







# GEOLOGICAL SERIES

## OF

## FIELD MUSEUM OF NATURAL HISTORY

Volume VI

CHICAGO, OCTOBER 31, 1939

No. 26

# NANODELPHYS, AN OLIGOCENE DIDELPHINE

BY PAUL O. MCGREW Assistant, Paleontology

In a former paper (McGrew, 1937) the genus Nanodelphys was described and tentatively referred to the subfamily Thlaeodontinae. The reference was based on its agreement in molar structure with the subfamily diagnosis given by Simpson (1929). However, small size, Oligocene occurrence, and lack of knowledge of premolar structure tended to make this assignment somewhat dubious.

In the course of sorting and cataloguing a collection of micromammals from the Brulé of northwestern Nebraska, three additional specimens were encountered which are unquestionably referable to Nanodelphys minutus. Teeth previously unknown in this genus,  $M^{\perp}$  and  $M^{\pm}$ , are preserved on the new specimens and offer further material for comparison.

A review of the existing Didelphidae has shown that the dentition of Nanodelphys is similar in many respects to that of certain living forms, especially Marmosa and Dromiciops. The only constant difference between Nanodelphys and all species of Marmosa is the subequal para- and metacones in the former genus. Although fairly constant within any given species of Marmosa, the stylar cusps vary greatly between different species. In some (especially M. *beatrix*) the stylar cusps are almost exactly like those of *Nanodelphys* and the Thlaeodontinae.

Except for their reduced stylar cusps, the molars of Dromiciops are almost identical with those of Nanodelphys in size and shape, and in having subequal para- and metacones. The metastylar spur is variable among the Didelphinae, being slight in some and prominent in others. Thus each character which was thought to be diagnostic of the molars of the thlaeodontines (see Simpson, 1935) may be matched in one or another living didelphine. The only character remaining which is truly diagnostic for the Thlaeodontinae seems to be the bulbous premolars. Although these teeth are unknown No. 455

393

Valm 10 - -

. · - Y=Y

# 394 FIELD MUSEUM OF NATURAL HISTORY—GEOLOGY, VOL. VI

in *Nanodelphys*, the agreement of this form with certain members of the Didelphinae, coupled with its Oligocene age, makes its reference to that subfamily almost certain.

## Nanodelphys minutus McGrew.

Holotype.—F.M. No. 25708, <sup>1</sup> portion of left maxillary with  $M^{2-3}$ .

Referred specimens.—F.M. No. P25709, portion of right maxillary with  $M^{\underline{2}}$ . F.M. No. P25719, portion of left maxillary with  $M^{\underline{2}^{-}\underline{4}}$ . F.M. No. P25720, portion of left maxillary with  $M^{\underline{1}^{-}\underline{2}}$ .

Description.-With the exception of its greater antero-posterior length in relation to transverse diameter. M<sup>1</sup> agrees exactly with M<sup>2</sup> and M<sup>3</sup>. In relation to their antero-posterior diameters, M<sup>2</sup> and M<sup>3</sup> are wide transversely, with large external shelves. From the anteroexternal corner of each, projects a spur, the outer border of which is confluent with the outer border of the tooth. This spur bears the relatively low stylar cusp A.<sup>2</sup> Immediately posterior to A and external to the paracone is the very prominent stylar cusp B. Behind B is a rather deep U-shaped valley (deeper on  $M^3$ ) in which stylar cusp C may be greatly reduced or absent. Stylar cusp D is rather low and unites posteriorly with cusp E to form an anteroposterior crest external to the metacone. The paracone and metacone are nearly or quite equal in size and height and the V between them is shallow. The antero-external crest of the paracone unites with the anterior slope of stylar cusp B, and the postero-external crest of the metacone unites with stylar cusp E. The paraconule and metaconule are absent. The protocone is very high and is situated antero-internally, lying immediately mesial to the paracone. The protocone, paracone and stylar cusp B lie in an almost straight transverse line. The antero-external crest of the protocone extends in front of the paracone to meet the inner base of stylar cusp A; its postero-external crest terminates at the base of the metacone.  $M^{4}$  is very narrow antero-posteriorly and has stylar cusps A and B as well as the paracone and protocone well developed, but the posterior stylar cusps are lost, and the metacone is greatly reduced.

Comparisons.—Nanodelphys is closer in molar structure to Marmosa beatrix than to any other known didelphid, living or fossil. It differs, however, in certain significant characters which appear

<sup>2</sup> Simpson's (1929) designations of the stylar cusps are followed.

<sup>&</sup>lt;sup>1</sup> This specimen previously bore the number Walker Museum No. 1545. In the interest of keeping the collection as a unit, however, the specimen was transferred, together with certain others which I had collected, to Field Museum.

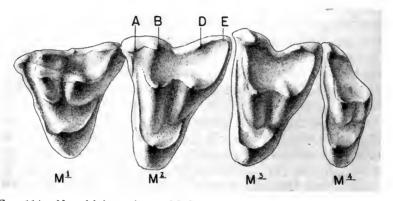
# AN OLIGOCENE DIDELPHINE

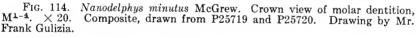
~ ~ `

COL .+

to be primitive. The molars of the Oligocene form are smaller and more compressed antero-posteriorly; the external shelf is broader and more deeply cleft; the stylar cusps are similarly arranged, but the second (B) is larger and higher. The paracone is reduced in M. beatrix and not in Nanodelphys. The V-shaped notch between the paracone and metacone is shallow in Nanodelphys and deep in the living species. The protocone shelf of Nanodelphys is more compressed antero-posteriorly.

Nanodelphys differs greatly from the contemporary Peratherium. It is much smaller and the molars are more compressed anteroposteriorly. Stylar cusp B is much more strongly developed, and





cusp C, which is rather strong in *Peratherium*, is either greatly reduced or absent. The paracone of *Peratherium* is reduced and the V between the paracone and metacone is much deeper. The protocone of *Nanodelphys* is much higher than that of *Peratherium*. In the latter the protocone shelf is V-shaped, whereas it is U-shaped in *Nanodelphys*. The external shelf is more deeply cleft in *Nanodelphys* and the metacone of  $M^{\pm}$  is more reduced.

Nanodelphys differs from Peradectes as it does from Peratherium and Marmosa. Thylacodon, being known only from a lower jaw, can not be directly compared, but it is much larger than Nanodelphys.

*Relationships.*—Without more complete material it would be premature to speculate about the exact position of *Nanodelphys* within the Didelphinae. The molar teeth are sufficiently primitive for the genus to be regarded as structurally ancestral to any or all

395

R

## 396 FIELD MUSEUM OF NATURAL HISTORY-GEOLOGY, VOL. VI

of the living forms. The fact that *Peratherium* is closer in tooth structure to most living murine opossums and to *Didelphis* suggests that *Nanodelphys* did not hold so central a position. By reduction of the stylar cusps *Nanodelphys* might have given rise to *Dromiciops*, but it seems improbable.

## TERTIARY DIDELPHIDS AND PHYLOGENY

The consensus among most writers on didelphid phylogeny has been that *Marmosa* represents the most primitive, structurally ancestral form. Dollo (1899), Bensley (1903), and Gregory (1910) have expressed the belief that all living genera of Didelphidae are structural descendants of that genus. Tate (1933) suggested two principal lines of descent, one giving rise to *Didelphis, Chironectes, Lutreolina* and *Metachirus*, and the other to *Monodelphys, Philander, Marmosa, Glironia*, and *Dromiciops*, the latter two branching from a marmosoid ancestor. The evidence for regarding *Marmosa* as prototypal is found in its unmodified prehensile hands and feet, accompanied by generally arboreal habits—conditions which are presumably primitive.

Several writers—Bensley (1906), Gregory (1910), and Simpson (1928)—have implied that the separation into the several existing genera occurred relatively recently (i.e. post-Oligocene), from *Peratherium*. As there is little or no structural change observable from *Peratherium* to *Marmosa* this view is supported by the paleontologic evidence.

Only one limited group of opossums in North America appears to have escaped extinction at the end of the Cretaceous (Simpson, 1928). This apparently direct, conservative line has been known from two Tertiary genera,<sup>1</sup> Peradectes of the Paleocene, and Peratherium of the Eocene, Oligocene, and Miocene (McGrew, 1937). The dental structure of these forms is very close to certain species of Marmosa; so close, in fact, that satisfactory characters with which to separate them from the recent genus are hard to find. Peradectes is slightly more primitive in that it retains the para- and metaconules and the somewhat less reduced paracone. The stylar cusps of any of the Tertiary species may practically be duplicated in living forms.

The almost complete absence of aberrant species or genera among the fossils seems to have led to the assumption that there

 $<sup>^1</sup>$  Thylacodon pusillus Matthew and Granger has been described from the Puerco. On the basis of the holotype of this species it is not possible to be certain of its relationships.

was little or no diversification among Tertiary didelphids. The conclusion reached from this reasoning has been that the relatively diverse living didelphids represent incipient branches of a new expansion. Bensley (1906) stated this view as follows: "The existing Didelphyidae of South America, which might at first sight be regarded as surviving remnants of the original didelphyid radiation, may be shown to represent a third radiation which is at the present time in its very incipient stages. Of these three radiations the Australian, and the existing South American ones are directly traceable to minute primitive didelphyid forms like the existing genera *Marmosa* and *Peramys*, or *Peratherium*." I am of the opinion, however, that such conclusions may well prove erroneous when additional specimens are known.

The living opossums show rather wide diversity of habit with corresponding foot adaptations. Thus, *Marmosa* is typically arboreal with complete opposable hallux and well-developed plantar pads; *Monodelphys* is terrestrial with reduction of the fifth digit and planter pads; *Chironectes* is aquatic and has highly modified webbed feet. The dentitions of these genera are very similar, however, and no differences in tooth structure may be correlated with the widely different habits. Foot structure, then, is a most important factor in the determination of exact relationships and phylogeny within the Didelphidae—apparently more important than dental structure in such a conservative group. The fact that we do not know the feet of the Tertiary didelphids is, therefore, a serious handicap in the attempt to fit the fossil forms into a phylogenetic picture.

Although the dentition of *Peratherium* is usually regarded as primitive and prototypal (Winge, 1893; Bensley, 1903; Gregory, 1910; Tate, 1933) there is some indirect evidence which indicates that this genus was actually so specialized in foot structure that it could not have been ancestral to most living didelphids. This possibility is suggested by two rather striking facts: (1) *Peratherium* is abundant<sup>1</sup> and is found commonly in the clays of the White River Oligocene. (2) It is nearly always found in direct association with such mammals as rabbits (*Paleolagus*), terrestrial rodents (*Ischyromys, Eumys, Heliscomys*, etc.), small artiodactyls (*Leptomeryx, Hypertragulus, Hypisodus*), horses (*Mesohippus*), and camels (*Poëbrotherium*).

<sup>&</sup>lt;sup>1</sup> In a few Sunday collecting trips I have found more than seventy-five jaws and maxillaries of *Peratherium*. When its small size is considered, with the consequent probability that many specimens were overlooked, such abundance is striking.

## 398 FIELD MUSEUM OF NATURAL HISTORY-GEOLOGY, VOL. VI

Study of the mammals contained in the clays of the White River series led Matthew (1901) to regard them as "strictly terrestrial." He stated: "The analogy of the clay fauna is with that of the modern plains, of the sandstone fauna with that of the modern forests (with some aquatic forms)." The abundance of specimens of *Peratherium* in the White River clays indicates that it formed an important component of the life of the time. The number of specimens of the genus is greater than that of all insectivore genera combined.<sup>1</sup>

It is highly unlikely that an arboreal creature would occur in such abundance with terrestrial forms. The obvious conclusion, therefore, is that *Peratherium* was terrestrial. Further, to maintain existence so successfully<sup>2</sup> among the numerous placentals it must have been considerably modified in foot structure. This would suggest that *Peratherium*, although having the generalized didelphine dentition, could hardly have been ancestral to all of the recent Didelphidae, unless arboreal modifications were secondary, which, according to Dollo (1899), does not seem to be the case. It is possible, of course, that a terrestrial form such as *Monodelphys* could be a direct descendant of *Peratherium*.

The presence of *Nanodelphys* in the Oligocene proves that there was at least some diversity of marsupials in the middle Tertiary of North America. It is very probable that many and diversified arboreal opossums lived throughout the Tertiary, but their habitus so rarely occasioned their presence in areas of deposition that we do not know them as fossils. It may be that the Tertiary didelphids would not be regarded as "stereotyped" or "monotonously unvaried" (Simpson, 1928) if they were adequately known.

It seems likely that many of the recent genera of didelphines originated in the early Tertiary. This view is supported by the occurrence of *Lutreolina* and *Didelphis* in the lower Pliocene of South America (Patterson, 1937). These early Pliocene forms are almost identical with living species of the same genera and do not appear to be more primitive.

<sup>1</sup> This statement is based upon more than three thousand specimens of small mammals which I personally have collected in the Brulé of northwestern Nebraska.

<sup>&</sup>lt;sup>2</sup> This fact may help to account for the complete absence of indigenous placental mammals in Australia. If marsupials did originally migrate to Australia over a land bridge, instead of being "waif" immigrants, they were probably accompanied by insectivores. The ability of *Peralherium* to thrive in competition with the White River insectivores suggests the possibility that the original Australian marsupials may have been victorious over the placentals in the struggle for existence.

Dromiciops has usually been regarded as a descendant of a marmosine. The paracone of this genus, however, is not reduced—a very primitive character—suggesting an early pre-Peratherium separation of this genus. It seems improbable that the paracone was secondarily enlarged, since it is known to have been equal in size to the metacone in primitive genera such as Pediomys and Nanodelphys.

It seems probable that the modifications seen in living didelphines may have had their origin at the time of the original early Tertiary mammalian expansion. After that time the limits of expansion in North America would have been determined by the few ecologic niches left open by placentals. In Australia and, to a lesser degree, South America similar obstacles to expansion were not encountered.

#### MEASUREMENTS (In millimeters)

P25708 P25709 P25719 P25720 M<sup>1</sup> A-p.... 1.5 . . . . . . . . .  $\begin{array}{c}
 M^{1} & Tr. \\
 M^{2} & Tr. \\$ 1.31.5 . . . 1.5 1.6 1.6 1.7 1.6 2.01.6 M<sup>3</sup> A-p..... 1.6 1.3. . . . . . M<sup>3</sup> Tr..... 1.8 1.7 . . .  $\widetilde{M}^{4}$  A-p.... 1.0 . . . . . . . . . M<sup>4</sup> Tr..... 1.9 . . . . . . . . .

### LITERATURE CITED

#### BENSLEY, B. A.

1903. On the Evolution of the Australian Marsupialia; with Remarks on the Relationships of the Marsupials in General. Trans. Linn. Soc. Lond., (2), 9, pp. 83-217, 3 figs., pls. 5-7.

1906. The Homologies of the Stylar Cusps of the Upper Molars of the Didelphyidae. Univ. Toronto Studies, Biol. Ser. No. 5, pp. 149-159, figs. 1-6.

Dollo, L.

1899. Les ancêtres des Marsupiaux, étaient-ils arboricoles? Miscellanées Biologiques, pp. 188-203.

GREGORY, W. K.

1910. The Orders of Mammals. Bull. Amer. Mus. Nat. Hist., 27, pp. 1-524, figs. 1-30.

MATTHEW, W. D.

1901. Fossil Mammals of the Tertiary of Northeastern Colorado. Mem. Amer. Mus. Nat. Hist., 1, Part 7, pp. 355-447, figs. 1-34, pls. 37-39.

McGrew, P. O.

1937. New Marsupials from the Tertiary of Nebraska. Jour. Geol., 45, pp. 448-455, figs. 1-4.

PATTERSON, B.

1937. Didelphines from the Pliocene of Argentina. Proc. Geol. Soc. Amer., 1936, p. 379.

### 400 FIELD MUSEUM OF NATURAL HISTORY-GEOLOGY, VOL. VI

SIMPSON, G. G.

1928. American Eocene Didelphids. Amer. Mus. Nov., No. 307, pp. 1-7, figs. 1-5.

1929. American Mesozoic Mammalia. Mem. Peabody Mus., 3, pp. I–XV, 1–171, figs. 1–62, pls. 1–32.

1935. Note on the Classification of Recent and Fossil Opossums. Jour. Mamm., 16, pp. 134–137.

Тате, G. H. H.

1933. A Systematic Revision of the Marsupial Genus Marmosa, with a Discussion of the Adaptive Radiation of the Murine Opossums. Bull. Amer. Mus. Nat. Hist., 66, pp. 1-250, figs. 1-29, pls. 1-26.

WINGE, H.

1893. Jordfundne og nulevende Pungdyr (Marsupialia) fra Lagoa Santa, Minas Geraes, Brasiliens. E. Museo Lundii, 2, pp. 1-132.

### INDEX

#### VOLUME V

Ahumada meteorite, 1 Arispe meteorite, 2

Bishop Canyon meteorite, 3

Davis Mountains meteorite, 4

- Davis Mountains meteorite, analysis of, 9
- Greenland, composition of sands from, 24

Labrador, composition of sands from, 22

Macquarie River meteorite, 12

- Macquarie River meteorite, analysis of, 14
- Mineral composition of sands, laboratory procedure for determination of, 17-20

Quebec, composition of sands from, 20

Rawson-MacMillan Expedition, 17

South Bend meteorite, 14

#### VOLUME VI

Adinotherium, 17-21, 95, 107, 114, 210, 212, 214, 221, 222, 276, 278, 279, 286, 298 ovinum, 17, 212-214, 220-222, 282. 286-288 Adpithecus; see Notopithecus Aelurodon, 329 Ailuropoda, 325, 333, 334, 336, 337, 338 melanoleuca, 334 Ailurus, 325, 333, 334, 336, 337 fulgens, 333 Aletocyon, 337 multicuspis, 331 Allognathosuchus, 315, 318 mooki, 318 Amblypoda, 352, 373, 381 Ameghinotherium, 132 Amherst brain cast, 279 Amphicyon americanus, 349 amnicola, 348 aurelianensis, 349 frendens, 348 idoneus, 348 ingens, 348 palaeindicus, 349 pontoni, 349 reinheimeri, 348 riggsi, 341-350 shabbazi, 349 sinapius, 348 Ancylocoelus, 166, 215 frequens, 15-17; (Colpodon sp.), 94, 166, 215-216 Ankylodon, 267, 269, 271 annectens, 269-271 Araucanian-Entrerian series, 132 Archaeohyracidae, 131 Arctostylopidae, 108

Argyrohippus, 96, 97, 98, 161, 162, 164, 281 boulei, 161, 162 fraterculus, 96–98, 161, 162, 164, 166 praecox, 161–165 sp., 97, 109 Argyrohyrax, 21; see Plagiarthrus proavus; see Plagiarthrus proavus Arsinoitheria, 373 Artiodactyla, 373 Asmodeus, 100, 299 sp., 100, 107 Astrapotheria, 24, 25, 110, 176, 373 Astrapothericulus, 170 Astrapotherium, 110, 167, 173, 175-176 magnum, 167, 175 Bacteria in stony meteorites, 179 Barylambda, 229 (Titanoides), 173, 174, 229–230, 361– 364, 365, 367, 369, 370, 371, 372 faberi, 230, 365, 372 group, 370–371 Barylambdidae, 361, 371, 372 Barylambdinae, 230, 372 Bassariscus, 325, 326, 327, 331, 332, 335, 336, 338 astutus, 326 Bathmodon; see Coryphodon Bathyopsis, 374, 376, 377, 382 fissidens, 374 Bathyopsoides, 373-374, 376, 377, 379, 380, 382 harrisorum, 373, 374-378, 379, 380, 381 Bathyurus sculpinensis, 40, 59 sp., 44, 59 Bessoecetor, 268

402 FIELD MUSEUM OF NATURAL HISTORY—GEOLOGY, VOL. VI

Bison bison, 308, 310, 311 Blarina, 247, 249, 256 Blarina brevicauda, 247, 306 "Blarinae," 256 Borhyaena, 63, 64 Borhyaenidae, 65 Borhyaeninae, 65 Braincasts, method of making, 273 Canis, 329, 330, 346, 347, 348 familiaris, 307–308 latrans, 306, 311 lupus, 309, 311 Carnivora, 373 Casamayor formation, 132 Castoroides ohioensis, 306 Ceratiocaris leesi, 155 markhami, 142 Ceratosuchus, 315–316 burdoshi, 316–318 Cervalces roosevelti, 310 scotti, 310 Cervus canadensis, 310 Chert, analysis of, 82 Chironectes, 396, 397 Chrysemys sp., 311 Cochilius, 21, 86, 121, 134, 135 volvens, 23, 88 Colhué-Huapí formation, 132 Collon-Curá formation, 132 Colpodon, 15, 165, 166 propinquus, 94 sp., 166; see Ancylocoelus frequens Condylarthra, 369, 373, 382 Conularia manni, 147 Coresodon, 109, 281 Coryphodon, 172, 173, 174, 352, 353, 354, 356, 357, 358, 359, 360, 361, 362, 364, 365, 369, 370, 371, 372 group, 370-371 wortmani, 357 Coryphodontidae, 370, 371-372, 381 Creodonta, 382 Crinoidal stems, Labrador, 36 Crista meati, 85 Crocidura russula russula, 254 group, 254 Cynarctinae, 337 Cynarctoides, 324, 325, 327, 328, 330, 331, 336, 337, 338 Cynarctus, 323, 324, 325, 327, 329, 331, 335, 336, 337 acridens, 323, 324, 328, 336 crucidens, 323, 324, 329 saxatilus, 323, 324 Dalmanites pratteni, 67 Daphaenodon, 346, 347, 348, 350 Daphaenus, 346, 347 Deep River beds, 341

Deseado formation, 132, 299 Didelphidae, 393, 398

Didelphys, 396, 398 Dinoceras, 377 Domnina, 246–248, 250, 255–256 gradata, 246, 247, 248–255 Dromiciops, 393, 396, 399 Echinosoricinae, 268 Elachoceras, 382 Elasmosaurus platyurus, 385 serpentinus, 385–390 Embassis, 246 Entelonychia, 6, 23, 24, 297, 298 Entomolestes, 268 Eobasileus, 382 *Epitypotherium*, 132, 135 Erinaceidae, 245, 267, 268 Erinaceinae, 268 Eumys, 397 Eurygeniops, 108 Eurygenium, 108 Eurygenius, 108 Eutrachytherus, 119, 130 modestus, 133 Eutypotherium, 130, 131, 293 Ferrissia fusca, 304 Fossaria obrussa, 304 Fossils of Northeast Labrador, sources of, 49 Gastroliths of Elasmosaurus serpentinus, 390 Glironia, 396 Goldman, E. A., on Post-Glacial Indian Dog, 307 Goniobasis livescens, 304 Gravel pits in Coles County, Illinois, geology of, 303-304 Gypsonictops, 267 Gyraulus altissimus, 304 Haplolambda, 365, 367, 370 quinni, 365–367 Hegetotheriidae, 108, 131, 134–135, 136, 222, 224, 297, 299–300 Hegetotherium, 128, 129, 200, 204, 205, 212, 222, 223, 274, 276, 277, 293, 294, 295, 296, 297 mirabile, 200–203 Helicotoma rawsoni, 39, 59 Heliscomys, 397 Helisoma anceps, 304 Hemiechinus, 268 Heterosorex, 255, 256 Hicoria sp., 304, 312 Homalodotheriidae, 108, 225, 298-299 Homalodontotherium; see Homalodotherium Homalodotherium, 100, 106, 207, 215, 216, 224, 225, 242–243, 288, 291, 292, 293, 297, 298, 299

cunninghami (segoriae), 6-9, 99, 113-117, 216–220, 222, 223, 225, 233– 242, 288–293 segoviae; see H. cunninghami Homo sapiens, 308, 309 Hormotoma labradorensis, 37, 59 Hormotoma minuta, 38, 59 Hylomys, 268 Hypertragulus, 397 Hypisodus, 397 Hyracoidea, 222, 373 *Ictops*, 264 Illinois, fossil vertebrates from, 303 Interatheriidae, 108, 131, 134, 136, 222, 224, 297, 299 Interatherium, 134, 136, 204, 206, 207, 209 robustum, 204-206, 224 Interhippus, 109 Ischyromys, 397 Isoproedrium, 132 Isotemnidae, 108, 299 Isotypotherium, 132, 135 Labrador fossils, 33 La Flecha deposit, 165, 166 Larix sp., 304 Leidyosuchus, 318, 319, 320 acutidentatus, 320 multidentatus, 320 riggsi, 318-320 sternbergii, 320 Leontinia, 92, 99, 105, 108, 126, 216 gaudryi, 92-94, 105, 107 sp., 93 Leontiniidae, 108, 225, 297 Leptacodon, 266, 267 tener, 267 Leptictidae, 266, 267 Leptomeryx, 397 Limestones, Sculpin Island, 62 Silliman's Fossil Mount, 52 Limnoecus, 254 Loxolophodon, 352 Lutreolina, 398 *Lynx* sp., 311 Macrauchenia, 172 Mammut americanum, 308 Marmosa, 393, 394, 395, 396, 397 beatrix, 394, 395 Marsland beds, 324 Marsupial sabertooth, 61 Megalonyx jeffersonii, 306 Megalonyx? sp., 306 Meleagris galloparo, 311 Mesohippus, 397 Metacodon, 257, 263, 266–268, 269, 270, 271magnus, 257-258, 264 mellingeri, 258-266, 269, 270

Metamynodon, 168, 176 Miniopterus, 256, 257 Miothen crassigenis; see Domnina gradata "Miothen" gracile; see Peratherium huntii Monodelphys, 396, 397, 398 Moropus, 243 Morphippus, 96, 108, 109, 110, 164, 165, 281 Muñizia, 136–137 Muñiziinae, 136 Musters formation, 299 Mystipterus vespertilio, transferred from Chiroptera to Soricidae, 256-257 Nanodelphys, 393-399 minutus, 393, 394 Nectogale, 255 Neomys, 248 Nesodon, 17–21, 95, 107, 130, 200, 209, 212, 214, 221, 222, 276, 279, 281, 282, 284, 285, 286, 287, 288, 290, 291, 292, 298 imbrietus 10, 06, 200, 210, 200, 201 imbricatus, 19, 96, 209-212, 220, 221, 222, 281-286 Nesodontidae, 109, 286 Nesohippus, 109 Nothocyon annectens, 337 Notioprogonia, 223 Notohippidae, 108–110, 279, 281, 297, 298gen. et sp. indet., 279-281 gen. et sp. maet., 213-201 Notohippus, 109, 164 Notostylopidae, 108 Notostylopidae, 108 Notostylops, 129, 284, 290, 299 aspectans, 9-12 brachycephalus, 9 sp., 105 Notoungulata, 6, 23, 25, 91, 92, 107, 110, 199, 223-224, 273, 297-300, 373Odocoileus virginianus, 308, 310, 311 Oldfieldthomasia, 203, 207, 214, 215, 223, 279 Ondatra zibethica, 309 Orohippus, 107 Ovibos, 308, 312 Ovibovinae, 308 Pachyrukhos, 203, 207, 209, 222 moyani, 203–204, 224 typicus, 19 Palaeostylops iturus, 103-105 macrodon, 104 Paleolagus, 397 Pantodonta, 352, 353, 369-373, 382 Pantolambda, 230, 352, 353, 354, 356, 357, 358, 359, 360, 361, 364, 367, 369, 370, 371

#### 404 FIELD MUSEUM OF NATURAL HISTORY—GEOLOGY, VOL. VI

attenuatum, 207

Pseudemys sp., 311

australe, 206-207, 209, 224

Pantolambda bathmodon, 356 cavirictus, 356, 367 Pantolambdidae, 369, 370, 371, 381 Pantolambdinae, 372 Pantolambdodontidae, 370 Parastrapotherium, 110, 167 Parictis dakotensis, 324, 338 Patagonian formation, 132 Peradectes, 395, 396 Peramys, 397 Peratherium, 246, 247, 395, 396, 397, 398, 399 huntii, 248 sp., 247 Periphragnis, 299 Periptychidae, 369, 373, 382 Perissodactyla, 373 Phenacodus, 284, 291 Philander, 396 Phlaocyon, 325, 327, 330, 331, 335, 336, 337, 338 leucosteus, 324, 331 Phyllopod mandible, 155 Phyllopodous crustacean, new Silurian, 141Physa gyrina, 304 integra, 304 Pisidium sp., 304 Plagiarthrus (Argyrohyrax), 21, 107, 121, 131, 134, 135 proavus, 21–23 Plateau Valley beds, Tiffany age of, 351 vertical distribution of fossils in, 380 Pleurostylodon, 100, 105, 293, 299 biconus? 100 Poebrotherium, 397 Post-Glacial of Coles County, Illinois, 312Proadinotherium, 94, 98 leptognathum, 95, 97 muensteri, 94–96 Probathyopsis, 373, 374, 376, 378, 382 newbilli, 378–381 praecursor, 378, 380, 381 Procavia, 222 Procyon, 325, 330, 332, 333, 334, 335, 336, 337 lotor, 309, 332 priscus, 309 Procyonidae, 323, 324, 335, 338 Prodinoceras, 373, 374, 380 Proedrium, 132 Proedrus, 132 Prosotherium, 102, 103, 134, 136 Proterixoides, 267 davisi, 267 Proterotherium, 276 Protosorex, 246, 247 crassus, 247, 248, 255 Protypotherium, 23, 128, 129, 134, 136, 204, 205, 206, 209, 212, 293, 295, 296, 297

Pseudocynodictis, 325, 326, 327, 328, 331, 335, 338 gregarius, 325 Pseudotypołkerium, 83, 88, 120, 121, 122, 123, 124, 125, 126, 133, 208, 209, 224, 293 pseudopachygnathum, 83-89, 116, 208, 209Pyrotheria, 373 Rhynchippidae, 108-110 *Rhynchippus*, 108, 109, 110, 214, 215, 274, 276, 278, 279, 280, 281, 282, 284, 285, 286, 287, 290, 291, 292, 298, 299 equinus, 12-14, 98, 214-215, 274-279 pumilus, 98–99, 214–215, 218, 225 Rhyphodon, 274, 275, 276, 277, 278, 279, 290, 291, 292, 298, 299 Río Frías formation, 132 Río Mayo formation, 132 Rubber technique for braincasts, 273-274Santa Cruz formation, 132 Sculpin Island, Labrador, 35 Septum in notoungulate bulla, 222-223 Sorex, 247 Soricid, 245 Soricidae, 255 Sparactolambda, 352-354, 356, 357, 358, 359, 361, 362, 367, 370, 371, 372, 373 looki, 354-361 Sphaerium sulcatum, 304 Stagnicola reflexa, 304 Stilhippus, 109 Stratigraphy of northeast Labrador, 44 Sylvilagus floridanus, 306 Tachytypotherium, 130 "Taligrada," 373 Terrapene ornata, 311 Thomashuxleya, 236, 299 Thylacodon, 396 Thylacosmilinae, 65 Thylacosmilus, 61-65, 114 atrox, 62 lentis, 62 Tinoceras, 377 Titanoideidae, 372 Titanoides; see Barylambda faberi; see Barylambda faberi gidleyi, 229, 230 primaevus, 229, 230 Titanoidinae; see Barylambdinae Toxodon, 19, 20, 21, 85, 96, 275, 282, 284, 286, 287 burmeisteri, 18, 19, 285

## INDEX

platensis, 18, 87 Toxodonta, 12, 23, 24, 207, 222, 223, 224, 225, 297, 298 Toxodontia, 6, 23-25, 110 Toxodontia, 6, 23-25, 110 Toxodontidae, 108, 109, 225, 297, 298 Trachytherium, 101, 119 Trachytherus, 130–137, 280, 293, 294, 295, 296, 297 conturbatus, 120 grandis, 120 spegazzinianus, 101-102, 119-129, 274, 279, 293 Trachytypotherium internum, 21 Trigodon, 96 Trilobite, New Devonian, 67 Trinacromerum, 385 Tupaiodon, 260, 261, 267 Tupaiodon? minutus, 267, 268 Tupaiodon morrisi, 264 Turritoma cf. T. ada, 39, 59 Typotheria, 21, 23, 24, 136, 297, 299–300 Typothericulus, 133 Typotheriidae, 108, 132-133, 222, 224, 297, 299

Typotheriopsis, 84, 88, 122, 126, 133, 135, 224, 293, 294, 295, 296, 297

chasicoensis, 293 internum, 21, 293–297 studeri, 293 Typotherium, 19, 21, 86, 122, 129, 130, 131, 132, 133, 135, 295 cristatum, 18, 19, 83, 84, 85, 86, 87, 88, 131 Uintatheriidae, 381 Uintatherium, 352, 377, 382 Ulmus sp., 304 Ungulates, 373 Upper Canadian fossils, Labrador, 31 Upper Harrison, 323 Uquía formation, 132 Ursus (Euarctos) americanus, 309 Ursus gyas group, 309 horribilis group, 309 procerus, 309 sp. cf. U. horribilis, 309 Vulpes, 330

Xotodon, 236

"Zotodon;" see Xotodon

-.



