one radial vein. No pecten; palpi slender, third segment nearly as long as second. Male antenn ∞ with long cilia; tongue developed, M_3 and Cu_1 of hind wing connate.

1. F. shalleriella Chambers. Dark brown, broadly shaded with golden yellow, but leaving the edges of the markings narrowly brown. A white antemedial fascia, broadening regularly to inner margin, erect, and with brilliant blue iridescence toward costa; three or four large, partly confluent, black spots in lower half of median area, each centered with a metallic blue spot. An erect blue subapical fascia across the wing, followed by a large cream-white spot on costa. Fringe dark brown, whitish at base toward anal angle; hind wing and body dark brown. 12 mm.

May southward, June in Pennsylvania.

New Jersey to western Pennsylvania and Georgia.

14. *ŒCOPHORA* Latreille

(In part; *Dasycera* Haworth)

Similar to Borkhausenia; no ocelli; eves small; antenna thickened more or less with scales toward base, and heavily ciliate in male (cilia three times as long as segment); palpi smooth, maxillary palpi distinct; tongue weak, scaled to its tip. Fore wing broad toward tip and bluntly rounded; hind wing broad-lanceolate; \mathbf{M}_2 quite variable, often nearer \mathbf{M}_1 than \mathbf{M}_3 in both wings.

7. C. newmannella Clemens. Bronze-black; second segment of palpi yellow; a yellow streak in base of fold and one near base of cell of fore wing. Antennæ purple-black, white-tipped. 15–18 mm.

The moth flies in June in the North, in May in North Carolina. It is generally distributed and not rare.

New York: North Elba, Batavia, Rock City (Cattarangus County), Portage, Ithaca, Trenton Falls, Albany, New Windsor.

Typically, both yellow spots are large, being separated only by the black vein, and the terminal segment of the palpus is mostly yellow. I have seen a variety from North Carolina with only a narrow basal yellow streak, or, at most, a few yellow scales in the cell. This form seems also to have far less rough scaling on the male antennæ, which makes the ciliation appear longer.

15. TRICLONELLA Busek

Similar to Borkhausenia, except for the stalking of \mathbf{M}_2 in the hind wing; \mathbf{M}_2 rising from near middle of end of eell in fore wing.

1. T. pergandeella Busck. Othre yellow; head brownish black with a very thin white line over eye; second segment of palpi with four broken silvery lines, and third segment with one in front. Fore wing with a minute black point in middle of cell. Outer two-fifths of wing purple-gray, the boundary edged with white, and running out on costa. A little white scaling on the gray. Fringe duller, pale-tipped. Hind wing and abdomen fuscous. 15 mm.

The moth has been obtained in late June and late August. It hibernates in the imago. There are two or three broods. The caterpillar feeds on Desmodium, in a case made of two leaflets. It is black, with the head and anterior half of prothorax yellow; a black spot over the eye, and two on the vertex. There are a dorsal and two lateral yellow spots on the meso- and metathorax, small dots on the first three segments of the abdomen, and large patches on the fourth and fifth segments. When young, the larva is predominantly yellow. The cocoon is translucent and oval, and is suspended in a case of three leaflets. There appears to be a form of this species in Arizona.

District of Columbia.

 \cdot 2. T. determinatella Zeller. Deep purple brown; a large, antemedial yellow area, nearly as long as wide, and reaching both margins broadly, with straight onter boundary; and an oblique postmedial area half as large, touching only the costa. Fringe and body dark. 10 mm.

June.

Missouri to Louisiana and west.

16. BORKHAUSENIA Hübner

(*Ecophora*, in part)

Palpi moderately long, curved (but less so than in the preceding genera); second segment thickened but smooth, and third, in *ascriptella*, about as long. Tongue developed; pecten strong. Antennæ ciliated in male, the cilia long in *ascriptella*, which belongs to subgenus Crossophora Meyrick. Fore wing (fig. 143) lanceolate, normal; hind wing with all veins present, **R** and **M**₁ parallel; **M**₃ and **Cu**₁ connate.

1. B. ascriptella Busek. Straw yellow, sparsely dusted with black seales. Palpi light cehreous; fore wing with base of costal edge black; a streak or spot in cell and one at end of cell. Hind wing pearl white with yellowish fringe. Fore legs black in front. 10 mm.

legs black in front. 10 mm. The moth looks like a Tinea but is easily recognized by the smooth head. It has been taken from June to August.

New York to western Pennsylvania and District of Columbia. New York: 'thaca.

17. ENDROSIS Hübner

Palpi long, upturned, smooth; second and third segments equal; antennæ with large scape bearing a peeten. Fore wing with all veins present; laneeolate, normal. Hind wing with \mathbf{M}_3 lost and \mathbf{M}_2 commate or short-stalked with \mathbf{Cu}_1 ; \mathbf{R}_1 as strong as any vein, but close to base (fig. 145).

The genus seems transitional to the Blastobasidae, and in the past has also been placed in the Elachistidæ. The larva appears to be as in the Blastobasidæ.

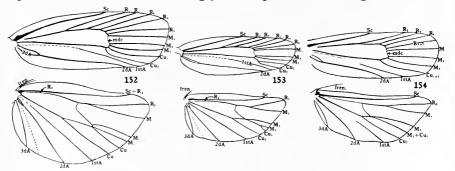
1. E. lacteella Schiffermüller. Dirty white, mottled and dusted with fuscous. Head, thorax, and extreme base of fore wing pure white, followed by a broad fuscous band. Base, middle, and apex of costa also blackish. Three vague blackish dots on disc,— in middle and at end of eell, and below and before the first in fold. 15–18 mm.

A general household insect, feeding on fruit, fungi, meal, etc., Caterpillar yellowish white with yellow-brown head and cervical shield. Injurious in Europe and on the west coast of the United States. Only sporadic specimens have been reported from the East.

Family 20. XYLORICTIDÆ

(Stenomida; Uzuchida)

Head smooth. Palpi smooth and upturned to vertex or beyond, with second joint slightly thicker and squarely cut off. Thorax smooth-scaled; hind tibiæ with loose hair. Fore wing typically with all radials preserved (one lost in Ptochoryctis and Menesta), \mathbf{R}_5 often free, running either to costa or to outer margin. \mathbf{Cu}_1 and \mathbf{Cu}_2 often stalked; completely united in Menesta; in the typical group widely separated; \mathbf{M}_3 in some forms from the base of their stalk. The other veins all present and free. 1st A strongly developed. Hind wing with Sc and



FIGS. 152-154. XYLORICTIDÆ

152, Stenoma algidella, venation; 153, Stenoma humilis, venation; 154, Menesta melanella, venation

R connected by a more or less distinct cross vein (\mathbf{R}_1) ; **R** and \mathbf{M}_1 long-stalked, the other veins normal; one medial lost in Menesta. Fore wing normally much more than twice as long as wide; shorter in Menesta and Setiostoma. Hind wing typically much broader, sometimes more than twice as broad.

Caterpillar with front less than half height of head; head held more or less horizontally; adfrontals narrow, and not extending much above front. Body with primary setæ only, above vii; vii represented by 3 to 6 setæ. Thoracic legs adjacent, prolegs normal, short, each with a complete ellipse of bi- or triordinal hooks.

Ptochoryctis, with its case-bearing larva, and its multiple setæ on the prolegs, approaches the Lacosomidæ, but differs in lacking the characteristic, rough, enlarged head, and swollen abdomen thicker in the middle. The pupa of Stenoma is short and flattened, and has minute maxillary palpi lying close to the antennæ. The clypeal suture is not distinct. The antennæ meet on the midventral line beyond the tip of the tongue and then diverge again. The labial palpi are covered. The ninth segment of the abdomen has a tuft of hooked set on the venter, besides the cremaster. In Menesta this tuft of setæ is on a distinct lobe.

The family divides into two well-marked groups, sometimes considered to be separate families. The Xylorictinæ (Ptochoryctis), which are almost confined to the Old World, have R5 running to the outer margin and long-stalked, and Cu_2 widely separated from Cu_1 . Their larvæ are borers or case-bearers. The Stenominæ are found mostly in the New World; they have \mathbf{R}_5 most often running to costa and almost always free, and \mathbf{Cu}_2 usually connate or stalked with \mathbf{Cu}_1 . Their larvæ are brilliantly marked and live exposed. The larva of one neotropical species bores in fruit. Besides the species here included in the family, some authors would add Strobisia, and perhaps other genera from the Gelechiidæ.

Key to the genera

1. Cu_1 and Cu_2 preserved, often stalked. 2. R_3 and R_5 long-stalked; R_4 lost..... 2. R_4 and R_5 separate or hardly stalked; R_3 free.4. Ptochoryctis.

1. STENOMA Zeller

(With Ide Chambers, Brachiloma Clemens, Harpalyce Chambers)

Palpi smooth, upturned beyond vertex; ocelli absent; male antennæ heavily ciliate, without pecten. Fore wing about two and a half times as long as wide; costa arched, apex rounded, outer margin strongly convex, nearly erect and short; inner margin more nearly straight. \mathbf{R}_{s} normally free and running to costa, in one of our species running to outer margin and connate or very shortly stalked with \mathbf{R}_{i} , which in all the United States species runs to the costa. \mathbf{M}_{i} and M_2 free, nearly straight; M_3 nearly straight, rarely arising from base of stalk of Cu. Cu₁ stalked, connate, or barely separate from Cu₂, varying somewhat in a single species. Cu₂ starting from cell or from Cu-stem almost perpendicularly and then curving around parallel to Cu_1 and M_3 ; 1st A long. Hind wing much broader than fore wing, folded under it in the resting position, the wings rolled about the body. Hind wing with R and M_1 stalked, often very shortly; M_2 slightly separated from M_3 , or shortly stalked when M_3 is longstalked; M_3 stalked or connate with Cu_1 , Cu_2 arising well back from end of cell, and straight.

The genus is very large in South America, and varies enormously in structure as well as in pattern; but has not as yet been successfully divided. We have three main subgroups 1, Stenoma proper, with Cu_2 normally free in the fore wing, M_a and Cu_1 counate in hind wing, and white wings characteristically marked with gray; the thorax with a metallic posterior tuft; 2, Brachiloma Clemens (*Ide* Chambers) with Cu_1 and Cu_2 stalked, M_3 and Cu_1 of hind wing typically stalked, and no metallic tuft; 3, a group containing *S. mistrella* Busek, with marked apex and with R_1 and R_5 forking over it.

The caterpillar is as noted in family description. It has primary set: only, and tubercle iv+v of the abdomen directly below iii. The pupa is described under the family.

Key to the species

I. White marked with gray.
2. Larger; terminal line light gray, continuous, followed by black bars in
base of fringel. schlægeri.
2. Smaller; a series of black terminal bars on membrane, not preceded by
a fine gray line
1. White, practically immaculate4. restalis.
1. Gray with complex darker pattern
1. Not white; with darker discal dot only.
2. R ₅ to outer margin, apex marked
2. \mathbf{R}_5 to costa; apex often bluntly rounded.
3. Ground light dull ochreous, with strong discal dot5. unipunctella.
3. Ground chocolate brown, hind wing cream, contrasting6. decorosella.
3. Ground gray

1. S. schlægeri Zeller. White; fore legs smoky; middle tarsi ringed with gray. Fore wing with a double gray basal line from costa; a conspicuous, mixed gray and black, somewhat raised patch on inner margin from basal angle almost to middle of fore wing, formed of more or less confluent waved bands; discal bar gray, edged finely with white, and then broadly and irregularly with gray, and usually with a gray band running to inner margin; broad, even, pale gray postmedial and subterminal bands, and a very narrow terminal one cut into spots. A black line, usually broken, in base of friuge; much more contrasting than the terminal line. Hind wing pale gray with white fringe. Thorax with prominent lead-colored posterior tuft. Male with simple curved uncus and very broad subscaphium broadly notched at tip. Fore wing with Cu_1 and Cu_2 usually connate, separating at right angles; M_a and Cu_1 of hind wing connate. 30 mm. (H 48:41).

General in distribution and not rare, in May and June. Caterpillar on oak. New York: Mt. Marcy, Rock City (Cattaraugus County), Ithaca, McLean, Big Indian Valley, Bethlehem, New Windsor.

2. S. algidella Walker. Very similar to S. schlægeri, with very little or no black toward base of inner margin, but in the northern form with a large gray patch. Terminal line broken into dots, dark gray and contrasting; dark dots in base of fringe obscure, paler. Uncus of male with broadened bifd tip, subscaphium simple, slender, ending in a sort of beak. Cu_2 of fore wing well separated from Cu_1 (fig. 152) 15-23 mm. (*leucillana* of authors, not Zeller).

The distribution is general and the species is not rare in June and August. The caterpillar is common on maple and various trees and shrubs, and has been found

boring in an apple. It is not certain that this name is correctly applied to this species, but there seems to be no other available.

New York: Rock City (Cattaraugus County), Ithaca, Trenton Falls, Big Indian

Valley, Fort Montgomery. 3. S. humilis Zeller. Cu_1 and Cu_2 stalked in fore wing; M_3 and Cu_1 long stalked in hind wing (fig. 153). No metallic tuft on thorax. Fore wing ash-gray, dusted and shaded with white. Three diffuse blackish spots on costa and two transverse fasciæ resting on inner margin, the inner starting from a black dot in fold. Three dark dots in cell, the middle one not so intense. A faint curved dark subterminal line and a series of terminal dots. Hind wing browner than fore wing. 15 mm. (canusella Chambers).

March to May; July and August.

Southern New Jersey, Missouri, and southward. 4. S. vestalis Zeller. White, with a few yellowish or black scales forming a slight discal dot. Hind wing and under side slightly yellowish. 18 mm. (albella Chambers).

This moth is well known in the Southern States but may not exist in our territory.

5. S. unipunctella Clemens. Straw color, dusted heavily and evenly with pale yellow-brown, producing an effect of deep ochre; the dusting gathering to form a dark discal dot. Cu_1 and Cu_2 stalked. Hind wing yellowish white. M_3 and Cu_1 connate. 20 mm. (lithosina Zeller).

Maryland to Illinois, Missouri and southward. Larva on laurel oak.

6. S. decorosella Busck. Rich brown, rather like S. unipunctella, but more deeply colored. Palpi brownish ochreous, with tip of second segment and base of third whitish. Head paler, in some lights yellowish white; antennæ and thorax light brown. Fore wing deep brown, silky; costal edge often showing the yellowish-white under side. Discal dot darker. Hind wing whitish fuscous; fringe yellowish. 22-24 mm.

New Jersey; North Carolina. 7. S. querciella Busck. Antennæ as in S. decorosella; palpi whitish, second segment light gray on outer side. Thorax dark gray. Fore wing silky mousegray, slightly darker on fold, with two obliquely placed dark discal dots. Hind wing paler yellowish gray. 22-24 mm. Caterpillar on oak in July.

New Jersey and Pennsylvania to Texas.

8. S. mistrella Busck. Fore wing dull light yellow-brown, slightly powdery, with obscure dark discal dot and antemedial spot in fold, and obscure terminal dots. Hind wing dark gray with light brown fringe. \mathbf{R}_4 and \mathbf{R}_5 usually connate or slightly stalked, R5 running to outer margin; hind wing with M3 and Cu1 perceptibly separate. 18 mm.

Larva on timothy. Moth in July and September.

Western Pennsylvania to northern Illinois and Missouri.

2. SETIOSTOMA Zeller

Antennæ heavily ciliate; palpi very slightly thickened with rough scales, upturned beyond vertex, with third joint a little longer than second; no pecten. Legs distinctly tufted at spurs. Fore wing but little longer than wide, with Cu, arising well before end of cell; Cu_2 at two-thirds way out on the cell; R_6 in some specimens running to costa. Hind wing not large; M₃ stalked, rarely united with Cu₁.

This genus was formerly placed in the Glyphipterygidæ, but is obviously Stenomid. 1. S. xanthobasis Zeller. Basal third of fore wing lemon yellow, with black costa and inner margin; the rest iridescent black, bounded by an iridescent blue fascia. Hind wing black. 12 mm.

July. Larva shaded with green and crimson. Head, cervical shield, and true legs pale brown; a dark spot on center of head; cervical shield narrow; anal plate brown. Larva in a nest about 2 cm. in diameter, formed by an oval wall of silk between two slightly separated leaves; feeding on the lower parenchyma only. On *Quercus stellata* and related species, in May.

Massachusetts to Florida.

3. MENESTA Clemens

(*Hyale* Chambers; *Strobisia*, in part)

Male antennæ heavily ciliate; palpi smooth, second segment thickened, third a little shorter; fore wing (fig. 154) twice as long as wide, with square outer margin; 10 veins, all separate; \mathbf{R}_4 and \mathbf{R}_5 united, one dorsal vein absent, 1st A well developed. Hind wing slightly broader, \mathbf{M}_2 absent, \mathbf{R} and \mathbf{M}_1 stalked, \mathbf{M}_3 and \mathbf{C}_4 , connate.

This is an aberrant and isolated genus, with the superficial appearance, genitalia, and pupal characters of the Stenoma group. The preservation of 1st A in the imago also shows that it belongs rather in this neighborhood than among the Gelechiidæ, where it is commonly placed.

Key to the species

	large white costal spot
1. A	minute costal point or none.
2.	Fringe white-tipped at apex
2.	Fringe not white-tippedl. tortriciformella.

1. M. tortriciformella Clemens. Black-brown, with a purplish hue; head and antennæ deep yellowish fuscous; face pale below; palpi fuscous, whitish toward base. Fore wing with a small lunate white spot, convex outward, at end of cell; fringes and hind wing fuscous; feet pale. 9 mm. (*liturella* Walker, coryliella Chambers.)

Caterpillar white; on hazel, in September and October; under a thin silken web placed in the angle between a vein and the midrib, on the under side of a leaf. Pupa in a cocoon formed by crumpling up this web. Moth in April.

Massachusetts to Kentucky.

2. M. melanella Murtfeldt. Shining black; face, palpi, under side, a spot on metathorax; a large spot on costa of fore wing at middle, and apical half of fringe, pure white. 8 mm.

Caterpillar with the same habits as M. tortriciformella; on oak. Moth in May and August.

Virginia; Kentucky; Missouri.

3. M. albaciliella Chambers. Purple-black; face, most of palpi, under side, a small and variable irregular spot at end of cell (usually a short discal streak), and outer half of fringe except at anal angle, contrasting white; thorax wholly black. 9 mm. (argenticiliella Chambers.— lapsus calami, Strobisia Chambers.)

Caterpillar with the same habits as the other species; on under side of blackberry leaf, in June; skeletonizing the leaf and later eating holes through it. Two broods; moth in July and in early spring.

New York to Pennsylvania and Ohio. New York: Peru.

4. *PTOCHORYCTIS* Meyrick

Fore wing with 11 veins, \mathbf{R}_s and \mathbf{R}_{4+5} stalked, both running to costa. \mathbf{Cu}_2 arising from cell four-fifths way out, straight; \mathbf{M}_s and \mathbf{Cu}_1 short-stalked. Hind wing hardly broader, with seven veins; \mathbf{M}_{2+3} and \mathbf{Cu}_1 connate; \mathbf{R} and \mathbf{M}_1 connate. Ocelli

present; male antennæ bipectinate, without pecten on scape. Larva a casebearer on evergreens, with six setæ on prolegs; flattened, widest at first segment of abdomen; setæ iii of spiracle-bearing segments with a double annular tubercle, apparently movable. Pupa in the cocoon.

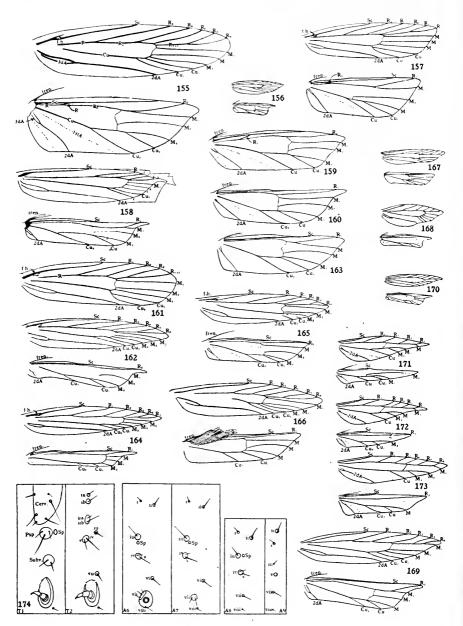
1. P. tsugensis Kearfott. Silvery white, veined with gray; with a narrow terminal gray shade immediately preceded by a blackish line. Hind wing yellowish white. 21-24 mm.

Larva on Tsuga (Japanese hemlock); cream-white, speckled with pink, with blackish head and cervical shield; in a case covered with needles and frass. New Jersey. Introduced from Japan (?)

Family 21. GELECHIIDÆ

Moderately small moths, sometimes minute. Head typically smoothly scaled, clothed with long scales curving forward and down, and sometimes ruffled. Labial palpi normally as described for the superfamily; second joint often with a triangular tuft. The third joint rudimentary in male of Anarsia only. Antennæ as in Ecophoridæ; very rarely with pecten (Sitotroga, Symmoca ?). Fore wing narrow, with rounded, rarely caudate (fig. 158) outer margin. \mathbf{R}_{z} running to costa, stalked, or, rarely, completely fused with \mathbf{R}_4 ; \mathbf{R}_3 usually free; M, from near upper angle of cell or stalked with \mathbf{R}_s ; Cu, and \mathbf{Cu}_2 often stalked, never very widely separated; the other veins mostly free, and hardly ever with more than one vein lost. M, arising nearer Cu-stem than R-stem. 1st A absent (except the extreme tip in some exotic Symmocas); 2d A strongly forked at base. Hind wing more or less trapezoidal, the outer margin more sharply bent over \mathbf{M}_{a} and \mathbf{Cu}_{a} , and often concave above, or with projecting apex; rarely bifid; lanceolate in some Helices. Sc and R connected by a well-developed \mathbf{R}_1 near base; \mathbf{R}_1 and base of \mathbf{R}_s both lost in Helice; R and M, approximate at origin or stalked, except in a few genera with markedly concave outer margin; \mathbf{M}_2 various; \mathbf{M}_3 and \mathbf{Cu}_1 connate or stalked, rarely separate. 1st A lost, and 3d A very weak. M_1 , or \mathbf{M}_{2} , or both, sometimes disappearing in forms with concave outer margin; and \mathbf{M}_3 rarely fused with \mathbf{Cu}_1 .

Larva (fig. 174) typical of the superfamily, as a rule; coxæ often a little separated, but by less than their width; adfrontals reaching nearly or quite to vertex; and front two-thirds as far, or ending in an attenuated point; prolegs almost always with biordinal hooks, either in an ellipse or in two transverse bands; in the latter case with two groups on anals. Second, third, and fourth ocelli not distinctly grouped. Prolegs lost and true legs rudimentary in a few genera. Pupa with elypeal suture complete and usually straight; labial palpi concealed, or showing a small triangle next to labrum; femora concealed. Antennæ contiguous for some distance behind tongue, and then again diverging before apex; maxillary palpi present and usually large, reaching from



FIGS. 155–174. GELECHIIDÆ

155, Gelechia lugubrella, venation; 156, Stomopteryx taniella (Europe), venation; 157, Trichotaphe fernaldella, venation; 158, Polyhymno luteostrigella, venation (the outline of the apical part of the fringe is also shown); 159, Gnorimoschema gallæsolidaginis, venation of hind wing (typical of the normal group of the genus); 160, Gnorimoschema scutellariæella, or a closely related species (typical of the reduced group of the genus); 161, Glyphidocera species, venation of fore wing; 162, Evippe leuconota, venation; 163, Telphusa longifasciella, venation of hind wing; 164, Aristotelia disconotella, venation (figured from a specimen from Rock City, New York); 167, Recurvaria leucatella (Europe), venation; 168, Chrysopora eppelsheimi (Europe), venation; 169, Epithectis subsimella, venation; 170, Metzneria carlinella (Europe), venation; 171, Theisoa constrictella 3, venation (after Braun); 172, Theisoa constrictella Q, venation (after Braun); 173, Theisoa pallidochella Q, venation (after Braun); 174, Dichomeris verbascella (Europe), seta map of larva

9

antenna to tongue. Cremaster present and sometimes with some hooked setæ on venter of ninth segment of abdomen as well.

The family is probably the largest of the Tineid series, and is especially developed in the tropics. Most of the subordinate groups are widespread, but the forms with a pecten are of Old-World affinity, and the Strobisia type is South American, like the Stenoma group of Xylorictidæ, which they resemble.

Key to the genera

Moth

11 0 0 10
1. Fore wing with M_1 stalked with R_5 beyond origin of R_4 , which is distinct (fig. 170).
2. Pecten present
3. Fore wing with a vein absent.
4. \mathbf{R}_{1} absent, the first developed radial arising from cell close to end. 32. Agnippe.
 4. A dorsal vein absent; R₁ from cell two-thirds way out. (figs. 171, 173). 5. Hind wing with R or M₁ lost
3. Fore wing with all veins preserved.
 Hind wings with M₃ and Cu₁ widely separate (fig. 170). Second joint of palpus with long spreading hairs or a triangular tuft.
31. Ptycerata.
5. Second joint of palpus rough-scaled; expanse over 12 mm. 30. Metzneria.
5. Second joint of palpus not very rough; expanse under 12 mm.
4. Hind wings with M_3 and Cu_1 connate, stalked, or barely separate (fig. 169).
5. Hind wings with M_1 absent (R-stem apparently simple) (fig. 162). 28. Evippe.
5. Hind wings with \mathbf{M}_1 present, attached to R-stem. 6. Fore wings with \mathbf{M}_3 and \mathbf{Cu}_1 stalked27. Trypanisma.
6. \mathbf{M}_{s} not stalked with \mathbf{Cu}_{1} . 7. Second joint of palpus with a long, expansible tuft on inner
side25. Eucordylea. 7. No such tuft.
8. R and M_1 of hind wing approximate at base24. Recurvaria. 8. R and M_1 stalked
1. Fore wings with \mathbf{M}_1 arising from cell, or more shortly stalked than \mathbf{R}_4 (or with \mathbf{R}_4 absent).
2. Cu_1 and Cu_2 stalked (fig. 157).
3. Second joint of palpus with a long triangular tuft below. 9. Dichomeris, 11. Anorthosia.
3. Second joint of palpus without such a tuft; sometimes broad-scaled on
both sides. 4. Hind wings narrower than fore wings12. Strobisia.
4. Hind wings broader than fore wings.
5. Fore wings with 11 veins (\mathbf{R}_s united with \mathbf{R}_4 as shown in fig. 161). 10. Glyphidocera.
5. Fore wings with all veins present; \mathbf{R}_4 and \mathbf{R}_5 stalked. 6. \mathbf{R}_2 shortly stalked with \mathbf{R}_{4+5}
6. $\mathbf{R}_{\mathbf{s}}$ free

2. Cu_1 and Cu_2 arising from cell separately.

- 3. Hind wings only, with one vein (M) lost (fig. 168).....23. Chrysopora.
- 3. Both wings with all veins present.
 - 4. Hind wing with **R** and \mathbf{M}_1 parallel, or not more than twice as far apart at tip as at origin.
 - 5. Hind wings with M_3 and Cu_1 separate.
 - 6. Costal margin of fore wings concave before apex......22. Enchrysa.
 - Costal margin of fore wings normal.
 Fore wing smooth-scaled; hind wing with R and M₁ parallel; M₂ distinctly nearer to M₁ than to M₃ (fig. 164)....21. Aristotelia.
 R and M₁ of hind wing divergent; M₂ nearer M₃ (fig. 166).
 - 8. Fore wing with raised scale tufts; ground nearly white.

8. Fore wing not tufted; ground dark gray.....19. Glauce. 5. Hind wings with M_3 and Cu_1 connate or stalked.

- 6. Hind wings with large costal hair-pencil.....4. Phthorimœa (male). 6. Hind wings without hair-pencil.
 - 7. Second joint of palpus with divided scale-ridge, third joint thickened with scales toward base.
 - 8. Thickening of third joint, of appressed scales; fore wings normally without black, white-tipped scales; larvæ on Solanaceæ4. Phthorimœa (female).
 - 8. Thickening of third joint reaching nearly to apex, leaving a short fine apex; fore wing usually with a large proportion of finely white-tipped scales; larvæ almost always on Compositæ3. Gnorimoschema.

7. Second joint of palpus without divided brush; third joint thin. 16. Paralechia.

- 4. Hind wings with R and M_1 approximate, connate, or stalked at origin (figs. 155, 163).
 - 5. Hind wings with M_3 and Cu_1 separate (fig. 163).
 - 6. Second joint of palpi without such tuft.
 - 7. Male normal; M_1 of fore wing free or barely stalked with R_{4+5} . 18. Telphusa.

7. Male with large tuft on costa of hind wing; M_1 often stalked almost out to origin of R₄.....19. Glauce. 5. Hind wings with M_3 and Cu_1 connate or stalked (fig. 155).

6. Terminal joint of palpi about as long as second.

7. Second joint of palpus with large triangular tuft.

- 7. Anarsia (female). 7. Second joint of palpus rough-scaled beneath, often with divided brush2. Gelechia. 7. Second joint of palpus smooth scaled.

8. Hind wing with outer margin sinuate, concave below apex. 9. Fore wing not caudate.

10. Hind wing with R and M₁ stalked.....14. Stomopteryx.

9. Fore wing candate (fig. 158).....15. Polyhymno. 8. Hind wing somewhat trapezoidal, but with outer margin not

concave below apex. 9. Hind wing much broader than fore wing, R and M, approxi-

^{17.} Arogalea.

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9. Hind wing hardly as broad as fore wing.

- 10. R and M₁ long-stalked.....l. Symmoca. 10. R and M₁ connate or approximate; M₂ and M₃ very widely

Key to known genera

Larva (Fracker)

1. No prolegs.

2. Body swollen and strongly tapering to ends (in burdock heads).

30. Metzneria. 1. Prolegs present.

- 2. A complete circle of hooks.
 - 3. Seta iii of eighth segment of abdomen in front of spiracle (rarely obliquely above it); setæ large, body striped.
 - 4. Seventh and eighth segments of abdomen both with setw ii farther from mid-dorsal line (and from its mate), than i.....17. Arogalea.
 - 4. Seventh segment of abdomen with seta ii closer to mid-dorsal line than i, eighth segment normal.
 - 5. Triangle formed by fourth, lower, and posterior ocelli, with a right angle at the fourth; pattern wholly of transverse stripes.

18. Telphusa.

5. Triangle formed by the three lower ocelli with an acute angle at fourth; the posterior as near the lower as the fourth; pattern usually partly or wholly of longitudinal stripes.

24. Recurvaria, 2. Gelechia.

- 3. Seta iii of eighth segment of abdomen above, usually directly above spiracle; body pale, not striped.....5. Gnorimoschema, 4. Phthorimœa.
- 2. Two transverse bands of hooks, spiracle of eighth abdominal segment very large9. Dichomeris.

Key to the genera

Known pupæ (Mosher)

- 1. Body-setæ strong, often as long as a segment, and truncate at tip; second, third, and fourth segments of abdomen each with a subdorsal projection at its anterior edge and edged in front with a fringe of whitish hairs; and behind this with a prominent hump tipped with a similar fringe of setæ. Clypeal suture strongly curved forward; cremaster always present.
 - 2. Movable segments of abdomen each with a pit with chitinized edges, on mid-dorsal line at front edge......8. Trichotaphe (flavocostella).
 - 2. No such pit; a deeply punctate band, followed by a groove, a band of short spines, and a strong ridge.....9. Dichomeris.
- 1. Body setse not modified; second to fourth segments not as described in the alternative.
 - 2. Pupa densely covered with white setæ.
 - 3. Maxillary palpi reaching tongue, antennæ broadly in contact on middle line; cremaster short and blunt, with hooked setæ at tip..5. Anacampsis.
 - 3. Maxillary palpi not reaching tongue; antennæ meeting at a point only; cremaster sharply curved at tip, with spines on sides.

21. Aristotelia (salicifungiella).

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2. Pupa not setulose.

- 3. Seventh segment of abdomen with a dense fringe of seta on some portion. 4. The fringe on the front and sides of a prominent lateral cavity only.
 - 5. Body smooth, not depressed; front edge of cavity trilobed. .28. Evippe.
 - 5. Body strongly depressed, spinulated; front edge of cavity bilobed.

18. Telphusa.

- 4. Fringe extending around the segment or nearly so.
 - 5. Fringe extending around the segment in a straight line; body smooth
 - 5. Fringe extending around in a wavy line; body depressed; surface with punctures or spines.
 - 6. Fringe edging two very large lobes on dorsal surface; last three
 - 6. Fringe not edging two lobes; last three segments not tapering.

2. Gelechia.

- 3. Seventh segment without a fringe of setæ.
 - 4. Cremaster with short projecting spines.

5. One spine on mid-dorsal line; clypeal suture nearly straight.

4. Phthorimœa.

- 5. An additional spine on each lateral margin; clypeal suture strongly curved forward 4. Cremaster with curved or straight setæ.
- 5. With hooked setæ.
 - - 6. Antennæ reaching tips of wings; divergent at tips, uncovering tips of legs.

 - 7. Two pairs or less of long curved setæ, front end of body rounded. 21. Aristotelia (physaliella).

6. Antennæ not reaching caudal end of wings, nor diverging at tips. 3. Gnorimoschema (lavernella).

5. Short straight setæ only......3. Gnorimoschema (gallæsolidaginis).

1. SYMMOCA Hübner

(*Ægoconia* Stainton)

No ocelli; head normal; antennæ subserrate; palpi with second segment long; smoothly but densely scaled below; third long, thin, and acute; wing form about like that of Ecophora, but hind wing very slightly trapezoidal; fore wing with R_s , in our species, running to costa, Cu_1 and Cu_2 free in the American species; hind wing as wide as fore wing, R and M_1 stalked, in *S. novimundi*, halfway to apex; M_s and Cu_1 connate; 1st A distinct in both wings. Larvæ hardly known, apparently feeding on lichens. This is a primitive genus, transitional from the Ecophoridæ and by some authors associated with them. Our species is an apparently feeding on lichens. This is a primitive genus, transitional from the Ecophoridæ, and by some authors associated with them. Our species is an

Ægoconia and very near to S. (*Œ*.) quadripuncta Haworth. 1. S. novimundi Busck. Dark fuscous; second joint of palpus dark brown, with yellow inner side and apex; extreme tip of third ochreous. Face, tips of tegulæ, and two small posterior dots on thorax, ochreous. Fore wing with three ochreous costal spots, the one at end of cell largest; sometimes these spots are lengthened into irregular bands, and sometimes there is a fourth on inner margin opposite the last costal. Hind wing and anal tuft lighter. 12-13 mm.

August and September.

Montclair, New Jersey; Roxborough, Pennsylvania.

2. GELECHIA Hübner

(Cirrha Chambers; Eseis Chambers; Pseudochelaria Walsingham; Teleia Heinemann; Lita Treitsehke)

Palpi with second joint rough-scaled; most often with a divided brush or tuft which is sometimes very large; third smooth, more slender than in Gnorimoschema. Fore wing (fig. 155) normal, oblong or pointed, variable in breadth; M_3 and Cu_1 sometimes stalked in *G. bosquella*; hind wing never much narrower and usually rather broader than fore wing, typically trapezoidal, strongly sinuate below apex in subgenus Lita; **R** and M_1 sometimes stalked, M_3 and Cu_1 connate or short-stalked; M_2 approximate to M_3 .

Eseis Chamhers includes the species with very heavy palpal tuft; **Teleia** those without ocelli, and otherwise normal. This is a somewhat heterogeneous genus, which should be subdivided. It includes forms transitional to several other genera, notably Anacampsis, Gnorimoschema (or rather, the primitive aberrant Gnorimoschemas which are themselves transitional to Phthorimœa) Telphusa, and Duvita.

Key to the species

I. Ground color black (or dark brown or gray, with contrasting pale markings).
2. Inner margin yellow to rust red10. bosquella.
2. With white or whitish markings only.
3. Head pure white.
4. White markings diffuse
4. White markings clean-cut.
 Two white fasciæ and a triangular spot between them9. viduella. One incomplete fascia and several spots1. cercerisella. Head more or less mottled or dark.
4. A white oblique streak or fascia from costa a third way out.
5. A fascia also at two-thirds way out.
6. A triangular white spot at middle of costa
6. No spot at middle of costa
4. No such streak.
5. An angulate white fascia at apical third6. bimaculella. 5. No such fascia.
6. A white posterior spot on thorax
6. No white spot on thorax.
7. Second segment of palpus white or yellow.
8. Ordinary spots white
8. Ordinary spots black, hardly visible
7. Second segment of palpus dark
1. Ground color not black or blackish with pale markings.
2. A contrasting brown-black bar or patch at basal angle.
3. An oblique light bar a third way out; patch larger30. walsinghami.
3. No such bar; patch smaller
2. No black patch at basal angle.
3. A black or contrastingly dark oblique bar running to costa near base;
the ground before it pale fuscous, luteous, or cream white.
4. Bar diffuse on outer side; no decided outer fascia29 mediofuscella.
4. Bar sharply defined on outer side; a similar black postmedial fascia.
5. Outer margin heavily edged with powdery black; the orbicular a
black point, well separated from the outer fascia28. pseudofondella.
5. Outer margin without definite black border; orbicular large and fused
with outer fascia, or at least with reniform

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- 3. No black antemedial oblique fascia running to costa; or else with ground powdery gray to base, and fascia diffuse. 4. Ground dominantly luteous; pale moths.
 - - 5. Ground very slightly mottled, with three to six black dots.
 - 35. branella.
 - 5. Streaked with a darker shade of light gray......34. arenella. 5. Dusted and mottled with yellow-brown and black; tending to diagonal
 - 4. Ground dark, or so heavily dusted as to appear dark gray.
 - 5. Dorsum of thorax luteous, contrasting with the blackish tegulæ. 6. Sides of disc of thorax and third joint of palpi black.

(12). ochreostrigella.

- 6. Disc of thorax wholly yellowish white; palpi yellow, banded with12. hibiscella. black
- 5. Dorsum of thorax evenly colored, or nearly so, dorsal half of fore wing not contrastingly pale. 6. Outer discal dot black, ocellate with white or yellowish.

13. discoocellella.

- 6. Outer discal dot not ocellate, or else with the center pale. 7. Fore wing markedly darker-streaked on the veins, at least outwardly, and not notably powdery.
 - 8. Stripes black on luteous, and fine in the cell.

10. ochreosuffusella.

- 8. Stripes blackish on fuscous brown, not contrasting; usually with a thick bar in outer part of cell......11. fluvialella.
- 7. Not dark-streaked on veins; ground often powdery.
 - 8. A large part of scales with fine black tips and longitudinal lines on a gray base, producing an almost evenly gray effect; the wing under a lens apparently more finely pow-dered than usual13. anarsiella.
 - 8. Scales mostly unicolorous, or broadly black- or white-tipped.
 - 9. Crisply powdered with black or dark gray on light bluegray or white; often white-shaded toward costa; or with broad areas of light bluish-gray.
 - 10. A contrasting blackish longitudinal dash in outer part
 - 10. No such marking.
 - 11. Antemedial line represented by two fine strongly outwardly oblique lines to costa, contrasting when costa is whitish; very obscure or even obliterated when costa is dark brown.

12. Costa heavily shaded with whitish; larva on Robinia. 25. pseudoacaciella.

12. Costa concolorous, or brown; larva on Prunus. 26. serotinella.

- 11. Antemedial line nearly transverse and thick; rarely absent.
 - 12. Postmedial line whitish, contrasting, nearly straight
 - 12. Postmedial strongly bowed out in middle, when distinct.
 - 13. Second segment of palpi solid black, except extreme tip; tuft in fold preceded by a well-marked yellow bar; ground suffused with blue-gray except

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- 13. Second segment of palpus powdery gray; tuft in fold blackish, preceded by a few yellow scales at most; wing with strong contrasts.
 - 14. Moth whitish, the markings formed of black powdering on a white base, without any gray; a distinct, outwardly oblique median fascia.

17. lynceella.

- 14. More or less gray or fuscous scaling; the effect less whitish.
 - 15. Scutellum contrasting, light yellow.

19. maculimarginella. 15. Scutellum concolorous, powdery gray.

18. bicostomaculella.

13. Second segment of palpus and whole wing nearly evenly powdered black on blue gray.

15. inquilinella.

9. Fuscous, nearly even or obscurely dusted and shaded.

10. A heavy but irregular black antemedial bar..21. dyariella. 10. No such marking.

- 11. Heaviest black spots are first discal dot (orbicular), which is a bar, and antemedial dots on costa and in
- 11. Outer discal dot most strongly marked, or the discal spots obsolete.
 - 12. Second segment of palpus deep black.

13. Fore wing with indistinct white markings.

14. albisparsella.

Fore wing with faint dark markings only.
 16. unctulella, rileyella.

12. Second segment of palpus powdery gray.¹⁸ 11. *fluvialella*, etc.

1. G. cercerisella Chambers. Velvety black; palpi, head, and collar white; third segment of palpi white except at base; antennæ dark. Fore wing with a few ochreous scales, slightly bronzed; three costal spots, the first reaching fold, the second and third on inner margin; and some white terminal points. Costal cilia brown, dorsal white, with a brown line at base. Hind wing pale drab, faintly pink tinged. 14 mm.

Two or more broods. Moth from May to September. Caterpillar on *Cercis* canadensis, spinning the leaves together; white, a broken black band on face and a bowed one on vertex; posterior half of each segment above, and true legs black. Young larva wholly white; sometimes living in a single folded leaf.

Distribution general, north to central Illinois and Maryland.

2. G. coloradensis Busck. Deep black; antennæ black; second joint of palpi white with dark scales above; third mostly black with white tip. Face white; fore wing with an oblique antemedial costal streak reaching to fold, an elliptical white spot on disc, an angulate costal spot at beginning of fringe, a dorsal spot

¹⁸ G. fluvialella will run to the last alternative of the key if the darker veining, which is often obscure, is not noted. Its large size and broad wings distinguish it from most of the plain Gelechias. G. discoocellella will also run there, in the case of specimens that have lost the black dot in the reniform. It may be distinguished by its purplish overcast and pale, dot-like reniform. There are several other smallish species which have never been worked out.

opposite the last, and one less than halfway out on fold. A few white scales near apex. Hind wing dark fuscous. 15 mm.

May.

New Jersey (Benjamin) to Colorado, south to Florida.

3. G. argentipunctella Ely. Purple-black; antennæ with a yellow ring at end of scape, second joint of palpus flecked with white, and white in front; third joint white at base and tip. Thorax with a white posterior tuft; fore wing with a white costal spot two-thirds way out with black before it; two black, white-shaded, discal spots, and a third before them in the fold. Abdomen dark, legs with yellow annulations. Hind wing with fringe and hind tibial tuft yellowish; a large yellow pencil on hind wing of male. 17 mm.

July.

East River, Connecticut.

4. G. trialbamaculella Chambers. Black-brown. Second segment of palpus and face yellow. Fore wing with small white spots opposite each other two-thirds way out, and several minute white points, mostly in cell and fold. Hind wing light fuscous. 15 mm.

Caterpillar on many shrubs (locust, oak, Comptonia, Epigæa, Vaccinium) in a nest between two leaves, forming a frass tube, open at both ends, and skeletonizing the leaves. Caterpillar dirty yellow, with head, cervical shield and six longitudinal stripes darker yellow. Moth emerging in late July and early August.

I have specimens from Virginia in May and Massachusetts in June, indicating an early brood.

Maine to Virginia and Missouri. New York: (Beutenmüller). 4½. G. psiloptera Barnes and Busck. Black-brown, the head and thorax with a purplish sheen, and fore wings with contrasting yellow scale-bases visible under a lens. Second segment of palpus yellow, dusted with black; third segment black with a few yellow scales. Fore wing with ordinary spots black; almost invisible; fringe blackish fuscous. Hind wing blackish fuscous with somewhat paler fringe. 16-17 mm.

Meach Lake, Laurentians, Quebec. New York: Peru (variety).

5. G. confusella Chambers. Similar, face whitish below only; palpus dark, lightly dusted with white; fore wing with postmedial dot in fold. 15-18 mm. (persicaella Murtfeldt).

Larva on peach.

Michigan.

6. G. bimaculella Chambers. Dark purplish brown or black; extreme tip of palpus yellowish white; a white spot beyond middle of wing and a bar from base of costal fringe, not reaching inner margin. 12 mm. (ternariella Zeller, sylvæcolella Chambers).

Locally common.

New Jersey; Kentucky; and Texas.

7. G. continuella Zeller. Blackish gray, dusted with white, the white tending to form a couple of oblique bands; the large black discal dots more or less ringed with white or with the space between them white; normally with a white spot in fold beyond the first fascia, costal and dorsal ones opposite each other at beginning of fringe, a small spot at apex, and a couple on margin below. Hind wing with **R** and **M**₁, **M**₂ and **Cu**₁ stalked. 18-19 mm. (trimaculella Packard, albomaculella Chambers).

July.

Labrador; Europe. New York: Lake Tear (Mt. Marcy), Ithaca.

8. G. lugubrella Fabricius. Black, face and second segment of palpus white. Fore wing with an antemedial white fascia, running to fold, and a postmedial fascia crossing the wing, less oblique than outer margin, and more or less con-stricted at middle. 15 mm.

June.

Circumpolar; south to Maine, northern New Jersey, and Manitoba. New York: Peru

9. G. viduella Fabricius. Black, amount of white on head variable; sometimes with white on thorax also. Antemedial fascia as in lugubrella, postmedial parallel to outer margin, and often interrupted for a very short distance at middle. Fringes white-tipped. Typically with the white covering about a fourth of the wing surface; in var. labradoriella Clemens covering about half. 15 mm.

July.

Circumpolar; south to Mt. Washington, New Hampshire, and Alberta.

10. G. bosquella Chambers. Thorax and fore wing rich chocolate brown, face yellowish white; antennæ brown with extreme tip of scape white; palpi dark brown with two pale rings each on the second and third joints. Fore wing mixed brownish and bluish black, with a large, irregular, bilobed, yellow or orange dorsal patch, normally extending to costa at a third way out, and a white, postmedial costal spot. A jet black spot in cell. 12 mm. (costipunctella Möschler; Parastega Mayrick).

August and September. Caterpillar on Cassia chamæcrista, green, with head, cervical shield, feet, and the small tubercles black; thorax mostly deep purplish red.

District of Columbia to Chicago, Illinois; Kansas; and south.

G. ochreosuffusella Chambers may approach our territory on the southwest. It is dark brown; third segment of palfus light, with two narrow brown rings; thorax, base of fore wings, and a streak in fold yellowish. Fore wings finely veined with dark brown, with four fine dark lines in the cell. 18 mm. July to September.

11. G. fluvialella Busck. Brown, somewhat yellowish, the veins broadly and diffusely lined with dull black; often with the blackish color suffusing considerable areas of the wing. Discocellular vein marked with a blackish spot. Hind wing somewhat paler gray; rather broader than fore wing; palpi with second joint somewhat dusted with white, no darker than third, but darker than fore wing, brush as wide as eye, widest about the middle. Wings relatively broad, with broad fringes. 18-22 mm.

June to early July.

Pennsylvania. New York: Peru, Rock City, Ithaca.

12. G. hibiscella Busck. Dark Brown, antennæ concolorous; palpi with large spreading brushes on second segment, yellowish white with a few black scales outside, extreme tip of second segment and an annulation of third black. Face, head, and thorax yellowish white; shoulders and sometimes entire tegulæ blackish. Fore wing with brown or brown-black costal half, and ochreous dorsal half and apex; the boundary slightly diffuse. Dark part showing obscure pale spots and pale part with dark bars and striæ. Hind wing a little broader, light bluish fuscous with yellower fringe. 16 mm.

May to June; August. Caterpillars on leaves and capsules of rose mallow (Hibiscus), semisocial, when mature often cutting a leaf to make it wither, and form a shelter for pupation. Head and cervical shield black; thorax reddish brown with third segment and front of second white; abdomen white with three pairs of reddish dorsal lines. Tubercles black. Two broods; the larva hibernating.

New Jersey to Illinois.

G. ochreostrigella Chambers is a western species, or western race of G. hibiscella, which has been reported from the northeast.

13. G. discoccellella Chambers. Dark purplish brown, as a rule, shading into wood-brown toward inner margin. Veins sometimes darker. Outer discal dot yellow, usually with a black point in the center, at least in the female. Hind wing pale and yellowish, palpus light dull yellow, with second and base of third segment nearly black. 18 mm.

Caterpillar on yellow dock (Rumex), and Polygonum; green, with yellow-green

head and cervical shield; in a slight web on under side of a leaf; skeletonizing the leaf. Moth flying in late June and July and again in September.

Northern New Jersey and District of Columbia to Kansas and Texas.

A. anarsiella Chambers, an immaculate, powdery, steel gray species without any definite markings, probably occurs in the northeast; its larva feeds on Ceanothus.

14. G. albisparsella Chambers. Brush on palpus long, ragged, but not distinctly divided. Moth dark gray-brown, the head a little paler and somewhat iridescent. Palpi and antennæ dark brown. Fore wing with small pale spots at middle and end of cell, and an indistinct, sometimes broken, whitish fascia beyond. 15 mm. (platanella Chambers).

Caterpillar on Platanus, in a roll made of the down on the under side of the leaf. Western Pennsylvania; Kentucky. New York: Ithaca.

G. fluvialella has been determined as the same as this species hut it appears quite distinct.

15. G. inquilinella Busck. Palpus with second segment white, dusted with black; third, blackish fuscous, somewhat dusted with white. Antennæ blue black, with white dots on segments below. Head dark gray, with black-tipped vestiture. Face light steel gray; thorax blackish fuscous. Fore wing whitish, heavily dusted and suffused with blackish fuscous, appearing dark gray. Discal dots obscure, normal, black. Fringe light gray. Hind wing wider than fore wing, light shining fuscous with paler and yellower fringe; with R and M_1 short-stalked and M_2 and Cu_1 14-15 mm. connate.

Larva inquiline in brassicoides gall on Salix; moth in the spring. Karner, New York.

16. G. unctulella Zeller. Evenly fuscous, including thorax. Orbicular and reniform blackish, not prominent; the other dots very faint; a row of black basal scales in fringe. Head concolorous. Second segment of palpus deep black, with out a strong tuft. 15-18 mm. Caterpillar webbing leaves of locust (Robinia), sometimes in injurious num-bers. Gray; dorsum finely striped with dull red; head and cervical dull yellow;

cocoon between two or three leaves; pupa chestnut brown, very broad and flat.

North Carolina; Colorado; Texas; Arizona; and probably more generally distributed.

17. G. lynceella Zeller. White, dusted with blackish; with parallel oblique blackish shades; 1, along base of inner margin, 2, usually obscure, from near base of costa to inner margin, closely parallel to 1; 3, across middle of wing, strong; 4, more or less obscure, half way across wing two-thirds way out, then zigzag to inner margin. A chevron of black dusting over apex. Fringe pale, slightly dark dusted; head, thorax, and palpi powdery, nearly concolorous. 15 mm.

May.

Texas; Cincinnati, Ohio.

18 G. bicostomaculella Chambers. Ground dark gray with a purple luster, formed of heavy black dusting on a pale blue-gray ground. Palpus with second and third segments dark, powdery, and with a few pale scales, leaving two darker bands on second segment. Head, whitish; a few dark scales on face and many on vertex; antennæ dark. Thorax gray, evenly powdery, with only the usual basal hair on metathorax yellow. Fore wing with small, fugitive, white tufts, with some yellow scales, especially near the tufts. Most distinct marks of fore wing black spots near middle and at end of cell, a bar from the first to the costa, with a spot in the fold below it, and a pale postmedial fascia, all formed of the powdering and evanescent under a lens. Hind wing with R and M_1 stalked, M_s hardly connate with Cu_i; male with a strong hair pencil near inner margin. 14 mm. (Adrasteia quercifoliella Chambers.)

Caterpillar pale yellowish; head and thorax dark brown, with shining cervical shield; in a web on either side of leaves of black oak; skeletonizing the leaf around its web.

I have seen authentic material only from Missouri. It is doubtless general. "New York" (Beutenmüller).

19. G. maculimarginella Chambers. Closely similar to G. bicostomaculelle. Head usually more dominantly white; scutellum with a contrasting yellowish white tuft; fore wing more crisply powdered, usually with postmedial line represented by prominent white spots on costa and inner margin. The yellow on fore wings more distinct. 13 mm.

Caterpillar with head, feet, and cervical shield polished black; pro- and mesothorax paler, white in front; abdomen with four purple stripes posteriorly on segments, joining a purple transverse band in front. Two or more broods; on oak; the first brood feeding in the buds, and emerging in May, the second between spun leaves in June, emerging in July.

Distribution general. New York: Buffalo (Wild), Ithaca. 20. G. vernella Murtfeldt. Violet-gray, more or less dusted with blackish; each scale of ground violet with black tip. Palpus with second segment yellowish with a strong yellow brush, and third blackish, contrasting. Thorax with small yellow posterior tufts. Fore wing with a strong black antemedial patch over cell and fold, with yellow dots before and beyond it, connected by an oblique black bar to costa. Outer part of wing blackish, with contrasting but diffuse white postmedial costal and dorsal spots; extreme margin and fringe powdery gray like base. First dorsal dot black, more intense than the patch in which it lies. No tufts. Hind wing paler, with a weak pencil near inner margin. Easily distinguished from the other species of this group by the yellow palpus. R and M, of hind wing separate. 15 mm. (formosella Murtfeldt. not Hübner).

Larva on laurel oak, in May, rolling the leaves. Head and cervical shield black; body gray with six or eight purplish stripes; meso- and metathorax choco-

late, their anterior two-fifths white. Pupa in a folded leaf. Moth in July. This species is perhaps nearest G. gilvomaculella, which has been confused with it. It has been bred in New Jersey and Missouri and emerges in June.

New York: Ithaca.

21. G. dyariella Busck. Palpus jet black on outer side. Fore wing fuscous, somewhat mottled; head and thorax concolorous. Fore wing marked with black as follows: a dot or small streak on base of R and of A, a noticeably irregular, black bar running obliquely out from costa a fourth way out, to fold, connecting with the orbicular and claviform dots, which form extensions of its distal side, a black bar at end of cell, connected to a vertical bar resting on the anal angle, and black dots in cell before this. A fuscous subterminal fascia. Hind wing paler toward base. 18 mm.

June; larva on poplar.

Western Pennsylvania to Colorado.

23.¹⁹ G. nigrimaculella Busck (Chambers ms.). Fuscous, sprinkled with dark brown, black, and whitish; palpi with a well developed brush; head and thorax largely brown, the face not pale. Fore wing with costal edge black, whitish below; two short oblique dashes on disc; large black costal and dorsal spots at beginning of fringe, with an obscure pale angulate fascia beyond. Abdomen white in middle below. 13 mm.

July to August. Closely similar to G. vernella; most easily distinguished by the dark face.

Southern Massachusetts to western Pennsylvania. New York: Ithaca.

24. G. gilvomaculella Clemens. Ground fuscous, somewhat bluish, and a little powdery, but less than in G. nigrimaculella. Head concolorous; face pale below; second segment of palpus with large brush, blackish, lightly dusted with pale vellow; third slender and mostly black. Fore wing somewhat shaded with yellowish, especially on costa; the costa with broad obscure blackish bars, and a pale

19 Number 22 is vacant.

streak from beginning of fringe nearly meeting one from inner margin. A large blackish spot in cell and a smaller one before it in fold, both more or less flanked with yellow scales and minute tufts. Second discal dot much smaller, inconspicuous. Abdomen mottled below. 18 mm. (biminimaculella Chambers).

Caterpillar on oak. Moth in July.

Penneylvania (?); Missouri; Texas. New York: East Aurora, Ithaca. 25. G. pseudoacaciella Chambers. Dark purplish brown, somewhat powdery, flecked and streaked with ochreous and white; head, antennæ and palpi mixed brownish and white. Fore wing with a pale subcostal streak from base to apex, sometimes obscure, a pale spot at beginning of costal fringe and a dorsal one opposite, sometimes joined into an obscure fascia. First three segments of abdomen dorsally with finely toothed contrasting yellow scales. 18 mm. (cæcella Zeller).

The markings are strongly variable but the species may be distinguished from most Gelechiidæ by the yellow base of the abdomen (which occurs more often in Gnorimoschema than Gelechia). Often the costal region is shaded or suffused with brown, and the antemedial oblique bars may be absent.

Caterpillar on Robinia, often invading mines of other Tineids, especially when young; later between leaflets sewed together. Green with light brown head and cervical shield; abdomen with subdorsal and stigmatal stripes and addorsal and lateral spots; larva, when young, black. Moth in May and June, and August.

Western Pennsylvania; southern Ohio; Illinois.

26. G. serotinella Busck. Very similar to G. pseudoacaciella and not always distinguishable in the imago, but usually with the costa practically concolorous, or brown; never whitish; less streaked looking. 16-21 mm.

Larva on Prunus serotina, in a nest formed of a folded leaf, the caterpillar feeding in a chamber roofed with a heavy sheet of white silk, and living in a black frass tube, much like Catastega. Cocoon within the frass tube. Caterpillar whitish, with brown head and cervical shield, with brown narrow addorsal lines on body, and four broader lines on sides; when young greenish white with black head and cervical shield.

Moth in July and August. District of Columbia; Virginia; Illinois; Missouri; Colorado. 27. G. fondella Busck. Powdery light fuscous. Antennæ annulate black and ochreous; palpus sprinkled with black; head and thorax concolorous; fore wing with each scale darkening to tīp, faintly roseate; two conspicuous black spots trisecting the costa, and converging below, the outer strongly oblique, and covering orbigular as well as resificant. orbicular as well as reniform. Extreme apex dusted with black, not contrasting. Hind wing light ochreous fuscous. Legs pale, with black bars on outer side. Sometimes with the orbicular and reniform separate from the postmedial bar, but the orbicular always very large and touching the reniform. 13-14 mm.

June.

Maryland to western Pennsylvania.

28. G. pseudofondella Busck. Dull ochrcous, slightly dusted with black. Palpi with slight black mottling and bar near tip; head pale; fore wing with black dots on edges at extreme base and dot in fold near base; an outwardly oblique antemedial fascia from costa to fold; a more diffuse postmedial fascia reaching inner margin and covering the reniform; and a small blackish orbicular spot between them. Apex heavily shaded with blackish. 14 mm.

June to July.

Northern New Jersey to North Carolina and Illinois. New York: Ithaca.

29. G. mediofuscella Clemens. Clay color, dusted in varying proportion with dull black; antennæ annulate with white and blackish; palpi dominantly dark. Fore wing with base dominantly of the ground color, ending at a black oblique fascia from costa to fold, defined on inner side. Outer part mottled and dusted with black, gradually shading into the black fascia; usually without definite markings.

This species is rather similar to some forms of the bicostomaculella group, but with no blue tint, and with the hair pencil very weak. In dark specimens the yellow base may be almost obliterated; in light ones it is continued by a band along the inner margin to beyond its middle. This is one of our species which come closest to the subgenus Lita, but its hind wings are too brond to be typical. 10-15 mm. (vagella Walker, fuscoochrella Chambers, Lita liturosella Zeller).

Early spring; July. Larva in seeds of Ambrosia trifida.

General in distribution and not rare. New York: Peru, Rock City, (Cattaraugus County), Crosby, Potter Swamp (Yates County).

30. G. walsinghami Dietz. Palpi ash gray; outer side of first and base of second joints fuscous; tuft divided, dusted with fuscous; third joint dusted with fuscous, with a broad dark ring. Antennæ faintly annulate. Head, body, and fore wings gray; a dark brown posterior spot on thorax. Fore wing marked with dark rich brown; a triangular sub-basal patch, not reaching costa; a longitudinal stripe through middle of apex, defined below and interrupted by a pale postmedial fascia which is defined inwardly and preceded with brown at costa. Some raised white scales in fold. 15 mm. (Pseudochelaria Dietz).

Late May to July. Caterpillar pale green, in a white web on under side of leaves and along petioles of *Rhus typhina* in late August and September. Western Pennsylvania; Anglesea, New Jersey. New York: Ithaca.

31. G. pennsylvanica Dietz. Similar to G. walsinghami. Dark basal area extended outward toward end of cell, where it joins the longitudinal streak, and reaching base of inner margin; the transverse fascia less conspicuous. 15 mm.

July.

Western Pennsylvania.

32. G. tephriasella Chambers. Pale gray, dusted with white; palpi dark brown, second joint white-tipped, third white-dusted, with a white annulus; antennæ light gray and white, with three or four strong annuli toward tip (like Aristotelia absconditella and Duvita conclusella). Fore wing with each gray scale whitetipped; wing darker toward apex; a narrow, white fascia at beginning of fringe, an oblique streak from costa before middle, and obscure darker terminal dots. 10 mm.

Kentucky. Type lot only known.

33. G. nundinella Zeller. Pale buff or cream, with yellow-brown shading and sparse black dusting, tending to form longitudinal striation toward border and oblique shading on disc. Palpus pale with two brown patches each on second and third segments. Hind wing somewhat grayer. 20 mm. (beneficentella Murtfeldt).

Caterpillar on Solanum carolinense, the first brood webbing the terminal leaves into a hollow ball, the second brood in the berries. Pupa in the nest. Larva with head and cervical shield black when young, olive-brown when mature. Body yellowish green with dark glaucous dorsal stripe.

District of Columbia; Texas; Missouri.

As the moth has R and M_1 quite widely separate at origin, though divergent, and the third segment of the palpus is rough, it would probably be better considered a Gnorimoschema or primitive Phthorimoea without pencil, but is usually placed here. The pupa also is unlike other true Gelechias, though far from the other Gnorimoschemas and Phthorimœas studied.

34. G. arenella Forbes. Pale clay-color, the veins perceptibly paler, with some scattered black scales; black discal dots and a dot in the fold; black terminal dots. 20 mm.

May, June, and August.

Massachusetts, on the coast. New York: Rockaway Beach.

The species is not close to Gelechia petasitis, of Europe, with which it has been confused

35. G. branella Busck. Dull dirty ochreous, a little uneven; face paler; antennæ paler, annulate with black; palpi fuscous, the inner side and apex of second seg-

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ment pale. Fore wing with extreme base of costa black, and some scales on inner margin; a black dot near base; two discal dots, and a contrasting dot in fold before the first one. A series of diffuse black terminal dots. 13 mm. This moth is superficially much like Aristotelia quinquepunctella and some Tineas. G. (Teleia) sequax of Europe was reported by Zeller from Massachusetts. It

may be distinguished by the hind wing narrower than the fore wing and narrower than its own fringe, and by the black dash in the apex.

The following nominal species are unknown to me. Many of them have not been recognized since their description, and a fair proportion are likely not to be true Gelechias.

- G. ambrosiæella Chambers. Cincinnati Quart. Journ. Sci., vol. 2, p. 239. 1875. Kentucky.
- G. angustipennella Clemens. Proc. Ent. Soc., Philadelphia, vol. 2, p. 119. 1864. Pennsylvania?

- G. aurimaculella Chambers. Canad. Ent., vol. 4, p. 172. 1872. Kentucky.
 G. badiomaculella Chambers. Canad. Ent., vol. 4, p. 192. 1872. Kentucky.
 G. brackenridgella Busck (detersella Clemens, not Zeller). Proc. Acad. Nat. Sci., Philadelphia, 1860, p. 164. Pennsylvania.
 G. brumella Clemens. Proc. Ent. Soc., Philadelphia, vol. 2, p. 416. 1864. Labrador.
- G. capiteochrella Chambers. Cincinnati Quart. Journ. Sci., vol. 2, p. 252. 1875.
- G. caryævorella Packard. Rept. U. S. Dept. Agr., 1885, p. 331. 1885. Rhode Island.
- G. cassella Walker. List Lepid. Ins. Brit. Mus., part 29, p. 594. 1864. North America.
- G. discoanulella Chambers. Cincinnati Quart. Journ. Sci., vol. 2, p. 254. 1875.
- G. flavicorporella Walsingham. Trans. Amer. Ent. Soc., vol. 10, p. 177. 1882-3. Massachusetts.
- G. flexurella Clemens. Proc. Acad. Nat. Sci., Philadelphia, 1860, p. 163. Pennsylvania.

- G. fuscoluteella Chambers. Canad. Ent., vol. 4, p. 106, 147. 1872. Kentucky. G. fuscomaculella Chambers. Canad. Ent., vol. 4, p. 170. 1872. Kentucky. G. fuscopunctella Clemens. Proc. Ent. Soc., Philadelphia, vol. 2, p. 12, 121. 1864. Pennsylvania?
- G. grissaella Chambers. Canad. Ent., vol. 4, p. 88. Canada. (grisseella Chamb. Bul. U. S. Geol. and Geog. Surv. Terr., vol. 4, p. 144. 1878.)
 G. grisella Chambers. Canad. Ent., vol. 4, p. 171. 1872. Kentucky.

- G. immaculella Kearfott. (Not published?) G. labradorica Mæschler. Canad. Ent., vol. 4, p. 125. 1872. Labrador. G. liturella Walker. List Lepid. Ins. Brit. Mus., part 29, p. 591. 1864. Nova Scotia.
- G. milleriella Chambers. Cincinnati Quart. Journ. Sci., vol. 2, p. 253. 1875.
- G. mimella Clemens. Proc. Acad. Nat. Sci., Philadelphia, 1860. p. 163. Pennsylvania?

- G. obscurella Chambers. Canad. Ent., vol. 4, p. 170. 1872. Kentucky.
 G. obscurusella Chambers. Canad. Ent., vol. 4, p. 106, 128, 148. 1872. Kentucky.
 G. ornatifimbriella Clemens. Proc. Ent. Soc., Philadelphia, vol. 2, p. 420. 186 1864. Illinois.
- G. palpialbella Chambers. Cincinnati Quart. Journ. Sci., vol. 2, p. 253. 1875.

- G. pullifimbriella Clemens. Proc. Ent. Soc., Philadelphia, vol. 2, p. 120. 1864. Pennsylvania?
- G. punctiferella Clemens. Proc. Ent. Soc., Philadelphia, vol. 2, p. 119. 1864. Pennsylvania?
- G. rileyella Chambers. Canad. Ent., vol. 4, p. 106, 147. 1872. Kentucky.
- G. simpliciella Chambers. Cincinnati Quart. Journ. Sci., vol. 2, p. 238. 1875. Kentucky.

- G. suffusella Chambers. Canad. Ent., vol. 4, p. 171. 1872. Kentucky. G. thoraceochrella Chambers. Canad. Ent., vol. 4, p. 169. 1872. Kentucky. G. unistrigella Chambers. Canad. Ent., vol. 5, p. 176. 1873. Kentucky. G. versicolorella Chambers. Canad. Ent., vol. 4, p. 127, 147. 1872. Kentucky.

3. GNORIMOSCHEMA Busck

(Lita: Gelechia, in part)

Closely similar to Gelechia. Third joint of palpus thickened with rough scales except at extreme tip; second joint with distinct divided brush, variable in size. Fore wing as in Gelechia, a considerable proportion of the scales usually finely tipped with white. Hind wing (figs. 159, 160) with acute subfalcate apex, long-drawn-out in the smaller species. **R** and \mathbf{M}_1 well separated at base, more than half as far apart as at margin, but divergent. No hair pencil. This is a medium-sized genus, composed of two well-distinguished groups. In the large typical forms the hind wing is only subfalcate and is broader than its fringe; their larvæ are gall-makers in various Compositæ, cutting an exit hole just

before pupation and pupating in the gall. The smaller group have narrow, long-drawn-out hind wings, with a fringe broader than the membrane. The biology of this group is none too well known, but a couple of species, at least, feed on Solan-aceæ like the Phthorimœas. Only members of the second group are known in Europe, where they are not generally separated from Lita. The species run very close, and the key will not be wholly trustworthy.

Key to the species

- 1. Scales practically all dark-tipped.
 - 2. Fore wing almost evenly black, with a few scattered, white-tipped scales and
- - postmedial line and fringe in G. banksiella).
 - 2. Fore wing with inner fourth distinctly paler than costal fourth.
 - 3. Head and palpi whitish (except outer half of third segment of palpus).
 - 3. Head and palpi rather dark powdery gray, about like the gray parts of
 - 2. Costal and dorsal parts concolorous; mostly smaller species.
 - 3. Fore wing heavily streaked with clay-color, leaving a broad, longitudinal dark streak from base to apex.....ll. henshawiella. 3. No dark longitudinal streak.
 - 4. Fore wing with three ocellate, brown, dark-centered discal spots.

10. triocellella.

4. With obscure ocellate spots or none.

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- 5. A heavy, blackish, oblique bar from costa near base, crossed by a more or less distinct dark streak running from middle of costa toward base of fold.....12. lavernella.
- 5. No heavy, black, oblique bar in antemedial region.
 - 6. Expanse about 18-25 mm. Obscurely streaked longitudinally with 6. Expanse about 12 mm., or ground brown and not streaked.

 - 7. With yellow longitudinal streaks between veins......13. axenopis. 7. Yellow longitudinal streaks absent or rudimentary (about as wide as a scale).
 - 8. Ground white, lightly dusted with black-barred scales; antennæ with white annulations broader than dark ones.
 - 9. Slightly yellowish, with very irregular powdering.

14. batanella

- 9. No yellow shade; with even powdering; larger than G. bata-nella
- 8. Fore wing not mostly white; white annulations on antennæ narrow.
 - 9. Fore wing, thorax, head, and palpi suffused with pink.

9. brackenridgella.

- 9. Not suffused with pink.
 - 10. Black-dusted on white; each scale with a heavy black bar and only a few showing the white tip....8. busckiella.
 - 10. A large, irregular, evenly colored, chestnut-brown area, contrasting with the black ground 17. banksiella.
 - 10. Broadly shaded with brown and powdery blackish; with no contrasts at all and no sharp boundaries.

7. subterranea.

10. Nearly unicolorous fuscous (not seen).

16. scutellariæella.

10. Evenly powdered dull fuscous with four black dots on discchenopodiella.

1. G. detersella Clemens. Head grayish fuscous; palpi pale yellowish white, with two fuscous patches on second segment; a very narrow ring near base and a broad one near tip of third segment. Antennæ annulate, lighter and darker fuscous, without any white. Fore wing grayish, each scale outwardly fuscous, without any white tips; obscure darker dots at middle and end of cell and one in fold. Fringes and hind wing pale ochreous gray; fringes of hind wing yellower. 11 mm. (Not seen).

Pennsylvania.

2. G. gallædiplopappi Fyles. Deep Indian red, including thorax; unspotted, but with a divided pale fascia near hind margin. Head white; upper part of palpus brown, with a white ring on tip of second segment. 22 mm.

The caterpillar forms a gall well up in the main stem of Diplopappus umbellatus. The pupa is suspended in the gall, but the exit hole is not plugged nor the gall lined with silk. (This may be merely a stained specimen of G. gallæasteriella.)

Canada.

3. G. pallidochrella Chambers. Pale ochreous all over; palpus with two dark rings on third segment; antennæ annulate at tip only, brown; fore wing sparsely dusted with fuscous, and with an oblique line across base of wing; a second line a fourth way out; apex fuscous. Hind wing duller. 19 mm. (Not seen.)

Kentucky.

4. G. gallæasteriella Kellicott. White, spotted with brown and blackish, and lightly dusted with black; with only the middle of the costal half dominantly dark, as a rule; a large central costa patch running down to a small bar in fold. Palpi white, second segment with a little gray, third with outer half except extreme tip black. 22 mm. (asterella, in error).

Caterpillar in a more or less triangular gall on top of dwarfed stems of Solidago latifolia, cæsia, axillaris, and Aster divaricatus, the forms of the gall differing with the plant. Hole cut and plugged before pupation.

Canada to Pennsylvania and Michigan. New York: Ithaca.

5. G. gallæsolidaginis Riley. Powdery grav, including head and palpus; most of the scales pale at base with a heavy black bar and fine white tip. Basal half of third segment of palpus contrastingly pale; fore wing with middle half of wing toward costa heavily shaded with brown or brownish black, contrasting, and with the boundary toward base oblique and fairly definite. Some dark shading also near base of inner margin and toward apex. 22 mm.

Caterpillar in stems of goldenrod, not stopping the growth of the plant, but often slightly dwarfing it; forming a fusiform gall. Pupa in a silk hanmock suspended opposite the emergence-hole, which is plugged. Parasitized caterpillars (*Copidosoma* gelechiæ) grow abnormally large and die before cutting the exit-hole. The moth emerges in late August, and is said to hibernate occasionally. The eggs are normally laid in the fall, and hibernate.

General in distribution and not rare. New York: Vicinity of Buffalo, Ithaca, Big Indian Valley, Albany; Richmond Hill, Long Island.

6. G. salinaris Busck. Closely similar to G. gallæsolidaginis but averaging slightly smaller and broader winged; band on third segment of palpus less definite; fore wing with markings slightly more diffuse and tending a little to longitudinal striation. 20-22 mm.

The moth flies early in September. The larva hatches the same year, but does not begin to form the gall till the following spring. The gall is similar to that of *G. gallæsolidaginis*, but usually nearer the ground; in *Solidago salinaris*.

Coast of Massachusetts.

7. G. subterranea Busck. Body blackish, dusted with white. Antennæ with black, brown, and white; fore wing rich reddish to chocolate brown, irregularly sprinkled with blackish, white-tipped scales; fringe whitish, brown-powdered. Hind wings yellowish fuscous with grayer fringe. Abdomen as usual, with yellow at base. 18 mm.

Early September. Galls small, about 15 mm. long, at the bases of the clustered stems of Aster multiflorus; usually in a cluster.

Boston, Massachusetts.

8. G. busckiella Kearfott. Dull black, dotted all over evenly with the white scale-bases; a few of the scales also narrowly white-tipped. Head, thorax, and palpi also mainly black. Hind wing blackish fuscous. 18 mm.

The caterpillar forms a long, cylindrical gall in the side-branches of Aster patens dwarfed by Thiodia radiatana, and stops the growth of these side-branches beyond the galls. The larva may be found in July and August, the moth in November.

Northern New Jersey.

9. G. brackenridgella Busck. Moth similar to G. subterranea but with the whole wing, including the fringe, brownish, and strongly suffused with pink, and also the head, palpi, and thorax. No definite markings but traces of a longitudinal shade rather below middle of wing outwardly. 20 mm.

Gall quite like G. $gall \alpha solidaginis$, but smaller; in stem of aster. Moth in October. This may be the same as G. septentrionella Fyles.

Magnolia, Massachusetts.

10. G. triocellella Chambers. White, contrastingly shaded and dusted with black; whitish ante- and postmedial white fasciæ, converging toward inner margin, a little diffuse, and defined with black shades. Large black antemedial and discal dots, and a similar dot in fold, ringed with white and buff, and with some buff shading near base. 12 mm.

Colorado; reported by Dietz from New Jersey.

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11. G. henshawiella Busck. Whitish, with scattered fuscous-barred scales which gather into a contrasting dark band from base to apex, a third as wide as the wing or more, leaving the costal side of the streak much wider, and suffused and streaked with brown. A couple of black dashes on the upper edge of this band and one in the fold on its lower edge. Irregular, black terminal dots. Head and thorax pale; palpi with four more or less distinct dark bands. 12 mm.

Eastern specimens are not typical and may represent a new species. Barnes and McDunnough consider this a synonym of G. ochreostrigella.

May and June.

Hampton, New Hampshire; western Pennsylvania; also South Dakota and west. 12. G. lavernella Chambers. Fore wing black-powdered on pure white, shaded and noticeably streaked with yellow-brown; palpus yellowish with two dark rings on third segment. Fore wing mottled with black; a heavy fascia from costa near base obliquely out to fold, where it may enlarge into a patch, and is met by a more or less distinct dark streak from costa near middle to base of fold. The latter often broken, or partly obsolete. Postmedial fascia distinct, defined with dark. 12 mm. (physalivorella Chambers).

Larva in fruit of Physalis.

District of Columbia; Ohio; Michigan; and west.

13. G. axenopis Meyrick. Palpus with second segment powdery gray, third white with two slender, blackish bands. Shoulders tinted with yellow; fore wing powdery gray, with several rusty yellow streaks between the veins, toward apex breaking up into a series of oblique streaks between the veins. 11 mm. (artemisiella Kearfott, not Treitschke).

Meyrick considers this a Phthorimaa. It will almost certainly be congeneric with "Lita" artemisiella of Europe.

Caterpillar webbing together the young terminal leaves of Artemisia canadensis, and eating back into the stem.

Atlantic States.

G. saphirinella Chambers is similar, but with nearly white second segment of palpus, blackish shoulders, and more brilliant orange-red stripes on wing. Eastern records should probably be credited to *G. axenopis*.

14. G. batanella Busck. Antennæ annulate with black and white, the white annulations as wide as the black; and without distinct brown ones. Palpi obscurely annulate. Fore wing white, usually lightly dusted with black; the black gathering into a shade at apical third, or defining an angulate pale postmedial line; four small yellow and light brown spots at middle and end of cell, and middle of inner edge, and two more farther out, all obscure and variable; fringe white, black dotted, and with a broken black line. 12 mm.

June.

New Jersey to western Pennsylvania.

15. G. petrella Busck. Head, palpi, thorax, and wings white; almost evenly and moderately dusted with fuscous, the white dominant. Antennæ white, ringed with black. Discal dots blackish, not contrasting. 17 mm.

May:

Hampton, New Hampshire.

16. G. scutellariæella Chambers. Fore wing dark powdery fuscous, each scale bluish at base and finely tipped with white; apex paler, with ill-defined whitish costal and dorsal streaks; palpi yellowish within. 9 mm.

The caterpillar is white, with a pale yellow head; and lives in a flat, curved case covered with frass, which it leaves to mine in a leaf of the food plant. Apparently, a single case is used, which is permanently attached by the smaller end. Food: Scutellaria.

Only the type is known.

Kentucky.

17. G. banksiella Busck. Blackish. Palpus light gray on dorsum of second segment and extreme apex of third; antennæ narrowly ringed with white. Fore wing with white powdering at apex, and indicating a broken postmedial fascia; disc of wing occupied with several rounded, even, chocolate or yellow-brown spots, which are often confluent into an irregular patch; and a more diffuse band on fold and inner margin. First two segments of abdomen yellow on dorsum (as Moth in July and August. Caterpillar on Solidago.
 Massachusetts to western Pennsylvania. New York: Essex County, Batavia,

East Aurora, Otto, Ithaca, Sea Cliff.

18. G. alaricella Busck. Powdery fuscous gray, the dominant scales with dull luteous base and white tip; intermixed with yellow scales, which tend to gather in numerous obscure longitudinal streaks. Discal dots black, rather elongate, obscurely ocellate with the yellow. Costal edge blackish, contrasting. Palpi powdery gray with a narrow basal and a broader outer ring on third segment. Antennæ annulate. 18-25 mm.

Western Pennsylvania.

4. PHTHORIMEA Meyrick

(Bryotropha; Lita; Gelechia, in part)

This genus is closely similar to Gnorimoschema. The white-tipped scales are perhaps less frequent. The third segment of the palpus averages more smoothly scaled. The male has a heavy pencil of hair at the base of the hind wing above, distorting the venation. The known caterpillars all feed on Solanaceæ and appear to be rather similar in habits. The names "potato tuber moth" and "tobacco splitworm" apply especially to P. operculella, but some of the accounts of injury seem to be based on other species. The species are close and difficult to recognize, P. operculella and glochinella being often indistinguishable in the female.

Key to the species

- 1. Fore wing not black-striped.
 - 2. Strongly mottled with dark brown, with a series of dark spots along costa

- 4. Normally larger, yellow scaling more distinct and forming well marked longitudinal streaks; male with large dorsal and ventral flat lobes to genitalia, covering the valves.....l. operculella.
- 4. Normally smaller; yellow scaling obscure and usually not gathering into longitudinal lines, sometimes practically absent; male with dorsal and ventral lobes small and tapering; valves slender, claw-like,

1. P. operculella Zeller. Powdery gray, streaked between the veins with pale ochreous. Head pale, usually cream white; palpi similar, with a few gray scales. Antennæ grayish, annulate with dull black. 12-16 mm. (Bryotropha solanella Boisduval).

This species breeds continuously in the warmer season in stored potatoes, having many broods a year. The caterpillar is whitish, often with some pink or green shade, with a black head and cervical shield. Typically, when young it is a leaf-

20 See also Gnorimoschema axenopis.

miner; later it enters the large veins of tobacco, or the shoots of potato. Eggs are also sometimes laid on the surface of potatoes if left exposed above ground in the field or in storage. In that case the young larve feed at first just under the skin, and later bore through the potato in all directions. Pupation takes place in a cocoon outside the burrow, as a rule. The larva also works in leaves, stems, and fruits of other Solanacee.

The distribution is general southward.

P. minor Busck is an exactly similar, but much smaller, form, or species, from the far South.

2. P. striatella Murtfeldt. Antenna blackish. Fore wing light wood-brown, streaked with dark brown and with some powdery gray, especially at apex and in fringe. A black streak from base through upper part of cell to apex, and a broader irregular and diffuse one running above inner margin to anal angle. Genitalia much as in the following species. 12 mm.

Larva in herries of Solanum nigrum; pale greenish yellow with five irregular and interrupted crimson stripes, the lateral ones formed of series of spots; head, cervical shield, and true legs proportionately small, shining dark brown.

St. Louis, Missouri; California.

3. P. glochinella Zeller. Closely similar to *P. operculella*. Fore wing pale yellowish gray to the naked eye; under the lens the color made up of a mixture of black-and-white scales and pale ochreous scales, the latter often very few and usually scattered. Head usually powdery light gray, including palpi. Male genitalia with a short, triangular, dorsal plate, a slightly larger, trough-shaped, ventral one, and two slender, curved valves. Usually distinguishable from operculella by the evener coloration, but occasionally indistinguishable in the female. 12-14 mm. (solaniella Chambers, in part; similiella Chambers, in part, not Ptycerata similiella).

Larva green, becoming almost blue when mature; in a dense, silken, frasscovered tube, in a mine in leaves of *Solanum carolinense*. Pupa in a cocoon at surface of ground.

Missouri; Texas.

4. P. marmorella Chambers. Light yellowish fuscous, irregularly spotted and mottled with dark brown. 9 mm.

Types only known.

Kentucky.

5. ANACAMPSIS Curtis

(*Tachyptilia* Heinemann; with *Compsolechia* Meyrick)

Palpi with second segment smooth or slightly roughened above the middle; third longer, slender, and pointed; fore wings long with blunt apex, rounded outer margin, and well-marked anal angle above Cu_2 . Venation as in Gelechia. Hind wing much broader (often nearly twice as broad as fore wing), rounded-trapezoidal, not sinuate, with normal venation; R and M, approximate; a fringe on Cu.

The moths of A. agrimoniella and A. levipedella, at least, walk in a circle on alighting, like the Choreutidae.

Key to the species

1. Fore wings without any markings on basal half; though usually with light outer markings.

2. With a white transverse fascia.

- 3. Two or three longitudinal white apical dashes, sometimes fusing into a white terminal bar......8. tristrigella.
- 3. Without white terminal dashes.

2. No transverse fascia.
3. One white costal streak or none.
4. Palpi yellow
4. Palpi dark brown
3. Two white costal spots9. levipedella.
1. Basal half of fore wing more or less mottled.
2. Crisply dusted and marked with black on white
2. Ground practically even.
3. Postmedial line indicated by a white dot at costa and a yellow one on
inner margin; palpi unicolorous whitish, more or less contrasting
4. crescentifasciella.
3. Postmedial line all one color and continuous or nearly so, often faint.

- 4. Ground ash gray. Larger. Second segment of palpus blackish with

I. Second segment of palpus with a loose tuft above (Anacampsis).

1. A. innocuella Zeller. Ash gray, postmedial line paler, transverse, well out, waved, and iollowed by a blackish shade; three blackish dots in cell and one or two in fold. Palpus somewhat paler, with basal two-thirds of second segment blackish, contrasting. A slight tuft on upper side of second segment. 20-24 mm. Common in July. The caterpillar forms a cylindrical roll of a leaf of poplar.

It usually cuts the petiole early in the last stage, and finishes its feeding in the decaying leaf lying on the ground It may be found in June. It is transparent, the white fat and green food showing through; head black or black-brown, cervical shield brown, with black sides and posterior edge, true legs and tubercles black. Massachusetts to Colorado. New York: Ithaca, Ilion, Pearl River. Cold Spring

Harbor, Long Island.

2. A. niveopulvella Chambers. Palpi blackish, white at tip of second and upper side of base of third segments. Fore wing brownish black, heavily dusted with white, the markings as in A. innocuella but strongly contrasting, formed of the white powdering. 20 mm. Caterpillar on willow. Moth in July and August. Probably a contrasty variety of innocuella.

Quebec to Wisconsin and British Columbia.

II. Second segment of palpus smooth above (Compsolechia).

3. A. rhoifructella Clemens. Grayish fuscous, marked as in A. innocuella, the dark discal spots variable in size and often a much warmer yellow-brown. Palpi warmer yellow-brown, paler toward tip, but without any sharp contrast. 15-18 mm. (consonella Zeller, ochreocostella Chambers, quadrimaculella Chambers 1874, not 1875).

General from May to August. Caterpillar in the spring in the fruit-spikes of sumach (*Rhus typhina*), feeding on the crimson hairs and exterior coat of the drupes, living in a silken gallery within the cluster, and leaving strings of frass outside. Cocoon in a slight silken web near the surface, in the frass. Caterpillar immaculate, of various shades of brown with darker head and tubercles, and blackish cervical shield. There is a species apparently the same as this on Viburnum.

New York: West Farms (the Viburnum species).

4. A. crescentifasciella Chambers. Ash gray, finely dusted with brown; a very indistinct, pale, crescentic fascia at beginning of fringe, strongly concave toward base, and rarely absent. One or two minute dark spots in disc and one at apex. 12 mm.

April and May. Caterpillar on Krameria. This species has been confused with Duvita conclusella.

Kentucky; Texas.

5. A. nonstrigella Busck. Antennæ black, annulate with silvery white; palpi bright deep ochre, the joints black-shaded at tip; eyes dark red; face white, iridescent, shading into the dark olive brown, iridescent vertex and thorax. Basal two-thirds of fore wing dark olive, apex blackish with a few golden brown scales. Hind wing dark purplish fuscous; fringe of inner margin white. Abdomen yellow below, black and white annulate above. 15 mm.

July and August.

Northern New Jersey; western Pennsylvania. 6. A. lupinella Busck. Brown-black with a straight, dull white postmedial fascia, sometimes interrupted or nearly obsolete. Wing sometimes streaked with brown; palpi brown, shaded with black. 12 mm.

June and July. Caterpillar dull green; head testaceous with a black spot on each side near eyes, cervical shield of the same color, partly edged below with black; tubercles black; in a slight web between leaves of lupine.

7. A. agrimoniella Clemens. Similar to A. lupinella, the basal half of the wing

powdery ash gray; fascia apparently always complete, and palpi pale. 12 mm. June and August. Caterpillar between leaves of agrimony; pink when young, later dull green with black tubercles and pale brown head and cervical shield.

New York to Illinois and Georgia. New York: Otto, Taughannock Falls, Sea Cliff, Long Island.

8. A. tristrigella Walsingham. Closely similar to A. agrimoniella with two or three short, thick, longitudinal white dashes at outer margin, sometimes confluent in a short bar.

Caterpillar on Corylus americana.

Connecticut; eastern Pennsylvania; Manitoba.

9. A. levipedella Clemens. Antennæ dark brownish; head whitish beneath; palpus with a brown exterior spot on third segment. Fore wing dark brownish, coppery, paler toward base; an oblique white costal patch at middle with a longitudinal dash in fold opposite, and a second more triangular costal streak threefourths way out, with two longitudinal dashes below it. Fringe with violet iridescence. Below, with white costo-apical dots on all wings. 10 mm. (Strobisia Clemens).

June.

Pennsylvania; District of Columbia.

6. UNTOMIA Busck

Palpi long, second segment slightly thickened, truncate at tip, third smooth, longer. Fore wing with R, and R, united, M, and Cu, stalked. Hind wing about as wide, slightly notched below apex; R and M, short stalked; M, and Cu, connate; cell open.

1. U. albistrigella Chambers. Blackish brown, slightly bronzy; a small, oblique, white costal streak before apex, running into fringe, and a few white scales at apex, near dorsal fringe. Fringe pale fuscous with a dark line. Hind wing rather darker. 8 mm. (Gelechia Chambers).

June.

Western Pennsylvania; Kentucky.

7. ANARSIA Zeller

Palpus of female about like Dichomeris, with third segment hardly as long; in male, with a more or less expansible ridge of long, loose hair on inner side of second segment as well as the triangular tuft of Dichomeris, and third segment

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rudimentary, concealed in the vestiture of the second. Fore wing normal; Cu, and Cu_1 remote, M_1 out of base of stalk of R_4 and R_5 ; hind wing trapezoidal, normal, R and M₁ short stalked. The male on account of its rudimentary third joint of the palpus might run to the Phaloniid Tortricids, but is distinguished by the neater triangular palpal tuft, and stalked R, and R5.

1. A. lineatella Zeller (Peach bud moth). White, dusted and mottled with gray; palpal tuft blackish on outer side. A couple of black spots on costa of fore wing, and sometimes blackish streaks on disc on R, Cu, fold, and toward tip of wing. 10-15 mm. Female larger (pruniella Chambers).

Caterpillar chestnut brown with white incisures, head, cervical shield, and feet black, anal plate black and whitish. It works in the buds and tips of the young

twigs of peach and plum in the spring, killing the twigs; the later broods bore in the fruit and eat the stone. (H p. 426 f. 245-6.) General in distribution; injurious southward, and in many foreign countries; probably introduced from the Old World, where there are several close relatives. New York: Rochester, Lockport, Clifton, Jordan Station, Ithaca, Schoharie.

8. TRICHOTAPHE Clemens

(Begoë Chambers; Sagaritis Chambers [?]; Dichomeris; Brachmia; Nothris, in part)

Like Dichomeris, but with second joint of palpus at most only moderately thickened, and usually more strongly so above than below; without a triangular tuft: the third segment apparently arising from its end; palpus sometimes evenly curved as in Anacampsis; more often with the third joint set on at an angle. A slight variant of Dichomeris (fig. 157).

Key to the species 1. Ground color nearly black. 2. Head and costal edge cream color, contrasting. 3. Costal cream-colored band sending a curved pointed process into the dark dorsal regionl. flavocostella. 2. Costal edge not light. 3. With strong iridescence. 4. A light ochreous dot at apical third of costa......4. alacella. 4. No such dot. 3. Not iridescent. 4. Palpi yellow. 5. Three prominent yellow dots on fore wing......8. trinotella. 2. No heavy black markings on disc. 3. Fore wing overlaid with dark scales. 4. An obscure dark and pale postmedial fascia, no longitudinal streaking, 4. No postmedial fascia, powdering gathering in streaks on veins, terminal line continuous, at least toward costa.....10. chambersella. 3. Fore wing at most lightly sprinkled with dark scales.

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 With dark spot on fold......12. trimaculella.
 No dark spot on fold; head whitish......13. fernaldella. 2. A heavy black V-mark on disc.....14. setosella.

1. T. flavocostella Clemens. Palpi with second segment loosely hairy on upper side (Begoë Chambers). Head and palpi light ochre; tegulæ cream white, fore wing purple-black, with costa cream white almost to apex, narrowing a little at middle of wing, and broadening and sending a long spur toward anal angle at end of cell. 15-18 mm.

June and July. Caterpillar on Solidago and Helianthus. Maryland to Georgia and Missouri. New York: Ilion, Big Indian Valley, Poughkeepsie, New Windsor, New York City.

2. T. inserrata Walsingham. Very close to the last species and perhaps a form of it, but with the costal cream colored stripe narrowing more abruptly at middle of wing. and widening roundly over end of cell without any spur. 15 mm.

May to July; Caterpillar in immature fruit of Solidago; olive green.

District of Columbia: Missouri, and elsewhere.

3. T. serrativittella Zeller. Similar to T. flavocostella, the costal cream stripe fully as broad, even in width almost to apex. except for two minute teeth, and then abruptly running to a point just before apex. 10-15 mm. (plutella Chambers).

May; July to September.

New Jersey to Florida, Colorado, and Texas. 4. T. alacella Clemens. Chocolate brown; palpi and face bright yellow. Fore wing with terminal margin metallic lead colored or greenish. Costa also lead colored out to a cream spot before the apex, the area extending down into cell in a tooth at a third way out, narrowed beyond the tooth, and then with a second process at end of cell running across wing to inner margin. Vertex and tegulæ dark. 15 mm. (ochripalpella Zeller; goodellella Chambers).

July.

Massachusetts to District of Columbia; west to Missouri. New York: Ithaca; Sea Cliff, Long Island.

5. T. purpureofusca Walsingham. Purplish black (becoming brownish when rubbed); face paler. steel gray; palpi and tongue bright yellow; immaculate. Antennæ normal. 18 mm.

June and July.

Ottawa. Ontario. to Wyoming County, Pennsylvania; Manitoba.

6. T. nonstrigella Chambers. Antennæ verv denselv clothed on basal half with overlapping scales, as in Dasycera. Nearly black: a little iridescent; lower part of face and palpi bright vellow; outer side of third segment blackish. Hind wings fuscous as usual but with narrow white costal edge.

June, one brood. Caterpillar spinning together terminal leaves of Aster shortii in late April and early May; head black. cervical shield blackish, mesothorax and front of metathorax plum-color: rest of body gravish white with seven plumcolored stripes as wide as the spaces between them. Dorsal tubercles of thorax enlarged, black; legs black.

Western Pennsylvania to Kentucky.

7. T. juncidella Clemens. Dull purple-brown, with obscure darker postmedial fascia and spot in fold near base. Discal dots (reniform, orbicular, claviform) black, sometimes very lightly defined with vellow, and with a few vellow scales on costa before apex. Palpi and lower part of face contrasting, yellow. 15 mm. (dubitella Chambers).

June to September. Apparently two broods. Caterpillar a leaf-roller on Soli-dago, Aster, Artemisia, and other composites. Head and cervical shield polished black; mesothorax velvety black, white on posterior half, with five linear black marks; rest of body white and obscurely striped, tubercles small, black.

Canada and Maine to Virginia and Kansas. New York: Ithaca, Rhinebeck. T. leuconotella Busck, a slightly paler species with a single conspicuous, not ocellate, canary-yellow dot at the end of the cell, and a faint light brown costoapical spot, has been determined by Busck from East Aurora, New York. It was described from Washington State.

8. T. trinotella Busck. Antennæ dark purple, palpi light ochreous, face ochreous, thorax and fore wing brown, prominent ochreous dots at middle of wing, end of cell, and in fold; a few ochreous scales at beginning of costal fringe. 15 mm.

May. Caterpillar, apparently of this species, on Enothera in Missouri.

Western Pennsylvania.

9. T. levisella Fyles. Palpi fuscous with third segment paler. Fore wing clouded brown, a little lighter than T. *juncidella*, with a darker postmedial band tending to break into spots; and an obscure, pale brown, horse-shoe mark at end of cell. 15-22 mm.

June and August. Caterpillar folding and crumbling basal leaves of Aster macrophyllus in June and cating the parenchyma. Pale green, with darker dorsal, subdorsal, and lateral lines; head and cervical shield glossy black; mesothorax brown in front. behind with four conspicuous white patches; tubercles, spiracles and leg-plates black. Cocoon capsule-like, open at one end.

Canada; New York.

T. washingtoniella Busck, described from Ambrosia artemisia folia, and reported from the District of Columbia to Missouri, appears to be the same species.

10. T. chambersella Murtfeldt. Palpus nearly three times as long as head, second segment smooth-scaled above. Outer margin more oblique than usual, nearly straight. Fore wing clay-color dusted with light fuscous; head, thorax, and palpi concolorous. Fore wing with dusting gathering in blackish streaks between the veins; a blackish bar or streak in the cell beyond the middle, and a spot at the end of the cell sometimes also drawn out into a streak. Terminal line black, continuous; fringe whitish at base. Hind wing pale gray with whitish fringe. 9-13 mm. (*inæquepulvella* Chambers). End of May to September (2 broods?). Caterpillar in a fusiform case, on

End of May to September (2 broods?). Caterpillar in a fusiform case, on *Ambrosia artemisiæfolia*, webbing together the divisions of the leaf, and eating the upper parenchyma from the inside. Head polished, dark brown; cervical shield small, yellowish; prothorax otherwise dark brown; rest of thorax and third and fourth segments of abdomen brown-black with a white fold on the posterior edge of each segment; first two segments of abdomen wholly black; the rest with an oblique anterior lateral band on each segment. Caterpillar making several nests, and pupating in the last one.

Western Pennsylvania; Kentucky; Missouri; and south.

11. T. inversella Zeller. Second segment of palpus with a large loose tuft on upper side; dark gray on outer side; third segment very short, with a black ring beyond middle. Fore wing shaded powdery gray, with a diffuse whitish post-medial band, and a longitudinal shade through cell. Slightly diffuse black spots beyond middle and at end of cell, and near base and middle of fold; costal edge blackish at base. 18 mm. (*Epicorthylis* Zeller).

Texas (doubtful northward). There is a related but much smaller species known from Maryland and Virginia in June.

12. T. trimaculella Chambers. Palpus with third segment only half as long as second. Pale yellowish, minutely and sparsely dusted with reddish ochreous; head and palpi nearly white; third segment of palpus tinged with fuscous. Two small, nearly round, blackish spots at middle of wing, obliquely placed in fold and cell, a larger one at end of cell, and a minute terminal series. 12 mm.

April and May. Almost identical with *Dichomeris touceyellus*, except for the smoothish Trichotaphe palpus with short third joint.

District of Columbia to Texas.

LEPIDOPTERA OF NEW YORK AND NEIGHBORING STATES

13. T. fernaldella Busck. Palpus very long, second segment smooth on dorsal side. Cream white, including head and palpi. Fore wing lightly dusted with light dull gray between the veins, which stand out pale. A small blackish spot beyond middle and one at end of cell, sometimes with a few blackish scales in fold below the first. Outer margin strongly oblique. 18 mm.

June. Very similar to T. chambersella but paler and much larger; also easily confused with the Gelechias if the palpi are not noted.
Orono, Maine; Adirondacks; Manitoba. New York: Peru.
14. T. setosella Clemens. Almost exactly similar to Dichomeris eupatoriellus.

Fuscous gray; a costal cream-colored stripe, gradually narrowing to postmedial line, where it runs out. Postmedial line yellow, slightly concave, edged beyond with a blackish shade. A very heavy and widely open V-shaped mark in cell and fold from close to base almost to middle of wing, filled with an extension of the cream-colored costa. 18 mm. (Malacotricha bilobella Zeller).

July and August.

General and not rare. New York: Ithaca; Sea Cliff, Long Island.

81/2. BRACHMIA Hübner

Similar to Trichotaphe. Fore wing with R_3 shortly stalked with R_{4+5} . Palpi long, upcurved, and moderately thickened.

1. B. hystricella Braun. Brownish ochreous or fuscous. Face straw-colored; palpi with two fine longitudinal lines. Antennæ dark, banded beneath with whitish. Fore wing with veins distinctly outlined with pale straw color, costal and inner margins, and a streak in cell also whitish. Ordinary spots black, outlined with white, the one corresponding to the claviform elongate. Costal fringe straw-colored, dorsal fuscous, contrasting. Terminal line blackish, fringe with two dark lines.

Hind wing yellowish white or light gray. 14 mm. Several broods a year; larvæ in rolled leaves of *Hystrix patula*. Larvæ of the last brood wintering in the rolled leaves. Larva with head brownish ochreous, the lateral margins dark reddish brown; prothorax whitish, with a continuation of the lines of the head, part four comparison dark brown the antrior of each protection. of the lines of the head; next four segments dark brown, the anterior edges of the first two whitish; rest of body whitish with a brown, subdorsal line and oblique lateral stripes.

Cincinnati, Ohio.

9. DICHOMERIS Hübner

(*Ypsolophus* auct., not Fabricius; *Nothris* auct., etc.)

Palpus with a prominent triangular tuft on ventral side of second segment, extending beyond the attachment of the third, which is longer than the second without its tufts, and slender. Fore wing long, normal; R_4 and R_5 stalked, Cu_1 and Cu_2 stalked, the other veins free. Hind wing with R and M_1 approximate or shortly stalked; M_3 and Cu_1 connate or stalked; M_2 associated with Cu-stem.

Key to the species

- 1. Fore wing very long and narrow (over four times as long as wide); hind wing bluish, translucent between the veins.....1. ligulella.
- 1. Fore wing usually about three times as long as wide and always less than four; hind wing fully scaled.
 - 2. Fore wing with dark dorsal spot two-thirds way out..........8. citrifoliella. 2. No dark spot two-thirds way out on inner margin.
 - 3. Two large black spots on disc.....11. eupatoriella.
 - 3. No large black spot on disc.

4. Margins concolorous with the ground.

5. Usual discal dots distinct (reniform and claviform at least).

- 6. These dots not light brown.
 - 7. Head and outer and inner margins blackish, more or less con-
 - 7. Head not contrastingly blackish.
 - 8. Ground red-brown with coarse paler (pinkish) powdering. 6. trinotella.
 - 8. Ground not red-brown.
 - 9. Expanse about 12 mm.
 - 10. Ground rather light brown with blackish dusting; black terminal bars........... (Anorthosia punctipennella). 10. Ground blackish brown.
 - - 11. Scattered black scales only in fringe.....3. touccyella.

11. Regular blackish bars in base of fringe....4. hirculella. 9. Expanse about 20 mm.

10. More or less distinct longitudinal streaks5. bipunctella.
10. Not streaked10. ventrella, georgiella.
5. No distinct discal spots9. caryæfoliella.
4. Margins pale, contrasting12. marginella.

1. D. ligulella Hübner. Fore wing about four and a half times as long as wide; hind wing notably shorter, with disc translucent; bluish between veins, the scales reduced and not meeting each other. Fore wing normally fuscous brown, with discal dots and postmedial band normally somewhat darker, but sometimes obsolete. Hind wing bluish, with fusçous brown veins. 15 mm.

This is var. pometella Harris. In the typical form the costa is broadly creamwhite, contrasting with the ground color. (The names Chætochilus contubernatellus Fitch, pauciguttellus and flavivittellus Clemens, and recdella, ruderella, and quercipomonella Chambers, mostly represent variant forms.)

Common and generally distributed; often in injurious numbers on apple (the Palmer-worm) but sometimes rare for years together. Caterpillar green, less often brown, head yellow-brown, black-brown in dark specimens; cervical shield concolorous with head, often heavily marked with black in dark specimens. Tubercles black; body with a narrow dorsal and very broad subdorsal diffuse dark bands, separated by whitish stripes. A leaf-roller on apple and oak.

New York: Genesee County; Geneva, Rock City (Cattaraugus County), Ithaca, Nassau, Byron, Albany.

D. malifoliella Fitch, of which no material is known to exist, is probably a form of ligulella. It was described from New York.

2. D. punctidiscella Clemens. Obscure fuscous, varying in shade, with four broken brown transverse bands, the second formed by the rounded orbicular and claviform spots, the third including the discal bar. Hind wing fuscous, very slightly translucent between the veins. Wing about three and a half times as long as wide. Second segment of palpus black on outer side, finely edged with white; third pale ochreous; antennæ annulate with pale yellowish brown and deep brown; head light yellowish. 18 mm.

End of May to June.

Parry Sound, Ontario, to District of Columbia and Ohio. New York: Taughannock Falls (Ithaca).

3. D. toucceyella Busck. Palpi as in D. punctidiscellus; head and thorax pale ochreous, densely dark-dusted. Fore wing ochreous, dusted and suffused with brown; a small dark brown spot on fold before middle, and larger ones at middle and end of cell. Some scattered dark scales at base of fringe. 12 mm. (Anarsia trimaculella Chambers, not Clemens.)

Kentucky; Texas.

4. D. hirculella Busck. Wing form as in touceyella; hind wing fully scaled. Palpus with second segment blackish on outer side, third ochreous. Fore wing ochreous, heavily dusted and mottled with black scales,- in effect, blackish fuscous; and with blackish spots in cell, at end of cell, and sometimes a postmedial series of smaller ones. Fringe yellower with blackish bars in base. Hind wing lighter fuscous. 11-12 mm.

East River, Connecticut.

5. D. bipunctella Walsingham. Dull brownish yellow. Discal dots well marked; terminal points each at end of an obscure gravish streak, rarely obscure. Veins sometimes distinctly streaked. 22 mm."

April to July.

Digby, Nova Scotia, to Florida, along the coast. Hazelton, Pennsylvania.

6. D. trinotella Coquillett. Head pinkish brown; palpus with second segment dark brown on outer side; tuft tipped with pale pinkish; third segment dark brown. Antennæ annulate; fore wing reddish brown, inclining to pink; sparsely and coarsely dotted with black; often with three white spots near its center, the outermost spot crossed by a black dash, the other two edged within with black, sometimes with only the black spots. Hind wing dull leaden. 11 mm. (Nothris Coquillett.)

Caterpillar on hazel, in a folded leaf.

Illinois.

7. D. vacciniella Busck. Tuft of palpus dark brown, its apex sprinkled with white; third segment light brown, dusted with black, base white externally. Head blackish. Fore wings wood-brown, shaded with dark brown on outer and inner margins, and sparsely and irregularly strigose with black scales; three small, round, black, white-ringed dots, or a few white scales at middle and end of cell and in fold before the first dot; apex suffused with purple-black, becoming black at margins; fringes dark fuscous, with light ochreous brown tips; thorax concolorous. 15-20 mm.

Larva on cranberry.

Pemberton, New Jersey.

8. D. citrifoliella Chambers. Ochre yellow, densely and evenly dusted with gray. Palpus blackish, with tuft white tipped; third segment yellow. Thorax below, shoulders, and costal edge at base, black. Fore wing with two antemedial dark dots above and below fold; two dots in cell, in position of orbicular, and one dot in fold just beyond them; a pair of blackish discal dots and a stronger blackish shade on inner margin beyond them; postmedial line faint, pale, defined with dark, irregular. Hind wing gray. 18 mm.

June. Larva yellow with black head and cervical shield; a leaf-roller and budworm on citrus and prickly ash; sometimes injurious to orange in the South.

Cincinnati, Ohio, and south.

9. D. caryæfoliella Chambers. Palpi as usual; head and thorax golden or reddish iridescent, suffused with fuscous; fore wing iridescent, silky, suffused, showing golden, red-brown, and lead color; two or three minute dots on disc. Hind wing lead color. 22 mm.

End of June. Caterpillar green with six interrupted longitudinal white stripes. Head ferruginous; prothorax brown; true legs black. Sewing together leaves of hickory in early June. At maturity it turns white, striped and suffused with pink. Missouri; Kentucky; Texas.

10. D. georgiella Walker. (1866). Varying in color from light yellowish to dark purple-brown, dusted with a darker shade, and with scattered black scales. Costal edge pinkish, sometimes very narrowly; outer margin sometimes suffused with pink. With antemedial; median, and discal dots, which sometimes run together into three darker bands, parallel and slightly oblique inward. Orbicular and claviform in the median band; almost always with some white scales; also with some white scales, occasionally wanting, on inner side of discal dot. Fringe yellower; hind wing mouse gray. 17-20 mm.

May to July. Larva a leaf-roller on oak, skeletonizing the leaf. (Described from the same material as type of D. roseocostella Walsingham.)

Distribution apparently general.

D. ventrella Fitch (1854) appears to be the same species, so far as can be told from the description; and in that case would have priority (unicipunctella Clemens).

"New York" (Fitch).

11. D. eupatoriella Chambers. Male with a large expansible tuft, normally hidden behind fore coxa. Dull purplish gray or fuscous; costa whitish and lightly dusted with dull black, somewhat less contrasting than in *Trichotaphe* setosella. Blackish patch on cell and fold of fore wing not quite reaching middle of wing; its front edges with sharp angles near its tip and halfway between there and base of wing, with a right angled emargination between; shorter than in setosella. Postmedial line, whitish, fine, and nearly erect; the wing darkened beyond it to outer margin. 16 mm. (dolabella Zeller, setosella Walsingham, not Clemens.)

May to July; late September. Caterpillar on Eupatorium ageratoides and Vernonia, when on the former plant folding a leaf so that a large side vein lies against New York to Florida and Manitoba. New York: Crosby (Yates County).
12. D. marginella Fabricius, a pale even pinkish brown species, with the fore

wing margined all around with cream color, has been introduced from Europe at Westchester, Pennsylvania, and at Tarrytown and Plandome, New York. Larva on Juniperus communis, social in a web; brown, longitudinally striped. (Nothris auct.)

10. GLYPHIDOCERA Walsingham

(With Sceptea Walsingham)

Similar to Trichotaphe; palpus with second segment not swollen, evenly upturned; fore wing (fig. 161) with R_s and R_s completely united, and often stalked with R_s or M_1 ; Cu_1 and Cu_2 stalked, M_3 occasionally arising from the base of their stalk, typically nearer M_2 . Male antenna in some species with a notch in the shaft near the base, edged with rough scales, and running obliquely across several segments. Obscure ochre yellow species, dusted more or less heavily, with the ordinary dots alone indicated by a gathering of the dusting. G. lactiflosella only is pale yellow without dusting.

Key to the species

1. R_3 , R_{4+5} , and M_1 all free. 2. Antenna strongly notched; ground dull ochreous......2. æquepulvella. 1. R_3 short-stalked with R_{4+5} ; notch rudimentary or absent.

2. M_a free.

3. Antennæ annulate with black; a moderately powdered species.

4. meurickella

1. G. lactiflosella Chambers. Cream color. Thorax with three dark dots behind; palpus simple; second segment dark on outer side, except apex. Fore wing with a dot in base of fold, an antemedial dot in fold, dots opposite each other at middle of wing in cell and fold, sometimes fusing into a bar, and a bar at end of cell, all brown. Terminal line brown, at apex only. Ground lightly dusted with brown. 12 mm. (Trichotaphe).

It flies in Louisiana in June.

Texas; Louisiana; Plummer's Island, Maryland.

2. G. æquepulvella Chambers. Fore wing with all veins well separated except Cu_1 and Cu_2 ; male antennæ with notch cutting nearly halfway through stalk. Light dull ochre, dusted with fuscous in about equal proportions. Palpus with third segment darker; head and thorax pale. 14 mm.

Virginia; Kentucky; Texas; Colorado; and Utah. 3. G. speratella Busck. Fore wing with \mathbf{R}_{4+5} stalked with \mathbf{R}_3 . Antennæ light ochreous, in male with a slight indication of notch. Palpi slightly sprinkled with fuscous externally, face and head light ochreous. Fore wing thickly and evenly sprinkled with fuscous, and with four darker dots (the usual three and one near base). 18 mm.

Western Pennsylvania.

4. G. meyrickella Busck. Palpi ochreous, third segment slightly scaled with black on inner side; antennæ light ochre, annulate with black, simple. Fore wing evenly and profusely dusted with black scales, sometimes mostly fuscous, with the usual discal dots but no terminal dots. Venation as in G. speratella.

June in the South.

Maryland and to Louisiana and Mississippi. New York: Ithaca.

5. G. dimorphella Busck. R_{4+5} stalked with R_3 and M_3 with Cu_1 and Cu_2 . Palpi yellow, lightly mottled with black scales; antennæ yellowish fuscous, not strongly annulate, and without notch. Head and thorax clear straw yellow; fore wing light straw yellow, lightly dusted with black, with two blackish discal dots in cell and a well-marked terminal series. 10-11 mm.

Maryland.

6. G. aberratella Busck. Fore wing with R_{4+5} stalked with M_1 , forking over apex; M_5 free (Sceptea Walsingham). Antennæ dark fuscous and simple in both sexes. Head and thorax dark; palpi heavily dusted; fore wing heavily overlaid with fuscous scales, especially along edges and toward apex. The usual discal dots nearly covered over. 14 mm.

June.

Maryland; Virginia; Missouri.

There is an undescribed species from New York, near aberratella.

11. ANORTHOSIA Clemens

Male antenna with a deep oblique notch in shaft near base, the segments before it much thickened. Palpi with a triangular tuft on second segment as in Dichomeris, but hardly as broad at tip, and in male with a large expansible tuft above. Fore wing as in Trichotaphe and Dichomeris, Cu_1 and Cu_2 strongly stalked; outer margin oblique. Hind wing hardly as wide as fore wing, with apex more drawn out than in Dichomeris.

1. A. punctipennella Clemens. Rather dark ochreous. Palpi dark brownish externally; antennæ sharply annulate with dark brown. Ordinary spots dark, somewhat diffuse, contrasting, and with an additional antemedial dot or pair of dots. Fringe ochre yellow, with a series of black bars in its base. Hind wing 12 mm. grayer.

June and July.

Distribution general, north to Pennsylvania. New York: Taughannock Falls, Ithaca.

12. STROBISIA Clemens

Similar to Trichotaphe; palpi smooth, third segment as long as second or longer, both smooth and slender. Fore wing typically with R_4 and R_5 united, long-stalked in emblemella (subgenus Holophysis Walsingham), Cu_1 and Cu_2 stalked. Hind wing

trapezoidal, without decided produced apex; hardly as wide as fore wing; R and M_1 connate. Fore wing with metallic markings.

An exotic species has been bred from a fungus bed in a termite nest.

Key to the species

Narrow blue dots and dashes.....irridipennella. Broad spots and bands.....emblemella.

1. S. emblemella Clemens. Fore wing brown-black, with steel-blue markings: a basal band, and antemedial band from costa across cell, a very irregular, narrower medial band, partly fused with the discal dot and forked toward inner margin, a costal spot four-fifths way out to apex, and a subterminal band reaching twothirds way to costa and very irregular on outer side. The bands easily become dulled in old specimens. 9 mm.

June and early July. Caterpillar possibly on buckeye.

Western New York to District of Columbia, southern Ohio, and Kentucky. New York: Rock City (Cattaraugus County), Taughannock Falls (Ithaca).

2. S. irridipennella Clemens. Shining deep brown. Fore wing marked with bright iridescent blue. A strongly oblique line from costa a fourth way out across cell and fold; a slightly more erect median line running into cell, and a longer one, almost three-fourths way to apex. A short bar four-fifths way out and a bar on inner edge near middle; the longer lines sometimes broken into dots (about 10 in all). 10 mm. (aphroditeella Chambers, proserpinella Frey.)

June to August.

District of Columbia to Missouri. New York: Robinson Collection (presumably from New York).

13. DUVITA Busek

(Anacampsis, Aproarema, Untomia, in part; with Battaristis Meyrick)

Similar to Stomopteryx and Anacampsis. Palpus with second joint smoothly thickened and abruptly cut off, third slightly thickened. Fore wing normal, M_1 not stalked, M_3 and Cu_1 sometimes stalked, typically free. Hind wing about as wide as fore wing, trapezoidal, with somewhat produced apex; R and M_1 stalked, M_3 and Cu_1 connate, discocellular vein imperfect. Palpus shorter and hind wing less produced than in Stomopteryx. Fore wing in all our species with a black dot immediately below the apex as in *Epithectis subsimella*.

Key to the species

Dark smoky gray......4. tahavusella.

I. Apex of hind wing strongly produced (Duvita).

1. D. vittella Busck. Palpus with second joint black-brown outwardly, tipped with white; third whitish with broad black annulus; head and thorax whitish fuscous; face pale. Fore wing bright brown with white bands, strongly dusted with steel gray, much as in *Paralechia pinifoliella*, but without tufts; basal half of costal edge dark brown; two transverse gray and white fasciæ, the median one strong, defined on basal side; outer margin and apex also gray and white,

with a black dash at middle of outer margin. Terminal line black, white-edged: fringe gray, white-tipped. 10-11 mm. Larva in "stunted cones" of Scotch and Austrian pine, and in "Cecidomyid

gall" on Pinus tæda.

Long Island, New York; Maryland.

2. D. nigratomella Clemens. Head whitish; antennæ pale yellowish; palpi with second joint fuscous on outer side except extreme tip, third with fuscous outer line. Fore wing clay color, basal two-fifths of costa white, outer part alternately barred black and white, with a long, nearly longitudinal, black dash at middle of costa. Postmedial line white, straight, oblique outward, fading out below middle of wing near apex. Apex white, with a black spot below it. Hind wing a little darker. 9 mm. (apicistrigella Chambers, apicilinella Clemens).

May to July.

New Hampshire to Virginia and Colorado. New York: Rock City (Cattaraugus County).

3. D. concinusella Chambers. Evenly dull gray, head and costa paler, antennæ blackish; postmedial line white, the upper half as in the last species, then turning a right angle and running into inner margin, with white dots on costa and outer margin near apex, and traces of oblique white striæ at middle of costa, all defined with dark. Apical dot in a gray shade. 9 mm. Pennsylvania; Colorado; and Texas. New York: Rock City (Cattaraugus

County).

4. D? tahavusella²¹ Forbes. Antenna with a single bristle on scape, representing a pecten. Ædæagus simple, spine-like. Palpus with second segment thickened, but apex. Fuscous; inner side of palpus, scaling of tongue, and lower part of face pale. Fore wing with scale-bases pale, but hardly showing in a fresh specimen; with a whitish, diffuse, subterminal spot on costa, and a fainter one opposite it on inner margin. Hind wing light gray, with fuscous fringe. 10 mm.

June 8, 10; July 10. New York: Uphill Brook (Mt. Marcy), Peru (Adirondack Foothills).

II. Apex not produced, hind wing trapezoidal (Battaristis).

5. D. conclusella Walker. Gray, variable in shade, often shading into blackish before the postmedial line, and heavily shaded with white. A fair proportion of scales white-tipped. Antenna blackish, annulate with white, the outer third with annulations on alternate segments only. Palpus with a black bar or two on third segment, and second segment fuscous. Fore wing with a slender black dash along middle of costal edge, and another at origin of postmedial line. Postmedial line strongly curved out on upper half, then sinuous or obscure to inner margin. Terminal dots often strong, the dot below apex often obscure, or only slightly larger than the other terminal dots. Hind wing with apex marked but not extended, R and M, very shortly stalked or approximate; M, and Cu, of fore wing occasionally stalked. 9 mm.

June to August.

Canada to Pennsylvania. New York: Peru, Rock City, Ithaca.

14. STOMOPTERYX Heinemann

(Aproærema Durrant; Anacampsis auct.)

Palpi smooth, slender, with third segment longer than second. M, shortly stalked with R_4 and R_5 in fore wing (fig. 156), the other veins free; hind wing narrow, apex produced, R and M_1 stalked, M_2 free, associated with Cu-stem. This

21 Tahawus is the Indian name for Mt. Marcy.

10

genus is derived from the similarly marked group of Anacampsis, and differs mainly in the reduced hind wing. Duvita is a parallel development.

Key to the species

Third joint of palpus longitudinally striped with white.....l. palpilineella,
 Third joint of palpus not striped.

2. Fore wing with a curved white fascia at middle.....2. kearfottella.

2. Fascia two-thirds way out to apex, or obsolete......(1) nigrella.

1. S. palpilineella Chambers. Fore wing, head, and body dull black; extreme tip of second joint of palpus white, third joint jet black with fine, often broken, white stripes. Sometimes with traces of a pale postmedial line at costa and inner margin. 8 mm.

August. Caterpillar folding leaves of red clover.

New Jersey to Missouri and Louisiana.

A. nigrella is probably western only (east to Kansas). It is very like A. palpilineella, but with the palpus not striped.

2. S. kearfottella Busck. Purplish black; antennæ annulate with white; palpi silvery white, third joint dusted with fuscous, darkening to tip. Face white; fore wing with a slightly excurved median white fascia. Fringe white-tipped at middle. Hind wing slightly duller. Tibiæ silvery white; femora black; tarsi dusky.

July.

New Jersey.

15. POLYHYMNO Chambers

(Copocercia Zeller)

Palpus with second segment hardly thickened, third much longer. Fore wing narrow, rather oblong, caudate, the fringe strongly concave above and below the long-drawn-out apex. R_4 and R_5 very long-stalked in our species; hind wings narrower, apex much drawn out and almost caudate, R and M_1 long-stalked, M_3 and Cu_1 short-stalked, M_2 low, discocellular vein weak. Larva on Leguminose. The genus also occurs in Africa.

8

1. P. luteostrigella Chambers. Silvery white, thorax with four longitudinal golden brown lines; fore wing with three brown lines from base to apex, and some oblique ones at apex of costa, converging at apex of wing, and with anastomosing dorsal lines, leaving inner margin white; dorsal fringe yellow. 9 mm. (fuscostrigella Chambers).

August.

Connecticut to South Dakota (Upper Austral Zone) and south.

16. PARALECHIA Busck

(Gelechia, Aristotelia, in part)

Palpi with third segment only three-fifths as long as second, blunt at apex; fore wing (fig. 165) with three pairs of raised tufts; M_1 free; M_2 , M_3 , and Cu_1 long, approximate; Cu_2 shorter and distant; with a reniform area of blackish sex scaling on under side, in male. Hind wing three-fourths as wide; long-trapezodial; outer margin a little waved; R and M_1 very slightly divergent; M_2 , cubital; M_1 and Cu_1 almost connate, or stalked.

1. P. pinifoliella Chambers. Shining deep yellow-brown; inner edge of fore wing toward base, apex and apical fringes, and three bands, dividing the wing inte equal parts, white, dusted with gray; the bands rarely broad and suffused; the

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raised tufts on inner edge of bands, black. Outer band narrowest and paler. No apical dot. Hind wing fuscous. 9 mm.

June. Caterpillar a leaf-miner in pine needles.

General in distribution and common. New York: Round Lake, Ithaca, Karner.

17. AROGALEA Walsingham

(Paralechia, in part)

Hardly distinct from the European genus Stenolechia. Palpi with third segment as long as second, and acute; second segment rough-scaled. Fore wing dull, with slight raised tufts in fold at one-fifth and three-fifths way out, easily lost. Venation much like Paralechia; hind wing with apex a little drawn out; **R** and \mathbf{M}_1 well separated at origin, but strongly divergent; \mathbf{M}_3 and \mathbf{Cu}_1 shortly separated at origin.

1. A. cristifasciella Chambers. White, palpus with four blackish bands, fore wing with black dots near base. Black antemedial fascia from costa obliquely inward to inner margin, including first tuft; black postmedial spots beyond middle (the dorsal one including the second tuft); and some minute black dots. Hind wing yellowish. 13 mm. (*inscripta* Walsingham). April; July. Caterpillar on oak, skeletonizing leaves from under side, or

inquiline in galls.

General in distribution. New York: Crosby (Yates County).

18. TELPHUSA Chambers

(Xenolechia Meyrick)

Closely related to the tufted mediofasciella group of Gelechia, and derived from it. Fore wing (fig. 163) similar to Gelechia, usually with raised tufts; hind wing as in Gelechia, except that M_a and Cu_1 are well separated at origin; palpi as in Gelechia. The species average smaller than Gelechia.

Key to the species

- With contrasting, oblique, white or whitish fascia at basal fourth.
 Fascia narrower, outer half of inner edge white.....l. longifasciella.
 Fascia broader, inner edge not white......2. latifasciella (part).

1. No contrasting white fascia.

- 2. Ground color of fore wing white, not heavily dusted with gray.
 - 3. Markings black; antemedial fascia stopping at fold.....8. basifasciella.
- 2. Ground color of fore wing not white, or else heavily gray-dusted.

 - 3. Gray, often powdery.
 - 4. An oblique black antemedial fascia, marked by two raised scale-tufts, and often broken below costa.
 - 5. A broad lighter antemedial fascia, completely defined on both sides, and contrasting with the blackish base.....2. latifasciella, in part.
 - 5. Antemedial fascia defined toward base only, and of the same shade as ground color and base.
 - 6. Head and thorax light fuscous, not powdery. .7. palliderosacella.
 - 6. Head and thorax, like fore wing, heavily dusted with blackish on pale gray or white.
 - 7. Fore wing dusted with purple-black on white; antenna annulate

4. No oblique antemedial fascia; wings smoothly scaled.

5. A sharply defined blackish spot on fold near middle..11. glandiferella. 5. No such spot.

6. Palpus with third segment yellowish; brown shades on fore wing.

- 10. belangerella.
- 6. Palpus with third segment pure white; fore wing dusted with black and gray on a pearl-white ground... (Glauce pectenalæella).

1. T. longifasciella Chambers. Dark purple-brown. Head white; palpi purplish with white annuli on third segment at base and end; fore wing with a white fascia from costa near base, running obliquely across to inner margin, then along inner margin almost to apex, but constructed or interrupted at anal angle. 15 mm. (curvistrigella Chambers, obliquefasciella Chambers).

April and May.

New York to Oregon and Texas. New York: East Aurora, Crosby, Ithaca.

2. T. latifasciella Chambers. Powdery gray, the base blackish; a broad paler (typically white) antemedial fascia, as wide as breadth of wing, edged on both sides with raised black lines. An irregular postmedial line, weak or broken in middle, with a whitish shade beyond it toward costa, and typically a large white patch between it and end of cell; a double black tuft at end of cell. 13 mm.

May. Larva on oak; a leaf-roller.

Massachusetts to North Carolina and Missouri.

3. T. quinquecristatella Chambers. Deep brown, with a little purple iridescence; 3. T. quinquecristatella Chambers. Deep brown, with a little purple indescence; face dusted and shaded with whitish; palpi light powdery gray, with black bars; legs dark brown with white bars. Fore wing with three pairs of somewhat darker tufts; the anterior of each pair a little farther out, and each with some white-tipped scales; apical region dusted with white, indicating the postmedial line at the margins. No black spot at base of costa. 16 mm. May; July; October. Larva a bud worm on huckleberry. Slender; yellowish white; head and cervical shield pale yellow; body showing the green dorsal vessel. What is annarently the same thing also occurs on Comptonia

What is apparently the same thing also occurs on Comptonia.

Generally distributed eastward.

4. T. quercinigracella Chambers. Powdery gray: palpi and antennæ also banded in two shades of gray; fore wing with a heavy black bar from costa to fold near base, nearly interrupted in middle, and containing two large tufts. Black basal spots at costa and in fold; a black spot at middle of costa, with an oblique spot in cell and a streak in fold opposite; a black postmedial spot on costa, connected to a similar spot in fold by the black discal bar. Apex blackish, with an obscure, sinuous, pale, postmedial fascia. Fringe powdery gray. 12 mm.

Caterpillar on black-jack and other oaks, in late June and July. White with a black bar on each side of head; a yellow cervical shield with black posterior edge and black tubercles; at maturity developing red transverse bands on the middle of the segments.

Generally distributed. New York: Ithaca. 5. T. fuscopunctella Clemens. Very closely related to the last species and indistinguishable when rubbed. Antemedial band usually broken, the tuft nearer costa being weak and well separated from the other; ground slightly paler, and base more contrastingly marked.

Caterpillar with T. quercinigracella, similar to it, but with two ocellar dots on each side of head instead of a single bar, and incisures instead of mid-segments striped with red at maturity.

Pennsylvania and District of Columbia.

6. T. querciella Chambers. Very close to T. quercinigracella; ground whiter, antennæ and palpi more distinctly ringed with white; posterior tuft of thorax yellow. Fore wing marked as in the last two species; a large proportion of scales white, with a gray bar only. 12 mm.

Early June. Caterpillar whitish; head, cervical shield, and tubercles purplish brown; body with fine purple addorsal and broader subdorsal lines. A leaf-roller on oak in May. These three species are practically indistinguishable without breeding.

New Jersey; Kentucky.

7. T. palliderosacella Chambers. Similar to the last three species but with more purplish tint. Head hardly powdery, and pale in front. Fore wing with ground a light rather pinkish fuscous; the black spots on the costa as in quercinigracella, and antemedial bar strong, but tufts small, sharply defined, black, and well separated. 13 mm.

Larva in oak, sometimes inquiline in galls. New York to Texas. New York: Rock City (Cattaraugus Co.).

8. T. basifasciella Zeller. White; fore wings with a little gray dusting, palpi with two gray bands on second, and stronger ones on third segments; antennæ blackish. Fore wing with a black bar from costa at a fourth way out, running obliquely outward to below fold and connected along costa to base; black spots on costa before and beyond middle, with a dark gray spot on inner margin opposite the outer one; a black discal dot and bar, and heavier gray dusting at outer margin. Hind wing dirty white. 10-12 mm.

May to June; late July. Caterpillar on oak, skeletonizing the leaves from the under side.

New Hampshire to Missouri and Texas.

9. T. betulella Busck. White, mottled with pale pinkish brown; antennæ annulate with white, and pale and dark brown; fore wing with three obscure and broken oblique transverse fasciæ, including the large concolorous basal tuft, and with smaller outer ones; base of costa black. Fringe and hind wing whitish. 12 mm. August; early spring. Caterpillar in a rolled leaf of black birch, pupating in

the roll.

District of Columbia; Virginia.

10. T. belangerella Chambers. Fuscous, powdered on a slightly pinkish white ground, with slightly raised brown bars on disc, and a brown streak on fold. Terminal dots dark, obscure, diffuse. Antennæ and legs brown; palpi with second segment pale gray, third yellowish with brown rings. 14 mm. (oronella Walsingham).

May to July. Caterpillar a leaf-roller on alder.

Canada; eastern United States.

11. T. glandiferella Zeller. Pale, hardly powdery, ochreous gray; a velvety, sharply defined, dark brown spot in fold near base; a larger dark brown spot on fold, or typically, a bar reaching to inner margin, beyond it; and usually a small dot at end of cell: each spot lightly defined with paler brown. 12 mm.

July to September.

East River, Connecticut, to Kansas and south.

19. GLAUCE Chambers

Closely related to Telphusa, and probably a development of it. Palpus with second segment somewhat thickened, but nearly smooth, third segment longer; fore wing with R, and R, stalked, M, normally stalked more or less with R, sometimes to well beyond origin of R., as in the Sitotroga group, rarely free, connate. M_1 connate with M_3 and widely separated from M_1 , even at margin; Cu, long and parallel to M_2 , M_3 , and Cu. Hind wing with R and M_1 stalked, approximate, or widely separated, apparently varying locally; M_2 twice as near M_3 as M_1 . M_3 and Cu, separate. Male with a very large tuft of black bristles (fig. 166) projecting forward from front of hind wing, attached below Sc; the venation distorted; and with costal edge clothed with much-enlarged scales.

1. G. pectenalæella Chambers. Black, heavily powdered on a white base; head and thorax almost solid black; antennæ black, with a strong white ring at tip of scape; fore wing mottled gray and black, without definite markings. 7 mm.

July; June, southward. New York and District of Columbia to Kentucky and south. New York: Rock City (Cattaraugus County).

20. AUTONEDA Busek

(*Neda* Chambers, not Mulsant)

Palpus with a very short, triangular tuft on under side of second segment; third as long, slender. Fore wing with M_1 approximate to R_{4+5} ; M_2 separate; hind narrower than fore wing, with M_2 nearer M_1 than M_3 ; apex produced. 1. A. plutella Chambers. Brownish black; palpi above, head, thorax, and inner

1. A. plutella Chambers. Brownish black; palpi above, head, thorax, and inner margin of fore wing, cream white; shoulders black, shading into cream white; costa white; fringe cream white except at apex. 11 mm.

September. Type only seen.

Kentucky.

21. ARISTOTELIA Hübner

(Ergatis Heinemann, with Eucatoptus Walsingham)

Palpi small, often quite rough. Fore wing broad, lanceolate (fig. 164). M_1 free; M_2 and M_3 occasionally stalked; the other veins free. Hind wing falcate, with produced apex; cell large; R and M_1 divergent but widely separate at base; M_2 nearer M_1 than M_3 ; M_3 widely separate from Cu_1 . Hind wing narrower, sometimes much narrower, than fore wing. Male sometimes with a costal hair-pencil on hind wing, not disturbing the veins (Eucatoptus).

The palpi in this group are strongly divergent, and the patterns described are those of the inner face; the outer face is usually similar but with a little more black.

Key to the species

1. Ground not dull red (sometimes suffused with pink).

2. Ground nearly even, with small markings.

3. Ground white or yellowish to dull wood-brown.

- 4. Five black marks on disc.....14. quinquepunctella. 4. Not so marked.
 - 5. A light yellow or cream-white streak running from costa at beginning of fringe.
 - 6. Palpus with second segment and basal half of third segment black 15. robusta.
 - 6. Palpus less extensively black.
 - 7. Apex of palpus black; a paler species.....13. angustipennella.
 - 7. Apex of palpus not black......12. gilvolineella.
 - 5. No light costo-apical streak.....11. disconotella.
- 3. Ground black or fuscous.

4. Fifth and tenth joints from apex of antenna white above.

8. absconditella.

4. Antennæ without white joints
5. With subterminal dots9. minimella.
5. No subterminal dots10. physaliella.
Fore wing broadly and contrastingly marked in two or more colors.
0 117/11 / 11/ 1/

3. With metallic markings.

3. No metallic markings.

- 4. Fore wing with dark fascia at apical third......3. molestella. 4. No such fascia; usually roseate.
 - 5. With pure white costal markings.....l. roseosuffusella. 5. No pure white markings.
 - 6. A costal hair-pencil at base of hind wing....2. rubidella (male). 6. No costal hair-pencil.
 - 7. Fore wing strongly suffused with pink; third joint of palpus
 - with dark base and tip; three white bands, the first weak. 2. rubidella (female) and 4. pudibundella. 7. Fore wing with slight traces of pink or none; third joint of palpus with broad blackish base, the first white band of the

1. A. roseosuffusella Clemens. Palpus with third segment black, imperfectly banded with white on basal balf, the extreme tip white. Fore wing white, lightly dusted with gray, with three blackish bands about as wide as the spaces between them. First band strongly oblique outward, from costa at base, crossing fold, and fading out; second parallel to it, running from costa at a third way out, abruptly truncate in fold; third broader, transverse, at two-thirds way out to apex, with pink spots before and beyond it at inner margin, and containing a large yellowbrown spot or shade. Apical region nearly filled with a black patch, but leaving a white spot at apex. Inner margin shaded with yellow-brown on basal half or more. 10 mm. (bellella Walker).

May to October; commonest in July and August. Caterpillar a leaf folder on clover, also feeding on the flowers; also reported from sumach by Clemens, most likely in error.

Generally distributed and common. New York: Ithaca.

2. A. rubidella Clemens. Fore wing pink, dusted with fuscous, and somewhat shaded with yellow; head yellowish; palpus with third segment relatively short, with three white bands, near base, before middle, and before tip, the tip black. Fore wing with oblique fuscous bands, much like those of A. roseosuffusella, the first one ending at fold, the third dark fuscous toward the costa, shading into brownish yellow below and fading out on inner margin. Apex wholly shaded with blackish, leaving no apical spot. Inner margin broadly shaded with brown-ish yellow to the middle, interrupted with pink at a quarter way out. Thorax fuscous. Hind wing of male with a hair-pencil. 8 mm.

General in distribution; May.

A. rubensella Chambers, generally considered a synonym, was described as having the palpus white-tipped (perhaps damaged), the underlying ground color white instead of pink, tongue brownish (usually gray in this group), bands of fore wing brown, and thorax pale ochreous. 8 mm. Kentucky.

3. A. molestella Zeller. Palpus with third segment as in pudibundella. Ground of fore wing powdery gray, the paler scales white-tipped. Half of surface covered by fasciæ, which are even, checolate brown; first fascia paling out to base, with a couple of yellow dots; second broad, with a yellow spot in it below fold; third parallel, nearly interrupted by a yellow spot over cell; two dark brown dots on costa near apex; costal and dorsal fringe barred with pink in the base, the bars invading the membrane, at least on the costa. First discal dot large, tangent to second fascia; second obsolete; postmedial fascia followed by a black bar in discal fold. 12 mm.

District of Columbia in September; Texas.

4. A. pudibundella Zeller. Hardly distinguishable from rubidella and the several undescribed species closely related to it. Palpi pinkish white, with black bands before middle and before apex of third segment. Antennæ blackish, obscurely annulate. Head and thorax variable, usually gray, with pale face; fore wing with ground powdery brown-gray, suffused with yellow and shaded with pink; markings blackish, about as in D. roseosuffusella, not distinct below fold, and with a somewhat less distinct costal spot before the apex. Fore wing dark gray beneath, with a longitudinal pale streak along middle of costa, and pale subterminal spot. Hind wing with a streak of rough gray scales below costa. 11 mm.

July.

Massachusetts; Texas. New York: Rock City.

There are a number of undetermined species closely related to this, and which will run to this in the key, but which differ in details of markings and sex-scaling; one, usually considered the same, seems to differ only in the white-annulate antennæ, and eats apple. A. intermediella Chambers is a name belonging to a member of this group.

5. A. fungivorella Clemens. General pattern as in roseosuffusella, etc. Head whitish, tinged with fuscous; palpi white, second segment with three blackish rings, third also with three, leaving the extreme tip white. Fore wing pale luteous, the inner margin strongly dusted with testaceous brown from base to tip; the costal half with bands of fuscous brown, on a brownish-dusted, luteous base; second band ending in a point, opposite which is a blackish, white-ringed dot; a couple of black terminal dots near anal angle. Fringe testaceous with a white patch below apex crossed by a dark line, and a white patch near anal angle. 12 mm.

August. A leaf-roller on willow, often feeding in the various willow-galls.

District of Columbia; Illinois; and probably generally distributed. 6. A. salicifungiella Clemens. Very similar to A. fungivorella, of which it is possibly a red variety. Head reddish, palpi pale red with two black rings on second segment; third segment black-tipped; fore wings dark brick red, the outer half almost solid red, the basal half marked with whitish; a whitish band near the base, dusted with fuscous, and followed by a darker, red-brown band; three small white spots on costa; ground dusted with fuscous, and inner margin with whitish red and fuscous. Fringe red. 13 mm. August. Caterpillar in a loose open web between leaves of willow, not drawing

them together; eating parts of the leaves. Pale green, with many fine broken lines; head paler, straw yellow. July. Cocoon outside the web, in trash, etc.
Illinois; Texas, in June.
A. ivæ Busck, a Florida species without any pink shading at all, is to be

expected northward where its food, *Iva frutescens*, occurs. 7. A. elegantella Chambers. White. Four transverse, black-edged, rich brown bands; a black bar on middle of disc containing four or five metallic lead-blue spots. 13 mm.

June, in Texas.

Texas to Missouri, central Illinois, and west, doubtfully reported from Pennsvlvania.

8. A. absconditella Walker. Shining bronzy gray-brown; extreme tip of second segment of palpus white; third segment white with blackish shading at base and apex; fore wing with a black spot at end of cell and sometimes a less distinct one at middle; a fine outwardly oblique subterminal line from costa, fading out at middle of wing, and a similar line opposite it running from inner margin, the two at right angles. Fringe with more or less distinct, blackish basal line, cut with luteous. 8-10 mm.

May and June. Larva in stem of Polygonum, often causing a slight gall; and apparently the same species in Ampelopsis. Larva hibernating.

District of Columbia; Kentucky; Missouri.

9. A. minimella Chambers. Dark fuscous, indistinctly sprinkled with white, no annuli on antenna; face a little paler below; fore wing with obscure pale postmedial dots on both margins. 9 mm.

July and August. Caterpillar on oak.

New Jersey; District of Columbia to Kansas and Texas.

There is a closely related species on Polygonum associated with A. absconditella. 10. A. physaliella Chambers. Practically identical with A. minimella, but with the pale lower part of the face sharply defined, and contrastingly pale. 9 mm. May; July. Caterpillar a leaf-miner on under side of leaves of Physalis. Green

with yellow head and cervical shield; at maturity becoming crimson with greenish head and cervical shield; cocoon outside the mine. The moth of the second brood emerging the next spring.

Kentucky; Texas.

11. A. disconotella Chambers. Light dull ochreous, somewhat dusted with gray. Palpus brown, apex of second segment pale, third with a broad blackish band at middle, at least in outer side. Fore wing with a horizontal elliptical black spot at end of cell, and a smaller spot below costa half way to base. 8 mm.

June. Larva in stem of raspberry.

District of Columbia; Kentucky; Mississippi. 12. A. gilvolineella Clemens. Superficially very close to Paltodora striatella, but with simple palpi. Palpus with second segment dark on basal half and white on outer half within, mostly black outwardly. Fore wing pale yellowish, over-laid with fuscous brown, the outer part rather darkened. Apex crossed by a fine pale line running to a pale shade on outer margin. Discal dot black, distinct; fringe with a black basal line, cut by the pale terminal bars. Antennæ dark. There may be a black claviform spot also. 12 mm.

July.

Pennsylvania: Philadelphia, Hazelton, Easton.

13. A. angustipennella Clemens. Similar to A. gilvolineella, head and thorax and base of fore wing nearly white; outer part light brown. Antennæ white on basal two-thirds, fuscous-barred at tip; palpus white, with a black bar on second segment and apical half of third segment black, except extreme tip. Fore wing with some longitudinal brown streaks near base, dot at end of cell smaller than in the preceding species, and claviform dot obsolete; postmedial line shorter and without a pale spot at its apex. 12 mm. (kearfottella Busck.)

July.

New Jersey to Ohio.

14. A. quinquepunctella Busck. Antennæ light fuscous, annulate with luteous. Palpi with second segment fuscous with white apex, third white with fuscous apex. Face whitish, head and thorax dull luteous; fore wing luteous, with gray-ish dusting, becoming fuscous outwardly, with five black spots on disc, four in a rhomb (the basal sometimes weak) and one beyond. Hind wing paler. 12 mm. June.

Pennsylvania.

An apparently identical species occurs in southern California.

15. A. robusta Brann. Dull ochreous, rather densely overlaid with purplish fus-cous dusting, especially toward apex of fore wing. Palpus blackish, apical half of third segment, and sometimes extreme apex of second, white, extreme apex of third sometimes black. Base of antenna fuscous, annulate with white, apex blackish, with apical segment, and fifth and tenth segments from apex, white. Fore wing with a pale costo-apical streak, an elongate black discal dot and faint dark shades; fringe with a broken line in base. 11-12 mm.

Larva at first forming a small, transparent blotch mine in Scirpus atrovirens; then a linear green mine, and finally a broader transparent portion. Larva in the spring; moth in June.

Cincinnati, Ohio.

22. ENCHRYSA Zeller

Similar to Aristotelia, but with the outer half of the costa distinctly concave. Palpi entirely smooth, as long as head and thorax, and strongly divergent.

1. E. dissectella Zeller. Dark bronzy brown, nearly black. Basal half of fore wing brown with decided green iridescence, the brown area ending in a vertical yellow line. Outer half orange, with a large blackish patch, starting just beyond middle of costa over half as wide as the wing, and gradually tapering to apex. Fringes blackish. Apieal fourth of antenna white. 11 mm. (Aristotelia youngella Kearfott).

July. Rare.

Ontario to Ohio, west to Vaneouver Island. New York: Wells (New York State Museum).

23. CHRYSOPORA Clemens

(Aristotelia, in part; Nannodia Heinemann; Nomia Clemens, not Latreille)

Very close to Aristotelia. Palpus hardly longer than head, with third segment much shorter than second. Fore wing as in Aristotelia; hind wing narrow (fig. 168) with apex strongly produced, and \mathbf{M}_1 rudimentary.

1. C. lingulacella Clemens. Golden yellow; head and thorax blackish, iridescent; palpi paler with fuscous third segment; antennæ dark. A blackish, more or less metallic, patch on base of costa; an elongate one on basal half of inner margin, and a large trapezodial patch on middle of costa, all edged with brilliant violet-silver. Costa edge brown outwardly. Fringe dark brown, with silver in its base below apex. Hind wing dark. 7 mm. (hermanella Chambers, armeniella Frey and Boll).

May; August. Caterpillar in September, in a large blotch-mine on Chenopodium and Atriplex.

Pennsylvania and Kentucky to Michigan and Kansas.

2. C. hermanella Fabricius. Very similar, but with the silvery antemedial band extending clear across the wing, and the ground deeper orange. Caterpillar like *lingulacella*.

Ontario; Minnesota; Iowa; Missouri; possibly introduced from Europe. New York: Ithaca.

24. RECURVARIA Haworth

(Restricted, not Meyrick. Evagora Clemens; Eidothoa, Sinoë Chambers; Aphanaula Meyrick)

Palpi slightly roughened beneath, third segment nearly as long as second. Fore wing (fig. 167) with \mathbf{M}_1 stalked with \mathbf{R}_s nearly to apex, \mathbf{Cu}_1 longer than \mathbf{Cu}_2 and much more closely approximated to \mathbf{M}_3 ; usually with raised scale-tufts. Hind wing typically nearly trapezoidal, male often with costal hair-pencil. **R** and \mathbf{M}_1 only moderately approximate at base.

The caterpillars of one group are miners in the needles of various conifers. In the needle-conifers they pass from needle to needle in a thin silken tube along the stem; in the scale-type they eat out the spray as a whole. The moths emerge in June. The young larva hatch in July and form a minute mine in a single needle in which they hibernate, in the spring moving to a new place and feeding up rapidly. The deciduous feeders live in a silken tube on the suface of the leaf, and seem to have the same seasonal history. The species are close and not fully understood; many can hardly be determined without the food plant:

Key to the species

- 1. Fore wing with a longitudinal stripe through the middle from costa near base to apex.

2. Costal area about half covered with black mottling; on sweet gum. 12. dorsivittella.

- 2. Costa very dark, concolorous with median area, the black streak lying in
- the fold; on alder.....15. alnifructella. 1. Fore wing without longitudinal stripe, or (obliquistrigella) with a slight one on the apical half only.
 - 2. Blackish and black with a little white scaling and an angulate, white postmedial band.
 - 3. Head and inner margin dark-dusted and shaded on a luteous ground.

5. piceaella, variety obscurella.

- 3. Head and inner margin blackish, concolorous......14. nigra. 2. Not dominantly brownish black.

Fore wing more or less yellowish.
 An oblique antemedial fascia running from costa to fold.

- 6. Ground decidedly tinted with yellow; head straw yellow. Larva on spruce.
 - 7. Fasciæ broken, leaving the series of tufts in the cell showing as black dots about as conspicuous as those in the fold.

3. coniferella.

- 7. Ground duller; fasciæ covering the series of tufts in the cell. 4. piceaella.
- 6. Ground hardly tinted with yellow; head cream color.

 - 7. Second fascia running out in cell to join third; on juniper.

7. gibsonella.

5. A slight yellowish-white antemedial shade only; cell-tufts minute or absent; second segment of palpus yellowish white.

1. apicitripunctella.

- 4. Fore wing not tinted with vellowish. 5. Fore wing with a strong, oblique, white streak at a third way out.
 - 13. cristatella. 5. No strong white costal streak one-third way out.
 - 6. A black dorsal patch near base.....10. robiniella. 6. No such patch.
 - 7. Male with hair-pencil toward inner margin; palpus with second segment mostly blackish; first antemedial fascia short; larva on oak11. quercivorella.
 - 7. Hair-pencil not reported; palpus with second segment heavily dusted with gray; first fascia reaching to fold.
 - 8. Ground brown, suffused on a cream-colored base; larva on arbor vitæ5. thujaëlla. 7. No hair pencil; palpus with two black bars on second segment;

I. Evergreen feeders, ground strongly yellowish.

1. R. apicitripunctella Clemens. Pale yellow, shading into white; palpus with second segment handed with yellow and white, third white with two black bars. Antennæ annulate, pale yellow and blackish. Fore wing with a brown shade on base

of costa, some black scales before middle, and a slight oblique postmedial fascia from costa, of darker yellow, scaled with black, and edged with white. A black tuft in fold opposite this fascia. About five small black terminal dots, the apical largest. Sometimes with a complete angulate white postmedial line. Male with hair pencil. 9 mm. (*abietisella* Packard).

June and July. Caterpillar webbing needles of hemlock in early June. Green, with blackish head and cervical shield.

New Jersey to District of Columbia and Pennsylvania; probably more generally distributed. New York: Wells.

2. R. variella Chambers. Yellowish white, apical half of fore wings suffused with golden yellow or dusted with brown; three or four distinct black dots at base of costal fringe, which has some scattered black scales; occasionally with a couple of fine streaks along costa. Costa sometimes wholly brown or golden. Sometimes with a white angulated postmedial fascia broken at middle; a raised black tuft on fold at three-fourths way to apex. Head, including inner side of palpi and antennæ, white. Outer side of palpi yellow. 8 mm.

June and July. Caterpillar in a silk-lined tube between needles of bald cypress and evidently other trees. Mississippi valley specimens are yellower than eastern ones.

Montclair, New Jersey and District of Columbia to Kentucky. New York: Yonkers.

3. R. coniferella Kearfott. Similar to R. apicitripunctella; ground ochre, not lemon yellow, and much mottled with white; antemedial fascia nearly continuous as far as fold, but all fasciæ showing the tufts as partly or entirely separated dots. Apex and fringe more heavily gray-dusted; second segment of palpus rather heavily scaled with black, suggesting two fasciæ.

June. Larva on pine.

Ottawa; Ontario.

4. R. piceaella Kearfott. Similar to R. apicitripunctella; dark, dusted with fuscous on a straw-yellow ground; head straw yellow, concolorous with the ground. Fore wing with three fasciæ, like R. thujaëlla, the outer one white, and right-angled at the middle; normally a black dash in the terminal area, extending to the apex. Hind wing dark gray. Abdomen with dark dorsal patches. 10 mm.

Variety obscurella Kearfott (*nigra* Kearfott, not Busck), is suffused with blackish except on the inner margin, and on the costa beyond the median fascia, where it is powdery luteous; white postmedial fascia only, contrasting. Head luteous; the face lightly and vertex heavily dusted with black.

Caterpillar webbing and mining needles of red and black spruce. Red, with light brown head and cervical shield and dark green dorsal segmental patches. Crimson ventral spots on thorax between legs, and on fifth and sixth segments of abdomen. Moth in June.

New Jersey. Apparently the same moth occurs in Ontario.

5. R. thujaëlla Kearfott. Palpus with first segment black, second with a black streak below and dusted with black, third white with two black bars. Head creamwhite, paler than in *R. piceaëlla*. Fore wing cream-white, heavily dusted with blackish, appearing fuscous gray; with three oblique blackish fasciæ, the first reaching to fold, the other two shorter; apical region darker, with blackish streakt running out to the strong costal terminal dots. Tufts well marked, in three pairs, black. Hind wing light gray. Abdomen cream-white. 9 mm. June. Caterpillar on Thuja, eating out the sprays from inside, in May. Head

June. Caterpillar on Thuja, eating out the sprays from inside, in May. Head, cervical shield, true legs, and anal plate black-brown; body dull red with greenist incisures, pinkish below.

New Jersey and probably rather generally distributed. New York: Westbury Long Island.

6. R. juniperella Kearfott. Palpi with black on outer side; antennæ annulate scape white in front. Fore wing powdery gray, with a curved black band from

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base of costa to costa near apex, defined above except at the tip with a white streak; and containing three raised tufts. Costal area heavily mottled with blackish, forming three oblique fasciæ, the outer sometimes meeting the longitudinal streak. Dorsal region lighter gray, with three raised black tufts and three minute ones along outer margin. 10 mm. (obliquestrigella Chambers?). June; August. Caterpillar on red cedar and ground pine (Juniperus), in a web

or eating out the sprays; pale green with light brown head, cervical shield, and true legs; segments dorsally with pinkish open rectangles, filled with the ground color. Connecticut; New Jersey.

7. R. gibsonella Kearfott. Similar to R. thujaëlla. Ground yellowish white, heavily overshaded with black; the first fascia reaching to fold or beyond, second broadening at middle of wing and sending out a long spur to meet the third, which is angulate and reaches inner margin. Head whitish; palpus with two bands on third segment, sometimes fused into a broad area, leaving only the base and tip white; always rather dark. 11 mm. June. Larva on juniper in May. Head and cervical shield pale honey yellow.

Ottawa; Quebec.

(Number 8 is vacant.)

II. Larvæ deciduous feeders. Moths without yellow tint. Palpus with third segment more slender than in group I, twice as long as eye, more than half of its surface covered with two broad black fascia, second segment mostly brown. Face and scape white. No hair-pencil.

9. R. cratægella Busck. No hair pencil. Wings shorter than usual; white, dusted with black. Antennæ annulate; palpi white, with two black bars on each segment and white tip. Head powdery gray. Fore wing with two oblique dark streaks from costa, the first reaching inner margin, but sometimes broken; the second reaching to cell only; and a third angled on outer side and running to below tip of second, in fold, followed by a white line; a broad blackish shade running from its angle to the apex. Basal half of fringe powdered, outer half pale, with a gray line. 12 mm. May to July. Larva on Cratægus. Probably a race of *R. nanella* of Europe.

New Hampshire to District of Columbia and Ohio. New York: East Aurora, Batavia, Albany.

10. R. robiniella Fitch. Powdery fuscous gray, somewhat mottled, with only the angulate and incomplete postmedial line pure white. Palpi with outer side of second segme_t fuscous, except at tip, inner side with two bands indicated; third segment with two black annuli. Fore wing with a blackish bar extending up and out from basal angle; a small blackish costal spot before middle, and a larger one before the postmedial band; a raised black dot beyond middle of cell; another in fold at three-fourths way to apex. Head pale. 7 mm. (robiniella Fitch, imago, not larva; fuscopallidella, robiniæfoliella Chambers).

Caterpillar pale greenish; between two leaflets of locust.

Eastern United States south to Texas and north to District of Columbia. 11. R. quercivorella Chambers. Very irregularly dusted and mottled with black on white, leaving most of the white along costa and inner margin, and forming a more or less distinct, but quite irregular, longitudinal median fascia, at least toward base. Head powdery gray; male with a large, spreading, yellow pencil at anal angle. 11 mm. (gilviscopella Zeller.)

Larva on oak, in a silken tube on the under side of a leaf; white with red spots. Moth in June.

12. R. dorsivittella Zeller. Palpi annulate, tip white; face white; vertex darker, powdery; thorax and fore wing powdery dark brown and white, the dark pre-dominant; inner margin pale, a whitish streak near base of costa, a white spot

on costa beyond middle, white along costa outwardly and at apex, with a brown costo-apical spot. Fringe pale. 12 mm. (ragatioella Chambers.)

May. Caterpillar in a brownish frass-tube 1 mm. in diameter and 1 centimeter long, along a vein on under side of leaf of sweet gum, skeletonizing leaf about the ends of the tube. Tube with somewhat valvular openings at the ends, which are closed before pupation. September.

13. R. cristatella Chambers. Rather heavily powdered with black and white, more evenly than usual, leaving a broad, oblique, clear white, antemedial fascia from costa to fold, where it fills the space between the first two tufts; a blackish shade before it; three black tufts in fold; postmedial line pale, distinct, angulate. 7 mm.

Kentucky. Type only known.

14. R. nigra Busck. Black with a silvery glance; darker than obscurella; head dark, antennæ obscurely annulate. Fore wing with traces of the silvery-white ground color, and a V-shaped fascia at three-fourths way to apex, its two limbs paralleled by a fine line beyond. Tufts black. Hind wings blackish. Legs with white annulations and tufts. 11 mm.

May. Larva on Hypericum.

15. R. alnifructella Busck. Purple-black; the first two fasciæ from costa absent, or represented by a few white scales; inner margin below fold nearly white, contrasting; the outer margin as far as the apex also strongly mottled with white. Fold defined with black; the usual tufts black and white, the outer one showing as a distinct pale spot. Postmedial line whitish, irregular and broken, the upper half in part parallel to costa and outlining a pale patch, the lower half often lost. Head and thorax contrasting, nearly white. 10 mm.

Caterpillar in catkins of alder in late fall, emerging in the spring. Pale; head brownish yellow; cervical shield and anal plate dark brown; each segment with a transverse pink band on dorsum and mottling below; abdomen with pink incisures.

25. EUCORDYLEA Dietz

Palpi with a dense expansible tuft on upper side of second segment; third shorter. Fore wings slightly concave at Cu_2 , sinuate, apex rather obtuse, with all veins preserved. M_1 stalked beyond origin of R_1 ; M_2 . M_3 , and Cu_1 approximate, Cu_4 distant, running to concavity of inner margin. Hind wings about as wide as fore wings, trapezoidal, with all veins preserved. R and M_1 connate, M_2 and M_3 approximate, M_3 and Cu_1 nearly connate. A slight modification of Recurvaria.

1. E. atrupictella Dietz. Pale creamy yellow, palpi with dark base and two bands each on second and third segments (as often in Recurvaria). Antennæ annulate. Fore wing blackish-marked, an oblique band one-fourth way out, shaded inwardly; a heavy oblique band from middle of costa about to fold; and a thinner erect one beyond, connected to it by a bar on disc. A series of small black dots around apex. Hind wings cinereous. 12 mm.

August.

Hazelton, Pennsylvania; Ontario. New York: Ithaca.

26. EPITHECTIS Meyrick

(*Taygete* Chambers; *Parasia*, in part)

Similar to Recurvaria. Fore wing (fig. 169) with Cu_2 as long as Cu_1 and nearly parallel to it; M_3 , Cu_1 , and Cu_2 evenly spaced at origin. Hind wing with R and M_1 stalked, R running to costa before apex, M_1 usually to outer margin, rarely also running to costa; M_3 and Cu_1 connate or short-stalked.

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Key to the species

A black terminal spot just below apex, as in Duvita.....2. subsimella.
 No single black terminal spot.

2. Fore wing overlaid with dark scales.

3. A broken, oblique antemedial band, farther out on costa.

4. Band broken into contrasting spots; ground practically white.

3. saundersella.

3. Smaller; bands oblique out to inner margin; browner, with dark apex.

3. saundersella.

2. Fore wing whitish gray, with contrasting blackish spots.....l. attributella.

1. E. attributella Walker. Hind wing with \mathbf{M}_1 running to apex, \mathbf{M}_2 almost connate with \mathbf{M}_3 and \mathbf{Cu}_1 , which are long-stalked; cell open. Powdery pearl white (each scale fuscous and white). Palpi fuscous, tip of second joint and two bands on third white; antennæ dark. Fore wing with a large costal spot at base and one rather beyond middle, a small one between them, and two or three less intense ones on inner margin. Raised dark dots at end of cell and in fold. Apex lightly spotted with fuscous. 10 mm. (*difficilisella* Chambers.)

The moth is locally common, resting on the trunks of trees in July.

Massachusetts to North Carolina and Illinois. New York: Ithaca.

2. E. subsimella Clemens. R and M_1 normal. Head and thorax yellowish fuscous; palpus with second joint brownish with a white ring at tip, third joint white with outer half black; antennæ dark. scape striped with yellowish in front. Fore wing dark yellowish fuscous, brown along costa from the middle and toward the tip; ground much sprinkled with white outwardly, a short yellowish white streak at middle of costa and an angulate line at two-thirds way out; a blackish spot below apex; and a dark streak in fringe. 8 mm.

A southern species, doubtful in our territory.

3. E. saundersella Chambers. Pale creamy yellow. Palpi with the two terminal joints brown with white tips, and a band on the third. Head and thorax dusted with blackish. Fore wing densely dusted below the fold, with three blackish costal spots and two on disc, sometimes partly confluent into two oblique bands. Apex heavily suffused; a dark line in base of fringe. Hind wing pale slate color. 7 mm.

Kentucky. Types only known.

4. E. geminella Riley. Cream white; a basal, blackish fascia of two partly fused spots, and antemedial and postmedial fasciæ of about three spots each; all three parallel, and farthest out at costa. Fringe with a series of dark bars in base, and preceded by a fuscous terminal spot.

Larva on oak.

Only a single specimen seen, in National Museum, and marked "rileyella type." This is possibly the same as *E. gallægenitella* Clemens, of which I have not studied any authentic material.

5. E. sylvicolella Busck. Fore wing dirty white, thickly dusted and shaded with fuscous, and marked with black. No terminal dots. A raised, oblique, dark streak near base, farther out on costa, and broken into spots on costa, fold, and inner margin, sometimes obscure; another on costa two-thirds way out, defined outwardly by an outwardly oblique white line; and three bars along cell, sometimes running together into a line or obscure. 9 mm.

New York (Busck); Kentucky; Texas.

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27. TRYPANISMA Clemens

Body and hind wing similar to Recurvaria. Fore wing with M, closer to M. M_a and Cu_1 stalked. Hind wing with R and M_1 stalked, R running to apex

Key to the species

Head	white .			 	 	 2. fermia.
Head	suffused	l with	fuscous.	 	 	 l. prudens.

1. T. prudens Clemens. Head pale yellowish white; vertex with large, blackish spots on most of the scales; second segment of palpus with two dark brown spots on the outer side, or wholly dark, and third segment with rings. Automos slightly annulate. Thorax and fore wings powdery blackish, tinted with yellowish, darker than T. fagella, with a small ochreous dot below base of costa; a patch on middle of disc, often reaching inner margin, preceded by black dots in the cell, and a postmedial band, strongly angulated or broken into costal and dorsal burs

and a postmedial band, strongly angulated or broken into costal and dersal bors as in *T. fagella*. Hind wing fuscious. 9 mm. (quinqueannulella Chambers.) May. The caterpillar forms a web on the under side of an oak leaf near the tip, communicating by holes with the upper side where the caterpillar for 14. It occurs in July and August. The pupa is formed in a slight cocoon in the leaf District of Columbia; Pennsylvania; Ohio. 2. **T. fagella** Busck. Head white. Antennæ annulate with fuscous. Palpi with only one faint annulation on second segment, and two on third. Thorax and fore wing nowdery light gray. Two circular blacking spots (at middle and and of cell)

wing powdery light gray. Two circular blackish spots (at middle and end of cell) in a cream-yellow shade, which reaches the inner margin at the middle; a larger diffuse oblique streak at base of costal fringe and a spot on the inner margin opposite it, both edged beyond with cream. Hind wing, with fringe, and abdomen light silvery gray. 9 mm.

Caterpillar similar to T. prudens; on beech. District of Columbia.

28. EVIPPE Chambers

(Phaëtusa Chambers)

Similar to Epithectis and Recurvaria. Fore wing with all veins present; M_z and M_s connate or more often stalked; Cu_1 free, halfway between M_s and Cu_z , and equally long. Hind wing (fig. 162) nearly as wide as fore wing, with slightly produced apex, R sinuous and running to apex, M1 lost, cell very weakly closed or open. M_2 close to M_3 , which is almost connate with Cu_1 .

The moths are superficially similar to Recurvaria dorsivittella, but have cleanercut markings and lack the two black palpal bands of Recurvaria. 1. E. prunifoliella Chambers. Palpi white, with base of second joint and extreme

tip of third blackish. Head and thorax white, with a blackish shade above antennæ, and black tegulæ. Fore wing black, shading into dark gray on costa, powdery toward apex along outer margin. Inner margin broadly white, with a twice-waved upper boundary, which becomes diffuse at apex. A subterminal costal spot, more or less distinct. 9 mm.

May to June. Caterpillar under the folded tip of a leaf of plum or peach in late fall; apparently a leaf-miner when young; rarely on soft maple.

E. leuconota Zeller, a much smaller species, without any trace of the costal pale spot, has been reported from Albany, New York.

29. SITOTROGA Heinemann

Antenna with a sparse pecten, otherwise practically like Metzneria. We have only one, probably introduced, species, of Oriental affinity; the Angoumois grain moth.

1. S. cerealella Olivier. Fore wing silky luteous yellow; hind wing fuscous, fringes brownish fuscous. Ordinarily without markings, though some moths show a black discal dot and a streak in the fold, and some, streaky black scaling. 15 mm.

(Anacampsis; Gelechia). Caterpillar stout, legless, white, with yellow head, breeding continuously in stored grain. The moth flying at large in August. New York: Geneva, Ithaca, Albany; Smithtown, Long Island.

30. METZNERIA Zeller

(Parasia)

 R_4 shortly, and R_5 long-stalked, with M_1 (fig. 170); the other veins separate. Hind wing with all veins widely separate, R and M_1 somewhat divergent, M_1 and M_2 twice as far apart as the others; anal region much reduced.

This is a European genus of some size. We have one species probably introduced with its food plant.

1. M. lappella Linnæus. Fore wing dull olive brown with paler yellowish shades and streaks. Hind wing dark fuscous gray. 12-15 mm.

May to August. Caterpillar stout, white, with black head and divided brown cervical shield; legless; in burs of burdock, eating out the seeds and wintering as larva.

Europe; Canada to Pennsylvania. New York: Peru, Newcomb, Lewiston, Ithaca, Katonah.

31. PTYCERATA Elv

(Pyncostola Meyrick (?); Paltodora Meyrick, in part, not typical; Cleodora Curtis, not Peron and Lesueur, etc.)

Palpus typically with a long triangular terminal scale-tuft on second segment; in subgenus Isophrictis Meyrick with a looser brush of spreading hair, very loose in anteliella and busckiella. Hind wing with R and M_1 approximate or connate; otherwise like Metzneria.

Key to the species

1. A white oblique costal streak running down from beginning of costal fringe. 2. Head cream white.....l. striatella.

2. Head and thorax brown..... 1. No such streak.

2. Entire wing with white-tipped scales and longitudinal ochreous streaks.

3. similiella.

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...4. busckiella. 2. Whitish, with two or three blackish dots.....

1. P. striatella Hübner. Brown; a fine white streak extending obliquely across apex, with two white costal dots beyond it; an oblique whitish streak at anal angle, and more or less distinct pale longitudinal streaks. Fringe with a black bar across apex. Hind wing fuscous. Head, including palpi and more or less of thorax, contrasting, whitish. 12 mm.

September. Very likely this species is rather generally distributed with its food, tansy, but I find no authentic records.

Europe.

2. P. anteliella Busck. Head and thorax pale fawn color; concolorous. Fore wing darker reddish brown, two black streaks toward base, and a black discal dot; a fine white line across apex and a white bar in fringe at apex.

When fresh, this species shows the triangular palpal tuft, but it is easily lost. 12 mm.

New Jersey; western Pennsylvania.

3. P. similiella Chambers. R and M_1 of hind wing shortly stalked; palpus rough. Fore wing with the yellow streaks irregular and anastomosing, leaving a powdery band around the apex. Superficially, practically like the species of Phthorimœa. but distinguishable by the venation and the more bushy palpus. (Isophricits Braun; solaniella Chambers, in part, not larva; piscipellis Zeller.)

The larva breeds in heads of *Rudbeckia hirta*; pupating in the stem. Moth, June to August.

Kentucky; Ohio; Texas; Oklahoma.

4. P. busckiella Ely. Palpus with a triangular tuft; M_3 of hind wing lower than in the other species, arising from middle of end of cell. Antenna yellowish; palpus with second joint brown, third white, brown-shaded; head and tegulæ white; thorax darker. Fore wing light clay-color; two or three small, more or less elongate, brown spots; hind wing smoky with paler fringe. 12-15 mm. June and July. This species looks like a Gelechia, but structurally belongs to

June and July. This species looks like a Gelechia, but structurally belongs to this series.

32. AGNIPPE Chambers

Palpi with second segment rough-scaled, third about as long. Fore wings with 10 or 11 veins, \mathbf{R}_1 absent, \mathbf{M}_1 long-stalked with \mathbf{M}_3 , or absent, \mathbf{Cu}_2 long. Hind wing slightly broader. trapezoidal, pointed at apex, \mathbf{M}_1 absent, cell open, \mathbf{M}_3 to \mathbf{Cu}_2 evenly spaced. Fore wing with a tuft of raised scales in middle of fold. Moth resting with tail raised, like many other Gelechilds. A reduction-form of Recurvaria.

Key to the species

Head dusted with black; \mathbf{M}_2 present.....l. biscolorella. Head all white; \mathbf{M}_2 lost.....2. fuscopulvella.

1. A. biscolorella Chambers. Antennæ annulate. Head yellowish white. Vertex dusted with brown at sides. Palpi mostly blackish, annulate. Thorax and base of fore wings yellowish, with a blackish brown patch on front of thorax. Fore wing blackish fuscous with bluish iridescence, middle to apex heavily dusted with brown and some white. Tuft small, black and white. 12 mm.

May. Larva possibly on Gleditschia.

Southern Ohio and Kentucky.

2. A. fuscopulvella Chambers. Head white. Palpi yellowish fuscous at base and near tip. Antennæ annulate. Thorax and wings whitish yellow, densely dusted with fuscous, duller than in the other species, leaving the black tuft contrasting. 11 mm.

April. Rare, only one specimen seen. Kentucky.

33. STEREOMITA Braun

Similar to Metzneria. Palpus with second segment thickened and slightly tufted beneath; third equally long. Fore wing with one dorsal vein lost $(M_2 \text{ or }$

Cu, ?); the remaining cubital widely separated from the other two, which are connate. Hind wing with \mathbf{R}_{a} widely separated from stalk of \mathbf{R}_{4} to \mathbf{M}_{1} ; \mathbf{R}_{1} from beyond middle. Hind wing half as wide, with M3 and Cu1 widely separated, M2 nearest M_1 , M_1 connate with R.

1. S. andropogonis Braun. Pale ochreous, dusted with dark brown. Head paler; palpus with a brown spot on outer side of second segment, and annulus on middle of third. Fore wing with the dusting denser at apex, gathering in longitudinal streaks, and usually forming two oblique postmedial streaks on costa; costal fringe ochreous, dorsal brownish, with brown terminal dots and a bar at apex. Hind wing pale brown, fringes reddish ochreous. 9 mm.

Larva in inflorescence of Andropogon scoparius, showing its presence by a yellowish patch in the flower-spike. Moths in August, flying at dawn and evening, and resting head down. Clermont County, Ohio.

34. HELICE Chambers

(*Theisoa* Chambers; *Cacelice* Busck)

Palpus with third segment very long, much longer than the second, which is nearly smooth, curved, and fusiform. Fore wing lanceolate, with R_a lost in the male, usually stalked in the female, M2 lost, M1 long-stalked, and M3 and Cu1 stalked, with more or less distinct gray scale-tufts. Hind wing half as wide or a little more, lanceolate or notched below the apex, always very narrow. Sc twofifths length of wing, R often lost at base or entirely lost, and M₁ running to just below the apex, (male) or with R and M_1 represented by a single free vein running just above the apex (female); M₂ lost; M₃ and Cu, stalked (male), or cell open above M_3 (female).

The extraordinary sexual dimorphism in venation has had a curious effect on the synonymy of this genus. It is the most reduced of the recognized Gelechidæ.

I. Female with apex of hind wing produced, and outer margin concave below it (Helice).

1. H. pallidochrella Chambers (fig. 173 \mathcal{Q}). Light gray, powdery, a small costal dot near base, a large dark antemedial spot, crossing the cell but not crossing A, a third way out; usually with a smaller costal spot at three-fifths way out, a subterminal shade on costa and a complex of dark streaks across apex of membrane. 9 mm. (gleditschiæella Chambers; & Cacelice permolestella Busck). Larva probably on Gleditschia. Moth in May and late summer.

'Kentucky; Maryland; southern Ohio.

II. Female with lanceolate wings as in male (Theison).

2. H. constrictella Zeller (fig. 171 3, 172 2). Light wood-brown, shaft of antenna lighter, annulate with dark brown. Fore wing darker at base, especially before the antemedial line, which is excurved evenly, except for a sharp inward turn in at costa, and followed by a paler shade. Sometimes with a black discal dot; a blackish costal spot at two-thirds way to apex, followed by a pale one, the boundary sharp and oblique, sometimes extending into a short fascia. Dorsal fringe paler. 8 mm. (bifaciella Chambers).

May to July. Larva on elm, forming a web on the under side of the leaf, connected by a whitish tube of silk and frass to the base of the petiole; spinning a thin, oval, silken cocoon. July and September.

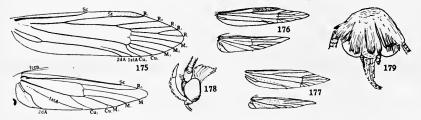
Massachusetts to southern Ohio and Texas.

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Family 22. BLASTOBASIDÆ

(Gelechiidæ, in part)

Head smoothly scaled, with the scales long and curving down over face, often diverging and covering the face and the base of the antennæ with a fanlike tuft (fig. 179). Antennæ long, scape broad, with a heavy pecten more or less mixed with scales (fig. 178) sometimes forming a definite eye-cap (Calosima). Shaft with two bands of scales to each segment, one more broadly interrupted below than the other; sensory hairs strong, especially in males. Many of the Blastobasinæ have the shaft notched at the base in the male. Palpi typically smoothly scaled and upturned beyond the vertex (fig. 178); reduced in the Pigritiinæ (fig. 179), never reaching beyond the middle of the front, but smoothly scaled or nearly so; in the latter often modified, with a specialized sensory area in the male. Tongue strong, scaled at base. Fore wing with \mathbf{R}_1 arising well toward base of wing, \mathbf{R}_2 near apex of cell, usually more than nine-tenths way out on the cell and much shorter than \mathbf{R}_1 ; \mathbf{R}_3 arising from upper angle of cell; \mathbf{R}_4 and \mathbf{R}_5 long-stalked, usually arising from end of cell, and well separated from \mathbf{R}_3 , \mathbf{M}_1 free, \mathbf{M}_2 to Cu₂ arising in a group from lower angle of cell, Cu₂ starting off at



FIGS. 175-179. BLASTOBASIDÆ

175, Holcocera purpurocomella (?), venation; 176, Blastobasis phycidella (Europe), venation; 177, Pigritia species, venation; 178, side view of head of Blastobasinæ; 179, front view of head of Pigritia, showing minute palpi and imperfect eye-cap.

right angles, and usually running straight across to inner margin, perpendicular to \mathbf{M}_2 ; \mathbf{M}_3 and \mathbf{Cu}_1 intermediate in direction. \mathbf{M}_2 and \mathbf{M}_3 often stalked; 1st A developed at margin, sometimes arising from 2d A near its tip; 2d A normal and forked at the base also. Fore wing with a thickened stigma between \mathbf{R}_1 and costa. Hind wing narrower than fore wing, Sc and R normally fused for a short distance at base; R and \mathbf{M}_1 well separated, parallel; \mathbf{M}_2 closely associated with Cu-stem; \mathbf{M}_3 often stalked with \mathbf{M}_2 or \mathbf{Cu}_1 or lost; \mathbf{Cu}_2 normal, much farther from end of cell than in fore wing. Anal

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region reduced, with more or less traceable veins. Fringe two-thirds to one and a half times as wide as membrane. Hind tibiæ with long hair; tarsi normal, with weak spinules. Female ovipositor slender. usually exserted.

Larvæ known only in the Blastobasinæ; prolegs short, with a complete series of uniordinal hooks; otherwise like the Ecophoridæ, and not distinguished in any way from those of Endrosis; tubercles i and ii distant, iii of eighth segment of abdomen above and behind spiracle; ocelli reduced. Pupa of Gelechioid type; not studied.

A small family, derived from the scavenger group of the Ecophoridæ, from which they differ only in the fusion of Sc and R of the hind wing, the rather heavier pecten, the closer grouping of the veins at the end of the cell of the fore wing, and the stigma. Endrosis is sometimes also included here, but is Ecophorid in the characters named.

The species are all obscure and very imperfectly known; the few known larvæ are borers in nuts (often scavengers after other borers), scavengers, and predaceous on scale-insects.

Key to the genera

- 1. Palpus not reaching beyond middle of front, often rudimentary; hind wing with 7 veins, Ms completely fused with Cu, (Pigritiinæ).
 - 2. Fore wing with R₃ stalked with R₄₊₅.....9. Pseudopigritia.
 - concealed in a groove in the front in life 6. Ploiophora.
 - 3. Fore wing with M, and M, not stalked; palpi minute in both sexes.

8. Dryoperia.

1. Palpus upturned beyond vertex; male. often with a notched antenna (Blastobasinæ).

2. Hind wings with seven veins; M₃ and Cu₁ fused (fig. 176)...5. Blastobasis.

4. Antenna with eye-cap.....4. Calosima.

1. EURESIA Dietz

Male antennæ deeply notched, not heavily ciliate. Fore wing with all veins present, all but \mathbf{R}_{4} and \mathbf{R}_{5} free; hind wing much narrower, all veins free and widely separate, lower discocellular vein nearly longitudinal and long, as well as m-cu.

1. E. pulchella Dietz. Fore wing light ochreous, trisected by two vertical whitish shades, the first broader and the second followed by a strong dark shade. 81/2 mm. June.

District of Columbia.

WILLIAM T. M. FORBES

2. VALENTINIA Walsingham

(Holcocera auct., not Clemens; Auximobasis Meyrick, in part)

Male antenna in our species deeply notched at base, scape broad, with a weak peeten. Fore wing normal, with \mathbf{M}_2 and \mathbf{M}_3 separate; hind wing with \mathbf{M}_2 and \mathbf{M}_3 stalked; nearly as wide as fore wing, with fringe rather wider than membrane.

1. V. glanduella Riley (The acorn moth). Light gray, more or less powdery and mottled; antemedial fascia moderately angled out on Cu, the following blackish shade broken there, so that there appears to be an extension outward of the antemedial line between its two halves. A blackish dash toward base below fold; erect costal and dorsal bars just beyond the distinct small black discal dots, and fairly defined on inner side. Margins more or less edged with blackish at apex; first discal dot also distinct in light specimens. Hind wing much yellower, olive gray, 15-25 mm.

July. Larva in acorns, apparently sometimes a scavenger after the acorn weevil. Distribution general. New York: Ithaca. 2. V. quaintancella Dietz. Much smaller than V. glandulella; hind wings nar-rower, fringes twice as wide as membrane. Duller, brownish, more evenly powdery; antemedial line followed by a strong continuous angulate dark shade, the middle part formed of the large diffuse first discal dot; outer discal dots smaller and well marked, the outer markings obscure. Moth resembling a Pigritia. 10-14 mm.

Larva in mummy apples.

Distribution uncertain.

3. HOLCOCERA Clemens (restricted)

Similar to Valentinia, except as noted in key. The notch varying in development and not always present.

The following key is incomplete.

Key to the species

1. Ground white, marked with black.

2. A complete heavy black antemedial band including first discal dot.

3. confluentella.

2. Antemedial band separated from orbicular; or band lighter brown. 3. No black powdering; a distinct dark streak on R and no antemedial band.

17. melanostriatella.

3. Heavily black-powdered; antemedial band present; dash, if present, short, clean-cut, and confined to antemedial region.

4. All black and white; black dashes conspicuous......13. elyella

4. With brown base of costa and antemedial shade, or suffused with gray. 7. busckiella. 1 Ground luteous, notch moderate

1. Ground luteous; notch moderate.
2. With strong, complex, and contrasting marks
2. Nearly evenly powdered
1. Ground reddish buff10. villella
1. Ground gray.
2. Male antennæ with deep notch; cilia as long as width of shaft.
3. Wing-scales not white-tippedl. dives
3. Scales white-tipped
2. Male antennæ with moderate notch; cilia half as long as width of shaft.
3. Larva on apple; moth immaculate except for three discal dots.

8. maligemmella. 3. Larva on chestnut; moth dark gray, but with ground contrasting luteous on basal third and here and there outwardly.....6a. minorella.

3. (H. quisquiliella also belongs in this group.)

2. Male antennæ without notch.

- 3. Even, dark mouse gray, with faint darker dots.....ll. funebra. 3. Paler, more or less dusted with white.
 - 4. Basal dash present.....14. zelleriella. 4. No basal dash.
 - 5. Evener, antemedial band distinct toward costa and inner margin, and pointing well before orbicular spot.....punctiferella.
 - 5. Rougher looking; antemedial band, when distinct, with its two halves pointing directly to orbicular.....16. tartarella.

I. Notch on male antennæ deeper than width of shaft, outwardly distorting the shaft; cilia as wide as shaft.

1. H. dives Dietz. Mouse gray, the costa distinctly browner; black discal dots only distinct; outer margin and fringe vaguely barred; no white-tipped scales. 16-19 mm.

July.

Toronto, Ontario; New York; Pennsylvania; Indiana. Var. basipallidella Dietz has a distinctly paler basal two-fifths, with angled outer

boundary; and the first discal dot indicated at its angle.
New Hampshire; Cohasset, Massachusetts; Pennsylvania.
2. H. aphidiella Walsingham. Similar to H. dives, many scales white-tipped, defining a vague pale antemedial region; base and middle of costa solid dark; also orbicular and discal dots, some apical shades, and a vague bar on Cu outwardly. 13 mm.

July. Larva in a Phylloxera gall on Carya.

District of Columbia; New York: Rock City.

3. H. confluentella Dietz. Ground pure white, crisply dusted and mottled with black. Base black, thorax with black anterior and posterior bars; antemedial fascia broad, complete, covering orbicular, angled out at middle; discal dots and postmedial bands fused into a Y-shaped black fascia, which may break into two costal spots and a discal and dorsal bar, or form two anastomosing bands, enclosing a pale spot beyond end of cell. Some black terminal suffusion.

June and July. Larva on pitch pine, associated with Rhyacionia frustrana and Recurvaria.

Central New York; Pennsylvania; Massachusetts.

II. Notch moderate, not as deep as width of middle part of shaft; shaft stouter, with cilia about half as long as width of segments.

4. H. purpurocomella Clemens. Mouse gray, dusted with paler and darker, with distinct first and second discal dots only; very slight purple iridescence. 18 mm. June.

Pennsylvania; Maryland. New York: Rock City, Vandalia. 5. H. ochrocephala Dietz. Mouse gray. Head and collar clay-color; strongly contrasting; middle half of thorax clay-color; front and back grayish; tegulæ clay-color, with gray spot and dusting; base of fore wings clay-color, the wing with obscure orbicular and outer discal dots, not defined with pale. 16 mm.

August. Type only seen.

Maryland; West Virginia.

6. H. chalcofrontella Clemens. Typically clay-color, more or less dusted with blackish; orbicular a black dot, with a blackish shade before it on costa and two obliquely placed and more or less confluent patches just beyond it on dorsal half, below Cu; two black discal dots, a blackish patch between the lower one and its anal angle; a curved blackish band half way between end of cell and apex, curving in below along outer margin and reaching nearly or quite to the anal patch. More or less distinct blackish terminal dots around apex. Fringe and hind wing

mouse gray. Outer markings sometimes replaced by a continuous, less-contrasting postmedial line, and the inner by an antemedial one interrupted over the cell. 12-16 mm.

Var. fumerella Dietz is nearly even mouse gray, powdered in two shades, the marks as in the type, with a continuous strongly curved postmedial line. Var. quisquiliella Zeller is pale, more evenly and less contrastingly powdery; the marks all obscure except the three discal dots (perhaps it is a good species).

General. Larva in sumac heads. Moth in July.

New York: Ithaca.

H. minorella Dietz is probably a good species. It is suffused with dark gray, except in irregular patches and on the basal third, the three discal dots contrasting. The larva lives in chestnut burs.

7. H. busckiella Dietz. White, more or less dusted with black and suffused with gray, but always with some pure white scales. Normally with base of costa light brown, antemedial fascia blackish on costa, weak or broken on cell, light brown and rather distinct below; orbicular a black dot, and rather beyond it a blackish patch resting on costa between orbicular and reniform, often strongly contrasting. Two normal discal dots; postmedial fascia with its ends just beyond the outer discal dot, its middle curving out nearly half way to apex, the fascia normally heavy and blackish. Terminal dots light. 16-20 mm.

July and August.

Plummer's Island, Maryland.

8. H. maligemmella Murtfeldt. Wings broad (fore wing less than four times as long as wide), light yellowish fuscous, with the discal dots distinct; otherwise practically immaculate. 14 mm.

Moth in July. Larva in buds of apple.

Columbia, Missouri.

9. H. gibbociliella Clemens. Luteous, lightly dusted with blackish, a little more heavily on outer half, leaving costa paler at two-thirds way out, and outer margin paler on both costa and dorsum; discal dots obscure, the orbicular rather more distinct, rarely absent. Head and thorax concolorous. 11-15 mm.

Atlantic States.

10. H. villella Busck. Unicolorous light yellowish brown, immaculate; palpi blackish on outer side, hind wings grayer, with yellowish fringes. Tarsi marked with blackish on outer side. 15 mm.

Larva on Andromeda ligustrina.

Maryland. New York: Ithaca.

III. No antennal notch, cilia normally short.

11. H. funebra Dietz. Mouse gray, slightly powdery, showing the usual three discal dots only. 12 mm.

Var. reductella Dietz is paler, less shining, and with the basal third slightly paler. Manitoba.

May to July.

Maryland; Pennsylvania. New York: Rock City.

Dives, aphidiella, funebra, and purpurocomella are very similar, practically identical in the female, but funebra is rather strongly frosted with white-tipped scales and the third joint of the palpus is notably shorter than the second (threefourths as long), while in the other three the segments are nearly even in length; purpurocomella and dives are larger and noticeably stouter than the other two (expanse normally over 15 mm. H. dives is the largest and purplest, and shows only the faintest trace of discal spots.

12. H. boreasella Dietz. Fuscous, the inner margin below the cell somewhat contrasting, whitish, crossed by an oblique fuscous costal shade from the middle of cell to the inner margin at a third way out. Usual dots diffuse and obscure. No

other distinct markings. The antemedial brown shade bends at an angle on the cell and is obscurely continued to the costa.

August. (One cotype only seen.) Montreal, Canada; New Hampshire.

13. H. elyella Dietz. White, lightly powdered with black; black dashes near base below Sc, and in fork of A, orbicular and discal dots horizontally elliptical, contrasting, black; a blackish shade in fold below orbicular, and one before it near costa; some dark gray shading outwardly near margin. Usually with distinct dashes between the radial veinlets beyond orbicular and upper discal dot. Hind wing light. 13-16 mm.

July and August.

Dublin, New Hampshire, to Maryland.

14. H. subsenella Zeller (as determined in United States National Museum). Pale powdery ash gray, the margins contrasting dark gray, especially outwardly; antemedial fascia moderately excurved, interrupted over cell, running well before the orbicular, which is a dot; two discal dots and one on Cu half way out from orbicular to lower discal dot; no other definite marks. 11-14 mm.

August.

Pennsylvania; Texas.

H. zelleriella Dietz, described from Texas, Iowa, and Louisiana, may reach our area. It is similar but with one or two dark dashes at the base, the stronger one in the fold.

15. H. vestaliella Dietz. White; antemedial fascia indicated by dashes in cell and fold, and shades on margins, angled out at middle or reduced, and pale brown, sometimes nearly absent; a dash beyond it on lower side of cell. Discal dots conspicuous, with a black spot below them and a small dash above. Middle of costa shaded with brown; outer margin somewhat shaded with black.

June and July.

Southern Massachusetts; Maryland; Pennsylvania.

16. H. tartarella Dietz. Mouse gray, somewhat powdered with white; occasionally with contrasting whitish basal third; antemedial fascia angled out, formed of separate shades running to costa and inner margin, and an orbicular dot; sometimes also with a separate dot in the fold. Discal dots and dash on Cu distinct; the other marks obscure. Thorax concolorous, light gray. Marks in the darkest specimens defined with whitish.

End of May to July; September.

Plummer's Island, Maryland.

17. H. melanostriatella Dietz. Pearl gray, somewhat shaded with light gray, but not dusted with black; a dark gray shade, variable in distinctness, on base of upper edge of cell, and sometimes at middle of wing. Orbicular black. Discal dots, at least the upper, black; the other marks gray and slight. 15 mm. (melonostriatella Dietz, misprint.)

August.

Southern Connecticut; Pennsylvania; Maryland.

The following species of Holcocera are wholly unknown to me; some may be synonyms.

H. modestella Clemens. Ground even, fuscous; basal part of fore wing paler and grayer; a dark fuscous orbicular and two discal dots. The outer boundary of the pale base angled out but not reaching the orbicular (group II). 16 mm.

Atlantic States.

H. inclusa Dietz. Similar to H. modestella; gray with paler base, and excurved antemedial fascia not reaching orbicular, but without antennal notch. No dot outward on Cu. Ground nearly white, but heavily overlaid with gray; no basal dashes. 15 mm.

Pennsylvania.

H. messelinella Dietz. Gray, sprinkled with whitish scales; thorax whitish, dark brown in front; fore wing with antemedial fascia strongly angled, partly breaking into costal and dorsal dashes; orbicular spot with two dashes before it, above and below fold; postmedial fascia dark, angulate. 15 mm.

Maryland; Florida.

H. m. var. spoliatella Dietz. More glossy; marks less distinct; the basal streaks obscure; front of thorax concolorous. 12-13 mm.

New Jersey; Pennsylvania.

H. clemensella Chambers. The unique type is now greasy, but this seems to be a white species.

Kentucky.

H. illibella Dietz. Gray, dusted somewhat with white and fuscous. Antemedial fascia reduced to dots on margins; orbicular and discal dots more distinct; postmedial fascia obsolete. 12-13 mm.

Maryland.

H. punctiferella Clemens. White, with five black bars, arranged 2, 1, 2; ground dusted with gray toward apex, and shaded with yellowish flesh color in fringe; and fringe at least of hind wing gray. (The five bars are the orbicular and one nearly opposite it in the fold, a spot on Cu, and the two discal dots.) 12 mm. Pennsylvania.

H. triangularisella Chambers. Grayish fuscous; third segment of palpus whitish. Antemedial fascia a broad brown-powdered whitish triangle, resting on the inner margin, followed by a dark brown triangle, resting on the costa. Outer part of wing powdered on a whitish ground; discal dots minute. 18 mm.

Kentucky.

4. CALOSIMA Dietz

Like Holeocera but with fully scaled eye-cap in place of a pecten on the antenna. Fore wing immaculate.

1. C. argyrosplendella Dietz. Yellowish white. Head white, shining; palpi dusted outwardly with fuscous. Antennæ gray; eye-cap white. Fore wing shading into pale reddish brown outwardly; hind wing pure white, nearly as wide as fore wing. 11.mm.

July.

Penusylvania; Louisiana; Florida.

5. BLASTOBASIS Zeller

(With *Epistetus* and *Prosthesis* Walsingham)

Similar to Holcocera, but with M_3 and Cu_1 completely united. Our species have male antennæ without a notch; cilia of male antennæ as long as width of shaft; fore wing with M_2 well separated, M_3 , Cu_1 , and Cu_2 connate; hind wing with M_2 short-stalked (fig. 176).

1. B. plummerella Dietz. Ground powdery smoky brown, strongly shot with purple; nearly even, but with a paler shade across the disc at two-fifths way out, and one above anal angle. No definite markings as a rule; the most definitely marked specimens with a small black orbicular and two discal dots. 10-14 mm. July.

Maryland. New York: Rock City.

Var. fuscopurpurella Dietz is large and practically immaculate, with the purple rather strong, and the dark element of the ground strongly predominant.

2. B. sagittella Dietz. Gray, speckled with dark brown; a pale antemedial fascia, a little nearer the base at costa, angled out, and sharply extended along Cu in a tooth; a heavy dark shade on its outer side. Base paler, edged along both

margins with blackish, and with a weaker streak below the fold. Outer part of wing a little darker along costa, with distinct discal dots and terminal dots. 11 mm.

August. Unknown to me.

Hazelton, Pennsylvania. New York: Rock City.

6. PLOIOPHORA Dietz

Palpi of male closely upturned nearly to middle of front, lying in shallow grooves on the lower part of the face, and roofed over by the long, overhanging vestiture (usually falling forward at death); second segment with a large obliquely oval sensory area on inner side; third segment small. Female palpi and wing characters as in Pigritia.

1. P. fidella Dietz. Fuscous gray, smooth and shining; antemedial fascia nearly straight, diffuse, and very slightly inwardly oblique from cell to inner margin, bent on cell and slanting in to costa also; the fascia formed of pale dusting, and variable in strength; discal dots dark, obscure; outer third of inner margin and outer sixth of costa with scattered white-tipped scales, extending somewhat into fringe. 12 mm. June.

I cannot distinguish P. ampla Dietz by the one rubbed type I have seen; fidella is possibly a synonym of it.

Northern New Hampshire and Parry Sound, Ontario, to Pennsylvania. New York: Rock City.

7. PIGRITIA Clemens

(With *Epigritia* Dietz)

Palpi of male practically obsolete (fig. 179), and of female short and porrect, with third segment variable in length but generally shorter than the second. Antennæ not modified. Fore wing (fig. 177) with cell set obliquely in wing, even more noticeably than in the Blastobasine; Cu_2 short and running straight to inner margin. Hind wing sharply lanceolate, half as wide, with **R** and **M**, decidedly divergent, M_2 connate with Cu_1 , the cell very weakly closed. M_3 and Cu_1 rarely very shortly separate at extreme margin, normally fused. The genus Epigritia was based on the relative shortness of the female palpi, a character that has proved intangible.

Key to the species

- 1. Base of wing, except costal edge, contrasting whitish (late July to August). 2. Its outer boundary distinctly excurved......11a. heidemannella.
- 1. With a base of ground color at least twice as wide as the antemedial fascia (June).
 - 2. Antemedial band even, usually straight.
 - - Band moderate, extending a third way to base.
 With a complete dark discal bar; antemedial band broad and con-.....4. mediofasciella. trasting

2. No antemedial band, or only faint traces

- - 3. Head and palpi pale......10. obscurella. 3. Head and palpi dark; wings purplish......2. purpurella.

1. P. confusella Dietz. Fuscous brown, the antemedial band broad, pale, erect, faint and diffuse; some pale scales at anal angle. No other distinct marks. This form looks very much like P. fidella but is browner.

June.

Pennsylvania; New Jersey.

2. P. purpurella Dietz. Blackish, shot with purple; with more or less distinct traces of the usual markings. Head and palpi concolorous. Male unknown. 12 mm. June.

Pennsylvania. New York: Rock City. 3. P. laticapitella Clemens. Dull fuscous, about the color of P. fidella; the scales largely pale-tipped, especially on outer half of wing. Antemedial fascia prominent, straight, erect, pale (the curve shown in Dietz's figure is probably an artefact), fairly defined on outer side, but diffuse on basal. Base more solidly fuscous; a broad shade of solid fuscous beyond the fascia to middle of wing. Two blackish discal dots, sometimes obscure, with a bar below them in the fold, all defined by a well-marked pale shade; the apex again darker. Some whitish dusting in base of fringe. 12 mm.

June.

New Hampshire; Pennsylvania; Iowa; Kansas.

4. P. mediofasciella Dietz. Practically like the last form, the marks less contrasting antemedial fascia narrower, discal dots fused into a vague vertical bar, only faintly defined with pale; white-tipped scaling at apex well marked. 12 mm. July 1.

New Jersey; Pennsylvania.

5. P. basilarella Dietz. Pale silvery gray, dusted with pale fuscous. Head, thorax, and costal half of fore wing at base with golden iridescence. Fore wing with base paler, and a dark shade beyond the antemedial fascia; a dark streak on fold at a sixth way from base; fascia with a curved projection at the middle; discal dots strong, with a dot below them in the fold, and followed by a waved postmedial shade. Female less iridescent, with third segment of palpus shorter than second; first segment fuscous-dusted, second and third pale. 12 mm. (Not seen.) Pennsylvania; Louisiana; Kansas.

6. P. ornatella Dietz. Similar to P. laticapitella and mediofasciella, the white antemedial fascia striking, and perceptibly excurved on outer side, almost pure white; discal dots separate, dark, faintly set off; the whole apical region with white-tipped scales, paler than the base. 11-12 mm.

Hazelton, Pennsylvania.

7. P. angustipennella Dietz. Fore wing narrower (less than one-fifth as wide as long). Lighter mouse gray with a slight violet tint; antemedial band erect, vague, slightly paler. A vague pale area over end of cell with obscure darker discal dots. 11 mm.

Pennsvlvania.

The one type I have seen looks too similar to P. confusella.

8. P. tristella Dietz. Mouse gray. Antemedial band, of white-tipped scales, but quite narrow; discal dots practically obsolete, vaguely defined by the well-marked white-tipped scales on the outer half of the wing. All the scales with more or less distinct pale tips. 11 mm.

Hazelton, Pennsylvania.

9. P. spoliatella Dietz. Identical with Dryoperia grisella (of which it may be a venational aberration), except for the stalking of M_2 and M_3 . 11 mm. (Unknown to me.)

Hazelton, Pennsylvania.

10. P. obscurella Dietz. Fore wing broad, four times as long as wide, mouse gray, immaculate, with ochreous bronzy head and pale palpi. 11 mm. July 4.

Northern New Jersey; Pennsylvania.

LEPIDOPTERA OF NEW YORK AND NEIGHBORING STATES

11. P. ochrocomella Clemens. Basal third of fore wing nearly white, gray at the extreme base, with contrasting blackish costa and thorax; outer two-thirds blackish. more or less powdered with white and often heavily shaded over the end of the cell, leaving the dark discal dots contrasting, and a darker bar halfway between them and apex; the region beyond the antemedial fascia contrastingly dark. Dark specimens have only the discal bar defined with whitish. The outer boundary of the fascia is typically erect, but markedly curved in the usually darker var. heidemannella Dietz. (Epigritia Dietz; E. pallidotinctella Dietz.)

Late July and August.

Southern Connecticut to Pennsylvania. A pale form occurs in Missouri.

8. DRYOPERIA Coolidge

(Dryope Chambers, not Desvoidy)

Exactly like Pigritia except for the separation of M₂ and M₃ in the fore wing. Palpi minute in both sexes, a little smaller in the male; first, and often second, joint with a triangular tuft below, rough-scaled, third joint shorter, and sometimes very small.

Key to the species

1. Immaculate ochreous6. ochreella.

1. A distinct antemedial fascia with zigzag outer boundary, followed by dark.

- 2. Fore wing gray 1. grisella. 2. tenebrella.

1. D. grisella Dietz. Light powdery gray, the costa darker, especially toward base, the dark showing as powdering on a paler ground. Base dark; antemedial fascia zigzag, not reaching costa, diffuse inward, outwardly defined by a contrasting blackish shade which is strong at costa and forms a patch in the fold, but is practically interrupted in the cell. Ground outwardly pale, powdery; the two discal dots formed of the powdering, distinct but diffuse, and followed by a blackish shade across the apex. 12 mm.

April.

Pennsylvania; Missouri; South Dakota.

2. D. tenebrella Dietz is very close to the preceding species, of which it may be a variety; apparently it is slightly paler, with the dark antemedial shade weaker and not forming so distinct a spot below the cell. 12 mm. Pennsylvania; Kentucky.

3. D. fuscosuffusella Dietz. Pale ochreous, with dark fuscous dusting and shading, grayer than D. murtfeldtella. Antemedial fascia of white dusting followed by a blackish patch on the costa, the patch rarely extended diffusely toward the apex; a similar patch on inner margin. Outer part of wing normally dusted with white; with weak discal dots. 11 mm. (Not seen.)

St Louis, Missouri.

4. D. murtfeldtella Chambers. Dull light ochre, shaded with pale brown; antemedial fascia pale, rather narrow, toothed at middle, heavily shaded with brown except on the tooth, extending faintly across the brown costal edge. Base shaded with light brown; subterminal region also shaded with light brown; and discal dots faintly indicated by a red-brown shade, all somewhat dusted with whitish. 12 mm. (ochrocomella Dietz, not Clemens; erratella Dietz).

Pennsylvania specimens are duller clay color, with dark shading and dusting, the discal dots distinct, the antemedial dark shade excurved, interrupted and

preceded by a longitudinal dark shade in the middle of the wing, the apical dark shade most distinct on the margins.

New Jersey to Texas and Nevada. 5. D. discopunctella Dietz. Grayish ochreous; autemedial fascia moderately excurved, faint, defined by a darker shade beyond it on costa; orbicular a strong but diffuse dot; two fainter discal dots. 12 mm.
Hazelton, Pennsylvania. (Unknown to me.)
6. D. ochreella Clemens. Shining dull ochreous, dusted with reddish scales.

Antennæ grayish ochreous; palpi dusted with brownish; fore wing with apex red-der; costa reddish fuscous with traces only of the fascia (unknown to me).

Pennsylvania; Texas; Alabama; South Dakota.

9. PSEUDOPIGRITIA Dietz

Similar to Pigritia, with similar sexually dimorphic palpi. Fore wing with R_3 shortly stalked with \mathbf{R}_{i} and \mathbf{R}_{5} . **1. P. dorsomaculella** Dietz. Powdery gray; antemedial fascia transverse, pale

gray, broad, and vague, bent out a little at middle discal dots more or less distinct, in a vague pale shade. Similar to spoliatella. 11 mm.

Dietz describes this species as having three darker diffuse spots along outer part of dorsal margin.

June.

Pennsylvania; New Jersey.

2. P. equitella Dietz. Similar to P. dorsomaculella, but with markings fainter and without any dorsal spots. Fascia at a third way out from the base. 11 mm. July.

Hazelton, Pennsylvania.

3. P. fraternella Dietz. Similar to P. equitella, but with the fascia farther out, at two-fifths. 10 mm. Hazelton, Pennsylvania. (Unknown to me.)

4. P. argyreella Dietz. Silvery white, tinged with ochreous; with the fascia barely traceable, partially defined with darker. 11 mm.

Hazelton, Pennsylvania. (Unknown to me.)

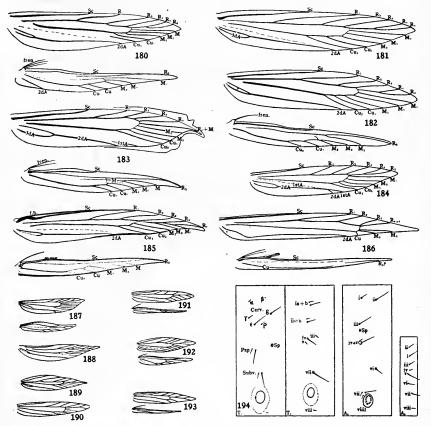
Family 23. LAVERNIDÆ

(Momphidæ; Cosmopterygidæ; Elachistidæ, in part)

Head smooth, the vestiture curving down over the face; eyes moderate or smallish; ocelli variable. Antennæ as in the Gelechiidæ; scape long and slender, often with a pecten, which is sometimes reduced to a single bristle at its base; never with eye-cap. Antennæ sometimes as long as the fore wing, never much longer or very short. Palpi divergent, with second segment smooth or tufted below, but never with a triangular tuft; third segment variable in length, smooth, upturned, slender, and regularly tapering; the palpus always upturned and much longer than the head.

Fore wing lanceolate, always narrow, the membrane rarely with a blunt apex; with broad dorsal and costal fringe. Anal angle never well marked: the apex occasionally caudate (fig. 183), as in the Lyonetiidæ, some Gracilariidæ, and Polyhymno. **R**, from cell less than three-

fourths way to the apex (except perhaps in Synallagma), but rarely arising before middle of the cell; \mathbf{R}_2 free, arising well before end of



FIGS. 180-194. LAVERNID.E

180, Walshia amorphella, venation; 181, Limnæcia phragmitella, venation of fore wing; 182, Perimede particornella. venation; 183, Lophoptilus eloisella, venation and outline of apical fringe; 184, Larerna species, venation of fore wing showing 1st A and 2d A anastomosing; 185, Cosmopteryx scribaiella (Europe), venation; 186, Batrachedra praangusta, venation (from a European specimen); 187, Psacaphora species, venation; 188, Stilbosis tesquella, venation of fore wing; 189, Stagmatophora graboviella (Europe), venation of fore wing; 190, Batrachedra trichella, venation of fore wing; 191, Eriphia albalineella, venation; 193, Eriphia concolorella, venation; 194, Limnæcia phragmitella, seta map of larva

cell (unlike the narrow-winged Blastobasidæ). \mathbf{R}_4 and \mathbf{R}_5 stalked, or rarely, united; \mathbf{M}_1 often, and \mathbf{R}_3 and \mathbf{M}_2 rarely stalked with them.

 \mathbf{M}_{s} , \mathbf{Cu}_{1} , and \mathbf{Cu}_{2} almost always parallel and subequal in length, from an oblique lower outer end of cell. 1st A rudimentary at the base, but frequently developed as a normal vein at the margin; this marginal piece being sometimes very short, and often apparently wholly separate rate from the rudimentary base, arising out of 2d A (fig. 184). 2d A forked at its base, the lower fork occasionally rudimentary. Hind wing narrower, lanceolate to linear, fringe one and one-half to eight times as wide as the membrane. A costal tuft of bristles toward the base, with the margin concave beyond it. Sc free from R, but occasionally connected by a cross vein (\mathbf{R}_1) . (In a couple of exotic species **Sc** is extremely short and \mathbf{R}_1 appears as a free vein running to the costa) **R** and **M**₁ more or less convergent at their origin; approximate (usually closely), connate, or stalked; \mathbf{M}_1 occasionally stalked with \mathbf{M}_{2} , and the cell often open below them; \mathbf{M}_{3} distinctly separated from Cu, usually connected by a long oblique cross-vein parallel to the dorsal margin. The anal region reduced, usually with rudiments of the normal veins: 2d A forked when distinct. (When the hind wing is linear the veins are much reduced and their connections often uncertain).

The caterpillars are quite insufficiently known; they include leafminers, stem-borers, seed-eaters, and, more rarely, leaf-rollers. Batrachedra and Pyroderces include some scavengers. A large proportion are associated with the Onagraceæ.

Caterpillar (fig. 194) with head small and depressed, adfrontal sclerites not quite reaching vertex, ocelli close together; body-setæ small; ninth segment of abdomen with setæ ii farther apart on middle line than they are from i, and i, ii, and iii in a vertical line; thoracic coxæ twice as far apart as their width, prolegs also far apart, with hooks in a complete circle, normally biordinal. Pupæ various, but normally with the clypeal suture complete, a quadrangular prothorax not narrowed at the middle, the palpi and the femora fully exposed, and maxillary palpi large, nearly or quite reaching the tongue. Antennæ meeting on the middle line well before the wing-tip, but not again diverging; cremaster with stout spines.

In Chrysopeleia the characters are nearly those of Aphelosetia, throwing some doubt on the position of the genus. In that genus the labial and maxillary palpi and the femora are wholly concealed; there are only a few short straight cremastral setæ, and the wing cases etc., are soldered to the body as far as the seventh abdominal segment, so that there is probably no motion; but the clypeal suture is as in the Lavernidæ. Cosmopteryx has the prothorax narrowing in the middle line, the maxillary palpi exposed, but the labials and femora concealed, the clypeal suture lost, and a single movable segment. Its cremaster has hooked setæ.

The family as it now stands is undoubtedly heterogeneous, being comprised of various reduced Gelechioidea, with perhaps a couple of genera which really belong to other superfamilies. The forms with 1st A preserved, or with a primitive pupa, are evidently correlated with the Œcophoridæ; others, like Ithome, seem to be derived from the Gelechiidæ by reduction; Batrachedra and Synallagma are most doubtful in position. A few genera of other families have been added to the kev for convenience.

Key to the genera

1. Hind wing immaculate; maxillary palpi minute, close-scaled.

2. Fore wing with one or more veins lost.

- 3. Cell set obliquely in the wing (fig. 186), squarely truncate at end, between \mathbf{R}_{1+5} and $\mathbf{Cu}_{1,1}$ cubital branches almost invariably running straight across to dorsal margin, much shorter than M₃.
 - 4. Fore wing with \mathbf{R}_4 and \mathbf{R}_5 stalked; hind wing narrow-lanceolate.

12. Blastodacna.

- 4. R4 and R5 completely united; hind wing linear (figs. 186, 190). 13. Batrachedra.²²
- 3. Cell central in the wing, the closing vein strongly oblique and nearly parallel to the dorsal margin from M_2 to Cu_2 ; M_3 to Cu_2 rather evenly spaced, subequal, and parallel.
 - 4. A radial vein lost; only four veins running to costa; fore wing falcate. 15. Synallagma.
 - 4. One or more dorsal veins absent; \mathbf{R}_1 always present.
 - 5. \mathbf{R}_{3} more or less stalked; \mathbf{M}_{1} often stalked.
 - 6. Wings broad; hind wing lanceolate and slightly trapezoidal, with two M's lost, and M_3 connate with $Cu_1, \ldots, (Gelechildæ - Helice.)$ 6. Wings linear; hind wing with all veins preserved.

(Heliodinidæ — Erineda.)

5. R₃ free; M₁ stalked.....ll. Chrysopeleia. 2. Fore wing with complete venation (cell always central, with oblique end). 3. M₁ stalked (fig. 185).

- 4. Scape of antenna as long as width of head, without a pecten; antennae longer, more slender. Fore wing with a silver streak in the apex; Cu_2
 - only four-fifths as long as fore wing, fore wing without a white streak in the apex; with Cu_2 arising more than three-fourths way out. 5. Fore wing with heavy, raised, metallic tufts; M_1 stalked nearly to

 - unless M₂ is also stalked.²³
 - 6. Wing dull, linear; pecten strong......7. Pyroderces. 6. Wing shining, lanceolate; pecten absent.

9 Stagmatophora, 10. Ithome.

²² Coleophora may be sought here, but is distinguished by its antennæ thrown forward in repose, its slender fore tibia without epiphysis at the middle; its upper spurs of the hind tibia above the middle, and its more angular-looking palpi. ²³ Laverna sexnotella also normally has M_1 very short-stalked, but the second segment of the palpus is much thickened, the wing has metallic tufts, and there is a weak pecten.

3. \mathbf{M}_1 free from \mathbf{R}_5 (fig. 184).

- 4. Hind wing with \mathbf{M}_1 and \mathbf{M}_2 stalked (fig. 183), in our species a third way to apex (when the cell is open, separating well beyond the separation of M₂ and Cu,).

 - 5. Fore wing not caudate. 6. Palpi smooth, **R** of hind wing running to costa less than three-quar-6. Palpi with second segment slightly tufted below; R of hind wing
- 4. Hind wing with \mathbf{M}_1 and \mathbf{M}_2 separate, rarely connate. 5. Scaling coarse and heavy, with large raised tufts; palpus tufted.

 - 6. Peeten normall. Laverna.
 - 5. Scaling fine and smooth; palpus smooth.
- 1. Hind wing linear with silver bars; maxillary palpi tufted. (Heliodinidæ-Idioglossa).

For an artificial key to the species related most closely to Laverna, see Laverna.

1. LAVERNA Curtis

(Mompha Herrich-Schæffer; Hübner, in part; Wilsonia Clemens)

Pecten well developed; palpi with a rough divided tuft on under side of second segment, third nearly as long; scape, tongue, and maxillary palpi normal. Fore wing (fig. 184) with \mathbf{R}_{4} and \mathbf{R}_{5} only stalked; \mathbf{M}_{3} , \mathbf{Cu}_{1} and \mathbf{Cu}_{2} equal, evenly spaced, and normal, from an oblique closing vein. Tip of 1st A normally appearing as a terminal fork of 2d A, more rarely free. A widely forked at base. Hind wing with R and M1*approximate at base, well separated from M2, which is associated with the Cu-stem; the wing lanceolate; hind wing, rarely, marrow with weak veins. Caterpillars mostly associated with the stems and fruit of Onagraccæ.

L. cephalanthiella Chambers is a Psaeaphora.

Key to the species

1. Head and thorax white.

2. White patch at base of wing continuous on inner margin to middle.

- 3. Apical fourth of fore wing dark 1. circumscriptella. 2. White basal patch confined to inner margin and extending to one-fourth only, followed by a smaller spot on inner margin.....3. murtfeldtella.
- 1. Head white, thorax blackish, contrasting,
 - 2. Fore wing evenly blackish with contrasting white and silver spots.

 - 2. Fore wing lighter brown, the seales white tipped.

(Psacaphora cephalanthiella.)

- 1. Head and thorax concolorous with fore wing. 2. Head and thorax, base of inner margin of fore wing, convergent faseiæ, etc., pinkish white, scaled with black......4a.
 - 2. Head, thorax, and general ground of fore wing wood brown. 3. Two black dashes on disc and smaller spots in fold5. brerivittella. 3. Tuft in middle of fold forming the largest black mark, no dashes. 4. stellella.

2. F	Iead	and	thorax	blackish.			decorella.
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1. L. circumscriptella Zeller. Head, including palpi, thorax, and basal half of fore wing, white; antennæ blackish, with tip of scape white. Outer half of fore wing typically rusty-brown (gray-brown in the more northern race). Costal edge brown all the way to the apex, broadly brown at base, the area widening into a triangle at a fourth way out and then very narrow to middle; a white costal fascia at three-fourths way to apex, typically connected by a curved white line with the white at the middle of the wing, where there is a black tuft above and below it. 9-12 mm.; larger northward.

June to October. Larva in seed-capsules of (Enothera, in August,

Southern States, north to southern New Jersey, Illinois, and Missouri.

2. L. definitella Zeller. Similar to L. circumscriptella; scape white; palpus with brown outer side of second segment; fore wing with the dark part fuscous brown, apical fourth white, with erect denticulate inner boundary, crossed by a blackish fascia from just below apex to anal angle, leaving a narrow irregular marginal white band; fringe gravish, with some white scaling at apex. 15 mm. (unicristatella Chambers; ruficristatella of the New Jersey list).

July and August.

Southern New Jersey; Texas.

3. L. murtfeldtella Chambers. Head, palpi, and thorax white, the palpus with two narrow black bars on the third segment. Fore wing powdery gray (black on white) and light wood-brown; a semicircular white patch on basal fourth of inner margin, with a black tuft on its outer edge, a smaller patch before middle; with a large black tuft, and a small white and black tuft in fold at three-fourths way to apex; small black and white tufts at middle and end of cell, the outer one sometimes extended into a black streak, and a similar black streak in the discal fold beyond it, opposite a white costal spot.

October. Larva in buds of (Enothera; dull brownish green, shaded with red, with brown head and plates.

Kentucky; Missouri; Colorado; Texas. 4. L. stellella Busck. Similar to L. brevivittella but more mottled looking; antennæ unicolored, dark brown; palpi powdery, with a black ring near tip of third segment. Head and thorax light ochroous; face silver white; fore wings light ochroous mottled with brown and black scales, costal edge evenly mottled with black and apical part evenly sprinkled with black scales; two indistinct oblique light brown shades crossing the wing, from base and middle of costa; six raised ochreous tufts, in cell and below fold, the middle one of the lower row the largest, terminating in black scales, forming the most contrasting markings.

August to October, more rarely in the spring. Larva in Œnothera; in the buds, flowers, and seed-capsules; maturing in August. Light yellow, transversely banded with pink; with brown head.

Ontario to District of Columbia.

L. ______. Head, thorax, and fore wing gray (black powdering on a white ground. Palpus with second segment scaled with gray and black; third segment black, with faseiae: the base lighter out to the first faseia, but half covered with a larger triangular dark brown area resting on the costa. Antemedial fascia defined beyond by a couple of black tufts; outer three-fifths of wing darker, and heavily scaled with wood-brown and chocolate brown; with a black shade through outer part of cell, and a vague one beyond the postmedial line to the apex; postmedial line followed by black scales on the discal fold and a black tuft in the submedian fold. 9 mm.

Octoher to April.

New York: Ithaca,

5. L. brevivittella Clemens. Wood-brown, the scales of fore wing and thorax finely white-tipped, those of head mostly solid brown. Palpns grayish with a heavy black har on third segment and black tip. Fore wing nearly evenly colored, but shaded with graver and yellower brown; short, longitudinal black dashes in fold near base, in cell at middle, and in middle of wing toward apex, each with some raised scales; with raised, black dorsal spots opposite the first two dashes, and a little before the third. 11 mm. (*ænotherivorella*, *ænotheriseminella* Chambers.)

October. Larva in seed-capsules of Enothera.

New Jersey to Missouri. New York: Ithaea, Albany.

6. L. sexnotella Chambers. M_1 shortly stalked, but pecten and tuftings as in Laverna. Brown-black; head white; palpus with gray at base, two black rings on third segment; thorax with some white scales; scape white-tipped, and apex of antennae with five white rings on alternate segments. Fore wing with an oblique white bar from the costa near base, an oblique triangle at middle, and a large triangular white spot before the apex; raised dorsal silver spots opposite the first two and just before the third; some white terminal dots, the largest at apex. 12 mm. (Stagmatophora.)

May to August. Caterpillar a gall-maker in stem of *Trichostema dichotomum* (a mint).

Massachusetts to Ontario and south.

7. L. wyattella Barnes and Busck. Similar to L. sexnotella, larger; middle and outer spots on inner margin of fore wing connected by white scaling; and white markings as a whole rather more extensive; hind wing much darker, purplish black with a purple sheen. 13-16 mm. (Stagmatophora Barnes and Busck.)

Chicago, Illinois.

8. L. decorella Stephens. Black-brown, with two narrow, broken, transverse, white fasciae and some scattered white spots. 11 mm. (unifasciella Chambers).

August and September. Larva light gray-green, with blackish head and brown cervical shield; in a stem gall on Epilobium, in July and August.

Quebee to West Coast. Eurasia.

2. LIMNÆCIA Stainton

(Lymnæcia auct.; Laverna, in part)

Head smooth; eyes large; scape slender, almost as long as width of head between insertions of antennæ; pecten of few bristles, fugitive. Palpi smooth; third segment twice as long as seeond. Fore wing lanceolate (fig. 181) as in Laverna, but with rudimentary tufts in cell only; 1st A completely lost. Hind wing two-thirds as wide; venation as in Laverna, but with eubitals relatively longer and closer together at origin. Fringe nearly twice as wide as membrane. A very distinct genus of Australian affinity; our species cosmopolitan with its food plant.

1. L. phragmitella Stainton. Clay-color. Palpus with two slender black lines on third segment; scape with a black line; shaft of antenna with a series of black dots. Fore wing with small black discal dots, ringed with white, and a weaker black point or dash in the fold; fringe barred with cream. Hind wing light gray with nearly white fringe. 20 mm.

Of general distribution, emerging in early July. Caterpillar whitish, with yellower head and several longitudinal pink stripes; eating in the seed-heads of eattail, and causing them to fluff out; wintering half grown. One brood.

New York: Lewiston, Ithaca.

3. WALSHIA Clemens

(Mompha auct.; Sorhagenia Spuler)

Similar to Laverna; peeten reduced to a single bristle. Palpus with second segment practically smooth, but fore wing heavily tufted. Fore wing (fig. 180) with venation as in Laverna, but Cu_1 nearer Cu_2 than M_a at origin; Cu_2 partly lost, easily

mistaken for 1st A, which is absent. Hind wing normal; Cu_1 and Cu_2 somewhat approximate. The European species (Sorhagenia Spuler) has apparently lost Cu₂ completely, and is a twig-miner and later a leaf-roller on Rhamnus.

1. W. amorphella Clemens. Head, thorax, and basal third of wing smoky blackish, somewhat mottled. Antennæ and palpi nearly concolorous. Fore wing with outer boundary of the dark area oblique outward, with a moderate blackish tuft in the cell and a very large one on the fold, just before it (unlike Laverna). Outer lart of wing paler, mottled elay-color and light brown, with some black scaling, tending to leave a somewhat paler, outwardly oblique median fascia, and a vague broken angulate subterminal one. Median fascia with a large transverse tuft across the cell, and a small one below the fold; a black dash in the fold just beyond them. A small tuft at end of cell. Terminal dots black. Many of the scales white-tipped, most strikingly at the margin. 15 mm. (miscecolorella Chambers.) May; July. The caterpillar forms a gall on Amorpha fruticosa, pupating in the

gall. (H. p. 430, f. 249.)

New Jersey to Minnesota and Missouri. New York: Ithaca.

4. STILBOSIS Clemens

No pecten, scape rather long and slender. Palpi long, smooth, and slender; third segment as long as second. Fore wing heavily tufted, in our species with strongly metallic vestiture, the tufts as in Walshia but larger, and tending to fuse in pairs. Fore wing narrow (fig. 188) four times as long as wide; costa slightly concave; \mathbf{M}_1 stalked with \mathbf{R}_2 nearly to apex; \mathbf{Cu}_2 obsolescent at base, but not approximate to Cu₁; 1st A lost. Hind wing a third as wide as fore wing, linear, with concave costa; **R** obsolescent, apparently long stalked with \mathbf{M}_1 ; the remaining veins separate; fringe five to six times as wide as membrane.

1. S. tesquella Clemens. Shining dark lead-color; antemedial tufts black; median and discal tufts in large straw-yellow patches, the latter connected with a strawyellow cesto-apical spot. 9 mm. (quinqueeristatella Chambers.) May to July; August. Caterpillar between leaflets of Amphicarpa monoica and

Lespedeza; light yellow, with concolorous head, black shields, tubercles and true legs. July.

New York and North Carolina to Minnesota. New York: Ithaca; Sea Cliff, Long Island.

5. LOPHOPTILUS Sircom

(Cyphophora Herrich-Schæffer; Leucophryne Chambers; Laverna, *Mompha*, in part)

Palpus nearly smooth, third segment three-fifths as long as second. Similar to Laverna. No pecten: 1st A of fore wing (fig. 183) stalked or free, well developed; hind wing three-fifths as wide as fore wing; R running to costa a short distance before apex; \mathbf{M}_1 and \mathbf{M}_2 stalked, widely separated from \mathbf{M}_3 ; upper and lower discocellular veins transverse. Tufts on fore wing normally heavy; fringe normally even.

L. eloisella and passerella are strongly aberrant in having minute raised tufts and a strongly caudate apex, and perhaps should be separated generically. The larvæ, so far as known, feed on Onagraceæ.

Key to the species

1. Ground white, apex caudate.
2. An antemedial faseia; smaller
2. Two antemedian black dots; larger2. eloisella.
]. Ground gray anex not candate] tricristatella

1. L. tricristatella Chambers. Dull brown, slightly shining, the scales of the paler portion pale-barred and tipped. Markings mostly indefinite. Inner margin slightly paler toward base, apex distinctly paler, tending to show a pale sub-terminal costal patch. The principal tufts are a median one below fold, and one at end of cell. 20 mm. (grandiscila Chambers, subiridescens Walsingham.)

May and June.

Ontario to District of Columbia and Colorado. New York: Uphill Brook (Mt. Marey).

2. L. eloisella Clemens. White. Palpus with two black bars. Fore wing with black dot at base, one on costa a fourth way out, and a small tuft on the fold opposite it. Outer half heavily streaked with yellow-brown; two broad streaks converging on apex from costa at three-fifths and four-fifths way to apex, each sometimes divided into two lines; two finer lines beyond middle of disc, meeting at the upper edge of a large brown, white, and gray tuft three-fourths way out. A smaller black spot near middle of inner margin, followed by a brown triangle just reaching this tuft; a powdery gray patch below apex. Fringe with a strong black peneil projecting from apex; with two triangular teeth on costal side, each edged with black. The yellow-brown may be partly replaced with black. 15 mm. (occasional dwarfs, 8 mm.)

Generally distributed; in June. Larva yellowish white, with red-brown head; in pith of Enothera stems, wintering in the dry stems.

3. L. passerella Busck. Venation as in Psacaphora; wing-form and pattern as in *L. cloisella*. Pecten present. White; palpus with a gray spot on second segment; fore wing with two black dots at base, a streak on costa a third way ont, a broad light brown median fascia, and a postmedian one separated by a narrow white area, both further out in the middle, irregular, and defined with black scales. Apex with mixed brown and black-and-white-barred scales, with a longitudinal black streak from the outermost dorsal tuft to the apex. A transverse black line across fringe at apex. Basal two-thirds of hind wing black, the rest fuscous.

East River, Connecticut. New York: New York City (Watson).

6. *PSACAPHORA* Herrich-Schæffer

(Lophoptilus, Laverna, Mompha, in part)

Peeten rudimentary. Palpi rather thick, but smooth, third segment much shorter than second; venation as in Lophoptilus, but with **R** of hind wing reaching costa only two-thirds way to apex, like \mathbf{R}_1 of Tinagma. 1st **A** stalked (fig. 187). Fore wing typically with slightly raised metallic markings.

Key to the species

1. P. (?) cephalanthiella Chambers. Powdery gray, obscurely mottled. Head eream-white, contrasting. Fore wing with markings all obscure, the distinctest being dark dashes in fold at base, and in middle of wing toward apex, of black scales and an antemedian silvery white spot in cell, one in fold before middle, and one at end of cell. Some of the dark scales also shining lead gray in some lights. Black dorsal tufts at two-fifths and three-fourths way to apex. 7 mm.

September and October. Caterpillar on button-bush in September and early October, in a blotch mine; pupating and emerging very soon. Larva with light.

dull-brown head, and body with crimson dorsal line and spots. Hibernation apparently in the imago.

This is probably a synonym of *luciferella* Clemens, described from New York and Pennsylvania, in June,

Southern Ohio.

2. P. argentimaculella Murtfeldt. Shining dark brown, with a good many of the scales toward apex finely white-barred; a broad lead gray antemedial fascia across middle of cell, with a large blackish tuft in fold beyond it; a strongly irregular postmedial fascia, sometimes Y-shaped, or with a separate costal spot before it, and sometimes broken in middle, with a large tuft beyond it on inner margin; with a small yellow area before it, or in the fork, if it is forked; a narrower strongly irregular subterminal band, produced out at middle. Fringe white-tipped. 7 mm.

End of August; December (forced?). Caterpillar a leaf-miner of Enothera, in August and October; mine a winding tract, often recrossing itself and sometimes becoming confluent into a blotch, with scattered frass; larva pale green with three indistinct pink dorsal stripes. Pupa in a dense white cocoon, usually outside the mine in a wrinkle of the leaf.

St. Louis, Missouri.

3. P. terminella Westwood, race engelella Busck. Deep bronzy brown; palpi pale powdery gray; autenne with a white band before apex. Fore wing bronzy on basal fourth, the rest bright yellow (metallic and changing from green-gold to copper), shading into dark brown at apex. An oblique silvery fascia at base, curving out to meet the antemedial fascia, one at a fourth way out below fold, and an equally long one at three-fourths way out; two large black tufts on dorsal half of wing, just beyond the two fasciæ, with a silver dash between them; a white costal dot before apex. Orbicular silver, slightly raised. Base of dorsal fringe lead gray. 8 mm,

May; June. Larva (Europe) whitish with yellow-brown head, in a greenish blotch mine on *Circua lutetiana*.

Connecticut to District of Columbia and western Pennsylvania. New York: Rock City (Cattaraugus County), Hemlock Lake, McLean.

7. PYRODERCE8 Zeller

(*Batrachedra*, in part)

Pecten strong, scape short: fore wing narrow-lanceolate and subfalcate: \mathbf{R}_i to \mathbf{M}_2 stalked. \mathbf{R}_s and \mathbf{M}_1 farthest: \mathbf{M}_s , \mathbf{Cu}_1 , and \mathbf{Cu}_2 parallel and equal; 1st A free, 2d A forked at base. Hind wing half as wide as fore wing, with concave costa and somewhat weakened veins; \mathbf{R} and \mathbf{M}_1 long-stalked; \mathbf{M}_2 somewhat approximate to them. CaterpiNars various. Palpi with second segment somewhat roughened, third a little longer.

1. P. rileyi Walsingham. Bright yellow-brown, palpus with third segment pale with three black rings. Fore wing with irregular and incomplete black bands at one-fourth and one-half way to apex, typically followed by luteous, and with a similar rudimentary mark on the costa before the first band, sometimes joined to it. Terminal region with two pale patches, each followed by a black one running out toward apex. A couple of black bars in fringe. Markings sometimes more obscure, the luteous being masked by a powdery extension of the black. Black powdering rarely gathering in a broken longitudinal streak.

Larva feeding in mummy fruits of loquat, rotten cotton bolls, stored corn, etc. . District of Columbia to Arkansas and south; common in the tropics.

WILLIAM T. M. FORBES

8. PERIMEDE Chambers

(*Mompha*, in part)

Similar to Laverna; scape rather long, without pecten; palpi smooth, with second segment slightly thickened, and third a little longer. Fore wing lanceolate (hardly four times as long as wide, fig. 182), normal, obscurely tufted, with 1st A lost; hind wing half as wide as fore wing, with R and M_1 and M_2 and M_3 connate.

Key to the species

1. Antenna white-tipped	
1. Antenna dark.	
2. Fringe wholly dark	
2. Fringe below apex white	

1. D menticements Provide Shining land groups immovalates terminal eight or

1. P. particornella Busck. Shining lead gray; immaculate; terminal eight or ten segments of antenna white. No other pale scaling. 12-15 mm.

End of May.

District of Columbia and vicinity; Texas.

2. **P. erransella** Chambers. Dark lead gray (with slight purple iridescence), the tufts very slightly defined with yellow or whitish scales; sometimes with a costal subterminal spot.

July and August.

New Hampshire to District of Columbia and Missouri. New York: Ithaca.

3. **P. falcata** Braun. Fore wing purplish fuscous, powdered, on a shining grayish white base. Similar to *P. erranscila* and *P. particornella*, but with a contrasting white patch in fringe below the apex, and with antennæ wholly dark. Hind wing mottled with white beneath. $11\frac{1}{2}$ to 14 mm.

June and July.

Cineinnati, Ohio; Pittsburgh, Pennsylvania; New York: Ithaca.

9. STAGMATOPHORA Herrich-Schæffer

Similar to Perimede; palpi more slender in our species, with second segment slender and as long as third. Fore wing with \mathbf{M}_{t} short-stalked (fig. 189); narrow-scaled and green-iridescent, with very obscure, practically obsolete tufts. For *S. sexnotella* see Laverna.

Key to the species

 \mathbf{M}_2 stalked (Cholotis).....2. ceanothiella. \mathbf{M}_2 free (Stagmatophora).....1. gleditschiæella.

1. S. gleditschiæella Chambers. Dark bronzy, with some greenish iridescence on thorax and fore wing; immaculate, the rudimentary tufts slightly duller. Anal tuft and hair on hind tibia orange in male. 12-15 mm.

May. Larva boring out thorns of Gleditschia.

District of Columbia; Ohio; Kentucky.

2. S. ceanothiella Cosens. Practically identical, except for the venation, with *S. gleditschiæella*. Basal joints of antennæ long and enlarged at tip (*Pyroderces*; *Cholotis* Meyrick).

End of May. Larva in a gall in a distorted terminal bud of Ceanothus (Ontario); also in a slight gall in the stem (Texas). In the former case wintering in the gall. Light yellow, with black head and two light brown triangles on neck.

Vieinity of Toronto, Ontario; Texas.

10. ITHOME Chambers

(Eriphia Chambers, in part; not Latreille, Meigen, or Felder)

Similar to Stagmatophora. Fore wing shorter, short-lanceolate pecten apparently present but fugitive. Venation apparently unstable in the number of medials preserved and in degree of stalking of M_1 . Cu_2 arising near angle of cell. Hind wing with R and M, long-stalked.

1. I. unimaculella Chambers. Dark smoky; a small subterminal pale spot on fore wing. Third segment of palpus with a series of pale dots on under side. 7 mm.

This species has been confused with I. (?) concolorella and even with the usually much larger Perimede erransella, but is distinguished by the palpi.

Southern States; northern distribution uncertain.

11. CHRYSOPELEIA Chambers

(*Æaa* Chambers)

Palpi rather long and slender, somewhat rough, third segment shorter than second. Wings with small turts; fore wing with \mathbf{M}_1 long-stalked with \mathbf{R}_6 ; \mathbf{M}_2 lost; Cu normal; cell central; hind wing linear-lanceolate; Sc obsolete; M_1 long-stalked with R, which runs to apex; the other veins present, apparently arising from Cu-stem.

The larva forms a large blotch mine between two veins, starting usually with a tract along the midrib. It forms a frass tube along the midrib, with two walls of frass extending from it to the margin of the leaf.

This genus is perhaps a reduced Gelechiid, and appears related to Helice.

1. C. ostryæella Chambers. Face, palpi, legs, and under side pale hoary gray; vertex, antennæ, thorax and fore wings dark powdery steel gray; base of inner margin paler. A paler fascia before the middle, farther out and wider toward inner margin, and a nearly parallel, doubly curved fascia beyond apical third; apex pale, with minute dark tufts in dorsal fringe. Four principal tufts on wing surface, arranged in a diamond, the two basal ones lying along the inner side of the first fascia, and the dorsal one farther out. 6 mm.

May to August; commoner later. Larva yellowish-white, with brown dorsal spots in front, and a spot on mouth parts; on iron wood (Ostrya) in September. Kentucky; southern Ohio.

2. C. purpuriella Chambers. Very close to C. ostryxella, the two basal tufts equidistant from base, the antemedial fascia weaker or absent, and the apical one more solidly pale. 6 mm.

June; end of July. Larva in July and late September; with habits like C. ostryæella, but the mine less regular in form; on red oak.

Kentucky.

12. BLASTODACNA Wocke

(Mompha, in part)

Similar to Laverna; pecten present, palpus with second segment a little roughened. Fore wing unlike all the preceding genera and like Batrachedra in having the cell squarely truncate; Cu_1 and Cu_2 connate from its lower angle. Two large dorsal tufts, 1st A stalked from 2d A. Hind wing with M_1 and M_2 stalked. The larvæ are bud-worms and fruit borers on Cratægus and related Rosaceæ,

and have dense secondary hair besides the usual setæ.

.1. B. curvilineella Chambers. Rather even, powdery, light gray, with two black tufts; sometimes with a dark dash beyond the first, but with no vellow shading. 8 mm.

Larva densely hairy, Platyptilia-like, in berries of Cratagus. The pupa hibernates and the moth emerges in the spring. Kentucky, New York: Ithaca (Wellhouse).

13. BATRACHEDRA Stainton.

Fore wing typically almost linear (fig. 186); but lanceolate in some of our species; palpi long, second and third segments equal; no pecten (except in some aberrant species); fore wing with cell oblique in wing, at its outer end separated from dorsal margin by only half its width, the end perpendicular; \mathbf{R}_{4} and \mathbf{R}_{5} completely fused, \mathbf{M}_1 typically stalked with them, free in \hat{B} , placendiella. One medial lost, \mathbf{Cu}_1 and \mathbf{Cu}_2 typically connate, and sharply divergent, \mathbf{Cu}_2 running directly across to inner margin; \mathbf{M}_3 closely approximate or connate, and much longer. (In B. trichella \mathbf{Cu}_2 is well back from the end of the cell, and \mathbf{M}_2 , \mathbf{M}_3 , and \mathbf{Cu}_1 are longitudinal, parallel, and more nearly equal than usual. \mathbf{M}_{t} is long-stalked.) Hind wing in B. placendiella as in Blastodacna; in the others linear, with reduced venation. Larvæ variable in habits, frequently scavengers. Epiphysis of fore tibia, in the typical group very small, at middle of tibia, and strongly prominent; more normal in B. placendiella.

The genus is heterogeneous and of doubtful relationship; it is sometimes associated with the Coleophoride. B. placendiella is clearly related to the Lavernidæ, but may not be very close to the other species.

I. Wings lanceolate; \mathbf{M}_1 free, \mathbf{M}_2 lost; epiphysis normal; pecten strong; antennor with raised whorls of scales; palpi not strongly divergent; oripositor exserted.

1. B. placendiella Busck. Palpi blackish fuscous, third segment with an ochroous annulation at base, and extreme tip ochroous. Antennæ dark fuseous with narrow black annulations. Thorax dark fuscous: head light and iridescent; fore wing blackish fuscous, nearly evenly powdered with pale; without a pale central stripe; small black tufts in fold at middle, on inner margin at three-fourths way to apex, and at end of cell; a black terminal line around apex, fringes light with a black line in them. Abdomen concolorous, with yellow tip. 11 mm.

June.

Western Pennsylvania,

11. For wing lanceolate (fig. 190) with \mathbf{M}_1 apparently stalked, \mathbf{M}_2 present, \mathbf{M}_3 and Cu, longitudinal; hind wing linear with erowded venation; maxillary palpi turned laterally; antenna smooth, scape short, no pecten.

2. B. trichella Busck. Palpi light ochreous with a small dark spot on side of second segment; antennæ rather darker; head and thorax light ochreous. Fore wing light ochreous at base, shading into purplish brown at tip, basal third of costal edge black; a short longitudinal black streak in middle of wing at threefourths way to apex, and an obscure oblique one at base of fringe; the wing sparsely dusted with black. Fringe, hind wing, and legs light ochroous; abdomen fuscous. 15 mm.

August.

Western Pennsylvania; southern Ohio.

111. Fore wing practically linear (fig. 186); Cu, very short and transverse: hind wing linear with renation almost obsolete and the scale tuft prominent. one-fifth way out from base; epiphysis small and prominent; palpi turned laterally; antenna smooth, without pecten; maxillary palpi porrect (Batrachedra).

3. B. salicipomonella Clemens. $R_{\rm y}$ and $M_{\rm t}$ separate; auteunae dark fuscous, with white annulations except at tip; palpi dark fuscous: second segment with a white ring at tip and sometimes at base; third with a more or less distinct central ring and whitish tip. Head fuscous, face white. Fore wing powdery fuscous, with a

paler, sometimes obscure stripe through the middle to outer margin and apex, with the blackish diseal dot on its upper edge, and a larger black spot on its lower edge at two-fifths way from base to apex. 10 mm.

Caterpillar inquiline in willow galls, especially the saw-fly gall on the leaves. White with a broad transverse black band on the front edge of each segment, the first band placed on the mesothorax, and interrupted in middle; eighth segment of abdomen with a band on posterior edge also. Head yellowish, venter pale; immaculate: legs normal. Hibernation as larva: image emerging in May. The larva apparently destroys the maker of the gall.

Generally distributed; a paler race (striolata Zeller) in Texas. New York: Larvæ at Ithaca.

4. **B.** præangusta Haworth. Luteons; head, body, palpi, and fore wings heavily dusted with blackish; not forming a distinct pattern, but distinctly darker on inner half of basal halt of fore wing. Typically with scattered spots of plain luteous, which are obsolete in our race.

June. Larva (Europe) white, with ocellate brown spots on sides, and a whitemarbled brown subdorsal band; dark brown head and black cervical shield; in the catkins and between leaves of poplar.

Western Pennsylvania.

IV. Fore wing very narrow clongate-lanceolate: \mathbf{M}_i free from \mathbf{R}_{5^*}

5. B. concors Meyrick. Head, with palpi, and abdomen whitish ochreous, antennæ and thorax somewhat darker; with tegulæ tinged with reddish. Fore wing pale ochreous, suffused and sprinkled with fuscous, the third fourth of the costa reddish brown, and apical part paler, more or less tinged with reddish; costal fringe with three dark basal dots, apical fringe with a dark basal line and a dark shade outwardly. Hind wing pale gray; fringe yellower. 16 mm. (Unknown to me.)

July.

Parry Sound, Ontario.

14. COSMOPTERYX Hübner

Antennae practically as long as fore wing, more slender than in the other species, and quite smoothly scaled toward base. Scape as long as entire width of head, and only a sixth as thick; without pecten; palpi as long as head and thorax, turned laterally, smooth, and very slender. Hind tibiæ with very little loose hair. Fore wing subfalcate (fig. 185) with apex of membrane produced, almost linear; with a silver streak in the apical fringe. M_1 long-stalked with R_{4+5} , and usually M_2 also, lower discocellular, and m-cu strongly oblique; Cu_2 long and longitudinal. Hind wing 'linear, with more or less rudimentary veins; R running to apex, long-stalked with M_1 ; the cell open below them; fringe five times as wide as membrane, or more.

Larva with strongly flattened head but otherwise fairly normal; forming a blotch mine with a hole in it through which the frass is ejected.

C. delicatella and fernaldella belong to the subgenus Lienigia Spuler, with eyes larger and rounded, front less prominent, scape and palpi rather shorter, spurs of hind tibia rather above than below its middle, and larva not always ejecting its excrement. The larva are to be sought in rush-like marsh plants.

Key to the species

- 1. Fore wing with fine longitudinal white lines toward base.
 - 2. Lines short, centering at one-fourth the length of the wing; ground dark brown.
 - 3. Inner margin very narrowly white toward base.

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4. Outer silver fascia of two obliquely placed vertical bars

4. gemmiferella.

3. Expanse usually under 8 mm.; apex of antenna white.....2. delicatella.

3. Expanse usually full 10 mm.; apex of antenna black.....1. fernaldella.

1. Fore wing with a double silver spot at one-fourth way out; but no lines. 7. clandestinella.

1. C. fernaldella Walsingham. Light brown. Head and thorax with three fine white lines; antennæ outwardly dark, with the fourth and tenth segments from apex white, and sometimes other isolated segments also. Fore wing with costal edge, at basal third, and dorsal edge, toward base, finely white; three white streaks on disc, the first running from costa, well out from base, and the lower reaching middle of wing. A yellow fascia covering third fifth of wing, bounded on each side with two silvery bars; the upper, inner one followed by a black spot; the outer two separated by a longitudinal yellow streak, which becomes white outwardly, is edged with dark brown, and runs through the fringe to the extreme apex. 8-10 mm. (foridanella Beutenmuller, nigripunctella Busck.)

Southern States; New Jersey; Ottawa, Ontario.

2. C. delicatella Walsingham. Similar. Antennæ with last four segments white, and normally also the sixth and tenth from apex. Fore wing with yellow fascia broader than in C. fernaldella; the upper anterior silver bar located farther toward the base; and the following black dot small, the yellow extending basally below the bar and touching the lower white streak. 8 mm.

Southern States, north to District of Columbia.

3. C. clemensella Stainton. Very similar to *C. gemmiferella*; ground of fore wing darker, orange fascia paler, less reddish, its bordering fascia pale golden rather than violet, the more apical one oblique and continuous. Antennæ with apex broadly white, and a broad white area about three-fourths way out, sometimes interrupted by a couple of black rings.

Larva on Ipomœa.

Massachusetts to Virginia and western Pennsylvania; "New York" (United States National Museum).

4. C. gemmiferella Clemens. Antennæ with apical three or four segments white, then four black ones, and one or two narrow white bands. Fore wing and thorax dark greenish brown; three longitudinal white lines on body; fore wing with three short white streaks at a fourth way out, the upper resting on the costa; and no white on inner margin; a broad orange fascia rather beyond the middle, with a large, oval, violet-silvery fascia before it, extending almost across the wing, and two separate spots beyond, the upper offset outward about its width and edged above with white; all the silver spots partly edged with black; a long silvery streak in apical fringe, starting from a silver-blue apical dot. 11 mm.

June and July. Larva a miner in Ipomea.

New York to Texas. New York: Sea Cliff, Long Island (Busck).

C. hermodora Meyrick (Exotic Microlepidoptera ii, 282) must be closely similar, but with one broad subapical band on the antennæ.

5. C. pulchrimella Chambers. Antennæ outwardly black, with fourth apical joint white, and a second group of two or three white rings at three-fourths its length. Fore wing much like C. gemmiferella; inner margin very narrowly white at base, yellow band duller and shading into brown toward base, rarely chocolate brown; silver fasciae bluish, the outer one continuous, oblique, starting from a white costal streak; apical white streak not reaching halfway in to the apical blue dot. 8 mm.

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June. Larva yellowish white with black head, and finely divided cervical shield. a miner on richweed (Pilea), making several successive mines and twisting and crumpling the leaves.

Minnesota to southern Ohio.

6. C. magophila Meyrick. Similar to C. pulchrimella; apex of antenna white, then five black segments, and two narrow white bands; dorsal edge rather strongly white; orange fascia rather pale, its silver outer boundary offset out on the costa. 7-8 mm.

August.

North Carolina; southern Ohio. New York: Ithaca.

7. C. clandestinella Busck. Antennæ blackish brown, last four joints white, then five black and two or three white joints. Fore wing blackish brown, with two or three short, longitudinally oval and usually confluent silver spots at basal fourth, orange area shortly and abruptly produced along costa. edged within by a narrow. complete, violet-silvery fascia, followed by a band of the ground color. Outer silvery fascia normal, its dorsal half stronger; streak in apical fringe small. 9 mm.

Late July. Larva a miner in grass (*Panicum clandestinum*), forming a clear, irregular blotch, and ejecting the frass through a hole at the end. Light green with yellow head and cervical shield; at maturity developing three brilliant wine red longitudinal stripes. Cccoon made of a bit of epidermis folded lengthwise.

District of Columbia, etc.

15. SYNALLAGMA Busek

Palpus with third segment about half as long as second and smoothly curved. Antenna with scape elongate. Fore wing lanceolate, with apex drawn out into a strongly sickle-shaped hook, and the outer margin below it nearly erect. Eleven veins; \mathbf{R}_1 lost, \mathbf{R}_4 and \mathbf{R}_5 stalked, running to costa, M₁ free, ending at the apex, 2d A strongly forked at base, 1st A doubtful. Hind wing half as wide as fore wing, lanceolate; fringe twice as wide as membrane, with all veins preserved; Sc and R approximate at base, \mathbf{R} and \mathbf{M}_1 connate, \mathbf{M}_3 and Cu, connate, M2 free, with the cell open below it, Cu₂ well separated. Fore wing with slight raised tufts. Maxillary palpi apparently absent. Hind tibiæ with short bristles above.

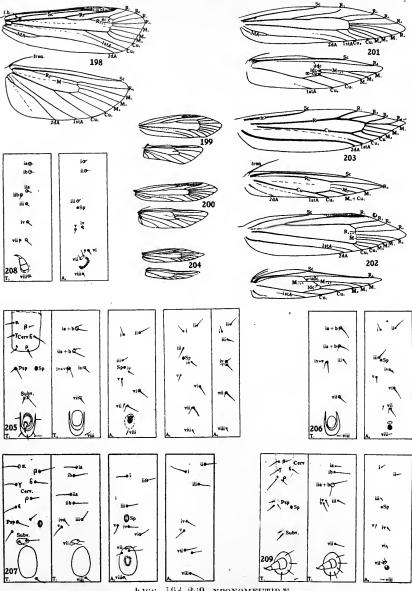
This genus is of very doubtful position; perhaps it is Heliodinid.

1. S. busckiella Engel. Dark brown. Antennæ annulate, dark and silvery gray; fore wing with coppery green iridescence; two dull white streaks in middle of wing, the upper one reaching the base; a double dark silver spot beyond a silver bar across base of hook, with two white spots in costal iringe near its end; apical fringe pale, fuscous-tipped. Hind wing dark. 9–10 mm.

. June to September. Western Pennsylvania.



195, Dysodia oculatana, venation; 196, Thyris maculata, venation of hind wing; 197, Thyris fenestrella, seta map of larva, ninth segment of abdomen (after Fracker)

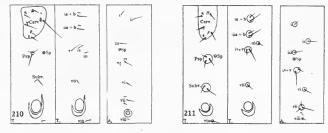


FIGS. 193–239. YPONOMEUTIDÆ 198, Attera aurea, venation; 199, Yponomenta padellus, venation (from a European specimen); 200, Plutella annulatella (Europe), venation; 201, Plutella maculipennis, venation; 202, Argyresthia oreasella, venation; 203, Scythris eboracensis, venation; 204, Epermenia illigerella (Europe), venation; 205, Yponomenta cagnagellus (Europe), seta map of larva; 206, Swammerdamia species (Europe), seta map; 207, Atteva aurea, seta may after Fracker); 208, Mieza species (Georgia), seta map; 209, Plutella porrectella, seta map (from a Enropean specimen).

SUPERFAMILY YPONOMEUTOIDEA

Head smooth, or nearly so, with a more or less distinct tuft on the vertex in Argyresthia, Zelleria, and Acrolepia, but with shorter vestiture than in any of the true Tineidæ. Eyes moderate or small; ocelli usually present and often conspicuous; antenna with pecten in some cases, without eye-cap, the scape of moderate size, as a rule; shaft with two rows of scales to each segment, the more basal always and the other sometimes interrupted ventrally, varying from the condition of the lower Tineids to that of the Macrolepidoptera. Palpi normally upturned (minute and drooping only in Heliodines) with third joint nearly as long as second, smooth sealed, long and conical or fusiform, never drooping; second joint smooth-scaled or with various ventral tuftings, but never with the roughly triangular vestiture usual in the Tortricidæ; without bristles. Maxillary palpi typically moderate and porrect, rather bristle-like, often minute, folded in Acrolepia. Tongue normally scaled, often strong, very rarely absent. Hind tibiæ various. No aculeæ or ovipositor. Fore wing with R, running to outer margin (except in the Heliodinidæ), but doubtful in some genera with reduced venation; \mathbf{R}_4 and \mathbf{R}_5 usually separate; 1st A free, usually distinct toward margin; 2d A usually forked at base; hind wing with Sc and R closely parallel at base, usually connected by \mathbf{R}_1 , but sometimes separate; never anastomosing. Female with two bristles in frenulum. Venation complete or nearly so.

Larvæ and pupæ various, though always showing the three prespiracular setæ in the larva. Cocoon very frequently lacelike of open meshes.



FIGS. 210-211. YPONOMEUTID.E

210, Argyresthia nitidella (Europe), seta map; **211,** Epermenia illigerella (Europe), seta map

This superfamily, as here delimited, is a rather heterogeneous series of forms which do not definitely belong to any of the other recognized groups; and the family separation here given is largely arbitrary

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(aside from the Ægeriidæ); but I believe it represents natural points of division. The whole series of isolated genera has been put here in the Yponomeutida; some of them are nearly Heliodinid while others would go as well in the neighborhood of the Tineidæ.

Key to genera of Yponomeutoidea (except Ægeriidæ)²⁴

- 1. Venation complete or nearly so.
- 2. Fore wing with five radials preserved, all running to costa (fig. 219). 2. Fore wing with only four radials running to costa. 3. Hind wing with \mathbf{M}_1 and \mathbf{M}_2 long-stalked (fig. 201). 4. Palpi with a triangular tuft on second joint; maxillary palpi porrect and 4. Palpi nearly smooth. 5. Head with a few erect hairs behind; maxillary palpi folded. Y 9. Acrolepia. 5. Head with a large tuft on vertex; maxillary palpi obsolete. Y 10. Argyresthia. 3. Hind wing with M, and M, separate; or a vein wholly lost. 4. \mathbf{R}_1 arising beyond middle of cell, which is lanceolate; without accessory 4. Cell ample, with large accessory cell; \mathbf{R}_1 almost always arising before middle of cell. 5. Palpi very short, drooping (fig. 222); hind wing lanceolate; M₃ long-5. Palpi upturned at least at middle of front; or wings ample, usually both. 6. Hind wing with complete venation (6 veins arising from cell). 7. With M₃ and Cu, separate or connate. 8. Vestiture hairy; palpi obsolete.........(Psychidæ-Solenobia). 8. Vestiture mostly scaly, palpi well developed. 9. Hind tibiæ bristled. 10. Hind tibiæ bristled both above and below; \mathbf{R} of hind wing ending at or above apex (fig. 204).....Y 8. Epermenia. 10. Ilind tibiæ bristled above only; R running to outer margin (fig. 220) 9. Hind tibia with long loose hair.
 - 10. Palpi upturned to vertex......(Xylorietidæ-Stenoma).
 - 9. Hind tibiæ smooth-sealed.

10. Palpi nearly smooth.

11. Hind wing with \mathbf{M}_{3} and \mathbf{Cu}_{1} separate (fig. 198); fringe less than half as wide as membrane; ocelli small.

Y 4. Atteva.

11. Hind wing with \mathbf{M}_3 and \mathbf{Cu}_1 approximate (fig. 214); ocelli very large.

12. Palpi with second and third joints equal (brown). G 4. Glyphipteryx.

²¹ In this key the letters prefixed to the genus numbers indicate the families: Y = = Y ponomeutidæ; G = G lyphipterygidæ; H = Heliodinidæ.

12. Palpi with third joint as long as first two together (steel blue).....G 1. Abrenthia.

10. Palpus with a triangular tuft; hind wing with \mathbf{R} and \mathbf{M}_1

7. M₃ and Cu₁ stalked; ocelli present.

8. **R** and **M**₁ parallel (figs. 214, 215).

9. Hind wing oblong, only about half as wide as fore wing.

G 4. Glyphipteryx. 9. Hind wing broader and rounded or triangular; nearly as wide as fore wing (fig. 213).

10. Second joint of palpus with long hair below (fig. 216).

G 3. Choreutis § Millieria.

10. Palpus smooth or somewhat rough.
11. R₄ and R₅ of fore wing stalked halfway to apex (fig. 242)......G 2. Simaëthis § Allononyma.
11. R₄ and R₅ separate.

12. Palpus with blunt truncate apex.

G 2. Simaëthis, typical.

12. Palpus with acute apex. G 2. Simaëthis § Brenthia. 8. R and M₁ of hind wing stalked...... (Xylorictidæ, Setiostoma). 6. Hind wing with a vein lost.

7. Hind wing ample, with broadly rounded apex.

8. No ocelli.

9. Palpi smooth-scaled; (ground white or gray).

10. Fore wing with all veins preserved (fig. 199).

11. A transparent spot at base of Cu of hind wing.

Y 2. Yponomeuta.

9. Palpi very rough; (two-thirds of fore wing black).

(Psychidæ, Kearfottia).

7. Hind wing lanceolate; ocelli present but not always large (fig. 215).

8. Apex rounded; palpi smooth...... G 4. Glyphipteryx.

8. Apex of membrane falcate; palpi rough, ending in a loose, trun-

Family 24. **YPONOMEUTIDÆ**

(With Plutellidæ, Argyresthiidæ, Acrolepiidæ, Scythrididæ, etc.)

Head smooth, except sometimes on vertex; oeelli small or absent; antennæ moderate, without eye-cap; scape moderate; each segment of shaft with two rows of scales, of which the outer is usually a complete whorl; the inner interrupted more or less broadly below, exposing an area of sensory setæ (in Argyresthia with both whorls complete as in some Tineidæ; in some specialized genera with both interrupted, and the sensory area continuous; as in Seythris etc.); but with the outer row always distinctly better developed than the inner, and the inner always present and well-exposed dorsally. Maxillary palpi various;

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labials moderate to long, upturned, without bristles, with the third joint long, and smoothly scaled; almost always straight and fusiform; tongue present, sometimes naked. Eyes good-sized. Thorax with scaly vestiture: legs various, typically smooth-scaled. Venation but little reduced; fore wing with \mathbf{R}_5 running to outer margin; usually with all veins separate, and usually with a stigma along costa from tip of **Sc** to \mathbf{R}_1 ; hind wing with \mathbf{R} and \mathbf{M}_1 well separated. except in Cerostoma; \mathbf{M}_1 and \mathbf{M}_2 often stalked; **1st A** distinct in both wings.

Larva quite variable (figs. 205-211) but always with seta beta on the cervical shield farther from the middle line than *alpha*, setæ iv and v widely separated on the abdomen, and the prespiracular wart bearing three setæ.

Egg, so far as known, of flat type and normal. Pupa obtect; with maxillary palpi distinctly preserved; with pilifers represented by distinct lobes only in the genus Atteva; front femora exposed except in Seythris. The pupa are less heterogeneous, apparently, than the larva, except for Seythris and Atteva.

The family as here limited is far from homogeneous: Yponomeuta. Prays, and Swammerdamia form the Yponomeutine, and are close relatives; Plutella, Cerostoma, and Trachoma form a second group: Plutelline; and Argyresthia and Zelleria, a third. The remaining genera are entirely isolated. Acrolepia partly fills the little gap between Argyresthia and the Tincide; Atteva is a fair Pyralid in the pupa, though nearly a normal Yponomeutid in the adult, and with several exotic genera may constitute the transition from the Tineids to the Pyraloids and Maeros. Seythris shows characters of the Gelechioidea, and may not belong here; Epermenia seems to belong in this region but is generically isolated.

Besides our miscellaneous assortment, several other types occur in Europe and in the tropics, some of the latter hardly to be distinguished from the Zygænidæ, and others connecting the Ægeriidæ and Heliodinidæ with this family (Tinægeriidæ). The Tinægeriidæ, as usually delimited, are mostly placed by Meyriek in the Heliodinidæ.

1. PRAYS Hübner

(*Yponomeuta*, in part)

Similar to Yponomeuta, but with fully scaled wings. \mathbf{R}_1 and \mathbf{R}_5 of the fore wing typically stalked, and basal fork of \mathbf{A} very large; our species with the venation of Yponomeuta, \mathbf{R}_4 and \mathbf{R}_5 being either stalked or closely approximated.

1. P. atomocella Dyar. Much like Y. multipunctella, black dots larger; abdomen, fringe, and hind wing dull salmon color, duller in male. 20 mm. (diaphorus Walsingham; Yponomeuta Walsingham and Dyar.)

April. Larva on Ptelea.

Southern Ohio; Illinois; Texas.

2. YPONOMEUTA Latreille

(Hyponomeuta)

Vestiture of head, body, palpi, and hind tibiæ entirely smooth, that on the head being of even, fine hair. No ocelli: male antennæ with the outer rows of scales raised, the sensory areas good-sized: scape large, cylindrical, with pecten; palpi reaching about to middle of front, slender, often drooping; maxillary palpi minute. Metathorax very large; scutum undivided. Fore wings (fig. 199) broadest four-fifths way out; with rounded, nearly erect outer margin, and well-marked anal angle at Cu_1 ; accessory cell large; M_a and Cu_1 arising rather close together, the other veins well spaced out. Hind wing about as broad as fore wing; oblong, with a transparent spot at base of $Cu; M_a$ lost; the other veins well separated.

Larvæ (fig. 205) social, in a loose web. With primary setæ only; posterior dorsal setæ on cervical shield farther apart than the anterior ones; ia and b obliquely placed on metathorax; iv and v widely separate on abdomen; prolegs with several complete circles of hooks. Pupa without fronto-clypeal suture; with prothorax much narrowed on middle line; antennæ shortish, and not meeting on middle line; maxillary palpi large, touching antennæ, middle legs, and tonguecase; abdominal spiracles produced and tubular. but not the thoracic ones; cremaster represented by four divergent setæ.

1. Y. multipunctella Clemens. White. A black spot on collar and a few on thorax; fore wing with rows of black spots, those on **A** and above fold quite regular. Male with hind wing gray except toward inner margin, base of costa. and under side; female with ground color wholly white; fringes all white; in aberration leucothorax Meyrick without dots on thorax. 20 mm. (orbimaculella Chambers, euonymella Chambers, ordinatellus Walker. δ scmialba Meyrick (H 48:44).

Larva social on Euonymus.

• Y. leucothorax Meyrick was described as a male with white hind wing.

General but local in distribution. New York (Edwards and Angus Collections). 2. Y. padella Linnæus, with fore wing lightly suffused with gray, or at least with a pale gray fringe, and wholly darker gray hind wing in both sexes, has been introduced at Rochester, Geneva, and Scholarie, New York, and is somewhat injurious to apple and other trees of the Rosaceæ.

3. SWAMMERDAMIA Hübner

Closely related to Yponomeuta; palpi short, projecting, rough, not unlike Argyresthia; no maxillary palpi; head slightly roughened, but less so than in Argyresthia. Wings narrower than Yponomeuta and more lanceolate, but with the same translucent patch. Larva (of European species, fig. 206) much like Yponomeuta, with seta ib behind ia on metathorax, and the anterior seta of the prespiracular wart lower (instead of higher) than the other two.

1. S. castaneæ Busck. Head and thorax cream-white, becoming gray behind; fore wing shining violet gray, heavily dusted on a luteous ground, leaving the costal fringe mostly pale, and pale striæ on the inner margin; wings becoming strongly coppery toward the apex. 12 mm. (pyrella Busck, not Villiers).

Larva solitary, in a loose open web on upper side of chestnut leaves. Green, shaded with darker; tubercles very dark in pale yellowish areas; i and ii in line, forming a broken subdorsal band. Head yellowish; cervical shield pale. Early July. Cocoon white, spindle-shaped, suspended in web. Moth emerging in August.

Dublin, New Hampshire; Connecticut; Pennsylvania.

There is a closely related, undescribed, oak-feeding species, with only a white bar in the fringe and the thorax wholly white.

4. ATTEVA Walker

(*Œta* Grote; *Paciloptera* Clemens)

Palpi closely upturned to vertex, and smooth-scaled; ocelli absent; tongue strong, naked; maxillary palpi obsolete; antennæ smoothly scaled, with short segments. Wing form much like that of Yponomeuta (fig. 198); the hind wing more or less translucent at the base, more ample than usual. All veins present and well separated. Hind tibiæ smooth in female, hairy in male.

Larvæ (fig. 207) social in a web on Simarubaceæ; much like those of Yponomeuta, but with two setæ instead of one on the leg-base of the mesothorax. Pupa aberrant, Pyraloid, with strong lobes representing pilifers, meeting in the middle line, so as to leave a suture running forward from the base of the tongue; labial and maxillary palpi almost completely covered; antennæ reaching tip of wing, separated by hind legs and tongue; frontoclypeal suture partly preserved, but epicranial suture absent; seventh segment of abdomen free in male, but no deep suture between ninth and tenth segments.

Atteva is a tropical genus of both hemispheres; our species was probably introduced from South America. It has been put in various groups, and even made the type of a family, Attevidæ. 1. A. punctella Cramer. Fore wing with alternate pale yellow and bright

1. A. punctella Cramer. Fore wing with alternate pale yellow and bright orange, rarely brown, bands; the yellow bands wider and eut into rounded spots by a fine black network. Hind wings translucent, smoky. 25-30 mm. (aurea Grote). (H 48:36.)

Larva social; in a web on Ailanthus.

New York to Illinois and south.

5. URODUS Herrich-Schæffer

(Trichostibas Zeller)

Antennæ without pecten, no ocellus; eyes very large; front strongly tapering below, the palpi hardly extending beyond it; tongue weak. Hind tibiæ with some loose hairs above, at base. Fore wing narrow, oblong; costa arched, apex bluntly rounded. All the veins preserved and free; \mathbf{R}_5 running to apex; accessory cell cutting off the upper angle of the discal cell obliquely; with \mathbf{R}_2 arising from beyond its middle, \mathbf{Cu}_1 and \mathbf{Cu}_2 somwhat approximate and strongly curved at base, starting off from the \mathbf{Cu} -stem at right angles; $2\mathbf{d}$ A forked. Hind wing ample, translucent; anal angle slightly lobed; all veins present and well separated; Sc and \mathbf{R} parallel, not closely approximated. \mathbf{Cu}_1 and \mathbf{Cu}_2 curved and widely separated from the rest. (H 29:66, as *Cydosia majuscula*).

The genus is a good sized one in South America, and looks more like a Pyromorphid than a Tineid.

1. U. parvula Henry Edwards. Blackish, immaculate. 20-25 mm. (calligera anet.)

Larva on Persea. With stiff bristles and yellow head. Cocoon oval, formed of regular, open trapezoidal meshes in oblique series; suspended by a thread which extends down along the side of the cocoon. The cast larval skin is ejected through a hole in the bottom; the pupa emerges through a similar hole in the top, before eclosion.

District of Columbia; Florida. The northern record is based on a single specimen which may have been a stray.

6. PLUTELLA Schranck

Head somewhat rough, but less so than in Argyresthia; ceelli present, small; maxillary palpi porrect, small in our species; labial palpi with a long triangular tuft on second segment; third smooth-scaled and sharply pointed, upturned; antennæ with pecten; more roughly scaled than in Yponomeuta; hind tibiæ smooth; wings broad-lanceolate, not distinctly falcate; \mathbf{M}_1 and \mathbf{M}_2 of hind wing approximate or fused on basal third (figs. 200, 201) (fused in our species), then separating; the other veins separate.

Larva (fig. 209) forming a slight web on Crucifera. Prolegs slender, with a single circle of hooks at the enlarged end; seta vii of metathorax single; abdomen . with seta iv and v remote. Pupa in a cocoon of regular, open meshes, which does not at all conceal it. Pupa similar to that of Yponomenta, but with the thoracic spiracle tubular, as well as the others, the cremastral setae hooked, and the tongue more than three-fourths the length of the wings.

1. P. maculipennis Curtis (The diamond-back moth.) Wings narrower and more lanceolate than *P. porrectella*; with \mathbf{M}_1 and \mathbf{M}_2 of hind wing fused two-thirds way to the margin. Fore wing of male with upper two-thirds mouse gray, lower third pale, the boundary usually marked by a darkening of the upper and a whitening of the lower ground color, with three sharply marked teeth. Female paler. without contrasts, but with the markings, so far as traceable, like those of the male. 13 mm. (cruciferarum Zeller; xylostella in part; male limbipennella, female mollipedella Clemens; female dubiosella Bentenmuller).

Caterpillar green with black head. On various Crucifera; often injurious to cabbage.

Generally distributed; probably introduced from Europe. New York: Ithaca, Albany, West Farms.

2. P. porrectella Linnæus. Wings broader; \mathbf{M}_1 and \mathbf{M}_2 fused only one-third way to margin; fore wing paler; luteous, with only traces of the diamond-back markings. Outer margin and fringe contrastingly blackish (vigilaciella Clemens). 14-15 mm.

Larva on the more slender Cruciferæ, such as Hesperis. Similar to P. maculipennis, but with brown head.

Europe; New Jersey; also on Pacific Coast. Probably introduced in the East.

7. CEROSTOMA Latreille

(With Trachoma, Harpipteryx, Periclymenobius, Credemna, etc.)

Similar to Plutella. Wings in our species falcate; R and M1 of hind wing, and sometimes \mathbf{R}_{i} and \mathbf{R}_{5} of fore wing, stalked; \mathbf{M}_{1} free and well separated from \mathbf{M}_{2} ; palpi with triangular tuft decidedly larger, and third joint proportionately quite small, but of the same character. Of the numerous subgenera which have been formed, our falciferella belongs to Trachoma, with R₄ and R₅ separated, and dentiferella and xylostella to Harpipteryx (Periclymenobius), with R, and R, stalked.

Larva similar to that of Plutella, with the hooks of the prolegs in a single row. 1. C. falciferella Walsingham. Wings very long and narrow, of almost even width, with falcate tip and nearly erect outer margin. Powdery grayish brown with two darker parallel oblique shades. (ordinalis Meyrick.)

Larva to be expected on Rosaceæ. Moth in August. Highlands of the Hudson, New Jersey; Ithaca, New York; Pacific States. 2. C. dentiferella Walsingham. Wings with extended falcate apex. Fore wing pinkish buff on costal two-thirds, pale yellow along inner margin; with a broken darker shade along the boundary, black in California specimens. Hind wing pale. (frustella Walsingham.)

Known from California; Douglas Lake, Michigan; and Ontario (where the pale form canariella has been taken in July).

3. C. xylostella Linnaus. Similar to C. dentiferella in structure. Fore wing very strongly falcate, dark brown, with inner margin contrastingly light yellow, its white edging sending a fine spur up to beyond end of cell. Hind wing dark brown. Head and top of thorax white.

June to July. Larva on Lonicera.

Hyde Park, Massachusetts (Haimbach). New York: Ithaca (W. T. M. F.).

8. EPERMENIA Hübner

(Chauliodus Treitschke)

Head smoothly scaled, rounded, without occlli; antennæ a little over half as long as fore wing, with pecten; palpi upturned to middle of front or beyond, with the second joint slightly rough-scaled; the third joint similar and rather shorter. broadly scaled to the apex; maxillary palpi very small. folded across the tongne: broadle; tarsi and maked. Hind tibias stiffly bristled above and below; spurs above middle; tarsi and mid-tibiae also bristled. Wings lanceolate (fig. 204), more or less falcate, almost as in Ccrostoma; fringe broad at anal angle and sometimes making the falcation much more marked. Fore wing with SC reaching to middle of costa; \mathbf{R}_i arising from before middle of cell, twice as long as \mathbf{R}_2 ; normal, with all veins present and well-marked accessory cell; \mathbf{R}_i and \mathbf{R}_3 sometimes stalked, but separate in *imperialella*, forking over apex; with small scale-tufts in dorsal fringe. Hind wing with SC reaching well toward apex, typically longer than \mathbf{A} of fore wing, \mathbf{R} and \mathbf{M}_i approximate at origin or stalked; the rest free: but the medials typically closely approximate; **m-cu** longish and parallel to outer margin; hind wing lanceolate, narrower than fore wing.

Egg rough, oval, horizontal. Larva (fig. 211) with setæ iv and v on a single large plate; prolegs with 16 hooks in a single circle; the anals with 12 hooks. Pupa perhaps the most primitive of the Obtectæ, with seventh segment of abdomen free in male, but third fixed; not progressing, with a large head and a narrow prothorax; a small maxillary palpus not reaching the tongue; and a specialized cremaster with about 10 hooked setæ; antennæ reaching to tip of wing but tongue falling short; two deep pits on sides of ninth segment of abdomen.

The genus is apparently of the same stock as the Yponomentidæ, but is quite isolated and doubtful in position. Superficially it may be mistaken for Acrocercops. but differs in the tufted wing and more extensively spined legs, as well as in the family characters.

Key to the species

- 1. Yellow, fringe falcate, more decidedly so than membrane.....6. imperialella. 1. Grayish, membrane lanceolate, fringe less falcate.
 - 2. Base of wing not noticeably paler.
 - A white discal dot, preceded by a heavy black bar....l. albapunctella.
 No white discal dot; at most, with some whitish scales outlining a black dot.
 - 4. Ash gray with yellowish streaks.....2. ramapoëlla.
 - 4. Reddish with sparse, irregular, white dusting only......3. pimpinella.
 - 2. Basal third somewhat paler gray.....4. cicutaëlla.
- I. Fore wing with a series of about six tufts on inner margin; the first one the largest, and the last ones in the fringe; \mathbf{R}_1 and \mathbf{R}_3 stalked; hind wing linear. more acute, a fourth as wide as its fringe, with \mathbf{M}_1 , \mathbf{M}_2 , and \mathbf{M}_3 approximate or stalked (Epermenia).

1. E. albapunctella Busck. Blackish gray, irregularly powdered with grayish white; two white dots on disc, the outer one contrasting; a thick black bar between them, which spreads out into a transverse bar at its anterior end. 12 mm.

Early spring.

Oak Station (Alleghany County), Pennsylvania.

2. E. ramapoëlla Kearfott. Palpi gray, dark-dusted on outer side, as usual; head paler. Fore wing somewhat streaked with luteous; black points at middle and at end of cell, more or less defined with pale yellow, and a slight dark oblique streak running up from inner margin to beyond the more basal black point. Wing outwardly blackish-dusted; the apical fringe quite dark gray. 13 mm.

Lighter than E. pimpinella and larger, shaded with luteous instead of the peculiar reddish gray of E. pimpinella.

Ramapo, New York (type).

3. E. pimpinella Murtfeldt. Powdery reddish gray; becoming almost chocolate brown on costal half; with a couple of unpowdered areas on the middle of the wing; sometimes with these areas extended, leaving the powdery ground only on the inner margin toward the base and an irregular, dominantly white, apical area and costal fringe. Three ocellate discal dots in a series. Five or six blackish tufts on the dorsal margin, regularly increasing in size toward the base. 9 mm.

Bred from Umbelliferæ.

Missouri.

4. E. cicutaëlla Kearfott. Ground luteous; basal third of fore wing largely so, but lightly strigose with black; outer two-thirds a mixture of brown, luteous, and black, the brown mostly in the middle of the wing. Dark specimens showing the brown more generally, especially toward base of costa. Base usually contrastingly pale to the naked eye, its outer boundary aligning with the largest dorsal tuit. Two longitudinal, barlike, ocellate discal dots.

• Larva in flowers and seeds of Cicuta (Umbelliferæ).

New Jersey.

5. E. canicinctella Clemens. Basal half of fore wing whitish; outer half fuscous; the boundary slightly oblique outwardly; a postmedial, slightly irregular, whitish band, better-defined on the outer side, followed by a raised black spot toward inner margin. Fringe broad without tufts in it.

I have seen only the unique type of this species; it probably does not belong here, but to the Lavernidæ (possibly to the genus Blastodacna).

II. Fore wing with only one really distinct scale tuft on inner margin; antennæ heavily pubescent, with reduced scaling; hind wing broad-lanceolate, with blunt apex; fringe only twice as wide as membrane, \mathbf{M}_1 , \mathbf{M}_2 , and \mathbf{M}_3 separate. R shortstalked. Fore wing with \mathbf{R}_4 and \mathbf{R}_5 separate.

6. E. imperialella Busck. Straw yellow. Thorax and head darker; palpi fuscous on outer side. Fore wing with a broad, oblique, ochreous shade extending from a third way out on inner margin to middle of costa; some less oblique streaks before apex and a more or less double blackish line in fringe beyond anal angle. Hind wing pale gray with yellower fringe. 15 mm.

June. Western Pennsylvania.

9. ACROLEPIA Curtis

Head moderately rough above; with ocelli; palpi upturned about to vertex, slightly tapering, the end joint long; maxillary palpi apparently rather longer than eye, slender, folded. Hind tibiæ smooth-scaled. Venation as in Argyresthia; accessory cell strongly marked; our species with fore wing a little falcate; Sc long; hind wing rather trapezoidal, nearly as wide as fore wing; M_1 and M_2 , M_3 and Cu_1 stalked. Early stages not studied structurally.

The genus is sometimes put in the family Tineidæ or Argyresthiidæ, sometimes in the Yponomeutidæ or Plutellidæ, and sometimes given family rank. It seems

a little lower than Argyresthia but closely related to it. The wing-form makes it look like a minute Cerostoma.

1. A. incertella Clemens. Fore wing gray-brown, somewhat iridescent with red bronze; with vague darker streaks and patches, especially one at middle of costa; an oblique white streak extending up from inner margin near base, to about middle of wing; hind wing pale. 12 mm. (Adela flamensella Dietz, not flammeusella Chambers).

April; July and August; October. Larva tying and skeletonizing leaves of smilax, or boring in bulbs of Lilium. Pupa in a lace cocoon, like those of the rest of this series.

New Hampshire to Kentucky; west to California.

10. ARGYRESTHIA Hübner

(With *Blastotere* Ratzeburg)

Vertex rough-hairy, though less so than that of Tinea; front smooth-scaled; no ocelli; antennæ normal; the shaft with two nearly complete circles of scales to a segment; peeten strong, sometimes almost an eye-cap. Labial palpi moderate and porrect, or upturned nearly to vertex, smooth-scaled or a little roughened; maxillary palpi obsolete; hind tibiæ with spurs near middle, usually rather sharp edged above, with some rough, bristly hair, which may also extend on the metatarsus, but otherwise smooth-scaled. Fore wing lanceolate (fig. 202), with Sc short, \mathbf{R}_1 arising from middle of cell or before; \mathbf{R}_4 and \mathbf{R}_5 typically free (stalked in group Blastotere); M_2 and M_3 free, stalked or rarely united (annettella, rileiella); accessory cell and usually base of M distinct; 1st Å distinct; 2d A shortly forked at base, long; a stigma on costa before \mathbf{R}_{i} , as in many Yponomeutidæ; hind wing with costal cell abruptly narrowing before the middle; Sc ending at middle of wing or before; M_1 and M_2 stalked, widely separate from R; M_3 and Cu_1 free, stalked, or, in one western species, united; hind wing lanceolate, narrower than fore wing. The imago differs from the typical Tineidæ mainly in the lack of palpal bristles, and in the fact that \mathbf{R}_{s} runs to outer margin, in both of which points it agrees with the Acrolophinæ.

The larvæ (fig. 210) bore in twigs, buds, and fruits, or occasionally are leafminers; the cocoon is of white silk, often spun in the tunnel. The pupa is obtect, with maxillary palpus reaching from antenna to the maxilla, which is extended laterally to meet it; middle leg not reaching maxillary palpus; tongue short; antennæ reaching to tip of wings; eremaster unspecialized, with eight hooks and setæ.

The moths of this genus rest with the head appressed to the substratum, and the tail held up at a sharp angle.

Key to the species

1. Fore wing silvery and golden or bronze brown.

2. With definite markings.

- 3. Ground wholly golden, with brown markings; head white.. 10. alternatella. 3. Part of ground, at least, white.
 - 4. Disc of thorax golden; head more or less yellowish; fore wing silver with golden fasciæ.....l. gædartella.
 - 4. Head and thorax white. 5. Markings outlined with dark brown; median fascia broader on inner
 - 5. Markings black-brown on silver.....4. media.

 5. Markings golden, not outlined. 6. Apical marks of three or four confused striæ; or suffused with golden
golden. 7. With a black apical spot; sometimes a single scalc. 8. The golden area dominant; markings slightly diffuse.
6. freyella. 8. White ground color predominant; markings clean-cut.
9. A forked median fascia
7. No black apical spot. 8. Dorsal margin white.
9. Inner margin narrowly white, with dark spots. 13. belangerella.
9. Inner half of wing white.
10. No dorsal dark spot9. subreticulata.
10. A dorsal spot or dot near middle of inner margin. 11. castaneella.
S. Dorsal half barred with silver, white, and golden.
5. pygmæella. 6. Apical marks simple, enclosing only three silver spots. 2. calliphanes.
2. Immaculate pale golden
2. Expanse 12 mm; white dorsal area narrow, not extending up to fold toward base.
 Spot on middle of inner margin rounded, separate13. belangerella. Spot on middle of inner margin, broadly connected to dark costal region. 14. conjugella.
 Expanse 9 mm.; white dorsal area often covering half of wing. Ground color of fore wing light brown except basal half of inner margin. <i>15. rileiella.</i>
 Ground color white, more or less marked with black-brown. A complete oblique transverse fascia.
 5. A large, dark streak preceding the fascia
I. Fore wing with metallic markings.
* \mathbf{R}_{*} and \mathbf{R}_{5} free; apical markings coarse (Argyresthia).

1. A. gædartella Linnæus. Thorax bright golden; fore wing white and coppery golden; first band running from costa at base to a quarter way out on inner margin; median band a widely forking upright Y; outer band similar, but inverted and sometimes joining the dark terminal patch. 11-13 mm.

April; June to August. Larva green or reddish, with dark brown head and cervical shield marked with black; in catkins and under bark of twigs of birch and alder.

Europe; reported from various places in United States, in part, at least, in error for A. calliphanes.

2. A. calliphanes Meyrick. Head white; thorax white, slightly gray-stained; wing markings exactly like those of A. gadartella, but on the average slightly more extensive.

Larva on alder.

1.

Toronto, Ontario, and Maine, to British Columbia and California. New York: Albany.

3. A. oreasella Clemens. Head and thorax white; fore wing silvery; marks golden, edged with brown, except the yellow shade at base of costa. Median band a half-crescent, somewhat irregular and rarely reaching costa; sometimes with two small dots opposite it on costa; outer third with an irregularly forked golden figure, enclosing four white spots, of which the one two-thirds of the way out on the costa is the largest; or with the three outer spots fused into one. 10-13 mm. (andereggiella of American authors).

July. Larva possibly on oak.

Distribution general. New York: Wells Rock City (Cattaraugus County), Ithaca, Poughkeepsie, West Farms.

4. A. media Braun. Silver; palpi, face, and front of tuft pale golden; antennæ annulate golden and pale brown, rather less contrastingly than in A. oreasella; tegulæ golden. Fore wing with costal edge suffused with pale golden; markings dark bronzy brown; a rather broad fascia from middle of dorsum almost to costa, widening upward, with a spur running obliquely outward to join the eleventh or twelfth of a series of dark costal striæ, apical third dark bronzy, extended inward acutely toward middle of wing and enclosing two white costal spots and three smaller dorsal ones. 9 mm.

This species is transitional to the following group.

Late May.

Cincinnati, Ohio.

5. A. pygmæella Hübner. Silver white, slightly yellowish; antennæ annulate; tegulæ golden; fore wing with golden fasciæ fading out above, the first one meeting a dash in the base of the fold, connected by suffusion to the base of the costa; the second fascia outwardly oblique and fading out at costa; the third confused and running out into the mottled apex. Costal region lightly strigose with golden. 12-14 mm.

July.

Ottawa, Ontario, to British Columbia; Europe.

If the American form is distinct, its name will be chalcochrysa Meyrick.

** \mathbf{R}_4 and \mathbf{R}_5 stalked in fore wing; with fine apical stria.

6. A. freyella Walsingham. Golden vellow and silvery white. Base of the fore wing streaked, the rest marbled, tending to form wavy anastomosing transverse bands; tegulæ golden; antennæ annulate. 8 mm. (abdominalis Zeller, in part, thuiella auct., not Packard.)

June. Larva on red cedar and arbor vitæ.

Ottawa, Ontario, to New Jersey and Texas. New York: Ithaca (United States National Museum).

7. A. annettella Busck. Tegulæ white; fore wing silvery white; a golden antemedial band; costa golden from it to base; a median band, wider on costa and enclosing a small white spot; the outer third with about three broken golden striæ. 9 mm.

June and July. The larva mines about four leaves at the tip of a juniper twig, passing through the stem from leaf to leaf, completely emptying the leaves, and scattering the frass. It hibernates in the mine; it pupates in May, in a cocoon of open meshes, formed outside the mine.

Connecticut to southern Ohio.

8. A. apicimaculella Chambers. Silvery white; fore wing with a more or less triangular dark brown apical spot, and with indistinct brownish streaks across the apex before it; a bright ochreous streak below base of costa, or suffusion on costa. 9 mm. July. Larva possibly on oak.

New Jersey to western Pennsylvania and Kentucky.

9. A. subreticulata Walsingham. Costal third light bronzy brown, outwardly reticulate on a white base; the rest of the wing pure white. 9 mm.

June. Larva on red maple.

New Jersey to Pennsylvania.

10. A. alternatella Kearfott. Head white; palpi golden; antennæ golden fuscous; scape paler; thorax white; posterior part and tegulæ golden. Fore wing golden ochreous, with oblique and anastomosing brown fasciæ, terminating in five evenly spaced costal and three dorsal patches. Fringe vellowish toward apex; fuscous at anal angle. 10-12 mm.

May; July. Bred from juniper berries.

Essex County, New Jersey. New York: Ithaca. 11. A. castaneella Busck. Near A. subreticulata; antennæ annulate, golden and brown; base of tegulæ pale golden; fore wing with costal edge golden, gradually widening beyond middle into the golden brown apical region, which is slightly reticulate with white. A golden brown spot at end of cell and one on middle of inner margin.

May and June. Bred from bark of chestnut infested with Sesia.

New Hampshire; Virginia.

12. A. laricella Kearfott. Head and palpi whitish ochreous; second segment of palpi somewhat fuscous toward tip; face, antennæ, and thorax white, the latter yellowish. Fore wing very pale shining ochreous, the basal half of the costal edge and fold narrowly darker.

Larva in terminal twigs of Larix americana, forming a burrow sometimes 15 cm. long; in injurious numbers at the Mer Bleue, near Ottawa, Ontario. Moth in July.

II. Without metallic lustre.

$*\mathbf{R}_{4}$ and \mathbf{R}_{5} free.

13. A. belangerella Chambers. Head and thorax white, with brown tegulæ; costal half of fore wing brown, the costal edge with darker dots, alternating with whitish ones. Dorsal third white, interrupted by a couple of dark spots. 13 mm.

The type shows considerable gloss and might possibly be put in the first group of the genus.

June.

Canada (Montreal and Ottawa).

14. A. conjugella Zeller. Closely similar to A. belangerella, but with the blackish spot near the middle of the inner margin squarish and broadly connected with the brown general surface of the wing. 12-13 mm.

July. Larva on fruit of apple and mountain ash.

New York to British Columbia; Europe. New York: Mt. Marcy (3000 feet), Trenton Falls.

A. belangerella may be only a variety of this species.

15. A. rileiella Busck. Light brown, somewhat shining; head and thorax, except tegulæ, white; dorsal margin on basal half white, at middle cut by a heavy brown bar, and outwardly more or less striated; apex striated as in freyella and its relatives, but less strikingly. No black apical dot. 9 mm. (mendica Walsingham, not Haworth).

This species may possibly be a dwarf form of A. conjugella.

District of Columbia.

** \mathbf{R}_{4} and \mathbf{R}_{5} stalked (Blastotere).

16. A. undulatella Chambers. White; antennæ annulate with black and white. Fore wing dusted with dull brown except on inner margin; base white except at costal edge, forming a white triangular area. An oblique, slightly irregular, dark median fascia, weaker at margins. and an apical patch formed of three partly fused fasciæ. 8 mm.

Larva a bast miner on trunks and larger branches of elm; maturing early in the spring. Cocoon on the bark. Moth in June; sometimes abundant.

Massachusetts to Kentucky and Missouri.

The stalking of \mathbf{R}_4 and \mathbf{R}_5 in this species is inconstant, in fact the Cornell University collection has a specimen in which \mathbf{R}_4 and \mathbf{R}_5 are stalked on one side only. Meyrick has based the name mesocausta on specimens in which R_1 and R_2 are separate. This appears to be the more usual condition.

17. A. austerella Zeller. White, with four, parallel, oblique, slightly irregular, dark fasciæ; the first usually not reaching the inner margin, the second complete, the third and fourth near the apex, obscurely forked at costa and lying in a fuscous-dusted area. Some brown also on base of costa. Fringe dirty white. 9 mm.

June.

Maryland to North Carolina, and Kausas to Texas. New York: Otto, Rock City (Cattaraugus County).

18. A. thuiella Packard. Similar to A. undulatella; the median band not so black, and broken into costal and dorsal spots, not oblique; an erect postmedial band. Oblique apical striation grayer, best-marked on the costa. 8 mm.

June. Sometimes injurious to arborvitæ, eating out the tips of the shoots.

Canada and Maine to Pennsylvania. New York: Ithaca, Hicksville, Westerly, Long Island.

11. ZELLERIA Stainton

Much like Argyresthia. Ocelli present; third joint of palpi broadly scaled. somewhat blade-like in Z. celastrusella; vestiture spreading in Z. retiniclla and haimbachi, the second joint also quite rough scaled; venation as in Argyesthia; Sc of hind wing short; M_3 wanting, M_2 free.

In Z. celastrusella the palpi are very rough and the wing has rough scales and tufts as in Xyrosaris Meyrick, but in the latter the antenna is as long as the fore wing, in our species much shorter. Neither of our species is typical of the genus.

1. Z. celastrusella Kearfott. Grayish, powdered with brown on a pale gray base; the wings loosely scaled and with scattered black dots formed of a single long scale each; head paler; palpi pale in front. Fore wing with a dark fascia at a fourth way from the base; white patches well toward the apex, and opposite each other on costa and on dorsal margin. Terminal line black, fringe dark. A long pencil of dark hair arising from base and extending along the under side of costa. 13-16 mm. (Xyrosaris Meyrick.)

End of May to June. Larva vivid green, cervical shield concolorous, shining; head olive; legs yellower. Webbing leaves together and feeding in terminal twigs of *Celastrus scandens*, in early spring. Moth resting with the head appressed to the substratum and tail raised, as in Argyresthia.

New Jersey; New York (Kearfott); Texas.

2. Z. haimbachi Busck. M_2 of hind wing distinctly nearer Cu_1 than M_1 . Wings smooth. Palpi ending in a large loose tuft of spatulate scales. Yellow ochre; head white; a broad, somewhat diffuse, white longitudinal band through fore wing. Hind wing pearl gray. 10 mm. July. Larva on short-needle pine, in June.

Wenonah, New Jersey.

3. Z. retiniella Kearfott. Structure much as in Z. haimbachi. Antennæ much shorter than fore wing. Fore wing bright golden, with confused white striæ on the disc, the strize formed of scales some of which have yellow-brown and light gray-brown spots and bars. Fringe white, gray-brown at apex. Hind wing white, gray-brown at apex, with white fringe. Head and thorax white, tegulæ golden. 15 mm.

July 4.

Lakehurst, New Jersey.

12. SCYTHRIS Hübner

(Butalis Treitschke; Arotrura Walsingham)

Head smoothly scaled; eyes small, ocelli and pecten variable, the latter typically present, but absent in our species (group Apostibes Walsingham). Palpi smoothly scaled, upturned about to vertex, or somewhat shorter, maxillary palpi extremely small and folded over base of tongue, as in the Gelechioidea. Shaft of antenna much as in Yponomeuta: scaled dorsally only; with strong bristles on sensory area; hind tibia hairy. Body stout. Fore wing lanceolate (fig. 203); \mathbf{R}_1 arising from well beyond middle of cell and short; cell narrow; without accessory cell; \mathbf{R}_{4} and \mathbf{R}_{5} more or less stalked, forking over apex; \mathbf{R}_{5} running to outer margin; M_s fused with Cu_i , and 1st A completely free, 2d A long, with the lower leg of its basal fork strong. Hind wing nearly as wide; costa not sinuate; Sc long; R and M, nearly parallel; anal region small but fully veined, as usual; M_2 and M_3

Connate or stalked in the typical group, but separate in our species. Larva with tufted hair from small warts; with from four to eight sets on the ventral leg plates; prolegs not especially long, with a complete circle of bi- or trito the seventh segment of the abdomen; antennæ meeting in the middle line, not reaching the end of the wings, which also meet; epicranial and frontoclypeal sutures complete. Prothorax Gelechiid, rather wide; maxillary palpi minute; babiele surgest of for former concerted abdoment of the seventh and the seventh and the seventh of the seventh and the seventh of the labials exposed; fore femora concealed. Abdominal spiracles tubular; setæ mostly hooked, including the cremastral ones.

This genus represents a small group with a curious mixture of Gelechiid and Yponomeutid characters in pupa and adult, which is sometimes placed in the Gelechiidæ, or made a separate family. There are but few genera, though Scythris has a large number of species in Europe. Most of our species will probably be removed on venational characters, but for the present the series had better be treated as a single genus. Colinita is very close, but has \mathbf{R} , in its normal position and M₃ preserved.

Key to the species

1. With erect postmedial and oblique antemedial pale bars.....l. impositella.

1. No pale bars.

2. Apex with golden scales; typically, with a golden streak in base of fold also.

5. basilaris.

1. S. impositella Zeller. Head, thorax, and forewings blackish, with some purple iridescence; tegulæ pale; two greenish white, somewhat diffuse fasciæ, the more basal one oblique and curved, with the lower part longitudinal, the outer erect and oval. A pale dot at base of wing. 12 mm. (matutella Clemens). General. New York: Rock City, West Farms. S. trivinctella Zeller, a related species with more white at the base and with a vigrage nestmedial fascia is to be expected in the wast of our territory.

zigzag postmedial fascia, is to be expected in the west of our territory. 2. S. graminivorella Braun. Dark brown, faintly brassy; palpi slightly paler inwardly and at bases of segments. Fore wing streaked with paler scales that gather to form distinct dots, the first in the fold at one-third, the second an oblique bar at two-thirds, and a third at the apex. Antemedial spot defined with a vague darker area, lacking the pale scaling, and a similar area between the outer bar and the apex. Hind wings darker, purplish. Abdomen pale below. 11 mm.

Larva a leaf-miner in grasses (Hystrix, and more rarely, Poa), making several

mines, whose entrance is guarded with a broad tube of silk. Larva in May; moth in early June. Cincinnati, Ohio.

3. S. fuscicomella Clemens. Black-brown, slightly iridescent, with a slight yellowish iridescence except on fringe.

This is probably the same species as the next. It was taken in June. Larva with S. eboracensis.

Pennsylvania.

4. S. eboracensis Zeller. Wholly blackish. Base of palpi whitish on outer side. 9-12 mm.

June and July; end of August and September. Larvæ in webs in tops of thistle. Maine to Texas. New York: Rock City (Cattaraugus County), Portage, Crugers, Sea Cliff, Long Island.

5. S. basilaris Zeller. R, and R, wholly united, running to apex. Purplish fuscous. Face ochreous at the sides in variety flavifrontella Clemens. Fore wing with a few golden bronze scales, forming a spot at the apex of the membrane, and often a streak in the base of the fold. 12 mm.

In variety flavifrontella the apical spot is also enlarged.

Distribution general in June and July. New York: Keene Valley; Cold Spring Harbor, Long Island.

6. A. pilosella Zeller. Brown. Palpi yellow-brown: inner third of fore wing with seattered, slender, yellow hairs. No iridescence. 10 mm.

This species is unknown to me.

Massachusetts.

7. S. aterrimella Walker, a plain blackish species from west of Hudson Bay, is unknown to me; it is possibly the same as S. eboracensis, but the male genitalia would have to be studied to make sure.

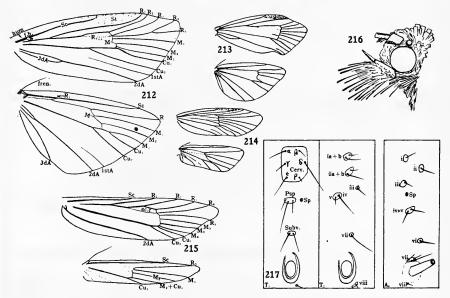
S. charon Meyrick, described from North Carolina, has a pale streak in the fold.

Family 25. GLYPHIPTERYGIDÆ

(Choreutida; Hemerophilida; Yponomeutida, in part)

Head smoothly scaled; ocelli in our species large and conspicuous; tongue scaled at base; maxillary palpi minute; labial palpi upturned to middle of front or beyond, sometimes beyond vertex; normally smooth scaled, but often with long loose hair on second joint or rough hair on second and third joints. Antennæ without pecten; with the outer row of scales of each segment usually complete, but narrow, and the inner row broadly interrupted below and replaced by conspicuous sense-hairs (minute in the Florida genus Tortyra). Middle and hind tibiæ with more or less hair above, which tends to be stiff and to gather opposite the spurs; or else smooth scaled. Wings exceptionally broad; macrolike in shape; sometimes with the fringes not wider than in some macros, and but little lengthened at the anal angle; only in Glyphipteryx, lanceolate with broad fringes, and even in this genus with rounded apex. Fore wing with a large accessory cell, and often with base of **M** more or less preserved; cell normally almost squarely cut off at end; \mathbf{R}_{\star} and \mathbf{R}_5 stalked only in Allononyma, \mathbf{R}_5 running to apex, or, usually, to outer margin; Cu, arising close to angle of cell; 1st A rather weak, free; 2d A with a large basal fork. Hind wing usually somewhat

smaller, ample except in the more reduced Glyphipteryxes; Sc connected to **B** by a more or less distinct crossvein; **R** and \mathbf{M}_1 separate, parallel or divergent; \mathbf{M}_3 and \mathbf{Cu}_1 approximate, stalked, or united. \mathbf{Cu}_2 farther from angle of cell than in fore wing, and without a fringe of



FIGS. 212-217. GLYPHIPTERYGIDÆ

212, Simaëthis diana, venation; 213, Choreutis pretiosana (Europe), venation; 214, Glyphipteryx loricatella (Europe), venation; 215, Glyphipteryx impigritella, venation; 216, Choreutis species, side view of head; 217, Simaëthis fabriciana (Europe), seta map of larva

hair on base of **Cu**; 1st **A** and 3d **A** sometimes weak; 2d **A** strongly forked at base, more so than in any other frenate known to me.

Egg of upright type. Larva (fig. 217) with front acute, extending well toward, but not reaching, vertex; ocelli normal; upper anterior seta of cervical shield farther from middle line than upper posterior seta; abdomen with seta i nearer middle line than ii, even on the eighth segment; iv and v close together; prolegs slender, with a complete, but often weak, circle of uniordinal hooks. Pupa (described by Miss Mosher as Heliodinidæ) incomplete, with some motion even between the second and third abdominal segments; with some cremastral setæ, but no true cremaster. Abdominal segments with anterior rows of fine spines only; wings reaching the fourth segment of abdomen; spiracles not distinctly tubular; maxillary palpi, labial palpi, and front

femora all exposed; antennæ not meeting in the middle line and not reaching the end of the wings; tongue moderate or reaching end of wings; prothorax and dorsal head piece both rather narrow, distinct, and sub-equal; mesothorax extending back as a long lobe in mid-dorsal line, nearly cutting the metathorax in two.

The Glyphipterygidæ are a small and homogeneous family, with a distinct relationship to the Tortricidæ with which they have sometimes been united. They are very close to the Yponomeutidæ in adult characters, but well separated from them in the early stages, especially by the normal incomplete pupa. There are about 550 species, largely Oriental. Setiostoma, formerly placed here, belongs near Stenoma.

Key to the genera

- 1. Third joint of palpus twice as long as second, smooth.....l. Abrenthia. 1. Third joint of palpus but little if at all longer than second.
 - 2. Second segment of palpus with long hair below, about as long as the segment
 - 2. Palpus not reaching vertex; with rough-scaled second joint..2. Simaëthis.
 - 2. Palpus smooth, upturned beyond vertex.....4. Glyphipteryx.

1. ABRENTHIA Busck

Wings ample; venation as in Glyphipteryx; fore wing with cell two-thirds as long as wing; hind wing widening outward, as in Glyphipteryx, with cell half as long as wing. Palpus with third segment longer than first and second together; smooth.

1. A. cuprea Busck. Deep purple; head and tegulæ golden bronze, thorax darker. 11 mm.

Late June and July.

Quebec; Pennsylvania; Virginia.

2. SIMAETHIS Leach

(With Brenthia Clemens, Orchemia Guenée, Hemerophila Hübner (Tentamen), Allononyma Busck)

Fore wing (fig. 212) triangular with moderately short outer margin and marked anal angle; hind wing rounded triangular, with anal region broad, and all veins developed; 3d A long. Cells of moderate length. Wings marked with metallic scales. Palpi with second and third joints slightly thickened with rough scales.

Our four species represent three different groups. The first three occur also in Europe, most of the relatives of the fourth are tropical.

I. Fore wing more perfectly triangular, with marked apex; outer margin concave above and below middle, the fringe marked there with white. Palpus with third joint cylindrical; thick in side view. Fore wing with \mathbf{R}_4 and \mathbf{R}_5 normally separate (Simaëthis).

1. S. fabriciana Linnæus. Mottled dull brown, with obscure markings suggesting a small noctuid; the two white streaks in fringe covering two-thirds of the width of the fringe, contrasting. 12 mm. June. Larva on nettle, etc. Dull yellowish; head black; clypeus and a couple

of spots on cheeks pale; cervical shield, anal plate, and true legs black; the shield

divided. In a web at tip of leaf in April, July, and August. Cocoon dense, white. Ottawa, Ontario; Montreal, Quebee; Europe. A paler form occurs in Colorado.2. S. pariana Clerck. Similar to S. fabriciana, but with the markings even more

2. S. pariana Clerck. Similar to S. *fabriciana*, but with the markings even more obscure, and the white in the fringe confined to the tips of the outermost scales. 10 mm.

August. Larva on apple: a leaf-roller.

Tarrytown, New York, introduced from Europe.

II. Fore wing triangular with marked apex, but even and slightly convex outer margin; the fringe wholly and evenly fuscous. Palpi as before. \mathbf{R}_4 and \mathbf{R}_5 stalked halfway to apex (Allononyma).

3. S. diana Hübner. Gray, powdery, and mottled with inscous; with three obscure, broken, and irregular transverse bands, partly defined with white. Sometimes distinctly greenish. 15 mm. (*Hemerophila vicarialis Zeller*; betuhperda Dyar.)

Larva translucent greenish yellow. Head pale reddish with a black line on sides behind; tubercles black; body with a clear white dorsal, and blurred lateral lines; under a web on upper side of leaf of birch; eating upper parenchyma only. Cocoon fusiform, with truncate ends, under a carpet, denser than the larval web

Nova Scotia to White Mountains, New Hampshire; west to British Columbia and Utah.

The status of this species is uncertain. The green (diana) and gray (betuliperda) forms are rarely found together, but each covers the range of the species. Viearilis was described as largely light brown. I have seen no such specimens and it may be stained betuliperda. Variants of both green and gray forms occur in Europe. If these European types prove to be a distinct species, diana will have to be used for them and vicarilis is available as the name for our species.

III. Fore wing with costa more arched, apex rounded, outer margin evenly convex, more upright than before, with metallic scaling. Palpi with third joint ehisel-shaped, appearing pointed in side view, but as broad as second, to its apex, in front view (Brenthia Clemens).

4. S. pavonacella Clemens. Fuscous. Lower part of face pale; palpi white with three fuscous rings; antennæ barred above; fore wing mottled with whitish, especially about the middle; a broad black terminal band, containing a streak of bright iridescent scales, often broken into spots, and with a few such scales at middle of costa. Hind wing fuscous, not powdery; shaded with whitish, with a short metallic band near apex. 8 mm.

May; July and August. Larva on Desmodium and Amphicarpa. The moth struts about on alighting, with hind wings displayed like Glyphipteryx, the smaller Anacampses, etc.

New York and Pennsylvania to Brazil, west to Kansas. New York: Ramapo.

3. CHOREUTIS Hübner

(*Porpe* Hübner)

Costa strongly arched (fig. 213), apex more or less marked; anal angle marked only in C. inflatella. Hind wing rounded, the first two anal veins not closely parallel. Palpi with long acute third joint; and long stiff hair-scales on the under side of the second joint (fig. 216), sometimes in two slightly divergent sets. Antennæ sometimes with a triangular mass of projecting scales on lower side of scape.

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Key to the species

1. Fore wing with two silvery streaks from costa; hind wing with a silver 1. Fore wing with silver broken into spots; hind wing with white only.

- 2. Fore wing ochreous and brown at base.
 - 3. With silver markings on costal half of wing outwardly.
 - 4. Fore wing with little or no yellow at base, but with two prominent transverse white fasciæ2. onustana.
 - 4. Fore wing with more yellow at base, and without white fasciæ.
 - 5. 9 mm. Medial area powdery like ground......3. gnaphaliella.

1. Fore wing triangular, with marked anal angle; hind wing with all veins prcserved; R running to apex (Choreutis).

1. C. inflatella Clemens. Dark brown, with some other toward apex; disc dusted with white; some silvery marks near base; a curved postmedial band and a straight subterminal one converging toward anal angle; a couple of white spots on costa. Fringe pale, with fuscous center-line. Hind wing fuscous, with a silver subterminal bar, and dot at anal angle. All the silver typically greenish but violet in the commoner variety virginiella Clemens. 9 mm. June and July; September. Larva skeletonizing Scutellaria lateriflora, in a slight web bending the leaf up and the edges together; in early September. Cocoon

usually in an uneaten leaf.

Toronto, Canada, to North Carolina and Washington. New York: Wilmington, Batavia, Ithaca, West Farms.

II. Fore wing blunter, with more arched margins, and anal angle not marked; \mathbf{M}_{a} of hind wing normally lost; **R** running to costa (Porpe).

2. C. onustana Walker. Brown; yellowish at base of cell, with two lead-gray streaks at base of wing and several spots outwardly in black patches; terminal line double, fuscous. 9 mm. (ohioensis Želler).

June and July.

Canada to Pennsylvania, British Columbia, and California. New York: Mt. Whiteface, Uphill Brook (Mt. Marcy), East Aurora, Ithaca.

3. C. gnaphaliella Kearfott. Light brown or fuscous, with bright othre yellow patches at base, defined by the lead-colored streaks on costa, Cu, and inner margin. Outwardly, with lead-colored and black patches as in C. onustana. With whitedusted areas, tending to form a band across apex from middle of costa to middle of outer margin, and a broader and less regular one from before middle of costa to anal angle, the two often connected along lower edge of cell. Hind wing fuscous, with a short white line. 9 mm.

Generally distributed. May to July; September and October. Larva on Antennaria and Gnaphalium polycephalum; a leaf miner when young later in a sticky web mixed with frass; gregarious in spinning its cocoon.

4. C. carduiella Kearfott. Closely similar to C. gnaphaliella; the white dash on the hind wing, on the average, weaker, sometimes wanting; the two powdery areas almost always entirely separate; the median area, and the subterminal region before the black mark, even bright chocolate brown. 13 mm.

Larva pale yellow. Head, cervical shield, and true legs light brown; a black spot low down on side of head; tubercles black, stronger on thorax. Social, in pith of stems of Carduus spinosissimus, in June. Two broods. Moth in July and in November. Very close to C. pretiosana of Europe.

5. C. extrincicella Dyar. Light brown. Head paler, fore wing with a broad whitish antemedial band, edged outwardly toward costa with silver, and filled with silver below. Outer half, except extreme margin, cream, shaded with ochreous and light brown; above, with black streaks; below, with a large black patch containing two silvery spots; a yellow-brown terminal band. Hind wing whitish. 12 mm. (Millieria).

June.

Western Pennsylvania; Wisconsin; Regina, Canada; California. 6. C. leucobasis Fernald. Head, thorax, and base of fore wing white; or pale gray with a white antemedial band. Outer two-thirds of wing dark fuscous, dusted with white, and shading into a white central patch; three oblique white streaks near apex of costa, the largest at apex. With scattered metallic spots, especially below the white patch. Hind wing fuscous. 12 mm. (*Millieria*).

June to September.

Vermont, Massachusetts, and Ontario to British Columbia.

4. GLYPHIPTERYX Hübner

Palpi upturned about to vertex, smooth; ocelli present. Fore wing long and narrow, with strongly oblique outer margin and sometimes with marked anal angle (figs. 214 and 215); the apex tending to be subfalcate, but rounded off. Hind wing narrower, and much shorter; typically oblong; sometimes lanceolate, but with rounded apex; R sometimes running to apex. Cell very small in ample-winged forms. M_a connate, stalked, or united with Cu_i . Hind tibiæ smoothscaled. Quinqueferella of California is not a Glyphipteryx but a Hilarographa.

1. G. saurodonta Meyrick. Dark bronzy gray. Head with a white line above eyes; palpus with three whorls of white-tipped black scales; apex of third segment black with white lateral lines. (Wing form and venation not stated.) Fore wing with a semioval white dorsal spot before the middle; six white, dark defined costal striæ, the first two oblique, extending half way across the wing, the first at a third way out, and the second at the middle; the others shorter and erect, the last two being approximated. A wedge-shaped dorsal streak beyond the middle, nearly meeting the second costal, and a shorter erect subterminal streak with a whitish area before it. Two lead-colored metallic spots on disc and three on dorsal half of outer margin. Apex blackish with a silvery dot. Fringe gray, dark at the base; apical hook black, with a white streak below it. 10 mm.

September.

Toronto, Ontario. This species is unknown to me.

2. G. circumscriptella Chambers. Hind wing narrow and short, but trapezoidal;

the fringe wider than the wing; venation normal; M_a and Cu_i connate. Dull grav-brown with slight iridescence; a large white dorsal triangular streak with straight outer, and slightly bent inner, edge; hardly dark-edged, and running two-thirds way across the wing; beyond, with two erect silvery costal streaks, extending half way across wing, and one dorsal one. Speculum black, extending to a rudimentary second outer dorsal streak, interrupted by two silver spots, containing also a couple of silvery and several yellow spots, and with a black-dusted, straw-yellow patch above it. A curved blue streak from middle of outer margin to three-fourths way out on costa and a white subterminal bar beyond it. 12 mm.

Fabiola shalleriella has been mistaken for this species, but can be easily distinguished by its much longer palpi and hairy hind tibia. July.

Mt. Wachusett and Amherst, Massachusetts: Essex County. New Jersey. The species is in the Robinson collection, presumably from New York.

3. G. quadragintapunctata Dyar. Palpus apparently like that of Abrenthia, but wing with markings of Glyphipteryx. Fore wing dark brown; the median area

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darker, sprinkled with golden dots on the dorsal three-fifths; apex ochreous, a series of yellowish white, more or less opalescent dots on costa, the second and fifth continued into complete transverse lines, the fifth extending to the anal angle, the fourth oblique to the middle of the wing, the seventh a short line, and the eighth to the middle of the outer margin; no spot at middle of inner margin. Illind wing brown. 14 mm.

This species may fit better in the South American genus Machlotica Meyrick. Onaga, Kansas,

4. G. impigritella Clemens. Fore wing (fig. 215) with sublanceolate subfalcate membrane; cell nearly two-thirds of the length of wing; 1st A wholly lost; basal fork of $\mathfrak{L}\mathfrak{d}$ A obscure; hind wing half as wide as fore wing, lanceolate, with fringe wider than membrane and reduced anal region; with base of R wholly lost; R and \mathbf{M}_1 connate; \mathbf{M}_3 lost. Olive brown; fringe paler, grayer, and glistening; apex of membrane with a black spot. Markings silvery white, edged more or less with black, especially on their inner sides; basal half of costa immaculate; outer half with five transverse bars, the first two converging at the middle of the wing and nearly meeting a very large crescentic one from before the middle of the inner margin and another running directly across from the inner margin; third costal bar smaller, solitary, opposite a small one at anal angle; fourth and fifth bars close together, near the apical spot, with a single corresponding one in the dorsal fringe. Fringe white with a heavy black line in the base. 7 mm. (*cxoptatella* Clambers).

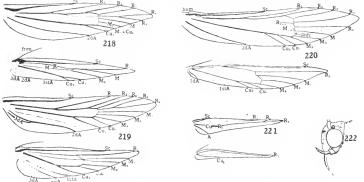
The moth flies in June in North Carolina.

New Hampshire to Texas and California.

Family 26. **HELIODINIDÆ**

(Lavernida, Elachistida, in part; Tinageriida, in part, etc.)

Head smooth-scaled, with rather small eyes; antennæ normally about as long as fore wing, (short in Cycloplasis), with small first joint and



FIGS, 218-222. HELIODINIDÆ

218. Heliodines resella (Europe), venation; 219. Euclemensia bassettella, venation; 220, Schreekensteinia species, venation; 221, Cycloplasis panicifoliella, venation; 222, Heliodines species, side view of head

annulate shaft; palpi varying from very short to moderate. sometimes drooping, apparently socketed in face; maxillary palpi obsolete; tongue

normally strong and naked; ocelli variable; hind tibiæ smooth-scaled or with stiff bristles; hind tarsi with more or less distinct groups of bristles at ends of segments. Fore wing lanceolate or linear; hind wing narrower; cell usually open below ${f M}$; and usually reduced in venation.

In delimiting this family, I have blindly followed Meyrick's revision. It appears homogeneous except for Euclemensia, which lacks the enlarged tarsal spinules and may prove to be a new family type. The most characteristic feature of this family is the resting position, the middle or hind legs being always, so far as known, conspicuously displayed, either raised, or held out laterally. Heliodines, Stathmopoda (Erineda). Schreckensteinia, and Cycloplasis are known to do this, as well as various exotic forms. Meyrick admits about 225 species. Stathmopoda is the only considerable genus.

The early stages are various. The egg of Schreckensteinia is upright; those of the rest I think are unknown. The larvæ are various (see Euclemensia, Schreckensteinia, Heliodines, Cycloplasis). The pupa of Schreckensteinia (q. v.) is known. It is formed in a lace cocoon. The parasitic Euclemensia pupates in the host.

Key to the genera

- 1. Hind wing banded with silver.....4. Idioglossa. 1. Hind wing immaculate.
 - 2. Fore wing with \mathbf{R}_5 preserved, running to costa (fig. 219).
 - 3. Hind wing nearly as wide as fore wing; hind legs unarmed.

1. Euclemensia.

2. Fore wing with four veins or less running from cell to costa.

- 3. Venation fairly complete.
 - 4. Hind wing lanceolate; M_3 lost......2. Heliodines. 4. Hind wing with all veins preserved, or linear and very narrow with all

1. EUCLEMENSLA Grote

(Hamadryas Clemens, not Hübner, etc.)

Head smooth; palpi upturned to vertex, smooth; maxillary palpi very minute, porrect. Tongue small, scaled; antennæ slightly rough-scaled. Tarsi with spinules very weak, tending to gather at tips of segments. Fore wing (fig. 219) with costa concave at middle, dorsal margin arched; hind wing nearly as broad, bluntly lanceolate, without emargination on costa; all veins widely separate. \mathbf{R}_{i} obsolete.

Caterpillar an internal parasite on the oak soft scale, Kermes; forming a crescentic chamber in the body of the host, which finally becomes very hard and gall-like. Caterpillar white, with brown head, very plump, and strongly coneave dorsally. Seta minute, not yet worked out. Prolegs rudimentary, with a circle of uniordinal books. Pupa in the infested gall; not studied.

The caterpillar cuts a sort of lid in the Kernes for the moth to emerge. This is a curious and entirely isolated genus, being the only known internal parasite in the order. It is usually placed in the family (Ecophoridae, as its

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venation would indicate, but the wing form and maxillary palpi are not (Ecophorid. 1. E. bassettella Clemens. Bronze-black; palpi yellow. Base of fore wing orange-red, bounded by a raised bar on inner half and a dot on costa, and marked with a black bar from base of costa, and streak on inner margin. Outer two-thirds nearly black with an orange-red band running through the middle of the wing to the costa before the apex, leaving only a slender black streak. 15 mm.

July and August.

New Hampshire, and Ontario to Texas. New York: Ithaca (Howard, W. T. M. F.), Nyaek (Zabriskie); Flatbush, Long Island.

2. HELIODINES Stainton

·(*Ætole* Chambers)

Palpi smooth, third joint pointed (fig. 222), antennæ about as long as body: hind tibiæ smooth-scaled; fore wing lanceolate (fig. 218); our species with 10 veins, the two apical forking over the apex. Cu_2 apparently lost. Hind wing narrower, lanceolate, with **R** and M_1 long-stalked, forking over apex; M_2 free, connected by a short cross vein to **R**-stem; cell open below it; our species with Cu-stem simple. Ocelli present. Larva of European species in a slight web, on Chenopodiaceæ.

1. H. bella Chambers. Fore wing deep orange; base marked with mouse-gray; also, with two gray spots on costa and two on dorsal margin; costa, outwardly. and inner margin, from beyond middle, edged with mouse-gray; fringes, body, and hind wings mouse-gray. The fore wing with the small gray spots raised, and with similar raised tufts in the gray border. 7 mm.

The moth has been taken on Chinquapin bloom.

Kansas and Kentucky to California and Texas.

2. H. nyctaginella Gibson. Similar, larger; costa with gray edge at base only. followed by four raised spots; inner margin with no separate spots; the antemedial tuft connected to the base, and the postmedial tuft to the apex by gray stripes. 10 mm.

August. Larva on Oxybaphus nyctagincus.

Madison, Wisconsin; Manitoba; Ames, Iowa.

3. ERINEDA Busck

(*Stathmopoda*, in part)

Antennæ two-thirds as long as fore wing, without pecten, but with whorls of long hair in male. Palpi upturned, slightly thickened, third segment rather shorter than second. Scales small, slender, and smooth. Tongue small; maxillary palpi apparently absent; hind tibiæ with tufts of spines above the spurs, the outer end-spur as long as the inner, and both long. Tarsi also with whorls of spinules. Fore wing slender, but wider than in the European genus Stathmopoda; lanceolate; with 11 veins; \mathbf{R}_1 and \mathbf{R}_2 approximate; \mathbf{R}_3 short stalked with \mathbf{R}_{1+5} all running to costa; \mathbf{M}_1 absent, \mathbf{Cu}_2 arising well back on the cell; \mathbf{M}_2 , \mathbf{M}_3 , and \mathbf{Cu}_1 equidistant. Hind wing half as wide, lanceolate; veins all free but crowded; cell open above M_2 . R_1 apparently free from the short Sc in the hind wing, as in Stathmopoda.

This genus is most probably Heliodinid, but will ordinarily be sought in the Lavernidæ. The moth of Stathmopoda rests with the middle legs extended laterally and the tufted hind legs raised.

I. E. elyella Busck. Brilliant golden bronze; the costa and inner margin con-trastingly pale golden in most lights. 9 mm.

East River, Connecticut.

4. IDIOGLOSSA Walsingham

(Metamorpha Frey and Boll, not Hübner, Walker, or Stainton; Idiostoma Walsingham)

Vertex and front smooth scaled; the front strongly narrowed below, as in Coleophora. No ocelli; scape of antennæ very long; pecten apparently absent; a notch in base of shaft. Palpi widely divergent, second segment slightly thickened, third as long as second. Maxillary palpi drooping, clothed with a conspicuous tuft of hair-scales; tongue scaled; hind tibiæ with two or three regular series of bristles. Mid-tibiæ smooth. Wings linear; \mathbf{R}_5 running to costa; cell low in wing, oblique, like Colcophora; one dorsal vein lost. Hind wing linear.

This is a very striking genus, distinguishable from all our other narrow-winged Tineina by the metallic bands on the linear hind wings. The relationship is entirely uncertain. The genus shows resemblances to Coleophora, Cosmopteryx, Batrachedra, and the Gracilariidæ. There is another species in South Africa, and two more in India.

1. I. miraculosa Frey and Boll. Straw yellow. Palpi and maxillary tuft nearly white. Fore wing with an oblique brownish fascia a third way out, nearer the base on dorsal margin, sending a point outward along the middle of the cell, and edged with violet-silver. Another fascia running obliquely outward from the beginning of the costal fringe, edged within with silver; and brownish fuscous streaks in the fringe below. Extreme apex silvery. Hind wings nearly concolorous, with three silvery fasciæ somewhat edged with brownish, the outer one followed by a stronger brown shade which extends into the fringe. Two brown scale-tufts in dorsal fringe of fore wing and one on hind wing. 10 mm.

scale-tufts in dorsal fringe of fore wing and one on hind wing. 10 mm. Caterpillar on *Panicum claudestinum*, in a translucent white silken tube on the under side of the blade, attached through a hole to the upper side of the leaf, which the larva skeletonizes. Pupation in the tube, which is trussed up and enclosed in loose silk.

Southern States, north to southern Ohio.

5. CYCLOPLASIS Clemens

Antennæ very short, hardly more than half as long as body; palpi short, divergent, oblique in life, slightly curved, with terminal bristles; with flat scale-masses beside the tongue, possibly representing the maxillary palpi; Tongue somewhat longer than face. Fore wing linear-lanceolate (fig. 221); R near costa, simple, fading out at base; a vein through middle of wing, furcate over apex (representing \mathbf{R}_s and \mathbf{M}); Cu very short and simple. Hind wing linear, abruptly broadened at base, with a simple vein running to apex, and a fragment of \mathbf{Cu} in the basal lobe. Middle and hind tibiæ and tarsi heavily spined; hind legs elevated in the

The larva lives in a long linear mine, which is later enlarged into a blotch. When mature, it cuts out an oval piece of the mine, and folds it lengthwise to form a cocoon, in which it drops to the ground and anchors itself. A mass of white froth is ejected from the end of the cocoon. I suspect the pupa is obtect. This isolated genus is placed here by Meyrick; the pcculiar cocoon would sug-

This isolated genus is placed here by Meyrick; the peculiar cocoon would suggest rather the Heliozelidæ, but the food and maxillary palpi may indicate more direct kinship with Idioglossa.

1. C. panicifoliella Clemens. Head and thorax lead-color; antennæ brown, with silver toward base; fore wing umber brown, more or less violet, with a broad straight bright silvery fascia a third way out, and suffused with silver in apical half, especially toward costa; fringe and hind wing violet brown; fringe of hind wing fuscous.

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The larva works in Panicum clandestinum early in July; the mine begins at the base of the leaf and runs to the tip and part way back before the enlargement begins to be formed.

Distribution probably general; moth in late July and August.

New York (?); Pennsylvania; Ohio.

6. SCHRECKENSTEINLA Hübner

(Chrusocoris Curtis)

Rather similar to Epermenia and possibly related to it. Head similar; no pecten; fore wing similar (fig. 220), with Sc much longer than 2d A, not distinctly falcate, but lanceolate; without dorsal tufts; all veins present; \mathbf{R}_3 ending just below apex. Hind wing with \mathbf{R} running to just below apex and never stalked with \mathbf{M}_{i} . Hind tibiæ with a single series of bristles above; palpi short, divergent.

Egg of upright type. Larva with set iv and v separated only in first stage, then on one tubercle; setæ i and ii glandular, as in some Pterophorids, adjacent at base. Prolegs long and slender, with only 4 to 6 hooks in a circle at the tip. Cocoon of regular meshes; the larval skin thrown out through a hole. Pupa incomplete; emerging from the cocoon; with segments 3 and 6 free, and 7 in male; with heavy dorsal spines and recurved hairs on fifth to ninth segments of abdomen, aud small ones on the third segment also; spiracles on high cones; maxillary palpi very small, dehiscing with legs; palpi and femur exposed; first legs touching antennæ at base; the others reaching tip of wings; prothorax minute; headpiece larger.

The genus curiously combines Yponomentoid and Pterophorid characters, and was for a time placed in the Pterophoridæ. Meyriek puts it here, however, but Spuler, in the Scythrididæ, with Epermenia.

1. S. erythriella Clemens. Reddish fuscous, with a more or less distinct greenish brassy hue; palpi ochreous with a fuscous tip; fringes fuscous. Hind wings reddish fuscous with concolorous fringes. 9 mm.

August. Larva in July in fruit racemes of sumae. Strongly moniliform, with raised tubercles; dark green, with rather small, pale brown head. Cocoon of large meshes; pupa green; on outside of raceme.

Distribution probably general. New York: Ithaca, Albany. 2. S. feliciella Walsingham. Deep bronze brown; browner and darker than S. erythiella; palpi with dark third joint. 9 mm.

This species was bred from Orthocarpus. The eastern record is very likely in error for an aberration of S. erythriella.

Pacific Coast; Hazelton, Pennsylvania.

.3. S. festaliella Hübner. Fore wing light olivaceous and brown, with a slight golden irideseence; contrastingly veined with dark brown. 12 mm.

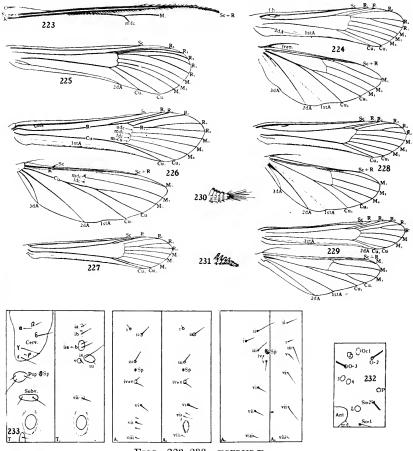
April and May; July and August. Caterpillar green with brown head, on Rubus leaves, in a slight web; the cocoon as in S. crythriclla.

Known from Massachusetts to western Pennsylvania and Michigan; Europe. New York: Ithaca, Rock City.

Family 27. ÆGERIIDÆ

(Sesiidæ; Sphingidæ, in part)

Head rather small; body often stout; mostly smooth-scaled; sometimes with rather finer hair-scales; ocelli present; palpi moderate; upturned often to vertex, normally smooth-scaled, without bristles. Antennæ usually fusiform, tapering to both base and apex, the terminal segment (fig. 230) ending in a minute tuft of bristles (terminal joint in Bembecia enlarged and public entire, fig. 231); sometimes pectinate,



FIGS. 223-233. ÆGERIIDÆ

223, Egeria apiformis: costa of hind wing, showing recurved spines on vein R; 224, Melittia cucurbitæ, venation. (In this and the following figures the costal region of the hind wing is shown a little more spread out than it is actually on the wings.); 225, Egeria apiformis (Europe), venation of fore wing; 226, Conopia (Sanninoidea) critiosa 2, venation; 227, Conopia (Chamasphecia) tipuliformis, venation of fore wing; 228, Albuna pyramidalis, venation; 229, Bembecia marginata, venation; 230, tip of right antenna of Egeria apiformis; 231, tip of right antenna of Bembecia marginata: 232, arrangement of ocelli of larva of Egeria apiformis, penultimate stage, typical of the family; 233, Memythrus tabaniforme (Europe), seta map

and when pectinate, with dorsum evenly scaled, with two widely overlapping rows of scales to a segment, often with only the tips of one row exposed; when not pectinate, with base usually fully scaled, middle of shaft with the outer whorl of each segment continuous, and the inner interrupted by a broad sensory area below; in both forms with most of the swollen tip covered by a sensory area, which bears only seattered scales. Maxillary palpi minute, of porrect type; tongue strong, naked; legs often with stiff bristly hair gathering in tufts at the spurs; sometimes with fine hair also. Fore wings (figs. 224-229) very narrow, with short outer margin, and well-marked anal angle; often transparent; \mathbf{R}_{5} stalked or united with \mathbf{R}_{4} ; accessory cell and base of medial lost; Cu, arising well out toward apex of cell; anal region extremely reduced, the anal veins weak or rudimentary. Hind wing usually broader, but narrow; more or less extensively transparent in our species, often with only the margins and veins scaled; costal edge (fig. 223) with Sc and R closely parallel, becoming coincident outwardly, and bearing a series of recurved spines, which interlock with a similar series on the inner margin of the fore wing. Cell usually squarely closed; anal region fully veined. Frenulum simple in both sexes. The sex is most easily determined by counting the number of visible abdominal segments, there being six in the female and seven in the male; or by the larger male antennæ. The body markings on the basal part of the abdomen are usually alike in both sexes, but the last band is very generally a segment farther toward the apex in the male than in the female. Anal tuft always conspicuous in male, simple or rudimentary in female.

Egg of the flat type, oval. Larva (figs. 232, 233) white; without markings; boring. Front nearly or quite reaching vertex; tubercles usually moderate (obsolete in Melittia). Posterior addorsal setæ on cervical shield nearer middle line than anterior; abdomen with tubercles i and ii separate, iv and v adjacent; spiracle of eighth segment above the level of the others; spiracles elliptical. Ventral prolegs with two transverse rows of uniordinal hooks; anals with a single row. Pupa incomplete, usually formed in the tunnel; with only a couple of basal segments immovable, and even these loosening on dehiscence. Labial and maxillary palpi exposed; tongue and antennæ not reaching the tips of the wings; the antennæ normally swollen toward tip; prothorax wider than dorsal head-piece. Last segment with ventral spines, but cremaster not even represented by specialized setæ. Two rows of spines to each segment dorsally.

The connection of this family with the Yponomeutoidea is unmistakable but, at least as represented in the United States, it is clearly defined both in adult and pupa.

Key to the genera

1. Antennæ with last segment minute; bearing a tuft of bristles (fig. 230).

- 2. Cu₁ in hind wing much nearer Cu₂ than M_3 at origin (fig. 224). 2. Melittia. 2. Cu_1 approximate, connate, or stalked with M_3 .
 - 3. M₂ and M₃ of fore wing strongly downcurved; Cu₁ and Cu₂ straight (fig. 225)
 - M₂ and M₃ practically straight (fig. 226).
 4. M₃ and Cu₁ approximate at origin in hind wing.....l. Memythrus.
 - 4. M₃ and Cu₁ stalked.
 - 5. Fore wing with $\mathbf{R}_{\mathbf{i}}$ absent, indicated only by a slight fold and thickening

Key to the known larvæ

(After Dyar)

A. Segments 3-annulate; tubercles distinct.
1. First annulet the highest.
2. A black band across bottom of clypeus
2. A brown shade instead; black spots on the sides.
Conopia exitiosa, Ægeria apiformis.
1. Second annulet highest.
2. Crotchets on abdominal feet 6 to 9, in a row.
3. Brownish, with distinct, paler tuberclesParharmonia pini.
3. White, tubercles indistinct
2. Crotchets 9 to 14 in a row.
3. Slender; head small
3. Stouter; head larger.
4. Lower ocellus developed, though light.
5. Crotchets normal
5. Crotchets very small, reducedBembecia marginata.
4. Lower ocellus replaced by a pale spotC. albicornis, C. rutilans.
2. Crotchets 15 to 18 in a row (epicranial lobes meeting in a point).
C. pictipes, Podosesia syringa.
2. Crotchets 18 to 22 in a row; epicrania broadly touching.
Memythrus 3-cinctus.
1. Third annulet higher than the others
B. Tubercles absent; annulets absent
SYNOPSIS OF FOOD-HABITS OF THE LARVA ²⁵

Boring in trees:

τ	'nder bø	ırk of	trunk	some	distance	from	\mathbf{the}	base	or in	the	branches:
	Maple										Conopia acerni.
·	Maple										Conopia corni
	Apple,	Pear									Conopia pyri.

²⁵ Benutenmüller, William. Monograph of the Sesiidæ of America, North of Mexico. American Museum of Natural History. Mem. 16 (215-352) (Reference on p. 229-230). 1901.

Dogwood, Oak, Chestmit
Cherry, Plum, Juneberry,
Peach, Cherry, Plum, Apricot, etc
Ash
Oak
Poplar
Willow and Poplar
Willow
Willow
In solid wood at base of trunks and roots of trees: Persimmon
Willow and Poplar
Willow
Boring in shrubs.
In solid wood:
Lilae
RhododendronConopia rhododendri.
In pith of stems:
Currant, GooseberryConopia tipuliformis.
In roots:
Blackberry and Raspberry
Blackberry and RaspberryBembecia marginata. Boring in vines and creeping plants.
In the stems:
Squash, Pumpkin, and other Cucurbs
In roots:
Grapevine
Clematis
Herbaceous perennial plants.
In roots:
Strawberry
Ileliopsis
Oak-gall (Andricus cornigerus)
Oak-gall (Andricus cornigerus)
Gall of Saperda concolor
Gall of Saperda concolor

1. MEMYTHRUS Newman

(Sciapteron Staudinger; Paranthrene Hübner, in part; Tarsa Walker; Fatua Henry Edwards; Albuna Henry Edwards)

Palpi upturned, normally with long hair below. Antennæ fusiform, normally bipeetinate in male, the pectinations clavate. Tongue weak. Hind tibiæ with a little longer hair, but not tufted. Abdomen with a bunch-like anal tuft, and short spreading side tufts, or four pencils. Fore wing with \mathbf{R}_{τ} sometimes shortly stalked, closure of cell oblique, hind wing with \mathbf{mdcv} more oblique and longer than \mathbf{ldcv} , and \mathbf{M}_{z} arising below middle of cell.

The two known American larva have brown-mottled heads with a black band across the face.

Key to the species

I. Male antennæ bipectinate except on the club (Tarsa Walker).

1. M. asilipennis Boisduval. Body, veins, and borders of wings dull brown; reddish in cell. Tegulæ rufous; segments of abdomen narrowly edged with yellow; legs orange; all wings of male and hind wings of female transparent; bar at end of cell oblique and rather heavy. Fore wings of female deep brown, with a transparent triangle at anal angle and a streak at base. 32 40 mm. (female larger).

Larva in ash and alder roots.

New Hampshire and Minnesota to Florida and Texas. New York: Rochester Junction, Buffalo, Staten Island; Brooklyn and Amagansett, Long Island.

2. M. simulans Grote. Blackish; face with two yellow stripes; front of palpi 2. M. simulans Grote. Blackish; face with two yellow stripes; front of palpi yellow; scape yellow; collar partly yellow; two yellow spots on tegulæ; and scutellum yellow. (In the type form the base of the abdomen is black, the next three segments yellow, narrowly black at base of each, the fifth and sixth black, overlaid with loose greenish-looking yellow hair; and the apical tuft yellow and reddish. Legs yellow, becoming orange on tarsi, the tibiæ concolorous. Fore wing brown, strongly reddish in middle of wing, with outer third transparent below \mathbf{M}_2 and with two transparent streaks on basal half. Hind wing transparent. This extreme form is little known and was described from Algonquin, Illinois, in June.) (**H 46**:10.)

a. M. simulans, var. luggeri Henry Edwards. Similar: male with transparent areas often meeting below cell. Abdomen black at base, with a narrow stripe on second segment only; the middle segments yellow on sides, and the tail solidly yellow; scape black, except below; fore tibia with considerable black hair above; middle and hind tibie with small spots. 27-32 mm.

Late May to July. Larva in wood of black and red oaks; very rarely in white oak or chestnut. (The single specimen I have seen bred from chestnut has the yellow replaced with orange.)

The larva takes two years to reach maturity; the moth flying in the vicinity of New York in odd-numbered years. Forms intermediate between variety *luggeri* and the type are not rare.

Maine to Maryland. New York: Staten Island; Jamaica. Long Island (all of variety luggeri).

3. M. palmii H. Edwards. Similar to *M. simulans*, but usually with a larger extent of more orange-yellow on the body. Spot on tegulæ inverted-comma-shaped, nearly reaching collar. Larva on oak.

New York City to Florida and the Pacific Coast. New York: Flatbush and Yaphank, Long Island.

4. M. scepsiformis H. Edwards. Dull black; antennæ in part, neck, palpi, legs in part, and base of hind wing rufous. Legs banded with black; second segment of abdomen with a yellow stripe. Border of hind wing very narrow, deeply extended in, along Cu, 1st A, and 2d A. 25-30 mm.

Male not seen; possibly a variety of polistiformis.

Maryland; Kansas; Texas.

5. M. polistiformis Harris. Brown, more or less iridescent with purple; with orange markings, as in *M. scepsiformis*, and also normally with orange bands on abdomen and almost wholly orange legs. A yellow band on fourth as well as second segment of abdomen; male with a transparent streak at base of fore wing, and with abdomen ending in five pencils, as in Sannina. 22-40 mm. (H 46:11 J 12 Q.)

Larva boring in grape roots, and sometimes injurious.

Vermont to Minnesota, South Carolina, and Missouri.

II. Male antennæ subpectinate and fasciculate; palpi hairy, but less so than in the simulans group.

6. M. tricinctus Harris. Brownish; front of palpi and neck yellow; legs black and orange; abdomen with yellow bands on second, fourth, and sixth segments, and in male on seventh also. Fore wing black with transparent streaks at base, the anal angle becoming transparent in rubbed specimens; hind wing transparent. Abdominal tuft short and massive. 25-28 mm.

June to July. Larva boring in willow and poplar branches.

Quebec to New York and Michigan. New York: Buffalo, Lancaster, Big Indian Valley, Karner, Long Island generally.

7. M. dollii Neumægen. Thorax and wings deep brown; abdomen deep brown, becoming more or less chestnut brown toward apex; the whole thorax and abdomen chestnut in variety castaneus Beutenmuller. Palpi dark. Fore wing a little transparent at base; hind wing with a broad brown border, somewhat diffuse, but with no special tendency to fill cell Cu_1 . Legs dull orange; hind tibia dull fuscous below. Strongly variable both in size and markings. 20-38 mm.

Larva in solid wood of young poplars.

Southern New York; New Jersey; Pennsylvania; Illinois. New York: Brooklyn.

III. Male antennæ somewhat servate, and less heavily fasciculate; palpi hairy; terminal tuft bifurcated (Albuna Henry Edwards).

8. M. pyramidalis Walker. Fore wing transparent, with reddish and blackbrown markings; margin extending about a third way into cell, rather wider toward apex, as in many Conopias. Bar at end of cell heavy and oblique. Border of hind wing narrower than fringe. Abdomen more or less striped with yellow. Palpi yellow in front. Legs black and yellow. 25 mm. (montana Henry Edwards.)

June to August.

Newfoundland and New York to the Pacific Coast. New York: Axton, Big Indian Valley; Brooklyn, and Bayshore, Long Island. The record for "morrisonii" from Watkins Glen doubtless refers to this species.

The female form coloradensis Henry Edwards, with a solid black body and legs, appears to be generally distributed with the species.

⁹. M. fraxini Henry Edwards (Q morrisonii ¹Henry Edwards), with solid black fore wings, abdomen, and legs, occurs east to Missouri. It may be distinguished by a bright red bar at the end of the cell.

2. MELITTIA Hübner

(*Trochilium*, in part)

Antennæ strongly flattened, with bristles along anterior edge; palpi like those of Memythrus, rather hairy. Hind tibiæ and tarsi heavily clothed with hair. Fore wing (fig. 224) with \mathbf{R}_s terminating below apex; hind wing with m-cu long, and set at an angle to base of Cu, so that the lower side of the cell appears curved. Larva boring in Cucurbitaceae; leaving the burrow to pupate in the ground. Segments not distinctly annulate; tubercles lost, and set \mathbf{m} induces the set \mathbf{M}_s is the transformed and the set \mathbf{M}_s is the transformation of transformation of the transformation of transformation of the transformation of the transformation of transforma

The genus also occurs in Africa and India. Some western species are fully scaled.

1. M. satyriniformis Hübner. Black with green iridescence; palpi and fore coxæ with orange and white markings; fore wing transparent at base; hind wing transparent; tibiæ and hind metatarsi orange and black; the rest of tarsi black and white. 25-30 mm. (H 46:1.)

Often injurious to squash, pumpkin, and other Cucurbitaceæ; but the imago rarely seen. June to August.

Generally distributed, New York: Wells, Albany, New York City, Staten Island, Long Island.

3. PODOSESIA Möschler

(Grotea Möschler, not Cresson)

Head normal; antennæ fusiform and fasciculate in male; normal. Tongue strong; abdomen with a short terminal tuft in both sexes; hind tibiæ slightly tufted at the spurs; tarsi extremely long, with rough bristling hair the whole length, becoming erect scales on the metatarsus. Fore wing with \mathbf{R}_s terminating rather above the apex; hind wing with \mathbf{M}_s and \mathbf{Cu}_1 stalked for a variable distance.

1. P. syringæ Harris. Deep brown, with more or less tawny, especially on palpi. An oblique yellow streak on side of abdomen at base, and a small lateral spot at middle; fore wings with purple iridescence, and often a red streak above \mathbf{R}_3 ; hind wing transparent, yellow, with a narrow, diffuse, scaled edge. Tibiæ black and orange; tarsi yellow and black. 25-35 mm. (Ægeria Harris.) (H 46:17.)

Larva in solid wood of ash and lilac.

Canada to Colorado and Texas. New York: Albany, Long Island.

4. SANNINA Walker

Palpi scaled, somewhat roughly; antennæ lightly fasciculate. Tongue moderate. Hind tibia with rough hair, which is longer at the spurs; metatarsus only with rough raised scales like Podosesia. Abdomen of male ending in five pencils, the two lateral short and stout, the middle one slender and easily lost. Wings almost fully scaled (unlike the few Conopias which have nearly the same hind tibiæ); fore wing with \mathbf{R}_5 running to outer margin, normal; hind wing with \mathbf{M}_3 and \mathbf{Cu}_1 rather long-stalked; cell short.

rather long-stalked; cell short. 1. S. uroceriformis Walker. Deep blue-black; palpi and base of tegulæ sometimes orange. Fourth segment of abdomen red dorsally, with a narrow blackish center line. Wings opaque, except for a small transparent area at base between Cu and 3d A of hind wing, the transparent spot in the female divided by a scaled area between 1st A and 2d A. 30 mm. (quinquecaudata Ridings). (H 46:7.) Boring in roots of persimmon, well below the ground.

District of Columbia to Kansas, and south. New York: Albany.

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5. PARHARMONIA Beutenmüller

(Harmonia H. Edwards, not Mulsant or Haswell)

Palpi almost straight, close-scaled; antenna finely public below. Abdomen flattened and rough-haired at sides; hind tilbie slightly tufted at spurs; \mathbf{R}_{s} running to apex, hind wing with **ldcv** long and oblique. Hardly distinct from Conopia, but with the tilbia less rough; our only species easily recognized by the orange under side of the abdomen. Larvae boring in conifers, with the skin darker than the tubercles.

1. **P. pini** Kellicott. Deep brown: fore wing slightly translucent; hind wing with scattered smoky scales; the cell and cell **1st A** not specially darkened. Abdomen brown above: fourth segment, above, sides, and under side mostly orange. 28 mm.

July. Larvæ boring in trunk of pine, usually in groups; a good deal of pitch oozing from the borings; the larva keeping an air-hole open through the pitch. Pupa in a cell in the mass of pitch. On pitch pine, white pine, and spruce.

Canada and New Hampshire to New Jersey. New York: Oswego County, Buffalo, Portage, Ithaca, Hudson, Hastings Center.

6. CONOPIA Hübner

(Sesia Fabricius, in part; Trochilium Meyrick, etc., not Staudinger; *Ægeria* Walsingham, not Beutenmüller, Meyrick, etc.; Synanthedon Hübner; Pyrrhotania Grote; Carmenta Henry Edwards; with Sanninoida Beutenmüller and Chamasphecia Spuler)

Palpi normal, with rather rough bristling vestiture; sometimes flattened below. Abdomen with terminal tuft of male moderate, spreading faulike, simple, biffd as in Albuna, or spearhead shaped (*exitiosa*); when simple, sometimes with small side-tufts; quite variable in appearance in most species, according to the state of expansion, and so hardly usable for the subdivision of the genus. Hind tibiæ with well-marked bristly tufts at middle and end spurs; smooth-scaled between, and at base; tarsi smooth-scaled. Fore wing with \mathbf{R}_1 and \mathbf{R}_2 typically free (fig. 226), becoming concident near margin in *C. tipuliformis* (Chamesphecia, fig. 227); \mathbf{R}_5 running to rather above the apex; hind with ldcv more transverse than in Parharmonia, often short. Several attempts to divide the genus have not been very successful, especially as some of the aberrant species are only known in one sex. *C. (Saminoida) critiosa* is perhaps most distinct, and shows the strongest sexual dimorphism; but each sex can be closely matched elsewhere in the genus, and there are no decided structural characters. Chamesphecia represents a natural group, but many of the species have not been examined as to \mathbf{R}_1 and \mathbf{R}_2 .

Key to the species

1. Fore wing largely hyaline, with the space below Cu hyaline.

2. Terminal tuft red.
 Interspaces near apex of fore wing light yellow
2. Terminal tuft black, or black and yellow.
 Palpi and middle and hind tibiæ wholly bright orange. 4. Tarsus with metatarsus only orange; wing with smoky fringe only. 9. fulripcs.
4. Tarsus wholly orange; fore wing with fairly broad dark tip. 10. saxifraga.
3. Hind tibiæ at least largely black or light yellow.
4. Abdomen with a single red transverse band.
5. Band on one segment; hind tibia with yellow band-(2.) culiciformis. 5. Band covering two segments.
6. Ground color of tip of fore wing red
6. Tip of fore wing wholly black
5. With seven hyaline interspaces toward margin, cell \mathbf{Cu}_1 being hyaline.
6. Stigma black, narrow. 7. Sides of front yellow; vertex between antennæ black.
11. pictipes.
7. Sides of front concolorous, black; a yellow tuft between antennæ 1. exitiosa.
6. Stigma largely red, contrasting12. rubristigma.
5. With six hyaline spaces, or fewer (Cu_1 scaled, with rare exceptions);
border of fore wing broader. 6. Discal stigma concolorous.
7. Yellow bands on fourth and fifth segments of abdomen, both
strong and equally strong
7. Fifth segment with a very weak band, or with none.
8. Hind metatarsus yellow; palpi yellow, or with some fuscous on upper side; abdominal tuft yellow at edges.
9. Abdomen with stripes on first and second segments nearly
equal; also third and seventh of male, and sixth of female. 15. bassiformis.
9. Abdomen with first segment not striped; a very strong
stripe on fourth segment; and a weaker stripe or none on
sixth or seventh segment
 8. Hind metatarsus black. 9. Abdomen with three strong yellow stripes, on second, fourth,
and sixth segments of female, and on seventh of male. 17. tipuliformis.
9. Abdominal stripes weak or absent; never very strong on
alternate segments.
10. Border extending more than half way in to end of cell. 20. $ithaca$.
10. Border narrower. 11. Abdomen striped more or less with yellow, at least with
a yellow bar on side of second segment; palpi largely
vellow
11. Abdomen wholly black above; palpi yellow below in male; wholly black in female19. albicornis.
6. Discal bar contrastingly red, at least in large part.
7. Stripes on second to sixth or seventh abdominal segments, equal
or nearly so; hind tibiæ yellow or largely so21. rilcyana.

- 7. Stripes on segment 4, or on 2, 4, and 6, much stronger; hind tibiæ largely black.
 - 8. Metatarsus black; moth smaller......13. decipiens.

1. C. exitiosa Say. Male with palpi black above and yellow below; antennæ black; a yellow tuft on vertex, as a rule; front black. Abdomen with variable fine yellow stripes. Hind legs black, narrowly yellow-ringed at spurs, and on segments of tarsi. Fore wing with border, aside from fringe, much narrower than an interspace; all cells below \mathbf{R}_2 hyaline. Scaling wholly black. Anal tuft lanccolate, the scales narrowly white-tipped. Female with fore wings, costa of hind wings, including base of cell, and body and legs purple-black. A red band across middle of abdomen, typically covering one segment (but two in var. edwardsii Beutenmüller). 20-30 mm.; female larger (Q fitchii H. Edwards).

July and August. Larva (the peach-tree borer) in the sap wood of peach, plum, and other Rosaceæ, usually close to the surface of the ground; often injurious in peach orchards. Pupation in a cocoon, rarely outside the burrow.

Generally distributed in the range of its food plant. New York: Buffalo, Ithaca and vicinity, Albany, New Windsor, Staten Island; Brooklyn, and Jamaica, Long Island. Var. edwardsi Beutenmüller is commoner than the typical female about Ithaca and also occurs at Phelps (Ontario County).

In aberration luminosa Neumogen, the head, most of the abdomen, and the costa, and part of the veins of the wings are bright golden yellow.

In the remaining species the sexual dimorphism, though often marked, is never as striking as in exitiosa.

2. C. rubrofascia H. Edwards. Blackish; mid-tarsi and last four segments of hind tarsi whitish; abdomen with two middle segments dull red. Fore wing of male hyaline, with blackish discal bar, and border extending halfway in to cell, leaving five hyaline cells. Hind wing with only the extreme edge blackish; the cell all hyaline. Anal tuft lanceolate. 22 mm.

South Carolina; Georgia; also very probably in our area. New York: reported from New York City.

C. culiciformis Linnæus, from birch, and the very similar C. americana, from alder, are definitely known only from the Rocky Mountains to the Pacific Coast. 3. C. bolteri H. Edwards. Almost like male of C. rubrofascia; tip of fore wing

red between veins; hind tarsus wholly blackish on outer, and paler on inner, side; sexes similar. 15-21 mm.

Larva in solid wood of willow, well above ground. Illinois. New York: Ithaca, Big Indian Valley, Mosholu.

4. C. pyralidiformis Walker. Blackish. Palpi yellow; fore wing obscurely hyaline at base and beyond cell in male, but not forming distinct spots; wholly opaque in female; hind wing like C. rubrofascia. Body fuscous, with a broad yellow band on fourth segment, and fine bands on second and sixth or seventh segments sometimes (usually in Illinois) with the three bands nearly equal. Legs blackish; a yellow spot at upper spurs, and last four joints of tarsus pale. Tuft as before, or spread fanlike (nigella Hulst).

Long Island, New York, to Illinois and Texas. New York: Fairport, Batavia, Staten Island; Amagansett and Woodhaven, Long Island.

5. S. sanborni H. Edwards. Blackish; palpi largely yellow; fore wing with a nearly round hyaline spot beyond cell, covering three cells, and a slight transparency at base. Border of hind wing rather wider than usual. Body with yellow stripes on segments 4 and 6. Female type only known; without hind legs. Possibly an aberration or a northern form of *pyralidiformis*.

Massachusetts.

6. C. rutilans H. Edwards. Palpi, occiput, and two stripes on thorax yellow. Segments 2 and 4 of abdomen with posterior half yellow, 6 a little more nar-

rowly striped, and the other segments narrowly striped if at all. Tuft, when perfect, bilobed, Albuna-like; largely black. Hind legs yellow, black between spurs of tibiæ. Fore wing normally with small hyaline areas in and beyond cell, fully scaled below cell in many females, and always with the outer part of the cell Cu fully scaled. Scaling of fore wing mixed black and yellow. Male usually with more hyaline and black, and less yellow. Border of hind wing wider than usual, with yellow and red scales. 15-22 mm. (*lupini, washingtonia, perplexa, hemizonix, impropria* Henry Edwards). (H. 46:31 J, 32 Q.) Light specimens are very close to C. bassiformis var. sexfasciata, but differ in

Light specimens are very close to *C. bassiformis* var. *sexfasciata*, but differ in that the stripe on the first segment is much weaker than on the second. *Scitula* may be distinguished by the much narrower black bar at the end of the cell (it is as wide as high in *rutilans*, and more than twice as high as wide in the *scitula* group); *ithaca* may be recognized by the black hind legs.

Larva in roots of Veronica, blackberry, raspberry, and strawberry; probably a general feeder.

Nova Scotia to New York and the Western States; doubtless also in intermediate localities in Canada.

7. C. corni H. Edwards. Black, slightly marked with yellow; hind tibia with a black patch; anal tuft contrasting, bright red, with a black center, often spread fanlike. Fore wing with scaling black, border extending only a third way in toward cell. 20 mm.

Larva in small branches on silver and red maple, causing a gall-like swelling.

Massachusetts to Parry Sound, Ontario, and western Pennsylvania. New York: Sharon Springs, Karner, Staten Island; Brooklyn and Newtown, Long Island. 8. C. acerni Clemens. Head and body shading from light tawny to straw yellow

8. C. acerni Clemens. Head and body shading from light tawny to straw yellow and light fuscous; anal tuft wholly light red, normally paler than in *C. corni*. Fore wing with border light yellow between veins (rarely with only a few yellow scales) extending two-thirds way in to the large black discal bar; hind wing also with discal bar large. 25 mm. (acericolum Germar).

Larva boring under bark of the larger branches and trunk of maple. Moth in June and July.

Montreal, Canada, to Long Island, New York, western Pennsylvania and Illinois. New York: Buffalo, Rock City, Ithaça, Ilion, Speculator, Schenectady, Albany, Staten Island; Brooklyn and Newtown, Long Island. 9. C. fulvipes Harris. Anal tuft lanceolate; hind tibia rough-hairy the whole

9. C. fulvipes Harris. Anal tuft lanceolate; hind fibiæ rough-hairy the whole length and only slightly more so at the spurs; transitional to Sannina. Black; fore wing with terminal edge, only, scaled; a few red scales beyond discal dot. Front of palpi, all tibiæ, and most of hind metatarsi orange. An orange band on under side of abdomen. 22 mm.

Hymers, Ontario, to Massachusetts. New York: Big Indian Valley, Catskills. 10. C. saxifragæ H. Edwards. Similar to S. fulvipes; border of fore wing extending in a third way to cell; middle and hind tarsi almost wholly orange; abdomen apparently solid black. 20 mm. (henshawi H. Edwards).

Labrador; Colorado.

11. C. pictipes Grote and Robinson. Sometimes like male *exitiosa*, except as noted in the key; typically smaller, and with outer half of most of the scales in the anal tuft yellow; palpi more yellow, \mathbf{R}_3 and \mathbf{R}_4 of fore wing normally more shortly stalked. Female like male, but usually with hind tibiæ and terminal tuft almost wholly black. 15-25 mm. (H 46:24.)

General in June. Larva usually under bark of trunk and large branches of cherry, plum, June-berry and other Rosaceæ; also on chestnut (*castaneæ* Busck). Not so common on peach nor so injurious as *C. exitiosa*, but much confused with it.

New York: Buffalo, Ithaca, Catskills, Schenectady, Albany, Poughkeepsie, Hastings Center, New York City, Staten Island, Brooklyn.

12. C. rubristigma Kellicott. Palpi black, the scaling in front yellow; antennæ black, and more strongly clubbed than usual; abdomen irregularly striped, the

stripes on base and fourth segment strongest; and tuft black and yellow, more broadly spreading than usual. Legs contrastingly banded with black and yellow, the hind metatarsus variable in proportions. Fore wing hyaline, like male *exitiosa* and *pictipes*, but with the black costal edge broader, because of the wider spacing of the costal veins; discal bar rather thick, and almost wholly bright red. 15 mm.

Larva inquiline in oak galls.

New York to Illinois, New York: Mosholu, Brooklyn,

13. C. decipiens II. Edwards. Similar to C. rubristigma; border of fore wing extending in a third way to cell, yellow between the veins. 15 mm. Probably a variety of C. rubristigma or its female. (nicotianæ H. Edwards, imperfecta H. Edwards.)

Mosholu, New York; Colorado; Texas.

14. C. rhododendri Beutenmüller. Palpi black above, straw yellow below; body black, with strong transverse stripes on segments 2, 4, and 5; anal tuft, in male, large and mostly black. Legs blackish bronze, pale at spurs and joints. Fore wing with scaling almost all black (a few yellow scales): outer margin extending two-fifths way in to cell, leaving five hyaline spots; discal bar narrow, black. 9–13 mm. Our smallest Sesian.

Larva on Rhododendron.

New York; Pennsylvania. New York: Ithaca, Brooklyn.

15. C. bassiformis Walker. Purple-black; palpi yellow with some dark on upper side of terminal segments: vertex with yellow line; female antenne, as generally in the following species, with the apical half largely white, but extreme tip black. Abdomen with fine yellow stripes on segments 1 to 4 and typically on 6 and 7: the anal tuft narrowly edged with yellow; in the female, with the stripes on 4 and 6 heavy, on 3 and 5 light. Hind tibiae yellow below, often mostly black above; the tarsi mostly yellow. Fore wing with border broad, and more or less yellow-scaled; typically with five hyaline cells. 20-25 mm. (*lustrans, cupatorii*, etc., H. Edwards). (**H 46**:21.)

In variety **sexfasciata** H. Edwards (*bollii* H. Edwards) there are only four hyaline cells, and the outer border is broader. Variety **consimilis** H. Edwards has segment 6 as well as segment 5 without yellow.

August; September. Larva in stem and root of Eupatorium.

Massachusetts to Wisconsin and Texas. New York: Tuckahoe, Staten Island; Sea Cliff, Long Island.

16. C. scitula Harris. Antennæ and palpi as in *C. bassiformis;* abdomen with the yellow hand on segment 4 typically much heavier than the others, often covering the whole segment; segment 2 with heavy band, but those on 1 and 7 very weak or absent. Hind tiblæ yellow, with a black band between spurs; metatarsi contrasting, yellow, Wings as in *C. bassiformis.* 15 mm. (**H 46**: 29.)

June and July. Larva reported from under bark of Cornus, oak, hickory, chestnut, Cratagus (Wellhouse), and in oak galls; but perhaps mixed with other species.— certainly from Andricus galls on oak.

Distribution uncertain: much confused with *pyri*, *bassiformis*, and other species in collections. Glen House, New Hampshire. New York: Hudson, Long Island.

17. C. tipuliformis Linnæus. Palpi black on outer face, vellow within. Abdomen with bands on segments 2, 4 and 6, and on 7 of male, that on 6 of male weak; anal tuft black. Hind legs black, with vellow at spurs and joints only. Fore wing like that of C. bassiformis. 20 mm. (H 46: 26.)

June. Larva in pith of currant and gooseberry stems: rarely injurious. This appears to be the only northern species which belongs to Chamæsphecia.

Generally distributed. New York: Plattsburg, Buffalo, Ithaca, Kendall, Albany, Staten Island; Pinelawn, Long Island.

18. C. pyri Harris Black; palpi yellow, with outer side mostly black; neck black; abdomen with a slender yellow line above, expanding on lower part of side into a patch which reaches base of body; segment four of abdomen narrowly yel-

low above and broadly so, below. Legs black; yellow at spurs and joints. Fore wing with border reaching about a third way in to cell: more or less yellow-scaled; discal bar narrow; black. Small, expanse typically about 12 mm., but quite variable in size. (kæbeli H. Edwards). (H 46:25, enlarged.)

May; late July. Larva boring under bark of trunk of apple, pear. etc.

White Mountains, New Hampshire, to California. New York: Honeoye Falls, Buffalo (Kellicott), Ithaca, Staten Island, Brooklyn.

19. C. albicornis H. Edwards. Black. Neck narrowly yellow; palpi black, yellow in front in male, but usually wholly black in female; face as in pyri. Antennæ black in male, with a white bar in female, as in related species. 15 mm.

June and July. Larva in solid wood of willow and poplar; also in galls.

Generally distributed. New York: Catskills, Karner, New York City, Brooklyn.

20. C. ithacæ Beutenmüller. Fuscous (probably black when fresh): palpi black below, at least in female; border of fore wing extending half way in to cell in male, and farther in female, leaving four or five hyaline spots; legs with a little pale at spurs and segments; abdomen apparently wholly fuscous (type lot only seen, all of them in bad condition). 15-20 mm.

End of June to early August. Larva in Heliopsis. Pupa in the ground.

Pennsylvania. New York: Ithaca.

21. C. rileyana H. Edwards. Antennæ of male blackish, of female reddish at base; palpi yellow, the outer face black in male. Fore wing with four to six transparent cells beyond the discal bar, which is bright orange red outwardly. Inner margin of fore wing also with red scales. Hind tibiæ vellow, black between spurs; metatarsi yellow: abdomen with yellow stripes on all but first segment. Border of fore wing quite variable, linear in var. hyperici H. Edwards. 20-30 mm.

In female aberration brunneipennis H. Edwards, the border reaches more than half way to the cell, leaving only three hyaline spaces.

July and August.

District of Columbia to North Carolina, and west.

22. C. sigmoidea H. Edwards. Palpi mostly yellow; hind tibiæ with black outer side; metatarsi vellow, contrasting with tibiæ and rest of tarsus; abdomen with bands stronger on alternate segments; the terminal tuft narrowly tipped with yellow, as in exitiosa. Fore wing with discal dot red, largely mixed with black; border not reaching halfway to cell; sometimes very narrow. 20 mm.

August. Larva in stems of black willow. A seaside species. Walpole, New Hampshire, to Long Island, New York. New York: Amagansett, Long Island.

7. ÆGERIA Fabricius

(Sphecia Hübner; Trochilium auct., Hübner, in part)

Palpi as usual, hairy below toward base; male antennæ unipectinate and laminate below; female antennæ simple. Tongue weak, completely covered by palpi when coiled; hind tibiæ loose-hairy, without tufts at spurs; abdomen with a small short tuft. Fore wing normal (fig. 225); \mathbf{R}_5 running to outer margin, \mathbf{M}_2 and \mathbf{M}_{3} more curved, and less widely separated from \mathbf{Cu}_{1} than usual; hind wing with \mathbf{M}_{a} and \mathbf{Cu}_{i} shortly stalked or rarely connate; the genus distinguishable from Memythrus in the latter case, by the erect discocellulars of the fore wing, and the strongly oblique and very long ldcv of the hind wing.

1. Æ. apiformis Clerck (The hornet moth). Sides of face, front of palpi, sides of back of head, vertex, front half or more of tegnlæ, and metathorax, yellow; the rest of head and thorax dark brown. Abdomen banded with blackish and yellow; with segments two and four normally with more black than the others. Tibiæ and tarsi more orange. Wings transparent, fore wing scaled along costa to middle of cell \mathbf{R}_{i} , and with slight streaks in middle of a few lower cells; otherwise with

scales on edges and veins only. Scaling brown; discal bar red in part. 30-40 mm. (H 46:8.)

Larva boring in base of trunks and roots of poplar and willow.

Europe; various stray records in United States (Michigan, Syracuse, New York, etc.): seen only from Long Island, New York, and Nevada.

2. E. tible a Harris. Similar to E. apiformis; yellow on vertex more limited; tegulæ with front quarter only, yellow, and part of that covered by collar; with a strong yellow stripe along dorsal edge. Fore wing scaled above R_s only. Rarely, with thorax and abdomen more extensively black; with narrow yellow lines only.

June to July. Larva like that of Æ. apiformis.

Montreal, Quebec, and New York to the Pacific Coast. New York: Plattsburg, Goat Island (Kellicott).

8. ALCATHOE H. Edwards

Palpi rough scaled only; male antennæ lightly fasciculate; tongue weak. Venation normal, except for loss of one radial. Hind wing with ldcv erect. Hind tibia with a heavy tuft at end, and an equally large tuft on metatarsus. Abdomen of male with a long hairy process, as long as the abdomen, besides short side tufts.

1. A. caudata Harris. Black. Fore wing scaled, except in and below cell in male; hind wing transparent, with a small black spot on mdcv. Antennæ, palpi, and neck orange; hind legs orange, with a black tibial tuft. Long process on tail orange; side tufts black. Female with hind legs black, except outer joints of tarsus. 25 mm.

June to August. The larva bores in the crown of clematis.

Canada to Florida, west to Michigan. New York: New York City, Brooklyn and Newtown, Long Island.

In variety walkeri Neumægen, the antennæ are mostly black, the palpi are black, and also the whole of the hind legs and the process on the abdomen.

Long Island, New York.

9. BEMBECIA Hübner

Antennæ bipcctinate in male, with oblique pectinations, and no terminal tuft (fig. 231); palpi with hair below. Abdomen stout, rather cylindrical, with short terminal tuft and a well-marked dorsal tuft on segment 3. Hind tibiæ loose hairy.

Fore wing normal (fig. 229) except for loss of one M; cell square at end; hind wing with M_3 and Cu_1 stalked half way to apex; ldcv transverse and more than twice as long as mdcv. Tongue moderate.

1. B. marginata Harris. Body black; palpi, neck, some markings on thorax, and a stripe on each segment of abdomen yellow; the abdomen becoming solid yellow at rear; fore wing with a mixture of brown and rusty scales, leaving cell and anal space clear, as well as three interspaces toward margin; the boundaries of the scaled areas not quite sharp. Hind wing with brown fringe only. Male 25 mm.; female mostly over 30 mm.

August to mid-September. The larva in blackberry and raspberry, at first in roots, then working up into the canes.

Generally distributed. New York: Wilmington, Lancaster, Ithaca, Staten Island, Long Island.

In variety albicoma Hulst the yellow is very pale cream color, except on the hind legs, where it is as dark as usual. I have seen it in recognizable condition from Big Indian Valley and Brooklyn.

SUPERFAMILY TORTRICOIDEA

Moths varying from very small to very large, some exotic Cossidæ being the heaviest known moths. Head with short erect hair, usually rough, but not high and bristling as in the Tineidæ; in a few exotic genera of Tortricidæ smooth-scaled. Ocelli usually present. Antennæ as in the Yponomeutoidea; palpi characteristic (figs. 240–243, 271–283). more or less rough, projecting forward, the second joint usually roughscaled or with a rough triangular tuft, and the third fusiform, porrect, often short, and rarely as long as second. Palpi in a few forms (for instance, fig. 272) upturned more as in the Tineoid series; but then never reaching the vertex and always with the third joint short. Palpi sometimes with roughly bristly vestiture, but never with the definite bristles of the Tineidæ. Maxillary palpi rudimentary or absent; the pilifer large, and concealing them in any case. Tongue usually developed, but often absent. Thorax with vestiture varying from simple scales to deeper spatulate scales or hair; the vestiture of the legs similar. Hind tibiæ hairy, even when the legs otherwise are scaled. Spurs weak and sometimes lost in the Cossidæ. Upper spurs of hind legs well below middle. Fore wing with all veins preserved or with a single vein lost; ample; \mathbf{R}_5 running to outer margin with rare exceptions, and free or stalked with \mathbf{R}_{4} ; the other veins usually free; accessory cell distinct, with its broad side resting on discal cell; the separating vein often weak, but strong in the Cossidæ and other primitive forms; 1st A distinct, at least at margin, in the Tortricidæ and Cossidæ, but not in the Phaloniidæ and Carposinidæ; complete in the Cossidæ; and, in some western and exotic Cossidæ, connected with 2d A by a crossvein; 2d A usually forked at base. Hind wing nearly as wide as fore wing but short in the Cossidæ; Sc and R separate, or connected by R1; often differing in closely related species. **R** and **M**, often stalked; \mathbf{M}_{a} and Cu, often stalked or united, but with two veins lost only in Carposina; 1st A as in fore wing, always free; 2d A forked at base, but less markedly than in the Choreutis group.

The wings rarely if ever show the typical macro maculation with ante- and postmedial lines and orbicular and reniform spots; in the lower forms the marks are usually a system of irregular spottings and anastomosing striæ.

Egg normally of flat type, that of the Cossinæ upright. In all the families, the larva typically is boring; but many of the Tortricidæ have come to live externally, protected in some sort of shelter, usually of a rolled or folded leaf, whence the name leafrollers. The larvæ (figs. 245, 284–287) never have conspicuous markings, except for the shields and tubercles. Head always exposed and fully chitinized, with normal mouth-parts (as in all the higher groups). Mandibles tending to point more forward than those of external feeders, but not different in structure. Front extending from one-fourth to three-fourths way to vertex; adfrontals almost always touching the vertex, sometimes very large; ocelli normal. Prespiracular wart with three setæ. Abdomen with setæ i and ii well separated, and iv and v approximate usually on one tubercle; prolegs with hooks in a complete circle, which is often broken in front and back,— except in a couple of Cossidæ; often with tubercles ii of ninth segment of abdomen approximated or united in mid-dorsal line, and often with a specialized multiple supra-anal spine. Many species are injurious as borers or external feeders,— probably a larger proportion than of any other superfamily.

Pupa of a normal incomplete type, with segments 3 to 6, and 7 of male, movable, with two rows of spines on each segment, as in the higher Tineoidea and Ægeriidæ. Head often with a cocoon-breaker; prothorax convex and wider than dorsal headpiece; maxillæ distinct, not divergent as in the Hepialidæ, but sometimes almost as small, and completely separated by the labial palpi, which are always exposed. Mentum well developed. Maxillary palpi distinct, separated by a suture; but in Zeuzera, at least, remaining with maxillæ on dehiscence.

Antennæ varying, following their character in the adult; rarely if ever as long as wings. Fore femora exposed. Cremaster of various types.

The four families are quite clean-cut; in fact, are united mostly by the combination of specialized micro larva with typical incomplete pupa. The Eucosminæ (Olethreutinæ) are often made a separate family, but have no elean-cut characters to distinguish them from the Tortrieinæ, though certainly a well-marked group in the imago. Several workers separate the Zeuzerinæ also as a family Zeuzeridæ. In this case intermediate forms are rarer if they exist, and the separation is very possibly justified; it has not generally been made by American workers.

Family. 28. TORTRICIDÆ

(With Eucosmida, Grapholithida, Epiblemida, Olethreutida)

Ocelli present (so far as looked for); antennæ rarely peetinate (never in our speeies); the sealing eonfined to dorsum of antenna, but the outer row stronger and longer than the inner; ventral surface pubeseent, more strongly so in male; palpus moderate, upturned to middle of front, or rough and porrect, often triangular; tongue present, usually rather

weak. Body slender; hind tibia hairy, the others smooth-scaled. Wings broad, the fore wing often abruptly widened at the base, giving the family its name of "bell moths," from the shape of the moths with their wings folded. Fore wing (fig. 252) with \mathbf{R}_5 rarely running to costa; base of **M** simple (representing \mathbf{M}_3) crossing the cell obliquely, or, more rarely, absent; accessory cell often ill defined; \mathbf{Cu}_2 arising two-thirds way out on cell, or less; distant from \mathbf{Cu}_1 ; **1st A** free and weak at base, absent in a few reduced forms. Hind wing ample, with fringe markedly widened at anal angle; scaling soft.

Egg flat. Larva (figs. 244–246, 284–287) with hooks of prolegs multiordinal, except in a few reduced forms; ninth segment of abdomen with tubercles ii usually united, always approximate; iv and v on abdomen obliquely or vertically placed; vii of seventh segment, of two or more setæ. Pupa with hooked spines either on last segment. or on the specialized cremaster, tongue well developed; maxillary palpi separating from tongue on dehiseence; antennæ reaching nearly to tip of wings.

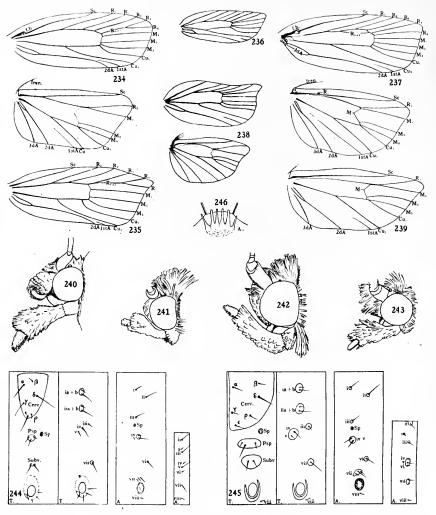
This is one of the large world-wide families, and very nearly represents one of Linnæus' subgenera of Phalæna (Tortrix). Several of the genera recognized here are based on secondary male characters only and have been rejected by Walsingham and Durrant, and others. Our series of forms is so unwieldly and other clean-cut characters are so scarce that I have retained them here with some modifications. The venational characters vary a little, and in some cases a series will be necessary to make sure that a specimen is not a venational freak. The costal fold in the male, when it occurs, is often very tightly closed, especially in Eucosma and Epiblema, and is easily overlooked; when it becomes rudimentary, as in some Archips, it is usually open and more easily seen.

Key to genera; imago

- 1. No fringe on base of Cu of hind wing (often with loose hair below Cu, or with a fringe on base of 2d A).
 - 2. Fore wing with R₄ and R₅ stalked half way to apex; R₅ running to outer margin (fig. 235).
 - 2. \mathbf{R}_{*} and \mathbf{R}_{5} very shortly stalked or free.

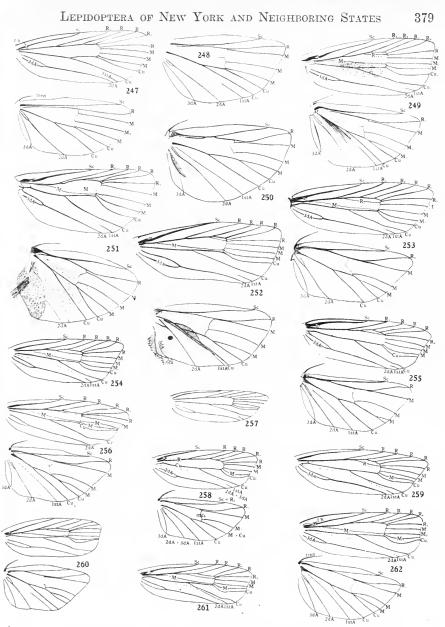
 - 3. M₃ and Cu₁ separate, rarely stalked.

 - 4. Fore wings with **R**₅ running to outer margin, or to the bluntly rounded apex (fig. 237).
 - 5. Palpi ascending (fig. 242); hind wing with R and M, approximate at base.



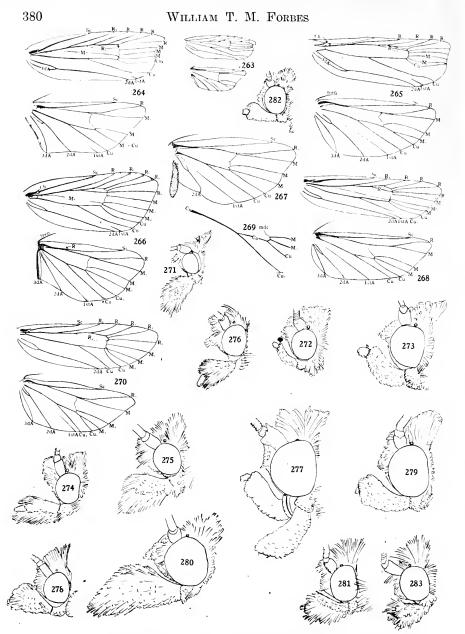
FIGS. 234-246. TORTRICIDÆ

234, Sparganothis sulfureana, venation; 235, Calostathma discopunctanum, venation of fore wing; 236, Peronea species (Europe), venation of fore wing; 237, Tortrix pallorana S, venation; 238, Archips podana Q (Europe), venation; 239, Cnephasia virescana, venation of hind wing; 240, Sparganothis (Cenopis) diluticostana, ventro-lateral view of head of male with palpi drawn down, showing the frontal vestiture characteristic of the subgenus; 241, Peronea variana, side view of head; 242, Archips rosaceana, head; 243, Tortria dohrniana (Europe), head; 244, Peronea species, seta map of larva; 245, Archips magnoliana, seta map of larva; 246, Archips magnoliana, dung fork of larva



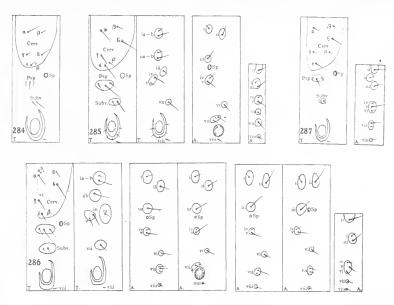


247, Hemimene bittana, venation; 248, Laspeyresia interstinctana, venation of hind wing; 249, Carpocapsa pomonella \mathcal{S} , venation and sex marks; 250, Ecdytolopha insiticiana \mathcal{S} , venation and sex pouch; 251, Gymnandrosoma punctidiscanam \mathcal{S} , venation and sex marks; Melissopus latiferreanus, \mathcal{S} , venation and sex marks; 253, Charlotta ratzeburgiana, venation; 254, Epinotia nanana, \mathcal{Q} , venation of fore wing; 255, Thiodia parmatana, venation; 256, Excutera spoliana, venation; 257, Thiodia striatana, venation of fore wing; 258, T. raracana, venation; 259, T. formosana, venation; 260, Gretchina deludana, venation; 261, Exentera maracara, venation; 262, Episimus argutanus, venation.



FIGS. 263-283. TORTRICIDÆ

263, Rhopobota ilicifoliana, venation; 264, Anchylopera nubcculana J. venation and anal fold; 265, Epiblema scudderiana J, venation and costal fold; 266, Olethreutes fuscalbana J, venation and anal fold; 267, Cymolomia species J, venation and lobe; 268, Bactra lanceolana, venation; 269, Polychrosis species, cubital stem and branches of hind wing; 270, Rhyacionia bushnelli, venation; 271, Hemimene dana, side view of head; 272, Laspeyresia caryana, head; 273, Carpocapsa pomonella, head; 274, Gypsonoma fasciolana, head; 275, Thiodia aspidiscana, head; 276, Charlotta ratzeburgiana, head; 277, Eedytlolopha insiticiana, head (the left palpus drawn down); 278, Epiblema occllana (Europe), head; 279, E. scudderiana, head, 280, Encosma dorsisignatana, head; 281, Olethreutes bipartitana, head; 282, Polychrosis viteana, head; 283, Rhyacionia comstockiana, head.



FIGS. 284-287. TORTRICID.E

284, Laspeyresia interstinctana, seta map of prothorax of larva; 285, Carpocapsa pomonella, seta map; 286, Tmetocera ocellana, seta map; 287, Anchylopera nubeculana, seta map.

6. Thorax smooth-scaled; M_a and Cu_i connate or shortly stalked.

44. Archips.

- 5. Palpi porrect, rough above as well as below (fig. 243).
 6. Hind wing with R and M₁ stalked (fig. 239); thorax normally smooth 40. Cnephasia, 42. Eulia.
 - - Fore wing smooth-scaled.
 Male antennæ with notch near base......45. Pandemis.
 - 8. Male antennæ not notched.
 - Thorax with posterior crest......42. Eulia (Argyrotænia).
 Thorax without crest.

2. Laspeyresia (lautana).

- 1. Cu of hind wing with a fringe or tuft of hair on upper side at base, besides the scattered hair below $\rm Cu.^{20}$
 - 2. Fore wing with \mathbf{R}_4 and \mathbf{R}_5 stalked.
 - 3. Apex of fore wing subfalcate with a notch just below it, above M_1 (fig. 263)ll. Rhopobota.
- ²⁶ For females of this group, see supplementary key, page 384.

3. Apex of fore wing not falcate. 4. M_{2} , M_{3} , and Cu_{1} closely approximate at margin of fore wing (like fig. 260). 5. Fore wing smooth-scaled......23. Exentera (and a few Epinotia). 4. M₂, M₃, and Cu₁ evenly spaced at margin. 34. Sparganothis, 35. Amorbia 🌻 2. Fore wing with \mathbf{R}_4 and \mathbf{R}_5 completely united; 11 veins only. 3. M_2 , M_3 , and Cu_1 approximate at margin of fore wing. 4. M. and Cu, stalked. 5. Costal fold present......18. Sonia. 5. Costal fold absent.....12. Kundrya. 2. Fore wing with \mathbf{R}_4 and \mathbf{R}_5 separate or connate. 3. Hind wing with R and M_1 remote at origin (fig. 247), more than half as far apart as M_2 and M_3 when the latter are also remote. 4. R separate from Sc in hind wing; eyes a little reduced, with a scaled band behind them (fig. 271).....l. Hemimene. 4. R of hind wing becoming coincident with Sc. $1\frac{1}{2}$. Pammene. 3. Hind wing with \mathbf{R} and \mathbf{M}_1 closely approximate or stalked. 4. Fore wing falcate, with a distinct point between \mathbf{R}_4 and \mathbf{R}_5 , broadly concave below.²⁷ 5. Hind wing with M_s and Cu_1 stalked, rarely separate.....8. Ancylis. 5. M₃ and Cu₁ united (fig. 264).....9. Anchylopera. 4. Fore wing not really falcate. 5. Hind wing with M_2 , M_3 , and Cu_1 all remote at origin from cell (fig. on superficial view, all connate; thorax smooth (fig. 268). 32. Bactra. 5. Hind wing with M_3 and Cu_1 connate; M_2 slightly separate (fig. 262). 28. Episimus. 5. Hind wing with M_2 and Cu_1 connate, or stalked; M_2 remote; male without costal fold. 6. Thorax with a posterior tuft. 7. Male hind tibiæ with metallic scales above, near base. 5. Gymnandrosoma. 7. Male hind tibiæ with normal loose hair. 8. Hind wing with a thickened area above base of 3d A only (fig. 250)6. Ecdytolopha. 8. Male with a fold on inner margin, normally containing a hair pencil, or a pencil arising from base of hind tibia, or, 6. Thorax smooth. 8. Male hind tibia and tarsus with coarse, divergent hair. 4. Melissopus. 8. Male hind tibia with loose hair; tarsus scaled. 9. Male with base of hair pencil on Cu covered with large shining scales; a groove along 2dA, containing the usual 9. Male without sex scaling toward inner margin of hind 2. Laspeyresia. wing. 27 In a few members of other genera the fore wing is more or less falcate, but the outer margin is hardly concave, or the apex is rounded over.

- 5. Hind wing with M₂ and Cu₁ counate, M₂ strongly curved and approximate to $M_3 + Cu_1$ at base; thorax crested; inner margin of hind wing sexually modified; male without costal fold; hind tibia almost always with a hair pencil at base.
 - 6. Inner margin of hind wing with a thickened free lobe.

31. Cymolomia.

6. Inner margin of hind wing with a more or less distinct fold and hair pencil.

7. Hind metatarsus of male with a tuft above....30. Phæcasiophora.

5. Hind wing with M_3 and Cu_1 stalked or united; M_2 strongly curved and approximate to their base; thorax rarely crested; male often

6. Fore wing with M_2 separate at origin from M_3 and curved.

7. Male antenna with a notch near base.

8. Fore wing with a large raised scale-tuft in fold.

17. Strepsicrates.

8. Fore wing smooth.

9. Costal fold present......Griselda (p. ---).

7. Male antenna without a notch.

8. Black sex-scaling toward costa of hind wing above.

21. Proteoteras.

8. No black sex-scaling on costa of hind wing. 9. Male with costal fold.

10. Sc absent $14\frac{1}{2}$. Hendecaneura. 10. Sc present...7. Epinotia, 13. Epiblema and 14. Eucosma.²⁹

- 9. No costal fold.
 - 10. Outer margin of fore wing evenly excurved; the veins evenly spaced.

11. Uncus slender, sometimes bifid at tip.

7. Epinotia, in part.

10. Outer margin concave toward costa; M3 somewhat upcurved toward margin; apex often with an ocellate black dot, making the apex appear more falcate than it is; \mathbf{R}_2 normally arising from cell before commencement of accessory cell. 11. Hind wing with \mathbf{R} and \mathbf{M}_1 stalked.....24. Gypsonoma.

11. Hind wing with **R** and M_1 approximate.....10. Norma.

- 10. Outer margin more or less distinctly notched near middle; M₃ strongly curved up to margin.
 - 11. Fore wing with more or less distinct raised scale-11. Fore wing smooth-scaled.

7. Epinotia, 15. Thiodia, 23. Exentera.³⁰

²⁸ In occasional specimens M_2 and M_3 are perceptibly separate; they can generally be distinguished by the palpi, which have the vestiture of the second joint abruptly cut off at the end; the third joint well defined and projecting free (fig. 283). ²⁹ For a key to these species see Eurosma. ³⁰ These genera are distinguished only by the mule genitalia. For a key to the species that may run here see Thirdia

that may run here, see Thiodia.

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10.	Outer	marg	in ev	enly.	incui	rved,	but ve	ins not	approxi-
	111	iate;	apex	ma	rked;	male	with	eighth	segment
	111	odifie	d. –					0	-

11. Fore wing with R₂ arising from accessory cell, approximate of R₃; valves divided......27. Hystrichophora.
11. Fore wing with R₂ arising from before accessory cell, halfway between R₁ and R₃; valve simple.
26. Pseudogalleria.

Partial key to females of Eucosmina

 R and M₁ of hind wing widely separate (fig. 247)
3. \mathbf{R}_4 and \mathbf{R}_5 long-stalked
 4. Hind wing with M₃ and Cu₁ stalked
 M₂, M₂, and Cu₁ of hind wing widely spaced at origin33. Polychrosis. M₂, M₂, and Cu₁ very shortly but equally spaced at origin, under low magnification appearing connate
 5. Fore wing with ground striate transversely; with speculum. 6. Fore wing more than twice as long as wide6. Ecdytolopha. 6. Fore wing less than twice as long as wide5. Gymnandrosoma. 5. Fore wing not striate transversely; without speculum; usually with "Exartema pattern" or transverse fascia.
4. Thorax smooth-scaled
tinctly stalked; wings with metallic bands. 2. Laspeyresia (fig. 248), 3. Carpocapsa (fig. 249), 4. Melissopus
(fig. 252). 3. M_3 and Cu_1 stalked or united; M_2 separated but curved and approximate to them at base.
 Fore wing with 11 veins; R_i and R_s united. 5. Hind wing with M_s and Cu_i united
 Fore wing with raised scale-ridges or tufts
5. Fore wing with a large tuft in outer part of fold17. Strepsicrates. 5. Fore wing with small tufts.
 6. Some green scaling
 6. R₂ arising from discal cell; accessory cell short. 7. Fore wing with a contrasting marginal lumulate fascia. 26. Pseudogalleria. 7. Fore wing with a dot at the slightly produced apex only.
8. Hind wing with R and M, stalked 14½. Hendecaneura, 24. Gypsonoma.
8. Hind wing with R and M_1 approximate10. Norma.

6. \mathbf{R}_2 arising from the large accessory cell.

7. Apex and outer margin evenly rounded (figs. 253, 254).

8. M_2 of fore wing connate with M_3 and nearly straight.

20. Rhyacionia.

- 8. M₂ and M₃ of fore wing approximate and curved.
- 7. Epinotia, 14. Eucosma, 23. Exentera, 25. Zeiraphera. 7. Apex marked; outer margin evenly concave; the veins not con-
- and veins \dot{M}_1 to M_2 approximating toward the concavity (figs. 255, 256).
 - 7. Epinotia, 13. Epiblema, 14. Eucosma, 15. Thiodia, 23. Exentera.

Key to genera: larva (after Fracker)

- 1. Ninth segment of abdomen with seta i close to iii and usually on the same tubercle; not associated with ii.
 - 2. Seta vi absent on ninth segment; tubercles black; crotchets uniordinal.

40. Cnephasia.

- Seta vi present on ninth segment, usually associated with iv and v.
 Adfrontal sclerites reaching vertex, and front reaching about two-thirds way.
 - Arms of epicranial suture concave above, meeting in an attenuate point.
 Abdomen with setæ iv and v on first segment in a diagonal line, horizontal on seventh.
 - 5. Abdomen with setæ iv and v on first segment nearly vertical, oblique on seventh.
 - Seta vi of ninth segment of abdomen on the same tubercle with iv and v16. Spilonota (fig. 286), 31. Cymolomia.
 Seta vi normally on a separate tubercle.
 - 3. Carpocapsa (fig. 285), 4. Melissopus. 4. Arms of epicrania straight above.

11. Rhopobota, 13. Epiblema, 21. Proteoteras. 3. Adfrontals extending two-thirds, and front one-half, way to vertex.

26. Pseudogalleria.

- 1. Ninth segment of abdomen with seta i as far from iii as from ii; iv-vi on one tubercle.

 - 2. Wart vii with three setse on seventh, and two on eighth, segment. 3. Middle sets of prespiracular wart dorsal of the other two.

34. Sparganothis.

- 3. Middle seta of prespiracular wart in line with, or lower than, the other two.
 - 4. At least three times as far from posterior as from anterior seta.
 - 5. Meso- and metathorax with vii single.
 - 6. Second ocellus much farther from first than from third.
 - 34. Sparganothis (group Platynota), 44. Archips in part (figs. 245, 246).
 - 6. Second ocellus as near first as third.
 - 9. Anchylopera (fig. 287), 28. Episimus, 29. Olethreutes, 34. Sparganothis in part, 43. Tortrix.

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- 4. Middle seta of prespiracular wart not more than twice as far from posterior as from anterior.
 - 5. Fourth ocellus much closer to third than to sixth, and behind the
 - 5. Fourth ocellus about halfway between third and sixth and in line

Key to genera: pupa (after Mosher)

1. With seta on anal rise; without a distinct cremaster.

2. Two long distinct setae on each side of anal rise.

- 3. One row of long strong flat set inserted along the row of spines at last segment.
 - 4. Dorsum of abdominal segments covered, more or less, with short triangular spines; the spines of the anterior row alternately of two
 - 4. With the two regular rows of spines only, the anterior all of one length. 5. Exposed part of fore coxæ more than half the length of the mid-
 - 5. Exposed part of fore coxæ shorter; body always slender.

16. Spilonota.

- 3. A second row of setæ at caudal end of body, the first row consisting of four setæ.

 - Second row also of four setæ; maxillary palpi touching maxillæ.
 Second row with middle setæ much slenderer than lateral ones; maxillæ less than a third the length of the wings; labial palpi about two-thirds the length of the maxilla; maxillary palpi touching front legs1. Hemimene.
 - 5. Setæ of caudal row all similar; maxillæ at least one-halt the length of the wings; palpi about one-half the length of the maxillae, maxil-
 - 4. Second row of two setae; maxillary palpi reaching maxilla.

7. Epinotia, 24. Gypsonoma.

- 2. Never with two long distinct set at each side of anal rise. 3. Lateral spines of last segment noticeably enlarged; seta at caudal end very weak and obscure; usually with two pairs of obscure setae on
 - 3. Lateral spines of last segment not noticeably larger than the others; setæ at caudal end long and heavily chitinized, on a distinct papilla on each side of anal rise.....2. Laspeyresia.
- 1. With a well-developed cremaster.
 - 2. Ninth segment of abdomen with a distinct row of spines, strongest in male; last segment sometimes with spines; cremaster broader than long; always with setæ on anal rise.
 - 3. Cremaster not curved ventrad; the corners not produced into prominent hooks, but usually ending in three short lobes; second segment of abdomen with anterior row of spines present, and posterior row strong; sette on anal rise lateral to anal opening.
 - 4. Last segment with spines, usually three or four crowded rows.
 - 5. With strong setar on anal rise.

6. Two sets on each side, much like those of cremaster...28. Episimus. 6. One seta on each side, smaller than those on cremaster.

29. Olethreutes in part.

5. No setæ on anal rise; spiracles at the bottom of deep pits.

33. Polychrosis.

- 4. Last segment without spines.
 - 5. Well-developed setæ on each side of anal rise.
- Cremaster curved ventral, the posterior angles ending in prominent hooks: second segment of aladomen with anterior row of spines absent, and posterior weak; setæ of caudal rise attached farther back.

 - 4. Maxillary palpi absent externally; anterior spines of segment nine rot on distinct ridges, the spines not extending as low as spiracles; setæ on ventral side of cremaster short and heavily chitinized. 39. Peronea.
- Ninth segment of abdomen with a few scattered spines, in male; no setæ on anal rise; cremaster nearly always longer than broad; last segment never with spines.
 - 3. Segmental membranes showing prominent dark brown spines scattered over a light brown surface.
 - 4. Cremaster much longer than broad, not flattened.
 - 5. With four setæ at end of cremaster.
 - 6. Anterior spines of segment two of abdomen weak in male and absent in female; head without an anterior projection.

41. Harmologa.

- - 5. Cremastral setæ flattened; anterior spines of second segment of abdomen wanting in female.....group Platynota.

Subfamily EUCOSMINÆ

(Grapholithing, Epibleming, Olethreuting)

This subfamily appears to represent a fairly marked group, though defined by no single character at any stage. The genera are in part ill-defined, or distinguished mainly by genitalic characters revealed only by dissection. Heinrich's recent study (United States National Museum, Bulletin 123) has been followed, but externally visible characters have been used as far as possible. In some cases it has been necessary to treat two genera as a unit for the sake of the specific key.

Valves strongly chitinized: with narrow basal articulation, articulating with the juxta; inner face of valve chitinized except at base; anellus consisting of a triangular plate with an external central arm supporting the ædæagus. Larva with setæ i and iii of ninth segment of abdomen normally on a single plate.

1. HEMIMENE Hübner

(Dichrorampha Guenée; with Lipoptycha, in part)

Antennæ simple; palpi moderate (fig. 271), porrect, long-hairy, with the third joint almost as long as the second but mostly concealed in its hair; head nearly smooth; eyes small, with a naked area behind them (fig. 271). Thorax smooth: Fore wing rather broad for a Eucosmid (fig. 247); the ofter margin not notice-ably oblique; more or less concave above the middle; tending to form a slight notch at M_1 ; veins all present and distinctly separate; the base of M_2 and R_{i+5} usually distinct. Cu running through middle of wing or rather above; R_3 arising at least $2\frac{1}{2}$ times as far from R_2 as from R_4 . Hind wing proportionate; with R and M_1 well separated, more than a third as far as M_1 from M_2 ; and more than half as far as M_1 from M_2 ; M_3 and Cu_1 connate or short-stalked.

This is the genus as defined by Busck. Meyrick and others have excluded the species without the costal fold, referring them to Lipoptycha. The larvæ are usually borers in stems and roots of Compositæ. This genus and Laspeyresia dispute the position of lowest of our North American Tortricidæ.

Key to the species

1. Fore wings with cream-white dorsal spot	
1. Fore wings with a yellow dorsal spot	
1. Fore wings with a vague yellow dorsal shade, or none; rarely, with short.	
well-separated, yellow striæ.	
2. Apex purplish, contrasting with the tawny basel. plummeriana.	
2. Apex not purplish and contrasting.	
3. Fore wing with undulating black transverse lines2. leopardana.	
3. Fore wing without such lines.	
4. Light golden yellow4. bittana.	
4 Dark fuscous without vellow 3. daya	

* No costal fold (Lipoptycha auct.).

1. H. plummeriana Busck. Palpi clay-color, with a good many black hairs mixed with some pale ones. Fore wing with base to middle of costa and three-fourths way out on inner margin dull clay-color, strigose, dusted with black; the outer half contrasting yellow-brown, with confused fasciae and marbling of lead-gray. and some black scales, which tend to gather as a lead-and-black fascia along the inner margin of the dark part. Some partly confluent black terminal dots, followed by a fine yellow line, below the notch, and a single black dot above it: with clay-colored costal striae on outer part of wing. Terminal dots and costal striae repeated on under side. Hind wing light gray. 10 mm.

April to June. Larva in flowers of pawpaw.

District of Columbia; Cincinnati, Ohio; Urbana. Illinois.

2. H. leopardana Busck. Bright orange-ochre, contrastingly marked with a mixture of lead-gray and brown-black streaks and partly confluent spots, tending to became oblique streaks along the costa. Terminal dots distinct and separate; below the notch. Fringe gray, cut with yellow at notch; hind wing dark. 8 mm.

May to August. Larva on Eupatorium.

Maryland to Cincinnati, Ohio.

3. H. dana Kearfott. Fuscous, with the usual yellow patch on side of palpi, but no other yellow. Fore wing with slight leaden lustre, tending to become

brighter on outer half; outwardly with many of the scales luteous-tipped, giving a frosted appearance; leaving several lead-gray bands of the ground, which are strongly oblique toward costa, parallel to outer margin below, and confused in the middle; with a few black spots and streaks in the yellow-dusted portion; with a series, usually of four strong terminal dots, below the notch, and sometimes one small one above it. Hind wing solid dark. 12 mm. (Enarmonia Kearfott.)

This species has been determined as Laspeyresia nigricana, but may be distinguished by the generic characters and by the terminal spots. Very close to *H. plumbana* Scopoli of Europe, and *H. sedatana* Busck, of Colorado, of which it is likely to prove the eastern race.

Late May and June.

New Hampshire to western Pennsylvania; common and generally distributed. New York: Black Brook, Batavia, Rock City (Cattaraugus County), Crosby, lthaca, McLean.

** With costal fold (Hemimene).

4. H. bittana Busck. Fold large, extending a third of the length of the fore wing. Fore wing golden or orange-ochre, more or less suffused with brown; the base almost always paler or darker brown; the outer half, especially toward inner margin, very rarely wholly brown except for a portion of the postmedial region. The markings tend to be black striations on the veins in the middle of the wing and curved transverse striæ at the middle of the inner margin; base almost evenly colored; a lead-colored postmedial band well out toward margin, sharply bent opposite notch and parallel to margin below, with a short lead-gray bar beyond it on costa; normally with fine broken black striæ between them. Black terminal dots conspicuous, at least below notch. 12–15 mm. (*alpinana* Fernald in part, not Treitschke).

June and July.

Rather common and general in distribution. New York: Ithaca.

5. H. simulana Clemens. Second joint or palpus with yellow body and brown hair. Fore wing gray-brown, with a large, half-crescent-shaped, bright yellow patch on middle of dorsal margin; outer han with grayer shining bands, separated by areas of yellow-tipped scales, and a few subterminal black dots as well

as the terminal ones. 10 mm. (alpinana auct., not Treitschke). End of May; end of August. Parry Sound, Ontario, to southern New Jersey, Pennsylvania, and District of Columbia. New York: Ithaca, West Farms.

6. H. incanana Clemens. Blackish, varied with dull white; with an oblique white dorsal patch, formed of two pairs of fused strike ending on the ochreous speculum, which has three black bars and lies at the middle of the subterminal region.

I have seen only the type. (Halonota Clemens).

Pennsylvania?

11/2. PAMMENE Hübner

Similar to Laspeyresia. Male with R of hind wing arising widely separate from M₁, running directly across into Sc, as in some Pyralids. Female as in Laspeyresia. 1. P. felicitana Heinrich (Kearfott, ms.). Purple-black, varied in median area with shining purple-gray, and sometimes with some whitish scaling toward base; the two outer lead-colored fasciæ showing a distinct purple shade, and the speculum narrow, with a few black spots rather than bars. Outer part of wing sometimes shaded with brown; sometimes with a brown terminal line; and usually with the ground in the speculum lighter, sometimes luteous. Inner margin to beginning of speculum broadly and irregularly cream-white, sometimes with a couple of spots of the ground color; the white area extending nearly to the costa at base. Hind wing mouse-gray. 13 mm.

June; August and September.

Western Pennsylvania; Montreal and St. Hilaire, Quebec.

2. LASPEYRESIA Hübner

(Grapholitha Rebel. etc.; Enarmonia Fernald, etc.; Ephippiphora, of the earlier writers)

Palpus with second joint merely rough scaled below (fig. 272), the vestiture not concealing the third joint, which is usually about a third as long as the second, and scaled. Thorax and fore wing like Hemimene; hind wing with **R** and \mathbf{M}_1 more approximate at base (fig. 248), usually somewhat sinuous, and diverging abruptly about a third way to margin. \mathbf{M}_2 strongly curved but well' separated from \mathbf{M}_4 which is connate or stalked with \mathbf{Cu}_1 . No sexual modifications, except for some dense scaling on the hind wing of L. pyricolana.

The species are numerons and close and the following key may not be always successful.

Enarmonia dana Kearfott is a Hemimene; L. lautana is strongly aberrant, and represents a new subgenus Sereda Heinrich; L. shawiana is a Hendecaneura. Aside from the slight difference in venation and palpi, the genus may be distinguished from Hemimene by the absence of terminal dots just back of the outer margin, below the notch; usually there are no terminal dots at all, but in L. lautana they are present exactly on the margin of the wing membrane, and are strongest on the apical part of the wing.

Key to the species

- 1. Two terminal dots at apex noticeably larger than the others.....l. lautana. 1. A single larger terminal dot at apex, or none.
 - 2. Hind wing half or more pure white.

3. Fore wing blackish.

4. Fore wing with a white crescent at middle of inner margin.

21. albimaculana.

4. Fore wing with only a faint gray crescent at middle of inner margin.

10. eclipsana.

2

3. Fore wing gray......18. multilineana.

- 2. Hind wing fuscous to blackish: sometimes with more or less of base dirty white.
 - 3. Ground of fore wing dull white.
 - 4. Outer half contrasting dark, largely blue......20. fletcherana.
 - 4. Base of inner margin and outer third spotted with blackish.
 - 22. gallæsaliciana.
 - 3. Ground of fore wing blackish.
 - 4. Two strongly irregular silver-white striæ clear across the wing. . 16. rana.
 - 4. A contrasting white crescent or group of striæ on middle of inner margin, reaching up about to middle of wing, but entirely separate from the costal striæ.
 - 5. Dorsal white extended in along inner margin to base.
 - (Pammene felicitana.) 5. Dorsal spot a group of four er more fine, more or less fused striæ, costa strongly striate with white......15. tristrigana.

Lepidoptera of New York and Neighboring States

5. Dorsal spot composed of two or three more or less fused striæ; or single and reaching well above fold, but not extended toward base; if of three striæ, costa not striate with white. 6. Speculum represented by a single lead-gray band; ground wholly 6. Speculum yellow-brown, regularly barred with black, the ground 4. No contrasting white mark at middle of inner margin. 5. Even blackish with white spotting; tending to form broken trans-..... albolineana. verse bands 5. Not evenly blackish without speculum. 6. Blackish. 7. Speculum absent; represented by a single lead-gray bar. 8. saundersana. 7. Speculum present; normally of two lead-gray bars, with a yellower area between them containing black markings. 8. With contrasting white costal striæ from a quarter way out on 8. No white costal striæ before middle of wing. 9. Ground brown, with scattered. Inteous-tipped scales, at least on outer part of wing; denser outwardly; the speculum often nearly solid luteous. 10. Speculum higher; its uppermost bar above or hardly below 10. Speculum of four black bars, distinctly farther from costa than the notch; hind wing solid blackish, except on covered part of costa. 11. With well-marked costal white striæ.....14. candana. 11. No distinct white costal striæ......13. dandana. 9. Fore wing with white-tipped scales.....17. molesta. 9. Ground solid black-brown, or nearly so. 10. Two median lead-gray fasciæ, nearly in contact, and so bread as tegether to be more than a third as wide 10. Median fasciae narrower and obscure or absent. 11. Hind wing very pale gray at base and beneath; black 11. Hind wing not paler at base; black dots distinct. 12. First line of speculum parallel to outer margin; 12. First metallic line of speculum erect; fringe of hind wing white except at base.....12. nigricana. Omitted from key: 4. nigromaculana. * Outer margin strongly oblique, hardly notched at \mathbf{M}_1 : \mathbf{R}_5 running to apex or very slightly below; hind wing with fringe on Cu very weak and almost always lost; head rough, with strong hair on palpi; eyes smallish, but round (Sereda Heinrich).

1. L. lautana Clemens. Powdery light gray, the outer half noticeably strigose with white, which tends to gather in a patch at middle of inner margin. Antemedial line strongly bent on cell, and oblique above; the base up to it more solidly dark; outer and costal parts of wing with lead-colored stripes, the next to last usually running to anal angle; all starting from paired white costal striæ. Five or six terminal dots on upper part of wing, the two at apex decidedly larger and a little separated from the others. 12 mm.

April and May.

This species is side-specialized in the direction of the Tortricine and Eucosma. New York and Pennsylvania to Manitoba. New York: McLean, Ithaea, Ramapo.

** Outer margin of fore wing nearly upright, with well-marked notch; \mathbf{R}_3 terminating well below apex; hind wing with strong fringe on \mathbf{Cu} ; head and palpi generally smoother, the eyes normally large (Laspeyresia).

† Male without sexual modifications.

2. L. caryana Fitch. Palpi whitish. Deep brown, with lead-colored stripes, not strongly contrasting, and distinct only on costa, except the last two, which enclose the speculum. Ground dusted in two or three areas with eream color; the dusting in the speculum dense; speculum with four or five black bars of unequal size, the first strong one pointing directly to the notch in the outer margin. White costal strike distinct outwardly, but not prominent. Hind wing fuscous, whitish on basal half; fringe whitish, with fuscous basal line. 10–12 mm.

May, July and August. Larva on hickory and walnut; the second brood eating out the young nuts; also inquiline in galls.

Canada to Missouri and Georgia. New York: Easton (type).

3. L. prunivora Walsingham (lesser codling-worm). Face and palpi whitish. Black-brown varied with ochre yellow; the outer margin below costa almost solid yellow in some specimens, except for some black bars and the two lead-colored bands; middle of wing more or less striate with black; middle of inner margin with confused oblique streaks. Costal edge outwardly with some white dots. Speculum of the yellow, with black bars, decidedly lower than the notch, as usual in the genus. Hind wing like L. caryana. 9 mm.

In dark specimens the yellow is reduced to some few scales on the disc of the fore wing, in the upper part of the speculum, and just before the apex.

Larva in young plums, Cratægus fruit, and crab apples.

Generally distributed. New York: Crosby (Yates County), Ithaca.

4. L. nigromaculana Kearfott. Palpi more than half hiteous, with dark tips. Fore wing cream color on basal half, the ground more than half covered with wavy brown transverse bands, which leave a vague pale area in the middle of the inner margin; outer half mixed bright ochre and black-brown, not contrasting to the naked eye; with some black dots and striæ, and with strong black terminal spots. 9 mm.

June.

Black Mountains, North Carolina.

Superficially very close to Laspeyresia prunivora, with which Kearfott has confused it. Types only seen.

5. L. packardii Zeller. Dull blackish; head and palpi gray (unlike *L. caryana* and *prunivora*); fore wing with lead-gray fasciae leaving a distinctly defined but not contrasting brownish erect median fascia, and a similar subtriangular patch at anal angle. Speculum with a couple of faint dots only. Hind wing very pale gray, with dark border and veins.

This species can apparently be distinguished by the partly pale hind wings and gray palpi. I have no notes on authentic material. L. pyricolana has been determined as this, in error.

Texas; doubtful northward.

6. L. interstinctana Clemens. Deep brown, about eight white costal striæ, on the outer three-fourths of costa; two larger curved striæ at middle of inner margin, reaching about up to cell; a lead-colored bar before anal angle. Hind wing solid black-brown. 9 mm.

Common in May and June; and again in late July and August. Larva (the clover seed caterpillar) in flower heads of clover, damaging the seed.

Generally distributed. New York: Newport, Ithaca, Big Indian Valley, New York City.

7. L. angleseana Kearfott. Basal half chocolate brown; outer half brown with strong golden iridescence. Costa with nine white striæ, the first two, fourth, seventh, and ninth forming the end of short lead-colored bars; the third weaker. Three lead-gray bars on inner half of wing, the area between the first two strigose with black; the speculum lying between the outer two, barred with black. Outer margin blackish. Hind wing solid dark. 10 mm. End of May to June; August. Locally common. Anglesea, New Jersey; Massachusetts.

8. L. saundersana Kearfott. Black-brown. Costa with about 10 white striæ, more oblique than in L anglescana; several of them ending in lead-colored bars. A vague pale crescent beyond middle of inner margin, extending half way to costa, formed of four pale striæ which end above in a lead-colored spot. A single lead-gray bar in position of speculum. 10 mm.

Toronto, Ontario. New York: Karner (Forbes). 9. L. fana Kearfott. Black-brown; costal marks about like L. angleseana. Middle of inner margin with a white spot, formed of two thick and partly fused white striæ. Speculum chocolate brown with four black bars, the lead-gray bar beyond it much shorter than the one before, being cut off below by the edge of the wing. 7½-10 mm.

May to August. Larva amber colored; a bud-worm on Meibomia (Kearfott). New Jersey to Ohio.

10. L. eclipsana Zeller. Deep brown; costal striæ as in L. angleseana, but all distinct, strongly oblique, the lead-gray stripes silvery and conspicuous, and the striæ which do not end in lead-gray streaks, running into shorter luteous ones. Speculum with the lead-colored bars rather close together, the black showing as spots rather than bars; or, speculum filled largely with black; with faint traces of a pale half-crescent on middle of inner margin, ending in a lead-colored spot at middle of wing. Hind wing white; the apical third blackish. 12 mm.

April to May. Larva on grape.

New Jersev and Ohio to Texas. New York: Ithaca.

11. L. youngana Kearfott. Fuscous, broadly banded with lead-color except toward base. The two median bands complete but irregular, and very broad, the outer ones narrower and more or less broken; all starting from slight pale costal spots. Speculum hardly paler, with four black bars, and with a lead-gray spot below it, as well as the bars before and beyond; a distinct narrow yellow terminal line before the black line in the fringe, but no other yellow. Line in fringe heavily cut with white at notch. 8-11 mm. (perstructana Walker?).

Larva in center of cones of *Picea alba*, hibernating as a larva and emerging in the spring.

Ottawa, Ontario.

12. L. nigricana Stephens. Black-brown, with some pale striation on outer part of costa; the two lead-colored stripes narrow and broken, the upright part outlining the speculum perpendicular to the inner margin, and separate from the costal part, which runs obliquely across the apex. Speculum with black bars, the series quite obscurely continued to costa, forming an oblique series between the two lead-colored stripes. Hind wing blackish brown, hardly paler at base, the fringe contrasting, white, with a gray line in the base. 12 mm. (novimundi Heinrich).

Almost identical in appearance with *Hemimene dana*, but differing in the palpi, the venation, and the submarginal, instead of marginal, position of the series of black dots.

June to August. Larva in pods and seeds of peas and other Leguminosæ; sometimes injurious.

New Jersey and western Pennsylvania to Manitoba; also reported as injurious in Nova Scotia.

13. L. dandana Kearlott. Dull dark brown, with some slight pale scaling on costa, and scales ontwardly showing pale tips in some lights. Two oblique bluish iridescent stripes running down and out from costa toward apex, with a fine black line half way between them; two purpler and duller bands before anal angle enclosing about four black spots, to form speculum. Hind wing concolorous. 10 mm.

End of August to September.

Essex County, New Jersey.

14. L. candana Kearfott. Much like L. dandana but larger; the metallic bands narrower and more broken; the black line between them obscure or lost; pale scaling more distinct, but more nearly confined to speculum. Hind wing with contrasting white fringe; without dark basal line. Possibly the spring brood of L. dandana.

May.

Western Pennsylvania; Virginia.

15. L. tristrigana Clemens. Dark brown, sometimes varied with black; speculum represented by a lead-colored bar only; about 10 or 12 closely crowded but cleancut white costal striæ, leaving only base of costa free; a group of four to six partially fused striæ on middle of inner margin, each one curved out in a quarter circue; their tips followed by some luteous scaling. Hind wing concolorous; outer part of fringe contrasting whitish. 12-15 mm.

Larva on "Tinctoria" (perhaps Baptisia tinctoria).

Massachusetts to Florida, Kansas, and Oregon.

16. L. rana Kearfott. Deep brown; a strong double medial, strongly waved but not distinctly broken, silvery band; and a single postmedial band, each of these starting from a pair of white costal dots. Another single white subterminal dot on costa, and a pair before apex. Speculum preceded by the postmedial band, followed by a vertical dark-silver bar, and crossed by four black bars; the ground concolorous. Hind wing dark. 14 mm.

May.

Black Mountains, North Carolina.

17. L. molesta Busck. Dark fuscous, with some white-tipped scales; head dark, including palpi. Fore wing with traces of the dorsal crescents showing as a gathering of the white-tipped scales. Speculum strongly white-dusted, with from three to six blackish dots. Hind wings blackish, with fringe whitish except at apex. 10-15 mm.

Two or three broods, breeding more or less continuously. Larva injurious to stone-fruits, especially peach; boring out or gnawing the surface of growing twigs. and boring in the fruit. Cocoon in any sort of shelter, that of the last brood often under bark.

Vicinity of Washington, District of Columbia; probably introduced from Japan. 18. L. multilineana Kearfott, with the fore wing powdery gray, and the hind wing white, has been taken at Jamestown, New York, in June.

19. L. garacana Kearfott. Light gray; the ground being made up of luteous mottling and dusting on a dark gray base, leaving the dark as a series of parallel but unequal and broken streaks. Base slightly darker; a distinct narrow dark median fascia running from middle of costa to before middle of inner margin, bent at a right angle on cell, the lower part more oblique than the outer margin. A

similar straight subterminal bar on inner half of wings only. Speculum obscure, high in the wing; black line in base of fringe clean-cut, and cut with white at the notch. 15 mm.

July. Types only seen, male and female.

Trenton, Ontario; Chicago, Illinois.

20. L. fletcherana Kearfott. Basal half of wing and body dirty white; outer half a mixture of blackish and steel-blue, with some wood-brown and a little whitish scaling: the terminal line normally brown. Line in base of fringe continuous, black, interrupted at notch with white. Hind wing fuscous, paler at base. 13 mm.

June.

Ottawa, Canada. A paler variety occurs in Washington.

21. L. albimaculana Fernald. Fuscous gray; costal edge black, irregularly barred with white, the white bars fading into the grayish ground below. Inner margin heavily shaded with black, especially beyond the white dorsal patch, which is moderate in size, rarely divided into two striæ, and extends obliquely out and up to middle of wing. Outer part of wing with some yellowish tint; the lead-colored stripes broken and irregular, with irregular black spots between them representing the speculum. 8-12 num. (articulatana Kearfott).

April 30.

Maine to Manitoba.

22. L. gallæsaliciana Riley. White. Head and thorax white; abdomen dark gray. Fore wing irregularly spotted with gray, shading into yellow brown or golden at the apex; the gray gathering to form a patch on outer margin, a third the length of the wing on the inner margin, but often hardly reaching the costa; and an antemedial patch toward inner margin. Speculum with heavy lead-colored bands, with some heavy black dots before and within it. Hind wing light gray, paler, and striate with dark gray beneath. 12 mm.

June and July. Larva in a slender twig-gall on willow, perhaps inquiline.

This species has been generally confused with a western Epinotia which has a costal fold in the male. The hind wing has \mathbf{R} and \mathbf{M}_1 , and \mathbf{M}_3 and \mathbf{Cu}_1 short-stalked and \mathbf{M}_2 more curved than usual in Laspeyresia.

Mt. Washington, New Hampshire, to Missouri. New York: Peru, Big Indian Valley.

^{††} Male with a patch of rough scaling on upper side of hind wing and one on under side of fore wing.

23. L. pyricolana Murtfeldt. Dark fuscous; head and palpi dark; fore wing with several transverse lead-gray fasciæ, the median ones nearly complete. Speculum with two lead-gray bars, parallel to outer margin, and also connected with oblique fasciæ running down from the costa. A distinct series of black dots or short bars in the speculum, and between the speculum and the costa, extending most of the width of the wing. Basal line in fringe unbroken. Hind wing in male light gray, no darker at the margin, with a blackish sex-patch covering most of cell and extending a little beyond it; in female blackish. Fringe gray. 10 mm. Male usually smaller. (*Penthina cyanana Murtfeldt in part, by type lot; packardi of collections; Steganoptycha, Epinotia.*)

May and June; August; _____. Three broods. Larva green.(?) the spring brood boring into the buds of rose and blasting them; the later broods normally feeding on the leaves. Pupa under the edge of a leaf folded over, or boring into pith. Also with similar habits on apple and peach. The larva is a little doubtful on account of confusion with O. cyanava, but the dates and food plants are certain.

WILLIAM T. M. FORBES

3. CARPOCAPSA Treitschke

(Cydia; Laspeyresia, in part)

Autenna smooth; palpi almost smoothly scaled (fig. 273), obliquely upturned, with short third segment, as in the smoothest species of Laspeyresia. Thorax smooth. Venation as in Laspeyresia, the notch at M hardly visible. Hind wing of male (fig. 249) with the fringe on Cu very strong, dark, and deflected downward by a mass of shining scales in the lower part of the cell, which turn down across the base of the fringe. A broad groove along 2d A, containing the peneil arising from the basal fork of the vein; anal region with slightly modified general scaling. Larva without anal fork.

The genus is of moderate size; the larva feeding in fruits and seeds. It is well represented in the West and in Europe, but has only a single introduced species in the Northeast, the famous codling worm. The sexual characters are different in almost every species and the genus is hardly distinct from Laspeyresia.

I have placed *C. toreuta* and *erotella* here temporarily, following Heinrich. They are not true Carpocapsas, but so far as I know they have no valid generic name. The whole group they represent are feeders on conifers. *Laspcyresia youngana*, also, should probably be grouped with them.

1. C. pomonella Linnæus (the codling-worm). Gray, with fine striation, showing under a lens as white tips to blackish scales; base, or a broad antemedial band, slightly darker, with excurved and scalloped outer boundary; speculum of two bronze-brown bars, the outer one more or less broken up. filled in with chocolate brown, the brown extending to form a large oval area almost reaching costa and outer margin; costa and extreme outer margin striate and gray like the base. No black in speculum, but speculum preceded by a heavy vertical black bar, ending in a point halfway to costa. Fringe with a black line, and sometimes cut with white at \mathbf{M}_1 . Hind wing brown; the enlarged scales covering \mathbf{Cu} in the male lead-color; the hairs of the fringe on \mathbf{Cu} blackish. 15-20 mm.

The codling moth is generally distributed and injurious wherever apples are grown; occasionally it feeds in the fruits of a few other Rosaceæ, especially pears. Northward there is but a single brood, the moths emerging in the spring (soon after the blossoms fall) and laving their eggs on the outside of the voung apples. The larvæ enter by the calvx-cup and feed on the inside of the fruit, including the growing seeds, growing slowly. They leave the fruit late in summer and form their cocoons under the bark or, more rarely, in trash on the ground, hibernating as larvæ and pupating in the early spring. Southward there is a second brood, the moths emerging in July and August, and laying their eggs on the well-grown apples. The larvæ of this brood often enter the side of the apple and feed less systematically, leaving the apple late in the fall and hibernating like the singlebrooded forms. They are sometimes known as "side-worms."

2. C. toreuta Grote. Dark gray; basal fourth, medial, and postmedial fasciæ lead-gray; also two subapical dots and a marginal line half as wide as the fasciæ.

Larva in cones of yellow pine. Falls Church, Virginia: North Carolina: and west. The moth is perhaps widespread in the East, but is rarely obtained save by breeding.

3. C. erotella Heinrich. Lead-color. Two shining fasciæ before and beyond middle of wing, each starting from a double pale stria at costa. excurved, and farther out on inner margin; a short bar on costa at three-fourths, also starting from a double stria; and an irregular and broken subterminal line, preceded by one or two black spots below middle of wing. Fringe with black basal line, cut with whitish below apex: outer part of fringe shining lead-gray. Hind wing concolorous. 9–10 mm.

March (perhaps forced): August. Larva in pitch nodule or gall on *Pinus* tada, the type bred in association with *Rhyacionia comstockiana*.

Maryland; Mississippi.

4. MELISSOPUS Riley

Metallic scaling more extensive than in *Carpocapsa pomonella*. Hind tibia of male with very long hair below and above toward its tip, and metatarsus with heavy hair above; hind wing (fig. 252) roughly and thickly scaled on inner margin and at base of Cu and 2d A, but without hair pencils. Otherwise like Carpocapsa and Laspeyresia.

1. M. latiferreanus Walsingham. Light red-brown, with a heavy erect median lead-colored fascia; a slightly narrower one three-fourths way out; and a slender and broken submarginal one, strongly curved below costa. Between the two outer fasciæ there are some broken black bars and streaks. Fringe lead-colored, with a reddish line in the base; hind wing mouse gray. 15 mm.

Two broods in Missouri, the second perhaps partial; July and August; late September. Larva on young acorns; cocoon formed between a leaf and a nearly circular piece cut out of another.

Our eastern form, as described above, is var. orichalceana Walsingham. Typically the second fascia is rudimentary (California).

Generally distributed north to Montreal. New York: Ithaca, Karner.

5. GYMNANDROSOMA Dyar

Similar to Ecdytolopha; palpi with third segment porrect, slightly more beaklike. Wings (fig. 251) more than half as wide as long, and rough-scaled; thorax tufted behind. M_2 of hind wing more curved than in Ecdytolopha. Abdomen of male with a naked patch above, flanked by a couple of pale hair-pencils; hind tibia with a fanlike tuft of hair, with metallic scales at its base. Hind wing with heavy tufts of hair and enlarged spatulate scales on inner margin; the fringe

on Cu extremely heavy, but not modified. 1. G. punctidiscanum Dyar. Irregularly mottled and strigose with fuscous, blackish, and some lead gray, the outer part often largely dirty white and con-trasting. Discal dot small, white, sharply defined in a black shade. No definite line in fringe. Hind wing mouse gray. 18-25 mm. The moth has been taken from May' to July.

New Jersey and northern Illinois to South Carolina. New York: Ithaca, New York City; Maspeth, Long Island.

6. ECDYTOLOPHA Zeller

Palpi rough-scaled; upturned to middle of front; with short third joint (fig. 277). Thorax tufted behind; wing rough with transverse ridges of slightly enlarged raised scales, two and one-half times as long as wide; with rounded outer margin and no distinct notch; all veins separate and normal. Hind wing (fig. 250) with R and M1 approximate, M2 widely separate, and parallel to M3, nearly straight; M3 and Cu, connate; the male with a thickened patch above 3d A at base, but not otherwise modified. Tibiæ with the usual hair only.

This genus seems intermediate between Laspeyresia and Olethreutes, and together

with Gymnandrosoma, has been sunk to the latter by Walsingham and Durrant. 1. E. insiticiana Zeller. Basal half fuscous, mottled with dull black; outer half dirty white, striate and mottled with fuscous and some black; the boundary somewhat oblique and nearer base on inner margin; sharply defined below, diffuse above; the outer markings usually forming a small triangular largely black anal spot. Hind wing mouse-gray. 25 mm. (H 48:29.) May to August. The larva may be found in August, in twigs of locust, making a fusiform gall which is sometimes very slightly swollen when in stout twigs.

.The passage to the exterior is kept open and the frass thrown outside. The larva

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leaves the gall to pupate, and forms a cocoon between two fallen leaves. It cuts a kidney-shaped piece nearly out of each leaf and sews them together at the edges, but leaves the cocoon suspended by a few uncut veins.

Generally distributed from Northern Vermont sonthward; replaced by a closely related species in Mexico. New York: Peru, Lewiston, Baldwinsville, Rochester Junction, Ithaca, Karner, Rhinebeck, and New Windsor.

2. E. islandana Kearfott. Pale dull brown, evenly and coarsely strigose with fuscons gray and blackish; the inner edge black toward the base, and usually with the somewhat darker base ending in a distinct black stria at the middle of the inner margin. Hind wing gray, paler at base. 15 nm. (Olethereutes Kearfott.)

May. Types only seen; in poor condition.

Plummer's Island. Maryland.

The generic position of this form is not yet certain, as it lacks the secondary sexual characters of Eedytolepha. It is certainly no Olethreutes,

7. EPINOTLA Hübner

(With Zeiraphera: Enarmonia: Steganoptycha: Thiodia: Eucosma, in part)

Fore wing with outer margin evenly excurved; more or less concave or notched, apex blunt; cestal fold sometimes present. All veins present, M_1 to Cu_1 convergent at margin in forms with a notch; R_1 from before middle of cell; R_2 slightly nearer R_3 than R_1 , arising from accessory cell. R_5 rarely short-stalked (fig. 254).

Valve simple; cucullus and anal indentation variable; the latter not densely spined, and usually narrow; sacculus with spines. Uncus strong and simple, or bifid, or reduced and simple; socii large, chitinized when stender; typically triangular and hairy.

This genus would appear to centain the most primitive of this series of Eucosmina. It is heterogeneous, but all the characters seem to intergrade. The species are not keyed separately, but are included in the keys to the genera Thiodia, Charlotta, and Eucosma, which they resemble in venation.

I. Outer margin evenly rounded out; the reins almost evenly spaced at outer margin; cell Cu_1 less than twice as wide as the others. Costal fold absent.

1. E. nanana Treitschke. Dark smcky brown; the head and thorax somewhat lighter fuscous. Fore wing with dcuble whitish striae, with paler fuscous areas between them; the striae sometimes so broad as almost to obliterate the pale areas, and sometimes reduced to a narrow edging of single scales. Base perceptibly darker, bounded by an excurved line. An oblique median fascia, of the ground color, from middle of costa to inner margin, before anal angle; much less oblique than in Z. pinicolana. Outer margin also solidly of the ground color. A continuous black line in fringe, sometimes cut with white at M₂. Hind wing nearly concolorcus. 9 nm. (domonana Kearfott; piceafoliana Kearfott).

June. Larva webbing terminal needles of spruce.

Maine to Ohio. New York: Portageville.

2. E. cruciana Linnæus. B'ack-brown, more or less heavily overlaid with claycolor or dull light ochre (in the Quebec specimen before me, mostly ochre). Two parallel oblique light fascia, one from middle of ecsta to outer third of inner margin, the other from cuter fourth of ecsta to anal angle, both edged with heavy but broken silver lines. Two silver dets at apex. Base of fringe mixed black, brown, and white: cuter part gray, white below apex. Hind wing mouse gray. 15 mm. (augustana Hübner).

July 6, 1916.

Megantic, Quebec; Europe.

The American form described above appears to differ a little from the European, and is probably a valid race.

3. E. septemberana Kearfott. Tawny brown with pink iridescence. A dull light gray area dusted with dark gray on inner margin from antemedial line to outer margin, over a third the width of the wing, into which projects a lobe of darker brown, covering the end of the cell. Fringe of the darker brown, with a pale basal line in the middle of wing; solidly pale at anal angle. The pale gray some-times extending narrowly to the base. 18 mm.

September. The larva of a closely related western species feeds on crab-apple.

Essex County, New Jersey; Scranton, Pennsylvania. 4. E. lindana Fernald. Costal half of wing black-brown; dorsal half pale powdery gray, as in E. septemberana; the boundary clean-cut from the base nearly to the apex, forming two large rounded projections of the dark area with a pale one between them. 20 mm.

September. Larva on Cornus.

New Brighton, Pennsylvania. New York: Ithaca.

II. Outer margin concave or notched at middle, the veins converging to the concavity. No costal fold (Catastega).

5. E. signatana Clemens. \mathbf{R}_{4} and \mathbf{R}_{5} separate. White dusted with gray; in effect, very powdery light gray; head light, with outer side of palpi gray; thorax strongly mottled with grav. Fore legs blackish with white bars on tarsal segments and two bars on tibiæ. Fore wing with markings formed by gatherings of the dark dusting. Antemedial line obscure above, extended and strongly angled at lower angle of cell, and a little concave and irregular or straight below, preceded by a strong blackish shade, and ground nearly white beyond. Numerous oblique costal white and gray striæ from base to apex; the extreme base and apex gray. (The striæ extend a third the width of the wing toward the base, but outwardly are no longer, and hardly enter the cell.) Some more nearly solid gray shades below the tips of these striæ, extending from the tip of the blackish antemedial shade to the outer margin below the apex. Speculum obscure, preceded by a couple of black spots; often suffused into a blackish triangle on inner margin. Fringe cut with white below apex; with two gray lines, distinct toward apex. Hind wing ash-gray. 15 mm. (Hedya Clemens, Catastega accriella Clemens.)

Sometimes this moth occurs in large numbers on the bark of trees in June. The larva forms a frass-tube in which it lives between two principal veins on the under side of a leaf of maple, and a silken web extending between the two veins. more or less erumpling the leaf. It eats only the parenchyma in the area covered by the web.

The name accrienta has priority but was based on the larva alone. This species and the next would form the genus Catastega Clemens. distinguished primarily by the peculiar larval habits.

Quebec to Pennsylvania. New York: Rock City (Cattaraugus County), McLean, New Windsor.

6. E. timidel'a Clemens. \mathbf{R}_{i} and \mathbf{R}_{i} separate. For legs banded as in T. signatana. Thorax powdery gray. Fore wing blackish; a large whitish area along inner margin, extending from basal angle to speculum, sharply defined above: quite narrow at base, to a third way out: where the boundary extends obliquely up to the middle of wing, then longitudinal in general direction, but sagging down in the midle, to the upper angle of the speculum, where it becomes ill-defined: the line defined above in the median area with black and below with very slightly raised silver-white scales. Costa outwardly striate as usual, the last stria almost eutting off a dark apical dot from the rest. Speculum whitish. dusted with light gray; with two broad, vague, silver bars. the outer one small; the whole preceded by two or three more or less confluent black spots. usually heavier than in .T. signatana. A clear blackish line in fringe, cut here and there with white. 18 mm. (Catastega Clemens; Gelechia.)

May. Larva on oak in September; with the habits of T. signatana. Pupa in the ground.

Minnesota; Manitoba. New York: Crosby (Yates County), Ithaca (Cornell, bred), Bellport, Long Island (Dyar).

111. Outer margin evenly rounded or stright; cell Cu, less than twice as wide as the others. Costal fold present in male (Epinotia).

7. E. solandriana Linnæus. Ground varying from pale gray, sometimes coarsely striate with whitish, to fuscous or wood-brown. Antemedial right-angled at middle of wing; strongly oblique to costa and to inner margin; normally defined by a blackish bar before it on dorsal half (obscure in many specimens but always traceable); postmedial fascia perpendicular to this, from middle of costa to before anal angle; its dorsal half often filled with blackish and the rest obscure (it is regularly twice as wide at middle as above and below, but only part of the enlargment will be dark-filled); often a dark shade in apex. Space on inner margin between the two bands often filled with white, with a few dark points on the inner margin; or the two bands and the space between them may be covered by a black patch; or more rarely with a contrasting white ray from base almost to outer margin. Outer half of costa, below, ochreous with four or five squarish fuscous patches. 20 mm.

Caterpillar a bud-worm on various trees and shrubs, especially Amentiferæ.

Ottawa, Ontario to British Columbia; Europe.

8. E. similana Hübner. Dull brown, mottled with blackish; with some black dots toward apex. A rounded white patch at middle of inner margin; speculum white with a yellowish shade in middle. Fringe with a brown line, slightly cut with white. 20 mm.

September. Larva in hazel and birch. Our specimens are distinctly yellower than European ones and are possibly distinct.

New Jersey and north; Europe. New York: Pearl River.

9. E. medioviridana Kearfott. Light dull gray, shaded with apple green; the median area almost solid green, especially toward inner margin; a blackish excurved basal line; antermedial line nearly erect, nearer base at costa, contrasting, black; wing outwardly with broken and confused fuscous strix. Speculum obscure. Line in fringe faint, gray. 16 mm.

August and September.

Ottawa, Canada; western Pennsylvania.

10. E. madderana Kearfott. Ground pale mottled gray, shaded with old rose; nearly covered by three large rounded patches; the basal one slightly darker and yellower brown, from a third way out in costa to middle of inner margin, with evenly excurved outer margin; a large transverse oval patch over end of cell, reaching costa; and a contrasting bright yellow-brown patch resting on apex and outer margin, reaching almost to hind angle, with its inner boundary even, defined with white, and bent at a right angle below costa. 15 mm.

June and July.

Ottawa, Ontario; Illinois; Manitoba.

11. E. laracana Kearfott. Light gray, somewhat strigose; base darker, bounded by an angulate antemedial line which is practically obsolete above the middle, and concave on the lower half, where it is defined with blackish. A blackish shade in base of fold. A blackish oblique fascia from middle of costa to middle of disc, followed by a spot in the subterminal region. Costa blackish, with paired white striæ, outer margin gray with a dark spot at anal angle, but the rest of the inner margin strigose on a white base. Line in fringe black, cut with white opposite the cell, and weak below. Hind wing light gray. 15 mm.

April.

Cincinnati, Ohio. Types only seen. Probably the same as E. celtisana.

12. E. vertumnana Zeller. Wings unusually narrow; fore wing pale gray with a slight bluish tint, lightly flecked with fuscous gray; rarely blackish. A slight blackish antemedial streak on dorsal half of wing, fading out above; perpendicular to inner margin; rarely heavier and practically reaching the costa. A similar heavier streak at two-thirds way to apex; oblique and a little excurved from just beyond middle of costa to inner margin at three-fourths way to anal angle, much thickened in the fold, and usually not quite reaching either margin; rarely thickened into a more even fascia, or sending a branch up from its outer side at Cu_z . Speculum with a few fuscous scales; line in base of fringe obscure, gray, and broken. 15 mm. (celtisana Riley, xandana Kearfott). March. The type with thickened bands is much smaller (9 mm.) than the

others, and may be distinct.

Cincinnati, Ohio; Missouri; Texas. 13. E. yandana Kearfott. Wing form as in E. zandana, of which it may be a darker variety. Ground crisply dusted and shaded with chalk white; blackish flecking conspicuous, clean-cut, and in the outer part of the wing forming sparse, but strong, strix, especially toward the costa. Antemedial band often a thick bar toward inner margin; postmedial band much thickened on outer side at fold, and usually sending off a spur toward the apex. 15 mm.

March.

New Brighton, Pennsylvania.

IV. Outer margin more or less concave, with veins converging to the concavity. Costal fold present in male.

14. E. zandana Kearfott. Closely similar to E. yandana; cell Cu_1 twice as wide as the others, but notch in outer margin obsolete; slightly smaller; oblique fascia rarely distinct; ground rather duller, dark smoky gray, dusted somewhat irregu-larly with black. Line in base of fringe broad, blackish, not contrasting. Hind wing dark mouse gray. 13 mm.

March. In this form the concavity of the outer margin is very slight, and it may be only a variety of E. yandana; in the remainder of the genus the concavity is marked.

Western Pennsylvania to Ohio.

15. E. nisella Clerck. Crisply powdered with brownish gray on bluish white; sometimes nearly evenly, showing only light and dark striation to the naked eye; usually with the base more or less contrasting, dark, its outer boundary sharply bent out at middle. Inner margin often brown in median area; speculum with a group of black dots or bars at middle of wing, and preceded by some black bars nearer inner margin; both sets obscure in the more evenly powdery specimens, and contrasting in luteous areas in the more contrasty ones. Lines in fringe obscure. 16 mm.

Larva in early spring on catkins of poplar, alder, birch, etc., falling to the ground with them, after which it is said to become a general feeder on the trash on the ground.

July and August. Extremely variable. Variety pavonana Donovan is the one with the brown patch on inner margin; in variety decorana Hübner the whole median area is yellow-brown, leaving the gray only on the base, apex, and speculum.

Generally distributed in Canada; western Pennsylvania. Europe.

16. E. walkerana Kearfott. Basal two-fifths blackish; with slightly excurved outer boundary, nearer the base toward the costa; median fifth white, toward costa filled somewhat strigosely with gray; outer two-fifths lighter brown with some short thick pale costal striæ; speculum a vague silver-gray area with some black dashes in its upper part, preceded by a black spot or two. A continuous dark line in fringe, cut with white below apex, brown rather than black. (hamptonana Kearfott, in part). 10 mm.

Late July to September. Larva in catkins of hazel.

Pennsylvania. New York: Ithaca.

17. E. hamptonana Kearfott. Ground wood-brown, without any white patch; a slightly darker patch before the speculum in place of the black dots; costal stria duller, and not continued by distinct gray lines; gray lines of speculum narrower than the enclosed space, which is heavily barred with black; line in fringe obscure. and not crossed by a white stria.

Hampton, New Hampshire (types only known).

18. E. momonana Kearfott. Similar to E. transmissana, but smaller; white dorsal patch shaped almost as in E. otiosana, but not defined with black. Ground gray, varied outwardly with dull yellow-brown. Dark base with oblique outer boundary on lower half of wing. Speculum with two broad lead-gray bars, both regular and subequal in width (unlike E. transmissana and otiosana); filling of speculum ochreous, with four or five black bars. Costal and marginal markings and fringe as in E. transmissana. 13 mm.

July.

Ottawa, Ontario. I have seen a type.

19. E. transmissana Walker. Base dark brown or blackish, striate, with excurved outer boundary, median area white or with a white patch; clouded and striate with fuscous toward costa; followed by a rather triangular brown area resting on the inner margin before the speculum. Juter part of costa narrowly brown, with geminate white striæ, defined with black; outer margin narrowly brown above, with a line below apex, running out through the fringe. Speculum confused, centering high up near the middle of the wing; mainly lead gray and white, with black bars; a black line in base of fringe. 15 mm.

July. Larva perhaps on birch.
Northeastern States; Nova Scotia to Illinois. New York: Saranac Inn, Summit of Mt. Marcy, Rock City (Cattaraugus County), Albany.
20. E. solicitana Walker. Dull brown, striate with violet-gray; with a slight golden lustre at apex. Base mostly brown, with slight gray striation, its outer strike with violet gray striation, its outer strike with violet gray striation. boundary markedly angulate at the middle; medial area broadly gray to middle of costa, and to beyond the second third of the inner margin, followed by an irregular oblique brown fascia. Costa outwardly with paired whitish striæ. Speculum of two more or less double silver-gray striæ, the ground between them brown with several black dots. Line in base of fringe black, cut with white. 12-15 mm.

May and June. Larva on white birch.

Seen from New Hampshire and Pennsylvania; type from Nova Scotia. "New York" (Fernald).

21. E. rectiplicana Walsingham. Head, thorax, and basal half of fore wing pure white; the fore wing with some gray flecking, especially along inner margin, and a large blackish splotch in fold. Outer half of wing mingled yellow-brown and blue-gray; the blue-gray shining in the speculum, where it takes the form of two broad, vertical bars. Filling of speculum yellow-brown, with black dots. Outer part of wing also with some white mottlings, especially along costa. Fringe fuscous with black basal line, white at apex. 13 mm. (gallæsaliciana Kearfott in part).

June in New York; November in California. Larva on willow.

Rocky Mountain and Pacific States. New York: Peru (Forbes).

A very distinct form which will prove to have a wide range in the North. The costal fold is slender and very long.

22. E. signiferana Heinrich (eastern form). Head and shoulders dark brown; thorax lighter, fore wing ash gray; the base somewhat darker, with oblique and excurved outer boundary; median fascia oblique out from middle of costa to

before anal angle; rich dark brown above, fading out below; its middle connected to apex by another less regular brown shade. A longitudinal black bar in outer part of cell and beyond, crossing the two brown fasciae. 14-18 mm, September 1.

New York; western States. New York: Hemlock Lake. 23. E. heucherana Heinrich. Black-brown: fore wing with some white streaking on the basal patch, a moderately broad, angulate antemedial fascia, and some white streaks on outer part of costa. Fringe with a black basal line; smoky fuscons, cut with white below apex. Hind wing very dark brown, with paler fringe. Head with white face, and base of palpi. 10-13 mm. This may be a race of the western *E. ruidosana* Heinrich, from the same food,

but it shows a genitalic difference.

Larva in a digitate mine in leaves of *Heuchera americana*; deep red, with black head and cervical shield. Moth in May and June.

Rosslyn, Virginia (near Washington).

8. ANCYLIS Hübner

(Ancylopera, Phoxopteris)

Eyes moderate; palpi porrect, somewhat rough-scaled, or more rarely quite hairy, often clavate, large. Wings smooth-scaled. Fore wing falcate, the outer margin usually concave from above M_1 to Cu_1 ; the concavity variable in character, but sharp and deep, as in Eudemis. All veins free; M_2 - Cu_1 more or less approximate at margin. Hind wing with M_3 and Cu_1 connate or stalked.

Valve slender; cucullus sharply defined, slender; anal indentation usually broad and smooth; sacculus only sparsely spined; costal hook weak or absent. Uncus

variable, bifid when present; socii very broad and hairy; gnathos slight, fused. I use the names Ancylis and Anchylopera as indicated by Walsingham and Durrant. In this country they have sometimes been interchanged, or, more often, the genera have been united.

Key to the species of Ancylis and Anchylopera

- At least apical part of costa brown or gray.
 Fore wing with a white triangle on outer part of costa, enclosed by a heavy angulate blackish band......8. torontana.
 - 2. White outer costal patch not edged on lower side with blackish, or absent."
 - 3. Fore wing evenly brown to the speculum, which is contrastingly pale. 1. tineana.
 - 3. Fore wing blue-gray and brown, with only costal striæ whitish.

3. carbonana. 3. Fore wing dull fuscous with only costal striæ whitish....4. unguicella.

- Basal or medial part of wing with contrasting markings.
 A longitudinal dark shade from base to apex; the wing light gray below it.
 - 5. Shade sharply defined on lower side, forming two shallow scallops.

2. diminutana. 5. Shade diffuse on both sides, broken at middle of wing...5. goodelliana. 4. No shade from base to apex; base of inner margin dark.

5. Base of wing dark from costa to inner margin, with a well-marked, transverse, excurved or angulate outer boundary; ground of outer part of wing yellow or yellow-brown.

³¹ A few Thiodias are likely to be sought here. They are distinguished by the white streak on the base of Cu.

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 6. Speculum absent; outer part of wing broadly yellow, with yellow fringe
cchreous9. cornifoliana. 5. Base of costa at most moderately paler than inner margin; the boundary diffuse; dark base below cell with strongly oblique outer boundary edged with a fine white line.
6. Yellow-brown 13. fragaria: 6. Chocolate brown 14. floridana 6. Umber brown 15. comptana 5. Base of inner margin ochre-yellow, contrasting with the whitish base
of costa, but not sharply set off from it
5. With a contrasting brown or blackish patch on basal half, more or less, of inner margin, with sharply defined boundary on upper and usually on outer side; the costal region pale and contrasting with it.
 6. Basal patch followed by dark gray markings extending it almost to anal angle; median fascia on costa weakl. nubeculana.³ 6. Basal patch ending abruptly about middle of outer margin. 7. Fore wing without a marked oblique fascia from middle of costa, rarely with a pale clay-colored one; outer part of wing without any bright yellow or brown; basal patch gray-brown lobed at its upper outer angle, extending far into cell. 2. subæquana.[*]
 Fore wing with a conspicuous oblique median fascia from costa or outer part of fore wing strongly shaded with ochre, or with a reddish basal patch. 8. Basal and outer markings of fore wing concolorous and black
ish fuscous. 9. Median fascia broadening below into a fuscous blotch cover ing whole region of speculum, followed by a trapezoida white patch on costa
 9. Region of speculum not suffused with a fuscous, the media costal fascia followed by a pair of striæ. 10. Basal patch and fascia chocolate brown, with clay color between them; small (10 mm.)6 angulifasciana.³ 10. Markings light fuscous on a whitish ground, only slightly tinted with clay-color; larger (15 mm.) 13³
 Median fascia largely ochreous or paler, when the basal patch patch is blackish, noticeably paler than it is. Basal patch blackish, contrasting with outer markings. <i>9. burgessiana</i>,³² 10. murtfeldtiana.³
 Basal patch ferruginous brown or paler. Specular region of fore wing typically with a small dark dot, sometimes even light gray; median fascia with its lower boundary sharply defined and rounded over, no containing distinct black dashes5. pulchellana.[*]
³² These species belong to Anchylopera.

- 10. Speculum without a dark dot, but frequently with more complex markings; median fascia normally diffuse below, and containing black dashes beyond cell.
 - 11. Costa luteous toward base, concolorous with ground of outer part of wing: basal patch edged with black above, its outer boundary meeting inner margin in a long slant......4. maritima.³²
 - 11. Costa whitish toward base, much paler than outer part of wing; outer boundary of patch meeting inner margin at right angles, the patch not outlined with black.
 - 12. Basal patch on its basal half without defined boundary, fading into the cream-white costal area.

7. platanana.³²

12. Basal patch defined, except sometimes at extreme base. 13. Fringe of fore wing below apex cream white, faintly suffused with ferruginous ochreous.

11. laciniana.²²

13. Fringe of fore wing below apex dominantly ferruginous ochreous12 fuscociliana.**

Omitted from key: Ancylis loricana; Anchylopera spiræifoliana, metamelana, discoferana, dubiana, lamiana.

I. Hind wing with \mathbf{M}_{0} and \mathbf{Cu}_{1} connate or barely stalked; for wing with apex strongly drawn out; the outer margin with a deep concavity, centering on M.

1. A. tineana Hübner. Brown with some silvery striation toward apex on costa; speculum white, shaded with light gray; terminal line dark brown; fringe below apex whitish. Hind wing dirty white. 15 mm. April to July. Two broods. Larva dull gray-green with yellow-brown head; on poplar and various Rosaceæ, in June and late fall.

Labrador to Massachusetts and Manitoba; also in Europe. 2. A. diminutana Haworth. Dull brown, shading into whitish on costa, espe-cially toward base. Inner margin and outer margin helow notch dull pale powdery gray; the upper boundary defined, and running in two shallow waves from base almost to outer margin, where two lead-colored lines converge to its tip from

costa. Apex wholly brown. 15 mm. April to July. Larva on willow. The American records for A. uncana are probably in error for this species. Uncana is a little larger and has M_3 and Cu, stalked.

Montreal, Quebec. and Vermont. to North Carolina and British Columbia; Europe. New York: Peru, Black Brook (Clinton County), Crosby (Yates County).

II. Hind wing with \mathbf{M}_a and \mathbf{Cu}_1 more or less stalked, rarely stalked half way to; aner.

3. A. carbonana Heinrich (Kearfott ms.). Dark brown with patches of somewhat shining dark gray: a large area on basal half of costa, diffuse below; a band of gray along inner margin, abruptly widened at middle of wing, and there ending; two or three small gray patches in speculum; some smaller oblique hars beyond middle of costa; leaving the brown ground as an oblique fascia at middle of costa. Costa with many short blackish striæ, becoming silvery white toward apex. Fringe dark, with two heavy white bars below apex. 12-16 mm.

May and June.

²² These species belong to Anchylopera,

Maine to Virginia, west to White River, Ontario.

4. A. unguicella Linnaus. Eyes smaller and palpi larger and more hairy than in any other Ancylis. Shining violet-gray, strigose with blackish; basal third more finely strigose, its outer boundary moderately excurved; a barely traceable darker fascia from middle of costa to before anal angle; fringe with whitish base, and cut with two white bars below apex. Hind wing paler fuscous. (plagosana Clemens.)

May; July. Larva on heath.

Labrador, Manitoba, and probably general in cooler parts of Canada; Mt. Washington, New Hampshire, at 1,600 feet; Europe.

5. A. goodelliana Fernald. Whitish, dusted with light brown, with a diffuse but contrasting brown shade from base to apex; usually nearly interrupted about a third way out, where it crosses from the fold into the cell; broadening gradually to three-fourths way out where it abruptly narrows and is thence narrow and sharply defined to the apex. Costal striation light. Fringe pale except at apex. Hind wings slightly browner. 15 mm. (Fernald says 28 mm., perhaps a misprint for 18 mm.)

Mav.

Maine to North Carolina and Washington.

6. A. albacostana Kearfott. Dull gray, becoming lighter blue-gray toward outer margin and blackish below the white costal stripe, which runs quite uninterrupted from base almost to apex, and is widest at about two-thirds way out. Outer margin white, except toward apex; fringe gray-tipped. 19 mm.

May.

Tryon, North Carolina; Colorado, in May. New York: Black Brook (Adiroudacks. June).

7. A. mediofasciana Clemens. Ground a mixture of dull gray and light bluegray, more or less mixed with white, especially toward outer margin. Costal third white, without definite boundary below, crossed by a blackish fascia at middle, as in torontana (which is perhaps an aberration of this species). Apex and apical fringe blackish; rest of fringe white.

May and June.

Maine to California. New York: Karner, Normaskill.

8. A. torontana Kearfott. Fore wing mottled gray (badly stained in the type. which is the only specimen I have seen). Costa white from base practically to apex, with blackish dots along the costal edge; crossed by a broad oblique fascia at middle, and with a blackish shade below it, at least on outer part. 19 mm. (Proteoteras Kearfott.)

There is some superficial likeness to Proteoteras crescentana, but the strongly falcate apex definitely places this species in Ancylis, where it appears very close to mediofasciana.

Toronto, Ontario.

9. A. cornifoliana Riley. Basal half of wing dark gray-brown with some leadgray iridescence; the outer boundary of the dark portion right-angled at middle of wing; median fascia dark chocolate brown at costa, where it runs along the edge of the dark base and is strongly oblique; much broader below cell; separated from the dark base by a paler gravish patch, upright, and composed of a mix-ture of yellow-brown and chocolate, forming a marked chocolate patch on inner margin. A gray band beyond the median fascia, strongly oblique in on costal half, and angled in below. Outer part of wing ochreous, with some golden lustre; the costal edge dark brown with paired whitish striæ. Fringe lead gray. 7-12 mm.

May and June. Not rare. Larva on blackberry, cornel, and birch. General in distribution. New York: Ithaca, McLean. 10. A. ______. Similar to A. cornifoliana; ground more evenly blackbrown, the markings largely finely edged with clay-color; base strongly suffused with purplish gray, median gray fascia broad and fusing with the basal gray

toward costa; costal half of postmedial gray line almost longitudinal, the ground chocolate brown above it; lower half expanding into a broad diamond-shaped patch. enclosing a blackish marginal spot, and leaving only a narrow clay-colored mar-ginal line beyond it. Fringe deep ochre, contrasting. Hind wing blackish. (cornifoliana Murtfeldt).

May. Larva a leaf-folder on rose.

Hazelton, Pennsylvania; Missouri. 11. A. apicana Walker. Deep yellow-brown, becoming chocolate brown on basal third and toward middle of costa. Thorax brown, head paler. Base of costal edge of fore wing gray; a broad lead-gray fascia just before middle, of almost even width and moderately angled at middle of wing; a lead-gray line from just beyond middle of costa, obliquely out two-thirds of the way to the outer margin, then middle of costa, obliquely out two-thirds of the way to the outer margin, then sharply angulate and straight, almost paralleling the outer margin, to the inner margin; the angulation preceded by a few black bars. Costal edge outwardly blackish, with double white striæ. A black terminal line along middle of outer margin. Fringe dark gray. Hind wing mouse gray with pale fringe. 10 mm. May and June. Larva on raspberry. Nova Scotia and Pennsylvania to British Columbia. New York: Ithaca, McLean, Terreter Ealls. Wellweille

Trenton Falls, Wellsville.

12. A. divisana Walker. Base deep red-brown, shading into violet-gray toward costa, and followed by a broad violet-gray fascia. Fore wing outwardly shading from light ochre to dull orange on costa, the speculum at most a slightly duller pale brown or cream area. Costa toward apex finely striate with gray, with a couple of white striæ close to apex. Fringe pale brown. Hind wing pale gravish brown. 12-15 mm. May. Larva a leaf-roller on oak and sycamore.

Generally distributed. New York: Crosby (Yates County), Trenton Falls.

13. A. fragariæ Walsh and Riley. Ground tawny brown, cut into oval patches by bands of powdery violet-gray, which are narrowly edged with white. Base, as far as antemedial line, brown; the line fine, even, white; preceded by a blackish shade; strongly outwardly oblique from just below costa to A; then abruptly curving and meeting inner margin at right angles, beyond middle. Outer part of the dull violet gray with a long brown streak from middle of costa tapering to a point just before the concavity in the outer margin; and with an oval longitudinal patch below it, more or less defined with white, the two often fusing into an irregular patch. Fringe light brown; outer part of costa obliquely striate. 9 mm. (amblygona Zeller).

July. Larva on blackberry.

District of Columbia to Louisiana and Colorado.

14. A. floridana Zeller. Deep reddish or chocolate brown; the basal patch and antemedial line as in A. fragaria; costa at base and band beyond the antemedial line blue-gray, somewhat iridescent; outer part of wing toward costa chocolate to tawny brown, finely striate with double white lines; middle of wing shading into blackish; dorsal part lighter blue-gray or blackish. Fringe pale brown with darker tips. Hind wing mouse gray. 10 mm.

Larva on bearberry, and doubtless on other swamp Ericaceæ.

I have seen specimens typical of this form from New Jersey, Illinois, Missouri, and Iowa; and it is doubtless wide spread, but generally confused with A. comptana or some undescribed species. It is injurious, at least in Iowa. 15. A. comptana Frœlich. Dull umber brown, the basal patch as in A. fragaria,

but not as clearly defined with white. Ground outwardly lighter fuscous, without blue iridescence, the markings dull brown, without any yellow tint. Besides the oblique costal bands separated by paired pale strix, as in A. foridana, there is a rather comma-shaped dark patch on the middle of the outer part of the wing, and a dark spot at the anal angle, the outlines of the two forming a well-marked double

whitish oblique stria toward the anal angle. Fringe fuscous, cut with two white bars at notch, below which it is crossed by a blackish shade, pale below.

Larva injurious to strawberries in Europe. Moth in May, in New Jersey.

New Hampshire; northern New Jersey; and perhaps widespread. New York: reported from Glens Falls, Nassau, and West Farms.

It is uncertain to what extent the injurious strawberry leaf-roller of this country is comptana or floridana, or one of two or three undescribed forms of the group which appear to exist. Specimens from Lakehurst. New Jersey, in the region of injury, are apparently constant and represent an intermediate form with practically the coloring of floridana, a little dulled, but with the outer patches of comptana distinctly marked. The same form occurs in Vermont and Manitoba and Colorado and has been confused with Anchylopera angulifasciana.

A. loricana Grote. Shining bronze brown, with bright yellow markings on outer part of costa and outer margin.

This species was described from Dayton, Ohio, but has not been rediscovered, and is probably exotic.

9. ANCHYLOPERA Stephens

(Restricted; Ancylopera auct.; Ancylis, in part; Phoxopteris in part)

Similar to Ancylis, but with M_3 and Cu_1 completely united (fig. 264). Palpi never as hairy as in some Ancylis. The falcation of the fore wing is always broad and well marked. Markings characteristic, a little modified in A. nubeculana. Fore wing with a dark basal patch not reaching the costa, and with its upper boundary curving into the oblique outer boundary; costa contrastingly pale. An oblique median fascia from costa to middle of outer part of wing, where it is usually somewhat widened and abruptly truncate, outlined on upper side with white, and usually also on lower outer side by a fine line which is often partly lead colored. Costa outwardly with white strize, more or less distinctly paired; usually ending a little above the oblique fascia. Fringe usually dark at apex and pale below.

The genus is quite homogeneous and derived from the *comptana* group of Ancylis; for a key to the species, see Ancylis.

1. A. nubeculana Clemens. White; outer part of wing shaded with light gray; a short oblique bar at middle of costa. Apex shaded with brown, with a couple of black subterminal dots before the notch. A blackish patch on inner margin, with quite irregular upper boundary, extending nearly across cell at a third way out; ending at middle of wing, but followed by a patch of dark blue-gray which is hardly distinguishable from it. Fringe broadly fuscous tipped, cut with white below apex, and broadly white at anal angle. 15 mm. Common and general in distribution in May and June. The larva a leaf-roller

on apple and other trees, but rarely in injurious numbers. New York: North Elba, Peru, Palmyra, East Bloomfield, Rock City (Cattaraugus

County), Crosby (Yates County), McLean, Ithaca, Niverville.

2. A. subæquana Zeller. Ground white, slightly tinted with clay-color outwardly and especially toward costa. Basal patch brown-black, irregular, its outer part extending up into cell as a more or less obvious lobe. Median fascia from costa weak or obsolescent; when strongest, clay-color. Speculum frequently inconspicuous, sometimes defined by clay shades before, and beyond, and above it, the former crossed by some black lines. A black apical dot; fringe sometimes cut with dark gray at middle. Hind wing light gray. 15 mm.

June and July.

Nova Scotia to the West Coast, south in the mountains to Virginia; in the Northwest replaced by a grayer form, kincaidana Fernald. New York: Black Brook (Clinton County).

Easily distinguished by the peculiar form of the basal patch, and lack of yellow coloring.

3. A. semiovana Zeller. Markings gray-brown with a slight golden iridescence, on a white ground. Dorsal patch large, extending two-thirds way out on inner margin, evenly rounded, and followed by a white fascia. Outer half brown, from middle of costa and from about three-fourths way out on inner margin; a large trapezoidal white costal fascia at two-thirds way to apex, containing a very slight brown costal stria; the costal striæ following it not distinctly in pairs. Speculum not distinct. Fringe contrasting, pale. 13 mm.

May and June.

Canada to North Carolina. New York: Ithaca.

4. A. maritima Dyar. Clay-color, with a slight olivaceous tint, the markings edged with white. Dorsal patch long and narrow, the outer side strongly oblique, and ending at the tip of the anal vein, where the spot shades into the ground. Costal end of median fascia clearly defined, and lower end defined by a curved white line; the fascia longitudinal in general course near middle of wing; the two black dashes in the outer angle of the fascia distinct. Costal striæ normal; apex dark; fringe concolorous. Hind wing mouse gray. 10-12 mm.

Larva in summer on beach pea; moth the following spring and again in August. The species will probably occur generally along the coast.

Kennebunkport. Maine, to Speonk, New York.

5. A. pulchellana Clemens. Ground whitish, shaded with light fawn; median fascia hardly darker, clearly defined on outer side by a silver-white line which forms a sharp angle opposite the notch; where it contains a couple of obscure brown dashes. Costal striæ fine, silvery, defining slightly darker yellow-brown bars. Speculum typically represented by a dark dot. Apex yellow-brown; dorsal patch yellow-brown, large and rounded. 13 mm.

May to July.

Massachusetts to Virginia and Illinois. New York: Ithaca.

6. A. angulifasciana Zeller. Ground dull luteous; dorsal patch and fascia blackish, the former rather short and wide, ending rather squarely on inner margin; the fascia broad, defined on outer side with an angulate dull silver line, with a couple of black dashes in the angle. Costa outwardly with irregular pale and dark short striæ, the ground of the whole outer part brown, but paler than the fascia. 8 mm. May to July; late Augnst. Larva on clover.

I have seen this species confused with A. floridana, from which it can be readily distinguished by the contrasting pale costa. It is easily recognized by its small size and heavy, sharply contrasting markings.

size and heavy, sharply contrasting markings. Generally distributed and not rare, westward to Illinois and Manitoba. New York: Crosby (Yates County), Ithaca.

York: Crosby (Yates County), Ithaca. 7. A. platanana Clemens. Ground cream-white, including thorax and whole base of wing. Dorsal patch defined only on its outer half, which is rusty ochre. Median fascia ochre, often interrupted by the cream-white ground, below costa; with two or three heavy black dashes in the angle, and often another in the speculum, which is more or less suffused with ochre. Costal striæ light ochre, in a cream-white area; all the markings tending to become diffuse. 10 mm.

a cream-white area; all the markings tending to become diffuse. 10 mm. Other specimens are larger and more diffuse in markings, with the dorsal patch almost completely lost, and with the black strime lighter. They may belong to A. platanana, but I suspect they are suffused forms of the burgessiana group. June.

Apparently general in distribution. New York: Rock City (Cattaraugus County), Big Indian Valley, New Windsor.

The remaining names have been used more or less interchangeably for specimeus in which the basal patch is well defined, of moderate size, and varying from yellowbrown to black in color; the faseia yellow-brown, usually markedly paler than the basal patch, defined on the outer side by an angulate silver or gray line, and con-

taining a couple of gray streaks in the angle; the costal striæ all paired, fine and short, on a yellow-brown ground, much darker than the whitish basal half of the costa. The specular region is yellow-brown or yellow, and is darker than the region just beyond the dorsal patch, but not sharply set off either from it or from the lower end of the costal fascia. The apex is a little darker, the fringe a little paler and duller brown. The hind wing is pale fuscous. I suspect that only one species, with a tendency to form numerous local strains, is represented in this series, but give the points of distinction as noted by their describers.

8. A. spiræifoliana Clemens. This is the oldest name of the series. As described, the fascia is slightly paler than the dorsal patch, both, however, being dark brown. The patch hardly enters the cell, and its outer boundary is but slightly sinuous. Larva on Spiræa opulifolia, folding a leaf longitudinally and living on the parenchyma within the fold.

Described from Pennsylvania.

9. A. burgessiana Zeller. Dorsal patch broad, variable in shape, brown, with 9. A. Burgessiana Zener. Dorsal patch broad, variable in shape, brown, with some golden gloss. Median fascia dark rusty brown, the whitish area before it more or less dusted with gray. Outer part of wing rusty. Hind wing typically blackish; nearly white in var. pruni Heinrich. 15 mm. Type locality, Massachusetts. June and July. Larvae on oak and plum (pruni). New York: Rock City (Cattaraugus County), Ithaca, McLean, and Karner. Generally identified with murtfeldtiana.

10. A. murtfeldtiana Riley. Small. Dorsal patch black-brown; median fascia yellower, reaching inner margin (though much paler below and diffuse on inner side), followed the whole distance by a whitish line, which is waved toward the inner margin. Speculum usually with a heavy black dash, preceded by a leadgray patch in the fascia. 12 mm.

May. Larva on oak.

Described from Missouri.

A. metamelana Walker is considered a synonym of *spiraifoliana* by Fernald. The dorsal patch is blackish, the fascia red-brown, apparently not reaching the inner margin. 17 mm. Type locality unknown.

A. discoferana Walker. Dorsal patch and fascia both brown, costal striæ black. This is also considered a synonym of spiræifoliana, but the dark coloring and small size (10 mm.) suggest rather angulifasciana.

11. A. laciniana Zeller. Patch and median fascia both ochre yellow, the wing toward apex heavily shaded with ochre yellow; with distinct black dashes in the angle of the fascia and some black scales in the speculum. 15 mm.

Described from Massachusetts.

This form seems identical with the following:

A. dubiana Clemens. Cream white, with all markings ochre yellow except the two black dashes.

This form was described from Virginia. I have seen similar specimens from Pennsylvania, Ohio, and elsewhere.

12. A. fuscociliana Clemens. Ground cream to light ochre; paler toward base of costa; dorsal patch large and rounded. Fascia hardly darker on costa, quite concolorous below, and shading into the ground color; defined on outer side with an angulate gray line, formed by the fusion of the first two costal striæ. Black dashes in angulation of median fascia strong. Inner margin generally suffused with dull brown beyond the patch; this suffusion slight in some females. Fringe brown, duller than ground color. 16 mm.

June. Larva on chestnut and elm.

Generally distributed and not rare. New York: Rock City (Cattaraugus County), Crosby (Yates County), Ithaca, McLean, Trenton Falls. This is hardly more than a subvariety of *burgessiana*, but appears to have a

different range of food-plants.

-. Light gray-brown, with some faint yellow tint over paler 13. A. fortions of outer half of fore wing. Markings as in A. burgessiana and A. fuscociliana, but easily distinguished by the lack of yellow or tawny. Costal striæ double, the first pair only moderately enlarged; fringe mixed whitish and ash gray. 15 mm. (spiræifoliana Heinrich, not Clemens; discigerana Walker ?). June and early July.

Parry Sound, Ontario, to Pennsylvania. New York: Newcomb, Rock City (Cattaraugus County), Ithaca.

The specimen now standing as type of *spiræifoliana* is this, but as Clemens' supposed types were not labelled till long after his death I prefer to give more weight to his description, which is of a yellow-marked species.

A. lamiana Clemens appears to be Ancylis floridana Zeller. A. parmatana Clemens is not an Anchylopera but the species more generally known as crispana.

10. NORMA Heinrich

(*Epinotia* Kearfott, in part)

Similar to Ancylis; fore wing with apex strongly produced, but not actually falcate, outer margin strongly concave. \mathbf{R}_2 of fore wing arising from cell.

Valve simple, with several long ventral spines from margin of cucullus; sacculus spinose. Uncus of two widely separated, weakly chitinized points. Socii long and broad, ribbon-like.

1. N. dietziana Kearfott. Basal third blackish, median area white; outer twofifths heavily shaded with light blue-gray. Antemedian line only a little excurved, the dark hase more or less strigose with pale gray. A dark gray spot in middle of wing two-thirds way to apex, often connected to inner margin by .a lighter gray shade; some oblique gray striæ on costa; and apex shaded with dark gray, with a black apical dot. A dark gray terminal line below notch, not reaching anal angle. Fringe mixed gray and white. 15 mm. (*Epinotia* Kearfott.) May and June. Larva on Cratægus.

New Hampshire; Pennsylvania; Arizona. New York: Rock City (Cattaraugus County).

11. RHOPOBOTA Lederer

(Eudemis auct., not Hübner)

Similar to Ancylis; palpi large, porrect, and clavate; as in some Ancylis. Thorax practically smooth. Fore wing smooth, with R_4 and R_5 long-stalked (fig. 263). A deep notch below apex, to which M_1 to M_2 converge. Cu, widely separated in the typical forms; convergent, but not actually running to the notch in R. *ilicifoliana*. Hind wing ample, with **R** and **M**₁ approximate; **M**₅ and **Cu**, stalked; in male, with a patch of black sex-scales over costa and cell. Larva a leaf-roller; with black head.

Valve of male with a row of stout marginal spines; uncus bifurcate, weak; socii large, with ends fusing in a hairy knob.

1. R. nævana Hühner. Dull brown with two very broad bands of lead-gray and a shorter narrower one near outer margin. Costa outwardly with heavy, paired, lead-gray striæ, often connected with the transverse bands, especially in the female, but in the male usually separated by a streak of the brown ground. Almost always with a narrow antemedial gray band, 8 mm. (vacciniana Packard).

Larva on cranberry and other dwarf shrubs; sometimes injurious to cranberry, where it is known as the "black-head," to distinguish it from *Peronea minuta*. Common, especially in peat-bogs; south to Pennsylvania, at least; also in

Europe.

2. R. ilicifolana Kearfott. Dull fuseons, somewhat powdery and strigose. Inner margin contrastingly pale beyond middle; the boundary erect. Costa pale at second fourth, then more or less suffused with dark in the form of a broad triangle whose apex is formed by a black spot at the end of the cell; this shade reaching from the middle of the costa to the apex. 10-13 mm.

June. Larva on terminal leaves of llex.

New Jersey, North Carolina, British Columbia. Heinrich considers this a food-variety of R. nævana.

12. KUNDRYA Heinrich

Similar to Norma. Fore wing with \mathbf{R}_4 and \mathbf{R}_5 completely united. \mathbf{R}_2 arising from discal cell. Male with only one long ventral spine on cucullus of valve.

1. K. finitimana Heinrich. Fuscous brown, more ferruginuous toward apex of forc wing. Base dark; a fairly broad antemedial and a median pale fascia, somewhat scaled with lead color; speculum pale, with the inner bar leaden. Usually a black dot in apex; fringe fuscous with a black basal line. Hind wing dark. 9-10 mm.

Larva on *Ilex verticillata*. Moth in June and July. Pupa with large protruding orange spiracles.

New Hampshire; Virginia.

13. EPIBLEMA Hübner

(Eucosma, Padisca, etc., in part, with Notocelia Hübner)

Similar to Eucosma. Fore wing (fig. 265) normally with \mathbf{R}_1 arising well before middle of cell. Hind wing frequently thickened at inner margin, with a more or less distinct hair pencil; strong in *E. suffusana* (Notocelia Hübner). Thorax often tufted (figs. 278-279).

Male with valve bearing a small but distinct clasper; no strong anal or lateral spines on cucullus; anal indentation sometimes densely hair-spined. Costal fold always present.

A few of the larvæ are gall-makers.

This genus merely represents the more primitive portion of the Eucosma series, and is possibly the most primitive of the Eucosminæ, rather than Hemimene; as the spined abdomen of the pupa would indicate. As there is no superficial character to separate the species from Eucosma, they are included in the key to that genus.

Key to the species

- - 2. A triangular patch on outer margin, narrowing to both costa and inner margin 6 culmingna

4. boxcana.

1. No contrasting blackish patch along outer margin or at anal angle.

- 2. Dark brown with contrasting pale speculum only.

 3. Smoky brown

 3. Purple-gray and brown

 2. abruptana
- 2. Otherwise marked.

3. A clean-cut, irregular, oblique, white patch at middle of inner margin.

3. otiosana.

3. No such patch.

- 4. Deep purple-gray and umber brown, with a clean-cut, contrasting, median white patch.
- 5. Patch covering a third of the area of the wing.....9. tripartitana.
- 5. Patch half as large 10. walsinghami.
- Lighter, at least outwardly; patch rarely well-defined.
 Fringe concolorous, powdery gray; a small species...5. abbreviatana.
 Fringe contrasting and dark, at least toward apex; anal angle often white like speculum.
 - 6. Hind wing and outer third of fore wing whitish; the base of fore wing contrasting, blackish16. illotana.
 - 6. Hind wing dark; fore wing darker, with much blue-gray, at least in postmedial region.
 - 7. Speculum mostly white, connected with dorsal patch.

11. dorsisuffusana.

- 7. Speculum with broad blue-gray bands. 8. Fringe all powdery fuscous; base of wing white-scaled;
 - speculum with a contrasting dark spot.....12. carolinana. S. Fringe white at anal angle; base not white-scaled; specular

region whitish.

9. Median area pure white; normally large species.

- 14. kennebecana, 13. scudderiana. 9. Median area pinkish white with seattered dark scales;
- 9. Median area somewhat pinkish or dirty white, with several strong gray striæ; speeulum obsolete 15. obfuscana.
- 9. Median area mixed white and gray, without pinkish tinge;

Unplaced: E. hirsutana, E. ochraceana,

1. E. strenuana Walker. Base of antenna sometimes black. Fore wing smoky brown, lightly and rather evenly dusted with luteous-tipped scales, except the dorsal part of the median area; which is sometimes much less dusted or clear. Antemedial line obscure, rarely fine; distinct and pale on dorsal half. A tri-angular area along outer half of costa and extending down on outer margin to \mathbf{M}_1 ; striate with whitish, lead-gray, and often light brown; a pair of apical striae palest, and often contrasting. Speculum a mixture of white and lead-gray, the white meetly in the upper upper portion end containing come black dots: the white mostly in the upper outer portion, and containing some black dots; speculum preceded by a fine broken black line or a couple of dots. Fringe broad, the basal half powdery. 10-17 mm. (*flavocellana* Fernald, *minutana* Kearfott). May to September. Larva boring in Ambrosia; typical specimens especially in A. trifida, and small ones (minutana Kearfott) in A. artemisicofolio.

General in distribution and not rare. New York: Stanley, Poughkeepsie (New York State Museum), New York City (Watson).

2. E. abruptana Walsingham. Base of antenna inky black. A mixture of dark purple-gray, brown. and black, without contrasting markings on the general surface. Costa with some white striæ, a strong one just before apex. Speculum strongly contrasting, white and silver, with a suggestion of black streaks. A little yellow iridescence in the speculum and between it and the apex. Fringe above blackish and obscurely banded; white at anal angle. 13 mm. (probably crispana Clemens).

June.

Northern New Jersey; Virginia; Texas.

3. E. otiosana Clemens. Gray, shaded with blackish; the base blackish; with postmedial patches towards costa and on inner margin, separated by a narrow paler oblique gray band, as in Sonia constrictana; but often obscure, and, more

rarely, partly filled with white; the outer boundaries of the dark base inwardly oblique to the dorsal margin; a white dorsal patch, filling the space between the dark base and portmedial patch, usually irregularly quadrangular, with anterior sides roughly parallel and cut off squarely in the middle of the wing; sometimes with a narrow extension between the two postmedial patches. Speculum light gray with some black in the middle; costa with double strive toward apex. 15 mm. (Monosphragis Clemens.)

June to September. Larva boring in pith and wood of stems of Bidens; hibernating as a larva in the dead stalks.

Common and general in distribution. New York: Peru, Ithaca. Big Indian Valley.

4. E. boxcana Kearfott. Dull gray-brown, shading into deep brown on inner margin, especially between the median dorsal patch and the speculum. Outer half of costa with paler gray and darker brown oblique striations, fading out below. Middle of inner margin with a contrastingly paler, sometimes whitish patch, made up of four or five whitish striations, converging to their tips near the middle of the wing, and more or less fused; practically as in several Laspeyresias. Speculum contrasting, pale blue-gray, with a couple of black dots. Rase of fringe more or less powdery. Hind wing concolorous. 15 mm.

May.

New Hampshire to Texas and Iowa.

5. E. abbreviatana Walker. Pale gray, dusted with blackish, with a slight blue tint; base and outer part a little darker. Antemedial line obscure toward costa. erect, and a little irregular, below. A few geminate white striæ toward apex. Speculum of two vague lead-gray bands, with some black dots between them. preceded by a brown area containing black dots. Fringe powdery gray. One of the smallest of the genus. 10 mm.

May and June.

New Hampshire to Toronto, Ontario, and Missouri; south to the District of Columbia.

6. E. culminana Walsingham. Light gray; strigose; the base slightly darker. A rounded, triangular, deep brown patch resting on costal two-thirds of outer margin, enclosing a pale gray marginal spot, and a small, subtriangular blackbrown spot on inner margin before the anal angle, the space between the two more shining. Fringe deep brown, contrasting, with black basal line; white at the anal angle. Hind wing pale gray. Head and collar deep brown. 16 mm.

August and September.

Massachusetts to District of Columbia, Manitoba, Washington, and Colorado. This species approaches *Rhyacionia picicola* closely in appearance, but is smaller. with browner patches, and has a strong costal fold. Its dark head is also distinctive.

7. E. brightonana Kearfott. Wings broad. Grayish brown, with dark brown patches; the one near the base of the inner margin large, as wide as high, and strongly contrasting; a narrow oblique fascia from middle of costa to end of cell. followed by some double pale costal striæ with dark streaks between them. Apical region brown, more widely on the costa where it contains a double pale stria. the brown tapering to a point near anal angle. A small spot on outer part of inner margin. Speculum obscure, somewhat shining, with a vertical darker streak in the middle. 12-15 mm.

August.

New Brighton, Pennsvlvania. New York: Ithaca.

8. E. tardara Kearfott. Wings rather broad, dull gray, like dorsisignatana. A triangular blackish antemedial patch on inner margin, reaching up to the fold: and a triangular apical patch; extending, on the costa, from four-fifths way out to apex, and down to Cu on the outer margin; but separated from the outer margin, except at apex and lower end, by a narrow strip of the ground color. 20 mm.

June and July. Montclair, New Jersey, to Plummer's Island, Maryland; west to Illinois.

9. E. tripartitana Zeller. Dark purple gray, mottled with blackish; the dis-tincter black spots more or less clearly edged with brown. Median area white except for the costal fold of the male, which extends two-thirds way to the apex. Antemedial line erect, a little wavy, and rather nearer base on inner margin; outer boundary of median white area oblique and irregularly excurved, nearer base on costa. Speculum a vague blue-gray area, preceded by black dots in the blue-gray ground. 20 mm. August. Larva in a gall on Rudbeckia Iaciniata and on Solidago.

Southern New York to Texas and Florida.

10. E. walsinghami Kearfott. Eyes rather small. Ground a confused mixture of deep brown, purplish, and black scales, the specular region rather tending to lead-color, and sometimes with light brown striæ toward apex. Middle of inner margin with a large semi-elliptical white patch, formed of four partly fused striæ. Fringe of fore wing coal black, with the usual black basal line not contrasting. 15 mm.

May.

Hemlock Falls, New Jersey (type); Oak Station, Pennsylvania; Alabama. New York: Rock City (Cattaraugus County).

11. E. dorsisuffusana Kearfott. Black, somewhat shaded with dark gray. Dorsal half of wing white from a third way out, contrasting; the boundary somewhat irregular and indented above at two-thirds, where the white area is divided by a group of black dots; a few black dots on middle of inner margin; outer part of costa brown, with paired white striæ; upper half of outer margin narrowly brown, cut with white at M₁, where the brown is preceded by a black dot; fringe grav above, whitish at anal angle. 20 mm.

In the male type, the black dots in the white area are almost lost.

June and July.

Cincinnati, Ohio.

12. E. carolinana Walsingham. Base and costa gray, with a slight blue tint, striated with blackish; medial area below middle of cell white or pale; outer part a confused mixture of blue-gray and white, with a group of black dots at apex, and two irregular vertical rows before the region of the speculum, finely defined with white. Fringe slightly powdery fuscous gray, without basal line. Hind wing mouse gray. 16-25 mm.

July and August.

Northern New Jersey to western Pennsylvania; south to North Carolina. New York: Ithaca.

13. E. scudderiana Clemens. Base dark purple-gray and blackish, the area darker and less extensive than in illotana, with sharply defined somewhat irregular outer boundary about a quarter way out; median area white, with a few gray striæ, light gray on costa; postmedial area mixed dull gray and light blue-gray, with some white; the speculum confused and slightly more white; preceded by a few black dots, one of them good-sized, and with a couple of fine black dots in its upper portion; a brownish area between speculum and apex. Fringe white toward anal angle only. Hind wing mouse gray. 20 mm.

Rather common. The larva bores in the crown or forms a slight fusiform gall in goldenrod; emerging the following June.

Generally distributed. New York: Lewiston, Buffalo, Ithaca, Albany, New Windsor.

14. E. kennebecana Kearfott. Fore wing white, shading into gray along costa; base dark gray, strigose with black; the antemedial line moderately angulate in

cell. Costa outwardly with paired fine white striæ; the ground toward apex shaded with yellow-brown. Speculum silver and cream-white, with some black scales; preceded by a region striate with black, and followed by a black bar; fringe light; the basal series of scales narrowly black-tipped, but hardly forming a line. Hind wing mouse gray, paler at base between yeins. 13 mm.

a line. Hind wing mouse gray, paler at base between vehics. 13 mm. August. Type only seen. The veins in the fore wing are distinctly approximate at the margin, which is concave.

Kennebunkport, Maine.

15. E. obfuscana Dyar. Base gray, transversely striate with blackish, its outer boundary about one-third way out, ercct, and very slightly excurved; somewhat diffuse. Outer part of wing white, striate with light gray, slightly more densely in the postmedial region. A black apical dot, and terminal bar at middle of outer margin; fringe dull black, whitish at anal augle. Hind wing dark gray. 16 mm. (obfuscata Riley, undescribed).

May to September. Larva in Solidago.

General in distribution. New York: Rock City (Cattaraugus County), Ithaca; Clove Valley, Staten Island.

16. E. illotana Walsingham. Wings broader than in *scudderiana*. Base blackish, mottled with dull black on blue-gray; outer part nearly white, the boundary diffuse and farther out on costa; wing shaded with light gray toward apex, and before anal angle; speculum represented by two or three black dots before outer margin; fringe gray on costal half. Hind wing nearly white. 15 nm.

May and June.

Probably generally distributed. New York: Kinderhook.

17. E. desertana Zeller. Basal third of fore wing blackish; strigose; with waved, erect, outer boundary; rest of wing white, with some gray strix, gathering to form a distinct gray and black triangular patch before the speculum, which is a mere silvery patch, with an oblique gray line bounding it above, resting on the outer margin. Terminal line and fringe blackish except at anal angle. Hind wing gray. 15-20 mm.

Larva in Solidago. Moth at end of May.

New York and District of Columbia to Texas. New York: Peru, Ithaca, New Windsor, Ramapo.

18. E. suffusana Zeller. Hind wing with a deep fold on upper side, below 3d A; a thickening between it and the inner deep margin, containing a hairy pencil arising near the base of the wing. Base, a postmedial patch toward anal angle, and apex dull gray; the medial area and speculum white, rather heavily shaded with light blue-gray, more heavily than in *Eucosma scudderiana;* costal edge dark gray with double white striæ on outer two-thirds; some small black dots in speculum, and a transverse series of stronger ones just before it; fringe gray toward apex, white below; hind wing mouse gray. 16 mm. (Notocelia Hübner).

European specimens usually show considerable brown shading toward the apex and before the speculum, but more rarely are quite like the American strain, which is probably introduced.

June. Larva a bud-worm on rose (fig. 278).

Maine; northern New Jersey, Maryland; Pennsylvania; Europe.

14. EUCOSMA Hübner

(With Pædisca, Steganoptycha, Halonota, Spilonota, etc., in part; Euryptychia; Monosphragis Clemens, etc.)

Palpi somewhat rough and beaklike (fig. 280); head rougher than in Laspeyresia and Hemimene, normal. Thorax smooth or tufted. Hind tibiæ with a moderate amount of loose hair only. Fore wing normally rather long and oblong, the outer margin typically smoothly rounded, but often concave in the middle, with M_1 to Cu_1 approximate toward margin. Apex rounded or acute, never strongly falcate, but occasionally subfalcate. All veins separate, the separation between M_2 and M_3 at base well marked. Male with a well-developed costal fold; so far as examined, containing a hair pencil. Hind wing trapezoidal, normal, without sexual modification; R and M_1 normally approximate at base, never separate; M_2 almost connate at origin with M_3 and Cu_1 , which are stalked, or rarely united.

Larvæ usually borers in the stems or roots of herbaceous plants, especially Compositæ.

The genus is too large and rather heterogeneous, but attempts to divide it have, so far, not been very successful. Presumably several genera are derived from it by the loss of the costal fold.

The following key to species must be used with caution, as many species, especially of Epiblema, run very close. Females of Thiodia and related genera must also be watched for; a few of these are included in this key for convenience.

Key to the Eucosminæ with costal fold

1.	With extensive silver or pure white areas2.
1.	Not largely silvery or pure white
2.	Not largely silvery or pure white
2.	Fore wing with more complex markings
3.	A transverse median or antemedian silver fascia4.
3.	A longitudinal silver streak from base to end of cell
	Basal half white, with an incomplete brown fascia29. Eucosma grotiana.
	With an upright silver bar between the small basal spot and the fascia.
	8. Eucosma robinsonana.
4.	No spot between the basal one and the fascia7. Eucosma quinquemaculana.
5.	A single silvery costal median spot, extending beyond the end of the streak
	along lower edge of cell
5.	along lower edge of cell9. Eucosma ridingsana. An antemedial spot, almost reaching the base on the costa, as well as the
	medial one11. Eucosma hipeana.
5.	Image and the strength of the strengh of the strength of the strength of the st
	streak10. Eucosma argentifurcatana.
6.	Ground color largely bright green9. Epinotia medioviridana.
8.	Without green
7.	Speculum with a group of regular rows of black spots8.
	Speculum with three or four streaks or spots, or absent
8.	Area above speculum evenly buffl. Eucosma circulana.
8.	Area above speculum dusted with black or brown9.
9.	Base of fore wing evenly yellow, not dusted10.
9.	Base of fore wing, at least along inner margin, dusted with brown12.
10.	With transverse silvery gray bands toward base5. Eucosma fraudabilis.
10.	Base wholly yellow
	wholly dusted with brown
u.	Less than half of this area dusted with brown
	Ground of apical half, and a streak in cell almost to base, yellow, not dusted. 2. Eucosma scintillana.
12.	Wing almost wholly dusted with brown
12.	Basal fourth dusted with brown, contrasting4. Eucosma fratruelis.
13.	M, and Cu, of fore wing becoming coincident at outer margin.
	19. Eucosma cataclystiana.
13.	M ₃ and Cu ₁ separate

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14. Ground color bright yellow, almost without marks....22. Eucosma bipunctella. 14. Ground pure white, but marked with blackish..29. Eucosma grotiana. 14. Ground pure white on basal half of wing, contrasting with the brown apical 14. Ground color varying from cream to buff or bright ochre, with black scates on speculum only15. 14. Ground brown, gray, or orange; or, if whitish, with considerable black scaling on general wing-surface, or black marking......20. 15. No speculum; fore wing lightly strigose with pale brown on pearl gray. 25. Eucosma engelana. 15. Speculum distinct, containing two or three black streaks or dots......16. 16. Base and most of costa whitish, contrasting with the brown area about the speculum 16. Ground color usually yellowish, the outer part not much darker 17. 17. Base, except toward costa, with scattered silver dots......18. 17. No silver dots on basal half of wing......19. 18. Bright ochre yellow, all white markings clean-cut and contrasting. 18. Eucosma graciliana. 18. Pale ochre yellow, shading into cream, the marks in part ill-defined. 17. Eucosma albiguttana. 19. Cream-color; longitudinally strigose with pale gray. 20. Eucosma pergandeana. 19. With rounded pale yellow areas separated by bands of powdery light brown. 21. Eucosma pallidipalpana. 20. With a distinct speculum, containing two or more black dots or streaks in a pale area between two silvery or lead gray bars, or with the speculum represented by a silvery patch, preceded by distinct black dots, toward the 20. No special marks at anal angle; at most, with a couple of silvery or leadcolored bands in species which have such markings on other parts of the wing also; white anal patch, if present, not preceded by black dots......54. 21. A white patch on middle of inner margin; sometimes lightly streaked with gray, but contrasting with the general surface; or with a larger whitish 21. No white at middle of inner margin; the patch sometimes traceable but so 22. Patch more than a third as long as wing, extending at least to the costal fold, and with sharply defined boundary.....9. Epiblema tripartitana. 23. Speculum also a large and clearly defined whitish patch, separate from the 24. Outer part bright brown......16. Epinotia walkerana. 24. Ground color all gray or dull brown, or with a very little brown at outer [25. Vacant.] 26. White patch pure white, with clean-cut boundary all around, mostly defined 27. Base of fringe powdery or diffusely gray, rarely fuscous; fringe often white

28.	Antemedial line running obliquely in to inner margin; inner lead-colored band
28.	of speculum definite
	irregular and broken
	A triangular blackish patch more than 1 mm. wide at the middle, resting on outer margin
29.	No triangular marginal patch
30.	Whole outer two-thirds whitish
30.	Whole outer two-thirds not whitish
31.	A squarish blackish patch before the speculum, as dark as the base; fringe
91	blackish
	costa
32.	Dorsal area, between white patch and speculum, of the fuscous ground color, somewhat shaded with black
32.	This area scaled with blue-gray and white; with clearly defined black
33	dots
00.	Speculum with broad blue-gray bands
33.	Speculum with broad blue-gray bands
34.	Fringe all powdery fuscous; base of wing white-scaled.
	12. Epiblema carolinana.
34.	Fringe white at anal angle; base not white-scaled
35.	Base deep brown and blue-gray; median area strongly contrasting, white, with
	a few gray striæ13. Epiblema scudderiana.
35.	Base of a less intense smoky gray and blue-gray; median area, half gray, less contrasting
36.	Dorsum wood brown, contrasting with the blackish costal region; speculum
.,	of the lightest wood brown, with a single clear black dash in its upper
	portion15. Eucosma rusticana.
36.	Dorsum not broadly wood brown, contrasting with a blackish costa
37.	Wings blackish, the speculum a contrasting pale patch
37.	Wings not solid dark with pale speculum
38.	Terminal joint of palpus and second joint of antennæ black. 2. Epiblema abruptana.
20	Terminal joint of palpus not black
39	Deen brown the specifium very obscure
39.	Usually light gravish brown or grav: never brown with obscure markings41.
40.	Costal strige at middle of wing blackish: region above speculum not dusted
	with white12. Eucosma mandana.
40.	with white
	with pale-tipped scales
41.	With sharply defined evenly colored dark patches42.
41.	No such patches
42.	No such patches
40	and near anal angle
	ground
43.	Somewhat less defined ones also at end of cell and along outer margin toward
40	costa
43.	With the two dorsal patches only
44.	With a clear black line in base of fringe, usually interrupted with white
11	opposite end of cell
44.	line

45.	Gray; the scaling wholly black and white, or with a brown patch on middle of inner margin
15	Brownish with a mildentile house a cline and in the line in the second state.
40.	Brownish, with considerable brown scaling, especially along outer margin. 46.
46.	Costal striæ conspicuous; outer two-thirds of wing pale gray.
	20. Epinotia solicitana.
46.	Costal striæ obscure; some lead color toward apex. 17. Epinotia hamptonana.
47	With central area of speculum luteous, and a luteous irregular patch along
	base of inner margin
	base of inner margin
41.	Without lutcous markings along inner margin or in speculum
48.	Ground nearly evenly blackish; speculum surrounded by three dark lead-gray
	patches
48.	Paler
40	Head and thorax conspicuously hoary
40	Head and thorax nearly even brown
40.	Heat and thorax hearly even brown
ə 0 .	Outer margin convex with a couple of black dots before speculum
	5. Epiblema abbreviatana.
50.	Outer margin concave; a dark vertical line or larger shade before speculum51.
51.	Outer margin evenly concave on upper half
51	Outer margin with a small notch as in Proteontery 2 Erenters improband
50	Ach may mile contracting doub has and chlique medica facia
0Z.	Ash gray with contrasting dark base and oblique median fascia.
	28.1. Eucosma palabundana, 28.2. Eucosma fiskeana. Not so marked
52.	Not so marked $52\frac{1}{2}$.
521	2. Speculum large, conspicuous; antemedial line concave below.
	31. Eucosma zomonana.
591	2. Speculum obscure;; antemedial line all convex30. Eucosma gomonana.
53.	Postmedial patch on inner margin obliquely truncate and clean-cut above.
	separated by a narrow even oblique stripe of the ground color, from the
	postmedial patch at the end of the cell; or with the two patches fused into
	a fascia which is sharply indented on its basal side at the fold.
	1. Sonia constrictana.
53	Otherwise marked
54	Hind wing bright orange
04.	Hind wing bright orange
54.	Hind wing not bright-colored55.
55 .	With sharply defined contrasting dark brown patches, but the base of the wing
	of the paler ground color
55.	Without such patches
56	Two subequal dorsal patches only19. Thiodia tomonana.
56	A patch near base of dorsum and one along outer margin toward apex.
50.	A patch hear base of dorsum and one along outer margin toward apex.
	8. Epiblema tandana.
56.	A large median patch like an inverted V, clearly defined on its lower part and
	strongly contrasting, but tending to fade out toward costa.
	24. Eucosma similana.
56	A conspicuous patch toward base of inner margin, and a dark, but not always
00.	clearly defined, patch over end of cell; the one at anal angle small or
	clearly defined, patch over end of cell; the one at anal angle small of
	absent
56.	A large patch on outer margin and a small one at anal angle
57.	Expanse 16 mm
57	Expanse 30 mm 10. Rhyacionia picicolana.
58	Height of antemedial spot more than one and a half times its width.
00.	var. diffusana.
- 0	
58.	Antemedial spot wider, often as wide as highTypical form.
59.	With a straight oblique shade from middle of costa to anal angle.
	23. Eucosma juncticiliana.
59.	A broad brown oblique median fascia, meeting another from apex at middle
	of wing

59.	No such shade
60.	Ground color rusty orange
60	Ground color not orange
61.	Ground color not orange
61	With transverse pairs of gray striæ, as in Evetria33. Eucosma monitorana.
62	With a white basal dash shaded below with black35. Eucosma bilineana.
62	No black and white basal dash
63	Ground largely whitish, or strigose with light blue-gray; the base darker 64.
63.	Not dominantly white or heavily striated with white
64.	Fringe dark on unper half of wing or toward apex only hind wing of male
•	Fringe dark on upper half of wing or toward apex only; hind wing of male mostly white
64.	Fringe and hind wing darker
65.	Fringe and hind wing darker
	solid dark
65.	solid dark
	third; hind wing mouse gray in both sexes15. Epiblema obfuscana.
65.	third; hind wing mouse gray in both sexes15. Epiblema obfuscana. Median area with a white fascia in basal portion; hind wing dark brown.
	23. Epinotia heucherana.
66.	Fringe with a continuous black basal line
66.	Fringe usually powdery gray toward base, without black basal line
67.	Wings broader, ground deep brown; contrasting with the black line in the fringe
	fringe1. Epinotia nanana.
67.	Yellow brown with ochreous fasciae
67.	Wings narrower; ground gray, not contrasting with the line in the fringe68.
68.	Dark smoky gray14. Epinotia zandana.
68.	Light gray11. Epinotia laracana.
6 9.	With an oblique lead-colored line running toward anal angle, converging with a similar subterminal one
	a similar subterminal one16. Eucosma sombreana.
69.	No oblique lead-colored line
70.	Ground blue-gray under a lens, appearing usually as two paired bands and an odd one
	an odd one
70.	Ground not blue-gray
71.	Apical third of costa, and outer margin to anal angle contrasting, pale, with
71	an even, straight, dark-gray subterminal lineHystrichophora kokana. No even dark subterminal line
71.	No even dark subterminal line
12.	Brownish, flecked with little groups of white-tipped scales; expanse about 25 mm
70	25 mm
14.	Ash grey nearly overly mettled
72	Ash-gray, nearly evenly mottled
74	Ground flecked with black,
74	Ground nearly even pale gray to blackish12. Epinotia vertumnana.
	Ground nearly compare gray to machine in the spinotial bertaining

1. E. circulana Hübner. Fore wing light brown on basal half, light ochre on outer. With some small silver streaks and spots in middle of wing and near apex; a silver streak from costa at three-fourths of the way out to outer margin, curving in its outer part so as to leave a clear yellow semicircle above the speculum. Speculum of two horizontal rows of alternate black and silver spots, the upper row normally containing five or six spots and the lower reduced. Fringe powdery gray. 15 mm.

North Carolina and Louisiana to Florida. New York: Staten Island (?) This species presumably occurs northward, but northern records, so far as I have seen the specimens recently, are based on other species of the group. The type is said to have come from Pennsylvania.

2. E. scintillana Clemens. Whitish, heavily dusted with brown, on basal half; light yellow on outer half. Base of inner margin less powdery than the general surface. A streak without the powdering, on the cell from base to middle of

wing. A silver bar across cell near end, and some smaller silver spots, but without a distinct silver circle on outer part of wing. A silver subterminal line ending at middle of outer margin. Speculum with four vertical series of three black spots each; with three vertical silver bars, one before them, one between the two middle rows, and the other beyond; the region above the speculum heavily dusted with black, and often in longitudinal streaks. 16 mm.

June; August.

I have seen this form only from Cincinnati, Ohio, Illinois, and westward. It was originally described from Pennsylvania. I suspect this is the "circulana" of most eastern records.

3. E. dodecana Zeller. Dull powdery brown-gray, with only slight yellow streaks along the outer margin. The pale streak in the cell faint or absent. Markings as in E. scintillana. 18 mm.

June. This is very probably a variety of E. scintillana; E. randana, from the Rocky Mountains, appears to be a larger race.

New York to Texas.

Basal fourth powdery gray; second fourth deep 4. E. fratruelis Heinrich. ochreous, bisected (toward costa) by a vertical lead-gray fascia; the ochreous fascia beyond it very narrow on the inner margin; outer part powdery gray; a lead-gray median fascia from costa perpendicularly to top of speculum with a little yellow beyond it at costa; speculum of five rows of partly fused black dots (normally 2, 2, 3, 3, 2), the first and second separated by lead-colored bars, and likewise the third and fourth. Apex ochreous, bisected by a lead-colored fascia. Hind wing blackish. Head, body, and fringe powdery gray. July to September. Described from the types in the United States National

Museum.

Southern Pines, North Carolina; Georgia.

5. E. fraudabilis Heinrich. Ochre yellow or yellow-brown, including head and thorax. Complete antemedial and medial lead-colored fasciæ, the antemedial at a third way out; three silver dots beyond, the second outwardly oblique and running down along upper outer side of the very large speculum, which almost reaches the costa; a lead-colored fascia on outer margin. Lower half of speculum with six rows of black dots and three lead-colored bars; the upper part of the speculum with alternate cream-colored lines and series of plack scales. Hind wing paler than in E. fratruelis.

June and July.

Skyland, Virginia; Southern Pines, North Carolina.

6. E. adamantana Guenée. Fore wing silver white, marked with golden brown; fold in male brown; a brown fascia from middle of costa to outer part of inner margin, crossed by one from inner margin at a third way out, to apex, reaching the outer margin near the middle and running along it to the apex; dorsal half of outer margin narrowly brown-edged; base of inner margin brown. 18 mm. September.

Coastal marshes from New York to Florida.

7. E. quinquemaculana Robinson. Fore wing gray-brown; a silver-white spot at middle of base; a broad irregular transverse fascia at a third way out, twice as wide at costa as at inner margin; a rounded-trapezoidal patch on costa at twothirds way to apex, and a smaller spot before the apex; a rounded spot opposite the postmedial patch, close to inner margin, but not quite reaching either the margin or the costal spot. 18 mm.

June: August and September.

Lucaston, New Jersey, to central Missouri, Arkansas, Louisiana, and Florida. New York: Wells (New York State Museum); Bellmore, Long Island.

8. E. robinsonana Grote. Gray-brown; silver spot at base less distinct than in E. guinguemaculana. An antemedial fascia a fourth way out not quite reaching the costa; a similar, broader one just before the middle of the wing, rarely reaching

the costa, irregular on the outer side, and rarely widening toward the inner margin. The two outer spots on the costa much as in E. guinguemaculana, more nearly equal, squarer, and each containing a minute brown dot on the costa; spot on inner margin further out, reaching the outer margin below, and less evenly rounded. A minute white streak on middle of outer margin. 13-18 mm. (quintana Zeller).

June and August.

New Jersey to Kansas, Alabama, Louisiana, and Florida.

Var. tryonana Kearfott, from North Carolina, is a variant with each of the two outer costal patches almost divided into a pair of bars, and the anal patch divided into a group of four partly fused spots. The antemedial fascia is even in width and does not reach the costa. I have only seen the types, which were taken in May.

9. E. ridingsana Robinson. Varying from pale ochreous to pale brown; the markings finely edged with black. A silver streak through middle of wing from base to end of cell, ending in a point; a silver streak along middle third of costa, leaving costa at two-thirds and ending in a point between the end of the first streak and the apex; a small costal dot beyond it; and a larger triangular patch before apex; an irregular silver mark at anal angle, and inner margin silvery; a small silver triangle on costal half of outer margin. 20 mm.

A specimen from Illinois differs only in the silver costal streak extending practically to the base, and outwardly joining the two outer costal spots (Conchylis Robinson).

August; September.

Manitoba to Illinois, Louisiana, and west.

Besides the two following, there are several other well-marked forms of this

group, which may be new species, or more local strains of *ridingsana*. 1.5. E. argentifurcatana Grote. Usually considered a variety of *E. ridingsana*, but at least a constant and clearly defined form. Similar to *E. ridingsana*; median stripe deeply notched on lower side; basal end of costal stripe curving down and

A form of this group from Southern Pines, North Carolina, and Texas agrees with this, except for its much smaller size. (20 mm.) It has been taken from August to October.

Ontario.

11. E. hipeana Grote. Markings mostly as in ridingsana; costal streak divided into a bar on basal two-fifths of costa, fading out just before reaching base, and an oblique outer fascia starting at middle of costa, and running out obliquely across the end of the cell, like the outer part of the costal stripe of ridingsana. 37 mm. This is also a possible form of ridingsana.

Port Stanley, Ontario.

A dwarf variant of this form also occurs in Nantucket, Massachusetts, North Carolina, Illinois, and Texas; it flies in August and September.

12. E. mandana Kearfott. Dull brown, not powdery, lighter than E. fulminana. Costa faintly striate with blackish on basal two-thirds and with lead-gray and dirty white toward the apex; the last stria at the apex distinctly whitish, large, and a little contrasting. A lead-colored line from costa at three-fourths way out, to outer margin about at M_2 . Speculum somewhat paler, with two lead-colored bars, more nearly parallel than in *E. fulminana*; the outer bars broken, enclosing three black longitudinal streaks. Fringe powdery white on blackish. Hind wing mouse gray; the extreme apex striate. 18 mm.

June.

New Jersev to Texas.

13. E. nandana Kearfott. Fore wing broad; outer margin slightly concave, but not notched. Fuscous, regularly dotted with little groups of white-barred scales; base slightly darker, its outer boundary oblique inwardly to inner margin; a suggestion of oblique dark shades over end of cell, and opposite it at anal angle. Fringe concolorous; hind wing slightly paler. 30 mm.

September.

North Carolina to Manitoba.

14. E. fulminana Walsingham. Deep dull brown, practically the same shade as in E. sombreana and mandana; more or less, though usually only slightly, mottled in two shades; often with an area of narrowly white-tipped scales below and beyond the end of the cell, and some scattered white scales. Costa with geminate paler striæ, becoming distinctly whitish toward the apex; the last one nearly I mm. back from the apex. Speculum composed of two broad, vague, irregular, lead-gray bars, the outer one normally broken in two, with, as a rule, some black scales between them. Hind wing mouse gray. 27 mm.

Late July.

Illinois and Wisconsin to Texas.

15. E. rusticana Kearfott. Fore wing luteous on dorsal half and toward outer margin, including apex; shading into blackish on costa, except at the apex; the darker parts more or less streaked with black. Speculum a vague, paler, and graver area, with a couple of long black streaks. Fringe luteous and light brown. June: August.

North Carolina to Illinois and south.

16. E. sombreana Kearfott. Dull gray-brown, mottled faintly, and becoming a little warmer brown toward the outer margin; with geminate pale costal striæ; base slightly darker, with oblique outer boundary; a lead-colored line along outer margin, nearly meeting a shorter one at anal angle; with some black scales in the space between, but no definite speculum. Fringe powdery. 16-22 mm.

July and August. Larva in Helianthus.

New York to Manitoba, North Carolina, and Arkansas. New York: Ithaca, Brooklyn.

17. E. albiguttana Zeller. Pale ochreous, with scattered silvery gray spots of a few scales each; costa obliquely striate with whitish, the outer striæ lightly edged with black; speculum of two vertical silver bars, connected by three con-trasting fine black streaks. Base of fringe dusted, black and white. Hind wing very pale grav. 12 mm. (ochraceana Fernald).

June and August.

New Hampshire to Virginia, Texas, and Kansas. 18. E. graciliana Kearfott. Doubtfully distinct from *E. albiguttana*; ground brighter ochre yellow; silver spots clean-cut and sharply gray-edged, as well as the striæ on the costa; one stria from the costa reaching practically to the outer margin, curving around the upper side of the speculum at a little distance. Hind wing darker gray.

July to September.

North Carolina.

19. E. cataclystiana Walker. M_3 and Cu_1 of fore wing becoming coincident at margin. Fore wing dull ochre, somewhat mottled, or more rarely, yellow-brown; costal striæ pale, somewhat silvery, distinctly defined with dark, the last one running along the outer margin, which is drawn out and subfalcate, about as in the lowest species of Ancylis. First silver bar of speculum slightly oblique, preceded by a brown shade and followed by two strong black dots; outer bar short, marginal. Base of fringe powdered, black and white. 16 mm.

Aberrant specimens have a brown ground color and the area above the speculum crisply dusted with white. In western specimens, the anterior bar of the speculum is more oblique.

August. Larva in Ambrosia.

Maine and Trenton, Ontario, to New Jersey, Colorado, and Utah. New York: Pern, Ithaea; Poughkeepsie; Clove Valley, Staten Island.

20. E. pergandeana Fernald. Pearl white, with pale gray striation, mostly longitudinal, but oblique on the costa except toward the apex, where the striæ converge toward the top of the speculum as usual, and are finely defined with black. Speculum of two vertical rows of two or three black points, with faint vertical silver bars before, between, and beyond them. Base of fringe dusted with black. Hind wing concolorous. 16 mm.

June and July.

Massachusetts to North Carolina, west to Colorado. New York: Pern; Lynbrook, Long Island, Gay Island (Long Island Sound). The paler race flavana Kearfott, occurs in Arizona.

201/2. E. comatulana Zeller. Buff-brown, coarsely flecked with clay-color; costoapical portion of wing vertically barred, and speculum filled with the clay-color. Lines mostly obscure; a strong lead-colored line from beyond middle of costa to below apex, separating the vertically barred region of the wing from the speculum. Speculum with three black bars, partly broken into dots, but not crossed by a leadcolored bar, the speculum with a vertical lead-colored bar on inner side and an oblique one on outer, 15-18 mm.

May and August.

Woods Hole, Massachusetts. to southern California; rare in the East.

21. E. pallidipalpana Kearfott. Light brown, more or less mixed with white; with brown and white costal striæ; and several round, pale yellow patches on basal two-thirds, two or three of them at the base fused into a larger irregular patch; all edged with white; speculum a silvery patch, divided by an irregular broken black line. Line in fringe nearly continuous. 10 mm.

July.

Connecticut to North Carolina, west to Iowa.

22. E. bipunctella Walker. Bright yellow; a brown dot at end of cell; fringe, abdomen, and hind wing brown. The yellow portion occasionally is suffused with pale dull brown except along the costa toward the base. Probably the largest and heaviest of our Tortricidæ. 35 mm. (worthingtoniana Fernald).

June and July. Larva in roots of Silphium.

Chicago, Illinois; Kansas and vicinity.

23. E. juncticiliana Walsingham. Light dull gray-brown; a nearly straight line from middle of costa to inner margin just before the anal angle; the ground before it much darker than beyond; sometimes with a blackish dot on this line at end of cell. Outer part with a large vague triangular dark shade resting on the outer margin. No definite line in fringe. 15 mm. July and August. Larva boring in goldenrod.

Maine to Florida and California. New York: Otto, Ithaca.

24. E. dorsisignatana Clemens. Dark ash-gray, with a very slight violet gloss. Markings dark brown, even-edged, and finely outlined with a narrow pale line. Dorsal antemedial patch strongly contrasting, half-elliptical, and reaching up to the cell. Postmedial patch contrasting and sharply defined below, but vague on the inner side at costa, elliptical; almost reaching the inner margin where it sometimes joins a brown dot on the inner margin just before the anal angle. A faint brown fascia from costa to outer margin above anal angle, and a slight brown apical dot. No speculum. Line in fringe faintly darker. 12-20 mm.

September. Larva in roots of goldenrod.

New Hampshire to British Columbia and Texas. New York: Essex County, Dansville, New Windsor, Pine Island, New York City, Staten Island.

Variety diffusana Kearfott is on the average smaller and darker, with the antemedial dark patch alone contrasting, narrow, about twice as high as wide, oblique, and in many cases concave on the outer side.

Massachusetts to Illinois, North Carolina, and Tennessee. New York: Coram, Long Island.

E. similana Clemens is evidently locally constant and may be a good species. Antennedial and medial patches are fused into a thick L-shaped mark, which is sharply defined all around, except on the inner side at the costa. The small spot on inner margin before the anal seems to be always absent. (confluana Kearfott).

August. This form or species occurs with the type in New Jersey, but is apparently earlier.

Massachusetts to District of Columbia, Illinois, and Missouri. New York: Big Indian Valley.

25. E. engelana Kearfott appears to be a distinct species of this group, but the type is so rubbed as to be almost unrecognizable. Apparently it is rather narrow-winged; dull gray-brown, with a dark antemedial spot on the inner margin as the most contrasting mark. It is likely to prove a synonym of *confluana* Kearfott. 15 mm.

August.

Pittsburg, Pennsylvania.

The female "type" from New Jersey is another species.

26. E. graduatana Walsingham (?). Fore wing dull gray; wings broader than in *E. dorsisignatana*; an antemedial half-oval spot resting on inner margin, and a large postmedial ellipse, reaching the whole width of the wing, from costa twothirds way out to near anal angle. Outer margin dark brown. All the markings darker and finely defined with pale, as in *E. dorsisignatana*. Hind wing orange; abdomen brown.

May and June. The form here described is quite distinct from typical graduatana from Texas, and may be a new species.

Vermont to Manitoba.

27. E. eumæa Meyrick. Umber brown, more or less scaled with pale fuscous; the type with the basal half apparently largely pale fuscous, leaving a patch of brown in the fold. Costa with geminate whitish striations, becoming silvery below the costal edge; one of the striæ extending obliquely out almost to the outer margin, about at \mathbf{M}_{i} , then sharply bent and running rather parallel to outer margin, to the inner margin; forming the outer line of the speculum. Inner line of speculum also narrow, silvery, and contrasting, the space between the two lines of the paler fuscous, crossed by strong black streaks. Fringe powdered; hind wing dark gray. 16 mm. (wandana Kearfott, not vandana Kearfott).

July. Type only seen, now in bad condition.

Cincinnati, Ohio.

28. E. womonana Kearfott. Black-brown, a little duller than E. wandana; vaguely mottled with areas of lighter fuscous dusting, the most definite one on irregular antemedial fascia, distinctly traceable only at the inner margin. Costa with paired striæ outwardly; speculum of three thick lead-gray patches, enclosing a narrow inverted Y-shaped portion of the ground, on which lie a couple of black spots. Base of fringe finely powdered with white. 15 mm. (wandana Kearfott, in part).

Maryland to Ohio and Texas.

28-1. E. palabundana Heinrich. Fuscous, powdery, and coarsely striate. Base dark, with some pale striæ, its outer boundary excurved, but strongly retracted in the cell below the dark costal fold. Antemedial area broadly pale, typically contrasting. A narrow, somewhat irregular and excurved, dark postmedial fascia, its upper part extending in obliquely to costa before the middle, its lower part lying along the speculum. Speculum with three broad, dull, lead-gray bars, leaving a small fuscous area between them, and perceptibly pinkish in some lights. Fringe powdery. 15 mm.

July and August. Described from an atypical specimen from Ithaca, probably representing a good race.

New York to Manitoba. New York: Ithaca.

28-2. E. fiskeana Kearfott. Ash gray, marked with darker gray-brown, somewhat mottled. Antemedial a broad dark shade resting on inner margin, defined on outer side; postmedial oblique out and excurved, preceded by contrasting brown shades over cell, and in and below fold. A blackish, diffuse, subapical patch. Speculum of three very broad, dull, lead-colored patches, with some black and luteous scaling between them. Fringe powdery gray, preceded by a white line above, and a black line opposite speculum. 20-29 mm.

Virginia and Illinois to South Carolina.

29. E. grotiana Kearfott. Pure white; head yellowish; shoulders and disc of thorax blackish. Markings of fore wing dark brown; a brown band along the costal edge, from the base to the apex, heavier on the costal fold of the male. Antemedial fascia running obliquely up from the basal angle across the fold, very irregular and broken, and enclosing more or less white. Some dark brown striæ toward base. Postmedial fascia oblique from middle of costa to upper angle of the speculum, then widening to inner margin; with its outer boundary erect and inner boundary oblique inward. Speculum slightly shining, with three black points in its upper part. Middle of outer margin brown, with two pairs of white striæ. Fringe dark, its base powdery. Hind wing gray. 15 mm.

July.

Chicago, Illinois, and west.

30. E. gomonana Kearfott. Costal fold long. Fore wing dark gray-brown, in some lights with yellow-brown iridescence; with broad, partly confluent, light blue-gray, somewhat shining bands. A narrow basal band; antemedial fascia excurved, broad, and defining a darker base, partly confluent with the smaller. more erect medial one; postmedial band narrower, outwardly oblique, almost straight, leaving a high rectangular area of the ground before it on the dorsal half; a short gray bar across the apex, and a similar oblique bar at the middle of the outer margin; speculum indicated by two broad gray bands, sometimes with some black scales between them. 9 mm.

May.

Northern New Jersey to Virginia.

31. E. zomonana Kearfott. Light gray, slightly bluish, dusted with blackish. A blackish antemedial bar on dorsal half, more distinct than in E. gomonana, with a well-defined, sinuous outer boundary, and more diffuse inner and costal boundaries. Speculum paler than that of E. gomonana, composed of a group of three thick silver-gray bars, separated by a Y-shaped blackish powdery mark; with heavy blackish shades before and above it, the latter reaching half way to the costa. Rest of wing with the black dusting gathering into vague dark gray striations, leaving the apex blackish. 10-15 mm.

August. Very closely related to E. gomonana.

Pennsylvania to Illinois, Missouri. and Tennessec.

32. E. giganteana Riley. White, with a little gray toward base of inner margin and along costal edge. A large gray patch on inner margin from just beyond middle to apex of wing; the more basal portion of the patch with thick raised lead-colored bars; speculum finely dotted with black on a mixture of light brown and white, with a region above it of mixed powdery gray and white; above this a triangular region, extending up to the apex, of mixed gray and brown; fringe powdery. Hind wing light fuscous, with white fringe. 25-35 mm.

June to August. Larva in roots of Silphium.

North Carolina to Illinois and Missouri.

33. E. monitorana Heinrich. Deep brown, with a blood-red overcast. Head dirty white, fuscous behind antenna; palpi fuscous. Fore wing with basal fourth of the ground color, scaled and shaded with lead gray, its outer boundary waved and farther out on inner margin; second fourth of wing broadly pale, mixed

silver and pale gray; median fascia of ground color, twice as wide at inner margin as at costa, normally enclosing a lead gray spot on inner margin, and interrupted on lower half of cell. Subterminal region pale, obliquely out from two-thirds way out on costa to anal angle, followed by a red-brown area with some whitish barring, especially on outer margin. Fringe with a weak dark basal line. Hind wing dark with a pale fringe. 15 mm.

Larva on pine, boring in the cones; moth emerging in May. Pennsylvania; Virginia.

E. cocana Kearfott, described from North Carolina, can apparently be distinguished by the darker brownish yellow head, heavily scaled with gray; and darker median fascia.

34. E. tocullionana Heinrich. Smoky violet-gray. Head ochreous, light in front: thorax brown in front. Ground of fore wing striate with black and shaded with brown; markings deep ochrc or yellow-brown, broadly edged with dark silver, and broken into double striæ at costa; antemedial band of even width, oblique out from a third way out on costa to inner margin at two-fifths, narrowed or interrupted on fold, and tending to form two rounded spots. Postmedial band diffuse on costal half, on dorsal half with a nearly round silver and ochre spot representing the speculum, without any black scales. Fringe dark, barred with white, and with a black-and-white checkered basal line, hind wing dark. 15-18 mm.

Larva on Picea, boring in the cones; the moth emerging in May.

Connecticut; Pennsylvania.

35. E. bilineana Kearfott. Fore wing gray, powdery, and varying strongly in shade, dark specimens tending to be strigose with blackish and white. Base of Cu white, with a strong black shade below it; a finer black streak running up to apex; and often a diffuse streak in outer half of cell, with some whitish above it. A series of dark terminal dots. Fringe concolorous. 25 mm.

July.

Hessville, Indiana, to Manitoba and Colorado.

14½. HENDECANEURA Walsingham

(Enarmonia, in part)

Similar to Eucosma. Sc of fore wing completely absent in male; \mathbf{R}_1 arising from cell close to base, easily mistaken for \hat{Sc} ; female with Sc and R_1 normal. M₂, M₃, and Cu₁ slightly approximate at margin, Cu₂ straight. Costal fold short. Hind wing normal, with R and M₁ stalked.

Male genitalia similar to Zeiraphera, but with the neck of the valve longer.

1. H. shawiana Kearfott. Deep brown, with some ochreous spotting; a more or less distinct white antemedial stria on inner margin; three irregular and more or less confluent striæ at middle of inner margin, continued above as a bluish metallic double fascia which reaches costa, ending in a few white scales on costal edge. Outer part of wing largely ochreous, but with a blackish subterminal shade between the two lead-colored lines above the speculum (opposite the notch). Speculum yellow with broken black bars. Costal edge outwardly barred black and white. 13 mm.

The larva is to be expected on the cone-scales of some conifer. June.

Hampton, New Hampshire; Newark, New Jersey.

Kearfott treats as a variety of this a form from Essex County, New Jersey, which is smaller (9 mm.), without decided ochreous areas, with the dorsal white spot less distinctly broken up into striæ, and the speculum quite obscure on account of the lack of yellow. Early July.

15. THIODIA Hübner

(Pædisca, in part; Ioplocama Clemens; Semasia, Hedya, Cydia, Epinotia, Enarmonia, in part)

Similar to Eucosma; \mathbf{R}_2 generally nearer \mathbf{R}_1 than \mathbf{R}_3 ; costal fold absent; hind wing frequently with M_3 and Cu_1 completely united.

Valve not strong, and shorter than in Eucosma; anal indentation usually with a pronounced cluster of hairlike spines.

The venation varies, but there is always at least some slight trace of the emargination of the outer margin, and \mathbf{R}_4 and \mathbf{R}_5 are normally separate. The genus is perhaps heterogeneous, being derived from more than one source in Eucosma and possibly also from Epiblema. The usual food, as in those two genera, is the Compositæ.

Key to the species

1. Speculum of four rows of two black dots each.

2. Area above the speculum (enclosed in a broken lead-colored circle) wholly dusted with brown.....l. annetteana.

2. Less than half the area above the speculum dusted with brown..2. refusana. 1. Speculum less complex.

2. With a rounded central dark patch on fore wing, defined below.

3. Base of fore wing pale.

4. Ground dull pale gray, with some darker dusting and brownish shading.

6. Exentera virginiana.

- - 4. Base of inner margin below fold contrasting pale gray, sharply sepa-
 - 4. Base of inner margin concolorous, or gradually shading into the light red-brown costal portion......3. Epinotia septemberana.
- 2. No such patch.

3. With a contrasting longitudinal white stripe in cell.

4. Basal half of costa solidly white......13. striatana.

4. Basal half of costa finely striate with brown......14. clavana.

3. Costal third broadly white and more or less contrasting.....15. kiscana. 3. No longitudinal white stripes.

- - 4. Inner margin contrastingly pale, the pale area sharply delimited in
 - followed with blackish.
 - 5. Fascia broad and complete.

6. Followed by a blackish fascia, the ground outwardly pale.

1. Gypsonoma fasciolana.

- 6. The whole outer half of wing dark....2. Gypsonoma substitutionis. 5. Fascia confined to dorsal half of wing, and sometimes narrow and diffuse on outer side.
 - 6. Apex of fore wing with bright yellow-brown overlay or iridescence.

5. marmontana.

- 6. Apex blackish, concolorons with other dark portions of the wing. 7. Median dorsal white patch small and well defined, separated from patch at anal angle by more than its width.....3. parmatana.
 - 7. Dorsal white patch, larger and more diffuse, separated from patch at anal angle by less than its width......4. alterana.

- 4. Antemedial region not white; ground, if white, heavily dusted with blackish, appearing gray.
 - 5. Dark, with more or less contrastingly pale speculum.
 - 6. Ground red-brown, extending along costa to apex....18. raracana.6. Ground black-brown.
 - 7. Apical region contrasting, yellow.....16. ochroterminana.
 - 7. Speculum only with deep yellow filling, not conspicuous.

17. perfuscana.

- 5. Without contrasting pale speculum; or, if the speculum is somewhat paler, ground either powdery gray, or strigose.6. Wing longitudinally striate with paler and darker, or tawny brown,
 - 3. Wing longitudinally striate with paler and darker, or tawny brown, and becoming darker toward apex; or moth large with almost evenly dull brown ground.

 - 7. Speculum less distinct or absent; sometimes with red-brown streaks.
 - 8. Males (with single frenulum).
 - 9. Tawny brown, shading into chocolate brown..7. ferruginana. 9. Ground luteous.
 - 10. Speculum obsolete, the region crossed by the uninterrupted luteous and brown striation of the wing.

12. radiatana.

- 10. Ground darkening and becoming solid brown toward outer margin; the outer silvery bar continuous, and lying in the solid brown portion, or interrupting the striations.
 - 11. Ground outwardly yellow-brown, streaked and shaded with chocolate brown.....9. umbristriana.
 - 11. Ground outwardly bright rusty orange, streaked with rusty brown8. roseoterminana.
- 10. Speculum with well-marked basal and outer lead-colored lines, and preceded and followed by distinct blackish areas.
 - 11. Blackish area at anal angle clean-cut above, stopping about at Cu₁.....10. essecana.
 - 11. Blackish area extending up to middle of wing and there joining the dark area above the speculum.

11. avcemeana.

8. Females (frenulum multiple).

9. Speculum indicated by lead-gray bars before and beyond it, usually distinctly paler and preceded by a dark shade.

10. essexana, 11. awemeana.

- 9. Basal lead-colored line of speculum absent, its ground concolorous.
 - 10. Ground even-colored, a little deeper at anal angle only. 12. radiatana.
 - 10. Ground of outer half decidedly deeper than base.
 - 11. Fringe contrastingly powdered with blackish and white. 9. umbristriana (?)
 - 11. Fringe concolorous with ground of wing.

8. roseoterminana, 7. ferruginana.

6. Fore wing heavily marked with green............ (Proteoteras).6. Fore wing pale yellowish, shaded and dusted with gray or brown.

7. Ground pale lemon yellow.

8. Costa appreciably barred with fuscous only on outer half. 21. olivaceana.

19. tomonana.

6. Ground gray, powdery; or dark fuscous, without brown or yellow markings; expanse over 10 mm.

7. Two prominent, obliquely placed raised tufts in fold.

6. Gretchina bolliana. 7. At most with a small continuous scale-ridge in fold.

9. A clearly marked, curved, angular, black marking above speculum, shaped like a bird's beak.

5. Gretchina deludana.

9. No such markings; with several blackish shades when there is any decided black marking above speculum.

10. Fore wing shaded with brown, at least in certain lights. 11. Smaller (14 mm.); hind wing white at base.

3. Gretchina watchungana.

11. Normally larger (18 mm.); hind wing all light fuscous. Exentera species.

10. Fore wing dark gray or blackish, heavily dusted with white, and without brown scaling or iridescence.

11. Face contrasting blackish; posterior tufts blackish. 1. Gretchina amatana.

11. Face concolorous or pale.

- 12. Thorax whitish, contrastingly but diffusely spotted with dark gray; fore wing powdery looking to naked eye; face whitish...5. Epinotia signatana.
- 12. Thorax light, white-mottled; face darker, fore wing light, powdery-looking, and with an indication of the Laspeyresia lunule at the middle of the inner margin (Laspeyresia garacana).

1. T. annetteana Kearfott. Basal half of fore wing dull ochre, shaded with light brown; outer half powdered with light brown and white, with some ochre and leadgray streaks extending down from white points on the costa; the boundary between the base and outer part also marked by a lead-colored fascia. Outer half of wing nearly covered by a large circle, brokenly outlined with lead-gray; its upper half filled with the white-dusted ground, the lower with four rows of black spots; each row composed of two large spots, and often a third spot below them; with a short, lead-gray bar between the second and third rows. Base of fringe also dusted with white. Hind wing light brown. 15 mm.

April.

Rhode Island; Ohio; Texas.

For points of distinction from the similar species of Eucosma, see the key to Eucosma.

2. T. refusana Walker. Light ochre-brown, shaded with gray; with pale streaks along costa, on Cu, and in cell. Outer part of wing with a large lead-gray broken circle, as in *annetteana*, and with some lead-colored costal striæ on the ochre ground. Upper half of circle with an area of brown, white-tipped scales, but only half filled with them; lower half with four rows of black spots, each row con-

sisting of two or three spots; and a lead-gray bar between the second and third rows. 15 mm.

April and May; September. Perhaps not rare, but confused with members of the circulana group.

Hudson Bay district to Virginia, west to Manitoba. New York: East New York, Long Island.

3. T. parmatana Clemens. Dull blackish, sometimes with some scattered whitish scales and with obscure black marks. Costa with some paired white striæ on outer half. Middle of inner margin with a white patch or a group of strong striæ, the most basal being strongest and defining the dark basal area. Speculum mixed white and silver, with a couple of black dots, its inner boundary erect: much straighter than usual. Base of fringe dusted with black and white. 10 mm. (crispana Kearfott etc., not Clemens).

June to September.

New Jersey to Missouri and probably generally distributed. "New York" (Edwards).

4. T. alterana Heinrich. Similar to T. parmatana. Ground somewhat paler fuscous; white markings more extensive, the median patch frequently diffuse on upper and outer side, its inner side and the speculum sharply defined. Apical region concolorous fuscous, cut, as usual, by fine paired white striations. 9-12 mm. August.

Massachusetts to Maryland. New York: Ithaca. 5. T. marmontana Heinrich. Similar to T. parmatana. Fore wing with ground perhaps slightly richer brown, the costal edge toward apex and upper part of outer margin overlaid with shining yellow-brown in favorable light, interrupted by the paired white striæ, which are defined with black.

July and August.

Manitoba. New York: Ithaca.

6. T. formosana Clemens. Male clay-color, striate or heavily shaded with brown; costa obliquely streaked with silver on outer two-fifths, the last streak parallel to outer margin and at a sharp angle to the others. Speculum well marked, vellow, almost surrounded with silver, and with three black streaks. Basal line in fringe weak. Hind wing mouse gray with outer part of fringe white. Female with the same markings, but almost wholly suffused with brown; with slight or no pale striation. Costal silver streaks weaker, and fringe more tinted with brown than in male. 20 mm.

End of May to June; locally not rare. This species is closely related to the type of Thiodia (aspidiscana).

Quebec to New Jersey. New York: Rock City (Cattaraugus County), Ithaca, McLean, Karner, Ramapo, Watchogue.

7. T. ferruginana Fernald. Sexes similar. Rusty brown, shading into chocolate brown outwardly; fringe concolorous with outer portion of wing. Speculum represented only by a couple of streaks of rather more shining scales. Hind wing chocolate brown, with paler fringe. 13-15 mm.

May and June.

The male of this species is easily recognized, but the female cannot be certainly distinguished from T. roseoterminana; it is usually smaller and brighter than the latter.

Maine to New Jersey. New York: Black Brook (Clinton County), Rock City (Cattaraugus County). Ithaca, McLean, Staten Island.

8. T. roseoterminana Kearfott. Male clay-color, lightly and irregularly streaked with light brown; the strongest streak below the cell. Outer part of wing shading into rusty orange, streaked with rust-brown. A continuous chocolate-brown ter-minal band. not contrasting, preceded below by a slight lead-gray streak. Fringe concolorous with outer part, or a little paler. Female with basal half rusty orange shading into chocolate brown outwardly, with the same marginal markings as the male; fringe yellow-brown. Hind wing dark mouse-gray, with pale fringe. 18 mm.

Late May. This species of this group are hopelessly confused in all collections; especially the females.

Certainly known only from Cincinnati, Ohio.

9. T. umbristriana Kearfott. Closely similar to T. roseoterminana, but a little duller and darker in both sexes. Base of fringe dusted with black and white in both sexes. 16–18 mm.

May and June.

I believe Kearfott had the females of these two species exchanged, to judge by the cotypes before me.

Western Pennsylvania to Manitoba.

10. T. essexana Kearfott. Male clay-color, streaked with brown, the streaks emphasized and partly fused in such a way as to form a dark streak from base to outer margin above speculum. roundly notched by the pale speculum. Outer two-fifths of costa with an irregular pale-outlined patch, containing three wellmarked darker pale-outlined costal spots. A heavy brown-black terminal line and a patch at anal angle before speculum, extending up to Cu_i but not joining the central shade. Speculum almost completely outlined with silver, and with some brown streaks. The female will probably show similar markings, but not contrasting, on a deep brown ground. 20 mm. (*radiatana* Kearfott, 1903, not Stephens).

The larva bores in stems of aster, hibernating as a larva and emerging in May. New Jersey.

11. T. awemeana Kearfott. Similar; the longitudinal streak stopping short before the middle of wing; the shade before speculum not decidedly blackish. extending up, though paler, to costa, before the peculiar outer costal mark, and joining a dark shade between the speculum and the costal mark. The latter shade, as seen under a lens, is formed of a couple of dark streaks. Speculum fairly well defined. but crossed by brown streaks. Terminal band brown, not blackish as in most specimens of the preceding form. Female chocolate brown, decidedly strigose with yellow-brown; base not much paler, but with the shades much as in the male, less contrasting, more diffuse, and often connecting to form a dark cross. 18 mm.

May and June.

New Hampshire; New Jersey; Manitoba. New York: Crosby (Yates County), McLean, Ithaca.

12. T. radiatana Stephens. Male elay-color, strongly striate with brown, forming a more or less marked darker shade from base to middle of outer margin. Speculum obsolete, marked by a few shining scales. Dark streaks heavy and tending to fuse on outer margin, and especially at anal angle. Fringe also dark gray. Female deep yellow-brown, with similar markings in black-brown. 20 mm.

End of May to June; not rare, being probably the commonest species of this series.

Generally distributed. New York: North Elba, McLean, Big Indian Valley, Ramapo, Bronxville, Oyster Bay.

13. T. striatana Clemens. Light gray; costa white from base to apex, only the outer half cut by light gray lines. A white streak shaded below with blackish, covering the lower edge of the cell to its end; a more or less distinct blackish streak on Cu_2 , shaded above with pale. Speculum with a white spot, edged above and below with black streaks. Fringe powdery gray. 15 mm.

End of May.

Common and generally distributed in grassland; flying up by day, about like the similarly marked species of Crambus. New York: Peru. Oswego, Portage, Crosby (Yates County), Karner, Nassau, Ramapo, New Windsor, Katonah.

14. T. clavana Fernald. Fore wing light powdery blue-gray, the basal half light brown except on the inner margin; costal edge white, more narrowly so toward the base than in T. striatana, and cut by brown bars its whole length, those on the basal half finer and shorter than those outward. A white band along lower edge of cell, outwardly broken into streaks extending to the speculum, and with a black dash below it ending abruptly at the middle of the wing. Speculum pale yellow, preceded by a fine black line and containing a couple of dots. Apex drawn out as in T. striatana. Hind wing brownish gray; fringes pale. 15 mm. (I have seen this species labelled tenuiana Walsingham).

Hampton, New Hampshire; Truro, Massachusetts. "New York" (Heinrich).

15. T. kiscana Kearfott. Nearly even, pale gray. Wing form as in T. striatana Fore wing with costa broadly but a little diffusely white; outwardly strigose with luteous and blackish; the white extending halfway across the cell on the basal half, but narrower outwardly. Inner margin more or less shaded with whitish; speculum white or pale yellow, preceded by a broken black line and containing two black bars. Fringe powdery. Hind wing with dark veins. 12 mm.

End of May and June.

Cincinnati, Ohio; New Jersey; District of Columbia.

16. T. ochroterminana Kearfott. Dull black; roughly scaled; outer fifth, including fringe, contrasting ochre yellow, somewhat mottled, preceded by a yellowbrown streak on dorsal half of wing. 10-15 mm.

August and September.

Kennebunkport, Maine, and Maryland to Manitoba and Illinois. New York:

Renneounkport, Maine, and Maryana of Maryana to Markova and Linear Potter Swamp (Yates County), Ithaca. 17. T. perfuscana Heinrich. Deep brown, mottled, made up of black-brown, chocolate brown, and some flecks of paler gray-brown; speculum lighter, deep ochre, flanked by two vertical lead-gray bars, and containing three black lines. Antemedial band broad, strongly excurved, formed of a gathering of the fuscous flecking; postmedial traceable at costa only. 12 mm.

August.

New York and western Pennsylvania. New York: Ithaca.

18. T. raracana Kearfott. Red-brown or, more rarely, chocolate brown, shading into light red-brown toward base, especially on costa. Costa with light graywhite striæ. Speculum contrasting pale yellow, with broad silvery bars and some broken black markings. Fringe powdery gray at apex, light yellow-brown below. 12 mm.

This form can be distinguished from the similarly marked Eucosmæ by its brown or reddish, rather than fuscous, color. In pattern it is very near T. formosana. It has been taken in August, and, southward, in May also.

New Jersey and western Pennsylvania to Texas and Florida.

19. T. tomonana Kearfott. Pale gray, somewhat strigose with duller light gray; the speculum represented by some black dots. Two contrasting blackish spots on inner margin, the basal one half-crescentic, the outer half-elliptical or more irregular, extending less than a third way across wing, but broad. 12 mm.

August and September.

St. Johns, Quebec; New Jersey; western Pennsylvania. New York: Ithaca.

20. T. imbridana Fernald. Ground pale yellow, overlaid, except toward base of costa, with rusty ochre; mottled irregularly with dark brown, with a strong purple iridescence in some lights; the purple-brown gathering to form a band from the middle of costa to inner margin before speculum, and another from this band where it crosses the cell to inner margin at a third way out, both sometimes broken up or obscured by brown suffusion. Outer part of costa also showing the yellow ground, and striate with brown. Speculum lutcous with three broken black bars. Hind wing light gray.

General in distribution; flying in August.

21. T. olivaceana Riley. Light lemon yellow, with a slight green tint, marked with light chocolate brown, so overlaid with yellow as to appear gray; the markings are largely longitudinal streaks, the principal ones being a broad one along lower side of cell to middle of wing, one below basal third of costa, a narrow and cleancut oblique one from middle of costa to speculum, a small spot on inner margin a quarter way out from base, and a large patch at anal angle; besides some small streaks on costa and a vague dark shade through outer part of wing. Speculum yellow, with some black dots. Fringe concolorous.

June to August.

New Hampshire to District of Columbia and Illinois. New York: Rhinebeck. 22. T. verniochrana Heinrich. Similar to T. olivaceana; palpus with a fuscous spot on outer side of second joint. Fuscous bars on costa of fore wing distinct from base to apex. 10-13 mm.

August.

Massachusetts; New Jersey.

23. T. tarandana Möschler. Light gray, powdered on a white base, with clear white streaks, and with the pattern more or less defined with silvery white. Antemedial band strongly oblique outward from beyond basal angle to middle of cell; light gray-brown, shaded with blackish on the fold; postmedial irregularly excurved and widening toward inner margin, running from middle of costa to outer third of inner margin, where it connects more or less distinctly with a similar narrow and irregular terminal band, three fuscous spots on costa outwardly, defined by white striæ. Speculum enclosed by the irregular silver lines defining the postmedial and terminal bands, filled with slightly yellowish white, mixed with gray, and with a few black scales. Fringe white with a gray center line. Hind wing darker gray. 25 mm.

August.

Manitoba, Labrador.

16. SPILONOTA Stephens

(*Tmetocera* Lederer; *Eucosma*, in part, Walsingham and Durrant)

Male without costal fold. Palpi beaklike, triangular, with third joint concealed in scales. Thorax smooth. Fore wing practically smooth-scaled, with outer margin concave at middle and M_3 and Cu_1 somewhat approximated toward the margin. Antennæ with notch more widely open than in Strepsicrates and nearer base; the segments concerned in it almost completely fused and difficult to count.

Valve very long and narrow; cucullus with a large apical spine. Uncus absent; socii short and broadish; gnathos weak.

A development of Thiodia.

1. S. oceilana Schiffermüller. Dull grayish brown, somewhat mottled; the median area typically contrasting dirty white, in variety lariciana Heinemann hardly paler. Basal area with outer boundary roundly and moderately angled. Outer margin shaded with gray-brown near apex; below, with about three black bars, followed by a gray streak; a black spot before anal angle, made up of partly fused dots, with a well-marked gray shade between it and the three black bars. Dark line in base of fringe grayish and not contrasting. 12-15 mm. (pyrifoliana Clemens).

Often injurious, the larva webbing together the opening buds of apple in early spring. Moth in June to early August.

General in distribution; also in Europe. Possibly introduced in this country. New York: Common and general.

WILLIAM T. M. FORBES

17. STREPSICRATES Meyrick

(*Phthinolophus* Dyar, *Eucosma*, in part, Walsingham and Durrant)

Palpi rather clavate, scaled, with prominent short porrect third joint. Thorax smooth. Fore wing with outer margin rounded, the cell with a heavy scale-tuft in fold at one-third way out. Costal fold of male very large. Hind wing with \mathbf{M}_2 markedly separate from \mathbf{M}_3 and \mathbf{Cu}_1 which are stalked, \mathbf{R} and \mathbf{M}_1 approximate. Male antenna with basal segments of shaft more or less finsed and distorted, about the sixth to tenth joints forming a notch, partly covered by a scale tuft.

Male genitalia similar to those of Spilonota. Pollex present: anal angle spinulose: cucullus with several marginal spines. Socii small and slender.

1. S. indentanus Dyar. Normally with costal two-thirds of wing brown, quite variable in shade, and the dor al third paler gray, often contrastingly pale, especially in males; the boundary wavy and more or less defined with black. Hind wing mouse gray. 12 mm.

July. Larva a leaf roller on Myrica. The Florida species on guava appears to be distinct.

Canada to Florida. "New York" (American Museum of Natural History).

18. SONLA Heinrich

(*Eucosma*, in part)

Fore wing with costal fold present in male, \mathbf{R}_i and \mathbf{R}_5 completely united; outer margin concave, with \mathbf{M}_2 to \mathbf{Cu}_1 approximated; \mathbf{Cu}_2 straight; \mathbf{R}_2 arising near \mathbf{R}_1 from the normal accessory cell; \mathbf{R}_1 from before middle of cell. Hind wing with \mathbf{R} and \mathbf{M}_1 often anastomosing and sometimes stalked.

Male genitalia similar to those of Suleima; clasper rudimentary; socii short and broad.

The single species is almost identical in pattern with *E. perplexana* Fernald, from the Gulf strip, but the latter has \mathbf{R}_{4} and \mathbf{R}_{5} separate.

1. S. constrictana Zeller. Ground light brown-gray; when fresh, with distinct violet iridescence; markings deep brown. Ease deep brown: antemedial line perpendicular to inner margin, except toward the costa where it often fades out; straight or slightly excurved below cell; an irregularly quadrangular dark brown patch on inner margin toward anal angle, with its upper boundary oblique and parallel to the lower boundary of a similar patch over end of cell; typically with the two patches fused, and the point of fusion marked by a strong notch on the anterior side of the combined patch, at the fold; postmedial patch on cell sharply defined on basal and lower sides, but above shading into a brown outer costal shade, which surrounds the upper sides of the speculum; costa with pale paired striæ outwardly; speculum blue-gray with some brown in cent r. Fringe powdery, 12-15 mm.

June to August.

Northern New Jersey to Florida, west to South Dakota and Texas.

19. SULEIMA Heinrich

(Thiodia, Nemasia, in part)

Similar to Sonia. \mathbf{R}_s arising from middle of cell or slightly beyond; costal fold absent; notch on outer margin variable. Hind wing with **R** and **M**₁ always stalked, **M**_s lost. Clasper absent.

1. S. helianthana Riley. Whitish gray. Fore wing with two quadrate blackish patches on inner margin, and a black dash and white triangle at the apex. Base

somewhat darker, with a paler space between it and the first blackish patch; speculum white, containing a black dot, and on its costal border two longitudinal black lines, and partly defined with brown. Fringe sometimes dusted with blackish. 15-20 mm.

Caterpillar forming a gall on Helianthus. Moth flying in Texas in August. Maryland to Texas, west to California.

2. S. cinerodorsana Heinrich (Kearfott ms.). Fore wing with costal two-thirds blackish, dorsal third grayish white, the boundary irregular and diffuse; outer third light wood-brown. Speculum white, with some small black spots, between two very broad silvery bars, edged before, above, and beyond, except at anal angle, with the wood-brown. Some dark gray and whitish striation on outer part of costa; one stria extending obliquely across the apex. the rest short and unequal. Fringe powdery gray. darkening to the apex. 13 mm.

July and August.

Maryland, Pennsylvania.

20. RHYACIONIA Hübner

(*Evetria* of authors, not Hübner; *Retinia* Guenée)

Palpi short, roughly clavate, with long, scaled, porrect third segment (fig. 283); thorax smoothly scaled. Fore wing rounded (fig. 270), with oblique, excurred outer margin; the radials all separate, \mathbf{R}_1 from middle of cell, \mathbf{R}_2 rather nearer \mathbf{R}_3 than \mathbf{R}_1 , \mathbf{M}_2 straight, connate or shortly stalked with \mathbf{M}_2 , except in the constockiana group, in which it may be perceptibly curved and well separated at origin; \mathbf{M}_3 curved. No costal fold. Hind wing as in Eucosma. Valve simple, pollex present, uncus absent, socii and gnathos weak or absent. Markings characteristic, formed of slightly raised shining silver gray transverse lines, edged with white, at least at the costal edge, in most of our species on an orange ground.

at the costal edge, in most of our species on an orange ground. R. comstockiana may run by the key to Charlotta, but by its genitalia, habits, and pattern belongs here. The palpi also are normal for Rhyacionia, and slightly aberrant for Charlotta.

The larvæ are of three types. Typically they bore in terminal twigs of pines, very often working in the leader. Some species kill the shoots outright, often causing the tree to fork; others. like the introduced R. buoliana, distort it, resulting in a tree with a crooked trunk. Some are seriously injurious, especially in nurseries, where the only remedy seems to be hand-picking the affected shoots, which show an exuding mass of pitch. The second group, **Petrova** Heinrich, live in pitch-nodules on the twigs; and the third, **Barbara** Heinrich (which contains no described eastern species) hore in the cones of spruce.

1. Ground dark gray.

Key to the species

- 1. Ground gray out to end of cell, the terminal third, more or less, yellowish.

 - 2. Male antenna lightly ciliate; a tawny patch in fold below end of cell, the solid gray extending only about to the middle of the inner margin

5. adana.

- 1. Ground red-brown or orange, at least on outer half of wing.
 - 2. Terminal space gray (except toward apex), or at least crossed by three heavy oblique gray strik.....2. frustrana.
 - 2. Terminal space red.

3. Transverse lines single, narrow, covering less than a third of the surface

- surface of the wing.

 - 4. Terminal band preceded immediately by the outermost gray stria.
 - 5. Base red-brown with a few whitish striæ.
 - 6. Basal line in fringe red......7. virginiana.

I. \mathbf{M}_2 straight, connate or stalked with \mathbf{M}_3 . Larvæ in buds; group vii with three setæ (Rhyacionia).

1. R. buoliana Schiffermüller. Bright orange, typically irregularly shaded with yellow, with six or eight narrow anastomosing pale silver-gray transverse lines, slightly edged with white at costa only. Fringe light gray-brown with a blackish line formed by bars near the tips of the first row of scales. Hind wing light gray-brown, with pale fringe. Head yellow and tegulæ red with powdery gray tips. 18-26 mm.

The larva bores in the tips of pine shoots, causing them to grow crooked.

Europe; occasionally introduced in nurseries. New York: Great Neck, Westbrook, Nassau, and Lynbrook, Long Island.

2. R. frustrana Comstock. Ground deep red, changing from crimson to red-brown, the thorax and base of wing shaded with blackish. Outer part of wing with four broad bands of silver-gray, obscurely divided by series of blacker scales. the last series terminal, except toward the apex, where it forks and one branch runs subterminally to the costa, the other remaining terminal, but falling short of the apex; sometimes with whole apical region gray, or with the gray broken into three bands, the uppermost one crossing the apex. Ground mottled with yellow patches, the dorsal half of the median area usually solid yellow, but with a narrow antemedial band toward costa. 10-13 mm.

Larva boring in terminal and lateral shoots of scrub pine and dwarfing them.

Nantucket, Massachusetts, to Florida and Texas; a larger race in the southwest. New York: Ithaca (Comstock). Karner (Felt). 3. R. rigidana Fernald. Fore wing similar to R. frustrana, the gray bands nor-

mally more extensive, the whole medial area whitish. Last gray band subterminal, followed by a yellow stripe and then a red terminal line. Fringe reddish purple. 18 mm.

April. Larva with the habits of R. frustrana, and apparently commoner inland, on Pinus rigida.

This species can be certainly separated from R. frustrana only by the genitalia.

New York to North Carolina. New York: Ithaca (Fernald).

4. R. busckana Heinrich. Grayish fuscous barred with gray-white. Head and thorax concolorous, the head with some reddish in vertex. Fore wing with outer fourth red, less on inner margin than at costa; a red terminal line and gray fringe. Hind wing smoky. 15 mm.

April.

New York to Pennsylvania. New York: Bellmore and Central Park, Long Island. 5. R. adana Heinrich. Similar to R. busckana; but with the reddish outer part invading the outer half of the inner margin, especially the region of the fold. 17 mm.

End of March and early April.

Massachusetts to Virginia.

II. M_2 well separated from M_3 , and normally curved. Larva nodule-makers with an extra seta on proleg (Petrova).

6. R. comstockiana Fernald. Head white; thorax light powdery gray, including whole of tegulæ. Fore wing light orange, striate with shining light gray, with dense antemedial and medial groups of striæ, covering most of the surface on the basal half of the wing. Fringe pale gray-brown. Hind wing very pale gray-brown, often nearly white, with whitish fringe. 16-22 mm.

Larva boring in younger twigs of Pinus rigida, forming masses of pitch.

Nantucket, Massachusetts, to Virginia. New York: Ithaca (Comstock), Karner (Felt).

7. R. virginiana Heinrich. Head more or less yellow; thorax pale orange, shaded in front with whitish. Fore wing light rusty orange, without any yellow, and heavily striate with somewhat yellowish silvery double striæ; somewhat irregularly striate, but as heavily on outer as on basal part of wing. Fringe very pale brownish, practically concolorous with the striæ; with an orange basal line cut with white; hind wing white, shaded with pale wood-brown. 16-22 mm. (*uenzeli*, Kearfott ms.).

May. Larva forming a large pitch nodule on Pinus virginiana.

Southern New Jersey to Virginia.

8. R. albicapitana Heinrich. Light reddish brown. Head and front of thorax cream white; base of tegulæ and rest of thorax orange. Fore wing with striæ a mixture of silver and lead-gray, leaving a larger area of the ground free at end of cell; some black on costal edge, and black dusting on the striæ. Fringe lead gray with black basal line. Hind wing dark. 16-19 mm.

Larva in young branches of Pinus divaricata.

Fort William, Ontario, to Wisconsin and Saskatchewan.

9. R. gemistrigulana Kearfott. Black, with most of surface covered with pairs of pale gray striæ, about 16 in all. Thorax gray with blackish collar; hind wing lighter brownish gray. 18 mm.

The larva bores in slender shoots of *Pinus virginiana*, the infested shoots being marked in the winter by a ring of pitch.

North Carolina and south; in May.

10. R. picicolana Dyar. Head and front of collar rusty ochreous; thorax and abdomen light brown. Fore wing mottled and strigose in two shades of gray; outer margin and fringe dark, cut with white at anal angle. Base of costa darker; a round blackish spot on inner margin before anal angle and a patch along the outer margin from the costa nearly to the anal angle, with regularly curved inner boundary. Hind wing dark gray with white fringe. (Male not seen.) 30 mm. (Eucosma Dyar).

Larva in a pitch mass on trunk of Abies.

Wisconsin to Washington.

21. PROTEOTERAS Riley

Thorax tufted. Fore wing tufted; outer margin with a notch, and \mathbf{M}_{1} to \mathbf{Cu}_{1} closely approximated, except in *P. claypoleana*, where the emargination is slight; accessory cell and venation otherwise normal; costal fold absent. Hind wing with a black area of sex scaling near the costal margin, except in *P. claypoleana* where there is perhaps a slight suggestion of thicker scaling.

Valve characteristic, with a series of long, flattened, blunt spines on the outer face of the sacculus, near the margin, continuous with the usual marginal spines, and very weak in *P. claypoleana*, where they are about as in *C. ratzeburgiana*. Socii short and stout, finger-like, with a long hair pencil; gnathos free, as in Eucosma.

The known larvæ all bore in petioles and twigs of Sapindaceæ; in fact, that is

the principal reason for putting *P. claypoleana* in this genus rather than in Charlotta. Most of the moths are scaled with green.

Key to the species

1. Ground nearly white	nigrana.
1. Ground gray or greenish.	
2. A blackish angulate bar from middle of costa to end of cell, and t	hence to
apex	centana.
2. No continuous heavy dark bar.	
3. Costa and speculum white $\ldots \ldots 4$. <i>n</i>	aracana.
3 Without broad areas of white	

4. Duller green or gray; male with black scaling on fore wing; very little black at base of wing above.

5. Costa evenly powdery dull gray; sex-patch on fore wing below more diffuse, fuscous......2. willingana.

4. A black dash in basal two-fifths of fold; no black sex-scaling.

1. claypoleana.

1. P. claypoleana Riley. Pale brownish gray, shaded heavily with sage green. Inner half toward base green, with a well-defined outer boundary, crossed by a black dash, which fades out at base; a black bar at base of inner margin. A green shade from tip of basal area to below apex, crossed by one from middle of costa almost to anal angle; both irregular and diffuse, and shaded with blackish where they meet; the blackish area extending outwardly almost to the outer margin. Hind wing mouse gray, with pale fringe. 16 mm. (*Epinotia, Enarmonia*).

This is the most primitive species of the series, having very slight wing-tufting, and the notch at the wing margin indistinct; but its appearance and larval habits place it here. The sex-scaling is present on the parts of the fore and hind wing which overlap, but it is concolorous and hardly visible.

Typically the young larva bores in the petioles of expanding leaves of buckeye; then deserts them, and feeds on the withered leaves; sometimes also in the flower. Mississippi Valley.

2. P. willingana Kearfott. Pale dull gray, with a faint green tint showing under the lens; shaded and mottled with contrasting dark shades. Wing-tufts well marked; notch on outer margin strong. Fore wing with an area of fuscous scales beneath, from Sc to \mathbf{R}_i ; hind wing with black scales on upper surface above the hair pencil only; below, with a black costal streak almost reaching from base to apex. 16 mm.

to apex. 16 mm. June. Larva forming a gall in twig of Negundo, in May.

Maryland to western Canada and Kansas.

3. P. crescentana Kearfott. Dull light gray, with a contrasting blackish band which runs from middle of costa to end of cell, then turns abruptly, running to the apex; half as wide at apex as at middle of costa. Sex-scaling slight, at middle of costa of hind wing above and below. 17 mm.

June. Larva with P. willingana in box-elder.

Maryland to Iowa.

4. P. naracana Kearfott. Fore wing rather smoothly light olive green, with some black and brown toward apex. Costa with a broad and irregular white area, tapering to a point at apex; speculum mostly white. Sex-scaling on under side of fore wing at middle of costa slight; hind wing without black scales, but with a heavy brown hair pencil on upper side. 16-20 mm.

End of May to June.

Western Pennsylvania to Ohio.

5. P. æsculana Riley. Darker olive green, more or less mottled with yellow and gray, and with some small black markings. The most distinct gray areas being a darker fascia from middle of costa toward anal angle, becoming diffuse below, and a shade running to apex; often with a fine longitudinal black streak at their junction. Fore wing below with black sex-scaling in a contrasting patch a third the length of the wing; hind wing with sex-streak, below, lying well away from costa; above, with one on each side of the costal hair pencil. 12-18 mm.

June. The larva bores in the tender terminal twigs of maple and buckeye, in May, causing them to wither.

The distribution is uncertain, as a large part of the records for the genus have been reported under this name. The species appears to be general. New York: Ithaca.

6. P. moffatiana Fernald. Bright green, mottled with black, heavily on basal third and in the form of a curved band from middle of costa to apex. Sex-scaling only along costal edge of hind wing, below; hair pencil concolorous and inconspicuous.

June. Larva in petioles of maple.

Canada to Pennsylvania. New York: Ilion, Lancaster, East Aurora, Ithaca, Big Indian Valley.

 $\overline{7}$. P. obnigrana Heinrich. Fore wing dull white, with olivaceous basal patch, a fascia from middle of costa to anal angle, and a shade below costa near apex. A thin black line from apex to near middle of wing, curving as in *P. \sigmasculana*. Sexscaling of male diffuse, on under side of both wings, dark and fairly conspicuous. 16 mm.

Dublin, New Hampshire; type only known.

22. GRETCHINA Heinrich

(Thiodia; Proteopteryx, in part)

Thorax with dorsal tufts; fore wing with more or less distinct tufts or transverse ridges, at least in the fold. Outer margin concave, normally strongly notched, with \mathbf{M}_1 to \mathbf{Cu}_1 closely approximate at margin; \mathbf{R}_4 and \mathbf{R}_5 stalked or separate; \mathbf{R}_1 from middle of cell; \mathbf{Cu}_2 sinuate; costal fold absent. Hind wing with \mathbf{R} and \mathbf{M}_1 approximate. Fore wing gray, largely formed of a mixture of black and white, with a characteristic cusped black mark, more or less suggestive of a bird's beak, below the apex.

Valve simple; cucullus moderate; spined area reduced and lateral and anal spines strong; sacculus densely hairy on outer side. Uncus absent; socii markedly chitinized, triangular, porrected; gnathos reduced and partly fused with socii.

This genus is well marked superficially, really resembling only the Catastegas; but in the forms with \mathbf{R}_{4} and \mathbf{R}_{5} separate, the only real diagnostic characters are in the genitalia.

Key to the species

1. \mathbf{R}_4 and \mathbf{R}_5 stalked (fig. 260); tufting stronger as a rule.

2. Hind wing white on basal half, contrasting with the blackish fore wing.

3. watchungana.

- 2. Hind wing darker, pale only when fore wing is equally pale.
 - 3. Gray without any brown tint.

2. delicatana.

1. G. amatana Heinrich (Kearfott ms.). \mathbf{R}_4 and \mathbf{R}_5 approximate; \mathbf{R}_5 to \mathbf{M}_2 markedly curved; \mathbf{M}_1 associated with the notch (unlike Exentera). Blackish, shaded with whitish, the dorsum beyond the antemedial band tending to show a rounded or broad triangular pale (whitish) patch, suggesting that of *Epinotia timidella*. Speculum of two very broad and dull bars, with a narrow line of the ground between them, often obscure, but normally pale, very often separated from the central patch by a distinct blackish bar. Fringe dark and obscurely barred, the longitudinal line in it usually more distinct. Hind wing dark gray, as in *G. bolliana*.

Moth locally common on tree trunks, in May and early June.

Forest Hills, Massachusetts, and south. New York: Ithaca, Crosby (Yates County), McLean.

2. G. delicatana Heinrich. Similar to G. amatana. Fore wing with \mathbf{R}_{*} and \mathbf{R}_{*} connate. Fore wing narrower, with the powdering more whitish gray, extending along the costal and subcostal region to the base, cutting off the darker basal patch; a tendency to a nearly continuous dark line from apex to upper side of cell, without the subterminal spot or hook of G. amatana. Hind wing paler toward base. 15 mm.

April and May.

New Jersey and Pennsylvania.

3. G. watchungana Kearfott. \mathbf{R}_4 and \mathbf{R}_5 very shortly stalked; fore wing a little rough scaled. Blackish gray, in some lights shot with brown, especially at apex; irregularly mottled with whitish. Costa with fine irregular oblique straition; inner margin with coarser and rougher oblique streaks and bands; with two strong pale streaks at middle of inner margin, tusing into a vague patch, as a rule; but when distinct, with the outer one decidedly shorter than the more basal. Speculum pale, a mixture of white and dull silver, with some black scales. Apex black, preceded by two strong white striae on costa. Fringe with a broad gray line, breaking up toward anal angle, and cut with white below apex. Hind wing white, shaded with fuscous outwardly and on veins, with pale fringe. 15 mm. (Thiodia).

May.

New Jersey to Pennsylvania.

4. G. dulciana Heinrich. Similar to G. watchungana, with the same contrasting pale speculum; ground more ferruginous brown, especially on the outer half of wing. Hind wing dark smoky. 14 mm.

June and July.

New Jersey and Maryland.

5. G. deludana Clemens. Tufts more distinct than in the preceding species, much weaker than in G. bolliana, the outer part of Cu_2 with a tuft like the others. Similar to G. bolliana, but with the ground more crisply powdered with white, and the brown areas small and crisply dusted with the yellowish scale-tips; blackish shades not obvious, but black lines strong; antemedial evenly concave up to the cell and then abruptly ending; the dorsal half of the base, when darkened, with an even straight upper boundary, not extended by an antemedial spur; black bar in end of cell not as strong as the one just beyond it, connected with the "bird-beak" marking; lower line of the "bird-beak" marking usually fine. and reaching down to the anal vein. Hind wing paler than that of bolliana, whitish at base. 15 mm.

May. Found in similar places to G. amatana and the Catastegas.

New Hampshire, to Plummer's Island, Maryland. New York: Crosby (Yates County).

6. G. bolliana Slingerland. R_4 and R_5 stalked halfway to apex; tufts at and along base of Cu_2 , a second obliquely in and below this one on fold and anal vein, and on inner edge of speculum. Fore wing powdery gray, suffused with black on basal third below fold, as far as the first tuft, sometimes sending a spur up across the cell near the base; a black area in outer part of cell, bounded above by a black bar along upper edge of cell, and a blackish crescent, convex up, covering the bird-beak marking, and connected by a narrow black bar to the apex of the Upper part of face blackish, overhung by a large gray tuft between fringe. antennæ; lower face gray. 18 mm.

Caterpillar injurious to pecan in the South; moth flying northward in September and southward in April.

New Jersey to Wisconsin, Florida, and Texas. "New York" (Kearfott).

23. EXENTERA Grote

(With Thiodia, in part)

Similar to Gretchina; fore wings smooth (figs. 256, 261).

Cucullus normal; anal indentation with short black spines; no strong lateral spines; anal spines weak. Socii as in Hulda.

The genus is close to both Gretchina and Thiodia, and the species which have \mathbf{R}_{\star} and \mathbf{R}_{s} separate differ from Thiodia only in their genitalia and appearance. They are narrow-winged forms that mostly fly in the early spring (whence the name of the type species). While the genus is easily recognized, after a little practice, the species are very close and not all are surely distinct. 1. E. maracana Kearfott. Notch well-marked. Fore wing whitish gray; dark

basal patch extending a quarter of the length of the wing on costa and inner margin, and running out in a right angle at middle; a blackish shade extending out from middle of base; an oblique dark fascia from middle of costa to inner margin at anal angle, with a brown shade crossing it in the middle. Median area also shaded with light brown. A large horizontal triangular dark spot below costa, its base resting on costa and apex on outer margin; speculum with or without a few black dots; costo-apical white spots conspicuous; fringe blackish outwardly. Hind wing light smoky brown. 13-161/2 mm. End of March to April.

Cincinnati, Ohio; western Pennsylvania. New York: Ithaca, McLean.

2. E. improbana Walker. R. and R. normally barely stalked. Fore wing normally nearly immaculate gray, often with a strong purplish or greenish iridescence, or duller fuscous gray without contrasts; the antemedial line marked as the boundary of the darker base, more oblique on the dorsal half than in E. spoliana, and usually straight or evenly and gently curved. 18-20 mm. (cressoniana Clemens, apriliana Grote).

The moth flies in early spring and is sometimes very common. The various forms seem more or less constant locally, and would represent distinct strains. Larva on hickory and perhaps apple (another species?).

Eastern States: exact distribution uncertain on account of confusion with the following species; the race oregonana extending west to the Pacific.

3. E. spoliana Clemens. Typically gray, with strong whitish contrasts; a blackish area before antemedial line, and one on inner margin before anal angle; the antemedial line on dorsal half of wing normally strongly excurved or waved, and more erect in general course. The commoner form with strong reddish shades, at least over costal half of wing. Genitalia distinct from those of E. improbana. 18 mm. (haracana Busck, not Kearfott).

Caterpillar on chestnut, rolling the tip of the leaf. Moth in early spring, locally very common.

Northeastern States: exact distribution uncertain.

4. E. haracana Kearfott. \mathbf{R}_4 and \mathbf{R}_5 not stalked. Head and thorax blackish. Fore wing blackish with obscure coarse violet-gray strine; becoming finer paired whitish ones at costa. An irregular black line from outer part of cell to apex, evanescent under high magnification. Speculum composed of two long lead-gray bars, the outer one broken; filled with clay-colored powdering, which also covers the region above it, below the dash. Three or four black bars in speculum. Fringe with faint longitudinal lines; the dash in the apex obscure. Hind wing rather pale. 15 mm. (faracana Kearfott?)

I have seen no decent material of this species; some specimens determined as *faracana* appear to show the longitudinal black streak in the axis of the wing. New Jersey; Pennsylvania; Missouri?

5. E. costomaculana Clemens. Fore wing light pinkish ochreous or cream color, paler about the edge of the dark patch; a few scattered black scales. A large rounded brown patch, extending down from middle of costa, sharply defined below, but somewhat diffuse above and shading into the pale ground. Patch shaded with black. Fringe grayer than ground, dark at apex. 16 mm.

This species and the next differ from the similarly marked Epinotias in the pale base of the fore wing.

Common and general in distribution, April and May.

New York: Gowanda, Rock City (Cattaraugus County), Portage, Crosby (Yates County), Ithaca.

6. E. virginiana Clemens. Closely similar to E. costomaculana but averaging a little larger; ground light violet-gray, shaded with brown, especially below apex; not paler than hind wing. Patch rather darker brown than in E. costomaculana. Speculum indicated by well-marked black dots in the brown outer shade. 18 mm.

April and May.

Pennsylvania. New York: Ithaca, Pearl River, Albany.

24. GYPSONOMA Meyrick

(*Epinotia*, *Hedya*, in part)

Fore wing with apex rather marked, but not distinctly falcate, usually marked by an ocellate apical spot; outer margin concave, but more broadly than in the preceding genera; in *H. haimbachiana*, suggesting Anchylopera; \mathbf{M}_1 to \mathbf{M}_3 slightly approximate at margin; \mathbf{R}_1 arising from cell at middle: \mathbf{R}_2 halfway between \mathbf{R}_1 and \mathbf{R}_3 ; accessory cell short, commencing before the point of separation of \mathbf{R}_2 ; \mathbf{Cu}_2 straight; no costal fold. Hind wing with \mathbf{R} and \mathbf{M}_1 stalked. (Fig. 274).

Valve club-shaped; cucullus moderate; clasper rudimentary; anal indentation smooth; sacculus sparsely hairy. Uncus wanting: socii as in Charlotta, articulated on a stem, the gnathos partly fused with them.

1. G. fasciolana Clemens. White, shaded with gray toward outer margin; a few black scales in position of speculum. Outer part of costa dark gray, crossed by paired white striæ. Base blackish, the outer boundary oblique outward and a little convex; a broad fascia of the same color from middle of costa to before anal angle. 15 mm. This is very similar in markings to some species of Exartema and Olethreutes, but may be distinguished by the stalked M_s and Cu_1 of the hind wing as well as by the lack of the fold on the inner margin in the male. There is a good deal of variation in the details of markings and the species should probably be divided. This species and the next represent Meyrick's genus Gypsonoma, but there do not seem to be any really tangible characters. (Epinotia auct., Anchylopera Clemens; blakeana Grote.)

July.

A northern species, ranging from Nova Scotia to Manitoba, and south to Penn-sylvania. New York: Peru, McLean, Karner, and New Windsor. 2. G. substitutionis Heinrich. Blackish. Head grayish fuscous; fore wing with

a moderately broad white median area, its inner margin regularly excurved and outer margin outwardly oblique and a little irregular; outer part of costa with four pairs of white striæ; the first and third pairs lying at the commencement of irregular excurved lead-colored bands, which approach and become broader-toward inner margin, forming the speculum. Speculum enclosing a few black dots or bars. Fringe dark, with a blackish basal line. Hind wing dark. 12 mm. July.

Manitoba; New York: Ithaca.

This is the American representative of G. incarnana, and may stand as it in some collections, but as a rule that name has been used for specimens of the transmissana group of Epiblema.

3. H. haimbachiana Kearfott. Dull gray, less brownish than H. salicicolana. Basal third blackish and strongly contrasting toward inner margin, becoming ighter toward costa, its outer boundary angled at Cu, often forming nearly a right angle. Often with a slight gray shade, but little darker than the general color, from middle of costa to inner margin, three-fourths way out, bent somewhat at middle, and somewhat irregular, twice as wide at inner margin as at costa; also with some vague striation. Apical dot black, contrasting, but a little blurred; line in fringe distinct toward costa. Concavity on outer margin a little shallower and broader than in *H. salicicolana*. 15 mm. (*Epinotia*? Kearfott).

Superficially H. haimbachiana is rather like Rhopobota ilici-June to August. foliana, but it is easily distinguished by the shallower notch in the margin as well as by the free \mathbf{R}_4 and \mathbf{R}_5 , and the lack of sex-scaling.

New Jersey to Wisconsin and southern Ohio.

4. H. salicicolana Clemens. Notch well marked and well above middle of wing. Dull gray-brown, when fresh with a slight purple gloss; the basal third contrastingly darker, obscurely mottled, the outer two-thirds sometimes frosted with whitish; outer boundary of dark base strongly outcurved at middle. A dark streak from costa beyond middle to middle of wing at two-thirds way to apex, sharply bent there, and very obscurely continued to inner margin. Dark apical dot obscure, preceded by a pale stria; dark line in fringe obscure below, distinct toward costa. 12 mm. (Epinotia, Hedya; saliciana Clemens).

June. Larva a leaf roller on willow; also in "pine-cone" gall. New Hampshire to Pennsylvania, and probably generally distributed. New York: Ithaca.

25. ZEIRAPHERA Treitschke

(*Epinotia*, in part)

Thorax and fore wing smooth. Fore wing (fig. 253) with outer margin evenly excurved, the veins not at all approximate at margin. \mathbf{R}_i and \mathbf{R}_5 sometimes connate, not stalked; Cu₂ slightly bent up toward margin; no costal fold.

Valve sickle-shaped; anal indentation densely spined; cucullus large; sacculus reduced, not strongly spined, but practically always with a series of bristles homologous with the enlarged ones of Proteoteras, but far less conspicuous. Uncus

rudimentary; socii broad, triangular; gnathos free, weak. This is a homogeneous genus, and perhaps more primitive than its position would appear to indicate, as it closely resembles the species of Epinotia which have a similar wing form. P. claypoleana is put in this genus by Heinrich, but differs markedly in wing form, and I would put more weight on the characteristic larva than on the small difference in genitalia.

WILLIAM T. M. FORBES

Key to the species

- 1. Costal half brown, contrasting with the pale dorsal half; the boundary on middle of wing sharply defined, forming a rounded lobe.
 - 2. Base of inner margin below fold contrasting pale gray, sharply separated from the black-brown costal portion......4. Epinotia lindana.
- 1. Costal half not contrastingly dark.
 - 2. No silvery fasciæ.

5. Expanse 10 mm.; blackish with dirty white striation.

- 1. Epinotia nanana.
- Expanse 15 mm.; base of fringe cut with white.
 Blackish with ash-gray striation.....l. diniana.
 With white fasciæ and striations......2. fortunana.
- 3. Conspicuous bright silver fasciæ.....2. Epinotia cruciana.

1. Z. diniana Guenée. Ash gray, powdery-strigose in two shades; fore wing with antemedial line right-angled in middle of wing, defining the darker base; a fascia from costa, just beyond middle, to anal angle; a little irregular, and widening slightly to inner margin; clearly defined below, but sometimes connected at costa to the dark base and outer margin by blackish suffusion. Outer margin dark shaded, with this shade not forming a definite fascia. Fringe cut with white twice below apex, and twice at anal angle. Hind wing somewhat browner. 18 mm. (pinicolana Zeller, pseudotsugana Kearfott).

August. Larva green-gray, with black head, dark brown cervical shield, and dark tubercles, between needles spun together of various conifers; fir, larch, and others. Pupa in the ground.

Arctic-alpine; Mt. Washington, New Hampshire, and north. "New York" (Fernald).

2. Z. fortunana Kearfott. Black-brown, sometimes dusted with wood-brown: with a broken double white fascia at base; an irregular but strong and complete excurved fascia just before middle; irregular white markings ontwardly, the most distinct usually a fascia starting from costa at three-fourths way to apex and running two-thirds way across to anal angle, with a patch on outer margin at its lower end. A few additional white strike at costa. Black basal line in fringe cut with white at fold and anal agle. Hind wing mouse gray. 15 mm.

End of June to July.

Ottawa, Ontario.

3. Z. ratzeburgiana Saxesen (in Ratzeburg). Yellow-brown, or light woodbrown; antemedian band dark brown, and defining a darker base. sharply angled at lower side of cell, and bent in, in cell; outwardly, with a fascia from middle of costa to anal angle, wider above; normally broken into three toward costa, and sometimes crossed by a black bar along lower side of cell. With a shade of mixed black and dark brown, extending obliquely down from apex, often preceded by a white snade and followed on middle of outer margin by a white stria. A dark brown line in fringe. This moth is variable in details of markings. 12 mm.

Larva webbing together the terminal needles of spruce in early spring. Pupation outside the nest. Moth in July and August.

Ottawa, Ontario, to Washington, and probably general where its food plant occurs; also in Europe.

26. PSEUDOGALLERIA Ragonot

Palpi porrect, clavate; the tip formed by the third joint but markedly set off only in rubbed specimens. Thorax tufted. No sexual modifications. Fore wing with all veins separate, and evenly spaced at margin; wing rather long and narrow, as in Thiodia; apex marked; outer margin with a strong concavity center-ing on M_1 and M_2 ; R_2 arising from discal cell. Hind wing normal, with R and M_1 approximate, M_2 close to M_3 at origin, M_3 connate or shortly stalked with Cu_1 . Eighth segment of abdomen somewhat chitinized and modified. Valve simple.

Uncus and socii absent. Gnathos weak.

An isolated genus. The wing form and the modification of the eighth segment suggest Hystrichophora, but in the remaining genitalic structures there is no likeness.

1. P. inimicella Zeller. Dull gray, more or less shaded with brown; the notch in the margin edged with a brown band. Costa heavily shaded with blackish. Hind wing practically concolorous. 13-20 mm.

June. Apparently rare. Larva boring in rootstocks of Smilax.

New Jersey to Manitoba and Texas. New York: Ithaca.

27. HYSTRICHOPHORA Walsingham

Fore wing similar to Pseudogalleria; Accessory cell normal.

Male genitalia with eighth segment of abdomen modified. Valves asymmetrical; a strong dorsal process arising from the upper articulation of the valve and extending out parallel to it; cucullus and sacculus not defined. Uncus long strong, triangular, bifid; socii and gnathos absent.

Three species of this genus are likely to occur in the territory considered. H. vestaliana Zeller is white with black points, and is known from Iowa; H. ochreicostana Walsingham is brown and yellow, and has been taken in Missouri and west; *H. kokana* Kearfott, a smoky species with contrasting pale border, occurs to the southward.

28. EPISIMUS Walsingham

(Enarmonia Walsingham and Durrant, in part)

Similar to Thiodia, but with Ms and Cu, of hind wing connate (fig. 262) instead of stalked. Fore wing smooth, with all veins free.

A transitional genus, connecting the Eucosmini and Olethreutini. 1. E. argutanus Clemens. Costa thickened, but not folded, in male. Dull red-dish or grayish brown, the color made up of fine mottling in wood-brown, graybrown, and blackish, overlaid when fresh with purple. Costa shaded with blackish along middle; speculum confused, dark gray toward base of wing, paler outwardly, and more brown in the upper part, where there are a couple of black dots. 13 mm. (Bactra Clemens).

May. Larva on various shrubs, especially Rhus and Hamamelis; on Euphorbia in the south. The larva lives between two leaves sewed together, or rolls a leaf into a cone. (Possibly Catastega hamameliella Clemens).

New York to Florida. New York: Speculator, Crosby (Yates County), Ithaca, New York City.

2. E. tyrius Heinrich. Whitish gray, tinted with ochre, and shading into orange ochre and reddish toward outer margin. Costal part on basal half dark brown, contrasting, the boundary sharp toward base, and strongly curving down in antemedial region, diffuse outward. Hind wing dark gray. 12-15 mm.

Larva on maple, with nearly the habits of E. argutanus.

Long Island, New York to Mississippi.

29. OLETHREUTES Hübner

(Argyroploce; Penthina; Sericoris; Eucosma, in part, etc.)

Palpi porrect (fig. 281), clavate or triangular, with short porrect third joint. Thorax tufted behind; fore wing smooth or with slight tufts or ridges at end of cell and near base of inner margin. Fore wing (fig. 266) with all veins free and well spaced at the margin, which is evenly rounded, M_2 a little spaced from M_3 and Cu_1 at origin; no costal fold. Hind wing with R and M_1 approximate or stalked; \mathbf{M}_2 distinctly separate from \mathbf{M}_3 , but rarely as widely as in Laspeyresia (0. hebesana); M_3 and Cu_1 connate, rarely stalked, or very slightly separate. Inner margin in male thickened and variously folded or rolled up; sometimes with a lobe as in Exartema, but if so, with the lobe concealed in the fold, and quite small; usually with a hair pencil. No thickening or dense scaling except on dorsal margin. Hind tibia of male with a hair pencil arising from base (rather fragile, and lost in rubbed specimens) but without long dense hair. Rarely, the dorsal fold or tibial hair pencil are rudimentary, but never both in the same species.

This is a large and well-defined genus (at least in the male) from which Phacasiophora is derived; it is closely related also to Cymolomia and, to judge by the occasional occurrence of a free lobe on the inner margin, is perhaps derived from it; more probably it is heterogeneous, composed partly of older and partly of newer species than Cymolomia. Bactra is also considered related, but lacks the thoracic tuft. The male is easily distinguished; the female is like that of Cymolomia and Phæcasiophora, but can be distinguished from other genera by the combination of tufted thorax, practically smooth wings, approximate R and M_1 , and M_2 shortly but distinctly separate from M_3 and Cu_1 . Ecdytolopha and Gymnandrosoma have ucarly the same combination of characters, and are united to it by Walsingham and Durrant, but \mathbf{M}_2 is more widely separated, the palpi are smooth, and the general appearance is different.

Key to the species

ı.	\mathbf{F}	ore v	ving	dark,	, with	moder	ate,	clearly	r-de:	fined	white	costal	patch.	
	2.	Pate	h a	third	the le	ngth of	the	wing,	its	edge	sharpl	y defin	ed and	even.

33. ehionosema.

2. Patch less than a quarter length of wing, irregular or diffuse on the lower outer side.

	3. Patch	pink .							. 31.	roseo	maculana.
	3. Patch	white							32.	cost	imaculana
1.	Fore wing	whitish	with	dark	shade	toward	middle	of costa		. 30.	albeolana.

1. Median area evenly yellow, contrasting.

- 1. Median area with a large, evenly colored patch of greenish or olive-gray.
 - 2. Patch on inner margin, between two black patches.
 - 2. Patch not resting on inner margin; outer part of wing deep rose.

36. malaehitana.

- 1. Ground yellow and yellow-brown or yellow-brown and black-brown, separated by brilliant blue metallic lines or series of dots.
 - 2. Ground two shades of dark brown; small (12 mm.).....19. coronana. 2. Paler portion of ground ochre yellow; large (18 mm.)

- 3. Hind wing beneath dark, concolorous with fore wing.....16. coruscana.
- 3. Hind wing beneath contrastingly pale, somewhat darkened at costa only.
 - - with the outer margin at an angle to the dorsal and outer margin of fore wing nearly straight......17. constellatana.
- 1. Fore wing with base dark gray, contrasting with the whitish apical third or half.
 - 2. Outer boundary of gray area, located near middle of wing, diffuse and farther out on costa than on inner margin...... (Ecdytolopha islandana.) 2. Outer boundary clean-cut, more transverse, and farther out.
 - 3. Smaller, ground of apical part pinkish; outer boundary of dark base usually striking inner margin well before anal angle; often followed
 - 3. Larger; ground pure or cream white; outer boundary running almost to anal angle, the stria being sometimes traceable, but fused in with the blackish base.
 - 4. Apical white ground extended as a hook in apex of cell, partly outlining a black dot in lower angle of cell.
 - 5. Expanse over 18 mm.....23. capreana.
 - 5. Expanse 15 mm.; apical white region less sharply defined.
 - 22. apateticana. 4. No white hook in end of cell, and usually no obvious black dot in lower angle.
 - 5. Outer boundary of black area, distinctly excurved at middle; sub-
 - 5. Outer boundary of blackish area nearly straight and erect, the apical
 - 5. Outer boundary of blackish area drawn in at cell, with a well
 - marked notch, surrounded with black.
- 6. Eyes large; a large mottled species......24. youngana. 1. Fore wing with ante- and postmedial costal spots on a blackish ground; eyes
- 1. Fore wing with a clear white antemedial fascia, sometimes lightly scaled with gray, or with whole middle costal area suffused with white, and the fascia distinct only toward inner margin; eyes large.
 - 2. Blackish median fascia only narrowly reaching costa, or falling short;
 - 15. aspasiana.
 - 2. Blackish median fascia of nearly even width, and on costa about a fourth as wide as length of wing.
 - 3. Apical portion of tegulæ and bands on disc of thorax white...3. carolana. 3. Thorax and tegulæ blackish.
 - 4. Postmedial fascia forking, and enclosing a triangular blackish dorsal spot (the inner fork gray, only the outer one white) dark markings
 - 4. Median area solid blackish; no triangular patch cut off on its outer side.
 - 5. Fore wing broader, the blackish median fascia normally nearly twice as high as wide, median area with large lead-gray flecks.

1. bipartitana.

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- 5. Fore wing narrower, the median fascia almost as wide as high, median area not flecked with lead-gray.
 - 6. White discal dot obvious, connected with postmedial line and flanked above and below with black teeth, as in Cymolomia.

4. polluxana.

6. White diseal dot obsolescent, not connected with postmedial white; the postmedial line nearly straight opposite cell.

2. fuscalbana, 3. carolana.

- 1. With antemedial fascia yellowish, gray-filled, or dull lead-gray; less contrasting, on a dark brown ground; sometimes reduced to costal and dorsal groups of striæ.
 - 2. Fascia yellowish, not shining.
 - 3. Median dark area about of equal width throughout.....14. instrutana.
 - 3. Median dark area half as wide at costa and inner margin as at middle. 15. aspasiana.
 - 2. Fascia lead-gray, or replaced by shining striæ.
 - 3. Expanse 16 mm. Fascia, when complete, of three or four partly fused striæ.
 - 4. Ground mostly dark brown.
 - 5. Brown terminal dots in base of fringe.....9. septentrionana. 5. No such marking......8. intermistana.
 - 4. Ground overlaid with tawny yellow.....10. schulziana.
 - 3. Expanse 10 mm. Fascia of two striæ......11. abietana.

- 1. Ground even rich dark brown, with narrow darker median fascia.
- 2. Fascia clean, cut, Cu, arising three-fourths way out on cell....41. hemidesma. 1. With "Cymolomia pattern." 33
 - 2. Ground rusty orange, marked with dull red; discal dot white.

(Phæcasiophora niveiguttana.)

2. Dull rose, shading into luteous; a black longitudinal dash in outer part of 2. Dark wood-brown, the subterminal and lower end of the median patch

- lead-gray marks, if present, not gathering into antemedial fascia; no white or pale colors.
 - 2. Fore wing all of dull colors (or with some slightly shining scales irregularly distributed).
 - 3. Clear powdery ash-gray; seales bicolored.
 - 3. Not clear powdery gray; the powdering, if present, yellow; individual scales unicolored.
 - 4. Lighter markings composed of paired lead-gray striæ and yellow scaling; larger forms.
 - 5. Fringe with brown dots in its base.....9. scptentrionana.
 - 5. No brown dots in base of fringe......8. intermistana.
 - 4. Fore wing with metallic scaling, if present, in broad areas; no yellow scaling.

³³ Ground of a lighter shade, usually brown; the markings mostly clean-cut, finely pale outlined, outwardly consisting of a strongly irregular median fascia. sometimes interrupted; an oblique subterminal spot resting on the middle of the outer margin, and a subtriangular spot at the anal angle, all rather evenly colored in a darker shade; the subterminal patch alone of these really distinct in *interruptolineana*.

5. Ground purple-black, not darker at middle of costa....43. dackeana. 5. Ground brownish, with a blackish shade at middle of costa.

42. hebesana.

5. Lighter dull fuscous with a vague transverse luteous band.

40. murina.

2. Fore wing with some brilliant blue iridescence.

3. With a well-defined, vertical, blue-gray bar before anal angle.

6. Fringe almost entirely dark 6. Fringe on costal half of outer margin mostly cream-white.

47. albiciliana.

4. Heavily dusted with cream and whitish 3. With a large blue patch before anal angle, ground blackish...44. cyanana.

O. conditana is prohably confined to California: eastern records are based on dwarfs of hcbesana, and specimens of Polychrosis.

I. O. bipartitana Clemens. Fore tibia with faint pale bands; mid-tibia usually all black, and hind tibia whitish. Palpus with more fuscous than in O. fuscalbana. Fore wing rather less than twice as long as wide: mixed dark shining blue-gray and black; a white antemedial fascia, with some black scales, a sixth as wide as and blacks, a write anteneous rasea, with some black scales, a sixth as wide as length of wing, erect and of even width, its boundaries slightly irregular. Beyond this a blackish fascia, slightly broadening toward inner margin, reaching ont to end of cell. The rest of wing white, shaded with gray on the apical half, more or less, or with a separate gray margin and subterminal patch. Costal edge gray, with paired white striæ. Hind wing typically whitish, gray in variety cæsialbana Zeller; fringe whitish. 16 mm. (Antithesia Clemens).

May and June; August.

Common everywhere. New York: North Elba, Fentons (Lewis County), Oswego, Jamestown, Rock City (Cattaraugus County), Millerville, Otto, McLean, Ithaca, Big Indian Valley, Albany, Ramapo.

2. O. fuscalbana Zeller. Fore tibia with three heavy white bands; mid-tibia with two or three weaker ones; palpus heavily mottled with white. Wings slightly narrower than O. bipartitana; antemedial faseia somewhat narrower. hardly more than half as wide as the blackish area following it, which is more or less dusted with broad yellow scales. Apical region more distinctly blackish, especially the apical dot; subterminal bar conspicuously dark. 15 mm.

June to August: common.

Generally distributed, south to North Carolina. New York: Peru, North Twin Brook (Mt. Marcy). Newcomb, Peru, Rock City (Cattaraugus County), Ithaca, Trenton Falls, Fort Edward.

Gypsonoma fasciolana is near this species in appearance, but differs in the stalking of \mathbf{M}_{3} and \mathbf{Cu}_{1} , the wing-form, and the narrower, decidedly oblique median dark fascia, preceded by a broader and purer white antemedial fascia. Cymolomia fasciatana has similar tibix, but the palpi are almost wholly whitish; and the antemedial fascia is considerably stronger than the postmedian.

3. O. carolana McDunnough. Tegulæ white, with a dark patch at the base; thorax banded with white. Blackish and olivaceous brown; antemedial white band slightly irregular; subterminal white band narrow, the terminal area being broadly suffused with olivaceous brown; anal spot fused in with the dark median area, with a point of the white color running between them above; two olive-brown costal spots, sometimes connected by hair-lines with the brown terminal area. 14 mm. (Not scen.)

June.

Ottawa and Trenton, Ontario.

4. O. polluxana McDunnough. Similar to O. fuscalbana. Ground blackish, without any yellow overscaling. Antemedial fascia about as in O. fuscalbana; postmedial more irregular, and broken into by patches of the black ground; the blackish medial area extending in two strong teeth at the end of the cell with a white indentation between them. Outer markings almost entirely brown-black and white, without the gray subterminal band of O. fuscalbana. 25 mm.

June and July.

New York to Alberta. New York: North Twin Brook (Mt. Marcy), Peru.

5. O. campestrana Zeller. Similar to O. fuscalbana. Ground overlaid with slender yellow scales. Antemedial band more distinctly made up of two pairs of fused white striæ. Postmedial white band on costal half formed of two pairs of white striæ; below the middle the anterior pair becoming gray and turning abruptly in to the middle of the inner margin, while the outer pair becomes a broad white band extending to the anal angle; the two enclosing a triangular blackish patch between them. Outer part of wing blackish, with strong paired white costal striæ, and a couple of longer white striæ, defining the usual subterminal patch. A white subterminal line toward apex. 15 mm. (dealbana Walshingham, not Walker).

June. Larva on cherry.

Generally distributed and not rare, but generally confused with O. fuscalbana. New York: Peru, Rock City, Ithaca, Trenton Falls, Albany (New York State Museum).

6. O. mengelana Fernald. Eyes very small. Ground powdery black on a bluegray base; antemedial whitish fascia distinctly composed of a group of three or four coarse striæ, fading out below; postmedial white fascia broad and distinct toward costa, of two pairs of striæ, becoming obscure below the middle of the wing. Outer margin with the blue-gray dominant. Outer half of fringe cut with white. 15 mm.

July.

Greenland. This is probably a widespread arctic species, but I have seen no other specimens.

7. O. turfosana Herrich-Schæffer. Fore wing with nearly even, broken, transverse striation of blackish, clay color and lead gray, but with the pale striæ more or less paired, especially at the costal edge. Outer part of wing normally more heavily shaded with the yellow. A white spot at end of cell, partly defined with blackish. Fringe blackish, irregularly cut with white. Hind wing dark gray. 16 mm.

June and July.

Labrador; Maine; Europe.

8. **O.** intermistana Clemens. Blackish gray with a variable amount of yellow and white scaling. Fore wing with confused paired white or light blue-gray double striæ, cutting up the ground into patches toward the outer margin, but leaving fairly definite narrow antemedial blackish bands, or with the striæ toward base confined to costal and dorsal edges. Scaling sometimes gathering to form a pale shade at anal angle. A white discal dot. Fringe black and white. 18 mm. (*tessellana* Packard).

August.

Labrador to Alberta; Mt. Washington, New Hampshire.

9. **O.** septentrionana Curtis. Blackish brown, somewhat variegated with gray, leaving a darker oblique fascia as in *O. intermistana;* costa with six or seven pairs of white striæ; fringe pale, spotted with brown at the base. Hind wings apparently variable. 15 mm. (Sciaphila primariana Walker; Penthina fulvifrontana Packard).

Arctic America south to Labrador.

10. 0. schulziana Fabricius. Fore wing black, heavily overlaid with narrow bright yellow scales, with somewhat confused and broken paired white striæ, the

subterminal ones not anastomosing as in O. intermistana, but leaving a clear postmedial band of the ground. A white spot at end of cell, partly edged with blackish. Fringe checkered black and white. 18 mm.

The larva is said to feed on pine or Vaccinium.

Europe; also reported from Arctic America.

11. O. abietana Fernald. Fuscous brown, marked with shining gray. A slight gray basal facia; a broad antemedial fascia, starting from two pairs of costal striæ. A postmedial fascia, also starting from two pairs of striæ, with a small dark dot between them, its outer side somewhat wavy and its inner side with two deep notches (above and below the cell), into which run teeth from the brown median area. (Two pairs of costal striæ toward the apex, from which runs a broken irregular subterminal fascia. Hind wing dark. 10 mm. (piceæ Busck).

Larva on spruce.

Maine to the Western States; generally distributed northward. New York: Ithaca, Woodmere, Long Island.

12. O. deceptana Kearfott. Finely dusted with white, gray, and black, the gray scales white-tipped; no yellow or other colors. Head and thorax concolorous. Fore wing with costa arched, transversely strigose; base slightly darker, with excurved irregular outer boundary; median fascia bluntly two-toothed, barely traceable; the notch between the teeth white-filled, but not forming the definite spot of the intermistana-group. A slight narrow fascia across apex. Hind wing duller; brownish gray. 18 mm.

June to July.

Peru, New York; Dickinson, Michigan, and westward. 13. O. removana Kearfott. Ash gray; each scale finely pale-tipped as seen under a lens; superficially similar to O. deceptana, but actual ground color much paler, with faint transverse bands; an oblique band at a fourth way out and one at middle of costa, and a distincter subterminal bar, sometimes defined with distinct black striæ. Hind wing a little browner. 17-21 mm.

When most distinct, the darker gray shows the Exartema pattern, with subterminal and anal patches, and with median band rather narrow and deeply dentate on the outer side, but all in shades of ash gray.

New Jersey to Manitoba.

14. O. instrutana Clemens. Dark brown mixed with blackish, heavily overscaled with rusty ochre; antemedial fascia of a group of broken luteous striæ; postmedial band a double, lead-gray stria, strongly and broadly curved out over middle of wing, and a little concave toward inner margin. Discal dot pale, well within the postmedial bands. Three pairs of white striæ on costa outwardly; an oblique lead-gray line across apex and a confused patch at anal angle. Hind wing dark. 15 mm.

Common and generally distributed. Larva reported on clover and buckeye. Much like species of Cymolomia, but distinguished by the browner color with yellowish antemedial line and discal dot, and continuous gray postmedial band. Moth in August.

New York: Peru, Saranac Inn, Newport, Fentons (Lewis County), Batavia, Otto, Ithaca, Schenectady, Rhinebeck, New Windsor.

15. O. aspasiana McDunnough. Similar to O. instrutana. Dark olivaceous brown, with a double whitish antemedial fascia, and some whitish scaling at base. Medial dark area twice as wide at middle as at costa and inner margin, its outer boundary strongly bowed out at the middle, and scaled there with black. Subapical band extending from costa to outer margin, and containing a leadencolored patch opposite the cell; anal spot long, narrow, triangular, brown. Apex also brown, cut off by a silvery streak. 11 mm. (Not seen.)

Early July.

Ottawa, Ontario.

16. O. coruscana Clemens. Fore wing ochre yellow, marked with bands and

patches of brown (formed of blackish dusting on an ochre ground), the markings edged with series of silver-blue raised dots, which form continuous lines on the outer part of the wing. Basal markings confused; median band broad, not quite reaching the inner margin, which is yellow with a few black scales; the band deeply notched on its inner side, at the cell; and opposite cell and on **Cu**, on the outer side; a good-sized spot at anal angle; a larger band across apex, and a small apical spot. Fringe mixed, yellow, brown, and gray, quite variable. Hind wing fuscous brown; fringe white with a brown line in base; below dark fuscous, concolorous with fore wing. 16 mm. (*argyroëlana* Zeller).

Jupe and July.

Common and generally distributed south to Pennsylvania. New York: Fentons (Lewis County), Newport, Rock City (Cattaraugus County), Portage, Big Indian Valley, Albany, Poughkeepsie, New Windsor, New York City; Lynbrook, Long Island.

17. O. constellatana Zeller. A little larger on the average than O. coruscana; the black mottling covering the whole surface except on the apical third, and even there somewhat encroaching on the yellow ground, but leaving at least the subterminal patch distinct. Hind wing a little paler on the average; whitish below, strongly contrasting with fore wing. 20 mm.

With O. coruscana; not quite so common. New York: Otto, Rock City, Portage, Ithaca, McLean, Trenton Falls, Big Indian Valley, Schenectady.

18. **O. astrologana** Zeller. Ochre, almost evenly strigose with blackish on the basal two-thirds, becoming pale ochreous with light dusting on the apical third. Blue dots on basal part of wing evenly scattered; on apical part arranged in three oblique series, which indicate the outer side of the median fascia, and the edges of the subterminal fascia of *coruscana*, but with these fasciae no darker than the ground. Fringe largely ochreous. Hind wing a little lighter than that of *coruscana*, 16 mm.

Valve below with a rough tuberele in place of a ventral spine.

Illinois; Texas. New York: Mt Marey, Newcomb.

O. albiciliana approaches this group in the scattered blue spotting, but is much darker without any areas of clear yellow, and the hind wing is contrastingly paler at the base.

19. O. coronana Kearfott. Closely similar to small specimens of O. astrologana; fore wing almost wholly suffused with black, leaving only some broken ochre faseiæ in postmedial and subterminal regions, especially toward costa. 12 mm.

Jime.

Canada; Trenton Falls, New York.

20. 0. major Walsingham. Fuscous, similar in color and wing form to 0. constellatana, but with the pale portions duller. Fore wing with a dull luteous, erect antemedial fascia, and a parallel postmedial fascia running from two-thirds of the way out on costa to anal, both evenly edged with slightly brassy or silvery continuous lines. Outer half of costa with four pairs of pale stria, the first two connected with the postmedial fascia, the third free, and the fourth running down into a rather pale subterminal fascia, which runs out at the middle of the outer margin. Hind wing fuscous with pale fringe. 22 mm.

July.

Northern United States and Canada. New York: North Twin Brook (Mt. Marey).

This species is easily recognized by the faint continuous silvery or pale golden borders to the pale fascia, visible only in a favorable light. Eastern specimens are much darker than the typical form from the west coast.

21. 0. frigidana Packard. Basal two-thirds dark gray, with obscure and confluent blackish and fuscous striæ and some scattered brown scaling, the outer boundary oblique from costa at two-thirds way to apex, to just before anal angle, running in distinctly to end of cell, and excurved or irregular from there to inner margin. Outer third white, shaded with blue-gray, and with a few black scales. Hind wing light fuscous gray. 15-18 mm.

July.

Labrador and White Mountains, New Hampshire, to British Columbia.

The Labrador form has perceptibly smaller eyes than the Mt. Washington race, and has also more solidly dark palpi and a more contrastingly pale apical third of the fore wing.

22. O. apateticana McDunnough. Fore wing with base brown, with scattered white scales, making a more or less distinct dash through center of wing. Basal side of white outer part not quite sharply defined, but with a white hook in end of cell outlining a black dot, as in *O. capreana* and *scparatana*. Apex shaded with brown; a distinct black subapical bar. Anal spot partly cut off from dark base by whitish scaling. 15 mm. (*deceptana* McDunnough, not Kearfott). (Not seen).

June.

Meach Lake and Ottawa, Canada.

23. O. capreana Hübner. Similar to O. frigidana. but with basal part of fore wing mottled, rather than striate, the brown and black tending to gather in the antemedial and medial region, and the intermediate portions typically shaded with whitish, gathering in two squarish patches on basal half of costa and a vague shade in base of cell. Outer boundary of the blackish base rather less indented at end of cell than O. frigidana, where the white forms a comma-like hook, and regularly convex from there to inner margin. (funerca Meyrick).

Typical *capreana* is European, and appears always to have the pale costal areas.

Labrador to Duluth. Minnesota. and British Columbia.

24. **O. youngana** McDunnough. Similar to *O. capreana*, Costa of fore wing on basal half heavily shaded with white, leaving a squarish dark antemedial spot. White apical area strongly angled in at end of cell, and more or less defined with black, but not extending hooklike around a black dot. 19-20 mm.

July and early August.

Laurentians, Quebec, to New York. New York: Wilmington.

25. O. nimbatana Clemens. Base solidly dark, a mixture of bluish and brownish black; apical third white with three or four light gray striæ on costa and a stronger gray subterminal streak. Outer boundary of the dark base sometimes regularly excurved, sometimes with a tooth projecting upward at fold, running from a little beyond the middle of the costa to just before the anal angle; fringe pale; hind wing light gray, or whitish with light gray border. 16 mm. (H 48:24, as *Platymota flaredama*.)

June to August. Common. Larva on rose.

General, at least from New Jersey northward. New York: Ithaca, Albany, Rhinebeck, Scarsdale.

26. O. tertiana McDunnough. Base of fore wing purplish brown, dusted with bright brown, and mottled with darker; a few white antemedial scales. Boundary of pale outer half oblique and rather irregular. Outer part shaded with bright brown toward apex; with a dark subapical bar, the region between the inner margin and the subapical bar largely filled with a grayish shade. 16 mm. (Not seen.)

June.

Ottawa, Ontario.

27. O. separatana Kearfott. Fore wing with basal three-fifths, mixed gray and blackish; the outer boundary nearly straight from costa, rather before twothirds, to inner margin, rather beyond two-thirds; followed by blackish spots at end of cell and on inner margin. Outer third pale pink, shaded with light gray. Basal half, except on base of inner margin and on middle of costa, also sometimes shaded with pale pinkish gray. Hind wing fuscous gray. 13 mm. (O. dimidiana anet., not Sodofsky).

May, July, and August. Larva on rose and blackberry.

Hampton, New Hampshire, to Missonri and Arkansas. New York: Ithaca, Big Indian Valley.

28. O. montanana Kearfott. Similar to O. nimbatana, the outer boundary of the dark base erect, nearly straight, and somewhat diffuse. Pale portion duller and more clay colored. Ithaca, New York; North Carolina.

29. O. duplex Walsingham. Base light gray, striate with blackish; the outer boundary oblique on costal two-thirds, sharply bent in on A and ont again to the inner margin. Antemedial area white, a large mottled blackish patch resting on inner margin from middle almost to anal angle, more or less divided by irregu-lar whitish marks, a more or less distinct gray or blackish fascia extending from the upper side of the patch obliquely to the middle of the costa. Outer part of wing white, the apical region shaded with gray and sometimes some brown. usually leaving the costa, except at extreme apex, and a band extending to anal angle, white; more rarely, with only the costa white, and the dorsal region strigose or suffused with gray or brown. Hind wing light gray. 18-27 mm. June and July. Larva a leaf roller on poplar, forming a roll very much like

that of Anacampsis innocuella.

New York to Pennsylvania, and west to California. New York: Peru, Ilion. Portage, Ithaca.

This species is intermediate between the *nimbatana* and *bipartitana* groups and

is possibly nearer the latter, with which it is almost always confused in collections. 30. **0. albeolana** Zeller. White, marked with dark gray. An oblique bar on costa at a fourth way from base; a much larger rounded patch at middle of costa: a gray shade along inner margin to middle, defined by a black dash in base of fold, but outwardly diffuse or breaking up into striæ; a black dot at lower angle of cell, and some lighter gray shading on outer third of wing. Hind wing brownish gray. 18 mm. (hartmanniana auet., not Linnæus.)

June and July. Larva a leaf roller on white birch.

Maine to New Jersey. New York: Big Indian Valley, Karner, New Windsor.

31. O. roseomaculana Herrich-Schæffer. Fore wing deep blne-gray, mottled with black, with an irregular pink fascia of moderate breadth from third quarter of costal margin to outer margin at anal angle, sometimes broken into costal and anal patches by a gray shade. Hind wing mouse gray. 15 mm.

Larva on Pyrola.

Europe; Labrador.

32. O. costimaculana Fernald. Blackish, shaded with blue-gray and whitish. Base, and especially second fourth of costa, strongly dusted with whitish; a large square white patch on costa at two-thirds way out and some white striæ at anal angle. Hind wing gray. 11-14 mm.

Maine; Massachusetts; Manitoba.

33. O. chionosema Zeller. Blue-gray, somewhat shaded with duller gray. A large half-pear-shaped white patch from middle almost to apex of costa, with smooth clean-cut boundary; the dark part of the wing blackish around it. Hind wing dull gray. 15 mm.

Larva on thorn and apple.

New Brunswick, and Montreal, Quebec, to Virginia and Western Pennsylvania.

New York: Ithaca, Schenectady, Albany. 34. O. impudens Walsingham. Typically cream-white, rarely dull gray; base blackish, with outer boundary angled on cell; costa usually diffusely shaded with blackish to beyond middle, obscuring the antemedial fascia, and toward the apex more narrowly dark, cut with paired white striæ. A broad, typically white, antemedial fascia, in gray specimens distinctly composed of about four strix,

followed by an irregular olivaceous patch on dorsal half of wing, beyond which there is a blackish spot on end of cell, and one farther out on the dorsal margin; middle of costa also with a dark patch, obscured by the blackish shading. Fringe dark; hind wing pale in the typical form, gray in the dark phase. 12 mm.

June and July.

New Hampshire to North Carolina and Manitoba. New York: Newcomb, Rock City (Cattaraugus County), Batavia.

35. O. grisecalbana Walsingham. Whitish; costal edge dark gray, cut with white toward apex; outer edge blackish, preceded by white striæ. Base dark gray, except at costa; a pale rounded gray patch at middle of dorsal margin, and a smaller black one before hind angle, with some black striæ on the white ground between them. Hind wing dark. 15 mm. (Exartema).

June to August.

New Hampshire and Massachusetts to southern Ohio.

36. O. malachitana Zeller. Dull crimson, a little powdery, thorax and base of wings dull powdery gray. A large olive patch reaching from end of cell almost to base of wing and down to below A, striate with whitish on costa, and defined on the whole lower side by an evenly curved whitish line. A horizontal brown crescent from end of cell to outer margin, concave downward, and a spot at beginning of dorsal fringe. Fringes concolorous; hind wing mouse gray. 15 mm.

Larva on persimmon.

Virginia and Pennsylvania to Quincy, Illinois, and south.

37. O. interruptolineana Fernald. Wood-brown, shaded with dull rose and violet. Basal half light grayish toward costa, ending abruptly at a square brown median costal patch, and edged below with a sharply defined dark brown bar extending from inner margin near base a third way to apex. An irregular brown patch extending from cell opposite the median costal spot to middle of outer margin,

extending from cell opposite the median costal spot to middle of outer margin, constricted so as to appear composed of three partly fused patches, of which the first may fuse with the median costal spot, and the middle one may be hardly darker than the ground. Apex often brown, cut off by a pale stria. 13 mm. May; July and August. Larva on Vaccinium. New Hampshire to District of Columbia. 38. O. osmundana Fernald. Dull gray-brown. A large yellow-brown patch extending from middle of inner margin two-thirds way to costa; a dark brown patch, half as large, at anal angle; a subterminal bar and a short oblique bar from middle of costa, all fairly clean-cut and finely pale-edged. Hind wing mouse gray. 10 mm. gray. 10 mm.

Larva on Osmunda regalis; also reported from seeds of Ambrosia.

Maine; District of Columbia.

39. O. ochromediana Kearfott. Base chocolate brown; the outer boundary irregular and farther out on costa; median area even ochre yellow, twice as wide on inner margin as on costa; outer two-fifths chocolate brown, a little mottled and shaded with gray, but without clearly defined patches. Hind wing mouse gray. 10 mm.

July. One New Hampshire specimen is suffused with ochre yellow, with the usual marks just traceable.

New Hampshire to Pennsylvania.

40. O. murina Packard. Mouse color, dusted thickly with luteous scales. Head and thorax darker; fore wing with small obscure pale luteous costal spots, becoming paler toward apex; forming a very vague fascia from beyond middle of costa to anal angle; tringe also pale. Hind wings pale, dusky toward apex. 20 mm.

Of this species I have seen only a single fragmentary specimen. It appears to be a suffused form of the capreana group.

Straits of Belle Isle, Labrador. Reported as common.

41. O. hemidesma Zeller. Cu₂ leaving cell three-fourths way out, deep red-brown, with a slight pink iridescence. A short fragment of an oblique antemedial chocolate-brown fascia on inner margin; a complete narrow median fascia, slightly irregular on outer side, from middle of costa, the band narrowing and fading out toward inner margin. Hind wing fuscous brown. 16 mm. (Euchromia Zeller).

July. Larva webbing leaves and flower heads of Spiræa.

Maine to Pennsylvania and California.

42. O. hebesana Walker. Dull fuscous brown, somewhat mottled; the ground with a slight purple tint, the bands perceptibly darker, and not iridescent, mixed with some black scales. Base of the darker duller brown, its outer boundary irregularly excurved; median fascia narrow at costa and especially at inner margin; broad above the middle, where it extends out in two broad blunt teeth at upper and lower sides of cell. With traces of gray strize outwardly and usually a faint grayer subterminal patch; often with a suffused blackish area over middle of costa. 9-16 mm. (*Penthina fullerea* Riley.) June to end of September. Larva on Tigridia, verbena, Antirrhinum, Stachys,

pitcher plant, and iris; doubtless a general feeder. Dwarf specimens of this species have been determined as conditana Walsingham, which apparently is really purely a western species.

Maine to North Carolina and California. New York: Crosby (Yates County), Ithaca, Nassau, New Windsor.

43. O. dæckeana Kearfott. Deep shining blue-gray, almost black; the markings a mixture of deep dull brown and black; so far as traceable as in O. hebesana. Hind wing blackish. 16 mm.

June. Larva on pitcher plant. This may be a variant form of O. hebesana, which has been bred from the same food.

Southern New Jersey and Chicago, Illinois.

44. O. cyanana Murtfeldt. Brownish black, heavily scaled with shining deep blue. Base largely blue, with groups of blue scales and striæ before and beyond middle of costa; a large patch near anal angle, more than half the width of the wing, formed of two or three partly confluent blue spots or streaks; a few blue strim of anow. Hind wing blockich blue spots or streaks; a few blue striæ at apex. Hind wing blackish, paler toward base in male. 15 mm. May to July. Larva on rose.

Quebec to Missouri and Pennsylvania.

45. O. agilana Clemens. Blackish, irregularly scaled with yellow-brown and a little chocolate brown, leaving a vague darker shade at middle of costa, and an oblique subterminal band resting on middle of outer margin. A vertical leadgray stria, perpendicular to costa, resting on anal angle and extending half way to costa; also with some scattered lead gray dots and striæ on outer half of wing. Fringe dark gray, with black basal line; hind wing blackish with pale fringe, becoming pale on basal half. 10-12 mm.

May to July. Larva not rare in stem of Impatiens.

General. New York: Portage, Potter Swamp (Yates County), Ithaca.

46. O. auricapitana Walsingham. Similar to O. agilana; head and collar contrasting ochre-brown; fore wing with rather more ochre scaling, especially at apex. Lead-gray scaling coarser and sparser, the principal marks being oblique anteand postmedial costal bars, a broken bar across the apex, and the one at the anal angle (which is heavier than in O. agilana). No black line in fringe, but with a contrasting series of alternating black and yellow terminal dots.

June; August 1. New York and New Jersey. New York: Ithaca.

47. O. albiciliana Fernald. Crisply mottled with bright ochre yellow and black, more coarsely than in the coruscana group, which it approaches. Numerous coarse scattered blue spots, partly defined with yellow; two oblique streaks toward apex on costal half of wing, tending to slightly overlap two parallel erect streaks

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on dorsal half of wing. Fringe on costal half nearly white, with faint basal line; on dorsal half dark gray, cut with white at anal angle. Hind wing dull gray with broad white fringe; basal half whitish. 12 mm.

June and July. Larva purple, a leaf roller on Spiræa. Maine and Ontario to Pennsylvania.

48. O. nubilana Clemens. Dull fuscous, sometimes with a distinct reddish tint; with clearly defined but not contrasting darker brown markings. Basal markings slight, the most distinct a streak in fold; median band broad, deeply dentate on outer side, ending in a right-angled point between fold and A; a subterminal spot rather above the middle of wing; usually also a right triangle in shape, but sometimes oblique and narrower; a slight spot at anal angle. Outer margin of the fore wing often perceptibly concave. Hind wing concolorous. 15-20 mm.

June to August.

Distribution uncertain. There are one or two closely related undescribed species, one of which has been taken at Ithaca, besides the following.

49. O. infuscata Heinrich. Wing form as in O. constellatana, with outer margin of fore wing nearly straight, but apex more rounded than in O. nubilana; hind wing more or less trapezoidal, with outer margin sharply bent near middle. Dark fuscous brown, a little powdery looking. Median fascia vague and slightly darker; subterminal bar across apex darker, a little more distinct; some dark shading at apex; and a series of dark spots along costa, separated by short paired pale striæ, all inconspicuous. Hind wing concolorous, with slightly paler fringe. 22 mm.

Ithaca, New York.

30. PHÆCASIOPHORA Grote

(Sciaphila; Sericoris; Olethreutes, in part)

General structure and pattern as in Olethreutes. M_a and Cu, of hind wing connate. Hind tibia of male with dense masses of long hair above, and between spurs below; hind metatarsus with long dense hair above.

1. P. confixana Walker. Ground light olive-brown, rarely red-brown, striate with pale luteous; typically blackish with longitudinal pale striæ toward the base. Antemedial band a bundle of striæ, strongly excurved, and breaking up toward inner margin; postmedial usually of two stronger partly fused striæ, outwardly oblique to beyond the cell, tangent to a white spot at the end of the cell, then sharply curved and inwardly oblique to the fold, and curving out again to the inner margin; some paired subterminal striæ on costa, and blackish terminal dots. Hind wing fuscous brown. Fore wing sometimes, especially in the female, of a warmer red-brown tint, with the basal portion concolorous. Outer striæ of the antemedial, and basal striæ of the postmedial line meeting just above the inner margin, in clearly marked specimens separating the darker median area from the margin. 18 mm.

May to August.

New Hampshire to Virginia and western Pennsylvania. "New York" (Grote). 2. P. niveiguttana Grote. Hind tibia but little cularged. Ground red-brown, cut into irregular spots and patches by bands of pinkish ochreous edged with yellowish white; the brown areas defined with black. The most continuous brown area a median fascia, deeply notched on the outer side above the cell and on the inner side below the cell, and extended out at end of cell, where it con-tains a white discal bar. Subterminal patch usually large and forked below, patch at anal angle small. Hind wing mouse gray. 17 mm. (Olethreutcs auct.).

Late May. Larva on sassafras and hamamelis. Massachusetts to Florida. "New York." (Fernald).

31. CYMOLOMIA Lederer

(Exartema Clemens; Eccopsis, in part)

Similar to Olethreutcs; hind wing with a thickened and specialized folded lobe on inner margin (fig. 267) projecting beyond the general outline of the wing, but quite variable in size. Hind wing with II_2 strongly curved, but often well separated from M_3 and Cu_1 , which are connate at origin.

The numerous species as a whole show excellent genitalic characters, but they have not been fully worked out, and the present arrangement is tentative, awaiting Heinrich's revision of the Olethreutes group. On the whole a single species seems to prefer a single food-plant, but the rule is not absolute, and too little breeding has been done.

The pattern in most of the species is characteristic. The ground is of a somewhat lighter shade, usually becoming quite pale along the edges of the markings and tending to be dusted with paler scales and broken into striæ. It is usually slightly shining or iridescent. The markings are usually laid on in broader shades and have definite boundaries, often edged with dark; they are often shaded with yellow, or paler brown or gray, but rarely powdery, and hardly ever striate. The thorax and basal third usually show a confused mixture of the two colors, ending in an excurved antemedial line, which may run to the costa, or turn in to the base below the costa; the median band is typically broad in the middle, where it extends out in two long teeth along the upper and lower edges of the cell; and is usually deeply constricted or divided below the lower tooth, often cutting off a large dorsal spot. The upper tooth and costal portion are also often cut off as a separate oblique patch. There is an oblique subterminal patch or fascia, running from below the costa to the middle of the outer margin, and also a series of small costal spots beyond the median band, the second one often joining the subterminal fascia, and the fifth one apical. There is a patch at the anal angle, sometimes connected with the dorsal end of the median patch, especially if the latter is separate from the rest of the median band. The hind wing is almost constant, being mouse-gray with a pale fringe. The basal line of the fringe on the fore wing is continuous, often fading out toward the anal angle; the outer part is usually blackish at the apex and at the end of the subterminal fascia, and pale between and at the anal angle, but is often wholly dark.

The genus is almost wholly North American, with a couple of species in Europe.

Key to the species

1. Fore wing rather evenly marked with numerous wavy transverse lines. monetiferana.

- 1. Fore wing irregularly marked.
 - 2. Large smooth brown patches on basal fourth and near apex of costa.
 - 34. ferriferana.
 - 2. Without two large smooth brown patches.
 - 3. Thorax red-brown, contrasting with ground color of wings; rarely, with only the posterior tuft brown.

 - 4. Base of costa grayish.
 - 5. Fore wing with stem of cubitus and three or four veins subterminally
 - 5. Fore wing with a broad pale shade on Cu only; base of inner margin
 - 3. Thorax concolorous. 4. Most of wing almost evenly blackish, contrasting with the pale, clearly marked outer margin......10. concinnana var. terminana.

- 4. Basal three-fourths not suffused with blackish.
 - 5. With a distinct pale fascia of even width, formed of a group of striæ.
 - 6. Ground black-brown or yellow-brown, lightly shaded with pale brown; the antemedial fascia white and strongly contrasting. brown; the antemedian lastia matter of the antemedian lastia matter of the antemedian lastia matter of the antemedian as the astronomy of the

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- 7. Fascia running to anal angle, white, broader and strongly contrasting4. micantana.
- 6. Ground brown, with pale ochreous ante- and postmedial fasciæ.

6. Ground half olive and half blackish, with no yellow or brown. 9. appendicea.

- 6. Ground light brown with olivaceous tint, often heavily flecked or suffused with cream; the fasciæ hardly paler than the rest. 8. olivaceana.
- 5. Antemedial fascia concolorous with outer markings generally, or absent.
 - 6. Ground pure white, the borders irregularly blackish, except on outer part of inner margin and base of costa......21. malana. 6. Ground not white.
 - 7. A black apical dot and sometimes an oblique bar at basal angle the only contrasting markings......24. exoleta.
 - 7. Apical dot minute or absent.
 - 8. Dorsal margin contrastingly dark, the boundary sharply defined, at least on basal half.
 - 9. Dorsum black or brown, reaching to anal angle.. 29. nigrana.
 - 9. Dorsal shade gray, fading out at middle of wing.

30. clavana.

- 8. Dorsal margin not contrastingly dark.
 - 9. An oblique blackish patch at basal angle; the other markings
 - 9. Patch at basal angle, if present, no more conspicuous than costal markings.
 - 10. Basal patch not reaching costa, and sharply defined on upper side.
 - 11. With a narrow streak in base of cell Sc, separate from the basal patch.

12. Ground rusty orange, shot with purple.

- 28. nitidana. 12. Markings light olive on basal half, bright ochre and contrasting on apical half.....27. fagigemmeana.
- 12. Markings bright ochre, with a slight olive tint, ground paler and scaled with gray.

26. ferruginana.

- 12. Ground varying from ochreous to gray with slight olive tint (like C. permundana).
 - 13. Dorsal part of median fascia contrastingly smoother-looking and paler than other markings. 25. tiliana.

13. Dorsal part of median area not contrasting. 29. nigrana.

- 11. Patch simple and extending at least up to \mathbf{R} .
- 12. Median fascia blackish at costa only, ground pale. 22. punctana.
 - Median fascia blackish, as a whole.
 13. Fascia almost, but not quite, reaching inner margin.
 - 35. footiana. 13. Fascia enlarged into a squarish patch at inner margin, narrow above.....20.
- 10. With dark base reaching costa, or a dark antemedial line running through to costa.
 - 11. Median dorsal spot separate from rest of dorsal fascia, united with anal spot to form a thick bilobed patch. 10. concinnana (part), 11. foedana.
 - 11. Median dorsal spot separate from anal one, the median fascia usually complete.
 - 12. With strong black bars in the two teeth on the costal half of the median fascia, the dorsal half pale and obscure; ground pale.....12. atrodentana.
 - 12. Without contrasting black streaks across the median fascia.

 - 13. Subterminal spot not more prominent than other markings.
 - 14. Fringe rusty orange toward base, contrasting with the gray ground.....14. corylana.
 - 14. Fringe concolorous, or gray, black, and white. 14½. Costal half of median fascia blackish, contracting

- 15. Bands bright rusty ochre, much brighter than the gray ground17. sericorana.
- 15. Ground and bands two shades of deep redbrown; rough-looking13. zelleriana.
- 15. Nearly even deep rusty brown, interrupted by narrow streaks of the lead-gray ground.

7. quebecensis.

15. Leaden purple, with chocolate brown markings. brunneopurpurata.

- 15. General effect grayish brown, variable in shade.
- 16. A distinct fine dark wavy stria just before outer margin, from apical dot to anal angle, interrupted only by the oblique subterminal bar.....15. hippocastana.

16. No continuous stria close to outer margin.

17. Thorax and dark bands of fore wing, especially the antemedial one, mottled with little groups of clay-colored scales. 10. concinnana.

17. Thorax more evenly colored; dark bands more smoothly overlaid with the paler color, or evenly colored.

- Olive, with contrasting blackish base and middle of costa...9. appendicea.
 Fuscous, with a slight olive tint, the dorsal half of the median area only, contrastingly paler......25. tiliana. 18. Fuscous, more or less olivaceous, but
- with the markings mottled and all of one color...5, furfurana, 6. fraternana, 16. permundana, 18. sciotana.
- I. Palpi closely upturned to beyond vertex: not clavate; hind wing with a welldeveloped process but not lobed at 2d A; fore wing without Cymolomia pattern.

1. E. monetiferana Riley. Wood-brown, with clay-colored thick double striæ, separated by wood-brown lines, and edged with black; the bands basal, antemedial, postmedial and subterminal, all irregular and broken. Palpi with a gray longitu-dinal streak or a shade at base only and dot at base of second segment. 15-20 mm. End of May to June.

Western Pennsylvania to Alabama.

II. Palpi clavate and obliquely upturned, the second joint rather tufted at the end above and below; hind wing often notched at 2d A; always with more or less complete Cymolomia pattern.

* Not notched at 1st A and lobed below.

† Palc or gray antemedial fascia most prominent.

2. C. fasciatana Clemens. Black-brown; a transverse white antemedial fascia more or less distinctly made up of strix, nearly even in width, but widening to the costa, its inner boundary sharply bent above cell, and running in to costa, or, the costa, its inner boundary sharply bent above cell, and running in to costa, or, more rarely, continued to the base along R, leaving the base of the costa white. Median area broadly blackish, the anal spot fusing in with the median band, sepa-rated from it by an obscure lead-gray bar, visible only in certain lights; sub-terminal fascia lighter brown, its upper part fusing with median area; the region between these two and anal angle nearly filled by a vertical lead-gray bar. Apical region whitish, with a black apical dot. Line in fringe brown-black. 15 mm. June and July. Larva on Rumex.

Common and generally distributed. New York: Otto, Rock City (Cattaraugus County), Ithaca, New Windsor.

This species is easily confused with Olethreutes of the *fuscalbana* group, they are duller fuscous and black, and have the outer fascia much sharper and more contrasting.

3. C. rusticana McDunnough. Similar to C. fasciatana and the fuscalban i group of Olethreutes. Blackish ground overlaid here and there with ochre-yellow scales, but dominantly chocolate or umber brown; antemedial fascia composed of four clay-colored striæ, not nearly as white as in *C. fasciatana*; postmedial similar, running to anal angle; the spot of the ground color before it on the inner margin cut off from the median area by a lead-gray band. Median band two-toothed oppo-site cell with a deep notch between, as in *O. polluxana*, and the other Cymolomias. 15 mm.

Julv.

Northern States. New York: Peru, Ithaca.

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31/2. C. electrofusca Heinrich. Similar to C. rusticana; but with the darker portions evenly overlaid with ochre, giving a bright, tawny brown effect; the pale ground with a suggestion of pinkish iridescence.

End of May to July. Larva on sweet fern.

New Hampshire to New Jersey. New York: Ithaca.

4. C. micantana Kearfott. Similar to C. fasciatana. Ground nearly even dark brown, the gray band between the anal spot and the median fascia, and the gray band at the anal angle both replaced by powdery whitish fasciæ which join the whitish apical suffusion and separate the subterminal fascia widely from the median area.

June. Larva on dogwood.

Apparently more common northward. New York: Ithaca.

5. C. furfurana McDunnough. Color and appearance as in C. permundana; genitalia as in C. rusticana. Slightly more greenish olivaceous than C. permundana, with the pale areas more strigate with darker lines and dashes, the markings distinctly outlined with pale ochreous. Teeth of the postmedial band very long, the upper one almost touching the subterminal band, which does not reach the costa. 16 mm. (Not examined.)

June.

Vicinity of Ottawa, Canada.

6. C. fraternana McDunnough, another species with the appearance of *permundana* and genitalia of *rusticana*, is unknown to me.

†† Almost wholly brown.

7. C. quebecensis Heinrich (Kearfott ms.). Rather bright deep tawny brown (the "Vandyke brown" of Smith's glossary); evenly colored; antemedial band of two narrow dark lead-gray striæ; median, subterminal, and anal spots separated by an irregular narrow lead-gray marking, the subterminal fascia not separate from the costal region, and reaching the anal angle, with only a short stria between it and the concolorous brown apex. The usual costal striæ hardly visible; fringe and hind wing unusually dark, mouse gray. 13 mm.

Quebec; type only seen.

+++ With normal Exartema pattern; a part of the markings sometimes somewhat suffused. Dark base reaching costs, or gradually fading out toward costs, without definite upper boundary (Exartema).

8. C. olivaceana Fernald. Markings almost completely overlaid with slightly duller ochreous, the fascia narrower, silver-gray and cream, not contrasting; outer markings similar, a little narrower, yellower, and less contrasting than in C. *linei*fasciana. 15 mm.

June and July. To the naked eye the whole Cymolomia pattern is more distinct than in other members of this group.

General in distribution. New York: Ithaca, New Windsor (Morton). 9. C. appendicea Zeller. Ash gray with a slight olivaceous tint, unlike all the similar species of the fasciatana and permundana groups by the complete lack of any yellow or brown tint. Base and median fascia above fold and apical dot blackish, strongly contrasting with the olive subterminal and anal markings, and the olive or obscure dorsal part of the median fascia.

June. Larva on oak.

Kearfott considered this a synonym of versicolorana Clemens. McDunnough calls attention to the whitish base of the costa mentioned in the original description of versicolorana, and treats it as distinct. I have not seen versicolorana as identified by McDunnough. Presumably it belongs to the malana or nigrana groups as defined by the pattern.

Canada to North Carolina, west to the Pacific. New York: Rock City (Cattaraugus Country; aberration with pale antemedial fascia), Trenton Falls.

10. C. concinnana Clemens. Pattern typical. Median band interrupted below lower tooth and, rarely, above it also; its dorsal portion normally attached to the anal patch, forming an M-shaped spot. Ground light gray-brown, somewhat shaded with blue-gray and, heavily mottled with luteous, in little groups of scales, the markings rather broadly edged with the same luteous; markings shading from fuscous brown to blackish, not contrasting, except usually for the blackish upper end of the median fascia. Palpi shaded on outer side with fuscous. Fringes of fore wing generally darker than in *C. permundana*. 14 mm.

In variety terminana McDunnough (doxcana Kearfott ms.), the fore wing is blackish with contrasting pale margin.

June and July. Larva on blackberry.

New Jersey to Ohio. New York: Ithaca, West Farms (Angus).

C. mediopartita Heinrich is a similar species described from Virginia. It appears to lack the dark shade at the middle of the costa.

11. C. foedana Clemens has not been certainly recognized. It is purple-gray, marked with dark brown, like C. permundana, and is even smoother looking, not even showing the usual darkening at the middle of the costa, but has the pattern and size of C. concinnana. The costal spots are blackish, and the subterminal fascia is usually crossed with fine black lines. The fringe is dark, with a blackish line. 18 mm.

August. Something like this in the National Museum has been bred from alder in Virginia.

Toronto, Ontario?, Manitoba?, western Pennsylvania.

11½. C. brunneopurpurata Heinrich. Ground metallic leaden purple; markings rich dark brown, narrowly edged with white; pattern as in *C. permundana*. Antenna with blackish spots on upper side of first four or five joints. 14 mm. August. Larvæ on alder in July; tying the young terminal leaves. Pale green-

August. Larvæ on alder in July; tying the young terminal leaves. Pale greenish yellow, with segments transversely banded with pink. Head and cervical shield pale yellow.

Virginia.

12. C. atrodentana Fernald. Ground pale clay-color, somewhat striate with black. Markings light gray-brown, with a slight olivaceous tint; basal third heavily shaded, striatc, and mottled with blackish on the clay-colored ground. Median band normal, but very narrow, and olive at the inner margin; the olive part sometimes a separate spot, its upper half, and especially the two teeth which are long and narrow, suffused with black, contrasting.

13. C. zelleriana Fernald. Ground dark shining gray, heavily mottled with redbrown, of the same shade as the markings; basal area strongly mottled with the ground color. Markings orange-brown with crimson iridescence, the base strongly mottled, the outer markings rather even. Median fascia with inner side more irregular than usual, teeth blunt and rounded off, the lower one usually scaled with black. Outer markings extensive and tending to join, cutting the ground into irregular streaks and spots, but sometimes normal. 16 mm.

July and August. Larva crumpling leaves of white birch, in May (Kearfott).

East River, Connecticut; New Jersey; Pennsylvania. New York: Portage. 14. C. corylana Fernald. Light clay-color, with a slight greenish tint, dusted and shaded with light gray, obliterating the basal markings and most of the median fascia, the gray shade running from the base to the middle of the costa, leaving the base of the costa and the middle of the wing light. Dorsal part of median fascia, and anal and subterminal patches evener, pale dull gray, not at all contrasting, but finely defined with pale; last three costal dots dark, the apical one contrasting. Fringe with strong pinkish-to-rusty iridescence in basal half. 15 mm.

June to July. Larva on hazel.

New Hampshire to New Jersey, Manitoba, and Missouri. New York: Ilion, East Aurora.

15. C. hippocastana Kearfott. Luteous, rather striate with gray, a little Markings brown, shaded with blackish, well contrasted, and all about shining. Thorax with well-marked transverse banding; median band strongly alike. indented, rarely broken, narrow toward inner margin. Palpi of the lighter ground color, with base of second segment black. 20 mm.

June. Larva on buckeye, in May.

Distinguished by its coarse mottling, and tendency of the markings to break up, especially toward the base, as in Phæcasiophora.

Black Mountains, North Carolina

16. C. permundana Clemens. Markings normal. Ground shining gray, somewhat mottled with brown and luteous, the markings edged with luteous, decidedly smoother looking to the naked eye than C. concinnana; the general appearance dull brown. Markings rather dark brown, with a slight olivaceous or tawny tint; even, occasionally, with the lower tooth a little shaded with blackish. Thorax mottled, but less strikingly than in the last two species. 16 mm.

This name has been generally used to include forms not distinctively named, and even after the removal of Kearfott's recent species, is still a little heterogeneous; but the differences are probably of strain rather than of species. Sciotana, hippocastana, and merrickana could be united with this species with very little violence.

Larva on raspberry and Opulaster (Rosaceae) and on huckleberry. Moth in July.

Generally distributed and not rare. New York: Ithaca.

17. C. sericorana Walsingham. Shining gray, marked with bright rusty orange, the marks covering fully two-thirds of the wing surface, and finely pale-edged. Base mixed gray and orange, the area reaching costa. Some black scaling in lower tooth of median fascia and in subterminal and anal spots. Occasionally with the blackish dominant, and orange only in the median faseia. Hind wing and palpi as in C. permundana. 16 mm.

July. This species appears to intergrade with C. permundana.

New Jersey and Pennsylvania. New York: Ithaca.

18. C. sciotana Heinrich (Kearfott ms.). Outer part of second segment of palpus heavily dark-shaded. Ground dark blue-gray, nearly even, the markings narrowly pale-edged. Markings dark brownish gray, not strongly contrasting, nearly normal; the base as usual a mixture of the colors of ground and markings; the median band deeply constricted on cell, and normally broken by extensions of the luteus edging, also quite obsoure toward the inner margin, where the pale edging disappears. 20 mm.

June and July.

Southern Ohio.

C. subnubila Heinrich, a similar species from hazel, is not now before me. It is described from New Jersey and Maryland.

181/2. C. melanomesa Heinrich. Light wood-brown, the ground somewhat gray and with a slight pink iridescence. Markings essentially as in *C. permundana;* darker brown, the median fascia at the costa, the teeth, and the middle of the subterminal fascia, strongly suffused with blackish; the lower tooth especially prominent and almost cut off from the medial fascia. 16 mm.

The darkening of the upper part of the median fascia is the most conspicuous character for this species. It is superficially intermediate between *permundana* and merrickana, and according to Heinrich has genitalia of the permundana type. July.

Maine to New Jersey. New York: Ithaca. 19. C. merrickana Kearfott. Light fuscous gray, the markings hardly darker than the ground. Base fuscous, less mottled than usual, the antemedial line bent at a right angle over the cell, and strongly oblique to the costa and inner margin.

Medial fascia normally broken up into three separate spots, the costal one quadrate and brown, the medial one much farther out and shading into blackish, usually widely separated from the other two, and the dorsal one rounded, light fuscous, and not always reaching the inner margin. Apical portion of wing usually shaded with brighter brown, the subterminal fascia normal, chocolate brown heavily shaded with black, contrasting. 20 mm. June and July. Larva on witch-hazel.

The breaking up of the median fascia would group this form with corylana, which is smaller and paler, and has the outer reddish shade confined to the fringe. The records for "Black Mountains, North Carolina," were apparently based on a misidentification which has since been corrected, as there are no specimens so labelled in the series at New York.

General, west to Wisconsin and south to Virginia. New York: Croshy (Yates County), Ithaca, Portage (red variety).

++++ With normal Exartema pattern. Base of costa of the ground color, with a contrasting dark basal patch resting on inner margin and usually sharply defined on upper side (diffuse in exoleta).

-. Grayish brown, with a distinct pink tint; rather smoothly 20. C. colored. Markings black-brown and strongly contrasting. Basal third of ground color, with an oblique blackish bar running up from basal angle to middle of wing one-third way out; normally sending a spur back toward the base along the fold. Median fascia black-brown, the inner boundary sharply bent at Cu, where a pale line crosses it along the lower edge of the cell and separates it into a costal and a dorsal portion; lower tooth long and narrow, arising from the point where the two portions meet, typically connected narrowly with the lower portion, which is a large squarish patch. Anal patch very small and not contrasting; subterminal fascia normal. Palpus largely dark. (zelleriana Kearfott, not Fernald).

21. C. malana Fernald. Pure white, somewhat mottled with fuscous and a little blue-gray. Inner margin shaded with fuscous to two-thirds, with a roughlooking black bar resting on basal angle; middle of costa shaded with fuseous. with costal part of median fascia showing darker; apex and onter margin shaded with fuscous. Usual marks obsolete. A dark dot at end of cell. 12 mm.

Larva on apple and plum, in terminal buds and on leaves. Moth in July.

Apparently general, but rare and local. 22. C. punctana Walsingham. Light clay-color, shading into whitish; a large blackish patch resting on base of inner margin, extending up over fold and base of cell to middle of wing at a third of the way out; connected by a vague dark shade to a quadrate blackish patch on middle of costa, this patch representing the costal end of the median fascia and the region just beyond it on the costa. Middle of median fascia obsolete, dorsal part, and anal patch represented by rounded smooth pale olivaceous patches, hardly darker than the ground. Apical part of wing suffused with fuscous, obscuring the subterminal fascia. Hind wing distinctly lobed, but not nearly as strongly so as in *C. footiana*. Black dot at end of cell contrasting in the typical race from California; often obscure in eastern specimens.

June to July. Larva on Cornus.

California; southern Connecticut; New Jersey. New York: East Aurora. 23. C. cornana Heinrich (cornutana Kearfott ms.). Clay-color, a little duller and more even than in C. punctana. A strong oblique black bar from basal angle to fold a third of the way out; a black dot at angle of cell; median band indicated by a faint darker shade, not forming a defined patch on the inner margin; anal

and subterminal patches also obsolete, the latter, when at all traceable, round rather than barlike. Hind wing distinctly paler at base. 16 mm.

July. Larva on Cornus.

Northern New Jersey. New York: Ithaca.

24. C. exoleta Zeller. Clay-color, with a more or less distinct oblique fuscous shade resting on basal angle; powdery at edges; without definite boundary. A contrasting round black dot at apex, twice as large as the usual apical dot, and only preceded by very faint paired striæ. 16 mm.

June and July; late August. Larva reported from hazel and gooseberry.

Generally distributed and not rare. New York: Ithaca.

+++++ Markings of usual Cymolomia pattern; basal area formed of a patch below R, and a slender dark stripe in base of cell Sc, separated by a distinct pale line; costa pale

25. C. tiliana Heinrich (Kearfott ms.). Ground blue-gray with a slight violet tinge, flecked with brown, the markings slightly olivaceous brown and edged with elay-color, exactly as in *C. permundana*. Median patch usually with costal portion, including the long upper tooth, separated from the second tooth and dorsal portion. Fringe of hind wing white. 18-20 mm.

June and July. Larva on basswood.

The basal patch in this species is sometimes as in *C. permundana*, but it is distinguished by the even and finely white-edged dorsal part of the median fascia, normally contrastingly paler than the other markings.

Northern New Jersey to Missouri. New York: Ithaca.

26. C. ferruginana Riley. Bright ochre, with a slight olive tint; the ground a little grayer, paler, and more powdery than the markings, which are crisply paleoutlined. Median fascia broken into costal and median streaks and a dorsal patch, the latter fused with the anal patch. 15 mm.

Originally reported as bred from Hydrangea; but the type in the United States National Museum is labelled "plum."

New Jersey to Missouri.

This form and the next two are very possibly mere color-varieties of a single species.

27. C. fagigemmeana Chambers. Like C. ferruginana, and with similar pattern. Markings on basal half of wing light olive, including the dorsal half of the median fascia and anal spot; costal part of fascia and apical spot light ochre, contrasting. Ground grayish, also paler in the apical region.

Larva a bud worm on beech.

Pennsylvania; southern Ohio; Kentucky.

28. C. nitidana Clemens. Ground rose, strongly shot with violet; markings bright tawny brown. Markings mostly normal; lower end of median band usually separated and fused with anal patch to form a large M-shaped patch. Hind wing of male with anal angle markedly lobed, though less so than in *C. footiana.* July.

Pennsylvania and southern Ohio; doubtless widespread.

29. C. nigrana Heinrich. Ground varying from light pinkish brown to dark purple-gray, with dark chocolate brown to black-brown markings, clean-cut and strongly contrasting. Markings normal, mostly as in *permundana*; anal patch triangular, often fusing with lower end of the median fascia, which touches the inner margin beyond, leaving a narrow triangular area of the ground color along the inner margin beyond it. Dorsal third of wing frequently contrastingly darker. Hind wing markedly paler toward base. 20 mm.

July and August.

Generally distributed. New York: Geneva, Ithaca.

C. ornatana Kearfott, reported from New Jersey in Smith's list, is unknown to me. It is presumably a manuscript name, and equal to one of the other names in this group.

30. C. clavana Walker. Clay-color, a little powdery; a contrasting fuscous shade along whole costa, becoming wider and blackish at middle; a fuscous shade along basal two-thirds of inner margin, sharply limited at fold toward base, becoming diffuse outwardly. Outer margin less strongly dark-shaded, but with dark fringe. Usual markings obsolete, the subterminal fascia only defined, but very slightly darker than the ground. Thorax blackish, but less intensely so than in the preceding form. 16 mm.

July and early August. Quebec; New York; New Jersey; Kansas. New York: Ithaca.

††††† Thorax partly or wholly bright brown, contrasting with the greater part of the fore wing; the brown also appearing along costa, and at basal angle of fore wing. Usual markings not contrasting, and medial band incomplete, or wholly obliterated.

31. C. ochrisuffusana Heinrich (Kearfott manuscript). Ground light gray-brown, markings dark mustard yellow or ochreous toward base, brighter ochre-brown toward margin. Head and thorax yellow-brown. Basal patch large, diffuse toward costa, when most distinct forming an oblique shade resting on basal angle. Base of costa down to cell and out to two-fifths, the palest part of wing. Medial fascia obscure, represented by spots at costa, end of cell, and on fold; subterminal and anal patches normal. 18 mm.

June.

There is also a dull brown form distinguished by the pale base of costa and obsolescent median fascia.

Southern Ohio, Illinois, Kansas.

32. C. quadrifida Zeller. Thorax dull gray anteriorly, becoming bright brown (burnt sienna) on the tufts, or all brown. Fore wing pale dull gray, somewhat powdery, with strong pale veins, especially on disc, costa, apex, and inner margin; shaded with rusty brown; the costa cut with fine paired striæ, but the other normal markings lost. Hind wing whitish at base. 18 mm.

June and July. Larva on Cornus.

Massachusetts to Illinois. New York: Peru, Ithaca. 33. C. inornatana Clemens. Thorax mostly rich red-brown, usually showing the gray ground only at the base of the tegulæ. Fore wing gray, shaded with a vague paler streak through the middle of the wing and running into a large pale area at the anal angle. Veins not paler. Costa a little shaded here and there with brown; basal angle with a large fleck of bright brown, extending up to A. Usual markings often indicated only by partial pale outlines, the pale, excurved, and waved antemedial line most distinct. Anal patch often perceptibly paler than ground. Median band, when traceable, broken up into spots. 18 mm.

June to August. Larva on wild cherry.

New Jersey to western Pennsylvania. New York: Ithaca.

<i>ititit Antemedial patch contrasting, bright brown, with straight outer boundary; Cymolomia pattern lost.

34. C. ferriferana Walker. Ground powdery light gray; thorax and base of fore wing mahogany brown; the outer boundary straight, or nearly so, from cell to inner margin; usually bent in toward costa. A large trapezoidal brown patch on outer half of costa, half as wide as the wing, not quite reaching apex, but touching outer margin at middle, enclosing a gray apical triangle. Sometimes with the dark patches black-brown. 16 mm. (gratiosana Clemens).

June. Larva on Hydrangea.

Pennsylvania to North Carolina.

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** Hind wing of male markedly notched at 1st A, with a strong lobe below, whose apparent length is increased by its much longer fringe; outer margin more oblique, and apex more extended than usual.

35. C. footiana Fernald. Fore wing fuscous, marked with black-brown. Thorax fuscous; basal patch large, prominent, not reaching costa; medial band broad toward costa, and especially over cell, the teeth well marked but not very long, then abruptly narrowing, and usually not reaching inner margin; the lower end lighter, chocolate brown. Anal patch narrow and normal, or absent; subterminal bar normal. 20 mm.

June to August. Larva on witch-hazel.

New York to western Pennsylvania and Virginia. New York: Hemloek Lake, Ithaca.

32. BACTRA Stephens

Palpi large, broadly hairy, and beaklike, about as in Ancylis; eyes normal; thorax smooth. Fore wing smooth scaled; outer margin oblique, slightly arched, or straight. All veins separate and evenly spaced; \mathbf{M}_2 perceptibly separate from \mathbf{M}_a at origin; \mathbf{Cu}_2 nearly straight. Hind wing normal (fig. 268), \mathbf{R} and \mathbf{M}_1 stalked at origin, then usually approximate a third way to margin; space between \mathbf{M}_1 and \mathbf{M}_2 wide, as usual; \mathbf{M}_2 , \mathbf{M}_3 , and \mathbf{Cu}_1 close together at origin, but perceptibly spaced. No secondary sexual characters.

The genus is nearly world-wide, and perhaps, like Limnæcia, Australasian in origin. All the known larvæ are borers in rushes (Juncus), but seem to stray to other marsh plants.

1. B. verutana Zeller. Clay-color to dark wood-brown; costa and dorsal margin with fine blackish dots and striæ, and slightly heavier bars representing the antemedial and postmedial bands. Wing surface with some fuseous flecking, a blackish shade or spot in fold at one-third way out and one over lower angle of cell, often enclosing a pale discal dot. Frequently a pale longitudinal stripe below the costa. Terminal line usually continuous and perceptibly wavy, followed by a series of very short white scales in extreme base of fringe. Fringe often showing several parallel gray lines. Wing often longitudinally streaked toward margin. 9-15 mm. (*lanceolana* of American authors.)

May and June; August and September. Larva normally in Juneus, but reported also from Cyperus and Seirpus.

Indiana to North Carolina and south.

2. B. furfurana Haworth. Wings a little narrower, with more oblique outer margin; fore wing clay-color to wood-brown, the markings tawny to gray-brown; base dark, with outer boundary abruptly bent in cell; an irregular postmedial fascia, sharply bent on cell or lost below middle of wing, enclosing a pale or white discal dot; the markings frequently barely traceable. Usually with a spot at anal angle; almost always with a shade running to the apex. Terminal line as before. 10-16 mm.

Larva on Juneus and Seirpus.

General in distribution; also occurring in Europe. New York: Ithaca.

3. B. priapeia Heinrich. Larger than B. verutana and furfurana, costal stria fine and obscure, not much emphasized at middle; fore wing frequently all dark brown, typically with a longitudinal blackish shade from base to apex, leaving the anal angle contrastingly pale. Palpus with a fuscous spot on second segment and dark third segment. 16 mm.

July; September.

Woods Hole, Massachusetts, to Utah.

4. B. maiorina Heinrich. Similar to B. priapeia, but paler. Ochreous; head and front of thorax paler, the palpus whitish; fore wing with a fuscous shade

from base to apex; costa faintly barred with fuscous; a small white discal dot. Fringe dusted with fuscous. 13-20 mm.

June and July; September. Larva on Scirpus and grass. Virginia; Indiana; Utah.

33. POLYCHROSIS Ragonot

(Eudemis auct., Chrosis Guenée)

Similar to Olethreutes; no secondary sexual characters; hind wing (fig. 269) with M_a and Cu_1 distinctly separated at origin.

The moths are three-brooded, typically leaf rollers in habit, and also feed on flowers and fruits when in season. The moths of the various species are very similar to each other and difficult, sometimes impossible, to name without the foodplant. They show the Cymolomia pattern, but the base is more consistently of the ground color, with a brown antemedial fascia only; the median band has only the lower tooth on the outer side, and is broad, gradually narrowing to the inner margin; the upper tooth is usually replaced by a small separate spot. The subterminal patch is usually very large and broadly oval. It is doubtful if all these names will stand as species; some of them are perhaps rather food-varieties. Polychroses have also been bred from several other plants, but have not been described as distinct (figs. 269, 282).

Key to the species

- 1. Hind wing white, fuscous at apex only; fore wing irregularly mottled and shaded with white.....l. yaracana.
- 1. Hind wing fuscous, sometimes darker toward margin, or blackish; no white. 2. Subterminal patch evenly broad-elliptical, with only a few black scales on
 - 2. Subterminal patch separated from outer margin by a streak or patch of the
 - ground, except at upper side, where it sends a spur obliquely down to margin; more rarely, entirely separated from margin.
 - 3. Basal half of fore wing blue-gray, with narrow dark brown antemedial and broken basal lines only.
 - 4. Fringe light brown, concolorous with paler parts of wing.
 - 5. Pale edging of lines gathered to form a pale dot above tooth of
 - 9. liriodendrana.
 - 4. Fringe dark blue-gray, sometimes with light spots in its outer portion.
 - 5. Hind wing contrastingly pale, whitish, at base....10. cypripediana. 5. Hind wing dark gray, hardly paler at base.
 - 6. With a dark postmedial patch or short fascia......7. rhoifructana.
 - 6. With confused markings beyond upper end of median fascia, in
 - broad dark brown antemedial fascia.
 - 4. Dorsal third or half of pale antemedial fascia heavily shaded with light luteous, showing to naked eye as a pale spot.....4. carduana.
 - 4. Antemedial fascia of ground color, wholly gray, or with the luteous only finely edging it.
 - 5. Dorsal half of median fascia ochreous, a third as broad as width of wing, showing as a contrasting pale patch......5. aruncana.
 - 5. Median fascia all dark, or narrowing to a point at inner margin.

- 6. Median fascia evenly black-brown, the markings very narrowly and incompletely pale-edged (larva on Rosaceæ)

6. spiræifoliana.

Polychrosis has also been bred from clematis, Kalmia, Amorpha, raspberry, sassafras, rose, Circium, and Monotropa. Some of these records are doubtless of stray larvæ, but some may represent new species or strains.

1. **P. yaracana** Kearfott. Ground blue-gray; markings blackish, broadly edged with white; basal third largely blackish. Costal part of median fascia nearly black, contrasting, followed by a darker blue-gray area toward costa, forming a patch visible to the naked eye; dorsal part of fascia obliterated by a white spot. Hind wing white, the fuscous apical shade only reaching half way to cell. 9 mm.

May and June. Larva unknown.

New York to Cincinnati, Ohio. New York: Gowanda.

2. P. slingerlandana Kearfott. Shining dark blue-gray; markings black-brown, heavily edged with wood-brown, and more or less suffused with the same woodbrown, especially on outer two-thirds of dorsal half. Base markedly grayer, but still heavily marked. Apical spot blackish, fringe dark gray; hind wing blackish, with dark fringe. 9 mm.

Larva on Eupatorium, working especially in flowers and seed.

New Jersey, etc.

3. P. ambrosiana Kearfott. Dark blue-gray, marked with blackish; the marks finely pale-edged, but antemedial space not contrastingly pale; base less darkened than in *P. carduana*. Markings normal, subterminal patch well set back from the outer margin, which bears a strong blackish streak, usually joined at one point to the subterminal patch; postmedial costal patch normally broken into a couple of oblique striæ. 9 mm. (vernoniana Kearfott).

Larva in flowers and seeds of Ambrosia trifida, and on Vernonia.

New Hampshire to District of Columbia, and west to Kansas.

4. **P. carduana** Busck. Dark blue or purple-gray; markings normal, black-brown, finely pale-edged, with a well-marked postmedial costal patch. Base almost solidly blackish, contrasting with the autemedial area, which is light clay-color on the dorsal half; median band a little paler toward inner margin and of moderate width; subterminal patch large and dark; fringes and hind wing dark gray. 9-12 mm.

Larva in the heads of thistle; semisocial.

Maryland; Normal, Illinois.

5. P. aruncana Kearfott. Dark blue-gray, marked with dark brown, the markings mostly normal and finely pale-edged. Base dominantly blue-gray, but with a broad antemedial fascia. Median fascia broad and light wood-brown to ochreous on dorsal half; more than a fourth as broad as wing at dorsal margin. Postmedial patch broken into striæ; brown. Fringe and hind wing dark gray. 7 mm. (Specimens perhaps dwarfed by breeding.)

Larva on Aruncus (Spiræa).

Maryland.

6. P. spiræifoliana Heinrich (Kearfott ms.). Larva on Spiræa salicifolia. Markings normal, black-brown on a dark blue-gray ground, with only slight traces of pale edging. Terminal fascia heavy. Median fascia wholly black-brown, the tooth not very strong; subterminal patch very high, joining one of the black-brown costal striæ (which are all thick bars) and reaching nearly or quite to anal angle. Fringe and hind wing dark. 9 mm.

May; August.

New Hampshire to Pennsylvania.

7. P. rhoifructana Kearfott. Normal in markings with dark base, closely similar to *viteana*, but with a vertical postmedial bar resting on the costa, and dorsal half of the median fascia much browner than the costal end. 11 mm.

The first brood in May. Larva in seed heads of Rhus, and on Cornus and Kalmia.

Kennebunkport, Maine; Ohio. New York: near Rochester.

8. P. viteana Clemens. Normal in markings. Base blue-gray, with basal line hardly distinct, and antemedial brown band narrow and weak and broken. Markings dull umber brown, shaded with blackish, the subterminal patch dark brown or blackish, markedly darker than the ground. Markings finely edged with luteous, the dorsal end of the median fascia narrow, and somewhat suffused with luteous, or wood-brown. Costal region beyond median fascia with confused markings, more or less defined by pale longitudinal streaks; without a distinct costal patch but with a couple of minute costal dots instead; apical dot black-brown; fringe deep shining blue-gray, hind wing fuscous, a little paler at base. 10 mm.

The larva lives on grape. The first brood usually works in the flower clusters, the other two, in the grapes (for this reason it is called the "grapeberry moth"). Usually the larva works in two or three grapes, webbing them together, or attaching a leaf to the hole. It may also live as a case-bearer, eating out the Anthonomus gall and utilizing it as a case (Clarke).

Generally distributed and injurious. New York: Chautauqua County (Felt), Ithaca, Karner, and Orient Point.

P. botrana Schiffermüller is probably confined to the Old World. It is a much paler species, with the costal half of the antemedial fascia whitish, leaving a gray dorsal antemedial patch. American records are based on various species of this genus.

9. **P. liriodendrana** Kearfott. Similar to *P. viteana*; the outer part of the wing with pinkish suffusion on the ground, and the markings, including the apical dot, light brown. Postmedial costal spot well defined; fringe light wood-brown. Hind wing rather light, with whitish fringe. 9-12 mm. Larva on Liriodendron and magnolia.

New Jersey to District of Columbia.

The variety magnoliana Kearfott has the pale edging of the markings heavier and forming a distinct pale spot above the tooth of the median fascia. This variety occurs with the type on both food-plants.

10. P. cypripediana Kearfott. Basal half mostly dark blue-gray, with the antemedial band of moderate width; markings dull fuscous, dominant on outer part of wing, with fine pale edging, but hardly any pale shading. Markings normal. postmedial costal spot distinct, subterminal spot narrow, mostly well separated from outer margin. Hind wing dirty white, noticeably darker than in *P. yaracana*, with the fuscous apical shading extending in to the end of the cell. 10 mm.

Larva on Cypripedium.

Seen from Manitoba only, but doubtless to be found in our area where its foodplant is found.

Subfamily TORTRICINAE

The arrangement of this subfamily follows approximately that of Meyrick in the Genera Insectorum.

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WILLIAM T. M. FORBES

34. SPARGANOTHIS Hübner

(*Enopthera* Duponchel; *Enectra* Guenée, not Hübner; *Cenopis* Zeller, etc.; with *Platynota*, *Leptoria* Clemens, *Epagoge* auct.)

Front with a projecting tuft, leaving a more or less circular, naked or closescaled area covered by the palpi (fig. 240); which are beak-like and typically as long as head and thorax together. Fore wing (fig. 234) normally with some raised scales, becoming regular tufts in group Platynota; \mathbf{R}_1 and \mathbf{R}_5 forking over apex; \mathbf{Cu}_2 arising beyond middle of cell. Hind wing with a distinct tuft or fringe of hair-scales on base of \mathbf{Cu} , weaker than in most Eucosminæ; \mathbf{R} and \mathbf{M}_1 approximate, and often shortly stalked; \mathbf{M}_2 closely grouped with \mathbf{M}_3 and \mathbf{Cu}_1 , and widely separated from \mathbf{M}_1 .

Key to the species

1. Ground white.
2. Immaculate
2. A large brown patch
1. Ground lemon yellow.
2. A brown spot on middle of inner margin 20. albicaudana.
2. No such spot.
3. Retienlate with orange or red-brown.
4. Hind wing blackish10. lycopodiana.
4. Hind wing light gray9. sulfureana.
4. Hind wing pale orange or white.
5. Fore wing with narrow bands of red-brown14. reticulatana.
5. Fore wing with large areas of red-brown15. ferreana.
3. Not reticulate with orange, or with only a few brownish seales.
4. Hind wing ochreous white18. karacana.
4. Hind wing pure white.
5. A broad curved band, both ends of which run to costa17. groteana.
5. Narrow broken bands, or none
4. Hind wing gray.
5. Fore wing with three rusty stripes11. tristriata.
5. Fore wing with an oblique rusty band
1. Ground duller or darker.
2. Base of fore wing pale orange, outer part dark.
3. Base defined by a blackish band; with a blackish triangular apical
shade2. flavibasana.
3. Markings wholly diffuse; no blackishl. violaceana.
2. Hind wing dull orange or orange brown, or white suffused with orange.
3. With obscure dull purplish reticulation only, not contrasting.
22. testulana.
3. With well-marked dark reticulation
3. Light yellow, with light brown patches
3. Dull light tawny and darker brown; sexually dimorphic26. flavedana.
2. Dull orange, hind wing white
2. Base concolorous with medial or terminal region; hind wing not orange or
2. Base concolorous with medial or terminal region; hind wing not orange or pure white.
 Base concolorous with medial or terminal region; hind wing not orange or pure white. Ground bright vellow or orange.
 Base concolorous with medial or terminal region; hind wing not orange or pure white. Ground bright yellow or orange. Not reticulate; with oblique dark band
 Base concolorous with medial or terminal region; hind wing not orange or pure white. Ground bright yellow or orange. Not reticulate; with oblique dark band8. puritana. Yellow, reticulate with orange; or orange, reticulate with yellow.
 Base concolorous with medial or terminal region; hind wing not orange or pure white. Ground bright yellow or orange. Not reticulate; with oblique dark band
 Base concolorous with medial or terminal region; hind wing not orange or pure white. Ground bright yellow or orange. Not reticulate; with oblique dark band
 Base concolorous with medial or terminal region; hind wing not orange or pure white. Ground bright yellow or orange. Not reticulate; with oblique dark band

4. Evenly reticulate
3. Ground duller brown or gray.
4. Pale brown, with blurred antemedial and postmedial costal patches, or
immaculate in female
4. Very small; brown, with darker, rather even and sharply defined
bands13. diluticostana.
4. Dull dark gray, heavily tufted, and marked with blackish.
5. Blackish with luteous outer margin24. semiustana.
5. Blotched with whitish
5. No pale areas
4. Pale powdery grav.
5. With brown inner margin (Amorbia humerosana female).
5. No brown inner margin 6 cana

I. Fore wing with isolated scales or raised strictions only, sometimes practically smooth.

* Front normal, the enclosing tuft of long. incurved hairs, not quite regularly arranged (Sparganothis).

1. S. violaceana Robinson. Fore wing with basal two-fifths light yellow, edged with copper-red iridescence; the next two-fifths dull gray with purple iridescence; and the apex brown with green and red iridescence, all shading into each other. Hind wing dark. 20 mm. (Tortria, Enectra). May to July.

Quebec to Pennsylvania and British Columbia. New York: Peru, Rock City (Cattaraugus County), Ithaca, Schoharie, Karner (Forbes).

2. S. flavibasana Fernald. Base coppery, notched by a very large square fuscous patch a third way out on costa; most of wing dull brown; apex triangularly blackish, and also hind wing and fringe. Normally with the square costal patch connected by a fuscous band to inner margin near base, crossing the coppery area.

June. Larva on Lonicera.

Ottawa, Ontario; Illinois; Texas.

[Number 3 is vacant.]

4. S. irrorea Robinson. Typically light straw yellow, much less brilliant than the sulfureana group; with a large purple-brown patch beyond middle of inner margin, a smaller one opposite it on costa, often connected to it by a curved line, and costal edge and fringes dark; the patches not sharply defined. Ground often dotted with purple-brown. Hind wing pale. 22 mm.

June to early August.

Quebec to Pennsylvania and British Columbia. New York: Otto, Ithaca.

5. S. xanthoides Walker. Pale brown, typically reticulate and finely veined with dark brown; in variety breviornatana Clemens, with orange-tinted ground, and more heavily reticulate; male with two distinct costal patches of the brown, the basal one tending to become an oblique fascia across the wing, or to join a patch over Cu at the middle of the wing. Female with the patches obseure. Hind wing variable, dirty white to gray. Very near S. irrorca, and perhaps intergrading with it, but more buff-brown, with the patches less clean-cut on inner side when present, especially the lower one.

New Hampshire and Ottawa, Ontario, to Pennsylvania and British Columbia. New York: Newcomh, Portage. Potter Swamp (Yates County), Liberty, Big Indian Valley, Putnam County, New Windsor.

6. S. cana Robinson. Pale gray, with dark shading forming more or less distinet quadrate patches on base, before and beyond middle of costa, and rather beyond middle of inner margin; ground striated with black, the striation more definitely transverse than in A. humerosana, and defining the antemedial costal,

and the dorsal, patch with black. Hind wing slightly browner. 20mm. (Tortrix, Cenopis.)

This species is superficially very like Tortrix afflictana.

Pennsylvania. New York (Fernald).

7. S. caryæ Robinson. Cream color, regularly reticulate with red-brown; not forming oblique bands; hind wing lighter red-brown. 22 mm. June to September. Larva a general feeder.

Maine; Illinois; Missouri; Alabama; Texas. New York: Lewis County. 8. S. puritana Robinson. Bright ochre yellow, with an oblique even red-brown fascia from costa before middle of inner margin; a spot on costa at two-thirds and one at middle of outer margin. Fascia usually broken in the fold, especially in the large western race, vocaridorsana Kearfott. Expanse typically 18 mm.; the race vocardidorsaua often over 25 mm. (Crasia.)

End of June; August.

Quebec to Massachusetts and west. New York: Newport, North Creek, Wells, Ithaca, Trenton Falls, Karner.

9. S. sulfureana Clemens. Lemon yellow, more or less reticulate with orange; the reticulation sometimes almost completely lost; base of costa, two rust brown or black fasciæ meeting at middle of inner margin at an angle of 60°, the outer one often angulate or forked, and often an irregular outer margin; hind wing gray, or more or less shaded with orange, never pure white. Quite variable and tending strongly to break up into strains. 10-15 mm.; southern specimens (variety belfrageana Zeller) averaging very small. (Epagoge.)

Generally distributed, flying from June to September and the commonest of the genus. Larva a general feeder.

New York: Newport, Honeoye Falls, Buffalo, Portage, Ithaca, Big Indian Valley, Liberty, Pearl River, New Windsor; East New York, Long Island.

10. S. lycopodiana Kearfott. Markings similar to S. sulfureana, heavier, the red-brown often suffusing the whole outer half of the wing, and the bands centered with dark brown (as in some specimens of S. sulfureana). Hind wing dark monse-gray. 12-13 mm. (Epagoge.)

July and August. Larva on Lycopodium.

This form is fairly constant; it may be an extreme strain of S. sulfureana, but is quite distinct from its ordinary forms.

Ottawa, Ontario; Hampton, New Hampshire; Sebec Lake, Maine.

11. S. tristriata Kearfott. Lemon yellow, all margins, and a band through cell from base to outer margin, red-brown. Outer margin rarely yellow, merely reticulate with orange. Hind wing reddish gray. Fore wing rarely rusty orange, with grayish streaks.

September. Very rare. Minnesota; New York; Maryland.

S. bistriata Kearfott ranges north to North Carolina. It has two wide stripes, and a weak one on the inner margin, and a light brown hind wing.

** Front in male flattened or slightly concave, close-scaled, and produced into a point between the palpi; with the hair overhanging it all cut off even, forming a regular horseshoe-shaped tuft, so as to make the front appear deeply concave (fig. 240); female with the same structure less developed, much as in some males of the typical group (Cenopis).

12. S. saracana Kearfott. Very dull light ochre with a square light brown patch a third way out on costa, and a band from outer third of costa to beyond middle of inner margin, widening into patches at the ends and sometimes broken at the middle, or broken into three spots. Hind wing browner. 18 mm.

July. Larva on sassafras.

Essex County, New Jersey.

There is a closely related undescribed species from Philadelphia, Pennsylvania. 13. S. diluticostana Walsingham. Red-brown, with even, alternating bands of faint purplish and golden iridescence, defined by narrow lines which are not iridescent; the bands of moderate width toward outer margin, about as broad as long toward base. Hind wing reddish gray. 13-16 mm. (quercana Fernald). June and July. Larva on oak, cherry, and syringa.

New York to Texas. New York: Newark, Batavia, Otto, Ithaca. 14. S. reticulatana Clemens. Anterior half of tegulæ red-brown; front of thorax with a transverse orange bar. Fore wing lemon yellow, reticulate with orange, sometimes almost entirely orange; a dark band from costa at one quarter way out to beyond middle of inner margin, more oblique than in sulfureana; a costal patch at two-thirds way out, sending a forked line from its lower side; one leg of this line joining the lower end of the first band, and the other running to near the anal angle and usually fading out; margin with a dark stria. Hind wing white, typically suffused with orange. Very close to some forms of sulfureana, but apparently distinguishable by the outer patch on the costa and the pale hind wing; also rather like Eulia quadrifasciana, which has separated orange dots instead of reticulation. 15 mm. Not rare in July and August. Larva a general feeder.

The dark, dominantly orange form is typical, the yellow one is variety gracilana Walsingham.

Generally distributed. New York: Wilmington, Fentons (Lewis County), Hope, Newport, Otto, Hemlock Lake, Ithaca, Albany (variety gracilana), North Creek (typical).

15. S. ferreana Busck. Near S. reticulatana. Ground reticulate with rusty brown on pale straw yellow, suffused with brown except on basal third; a large semicircle on middle of costal margin, leaving a narrow oblique antemedial band and a terminal band. Hind wing white with ochreous tint.

Julv.

Ilion, New York.

16. S. niveana Walsingham. Reddish brown with bright purple iridescence; a basal patch, a conspicuous spot on middle of costa, and outer margin, white. Hind wing pale ochreous brown, with white margin and fringe. 23 mm.

The type is unique and is possibly an aberration of S. groteand.

Canada.

17. S. groteana Fernald. Lemon yellow; a large triangular patch on middle half of costa, extending down to fold and enclosing a semicircular yellow spot; sometimes with a series of brown postmedial striations. Rarely, with middle half of wing suffused with yellow-brown, except the costal spot, or with the brown reduced to a couple of striæ on costa, which tend to converge, unlike maculate forms of pettitana and karacana. 30 mm.

Western Pennsylvania to Manitoba.

18. S. karacana Kearfott. Lemon yellow with transverse water-lining like that of S. pettitana, of which it may be a maculate variety, but unlike S. groteana; a narrow brown fascia from one-third way out on costa to beyond middle of inner margin, and a patch on costa at two-thirds way out tending to send out lines toward inner margin. Normally not at all striate. Fore wing rarely suffused with tawny brown, showing the yellow only on the basal half of the inner margin, the outer margin, and a couple of spots toward base of costa. 20 mm.

Bayberry and scrub oak.

New Jersey. New York: Batavia.

19. S. pettitana Robinson. Pale lemon yellow, water-lined with raised scales, typically immaculate, sometimes with two more or less complete parallel oblique lines. Ground, rarely, pure white or shaded with pale brown. 20-27 mm. (H 48:35.)

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Common and general in distribution. Larva on various plants, especially maple. New York: Rochester, Batavia, Rock City (Cattaraugus County), Little Falls, Scheneetady, Pine Island; Great River, Long Island.

20. S. albicaudana Busek. Lemon yellow, mottled with slightly darker yellow; antemedial line fine, pinkish brown, starting from a small triangle on costa a third way out, and running to the inner angle of an irregular pinkish brown patch on middle of inner margin. Postmedial line two-thirds way out, irregular, but roughly parallel to antemedial as far as Cu_{s} , then fading out, or curving in and joining the outer end of the dersal spot. A little brownish reticulation ontwardly. Fringe pinkish brown. Hind wing cream-white, slightly shaded with orange. 15–18 mm. July. Larva on maple. This form is doubtless the extreme maculate variety

of S. pettitana.

Notch, Pennsylvania.

21. S. chambersana Kearfott. Dull orange, reticulate with dull red-brown, the reticulations gathering into a broken faseia from before middle of costa to beyond middle of outer margin, and a vague spot at end of cell. Hind wing white in male, orange in female. 20-25 mm.

June. Larva on Cereis. This form is superficially very close to S. xanthoides, but is distinguished most easily by the lack of gravish tint on the hind wing. It seems to fit rather better in group Cenopis than in typical Sparganothis. It also resembles 8. testulana, but is brighter, and has a paler hind wing.

Cincinnati, Ohio.

22. S. testulana Zeller. Bright orange, reticulate with light brown; with a slight purplish iridescence, not at all contrasting. Indications of an oblique median faseia, and a discal spot. Hind wing light orange. 16-22 mm.

June to August. Larva on oak and walnut.

Missouri to New Jersey and south.

II. Fore wing with heavy tufts and scale ridges and a conspicuous costal fold, the strongest tufts being two in the fold and one at the end of the cell. Eyes large in male (Platynota).

* Costal fold moderate.

23. S. idæusalis Walker. Ash gray, irregularly shaded with blackish and dull brown; the striæ markedly raised and black; male tending to have the outer margin paler, with two or three fine striæ; female rather paler on the basal half, out to an oblique black stria across the middle of the wing. Hind wing fuscous. 12-25 mm.; female averaging much larger. (Hypena Walker; scntana Clemens).

General in distribution; June to August; not rare. Larva on Solanum, goldenrod, clover, and other herbs; in a folded leaf or between two leaves; cutting the petiole of one leaf and feeding on it when withered and dry.

New York: North Creek, Otto, Itnaca, McLean, Big Indian Valley, New Windsor. Clove Valley, Long Island.

24. S. semiustana Walsingham. Basal part of wing almost solid blackish, contrasting with the luteous margin, which has two black striæ. Some yellow brown, mixed with the blackish base. Hind wing a little browner than in S. idæusalis.

Male only seen. Possibly a variety of S. idausalis.

New Hampshire; North Carolina.

25. S. exasperatana Zeller. Similar to S. ideusalis, but with costal half and outer margin of male whitish, except for a semicircular dull black patch on outer half of costa. Female with basal half whitish, and middle of costa also frosted with whitish; hind wing whitish in male, mouse gray in female. 12-15 mm.

July and August.

New Hampshire; Texas. New York: Ithaca.

** Costal fold extending beyond middle of wing.

26. S. flavedana Clemens. Male black-brown, with contrasting reddish-ochreous outer fourth; female red-brown, almost evenly marked, as in S. $id\alpha usalis$; hind wing red-brown. 10-20 mm.

Larva a general feeder.

Common and generally distributed. New York: Putnam County, New Windsor, New York City; Clove Valley, Staten Island, Jamaica, Long Island.

35. AMORBIA Clemens

(Hendecastema Walsingham)

Similar to Sparganothis; \mathbf{R}_4 and \mathbf{R}_5 united in male. running to costa; palpi exceeding head by less than twice the width of the eye, even in female. Larvæ leaf-rollers, like Sparganothis, etc., general feeders.

1. A. humerosana Clemens. Whitish gray, slightly mottled in two shades, and dotted with minute black scale-tufts; inner margin brownish. 21-30 mm. (H 48:23.)

Rather common in May and June.

General in distribution. New York: Ilion, Crosby (Yates County). Ithaca, McLean, Big Indian Valley, New Windsor, Long Island.

36. CELOSTATHMA Clemens

(Amphisa)

Male antennæ heavily bristled and fasciculate; palpi moderate, beaklike; no costal fold. Fore wings with \mathbf{R}_3 shortly stalked (fig. 235); \mathbf{Cu}_1 well separated from \mathbf{M}_3 and both \mathbf{Cu}_1 and \mathbf{Cu}_2 strongly curved; base of \mathbf{M} and of \mathbf{R}_{4+5} lost. Hind wing with \mathbf{R} and \mathbf{M}_1 stalked; \mathbf{M}_2 and \mathbf{M}_3 connate; \mathbf{Cu}_1 widely separated from \mathbf{M}_3 . An isolated genus, resembling both Sparganothis and Capua.

1. C. discopunctanum Clemens. Fore wing distinctly activated role 23. In isolated genus, resembling both Sparganothis and Capua.

 C. discopunctanum Clemens. Fore wing distinctly falcate; dull luteous, typically with outer and inner convex fine brown lines, somewhat outwardly oblique and shaded beyond with dull light brown; the inner line tending to break into costal and dorsal patches, the outer to become diffuse. Sometimes with large brown shades beyond the inner line at costa and inner margin, and beyond the outer line at the costa. 15 mm.

June to August; apparently two broods with maxima in July and in August. Larva on clover.

Quebec to Florida and west. New York: Newcomb, Ilion, Otto, Ithaca, Schenectady, Rhinebeck.

37. ADOXOPHYES Meyrick

(Capua, in part)

Male antennæ ciliate; palpi beaklike, the third joint moderate and downcurved; thorax with a slight crest behind; fore wing of male with large costal fold; \mathbf{R}_4 and \mathbf{R}_5 stalked, forked over apex; \mathbf{Cu}_1 arising well before angle of cell; hind wing in our species with \mathbf{R} and \mathbf{M}_1 stalked; \mathbf{M}_3 and \mathbf{Cu}_1 separate.

This genus is rather close to Sparganothis, both in structure and markings; but the peculiar frontal scaling is not suggested, the frontal vestiture being smooth.

1. A. furcatana Walker. Pale straw yellow, reticulate with golden brown; a light brown, oblique and quite irregular fascia from before middle of costa to before anal angle, about as wide as two interspaces on the cell; below which it

sends off a fine branch toward the basal angle; a second oblique and nearly even fascia from three-fourths way out on costa to anal angle, obscurely joining the first one below; well defined on inner side, but gradually shading into the pale apex. Hind wing white. 18. mm. June to September.

Mississippi Valley; east to western New York: Ithaca.

Eastern race. Paler; the ground between the reticulations very nearly white; oblique fascia much narrower, and darker brown, practically reduced to a couple of anastomosing lines, with a strong branch extending to inner margin about a third of the way out; outer fascia farther out, excurved and connected to the inner one by irregular brown reticulations.

Maine to western Pennsylvania.

38. ARGYROTOXA Stephens

(*Tortrix*, in part)

Similar to Tortrix, but with scale tufts on the fore wings, the tufts being only a little raised, the one at the end of the cell perhaps most prominent; R, running practically to the apex.

Argyrotoxa is closely related to Peronea, which differs only in \mathbf{R}_{5} running clearly to the costa; and which has a similar pupa. The species are superficially very close, but have perfectly distinct genitalia. Larvæ of the genus are known from rose, oak, huckleberry and other foods.

Key to the species

- 1. Fore wing with discal spot inconspicuous.
 - 2. Fore wing with a subterminal band.
 - 3. Subterminal band closely parallel to outer margin, leaving a narrow yellow marginl. bergmanniana. 3. Subterminal band more strongly curved, drawing away from margin
 - toward costa, where it is about as far from the apex as twice the width

1. A. bergmanniana Linnæus. Bright lemon yellow; a brown transverse band from a third of the way out on costa to before middle of inner margin; one from two-thirds of the way out on costa to before anal angle, these two breaking into orange reticulations below costa; a third contrasting dark band closely parallel to the outer margin, leaving a narrow yellow margin; the wing sometimes suffused with dull orange, leaving the margin and costal region pale. Hind wing white or vellowish. 10-12 mm.

Larva greenish white, with black head, cervical shield, and true legs; webbing together the terminal leaves of rose.

I am not quite sure this species really occurs in America, as the records may be based on specimens of albicomana, which is very close.

2. A. albicomana Clemens. Closely similar to A. bergmanniana, save for the greater curvature of the subterminal band.

Larva on rose.

Distribution uncertain, from confusion with the other species. New York: North Elba, Batavia, Rock City (Cattaraugus County), Ithaca, Albany, Rhinebeck. Crugers, New York City, Staten Island.

3. A. curvalana Kearfott. Rich tawny brown, leaving the base and outer margin yellow, and a conspicuous longitudinally oval discal spot at lower angle of cell.

Mississippi Valley.

4. A. semipurpurana Kearfott. Typically purplish gray, with the base, outer margin, and a patch at middle of costa yellow; varying to forms that are wholly yellow, with the usual märkings showing as a different shade of yellow, or merely as waterlining. Sometimes with whitish markings, or with orange bands.

Larva on rose and oak.

General, and somewhat commoner, as a rule, than A. albicomana. New York: Ithaca, common.

Variety dorsipurpurana Kearfott. Mostly pale yellow, but with a narrow contrasting blackish shade along inner margin, widening more or less distinctly into teeth ante- and postmedially.

With the typical form. New York: Ithaca.

39. PERONEA Curtis

(Acleris, Teras, Acalla, Rhacodia, Teleia Hübner; Cheimatophila Stephens; Alceris Fernald, in Dyar's List)

Similar to Tortrix; as shown by the pupa closely related to Argyrotoxa. Fore wing with all veins separate, \mathbf{R}_5 running to costa (fig. 236), \mathbf{Cu}_2 arising from well before middle of cell; the lower boundary of the cell beyond it strongly upcurved; hind wing with \mathbf{R} and \mathbf{M}_1 approximate, \mathbf{M}_2 low, \mathbf{M}_3 and \mathbf{Cu}_1 connate or shortly stalked. Thorax sometimes tufted; costa normally nearly straight, sometimes arched, notched at middle in subgenus **Rhacodia** (*emargana*, *effractana*), especially in the female.

Variation in some of the species is extreme; and the key can apply only to the more usual forms. There are two or more broods, the adult hibernating. For the larva see figure 244.

Key to the species

1. Fore wing with a broad notch in middle of costa, and apex strongly falcate.. 24. effractana.

1. Fore wing with costa not notched, or slightly so in female; in the latter case, with rounded apex.

2. Fore wing smooth-scaled.

3. No localized markings; the wing regularly spotted or reticulate, or plain.

1. minuta.

- 3. With a well-marked longitudinal stripe from base to apex...2. angusana.

2. Fore wing with tufts, at least small ones in fold and beyond middle of cell. 3. Head and front of thorax contrastingly blackish.

- 4. Five large tufts, two antemedial, two at middle of wing, and one near end of cell; the pale parts white.
 - 5. With contrasting white or whitish base or broad antemedial fascia.

3. Head and front of thorax not dark; or somewhat darkened (logiana), but shading into the ground color.

4. With a clean-cut diagonal scale-ridge clear across wing.

4. With the scale ridge abruptly offset inward at lower edge of cell, and less below Cu_2 , outward; ground white, lightly dusted with gray.

9. nigrolinea.

- 4. With separated tufts; when more than four in line, quite small and clean-cut.
 - Most distinct mark a triangular patch on middle of costa.
 Ground white.

7. With connected brown reticulation and brown terminal line.

12. americana.

- Ground gray or brown.
 Inner boundary of dark costal patch clean-cut; head light, ground
 - broadly grayish16. schalleriana. 7. Inner boundary of dark costal patch blurred; head deep brown.

15. logiana.

- 7. Head, body, and ground of wings concolorous, dark fawn color. 23. ferrugana.
- 7. Evenly brown, varying from pinkish cream to deep yellow brown,
- 5. Most distinct mark an oblique antemedian fascia or a longitudinal stripe.
 - 6. With a distinct tuft in cell Cu_1 , below those lying beyond middle of cell, and with outer part rough-scaled or with raised tufts.
 - 7. Tuft in cell Cu_1 in line with the others.

19. maculidorsana, 18. maccana.

- - 8. Ground pearl gray.....15. logiana.

 - 8. Ground reddish brown or reddish gray......23. ferrugana.
 - 8. Ground dull fuscous, or yellower, with red-brown markings. 15. logiana.

6. Outer part of wing, and usually cell Cu_1 , quite smoothly scaled. 17. hastiana.

I. Fore wing with smooth scaling, rounded apex, and convex to straight costa.

1. P. minuta Robinson. Typically orange, the scales shining, but reticulate with dull scales; often yellow, reticulate with orange.

The moth becomes darker later in the season, successive specimens passing through red and crimson to gray with only a few scattered red dots and striæ (variety cinderella Riley, probably the same as *oxycoccana* Packard). The area of the dull scaling becomes less at the same time and finally disappears. 9-18 mm.; the early yellow and orange specimens usually the smaller.

Several ill-defined broods, from June on; hibernating as imagoes in the form cinderella. Larva with yellow head; a leaf roller in habits, injurious to cranberry; also on apple.

Quebec to New Jersey, and perhaps generally distributed. New York: McLean. 2. P. angusana Fernald. Varying from light yellow to tawny brown, with inner margin below \mathbf{A} always grayish, and in dark specimens with other gray shades below the stripe. Stripe darker than ground, red-brown to blackish, running from middle of base to apex; more or less edged below, and often above, with slightly frayed and irregular white lines. 13-15 mm.

End of July to September.

Meach Lake, and Ottawa, Quebec; Massachusetts; New Jersey. New York: Ithaca.

3. P. fishiana Fernald. Dull gray, more or less distinctly dotted with brown. Two parallel fine lines from two-fifths and three-fifths way out on costa toward anal angle, each defined with slightly paler before and with a brown shade beyond. Hind wing whitish, more distinctly dotted with dark. 25 mm.

Montreal, Quebee; Franconia, New Hampshire; Maine.

II. Fore wing with more or less raised scale tufts; the wing rounded or with subfalcate membrane only; the costa smooth-scaled, or with rough bristling scales, tending to form tufts before and beyond middle, especially in female (Acleris)

4. P. subnivana Walker. White, sometimes tinted with cream, especially in female; a triangular dark patch resting on middle half of costa, tending to be broken up into three spots in very lightly marked specimens, but usually solid or with a small central pale spot; a yellow shade extending from it to middle of inner margin, at least in female. Hind wing gray, markedly darker than ground of fore wing. Costa of female concave and strongly rough-scaled both before and beyond the concavity. 13-15 mm. (peculiana Zeller, deflectana Robinson). In a variety the lower two-thirds of the wing is regularly spotted with brown between the veins.

The tufts are moderate but well marked; the costal notch of the female is the only definite difference between this species and the next. Larva on Vernonia (Cacalia).

Common and generally distributed in late fall and early spring.

New York: Wilmington, Ithaca, Albany.

5. P. cervinana Fernald. Similar, the ground pinkish cream to bright buffbrown, or buff-brown reticulate with white, the dark color covering a larger area than in spotted forms of P. subnivana. Hind wing paler, whitish, almost always paler than ground of fore wing. Wing form similar in the two sexes, without costal concavity or decided rough scaling; costal patch tending strongly to be weak below or to break into spots. 13-15 mm.

General all season; wintering as adult.

6. P. heindeliana Fernald. Ash gray, slightly shaded with brown below the triangular costal patch, which extends from a third to four-fifths way out on costa, and is spotted with black tufts (gallicolana Clemens).

Larva in "pine-cone" gall on willow. Illinois to Manitoba. New York: Ithaea (?) 7. P. trisignana Robinson. White, irregularly dusted with gray, appearing pale gray, with the costal triangle often indicated by three partly confluent light gray spots. Fore wing with antemedial tufts on cell and in fold, and with rough scaling which tends to form a very irregular and broken postmedial band. A scale ridge running obliquely almost across the wing from before middle of costa to beyond middle of inner margin, narrowly interrupted on the veins, but not offset on Cu₁ or Cu₂. 15-20 mm.

This form is doubtfully distinct from boscana Fabricius of Europe, which is not close to our so-called boscana (placidana Robinson). Larra fastening together two overlapping leaves of white birch, living in a small white tube. Pale dull green; head and cervical shield black; also with blackish prespiracular and subventral warts on prothorax.

This may be merely a gray winter form of P. niveana.

Montreal to New Jersey and western Pennsylvania. New York: Big Indian Valley, Bronx (New York City), Staten Island, Long Island.

8. **P.** niveana Fabricius. Pure white, with a few gray scales; the tufting as in P. trisignana, but interrupted in cell Cu_i . 20 mm.

Larva on bireh.

Europe; reported from Ontario.

9. P. nigrolinea Robinson. White, irregularly dusted with gray, often gathering into spots at middle of wing; typically, but not usually with a fine black longitudinal line from base to middle of wing. Tufts numerous, the outer ones forming parts of three series; the median series strongly broken at Cu_1 , and less so at Cu_2 . 25 mm.

This form is very probably a mere variant of P. trisignana.

Montreal, Quebec, to western Pennsylvania, Missouri, and British Columbia.

10. P. chalybeana Fernald. Tufts somewhat reduced, with wider spaces between them, but with the one in cell \mathbf{Cu}_1 in line with the others. Costa not so roughly scaled as in *P. hastiana*. Dull gray, normally with darker shades, especially with traces of the costal triangle; sometimes with costal triangle well-marked, and a heavy blackish patch at base of inner margin. Tufts often defined with black. 22 mm.

August; October to May.

Maine to western Pennsylvania. New York: Ithaca, McLean.

11. P. comandrana Fernald. Tufts almost obsolete. Pale brown, with obscure striation and a darker terminal line; costal patch normally strongly contrasting, dark brown, with central triangle of the ground color, rarely suffused and broken up. 13 mm.

June; November. Larva on *Comandra bellardi*; light green with whitish protuberances, and black head and cervical shield; in early June.

Massachusetts to Texas.

12. P. americana Fernald. Head reddish white. Fore wing white, reticulate with bright brown, the transverse bars of the reticulation much stronger than the veining, which is only partly traceable. Costal triangle bright brown, slightly suffused below, with pale center; fringe deep brown at base, yellow-brown outwardly, contrasting. Hind wing mouse gray. 15 mm.

October.

Ontario and New Hampshire to western Pennsylvania.

13. P. nivisellana Walsingham. Light gray; costal part toward base and a shade about the contrasting black triangle white; apex coppery orange, with a couple of black spots. Five large transverse tufts of whitish scales and several small ones, partly black. Markings variable in extent but unusually constant for the genus. 15 mm.

Larva on Cratægus.

Generally distributed, west to British Columbia and south to Pennsylvania. New York: Ithaca.

14. P. variegana Schiffermüller. Head grayish or black, contrasting with the white thorax; fore wing white or yellow at base; typically, with the antemedian tuft below cell blackish, and the area between it and the inner margin gray; in the only specimen seen from this region, nearly solid white. Outer half of wing contrastingly dark, yellowish to nearly black. 13 mm.

July; larva on various trees and shrubs.

British Columbia; Europe. New York: Clinton.

15. P. logiana, Schiffermüller. Typically pearl gray, with a contrasting brown costal triangle, which extends nearly to apex, and tends to run down a little along the outer series of tufts; a double point below. Tufts small. Fringe darker in pale specimens, concolorous with hind wing. Fore wing in variety germarana Frölich gray, shaded with brown, the costal triangle not contrasting with the other brown shades; in variety famula Zeller, dull fuscous with the triaugle harely traceable; the fuscous ground, under the lens, showing as gray, more or less flecked with red-brown. Fore wing in other specimens strongly shaded with yellow on the outer half (variety viburnana Clemens).

This complexly variable species is best determined by breeding from its food plant, Viburnum.

General in distribution. New York: Ithaca, Karner, New Windsor. **P. stadiana** Barnes and Busck is unknown to me. It is described as similar to P. viburnana, but smaller (16 mm.) with the costal triangle relatively smaller. ground light reddish brown, rather heavily overlaid toward the apex with dark brown, and terminal edge dusted with black. It was described from Ottawa, Canada.

16. P. schalleriana Linnæus. Light dull gray; with scattered small black tufts. Costal patch a clean-cut triangle, twice as long on outer as on inner side, and truncate parallel to the costa along the middle of the cell; costal patch bright brown, with some black scaling on its edges, and with two blackish bars in the fringe at its apex. 18-20 mm.

Two broods, August to April; June and July. Larva green with darker sidestripes, head and cervical shield yellow-brown, with blackish spots on sides of shield; in flowers and young leaves on Symphytum, Salix, and Vaccinium.

Quebec to Massachusetts, West to British Columbia; Europe.

17. P. hastiana Linnæus. Fore wing with moderate antemedial and medial tufts in cell and in fold, the medials about three-fourths way out in the cell; with no corresponding tuft in cell Cu_1 . Outer part of wing smooth scaled. Markings extremely variable, and of two principal types; either with a longitudinal stripe, which may run through the middle of the wing or rather toward the costa, or may be represented by a broad pale costal area; or with mainly transverse markings, either contrasting or obscure, of which the most distinct is an oblique fascia between the two pairs of tufts in the cell. Ground usually violet- to brownish-gray, duller than in P. variana. 20 mm. (celiana Robinson).

Larva much like P. schalleriana, on Vaccinium and Andromeda.

Generally distributed and not rare, but thoroughly confused in collections with various of the following species: New York: Hemlock Lake, Ithaca, McLean, Albany. (Variety inana Robinson occurs at New Windsor, and variety flavivittana Clemens is also reported from New York by Robinson. A black strain occurs at McLean.)

The principal named varieties are the following:

coronana Thunberg, with whitish antemedial fascia.

byringerana Hübner, with a yellowish fascia on a red-brown ground.

psorana Frœlich, variable in ground color, with striation on the veins, and traces of dark costal triangle.

autumnana Stephens, immaculate yellowish gray to dark brown, with the dark raised tufts only.

aquilana Hübner, dark, with veins finely pale and pale head and thorax.

albistriana Haworth, with fewer or no pale veins, and a pale dorsal stripe.

combustana Hübner, with yellow dorsal stripe. radiana Hübner, dark, with pale veins on dorsal half, pale yellowish on costal half, with traces of dark veins.

divisana Hübner, costal half contrastingly pale, white, or yellow, separated from the brown dorsal half by a dark line.

apiciana Hilbner, light brownish yellow, with a dark line from base to apex.

mayrana Hübner, the reverse of *apiciana*, being blackish brown, with a fine, cleancut yellow or white stripe from base to apex.

centrovittana Stephens, with the yellow or tawny longitudinal stripe crossing the typical transverse pattern; which is rather clean-cut and moderately contrasting in shades of brownish gray, as in the typical form.

The forms in this list were all described from Europe, but most or all are represented in America; besides these, the following were originally described from American specimens:

flavivittana Clemens, with a yellow dorsal stripe below the fold: it may be considered the representative of *combustana*.

perspicuana Robinson, pale gray with contrasting blackish base.

celiana Robinson, red-brown with normal pattern in violet-gray; and yellowish, rather than black, tufts.

- hudsoniana Walker, from west of Hudson Bay; described as blackish, with a paler quadrate patch in the cell and no other decided markings. (I have not seen it, and Kearfott notes that it is not a form of *hastiana*. A black form of *hastiana* occurs, however, at McLean, New York.)
- ptychogrammos Zeller, perhaps also a distinct species, described from Texas; dwarf (15 mm.); gray, with well-marked brown costal triangle; a longitudinal black streak in fold to three-fourths, and the black triangle edged inwardly with black.
- albilineana Kearfott, violet-gray, with a white stripe from base to apex, leaving a gray costal edge, and edged below with blackish. (I am not at all sure that this is a variety of *hastiana*. Very possibly the tufting is unstable and several of these nominal species should be united.)

18. P. maccana Treitschke. Tufts almost obsolete, but with the antemedial one in the fold distinct; a black dot or streak, sometimes slightly raised, in cell Cu_{ij} , almost directly below the lower tuft in cell; unlike the *hastiana* series, where the tuft or dot if present is well basad of the discal ones. Dull brown, somewhat shaded and frosted with pale violet-gray; usually very slightly, but in the only specimen seen from the United States, heavily, except for the dark base and the triangular patch on costa. Two fine, somewhat irregular, oblique lines from costa, the inner at two-thirds way out, much as in *P. fishiana*, which however is perfectly smooth-scaled. 20 mm.

Larva on Vaccinium and Ledum,

Europe; Winchendon, Massachusetts.

19. P. maculidorsana Clemens. Light violet-gray, shaded with brown beyond the principal row of tufts, which are all practically in line. A triangular dark patch on inner margin near base, with the black tuft in the fold at its apex, as in some specimens of *hastiana*. 17-20 mm.

Variety inana Robinson is all light dull gray, and wholly without contrasts.

New Hampshire and New Jersey to Nevada; apparently only the variety in Nevada.

20. P. variana Fernald. Outer tuft in fold directly below the outer discal ones, which are more erect than in the *hastiana* group; tuft or black dot in Cu_1 far basad of them, resulting in a long inward-projecting tooth on the outer boundary of the pale antemedian fascia. Markings white, gray, and black; or often with yellow, either as a basal patch or a longitudinal stripe; antemedial fascia broad, paler, often pure white, preceded and followed by narrow blackish irregular bands, which often are contrasting.

New York to British Columbia; the larva on spruce and often very common northward. New York: Mt. Whiteface, just below the tree line.

21. P. permutana Dupouchel. Reddish wood-brown, shaded with yellow, especially toward base of wing; with a blackish patch near base of inner margin; and shading at apex and beyond the antemedial fascia. Tufts strong, the one toward

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the base of the fold, practically filling the space from Cu to A; the outer tufts strong, well separated, and not very numerous; the one in cell Cu, typically much out of line with the others, but variable. Fragariana Kearfott applies to speci-mens with the tuft in cell Cu, practically in line. 15 mm. Larva on Prunus, rose, and strawberry; also reported from willow.

New Hampshire to British Columbia.

P. robinsoniana Kearfott appears from the tufting to be a variety of permutana. The inner margin is contrasting light yellow, widening to half the width of the wing at outer margin; the rest wood-brown, shading into black on basal half toward apex.

Specimens seen from New Hampshire, New Jersey, and Manitoba.

P. clemensiana Kearfott is another probable variety; it is suffused with black-ish except the large tuft on the disc and a patch at the anal angle, which are ochreous.

Hampton, new Hampshire.

P. flavivittana of Robinson, not Clemens, is another form of very different appearance; red-brown frosted with gray and white; with the inner margin from base to anal angle heavily dusted with white and marked by a fine white line.

Several other unnamed forms occur.

New Hampshire.

22. P. pulverosana Walker. Violet-gray, with a good many small groups of black scales, rather loosely and roughly scaled, but with small tufts. Tuft in Cu, hardly out of line, but moth distinguished from maculidorsana by its even gray color, with a contrasting yellow-brown spot in the end of the cell, defined inwardly by the black tufts. (When yellow-brown is present in maculidorsana, it is more extensive and diffuse.) 18 mm. (brewsteriana).

Massachusetts; Manitoba.

P. hypericana Ely is closely similar. The base before the tufts is grayish white, with a strong blackish patch on the base of the inner margin, covering the first tuft. The oblique series of tufts is normally followed by a strong yellow shade.

Larva on Hypericum.

Connecticut.

23. P. ferrugana Schiffermüller. Brown, often shaded with pale dirty gray or blackish; rarely, almost wholly of the pale gray, with scattered browner scales. Costal triangle never complete, but often marked in blackish. Tufts normal, small and clean-cut, usually of black and white scales; that in cell Cu, out of line. Distinguished from the hastiana series by its brown or brownish color, and lack of contrasting gray or yellow marks, from the comandrana group, by its less cleaneut costal triangle, and usually by its more decided yellow-brown tint. 15 mm.

Larva on white birch.

Semiannula Robinson is the extreme, light form; pale gray with only scattered brown scales. P. gallicolana Clemens, I am unable to place, but from its food habit it is more apt to be an earlier name for heindeliana Fernald than a synonym of this species; the description also agrees. If so, the supposed type at Philadelphia is incorrectly labelled.

Generally distributed. New York: Wilmington, Ithaca (United States National Museum), Taughannock.

III. Fore wing with costa strongly concave, even in male, and roughly scaled before and beyond the concavity in both sexes; apex strongly falcate. Tufts as in the preceding group. (Rhacodia).

24. P. effractana Frælich. Gray, more or less shaded with reddish; sometimes largely pale reddish. 18 mm.

August. Larva green, with yellow-brown head; on poplar, willow, and birch. Europe; Ottawa, Ontario; etc.

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39½. TORTRICODES Guenée

Similar to Peronea. Palpi somewhat weaker and down-curved; tongue weak in our species, typically obsolescent. Fore wing with \mathbf{R}_s running to apex. Hind wing with \mathbf{M}_a and \mathbf{Cu}_1 completely united; \mathbf{R} and \mathbf{M}_1 long-stalked.

with M_s and Cu, completely united; R and M_1 long-stalked. 1. T. fragariana Busck. Ash gray; base somewhat darker, its outer boundary strongly excurved and angled over cell; median fascia but little darker, strongly oblique, its inner boundary strongly extended in, in fold, and its outer boundary dentate. Hind wing smoky. Wings narrower and more lanceolate than in Peronea, though less so than in the type of Tortricodes.

August. Larva on strawberry.

New York to the Pacific coast. New York: Ithaca.

40. CNEPHASIA Curtis

(Sciaphila, etc.; Tortrix, Capua, in part)

Very close to Tortrix; fore wing usually more pointed, with extended apex and convex costa, smoothly scaled; palpi beaklike, often long. Venation like that of Tortrix, but with **R** and \mathbf{M}_i of hind wing stalked (fig. 239).

As defined here this is a heterogeneous group, apparently related to the lower members of Tortrix. Venational aberrations occur with \mathbf{R} and \mathbf{M}_1 free, but they can generally be identified by having longer and more pointed wings than Tortrix.

Key to the species

1. Silvery white, immaculate1. argent	l. argentana.	. Silvery white, immacula
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- 1. Tawny brown to olivaceous, with a more or less distinct oblique fascia.

1. C. argentana Clerck. No costal fold. Fore wing and thorax silver white, on a fuscous base, becoming fuscous gray when rubbed; hind wing duller and yellower. 25 mm. (Argyroptera Duponchel.)

July.

Europe; Truro, Nova Scotia; Quebec; Colorado, and west. New York: Trenton Falls.

2. C. osseana niveosana Packard. No costal fold. Cream white, somewhat shining; the wings less pointed then in A. argentana. 20 mm. (Ablabia Hübner).

The larva forms a tube under stones and trash, and feeds on various low plants. The moth flies in June.

Labrador to Alaska, and southward in the Rocky Mountains. New York: Lewis County (Hill, determined by Fernald).

3. C. mæschleriana Wocke. No costal fold; usually with \mathbf{R}_4 and \mathbf{R}_5 free; rarely, shortly stalked. Sometimes with \mathbf{R} and \mathbf{M}_4 of hind wing free. Light yellow, usually suffused with dull olive, red-brown, or reddish fuscous; with a strongly oblique brown median fascia and a spot on the costa near apex, obscure in suffused specimens. (gelidana Mæschler; Tortrix, Capua).

April; August.

Arctic; and on Mount Washington, New Hampshire, and other alpine summits.

4. C. listerana Kearfott. Brown with distinct pink iridescence; not strigose. Markings very slightly darker; golden brown with fine paler edging. A narrow excurved basal fascia; median fascia narrower on costa, covering whole outer third of inner margin; a convex subterminal fascia from costa almost to inner margin, much narrower toward inner margin. Hind wing chocolate brown. 10-12 mm. (*Phalonia* Kearfott).

July.

Nicholson, Pennsylvania.

5. C. peritana Clemens. Dull ochreous brown, sometimes with a slight greenish tinge, slightly striate with fuscous; a blackish band from costa at two-fifths way out to before anal angle, usually complete; and a small costal patch at four-fifths, somewhat extended down by dark striations. Female not always distinguishable from *C. virescana.* 8 mm. (*Smicrotes* Clemens).

May and June; September. Generally confused in collections with C. virescana. Probably general. New York: Newcomb, Otto, Ithaca, Ramapo, West Farms.

6. C. virescana Clemens. Similar; the male costal fold well marked, and contrasting brown; the costal edge more or less brown in female, and not striated. Ground color more constantly olivaceous; oblique fascia usually reduced to a costal patch in male, and rather diffuse in female. 13-17 mm. (Archips, Cacacia, Smicrotes).

The larva of a related California species eats mealy bugs (Pseudacoccus).

Common and generally distributed, a variety occurring in British Columbia. New York: Rock City (Cattaraugus County), Portage, Crosby (Yates County), Ithaca.

41. HARMOLOGA Meyrick

(Tortrix, in part)

Vestiture rough; the thorax with a low posterior tuft; pulpus upturned to middle of front, but rough above as well as below; clavate, with short third joint. No costal fold; wings loosely scaled, but without the definite raised scaling of Argyrotoxa and Peronea.

This genus was proposed by Meyrick to include a primitive group from New Zealand. Our species is aberrant in having M_a and Cu_i connate or stalked, and combines characters of Tortrix, Eulia, and Archips. 1. H. fumiferana Clemens. Mottled with clay-color and fuscous or reddish

1. H. fumiferana Clemens. Mottled with clay-color and fuscous or reddish brown, the western race strongly reddish; reticulate with black. Hind wing gray. 20-25 mm. (*nigridia* Robinson.)

The larva is the spruce bud worm, and is often injurious.

Generally distributed in the spruce zone; south to Massachusetts and northern New York in the East. New York: Common in the Adirondack and Allegany Highlands.

42. EULIA Hübner

(Lophoderus Stephens)

Like Tortrix, but more generally with convex costa; with a well-marked posterior tuft on thorax.

Key to the species

1. Ground cream.

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1. With complex clean-cut markings, including a broad oblique fascia.

- Pale triangular area resting on outer margin merely striate..4. gloverana.
 This pale area containing a large dark patch, or broken up.

 - 3. Dark markings largely deep purple-gray.....2. velutinana.

I. Hind wing with \mathbf{R} and \mathbf{M}_1 stalked (Eulia).

1. E. ministrana Linnæus. Light yellow, dusted with gray so as to appear olivaceous; with vague shaded markings of orange, red, and brown, the outer margin shading into red, the most prominent brown mark a patch on middle of inner margin. Hind wing gray. 20-25 mm. (*ferrugana* Hübner). June. Larva green with brown head; in a folded leaf on many kinds of trees

and shrubs; wintering full-grown and pupating in the spring. Generally distributed south to New York; Europe. New York: Summit and North Twin Brook (Mt. Marcy), North Elba, vicinity of Indian Pass (North Elba Township), Ithaca, Big Indian Valley. Variety subfasciana Stephens, which I have not seen from America, is more

evenly colored, darker brown, and frosted with whitish.

II. Hind wing with \mathbf{R} and \mathbf{M}_1 separate (Argyrotænia Stephens).

2. E. velutinana Walker. Ground nearly white, striate with gray and shaded with pale reddish; markings red-brown, striate with black, and shaded with purplegray. A fascia from middle of costa widening to inner margin and covering the outer third of it; a blackish triangle beyond it on costa and a more or less distinct oval gray patch in the pale apical area left by the two markings. Base varying from the ground color to dark gray; when dark, defined by a well-marked excurved antemedial line; the inner margin on the basal half often shaded with gray. 12-16 mm. (triferana Walker, lutosana, incertana Clemens).

April to October. Two or three broods, with maxima in May, July, and late August. Larva a leaf roller on almost any plants except conifers; and even reported from balsam fir.

Common and generally distributed. New York: Newark, Lewiston, Honeoye Falls, Ithaca, Delmar, Albany, New Windsor, Florida.

There is a closely related, but somewhat larger undescribed species without any brown scaling. It occurs at Ithaca in May.

3. E. pinatubana Kearfott. Similar in markings; but with the dark marks light tawny brown or Indian red; hardly ever with gray or purplish scaling. Outer costal patch connected with the outer oval spot, so as to divide the ground into an oblique postmedial fascia, and a terminal one narrowing toward inner margin. 10-15 mm. (politana auct.).

Seasons as in E. velutinana. Larva on pines, making a cylindrical tube out of a couple of clusters of needles and feeding on the tips of the same needles, deserting the tube and forming another when the needles are eaten down to about an inch long. Pupation in the last tube or in another formed for the purpose, but without a cocoon; the pupa easily falling out. General in distribution. New York: Peru, Portage, Ithaca. Probably common

all over the State but overlooked.

Politana Haworth has the ground dull red-brown, and the fascia breaking into a small costal and a large quadrate dorsal patch; and the larva is a general feeder on deciduous plants. It is probably not American.

4. E. gloverana Walsingham. Similar to E. velutinana, larger, wing rather more acute. Ground whitish, shaded with pale red, especially toward costa, and lightly and almost evenly striate with gray. Fascia light brown, even in width; costal

triangle also light brown; and brown base, when distinct, of the same shade the whole width of the wing. No subterminal dark area. 15 mm.

May. Probably mixed with females of E. velutinana in collections.

Winchendon, Massachusetts; Pacific States; not seen from the Central States.

New York: Lewis County. 5. E. mariana Fernald. Whitish, shaded with light gray and ochreous, with a large blackish triangle on outer two-thirds of costa; often breaking into two a large blacking triangle on outer two-thirds of costa, orient breaking into two spots, representing the fascia and spot of the preceding forms. 20 mm.
May. Larva on Vaccinium.
Nova Scotia to Florida. New York: Ilion, Jamestown, Ithaca, Ramapo.
6. E. juglandana Fernald. Wings rather broader; the outer half of the costa

June and July. Larva on hickory and Viburnum, in May. The thoracic tuft

is well marked, but very easily lost.

Quebec to Pennsylvania and Minnesota. New York: Batavia, Ithaca, New York City, Brooklyn.

7. E. quadrifasciana Fernald. Lemon yellow, heavily dotted with orange; with two narrow oblique red fasciæ, and outer margin often shaded with brownish red. Hind wing gray or orange. 15-20 mm.

May to July.

Quebec to Pennsylvania and Missouri. New York: Lewis County, Batavia, Albion, Ithaca.

8. E. quercifoliana Fitch. Cream yellow, dotted with brown; with two narrow oblique brown fasciæ, and a curved subterminal streak, sometimes connected by a streak in the discal fold to the outer oblique fascia. Hind wing white. 16-20 mm. (trifurculana Zeller; Tortrix).

June and July. Larva on oak and buckthorn, greenish white with amber-yellow head, and pale legs.

Quebec to Texas. New York: Ithaca, Big Indian Valley, Albany.

9. E. alisellana Robinson. Light brown with a series of more or less confluent cream-white patches all around the margin. Hind wing white. (H 48:39.) June.

Quebec to Illinois. New York: Ithaca, Bethlehem.

43. TORTRIX Linnæus

(Restricted; with Archips, etc., in part)

Head (fig. 243) smoother scaled than in Eulia; thorax smooth-scaled. with scutellum rounded out, but no posterior tuft. Palpi with second joint porrect, more or less clavate, rough-hairy on upper side, and third joint porrect or beaklike. Fore wing with all veins separate (fig. 237), Cu_1 arising close to M_3 , strongly curved, Cu_2 from middle of cell, nearly straight. Hind wing with R and M_1 approximate at origin, M_2 close to M_3 and Cu_1 , which are connate at origin. Cu₂ straight, arising two-thirds way out on cell. Costal fold usually absent.

The type of the genus (T. viridana Linnæus) is bright green, and, except in palpi and lack of costal fold, resembles Archips; our species are all aberrant in one way or another.

Grisea and fractivittana, placed in Tortrix by Meyrick, are included here in Archips; for peritana see Cnephasia, and for quercifoliana, see Eulia.

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Key to the species

1. Ash gray, obliquely banded and reticulate.....1. afflictana. 1. Yellowish.

2. Reticulate with dark and obliquely shaded.

2. Not reticulate.

3. Male with costal fold; shading on under side of fore wing and upper side of hind wing light brown; wings more acute.....3. clemensiana.

3. No costal fold; hind wing shaded with gray, or pure white; fore wing with costa more arched and apex blunter.....4. pallorana.

I. Male with a short costal fold, hardly reaching base of wing, and bearing a fanshaped tuft of long scales at middle, extending down over costal part of wing.

1. T. afflictana Walker. Pale gray, with base, an oblique fascia across middle, a costo-apical triangle and a vague shade on outer margin darker; defined with fine black striæ. Antemedial line angled at middle. Ground more or less distinctly reticulate. Hind wing slightly duller gray. 18-22 mm. (*fuscolineana*). May. Caterpillar on fir.

Newfoundland to Texas and California. New York: Ithaca, Albany, New Windsor.

2. T. packardiana Fernald. Fifth report United States Entomological Commission (Forest Insects) 849. A black, gray, and white species from fir, which I have not seen.

3. T. clemensiana Fernald. Fore wing long and narrow, nearly three times as long as wide, and falcate in the female. Fore wing plain straw yellow, in variety nervosana Kearfott lightly veined with gray. Hind wing and under side of fore wing white, shaded, at least in male, with pale brown, leaving the costa white. 16-20 mm.

Common in May and June, and in August. Female somewhat narrower-winged than pallorana.

Generally distributed. New York: Peru, Newport, Lewiston, Honeoye Falls, Rock City (Cattaraugus County), Potter Swamp (Yates County), Ithaca, Big Indian Valley, Utica, Albany, New Windsor, Staten Island, Brooklyn. Variety nervosana Kearfott occurs at Peru and Ithaca.

II. No costal fold.

4. **T. pallorana** Robinson. Similar to *T. clemensiana*, but without costal fold; on the average little larger, especially in female (30 mm.); hind wing a little graver in male, normally white in female; fore wing a little duller, not veined with gray. 20 to 25 or 30 mm. (Male *lata*, female *pallorana* Robinson.)

June and July. Larva on cherry, Silphium, and Verbena.

Massachusetts to Illinois, Missouri, and Texas. New York: Newport, Honeoye Falls, Batavia, Potter Swamp (Yates County), Ithaca, Albany.

5. T. alleniana Fernald. Light yellow-brown, crisply reticulate with brown; with a vague darker oblique fascia and a costo-apical patch, the fascia tending to be lost in the middle, and both clean-cut only on the costa; sometimes suffused with dull gray. 20-25 mm. (*Cacceia*).

July. Larva a general feeder.

This species is superficially an Archips, but the palpi are markedly beaklike, and the costal fold is completely absent.

Maine to Manitoba. New York: Peru, Newcomb.

44. ARCHIPS Hübner (Tentamen)

(Cacæcia Hübner; Ptycholoma Stephens; Tortrix, in part, etc.)

Similar to Tortrix, with the same venation (fig. 238) and vestiture; palpi closescaled except for a little loose hair near the tip of the second joint; second joint strongly upcurved (fig. 242) and concave on upper side; costal fold almost always present, but in several species reduced to a tiny pointed flap which does not fold over; absent in *conflictana*, extremely rudimentary in *purpurana*. \mathbf{R}_{4} and \mathbf{R}_{5} frequently short-stalked in *C. rileyana*, especially in the male.

Key to the species

1. Ground color white; marked with black15. dissitana.
1. Ground color not white.
2. Streaked longitudinally with blackish between veins
2. Transversely marked or spotted.
3. A triangular pure white costal patch
3. No pure white markings.
4. Costa cream colored, except for a patch toward apex. 23. melaleucana.
4. Whole wing cream colored, with faint ochre reticulations and mark-
ings
4. Ground dark, with a quadrate or triangular cream patch beyond
middle of costa, and usually other patches.
5. Ground yellow-brown, of a mixture of contrasting brown, yellow, and
cream scales
5. Ground gray-brown in broad shades17. mortuana.
5. Ground light red to pinkish gray, the costal spot triangular.
18. semiferana.
4. No cream color on fore wing; the quadrate patch beyond middle absent.
or of the ochre or darker ground color, and not contrasting with
the general surface of the wing.
5. Hind wing yellow or orange, at least on costal half.
6. Fore wing with sharply defined red-brown dots12. rileyana.
6. Fore wing with irregular powdery red-brown patches, or all rusty
orange
orange
at least at costa
6. Fore wing with a narrower blurred dark fascia2. obsoletana.
5. Hind wing not largely yellow or orange.
6. Males (frenulum simple).
7. Costal fold absent or represented by a slight roughness and
curvature of the costa.
8. Hind wing fuscous, concolorous with fore wing.
1. conflictana.
8. Hind wing pale.
9. Fore wing reddish, with a distinct outwardly oblique ante-
medial line; median band complete, contrasting; subter-
minal band continued by a shade and stria on its inner
side to anal angle4. parallela.
9. Fore wing reddish, with a dark spot at middle of costa and
other faint markings, or immaculate2. obsoletana.
9 Fore wing with distinct olivaceous tint the median band

above cell, and subterminal spot distinct; large.

3. zapulata.

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- 7. Costal fold represented by a small point ending in a tuft of hair-scales, folded back on wing.
 - 8. Hind wing dark gray, median fascia represented by a large
 - 8. Hind wing light on costal half. 9. Striation in antemedial region distinct, nearly transverse;
 - 9. Striation strongly excurved or angulate on the fold, often
 - lost in the dark purplish ground color; ground not usually
- 7. Costal fold slender, extending nearly to middle of wing and fringed below with long hair-scales. 8. Ground purple, banded contrastingly with chocolate brown.

9. infumatana.

- 8. Ground yellow-brown, with a dark gray patch in fork of Cu. 10. fervidana.
- 7. Costal fold broad and clean-cut.

8. Wings evenly mottled in two shades of rusty orange.

- 20. georgiana.
- 8. Fore wing with a dark bar from below middle of cell to basal angle.

6. Females (frenulum of three bristles).

- 7. Ground color gray, with clean-cut markings......21. grisea.
- 7. Ground brown, varying in shade.
 - 8. Hind wing gray (except costa).
 - 9. Fore wing with an oblique dark bar below middle of cell
 - 9. No oblique bar running to basal angle.
 - - 10. Fore wing purple with brown fascia....9. infumatana. 10. Fore wing olivaceous, with clean-cut, but not contrasting
 - 10. Fore wing yellow-brown, with patch in fork of Cu. 10. fervidana.
 - 10. Fore wing evenly mottled in two shades of rusty orange. 20. georgiana.
 - 8. Hind wing with dorsal half shaded with light gray.
 - 8. Hind wing pale.....2. obsoletana, 3. zapulata, 4. parallela.

I. No costal fold.

1. A. conflictana Walker. Dull light gray, velvety looking; all the marks, even the reticulation, a little diffuse; an obscure darker shade from costa just before middle, reaching inner margin two-thirds way out, with a paler patch following it; less oblique than usual. Indications of a postmedial patch and curved subterminal shade. Hind wing concolorous. 25-35 mm.

June and July.

Labrador to Alaska, south to New York and Utah. New York: Peru, Batavia, Ithaca; Woodhaven, Long Island.

2. A. obsoletana Walker. Fore wing in male with costa convex except near apex, not sinuous; outer margin convex; very pale fawn color, striate with darker; the

fasciæ represented by two darker fawn patches on costa. Hind wing dirty white. Female similar, the hind wing usually yellow, and fore wing almost immaculate. 20 mm. (vesperana Clemens, sanbornana Robinson, female transiturana Walker).

A. obsoletana has a tendency for the inner half of the hind wing to darken, especially in the female, as in rosaceana, but it may be distinguished by its lack of clearly defined fasciæ. The next two species are practically the same in structure and may not be distinct.

The larva is a general feeder and is sometimes injurious to strawberries. Moth common in August, rarer in July.

General in distribution. New York: Peru, Essex County, Ithaca, Albany, Putnam County, Katonah, Clove Valley, Staten Island. 3. A. zapulata Robinson. Larger; generally a little lighter and less yellow;

fascia clearly defined on the costa, and represented by a slight vague patch in fold. Hind wing entirely cream color in both sexes. 25 mm.

This is probably merely the western form of A. obsoletana. Many eastern specimens show the same color and pattern but they are never quite as large.

Illinois and west.

4. A. parallela Robinson. Costa of fore wing distinctly sinuous, as well as outer margin, forming a small lobe at apex. Fore wing dull reddish brown, the fascia contrasting blackish, but not always sharply defined; base also dark, or indicated by a dark antemedial fascia; postmedial spot strong. Hind wing with more or less gray, never yellow. 20-25 mm. (H. 48:31.) Superficially, very like forms of *purpurana*, but distinguishable in male by the

lack of costal fold.

Apparently general in distribution. New York: West Albany; Clove Valley, Staten Island.

II. Costal edge folded over in a small lobe, the fold not reaching the base, and ending in a more or less marked tuft of hair-scales.

5. A. purpurana Clemens. Costal fold with shorter hair-scales than in A. rosaceana. Varying from light dull wood-brown to umber brown, with a slight purple iridescence in male; frequently leaving the outer margin contrastingly light; less distinctly purple than infumatana; costa and outer margin more or less sinuous. extremely variable; fore wing with well-marked reticulation in pale phases, the brown antemedial line fine, and strongly excurved or bent at almost a right angle in the fold, the upper part parallel with the fascia, which is usually well defined and contrastingly dark except in the cell, where it is represented by the dark outlines only; in very pale or very dark specimens, the fascia is obsoletc. Sub-terminal patch not defined below, lying at the foot of the costal notch. Hind wing gray on dorsal half, cream above. Female similar, rarely so dark as to obscure the pattern; with the median band marked by strong striæ but only a little darker than the ground; costa outwardly and outer margin above very strongly concave, cutting off a round apical lobe. 23-30 mm. (gurgitana Robinson; lintneriana Grote).

Larva on geranium.

New Hampshire to latitude of New Jersey. New York: Fentons (Lewis County), Ithaca, Schoharie, vicinity of Albany.

6. A. fractivittana Clemens. Dull olivaceous, paler on costal half; fascia fairly well defined on costa, interrupted by a pale shade on cell; enlarged into a large patch below; postmedial costal patch small, distinct, and often with an ante-medial costal patch besides. Reticulation weak, often absent in male; hind wing gray with pale costs is male in costal patch to be absent in male and the second sec gray with pale costa, in male; in female with costal half pale. 18-28 mm. (fumosa Robinson.)

June.

Western Pennsylvania. New York: Ithaca, Trenton Falls.

7. A. rosaceana Harris. Costa and outer margin somewhat sinuous, but less so in male than in male of *A. purpurana*, and strikingly less so in female, the sexes being nearly alike in wing form. Strongly reticulate with brown; median fascia clean-cut but not strongly contrasting, postmedian patch likewise; basal striation transverse, or rarely, parallel to fascia, and not strongly curved. Hind wing of male cream, somewhat shaded toward the inner margin with gray; in the female, yellowish to bright orange, with the dorsal half contrasting gray. 20-28 mm. (H. 48:32.)

Common. Larva general on trees and shrubs; rarely, even on clover; sometimes injurious to apple.

General. New York: Common everywhere.

A. hewittana Busck is unknown to me, and from the description I cannot distinguish it from *A. rosaceana*. It was described from Nova Scotia.

111. Fold narrow, gradually fading out near middle of wing; fringed more or less with hair-scales below; generally forming a distinct flat tuft a third way out on costa.

8. A. rosana Linnæus. Wings slightly sinuous before apex; sexes similar. Dull brown, with slight greenish gloss; hind wing mouse gray, not contrasting; fascia not strongly oblique, well marked, sharply defined on inner side; postmedial patch continued by a strong subterminal stria toward anal angle. Striation sparse. 15-22 mm.

Not very common. Larva a general feeder.

Massachusetts to Missouri and Europe. New York: Thousand Islands, Albany, West Farms.

9. A. infumatana Zeller. Purple (as a strong iridescence on a blackish undercolor); base, a broad, well-defined fascia, and a subterminal streak reaching nearly to anal angle, contrasting dark chocolate brown; all often defined with light yellow brown; wing form of male like that of *A. rosana*; the female with more strongly sinuous costa. 18-25 mm.

July. Larva on hickory.

Connecticut to Missouri and Wisconsin.

10. A. fervidana Clemens. Brown on basal half, gray on outer half, with heavy and sparse ochre-yellow striæ, partly edged with cream, and, especially outward, centered with blackish, giving a general yellow-brown effect; dark patches on costa at middle and three-fourths way to apex, with yellow between them; a curved blackish patch below outer end of cell, defined on inner side. Hind wing gray. 15-22 mm. (paludana Robinson.)

July and August. Larva on oak; somewhat social; in small webs full of frass. Generally distributed. New York: Ithaca.

11. A. cerasivorana Fitch. Fore wing dull orange and when fresh, with purple iridescence; hind wing immaculate bright orange. Male irregularly spotted and speckled with dark red-brown, gathering into patches at middle and three-fourths way out on costa, and below end of cell; female with more reduced speckling, or nearly immaculate. 18-25 mm. (H. 48:21.)

Larva social; in a large nest of mixed silk and frass; on wild cherry; pupating in the nest. Imago in July.

Generally distributed. New York: Peru, Essex County, Fentons (Lewis County), Honeove Falls, Ithaca, Big Indian Valley, Schenectady, Albany, New Windsor. 12. A. rileyana Grote. \mathbf{R}_{s} and \mathbf{R}_{s} stalked in male and sometimes in female. Dull

12. A. rileyana Grote. $\hat{\mathbf{R}}_{4}$ and \mathbf{R}_{5} stalked in male and sometimes in female. Dull orange, tending to be suffused somewhat with grayish brown; with small deep orange-red spots; two larger spots on costa; a spot a third way out, and a pair two-thirds way out, on fold; and a broken series of small spots near outer margin; often with part of the spots wanting, or with additional spots at a third and two-

thirds way out, forming short series. Hind wing light orange. 20-28 mm. (Homona Meyrick.)

June. Larva social, on horse-chestnut in early June; also reported from hickory, walnut, snowberry, and Vernonia. Pennsylvania; North Carolina; Washington. 13. A. striana Fernald. Umber-brown, with a broad luteous ray over cell from

base to apex; a similar streak below A to three-fourths way out on inner margin; and broad, partly confluent streak on veins toward outer margin; blackish streaks between veins. Hind wing gray. 25 mm.

Rare.

Quebec; Ontario; New Hampshire; Manitoba.

IV. Fold not reaching middle of costa; rather thick, and, on the outer half, fringed heavily with hair-scales.

14. A. persicana Fitch. Basal half of fore wing orange-ochre; outer half bright brown, with a silver-white costal triangle, concave on the outer side. Margin sharply defined, contrasting pale yellow with a couple of brown striæ. 20 mm. (blandana Clemens).

June. Larva on various plants.

Maine to Manitoba; a race with a smaller silver spot, in British Columbia. New York: Peru, North Elha, Fentons (Lewis County), Newport, Rock City (Cattaraugus County), Ithaca, Liberty, DeBruce, Big Indian Valley, Ballston, Schenectady.

V. Fold well formed, but only one-fifth of the length of the wing, rough-scaled on the edge.

15. A. dissitana Grote. White, with irregular angular black patches, the two largest forming a fascia from costa to before anal angle, interrupted over the cell. 20-25 mm.

July. Apparently rare. Larva probably on conifers. Maine to Ohio. New York: Uphill Brook, Mt. Marcy, Ilion, Buffalo, Jamestown, Ithaca.

16. A. argyrospila Walker. Irregularly mottled with a mixture of cream, straw yellow, red-brown, and often black-brown, scales, leaving a series of cream-colored quadrate patches on costa, the one just beyond the middle being the largest and most constant. Hind wing mouse gray. 20 mm. (furvana Robinson, signatana Packard). (H 48:34.)

Larva a general feeder, and often injurious to apples, being the commonest of the many apple leaf-rollers. Moth in June, rarely with a few stragglers in July. Very common and generally distributed. New York: Common and general. 17. A. mortuana Kearfott. Brownish gray, irregularly mottled with cream and

darker gray or blackish; with two contrasting quadrate cream-white spots on middle of costa, and sometimes considerable cream striation on outer margin. 18 mm.

June. Possibly a variety of A. argyrospila, which is markedly variable.

Western Pennsylvania to Wisconsin. New York: Ithaca.

18. A. semiferana Walker. Light Indian red, more or less shaded with gray; somewhat mottled with straw yellow, especially on basal fourth; costa with strawyellow patches at one-fourth way out at middle, and before apex, the middle one neat and triangular, and the other two irregular and broken up into spots. Female generally paler. Very pale specimens shading into cream color on the inner margin and apparently intergrading with A. negundana. Hind wing concolorous. 20 mm.

Common, the larva a general feeder, but most common on maple and Negundo. Generally distributed. New York: Newport, Ithaca, Albany.

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19. A. negundana Dyar. Light cream-yellow with a slight pinkish tinge, faintly reticulated with ochreous; the cream pattern of A. semiferana more or less traceable.

The larva occurs with A. semiferana on Negundo and maple.

New Jersey; Utah; Manitoba; Florida; etc.

20. A. georgiana Walker, a bright orange species, striate with red-brown, with cream-colored areas between the striations, especially toward the costa, and dark gray hind wing; has been taken at Lakehurst and Whitesbog, New Jersey, in July.

VI. Fold reaching practically to middle of wing, broad, thick, and scaled along the edge.

21. A. grisea Robinson. Light ash gray, a little powdery. A contrasting black-ish streak from middle of lower side of cell to inner margin near base. A blackish patch in outer part of cell, extended as a yellow-brown shade toward anal angle; a contrasting blackish patch on costa at three-fourths way out and a dark brown oblique subterminal lunule. Hind wing gray. Female similar, larger, the dorsal markings all obsolete, or barely traceable, and with an additional short oblique blackish bar from costa at two-fifths way out. A 18 mm., 9 23 mm. (A brauniana Kearfott).

Larva on oak and Rudbeckia.

Maine to Missouri.

22. A. magnoliana Fernald. Typically, fawn colored, with purple reflections on disc; varying to dark purple-brown; the markings much as in A. grisea, cinnamonbrown, faintly outlined with white; outer part of wing dull whitish, the oblique subterminal streak smoky brown. Hind wing pale fuscous. 20 mm.

Larva on Magnolia acuminata.

New York: Rock City, Ithaca.

VII. Fold thick and broad on basal half, as in group V, but narrowly extended to middle of wing.

23. A. melaleucana Walker. Costal third or more cream white, cut by the blackish costal fold and a vague postmedial patch. Dorsal half of the base gray, mixed with yellow-brown, followed by a very large rounded patch of mixed blackbrown and purple-gray, which extends nearly to, anal angle, the dark marks all edged with yellow-brown. Outer margin usually cream, sometimes suffused with dark gray, especially in the female. Hind wing whitish, shaded with gray. In the darkest specimens the fore wing almost all suffused with gray, leaving only the middle of the costa cream color. 20 mm.

May and June. Larva on Trillium and Polygonatum. Very common south to New Jersey. New York: Peru, North Elba, Batavia, Vandalia, Rock City (Cattaraugus County), Portage, Ithaca, Big Indian Valley, Schuyler, New Windsor, Staten Island.

45. PANDEMIS Hübner

Palpi triangular, beak-like; thorax Appearance and venation as in Archips. smooth; male antennæ with a few joints near base of shaft partially fused and notched on inner side.

1. P. limitana Robinson. Pale reddish brown, with some fine darker and grayerbrown striations. Three fine, rather contrasting pale lines from before middle of costa, running to one-third and two-thirds way out on inner margin, and to anal angle; the base and the space between the two outer lines perceptibly darker. A semicircular or somewhat pear-shaped subterminal costal patch, which also has a Hind wing paler, with some light reddish brown striation fine pale outline. toward apex, becoming gray toward inner margin. 20 mm.

June; August. Larva a general feeder, especially on trees.

Generally distributed. New York: Ithaca, Schenectady.

2. P. lamprosana Robinson. Slightly yellower, the darker base, medial fascia, and spot near apex more contrastingly dark, and edged with dark instead of pale. Hind wing of male all dirty white.

Rarer than P. limitana. June to August. Quebec to Pennsylvania. New York: Ithaca, Schenectady, Albany, Staten Island. **P.** albaniana Walker, described from Hudson Bay, is unknown to me, and I cannot distinguish it, by the description, from *P. lamprosana*.

3. P. canadana Kearfott. Darker reddish brown, the three lines strongly waved, especially the inner two, which are practically straight in the other two species; the lines themselves not notably darker or paler, but serving as the boundaries of the contrasting, fuscous gray base, median fascia, and apical patch. Hind wing almost wholly gray.

July and August.

Quebec and Manitoba.

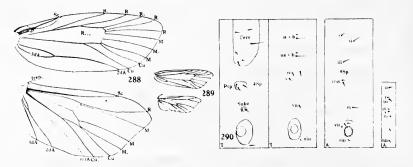
Family 29. PHALONIIDÆ

(Conchylidæ; Tortricidæ, in part)

Small, rarely medium-sized, moths. Head somewhat roughly scaled. Ocelli normally small, antennæ as in Tortricidæ; tongue weak or absent, when strongest about like the weakest tongues in the Tortricidæ. Palpi varying from moderate to long, porrect, beak-like; rough-scaled, the third segment long and porrect. Body small, scaled; thorax usually with a slight posterior tuft. Under side scaled. Legs scaled, the hind tibiæ with some rough scales or hair. Wings (figs. 288, 289) most often rather long and narrow; rounded; usually bent down at the apex. Usually with a distinct accessory cell, of Tineid type, and the base of M, when traceable, running obliquely across the cell as in the Tortricidæ. R. running either to costa or outer margin, stalked or free, but free in our species; the other veins all free. Cu, from three-fourths way out on cell or beyond; 1st A absent (distinguishing the family from the few Tortricidæ in which Cu₂ arises about three-fourths way out on the cell), 2d A with a large basal fork, with both branches equally strong. Hind wing trapezoidal; Sc normal, connected with **R** by a weak vein before middle of cell; **R** and \mathbf{M}_1 approximate or stalked; \mathbf{M}_2 free, rather close to \mathbf{M}_3 , which is separate or stalked with Cu_1 ; Cu_2 arising farther from end of cell than in fore wing; 1st A weak or absent, 2d A with a rather strong basal fork, 3d A very weak or absent.

Larvæ (fig. 290), so far as known, borers, or feeding in seeds, usually in herbaceous plants. Prothorax with seta delta (subdorsal posterior) below and in front of beta (addorsal posterior) and close to it; iv and v of abdomen in a nearly horizontal line; seventh segment of abdomen with vii of a single seta; prolegs poorly developed, with uniordinal hooks. Pupa of Phalonia approximating the Cossid type.

Vertex narrower than prothorax, very narrow on mid-dorsal line; front only slightly roughened, in *P. rana* with small eye spines; maxillary palpi not separate from tongue; indicated by a lobe at base of



FIGS. 288-290. PHALONIIDÆ

288, Hysterosia terminana, venation and costal fold; 289, Phalonia dubitana, venation; 290, Phalonia posterana (Europe), seta map of larva

tongue; labial palpi fully exposed but shorter than the tongue; tongue shorter than the fore legs. Antennæ long, not quite reaching to the end of the wings. Abdomen with two rows of spines on each segment from the first or second to the sixth or seventh; with single rows on the eighth to the tenth or the seventh and eighth segments; the anterior row on each segment heavier. Tenth segment without cremaster; obliquely truncate; sometimes with a circle of setæ around the edge; with the addorsal, subdorsal, and lateral pairs stronger, but knobbed rather than hooked; sometimes with a few weak setæ only; the edge also with angulations or short spines, of which a subdorsal pair is most prominent, or is alone present. Early stages studied mainly in Phalonia.

In Hysterosia birdana the pupa is rougher, rugose, and with the posterior portions of the movable segments shagreened with rough knobs. There are two rows of spines on the second to sixth segments of the abdomen inclusive, and one only on the seventh and eighth segments. The circle of spines at the end of the body is composed of three pairs on the supra-anal plate, and four ventral pairs. The setiferous tubercles are somewhat elevated. The front has an even-edged ventro-apical plate curving up at the ends and down at the base of the antennæ, and no eye spines. The maxillary palpi are more definitely indicated than in Phalonia.

This family is closely related to the Tortricidæ, but with definite points of distinction in both larva and imago.

Key to genera: imago

1. Fore wing with R_5 running to costa, or, rarely, to apex.

- 2. Hind wing with M_3 and Cu_1 separate; wings nearly smooth (fig. 289). 4. Phalonia.
- 2. Hind wing with M₃ and Cu₁ connate or stalked; wings with scale-tufts.

3. Commophila.

1. HYSTEROSIA Stephens)

(Idiographis Lederer)

Rather large, ample-winged moths with long beaklike palpi. Colors dull browns or grays; antemedial line running up obliquely from basal angle to cell at one-fourth way out, then obscure, or curving in to costa; postmedial line running obliquely across apex, with a dark shade beyond and a pale shade before it. Gen-eral wing surface obscurely mottled; hind wing light, distinctly reticulate.

Key to the species

- 1. Postmedial hand contrasting, clearly defined, and followed by a blackish shade; antemedial line less distinct.
 - 2. Head, except close to eyes, contrasting pale luteous......2. terminana.
 - 2. Head concolorous dark smoky grav; fore wing with slight purplish tint.

1. birdana.

- 1. Postmedial line inconspicuous or absent; if present, diffuse; antemedial line often distinct.
 - 2. Head luteous, fore wing shaded or dusted with yellowish, especially in fold. 3. Smaller; base of fore wing dull clay-color; ground gray-brown.

5. baracana.

- 3. Larger; base of fore wing bright ochre, followed by a yellow-brown
- 2. Head concolorous; fore wing all dark.
 - 3. With a clearly defined, but not pale-edged, dark apical patch....3. riscana. 3. Apex concolorous4. modestana.

1. H. birdana Busck. Dark fuscous gray, slightly purple-iridescent, dusted with black-brown. Head and palpi concolorous. Antemedial band fairly defined in inner side, showing as a dark shade not reaching costa. Postmedial band slightly concave outward, from costa at three-fourths way to apex toward outer margin just above anal angle, but not quite reaching margin; formed of a fine pale line followed by a heavy blackish shade. A fine black discal dot in a blackish shade, the dot partly defined with pale, not conspicuous. Hind wing much paler, reticulate with fuscous. Fore wing below dark, with reticulate costal edge; hind wing light, crisply reticulate, with a bar in outer part of discal fold. 22-27 mm.

August. Larva boring in roots of Helianthus, accompanying Papaipema nelita. New York to Delaware, especially along the coast. New York: Ithaca, Rye.

2. H. terminana Busck. Duller lighter fuscous, without purple tint; the mark-ings about as in *H. birdana*, but head, except above eyes, contrastingly pale. Beneath about as in *H. birdana*, but without the blackish har in the discal fold, a character which is perhaps more distinctive than the color of the head. 18-22 mm. (merrickana Kearfott).

Apparently general in distribution; usually reported as inopiana Haworth, which does not occur in the New World.

New York: Ithaca, Slaterville, Catskills (Pearsall).

3. H. riscana Kearfott. Ash gray, dusted with fuscous; antemedial band conspicuous, edged above with whitish, running from basal angle to upper side of cell, at one-third way out, fading out to costa. A distinct darker apical patch, its inner boundary running as in *H. birdana*, but not at all defined with pale or blackish. Hind wing hardly paler, reticulate; head and thorax hardly paler. Under side with costal edge of fore wing, and whole hind wing whitish, sharply reticulate with fuscous. 16 mm.

July. Types only seen.

Glenburn, Pennsylvania. 4. H. modestana Busck. Dull fuscous, mottled, but without any clean-cut powder-ing or markings, the two bands sometimes indicated by blurred dark shades. Discal dot black, in a pale spot. Hind wing grayish fuscous, not reticulate above, and obscurely so below. 16 mm. Maine; Iowa. New York: Ithaca. 5. H. baracana Busck. Fore wing a crisp mixture of fuscous, blue-gray, and

luteous or ochre-yellow, with the ochre often predominant in the fold and above the antemedial shade, where there is usually a yellowish streak. Postmedial band indicated by a gathering of the dark fuscous powdering, not sharply defined. Discal dot distinct, black. Hind wing pale, not distinctly reticulate. 12-16 mm. dot distinct, black. (tiscana Kearfott).

July and August.

Maine to Pennsylvania and Manitoba. New York: Ithaca.

6. H. cartwrightiana Kearfott. Wood-brown, shaded with bright tawny or deeper yellow-brown beyond the antemedial band. Basal area bright ochre, shaded with dark brown along costa, antemedial shade blackish, conspicuous; postmedial a vague darker area merely. 18-21 mm.

July.

New York to Manitoba. New York: Slaterville, Ithaca.

2. PHARMACIS Hübner

(Euxanthis Hübner)

Like Phalonia, except for the termination of R in the outer margin, and the connate or stalked M, and Cu, of the hind wing; superficially exactly like Phalonia.

Key to the species

1. Dull ochreous to gray.

	2	Ante	medial	fascia	complete,	rea	iching	inner	margin	 	2. sartana.
	2	Antei	medial	fascia	stopping	at	A			 l. b	imacula na .
1.	Br	ight	lemon	yellow	7					 3. 1	ritellinana.

1. P. bimaculana Robinson. Varying from light dull ochreous to dull grayish brown. Two thick dark brown crescents, of equal size, one running from below middle of costa to vein A at one-fourth way out, the other from below apex to lower angle of cell; both with the convex side down, and placed in paler areas. Hind wing light brown to mouse gray. 10-14 mm.

August and September.

Southern States north to St. Louis, Missouri. 2. P. sartana Hübner. Similar to P. bimaculana. Antemedial fascia running from middle of costa to inner margin at one-third way out, about twice as wide at inner margin as at costa, with irregularly excurved outer side.

August. I have seen no northern specimens.

Pennsylvania (type locality); Florida; Louisiana.

3. P. vitellinana Zeller. Lemon yellow, with transverse silver striæ or series of dots; ground sometimes shaded with red brown, or with the silver markings outlining red-brown bands; when most developed, with a brown oblique bar to the basal angle, one at right angles to the first from the middle of the costa to the inner margin at two-thirds way to the apex, and a couple of mixed brown and silver streaks across the apex. Hind wing dark gray, contrasting. 10-14 mm. June. Sometimes very common locally.

New York to Illinois and north. New York: Peru, Black Brook (Clinton County).

3. COMMOPHILA Hübner

(Phtheochroa, in part)

Similar to Hysterosia, but wings narrower, heavily scale-tufted; R, running to

costa. No costal fold (normally present in Hysterosia). 1. C. contrastana Kearfott. M_3 and Cu_1 of hind wing connate. Head and fore wing white; thorax and base and dorsal half of fore wing two-thirds way to apex, contrasting, black, mottled with blue-gray; with a couple of black dots. Hind wing pale brown, with white fringe. 20 mm.

End of May to June.

Connecticut; western Pennsylvania.

2. C. bana Kearfott. M_s and Cu_1 separate. Dark wood-brown, considerably scaled with black; the scaling as a whole rough, with larger rounded tufts at end of cell and before and beyond middle of A. Tufts more shining, slightly pinkish, and in most lights paler than the ground color. Hind wing a little duller and paler, with pale fringe. 13 mm. (Phalonia Kearfott).

Chicago, Illinois; in September. (The type has the date "June" corrected to "September," the description says "June.") Woods Hole, Massachusetts; in August.

4. PHALONIA Hübner

(Conchylis Treitschke; with Dapsilia, Eupæcilia, etc.)

Palpi beaklike but moderate, shorter than in Hysterosia and Commophila. Fore

wing rounded, rather narrow, with all veins separate and Commonnia. Fore apex; hind wing with \mathbf{R} and \mathbf{M}_1 stalked, \mathbf{M}_3 and \mathbf{C}_1 well separated. The species of Phalonia are often strongly marked in contrasting colors, some-times with silver, and are among the most brilliant of the Tortricids; but a few are dull-colored. The genus is already large, but probably a good many species Phalonia listerana Kearfott is a Cnephasia, P. vitellinana is a Pharmacis.

Key to the species

- 1. Fore wing with a sharply defined, nearly terminal darker band, more or less distinctly pale-edged; the median fascia rarely silver-edged, and in that case broken into spots.
 - 2. Yellow, with four red bands; the two outer ones joining toward the inner marginl. rutilana.
 - 2. Without red bands.
 - 3. Base solid dark, its outer boundary even, nearly straight, and only a little

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3. An oblique antemedial fascia resting on basal angle, or dark base with angulate outer boundary. 4. Apex sharp, forming an angle of over 60 degrees; ground, especially of antemedial area, normally shaded with yellow-brown, never dark. 11. obliquana. 4. Apex blunt, hardly forming an angle of 60 degrees, even in female. 5. Antemedial pale area toward dorsum evenly pale cream; markings also very pale.....10. promptana. 5. Antemedial area dusted with fuscous or shaded with brown. 12. angulatana, 13. bomonana. 1. With dark base, a median band, rarely broken, and subterminal darker fascia. all distinctly edged with silver (the silver very faint in P. labeculana). 1. With a fine contrasting dark subterminal line, parallel to the strongly oblique outer margin. 2. Expanse over 15 mm.; darker.....8. biscana. 2. Expanse under 15 mm.; paler.....variety giscana. 1. Terminal, or subterminal, dark band absent, or represented by scattered scales or a vague shade; markings rarely defined with silver. 2. A triangular or rounded dorsal postmedial patch, not reaching the cell; antemedial oblique fascia running up from basal angle to cell. 3. Ground dark gray.....14. rana. 2. An oblique antemedial fascia, extending up to cell, starting from inner margin at a point well beyond the basal angle, the upper part of the cell wholly of the ground color. 3. Ground pale gray; fascia extending into cell. 4. No costal spot opposite fascia; fascia mottled with yellow. 40. temcrana (cincinnatana). 4. Costal spot present; dark markings wholly gray......47. lavana. 3. Ground light yellow; fascia not entering cell. 4. A small costal spot opposite fascia.....15. smeathmanniana. 4. No costal spot; a series of subequal costal streaks......16. scissana. 2. A dark median fascia with clearly defined edge, reaching from costa to inner margin, sometimes interrupted part of the width of the cell. 3. Base and outer third of wing at least suffused with bright pink, median 3. No pink iridescence at base; sometimes a little outwardly in species with a blackish fascia. 4. Ground yellow; fascia H-shaped, enclosing squarish costal and dorsal patches; the costal ends of the fascia cream-colored, paler than the ground, but shading into the brown dorsal part......7. sublepidana. 4. Fascia cut by a pale spot; bar in cell nearly or completely interrupting it. 5. This spot yellow......17. fuscostrigana. 4. Fascia complete, simple, and wholly dark. 5. An oblique antemedial fascia resting on the basal angle; median fascia yellow-brown, followed by an oblique black streak. 22. romonana. 5. No oblique antemedial fascia; median fascia, when yellow-brown, not defined with black. 6. Median fascia brown, edged with blackish below; base yellow-

7. Fascia wholly yellow-brown, not very much darker than ground.

7. Fascia dark brown or heavily shaded with blackish.

8. Basal and outer thirds bright yellow; fascia brown.

9. Fascia black below the cell; outer third darker than base.

43. gunniana.

- 9. Fascia all brownish; basal and outer thirds concolorous. 23. hospes.
- 8. Base cream white; outer part dull ochre, at least on terminal margin and fringe, more or less distinctly shaded with pink. 29. hollandana, 30. bunteana.
- 8. Ground all luteous; a very small species with a distinct dark subterminal bar across apex......42. maiana.
- 8. Base straw yellow, except along costal edge; outer part gray and smooth looking to naked eye; without distinct subterminal bar.

24. straminoides.

- 8. Base with white ground but mottled with luteous and gray; outer part heavily mottled with gray.......47. lavana.
- 2. Fore wing with confused markings; with a more or less distinct diffuse median fascia; outer part of wing darker than base.
 - 3. Base pale gray, shading through chocolate brown to the pink apex.
 - 3. Base and outer part more or less suffused with pink......28. aurorana.
 - 3. Outer part of wing not suffused with pink.
 - 4. Fore wing heavily tufted, the tufts whitish on a brown ground.

(Commophila bana).

- 4. Fore wing smooth, or with small hardly raised tufts.
 - 5. Basal third or half of fore wing whitish, more or less contrasting.
 - 6. Base broadly ash gray to a fourth way out on costa, leaving a pale antemedial band.
 - Antemedial band partly white; markings gray....41. leguminana.
 Markings partly yellow-brown, and slightly brassy..2. labeculana.

6. Paler basal third extending almost to base of wing, at least in cell.

- - 8. Outer part with light black strize only......47. lavana. 8. Subterminal region with a large black patch.

44. hoffmanana.

8. Subterminal region with a .arge brown patch, concolorous with the paler scaling of the median fascia; markings all very indistinct and powdery......40. temerana.

5. Base of wing concolorous.

- 2. Ground whitish, strigose, with contrasting blackish base and outer margin,
- 2. Ground strigose, light gray, with a large square darker patch resting on middle of costa, extending more than halfway across wing 39. ednana.
- 2. Strigose, light gray on whitish, with a contrasting curved blackish fascia
- 2. Basal half contrastingly paler, and usually yellower, than apical half.

 - - landana, etc., see group with median fascia).
- 2. Base and thorax gray; medial area dark; outer part light pink or yellow. 3. Medial area brown, outer part pink; head concolorous.....27. viscana.
 - 3. Medial area blackish, outer part yellow-brown; head whitish, contrasting. 32. albidana.
- 2. Yellow without definite markings; inner margin and base paler than apex

1. P. rutilana Hübner. Bright yellow, with four even deep red bands, the outer-most not quite terminal, turning inwards at right angles just above inner margin and joining the third band; also often broadly connected with the third band at costa, enclosing a yellow patch. First band basal. Hind wing gray with a slight reddish tint. 9-12 mm.

June. Larva brownish yellow, with darker yellow-brown head and cervical shield. In a tubular web between needles of juniper, sometimes injurious. Maine; New Jersey; Europe. New York: Long Island (introduced about 1878).

2. P. labeculana Robinson. Cream white, with a few fuscous scales. Bands yellow-brown mottled with fuscous, sometimes dark-edged. Base fuscous, followed by a broad area of the white, extending two-fifths way out, and a little shining at the edges of the markings. Median fascia strongly constricted, and, almost cut in two at Cu₂; the lower part forming a more or less distinct triangular patch; outer part cream color, mottled with light and dark fuscous scales, the dark parts tending to form a subterminal costal spot and a nearly marginal fascia. Hind wing gray. 12 mm.

May. Described from a type at Philadelphia.

Pennsylvania; New Jersey. New York: Ramapo.

3. P. argentilimitana Robinson. Similar to P. labeculana; the bands distinctly brassy, with well-marked silver edging, much more shining than the ground, which is typically purer white. Median band broadly continuous across the cell and fold. 11 mm.

These two species are generally confused, and, in fact, may not be distinct. The type of argentilimitana is in very bad condition, leaving the possibility of incorrect identification.

Early September.

Pennsylvania to South Carolina and Missouri. New York: Ithaca.

4. P. interruptofasciata Robinson. Light ochreous, markings yellow-brown, the medial ones darker. Base brown more than a third way to apex, with clean-cut, slightly excurved outer boundary; median fascia represented by a large rounded dorsal spot and a less definite costal one, broadly separated from each other. A broad subterminal fascia from costa to dorsal margin, separated from the apex by less than its width, and gradually approaching the outer margin below. Disc sometimes shaded with gray. Hind wing mouse gray. 10-15 mm.

June and July.

Northern New Jersey and Pennsylvania.

5. P. aureana Busck. Pattern as in P. interruptofasciata; basal area blackish, heavily overlaid with olivaceous on lower half of wing, pale in cell and dark along costa; median spots dark brown, the dorsal one smaller than in P. interruptofasciata, subterminal band more curved, narrow, and silvery; preceded by a large blackish shade nearly connecting it with the median spots. 12 mm.

June; July.

The markings are obscure, giving the effect of a blackish species with a large yellowish patch covering the base, except the costa, and a narrow yellowish outer margin and fringe.

Western Pennsylvania.

6. P. lepidana Clemens. Deep ochre; median band a little darker, yellow-brown, sharply defined; extreme base light yellow. An antemedial light transverse line defining the median band, which is edged with somewhat powdery black scaling toward the inner margin; a similar edging on the outer side, both widening toward the costa, but narrower than in *P. sublepidana*. (The median band is oblique and even in width.) A pale line from lower angle of cell to before anal angle, cutting off a triangular spot of the ground color; this line often heavily dusted with blackish; a vertical subterminal line of blackish on a cream base, lying about in the middle of the yellow-brown outer region, its upper end sometimes joining the costal end of the postmedial line. 10 mm.

Intermediate between *P. sublepidana* and romonana. Described from the types at Philadelphia.

Pennsylvania.

7. P. sublepidana Kearfott. Fore wing bright ochre; broad antemedial and postmedial dark gray bands, both pale-edged, and becoming whitish toward costa; the two connected by a longitudinal band equally broad, in middle of wing, reducing the ground to costal and dorsal spots. Base and terminal margin and fringe much paler than median and subterminal areas. Hind wing fuscous brown. 9 mm.

July.

Northern New Jersey.

8. P. biscana Kearfott. Dull tawny brown to gray-brown, often dotted with cream or reticulate on a cream ground; a strongly oblique dark gray streak to basal angle, sometimes bent at right angles at middle of wing and faintly continued to costa; median line narrow, a little diffuse, bent at right angles at middle, where it may send lines to costa at three-fourths way out and to anal angle, forming an oblique cross; subterminal line narrow, parallel to outer margin below, running to apex, or, more rarely, fading out toward costa. Hind wing paler, dirty white. 15-23 mm.

July; August.

Maine to western Pennsylvania. New York: Peru.

Variety giscana Kearfott is dwarfed, with almost wholly cream-colored ground and markings heavy and dark gray, strongly contrasting. Intermediates are relatively common.

9. P. dorsimaculana Robinson. Luteous, with some scattered dark scales, especially with a dot or group of dots at lower angle of cell. A gray-brown streak extending obliquely up from basal angle, defined on outer side, but on inner side often gradually fading out toward base; a blackish irregular dorsal triangle on inner margin two-thirds way to apex; some black subterminal scales, normally grouped as a series of double dots between the veins, but rarely extending either to apex or dorsal margin. Fringe concolorous, hind wing grayish white to pure white. 10-16 mm. (angustana Clemens, not Hitbner).

July to September.

Nova Scotia; Quebec; Pennsylvania; and probably general. New York: Ithaca, Trenton Falls.

P. fernaldana Walsingham. Ground light yellow, including base of wings; markings yellow-brown; an angulate antemedial fascia; an oblique bar to middle of costa, not joining a large triangular patch on middle of inner margin; and confused outer markings, much as in *P. dorsimaculana*, but typically lighter. 17 mm. Described from the Pacific Coast and not certainly known from the East. Eastern records should probably be credited to *P. promptana*.

10. P. promptana Robinson. Pale cream, the markings darker; straw yellow with some black scales but not contrasting. Basal mark as in *P. dorsimaculana*, normally extending up from very close to base of wing on inner margin, or even touching base. Median triangle more rounded and not contrasting, with very few black scales; a well-marked oblique fascia from middle of costa two-thirds way to anal angle, extending down on the outer side of the dorsal triangle; a continuous subterminal band from near the apex almost to the inner margin, with considerable black scaling, which sometimes indicates the double dots of the preceding species. Hind wing nearly white. 15 mm. (angustana auct., not Clemens; dorsimaculana auct., in part).

June to August.

Canada to Texas. New York: Ithaca.

11. P. obliquana Kearfott. Light yellow, heavily shaded with yellow-brown, leaving well-defined pale edges to the markings, and a pale yellow fringe. Markings yellow-brown; an oblique streak in fold extending a third way to apex, ending in a point on Cu, strongly pale-edged; a large median dorsal triangle, a broad oblique fascia extending down from middle of costa, ending just beyond the tip of the dorsal triangle, more or less broken by pale shades on the veins. A costal triangle three-fourths way to apex and a yellow terminal band, finely edged with golden yellow; base of inner margin shaded with yellow-brown, without golden edging. Hind wing brownish white. 20-22 mm.

The fore wing is very acute, the costa and outer margin making an angle of less than 60 degrees, even in the male.

Florida; northern distribution uncertain, but described from North Carolina and Manitoba. I have seen a variety from Virginia in early June.

12. P. angulatana Robinson. Light clay-color, more or less dusted with gray, or, more rarely, with red-brown; markings grayish or reddish brown, nearly concolorous with the dusting. Basal area dark, its outer boundary bent at nearly a right angle at middle of wing, and running to inner margin well beyond basal angle; often with the lower part emphasized by a darker shade corresponding to the oblique band of the obliquana group. A broad contrasting median oblique fascia to costa, and spot on inner margin, the former not distinctly cut by pale veins; marginal dark band clean-cut, set back a little from the margin, a little irregular, running from apex to inner margin. Hind wing gray. 10-15 mm.

Not rare from July to October.

This species is strongly variable in ground color and slightly so in pattern. One specimen is suffused with fuscous, with the darker fuscous markings not at all contrasting.

Generally distributed. New York: Peru, Slaterville, Ithaca.

13. P. bomonana Kearfott. Similar to *P. angulatana*, ground light, lightly dusted with blackish; the markings only partly dark-shaded, partly of the ground color, and only defined by their paler edging; the median fascia cut by pale shades on the veins, and the subapical patch and subterminal fascia hardly paler than the ground, but with distinct pale defining lines and some black scaling. 15 mm.

ground, but with distinct pale defining lines and some black scaling. 15 mm. The type has the basal markings of the same form as in *P. dorsimaculana*, of which it is probably a dark variety, showing the coloring of *P. angulatana*. With this Kearfott has associated a Florida specimen which seems rather to be

a pale angulatana, with the normal angulatana pattern but with the coloring of dorsimaculana.

Chicago, Illinois.

14. P. rana Busck. Dull fuscous, with a slight purplish tint, mottled and dusted with dull black; base blackish with strongly excurved or almost right-angled outer boundary; a median oblique fascia running down from costa and a rounded spot on inner margin, about as in angulatana; subterminal spot on costa less distinct, and subterminal fascia barely traceable or absent. Hind wing nearly concolorous. 15-20 mm.

August and September. Larva on ironweed.

Connecticut, western Pennsylvania. New York: Rye. 15. P. smeathmanniana Fabricius. Straw yellow; markings deep brown, narrow and clean-cut, vaguely pale edged. A slight olivaceous shade near base of inner margin; a narrow brown oblique bar from inner margin at one-third way out to middle of cell two-fifths way out, with a small spot opposite its tip at middle of costa; three postmedial brown spots, forming a series parallel to this bar, the middle one sometimes diffuse and sometimes connected to the one on the outer margin. 15 mm.

July and early August.

Sherbrook, Quebec; Maine; New York; California. New York: Newcomb, Ithaca. The California record may belong rather to the closely similar P. parallelana Walsingham.

16. P. scissana Walker. Light yellow; costa with numerous oblique light brown striæ, but no rounded spots; inner margin with an oblique bar near base, and a rounded triangular spot beyond middle, with some gray shading between them. 15 mm.

Nova Scotia. Unknown to me.

17. P. fuscostrigana Clemens. Olivaceous, marked with darker olivaceous brown; the markings broadly edged with shining pale yellow. Base somewhat darkened. Median fascia from inner margin at one-third way out to just before middle of costa, less oblique than in P. scissana; dark brown, suffused with ochre yellow in the cell. A small blackish spot on inner margin at two-thirds way out; a nearly erect olivaceous subterminal fascia from costa to inner margin, less oblique than the outer margin, and narrowing toward the inner margin. Fringe checkered in lighter and darker gray. Hind wing gray. 15 mm. Labrador; northern New Jersey. Distribution uncertain.

This species is usually confused with *deutschiana*, of which it is probably only the eastern form.

18. P. deutschiana Zetterstedt. Similar in coloring to P. fuscostrigana, but without the bright yellow in the cell; the antemedial band more strongly oblique, reaching costa at middle, interrupted by a vague pale shade in cell only; spot on inner margin at two-thirds concolorous with it; subterminal fascia obscure, represented by a strong dark spot on costa only; pale edging of markings not silvery. 15 mm.

Europe; Colorado. Eastern records are probably based on fuscostrigana, if the two are really distinct.

19. P. floccosana Walker. Straw yellow, shaded with bright ochre on inner margin and disc of wing to two-thirds, and on costal edge to one-third, leaving a narrow yellow streak extending along cell to base. Hind wing almost white. 15 mm. (confusana Robinson).

June.

Nova Scotia to southern Ohio. New York: Ithaca.

20. P. atomosana Busck. Straw yellow; duller than P. floccosana, dusted or shaded with light wood-brown, and more or less dotted with black. A black dot, or usually a group of dots, at lower angle of cell, and a dot below them in the

fold; usually with a more or less complete subterminal series between the veins. Hind wing nearly concolorous. 15-20 mm.

August. This species looks like a Bactra, superficially.

New Jersey to Wisconsin.

21. P. louisiana Busck. Larger than P. romonana. Straw yellow, brighter thap P. romanana; markings deep ochre or yellow-brown. Base somewhat dark-shaded, especially at margins; median fascia running from middle of costa to inner margin one-third way out; somewhat bent and widened at cell, where it is paler, some times hardly darker than the ground; a small brown spot on inner margin at two-thirds; subterminal fascia narrow, irregular, and broken, sometimes suffused, sometimes with its lower end connected with the outer spot on inner margin; set well back from outer margin. Fringe pale. Hind wing gray.

• May to July.

Illinois; Missouri, Utah; New York: Ramapo (Kearfott).

22. P. romonana Kearfott. Straw yellow, marked with light olive brown; base somewhat darkened, or with an oblique bar extending up from basal angle; median fascia narrow on costa and at inner margin; more than twice as broad at middle of wing: the inner boundary nearly straight except at costa; the outer boundary outwardly oblique to end of cell, then angled or sharply bent and inwardly oblique to inner margin. An oblique fascia from costa at three-fourths way out toward outer margin, fading out below. Some confused marks on inner margin, and sometimes a distinct spot at two-thirds way to apex. Fringe concolorous; hind wing gray. 8-14 mm.

August.

New Jersey to Maryland and Manitoba.

23. P. hospes Walsingham. Ochre yellow; median area mixed yellow-brown and deep purple-gray, in varying proportion, especially toward the costa, forming a large triangular patch, starting narrowly at middle of costa and covering middle half of inner margin. Outer part with a curved yellow-brown shade starting from costa half way between the median patch and the apex, and joining the patch just below the cell. Fringe paler yellow. Hind wing brownish gray; the fringe a little paler, with a distinct dark line in its base. 10-15 mm.

August.

This form is *straminoides*, as identified by Busck, but Grote's original description applies better to the following form.

New Jersey to North Carolina and Wisconsin.

24. P. straminoides Grote. Ground light straw yellow; median fascia broad, oblique, narrowing somewhat to costa, from rather before middle of inner margin to rather beyond middle of costa; typically red-brown on dorsal half, becoming olivaceous and tending to fade out toward costa; sometimes entirely gray-brown. Outer part of wing broadly shaded with brown or olivaceous, with some blackish striæ along costa. Hind wing light gray. 9-15 mm. (schwarziana Busck, zaracana Kearfott).

Both names seem to apply to the form with the red-brown median fascia.

New York to Maryland and Illinois. New York: Buffalo, Ithaca, Trenton Falls. 25. P. plummeriana Busck. Similar to *P. straminoides*; rather brighter yellow, with apex of fore wing shading into a distinct red-brown patch. Hind wing white, nearly immaculate. Probably an extreme form of *straminoides*. 13 mm.

June.

Maryland to Illinois. New York: Peru.

Number 26 is vacant.

27. P. viscana Kearfott. Base of wing powdery gray; middle of costa fuscous; a chocolate-brown median shade, not reaching the costa; outer half of wing bright pink, crossed by a red-brown fascia aeross the apex; fringe bright ochre yellow,

yellow-brown at apex. None of the markings quite sharply defined. Hind wing light brownish gray. 12 mm.

May. Type only known.

Northern New Jersey.

28. P. aurorana Kearfott. Fore wing light ochreous with pink iridescence; median fascia somewhat oblique from middle of costa to inner margin at onethird way out, nearly even in width, somewhat excurved, and shaded with black on both sides at inner margin; outer part of wing with a triangle on inner margin, a fascia across apex and a large central patch lacking the pink iridescence, but not contrasting. Fringe concolorous, hind wing dirty white. 12 mm.

August to early September.

Maine and northern New Jersey.

29. P. hollandana Kearfott. Similar to P. aurorana; the outer part of the wing suffused with gray, leaving most of the pink along the costal edge; the median fascia more solidly blackish. Hind wing mouse gray. 13 mm. September. Not improbably a dark variety of *P. bunteana*.

Western Pennsylvania.

30. P. bunteana Robinson. Basal third cream white, median fascia even, oblique, fairly broad, abruptly narrowing to costa; mixed yellow-brown and black; outer part of wing yellow-brown, heavily shaded with pink; a brown fascia across apex, shaded with black, and with some black scaling between it and the end of the cell, often gathering in a distinct spot. Often with a mixed black and brown triangle beyond the middle of the inner margin, which may fuse with the fascia. 13 mm. (voxcana Kearfott).

June to September. Larva on Lactuca scareola. Common, south to North Carolina and west to Manitoba. New York: Wells, Ithaca, Poughkeepsie.

Number 31 is vacant.

32. P. albidana Walker. Head white, contrasting with the dark gray thorax. Base strigose blackish gray, the strigæ much stronger toward the inner margin usually crossed by a well-marked white antemedial shade and with some yellow scales in median area. Outer two-fifths ochre yellow, becoming white just beyond the gray base and on the costa, with a blackish area beyond it, wider on the costa and running out toward the anal angle, and crossed by a whitish transverse stria or band. Fringe yellow. Hind wing gray. 10-12 mm. (winniana Kearfott; nana auct., not Hübner.)

May to July.

New Jersey and north. New York: Rock City, Ithaca, Trenton Falls.

Number 33 is vacant.

34. P. cnotherana Riley. Base of fore wing ochre or olive yellow, outer half or rather more, deep rose pink; sometimes enclosing a patch of the ground color; the boundary nearly perpendicular to costa. Hind wing dark fuscous brown. 10 mm. General.

April; July; September. Larva a bud worm on evening primrose in June, and feeding on the rosette leaves in late fall.

Generally distributed. New York: Vicinity of New York City (Watson). 35. P. elderana Kearfott. Bright yellow-brown, shaded and reticulate with darker brown; a distinct but diffuse darker median shade, and bar across apex; fringe concolorous. Hind wing grayer brown. 12 mm.

July. Larva in pith of swamp elder, in June.

Southern New Jersey.

36. P. discana Kearfott. Fore wing wood brown, mottled with finely whitetipped black scales, sometimes forming a vague darker median shade. Fringe concolorous; hind wing brownish gray. 12 mm.

August.

Maryland to Ohio.

37. **P. foxcana** Kearfott. Dull fuscous brown, mottled with blackish, forming a vague darker median shade and band across apex. Hind wing nearly concolorous. 12 mm.

I suspect this is merely rubbed material of P. discana.

With P. discana.

38. P. wiscana Kearfott. Fore wing white, striate with light gray; the base of the costa darker; striæ forming a dark shade along middle of inner margin. A large lenticular postmedian gray patch from just below costa to inner margin, with a brown and blackish band curving around its upper end, running from costa to outer margin below middle. Apex with some brownish shading. Hind wing gray. 18 mm.

Type only seen.

Milwaukee County, Wisconsin.

39. P. ednana Kearfott. Pale gray, sparsely reticulate with dark gray. Basal fifth of wing darker, often contrastingly blackish; with somewhat excurved outer boundary; costal half of medial area occupied by a large, rounded, darker, often blackish, spot, covering end of cell. A stronger dark subterminal stria, starting from a distinct patch at costa; somewhat more sharply curved than the outer margin; typically reaching the inner margin well before the anal angle, but often fading out. Hind wing light gray. 12 mm.

June and July.

Mount Washington, New Hampshire, and Ottawa, Canada, to Pennsylvania; the specimens showing strong contrasts apparently commoner northward.

This is probably a Cnephasia near virescana.

40. P. temerana Busck. White, strigose with gray; the base paler than the median area; a mixed yellow-brown and black oblique fascia from inner margin at one-third to outer part of cell, slightly diffuse in outline, and fading out above; costal edge shaded with gray; outer third, beyond cell, heavily shaded with yellow brown, with a darker brown fascia across apex. Hind wing light gray. 9-15 mm. (cincinnatana Kearfott).

Western Pennsylvania to southern Ohio.

41. P. leguminana Busck. White, base light gray, middle third light gray on dorsal half of wing, defined with black on basal side, leaving a narrow white band between it and the gray base; costal part of median area somewhat shaded with gray. Outer part of wing mixed white, gray, fuscous, brown, and blackish; the white clear at end of cell, and usually also subterminally beyond an irregular and broken subterminal dark band, which is roughly parallel to the outer margin. Fringe gray with two darker lines. Hind wing brownish gray. 10-15 mm.

Larva in seed pods of Gleditschia.

Pennsylvania to Missouri and Texas.

42. P. maiana Kearfott. Dull straw yellow; median fascia contrasting, redbrown mixed with black, starting as an oblique fascia at costa, on reaching cell becoming a large triangular patch, almost as broad as its height on inner margin; the outer boundary on the whole transverse, the inner one strongly oblique, turning a right angle at **R**. Outer part of wing heavily shaded with fuscous and scaled with blackish, leaving a clear quadrate cream patch on costa behind the median fascia, and an obscure one on dorsal margin. Fringe with two obscure darker lines. Hind wing brownish gray. 8 mm.

May.

Northern New Jersey.

43. P. gunniana Busck. Similar to P. maiana; ground bright ochre yellow; base paler on inner margin and with a slight pink tinge beyond the median fascia; median fascia light yellow-brown, the part below Cu overlaid with black, forming a contrasting rounded patch. Outer part with blackish scaling heavier, and a dis-

tinct largely black band across the apex. Fringe with basal line stronger, but outer line weaker; outer part of fringe barred with grav. 10-12 mm. New Jersey to Maryland.

44. P. hoffmanana Kearfott. Base cream, its outer boundary oblique from middle of costa to inner margin one-third way out; outer part a confused mixture of vellow brown and black, with a white postmedial spot on costa, preceded and followed by chocolate brown; the outer chocolate brown bar tending to continue as a fascia across the apex. Fringe mixed dull yellow and gray. Hind wing dark grav. 12 mm.

May and June.

Black Mountains, North Carolina.

45. P. zoxcana Kearfott. Base of fore wing cream, the boundary running from before middle of costa to inner margin at one-third way out; outer part vellowbrown, mixed with blackish; forming a large blackish subterminal patch, with a cream-white postmedial costal spot, and a pale streak at middle of inner margin. Fringe with distinct dark basal line. Hind wing dark gray. 8-11 mm.

May and June; August.

Cincinnati, Ohio.

46. P. toxcana Kearfott. Base cream white, more or less scaled or shaded with dark gray; a vague yellow-brown median fascia, shaded with gray, nearer base on inner margin; outer third of wing heavily shaded with yellow-brown on a silvery gray base; the brown principally in two oblique fasciæ, the upper crossing the apex and partly filled with a small black patch; considerable white scaling in postmedial region. Fringe dark gray and dull white, with considerable yellowbrown in its base. Hind wing light gray, paler at base. 10-12 mm.

August and September.

Northern New Jersey to western Pennsylvania.

P. marloffiana Busck (nonlavana Kearfott) I am unable to distinguish from P. toxcana. The type differs only in the greater extension of the black and gray at the expense of the yellow-brown; but toxcana Kearfott is included in Busck's conception of marloffiana. Busck's name has a few weeks' priority.

April; August.

Western Pennsylvania; Maryland, southern Ohio. New York: Ithaca.

47. P. lavana Busck. Similar to P. toxcana, but lighter and apparently broaderwinged. Head pale; costal edge with black points, base much shaded with light gray; extreme base blackish; median oblique fascia narrow, blurred, and often obscure, mixed ochreous and light gray, the gray dominant; median fascia followed by a well-marked whitish area; outer part of wing mixed light gray and ochreous, with some black scaling; the bar across apex rather distinct and largely black-filled. Fringe dark gray with a little yellow in base (nonlavana Kearfott; dubitana auct., not Kearfott). 10 mm.

June; August. Larva on Pentstemon.

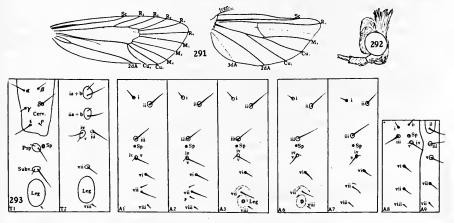
I have seen this species determined as campicolana Walsingham. Some specimens show practically no yellow.

Nova Scotia to British Columbia, and south to southern New Jersey and Missouri. New York: Ithaca.

30. Family CARPOSINIDÆ

(Tortricidæ, in part)

Head as in the Tortricidæ. Ocelli absent. Tongue rather weak; antennæ heavily ciliate in male. Palpi strongly dimorphie in the sexes; in male (fig. 292) upturned, with second joint heavily scaled on lower edge and a short close-scaled third joint well set off; in female, long and oblique, the second joint straight and smoothly scaled on the lower side but with long vestiture on the upper; the third joint short, nearly smooth, and porrect. Thorax and legs scaled, the hind tibiæ more or less hairy, as usual. Wings thin and soft, with long fringe (fig. 291); with arched costa and curved, oblique outer margin, as in the broad-winged (Ecophoridæ (Semioscopis)). Fore wing heavily tufted, with all veins preserved; \mathbf{R}_5 widely separated from \mathbf{R}_4 at origin, running to outer margin; \mathbf{R}_1 arising well out on cell; \mathbf{M}_2 to \mathbf{Cu}_2 inclusive, closely crowded at lower angle of cell; \mathbf{M}_3 and \mathbf{Cu}_1 perhaps closest together; \mathbf{Cu}_2 leaving cell almost at right angles,



FIGS. 291-293. CARPOSINIDÆ

291, Carposina fernaldana, venation; 292, Bondia crescentella, male, side view of head; 293, Carposina fernaldana, seta map of larva.

and then curving, as in many (Ecophoridæ. 1st A lost, 2d A shortly but strongly forked at base. Base of \mathbf{R}_{4+5} and of M obsolete. Hind wing with cell closed. Sc normal, apparently free, but closely approximate to R at base; R normal; \mathbf{M}_1 lost in the American species, represented by a fold running from just above the middle of the cell; \mathbf{M}_2 from, or from just above, lower angle of cell; \mathbf{M}_3 lost (perhaps completely fused with \mathbf{Cu}_1); \mathbf{Cu}_1 from lower angle of cell; \mathbf{Cu}_2 from twothirds or three-fourths way out on cell, normal, straight; the American species with a weak fringe of hair on Cu. Veins 1st A and 3d A rather weak; 2d A with a well-marked fork at base.

Larva (fig. 293) with only two setæ on prespiracular wart; vii of two setæ on prothorax, one on meso- and metathorax and eighth segment of abdomen, two setæ on first and seventh segments, and three on second as well as on the leg-bearing segments. Abdomen with

setæ iv above v; prolegs with a small complete ring of uniordinal hooks. Eighth segment much reduced dorsally, with spiracle enlarged, dorsal, on a level with setæ iii; ii rather higher than i. Ninth segment with setæ ii well separated, but on a single plate. Pupation in a cocoon of brittle silk, covered with sand, in the ground. Pupa not described.

The larvæ, as far as known, are borers in fruits, and one is injurious to peaches in Japan. The family is small and probably very old. It is of uncertain relationship but perhaps cognate with the Phaloniidæ. Most of the hundred known species are from Hawaii and Australasia.

Key to the genera

1. CARPOSINA Herrich-Schæffer

Male with a heavy tuft on lower side of second segment of palpus; the palpi closely upturned. Wings broader; fore wing (fig. 291) with an oblique series of three tufts near base, an antemedial series parallel to it, the middle one located on cell Cu, and very weak, the ones below costa and above A strong; strong tufts below Cu farther out, above and below cell, and a couple at end of cell. Hind wing translucent, with strong fringe of hairs on base of Cu; M_2 and Cu_1 connate or shortly stalked; costal margin strongly sinuate in female. Postmedial shade of fore wing wavy and indented opposite lower angle of cell.

1. C. fernaldana Busck. Palpus of male with tuft moderate and rounded at end; fore wing powdery gray, the costa concolorous, mottled with darker gray. Tufts blackish-tipped; a black bar or patch in end of cell; postmedial line diffuse, excurved, indented opposite lower angle of cell, and slightly dentate on veins. A series of fine black terminal dots. Hind wing translucent, pale gray. 18 mm.

Late July and August. Larva in nearly ripe currants.

New York to Illinois and Missouri. New York: Ithaca.

2. C. nicholsana Kearfott. Palpal tuft in male much larger, and triangular; fore wing with practically the same pattern, but with the base of the wing and the costa above Sc contrastingly dark, sometimes almost wholly dark gray. Blackish patch in end of cell in a dark shade. Terminal dots larger, lighter, and diffuse, often partly confluent. 15 mm.

Nicholson, Pennsylvania.

July.

3. Č. ottawana Kearfott. Wings narrower, though broader than in *B. crescentella*. Palpal tuft triangular. Fore wing duller fuscous, powdery, but not contrastingly so; antemedial region, from the first row of tufts to the middle, broadly shaded with white; blackish patch in end of cell and postmedial line nearly lost in the general fuscous ground; terminal dots obscure. Hind wing more opaque, darker gray. 16 mm.

June.

Ottawa, Ontario.

2. BONDIA Newman

(*Carposina*, in part)

Near Carposina. Palpi of male with a slight tuft at end of second segment only (fig. 292); wings narrower, scaling coarse, hind wing with M_2 and Cu_1 distinctly separate.

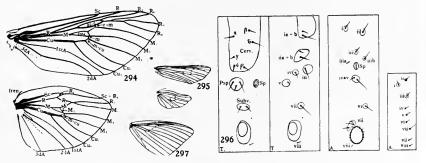
1. B. crescentella Walsingham. Powdery gray; markings much like *C. fernal-dana*; the postmedial shade strongly and evenly excurved; the larger tufts with contrasting yellow scales on their upper and outer sides, the one at the end of the cell showing as a yellow and white crescent to the naked eye. 13 mm.

May.

General in distribution, South of Pennsylvania. New York: Rock City (not rare), Big Indian Valley.

Family 31. COSSIDÆ (Zeuzeridæ)

Head with erect vestiture, sometimes forming a distinct tuft on vertex; no ocelli; male antennæ, in the North American species, pectinate, at least toward the base. Shaft scaled on the upper side, the scales irregular and often ephemeral. Palpi upturned to middle of front (Cossinæ) or very short (Zeuzerinæ); the other mouth-parts rudimentary. Thorax with spatulate vestiture, usually rather close, with a posterior tuft in the Cossinæ; vestiture loose and woolly in the Zeuzerinæ; pleuræ, lower side of femora, and abdomen also with spatulate vestiture, mixed with loose hair in the Zeuzerinæ. Legs stout, often with short spurs. Body very stout, far exceeding the hind wings, as in the Sphingidæ, but somewhat more clumsy. Fore wing (figs. 294, 295) with a large accessory cell, separated from the discal cell by a strong vein; \mathbf{R}_3 to \mathbf{R}_5 stalked from its tip, \mathbf{R}_4 and \mathbf{R}_5 the farthest; base of media well developed, forked, enclosing an intercalary cell in our species;



FIGS. 294-297. COSSIDÆ AND CASTNIIDÆ

294, Prionoxystus robinia, venation; 295, Zeuzera pyrina, venation; 296, Cossus cossus, seta map of penultimate stage of larva (Europe); 297, Castnia harmodius (Castniidæ), fore wing (South America).

1st A fully developed; 2d A strongly forked at base. Hind wing proportionately small; frenulum ordinarily well developed, rarely lost; humeral vein sometimes distinct, running across to base of frenulum; \mathbf{R}_1 variable; when present, arising near middle of cell or beyond; exceptionally long and strong in Prionoxystus, where it is well beyond the middle of the cell, sometimes supplemented by a

secondary cross vein beyond end of cell; base of **M** developed, forked; all three anals strong and complete; the second shortly forked at base. Egg of upright type, with longitudinal ribs and reticulate micropyle in the Cossinæ; of flat type, oval and smooth, in the Zeuzerinæ. Larvæ boring, usually in solid wood, the smaller species more often in herbaceous perennials; as a rule spending two or more years in the larva stage. Larva (fig. 296) stout, with a relatively small head, held horizontally; shields and tubercles large and heavily chitinized; labrum hardly notched, mandibles very large, and projecting forward. Ocelli normal. Front not reaching more than half way to vertex; adfrontals short in Zeuzera, reaching to the slightly cleft vertex in Cossus. Bodysetæ as in other Tortricoidea; setæ ii of ninth segment of abdomen at least as far apart as on other segments; setæ iii of abdomen duplicated in Cossus. Prolegs with hooks variable.

Pupa incomplete, with second segment of abdomen fixed; mandibles separately chitinized; head sutures mostly distinct; dorsal headpiece rudimentary but carrying the eye pieces on dehiscence; front with a specialized cocoon-breaker in the Zeuzerinæ; maxillary palpi separate in Cossus, imperfectly separated in Zeuzerinæ; antennæ not reaching end of legs, widely separated; tongue very short, the two halves separated by the labial palpi in the Cossinæ: Base of labium well developed, and palpi fully exposed. Cremaster not developed; end of abdomen more or less truncate, with a group of angulate projections. Early stages of Hypoptinæ not studied.

This is in many ways, notably in venation, one of the most primitive families of Lepidoptera, and at the same time is the base of a series leading directly to the butterflies. The only definite butterfly character in this family is the upright egg of the Cossine. In the Castniidæ (fig. 297) the tongue, which is lost in the Cosside, is preserved; the habits have become butterfly-like; the upper fork of **M** is lost; the wings have become ample and the cell relatively small; and the antennæ are clubbed. In the Neocastniidæ, which are in other ways aberrant, \mathbf{R}_1 has become stalked beyond the origin of \mathbf{R}_{4+5} , as in pupæ of butterflies. The Euschemonidæ have lost the base of **M** and **1st A** in both wings, differing from our local skippers only in the more woolly vestiture and the presence of the frenulum. They also have the skipper type of larva, with secondary hair, and well-set-off head.

Key to the genera

- 2. Both branches of base of **M** ending in outer margin of cell; male antennæ pectinate to apex.

^{1. 1}st A and 2d A connected by a cross vein near margin.....1. Givira. 1. 1st A and 2d A free.

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- 3. Sexes similar; male with rounded hind wings; female with simple or laminate antennæ......2. Acossus.
- 3. Sexually dimorphic; male with outer margin of hind wing nearly straight;

Subfamily HYPOPTINÆ

Smaller species, the larvæ probably normally in roots of perennial herbs; moth tending strongly to lose tibial spurs and frenulum. A strong cross-vein near outer margin, between 1st A and 2d A; other characters as in Cossinæ; larva not well known.

1. GIVIRA Walker

(Inguromorpha; Hypopta, in part)

Male antennæ pectinate to apex, in female normally very narrowly pectinate; palpi moderate, upturned; accessory cell small, but well set off from discal cell; intercalated cell inconspicuous. Hind wing with \mathbf{R}_{r} rudimentary, \mathbf{R}_{s} and \mathbf{M}_{l} stalked. Vestiture spatulate.

1, G. anna Dyar. Gray, evenly recticulate with lines of black powdering, discal lunule white; inner margin narrowly white at base. Hind wing darker, unmarked. 25-30 mm.

Florida; St. Louis, Missouri; and probably in intermediate country. The St. Louis race is darker than the type.

Subfamily COSSINÆ

Egg upright; strongly ribbed; with well marked micropyle. Larva with cervical shield moderate and smooth, spiracles in line; crotchets in our two genera trior-dinal, in a complete circle; in the genus Cossula (which is transitional to the preceding subfamily) crotchets uniordinal in two transverse rows. Pupa with maxillæ not meeting in middle line beyond end of labial palpi; maxillary palpi separately chitinized; head smooth. Imago with male antennæ pectinate to apex, cronticular single leminate: palpi moderate; interceleted cell triangular, permet or entirely simple, laminate; palpi moderate; intercalated cell triangular, normal; 1st A and 2d A free; hind wing with R and M_1 connate or stalked; M_2 , M_3 , and Cu₁ more or less approximate.

2. ACOSSUS Dyar

(Trypanus, Cossus, in part)

Vestiture of thorax deep, spatulate, but regularly imbricated, with posterior tuft; male antennæ pectinate to apex; female antennæ laminate below; palpi moderate, upturned. Fore wing with large accessory cell; R_1 arising from discal moderate, upturned. Fore wing with large accessory cell; \mathbf{R}_1 arising from discal cell, \mathbf{R}_2 from accessory cell, \mathbf{R}_3 connate with $\mathbf{R}_{1,33}$, which are stalked less than half-way to apex. Intercalated cell triangular, not reaching posterior margin of discal cell; anals not connected. Hind wing relatively large, with Sc and \mathbf{R} connected by a strong vein about two-thirds way out on cell; \mathbf{R} and \mathbf{M}_1 shortly stalked; \mathbf{M}_2 , \mathbf{M}_3 , and \mathbf{Cu}_1 approximate at base. 1. A. centerensis Lintner. Thorax and basal half, or rather more, of fore wing blackish; collar often partly light gray; thorax spotted with whitish. Outer part of wing contrasting, pale gray; the whole wing dusted and heavily reticulate with black. Hind wing translucent, very lightly reticulate, with blackish fringe. Hair in base of anal region blackish. 50-60 mm. (H 12:1.) Larva boring in poplar. Tubercle iii of metathorax fully as large as iia+b.

Montreal, Quebec, to New Jersey and Illinois. New York: Peru, Batavia, Lestershire, Karner (type), Brooklyn.

2. A. populi Walker. Fore wing light gray, strongly reticulate, the reticulation a little denser in the antemedial region. Hind wing strongly reticulate on outer half of wing, with checkered fringe. No dark hair at the base. 60 mm. July.

Hudson Bay; Hymers, Ontario.

3. PRIONOXYSTUS Grote

(Xystus Grote, not Hartig; Xyleutes, in part)

Similar to Acossus; sexes unlike; the male much smaller, with the outer margin of the hind wings nearly straight; female with bipectinate antennæ; vestiture slightly shorter, practically of plain scales on top of thorax. (Fig. 294.) Larva with tubercle iii of metathorax decidedly smaller than ii a+b.

1. P. robinize Peck. Fore wing nearly white, slightly translucent, irregularly striate with black; the middle of the wing covered by an irregular darker patch, especially in the male. Hind wing of male bright yellow, the basal half and costa above \mathbf{R}_1 blackish; outer margin narrowly black. Female with hind wing smoky gray, the reticulation of the under side showing through faintly. 50-75 mm. (plagiatus Walker). (H 41:11, 5; 10, 2.)

Female variety reticulatus Lintner lacks the blackish patches in the median area of the fore wing, which is only a little more heavily reticulate there; is a little smaller and more translucent and is commoner southward. In female aberration quercus Ehrman, the female hind wing is almost as bright yellow as that of the male.

Generally distributed. New York: Plattsburg, Peru, Ogdensburg, Brockport, Buffalo, Ithaca, Oneonta, Albany, Staten Island; Long Island, common at Brooklyn. One Ithaca specimen is practically var. reticulatus Lintner.

Not very rare, but easily overlooked, the moth being short-lived and not flying to sugar, or often to light. It is most easily found resting on the trunk of its food plants. Larva boring in many trees, preferring soft woods; perhaps most

often in poplar and locust; often doing serious damage to unhealthy transverse, in 2. P. macmurtrei Guérin. Fore wing with reticulation mostly transverse, in the female with one well-marked stronger stria from costa to anal angle just beyond the cell; no dark shading in median area. Hind wing of male nearly transparent, with a very slight yellow tint, blackish along the inner margin. 3 30, \Im 60-75 mm. (querciperda Fitch).

June. Larva boring in oaks. Rare in collections.

Widespread but apparently quite local in distribution, west to Minnesota. New York: Buffalo, Ithaca, Schoharie, Crugers, Newburgh, New York City, Staten Island; Brooklyn, Woodhaven, and Newtown, Long Island. The two species have been much confused, especially in the larva, making food

records a little uncertain.

Subfamily ZEUZERINÆ

Egg ovate, flat, smooth; larva with cervical shield extremely large, rough; last spiracle raised, as in Carposina; crochets slightly irregular, but nearly uniordinal, Pupa with maxillæ meeting in middle line beyond end of labial palpi; maxillary palpi not separately chitinized, separated by a partial suture; head with a characteristic toothed projection. Imago with male antennæ pectinate only on basal portion, the pectinations curved and forming a sort of eye-cap; vestiture deeper; base of M₃ transverse, and intercalated cell rectangular. Veins of hind wing, especially R and M₁, widely spaced.

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4. ZEUZERA Latreille

Male antennæ broadly bipectinate on basal two-fifths, the rest simple. Female antennæ simple; vestiture woolly, largely mixed with long spatulate scales and hair; both wings (fig. 295) with stem of M_a running directly across to the lower side of the cell, making the intercalated cell nearly rectangular. Hind wing with R_i rudimentary, but visible in a well-bleached wing, functionally replaced by a secondary crossvein at end of cell; all veins arising well separated, from end of cell. Fremulum strong; palpi rudimentary; hind tibiæ with end spurs only. Larva white, cervical shield very large, rough, and black; tubereles black, skin white; last spiracle raised. Larva boring in the twigs the first fall, then moving down to the trunk or larger branches (fig. 295).

1. Z. pyrina Linnæus (Leopard moth). White, regularly spotted with blueblack; the hind wings sometimes nearly all white; abdomen blue-black. 45-70 mm. (*æsculi* Linnæus) (H 9:9.)

Larva a general feeder on trees, but really attacking perfectly healthy ones; often a serious pest under city conditions.

Vicinity of Boston. Massachusetts, and New York City; introduced from Europe. New York: Vicinity of New York City, and north to Kensico, Westchester County.

SUPERFAMILY PYRALIDOIDEA

Slender, usually moderately small moths. Head prominent, usually with ocelli; antennæ almost always simple, ciliate, with two dorsal rows of scales on each segment; the reticulation of the unscaled surface usually (except Orneodes) forming hexagonal meshes. Male antennæ frequently with a process on the scape (Phycitinæ, Epipaschiinæ, Omphalocera) or a notch and tuft on the shaft (Desmia). Palpi usually moderate or long, very often projecting beak-like (whence the name " snout-moths " for the group); much reduced in males of Galleriinæ. Tongue almost always scaled at base. Wings usually close-scaled, the scales attached more firmly than in most of the lower families. Fore wing varying from narrow to ample; the discal cell always well-formed, but accessory cell always absent. Cu apparently 4-branched, base of media lost; 1st A usually lost. Hind wing with Sc and R closely parallel to beyond end of cell; often fused for a greater or less distance, Base of **R** often lost by atrophy, the tip of **Sc** very rarely lost, leaving the basal part of **Sc** and the tip of **R** to form the most anterior vein of the wing. 1st A almost always preserved. Legs usually long, sometimes extremely long.

Eggs of flat type. Larva with prespiracular wart with only two setæ, except in the few forms with tufted hair. Setæ iv and v approximate on abdomen, usually on the same tubercle. Rarely with tufted or secondary hair, and only in forms with round spiracles. Larvæ of almost all the species concealed feeders. Pupæ obtect, with apex of labrum bilobed (indicating the pilifers); and antennæ long, not swollen outwardly. Eighth segment of abdomen free from seventh in males of Pterophoridæ only.

This large group is formed mostly of the family Pyralididæ. The Pterophoridæ are undoubtedly related, though more specialized in some ways and more generalized in others. The Orneodidæ are of doubtful affinity, and are placed here mainly for convenience.

Family 32. THYRIDIDÆ^{33a}

(Thuridæ: Sphingidæ, in part, of the older authors)

Ocelli absent; eyes small; tongue and labial palpi strong; maxillary palpi obsolete. Hind tibiæ of male enlarged with a peneil of hair. Fore wing with all veins present, in the northeastern species all arising separately from the cell, or with \mathbf{R}_2 and \mathbf{R}_3 very shortly stalked. No distinct patch of spinules near base of inner margin; 1st A abse t.

³⁷a The figures for this family are on page 333.

Hind wing with Sc and R wholly independent, but approximate (Thyris, fig. 196); or fused (Dysodia, fig. 195) beyond middle of eell; the other veins normally all arising separately; 1st A lost but ,3d A strong. Cells open.

The species of temperate climates agree with the preceding descripstion, and are small, heavy-bodied moths with thick vestiture and gransparent discal spots. The tropical Rhodoneurinæ are more slenedler, with ample wings. They frequently have some radial branches stalked; **Sc** and **R** of the hind wing are approximate beyond the end of the cell; and the cells normally are closed. They link the typical members of the family with the Pyralididæ.

Caterpillars (fig. 197) superficially Pyraloid, but more primitive than any Pyralid in having three setæ on the prespiracular wart of the prothorax at least in some specimens (Fracker). Primary setæ only. Head smallish, and normal; front reaching more than half way to vertex. Hesothorax and metathorax with tubercle vii bearing two setæ. Abdoinen with setæ iv and v on a single tubercle, vii of two setæ on first and seventh segments, and single on eighth. Ninth segment with a preanal plate bearing setæ i in front of, and slightly above, ii. Prolegs vather long, with a complete series of biordinal hooks. Spiracles oval. Caterpillars concealed feeders; immaculate, except for the black chitinous parts. Pupa solidly obtect, not studied.

The family is a wholly isolated and very strange one, combining beharacters of the Pyralids and Macrolepidoptera with primitive feattures. Its nearest relatives seem to be the tropical Hyblæidæ. Fracker has placed the group in the Tineid series; but it appears on the whole pearer the Pyraloids.

20

1. THYRIS Laspeyres

. Palpi obliquely upturned, with broad, bladelike second joint; male antennæ 9 prismatic and thick. Hind tibiæ stout, with large, crowded spurs. Abdomen 9 meavily tufted at sides toward rear. Fore wing with \mathbf{R}_2 and \mathbf{R}_3 well separated. If find wing with Sc and R separate (fig. 196). Larva with adfrontal sclerites even in width.

Key to the species

1. T. maculata Harris. Outer margin strongly wavy. Black. Stripes on abdoymen, a subterminal series of spots on wings, and scattered spots, all bright tawny .yellow. A large, squarish, hyaline patch in the end of the cell of each wing, and a similar patch on inner margin of hind wing, sometimes fusing with the discal one. 12-15 mm. (H 47:30.)

one. 12-15 mm. (H 47:30.) .b5 End of May to July; not common and quite local in occurrence. Larva probably ha stem borer or leaf-roller on clematis.

Montreal, Ontario, to Montana; south to District of Columbia. New York: Lancaster, Newport, DeBruce, Kingston, Poughkeepsie, Esopus, Long Island.

2. T. lugubris Boisduval. Black, with translucent white markings; pale patches on tegulæ; two or three white bars on body; a hyaline patch below the cell on fore wing, as well as the one in the cell, often fused with it; hind wing with a broad median fascia; and a very irregular series of subterminal white spots, one opposite the cell of the fore wing largest. 15-23 mm. (H 47:31.)

June. Larva on grape.

Southern States, north to Ithaca, New York. New York: Ithaca (Eyer), Karner, Bronxville, Staten Island; Brooklyn, and Wading River, Long Island.

2. DYSODEA Clemens

(*Thyris*, in part; *Platythyris* Grote)

Similar to Thyris. \mathbf{R}_2 and \mathbf{R}_3 of fore wing at least somewhat approximate; Sc and \mathbf{R} of hind wing anastomosing (fig. 195). Larva with adfrontals wider toward the top.

the top. 1. D. oculatana Clemens. Yellow, reticulate with brown, with a premedial, post-medial, and subterminal darker brown bands. Transparent discal spot small and rather triangular. Hind wing similar, less obviously banded, with a large, lunate, discal spot. 20 mm. (fasciata Grote and Robinson). (H 3714, Job another the medial, and the searly in July. The larva appears to be a rather general feeder and lives in a dirty and foul-smelling nest. (It has object of the search of the search of the very lew records of the search of the search of the search of the search of the medial into beans, and is easily recognized by its shell. The distribution seems general but II have very lew records of the search of the Pennsylvania; West Virginia; and southern Canada. 2. D. vitring Guenée. Brown, with reddish areas near outful the search of t

2. D. vitrina Guenée. Brown, with reddish areas near outer therein and on hind wing only. Transparent spots as in *D. operational* with the distance of the di

Georgia.

This species or form appears not to have been taken since its original description.

6. (all of bin EADIDIDAE us Family 33. PYBALIDIDAE und for Mail of the State of the

Crambinge (Raphipt.

5 R. and R. stulked. (Snout moths)

1) heav tringe on base of Cu of hing wing, labbi path bask-like wing never divided in two shirts fathers; divided in two in a single Australasian gentis! Fore wing entire, except in the same genus as Fore wing with 1st A fisually lost." Hind wing without special scaling valong Cu beneath. Larva with, primary sete, and moderately developed prolegs mwithsibi- or triordinal hooks except in the Chrysaugrine motenue syswie teomic pur bud is 9 bus 22 : 00% gen athg subfamilies Glaphyrine, Pyratstine, Nynghuline, Scopa-

ringenGrambing; Anerastiing, and Phyciting, the antennal socket is normally separated from the eye by a row of scales; in the Pyralidina,

Epipaschiine and a few others, they are normally in contact. totalking about 10,000 species. The subfamily Pyraustine is predomiffahtly "tropical, the "Phycitina" mostly in the temperate zone. The Chrysauging and Macrotheeing are practically collined to the New Worldhewhile, the Endotriching and several small groups are almost confined to the OldizWorld. 8 mont betatages lies .M base of Rast as from Ma. p. 581 Scopariinge.

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Key to subfamilies; imago

- 1. Vein 1st A preserved in fore wing (fig. 298); tongue weak or absent; fringe on Cu weak or absent. p. 525..... Schenobiinæ.
- 1. Vein 1st A absent in fore wing.
 - 2. Vein 3d A of fore wing running into 2d A near base of wing (figs. 306 to 308, 312, 315).
 - 3. Tongue and ocelli absent; male with third segment of palpi rudimentary. 4. Front with a conical tuft; general vestiture deep and mixed. p. 532.
 - Galleriinæ. 4. Front and thorax smooth-scaled. p. 535......Macrothecinæ. 3. Tongue and ocelli present, palpi normal.
 - 4. Maxillary palpi rather well developed; Sc of hind wing free (fig. 362).

Chrysauginæ.

- 2. Vein 3d A of fore wing free, or curving into 2d A near middle of wing (fig. 320), forming a wide loop, often rudimentary.
 - 3. M₁ in hind wing arising from closing vein of cell, widely separated from Sc (fig. 367); R more or less weakened; cell finely closed. Labial palpi beak-like and maxillaries triangular. p. 588...... Ancylolomiinæ.
 - 3. M₁ closely approximate to **R**.
 - 4. \mathbf{R}_{5} stalked with \mathbf{R}_{3+4} ; one free vein only, below the forked vein running to the apex from the radial stem.
 - 5. \mathbf{R}_3 and \mathbf{R}_4 of fore wing completely united; fringe on \mathbf{Cu} of hind wing strong.
 - 6. Cell of hind wing closed by a fine but nearly complete vein; frenulum of female simple.
 - 7. Tongue strong, separating the palpi toward the base. p. 608.
 - Phycitinæ. 7. Tongue rudimentary, concealed by the palpi when coiled. p. 637. Anerastiinæ.
 - 6. Cell of hind wing widely open, frenulum of female multiple. p. 604. Crambinæ (Raphiptera).
 - 5. \mathbf{R}_{3} and \mathbf{R}_{4} stalked.
 - 6. A heavy fringe on base of Cu of hing wing; labial palpi beak-like
 - plumose, or small and concealed.

7. Fore wing with raised scale-tufts. p. 604..... Epipaschiinæ. 7. Fore wing smooth. p. 584..... Pyralidinæ.

- 4. \mathbf{R}_5 free; two free veins from radial stem below the forked one (fig. 320); Sc and R of hind wing almost always anastomosing.
 - 5. Fringe on Cu heavy; palpi beak-like, with trangular maxillary palpi.
 - 5. Fringe light or absent; palpi rarely beak-like; maxillary palpi usually moderate or small and not triangular.
 - 6. No loose spatulate scales on dorsal part of hind wing.
 - 6. Some loose hair near inner margin of hind wing, part of it forming a weak fringe on Cu which runs into a group of spatulate hairs or scales below Cu. p. 536.....Glaphyriinæ. 7. \mathbf{R}_2 stalked with \mathbf{R}_3 and \mathbf{R}_4 (fig. 346) p. 574..... Nymphulinæ. 7. \mathbf{R}_2 free.
 - 8. Labial palpi beaklike; maxillary palpi large and triangular (fig. 354); fore wing usually slightly rough-scaled; with M_1 well separated from R_3 at origin, about as far from

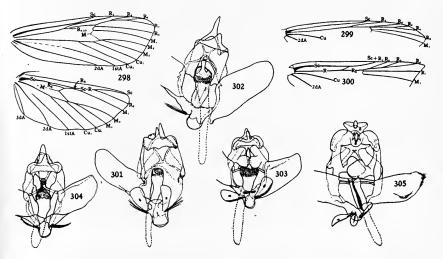
8. Labial palpi often upturned; maxillary palpi (fig. 339, etc.) yery rarely large and triangular (Loxostegopsis), and in that case with \mathbf{R}_{3+4} , \mathbf{R}_{5} , and \mathbf{M}_{1} closely approximate. p. 541.

Pyraustinæ.

Subfamily SCHCENOBIINÆ

(Crambinæ, in part)

Male antennæ simple, laminate, except in a few exotic species; palpi long, almost always porrect; maxillary palpi large, triangularly tufted; tongue very weak or absent. Thorax often with vestiture of fine loose hair. Fore wing (fig. 298) with outer margin curving more evenly into inner margin than in most Crambinæ; **R** with all five branches preserved; \mathbf{R}_5 free in our species, and \mathbf{R}_2 frequently free; 1st **A** a developed tubular vein at margin, with interspaces of the normal width between it and the veins above and below it; usually curving down to inner margin; 2d **A** almost always ending in inner margin; 3d **A** short and free, obscure. Hind wing with a slight fringe on base of **Cu**, or none; female with frenulum of several bristles; **Sc** and **R** anastomosing beyond end of cell; **M**₁ from upper angle of cell or shortly stalked with **R**; dorsal veins all present and normal.



FIGS. 298-305. PYRALIDIDÆ (SCHÆNOBIINÆ)

298, Rupela albinella, J, venation; 299, Schanobius melinellus, venation of costal half of fore wing; 300, Patissa parthenialis, venation of costal part of fore wing; 301, S. sordidellus, J, genitalia (right valve removed) seen from ventral view; 302, S. nitidellus, J, genitalia; 303, S. unipunctellus, J, genitalia; 304, S. mellinellus, J, genitalia; 305, S. longirostrellus, J, genitalia.

The Scheenobiinæ are a small group of more or less aquatic moths, much like the Crambids, but weaker-winged. The wings are normally more loosely rolled in repose. The moths are locally common in wet places. The larvæ are hardly known, structurally, and are borers in marsh and aquatic plants. In one European genus, Acentropus, the larva is completely aquatic and the female of the summer brood is wingless and lives under water.

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Key to the genera

Palpi upturned; fore wing with \mathbf{R}_2 to \mathbf{R}_4 stalked.....l. Rupela. Palpi porrect, beak-like; R₂ free.....

1. RUPELA Walker

(Scirpophaga, in part)

Palpi upturned, loosely held. Thorax with loose soft hair. Fore wing (fig. 298) with \mathbf{R}_2 shortly stalked with \mathbf{R}_3 and \mathbf{R}_4 ; \mathbf{R}_1 anastomosing shortly with Sc in our species and hind wing with \mathbf{M}_2 and \mathbf{M}_3 stalked. \mathbf{M}_1 long stalked with $\mathbf{Sc}+\mathbf{R}$. 1. **R. albinella** Cramer. White, immaculate. 25 mm. (*nivea* Walker).

June. Northern specimens are small.

New York, Snake Hill, New Jersey, and south. New York: Staten Island.

Patissa xantholeucalis Guenée ranges north to North Carolina. Its palpi are beak-like tut not long, \mathbf{R}_{2} is stalked, and \mathbf{R}_{1} nearly obsolete (fig. 299). The wings are white, with two yellow bands.

2. SCHENOBIUS Duponchel

(Chilo, in part)

Palpi beaklike, as long as head and thorax, and down-curved at apex. Thorax without long loose hair; fore wing (fig. 299) with \mathbf{R}_1 joining Sc, \mathbf{R}_2 and \mathbf{R}_5 free.

The species are both close and variable and not fully understood. Some species are sexually dimorphic, the female being paler, and often narrower winged than the male. There appear to be good structural characters, but they have never been worked out. The following key is incomplete but may be of some use as a guide. The genus is a very difficult one in the South and is not well understood. but in New York and the North generally only the well-characterized S. sordidellus, melinellus, and longirostrellus seem to occur.

Key to the species

1. Hind wing infuscated, fore wing fuscous (males).
2. Expanse 25 mm
2. Expanse 40 mm2. sordidellus.
1. Hind wing white.
2. Expanse over 50 mm.; yellow with brown markingsl. maximellus.
2. Smaller.
3. Postmedial line straight, running from apex to a dot in the fold two-
thirds way to margin; wings broader; sexes similar6. longirostrellus.
3. Postmedial line fading out, or running into a longitudinal streak; wings
narrower, especially in females.
4. Males.
5. Fore wing smoky4. unipunctellus.
5. Fore wing yellow
4. Females.
5. Expanse over 35 mm2. sordidellus.
5. Expanse under 35 mm.
6. Śmoky or straw yellow4. unipunctellus.
6. Bright ochre, or overlaid with brown, leaving at least a yellow
costa

1. S. maximellus Fernald. Male unknown. Dull ochre with a blackish longitudinal shade from base to apex; an oblique series of blackish postmedial dots

on veins, the uppermost 1 mm. before the apex; those on Cu_1 , Cu_2 , and A obscure; sub-basal, antemedial, and postmedial black dots in fold, the latter well beyond, the series on the veins; and black terminal points. 55 mm.

Southern States in June.

2. S. sordidellus Zincken. Male with blunt squarish wings; dull fuscous; fore wing with black discal and terminal dots; a postmedial series from just before, apex to three-fourths way out on inner margin; obscure below. Hind wing con-

colorous, slightly paler at base. 30-25 mm. Uncus pointed; gnathos emarginate; bearing a spine nearly as long as the suncus; transatilla with rudimentary free lobes; juxta with a short, pointed ventral spine. (Fig. 301).

Female with narrow, lanceolate wings, the postmedial series reaching inner margin three-fifths way out, normally obscure, or replaced by an obscure dark line, irregular below. Fore wing with ground sometimes heavily dusted on an ochreous base. Hind wing white, shaded with gray toward margin. 35–48 mm. August; June in the South.

Southern States; Illinois, Woods Hole, Massachusetts (dwarf). 3. S. nitidellus Dyar. Male superficially like that of S. sordidellus, but smaller; fore wing striate in two shades of fuscous; practically immaculate otherwise. 25-30 mm. Gnathos not emarginate, running into a short broad spine, half as; long as the uncus; transatilla with two heavy backward-directed hooks, juxta

with a large blunt spine swollen at the tip. (Fig. 302). Female rather larger than male with narrow lanceolate wings, fuscous, or, rarely, clay-color, dusted with fuscous; with a blackish shade from base to apex. Postmedial line represented by an obscure shade, and a more distinct dot in the fold. Terminal dots weak. 35 mm. (dispersellus Robinson in part?).

Hampton, New Hampshire, to Texas.

S. clemensellus Robinson is unknown to me; as described, it must be closely similar to the preceding species, but perhaps with more distinct pale streaks on (Size and locality not given). the veins.

S. roscidellus Dyar, from the Gulf Strip, is also blackish with pale yellow veins, but the hind wing is almost wholly white. If I have it correctly determined the uncus is blunt, the spine on the gnathos almost as long as the uncus and well set off, the hook on the transatilla strong, and the spine on the juxta even in width for most of its length.

4. S. unipunctellus Robinson. Unicolorous dark fuscous, with a conspicuous black discal dot. Uncus shorter than spine of gnathos, which is even in width and arises from a deep notch in the gnathos. Spine of juxta very large and pointed; transatilla with a pair of large rounded lobes notched at the apex. The female appears to be dimorphic, as both fuscous and straw-yellow females have been taken with males of this appearance. (Fig. 303). Male 20 mm., female 25 mm.

Southern States.

S. tripunctellus Robinson. Powdery fuscous on a cream-colored base, show. ing two distinct blackish dots in the fold as well as the discal dot. Uncus short, blunt, and very broad; gnathos deeply emarginate, with a short tapering spine; transatilla bearing very large hooks, and spine of juxta swollen toward the tip. Female straw yellow, apparently indistinguishable from the yellow form of S. unipunctellus. 20-25 mm.

Southern States.

5. S. melinellus Clemens. Male straw yellow, with a faint grayish postmedial line extending down from apex, and fading out below; slightly narrower winged than S. longirostrellus, but not constantly distinguishable on superficial characters. A discal black dot and two dots in fold. Genitalia similar to those of S. tripunctellus, but uncus narrower toward the base, spine of gnathos stouter, hooks of transatilla hardly as large, and spine of juxta more swollen at the tip. (Fig. 304).

Female variable. Very narrow-winged. Ground normally light yellow, with a more or less distinct brown shade through the middle of the fore wing from the base to the apex 20-28 mm. Hind wing white in all forms; rarely, with black terminal points.

Typically, the longitudinal shade is moderately developed and diffuse; in variety albocostellus Fernald, it is defined on the upper side, setting off a clean-cut pale yellow costal stripe from base to apex; in variety dispersellus Robinson, the ground is more or less dusted and suffused with fuscous; variety pallulellus Barnes and McDunnough is immaculate pale yellow; and variety uniformellus Dyar is fuscous, immaculate except for the blackish discal dot.

June to early August.

Northern States; Nova Scotia to Virginia. New York: Buffalo, Otto, Ithaca. 6. S. longirostrellus Clemens. Light straw yellow, more or less dusted and shaded with fuscous. A grayish postmedial dot in the fold; a blackish antemedial dot in the fold; and discal dot. Hind wing white. Uncus bluntly truncate, somewhat heart-shaped in posterior view, with a small nodule in place of a spine on its ventral side, transatilla bearing a pair of very large circular lobes; penis much more coarsely spinulated than in the rest of the genus, and containing a very heavy spine. Juxta apparently unarmed. Gnathos much modified, without a spine. (Fig. 305).

Female similar to male, the wings slightly narrower.

This species is easily distinguished by the blunt uncus in the male, which is almost always visible without dissection, and by broad blunt wings in the female. It is superficially close to the European S. forficellus, for which it has been commonly mistaken, but shows no resemblance in structure. I cannot tell by the description whether S. amblyptepennis Dyar is the same.

June and July.

Northern States; Canada (Montreal and perhaps Quebec) to Pennsylvania. New York: Newport, Newcomb, North Creek, Niagara Falls, Otto, Ithaca, Little Falls, Albany, New Windsor,

Subfamily CHRYSAUGINÆ

Antennæ normally laminate and fasciculate in male, rarely with any special rantenne normaty laminate and fasciculate in male, rarely with any special modification. Palpi various, not strikingly dimorphic in the sexes, third segment normal, fully scaled; maxillary palpi completely absent; tongue developed. Ocelli most often present. Fore wing in female with \mathbf{R}_{s-s} stalked, typically similar in male, but almost invariably more or less modified sexually; with distorted venation and costal lobes, tufts, etc.; often with a funnel-like structure at base of costa (fig. 306). Male retinaculum often modified into a loop. \mathbf{M}_{z} and \mathbf{M}_{s} free, stalked or united. Let \mathbf{A} completely absent 2d A minimized the model. stalked or united; 1st A completely absent, 3d A primitively forked, the upper branch becoming coincident with 2d A. (In specialized forms 3d A runs up into 2d A, and then the lower fork separates again from 2d A as a free spur.) Hind wing with no fringe on base of Cu; Sc and R anastomosing in our species; 1st A completely lost, and frenulum often thickened and modified in male, multiple in female.

Larvæ normally leaf rollers, with uniordinal hooks in two transverse bands, on prolegs; adfrontal sclerites reaching vertex, and front, nearly to vertex; vii represented by a single seta on meso- and metathorax; ninth segment of abdomen with seta i about midway between ii and iii; with three well-separated and welldeveloped lateral setæ. Pupa not fully studied.

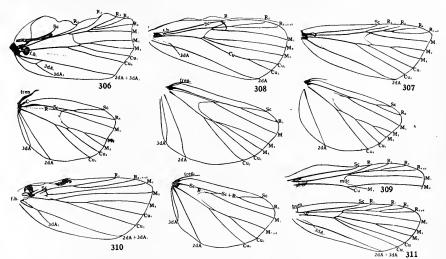
Key to the genera

- 1. Hind wing with all eight veins preserved.
 - 2. Fore wing with Cu_1 and Cu_2 stalked; M_2 and M_3 arising from cell sepa-
 - 2. Fore wing with \mathbf{M}_2 to \mathbf{Cu}_2 all arising separately from cell......4. Tosale.
 - 2. Fore wing with \mathbf{M}_2 and \mathbf{M}_3 stalked.

Outer margin irregular, all veins present (fig. 306).....3. Clydonopteron.
 Fore wing with even margins; 11 veins only (fig. 307).....6. Arta.
 Hind wing with M₂ and M₃ completely fused; 7 veins.
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2. Hind wing with Sc and R fused halfway to apex (fig. 310).

2. Hind wing with Sc and R completely fused......8. Polloccia.



FIGS. 306-311. CHRYSAUGINÆ

306, Clydonopteron tecomæ, \mathcal{J} , venation; **307,** Arta statalis, \mathcal{J} , venation; **308,** Galasa rubidana, \mathcal{J} , venation; **309,** G. rubidana, \mathcal{Q} fore part of wing; **310,** Condylolomia participialis, \mathcal{J} , venation; **311,** Polloccia alticolalis, \mathcal{J} , venation of fore wing.

3. CLYDONOPTERON Riley

(Salobrena, in part)

Palpi oblique, as long as head and thorax together; second segment heavily tufted above and below, toward tip; third segment blade-like, long and heavily tufted above and below. Middle and hind tibia and metatarsi tufted. Fore wing (fig. 306) sexually dimorphic; male with a hollow eone at base of costa; retinaculum modified into a ring; costa with a deep triple noteh at middle; apex subfalcate, and outer margin bent at \mathbf{M}_3 ; wings very broad; Sc short, but well developed, free; \mathbf{R}_1 running to the middle notch on the costa; \mathbf{R}_2 arising near end of cell; \mathbf{R}_3 to \mathbf{M}_1 stalked, \mathbf{R}_4 and \mathbf{R}_5 the farthest; \mathbf{M}_2 and \mathbf{M}_3 stalked (unlike Salobrena); 3d A strongly forked, the upper fork distinct and running in to 2d A; cell half as long as wing. Female without the cone; costal notehes and falcations of apex weaker, venation much like that of the male. Hind wing in both sexes with Sc and R shortly fused, \mathbf{M}_2 and \mathbf{M}_3 stalked, otherwise normal; lst A obsolete. The genus is hardly distinct from the South American and Texas genus Salobrena.

1. C. tecomæ Riley. Fore wing with basal two-thirds mottled with yellow and brown, and shaded with orange; postmedial line a short fine white streak at

costa, then evenly and slightly concave, slightly paler and fine, to inner margin; defined by a heavy dark shade before and a fine line beyond it. Wing beyond postmedial line more nearly even purplish fuscous. Hind wing dark. (H 48:11.) Larva a leaf roller on trumpet creeper.

District of Columbia, Illinois, Missouri, and south.

4. TOSALE Walker

(Siparocera Grote)

Palpi obliquely upturned to middle of front, short; tibiæ tufted as in Clydonopteron, but the hind tibiæ less heavily so; fore wing of male with a very large hollow cone at base of costa, retinaculum ring-like, and frenulum thickened and modified, the outer margin nearly even and apex not falcate; \mathbf{R}_2 stalked in female, free in male; \mathbf{M}_1 from cell; \mathbf{M}_2 and \mathbf{M}_3 connate; **3d A** swinging up to meet **2d A** and obliterating **3d A**₁. Hind wing with **Sc** and **R** shortly anastomosing; all the other veins free. Female similar, without the cone and modified frenulum and retinaculum. In *T. oviplagalis* the male has a large oval patch of black sex scales on upper side of hind wing and one on under side of fore wing.

1. T. oviplagalis Walker. Antemedial space yellow-brown to olive, bounded by a white, evenly excurved antemedial line; basal sixth pinkish, the rest of the wing dull light brown; median area paler and grayer, especially toward costa; postmedial line twice as far from apex as from anal angle, convex in the middle and concave toward the margins; even. Some gray shading before postmedial. line and some brown beyond it toward costa. Hind wing fuscous, with a large black patch in male. 16 mm. (S. nobilis Grote, Asopia anthecioides Grote.) (H 48:33).

May to July.

New York to Illinois, and south to South America. New York: Ithaca, Long Island.

5. GALASA Walker

(Cordylopeza Zeller)

Palpi rather short, beaklike, second segment tufted on upper side. Middle tibiæ and metatarsi very heavily tufted with black scales; hind metatarsi very heavily tufted, and tibiæ and second joint of tarsi more lightly; fore wing of male (fig. 308) without a cone at base of costa (a rudiment visuble on bleaching), retinaculum large but normal, buried in loose scaling; Sc weak, normal; \mathbf{R}_1 and \mathbf{R}_3 lost (Hampson figures \mathbf{R}_1 as present, but I cannot find it), cell two-fifths length of wing, \mathbf{M}_1 free; \mathbf{M}_2 and \mathbf{M}_3 connate or shortly separate; \mathbf{Cu}_1 and \mathbf{Cu}_2 long-stalked; **2d** A and **3d** A anastomosing at a point, obliterating **3d** A₁. Hind wing with fremulum not modified; Sc and R anastomosing, lower part of discocellular vein closely parallel to lower edge of cell, and \mathbf{M}_2 and \mathbf{M}_3 more or less distinctly stalked. Fore wing with two shallow notches, with a curved pencil of hair (concealed by the scaling) curving down across the surface of the wing from the second one. Female with palpi longer, curved down at tip, tibiæ and tarsi less heavily tufted; fore wing (fig. 309) with a single broad shallow notch; \mathbf{R}_1 arising near the tip of the cell; \mathbf{R}_2 stalked; \mathbf{R}_3 lost; **3d** A connected to **2d** A by a short section of **3d** \mathbf{A}_1 ; frenulum triple.

1. G. nigrinodis Zeller. Fore wing orange-red, at base, dull crimson outwardly; the concave part of costa finely gray-edged; antemedial and postmedial lines white, normally defined with fuscous, sometimes distinct at costa only. Antemedial line waved, postmedial finely dentate. Hind wing dirty white, shaded with fuscous. often with a distinct postmedial gray shade. 15-18 mm. (*rubidana* auct. nor Walker).

June and July: September.

New Hampshire and Ontario to Illinois. Missouri, and south. New York: Buffalo, Ithaca, Big Indian Valley, Schenectady. Nassau. Highland, New Windsor, Florida, Katonah, New York City.

6. ARTA Grote

(*Heliades* Ragonot)

Palpi quite short and porrect or drooping; front smooth-scaled, tibiæ practically smooth-scaled; fore wing (fig. 307) triangular, and without striking sex modifications; Sc long, normal, reaching well beyond middle of costa; one branch of **R** free, three stalked, one lost; \mathbf{M}_1 free; \mathbf{M}_2 and \mathbf{M}_3 stalked; 2d A and 3d A becoming coincident. Hind wing with Sc and R anastomosing more than half way from cell to apex; \mathbf{Cu}_1 shortly stalked with \mathbf{M}_{2+3} , and 1st A lost.

1. A. statalis Grote. Light pinkish brown, with a fine straight pale line at three-sevenths, and a similar slightly waved one at five-sevenths; hind wing grayish, with traces of postmedial line. 15 mm.

The moth flies about Myrica in July and August, and is locally common in Massachusetts. The southern distribution is uncertain on account of confusion with A. olivalis and other related species.

New Hampshire to Illinois and south. "New York" (Grote).

2. A. olivalis Grote. Light olivaceous, with faint white lines. 14 mm.

July. and August.

Maryland; North Carolina; Texas.

7. CONDYLOLOMIA Grote

Front slightly tufted; legs with thick scaling but without definite tufts; palpi short, porrect, and hairy below. Fore wing (fig. 310) narrower than in Arta; in male, with a small portion folded over a third way out on costa, bearing a blackish hair-tuft, and with a rudiment of a basal cone; retinaculum heavy, frenulum normal; fore wing with cell minute, one-fourth length of wing, very short toward costa; \mathbf{R}_1 lost; \mathbf{R}_2 running into Sc, which bifurcates, both branches running to costa; \mathbf{M}_1 widely separated from \mathbf{R}_{3-3} ; \mathbf{M}_2 from lower angle of cell; \mathbf{Cu}_2 shortly stalked with $\mathbf{M}_3 + \mathbf{Cu}_1$; 3d A fused for a short distance with 2d A; with a strong free tip. Hind wing with Sc and R moderately anastomosing; \mathbf{M}_1 arising from the free sector of R, Cu, short-stalked with \mathbf{M}_{2+3} . Female with \mathbf{R}_1 becoming coincident with Sc; the other three radials stalked; one lost; \mathbf{M}_2 and \mathbf{M}_3 long-stalked.

1. C. participialis Grote. Pale grayish olivaceous, darker toward the margin; with a diffuse darker antemedial line, and a darker, followed by a pale luteous, postmedial line, both slightly irregular. Hind wing darker gray. 12-15 mm.

Moth flying over Myrica in July and August. Massachusetts to Pennsylvania and Illinois. New York: Rock City (Cattaraugus County), Ithaca, Katonah.

8. POLLOCCIA Dyar

Palpi much longer than head, beaklike, drooping, legs slender. Wings (fig. 311) not much modified sexually; fore wing with Sc free, one R from tip of cell, three stalked and one lost; M_1 separate; M_2 and M_3 stalked; Cu, and Cu₂ approximate from lower angle of cell; cell one-third length of wing. 3d A shortly anastomosing with 2d A, with free tip. Hind wing with Sc and R completely fused, M_1 moderately stalked with them; Cu₁ and Cu₂ free; 1st A preserved.

1. P. alticolalis Dyar. Clay-color. more or less suffused with pinkish, and shaded with light gray, partly defining the ante- and postmedial lines; antemedial line

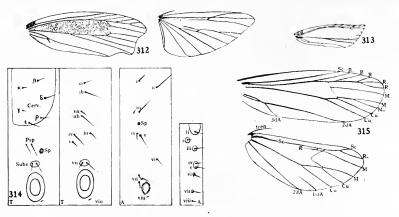
dentate, black, preceded with luteous, and rather broad at costa, interrupted by a fovea in cell of male; postmedial line blackish, followed by luteous; perpendicular at costa, then strongly excurved opposite cell, and sinuate below. Hind wing pale. 13 mm. July and August.

New Hampshire to Virginia and northwestern Ontario. New York: Jamestown (McElhose, Dyar det.), Ithaca (W. T. M. F.).

Subfamily GALLERIINÆ

Male antennæ simple. not modified; ocelli always absent, maxillary palpi obsolete in male; often distinct but small, in female; tongue very weak or obsolete. Body normally stout, with deep vestiture. Palpi sexually dimorphic, except in a few primitive exotic species, upturned in male, rather short, with third segment more or less rudimentary; upturned or porrect in female, and normally developed, sometimes long and beaklike. Fore wing with R_3 to R_5 stalked, 3d Å forked, with the upper fork joining 2d A, or simple (the lower fork lost) and becoming coincident with 2d A. 1st A lost. Hund wing with male fremulum normal, female multiple; Sc and R usually very shortly anastomosing; middle discocellular vein curving far in toward base of wing, the upper and lower parts nearly parallel with R and Cu, the middle part short and transverse, sometimes nearly obsolete. Base of **M** usually preserved as two thickenings connected with the deepest part of the curve of the discocellular. M_2 and M_3 stalked or fused; Cu_1 sometimes stalked; 1st A preserved; a strong fringe on base of Cu.

The larvæ (fig. 314) are normally scavengers, or inquilines in bee and wasp nests. They include a few species injurious to dried food, and the well-known bee moths, which sometimes do a good deal of damage by cating the wax, etc., and destroying and dirtying the comb in ill-tended beehives.



FIGS. 312-315. GALLERIINÆ AND MACROTHECINÆ

312, Galleria mellonella, J, venation and sex patch; 313, Aphomia sociella, J, venation of fore wing; 314, Galleria mellonella, seta map of larva; 315, Macrotheca flexilinealis, venation

Prolegs with uni- or biordinal hooks, in a complete ellipse; prespiracular setæ of prothorax and iv and v of abdomn in a horizontal line; vii of meso- and metathorax of two setæ; i to iii of ninth segment of abdomen forming an equilateral

triangle, with iii directly below ii. Pupæ with tongue short, and pilifers rudimentary, maxillary palpi present, small; dorsum of body with a prominent median ridge, and segments covered with small spines.

The subfamily is a small one, widely separated from the other Pyralids (except the Macrothecinæ). Our few genera are closely related and do not fully represent the group. It has been suggested that Linnæus' name "Tinea" should be applied to this group, as the bee moth was included in his genus, and was the form best known as "Tinea" by the ancients; but I have preferred to keep to familiar usage, especially as Tinea was regularly used in an inclusive sense for a variety of stored food pests and clothes moths.

Key to the genera

- - 2. Fore wing with all veins; R_2 from cell.
 - 3. Cell about two-thirds length of wing in both sexes; M_2 and M_3 stalked.
 - 10. Melissoblaptes.
 - 3. Cell nearly reaching outer margin in male; with M_2 and M_3 well separated and often rudimentary; in female, M_2 and M_3 connate from lower angle of cell.

 - 2. Fore wing with a dorsal vein lost; no sex scaling in cell.....(12) Corcyra.

9. GALLERIA Fabricius

Antennæ simple with a scale-tuft on scape in both sexes; palpi of male upturned, minute, hidden in frontal tuft; palpi of female forming a short beak. with third segment short; fore wing (fig. 312) in male with a slight thickening at base of costa, cell thickened and with sex scaling below, three-fourths length of wing; middle discocellular distinct and angled in; M_1 and M_2 well-developed, free; R_2 free; apex bluntly subfalcate (at R_5 and M_1); outer margin produced at Cu_2 , concave above and below; tip of 3d A free. Hind wing with M_2 and M_3 longstalked; Cu_4 free; discocellular extending more than half way to base of wing. Female without thickenings or sex scaling, the tooth on Cu_2 and concavity of the outer margin less distinct; cell rather broader and shorter. A row of small raised tufts in the fold.

1. G. mellonella Linnæus. Dull gray, strigose; inner margin, below fold, ycllower; tufts on fold often blackish; postmedial line represented by a series of obscure blackish bars. Hind wing fuscous, pale at base. 25-35 mm., female larger. (*Tinea* Linnaeus; *Tortrix cercana* Linnaeus.) (H p. 406, f. 226.)

The larva is a serious pest in ill-kept beehives, eating and webbing up the comb; also a scavenger in waste wax, etc.

World-wide in distribution. New York: Fentons (Lewis County), Vicinity of Buffalo, Ithaca, New Baltimore, Flatbush. A common species wherever looked for.

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10. MELISSOBLAPTES Zeller

Palpi much as in Galleria; scape large, but smooth-scaled; fore wing with a large costal pocket in male, but cell not enlarged; \mathbf{R}_1 and \mathbf{R}_2 free, \mathbf{R}_3 to \mathbf{R}_5 stalked; \mathbf{R}_4 and \mathbf{R}_s the farthest; \mathbf{M}_1 arising distinctly below angle of cell; \mathbf{M}_2 and \mathbf{M}_3 in our species connate, typically stalked; Cu_2 arising well toward base. Free tip of 3d A lost. Hind wing with cell extremely short in middle, M_{2+3} stalked with Cu_1 . The larva lives in a sand tube at the surface of the ground. The normal food is unknown, but it will eat dried insects, etc.

1. M. fuscolimbella Ragonot. Whitish, shaded with brown; marginal area darker brown; lines fine, dark, defined, the antemedial line edged before, and postmedial line beyond, with pale; antemedial line obtusely angled below cell; postmedial dentate; discal spots fusing into an irregular triangle. Hind wing fuscous, dark at apex. Female unknown. 24 mm.

North America.

11. PARALISPA Butler

(*Paralipsa*, by misprint)

Similar to Melissoblaptes; scape somewhat tufted; palpi moderate, porrect, about as long as the large frontal tuft. Fore wing rounded-lanceolate, with convex costa; cell long, broad in male, and extended out below well toward margin; scaled as in Galleria; large but normal in female.

This genus is hardly distinct from the preceding and following, which are combined with it by Hampson. The three nominal species are not well known and

may be only varieties. 1. P. terrenella Zeller. Cell two-thirds length of wing (\mathfrak{Q}) . Brownish gray, somewhat streaky; with a large blackish basal dash, more than one-third length of wing, and stronger in the male; a fine irregular outer line, deeply rounded out on middle half, the wing shaded with brown before it, and grayer beyond. Reniform represented by a black stria. Hind wing fuscous in male, pale in female. 28 mm. (Melissoblaptes furellus Zeller.)

End of June to August.

New York; Georgia. New York: Ithaca, vicinity of Albany.

2. P. decorella Hulst. Similar to P. terrenella; described as having a distinct dentate antemedial line, but postmedial line obsolete. 28 mm.

Unknown to me.

Southern Ontario. New York: Buffalo (Grote).

3. P. fulminalis Zeller. A more contrasty form than the two preceding, the markings as in *terrenella*, but strong on a pale luteous ground; more or less emphasized with dark gray; the cell often filled out with a dark gray blotch, and outer margin more or less shaded with dark gray. Hind wing darker and grayer, ¿ 22 mm. Permale

Marvland to Pennsylvania.

anter margine eit offt ni "Hut the

L. G. mellonella

lower, trift, on fold

12. APHOMIA Hübner

Male palpi as in Galleria, those of female rather long, porrect: Todgue and maxillary palpi relatively well developed. Fore with "of male "flge" 313) with medials and cubitals all well separated; the cell extending "almost "to" margin" at M_2 and M_3 , nearly obliterating them. Cell, very broad, "Hind" wing "very broad; trapezoidal. Otherwise like Melissoblaptes." Way method with "to be oblighted" in the following "almost "to be the first of the first

female pale gray shaded with olivaceous and russet; the lines strongly dentate, close together, black when distinct; discal dots strong and black, or lost in a blackish shade; hind wing gray in both sexes. 30 mm., the female usually larger. (colonella Linnæus.)

Larva in nests of bumblebees and wasps, but not with honeybees; more rarely in dried stores, cotton, books, borings of wood-borers, etc.

Nova Scotia; Massachusetts; probably generally distributed; introduced from Europe.

Corcyra cephalonica Stainton (*Tineopsis theobromæ* Dyar) is a common European dried-food pest, to be expected in our territory.

13. ACHROIA Hübner

Easily distinguished by the want of the frontal tuft. Antennæ long, with a scale-tooth on scape; palpi minute in male, shorter than eye in female; legs slender. Fore wing of male oval; broadest toward base; 11 veins, one radial lost, all the others free; \mathbf{R}_1 arising opposite \mathbf{Cu}_2 , \mathbf{Cu}_1 as near \mathbf{Cu}_2 as \mathbf{M}_3 , \mathbf{M}_2 well separated. Female with fore wing more evenly elliptical, with \mathbf{M}_2 and \mathbf{M}_3 stalked; hind wing with \mathbf{M}_{243} stalked with \mathbf{Cu}_1 .

1. A. grisella Fabricius (The lesser bee moth). Pale gray-brown; the head light yellow, contrasting; immaculate. Hind wings pale, with gray fringe. \mathcal{J} 17, \mathcal{Q} 21 mm.

Caterpillar with the habits of *G. mellonella* and associated with it. It will also eat dried apples, raisins, crude sugar, and apparently also dried insects.

World-wide in distribution.

Subfamily MACROTHECINÆ

Near the Gallerinæ, but slender species with close-scaled bodies. Palpi in male very short, with a heavy tuft on second segment above and below; third segment very short, naked, and turned inward; palpi in female very long and beak-like; maxillary palpi and tongue absent. Fore wing (fig. 315) narrow and thin, with arched costa and rounded apex and outer margin. Macrotheca has \mathbf{R}_2 shortly stalked with \mathbf{R}_{a-5} , \mathbf{R}_4 and \mathbf{R}_5 the farthest; \mathbf{M}_1 widely separated; \mathbf{M}_2 and \mathbf{M}_3 stalked; cell about two-thirds length of wing; 1st \mathbf{A} not chitinized, but with a wide space in its position; 3d \mathbf{A} anastomosing with 2d \mathbf{A} but with a strong free tip, reaching margin. Hind wing with cell extremely large, two-thirds length of wing; Sc and \mathbf{R} strongly anastomosing; the base of \mathbf{R} reduced to a short spur; **Mcv**, sharply right angled in middle, with a single rudiment of the base of **M** running to its angle; \mathbf{M}_2 lost; \mathbf{M}_3 and \mathbf{Cu}_1 nearly connate; fringe on **Cu** obsolete; anal region reduced, especially toward the base, and the veins very weak.

The only known larva of the group is predacious on scale insects.

14. MACROTHECA Ragonot

1. M. unipuncta Dyar. Powdery gray; antemedial line far out, sometimes obsolete, black and dentate when distinct; postmedial obscure, irregularly sinuous, followed by an imperfect second line. Discal dot a large black irregular spot. Hind wing fuscous in male; female not seen.

July.

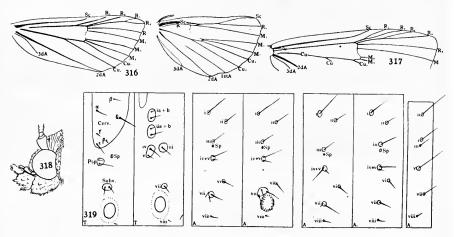
Western Pennsylvania. Apparently a form of this species occurs in Arkansas, but its discal dot is small. It is not known from the intervening territory.

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Subfamily GLAPHYRIINÆ

(Homophysidæ; Pyraustinæ; Crambinæ, in part)

Head smooth (fig. 318); ocelli well developed; antennæ of male not modified, with a rather small smooth scape; tongue normal, scaled at base (absent in the western genus Chalcoëla; labial palpi moderate or rather long, the segments almost equal in length, the first two broadly scaled, the third much more slender, often pointed, upturned in the eastern species, porrect in Chalcoëla. Maxillary palpi typically as long as a segment of the labials, obliquely porrect and rough scaled; rarely rudimentary. Wings usually broad and ample, coarsely scaled, sometimes with metallic scaling at the margin of the hind wing, as in some Nymphuline. Fore wing with \mathbf{R}_1 and \mathbf{R}_2 free, or stalked with each other (figs. 316, 317), rarely united; \mathbf{R}_3 and \mathbf{R}_4 stalked, \mathbf{R}_5 free, divergent from \mathbf{R}_{3+4} ; rarely with



FIGS. 316-319. GLAPHYRIINÆ

316, Lipocosma fuliginosalis, venation; **317**, Glaphyria glaphyralis, venation of costal part of fore wing; **318**, Dicymolomia julianalis, head; **319**, D. julianalis, seta map of larva (ex coll. Claassen)

 \mathbf{R}_2 stalked with \mathbf{R}_{3+i} . 1st A lost; 3d A free, weak. Hind wing ample, with a large broad cell; Sc and R strongly anastomosing; \mathbf{M}_1 connate with \mathbf{R} ; \mathbf{M}_2 and \mathbf{M}_3 closely approximate or shortly stalked; middle discocellular long and moderately bent, not tubular; frenulum of female multiple. Upper side of hind wing with a more or less defined fringe of long hair just below Cu, ending in a larger or smaller number of spatulate hairs or scales in the fold, near or below end of cell: a similar series of hairs on 2d A, also usually ending in a group of scales.

smaller lumber of spatialize lumber of A show usually ending in a group of scales. The spatulate hairs or scales are more or less deciduous, but distinct in fresh specimens of all species; the characteristic palpi will distinguish them from the most similar Pyraustinæ, the broad rounded wings from all our Nymphulinæ, to which they are probably most closely related. The Crambinæ and the Ancylolomiinæ are not, in fact, close relatives, and all the species known to me may be distinguished by the open cell or widely spaced **R** and **M**, of the hind wing. (Larva, fig. 319.)

Key to the genera

1. Fore wing with R_1 and R_2 stalked, rarely united (fig. 317); no metallic scaling 1. Fore wing with R_2 free from R_1 (fig. 316), rarely short-stalked with R_{3+4} .

16. Lipocosma.

15. GLAPHYRIA Hübner

(Homophysa Guenée)

Characters of the subfamily. Palpi with slight projecting angles formed by the vestiture of the first two joints, third joint obliquely upturned. Hind wing with \mathbf{M}_2 and \mathbf{M}_3 approximate. Fore wing with apex rounded or marked. Maxillary palpi always moderate in size, rough-scaled, and porrect. Fore wings with a yellow or yellowish ground color, with two white transverse lines; the antemedial line usually waved, the postmedial one denticulate, and both normally finely dark-edged.

Key to the species

- 1. Antemedial space below costa, at least, and sometimes whole disc brown or fuscous.
- medial area.

 - 3. Fringe mixed fuscous and white, appearing striate in perfectly fresh

1. G. glaphyralis Guenée. Lemon vellow, more or less marked with cream-white: sometimes cream-white with the yellow only defining the markings. Antemedial line waved; postmedial sinuous and denticulate; white terminal dots. Fringe somewhat paler. Sometimes lightly shaded with pale brown. Hind wing similar on the dorsal part, toward the costa shading into cream-white. 15-18 mm.

July and August. Florida specimens are usually bright yellow; northern ones mostly cream-white.

New York to Illinois and southward. New York: Otto. Ithaca.

2. G. sesquistrialis Hübner. Ochre-yellow; duller than G. glaphyralis, the lines similar, distinctly defined with light brown, and the ground somewhat shaded with light brown, but without whitish shades. A distinct broken black terminal line, preceded by white dots; fringe ochre to light brown. 15-18 mm.

June to August; October.

Massachusetts to Illinois and south and west. New York: Ulster County.

3. G. psychicalis Hulst. Bright ochre yellow; lines very fine, normal, sometimes partly defined with brown, and more or less broken; more wavy than in the last two species. Terminal dots strong, black and white; fringe mixed fuscous and white. Hind wing frequently more or less suffused with light fuseous. 12 mm.

July and August.

Massachusetts to Ontario, Illinois, and Florida.

4. G. lentiflualis Zeller. Ochre: antemedial line obscure. not dentate in the fold; postmedial line rather evenly sinuate, only slightly paler, and preceded by a darker

line; terminal dots obscure; fringe fuscous on basal half, white-tipped. Hind wing similar toward margin, pale at base. 15 mm.

June. Perhaps a synonym of G. invisalis Guenée, of South America.

Central Illinois; North Carolina; Iowa; Texas. 5. G. fulminalis Lederer. Disc of wings yellowish to dark umber brown. Base of wings contrasting, pale yellow, brighter than in L. fuliginosalis, the paler area extending out to the antemedial line along the costa. Outer margin yellow to postmedial line, except at anal angle. Costal edge sometimes wholly yellow. Lines normal, white, defined with brown. Hind wing pale, with a brown patch crossed by a white line toward the inner margin. 10-15 mm.

Late June and July.

Connecticut to Illinois, Florida, and Texas. "New York" (Grote.)

I have examined a specimen in which the radial veins were reduced to three: \mathbf{R}_{1+2} , \mathbf{R}_{3+4} and \mathbf{R}_{5} .

6. G. peremptalis Grote. Similar to G. fulminalis, the fuscous area extending to costa, almost to base, and diffusely shading into the lighter brown outer third. 12 mm.

July.

Western Pennsylvania to North Carolina and Texas.

16. LIPOCOSMA Lederer

(With Symphysa Hampson, in part; Egesta Ragonot)

Intermediate between Glaphyria and Dicymolomia. Maxillary palpi moderate to rudimentary; labials oblique, with the three segments equal, or with the first two segments decidedly longer; closely upturned beyond the vertex. Tongue and wing form as in Glaphyria; \mathbf{R}_2 well separated from \mathbf{R}_1 (fig. 316), sometimes stalked with \mathbf{R}_{a} . Spatulate scale-tufts on hind wing well developed. Ocellus larger than in Nymphula.

Key to the species

1. Ground even fuscous b	rown.	
2. Reniform a large wh	ite lunulel.	reniculalis.
2 Reniform obsolute		2 orinalie

1. Base white; apex at least whitish.

2. Median area yellow to fuscous, between cell and inner margin....4. sicalis.

1. L. reniculalis Zeller. Powdery dull brown; antemedial line dentate, postmedial sinuous. white, defined with dark brown; a large white discal lunule, with a small white spot below it in the fold; terminal line black, broken, with some white before it. Fringe concolorous. Hind wing similar, becoming whitish at costa. Palpi closely upturned, maxillary palpi obsolete; spatulate tufts on hind (Homophysa Hampson, Symphysa Hulst.) wing weak. 15 mm.

June and August.

Central Illinois and North Carolina to Texas; a race in Arizona.

2. L. eripalis Grote. Similar to L. reniculalis, with a slight violaceous tint when fresh; lines finer, antemedial extended far out in cell, and more dentate. No reniform spot or dot below it. Hind wing with only part of outer line visible. Fringes with a strong white line in outer part. Palpi obliquely porrect, rough; maxillary palpi rough and well developed; raised tufts on hind wing slight. Front more oblique than usual. 18 mm. (Symphysa Hampson; Egesta Barnes and McDunnough.)

Late June.

Virginia to central Illinois and Missouri, south and west.

3. L. adelalis Kearfott. Raised tufts of spatulate scales on hind wing large and black; palpi rough, oblique, beak-like, with large maxillary palpi. Ground white.

Fore wing with obscure dark ante- and postmedial lines, running about as usual; slight gray shades before antemedial line and beyond postmedial. Discal dot black, strong; one or two black terminal dots near apex. Hind wing white, with a small gray subterminal shade near the black tufts. 15 mm. (Symphysa Kearfott.)

Late July and early August. Larva in a flat oblong case; constricted near each end; on lichens.

Anglesea, New Jersey.

4. L. sicalis Walker. Palpi oblique, maxillary palpi very small; tufts strong. Fore wing cream-white, shaded with clay color in outer part, leaving some white along postmedial line and on margin; medial area largely clay-color, more or less dark; postmedial line, when complete, with a long tooth in the fold running up toward the lower angle of the cell. Hind wing similar, with a distinct black postmedial line; median area dusted with black, usually leaving a distinct white discal dot. 18 mm.

May to July.

New York to Illinois, Iowa, and Texas. "New York" (American Museum of Natural History.)

5. L. fuliginosalis Fernald. Maxillary palpi distinct, labial palpi oblique; tufts on hind wing strong, whitish, heavily shaded with fuscous, leaving only the base halfway out to the antemedial line, apex, and sometimes onter part of costa and outer margin, white; lines blackish as before, usually with broken terminal line. Hind wing normally with blackish below the end of the cell to the inner margin, and a marginal shade toward inner margin; sometimes mostly fuscous. 12 mm.

June and September.

Southern New Hampshire to Michigan, south to New Jersey and Missouri. New York: Otto.

17. DICYMOLOMIA Zeller

Venation as in Lipccosma; tufts on upper side of hind wing strong; labial palpi upturned (fig. 318), with more or less rough hair, as in Lipocosma; maxillary palpi decidedly longer than third joint of labials, rough-scaled, and truncate. Hind wing more or less distinctly notched opposite cell, with brilliant metallic scaling along outer margin below the notch.

Larva (fig. 319) with setæ vii on meso- and metathorax single; prolegs with crotchets biordinal; in a circle, shortly open on the outer side. Seta v of abdomen directly above iv; the tubercles much reduced; a single lateral seta on segment 9.

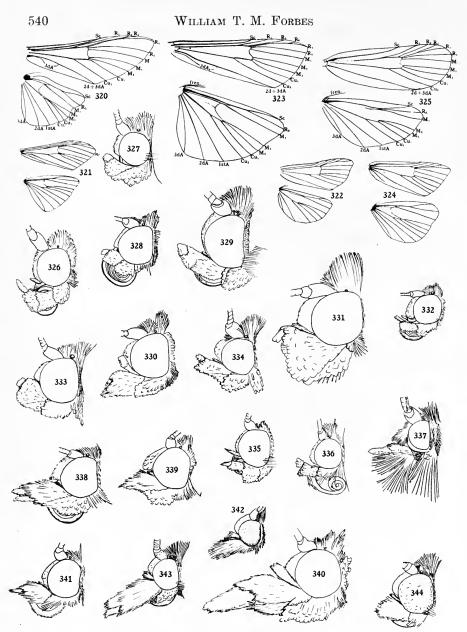
This genus, and the related western Chalcoëla, have generally been put in the Crambinæ, but they are very closely related to Glaphyria and Lipocosma in every way. The larva also appears to be near that of the Pyraustinæ, but wholly unlike the Crambinæ.

Key to the species

A single row of black dots on outer margin of hind wing.....l. julianalis. Two rows of alternate black dots.....

1. D. julianalis Walker. Light brown, shading into bright yellow at the base; median area white, dusted and suffused with fuscous brown; lines white, rather obscure, except as a definition of the median area; running as in Glaphyria; discal lunule white, followed by black dusting; some white before terminal line; fringe brown. Hind wing similar on inner half, median area dusted with black; with two strong raised tufts; the basal and costal portion shading into white. A brilliant lead colored marginal line below the notch opposite the cell, cut by four black dots in interspaces. 15-18 mm. (decora Zeller.) Late June and August. Larva in cat-tail heads; stout and unicolorous whitish,

with dark head.



FIGS. 320-344. PYRAUSTINÆ (See bottom of opposite page for descriptions)

Connecticut to Illinois, Florida, and Texas. New York: Portage, Ithaca, New Windsor, Riverdale, Bronxville.

2. D. pegasalis Walker. Base and outer margin more evenly chocolate brown; dorsal half of hind wing more evenly dusted with black, the tufts weaker. Marginal line cut by five or six black spots, and with a series of black dots on the veins, along its inner edge. 20 mm. (principialis Walker.)

July and August.

Central Illinois to North Carolina and south.

Subfamily PYRAUSTINÆ

Head smooth; ocelli present. Male antennæ normal with a few exceptions (the most striking ones in our fauna are Pilocrocis and Desmia). Front very often

oblique (fig. 344, etc.), extending out below, in a few genera conical (fig. 339), and in some western and exotic forms with a specialized process. Tongue strong, scaled; maxillary palpi small but distinct, rarely if ever as long as second segment of labial palpi. Labial palpi of various shapes (figs. 326 to 344), usually with short third joint. A distinct row of scales between antennal socket and eye. Hind tibiæ sometimes with the outer spurs in the male rudimentary. Fore wing usually triangular, ample, rarely as bluntly rounded as in the Glaphyriinæ, narrow in Nomophila (fig. 323). Hind wing of moderate size, ample in Nomophila. Fore

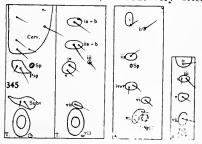


FIG. 345. HYMENIA PERSPECTALIS Seta map of larva

wing with \mathbf{R}_1 and \mathbf{R}_2 free, \mathbf{R}_3 and \mathbf{R}_4 stalked except in a couple of exotic genera; \mathbf{R}_5 free, often approximate to \mathbf{R}_{3+4} at the base, \mathbf{M}_1 approximate to \mathbf{R}_5 . Hind wing with Sc and R anastomosing except in some specimens of Loxostegopsis (fig. 325) and a few exotic forms; \mathbf{M}_1 arising from upper angle of cell; Cu with a very slight fringe at base, or none; frenulum of female multiple. All veins present in both wings.

This is the largest and the most varied subfamily of Pyralididæ, but very few

FIGS. 320-344 PYRAUSTINÆ

(See opposite page for figures)

320, Desmia funeralis, J, venation; 321, Glyphodes unionalis (Europe), venation; 322, Blepharomastix stenialis, venation; 323, Nomophila noctuella, venation; 324, Pyrausta insequalis, venation; 325, Loxostegopsis polle, venation; 326, Hymenia fascialis, side view of head; 327, Desmia funeralis, side view of head; 328, Samea ecclesialis, side view of head; 329, Pantograpta limata, side view of head; 330, Glyphodes quadristigmalis, side view of head; 331, G. hyalinata, side view of head; 332, Blepharomastix stenialis, side view of head; 333, Pilocrocis ramentalis, side view of head; 334, Diastichtis argyralis, side view of head; 335, Evergestis straminalis, side view of head; 336, Hellula undalis, side view of head; 337, Titanio pollinalis, side view of head; 338, Nomophila noctuella, side view of head; 341, Crocidophora serratissimalis, side view of head; 342, Autocosmia (?) helianthales, side view of head; 343, Phlyctania ferrugalis, side view of head; 344, Pyrausta pertextalis, seta map of larva. of the genera show striking structural characters. Superficially, the species often resemble Noctuidæ, Geometridæ, or even Pterophoridæ; but can generally be recognized by their thinner, more ample hind wings, besides the different venation. The venational characters used in this key are somewhat unstable, and the palpal characters are often almost intangible, so a certain proportion of specimens may run to the wrong genus. The "oblique" front is less convex than the "rounded" one, and gets steadily farther from the eye clear to its lower edge; the "rounded" front is strongly convex, and its lower half, at least. parallel to the curvature of the eye. (Larva, fig. 345.)

Key to the genera

- 1. Labial palpi (figs. 332, 348) obliquely upturned, with segments sharply set off, the third segment nearly as long as the second and closely scaled; maxillary palpi rough-scaled, with truncate tip, almost as long as second joint of labial palpi and much longer than third.
 - 2. Tongue very strong, much wider than palpi. .21. Blepharomastix (stenialis).
 - 2. Tongue weak, much narrower than palpi. (50. Nymphula Nymphulinæ).
- 1. Labial palpi with third joint usually smaller, sometimes very small, attached to upper side of end of second joint; sometimes lying along the tuft when
 - the second joint is tufted, but never forming the end of the tuft; maxillary palpus smaller, not broadened at end, sometimes minute (fig. 326).

 - 2. Third segment of palpus short, lying along upper side of tuft on second (fig. 334).
 - Maxillary palpus reaching to tip of labial on upper side (though not to the tip of its tuft) and more or less truncate.....21. Blepharomastix.³⁵
 Maxillary palpus shorter.
 - 4. Third segment of palpus well set off. truncate, and broadened or with a triangular tuft at tip (fig. 334); \mathbf{R}_5 well separated from \mathbf{R}_{344} .

20. Diastichtis.

- 4. Third segment of palpus not broadened at tip.
 - 5. Second segment of palpus, with its vestiture, almost as large as eye, in side view (fig. 330, 331).....27. Eudioptis, 28. Condylorrhiza.
 - 5. Second segment of palpus rarely half as large as eye in side view. 6. Fore wing with \mathbf{R}_5 approximate to stem of \mathbf{R}_{3+4} at origin; palpus

 - 7. Male without modification at middle of antennæ; Cu_1 usually nearly straight.
 - 8. Palpus projecting the width of the head beyond the head, with a long third segment; front rounded; wings blunt.

Polygrammodes.

- 8. Palpus shorter, with shorter third joint. 9. Palpus with third segment shorter than wide buried is
 - 9. Palpus with third segment shorter than wide, buried in the short vestiture of the second.

 $^{\varpi}$ Eudioptis quadristignalis may run here, but is distinguished by R_5 being closely approximate to R_4 at base.

9. Third segment of palpus longer.

10. Front oblique.

- 10. Front rounded (larger species).
 - 11. Maxillary palpi small, shorter than width of tongue; second segment of labials broader.....24. Sylepta.
- 6. Fore wing with \mathbf{R}_5 divergent from \mathbf{R}_{3+4} , lying about half way between \mathbf{R}_{3+4} and \mathbf{M}_1 ; somewhat approximate to \mathbf{R}_{3+4} in Pachyzancla, which has a triangular second segment of the palpus, unlike Sylepta.
 - 7. Second segment of palpus trapezoidal, the third lying along the upper side on the tuft.
 - 8. Male with a fold on inner margin of hind wing; ground dark. 38. Pachyzancla (perfusalis).
 - 7. Second segment of palpus rounded; the third well set off.but small.
 - 8. Third segment a third as long as second, fine-scaled and rounded.

 - 9. Maxillary palpi obsolete; cell of hind wing reaching hardly half way to outer margin.....26. Lygropia.

8. Third segment of palpus broad and coarsely scaled, triangular. 22. Conchylodes.

- 1. Palpus more or less beaklike, porrect or oblique, with third joint porrect, and lying in the center of its terminal tuft, not on the upper side (figs. 339 to 344); upper side of second joint always convex; \mathbf{R}_{3} usually divergent from \mathbf{R}_{3+4} .
 - 2. Eyes much narrower than front; truncate behind, with a wide hairy and scaly area of the head behind them (fig. 337).....41. Titanio.
 - 2. Eyes fully as wide as front, round, reaching practically to back of head.
 - Apex of hind wing angulate, subfalcate......40. Diasemia.
 Apex of hind wing rounded.

4. Front conical (fig. 339).

4. Front oblique or rounded, without a point below; maxillary palpi never as long as second joint of labials, triangular.

5. Palpi twice as long as head (figs. 340, 343).

6. Front rounded out half the width of the eye, or nearly so (fig. 342); male with a blister between \mathbf{R}_4 and \mathbf{R}_5 of fore wing.

43. Autocosmia (helianthales). 6. Front moderately rounded out or flat and oblique.

- 7. Hind legs twice as long as body......49. Lineodes.
 7. Hind legs moderate.³⁶

 - 8. Maxillary palpi slender but widened by rough scaling at the tip; more or less triangular (fig. 335)........31. Evergestis.
 - 8. Maxillary palpi small and slender.
 - 9. Fore wing half as wide as hind wing; outer margin half as long as inner; M_3 and Cu_1 separated at margin by from a third to half an interspace (fig. 323)....36. Nomophila.
 - 9. Wings proportionate, fore wing nearly as wide (fig. 324) as hind wing.

10. Male with a bulla on fore wing and distorted venation. 32. Crocidophora.

- 10. Male and sometimes female with a hyaline discal spot; brown with falcate fore wing......44. Perispasta.
- 10. Male without sexual modification.

11. Fore wing falcate; both wings half orange.

45. Cindaphia.

11. Fore wing with marked or rounded apex.

18. HYMENIA Hübner

(Zinckenia Zeller)

Palpi upturned (fig. 326) with the three segments nearly equal in length, the first two well set off and broadly scaled, the third much slenderer, upturned beyond vertex, smooth-scaled and acute. Male antennæ somewhat thickened and obliquely notched at base. Maxillary palpi long and fusiform. Fore wing with \mathbf{R}_2 approximate to \mathbf{R}_{3+4} , \mathbf{R}_5 divergent from base; \mathbf{M}_2 and \mathbf{M}_3 somewhat approximate, \mathbf{Cu}_1 well separated. Hind wing with cell, measured on the costal side, hardly a third the length of the wing, extending out below as usual; \mathbf{M}_2 and \mathbf{M}_3 approximate.

Key to the species.

A continuous white fascia from within cell to inner margin.....2. fascialis. Median fascia starting below cell; the spot in cell separate, often obscure.

1. perspectalis.

1. H. perspectalis Hübner (Spotted beet worm). Brown, somewhat mottled with ochre yellow. Fore wing with a squarish spot in cell; a weak antemedial line across the wing; a postmedial fascia from costa at three-fourths way to apex, extending a third way across the wing, with two small dots below and beyond

 $^{^{36}}$ A few members of the first group with triangular palpi, may run out here, but may be distinguished from most of the following genera by $\rm R_5$ which is curved and approximate to $\rm R_{344}$ at base.

³⁷ Sameodes may run here, but differs by its close-scaled palpi with well-set-off third joint.

its lower end; a nearly straight, but waved, fascia from below the spot in the cell to the middle of the inner margin; hind wing with a median fascia, entirely very narrow, or irregular and widened to twice its width above and below cell. All the markings either translucent white or obscure yellowish, defined with dark brown. 22 mm. (animalis, exportalis, Guenée, primordialis Zeller).

brown. 22 mm. (animalis, exportalis, Guenée, primordialis Zeller). Late autumn. Larva on beet and chard; in greenhouses also on Alternanthera. Green with black tubercles. Several broods, breeding continuously in greenhouses. New York to Illinois and south; also world-wide in the warm-temperature and tropical regions. New York: Ithaca.

2. H. fascialis Cramer (Hawaiian beet webworm). Brown, not varied with yellow; antemedial line obsolete; a broad white patch running up from middle of inner margin to middle of cell, obliquely truncate above, and often sending out a tooth toward anal angle; a large, white bar from costa, at three-fourths, halfway to anal angle; with two white dots beyond its lower edge. Hind wing with a broad, nearly straight, white median fascia, slightly broadened at the middle. 20 mm. (recurvalis Fabricius). (H. 47:28.)

Larva on beet, chard, mangels, Amaranthus, and various weeds; sometimes injurious. The moth is taken in the North, late in the fall.

Subtropical, straying north to New York and western Pennsylvania.

19. DESMIA Westwood

Palpi with first two joints broadly scaled (fig. 327), the second rounded at end; third joint short, with a more or less triangular tuft in front (less marked than in Diastichtis) well set off, attached to upper side of second joint. Fore wing of male (fig. 320) with outer margin oblique and about as long as inner; hind wing relatively small, with lobed and sinuate costa and more or less extended anal angle; female more nearly normal; fore wing with marked apex; hind wing with waved outer margin. Fore wing with \mathbf{R}_3 and \mathbf{R}_4 very long-stalked. Hind wing with a notch at middle of shaft, preceded by a scale tuft. The species are black with white markings.

1. D. funeralis Hübner. Black, fringe white; fore wing with two large oval spots; hind wing with a broad oval transverse band in middle, not quite reaching costa or inner margin; in the female partly or completely divided into two spots. 20-28 mm. (H 47:37.)

Rather common. Moth from May to October, with two or three broods. Larva (grape leaf folder) sometimes injurious to grape; more rarely on Enothera and Cercis.

Generally distributed. New York: Fentons (Lewis County), Lewiston, Buffalo, Niagara Falls, Ithaca, Big Indian Valley, Onteora Mountain, Schenectady, Rhinebeck, New Windsor, Pearl River, Katonah.

20. DIASTICHTIS Hübner

(Botis, in part; Bocchoris Moore)

Front flat, oblique (fig. 334); palpi obliquely porrect, with a triangular tuft on second joint; third joint well set off, set on upper side of second, with a distinct terminal triangular tuft, rarely appressed against the tuft of second joint. Fore wing pointed, broad; \mathbf{R}_s moderately curved, divergent; hind wing ample; \mathbf{Cu}_1 divergent; fringe on \mathbf{Cu} more distinct than usual; cell nearly two-fifths of length of wing.

1. D. argyralis Hübner. Bright ochre vellow (leather-brown in the northern variety ventralis Grote), usually with an irregular series of dark brown ringed,

silver-white postmedial dots, the three middle ones much farther out than the others; rarely, with markings obsolete. Hind wing a little duller, usually immaculate. 25 mm.

July.

The two varieties overlap widely and intergrade. Westward the species seems also to intergrade with *D. fracturalis.*

Generally distributed, north to New Hampshire and Wyoming. New York: Albany, New Windsor, Poughkeepsie, Crugers, Pearl River, Clove Valley, (Staten Island) East New York (Long Island).

2. D. talis Grote. Crimson; median area translucent yellow, divided by a squarish crimson spot over the end of the cell, reaching the costa, and also touching the crimson base at the costa. A minute, hyaline antemedial spot. Hind wing with the crimson confined to the base; a small quadrate spot on end of cell, only a third way out, touching the basal crimson area, and a broad irregular border. 18 mm.

June.

Virginia; south to the tropics.

21. BLEPHAROMASTIX Lederer

(Nacoleia, in part; Lamprosema, in part)

Typically with male antennæ simple and ciliate, smooth in *B. rivulalis;* front somewhat rounded out; palpi trapezoidal, beak-like, the third segment lying along the upper surface of the second, or somewhat prominent, obliquely divided in two colors by a line running to the tip of the tuft; maxillary palpi slender, slightly truncate at tip when perfectly fresh, extending to the tip of the upper surface of the second joint of the labials. Fore wing fully twice as long as wide, with marked apex and anal angle, and even, slightly excurved onter margin. Venation normal (fig. 322), \mathbf{R}_5 divergent from $\mathbf{R}_{3:4}$. Hind wing with all veins divergent.

Ranalis is our only typical species. Magualis and rivulalis are transitional to Lygropia; stenialis is apparently a Nymphuline, near Nymphula, but \mathbf{R}_2 is constantly free and it is usually placed in Blepharomastix.

Key to the species

1. Evenly luteous to light brown	1. ranalis.
1: Ground yellow and white.	
2. Median area yellow	2. magualis.
2. Median area white, except toward costa	.3. rivulalis.
1. Ground yellow, heavily shaded with umber brown	.4. stenialis.

1. B. ranalis Guenée. Clav-color, occasionally light brown, especially in western specimens; markings darker dull brown; costa shaded with brown; antemedial line excurved, often followed by a dark dash (claviform) in fold; orbicular a well-marked ring; reniform a double bar or oblong ontline; postmedial line in male normally of two parts, the upper part running straight from the costa to the outer margin at Cu_2 , the inner from the discal dot to the anal angle; the two sectors sometimes connected by a line along Cu_2 . Female normally with a continuous line, strongly sinuate on Cu_2 . A broken brown terminal line. Hind wing similar, with a single dark discal bar. 20 mm.

Larva webbing leaves of Chenopodium.

New York to Missouri and south. New York: Buffalo, Ithaca, Poughkeepsie, New Windsor, New York City.

2. B. magualis [sic] Guenée. Yellow, base pale toward inner margin; sub-

terminal space contrasting, white; lines fine, nearly black; orbicular and reniform as before, heavy, and nearly filled with brown; antemedial line excurved from orbicular to inner margin, an oblique basal line before it; postmedial line as in *B. ranalis*, but heavier, the horizontal part slightly oblique above Cu, and touching the lower side of the reniform; subterminal line nearly parallel to postmedial, often touching the outer margin at Cu_2 , and the postmedial at its angle; twice as far from postmedial below the horizontal part as above. A continuous brown terminal line and a broken line in fringe; a brown bar across the terminal space, opposite the cell. Palpi less beaklike at the tip than in *B. ranalis*, the entire tip brown; third segment short and blunt; maxillary palpi rather shorter. This species and the next might go better in Lygropia. 15 mm. (magnalis auct.)

June.

Central Illinois and southward.

3. B. rivulalis Hampson. Fore wing with a fringe of rough hair on under side of costa, fore coxa and femur with expansible tufts, head as in *B. magualis*. White; more or less of base, costa, and whole outer margin pale yellow; markings brownblack. Hind wing similar, hase with four irregular and more or less interlocking lines; the first and fourth typically complete and excurved, fifth line running to orbicular; postmedial line much as in *B. magualis*, the offset running along Cu_2 and not touching the reniform, the space beyond it not white; subterminal line well separated, except the horizontal portion, which is fused with the postmedial on Cu_2 . Reniform a double brown bar; a brown bar beyond it in the postmedial space below which there is an oval white patch above Cu_2 (this region is yellow in *B. magualis*). Terminal line strong. 20 mm.

Rare; in June and in late August.

Quebec and Illinois, south to Pennsylvania and North Carolina.

4. B. stenialis Guenée (figs. 322, 332). Light yellow, shaded heavily with brown; the yellow ground most constant before the postmedial line, and the brown filling the terminal area, except for a narrow line just beyond the postmedial line and the costa toward the apex. Lines as in *B. ranalis*, thicker, but broken; orbicular and reniform thick, obscured by the brown shading; basal line represented by a brown shade. Hind wing similar. Palpi obliquely upturned with segments well set off; the third segment a third as long as the second and broadly scaled; maxillary palpi truncate, subtriangular, and as long as second segment of labials. 18 mm.

Not rare; in swamps in June and July. Larva unknown, possibly aquatic. Closely related to *Geshna primordialis*, and very near the point of separation of the Nymphulinæ and Pyraustinæ.

Generally distributed. New York: Gowanda (Wild), New Windsor.

22. CONCHYLODES Guenée

(Spilomela Guenée)

Labial palpi thick and smoothly upcurved; the first two joints rather marked, not triangular, and third joint short and triangular. Maxillary palpi minute; antennæ prismatic, front flat, oblique, prominent. Fore wing with \mathbf{R}_s slightly approximate to \mathbf{R}_{3+4} ; hind wing with \mathbf{M}_1 shortly stalked, \mathbf{M}_2 and \mathbf{M}_3 approximate.

The southern species C. diphteralis eats pecan.

1. C. ovulalis Guence. White, abdomen with fifth to seventh segments yellow, second and fourth with black bands. Fore wing with straight basal and antemedial lines; postmedial line excurved from costa at four-fifths to inner margin at four-fifths, fine except on costa; orbicular a black patch; reniform a heavy black ellipse with a white line in the center; subterminal excurved, parallel to outer margin, much heavier than postmedial line. Terminal line rather strong, black.

A black line from inner margin at one-third up to cell beyond orbicular, then along lower side of cell to reniform, then much finer and running to inner margin at lower end of postmedial line (of which it is morphologically a part); more rarely, with its lower end joined to the lower end of the postmedial line and not going below **A**. Hind wing with black discal bar, postmedial, subterminal, and terminal lines, and loop, much as in fore wing. 25 mm. (*platinalis* auct., not Guenée). (H 47:60.)

June and July; September. Larva probably on sycamore. In C. platinalis. which was described from Missouri but has since only been taken in South America, the subterminal line is irregularly thickened, and the postmedial line joins the looped line above the fold.

North Carolina to central Illinois, Missouri, and south.

23. PANTOGRAPTA Lederer

(Sylepta, in part)

Similar to Conchylodes; front less prominent below (fig. 329); \mathbf{R}_5 more strongly curved and approximate to \mathbf{R}_{a+4} . Hind tibia of male of our species clothed with long hair on outer side.

P. limata Grote and Robinson. White, shaded with pale yellow; marked with dull brown. Basal line excurved, much nearer to base than to antemedial; antemedial line excurved and slightly wavy; orbicular higher than wide; a brown outline, constricted in the middle; claviform a brown ring, separate from ante-medial line, with a line running from it to inner margin; reniform a strongly irregular brown outline; postmedial line heavy, distinct on costa, obscured below. but when traceable, wavy and parallel to outer margin; subterminal wavy and parallel to outer margin, even at costa. Terminal line even, continuous. Outer third from \mathbf{M}_1 to inner margin, and inner margin beyond middle suffused with dull brown, obscuring the marks. Hind wing somewhat suffused with brown; with a pale spot beyond the double discal bar, and a strong deeply serrate double postmedial line. 37 mm. (H 47:38.)

Moth in June to August. Larva a leaf roller on linden. General in distribution. New York: Vicinity of Buffalo, Ithaca, Big Indian Valley, Ilion, Poughkeepsie, New Windsor, Long Island.

24. SYLEPTA Hübner

Similar to Pantograpta; male hind tibiæ normal; maxillary palpi and third segment of labials rather smaller.

1. S. penumbralis Grote. Light fuscous, shaded with slightly darker fuscous toward margin and slightly paler beyond the postmedial line. Palpi wholly concolorous. Lines diffuse, a little darker; antemedial strongly oblique outward, obsolete on costa; discal mark a vertical bar; postmedial line subdentate, excurved a little below cell, running far in on Cu_2 , but rarely as far as opposite the discal bar. Hind wing similar; discal bar less than a third way to apex, postmedial line much as in fore wing. 30 mm. (silicalis Guence.)

May; September and October.

Ohio to Illinois, Missouri, and Florida. 2. S. obscuralis Lederer. Identical in markings with S. penumbralis, but ochreous, veined and shaded with fuscous, the shading especially beyond the reniform along the outer margin; rarely nearly even fuscous. Orbicular a small dark circle, reniform a lunule, both pale filled; dark terminal bars more distinct.

Larva on Phytolacca decandra.

St. Louis, Missouri; Florida; an orange race in Arizona, with intergrades in Texas.

25. PILOCROCIS Lederer

(Zinckenia, in part)

Similar to Pantograpta and Sylepta. Third joint of palpus rather smaller, in some species with a tfiangular tuft, but smooth in the typical group (fig. 333). Antennæ of male in our species (typical Pilocrocis), with a notch at the base of the shaft, and thickened and roughly scaled beyond. Fore wing with a thick fringe on basal half of costa above.

The known larvæ eat Convolvulaceæ.

1. P. ramentalis Lederer. Fuscous brown; lines as in Sylepta penumbralis, cleancut, whitish, slightly defined on the side toward the median area, with darker brown. 25-28 mm. (perfuscalis Hulst.)

Northward this species has only been taken in the fall.

New York to Central States and south. New York: Ithaca.

26. LYGROPIA Lederer

Palpi similar to those of Pantograpta, upturned, somewhat broadly scaled, with marked first and second segments; third segment small, scaled, and attached to upper side of second. Fore wing with \mathbf{R}_{5} divergent from \mathbf{R}_{3+4} . Moths slighter than in the preceding genera; normally yellowish with contrasting markings; similar on both wings. No secondary sexual characters in our species.

A large tropical genus to which octonalis doubtfully belongs; nymphulalis and magualis, treated as Blepharomastix, might be better placed here.

1. L. octonalis Zeller. Pale lemon yellow; fore wing with a rounded light red patch at base of costa; antemedial patches on costa and inner margin, median ones on costa and cell, and sometimes a terminal bar on costal half of wing, all finely edged with black. Traces of black transverse lines. 15 mm. (Eustixia, Orobena; sexmaculalis Grote.)

Southern States; doubtful in our area.

27. EUDIOPTIS Hübner

(Margaronia Hübner; Phakellura Poey; Glyphodes Guenée; Margarodes Guenée, not Guilding)

Male antennæ normal in our species; palpi oblique, the second joint in hyalinata and mitidalis (fig. 331) rounded, practically as large as the eye; third joint minute but distinctly set off. Second joint of palpus in quadristigmalis (fig. 330) rather smaller, trapezoidal, with triangular terminal tuft, the third nearly concealed in the vestiture of its upper side and not reaching the point; the vestiture at the tip of the palpus wholly borne by the second joint. Palpus obliquely divided into brown and white, the line of division running almost to the tip of the tuft. Abdomen of male, in our species, with large fanlike terminal tuft. Maxillary palpi large, triangular, almost as in *Blepharomastix stenialis*. Wings (fig. 321) very finely scaled, translucent; \mathbf{R}_5 curved and approximate to \mathbf{R}_{644} . Margins even.

Key to the species

1. E. nitidalis Cramer. Fore wing brown, with a translucent irregular yellow patch extending from beyond and below end of cell to inner margin. Hind wing

translucent yellow, with marginal third brown. Anal tuft yellow and brown. 30 mm. (H_47:43.)

August. Larva in stems and fruit of the melon family.

Massachusetts to Missouri and south to the tropics. New York: Albany, 2. E. hyalinata Linnæus. White. Costa with an even black-brown band reach-ing inner margin at base, and covering cell; outer margin with an equally broad band; hind wing with blackish band evenly tapering to anal angle, the fringe whitish. Tuft of male black and yellow (in aberration niveocilia Hampson wholly black). 28 mm. (H 47:39.)

Larva with E. nitidalis; sometimes injurious.

Quebec to Colorado and south. New York: Buffalo, Brooklyn.

3. E. quadristigmalis Guenée. Almost transparent; white, costa brown above R; two black discal dots, and one representing orbicular, rarely absent. Tuft white. 25 mm. (H p. 394 f. 217.)

Not rare.

Quebec to Colorado and south. New York: Ithaca, New Windsor.

28. CONDYLORRHIZA Lederer

Similar to Eudioptis; palpi beaklike, the third joint rather long and lying along the upper face of the second, reaching its tip.

Perhaps E. quadristignalis would be better transferred to this genus.

1. C. vestigialis Guenée. Typically very pale brown; in variety tritealis Walker, translucent light lemon yellow, with lines brown; antemedial strongly oblique out toward inner margin; orbicular a dot, reniform a short bar. Postmedial line strongly excurved at middle, sinuous, bent at a right angle below cell, and then ending perpendicular to inner margin. Terminal line and fringe brown. Hind wing concolorous, with discal bar, an irregular postmedial line, and partly fused subterminal and terminal lines, most distinctly separate toward costa. Anal tuft fanlike, black. 30 mm.

Larva on willow.

Subtropical, straying in late fall north to New York and western Pennsylvania. New York: Ithaca.

29. METREA Grote

Palpi upturned and not very broadly scaled, with porrect third joint, smoother than usual in Pyrausta; maxillary palpi filiform; venation normal, \mathbf{R}_{s} divergent; wings more rounded than usual.

1. M. ostreonalis Grote. White, a little translucent, especially the hind wing. Fore wing with a diffuse dark gray patch at end of cell (rarely absent) enclosing a diffuse blackish discal dot. Two large, partly confluent, obliquely placed gray patches in fold and below A, below end of cell; a gray subterminal shade below M₁, resting on inner margin. 30 mm.

July. Rare.

Quebec to Connecticut and western Pennsylvania. New York: Peru (Everett).

30. HELLULA Guenée

(Cabbage web-worms)

Palpi obliquely upturned (fig. 336), the first two joints marked, the third fairly long, closely scaled, oblique, well set off; maxillary palpi filiform. Venation nor-mal, the veins well spaced; the scales on the discal bar very slightly raised, and often slightly metallic.

The species are not well understood.

1. H. undalis Fabricius. Pale luteous, usually heavily shaded with fuscous; lines white, wavy, defined with blackish; basal line fragmentary; antemedial com-

plete, transverse; postmedial complete, strongly sinuous, and distinct, sharply bent in on R, and on Cu, and A; a series of black terminal dots, immediately preceded by an even whitish line, defined on inner side with darker. Discal lunule black, with gravish white center kidney-shaped, concave on outer side, with more or less distinct blackish shades before and beyond it. Hind wing cream-white, with a broad, even but diffuse, darker border, and dark gray terminal line. 15 mm. (rogatalis Hulst.)

Apparently introduced from the Old World. Larva most often on Cruciferæ and sometimes injurious in warm countries.

Southern States, probably not quite reaching our area. 2. H. phidilealis Walker. Similar to H. undalis; clay-color; hardly, if at all, shaded with blackish; the lines not quite so irregular and defined with slightly darker elay-color; the region about the reniform concolorous; reniform roundedtrapezoidal, oblique; rather smaller than in *H. undalis*, not concave on outer side and strongly iridescent with violet. Hind wing whiter, its border less distinct.

This form appears to be the native representative of H. undalis, and possibly interbreeds with it. Intermediates appear to be rare.

Southern States and southward; doubtful in our area.

31. EVERGESTIS Hübner

(Pionea, Mesographe)

Similar to Pyrausta; maxillary palpi extending fully to end of second segment of labials, rather truncate at tip (fig. 335). Lahial palpi rather more closely scaled, the third joint porrect, moderate, pointed, its base hardly concealed in the vestiture of the second joint.

1. E. straminalis Hübner (Purple-backed cabbage worm). Straw vellow, somewhat dusted with brown; basal line obscure, angulate at middle when most distinct; antemedial line from costa at two-fifths, perpendicular to cell, deeply concave across cell, forming a sharp tooth on Cu, the tooth touching the lower end of the reniform, then strongly oblique to its inner margin; postmedial line excurved above; below, parallel to outer margin and slightly waved; reniform a brown outline, deeply constricted at middle, normally 8-shaped. Subterminal region shaded with brown, outlining a triangular yellow patch on upper part of outer margin; terminal line brown. Hind wing translucent whitish, yellower toward margin, with traces of a fine brown postmedial line, and a narrow brown terminal band. 20-25 mm. (H 49:45.)

May to September; two broods, flying mainly in June and August. Quite variable in the amount of brown shading. Larva dark green or violet, shaded with yellow on sides and paler below; with black head and tubercles; on Cruciferæ in September: rarely injurious.

Very common northward; also in Europe. New York. Common everywhere.

2. E. rimosalis Guenée (Cross-striped cabbage worm). Fore wing rather longer than E. straminalis; hind wing with marked apex, less falcate than in Diasemia, Fore wing light vellow, shaded with brown along the costa, over end of cell, and more heavily on outer third; the upper boundary of the latter shade forming an oblique streak running to the apex; antemedial and postmedial lines brown, weak, subparallel to outer margin, wavy, nearly trisecting the wing; with faint lines between them; reniform spot dark, obscure. 27 mm.

Eggs in a cluster, overlapping. Larva bluish gray with three or four transverse black stripes on each segment, and a bright yellow stigmatal band. Probably three broods; pupa in the ground; the third brood wintering and emerging in April.

West Virginia to Illinois, Colorado, Washington, and south, New York: Long Island.

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32. CROCIDOPHORA Lederer

Similar to Evergestis and Pyrausta; the palpi typically with the hair on the second joint longer, covering base of third (fig. 341), but short in *C. tuberculalis.* Fore wing with a bulla at end of cell, between \mathbf{R}_4 and \mathbf{R}_5 , \mathbf{R}_5 curved down to make room for it in males; sometimes with highly developed sexual modifications. Antenuæ normal in our species.

The style of markings associates this genus with Pyrausta rather than with Evergestis; but it is customarily put here.

Key to the species

1. C. serratissimalis Zeller. Male retinaculum formed into a large blackish scale tuft, covering a fovea; the base of Cu curved up around it; no distinct fovea in cell Cu₁, but with wing rather thinly scaled. Fore wing pale straw yellow with some light brown on veins; antemedial line somewhat waved and oblique, obscure in male, postmedial line sinuous, rather broad and even, deeply bowed in opposite cell and across Cu_2 and A; with more or less distinct brown shades before it; reniform a dark bar; subterminal deeply denticulate, nearly parallel to outer margin; terminal line fine, continuous, brown. Hind wing with postmedial and subterminal lines as in fore wing, but the postmedial less sinuous. 18-25 mm.

Superficially this species is very near *P. penitalis*, but is distinguished by the servate subterminal line on the hind wing.

June to September.

New Jersey. New York: Newport, vicinity of Buffalo, Otto, Potter Swamp (Yates County), Ithaca, Schenectady, Poughkeepsie, New Windsor, New York City.

2. C. tuberculalis Lederer. Male with foveæ much larger, tuft representing retinaculum much larger, covering a fovea; a large fovea, in cell Cu at base, distorting Cu_2 and lower edge of cell; which runs almost transverse from Cu_2 to Cu_1 . Light straw vellow, no dark on veins; translucent. Antemedial line about as in *C. serratissimalis*, less wavy; postmedial sinuous below cell, but evenly convex opposite cell; subterminal shade much broader, even, sometimes suffused out to outer margin; terminal line obscure; discal dot faint, vertical. Hind wing with even excurved postmedial line, and broad subterminal shade. 15 mm.

June and July.

New York to central Illinois and south. New York: vicinity of Buffalo, Lancaster, Ithaca, New Windsor, Long Island.

C. pustuliferalis Lederer has narrower wings, a dentate postmedial line, and sometimes an inconspicuous dentate subterminal line. It occurs in North Carolina and southward in May. 25 mm.

33. SAMEA Guenée

Palpi moderate (fig. 328), upturned, the first two joints well marked off and broadly scaled, the third small, close scaled, and lying on the upper surface of the end of the second; maxillary palpi small, porrect. Fore wing subfalcate, somewhat translucent; \mathbf{R}_{5} curved and somewhat approximate to \mathbf{R}_{3+4} . Hind wing subfalcate; a prominent tuft of scales at base of **Cu** above, representing the usual fringe.

S. ecclesialis Guenée, a widespread tropical species, may possibly occur in our area. The abdomen of the male has long curved tufts on the sides of the

fourth segment. It certainly occurs north to the coast of North Carolina. (H. 48:2.)

34. SAMEODES Snellen

(*Pyrausta*, in part)

Palpi nearly as in Pyrausta with very little loose hair, the third joint practically . terminal. Fore wing with \mathbf{R}_{5} curved and more or less approximate to \mathbf{R}_{3+4} , apex marked. Markings as in Samea, but without the tuft on base of Cu of hind wing.

1. S. adipaloides Grote. Abdomen brown, in the female with some yellow scales on the base and white ones at the apex of the segments, the first segment often yellow. Male without a dorsal tuft. Straw yellow. Costa gray-brown above ${f R}$; outer margin broadly gray-brown, with sharp inner boundary two-thirds way in to the postmedial line and roughly parallel to it; orbicular, claviform, and reniform spots represented by large gray-brown patches, the first two broadly in contact, antemedial line fine, gray-brown, excurved, running shortly before the orbicular and claviform; postmedial deeply sinuate below cell, extending in along Cu_2 to below end of cell; scalloped on veins; the veins more or less defined with brown; hind wing similar, without brown patches in disc; postmedial line confluent with border at Cu_2 . 20 mm.

July to September.

Generally distributed. New York: vicinity of Buffalo, McLean Bogs, Big Indian Valley, Schenectady, Rhinebeck.

I have seen S. elealis Walker from North Carolina and south (adipaloides Barnes and McDunnough, but not of Grote's original figure and description). The costa is mostly yellow; the male has a black dorsal tuft on the abdomen, which is more largely yellow.

35. STENOPHYES Lederer

(*Crocidophora*, in part)

Palpi trapezoidal, closely and a little roughly scaled, with third segment terrapit trapezoidat, closely and a fittle foughly scaled, with third segment ter-minal; maxillary palpi moderate; front oblique. Fore wing with \mathbf{R}_{s} approxi-mate to $\mathbf{R}_{s_{14}}$; costa practically straight; fore wing more than twice as long as wide, much more in male, with apex marked; outer margin strongly oblique and nearly as long as inner; hind wing much longer to apex than to anal angle; abdomen long and conical. Wing translucent. Markings as in Sameodes. 1. S. huronalis Guenée. Yellowish white; lines fine, light brown; orbicular, elaviform and roniform large squarish orange-colume patches adapt with brown.

a yellow basal shade; antemedial line erect, wavy, nearly meeting lower angle of claviform; postmedial oblique, but less so than outer margin, offset inward on Cu_2 , and then oblique to inner margin. Outer margin light brown, sending an extension to the postmedial line opposite the cell and at the anal angle. Hind wing with two broken gray transverse lines, the outer meeting margin near Cu_z ; outer margin broadly gray at apex, tapering off to anal angle. Male 25 mm; female 20 mm. (*serinalis* Walker). September and October. Described from Canada but only seen from North Caroling and anothered

Described from Canada but only seen from North Carolina and southward.

36. NOMOPHILA Hübner

Palpi as in Pyrausta, rather closely scaled (fig. 338); fore wing over $2\frac{1}{2}$ times as long as wide (fig. 323); the outer margin less than half as long as the inner; hind wing ample, nearly twice as wide. 1. N. noctuella Schiffermüller. Fore wing dull gray, orbicular, reniform, and

claviform large, darker, but rarely contrasting, outlined with black; orbicular and claviform horizontally oval and often slightly separated; postmedial finely dentate when distinct; subterminal space finely strigose, with two streaks to an interspace. Illind wing contrasting, pale luteous at base. 28 mm.

June to October.

The narrow wings and dull coloring make this species look like a Phycid. It is one of the commonest Pyralids. The larva is a general feeder on low plants, and has been reported definitely from Polygonum. It is yellowish green, with darker tubercles.

Distribution world-wide. New York: common everywhere.

37. POLYGRAMMODES Guenée

Wings ample, the fore wing less than twice as long as wide, with strongly curved outer margin, stumpy looking. Wings transheent: \mathbf{R}_s strongly curved up and approximate to \mathbf{R}_{s+4} ; hind wing with a black scale tuft at base of \mathbf{Cu} , as in Samea. Body stont, showing a strong tendency to grease; palpi as in Stenophyes.

1. P. capitalis Grote. White with some violet iridescence; with white fringe. Fore wing overlaid with brown wool at base and inner margin. A fine, incomplete, black, antemedial line, a rather irregular postmedial; basal line sometimes visible; orbicular a small ring; reniform a double black bar with a blackish patch below it; a blackish patch at apex. Hind wing similar. 37 mm.

June.

Mississippi Valley.

Variety **posticata** Grote and Robinson is practically immaculate, with yellowish fore wing and white hind wing. The typical *P. hirtalis* Guenée, from Texas and south, is much smaller. The names have sometimes been interchanged in collections, but I think the present use is correct.

38. PACHYZANCLA Meyrick

Structure much as in Nomophila and neighboring genera. Palpi rather smoothscaled, somewhat beaklike, with third segment very small, on upper side of second. Fore wing triangular, twice as long as wide; M_2 concave up at base, and closely approximate to M_3 . (I very much doubt 14 this genus is either natural or in the right place here.)

1. P. perfusalis Walker. Fore tibia with a terminal tuft. Inner margin of hind wing distorted; anal angle lobed in male, with a short, stiff tuft, and an area of black raised scales above Cu_2 , which is distant from Cu_1 at margin. Dark fuscous, antemedial line darker, excurved; postmedial running in a broad sweep from costa, far out toward outer margin, then in below cell to opposite reniform, then nearly erect to inner margin. A blackish discal bar. Hind wing duller, with a slight postmedial line only. 22 mm.

Larva on eggplant.

Southern States; not seen from our area.

2. P. bipunctalis Fabricius. No secondary sexual characters. Ground color dull luteous, darkening on costa and onter margin, and on apical half of outer margin of hind wing; with a short thick solid black reniform spot hardly higher than wide, and smaller round orbicular dot. Subterminal line absent, and post-medial indented opposite the cell. 25 mm.

Larva (southern beet webworm) on beet and a great variety of other herbaceous plants; injurious in the Gulf Strip.

Decatur, Illinois, and south. The eastern records, so far as traced, are in error for species of Phlyctænia and Pyrausta, especially *Pyrausta nertextalia*.

I believe this species would go better in Sylepta.

39. LOXOSTEGE Hübner

(*Phlyctanodes* Guenée, *Eurycreon* Lederer)

Similar to Pyrausta, but with the lower part of the front drawn out into a sharp cone, or, in some western species, a more complex process (fig. 339). Hind wings usually more simply marked than fore wings. Eyes normal in our species, reduced in some western ones.

Key to the species

1. Expanse 10 mm10. ophionalis.
1. Expanse 15 mm. or more.
2. Fore wing falcatel. externalis.
2. Fore wing with apex at most marked.
3. Fore wing mostly white, with yellow costa
3. Fore wing yellow, nearly or quite immaculate.
4. Hind wing concolorous8. marculenta.
4. Hind wing white
3. Fore wing contrastingly marked, or fuscous.
4. Fuscous with a contrasting light yellow terminal band and square
between orbicular and reniform.
5. Postmedial line denticulate and fine in its upper course.
4. sticticalis.
5. Postmedial with its outer side even, often represented by a series of
black wedges
4. Less mottled, terminal space and space between orbicular and reniform
not contrastingly pale.
5. Cream-white, with contrasting gray or olivaceous veining, and dentate postmedial and subterminal lines, checkering the outer part of the
wing
5. Veins concolorous; postmedial line evenly sinuous; ground not cream-
white with contrasting dark markings.
6. Lemon yellow, with broad contrasting rusty marks, including a
terminal line on hind wing but no subterminal2. helralis.
6. Straw yellow, with light brown to obsolescent markings, and sepa-
rate subterminal and terminal lines on hind wing, or with the
subterminal line only.
7. Postmedial line strongly denticulate; hind wing with postmedial
line distinctly euryed above Cu
7. Postmedial very slightly denticulate or even, on hind wing
straight to below \mathbf{Cu}_2
6. Fuscous to ochreous, hind wing with, at most, slight diffuse mark-
ings, usually immaculate in pale forms
I i the liter with M and M closely approximate Evental
Fore wing falcate; hind wing with M ₂ and M ₃ closely approximate. Frontal prominence broad at the end, somewhat chisel-shaped.
I T anternalia Warman Frank with white side lines. Light dull brown: lines

1. L. externalis Warren. Front with white side-lines. Light dull brown; lines fine, single, dark; antemedial irregular, excurved; postmedial sinuous, denticulate, not bent in opposite cell; orbicular a dot and reniform a curved vertical line, all faintly defined with paler; no subterminal line. Hind wing slightly paler, with obsolescent postmedial only. 22-25 mm. (macluræ Riley).

Larva on Osage orange.

Plummer's Island, Maryland; western Pennsylvania, Illinois, and St. Louis, Missouri, south to North Carolina and Texas.

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II. Fore wing not falcate, front definitely conical, usually with a sharp point; hind wing \mathbf{M}_2 and \mathbf{M}_3 closely approximate.

* Hind tibiæ with outer spurs about half the length of the inner.

2. L. helvalis Walker. Bright yellow; lines clean-cut, yellow-brown, single and not denticulate; costa yellow-brown; antemedial excurved; postmedial broadly excurved above, with a blunt inward tooth on fold; orbicular a dot, reniform a curved line; no subterminal. Hind wing paler, with a short straight postmedial line pointing toward anal angle; both wings with even terminal lines. 15-20 mm. (citrina Grote and Robinson.)

September.

Coast of Massachusetts to Florida and Texas. New York: Long Island.

3. L. similalis Guenée. Typically dull ochre, somewhat shaded with fuscous; lines diffuse, dark, shading imperceptibly into the median area, usually more sharply defined on the side away from the median area, and defined with pale. Terminal line waved, postmedial somewhat sinuous and denticulate, when distinct. Orbicular a blurred spot; reniform an oval spot or lunule, both dark and diffuse. Hind wing somewhat paler, dark toward margin, translucent at base; in fully marked specimens, with broad, vague, dark border, and sometimes a faint dark postmedial line. 20 mm. (*licealis* Walker, *communis* Grote, *caffreii* Flint and Malloch.

Variety intractella Walker is the dark, fuscous form; *rantalis* Guenée is probably the same form; crinitalis Walker is brighter rusty orange, and **posticata** Grote and Robinson (*miserulata* Grote and Robinson) is nearly immaculate yellow with an immaculate white hind wing. Intergrades are common.

yellow with an immaculate white hind wing. Intergrades are common. Larva (the garden webworm) a general feeder, rarely injurious. Dull green above and yellowish below, with a double pale dorsal, and whitish lateral, line. Tubercles black. Four or more broods.

Generally distributed. New York: Saranac Inn, Ithaca, Albany, Nassau; Woodhaven, Long Island.

4. L. sticticalis Linnæus. (Sugar-beet webworm.) Superficially very close to L. commixtalis; upper outer spur of male hind tibiæ nearly half as long as lower one. A little smaller than L. commixtalis and more lightly built. Fore wing with basal dash and streaks on veins nearly obsolete, light fuscous; orbicular thicker and more obscure; postmedial line distinctly and finely denticulate; yellow terminal shade widened opposite the cell, followed by a single dark terminal line. Hind wing nearly concolorous, with a vague pale postmedial shade; yellow marginal shade evener; terminal line single. 25 mm.

medial shade; yellow marginal shade evener; terminal line single. 25 mm. Larva gray-green, yellower on sides, with black head and pale, dark-ringed tubercles; a general feeder and sometimes injurious in gardens.

Nantucket, Massachusetts; Madison, Wisconsin; Western States; Europe; probably introduced in the East. New York: Ithaca, Big Indian Valley, East New York.

** Upper outer spur of male hind tibia reduced, less than a quarter the length of the inner spur.

5. L. commixtalis Walker. Middle tibiæ with a groove and pencil. Luteous, heavily streaked with fuscous, especially on veins; terminal space vellow, not broadened opposite cell; a long black basal dash with a pale shade above it; antemedial line far out, deeply dentate, the longest tooth in the fold; orbicular a horizontal ellipse when distinct, often confused with antemedial line; followed by a yellow patch; this patch sometimes divided by a black dash connecting the orbicular and reniform spots; postmedial line represented by a series of black wedges, conspicuous beyond the cell, where their outer sides form an even line, and their points extend in along the veins to the

reniform, which is confused; black postmedial line deeply dentate below and finely so toward costa. A fuscous subterminal shade; terminal line double, the outer line in the fringe. Hind wing paler fuscous, sometimes with blackish veins on disc, sometimes with a vague vellowish postmedial shade, and nar-rower irregular yellow terminal band; blackish terminal line double. (A specimen from Newfoundland has all the yellow shades on the fore wing, except

the terminal band, replaced by light blue-gray.) 25-28 mm. (crearalis Zeller.) Usually rare; sporadically injurious; usually confused with L. sticticalis, but easily distinguished in the male, by the hind tibiæ. Larva about like that of L. sticticalis.

General in distribution. "New York" (Grote). 6. L. chortalis Grote. Pale yellowish, marked and shaded with light gray, appearing greenish on account of the yellow underlay. Veins contrasting gray; postmedial line serrate; subterminal gray, normally serrate; basal markings confused, largely lost in gray shading; hind wing whitish, with wavy subterminal and variable, often obscure, postmedial lines; terminal line dark gray. 25 mm.

Not rare from May to August. General, north to Northern Ontario and west to California. New York: Black Brook, Uphill Brook (Mt. Marcy), Clayton, Buffalo, Ithaca, Big Indian Valley, Onteora Mountain, Karner, Albany, Rhinebeck, New Windsor.

7. L. dasconalis Walker. White, with traces of the markings of L. chortalis and a faint light gray terminal line; front of thorax and base of costa bright vellow.

Apparently rare. June to August. Larva on Baptisia. Massachusetts to Illinois and south. New York: New Windsor. 8. L. marculenta Grote. Upper outer spur minute. Pale lemon yellow; head deeper ochre; markings pale brownish gray, very slightly diffuse; antemedial line excurved and somewhat wavy; postmedial sinuous and moderately dentate, most deeply on Cu2; subterminal a vague shade; terminal fine; orbicular and reniform small and diffuse. Hind wing similar toward margin, nearly hyaline white toward base; postmedial straight or curved; subterminal close to margin, sometimes fused with terminal line or absent in pale specimens. Markings often nearly obsolete. 20 mm. (probably obliteralis Walker.)

May to August. Larva on Solidago.

Massachusetts to Texas. New York: General.

9. L. mancalis Lederer. Upper outer spur one-quarter the length of the inner. Similar to L, marculenta and usually confused with it. Rather duller straw yellow; rarely, with brown veins as in L. chortalis; postmedial line almost evenly sinuate on fore wing; on hind wing straight from below costa to fold, and stopping there. Subterminal distinct, close to margin. Front brown with whitish shades; palpi light brown.

June; August. Caterpillar on Convolvulus, Rumex, and mint.

Maryland to Missouri and south ("Maine" in error).

III. Very small species; costa and outer margin nearly straight, meeting at a sharp angle; front as in L. externalis; palpi twice as long as head; maxillary palpi large, triangular, truncate at tip, hind wing with M. and M₃ widely separate, parallel at base.

Light yellow, suffused with tawny and brown; 10. L. ophionalis Walker. with a single straight red-brown and yellow subterminal band. 10 mm. North Carolina and south.

A closely related form, with crimson ground-color, occurs on the sand barrens of Illinois. It may be a race of ophionalis.

40. DIASEMIA Guenée

Palpi about as in Loxostege; rather short. Maxillary palpi large, trian-gular; male antenna broad and typically subservate in male. Fore wing typically with falcate apex; venation normal, with R₂ divergent. Hind wing with apex subfalcate, outer margin excavate below it; \mathbf{M}_{a} and \mathbf{Cu}_{i} approximate; cell two-fifths length of wing; front rounded.

Only the type species is normal in structure, *janassialis* is probably not really congeneric.

1. D. ramburialis Duponchel. White, striate with yellow and dusted and mottled with brown-black; the white showing especially as a very irregular postmedial line, widened on costa and forming a contrasting patch on inner margin. Fringe cut with white. Hind wing with white mostly in broad antemedial and medial bands, with a solid black band between them. 15 mm.

Europe; Atlantic States; seen only from Florida.

2. D. janassialis Walker. Male antennæ strongly annulate, with rows of raised scales; hind wing excavate opposite cell, but not falcate; venation as in Pyrausta; palpi long, beaklike. Black, with heavy white postmedial line, discal spot, partial antemedial line and other usual markings; not at all powdery or striate.

Southern States, north to North Carolina; not known in our territory. 3. D. magdalena Fernald occurs north to North Carolina. It is yellow, with fine silvery ante- and postmedial lines converging to the inner margin, and with a similar terminal line and confused discal marks.

41. TITANIO Hübner

(Botis, in part)

Similar to Pyrausta; front more prominent, rounded out and rough-bairy (fig. 337); gena a third as wide as eye, pale and conspicuous in *pollinalis*; eye narrower than front, with a chitinized band behind it; vestiture rough and more hairy than usual; with tufts on the tips of the large maxillary palpi.

This is a subarctic development of Pyrausta, and apparently intergrades with it. Two or three species placed here in Titanio have been generally considered Pyraustas, but have the characteristic small diurnal eves of this genus.

Key to the species

1. Black with two white spots on fore wingl. pollinalis. 1. Gray.

2. Postmedial line straight, whitish4. marginalis.
2. Postmedial line dark, denticulate and sinuate
2. Markings obsolete
1. Ground of hind wing yellow (insequalis group of Pyrausta).

1. T. pollinalis Schiffermüller. Black; two white spots on fore wing and a broken median fascia on hind wing. 20 mm.

May and June; August. Larva on broom and other Leguminosæ.

Europe; reported from Pennsylvania.

2. T. ephippialis Zetterstedt. Gena black, naked. Dark ash gray, somewhat powdery, especially the darker base; antemedial line wavy, oblique outward, dark; postmedial well out; excurved above, extending far in below cell, enlarging into dots between veins; median area sometimes paler; ordinary spots small, solid, dark, Hind wing dark gray with more or less distinct dark and pale postmedial line and pale costa. 20 mm.

July.

Holaretie, south to Labrador.

3. T. torvalis Mceschler. Dark fuscous, with some white scales, markings almost obsolete, so far visible like T. ephippialis. Hind wing much paler. 20 mm. Greenland to Labrador.

4. T. marginalis Walker. Powdery mouse gray; blackish toward base and beyond postmedial line; under the lens showing blackish, white, and orange-brown scales. Antemedial line obscure. outwardly oblique, waved, postmedial line nearly straight. white, contrastingly defined with blackish; orbicular a black spot, and reniform a small black patch; median area slightly paler, graver, and more powdery. Hind wing with black-brown powdering on a luteous base; with a short straight postmedial line. 18 mm. (stenopteralis Grote.)

May.

Maine to Alberta.

42. THOLERIA Hübner

(Mecyna Guenée)

Structure as in Phlyctænia, the third segment of the palpus not well set off, somewhat down-curved (fig. 340); midtibia with a groove and tuft of hair in male. Hind tibia normal; \mathbf{M}_2 and \mathbf{M}_3 more closely approximate than normal in Phlyctænia and Pyrausta.

1. T. reversalis Guenée. Fore wing brown with faint markings; hind wing bright yellow, with blackish apex. 30 mm.

June and July. Larva on Baptisia and Louicera.

New York and Illinois to Florida and Colorado. New York: Ithaca, Oneonta. New Windsor, Staten Island; Glencove, Long Island (in a conservatory).

43. AUTOCOSMIA Warren

(*Titanio*; *Metasia*, in part)

Front typically conically prominent, in our species (fig. 342), which belongs doubtfully to the genus, oblique and rounded out. Our species with a bulla between \mathbf{R}_{i} and \mathbf{R}_{j} of the male, as in Crocidophora; palpi moderate, beaklike, the third segment not well set off; hairy, normal; eyes wider than front; maxillary palpi larger, triangular; no veins approximate, not even M_2 and M_3 ; wings rounded, with costa more arched than usual; normal.

I. A. helianthales Murtfeldt. Ash gray, shaded with whitish, especially over the end of the cell, and costa opposite; postmedial line fine, white, convex above, more oblique than outer margin below, even, nearly touching terminal line opposite cell; a more or less irregular streak on A out to postmedial line, forked near its tip; a white line in base of fringe, preceded by a dark terminal line. Hind wing pale gray, with a white postmedial line far out, but receding toward anal angle. Upper tibial spurs more than three-fourths way to apex of tibia, very small. 12 mm.

April to July. Larva forming a large blotch mine in leaves of sunflower, the scattered frass mostly on one side. Whitish green, sometimes shaded with rosy. Head mottled brown, with whitish front; cervical shield whitish green with two large brown spots covering most of its surface. Tubercles dark. Pupa usually in the mine. Three broods, the last hibernating as larva in the cocoon.

Northern and central Illinois; St. Louis, Missouri; Texas.

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44. PERISPASTA Zeller

(*Pionea*, in part)

Similar to Pyrausta; fore wing in male with \mathbf{R} , divergent from \mathbf{R}_{3+4} , running through the middle of a large hyaline patch beyond the cell, \mathbf{M}_1 in male below the hyaline patch, near lower angle of cell; in female from middle of end of cell; fore wing falcate.

1. **P. cæculalis** Zeller. Dark fuscous, with a faint excurved and sinuous darker postmedial line; fringe white in notch; hind wing nearly concolorous, with faint postmedial line and fringe white toward apex.

Not common. June to August.

Quebec and New Jersey to Manitoba, Colorado, and Texas. "New York" (Grote).

I have taken this species within two miles of the New York boundary, at Ramsey, New Jersey.

45. CINDAPHIA Lederer

(*Phlyctania*, in part)

Similar to Pyrausta; fore wing subfalcate; abdomen rather long. Hind tibiæ of male much swollen, with both outer spurs reduced; mid-tibiæ slender.

1. C. bicoloralis Guenée. Orange; outer two-fifths brown, the boundary marked by the sinuous darker brown postmedial line. Hind wing similar. 18 mm. (H 47:40.)

May and June; late July to September.

Generally distributed. New York: Lancaster, Ithaca, Big Indian Valley, Albany, Rhinebeck, Poughkeepsie, New Windsor, New York City.

46. PHLYCTÆNIA Hübner

(Pionea Guenée; Udea Guenée)

Similar to Pyrausta; palpi usually longer (fig. 343); middle tibiæ usually swollen; hind tibiæ usually with upper outer spurs rudimentary.

The genus is hardly distinct from Pyrausta. For key to species see Pyrausta.

* Upper outer spur of hind tibia much reduced.

1. P. ferrugalis Hübner (Celery leaf tier). Palpi twice as long as head. Hind tibia with upper outer spur obsolete. Dull light brown, sometimes reddish, or dusted with black; orbicular a slightly darker circle, reniform an hourglass-shaped spot; postmedial line parallel to outer margin from \mathbf{R}_3 to \mathbf{Cu}_2 , somewhat offset in on \mathbf{R}_3 , and deeply looped in on \mathbf{Cu}_2 and fold; with fine dark terminal dots and a double gray line in fringe. Hind wing much paler; fuscous toward outer margin, with a fuscous bar on upper half of discocellular; a spot on crossvein \mathbf{m} -cu, distincter below, and a fine dark gray postmedial line. 18 mm. (*rubigalis* Guenée).

Common all the year. Larva transparent green, with a white subdorsal stripe, and a pair of black spots on cervical shield. Head and tubercles pale. A general feeder on herbs, often injurious in greenhouses. Several broods.

feeder on herbs, often injurious in greenhouses. Several broods. Massachusetts to Missouri and south. New York: Rochester, Brockport, Buffalo, Crosby (Yates County), Ithaca, Big Indian Valley, Albany, Pearl River, New Windsor.

2. P. acutella Walker. Straw color, somewhat shaded with pinkish, broadly

streaked on the veins and more or less suffused with pale brown; no transverse markings, not even terminal dots. Hind wing yellowish, unmarked. 25 mm. (venalis Grote).

June to August.

Quebec to Wisconsin, south to southern New Jersey and Colorado. New York: vicinity of Buffalo, Potter Swamp (Yates County), Ithaca, Karner, Nassau, New Windsor.

3. P. terrealis Treitschke. Palpi as long as head. Pale fuscous, heavily powdered on a luteous ground; markings darker, faintly defined with pale, diffuse, obscure. Antemedial line excurved, postmedial sinuous, running in to below the cell at Cu and then obliquely into inner margin at two-thirds way to hind angle; orbicular a vague dot, reniform a dark bar. Hind wing paler and translucent, except at margin, with diffuse darker postmedial line. A distinct gray ter-minal line and two dark lines in fringe on both wings. 25 mm.

Two broods, flying mainly in May and August. Larva green, with black-dotted head and small black tubercles; on Solidago.

Quebec, south to northern New Jersey, west to British Columbia and Cali-fornia; Europe. New York: Peru, North Creek, Lancaster, Jamestown, Crosby (Yates County), Ithaca, Big Indian Valley, Onteora Mountain, Fort Edward, Schenectady, Rhinebeck, New Windsor.

4. P. extricalis Guenée. Cream color, dusted with dull brick-red, more coarsely than in P. terrealis. Antemedial line dentate, postmedial deeply and sharply dentate, drawn in below cell on Cu_2 , as usual. Subterminal absent. Orbicular an obscure point, and reniform a vague lunule. Hind wing similar, postmedial line far out across M₂ to Cu₁, as usual in the obscure Pyraustas; nearer outer margin than end of cell, then sharply angled in at Cu_2 ; usually strongly dentate; often distinct only opposite the end of the cell. 25 mm. (dionalis Walker, oppilalis Grote).

May and June; July; September. Larva a leaf roller on alder.

Generally distributed. New York: Peru, Newcomb, Big Indian Valley, Schenectady.

The coarser, less regular dusting, obvious to the naked eye, will distinguish this species from P. terrealis and theseusalis.

4½. P. helvalis Walker. Similar to P. extricalis; normally more powdery. rougher looking, with orbicular a ring and reniform at least a double bar; hind wing paler, with more dentate postmedial line.

This species is only certainly distinguishable from *extricalis* on genitalic characters, and I am not at all sure I have credited the various New York records to the right species. Both tend to be redder in the southern part of their range and grayer in the north. The caterpillar of helvalis is a leaf roller on poplar, emerging in July.

Nova Scotia to Missouri. New York: Black Brook, Ithaca, Neversink.'

5. P. theseusalis Walker. Palpi exceptionally short. Wings dull ochreous, shaded with dull light brown, or, more often, almost wholly light brown, except narrowly beyond the postmedial line. Lines contrasting, hardly diffuse, dark brown, antemedial excurved, practically even; postmedial waved and sinuous; orbicular a dot, and reniform a brown bar. Terminal line continuous. Hind wing similar. Discal dot of hind wing below obscure. 25 mm. (feudalis Grote). July. Larva rolling the tips of various ferns, in June.

General in distribution; Canada to Florida. New York: Mt. Marcy, Lewis County, Lewiston, Ithaca, Karner, Rhinebeck, Katonah, New Windsor.

6. P. tertialis Guenée. Pale straw yellow or luteous, marked with dull brown; lines single, contrasting, brown; antemedial excurved, postmedial sinuous and strongly bent in below end of cell, strongly dentate; subterminal in light specimens deeply dentate, the teeth touching the postmedial and terminal lines alter-

nately, enclosing yellowish lunules beyond the postmedial and a fine dentate yellowish line before the terminal dots, which are triangular and alternate with the teeth of the subterminal line; a continuous brown line in base of fringe; hind wing similar. Orbienlar and reniform usually suffused brown patches. 15–20 mm. (*pleetilis* Grote and Rohuson; syringicola Packard). (H 47:47.)

In dark specimens the wing suffused with brown, the yellow only showing in small spots and streaks, except a postmedial patch on the costa, one between the orbicular and reniform, and a larger one between the reniform and postmedial line beyond cell.

May to July; August. Larva in pith of elder.

Generally distributed; very common northward. New York: common every-where.

** Outer upper spur of hind tibia of male almost half as long as inner, about as in female; palpi beaklike, exceeding the head by twice its length.

+ Fore wing exceptionally broad and squarish.

7. P. fumalis Guenée. Fawn to chocolate brown; lines dark, not strongly dentate, single and contrasting; antemedial convex; postmedial somewhat excurved above; well out, nearly parallel to outer margin; angled on Cu, and sharply concave to inner margin, but only running halfway into cell; reniform a hunule or obscured by a blackish patch. Hind wing grayer, with a slightly wavy postmedial line parallel to outer margin, the disc often suffuse with dark gray. 22 mm. (H 47:55.)

June to September.

Nova Scotia to Pennsylvania and west. New York: Wilmington, Saranac Lake, Old Forge, Newcomb, North Creek, Fentons (Lewis County), Honeoye Falls, Oak Orchard Swamp, Ithaca, Big Indian Valley, Karner, New Windsor.

†† Fore wing narrow, with short outer margin and arched costa.

8. P. itysalis Walker. Luteous, the costa shaded with blackish, and orbicular and reniform large and blackish; hind wing with obscure markings, whitish, with a black dot on m-cu, as in *P. ferrugalis.* 28 mm. (*P. turmalis* Grote, *hyperborealis* Möschler, *tillialis* Dyar.)

July.

This species is transitional to Nomophila, but is easily distinguished from *N. noetuella* by its light ground color and its less exaggerated wing form.

Labrador; Mt. Washington, New Hampshire (5000 feet). Widespread in the Western States.

+++ Fore wing narrow, triangular, with straighter costa.

9. P. roseopennalis Hulst. Fore wing with oblique outer margin and subfalcate apex; hind wing rounded; antennæ annulate with rows of raised scales; front oblique, palpi fully twice as long as head; hind wing with \mathbf{M}_2 and \mathbf{M}_3 well separated, as in the last group of Loxostege. Fore wing pale yellow, with pink costa, except sometimes in the subterminal region; with the orbicular and reniform partly confluent with it; pink outer margin, and a large patch on outer part of inner margin, sometimes touching the reniform. Hind wing white, slightly yellowish at the apex. 18 mm.

May, June, and September.

Virginia; North Carolina; New Brighton, Pennsylvania; Arizona. P. radiosalis Mösehler, from Labrador, is unknown to me.

47. PYRAUSTA Schranck

(*Botis*, etc., in part)

Palpi beak-like, exceeding head by rather more than its length, usually more or less hairy (fig. 344); the third joint not well set off; eyes normal or nearly so; maxillary palpi porrect; front oblique but not conical or chisel-shaped. Hind tibiæ of male with spurs normal. Wings ample (fig. 324). Fore wing with all veins normal, divergent; \mathbf{M}_2 not at all concave at origin, hardly approximate to M_a ; frenulum-hook often absent; hind wing with M_a and M_a only shortly approximate. No decided secondary sexual characters. Markings various.

In the first group of the genus the markings are, except as noted, single, dark, on a somewhat lighter ground, with a more or less distinct paler shade beyond the postmedial line; antemedial line excurved, more or less waved, from costa at a quarter way out to inner margin before a third way out; the postmedial waved or dentate, well out, and concave opposite cell; excurved from M_1 to Cu_1 , then running in deeply to below end of cell above Cu2; nearly perpendicular to inner margin; the subterminal line distinct, diffuse, or absent. The orbicular and reniform are normal in position, the former circular and the latter kidney shaped. The hind wing is similar or somewhat paler, with the antemedial line absent; a curved discal bar confined to the discocellular vein; no dark spot on m-cu; postmedial line running much as in fore wing, nearer outer margin than end of cell across M_2 to Cu_1 , concave above and retracted below Cu_2 ; subterminal as in fore wing. The under side is much like the upper side, but with the markings more diffuse and the fore wing shaded with fuscous. The species are very close and some are doubtful. Of the Phlyctænias, P. terrealis, extricalis, theseusalis, and helvalis have similar patterns except as noted, also the Crocidophoras, Pilocrocis ramentalis, Pachyzancla perfusalis and bipunctalis, the Syleptas, etc.

Key to species of Pyrausta and similar forms

1. Both wings with basal half yellow and outer half brown.

(Cindaphia bicoloralis).

- 1. Basal half of wings not contrastingly paler than outer half. 2. Ground black; rarely, somewhat grayish on fore wing.
 - 3. White fringe only, or a faint postmedial shade also 30. niveicilialis.
 - 3. Gravish black, with darker postmedial line...... (Perispasta caculalis).

 - 3. A white sinuous postmedial line and other markings.

(Diasemia janassialis).

- 2. Immaculate dark lead gray; base dusted with white......32. demantrialis. 2. Ground not black (rarely, purple-black marked with yellow).
 - 3. Fore wing with longitudinal streaks only (Phlyctania acutella). 3. Fore wing with transverse markings, with spots, or immaculate.
 - 4. Hind wing nearly immaculate yellow, contrasting with the brown fore
 - 4. Hind wing not yellow and contrasting with the dark fore wing.
 - - 6. Hind wing with dark border and fine postmedial line only; fringe of fore wing dark.
 - 7. Fore wing with a yellow patch beyond ccll, before the post-medial line, which is often obscure above, but always distinct and strongly excurved on under side 20. ochosalis.

- 7. Fore wing with yellow postmedial markings beyond the postmedial line, which is always distinct.
 - 8. Hind wing with two yellow bands parallel to outer margin.
 - 28. subsequalis, borealis.

 - 8. Hind wing with postmedial yellow band straight as far as Cu_{2} , and pointing to anal angle.
- 5. Hind wing pale yellow, without broad and clean-cut contrasting markings, or not yellowish (sometimes with a pattern of fine lines or dots).
 - 6. Fore wing marked with pink, crimson, or purple (reduced to slight shades in some *acrionalis*).
 - 7. Fore wing dominantly light yellow, with pink borders and a large patch at anal angle......(*Phlyctania roscopennalis*).
 - 7. Fore wing half pink and half yellow, the pink not forming an anal patch or postmedial fascia......22. laticlaria.
 - 7. Fore wing olive, more or less shaded with purple. 22. laticlavia, variety cinerosa.
 - 7. Fore wing varying from pink to violet; sometimes suffused with gray; often with a pale yellow terminal band and fringe.
 - 8. Hind wing with a diffuse gray shade at border, or all gray; yellow postmedial spot beyond the postmedial line.

19. acrionalis.

- 8. Hind wing with a definite dark border, suffused with purple; yellow postmedial spot beyond the postmedial line.
 - 9. Under side of cell with a square yellow patch surrounded on three sides with gray; a similar larger patch beyond cell. 17. phænicialis.
- 7. Fore wing with small white markings only.

8. Postmedial line reduced to a costal bar; antemedial absent. (24). angustalis.

- 8. Antemedial line more or less distinct; postmedial complete. 24. signatalis.
- 6. Fore wing without pink, crimson, or purple markings.
 - 7. A contrasting, sinuate, yellow postmedial line, sometimes interrupted on the middle third......26. nicalis.
 - 7. A large pale yellow postmedial patch only......27. unimacula.
 - 7. Markings more complex, with distinct dark postmedial line, or ground pale.

 - 8. No contrasting pale spots on fore wing, or with both spots beyond the postmedial line.
 - 9. Outer margin broadly and evenly fuscous beyond subterminal line.
 - 10. Subterminal line deeply dentate, forming strong teeth; or (*nubilalis*) overlaid with the marginal shade.

11. Ground straw yellow, shaded with reddish and choco-

10. Subterminal line slightly irregular, sinuous, clean-cut (Sameodes species).

- 9. A broad subterminal band, contrasting with the yellowish outer margin; ground yellowish. 10. Subterminal band much broader than margin, clean-cut

16. fodinalis.

- 9. No subterminal band; ground broadly dark beyond the postmedial band, which is near the margin....31. inquinatalis.
- 9. Not yellow; or white with a broad dark terminal or sub-terminal band.
 - 10. Fore wing with white ground, more or less heavily dotted with black; often with large brown or black orbicular and reniform spots.
 - 11. Palpi twice as long as head.... (Phlyctania itysalis).
 - 11. Palpi projecting beyond head by a little more than its length11. illibalis.
 - 10. Fore wing not white and dotted or suffused with black. 11. Palpi twice as long as head, reniform constricted at
 - middle, hourglass-shaped...(Phlyctænia ferrugalis). 11. Palpi shorter; reniform not hourglass-shaped.
 - - 12. Hind wing with postmedial line even and nearly straight, from near costa to Cu close to lower angle of cell, tending to become obsolete; ground pale vellow.
 - 13. Postmedial evenly sinuate, almost always with dark brown spots before it opposite cell and over the roots of M₂ to Cu₁..... (Crocidophora species).
 - 13. Postmedial regularly dentate; at most with a diffuse brown shade before it.
 - 14. Ground bright yellow; in male, overlaid with brown; with clear white stripes on sides of front10. nubilalis.
 - 14. Ground paler straw yellow; male dusted with rusty or shaded with light brown; stripes on sides of face paler straw yellow, obscure.

8. penitalis, 9. ainslei.

- 12. Fore and hind wings with similar complex markings; the hind wing often paler, with the markings somewhat simplified, but postmedial line at least distinct, and offset out. or running far out, as near margin as to cell, opposite lower angle of cell. 13. Straw to lemon yellow, marked with brown.
 - 14. Subterminal line much heavier than postmedial. more or less diffuse, and suffused with the red-brown terminal shading (6). oxydalis.

14. Subterminal line not heavier than postmedial;

13. Not bright yellow.

- 14. Reniform a solid bar, sometimes nearly obsolete; subterminal line obscure or absent.
 - 15. Subterminal a distinct dark shade; median area more or less suffused with fuscous, leaving the space between orbicular and reniform contrastingly pale....4. *aglealis*.

 - 16. Postmedial line obscurely or not at all dentate.
 - Wings very broad; postmedial line in fore wing crossing Cu₂ nearly at right angles, well beyond cell. (*Phlyctania fumalis*).
 - 17. Wings normal; postmedial line retracted in on Cu_2 to below end of cell.
 - 18. Ground dull brown or ochreous to light fuscous.
 - 19. Discal dots and lines blackish, contrasting.: (Phlyctania theseusalis).
 - 19. Discal dots and outer line obscure.

(Phlyctænia terrealis).

- 18. Ground cream color.....7. submedialis.
- 14. Keniform more or less distinctly double, with pale center; always thick and squarish; subterminal line distinct, dentate.

 - 15. Postmedial line continuous, subdentate; subterminal continuous, dentate, delicate; ground vellow.
 - 16. Expanse under 25 mm.; less iridescent.

1. pertextalis.

16. Expanse over 25 mm.; markings coarser; ground whiter, with bluish iridescence.

- 14. Reniform double; subterminal line completely absentPhlyctenia helvalis.
- 12. Hind wing hardly, or not at all, marked.
 - 13. Immaculate ochreous, or with normal markings somewhat ill-defined and light red-brown.
 - 14. Hind wing concolorous.....13. inconcinnalis.
 - 14. Hind wing nearly white 12. futilalis variety.
 - 13. Fuscous; nearly immaculate, or with inconspicuous darker markings.
 - 14. Hind wing evenly dark fuscous, or with a pale postmedial shade.....15. unifascialis.
 - 14. Hind wings whitish.....12. futilalis.
 - 13. Dull olive, immaculate; small, with pointed fore wing......22. laticlavia, variety cinerosa.
 - 13. Dark olivaceous, with diffuse pale postmedial band. 15. unifascialis.
 - 13. Violet-gray with yellow costo-apical bar. 24. angustalis.

^{2.} thestialis.

1. P. pertextalis Lederer. Cream-white; markings light gray-brown, clean-cut; postmedial line hardly waved, sinuous; subterminal clean-cut, contrasting, deeply dentate; orbicular a distinct ring, reniform a double bar (unlike (rocidophora serratissimalis, which is superficially very similar); some light brown streaks on veins, and also between them along outer margin; terminal line continuous, dark brown. Hind wing closely similar. Discal dot practically obsolete on under side, distinct above. 20-28 mm. (H 47:54.)

Two broods, flying mainly in June and August.

General in distribution; not rare. New York: Fentons, Honeove Falls, vicinity of Buffalo, Ithaca, Big Indian Valley, Albany, New Windsor, New York City; Lynbrook, Long Island.

P. gentilis Grote appears to be the name for the smaller yellower specimens, with clean-cut markings; feeding on Erigeron. Large variants of this type, approaching P. thestialis and P. fissalis, have been bred from Ilex and Clethra.

2. P. thestialis Walker. Markings as in P, pertextalis, ground paler, with decided violet iridescence; orbicular and reniform often filled with brown, contrasting; terminal space often suffused with brown, slightly paler than the subterminal line. Hind wing with a blackish discal spot, contrasting below as well as above. 28-35 mm.

Very close to the preceding species but apparently distinct. June and July. Larva on Corylus, linden, and Euonymus. Generally distributed. New York: Mt. Marcy, Lewis County, Lewiston, Ithaca,

Karner, Rhinebeck, Katonah, New Windsor.

3. P. fissalis Grote. Ground nearly pure white. Pattern as in P. pertextalis, but more or less broken up into arrowhead marks on the interspaces. Discal spot on hind wing distinct above, completely absent below, except as it shows by transparency. 25-30 mm.

Larva on sassafras and goldenrod in late May; long, fusiform, grayish yellow, shining; tubercles large, shining, brown-stained around the edges; ia and ib of thorax partly fused. Head and cervical shield pale brown. Pupa in a slight web in crumpled leaves. Moth from June to August; commonest in July.

Atlantic States north to Maine. New York: Otto, Ithaca, Utica, Little Falls, Catskills.

4. P. æglealis Walker. Cream color, heavily shaded with dull brown; often with only the space between the reniform and orbicular spots pale, and a rather clearly defined pale shade beyond the postmedial line; in light specimens with the subterminal line represented by a vague dark shade. Veins more or less dark-streaked; terminal line usually marked by a series of darker dots. Discal dots of fore wing above simple, of hind wing below nearly obsolete. 30 mm.

Larva on Phytolacca; also reported from Osmunda, possibly confused with *P. theseusàlis.* Moth from June to August; common in July. General in distribution. New York: Buffalo (VanDuzee), Liberty (Morton),

Ithaca; Bucks Hollow, Staten Island.

5. P. langdonalis Grote. Cream color; antemedial line obscure; postmedial line merely the boundary of the very broad dark brown subterminal space, normal in course. Reniform a thick spot. Outer margin narrowly cream color. Hind wing with a similar smaller discal dot, and equally broad border. (H 47:52.)

Late June and July.

New Jersey to central Illinois, south to Tennessee.

6. P. flavidalis Guenée. Pale lemou yellow, shading into cream color, usually bright yellow along the margins; sometimes with only the costal margin bright yellow; markings red-brown, sometimes mixed with blackish; normal, tending to break into spots; the horizontal part of the postmedial line usually absent, and the part of the postmedial from Cu₂ to the inner margin apparently continuous with the reniform. Subterminal line thick, irregularly offset on veins; terminal line fine. 28-35 mm.

Late July to August. Larva a root borer in Vernonia, usually pupating in stubs of plants of the previous year in June.

Long Island, New York, to Illinois and south. New York: Florida, New Windsor, Long Island.

P. oxydalis Guenée, which is similar to *P. flavidalis*, but with heavy, suffused, red-brown marginal markings, is probably confined to the Southern States.

7. P. submedialis Grote. Cream color; markings normal, pale grayish, faint; orbicular a small circle, reniform when most distinct, with pale center; no traces of subterminal line. Hind wing whitish with distinct broken postmedial. Under side with a broad gray subterminal band on fore wing; discal dot of hind wing obscure. 25 mm. (dissectalis Grote, pilalis Hulst.)

Canada; Illinois; Florida; Texas; Arizona.

8. P. penitalis Grote. Pale yellow, with contrasting and not very diffuse rusty ochre, rarely grayish, markings and dusting; antemedial line erect, sharply dentate, not running out to inner margin; reniform a slight vertical bar, followed by more or less obvious rusty or grayish streaks or shades; postmedial line fine, moderately dentate, sometimes obscure in female, drawn in below cell as usual, produced out roundly in submedian space, but running in again to inner margin. Subterminal a broad but distinct dentate shade, or, more rarely, a deeply dentate line; hind wing not dark at base; with slight discal bar and even postmedial and terminal shades, the latter slightly paler on the margin, or with the postmedial shade indicated only as the outer boundary of a darker discal area; in the female often with only traces of the middle part of the postmedial line, which is near the end of the cell, unlike the similar, nearly immaculate yellow specimens of *P. inconcinnalis* and *futilalis*. Male with a patch of sex-scaling on iuner margin of fore wing below. Juxta with a single, stout, ventrally projecting spur. 25 mm. (*nelumbialis* Smith.)

The larva is aquatic, on water lily and Nelumbo; rarely in other aquatic plants. Pupa without a frontal hump; with cremaster broader than long.

Apparently general in distribution, but obscure and usually misidentified. New York: Lynbrook, Long Island.

9. **P.** ainslei Heinrich. Similar to *P. penitalis;* fore wing without reddish shading and powdering, no sex-patch on inner margin, on under side. Juxta with two slender dorsally projecting arms embracing the ædœagus; apex of uncus rounded. 20-27 mm. (obumbratalis Lederer ?)

Larva boring in marsh plants, normally only in Polygonum, but frequently straying to other foods. Pupa with a prominent projecting frontal knob.

Generally distributed; widespread in New York, but local.

10. P. nubilalis Hübner (European corn borer). Pattern exactly as in *P. penitalis*; ground brighter straw yellow; male heavily overlaid with brown, leaving yellow areas in the antemedial region, between the ordinary spots, and beyond the postmedial line. Under side of fore wing with a sex-patch, as in *P. penitalis*. Juxta with two dorsal arms, embracing the ædæagus. Uncus trifurcate. Female not certainly distinguishable from the two preceding species without dissection; normally slightly brighter in coloring. 30 mm.

Larva a serious pest in corn and other coarse herbaceous plants; easily distinguishable from the other corn-pests by the tubercle pattern of the genus, with ii much smaller and farther lateral than i; distinguishable from the two preceding species only with the microscope. Massachusetts strain with two broods, and showing a strong tendency to eat a great variety of plants; western strain singlebrooded in normal seasons, and practically confined, so far as now known, to corn. Hibernation in the well-grown larva, in the stubble. Pupa without a frontal hump; with cremaster longer than broad. Probably the most serious known corn pest.

Eurasia; recently introduced in eastern Massachusetts; and in the vicinity of Albany, New York, and the Lake Erie basin.

For a full account of these three species see the Journal of Agricultural Research; vol. 18, p. 171.

11. P. illibalis Hübner, race arsaltealis Walker. White, sometimes suffused with fuscous; shaded and spotted with blackish; orbicular a black spot, larger than in those species of the preceding group where it is solid; reniform a narrow vertical blackish bar, often forming the center of a blackish shade; both rarely obsolescent. Ordinary lines absent, terminal line broken into black bars, with a gray bar in the fringe opposite each. Hind wing usually a little suffused with clay color, and normally very lightly dotted with blackish and shaded with fuscous at margin; rarely, with a dark discal bar. 25 mm. (H 47:48.)

The typical form, with both wings almost pure white, with very little black or brown dotting, except for the broken black terminal line, and with contrasting large brown orbicular and reniform spots, appears to be confined to the South.

May and June.

Generally distributed north to Hymers, Ontario. New York: Buffalo, Ithaca, Big Indian Valley, New Windsor.

12. P. futilalis Lederer. Typically powdery dull fuscous; in the Mississippi Valley race, powdery ochre yellow; markings obscure; postmedial, when distinct, moderately dentate and sinuous, fine and dark; sometimes followed by a paler shade; terminal line often showing two dots to each interspace. Hind wing white, sometimes with traces of the postmedial line far out toward the margin, lightly shaded with fuscous (in the western race ochreous) along the outer margin. 30 mm. (erectalis Grote. The yellow form is penitalis Hulst, not Grote.)

June.

General in distribution. New York: Peru, Mt. Marcy, Lewis County, Lewiston, Ithaca, Karner (yellow form), Albany, Poughkeepsie, New Windsor.

13. P. inconcinnalis Lederer. Orange-ochre, practically immaculate; fore wing and margin of hind wing powdery; base of hind wing slightly paler. 28 mm. (crocatalis Grote, festalis Hulst.)

Western States; reported from Illinois, probably in error for the yellow form of P. futilalis.

14. P. fumoferalis Hulst. Powdery ash gray; lines deeply dentate, antemedial erect, postmedial moderately sinuous and followed by a dentate pale line or shade; the margin beyond perceptible darker. Terminal line black, usually broken. Hind wing light fuscous, with an even, slightly diffuse, darker border, often preceded by a pale shade. Veins more or less distinctly darker, often darkening into a series of short postmedial streaks; rarely, with a wavy postmedial line. 30 mm.

Cocoon under bark of hickory; characteristically formed of a fusiform central cocoon enclosed in two or three oval walls. Larva presumably on hickory. Moth in June.

Generally distributed; south to Pennsylvania and Illinois and west to California. New York: Mt. Marcy, Lewis County, Ithaca, Schenectady.

15. P. unifascialis Packard, race subolivalis Packard. Dull olivaceous, powdered with cream-white on dark brown, with a more or less distinct. light, moderately sinuous postmedial band, parallel to outer margin, not bent in at all below cell, defined on inner side, diffuse on outer; sometimes obsolete, and sometimes very conspicuous. Sometimes with a more or less distinct pale patch in outer part of cell, the other markings obsolete. Hind wing duller, without greenish tint; with or without an irregular pale postmedial shade or bar, sometimes defined with a darker line on inner side. 20-25 mm. (hercynalis Grote, obnigralis Hulst.) (H 47:51.)

June to July.

Maine to Illinois, and south to New York; the typical form in California. New York: Peru, Axton, Fentons (Lewis County), Lancaster, Liberty, Portage, Potter Swamp (Yates County), Ithaca. Big Indian Valley, Karner, Nassau, Rhinebeck, New Windsor.

16. P. fodinalis Lederer. Onter margin straighter than usual in the larger Pyraustas, often a little concave; apex marked. Fore wing light oclire, often shaded with olivaceous or reddish; orbicular a gray dot, reniform an oblong patch. Ante-medial excurved, and waved; postmedial finely dentate, running in along Cu_2 to before end of cell, very fine, gray; a broad gray subterminal shade from apex to anal angle, concave on outer side, sometimes wanting. Hind wing with a straight subterminal shade from near apex, usually fading out toward anal angle; post-medial line usually short and straight or absent. 20 mm. (socialis Grote.)

Wisconsin and west; rare eastward. New York: Lewiston, Lancaster, Buffalo, Ithaca, Big Indian Valley, Onteora Mountain.

17. P. phœnicialis Hübner. Bright straw yellow. Fore wing with a nearly marginal light brown or crimson border, reaching in almost, but not quite, to postmedial line, nearly even in width; postmedial very fine, deeply sinuous, and curving in below end of cell, but not dentate; brown, with a brown shade between it and end of cell and usually before it to inner margin; orbicular a gray dot; reniform a vertical lunule; antemedial line fine, excurved, with more or less brown shading before it. Fringe gray, or more or less yellow. Hind wing concolorous; border similar; postmedial nearly straight from costa at two-thirds to near anal angle, sometimes preceded by a brown shade. Beneath, with costa gray or crimson, extending across cell on basal half and at end, leaving a square yellow patch. Outer margin also dark. 15 mm.

Larva on apple.

Hemlock Falls, New Jersey; Florida. "New York" (Walker).

Variety flegialis Walker is larger and more deeply and richly colored, with a pale costa below. Various other varieties occur.

18. P. onythesalis Walker. Fore wing about as in *P. phænicialis*, the brown usually lighter and grayer; the fringe, and often a little of the outer margin, pale yellow, often crossed by a deep red, broken terminal line. Hind wing pale yellow with a fine straight postmedial line running to the outer margin near the anal angle, and a broad purplish terminal band, tapering off below. Fringe yellow. 20 mm. (probably *insignatalis* Guenée; *phænicialis* auct., in part, not Hübner.)

Larva perhaps on Salvia.

Illinois; Missouri; Texas; Arizona.

19. P. acrionalis Walker. Base mixed rose or light violet and light yellow, the lines and orbicular and reniform spots violet, partly obliterated by the violet shading; postmedial region from postmedial line almost to margin, solidly rose or violet, leaving a narrow pale yellow terminal band, and yellow fringe, sometimes separated by a broken red terminal line. Under side dark. Hind wing typically whitish, shading into gray toward outer margin; with some violet iridescence; in variety rufifimbrialis Grote mostly or wholly gray, usually with a fine clear yellow postmedial line; in variety haruspica Grote and Robinson with a dark fringe. 18 mm. (phanicialis auct., in part, not Hübner.)

July to September.

General in distribution. New York: Old Forge, Newport, Speculator, Lancaster, Otto, Ithaca, Big Indian Valley, Utica, Rhinebeck, New York City, Fort Montgomery.

20. P. ochosalis Holland (Fitch ms.). Deep purple-brown, with obscure pale markings, defining more or less distinctly the antemedial and postmedial lines; a conspicuous round yellow postmedial patch, not reaching costa; usually with a separate yellow costal dot at its upper outer angle. Margin and fringe dark. Hind wing blackish with a broad postmedial yellow band. 18 mm. (H 47:57.)

June to August.

Quebec to Arizona; a yellower variety in California. New York: North Elba, Oak Orchard Swamp, Ithaca.

21. P. generosa Grote and Robinson. Deep brown; postmedial yellow patch higher than wide and running through to costa, and sometimes narrowly to inner

margin; sometimes with a narrow postmedial line and a distinct vellow spot in cell. Hind wing about as in P. ochosalis. 10 mm.

Western Pennsylvania to Illinois and Manitoba. New York: Feutons (Lewis County), Newport, Lancaster, Buffalo, Karner, Sloatsburg. I have verified none of these records, which may belong wholly or in part to the preceding species.

22. P. laticlavia Grote and Robinson. Yellow, with a broad even rose-colored border; costa rose, down to \mathbf{R} ; and with a broad median area, of moderate width, constricted somewhat at fold, and containing a small yellow spot in cell. Hind wing duller luteous, with purplish border. 15 mm. In variety cinerosa Grote and Robinson, the yellow is replaced by olive, and the

rose by dark purple, or sometimes the whole moth is dull olive.

August.

Southern States, north to Five-mile Beach, New Jersey. "New York" (Grote).

23. P. tyralis Guenée. Fore wing typically deep crimson, sometimes fuscous suffused with crimson; an excurved antemedial fascia, and a somewhat sinuous postmedial one, yellow in crimson specimens, edged with crimson or wholly crimson in dark ones; sometimes with a small yellow spot at end of cell. Hind wing orange-red at base, shading into crimson at margin, rarely with a yellow postmedial bar. 15 mm. (H 47:44.)

September.

Southern States, north to Missouri and Illinois. 24. P. signatalis Walker. Rose pink; the ante- and postmedial lines narrow, pale yellow, and often partly obsolete; antemedial line erect, dentate; postmedial line sinuous and somewhat dentate, the strongest tooth on A; sometimes with yellow spots in median area; median area sometimes suffused with gray. Fringe ochreous. Hind wing pale yellowish, with a brownish shade toward margin. 18 mm. (virulenta Grote and Robinson.)

May, August and September. Larva on horsemint.

Southern New Jersey and central Illinois to Texas. New York: Lewiston, New Windsor (Morton).

P. angustalis Grote is rather darker crimson, shaded with brownish gray; the ordinary lines are more reduced; usually with only a short straight whitish postmedial bar on costa at three fourths way to apex. The hind wing is mostly brownish gray, with dark fringe. This species appears to be purely western.

25. P. inveterascalis Barnes and McDunnough. Evenly powdered with blackish on a dull crimson base. Costa more solidly crimson, antemedial line fine, pale yellow, zigzag, stopping at vein R; postmedial perpendicular to costa, parallel to outer margin below, fine, yellowish, often reduced to scattered scales except for a yellow bar at costa. Margin and fringe narrowly yellow. Hind wing duller, without red; with a pale postmedial line from middle of M_2 to near outer margin at Cu_2 , then angled and faintly extended toward anal angle. Below, dark fuscous with a pale margin and a postmedial costal dot on the fore wing; and with a postmedial shade and basal shading on hind wing. 15 mm. (angustalis of collections, not Grote.)

June and July. Only types seen. New Brighton, Pennsylvania.

26. P. nicalis Grote. Fore wing fuscous, with a slight dull purple tinge; postmedial line narrow, contrasting, whitish, moderately sinuous, twice as wide at costa and at inner margin as at middle, sometimes practically interrupted at middle, antemedial usually absent. Hind wing duller tuscous, with somewhat darker outer margin; postmedial line somewhat waved, erect, approaching outer margin near anal angle. 22 mm.

July.

Hymers, Ontario, to Colorado and California.

27. P. unimacula Grote and Robinson. Mouse gray, with a large pale yellow postmedial patch, resting on costa; sometimes with a trace of the postmedial line on its outer side. A small yellow dot in fold. Hind wing similar, the whole median area yellowish. 18 mm. (H 47:56.)

End of May to August.

New York to Illinois and Florida. New York: Ramapo, Brewster's (Grote).

28. P. subsequalis Gnenée. Typical female: fore wing ranging from ochre yellow to dull orange; antemedial line wavy; postmedial sinuous, and widened into a series of black wedges; both lines rather thick and somewhat irregular; large black orbieular and reniform spots; subterminal a broad shade, narrowed or interrupted in middle: often with a blurred blackish median line, or the median area somewhat suffused with blackish. Hind wing bright orange, with a large black discal dot; two outer black bands parallel to outer margin, and some longitudinal streaks on basal half of wing along Cu and the anals. Fringes black. Typical male: fore wing suffused in two shades of fuscous, and scaled with dull red; with traces of the markings of the female; the hind wing black, with short postmedial and terminal orange lines only. 20 mm. (insequalis Guenée.)

Larva on thistle. Moth in June and late July.

Northern New York to Illinois and south. New York: Black Brook (male).

The northern variety **borealis** Packard, is a little smaller than the female of the preceding form, with the same markings: fore wing light oehreous and fuscous; hind wing yellow, with the basal half almost solidly fuscous, obscuring the discal dot; outer markings rather light. Nova Scotia to British Columbia.

dot; outer markings rather light. Nova Scotia to British Columbia. Specimens from Massachusetts, New York, etc., are generally intermediate, with the sexes similar, but with the base of the hind wing orange with gray streaks as in subsequalis. These are var. madetisalis Walker (*efficitalis* Walker.) (H 47:41.)

April to July.

Generally distributed and not rare locally. New York: Black Brook, Saranac Lake, Ithaca, Big Indian Valley, Ilion, Schuyler, Nassau, Rhinebeck, New Windsor, Suffern, New York City; Tottenville, Staten Island.

29. P. funebris Ström. Black. Tegulæ yellow. Two large rounded pure white spots on each wing. Fore wing often with a third small spot below the costa between the two large ones; abdomen finely striped with white. 20 mm. (octo-maculata Linnæus.) (H 47:50.)

June and July. Larva light yellow-green, with yellowish head; living on the under side of the basal leaves of Solidago.

Generally distributed northward, south to New Jersey at least; Eurasia. New York: Common and generally distributed.

30. P. niveicilialis Grote. Dark, smoky gray, nearly black, with a pale postmedial shade, usually distinct only near the costa; fringe white, contrasting. Hind wing black, with white fringe. 22 mm.

May and June; August.

Canada to northern New Jersey, Pennsylvania, and Colorado. New York: Crosby (Yates County), Ithaca, Albany, Poughkeepsie, and New Windsor.

31. P. inquinatalis Zeller. Palpi moderate, tibial spurs normal. Pale powdery gray, shaded heavily with dark gray at base and outer margin. Antemedial line hardly one-fourth way out, slightly outwardly oblique and dentate; postmedial line blackish, finely dentate, and slightly excurved from costa to Cu, then in to below end of cell, where it forms a dark bar, and out again on A; below A, about in line with its position above, two-toothed. A pale shade between the postmedial line and the dark border. Costa more or less gray shaded; orbicular a gray patch, reniform kidney shaped, and dark gray, separated from orbicular by its width. Hind wing paler toward base, with a waved weak postmedial line; discal bar and dot at lower angle of cell, and dark border. 20 mm. (glacialis Packard; Phlyctania auct.)

May to July. Male unknown to me.

Ottawa, Ontario; Labrador; Europe.

32. P. demantrialis Druce. Antennæ rather thick and rough-scaled at base. Front strongly oblique, but without a cone; palpi rather long; fore wing with \mathbf{R}_{s} more curved than usual, the wings narrow and subfalcate. Upper outer spur of hind tibia of male only one-fifth the length of the inner.

Fore wing shining lead gray, immaculate, the basal half overlaid with blacktipped whitish scales. Hind wing somewhat paler toward base, not powdered. 20 mm. (Blepharomastix Druce; Bæotarcha Hampson; chalybealis Fernald manuscript.)

July and August. New York, to Arizona, south to Central America. New York: Ithaca. **P. singularis** Lederer, from "North America," is probably a stray from the tropics. It is described as flesh color with a black tuft at basal angle. 33 mm.

P. venalalis Hulst and abdominalis Zeller are unknown to me; by the original descriptions the first suggests Loxostege helvialis, and the second P. fissalis. Torvalis and marginalis have been treated as Titanios.

48. LOXOSTEGOPSIS Dyar

Wings as in Pyrausta; Sc and R of hind wing rarely approximate, instead of anastomosing (fig. 325). Palpus beak-like, extending barely its length beyond the head; maxillary palpi large and triangular, filling the space above the labials. Tongue weak, typically obsolete.

This may be the primitive genus of Pyraustinæ, but in our present uncertainty as to Pyralid evolution, I shall not disturb the customary order to put it at the head.

1. L. merrickalis Barnes and McDunnough. Brown-black; ante- and postmedial lines darker, obscure on the dark ground, the postmedial sinuous. Head and top of palpi clay yellow, contrasting. 15 mm. (Pyrausta Barnes and McDunnough). July.

New Brighton, Pennsylvania. New York: Ithaca.

49. THELCTERIA Lederer

(Eustixia Hübner?)

Palpi rather long and porrect obliquely, the third segment not long but well marked and porrect; much shorter than in Scoparia. Maxillary palpi very large and broadly triangular, as in Scoparia; front with a conical prominence; wings broad with normal venation; all veins spaced.

This genus is possibly an aberrant Scopariine.

1. E. pupula Hübner. White with about ten jet-black dots, some of them, rarely, partly fused into ante- and postmedial lines; one spot basal, two terminal, and one representing the reniform. Hind wing white, or with slight fragments of postmedial and subterminal lines. 15 mm. (H 47:58.)

Locally common all season.

Massachusetts to Texas. New York: Poughkeepsie, New Windsor; Brooklyn, and East New York, Long Island.

Octonalis appears wholly unrelated, and is removed tentatively to Lygropia. Lineodes integra Zeller, easily recognized by its entirely Pterophorid appearance of wings, body, and legs, is a western species ranging east to Missouri. The larva is pale green with a pale brown head and black spots on either side of the prothorax. It webs the leaves of Solanum nigrum and other Solanacce.

WILLIAM T. M. FORBES

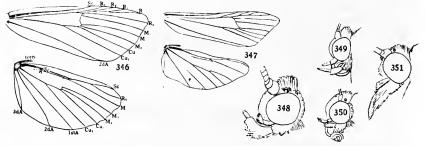
Subfamily NYMPHULINÆ

(Hydrocampinæ)

Similar to the Pyraustine, except for the usual stalking of \mathbf{R}_2 with \mathbf{R}_{3+4} (fig. 346). Maxillary palpi always well developed (figs. 348 to 351); labial palpi normally upturned with well-marked segments, and a good-sized third segment. Caterpillars of all our known native species aquatic, forming some sort of a

Caterpillars of all our known native species aquatic, forming some sort of a case or shelter; frequently with tracheal gills, and spiracles reduced. Lateral setæ on ninth segment of abdomen obscure, and i close to iii; hooks arranged in a complete ellipse, alternately of two lengths.

Those caterpillars without gills show no single difference from the Pyraustinæ.



FIGS. 346-351. NYMPHULINÆ

346, Nymphula icciusalis, venation; 347, Elophila fulicalis, venation; 348, Nymphula gyralis, side view of head; 349, Elophila fulicalis, side view of head; 350, Geshna (?) primordialis, side view of head; 351, Diathrausta reconditalis, side view of head

The pupe are very thin-skinned, and often with spiracles of two sizes, only the larger ones being functional. The cocoon is filled with air even in those species that breathe water in the larva; but is often completely submerged. So far as known to me the species are two-brooded, hibernating under water as part-grown larve. Some may feed a little through the winter.

Key to the genera

(Imago)

- 1. Hind wing with a vein lost (\mathbf{M}_2) .
- 1. Hind wing with all veins preserved.
 - Palpi porrect, with scaling continuous on second and third joints; terrestrial
 Palpi obliquely porrect, with well-set-off segments (fig. 350); tongue

 - 2. Palpi upturned, with strongly curved second joint, and close-scaled third, well set off; aquatic.
 - 3. Maxillary palpi regularly tapering to apex (fig. 349); outer margin of hind wing with a metallic and black border extending to Cu_2 .

(Larræ)

I. Aquatic.	
2. Without tracheal gills.	
3. Case ovate, on Lemna	
3. Case sharp-edged, on other plants	
2. With simple tracheal gills, under a web on rocks	
2. With branching tracheal gills, in a floating case	
50. Nymphula (Parapoynx)	
1. Terrestrial, on nettle	

50. NYMPHULA Schranck ·

(Hydrocampa Latreille, Parapoynx Hübner, Paraponyx auct., Oligostigma Guenée, etc.)

Palpi obliquely upturned, with second joint well set off and rather broadly scaled, third conical, much slenderer, and nearly as long (fig. 348); maxillary palpi rough-scaled, and truncate at tip or triangular; large, except in N. ekthlipsis; tongue not very strong; ccelli small, but present. Antennæ usually annulate, with the outer row of scales on each segment raised; legs long and slender.

Fore wing (fig. 346) with \mathbf{R}_1 free, \mathbf{R}_2 usually stalked, free in odd specimens of several species; \mathbf{R}_s free, divergent; hind wing with Sc and R fused a considerable distance, the other veins free; \mathbf{M}_2 and \mathbf{M}_3 approximate at base. Sexes often strongly dimorphic in size and markings. Female with exserted ovipositor. Caterpillar aquatic, in a case, or under a piece of a leaf cut out and attached

to the surface of the leaf (when a case is made, it is of two pieces of leaf, and has sharp edges). Larva often with branched tracheal gills, and breathing water. Pupa in the case, which is more heavily lined with silk than while used by the larva, and in the species whose larvæ breathe water, is emptied of the water.

Key to the species

1. Hind wing wholly white, or with one or two faint lines.

2. Fore wing yellow or brown, markings on hind wing more distinct.

4. gyralis, in part. 2. Fore wing mouse gray, or white, or gray and white with a contrasting 1. Hind wing with diffuse gray lines, with diffuse grayish discal spot.

4. gyralis, in part.

1. Hind wing with two fine, closely parallel, wavy black lines.....9. allionealis. 1. Hind wing with two even and parallel mesial bands.

2. Fore wing coppery and gray, with broad even white postmedial fascia.

6. seminealis.

- 2. Fore wing dull fuscous brown, with a narrow wavy postmedial white line
- 2. Fore wing largely coppery, with obscure pale postmedial band, or none; with four black enlarged dots on outer margin of hind wing.
 - 6. seminealis Q.
- 1. Hind wing with two irregularly waved black lines, with a yellow discal spot between them.

	2. Discal	spot	edged	with	black.			 	1	. ekthlipsis.
	2. Discal	spot	not day	rk-edg	ed			 		2. icciusalis.
1.	Hind wi	ing bla	ckish v	with o	bscure	markii	ngs	 	3	. obliteralis.

Larva

- 1. With tracheal gills.
 - 2. Anterior and posterior infrastigmatal gills with the same number of fila-
 - - 6. seminealis.
- 1. No tracheal gills; second abdominal spiracle decidedly smaller than third; caterpillar stout and flattened, with head darker than body.
 - 2. Head dark chitin-yellow; case nearly circular.
 - 3. Body whitish; thoracic tubercles more distinct than abdominal ones; head with a lateral brown stripe; on Potamogeton......3. obliteralis.
 - 3. Body pale yellow; tubercles all similar; head with dark spot confined to mouth parts; on Nuphar.....4. gyralis (?).
 - 2. Head dark brown, body dirty gray; in an oblong case on Potamogeton and

I. Larva without tracheal gills, breathing dir; the case full of air; pupa with first open spiracle smaller than the other two; moth with fore wing typically subfalcate; Sc separating from \mathbf{R} halfway between cell and apex, the free part at least two-thirds as long as free part of \mathbf{R} ; palpi normally hairy below; antemcdial line of hind wing crossing cell perpendicularly (Hydrocampa, Nymphula).

1. N. ekthlipsis Grote. Maxillary palpi smaller than in the other species; half as large as third segment of labials; fore wing subfalcate. White, with three confused bands on basal half, the outer one angled out on Cu, and touching the lower end of the large kidney-shaped reniform; postmedial band yellow, its inner margin running in a curve from costa to lower outer angle of reniform, and in a shorter curve from inner angle of reniform to inner margin, with the lower part of the antemedial band enclosing a circular white spot. Subterminal band white; all the markings heavily edged with brown, sometimes largely covering the yellow; subterminal line brown, closely parallel to margin, followed by a light brown and yellow stripe and white fringe. Hind wing similar, the basal marks replaced by a straight antemedial band, edged outwardly with dark brown. 18-25 mm.

Larva in an oblong case on sedge.

General but local, south to New York and Illinois. New York: Ithaca, Michigan Swamp (Tompkins County), Albany.

2. N. icciusalis Walker. Yellow or brown, shaded with silvery white; base with considerable white; a white triangle at middle of costa with a black bar on its front edge; a white discal crescent, edged outwardly with black; a white circle in front edge; a white discal crescent, edged outwardly with black; a white circle in middle of fold, strongly outlined with black, and variable in size; subterminal line strongly irregular, sometimes edged outwardly with black, at least toward apex; fringe barred, brown and white. A fine white postmedial line following the outer edge of the reniform lunule and the circle on the fold. Hind wing similar; antemedial band as in *N. ekthlipsis*, postmedial line irregular, edged before, and sometimes beyond, with black; discal spot large, yellow; subterminal line as on fore wing. Ground of basal two-thirds white, of outer third, yellow or brown. Ground rarely both yellow and brown the region beyond the cell or brown. Ground rarely both yellow and brown, the region beyond the cell, and the base of the fore wing being contrastingly light; white and black markings of the fore wing sometimes suffused. 12-20 mm. (formosalis Clemens, genuialis Lederer.)

Caterpillar in an oblong case on Potamogeton, sedge, Lemna, Menyanthes, and other aquatic plants. Cocoon sometimes anchored under water. Moth in August. Common everywhere, flying over the water and along the shore of ponds. New York: Saranac Lake, Saranac Inn, Lancaster, Otto, Potter Swamp (Yates County), Ithaca, Big Indian Valley, Nassau, Rhinebeck, New Windsor.

3. N. obliteralis Walker. Fore wing not falcate. Dull black, obscurely marked with ochre and a little white, with obscure light blue iridescence; hind wing similar, or almost completely suffused with black; discal unule white on both wings. Female lighter, browner, with more diffuse markings, and much larger; discal lunule often very weak. 10-16 mm. Larva on water lilies (Nymphæaceæ); sometimes a minor pest in greenhouses.

Larva on water lilies (Nymphæaceæ); sometimes a minor pest in greenhouses. Quebec, and south; common southward. New York: Hemlock Lake, Rhinebeck. (Usually a greenhouse pest northward, but the Hemlock Lake specimens, at least, were taken outdoors.)

4. N. gyralis Hulst. Male fuscous or light brown; base a little darker, bounded by a waved white line; costal triangle a white outline, filled with blackish and edged with white, the outline often broken; reniform lunule a curved black line, edged with white and often connecting with a short black and white subterminal streak on costa; subterminal line white, irregular, edged with brown. Hind wing white, with vague fuscous ante- and postmedial lines, often fusing on dorsal half of wing; postmedial line followed with white, and strongly irregular; discal spot vague, grayish; subterminal line as in fore wing, in a brownish or yellowish shade. Female with markings the same as far as traceable; ground lighter brown, or dirty yellow, the markings often suffused; hind wing white with slight gray markings. 20 mm.

General, but relatively rare; larva on water lily.

New York: Big Tupper Lake, Saranac Lake, Lewiston.

In southern specimens portions of the ground are light ochre yellow; northern ones are usually wholly fuscous brown and white.

II. Larva with tracheal gills, two subdorsal, two subventral, and a pedal pair on most segments; the subventrals with an equal number of filaments; second and third abdominal spiracles equal, all very small; case filled with water; pupa with the three open spiracles equal in size, with a median longitudinal carina on ninth segment of abdomen. Imago with venation as in the last group; palpi merely rough-scaled; hind wing immaculate, or with a faint postmedial line only (Nymphxella Grote).

5. N. maculalis Clemens. Normal male. Fore wing not falcate; maxillary palpi rough and larger than third segment of labials; fore wing white, marked with dark gray, leaving the white mostly as ante- and postmedial patches in the fold; a vaguer one in the outer part of the cell, and two confluent ones near the outer margin. All the patches connected in light specimens; in dark ones only the two on the fold contrasting. Hind wing white, sometimes with a gray postmedial line on dorsal part. Normal female. Fore wing immaculate mouse gray, and hind wing white; much larger. 18-25 mm. (seminivella and dispar Walker.) Caterpillar on the water lilies, and rarely on Brasenia; in the last stage, with five filaments on the posterior subdorsal and all subventral gills of most segments, the anterior subdorsal usually with four Eirst stage without cills and the

Caterpillar on the water lilies, and rarely on Brasenia; in the last stage, with five filaments on the posterior subdorsal and all subventral gills of most segments, the anterior subdorsals usually with four. First stage without gills, and the intermediate stages with smaller numbers of filaments. Caterpillar immaculate, transparent till just before pupation, then turning yellow; pupa generally under a piece of leaf on a floating lily pad.

Common and generally distributed, flying over water. New York: Saranac Lake, Big Tupper Lake, Lewis County, Spencer Lake. The male variety feminalis Dyar, is dark gray, with obscure markings, much like the female; the female variety masculinalis Dyar is white with only traces of dark markings; the female is rarely brown, and often has a pale costa.

III. Larva as in the last group, but with one less filament on anterior than on posterior infrastigmatal gills; pupa as before. Imago with palpi merely rough-scaled; Sc and R of hind wing fused decidedly more than halfway to apex; the free part of Sc not more than half as long as R and sometimes very short; apex of hind wing more or less distinctly truncate. Antemedial line of hind wing strongly oblique, parallel to postmedial line, cutting across end of cell. (Parapoynx, Oligostigma.)

6. N. seminealis Guenée. Male whitish, powdered with dark brown, giving a chocolate brown effect. Outer line broad, even and white, strongly contrasting, nearly straight from costa to fold, then turning inward almost to middle of wing; preceded by an equally wide band of brilliant bronze. Subterminal space bronze, terminal space tawny; subterminal line grayish white and black. Female similar, but less brilliantly bronzy; postmedial line gray, inconspicuous. Hind wing with a narrow inner and broader outer line, and a bronzy band; several small black marginal dots in a group, below apex. 20-30 mm.

The caterpillar feeds on Limnanthemum, and perhaps Potamogeton. In the second stage it forms a trumpet-shaped mine on the lower surface of the leaf; and then has simple gills; the number of filaments gradually increasing to four in the last stage. The caterpillar eats more or less of the parenchyma within the case. This species doubtfully distinct from the South American *N. juncealis* Guenée.

Massachusetts to Florida.

7. N. obscuralis Grote. Ground color whitish, powdered with black-brown, as in the male of N. seminealis, but much less brilliant, the powdering forming a darker median shade; postmedial line white, narrow in female, broader in male, more or less toothed in, below cell in female, and often obsolete below M_3 . Subterminal space a little warmer brown, hardly contrasting. Hind wing marked as in N. seminealis, but entirely without bronze reflections; the black marginal points not emphasized below the truncation. 20-38 mm. (H 48:10.)

Caterpillar with a maximum of six gill-filaments; on Potamogeton and Vallisneria.

Generally distributed. New York: Waddington, Potter Swamp (Yates County), Ithaca.

8. N. badiusalis Walker. White; base of costa blackish; a short oblique dark antemedial band; a postmedial band running from costa two-thirds way out, perpendicularly down to Cu, then in along lower edge of cell to before middle of wing, and obliquely into inner margin; the band grayish, normally partly filled with yellow. Subterminal band yellow, edged with brown, roughly parallel to outer margin, running almost to inner margin, then turning in along the inner margin a quarter way to the base; marginal band yellow, edged within with brown; fringe pale. Hind wing with parallel antemedial, subterminal, and terminal bands, of nearly equal width, the two outer heavily filled with yellow. Male usually smaller, the subterminal band of both wings usually black, emphasized and contrasting; only the marginal band yellow. 15-20 mm. (albalis Robinson).

Moth in August. Early stages unknown. Common north of West Virginia and Illinois.

New York: Saranac Lake, Waddington, Lake Bluff, North Fairhaven, Buffalo, Hemlock Lake, Ithaca, Rhinebeck, Ulster County (Dyar).

9. N. allionealis Walker, race itealis Walker. Fore wing white to luteous, with fine fragmentary black markings; hind wing white or slightly shaded with lute-

ous; with from two to four fine irregular parallel black lines, the outermost when present, parallel to outer margin. 14-18 mm. The typical form, from Florida. is much darker.

The caterpillar is unknown. The moth is near to the European N. stratiotata, which eats submerged plants and has one more filament in the posterior subdorsal gills, than in the infrastigmatals.

Massachusetts to Florida.

51. ELOPHILA Hübner: Tentamen

(Cataclysta Hübner, Chrysendeton Grote, Anydraula Meyrick)

Labial palpi about as in Nymphula; maxillaries more slender, tapering to a point (fig. 349); antennæ annulated; ocellus obscure or absent. Fore wing (fig. 347) subfalcate, with oblique sinuous outer margin; \mathbf{R}_2 always stalked in the American species (free in group Anydraula). Hind wing with Sc and R fused halfway to apex; M_2 free and normal or obsolete (not fused with M_3 , as the position of the spots shows). Hind wing always with a marginal chain of metallic and black spots, from apex to Cu₂ in our species. Caterpillar typically an air breather; in an ovate case of Lemna leaves; feeding on Lemna; cylindrical and moniliform; with head yellow, paler than the grayish body; with spiracles functional, and those of third to fifth segments of abdomen enlarged. Larva of E. fulicalis under a silk web, on the surface of stones in swift-flowing brooks; feeding on the diatoms and other microscopic forms, growing in the neighborhood of its web or caught in it; with simple tracheal gills, and breathing water. Pupa in E. fulicalis in the web, which is strengthened, and whose openings are arranged to catch the air bubbles carried down by the water.

Key to the species

- 1. Hind wings with a black-dusted area beyond cell, and no lines.
 - 2. Fore wing with a crescentic submarginal spot, not reaching apex, and two
 - or three rounded spots......2. medicinalis. 2. Fore wing with a submarginal streak broadening toward costa, without contrasting rounded spots.
 - 3. Black spots of the terminal series on hind wing opposite each other in pairs, between veinsl. magnificalis.
 - 3. Black spots alternating, the inner series on the veins......4. fulicalis.

1. M₂ preserved; larva air-breathing; on Lemna (Elophila).

1. E. magnificalis Hübner. Fore wing silver white, with light gray-brown lines; costal region shaded with gray; with a waved diffuse antemedial band and irregular postmedial lines, leaving a rounded or quadrate white area on middle of inner margin; three outer gray streaks, converging to a gray patch at anal angle. Hind wing with a gray antemedial shade and bar across cell; postmedial region broadly dusted with black-tipped scales; five marginal ocellate spots between veins, enclosed with yellow, and with a gray marginal streak below the last one. 18-23 mm. (lamialis Walker, helopialis Clemens.)

July.

Quebec to Pennsylvania. "New York" (Grote). 2. E. medicinalis Grote. Brownish gray; a large obliquely oval antemedial patch, resting on inner margin; a more rounded oval median patch, its apex resting on inner margin, frequently with a small white streak beyond it on inner margin; a white postmedial streak from costa at two-thirds, almost to anal angle; and a crescentic submarginal white streak, narrowing more or less to costa, and usually not reaching it; a yellow marginal stripe, edged with gray, and sometimes other yellow streaks and shades between the white spots. Hind wing white on outer half, fuscous on basal half, the boundary a straight perpendicular brown band followed by a lead-colored line; postmedial region irregularly black-dusted. Ocellate marginal spots confluent, with excentric blue pupils, lying below the veins; preceded by a white band and followed by a series of bright yellow bars. 12 mm. (claudialis auct., not Walker.)

July.

District of Columbia to Illinois and Missouri, and south.

II. M₂ lost, cell longer (fig. 347); larva in a web on submerged stones; with tracheal gills (Argyractis Hampson, in part).

3. E. bifascialis Robinson. White, lightly dusted with black; base of costa darkshaded; a broad dark vertical fascia just before middle, followed by a parallel dark line; apical region with three broad dark fasciæ, tapering and converging to anal angle, the first broader, darker, irregular, and bisected by an oblique white or silver crescent. Markings sometimes almost wholly yellow, sometimes gray-brown, with more or less yellow scaling, especially in the two outermost bands. Hind wing with a broad transverse lead-gray fascia in the middle, ending abruptly at the fold, preceded by a shorter yellow fascia, and the latter usually by a lead-gray bar; an oblique black bar halfway between cell and apex; in the variety kearfottalis Dyar, divided into two strize. A marginal waved lead-colored line, partly broken into spots, preceded and followed by alternating black spots; the inner on the veins and much the larger, the outer alternating with yellow bars. 12–15 mm.

August and September. The variety is only known from the Southwest.

New York to Wisconsin and south. New York: Waddington, Honeoye Falls, Ithaca.

4. E. fulicalis Clemens. Fore wing gray-brown; an antemedial shade, oblique inward, and a median line parallel to it; a short white bar before anal angle; a pale shade in end of cell; a white perpendicular line from costa at two-thirds, and a similar subterminal line, converging to a point above anal angle; a bright yellow terminal line, edged with brown. Median area more or less powdery. Hind wing white with a broad curved blackish antemedial band, followed by a finer line parallel to its outer edge; postmedial region broadly dusted with black scales; marginal markings as in *E. bifascialis*, preceded by a fine line of black scales. 10-18 mm. (angulatalis Lederer, confusalis Walker).

Larva on rocks, as described under the genus; moth in June and early July. Common and general in distribution. New York: Peru, Waddington, Lancaster, Niagara Falls, Jamestown, Honeoye Falls, Ithaca.

There is considerable variation in detail, and more than one species may be included. The names *fulicalis* and *angulatalis* apparently apply to large, rather pale and evenly brownish-gray specimens, with a continuous fine black line before the marginal lunules of the hind wing; and *confusalis*, to a small form with much yellow shading, a fine black line on the hind wing, widely broken in the middle, and a straighter dark median fascia.

52. DIATHRAUSTA Lederer

Palpi porrect, fairly long, triangularly scaled (fig. 351); maxillary palpi triangular, hind wing with one median absent; otherwise like Nymphula.

1. D. reconditalis Walker. Brown-black; faint yellowish antemedial and sinuous postmedial lines (white on lower half of hind wing) apparently running directly from the discal dot to above the anal angle; the reniform, orbicular, and claviform, and the discul dot of the hind wing represented by quadrate hyaline white

spots. Fringe with a dark line near base, dark-shaded below middle and at apex. 15-18 mm.

June and July; September.

General. New York: Niagara Falls, Ithaca (aberration with R₂ free), Albany, Pearl River.

2. D. dæckealis Haimbach. Similar to D. reconditalis; differing in the unbroken median line of the hind wing. Black; a faint white line from costa to middle of wing at three-fourths way to apex, and another evenly curved line across both wings; a large white spot in fringe of fore wing. 13 mm. June.

Brown's Mills, New Jersey.

53. GESHNA Dvar

Our species (which belongs doubtfully to this genus), with palpi oblique, shorter than those of the other Nymphulinæ, the third joint well set off and rather blunt (fig. 350); tongue much thicker than palpi; maxillary palpi large and triangularly scaled; fore wing with \mathbf{R}_s free. Outer margin about as in Elophila. (Typically with \mathbf{R}_s stalked, and the outer margin rounded.) 1. G. (?) primordialis Dyar. Fuscous, with the exact pattern of *Blepharomastix* stenialis, but usually without any yellow, or with very little; distinguishable, at last resort, only by the stalking of \mathbf{R}_2 . 10-15 mm. Very common in damp places in June and early July. Generally distributed. New York: Vandalia, Crosby (Yates County), Ithaca, Big Indian Valley.

54. EURRHYPARA Hübner

E. urticata Linnæus (Palpita hortulata), a black and white European species with yellow thorax; has become established at MacNab's Island, and at Truro, Nova Scotia. The larva is found on nettle.

Subfamily SCOPARIINÆ

Similar to the Pyraustinæ, but with labial palpi porrect and beak-like, and maxillaries large and triangularly scaled, as in the Crambinæ (fig. 354). Fore wing (figs. 352, 353) with more or less distinct raised scaling; with R₃₊₄, R₅, and \mathbf{M}_1 more widely spaced at origin than usual in the Pyraustinæ.

55. SCOPARIA Haworth

Front flat; antennæ cilate, more or less distinctly annulate; fore wing (figs. 353, 354) oblong with short outer margin, \mathbf{R}_s divergent; hind wing with Sc and R fused, but often for a very short distance, R and M_1 shortly stalked, M_2 stalked. Slender forms with moderately long legs.

Caterpillars (fig. 355) not well known; the group Eudoria found under moss, etc., on trunks of trees; the typical group apparently in roots of Compositæ.

Key to the species

1. Expanse over 15 mm., typically over 25 mm. Fore wing dark, sometimes shaded1. centuriella. with gray....

1. Expanse under 15 mm.

2. Wings light powdery ash gray, with contrasting dark markings.

3. Orbicular, claviform, and reniform represented by longitudinal black bars.

5. strigalis.

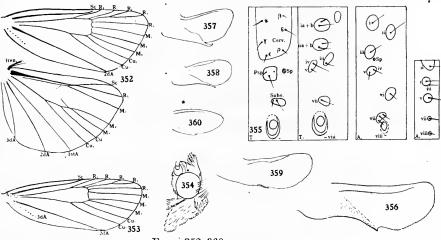
WILLIAM T. M. FORBES

- 3. Reniform hourglass-shaped, or broken into two wedges; rarely, reduced to a dot at lower angle of cell.
 - 4. With a broad, contrasting paler median area.

4. cinercomedia, 7. truncatalis. 4. Median area less contrastingly pale (not at all pale in male).

I. Ventral edge of valve not thickened, but bearing a pointed projection near middle; uncus tapering, with blunt apex (Scoparona Chapman).

1. S. centuriella Schiffermüller (fig. 352, 356). Varying from light ash gray to nearly black; lines dark, diffuse; the outer one convex opposite cell, pale edged;



FIGS. 352-360. SCOPARIINÆ

352, Scoparia centuriella, venation; 353, S. basalis, venation; 354, S. basalis, side view of head; 355, S. truncicolella (Europe), seta map of larva; 356, S. centuriella, left valve of male, side view; 357, S. penumbralis, left valve of male, side view; 358, S. basalis, left valve of male, side view; 359, S. cinereomedia, left valve of male, side view; 360, S. lugubralis, left valve of male, side view. (These five figures are drawn to the same scale from average-sized specimens of the species.)

the inner line often indented on costa; claviform black, linear; orbicular linear or a narrow ellipse; reniform irregular, or broken in two. Dark subterminal shades toward costa and inner margin, sometimes narrowly connected. Hind wing clay color, with blackish border and veins. Female usually darker, sometimes wholly blackish, with obscure markings. 17-27 mm.

Common locally. End of June and July. Caterpillar unknown.

Massachusetts and north, southward in the Rocky Mountains to Arizona. New York: Plattsburg, Peru, Newcomb, Essex County, Saranac Inn, Fentons (Lewis County), Rochester, Buffalo, Ithaca, New Windsor.

II. Ventral edge of valve with a strong chitinized ridge, ending in a heavy spine; uncus broad, membranous, and hoodlike, but pointed; larvæ apparently root feeders (Scoparia).

2. S. penumbralis Dyar (fig. 357). Dull powdery gray (black on elay color), the markings formed of the black powdering, diffuse, and evanescent under a lens; antemedial line pale, followed by dark, transverse, well out; postmedial line slightly dentate on veins, excurved opposite lower angle of cell; followed by a distinct pale line, and a vague darker subterminal shade. Reniform a blackish X; orbicular and elaviform obsolete; blackish terminal dots, alternating with pale ones. 12-15 mm.

Ontario to Pennsylvania. New York: North Creek, Rock City, Vandalia, McLean.

III. Valve simple with a slight and slender ventral thickening, usually ending in a less prominent spine, somewhat trapezoidal; uncus membranous, hoodlike, with a blunt, and sometimes bifid apex; larvæ moss feeders (Eudoria Chapman).

3. S. basalis Walker (figs. 353, 358). Powdery ash gray, on white; a blackish basal dash; a strongly excurved antemedial line, with a pale shade before it, and an irregularly developed darker shade beyond; claviform a black dash; orbicular a short bar, separated from the antemedial line by its length; reniform a black cross, sometimes imperfect, more or less shaded with brown; postmedial line strongly and sharply excurved opposite lower angle of cell, almost angled, pale, defined with gray shades, especially on outer side at costa and inner margin; a gray shade along middle of outer margin. Hind wing grayish white. Male with a short small valve, convex on its dorsal side, with a weak, but distinct, spine on its ventral edge, rather near its apex than its base $(\frac{1}{4}$ mm, from apex). 11-16 mm.

June to August.

Generally distributed; common on tree trunks. New York: Niagara Falls, Lewiston, Hemlock Lake, Rock City (Cattaraugus County), Otto, Crosby, Ithaca, Big Indian Valley, Rhinebeck, New Windsor, New York City. I have also unverified records from Essex County, Old Forge, Newport, Fentons (Lewis County), Buffalo, Wells, and Albany, most of which are doubtless correct.

4. S. cinereomedia Dyar (fig. 359). Powdery ash gray without any brown; the median area, except beyond the reniform, more or less distinctly paler, sometimes white. Lines as in *S. basalis*, the postmedial perhaps a little less irregular. Ante-medial line with its dark outer shade more nearly continuous; the dash in the fold rarely distinct. Male with valve much longer than in *S. basalis*; concave on the upper edge, with the spine weaker, and much further from the apex (nearly $\frac{1}{2}$ mm). 15 mm.

Found with S. basalis, in July.

New York and western Pennsylvania. New York: Otto, Ithaca.

5. S. strigalis Dyar. Light powdery gray, much like male basalis; orbicular, reniform, and claviform nearly equally large, heavy black bars, the orbicular touching the obscure antemedial line, and the reniform a little thicker; a series of black terminal bars, those at apex, opposite cell, and in fold, much heavier; basal dash long. Genitalia not studied. 10-17 mm.

July and August.

Ontario and southern New Hampshire to Georgia. New York: Ithaca.

6. S. lugubralis Walker (fig. 360). Very close to S. basalis. Outer dark shade of antemedial line continuous, and entirely separate from the rounded claviform spot. Frequently a black bar just above Cu, connecting orbicular and reniform. Postmedial line less deeply angled, but just as sharply. Male with valves long and slender, without a distinct spine on ventral edge. 18 mm.

June to early August.

Bretton Woods, New Hampshire; Canada; Western States.

June.

7. S. truncatalis McDunnough. Similar to S. basalis, but without any brown markings, the scaling wholly black and white. Claspers much broader than in S. basalis. 15 mm.

July.

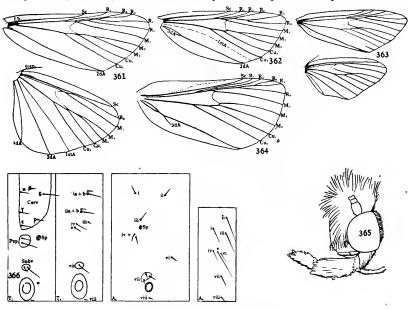
Norway Point, Lake of Bays, Ontario.

This form is obviously distinct from *basalis*, but not distinguishable (on the basis of the single specimen before me) from dark specimens of *S. cinereomedia*, with which McDunnough does not compare it.

There are probably several other undescribed species of this genus in our territory, but they need a full study of the structure and variation, to be worked out.

Subfamily PYRALIDINÆ

Antennæ rarely with modification; ccelli variable, normally concealed or absent; antennal socket rarely separated from eye by a line of scales; front nearly smooth (fig. 365); maxillary palpi good sized, but not triangular, and well developed even when tongue is rudimentary; tongue variable; fore wing (fig. 361) with \mathbb{R}_5 stalked with \mathbb{R}_4 and \mathbb{R}_4 ; \mathbb{R}_2 free; 1st A lost, but represented by a wider space between $\mathbb{C}u_4$



FIGS. 361-366. PYRALIDINÆ

361, Pyralis farinalis, venation; 362, Hypsopygia costalis, venation of fore wing; 363, Aglossa pinguinalis (Europe), venation; 364. Omphalocera dentosa, venation and costal fold of male; 365, Aglossa cuprealis, side view of head; 366, Cledeobia angustalis (Europe), seta map of larva.

and 2d A; 3d A usually obsolete, becoming coincident with 2d A in Hypsopygia (fig. 362). Hind wing without a fringe on base on Cu; Sc and R parallel, sometimes no closer beyond the cell than along it.

Larva essentially as in the Phycitinæ (fig. 366); pupa with a dorsal furrow between ninth and tenth segments of abdomen, with crenulate margin; thoracic spiracles slit-like and inconspicuous; end of abdomen with a transverse row of six or eight hooked setæ, without a specialized cremaster.

Key to the genera

1.	Palpi upturned.
	2. Tongue very weak (about as long as eye or shorter)
	2. Tongue well developed.
	3. R ₃ and R ₄ stalked farthest
	3. \mathbf{R}_{\bullet} and \mathbf{R}_{\bullet} stalked farthest
1.	Palpi obliquely upturned, with porrect third joint
1.	Palpi long, porrect, beaklike

56. PYRALIS Linnæus

Tongue strong, scaled at base; palpi typically upturned to vertex; antennæ typically with short ciliation (shorter than length of segments); fore wing normal (fig. 361), M_2 and M_3 stalked; hind wing with M_2 and M_3 stalked; Sc and R typically very closely parallel.

The second group is transitional to Aglossa.

Key to the species of Pyralis and Aglossa

1. Fore wing evenly colored, with white ante- and postmedial lines only.

P. 1. farinalis.

- Fore wing more or less mottled; no pure white; usually with discal spots.
 With a blackish discal spot, and usually costal spots at base and before apex.
 - 3. Basal blackish spot broad, usually reaching fold; subapical spot strong, dorsal two-thirds of wing contrasting, pale.....P. 2. costiferalis.

 - patch usually prominent.....P. 3. disciferalis. 2. Ground dark with paler markings; discal spot pale when distinct.
 - Ground pinkish coppery; antemedial tooth on fold strong, and usually filled with blackish; reniform usually a broad, pale, horizontal ellipse, strongly contrasting; tongue as long as eye.....A. 1. cuprima.
 Usually without red tint; blackish, with confused luteous markings only;

1. P. farinalis Linnæus. Brown, somewhat olivaceous when fresh; median area paler, twice as wide as basal and terminal areas, the latter shaded with gray. Lines white, defined, the antemedial strongly excurved; postmedial concave on upper two-fifths and in the fold; curved far out between. Terminal line pale; fringe grayer. Hind wing with obscure antemedial, and fine, clear postmedial, lines on a ground shading from white to fuscous gray. 15-25 mm. (H 47:53.) Larva injurious to meal etc. Dirty white, with a dark brown head; hooks on

Larva injurious to meal etc. Dirty white, with a dark brown head; hooks on prolegs biordinal, the shorter ones a quarter as long as the longer ones; prothorax with posterior setæ on cervical shield approximately in a vertical line, but with the uppermost well in front of the others (nearer to them than to the anterior series); front extending halfway, and adfrontals three-fourths way to vertex (unlike Plodia); the three ventral ocelli in a right triangle.

Common and generally distributed. New York: Generally distributed and not rare.

2. P. costiferalis Walker. Clay color, shaded and dusted with dull rose; with contrasting blackish base (above fold), patch on cell, and subapieal costal patch; postmedial line excurved and serrate, diffuse, clay color; often defined on inner side with a blackish shade. Antemedial line similar, zigzag, or obscure. Hind wing pale, with a faint darker postmedial and rarely a subterminal line. 15-24 mm.

July to August.

In this and the next species, the antennal eiliations are longer than the segments, the tongue is very weak and half as long as the thorax, and Sc and R are farther apart in the hind wing. These species approach Aglossa.

July.

Ontario to Pennsylvania. New York: Saranac Inn, Ithaca, Big Indian Valley, Albany.

3. P. disciferalis Dyar. Shining dull rose, much darker than P. costiferalis. Costa barred with black on basal two-thirds, heavily toward base. Lines broad, pale, diffuse, heavily edged toward median area with blackish; the antemedial line zigzag and the postmedial excurved opposite the cell and concave toward the costa and in the fold. Hind wing much as in P. costiferalis, but darker, with a pinker costa. 15 mm.

July.

Maryland to Missouri. New York: Ithaca.

57. AGLOSSA Latreille

(Grease moths)

Similar to Pyralis, and only differing from the latter group of Pyralis in the weaker tongue (fig. 365). Antennal ciliation longer than the segments, fasciculate; tongue shorter than eye; M_2 and M_3 sometimes approximate only (fig. 363), but stalked in our species; Sc and R well separated beyond cell in hind wing; scaling shining, greasy looking. 1. A. cuprina Zeller. Tongue coiled and as long as eye. Clay color, heavily

shaded with dull rose; the median area, and frequently the base and outer margin, shaded with dark gray. Antemedial line zigzag and postmedial line strongly dentate, both edged with blackish toward the median area; tooth on antemedial line on fold often strongly contrasting, filled with a blackish patch, and capped with a blackish arrow-head mark. Reniform horizontally oval, often with a diffuse black spot beyond it. Hind wing luteous, nearly immaculate. 18-23 mm.

June to September.

Generally distributed. New York: Ithaca, Rhinebeck.

2. A. cuprealis Hübner. Tongue reduced to two scaly lobes; fore wings dark luteous gray, marked with luteous, with little or no pinkish shading; lines as in A. cuprina; the antemedial, on the average, less deeply dentate, not defined with darker; reniform irregularly invaded by the dark ground, or obsolete. 18 mm.

Caterpillar gray, with brown head; a scavenger. General; also in Europe. New York: Mt. Marcy (Hill), Ithaca, Poughkeepsie (New York State Collection), New Windsor (Morton), Brooklyn.

58. HYPSOPYGIA Hübner

(Pyralis, in part)

Like the typical group of Pyralis, except for the longer stalking of R_4 and R_5 (fig. 362); 3d A preserved, running into 2d A. Caterpillar as in *Pyralis farinalis*. 1. H. costalis Fabricius (Clover-hay worm). Light dull rose; fringe very broad and pale yellow; lines rather fine, pale yellow, enlarged into large patches on costa of fore wing. 13-18 mm.

Very common, and generally distributed, sometimes injurious to stored hay. May to September.

New York: Ogdensburg, Lewiston, Sherwood (Cayuga County), Ithaca, Big Indian Valley, Onteora Mountain, vicinity of Albany, Rhinebeck, North Hillsdale and New Windsor.

Variety hyllalis Walker has the ground pale yellowish, the lines almost lost.

59. HERCULIA

(Pyralis, in part)

Similar to typical Pyralis, except as noted in the key; M2 and M3 free; 3d A preserved.

Key to the species

Outer line finely dentate, dark, far out.....l. intermedialis. Outer line even, nearly straight, often obscure.

1. H. intermedialis Walker. Wings narrower, suggesting Aglossa. Dull rcdbrown; terminal space distinctly redder and paler. Base pale; antemedial line strongly irregular, toothed far out on Cu, and less on A; pale followed with black; postmedial four-fifths way out, black, offset in to costa, and followed by a dis-tinct pale patch; below, narrowly pale shaded. Hind wing paler, graver, with the faint darker inner and outer line widely separated, and the outer nearly even. 22 mm. (cohortalis Grote.)

June and July.

Generally distributed. New York: Mt. Marcy, Fentons (Lewis County), vicinity of Buffalo, Ithaca, Hastings, New Windsor, Katonah.

2. H. olinalis Guenée. Dark gray with more or less violet tinge, and pinkish fringe, tipped with yellow; marked like H. costalis, but the lines straighter, and often obscure below the costa. Outer line on hind wing hardly beyond middle of wing. The ground sometimes much paler, rose pink (*himonialis* of authors, but not of Zeller, who had the typical form). 22 mm. (H 47:46; 48:13.)

June and July.

New York: Lancaster, Peru, Ithaca, Big Indian Valley, Schenectady, New Windsor, New York City; Clove Valley, Staten Island.

Variety infimbrialis Dyar is ochreous with only a slight pinkish tinge, almost as light as *H. thymetusalis*. It occurr in southern Massachusetts, and is probably somewhat general in distribution.

3. H. thymetusalis Walker. Simil r to H. olinalis; pale luteous, dusted and shaded with dull light gray, emphasi ing the luteous lines on the side toward the median area, where there are distinct gray lines. Lines slightly dentate; antemedial excurved, broad; postmedial sometimes shortly interrupted, starting from a triangular costal patch. Hind wing paler, the lines close together; antemedial slightly irregular, and postmedial excurved. 22 mm.

Perhaps a variety of \hat{H} . olinalis, with which it seems to intergrade.

A northern species, ranging south to the Adirondacks. New York: Saranac Inn, Mt. Marcy, Fentons (Lewis County), Albany, New Windsor.

60. OMPHALOCERA Lederer

Palpi beaklike, clavate, the second joint long, straight and blade-like, the third long and porrect, sharply marked off. Antennæ simple, with scape long and modified. Abdomen tufted dorsally. Fore wings triangular (fig. 364), moderate,

broad at base. R_3 , R_4 , and R_6 forking at nearly the same point; M_2 and M_3 free and well separated in both wings. Moths large and Noetuid-like in appearance.

Caterpillar exactly as in the Phycitinæ.

An aberrant genus, perhaps nearer to the Epipaschiinæ and lower Phycids than to the true Pyralinæ.

1. O. cariosa Lederer. Dull dark red brown; base paler, and median area usually larker, or even blackish; usually concolorous to the inner margin; bounded by a rather evenly excurved antemedial line, whose upper part is oblique outward, and by a sinuous postmedial. Terminal space practically even, without the streaks and wedges of O. dentosa. Hind wing darker and more fuscous. 30 mm.

Larva a borer in fruit of papaw; blackish, with a broad red band on dorsum, and a lateral red band, separated by an equally wide black subdorsal area from the dorsal; red bands mottled with brown; tubercles white.

District of Columbia, southern Illinois, and southward.

2. O. dentosa Grote. Clay color, dusted and streaked outwardly with dark redbrown; a large, nearly square, dark-brown patch on middle of costa, extending halfway across the wing, shaded with black, and running out into teeth on the veins. Ordinary lines obscure, bordering the patch. Hind wing blackish, redder at margin. 35-40 mm.

June. Caterpillar black, dotted with white; without red bands; on barberry. Southern Connecticut to Ohio, Iowa, and south.

Subfamily ANCYLOLOMIINÆ

Similar to the Crambinæ. Fore wing very frequently notched at the middle, and twisted in the resting position. Hind wing (fig. 367) in female with a simple frenulum; cell closed by a weak but distinct, middle discocellular vein; \mathbf{M}_1 more or less weakened; widely separate from R at origin. Basal portion of Sc and R also widely separated in some cases.

This is a primitive type, related to the common stem of the Crambinæ and Phycitinæ. There are only a few forms in all, the majority in the Old World. The early stages are but little known.

61. PRIONAPTERYX Stephens

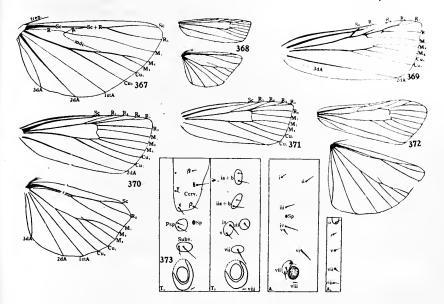
Front slightly conical, ocelli absent; male antennæ serrate; palpi about twice as long as head; tongue well developed. Fore wing with apex rectangular, abruptly offset inward at middle of outer margin, between M2 and M3, and crumpled at that point in the resting position. \mathbf{R}_1 becoming coincident with Sc in our species. \mathbf{R}_6 connate or short-stalked with \mathbf{M}_1 in our species, but completely fused with it in the western P. achatina; M_2 and M_3 short stalked. Hind wing with Sc and R moderately anastomosing; the upper discocellular erect, middle discocellular complete, but rather weak, and oblique outward, but not strongly angulate, M_2 and M₃ fused in our species.

1. P. nebulifera Stephens. White, base blackish, costa shaded with fuscous; median area blackish, crossed by the zigzag white median line; subterminal line fine, white, defined on outer side with a fine dark line, and on inner with a blackish shade, with a tooth opposite cell; waved and concave below. Apex white; with a blackish ray running to the tooth of the outer margin, and with a black bar below it. Terminal line fuscous, and fringe pale, except on the tooth. Hind wing pale gray. 25 mm. May to August; October.

The larva lives on the ground and forms a tunnel of sand and silk extending

up the stem of its food-plants to near where it feeds. It may be found on various sand-barren Ericaceæ.

Massachusetts to Texas, along the Coast. New York: Karner, Yaphank.



FIGS. 367-373. ANCYLOLOMIINÆ AND CRAMBINÆ

367, Eufernaldia cadarella (Ancylolomiinæ), venation of hind wing (Western States); 368, Crambus girardellus, venation; 369, C. luteolellus, venation of fore wing; 370, Argyria nivalis, venation; 371, Platytes multilineatella, venation of fore wing; 372, Ommatopteryx ocelleus (Texas), venation; 373, Crambus species, seta map of larva

Mesolia incertella Zincken reaches north to North Carolina. It has the same notched wing and a rather similar pattern; but a more conical front, and the subcostal cell of the hind wing nearly as wide as the discal $-M_1$ arising opposite the tip of the base of R, so that the upper discocellular is almost obliterated, but well separated from the stalk of Sc+R.

Subfamily CRAMBINÆ

Antennæ simple or pectinate, without modification at base; their sockets separated from the eyes by a band of scales. Ocelli most often present. Tongue variable; labial palpi beaklike, porrect; maxillary palpi large and triangularly dilated with scales. Tibiæ with all spurs. Fore wing (figs. 368 to 372) narrow, except in Argyria; with short outer margin; usually with all veins preserved; **R**, often anastomosing with Sc, \mathbf{R}_{4} and \mathbf{R}_{4} stalked; and \mathbf{R}_{2} and \mathbf{R}_{3} almost always stalked with them; ist A completely lost, the distance between 2d A and Cu_z at the margin being hardly wider than between any two veins. 3d A free, weak. One or two radials or one median rarely lost. Hind wing ample, much folded;

Sc and R strongly anastomosing beyond the cell; female frenulum multiple, cell open, M_2 most often stalked, sometimes lost; a heavy fringe of simple bristles along base of Cu. Wings always smooth-scaled. Larvæ and pupæ of two radically different types, as described under Crambus and Diatræa. The subfamily is fairly homogeneous.

Key to the genera

1. Fore wing with all radials preserved; \mathbf{R}_1 sometimes very short and becom-
ing coincident with Sc.
2. R ₂₋₄ stalked64. Eoreuma.
2. R _{s-5} stalked.
3. Male antennæ uni- or bipectinate
3. Male antennæ laminate, simple
2. R_s and R_4 only stalked.
3. R_1 becoming coincident, or anastomosing, or connected by a crossvein
with Sc.
4. Palpi twice as long as head; front more or less conical66. Argyria.
4. Palpi much longer.
5. Ocelli present
5. Ocelli absent
3. \mathbf{R}_1 free.
4. Ocelli absent
4. Ocelli present.
5. Front conical
5. Front rounded.
6. Palpus projecting its length beyond head
6. Palpus projecting twice its length beyond head65. Platytes.
1. Fore wing with two radials and a median lost

62. DIATRÆA Guilding

Palpi beaklike, extending two or three times the length of the head; antennæ nearly simple, ocelli absent. Front usually conical. Body normally rather stout. Fore wing moderately broad; R_1 typically becoming coincident with Sc; R_3 and R_4 stalked; the rest free. Hind wing somewhat broader, with Sc and R rather strongly anastomosed; subcostal cell narrow, normal; M_1 from upper angle of discal cell, and M_2 and M_3 connate.

Caterpillars, so far as known, borers in Gramineæ, and most commonly in wet places, slender, with strong tubercles; prolegs with an ellipse of triordinal hooks; vii with two set on meso- and metathorax, as in the Galleriinx; set wi and v of abdomen obliquely placed, i and iii of ninth segment of abdomen approximate, and ii of eighth and ninth segments forming single dorsal plates. Pupa usually cylindrical, truncate at anterior end; tongue short.

The first group is ancestral to both Chilo and Diatræa, and the species have been variously treated in the past.

Key to the species

1. Wings narrower; fold with alternating pale and dark shades.

2. Hind wing white.....2. forbesellus. 2. Hind wing clay-color or brown.....l. comptulalis.

1. Wings broader; fold evenly colored or streaked longitudinally.

2. Fore wing mouse gray, with traces of postmedial dots or immaculate.

3. idalis.

2. Fore wing yellow or brown.

3.	Fore	wing	with	a terminal	l dark	line	4.	alleni.
3.	Fore	wing	with	dark tern	inal d	lots	5. zea	colella.

I. Sc and R_1 merely approximate.

1. D comptulalis Hulst. Male dull brown, with grayer margin; a diffuse pale crenate and broken antemedial line; a finely dentate postmedial line roughly parallel to outer margin, toothed inward on A, and running in to costa, and a pale discal spot. Hind wing fuscous. Female light clay color, shaded lightly with brownish, leaving the whole cell pale, but with dark patches in the base and middle of the fold, and near the middle of the inner margin. Discal dot dark. A diffuse pale subterminal line, preceded by a well-marked dark band, defined on the outer side only. A slight, dark terminal shade. Hind wing slightly paler, with a corresponding terminal shade. 25 mm. (Chilo auct.) Michigan to Missouri and west. New York: Ithaca.

2. D. forbesellus Fernald. Similar to D. comptulatis. Fore wing with more or less white scaling on disc, setting off the black discal dot; a streak of white scales in fold, interrupted by black bars near base and middle of fold; subterminal markings and terminal dots obscure. Hind wing white, in the male light fuscous apically, and with a distinct dark terminal line; in the female almost immaculate. Fore wings in some specimens much paler, light ochre yellow. 23-38 mm. (Chilo auct.)

Larva a borer in Scirpus.

Illinois; Missouri; New York: Buffalo.

3. D. idalis Fernald. Fore wing gray, obscurely or not striate, with traces of markings or none; hind wing white in female, pale gray in male. 25-34 mm. New Jersey; Georgia.

4. D. alleni Fernald. Clay-color to brown, finely dark-streaked on and between the veins, the venular streaks even in width. A distinct discal dot. Terminal line obsolescent, but not breaking into dots. Inner margin sometimes contrastingly pale. Hind wing fuscous outwardly. 30 mm.

Maine. New York: Newport, Big Indian Valley.

II. Sc and \mathbf{R}_1 anastomosing.

5. D. zeacolella Dyar. Clay color, finely streaked with dark on and between the veins; the venular streaks enlarging to form a very oblique series of postmedial dashes, and a subterminal series which is very oblique series of postmetial angle of cell, and slight terminal dots in interspaces; slight dark shades in upper part of cell, below cell, and extending obliquely to apex. Female with markings nearly lost. Hind wing white. 25-30 mm. (saccharalis auct., not Fabricius.) Larva a borer in corn (not normally in cane); sometimes injurious.

Virginia and south.

63. CHILO Zincken

Similar to Diatræa. Ocelli prominent, front always conical. R, closely approxi-

mate to Sc for a distance, but free. Larvæ borers in water plants. 1. C. plejadellus Zincken. Whitish yellow: fringe, a broken line before it. a short, irregular, oblique median band, and scattered scales on disc, all lead-colored or silvery; black terminal dots. Hind wing white. 22-32 mm.

July.

Larva a borer in stems of rice, and presumably other swamp grasses; yellowish white, head dark brown. cervical shield light brown, body with four vague purplish stripes. Pupa with conical front.

New York to Wisconsin and Georgia. New York: Ithaca.

64. EOREUMA Ely

(Chilo, in part)

Similar to Diatræa and Chilo. Ocelli present; palpi as long as head and thorax, tongue weak. \mathbf{R}_2 stalked with \mathbf{R}_{2+4} , instead of approximate; front only slightly rounded out.

1. E. densella Zeller. Yellowish, dusted with brown, veins white, and with clearer yellow line halfway between the veins, alternating with brown lines; a strong black discal dot; black terminal dots and dark streaks opposite the cell and in the fold. Hind wing nearly white. 15-18 mm.

June and July; September. Connecticut to Florida, Illinois, and Texas.

65. PLATYTES Guenée

(Chilo, in part)

Similar to Argyria; fore wing longitudinally marked, and typically narrower (fig. 371), palpus as a long as head and thorax. M_2 and M_3 of hind wing some-

times approximate (varying in our species). The only known larva in moss. 1. P. panalope Dyar. R_2 sometimes partly fused with the stem of R_3 and R_4 Cream color, with contrasting fuscous streaks. Outer side of palpi fuscous; fore wing with fine dark streaks on veins and heavy ones on interspaces, leaving white lines in the middle of the cell and on the fold. Diseal dot black in a white ring. Terminal dots black. Hind wing nearly white. 12-15 mm. (multilineatella auct., not Hulst; densellus Fernald, not Zeller, at least in part).

Early stages unknown. Moth in June.

Southern States: Connecticut.

2. P. vobisne Dyar. Silvery white, with several dark brown lines, those in basal part of wing nearly longitudinal, and two zigzag postmedial and subterminal ones; a dark terminal area, and some dark postmedial shading. Hind wing blackish; fringe white with a dark line. 13 mm.

June; August.

Connecticut; South Dakota.

66. ARGYRIA Hübner

(*Platytes*, in part)

Ocelli present. \mathbf{R}_1 normally straight, and parallel to Sc and \mathbf{R}_2 (fig. 370), \mathbf{R}_3 and \mathbf{R}_{c} only, stalked; \mathbf{M}_{2} and \mathbf{M}_{3} separate. Fore wing twice as long as wide, with moderate, somewhat excurved outer margin. Hind wing but little broader, with Sc and R strongly fused; M_2 and M_3 more or less stalked, often strongly. Front normally nearly flat. Palpi variable, normally projecting beyond head for a dis-tance about equal to its length, rarely, nearly twice its length. Larva unknown, probably much like those of Crambus.

The tiny subtropical species C. lacteella, has been reported from Saranac Inn.

Key to the species

 Fore wing with a yellow median fascia. Inner margin yellow on outer half 	3. auratella.
2. Inner margin white on outer half	4. critica.
1. No yellow fascia.	
2. Inner margin finely edged with yellow2	. argentana.
2. Inner margin white.	
3. Head white	l. nivalis.
3. Head and palpi deep yellow	rileyella.

I. Front nearly flat, rounded; palpi as long as head; tongue stronger; R, free.

1. A. nivalis Drury. Silver white. Palpi yellow on outer side, except on upper edge, sides of occiput yellow; a longitudinal stripe on thorax. Base of costal edge brown, terminal line black; fringe yellow at base, with whitish tips. A black dot at middle of inner margin, rarely lost. 20 mm.

Very common in grass land in June and July. Generally distributed. New York: Newcomb, North Creek, Newport, Locust Lake, Lewiston, Lancaster, Ithaca, Big Indian Valley, Utica, Schenectady, Little Falls, Albany, Rhinebeck, New Windsor, Katonah, New York City.

2. A. argentana Martyn. Silver white; head, palpi, scape, and pedicel wholly brownish yellow; antennæ otherwise gray. Thorax with a yellow central stripe, fore wing with costal edge narrowly yellow, inner edge a little more broadly so, especially at middle; terminal line black, followed by a fringe shading from brown at base to pale yellow at apex. Hind wing straw color. 20 mm,

July.

Southern New Jersey; Pennsylvania; and south.

A. rileyella Dyar ranges north to North Carolina. It has a faint, slightly waved, dark postmedial line.

II. Front distinctly conical; palpi twice as long; tongue weaker; \mathbf{R}_1 anastomosing with Sc.

3. A. auratella Clemens. Silver white; head and palpi white above, golden yellow on sides; thorax yellow, with white tegulæ, except the shoulders, which are yellow. Fore wing with a broad yellow median fascia, widening to inner margin, and fringe yellow, connected to the median fascia by a fine yellow line along outer half of inner margin; terminal dots black. 18 mm.

Julv.

Maine to California and south. New York: Ithaca, Albany, Rhinebeck, Katonah. The southern race, pulchella Walker, is much smaller.

4. A. critica Forbe's. Similar to A. auratella; front much less prominent (onefourth width of eye in side view); fore wing with the median band not widening toward the inner margin, which is white between the median and the yellow terminal band. 18 mm.

July. (April, in Florida.)

Trenton, Ontario, and New Jersey, to Florida. New York: Utica, Ithaca.

67. HAIMBACHIA Dyar

(Crambus, in part)

Similar to Crambus. R anastomosing with Sc, R2 and R5 free, Cu2 widely separated from Cu_1 . Hind wing with M_2 and M_3 connate or very shortly stalked; Sc and R stalked almost to apex.

1. H. placidella Haimbach. Front rounded. Cream white, dusted with blackish out to the subterminal line; a darker pale brown medial shade, angulate on the cell, and a fine, sinuous, double subterminal line reaching almost out to the margin, filled with a slightly paler cream white. Terminal line continuous above, dots below; fringe lead colored. 16 mm.

July.

Northern New Jersey.

A second species of this genus ranges from New Jersev to Florida; it is closely similar to the Texan H. squamulellus Zeller, with a largely white fringe, and with yellow bars in the terminal space, but has a conical front. It appears to be undescribed.

68. THAUMATOPSIS

(Propexus Grote; Crambus, in part)

Closely related to the *mutabilis* group of Crambus, but with antennæ of male pectinate, or at least with slender laminations on the lower side twice as long as the segments. Fore wing normally with a slender longitudinal white streak on Cu, and sometimes on its branches toward margin, unlike all our species of Crambus. R_1 free.

T. repanda is reported from Saranac Inn and Newport, doubtless in error.

Key to the species

1. A contrasting white streak on Cu, the wing more or less darkened above it.
2. Fore wing dark brown
2. Fore wing clay color.
3. A defined black streak running obliquely to apex4. fernaldella.
3. No such streakl. pexella.
1. No white.
2. Mouse gray
2. Yellow

* Male antennæ bipectinate.

1. T. pexella Zeller. Dull clay color, shaded below and outwardly, and somewhat striate with gray. A fine, somewhat sinuous white streak along lower side of cell, to end, fading out at base and rarely obsolete, with a dark, sometimes partly black, shade below it at base, and above at middle; an irregular series of oblique shades from opposite cell to inner margin. continued by a vague shade to apex. Terminal dots fine, dark. Hind wing nearly concolorous. 30 mm.

New Jersey; Connecticut; Illinois, and west. New York: New Windsor (Morton).

2. T. gibsonella Kearfott. Powdery mouse gray, strongly yellowish toward costa and base, with slightly paler Cu, and with traces of the dark shading of the last species. Subterminal markings obsolete, terminal dots minute or lost. Traces of an oblique median shade at inner margin. Hind wing nearly concolorous, with paler base. Female rather narrower-winged, with much paler and slightly yellower fore wing. 26-33 mm.

September.

Ontario.

3. T. edonis Grote. Closely similar to T. gibsonella but light ochreous yellow (when fresh, strongly pinkish). 35 mm.

Nantucket, Massachusetts; North Carolina; Kansas.

** Male antennæ unipectinate.

4. T. fernaldella Kearfott. Similar to *T. pexella*. Brighter yellowish, with obsolete subterminal markings; the black streak above the white line on Cu extending to the subterminal region and separated by a narrow longitudinal pale line from a fine curved oblique streak that runs to the apex. Black streak defined below, and pale edged below outwardly. 25 mm.

A western species reported doubtfully from Anglesea, New Jersey, and from Florida.

5. T. dæckella Kearfott. Fore wing rather dark brown, hind wing blackish gray; a longitudinal whitish streak on Cu, fading out near end of cell. 22 mm. (Crambus haytiellus Hart, not Zincken.)

Lucaston, New Jersey; Havana, Illinois.

69. CRAMBUS Fabricius

Tongue moderate; palpi projecting as far as length of head and thorax; male antennæ simple, laminate below, the laminations usually in contact; rarely more slender (*mutabilis*, etc.), but always more than half as wide as the length of a segment. Front nearly flat. Ocelli present. Fore wing (figs. 368, 369) narrow (normally about two and one-half times as long as wide), R_1 free or anastomosing with Sc; R_2 free; R_5 stalked, in our species shortly, with R_{3+4} ; M_2 and M_3 free or short-stalked. Hind wing much broader, with Sc and R fused about three-fourths way to apex. M_1 from upper angle of cell, M_2 and M_3 almost always stalked.

Caterpillar (fig. 373) usually with good-sized tubercles, vii double on prothorax, single on meso- and metathorax; only a single lateral seta on ninth segment of abdomen; prolegs with an unbroken ellipse of the triordinal hooks.

The caterpillars live mostly in tubes of silk and earth, and feed on the roots and leaves of grass and other low plants. Several species have appeared from time to time in injurious numbers, and many are very common.

Pupa smooth, maxillæ reaching almost to tips of wings, but with the mid-legs meeting behind them; dorsum of thorax and abdomen not spined; sides of last segment with deep longitudinal furrows; labial palpi often largely exposed, normal. Caudal end usually truncate, with blunt angular spines; without a developed cremaster.

Key to the species

- 1. Ground color silver-white (including cell and costa, and outer and inner margins).

 - 2. With black terminal, and usually subterminal, dots only....20. turbatellus.
 - 2. At least with a complete and regular subterminal line.
 - 3. Subterminal evenly excurved; seven terminal dots......19. elegans. 3. Terminal line above, and dots below.
 - 4. Subterminal line slightly curved opposite end of cell. . 16. lyonsellus var. 4. Subterminal line sharply angulate.

1. Terminal space silvery; subterminal line erect, curved; median shade angulate. 19. elegans.

- 1. Inner margin, or terminal area, or both, dark; the ground not white.
 - 2. A fairly broad, white, more or less silvery stripe in cell from base, covering most of the width of the cell.
 - 3. White stripe practically reaching outer margin, but cut into trapezoidal patches by median and subterminal brown bands; the outermost patch almost terminal2. myellus.
 - 3. Postmedial silver trapezoidal patch absent, or not an even continuation with the silver stripe.
 - 4. Silver stripe continuing uninterrupted, and hardly narrowed to outer marginll. unistriatellus.
 - 4. Stripe at least constricted to half its width in subterminal region.
 - 5. Fore wing strongly falcate, with apex drawn out; an oval silver 5. Fore wing rectangular or subfalcate; no oval subapical spot.
 - 6. Terminal space crossed by fine black lines from subterminal line to margin; a strong costal silver streak, well separated from the subcostal one; ground cream color.....l. laqueatellus.

- 6. Terminal spots with black dots or short bars, longest in *C. quin-quareatus*, which has only a single silver streak and dark brown ground.

 - 7. White streak simple, leaving at least the costal edge brown. S. A white or silver patch in middle of wing before the sub
 - terminal line, below or beyond the tip of the streak.
 - 9. Apex forming an angle of 60°, the upper part of the outer margin at an angle to the lower; inner margin white, ground pale and streaked with lead color; brown terminal line and white in base of fringe ending abruptly at the angle of the outer margin; terminal dots marginal.
 - Inner margin pure silver white, with contrasting black edge, outwardly; expanse 28 mm.; lines on veins all lead-gray13. dxckellus.
 - 10. Inner margin powdery and less contrasting, lines on veins a mixture of silver and blackish; expanse 22 mm.

14. floridus.

- 9. Apex more nearly a right angle, except in C. exsiccatus; outer margin even, the blackish terminal line and white base of fringe fading out gradually below.
 - 10. Part of inner margin white, contrasting (sometimes only a small spot in *C. labradoriensis*, which is distinguishable by its very dark ground-color).

 - 11. Ground bright yellow-brown, with a narrow silvery streak on inner margin......7. youngellus.
 - 10. Inner margin at most pale straw yellow, and in that case shading into the darker yellow ground. 4. alboclavellus.
- 8. No such patch; ground continuous to subterminal line.
 - 9. A contrasting pale spot in middle of terminal space; tooth on lower side of silver streak strong, and more than twothirds way out6. bidens.
 - 9. A pale spot below apical dash only, obscure in *C. lyonsellus*; tooth usually three-fifths way out, running into a slender line or obsolescent.
 - Silver patch only covering a little of costal region (above R), toward base.

8. præfectellus.

- 10. Silver stripe broader, toward the base covering all but . the costal edge, and covering \mathbf{R} to end of cell.
 - 11. Ground cream and yellow; silver streak lying along costa three-fifths way to apex.....16. lyonsellus.
 - 11. Ground dark brown; streak gradually diverging from costa.

12. A silver stripe on inner margin; apex more acute.

10. quinquareatus.

12. Inner margin concolorous, brown; apex blunter.

- - 3. Terminal line continuous on costal half of wing, contrasting with the yellow terminal space, represented by three black dots below.

27. hortuellus.

- Terminal line all broken into dots, or obscure.
 Eyes less than one-half as wide as front; palpi hairy; a heavy oblique
 - - fascia, or none.
 - 5. Fringe bright golden bronze; six or seven black terminal dots.
 - 5. Fringe dull, or dark lead-gray, rarely shining.

 - 6. Subterminal line irregular, diffuse, and much more oblique than outer margin, or obsolete.
 - 7. A distinct blackish spot or oblique streak in fold below end of cell; a longitudinal pale shade through middle of wing.
 - .8. Male antennæ with laminations narrow, separate; fore wing with six or seven terminal dots; pale shade whitish.

26. mutabilis.

- 7. Median shade but little darker than ground, or obsolete; no discolorous longitudinal shade through cell.

 - 8. Terminal line very weak and continuous, or enlarged into slight dots on all the veins.

 - 9. Ground fuscous, sometimes heavily dusted with fuscous on a dirty white ground......30. caliginosellus, 31. zeellus.

Thaumatopsis gibsonella will run to near lutcolellus, but the subterminal line is obsolete, edonis may be distinguished by its large size (35 mm.), also.

I. Palpi moderately hairy, eyes about as wide as front (Crambus).

A. Fore wing with Sc and R_1 separate, normally approximate (fig. 368).

* Fore wing with a silvery stripe.

+ Fore wing normal in form.

1. C. laqueatellus Clemens. Straw color, costal and cellular silver stripes rather widely separated; a large triangular costal subterminal streak. More or less silver on veins, edged with black; terminal space pale, with five fine

black lines across it. Terminal line black on costal half of wing, preceded with silver, subterminal line also silvery. 23 mm.

Common and generally distributed, in June. Larva on moss; refusing grass, at least when young. New York: common everywhere.

2. C. myellus Hübner. Chocolate brown, shading into ochre yellow at margins; three silver-white patches in middle of wing, separated by brown fasciæ, the first running to the middle of the wing, obliquely triangular, the second half as long and rather trapezoidal, the third a narrow subterminal bar. Terminal line obscure, continuous; fringe gray, cut with white opposite the cell. 22 mm. (luctuellus Fernald, not Herrich-Schæffer).

July. Caterpillar under moss; brown, with black head and cervical shield.

Labrador to Carbondale, Pennsylvania, and west to British Columbia. New York: Karner.

3. C. agitatellus Clemens. Fore wing broader than that of C. laqueatellus, of the same pale yellow, becoming darker along the markings, and nearly white on the inner margin. Silver streaks broader and separated only by the fine, often incomplete line along R; the longer streak less than three-fourths of the length of the wing. Middle of wing silver white from cell to outer margin, cut by the silver and yellow subterminal line. Apical marks as in laqueatellus; terminal line fine and black on costal third, replaced by five thick dots along outer margin below; base of fringe silver white toward costa, the rest silver gray. Hind wing cream white, lightly shaded with gray. 20 mm. (saltuellus Zeller.)

Generally distributed and common, especially northward; flying mostly in July. New York: North Elba, Summit of Mt. Marcy, Newcomb, North Creek, Rock City (Cattaraugus County), Ithaca, Trenton Falls, Schenectady, New Windsor, Crugers; Lynbrook, Long Island.

4. C. alboclavellus Zeller. Identical with C. agitatellus except for the lack of the brown line on \mathbf{R} , probably a variety of it. Ground color usually a little darker, and sometimes a little shaded with dark brown; trapezoidal patch beyond the silver streak usually brighter silver and standing out more distinctly (caro-linellus Haimbach).

July and August. Larva on grass.

Common and generally distributed. New York: Newport, Newcomb, Lewiston, Potter Swamp (Yates County), Ithaca, Big Indian Valley, Utica, Schenectady, New Windsor, Fort Montgomery, Katonah.

5. C. labradoriensis Christoph. Dark brown, diffusely shaded with whitish, especially opposite the cell, over M_2 to Cu_1 , and in areas extending to the inner margin before and on the middle of the wing; a silver bar in the cell, broad outwardly, but hardly reaching beyond middle of wing, separated from the whitish area beyond by a broad zigzag median band, the tip of the silver streak forming two teeth, the lower of which is only a little smaller than the upper; costa broadly brown; subterminal line normally silvery, a little enlarged at costa. A triangle before and one beyond it at costa. A terminal line above, and dots below, preceded narrowly with white. Hind wing fuscous. 20 mm. (*luctuellus* auct., not Herrich-Schæffer.)

This species may prove to be a race of C. luctiferellus, of Europe, which has Sc and \mathbf{R}_1 anastomosing. I have not examined the venation. C. dissectus Grote appears to be the same, both names applying to light forms.

New York (type of dissectus) and Mer Bleue, Ontario, to Oregon, and north. 6. C. bidens Zeller. Similar to C. alboclavellus and C. agitatellus; silver streak three-fifths length of wing, its tip bluntly angled, reaching costa, leaving only a fine brown costal edge, and strongly toothed below, at origin of Cu. Ground brownish ochre; no silver patch in middle of wing beyond cell; but middle of terminal space white; black terminal line extending below middle of wing; the dots somewbat elongate and distinctly farther from the margin than the terminal line is. Sometimes with a broad brown ray in the silver streak. Hind wing white, or cream with a white fringe. 18 mm.

This species is usually found in peat-bogs. Quebec_to Massachusetts, New York, and northern New Jersey. New York: Niagara Falls.

7. C. youngellus Kearfott. Closely similar to C. bidens, but smaller; brown costal edge slightly wider, ground yellow and somewhat dusted with light brown; a silver postmedian patch and a silver stripe on inner margin; terminal line stopping above middle of wing, and dots only below; more or less silver along the veins. Apical markings as in the preceding group; the terminal line and white streak in the fringe not ending abruptly as in *C. floridus*. Subterminal line broader than in *C. alboclavellus*. 18 mm.

Mer Bleue, Ottawa, Ontario. American records of C. hamellus appear to belong in part to this species.

8. C. præfectellus Zincken. Chocolate brown; inner margin hardly paler. Silver streak covering cell, and extending well beyond it, but not reaching the subterminal line, and separated from the costa by half its width; a slight silver streak above its apex, but the veins on the disc hardly, or not at all, marked with silver; subterminal line silvery, as usual; moderately bent opposite cell, but little widened at the costa. Terminal space brown, with silver triangles above and below an oblique apical dash; a few white scales in lower part of wing, but no spot. Terminal line brown, hardly darker; dots below oval, and set well back from margin. Fringe shining brown-gray, becoming white in its base, toward the apex. Hind wing cream white. 2 mm.

Two broods, flying mainly in June and August; common. New York. Common everywhere.

Generally distributed.

Hardly distinct from C. leachellus. In the Colorado race, oslarellus Haimbach, the hind wing is dark.

9. C. leachellus Zincken. Like C. præfectellus, but with the costal edge only very narrowly brown toward the base; apex normally slightly more acute; region beyond tip of silver streak distinctly yellow. Frequently larger; expanding up to 27 mm.

June and July.

Common and generally distributed, flying with C. præfectellus. New York: Wilmington, Saranac Lake, Newport, Ithaca, New York City, Yaphank.

This species is probably the true hastiferellus of Walker, rather than the following, which may be a southern race of it.

10. C. quinquareatus Zeller. Closely similar to C. leachellus, the light area beyond the tip of the silver stripe nearly white, and connecting it with the subapical silver triangle, which is a little larger than in C. leachellus. Apex a little more strongly pointed, but terminal line and white streak in fringe not ending abruptly; terminal dots replaced by slender bars nearly as long as those of C. laqueatellus, on a fuscous ground. No silver line above the tip of the streak in cell. (extorralis Hulst; hastiferellus auct., not Walker.)

Southern States, north to Pennsylvania; northern distribution uncertain on account of confusion with C. leachellus (hastiferellus Walker.)

11. unistriatellus Packard. Chocolate brown; a broad silver white stripe from base to costal half of outer margin, leaving a brown costal stripe which is extremely narrow at the base and apex, but reaches down to the cell between, and is widest at three-fourths way out. Terminal line and dots very weak; costal half of fringe white. 25 mm. (exesus Grote.)

Apparently rare.

Labrador to Pennsylvania; west to California. New York: Saranac Inn, North Creek, Newport, Trenton Falls, Schenectady.

12. C. girardellus Clemens. Silver white; a bright yellow stripe from base to postmedial region, below cell, in typical specimens widening and then fading out at onter end; defined with a black line above; a little yellow on base of costal edge; and sometimes beyond middle also, rarely forming an angulate fascia, bounded by fine black postmedial and subterminal lines; terminal line fine, black, preceded by black dots on dorsal half. 25 mm. (nivihumellus Walker.)

Common in June and July.

Generally distributed south to Pennsylvania and Ohio. New York: Saranac Inn, Newcomb, North Creek, Fentons (Lewis County), Batavia, Rock City (Cattaraugus County), Otto, Ithaca, Trenton Falls, Liberty, Big Indian Valley, Schenectady, Nassau, New Windsor, Katonah, New York City, Fort Montgomery.

13. C. dæckellus Haimbach. Near C. floridus, but much larger, deeper yellowbrown, the veins streaked with lead-gray only; the brown line between the silver cell and postmedial patch, twice as wide as a vein; terminal space more evenly yellowish in color, and narrower, even, than in floridus; inner margin, below A, pure white, though with diffuse upper boundary; the outer half of the inner edge narrowly blackish. Terminal markings as in floridus. 27-31 mm.

September.

New Jersey generally; perhaps elsewhere confused with C. floridus.

14. C. floridus Zeller. Closely similar to C. alboclavellus (and agitatellus); apex a little more acute, with a distinct inward angle opposite the cell, at which the black terminal line stops abruptly. Silver streak diverging gradually from costa, and acute, as in *leachellus*. Inner margin silver white, but not contrasting nor sharply defined; apex more contrastingly white. 22 mm. June and July. Caterpillar on grass. Atlantic States (apparently not common) to California. New York: Ithaca.

Very close to both alboclavellus and the European C. pascuellus, of which latter it is probably a race.

15. C. hamellus Thunberg. Fuscous; apex a little produced; silver stripe very narrow, being not wider than the brown costa, outwardly; terminal dots on dorsal half of wing a little elongate, followed by a fine terminal line. Hind wing grayish; inner margin not pale. 20 mm.

August.

Our form is very dark on the costa above the silver streak. Kamouraska, Quebec; Maine; Mt. Wachusett, Massachusetts (Forbes); Europe. New York: Saranac Inn and Albany. (New York State Collection.)

16. C. lyonsellus Haimbach. Similar to C. floridus; apex squarish; silver streak almost touching the costa to beyond the middle, then abruptly leaving it, as in C. bidens; no distinct silver postmedial spot; subterminal line regularly curved, meeting the costa almost at right angles; terminal line not ending abruptly at middle of outer margin. Subterminal and terminal spaces typically filled with bright ochre. 21 mm.

Late June.

Ottawa, Ontario; New Jersey.

17. C. albellus Clemens. White, very rarely pale gray; outer margin somewhat shaded with pale yellow. Medial line dark brown, fine, strongly angled out on discal fold, forming an acute tooth, obsolete on dorsal half of wing, or represented by some black scales. Postmedial line indicated by a costal stria; subterminal line double, light brown, silver-filled, angled opposite the cell, where its outer line may touch the margin. Fringe silver gray, with a white base on the costal third, ending abruptly at the tooth of the subterminal lines. Hind wing gray. 15 mm.

June and July. Larva on grass.

Common in wet places.

New York: Trenton Falls, Rock City (Cattaraugus County), Ithaca, Big Indian Valley, Scheneetady, Poughkeepsie, New Windsor.

†† Fore wing with long-drawn-out, falcate apex.

18. C. satrapellus Zincken. Silver streak broad, but widely separated from costa; ending in a sharp point seven-eighths way to outer margin, with a large oval silver spot above its tip, and with a slender spur from its lower side, running along Cu_2 . Ground yellow, becoming brown along the veins and edges of the markings. Terminal space powdery fuscous, cut by short black streaks below: terminal line obsolete below; subterminal line lead gray, running across the apex, obsolete below; fringe white, brown-tipped on costal half. A white oblique streak running to the apex. Hind wing pale gray, 25-35 mm.

July.

Southern States, north to New Jersey; South America.

****** Fore wing without a silvery stripe.

19. C. elegans Clemens. Ground white; more or less shaded with dull brown, always brown before the subterminal line, and more or less so along the costa toward the base, and on the outer margin; sometimes dominantly brown, but always with a large part of the terminal space white. A brown median oblique shade to beyond middle of costa, often forming the inner boundary of a brown area; and a blackish oblique shade running to the middle of the inner margin; the two sometimes connected by a brown line which forms a long tooth opposite the end of the cell, and is preceded by a white line. Subterminal line characteristic, white, followed by brown, nearly erect, but slightly excurved. Seven black terminal dots. 12-15 mm.

Larva on grass. Moth from July to September. Common and generally distributed. New York: Wilmington, Speculator, Otto, Ithaca, Katonah, New Windsor, New York City, Staten Island.

This is a broad-winged primitive species of doubtful affinity, superficially very closely resembling albellus, but apparently not related. Sc and R_1 are closely approximate but not fused.

A form of C. polingi Kearfott has been taken a few times at Ithaca. New York. in July. It is grayer than C. elegans. with the double dark subterminal band markedly angled opposite the cell, and inwardly oblique to costa, filled with white at costa, but below more or less definitely dentate, and suffused; with the black crescent on the inner margin constricted on A; the terminal dots enlarged, and typically fusing into a line; and the labial palpi heavily handed with white. It

was described from Arizona. 20. C. turbatellus Walker. Silver white; a black dot at lower angle of cell; one below cell beyond middle, and a subterminal series; a sinuous postmedial series, very often lost, and a terminal series of seven dots. All the dots except the terminal ones and the one below the cell frequently obsolete. 20-25 mm. (bipunctellus Zeller.)

July.

Canada to Pennsylvania and Illinois. New York: Otto, Rock City (Cattaraugus County), Ithaca. Big Indian Valley.

21. C. perlellus Scopoli, race innotatellus Walker. Silver white, without markings; the female often silver-gray; hind wing sometimes pale gray in the male also. 20 mm. (sericinellus Zeller, inornatellus Clemens.)

July and August. Larva on grass.

Northern States, south to northern Pennsylvania, Illinois, and California; com-moner northward. New York: Wilmington, North Elba, Saranae Inn, Newcomb. Newport, Otto, Big Indian Valley, Onteora Mountain. The typical form from Europe is darker.

22. C. teterrellus Zineken. Fuseous brown, strongly shaded with violet-grav between the veins; fresh specimens often dominantly violet-gray. Median line

brown, running across the end of the cell and bent at a right angle on the cell; irregular and black across the fold, where it usually forms a short tooth; sub-terminal line dark brown, followed by lead gray, and then again with brown toward the costa, somewhat excurved, bending away from the outer margin toward the costa. Terminal space powdery fuseous, each scale palc with a broad brown tip; seven black terminal dots; fringe shining lead-gray, without golden tint. 15-21 mm. (camurellus Clemens, terrellus Zeller.)

June and July. Larva on grass; ycllowish or greenish white, with darker dorsal vessel; tubercles and set a large; head rough, dull light brown; cervical shield darker, inconspicuous.

Common north to New York; rarer in Maine. "New York" (American Museum Natural History.)

23. C. decorellus Zincken. Clay color, broadly shaded with shining ash gray between the veins, much more conspicuously streaked in appearance than the last species. Median line brownish yellow, a little irregular and excurved, partly black in the fold; subterminal line brownish yellow, followed by lead gray; evenly excurved, very closely parallel to outer margin except at costa; the terminal space ochre yellow, often containing a gray streak toward the costa, but not powdery. Seven black terminal dots; fringe golden bronze. 19-27 mm. (polyactinellus Zeller, goodellianus Grote, bonusculalis Hulst.)

July and August.

Massachusetts to Georgia and Texas. New York: Ithaca and New Windsor.

24. C. vulgivagellus Clemens. Clay color, broadly striped with dull gray between the veins; no transverse markings; seven black terminal dots; fringe golden bronze. 20-39 mm. (usually 25 mm.).

Late August and September. Larva on various grasses and grains, brown with darker tubercles, and a deep brown cervical shield. Head black. Hibernation in the half-grown larva; cocoon spun in May; pupation in July. Common and generally distributed. New York: Common and general.

25. C. ruricolellus Zeller. Clay color, shading into ochre at costa and outer margin; lightly dusted with deep brown, usually leaving the costa nearly clear; median and subterminal lines indicated by a gathering of the dusting; the median short, thick, oblique, in the middle of the disc, sometimes weakly continued to the costa and inner margin; the subterminal oblique, sinuous, nearest outer margin opposite the cell; veins toward the apex often streaked with pale gray and edged with the brown scaling. Seven terminal dots; fringe golden bronze. 20 mm.

Larva on grass and sorrel. Moth in August and September.

Common south to Pennsylvania and Missouri. New York: Ausable Lake, Wilmington. Saranac Inn. Honeove Falls, Batavia, Hemlock Lake, Ithaca, Liberty, Big Indian Valley, and Onteora Mountain.

26. C. mutabilis Clemens. Male antennæ with narrow, well-separated laminations. almost appearing unipectinate. Fore wing dull ash gray, with whitish streak from base, through cell to costal half of outer margin; streaked outwardly with the ground color; and with scattered black scales. Median line usually reduced to a black patch in upper part of fold near end of cell, irregularly oblique when more distinct; subterminal line usually reduced to dots, nearly parallel to outer margin, and regularly dentate on the veins when distinct. Seven minute black terminal dots; fringe powdery fuscous, hardly shining. Hind wing light fuscous.

Larva on grass. Moth in August.

Common and generally distributed. New York: Peru, Newport, Lancaster. Otto, Rock City (Cattaraugus County), Ithaca, Big Indian Valley, New Windsor, New York City.

B. Fore wing with R₁ anastomosing, or becoming coincident, with Sc (fig. 369).

* Terminal line continuous above, replaced by black dots below.

27. C. hortuellus Hübner, race topiarius Grote. Clay color, becoming clear yellow toward the outer margin; with dark fuscous, more or less shining stripes between the veins, leaving a clear yellow space before the subterminal line; two such stripes in the cell, and three oblique ones running down from the costa outwardly, the first nearly longitudinal and extending along the costa to the base; subterminal line silver gray, angled moderately opposite the cell, often edged with brown, and followed by a second silver streak at the costa; terminal line and dots contrasting on the yellow ground; fringe shining, somewhat bronzy. Hind wing gray, with a pale fringe as usual in the genus. 18 mm. Very common in June and July. Larva on various low plants; sometimes

Very common in June and July. Larva on various low plants; sometimes injurious to cranberry. Larva dirty gray with light yellow-brown head, and with black clypeus and labrum; cervical shield inconspicuous.

Generally distributed. New York: Common and general.

** Terminal line formed of dots or inconspicuous.

28. C. trisectus Walker. Light grayish luteous, shaded with dull gray between veins, leaving fine pale streaks on the veins, and outwardly in the middle of the interspaces; costal region more or less streaked with yellowish, sometimes clear light ochre. Median and subterminal lines represented by blackish oblique shades across the middle of the wing; terminal dots weak, often obsolete; fringe fuscous, strongly cut with white between the veins. 28 mm. (biliturellus Zeller, exsiccatus Zeller, interminellus Walker.)

Larva on grass. Two broods, the moth flying in June and August.

Generally distributed and not rare. New York: Newport, Fentons (Lewis Co.), Newcomb, North Creek, Honeoye Falls, Batavia, Lewiston, Otto, Ithaca, Big Indian Valley, Little Falls, Schenectady, Nassau, Rhinebeck, New Windsor, and Lynbrook, Long Island.

Inornatellus Walker may be a pale or rubbed specimen of this species.

29. C. laciniellus Grote. Fore wing pale cinerous ochreous, especially on the costal and outer portion of the wing; a few scattered brown scales. Median and subterminal lines faint, of ochre scales, partly overlaid with brown; three terminal dots on dorsal part of wing; fringe evenly gray, pale at base. 27 mm.

Maine.

30. C. caliginosellus Clemens. Smoky brown or fuscous; somewhat powdery and showing traces of a luteous ground. Median and subterminal lines blackish, underlaid with yellow-brown, irregularly dentate, and roughly parallel; often broken and obscure, the median line sometimes obsolete; terminal line faint, widening into dots between the veins in some specimens. Fringe and hind wing comcolorous. 13-25 mm.

Larva pinkish white, tinged with brown; with a dark brown or black head; sometimes injurious to young corn, girdling it and eating out the growing point; also on grass. Two broods, June to end of August.

Common and generally distributed. New York: Lancaster, Otto, Ithaca, Nassau, Greenwich, New Windsor.

31. C. zeellus Fernald. Closely similar to C. caliginoscilus, of which it may be a pale variety; ground luteous, considerably overlaid with smoky brown; with the markings of C. caliginoscilus in blackish brown. 18-24 mm.

Larva like C. caliginosellus; likewise injurious to corn.

Maine to West Virginia, Illinois, and Missouri. New York: Batavia, Ithaca, Big Indian Valley, Little Falls, Albany, Rhinebeck, Katonah, Staten Island.

32. C. luteolellus Clemens. Pale ochre yellow, marked with tawny brown; the markings exactly as in *caliginosellus*, but weaker, and often nearly obsolete. Fringe pale brown, not shining. Valves more slender than in *caliginosellus* and *zeellus*. 23 mm.

Common and generally distributed. New York: Peru, Newcomb, Ithaca, Little Falls, Katonah.

The variety ulæ Cockerell, from Colorado and Arizona, is much more contrasty, and has black terminal dots.

II. Eyes half as wide as front, or slightly less; palpi very hairy; venation as in the first group.

33. C. trichostomus Christoph. Powdery dull light gray; head and thorax blackish; a broad oblique blackish antemedial hand from middle of costa to a third way out on inner margin, followed by a white shade; a similar, but usually narrower, subterninal band, roughly parallel to the outer margin, but sinuate below the costa, followed by a stronger white shade. Terminal line practically continuous, heavy, black; fringe gray, cut with white. Hind wing slightly browner. 20 mm.

Labrador.

70. RAPHIPTERA Hampson

(Crambus, in part)

Similar to Crambus; \mathbf{R}_1 lost and \mathbf{R}_3 and \mathbf{R}_4 completely fused, also \mathbf{M}_2 and \mathbf{M}_3 , leaving no stalked veins in the fore wing. Hind wing with \mathbf{M}_2 and \mathbf{M}_3 completely fused; fore wing strongly falcate.

'l. R. minimella Robinson. Mouse gray; cell white, slightly silvery, the streak ending in a point five-sixths way to apex, with a white chevron over its apex, and a triangular patch running to tip of wing; costa shaded with luteous as far as the postmedial line; postmedial line oblique outward to opposite cell, then brown, nearly even and parallel to outer margin, aeross to the inner margin. Hind wing fuscous, sometimes pale at base. 12 mm.

Southern States and Illinois.

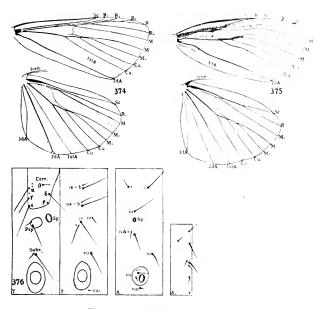
2. R. argillaceellus Packard. Similar, slightly larger; costal region not contrastingly pale, there being strong dark stripes both above and below \mathbf{R} , the lower one noticeably narrowing the stria in the cell. 10–15 mm.

This is probably a northern race of R. minimellus.

Labrador, Mer Bleue; Ontario; Wisconsin; Connecticut. New York: McLean Bogs.

Subfamily EPIPASCHIINÆ

Antennal socket more or less completely surrounded by scales, in the type genus with a wide gap on the lower side of the circle, leaving the socket in contact with the eye, but more often with the antenna and eye wholly separated by a scaled area. Ocelli present; antennæ normally fasciculate in male, often with a long plume-like process on scape. Labial palpi large, upturned, in many males hollowed out, and enclosing the plume-like maxillary palpi. Maxillary palpi of females and of a few males normal, somewhat triangularly scaled; always well developed. Tongue developed. Fore wing with \mathbf{R}_1 free, \mathbf{R}_5 stalked with \mathbf{R}_3 and \mathbf{R}_4 . Male often with fovea and distorted venation; 1st A absent; 3d A free and weak. Frenulum simple in both sexes. Hind wing with Sc and R approximate or anastomosing; \mathbf{M}_1 from near upper angle of cell; cell distinctly elosed by a strongly curved vein, M_{\circ} from lower angle or stalked; Cu with weak fringe or none.



FIGS. 374-376. EPIPASCHIINÆ

374, Epipaschia superatalis, venation; 375. Tetralopha asperatella, venation, and sex characters of male; 376, T. militella (?), seta map of larva

This is a small group, transitional between the Pyralidinæ and Phycitinæ. The larvæ (fig. 376) are normally leaf rollers much like those of the Phycitinæ; one Indian species, at least, is social.

Key to the genera

Sc and R of hind wing approximate (fig. 374).....71. Epipaschia. Sc and R fused for a distance beyond end of cell (fig. 375).

73. Tetralopha.

71. EPIPASCHIA Clemens

(Macalla, in part)

Male with fasciculate antennæ; process on first joint very long and heavily fringed with hair below, turned back over thorax; palpi not modified; fore wing (fig. 374) with \mathbf{R}_2 and \mathbf{M}_1 free, \mathbf{M}_2 and \mathbf{M}_3 closely approximate at base or stalked; hind wing with \mathbf{M}_2 and \mathbf{M}_3 closely approximate or stalked; \mathbf{M}_1 when approxi-

60.5

mate appearing to be a continuation of the discocellular vein, as in many Phycitinæ.

1. E. superatalis Clemens. Clay color, often more or less dusted with gray and black; antemedial line dentate and broken, or reduced to dots; postmedial oblique above, triangular and sinuous below, black, subdentate and tending to break up into dots, followed by a narrow pale area; the outer margin largely or wholly dull red-brown to gray. Black terminal bars. Hind wing fuscous, somewhat paler at base. 20-25 mm.

Generally distributed. New York: New Windsor; Chappaqua, Long Island.

2. E. zelleri Grote. Base dull gray, ending in a broad, straight, erect, antemedial line, slightly dentate in female; postmedial line sinuous, meeting costa and inner margin at right angles; with a little whitish beyond it on costa. Median area whitish, more or less suffused with gray, especially in the female; a more or less distinct blackish discal bar. Terminal space concolorous with base. Hind wing gray. 23 mm.

July.

New Jersey to Florida and west.

72. ONEIDA Hulst

(*Tioga*, in part)

Similar to Tetralopha, except as noted in the generic key. The position of \mathbf{M}_1 , which has been used for the separation of Oneida from the Tetralopha group, is variable in both genera. The male maxillary palpi and wings are apparently not modified.

1. O. lunulalis Hulst. Bluish gray, rather pale before the scale ridge, which is of brown-tipped scales. Antemedial line far out, excurved, double, obscure. A small whitish tuft toward end of cell, and a pale brown raised discal bar. Postmedial concave on upper half of wing, contrasting, pale, followed by a series of blackish streaks on a brown patch, and preceded by a dark brown line; rest of the line very weak and nearly parallel to outer margin. A broken dark terminal line and a pale line in base of fringe, followed by a gray shade. Hind wing light fuseous. 22 mm.

June to August.

Massachusetts to Illinois; Canada. New York: Ithaca.

73. TETRALOPHA Zeller

(Pococera, in part; Lanthaphe Clemens; Benta Walker; Ratona, Saluda, Loma, Wanda, Attacapa, Tioga Hulst; etc.)

Scale ring about socket of antenna complete; male antenna fasciculate, with a more or less developed process on the scape; maxillary palpus of male plumelike, enclosed in a groove in labial palpus. Fore wing (fig. 375) with small separated antemedial tufts; venation unstable, but normally with \mathbf{R}_2 stalked, \mathbf{M}_1 free, \mathbf{M}_2 and \mathbf{M}_3 approximate or shortly stalked; male with a more or less hyaline area in the cell, with a large tuft of enlarged scales projecting down from costa on the under side and partly covering it. Hind wing with Sc and R fused beyond cell, \mathbf{M}_2 and \mathbf{M}_3 approximate, or normally stalked.

The species are almost hopelessly confused, and the analysis below may not be wholly correct.

Key to the species

1. Antemedial and subterminal spaces suffused with light reddish brown.

5. baptisiella.

- 1. Outer part without strong pale red suffusion.

 - 2. Head, collar, and base of wings suffused with pinkish.....4. militella.

2. Head, thorax, and base of wings powdery gray.

- 3. Thorax and base of wings brown; black scaling of wing with purple
- 3. Thorax and base paler luteous or gray; purple iridescence, when present, on the paler scales of outer part of wing. 4. Ante- and postmedial lines both more or less diffuse; the ground

 - 4. Postmedial line more diffuse, less contrasting than the outer antemedial, which is followed by a whitish shade.....2. subcanalis.

1. T. asperatella Clemens. Rather even powdery gray (black on white). Outer half of wing sometimes distinctly paler, especially in female; tufts small, well contrasted, black; antemedial line doubled, darker gray, outwardly oblique, interrupted in cell by the translucent area in male; postmedial irregularly sinuous, denticulate, followed by a paler shade, inconspicuous. Process on antennæ longer than scape; normal; plume variable, yellow or blackish. 20-25 mm.

Larva green, granulated with yellow; on oak, maple, elm, and beech. Moth in June and July.

Generally distributed. New York: Rock City (Cattaraugus County), Ithaca, New Windsor (Morton).

In var. nephelotella Hulst (clemensalis Dyar), the thorax and base of wings is pale and pinkish, almost as in platanella, but the darker, rather than paler, outer part of the wing and the narrower wing will distinguish it. Var. expandens Walker is transitional.

2. T. subcanalis Walker. Similar to the preceding species, antemedial line contrasting, blackish, preceded by a heavy blackish shade covering the tufts and the first antemedial line. Plume blackish. 22 mm. (*militella* Hulst, not Zeller.)

Illinois to Texas.

This is probably merely a variety of T. asperatella.

3. T. melanogrammos Zeller. Plume variable in color. Ground of thorax and base and outer part of wing dull gray, somewhat suffused with reddish and dusted with black; of median area white, dusted with black. Antemedial line double, black, white-filled, contrasting, a little irregular and a little farther out on inner margin; interrupted by the translucent patch in the cell of the male. Postmedial similar and equally distinct, its outer line suffusing into the gray outer region; somewhat sinuous and a little irregular, but not dentate; ante-medial tufts black-tipped; medial tufts white, with some black scales at end of cell. 20 mm.

Larva on shrubby Leguminosæ (Gleditschia, Prosopis).

New Jersey, south and west. New York: New Windsor (Morton). In the typical form, from the Southwest, the contrast between the dark base and pale median area is not striking, only the antemedial space being distinctly pale. Our specimens are variety diluculella Grote (*talleolalis* Hulst). 4. T. militella Zeller. Wings broad; plume black. Head, thorax, and base of

wings dull red-brown, with a pinkish tint, and much darker than the outer part, which is also a little pinkish; many scales, especially on the under side, crimsontipped. Antemedial line blackish, separated from base by a white line; straight and erect; no nearer the base on the costa. Tuits mostly brown; median area nearly white, especially toward the antemedial line; postmedial line a little paler gray and more diffuse, somewhat sinuous, and, when most distinct, dentate in the middle, followed with white, but with the outer part hardly darker and not contrasting. Tufts in median area pale; no distinct discal dot. 22 mm. (platanella Clemens.)

Larva on Platanus.

Generally distributed. New York: Hion, Rhinebeck (Dyar), Onteora Mountain. 5. T. baptisiella Fernald. Wings not very broad, plume blackish or yellow. Thorax and basal half of wing white, somewhat shaded with light red, especially the antemedial region; antemedial line white, dentate, mostly lost in the male; outer half of wing light red; postmedial deeply dentate, the teeth defined on the inner side with blackish, on the outer with white, especially in the middle of the wing; all tufts white, black-tipped; the antemedial one below A absent. Terminal line practically complete. Fore wing sometimes more or less suffused with gray, but the species always distinguishable by the deeply dentate postmedial line and the dentate antemedial, ending near the middle of the inner margin. No distinct process on male antenna; a small scale-tuft only. 20 mm.

Larva on Baptisia.

Seen from New Jersey, West Virginia, and west. "New York" (Fernald).

6. T. humerella Ragonot. Shining powdery gray. Postmedial far out, pale, defined with gray, with a blackish shade on the costa beyond it; antemedial less distinct and far out; tufts white, heavily black-tipped, contrasting, the base of the wing before them contrasting, whitish. Terminal dots heavy, sometimes confluent, alternating with heavy dark gray bars in the basal half of the fringe. 18 mm. (formosella Hulst).

Missouri.

7. T. robustella Zeller. Wings broad. Basal segment of antenna much enlarged, without a process; maxillary plume yellow in male. Basal third purple black, paler at extreme base, followed by an even, usually concave, fine, whitish antemedial line, cut by a small blister in the cell in the male; antemedial space pale gray, shading into the darker gray outer half. Postmedial strongly sinuous, pale, diffuse, terminal fine, broken; tufts black and white. 22 mm. (scortealis Lederer, diluculella Grote.)

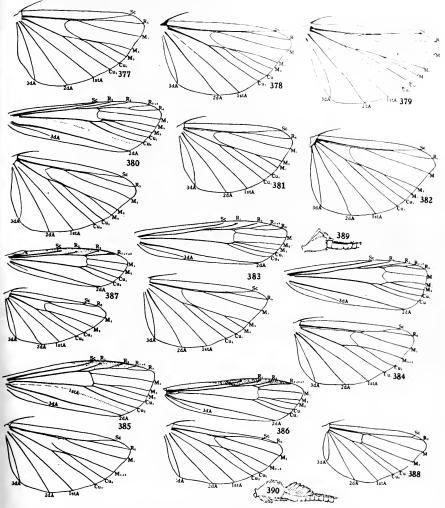
Larva on pine.

Apparently general but rarely taken. Cœnodomus hockingii Walsingham (Dyaria singularis Neumœgen), a heavy Noctuid-like species with pectinate antennæ, from India, has been taken but once and is doubtless a stray introduction.

Subfamily PHYCITINÆ

(Phycinæ)

Head with antennal socket separated from eye by a complete line of scales; ocelli present; male antennæ often fasciculate, with the base of the shaft often curved and bearing a tuft of scales on the concave side (fig. 390); scape very rarely with a projection; antennæ often normal. Palpi often upturned, always large; maxillary palpi always well developed, in the male of some genera with long hair scales on the outer joints, forming a plume-like structure, concealed in a groove in the second segment of the labial palpus, which is in that case almost always closely upturned; tongue well-developed, the palpi curving out a little near the base, to make room for it. Fore wing with one radial lost, R_3 and R_6 stalked, \mathbf{R}_2 usually free; rarely, with \mathbf{R}_3 and \mathbf{R}_6 completely united; 1st A lost, 3d A weak and free. Hind wing with Sc and R usually fused for a distance beyond the end of the cell, rarely closely approximate only; base of R as far as end of cell, often lost, making Sc appear to rise from the end of the cell; the free tip of Sc occasionally lost, and often very short, running directly across to the costa. Frenulum of female simple. Hind wing with a strong fringe of straight stiff hairs on base of Cu above; in many genera with one or two medials lost; middle discocellular typically strongly curved, the lower part often very closely approximate with the lower side of cell (base of Cu_1 and M-Cu), always formed as a normal tubular vein at the two ends; in the middle, sometimes very weak but never lost. M_1 usually stalked with R before R joins Sc, M_2 usually stalked



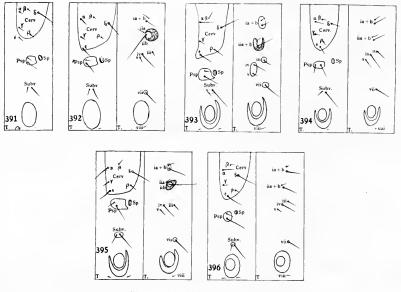
FIGS. 377-390. PHYCITINÆ

377, Rhodophæa hystriculella (Western States), venation of hind wing; 378, Aorobasis zelleri (Europe), venation of hind wing; 379, Dioryctria species, vena-tion of hind wing; 380, Laodamia fusca, venation; 381, Salebria contatella, vena-tion of hind wing; 382, Etiella zinckenella, venation; 381, Salebria contatella, vena-tion of hind wing; 382, Etiella zinckenella, venation; 381, Canarsia ulmi-arrosorella, venation; 386, Moodna ostrinella, venation; 387, Plodia interpunctella, venation; 388, Homœosoma masinella (Europe), venation of hind wing; 389, Acro-basis, base of male antenna; 390, Loodamia fusca, base of male antenna. basis, base of male antenna; 390, Laodamia fusca, base of male antenna

20

with \mathbf{M}_{a} ; \mathbf{M}_{2} occasionally continuous with the outer part of the middle discocellular, and running closely approximate to \mathbf{M}_{a} for a considerable distance, but not fusing with it.

The larvæ (figs. 391 to 396) are very generally leaf rollers and crumplers, often living in a tube of silk mixed with frass, within the folded leaf; several species eat stored food etc., and become serious pests. They are distinguished from most



FIGS. 391-396. PHYCITINÆ

391, Myelois cribrella (Europe) seta map of prothorax of larva; **392**, Acrobasis sodalella (Europe), seta map of thorax of larva; **393**, Dioryctria zimmermanni, seta map of thorax; **394**, Etiella zinckenella, seta map of thorax; **395**, Vitula edmandsii, seta map of thorax; **396**, Hom ∞ osoma nimbella (Europe), seta map of thorax

other Pyralid larvæ by tubercle iii of the metathorax, which is normally enlarged, black with a white center.

The subfamily seems to be derived from the Pyralidinæ, perhaps through the Epipaschiinæ; the genus Omphalocera is a synthetic one, with the venation of the Pyralidinæ, the sex-scaling on the wings most characteristic of the Epipaschiinæ, and a common Phyeid type of male antenna. Its larva is also rather Phycid than Pyraline.

The middle discocellular of the hind wing is of four principal types:

(1) Discocellular eurved strongly above middle, the lower part long, oblique, meeting lower side of cell at a sharp angle, well beyond the origin of Cu_1 , so that M_2 and M_3 appear connate. This type is represented by the genus Trachycera in Texas, but appears not to reach our area.

(2) Discoellular strongly curved, the lower part somewhat longer, meeting lower side of cell at a sharp angle not far from origin of Cu_1 (normally a little

beyond), not running closely parallel to lower side of cell. M2 and M2 more or less stalked. Cell moderate (for example, Aerobasis, fig. 378).

(3) Discocellular very strongly eurved, its lower part approximate to the lower side of the cell for a greater or less distance before the origin of Cu, making Cu. appear stalked (for example, Dioryctria; Nephopteryx). The group may be divided into two subtypes; a, in Dioryctria (fig. 379), Ulophora, and Monoptilota, Cu_z separates the subtype states of the states of t rates from the cell at a third of the length of the wing; b, in most of the other genera it separates two-fifths way out (fig. 380).

(4) Discocellular practically straight and transverse, a little broadened, and not tubular $(\mathbf{M}_2 \text{ lost})$ (fig. 388).

Many genera which have lost M_2 show a condition intermediate between types (2) and (4) (fig. 386).

Key to genera

- 1. Hind wing with all veins present, or with Sc weak (Cu apparently 4-branched); fore wing with all veins present (except the missing radial characteristic of the subfamily).

 - 2. Male antennæ not unipectinate.
 - 3. Palpi upturned or recurved, the last joint rarely porrect.
 - 4. Discocellular of type (2) (fig. 378).

5. Male antennæ simple.

5. Scape of male antenna strongly dilated or angulate at apex; rarely

- 4. Discocellular of type (3) (figs. 379, 380).
 - 5. Eves small: palpi with first two segments long-hairy below.

81. Polopeustis.

5. Eyes normal; palpi sealed.

- 6. Male antennæ practically straight, but a little thickened with scales; palpi scaled, maxillary palpi without plume. 7. Cu₂ of hind wing arising one-third way out (fig. 379).

 - 8. Maxillary palpi of male flattened, closely appressed against the

7. Cu₂ of hind wing arising two-fifths way out. 8. Maxillary palpi flattened, closely appressed against the face. 79. Glyptocera.

- 6. Male antennæ more or less, usually strongly, curved at base, with a tuft of seales filling the concavity; maxillary palpi usually plume-like; Cu2 arising from cell two-fifths of the way to margin of wing.
 - 7. Maxillary palpi filiform; labials without groove.
 - 8. Fore wing with \mathbf{M}_2 and \mathbf{M}_3 approximate.
 - 7. Maxillary palpi plume-like; rarely, forming a sealy tuft, enclosed in a groove in labial palpi.
 - 8. Fore wing with \mathbf{R}_2 shortly stalked, \mathbf{M}_2 and \mathbf{M}_3 stalked or connate; hind wing usually with Cu, apparently stalked.

85. Meroptera.

8. **R**₂ from cell; **M**₂ and **M**₃ often separate.

9. Middle discocellular curved far in, so that the lower part of it is continuous with the general direction of Cu_2 ; male antennæ eurved at base.

- 9. Discocellular decidedly more transverse than Cu₂.
 - 10. Male antennæ flattened, with a strong tuft; front generally smooth; \mathbf{M}_2 and \mathbf{M}_3 of fore wing often closely approximated.
 - 11. Patpi reaching barely up to base of antennæ. .88. Myrlæa.
 - 11. Palpi strongly overlapping base of antennæ.

89. Laodamia.

- 10. Male antennæ hardly flattened, with a weak scale-tuft; front with a strong tuft; M_2 and M_3 well separated (fig. 383).....90. Elasmopalpus.
- 3. Palpi oblique or porrect, with porrect last joint.

4. Second joint oblique, without a plume in male.

- 4. Second joint very long and straight, with a groove enclosing a plume.

92. Etiella.

- 1. Hind wing with M_2 lost (completely fused with M_3); Cu therefore apparently trifid.
 - 2. R_3 and R_5 of fore wing stalked; Sc of hind wing generally distinct.
 - 3. Hind wing with Cu_2 arising well back from angle of cell (five-sixths way out).
 - 4. Palpi porrect or oblique, the third joint drooping or porrect.
 - Maxillary palpi large, scaly, exposed; basal joint of labials with a projecting tuft; tongue hidden; male antennæ pectinate, female subpectinate
 Maxillary palpi cylindrical; basal joint of labial palpi not projecting;
 - 4. Palpi ascending, sometimes recurved.

- 3. Hind wing with Cu_2 arising very near angle of cell.
 - 4. Fore wing with Cu apparently quadrifid.

5. Palpi ascending, curved, with ascending third joint.

6. Fore wing with M₂ and M₃ not in line with Cu.

7. R₂ from cell, M₂ and M₃ stalked; hind wing with Sc short.

97. Lætilia.

- 5. Palpi oblique or porrect, third joint drooping.
 - 6. M_2 and M_3 in line with stem of Cu.....102. Honora. 6. M_2 and M_3 distinctly out of line with stem of Cu, long-stalked.

100. Valdivia.

3. Middle discocellular vein of hind wing of type (4) (fig. 388).

104. Homœosoma.

3. Middle discocellular of hind wing more normal, oblique and not flattened.
4. Hind wing with Cu_2 from angle of cell; Cu_1 and M_2 long-stalked.
106 Enhestindes
4. Hind wing with Cu_2 distant from angle of cell; cell mostly short; M_2
mostly free (fig. 386).
5. Fore wing with Cu quadrifid107. Moodna.
5. Fore wing with Cu trifid.
6. Palpi recurved105. Ephestia.
6. Palpi drooping.
7. Cell of hind wing short (fig. 387) 105. Plodia.
7. Cell half the length of the wing109. Caudellia.

74. MYELOIS Hübner

Antennæ simple, pubescent; front smooth, palpi smooth, upturned to vertex. tapering; maxillary palpi slightly rough, normal. Fore wing smooth, M_2 and M_3 distinctly stalked, not in line with Cu; hind wing with long cell; middle discocellular meeting lower side of cell at 30 degrees, just beyond origin of Cu.

Key to the species

1. Sc of hind wing long, nearly parallel to R.

2. Fore wing blackish; the base a little paler; markings obsolescent.

1. obnupsella.

- 2. Fore wing blackish, with distinct lines.
- 1. Sc of hind wing very short, at a sharp angle to R.
 - 2. Antemedial line straight, continuous, almost perpendicular.....5. alatella.

1. M. obnupsella Hulst. Gray, hardly brownish, lightly dusted with white; gathering to form a distinct, upright, antemedial pale shade, followed by an equally wide shade without any dusting. Fringe similar. Hind wing whitish, with dark veins, outer edge, and costa. 20 mm. (subtetricella Ragonot.)

May.

Canada to Florida.

In many specimens the postmedial line is distinctly indicated by a gathering

of the white dusting. 2. M. żonulella Ragonot. Blackish gray; lines pale, edged with blackish; aute-medial oblique, straight, reaching almost to middle of inner margin; the dark edging on outer side heavy; base of wing and outer margin paler. Postmedial line oblique, sinuous and denticulated; discal dots distinct, black; no terminal dots. Hind wing dark brown-gray, but paler than fore wing. 22 mm.

May. (Not seen.)

Northern Illinois.

3. M. bilineatella Ragonot. Fore wing blackish gray; antemedial line whitish, followed by a dark shade, nearly straight and erect; postmedial line sinuous and denticulate, toothed out at middle; discal dots forming a slight blackish bar; terminal dots obscure. Hind wing translucent with a little gray, like obnupsella. 21 mm.

June. (Not seen.) New York.

4. M. bistriatella Hulst. Fore wing dark brownish gray, almost even; lines whitish, diffuse; antemedial line somewhat outwardly oblique, and widening to the inner margin; base dusted with white; postmedial line diffuse, nearly parallel to outer margin, distinct; discal dots of the ground color, large, defined by white dusting. Hind wing like that of M. obnupsella. 20 mm.

New York to Florida, Wisconsin, and Colorado.

5. M. alatella Hulst of California has been reported from New Jersey, probably in error for some species of Homœosoma. It is powdery gray, with dentate antemedial and zigzag postmedial lines; outer part of cell whitish, with a dark shade below it.

6. M. ceratoniæ Zeller is injurious to dried figs and other stored food, in the Old World, and is to be expected in our territory. It is gray, with the usual pale lines, shaded with blackish; the inner line straight except for a large inward angle on **A**. 22 mm.

The last two species have more pointed wings than the typical group.

Rhodophæa exulella Zeller approaches our area on the south. It is near Aerobasis, but without the modified male antennæ. It is gray with crimson base; black discal dots; antemedial line pale and far out; and postmedial line heavily edged on both sides with fuscous.

75. ACROBASIS Zeller

(With Mineola Hulst)

Male antennæ with scape enlarged, with a pointed process or angle on inner side (fig. 389), sometimes produced as a short spur; shaft pubescent in male, sometimes sinuous at base, and slightly thickened with scales, often normal. Fore wing normal in venation, often with a heavy raised scale-ridge a short distance before the antemedial line, the space between it and the antemedial line often tinted with red. Hind wing (fig. 378) with discocellular vein of type (2), meeting the lower side of the cell at a sharp angle, but not always opposite the origin of Cu_i . Under side in many species with patches of black sex-scaling in the male. Palpi as in Myelois.

Acrobasis is a very difficult genus to work with. Several of the species are misidentified more often than not, and some may not be correctly placed in the analysis below.

Key to the species

- 1. Fore wing with a transverse antemedial scale ridge.
 - 2. Fore wing of male, beneath, with a small black dash at base of costa (above Sc), or a larger streak not reaching the base.
 - 3. Hind wing with black sex-scaling beneath.
 - 4. Hind wing with two black streaks (on R and Cu, sometimes partly confluent).
 - 5. Patch on **R** extending from base to beyond middle of wing, the outer half rather thicker.....l. angusella.
 - 5. Patch on \mathbf{R} not connected to base, or with the basal half represented by a very fine line; streak on \mathbf{Cu} heavier.....2. demotella.
 - 4. Hind wing with the streak on **R** only.
 - 5. Fore wing with a streak on R from base to middle....3. minimella.
 5. Fore wing with a short bar at base only.
 - 6. Black streak on hind wing from base to beyond middle..4. eliella.6. Hind wing with an oval spot at middle of costa.

5. aurorella.

- 3. Hind wing without black markings below in male.

 - 4. Fore wing with a short subcostal bar at base only. 5. Fore wing whitish to about three-fourths way out on the costa, cross-
 - ing the antemedial band 10. kcarfottella. 5. Fore wing whitish at base only, out to the antemedial band, or all
- 2. Fore wing without any sex mark below.

3. Venation normal in both sexes.

- 4. An oblique white band from middle of wing beyond antemedial line to costa before postmedial line, containing the contrasting black discal
- costa.
 - 5. Thorax, base of wing, antemedial area, and postmedial area toward inner margin all shaded with red. Larva on hickory

12. latifasciella.

- 5. Red practically confined to antemedial space; sometimes replaced with chocolate brown, or absent.
 - 6. Larva on Comptonia; postmedial line almost obsolete; antemedial space usually bright Indian red in northern form.

15. comptoniclla.

6. Larva on alder; antemedial area usually bright brick red; postmedial line distinct but a little diffuse; discal dots distinct.

14. rubrifasciella.

- 6. Similar; larva undescribed 18. irrubriella.
- 6. Larva on birch; moth like rubrifasciella, but usually with the antemedial space deep brown; and the dark areas of the ground deeper brown, with distinct purple iridescence......16. betulella,
- 6. Larva on hazel; moth nearly uniform gray, not shaded with white: antemedial space somewhat browner......17. coryliclla.
- 1. Fore wing smoothly scaled, the tuft replaced by a blackish first antemedial line.
 - 2. Antemedial area with a red band.

2. Aptemedial area gray-brown and white only.

- 3. Dark gray, the discal points separate, in a contrasting white patch.
- 3. Lighter; the discal spots not in a small patch, but often in a broad whitish area.
 - 4. Antemedial lines starting from a heavy black triangle on costa; larva

I., Fore wing with an antemodial scale ridge (Acrobasis).

* Male with black sex scaling beneath.

1. A. argusella Grote. Head. thorax, and hase of fore wing pale reddish. Scale ridge partly blackish. Antemedial line excurved, oblique outward to inner margin. Median area dark grav; marginal area shaded with reddish. Diseal dots separate. darker: postmedial line fine, pale, excurved in middle, and indented in discal and submedian folds. Hind wing smoky, somewhat translucent. 22 mm.

Larva eating into growing points of hickory in the early spring, causing them to wither. Moth in June.

New York: West Farms? (type), Brooklyn.

This species has been called *nigrosignella*, and the species here discussed under the name of *cliclla*, *angusella*; but in each case the sex-scaling of the under side of the hind wing was mentioned in the original description, making the present determination fairly certain.

2. A. demotella Grote. Similar to A. angusella; head white, thorax very pale; base of fore wing contrastingly paler than median area. Tuft gray, the gray area shading into the red about it. Antemedial band outwardly oblique, light red followed with cream color. Median area less mottled than in A. angusella; discal dots obscure; postmedial line fainter and marginal area less tinted with red. 20-24 mm.

Larva on black walnut, killing the buds as A. angusella does; dark olive brown; head, cervical shield, and tubercles shining dark brown. Moth in June and July.

Ontario to North Carolina, Pennsylvania, and Missouri. New York: Ithaca. West Farms.

This species is regularly correctly determined.

3. A. minimella Ragonot. Front and palpi reddish; thorax dark fuscous. Fore wing heavily overlaid with blackish on a light gray base; the blackish gathering in a large triangular costal patch beyond the antemedial line, which is pale gray, preceded by a black-centered, reddish ridge. Discal dots distinct; postmedial line obscure. 13-16 mm. (& nigrosignella Hulst.)

August.

Texas; also reported from North Carolina.

I suspect that northern records under these names are based on other related species.

4. A. eliella Dyar. Gray, powdered on a white base. Thorax and base of fore wing shaded with red; tuft blackish, followed by a broad orange-red and white antemedial band, followed in turn by a slight blackish shade on the costa. Discal dots joined; postmedial offset out in the middle, and finely denticulate, followed by a red shade concolorous with the antemedial one, except at the apex, where there is a weak oblique gray shade. 18 mm.

June and early July; August.

East River, Connecticut; New Brighton, Pennsylvania. New York: Ithaca.

5. A. aurorella Ely. Head and thorax gray. Basal third of fore wing blackish, strongly contrasting, its outer boundary but slightly oblique outward, reaching inner margin only a third way out. Outer part of wing pale gray, a little shaded with pinkish, and darkening to the outer margin. Discal dots small, blackish; postmedial line faint, paler. 20 mm.

July.

New York: Ithaca, Ilion.

6. A. stigmella Dyar. Purple-gray, much darker than A. aurorella. Head and middle of collar white. Scale-ridge slightly darker, followed by a faint reddish shade. Discal dots double. Postmedial line faint, paler in a darker shade. Disc of hind wing, below, overlaid with yellow scales. 18 mm. May to August. Caterpillar a bud worm on hickory; gray green, with black head,

cervical shield, and true legs; and with brown tubercles and anal plate.

East River, Connecticut; Fort Lee, New Jersey. 7. A. caryivorella Ragonot. Dark blue gray, rather even, without any reddish or yellowish shading. Lines diffuse; paler, defined with dark, especially toward the median area; the antemedial bent at the middle. Discal dots strong, separate. 23 mm.

The caterpillar is found on hickory in early spring; at first working in a petiole, then in the top of a twig. It lives in a frass tube, the earlier, slender portion of which is serpentine; and transforms in a large oval cocoon.

Caterpillar dark gray or greenish black, with granulose red brown head and yellow-brown cervical shield, and a large black and red subdorsal plate (ib) on the mesothorax. Moth in June.

East River, Connecticut, to Illinois, Missouri, and Texas.

8. A. caryæ Grote. Dark fuscous; thorax concolorous; base of fore wing some-what paler. A slender pale-pinkish antemedial band beyond the scale ridge, but no other pinkish or yellowish shading. 18 mm. June. Caterpillar a bud worm on Carya porcina in May, living in a slender

frass tube; greenish gray, with dark brown head, shining yellow-green cervical shield, and light brown subdorsal plate on mesothorax.

Illinois. "New York" (Grote).

The original description and Ragonot's notes on the type make no mention of sex scaling. As determined in the Barnes collection, the male has a short black bar below the costa, followed by yellow scaling, and a streak the whole length of the cell, its base overflowing the cell and touching A. The species should be bred again from the characteristic larva.

9. A. hebescella Hulst. Dull yellowish gray-brown; head and thorax concolorous; scale ridge black; the other markings obscure. 18-20 mm. Larva on oak (apparently in a folded leaf), pupating in a large egg-shaped

cocoon.

East River, Connecticut, to Texas.

10. A. kearfottella Dyar. Similar to A. palliolella, with a large whitish basal area, extending across the antemedial line to the middle of the wing, giving a suggestion of A. indiginella, but without the black antemedial costal triangle. Some reddish scaling about the scale ridge.

July and August. Larva leathery, brown; in a somewhat flattened case of grayish silk with some pellets of frass. Cocoon at the end of the case, oval, 20 mm. by 6 mm. by 4 mm. Food, hickory.

Distribution uncertain. New York: Ithaca.

11. A. palliolella Ragonot. Dark mouse gray, becoming blackish on the middle of the costa. Head, thorax, and base of fore wing typically white, somewhat shaded with some buff or orange shading; the outer boundary nearly straight and strongly oblique, to middle of inner margin. In a variety which commonly passes for nebulella the base is nearly concolorous, leaving a reddish antemedial area on the inner margin. 20 mm. (juglandis LeBaron; albocapitella Hulst, nebulella Hulst, etc., not Riley).

July. Larva on walnut and pecan.

Ontario to Florida, Illinois, and Texas.

The gray form of this species may be A. sylviella Ely, which is unknown to me.

** Male without black sex-scaling below.

12. A. latifasciella Dyar. Superficially exactly like A. eliella, but without sex scaling below. 19 mm.

This species has passed for caryx in some collections.

June and early July; August. Larva on hickory. New Brighton, Pennsylvania; Maryland; Virginia; Missouri.

13. A. normella Dyar. Pale gray with a slight reddish suffusion; scale ridge large and dark, with a slight pale reddish shade beyond it; a diffuse antemedial costal gray triangle; costa and cell shaded with white before and beyond it, as far out as the postmedial line, which is normal; discal dots black, contrasting, separate. 19 mm.

End of July and early August. East River, Connecticut.

14. A. rubrifasciella Packard. Closely similar to A. comptoniella, but on the average a little smaller, narrower-winged, with rather brighter markings, and more frequently (but not always) with a red antemedial band. 21-24 mm.

Caterpillar brown with a rosy tint; with rough darker head, and shining black mesothoracic plates; on alder and hazel, living between two or three leaves in a conical case of frass lined with silk.

Distribution unknown. Indistinguishable except by breeding.

15. A. comptoniella Hulst. Fuscous gray, rather lighter and bluer toward the base, and on the costal half of the median area. Tuft blackish-brown, followed by a dull Indian-red shade, and sometimes by a pale antemedial line. Discal dots separate and black; postmedial line faint. 25 mm.

Caterpillar blackish, with broad diffuse pale reddish dorsal and lateral bands; head dark red, black in the sutures; cervical shield pale red, contrasting; feet black. It feeds on sweet fern and on bayberry; in an oval case with a soft slender neck; lined with silk and covered more or less with bits of leaves, etc.

Maine to New Jersey. There is a race, or closely related species, on Myrica in Florida. This form is doubtfully distinct from A. rubrifasciella, and determinable only by breeding.

New York: Rhinebeck.

16. A. betulella Hulst. Similar to A. comptoniella. General wing surface shaded with reddish and dusted with white, but without a red antemedial band.

End of July. Caterpillar on black and white birch, in a case similar to that of comptoniella but almost wholly of silk.

Maine to New York; Colorado; California. Probably a food variety of A. comptoniella. New York: Ithaca.

17. A. coryliella Dyar. Gray, not shaded with white, nearly even; antemedial band slightly browner, pale-edged, with a darker shade beyond it at costa. Postmedial line a little more distinct, finely crenulate.

June to August. Larva in the usual case on hazel (Corylus); pupa in a dense hard oval ichneumon-like cocoon.

18. A. irrubriella Ely. Similar to A. latifasciella; head and thorax not so reddish; band beyond the scale ridge narrow; only slightly marked with orangered; outer line more even, not followed by an orange-red shade. 19 mm.

July. East River, Connecticut.

19. A. malipennella Dyar. Male fore wing short and broad, with distorted venation and m-cu very long. Gray thorax tinged with copper-red; scale ridge short and black; a white triangular shade in median area, as in A. normella, containing a single black discal dot. 12 mm.

East River, Connecticut.

This type is unique, and may be a monstrosity of A. normella.

II. Fore wing smooth-scaled (Mineola).

20. A. tricolorella Grote. Wings irregularly black-dusted on a white base, appearing mottled gray. Antemedial line black, forming a triangular shade on costa; a triangular reddish patch before it on inner margin, deepening to black, basally, and preceded by a white line. Discal lunule contrasting, black, in a pale shade; a darker medial shade joining the postmedial line at the discal fold; post-medial line pale, edged within with black, and followed by a chestnut band. A blackish apical streak to costa. Head and collar dark red; thorax grayer.

Quebec and Maine to California. New York: Catskills. 21. A. rubescentella Hulst. Gray, shaded with reddish, with two inconspicuous darker discal dots, and a reddish antemedial band. 25 mm.

Tennessee.

22. A. amplexella Ragonot. Blackish gray, with a large well-defined white patch at middle of costa, containing the sharply defined black discal points. Antemedial line single, diffuse, white, oblique outward; postmedial black, indented in the discal fold as usual, followed by white scales, and with a little white scaling just before the margin. Hind wings and body browner. 15 mm.

Quebec to Texas.

23. A. vaccinii Riley. Similar to *M. indiginella*; rather smaller; under side of costa of male with yellow sex scaling. Upper side more contrastingly marked, with considerable white on costa, and inner autemedial band heavily black, the costal black antemedial spot not so neatly triangular. Discal dots well separated. Red shading more diffuse than in *A. indiginella*. 16 mm.

Red shading more diffuse than in A. indiginalla. 16 mm. August. Larva variable in color, with shining yellow head, and paler cervical shield and anal plate; in berries of Vaccinium (blueberries and huckleberries); often injurious. The moth flics over the bushes, and is easily flushed in the daytime.

Massachusetts to Texas.

24. A. indiginella Zeller. Pale gray, dusted on a white base; a broad antemedial black triangle on costa, with two fine divergent lines running from it to the inner margin, enclosing a red-shaded area. Base also shaded with red. Median area shaded with dark gray below, and toward the postmedial line; which is pale, defined with dark gray and moderately irregular. A black discal bar. in variety nebulella Riley broken into two dots. A small triangular subterminal black shade at costa. 17-22 mm. (*Phycita nebulo* Walsh). (H. p. 409 f. 228-229.)

Caterpillar dull olive, with dark red-brown head and blackish shields; webbing the leaves together in the spring, and living in a frass tube. The young caterpillar hibernates. The species is sometimes injurious to apple, and eats most other Rosales also.

Moth in July.

Generally distributed. New York: Ithaca, Big Indian Valley, Schenectady, Albany.

76. ULOPHORA Ragonot

Similar to Dioryctria. Antennæ simple, a little thickened toward the base, not curved; palpi upturned beyond vertex, maxillary palpi in the male rough. flatly scaled, compressed against the smoothly scaled front. Fore wing with a scale-tuft before the antemedial line.

1. U. groteii Ragonot. Fore wing brown at hase, shaded with dark gray-green. especially beyond the tuft; outer part light gray, powdery, heavily shaded or overlaid with blackish; lines concelorous. heavily edged on both sides with dark slate grav; the antemedial convex, at the middle of the wing, the postmedial strongly bent out at middle, concave above and below, located far out: raised tuft very heavy, blackish; discal dots black, the lower one contrasting. Hind wing of male bright copperv luteous, with dark margin, of female, dull and darker. 18 mm. July and August.

North Carolina; Florida.

2. U. tephrosiella Dyar. Base powdery light gray, shaded with dark gray: a brown band before the antemedial line, which is at the middle of the wing, and is paler, followed with blackish; outer half deep gray, with paler normal postmedial line, both lines defined with blackish. 15 mm.

Larva on Tephrosia.

North Carolina.

U. brunneella Dyar, also from North Carolina. the area before the antemedial line is black.

77. DIORYCTRIA Zeller

(With *Pinipestis* Hulst)

Male antennæ nearly straight, usually with a little raised scaling on the inner side at the base of the shaft; sometimes serrate; scape rounded, curved, with a slight scale tuft only. Palpi upturned to beyond vertex; maxillary palpi cylindrical, porrect, rough-scaled. Fore wing normal: in group Pinipestis with raised scaling, normally forming antemedial and medial rough ridges. Hind wing of type 3a (fig. 379); discocellular approximate to lower side of cell from near origin of Cu_2 , or just beyond. Sc and R approximate.

The larvæ (fig. 393) so far as known are all borers in Coniferæ, usually under the bark of the twigs, or in the concs; causing more or less exudation of pitch. Sometimes they feed externally, concealed in a tube formed of pitch and frass. Several are serious pests.

Key to the species

1. Fore wing smoothly scaled (Dioryctria) 4. reniculella, 5. abietella.

1. Fore wing with raised scale ridges or tufts; gray shaded with reddish (Pinipestis).

- 2. Postmedial line with indentations in discal and submedial folds nearly equal; small with slight median scale ridge (18 mm.).....3. pygmxella.
- 2. Postmedial line with discal notch slightly deeper; a very heavy median scale ridge; large (25 mm.).....l. zimmermanni.
- 2. Postmedial line deeply toothed in opposite cell; antemedial line reaching margin at middle of wing, strongly oblique (25 mm.).....2. clarioralis.

1. D. zimmermanni Grote. Gray, shaded with reddish, especially toward the base, and with blackish; the two scale ridges blackish, the median one very heavy. Lines pale gray, defined on both sides with blackish, more strongly toward the median area. Antemedial line zigzag, hardly farther from base on inner margin than on costa; discal dot pale, diffuse, but contrasting; postmedial line denticulate, moderately angled in opposite cell, and very slightly so on fold. Terminal space rather paler and more gray; terminal line somewhat broken, black. 28 mm.

August.

Larva injurious to pine, working by preference under the bark of small branches, causing the pitch to ooze out. Cocoon usually formed in a mass of pitch. Sometimes seriously injurious. July. Doubtless there is a second brood in the fall, emerging the following spring.

Hampton, New Hampshire, to Pennsylvania. New York: reported from Oswego County, Gowanda, Cheektowaga, Hamburgh, Clarence Center, Buffalo, Schenectady, Karner, Hastings.

2. D. clarioralis Walker. Similar to *D. zimmermanni*; antemedial line strongly oblique, only a little sinuous, and not angled on fold, reaching inner margin at middle; outer line with tooth in discal fold almost as deep as wide, merely dentate below. Discal spot obscure. 28 mm. (zimmermanni Hulst, not Grote.)

"United States" (Walker). Seen only from Florida, and probably confined to the South.

3. D. pygmæella Ragonot. Similar to *P. zimmermanni*; much smaller, median scale ridge hardly raised; a distinct blackish shade between the discal bar. which is pale, and the costal end of the outer line; antemedial line zigzag, erect, preceded by a broad deep orange band toward the inner margin. 18 mm.

North Carolina; Florida.

4. D. reniculella Grote. Ash gray, crisply powdered with black on white, with a slight brown tint to the darker parts; especially in rubbed specimens; hase and terminal space paler; antemedial space solidly dark gray; the lower half of the median area also with a dark patch. Lines whitish, somewhat dusted with gray, heavily defined with dark gray; antemedial line erect, somewhat dentate; post-medial line diffuse, notched in the discal fold, and slightly concave below; discal dot a distinct upright oblong spot. 25 mm. (abietella of Hulst, in part.)

Larva injurious to spruce: its habits much like D. zimmermanni; very often working in the young cones. Moth in August.

White Mountains, New Hampshire, to Colorado and south. A variety in Florida. New York: Ithaca.

5. D. abietella Fabricius. Closely similar to D. reniculella, but averaging a little larger. 25-28 mm.

Distribution uncertain; well-known in Europe, where it has the habits of the preceding.

Two broods, the second perhaps partial. Our records are partly based on the preceding species; part are correct. 1 can give no tangible differences between the two species.

78. MONOPTILOTA Hulst

(Dioryctria, in part)

Similar to Dioryctria; maxillary palpi small, but slightly plumose; male antennæ with scape enlarged, shaft strongly curved at base, the curvature filled with scales on the inner side, unipectinate on outer side nearly to apex. Fore wings smooth.

1. M. nubilella Hulst. (Lima-bean vine borer). Fuscous, shaded with whitish, especially over and beyond the cell, and somewhat streaked with blackish; antemedial line indicated by blackish streaks; postmedial line normal, very faint, pale; discal dots a little lengthened, not contrasting. Hind wing translucent in male, dark in female. 23 mm.

Larva producing gall-like swellings in the stems of lima beaus, usually about two or three feet above the ground. Stout, blue-green, with a pinkish overlay on the dorsum; head brown, shading into black on the mouth parts; cervical shield olive brown; anal plate pale yellow, with four black dots. Tubercles pale. Pupa in the earth. Larva mature in July; moth in August and September. There is a partial second brood late in the fall.

Maryland to Florida and Arizona.

79. GLYPTOCERA Ragonot

(*Nephopteryx*, in part)

Male antennæ with shaft toward base slightly hollowed, forming a longitudinal groove, edged on both sides with scales. Labial palpi normal; maxillary palpi quite large, rough and thick; appressed to the frontal tuft. Venation normal; hind wing of type 3b.

1. G. consobrinella Zeller. Tuft represented by a few raised seales. Fore wing pale ash gray; base whitish, more or less overlaid with Indian red; a strong blackish shade before the antemedial line; which is pale, bidentate irregularly, and defined with black, two-fifths way out. Postmedial line similar, somewhat sinuous and crenulate, at four-fifths the length of the wing; a heavy black discal bar, with some blackish and dull red shading below it. 20 mm.

End of May to August. Quebec to Texas. New York: McLean.

80. TACOMA Hulst

Wings normal; male antenna with a curve in the shaft, filled with a large scale tuft; scape simple. Palpa upturned to vertex; maxillary palpi large, filiform. Fore wing with M_2 and M_3 stalked; discocellular vein bent before origin of Cu_2 .

1. T. nyssæcolella Dyar. Powdery gray; antemedial line whitish, offset a little on Cu. with a more evenly gray patch before it on inner half, and beyond it on costal half, the rest of the line defined with dark; a pale shade beyond it on inner

margin. Postmedial line pale, defined with dark, normal, with a large dark patch

before it on inner margin. Hind wing paler. 20 mm. May; July and August. Larva black, with a yellow head; in a loose web con-taining suspended frass, in a folded leaf of Nyssa.

Southern New Jersey to western Pennsylvania.

81. POLOPEUSTIS Ragonot

Male antennæ rough and obscurely dentate, pubescent; palpi relatively short, upturned, not exceeding the vertex; first two joints long-hairy, the third short, and cylindrical; maxillary palpi slender. Fore wing more triangular than usual, with \mathbf{M}_2 to \mathbf{Cu}_2 free and parallel; hind wing with moderate cell (\mathbf{Cu}_2 at two-fifths), \mathbf{M}_2 and \mathbf{M}_3 very long-stalked, angle of discocellular between \mathbf{Cu}_1 and \mathbf{Cu}_2 . Moth diurnal.

1. P. annulatella Zetterstedt. Black, dusted with white, appearing light gray; lines pale, defined with dark, converging toward inner margin; antemedial line nearly straight; postmedial irregular. Hind wings very pale brownish, darker in female. Male 24, female 20 mm.

Arctic Eurasia; Altai; Labrador (?).

The Labrador specimen, determined as annulatella in the Hulst collection, is a Nephopteryx, and apparently a small eastern race of N. fasciolella.

82. AMBESA Grote

Like Nephopteryx, male antennæ flattened and slightly thickened at base, but without the notch and tuft of scales.

1. A. busckella Dyar. Powdery gray, slightly reddish, especially at base and on thorax. Base pale, followed by a diffuse blackish band, wider on the inner margin, the band lying before the antemedial line toward the inner margin and beyond it at costa, sometimes obliterating it. Lines doubled, dentate, antemedial line fully two-fifths way out, and farther out on inner margin; postmedial line well out, normal, its blackish edging somewhat wider on costa. A black reniform, and some blackish postmedial shading. Hind wing translucent at base. 22 mm.

July.

Maryland; western Pennsylvania.

83. NEPHOPTERYX Hübner

Male antennæ simple or serrate, with a more or less developed curve in base of shaft, filled with a mass of large scales. Front swollen, rough-scaled; palpi upturned beyond vertex, normal, the maxillary palpi normal, or with a flat tuft of scales applied to the face as in Ulophora. Wings smooth-scaled. Fore wing with R_2 free in our species, M_2 and M_8 separate but rather approximate, not in line with Cu. Hind wing with Sc and R closely parallel; M_1 stalked, M_2 and M_3 stalked; dcv of type 3b, becoming approximate to lower side of cell at or beyond origin of Cu₂, which is at two-fifths the length of the wing. Prothorax below with large scales, as in Ambesa, but unlike Dioryctria.

Key to the species

1. Ground dusted with white on clear black, appearing bluish ash gray.

- 2. Each scale with a white base and a fine white tip; apparently very finely
- 1. Ground fuscous, dusted with black, and broadly white-tipped, scales.

1. N. ovalis Packard. Tuft on male antennæ twice as large as scape in front view. Powdery ash gray, rather coarse; the antemedial space typically, but not Wew. Powery as gray, rather coarse, the antemedial space typically, but not always, brown or yellowish; antemedial line waved, defined on inner side by the brown antemedial area, on outer side mostly by a blackish bar at costa and dots on veins; postmedial line normal, pale, only partly defined with blackish before it and fuscous beyond, but with heavier black edges at costa; some blackish shading near middle of wing; two black discal dots. Hind wing fuscous, not paler. 25 mm. Late July and August.

Maine to California. New York: Rhinebeck, Delmar.

Variety geminipunctella Ragonot, with ground largely white, was described from Washington.

N. fascicolella Hulst, described from British Columbia, is similar to N. ovalis. but without any brown shading, and with only a little blackish shading along the antemedial line. Tuft on antenna no larger than scape. 30 mm. A small form of this (22 mm.) appears to occur in Labrador, and has been mistaken for Polopeustis annulatella.

2. N. rhypodella Hulst, another western species, almost evenly and very finely powdered black and white, with traces of markings; has been reported from Illinois.

3. N. inquilinella Ragonot. Similar to N. modestella; the markings more diffuse, antemedial band nearly erect and even. 18-24 mm.

Larva in willow galls (Salicis nodum).

Wisconsin.

4. N. modestella Hulst. Tuft on antenna large. Fore wing fuscous, somewhat dusted with white, especially about the end of the cell, and with a few black scales, antemedial line pale, diffuse, well out, reaching the inner margin almost at the middle of the wing, twice dentate, defined with blackish on outer side, especially on the veins, and with a broad brownish band before it. Discal dots black, distinct but not strong; postmedial line pale, normal, hardly defined with dark. Outer part of wing slightly reddish shaded. 20 mm.

Massachusetts.

5. N. rubrisparsella Ragonot. Ochreous brown, more or less suffused with pinkish; base solidly light; antemedial region suffused with blackish; antemedial and postmedial lines luteous, black edged; median area more or less shaded with black; discal dots black, strong; terminal space evenly pale, with black terminal dots, and a line in the fringe. 18 mm.

The typical form, from Texas, has the blackish shadings replaced with lighter ash gray. I have not seen the male. August. Larva on Celtis.

Jefferson County, West Virginia; Texas.

84. TLASCALA Hulst

(Nephopteryx, in part)

Similar to Nephopteryx. Fore wing with a more or less developed ridge or scale-tuft before the antemedial line, sometimes with other markings more or less raised.

Key to the species

1. T. finitella Walker. Dark powdery gray, somewhat brownish; antemedial line raised, blackish, obscure; tuft heavy, blackish; a series of small median blackish tufts; discal dots black and tending to be confluent; postmedial line normal, pale, defined with dark, not conspicuous. A broken black terminal line as usual. Tuft on antenna very large, much more than filling the notch. 25 mm. May.

Nova Scotia to Florida and Colorado.

2. T. reductella Walker. Wings pale gray, in part often almost white; postmedial line slightly paler, normal, dentate, defined by slightly darker gray on outer side. Base of wing and thorax brown (in variety gleditschiella Fernald, mouse gray); the tuft black, and not strong. Antemedial line heavy, black, outwardly oblique, and a little wavy, preceded by a pale line; discal dots black; no raised median tufts. 22 mm.

The larva webs together leaves of Gleditschia in September, emerging in late fall or the following May. Cocoon in rubbish on the ground.

New York to Ohio. New York: New Windsor.

85. MEROPTERA Grote

Like Salebria, our species with \mathbf{R}_2 stalked, \mathbf{M}_2 and \mathbf{M}_3 clearly stalked; wings smooth-scaled.

1. M. pravella Grote. Ash gray, base contrastingly pale gray, with an even, slightly oblique outer boundary, followed by a strong blackish shade. Basal angle reddish. Discal dots blackish, tending to fuse, in a paler area; postmedial line pale, defined with darker gray, distinct but not contrasting. Terminal line continuous, black, weak. 20 mm.

March to May. The larva is said to be brown, with dorsal and lateral green lines, plain brown cervical shield, and head green, brown, or reticulate with black; in a web between two leaves of willow, or on *Rhus Cotinus*, in September.

Maine to British Columbia and Texas. New York: Lancaster, and New Windsor.

2. M. unicolorella Hulst. Mouse gray, slightly violaceous; markings obscure, but normal so far as traceable; antemedial line darker; postmedial paler, both diffuse. 22 mm.

Late May to August.

Montreal, Canada, to Florida and Texas (not Washington, as stated by Ragonot). New York: Ithaca.

3. M. uvinella Ragonot, as described, is paler than M. pravella, with the postmedial line white, clear, and straighter in the middle part. 16 mm.

United States.

Probably a mere variant of *M. pravella*. *M. cviatella* Dyar appears to be a Salebria.

86. SALEBRIA Zeller

(Meroptera, in part; with Pempelia)

Similar to Nephopteryx. Maxillary palpi yellow, plume-like, enclosed in a groove in the second joint of the upturned labial palpi. Fore wing normal; hind wing (fig. 381) with moderate cell, Cu_2 apparently from angle. Wings smooth in our species. Antennæ in our species smooth beyond the tuft.

Key to the species

- 1. Antemedial line followed by a pale patch on inner margin (obscure in some specimens of contatella).

 - 2. Basal area contrastingly darker gray, at least toward the inner margin.

2. annulosella.

2. Markings very clean-cut; extreme base blackish, with a red shade before the antemedial line
2. Whole basal area reddish
2. Base mostly light gray, reddish at basal angle
1. Antemedial line not followed by a pale patch on inner margin.
2. Base distinctly paler.
3. Base yellow or reddish.
4. Followed by a strong antemedial line, outer part blue-gray.
5. Costa and inner margin broadly brown11. cviatella.
5. Outer two-thirds entirely gray 10. basilaris.
4. Antemedial line obscure,
5. Base mostly yellowish
5. Base yellowish toward inner margin only
3. Base lighter gray without any yellow or brown.
4. Antemedial space blackish
4. A double black antemedial line toward inner margin only.
13. purpurella.
2. Base not paler than outer part of wing.
3. Antemedial region, and dark shades defining the postmedial line, strongly
shaded with crimson12. carneella.
3. At most, with crimson streaks between the veins.
4. Pale antemedial line strongly waved on fold and on A5. contatella.
4. Antemedial line almost straight on lower half of wing4. afflictella.

1. S. turpidella Ragonot. Powdery gray, the lines hardly paler, sharply edged on both sides with black; the first line zigzag, normal, the second normal, rather more irregular than usual. Discal bar continuous, black; a blackish median shade below it toward inner margin; terminal line continuous, black; a slight brownish shade in the fold before the antemedial line. 18 mm.

Massachusetts and Colorado.

Distinguished from all other species in the genus by its absolutely clean-cut markings.

2. S. annulosella Ragonot. Mid-tibia blackish with a contrasting pale tip, basal segments of abdomen transversely banded with black. Scale tuft on antenna twice as long as scape. Fore wing light gray, with blackish shades beyond the antemedial line at costa, before it at inner margin, over costal end of postmedial line, and sometimes covering most of the base; antemedial line obscurely paler; postmedial faint, normal, preceded by a slight darker shade. Discal dots separate, black, contrasting; terminal dots separate. 18 mm.

The pale patch beyond the antemedial line is often obsolete in light specimens, but the species differs from *afflictella* in the nearly obsolete, diffuse, antemedial line, and the much fainter postmedial.

Massachusetts to Texas and Colorado.

Northern records are possibly in error for *S. engeli*. The form with the base shaded with brown is variety pumilella Ragonot; the type form has a blackish base, while those with concolorous base are variety nubiferella Ragonot.

3. S. engeli Dyar. Abdomen not banded with black. Dark fuscous, the markings more or less obsolescent, except for a large whitish dorsal patch over and beyond the antemedial line, sometimes continued across to the costa. Some pale scaling about the discal dots. Postmedial line traceable, pale. Tuft on antenna hardly larger than scape. 18 mm. (annulosella, in part).

May to July.

New Jersey to Texas.

4. S. afflictella Hulst. Base fuscous, out almost to the middle of the wing; crossed by a strong whitish basal line; antemedial line whitish, fine, a little waved,

followed by a clean-cut fuscous line. Median area whitish, outer third fuscous, crossed by a fine wavy normal postmedial line, the outer margin gray. Discal dots black, distinct. Terminal line black, almost continuous. Tuft on antenna more than twice as long as scape, not very broad. 16 mm. (liquidambarella Dyar).

The fuscous areas all show a slight reddish iridescence.

Larva on sweet gum.

Northern New Jersey to Florida.

5. S. contatella Grote. Middle tibiæ pale, with a dark annulus toward tip. Fore wings powdery gray, base shading into reddish near basal angle, median area typically suffused with mouse gray, leaving a vague pale patch on inner margin beyond the antemedial line; in light specimens, concolorous with the base, with only a vague gray median shade from the discal dot to the inner margin. Antemedial line whitish, zigzag, preceded by a heavy blackish shade and followed by a fine blackish line, thickened on Cu and A; or reduced to black wedges on Cu and A; postmedial line normal, doubly defined with darker gray; discal dots a little diffuse and tending to fuse. A broken black terminal line. Ordinary lines often nearly obsolete. Hind wing pale yellowish. 20-28 mm. (Probably subcasiella Clemens.)

In variety quinquepunctella Grote (virgatella Clemens) there are brown, reddish, or yellowish streaks between the veins, cutting up the blackish markings into dots or streaks on the veins.

May to early July; August. Larva green, head black or reticulate with brown; cervical shield black or spotted with black; body turning red before pupation. On locust and wistaria between two leaves, in June and July.

Maine to West Virginia and west to Colorado; the variety with as wide a range

as the type form. New York: Peru, Ithaca, Albany, New Windsor. 6. S. vetustella Dyar. Gray, somewhat paler on the inner margin beyond the antemedial line. Base dull orange-brown; a large blackish fascia before the antemedial line, narrow on costa, separated from the brown base by a narrow pale band. Antemedial line outwardly oblique, a little dentate, pale filled, its inner line fused with the black antemedial space. Wing beyond cell a little reddish, with an obscure normal, pale postmedial line, two darker discal dots, and nearly continuous terminal line. 22 mm.

New York and Maryland to Illinois.

7. S. celtidella Hulst. Ground light ochreous, the whole middle half suffused with fuscous gray; base toward inner margin shaded with tawny, followed by a luteous shade; antemedial line fully out to middle of wing, pale, defined with blackish, abruptly offset in, and defined by black dots on each vein. Discal dots black, well separated, the lower one in the upper part of a vague area of the pale ground color; postmedial line irregular and deeply dentate, defined on inner side by black dots and on outer by strong black wedges; terminal space contrastingly pale; a strong broken black terminal line. 18 mm.

August. Larva joining leaves of Celtis; not forming a tube. Body striped evenly in two shades of green; head and cervical shield piceous black, with two whitish stripes on each; tubercles black.

New England and central Illinois to Florida.

8. S. lævigatella Hulst. Almost even fuscous, slightly powdery, with a few white scales at lower angle of cell; antemedial line indicated by a double or triple blackish shade on inner margin at one-third way out, separated by whitish scales; base before it reddish; discal dots, postmedial line, and terminal line obscure. 25 mm. Male not seen.

Quebec; Massachusetts; Wisconsin.

9. S. semiobscurella Hulst. Slightly smaller than Meroptera pravella, but practically identical in markings; the blackish shading a little lighter, and the antemedial line usually distinct. Base of inner margin distinctly reddish. 20 mm.

July. Larva on sumac. Perhaps one of the two very different larvæ described for M. pravella belongs to this species.

New York and Texas.

10. S. basilaris Zeller. Bluish ash gray, base light wood-brown or orange, shading into blackish along the inner margin, or followed by a black shade; antemedial line strong, whitish, a little wavy, defined on outer side with a black line, fading out at costa. Outer margin shaded with brown, the brown varying a good deal in amount; discal dots and postmedial line obscure; terminal line distinct, broken, black. 25 mm.

July.

New Hampshire and northern Ontario to Colorado and Texas.

11. S. cviatella Dyar. Base, costa, and inner margin broadly bright reddish brown, disc and outer margin bright purplish gray with paler veins; a broad black inner band, cut by a fine white irregular line; discal dots confluent, clouded; outer line whitish, diffuse, sharply bent opposite cell and in fold. Black terminal dashes. 22-25 mm.

Chicago, Illinois.

12. S. carneella Hulst. Powdery gray; the thorax, base of wing, antemedial band, and postmedial region suffused with crimson; median area with crimson shades between the veins; lines normal; antemedial excurved and hardly dentate, gray, defined mostly by the lack of crimson suffusion; discal dots obscure, blackish, terminal line obscure and fine. 22 mm.

Larva on willow.

Maine and Massachusetts.

13. S. purpurella Hulst. Paler powdery gray, almost wholly suffused with light red, leaving a whitish band before the antemedial line, which is pale, heavily defined with blackish on both sides, toward inner margin, and erect; outer part of costa also without red; postmedial line obscure, indicated by the lack of red shading. 24 mm.

New Mexico, the Illinois record probably in error.

87. IMMYRLA Dyar

Similar to Salebria, with an antemedial scale tuft.

1. I. nigrovittella Dyar. Dark gray, median area slightly paler; ridge deep black, farther out toward costa, not reaching costa; antemedial line just beyond it and parallel, faint; postmedial line pale, faintly bowed at middle, dark-edged and running in a darker shade. 20 mm.

May and June.

Western Pennsylvania. New York: Ithaca.

88. MYRLÆA Ragonot

Similar to Laodamia; palpi shorter, fore wing much broader, with strongly arched costa.

1. M. vetustella Dyar. Light ash gray; head, collar, and base of fore wing Indian red, contrasting; antemedial line defined on the outer side with a blackish line; within, by a heavy blackish shade, the two fusing on the costa and obliterating the antemedial line. Discal dots gray, obliquely placed; postmedial line faint, emphasized by dark shades before it on the veins. 25 mm.

May.

Ithaca, New York, and south.

89. LAODAMIA Ragonot

(*Pinipestis*, in part)

Venation (fig. 380) and habitus practically as in the genus Dioryctria; Cu_2 of hind wing just about at one-third, discocellular vein separated by a distinct space from lower side of cell, approaching it on a long slant. Male antennæ (fig. 390) and palpi about as in Salebria, but male palpi longer, and shaft of antennæ more laminate than in our Salebrias; front rougher.

1. L. fusca Haworth. Blackish, lightly powdered with gray except on veins and lines; lines gray, normal, inconspicuous; antemedial zigzag. Discal dots large, black. 28 mm.

In variety frigidella Packard, the ground is gray, black only along the lines.

Locally common in July and August, especially on burnt-over heaths. The larva eats Vaccinium and willow.

Arctic America, south to Northern New Jersey and British Columbia. New York: Waterville, Rochester Junction, Ithaca.

90. ELASMOPALPUS Blanchard

Hind wing (fig. 383) with Cu_2 arising about a third way out on the wing, the discocellular vein well beyond it, and well separated from the lower side of the cell. Palpi as in Salebria, more oblique and straighter, with well-developed plume. Male antennæ strongly flattened, with hardly any notch, but a wellmarked scale tuft not reaching the base of the shaft.

Key to the species

1. Fore wing with raised scales before the antemedial line; gray, with a

1. Fore wing smoothly scaled.

2. Markings distinct, normal, gray.....2. petrellus. 2. Markings strigose and broken; or obsolete; male light wood-brown; female dark

1. E. decoloralis Walker. Pale gray with a paler postmedial line; antemedial indicated by a few dots, with a small, slightly raised black patch before it toward the inner margin; black discal and terminal dots. 26 mm.

Not seen.

United States (Florida?)

2. E. petrellus Zeller. Head, thorax, base of fore wings, antemedial and subterminal spaces light wood-brown, the rest white, dusted and shaded with light dull gray, darker toward the inner margin; fold suffused with wood-brown in the median area; lines whitish, dentate, defined with dark gray dots, or broken dentate lines; discal dots partly confluent, darker gray, with some brown scales around them. Black terminal dots. δ 20 mm., 9 25 mm. In variety hapsella Hulst the areas normally brown are only slightly browner

than the rest.

New Jersey to Colorado, and south.

3. E. lignosellus Zeller. Male typically ochre yellow or light wood brown, less rusty than the brown parts of the last species; costa and outer and inner margins normally shaded with fuscous brown, dusted with white, but sometimes concolorous; the borders disappearing at the base of the wing; ante- and postmedial lines of a few dots; only lower discal dot distinct, black; Cu sometimes narrowly shaded with powdery fuscous. Hind wing translucent white, with fuscous border. Typical female blackish, varying in details, and sometimes with a yellow bar on the disc. 15-25 mm.

The antenna is as in normal Salebria, with a large scale tuft, but the venation places the species here, and the palpi are even longer and more oblique than in E. petrellus.

The female variety tartarella Zeller is dwarf and wholly dark gray (carbonella Hulst). Female variety incautella Zeller is similar to the normal male. The amount of the whitish powdering on the fuscous border varies from practically none to a complete suffusion.

Southern States, south to Chili, straying north to Maine.

Pyla aëneoviridella Ragonot was described from New York in error; the type was from Wyoming.

91. EPISCHNIA Hübner

Palpi with stout oblique second segment; third segment long and porrect; maxillary palpi moderate, scaled; male antennæ, in our species, dentate, with a slight sinus at the base of the shaft. Fore wing exceptionally long and narrow. with normal venation, smooth; hind wing with discocellular closely parallel to

with normal venation, smooth; hind wing with discocellular closely parallel to lower side of cell from origin of Cu_2 to origin of Cu_1 . **1. E. boisduvaliella** Guenée. Fore wing typically pinkish ochreous, a little shaded with blackish and dusted with blackish along outer margin. Costa con-trasting, white, from just beyond base to apex, the lower boundary practically straight and running through upper discal dot. Postmedial line usually indi-cated by a few blackish striæ; lower discal dot black; the other markings obso-lete. 18-26 mm. (*farrella* Curtis, *lafauryella* Constant, etc.) In variety albocostalis Hulst, the ground is dark fuscous, heavily dusted with white toward inner margin and apex, and the white costal stripe becomes diffuse and fades out at the apex.

and fades out at the apex.

Larva russet, with a greenish tint, and with obscure lateral reddish striæ; yellow-green beneath; in pods of Leguminosæ (Ononis, Anthyllis, Lotus, Astragalus), hibernating full-grown; moth in July. New Hampshire and Massachusetts to British Columbia and Texas; Eurasia.

92. ETIELLA Zeller

Palpi with second joint very long, the lower side, and upper side except at extreme base, straight and regularly converging; third segment rather short, fine, and porrect; maxillary palpi with a plume, concealed in a groove in the second joint of the labials. Male antennæ hardly notched, with a heavy tuft of dark gray scales on inner side of shaft near base, and a tuft of finer pale hair scales on outer side, lying against it. Fore wing very narrow, with a slightly raised golden antemedial bar; hind wing ample, discocellular of type 3 (fig. 382) angulate at a point halfway between Cu_1 and Cu_2 then approximate to lower side of cell.

1. E. zinckenella Treitschke. Light gray, dusted heavily with light pinkish ochreous; a white costal stripe from base to apex, leaving the costal edge gray in the middle of the wing; antemedial band yellow, just beyond the deeper orange raised ridge; both cut off by the white costa. 18-25 mm. (exiella Treitschke, etc.).

Larva (fig. 394) in pods of Leguminosæ, sometimes injurious. Apple-green, reddish, or brown; tubercles yellow, with a black setigerous puncture; no enlarged one on mesothorax. Head amber yellow with a brown posterior line; prothorax green with three pairs of black dots, the lateral ones in a reddish shade.

World-wide, but rare in this country.

93. MELITARA Walker

(Megaphycis Grote)

Tongue short, weak, not always exposed between the bases of the palpi; antennæ not modified at base, pectinate in male, subpectinate in female; palpi massive,

porrect, the joints with separate short tufts on under side, that on the first the largest. Maxillary palpi large, flattened against the face. Fore wing rather narrow, with short erect outer margin \cdot venation normal; M_2 and M_3 stalked. Hind wing with Sc approximate to R; M_2 wanting; M_3 shortly stalked, discocellular of type 2 (meeting lower side of cell at 60 degrees; Cu_2 given off at two-fifths length of wing).

Very large species, the larva boring in the phyllodia of Cacti, and more or less social. Ours works in the prickly pear, Opuntia. 1. M. prodenialis Walker. Whitish, shaded with mouse gray, except toward the costa; antemedial line with outer component strong, blackish, with a long tooth out on fold, the inner line faint, but traceable at the inner margin; postmedial line double, blackish, fading out below, strongly dentate except toward costa. A vertical black discal bar, and black terminal dots. 27-40 mm. Female larger.

End of July.

Southern New York to Florida and Texas; on the barrens northward.

94. ZOPHODIA Hübner

(Pempelia, in part; Dakruma Grote)

Front rounded out; antennæ nearly simple; tongue strong; palpi porrect, thickly and evenly scaled; more oblique in female. Maxillary palpi simple; venation about like that of Melitara; the fore wing narrower, with more oblique outer margin.

1. Z. grossulariæ Riley. Brownish gray, whitish toward costa; antemedial line erect, dentate, forming a W; postmedial line oblique and nearly straight to M_{22} then dentate to inner margin. Wing more or less striate, especially toward base. Male much less contrastingly marked than female. 25 mm. (turbatella Grote.) (H. p. 411 f. 230.)

Larva injurious to currants and gooseberries, feeding on the berries and webbing them together.

Canada and Maine to Illinois. New York: Lancaster (VanDuzee). 2. Z. bella Hulst. Similar to Z. grossulariæ, powdery pale gray; more strigose; antemedial line represented by a diffuse dark shade, running out at middle and on inner margin; postmedial line strongly oblique with a single large tooth at M₂. 28 mm.

Massachusetts.

Typically the two lines of the postmedial line are about equal, but the inner is much stronger in the Colorado form.

95. EUZOPHERA Zeller

Palpi erect, second joint scaled, third erect and acuminate; maxillary palpi broad-scaled and appressed to front. Fore wing (fig. 384) with arched costa and rounded apex, Cu_1 well separated, M_2 and M_3 long-stalked; R_2 rarely stalked. Hind wing with Sc strongly anastomosing with R, discocellular vein moderately curved; M_2 stalked or connate with Cu_1 .

Key to the species

1. Lines, at least the postmedial one. clear and continuous.

2. First line well before middle of wing; nearly straight and vertical, almost obsolete

First line close to middle of wing, distinct, pale; the wing reddish before it.
 Median area broadly fuscous, shaded with red; 28 mm...2. ostricolorella.

- 3. Median area crisply dusted with black on white; 22 mm. . 4. semifuneralis.

3. Median area blackish, or suffused with red-brown; 12 mm.

3. ochrifrontella

1. E. franconiella Hulst. Light gray, strigose, dusted, and shaded on a white base, the costa paler; antemedial line obscure; discal spot rather H-shaped (turned on its side), postmedial line oblique, obscure, and coarsely toothed; terminal dots blackish, a little diffuse. 25 mm.

Franconia, New Hampshire.

2. E. ostricolorella Hulst. Dark gray, suffused with dull Indian red; the base mostly reddish; antemedial line at middle of wing; diffuse, pale, nearly even, a little bent out at inner margin; postmedial line rather beyond three-fourths, more erect than outer margin, pale and diffuse, somewhat toothed opposite lower angle of cell; terminal line blackish, almost continuous, discal spot obscure. 28 mm.

Larva mining in the bark of the trunk of tulip tree, usually near the ground, and only where the bark is damp; maturing in the spring. Moth from May to July.

New York (Hulst). Was described from the State. Type only seen.

3. E. ochrifrontella Zeller. Fuscous gray, the lines luteous or indicated by white powdering; ground a little powdery at end of cell and along outer margin; base and sometimes median area suffused with reddish. Antemedial line broadened and more diffuse; postmedial line clearer, a little wavy, a little more oblique than in E. ostricolorella. 12 mm.

May; July and August. New York to Iowa. New York: Ithaca.

4. E. semifuneralis Walker. Median area blackish, sometimes dusted or mottled on a white ground. Basal half and outer margin light reddish gray, heavily dusted with white, especially along the outer margin and on the costal half of the base. Lines white, powdery, wavy, normal, the antemedial line near the middle of the wing. 20-25 mm. (pallulella Hulst, impletella Zeller).

Larva under bark of peach and plum; sometimes injurious.

Massachusetts to Colorado and south. New York: Lancaster (VanDuzee), Kinderhook.

5. E. inornatella Hulst. White, lightly dusted with gray; two or three blackish dots at middle of wing representing the antemedial line, and four or five the postmedial line; two discal dots. 20 mm.

May.

Anglesea, New Jersey.

96. VITULA Ragonot

Similar to Euzophera. Front tufted; wings of male with a tuft of scales at base of costa below; M_2 and M_2 stalked; hind wing with cell short; discocellular vein not much curved; M_{2+3} and Cu_1 connate or nearly so; Cu_2 at one third the length of the wing.

1. V. edmandsii Packard. Gray, powdery, somewhat shaded with brownish; first line well toward middle of wing, oblique, a little wavy, diffuse, broad and blackish; postmedial line at four-fifths way to apex, paler, defined with fuscous, diffuse, a little irregular, and angled opposite lower angle of cell; a dark discal bar. 20 mm.

June to October. Larva (fig. 395) in bumblebee nests. Generally distributed. New York: Buffalo, Ithaca.

97. LÆTILIA Ragonot

(Dakruma Comstock)

Tongue weak; labial palpi oblique, reaching about to vertex; maxillary palpi normal; front smooth; male antennæ ciliate; fore wing smooth, with R_s free, M_s and M_s stalked, Cu_s from angle of cell; hind wing with discocellular moderately curved, Cu_s arising from near angle of cell; M_s and M_s and Cu_s long-stalked; Sc and R fused to very near apex.

Larva feeding on soft scales (Pulvinaria) on Negundo and other trees, spinning a light silken tube not unlike that of Feniseca.

1. L. coccidivora Comstock. Light gray, heavily shaded with white on costs and outer margin; antemedial line white, narrow, erect, and a little excurved with a fine gray line before and a heavy shade behind it; postmedial line a little waved, fine toward costa, heavily defined with blackish, becoming broader and defined with broader shades of light fuscous toward inner margin. Discal dots often fused, black; terminal dots black. 15–18 mm. General southward. New York: Albany.

2. L. myersella Dyar. Similar to L. coccidivora, but without gray suffusion on the inner half of the fore wing, the white reaching the inner margin. Lines clean-cut and heavily gray-edged, the antemedial more waved than in L. coccidivora. September.

Southern Pines, North Carolina.

L. fiskella Dyar, from Pennsylvania and North Carolina, is unknown to me.

98. CANARSIA Hulst

Tongue strong; palpi with second joint reaching vertex and third slender; maxillary palpi normal; front tufted; male antennæ compressed, with sinus and strong scale tuft at base of shaft. Fore wing (fig. 385) normal; Cu_1 from angle of cell, M_2 and M_3 stalked; hind wings with Cu_2 a little before angle of cell; discocellular as in Lætilia; Sc anastomosing with R. 1. C. ulmiarrosorella Clemens. Fuscous gray, more or less dusted with white, especially on the disc and outer margin. Lines double, darker gray, filled with whitehe optemodel lines well out program.

whitish; antemedial lines well out, wavy, postmedial line somewhat wavy and irregular; the outer line of the antemedial often blackish, clean-cut and contrasting; a whitish patch or shade beyond it on inner margin. Fine separate terminal dots; discal bar high and narrow. 13-16 mm. (pneumatella Hulst, ulmella Ragonot, fuscatella Hulst).

End of June to August. Larva green with paler dorsal and stigmatal lines; several segments with brown subdorsal dots; head pale brown with darker stains; found on elm and, rarely, on hickory in a silk nest between leaves, in August.

Canada to Texas. New York: Peru, North Creek, Otto, Ithaca, New Windsor. 2. C. gracilella Hulst. Similar to C. ulmiarrosorella; paler, the markings slightly more obscure.

The types from northern New Jersey, appear to be merely somewhat pale and rubbed specimens of C. ulmiarrosorella.

99. PSOROSINA Dyar

Fore wing with M_2 and M_3 separate; hind wing with Cu_2 at angle of cell, M, and Cu, stalked. Male antennæ bent, with a tuft in the bend as in the last genus; tongue moderate; palpi upturned to vertex; maxillary palpi filiform.

1. P. hammondi Riley. Wings very broad, brown-black with some white scales, the two usual lines nearly erect, fairly even and of white powdering, the postmedial fading out at the costa, where there is a powdery white patch before it. 12 mm. (Canarsia auct.)

Larva dull green or brown, with black tubercles; head pale with greenish face, cervical shield black; feeding on pear, and also bred by Miss Murtfeldt from an acorn.

2. P. angulella Dyar is unknown to me. It is described as practically like P. hammondi, but with an angulate antemedial line.

Iowa.

100. VALDIVIA Ragonot

(Maricopa Hulst)

Palpi long, porrect, second joint bowed above, maxillary palpi invisible. Tongue developed. Wings sublance late, fore wing with R_2 arising from cell, M_2 and M_3 long-stalked Cu₂ from angle of cell.

1. V. albocostella Hulst. Palpi blackish. Fore wing with anterior third whitish, with a vinous tint, somewhat mixed with dark scales, the rest fuscous, vinous toward inner margin; large black discal dots. Hind wing fuscous, paler at base; 18 mm.

August. Unknown to me. Anglesea, New Jersey.

101. HULSTIA Ragonot

Palpi as in Canarsia. Front smooth, antennæ normal. Cu, and Cu, of fore wing arched, from very near end of cell; M_2 and M_3 stalked, on a line with base of Cu; hind wing with Cu₂ from angle of cell; M₁ stalked; Sc and R anastomosing; M₂ stalked with Cu_1 . In one specimen before me M_3 is preserved, long-stalked with M2.

1. H. undulatella Clemens. Brownish ash gray, a little shaded with light brown, and powdery; ground sometimes mixed with yellow-brown; antemedial line white, fine, wavy and outwardly oblique, the fine dark gray line on its outer side thickened at costa; antemedial space broadly mouse gray, especially toward inner margin where it is preceded by some white scales. Postmedial line wavy and irregular, fine, pale, with a sharp dark gray line on inner side, and a more or less distinct broad, gray subterminal shade on its outer side. Discal dots confluent into a bar; terminal dots fine. Postmedial line not distinctly bowed at the middle. 15-20 mm.

July and August; not rare locally.

Quebec to California. New York: Niagara Falls, Kinderhook.

102. HONORA Grote

Palpi oblique to above vertex; second segment slender and evenly scaled, third porrect and rather large. Maxillary palpi short and thickened with scales at tip. Antennæ simple. Wings narrow; venation as in Hulstia.

1. H. fumosella Hulst. Dull black, shaded with ochre yellow between the veins. Lines fine, white, a little diffuse under a lens. Basal line distinct, at one-sixth way out, incomplete. Antemedial line from costa at one-third to middle of inner margin, nearly straight; postmedial line well out, nearly straight, and distinct toward the inner margin only; erect. 15 mm. July. Type only seen.

Newark, New Jersey.

103. DIVIANA Ragonot

Palpi slender, recurved, second segment tufted in front; maxillary palpi fusiform; antennæ a little curved at base of shaft in male, with teeth in the sinus. Fore wing with M₃ wanting, R₂ free, M₂ and Cu₁ from angle of cell; hind wing with cell moderate, Cu_2 arising near angle, M_2 stalked; M_1 stalked, Sc and R anastomosing.

1. D. eudoreella Ragonot, a brownish gray species, with fairly normal markings, occurs from North Carolina to Florida.

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104. HOMEOSOMA Curtis

Palpi obliquely upturned to level of vertex, or, rarely, shorter and rough-scaled; maxillary palpi slender, normal; male antennæ rather thick. Fore wing with R_2 free, R₃ wanting as well as R₄, M₂ and M₃ stalked or united, Cu₁ and Cu₂ approximate and strongly curved; hind wing with discocellular vein straight (fig. 388), a little flattened and widened, normally not tubular, Sc anastomosing strongly with R, its tip often very short; M, normally shortly stalked, M2 and M3 united, Cu, from cell. (Larva, fig. 396.)

Key to the species

1. Antemedial line indicated by two or three black dots (on R, Cu, and A).

1. electellum.

1. Antemedial line distinct, sometimes interrupted.

1. H. electellum Hulst. Powdery light gray, with a white subcostal streak; lines represented by a few darker dots, or obsolete, the antemedial dots in a vertical series when distinct; at least one dark discal dot present, sometimes both. Terminal dots dark, often obscure. 15-22 mm.

July and August. Larva on buds of Grindelia and in seeds of sunflower. Light, with purple dorsal, subdorsal, and substigmatal lines, twice as wide as the space between them. Pale below; head light brown, with dark lateral band from back of head to eyes.

Southern New Jersey; Iowa, and west.

2. H. stypticellum Grote. White, more or less dusted and shaded with gray. Antemedial line white, obscure, erect, followed by a heavy dark gray shade; postmedial line oblique, parallel to outer margin, diffuse but distinct, with a strong transverse dark gray shade between it and the end of the cell; discal dots blackish. Hind wing gray. 20 mm. June and July. Larva apparently in the panicles of *Rhus glabra*.

Maine and northern Ontario to Costa Rica. New York: Fentons, Ithaca.

H. anguliferellum Ragonot, an Indian species recorded from North America, probably in error, is similar to H. stypticellum, but with the antemedial shade sharply angled and the discal points well separated. 30 mm.

3. H. mucidellum Ragonot. Pale powdery gray; antemedial line dark gray, strongly and acutely angled at middle: or represented by three dark gray dots, the middle one much farther out than the other two; postmedial line pale, defined with gray shades, parallel to outer margin. Discal dots distinct, sometimes fused. Hind wing translucent whitish, with darker veins. 20 mm.

July; September. Larva in heads of Gnaphalium and apparently in chicory seed. New York to Florida and west. New York: Ithaca.

H. reliquellum Dyar, from New Hampshire, appears to be a variety of H. mucidellum; it has a somewhat less angulate antemedial line, and rather darker hind wing.

105. EPHESTIA Guenée

Palpi upturned beyond vertex, third segment long; maxillary palpi normal; antennæ, in our species, simple laminate. Fore wing with R_2 from cell; R_3 and M_3 wanting, M_2 and Cu_1 from angle of cell. Cu_2 arising well before angle. Hind wing with cell short; with middle discocellular short, but decidedly curved, Sc with a

very short free tip, R shortly stalked. Wings sometimes tufted. The larvæ are pests in dried-food products. The four species noted below have been introduced here and there, from Europe, but E. kuchniella has become very common.

Key to the species

- 1. Hind wing with cell short, M_2 free from the angle; Sc rudimentary; fore wing with postmedial line sinuous, oblique.
- 2. Antemedial line angulate or dentate; a black discal lunule or point.

1. kuehniella.

2. Antemedial line very slightly angulate or dentate, or even. 3. Antemedial line oblique from costa to Cu, then erect to inner margin.

3. figuliella. 3. Antemedial line vertical......2. cautella. 1. Cell long, M₂ stalked with Cu₁.....4. elutella.

1. E. kuehniella Zeller (Mediterranean flour moth). Typically even pale or bluish gray; lines paler, outlined with darker, often faint; antemedial line zigzag; postmedial dentate; a slightly darker diseal lunule. Hind wing mostly translucent white. 22 mm. (H p. 412, f. 232-233.)

Larva white to red, with yellow-brown head; usually in meal, and showing a decided preference for rolled wheat.

Generally distributed and rather common; sometimes in injurious numbers. New York: Ithaca.

There are several inheritable mutations of this species, the most striking being a black one, and one with blackish ante- and postmedial bands.³⁸

2. E. cautella Walker. Brownish or blackish gray; similar to E. kuchniella except for the direct, rather distinct, pale antemedial line; defined with dark on the outer side. 15-20 mm. (H. p. 414, f. 235.)

Larva with contrasting black tubereles; white, with amber head; commonest in dried figs.

New York: Albany.

3. E. figuliella Gregson. Antemedial line oblique from costa to Cu_1 , then upright to inner margin. Dark gray, similar to the last two species. 20 mm.

Larva with dark brown head, cervical shield, and tubercles. Injurious to dried fruit.

4. E. elutella Hübner. Blackish to light gray-brown, tinted with ochreous and reddish. Lines formed of white powdering, moderately wavy; antemedial line at middle of wing; often obsolete. 15 mm. Larva light flesh color, with red-brown head, cervical shield, and supra-anal

plate, and small brown tubercles; found in rotten wood. New Jersey, western Pennsylvania, etc. Europe. Probably native. New York: Albany.

106. EPHESTIODES Ragonot

Palpi upturned to vertex, third segment longer than second, broadly scaled; antennæ simple; maxillary palpi normal. Fore wing with 10 veins; M_2 and M_3 stalked. Hind wing with Sc minute, M_2 and Cu_1 long-stalked; Cu_2 from angle of cell.

1. E. infimella Ragonot. Wings narrow, reddish luteous, yellower at the base, and shaded with violet gray in the middle above the fold. Antemedial line diffuse, nearly erect, obscure, followed by a blackish shade. Postmedial line oblique, preceded by a dark shade; discal points distinct, sometimes fused. Terminal space decidedly reddish. 10 mm.

June in the North, earlier in the South. Larva in seeds of Ambrosia.

Virginia to Texas and California; North Carolina; Columbia. New York: Ithaca (?)

³⁸ Details have been published by P. W. Whiting in the Journal of Experimental Zoology 28: 413. 1919

107. MOODNA Hulst

(Manhatta Hulst; Hornigia Ragonot)

Palpi in our species oblique, beak-like, with third segment almost as long as second; antennæ typically a little sinuous toward base. Fore wing (fig. 386) narrow, with a costal fold and hair pencil on under side in male. \mathbf{R}_2 free, \mathbf{R}_3 lost; \mathbf{M}_2 and \mathbf{M}_3 stalked; \mathbf{Cu}_1 arising from angle of cell, and \mathbf{Cu}_2 well before it. Hind wing with \mathbf{M}_3 wanting, \mathbf{M}_2 and \mathbf{Cu}_1 connate or short-stalked; discocellular short, curved; \mathbf{Cu}_2 arising well before angle. The larva of the type species (in Europe) feeds on pine.

1. M. ostrinella Clemens. Male antennæ with very slight sinus. Fore wing crimson, sometimes suffused with gray; median area shaded gray, suffused with black, except beyond the discal dots; lines well marked, pale; antemedial line well out, postmedial slightly waved. 15 mm. (obtusangulella Ragonot.)

Larva in sumac heads (Crosby) and in acorns (Murtfeldt). Moth in late July and August.

Pennsylvania; Texas. New York: Rochester, Otto, Ithaca.

2. M. pelviculella Hulst. Gray, much lighter than M. ostrinella; base washed with russet; antemedial line erect; pale, followed by a darker shade; outer weaker, denticulate. 15 mm.

Pennsylvania; doubtfully distinct from M. ostrinella.

108. *PLODIA* Guenée

Front strongly tufted; palpi beak-like, porrect, about as long as head; antenna simple; male with a small costal fold and pencil on fore wing. Fore wing (fig. 387) with \mathbf{R}_3 and \mathbf{M}_3 lost, \mathbf{Cu}_1 separate, \mathbf{Cu}_2 arising well before angle of cell. Hind wing with Sc free only at extreme tip; \mathbf{R} short-stalked, discocellular oblique, \mathbf{M}_3 wanting and \mathbf{Cu}_2 arising well before angle of cell.

1. P. interpunctella Hübner. Base clay color, greenish when fresh; outer threefifths dull red-brown, contrasting with three or four shining lead-gray bands, the last one terminal. 15 mm.

Larva injurious to stored grain, and occasionally on other dried foods; pale yellow with russet head and shields. (H p. 415 f. 236.)

Generally distributed and world-wide. New York: Common generally.

109. CAUDELLIA Dyar

Fore wing with only nine veins; \mathbf{R}_4 and \mathbf{M}_2 lost, \mathbf{M}_3 and \mathbf{Cu}_1 arising separately. Hind wing with seven veins; Sc very short, \mathbf{M}_3 and \mathbf{Cu}_1 separate; cell half as long as wing. Tongue well developed, palpi oblique, projecting twice the length of head, third segment deflexed, maxillary palpi filiform. Ocelli present. Male antennæ with a slight flexure on shaft. Fore wing with an oblique scale tuft on under side of costa near base.

1. C. apyrella Dyar. Dark vinous brown, heavily shaded with black. Marks obscure, of vinous shades. Antemedial line a vague paler shade; an obscure light postmedial patch, reaching up to the discal dot; a black shade at apex, and terminal line. Costal tuft vinous. Hind wing translucent whitish, shaded with fuscous. 14-15 mm.

June and July.

Plummer's Island, Maryland.

2. C. albovittella Dyar. Third segment of palpus shorter; a costal fold in male besides the tuft. Fore wing vinous brown, shaded with black. Antemedial line white, oblique, straight, almost forming a blotch subcostally; postmedial

whitish, well out; discal dots small, black, followed by white scales. Hind wing as in C. apyrella. 13 mm. July.

Plummer's Island, Maryland.

Subfamily ANERASTIINÆ

(Peorinæ; Phycitinæ, in part)

Closely similar to the Phycitinæ, and hardly worth separating; more frequently with porrect palpi and dominantly longitudinal markings. Ocelli present in our species. Tongue weak, not separating the palpi. There are a couple of genera which seem to be closely related to the normal

Phycitinæ but with a weak tongue. They are taken up with the Phycitinæ, but included in both keys to genera, for convenience.

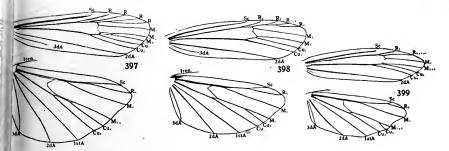
Key to the genera

1. Cu of hind wing apparently trifid (one medial lost).

2. Cu of fore wing quadrifid (M_2 preserved).

3. Palpi porrect and beak-like.

4. \mathbf{R}_2 stalked, \mathbf{M}_2 and \mathbf{M}_3 stalked or connate; hind wing with Sc and R anastomosing110. Aurora.



FIGS. 397-399. ANERASTIINÆ

397, Pectinigeria gemmatella, venation (The cross-vein between M₂ and M₂ is probably an individual aberration.); 398, Peoria hamatella, venation; 399, Tampa dimediatella, venation

4. R₂ generally free; hind wing with Sc and R approximate.

2. Cu, and Cu, stalked; fore wing with nine veins only......115. Cabnia.

110. AURORA Ragonot

Similar to Peoria. Palpi as long as head and thorax; maxillary palpi small, normal; fore wing with 11 veins, R2 stalked, M2 and M3 stalked, Cu1 from angle of cell. Hind wing with short cell; Sc and R anastomosing, M_1 stalked, M_2 and Cu_1 long-stalked, Cu, arising before angle of cell.

1. A. longipalpella Ragonot. Gray, with a whitish shade toward costa; antemedial line shaded with blackish, running obliquely up and in from inner margin toward costa; outer line similar; below, nearly parallel to outer margin, angled toward costa; no discal points. 22 mm.

October.

"North America" (Ragonot), perhaps western.

111. POUJADIA Ragonot

Palpi oblique, third segment porrect, second with a cavity containing the small maxillary plume. Front with a conical tuft. Antennæ with a sinus and a small scale tuft. Fore wing with R_2 free, R_3 and R_5 stalked, M_2 and M_3 stalked, Cu_1 near angle of cell. Hind wing with Sc free, R and M_1 connate, M_2 and Cu_1 stalked, M_3 lost, and Cu₂ well back from angle of cell.

1. P. glareosella Zeller. Dull, somewhat pinkish gray, with a darker shade below the whitish costa toward the base; costa diffusely white to the apex; discal bar faint, darker. Hind wing pale and translucent. 18 mm.

August.

East River, Connecticut, to Texas.

112. PECTINIGERIA Ragonot

(Cayuga Hulst; Spermatophthora, in part)

Palpi very long, porrect; maxillary palpi small, normal; tongue weaker than in Melitara; ocelli distinct. Male antennæ broadly laminate below, with a longitudinal groove in the base of the shaft above, flanked by scale ridges; basal segments of shaft partly fused. Fore wing (fig. 397) with 11 veins, \mathbf{R}_2 closely approximate (sometimes partially fused) to stem of R_{3-5} , M_2 and M_3 stalked, Cu_2 arising well before angle of cell; hind wing with M_a lost; Cu_2 distinct from angle of cell; M2 and Cu1 stalked; Sc and R closely approximate or partly fused.

1. P. gemmatella Hulst. Pinkish, about the color of Peoria approximatella, but strigose; costal stripe paler pink, streaked with white on Sc and R, with a blackish shade below it running along lower side of \mathbf{R} to end of cell and then to apex. 25 mm.

Coast of Massachusetts and New Jersey; Illinois and west.

113. PEORIA Ragonot

(Eurhodope, Anerastia, in part)

Male antennæ slightly curved at base of shaft, only slightly laminate; palpi porrect, long, and pointed; maxillary palpi very small. Fore wing (fig. 398) with 10 veins: \mathbf{R}_2 stalked. \mathbf{M}_2 and \mathbf{Cu}_1 connate, \mathbf{Cu}_2 arising near angle of cell. Hind wing with Sc and R anastomosing; \mathbf{M}_2 and \mathbf{Cu}_1 long-stalked.

1. P. approximella Walker. Fore wing dull rose to deep purple, shading into brown on cell, and pale below; costa contrasting, white; the stripe covering part of the cell at the base, and running to apex, shaded below with blackish. 20 mm. (hæmatica Zeller, roseatella Packard). June and early July.

Common and generally distributed. New York: Peru, Lancaster, Ithaca, Big Indian Valley, Rhinebeck, New Windsor, Katonah.

2. P. bipunctella Ragonot. Similar, with costal half of fore wing white, including the cell; ground generally paler. 15 mm. North Carolina.

114. CALERA Ragonot

Similar to Peoria; scape broadened and flattened; M2, M3, and Cu1 fused.

1. C. punctilimbella Ragonot. Marked like P. approximella; more delicate, as a rule without any blackish shade. Very possibly a venational aberration of approximella.

North Carolina; Texas; Louisana.

115. CABNIA Dyar

Fore wing with Cu_1 and Cu_2 stalked, M_2 absent, as well as M_3 ; three radials only; a costal fold beneath, at base. Palpi upturned, second segment with a tuft below; maxillary palpi simple; male antennæ with a slight process on scape; shaft sinuous at base and slightly thickened, with a few hairs in the bend. 1. C. myronella Dyar. Dark cinereous, even; lightly dusted with white; lines obscure, whitish, sinuous, distinct only toward the inner margin. 11 mm.

June.

District of Columbia.

116. VARNERIA Dyar

Fore wing with nine veuns; R_2 and R_4 separate, M_2 lost, M_3 and Cu_4 stalked; hind wing with six veins, Sc and M_2 lost, M_3 and Cu_4 stalked; Cu_4 near angle; discocellular short, slightly curved. Tongue as long as head; palpi upturned above vertex, third segment more than half as long as second.

1. V. postremella Dyar. Crimson, immaculate. Head, thorax, and costa of fore wing somewhat suffused with blackish; sometimes with blackish shades on the veins beyond the cell. Hind wing fuscous gray. 10 mm.

July and early August.

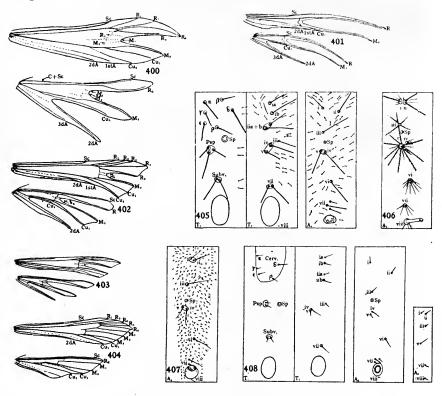
East River, Connecticut, to Kentucky.

FAMILY 34. PTEROPHORIDÆ

(Alucitidæ)

Head prominent; palpi often rather long, tongue functional; maxillary palp: obsolete; ocelli weak or absent; antennæ long, with two rows of scales to a segment above; pubescent below, simple. Legs very long and slender, the hind legs normally much longer than the entire body; with strong scale tufts at the spurs; spurs all present and exceptionally long. Fore wing, in the northeastern species, deeply cleft at middle of outer margin; hind wing cleft into three narrow feathers, often nearly to the base (in the Agdistinæ (fig. 404) the wings are entire). Venation of fore wing (figs. 400 to 403) often reduced in the radial region, \mathbf{M}_1 and \mathbf{M}_2 very short and weak, running to the notch, 1st A generally well developed, much longer than 2d A, anal angle located if developed at all, at Cu₂. Inner margin of fore wing and costa of hind wing folded over near middle and interlocking. Hind

wing with Sc and R closely parallel to beyond the end of the cell, then diverging and supporting the first feather; \mathbf{M}_1 and \mathbf{M}_2 as in the fore wing, \mathbf{M}_3 and \mathbf{Cu}_1 supporting the second feather, \mathbf{Cu}_2 closely parallel to \mathbf{Cu}_1 but much shorter, often fading out, the two veins marked by a groove on the under side of the wing and bearing a double series of dark spatulate scales, which is the surest characteristic of the family. One or two developed anals, supporting the last feather, which often bears tufts of dark-colored scales in the fringe.



FIGS. 400-408. PTEROPHORIDÆ AND OBNEODIDÆ

400, Pterophorus elliottii, venation; 401, Alucita pentadactyla (Europe), venation; 402, Platyptilia cosmodactyla, venation; 403, Stenoptilia pelidnodactyla (Europe), venation; 404, Agdistis statices (Europe), venation; 405, Platyptilia rhododactyla, seta map of larva; 406, Pterophorus lienigianus (Europe), seta map of a middle segment of abdomen; 407, Oxyptilus hieracii (Europe), seta map of a middle segment of abdomen; 408, seta map of larva of Orneodes hexadactyla

The moths rest with the fore wings folded, enclosing the hind wings in a tight roll, the whole held at a sharp angle, often at right angles, to the body. The flight is very weak and suggests that of a crane fly.

Egg of flat type. Caterpillar (fig. 405-407) with prolegs slender, often almost stalked, with a few hooks in a single band; the hair often glandular; usually with secondary or tufted hair; warts **iv** and **v**, as well as **i** and **ii** tending to unite, but quite variable. Spiracles small and circular, unlike the Macrolepidoptera. They are mostly leaf rollers, a few are borers.

Pupa usually suspended by the tail, but of the incomplete type in the number of free segments; maxillary palpi absent; fore and midlegs extending between eyes and antennæ; never with a deep groove between ninth and tenth segments of abdomen; rough or with tufted setæ, often very angular. Cremaster elongated, covering a ventral strip on last three segments, or divided in two.

The Pterophoridæ are an isolated group, but evidently nearest to the Pyralididæ; over 350 species are known.

The northeastern genera represent two tribes. In the Platyptiliini there are almost always specialized scales in the fringe; the second feather of the hind wing contains three veins (save in Trichoptilus); the head usually bears bifid scales, and the female frenulum is usually simple. The larva almost always has secondary hair. The Pterophorini never have specialized scales in the fringe, there are only two veins in the second feather, the head rarely bears bifid scales, and the female frenulum is usually of two bristles.

In this family the revision by Barnes and Lindsey has been followed in the delimiting of genera and species, but it has seemed better to leave the more familiar generic names undisturbed, retiring the two oldest, Pterophorus and Alucita, temporarily, as of uncertain application. If Hübner's "Tentamen" is taken at face value as a real attempt to define a series of genera by fixing types, Pterophorus would fall to a group not known from the northeastern States, in any case. The key to Oidæmatophorus is based primarily on Barnes and Lindsey's key. New York records in some cases are few on account of the impossibility of verifying records at present.

Key to the genera

1. Hind wing with a black scale tuft in third feather.
2. Fore wing with marked anal angle; second feather truncate.
3. First feather of fore wing also truncate, with marked anal angle; \mathbf{R}_1 and
\mathbf{R}_2 both free (fig. 402)l. Platyptilia.
3. First feather lanceolate; \mathbf{R}_1 only free
2. Both feathers linear

1. Hind wing without any black scaling on third feather.

- 2. First lobe of hind wing lanceolate; one radial missing, there being only four veins in the first feather (fig. 400).
- 2. First lobe of hind wing obliquely truncate; fore wing with all veins preserved (M_1 and M_2 rudimentary, as usual).
 - 3. Front conical (in our species); fore wings narrower.....4. Stenoptilia.
 - 3. Front flat, with a tuft of scales only, fore wings broader (fig. 402).

1. Platyptilia.

1. PLATYPTILIA Hübner

(Amblyptilia Hübner, Cnæmidophorus, Eucnæmidophorus Wallengren, Sochchora Walker, Gilbertia, Crocidoscelus Walsingham, Gillmeria, Fredericina Tutt)

Fore wing only moderately cleft (fig. 402); both feathers broadening to obliquely truncate apices, and quite broad; front more or less tufted, but without a horny cone; palpi fairly long and oblique. Tibiæ normally tufted, smooth in *P. punctidactyla*. The black scaling on the third feather is variable from species to species, and is useful for distinguishing the species; it is rarely absent.

The larvæ (fig. 405) are borers, at least when young, and have simple primary and fine secondary hair, or hair-like granulations. They hibernate part-grown.

Key to the species

- 1. Basal portion (more than half) of third feather of hind wing white, contrasting with the rest of the wing; fore wing bright ochre..9. rhododactyla.
- 1. Third feather concolorous, sometimes with pale fringe; fore wing duller.

2. Ground clay color to tawny.

- 3. Tuft conspicuous, dark, a chocolate brown costal triangle on fore wing. 6. carduidactyla.
- Tuft weak or absent, no costal triangle on fore wing, but an oblique shade only; frontal tuft and palpi very long.

4. Tuft rather diffuse, at middle of third feather.....l. pallidactyla.

4. Tuft near apex of third feather, sometimes absent.....2. carolina. 2. Ground gray to chocolate brown.

3. Tuft three-fourths way out on third feather.

- 4. Dark gray, with markings scarcely darker; tuft triangular.

4. auriga.

- 3. Tuft at middle of feather.
 - 4. Tuft inconspicuous, of scales both before and beyond middle of feather. 5. tesseradactyla.
 - 4. Tuft conspicuous and triangular, beginning at middle of feather. 5. Fore wing shaded subterminally with wood-brown...8. acanthodactyla.

1. Tuft on inner margin very slight, brown, near middle of third feather; palpi and frontal tuft very long (Gillmeria).

1. P. pallidactyla Haworth. Head, body, and fore wings clay color or pale ochre yellow; fore wing more or less shaded and banded with ochre-brown, not

forming a costal triangle, but usually a distinct marginal band. Hind wing dark yellowish fuscous, the tuft light brown and variable in distinctness. Hind tibiæ white at base, dull brown outwardly; metatarsi brown-tipped. 22 mm. (marginidactyla Haworth.)

Common in June, flying into July. The caterpillar bores in the stems of yarrow, in the fall, hibernates when half-grown, and feeds more or less exposed in the crown of the plant, in the spring. It is green, with three pairs of white stripes, the subdorsal being the strongest. When half-grown, it has purple-brown stripes. The secondary hairs (or possibly enlarged skin-granulations) are minute; the primaries simple, iv and v separate, vi of two setæ. It is also reported on leaves of Senecio.

Generally distributed. New York: Peru, Newcomb, North Creek, Black Brook, vicinity of Buffalo, Rock City and Vandalia (Cattaraugus County), Potter Swamp (Yates County), Portage, Ithaca, Karner, Albany.

II. Hind wing with a small tuft more than three-quarters way out, rarely absent; and no distinct one at apex; fore wing with scattered black marginal scales, but no tuft; head as before. Larva unknown.

2. P. carolina Kearfott. Frontal tuft and palpi long. Ochre yellow, fore wing with a little darker dusting; costa dusted with black from one-third to three-fourths way out; dark dots below costa at one-third and on lower lobe just before the cleft. Terminal line blackish brown. A lighter brown line in base of fringe, the rest of the fringe slightly purplish. Dark specimens with the usual pattern of the genus in brown. Hind wing slightly pinker, with tuft very small or absent. Legs nearly concolorous. 18-26 mm.

End of May to September. New Jersey; Valley of Black Mountains, North Carolina; Utah; California. 3. P. edwardsii Fish. Tawny red-brown, costa blackish on basal two-thirds; triangle black-brown, followed by a white streak and a white bar in the costal fringe. Subterminal line white, with a blackish shade before it, preceded by dark streaks or shades on the costa and at the middle of the first lobe. Fringes black and white. Hind wing red-brown or chocolate brown, with a smallish tuft fourfifths way out on the third feather. 19-29 mm.

May to August.

Quebec to North Carolina and the Pacific.

4. P. auriga Barnes and Lindsey. Dark brownish gray with normal markings (as in P. edwardsi). Inner margin of third feather with a triangular tooth, the scales successively shorter outward. Abdomen with oblique pale and dark stripes above and longitudinal ones below. 18 mm.

May to September.

New York to North Carolina. New York: Ithaca. **P. petrodactyla**, of the Arctic region, belongs to this group. White, dusted with gray-brown; terminal line gray; fringes white, slightly darker outwardly on hind wing. No triangle of fore wing. Legs gray. Tuft very weak or absent.

III. Tuft on inner margin distinct, at middle of third feather; palpi and frontal tuft short. (Platyptilia; Fredericina Tutt).

5. P. tesseradactyla Linnæus. Ash-gray, dusted with white, and somewhat shaded with brown. A white subterminal line across both feathers, and a faint trace of the dark costal triangle, opposite the bottom of the notch, with a small white bar beyond it; a few spatulate black scales on inner margin. Hind wing darker, with a rather diffuse tuft of black scales. 15 mm. (*Fredericina* Tutt.) May. Caterpillar boring in stem and flowers of Gnaphalium, hibernating part-

grown, and pupating upright in the boring, the pupa normally emerging. Larva

brown, with large white dorsal patches, with primary setæ only; iv and v on one tuberele, vi of two setæ.

Northern; south to Pennsylvania and Colorado; also in Europe. New York:

('rosby (Yates County), Ithaca, West Danby, Karner. 6. P. carduidactyla Riley. Fore wing dark wood-brown or tawny brown; an irregular antemedial oblique blackish shade; a contrasting chocolate-brown tri-angle over end of cell, touching the costa, at least at its outer end, with a paler band beyond it; a more or less distinct, paler, subterminal band across both feathers. Fringe of inner margin white, cut with black. Hind wing with a wellmarked triangular black-tipped tuft at middle of third feather, and some scattered scales. 20-27 mm.

May to September. Larvæ several together in a web in heads of thistle. Light straw yellow; head, cervical shield, thoraeic legs, tubercles, and anal plate black; eleventh segment with two transverse black marks. Pupa very slightly angular.

Ğeneral.

IV. Hind wing with a strong tuft about three-fifths way out on third feather, and a very small one at apex; fore wing with a strong tuft beyond middle of inner margin. Palpi and frontal tuft short (Amblyptilia).

7. P. punctidactyla Haworth. Fore wing mottled, brownish black and creamwhite, with a blackish triangle and band before the subterminal line. 16-18 mm. (cosmodactyla Hübner, monticola Grinnell).

Larva green, with green prothorax; secondary hair fine, not clubbed. dorsal primaries single-haired, subventrals with two hairs; on flowers and young seeds of Stachys, columbine, geranium, and other herbs.

Northern Illinois and west; doubtful eastward. Europe.

8. P. acanthodactyla Hübner. Similar to P. punctudactyla, less mottled, and strongly shaded with wood-brown and gray.

Larva with several-haired warts, the longer hairs swollen just before their tips, and with fine, somewhat clubbed, secondary hair; tubercles iv and v united. On mints, Ononis, Pelargonium, Euphrasia, and others.

Europe; California. All castern records are doubtful. New York: Otto and West Farms (doubtful).

V. Third feather of hind wing relatively short, with a very large tuft well beyond the middle. Front only a little rough-scaled, without any conical tuft, (Eucnæmidophorus Wallengren).

9. P. rhododactyla Fabricius. Bright ochre yellow; fore wing with an oblique white streak reaching inner margin at one-third, a patch in the cell beyond the middle, and a postmedial line, touching the base of the notch; with a little dark brown before it. Hind wing gray-brown. 20 mm.

June. Larva a leaf roller on rose; with fine clubbed secondary hair; primaries distinct, but somewhat associated with secondaries to form rudimentary warts; iv and v united.

St. Louis, Missouri; probably introduced from Europe.

2. OXYPTILUS Zeller

(*Pterophorus* Latreille, not Hübner; Geoffroy, in part)

Similar to Platyptilia; no distinct frontal tuft; palpi slender, second segment with a slight apical tuft, third pointed. Fore wing with first feather in our species simply lanceolate, with the hind angle only indicated by a black tuft

in the fringe; second feather strongly falcate. One radial sometimes lost; \mathbf{R}_2 stalked; \mathbf{M}_3 stalked with \mathbf{Cu}_1 . Hind wing with a heavy black tuft in the dorsal fringe on the apical fourth of the third feather, and in our species with a heavy tuft opposite it in the costal fringe of the feather. (Larva, fig. 407.)

Key to the species

1. Ground ochre yellow..... 2. Second segment of palpus not tufted.

1. O. periscelidactylus Fitch. Tawny or ochre yellow, including the head and thorax. Palpi orange and white; antennæ white above, barred with brown. Fore wing with postmedial line white, crossing the two feathers well beyond their separation, and continued as a fine and broken line around the base of the notch from one feather to the other. Subterminal line white, practically crossing the first feather, but not reaching the inner margin of the second. Hind wing with first

two feathers chocolate brown, third nearly white, with a blackish apex. 18 mm. Rather common from the end of May to July. Caterpillar with tufted hair and fine clubbed secondary hair; webbing together the growing tips of grape, but injuand ectablished secondary har, websing together the growing tips of grape, but infurious only in localities where it occurs exceptionally early, as otherwise it only damages the shoots which are soon to be pruned away. Pupa suspended in the web. Generally distributed. New York: Vicinity of Buffalo, Union Village, Batavia, Ithaca, Schenectady, Albany, Menands, Poughkeepsie, New Windsor, Scarsdale.
2. 0. tenuidactylus Fitch. Antennæ black and white. Dark brown with some

coppery tint, and often more or less tawny, or heavily mixed with black; postmedial and subterminal lines as in O. periscelidactylus. Apical fringe sometimes pale yellow (*delawaricus* of collections). Hind wing blackish, the third feather with some white near the middle; under side of first feather with fine postmedial and subterminal lines. Hind tibiæ with spurs at middle; hind tarsi lightly barred with black, except at base. Abdomen brown, third segment with diverging white stripes, fourth all brown, fifth mostly white above; below, with fourth segment mostly white, and much white on the other segments. 12 mm.

May; July and August. Caterpillar on blackberry.

Generally distributed and common. New York: Keene Valley, Geneva, Wells, vicinity of Buffalo, Rock City (Cattaraugus County), Ithaca, Schenectady, Albany, Poughkeepsie.

In 0. cygnus Barnes and Lindsey, from Iowa, each segment of the abdomen is marked with white, above, the fourth as heavily as the others.

0. raptor Meyrick, from Colorado and Hessville, Indiana, differs from tenuidactylus in the upper spurs of the hind tibiæ being well beyond the middle. Abdomen with some white scales on hind margins of segments only. June to September.

3. O. delawaricus Zeller. Ochre yellow, like O. periscelidactylus. Second feather of fore wing less markedly falcate, and markings less clean-cut. Tuft on second feather of hind wing weaker. 13-20 mm. (bernardinus Grinnell, and raptor Meyrick, in part).

June and early July; apparently rare.

Quebec to New Jersey; west to the Pacific.

3. TRICHOPTILUS Walsingham

(Buckleria Tutt)

No frontal tuft; palpus with second segment tufted; fore wing more deeply cleft, full two-fifths the length of the wing; both feathers linear, much narrower than their own fringes, and without dorsal angles. Hind wing with feathers linear, the third with a strong tuft two-thirds way out.

1. T. lobidactylus Fitch. Fore wing with basal half mixed clay-color, blackish, and white, shading into solid blackish on onter half. Each feather with two white bars; fringe blackish, white toward apex of costa, and with some white scales in dorsal fringe. Hind wing dark. 15-20 mm.

June. Larva on Solidago; green with ochreous head and some black points; with subdorsal chitinous plates bearing setæ i and ii, which are single and clubbed, arising from a large brown chitinous plate; iv and v united; a few pale secondaries, partly associated with the primaries. Pupa truncate, with four short anterior horns, and subdorsal ridges reaching to fourth segment of abdomen, bearing setæ i and ii, green, marked with pink. The species is somewhat variable locally.

Massachusetts to Florida. New York: Ilion, Ithaca, New York City.

4. STENOPTILIA Hübner

Similar to Pterophorus in appearance, or with the lobes distinctly and very obliquely truncate. Front with a tuft of scales, normally covering a strong horny cone, ocelli present, palpi long and porrect in our species. Tibiæ smooth. Fore wings cleft a third way to base (fig. 403). Venation practically complete.

A cosmopolitan and very primitive genus, running into the lower forms of Platyptilia, and leading to the Pterophorini. Larva (of *pterodactyla*) with dense secondary hair on the cervical shield, unlike the preceding group; with warts and secondary hair on body, the latter clubbed. Two distinct subprimaries behind the spiracles. The larva are borers in the fall, and external feeders in the spring. They hibernate part-grown.

1. S. pterodactyla Linnæus. Body ashy; legs yellowish; tips of palpi white; fore wings and thorax reddish brown; costa and apices of lobes heavily scaled with dark brown; a reniform dark fissural spot; fringe and hind wing darker fuscous; terminal line paler. Nearly 25 mm.

Caterpillar green; on Veronica.

Europe. Reported from West Farms, New York.

2. S. exclamationis Walsingham. Lobes of fore wing distinctly obliquely truncate. Gray-brown; a distinct subterminal blackish shade on first feather, and a white subterminal line, besides the usual dark antemedial dot in cell and discal bar. 18-24 mm.

This species is obviously a transitional form to Platyptilia; it flies in July and August.

Ottawa, Ontario, and west.

S. mengeli Fernald, an ash-gray species without the heavy costal shade or dash, is known from Greenland and British Columbia.

5. MARASMARCHA Meyrick

Similar to Stenoptilia, \mathbf{R}_2 stalked, \mathbf{R}_3 lost. Our species (at least) with wellmarked scale tufts in inner fringe of fore wing, but none on hind wing.

1. M. pumilio Zeller. Light brown, dusted with white, especially toward the dorsal margin; posterior half of thorax, with tips of tegulæ, abdomen, and base of inner margin of fore wing, pale yellow. Fore wing with a blackish antemedial spot in the fold, and one in the cell; a spot at end of cell. Fringes and hind wing grayer. 15 mm. (ambrosiæ Murtfeldt, in part; liophanes Meyrick.)

Larva possibly on Ambrosia or some Papilionaceous plant; moth flying northward in August and September.

New Jersey to Missouri and south; Old World tropics.

6. OIDÆMATOPHORUS Wallengren

(Pterophorus Geoffroy, in part, not Hübner; Leioptilus Wallengren, in part; Alucita of Meyrick; Linnæus, in part, etc.)

No ocelli. Front without definite tuft, or more or less loosely conically tufted; palpi moderate, sometimes rough-scaled or tufted at apex; hind tibiæ and tarsi most often smooth-scaled. Fore wing (fig. 400) normally cleft a third way to base, or rather more; with two lanceolate lobes. Hind wing with the first feather lanceolate, the third linear, without any black scales in fringes. Fore wing with \mathbf{R}_3 lost, the other veins free; \mathbf{M}_2 running to the lobe, well above the notch; hind wing with one \mathbf{Cu} lost, first feather without dorsal angle, \mathbf{R} running almost to its apex; second feather also lanceolate.

A large and moderately varied genus, dominant in North America; absent from Australia.

Caterpillar (fig. 406) normally with distinct warts, and living more or less exposed A few species are said to be borers, with simple hair only (*Adaina*?). There may be a few subprimaries, but never secondary hair. Pupa suspended.

Key to the species

1. Fore wing lemon yellow, contrasting with the chocolate-brown hind wing.

15. sulphureodactylus.

- Fore wing not yellow, or hind wing hardly darker.
 Fore wing with a dark postmedial costal dot or patch opposite the base of the cleft, sometimes connected with a spot before the base of the cleft.

 - 3. Abdomen otherwise marked, often with unpaired dots.
 - 4. Ground white; middle tibiæ with a well-marked median tuft..9. elliottii.
 - 4. Ground not white, or tuft on mid-tibiæ slight or absent.
 - - - 6. Fore wing not white, or tuft on mid-tibiæ strong.
 - Fringe on inner margin of fore wing white or with white tufts; hind legs annulate; mid-tibia with strong tuft....3. eupatorii.³⁰
 Fringe with traces of white, or none, on inner margin.
 - 2. No dark mark opposite base of notch, on costa, but sometimes one halfway between this point and apex.

 - 3. Fore wing broader with narrower fringe; no crest on hind tarsus.
 - 4. Snow white, with no tuft on mid-tibia; fore wing with at most a slight dark dot at notch......8. homodactylus.

³⁹ O. mathewianus is always paler, with white hind legs and weaker tufts.

4. Cream-white or darker.

- 5. Spot at cleft an oblique shade, extending toward costa; fringe gravish
- into upper lobe, or double.
 - 6. Brownish, often with a double diseal dot at lower angle of cell; a blackish shade in fold to middle of wing; expanse 30 mm. 14. balanotes.
 - 6. Not brownish, or small; often with dark points at ends of veins. 7. With a dark dot near base of cleft.
 - 8. Dot lying before base of eleft, terminal dots on second lobe lacking or faint. Hind wings grayish.....11. paleaceus. 8. Dot lying at base of cleft.
 - 9. Fore wings more or less tinged with clear yellow; at most with one terminal dot at apex of second lobe.

10. stramineus.

- 9. No trace of clear yellow. Terminal spots evident, sometimes conspicuous......13. kellicottii.
- 7. No dot at base of cleft, or faint traces; outer margin with more or less distinct marginal dots or bars...12. lacteodactylus.
- 1. First two feathers of hind wing lanceolate, third with relatively narrow fringe; hind tibix with upper spurs subequal, typically without a small tuft of hair scales on dorsal side a little above the spurs; hind tarsi without crest (Oidæmatophorus, Leioptilus).

1. O. cretidactylus Fitch. Brownish or creamy white; head, except between antennæ, and sides of abdomen, brown; legs white with some light brown; not annulate. Fore wing shaded with brown on apex and dorsal margin, a dark fawn fissural spot, sharply defined on the outer side, connected by a bar to costa. Hind wing mouse gray; fringe not so dark as in P. cupatorii. 25 mm.

Late June and July.

New York and Essex County, New Jersey. New York: Vicinity of Buffalo. 2. **0. mathewianus** Zeller. White, more or less shaded with tawny brown and dark brown scales; sometimes forming spots on the costa and an oblique shade at the end of the cell, followed by a white line. Fringe on inner margin of first feather with a brown patch, followed by a white bar. 21-27 mm.

Western States. A very pale specimen apparently of this species has been taken by McDunnough at Sebec Lake, Maine.

3. O. eupatorii Fernald. Legs brown; hind tarsi mostly white, but annulate with brown, fore and middle tarsi white. Fore wing pale gray-brown, dusted with dark brown or yellowish white. Fissural spot diffuse, oblique, tending to form an oblique streak by joining with the spot on costa of first feather; a pale streak from base below costa almost to apex, not contrasting in very pale specimens. Costal edge marked with a heavy black bar opposite the fissural streak, and two smaller bars beyond. Second feather also sometimes streaked. Fringes blackish, the dorsal very heavy and contrasting, with black dots at base, cut with white, Hind wing dark grav. 22 mm.

Caterpillar social, on Eupatorium purpurascens. Pale green; when nearly mature, shaded with reddish, with an orange-ochre head, a yellow dorsal, a faint and broken subdorsal, passing over tubercles i and ii, a narrow lateral, above iii, and a broken stigmatal line. Principal dorsal setæ long, black, rough, wart iii with two long set $\tilde{\mathbf{x}}$; $i\mathbf{v}+\mathbf{v}$ with five longer white hairs, vi similar, a small subprimary behind the spiracle. Cervical shield with a fringe of white hairs overhanging the head, in front, and scattered black hairs behind. Pupa normal, green with subdorsal reddish shades. Moth in July.

Montreal to Pennsylvania, Illinois, Vancouver, Idaho, and California. New York: Ithaca, Van Cortlandt Park, New York City.

4. O. cineraceus Fish. Gravish to brownish white. Thorax often with the tips of the tegulæ darker; abdomen darker with a pale dorsal band containing dark dots; tibiæ tufted; tarsi usually annulate. Wings narrow. Fore wing, in dark specimens, with pale veins; dusted with black scales, sometimes very sparsely; base of cleft white, preceded by the usual black dash; with a spot opposite it and dark subterminal dots on the costa. 28 mm.

O. cineraceus differs from O. cretidactylus, and from Adaina ambrosiæ, which may run to it in the key, by the narrower wings not mottled on the basal half. Pennsylvania; western States.

5. 0. brucei Fernald. Pure white. Scale tufts on fore and middle tibiæ nearly obsolete. Fore wing with more or less gray-brown scaling, becoming a strong gray mottling in dark specimens, with the usual dark spots on cell and costa. Subterminal costal spot confined to membrane. 22-27 mm. (chionastes Meyrick.) June to August.

Pennsylvania, western States. Massachusetts (?; my specimen is not at hand and may belong to another of the light gray species of this genus).

6. O. inquinatus Zeller. Similar to Adaina ambrosiæ, normally larger, without any brownish tint; costal markings similar, but practically confined to the fringe; the dorsal fringe with clear white bars, two of those in the dorsal fringe of the second feather conspicuous. Abdomen with paired black dorsal dots, mottled with white. 20 mm.

June to September. Caterpillar apparently undescribed, on Ambrosia. Of the species confused with this by Miss Murtfeldt, and apparently bred by her from the same food plant, Adaina ambrosia is distinguished by the clay color or light wood-brown shading in the axis of the first feather of the fore wing, and by the unpaired black dorsal dots on the abdomen; and Marasmarcha pumilio by the black tufts on the inner margin, as well as by the structural characters.

New York to Illinois, Iowa, California, and south. New York: Ithaca.

7. O. linus Barnes and Lindsey. White. Fore and middle tibiæ with longitudinal gray stripes, not tufted; hind tibiæ with gray on spurs, and sometimes at tarsal joints; abdomen with a series of mid-dorsal dots, sometimes connected. Fore wing narrow, more or less dusted with black, especially at the apices of the feathers and on the inner margin; usual postmedial markings present; a few blackish dots at tips of veins, sometimes lost in the black dusting; fringe grav, cut with white at the veins. Hind wing brownish gray. 22 mm. (brucei in part ?; lienigianus Zeller ?.)

The coarse black dusting is characteristic.

June and July.

Hampton, New Hampshire, to Pennsylvania and Ohio.

8. 0. homodactylus Walker. Pure white. Fore wing with a little grav shading on under side, and with traces toward costa above. Sometimes with a faint darker dot at base of cleft, and dots at tips of veins. Middle tibiæ with a fringe of scales, but no tufts. 25 mm. (elliottii, in part).

This species is usually confused with O. elliottii, from light specimens of which it is sometimes only distinguishable by the different tufting of the mid-tibia. The single New York record below is based on specimens determined by Barnes and Lindsey; it is probably not rare, but confused with elliottii.

Larva on Solidago. Green with yellowish head and a broken irregular yellow dorsal line; a yellow lateral line, above iii, and traces of a stigmatal on the thorax. Hair whitish, slightly barbed; tubercles i and ii contiguous, i with 4 setæ and ii with 2 long and 2 shorter ones, ii with two short and a long; iv + v and vi each with about 12 hairs; posterior part of segment with a single hair behind iii and a small wart with 4 hairs behind iv+v. Prothorax with two patches of hair point-

ing forward, and some posterior hairs, mostly on three warts. Pupa green with white hair, the wing eases smooth, with one row of short hairs; with cream dorsal, subdorsal, and subventral stripes, and a broken lateral, the subventral on the ridge.

Quebec to New Jersey, British Columbia, and California. New York: Ithaca.

9. 0. elliottii Fernald. White; head brownish. Fore wing normally with traces of the usual spots and oblique bar at the base of the noteh; sometimes practically immaculate. Fringe brownish except in the cleft. Middle tibia with a strong tuft. 22-25 mm.

June and July. Larva on Eupatorium; light green, with smooth shining white hairs; head pale ochreous. A broken, yellow dorsal line, lateral, stigmatal, and traces of a subdorsal; tubercles i, ii, and iii with one strong hair each, iv+v with four long hairs, the wart behind it with a couple of short ones; only vi with twelve hairs. Pupa similar to that of homodactylus, with several rows of hairs on the wings, shorter hair generally, and with a mid-dorsal hair on several segments.

Quebec to New Jersey; west to Illinois and Manitoba. New York: Ithaca, Nassau (?).

10. O. stramineus Walsingham. More or less ochreous yellow; head brown in front and above; abdomen with brown dorsal and ventral lines. Fore wing usually with a distinct brown spot at the cleft; tips of veins sometimes brown; frequently with a brown shade from the base near the inner margin to the apex of the first feather, the apical part of the shade most persistent. Fringe and hind wing grayer. 15-21 mm.

July to September. A larva, possibly of this species, occurs on Anaphalis in Colorado. It is white, heavily granulose with black, and with a brownish black head.

Eastern Canada to New Jersey, and west to the Pacific. The specimens recorded below as of sulphureodactylus may belong here, as they are small, but all three have lost their abdomens.

11 0. paleaceus Zeller. Very pale brownish gray; head brown, pale between antennæ. Fore wing shaded with pale fuscous on costa outwardly, and tending to show a pale longitudinal streak below the costa, rarely suffused with brown; a darker spot at base of fissure. Legs yellowish white, shaded with fuscous below. (The ground is usually the light buff of Ridgway's Nomenclature of Colors for Naturalists,— 17'f or 17"f.) 19-26 mm. (sericidactylus Murtfeldt.) Two broods. Caterpillar on Vernonia, varying from greenish white to dull

salmon, the shorter hairs sticky. Pupa varying in color.

General south of Canada.

12. O. lacteodactylus Chambers. Cream white, with more or less distinct dark dots at tips of veins, but at most a faint dot at the cleft. 25-28 mm. (kellicottii of collections, in part.)

May to July.

Nova Scotia to North Carolina and California. New York: Portage, Potter Swamp (Yates County), Ithaca; Lynbrook, Long Island.

13. O. kellicottii Fish. Cream color, slightly streaked with brown, and sometimes with some scattered black scales; dark brown fissural dot; brown terminal dashes, usually one above the apex and two below on the first feather, and four on the second. Wing slightly translucent between the veins. Hind wing silky browngray; legs whitish. 30 mm. (chlorias Meyrick.) June. Larva unknown. The larva described as of this species is obviously a

Platyptilia.

Laurentians, Quebec to Pennsylvania. New York: Ithaca.

14. 0. balanotes Meyrick. Brownish white, suffused more or less with brown; abdomen striped. Palpi exceptionally long, long enough to reach base of antennæ if turned up. Fore wing, when fully marked, with dark brown spots at tips of veins, a double spot at lower angle of cell, and a powdery streak from base of

fold to below end of cell, offset upward at half its length; these markings all variable. (25) 30-41 mm.

Larva in stem of Myrica.

New Jersey; southern United States.

15. O. sulphureodactylus Packard. Bright sulphur yellow, slightly tinged with brown; with more or less distinct traces of the usual markings. Hind wing chocolate brown, contrasting. 25 mm. (sulphureus Walsingham.)

September.

Western States. Woods Hole, Massachusetts, and Hemlock Lake, New York (?). The eastern specimens are undersized, and their determination not certain.

0. inconditus Walsingham has been reported from the District of Columbia, presumably in error for one of the species just described. It is somewhat similar to lacteodactylus, but without the dark points at the ends of the veins, and is typically grayer.

II. All three feathers of hind wing linear, the third with a very broad fringe; hind tibiæ with upper outer spur only three-fifths as long as inner; with a small tuft of hair scales a short way above them. (Pterophorus; Emmelina Tutt).

16. O. monodactylus Linnæus. Pale grayish or pinkish brown (in the typical form from Europe with a decided yellow tint). Fore wing sometimes with a little black and white dusting; with a dark fissural spot or wedge; a dark sub-terminal bar on the costa, and two smaller spots beyond it; and dark spots on both feathers; all relatively slight. Hind wing and fringes normally chocolate brown. 28 mm. (pterodactyla Hübner not Linnæus, cinereidactylus Fitch, per-magiliactulus Bachard etc.) gracilidactylus Packard, etc.).

Caterpillar on Convolvulus and several other herbs. Bright yellowish green; head pale yellow. A narrow dorsal and broader stigmatal yellowish white stripe. Hairs grayish, in tufts from tubercles. Under side paler. Pupa green with blackish brown dorsal and lateral lines, and streaks on wing-cases; head flattened, hairy, marked with brown; part of dorsal hairs blackish. Nearly world-wide, but running to well marked local forms. New York:

Wilmington, vicinity of Buffalo, Ithaca, Schenectady, Albany.

7. ADAINA Tutt

(Pterophorus, Alucita, Œdematophorus, in part)

Similar to Oidæmatophorus, but with R, and R, stalked. Tibiæ not tufted, wings, on the average, narrower.

Neither of our species is typical of the genus, and they should perhaps be restored to Oidæmatophorus. In the typical species the larva is a borer, with simple hair, except for a couple of extra hairs associated with seta vi; our species agree with those of Oidæmatophorus.

1. A. montana Walsingham. White, lightly dusted with brown, the dusting gathering into a bar at end of cell, and a subterminal shade on costa. Fringe white, cut with brown below apex of first feather, and above apex of second. Hind wing browner. 16 mm.

July. Caterpillar on leaves of Solidago, full-grown in June. Pale green with a triple fine white dorsal stripe, and "seventh to ninth rings" yellow. Tubercle i with one hair, ii with two. iv and v as warts. Pupa green, sometimes with a red dorsal stripe; suspended. The egg hibernates. California. New York: Buffalo.

A. declivis Meyrick is considered by Barnes and Lindsey to be a variety of montana. Moth somewhat darker, shaded with brown dusting, and with more fully developed brown markings.

June and July. Larva in heads of Helianthus in late May. Warts i and ii approximate with one long and several short hairs, iii simple, iv+v a regular wart, the wart behind it with several hairs. Warts black; two black lines on dorsum of prothorax. Hairs white, spinulose. (This description is probably more accurate than the one above.)

Ontario; Manitoba; Colorado.

2. A. ambrosiæ Murtfeldt. White, more or less irregularly dusted with black, with a more or less distinct luteous to brown streak through the middle of the first feather, the feather above it and costal fringe white, with a heavy black patch near base of first feather and a narrow one near tip; the brown streak defined below with some black scales. Dorsal fringe of first feather dark gray, cut with two or three paler back scales. Dotsail ringe of mist leather dark gray, cut with two or three paler bars. Notch edged with white, preceded by a strong blackish shade, which is extended toward the inner margin by dark suffusion. Hind wing gray, base of fringe somewhat yellower. 15 mm. (*inquinatus*, in part.) Late June: late August and September. Larva on Ambrosia, with barbed sets,

those of warts i and ii short and recurved; subprimaries near i and ii; warts i and ii conical, brown, i with a tuft of long hair, with a black lenticle above it; dorsal line yellow, faint, sides with \mathbf{V} -shaped oblique stripes, pointing forward. Pupa green with a brown dorsal stripe.

Illinois; Missouri; Florida.

Family 35. **ORNEODIDÆ**

(Alucitidæ)

Moderately slender moths, with the wings each divided into six feathers; in our species, nearly to the base. Head prominent, scaled, with ocelli and tongue; palpi long and rough-scaled; maxillary palpi rudimentary. Each wing broad as a whole, but each feather linear, much narrower than its fringes, and each with a single strong vein, (these being: **R**, **M**₁, **M**₂, **M**₃, **Cu**₁, **Cu**₂ or **A**). Female frenulum of two bristles; male with a scent-pocket in last feather.

Egg rough, of flat type; larva rough and granulose (fig. 408), with small head, held nearly horizontal; primary setæ only; prespiracular wart and subventral of prothorax each with two setæ; subventral of meso- and metathorax with one seta; prolegs with a complete circle of uniordinal hooks. Spiracles circular. Pupa obtect, with Adelid type of prothorax; maxillary palpi concealed; dorsal head-piece enormous; seventh segment of abdomen immovable in both sexes. Dehiscence of macro type: with the antennæ attached to the legs; but the dorsal head-piece coming away separately; appendages nearly reaching tips of wings, second leg touching head, tips of third legs exposed; cremaster with 10 or 12 recurved spines. Setæ iii and iv distinct.

1. ORNEODES Latreille

(Alucita Curtis, and others⁴⁰)

Characters of the family. Wings cleft nearly to the base. 1. **O. montana** Cockerell. Dull light gray, each feather barred with blackish; with narrow white bands before and beyond each bar. 14 mm. (hexadactyla of authors, not Linnæus; huebneri Wallengren?)

⁴⁰By the type-fixation rule, hexadactyla would become the type of Alucita as fixed by Curtis.

April. (In Europe O. hexadactyla is reported as emerging in July and overwintering). The caterpillar of the European O. huebneri is translucent red, with darker shining head and cervical shield; reported from flowers of Centaurea and Knautia; also, perhaps in error, from honeysuckle and scabious. Pupa in a cocoon.

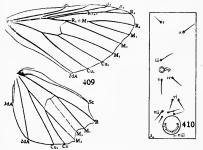
New York to Western States. New York: Ithaca. (Possibly introduced).

SUPERFAMILY URANIOIDEA

This superfamily is, in the main, made up of rather slender moths, with vestiture mostly of scales, and thin ample wings; the North American species closely resembling the geometers. Antennæ normally simple, prismatic, pectinate in a few exotic forms; palpi slender and upcurved; maxillary palpi minute, of porrect type, apparently absent in the northeastern species; tongue coiled, somewhat weakened. No ocelli. Fore wing with \mathbf{R}_3 and \mathbf{R}_4 stalked (fig. 409), widely separated from \mathbf{R}_5 which is approximate or stalked with \mathbf{M}_1 ; 3d A running into 2d A near base, or obsolete; 1st A lost. Hind wing with enlarged humeral angle, supported by a more or less distinct humeral vein; its outline usually irregular, angled or tailed; 1st A lost, 3d A normal. Other characters as given in synopsis. The superfamily includes the exotic Uraniidæ and Epicopeiidæ, as well as the Epiplemidæ, and is a characteristically tropical group.

Family 36. **EPIPLEMIDÆ**

Frenulum present, fully developed, attached to the tip of the



FIGS. 409-410. EPIPLEMIDÆ

409, Venation of Callizzia amorata; 410, Callizzia amorata, seta map of middle segment of abdomen (after Fracker)

humeral angle; retinaculum of bar type (as in the Arctiidæ). Caterpillar (fig. 410) of our species microlike in appearance; when young, social in a web; later, feeding more exposed; head normal with front extending about halfway to vertex; without tufted or secondary setæ, but with a subprimary seta associated with vi; vii represented by two setæ on prothorax, one on first segment of abdomen, two on second, and four on the leg-bearing segments; setæ iv and v on a level, and well below the level of the spiracles; approximate on first three segments

of abdomen but on the others widely separated; prolegs normal, with biordinal hooks in a sharply curved band. Pupa in a cocoon, not studied. Our two genera are typical of the family, though unusually small. Some of the exotic forms are striking.

Key to the genera

Fore wing deeply notched, toothed at middle and anal angle...1. Calledapteryx. Fore wing with even outer margin......2. Callizzia.

1. CALLEDAPTERYX Grote

Fore wing with arched costa, sharply curved down at the apex; outer margin toothed below \mathbf{M}_3 , deeply excavate above, and slightly below the tooth; anal angle broadly toothed, and inner margin sinuate. Hind wing with very strong teeth on **R** and \mathbf{M}_3 , and lesser ones on the other veins, especially \mathbf{M}_1 . Fore wing with \mathbf{R}_1 and \mathbf{R}_2 well separated, arising from cell; \mathbf{R}_3 and \mathbf{R}_4 long-stalked from well before end of cell; and \mathbf{R}_5 , \mathbf{M}_1 , and \mathbf{M}_2 from the apex. Body slender, antennæ simple, a little prismatic, palpi small; hind tibiæ swollen in male. The moth rests with the fore wings held somewhat rooflike, covering the hind wings, which are wrapped about the body.

1. C. dryopterata Grote. Light brick red to dull brown; ground narrowly blackish along the margin; two roughly parallel dark lines, with a brown shade between them at inner margin; a dark brown line just within the outer margin on upper half of wing. Hind wing similar, the two lines often distinct only at the inner margin. 18 mm. (H 42:17.)

May to early June; August. Caterpillar on Viburnum prunifolium (but not dentatum or Lentago). Head light brown, heavily spotted with black; body greenish white, more or less spotted with black, the black tending to form a dorsal line and a subdorsal band, especially toward the rear. Pupa in a slight web at surface of ground.

St. Johns, Quebec, to Alabama; west to Manitoba. New York: West Farms, (Angus); Corona, Long Island.

CALLIZZIA Packard

Head broader than in Calledapteryx; fore wing (fig. 409) with arched costa, outer margin only slightly bent at middle. Inner margin and hind wing not quite so irregular as in Calledapteryx.

This genus is hardly distinct from Epiplema.

1. C. amorata Packard. Ash gray, dusted and shaded in two shades; two fine dark brown lines, joining each other above inner margin, or connected by a bar; a dark patch above middle of outer margin, defined by an angulate dark line; hind wing with inner line almost evenly curved, and outer irregular, with a dark gray shade on disc between them. 18-22 mm.

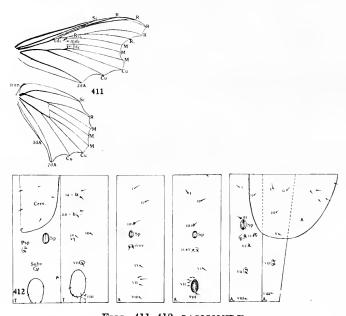
End of June and July Caterpillar apparently concealing itself at the foot of the plant when not feeding; on *Lonicera dioica*. Head whitish with two brown bands, tubercles a little raised; body dull brown with black lateral band, pale subventrally.

Quebec to Virginia, and west to the Pacific. New York: Keene Valley, Mt. Marcy, North Elba, Indian Pass, Newcomb, Ithaca, Big Indian Valley, Albany.

Family 37. LACOSOMIDÆ

(Perophoridæ)

Stout, bombyx-like moths, of quite uncertain affinity. Male antennæ broadly peetinate, abruptly narrowing at two-thirds their length. Female narrowly peetinate. Palpi and tongue rudimentary; vestiture deep and mixed. Fore wing falcate, and more or less bent at middle; with thick sealing and heavy veins; \mathbf{M}_2 arising from middle of end of eell; \mathbf{R}_2 and \mathbf{R}_3 long stalked, and close to costal edge, separated by a wide space from \mathbf{R}_4 and \mathbf{R}_5 (fig. 411), which are also stalked; \mathbf{M}_1 free, connate; \mathbf{Cu}_2 arising only halfway out on eell. Hind wing with humeral angle enlarged and strengthened, bearing a minute fremulum or none; Sc fused for a short distance at base with \mathbf{R}_5 , then abruptly



FIGS. 411-412. LACOSOMIDÆ 411, Lacosoma chiridota, venation; 412, Cicinnus melsheimeri, seta map of larva

and widely divergent, as in the Epiplemidæ and Saturnioidea; \mathbf{M}_2 arising from middle of end of cell. Fore wing with **2d A** only; hind wing with **2d A** and **3d A** normal, and **1st A** more or less distinctly indicated at margin.

The caterpillar (fig. 412), when young, makes a lace-like nest between two leaves; later it forms a portable house of two pieces of leaf, which is lined with heavy silk, very roomy inside, and has a eircular opening at each end. When not moving about, it anchors the case with silk, often in a slight nest formed of leaves drawn together, closing one opening of the ease with its head, and the other with the eircular thickened posterior end of its body. Head rough, more rounded than usual and heavily ehitinized, wider than prothorax; body thin-skinned, swollen at middle, and a little flattened; abdomen with seta i nearer to middle line than ii, even on ninth segment; iv and v well below level of spiraele and more or less approximate; vii composed of 2 or 3 setæ on first, second, seventh, and eighth segments; four (Laeosoma), or seven (Cieinnus) set on leg-bearing segments. Prolegs rather short, normal, with a complete circle of biordinal hooks. The full-grown eaterpillar hibernates in its ease at the surface of the ground, and pupates in the spring, like many micros.

Pupa obtect, of macro type; heavily chitinized, with three movable segments. Head missing in the only specimen before me; tongue rudimentary, as wide as long, with a minute rhomboidal labium between the bases of the maxillæ; fore femur broadly exposed, apparently not quite reaching the eye; fore tibiæ broadly abutting on the eye, middle tibiæ falling far short; the tips of the hind tarsi visible beyond the tips of the middle ones. Antennæ broadly peetinate; no spiraeular furrows; abdominal segments 2 to 7, with a double set of alternating teeth near front edge, dorsally, those on segment 7, much the strongest; first segment much reduced. This pupa seems distinctly Saturnioid in character, and agrees with the general impression that this family is related to the ancestral Saturnioidea.

The family is a rather small one, and wholly American.

Key to the genera

Margin of fore wing deeply scalloped (fig. 411).....l. Lacosoma. Margin of fore wing even.....

1. LACOSOMA Grote

1. L. chiridota Grote. Deep ochre yellow, red-brown, or dark brown, the male usually darker, with dark brown and white fringe, and more or less distinct postmedial band and discal dots on both wings. 22-30 mm. (H 41:21.)

June. Caterpillar in a somewhat clumsy and nearly circular case, on oak, in the late fall. Without clubbed setse on head. Massachusetts to Pennsylvania, Iowa, and south. New York: Binghamton,

Poughkeepsie, Staten Island, Yaphank.

WILLIAM T. M. FORBES

2. CICINNUS Blanchard

(Perophora, in part)

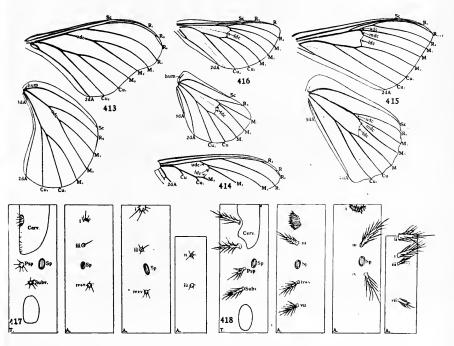
1. C. melsheimeri Harris. Wings light gray, shaded with rose, especially toward margin and on under side of fore wing, and dotted with black. A blackish postmedial line, bent sharply at \mathbf{R}_s , and more or less traces of an antemedial line on fore wing, and a straight postmedial line on hind wing. A black discal bar. 35-45 mm. (H 41:17.)

June. Commonest on sandy barrens. Caterpillar on oak, usually on scrub oak, in an ellipsoidal and rather neatly made case, covered with two pieces of leaf; caterpillar with two long clubbed setæ on the head.

Massachusetts to Wisconsin and southward. New York: Hempstead Plain, Yaphank, Long Island. Will probably prove widespread in sandy territory.

SUPERFAMILY SATURNIOIDEA

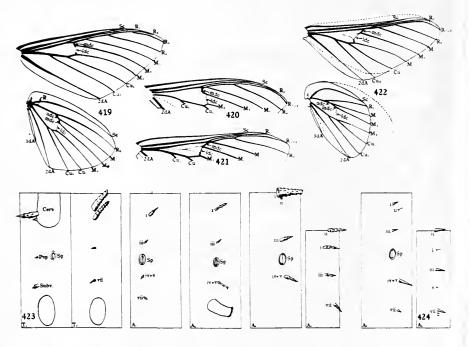
Large or very large moths. Body stout, relatively large as a rule, with hairy vestiture; head relatively small, more or less retracted,



FIGS. 413-418. SATURNIOIDEA

413, Callosamia promethea, venation of female; 414, Tropaea luna, venation of costal portion of fore wing; 415, Antomeris io, venation of male; 416, Hemileuca lucina, venation of male; 417, Telea polyphemus, seta map of larva, indicating only the setæ arising from warts, not the secondary setæ; 418, Hemileuca maia, seta map, omitting secondary setæ

often tapering ventrally; ocelli absent, mouth parts reduced, non-functional for feeding, sometimes almost completely absent. Labial palpi small to minute in the North American species. Antennæ bipectinate in the male in all the North American species, and most often in the female also; sometimes extremely broad; normally doubly bipectinate, especially in the male. Scape densely scaled; but shaft in the North American forms with a few scattered and fugitive scales (Automeris) or none.



FIGS. 419-424. SATURNIOIDEA

419, Citheronia regalis, venation: 420. Eacles imperialis. venation of costal part of fore wing; 421, Adelocephala bicolor, venation of costal part of fore wing; 422, Anisota virginiensis, venation (The dotted outline indicates the wing form of A. rubicunda, magnified to approximately the same proportions.); 423, Citheronia regalis, seta map, showing spines only, 424, Anisota rubicunda, seta map, showing spines only.

Legs stout, the spurs inconspicuous or absent. Abdomen usually relatively short, but exceeding the hind wings in the more primitive species. Fore wings densely scaled, with heavy veins; one radial at least lost (possibly \mathbf{R}_5 if the family is related to the Epiplemidæ); \mathbf{M}_2 more or less definitely associated with **R**-stem; the cell weakly closed or open below it. Bases of **R** and **Cu** closely approximate, often for a considerable distance; **1st A** lost, **2d A** normal. Hind wing without any trace of frenulum, with a broadly expanded humeral

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angle, supported by a more or less distinct humeral vein; Sc and R widely divergent almost from base, connected close to base by a rudiment of R_1 in Citheronia; M_1 and M_2 connected with R-stem by a full-strength longitudinal vein, the cell weakly closed or open below M_2 ; 1st A lost, 2d A normal; 3d A very short and rudimentary in the typical Saturniids, which have the inner margin more or less concave toward the body, and not folded.

Egg of flat type, usually thick-shelled, ovoid, without definite sculpture, but often with characteristic markings. Larva in first stage with branching spines bearing several setæ, but with the primary setæ distinct; or, for the major part, with primary setæ only, but with one or more pairs of thoracic spines each bearing two primaries; setæ ia and ib, iia and iib of thorax on single tubercles. i and ii of abdomen separate. iv and v of abdomen on a single bifurcated tubercle. Later stages with abundant but usually fine and inconspicuous secondary hair on body, but little or none on head, warts all several-haired, and usually modified into branching spines (fig. 418) or knoblike structures (fig. 417) often with the hairs themselves rudimentary. The spines or knobs often unequal, and some of them often rudimentary. Warts iii and iv+v usually forming spines; but i of the ninth segment of the abdomen single-haired and soon lost in the general mass of secondary hairs. Seta ii similarly lost except on the ninth and sometimes eighth segment; i of the eighth segment, or ii of the ninth, or both, usually (in our species always) fused with their mates on the middorsal line, those of the eighth forming the so-called "caudal horn." The spines of the Hemileucinæ are poisonous (like nettle), and have the setæ modified into conical caps which easily break off, setting free the poison.

Pupæ moderately to heavily chitinized, always obtect. but capable of considerable progression in the Citheroniidæ. Head without transverse sutures; mouth on ventral surface of body; tongue never reaching tip of wings, and normally only about a fifth as long as wings: labial palpi wholly concealed; no maxillary palpi; fore femora exposed: fore and middle legs usually meeting on the middle line behind the tongue. and the wings meeting on the middle line behind them; antennæ very broad, usually more than a fifth as broad as long. larger in the male, where they may almost completely cover the other structures. Abdominal segments sometimes spined but never (as in the Lacosomidæ) with an anterior and no posterior row of spines. Pupa clothed with microscopic secondary hair.

I here recognize two families. Besides these there is a third family in South America, characterized by fully scaled antennæ, and apparently transitional to the Uranioidca. Many authors consider the Hemileuca group also a distinct family, some including and some excluding Automeris. It is certaintly transitional between the Citheroniidæ and Saturniidæ.

Key to the genera, imago

- 1. M, of fore wing distinctly stalked with R-stem (figs. 420-422); antennæ pectinate with simple apex, or simple.
 - 2. Small species, expanse under 75 mm.; middle discocellular short and oblique inward.
 - 3. Inner margin of fore wing longer than outer margin; two white discal dots, or none; northern species shaded with crimson on the hind wing
 - 2. Large species, expanse over 75 mm.; middle discocellular oblique outward in general direction.
 - 3. Mid-discocellular much shorter than lower discocellular, and straight (fig. 419), rudiment of stem of M_{1+2} running into M_2 ; R_2 present; palpi largerC. 1. Citheronia.

1. M_i free; upper discocellular nearly longitudinal, and much longer than middle discocellular, which may be obliterated (figs. 413, 414).

2. 3d A of hind wing more than half as long as 2d A (fig. 416).

S. 1. Hemileuca.

- 2. 3d A of hind wing rudimentary. 3. Hind wings long-tailed......S. 3. Tropæa. 3. Hind wings rounded.
 - 4. Abdomen with small raised tufts of hair.....S. 7. Philosamia. 4. Abdomen with smooth hair.
 - 5. Cells closed by a fine vein passing through a transparent spot.

S. 4. Telea.

- 5. Cells open.
 - 6. R₂ preserved; sexes strongly dissimilar (fig. 413)...S. 5. Callosamia.
 - 6. R₂ lost, sexes alike.....S. 6. Samia.

Key to genera, larva

1. A single pair of long spines on thorax.....C. 4. Anisota. 1. Two pairs on thorax, and caudal horn much longer than the subdorsal ones on remaining segments of abdomen (fig. 423). 2. Secondary hair microscopic. 3. Spines cylindrical. 4. Five principal spines short and blunt, the others rudimentary. (Also young of Eacles and Adelocephala).

1. Spines subequal in length, at least several pairs of subdorsal ones.

2. Spines minute, mere tubercles, surmounted by long setæ (fig. 417).

- 3. Secondary hair more conspicuous; no secondaries on labrum; head green,
- body with oblique stripes on sides......S. 4. Telea.
- 2. Spines long, heavily spinulated, the caterpillar as a whole appearing hairy. 3. Ninth segment of abdomen with a small spine on mid-dorsal line; spines

poisonous (fig. 418).

- 4. Subdorsal spines of abdomen truncated, and discolorous in last stage; laterals sparsely spinulated.....S. 1. Hemileuca.
- 4. Subdorsal spines rather longer than laterals; and both densely spinulatedS. 2. Automeris.

3. Ninth segment without a mid-dorsal spine....young of various Saturniinæ.

Spines small, pointed, and strongly flattened.....C. 3. Adelocephala.
 Spines modified into knobs, with rudimentary conical setæ.
 Four knobs on thorax and caudal horn slightly enlarged, discolorous.

S. 6. Samia.

3. Knobs all equal, blue......S. 7. Philosamia.

Key to genera, pupa

1. Pupæ with flanges on anterior edges of movable segments; not telescoping when dried; cocoon slight or absent; cremaster present.

2. Cremaster distinctly bifurcate, though sometimes shorter than wide; metathorax with a pair of prominent dorsal tubercles; pupa in ground.

(Citheroniidæ).

- 3. Pupa not spinose, cremaster broader than long, not over 2 mm. long. C. 1. Citheronia.
- 3. Pupa more or less spinose; cremaster at least twice as long as broad. 4. Pupa sparsely spinose; posterior row of spines on segments 5, 6, and 7
 - 4. Pupa densely spinose, anterior row of spines on segments 5 to 7 usually longer than posterior.

5. Pupa with a raised transverse ridge at middle of eighth segment

of abdomen, dorsally.....C. 4. Anisota. 5. No such ridge.....C. 3. Adelocephala.

2. Cremaster short, more or less rudimentary, not bifurcate.... (Hemileucinæ). 3. Front edge of segments, above flange-plates, with sharp transverse ridges;

3. Front edge of segments above flange-plates smooth, or with fine longi-

tudinal striation; pupa uncovered.....S. 1. Hemileuca.

1. No flanges at anterior edges of abdominal segments, the segments telescoping within each other in a dried specimen; pupa always in a cocoon.

(Saturniidæ).

2. Movable part of abdomen (segments 5 to 7) strongly tapering toward rear; pupa strongly humped; cocoon in our species without trapdoor.

3. Anterior slopes of movable segments of abdomen with three transverse ridges on sides, cremastral hooks in a circular group....S. 3. Tropæa.

- 3. Anterior slopes of movable segments merely granulose on sides; cremastral hooks in two circular groups......S. 4. Telea.
- 2. Movable part of abdomen nearly cylindrical, the pupa less humped; cocoon with a trapdoor.

- 3. Maxillæ a fourth the length of wing or less, not strongly concave on outer side; anal angle of fore wings opposite incisure between third and fourth segments, or further back.

 - 4. Maxillæ about one-fourth length of wings; male antennæ even in width most of their length, abruptly narrowing to the blunt apex; female with part of glazed eye exposed; very large species

S. 6. Samia.

Family 38. CITHERONIIDÆ

(Ccratocampidæ)

Male antennæ doubly bipectinate about halfway to apex, with simple laminate apical fourth or more. Fore wing with \mathbf{M}_1 more or less stalked with \mathbf{R}_{2-4} , parallel to \mathbf{M}_2 , middle discocellular vein well developed, cell closed in both wings. Hind wing with 2d A preserved. Abdomen more or less conical, extending beyond hind wings, sometimes far beyond. Female with antennæ simple; rarely singly bipectinate, with simple apex (*Adelocephala bicolor*); abdomen stouter than in male, and wings often more rounded.

The moth rests with wings folded in a triangle, somewhat rooflike. Egg with rather thin translucent shell, with faint, fine hexagonal sculpture, most distinct in Citheronia. Larva in first stage with primary hair, except in Citheronia; ninth segment of abdomen with a middorsal spine (often rudimentary), anal plate tuberculate or spined; spines more or less horn-like, never with long spinules, and always strongly unequal. Pupa active, formed in the ground, with bifurcate cremaster without hooks, and flanged segments. The pupa hibernates.

A wholly American family, as defined here. Some authors would add to it various genera (mostly exotic) associated with Automeris and Hemileuca, and use the lack of a cocoon as the primary family character.

1. CITHERONIA Hübner

(*Ceratocampa* Harris)

Palpi projecting slightly beyond front; tongue rudimentary but coiled. Fore wing with middle discocellular very short, straight and oblique outward; hind wing with a weak rudiment of \mathbf{R}_1 , running from \mathbf{R} to Sc (fig. 419). Larva (fig. 423) with numerous strong recurved branching spines, the subdorsal ones on meso- and metathorax and caudal horn, very large; the subdorsals on prothorax and laterals of meso- and metathorax also large, and all well developed. Spines in younger stages relatively longer, with fewer spinules.

1. C. regalis Fabricius. Fore wing grayish olive, more rarely purple, with orange-red veins and pale yellow spots; a yellow spot at base, one near end of cell, and a broken postmedial series; hind wing mostly orange, somewhat redder on the veins, with a large yellow area toward costa and a smaller one toward inner margin, neither reaching outer margin; rarely, with some olive-gray shades between veins. Body orange-red, striped with yellow. In variety infernalis Strecker, the yellow on the wings and body is replaced with orange-red; in variety sængeri Neumægen, the body and markings of the fore wings are wholly yellow. 110-160 mm. (H 10:3. H 1:4, larva).

Caterpillar blue or green; more rarely, brown or pinkish, with oblique dark and light lateral shades, which are often more or less orange, and black horns, the longer of which are orange at the base. Leg-plates orange. A pair of large squarish black dorsal patches on second incisure of the body, separated by a fine mid-dorsal line; smaller patches on the incisures in front and behind. Food, many shrubs and trees; including the walnut family, persimmon, sweet gum, and cotton. There is one brood, the moth flying in June.

Massachusetts to Illinois, Missouri, and south; rare northward. New York: Brockport, Utica, Lansingburg, Albany, Oak Hill, Clermont, Poughkeepsie, New Windsor, Peekskill, South Nyack, Staten Island, Long Island.

2. C. sepulcralis Grote and Robinson. Dull brown with a darker shade over the end of the cell and usually a small yellow spot at the base of the fore wing; hind wing with a little reddish shading. 75-100 mm. (H 41:5). June; one brood. Caterpillar dull brown with irregular mottled oblique stripes

June; one brood. Caterpillar dull brown with irregular mottled oblique stripes on sides, and rather short orange horns, the front ones being little if at all longer than the eaudal horn. Black spot on incisure of thorax smaller than in *C. regalis* and not divided. Food, pine (pitch and white).

Southern Maine and New York to Florida; very rare north of Florida.

2. EACLES Hübner

(Basilona Boisduval, Dryocampa)

Palpi smaller than in Citheronia, not at all projecting beyond the front; male antennæ with a couple more segments peetinate (about 20 in all). Fore wing with \mathbf{R}_2 lost (fig. 420), middle discocellular right-angled at middle and relatively long, in some species longer than lower discocellular. Hind wing with \mathbf{R}_1 lost. Female much larger than male.

1. E. imperialis Drury. Lemon yellow with diffuse dull-rose markings; antemedial line far out, strongly irregular, and excurved; postmedial line oblique, running to apex, on fore wing, and straight across hind wing. Discal spots annulate. Disc of thorax, tegulæ, and dorsum of abdomen largely dull rose. Wings with scattered blackish points. Male with considerable pinkish shading on basal part of fore wing, and a triangular area on outer margin, its upper half bounded by the postmedial line. Variety **punctatissima** Neumogen is very heavily dusted with blackish, the dark dominating the yellow; race **didyma** Beauvois, from the Gulf strip, in the male has the yellow of the fore wing confined to a triangle resting on the middle of the costa, and the outer part of the hind wing brown; in the female, the base of the fore wing and outer part of both wings brown. (H 11:13 d; 10:2 Q).

June; one brood. Caterpillar with sparse but long and conspicuous white secondary hair; spines about 3 or 4 mm. long, on second and third segments of body, and caudal horn, the others rudimentary. Color green, brown, or nearly black, the dark forms often with large russet patches about the spiraeles; horns usually yellow, and anal and leg plates dusted and edged with yellow. Food, many trees, especially pine, maple, oak, cherry, cedar.

Ottawa, Canada, and south. New York: general from Peru southward.

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3. ADELOCEPHALA Herrich-Schæffer

(Sphingicampa Walsh, Othorene Boisduval)

Imago practically indistinguishable from that of Anisota; average size a little larger, discal dot of fore wing, when visible, double; inner margin of fore wing, measured to tip of A, fully as long as outer margin. Hind wing shaded with deep rose in our species. $\dot{\mathbf{R}}_{i}$ free in the few specimens examined (fig. 421) but from extreme end of cell. Caterpillar entirely unlike that of Anisota, with a variable number of strongly flattened conical horns which are silvery on outer side, the two on the mesothorax not much longer than the others. Pupa smoother than that of Anisota.

* Female antennæ bipectinate, wings blunter; pupa with hooked spines on shaft of antennæ (Sphingicampa).

1. A. bicolor Harris. Dull other to fawn brown, dotted with blackish; usually with two superposed silver-white discal dots in a darker shade; postmedial line diffuse, running to costa about 3 mm. before the apex when distinct, sometimes hardly traceable. Hind wing crimson, shading into brown on outer margin, with a vague darker crimson postmedial band. Variety immaculata Jewett lacks the dark dusting --- wholly in the female, and almost completely in the male. Variety suprema Neumogen has the outer third of all wings and the body at least with ash-gray ground; the name is often used in error for a variant with more black dusting than usual. 60-75 mm., male usually smaller. (H 10: 53, 69.)

Caterpillar granulose, usually green with a bicolored lateral stripe, reddish anterior spines, and caudal horn, and a variable number of silver spines, usually developed on alternate segments; on locust and Kentucky coffee tree. Three broods, May; July; August; hibernating in pupa. Mississippi Valley, north to Ohio and Iowa, a stray specimen taken in New Jersey.

New York: (Grote, presumably from Buffalo).

** Female antenna simple, wings more pointed, pupa with blunt tubercles on shaft of antenna (Adelocephala).

2. A. bisecta Lintner. Fore wing ochre yellow, brighter than in A. bicolor, and never with a decided gray border; typically without any blackish dots, but well sprinkled in variety nebulosa Neumorgen. Inner line usually obscure, especially in male, outer strong, straight, pinkish, running practically to apex; discal dots small, white, or more often obsolete. Hind wing lighter ochre, heavily shaded with crimson; darkest in fold, with a faint postmedial shade or none. 55-75 mm.

Two broods-late May and July. Caterpillar similar to that of S. bicolor, but on the average with fewer spines; sometimes with very few (a few pairs on the thorax, two on the sixth segment of the abdomen, and the caudal horn); the spines of the thorax paler, the lower on the metathorax silvery; a brown and yellow lateral stripe. Larva social when young. Eggs very flat, laid in a cluster, browner than those of A. bicolor. Food Gleditschia.

4. ANISOTA Hübner

.(Dryocampa Harris)

Similar to Adelocephala, with head a little more prominent, antennæ with about sixteen pectinate segments, simple in female; fore wing typically markedly triangular, with inner margin distinctly shorter than outer; but always more rounded in female, and in both sexes of *A. stigma* practically like that of *Adelocephala* bicolor, even less triangular than A. bisecta. R_2 lost; M_1 , in the specimens exam-

ined, very shortly stalked (fig. 422). Hind wing more or less extended at inner margin, and sometimes lobed.

Larvæ (fig. 424) with long cylindrical subdorsal horns on mesothorax, in all stages, and with the other horns reduced to short spines or nodules, longest in *A. stigma*. Pupa very rough and spiny, with a strong servate ridge across middle of segment eight of abdomen; formed in the ground.

The moths of the three latter species are not always distinguishable, but the larvæ are strikingly different.

Key to the species

(Adult)

1. No discal spot.....l. rubicunda. 1. A white discal spot.

2. Males (antennæ pectinate).

3. Fore wing opaque, and sprinkled with brown dots......2. stigma. 3. Fore wing translucent on disc, and not sprinkled.

4. senatoria, 3. virginiensis.

2. Females (antennæ simple).

- 3. Fore wings translucent, not sprinkled with brown dots....3. virginiensis.
- 3. Fore wings opaque, sprinkled with brown dots....2. stigma, 4. senatoria.

(Larva)

Green	ı.
Yellow-brown	ı.
Pink and gray	s.
Black and yellow	ı.

I. Moth without discal dot; larva with short spines, pupa with hooked spines on antennal shaft (Dryocampa).

1. A. rubicunda Fabricius (Spiny maple worm). Bright straw yellow (nearly white in the western variety alba Grote), with broad diffuse pink ante- and post-medial lines, or (in the eastern form only) with the basal and outer thirds more or less completely filled with pink; hind wing with a slight pink postmedial shade, or wholly yellow. Male 40, female 50 mm. (H 8:11.) One broad in the north (June and July), two broads southward (May and June; August). Cotenillar stringd in two shades of green with a pink subventral bar

August). Caterpillar striped in two shades of green, with a pink subventral bar toward tail; spines black; feeding on maple, rarely in injurious numbers, occa-sional on oak. Just before pupation the caterpillar turns dark brown with obscure transverse banding.

Generally distributed, north to Quebec. New York: general.

II.. Moth with white discal dot; larva often with long spines; pupa with blunt tubercles on antennal shaft (Anisota).

2. A. stigma Fabricius. Sexes nearly alike, but wings a little squarer in male. Bright ochre yellow, dotted with brown, often heavily; outer margin sometimes with a little gray-brown shading. Antemedial band obscure, postmedial straight, on fore wing running almost to apex, somewhat diffuse. Male 45, female 60 mm. (H 11:9 male, 10 female.)

One and two broods, the first in May and June. Caterpillar dull yellow-brown, spines strong, even on prothorax. Food, oak, and rarely, hazel. Massachusetts to Ontario, Illinois, Kansas, and south. New York: Karner, Lansingburg, Poughkeepsie, Staten Island, Brooklyn. Records in this genus to be

quite safe should be based on the larva.

3. A. virginiensis Drury. Male with outer margin of fore wing practically straight; of hind wing, normally distinctly concave; hind wing much longer on inner margin than on costa. Ground deep red-brown, often shaded with yellow-brown beyond middle of costa; disc of fore wing with a nearly transparent patch beyond the large white discal dot, clothed with widely spaced rudimentary scales. Postmedial line obsolete across the transparent patch, distinct toward the costa, antemedial line sometimes traceable. Postmedial line of hind wing below narrower, better-defined, passing well beyond the cell. Body deep yellow-brown. Female with \mathbf{M}_{s} and \mathbf{Cu}_{1} separate; fore wing transparent, otheroous, the outer margin of both wings and the base of the fore wing somewhat suffused with pinkish; postmedial line rather weak, discal dot often small. Male 40, female 50 mm. (H 8:9 male, 10 female.)

One brood in June. Caterpillar blackish, heavily dusted with cream-white tubercles; with two pairs of broad pink stripes; smaller spines nearly 2 mm., long in large part, but reduced to tubercles on prothorax. On oak.

Quebec to Minnesota and sonth. New York: Plattsburg, Buffalo, Lancaster, Ithaca, Salem, Albany, Staten Island, Brooklyn.

4. A. senatoria Smith and Abbot. Male practically like male *virginiensis*, normally with fore wing a little shorter, with more convex outer margin, somewhat less translucent disc, smaller discal dot, postmedial region of costa no yellower than the base and outer margin, and no trace of antemedial line; hind wing shorter, more rounded than in *A. virginiensis*, with nearly straight outer margin. Hind wing below with postmedial line more outfine, crossing lower angle of cell. The range of variation of this species and of *A. virginiensis* seems to overlap. Female indistinguishable from female *A. stigma*, but on the average smaller, narrower winged, and with fewer brown dots; normally with M_3 and Cu_1 short-stalked. Male 40, female 50 mm.

One brood in June. Caterpillar on oak, sometimes on beech; black, striped with bright yellow, with black head; all the lesser spines reduced to tubereles less than 1 mm. long.

Quebee to Minnesota and south, rarer westward. New York: Karner, Bronxville, Staten Island; Yaphank, Long Island.

Family 39. SATURNIIDÆ

Moths large or very large, some exotic forms being almost the largest of known Lepidoptera. Body stout and heavy, but typically small in proportion to the enormous wings, relatively large in the Hemileueinæ. Vestiture hairy and dense. Antennæ plumose in male, in none of our species with simple apex; typically pectinate in female. Mouth parts much reduced, the tongue, when recognizable, usually too short to coil. Fore wing with M, free from R-stem in all the eastern species (stalked in the western genus Coloradia), typically elosely associated with M₂; cell often open. Hind wing with 3d A rudimentary except in Hemileuca. Egg with heavier shell than in the Citheroniidæ, usually somewhat rough. Larva with densely bristly spines, at least in younger stages, never with primitive first stage; tubercles i of segment eight of abdomen fused into a caudal horn in all our species, separate in Saturnia; ii of segment eight rarely developed into a spine; ii of segment nine fused on middle line only in the Hemileucine, which have subequal bristly spines and a smooth anal

plate. Pupa in a cocoon (except in Hemileuca), which is often complex in structure. Cremaster simple when present, often represented merely by a tuft of hooked setæ or absent; surface not spinose; metathorax without subdorsal tubercles, but often with a broad depression where it abuts on the wing. Abdominal segments frequently without flanges and telescoping when dried.

A large tropical family of very striking moths. Automeris is in many ways (save in the loss of 3d A), the most primitive genus. Hemileuca is derived from nearly the same point, the others (Saturniinæ or Attacinæ) are more specialized and rather closely related. All but Hemileuca hibernate in the pupa.

1. HEMILEUCA Walker

(Euchromia Packard; Saturnia, in part)

Male antennæ singly bipectinate, of about 40 segments, female shortly pectinate, with about four simple joints at base. Vestiture of longer and losser hair than usual; fore tibia with a terminal claw. Abdomen usually slightly exceeding hind wings, with a terminal red tuft in male. Fore wing (fig. 416) with evenly rounded apex and outer margin, \mathbf{R}_2 lost, upper discocellular short, but longer than middle discocellular, and longitudinal. 3d A tubular, joining 2d A. Hind wing with 3d A more than half as long as 2d A, normal, the wing croupled next to body. Scaling thin a mixture of forked and trifd scales and hair scales making the wing transthin, a mixture of forked and trifid scales and hair scales, making the wings trans-

lucent, especially in *H. lucina*. Markings characteristic. Eggs laid in a large cluster (100) about a twig. Larvæ (fig 418) social and solitary in last stage. In all stages (except possibly when very small) with bushy branching spines tipped with nettling hairs, the two subdorsal rows on abdomen distinctly shorter than the others; and in the first stage each tipped with a long primary hair; mid-dorsal spines on both eighth and ninth segment of abdomen. Our species both have a black ground color, with yellow dots at the bases of the secondary hairs, most of the spines black, but the subdorsal row mostly rust-

red. In some specimens the light dots become confluent.

Pupa with flanges on anterior edge of movable segments, normally with cremasteral hooks, but pupating at the surface of the ground without forming any Hibernation in the egg. cocoon.

Sometimes pupze go into hibernation and do not emerge until the following year. 1. H. maia Drury (Crape moth, buck moth). Smoky black, a cream median band, normally covering a third, buck model. but often narrow, and absent on the fore wing in ab. lintneri Packard. Discal spots large, black with central white lunules. Hind wing similar. Thorax and abdomen black, with very little or no white hair; posterior thoracic tuft and anal tuft of male red; collar partly yellow. 50-65 mm. (**H**[·] 11:1.)

Common locally on oak barrens, flying very swiftly by day in September and October. Caterpillar almost always on scrub oak, black, dusted with light vellow, the dusting heavier above and gradually fading out on the sides; dorsal tubercles largely black, but red at base. In the South a yellow form of the caterpillar seems to occur with the black concentrated into a subdorsal band, and contrasting red dorsal spines. Head red.

Massachusetts to Colorado and south. New York: Karner, Albany, and general on Long Island.

H. latifascia Barnes and McDunnough appears intermediate between maia and

lucina; it has the thin wings of *H. lucina* and the light band even broader, but is as large as normal maia, and looks somewhat suffused. Larva like lucina; apparently on willow. Manitoba; Wiseonsin; South Dakota.

2. H. lucina H. Edwards. More translucent than *H. maia*, paler, the median band normally occupying fully a third of the length of the wings; the body with much white hair on the black portions. 50-60 mm.

Not rure in woods northward, replacing *H. maia*, and overlapping it in Massachusetts and Colorado. September. Caterpillar black, dusted with yellow, the yellow gathering into a broad stigmatal band, as in some southern larvæ of *H. maia*. Food, Spiræa. The larva has never been found on oak.

In aberration obsoleta Reiff, the pale band is interrupted on the fore wing, in aberration lutea it is yellower than usual.

2. AUTOMERIS Hübner

(Hyperchiria Hübner, Io Boisduval)

Fore wing with strongly arched costa, and, in our species, right-angled apex; hind wing rounded, slightly exceeding abdomen, at least in female. Male antennæ doubly bipectinate to apex, the outer pectinations rather shorter; female nearly simple. Palpi extending distinctly beyond the front, which is closer-scaled than in Hemileuca. No claw ou tibia. Fore wing (fig. 415) with \mathbf{R}_2 lost, upper discoccellular short and oblique, middle discoccellular transverse, but rather shorter than the weak and ben lower discoccellular. \mathbf{M}_1 and \mathbf{M}_2 divergent. Hind wing with cell closed, 3d A almost obsolete, as in the rest of the family. The moth rests with the wings folded in a triangle. A large genus in South America.

Eggs ovate, white, with a black spot at one end, laid in a flat cluster. Larva processionary when young, the families gradually breaking up, as in Hemileuca. Spines like those of Hemileuca in arrangement and structure; poisonous. Subdorsal spines rather longer than lateral; in stage 1, with a group of terminal setæ. Pupa flanged, in a thin cocoon of a single layer of rather brittle silk, between leaves on the ground. Hibernation in pupa.

1. A. io Fabricius (Corn emperor). Fore wing of male bright yellow, of typical female dull pinkish brown; antemedial line zigzag and outwardly oblique; postmedial line waved and nearly parallel to outer margin; both dark in male and in typical female. A dark brown discal bar, surrounded by a series of dark points, or the whole fused into an irregular patch; the female with more or less whitish scaling on the antemedial and postmedial line. Hind wing bright yellow, a little darker in female; a large blue ocellus on end of cell, centered with a white bar and encircled in a heavy black ring; a fine black postmedial line and a crimson subterminal band; inner margin broadly crimson. In the female variety lutheri Cockerell (fusca Luther, not Walker), which is dominant northward, the ground is deep purple, shaded with olive, with strongly contrasting greenish white ordinary lines and dots about the discal bar. In the Florida race, lilith Strecker, the ground of the fore wing is red in both sexes, and the hind wing orange. Under side yellow, shaded with crimson, with straight dark postmedial lines on both wings, and a large white-pupilled black ocellus on fore wing. Male aberration **argus** Neumoegen and Dyar has all the markings lost except the ocelli on the upper side of the hind wing and under side of fore wing. Male 60, female 80 mm. (H 9:4 male, 5 female.)

June to August, one brood. Caterpillar brown when young, with brown and black spines, becoming dull yellow and then green, in the later stages with a red and white stigmatal line. A general feeder. Cocoon brown, pupa black.

Common and generally distributed. New York: Wilmington and North Creek (Adirondacks), Buffalo, Ithaca, McLean, Oneonta, Albany, and south. One Ithaca specimen (male) is close to variety lilith Strecker.

3. TROPÆA Hübner

(Actias, in part; Attacus, in part)

Male antennæ very broad and plumose, doubly bipectinate, with equal branches; narrow, with alternate branches only half as long in female. Front with conical tuft, palpi distinct. Fore wing with arched costa, rounded apex, and somewhat concave, rarely scalloped, outer margin; hind wing with general form an equilateral triangle, but inner margin drawn out into a long twisted tail, supported by veins M_{s} , Cu and A. R, long-stalked (fig. 414) with R_{2+33} , R_2 distinet, R_4 shortly stalked. Upper discoellular rather short (unlike Actias), M_1 and M_2 strongly divergent, eell closed, the vein erossing a transparent spot. Hind wing with dorsal venation somewhat crowded, costal widely spaced; middle discocellular very long and longitudinal, as well as upper discocellular; cell closed like that of fore wing. Male abdomen rather small and conical in repose; female abdomen more eylindrical. Fore wings thrown back over hind wings in repose. The moth hybridizes with Actias selene, and the genera should probably be united.

Eggs laid in small groups, dark brown, blotched with white. First-state larvæ densely spinose, but with no unpaired spine on ninth segment of abdomen; green, rarely longitudinally striped with black; with black-barred head, secondary hair notably denser than in Telea; in later stages, body less humped than in Telea; turning dark brown immediately before pupation.

1. T. luna Linnæus (Empress, luna moth). Wing membrane bright green, partially covered with scales which are almost all bright yellow in male, largely white in female. Body white, a pinkish powdery band across collar, and on costa of fore wings, joined by a bar to diseal occllus; a waved erimson stripe on abdomen; fringes typically yellow. A black yellow, crimson, white, and transparent diseal eyespot on each wing. The earlier spring specimens are variety rubromarginata Davis, and have the margins broadly edged with erimson, the costal stripe deeper red, and the postmedial line almost always distinet. The subtropical race dictynna Walker has shorter tails. 100 mm. (H 12:7.) May and June; August. Caterpillar at first green with black and white head,

May and June; August. Caterpillar at first green with black and white head, later usually with green head (rarely reddish like *T. polyphemus*); each segment of body with a fine white line around the sides and back. Food walnut, birch, beech, and many other trees. Cocoon thin but tough, of a single layer of brown silk (heavier and white in *dictynna*).

Common and general, especially in wooded regions. Two broods (the second apparently partial). New York: Buffalo, Roek City, (Cattaraugus county), Ithaca, Trenton Falls, Oneonta, Schoharie, Albany, Saratoga Springs, New Windsor, Staten Island, Brooklyn.

4. TELEA Hübner

(Attacus, in part)

Male antennæ plumose, wider than in any other Saturniid, equally doubly bipectinate, female antennæ narrower than in Tropæa, with alternate peetinations vestigial, palpi minute. Body larger than in Tropæa. Fore wing with rounded, but strongly falcate apex. \mathbf{R}_1 free, \mathbf{R}_2 lost, upper diseoeellular very long, longer than in hind wing, and longitudinal, middle discocellular nearly obsolete. Hind wing with outer margin bent at middle, and often somewhat waved, apex often a little produced. Cells closed. Eggs white, with a brown band around the edge, laid in small groups, Cater-

Eggs white, with a brown band around the edge, laid in small groups, Caterpillars (fig. 417) very strongly humped, hardly more than twice as long as high when at rest. Cocoon ovoid, densely woven of two firm layers with a little flosssilk between, without trapdoor; rarely, suspended by running a band of silk up to the stem along the petiole of one of the leaves in which it is enclosed, and

forming a ring about the stem; ecocoon usually falling to the ground. The moth secretes a good deal of acid from the mouth before emerging, with which the silk is softened; and then cuts the silk with a hook developed out of one of the basal sclerites of the fore wing.

1. T. polyphemus Cramer. Grounds typically tawny yellow, dusted with blackish, but varying from cream color to olive or black-brown; outer margin and base rarely darkened. Collar and costa ash gray; antemedial line red and white, offset on Cu; postmedial line more than three-fourths way to margin, pinkish white, preceded by a blackish shade, and by a double black spot at costa; median shade faint, dark. Discal eyespot of fore wing with a good-sized hyaline center, ringed with yellow and finely with black; hind wing with a large black patch, enclosing the discal eyespot in its outer part; and largely filled with a blue shade; separated from the ocellus by a fine black line. Postmedial line heavier than in fore wing. Under side marbled in several shades of brown, quite unlike upper side; the wings folded over the back at rest. 125 mm. Dwarfs (down to 65 mm.), and giants (150 mm.) occur. (H 9:1.)

One brood in the north, in June and July; two southward. Caterpillar on maple, birch, and many other trees; translucent grass green, with pinkish head, more or less pinkish shining tubercles and thin slightly oblique whitish bars on sides. Young caterpillars have a darker brown head, and relatively longer spine. The color does not change before pupation.

Common and generally distributed. New York: Common everywhere in the State.

5. CALLOSAMIA Packard

(Samia; Attacus, in part)

Male antennæ plumose, the pectinations alternating in color, but scarcely in length; female antennæ about half as wide, with shorter pectinations also nearly as long as the longer ones. Mouth parts obsolete; male with very small body, female body large, but much shorter than the wings, which are strongly concave along the inner margin. Fore wing with rounded but produced apex, especially in male, and concave outer margin; hind wing produced at anal angle in male, margin rounded. Fore wing with \mathbf{R}_2 preserved, \mathbf{R}_1 free (fig. 413) upper discocellular very long, cell open, and \mathbf{M}_1 and \mathbf{M}_2 stalked in both wings. Wings folded over back at repose.

Eggs whitish, blotched with dull red cement, more rounded than in Telea and Tropæa. Larva, when young, as usual, with banded head; full-grown larva with two pairs of large blunt tubercles on meso- and metathorax, and one dorsal one on segment eight of abdomen; the others reduced to minute black points. Pupa nearly cylindrical, largely bright yellow-brown. Cocoon in a folded leaf, which is normally attached to its twig by enclosing the petiole and a short bit of the twig in silk. Cocoon proper oval, double, with a filling of floss silk between the two denser layers; each layer with a trapdoor at head-end of cocoon for emergence of moth, formed by a group of converging longitudinal loops of silk, which spring together again after emergence. The pupa rarely passes two years before emergence.

Key to the species

- 1. Outer margin bright ochre yellow; base of hind wing; below, largely rusty
- without yellow tint.
 - 2. Fore wing, above, blackish; antennæ broader (males).
 - 3. Discal spots above large and angulate.....2. angulifera. 3. Discal spots faint or absent.....l. promethea.

2. Fore wing above blackish or red-brown toward base, without decided yellow tint; pinkish red-brown in postmedial area.....1. promethea Q.

2. Fore wing broadly shaded with othre yellow, especially toward base.

2. angulifera Q.

1. C. promethea Drury. Male nearly black, a little shining when fresh; postmedial region slightly paler; antemedial line lost; discal dot usually lost, or represented by a faint pale spot; postmedial line waved, fine, pale, a little more erect than outer margin; margin clay color, with a fine deeply waved black subterminal line; apical region, except beyond subterminal line, shaded with crimson; a waved oblique white apical line in cell \mathbf{R}_{a} and a large rounded black, brown, and blue eyespot in cell \mathbf{R}_i . Hind wing similar, without the apical marks, with a chain of blackish spots before the subterminal line. Under side of fore wing similar, the postmedial area much paler, especially near the postmedial line, pinkish, and powdery. Hind wing with ground deep red-brown, the costa and postmedial space as in fore wing; discal bar often contrasting, but not large. Female with base of wings dull red, sometimes with a slight orange tint, shaded with blackish; antemedial line pale, defined with blackish, bent at a right angle over cell; postmedial line white, waved, preceded by a strong black shade; discal spot pale, edged with blackish, bar-shaped or angulate, not touching postmedial line. Postmedial region much paler, pinkish, and powdery; marginal marks like those of male; hind wing similar to fore wing, the dark blotches before the waved subterminal line dull red. Under side dull red, marked like male, the markings

lightly defined with blackish, especially on fore wing. 75 mm. (**H 8**:3 β , 4 9.) June to August, the second brood partial. Male flying by day, female by night, and not normally flying at all until after mating. Caterpillar on many trees, preferring sassafras and spicebush, wild cherry, and tulip tree. On hatching, yellow, with head and body heavily striped transversely with black; soon turning green. When full-grown, blue-green, somewhat pruinose, with the four thoracic humps red and the caudal one yellow, their bases ringed with black. No stigmatal line (**H 1**:2). Cocoon always suspended, twice as long as wide.

Common and generally distributed, north to Montreal, Quebec. New York: Buffalo, Ithaca, Elmira, Saratoga Springs, Albany, and common southward.

2. C. angulifera Walker. Pattern similar to C. promethea; male somewhat lighter umber brown, with strong angulate discal white spots on both surfaces; female heavily shaded with yellow, the angulate discal spots very large, and often crossing the postmedial line. Male as well as female nocturnal. (H 11:11 σ_{1} , 12 Ω .)

Two broods, common southward. June; August. Caterpillar similar to that of *C. promethea*; when young, normally with head only striped, later with a clear yellow substigmatal stripe; on various trees, but especially tulip tree. Cocoon usually not attached to twig, and so falling to the ground with the leaves.

New England and south. New York: Buffalo, Ithaca (rare), Dutchess County, Staten Island, Brooklyn.

C. carolina Jones (*securifera* Maassen and Weymer?) approaches our southern boundary. The moth is suffused on both sides in both sexes with bright ochre yellow; discal spots of male small, as in *C. promethea*. Caterpillar like that of *C. promethea* in all stages; cocoon suspended, very large, the outer cocoon inflated and three times the diameter of the inner. Food *Magnolia glauca* only (refusing other Magnolias, but accepting tulip tree). South Carolina; Alabama; Florida (figured by Packard as *C. angulifera*).

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6. SAMIA Hübner

(*Platysamia* Grote; *Attacus*)

Very close to Callosamia; larger, sexes alike, outer margin less concave; \mathbf{R}_2 of fore wing lost; palpi recognizable. Egg like that of Callosamia (but larger); larva also similar, almost identical when young, but when mature with all the warts good-sized, rounded, studded with minute conical black setæ, which, on most of the warts, form rings about their tips; subdorsal warts of second to fourth segments of body more rounded, swollen, and tending to be discolorous; homologue of the eaudal horn also swollen. Cocoon double, with a trapdoor at the anterior end, as in Callosamia, but fastened longitudinally to a twig; when first spun surrounded by an enclosure of leaves which soon weathers away.

1. S. cecropia Linnaus (Emperor). Body red, collar and abdomen striped with white; wings blackish, powdery, more or less shaded with red, lines and marginal markings about as in female promethea; antemedial line pale, defined with a dark shade; postmedial white, nearly even, preceded with blackish and followed by a wide red band; discal spots reniform, red and white, opaque; apical and marginal marks about as in promethea, but with less erimson at apex, and with some violet toward costa. Postmedial region blackish, with black spots along its outer margin. Under side similar, much more powdery, the costa of hind wing nearly white. 125-165 mm. (H 8:1.)

One brood in June. Caterpillar on many trees and shrubs: apple, elm, wild cherry, Spiræa, etc. When first hatched, black, gradually becoming red, orange, yellow, yellow-green, and apple-green, the color-changes not always occurring at The enlarged warts are larger from the beginning. Full-grown larva moults. with subdorsal warts of meso- and metathorax dull red, spotted with black cones, those on first segment of abdomen orange-yellow, the rest pale yellow; laterals all pale blue. (H 1:8). Skin grass-green in next to last stage, and the anterior warts scarlet, as in S. columbia. Cocoon fusiform, pointed at both ends, and usually with one or both ends attached to the twig; the outer cocoon sometimes much inflated, especially when spun on low shrubs. Silk reddish when fresh. Generally distributed, north to Quebec. New York: general, not rare.

2. S. columbia Smith. Similar to S. cecropia, much duller, with red shadings reduced, body with a good deal of gray-brown; no red beyond postmedial line; discal spots on fore wing obscure or wanting; less red in apical region; ocellus usually with a small blue lunule only, and no brown, mostly black. Markings often suffused in northern specimens. 100 mm. (H 8:8.)

One brood, June and July. Caterpillar similar to that of cecropia, but grassgreen in last stage, the anterior tubercles bright coral-red with black rings at their bases. Food, larch only. Cocoon ash gray, mottled with silvery streaks, more rounded than that of *cecropia*, and free at both ends. The western \vec{S} . nokomis, described as a variety of this species, belongs more probably to S. gloveri. The western limit of the true conifer-feeding columbia is uncertain.

Southern New Hampshire to Michigan and northward.

7. PHILOSAMIA Grote

(Samia; Attacus, in part)

Near Callosamia and Samia. Female antennæ broader than usual; palpi slender and reaching front; fore wing with apex more produced than in Samia, fully as much as in male promethea, with outer margin sharply bent in above middle. \mathbf{R}_2 very short, \mathbf{M}_1 and \mathbf{M}_2 hardly stalked, cells open, discal lunules transparent; hind wing long on inner margin, especially in male. Abdomen with three rows of small raised white tufts (in an exotic species with raised ridges).

Eggs relatively small, ovoid, without dark cement. Caterpillars as usual at hatching; when full-grown, with all the tubercles blunt, somewhat tapering, subequal, and of the same color (blue). Cocoon constructed like that of Callosamia, between two or three leaflets of the food plant, the sheathing of silk extending up the midrib of the compound leaf to the twig; cocoon more fusiform than that of Callosamia.

1. P. walkeri Felder. Olive (readily fading to yellow-brown); antemedial band white, running out on forks of $\hat{C}u$, and touching the discal lunule; postmedial white, with black before it, and a broad pink shade beyond, bent out at discal lunule, concave above and below; outer part of wing powdered, black and yellow or olive, shading into pink near apex. Apical eyespot small with a white crescent on its inner side; subterminal line close to margin and nearly even. Hind wing with postmedial line running to anal angle, and subterminal line preceded by a broader inner line. Discal lunules, starting from basal side, black, white, transparent, white, and yellow. Under side similar. (cynthia auct., not Drury). (H 9:2 as P. cynthia.)

Caterpillar on Ailanthus; rarely straying to wild cherry and other *promethea* foods; at first green, with black head and tubercles; then yellow with pale head and tubercles; finally, light greenish with blue tubercles.

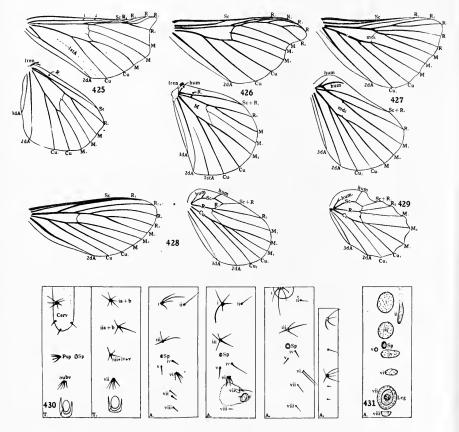
Sometimes several caterpillars spin on one leaf, and their cocoons have a common stem. The moth does not exactly match any of the described races of *P. walkeri*, and may have come from some not fully explored part of China, or be a domesticated mongrel form. It was originally introduced as a silk-worm. Vicinities of New York, Philadelphia, and Washington, District of Colum-

Vicinities of New York, Philadelphia, and Washington, District of Columbia; introduced from the Orient. New York: Livingston, New York City, Staten Island, and vicinity.

SUPERFAMILY BOMBYCOIDEA

(Lasiocampina)

Heavy-bodied and usually medium-sized moths (a few exotic forms larger-bodied than most Saturniidæ). Head relatively small, retracted;



FIGS. 425-431. BOMBYCOIDEA

425, Apatelodes torrefacta (Eupterotidæ), venation (The figures 1 and 2 indicate accessory branches of Sc.); 426, Bombyx mori (Bombycidæ), venation; 427, Malacosoma americana (Lasiocampidæ) male, venation; 428, Heteropacha rileyana (Lasiocampidæ), venation; 429, Epicnaptera americana (Lasiocampidæ), venation of hind wing; 430, Bombyx mori (Bombycidæ), seta map of first-stage larva, showing the characteristic arrangement of tufted setæ; 431, Chondrostega (a primitive, Old-World Lasiocampid), showing arrangement of warts

front normally smooth, with a tuft of hair. Ocelli absent; mouth parts almost completely absent; only the palpi developed, and never large; clypeus sometimes inflated. Antennæ bipectinate, both sets of pectinations ventral; shaft somewhat irregularly but densely scaled above; pectinations often scaled. Female antennæ much narrower, but pectinate like those of male. Body with deep hairy or mixed vestiture, usually stout and woolly looking; abdomen large, often exceeding hind wings, and roughly cylindrical. Legs short, the femora and tibiæ densely hairy, and tarsi somewhat hairy; spurs generally normal. Wings most often broad, loosely and heavily scaled, with heavy veins. Fore wing with \mathbf{R}_2 and \mathbf{R}_3 stalked beyond their separation from \mathbf{R}_4 and \mathbf{R}_5 in the North American species; given off successively from \mathbf{R}_{4+5} in Bombyx, and united in the Eupterotinæ. Accessory cell always absent, and \mathbf{R}_3 and \mathbf{R}_4 never stalked the farthest; \mathbf{M}_1 often stalked with \mathbf{R}_5 or \mathbf{R}_{4+5} , \mathbf{M}_{2} variable in position; 1st A lost, 3d A sometimes distinct and running into 2d A (Apatelodes). Hind wing in primitive forms with Sc and R closely parallel toward base, diverging before end of cell, and connected to \mathbf{R} by a very distinct \mathbf{R}_1 , in Apatelodes (fig. 425) with \mathbf{R}_1 lost, and the veins very closely parallel, in the Lasiocampidæ with the veins tending to withdraw in the higher genera; \mathbf{R}_1 as strong as any vein, and far out toward end of cell; occasionally even short-stalked with \mathbf{R}_s . \mathbf{M}_2 as in fore wing. 1st A lost, 3d A normal. Wings folded triangular at rest, the costa of the hind wing uncovered in the Lasiocampidæ.

Egg normally of flat type; very thin and flat in Apatelodes; laid endwise in a cluster in Malacosoma. Larva with dense tufted and secondary hair (fig. 431), the tufts strong in stage 1 (fig. 430) where an additional subdorsal tuft is often distinctly developed, and iv and v form separate warts; later with tufts reduced and difficult to recognize; hair rudimentary in Bombyx. Many Eupterotidæ (including *A. angelica*) and also some primitive Lasiocampidæ, have scales as well as hairs. Front small, head with dense secondary hair, even on some or all of mouth parts. Prolegs normal, with a single band or biordinal hooks; often more or less spread out laterally. Skin thin and soft. Warts i of eighth abdominal segment often fused in a tubercle or caudal horn, very strong in Bombyx. Pupæ obtect, of normal macro type, varying in details in the families; typically enclosed in a dense silken cocoon. Cocoon cutter and fluid secretion normally as in Telea and its kin (p. 671).

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Family 40. EUPTEROTIDÆ

(Lasioeampidæ, Notodontidæ, Bombycidæ, in part)

Head and body normal for the superfamily; thorax with a longitudinal erest in Apatelodes. Fore wing in the Apatelodinæ (fig. 425) with all veins preserved, \mathbf{R}_2 and \mathbf{R}_3 , and \mathbf{R}_4 and \mathbf{R}_5 long-stalked; \mathbf{M}_1 very shortly stalked; \mathbf{M}_2 nearer **R**-stem than **Cu**-stem, the discocellulars transverse; Cu₂ arising more than halfway out on cell. Hind wing in our species with Sc and R very elosely approximate on basal third of cell, almost obliterating \mathbf{R}_1 ; frenulum normally developed; \mathbf{M}_1 short-stalked, \mathbf{M}_2 and \mathbf{Cu}_2 as in fore wing.

Caterpillar strongly flattened, with at least a few seales or hair scales; prolegs more or less stretched out laterally. Secondary hair (in our species) on mandibles, base of maxillæ, and labium, but not on distal part of maxillæ and labium, elypeus, or labrum. No caudal horn. Pupa resembling that of the Notodontidæ; maxillæ very short, the fore and middle legs meeting on the middle line behind them; abdomen finely punctate, with a row of beading on each movable segment; not clothed with fine setæ; second to seventh segments of abdomen with flange plates at anterior and posterior margins; cremaster short or absent.

Our single genus represents the Apatelodinæ, which are mainly South American; in the Eupterotinæ, of the Old World, \mathbf{R}_2 and \mathbf{R}_3 are completely united, Sc and R of the hind wing are more promptly divergent, and the frenulum is weaker.

APATELODES Packard

Palpi barely exceeding front (torrefacta), or moderate and upturned (angelica); fore wing subfalcate (fig. 425), with outer margin bent at middle and concave on upper half, costa nearly straight. Our species have one or two transparent subterminal spots.

1. A. torrefacta Smith and Abbot. Outer margin even. Pale gray, disc below costa and outer margin somewhat darker; lines fine, dark brown, continuous; antemedial excurved, postmedial bent out below costa and waved somewhat on veins; two fainter lines across median area; a brown patch on inner margin before A; rarely almost obsolete. Hyaline spot small and single. Hind wing redbrown, with a dark and pale postmedial shade parallel to outer margin and ending in one or two deep brown bars at inner margin. Upper side with a strong dark brown shade in cell. 35-40 mm. (H 40:20.)

The light form floridana H. Edwards with the spot on inner margin smaller and light chestnut brown, in place of black-brown, is commoner southward. June and July. Caterpillar with dark body and fine and dense light yellow

hair, with a series of small mid-dorsal black pencils, those on meso- and meta-

thorax and segment 8 of abdomen much longer. It feeds on many trees and shrubs.

New Hampshire to western Pennsylvania and south. New York: Kingston, Pcughkeepsie, New Windsor, Greenwood Lake, Staten Island, Brooklyn and Lynbrook, Long Island.

2. A. angelica Grote. Outer margin scalloped, especially in female; thoracic crest stronger, palpi longer; usually with two hyaline spots. Pale powdery gray; ordinary lines reduced to blackish points on veins, the antemedial followed and the postmedial preceded by shaded brown lines; transparent dots finely edged with brown, with a vague brownish shade beyond them. No darker patch on inner margin. Hind wing browner, the base luteous, with dark median and postmedial shaded lines; outer margin brown. Under side pale, without dark shade in cell. 35 to nearly 50 mm. (H 40:21.)

May to July. Rarer than A. torrefacta. Caterpillar strongly flattened, gray, barklike, with a rough contrasting dark tuft on thorax, and the small dorsal pencils, but no long ones; on ash and lilac, resting on the bark. Southern Maine to Texas. New York: Plattsburg, Bath, Sharon Springs, Albany, Poughkeepsie, West Point, Coldenham; Jamaica, Long Island.

The Chinese silkworm belongs to the family Bombycidæ (fig. 426, 430). Bombyx mori Linnæus.

Moth mostly white, with falcate wings; \mathbf{R}_1 , \mathbf{R}_2 and \mathbf{R}_3 successively given off from stalk of \mathbf{R}_{4+5} ; \mathbf{M}_2 nearly central in both wings; Sc and R somewhat divergent, even from the base, but \mathbf{R}_1 placed well out and distinct; frenulum very weak. The moths have fully-developed wings, but fly little or not at all. Cater-pillars white, strongly humped on thorax, with a more or less distinct lateral eyespot; with a strong caudal horn, and microscopic secondary hair. Cocoon dense, normally of yellow silk. The silkworm is occasionally cultivated in our territory, but is not known wild anywhere. Food black and white mulberry; it will also cat Osage orange, and will also feed, especially when young, on lettuce. (H p. 316 fl. 191-193.)

Family 41. LASIOCAMPIDÆ

(Bombycidæ, Lachneidæ)

Body and appearance about as in Apatelodes; normal. Spurs small. Fore wing normally with all veins preserved, \mathbf{R}_2 and \mathbf{R}_3 long-stalked, \mathbf{R}_{5} and \mathbf{M}_{1} more shortly stalked from end of cell, \mathbf{R}_{4} often arising out of the base of their stalk; \mathbf{M}_2 from **Cu**-stem, middle discocellular vein long and angled. Cu, arising only one-quarter to one-third way out on cell. Hind wing without any trace of frenulum, with humeral angle broad and expanded, extending in front of fore wing at rest and supported by one or more well-developed humeral veins; Sc and R separate at base, then anastomosing or connected with R, forming a cell of variable size; \mathbf{M}_2 arising from **Cu**-stem, \mathbf{Cu}_2 attached rather before middle of end of cell, not so near the base as in the fore wing.

Eggs, when laid singly, with micropyle on side, ovate; eggs of Malacosoma clustered in a ring about a twig, such egg being laid on the previous one so that the micropyle is at the exposed end; the whole

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mass covered by a sort of varnish. Caterpillar (fig. 431) very hairy, with dense secondary hair on all parts except tips of palpi, even on antennæ. (Some primitive forms are like Apatelodes.) Form varying from nearly cylindrical (M. americana) to very much flattened, with lateral lappets on segments (Tolype and Epicnaptera). Notch of labrum not so deep as in Apatelodes. Pupa in a cocoon, which is dense except in M. disstria; more or less densely clothed with secondary hair; palpi well exposed; no cremaster; fore femora covered, and maxillary palpi absent. Epicranial suture distinct.

Key to the genera

1. Humeral cell of hind wing about as large as discal (fig. 429).

- 2. Fore wing with all veins preserved; wings very irregular...4. Epicnaptera. 2. Fore wing with \mathbf{R}_2 and \mathbf{R}_3 completely united; wings nearly even (fig. 428).
- 5. Heteropacha.
- 1. Humeral cell small and inconspicuous.
 - 2. Outer margin of fore wing distinctly concave at middle; cell of hind wing
 - 3. Thorax with even hair.....2. Artace. 3. Thorax with a strong median crest of metallic scales.....l. Tolype.

1. TOLYPE Walker

Body very stont and long, with dense woolly vestiture, looser on tegulae; disc of thorax clothed with shining spatulate hairs; end of abdomen with a loose terminal tuft. Fore wing evenly rounded; cell scarcely two-fifths its length; **R**, short-stalked. Hind wing rounded, with small humeral cell, and a single long humeral vein, curved out at tip; the other veins arising from the cell. Eggs laid in a row, covered with hair from the abdominal tuft. Caterpillar strongly flattened, with lateral lappets, and well-marked subdorsal warts; those

on eighth segment of abdomen not enlarged. Cocoon dense, strongly flattened; on bark. Pupa with secondary setæ very weak; cremaster indicated by an abrupt narrowing of the end of the last segment.

Key to the species

- 1. Smoky gray, the markings nearly obsolete, except for the pale subterminal
- 2. Subterminal band dark gray, contrasting with the whitish base and median 2. Median area dark, concolorons with the subterminal band.....l. velleda.

1. T. velleda Stoll. Head and thorax white, abdomen gray, disc of thorax and sometimes base of abdomen contrasting, blackish. Fore wing ash gray, with white veins, and fine, waved, white, ordinary lines, the transverse anterior and transverse posterior often double; terminal line white, followed by gray bars in the fringe. Base, margin, and filling of lines sometimes pale. Hind wing with vague bands. 30-50 mm. (H 11:7 3, 8 9.)

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Caterpillar gray, finely and obscurely striate, with a dorsal pair of warts on metathorax and a narrow black band between thorax and abdomen, concealed at rest; on apple, poplar, and other trees.

Montreal, Quebec, to Pennsylvania and Michigan. New York: Spier Falls. Rochester, Lancaster, Buffalo, Ithaca, Oneonta, Albany, Bronxville, Staten Island; Southold and Bellport, Long Island.

2. T. laricis Fitch. Male deep smoky gray; head and collar usually white, contrasting; fore wings with traces of the markings of *velleda*, but usually with only the waved whitish subterminal line distinct; veins dark. Female white, with several more or less distinct waved gray bands across basal two-thirds of fore wing; subterminal space broadly ash-gray, terminal space paler gray, separated from it by the wavy white subterminal line. Veins wholly white. Hind wing shading into gray on outer half. $\checkmark 25, 0.25$ mm. Caterpillar dull brown, similar to *velleda*, but with the warts on the fifth seg-

ment of the abdomen stronger than those on the metathorax; on larch. Occasional transitional specimens occur, suggesting that the species may hybridize.

Distribution with T. velleda, but more local. New York: Plattsburg, Rochester, Coldwater, Buffalo, Bath, Ithaca, Albany, Poughkeepsie.

2. ARTACE Walker

Similar to Tolype, but without the metallic hair on the disc of the thorax. Caterpillar less flattened.

I. A. punctistriga Walker. White, with the usual lines represented by four or five rows of black dots on veins of fore wing. 25-35 mm. (H 12:5.)

Caterpillar gray, mottled, with an orange transverse band behind thorax, and a distinct raised lappet on dorsum of eighth segment of abdomen, only. New York and south; west to Mississippi Valley and Arizona. New York:

Brooklyn (Ottolengui).

3. MALACOSOMA Hübner

(Clisiocampa)

Body less loosely hairy than in Tolype, without any central thoracic tuft. Fore wing bluntly rounded (fig. 427), much longer in female than in male, the slight concavity of the outer margin nearly evened up by the fringe; \mathbf{R}_i free, \mathbf{M}_1 only very shortly stalked. Hind wing with very small humeral cell, bearing 2 humeral veins. Caterpillar cylindrical (americana), or slightly flattened (disstria), without lappets or distinct enlarged tubercles; social. Cocoon double, the inner cocoon with the interstices filled with yellow powder.

Key to the species

Two transverse white lines across wing.....l. americana.

1. M. americana Fabricius (Tent caterpillar). Brown (walnut brown of Ridgway) with two white lines, trisecting the wing, fringe cut with white, most distinctly in the male. Median area sometimes paler, or base with whitish suffusion. 325 235 mm. (H 10:12.) Very common and general in distribution, often injurious to apple; also on wild cherry and other Rosacee. When very common, it will eat almost any deciduous tree, but then usually dies before maturity. The caterpillars form a conspicuous to the fork of a breach and in a considered to the fork of a breach and in a constant of silv in the fork of a breach and sing no here a breach and the form of a breach and the form of a breach and the former of a breach and the former of a breach and the former of the former of the former of the former of the breach and the former of the form

conspicuous tent of silk in the fork of a branch, enclosing no leaves, and feed

outside. Eggs in a belt around a twig, the belt more than twice as long as wide (unless dwarfed in some way). Caterpillars mottled and striped with blue, tawny yellow and black, with a continuous white dorsal line. Cocoon dense, of white silk filled with yellow powder, the inner cocoon opaque and outer very slight; usually formed at a distance from the food; but in captivity, often in the tent.

New York: Common throughout the State.

2. M. disstria Hübner (Forest tent caterpillar). Light grayish brown; typically, with two brown lines replacing the white ones of M. americana. In variety sylvatica Harris, the median third is contrasting dark brown; variety thoracicoides is practically immaculate, with traces of pale lines; in variety perversa Neumoegen and Dyar the base and outer margin are darker than the median area; aberration astricta Reiff is immaculate straw yellow, and aberration anita Reiff chocolate brown, with traces of pale lines. 20-37 mm., female larger (H 10:9.)

Caterpillar living in a colony on the trunk or a large branch of the food tree, but spinning only a slight carpet; pale grayish blue, with fine orange and black lines, and a dorsal stripe broken into a series of cream-yellow spots; normally with a larger and a smaller one on each segment. Outer cocoon relatively strong, and inner light, translucent, with only a little yellow powder; often spun on the food plant. Food, forest trees, especially maple; often injurious.

Common and general in distribution; a variant form on the Pacific Coast. New York: Common throughout the State, even on the top of Mt. Marcy. I have seen variety sylvatica from Ithaca, Geneva, and Peru, and variety thoracicoides from Peru.

4. EPICNAPTERA Rambur

(Gastropacha; Phyllodesma, in part)

Palpi moderate, longer than in the preceding genera; vestiture closer; fore wing with margin scalloped and deeply notched at anal angle; \mathbf{R}_4 hardly stalked (fig. 429); hind wing scalloped and deeply notched on costa, humeral cell as large as discal; two humeral veins, one from point of separation of Sc and R, and the other well out; \mathbf{R}_1 long, transverse, arising from R after its separation from cell, \mathbf{M}_2 and \mathbf{M}_3 short-stalked; caterpillar much like that of Tolype, without enlarged warts in our species; two transverse red bars on incisures of thorax, each marked with three black dots, and concealed at rest. Cocoon on bark, flattened like Tolype. Pupa bluntly rounded at rear, less hairy than that of Malacosoma, but more so than in Tolype.

1. E. americana Harris (American lappet). Bright brown, shaded with dark brown and frosted with white, resembling a crumpled dead leaf. Veins rather darker; transverse anterior and transverse posterior lines dark, irregular, and quite incomplete or waved on veins, followed by the whitish shades. Summer form ferruginea Packard much redder; the ground nearly even, with some white scaling, but no contrasting pale shades. 330, 945 mm. (H 41:19 520 9.)

Not common. Caterpillar mottled, dark gray, on apple, oak, maple, and other trees; resting on the bark by day. Two broods; moth in May and early June; July and August.

Generally distributed; a slightly variant race in the west. New York: Old Forge, Rochester, Lancaster, Buffalo, Ithaca, Oneonta, Rhinebeck, Poughkeepsie, Long Island. I have seen variety *ferruginea* from Ithaca and Karner.

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5. HETEROPACHA Harvey

Similar to Epicnaptera, but with the margins nearly even (fig. 428), the anal angle of fore wing not notched, and the costa of the hind wing evenly rounded; fore wing with R_2 lost, R_4 strongly stalked, hind wing with three humeral veins; two arising from the very large humeral cell. 1. H. rileyana Harvey. Dark ash gray, powdery and mottled, the median area rather darker, bounded by wavy and often obsolete wavy pale and dark transverse anterior and transverse posterior lines; subterminal of dark dots; fringe checkered. 30 mm. (H 8:7.) Caterpillar much like that of *E. americana*, but without any wart on eighth segment of abdomen.

segment of abdomen. Western Pennsylvania to Kansas and south; Florida.

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SUPERFAMILY DREPANOIDEA

Ocelli and maxillary palpi absent; tongue not scaled, variable in strength, absent in Oreta; antennæ normally deeply prismatic, rarely slender or pectinate, sometimes with very little scaling on shaft; palpi small or moderate. Body relatively small and stout, stumpy looking, appearing larger in the Thyatiridæ on account of the deep vestiture, abdomen broadly attached to thorax, legs rather weak. Tergopleural groove of basal segment of abdomen much enlarged, opening into a large cavity covered by a lateral chitinous area representing the tympanic structures; our species also with a chitinized subventral bulla at the base of the abdomen, divided in two parts; thorax not modified. Fore wings typically with a long slender accessory cell, lost in a few exotic forms; sometimes with \mathbf{R}_{a} and \mathbf{R}_{a} , \mathbf{R}_{4} and \mathbf{R}_{5} separately stalked together, \mathbf{R}_1 free, and all radials present. \mathbf{M}_2 variable in position; in our species always at least twice as near \mathbf{M}_3 as \mathbf{M}_1 in the hind wing and often in the fore wing; 1st A wholly lost, 2d A and 3d A normal. Hind wing with \mathbf{R}_1 obsolete at extreme base of wing, Sc and **R** closely parallel to well beyond end of cell, or anastomosing beyond end of cell; humeral angle sometimes expanded, and frenulum occasionally lost; 1st A lost, 2d A normal, 3d A often very weak. Base of Sc much swollen in Oreta.

Egg of flat type; larva with hooks of prolegs biordinal, except in Eudeilinea, normally with a few outer hooks also. Front small; head more or less bilobed, and often decidedly wider than high, with primary set only. Body with a few subprimaries, sometimes with only vi duplicated; in other forms with a few subdorsal warts; vii composed of two or more setter on the meso- and metathorax (unlike the simple-haired members of the Noctuidæ and related families). Ventral prolegs equal, anals somewhat reduced, and raised at rest (Thyatiridæ, Brahmæidæ), or absent (Drepanidæ). Pupa not well known, thin-skinned, in Oreta rosea cylindrical, tapering very abruptly at posterior end; no sutures on head; prothorax of macro type, broad and rounded out; labial palpi showing as a minute area at base of tongue, which is about a third the length of the wing (doubtless longer in other genera); femora covered; fore and middle tarsi meeting on middle line (probably a merely generic character); antennæ shorter than middle legs, filiform; and neither middle legs nor antennæ reaching tip of wings. Tips of hind tarsi exposed; some rudiments of abdominal spining; cremaster rudimentary, without setæ. Setæ indistinct.

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The superfamily on the whole is an isolated member of the general Bombycid-Saturniid series, but not especially close to any particular group; the likeness to Geometridæ, Notodontidæ, and Noetuidæ being mostly superficial, or due to the persistence of primitive characters. They may possibly be nearest the Geometridæ. The Oriental genus Euchera connects the two families. Besides the two North American families, the Brahmæidæ also appear to belong to the same stock.

Family 42. THYATIRIDÆ

(Cymatophoridæ)

North American species with deeply laminate antennæ, tongue developed but not very strong, and moderate palpi; deep hairy or mixed vestiture, with a truncate tuft, or rather, mass of hair-scales, on disc of thorax, and hairy legs. Abdomen with small dorsal tufts. Fore wing with \mathbf{R}_1 free, \mathbf{R}_2 from accessory cell near tip, \mathbf{R}_3 and \mathbf{R}_4 , or \mathbf{R}_4 and \mathbf{R}_{a} shortly stalked from tip of accessory cell; in the latter case, with the cell closed by a crossvein, r. Accessory cell extending nearly halfway to apex; \mathbf{M}_1 normally from it, \mathbf{M}_2 from middle of end of cell; both middle and lower discocellular veins weak and bent; 3d A distinct. Hind wing with middle discocellular at least twice as long as lower discocellular, and bent; 3d A well developed; frenulum strong, and, in the male, knobbed at tip.

Caterpillar (fig. 434) with head decidedly wider than high, labrum deeply notched, vi on abdomen represented by two well-separated setæ; vii of 3 setæ on leg-bearing segments, and 2 or 3 on the other segments of abdomen. Seta iv higher than spiracle, sometimes almost directly over spiracle and close to iii, v normally associated with one or two subprimaries. Prolegs sometimes with rudimentary outer hooks. The caterpillars live in a loosely rolled leaf. Pupa not studied.

A small family, formerly usually placed with the Noctuidæ or Notodontidæ, which the species resemble superficially. In the Old World, forms occur with **M**, either low or high in both wings, but otherwise typical, even in pattern.

Key to the genera

1. Fore wing with a tuft a third way out on fold; R_3 and R_4 shortly stalked

free (fig. 432).

2. Fore wing with anal tuft; more than twice as long as wide.

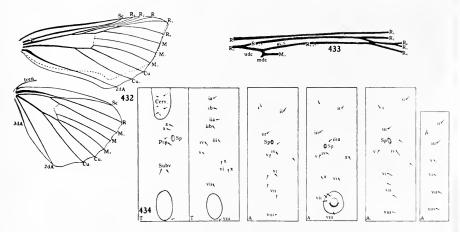
2. Pseudothyatira.

2. Fore wing without anal tuft; hardly twice as long as wide....l. Habrosyne.

1. HABROSYNE Hübner

(*Thyatira*, in part)

Fore wing about twice as long as wide, with a slight anal lobe and tuft; accessory cell relatively short, closed by crossvein \mathbf{r} ; \mathbf{M}_1 arising from accessory cell; wing smooth-scaled; hind wing with \mathbf{M}_2 about twice as near \mathbf{M}_3 as \mathbf{M}_1 , (Figs. 432, 433.)



FIGS. 432-434. THYATIRIDÆ

432, Habrosyne scripta, venation. (The form of the fore wing of 'Pseudothyatira is also shown in outline); 433, Euthyatira pudens, accessory cell of fore wing, showing connections of neighboring veins; 434, Habrosyne derasa (Europe), seta map.

Caterpillar mottled, dead-leaf brown, with dark dorsal line and rather smooth head, the dark dorsum sharply set off from the pale venter.

1. H. scripta Gosse. Light gray-brown. A large smooth brown patch toward base, below Cu, its upper boundary sharp, along Cu; inwardly, with a sharp oblique boundary from Cu to A, then shading into base; outer boundary outwardly oblique from Cu to A, and only a little indented; then inwardly oblique to inner margin; patch outlined with pinkish white, expanding into a large triangular area on cell and costa, but leaving base dark. A group of faint oblique striæ across middle of cell, followed by the oblong dark, pale-outlined orbicular and reniform spots. Transverse posterior line deeply but roundedly waved; of three or four parallel deep-brown lines, not reaching costa; a broad curved pale subterminal shade from apex to anal angle, with a dark shade before it toward costa; middle of costa pale. Hind wing clay color, with shaded postmedial and outer bands. 30-35 mm. (H 40:22.)

June. Caterpillar with dull, slightly rugose head; on blackberry, thimbleberry, and other Rosaceæ.

New Jersey to Alberta; north to Gaspé and Alaska. New York: Plattsburg, Peru, Essex County, Lake Pleasant, Evans Center, Rock City (Cattaraugus County), Ithaca, Big Indian Valley, Onteora Mt., Schenectady, Poughkeepsie, Brooklyn.

2. H. rectangula Ottolengui. Outer boundary of sub-basal patch starting perpendicularly down from Cu, and then right-angled; inner boundary also more perpendicular. Ground darker brownish gray; pale patches above antemedial patch, and at middle of costa, more decidedly pinkish.

Caterpillar similar to that of H. scripta, with shining head.

Maine; New York; Pennsylvania; Illinois; Rocky Mountains; a paler race in Arizona. New York: Brooklyn (type).

2. PSEUDOTHYATIRA Grote

Accessory cell closed by a short crossvein \mathbf{r} , running from \mathbf{R}_3 to stem of \mathbf{R}_{4+5} , as in Habrosyne. Similar to Habrosyne, but with the fore wing relatively longer, with a stronger tooth and tuft at anal angle, and costa more arched. Caterpillar similar, but with the brown dorsum shading gradually into the paler venter.

1. P. cymatophoroides Guenée. Ash gray, somewhat mottled and shaded with pink in antemedial region and over end of cell; base typically blackish; transverse anterior line of a group of three waved blackish lines; transverse posterior of a similar series of waved lines. more sinuous, and blackish only at costa; with a blackish patch beyond it on inner margin, preceded by a white line; a dark patch on costa before apex, crossed by a white lunule; subterminal line of small, wedgeshaped marks between the veins. Orbicular circular; reniform oblong, both small; with raised central dark dots and gray rings; fringe pinkish. Hind wing pale brownish gray. 42 mm. (H 40:25.)

Variety expultrix Grote is similar, but lacks the blackish shades, the antemedial group of lines being light gray, and the hind wing rather darker gray. (H 40:26.) June to August. Caterpillar often with one or a series of lateral white spots,

especially in the type form; on oak and thimbleberry.

Generally distributed; the variety rather commoner than the type. New York: Wilmington (larva), Newcomb. Plattsburg, Mt. Marcy, Fentons (Lewis County), Clayton, Lancaster, Otto, Ithaca, DeBruce, Sharon, Schenectady, Poughkeepsie, Staten Island, Newtown, Long Island. Form *expultrix*: Plattsburg, Fentons (Lewis County), Mt. Marcy, Newcomb, Clayton, Buffalo, Lancaster, Otto, Rock City, Ithaca, Sharon, Schenectady, Long Island.

3. EUTHYATIRA Smith

(*Thyatira*, in part)

Fore wing rather long and narrow, with arched costa, without a scale tuft at anal angle; hind wing longer on costa; accessory cell formed by the anastomosis of \mathbf{R}_3 and \mathbf{R}_4 (fig. 433); a well-marked tuft on fold before transverse anterior line, but no tufts on orbicular and reniform spots.

The caterpillar lives concealed in a loosely folded leaf; it is translucent, with four black spots on the head.

1. E. pudens Guenée. Gray, with large pink spots, the basal one large and oblong, with the tuft in its outer end; a large one on middle and one at apex of costa, and a smaller and browner one at anal angle. Orbicular horizontally elongate, small, touching reniform, both pink-filled; the other markings obscure. 45 mm. (H 40:23.) Variety pennsylvanica Smith (H 40:24) from western Pennsylvania, is wholly

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ash-gray, with inconspicuous markings; in variety anticostiensis Grote, from Anticosti and the neighboring mainland, the pink spots are reduced, and the ground strongly hoary.

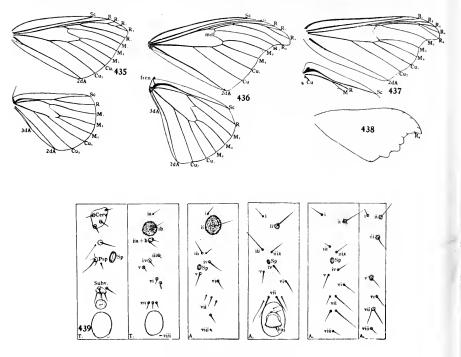
Caterpillar on cornel. Moth in May.

Newfoundland to West Virginia and British Columbia. New York: Brockport, Lancaster, Ithaca, Dutchess County. I have not seen the variety *pennsylvanica* from the State.

Family 43. DREPANIDÆ

(Drepanulidæ; Platypterygidæ; with Auzatidæ)

Slender broad-winged moths, similar to Geometridæ, but distinguished from them by the stumpy body. Male antennæ often pectinate; palpi



FIGS. 435-439. DREPANIDÆ

435, Eudeilinca herminiata, venation; 436, Drepana arcuata, venation; 437, Oreta rosea, venation, showing only costa of hind wing; 438, Falcaria bilincata, outline of fore wing; 439, Drepana falcataria (Europe), seta map of larva

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minute in our species; tongue weak or absent. Fore wings normally with falcate apex; with accessory cell very long and slender, closed by the anastomosis of \mathbf{R}_3 and \mathbf{R}_4 ; \mathbf{M}_2 well separated from \mathbf{M}_3 , but distinctly arising from Cu-stem in both wings; M₃ and Cu₁, separate; Cu₂ arising well out on cell. Hind wing with humeral angle enlarged, frequently supported by a short but distinct humeral vein; frenulum more or less reduced, or lost; humeral vein distinct in fore wing.

Caterpillar (fig. 439) with more secondary hair than in the Thyatiridæ, ventrally, but none dorsally except on the few enlarged tubercles; normally with hair-like skin granulation. Ventral prolegs normal, with rudimentary outer hooks; anals completely lost; a tubercle or spine on anal plate.

There are two broods a year, often differing in color.

Key to the genera

1. Fore wings blunt; hind wing with Sc and R anastomosing (fig. 435).

1. Fore wings falcate; Sc and R of hind wings separate.	1. Eudemnea.
2. Outer margin dentate (fig. 438)	3. Falcaria.
2. Outer margin even.	4 0
3. Frenulum and tongue lost	

1. EUDEILINEA Packard

(Corycia, in part)

Antennæ simple, laminate. Fore wing blunt (fig. 435), oblong with arched costa, translucent. No accessory cell. Hind wing with Sc and R anastomosing, female frenulum distinct, of many bristles. Larva with a mere tubercle on supra-anal plate; with uniordinal hooks.

plate; with uniordinal hooks.
I. E. herminiata Guenée. White; some blackish on legs; normally with ante-and postmedial series of grayish points. Two discal points, obliquely placed, on each wing, below. 28 mm. (biseriata Packard). May and June; July and August. Caterpillar on cornel. Generally distributed. New York: North Elba, Fourth Lake, Fulton Chain, Newcomb, Batavia, Oak Orchard Swamp, Otto, Potter Swamp and Crosby (Yates County), Taughannock Falls, Ithaca, Big Indian Valley, Nassau, Albany, Rhine-beck, New Windsor, Staten Island.

2. DREPANA Schranck

(*Platypteryx* Laspeyres)

Fore wing falcate (fig. 436) below \mathbf{R}_{4} , with evenly sinuous outer margin; \mathbf{R}_{2} long-stalked with \mathbf{R}_{3+4} ; \mathbf{M}_{1} , arising separately from cell, hind wing with Sc and R closely approximate beyond end of cell; humeral angle moderately enlarged, with a weak but distinct frenulum; 3d A rudimentary. Antennæ pectinate, narrowly in female; palpi minute, tongue well-developed.

Caterpillar (fig. 439) with a caudal process about as long as head, with subequal hairy warts on meso- and metathorax and second segment of abdomen, and minute ones on eighth segment.

1. D. arcuata Walker. Typically cream-white, the area before the postmedial line with fine brown wave lines; postmedial in a smooth curve from just beyond middle of costa to apex, then in an even curve to outer third of inner margin, with a blackish shade below it at apex; discal dot fine, black. Hind wing similar toward inner margin, pale on costal half. Summer form genicula Grote (H 41:22), straw yellow, and less strongly striate, the upper leg of the postmedial line often lost. 30 mm. (H. 41:23.)

The forms are not entirely seasonal and intergrade.

May and June; August. Caterpillar on birch and alder.

Pennsylvania to Indiana and north. New York: Plattsburg, Peru, Old Forge, Mt. Marcy, Lewis County, Lancaster, Ithaca, McLean, Trenton Falls, Rhinebeck, Big Indian Valley, Poughkeepsie, New York City, Staten Island; general on Long Island.

3. FALCARIA Haworth

(Edaptera Packard; Drepana, in part)

Similar to Drepana, but with the outer margin regularly scalloped (fig. 438), the deepest scallops on Cu_1 and Cu_2 (not as usual between veins); and hind wing slightly waved. Caterpillar with strongly enlarged warts on meso- and meta-thorax and on eighth segment of abdomen; caudal process about as long as the height of the head.

1. F. bilineata Packard. Typically cream-white, heavily striate with brown (in the late-summer variety levis Hudson, bright yellow and hardly striate); with two parallel and nearly straight oblique brown lines, the outer cutting the lower angle of the cell; discal dot minute. 28 mm. (H 41:7.)

Caterpillar on birch.

New Jersey and north. New York: Plattsburg, Wilmington, Albany, Brooklyn, Staten Island. Variety *levis* Hudson is known definitely from Plattsburg, Wilmington, Alpine, Saratoga, Westchester County, and Brooklyn.

4. ORETA Walker

(Drepana, in part; Dryopteris)

Eyes relatively large; palpi rudimentary; tongue invisible; antennæ deeply laminate, not pectinate. Fore wing falcate (fig. 437), extreme apex rather blunt, on \mathbf{R}_i ; outer margin more abruptly notched below it. \mathbf{R}_2 arising from the accessory cell, which is extremely long; hind wing without frenulum, with a rudimentary humeral vein arising from the much-thickened base of Sc.

Caterpillar with a prominent unpaired hump on metathorax; caudal process twice as long as head. Pupa as described in the superfamily, showing the short tongue and large eyes. The two nominal species are doubtfully distinct. 1. O. rosea Walker. Pinkish or purplish brown, striate with darker brown, out to postmedial line; subterminal region bright yellow; outer margin dark

1. O. rosea Walker. Pinkish or purplish brown, striate with darker brown, out to postmedial line; subterminal region bright yellow; outer margin dark brown; hind wing with outer third yellow and only apex dark brown; antemedial line obscure, darker, distinct, and parallel to postmedial on lower part of wing; postmedial from three-fourths way out on costa, acute-angled on costa, and then oblique in, to two-thirds way out on inner margin. Subterminal line sometimes indicated by blackish dots. Under side similar. Variety marginata Walker is

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yellow, except for the dark brown border, and intergrades with the type form. 25-35 mm. (**H 41**:24.)

May to September; two broods. Caterpillar on Viburnum. New Jersey to Ohio and north. New York: Peru, Fentons (Lewis County), Mt. Marcy, Ithaca, Big Indian Valley, Albany, Rhinebeck, Poughkeepsie, Staten Island, Brooklyn. I have seen variety marginata Walker from "New York City and vicinity" (Eliot), and a transitional specimen from Ulster County.

2. O. irrorata Packard. Ground wholly strigose dull brown or brownish crim-son, without any yellow; markings as before, the postmedial sometimes double or almost obsolete. 25-35 mm. (H 41:6.)

With the preceding form, but apparently much rarer and local.

Massachusetts and Pennsylvania to Manitoba. New York: Mt. Marcy, Ithaca.

Memoir 66, Simplified Apparatus and Technique for the Electrometric Determination of Hydrogen Ion Concentration in Milk and Other Biological Liquids, the third preceding humber in this series of publications, was mailed on June 26, 1923.

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FOOD INDEX

(This index is arranged 'systematically, following the Robinson and Fernald edition of *Gray's Manual*. In looking up a caterpillar, it is best to look first for the general feeders and the forms already reported from the plant on which it was found, and then for other members of the same family, as caterpillars rarely confine themselves strictly to a single food plant.)

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(These three types of habit intergrade so perfectly that it is not convenient to list them separately. The majority of species listed here would naturally belong to at least two of the three categories.)

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Lycopodium: lycopodiana 476.

Lichens

Eudarcia 77, walshella 144, rileyi ? 144, Symmoca 261, adelalis 538.

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Ferns (Filicales)

General: gracilis ? 69, theseusalis 561. Osmunda: osmundana 457, æglealis ? 567.

Conifers (Pinaceæ)

General: variella ? 300, shawiana ? 428, diniana 446, dissitana ? 497, imperialis 665. Pine (Pinus): fuliginosella 124, borkhausenii 248, vittella 288, pinifoliella 290, cond-ferella 300, confluentella 311, haimbachi 348, pini 368, toreuta 396, erotella 396, moni-torana 427, buoliana 438, frustrana 438, rigidana 438, comstockiana 439, virginiana 439, albicapitana, 439, gemistrigulana 439, schulziana ? 452, pinatubana 490, robustella 608, zimmermanni 620, sepulcralis 665. Larch or tamarack (Larix): laricella 209, laricella 347, columbia 674, laricis 681. Spruce and fir (Picca and Abies): piceaella 300, youngana 393, nanana 398, tocul-lioaene 429, pickolame 420, ratabuscione 416, ablotama 432, variane 486, fumiforaul-

Sprice and nr (Price and Ables): piceacha 300, youngana 303, hanana 303, hoch-lionana 428, pickolana 439, ratzeburgiana 446, abietana 433, variana 486, fumiferana 489, afflictana 492, packardiana 492, reniculella 620, abietella 621. Henlock (Tsuga): tsugensis 255, aplicitripunctella 299. Cypress (Taxodium): variella 300, freyella 346, thuiella 348.

Red cedar and juniper (Juniperus): marginella 286, juniperella 300, gibsonella 301. freyella 346, annettella 346, alternatella 347, rutilana 506.

Typhaceæ

Cat-tail (Typha): phragmitella 324, julianalis 539.

Najadaceæ

Pondweed (Potamogeton): seminealis ? 578, obscuralis 578,

Hydrocharitaceæ

Eelgrass (Vallisneria): obscuralis 578.

Grasses (Gramineæ)

General: vagaus 215, graminivorella 349, maiorina 470, Crambus spp. 575-604. (Many of the species listed below will doubtless also turn out to have a variety of food-plants (Many of in the fami y.)

Beard grass (Andropogou) : andropogonis 307. Witch grass etc. (Panicum) : clandestinella 333, miraculosa 359, panicifoliella 359.

Rice (Oryza) : plejađellus 591. Brachvelytrum : brachyelytrifoliella 221.

Timothy (Phleun) : mistrella 253. Meadow grass, blue grass (Poa) : albicapitella 222, sylvestris 222. Glyceria : irrorata 223.

Wild rye (Elymus): leucofrons 222, præmaturella 223. Bottle-brush grass (Hystrix): orestella 220, prælineata 222, leucofrons 222, hystricella 283, graminivorella 349. (°orn (Zea): arcanellus 121, nubilalis 568, zeacolella 591.

Sedges (Cyperaceae)

General: verutana 470, ekthlipsis 576. Bulrush (Scirpus): enitescens 221, madarella 221, robusta 297, furfurana 470, majorina 470, forbesellus 591.

Sedge (Carex): madarella 221, argentosa 221, eucullata 222,

Elophila spp. 579.

Duckweed (Lemnaceæ)

Rushes (Juncaceæ)

Juncus: unicolorella 209, quadrilineella 213, cratipennella 214, cœspititiella 214, fagi-costicella 215, verutana 470, furfurana 470. Luzula: biforis 214.

Liliaceæ

Lily (Lilium): incertella 344. Yucca: quinquepunctella 74, yuccasella 75.

Solomon's-seal (Ploygonatum): melaleucana, 498.

Trillium: meleleucana, 498.

Cat-briar (Smilax): smilaciella 151, smilacisella 183, incertella 344, inimicella 447.

Orchidaceæ

Lady's-slipper (Cypripedium): cypripediana 473.

Salicaceæ

(Most of the species will eventually prove to occur on both willow and poplar.)

(Most of the species will eventually prove to occur on both willow and poplar.) General: albela 151, purpuriella 175, stigmatella 175, salcifoliella 192, tricinetus 366, albicornis 373, apiformis 374, tibialis 374, nisella 401, effractana 487. Wil ow (Salix): thule 68, fuscotibiella 92, pallida 95, saliciella 152, saliciella 28, in-quilinella 267, niveopulvella 278, fungivorela 296, salicifungiella 296, salicipomonella 330, bolteri 370, sigmoidea 373, gallæsaliciana 395, rectipilcana 402, diminutana 405, salicicolana 445, heindeliana 483, schalleriana 485, permutana ? 486, participialis 550, inquilinella 623, pravella ? 624, carneella 627, fusca 628, latifascia 669. Poplar (Populus): populella 83, populetorum 95, populiella 154, populiella 192, fletch-crella 236, dyariella 268, innocuella 278, præangusta 331, dollii 366, tineana 405, duplex 456, centerensis 518, populi 519, helvalis 561, velleda 680.

Myricaceæ

Bayberry (Myrica): obscurella 89. flavella 173, picturatella 198, walsinghamella 239, indentanus 436, karacana 477, participialis ? 531.

Sweet fern (Comptonia): electrofusca 464, comptoniella 618.

Juglandaceæ

General: juglandifoliella 90, caryæfoliel a 197, caryana 392, juglandana 491, palliolella

Walnut (Juglans): juglandiella 172, juglandivorella 176, nigralineella ? 216, juglandiella 228, testulana 478, demotella 616.
Hickory and pecan (Carya): trinotata 87, y-inversa 110, clemensella 159, olivæformls 191, caryæalbella 191, caryæfoliella 211, lucifluella 228, ella 228, caryæfoliella 285, aphidiella 311, bolliana 443, improbana 443, infumatana 496, fumoferalis 569, angusella 616, stigmella 616, caryivorella 616, caryæ 617, kearfottella 617, latifasciella 617, ulmi-arrosorella 632, luna 671.

Betulaceæ

General: corylifo iella 89, nisella 401.

General: corylifo'iella 89, nisella 401. Hazel (Corylus): quadrinotata 83, corylisella 199, corylifoliella 212, grotella 242, tortriciformella 254, tristrigella 279, trinotella 285, similana 400, walkerana 401, corylana 465, exoieta ? 468, thestialis 567, rubrifasciella 618, coryliella 618. Ironwood, hop hornbeam (Ostrya): ostryæfoliella 89, ostryæella 175, obscuricostella 191, ostryæfoliella 191, tritænianella 194, hamadryadella 196, lentella 197, ostryærella 199, ostryæ 211, ostryæfoliella 228, ostryæella 329. Ironwood, blue beech (Carpinus): quadrinotata 89, y-inversa 110, ostryæella 175, carpinella 211 Psilocorsis sp. 236

- Ironwood, blue beech (Carpinus): quadrinotata 89, y-inversa 110, ostryæella 175, carpinella 211, Psilocorsis sp. 236.
 Birch (Retula): ostryæfoliella 89, mora 121. coronatella 158, canadensisella 158, conspicuella 167, vicinella 168, oblitere.la 168, coroniella 174, martiella 103, lentella 197, betulivora 198, cincrella 210, lentella 212, betulella 242, betulel a 293, gœdartella 345, diana 353, culciformis 370, similana 400, transmissana ? 402, sollcitana 402, albeolana 456, zelleriana 465, trisignana 483, niveana 484, ferrugana 487, affractana 487, betulella 618, luna 671, polyphemus 672, arcuata 600, bilineata 680.
 Alder, (Alnus): argenteomaculatus 68, elongella 172, glutinella 176, auronitens 192, cincrella 210, alniel'a 212, belangerella 293, alnifructella 302, gœdartella 345, calliphanes 345, asilipennis 365, americana 370, brunneopurpurata 465, extricalis 561, rubrifasciella 618, arcuata 690.

Fagaceæ

General: packardella 158, strigifinitella 179, asperatella 607. Beech (Fagus): faginella 236, fagella 304, fagigemmeana 468, luna 671. Oak and chestnut: auricyanea 66, castaneæfoliella 74, saginella 95, latifasciella 94, castaneæella 198, quercicella 235, obsoletella 236.

Chestnut (Castanea): castaneæ 83, phleophaga 83, castaneæella 147, kearfottella 191, leucochrysella 216, castaneæ 239, castaneella 347, pictipes 371, fuscociliana 410, spoliana 443

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(ak (Quercus): heinrichi 83, terminella 88, altella 89, quercipulchella 90, similella 93, flavipedella 94, anguinella 97, platea 97, nasoni 109, biguttata 110, pallida 111, fusco-marginella 147, tinctoriella 148, badiiella 148, citrinipennella 148, albostraminea 148, zelleriella 147, tinctoriella 148, badiiella 148, citrinipennella 148, albostraminea 148, zelleriella 148, litigiosella 157, ainsliella 159, alchimiella 176, quercinigrella 177, strigosa 179, albinotella 180, fugidel:a 182, fitchella 180, bataviella 189 (footnote), querciabella 190, argentifimbriella 197, uterciola 148, ibiotella 196, albanotella 190, hageni 190, arciterella 197, conglomeratella 197, quercivorella 197, atanoidella 196, incinatiella 196, macrocarpella 197, conglomeratella 198, obstrictella 198, tubifereila 199, atromarginata 217, querciella 253, vanto-basis 253, melanela 254, bicostomaculella 267, maculimarginella 268, vernella 254, georgiella 282, gristifasciella 291, latifasciella 292, quercinigracella 292, fuscopunctella 292, guanduella 310, purpuriella 223, summerdamia sp. 339, oreasella ? 346, apicimaculella ? 346, simulans 365, palmii 366, rubristigma 371, latiferrennus 397, timidella 99, divisana 407, burgessiana 410, murtfelditana 410, appendicea 464, diluticostana 477, karacana 477, testulana 478, semipurpurana 481, quercifollana 491, fervidana 496, grisea 498, macmurtrei 519, hebescella 617, hammondi 632, ostrinella 636, cuiridota 657, melsheimeri 658, stigma 667, virginiensis 668 senatoria 668, maia 669, cymatophoroides 687.

Urticaceæ

General: fabriciana 352. Elm (Ulmus): apicialbella 90, ulmella 92, Marmara sp. 182, argentinotella occitanica 192. ulmella 197, limosipennella 212. costrictella 307, undulatella fuscociliana 410, asperatella 607, ulmiarrosorella 632. Hackberry (Celtis): ovina 102, celtisella 193, celtifoliella 194, rubrisparsella celtidella 626. 192. 347

623.

Osage orange (Maclura) : externalis 555.

Nettle (Urtica): urticata 581.

Richweed (Pilea) : pulchrimella 332.

Santalaceæ

Bastard toadflax (Comandra): comandrana 484.

Polygonaceæ

Sorrel, dock (Rumex) : discoocellella 266, fasciatana 463. Smartweed, knotweed (Polygonum): borea 214, shaleriella 214, discoocellella 266, absconditella 296, minime.la 296, ainslei 568.

Chenopodiaceæ

Pigweed (Chenopodium): lingulacella 298, hermannella 298, ranalis 546. Beet and chard (Beta): perspectalis 544, fascialis 545, bipunctalis 554, sticticalis 556.

Amaranthaceæ

Pigweed, Amaranth (Amaranthus): amaranthella 215. Alternanthera: perspectalis 544, fascialis 545.

Phytolaccaccæ

Poke (Phytolacca): obscuralis 548, æglealis 567,

Nyctaginaceæ

Oxybaphus: nyctaginella 358.

Caryophyllaceæ

Chickweed (Stellaria) : cœnosipennella 214. Campion (Silene) : apicialbella 210.

Water lilies (Nymphæaceæ) penitalis 568, obliteralis 577, gyralis 577, maculalis 577.

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Ranunculaceæ

Meadow rue (Thalictrum) : fuscipedella ? 245. Clematis : caudata 374, maculata ? 522 Marsh marigold (Caltha): Opostega ? 160.

Magnoliaceæ

General: liriodendrana 473, angulifera 673. Tulip tree (Liriodendron): liriodendrella 154, ostricolorella 631, promethea 673. Magnolia: magnoliella 154, hamadryadelia 196, magnoliana 498, carolina 673.

Anonaceæ .

Papaw (Asimina) : plummeriana 388, cariosa 588.

Berberidaceæ

Barberry (Berberis) : dentosa 588.

Lauraceæ

Red bay (Persea) : parvula 340. Sassafras: sassafrasella 172, niveiguttana, 459, saracana 476, fissalis 567.

Mustard family (Cruciferæ)

General: maculipennis 341, porrectel.a 341, undalis 550, straminalls 551, rimosalis 551.

Sarraceniaceæ

Pitcher plant (Sarracenia): dæckeana 458.

Saxifragacese

Saxifrage (Saxifraga): saxifragæ 371. Alum root (Heuchera) heucherana 403.

Hydrangea: hydrangiæella 226, ferruginana ? 468, ferriferana 469. Currant, gooseberry (Ribes): taylorella ? 75, albogalleriella 161, tipuliformis 372, exoleta ? 468, fernaldana 515, grossulariæ 630.

Hamamelidaceæ

Witch-hazel (Hamamelis): superbifrontella 176, hamameliella 199, argutanus 447, nivelguttana 459, merrickana 466, footiana 470. Sweet gum (Liquidambar): liquidambarisella 154, dorsivittella 301, afflictella 625, regalis 665.

Platanaceæ

Sycamore (Platanus): platanella 92, clemensella 93, albisparsella 267, divisana 407, platanana 409, ovulalis ? 547, militella 607.

Rosacea

(In this family especially, the species tend to feed on members of several related genera. The Ericacee seem to be closely related to this family biologi-cally, and there is a tendency for the forms feeding on either to stray to the other.)

Geueral: cratægella 191, fletcherella 209, Tinagma 224, splendoriferella 227, pade'la 339, falciferella ? 341, exitiosa 370, pictipes 371, pyri 372, prunivora 392, pomonella 396, tineana 405, permundana 466, permutana 486, indiginella 619, americana 681, scripta 686, rectangula 687. Ninebark (Physocarpus or Opulaster): opulifoliella 90.

Meadow sweet, Steeple-bush, Hardhack (Spiraea): spiræifoliana 410, hemidesma 458, albiciliana 458, aruncana 472, spiræifoliana 472, lucina 670. Apple, pear, chokeherry, mountain ash, (Pyrus): pomivorella 93, chalybeia 93, mall-foliella 147, pomifoliella 158, guttea 166, geminatella 167, quadripunctella 168, sorbi-

vorella 168, arbutifoliella 168, fraxinella 177, elotella 182, pomonella 183, malimali-foliella, 191, cerasivorella 208, maiivorella 217, ligulella 284, quaintancella 310, mali-gemmella 312, conjugella 347, parlana 353, pomonella 366, ma ana 467, minuta 482, pheniciallis 570, hammondi 632, velleda 680, americana 681. Quince (Cydonia): geminatella 167, malimalifoliella 191. S'adbush (Amelanchier): amelanchierella 97, Psilocorsis sp. 236. Thorn (Crataegus): scintillians 86, cratægella 301, curvi ineella 329, dietziana 411, chionosema 456, niveiguttana 484. Strawberry (Fragaria): anglicella 167, combtana 408 (floridana 407?). fragariana 488. Raspberry, blackberry (Rubus): villosella 88, rubifoliella 91, aënea 147, cretaticostella 213, albaciliella 254, disconotella 277, festalella 360, marginata 374, apicana 406, fra-gariæ 407, separatana 455, concinnana 405, tenuidactylus 645, cymatophoroides 687. 'grimony (Agrimonia): agrimoniella 279. Rose (Rosa): roszefoliella 91, roszeticola 147, rosacella 210, roszefoliella 217, pyricolana 95, Ancylis sp. 406, suffusana 416, nimbatana 455, separatana 455, cyanana 458, berg-manniana 480, albicomana 480, semlpurpurana 481, rhododactyla 644. Cherry, plum, peach (Prunus): bifasciella 57. slingerlandella 91, geminata 112, quadri-punctella 168, serotinella 178, serotinella 183, propinquella 91, cerasivorella 208, occi-dentis 209, pruniella 209, umbratica 211, lapidicornism 215, at anica 212, confusella 265, serotinella 269, lineatella 280, prunifoliella 304, exitiosa 270, molesta 394, bur-gessiana pruni 410, campestrana 452, malana 467, ferruginana ?468, inornatana 469, pallorana 492, cerasivorna 496, semlfuneralis 631, promethea 673, americana 681.

Leguminosæ

General: desmodiella 193, lecontella 241, Polyhymno 290, nigricana 393, pollinalis 558, melanogrammos 607, contatella 626, boisduvaliella 629, zinckenella 629, pumilio ? 646,

hicelor 666. Honey locust (Gleditsia): biscolorella ? 306, pallidochrella 307, gleditschiæella 328, leguminana 512, reductella 624, bisecta 666. Judas tree (Cercis): cercerisella 264, chambersana 478, funeralis 545. Wild indigo (Baptisia): tristrigana ? 394, baptisiella 608.

Lupine (Lupinus): lupinella 279. Clover (Trifolium): palpilineella 290, roscosuffusella 295, interstinctana 292, angulifasciana 409, instrutana 453, discopunctanum 479.

Amorpha : uhlerella 192, amorphella 325. Tephrosia : tephrosiella 619.

Locust (Robinla): robiniella 184, robiniella 192, gemmea 193, ostensackenella 193, robiniella 241, unctulella 267, pseudoacaciella 269, robiniella 301, insiticiana 397. Tick trefoil (Desmodium, Meibomia): violacella 173, pergandeella 249, pavonacella 353,

fana 393. \ Bush clover (Lespedeza): lespedezæfoliella 185, tesquella 325.

Beach pea (Lathyrus): maritima 409. Bean: oculatana 523, vitrina 523. Lima bean (Phaseolus): nubilella 621.

Hog peanut (Amphicarpa): amphicarpeæfoliella 181, morrisella 192, tesquella 325, pavonacella 353.

Krameria : crescentifasciella 278.

Geraniaceæ

Geranium: purpurana 495.

Rutaceæ

General on woody members: citrifoliella 285. Ptelea: pteliæella 86, pteleæ 239, nigrinotella 240.

Simarubaceæ

Ailanthus: punctella 340, walkeri 675.

Anacardiaceæ

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Sumach, poison ivy (Rhus): intermedia 87, rhoifoliella 87, rhoifoliella 172, gutti-finitella 198, walsinghami 270, rhoifructella 278, roseosuffusella ? 295, chalcofrontella 311, erythriella 360, argutanus 447, rhoifructana 473, pravella ? 624, semiobscurella 626, stypticellum ? 634, ostrinella 636.

Aquifoliaceæ

Holly, black alder (Ilex): ilecella 159, cryptolechiella 236, ilicifoliana 412, finitimana 412, pertextalis 567.

Celastraceæ

Evonymus: multipunctella 339, thestialis 567. Bitter-sweet (Celastrus): celastrusella 348.

Aceraceæ

Maple, box elder (Acer): acerifoliella 76, saccharella 82, aceriella 171, bimaculatella 174, Gracilaria A 174, packardella 174, negundella 175, trinotella 189, clemensella 190, lucidicostella 190, saccharella 197, aceriella 199, subreticulata 347, corni 371, acerni 371, signatana 399, willingana 440, crescentana 440, aesculana 441, moffatiana 441, tyrius 447, pettitana 477, abicaudana 478, semiferana 497, negundana 498, asperatella 607, rubicunda 667, polyphemus 672, disstria 682.

Sapindaceæ

Horse-chestnut, buckeye (Æsculus): æsculisella 199. emblemella ? 288, claypoleana 440, æsculana 441, instrutana 453, hippocastana 466, rileyana 496.

Balsaminaceæ

Touch-me-not, jewel weed (Impatiens): agilana 458.

Rhamnaceæ

Buckthorn (Rhamnus): rhamnicola 87, pullata 180, quercifoliana 491. New Jersey tea (Ceanothus): anarsiella 267, ceanothiella 328.

Vitaceae

General: falsarius 114, americana 114, texana 114, vitigenella 154. Woodbine (Ampelopsis, Psedera): flavescens 106, ampelopsiella 154, quiquenotella 158, apmelospsiella 226, absconditella 296.

Grape (Vitis): speculella 152, vitifoliella 154, isabella 226, viticordifoliella 226, æsella 227, pollstiformis 366, eclipsana 393, viteana 473, botrana 473, lugubris 523, funeralis 545, periscelidactylus 645.

Tiliaceæ

Basswood, Linden (Tilia): argentifasciella 86, tiliella 91, clemensella 159, lucetiella 193, tiliacella 194, tiliæfolie la 216, tiliana 468, limata 548, thestialis 567.

Malvaceæ

Rose mallow (Hibiscus): Marmara sp. 182, hibiscella 266. Cotton (Gossypium) regalis 665.

Hypericaceæ

St.-John's Wort (Hypericum): hyperella 240, nigra 302, hypericana 487.

Cactaceæ

Prickly pear (Opuntia): opuntiella 183, prodenialis 630.

Thymeleaceæ

Leatherwood (Dirca): dircella 181.

Elæagnaceæ

elæagnisella 217.

Lythraceæ

Swamp loosestrife (Lythrum): lythrella 240.

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Onagraces

Willow-herb (Epilobium) : decorella 324.

Evening primrose (Chothera): trinotella ? 282, circumscriptella 323, murtfedtella 323, stellella 323, brevivittella 323, eloisella 326, argentimaculella 327, œnotherana 511, funeralis 545.

Enchanter's nightshade (Circaa): terminella 327.

Umbelliferæ

General: argillacea ? 238, applana clemensella 239, heracliana 243, pimpinella 343. Hemlock (Conium): canella ? 238.

Water hemlock (Cicuta) : cicutaëlla 343. Carawny (Carum): cinereocostella 242.

Water parsnip (Sium) : cinereocostella 242.

Cornsees

Cornel, dogwood (Cornus): dietziella ? 76, cornusella 173, belfrageella 174, alblan-tennaëlla 211, cornifollella 226, lindana 399, micantana 464, punctana 467, cornana 467, quadrifida 469, rholfrüctana 473, pudens 687, herminiata 689. Tupelo, sour gum (Nyssa): nyssæfollella 91, nyssæfollella 226, nyssæcolella 621.

Ericaceæ

General: unguicella 406, floridana 407, hastiana 485, maccana 486, nebullfera 588. Sweet pepper bush (Clethra): pertextalis 567.

Pyrola: roseomaeulana 456.

Rhododendron: latistrigella 152, rhododendri 372.

Azalea : azaleæ 175. Laurel (Kalmia) : kalmiella 166, rhoifructana 473.

Andromeda (Andromeda, Lyonia) : villella 312.

Andromeda (Andromeda, Dyona): diversella 193.
Borrel tree (Oxydendrum): diversella 193, gaylussaciella 212, magnella 228.
Blueberry, cranberry (Vaccinium): paludicola 89, preciosella 167, arbitrella 167, vacciniella 174, negligens 228, vacciniella 285, quinquecristatella 292, nævana 411, schulziana ? 452, interruptolineana 457, permundana 466, minuta 482, schalleriana 485, mariana 491, vaccinii 619, fusca 628.

Primulaceæ

Loosestrife (Lysimachia): lysimachiæella 194.

Ebenaceæ

Persimmon (Diospyros): diospyriella 228, uroceriformis 367, malachitana 457, regalis 665.

Oleaceæ

General: syringæ 367, angelica 679. Privet (Ligustrum): cuculipennella 177. Ash (Fraxinus): fraxinella 177, asilipennis 365.

Gentianaceæ

Floating heart (Limnanthemum, Nymphoides) : seminealis 578.

Apocynaceæ

Dogbane, Indian hemp (Apocynum) : apocynella 183.

Convolvulaceæ

Morning-glory (Ipomea): somnulentella 153, clemensella 332, gemmiferella 332. Bindweed (Convolvulus): monodactylus 651.

Polemoniacea

Greek valerian (Polemonium) : polemoniella 213.

Hydrophyllaceæ

Phacella: macelhosiella 245, zellerlella 246.

Boraginaceæ

Comfrey (Symphytum): schalleriana 485. Stoneseed (Lithospermum): longimaculella 246. Onosmodium: onosmodiella 179.

Mints (Labiatæ)

General: acanthodactyla 644. Bastard pennyroyal (Trichostema): sexnotella 324. Skullcap (Scutellaria): scutellariæella 275, inflatella 354. Sage (Salvia) : onythesalis ? 570. Horsemint (Monarda): signatalis 571.

Solanaceæ

General: operculella 276. Nightshade, potato, eggplant (Solanum): nundinella 270, striatella 277, glochinella 277, perfusalis 554. Ground cherry (Physalis): lavernella 275, physaliella 297.

Scrophulariaceæ

Beardtongue (Pentstemon): lavana 513. Speedwell (Veronica): pterodactyla 646. Orthocarpus: feliciella 360.

Bignoniaceæ

Trumpet creeper (Tecoma): tecomæ 529.

Plantaginaceæ

Plantain (Plantago): plantaginisella 185.

Rubiaceæ

Buttonbush (Cephalanthus) : cephalanthiella 326.

Caprifoliaceæ

Honeysuckle (Lonicera): fragilella 194, xylostella 342, flavibasana 475, amorata 655. Snowberry (Symphoricarpus): symphoricarpella 193, mariæella 194. Viburnum: viburnella 216, logiana 485, juglandana 491, dryopterata 655, rosea 690. Elder (Sambucus): argenticinctella 247, tertialis 561.

Cucurbitaceæ

Squash, melons, cucumber: satyriniformls 367, nitldalis 549, hyalinata 550.

Compositæ

General: crescentella 159, ignota 186, juncidella 281. Ironweed (Vernonia): vernoniæella 215, ambrosiana 472, subnivana 483, rana 509,

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