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EXPERIMENTAL MORPHOLOGY

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# EXPERIMENTAL MORPHOLOGY

BY

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## PART FIRST

*EFFECT OF CHEMICAL AND PHYSICAL AGENTS  
UPON PROTOPLASM*

New York

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**Dedicated**

TO THE MEMORY OF THE FIRST AND MOST IMPORTANT  
OF MY TEACHERS IN NATURAL HISTORY  
MY MOTHER

Die morphologische Betrachtung setzt also eine genaue chemisch physikalische Kenntniss, 1. des betreffenden Körpers selbst, und 2. aller der bei seiner Entstehung auf ihn einwirkenden Stoffe und Körper voraus. — JAEGER, *Zoologische Briefe*, p. 9.

La vie ne se conçoit que par le conflit des propriétés physico-chimiques du milieu extérieur et des propriétés vitales de l'organisme réagissant les unes sur les autres. — BERNARD, *Rapport sur les progrès de la physiologie générale en France*, 1867, p. 5.

There can be little doubt, indeed, that every science as it progresses will become gradually more and more quantitative. Numerical precision is doubtless the very soul of science, as Herschel says. — JEVONS, *Principles of Science*, chap. xiii.

## P R E F A C E

THE problem which, since the time of Aristotle, has stood first in interest and importance among the great questions of Biology is that of the causes which direct the development of the individual—that marvellous process by which the germ is built up into the complex organism, by which the embryo clothes itself with the characters peculiar to its species, by which even minute individual traits of form and action are exactly reproduced in the offspring from its parents.

The burden of clearing up this problem has fallen, naturally enough, upon the shoulders of students of morphology. For since morphologists deal with form, they are properly especially concerned with the interpretation of form—they may well be asked to account for it. Thus the problem of development is an acknowledged morphological problem.

Several distinct steps can be recognized in the progress which has been made in the interpretation of form. The earlier studies were concerned chiefly with answering the question, *What* are the differences between the various adult forms? The results of observations and reflections relating to this question constitute the sciences of descriptive and comparative anatomy. Next, a more fundamental inquiry was entered upon: *How* are these forms produced or developed? The results of observations and reflections upon this subject constitute the science of comparative embryology. Finally, in these later days a still more fundamental question has come to the front: *Why* does an organism develop as it does? What is that which directs the path of its differentiation? This is the problem which the new school of “Entwicklungsmechanik” has set for itself—it is likewise the problem with which this book is concerned.

The causes which determine the course of an organism's development are numerous, but fall into two general categories; namely, internal causes, which include the qualities of the developing protoplasm; and external causes, which include the chemical and physical properties of the environment in which the protoplasm is developing. The internal and external causes may be studied separately, and in order to disentangle their effects they must needs be studied separately. It is the purpose of the present work to consider the effects resulting from external causes.

When we wish to isolate the separate effects in any complex of causes, we must resort to the well-known procedure of experimentation,—and we find, indeed, that these external causes lend themselves readily to this method of treatment. Accordingly we call in experiment to get an insight into the causes of organic form, and thus justify the name which we have applied to our study,—*Experimental Morphology*.

The primary subdivision of the subject is based upon the morphogenic processes to be treated of; and of these, four principal classes may be recognized. The first includes those processes which are characteristic of all living *protoplasm*; the second, those connected with *growth*; the third, those involved in *cell-division*; and the fourth, those producing *differentiation*. It is proposed to devote one part of the work to each of these four classes of processes.

The secondary subdivision may be based upon the chemical and physical agents whose effects we wish to isolate. These may be grouped into eight categories, determined largely by convenience; namely, 1, chemical substances; 2, water; 3, density of the medium; 4, molar agents; 5, gravity; 6, electricity; 7, light; and 8, heat. It is proposed to devote one chapter to a consideration of the effects of each of these agents upon protoplasm, upon growth, upon cell-division, and upon differentiation.

Two words should be said about the point of view from which this book has been written. In the first place, the developing organism is regarded as a *living* organism, and as such endowed with irritability and capacity of response; consequently, at the outset, we must especially consider the phe-

nomenon of response to external stimuli. Again it is with living *organisms* that we have to deal, and, accordingly, no distinction should be made between animals and plants. I have, indeed, made no such distinction; nevertheless, tastes and training have led me to lay especial stress upon animals. Even this is unfortunate, for the problem with which we are concerned is precisely the same problem in all *living organisms*.

In the second place, much stress is laid upon the quantitative measurement of agents and effects. The lack of precision in many investigations can hardly be too strongly decried; for it often results in confusion and useless disputes. On the other hand, there is good reason for believing that exact measurement is the key to many of the most puzzling of our problems, and important results are to be expected from its use.

As for the aim of the book, it is twofold. I have hoped on the one hand that it might be readable to those who are interested in the general matters of which it treats — matters of importance for philosophy, for psychology, and for pedagogy. For man is an organism, and the development of his qualities is modified by just those agents which guide the development of other organisms. My primary aim, however, has been a different one. It is this aim to which other purposes have been made subservient, which justifies the historical treatment that has been often adopted, and justifies also the detailed descriptions of methods which the lay reader will, naturally, omit. This aim is so to exhibit our present knowledge in the field of experimental morphology as to indicate the directions for further research.

A few words of explanation and acknowledgment are necessary: It was planned at the first to issue all four parts of the work at once; but the task grew in the doing, while the need of its publication became more pressing. So it was decided to issue the work in parts as soon as each should be done. Even under this arrangement it has not been possible to include some of the papers of the last six months; especially I regret the omission of important papers by VERWORN and LOEB upon Galvanotaxis. In writing a book of this sort, which draws upon several sciences, I have had recourse to the kind assistance of several of my colleagues in the physical and

chemical departments of the University. I must especially thank for favors Professor W. C. SABINE, Dr. G. W. COGGESHALL, and Dr. H. E. SAWYER. Of my zoölogical associates, I am greatly indebted to Dr. G. H. PARKER, who has read nearly the entire manuscript and has offered valuable criticisms on it, and to Professor E. L. MARK, who has read parts of the manuscript and proof and has made important suggestions and emendations. I am also greatly indebted to Mr. CHARLES BULLARD for his kindness in making photographs of figures from which most of the illustrations of the First Part were reproduced. Finally, I cannot forbear to mention the painstaking work of my wife, GERTRUDE CROTTY DAVENPORT, in preparing the manuscript for the press and revising the proofs.

As I send out this work I do so with the hope that it may stimulate to even greater activity in the field of experimental morphology. The subject is new, its importance hardly yet generally recognized, its needs incompletely appreciated. In its scope it embraces much of physics and chemistry, for life and development are to be studied as the physicist studies light and heat, or as the chemist studies solutions and combustion. They are phenomena which must be analyzed by the use of instruments of precision to determine the quality and quantity of the acting agents, and to measure the change in the phenomena resulting from a change in these agents. No other field offers a better opportunity for the utilization of a broad scientific training. The times, too, are auspicious. Biology has never before attracted so many enthusiastic workers as it does to-day. As DRIESCH has said, "Die Lust an thatsächlicher exacter biologischer Forschung ist erwacht"; and the greatest problem of morphology is ever more and more the object of this biological experimentation.

CHARLES BENEDICT DAVENPORT.

CAMBRIDGE, MASS., Dec. 1, 1896.

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# EXPERIMENTAL MORPHOLOGY



## CHAPTER I

### *ACTION OF CHEMICAL AGENTS UPON PROTOPLASM*

IN this chapter it is proposed to consider (I) the effect of the various chemical agents upon the chemical constitution of protoplasm, as revealed by the results of their application, — death, modification of the metabolic processes, and of rate of movement; (II) the phenomena of acclimatization to chemical agents; and (III) the effect of such agents in determining the direction of locomotion, — chemotaxis.

#### § 1. MODIFICATION OF VITAL ACTIONS\*

The vital processes are chemical processes, taking place in a highly complex, very unstable, constantly changing substance, whose activities we call life. It is not easy to study this living substance chemically by the ordinary methods; for these usually, first of all, kill the substance. That the living substance and the dead are quite different is illustrated, for example, in the action of diamid ( $N_2H_2$ ) and hydroxylamine ( $NH_2 - O - H$ ), which show no action upon dead protoplasm, but are powerful poisons for all living plasm. The instability of protoplasm enables us, on the other hand, to make use of

---

\* In the preparation of this section, much use has been made of the admirable work of LOEW ('93). Not only is the adopted classification of poisons for the most part his, but also, in a few cases, passages from his book have been translated *in toto* here. Most of the determinations of killing strengths of the various reagents for which no other authority is given have been taken from LOEW'S book.

certain indirect means for determining its constitution. Since death is due to chemical change, we ought to determine what substances are fatal poisons to protoplasm; and since every activity of protoplasm is a chemical process, we ought to study the modifications of these processes by the action of various chemical reagents.

In studying the behavior of protoplasm in the presence of various reagents, we shall make use especially of observations upon Protista, sexual cells, and tissue cells. In cases where sufficient observations on isolated plant or animal cells are wanting, use will be made of observations upon Metazoa.

At the outset, attention should be called to the necessity of a more quantitative study of the subject. A quantitative study demands, especially, a careful noting of the conditions of the experiment; for the various physical conditions under which the reagent is applied modify the result. Thus, it has been shown, for example, by RICHET ('89, p. 212) that with various poisons the toxic dose diminishes in amount with the elevation of the temperature of the body.

1. **Oxygen.**—It is almost certain that no protoplasm can long survive in the absence of oxygen. Apparent exceptions are found in the case of the anærobic bacteria, some of which are killed in the presence of free oxygen, but multiply rapidly when the oxygen supply is cut off. It has been suggested that, in the case of these and some other parasitic organisms, oxygen is derived from the breaking down of O-containing compounds in the nutritive medium. (Cf. LOEW, '91, p. 760.)

The effect of *diminished oxygen* upon protoplasm is described by CLARK ('89, pp. 370, 371) and by DEMOOR ('94, p. 191). CLARK determined the minimum oxygen pressure necessary for the vital movements of the plasmodia of Myxomycetes and the protoplasm of plant hairs and tissue cells. This he found to range from 1 mm. (plasmodia of Myxomycetes) to 3 mm. (leaf hairs of *Urtica*) of mercury.

DEMOOR\* subjected *Tradescantia* stamen hairs, in water, to a

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\* In studying the effect of a vacuum, DEMOOR employed a piece of apparatus constructed essentially on the plan of an ENGELMANN'S chamber. This consists of a box whose top is a centrally perforated metallic diaphragm and whose bottom is a circular glass. The vertical walls consist of an outer cylinder, at

pressure of 6 to 8 cm. of Hg, or 0.08 to 0.1 of an atmosphere (p. 71). In one hour, on the average, the protoplasmic movements were affected, and in most cells ceased in 2 to 3 hours, slight oscillations only of granules occurring. Thus, not death, but arrest of activity, occurred during this period as a result of reduction of atmospheric pressure — upon which the amount of oxygen held in the water depends. That death did not occur is shown by the fact that when air was readmitted at the normal pressure the protoplasm promptly regained its normal activity. Cells immobilized during 24 hours regain their movements in less than 5 minutes, and these become normal in from 10 to 20 minutes.

*Pure oxygen* acts in an opposite fashion from diminished oxygen tension, exaggerating the activity of protoplasm. Under its action the protoplasmic movements are much accelerated, but preserve, meantime, their normal character. (*Tradescantia* hairs, leucocytes; DEMOOR, '94, pp. 192, 218.) In Ciliata the rate of the contractile vesicle does not, however, seem to be altered. (ROSSBACH, '72, p. 40.)

*Ozone* and *hydrogen peroxide* produce atomistic "active" oxygen by becoming split up in the plasma. Ozone ( $O_3$ ) is said to kill quickly bacteria in water, if the latter does not contain too much organic substance; in the dry state, however, bacteria are injured only slowly by it. (OHLMÜLLER, '92, p. 861.)

Other substances which, with a greater or less degree of probability, may be said to act through oxidation of the protoplasm, may be treated of here.

*Hydrogen peroxide* ( $H_2O_2$ ).—PANETH ('89) added one part of neutralized  $H_2O_2$  to 10,000 (0.01%) of hay infusion, and found that all Ciliata were dead within 15 to 30 minutes. Stronger solutions act more rapidly; and even in a 0.005% solution, only part of the animals survived. Algæ survived only 10 to 12 hours in a completely neutral 0.1% solution. A 10% solution is fatal in a few minutes. (Cf. BOKORNY, '86, p. 355.)

*Salts of chromic, manganic, permanganic, and hypochlorous*

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the periphery of the diaphragm, and an inner cylinder at the inner margin of the diaphragm. An inlet and an outlet tube communicate with each of the spaces, — the central space and that between the two cylinders.

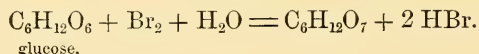
*acids* act as intense poisons, apparently by directly yielding oxygen atoms to the plasma proteins.

*Sodic chromate* ( $\text{Na}_2\text{CrO}_4$ ). — Many anærobic Schizophytes are killed even by a 0.05% solution of this salt. Splenic fever bacteria do not develop in a 0.05% solution in bouillon; in an agar-agar solution, they do not cease to develop until a concentration of 0.5% is reached, although they no longer produce spores in a 0.05% solution. Sodic chromate also acts strongly on algæ. (LOEW, '93, p. 16.)

*Potassic dichromate* ( $\text{K}_2\text{Cr}_2\text{O}_7$ ). — A 0.1% solution kills algæ (*Spirogyra*) in a few hours.

*Potassic permanganate* ( $\text{KMnO}_4$ ) is an energetic poison for algæ and Infusoria. A 0.2% solution kills Infusoria (*Paramecium*) in one minute.

Chlorine, bromine, and iodine, as well as hypochlorous acid salts, act, even in very considerable dilution, fatally upon all organisms, by splitting water, forming hydro-halogen compounds, and leaving the oxygen to unite with the living protoplasm. The action of bromine upon glucose may be written —



(LOEW, '93, p. 15.)

BINZ has pointed out that (on Infusoria) the poisonous action of these three halogens, like their other chemical properties, diminishes with increase of atomic weight, in the series Cl, Br, I. (Compare the *osmotic* effects of the halogens, p. 72.)

*Potassic chlorate* ( $\text{KClO}_3$ ) — also similar salts of I and Br — oxidizes in an essentially different fashion from the permanganates. For the latter oxidize even dead organic matter, but the former does not. This reagent may be considered a passively oxidizing one. Concerning its visible effects, we find that bacteria in general are injured by a 2% solution; with weaker solutions in nutrient media the bacteria reduce it to KCl. The anærobic forms are affected by a 0.5% solution; the aërobic withstand up to 3%. Algæ (*Spirogyra*) die after a few days in a 0.01% solution of the salt. (LOEW, '93, p. 17.)

*Arsenious acid* ( $\text{H}_3\text{AsO}_3$ ) and to a less degree arsenic acid ( $\text{H}_3\text{AsO}_4$ ) are poisons which BINZ and SCHULZ ('79) believe

to act by oxidizing the protoplasm. Thus  $H_3AsO_3$  can take up free oxygen as it would be found in water, and it can part with it readily to the protoplasm, thus oxidizing and eventually wholly consuming it. Such is one theory of its action. A few words as to effects upon Protista: Infusoria survive in 0.1% potassic arsenite in spring water only a short time, but live for weeks in a 0.1% solution of the potassic arsenate. (LOEW, '83, p. 112.) Algæ (*Spirogyra*) are killed by a 0.1% solution of potassic arsenite in six days, — the protoplasm contracts and shows formation of granules, the death of the chlorophyll bands preceding that of the cytoplasm. The same solution of potassic arsenate, meantime, shows no injurious action, (LOEW, '87, p. 445.) Still other arsenious acid salts tried upon other algæ (*Zygnema*, *Diatomacea*), upon Infusoria, and upon tadpoles showed themselves, uniformly, more powerful agents than the corresponding arsenic salts. The lower fungi are only slightly affected by arsenious salts; not at all by those of arsenic acid.

2. **Hydrogen.** — KÜHNE ('64, p. 52) subjected *Amœba* to H for 24 minutes. At the end of that time, some individuals had assumed a spherical shape, others appeared unchanged in form, but were motionless. Similar results were obtained with *Actinophrys*, the plasmodium stage of *Myxomycetes*, and with the stamen hairs of *Tradescantia*.

DEMOOR ('94, p. 190) also experimented upon the latter object, and his results are worth giving in detail.\* During the first moments of the passage of the gas, the protoplasmic movements are slightly accelerated. Soon the protoplasm becomes very granular, and, after a variable time, 15 to 40 minutes, is quiet. The aspect of the protoplasm at this time varies with the character of the cell. If it is young, having a large nucleus and without a primordial utricle, the protoplasm appears uniformly granular. If, on the contrary, the cell possesses a great reserve of water, with long, protoplasmic filaments, the protoplasmic granules become more refringent, increase in volume, and accumulate around the nucleus, — the peripheral

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\* Method: The hydrogen gas may be generated in a KIPP'S apparatus, and should pass through a series of washing flasks containing, *e.g.*, solutions of potash and acetate of lead. See VERWORN: *Allgemeine Physiol.*, p. 285. DEMOOR kept his stamen hairs in sugared water in an ENGELMANN'S chamber.

protoplasm appearing hyaline. The living substance is in repose. The hydrogen may be passed through the apparatus containing the stamen hairs for from 1 to 5 hours without any movement or other change appearing in the protoplasm. Air is now admitted. The protoplasmic movements rapidly return; the granules at first oscillate in their places, then gradually extend the range of their movement. In 5 to 6 minutes the cell has regained all of its anatomical and physiological characters. A similar immobility affects also leucocytes subjected to hydrogen. This occurs in about an hour; but there is great individual variation in this respect. Upon substituting air, the activity of the protoplasm is resumed in from 10 to 20 minutes. Protoplasm which has been subjected to the action of hydrogen thus appears not to be permanently modified, since normal movements recur rapidly upon readmitting air. It seems probable, therefore, that the temporary cessation in movements in the presence of hydrogen is due to the exclusion of oxygen from the protoplasm.

3. The two **Oxides of Carbon**,  $\text{CO}_2$  and  $\text{CO}$ , have very different effects upon protoplasm. Thus DEMOOR ('94, pp. 191, 202, 219) found that whereas the former immobilizes quickly, but kills very slowly, perhaps chiefly by asphyxia, the latter seems in some cases actively to attack the protoplasm. In leucocytes, the ectosarc is separated from the endosarc in a number of completely hyaline fragments; the endosarc becomes vacuolated, and death ensues in from 20 to 60 minutes. Many bacteria are only slightly affected by  $\text{CO}$ .

4. **Ammonia** ( $\text{NH}_3$ ). — A 10% solution provokes vacuolization, partial coagulation, and irregular movements in the protoplasm of the *Tradescantia* hair. The cell finally enters into repose, all the granules accumulating around the nucleus. Washing the preparation with water restores the original characters of the protoplasm. Thus, ammonia at first energetically excites protoplasm, later producing anæsthesia. (DEMOOR, '94, p. 193.) Even with very weak aqueous solutions (0.005%), which do not kill the protoplasm, BOKORNY ('88) has observed the production in *Spirogyra* cells of granules, which process does not, however, seem to modify the normal activities of the cell. These granules, "proteosomes," are intensely blackened



by alkaline (0.001%) silver solutions. Other basic substances, like potash and organic amine bases, and various alkaloids, produce the same effect. (Cf. LOEW and BOKORNY, '89.)

*Azoimid.* — LOEW ('91) has studied the effect upon protoplasm of this somewhat close ally of ammonia,  $\begin{array}{c} \text{N} \\ \parallel \\ \text{N} \end{array} \rangle \text{NH}$ . The poisonous action of this substance seems to depend upon its excessively unstable structure, for it easily disintegrates with violent explosion and production of ammonia. This latter then produces the characteristic granulations. Infusoria are killed in 2 to  $2\frac{1}{2}$  hours by a 0.1% solution of  $\text{N}_3\text{Na}$ , and the water-living Nematodes, Planaria, Ostracoda, Copepoda, and young Planorbis and Lymnæa are killed by a 0.05% solution in 30 to 40 minutes. Algæ are more resistant.

5. **Catalytic Poisons.** — There is a large number of unstable C-compounds which are neither acid nor basic nor characterized by chemical energy, which are, nevertheless, intense poisons for all living cells. Here belong the anæsthetics — ethylether, chloroform, chloral, carbontetrachlorid, methylal, alcohols, carbon disulphide, etc.\*

NÄGELI believes these to act as poisons by virtue of an inherent lively condition of molecular movement, which disturbs the normal condition of movement in the living plasma body, and, on that account, produces death. LOEW believes, more precisely, that the transmitted condition of violent movement leads to chemical transformations in the unstable albumen of the protoplasm.

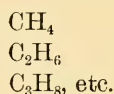
As examples of the effect of the mere presence of many unstable carbohydrates upon chemical processes, it has been found that HCl and prussic acid, which unite alone only at a high temperature, unite in the presence of various ethers at  $-15^\circ$ . Again, the mere presence of some CH compounds transforms a substance into its isomeric condition. Thus, thiourea is transformed by an alcoholic solution of amylnitrite into its isomer rhodanammium. Such poisons, which change the protoplasm by transmission of molecular movements, may be called cata-

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\* This paragraph and the two following are largely translated from LOEW ('93).

lytic poisons; the process which they inaugurate being known as catalysis (apparently produced by mere contact).

First will be considered the laws of *relation between molecular composition and strength of action*. We may begin with the methan series. This series, which has  $\text{CH}_3$  — as its base, runs as follows:—



In the members of this series, *the poisonous action increases up to a certain limit, with the number of C atoms*; above that limit the compounds are more stable and are more indifferent; *e.g.* paraffine ( $\text{C}_{21}\text{H}_{44}$  to  $\text{C}_{27}\text{H}_{56}$ ).

Beginning with methan,  $\text{CH}_4$ , we find this substance — marsh gas — innocuous when mingled with air. *As the H atoms become replaced by one or more chlorine atoms, the poisonous qualities increase,* —

$\text{CH}_3\text{Cl}$  is slightly anæsthetic,  
 $\text{CHCl}_3$  = chloroform,  
 $\text{CCl}_4$  is very dangerous, stupefying involuntary muscles.

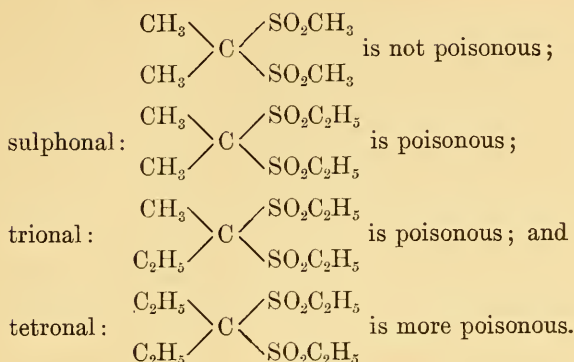
If the H atoms are replaced by any other halogen, — *e.g.* I, — anæsthesia is produced among some Vertebrates. Thus, 0.5 to 1 grain of  $\text{CH}_2\text{I}_2$  kills a rabbit.

In ethan ( $\text{C}_2\text{H}_6$ ) also, when Cl replaces H, the substance becomes a more active poison; *e.g.*  $\text{C}_2\text{H}_5\text{Cl}$ , ethyl chloroform, acts like chloroform.

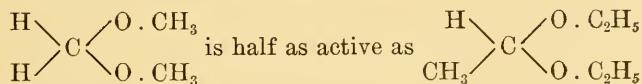
Also, among the sulphur hydrocarbons we observe the same fact of increase of poisonous action with increase in the number of Cl atoms to the molecule; thus, —

sulphur ethyl,  $\text{C}_2\text{H}_5-\text{S}-\text{C}_2\text{H}_5$  is a weak poison.  
 monochlorsulphurethyl,  $\text{C}_2\text{H}_5-\text{S}-\text{C}_2\text{H}_4\text{Cl}$  is a stronger poison.  
 dichlorsulphurethyl,  $\text{C}_2\text{H}_4\text{Cl}-\text{S}-\text{C}_2\text{H}_4\text{Cl}$  is a very powerful poison.

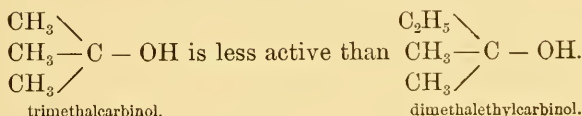
In the more complex sulphonic hydrocarbons of the methan series, where the alkyls  $\text{CH}_3$  —,  $\text{C}_2\text{H}_5$  —, etc., are introduced, the rule holds that *the more atoms in the alkyl the more active the substance as a poison*; thus, —



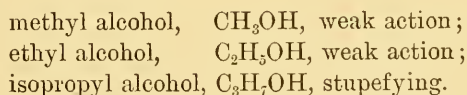
The same holds for the acetals; thus, —



We find the same thing in the ethyl group, —



And also in the alcohols, —



A few words now concerning the *morphological changes* observed in protoplasm subjected to the action of poisons belonging to this group. Here belong especially the various anæsthetics.

*Chloroform* and *ether* seem to affect all protoplasm anæsthetically, that of the higher plants as well as that of the higher animals. (BERNARD, C., '78, and ELFING, F., '86.) KÜHNE ('64, p. 100) first studied the effect of chloroform vapor upon *Tradescantia* hairs, but DEMOOR ('94, p. 193) has since described the action of this reagent in much more detail.  $\frac{1}{4}$  chloroform water at first (2 to 5 minutes) produces a very intense excitement in the movements of the protoplasm, a strong vacuolization occurs, and then the cytoplasm gradually

becomes immobile and dies in from 15 to 30 minutes. The nucleoplasm is less energetically acted upon than the cytoplasm. Upon swarm-spores, which are highly responsive to light (p. 182), weak solutions of ether and chloroform have such an effect that without preventing locomotion they destroy the power of responding to the stimulus of the external agent. So, too, the migrations of the chlorophyll in *Metaphyta*\* under the influence of light (see p. 189) is prevented. Ciliata are slightly paralyzed, for the period of the contractile vacuole is diminished. The whole cell body becomes distended with water, and the trichocysts are exploded. (SCHÜRMEYER, '90, p. 453.)

When chloroform water is slowly applied to leucocytes they acquire a spherical form; when quickly applied they become immobile without change of form. The first effect is a very intense increase in movements, especially of the ectosarc. Ultimate washing in serum suffices to restore the leucocyte to its wonted activity; so that it has not been killed, but only anesthetized. (DEMOOR, '94, p. 217.)

*Chloral hydrate*,  $\text{CCl}_3 - \text{C} \begin{array}{l} \diagup \text{OH} \\ \text{---} \text{OH} \\ \diagdown \text{H} \end{array}$ , which is closely related to

chloroform,  $\text{Cl} - \text{C} \begin{array}{l} \diagup \text{Cl} \\ \text{---} \text{Cl} \\ \diagdown \text{H} \end{array}$ , acts similarly as a protoplasmic anæ-

thetic. A 0.1% solution kills Infusoria, Rotifera, and diatoms in 24 hours, but filamentous algæ and Nematoda withstand it.

*Sulphonal* in 0.1% solution is less injurious than the preceding, since during 24 hours the above-mentioned organisms are uninjured. (LOEW, '93, p. 25.)

Upon the effect of *alcohols* on protoplasm, extended experiments have recently been made by TSUKAMOTO ('95). These reveal in much detail the peculiarities of action of the different kinds. I give three tables showing the time of resistance in hours of various organisms to the various alcohols, constructed from data furnished by his paper.

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\* ELFING, F. ('86, pp. 47-51). The swarm-spores employed belonged to the species *Chlamydomonas pulvisculus*. The strengths of the solutions which inhibit their response without stopping locomotion are: of ether, 1% to 5%; of chloroform, 12% to 25%. The migration of chlorophyll in *Mesocarpus* is inhibited by a 1% to 2% ether solution.

TABLE I

TIME (IN HOURS) OF RESISTANCE OF TADPOLES OF *BUFO VULGARIS*, LAUR.  
(HIND LEGS HAD JUST APPEARED) TO VARIOUS ALCOHOLS

STRENGTHS.	0.01%	0.1%	0.3%	0.5%	0.7%	1.0%	1.5%	2.0%	2.5%
methylic* . . . . .								0.3-0.8	0.07-0.3
ethylic . . . . .							stupor	0.16	
propylic, norm. . . . .				0.6	0.3	0.15			
propylic, iso. . . . .				0.5	0.2-0.3	0.15			
butylic, norm. . . . .		1.0	0.08						
butylic, iso. . . . .			0.15-0.25	0.05-0.15					
butylic, tertiary. . . . .				0.15-0.33	0.05-0.25				
amylic, norm. . . . .		0.2-0.5							
allylic . . . . .	4-5	1.5-2.0							

TABLE II

TIME (IN HOURS) OF RESISTANCE OF INFUSORIA AND OSTRACODA TO VARIOUS  
ALCOHOLS

STRENGTHS.	0.005%	0.1%	0.5%	1.0%	3.0%
methylic . . . . .					20
ethylic . . . . .					4
propylic, norm. . . . .				72†	
propylic, iso . . . . .				18	
butylic, norm. . . . .			48	18	
butylic, iso . . . . .			72	48†	
butylic, tertiary. . . . .				48+†	
amylic, norm. . . . .		24+	24		
allylic . . . . .	24				

\* The structural formulas of these alcohols are given here for reference:—

methylic . . . . .	H — CH <sub>2</sub> OH
ethylic . . . . .	CH <sub>3</sub> — CH <sub>2</sub> OH
propylic, norm. . . . .	CH <sub>3</sub> . CH <sub>2</sub> — CH <sub>2</sub> OH
propylic, iso. . . . .	(CH <sub>3</sub> ) <sub>2</sub> — CHOH
butylic, norm. . . . .	CH <sub>3</sub> . CH <sub>2</sub> . CH <sub>2</sub> — CH <sub>2</sub> OH
butylic, iso. . . . .	CH <sub>3</sub> > CH — CH <sub>2</sub> OH
butylic, tertiary . . . . .	CH <sub>3</sub> > COH — CH <sub>3</sub>
amylic, norm. . . . .	CH <sub>3</sub> . CH <sub>2</sub> . CH <sub>2</sub> . CH <sub>2</sub> — CH <sub>2</sub> OH
allylic . . . . .	. CH <sub>2</sub> . CH — CH <sub>2</sub> OH

† Ostracoda only ; the Infusoria died after 18 hours.

TABLE III

TIME (IN HOURS) OF RESISTANCE PERIOD OF SPIROGYRA COMMUNIS TO VARIOUS ALCOHOLS

STRENGTHS.	0.005%	0.01%	0.05%	0.1%	0.5%	1.0%	2.0%	3.0%	4.0%
methylic . . . . .							120	96	48
ethylic . . . . .							72	72	48
propylic, norm. . .						72			
propylic, iso. . . .						48			
butylic, norm. . . .					72	48			
butylic, iso. . . . .					96	48			
butylic, tertiary.						48			
amylic, norm. . . . .					24				
allylic . . . . .	66	72	24	24					

From these experiments it appears that allylic alcohol is more injurious than the others, so that TSUKAMOTO ('95, p. 281) believes it to attack the protoplasm directly rather than to act merely catalytically. We see also that the rule enunciated above about the greater activity of substances with more complex alkyls holds true in general. Of the butylic alcohols the normal is the most poisonous; the tertiary, least.

*Carbonic disulphide* ( $CS_2$ ) is one of the more powerful catalytic poisons. A saturated aqueous solution, which contains only a trace of  $CS_2$ , nevertheless kills quickly algæ, bacteria, and the lower water animals. (LOEW, '93, p. 29.)

6. **Poisons which form Salts.**—This is the third group recognized by LOEW. In this case we have to do with acids and bases which unite with the protein substances of the protoplasm-producing salts. Thus disturbances leading to death are produced. In addition this group comprises the poisonous metallic salts. So we may recognize three groups: *a.* acids; *b.* the soluble mineral bases; *c.* salts of heavy metals.

*a. Acids.*—The strong *inorganic* acids act, in general, more powerfully than the organic. Most bacteria, algæ, and Infusoria are very sensitive to inorganic acids (see MIGULA, '90), but splenic fever bacteria resist 1% HCl for 24 hours, and their spores 2% HCl for 48 hours. Mold withstands 1% phosphoric acid. Certain tissues have gained a high resistance capacity to inorganic acids. Thus, the gland cells of marine

Gastropoda (Dolium, Cassis, Tritonium, Natica heros) secrete 2% to 3%  $H_2SO_4$ . (LOEW.)

To *organic* acids many algæ are little resistant. Thus Spirogyra and Sphæroplea die in 0.1% malic or tartaric acid after 30 minutes; in 0.05% malic or tartaric acid after 24 hours; in 0.01% of these same acids in a few days. Formic acid prevents development of bacteria even in small percents — 0.05% to 0.006%. On the other hand, some protoplasm has acquired a resistance to organic acids. The vinegar eel — Rhabditis aceti — lives in 4% acetic acid. The protoplasm of the Drosera tentacle resists 0.23% tartaric, citric, and other organic acids.

b. *Soluble mineral bases*, including those of corrosive alkalies and the alkaline earths: Ca, Ba, and Sr. The corrosive alkalies cause a swelling of the protoplasm, but the primary effect is rather a chemical one. (Cf. FROMANN, '84, p. 90.)

The lower water animals and plants are quickly killed by 0.1% potassic or sodic hydrate. Thus, the movements of Chara cease in 0.05% KOH in 35 minutes. Bacteria are more resistant; the limit for the typhus bacilli being between 0.10% and 0.14%, and for the cholera bacillus, between 0.14% and 0.18%. Ascaris is still more resistant, living for 20 minutes in a 2% solution of NaOH.

CaO is still more powerful. A 0.007% to 0.025% solution in bouillon kills bacilli. A 0.013% solution is fatal to algæ like Spirogyra.

$K_2CO_3$  kills bacteria in 0.8% to 1.0% solutions.

$Na_2CO_3$  kills Ascaris in a 5.8% solution after 5 to 6 hours. (LOEW, '93, pp. 33, 34.) FROMANN has discussed the histological changes in protoplasm after treatment in  $Na_2CO_3$ .

It is difficult to say whether the action of some of these reagents may not be an osmotic, rather than a chemical one. The action of  $Na_2CO_3$ , for example, as described by FROMANN, is very similar to that of NaCl, whose action is probably solely osmotic.

c. *Salts of Heavy Metals*. — The method of action of these poisons has been accounted for upon the following grounds: When amido-acids (which are found as disintegration products of all animal tissues) are treated with salts of the heavy metals,

the hydrogen in either the carboxyl-group or the amido-group can be replaced by the metal. Likewise the hydrogen of the amido-group in urea derivatives and many bases are replaceable by metals. In the still more complicated protein stuffs the H bound to the N or O can be replaced. Many metals, indeed, like silver or mercury replace *preferably* the H of the amido-groups, and on this account, perhaps, their salts are especially poisonous. (LOEW, '93, p. 34.)

Salts of Hg, Ag, and Cu cause death to Spirogyra even in a dilution of 1 : 1,000,000; the chlorophyll bodies being first affected.\* Upon the bacteria of splenic fever the double cyanides of Ag, Hg, and Au are the most injurious, next those of Cu, Pb, and Zn, and, finally, those of Pt, Ir, and Os. Tadpoles and Tubifex are killed in 24 hours by solutions of  $\text{CuSO}_4$  weaker than 0.00005%. (LOCKE, '95, p. 327.) Among mercuric salts, splenic fever bacteria do not develop in 0.0003%  $\text{HgCl}_2$  in nutritive bouillon, nor 0.0125% in blood. Lactic acid bacteria do not reproduce in 0.0007%. Mold spores are killed in

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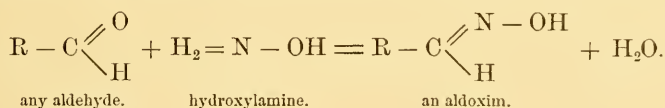
\* In this connection reference must be made to the posthumous paper of NÄGELI ('93), "Ueber oligodynamische Erscheinungen in lebenden Zellen." This author found that water distilled in copper vessels or 1 litre of water in which 12 clean copper coins had stood for four days acted fatally upon Spirogyra. The water was found in one such case to contain 1 part Cu in 77,000,000 of water. It was believed to be in solution in the form of the hydroxyd ( $\text{CuH}_2\text{O}_2$ ). Similarly produced solutions of other metals, Ag, Zn, Fe, Pb, Hg, had a similarly fatal effect upon Spirogyra. NÄGELI believed that the effect of the metals was not a chemical one, but was due to a new force — "oligodynamic." Besides the fact of the action of very dilute solutions, the only evidence he adduced for the new force was based on the difference of action on the chlorophyll bands of solutions of 1:1000 or 1:10,000 and 1:10,000,000. In the weaker solutions ("oligodynamic" action) the bands alone were drawn away from the cell-wall, in the stronger solution (chemical action) the whole peripheral protoplasm was shrunk away. It does not seem necessary to invoke a new force to explain the action of weak solutions: first, because the two actions are not sharply separated, according to NÄGELI's own data; secondly, because the chlorophyll bands are in general more sensitive than the rest of the protoplasm (p. 5); and, thirdly, because the action of so weak a solution is not surprising in view of the fact that Spirogyra is one of the least resistant of organisms. Even in a comparatively resistant organism, like Stentor, a solution of 1 : 80,000,000  $\text{HgCl}_2$  produces acclimatization to the poison and 1 : 10,000,000 has an injurious effect. Yet between the action of such solutions and those of 1 : 1000 there is a complete graduation in increasing effect. (See p. 30.) Even in NÄGELI's experiments solutions of less than 1 : 100,000,000 had little action.



0.1%. NEAL and I have found that *Stentor cœruleus* is killed by a 0.001% solution  $\text{HgCl}_2$  in a few seconds. (Cf. p. 30.) Ascarids die in a 0.1% solution in an hour. (SCHRÖDER, '85.)

Silver salts occasionally act upon bacteria more energetically than those of Hg. Cadmium and zinc salts are poisonous—the former more so than the latter. Thus, whereas 0.015% of cadmium sulphate inhibits reproduction of lactic acid bacteria, 0.1% of zinc sulphate is not injurious. Many salts of thallium are likewise active. Thus LOEW ('93, p. 37) found that in 0.1% thallium sulphate *Spirogyra* died in 4 to 6 hours.

7. **Substitution Poisons.**—In this group LOEW places certain nitrogenous substances which attack the amido and aldehyde groups of living protoplasm. These are extremely unstable substances and may therefore be transformed by agents which have no effect upon dead protoplasm. The supposed method of action of a poison upon an aldehyde may be illustrated in the case of the poison hydroxylamine ( $\text{H}_2\text{N} - \text{OH}$ ); which justifies at the same time the term “substitution poisons.”



*Hydroxylamine.*—This is a general and powerful poison. Thus, among the lower organisms, a solution of neutral hydroxylamine of —

0.001% kills diatoms within 24 hours. (LOEW, '85<sup>a</sup>, p. 523.)

0.005% kills in 36 hours Infusoria which withstand a similar concentration of strychnine. (LOEW.)

0.01% kills diatoms in something less than 15 hours; Planaria and leeches in 12 to 16 hours. (LOEW.)

0.1% paralyzes the muscles of Rotifera in 10 to 15 minutes; those of Nais in 20 to 30 minutes. (HOFER, '90, pp. 324, 325.)

0.2% kills Rotifers, Copepoda, and Isopods in 1 hour (LOEW); stupefies *Vorticella* in from 2 to 10 minutes. (HOFER, '90, p. 325.)

0.25% stupefies *Stentor* in 10 to 20 minutes. (HOFER.)

*Benzenylamidoxim* and *acetoxim*, more complex derivatives of hydroxylamine, are somewhat less poisonous.

*Diamid*, or *hydrazin* ( $\text{H}_2\text{N} - \text{NH}_2$ ) in the form of neutral solutions of the sulphate is a rapid poison. A solution of —

0.01% kills various alga species in 1 to 2 days.

0.02% is injurious to bacteria.

0.05% kills various water animals within 12 hours. (Loew, '93.)

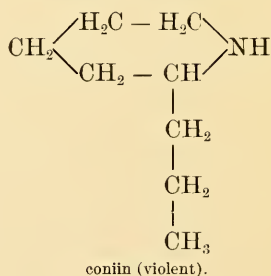
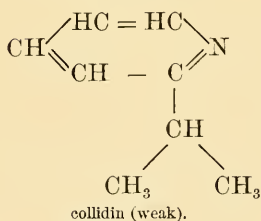
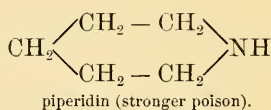
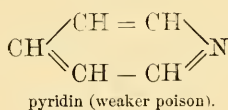
*Phenylhydrazin*  $\left\{ \begin{array}{l} \text{C}_6\text{H}_5 \\ \text{H} \end{array} \right\} \text{N} - \text{NH}_2$  is more powerful. A solution of —

0.0067% kills Infusoria and algæ within 18 hours.

0.05% prevents the development of bacteria and mucors.

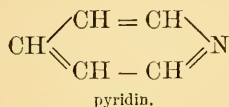
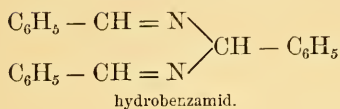
As free ammonia ( $\text{NH}_3$ ) is a far weaker poison than diamid ( $\text{H}_2\text{N} - \text{NH}_2$ ), so *anilin* ( $\text{C}_6\text{H}_5\text{NH}_2$ ) is far weaker than phenylhydrazin ( $\text{C}_6\text{H}_5 \cdot \text{NH} \cdot \text{NH}_2$ ).

Passing now to the more complex nitrogenous compounds, we find, first, that bodies which possess slight or no poisonous power, and contain tertiary N, can become strong poisons by addition of H and formation of imido-groups (*i.e.* groups which can be derived from ammonia by the substitution for two H atoms of bivalent acid radicals); thus, —

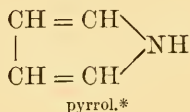
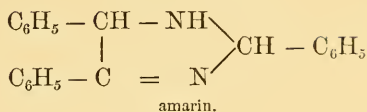


In the preceding and the two following cases it is seen that, in general, when there is a hydrogen atom of the amid radical ( $\text{NH}_2$ ) unreplaced by an alkyl, the substance is poisonous.

## ONLY SLIGHTLY POISONOUS.



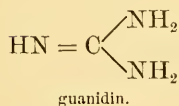
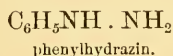
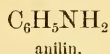
## VIOLENT POISONS.



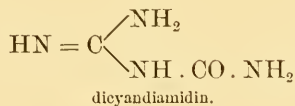
This increased poisonousness, correlated with the presence of H joined to N, may be accounted for by the union of this H with the O of the ketons or aldehydes of the living substance.

Likewise when one or more H atoms of the amido-group are replaced by an acid radical (*e.g.* that of acetic acid,  $\text{CO} \cdot \text{CH}_3$ ), the poisonous qualities of the substance are considerably diminished; thus, —

## MORE POISONOUS.



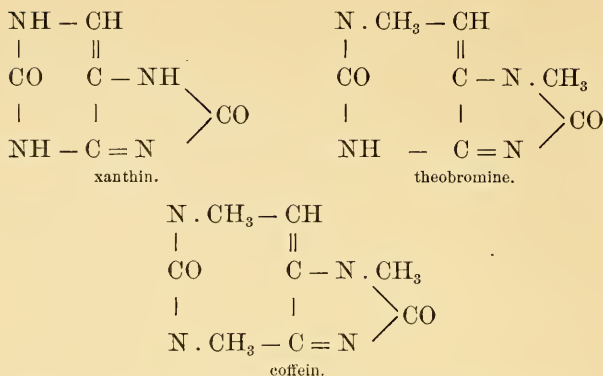
## LESS POISONOUS.



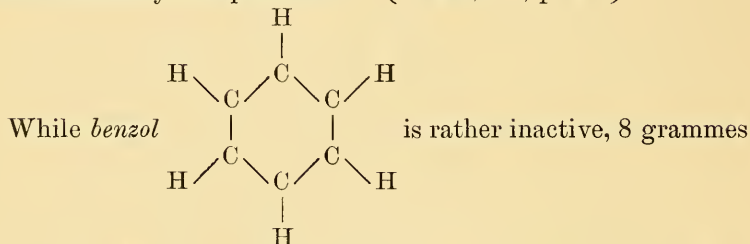
In like manner when, in an imido-group, the H (of the NH radical) is replaced by alkyls (*e.g.*  $\text{CH}_3$ ), the substances become less poisonous; thus, —

\* While a 0.07% solution of pyrrol kills Isopods, Rotifers, Planaria, etc., in about 1 hour, these organisms withstand a solution of pyridin of the same strength. (LOEW, '87, p. 444.)

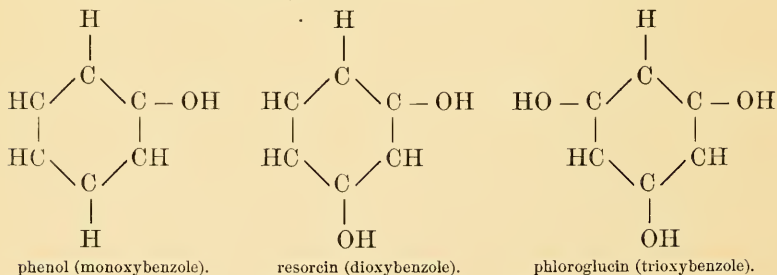
† SCHÜRMEYER ('90, p. 445) finds that upon Ciliata (Carchesium) a 0.1% solution slightly accelerates at first the action of the cilia, diminishes the rate of succession of the phases of the contractile vacuole, and leaves the protoplasm permanently more or less paralyzed.



are successively less poisonous. (Loew, '93, p. 46.)



per day being withstood by the human organism, with the replacement of the H atoms by OH the substance becomes more poisonous in direct proportion to the number of H atoms thus replaced. (Loew, '87, p. 440.) Thus there follow in order of poisonousness:—



Phenol (or carbolic acid) and its derivatives attack unstable substances, especially aldehydes, forming insoluble products.

*Phenol* itself produces in the higher animals a paralysis of the nerve centres. Algae die in a 1% solution after 20 to 30

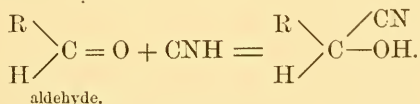
minutes; in a 0.1% solution after 3 days. Infusoria die quickly in a 1% solution. *Ascaris* lives only 3 hours in a 0.5% solution.

*Resorcin* (0.5 g.) is hypnotic to man; 0.085 g. per kg. is fatal to dogs. Its isomer — pyrocatechin — is more active. 0.1% of it in spring water kills diatoms and Infusoria after a few minutes, and filamentous algæ in a few hours; while, with resorcin, Infusoria, diatoms, and green algæ live several hours — even as long as 18 hours.

By replacing one of the H atoms of phenol by COOH (or carboxyl), thus producing salicylic acid, the poisonous qualities are reduced.

*Hydrocyanic Acid*, CNH. — The action of this substance is peculiar in that, acting on the central nervous system, it is in small quantities a more violent poison for Vertebrates than for Invertebrates. Hydrocyanic acid acts upon aldehydes — in dilute solutions upon the most unstable compounds only; in stronger concentration upon all aldehydes. Its peculiar working may be hypothetically explained by assuming the aldehydes of the ganglion cells to be more unstable than those of other cells, so that traces of CNH which do not injure other cells destroy quickly the nerve cells. (LOEW.)

The action may be shown by the equation, —

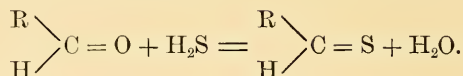


The degree and variety of its action may be inferred from the following data, taken from LOEW ('93): Infusoria die quickly in a 0.1% solution, but *Ascaris* resists a 3% solution for 75 minutes. The resistance of the hedgehog to CNH is remarkable; five times the dose which killed, in 4 minutes, a cat weighing 2 kg. produced in the hedgehog only a slight sickness. A myriapod (*Fontaria*) excretes CNH when irritated. Certain salts of CNH act as poisons; *e.g.* (CN)<sub>2</sub>Hg, Na<sub>2</sub>Fe(CN)<sub>5</sub>NO.

*Hydric sulphide* acts as a poison either by deoxidizing the plasma,

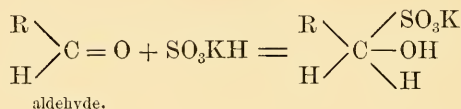


or by acting on the aldehydes,



It acts rather energetically upon algæ and Infusoria. In Vertebrates, the central nervous system is attacked and the oxy-hæmoglobin of the blood is altered.

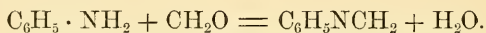
*Sulphurous oxide* ( $\text{SO}_2$ ) attacks members of the aldehyde group, —



0.1% kills lower fungi in a few minutes, 0.01% in a few hours.

*Selenous oxide* ( $\text{SeO}_2$ ), which acts chemically much like  $\text{SO}_2$  and has a much greater molecular weight (64 : 111), acts less energetically as a poison. A 0.1% solution kills *Spirogyra* and *Zygnema* in 3 hours, while 0.01% is scarcely injurious. Tellurous oxide ( $\text{TeO}_2$ , mol. wt. = 157) is non-poisonous, although chemically closely allied to the two preceding. (BOKORNY, '93.)

*Aldehydes.* — The poisonous action of these substances derived from oxidation of alcohol is dependent upon their instability. So we find that an aldehyde, which, like grape sugar, is fairly stable, is likewise non-poisonous; while formaldehyde, which is very unstable and active, is correspondingly poisonous. Aldehydes attack especially the unstable amides, affording nitrogenous compounds; *e.g.* —



Now, even in passive albumens, part of the N is in the form of amido-groups; for, in treating with nitric acid, much nitrogen is set free, which would not occur were all of the N secondarily or tertiarily bound up. (LOEW, '93, p. 58.) Hence the poisonousness of aldehydes for living albumens.

*Formaldehyde.* — This substance ( $\text{H} - \text{CH} : \text{O}$ ) acts upon propeptones and upon albumen, affording compounds which are not readily soluble. An aqueous solution of —

0.01% is fatal to bacteria.

0.05% kills worms, molluscs, and isopods in 2 hours. (LOEW, '88, p. 40.)

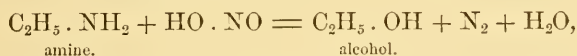
1.00% kills Spirogyra very quickly. (COHN, '94, p. 5.)

A weak solution seems to act anæsthetically upon Noctiluca. (MASSART, '93, p. 65.)

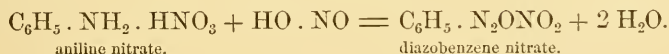
When various radicals are substituted in  $H - CH : O$ , the substance acts more like a catalytic poison. Thus, ethylaldehyde ( $CH_3 - CHO$ ) is anæsthetic; and paraldehyde ( $(CH_3 - CHO)_3$ ) kills algæ in a solution of 0.002% in 24 hours and causes protoplasm to become immobile either (leucocytes) after momentary stimulation (DEMOOR, '94, p. 218) or (Noctiluca) at once (MASSART, '93, p. 66).

Several derivatives of ethylaldehyde are poisonous. LOEW ('93, p. 60) has shown that in a 0.1% solution of the neutral sulphate of  $NH_2 - CH_2 - CH : (OC_2H_5)_2$ , amidoacetal, Infusoria, and diatoms die within 15 hours, and, somewhat later, filamentous algæ.

*Nitrous acid*, as is well known, produces, even in great dilution, OH-compounds from amido-compounds ( $R - NH_2$ ); or else, under certain conditions, especially with aromatic amido-compounds, diazo-compounds result; *e.g.* —



and



Thus a solution of 0.001% of free nitrous acid is poisonous to algæ, and more so than nitric acid. The lower fungi are also very sensitive to nitrous acid. Its action as an *acid* is weak, so that its salts are set free in the presence of even the weaker organic acids. On this account, even neutral nitrates kill such plants (some algæ, *e.g.* Spirogyra) as have an acid cell-sap.

8. **Sodic Fluoride.** — The poisonous action of the fluorides of the light metals, and especially sodium, has not hitherto been explained. A 0.2% solution of NaF kills various algæ (Oscillaria, Cladophora, Edogonium, diatoms) within 24 hours,

producing a change in size of the nucleus. (LOEW, '92.) A 1% solution kills the nerves of a frog in 2 hours.

9. **Special Poisons.** — *Toxic Protein Compounds.* Little need be said here concerning the recent discoveries of poisonous albuminoids excreted by disease-producing bacteria, or of those secreted by the parasitized body (alexines). Similar compounds, highly poisonous to Vertebrates, have been extracted from the seeds of some Phanerogams, *e.g.* ricin, from the seeds of *Ricinus communis* (castor-oil bean); abrin, from the seeds of the leguminous *Abrus precatorius*, L.; and phallin, from the toadstool *Agaricus phalloides*, Fr. Finally, in this group may be placed a large number of protein substances derived from animals, which are more or less poisonous to a greater or smaller number of kinds of protoplasm. The poison of the rattlesnake (*Crotalus*) and of the cobra (*Naja*) is fatal to Vertebrates in small, hypodermically injected, doses. Hydra, Turbellaria, Rotifera, and Crustacea are also affected by it; but Infusoria and Flagellata are apparently unaffected. (HEIDENSCHILD, '86, p. 330.)

It is important that, according to the experiments of several investigators, among the earlier of whom may be mentioned DAREMBERG ('91) and BUCHNER ('92), the various species of Vertebrates possess protein substances in their blood serum which are to a certain extent injurious to other species, since the blood serum of any one species will destroy the red and white blood corpuscles of another. The poisonous action of these animal protein substances seems to be due to their unstable character, whereby they easily form unions with the unstable groups of the protoplasm, frequently producing thereby violent poisons which work as substitution poisons. (LOEW, '93, pp. 81-84.)

*Alkaloids.* — These basic, nitrogenous compounds have, for the most part, very complex molecules, so that their structure has, in many cases, not been determined. Consequently the nature of their chemical action upon protoplasm is, in general, unknown.

LOEW suggests ('93, p. 85) the following theory of action of alkaloids. The bases unite with the active protein substances of the cell, and thereby introduce a disturbance of equilibrium



in the plasma body — a disturbance which is especially manifest in their action upon the protoplasm of ganglion cells. The capacity of this union is influenced by various factors: by the configuration and degree of dilution of the poison; by the degree of instability of the kind of protoplasm acted upon; by the configuration of the molecule of the active protein substance in the cells; and by the specific (micellar) structure of the plasma body. That organic bases can unite with active albumen is known from observations upon plant cells containing stored-up active protein substances. If the configuration of the albumen molecule and the general texture of the protoplasm favor the attack by the base, a disturbance of the equilibrium of the protoplasm will result, even in considerable dilution of the poison.

The alkaloids affect chiefly the nervous tissue in the higher animals, producing, in some cases, paralysis; in others, increased activity. Thus, curarin is paralytic in its action, while the closely allied strychnin is (in dilute solutions) stimulating upon nerve cells. Since action is almost confined to nerve tissue, additional evidence is afforded of the extraordinary instability of the nervous protoplasm.

The dissimilar effect of an alkaloid upon the different substances constituting nerve protoplasm gives an idea of the complexity of the latter. Thus, an alkaloid may stimulate nerves with certain functions to increased activity, and may reduce nerves in the same body, having other functions, to depression and paralysis; *e.g.* nicotin excites sensory nerves, and depresses the activity of the cardio-motor nerves.

It is important that many of these alkaloids act also upon Protozoa and the lowest plants, in which nervous substance is still undifferentiated. Other of these alkaloids, however, do not act upon Protista.

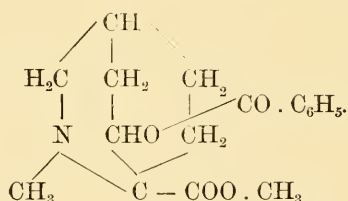
We will now proceed to an examination of the action of the principal vegetable alkaloids, arranged according to the systematic position of the plants from which they are obtained.

*Nicotin.* — The effect produced by nicotin is directly proportional to the differentiation of nervous substance; thus, it is almost inoperative on Protozoa and Actinia. Hydra is not very sensitive, 0.5% being a fatal dose. A solution of 0.05%

causes Medusæ to become quiet in 30 minutes, and is fatal to the earthworm in a few hours; Echinoderms are paralyzed by a 0.05% solution in 30 minutes; Palæmon (after temporary stimulation) is paralyzed by a 0.01% solution in 30 minutes; Sepiola is killed by 0.005% in less than a minute. (GREENWOOD, '90.)

*Veratrin*, *Atropin*, and *Cocaine* act upon Vertebrates so as to excite the central nervous system at first, and then to paralyze it. All act, however, as poisons upon undifferentiated protoplasm (Protozoa). Thus, ROSSBACH ('72) found that when Ciliata were subjected to veratrin chloride and to atropin sulphate, a peculiar rotary movement took place about one end as a fixed axis. Then imbibition of water with great vacuolation of the protoplasm occurred. Later, the contractile vacuole fails to contract, and protoplasmic movements cease a few seconds after. (Cf. KÜHNE, '64, pp. 47, 65, 100.)

*Cocaine* is apparently a benzol derivative, closely related, chemically, to atropin. Its formula is thus given:—



Its action upon Protista has been studied by CHARPENTIER ('85), ADDUCO ('90), SCHÜRMEYER ('90, pp. 438-448), ALBERTONI ('91, p. 318), DANILEWSKI ('92), and MASSART ('93, p. 66); upon sexual cells, by O. and R. HERTWIG ('87, p. 159) and ALBERTONI ('91, p. 309); and upon tissue cells by ALBERTONI. The result has been to show that cocaine first stimulates for a very short time to excessive activity, and then stupefies and paralyzes. With the paralysis, a strong vacuolation of the protoplasm occurs, since the excretory function of the contractile vacuole is inhibited (SCHÜRMEYER, '90, p. 439). Cocaine acts similarly upon the nerve centres and muscles of the more differentiated animals.

*Morphin* acts less violently upon the nervous tissue of Vertebrates. It has a very weak action upon Protista.

*Strychnin*, chemically considered, is an alkaloid with the formula:  $C_{21}H_{22}N_2O_2$ ; specific gravity, 1.359 at 18°; soluble to about 0.025% in water at 14.5°; has a very bitter taste. The nitrate is generally employed. The action of strychnin upon Protista is known through the studies of MAX SCHULTZE ('63, p. 32), BINZ ('67, pp. 384-389), ROSSBACH ('72, pp. 52-54), and SCHÜRMEYER ('90, pp. 423-433). KRUKENBERG ('80) has studied its effects upon higher Invertebrates. Its action upon sexual cells has been studied by the brothers HERTWIG ('87, pp. 153-156, 164).

Although not fatal to bacteria and only in strong solutions fatal to the large fungi, strychnin is a nearly universal protoplasmic poison. It kills the protoplasm of the *Drosera* tentacles and hinders the development of peas, corn, and lupines. The amount of strychnin that Protozoa can withstand has been variously stated, while all authors admit considerable individual variation in this respect. Probably Protozoa cannot ordinarily resist a saturated solution for one minute. ROSSBACH ('72, p. 52) found that no infusorian of his cultures survived a "0.1% solution" long enough to be placed under the microscope. A 0.02% or 0.01% solution can be withstood for a few minutes (0.01% solution was withstood for 5 minutes, SCHÜRMEYER). As for the weakest solution that will kill, SCHÜRMEYER found that a *Paramecium* resisted for only 15 to 20 minutes so weak a solution as 0.0005%, while ROSSBACH found *Stylonychia* little affected by a 0.0055% solution. The spermatozoa of Echinoids, according to the HERTWIGS ('87, p. 164), so resist a 0.01% solution that after 180 minutes the movement is only somewhat retarded. Echinoid eggs are injured in a few minutes by 0.005%.

The injurious action of strychnin on Protozoa varies in the different groups, the resistance capacity increasing, in general, with the height of systematic position of the group. The first effect in Ciliata is an increased activity of the cilia; if cirri are present, these strike more powerfully; locomotion is abnormally rapid; but the movements lack coördination, and a rotation takes place about the axis of progression. Next,

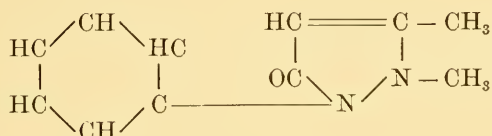
the movements become so disorganized that locomotion is impossible, despite accelerated cilia-motion. Finally, the movements suddenly cease, death intervening. (SCHÜRMEYER, '90, pp. 423-426.) The process of excretion seems to be especially affected. Immediately after the addition of a 0.02% solution, the contractile vesicle increases to from 4 to 10 times its normal diameter, and loses its spheroidal form. In a slightly greater dilution, 0.014%, the contractile vesicle momentarily constricts, but in diastole gains twice its normal diameter, and the time between phases of contraction is greatly increased. Thus, the normal rate of contraction for *Euplotes* at 15° C. is between 30 and 35 seconds; but in a 0.014% solution of strychnin this is diminished to 1 in 500 seconds. Frequently, several vacuoles are formed, and, eventually, the whole body becomes greatly vacuolated, and death intervenes. (ROSSBACH, '72, pp. 52, 53.) Many higher organisms are not very sensitive to strychnin. Thus, *Ascaridæ* have a considerable resistance, which SCHRÖDER ('85, p. 307) ascribes to their not opening their mouths in the solution, the poison being thus obliged to pass through the skin. So, too, snails were found by KRUKENBERG ('80, p. 100) to be very resistant to strychnin.

*Curare*, or *urare*, is an alkaloid, derived from *Strichnos* species. The commercial substance is very variable in composition. NIKOLSKI and DOGIEL ('90) have studied the effects of this drug upon various organisms. Upon adding a few drops of a 0.8% solution of curare to water containing an amoeba, the first effect is a shrinking towards a spherical form and a cessation of all movements. Subsequent washing in water ultimately restores the normal movements. As is well known, it paralyzes also the protoplasm of the nerve endings.

*Quinine*, or chinin ( $C_{20}H_{24}N_2O_2$ ). — The "sulphate," which is first produced in the process of extraction, is commonly employed. This is, moreover, more soluble than the pure alkaloid, 1 part dissolving at 9.5° in 788 parts of water. Investigations on the action of this poison upon protoplasm have been made especially by BINZ in a series of papers beginning with '67; by ROSSBACH ('72) on Protozoa; by TEN BOSCH ('80) on leucocytes; by O. and R. HERTWIG ('87) on sexual cells; and by KRUKENBERG ('80) on higher Invertebrata.

Amœba, Actinophrys, and various Infusoria are killed by a 0.1% solution in a few minutes, and leucocytes and eggs of Echinoids are paralyzed even by a 0.005% solution. Its action is thus more powerful than that of strychnin. The protoplasm at first contracts, then gradually dissolves and streams away. Upon the higher animals, quinine so acts as to paralyze the central nervous tissue (in Mollusca, KRUKENBERG, '80, p. 10), and it affects the cerebrum and heart ganglia of mammals.

*Antipyrin*, or phenyldimethylpyrazolon, is an alkaloid derived from and belonging clearly to the benzol type, in which one atom of H is replaced by a complex atom-group, as may be seen from the formula —



The effect of this agent upon Protozoa has been studied by SCHÜRMEYER ('90, pp. 434-437) and MASSART ('93, p. 64). A solution of 0.1% acting for 80 minutes, caused *Oxytricha* at first to move more rapidly, but eventually to become transformed into a shapeless mass, whose protoplasm disintegrates. Acting upon *Noctiluca*, a 0.25% solution causes a bright glimmer immediately after applying, followed by darkness again. Thus there is here a momentary hyperesthesia.

## § 2. ACCLIMATIZATION TO CHEMICAL AGENTS

It is clear that the protoplasm of different organisms is dissimilar. We see this in the different reactions to the same chemical agent. Not only is the reaction of the various species unlike, but individuals of the same species from different localities differ widely. (Cf. LOEW, '85.)

We are, naturally, most familiar with this phenomenon in the case of man. Thus, the common North American poison ivy (*Rhus toxicodendron*) produces, in some persons, extensive inflammation in parts which have come even indirectly in con-

tact with it; while, by other persons, it may be taken into the mouth with impunity.

The phenomenon shown by man is found in other animals also. Thus, among Invertebrates, although few bacteria can resist 1%  $\text{Na}_2\text{CO}_3$ , and even the extremely resistant *Ascaris* lives only 5 to 6 hours in a 5.8% solution of this salt, LOEW ('77, p. 137) has found in Owen's Lake, California (an alkaline water containing among other things 2.5%  $\text{Na}_2\text{CO}_3$ ), numerous living Infusoria, Copepoda, larvæ of Ephydra, and molds. Again, the vinegar eel, *Rhabditis aceti*, lives in a 4% solution of acetic acid, although this strength is usually fatal; *e.g.* a 0.23% solution of acetic acid kills the tentacles of *Drosera*. (DARWIN, '75, p. 191.)

What is true of the whole organism is true also of its parts. The gland cells of some marine Gasteropoda (*Dolium*, *Cassis*, *Tritonium*, *Natica heros*) secrete  $\text{H}_2\text{SO}_4$  of a strength (2% to 3%) which is fatal to most protoplasm; the myriapod *Fontaria* excretes, when irritated, the extremely poisonous CHN; and, according to LOEW ('87, p. 438), the plant *Oxalis* produces potassic oxalate, which is a violent poison to most protoplasm.

One general law of high resistance is worthy of notice: an organism which produces an albuminoid poison is strongly resistant to that poison. Thus, FAYRER ('74) has shown that venomous serpents are not destroyed by the secretion of their poison glands when it is injected into them; and BOURNE ('87) has shown that scorpions are not injured by their own venom.

An explanation of the facts of varied resistance capacity is first gained through experiment. We all know that, among men, a high resistance capacity to a poison may be acquired by taking a small quantity of it at frequent intervals. Thus, users of tobacco, alcohol, and various alkaloids become, in time, capable of taking, without apparent injury, quantities which would at first have proved fatal. Arsenic eaters may eventually swallow without injury four times the ordinarily lethal dose, *i.e.* as much as 0.4 gramme. (BINZ and SCHULZ, '79.)

Results similar to those observed in man have been obtained by experiment upon other animals. Thus, SEWALL ('87) inoculated a pigeon hypodermically with sub-lethal doses of

rattlesnake poison (*Crotalophorus tergeminus*). While no unacclimatized pigeon could resist 1 drop of a 6.8% solution of venom in glycerine, pigeons inoculated with at first weak, then gradually increasing solutions, came at last (after 178 days) to resist 4 drops of the glycerine venom mixture. Likewise KANTHACK ('92) succeeded in acclimatizing two rabbits and a hen to serpent's venom.

Very similar are the experiments of EHRlich ('91). This investigator fed white mice (which are killed by  $\frac{1}{20}$  of their weight of a 0.0005% solution of ricin, hypodermically injected) upon food cakes soaked in a weak solution of the poison. After feeding them for a varying length of time upon constantly increasing solutions, he determined the maximum solution which, hypodermically injected, they would withstand. If we call the maximum solution which the unacclimatized organism will withstand our unit of immunity, we can express the degree of immunity of the acclimatized organisms by the strength of solution (expressed in terms of that unit) which they can resist.

The following table, taken from EHRlich's paper, shows the gradual increase of immunity as a result of feeding on the poison : —

TABLE IV

NO. OF EXPERIMENT DAY.	STRENGTH OF LAST DOSE GIVEN, IN MG.	NUMBER OF INDIVIDUALS EXPERIMENTED ON.	MAXIMUM INJECTED SOLUTION BORNE, %'S.	DEGREE OF IMMUNITY.
IV . . . . .	4	8	} Die in 0.0005	1
V. . . . .	5	16		0.0007
VI . . . . .	6	23	0.0066	13.3
VII . . . . .	7	5	0.005	10
VIII . . . . .	8	18	0.01	20
X . . . . .	12	9	0.02	40
XII . . . . .	20	3	0.033	66.6
XV . . . . .	50	1	0.05	100
XVIII . . . . .	80	4	0.1	200
XXI . . . . .	80	1	0.2	400

Thus after the first 4 or 5 days the immunity rapidly increased; so that, while the solution of  $\frac{1}{200000}$  kills normal

mice, those acclimatized during 21 days resist  $\frac{1}{1000}$  to  $\frac{1}{500}$ , occasionally  $\frac{1}{250}$ , corresponding to a grade of immunity of 200 to 800.

By fundamentally similar procedures CALMETTE ('94) has rendered rabbits immune to the action of strong doses of the venom of *Naja tripudians* (cobra) and of *Pelias berus*.

Still more recently MARMIER ('95) has isolated a toxin produced by anthrax bacteria reared in a peptone-glycerine solution. Inoculated into an animal sensitive to anthrax, this toxin produces, in certain doses, death by cachexy. By employing suitably *graduated* doses, however, one can obtain immunity of the organism to anthrax, as one does to the venom of serpents.

Some attempts to produce acclimatization in lower organisms have been made by Dr. H. V. NEAL and myself. *Stentor* was employed as the object of experimentation. We reared two lots of *Stentors* under similar conditions except that Lot 1 was cultivated in water and Lot 2 in 0.00005%  $\text{HgCl}_2$ . After the lapse of two days both were put into a killing solution of 0.001%  $\text{HgCl}_2$ , and the second lot was found to survive longer than the first. The mean resistance period to the killing solution of the lot reared in water was 83 seconds; of that reared in 0.00005% corrosive sublimate, 304 seconds. Similar results were obtained in other experiments. In Fig. 1 a curve is given showing the relation between strength of culture solution and period of resistance. From this curve, based upon 132 determinations, it appears that *the resistance period varies directly with the strength of the solution in which the protoplasm has been cultivated.*

This law holds good, however, only within certain limits. If the culture solution is too strong, above 0.0001%, the organism will be weakened by it so that it cannot resist the killing solution so long as those reared in water can.

A similar effect of heightened resistance to quinine is obtained by cultivating organisms in quinine.

Experiment shows that a slight increase of the resistance period follows subjection to the culture for one hour only; and that the degree of acclimatization varies directly as the time of subjection.



The facts obtained by us clearly indicated, then, that, without selection,—for no deaths occurred in our culture solutions,—the protoplasm may become modified merely by subjection to the poison, so as to gain an increased resistance to it. Hence the acclimatizations that we find in nature need not have been brought about by natural selection—*must* have occurred, indeed, even without selection, if the organisms had been gradually subjected to their environment.

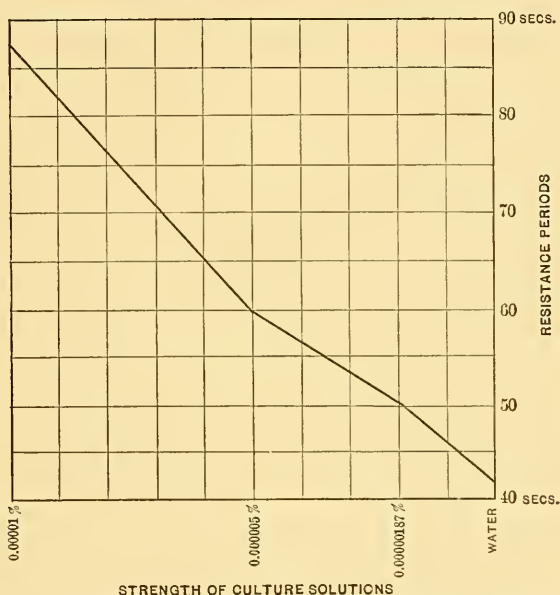


FIG. 1. — Curve of resistance periods to a 0.00125% solution of  $\text{HgCl}_2$  of Stentors reared in various solutions of  $\text{HgCl}_2$  during 20 to 96 hours.

We did not determine for how long a time the acclimatized Protozoa retained their heightened resistance capacity. The only data we have upon the subject of persistence of acclimatization is derived from studies on Vertebrates.

Thus it is the familiar experience of arsenic eaters that, after they have broken off their habit, the body does not quickly return to a normal condition. Even after a considerable period of self-denial the taking of large doses may be recommenced—must be recommenced, indeed, or illness sets in.

EHRlich ('91) has studied experimentally the phenomenon

of acclimatization to ricin. Mice which had gained an immunity of over 200, and were then kept for 6.5 months on normal food, had still a resistance, although not precisely determined, certainly far above 50.

It is an important question: Is an organism acclimated to one poison thereby rendered more resistant to poisons in general, or only to the specific poison to which it has been acclimated? EHRlich found that mice acclimated to ricin were just as sensitive to abrin as the normal animals, and the same is true, *mutatis mutandis*, for mice which resist abrin.

Concerning the changes in the protoplasm brought about by acclimatization little is known.

EHRlich ('91) and CALMETTE ('94) have shown that in the blood of the immunized animal a substance, antitoxic to the specific substance employed, is produced, and this apparently prevents the action of the strong poison by transforming its molecules. The antitoxic substance is of such a nature that when blood containing it (from an acclimatized animal) is injected into an unacclimatized one, the latter becomes immune to the poison.

For Protista another hypothesis is admissible; namely, that the weak solution of the poison, which is used in acclimatization, *gradually* destroys those compounds upon which the strong solution would have acted suddenly and, therefore, fatally. The gradual destruction is not fatal because of its slowness. At the same time it prevents the violent action of the strong poison, since it leaves it nothing to be acted upon.

The parallelism between the results of experiments upon acclimatization to poisons and those upon immunization through vaccination, leads to the suspicion that, at bottom, the two processes are closely akin.

### § 3. CHEMOTAXIS

ENGELMANN ('81) seems to have been the first to show that the direction of locomotion of simple protoplasmic masses is determinable by chemical agents in the environment. He found that *Bacterium termo* is thus acted upon by oxygen which is not uniformly distributed. Like many Infusoria, these bacteria

gather at the margin of the cover-glass, where oxygen is more abundant than elsewhere. If oxidized blood is introduced under the cover-glass, they move toward it, but not toward blood containing much  $\text{CO}_2$  in place of oxygen. If green algæ are introduced, the bacteria move towards them so long as they, under the influence of sunlight, are producing oxygen. In the dark the algæ have no effect.

During the decade and a half which have elapsed since ENGELMANN'S first paper appeared, chemotactic phenomena have been observed among nearly all kinds of motile organisms and with reference to the most diverse kinds of chemical substances. ENGELMANN ('82) has studied the chemotactic movements of diatoms; STAHL ('84), of Myxomycetes; PFEFFER ('84, '88), of plant spermatozoids, zoöspores, Flagellata, Infusoria, and bacteria; ADERHOLD ('88), of *Euglena viridis*; VERWORN ('89, p. 107), of *Cryptomonas*; STANGE ('90), of zoöspores and Myxomycetes; and MASSART ('91), of *Spirillum*, *Heteromita*, and Ciliata.

Within the last five years a voluminous literature has grown up on the medical side relating to chemotaxis in leucocytes and pathogenic bacteria. Into this literature we cannot penetrate deeply, but refer to some of the principal papers: LEBER, '88; BUCHNER, '91; ROEMER, '92; METSCHNIKOFF, '92.

It thus appears that chemotactic phenomena show themselves among all the groups of lower motile organisms: Rhizopoda (Myxomycetes), Flagellata, Ciliata, bacteria, diatoms, zoöspores, and spermatozooids. It can hardly be questioned that the phenomena shown by these organisms are of the same order as those seen in Metazoa—in those ants which LUBBOCK ('84, p. 233) has shown to move from chemical agents (essence of cloves, lavender water, and other scented stuffs), saturating a camel's-hair brush placed about  $\frac{1}{4}$  inch above their path; and in the larvæ of flies with which LOEB ('90, p. 79) has experimented. LOEB found that these crept towards a piece of flesh brought nearer to them than the distance of 1.5 cm. Even just hatched larvæ (which had therefore never been stimulated by meat) reacted in this way. Not meat only, but a trace of meat juice on glass attracted the larvæ strongly. While decaying flesh and cheese allure, neither fat, asafœtida, nor ammonia do so.

Returning now to the simple organisms, let us consider the kinds of chemical substances which incite to a response.

*Oxygen* is for almost all organisms a means of attraction. Various methods of demonstrating this have been used. Thus STANGE ('90, p. 139) filled capillary tubes with pure oxygen, under an air-pump, and brought them to the water containing zoöspores, which then penetrated into them.

The aggregation of zoöspores and bacteria to the edges of the cover-glass, to the open end of a capillary tube (ADERHOLD,

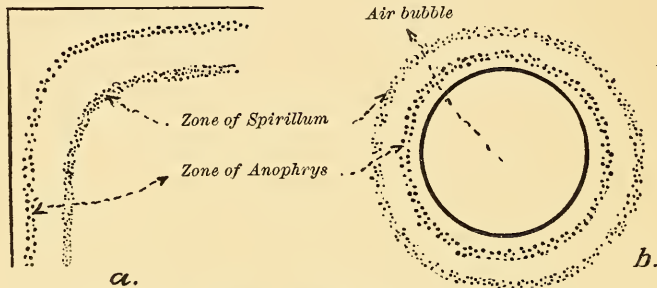


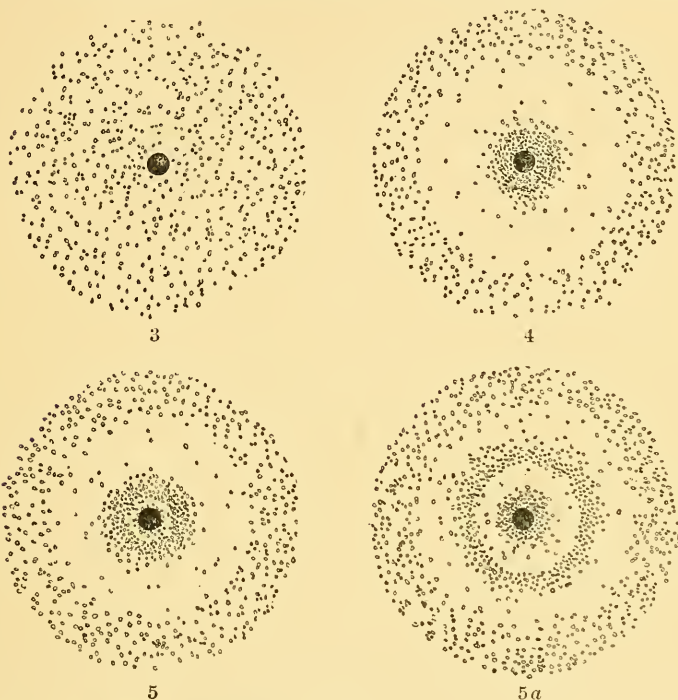
FIG. 2. — *a.* Corner of the glass slip covering a drop of liquid containing *Spirillum* and *Anophrys*, showing their aggregation with reference to the aerated bounding film of the drop. *b.* An air-bubble in the drop, showing aggregation of the organisms about it. (From MASSART, '91.)

'88, p. 314), or to an enclosed air-bubble, are well-known phenomena. (Cf. MASSART, '91, p. 159; VERWORN, '89, p. 107; and see Fig. 2.)

ENGELMANN ('94) has employed a still more refined method of studying attraction towards oxygen. A drop of foul water is put on a glass slide with an alga cell in the centre, and is covered by a cover-glass whose edges are hermetically sealed by vaseline. The bacteria are uniformly distributed in the water, moving in a lively manner, since they gain oxygen everywhere. If the slide thus prepared is kept in the dark, the oxygen is gradually consumed and the bacteria become quiescent, showing no distribution with reference to the central chlorophyllaceous body (Fig. 3).

If the slide is now exposed to the light, oxygen is produced by the alga and a regular distribution of the bacteria in two distinct regions — in a mass around the central alga, and in a

peripheral zone — is apparent (Fig. 4). The peripheral zone contains bacteria which are beyond the tactic action of the oxygen. The central mass of bacteria have emigrated from what is now a clear ring between the centre and the peripheral zone. If the light be temporarily cut off, the central bacteria



FIGS. 3-5*a*. — Bacteria surrounding an algal cell. Fig. 2 shows the uniform distribution of the bacteria when the drop of water is kept in the dark. Fig. 3 shows the aggregation of the bacteria towards the algal cell when this has emitted oxygen rapidly in the strong light for two minutes. Fig. 4 shows the same preparation shortly after the light has been cut off. Fig. 5 shows the same preparation when a fainter light is now permitted to fall upon the green cell. Magnified about 170 diameters. (From ENGELMANN, '94.)

begin to disperse (Fig. 5). If, a minute after, a *dim* light be let through, the radius of its activity will be relatively small, so that a central aggregation will be found and also an inner peripheral zone, comprising those dispersing central bacteria which are not affected by the small oxygen tension resulting from the dim light (Fig. 5*a*).

*Inorganic Salts.*—PFEFFER\* ('88, p. 601) tried various salts of potassium; viz. chloride, phosphates, nitrate, sulphate, carbonates, chlorate, ferrocyanide, and tartrate, and found that all attracted various bacteria (*B. termo*, *Spirillum undula*), and the flagellate *Bodo saltans* with greater or less strength, when the solution in the capillary tube contained 0.1% K. So, likewise, various salts of sodium, rubidium, caesium, lithium, ammonium, calcium, strontium, barium, magnesium, especially the chlorides, were employed. All of these solutions at a concentration of 0.5% exhibited a marked attractive influence upon *Bacterium termo*; a weaker one, upon the two other species.

STANGE ('90) experimented with the action of various phosphates upon zoöspores of a *Saprolegnia* belonging to the ferax group of DE BARY.† Sodie, ammonic, lithic phosphate, calcic phosphate held in solution by CO<sub>2</sub>, as well as phosphoric acid were employed and found to act attractively. Other salts, KNO<sub>3</sub>, K<sub>2</sub>SO<sub>4</sub>, KCl, HKCO<sub>3</sub>, BaClO<sub>3</sub>, SrCO<sub>3</sub>, MgSO<sub>4</sub>, had either a negative or indifferent action upon the zoöspores.

The attractive action of the phosphates is correlated with the fact that phosphates are abundant in the muscles of insects. The following table shows the effect of the different strengths of solutions of four phosphates upon zoöspores of *Saprolegnia*. In this table, constructed from STANGE, the symbol *r* indicates repulsion; 0, no action; *a*, attraction; *a*<sub>1</sub> indicates a slight attraction; *a*<sub>2</sub>, a strong attraction; *a*<sub>2</sub>*r*<sub>1</sub>, an attraction which is partly balanced by a repulsion due to density, so that the

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\* The method employed by PFEFFER in his experiments was as follows: Glass capillary tubes with a lumen of from 0.03 to 0.14 mm. diameter and a length of 7 to 12 mm., and sealed by fusion at one end were employed. To fill the capillary tube, it was placed in a watch-glass containing the experiment solution, and the whole was placed in a vessel from which air was pumped. Under the diminished atmospheric pressure, 2 to 4 mm. of fluid entered the capillary tube; the rest of the tube contained air, which kept the fluid oxidized. After rinsing, the free end of the tube was plunged into the drop culture, whence the solution diffused out.

† The species were cultivated upon carcasses of flies thrown into glasses filled with bog water. After good colonies were obtained, the carcasses were washed, to rid of Infusoria. Such colonies may be employed to infect sterilized flies' legs placed in sterilized bog water, or they may be transferred directly to wounds in flies' legs.

organisms pass only into the first part of the tube ;  $a_3r_3$ , such a balancing of the opposing forces that the organisms stand before the mouth of the capillary tube.

STRENGTH OF SOLUTION, % 's.	SODIC DIPHOS- PHATE ( $\text{HNa}_2\text{PO}_4$ ).	POTASSIC MONO- PHOSPHATE ( $\text{HKPO}_4$ ).	AMMONIUM PHOS- PHATE ( $\text{H}_2\text{NH}_4\text{PO}_4$ ?).	PHOSPHORIC ACID ( $\text{H}_3\text{PO}_4$ ).
0.8 to 0.4 . . . .	$a_3r_2$	$a_3r_2$	$a_3r_3$	$r$
0.4 to 0.08 . . .	$a_2r_1$	$a_2r_1$		
0.08 to 0.04 . . .	$a_2$	$a_2$	$a_2$	$a_1r_2$
0.01 to 0.02 . . .	0	$a_1$	$a_1$	$a_1r_1$
0.02 to 0.008 . . .		0	0	$a_2$
0.008 to 0.004 . . .				$a_1$
0.004 to 0.002 . . .				0

It will be noticed that the various substances produce different effects in the same strength of solution ; and it is interesting to observe (a point to which further reference will be made) that the strength of solution required to produce a given response is roughly proportional to the molecular weight of the substance employed.

*Inorganic acids and hydrides* seem, in general, to act repulsively, but phosphoric acid is an important exception to this rule. DEWITZ ('85, pp. 222, 223) states that mammalian spermatozoa are attracted by KHO.

*Organic Compounds.*—*Alcohol*, in grades between 10% and 1%, acts repulsively towards bacteria. *Glycerine* is neutral to the same organisms and to zoöspores of Saprolegnia. (STANGE, '90.) The *sugars*, etc., dextrose, milk sugar, dextrin, act attractively upon *Bacterium termo* in 10% or weaker solutions. Many *organic acids* are among the most attractive reagents. It was with malic acid that PFEFFER ('84) tried his earlier fundamental experiments upon the spermatozoids of ferns. The attraction exerted is very great, so that a capillary tube of 0.1 to 0.14 mm. calibre, containing a 0.05% solution of malic acid, attracts from a drop of water full of spermatozoids at the rate of 100 individuals in one hour. Even a 0.001% solution acts chemotactically. Now, malic acid is of very wide distribution among plants, and it occurs in the fern prothalli upon which the sexual

organs arise, so that it seems probable that it occurs in the mouth of the archigonium, and that by its presence spermatozooids are attracted towards the egg cell.

STANGE ('90, p. 155) has experimented much more fully with the action of organic acids upon zoöspores of *Saprolegnia* and upon myxamœbæ. To the former, acetic acid (0.01%) and tartaric acid (0.0125%) act attractively. Upon the latter, still other acids were tried; butyric, lactic, and valeric acids cause response in concentrations between 0.2% and 4%; malic acid, between 0.5% and 4%. Other attracting acids are: propionic, citric, tartaric, and tannic. Acetic acid repels the amœbæ of *Æthaliu*m, its repellent action being about equal to the attractive action of an equal amount of butyric acid.

*Nitrogenous Compounds.*—*Urea*, *asparagin*, kreatin, taurin, hypozanthin, carnin, and *peptone* have been found by PFEFFER ('88) to exert an attractive influence, especially in the case of the reagents *italicized*.

*Benzol Derivatives.*—PFEFFER found that sodium salicylate and (commercial) sulphate of morphine are clearly attractive to *Bacterium termo* in 1% solutions.

From the foregoing list of organic compounds whose effect upon Protista has been tested by PFEFFER and STANGE, it appears that except alcohol and sometimes acetic acid, none acts repulsively, and that glycerine alone is neutral to all protoplasm. It is further true that we do not find here any strict relation between the chemotactic action of a substance and its advantage to the organism. Substances which have a nutritive value for the organism, such as glycerine has for bacteria, may be wholly neutral, while solutions which act fatally, like 1% sodic salicylate and 1% morphine, attract. In the same way, many of the organic salts which act attractively cannot be considered as of importance to the organism. On the other hand, as already pointed out, the attraction of most Protista to oxygen, of *Saprolegnia* zoöspores to phosphates, as well as the cases of attraction of bacteria (PFEFFER, '88, p. 605) and of fly larvæ (LOEB, '90, p. 79) to meat extract, and of *Myxomycetes* to bark extract (STAHL, '84; STANGE, '90), is advantageous. Chemotaxis is, therefore, in some cases, a response to the stimulus afforded by substances which can be employed by



the organism as food; under which circumstances it can be called "Trophotaxis."\* In other cases it is a response to chemical substances which have no significance as food, and have no other importance for the organism.

It is clear that we cannot assume that response to injurious substances is an adaptation which has been brought about by natural selection. If a response occurs in one case independent of the action of selection, we should hesitate to ascribe to this cause the origin of other, even favorable, responses.

*General Remarks on the Relation between Molecular Composition and Response.* — PFEFFER ('88, pp. 608-612) has pointed out that the capacity of any substance to stimulate cannot be inferred from its chemical constitution and relationships. Thus, the minimum concentration of milk sugar which will produce a response is 1%, while in the case of the closely allied grape sugar it is 10%; but in the very different kreatin it is also 1%. Also, the action of any chemical compound is determined not by the elements, such as C, H, O, which it contains, but by the entire molecule; in other words, the atomic composition is less important than the structure of the molecule or the arrangement of its atoms. Thus, malic acid and its compounds with neutral ammonium, sodium, barium, and calcium containing 0.001% parts of the acid, have an equal action upon the spermatozooids of ferns, which do not react to the diethylester of malic acid, even in strong solutions. (PFEFFER, '88, p. 655.) Again, nitrogenous organic compounds are, in general, more active than the non-nitrogenous ones; but this cannot be held to be due alone to the presence of N; for dextrin ( $C_6H_{10}O_5$ ) is nearly as active as the nitrogenous peptone, and, on the other hand, the nitrates of metals are not more active than their chlorides, while the ammonia salts are relatively weak.

*Relation between the Strength of the Stimulus and that of the Response.* — When we say that malic acid attracts spermatozooids we mean that under certain physical conditions of the water which we may call normal it does so. And under normal conditions of the water, it is only within certain limits

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\* STAHL ('84, p. 164) called the attraction of plasmodium of myxomycetes to bark extract "Trophotropism."

that malic acid attracts. The strengths of solutions which attract under such conditions lie between 0.001% and 10%. The weaker solution may be designated the minimum; the stronger, the maximum concentration which provokes a response. The minimum solution provoking response is also often called by the Germans the "Reizschwelle," or "stimulation threshold";\* the optimum, the "Reizhöhe," or "stimulation acme"; the range, the "Reizumfang."

The character of the responses observable at the two limits is very different; at the minimum, attraction is very feeble; thus, while a capillary tube containing 0.01% neutral sodic malate, plunged into water at 14°-20° C., swarming with spermatozoids, attracts 400 of them in 10 minutes, a 0.001% solution attracts only 10-25 individuals during the same time, and a 0.0008% exerts little attractive effect, the spermatozoids remaining undirected in their movements. At the maximum, on the contrary, repulsion is observed. The spermatozoids move from the mouth of the capillary tube. Between the two extremes lies the concentration of greatest attraction—the acme. As we pass from the acme towards the minimum, the attraction becomes less and less. As we pass towards the maximum, the attraction remains the same, or increases; but repelling influences are now at work, which eventually entirely counteract the attractive influences.

A satisfactory method of expressing quantitatively the facts just mentioned has not been invented. PFEFFER ('88, p. 599) has employed the nomenclature which we have used above (p. 36) —  $a_1$  to  $a_3$  being combined with  $r_1$  to  $r_3$  to indicate the coworking in varying proportions of attraction and repulsion. Using this nomenclature, we may illustrate the statements made in the last paragraph with examples taken from PFEFFER'S work: —

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\* The following substances at the solutions named produce the threshold attraction ( $a_1$ ) in Bodo saltans: KCl, 0.02%;  $K_3PO_4$ , 0.002%;  $KH_2PO_4$ , 0.0035%;  $KNO_3$ , 0.26%;  $K_2SO_4$ , 0.22%;  $KClO_3$ , 0.3%;  $K_4(CN)_6Fe$ , 0.235%;  $K_2 \cdot C_4H_4O_6$ , 0.02%; RbCl, 0.14%; LiCl, 0.6%;  $LiNO_3$ , 3%;  $NH_4Cl$ , 0.3%; neutral ammonium phosphate, 0.08%;  $SrCl_2$ , 0.2%;  $Sr(NO_3)_2$ , 0.4%;  $BaCl_2$ , 0.17%; dextrin, 0.1%; urea, 1%; asparagin, 0.1%; taurin, 1%; sarkin, 0.33%; pepton, 0.01%; meat extract, 0.01%.

GRADE OF SOLUTION.	RESPONSE OF	
	BACTERIUM TERMO.	SPIRILLUM.
9.53 % KCl = 5 % K	$a_3$	$a_3r_3$
1.906 % KCl = 1 % K	$a_3$	$a_3r_1$
0.191 % KCl = 0.1 % K	$a_3$	$a_3$
0.019 % KCl = 0.01 % K	$a_2$	$a_1$
0.0019 % KCl = 0.001 % K	$a_1$	0
	BODO SALTANS.	
3.48 % $\text{KH}_2\text{PO}_4$ = 1.0 % K	$a_3r_3$	
0.348 % $\text{KH}_2\text{PO}_4$ = 0.1 % K	$a_3r_2$	
0.035 % $\text{KH}_2\text{PO}_4$ = 0.01 % K	$a_2$	
0.0035 % $\text{KH}_2\text{PO}_4$ = 0.001 % K	$a_1$	
0.00067 % $\text{KH}_2\text{PO}_4$ = 0.0002 % K	0	

Compare also the table on p. 37.

For all reagents which exert an attractive influence there exists the maximum (repelling) and minimum (indifferent) limits referred to. In the case of reagents, which, like alcohol, repel bacteria at between 1% and 10%, there is doubtless an indifferent limit, but it is not necessary that there should be a degree of concentration at which attraction takes place. In the one case, then, the phenomena of indifference, attraction, repulsion, follow each other with increasing concentration; in the other case, only indifference and repulsion. The difference in action of the two cases is due, in part at least, to the fact that all solutions, independently of their chemical constitution, become repellent when they become concentrated enough. The repulsion, then, of high grades of chemical solutions is purely an osmotic phenomenon, and, as such, will come under discussion in the third chapter. It follows, also, from what has been said, that, in the case of those reagents which exert no attraction at any concentration, the acme and maximum coincide and lie at the saturation point of the solution.

Finally, we may discuss the third case in which the reagent acts indifferently, as glycerine does upon bacteria between 17% and 0.86%. It is clear, that if the density of the solution can

be rendered great enough, a repulsion due to osmosis must occur. If the substance is, however, only soluble slightly or miscible, it may be that repulsion will never occur. Whether or not attraction will occur before the repulsion point is reached will have to be determined experimentally for each reagent.

Thus the action of an untried substance upon any organism may be any one of three kinds: (1) It may be indifferent at all grades; (2) it may be indifferent at lower and repellent at higher grades; (3) it may be indifferent, attractive, and repellent at successive grades. Of two substances belonging to the second or third class, one may act upon an organism at a certain concentration with indifference, the other at the same concentration with repulsion. Likewise the same solution of a substance may attract one kind of protoplasm and repel another, under otherwise similar conditions.

We have seen above that the same reagent acts upon the same kind of protoplasm similarly only when the other conditions of the experiment are also the same. Among the varying conditions which have been especially investigated is that of the chemical constitution of the medium. The experiment has generally been made as follows: A particular species, let us say *Bacterium termo*, is to be subjected to the action of a particular reagent, *e.g.* meat extract. The bacteria are reared in cultures containing a varying quantity of the meat extract, and the concentration of the capillary fluid producing the threshold stimulation is measured in each case. We may compare not only the threshold stimulations but also the concentrations necessary to produce the response indicated by  $a_1$ ,  $a_2$ , etc. The following table, from PFEFFER ('88, p. 634), gives some of such determinations:—

CULTURE FLUID— MEAT EXTRACT.	CAPILLARY FLUID—MEAT EXTRACT.			
	3 × cult. conc.	5 × cult. conc.	8 × cult. conc.	10 × cult. conc.
0.01 %	0.03 % (?)	0.05 % ( $a_1$ )	0.08 % ( $a_2$ )	0.1 % ( $a_2$ )
0.1 %	0.3 % (?)	0.5 % ( $a_1$ )	0.8 % ( $a_2$ )	1 % ( $a_2$ )
1 %	3 % (?)	5 % ( $a_1$ )	8 % ( $a_2$ )	10 % ( $a_2$ )

From this table it appears that the strength of solution necessary to provoke a certain response depends upon the strength of solution to which the protoplasm has been previously subjected and increases proportionately with it. It is clear that the capillary solution of 0.1%, which produces a marked chemotaxis in bacteria reared in a culture solution of 0.01%, would awaken no response in bacteria reared in 0.1%.

We are now in a position to appreciate the importance of still another addition to our terminology of stimuli—the differential threshold stimulation (*Reizunterscheidsschwelle*)—which may be defined as the minimum increase of a preëxisting stimulus which is capable of calling forth a just noticeable reaction. In the case just cited, the differential threshold stimulation lies just above 3 times the preëxisting (culture) stimulus, and this is true whatever the degree of the preëxisting stimulus; and it is shown by experiment that, in general, as the preëxisting stimulus increases, the differential threshold stimulation increases in the same proportion. This observation is in perfect accord with the law formulated long ago by WEBER with especial reference to sight. This law runs: The smallest change in the magnitude of a stimulus which will call forth a response (differential threshold stimulation) always bears the same proportion to the whole stimulus. We may express this law mathematically, as FECHNER has done, by the following considerations: Let us take the case of a protoplasmic body, as, for example, that of a spermatozoid, living in a stimulating medium ( $s$ ) of a certain concentration and experiencing a certain reaction  $r'$ ; then  $s$  corresponds to  $r'$ . In order just to get a chemotactic response (threshold stimulation), a solution of say 30 times the concentration must be brought to the solution affording the stimulation  $s$ . This will give a reaction which is greater than  $r'$  by a quantity which we may designate  $r$ , so that the quantity of the whole reaction may be designated as  $r' + r$ . If the organisms are now placed in this stronger solution ( $31s$ ), the solution in the capillary tube must be 30 times stronger ( $30 \times 31s$ ) in order to give the differential threshold stimulation. The reaction following this stimulation may be designated, according to FECHNER's conception, as  $r' + r + r$ . The relation of the successive stimuli and the reac-

tions may be shown by the following table, following one given by PFEFFER ('84, p. 401) :—

$$\begin{array}{rcl}
 & & s \text{ corresponds to } r'. \\
 s + & 30 s = & 31 s \text{ corresponds to } r' + r. \\
 31 s + & 30 \times 31 s = & 31 \times 31 s \text{ corresponds to } r' + r + r. \\
 31 \times 31 s + 30 \times 31 \times 31 s = & 31 \times 31 \times 31 s \text{ corresponds to } r' + r + r + r.
 \end{array}$$

That is to say, while the stimulation increases geometrically, the reaction increases arithmetically. In the preceding table the second term of the left-hand member of the equation is always the differential threshold stimulation.

The most important objection that can be urged against this formula of FECHNER is that there is not sufficient reason for believing that the various reactions ( $r$ ) to the differential threshold stimulations of various strengths are equal, nor that the stronger reaction to the strong stimulus is composed of many weak reactions. If these assumptions were true, it would follow that when the successively higher stimuli increase as a series of numbers the reactions increase as the logarithms of these numbers. If now we adopt as a unit in this phenomenon the quantity of the threshold stimulation (estimated in units of concentration of solution, of mass, light intensity, heat intensity, etc.), which we may call  $s$ , the strength of any stimulus ( $S$ ) may be estimated in those units, and the strength of the corresponding reaction ( $R$ ) will be indicated by the equation  $R = c \cdot \log S$ , in which  $c$  is a constant to be determined empirically, and  $S$  the strength of the stimulus expressed in units of the threshold stimulation.\*

While we are not yet in a position to understand the significance of WEBER'S law, we cannot fail to be struck with the resemblance of the phenomena with which it concerns itself to those of acclimatization referred to in the second section of this chapter. We there showed that organisms subjected for a while to a chemical agent no longer reacted as at first to that reagent. We have here shown that organisms subjected for

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\* Since the German word for stimulus is *Reiz* (initial  $R$ ), and since the reaction is usually indicated by the initial letter in *Empfindung*, in German textbooks this formula usually runs  $E = c \cdot \log R$ , which differs from the above equation only in the symbols employed.

a while to the action of a certain stimulating agent respond no longer to a concentration which would at first have provoked a reaction. In both cases it is the action of the chemical agent which modifies the subsequent action of the protoplasm, without doubt by changing the chemical constitution of the protoplasm.

*Mechanics of Response.*— Having considered the general relation between strength of stimulus and of reaction, it now becomes necessary to examine more in detail into the way in which the reaction takes place.

A variety of kinds of locomotion exists among chemotactic organisms — that of the Myxomycetes is amœboid, that of Infusoria is by flagella or cilia. In all cases the first and perhaps the only effect of the acting reagent is to determine the position of the axis of the body, in the case of bodies with fixed form; or to determine the pole of outflow in the case of amœboid organisms. The axis lies in the line of flow of the diffusing solution or perpendicular to the isotonic lines, or lines of equal concentration. Whatever movement now occurs must be either towards or from the source of stimulus.

I have said above that the axis orientation is perhaps the only effect of the acting reagent. PFEFFER ('84, p. 463; '88, p. 631), indeed, maintains that the stimulus does not directly cause a markedly more rapid locomotion in the case of bacteria and Flagellata; but in the case of plasmodia it seems possible that such a hastening of movements occurs. (STAHL, '84.) However, it is necessary that *measurements* should be made in this matter. Further observations on the mechanics of taxis must be deferred to the general treatment of the subject in Chapter IX.

#### SUMMARY OF THE CHAPTER

We attempted in the first section to bring together observations relating to the action of various chemical substances upon protoplasm with the aims of discovering the general laws of poison-action on protoplasm and of gaining an insight into the chemical structure of protoplasm and the chemical operations involved in the elementary vital processes. We ought now, therefore, to attempt to draw such conclusions as the imperfect and often confusing data we have collected will permit.

All the reagents with which we have dealt have been substances capable of absorption by, mixture with, or solution in, water; and the reason for this is that almost all protoplasm is itself enveloped by water and largely composed of water.

For the most part we have dealt with mixtures or solutions. Now the action of these is a double one. They exhibit, first, an osmotic action, and, secondly, they may attack the molecules of the protoplasm, transforming them. The osmotic action we will consider in the third chapter; the transforming one alone concerns us now. It is not easy, without experiment, to say to which of these two categories of action the change wrought by any substance is due. To determine between the two possible causes it would be desirable in each case to treat the protoplasm to a control solution having the same osmotic action as the first, but no transforming effect. If such a solution produces no modification of the protoplasm, then the effect wrought by the first reagent is due purely to molecular transformations. It is not easy to find a reagent of which we may be certain that it acts only osmotically. NaCl is probably more generally useful in this way than any other substance. In the experiments which have hitherto been made, this double action of solutions has not been sufficiently considered. Hence, a doubt concerning the immediate cause hangs about many of the phenomena described in the first section.

The first principle which the data collected establish is that the protoplasm of different organisms is dissimilar. This is shown by the diversity in their chemical reactions; by the fact that whereas, in one case, a certain percent solution causes so extensive a molecular transformation as to result in death, in another, no injurious effect is produced.

Thus, according to BOER ('90, p. 479), it takes of gold chloride to kill —

TABLE V

Anthrax bacillus . . . . .	0.0125%
Cholera spirillum . . . . .	0.1%
Diphtheria bacillus . . . . .	0.1%
Typhoid bacillus . . . . .	0.2%
Glanders bacillus . . . . .	0.25%



Thus the weak solution, 0.0125%, of  $\text{AuCl}_3$ , which is fatal to anthrax, does not injure the glanders bacillus, which requires a solution 20 times as strong; and we conclude that the chemical constitution of the glanders bacillus must be different from that of anthrax.

The dissimilarity of the different protoplasms may be either a qualitative or a quantitative one. That is to say, the *kinds* of molecules, or the *proportions* of the different molecules, may differ in the two cases. If we assume that gold chloride acts upon protoplasm by the Au replacing some of the H in an amido-acid, then the diversity in action of  $\text{AuCl}_3$  upon anthrax and typhoid may be accounted for by assuming that the amido-acids are dissimilar in anthrax and typhoid or that the proportion of the kinds especially affected is different in the two cases. To which of these two causes the diverse reactions to  $\text{AuCl}_3$  are due cannot yet, in any given case, be determined.

A second principle which we may draw from our data is this: Some kinds of protoplasm have a *general* high resistance to all chemical agents, while other kinds have a high or low resistance to particular agents only (*specific* high or low resistance). Thus, in the case of pathogenic bacteria, the experiments of BOER ('90) show that, in general, the anthrax bacillus has a low resistance, and glanders a high one. His experiments were made with 10 reagents upon five kinds of bacteria. Table VII gives in modified form the results obtained by BOER. His results are given in the form of Table V; the present table is constructed from the original by making the mean of the five observations in each column unity and reducing the separate observations proportionately. Thus Table V becomes —

TABLE VI

Anthrax bacillus . . . . .	0.09
Cholera spirillum . . . . .	0.75
Diphtheria bacillus . . . . .	0.75
Typhoid bacillus . . . . .	1.51
Glanders bacillus . . . . .	1.88

All the other determinations have been treated in like manner. Throughout the table the numbers in each column stand for

*relative* resistance capacity. The reagents are placed with the weakest-acting first.

TABLE VII

SUBSTANCES.	CAUSTIC SODA (NaHO).	CARBOLIC ACID (C <sub>6</sub> H <sub>6</sub> O).	MURIATIC ACID (HCl).	SULPHURIC ACID (H <sub>2</sub> SO <sub>4</sub> ).	METHYL VIOLET.
MOLECULAR WEIGHTS.	<b>40</b>	<b>94</b>	<b>37</b>	<b>98</b>	
Anthrax bacillus . . . .	0.46	0.95	0.40	0.37	0.07
Cholera spirillum . . .	1.38	0.71	0.32	0.37	0.33
Diphtheria bacillus . .	0.69	0.95	0.63	0.94	0.17
Typhoid bacillus . . . .	1.09	1.43	1.46	0.94	2.22
Glanders bacillus . . . .	1.38	0.95	2.19	2.36	2.22

SUBSTANCES.	SILVER NITRATE (AgNO <sub>3</sub> ).	GOLD CHLORIDE (AuCl <sub>3</sub> ).	MALACHITE GREEN.	OXYCYANIDE OF Hg.	AVERAGE.
MOLECULAR WEIGHTS.	<b>170</b>	<b>304</b>			
Anthrax bacillus . . . .	0.20	0.09	0.02	0.93	0.388
Cholera spirillum . . .	1.04	0.75	0.17	0.62	0.632
Diphtheria bacillus . .	1.67	0.75	0.11	0.93	0.760
Typhoid bacillus . . . .	1.04	1.51	1.75	1.25	1.415
Glanders bacillus . . . .	1.04	1.88	2.92	1.25	1.802

From this table we see that the bacillus of glanders is more resistant than that of anthrax (except in one instance, in which the resistance is equal in the two cases) whatsoever be the poison employed. The bacillus of glanders affords, thus, a good illustration of an organism with a general high resistance capacity.

The diversity in general resistance capacity which is found among bacteria exists also among other organisms. Thus, the parasitic *Ascaris* has shown itself highly resistant in all cases in which the action of a poison on it has been compared with that on another species; for instance (p. 10) 0.1% chloral hydrate kills Infusoria, Rotifera, and diatoms in 24 hours, but *Ascaris* withstands this solution. Again, while 0.1% HCN kills Infusoria quickly, *Ascaris* resists 3% for 75 minutes. The general higher resistance may be due to one of three causes:

either to the fact that the protoplasm is protected from attack, as is the case with the encysted forms of Protozoa, which are very resistant; or to the fact that the protoplasm is not so readily acted upon by reagents brought actually in contact with it, due to diminished amount of water or other structural modifications; or, finally, that the protoplasm has a different composition, certain unstable molecules found in other kinds of protoplasm being absent.

We will now consider the phenomena of diversity in *specific* resistance of protoplasm. A case of specific *low* resistance is found in the nervous tissue. Thus many of the alkaloids, *e.g.* nicotine and cocaine, are almost indifferent to the protoplasm of Protista, but act towards the nervous system as powerful poisons. Hence we are led to conclude that nervous protoplasm contains especially unstable compounds, upon which its action depends. When they are subjected to the action of very weak —towards most substances, indifferent— reagents, extensive and fatal transformations occur.

Cases of specific *high* resistance are apparently found in some glands which secrete intense poisons, or in some organisms which live in solutions of some usually poisonous agent. Examples of this class are the HCl-secreting glands of the Vertebrate alimentary tract, the poison-glands of venomous serpents, and the H<sub>2</sub>SO<sub>4</sub>-secreting glands of Gasteropoda; also the vinegar eel, which lives in 4% acetic acid. It ought to be said that it is largely an inference based upon experiments on acclimatization, that these glands or organisms will not show a *general* high resistance. Experiments are needed to determine this point. As to the cause of specific high resistance, I believe that much light is gained from the facts of acclimatization, and that any sufficient theory of the latter would serve also to explain the former (see p. 30).

Under the general poisons we have distinguished four main groups: *a.* oxidizing poisons; *b.* salt-forming poisons; *c.* substitution poisons; *d.* catalytic poisons. I will comment briefly upon the action of the poisons of each of these groups.

*a. Oxidizing Poisons.*—The ordinary oxidation processes in living protoplasm involve the consumption not of the proto-

plasm itself, but of the thermogenic substances stored therein (sugar, yolk). After these have been consumed in starvation, or when the organism is subjected to the action of oxidizing poisons, the molecules of the protoplasm become oxidized. All protoplasm which is readily accessible must be injured by the direct attacks of "active" oxygen.

*b. Salt-forming Poisons.*—The facility with which an acid or a base forms salts with the protein substances of the protoplasm must depend, in large part, upon the quality of the protein molecules. It is well known that certain protein substances, such as keratin, chitin, and fibrin, are not readily acted on by acids or bases, and it seems necessary to suppose that some such resistant proteids are the essential parts of glands which secrete these reagents. Into this group fall the salts of heavy metals characterized by their extraordinary fatalness.

*c. Substitution Poisons.*—This group comprises, besides a few sulphur compounds, almost exclusively nitrogenous substances, and among these a large proportion of compounds with closed chains. As many of these are indifferent to dead albumen, but violent poisons to living protoplasm, it is clear that the latter must contain certain extremely unstable groups (amido-, aldehyde-, and keton-groups, LOEW, '80). Among these poisons the relation between molecular structure and poisonous action is very marked, especially in the nitro-compounds. Thus, bodies containing H united with N are poisonous in direct proportion to the number of H atoms so combined. It seems probable that H so combined is very easily given up to the molecules of the living substance, destroying them. H in the hydroxyl radical seems also more easily parted with than H joined to C.

*d. Catalytic Poisons.*—Chiefly organic compounds of the fat series, which have little chemical energy and produce, for the most part, anæsthesia. The poisonous action seems here proportional to the complexity and instability of the compound. Thus, in many groups, when the alkyls  $\text{CH}_3-$ ,  $\text{C}_2\text{H}_5-$ , etc., are successively introduced, the substance grows more poisonous as the number of atoms in the alkyl increases. In the methan series and among sulphureted compounds the substitution of Cl for H increases the poisonous action.

The study of the action of poisons upon protoplasm gives us an insight into the extreme complexity of the living substance — its composition out of numerous kinds of compounds, many of which are extremely unstable. Not all protoplasm contains the same compounds, hence it must be a very dissimilar thing in different organisms. Not all of the compounds in any protoplasmic body are essential to life, for we may act upon a protoplasmic body by a weak reagent, and gradually change its composition so that it will no longer be killed by the strong solution, and all of this without perceptible injury — at least, this is the conclusion to which the study of acclimatization of Protista leads us. The altered chemical constitution will be transmitted in the division of the individual, and thus the composition of the protoplasm of a race will have been determined by the medium in which it and its ancestors have been living.

Finally, we may consider what light the action of reagents throws upon the processes involved in the elementary vital functions. The normal *movement* of protoplasm is profoundly modified by interfering with the oxygen supply. Thus, when the oxygen pressure is diminished, movements are retarded ; in the presence of pure oxygen they are accelerated. Some anæsthetic or paralyzing agents — *e.g.* chloroform and some alkaloids, veratrin, atropin, cocaine, strychnin, and antipyrin — give rise first to acceleration, then to disappearance of movements in the protoplasm. Protoplasmic movement is, consequently, closely associated with oxidation, and it does not occur in the absence of irritability.

Normal *locomotion* is interfered with by strychnin and cocaine. Their stimulating action produces accelerated movements, and these are accompanied by loss of coördination.

Since many catalytic poisons (anæsthetics) destroy *irritability*, one may conclude from the action of these chemical agents that (p. 7) stability of molecular movement is essential to the performance of this function.

Disturbance of the *excretory* function results from the action of CO, NH<sub>3</sub>, chloroform, cocaine, strychnin ; at least, an excessive vacuolation of the protoplasmic body occurs under the action of these agents.

Experiments on chemotaxis show that many substances brought near to protoplasmic bodies control their locomotion. The effect upon locomotion depends both upon the kind of protoplasm and the strength of the reagent. In many cases, a certain strength of reagent attracts an organism, while a stronger solution repels, and a weaker solution is indifferent. In such a case we may speak of the protoplasm as being attuned to the attracting strength of the reagent. We find great diversity in the strength of solution of a reagent to which different protoplasms are attuned. This difference of attunement to chemotactic reagents is parallel to the difference in strength of the killing solution of various protoplasms. As the latter is probably due to the past action of chemical agents upon the protoplasm, so is also the former.

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#### APPENDIX TO CHAPTER I

##### *Cytotaxis (= Cytotropism)*

ROUX ('94) has given the latter name to a phenomenon which is probably only a special case of chemotaxis, but which may be better considered apart. He isolated, in an indifferent medium, two or three cells from the egg of a frog (*Rana fusca*) at the morula or blastula stage of development. These he placed near each other upon a glass slide, and found that they moved slowly, and that the direction of the movement of any one cell was, under certain conditions, determined by the position of the other cell or cells.

In order to perform the experiment a proper medium in which to study the movement of the cells must be prepared as follows: a small quantity—5 to 10 ccm.—of fresh egg albumen (not cut up, but with the albumen threads intact) is filtered through clean wadding, and the completely clear filtrate is used. In other cases a more or less strong salt solution is employed. The cleavage cells are isolated in the filtrated albumen, on a glass plate, by means of needles. To diminish evaporation the glass plate is put into a shallow glass vessel containing several drops of water.

When two cells were placed near each other (about one-fourth of their diameter apart), the distance between them diminished. The approach took place along a line joining the two cells;

and when several pairs of cells were in the field, this movement took place in various directions, indicating that their movement was not determined by conditions outside the approaching cells. To get further light on the migration of the cells, their distance apart was measured at short intervals of time. The results of two series of such measurements are represented graphically in Figs. 6 and 7. In both of these diagrams the heavy lines indicate the successive positions assumed by four points; namely, the points of the two cells which are nearest each other and those which are most distant. In the first case the cells traverse the distance of their diameters ( $58 \mu$ ) in about 10.5 minutes. The rate of migration is, however, extremely variable. In some cases the cells seem even to move apart (negative cytotoxic?).

Certain special cases are worthy of consideration. When a third cell lies near an approaching pair, the path of migration of the pair may become convex towards the third cell. Two cell-complexes, each composed of three or four cells, may approach and connect. But masses composed of a larger number of cells form "closed complexes" which show no cytotoxic activity. The isolated cells of differ-



FIGS. 6, 7.—Two sets of curves, showing the course of "cytotoxic" movements of the cleavage cells of the frog. In each figure the dotted line represents a diameter of the cell. The full line represents the successive positions of the extremities of the diameters as the cells approach. The distances between horizontal lines =  $4 \mu$ ; between vertical lines, 75 seconds. (From Roux, '94.)

ent eggs of the same species behave like cells from the same egg.

By these important experiments it is established that, inside of the body, parts may act upon parts, determining the direction of motion. The importance of this fact will be discussed in a later Part of this book.

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## CHAPTER II

### *EFFECT OF VARYING MOISTURE UPON PROTOPLASM*

IN this chapter it is proposed to speak (I) of the amount of water in organisms; (II) of the effect of desiccation upon the functions of protoplasm; (III) of the acclimatization of organisms to desiccation, and (IV) of the control of the direction of locomotion by moisture — hydrotaxis.

#### § 1. ON THE AMOUNT OF WATER IN ORGANISMS

Any theory of the structure of protoplasm must recognize that water forms the greater part of the whole mass; between 60% and 90%. In the case of dry seeds and grains, however, it may fall below 15%. Many determinations have been made of the proportion of water in the body of entire organisms and in their organs. I give in tabular form some of these determinations, which were made by BEZOLD ('57), designated by (B); KRUKENBERG ('80), designated by (K); and LIEBERMANN ('88), designated by (L).

TABLE VIII

SPECIES.	CONDITIONS OF WEIGHING.	% WATER.
Various sponges (K) . . . . .	In most cases, kept a short time in fresh sea water; dried on surface and weighed.	84.0 to
		74.5
Medusa: Rhizostoma Cuvieri (K).	Whole animal, directly from water Piece of disc.	95.4
		95.0
Various Actinia (K) . . . . .	A few minutes after removal from sea. A little water lost from central cavity	87.7 to
		83.2
Acyonium palmatum (K) . . . . .	Weighed when fresh, 73.5 g.	84.3
Asteracanthion glacialis (K) . . . .	Weighed 850 g.	82.3
Lumbricus complanatus (K) . . . . .	2 large specimens	87.8

SPECIES.	CONDITIONS OF WEIGHING.	% WATER.
<i>Oniscus murarius</i> (B) . . . . .	200 young individuals	68.1
<i>Squilla mantis</i> (K) . . . . .	1 individual	81.9
<i>Astacus fluviatilis</i> (B) . . . . .	3 individuals weighing from 16.6 to 27.4 g.	71.1
<i>Doris tuberculata</i> (K) . . . . .		88.4
<i>Doriopsis limbata</i> (K) . . . . .	3 individuals	86.5
<i>Arion empiricorum</i> (B) . . . . .	6 individuals weighing from 4.3 to 27.1 g.	86.8
<i>Limax maximus</i> (B) . . . . .	4 individuals weighing from 0.1 to 17.1 g.	82.1
<i>Botryllus</i> (K) . . . . .	4 individuals weighing from 111.2 to 35.2 g.	93.6
Various Vertebrates (B) . . . . .		58.4 to 80.1
Chick (L) . . . . .		
7 days old . . . . .	Embryo only, yolk removed	92.8
21 days old . . . . .	Embryo only, ready to hatch	80.4
Turnip (root) . . . . .	From Goodale's <i>Physiolog. Bot.</i> , p. 236	91.0

These determinations suffice to show that water immensely predominates over any other substance in active organisms, and indicate that it plays an important role.

The role played by water is, in fact, extremely varied. It serves to maintain that unstable, foam-like structure of the protoplasm upon which its capacity for movement depends; it acts as a solvent for matter taken into the protoplasmic body; and it serves to transport dissolved substances from place to place in the organism. In a word, it is essential to movement and to those chemical processes which constitute metabolism.

## § 2. ON THE EFFECT OF DESICCATION UPON THE FUNCTIONS OF PROTOPLASM

We may consider this topic under the following heads: (1) effect upon metabolism; (2) effect upon the motion of protoplasm; and (3) the production of desiccation-rigor and death.

1. **Effect of Dryness on Metabolism.** — Since water is so essential to metabolism, we should expect that a diminution of metabolism would accompany dryness. And this is clearly the case. Thus dry seeds, in which the water is reduced to only 10

to 15%, when placed under conditions of temperature favorable to metabolism, show almost no change in the course of days. This has been indicated also by an experiment of KOCHS' ('90, p. 685), who placed seeds, which had been dried in a vacuum, in a receptacle connected with a GEISSLER'S tube, such as is used in the spectroscopic study of gases. The air was completely pumped out of both vessels, and after some months a spectroscopic study of the gases in the GEISSLER'S tube showed no trace of nitrogen or carbon, yet the seeds later germinated. This experiment can hardly be considered to demonstrate KOCHS' point, however, since the seeds were deprived of oxygen, as well as moisture.

The act of drying may, on the contrary, induce the manufacture and elimination of certain secretions. This occurs apparently in many Protista, which form cysts in the drying pools. This phenomenon is seen again in some of the higher animals, — such as our garden slugs, — which secrete slime in large amount when kept for a short time in a dry place. In both cases the result is of immense importance for the continued life of the organism, — in both cases it is to be considered a response to the stimulus afforded by evaporation of water.

**2. Effect of Dryness upon the Motion of Protoplasm.** — We have seen that water plays an important role in the movement of protoplasm. When by any means the water is partly withdrawn, the protoplasmic currents will be slowed. When, on the contrary, protoplasm, which is lying in an "indifferent" medium, such as blood serum, is placed in distilled water, unusually active movements occur. This has been shown by ENGELMANN ('68, p. 446) in the case of the spermatozoa of the frog, and the ciliated epithelium of the frog's œsophagus just removed from its body. Similarly, DEHNECKE ('81) found that protoplasm of the tissue cells of the higher plants exhibited abnormally rapid movements upon adding water. These observations indicate that water may act as a stimulus to the movement of protoplasm.

**3. Desiccation-rigor and Death.** — It is a familiar fact which has been established by over a hundred and fifty years of experimentation, that some organisms, when gradually dried, may cease from movements. This immotile condition has

sometimes been regarded as death. By PREYER ('91) it has been called "anabiosis." I shall call it desiccation-rigor, and correlate it with phenomena, produced by various other agents, which cause the cessation of a movement that is restored again when the action of the untoward agent is withdrawn.

And these are just the conditions we meet with here, — cessation of activity without loss of power of revival. This was very evident from the work of SPALLANZANI (1787, Tom. II, pp. 212, 213). This author showed clearly that one and the same adult rotifer can be observed in the evaporating drop, until all the water is gone, and it has lost all movement and its normal form. If, after an hour, the slide is moistened again, the rotifer reassumes, by degrees, its natural form and activities. SPALLANZANI noticed, what has been the nearly unanimous testimony of subsequent observers, that a rotifer dried for hours on clean glass does not revive; revivification occurs only when the rotifers have crawled into sand. As for the length of time during which desiccation-rigor may persist in rotifers without death occurring, we know only that it may be considerable, extending through months, and even years.

Similar phenomena to those observed in rotifers have been described for tardigrades and certain nematodes, although these organisms have not been studied in so much detail. Among the tardigrades only those species which live in moss, and are thus especially liable to desiccation, withstand drying. (LANCE, '94.) Among nematodes, *Tylenchus devastatrix*, KÜHN, which lives in grains of wheat, is a classic object of study. *Strongylus rufescens* is, according to RAILLIET ('92), capable of resisting dryness for 68 days or more. We may thus conclude that adult organisms of certain species may be subjected to desiccating influences, and that those same individuals may resist them so as to reëxhibit activities after the return of favorable conditions.\*

While the results of these drying experiments are scarcely doubted, much difference of opinion has arisen concerning the interpretation of the results. The first moot point is the degree of desiccation which the protoplasm of the organisms

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\* See in connection with this the valuable report of BROCA ('61).

experiences. Many writers have assumed that this has been rendered in their experiments very great or nearly perfect. Thus DOYÈRE ('42, p. 28) says: "What is the condition of the animalcules in the dried sand of the gutters? I have never seen them, at such times, in any other condition than reduced to spangles as fragile and more deformed than when dried free on the glass. I have never discovered a single one which manifested any traces whatever of life, or which did not present all the appearances of a *complete* desiccation. Nevertheless, I do not pretend by this to invalidate all contrary assertions; the principal fact, that of the return to life after an *absolute* desiccation, is not affected thereby." The physicist GAVARRET ('59, p. 317) subjected, for 34 days, moss containing rotifers to a vacuum having a pressure of only 4 mm. of mercury, and other experimenters have likewise employed a similar "chemically drying" device, which they believed capable of extracting all water from the protoplasm.

The evidence that all water is withdrawn from the body of the organism is often very slight. The fact that the seeds or plant tissues in which nematodes or tardigrades are living, have been dried until they lose no appreciable weight, is not sufficient evidence that their inhabitants are completely dried.

On the other hand, there is positive evidence that one, at least, of the organisms which has been considered as having been absolutely dried, can protect itself from this condition. It is especially DAVIS ('73) who has shown this. This author has experimented with the rotifer *Philodina roseata*. When dried on a glass plate with sand, it assumes a spherical form. At the same time, however, it secretes a gelatinous envelope. Thus encapsuled it may rest for days until upon the addition of water it reassumes its active, adult form. That a layer of such gelatinous substance is sufficient to resist the drying action of a vacuum-chamber with sulphuric acid, was illustrated by putting grapes varnished with gelatine in such a dry chamber for one week. They emerged in a fresh, juicy condition. One of the encapsuled rotifers was crushed after "desiccation" and yielded under the cover-glass a drop of fluid. In this case then the rotifer was not fully dried. DAVIS accounts for the fact that isolated rotifers dried on a clean



slide will not survive desiccation, on the ground that the sand forms a necessary retreat in which the organism can quietly encapsule itself. Of the fact of such encapsuling there can be no doubt; it is abundantly substantiated by the testimony of HUDSON ('73 and '86). There is a doubt, however, whether this encapsuling is a phenomenon common to all organisms which can resist desiccating influences, and, therefore, whether DAVIS's explanation is generally applicable. To sum up: I believe there is no sufficient evidence that an adult organism or active protoplasm of any sort can rapidly lose all of its "free water" without such a destruction of its finer structure as would make it incapable of exhibiting vital activities upon moistening again.

A much greater capacity undoubtedly inheres in spores and seeds. Thus KOCHS ('90) subjected perforated seeds of *Zea mais*, *Phaseolus*, and *Triticum vulgare* to an almost perfect vacuum (made by a mercury pump) for 8 days, and they nearly all germinated. Even the small radish seed, with part of the cuticula removed, subjected to a vacuum for three weeks germinated perfectly. Probably there is no limit to the amount of desiccation which seeds and other masses of protoplasm especially adapted to resist desiccation can withstand.

The second moot point is this: Is the protoplasm, rendered immotile by drying, living or not? SPALLANZANI prejudiced the question by the title of his chapter on this matter.—"Observations and experiments on some marvellous animals which the observer can at his will make pass from death to life"; and he and many of his successors argue that death has truly occurred. PREYER ('91), however, prefers to reserve the term "dead" for protoplasm which is at the same time lifeless and incapable of life; while to protoplasm which is lifeless but capable of revivification he applies the term "anabiotic." The question then is this, is life truly suspended during the immotile state? If we think of life as the sum of the chemical changes occurring in the protoplasm, we shall realize that all degrees of vitality, even to complete cessation of activity, may occur without our being able anywhere to say at this point life becomes extinct. We can hardly hope ever to deny that minimum vital changes are occurring; since the minimum changes

must be beyond our ken. That these vital changes are sometimes exceedingly slight, is sufficiently indicated by the experiments upon seeds performed by KOCHS ('90), and referred to on p. 60. That, however, slight changes are occurring even in seeds is indicated by the fact that dessication-rigor cannot continue indefinitely without loss of the power of revivification.

In the case of the rotifers, tardigrades, and nematodes, months, and even years, may elapse without complete loss of capacity for revivification. It is generally admitted, however, that in cases of long-continued drought, the chances of revitalizing upon moistening are much diminished.\* In the case of seeds it has been maintained that under certain conditions, such as are realized in the mummy-graves of Egypt, life may persist for more than a thousand years. However, the experiments of MÜNTER ('47), and more especially of KOCHS ('90, p. 683), throw doubt upon this assertion, since they found that the ancient, charred seeds fell to pieces in water like lime. As for seeds preserved above ground in the ordinary way, KOCHS was assured by seedmen that they could not remain capable of germination over 10 years. These facts go to show that gradual changes occur in the dry protoplasm which are probably metabolic changes, *i.e.* vital changes; and that therefore life is hardly extinct in the very dryest protoplasm.

The whole matter of desiccation-rigor is, after all, one with which we are familiar in nature's larger laboratory. Many Protista, when the ponds in which they live dry out, encyst themselves and enter into a motionless condition in which they resist the hot and dry summer winds. Thus, they may lie for weeks, and, as experimentation has shown, they may be dried for several years (see BÜTSCHLI, '89, p. 1663, for references) without loss of capacity for revivification. The same device

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\* Thus RAILLIET ('92) says of *Strongylus rufescens*: "I have seen them regain their activity after 42 and even 68 days of desiccation. However, this activity is much slower in manifesting itself. After the course of a month a contact of 8 to 10 minutes is sufficient to bring them back to movement. . . . After 68 days at least 50 minutes are required, and certain individuals have shown activity only after 1 hour and 20 minutes. Moreover, the movements were limited, and only a small number of cases contorted themselves like ordinary *Anguillulidæ*."

for resisting desiccation is seen in the gemmules of sponges, and Bryozoa, the eggs of many animals, and the spores of many plants. Thus some protoplasm normally responds to the stimulus of drought by going into desiccation-rigor.

While, as we have seen, some protoplasmic bodies may be dried as far as possible by the ordinary methods used in chemistry without death ensuing, other bodies, especially the adult forms of higher organisms, whose cellular respiration is dependent upon a circulating fluid, are killed by desiccation. For loss of this fluid or desiccation-rigor in the pumping muscles will produce asphyxia. But these conditions do not militate against the belief that there is no necessary lower limit to the amount of water which must occur in quiescent protoplasm in order that it may retain vitality during a limited period.

### § 3. ON THE ACCLIMATIZATION OF ORGANISMS TO DESICCATION

We have seen in the last section that certain organisms are more capable of resisting desiccation without fatal effect than others; *e.g.* rotifers, tardigrades, and *Tylenchus*. Now it is clear that these organisms are especially apt to become dried, so that it is possible that their high capacity for resistance has been produced by acclimatization without selection. I shall here add certain other cases of resistance to dryness which I believe, but cannot prove, to have been thus produced. LANCE ('94) has mentioned, as already stated, that only those tardigrades which live in the moss of gutters (where they are alternately wet and dried), and not those living in water, show the phenomenon of revivification. CERTES ('92) has found that, although marine Ciliata cannot, in general, withstand desiccation, those from the chotts and saline lakes of Algeria may be dried like those from fresh-water ponds and swamps. The difference in resistance between the forms dwelling in the sea and in inland salt-water ponds is doubtless due to the fact that the former are not regularly desiccated, while the latter are; consequently the latter alone have had a chance to become acclimated to desiccation.

#### § 4. THE DETERMINATION OF THE DIRECTION OF MOVEMENT BY MOISTURE, — HYDROTAXIS

This phenomenon has been described by STAHL ('84) in *Æthelium*. When this Myxomycete is placed in the dark upon a glass plate covered with several layers of moistened filter paper, it expands uniformly over the homogeneously moistened substratum. If, now, the plate be placed in a drying chamber, the paper dries slowly, and one can see that the mass of the plasmodium draws towards those places which remain longest damp. If a dilute gelatine jelly is smeared upon a glass slide supported in a horizontal position about 2 mm. above the plasmodium, still in the dark chamber, the plasmodium sends up branches, some of which may touch the gelatine and spread out over it. If the water dries still further, the entire Myxomycete may become transferred to the slide above. If, now, the paper be moistened again, the plasmodium sends branches down to it. STAHL'S explanation is that of the old mechanical school. He says, the peripheral protoplasmic layer lying next the dryer region is poorer in water; while that next the damper part of the substratum contains much water. If it be assumed that the internal streaming tends to occur uniformly towards all points of the periphery, it is clear that the dryer, more consistent part will offer greater resistance than the more fluid part, and in this part, therefore, branches will tend to arise. In correspondence with the interpretations which we have hitherto placed upon similar phenomena I prefer to call this a case of response to the stimulus of excessive moisture — in any case it may be designated positive hydrotaxis.

When, however, the plasmodium of *Æthelium* is in the fruiting stage, it retreats from the moister part of the substratum, and other Myxomycetes in the fruiting stage show the same negatively hydrotactic tendency. Thus the same agent, water, stimulates the same organism, at different stages, to reverse movements.

I will now summarize the conclusions concerning the effect of water upon protoplasm. Water constitutes by far the larger part of protoplasm and of all active organisms. Metabolism is

directly dependent upon it, and certain excretory processes are stimulated by it. The motion of protoplasm is likewise dependent upon water, which determines the unstable condition of that substance. Desiccation, therefore, produces a rigor, and this may continue, in the absence of water, for months, and even years, the organism being, meanwhile, ready to awake to activity upon the return of moisture. The degree of desiccation which organisms can resist varies. In the case of the higher organisms, it is slight; in the case of bodies especially adapted to resist dryness (spores, seeds, statoblasts), there is, perhaps, no practicably attainable limit to the dryness which their protoplasm may undergo without loss of power of revivification. The condition of desiccation-rigor is not known to be one of death which is replaced by life upon return of moisture. It is probably rather a condition of minimum metabolism. The great resistance capacity exhibited by certain organisms is correlated with their liability to desiccation in their natural surroundings. Finally, Myxomycetes (and probably other organisms) respond to inequalities in the amount of moisture in their environment, moving either towards or from greater moisture. We recognize thus that the activities of protoplasm are to a large extent dependent upon the existence of water in it; and that protoplasm reveals itself as sensitive to differences in the amount of moisture, responding by secretions, by the assumption of a quiescent condition, and by locomotion with reference to water.

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## CHAPTER III

### ACTION OF THE DENSITY OF THE MEDIUM UPON PROTOPLASM

IN this chapter we shall consider (I) the structure of protoplasm and the physiological action of solutions; (II) the effect of density upon the structure and general functions of protoplasm; (III) acclimatization to solutions of greater or less density than the normal; and (IV) control of the direction of locomotion by density — tonotaxis.

#### § 1. INTRODUCTORY REMARKS UPON THE STRUCTURE OF PROTOPLASM AND THE PHYSICAL ACTION OF SOLUTIONS

It is now generally recognized that protoplasm consists of two substances closely interwoven: the living plasma and a watery chylema. The relation of the plasma and the chylema is still a debated matter. Since the only theory of the structure of protoplasm which has been experimentally tested is that of BÜTSCHLI, his theory is especially worthy of recognition. According to this theory, the relation of plasma and chylema is that of water and air in a foam-work. The whole protoplasmic mass is bounded and penetrated through and through by plasma films which envelop watery globules. It is with membranes constructed of such protoplasm that the physical phenomena of osmosis are exhibited.\*

Osmosis occurs when two aqueous solutions of different density are separated by an animal membrane.† Such a mem-

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\* Excellent treatises on the physical and chemical nature of solutions, including a discussion of osmosis, are: OSTWALD, '91, and WIETHAM, '95.

† Osmosis occurs likewise when such solutions are separated by inorganic walls containing pores of extreme fineness; *e.g.* a wall of porous clay in which copper ferrocyanide has been precipitated.



brane permits the free passage of water, but not of the dissolved substance, or rather, of the dissolved substance but slowly. Under these conditions, the water flows more rapidly towards the solution containing the greater number of molecules (per cc.). The theory of this movement is that upon the side containing the greater number of molecules of salt fewer water molecules will in a given time strike the membrane than upon the other side; and since the number passing through is proportional to the number striking, relatively fewer molecules of water will consequently pass out, and so there will be a resultant flow of water to that side; and if the mass of water is confined, it will exert great pressure.

This phenomenon of osmosis plays an important part in organic life. Thus, under certain conditions, cells take in the surrounding water, so that their walls are put under tension (turgescence). The tension thus gained may be considerable, amounting to 6 or 7 atmospheres. Under other conditions the cells give up their water to the surrounding medium, thus losing their turgescence. This occurs when they are put into certain solutions of  $\text{KNO}_3$  or  $\text{NaCl}$ . The relation between the density of the internal and external fluids thus determines the internal pressure experienced by the cell.

A quantitative method of determining this pressure in the presence of various solutions has been employed by PFEFFER ('77). Solutions of different *dry* salts in different proportions, enveloped by a semi-permeable membrane, were placed in pure water, and the pressure upon a column of mercury determined. It was found, for example, that with a 1% solution of cane sugar a pressure of 47.1 cm. of mercury\* was produced; with a 1% solution of  $\text{K}_2\text{SO}_4$ , a pressure of 193 cm. of Hg. He concluded, as a result of his various experiments, (1) that the pressure is proportional to the concentration of the solution, and (2) that as the temperature rises the pressure increases.

DE VRIES ('84) made a noteworthy advance, using plant cells as objects of experimentation and subjecting them to various solutions of substances freed of water. He determined the degree of concentration which a solution of  $\text{KCl}$  must have

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\* The pressure of 76 cm. of mercury equals that of 1 atmosphere.

in order that no endosmosis or exosmosis should occur through the cell wall.\* He next determined the same thing for some other substances, *e.g.* KI, and found that the degree of concentration which produces no osmosis is, for two different solutions, proportional to the molecular weights of the salts dissolved in them. Solutions which produce the same osmotic effect DE VRIES called isotonic. A solution of 0.746% KCl is isotonic with a solution of 1.661% of KI, for the molecular weight of KCl is 74.6, and that of KI is 166.1. Thus the first result which DE VRIES gained was that the osmotic effect of solutions of salts of similar structure depends upon the number of their molecules in the solution.

The second conclusion of DE VRIES was that salts of *dis-similar structure* have different osmotic properties, even when the number of molecules in the two solutions is the same. Thus, he found that with an equal number of molecules to the solution (molecular-weight solutions †) :—

- (1) All salts of alkalis with one atom of metal to the molecule are isotonic (formula,  $R'A'$  [composed of a monad metallic radicle,  $R$ , and a monad acidic radicle,  $A$ ]);
- (2) All organic compounds with no metal radicle have two-thirds the osmotic action of the first group; *e.g.* cane sugar,  $C_{12}H_{22}O_{11}$ . ‡

\* As is well known, when a fully developed plant cell is put into a strong saline solution the living plasma sac separates from the cell wall and contracts, eventually, into a ball,—the result of the chylema flowing out of the protoplasm (plasmolysis). The weaker the concentration, the less marked the plasmolytic phenomena. Finally, a concentration is reached so weak that the separation of the plasma sac hardly occurs or is limited to a single corner. This concentration may be regarded as equal to that of the cell-sap—as that at which no osmosis occurs. (See Fig. 8.)

† I shall use the phrase “molecular-weight solution” to indicate solutions in the making up of which the molecular weight of the substance in grammes, dissolved in 100 g. of water, is used as the unit of concentration. It will often be convenient to abbreviate it as MW % sol. Chemists frequently use as a unit solution, called “normal” solution, the molecular weight in grammes dissolved in 1000 g. of water. Our MW % sol. is therefore equal to one-tenth of a “normal” solution.

‡ The fact that glycerine can be absorbed by some plants has introduced a complexity into the determination of its isotonic coefficient. This determination has been made the subject of a special investigation by DE VRIES ('88), who, by the use of slowly absorbing plants, has found the isotonic coefficient to be 1.78, which agrees approximately with the number given above for organic compounds.

- (3) All salts of alkalis with two atoms of metal to the molecule have four-thirds the osmotic action of (1) (formula,  $R_2'A''$ ); e.g.  $K_2SO_4$ .
- (4) Salts of alkalis with three atoms of the metal to the molecule have five-thirds the osmotic action of (1) (formula,  $R_3'A'''$ ); e.g.  $K_3(C_6H_5O_7)_x$ .

In other words, the osmotic action of groups (2), (1), (3), and (4) are in the proportions of 2, 3, 4, 5. These last numbers are the *Isotonic Coefficients* of DE VRIES. In addition to these substances, DE VRIES determined that the isotonic coefficient is, in the case of —

salts of earthy alkalis with 1 acid radicle; e.g.  $MgSO_4$  . . . 2  
 salts of earthy alkalis with 2 acid radicles; e.g.  $CaCl_2$ . . . 4

In the third place DE VRIES established the law that each acid group and each metal has, in all compounds, the same partial isotonic coefficients; the coefficient of any salt is the sum of these partial coefficients of the constituent components. These partial coefficients are : —

for each atom-group of an acid . . . . . 2  
 for each atom of an alkaline metal (Li, Na, K, Rb, Cs) . . . 1  
 for each atom of an earthy metal (Ca, Sr, Ba, Mg) . . . . 0

while of the compounds the isotonic coefficients are —

$$\begin{aligned} KCl &= 1 + 2 = 3, & MgSO_4 &= 0 + 2 = 2, \\ K_2SO_4 &= 2 \times 1 + 2 = 4, & MgCl_2 &= 0 + 2 \times 2 = 4, \\ K_3(C_6H_5O_7) &= 3 \times 1 + 2 = 5, \text{ etc.} \end{aligned}$$

The determination of isotonic coefficients has subsequently been extended by several authors, especially by HAMBURGER ('86 and '87) and by MASSART ('89).

The work of HAMBURGER was done upon blood corpuscles. The method employed by him was as follows: In certain weak solutions the hæmoglobin passes out of the red blood corpuscles of ox blood. The concentration at which it just began to extrude was determined for various salts, and it was found that these concentrations were usually proportional to the molecular weights of the substances divided by certain whole numbers, which are the same as the isotonic coefficients of DE VRIES.

The work of MASSART was done chiefly upon bacteria. He made use of the fact demonstrated by PFEFFER (see p. 41) that substances, which at a low concentration attract bacteria chemotactically, at a higher concentration repel them. He found that, in general, the repulsions exercised by the various dissolved substances are proportional to their isotonic coefficients, when the solutions are made up as MW solutions. Thus, when a 10 MW % concentration of a substance with isotonic coefficient 2 just begins to repel bacteria, a substance which just begins to repel in a 5 MW % concentration has an isotonic coefficient of 4.\*

## § 2. EFFECT OF VARYING DENSITY UPON THE STRUCTURE AND GENERAL FUNCTIONS OF PROTOPLASM

Under this head we may consider, (*a*) the effect upon the general structure of protoplasm; (*b*) the modification of general functions, and (*c*) the production of death.

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\* Starting from the observations of PFEFFER and DE VRIES the modern school of physico-chemists has greatly extended our knowledge of solutions. As a result of their work it appears that the validity of DE VRIES' law will not hold strictly for all solutions at all concentrations. For the number of effective particles in every solution of electrolytes, namely, of salts, bases, and acids, is greater than the number of molecules put into the solution; because a certain proportion of the dissolved molecules break up or *dissociate* into their constituent *ions*, and the osmotic pressure is determined by the number of both molecules and free ions in the solution. In the case of sugar, the alcohols and non-electrolytes in general, no dissociation occurs. In a normal solution of potassic chloride, on the other hand, 75.5% of the molecules dissociate, each forming two free ions. Since 24.5% of the molecules are intact and there are 151 free ions percent of the molecules introduced, the total number of molecules and free ions in the solution is 175.5% of the molecules introduced and the osmotic effect of a normal solution of KCl is 1.755 times that of a normal solution of sugar. The percentage of molecules of any electrolyte, as for instance KCl, which dissociate in solution increases as the strength of the solution diminishes, eventually becoming 100. Thus, in one-half the normal solution, 78% of the molecules of KCl dissociate; at 0.1 times the normal solution, 86%; at 0.01, 94%; at 0.001, 98%. Also, the percentage of molecules dissociated in normal solutions of different electrolytes varies. Thus, in such a solution of NaCl, 67.5% of the molecules are dissociated; of LiCl, 61%; of CaCl<sub>2</sub>, 53% (each into 3 ions); of MgCl<sub>2</sub>, 40%; of KI, 79%; of MgSO<sub>4</sub>, 19%; of Na<sub>2</sub>SO<sub>4</sub>, 35.6% (each into 3 ions); and so on. Valuable and extensive tables for the determination of the percentage of dissociation at different concentrations will be found in WHEATHAM, '95.

a. Since a protoplasmic mass is bounded by a film, permitting osmosis, it is clear that its characters may be greatly altered by varying the degree of concentration of the solution in which it lives; and we have already seen that they are so altered.

When plant cells, with a rigid cell-wall, are put into dense solutions, the water is drawn from the protoplasmic sac which, contracting, is torn from the cell-wall. The salt solution penetrates through the latter, but cannot enter the bounding plasma-film, which continues to contract around the diminishing globule of water until only a small ball remains.

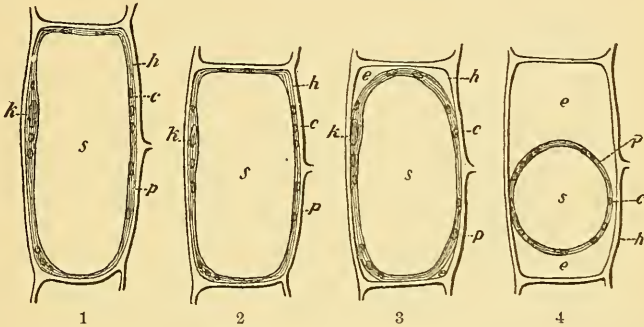


FIG. 8.—1. Young, not more than half-grown, cells from the cortical parenchyma of *Cephalaria leucantha*. 2. The same cell in a 4% solution of potassium nitrate. 3. The same cell in a 6% solution. 4. The same cell in a 10% solution. 1 and 4 from nature, 2 and 3 diagrammatic, all in optical longitudinal section. *h*, cell membrane; *p*, lining layer of protoplasm; *k*, cell nucleus; *c*, chlorophyll bodies; *s*, cell-sap; *e*, salt solution which has penetrated within the cell-membrane. (From SACHS: Pflanzenphysiologie, after DE VRIES.)

Put into pure water, on the contrary, the protoplasmic sac becomes distended, provided the cell sap contains an appropriate solution, generally a plant-acid. Thus turgescence is brought about.

The same effect of varied density upon the structure of protoplasm is observable among animals also. Thus, KÜHNE ('64, p. 48) and CZERNY ('69, pp. 158, 161) found that *Amœba* shrinks into a spherical mass when put into a 1% to 2% NaCl solution, and, when returned to fresh water, swells. Also, the character of the pseudopodia of *Amœba* and *Myxomycetes* changes. They become more numerous and attenuated, so that

the whole form of the organism has been likened to a horse-chestnut with its shell on. (KÜHNE, '64, pp. 48, 83; CZERNY, '69, p. 159.) ZACHARIAS ('84, p. 254, and '88) has described a similar phenomenon in the spermatozoon of *Polyphemus pediculus*. When put into a 3% NaCl solution the spermatozoa lost their cylindrical form and protruded long pseudopodia. A remarkable fact about the pseudopodia, moreover, was that in locomotion they were used like flagella. Likewise, FABRE-DOMERGUE ('88, p. 102) and MASSART ('89) have observed that the protoplasm of encysted Ciliata swells or contracts according as it is placed in a less or more dense medium; the cyst thus being perfectly permeable by water. MASSART has, indeed, obtained a rough quantitative expression of this statement, which is given in Tables X and XI, p. 87. HAMBUR-

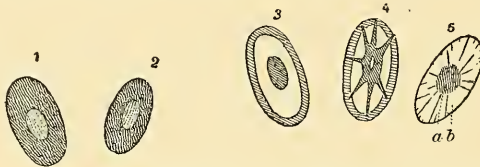


FIG. 9.—Blood corpuscles of the frog. 1, 2, normal; 3, 4, 5, various degrees of plasmolysis by solutions. *a*, nucleus and shrunken plasma; *b*, water-filled spaces. (From HAMBURGER, '87.)

GER ('87) has found that dense solutions produce the same modifications upon blood-corpuscles (see Fig. 9).

Again, GRUBER ('89) has found those individuals of the heliozoon *Actinophrys sol* which live in fresh water different from those which live in the sea, and he has produced that difference artificially. In the marine variety the plasma is dense, granular, free from vacuoles; while that of the fresh-water kind is extraordinarily rich in vacuoles, and has even a foamy appearance. If a marine form is gradually accustomed to fresh water its protoplasm soon acquires a vacuolated structure which renders it indistinguishable from the fresh-water one. GRUBER also accustomed fresh-water *Actinophrys* to sea water, when it acquired the structure of the normal marine form. Likewise the marine *Amoeba crystalligera*, which has a dense protoplasm, becomes vacuolated after being accustomed to fresh water. Also, SCHMANKEWITSCH ('79) has found that when the fresh-

water flagellate *Anisonema acinus*, BÜTSCHLI, is cultivated for many generations in water to which sea salt is gradually added, its structure is modified with the increasing density. The individuals become smaller and their feeding canal is not well-formed. Another change, which has been studied only in Vertebrates, is loss of weight. BERT ('71) found that a goldfish plunged into sea water loses 67% of its weight, and that young eels lose 10% to 17%. This fact also is clearly what we should expect from the theory of action of solutions, according to which the weak solutions of the body cavity should lose water. Thus, the changes produced in the structure of protoplasm by more or less dense solutions are chiefly the results of osmosis.

*b.* Among the functions of protoplasm, general movements (with locomotion) and excretion seem to be most markedly affected by density. Thus, KÜHNE ('64, p. 48) found that, when first subjected to a 1% NaCl solution, the movements of *Amœba* became more lively for a moment. ENGELMANN ('68, p. 343) noticed the same acceleration in movement in the cilia of the epithelium lining the frog's œsophagus when subjected to pure water—hence, to a weaker solution than the normal cell fluid. Even after death, fresh water causes a transitory activity in the cilia. In all cases, after a minute or two (1% solution) the movements begin to diminish, until at last they cease. This cessation of movement, whether due to loss or imbibition of water, is not necessarily death. For, if the abnormal concentration has not acted for too long a time, the movements return when the protoplasm is placed again in its normal fluid. (KÜHNE, '64, p. 48; ENGELMANN, '68, p. 343.)\* At a certain

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\* A similar cessation of movement occurs when the lower organisms are subjected in water to very great pressures. Experiments upon this phenomenon have been made chiefly by REGNARD ('84, '84a-'84d, and '86), CERTES ('84, '84a), and ROGER ('95). REGNARD was able, by the use of a special apparatus, to subject beer yeast, in water, during 1 hour, to a pressure of 1000 atmospheres (about 1000 kilograms per sq. cm.). When yeast so subjected was then placed in sugared water, it showed at first no activity. It was not dead, however, but had fallen into a latent life; for 1 hour after it had been relieved from pressure it revived and fermentation set in. Some algæ, Infusoria, and actinians, subjected to 600 atmospheres during 10 to 60 minutes, or to 300 atmospheres for 24 hours (CERTES, '84), exhibited a similar temporary rigor. Likewise muscle at 200 to

strength, however, varying for different individuals (ENGELMANN, CZERNY), death rapidly ensues. Thus *Amœba* quickly breaks up in a 10% solution, and the ciliated epithelium of a frog's throat in a 2.5% solution. Four phases in the action of concentrations may thus be observed: stimulation, retardation, density-rigor, and death. Even in concentrations at which motion is not entirely inhibited, locomotion may be interfered with. Thus, RICHTER ('92, pp. 37-40) found that while normal *Tetraspora* swarm-spores move at the rate of about  $60\mu$  per second, or else rotate about 100 times per minute, those in an 11% solution hardly move from their place, or sometimes move one-eleventh as fast as the normal swarm-spores. While it is possible that the dense water affords a mechanical obstacle to locomotion, it seems more probable that it is the general diminution of activities which causes the slow migration.

The modification of excretion by abnormal concentrations has been studied especially by ROSSBACH ('72). This experimenter worked upon fresh-water Ciliata (which alone possess a contractile vacuole) by subjecting them to a 0.5% solution of NaCl. The contractile vacuole became diminished almost to invisibility, and the interval between contractions was increased. In a 1% solution of sugar a reduction in size of the contractile vacuole occurred, but this was not so marked as in the case of the 1% NaCl solution. This is what we should expect according to theory, for the number of molecules in a 1% solution of sugar (mol. wt., 342) is much less than in a 1% solution of NaCl (mol. wt., 58.5), and their relative osmotic action is as  $\frac{2}{3 \times 342} : \frac{3}{3 \times 58.5}$ , or as 0.002 : 0.017, or as 2 : 17.

The phenomenon of contracting vacuoles seems not to be confined to Protozoa. It occurs in the embryos of some Mollusca, especially the stages of fresh-water Pulmonates, upon which my friend, Dr. KOFOID, performed some density experiments. The early cleavage and blastula stages of many fresh-water Pulmonates contain a central fluid-filled vacuole, which

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300 atmospheres loses its contractility, and at 400 atmospheres becomes rigid and hard. It also increases immensely in weight by the addition of water. ROGER pointed out that, subjected for 2 minutes to a pressure of 3000 kilograms per sq. cm., certain bacteria (*Streptococcus*) are even killed.



undergoes periodic enlargement and discharge as in the case of the contractile vacuole. KOFOID ('95, p. 104) found that when eggs of *Physa* and *Amnicola* were placed in a 0.19% or 0.10% NaCl solution, the contents of the central cavity, once extruded, were not so quickly restored as in the control eggs, and that the maximum volume attained by the vacuole in the salt solution was less than that attained in fresh water. For example, "the cavity of the control eggs attained a diameter of 5 to 7 units, while that of the eggs in the salt solution was only 3 to 4 at the time of elimination. There were, however, a very few cases in which the cavity reached a diameter of 5 units." Of interest is the additional fact that marine *Gastropoda* do not seem to have a "cleavage" cavity, but that this is confined to eggs developing in fresh water or moist situations.

The effect of density upon the higher animals is very complex, according to the observations of BERT ('71) upon the gold-fish. Plunged into sea water it shows violent, uncoordinated movements; then it becomes immobile, and rises to the surface by virtue of its relatively lower specific gravity.

The effect of fresh water upon marine organisms is equally striking, as GOGORZA\* ('91) has shown. They go immediately to the bottom and move with difficulty. Swimming animals swim badly if at all, and small fishes have to make much exertion to rise to the surface. The sensibility also undergoes great changes. Many animals soon become lethargic. Echinoderms and molluscs act as if anesthetized, since they do not respond as quickly as usual to external stimuli, and, finally, pass into complete paralysis. The action is slower upon Crustacea and fish; but here, too, fresh water acts as an anæsthetic. The respiratory movements become deep and rapid, bivalves extend their branchiæ, and Crustacea beat the water rapidly with their appendages in order to renew the supply. Animals ordinarily transparent, like medusæ, become opaque: first externally, then internally. The cornea of fish becomes opaque and the external slime coagulates. The tissues become swollen, so that soft-bodied animals are visibly deformed — in

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\* For an abstract of the work of GOGORZA, I am indebted to the kindness of Mr. F. C. WAITE.

fishes the eyes are forced out, the foot of gastropods swells, the blood corpuscles swell up and burst, and muscular tissue may increase as much as 6 times in volume. The enlargement of the different tissues is exhibited in the following table, which shows the percentage increase in weight and volume of organs of *Scyllium canicula*, placed in fresh water.

	AFTER 2 HOURS.		AFTER 24 HOURS.	
	% Increase in Weight.	% Increase in Volume.	% Increase in Weight.	% Increase in Volume.
Muscular tissue . . . .	15	50	20	200
Glandular tissue . . . .	12	0	19	100
Nervous tissue . . . .	20	50	60	250

All of these phenomena are clearly explicable upon the assumption of their production by endosmosis.

*c.* The pressure due to dense solutions may become very great, amounting, as I have said, to many atmospheres. So it is not surprising that a change of medium may rend cells or at any rate kill organisms. This result may be brought about either when the denser solution is inside of the body or outside; the former case is realized when marine animals are plunged into fresh water, the latter when fresh-water animals are plunged into solutions of salts. Studies upon the fatal effects of varying the concentration of solutions have been made by BERT ('66), PLATEAU ('71), COUTANCE ('83), RINGER and BUXTON ('85), DE VARIGNY ('88), MASSART ('89), GOGORZA ('91), and RICHTER ('92).

BERT'S ('66) studies were made upon marine fish which he plunged into fresh water. In a vessel holding 4.8 litres of fresh water, a mullet died in 44 minutes, and a Sparus in 86 minutes. Since the fishes lived longer in a sugar solution, BERT concluded that death was due to diminished density of the medium. In this conclusion he was nearer the truth than his immediate followers in this work.

PLATEAU'S ('71) observations were made upon all classes of Invertebrates, but especially upon Arthropods. He subjected

them to solutions both more and less dense than the normal, and determined their resistance periods. As a result of subjecting fresh-water animals to salt solutions, he found that their *resistance period diminished approximately as the thickness of the skin (and cuticula) diminished*. Thus, when plunged into sea water (3.046% salts) adult water insects resisted indefinitely; insect larvæ 6 to 4 hours; Entomostraca less than an hour; Nephelis, 5 to 7 minutes; Planaria, 4 minutes; and Hydra only 1 minute. GOGORZA ('91) got similar results, finding the resistance capacity of the different groups to diminish in this order: molluscs, crustaceans, fish, worms, tunicates, echinoderms, cœlenterates. So we may consider this relation between resistance period and thickness of covering a general law of resistance; and it is what we should expect upon the theory that the solutions act osmotically.

By subjecting organisms to separate solutions, each containing 3% of the various salts found in sea water, PLATEAU was able to show that NaCl produced the most important effect, MgCl<sub>2</sub> the next most important effect, and MgSO<sub>4</sub> still less. This is shown by the following —

TABLE IX

RESISTANCE PERIODS OF FRESH-WATER CRUSTACEA TO VARIOUS CONSTITUENTS OF SEA SALT. TEMPERATURE NOT GIVEN

(Numbers indicate minutes elapsing before death occurred)

SPECIES.	3% NaCl.	3% MgCl <sub>2</sub> .	3% MgSO <sub>4</sub> .	SEA WATER.
	MOL. WT., 58.5; I.C., 3; OSMOTIC INDEX, $\frac{3}{58.5}$ .	MOL. WT., 95; I.C., 4; OSMOTIC INDEX, $\frac{4}{95}$ .	MOL. WT., 120; I.C., 2; OSMOTIC INDEX, $\frac{2}{120}$ .	
Gammarus roeselii. .	105	131	520	230
Asellus aquaticus . .	155	1162	2000	160
Daphnia sima . . . .	7.8	19.5	87	22
Cyclops quadricornis	12.1	37	690	257
Cypris fusca . . . . .	26.7	223	460	36

Although from this table it seems clear that there is an inverse relation between resistance period and osmotic index, PLATEAU did not believe that the death of the animals experi-

mented upon was due alone to the osmotic action of the salts. To this conclusion he was led by an unfortunately devised experiment. He compared the action of several pairs of solutions, one of the members of the pair being a salt, and the other member sugar, the dissolved substance of each having the same gross weight. In all cases the action of the salt was the more powerful. But this is what we should expect upon the theory that death is caused by osmosis, since the osmotic index of sugar is far lower than that of any of the salts with which comparison was made.

Let us now ascertain the relation between the resistance period and the "osmotic index." To determine the *relative* resistance periods for any species in the different salts, we may take as our unit the average resistance period to all the salts, and express the separate resistance periods in terms of that unit. To determine the osmotic index, we divide the isotonic coefficient by the molecular weight. The resistance periods will vary inversely as the osmotic indices. For the salts, NaCl, MgCl<sub>2</sub>, MgSO<sub>4</sub>, the reciprocals of the osmotic indices are: 19.6, 23.8, 58.8; and the mean relative resistances are: 19, 63, 217. From this comparison it is seen that while the reciprocals of the osmotic indices increase roughly from 1 to 3, the relative resistance period increases from 1 to 11; or the resistance period increases more rapidly than the reciprocals of the osmotic indices, and roughly as the squares of those reciprocals.

At about the same time with PLATEAU'S work was published that of BERT ('71). The work of the latter was done chiefly upon fresh-water fishes; incidentally, upon frogs and some fresh-water Arthropoda. These were plunged directly into sea water, and their resistance periods determined. Some species showed an extraordinary variability in their resistance period; sticklebacks (*Gasterosteus leiurus*) from the same locality (about Paris) resisting for from 2 hours to 1 month or more.

A decided advance was made by BERT in observing that the resistance period varies with the temperature; thus, the European minnow (*Phoxinus lævis*) died in sea water—

at 9° C. in 30 minutes,	at 22° C. in 14 minutes,
at 14° C. in 25 minutes,	at 28° C. in 9 minutes.

Thus, in this case there is a diminution in the resistance period of approximately 1 minute for every degree of increase in the temperature.

Similar observations have been made by others. GOGORZA ('91, p. 242) finds that in all animals, at a low temperature, the resistance period is 2 to 3 times as long as at a high temperature.

In connection with these facts, it is to be noted that osmotic pressure increases with temperature, indeed, is proportional to the absolute temperature. (OSTWALD, '91, p. 114.) But as we are not able to say what relation exists between osmotic pressure and resistance period, we cannot say whether the above table agrees with the physical law.

Finally, we may discuss the question of the relation between the strength of the solution and the length of the resistance period. Data for this discussion are afforded by the extensive observations of GOGORZA. This author disclaims having found any mathematical relation, but his tables, properly treated, do show such a relation. The resistance periods depend upon so many factors that the times obtained by subjecting one animal to different concentrations of a salt cannot be directly compared with those obtained from another animal. It is the *relative* resistance periods only that can be thus compared.\* GOGORZA'S concentrations were obtained by subjecting marine animals to mixtures of marine and fresh water. No. 1 contained 100% sea water; No. 2, 75%; No. 3, 66%; No. 4, 50%; No. 5, 33%; No. 6, 25%; No. 7, 0%. Averaging the relative lengths of life of 22 species which died in 75%, or weaker percents of sea water, and comparing with the percentage of salts in various concentrations (the density of Mediterranean sea water being taken as 1.037), we get—

NO. OF SOLUTION :	1	2	3	4	5	6	7
% of salt in solution . . . .	3.7	2.8	2.5	1.9	1.2	0.9	0.00
Rel. resist. per. . . . .	Indef.	50.0	28.3	10.83	5.44	3.46	1.84
Log. of rel. res. per. . . . .		1.7	1.45	1.04	0.73	0.54	
Log. rel. res. per. $\times 1.7$ . .		2.9	2.5	1.7	1.2	0.9	

\* The relative resistance periods are calculated by the method described on p. 82.

The curve shown in Fig. 10 is constructed from the second and third lines of this table. The table shows that, within the limits of 2.8% and 0.9% concentration, the curve is a logarithmic one, *i.e.* as the ordinates increase the abscissæ increase as the logarithms of the ordinates.

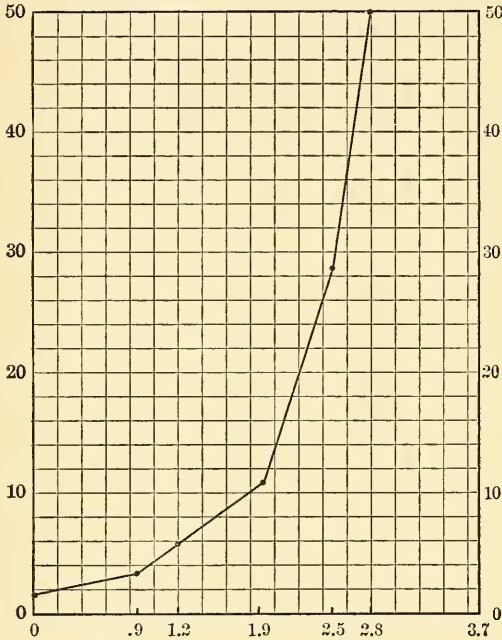


FIG. 10.—Curve showing relation between the percentage of salt in mixtures of fresh and salt water (abscissæ) and the mean resistance periods in hours of various organisms plunged therein (ordinates). Constructed from the table. (After data of GORZA, '91.)

In line 4 are given the (BRIGGS') logarithms of the numbers in line 3, and in line 5 these logarithms are each multiplied by a constant, 1.7, which gives a series of numbers closely similar to that of line 2. The relation between density and resistance period can thus be expressed by the equation

$$D = k. \log. R,$$

in which  $D$  stands for density;  $R$ , for resistance period; and  $k$  is a constant whose value depends upon the system of logarithms employed. This formula may be

transformed into the equivalent:  $R = e^{\frac{D}{k}}$ , in which  $e$  is the base of the NAPERIAN system of logarithms. Since the osmotic pressure is proportional to the concentration (p. 71), it follows also that  $R = e^{\frac{O}{k'}}$  where  $O$  stands for the osmotic pressure and  $k'$  for a new constant. The same relation holds when we compare the reciprocals of the relative resistance periods—or the relative rapidity of killing—and the absolute diminution of concentration.

### § 3. ACCLIMATIZATION TO SOLUTIONS OF GREATER OR LESS DENSITY THAN THE NORMAL

In the preceding section we saw that different organisms had a diverse resistance period to the same density of solution. In part, this may be accounted for, as we have seen, on the ground of a difference in the rapidity of osmotic action — thick-skinned animals resisting longer than thin-skinned ones. All diversity in the effect of solutions, cannot, however, be accounted for on this ground. Thus, the molluscs of the sea and those of fresh water appear to have an equally pervious epidermis, yet the former will, of course, withstand a much stronger solution of salt than the latter. This difference in resistance capacity seems closely correlated with the conditions of the medium in which the organism has been reared. Thus, BEUDANT ('16) found that littoral species (living, therefore, in a part of the sea where the water is much diluted by rivers), *e.g.* *Ostrea*, *Mytilus*, *Patella vulgata*, resist fresh water better than deep-sea species; and this discovery has been abundantly confirmed by DE VARIGNY ('88).\*

That the conditions of density of the culture medium determine the resistance capacity is proven by experiment, for, by varying the density of the culture solution, we may vary the resistance period of the individuals experimented on. BEUDANT ('16) was the first to show this. He used *Lymnea*, *Physa*, *Planorbis*, *Ancylus*, *Paludina*, and some other fresh-water Mollusca. He began in April by putting these organisms into a 1% NaCl solution, and, continuing to add salt slowly, by September many of these withstood a 4% solution — a solution which kills animals suddenly subjected to it. He performed likewise the reverse experiment upon marine Mollusca (*Patella*,

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\* The extremes of density in which organisms are capable of living are often considerable. On the one hand, the individuals of some species, especially fish, are able to migrate from fresh to salt water and back, with impunity. On the other hand, many species of a family, the other members of which are marine, have become accustomed to fresh water. Examples of this last case are the hydroid *Cordylophora lacustris*, the mollusc *Dreissena*, and the endoproctan bryozoan *Urnatella*. Likewise, some marine species have come to live in excessively salt water. Such, for example, is the case with *Artemia salina* which lives in Salt Lake, Utah, containing over 22% of salts. (LEIDY, '72, p. 165.)

Turbo, *Arca*, *Cardium edule*, *Mytilus edulus*, etc.) bringing them to live in fresh water by gradually diluting the medium.

PLATEAU ('71) gradually accustomed the fresh-water *Asellus aquaticus* to pure sea water, so that even in mixtures containing between 20% and 80% of sea water they laid eggs and produced a second generation. The second generation lived 108 hours in pure sea water, while *Asellus* freshly taken and plunged into sea water live only about 5 hours.

Not only the larger organisms, but also tissues and Protozoa may become acclimated. ROTH observed in '66 (p. 190) that cilia become "accommodated" to gradually increasing densities; ENGELMANN ('68, p. 343), however, denied, though without critical experiments, the validity of this conclusion for the case of the ciliated epithelium of the frog's throat. Later, CZERNY ('69, p. 161) succeeded in acclimating *Amœba* to a 4% solution of NaCl, although *Amœba* rarely resists 1% when suddenly subjected to it.

These early experiments have since been greatly extended, observations having been made upon nearly all groups of organisms — upon algæ, by RICHTER ('92); upon Myxomycetes, by STAHL ('84); upon *Actinospherium*, by VERWORN ('89, p. 10); upon bacteria, Flagellata, Ciliata, and Hydra, by MASSART ('89); upon Ciliata, by FABRE-DOMERGUE ('88); upon Crustacea, by PLATEAU ('71), SCHMANKEWITSCH ('75 and '77), and BERT ('83); upon the tadpoles of frogs, by YUNG ('85, p. 520); and upon representatives of almost all of the principal groups, by DE VARIGNY ('88) and GOGORZA ('91).

The aims and methods of these experimenters have been very diverse. Some have sought merely to illustrate how marine organisms may have come to live in fresh water, or the reverse. Such have usually made mixtures of fresh and salt water, the proportions of the one gradually increasing (DE VARIGNY, SCHMANKEWITSCH, GOGORZA), or they have added sea salt, dry or in solution, to the normal fresh-water medium of the organism (YUNG). MASSART, on the other hand, having in mind the more fundamental problem of the action of density upon protoplasm, has employed solutions of a single salt at a time — solutions, moreover, based usually upon the osmotic index of the salt as a unit of concentration.



Although there has been a gradual improvement in methods, the conditions other than that of concentration have too often been omitted from consideration. The omission of the temperature of the experiment solutions is especially unfortunate, for according to GOGORZA ('91, p. 270), acclimatization is more easily effected at a low temperature than at a high one.

Of the papers mentioned above, that of MASSART is especially worthy of extended notice from its quantitative nature. He subjected cysts of Ciliata to various concentrations of  $\text{KNO}_3$  and noted the effect upon the protoplasm. In the following tables, the first line of numbers names the solution in parts of the molecular weight expressed in grammes. The symbols in the columns headed by these numbers have the following significations: 0, no effect; *v*, the cysts possess a large vacuole whose pulsations are infrequent; *vp*, the vacuole is still prominent but plasmolysis is occurring; *p*, the plasmolysis is more marked and the vacuole is gone; *P*, the plasmolysis is so marked that the form of the infusorian is lost. The results given in the third and fourth lines were obtained from individuals acclimated for 22 hours to a 1.8 MW % and to a 3 MW % solution of  $\text{KNO}_3$  respectively. The observations were made immediately after immersion of the cysts. No mention is made of the temperature.

TABLE X—VORTICELLA

HUNDRETHS OF MOLECULAR WEIGHT.	0.5	1.0	1.5	2.0	2.5	3.0	3.5	4.0	5.0
Unacclimated . . . . .	<i>v</i>	<i>v</i>	<i>vp</i>	<i>vp</i>	<i>p</i>	<i>P</i>		<i>P</i>	
Acclimated to 1.8 % . . . . .				<i>v</i>	<i>vp</i>	<i>vp</i>	<i>p</i>	<i>P</i>	<i>P</i>
Acclimated to 3.0 % . . . . .						<i>vp</i>	<i>p</i>	<i>P</i>	<i>P</i>

TABLE XI—COLPODA

HUNDRETHS OF MOLECULAR WEIGHT.	0.5	1.0	1.5	2.0	2.5	3.0	3.5	4.0	5.0
Unacclimated . . . . .	0	<i>v</i>	<i>v</i>	<i>v</i>	<i>v</i>	<i>v</i>		<i>v</i>	
Acclimated to 1.8 MW % . . . . .				0	<i>v</i>	<i>v</i>	<i>vp</i>	<i>p</i>	<i>P</i>
Acclimated to 3.0 MW % . . . . .						<i>v</i>	<i>p</i>	<i>P</i>	<i>P</i>

If we take as our unit in Table X the concentration represented by  $v p$ , and in Table XI the concentration represented by  $v$ , we may conclude that the subjection for 22 hours to a 1.8 MW % or to a 3 MW % solution of the salt has given a resistance capacity of between 2 and 3 times the normal.\*

The question now arises, what is the cause of this increased resistance capacity? It is not merely apparent, resulting from the selection of the more resistant individuals, thus elevating the mean. It is clearly due to a diminution in the intensity of osmosis; and this must be due to the establishment of an equilibrium between internal and external osmotic pressures.

Now, this equilibrium can only be brought about by the density of the internal fluids becoming equal to that of the external medium; and this requires that the salt held in solution shall traverse the bounding protoplasmic films, gaining the interior. That such a traversing occurs has been argued by MASSART ('89), who has himself produced new evidence for this conclusion. As is well known numerous pigments in solution penetrate to the nucleus of the living protist. Potassic nitrate (JANSE, '87, p. 22), glycerine, and urea (DE VRIES, '88 and '89) have been observed to penetrate protoplasm.† That NaCl does the same thing has been shown by many observers. Thus, EMERY ('69) found that when a frog is placed in a salt solution and is left there for some time, then rinsed in water until no salt appears in the washings, and, finally, put into pure water, salt is given forth from the epidermis (precipitation on adding silver nitrate). Likewise PLATEAU ('71, p. 20) found that various fresh-water Arthropods reared in a salt solution excreted an unusual amount of salt; and FREDERIC ('85) has determined that the quantity of salt in the blood of *Carcinus* varies from 3.1% to 1.5%, according to the

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\* A few data concerning proper acclimatization cultures to NaCl may be found useful. To acclimate bacteria 0.003 to 0.009 MW % may be added daily; *Oscillaria*, 0.01 MW %, added monthly; *Anabæna* and *Tetraspora*, 0.018 MW %, added monthly; *Ciliata*, 0.003 MW %, daily; *Hydra viridis*, 0.001 MW %, daily for 6 days; *Tubifex*, 0.02 MW %, daily; tadpoles, 0.004 to 0.014 MW %, daily.

† A fact observed by BERR ('71) suggests that some solids are taken into the body in acclimatization; for, he says, fresh-water fishes acclimatized to sea water gain in weight, and when placed in fresh water fall to the bottom. The fact that they fall to the bottom indicates that their specific gravity is increased.

density of the salt solution in which it has been reared. Finally, MASSART has shown, by a new method, that several soluble organic compounds can permeate the bounding cell-film of Flagellata. Thus, if after permanently plasmolyzing *Polytoma uvella* by a 0.02 MW % solution of  $\text{KNO}_3$ , a 0.01 MW % solution of saccharose be added to the solution, the protoplasm soon regains its normal form, apparently by absorption of saccharose, since the cell-wall is impermeable to  $\text{KNO}_3$ . By the same method, potassium acetate, calcium butyrate, calcium phosphate, glycerine, ammonium tartrate, asparagine, glycose, sodium benzoate, salicin, and phloridzin can be shown to permeate the protoplasm of this flagellate. All these facts point to the conclusion to which physicists had arrived concerning dead animal membranes, that protoplasm admits the slow penetration of the dissolved salts, and thus effects the eventual equilibration of internal and external densities.

In conclusion, a word may be said concerning variability in capacity of acclimatization. The data afforded upon this subject by RICHTER ('92) are the most valuable. He was able to acclimatize *Tetraspora* to 16% (0.27 MW %) NaCl, while *Spirogyra* would not withstand, under like treatment, 0.5% (0.0085 MW %). It is clear then that, just as the resistance capacity varies, so also does the acclimatization capacity.

#### § 4. CONTROL OF THE DIRECTION OF LOCOMOTION BY DENSITY: TONOTAXIS

Three authors only, so far as I know, have concerned themselves with this phenomenon,—STAHL ('84), PFEFFER ('84, '88), and MASSART ('89, '91). STAHL ('84) observed that plasmodia of *Myxomycetes* withdrew from solutions either denser or less dense than the normal, and concludes that the action is not a simple, directly explicable one, but is rather a highly complicated irritability phenomenon. The observations of PFEFFER were incidental to his study of chemotaxis. He found that high concentrations of many substances acted repulsively, and he was at first ('84, p. 455) inclined to attribute this repulsion to osmotic action, but later ('88, p. 624) he believed this view disproved. The disproof he considered to lie in this, that

the repulsive quality varies with the quality of the substance — may occur even in substances which are not attractive at any concentration. PFEFFER is, therefore, inclined to regard strong, repelling solutions as acting in a different fashion from attractive ones. Just as strong sunlight may repel organisms attracted by weak light, — both phenomena being light phenomena, — so the repulsion and attraction of solutions may both be regarded as chemical phenomena.

The work of MASSART brought evidence against PFEFFER'S conclusions, and added many important data. His studies were made chiefly upon bacteria, to a less degree upon Flagellata, Hydra, the frog, and the human conjunctiva. The results of the studies showed that neutral solutions of a certain concentration repel, and that the repulsion is proportional to their isotonic coefficients and inversely proportional to their molecular weights, and, therefore, that the repulsions are purely osmotic phenomena.

The conclusions of MASSART thus summarized were obtained by the use of special methods, which gave quantitative results. So they are worth detailed consideration. A drop of liquid containing bacteria is suspended from the under side of a cover-glass in a moist chamber whose side walls are formed of cardboard, and whose top is the cover-glass. Into the drop, glass capillary tubes similar to those used by PFEFFER are introduced, filled with the solution whose action is to be studied. In addition to this solution all the tubes should contain  $\frac{5}{100000}$  MW % (0.00691 gr. %)  $K_2CO_3$  for the purpose of attracting the bacteria. When a tube containing only this dilute solution of  $K_2CO_3$  is put into the drop, bacteria crowd into it and literally fill it in from 20 to 30 minutes. But when a series of increasing solutions of a neutral salt like NaCl is added to the  $K_2CO_3$ , the organisms at first do not crowd in so rapidly, then remain at the mouth, and, finally, are repelled from the tube opening. MASSART has tabulated the results obtained with *Spirillum* upon using tubes containing different chemical substances in different degrees of concentration. One of these tables, in slightly modified form, is reproduced here. In this table, *A* indicates that the bacteria entered the tube readily; *a*, that they merely gathered about its mouth; *o*, that they

were repelled. The numbers at the heads of the columns are the different values of  $n$  in the formula,  $\frac{n}{1000}$  MW %. Since the different solutions were made up on the basis of molecular weights, all solutions of a given concentration contained the same number of molecules.

TABLE XII

ISOTON. COEF. 3.	1	2	3	4	5	6	7	8	9	10
NH <sub>4</sub> Cl . . . . .	A	A	A	a	a	o	o	o	o	o
NaCl . . . . .	A	A	A	A	a	a	o	o	o	o
KCN . . . . .	o	o	o	o	o	o	o	o	o	o
KCl . . . . .	A	A	A	A	a	a	o	o	o	o
NH <sub>4</sub> NO <sub>3</sub> . . . . .	A	A	A	A	a	a	o	o	o	o
NaNO <sub>3</sub> . . . . .	A	A	A	A	a	a	a	o	o	o
KNO <sub>3</sub> . . . . .	A	A	A	A	a	a	o	o	o	o
KBr . . . . .	A	A	A	A	a	o	o	o	o	o
KClO <sub>3</sub> . . . . .	A	A	A	A	a	o	o	o	o	o
KI . . . . .	A	A	A	A	a	a	o	o	o	o

From this table it appears that, as a rule, solutions of  $\frac{7}{1000}$  MW % and over are repelled, while those of  $\frac{4}{1000}$  or under, except in the case of KCN, permit the free migration of the bacteria into the tube.

In the case of those substances whose isotonic coefficient is 4, solutions of  $\frac{6}{1000}$  MW % and over always repel, and those of  $\frac{3}{1000}$  in the majority of cases permit free migration. In the case of those substances whose isotonic coefficient is 2, solutions of over  $\frac{10}{1000}$  MW % repel, and those of under  $\frac{8}{1000}$  usually permit free migration. The solutions at which repulsion just occurs in the three cases are in the ratio 10 : 7 : 6; which is nearly the same ratio as the reciprocals of the isotonic coefficients, which, multiplied by 2 run, 10 : 6.6 : 5. Thus the conclusion seems justified that the repelling action of these substances is proportional to their isotonic coefficients, and is, therefore, probably osmotic in its nature.

In a second work, MASSART ('91) has studied this matter with the aid of new methods. A drop of sea water containing bacteria is prepared as before, on a cardboard ring, but, in place

of a capillary tube containing a dense solution, grains of NaCl are placed at one point of the margin of the drop. These grains gradually dissolve, their molecules gradually diffuse through the drop, and as they do so the bacteria retreat before them, remaining in the zone of least concentration. Again, a drop of distilled water was placed alongside of the drop of sea water containing *Spirillum*, and the two drops were connected by a communicating canal of water (Fig. 11). As the dis-

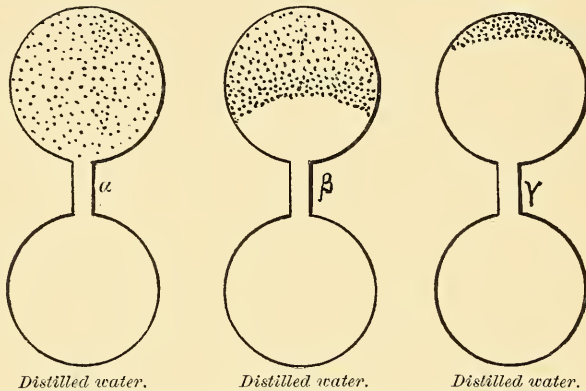


FIG. 11. — A drop of sea water connected with a drop of distilled water (in lower part of diagrams). The marine bacteria of the former retreat before the encroachment of the latter. (From MASSART, '91.)

tilled water mingles with the sea water at one mouth of the communicating canal, the bacteria retreat further and further from that mouth, keeping in the most concentrated part of the drop. Finally, when a drop of sea water is connected with one of distilled water, and granules of NaCl are placed in the drop of sea water, the bacteria, retreating from the zone of too great concentration penetrate into the drop of distilled water (Fig. 12), where they now find the proper concentration. Thus, *Spirillum* is sensitive both to solutions denser and to those weaker than the normal (hyperisotonic solutions and hypisotonic solutions, MASSART).

In summing up the observations of this section, we notice that some organisms (for MASSART found some non-sensitive bacteria) are sensitive to concentration. This sensitiveness is

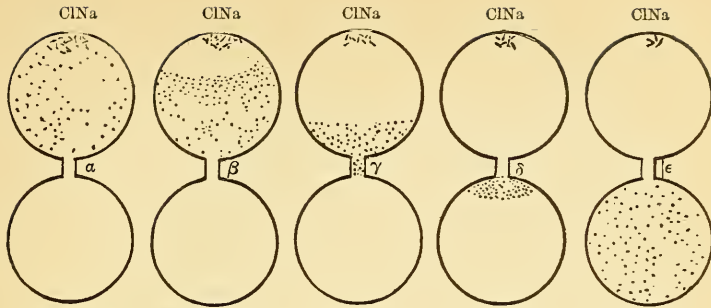


FIG. 12. — A drop of sea water joined by a canal with a drop of distilled water. The density of the sea water is being gradually increased in the successive figures by the dissolution of grains of salt placed at one edge. As the solution thickens, the organisms (marine Anophrys, represented by dots) retreat towards the distilled water. (From MASSART, '91.)

such that they are repelled by either hyperisotonic or hypisotonic solutions; only in a certain concentration do they come to rest. We may speak of these organisms as attuned to this concentration. Different organisms are attuned to different concentrations, and there can be no doubt that the degree of concentration to which they are attuned is determined by the past experience of the organisms, as the facts of acclimatization indicate.

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## CHAPTER IV

### *ACTION OF MOLAR AGENTS UPON PROTOPLASM*

THIS subject is so ill-defined that it is impossible to draw any line of distinction between contact on the one hand and a crushing pressure, or wounding, on the other. The molar agents may be solid or fluid. The methods of application may vary from a blunt contact or a sharp cut or puncture to the impact of flowing liquid. All these agents have this in common, however, that they act in a gross, mechanical way. The subject will be discussed under the following heads: (I) The effect of molar agents upon lifeless matter; (II) effect upon the metabolism and movement of protoplasm; and (III) effect in determining the direction of locomotion, — thigmotaxis (stereotaxis) and rheotaxis.

#### § 1. EFFECT OF MOLAR AGENTS UPON LIFELESS MATTER

Mechanical disturbance can induce in certain lifeless compounds violent chemical changes. Compounds which are so affected are preëminently unstable. This instability, however, varies greatly in degree. In some cases, the blow of a hammer is required to upset the molecules; the result being often a violent explosion. In other cases (*e.g.* chloride or iodide of nitrogen), the slightest touch of a feather suffices to produce an explosion. Now, most of the substances which explode upon impact, and which are used in the arts, are organic compounds, — fulminate, nitro-glycerine, gun-cotton, and picric-acid derivatives, — and therefore it is not surprising that we find the notoriously unstable protoplasm violently affected by contact.

Especially important for biology is the fact that undulatory motions and other *periodic* disturbances produce very important

molecular changes in chemical compounds. Certain substances have a specific rate of vibration, so that when this is reproduced by a vibrating cord or plate, explosion of the substance may occur. Iodide of nitrogen is one of these substances which is exploded by a high note. (CHAMPION and PELLET, '72, p. 212.) Upon this property of explosive compounds depends, apparently, the efficacy of "detonators," the explosion of a small quantity of which is capable of producing the explosion of a great mass of a second compound. Living protoplasm is, likewise, especially affected by periodic disturbances, and it is doubtless due to the peculiarities of its chemical structure that the auditory epithelium is so affected by sound waves in all their modifications of pitch, volume, and timbre.

## § 2. EFFECT OF MOLAR AGENTS UPON THE METABOLISM AND MOVEMENT OF PROTOPLASM

We shall first consider the effect on metabolism, and then on movement. The principal metabolic effects that will be considered are phosphorescence and secretion.

The *phosphorescence* of organisms is usually regarded as a slow combustion (oxidation) of organic substances. This chemical process is apparently accelerated by mechanical irritation, as every one must have noticed who has rowed a boat on a quiet summer's evening upon the sea. At every stroke of the oar, a gleam is sent along its length. An analytical study of this phenomenon has been made by MASSART ('93, p. 62). When a drop of water containing *Noctiluca* is put on filter paper, and the liquid is absorbed, there comes a moment when the surface film of the water flattens the spherical body of *Noctiluca*. At that moment of pressure light is emitted. If, however, the water is put into a slight vibration by a needle attached to a tuning-fork, and if the agitation is insufficient to deform the body, no light will be given forth. Deformation of the body, but not slight agitation, is, consequently, accompanied by those metabolic processes which result in the production of light.

Secondly, contact may induce the production and discharge of secretions. VERWORN ('89, p. 81) has called attention to this phenomenon in the cases of *Actinosphærium* and *Thalassi-*

cola. When Actinosphærium is subjected to a slight stimulation, such as would be produced by other Protozoa wandering among its pseudopodia, it shows no response. But when an infusorian or a rotifer swims against the pseudopodia with force, they discharge a sticky substance which holds the disturbing organism fast. The same result follows the irritation of one of the pseudopodia by touching it with a fibre of cloth or filter paper. Like effects follow the irritation of *Thalassicola*. Thus, some Protista respond to particular kinds of contact by the excretion of a sticky substance.

In the higher animals, also, contact may call forth secretions; thus, the stolons of many hydroids secrete a cement from the surface applied to the substratum.

Among the higher plants, also, contact has sometimes a similar effect. Examples appear in DARWIN'S ('75, p. 393) work on the gland cells of insectivorous plants. In many species, to be sure, *e.g.* *Drosera*, *Dionæa*, *Drosophyllum*, mere contact of inorganic bodies has no effect upon the secretions of the glands of the leaves. In the case of *Pinguicula lusitanica*, however, fragments of glass, as well as seeds and albumen, caused the glands with which they came in contact to secrete more freely than before.

This response to contact by secretion is, for the most part, an *advantageous* one. It enables the Protista and the insectivorous plants to hold their prey or their enemy, as the case may be; and it enables the stolon to hold fast to the substratum.

The change in metabolism may be so profound as to lead to death. HORVARTH ('78) and MELTZER ('94) have shown that when bacteria are violently shaken, not only is growth interfered with, as we shall see in the second part of this book, but death may ensue, so that cultures of bacteria may be sterilized.

We now turn to consider the *modification of movement* by molar agents. The general phenomena are familiar. An amœba, any other rhizopod, or a white blood corpuscle contracts when the cover-glass over it is disturbed. The streaming in the plasmodia of *Myxomycetes* is retarded or inhibited

upon shaking. When alga cells, such as those of *Chara* or *Vallisneria*, are freshly transferred to the slide, the disturbance causes cessation of movements (HOFMEISTER, '67, p. 50). When the stamen hairs of *Tradescantia* are crushed, the streaming of the plasma ceases. When *Chara* is cut across or punctured, rotation stops for a longer or shorter time (DUTROCHET, '37, p. 780). Even when a stem of *Chara* is pricked at the node by a needle, without penetrating into the cavity, movement ceases for a minute

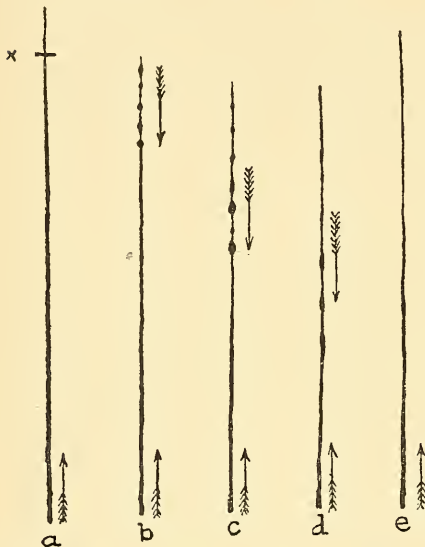


FIG. 13. — Pseudopodium of *Orbitolites*, retracting as a result of local stimulation. The arrows give the direction of the streaming of protoplasm. At the left is shown the beginning of the excitation; at the right, its end. (From VERWORN, '92).

or two. Thus, mechanical disturbance profoundly affects protoplasm.

Let us now consider more in detail the changes which take place in the protoplasm. VERWORN ('92, p. 24) has given us data on this matter. *Orbitolites* is a rhizopod having extremely delicate, filamentous pseudopodia. If one of these pseudopodia be cut across as at *x*, Fig. 13, *a*, the following changes occur:

the protoplasm lying next the cut directly collects into small spherical or fusiform masses which begin to migrate centripetally (Fig. 13, *b*). This movement meets with the normal centrifugally migrating plasm and turns the latter towards the centre again (Fig. 13, *c*). Gradually the thickenings elongate until, before they have reached the central body, they are no longer visible (Fig. 13, *d*). In about 2 minutes normal movements are completely restored (Fig. 13, *e*). Slightly different results are gained from *Cyphoderia* (Fig. 14). When the large pseudopodium of this organism is touched with a needle near its distal end, it thickens (as in the case of *Orbitolites*) and

the thick region, together with all the proximal lying protoplasm, begins to flow towards the centre. The whole plasma thread retracts.

Again, if an individual of *Diffugia* (Fig. 15) be slightly shaken, the pseudopodium contracts into the shell; if it be

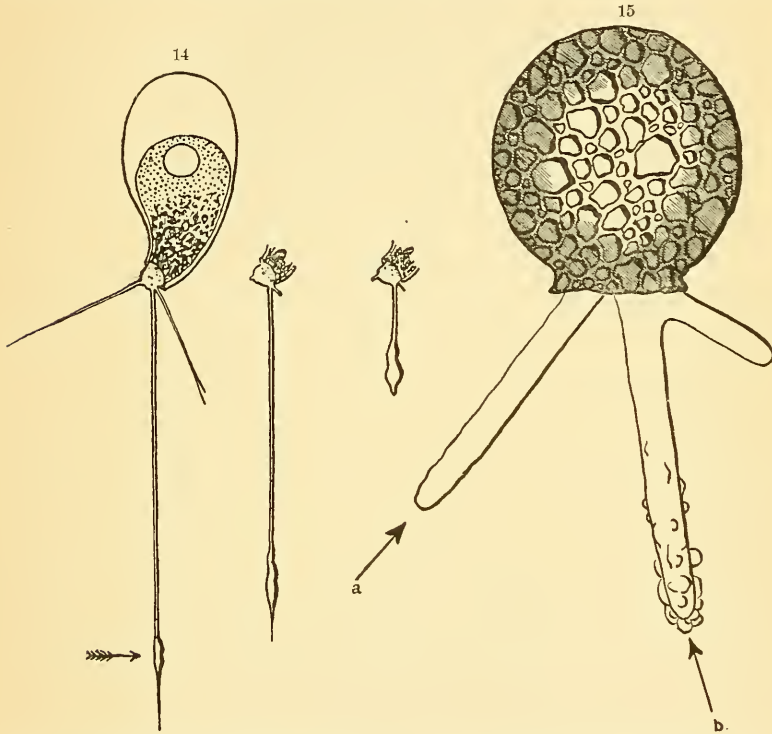


FIG. 14. — *Cyphoderia margaritacea*, showing the retraction of its pseudopodium as a result of irritation at the point indicated by the arrow. (From VERWORN, '92.)

FIG. 15. — *Diffugia urceolata*: at *a*, stimulated by a weak local irritation; at *b*, by a somewhat stronger one. (From VERWORN, '89.)

violently shaken, the following changes occur: drops of a less highly refractive substance seem to gather on the surface of the filamentous pseudopodium and unite to form a sheath surrounding a more highly refractive axis. At the same time, axis and sheath retreat into the central mass. In this case, then, we have a segregation of dissimilar protoplasmic substances, and a tendency to collect about centres along the

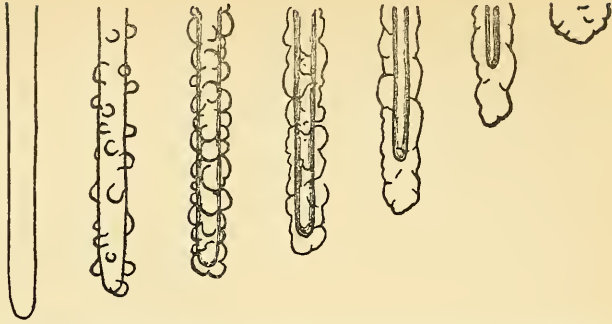


FIG. 16.—A series showing seven phases in the contraction of a pseudopodium of *Diffugia lobostoma*, following total stimulation. The series passes from left to right. (From VERWORN, '92.)

pseudopodium and in the whole mass. The same thing is seen in the widely dissimilar *Actinosphaerium* (Fig. 17). Here is

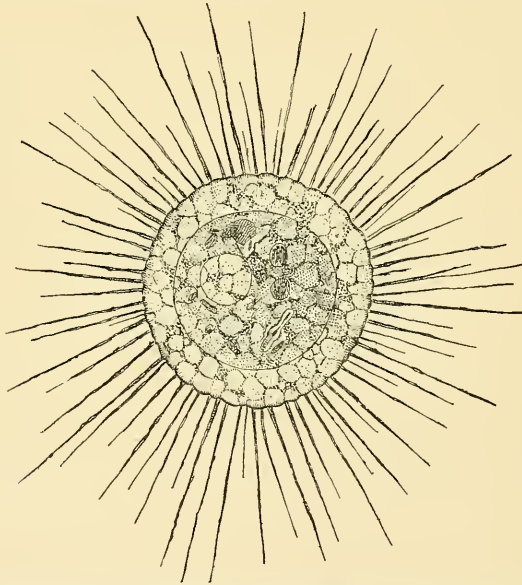


FIG. 17.—*Actinosphaerium Eichhornii*, unirritated. Natural size about 0.5 mm. (From VERWORN, '89.)

especially noticeable (Figs. 18, 19) the tendency to produce fusiform or spherical aggregations, and to retract the pseudopodia. So, too, in the irritated stamen hairs of *Tradescantia*;



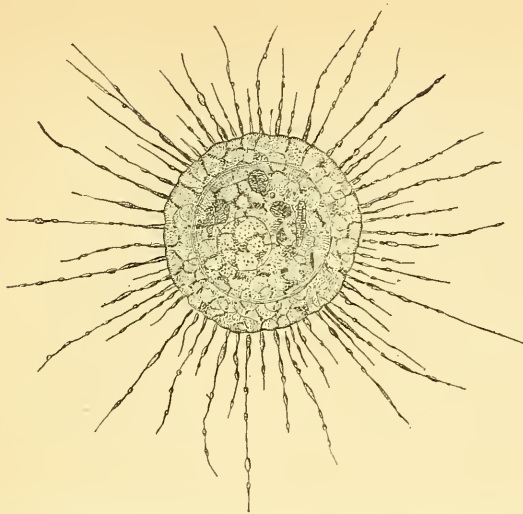


FIG. 18. — *Actinosphaerium Eichhornii*, at the beginning of irritation. The protoplasm is accumulated along the pseudopodia in drops and spindles. (From VERWORN, '89.)

says HOFMEISTER ('67, p. 50), "The threads become knotty, tear apart, draw together into short clubs or balls, and fuse

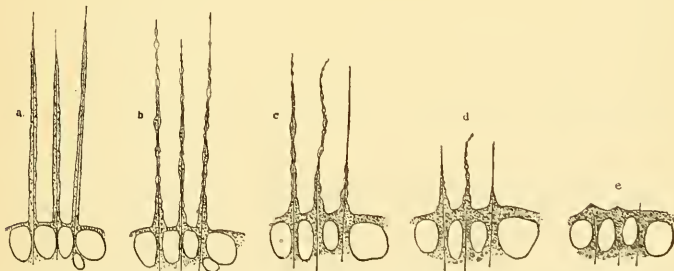


FIG. 19. — Three pseudopodia of the same individual, much enlarged. *a*, normal condition; the axial thread is seen, surrounded by protoplasm. *b*, the pseudopodia at the beginning of stimulation. *c*, *d*, the stimulation is continuing, and the axial thread is shortening. *e*, the three pseudopodia are almost completely retracted. (From VERWORN, '89.)

partly with the collection of protoplasm lying about the cell-nucleus and partly with the peripheral protoplasmic layer."

These similar phenomena from various organisms are fundamental; how are they to be interpreted? It is well known

that non-vital semi-fluid substances tend to assume a spherical form by virtue of the property of surface tensions. That protoplasm does not always assume this form is due to special causes. When a Protist or one of its pseudopodia is irritated by contact, it tends to assume a spherical form or a thread tends to aggregate into spherical drops. It seems probable, we cannot say more than that, that this aggregation is due to a diminution in the activity of those causes which oppose the action of surface tension; and so the latter reasserts itself. It is likewise possible that new attractive centres arise. That a thread should break up into drops indicates, moreover, a loss in cohesion. Loss of cohesion, formation of new centres of attraction, and diminution of the form-maintaining forces,—these seem to be the effects of contact. They must be due to the chemical changes wrought by contact.

The changes just referred to constitute the essence of contraction, a phenomenon of widespread occurrence not only among Protista, but among the higher plants and animals; for example, in the sensitive plant and in Vertebrate muscle. Into these contraction phenomena which follow contact in the higher organisms we cannot go; their study belongs to the field of plant and animal physiology. At bottom, however, we must believe many of these phenomena in the higher organisms to be due to the same causes as contraction in Protista.

A few words concerning rhythmically repeated disturbances. A single disturbance gives rise, as we have seen, to a series of phenomena producing contraction; but in a few seconds the effects of the disturbances are past and the protoplasm returns to its uncontracted form. If, however, the shock is repeated before relaxation has fully occurred a new contraction is superimposed on the first, and the resulting contraction is more violent than a single one. If now shock follow shock in quick succession, a violently contracted condition, known as tetanus, results. Under the condition of tetanus the amœba becomes a spherical mass, *Actinosphærium* retracts all of its pseudopodia, a branching *Carchesium* stock forms a little ball, and muscle fibres are greatly shortened. In a word, rhythmically repeated shocks are accompanied by an exaggeration of those changes which result from a single shock.

§ 3. EFFECT OF MOLAR AGENTS IN DETERMINING THE DIRECTION OF LOCOMOTION—THIGMOTAXIS (STEREOTAXIS) AND RHEOTAXIS \*

We have already seen that when a pseudopodium of an amœba is touched by a solid body it retracts. In this retraction the centre of mass is transferred to a new point. If the stimulation is often repeated upon the same side, contraction continues on that side, until eventually the amœba will have migrated a considerable distance. In this case the determination of the direction of locomotion is closely allied to the phenomena of contraction as a result of stimulation, considered in section 2. The retraction of the protoplasm which follows its irritation is the cause of the migration of the amœba in a definite direction. This direction is away from the touching body. The response may consequently be called negative thigmotaxis.

The phenomenon of negative thigmotaxis is widespread. There are almost no free-moving organisms which do not move away from contact or molar disturbance of an unusual or violent sort. Thus you may very definitely control the direction of movement of a planarian or a slug by touching the body upon the side opposite the direction in which you wish it to move. In such cases, also, there is first a contraction of the body upon the irritated side.

The opposite phenomenon of movement towards, or clinging to, the irritating body — positive thigmotaxis — is less common and therefore more striking. It has long been known, I imagine, — it certainly is an observation easily made, — that an amœba which has come in contact with a solid body clings close to it and moves over its surface. LE DANTEC ('95, p. 211) has described the action in much detail. An amœba descending in the drop touches the glass slide first by a single protruding pseudopodium. Next, the pseudopod elongates horizontally, and at the same time affixation takes place, so that the organism does not roll about when the water is agitated. The

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\* Thigmotaxis, under the different form "thigmotropism" (from *θίγμα*, "contact") was first applied to these phenomena by VERWORN ('89, p. 90); stereotaxis, under the form "stereotropism" (from *στερεός*, "solid"), was introduced by LOEB ('90, p. 28), and is practically synonymous with thigmotaxis.

pseudopod gradually extends itself, and new ones are formed, until at last the whole substance of the amœba is spread out parallel to the glass, over whose surface it moves. That there is a considerable adherence is shown by the fact that the amœba is not disturbed by an appreciable current. If, however, it is made to contract, it loses its hold at once.

Very similar phenomena occur, according to VERWORN ('95, p. 429), in *Orbitolites* also. Such an organism lying in a watch glass begins to send out pseudopodia which, so long as they move free in the water, are simple straight threads; but when they touch the glass they adhere to it, stream out along it, and send out branches. In these *Rhizopoda*, consequently, the presence of a solid body is a stimulus to the spreading out of the pseudopodia and to those changes by which close adhesion is effected.

We now pass to the other simple organisms. Among Infusoria, PFEFFER ('88, pp. 618-621) has found that *Glaucoma scintillans* and, to a less degree, *Colpidium colpoda*, *Paramecium aurelia*, and *Stylonychia mytilus* aggregate about solid bodies in the water, such as fragments of soaked filter paper or particles of barium sulphate. Since these cannot supply oxygen or soluble substances, the effect produced is doubtless due to contact.

The aggregated organisms tend, in moving, to keep upon the surface of the solid. Thus PFEFFER ('88, p. 619) found that *Urostyla weissii*, coming in contact with glass threads, moved along them on their ventral surfaces; and MASSART ('91) observed some *Chlamydomonades* remain hanging to objects with which they came in contact. VERWORN ('95, p. 431), likewise, finds that *Oxytricha* travels over the surface of *Anodonta* eggs or particles of detritus which it happens upon in the water. In one instance, the organism ran for some time over the surface of an egg of *Anodonta* without being able to leave it. After four hours, it was able, by the aid of a piece of slime which came in contact with the egg, to free itself from that body.

Phenomena similar to the above-described for bacteria and Infusoria are found in spermatozoa also. DEWITZ ('85 and '86) first noticed this in the case of the cockroach, *Periplaneta*

orientalis. When an 0.8% or 0.9% NaCl solution containing spermatozoa was put under a cover-glass, the spermatozoa arranged themselves in two layers, one in contact with the cover-glass, the other in contact with the slide. By isolating some of the spermatozoa at the upper surface and putting them under a cover-glass, he found that they likewise distributed themselves at both upper and lower surfaces. Hence the segregation into two layers was not due to a difference in kind between the spermatozoa occupying the two positions, but to the fact that there were here two surfaces of contact, separated

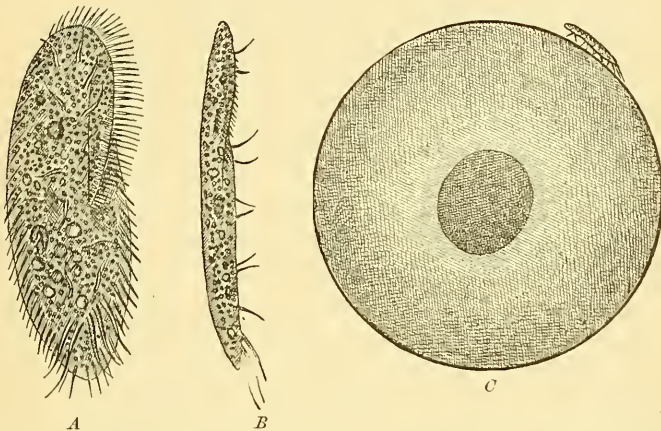


FIG. 20.—*A*, *Oxytricha* seen from below; *B*, from the side; *C*, crawling over the egg of *Anodonta*. (From VERWORN, '95.)

by a water-film. If a spherical grain be placed in the drop of water, aggregation takes place about that also. A similar experiment, with similar results, was made by MASSART ('88) with frog spermatozoa. Here, too, the active spermatozoa kept in contact with the upper and lower glass surfaces, whilst the weak forms lay midway between. The fact that only active spermatozoa show this tendency to keep in contact with solids, indicates that we are here dealing with irritability to contact.

The quality of the surface influences its capacity for stimulating to positive thigmotaxis. Thus, while mere roughness has no effect, if the surface of glass be smeared with a slimy mass, so thick that the spermatozoa can hardly penetrate it

they may no longer cling to the glass, but wander, undirected, through the water. Again, while the surface film of water often acts thigmotactically, if the surface tension is reduced by a thin covering of oil, it no longer holds the organisms. It would seem that a certain minimum difference in rigidity, between any surface and the medium, is necessary in order that the surface should act thigmotactically.

Once in contact with a sufficiently attracting surface, the organism may move to and fro over it, but it can hardly leave it. It is, as DEWITZ ('86, p. 366) says, as though the spermatozoa were attracted by a magnet. This close adhesion of the organism to the irritating surface is a remarkable phenomenon. LE DANTEC ('95) suggests that the amœba adheres to the glass by *molecular attraction*. On the other hand, it may be doubted whether the close adhesion signifies anything else than the absence of a sufficient stimulus to leave the surface of contact.

When an organism has been stimulated by contact for some time, it at last becomes changed so that it no longer responds as it did at first. Thus Dr. W. E. CASTLE has informed me that he has seen a colony of Stentors, in an aquarium, being constantly struck by Tubifex waving back and forth, yet the Stentors did not contract as they usually do when struck. PFEFFER ('88, p. 619) has observed that Urostyla retreats, after a time, from the surface with which it was in contact. These facts indicate that protoplasm can become acclimatized to contact so as to be no longer stimulated by it.

We now turn to the consideration of *Rheotaxis*, which may be regarded provisionally as a form of thigmotaxis, although the possibility of its being rather a case of chemotaxis is not excluded.

ROSANOFF ('68) was the first to notice the rheotaxis of the large plasmodium of *Æthaliium septicum*, but he ascribed it to geotaxis. The correct interpretation was first given by STRASBURGER ('78, p. 62), and has been confirmed by JÖNNSON ('83), and STAHL ('84). When *Æthaliium* is placed on a strip of saturated filter paper, the upper end of which is dipped in a beaker of water, it is subjected to a current of water in the substratum. At the same time it moves

against the current. The current controls the direction of locomotion.

The evidence that it is indeed the current is partly gained by exclusion. It cannot be geotaxis, for if the current is flowing upwards on any arm of the strip, the plasmodium flows down. It can hardly be hydrotaxis, for the strip is uniformly saturated throughout. The action of light may be excluded by shutting the whole apparatus in the dark, when the same response occurs. When the direction of the current in the strip is reversed, the movement of the plasmodium is reversed also. Thus no other cause will explain the result but that of the moving water.

Satisfactory evidence that it is the current as such which acts will not be forthcoming until it has been shown that other fluids than water, *e.g.* oil, provoke a similar response. Until such an explanation has been tried, it must remain uncertain whether the phenomenon is not perhaps due to a difference in the quality of the afferent and the efferent water.

Finally, it must be mentioned that higher organisms, especially fish, are rheotactic. Whoever has seen fish ascending streams from the sea in the spring has had this vividly impressed upon him. Before some dam thousands of fish will be seen, all facing the torrent of water against which they can hardly hold their own. It is the current which determines their position. They are responding to the direction of flow of the waters.

To recapitulate: In many non-living substances, especially organic compounds, violent chemical changes (explosions) are brought about by contact and especially by repeated vibrations. So, too, in protoplasm, chemical change, exhibiting itself in modified metabolism, frequently follows contact. The explanation adapted to the non-living series of phenomena is adapted to the living series also,—the molecules of the substance are complex, loosely associated, very unstable, so that even a slight mechanical disturbance will serve to dissociate their atoms. Protoplasm is a mixture of so many substances that the whole mass does not become changed at once; but continued stimulation may eventually produce such widespread changes as to lead to death. One of the most evident

results of contact upon protoplasm is modification of movement, — momentary quiet, followed by contraction. Rapidly repeated shocks lead to a summation of responses called tetanus. Slowly repeated shocks may lead to acclimatization to contact. Finally, the direction of locomotion is in some cases controlled by contact; many organisms move from the touching body — negative thigmotaxis; others may face the impact of flowing water or keep close, as though attached, to the rigid surface — positive thigmotaxis. If the changed chemical condition following contact be called the “response,” then all changes wrought by contact on protoplasm may be considered as responses.

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## CHAPTER V

### *EFFECT OF GRAVITY UPON PROTOPLASM*

WE shall consider this subject under three heads: (I) Methods of Study; (II) Effect of Gravity upon the Structure of Protoplasm; (III) Control of Locomotion by Gravity — Geotaxis.

#### § 1. METHODS OF STUDY

Under normal circumstances gravity acts upon organisms continuously, uniformly, and in one direction only at a time. In this respect it is widely different from most of the agents

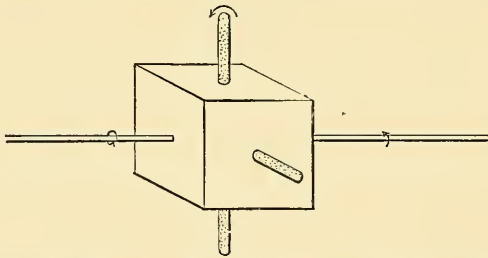


FIG. 21. — Diagram of the essential part of a klinostat. A rotating block or drum, to which tubes containing the geotactic organisms may be attached in the position indicated.

which we have to consider. Since its action is uniform it can be varied only in an indirect way; *i.e.* by turning the organism or by replacing gravity in part by a force working in another direction. One of the simplest ways of turning the organism so as to eliminate gravity is by means of the klinostat (Fig. 21). This is made in various forms, and consists essentially of a horizontal rod supported near the ends and made to revolve about its long axis by clockwork. Towards the middle of the rod, or at one end, is rigidly affixed a block to which may be

fastened, radially, the vessels containing the objects of experimentation. When the rod revolves, all sides of the object are brought successively and equally under the influence of gravity's pull. By this means the *directive* action of gravity is eliminated.

In the case of organisms living in water, the effect of gravity may be overcome by the buoyancy of the medium. It is clear that an organism floating in a medium of its own weight cannot be affected by gravity. This condition can be brought about by increasing the specific gravity of water by adding soluble substances such as gelatine and gum arabic. Since the specific gravity of the organism tends gradually to change with that of the medium, this method does for rapid experiments only.

In dealing with larger organisms, which, like slugs, can keep affixed to glass or other smooth surfaces, the inclination of the surface may be varied from a vertical position to a horizontal one, thus varying the active component of gravity. Finally, gravity may be replaced by centrifugal force by rapidly rotating either about a horizontal or a vertical axis. By varying the rate of rotation the centrifugal force will vary, in accordance with the formula,  $f = \frac{2\pi^2r}{t^2}$ , in which  $r$  is the rotating radius (in meters) and  $t^2$  the square of the time of a rotation (in seconds). This varying centrifugal force will act exactly in the same way as gravity, only *from* the centre of rotation.

## § 2. EFFECT OF GRAVITY UPON THE STRUCTURE OF PROTOPLASM

Very few observations have been made upon this subject, and yet indications are not wanting that the field would well repay working. Thus, where the cell contains specifically heavier and lighter substances the two will be separated by the action of gravity. This occurs in plant cells in which, according to DEHNECKE ('80), various contained bodies, *e.g.* chlorophyll granules and starch grains, tend to sink to the lower side of the cell. This result is produced in from a few minutes to several hours. This effect is likewise seen in many

ova in which the yolk sinks to the lower pole and the protoplasm floats on top, in whatever position the egg may be held. This fact undoubtedly has an important effect upon development, as we shall see later.

Of the specifically heavier bodies above referred to, the nucleolus is a striking example, as HERRICK ('95) has recently shown. Thus, when the ovary of a lobster is killed, the nucleoli of all the nuclei are found in contact with that part of the nuclear membrane which was the lowest at

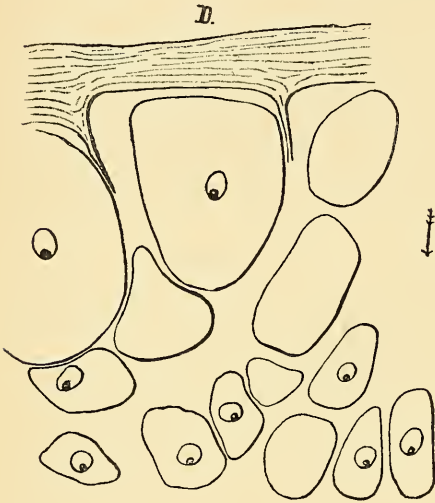


FIG. 22. — Section through the ovary of a lobster hardened with its dorsal surface (*D*) uppermost. The nucleoli lie against the ventral surface of the nucleus. Magnified 50 diameters. (From HERRICK, '95.)

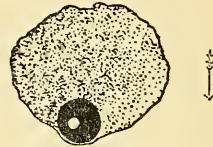


FIG. 23. — Section through the nucleus of a young ovum ( $\frac{1}{3}$  mm. in diameter) showing the nucleolus, which has, apparently, caused a distention of the nuclear membrane by the pressure of its own weight. Arrow shows the direction of the earth's centre. Magnified 248 diameters. (From HERRICK, '95.)

the moment of killing (Fig. 22). The weight of the nucleolus is relatively so great as sometimes to cause a depression in the part of the nuclear membrane upon which it rests (Fig. 23).

### § 3. CONTROL OF THE DIRECTION OF LOCOMOTION BY GRAVITY — GEOTAXIS \*

The control of the movements of Protista has been investigated chiefly by four naturalists: SCHWARZ ('84), who studied *Euglena* and *Chlamidomonas*; ADERHOLD ('88), who studied

\* So called by SCHWARZ ('84, p. 71).

Euglena and desmids; MASSART ('91), who worked upon bacteria, and ciliate and flagellate Infusoria; and JENSEN ('93), who experimented with Euglena, Chlamydomonas, and eight species of Ciliata.

The observation that led SCHWARZ to his study was that Euglena and Chlamydomonas, shaken up with sand and covered by it, constantly, even in the dark, rose to the surface. The experiments now made by SCHWARZ to determine the true cause of the phenomenon were a model of experimental investigation. In the first place only fresh and actively moving individuals were used, and light was carefully excluded, either by enveloping the culture vessel in black paper, or by working in a dark chamber. I shall now give in detail the experiments and their results.

When the Flagellata were placed in water they responded like those in sand — they soon came to the upper surface. But may not this upward movement be purely passive due to the small specific gravity of the algæ or to currents in the water? To get an answer to this question SCHWARZ heated the sand to 70° C — a fatal temperature — and no aggregation occurred. Again, the algæ were subjected to vapor of chloroform; no aggregation. Again, to a low temperature (5° to 6°); no aggregation. An aggregation occurred, however, when the temperature of the same culture was raised to 22°. Finally, Lycopodium spores and Euglena in the resting stage do not move upwards; hence no currents are passing in this direction. On the contrary, these experiments show that the upward movements of the algæ are the results of its own active locomotion.

Nor can it be that anything else than gravity determines the direction of the locomotion. That the greater amount of oxygen at the upper level is not the controlling agent was shown by smearing the sides of a glass cylinder with a thin layer of sand containing the algæ. In this thin layer, permeated by oxygen, they still accumulated at the upper margin. That the locomotion was not directed by currents in the water (Rheotaxis, p. 108) was indicated by the fact that whether the free end of the tube, at which evaporation is occurring, be up or down, migration is always upwards. Thus, since the stimulus

of chemical agents and currents was eliminated, gravity seemed to remain as the only directing force.

It only remained to show that the attractive force of the earth can be replaced by centrifugal force, and this SCHWARZ was able to do by means of the klinostat. By varying the rate of rotation of this machine he varied the centrifugal force and was able to determine the limits within which the Infusoria move against an opposing force. The acceleration of the rotation-force may be expressed in terms of the attraction of gravity as a unit by the formula  $c = \frac{f}{g}$ , when  $c$  equals the acceleration of centrifugal force in the required units;  $f$ , the centrifugal force found, as on p. 113; and  $g$  the acceleration due to gravity. It appeared from the experiments that, in both living *Euglena* and *Chlamidomonas* migration took place towards the central end, thus against the centrifugal force, when the latter was over 0.5 g., and under 8.5 g. Under the lower limit no migration occurred; near the upper limit aggregation occurred at both ends; above the upper limit aggregation took place at the peripheral end—that is to say, *with* the centrifugal force. Clearly, then, geotaxis is in these cases a movement against an opposing force, provided that force is considerable (over 0.5 g.) but not too great (over 8.5 g.).

The workers in this field who followed SCHWARZ advanced our knowledge of geotaxis in two principal ways: first, by increasing the number of organisms known to be geotactic, and, secondly, by revealing the fact that closely allied species may have geotaxis of opposite sense.

MASSART ('91, pp. 161–167) employed a simple but satisfactory method. He placed Protista in a capillary tube which was open, hence equally oxygenated at the two ends. By inverting the tube the ends were brought into different relative positions with respect to the earth, causing the geotactic organisms to migrate throughout its length. As a result of his experiments it appeared that *Spirillum*; the flagellata, *Polytoma*, *Chlamydomonas*, and *Chromulina*; and the ciliata, *Anophrys* and *Euplotes*, are geotactic. The sense of geotaxis may be different between individuals of the same genus; thus, under similar conditions *Spirillum* separated into a lot lying at the

upper part of the tube and a lot at the lower part; and the individuals of both the upper and lower lot were active. The sense of response depends upon temperature also. Thus *Chromulina woroniniana* is negatively geotactic at 15° to 20° C., and positively geotactic at 5° to 7° C. The other species mentioned above are negatively geotactic — *i.e.* move in the direction opposite to that in which the force tends to carry them.

JENSEN\* ('93) finally has greatly extended our knowledge of the species responsive to gravity, has shown the necessity for regarding carefully the other agents acting during the experiment, and has entered more carefully into the cause of the phenomenon than previous authors. The new forms which JENSEN worked with were these Ciliata: *Paramecium*, *Urostyle*, *Spirostomum*, *Colpoda*, *Colpidium*, *Ophryoglena*, and *Coleps*; also the more commonly used species, *Euglena* and *Chlamydomonas*. The other agents whose action may modify that of gravity are chemical stuffs, density, warmth, light, etc. Light may be easily excluded. On warm days the typical geotactic phenomena are often absent, the *Paramecia* sinking to the deeper, cooler layers. The *Infusoria* aggregate around bacteria in the water, — chemotaxis (Fig. 24,

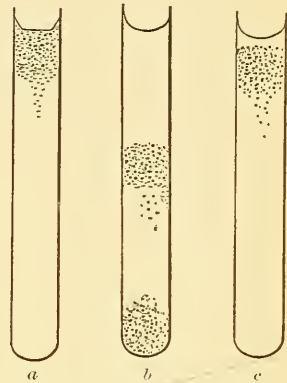


FIG. 24. — Glass tubes, about 0.5 cm. in diameter and 20 cm. long, fused at one end, and filled with water containing *Paramecium* (represented by points). *a* shows aggregation of the *Paramecium* at upper end of tube; *b*, aggregation of *Paramecium* around bacteria suspended in the water — chemotaxis veiling geotaxis; *c* shows that occasionally the *Paramecia* avoid the uppermost layer of the water. (From JENSEN, '93.)

*b*), — and they shun the uppermost layer, apparently because, owing to evaporation, this layer is denser — tonotaxis (Fig. 24, *c*). Whether light inhibits the geotactic response was one of the questions asked and answered by JENSEN. When

\* JENSEN used glass tubes of 0.5 to 1 cm. diameter, and 5 to 100 cm. length, fused at one end. To prevent the free end becoming richer in oxygen, a layer of oil 2 to 3 cm. high was poured over that end, or, air being carefully excluded, it was sealed by an impermeable plug of wax.

the centrifugal machine was used in the sunlight, movements towards the centre clearly appeared. It was thus proved that negative geotaxis (which is the same as centrotaxis) may occur in the sunlight.

The data of geotaxis are incomplete without a consideration of this phenomenon in the higher animals. LOEB is one of the first investigators in this field. In 1888, he found that flies, deprived on both sides of the free ends of the balancers or the wings, and placed upon a board, move always upwards upon it. If the plane of the board is held oblique to the horizontal, the fly always moves along that line which makes the smallest angle with the vertical. Likewise cockroaches seem to be stimulated by gravity when this acts perpendicularly to their ventral surface, so that they tend to move off from a horizontal surface and do not come to rest until they are on a more or less nearly vertical one. Thus, LOEB put twenty-one cockroaches into a truncated, pyramidal box, one of whose sides made an angle of  $80^\circ$  with the horizontal; another  $60^\circ$ , the third  $45^\circ$ , and the fourth  $25^\circ$ . After an hour, the number of individuals on each face of the box was counted at intervals of 10 minutes. Adding together the results of 10 such counts he found on the steepest wall 94 cases; on the wall inclined at  $60^\circ$ , 61 cases; on that inclined at  $45^\circ$ , 28 cases; on that at  $25^\circ$ , 25 cases; and on the horizontal surfaces, 2. After several hours 75 to 80% of the animals were found on the steepest side, although it had the smallest area. Later, LOEB ('90 and '91) showed that the holothurian *Cucumaria cucumis*, the starfish *Asterina gibbosa*, and the lady-bird beetles (*Coccinellidæ*) are likewise geotactic. Finally, it may be mentioned that several species of the slug, *Limax*, are geotactic.

There is in Protista, as already mentioned, a limit to the intensity of the attractive force below which no response will occur. Is there such a limit in Metazoa likewise? Experiments upon this point have been made by Miss HELEN PERKINS and myself in connection with my experimental course at Radcliffe College. We have also been able to answer the question, what difference in effect is produced by different intensities of gravity's action. We experimented with the great slug, *Limax maximus*, which crawls readily upon a glass plate placed



at any angle. As explained on p. 113, the intensity of gravity's action will diminish as the sine of the angle of inclination of the plate is diminished from 90° to 0°. We determined the deviation of the slug from a vertical position upon plates at various inclinations, and after the lapse of a constant time (45 seconds). The experiments were performed in a dark box. The number of tests made at each inclination was sixty. The time required to respond fully to gravity did not vary appreciably with the angle of inclination. The results obtained indicated that the deviation of the slug from verticality diminished with the cosine of the angle made by the plate. The relation between the angular deviation from verticality and the sine of inclination of the glass plate is graphically represented in Fig. 25. The conclusion from the experiments is that the lower limit of the sensitiveness of *Limax* to gravity is extremely small, below 0.13 g., and that as the angle of inclination of the plate diminishes the deviation from 45° towards verticality diminishes in accordance with the relation:  $\delta = a \cdot \sin \theta$ , in which

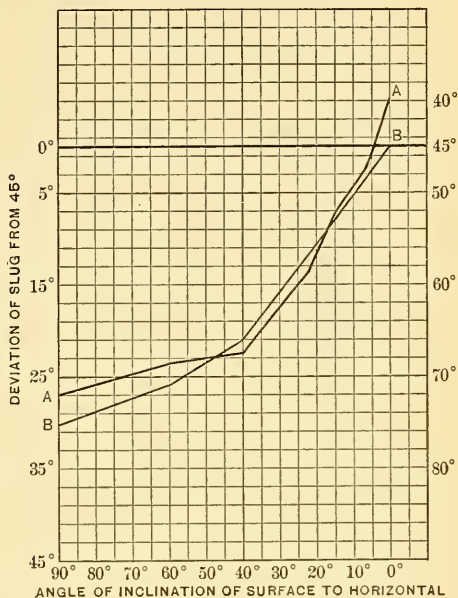


FIG. 25.—Curves showing relation between the sine of the angle of inclination of a glass plate and the angular position of a slug which was first placed on it in a horizontal position and then left for 40 seconds in the dark. Curve *A* is constructed by drawing ordinates from the heavy horizontal line, 0°–45°, corresponding to each angle of inclination of the surface (laid off as abscissæ). The lengths of the ordinates are determined by the number of degrees of deviation of the axis of the slug from 45° towards 90°. 45° is taken as a base, since it would be the mean angular deviation from the initial position of a slug crawling undirected upon a horizontal plate. Curve *B* is constructed by drawing down from the base ordinates proportional to the natural sines of the different angles of inclination of the glass plate.

$\delta$  is the angular deviation of the slug from  $45^\circ$  towards  $90^\circ$ , expressed in degrees;  $\theta$  is the angle of inclination of the plate to the horizontal, and  $a$  is a constant.

In inquiring into the cause of geotaxis in animals it seems best to consider chiefly the phenomenon as exhibited in Protista, for in the higher animals this capacity seems bound up with the possession of special organs of orientation. In this group the first and apparently most important part played by gravity is the determination of the axis of the individual, which comes to lie vertical and with the head end up or down according to the conditions of the protoplasm. After the positions of the axis and poles are determined, ordinary locomotion produces the geotactic phenomena. That gravity may determine a vertical position without locomotion occurring is shown in the ciliate infusorian *Spirostomum* (Fig. 26), which at times occurs in large numbers in ordinary aquaria, suspended almost motionless in mid-water, having a distinctly vertical position and with the head end directed upward. They cannot be said to be strictly motionless, since by carefully attending to them one can see them slowly rising or falling

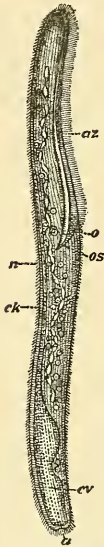


FIG. 26. — *Spirostomum ambiguum*, side view. *az*, adoral zone of cilia; *o*, mouth; *os*, gullet; *n*, nucleus; *ck*, contractile canal; *cv*, contractile vacuole; *a*, anus. Magnified about 120 diameters. (From BÜTSCHLI [BRONN'S Thier-reich: Protozoa], after STEIN.)

or alternately, perhaps, rising and falling in their almost imperceptible movements. Miss JULIA B. PLATT, who has studied carefully the movements of *Spirostomum*, found that of 78 individuals observed all but 7 had the anterior extremity directed upwards and the 7 exceptional individuals were all moving downwards. It therefore seems quite certain that *Spirostomum* tends in water to orient itself with reference to gravity, although without aggregating at the upper surface.

To explain the phenomenon of axis-orientation, two principal theories have been advanced. The first may be called the *mechanical* theory; the second the *response-to-stimulus* theory. The first theory is that once suggested by VERWORN ('89, p. 122). It appeared to him that it was self-evident from purely physical grounds that, in complete quiescence of the flagellum, the hinder end of the protist should be directed downwards, and not the anterior flagellum-bearing end. If one conceives such an individual to move its flagellum, which precedes in locomotion, it must move towards the surface of the water; thus against gravity. VERWORN finds the stimulation theory inconceivable, since gravity cannot even be compared with stimuli. In falling, the body of the protist might rub against the water particles, which would offer a stimulus, but this would be more allied to rheotaxis.

It might seem an easy thing to determine whether geotactic Protista artificially rendered quiescent (*e.g.* killed or stupefied) would stand with their anterior ends uppermost; but the killing is apt to distort the form, and the organisms being heavier than water \* fall to the bottom. Something might be gained from an observation of how they fall, but there is very great discordance among authors upon this point, probably in part due to difficulties of observation. Thus SCHWARZ ('84, p. 68) says that both *Euglena* and *Chlamidomonas* assume all positions in falling; MASSART ('91, p. 164) finds that *Chlamidomonas* falls with flagellum directed upwards and JENSEN ('93, p. 451) declares that *Euglena viridis* killed by iodine falls

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\* Few determinations seem to have been made of the specific gravity of living Protista. JENSEN ('93<sup>a</sup>) attempted to do this for *Paramecia*, but his method was bad and his results bad likewise. He made solutions of potassium carbonate, of varying specific gravity, and found that *Paramecium* just floats in a solution whose sp. gr. is 1.25. The difficulty of the method is that solutions of salt having a relatively small molecular weight act so powerfully in withdrawing water from the organism as to cause it to shrink and increase in relative weight. Miss PLATT has used solutions of gum arabic whose osmotic action is so slight that organisms live in it for hours. In such solutions, paralyzed but living *Spirostoma* and *Paramecia* neither sank nor rose when the specific gravity was between 1.016 and 1.019; so that it seems probable that the specific gravity of Infusoria lies near 1.017. Tadpoles recently hatched and having a length of 9.5 mm. had a sp. gr. of 1.044, while those 12 mm. long had a sp. gr. of 1.017.

almost without exception with the broader flagellate pole downwards. Both from the fact that it can be easily demonstrated that when a body heavier than water falls in that medium its larger end will precede, and from the fact that JENSEN was especially careful that the killed organism should not be deformed, his results must be considered the best established. Now, since the dead *Euglena* tends to sink with flagellum downwards whereas the active *Euglena* stands flagellum upwards, we must conclude that the orientation of *Euglena* and probably other Protista is not passive but due to their activity and must be regarded as a response more or less directly due to gravity.

But just how does gravity act as a stimulus to determine the direction of orientation of the body? We have two principal theories to examine. First, that of JENSEN, that gravity acts indirectly on the organism by directly causing a difference in pressure in the water at different levels. This difference in water pressure, at various levels, affects directly the two poles of the organisms, which stand at different levels, and the organism responds to this difference in pressure. The second theory, which I adopt, is that the organism, owing to its specific gravity being greater than the medium, experiences greater resistance (friction + weight) in going upwards even to the slightest extent than in going downwards (friction - weight). Another stimulus, which is probably associated with this, depends upon the fact that an unsymmetrical body, heavier than water, tends to fall with its larger end down. Those negatively geotactic organisms, which stand with their larger end up, will be consequently in a condition of unstable equilibrium; those organisms which stand with their larger end down will be in stable equilibrium. In the first case a deviation from verticality would be accompanied by relatively diminished resistance on one side; in the second by relatively increased resistance on one side. In either case, the distribution of the mass of the animal may give the organism the means of determining, but not in a mechanical way, the position of its axis.

The evidence for the first theory JENSEN finds especially in a fact which he believes opposes the second. Negatively geotactic organisms, placed in an inclined tube, move towards

the upper side and then travel obliquely, not vertically, along it toward the upper part of the tube, thus into strata of constantly diminishing pressure. If weight controlled in any way their movements, they should move vertically as from 1 to 2 (Fig. 27) until they meet the side of the glass. Then they should move off, as to 3, then vertically to 5, and so on. Since they do not so move, gravity, JENSEN thinks, cannot be said to act directly. In criticism of this conclusion it may be urged that it is without proper foundation, for if an organism whose irritability (instincts) would lead it to move vertically is mechanically unable to do so exactly, it will do so as far as practicable. This observation cannot, therefore, be said to militate against the second theory. Finally, there is this positive objection to JENSEN'S theory that it is applicable only to geotaxis in water animals, and can therefore be only a special explanation of geotaxis.

On the other hand, there is evidence which is opposed to the first theory and favors directly the second. And JENSEN has himself contributed some of this evidence. He put *Urostylela* into a glass tube containing a 0.5% aqueous gelatine solution. They showed no tendency to go upwards. At the expiration of 20 hours many deaths had occurred, but some normally active individuals were still at the lower end of the tube.

Why this loss of geotaxis? JENSEN believes it due to the fact that the difference in pressure of the successive layers did not increase proportionally to the increase in resistance of the solution. I would suggest that it may be due to the fact that the weight of the body of the Protista is now relatively less than that of the solution, so that the organism, tending to move against resistance, comes to lie at the bottom of the buoyant fluid, hence appears positively geotactic.

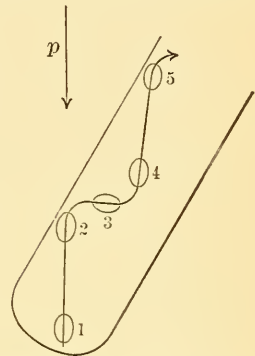


FIG. 27.—Hypothetical line of migration of *Paramecium* in an inclined tube, upon the assumption that gravity acts directly to determine direction of locomotion, according to the conception of JENSEN. The arrow at *p* indicates the direction of the pull of gravity; 1, 2, 3, 4, 5, successive positions occupied by the *Paramecium*. (From JENSEN, '93.)

Geotaxis in the higher organisms, especially Vertebrates, cannot here be discussed at length. It is sufficient to state that as LOEB ('91, p. 189) concludes, it is probably dependent upon the internal ear. Miss PLATT has, at my suggestion, subjected young negatively geotactic tadpoles to solutions of gum arabic of the same specific gravity as themselves, and has found that they still migrate upwards. This result makes it probable that here also orientation is effected by the internal ear, and hence is independent of the action of gravity upon the entire body.

Finally must be mentioned the phenomenon of *acclimatization to a central pressure*. This has been observed by JENSEN ('93, p. 470), who says, when *Paramecium* or *Urostyla* has been strongly "centrifugated" towards the peripheral end of the tube, where it is subjected to a high pressure, it shows, when the tube is then placed vertically, a much livelier geotaxis than it would have done without "centrifugating." Clearly the temporary action of the high pressure has increased the irritability to gravity.

To recapitulate: Gravity affects the structure of protoplasm by separating the lighter and heavier substances. It may determine the direction of locomotion by determining the verticality of the axis of the body. Varying the intensity of gravity's attraction diminishes the *precision* with which this determination takes place. The determination of the vertical position is, in the lower organisms, probably due to difference in ease of movement when going up and going down.

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## CHAPTER VI

### *EFFECT OF ELECTRICITY UPON PROTOPLASM*

IN this chapter we shall consider (I) some methods employed in the investigation of this subject; (II) the effect of electricity upon the structure and general functions of protoplasm; and (III) the effect of electricity in determining direction of locomotion — electrotaxis.

#### § 1. CONCERNING METHODS

While the phenomena of magnetism and electricity are closely allied, their effects upon protoplasm seem to be widely dissimilar. Thus no certain action of magnetism has hitherto been observed, but electricity, however produced, causes nearly uniformly an effect.

Any experimental work with the electric current involves apparatus for its production, application, and measurement; namely, batteries or other sources of electricity; electrodes for applying the current to the organism; troughs to contain the free swimming animals used for experimentation; a galvanometer for measuring the current; a rheochord for varying the intensity of the current; a reversing key; and, for interrupted currents, an induction machine with interrupter, and an electrometer for measuring such currents. A description of the principal forms of these instruments and the methods of constructing some of them will be found in VERWORN, '95, Chapter V, and in OSTWALD, '94, Chapter XV.

Since the works just named are easily accessible, it will be unnecessary here to describe these instruments in detail. A few *additional* suggestions, the result of my experience, may, however, be found helpful. Concerning *batteries*, first; accumulators are without doubt to be preferred, where practicable, on



account of the strength and continuance of their currents. In other cases, CLARK or DANIELL elements, if enough of them are united in series, will meet the requirements. The character of the *electrodes*, next, will depend upon the nature of the investigation. Nonpolarizable ones of hair (camel's-hair brush), clay, or paper (plug of filter paper in glass tubing drawn out to a cone) are usually employed, but all of these offer considerable resistance. The *troughs* will vary in form and size with the organisms to be contained in them; some of them will be described in connection with the experiments in which they have been employed. They are all rectangular enclosures having clay ends when it is desirable that these should be nonpolarizable. For large troughs, sheet-zinc electrodes are used, covering the smaller sides of the trough. Although some of the reflecting *galvanometers* are more sensitive, a "millammeter" such as that made by the WESTON Electrical Works is a much more convenient instrument and sensitive enough for most work of this sort. The *rheochord* is practically a low-resistance box, capable of indefinitely fine gradations. This is introduced into the short branch of a divided circuit, so that by varying its resistance a varying share of the current shall be forced into the longer circuit. A very simple and excellent device for altering the strength of current is the "Compression-rheostat" of BLASIUS and SCHWEIZER ('93). This consists of a piece of rubber tubing filled with zinc sulphate, stopped at the ends and introduced into the circuit. By means of a thumbscrew the walls of the middle of the tube may be pressed together, the lumen correspondingly reduced, and the resistance increased. The *induction apparatus* usually employed is one invented by DU BOIS-REYMOND. In this the secondary coil may be withdrawn from the primary coil to any desired distance, thereby diminishing the intensity of the induced current. Through the action of such an instrument the current is alternately made and broken, and each electrode becomes in quick succession anode and kathode. Since alternating currents cannot be measured by an ordinary galvanometer, an electrometer must be employed. So much concerning apparatus.

A word should be said about the method of stating the current employed. Very many authors have been satisfied with

saying that the current was strong or weak, others have given the kind and number of elements employed. Such statements are wholly inadequate to give an accurate idea of the strength of current to which the organisms under experimentation were subjected. Even merely to state the galvanometer reading in milliamperes is insufficient. We must know as nearly as possible what strength of current is passing through the organism, and this involves knowing the density of the current passing through the water in the trough. Now it is obvious that a current passing through a mass of water of small cross-section is stronger per square millimeter than an equal current distributed over a large cross-section. It is necessary, consequently, to know the cross-section of the mass of water through which the stimulating current is passing, in order to determine the "density" or strength at any point. For technical purposes the unit of current-density is taken at 1 ampere to the square millimeter. HERMANN and MATTHIAS ('94, p. 394) propose for physiological purposes a unit one-millionth as great, to be designated as  $\delta$ .  $\delta$  then indicates a current of  $\frac{1}{1000}$  milliampere per square millimeter of cross-section. It is very desirable that, when practicable, currents should hereafter be expressed in  $\delta$ 's. More than one useless discussion has been precipitated by not giving a sufficiently accurate quantitative expression to the current employed. (See, for illustration, below, p. 149.)

Finally, the strength of current necessary to produce a certain result depends upon the relative conductivity of the organism and the surrounding water. If, through the presence of substances in solution, the conductivity of the water is abnormally great, one must use a greater current (as read off from the galvanometer) than otherwise to produce a certain effect. (WALLER, '95, p. 97.) It would probably be best, when possible, to use in the trough the water in which the organism has been living, since the quantity of salts in the organism has been shown to vary with that of its medium. (See p. 88.)\*

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\* See KAISER, Wien Akad. CIV, p. 17, 1895, for a new trough adapted to the stage of the microscope.

## § 2. THE EFFECT OF ELECTRICITY UPON THE STRUCTURE AND GENERAL FUNCTIONS OF PROTOPLASM

The fundamental phenomenon of the action of an electric current upon protoplasm may be seen while watching a heliozoan (*Actinosphærium*), lying in a drop of water, through which a weak, constant current is "made." We find that the filamentous pseudopodia begin quickly to retract at the two poles lying in the axis of the current; and as the current continues, this contraction continues likewise. The primary effect of a weak constant current is thus a centripetal flowing of the protoplasm. The current stimulates to contraction.

If, now, the current be increased, or be longer continued, further changes occur. The pseudopodia lying in the current become varicose, and break up into a chain of drops; the vacuoles on the periphery begin to burst, emptying out their fluids; and in these regions the protoplasm collapses. Thus, the stronger current produces continued contraction, accompanied by collapse of the protoplasmic foam-work.

Finally, the plasma itself begins, upon the anode side, to disintegrate, and the loose particles to move towards the positive electrode. As the plasma of this side is gradually eaten away, the outline of the *Actinosphærium* passes through phases like those of the waning moon, until, finally, the last thin crescent fades away. The particles of the mass have wholly lost their cohesion (Fig. 28).

The facts just given concerning the behavior of *Actinosphærium* to the constant current are gathered from the observations of KÜHNE ('64, p. 59) and VERWORN ('89<sup>a</sup>, pp. 8, 9). Fundamentally similar observations have been made by KÜHNE ('64, p. 79) and VERWORN ('89<sup>b</sup>, p. 274) on *Myxomycetes*, and by VERWORN ('89<sup>a</sup>, pp. 13, 17) on the rhizopods, *Polystomella* and *Pelomyxa*. So these data may be considered as of general worth for naked protoplasm.

Also upon ciliated epithelium, the constant current acts as a very strong excitant, producing an active movement in cilia which had previously nearly ceased to beat. This excitation occurs, especially about the two poles, immediately upon "making" the current. (KRAFT, '90, pp. 234, 235.)

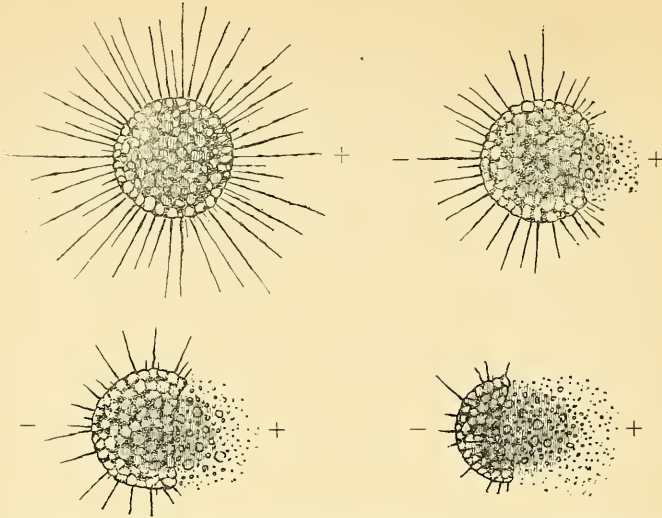


FIG. 28. — *Actinosphaerium eichhornii* in four successive stages of polar excitation by means of the constant electric current. Disintegration begins at the anode (+) pole. (From VERWORN, '95.)

Allied, apparently, to the foregoing phenomena are the protoplasmic changes which follow the sudden *breaking* of the current. Unless the current has been very feeble, the pseudopodia of *Actinosphaerium* begin, at the moment of breaking, to contract and become varicose upon the kathode side, while the formerly irritated anode side is quiet. Thus, the breaking of the current also acts as a stimulus, but this is, in general, weaker than that caused by making.

If, now, a current which endures for only an instant — if a single induction shock — is sent through, the making and breaking stimuli are practically coincident, and a violent response may be called forth. Thus, ENGELMANN ('69, p. 317) found that *Amœba*, subjected to a strong shock, retracted its pseudopodia, and assumed a spherical form within two seconds; and GOLUBEV ('68, p. 557) has described a similar response in leucocytes. Under similar circumstances, the flagellum of the flagellate *Peranema* (Fig. 29) made an energetic stroke. (VERWORN, '95, p. 414.) I have spoken above as though there were both a making and a breaking stimulus; but this is not known to be the case. It is generally recognized from experiments

on muscle, that it is the "making" only of a single induction shock which produces the response; but VERWORN ('89<sup>a</sup>, pp. 19-22) has found that in the rhizopod *Pelomyxa* it is, on the contrary, the breaking excitation which causes the response. The subject deserves further study.

Finally, the effect of an alternating current must be considered. This current is characterized by the fact that it is composed of a series of rapidly repeated instantaneous shocks

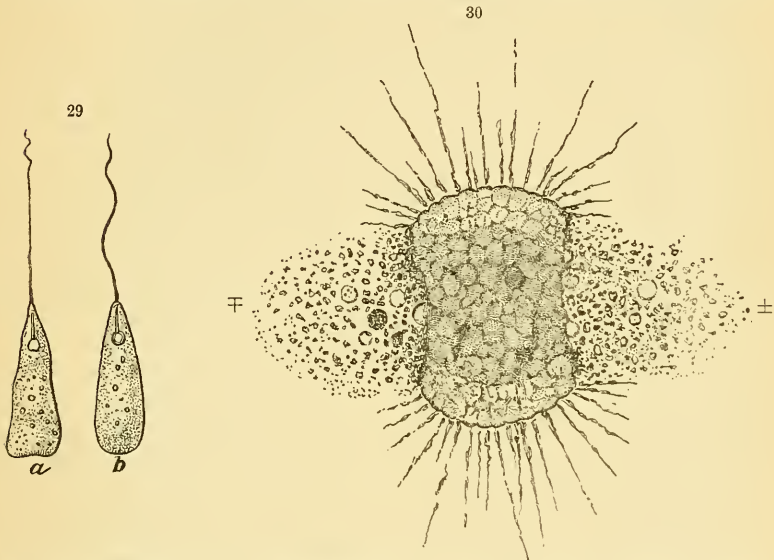


FIG. 29. — *Peranema*. *a*, quietly swimming; *b*, irritated by an induction stroke. (From VERWORN, '95.)

FIG. 30. — *Actinosphaerium eichhornii*, STEIN. Showing effect of the alternating current. At both poles the pseudopodia are undergoing a disintegration, which proceeds equally at the two poles. (From VERWORN, '89.)

which alternately reverse their direction. Thus, each pole of the organism subjected to such a current receives alternately the making (or breaking) effects at anode and kathode. The maximum action is thus obtained. When an *Actinosphaerium* is stimulated by such a current, the pseudopodia at both poles contract and become varicose; and, finally, the protoplasmic substance begins to disintegrate and to flow out from the cell towards the two electrodes, until the body acquires a biconcave form. (VERWORN, '89<sup>a</sup>, p. 11.) In this case the disintegra-

tion takes place at both poles, since both are, alternately, anodes (Fig. 30).

Similar effects have been observed in other cases. Thus, when an amœba is subjected to an alternating current, it becomes spherical; the protoplasmic streaming of the plasmodia

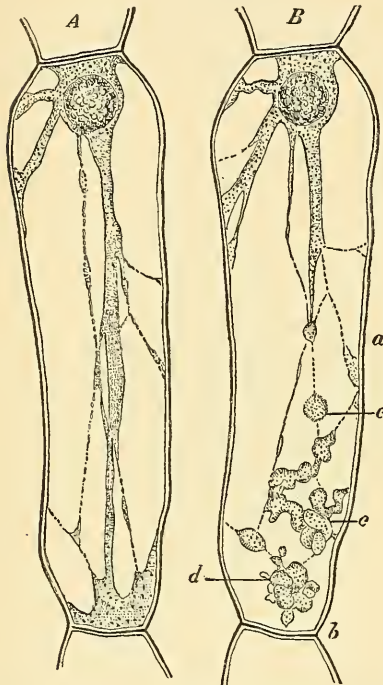


FIG. 31.—One of the cells of a stamen hair of *Tradescantia virginica*. *A*, unstimulated; *B*, stimulated by an induction current. At *a*, *b*, *c*, *d*, the protoplasm has aggregated into drops and clumps. (From VERWORN, '89, after KÜHNE, '64.)

of a myxomycete ceases, and, with stronger currents, the whole mass contracts, water being forced out. Finally, an attempt at a similar result is seen in the stamen-hair cells of *Tradescantia*, in which, under stimulation, the protoplasmic threads segregate into irregular or spheroidal clumps. (KÜHNE, '64, pp. 30, 31, 75, 99.) In all these cases we see that the action of a violent current, like repeated contact, leads (as ENGELMANN, '69, p. 321, has suggested) to results which can be accounted for on the ground of reduced cohesion,—first, tendency to spherical aggregation, and, finally, disintegration (Fig. 31).

After having studied the effect of the electric current upon Protista and simple cells, it remains to consider,

very briefly, its effect upon muscle and upon nerve. Since CALDANI discovered, in 1756, that frogs, shortly after death, could be stimulated to movement by frictional electricity, and GALVANI and VOLTA, towards the end of the last century, discovered, by the same response, the phenomenon of galvanism, these tissues have frequently been made the subject of careful experimentation. It has been shown, not merely that

the nerve can be stimulated to its functions, but that muscle from which the activity of the nerve has been excluded by the use of curare (which inhibits the action of the nerve, but not of the muscle), will contract upon the passage of a current.

Upon the character of the current, however, depends that of the response; thus, although, as we have seen, a closed constant current continues to stimulate Protista, it has been said not to stimulate nerve or muscle. A contraction follows, it has been maintained, only upon considerable variations in the electrical condition, such as result from making or breaking the current. It is probable, however, that there is not so great a difference in responsiveness of muscle and Protista as would seem to be implied, for BIEDERMANN ('83) has shown that the constant current produces a whole series of slight contractions in muscle which cannot be regarded merely as a secondary result of the making shock; and FICK ('63) has observed contraction due to the constant current in muscles of Lamelibranchia. So that even in muscles, there is an actual, though weak, response to a steady, constant current.

There are two phenomena following momentary shocks applied to muscles which deserve notice in passing. First, when a single induction shock is passed directly through a muscle, we notice that the contraction is not simultaneous with the shock, but follows only after the lapse of a certain "latent period." This latent period represents, it is believed, time spent in transformations going on in the plasma preparatory to contraction. Secondly, when we pass (especially in a muscle-nerve preparation) a series of induction shocks, closely following one another, as in the alternating current, a very violent contraction is produced, since the new shock comes to the muscle before it has had time fully to relax, and causes a contraction of the already contracted tissue. Thus stimulus is superimposed upon stimulus, and a summated response (tetanus) takes place.

We must now consider more carefully a subject to which we have hitherto merely alluded, namely, the relation to the electrodes of the point of the organism at which the response first appears. Thus, when the amœboid *Pelomyxa* is subjected to the constant current, a contraction appears, at the time of

making the current, at that pole only which is turned towards the anode. When the current is broken, on the contrary, a contraction occurs at the kathode, the pseudopodia next the anode becoming quiet. (VERWORN, '89<sup>a</sup>, p. 19.) This relation may be expressed in tabular form as follows:—

	AT ANODE.	AT KATHODE.
Upon making . . . . .	excitation	rest
Upon breaking . . . . .	rest	excitation

All Protista do not, however, according to VERWORN, respond in the same way as *Pelomyxa*; thus, with the constant current of a certain intensity, he got in both *Polystomella crispa*, and *Actinosphærium*, the following reaction:—

	AT ANODE.	AT KATHODE.
Upon making . . . . .	excitation	rest
Upon breaking . . . . .	rest	rest

It must be said, however, that the reactions obtained in any case are dependent upon the strength of current employed; thus, with a stronger current, the following result was obtained with *Actinosphærium*:—

	AT ANODE.	AT KATHODE.
Upon making . . . . .	excitation	excitation
Upon breaking . . . . .	rest	excitation

A comparison of the last two tables seems to indicate that, very probably, with a current intermediate between the weak and the strong current employed, we should get a result like that obtained with *Pelomyxa*. At any rate, we may say that all these cases tend to group themselves about the *Pelomyxa* formula;—making: anode, excitation; kathode, rest; breaking: anode, rest; kathode, excitation. A brief designation of this type is desirable. Since the condition at the anode upon



making is distinctive, we may call this the anode-excitation type, or, briefer still, *anex* type.

Turning, now, to nerve and muscle tissue, we meet with a type of response, on making and breaking the current, altogether irreconcilable with this. As is well known, when a constant current is made or broken, all the tissue lying between the electrodes is not stimulated at one time, but the excitation makes its appearance at the anode or kathode, and thence is transmitted to the other pole. One can demonstrate this on slow-moving (*e.g.* extremely tired or dying) muscle at the extremities of which the electrodes are placed. The contraction begins at one electrode, and travels towards the other. By using more refined methods, this relation, which holds for nerves, striated and smooth muscle (cf. ENGELMANN, '70, p. 302) has been formulated as follows:—

	AT ANODE.	AT KATHODE.
Upon making . . . . .	rest	excitation
Upon breaking . . . . .	excitation	rest

This is seen to be the very opposite of the response given by *Pelomyxa*. It may be called the kathode-excitation type, or, in brief, the *katex* type.

Having now seen that two fundamentally different types exist in the response of the two extreme groups of the animal kingdom, the question arises, what is the distribution of these types amongst the intermediate forms—the Invertebrate Metazoa? Fortunately, through the investigations of NAGEL ('92 and '92<sup>a</sup>), we have data upon this subject. In NAGEL'S experiments, the whole animal was employed, the two electrodes were placed at the opposite ends of its long axis, the metallic circuit was then made or broken as required, and the pole (anode or kathode) at which contraction first occurred was noted. Thus, NAGEL found that when the current was made through the sea-hare, *Aplysia*, there was strong excitation and momentary retraction of the parts next to the anode, while next to the kathode the body showed a considerably weaker contraction. Upon breaking the current, there was some excitation of the parts of the

body next to the kathode, but none at the anode end. The result that one obtains depends, however, to a certain extent, upon the strength of the current that one employs. But NAGEL did not, apparently, measure his currents, so there is no certainty that his results can be at once duplicated. Taking the results for various Invertebrates as they are given, however they are instructive.

TABLE XIII

	UPON MAKING.		UPON BREAKING.	
	AT ANODE.	AT KATHODE.	AT ANODE.	AT KATHODE.
<i>Limnæus</i> . . . . .	excitation	rest	rest	excitation
<i>Planorbis</i> . . . . .	excitation	rest	rest	excitation
<i>Aplysia punctata</i> . .	excitation >	excitation	rest	excitation
<i>Schæurgus</i> (octopod)	excitation >	excitation	rest	excitation
<i>Helix hortensis</i> . . .	excitation >	excitation	rest	slight excitation
<i>Ciona intestinalis</i> . .	excitation >	excitation	rest	rest
<i>Janus cristatus</i> . . .	excitation =	excitation	rest	slight excitation
<i>Pleurobranchia</i> . . .	excitation =	excitation*	rest	slight excitation
<i>Nassa reticulata</i> . . .	weak excit.	rest	rest	rest

All the species in this table (all of which, except *Ciona*, are Mollusca) show in their response a more or less close approach to the type of *Pelomyxa* (excitation, rest; rest, excitation); and we may believe that with appropriate stimulus they would respond in precisely that way.

In a second class of cases the response of the whole animal belongs to the katex type; thus two species examined by NAGEL showed the following responses:—

TABLE XIV

	UPON MAKING.		UPON BREAKING.	
	AT ANODE.	AT KATHODE.	AT ANODE.	AT KATHODE.
<i>Pagurus striatus</i> . . .	rest	excitation	excitation	rest
<i>Triton cristatus</i> . . .	excitation <	excitation		

\* Inconstant in occurrence.

These are representatives of the groups Crustacea and Vertebrata.

Among all the species studied by NAGEL there was only one which gave results not easily assignable to either of the two types. This organism is the larva of a dragon-fly, *Æschurea*. Its formula is excitation = excitation; rest, rest. NAGEL says, however, that these results were uncertain and variable.

In some other groups studied — Cœlenterata and Echinodermata — the current used provoked no response at either pole; while with *Amphioxus* the current employed produced excitation at both poles on both making and breaking the circuit. Such variations as these, are, however, easily accounted for on the ground that different species require currents of different strengths to call forth what may be termed the typical response.

Not merely between different groups do we find a difference in the type of response, but even inside the group of Protozoa dissimilarity has been shown to occur. Thus VERWORN ('89<sup>b</sup>, p. 301) has given reasons for believing that three flagellate species, the ciliate *Opalina*, and some bacteria belong to the katex type, although as just stated (p. 133) other Protozoa exhibit the anex type of response.

Finally it appears that individuals of one and the same species subjected to different intensities of current may give rise to responses belonging to the opposite types. Thus when a medium current is "made" through *Triton cristatus* the excitation is greater at the kathode than at the anode; but when the weakest current is employed a making response occurs at the anode only, and when a slightly greater intensity is used the continuance of the current provokes a continuance of the excitation at the anode. (NAGEL, '92<sup>a</sup>, p. 341.) Likewise the reaction of Vertebrate muscle varies with its internal condition. Thus degenerated or over-stimulated muscle shows predominating anode stimulation on making the current; and transverse stimulation of the muscle fibre gives the same result.\*

To sum up, two principal types of response to the electric current may be distinguished: the first or anex type characterizing most Protozoa, Mollusca, Vertebrates (slightly stimulated),

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\* For a discussion of these cases see VERWORN ('89<sup>a</sup>, p. 24).

and weak muscle fibres; the second or katex type found especially among some Flagellata, Arthropoda, and Vertebrata. A third possible type (katanex type) is certainly of very limited distribution. Between the two types we notice this connecting link, that in some Vertebrates a weak current produces one type of response; a strong current the other. The reason for the existence of these two distinct types — one of which characterizes animals with less differentiated, the other those with more differentiated, muscular and nervous systems — is still greatly in need of investigation.

The nature of the protoplasmic change wrought by the current is an important matter. We have already accounted for the effect that we see in the Protista, on the ground of reduced cohesion. It is probable also that the current gives rise in the cytoplasm to chemical changes which are different at the two poles. It is well known that when a current is passed through a neutral solution of a salt there is produced an acid at the anode, and an alkali at the kathode. Since in the higher animals, at least, the body contains such solutions, it seems probable that acid and alkaline substances are here likewise produced by the passing current. This probability is supported by an observation of KÜHNE ('64, p. 100), who found that the violet coloring matter of the stamen hairs of *Tradescantia* become changed by the action of a very strong induction shock. He says that a change in the violet fluid, like that which occurs at the anode, can be brought about by dilute hydrochloric acid; while a change like that appearing at the kathode can be produced by potassic hydrate. An observation of NAGEL ('92<sup>a</sup>, p. 346) suggests also that the current acts by producing a chemical change. He finds that that part of the body of the snail and the leech which shows most markedly the anode-making excitation is coincident with that which is most sensitive to chemical substances. So that the reaction to a galvanic, still more to a faradic, stimulation resembles that to a strong, disagreeable taste (quinine). In this case the response may result either from the chemical substance acting directly on the muscles or attacking first the sense organs. In the latter case the response would occur through the mediation of the nervous system.

So far then we have distinguished two chief effects of the current on protoplasm—a dissociation effect and a chemical effect. It may now be worth while to mention that there is good reason for believing that in the more highly differentiated animals, like Vertebrates, not all protoplasm is affected to the same degree nor in the same way. Thus a nervous and a muscular effect can be clearly distinguished in frogs, for example. The principal effect is exerted upon the central nervous system, for HERMANN ('86, p. 415) found that the tail of tadpoles is responsive only so long as it contains a piece of spinal nerve; and upon frogs subjected to curare, which inhibits the action of the nerve alone, the current produces a much-diminished effect, giving rise merely to muscular twitchings. (BLASIUS and SCHWEIZER, '93, p. 528.) Very little progress has been made, however, upon the determination of the action of different intensities upon the different tissues of which the Vertebrate body is composed.

We have seen that the electric current provokes a response, and we have seen also that organisms vary in their responsiveness so that a current strong enough to call forth a response in one species is not sufficient to excite another species. We may say that the one species is *attuned* to a different strength of current from the other. This difference in responsiveness indicates, of course, a corresponding difference in composition of the protoplasm. Such a difference may, moreover, be produced in a single individual by artificial means. These means are the subjection for a considerable period to the electric current. Suppose we subject an organism to a current of a strength only slightly greater than that just necessary to provoke a response. After the current has acted for some time we find that it no longer excites. This phenomenon of acclimatization to the galvanic current was first observed among the Protista, so far as I know, by KÜHNE ('64, pp. 76, 78), who found that in Myxomyces, after a few induction shocks had been sent through the plasmodium, additional shocks of the same intensity were without effect, and stronger shocks had to be sent through to cause contraction. Similar results were obtained by VERWORN ('89<sup>a</sup>, p. 10; '89<sup>b</sup>, p. 272) in subjecting *Actinosphaerium*

and Amœba to a weak constant current. At first the pseudopodia on the anode side plainly retracted, but later ceased to do so, and, finally, the current still passing, the retracted pseudopodia began to extend again. The action of the current so modified the protoplasm as to change the attunement of the organism.

### § 3. ELECTROTAXIS

In studying the subject of aggregation with reference to the electric current, we shall consider first the simplest case of this phenomenon as it is exhibited in Amœba; then pass to the more complex forms of Protista, especially the Ciliata, and after

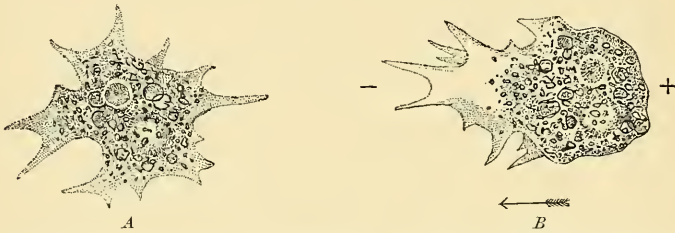


FIG. 32.—Galvanotaxis of *Amœba diffluens*. *A*, *Amœba* creeping, unstimulated; *B*, after closing of the constant current. The arrow indicates the direction of locomotion. (From VERWORN, '95.)

that to the Metazoa. After dealing with the phenomena we must attempt to explain them.

When such an amœba as that shown in Fig. 32, *A*, is subjected in a drop of water to the action of a weak constant current, as already indicated (p. 129), it contracts, especially upon the face turned towards the anode. If the current is not strong enough to produce disintegration at that pole, but only repeated contraction, and if meanwhile the kathode pole retains its power of throwing out pseudopodia, the amœba must gradually move from the anode (Fig. 32, *B*), and if several Amœbæ are under the cover-glass, they will eventually aggregate about the kathode. Here we have, then, in its simplest form, a case of electrotaxis, and, since the organism moves toward the negative electrode, we may call it negative electrotaxis.

If now, instead of an amœba, we watch a free swimming flagellate Infusorian, — *Trachelomonas hispida* (Fig. 33), — we

see the long flagellum which precedes in locomotion coming to lie in the current and directed towards the kathode, so that the animal migrates in that direction. The following explanation of the observed fact that the flagellum becomes directed towards the kathode has been offered by VERWORN ('89<sup>b</sup>, p. 298). The flagellum and the pole from which it arises constitute the most sensitive end of the body. When the flagellum is stimulated it beats violently, and since it is stimulated most when turned towards the anode, it beats most violently when in this attitude. A position  $180^\circ$  from this is one of comparative rest. In intermediate positions the degree of stimulation is intermediate. After a few strokes the body will "naturally" come to assume and to retain that position in which the flagellum is least stimulated.

More detailed still is our knowledge of electro taxis among

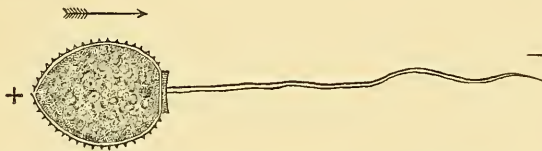


FIG. 33.—*Trachelomonas hispida*, swimming towards the kathode (—) upon closure of the current. The arrow shows the direction of locomotion. (From VERWORN, '89.)

the ciliate Infusoria. The authors who have worked upon this group are chiefly VERWORN ('89<sup>a</sup> and '89<sup>b</sup>) and LUDLOFF ('95). The work of the former shows that the phenomenon of electro taxis is exhibited by many species, especially *Paramecium aurelia* and *P. bursaria*, *Stentor cœrulens* and *S. polymorpha*, *Pleuronema chrysalis*, *Opalina ranarum*, *Bursaria truncatella*, *Halteria grandinella* and *Stylonicchia mytilus*. LUDLOFF employed only *Paramecium*, but studied it much more completely, especially using various currents of known relative intensity.\* He found that the precision with which

\* LUDLOFF employed a trough with wax walls, clay ends, and glass bottom, and used brush electrodes. The intensities of current given by him are the readings of the galvanometer. The cross-section of the water mass in which the *Paramecia* were, and over which the current spread itself, is not exactly given, but was probably about 20 sq. mm. If we employ the unit of strength recommended by HERMANN and MATTHIAS (p. 128), namely, 1 one-millionth of

the Paramecia aggregated at one pole was determined by the strength of the current, as follows: A current of  $3\delta$  caused in general a movement towards the kathode, although many individuals appeared not to be affected by it. In 20 seconds the anode end of the fluid was almost free from Infusoria. With currents of  $6\delta$  and  $15\delta$  the aggregation at one pole became more complete and took place in a short time. Indeed, there was a relation found between the time required for

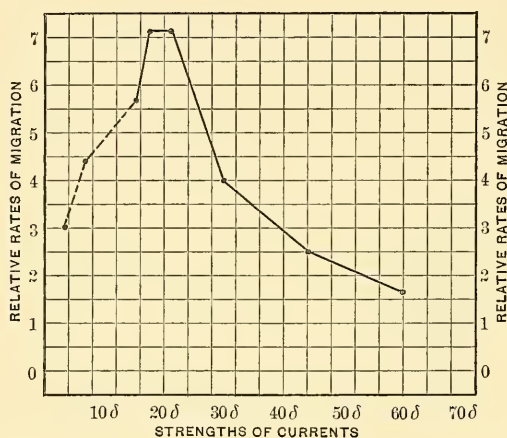


FIG. 34.—Curve showing relation between strength of currents and relative time elapsing before Paramecia have aggregated at the kathode. The ordinates are measured by the reciprocals of the number of seconds elapsing; the abscissæ, by the strength of current in  $\delta$ 's.

aggregation at the kathode and the strength of current employed, which is instructive, and is given above in graphic form (Fig. 34).

This curve shows that as the current increased from  $3\delta$  to  $21\delta$  the rapidity of aggregation increased, but as the current increased still further this rate diminished until locomotion nearly ceased at above  $60\delta$ . The intensity, therefore, of  $21\delta$  produced the most rapid movements.

Upon opening the current, the Infusoria in all cases swim,

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1 ampere per sq. mm. (designated  $\delta$ ), then we must divide LUDLOFF'S galvanometer readings (given in milliamperes) by  $\frac{20}{1000}$ , or (which is the same thing) multiply them by 50. That will give us the current in  $\delta$ 's per sq. mm. All of the numerical data given in the text have undergone this operation.



rapidly for a moment, towards the opposite pole (anode), but then quickly begin to redistribute themselves throughout the water. This redistribution has occurred in about 20 seconds

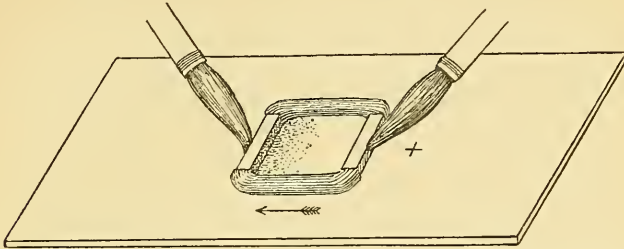


FIG. 35. — Apparatus for studying electrotaxis of *Paramecium*. A rectangular trough, whose ends are of clay and sides of wax, is built upon a glass plate. The current is applied by means of brush electrodes. The direction of the migration of the paramecia is indicated by the arrow. They move towards the kathode. (From VERWORN, '95.)

after the current is broken, and the time is independent of the strength of the preëxisting current.

The movement of the Infusorian from one pole to the other takes place along the lines of flow of the current. If the terminals are two parallel plates, these lines are about parallel (Fig. 35); if they are two points near the opposite sides of

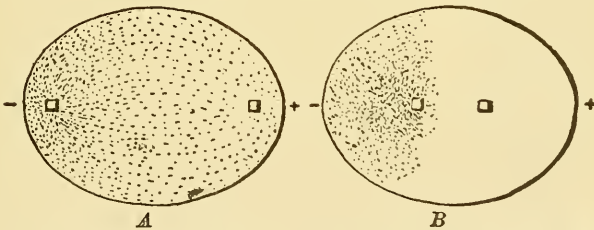


FIG. 36. — Curves made by *Paramecia* in its galvanotactic response when pointed electrodes are used in the drop of water. *A*, beginning of migration; *B*, complete aggregation. (From VERWORN, '95.)

a water drop, the lines have the direction of the lines made by iron filings scattered on a plate over the two poles of a magnet (Fig. 36).

Besides this path of general migration, the form of the path followed by individuals varies with the current. Normally *Paramecium* moves in a long spiral. As the current is in-

creased, however, this spiral becomes shorter, *i.e.* has more turns per centimeter of progression from one pole to the other

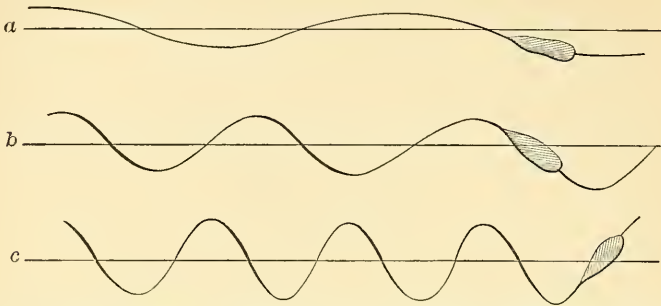


FIG. 37.—Form of the path of Paramecium under different conditions. *a*, when not subjected to the constant current; *b*, when subjected to a slight current; *c*, when subjected to a still stronger one. (From LUDLOFF, '95.)

(Fig. 37), until at  $60 \delta$ , in making one turn of the spiral, the organism progresses hardly more than its own length.

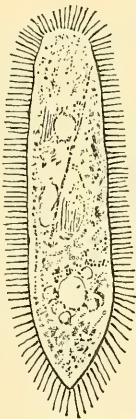


FIG. 38.—Paramecium, showing position of cilia when unstimulated. The blunt end is anterior. (From LUDLOFF, '95.)

Finally, the effect of the current upon the movement of the cilia must be considered.\* In the resting Paramecium the cilia rise perpendicularly from the surface (Fig. 38). If an individual stands with its anterior (blunt) end towards the anode, and a current of  $8 \delta$  passes through, the cilia at the posterior (kathode) end begin to vibrate. If the individual lies transverse to the current and the current is closed, the cilia on the kathode side vibrate, those on the anode side being quiet. With a current of  $16 \delta$  one can see that the kathode stimulation increases the *forward* (anteriad) phase of the cilium movement (the "recovery"). With an intensity of  $24 \delta$ , vibration of cilia occurs at both kathode and anode. It is, however, more

\* LUDLOFF was enabled to make a careful study of the effect of the current on the cilia by making use of gelatine solutions such as have been recommended by JENSEN ('93, p. 556).

intense at the kathode and is also in opposite directions at the two poles. This law is important, and may be thus formulated: *The current intensifies at the anode the backward movements and at the kathode the forward movements of the cilia, and the latter are more intensified than the former*; or, in other words, the anode stimulation increases the effectiveness of the normal stroke, the kathode stimulation diminishes the effectiveness of the normal stroke, and the diminishing effect is the greater of the two.

On the basis of these observed facts, LUDLOFF has proposed a theory which accounts for several of the electrotactic phenomena in the Ciliata, especially the fact that with strong currents there is a diminution in the rate of locomotion, and that at a lower intensity the axis of the organism is placed in the axis of the current, with its anterior end towards the kathode. This theory may be stated as follows: In every complete swing of a cilium two phases may be distinguished—the backward “stroke” and the forward “recovery.” Normally the stroke is the more effective, otherwise forward locomotion would not occur. The excess in effectiveness of the stroke may be designated by the quantity  $x$ . Let us assume a Paramecium lying in the axis of the current with its anterior end towards the kathode. Then the stimulus received at the anode or hinder end increases the effectiveness of the stroke by a quantity which we may designate  $m$ . Thus the excess energy of the stroke over recovery is for these hinder cilia  $x + m$ . The stimulus received at the kathode or anterior end diminishes the effectiveness of the stroke by a quantity which we may call  $n$ , which is larger than  $m$ . Here the excess energy of stroke over recovery is  $x - n$ . If at any intensity of current  $n$  exceeds  $x$ , the anterior cilia will work to oppose the forward motion of the individual, and when  $n - x = x + m$  locomotion will not occur. Such a strength of current probably occurred in the experiment given on p. 142, where locomotion ceased at above  $60 \delta$ .

To account for orientation of the axis and its anterior end, we have merely to apply the general law given above. Let us suppose that we are observing a Paramecium lying in the axis of a current of medium intensity, with its anterior

end towards the anode. Since  $n$  is here supposed to be less than  $x$ , the resultant effect is to move the animal forward. In moving forward in its spiral course, one side becomes presented to the anode. On this side the excess of energy of the stroke is  $x + m$ , while on the kathode side it is  $x - n$ . The resultant effect of the cilia on the two sides forms a couple which revolves the organism about one of its short axes until it comes again into the axis of the current, but with its anterior end towards the kathode. The beating of its cilia must now carry it towards the kathode (Fig. 39).

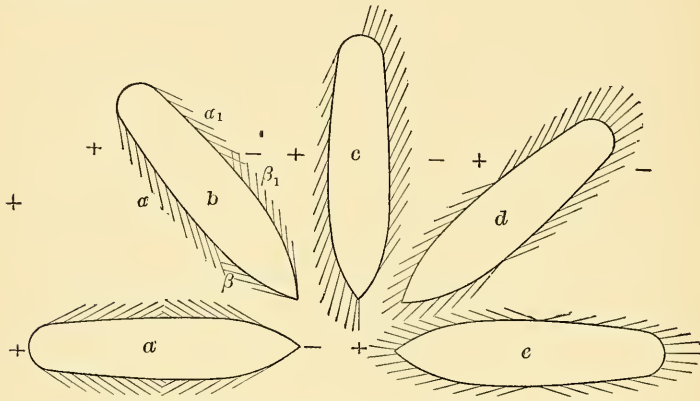


FIG. 39.—Diagram showing the successive attitudes (*a*, *b*, *c*, *d*, and *e*) assumed by *Paramecium* when its head is turned towards the anode at the beginning (*a*). It rotates till its head is next the kathode (*e*). (From LUDLOFF, '95.)

Before leaving the Protozoa, we ought to look over the whole field. We have hitherto considered only cases of migration towards the kathode—negative galvanotaxis. VERWORN ('89<sup>b</sup>), however, has found that some Protista are positively electrotactic; namely, the flagellata, *Polytoma uvella*, *Cryptomonas ovata*, and *Chilomonas paramecium*; the ciliate, *Opalina*; and some bacteria. Finally, VERWORN ('95, p. 446) describes one of the elongated Ciliata—*Spirostomum ambiguum*—which places its long axis across that of the current and migrates towards neither pole, a condition which may be called (after VERWORN) transverse electrotaxis.

Passing now to the Metazoa, we find investigations concerning electrotaxis among Invertebrates by NAGEL ('92 and '92<sup>a</sup>

and '95), and BLASIUS and SCHWEIZER ('93); and among Vertebrates by these authors and also HERMANN ('85 and '86), EWALD ('94, '94<sup>a</sup>, and '94<sup>b</sup>), HERMANN and MATTHIAS ('94), and WALLER ('95). The number of species investigated has been considerable. I give below a table of the Invertebrate genera studied and the sense (+ or -) of their response at the given intensity of currents. Barely rough quantitative expression of strength of current can be deduced from NAGEL's paper. Where no data are given the currents are supposed to be of intermediate strength. N. stands for NAGEL, B.S. for BLASIUS and SCHWEIZER. The numbers which follow give the year of publication and the page.

TABLE XV

SPECIFIC NAME.	STRENGTH OF CURRENT.	SENSE OF RESPONSE.	AUTHORITY.
Mollusca :			
<i>Limnæa stagnalis</i> . . . . .	weak	-	N. '95
Var. other Gastropoda (probably) .		-	N. '92 <sup>a</sup> ; '95
Annelida :			
<i>Lumbricus</i> . . . . .		-	N. '95, 626
<i>Tubifex rivulorum</i> . . . . .		-	N. '95, 631
<i>Hirudo medicinalis</i> . . . . .	0.8 δ	-	B.S. '92, 516
<i>Branchiobdella parasitica</i> . . . . .		-	B.S. '92, 516
Crustacea :			
<i>Cyclops</i> . . . . .	strong	+	N. '92, 629
<i>Asellus aquaticus</i> . . . . .	strong	+	N. '95, 633
<i>Astacus fluviatilis</i> . . . . .	0.4 δ	+	B.S. '92, 518
Insecta :			
<i>Notonecta</i> . . . . .		+?	N. '95, 636
<i>Corixa striata</i> . . . . .		+	N. '95, 636
<i>Dytiscus marginalis</i> . . . . .	1.9 δ	+	B.S. '92, 519
<i>Hydrophilus piceus</i> . . . . .	1.9 δ	-	B.S. '92, 519

From this table it appears that Mollusca and Annelida are usually negatively electrotactic to a current of medium intensity, while Arthropoda are mostly positively electrotactic to such a current. It is noteworthy, however, that two quite

closely allied beetles like *Dytiscus* and *Hydrophilus* should by the same observers be found to react in different ways.

In addition to the species named, some have been studied which have given no results. Thus NAGEL ('95, p. 639) obtained no response from the larva of *Libellula depressa*, even with a wide range of current-intensities.

Passing now to the Vertebrata, we enter a region in which, as a result of more numerous studies, the data are more voluminous, but at the same time less in accord. Since many matters are here still in dispute, we may best consider historically, *i.e.* in chronological sequence, what has hitherto been done in this field.

The first person to describe the phenomenon of electrotaxis in Vertebrates — as, indeed, in any organism — was HERMANN ('85, '86). He used frog larvæ 14 days old, held in a shallow rectangular porcelain trough, along the two small sides of which thick zinc wires were placed, connected with a chain of 20 small zinc-carbon elements. No mention of the strength of the current except such as can be gained from these facts was made. This omission of quantitative details is to be regretted, since had the strength of current to which the organisms were subjected been given, much subsequent confusion might have been avoided. With this current, then, of unknown intensity, flowing through the water containing the larvæ, all of the latter were seen to place themselves in the axis of the current with their heads directed toward the anode. This orientation was the consequence of the fact that a current passing cephalad through the larva acted as a violent stimulus; but while passing caudad it brought stupefaction or even (temporary) paralysis.

The results obtained by HERMANN were now confirmed and extended by BLASIUS and SCHWEIZER ('93). They employed a wooden trough with sheet zinc electrodes of nearly the cross-section of the smaller ends of the trough, and experimented upon fishes, *Salamandra*, and the frog. The weakest current employed was  $0.35 \delta$  to  $0.47 \delta$ , which merely affected the character of the swimming of the fish subjected to it, without determining its direction. The next stronger current mentioned was  $1.58 \delta$ . With this current a marked orientation of

the fish with their heads to the anode was noticed. With *Salamandra* larvæ currents of  $2.3\delta$  to  $4.7\delta$  were chiefly employed. In the experiments of BLASIUS and SCHWEIZER the organisms sometimes migrated toward the anode, if the current was not so strong as to stupefy, but they lay stress upon the point that the migration is a secondary phenomenon — that the orientation is the primary effect of the current.

Next came the observations of EWALD ('94), who used very young tadpoles (5 days) and non-polarizable *points* as electrodes. Thus he was not able to give the strength of current to which the individuals were subjected. His results seemed directly to oppose those of the two preceding authors, for with his (unknown) current, the tadpoles were stimulated when the current passed caudad and stupefied by one passing cephalad; also they placed themselves in the axis of the current with their heads towards the kathode. The larvæ did not seem to find this position as a direct response to stimulus, but whenever an individual, in its turnings to the right and left, fell into this — electrotactic — position it was no longer stimulated, but stupefied, and so came to rest.

These discordant results of EWALD led HERMANN, with his student MATTHIAS, again into the discussion. By careful measurements of the strength of current, they found that between  $0.3\delta$  and  $1.5\delta$  frog tadpoles of from 1 to 3 weeks old did face the kathode, as EWALD found, and did move towards it. But HERMANN and MATTHIAS ('94) believed this result to be due to the fact that only cephalad-flowing currents of this intensity excite, and thus only such currents are able to produce locomotion.

EWALD ('94<sup>b</sup>), however, cannot accept their idea that a caudad-passing current of small intensity produces no excitation, for, he says, he has seen small fish, lying with face to the anode, made to move towards the kathode by the action of the weak current. Since HERMANN and others have shown that very strong currents cause paralysis even when flowing cephalad, EWALD concludes that we must recognize the existence of three different effects at three intensities; weakest, medium, and strongest. The medium current (which has the broadest range) is that by which the organisms are irritated as it flows

cephalad, so that they come to lie with head towards anode; the weakest current is that by which (following EWALD) the organisms are irritated as it flows caudad, so that they come to lie facing the kathode; finally, the strongest current is that at which a violent stimulation leading to paralysis is produced by the cephalad-flowing current (as well as the caudad?).

In seeking for an explanation of electrotaxis in Metazoa, it is necessary, first of all, to notice that there is a close relation between response to the make-shock (as described on pp. 136, 137) and the direction of orientation of the body in electrotaxis. Thus all gastropods studied are, upon making, excited chiefly at the anode, and, correspondingly, all gastropods hitherto studied when subjected to the current face the kathode; so on the other hand, such Crustacea as have been studied are stimulated at the kathode, and they accordingly come to face the anode. In regard to Vertebrates, we have apparently a double electrostatic orientation varying with the current, and correspondingly we have, as NAGEL has shown (p. 137), a double irritability depending on the current. A medium current produces a kathode excitation and an anode orientation; while the weakest current produces an anode excitation and a kathode orientation. So we may lay it down as a general law: *Positively electrostatic organisms exhibit the katex type of irritability; and negatively electrostatic organisms exhibit the anex type or, in general, the electrostatic organism turns tail to the exciting pole.*

EWALD ('94, pp. 611-615) accounts for this difference of response of Vertebrates to weak and strong currents, by the aid of certain observations that he made upon the excitation of the nerve cord. We have already seen that the making of the medium constant current stimulates at the kathode, so that an animal turns tail to the kathode. EWALD found that the two parts of the dorsal nerve were differently stimulated by the current; the brain was stimulated chiefly by a caudad-passing current; the spinal cord chiefly by a cephalad-passing current. This conclusion was established by two experiments. First, the two electrodes, placed a few millimeters apart, are brought into contact with different parts of the body of a fish.



Let the current be passing through the fish from the anterior to the posterior electrode. At the tail end we get no excitation; and, as we pass forward, the body remains quiet until, passing the region of the medulla oblongata, an excitation appears. If the operation is repeated with a reverse current, we get excitation behind the head and quiet on the head. Secondly, if a frog larva be cut in two transversely at the root of the tail, the head end is irritated only by a caudad-flowing current; the tail end only by a cephalad-flowing current; while in both cases the opposite current quiets. (Cf. also EWALD, '94<sup>b</sup>, p. 162.) These observations were now made use of to explain the opposite orientation of the tadpole in the presence of weak and strong currents. NAGEL assumed that the weakest currents can affect the brain only. Now if that current runs caudad, it will strongly stimulate the body so that it turns tail to the anode. The medium currents, however, stimulate the whole dorsal nerve, but the spinal cord to a preponderating degree, so that a cephalad-passing current irritates more than a caudad current, and the animal will turn tail to the kathode. Thus weaker or stronger current will determine — or + electrotaxis.

#### SUMMARY OF THE CHAPTER

Electricity affects protoplasm in two principal ways: first, by causing contraction; second, by determining orientation. We can distinguish two principal types of contraction phenomena and, corresponding to and dependent upon these, two types of orientation phenomena. The first type of contraction is that which is produced, upon making the constant current, chiefly at the anode; the second is produced chiefly at the kathode. The corresponding orientation or migration types are, facing the kathode and facing the anode. Since the orientation phenomena are dependent upon the contraction phenomena, the most important causes to be investigated are, first, that of contraction, and second, that of the difference in type of contraction exhibited by different organisms. The fundamental teaching of this chapter is that the electric current acts as a stimulus upon protoplasm, and may determine the character of its activities.

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## CHAPTER VII

### *ACTION OF LIGHT UPON PROTOPLASM*

IN this chapter it is proposed to discuss (I) the application and measurement of light; (II) its chemical action; (III) the effect of light upon the general functions of organisms; and (IV) the control of locomotion by light—phototaxis and photopathy.

#### § 1. THE APPLICATION AND MEASUREMENT OF LIGHT

Light, which as a form of radiant energy is closely related to radiant heat, is always accompanied by a certain quantity of heat, whose action (in at least one control experiment in every set) should be eliminated. To cut out heat without great loss of light, we must employ transparent adiathermal media. Of these, a plate of ice is the most effective, but alum, on account of its higher melting point, is more convenient. A parallel-sided vessel full of distilled water, or, still better, a saturated aqueous solution of alum, forms an inexpensive, highly adiathermal screen.

The quality of the light used in any experiment should be carefully determined. If any other light than that of the sun or incandescent solids is employed, it should be subjected to spectroscopic analysis. For biological purposes a direct-vision hand spectroscope, such as BROWNING'S, is convenient and adequate.

Often *monochromatic light* or a definite range of the spectrum is desired. This may be obtained in various ways. The purest monochromatic light can be got by making a long spectrum and using the desired part of it. To make such a spectrum one may employ, in a dark room, a lamp, followed in succession by a slit and a lens to form an image of the slit

on a prism of bisulphide of carbon,\* which gives very great dispersion of the rays.

In defining the regions of the solar spectrum which are employed in any study, it is usual to make reference to the dark absorption bands (FRAUENHOFER'S lines) which cross the solar spectrum. The largest of these are lettered, beginning with *A* in the visible red and ending with *H* in the visible violet. At other times it may be more convenient to define any part of the spectrum by means of the extreme wave lengths between which it lies. Lithographs showing the spectral colors and the wave lengths corresponding thereto are given in encyclopædias and most of the text-books on physics. A crude attempt is made to show the relation between color and wave length in Fig. 40. The wave lengths at

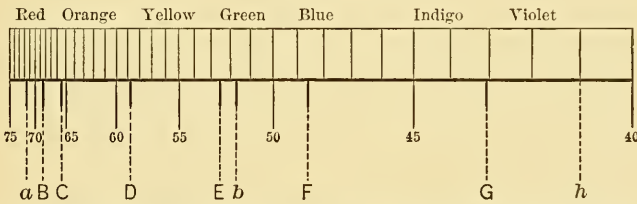


FIG. 40.—Diagram of the solar spectrum showing the main absorption bands and the range of the various spectral colors. The numbers are wave lengths in hundred-thousandths of a millimeter. (From REINKE, '84.)

the different absorption bands are given more exactly (in thousandths of a millimeter, =  $\mu$ ) in the following table, and also the number of waves per second in  $10^{12}$ ths.

TABLE XVI

ABSORPTION BAND.	WAVE LENGTH, $\lambda$ .	VIBRATIONS PER SECOND $n \times 10^{12}$ .	ABSORPTION BAND.	WAVE LENGTH, $\lambda$ .	VIBRATIONS PER SECOND $n \times 10^{12}$ .
A . . . . .	0.760 $\mu$	392	E . . . . .	0.527 $\mu$	566
B . . . . .	0.687 $\mu$	433	F . . . . .	0.486 $\mu$	613
C . . . . .	0.656 $\mu$	454	G . . . . .	0.431 $\mu$	692
D . . . . .	0.589 $\mu$	506	H . . . . .	0.397 $\mu$	751

\* The bisulphide prism may be made as follows: Upon a thick glass plate three rectangular pieces of glass of equal size are placed perpendicularly, so as

Extreme ultra-violet  $\lambda = 0.295 \mu$ ;  $1010 \times 10^{12}$  vibrations per second.

To obtain monochromatic light from the spectrum, REINKE'S ('84) spectrophor will be found useful (Fig. 41). In this instrument a beam of sunlight cast by a heliostat through a slit at  $H$  is converged by means of an interpolated lens,  $O$ , upon a prism,  $P$ , set at the angle of minimum deviation. Passing through this prism the rays are dispersed and a spectrum is formed upon a diaphragm  $D$ .  $D_1$  composed of halves bounding a second slit whose position and width may be varied at will so as to include any desired part of the spectrum. A large lens  $C$  known as the collector brings the rays which have passed through the second slit to a focus at  $E$ . Just in

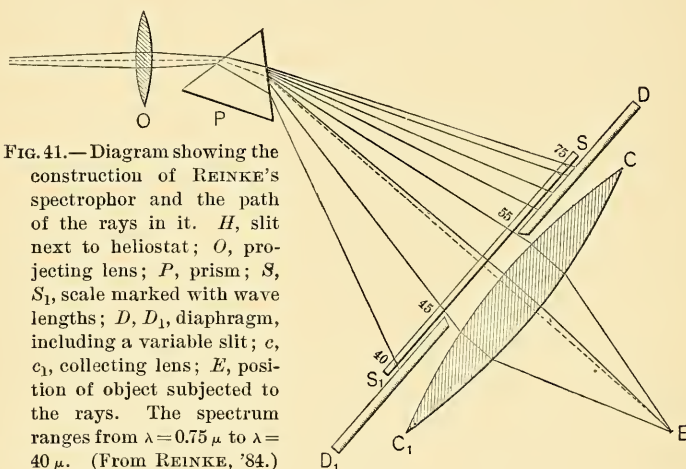


FIG. 41.—Diagram showing the construction of REINKE'S spectrophor and the path of the rays in it.  $H$ , slit next to heliostat;  $O$ , projecting lens;  $P$ , prism;  $S$ ,  $S_1$ , scale marked with wave lengths;  $D$ ,  $D_1$ , diaphragm, including a variable slit;  $c$ ,  $c_1$ , collecting lens;  $E$ , position of object subjected to the rays. The spectrum ranges from  $\lambda = 0.75 \mu$  to  $\lambda = 40 \mu$ . (From REINKE, '84.)

front of the diaphragm is placed a scale  $S$ ,  $S_1$  of wave lengths fitted to the spectrum obtained. Such a scale may be constructed with reference to the position of FRAUENHOFER'S lines by interpolation from Fig. 40.\* To include rays whose wave lengths differ by exactly  $0.05 \mu$  the slit must be wider when at the blue end than when at the red end of the spectrum (Fig. 40).

to form a hollow, triangular ( $60^\circ$ ) prism. The plates are fixed to each other and to the glass base by a pasty cement made by mixing plaster of paris and liquid glue. This cement soon hardens, and is not attacked by the carbon disulphide. The hollow prism is now filled with fluid, and a triangular glass plate is cemented on as a cover.

\* If artificial light is used, two points on the scale can be obtained as follows: Volatilize in a Bunsen flame, temporarily replacing the lamp, a salt of barium and one of calcium, and note the position of the extreme blue band on the former (line  $F$ ) and the yellow band of the latter (line  $D$ ). After determining these two points the remaining lines can be plotted upon the scale.

A second method of getting monochromatic light is by the use of flames tinged with various volatilized metals. Of these, lithium salts give reds at  $\lambda=0.67 \mu$  and  $\lambda=0.61 \mu$ , sodium salts give a pure yellow light of  $\lambda=0.59 \mu$ , thallium salts (poisonous vapor) a green at about  $\lambda=0.54 \mu$ , and indium salts a blue and a violet, both beyond  $\lambda=0.46 \mu$ . The number of metallic vapors which give even nearly monochromatic light is, however, not large.

A third method for producing monochromatic light is found in the use of solutions of pigments. Such solutions may be held in deep glass vessels whose back and front glass surfaces are parallel and near together. In the following table are given in the first column the names which may be applied to different parts of the spectrum, following HELMHOLTZ (Handbuch, p. 251); in the second column the pigments which while dry give corresponding spectral colors in diffuse daylight and which may also be used in making solutions; and in the third column certain pigments which in solution YUNG ('78, p. 251) found to transmit almost exclusively the part of the spectrum named in the first column.

TABLE XVII

From outer limit to line <i>C</i> , red . . .	Cinnibar, HgS (vermilion)	Alcoholic solution fuchsin *
<i>C</i> to <i>D</i> , { orange . . . . .	Minium	
{ golden yellow . . .	Litharge, PbO	
<i>D</i> to <i>E</i> , { yellow . . . . .	Chrome yellow,	Concen. Sol. potassic chromate
{ yellow-green . . . .	PbO <sub>2</sub> CrO <sub>2</sub>	(a little red and green)
<i>E</i> to <i>b</i> , green . . . . .	cupric arsenite, SCHEEL's green	Nickel nitrate, NiO <sub>2</sub> (NO <sub>2</sub> ) <sub>2</sub>
<i>b</i> to <i>F</i> , transition from blue-green to blue		
<i>F</i> to <i>F</i> $\frac{1}{2}$ <i>G</i> , cyanite blue . . . .	Berlin blue	Bleu de Lyon (a little V) †
<i>F</i> $\frac{1}{2}$ <i>G</i> to <i>G</i> , indigo blue . . . .	Ultramarine	
<i>G</i> to <i>H</i> , violet . . . . .		Violet de Parme

Solutions made up from these pigments should, however, be examined spectroscopically before using to make sure of the purity of the color.

\* Also a solution of iodine in carbon disulphide. (PRINGSHEIM, '80, p. 409.)

† *F* to *H* is given by ammoniated copper sulphate, CuSO<sub>4</sub>·4 NH<sub>3</sub> + H<sub>2</sub>O (PRINGSHEIM).

Finally, a fourth and decidedly practical way of obtaining pure colors is by the use of transparent plates of colored glass or other transparent solids. It is very difficult to get monochromatic glasses of certain colors in the market. A pure red is easily obtainable; the blue is apt to contain some red also; and the green, both blue and yellow. Lord RAYLEIGH ('81, p. 64) has used "films of gelatine or of collodion, spread upon glass and impregnated with various dyes, gelatine being chosen when the dye is soluble in water and collodion when the dye is soluble in alcohol." This method seems to me to be of wide applicability in our light experiments. For solid media are, after all, far less troublesome than fluids, vapors, or spectra; and convenience is one of the most valuable qualities of a method.

A brief statement must be made concerning the physical properties of the different light waves. An inspection of any prismatic solar spectrum shows that certain parts are brighter to our eyes than others, and a thermometer placed in different parts of the spectrum indicates a higher temperature towards the red end. Curves are given in Fig. 42 which show the *relative* warmth of different parts of the visible spectrum both when the spectrum is a normal one (*i.e.* such as is given by a diffraction grating, where all rays differing in wave length by  $0.1 \mu$  are equally distant) and when it is prismatic (in which there is a crowding of rays at the red end). Curves of relative brightness and of relative chemical (actinic) activity, so far as can be judged from the union of chlorine and hydrogen, are also given, for the *prismatic* spectrum. Being laid off on the "normal" scale the curves last mentioned are somewhat distorted. From these curves it appears that the brightest part of the spectrum lies between lines *D* and *E*, at  $\lambda = 0.59 \mu$ ;<sup>\*</sup> the warmest part is, in the normal spectrum, near  $\lambda = 0.60 \mu$ , but in the prismatic spectrum, beyond the visible red, at about  $\lambda = 1.00 \mu$ . Finally, the chemical activity of the rays increases towards the blue end of the spectrum, but the relative activity is different for the different substances acted upon. Measured by their ability to unite chlorine and hydrogen, the rays having

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\* MENGARINI ('89, p. 135) finds the point of maximum brightness to lie at about  $0.57 \mu$ .



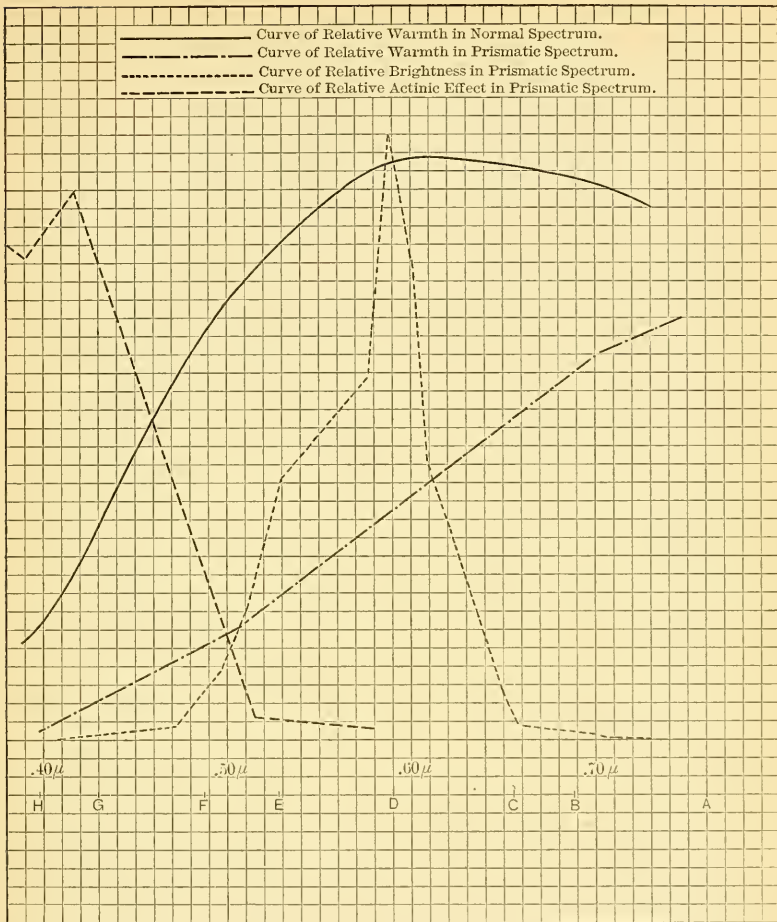


FIG. 42. — Scale of *normal* solar spectrum, above which is drawn the normal curve of relative warmth; also the curves of relative warmth, brightness, and actinism of the *prismatic* solar spectrum. The curves of warmth are taken from LANGLEY ('84, p. 233); the curve of brightness is constructed from the data of VIERORDT ('73, p. 17); that of actinism is taken from BUNSEN and ROSCOE ('59, p. 268) and indicates the relative efficiency of the different rays of the midday sun in causing the union of chlorine and hydrogen. The absolute value of the ordinates is entirely arbitrary.

a longer wave length than  $0.51 \mu$  have feeble chemical action; at about  $\lambda = 0.42 \mu$  this action reaches a maximum.

Not only the quality but also the intensity of the light with which we experiment must be known. It is, fortunately, quite

easy to determine the intensity of white light in terms of a recognized unit; namely, a paraffine candle burning at the rate of 7.78 grammes per hour. A paraffine candle burning at this rate has one candle power (C. P.); burning at twice this rate, 2 candle power, and so on. A comparison of any other light with this standard may be made by means of any of the well-known photometers, of which text-books of physics give a description.

The determination of the intensity of a *colored* light requires an additional piece of apparatus; namely, a spectrophotometer. The two principal types of spectrophotometer are that of VIERORDT ('73) and that of GLAN ('77), both of which have undergone important improvements. The principle in both types is the same. A spectrum of both the unmodified (standard) light and that which has passed through the colored screen are made side by side, so that their corresponding colors can be compared. Since the source of light is the same, every part of the spectrum of the unmodified light will be brighter than the corresponding part of the spectrum of the colored light. To bring the corresponding colors in the two spectra to the same intensity, the unmodified light must be made less intense to a measurable extent. In VIERORDT's spectrophotometer this result is brought about by narrowing that half of the slit through which the unmodified light passes to get to the prism. In GLAN's apparatus the diminution in intensity is gained by the polarization of both lights and the obscuring of the brighter by the rotation of its analyzing NICHOL prism, until equality of brightness is obtained. A modified form of VIERORDT's convenient instrument is made by H. KRÜSS of Hamburg, Germany. A modified form of GLAN's photometer is described by VOGEL ('77).

VOGEL's apparatus (Fig. 43) consists essentially of a collimator containing (1) a slit of changeable width, separated by a band  $q$  into an upper and a lower part to receive respectively the modified and the normal light; (2) a lens to render the rays parallel before they impinge upon (3) a doubly refracting quartz prism, by which both upper and lower rays are broken into two polarized rays. Of these four rays the uppermost and the lowest are cut off by a diaphragm near  $F$ , so that only the middle two, which lie near together, pass eventually to the eye. These two rays are oppositely polarized and come, one from the upper, the other from the lower slit. The two

rays now pass through (4) a NICHOL prism (capable of being rotated alongside a graduated arc) set at  $45^\circ$ , in which position both rays pass through without changed relative intensity. The rays emerging from

the collecting telescope are now dispersed by passing through the vertically placed prism, and the adjacent parallel spectra are observed through a telescope. By a rotation of the

NICHOL prism through an observed number of degrees the stronger light may be brought to the intensity of the weaker. The relative intensity of two lights with reference to a third (constant) is as the squares of the tangent of the angle through which the NICHOL prism has been rotated. Other modifications of

GLAN'S photometer are those of Lord RAYLEIGH ('81) and of LEA ('85), upon which the spectrophotometer of the Cambridge (Eng.) Scientific Instrument Co. is based.

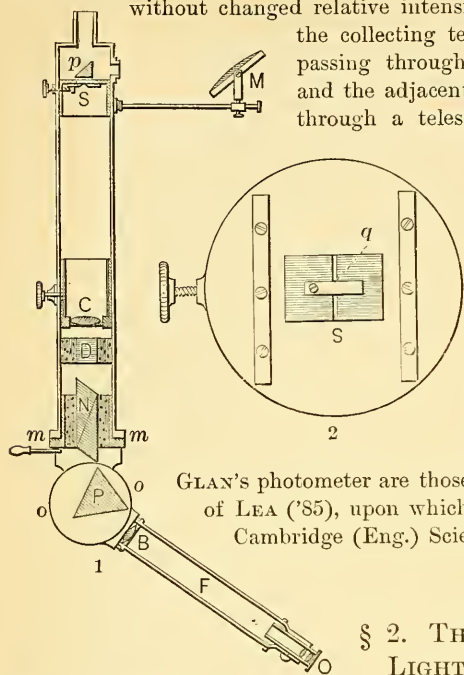


FIG. 43. — Diagrams showing construction of VOGEL'S spectrophotometer. 1. Horizontal section through the optical axis. *M*, mirror to reflect standard light, by aid of a totally reflecting prism *p*, into the optical axis. *S*, shutter with slit divided into an upper and a lower half by means of a band *q*; *C*, collimating lens. *D*, doubly refracting quartz prism; *m*, *m*, the holder of the NICHOL prism *N*, which can be rotated through an arc that can be read off from a graduation on *m*, *m*; *P*, a flint glass prism; *B*, *F*, *O*, observing telescope, in which, at the focal point, near *F*, is a diaphragm cutting out the two outermost of the four spectra coming through *B*. 2. Front view of the shutter. (From VOGEL, '77.)

## § 2. THE CHEMICAL ACTION OF LIGHT UPON NON-LIVING SUBSTANCES

The process of photography has made us familiar with the fact that daylight acts upon the halogen salts of silver, gold, platinum, and other metals, although the nature of the chemical change wrought by the light is uncertain. It is not, perhaps, generally appreciated, but it is well known to chemists, that light can produce or further very many chemical changes, particularly among organic compounds. The effects are mainly due to the blue and violet rays, hence are

not the results of the heat of sunlight. Most of these chemical effects may be grouped under four heads. 1. Synthetic; 2. Analytic; 3. Substitutional; and 4. Isomerismic and Polymerismic. A few others may be classed (5) as fermentative. Let us now consider each of these five classes.\*

1. **The Synthetic Effects of Light** will be considered chiefly with reference to organic compounds. All the cases I have gathered fall into three groups: addition to the organic compound either ( $\alpha$ ) of oxygen, ( $\beta$ ) of chlorine or bromine, or ( $\gamma$ ) of another organic compound.

Among the compounds which take up oxygen is bilirubin,  $C_{32}H_{36}N_4O_6$ , a solution of which, in sunlight, even when air is excluded, oxidizes to biliverdin,  $C_{32}H_{36}N_4O_8$ . In the absence of sunlight this change requires air (B. III, 418). DUCLAUX ('87, p. 353) finds that vegetable oils, such as olive or palm oils, are rapidly oxidized if exposed to light. CHASTAIGN ('77, p. 198) believes this oxidizing action of light upon organic compounds to be of very wide-spread occurrence; the blue-violet part of the spectrum being, in this respect, the most active.

The direct combination by means of light of a halogen and another substance is also not rare. Thus, in daylight, hydrogen unites with chlorine explosively. It unites with bromine also, although with difficulty. Similarly, equal volumes of chlorine and carbon monoxide unite quickly in the sunlight or magnesium light to form carbon monoxid chloride,  $COCl_2$  (B. I, 546). Again, when chlorine is passed through alcohol under the influence of strong sunlight or magnesium light the two substances unite and produce chloral hydrate (STREET and FRANZ, '70). Likewise, when chlorine is passed, in sunlight, through a solution of  $C_3H_2Cl_2O_2$  in  $CS_2$ , there is formed  $C_3H_2Cl_4O_2$ , two atoms of Cl having been added. Finally,  $C_2Cl_6$  may be made by uniting  $C_2Cl_4$  and  $Cl_2$  in sunlight (B. I, 158); and the compound  $C_2H_4 \cdot FeBr_2 \cdot 2H_2O$  may be made by passing, in sunlight,  $C_2H_4$  through a concentrated aqueous solution of  $FeBr_2$  (B. I, 113).

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\* Most of these cases were obtained by searching through BEILSTEIN ('86-'93). References to this book will be made throughout this section by the letter B, followed by the number of the volume and page upon which the statement may be found.

Important cases of the direct synthesis of organic compounds are given by KLINGER and STANDKE ('91). These authors have shown that in sunlight (and not in the dark) phenanthrenchinoxin unites directly with benzaldehyde to form a third compound phenanthrenhydrochinonmonobenzoat, in accordance with the formula:  $C_{14}H_8O_2 + C_6H_5CHO = C_{14}H_8(OH)(O \cdot CO C_6H_5)$ . Again, chinon (benzochinon) and benzaldehyde may unite in the sunlight to form benzohydrochinon, according to the formula:  $C_6H_4O_2 + C_6H_5CHO = C_6H_5CO C_6H_3(OH)_2$ . Finally, benzochinon and isovaleraldehyde may similarly unite to form isovalerochinhydrin, thus:  $C_6H_4O_2 + C_4H_9CHO = C_4H_9CO C_6H_3(OH)_2$ . These cases, then, are examples of organic compounds which are wholly indifferent in the dark, but which, subjected to strong sunlight, lose their identity by uniting directly; they may suffice to illustrate the important synthetic effect of sunlight on non-living, organic compounds.

2. **Analytic Effect of Light.** — Cases of this effect are numerous, varied, and striking. I will cite a few. The organic dibasic acids  $C_nH_{2n-2}O_4$  break up in the sunlight and in the presence of a small quantity of uranium oxide, into  $CO_2$  and an acid  $C_nH_{2n}O_2$  (B. I, 63). For example, oxalic acid,  $C_2H_2O_4$ , breaks up thus into formic acid,  $CH_2O_2$  and  $CO_2$ . Also an aqueous solution of butyric acid,  $C_4H_8O_2$ , in the presence of uranyl nitrate, breaks up, in the sunlight, into  $CO_2$  and  $C_3H_8$  (B. I, 422). We have seen that chlorine will unite directly with organic compounds under the influence of light; on the other hand, compounds containing chlorine may lose it in the sunlight. Thus under these conditions the ketone  $(C_8H_{17}NO \cdot HCl)_2PtCl_4$ , an ammoniacal derivative of acetone, becomes  $(C_8H_{17}NO \cdot HCl)_2PtCl_2$ ; and  $(C_9H_{17}NO \cdot HCl)_2PtCl_4$  becomes  $(C_9H_{17}NO \cdot HCl)_2PtCl_2$  (B. I, 982, 983). Again, chlorine acetate,  $Cl \cdot O \cdot C_2H_3O$ , undergoes slow decomposition in the light (B. I, 462);  $C_5H_6Cl_2$ , a derivative of pentine,  $C_5H_8$ , does the same; and ethylester,  $ClO \cdot C_2H_5$ , explodes in sunlight. Similarly explosive in sunlight is the greenish oil distilled when absolute alcohol is poured over dry calcium chloride (B. I, 223). Finally, sugar (DUCLAUX, '86, p. 881) and oxalic acid (DOWNES and BLUNT, '79, p. 209) are oxidized and break up into water, carbon dioxide, and other compounds. These cases may serve

to show the important chemical effects of sunlight in the disintegration of organic compounds.

3. **Substitution Effects of Light.** — The principal substitution effect of light is the replacement of hydrogen in an organic compound by either chlorine or bromine. This occurs so frequently that examples are superfluous. The substitution takes place most rapidly and completely in direct *sunlight*, and it has been shown that the rays at the blue end of the spectrum are the most active in this process. The compounds affected belong to the most varied groups of both the fatty and aromatic series — carbohydrates, acids, aldehydes, ketones, and sulphides.

4. **The Isomerismic and Polymerismic Changes produced by Light** are among the most interesting. I will cite some examples. In the first place, it may be said that the changes in the elements phosphorus and sulphur by which they assume their red form have been ascribed to sunlight. Elæomargin acid,  $C_{17}H_{30}O_2$ , is a compound found in connection with glycerine in the oil of the seeds of *Elæococca* (*Aleurites*) *vernica* — Chinese oil tree — one of the *Euphorbiaceæ*. This acid crystallizes in rhombic plates which melt at  $48^\circ$ . When an alcoholic solution of this acid is placed in a bright light, leaf-like crystals of its isomere, elæostearin acid, which melt at  $71^\circ$ , are produced (B. I, 535). Again, thymochinon forms yellow crystals, which are soluble in alcohol. Subjected to a strong light, opaque, whitish-yellow crystals are produced, which are insoluble in alcohol. This substance, which does not arise in the dark, and is hence not merely the result of oxidation, is called polythymochinon (B. III, 180). Again, among the derivatives of ethylene,  $C_2H_4$ , is chlorethylene,  $C_2H_3Cl$ , a gas. When placed in the sunlight this passes into a polymere, which forms a viscous, amorphous, insoluble mass (B. I, 158). In like fashion, bromethylene,  $C_2H_3Br$ , a fluid, is rapidly transformed in the sunlight into a polymere, which is solid, amorphous, and insoluble in water, alcohol, or ether (B. I, 181), and bromacetylen,  $C_2HBr$ , a gas, is gradually transformed, in the light, into a solid polymere. Finally, very many substances undergo a gradual change of color in the sunlight, but the nature of the accompanying molecular change is unknown.

5. Changes resembling those brought about by *fermentation* are produced by light. Thus NIEPCE DE SAINT VICTOR and CORVISART ('59) have found that a 0.1% solution of starch, exposed during 6 to 18 hours to the summer sun, becomes transformed into sugar, while in the dark no such change occurs. The change is favored by a small quantity of uranium nitrate. In a similar fashion glycogen is transformed into sugar more rapidly in the light than in the dark. On the other hand, GREEN ('94) finds that the ferment which normally transforms starch into sugar is destroyed by subjection to a strong light, the violet rays being especially active in this process. Likewise, ptyalin, the ferment of saliva, is destroyed by light.

To sum up, light affects organic compounds in very varied and important ways. We are, accordingly, prepared to find that light exerts a very important influence on the activities of protoplasm. Nor is the influence necessarily confined to the surface, for most protoplasmic bodies are more or less translucent. Thus SACHS ('60) found by looking through a tube with one end fitted to the eye and the other directed towards the sunlight, that considerable layers of plant tissue, for example over 32 mm. of the tissue of the potato tuber, did not cut out all the light, and that red had the greatest penetrating power, violet the least. Even the epidermis of man permits light to pass, and OXIMUS ('95) asserts that light can pass through the hand to such an extent as to affect during 26 to 30 minutes an orthochromatic plate kept in a tight wooden box perforated only by the opening which is covered by the hand. Whether the "RÖNTGEN rays," which have so striking a power of penetrating organic matters, are more of the nature of light than of other physical agents, is still a subject of debate. Whether they produce any important chemical changes in protoplasm has not yet been fully determined.\*

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\* During the six months which have elapsed since the above was written, accounts of marked physiological effects of the RÖNTGEN rays have been published. Thus, exposure of the skin to them for an hour frequently causes loss of hair and finger nails, and produces symptoms resembling those of sunburn. AXENFELD (Centralbl. f. Physiol. X, 147) finds that many insects and a crustacean, Porcellius, kept in a box only one-half of which is penetrated by the rays, aggregated in this part. Several experiments upon the tropic influences of the rays have resulted negatively.

### § 3. THE EFFECT OF LIGHT UPON THE GENERAL FUNCTIONS OF ORGANISMS

In this section we shall consider in succession (1) the effect of light upon metabolism; (2) the vital limits of light action on protoplasm; and (3) the effect of light upon the movement of protoplasm.

1. **Effect of Light upon Metabolism** (including Assimilation).—Metabolism is a complex of chemical processes. Since, as we have already seen, light has important chemical effects, we are not surprised to find that it plays an important role in metabolism. The effects of light are, however, of two distinct kinds. One is a thermic effect, due to the heat rays of white light; the other is a chemical effect due to the “actinic rays” of the spectrum.

*a. The Thermic Effect of Light on Metabolism* is shown chiefly in the assimilative processes of chlorophyllaceous plants. The facts of this assimilation are chiefly these: various simple compounds, water, carbon dioxide, salts of ammonia and nitrates, are used as food by plants. For every volume of the gas—carbon dioxide—taken in, one volume (nearly) of oxygen is excreted. Starch ( $C_6H_{10}O_5$ ) is the first visible product of the water and carbon dioxide taken in. Chlorophyll is essential to the absorption of carbon dioxide, to the giving forth of oxygen, and to the formation of starch. Finally, chlorophyll can assimilate only in the presence of sunlight and at a proper temperature.

Now, not all the rays of sunlight with their varied wave lengths are essential to this process. Just what rays are the essential ones has been a point of some dispute. The earlier studies on the subject, made chiefly by DRAPER ('44), SACHS ('64), and PFEFFER ('71), were unanimous in declaring that the most active rays in assimilation were those occupying the yellow part of the spectrum at about line *D*—the region of maximum brightness to our eyes (Fig. 42). But these observers were at fault in that, while they carefully determined the *quality* of light and the corresponding quantity of assimilation, none of them gave, in the experiments with color screens, any adequate data upon the *intensity* of the diversely colored



lights employed; and this is a fundamental matter, for it has been shown, for instance by REINKE ('83 and '84), that, within certain limits, the rate of assimilation increases with the intensity of the light (Fig. 44). Even in experiments with the colors of prismatic spectra one must remember that the rays are crowded together at the red end, so that a given length of the spectrum contains more rays at that end than at the other (cf. Fig. 40).

Later investigations with improved methods have shown quite conclusively that it is especially the rays with  $\lambda = 0.68 \mu$ , or those very close to the absorption band  $B$ , which are most

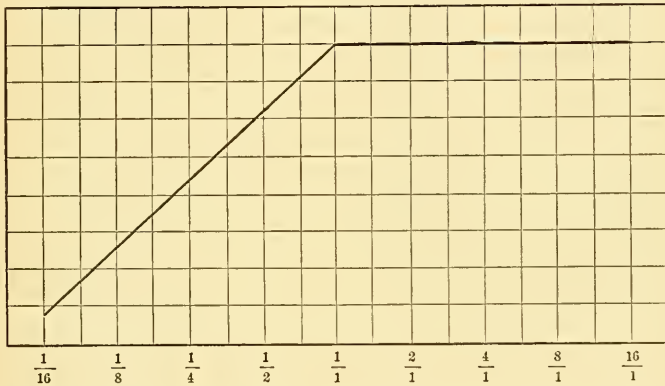


FIG. 44.—Curve showing the relation between intensity of light (abscisse) and quantity of oxygen set free by *Elodea canadensis*.  $\frac{1}{16}$  indicates the unit intensity of the light from the heliostat. (From REINKE, '84.)

active in assimilation. The methods employed have been most diverse, but they have yielded the same result. TIMIRIAZEFF ('77) studied the assimilative power of the different parts of the solar prismatic spectrum, determining by gasometric methods the quantity of gases decomposed in a given time. REINKE ('84) also used the spectrum, but by means of his spectrophor was able to get more strictly monochromatic light, to use more nearly comparable extents of the spectrum, and, especially, to get a more exactly comparable (in this case, optimum) assimilative intensity for each part of the spectrum than his predecessors. (See p. 156.) As the measure of assimilation, REINKE used the number of gas bubbles set free

per minute by the submerged, illuminated plant. As is shown in Fig. 45, the maximum of gas production occurred at about

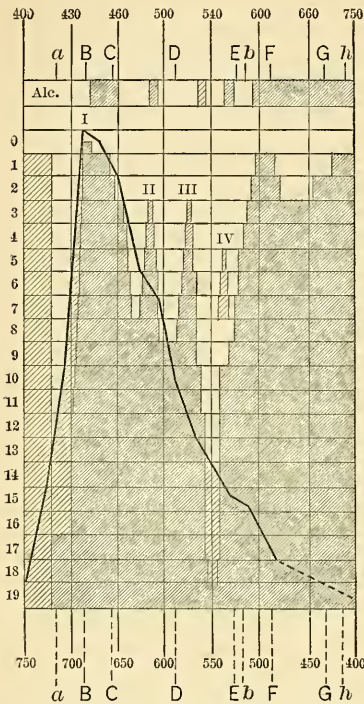


FIG. 45.—Curve whose ordinates are proportional to the number of gas bubbles eliminated per minute by leaves illuminated by the various rays whose wave lengths are given at the bottom of the diagram, and whose number of vibrations per second are given at the top. The background of the figure is composed of the absorption spectra of the chlorophyll in living leaves. 1, 2, etc., at the left, indicate the number of leaves of *Impatiens parviflora*, which, when superimposed, give the corresponding spectrum at the right of these numbers. The absorption at 0 is from a fern prothallus, that at *Alc* is derived from an alcoholic solution of chlorophyll. I to IV indicate absorption bands. Beyond *F* there is very general absorption of the highly refractive rays. (After REINKE, '84.)

absorption line *B*—and this is the more marked of the absorption bands of chlorophyll.

A similar result was reached by ENGELMANN and set forth in a long series of papers ('81, '82, '82<sup>a</sup>, '83, '83<sup>a</sup>, '84, '86, and '87). He found that certain bacteria are extremely sensitive to oxygen, moving in the direction of small increments of the oxygen density. Now, by putting a thread of alga in the same water with bacteria and subjecting the thread to a "microspectrum," that part in which assimilation is proceeding most rapidly, and in which, therefore, oxygen is being most rapidly excreted, will be indicated by the greatest aggregation of bacteria. The microspectrum was produced by means of an apparatus especially designed by ENGELMANN for his work and now manufactured by the ZEISS firm in Jena. The appearances seen under the microscope when the spectrum falls upon the alga in the bacterium-water are shown in Fig. 46. The maximum aggregation (hence, maximum assimilative activity) at the red end occurs

close to the absorption band of chlorophyll. Observations upon the chlorophylls of brown, blue-green, and red cells, which, as ENGELMANN'S microspectro-photometer indicated, have a maximum absorption at other points, showed a maximum of assimilative activity at these other absorption points. In bacterio-purpurin also, in which some of the most active assimilative rays are those of the invisible red at about  $\lambda = 0.85 \mu$ , most oxygen is produced at this point. (ENGELMANN, '83, p. 709.)

Finally, by an ingeniously devised experiment, TIMIRIAZEFF ('90) has settled this matter in the most direct and indubitable

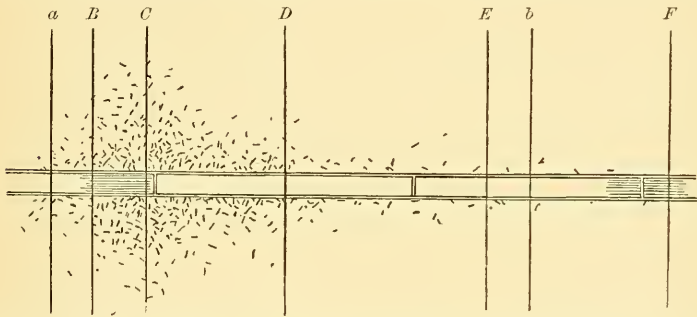


FIG. 46.—Piece of *Cladophora* with swarming bacteria in the microspectrum (gas-light). The chlorophyll grains which fill the cells very uniformly are omitted; and, instead, the absorption band between *B* and *C*, and the tolerably pronounced band at the violet end between *E* and *F*, are indicated by shading. (From ENGELMANN, '82.)

fashion. He kept a plant for two or three days in the dark, until the starch in its leaves had gone; then, in a dark room, a prismatic spectrum was thrown upon the leaf and the position of FRAUENHOFER'S lines indicated on the leaf. After from three to six hours, starch had formed, under the influence of the light, only in the region of the absorption bands of chlorophyll lying between *B* and *D*. This was determined by plunging the leaf into boiling alcohol, thus decolorizing it, and then staining in tincture of iodine, which combines especially with the starch. The deeply dyed places, where starch had been formed, reproduced the absorption spectra of chlorophyll.

The concurrent testimony of these and other observers working upon so diverse material and with such excellent methods

justifies the conclusion that it is the rays absorbed by the plant pigments which enable them to do their work in the decomposition of carbon dioxide. The effective absorbed rays are, moreover, chiefly those towards the red end of the spectrum, those having over  $525 \times 10^{12}$  vibrations per second (*i.e.* below the *D* line).\*

In conclusion it may be said that the greater proportion of the radiant energy entering the plant tissue is absorbed. Thus MAYER ('93) has shown that of dark radiant heat at  $100^{\circ}$  about 80% is absorbed by a leaf through which it passes, and this proportion is about the same whether the leaf is thick or thin. Of this absorbed heat perhaps less than 10% is absorbed by the chlorophyll. The rest must be used up in the vital processes other than assimilation.

*b. The Chemical Effect of Light on Metabolism* must now be considered; and of this we must notice at the outset two degrees. The greater effect, which is a fatal one and the better known, will be treated of further on. The lesser effect is less striking, yet it must be included in the greater. It shows itself in a disturbance of metabolism.

This disturbance of metabolism is evinced in some green plants by heightened production of carbon dioxide and the formation of chlorophyll; and it is noteworthy that a similar result occurs among Infusoria, according to the observations of FATIGATI ('79), who finds the violet rays more active than the green in this process. Among the Metazoa light produces important chemical changes in the retina of the eye, and especially in the skin, facilitating the production of pigment. That important chemical changes take place in the illuminated retina follows from the experiment of placing the electrodes at opposite surfaces of the frog's retina. The galvanometer shows in the darkened eye a slight "current of rest" flowing from the front face to the deeper-lying part, containing the cones. If now the retina be suddenly illuminated by blue, green, yellow, red, or white light, a current, the result of chemical action, appears flowing in the opposite direction; this continues for

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\* Other, less important, thermal effects of light on plants are found in the formation of chlorophyll and in the quickening of transpiration, which seem chiefly due to the red and ultra-red rays.

some time, slowly diminishing, however, in intensity. Certain chemical changes in the living retina may, indeed, be studied optically. These especially concern the visual purple. This is a substance lying in the outer ends of the rods of the retina, which, under the action of light, becomes bleached, but regains its color in the dark (HELMHOLTZ, Handb., pp. 265-273). These facts serve to indicate that light may influence metabolism even in organisms destitute of chlorophyll.

2. **Vital Limits of Light Action on Protoplasm.**—We have seen above (p. 167) that the rate of assimilation diminishes in chlorophyllaceous plants with a diminution in the intensity of the light. At last a point is reached where the intensity is so low that no further assimilation can occur, and after the consumption of the stored-up food-stuffs, starvation and death must eventually ensue. For non-chlorophyllaceous organisms, however, no such lower limit exists. Many, as parasites or cave dwellers, live in complete darkness, even through many generations. A lower vital limit to the action of light exists only in the case of chlorophyllaceous plants.

With the *upper vital limit*, it is, however, quite different. This is found in the most diverse groups. Its occurrence in *bacteria* being of especial hygienic importance, these organisms have been made the object of exhaustive studies. MONTEGAZZA (see NICKLÈS, '65) was perhaps the first to discover that strong light kills bacteria, but DOWNES and BLUNT ('78 and '79) were the first to study the matter thoroughly. Since their time, numerous experiments have been made upon bacteria, as well as the higher fungi. For literature, see FRANKLAND and WARD ('92), and WARD ('93, p. 309). Even the earliest observers found that, while cultures of bacteria reared in the dark rapidly flourished, they not merely did not thrive when subjected to sunlight, but actually became sterilized. That the sterilization was complete was shown by the fact that when the culture was placed again in the dark, no bacteria developed in it. This result is most striking when certain bacteria, say of the species *Bacillus anthracis*, are mixed with gelatine or agar-agar, poured uniformly over a glass plate. If the glass plate is then covered by a black paper stencil containing some character, *e.g.* the letter *E*, and exposed to a November sun-

light for 6 hours, and if then the whole plate is placed in a dark incubator at 20° C. for 48 hours, the bacteria will be found to have developed in all parts of the plate except in the *E*-shaped area sterilized by the light (Fig. 47). Compare the earlier results of BUCHNER ('92). That in these cases it is the light and not a high temperature which induces the sterilization in the illuminated region is shown by the fact that BUCHNER ('92) obtained even more striking results when parts of the culture plate were exposed under 50 cm. of water, which cuts off the heat.

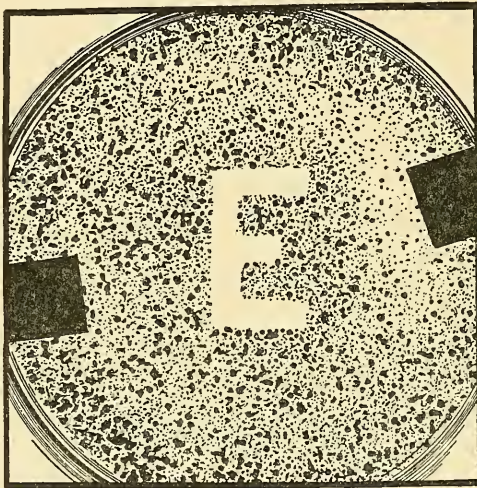


FIG. 47. — Appearance of a gelatine culture of *Bacillus anthracis*, exposed to the light over only the area *E*, and then incubated for 48 hours. In the area *E* no colonies have developed. (From WARD, '93.)

Not all rays have this bactericidal property. DOWNES and BLUNT ('78) found that only the blue rays were thus active, for behind red or yellow glass the bacteria readily developed. WARD ('94) threw a solar spectrum upon an agar film in which bacteria were developing in a dark chamber. He found that the bactericidal effect was greatest in the region of the blue-violet rays (about  $\lambda = 0.43 \mu$ ) and diminished towards the extreme violet and the yellow, where it had almost disappeared. These facts were ascertained by incubating the bacteria for 48 hours after insolation, when certain parts affected by the spectrum were found to remain clear (Fig. 48). When an electric

spectrum (obtained by the use of a quartz prism) was employed, a bactericidal effect was obtained (provided no glass intervened) in the ultra-violet. That the action of the light was not in these cases primarily upon the food-film was shown by the fact that a plate of sterile agar, exposed behind a stencil plate, and then laid flat on a film of dried unexposed spores, permitted the uniform growth of the spores, in the illumined as

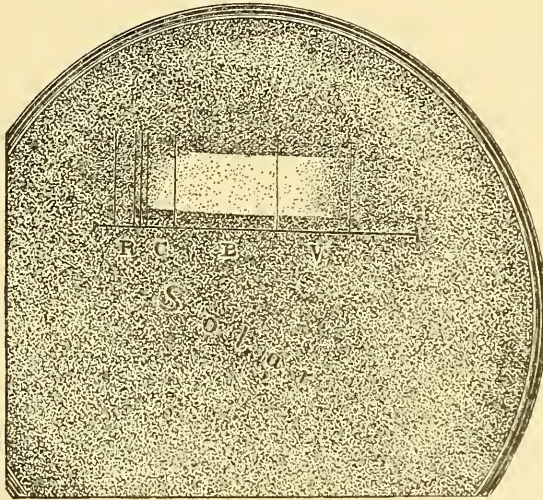


FIG. 48.—Plate of anthrax spores, exposed for 5 hours to the solar spectrum in August, and incubated for 48 hours. The horizontal line shows the length of the spectrum. The vertical lines are not FRAUNHOFER'S lines, but serve to show the limits of the principal regions of the spectrum. The clearest area is that where fewest spores have developed in the incubation—where, consequently, the bactericidal effect was greatest. (From WARD, '94.)

well as in the unillumined region. All these observations show that the bactericidal action of light is due to the action of the chemical rays on the protoplasm.

Another fact of importance, first discovered by DOWNES and BLUNT, is that light has no effect upon bacteria when they are in a vacuum. This abundantly confirmed observation indicates that death only secondarily results from light. The primary cause of death is an oxidation process,—a process rendered possible by the mediation of light. As we have seen (p. 162), many organic compounds undergo oxidation in the highly refracted light rays. Probably there are in bacteria such com-

pounds, the rapid oxidation of which is incompatible with life. In any case it is clear that the bactericidal effect of light is a chemical one.

Concerning the range of organisms which are thus affected, it must be said that chiefly pathogenic species, such as the bacteria of anthrax, of typhus fever, and of cholera have been experimented with and have shown themselves most susceptible. Other bacteria are, however, likewise affected. Among the other fungi, WETTSTEIN ('85) found that the conidia of *Rhodomycēs Kochii*, a human intestinal parasite, did not develop in the light. KLEIN ('85) found\* the same thing to be true for the conidia of *Botrytis cinerea*, and showed that the blue-violet rays were the most effective ones. ELVING ('90, p. 105) gained similar results with *Aspergillus*, although several days or weeks of insolation did not *kill* the fully ripe spores. WARD ('93) determined that insolated spores, cultivated on agar or gelatine plates, of *Oidium lactis* (5 cases), *Saccharomyces pyriformis* (4 cases), and "a 'Stysanus' conidial form" found as a saprophyte on the screw-pine, *Pandanus*, (2 cases) became injured. These are all hyaline and colorless except *Stysanus*, which is nearly so. Certain *colored* spores which WARD experimented with gave negative results, and WARD concluded that this is because the blue end of the spectrum is cut off before reaching the deeper protoplasm. However this may be, we actually find that in many, but not all, fungi the metabolic processes of the spores are disturbed and even death is provoked by intense light.

Why the spores should be especially susceptible to the action of light is an important inquiry. WARD believes the answer to be that the spores contain oily substances, which are especially liable to oxidation in light, as we have already seen.

Finally, we have to consider the experiments which demonstrate that a strong sunlight may be injurious even to green plants. This result follows clearly from the work of PRINGSHEIM ('81). When strong sunlight is focussed for a short time upon cells of *Spirogyra*, *Nitella*, *Mesocarpus*, or *Tradescantia* stamen hairs in atmospheric air (5 to 15 minutes), they are killed. No result occurs, however, when the same light falls upon green cells in which the atmosphere has been replaced by



hydrogen (Fig. 49). That it is here also the oxygen (and not the carbon dioxide) of the air which is the destructive agent is shown by subjecting the plants to air freed of carbon dioxide, when they are killed by light as before.

The most important results following from the conclusions of this sub-section are: a minimum vital limit of light action exists only in the case of those organisms (chlorophyllaceous plants) which depend upon light for assimilation; a maximum limit is found among the most diverse organisms, those with chlorophyll and those without. The rays which have the more rapid vibrations are the more active. They produce chemical changes to which death is primarily due.

3. **Effect of Light upon the Movement of Protoplasm.**—Under this head we shall consider only those protoplasmic movements which may not be grouped under Locomotion, and shall discuss three classes of cases:

(a) effect of low intensity of light upon movement; (b) effect of high intensity of light upon movement; and (c) effect of change of intensity on contraction.

a. *Effect of Low Intensity of Light on Movement—Dark-rigor.*—We have already seen that chlorophyllaceous plants must eventually die if kept in the dark. Some time before death occurs the plants go into a condition of immobility, which may be called dark-rigor, since return of light brings a return of movements. Dark-rigor is very marked in the sensitive plant.

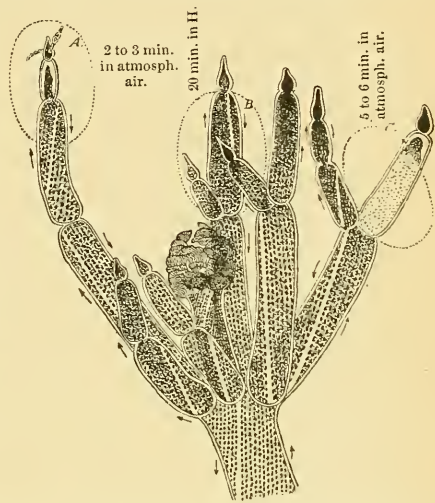


FIG. 49. — Piece of a sprout of *Nitella mneronata* which was subjected in a gas chamber to a green light in three successive experiments A, B, C. In experiment A the insolation lasted 2 to 3 minutes, the gas chamber being filled with atmospheric air. In experiment B the insolation lasted 20 minutes in the presence of hydrogen. In experiment C the insolation occurred again in the presence of atmospheric air and lasted 5 to 6 minutes. (From PRINGSHEIM, '81.)

If this plant is kept for several days in darkness, the usual response to touch does not occur. From some observations of BERT ('70, p. 338), it appears that it is the absence of the blue-violet and orange-red rays which brings about this dark-rigor; for it occurs nearly as rapidly in green light as in the dark. In these cases the absence of movement in the dark might seem to be the result of diminished assimilation.

But dark-rigor occurs under conditions which destroy the general validity of this conclusion; - for example, in the reddish-purple bacteria \* whose reactions have been studied chiefly by ENGELMANN ('83 and '88). It appears that in these organisms light is essential to movement; for, after having been kept over night in the dark, they are found in the morning at first motionless; only later, after 5 to 10 minutes of illumination, do they awaken to activity. If now, after keeping for a time in the light, the organisms are brought again into the dark, their movements gradually diminish until, in a few hours, they have ceased. We have seen above (p. 51) that oxygen is necessary to movement, and we know that many plants excrete oxygen in the light. We might expect that the quiescence of these organisms in the dark is a consequence of their failure to produce the oxygen necessary to locomotion, and indeed they do produce in the light a slight quantity of oxygen, by virtue of their chromophyll (bacterio-purpurin, LANKESTER). But that it is not merely oxygen which induces movement is shown by the fact that when an abundant oxygen supply is artificially furnished, no movement occurs in the dark. Thus light, in the presence of oxygen, is essential to movement; it seems to be necessary to the irritable condition upon which locomotion depends. This irritable state of the protoplasm conditioned upon a certain intensity of light ENGELMANN calls *phototonus*. †

The analysis of this matter has been carried further. It has been found that a perceptible time (latent period) elapses

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\* This term includes bacteria known as *Bacterium photometricum*, *Bacterium roseopersicinum*, *rubescens*, etc., *Monas okeni*, *Spirillum violaceum*, and by other names.

† The term was applied to this phenomenon by ENGELMANN on account of its resemblance to that already described for the higher plants, and to which SACHS had previously given this name.

between the illumination of the organism and the occurrence of movement. Also, the ultra-red rays produce most rapid locomotion, next the orange-yellow, and weakest the violet-blue and violet-red. Spectrum analysis shows that the most active rays are the ones absorbed by the chromophyll (Fig. 50).

This phenomenon of phototonus is not confined to the purple bacteria. Thus, FALINTZIN ('67, pp. 29-31) has shown that the movements of the closely related *Oscillaria* are diminished in the dark. SOROKIN ('78) found that protoplasmic streaming in the plasmodium of *Dictydium* ceases at night, being awakened to movement by the light. Finally, VER-

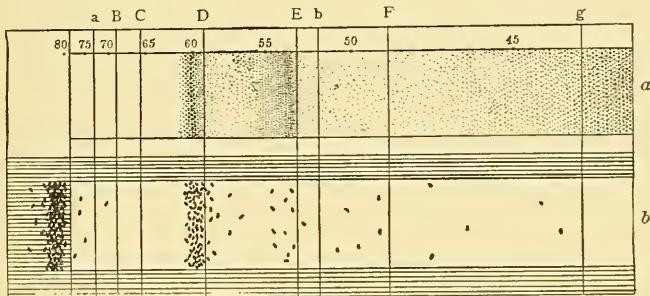


FIG. 50. — *a*. Spectrum of the chromophyll of bacterio-purpurin, showing absorption bands at  $\lambda = 0.59 \mu$  and  $\lambda = 0.53 \mu$ . An (invisible) absorption band has been determined by means of the bolometer at  $\lambda = 0.85 \mu$ . *b*. The bacteria are seen aggregated chiefly in the regions of the absorption bands. The accumulation of bacteria in these regions of absorbed energy seems due to the fact that the moving bacteria cannot pass from a region of high energy to one of low without a violent stimulus which impels them back again. (From ENGELMANN, '83a.)

WORN ('89, Nachschrift, and '95, p. 393) finds that, in the dark, the ciliate *Pleuronema chrysalis* rests quietly in the water, only occasionally making its peculiar spring. But when diffuse daylight is focussed upon it, for instance by the mirror of a microscope, it springs rapidly by the movement of its long cilia; and this movement is often repeated, so long as light continues to fall. The movement is produced by blue or violet rays; red rays have little or no effect. A latent period of from 1 to 3 seconds elapses before the response occurs.

These cases serve to show that light, in the presence of other suitable conditions, is, both for some chlorophyllaceous organisms and plant tissues and for some organisms destitute of

chlorophyll, nearly or quite essential to movement. Phototonus is a convenient name for the condition induced by light.

*b. Effect of High Intensity of Light on Movement—Light-rigor.*—We have just seen that in some organisms the most vigorous movements occur at an optimum intensity of light, which produces phototonus. At a lower intensity there is no movement. It appears, furthermore, that there is for many organisms a maximum intensity of light, below that which produces death (ultramaximum), which causes a cessation of movement that may be called *light-rigor*. This condition is distinguished from that of death by the fact that diminished light brings return of activity. ENGELMANN ('82, p. 109) observed this condition in his *Bacterium photometricum*, and remarks that it is common to all bacteria. Similar light-rigor has been observed in green plants also. PRINGSHEIM ('81, p. 516) found that when, in the presence of oxygen, strong sunlight was let fall upon *Nitella*, the movements ceased after  $1\frac{1}{2}$  minutes. If the insolation was now interrupted, normal movements were resumed.

Summing up the effects of varied intensities of light, it appears that for many organisms there is an optimum, which produces a condition of phototonus, in which the organism moves and responds regularly to stimuli. As the light intensity falls below, or rises above this optimum, the activity of movement diminishes, ceasing at certain points in the conditions of dark-rigor and light-rigor. Beyond each of these points, again, is the point of death.

*c. Contraction produced by Change in Intensity of Illumination.*—We here consider a number of cases not closely related except in this, that quick movements are produced after stimulation by change in the intensity of the light. The cases are found both among Protista and Metazoa.

Among the sulphur-bacteria ENGELMANN ('88, p. 665; 88\*) has noticed that a sudden diminution in the intensity of the light, produced by shading the mirror of the microscope, is followed by a spring backwards, often to the distance of 10 to 20 times the organism's length. This reaction ENGELMANN has called "Schreckbewegung." When the light is suddenly increased, a forward movement takes place, but this is

less marked. Among the Myxomycetes, ENGELMANN ('79) has found that the amœboid *Pelonyxa*, when suddenly subjected to a strong light, contracts into a spherical mass. Sudden darkening or gradual illumination produces no such contraction. Among swarm-spores, STRASBURGER ('78, pp. 575, 576) has noticed that a sudden diminution of the light puts the quiet *Hæmatococcus* spores again in motion, and makes the *Botrydium* spores start as though disturbed. Such violent movements of the protoplasm indicate that a very considerable chemical change has taken place in it.

Passing, next, to the Metazoa, we find that certain smooth muscle fibres are made to contract by the direct action of light; thus, STEINACH ('92) has offered most convincing evidence that the contraction of the iris, in the lower vertebrates at least, may occur as a direct reaction to illumination, even when the eyeball is cut out, and the iris, indeed, separated from connection with the ciliary part of the eye.

Some of the higher animals react strikingly like ENGELMANN'S bacteria. Thus, LOEB ('93, p. 103) found that *Serpula uncinata* retracts into its tube when the hand is passed between it and the light; but sudden increase of illumination has no effect. NAGEL ('96, p. 76) finds the same thing in *Spirographis*, and ANDREWS ('91, pp. 285, 296) has observed the same phenomenon in the eyeless *Hydroides dianthus*. In these cases the branchiæ seem to be the sensitive organs. Adult barnacles show a similar sensitiveness to light; for POUCHET ('72, p. 111) found that momentary cutting off of the light, as by the shadow of the hand, caused arrest, for several seconds, of the rhythmic movements of protrusion of the appendages from the shell. In this case, the sensitive region has not been located. Some lamellibranchs (NAGEL, '96, p. 50) react similarly to increased light. These are examples of a phenomenon which we shall meet with again in considering growth. They serve to show that there is a wide-spread irritability of protoplasm to changes in intensity of light.

Let us now review the conclusions of this section. Light — especially the thermic rays — is essential to the decomposition of carbon dioxide by chlorophyllaceous plants. The only effective rays are those absorbed by the chlorophyll. The rate

of assimilation is increased by increased intensity of light. The chemical rays act to increase metabolic changes, and the output of carbon dioxide. As these rays become more intense, the metabolic changes go on with abnormal rapidity, until, finally, death ensues; thus, intense light is fatal to many, perhaps to all, organisms. Absence of light, however, is injurious only as preventing assimilation in chlorophyllaceous organisms; but these supply the food for other organisms, so that continued darkness in any environment must likewise be eventually fatal to all life. All organisms, before succumbing to darkness or to light, enter into a condition of rigor, from which they may return to activity if favorable conditions are restored. Sudden change of intensity often produces violent protoplasmic changes, awakening quiescent organisms to activity, or causing, in the higher organisms, violent contractions.

All of these effects of light, whether produced by the thermic or chemic rays, probably give rise to great chemical changes by which disturbances of metabolism, and eventually death, may be produced. Not all organisms find light immediately necessary to their existence; but very powerful light, long continued, proves fatal to most protoplasm.

#### § 4. CONTROL OF THE DIRECTION OF LOCOMOTION BY LIGHT — PHOTOTAXIS AND PHOTOPATHY \*

In this section we shall (1) distinguish between false and true phototaxis; (2) consider the observed cases of phototaxis among Protista, the parts of higher organisms, and the Metazoa as entire organisms; and (3) discuss the general laws of phototaxis and photopathy.

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\* In this section we shall deal with two sets of phenomena which very likely are different, but which, in our ignorance, we cannot always distinguish. The first includes that active migration of organisms whose direction is determined by that of the rays of light. This is phototaxis. The second includes the wandering of organisms into a more or less intensely illuminated region, the direction of locomotion being determined by a difference in intensity of illumination of the two poles of the organism. This is photopathy. According as the migration is towards or from the source of light, we can distinguish positive (+) and negative (-) phototaxis. According as the migration is towards or from the more intensely illuminated area, we can distinguish positive (+) and negative

1. **False and True Phototaxis.** — It must certainly be a very old observation that when small organisms are placed in a vessel in front of a window, they are soon found arranged with reference to the window; some lying on the nearer side, some on the further side, and others swimming indifferently back and forth through the vessel. The conclusion is near at hand that this arrangement of the organisms is determined by the light. This conclusion is, however, not necessarily correct. Thus, SACHS ('76) showed that, under certain conditions, wholly passive substances — oil drops, in a mixture of water and alcohol — might exhibit a similar aggregation towards the window or away from it. These conditions are that the vessel should be cooler next the window. Then, on the cooler side, there will be a descending current; on the warmer side, an ascending current; on the surface, a current towards the win-



FIG. 51. — Vertical section through a dish showing distribution in water of passively suspended bodies, as a result of difference of temperature at the two sides of the vessel. *A*, warmer side; *B*, cooler side. Arrows show the direction of movement of currents in the water. The objects lighter than water are grouped at *b*; those heavier than water, at *a*.

dow; and on the bottom, a current from the window. If the passive bodies are such as float, they will thus be carried towards the window, and will exhibit a false phototaxis (in the positive sense); if, on the contrary, they tend to sink, they will be carried from the window, and show false negative phototaxis. Now this appearance, due to passive transportation by currents, may likewise, under the given conditions, be exhibited by *organisms* — but the phenomenon is not due to light (Fig. 51).

There is at least one other kind of pseudophototaxis. This

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(-) photopathy; and correspondingly we can in this second case speak of the organisms themselves as photophil or photophob. In this nomenclature I follow GRABER. STRASBURGER used *photometry* for what I here call *photopathy*, but OLTMANN'S ('92, p. 206) has employed photometry to indicate the capacity of organisms to perceive different degrees of intensity of light. So, perhaps, the terminology here employed may lead to the least confusion.

may occur when there are in the vessel chlorophyllaceous organisms producing oxygen in the sunlight. The oxygen, more abundantly formed on the sunny side of the vessel, becomes, then, a means of attraction to other (chemotactic) organisms, whose position seems thus to be determined directly by relative brightness.

A good example of this kind of pseudophototaxis is described by ENGELMANN ('81<sup>a</sup>). He found that the Schizomycetes in a certain drop of water, partially illuminated, were aggregated toward the illuminated side. Examination revealed the presence of a chlorophyllaceous schizomycete — *Bacterium chlorinum* — in the drop, and the apparent phototactic appearances were easily accounted for as follows: Under the influence of light the *Bacterium chlorinum* secreted oxygen, and this acted chemotactically to attract the bacteria, which thus moved, at the same time, towards the illuminated area. That it was the oxygen produced in the sunlight rather than the light itself which attracted, was evinced by the fact that, when the supply of oxygen is abundant in all parts of the drop, or if the *Bacterium chlorinum* is removed, no aggregation takes place at the bright point.

**2. Distribution of Phototaxis and Photopathy.** — *a. Protista.* — We now come to the consideration of the cases of true phototaxis and photopathy, and shall first discuss the distribution of the phenomenon in the different groups of Protista. Of the Protista we may take up first the chlorophyllaceous forms.

*Flagellata and Swarm-Spores.* — In no other group does phototaxis show itself more clearly than in this. The earliest studies were made here, but despite the ease of gaining results they were mostly fragmentary and uncritical. A simple experiment of NÄGELI ('60) had, indeed, showed conclusively that swarm-spores are responsive to light. A glass tube three feet long and held vertically was filled with alga-water. When the upper end of the tube was enveloped by black paper the organisms moved to the lower end, and conversely. A difficulty was encountered, however, in the fact that when zoöspores were placed in a plate by a window, the organisms gathered at the edge next the window, which, since the edge of the plate threw a shadow there, was the darkest part of the surface. In consequence some authors had concluded that swarm-spores shun the light; whereas COHN asserted, all too briefly, that they move in the direction of the rays and toward the source of light. Finally, FAMINTZIN ('67) had discovered that swarm-



spores which moved towards a light of a certain intensity would move from light of a certain greater intensity. That was the condition of knowledge on this subject when STRASBURGER'S ('78) epoch-making paper appeared.

STRASBURGER worked with swarm-spores of various species of algæ, and with the flagellate *Chilomonas* and *Euglena*. He observed again the phenomenon that the sense (+ or -) of response depends upon the intensity of the light. He also showed that the rate of movement is quicker in stronger light on account of the fact that the path taken by the organism is straighter; and (p. 586) that phototaxis is the result of the organism putting its long axis in the axis of the infalling rays. STRASBURGER found also that, in general, the smaller species of swarm-spores and the smaller individuals are more responsive than the larger ones.

Later studies have extended our knowledge of the distribution of phototaxis in this group. Swarm-spores have been studied by STAHL ('78, '80); *Euglena*, by ENGELMANN ('82<sup>a</sup>, p. 396); and *Volvox*, by CIENKOWSKI ('56), VERWORN ('89, p. 45), and OLTMANN ('92). Especially interesting is the fact that colorless swarm-spores, like those of *Chytridium*, which are parasitic upon chlorophyllaceous forms, respond like the green organisms. (STRASBURGER, '78, p. 508.)

*Desmids*, especially *Closterium*, have been experimented with by STAHL ('78 and '79), KLEBS ('85), and ADERHOLD ('88). All are markedly phototactic in moderate, diffuse daylight. This phototaxis is the more striking since the method of locomotion of these forms is peculiar. The crescentic *Closterium moniliferum*, for example, stands inclined and glides along, one extremity touching the substratum, the free extremity in advance. The gliding seems to result from the secretion of a stream of mucus along the substratum. Now STAHL believed that the angle of inclination of the *Closterium* is dependent upon the direction of the infalling rays of light, being parallel thereto. This relation has been denied by KLEBS, but ADERHOLD, by varying the direction of the infalling rays, has shown that the azimuthal position is determined by light. Under certain conditions *Closterium moniliferum* moves by a sort of head-over-heels motion, since the free end bends down to the

substratum and becomes attached, and the former attached end becomes free. STAHL explains this on the ground that the ends of *Closterium* periodically exchange their tendency to point towards the light. The appearances just described are found in diffuse daylight. In stronger light the azimuthal position is  $90^\circ$  from the infalling light. If direct sunlight falls upon desmids, they move from the light (negative phototaxis).

*Diatoms* have been studied by STAHL ('80) and VERWORN ('89, p. 47). Locomotion is effected in these organisms as in desmids by the secretion of mucous threads. The movement towards diffuse daylight (*Navicula*, *Stauroneis*) takes place slowly, but it often affects nearly all the individuals. The long axis does not seem to be clearly oriented in the direction of the infalling rays, which may be partly accounted for by the normal zigzag method of locomotion. Under strong sunlight diatoms appear negatively phototactic. Occasionally a culture will be found whose individuals are separated into two groups — one next the positive side, the other next the negative side of the vessel.

*Oscillaria*. — VERWORN ('89, p. 50) has made experiments on the reaction of these organisms, whose method of locomotion is probably similar to that of desmids. They are markedly positively phototactic from half darkness to direct sunlight; only in intense sunlight do they fail to accumulate at the positive end of the vessel. The aggregation at the positive pole takes place by the threads assuming a direction parallel to the rays of light and creeping forward thus, side by side. VERWORN states that after all have attained the + edge, rotation of the slide or vessel through  $180^\circ$  does not cause a prompt transfer of all individuals towards the light side — at least during the time of his observation only a few had crawled towards the light in its new position. According to WINOGRADSKY ('87), *Beggiatoa* is generally negatively phototactic.

*Myxomycetes*. — In its amœboid form and when subjected to strong sunlight *Æthelium septicum* retreats into the substratum, but while in the dark it comes to the surface (HOFMEISTER, '67, p. 625; STRASBURGER, '78, p. 620). Also, when the plasmodium is partially illuminated, the protoplasm tends to flow from

the illuminated region. (BARANETZKI, '76, p. 328, and STAHL, '84, p. 167.)

BARANETZKI proceeded as follows: a glass plate was placed in a saucer so that its surface was 2 or 3 mm. below the rim. The plate was covered by filter paper which extended over the rim and here dipped into water, by which means it was kept moist. Over the saucer was laid an opaque cover, blackened below and provided with a narrow slit. The plasmodium was placed on the filter paper and diffuse daylight was thrown upon the slit by means of a plane mirror. In less than half an hour the illuminated threads of the plasmodium had become very thin, owing to the retreat of the protoplasm from under the slit to the darker region (Fig. 52).

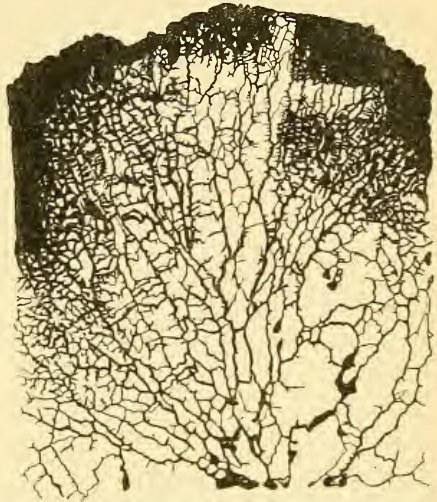


FIG. 52.—Plasmodium of *Æthalium septicum*, after having been kept in the dark for some time and then illuminated, for half an hour, over a cross-shaped area, only. The illuminated area is on the upper part of the figure. The protoplasm has retracted from it, leaving a partially clear region in the form of a cross. (FROM BARANETZKI, '75.)

*Rhizopoda*.—Although, as we have seen, *Pelomyxa* is irritated by a sudden illumination, a phototactic or photopathic response has not hitherto been certainly observed in this group. VERWORN ('89, pp. 40, 41), indeed, experimented, but with negative results, upon *Amœba limax*, *Amœba princeps*, *Actinosphærium*, and *Actinophrys*. Only in *Polystomella crispa* did he notice a slow wandering towards the source of light; but he was uncertain whether this was due to light.

VERWORN'S method was not well devised, however, for bringing out phototactic response. The Protista were placed on the slide, and, after cutting out heat rays by means of a plate of ice, were subjected to the light or to the ENGELMANN microspectrum, and illuminated at different intensities either over the whole body or over only a part. All disturbing influences, he says, were as far as possible eliminated: gravity, by an exact horizontal position of the microscope on a table with three screw-feet; the action of the edge of the drop, by using a very broad drop; and, finally, the laterally

*impinging rays of light, by means of a black cardboard box placed over the slide.* Thus it is clear that all of his light fell upon the organism in perpendicular rays from below. This method of experimentation would clearly not show whether *Amœba* is phototactic or not.

I have experimented with *Amœba proteus*, using methods resembling VERWORN'S, and likewise dissimilar ones, and have reached new results. In the first place, I have proceeded somewhat after the fashion of VERWORN to determine whether the amœba in a field illuminated from below, and separated by a sharp line into a light and dark half, showed any change of movement in passing from dark to light or from light to dark; also, whether an amœba moving in a uniformly illuminated field changed its direction when half of its body was darkened. Nearly all such experiments were negative. No effect resulting from the change from light to dark or the reverse could be detected. Thus far my results agreed with VERWORN'S.

In a second set of experiments, I proceeded differently. Usually one amœba was isolated by means of a capillary tube. It was then introduced, with a drop of clear water, between two slips of glass, each about 25 by 50 mm., which were kept 2 mm. apart, and at the same time cemented together, by glass strips of equal thickness placed near the ends. By this means a broad field for movement with uniformity of conditions of contact was ensured. The whole space between the two glass plates being now filled with clear water, the entire apparatus was submerged in a vessel which contained water about 2 cm. deep, and which was slightly smaller than the stage of the microscope. Finally, the entire stage, but not the substage optical apparatus, was kept in the dark by means of a cone made of several thicknesses of dense black cloth fastened by a slip-noose to the objective, and folded below the stage so as completely to exclude all extraneous lateral light. Light from the mirror was cut off by an interposed card. Through a slit in the cloth on the side next the window,—a west window,—a beam of direct sunlight, or of reflected light from the morning sky, was admitted to the amœba. The plates of glass being as nearly as possible horizontal and occasionally rotated, the directive action of gravity was eliminated. Since, so far as could be seen with

the microscope, no local sources of food or oxygen occurred in the water between the plates of glass, chemotactic influences were uniformly distributed. From the conditions of the experiment already described, a difference in temperature or of illumination at the two poles of the amœba is scarcely conceivable. The rays of radiant energy were the only directing agent. Under these conditions the amœba nearly uniformly showed itself negatively phototactic to light of an intensity varying from strong diffuse light to direct sunlight. The absence of uniformity is to be ascribed to the accidental presence of some disturbing agent. The movements made by the amœba were represented graphically by making at intervals a camera drawing of its outline. Two such graphic representations are reproduced in Figs. 53 and 54. It must be said that it is difficult to get so extended a series of changes in light as is shown in Fig. 54, for the phenomenon of acclimatization comes in and the responses become irregular. But, despite such irregularities, my studies lead me unhesitatingly to conclude that Amœba, although not at all photopathic, is strongly phototactic. This result is important, for, since Amœba is responsive to light, it may very well be that such responsiveness is a general property of protoplasm.

*Ciliata.* — A double action of light must be here taken into account. ENGELMANN ('82<sup>a</sup>, pp. 391–395) states that those Ciliata which contain chlorophyll (algæ) — *e.g.* Paramecium bursaria, Stentor viridis, Bursaria — move towards the light, but only when the oxygen tension in the water is low. Also when the water drop is illuminated by a microspectrum, instead of white light the organisms aggregate towards the red end. Here are the rays by which most oxygen is produced from the chlorophyll, since assimilation takes place fastest here. When the organisms are placed in excessively oxidized water they move from the light. The conclusions to which ENGELMANN arrived from these and other facts were that these species have a very delicate sensitiveness to variations in oxygen tension, and that it is through this sensitiveness that light influences movement. Accordingly, it would seem that the apparent phototaxis is truly a case of chemotaxis; but this conclusion requires better evidence.

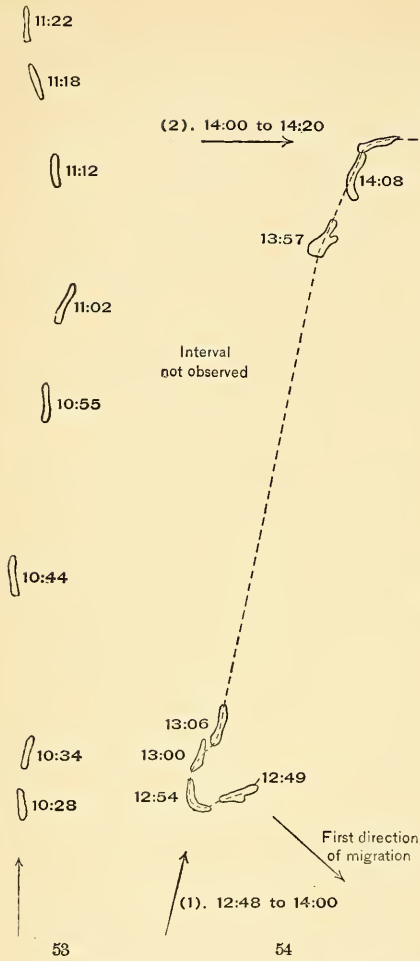


FIG. 53. — Camera drawing, showing the successive positions assumed by an amoeba subjected to light falling upon it from one side. The arrow lies in a horizontal projection of the sun's rays. The amoeba retreats from the source of light. The numbers to the right of the outlines of the amoeba give the observed times between 10:28 and 11:22 A.M. Magnified 16 diameters.

FIG. 54. — Camera drawing, showing the successive positions assumed by an amoeba retreating from the light. The position of the infalling ray was successively changed from (1) to (2), (3), and (4). The arrow labelled "First direction of migration" shows the direction of locomotion of the amoeba before the light fell upon it at the beginning of the experiment. The numbers indicate hours and minutes. During the interval from 13:06 (=1:06) P.M. to 13:57, the amoeba was not under direct observation, since I was called away. Magnified 16 diameters.

Cases that can be explained only on the ground of the immediate effect of light upon the direction of movement are certainly rare. ENTZ ('88), indeed, has intimated that *Opalina* flees from light, but VERWORN ('89, pp. 53-57) was not able to confirm him in this point. VERWORN'S method was here, as in the case of *Amoeba*, not satisfactory. Instead of having the light fall from one side only upon the drop containing the *Opalinas*, he let the light pass vertically from below through a small hole, and could observe no tendency to avoid the illu-

minated spot. The light in this case clearly did not act from one side, and the test of phototaxis can therefore hardly be said to have been critically made. Likewise, even with unilateral illumination, VERWORN was unable to gain a phototactic response with *Stentor roeselii*, *St. cœruleus*, *Carchesium polypinum*, and *Uroleptus musculus*. On the other hand, we have often noticed here in Cambridge that our *Stentor cœruleus* is (rather indefinitely) negatively phototactic to diffuse daylight. Thus, an individual swimming free in a bit of glass tubing pointing horizontally towards the window only very slowly wanders away from the light. In conclusion, then, we must admit that Ciliata are not markedly phototactic, but more refined methods must be used before we can say of any of them that they exhibit no trace of this response.

Let us summarize briefly the results obtained from Protista. Phototaxis is most marked among actively motile, chlorophyllaceous forms. Many colorless forms are, however, also phototactic — *Beggiatoa*, *Amœba*, plasmodia of *Myxomycetes*, and swarm-spores of *Chytridium*. The phenomenon is thus widespread, if it is not universal.

*b. Cells and Cell-organs.*—Under this head will be considered, (*a*) the rearrangement of chlorophyll corpuscles, (*β*) the rearrangement of pigment in animal cells, and (*γ*) the migration of pigment cells in the metazoan body.

*a.* That the *chlorophyll bodies* of the higher plants change their position in the cell according to the intensity of the light to which they are subjected has been made known chiefly through the labors of FAMINTZIN ('67), BORODIN ('69), FRANK ('72), STAHL ('80), and MOORE ('87). If one fastens a strip of black paper upon a leaf on which the sun's rays are falling, one will find, upon removing the paper after a time, that the darkened part is dark green whilst the brightly illuminated part is considerably lighter, so that an image of the form of the dark paper is produced upon the leaf. This image is, however, only temporary. A few hours after the removal of the paper the leaf is of a uniform green again. Sections through a leaf thus affected show that in the dark green (shaded) part of the leaf the chlorophyll lies on those walls of the cells which are perpendicular to the incoming rays.

whilst in the light green (illuminated) part of the leaf the chlorophyll lies upon the walls parallel to the rays. When

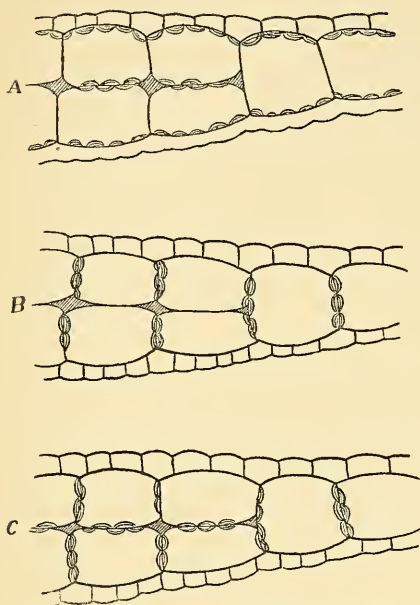


FIG. 55.— Cross-section through the leaf of *Lemna trisulca*. A. Position of the chlorophyll grains in diffuse daylight—epistrophe. B. Position of the chlorophyll grains in intense light—positive apostrophe. C. Position of the chlorophyll grains in darkness—negative apostrophe. (After STAHL.)

the grains are upon the superficial face of the cells they are said to be in *epistrophe*; when they have turned away from this face they are in *apostrophe*. This apostrophic position is found under two opposite conditions of illumination: under intense light, as we have just seen (positive apostrophe, MOORE), and upon prolonged standing in the dark (negative apostrophe). (Fig. 55.)

It appears, then, that epistrophe occurs only within certain limits of light intensity. The intensities included between these limits constitute what MOORE calls the epistrophic interval. The epistrophic interval varies in position and in extent in different species.\* It has been found that in the case of plants which normally live in the bright sun the epistrophic

\* The limits were determined by MOORE, in a roughly quantitative way, by means of his *photrum*, constructed as follows. A room with a single window illuminated by the sun was chosen and 12 feet spaced off from the window back into the darkness. The intensity of the light diminished of course as one retreated from the window. Plants of various species were allowed to stand, simultaneously, at varying distances from the window, and the distance back at which epistrophe began to appear, and, finally, at which negative apostrophe came in, were noted. Then a diagram  $\frac{1}{18}$  the actual scale was made (Fig. 56), showing the position of the points of beginning and ending of epistrophe (so-called positive and negative critical points).



interval is a region of relatively high intensity (Fig. 56, 6); in aquatic plants the epistrophic interval occurs in a region of low intensity (Fig. 56, 1 and 2); and in shade-loving aërophytes in an intermediate position (Fig. 56, 4 and 5). One may say that every species is attuned to a certain intensity and range of light, in which epistrophe occurs, just as in swarm-spores there is a certain intensity and range of light in which positive phototaxis occurs, and that attunement depends upon the conditions to which the organism has adjusted itself through living in them.

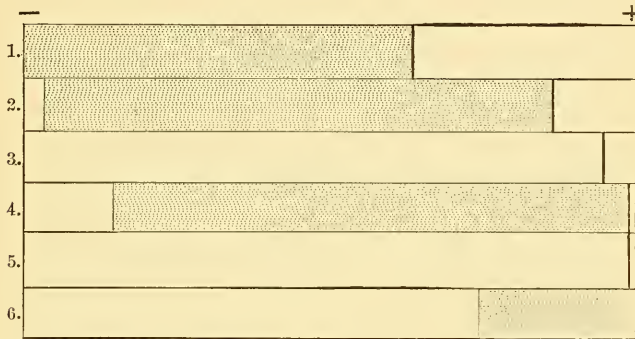


FIG. 56. — A diagram of MOORE'S photrum, showing for six spaces the epistrophic interval (shaded region). 1. *Anacharis (Elodea) canadensis*, "water-weed." 2. *Lemna trisulca*. 3. *Saxifraga granulata* (position of positive critical point). 4. *Oxalis acetosella* (position of negative critical point only approximate). 5. *Pteris critica* (positive critical point). 6. *Pyrethrum sinense* (garden Chrysanthemum). + indicates the brighter end of the photrum.

Concerning the question of the mechanism of the movement of the chlorophyll grains, there is much difference of opinion. It is urged on the one hand that the chlorophyll grains move actively to attain their new positions, and, on the other, that they are passively carried by the cell currents. Of these two views analogical reasons are perhaps the strongest for preferring the second.

As to the question in how far these movements can be regarded as adaptive, we may say that STAHL believed that they regulate the relation between intensity of sunlight and assimilating area, so that the quantity of assimilation shall not become too great. HIERONYMUS ('92, p. 466) considers them to be for the purpose of screening the nucleus. MOORE (p. 222),

however, chiefly from a consideration of vegetative apostrophe, has been led to the conclusion "that the movements of chlorophyll have no relation whatever to benefit or injury experienced by the grains, nor necessarily to the well-being of the protoplasm."

*β. The Rearrangement of Pigment in Animal Cells in Response to Light.*— One of the striking cases of this effect of light is seen in the pigment cells of the skin of the chameleon, as described by KELLER ('95, pp. 144, 162). He has found that the dark color of the (illuminated) skin is due to the rich

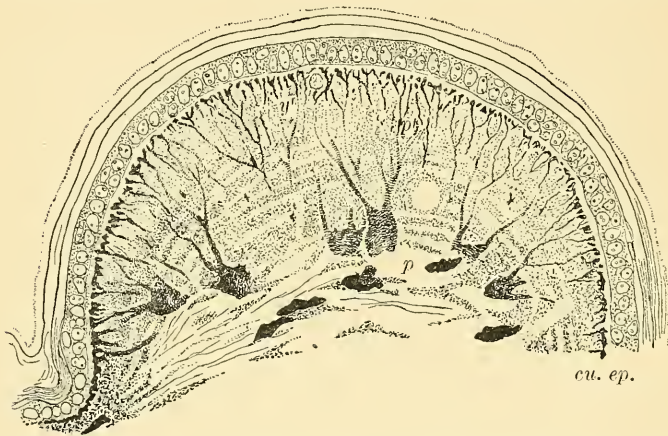


FIG. 57.— Vertical section through a black dermal papilla of *Chamaeleo vulgaris*. *ep*, epidermis; *cu*, cutis; *p*, black pigment cells; *p'*, processes of the cells containing pigment; *y'*, yellow pigment cells. (After KELLER, '95.)

branching at the base of the epidermis of black pigment cells lying deep in the cutis (Fig. 57). In the dark, the pigment granules stream out of the branches into the cell body, but the branches themselves are undisturbed (Fig. 58). So long as the black pigment has this central position, the skin appears whitish. The light, on the contrary, causes the pigment, which is probably carried passively in the plasma, to move centrifugally. Whether the direct response to light of the pigment cells of the frog, as described by STEINACH ('91), is of the same nature, or due to contractions of the pigment cells, remains to be determined.

Again, in the retina of the compound eyes of Arthropoda,

we find this capacity for rearrangement of pigment granules, as EXNER ('89 and '91, p. 104), STEFANOWSKA ('90), SZCZAWINSKA ('91), PARKER ('95), and others have shown. In the higher Crustacea, for example, the pigment granules of the pigment cells surrounding the rhabdome (or "spindle") are, in the dark, below the level of the spindle. Upon illumination, however, these granules migrate (or are carried) upwards, and partly envelop the rhabdomes. I believe it has not been determined what rays are involved in producing this result. This response to light is considered to be an advantageous one, since

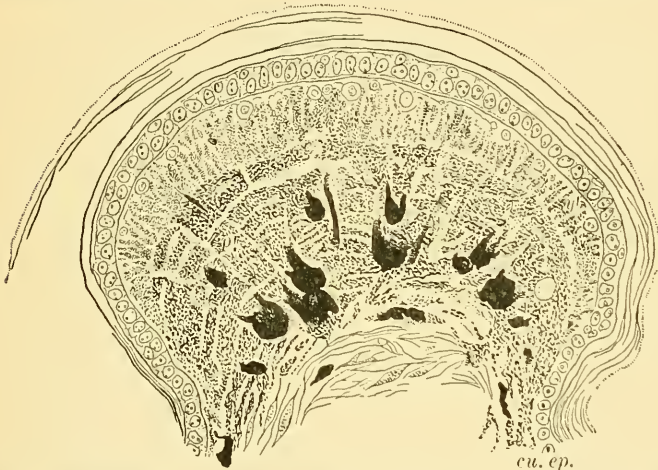


FIG. 58. — Vertical section of a whitish-yellow dermal papilla; lettering as in Fig. 57. *p'*, processes of black pigment cells containing no pigment. (After KELLER, '95.)

the pigment thus cuts off side rays from the perceptive organ — the rhabdome.

It is interesting that we should find cells containing two so diverse kinds of pigment as chlorophyll and the retinal pigment responding to light in so similar a fashion. In most of the cases, if not all, this response is an adaptive one.

*γ. The Migration of Pigment Cells in the Metazoan Body.* — It has been shown, apparently first by ENGELMANN ('85), that the pigment cells of the retina vary their movements with the light. Thus, when a strong light is thrown upon the retina of the frog, the pigment cells send out pseudopodium-like processes between the rods and cones, whereas in the dark the

pigment lies behind all these elements. Also, among the Crustacea, the protoplasm of the outer pigment cells, surrounding the cones of the compound eye, migrates centripetally in a strong light, to return again to its peripheral position in darkness. These movements may also be considered adaptive. They are, in addition, movements which are discharged only by light.

*c. Metazoa.* — We may treat of the control by light of the movements of the higher animals somewhat more summarily than we have the preceding classes. The facts will be arranged by groups in systematic order.

Among *radial* animals, Hydra has perhaps been for the longest time an object for photopathic study. TREMBLEY (1744, p. 66) had noticed that *Hydra viridis*, and even mutilated pieces of it, came to the light side of the vessel. When the light was admitted only through a chevron-shaped slit, the Hydroids were later found aggregated opposite the slit in the form of a chevron. That it was not the warmth of the sunlight that attracted was shown by turning the slit towards the cooler air, whereupon the same response occurred. The observations of TREMBLEY showed that the Hydroids did not move in as straight a line as possible towards the light (they must, of course, follow a firm substratum), but gradually wandered towards it. The response of Hydra must therefore be considered as photopathy. More extensive studies were made upon Hydra by WILSON ('91), who found that *Hydra fusca* is likewise responsive to light, and, indeed, photophil with reference to diffuse daylight, and photophob to direct sunlight. And it can be shown that it is an advantage to Hydra to be photophil, since many of the Entomostraca upon which it feeds are phototactic.

Besides Hydra, I know of only one case of response by Cœlenterata to light. The larvæ of the *sponge* *Reniera* are said (MARSHALL, '82, p. 225) to flee from the light, — probably negative phototaxis.

Among *Echinodermata*, *Asteracanthion rubens* (GRABER, '85, p. 155) appears to be photophil, and *Asterina gibbosa* (DRIESCH, '90, p. 155) to be photophob.

Although, as we have seen, some radial animals may respond to light, the phenomenon is more wide-spread in the bilateral groups, — flatworms, annelids, crustaceans, insects, molluscs,

and Vertebrates. The results of experiments may here be given in tabular form. Unless otherwise stated, the light is supposed to be diffuse daylight, and the response to be phototactic.

TABLE XVIII

ORGANISM.	SENSE OF RESPONSE.	AUTHORITY	REMARKS.
Fresh-water planaria. . . . .	—	LOEB, '90, p. 95	
Polynoë sp. . . . .	—	DRIESCH, '90, p. 155	
Polygordius, larva . . . . .	—	LOEB, '93, p. 90	see p. 200
Earthworm . . . . .	—	{ DARWIN, '81, p. 21 GRABER, '83, p. 210 HESSE, '96	{ photophob
Leech. . . . .	—	LOEB, '90, p. 96	
Daphnia . . . . .	{ + +	{ TREMBLEY, 1744, p. 96 BERT, '78, p. 989 LUBBOCK, '82, '83 DAVENPORT and CANNON	{ photophil (?) phototactic
Many marine copepoda . . . . .	—(or +)	LOEB, '93, p. 96	see p. 200
Balanus, larva . . . . .	—(or +)	GROOM and LOEB, '90, p. 160	see p. 200
Limulus, larva . . . . .	—	LOEB, '93, p. 83	
Idotea tricuspidata . . . . .	+	GRABER, '85, p. 141	photophil
Diastylis (Cuma) rathkii . . . . .	+	LOEB, '90, p. 91	mud-inhabiting
Carcinus menas. . . . .	—	DRIESCH, '90, p. 156	photophob
Homarus americanus, larva . . . . .	+	HERRICK, '96, p. 189	
Plant lice . . . . .	+	LOEB, '90, p. 55	
Blatta germanica (blinded) . . . . .	—	GRABER, '83, p. 235	photophob
Musca dom. (?), larva . . . . .	—	LOEB, '90, p. 69	
Musca, adult . . . . .	+	LOEB, '90, p. 81	
Musca vomitoria, larva . . . . .	—	DAVIDSON, '85, p. 160	
Musca cæsar, larva . . . . .	—	POUCHET, '72, p. 113	
Eristalis tenax, larva . . . . .	—	POUCHET, '72, p. 129	
Lepidoptera, adult . . . . .	+	{ SEITZ, '90, p. 337 LOEB, '90, p. 46	see p. 197
Lepidoptera, larva . . . . .	+	{ LOEB, '90, p. 51 POULTON, '87, p. 315	
Ants, after gaining wings . . . . .	+	LOEB, '90, p. 63	
Melolontha vulgaris. (May beetle) . . . . .	—	LOEB, '90, p. 86	
Tenebrio molitor, larva . . . . .	—	LOEB, '90, p. 84	
Dentalium . . . . .	—	LACAZE-DUTHIERS, '57, p. 25	photophob
Rissoa octona . . . . .	+	GRABER, '85, p. 144	photophil
Littorina rudis . . . . .	—	DRIESCH, '90, p. 155	photophob
Gasterosteus spinachia . . . . .	—	GRABER, '85, p. 148	
Triton . . . . .	—	GRABER, '83, p. 221	photophob
Frog . . . . .	—	LOEB, '90, p. 90	

A study of this table reveals the fact that, in general, organisms which live in shady places or in the dark are negatively phototactic or photopathic, while those living in the light are positively phototactic or photopathic. Thus most fresh-water planarians and leeches are inhabitants of shady pools. Polynoë is generally found in dark retreats, the earthworm and fly larvæ are lovers of the dark, and shell-molluscs are for the most part enclosed in cases impervious to light. On the other hand, *Daphnia* is found largely in open pools, larvæ of Lepidoptera and many adult insects live in the sun. Into this general rule there are some cases which do not so obviously fall. But we have little data concerning the habits of the races employed and the absolute intensity of light used, so that these cases may perhaps be only apparent exceptions. One clear exception is that of the mud-inhabiting *Diastylus*, which is + phototactic.

3. **The General Laws of Phototaxis and Photopathy.** — Under this head we shall consider: (*a*) the sense of the response; (*b*) the effective rays; (*c*) phototaxis *vs.* photopathy; (*d*) the mechanics of response to light.

*a. The Sense of the Response.* — In considering, now, more generally, the effect of daylight upon the direction of locomotion of organisms, we must recognize that the sense of response (whether + or -) depends upon internal conditions and external conditions — upon the quality of the protoplasm and the nature of the environment. Let us consider, first, the dependence upon *internal conditions*.

We find that, under similar external conditions, different organisms respond differently. For example, many *Oscillariæ* (p. 184) are positively phototactic even in direct sunlight; whilst even moderately strong light will repel many diatoms. In fact, we find that a positively phototactic or photopathic organism is such only in the presence of a certain intensity of light. When the intensity is diminished below a certain point, no response will occur. When, on the other hand, the intensity is increased above a certain point, the organism moves away from the source of light. There is a certain range of intensity in which alone the positive responses occur. The position and the extent of this positively phototactic range vary for the

different species, — they are closely correlated with the conditions of light in which the organism has been reared. As a result of these conditions, we may say that each organism is attuned to its peculiar range and intensity of light.

Upon the ground of this difference in attunement may be explained the remarkable difference in behavior of butterflies and moths to light. It is well known that butterflies fly towards even the strongest sunlight, whilst moths are secluded during the daytime, but at night fly towards the candle-light. LOEB ('90, p. 46) has performed some experiments with these insects, which I will cite in detail.

EXPERIMENT 1. — (*a*) Sphinx, Bombyx, and other moths were kept in a large glass cage in a room illuminated only by daylight. As darkness came on, the moths began to fly towards that side of the cage which was next the window. Again (*b*), pupæ of nocturnal moths, left in a room, emerged during the night, and were always found in the morning at the closed window of the room. Finally (*c*), a nocturnal moth, made to fly in the daytime, directed its way to the window. Thus, nocturnal moths are positively phototactic to diffuse daylight as well as candle-light.

EXPERIMENT 2. — Hawk-moths were brought into a room with the single window at one end, and a petroleum lamp at the opposite end. It was found that, as twilight came on, the moth flew to the window, or to the light, according to the relative intensity of the one or the other at the point where the moth was liberated. Thus, there is no preference for artificial light.

The conclusion at which LOEB arrived was that these moths undergo a diurnal variation in responsiveness to light, which corresponds to the change from day to night. But the fact that, in experiment 1 *c*, nocturnal moths flew, in the daytime, towards the diffusely lighted window, throws a doubt upon this interpretation. All the facts are equally well explained upon the following ground: Butterflies are attuned to a high intensity of light, moths to a low intensity; so that bright sunlight, which calls forth the one, causes the other to retreat. On the other hand, a light like that of a candle, so weak as not to stimulate a butterfly, produces a marked response in the moth. We shall consider, in a moment, the cause of these differences in light attunement.

We have seen that one internal condition modifying response is the racial quality of attunement. A second is that of *period of life*. Thus, LOEB ('90, p. 56) has found that, at the intensities

employed, the wingless plant lice were hardly responsive to light. The winged form was markedly positively phototactic. So, likewise, in the case of ants (LOEB, '90, p. 63), during the period of the marriage flight the males and females (but not the workers) are strongly positively phototactic, but after that period they show themselves neutral. The case of the house-fly, *Musca*, is interesting, since the larva and adult are phototactic in opposite senses (see table). In most of these cases, the difference in responsiveness is associated with a difference in habit.

The sense of response depends, also, as we have seen, upon *external conditions*. In this regard, the immediately preceding conditions of light, the temperature, the concentration, and the supply of oxygen have important effects. We shall consider, in order, the action of these conditions.

*Light* can modify the response to light; thus, GROOM and LOEB ('90) have shown that the nauplii of *Balanus*, as well as other pelagic animals, come to the surface of the sea during the night, but descend before the strong sunlight. This does not indicate merely a low light-attunement of the race; for nauplii exposed to sunlight in the early afternoon are all positively phototactic, and only gradually, as the day progresses, move from the sunny window, until, finally, even as dusk approaches, all are found on the side away from the window. Nor have we here to do with a diurnal change in the sense of the response. For if a culture is kept in the dark, it is found to be at first positively phototactic at whatever time of day it is exposed; only later acquiring the negative phototaxis. In the same way, when the young *Balanus* larvæ leave the interior of the shell of the parent, they are at first positively phototactic; but after being in the light for from  $\frac{1}{4}$  to 2 hours, they become negatively phototactic. The more intense the light, the quicker its effect.

Another observation upon the nauplii is representative of a new class of light effects. When nauplii which have become negatively phototactic through exposure are covered for a few minutes, and then suddenly again exposed to light, they move momentarily towards the light, and then begin their negative movement again. Somewhat similar are the results obtained



by STRASBURGER (78, p. 574) on *Ulothrix* spores, which are positively phototactic in a weak light. While responding to such a light, they do not, however, turn at once when a light of repelling intensity is thrown upon them. So, also (p. 600), when *Hæmatococcus* is responding to indigo light, the interposition of red glass does not at once cause it to turn from its path. In all these cases, the immediately preceding condition of light continues to exert an action which modifies the response.

Closely allied are the results obtained by VERWORN (89, p. 50) upon the diatom, *Navicula brevis*, which is attuned to only the faintest light. When, however, a culture had been reared by a window for two weeks, the attunement to light had been so raised that now a slight degree of positive phototaxis took place in diffuse light. We have in these facts examples of a phenomenon which we have observed in the action of other agents. It is one expression of the acclimatization of organisms to the peculiar conditions of their environment. We have just seen that every organism has its optimum intensity of light for metabolism and response, and that this optimum is very varied; but, throughout, one law holds. Organisms which are accustomed to live in strong light have a high optimum intensity; and those accustomed to live in a weak light have a low optimum intensity. This relation is, indeed, so close as to raise the suspicion that the normal intensity of the light has determined the optimum. And this suspicion is confirmed by the experimental evidence just cited. Now, since the position of the optimum is usually advantageous, we may conclude that light can so modify protoplasm as to adapt it for the conditions in which it is living.

We now pass to the consideration of the effect of *temperature* upon response. This effect was noticed by STRASBURGER (78, p. 605) in the swarm-spores of *Hæmatococcus*, *Ulothrix*, etc., which, at a temperature of 16° C. to 18° C., gather at the side of the drop next to the window. If, now, they are subjected to a temperature of 40° C., the intensity of the light being constant, they migrate to the opposite side. On the other hand, at a temperature of 35°, the + aggregation is more complete than at 16° to 18°. Control experiments with emulsions satisfied STRASBURGER that this change is not due to

currents in the water, but is a truly vital phenomenon. That it is such is indicated also by the following curious behavior. If swarm-spores which normally aggregate at  $18^{\circ}$  towards the positive side of the drop, are suddenly brought to  $18^{\circ}$  from  $30^{\circ}$ , they appear, for a moment, negative. Conversely, if swarm-spores which normally aggregate at  $30^{\circ}$  towards the negative side of the drop are suddenly brought from  $8^{\circ}$  to  $30^{\circ}$ , they appear, for a moment, positive. Thus, the immediately preceding culture-temperature affects the sense of the response.

The results obtained by STRASBURGER have been in part confirmed by other authors in other species.

GROOM and LOEB ('90, pp. 166, 172) state that in the case of the nauplii of *Balanus* — “at a higher temperature, for instance  $25^{\circ}$  C., the phenomena [of phototaxis] are run through more sharply and quickly than at a temperature of about  $15^{\circ}$ ”; and again, “we often succeeded in suddenly changing the sense of the heliotropism of the larva by a sudden change, of only a few degrees, in the temperature of the water.” This statement is unfortunately so vague as to say little more than this, that temperature influences the response. MASSART ('91, p. 164) remarks, incidentally, that the flagellate *Chromulina* is + phototactic at  $20^{\circ}$  C., but - phototactic at  $5^{\circ}$  C. LOEB ('93, pp. 90, 96) obtained a result with *Polygordius* larvæ and Copepoda which seems, at first sight, the opposite of STRASBURGER'S. *Polygordius* larvæ, negatively phototactic at  $16^{\circ}$ , were gradually cooled to  $6^{\circ}$ , at which temperature they began to move rapidly towards the + side of the vessel. As the temperature gradually rose they became - phototactic again. Individuals which were (abnormally) + phototactic at  $17^{\circ}$  to  $24^{\circ}$ , when raised gradually to  $29^{\circ}$  became - phototactic. Sudden diminution of temperature within the limits at which response occurs did not change the sense of their response. Thus, negative individuals brought suddenly from  $23^{\circ}$  to  $13^{\circ}$  remained negative. Exactly parallel results concerning the relation of temperature and response were obtained by LOEB from Copepoda.

All results may be harmonized in the expression: Diminution of temperature below the normal causes reversal of the normal response; elevation of the temperature to near the maximum accelerates the normal response. The point of light attunement varies with the temperature.\*

Not only light and heat, but also the *concentration of the medium* affects light attunement. We are indebted to LOEB

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\* It follows from these experiments that it is necessary in any phototactic investigation to regard not only the intensity of the light, but also the temperature.

('93, pp. 94, 96) for information on this subject. Negatively phototactic *Polygordius* larvæ were placed in sea water to which 1% to 1.3% NaCl had been added. They now appeared positively phototactic. Positively phototactic individuals, on the other hand, placed in sea water diluted with 40% to 60% fresh water became negatively phototactic. Similar results were obtained with Copepoda. Thus increased concentration rendered + phototactic (raised light attunement), while diminished concentration rendered - phototactic (lowered light attunement). Increased concentration works, therefore, upon *Polygordius* and Copepoda, according to LOEB, like diminished temperature.

Finally, the *chemical condition* of the medium has an important effect on photopathy, as can be judged from certain observations of ENGELMANN ('82<sup>a</sup>, p. 391). Various chlorophyllaceous Ciliata, *e.g.* *Stentor viridis* and *Paramecium bursaria*, are photopathic only when the oxygen supply in the medium is below the normal. In such media they are strongly photophil. This case is clearly not a case of phototaxis, as we have just seen (p. 187). The response is advantageous since it brings these organisms into the sunlight, where chlorophyll can produce oxygen.

To recapitulate: the sense of response in phototaxis is modified by previous subjection to light, by temperature, and by concentration. These agents modify the attunement of the organism. Any quantitative experiments upon phototaxis must therefore take all of them into account. Certain chlorophyllaceous organisms exhibit + photopathy, but only in an insufficiently oxygenated medium.

From the foregoing considerations we conclude that for every phototactic organism there are three ranges of intensity to be distinguished: the positively phototactic range in which the organism moves towards the light; below this, the indifferent range extending to darkness; above it, the negatively phototactic range extending up nearly or quite to the point of light-rigor. The limits of these ranges vary with both external and internal conditions.

*b. The Effective Rays.* — We have hitherto considered chiefly the action of white light, merely referring casually to the

action of the different rays of which it is composed. We must now answer the question: What different effects do the different rays have?

The effect of the different rays in phototaxis is very clearly seen in the various groups of Protista and among the Flagellata and the swarm-spores, there is entire uniformity of response according to the testimony of COHN ('65, p. 36), STRASBURGER ('78, pp. 593-599), ENGELMANN ('82<sup>a</sup>, p. 398, in *Euglena*), and VERWORN ('89<sup>a</sup>, p. 49, in *Navicula*). Here the more actinic rays with shorter and more rapidly vibrating wave, act exactly like white light, whilst the rays from the opposite end of the spectrum have no more effect than darkness. More precise determinations were made by STRASBURGER ('78, p. 597), who found that the swarm-spores of the alga *Botridium* responded to the blue and violet, but especially to the indigo, whilst the green and ultra-violet were alike without effect. And ENGELMANN, by means of his microspectral apparatus, was able to determine that *Euglena* responded chiefly to the rays  $\lambda = 0.47\mu$  to  $\lambda = 0.49\mu$ ; that is, rays very near FRAUENHOFER'S line *F*. The colorless *Myxomycetes* agree with the chlorophyllaceous forms, according to BARANETZKI ('76, p. 332), in responding to blue rays only.

Among the higher organisms, *Hydra*, according to WILSON, accumulates especially behind blue glass, to a small extent behind green glass, and is entirely indifferent both to the upper violet rays and those below the green. The photophil starfish *Astracanthion rubens*, even when deprived of its eyes, was found by GRABER to be "cyanophil"; even, though in slight degree, to a low intensity of light. Among Mollusca, GRABER found that the photophil *Rissoa* moved towards the blue even when the intensity of the blue light was less than that of the red, and DRIESCH asserts that the photophob *Littorina rudis* shuns only blue rays. Thus, without multiplying cases, the results of experiments may be summed up as follows: positively phototactic or positively photopathic organisms are such only in the presence of the blue rays.

There are some few observations which are in apparent discord with this conclusion. Whether the ultra-violet rays are ever active is a fairly debatable question. LUBBOCK ('82,

p. 127; '84, p. 137) showed that *Daphnia* and some ants are very sensitive to the violet rays, and GRABER ('83, p. 214) found that the photophobic earthworm withdraws from ultra-violet rays. This result is unusual, however, for most experimenters have agreed with this much of BERT'S ('78, p. 989) conclusions, that "the animals see . . . only those rays which we ourselves see," or, better, that the range of irritability of the protoplasm of our retina is as great as that of any other protoplasm.

Below the blue, some authors have believed the yellow rays, the brightest of the spectrum, to be prevalingly photopathic. Thus both BERT ('68, p. 381) and LUBBOCK ('83, p. 214) find that *Daphnia* accumulates especially in the yellow and green parts of the spectrum. Regarding these results I have only the comment that they need further confirmation.

*c. Phototaxis vs. Photopathy.*—We have hitherto assumed the existence of two dissimilar sorts of locomotor response to light—phototaxis and photopathy. Phototaxis we defined as migration in the direction of the light rays, and photopathy as migration towards a region of greater or less intensity of light. Are we justified in making this distinction?

The chief ground for this distinction is the existence of two sorts of phenomena which, not having been generally recognized as different, have led to extensive discussion. The best-established of these phenomena is phototaxis, which was proved to exist by certain crucial experiments of STRASBURGER on Protista, and of LOEB on Metazoa. Mr. W. B. CANNON and I have used STRASBURGER'S methods on *Daphnia*, and confirmed his results. Figure 59 gives a view of our apparatus, which was essentially the same as STRASBURGER'S. It consisted of a hollow prism *P*, containing a dark solution and placed over the trough *T*, with its organisms. STRASBURGER ('78, p. 585) put swarm-spores of *Botrydium* and *Bryopsis* into the trough, and reflected the light perpendicularly through the prism upon the trough. There was now a perfect gradation in intensity from the thick end to the thin edge of the prism. Yet the organisms showed no tendency to aggregate at the clearer end. The light was now permitted to enter the trough obliquely, the thicker end of the prism being next the source

of light, as in the figure. The spores now moved towards the source of light, *i.e.* in the direction of the infalling rays but constantly into a region of less intensity of light.

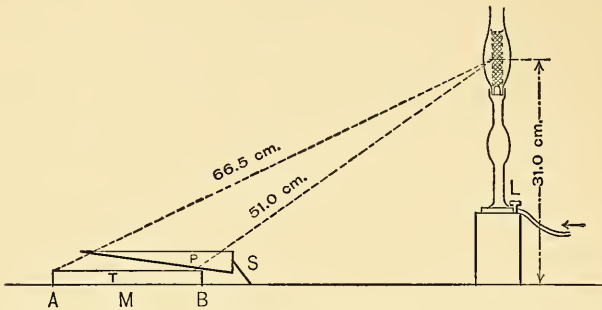


FIG. 59.—Diagram showing the position of apparatus and the direction of the rays in an experiment in phototaxis. *T*, trough of water containing organisms, *A* and *B* its two ends, *M* its middle. *P*, a prismatic box containing a solution of India ink. *S*, screen to cut off extraneous light. *L*, gas-lamp having a WELSBACH burner. Drawn to scale.

LOEB'S ('90, p. 32) results were obtained by the use of quite different methods. In one case he employed a chamber made of two test tubes placed with their mouths together. One of the tubes was darkened except for a clear streak at one end, *c*; and this darkened tube was pointed towards the light, so that the rays fell through its axis. Although the clear chamber was evidently the brighter, the *Porthesia* larvæ with which he experimented moved into the darkened chamber and thus towards the source of light (Fig. 60). Again, a clear test tube containing larvæ (Fig. 61) was placed so that its closed end *b* was directed towards the window *FF*. A bundle of sun's rays *SS* struck nearly perpendicularly the mouth of the tube *a*, when the larvæ were aggregated at the beginning. Nevertheless the larvæ, since their progress in the direction of the perpendicular rays was soon interrupted by the walls of the tube, moved towards the window, from the region of greater intensity of light in the direction of rays which passed more nearly in the axis of the tube. That this is not negative photopathy to strong light is indicated by the fact that the *Porthesia* larva is attuned to a high intensity of light. The evidence would thus seem satisfactory that the direction of migration of certain

organisms is determined by the direction of the light rays. There is, then, such a thing as phototaxis.

But is the direction of locomotion ever determined by a difference of intensity of light in adjacent regions, without reference to the direction of the light rays? Whole series of observations make this probable; for a migration to a definite part of the trough has followed unequal illumination by rays perpendicular to the trough. Thus LUBBOCK found that

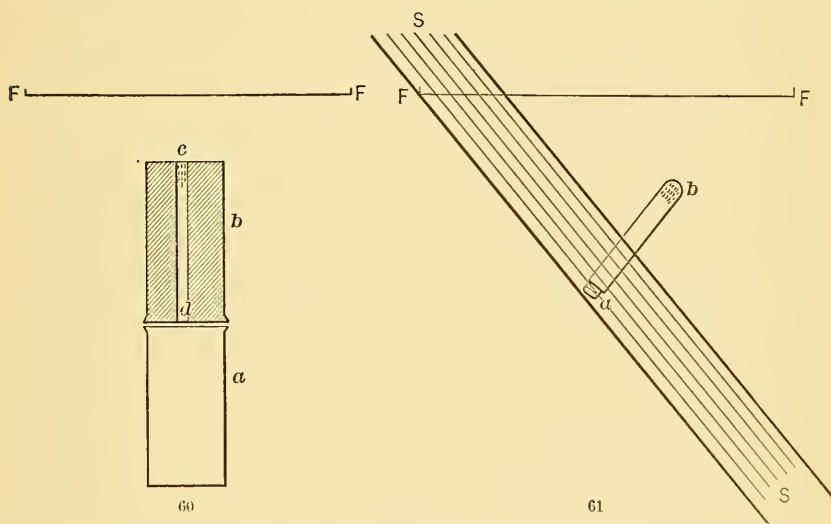


FIG. 60.—Two test tubes *a* and *b*, containing *Porthesia* larvæ *c*, which move towards the window *FF'*, although in doing so they pass from a brighter to a darker region. (LOEB, '90.)

FIG. 61.—Diagram to show how *Porthesia* larvæ move in the test tube *ab* towards the window *FF'*, although in doing so they leave the part of the tube more brightly illuminated by the sun's rays *SS'*. (LOEB, '90.)

*Daphnias*, placed in a trough nearly perpendicular to the rays dispersed by a prism, moved towards the brighter part of the spectrum. GRABER employed screens of diverse translucency and color, which were placed adjacent to one another, and found that the organisms tended to aggregate opposite the one or the other. OLTMANN'S ('92, p. 195) has offered certain new experiments pointing in the same direction. These experiments were made upon *Volvox minor* and *Volvox globator*, which were placed in a trough between which and the source

of light a vertical screen was interposed. This screen formed one side of a wooden box and consisted of two glass plates making an angle of  $2^\circ$  with each other, the interspace being filled with a solution of India ink in gelatine. When the sunlight was let through this screen, the individuals in the trough behind it sorted themselves into two groups; the parthenogenetic individuals, which collected opposite the clearer part of the screen, and the female individuals, with fertilized eggs, which collected behind the darker part of the screen, each suiting itself to the intensity of light to which it was attuned. When the intensity of the light was changed, the organisms also changed their positions.

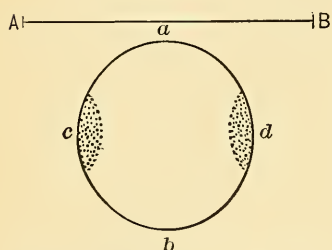


FIG. 62. — Diagram showing position taken by *Planaria torva* in a shallow cylindrical glass vessel *a, b, c, d*, placed opposite a window *AB*. (LOEB, '93.)

Finally, LOEB ('93, pp. 100-103) has found that fresh-water planarians (*Planaria torva*) gradually accumulate in the darker parts of the vessel, since the light constantly stimulates them to movement, and in their wanderings they gain the dark places by accident and there are at rest. So it comes about that when these *Planaria* are in a shallow cylindrical vessel (Fig. 62, *a, b, c, d*) in front of a window *AB*, they accumulate neither at the

side towards the window nor that away from it, but at *c* and *d*, where the side walls of the vessel cut off much of the light. All these cases, then, lead to one conclusion, that organisms may move with reference to more or less intense light — that there is such a thing as photopathy.

Indeed, a phototactic and a photopathic response may be exhibited by the same organism. Thus in CANNON'S and my experiments *Daphnia* was found to be phototactic, although other observers have clearly shown it to be photopathic, a result which we have not been able to disprove. We conclude, then, that some organisms have this double response to light that they may move in the direction of its rays, and that they may keep in a certain intensity of light to which they are attuned.



*d. The Mechanics of Response to Light.*—Under this head I shall speak of the part of the protoplasmic body most sensitive to light, of the immediate effect of the light, and of the cause of this immediate effect.

There can be no question that radiant energy with rapidly vibrating waves produces upon all protoplasm a profound effect. The question arises, however, to what extent in organisms a special kind of protoplasm is differentiated for the reception of rays which result in a discharge of the locomotor response. Certainly such a differentiated protoplasm can hardly be considered necessary to the discharge of such a response, since there is no morphological evidence of its existence in the responsive amœba. However, even in the swarm-spores and Flagellata such a specialized protoplasm is clearly indicated. Thus ENGELMANN ('82<sup>a</sup>, p. 396) found that when a dark band fell across the body of a swimming Euglena, no reaction occurred so long as the hinder chlorophyllaceous part alone was shaded. When, however, the clear area at the base of the flagellum was shaded, a marked reaction occurred. Here, near the pigment spot, if not at it, is the specialized light-perceiving protoplasm.

Similarly specialized protoplasm occurs extensively in the higher groups in the form of retinas; but there is much evidence that in many eyeless Metazoa the whole surface contains such light-perceiving substances. This is well known to be the case in the earthworm (cf. HESSE, '96). According to DUBOIS, ('89, p. 233), the siphon of the boring mussel *Pholas dactylus* contracts at the least variation of light intensity upon the skin. Similarly, other Lamellibranchia (*Ostea*, *Unio*, *Venus*) close their valves (NAGEL, '96, p. 58). Blinded *Helix* are said by WILLEM ('91, p. 248) and NAGEL ('96, p. 19) to be similarly sensitive. The lamellibranch *Psammobia*, the blind *Proteus anguinus*, and the blinded *Triton cristatus* are irritated by rays of light, especially the blue rays, falling upon the skin (NAGEL, '96, p. 22; GRABER, '83, p. 233; DUBOIS, '90, p. 358). Thus, in many Metazoa, protoplasm sensitive to light is of widespread occurrence, outside of the retina.\*

The *immediate visible effect of light* upon the organism differs

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\* For an extended list of cases of such dermatoptic reaction, see NAGEL, '96.

according to the form of the organism. In elongate, antero-posteriorly differentiated animals the first visible phototactic response is the orientation of the organism's axis in the direction of the impinging ray, and with the head end directed towards the source of light, or from that source, according as the organism is positively or negatively phototactic. The orientation is the more precise and the retention of the position the more sure the nearer the light approaches, the optimum (attractive) or maximum (repellent) intensity, as the case may be. If two rays of different intensities making an angle with each other fall upon the organism, it apparently moves in the direction of the intenser ray, if free to do so.\*

In Amoeba, without differentiated axes, the effect of the ray of light is to determine the position of the centrifugal streaming by which a pseudopod is thrown out away from the light; and the streaming continues in this single direction so long as conditions do not change. Thus the locomotion is in a straight line, lying in the ray of light.

Light not merely determines the direction of the axis but the position of the head end. As we have seen (p. 196) this determination of the position of the head depends upon the attunement of the organism, a quality which in turn varies with certain internal and external conditions. Acting upon a "highly attuned" protoplasmic mass, light will cause orientation in one sense; upon "lowly-attuned" protoplasm, an orientation in the opposite sense.

Whether light has any other effect than that of orientation of the body is a mooted question. STRASBURGER ('78, p. 577) and LOEB ('90, p. 109) recognize that migration from one point to another is more rapid in strong light than in weak, but believe this difference in rate of migration is wholly explicable upon the ground that the orientation is more precise in the stronger light, that there is less wandering from side to side. Some experiments made by Mr. CANNON and me upon *Daphnia* seem to confirm this view and at the same time afford quantitative data upon the degree of hastening. Thus in 18 trials *Daphnia*

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\* This statement is provisional only. It seems to follow from the experiments of LOEB made upon moths and described on page 197. The point is worthy of detailed comparative study.

required, to travel 18 cm. in full light, 15% longer time than the same individual required in light  $\frac{1}{4}$  as strong. Since the increased time was only 15% instead of 300%, as it should be were rate proportional to intensity, it seems probable — a conclusion confirmed by the direct observation of the organisms in the trough — that the slower rate in the weaker light is due to less precise orientation. How would the rate be influenced by two lights of different intensities acting from opposite directions? Upon this matter we have no experimental data.

Light, then, serves to orient the organism; but how? This again leads us to the general question of the cause of the tactic response, — a question which must be referred to a later chapter. Certain special considerations may, however, be introduced here. Let us first think of the way in which light acts on the negatively phototactic (and photopathic?) earthworm. Repre-

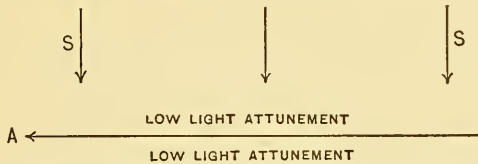


FIG. 63. — Diagram, representing sunlight (*SS*) falling upon an elongated, bilateral organism (represented by the arrow) whose head is at *A*. (Original.)

sent the worm by an arrow whose head indicates the head end (Fig. 63, *A*). Let solar rays *SS* fall upon it horizontally and perpendicularly to its axis. Then the impinging ray strikes it laterally, or, in other words, it is illuminated on one side and not on the other. Since, now, the protoplasm of both sides is attuned to an equal intensity of light, that which is the less illuminated is nearer its optimum intensity. Its protoplasm is in a phototonic condition. That which is strongly illuminated has lost its phototonic condition. Only the darkened muscles, then, are capable of normal contraction; the brightly illuminated ones are relaxed. Under these conditions the organism curves towards the darker side; and since its head region is the most sensitive, response begins there. Owing to a continuance of the causes, the organism will continue to turn from the light until both sides are equally illuminated; *i.e.* until it is in the light ray. Subsequent locomotion will carry the organism in a

straight line, since the muscles of the two sides now act similarly. Thus orientation of the organism is effected. The same explanation, which is modified from one of LOEB ('93, p. 86), will account, *mutatis mutandis*, for positive phototaxis.

Such an explanation can serve only for elongated organisms. The case of the amœba is quite different. Here we must think of the protoplasm as being modified by a light ray so as to flow centrifugally especially in that ray, perhaps through peculiar molecular disturbance wrought by the ray.

As for photopathic response, that is probably to be accounted for on the ground upon which ENGELMANN has explained the arrangement of *Bacterium photometricum* in the microspectrum; on the ground, namely, that increased brightness causes a movement forwards, that a diminution in brightness causes a movement backwards, or *vice versa*, thus resulting in the accumulation of the organisms in the darker or lighter parts of the field.

To summarize, then, light acts directly either through difference in intensity on the two sides of the organism, or by the course the rays take through the organism. Difference in intensity of light may also determine the position of organisms with reference to light by virtue of the irritation produced by rapid change of intensity.

#### SUMMARY OF THE CHAPTER

The study of the effect of light on protoplasm must be made quantitative as well as qualitative, and demands the use of apparatus for determining the quality and intensity of the light employed. The reactions produced by light upon protoplasm are undoubtedly of a chemical character, and, indeed, experiments with non-living organic compounds show that it has an important effect in synthesis, in analysis, in substitution, in the production of isomeric or polymeric conditions, and in fermentation. Since protoplasm consists of a large number of kinds of organic substances, we should expect light to produce far-reaching results, the more so as it can penetrate deep into the tissues of the organism.

The effect of light upon the general functions of organisms is

revealed in modifications of metabolism and of movement, and in the production of death. Upon metabolism we can distinguish an effect of the red rays, which are greatly absorbed by chlorophyll and are chiefly active in assimilation, and an effect of the blue rays, which seem to produce important chemical changes, increasing the production of carbon dioxide in plants, creating an electric current in the retina as it falls thereon, and bleaching visual purple. These chemical changes become more vigorous with increased intensity of light and may lead to death; while, at the opposite extreme, complete absence of light may prove fatal by withdrawing the necessary thermic and chemical energy. Again we find light sometimes necessary to movement in protoplasm, at other times by its absence or too great intensity inhibiting movement, or, again, by sudden change in intensity, creating abrupt changes in movement. Thus light undeniably has a great effect upon the processes of metabolism and movement.

Finally, in those complex processes involved in locomotion, light produces very widespread effects; for the direction (and, though only indirectly, perhaps, the rate) of locomotion is influenced in so important a way that when light is withdrawn the organism wanders aimlessly about. Of the various rays, those with wave length =  $40\mu$  to  $49\mu$  are the most active in controlling locomotion. Movement towards the light takes place at intensities of light varying greatly with the species and also with the conditions other than light in which the individual finds itself, — two factors upon which depends the degree of attunement. Light having an intensity above that to which the organism is attuned repels the organism. Two kinds of effects are produced by light: one by the direction of its ray — phototactic; the other by the difference in illumination of parts of the organism — photopathic.

We thus see that organisms respond to light, and that this response, exhibited in movements, is not of a widely different order from the disturbances produced in metabolism, which in turn are of the same order as the chemical changes produced by light in our laboratories upon non-living substances. In a word, response to light is the result of chemical changes in the protoplasm wrought by light.

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## CHAPTER VIII

### *ACTION OF HEAT UPON PROTOPLASM*

IN this chapter it is proposed to consider (I) briefly, the nature of heat and the general methods of its application; (II) the action of heat upon the general functions of organisms; (III) the temperature-limits of life; (IV) the acclimatization of organisms to extreme temperature, and (V) the determination of the direction of locomotion by heat — thermotaxis.

#### § 1. NATURE OF HEAT AND THE GENERAL METHODS OF ITS APPLICATION

Heat is believed to be due to the vibrations of the molecules of bodies. In any heated solid, fluid, or gaseous mass the molecules are in constant motion. When the temperature is increased, the motion is increased, and the impacts of the flying molecules become more frequent. If a vessel containing water is brought into contact with warmer air or warmer fluid, its molecules fly faster, its temperature is raised. As the motion of the molecules in the walls of the vessel increases, the increased motion is transmitted to the contained water, and finally to the objects in the water. Thus the motion of the molecules of an organism in the water is increased with the increase of the temperature of the water. Heat, as so-called radiant heat, is transmitted through space in straight lines, and follows all the laws of light, into which it passes when the rate of wave vibrations becomes rapid enough to affect the retina. The chief effects of radiant heat were considered in the last chapter.

Heat is an important element in all chemical processes. The state of cohesion — solid, liquid, gaseous — and the ease with

which molecular decomposition and synthesis occur, vary directly with it. This is an important consideration in our study of protoplasm, for most of its changes are chemical changes.

A word should be said concerning general methods of applying heat to protoplasm. In the case of the higher plants and

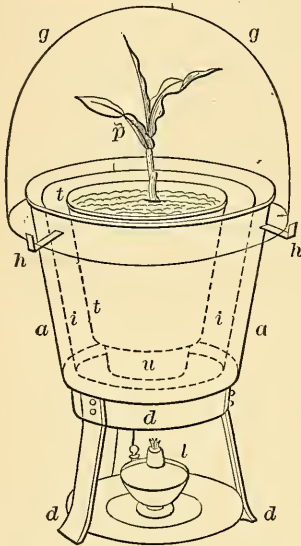


FIG. 64. — Apparatus for studying the effect of heat upon germination in phanerogams. *a*, the external; *i*, the internal vessel, between which is a water space; *t*, flower-pot filled with earth and containing a seedling of maize *p*; *h*, three supports for the glass bell *g*; *u*, support for the flower-pot; *d*, tripod iron stand carrying the spirit-lamp *l*. (From SACHS, '92.)

seedlings, the device of SACHS ('92, p. 117) may be employed. This consists of two metallic vessels, *a* and *i* (Fig. 64), of similar form, one placed inside of the other, the interspace being filled with water. Within the inner vessel is placed the pot (*t*) with the object of experimentation. The whole is covered over by a half globe of glass (*g*), extending down to below the level of the top of the pot. The water is heated by a lamp (*l*) below, by which means moisture and warmth are carried to the plant.

In the case of the lower organisms, brief experiments may be conducted in shallow aquaria for the horizontal microscope (Fig. 65), like those devised by CORI ('93). It is preferable to put inside of the outer vessel a smaller glass vessel, which shall contain the organisms and the thermometer marking the temperature of the water. For long-continued experiments where constant high temperature is required,

a warm oven, such as is used in bacteriological work, is essential.

The production of extremely low temperatures offers special difficulties. For temperatures to  $-40^{\circ}$  or so, various freezing mixtures can be employed. Of these chopped ice and common salt in equal parts give a temperature of  $-18^{\circ}$ ; calcium chloride and snow, in proportions of 3 to 2, give  $-33^{\circ}$ ; and calcium chloride and snow, in the proportion of 2 to 1, give  $-42^{\circ}$ ,

the initial temperature being always supposed to be  $0^{\circ}$ . Far lower temperatures than these have been obtained by physicists, notably PICTET, to whose work we shall refer again

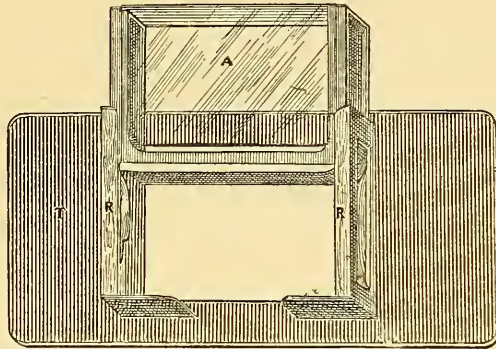


FIG. 65. — CORI'S stage aquarium. *A*, the aquarium proper; *T*, holder for the aquarium; *R*, *R*, slides, with springs. (From CORI, '93.)

(p. 240). The organisms to be acted upon may be kept in an apparatus (Fig. 66) like that employed by POUCHET ('66).

Throughout this chapter, as indeed throughout the whole book, thermometric readings are given in Centigrade scale,

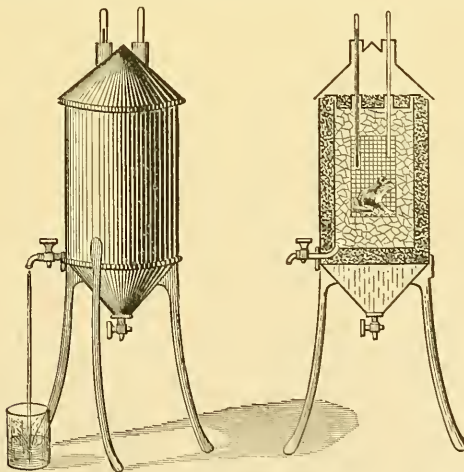


FIG. 66. — Cold chamber seen in external view and in section. The wall is composed of an inner cylinder containing the freezing mixture and the receptacle for the object of experimentation, and an outer cylinder separated from the inner by a packing of fragments of charcoal. The receptacle containing the organism is provided with a thermometer and an air tube. (From POUCHET, '66.)

unless otherwise stated. All readings not designated by the — sign are above the Centigrade 0 point. The point of absolute 0, to which we may have occasion to refer, is  $-273^{\circ}$  C.

## § 2. THE EFFECT OF HEAT UPON THE GENERAL FUNCTIONS OF ORGANISMS

Under this topic will be considered (1) the effect upon metabolism, and (2) the effect upon movement and irritability.

1. **Effect of Heat upon Metabolism.** — Within certain limits the relative increase of temperature leads to a relative increase in the activity of the various metabolic processes. This is well seen in those chemical changes which produce so-called *phosphorescence*. Many years ago MACAIRE ('21, p. 157) showed for fireflies, and ARTAUD ('25, p. 372) for the organisms of the sea, that light begins to appear shortly above  $20^{\circ}$ , reaches its maximum intensity at  $40^{\circ}$  in the fireflies and  $35^{\circ}$  in the water organisms, and entirely disappears at  $59^{\circ}$  to  $62^{\circ}$  in the first case, and  $43^{\circ}$  in the second. The temperature of these three points — lowest temperature of metabolic activity, temperature of greatest activity, and highest temperature permitting of activity — may be called, respectively, the minimum, optimum, and maximum temperatures for phosphorescence.

The effect of temperature on metabolism is seen in the *absorption of oxygen* by organisms. Thus VON WOLKOFF and MAYER ('74) found that more oxygen is absorbed by seedlings, as the temperature is increased, from  $0^{\circ}$  to about  $35^{\circ}$  C. This is shown in the following table. (From VINES, '86, p. 198.)

FIVE NASTURTIUM SEEDLINGS.		FOUR WHEAT SEEDLINGS.	
TOTAL AMOUNT OF O ABSORBED PER C.C.	TEMPERATURE C.	TOTAL AMOUNT OF O ABSORBED.	TEMPERATURE C.
0.60	$22.4^{\circ}$	0.10	$15.6^{\circ}$
0.77	$27.0^{\circ}$	0.038	$4.4^{\circ}$
0.76	$30.5^{\circ}$	0.067	$9.8^{\circ}$
0.77	$30.0^{\circ}$	0.088	$15.4^{\circ}$
1.04	$35.0^{\circ}$	0.022	$0.3^{\circ}$
0.91	$38.2^{\circ}$	0.010	$0.1^{\circ}$



Likewise MOISSAN ('79, p. 296) found that, in the dark, the amount of oxygen absorbed by a branch of certain plants varied with the temperature. Thus there was absorbed per hour by

Pinus pinaster (30 grammes) . . .	{ at 0° C.,	0.32 c.c.
	{ at 13°,	1.30 c.c.
	{ at 15°,	1.90 c.c.
Agave americana (70 grammes) . . .	{ at 11° C.,	0.54 c.c.
	{ at 40°,	5.56 c.c.

These experiments serve to show clearly that in plants more oxygen is absorbed as the temperature is raised to the optimum.

The same result is obtained from animals also. Thus TREVIRANUS ('31, p. 23) found that the honey bee *Apis mellifica* absorbed at 14° C. 1.35 Paris cubic inches, and at 27.5°, 2.77 cubic inches of oxygen. At the higher temperatures the bee was very active, so that the result seems here somewhat complicated by the increased muscular activity accompanying a higher temperature, which invokes a more rapid respiration. Nevertheless, the phenomena of increased oxygen absorption with higher temperature are fundamentally the same in plants and animals.

Turning now to the process of *excretion*, it appears that the amount of CO<sub>2</sub> evolved by seedlings varies with the temperature. On this point we have data by DEHERAIN and MOISSAN ('74, p. 327), RISCHAWI ('77), and others. DEHERAIN and MOISSAN experimented with leaves of tobacco kept in the dark. The same plant was used throughout the experiment, and it remained throughout in good condition. In the following table the temperatures are given in the first column, and, in the second, the number of grammes of CO<sub>2</sub> produced per 100 grammes of leaves:—

TEMP.	GMS. CO <sub>2</sub> .	TEMP.	GMS. CO <sub>2</sub> .	TEMP.	GMS. CO <sub>2</sub> .	TEMP.	GMS. CO <sub>2</sub> .
7	0.031	15	0.165	20	0.263	40	0.961
13	0.139	18	0.178	21	0.289	41	1.132
14	0.157	19	0.193	32	0.514	42	1.325

When plotted (with the temperatures as abscissæ) the relation between temperature and weight of CO<sub>2</sub> produced is expressed

by a line which is slightly steeper at the higher temperatures than at the lower. This change of steepness is, however, much less striking in the case of the etiolated wheat seedlings studied by RISCHAWI, where the following series was obtained:—

TEMPERATURE.	WEIGHT CO <sub>2</sub> IN MG.	TEMPERATURE.	WEIGHT CO <sub>2</sub> IN MG.
5°	3.30	25°	17.82
10°	5.28	30°	22.04
15°	9.90	35°	28.38
20°	12.54	40°	37.60

The evidence from excretion thus also confirms the conclusion that the metabolic processes are accelerated by raising the temperature to a certain limit.

The effect of heat in the metabolic process of chlorophyll formation is shown in some plants upon which SACHS ('64) experimented. He prepared three culture chambers, all illuminated by a north light. *A* was kept at a high temperature, namely, 30° to 34° C.; *B* was kept at a temperature of 16° to 20° C., and *C* at 8° to 14° C. Into these chambers were put etiolated seedlings of *Phaseolus multiflorus* (bean) and *Zea mais* (maize) which had been reared in the dark. The first traces of turning green appeared in *A* after 1½ hours; in *B* after 2 to 5 hours; whilst in *C* no trace of greening appeared until several days had passed. Thus it appeared that at the temperature of 8° to 14° C. chlorophyll is hardly produced.

We now pass to the consideration of some Protista. An indication, at least, that the rate of metabolism is increased with temperature is gained from the increased rapidity of formation of the contractile (excreting) vacuoles of Ciliata under these conditions. Thus, ROSSBACH ('72, p. 33) found that the rapidity of the rhythmic movements of the contractile vacuole is most intimately related with the temperature of the body, so that one and the same species of animal under normal conditions always has, at a given temperature, the same number of contractions. From the number of the rhythmic contractions one can therefore draw a certain conclusion concerning the existing degree of temperature. This relation between

temperature and interval between contractions is given in Fig. 67. We cannot say that the increment of excretion is exactly equal to that of contraction, but there is doubtless a correlation between the two activities.

From all these facts we may conclude that, within certain limits, an increase of temperature increases metabolism, and a diminution of temperature diminishes it. But the increment in metabolic processes soon finds a limit at a temperature above which the metabolic processes begin to diminish.

2. **Effect of Heat upon the Movement of Protoplasm and its Irritability.**—All observers (DUTROCHET, '37, pp. 777, 778; NÄGELI, '60, p. 77; SACHS, '64; HOFMEISTER, '67, pp. 53, 54; and COHN, '71, for plant cells; and KÜHNE, '59, p. 821; and SCHULTZE, '63, p. 46, for Protozoa) agree that a gradual increase in temperature above that of the ordinary living room results, within certain limits, in an increase in the rate of movement of the protoplasm. A diminution in temperature, on the contrary, causes a decrease in the movement.

For this acceleration with increased temperature, NÄGELI sought to obtain a quantitative expression. He measured the time consumed at different temperatures in the migration through 0.1 mm. of the granules floating in the stream of protoplasm seen in the end cells of *Nitella syncarpa*. Some

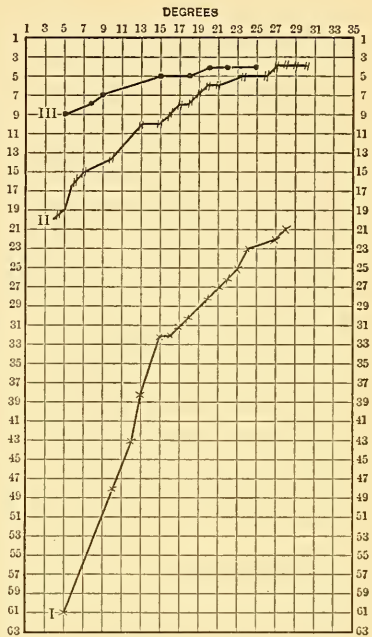


FIG. 67.—The mean thermal curves determined by ROSSBACH for the contractile vesicle of Infusoria. I, for *Euplotes charon*; II, for *Stylonychia pustulata*; III, for *Chilodon cucullulus*. The abscissæ indicate degrees of temperature, Centigrade, in two-degree intervals; the ordinates give the number of seconds elapsing between successive contractions of the vesicle, in two-second intervals. (From SEMPER, "Animal Life.")

measurements were made, also, by SCHULTZE, on *Tradescantia* hairs. But the work of neither of these equals in importance the determinations of VELTEN ('76), which I propose to give in some detail. He determined the time consumed by the

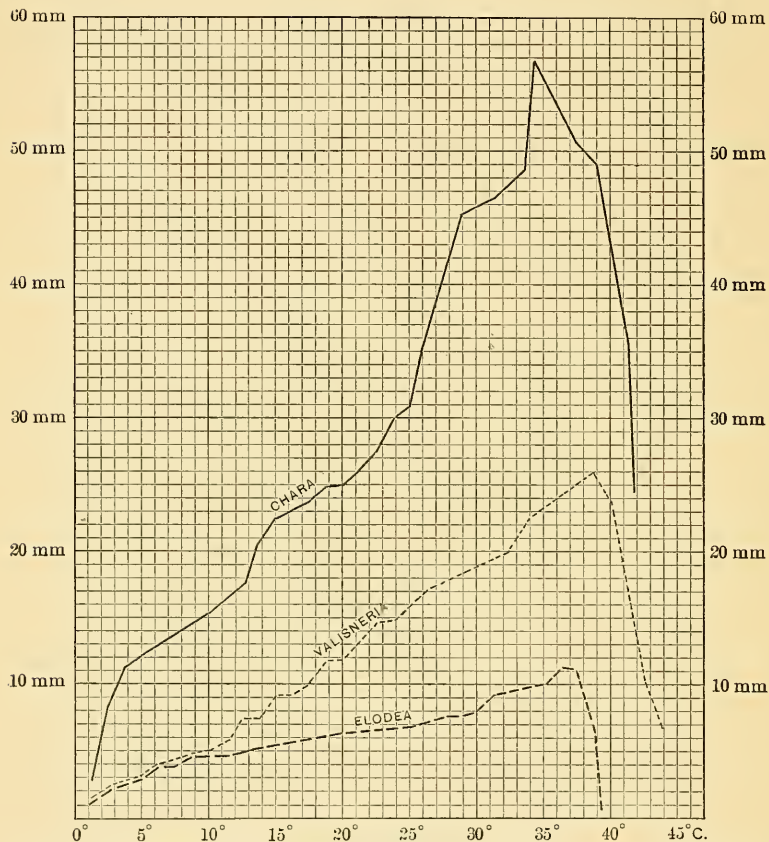


FIG. 68.—Curves showing the relation between temperature (abscissæ) and rate of movement per minute of the chlorophyll grains floating in the protoplasm of the cells of three species of green plants. (Data from VELTEN, '76.)

floating chlorophyll grains in cells of *Elodea canadensis* and *Vallisneria spiralis*, or the small granules near the wall of cells of *Chara* in traversing 0.1 mm., at various temperatures. The results in modified form are given graphically for these three species in Fig. 68. In this figure, the ordinates represent distance (in millimetres) traversed in 1 minute. The abscissæ

represent temperatures (Centigrade) from 0° on the left to 44° on the right. The rate of movement increases regularly up to a maximum (the optimum), a rise of 1° C. being associated with an increased rate of movement in *Chara* of 1.4 mm.; in *Vallisneria* of 0.62 mm.; in *Elodea* of 0.26 mm. The rate of increase is, in general, slightly greater near the optimum. The optimum varies, in the three species, from 34° to 39°. Beyond the optimum the rate rapidly decreases, cessation of movement being reached, in the case of *Elodea*, in 3.7°, in *Vallisneria* in 6.2°, in *Chara* in 8.4° beyond the optimum. The curves exhibited in Fig. 68 are characteristic of other vital functions besides motion, and illustrate this general law, that the optimum temperature for the vital activities lies much nearer to the maximum vital temperature than to the minimum.

After having seen that the rate of flow of protoplasm is dependent upon temperature, we should expect to find, as we do, that that other form of protoplasmic motion, cilia vibration, would be likewise dependent. Some quantitative data concerning relation of temperature and rate of vibration were gained as early as 1858 by CALLIBURCÈS. He placed a bit of ciliated membrane from the frog's œsophagus in a moist chamber, and in contact with the cilia he laid a small glass cylinder, horizontally supported, and provided with a dial by which its revolutions could be counted. He found that the mean time for a revolution was, —

at 12° to 19° C. . . . .	22 minutes, 3 seconds;
at 28° C. . . . .	3 minutes, 7 seconds;

thus, in increasing the temperature from 15° to 28° C., the rate of vibration is increased sevenfold.

Essentially similar results were obtained, through the use of new methods, by ROTH ('66, pp. 185-189), upon the ciliated epithelium of the frog's uterus, rabbit's trachea, and gill of *Anodonta*, and by ENGELMANN ('68, pp. 381-384; 443, 444; 454, 455; and '77), upon the frog's œsophagus. The simplest of these methods was to determine the rate of the transportation of fine particles over the surface of the tissue. The result of these studies showed that the optimum temperature for the

movements of Vertebrate cilia lies between  $35^{\circ}$  and  $40^{\circ}$  C., and that a gradual elevation of temperature to this point is accompanied by gradual increase in the rapidity of the stroke, the law of which is exhibited in the curves shown in Figs. 69 and 70.

This variation in rate and regularity of cilia movement with change in heat is marked in Infusoria, as ROSSBACH ('72, p. 312)

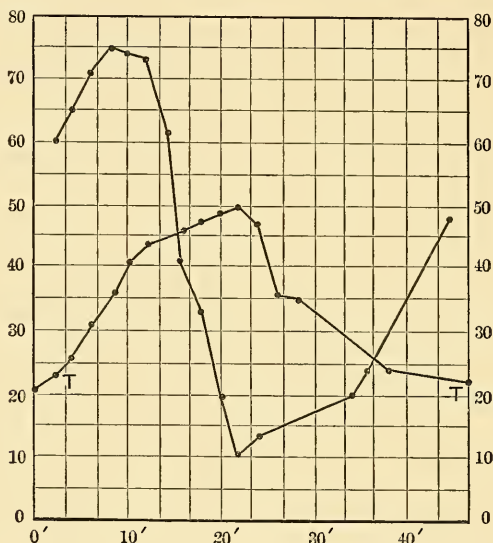


FIG. 69. — Curves showing relation between temperature (curve *TT*) and rapidity of movement of the cilia of the mouth and œsophagus of the frog. The abscissæ give, in minutes, the lapse of time from the beginning of the experiment. The ordinates give, for the temperature curve, the degrees Centigrade, and, for the other curve, the corresponding number of units of motor activity for the immediately preceding 2 minutes as registered by ENGELMANN'S apparatus. (From ENGELMANN, '77.)

and SCHURMAYER ('90, pp. 411, 412) have shown. The lower the temperature falls below  $15^{\circ}$  C., the slower the locomotion, almost ceasing at  $+4^{\circ}$ . Upon raising the temperature above  $15^{\circ}$ , motion quickens, until, between  $25^{\circ}$  and  $30^{\circ}$ , motion reaches a maximum, the Ciliata shooting back and forth with the quickness of an arrow. Between  $30^{\circ}$  and  $35^{\circ}$ , the movements become still more violent, and take on a new character. They are no longer coördinated. Towards  $40^{\circ}$ , the progres-

sive movement becomes slower, and finally ceases, while the rotation continues, but in ever diminishing rapidity. A new axis of rotation is assumed, running lengthwise and obliquely, or running transversely, in the short axis of the body. Finally, somewhere between  $38^{\circ}$  and  $40^{\circ}$ , motion ceases. Thus, the optimum temperature for the activities of the protoplasm lies at about  $30^{\circ}$  C., and the maximum temperature is perhaps  $10^{\circ}$  higher.

The experiments upon plant protoplasm, amœboid organisms, and ciliated cells thus agree in demonstrating a close relation be-

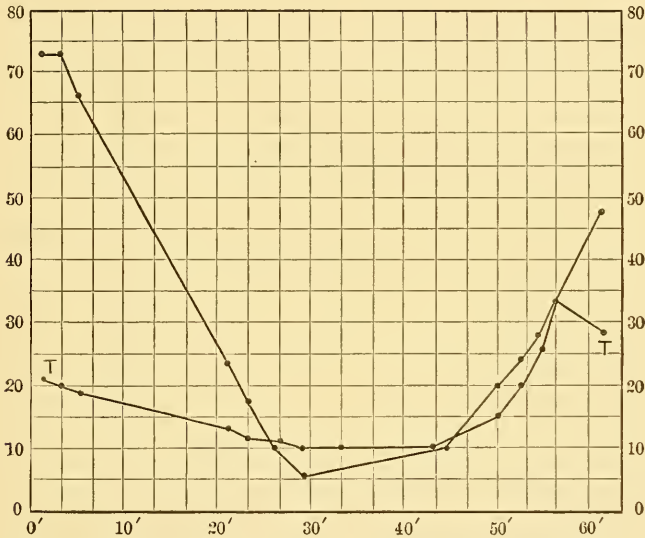


FIG. 70. — A set of curves having the same meaning as those of Fig. 69. In this case, however, the temperature first falls and then rises, instead of rising first and then falling. (From ENGELMANN, '77.)

tween temperature and protoplasmic movement. This relation is such that, as the temperature is elevated above the freezing point of water, the movements regularly increase, and reach their greatest activity at about the temperature of slow-running waters in the midst of summer, namely, about  $25^{\circ}$  to  $30^{\circ}$  C.\*

\* The maximum temperature attained by bodies of ordinary water inhabited by organisms seems to be close to the position of the optimum temperature of organisms. Yet, temperature data concerning the waters from which the organ-

Above this temperature, the rate of protoplasmic movement rapidly decreases.

That temperature influences the *irritability* of protoplasm is demonstrated by many facts. Thus, STRASBURGER ('78, p. 611) found that, at a high temperature, the light attunement of swarm-spores changes. For example, swarms of *Hæmatococcus* and *Ulothrix*, which are — phototactic, move at 30° C. from the — to the + side of the drop. LOEB ('90, p. 43) has found that *Prothesia* larvæ do not respond to light at a temperature below +13° C. Similarly, in respect to geotaxis, SCHWARZ ('84, p. 69) found that *Euglena* did not respond at below 5° to 6° C. So, too, CAMPBELL ('88, p. 130) has shown that the response of muscles to electric stimulus varies with the temperature. Thus, with the neck muscles of the tortoise at —

TEMPERATURE.	NUMBER OF SHOCKS PER SECOND REQUIRED TO TETANIZE.
4° C.	1
9° C.	5
21° C.	25
28° C.	34

isms of the experiment have been taken are, unfortunately, rarely given in experiments on heat. From observations made by the Massachusetts State Board of Health (Report on Water Supply and Sewage, 1890, Part I, p. 660), it appears that the various ponds and reservoirs in the state, having a depth varying from 19 to 5 metres, had a mean August (maximum) temperature ranging (in the different ponds) from 24° to 21° C. Various rivers, mostly not of mountain origin, had, in 1887, a mean July (maximum) temperature, varying (in different cases) from 26.5° to 24° C. Even in those ponds and streams in which the surface temperature is over 25°, a lower temperature can be found below the surface. Thus, from the report referred to, it appears that, while at 3.3 metres below the surface we have nearly the surface temperature, at 6.6 metres below the surface, we find a decrease of 3° to 10°, and at 10 metres a decrease of from 9° to 17° below the surface temperature in different ponds. As for the sea, the highest recorded surface temperature is about 32° C. (Red Sea, Gulf of Mexico.) See A. AGASSIZ, "Three Cruises of the Blake," Bull. Mus. Comp. Zoöl., XIV, p. 301. It would be a valuable piece of work to determine the maximum summer temperature attained by the waters of shallow ponds, pools, and marshes inhabited by organisms.



This shows that the muscle responds, and tends to return to its uncontracted condition less quickly at a low than at a higher temperature. In general, then, protoplasm is more responsive, the closer we approach its optimum temperature.

### § 3. TEMPERATURE-LIMITS OF LIFE

In the preceding sections we have seen that, as the temperature is raised above the optimum, or as it approaches  $0^{\circ}$  C., the vital activities begin to diminish. Finally, we meet with a higher or a lower limit, at which all movement and the processes of metabolism cease. This point may be called, in the case of the higher limit, the maximum, and in the case of the lower limit, the minimum. The maximum and the minimum are not points of death, but merely of cessation of activity, lasting while the temperature endures, but being replaced by renewed activity when the temperature is shifted towards the optimum. This quiescent, or latent, condition of the protoplasm near the vital limits of temperature may be called temporary rigor (German "Starre") to distinguish it from death. Death occurs at a very few degrees beyond temporary rigor.

1. **Temporary Rigor and Death at the Higher Limit of Temperature, Maximum and Ultramaximum.** (ENGELMANN, '79, p. 358.)—The occurrence, at a high temperature, of a condition resembling death, except that the organism may revive from it, seems first to have been noticed by P. DE CANDOLLE ('06, p. 346). He found that a Sensitive plant, kept 11 hours at  $37^{\circ}$  C., lost all sensibility to touch, and did not close with the coming on of night. Maintained, during the following day and night, at a temperature of about  $20^{\circ}$ , it remained insensitve during that period; but on the succeeding night it closed its leaves, and on the following day had regained its sensitiveness to touch. Thus, the high temperature of  $37^{\circ}$  had produced an immotile state which was not death, since it was only temporary. It may be called the state of temporary heat-rigor.

The earliest record which I have found of a similar observation among animals is that of PICKFORD ('51). He states that, at a high temperature, muscle went into a rigid con-

dition ("Scheintodtenstarre") from which it might return to a normal condition of sensitiveness. Such a rigid state was brought about by subjecting a decapitated frog in water to 35° R. (43.8° C.) for 1 minute. I will now add some additional cases of production of temporary heat-rigor in protoplasm which I have found in the literature.

In 1863, MAX SCHULTZE (pp. 33, 34) found temporary heat-rigor in *Actinophrys*, which retracts its pseudopodia and appears as a lifeless mass at 35° to 38°, but is not killed until 43° is reached. In the same year SACHS ('63, p. 453) repeated more fully the experiments of P. DE CANDOLLE on *Mimosa pudica*. He found that a temperature of 30° C. for 3 hours did not produce rigor. A temperature of 40° for 1 hour produced loss of sensibility during 20 minutes. Raised slowly even to 50°, sensibility was only temporarily lost, but 52° proved fatal. Immersed in water, heat-rigor occurred at a temperature 5° to 10° lower. SACHS clearly distinguishes a "vorübergehende Wärmestarre" from death.

KÜHNE ('64, pp. 45, 67, 87, 103) drew a sharp contrast between the rigidity of death, which he calls "Wärmestarre," and the transitory immobile condition or "Wärmetetanus." He found this latter condition to occur in *Amœba* subjected to 35° for 1 minute, in *Actinophrys* subjected to 35°-40° for several minutes, in motile *Myxomycetes* (*Didymium serpula*) subjected to 30° for 5 minutes, in *Tradescantia* stamen hairs at over 45°, when gradually brought to that temperature. In all cases there is such a relation between temperature and time of subjection that the greater the one is the less need be the other in order to produce heat-rigor.

Very instructive also are the observations of HOFMEISTER ('67, pp. 54, 55) which I briefly summarize: Hairs from the stem and leaf of *Ecbalium ageste* showing lively movement were gradually raised from 16°-17° C. to 40° C. They became motionless at 40° C. After 1 to 2 hours, movement returned, and was very violent. Cooled and raised again to 45° C., the protoplasm was motionless at first, but after 17 minutes movements recurred but were not rapid. Put again into 47.5° (after first cooling) heat-rigor occurred in 5 minutes, but upon cooling, movements return.

Very similar experiences have befallen subsequent investigations which unite in supporting the conclusion that at a certain temperature, slightly below the death point, protoplasm becomes immobile, but retains the capacity for subsequent reacquisition of movement upon lowering the temperature.

Finally, studies upon muscle, especially those of CHMULEVITCH ('69), SAMKOWY ('74), MORIGGIA ('91), GOTSCHLICH ('93), and others, have shown that as the temperature is elevated up to about 30° C., the muscle contracts more and more,

lengthening again as the temperature falls. If, however (GOTSCHLICH), the temperature is raised in about 60 seconds to 38° and then lowered, the elongation of the muscle takes place only very slowly. This is the condition of "thermische Dauerverkürzung," and is probably the same as the condition of temporary heat-rigor of SACHS. When, however, the temperature is raised rapidly to 45° to 50° (or slowly to 35°), death-rigor appears, accompanied by a coagulation of the protoplasm which renders the whole mass opaque and permanently contracted. The rapidly replaced contraction accompanying elevation to about 30°, the slowly obliterated contraction of 38°, and the permanent contraction of 45° are then three stages in a series of effects of heat on muscle.

If now, contraction, heat-rigor, and death-rigor are merely three stages in a series of effects of increasing temperature, they probably have related immediate causes. Heat-rigor is certainly a condition of tetanus, but the fact that the protoplasm in this condition is not sensitive and cannot quickly return to the relaxed condition indicates that some of those changes that produce death-rigor have already occurred, but not to such an extent that the organism cannot recover from them. As GOTSCHLICH says ('93, p. 154), "Die thermische Dauerverkürzung ist also eine qualitative unvollendete Starre" (*i.e.* death-rigor). From this point of view there is no exact point at which heat-rigor occurs, since the period of persisting rigidity varies in extent from 0 to many hours, and thus passes by almost imperceptible gradations from a contraction in response to heat on the one hand to death-rigor on the other. The muscle increases in sensitiveness as the temperature rises to the optimum, just as the movements of plasma in Chara do. Beyond the optimum, sensitiveness diminishes, and this leads to a condition of heat-rigor which becomes the more pronounced the higher the temperature, until, through completed coagulation, death occurs.

We must now consider this point at which death occurs from heat; and, as an introduction to this discussion, we may tabulate the results of experiments by numerous observers who have attempted to determine the ultramaximum temperature.

TABLE XIX

RESULTS OF EXPERIMENTS TO DETERMINE THE ULTRAMAXIMUM TEMPERATURE OF ORGANISMS IN WATER, OR THE TEMPERATURE AT JUST ABOVE WHICH ORGANISMS REARED UNDER NORMAL CONDITIONS WILL DIE

SPECIES.	MAXIMUM TEMPERATURE.	CONDITIONS OF EXPERIMENT.	AUTHORITY.
<i>Cryptogams.</i>			
Bacteria . . . . .	45° C.	Maximum temperature of growth in liquid	COHN, '77, p. 253; '94, p. 150
Yeast . . . . .	53°	Moist; average max.	SCHÜTZENBERGER, '79, p. 162
Oscillatoria . . . . .	45°	} Death point	DE VRIES, '70, p. 388
Nostoc . . . . .	42°		
Spirogyra } . . . . .	44°		
Œdogonium } . . . . .	46°		
Hydrodictyon . . . . .	45° to 60°	} Gradually raised	SACHS, '64, p. 5 DUTROCHET, '37, p. 777
Cladophora . . . . .	45°		
Nitella flexilis . . . . .	45°		
Funaria hygrometrica . .	43°		DE VRIES, '70, p. 388
Marchantia polymorpha } Lunularia vulgaris }	46°		" "
<i>Phanerogams.</i>			
Vallisneria spiralis . . . .	45° to 50°	} Gradually raised Suddenly immersed for 10 minutes	SACHS, '64, p. 5
Ceratophyllum demersum	45° to 50°		
Various plant cells . . . .	47° to 48°	Died (suddenly subjected)	SCHULTZE, '63, p. 48
<i>Protozoa.</i>			
Æthelium sept. . . . .	40°	Plasmodium died after 2 minutes	KÜHNE, '64, p. 87
Amœba . . . . .	40° to 45°	Death point	" "
Actinophrys . . . . .	42°	Death point. Activity lost at 38°	SCHULTZE, '63, p. 34
Miliolida . . . . .	43°	Death point	SCHULTZE, '63, p. 38
Various Flagellata and swarm-spores . . . . .	40° to 60°	45° to 60° most usual. Heat-rigor usually occurs between 40° to 50° and is lower for marine than for f. w. species. These temperatures for the motile stage	BÜTSCHLI, '84, p. 860; STRASBURGER, '78, p. 611; DALLINGER, '80, p. 10
Various Infusoria . . . . .	45°	Can withstand only a short time	SCHÜRMAYER, '90, p. 412

SPECIES.	MAXIMUM TEMPERATURE.	CONDITIONS OF EXPERIMENTS.	AUTHORITY.
Paramecium . . . . .	42° to 46° C.	Gradually subjected	MENDELSSOHN, '95, p. 19
Stentor . . . . .	44° to 50°	Heat-rigor point, temperature raised gradually. From a pool kept warm by boiler waste	DAVENPORT and CASTLE, '95, p. 229
Vorticellidæ . . . . .	41° to 42°		SCHULTZE, '63, p. 49
<i>Celenterata.</i>			
Actinia . . . . .	38°	Gradually raised (1 hour)	FRENZEL, '85, p. 464
Beroe ovatus . . . . .	40°	Death point, suddenly subjected	DE VARIGNY, '87, p. 63
<i>Mollusca.</i>			
Various Mollusca . . . . .	30° to 40°	Suddenly immersed	FRENZEL, '85, p. 461-466
Pleurobranchæa . . . . .	33°	Temp. gradually raised	" "
Aplysia . . . . .	33°	Died in 3 hours	" "
Eledone . . . . .	35°	Died	" "
Young squids . . . . .	37°	Heat-rigor; died at 41°	BERT, '67, p. 135
<i>Vermes.</i>			
Turbellaria . . . . .	44.5°	Death point	SCHULTZE, '63, p. 49
Anguillulidæ . . . . .	44.5°	" "	SPALLANZANI, 1787, Tom. I., p. 56 SCHULTZE, '63, p. 49
Rotifera } . . . . .	45° to 48°	Moist	DOYÈRE, '42, p. 29
Tardigrada } . . . . .	98°	Dried	BROCA, '61, p. 44-46
Diopatra . . . . .	40°	Suddenly immersed, died quickly; at 30° lived indefinitely	FRENZEL, '85, pp. 461-465
Terebella . . . . .	27° to 30°	Suddenly heated; slowly warmed, resisted 30°	" "
Naididæ . . . . .	44.5°		SCHULTZE, '63, p. 49
" Bloodsucker " . . . . .	44°	Death point	SPALLANZANI, 1777, Tom. I., p. 56
<i>Crustacea.</i>			
Daphnia sima . . . . .	33.5°	Suddenly subjected	PLATEAU, '72, p. 316
Cyclops quadricornis } . . . . .	36°	" "	" " 317
Cypris fusca } . . . . .	36°	" "	" " 316
Gammarus rösели . . . . .	36°	" "	" " 316
Asellus aquaticus . . . . .	43.5°	" "	" " 316

SPECIES.	MAXIMUM TEMPERATURE.	CONDITIONS OF EXPERIMENT.	AUTHORITY.
Palæmon . . . . .	26° C.	Died in 2 hours (suddenly subjected)	FRENZEL, '85, p. 463
Scyllaris . . . . .	30°	Died in 1 hour (suddenly subjected)	" "
Pagurus prideauxii . . . . .	36°	} Death point	DE VARIGNY, '87a, p. 173
Dromia vulgaris . . . . .	38°		
Pisa gibbosa . . . . .	36°		
Portunus puber. . . . .	34°		
Carcinus sp. . . . .	38°		
Grapsus sp. . . . .	38°		
<i>Arachnida.</i>			
Argyroneta aquatica . . . . .	38.5°	Suddenly subjected	PLATEAU, '72, p. 316
Hydrachna cruenta. . . . .	46.2°	Submerged (?)	" "
<i>Insecta.</i>			
Podura . . . . .	27°	Suddenly subjected; died slowly. At 36° died at once	NICOLET, '42, p. 11
Agabus bipustulatus . . . . .	38°	} Death point	PLATEAU, '72, p. 316
Hydaticus transversalis . . . . .	39°		
Culex pipiens, larva . . . . .	40°		
Hydrophilus caraboides . . . . .	42°		
Hydroporus dorsalis . . . . .	42°		
Nepa cinerea	} 44° to 45°		
Notonecta glauca . . . . .			
Cloë diptera, larva			
Musca vom. (?) . . . . .	37.5°	} Death point	SPALLANZANI, 1787, Tom. I, pp. 56-58
Musca vom., larva . . . . .	42.5°		
Musca vom., pupa . . . . .	43.7°		
Silk worm larva . . . . .	42.5°		
"Butterfly" larva . . . . .	42.5°		
Culex larva . . . . .	43.7°		
<i>Echinodermata.</i>			
Antedon . . . . .	30°	Died rapidly (suddenly subjected)	FRENZEL, '85, pp. 460-463
Holothuria . . . . .	30° to 40°	Died in several hours (suddenly subjected)	" "
<i>Vertebrata.</i>			
Many fresh-water fishes . . . . .	40°	Survived only a few seconds	EDWARDS, '24, p. 114
	36°	In pond out of doors.	KNAUTHE, '95, p. 752
Fish . . . . .	33°	Temperature elevated gradually	BERT, '76, p. 169
	27° to 38°		DAVY, '63, p. 125

SPECIES.	MAXIMUM TEMPERATURE.	CONDITIONS OF EXPERIMENT.	AUTHORITY.
Hippocampus . . . . .	30° C.	Lived half an hour	FRENZEL, '85, p. 462
Salamander . . . . .	44°	Death point	SPALLANZANI, 1787, Tom. I, p. 56
Frog . . . . .	40° to 42°	Suddenly subjected in water: death at once	EDWARDS, '24, p. 374
Frog, adult (summer) . .	42° to 43°	Death-rigor in 7 to 14 minutes	MORIGLIA, '91, p. 385
Frog, adult . . . . .	43.8°		SPALLANZANI, 1787, p. 55
Frog, tadpoles . . . . .	41°	Raised in from 5 to 10 minutes	See p. 253
Rabbit } . . . . .	44° to 45°	Death point when raised gradually; convulsions at 42°	OBERNIER, '66, p. 22
Dog }			
Mau . . . . .	45°	In water; giddiness in a few seconds	EDWARDS, '24, p. 374
Human spermatozoa . . .	50°	Died in 10 minutes	MANTEGAZZA, '66, p. 186
Vertebrate muscle . . . .	40° to 50°		KÜHNE, '59, pp. 784-804
Vertebrate muscle (frog) {	45° to 50° 35°	Raised in 30 seconds	GOTSCHLICH, '93, p. 123
		Raised in 18 minutes	

The determinations given in the above table may be compared only with caution, for diverse conditions give results which cannot be directly correlated. Thus individuals of the same species, but reared under diverse environment, have a different resistance period to heat. The lethal temperature varies according as the organisms are suddenly or gradually subjected to the high temperature. Also, the individuals of the same species will die at different temperatures according as they are rapidly subjected to the high temperature or gradually accustomed to it, and, as we have seen, a lower temperature, long continued, often produces the same result as a higher temperature during a brief period. Finally, too little care has been exercised in most cases to determine the temperature of the water immediately upon, and a few minutes after, placing the animals in it,—an operation which lowers the temperature of the water. In the experiments cited, unless otherwise stated, the conditions were gradual subjection continued for a short time only. The quality of the water in which the

experiments were carried on is supposed to be, except for its temperature, normal for the species. Summarizing the table, we find that the Protista have the highest maximum temperature of any group, it being in extreme cases about 60° for active organisms, but generally between 40° and 45°. Among the Metazoa, the highest maxima recorded (excepting Rotifera and Tardigrada) are 44° to 45° for Turbellaria, *Anguillula*, Naïdidae, Nepa (water-scorpion), Notonecta (water-boatman), Cloë larva, Salamander, and mammals. A water-mite (*Hydrachna*) is said to have withstood up to 46.2°, and some vertebrate tissues resist up to 50°. For the great majority of Metazoa, the maximum temperature lies below 45° and, in the case of marine species, below 40°. The low maximum temperature of marine species is probably due to the low maximum temperature of the sea as compared with ponds. We may consequently conclude from the foregoing that the maximum temperature for protoplasm lies generally between 35° and 50°, the lower limit being characteristic of organisms living in a medium of low temperature (the sea), the latter, of organisms reared in warm pools or of organs (vertebrate muscle) in a body kept at a high temperature.

The question now arises, what is the cause of the death of protoplasm at high temperatures? To get some insight into this matter, let us examine the phenomena accompanying death of protoplasm from overheating. KÜHNE ('64, p. 44) thus describes the appearance of an *Amœba* subjected for a moment to a fatal temperature (45°). The structure is entirely altered since it has become transformed into a mass of knobbed, opalescent, solid lumps, which, even in transferring to the slide, become easily broken apart. This appearance is clearly due to a coagulation of the protoplasm. A similar coagulation takes place in *Actinophrys eichhornii* (KÜHNE, '64, p. 67) at 45°. "The sphere shrinks into a flat, hardly transparent, cake, no longer reacts to the strongest induction shocks, and breaks up after 24 hours into a heap of small granules and irregular pieces." Likewise, in muscles a change is produced by heat which is evidently a kind of coagulation. A coagulation then seems to be the immediate cause of death at high temperatures.

But just what is the component of protoplasm which co-



agulates at the death point of organisms? As KÜHNE ('64, p. 1) pointed out, it cannot be ordinary egg albumen; for, excepting the contractile substance, we know of no native albumen which coagulates between 35° and 50° C. It was KÜHNE'S great service to show that there is a substance which can be pressed out of frozen, triturated, and then thawed muscle, which becomes quickly opalescent at 40°, through the separation of the muscle plasma into myosin and a serum. This serum, in turn, contains an albuminoid which coagulates at 47° (DEMANT, '79 and '80). Now, since there are proteids in muscle which coagulate at about the point at which muscle goes into permanent heat-rigor, and since these proteids can no longer be squeezed out of rigid muscles, the conclusion seems justified that permanent heat-rigor in muscle is due to the coagulation of these proteids.

Related, easily coagulable proteids occur in widely dissimilar organisms. For example, myosin has been found in vegetable protoplasm (WEYL, '77, p. 96), and HALLIBURTON ('88) has described a globulin from blood corpuscles which coagulates at 48° to 50°. Their distribution in protoplasm is, therefore, probably general, and so we are justified in concluding that the death of protoplasm by heat is, in general, the result of the coagulation of a proteid (globulin). Death occurs because the vital machinery has been broken down.\*

**2. Temporary Rigor and Death at the Lower Limit of Temperature, Minimum and Ultraminimum.**—Whilst towards the upper limit of ordinary terrestrial temperatures (35° to 40° C.) molecular changes in organic compounds are hastened, towards the lower limits (−40° to −50°) molecular changes are slow, being principally confined to the transformation from the liquid or gaseous to the solid or liquid condition. This transformation does occur in the water of protoplasm, but the colloids,

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\* At this place reference may be made to the fact that protoplasm subjected to a high temperature sometimes breaks to pieces with the suddenness and completeness of an explosion. Thus STRASBURGER ('78, p. 611) found that *Chilomonas curvata* was uniformly killed at 45° C. by the explosion of the body, and Dr. W. E. CASTLE tells me that he has observed the same phenomenon under like conditions in *Stentor*. An investigation of this profound change in protoplasm would be sure to throw valuable light upon the nature of the living substance.

which constitute the living part, are not modified by even the lowest of the terrestrial temperatures, except that the molecular changes which they undergo are very slow. This being true, protoplasm which contains no water, or very little, ought not to be changed by low temperatures, — that is to say, the *machine* will not be injured.

With the activities of the machine — with the vital processes — it is, however, quite different. They are essentially chemical processes, and hence we should expect them to be diminished at a low temperature. If, as PICTET ('93) maintains from an extensive and highly important series of experiments, no chemical processes take place at temperatures below  $-100^{\circ}$  C., then protoplasm ought to exhibit no vital processes at this temperature, and, indeed, experience shows that, as we have already seen, as the temperature is lowered below the optimum, all manifestations of activity diminish. It is clear that at a certain point they must entirely cease. And at that point death, following the usual definition of the word, would ensue.

But does the cessation of the vital processes, without injury to the mechanism, necessarily preclude the possibility of a return to activity? Let us examine the experimental evidence on this point. SCHUMACHER ('74, p. 179) subjected yeast to cold and found that, at the lowest temperature produced ( $-113.7^{\circ}$ ), the yeast cells were not completely killed. More recently, PICTET ('93<sup>a</sup>, cf. also C. DE CANDOLLE, '84) has submitted various dry seeds and spores of bacteria to a temperature of nearly  $-200^{\circ}$ , at which temperature the atmosphere becomes liquefied, but without fatal effects. Other results were still more remarkable: vibratile cilia from the mouth of the frog were cooled to  $-90^{\circ}$ , and recovered their movement upon raising the temperature. Some Rotifera and Infusoria were frozen in their native water at  $-60^{\circ}$ , and kept at that temperature, apparently, for nearly 24 hours. Most have subsequently regained their activity. Eggs of the frog, lowered slowly to  $-60^{\circ}$ , can revive. Eggs of the silk-worm can resist to  $-40^{\circ}$ . Other experiments of PICTET\* will be referred to

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\* This general criticism of PICTET's paper is, I believe, valid. He does not give us data enough upon time of subjection to the low temperature, time employed in reducing the temperature, and other details. Thus, concerning his

later. As a result of these and others' investigations we may conclude: Protoplasm may under certain circumstances, of which one of the most important is the absence of water, resist, uninjured, the lowest temperatures. *There is no fatal minimum temperature for dry protoplasm.*

We must now turn our attention to those cases in which the phenomena of cessation of activity or death appear, and seek to determine their causes; and first concerning temporary cold-rigor. We have already seen that as the temperature is lowered, the rate of metabolic processes and protoplasmic movements is lowered. What happens at the lower limit of activity, and where does this lie? The chlorophyll granules of *Vallesneria* move (according to VELTEN) only about 1 mm. per minute at 1° C. and not at all at 0°; the rotation of *Nitella* ceases (NÄGELI, '60, p. 77) at 0° C.; in *Tradescantia* hairs, movement is wholly arrested on freezing the cell sap (KÜHNE, '64, p. 100, and DEMOOR, '94, p. 194). Even in seeds and bacteria, which are not killed by the lowest temperatures, all vital activities have probably ceased at 0°, for DE CANDOLLE ('65) found that in only one species out of ten could he get a seed kept at 0° to germinate, and even then germination was so retarded that it took from 11 to 17 days as opposed to 4 days at 5.7°. Likewise, bacteria do not multiply below +5° to +10° (BONARDI and GEROSA, '89). Among animals, KÜHNE ('64, p. 46) found *Amœba* cooled to near 0° almost motionless. PURKINJE and VALENTIN ('35) first noticed that the ciliated

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experiments on *Scolopendra*, he merely says: "I have frozen to -40° three *Scolopendras* which perfectly resisted the treatment and lived after thawing out. Submitted to -50°, they have also resisted. Frozen a third time to -90°, they are all three dead." Now, in the absence of further data, it is quite possible that the heat of metabolism kept the internal body temperature considerably above that of the chamber, the thick cuticula preventing rapid loss of heat, very much as a man's clothing enables him to withstand the -40° of an arctic winter. Another experiment of PICTER's lends greater probability to this explanation of some cases of great resistance. Three snails were subjected to a temperature of from -110° to -120° during several days. The operculum of two of these was not intact, so that it did not close the orifice. These two individuals died; but the third, which was completely sealed up, survived. Those which were not sufficiently clad, so to speak, lost so much internal heat that their internal fluids were frozen. Of course this criticism cannot apply in the case of those organisms mentioned in the text which are without a thick cuticula.

epithelium of the frog ceased its movements at  $0^{\circ}$ . Muscles of the frog were found by KÜHNE ('64, p. 3) to become at  $-3^{\circ}$  to  $-7^{\circ}$  a solid lump which did not, however, wholly lack irritability. The evidence of all these cases shows that activity nearly ceases in protoplasm at or near  $0^{\circ}$  C.

Another effect produced on protoplasm by cold — an effect which often immediately precedes quiescence — is violent contraction. This has been repeatedly observed. The protoplasm of *Tradescantia* hairs, which has been in cold-rigor, was found by KÜHNE ('64, p. 101) to lie in separated rounded drops and lumps, — an appearance like that resulting from excessive stimulation. The rapid freezing of muscle gives rise, according to HERMANN ('71, p. 189), to violent contractions. The sciatic nerve of the frog's leg when cooled to  $-4^{\circ}$  to  $-8^{\circ}$  causes clonic contractions of the muscle, lasting two minutes. (AFFANASIEFF, '65, p. 678, and others.) It is clear, then, that cold acts as a violent stimulus to protoplasm.

The final result of temporary rigor is thus clearly brought about by the coöperation of two causes: (1) the diminution in the chemical processes upon which metabolism and movement depend, and (2) the directly stimulating effect of the cold, which acts like contact or excessive heat. Both causes work to produce a quiescence which may be replaced by activity when the causes are withdrawn.

The fact that cold-rigor usually occurs close to the zero-point indicates that the activities of protoplasm are closely determined by the fluid state of water. This fact is not to be explained on the ground that freezing prohibits all chemical change — many chemical changes take place below the freezing point of water, but, apparently, few of those which are involved in metabolism. Nor is the rigor due to the change which the freezing of the protoplasmic fluids brings, because as the temperature approaches the zero-point, but while the water is still perfectly fluid, metabolism diminishes; and it diminishes at such a rate as to cease just where water begins to freeze. The critical point for vital activity has been adjusted to this critical point of water.

So, too, the composition of protoplasm is such that at a temperature, lying below the normal and above the freezing point

of water, those chemical changes rapidly occur which we designate response to the stimulus of cold. This composition of protoplasm, upon which cold can work such important modifications, is a quality of immense importance in the economy of the organism, as the changes of each autumn testify.

Below the point of temporary cold-rigor lies that of death, if death point there be. The position of the death point is, however, very diverse in different organisms. Part of the diversity in the death points assigned by different authors is, however, due to the fact that in the methods of determining the death points there has been a lack of uniformity.

Five elements ought always to be regarded in experiments on the ultraminimum temperature. (1) History of the temperature conditions in which the individual or its race had lived before experimentation; (2) rate at which the organism has been cooled, — if possible, the temperature of the organism itself rather than that of the medium; (3) intensity of cold just sufficient to kill; (4) duration of application of the cold and the kind of medium in which the organism is subjected to the cold;\* and (5) the rate of thawing out.† These elements have been too much neglected in the past.

I shall now present in tabular form some of the more reliable determinations of the death point of organisms, prefacing with the caution that the results are not closely comparable.

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\* The duration of application and intensity of the fatal cold stand in an inverse relation, so that organisms which resist a temperature  $-A^{\circ}$  for  $X$  minutes will resist a lower temperature,  $-(A + a)^{\circ}$ , for a shorter time,  $X - x$ . Thus, *Clepsine complana* resists  $-8^{\circ}$  C. for 15 minutes;  $-5^{\circ}$  C. for 90 minutes. So *Planorbis corneus* resists  $-7^{\circ}$  for 5 hours, but  $-5^{\circ}$  for 48 hours. Again, *Musca domestica* can resist  $-12^{\circ}$  for 5 minutes;  $-8^{\circ}$  for 20 minutes; and  $-5^{\circ}$  C. for 40 minutes. (ROEDEL, '86.) Since many authors have little regarded the duration of action of the cold, their determinations have little scientific value.

† The importance of this is illustrated by some experiments of SACHS ('60, p. 177), who found that the leaves of the beet or cabbage frozen at from  $-4^{\circ}$  to  $-6^{\circ}$  died if they were thawed in air at  $2^{\circ}$  or  $3^{\circ}$ , or in water at  $6^{\circ}$  to  $10^{\circ}$ ; but lived when slowly thawed in water at  $0^{\circ}$ . In general, the more gradual the thawing, the lower the fatal temperature.

TABLE XX\*

DETERMINATIONS OF THE ULTRAMINIMUM OF ORGANISMS REARED UNDER NORMAL CONDITIONS

SPECIES.	MINIMUM TEMPERATURE.	CONDITIONS OF EXPERIMENT.	AUTHORITY.
<i>Plant cells.</i>			
Tradescantia . . . . .	-14° +	In water, rapidly frozen	KUHNE, '64, p. 100
Hair cells . . . . .	-14° -	In air, rapidly frozen	
Swarm-spores . . . . .	- 1°	Fully frozen, cautiously thawed	STRASBURGER, '78, p. 612
Swarm-spores (Proto-coccus) . . . . .	0° to -1°		COHN, '50, p. 720
<i>Protozoa.</i>			
Amœba . . . . .	0° -	Rapidly frozen on slide over ice and salt	KÜHNE, '64, pp. 46-47
<i>Animal tissues.</i>			
White blood corpuscles:			
of Amphibia . . . . .	-2° to -3°	During 8 hours; warmed rapidly	SCHENK, '69, p. 26
of rabbit . . . . .	- 7°	For a short time; warmed rapidly	" " 26
Saliva corpuscle . . . . .	- 3°	During 15 minutes	" " 26
Red blood corpuscle . . . . .	-6° to -8°	Over 60 minutes	" " 27
	-15° +		POUCHET, '66, p. 18
Spermatozoa:			
of Amphibia . . . . .	-4° to -7°		SCHENK, '69, p. 29
of Mammalia . . . . .	- 6° -	Returned to activity on thawing	" " 30
of frog . . . . .	- 8° to -10° -	Frozen in testis	PREVOST, '40.
of frog . . . . .	-10° to -12°		QUATREFAGES, '53, p. 353
of man . . . . .	-17°	Gradually thawed	MANTEGAZZA, '66, p. 183
Eggs of Amphibia . . . . .	- 7°	During 1 hour	SCHENK, '69, p. 28
Ciliated epithelium of Anodonta . . . . .	- 6°	Subjected a very short time	ROTH, '66, p. 189
	- 3°	Subjected during 6 minutes	" " 189

\* Temperatures all in degrees Centigrade. — before a number indicates below zero, — or + after a number indicates that the true lethal temperature lay slightly below or above that number. (A) indicates that the organism was in air; (W), in water.

SPECIES.	MINIMUM TEMPERATURE.	CONDITIONS OF EXPERIMENT.	AUTHORITY.
<i>Platyhelminths.</i>			
<i>Dendrocælum lacteum</i>	0° to -1°	Suddenly or gradually subjected, till ice forms	ROEDEL, '86, p. 207
<i>Mollusca.</i>			
<i>Helix hispida</i> . . . . .	- 8°	During 30 minutes	" " 191
<i>Helix pomatia</i> . . . . .	-10°	During 600 minutes	" " 192
<i>Helix pomatia</i> . . . . .	(-14° to -18°)+	Gradually frozen for 180 minutes, and then thawed	POUCHET, '66, p. 28
<i>Helix hortensis</i> . . . . .	(-14° to -18°)+	During 180 minutes	" "
<i>Helix aspera</i> . . . . .	(-14° to -18°)+	" 180 "	" "
<i>Planorbis</i> . . . . .	- 7°	" 300 " (A)	ROEDEL, '86, p. 212
<i>Limnæa</i> . . . . .	- 7°	" 180 " (A?)	" " 212
<i>Pulmonate embryos</i> . . . . .	0° to -1°	"Died upon freezing" (W and A)	" " 212
<i>Limax</i> . . . . .	-17°+	During 2 hours (A)	POUCHET, '66, p. 26
<i>Annelida.</i>			
<i>Aulostomum gulo</i> . . . . .	- 2°	During 12 to 15 hours (W)	ROEDEL, '86, p. 206
<i>Clepsine complanata</i> . . . . .	- 5°	During 90 minutes (W)	" " 213
"Leech" . . . . .	- 6°	" "some minutes" (W)	DOENHOFF, '72, p. 725
<i>Insecta.</i>			
<i>Apis mellifica</i> . . . . .	-1.5°	During 210 minutes	ROEDEL, '86, p. 212
<i>Apis mellifica</i> . . . . .	-1.5°	" a few minutes	DOENHOFF, '72, p. 724
<i>Formica rufa</i> . . . . .	-1.5°	" 180 minutes	ROEDEL, '86, p. 196
<i>Pelopæus (chrysalis)</i> . . . . .	-28°-	Out-of-doors, withstood this temperature	WYMAN, '56, p. 157
<i>Lema sp.</i> . . . . .	- 6°	During 30 minutes	ROEDEL, '86, p. 197
<i>Pæderus riparius</i> . . . . .	- 4°	" 45 "	" " 197
<i>Phytonomus sp.</i> . . . . .	-12°	" 90 "	" " 197
<i>Melolontha</i> . . . . .	-18°+	" 120 " (A)	POUCHET, '66, p. 26
<i>Melolontha (larva)</i> . . . . .	-15°+	" 180 " (A)	" " 26
<i>Cetonia</i> } . . . . .	-17°+	" 120 " (A)	" " 26
<i>Hydrophilus</i> }			
<i>Dytiscus</i> . . . . .	- 4°	" 60 " (A)	KOCHS, '90, p. 682
<i>Vanessa cardui, larva</i> . . . . .	-15°	" 600 "	ROEDEL, '86, p. 212
<i>Vanessa io, larva</i> . . . . .	-17°+	" 120 "	POUCHET, '66, p. 27
<i>Smerinthus populi</i> . . . . .	-10°	" 150 "	ROEDEL, '86, p. 212
<i>Ocneria dispar</i> . . . . .	- 4°	" 30 "	" " 212
<i>Culex pipiens, larva</i> . . . . .	- 4°	" 60 "	" " 212
<i>Musca</i> . . . . .	-6° to -10°	" 180 "	DOENHOFF, '72, p. 725
<i>Musca dom.</i> . . . . .	- 5°	" 20 "	ROEDEL, '86, p. 201
Various insects . . . . .	0°	" 2 to 30 minutes (on ice)	PLATEAU, '72, p. 98

SPECIES.	MINIMUM TEMPERATURE.	CONDITIONS OF EXPERIMENT.	AUTHORITY.
<i>Arachnida.</i>			
Phalangium opilio . .	— 9°	During 60 minutes	ROEDEL, '86, p. 201
Tegenaria domestica .	— 6°	“ 60 “	“ “ 201
Argyroneta aquatica .	— 4°	“ 180 “	“ “ 201
Hydrachna cruenta . .	— 4°	“ 30 “	“ “ 201
“ Spider ” . . . . .	— 2° to — 3°	“ 480 “	DOENHOFF, '72, p. 724
<i>Crustacea.</i>			
Cyclops quadricornus .	0°	“ 1 “ (W)	PLATEAU, '72, p. 300
Cyclops spirillum . . .	— 6°	“ 120 “ (W)	ROEDEL, '86, p. 201
Daphnia pulex . . . . .	0°	“ “ (W)	“ “ 201
Gammarus pulex . . . .	0°	“ 30 “ (W)	PLATEAU, '72, p. 300 ROEDEL, '86, p. 205
Asellus aquaticus . . .	0°	“ “ (W)	PLATEAU, '72, p. 299 ROEDEL, '86, p. 205
Astacns fluviatilis . .	— 11.5°	“ a day (A)	POUCHET, '66, p. 32
<i>Vertebrata.</i>			
Rana esculata . . . . .	— 4° to — 10°	“ 180 minutes (A)	“ “ 19

Summarizing these conditions, we find that the following organisms, even when thawed out carefully, are killed by subjecting to a temperature of between 0° and — 5° for 60 minutes: Amœba, swarm-spores, white blood corpuscles of Amphibia, planarians, pulmonate embryos, small leeches, certain entomostacans, and some insects and spiders. These organisms are all either soft bodied or of small size, and, excepting animals which, like the bees, live in protected situations, they do not winter over in our northern temperate countries. The following species, on the other hand, resist — 10° for at least 60 minutes: some snails (possibly), the beetles *Melolontha* (perhaps) and *Phytonomus*, the *Vanessa* larva, *Pelopæus* chrysalis, and the crayfish, — all protected by a thick covering, or of rather large size. The reason why large size and thick covering should increase resistance is not far to seek, — both conditions tend to prevent the rapid loss of heat, — to defend the body from freezing through and through.

Another cause of variation in resistance to cold is, doubtless, the amount of water in the protoplasm. I have already referred to this cause as explaining the fact that dry seeds and spores



can withstand almost any temperature.\* MÜLLER-THURGAU ('80) found that the "succulent labellum of *Phajus* freezes at  $-0.56^{\circ}$  C.; the succulent leaf of *Sempervivum*, at  $-0.7^{\circ}$ ; the potato tuber, at  $-1^{\circ}$ ; the leaf of *Tradescantia mexicana*, at  $-1.16^{\circ}$ ; the ivy leaf, at  $-1.5^{\circ}$ ; the leaves of *Pinus austriaca*, at  $-3.5^{\circ}$ ; young shoots of *Thujaopsis*, at  $-4^{\circ}$ ." (VINES.) In this series of plant tissues, we see that the more succulent the tissue, the higher its ultraminimum. Possibly the reason why spermatozoa have so low an ultramaximum, despite their small size, is on account of the denseness of their protoplasm.

Not all variations in ultraminimum temperature are, however, explicable upon the ground of difference in size, body-covering, or density of plasm. The interpretation of the difference in sensitiveness to cold of the honey bee (*Apis mellifica*) and the red ant (*Formica rufa*), between *Bombyx* on the one hand and *Smerinthus* and *Vanessa* on the other, must wait for further knowledge.

The question now arises, what is the cause of death in organisms and protoplasm which succumb to low temperatures? With the higher animals the immediate cause is doubtless in part asphyxia resulting from a stoppage in the flowing of the frozen blood plasma, and in part the destruction of the red blood corpuscles, as well as the white.† With the simpler organisms, like planarians, Protozoa, or *Tradescantia* hair-cells the case is different. An insight into the changes which produce death in such organisms may be gained from KÜHNÉ'S ('64, p. 101) description of the effect of a temperature of  $-14^{\circ}$  on *Tradescantia* hair-cells. The frozen hairs were placed in water and observed under the microscope. "The appearance."

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\* Striking cases are on record of the resistance of gemmules, or "animal spores," to cold. Thus, WELTNER ('93, p. 276) saw gemmules of *Spongilla fragilis* frozen in an aquarium from December 26 to January 24, from the end of January to February 5, from February 20 to March 6, and from March 12 to 24; the intervals being occupied by thawings. Yet these gemmules produced young sponges. In other cases, a certain amount of freezing favors the subsequent development of gemmules, e.g. those of fresh-water Bryozoa (BRÆM, '90, p. 83) and the eggs of the silk-worm (DUCLAUX, '71).

† POUCHET ('66, p. 18) found that when blood of the frog was let fall into a capsule at a temperature of  $-15^{\circ}$  few of the red corpuscles were uninjured. In most the nuclei had been cast out into the plasma.

says KÜHNE, "was very remarkable, for there was no trace of the protoplasmic network; but the violet cavity of the cell contained, in addition to the naked nucleus, a large number of separate round drops and lumps." In this case, the separate pieces eventually became active again, so that the protoplasm, though nearly killed, was not quite so.

The phenomena seen by KÜHNE so closely resemble those produced in the same kind of cells by the galvanic current and other strong irritants as to indicate that cold acts as an intense irritant. We cannot, however, conclude that cold acts in no other way. It is clear that the expansion of forming ice in the vacuoles of the protoplasm must seriously disturb the structure, and, since the whole matter has received little attention, it is possible that a molecular change of some sort takes place when there is much water in the freezing protoplasm. To summarize: Death by freezing results in the higher animals largely from asphyxia, and in the simpler organisms from excessive irritation, mechanical rupture, and, perhaps, other causes.

I shall now sum up this section on the effect of extremes of heat and cold. As the temperature is elevated above the optimum, molecular changes occur in the protoplasm leading to its contraction. The contraction becomes more violent as the temperature is still raised, until, finally, a new series of molecular changes occur by which the protoplasm begins to coagulate. At this point the protoplasm begins to lose its irritability. If this process has not proceeded far, the vital activities may, under favorable conditions, return (temporary heat-rigor). Beyond a certain point (death point) recovery is impossible. The death point varies with the species, but lies not far from the maximum natural temperature attained by the medium in which they live. On the other hand, as the temperature is diminished from the optimum, the chemical processes of metabolism decrease in vigor and come to a standstill at about the freezing point of water. Violent contractions accompany the cooling process, concomitantly with which the protoplasm breaks down. From this condition of temporary cold-rigor recovery is still possible; but a little below, at a point dependent upon the size of the body and the diathermous qualities of its

covering, the water of the body begins to freeze, and in that process, or the subsequent thawings, the protoplasm undergoes a (partly mechanical) change resulting in death. If the body, however, contains no water, freezing cannot kill it.

Thus the effect of high temperatures is principally chemical, involving the living plasma; that of low temperatures is principally mechanical, involving the water of the body. Both raising and lowering the temperature act also as irritants.\* Finally, the positions of the maximum and minimum stand in most intimate relation to the inorganic environment of the organism and have been molded to that environment.

#### § 4. ACCLIMATIZATION OF ORGANISMS TO EXTREME TEMPERATURES

The phenomena to be discussed in this section fall naturally into two subsections: (1) acclimatization to heat and (2) acclimatization to cold. They will be considered in that order.

1. **Acclimatization to Heat.**—Our study of the maximum temperature which organisms reared under ordinary circumstances can withstand, led us to the conclusion that few active organisms can resist a temperature of over 45°, and for whole groups like Cœlenterata, marine Mollusca, and Crustacea, and the fishes, 40° is a point of death. Yet, on the other hand, it has long been known that there are organisms living in certain hot springs in waters of considerably higher temperature. I shall now give in tabular form some cases which I have collected of organisms living at or above the normally lethal temperature of the species. †

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\* Whether *sudden* change of temperature has an especial effect upon the movement of protoplasm is a disputed question, which has been answered positively by DUTROCHET ('37, p. 777) and HOFMEISTER ('67, p. 53) for *Nitella*, and DE VRIES ('70, p. 394) for root hairs of *Hydrocharis*, but has since, as a result of careful experiments, been denied by VELTNER ('76, p. 214) for *Nitella* and other plant cells.

† It is desirable that accurate data concerning the temperature of organisms in hot springs should be made, and we have, in this country, unusually favorable conditions offered for this study, especially in Arkansas, California, and the Yellowstone National Park. It is to be hoped that persons who have had the proper training should, when contemplating a visit to hot springs, provide them-

TABLE XXI

LIST OF SPECIES FOUND IN HOT SPRINGS, WITH THE CONDITIONS UNDER WHICH THEY OCCUR

No.*	SPECIES.	TEMP. C.	LOCALITY AND CONDITION OF LIFE.	AUTHORITY.
1	Chroococcus	51° to 57°	Benton's Hot Springs, Cal.	WOOD, '74, p. 34
2	Nostocs or Protococcus	93°	Geysers, Lake Co., Cal.; not abundant at this temperature	BREWER, '66, p. 391, also WYMAN, '67, p. 155
3	Nostocs	51° to 57°	Benton's Hot Springs, Cal.	WOOD, '74, p. 34
4	Anabaena thermalis	57°	Dax, warm springs	SERRES, '80, pp.13-23
5	Leptothrix	44° to 54°	Carlsbad Springs	COHN, '62, p. 539
6	Oscillaria or "Confervæ"	54° to 68°	Yellowstone Nat. Park, U.S.A.	WEED, '89, p. 399
7	"	54.4°	Springs, Bernardino Sierra, Cal.	BLAKE, '53, p. 83
8	"	57°	Algeria, Constantine province, waters of Hammam-Meskhoutin	GERVAIS, '49, p. 12
9	"	57°	Hot Springs, Taupo, New Zealand	SPENCER, '83, p. 303
10	"	60° to 65°	Geysers, Lake Co., Cal., U.S.A.	BREWER, '66, p. 392
11	"	60° to 65°	Hot Springs, Ark., U.S.A. (Long)	JAMES, '23, II, p. 291
12	"	71°	Hot Springs at Baños Luzon, Philippines	DANA, '38-'42, p. 543
13	"	75.5°	Soorujkoona Hot Springs	HOOKEE, J. D. '55, I, p. 24
14	"	81° to 85°	Ischia	EHRENBERG, '59, p. 493.
15	"	98°	Iceland	FLOURENS, '46, p. 934
16	"	?	Outlet of Lake Furnas, Azores.	DYER, '74, p. 324

selves with a hand lens, bottles of alcohol for preserving organisms for further study, and an accurately calibrated thermometer. A source of error to be guarded against lies in the precise determination of the temperature of the water immediately surrounding the organism observed; for in some warm springs or their outlets the surface water is said to be much warmer than the deeper layers in which the organisms are found. Finally, if possible, it would be desirable to determine on the spot, experimentally, the maximum temperature which these organisms can withstand. For this determination some of the methods referred to on p. 220 should be used.

\* Notes on each of these cases will be found at the end of this chapter, pp. 263-267.

No.	SPECIES.	TEMP. C.	LOCALITY AND CONDITION OF LIFE.	AUTHORITY.
	Diatoms		Frequently associated with other algæ in hot springs	
17	<i>Physa acuta</i>	33° to 35°	Sources of Dax, St. Pierre, France	DUBALEN, '73, p. iv
18	<i>Paludina</i> sp.	50°	Thermal waters, Abano, Padua	DE BLAINVILLE, '24, p. 141
19	"Bivalve testaceous animal"	?	Hot Springs, Ark.	MITCHILL, '06, p. 306
20	Rotifera and Anguillulidæ	44° to 54°	Carlsbad Springs, Bohemia	COHN, '62, p. 539
21	Anguillulidæ	45°	Aix, springs	DE SAUSSURE, 1796, V, p. 13, § 1168
22	"	81°	Ischia, in hot springs	EHRENBERG, '59, p. 494
23	<i>Cypris balnearia</i>	45° to 50.5°	Hammam-Meskhontin	MONIEZ, '93, p. 140
24	<i>Stratiomys</i> larva	69°	In hot spring, Gunnison Co., Col.	GRIFFITH, '82, p. 599
25	"	?	In hot spring, Uinta Co., Wyo.	BRUNER, '95
26	"Water beetle"	44.4°	In warm spring, India; abundant	HOOKEE, J. D. '55, p. 24
27	"	?	Hot spring, Port Moller, Alaska	DALL, W. H. (personal letter)
28	Barbels	34°	?	BERT, '77, p. 169
29	Frogs	38°	"Baths of the Pise"	SPALLANZANI, 1787, Tom. I, p. 55

To summarize: Protista are stated to have been found in nature in water at temperatures far above 60° C. The most striking cases are of *Oscillaria* and "Conferva" from several localities, which resist nearly up to the boiling point of water. The closely allied *Nostocs* are, perhaps, next most abundant and resistant, reaching 93° (possibly *Protococcus*) in the California geysers. Metazoa are stated to live at temperatures far above 45°. Although some doubt has been cast on No. 22 by HOPPE-SEYLER'S inability to confirm EHRENBERG'S observations, the case seems established of *Cypris* thriving at 45° to 50.5°. Very extraordinary are the observations Nos. 24 and 25 on *Stratiomys* larvæ, which, however, are sadly in need of confirmation by competent observers. Leaving out of account, for the moment, the less well established cases, there still remains abundant evidence that organisms can live and thrive

in hot springs at a temperature near or above that which proves fatal to their close allies.

No one doubts that in all the cases cited above the individuals living in hot springs have been derived from ancestors which lived in water whose temperature rarely exceeded 40° C. The race has therefore become acclimatized, and the question arises: How has that acclimatization been effected?

Now experiments have shown that organisms, when gradually accustomed thereto, may resist a temperature which would have killed them if they had been suddenly subjected to it. Therefore it seems probable that the acclimatization of organisms to hot springs has been a slow, long-continued process, during which they have become gradually accustomed to higher and higher temperatures, probably attaining the hot springs by slowly advancing up their effluent streams.

This adaptation may have taken place without selection, purely by the capacity of individual adaptation which organisms possess. That individual adaptation is sufficient to account for the vitality of organisms in hot springs has been shown by experiment. DUTROCHET ('37, p. 777) observed, long ago, that an organism which at first seemed injured by a high temperature gradually regained activity while still subjected thereto.

Thus, he found that the current of *Nitella* was at first diminished by raising it to 27° C., but it soon became rapid again; raised, now, to 34°, the circulation began to fall off again, but in a quarter of an hour, the same temperature continuing, the circulation became very rapid. This phenomenon was repeated, also, at 40°. Similarly, HOFMEISTER ('67, p. 53) brought *Nitella flexilis* suddenly from + 18.5° to + 5° C. The streaming movements ceased. After staying 15 minutes in the cooler room, however, the rotation of protoplasm recovered.

Much more important, however, are the remarkable experiments of DALLINGER ('80). He kept *Flagellata* in a warm oven for many months. Beginning with a temperature of 15.6° C., he employed the first four months in raising the temperature 5.5°; this, however, was not necessary, since the rise to 21° can be made rapidly, but for success in higher temperatures it is best to proceed slowly from the beginning. When the temperature had been raised to 23°, the organisms began

dying, but soon ceased, and after two months, the temperature was raised half a degree more, and eventually to 25.5°. Here the organisms began to succumb again, and it was necessary repeatedly to lower the temperature slightly, and then to advance it to 25.5°, until, after several weeks, unfavorable appearances ceased. For eight months, the temperature could not be raised from this *stationary point* a quarter of a degree without unfavorable appearances. During several years, proceeding by slow stages, DALLINGER succeeded in rearing the organisms up to a temperature of 70° C., at which the experiment was ended by an accident.

In this case it is plain that the high temperature acted upon the same protoplasm at the end of the experiment as it did at the beginning. But while the protoplasm at the beginning of the experiment was killed at 23° C., at the end it withstood 70°. It will be seen that, by gradual elevation of the temperature, Flagellata may become acclimated to a temperature of water far above that which they can withstand when taken directly from out of doors, and approaching that of the hottest springs containing life.

A series of experiments, less extensive than that of DALLINGER, was carried on by Dr. CASTLE and myself ('95, pp. 236-240) upon the tadpoles of our common toad, *Bufo lentiginosus*. Recently laid eggs were divided into two lots: one lot was kept in a warm oven at a constant temperature of 24° to 25°, others at about 15° C. Both lots developed normally, but the former much the more rapidly. At the end of 4 weeks the point of heat-rigor was ascertained for each lot, by gradually heating (in from 5 to 10 minutes) the water containing them, until the tadpoles showed no response to stimulus, but, upon cooling, regained activity. The result was that the toad tadpoles had gained an increased capacity to heat. For when they were reared at a temperature of about 15° C., every tadpole went into heat-rigor at 41° C., or below; whereas, when they were reared at 24° to 25°, a temperature 10° higher, no tadpole died under 43°, the average increase of resistance being 3.2°. This increased capacity of resistance was not produced by the dying off of the less resistant individuals, for no deaths occurred in these experiments during

the gradual elevation of the temperatures in the cultures. The increased resistance was due, therefore, to a change in the protoplasm of the individuals.

The question now arose: In how far is this change in the protoplasm permanent? Will a return of the individuals to cool water cause a return to the old point of heat-rigor? We made a few experiments on this subject which showed that tadpoles which during 33 days in warm water have acquired an increased resistance of  $3.2^{\circ}$  lose part of that acquired resistance during 17 days' sojourn in cooler water. But the loss is a very slow one. The effect of the high temperature on the tadpoles is not, therefore, transitory, but persists—we have not been able to determine how long—after the cause has been removed.

So we may conclude: Individual organisms have the capacity of becoming adapted to a high degree of temperature, so that a temperature which normally is fatal may be withstood. This adaptation of the individual accompanies the subjection of organisms to temperatures higher than those to which they have already become accustomed. This capacity exists among both Protozoa and Metazoa. The effect of the elevated temperature persists (though in diminished degree) a considerable time after the individual has been restored to a lower temperature.

Acclimatization may show itself not only in the change of the maximum temperature, but also in the elevation of the optimum. This is shown by the following experiments of MENDELSSOHN ('95, p. 19). When *Paramecia* are placed in a trough whose temperature is  $24^{\circ}$  to  $28^{\circ}$  at one end and  $36^{\circ}$  to  $38^{\circ}$  at the other, they are found to collect at the cooler end, which indicates that the temperature of that end lies nearer their optimum. If, however, the *Paramecia*, while uniformly distributed in the trough, are subjected to a uniform temperature of  $36^{\circ}$  to  $38^{\circ}$  for from 4 to 6 hours, and then, in the same trough, to a temperature varying from  $24^{\circ}$  at one end to  $36^{\circ}$  at the other, they no longer collect at the usual optimum of  $24^{\circ}$  to  $28^{\circ}$ , but at  $30^{\circ}$  to  $36^{\circ}$ . Thus in 4 to 6 hours, by the action of a temperature of  $36^{\circ}$  to  $38^{\circ}$ , the optimum has been raised  $6^{\circ}$  to  $8^{\circ}$ .



Since experiments have proved the fact of acclimatization, it now remains to determine, if possible, its cause; to answer the question, by virtue of what property can organisms which, like Flagellata, normally perish at 45° C. come to live at 70° or even higher temperatures? We have seen that death at high temperatures is apparently due to coagulation of certain proteids in the protoplasm which undergo a chemical change at between 45° and 50° C. Now, although the matter has not yet been studied in these proteids, it has been shown for egg albumen that in proportion as it is dried its coagulation point rises, as the following table from LEWITH ('90) shows:—

EGG ALBUMEN.	COAGULATION TEMPERATURE.
In aqueous solution	56° C.
With 25 % water	74° to 80° C.
With 18 % water	80° to 90° C.
With 6 % water	145° C.
Without water	160° to 170° C.

Since the coagulation point of egg albumen is raised by dryness, it is very probable that a similar cause may act to raise the coagulation point of protoplasm in organisms of hot springs. Experimental studies are much needed upon this point. Meanwhile it can be said that *one* of the qualities which gives capacity of resistance to high temperatures is dryness. I shall now cite some cases that I have collected, which prove this point. It has been found that while moist yeast is killed at a temperature below 60°, dry yeast may be heated to 100° C. without losing its vitality (SCHÜTZENBERGER, '79, p. 162). Damp uredo-spores are killed at 58.5° to 60° C., but dry ones withstand up to 128° (HOFFMAN, '63); and dry spores of some molds up to 120° (PASTEUR, '61, p. 81). According to DALLINGER ('80, pp. 11-14), the dry spores of various Flagellata are capable of withstanding a temperature from 10° to 27° C. higher than that which these spores can resist in fluid. According to DOYÈRE ('42, p. 29), various animalecules (Rotifers, Tardigrades) which cannot in water withstand a temperature of 50° C. may, after long drying, be heated in air to 120° C.

(rarely to 125°) without all dying. The foregoing cases show clearly that increased resistance capacity is frequently gained by subjecting the protoplasm of the organism to dryness.

But there are other conditions under which the living substance shows extraordinary resistance capacity. In general, as is well known, the spores of organisms withstand higher temperatures than the motile stage, when both are in water. This rule holds for many cases: The spores of some bacteria may be heated for a time above 100° C. without killing them, although their motile stage is killed by 50° to 52° (LEWITH, '90). DALLINGER and DRYSDALE ('74, p. 101) and DALLINGER ('80, pp. 13, 14) have determined maximum temperatures for several Flagellata and their spores in water. While none in the motile stage could withstand a temperature higher than 61°, the spores in water withstood maximum temperatures varying between 65.5° and 131° for the different species.

Have the high resistance capacity of dry protoplasm and that of spores a common cause? Or, in other words, is the protoplasm of spores especially free from water? Many observations make it appear probable that this is so.

Thus in the case of bacteria, the protoplasm of the spore stage is optically denser and occupies less space than in the motile stage. (Cf. LEWITH, '90.)

In the case of the ciliate Infusoria, the larger size of the protoplasmic mass makes the comparison of the condition of the protoplasm in the two stages easier. We glean the facts from BÜTSCHLI ('89, pp. 1652-1654). As the process of encystment proceeds, the contractile vacuole continues to function, the intervals between its contractions gradually increase, and finally it disappears some time after the encystment is completed. Hand in hand with these changes goes a gradual condensation of the protoplasm. This condensation BÜTSCHLI believes to be due to an excretion of water from the protoplasm.

In *Actinosphaerium* the change from the richly vacuolated motile form to the encysted condition is even more marked. As BRAUER ('94, p. 193) has shown, the protoplasmic mass becomes, during the process of encystment, smaller and denser. The loss of water from the protoplasm is without doubt due to

the continued activity of the contractile vacuole at a time when no fluids are being taken into the protoplasmic body.

From the foregoing considerations it appears probable that one of the important characters of "spores" is the diminished amount of free water held in the protoplasm; or, in other words, its dryness. This dryness of the coagulable substance would seem to be cause of its higher resistance.

So far the evidence seems complete. Whether, however, loss of water is the ultimate cause of the high resistance capacity of hot-spring organisms or of those gradually acclimatized is still uncertain. Analogy renders it highly probable that such is the case.

2. **Acclimatization to Cold.**—Just as organisms may become acclimatized to high temperatures, so also may they live in very cold regions. I cite a few examples: Several species of Protista are said to live in the Alps above the snow line, coloring the snow red. (SHUTTLEWORTH, '40.) A tardigrade is found in the same locality. Certain insects live on or in the snow or ice. Thus *Desoria glacialis* (or glacier flea) lives on the Swiss glaciers, and on the snow live *Podura hiemalis*, *Trichocera brumalis* (when the temperature is "below the freezing point," FITCH, '46, p. 10), and other species of *Trichocera* and *Podura*. Cf. also *Boreus hiemalis* and *B. brumalis* (FITCH). Although swarm-spores are usually extremely sensitive to cold, STRASBURGER ('78, p. 613) cites a case of a marine alga in which they were being formed and thrown out when the temperature of the water was between  $-1.5^{\circ}$  and  $-1.8^{\circ}$  C.

Increased resistance to cold seems often the result of the action of cold on the organism. Thus, while SCHWARZ ('84, p. 69) found that *Euglenæ* gathered in the *summer* time were not responsive below  $+5^{\circ}$  to  $+6^{\circ}$  C., ADERHOLD ('88, p. 320) found that *Euglenæ* gathered in the *winter* would respond even at  $0^{\circ}$ . We may say, the winter cold had in some way lowered the heat attunement of these Protista.

In seeking for an explanation of acclimatization to cold we should recall that the cause of death from cold is chiefly the freezing of water in the protoplasm, and the irritation of excessive cold. Accustomed to great cold, protoplasm would doubtless be no longer irritated by it; whether under these circumstances

it would contain less water is a question which lacks an experimental answer.

The conclusion from the results offered in this section is this: protoplasm may become so modified through the action of excessive heat or cold that it is no longer killed at the ordinary fatal temperatures. This result is partly due to the fact that it is then not so strongly irritated by these extreme temperatures, and partly owing to the fact that the coagulation and freezing points have been shifted, possibly through loss of water.

#### § 5. DETERMINATION OF THE DIRECTION OF LOCOMOTION BY HEAT — THERMOTAXIS

Our knowledge of this subject is still in its infancy and depends chiefly upon the observations of STAHL ('84), VERWORN ('89, pp. 67-68), GRABER ('83 and '87), LOEB ('90, p. 43), DE WILDEMANN ('94), and MENDELSSOHN ('95). The first two and the last two mentioned have employed Protista, and we may consider their work first.

STAHL'S studies were made upon Myxomycetes. He used two beakers, of which one was filled with water at 7°; the other with water at 30°. These were placed near each other, and a strip of filter-paper, on which lay the plasmodium of *Æthelium septicum*, was stretched between them. The two ends of the strip with the corresponding ends of the plasmodium hung into the two glasses. The result was that the plasmodium moved from the colder water toward the warmer, although before the experiment it was moving in the opposite direction. WORTMANN ('85) added the observation that when the warmer temperature rose above 36° a repellent action of the warmer water was discernible.

VERWORN experimented chiefly with *Amœba*. The difficulty in this operation depended upon the necessity of warming only a part of the body of so small an animal. He used a glass plate of 5 sq. cm. area, to the upper surface of which was glued a piece of black paper, in which had been cut a rectangular opening, 3 sq. mm. large, and with very sharp edges. This plate was placed on the stage of the microscope so that the hole lay

in the rays of the infalling, concentrated light of midsummer, reflected from the mirror. Upon the black paper was placed the cover-glass with the amœba in a drop of water. The light from the mirror was cut off until the amœba, in its migrations, lay half-way over the edge of the orifice. Then concentrated light was let through the slit. A small part of the body was still moved across the line of demarcation; then for a moment movement ceased and a few seconds after the protoplasm of the amœba began to flow backwards. In from 10 to 30 seconds the amœba was wholly in the dark again. Similarly, when the cover-glass was moved so that the amœba was brought half-way over the open orifice it retreated into the dark. Direct measurement showed that the temperature at the illuminated part was 40° to 50° C., whilst over the black paper it was 15° to 20° less. That the movement was not due to the light was shown first by cutting out, by means of ice, the heat rays only. No reaction occurred. Secondly, by cutting out the light but not the heat, by passing the light through a solution of iodine in CS<sub>2</sub> so that only the ultra-red rays (which act like darkness to all organisms) went through; the typical reaction occurred when the temperature over the slit was 35°. From all of these experiments the conclusion seems justified — Amœba is positively thermotactic towards that temperature.

Similar results were obtained by VERWORN with the shelled Rhizopod *Echinopyxis aculeata*, and later (see JENSEN, '93, p. 440) with *Paramecium*. More complete studies on the latter were, however, made by MENDELSSOHN, who worked in VERWORN'S laboratory. MENDELSSOHN devised an excellent method of study. A brass plate 20 cm. × 6 cm. and 4 mm. thick is properly supported in a horizontal position, and to its under face are affixed, transversely, tubes through which hot or cold water may be run from a reservoir placed at a high level. In the middle of the plate a space 10 cm. × 2 cm. and 2 mm. deep is cut out and into it is fitted a glass or ebonite trough. Special thermometers whose bulbs are coiled in the plane of the trough, and hence perpendicularly to the stem, serve to measure the temperature of the water in the trough at any point. By means of water running through the transverse tubes either end of the trough may be heated or cooled as

desired. Starting now with the trough filled with infusion water, the *Paramecia* are seen to be uniformly distributed (Fig. 71, *a*). Hot water is run through tubes under the right end of the trough. After 10 minutes the thermometers show the temperature of the water at the right end to be  $38^{\circ}$ , at the left end  $26^{\circ}$ . At this moment, all *Paramecia* are in the left third of the trough (Fig. 72, *b*). If now the hot water be passed through the left tube only, the temperature rises at that end to  $36^{\circ}$  or  $38^{\circ}$ , falling to  $27^{\circ}$  or  $28^{\circ}$  at the other, and

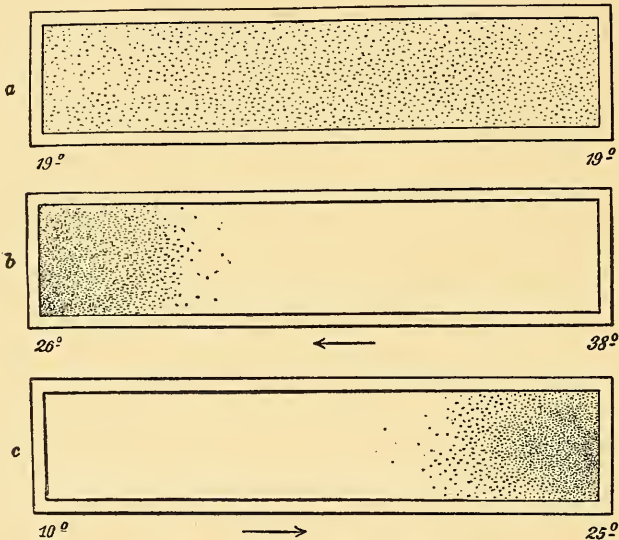


FIG. 71.—Distribution of *Paramecium* in a trough of water with variable temperature at the ends. (From MENDELSSOHN, '95.)

the *Paramecia* swim to the right end. Thus, with reference to a temperature of  $38^{\circ}$ , *Paramecia* are negatively thermotactic.

If, now, cold water be passed through the left tube so that the temperature of the left end of the trough falls to  $10^{\circ}$ , while the right end is at  $25^{\circ}$ , the *Paramecia* migrate to the right end. Towards a temperature of  $25^{\circ}$  *Paramecium* is thus positively thermotactic (Fig. 72, *c*).

Finally, if cold water be passed through the tube at one end of the trough and hot water at the other, the organisms will be found accumulated in the middle of the trough where the tem-

perature ranges from 24° to 28° C. This temperature is thus the optimum temperature for Paramecium, the temperature towards which it tends to move when the extremes are offered to it. Using another nomenclature, we may say, Paramecium is *attuned* to a temperature of 24° to 28° C.,\* and tends to keep in the temperature to which it is attuned.

Similar results have been obtained by DE WILDEMANN ('94) from Euglenæ which were kept in damp sand and in the dark, in a horizontal test-tube warmed at one end. Under these conditions they migrated towards the temperature of 30° rather than that of 15 to 22°.

Finally, we may consider thermotaxis as it is revealed in the higher animals. LOEB ('90, p. 43) enclosed the larvæ of the bombycid moth *Porthesia* in an opaque box, one end of which was next the stove. The animals moved to the warmer end of the box. The migration differed, however, from migrations with reference to light in that the body was not definitely oriented with reference to the source of heat, but the larvæ *wandered* thither. Similarly some ants (*Formica sanguinea*) are thermotactic according to WASMANN ('91, p. 22); and the cockroach (GRABER, '87, p. 254) moves towards that temperature which is more nearly normal for it. GRABER ('83, p. 230) has likewise shown that the salamander *Triton* is similarly responsive. Thus some Metazoa as well as Protista are clearly thermotactic.

Looking now for the cause of thermotaxis, we see at the outset that it is necessary to distinguish between two possibilities: a movement towards a greater or less intensity of heat, and a movement with reference to the direction of the heat rays in radiant heat. Now we have seen in earlier chapters, in considering the action of gravity, the electric current, and light, that these agents determine the direction of locomotion by determining the orientation of the axis of the body; and since radiant heat passes in lines, it might be possible to have a similar effect here. But there is no evidence that radiant heat acts here. In the case of the *Myxomycete*, it is clear that

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\* Adequate control experiments with dead Paramecia and fine suspended particles demonstrated that the movement was not purely passive; *i.e.* due to currents in the water.

a difference in the temperature of the two ends is sufficient to determine direction of locomotion. In the case of *Paramecium*, it is improbable that radiant heat acted, since this passes with difficulty through water. Finally, in the case of the insects enclosed in a box there is evidence of no axis orientation, and in the case of VERWORN'S experiments with the *Amœba* the action of direction is clearly shut out. We must therefore conclude that direction of locomotion in thermotaxis is not usually, if ever, determined by direction of heat rays.

Since it is not direction of heat rays, it is probably difference of intensity of the agent at the two poles of the organism which is the determining factor. This is clearly so in the case of the *Myxomycete* and *Amœba*. In the case of insects also, it is clear that one part of the body being appreciably nearer the source of heat would be appreciably warmer than the other, and this difference in temperature might serve as an indication to the organism of the direction of the source of heat. But when we come to consider MENDELSSOHN'S experiments on *Paramecium*, we pause to think of the organism being so sensitive as to be affected differently at the two poles by so slight a difference of intensity as these poles must experience. MENDELSSOHN has found that the least difference of intensity at the ends of his 10 cm. long trough which will call forth a thermotactic response is 3° C. The length of a *Paramecium* is about 0.2 to 0.25 mm., which corresponds to a difference of 0.01° C. of temperature at its two poles. This is the minimal temperature-difference which acts as a stimulus to *Paramecium* and calls forth a thermotactic response. Although this difference is small, we must, with MENDELSSOHN, in the absence of opposing data, consider it the determining factor in thermotaxis, and conclude that, in general, the thermotactic response is a response to differences in the intensity of heat to which the two poles of the body are subjected.

Let us now sum up the results of our study of the effect of heat on protoplasm. The rates of the metabolic processes and of protoplasmic movements are controlled by temperature, since they diminish from the optimum slowly towards the minimum and rapidly towards the maximum. At these points movement and irritability cease as a result of excessive stimulation, and



either the beginning of coagulation (maximum) or the cessation of chemical change (minimum) appears. Finally, death takes place at the ultramaximum through coagulation of the proteids, while it may occur beyond the minimum also, if the protoplasm contains much water. The optimum temperature is unlike in the different species, and certain individuals even may gain a very high or low optimum (lying even beyond the normal extremes) through the process of gradual acclimatization. The acclimatization seems to be due to the direct action of the varying environment upon the constitution of protoplasm. Finally, many simple organisms (probably all protoplasm) respond to heat by a locomotion which is adapted to keep them at the temperature to which they are attuned. The movement seems to be determined by the difference in intensity of heat at the different parts of the body. This whole chapter reveals protoplasm as a substance whose integrity is limited by chemico-physical conditions. Within those limits, however, it is highly sensitive to changes in temperature, becoming so altered by an untoward temperature as not to be injured by it, or migrating, if possible, so as to keep in the temperature to which it is already attuned. In a word, protoplasm shows itself to be a highly irritable, automatically adjustable substance.

## NOTES TO TABLE XXI (p. 250)

1. The *Chroococcus* was found in some of the fronds of *Nostocs* from Owen's Valley (see No. 3); but it is not stated whether they were in those *Nostocs* which were derived from the hottest springs.

2. No details are given by BREWER concerning the method of determining temperature. "In these warm mineral waters low forms of vegetation occur. The temperatures were carefully observed in many cases. The highest temperature noted, in which the plants were growing, was 93° C. (about 200° F.). But they were most abundant in waters of the temperature 52° to 60° (125° to 140° F.). In the hotter springs the plants appeared to be of the simplest kind, apparently, simple cells of a bright green color; but they were examined only with a pocket lens. In the water below, about 60° to 65° C., filamentous *Confervæ* formed considerable masses of a very bright green color." In a letter to WYMAN, however, BREWER says, concerning the same locality and determinations: "The temperatures given here were carefully observed with a standard Centigrade thermometer, with a naked elongated bulb," and "at the higher temperature [93° C.] they [the vegetable forms] were not abundant and existed as grains like *Nostoc* or *Protococcus*, intensely green and rather dark."

3. The temperature determinations, like the organisms, came from a Mrs. PARTZ, who is vouched for as reliable. No details concerning the method of obtaining temperatures are, however, given. The springs form a basin from which flows a creek. "In the basin," says Mrs. PARTZ, "are produced the first forms [Nostoc] partly at a temperature of 124° to 135° Fahr. Gradually in the creek, and to a distance of 100 yards from the spring, are developed, at a temperature of 110°-120° Fahr., the algæ," etc.

4. Not seen by me.

5. COHN states: "Thermometerbeobachtungen zeigten in verschiedener Temperatur des Wassers verschiedene, schon durch die Farbe erkennbare Arten; zwischen 43° und 35° R., die hellgrüne Leptothrix, zwischen 35° und 25° die Oscillarien, Mastichocladen, etc., gesellt mit Rädertieren, Infusorien und Wasserälchen; in noch abkühlterem Wasser die farblose *Hygrocrocis nivea*; Wasser über 44° enthält keine lebenden Organismen. Ganz dasselbe fand AGARDH 1827."

6. No statement as to the method of determining temperature. The measurements were made in the outlet to a hot spring. In this outlet *Hypeothyrix laminosa* flourished at 68° and occurred at even a higher temperature.

7. This account also leaves something to be desired as to definiteness: "Small springs rise at intervals of 10 to 20 feet along a distance of 30 to 40 rods. Their waters unite and form a little stream that empties into a brook a short distance below. . . . A dense mass of beautiful green confervæ grew about the bottom and sides of the channel, and floated in rich waving masses in the hot water. In the immediate vicinity of the springs, however, no vegetable growth appears. . . . The temperature of the hot stream, below all the springs, was found to be 130°."

8. GERVAIS' account is detailed, but the method employed in determining temperature is not given. The principal sections of interest are as follows: "Nous avons dit que l'eau au moment où elle s'échappe des sources avait donné à notre thermomètre + 95° cent." [It cooks eggs, meat, beans, etc.] "Il est inutile de dire qu'on ne trouve en cet endroit aucun animal ne aucun végétal aquatique vivant. Cependant on voit courir sur les cônes d'où jaillit l'eau bouillante, et en des points où le pied éprouve, même à travers la chaussure, un sentiment de vive chaleur, de petites Araignées qui m'ont paru être du genre *Lycose*. Quelques-unes s'aventurent même et cela sans inconvénient à travers la surface des petits cratères remplis d'eau chaude que présentent les cônes dont il s'agit. Dans la substance calcaire également fort chaude d'un de ces cônes que nous percions à coups de pioche pour en faire sortir l'eau bouillante par le flanc, nous avons trouvé plusieurs exemplaires vivants d'un petit Coléoptère de la famille des Hydrophiles, l'*Hydrobius orbicularis*, qui y avaient fixé leur demeure.

"L'eau à + 95° qui sort de différents points d'Hamnam-Meskhoutin perd assez rapidement cette température élevée. Elle n'a déjà plus que 57° dans les vases du second tiers de la cascade, dans lesquelles on commence à trouver des productions cryptogamiques. Celles-ci sont en partie couvertes d'un enduit ferrugineux assez épais."

9. Plants found in samples of water from Taupo, "growing in water the temperature of which varied from 105° F. to 131°." Two individuals are given as occurring at "temp. 136°"; two at "temp. 116°."

10. See note 2.

11. The reference reads: "Not only confervas and other vegetables grow in and about the hottest springs, but great numbers of little insects are constantly sporting about the bottom and sides." The temperature of the various springs runs from 92° to 151° F.

12. DANA says: "A species of feathery vegetation occurs also upon them [the stones of the brook], bordering the streamlets where the temperature is 160° F., and presenting various shades of green and white."

13. Data concerning temperature incomplete. "Confervæ abound in the warm stream from the springs, and two species, one ochreous brown, and the other green, occur on the margins of the tanks themselves, and in the hottest water; the brown is the best salamander, and forms a belt in deeper water than the green; both appear in luxuriant strata, wherever the temperature is cooled down to 168°, and as low as 90°."

14. This seems a carefully observed case. Hot water flowed from the clefts of the rock. "Die flache und schroffe Felswand worauf das heisse Wasser rieselnd und tropfend herabfloss war mit 2 fingerdicken hell und dunkelgrünen oder auch gelben, röthlichen und braunen Filzen überdeckt. In diese Filze an den Spalten eingesenkt zeigte das Thermometer 65 bis 68° R., entfernter von der Spalte schnell abnehmend weniger. Die organischen Filze waren so heiss, dass sie mit den Fingern nicht fassbar waren." Examined microscopically, the mass was found to consist partly of dead, partly of living "Eunotia forms" overgrown with *Oscillaria*. A similar condition was found also "in der Schlecht der Acqua della Rita bei eine Temperatur von 59° R. . . . Ich untersuchte in Serravalle aufgefangenes Wasser von 65° R. Wärme, welches ich in ein Glas laufen liess, während ich die felzige Masse drückte. Es war sehr voll von vielartigen lebenden kleinen Thieren. Darunter war 4 Arten munter bewegter Räderthiere, nämlich *Diglena Catellus*, *Conurus uncinatus*, die Abänderung des *Brachionus Pola* mit kleinen Stirnzähnen am Schilde, auch *Philodina erythro-phthalma*, ausgebildete Eier im Innern führend. Von Polygastern fanden sich in frischer Lebensthätigkeit eine noch unbekannte eigenthümliche, kleine Nasula, Formen von *Enchelys* und *Amphileptus* von weniger sich auszeichnender Gestaltung. Besonders auffallend war die lebende *Eunotia Sancta Antonii* der Capverdischer Inseln, deren Lebenszustand und Lebensbedingungen hierdurch zum erstenmale bekannt werden." Despite the evident care taken to obtain accurate results, EHRENBURG's observations have not been confirmed by HOPPE-SEYLER ('75, pp. 119, 120), who examined Ischia, but found no algæ living at a temperature much above 60°.

15. The entire reference is this: "M. FLOURENS met sous les yeux de l'Académie des *conferves* recueillies en Islande par M. DESCLOIZEAUX, qui les a trouvées végétant dans la source thermale de Gröf, à une température de 98 degrés" [of course, C.]. HOOKER ('13, p. 160) is often quoted as having obtained vegetation in Icelandic hot springs. Unfortunately, he gives no temperature determinations. He says: "Close to the edge of many of the hot springs [vicinity of the Great Geyser], and within a few inches of the boiling water, in places that are, consequently, always exposed to a considerable degree of heat, arising both from the water itself and the steam, I found *Conferva limosa* Dillw. in abundance." Again, "In water, also, of a very great degree of heat, were, both abundant and luxuriant, *Conferva flavescens* of Roth and a new species allied to *C. rivularis*."

16. The temperatures were not taken on the spot. MOSELEY says: "The water from which the Algæ were gathered was in the pools from which the *Chroococcus* was collected as far as I can now [*i.e.* many months after (?)] judge after testing water of successive temperatures with my finger, about 149°-158° F. The water of the sulphur-springs, in the area splashed by which the *Oscillatoria* are found, is quite scalding to the hand, and probably between 176° to 194° F."

17. The reference reads: "Dans celle [source chaude] de Saint Pierre, dont la température varie de 33 à 35 degrés, les *Physa acuta*, Drap. sont en nombre si considérable qu'elles forment un véritable fond mouvant dans les canaux." These hot water molluscs, as experiment showed, were killed at about 43°, while *Physa* from ordinary sources die at once at 35°. They perished after a few hours at 5° or 6°.

18. Merely the note: "On en [living mollusca] trouve aussi dans des eaux thermales: par exemple le *turbo thermalis*, espèce de paludine sans doute, vit dans celles d'Abano, dont la température est de 40° R."

19. The statement is not critical: "Their heat [hot springs] is too great for the hand to bear; the highest temperature is about 150°." "In the hot water of these springs a green plant vegetated, which seemed to be a species of *conferva* growing in such situations: probably the *fontenalis*. But what is more remarkable, a bivalve testaceous animal adhered to the plant, and lived in such a high temperature too."

20. See note 5.

21. "J'ai mesuré plusieurs fois & en diverses saisons, la chaleur de ces eaux, & je l'ai toujours trouvée à très-peu près la même; savoir, de 35 degrés dans celle du soufre, & de 36½ ou 36.7 [R. from context] dans celle de St. Paul. Malgré la chaleur de ces eaux, on trouve des animaux vivans dans les bassins qui les reçoivent; j'y ai reconnu des rotifères, des anguilles & d'autre animaux des infusions. J'y ai même découvert en 1790, deux nouvelles especes de tremelles douées d'un mouvement spontané."

22. See note No. 14.

23. Many individuals collected by R. BLANCHARD "dans les eaux de thermes du Hamman-Meskhoutine, près Guelma, dans les premiers jours d'avril; l'eau des thermes, au point de la récolte, a une température de 45° et de 50.5° C. Les Cypriis formaient une sorte de zone continue, de couleur chocolat, sur le bord de l'eau."

24. Found "in a hot spring, temperature 157° F., attached to the rock by the long end at about an angle of 45° and continually moving. . . . The rocks were covered with them."

25. The note in "Insect Life" is abstracted from a longer article by BRUNER in the newspaper called the Lincoln (Nebraska) "Evening Call," for April 6, 1895. The article, through the kindness of Professor H. B. WARD of the University of Nebraska, I have now before me. The larvæ were sent to Professor BRUNER by JOHN C. HAMM, of Evanston, Wyoming, upon whom this statement of the conditions of life of the organisms depends. The larvæ were found in a cup-shaped depression in the top of a small isolated cone about 20 inches high, situated about a few feet from a large sulphur mound or "dune," under which one could hear the rumbling of boiling water. Through apertures in the bottom the almost boiling water came up into the cup and ran over the edge of the pot.

The larvæ were actively moving. Mr. HAMM writes to Mr. BRUNER: "I did not have a thermometer with which to take the temperature, but . . . the water was so hot when I saw them that I could not hold my hand in it. My best judgment is and was at the time that the water was not more than twenty or thirty degrees [Fahr., of course] below the boiling point."

26. "A water beetle abounded in water at 112°" [F.].

27. Too hot for the hand to bear more than a moment or two.

28. The reference is to a discussion of a paper by BERT. "M. PAUL BERT a vu des barbellons dans de l'eau à 34° C."

29. "Mon ami Mr. COCCHI," says SPALLANZANI, "raconte que les Grenouilles ne souffrent point dans les bains de Pise, quoiqu'elles soient exposées à une chaleur indiquée par le 111° du Thermometre de FAHRENHEIT qui correspond au 37° du Thermometre de RÉAUMUR [! sic]."

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## CHAPTER IX

### GENERAL CONSIDERATIONS ON THE EFFECTS OF CHEMICAL AND PHYSICAL AGENTS UPON PROTOPLASM

IN the present chapter it is proposed to consider certain general matters upon which the facts given in this First Part throw light: namely, (I) the structure and composition of protoplasm; (II) the limiting conditions of metabolism; (III) the dependence of protoplasmic movement upon metabolism and external stimuli; and (IV) the determination of the direction of locomotion.

#### § 1. CONCLUSIONS ON THE STRUCTURE AND COMPOSITION OF PROTOPLASM

The question of the *structure* of protoplasm is preëminently a histological one. Microscopical study must eventually be relied upon to settle it. However, the results of experimental work seem to favor, as we have already pointed out (p. 70), BÜTSCHLI'S view of a honeycomb, or foam structure, of protoplasm.

The problem of the *constitution* of protoplasm is, on the other hand, preëminently a chemical one, and it must be solved by experimental methods. Our results can lead us to certain *qualitative* statements on this matter.

The chemical composition of protoplasm is immensely complex. Just as the geologist is forced by the facts to assume a vast, but not infinite, time for earth building, so the biologist has to recognize an almost unlimited complexity in the constitution of protoplasm.

The evidence that protoplasm is so complex is gained partly from the results of micro-chemistry. Many staining fluids act upon only a small part of the protoplasm of a single cell, so that a mixture of stains may be used, each component of which

attacks a different constituent of the protoplasm. In this way the dissimilar substances in protoplasm are made strikingly apparent. Not only does the protoplasm of one cell show this differentiation, but that of different cells of the body stains very diversely. Another line of evidence for the complexity of protoplasm is gained from the study of the effect of poisons. We have seen that the same poisonous substance acts very differently upon allied species of organisms (*e.g.* of bacteria) and upon the various organs of the body, — a fact which in many cases can only be accounted for on the ground of dissimilar composition. Again, most protoplasm must contain substances which are acted upon specifically by the different agents; for instance, certain highly explosive compounds which are set off by contact, certain others which are disturbed by light, and still others which are especially changed by heat. Each compound, again, must form an inconsiderable part of the whole, for (if the action be not too intense or prolonged) the “stimulus” of the agent results in no disturbance of the activities in general. Likewise, the facts of acclimatization, according to which, apparently, certain substances in the protoplasm may be destroyed without other important change in activities, give additional insight into protoplasmic complexity. Finally, the same agent acts in varying degree on closely related protoplasm, and this indicates that, even when the general composition is the same, the proportions of the different substances vary. From the facts of protoplasmic staining and of the varied effects of poisons, from the diverse effects of other stimulating agents, and from the facts of acclimatization of organisms, we conclude that in dealing with protoplasm we are not always dealing with the same thing, but, on the contrary, with very diverse combinations, which have this in common, that they exhibit life.

## § 2. THE LIMITING CONDITIONS OF METABOLISM

Metabolism is life. To know the limits within which it can occur is to know the vital limits. It is impossible to define these limits closely, however, for, at either extreme, metabolism graduates insensibly into inaction. It will be necessary, consequently, to place our limits very far out.

The limiting conditions at which inaction occurs are of two sorts. These may be termed respectively structural and dynamical. These two sorts of limiting conditions may be illustrated by comparing the protoplasmic mass to a factory, with many boilers and engines, much shafting and belting, and countless machines doing the most varied work. The amount of energy developed in the boilers and the efficiency of the engines and machines varies with certain conditions, such as the amount of heat applied to the former, and the friction and waste in the latter. The limiting mechanical conditions are reached when the boiler is rent by the steam pressure, a break-down is caused by friction, or a part rusts through and crumbles away. The limiting dynamical conditions are reached when the heat no longer suffices to form steam in the boiler, or the power is insufficient to run the machines. In either case, at the structural, or at the dynamical limit, work ceases. It may be the work of a small part of the factory, so that the cessation is hardly noticed; or it may involve all the machines, producing complete cessation of activity.

To return to the protoplasm: the *structural* limiting conditions are of two main sorts, — mechanical and chemical. The mechanical limiting conditions are those in which the gross structure becomes broken down, while the chemical limiting conditions are those in which the composition becomes changed. To the mechanical group belongs the breaking down of the plasma films, either by drawing out the water of the protoplasm (by osmosis or by drying) or by the expansion due to the freezing of the chylema. To the chemical group belong, for example, the reactions upon protoplasm of the halogen salts of the heavy metals, and of complex nitrogenous organic compounds in whose molecules hydrogen is unstably joined to nitrogen, also the coagulation of the plasma by high temperatures and the destruction of molecules by contact, by the electric current, or by light. The *dynamical* limiting conditions, on the other hand, are the absence of oxygen or other food-stuffs, the absence of the water necessary to the solution and circulation of the food, absence of light, in the case of chlorophyllaceous organisms, and a temperature much below 0° C. Thus, the conditions essential to metabolism are the

absence of causes mechanically rupturing the machine, the absence of agents of such intense activity as to change profoundly its molecular constitution, and the presence of those agents—food, heat, light, and water—which supply or distribute the energy of metabolism.

Given protoplasm under these conditions, and normal metabolism must occur; without them, there is no metabolism. Vary the dynamical conditions quantitatively, and a quantitative variation in metabolism will ensue. Approach a structural limiting condition, and metabolism begins to cease. This conclusion, important for experimental morphology, is now reached: *A vital phenomenon occurring in a given protoplasmic mass can be reproduced only when the dynamical conditions are reproduced, and the structural limiting conditions are in no wise closely approached.*

### § 3. THE DEPENDENCE OF PROTOPLASMIC MOVEMENT UPON METABOLISM AND UPON EXTERNAL STIMULI

I do not propose to enter the debated ground of the cause of protoplasmic motion; but shall merely summarize the results of our studies on this subject. First, protoplasmic movement is closely related to metabolism and is probably dependent upon it. This is indicated by the fact that cessation of movement always occurs before the vital limit is reached. Rigor always precedes death. A second series of facts indicating the same thing is found in the closeness with which the optimum for metabolism agrees with the optimum for movement. Thus at about 35° C. both the metabolic processes and the movements of protoplasm find their optimum. These two results, then, that movement is impossible in dead protoplasm even when its structure is seemingly unaltered, and the close approximation of the optimum points for metabolism and for movement, are the best justification for the belief that movement is dependent upon metabolism.

But are the conditions essential to metabolism the only conditions necessary for movement? In other words, will movement always accompany metabolism, or are external stimuli

essential to its production? There is in biology no question more important than this, and the answer is not so certain as it ought to be. The fact that rigor occurs at a point at which recovery of movement is still possible is not sufficient evidence that metabolism has not ceased with the motion; for I think it has not been shown that with rigor "latent life" does not come in. On the other hand, the fact that some bacteria are motionless in the absence of light (which can hardly be essential to metabolism) would seem to indicate that conditions other than those of metabolism are necessary to movement. This single fact cannot, however, lead us to a definite answer, and our inquiry, whether or not "stimuli" are essential to protoplasmic movement, must still be regarded as unanswered.

#### § 4. THE DETERMINATION OF THE DIRECTION OF LOCOMOTION

As we watch an animalcule swimming across the field of view, or as we see a larger organism moving, perhaps in a broken line, towards any point, we think of its movements as controlled from inside. Yet it is clear that if an organism is moving definitely towards a point, it must be on account of some influence emanating from that point and falling upon the organism. Without external directive influences of some sort there can be no directed movements.

This conclusion is confirmed by experiment. I have put an amœba into the apparatus already described (p. 186), so that the chemical conditions of its environment were uniform; contact and temperature were also similar on all sides; the directive action of gravity was annulled and all light was cut off. At intervals the position of the amœba was platted by the aid of light reflected momentarily from below the stage of the microscope and by means of a camera. Thus the path of the amœba was traced. A typical tracing made in this way is reproduced in Fig. 72. Compare the devious path made under these conditions with the straight path taken in response to light (Fig. 53). The curious spiral twists and the turning of the line upon itself are characteristic of all the tracings which I have made under these conditions. Important also is the



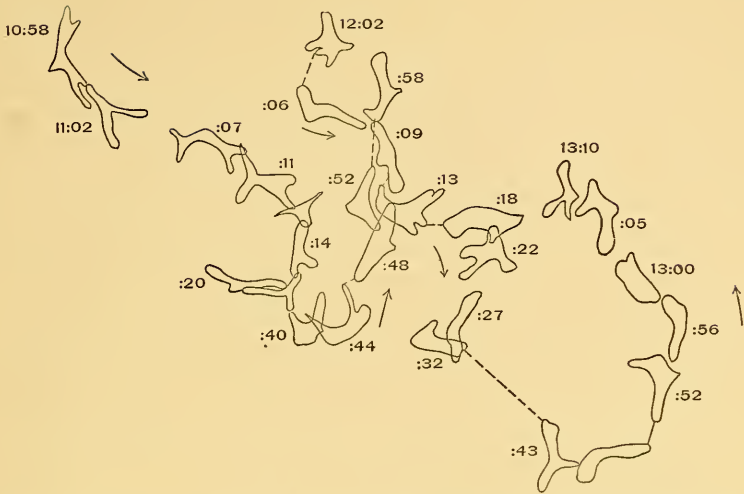


FIG. 72.—Camera drawing, showing the successive positions assumed by *Amœba proteus* when acted upon by external agents uniformly in all directions. Magnified 16 diams.



FIG. 73.—Tracks made on paper by larvæ of *Musca cesar* moving in the dark from a central spot of colored fluid. (From POUCHET, '72. *Revue et Mag. de Zool.* (2) XXIII.)

fact that whereas the amœba responding to light is constantly elongated in the direction of the infalling ray, the amœba which is not stimulated from one direction exhibits, most of the time, a stellate appearance. The wandering of the undirected organism is illustrated again in the experiment of Pouchet on the larvæ of *Musca* (*Lucilia*) *cæsar*, kept in the dark (Fig. 73; compare Fig. 74, where the same larvæ are migrating under the directive influence of light).

From these experiments we make the deduction that external agents play a role of the utmost importance for morphology, — of the utmost importance because by them alone is determined the *direction* of migration of the motile cells or the migrating



FIG. 74. — Tracing made like that of Fig. 73 by fly larvæ, when the light falls upon them in the direction of the arrow. *A*, the first direction of the light; *B*, the second direction. To be compared with the undirected movements of Fig. 73. (From POUCHET, '72.)

protoplasm of whatever sort in the organism. The *sense* of that migration depends in part upon the internal condition of the protoplasm. The *mechanism* by which locomotion is effected — that is wholly internal. The mechanism and the energy necessary to make it go are alone impotent to determine any adaptive movement or any other predictable result. To mechanism and energy must be added a stimulus external to the responding protoplasm in order that an adaptive or orderly result should occur.\*

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\* There are several other important matters upon which the results of this First Part throw light, such as the Mechanics of Response and the Origin of Adaptation in Response. Since additional facts for the discussion of these topics will be gained from the succeeding Parts, that discussion will be deferred.

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