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# NOAA TECHNICAL REPORTS

## National Marine Fisheries Service, Circulars

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387 Marine flora and fauna of the northeastern United States. Crustacea: Stomatopoda. By Raymond B. Manning. February 1974. iii + 6 p., 20 figs. For sale by the Superintendent of Documents, U.S. Government Printing Office, Washington, D.C. 20402.

NOAA Technical Report NMFS Circular 398



Marine Flora and Fauna of  
the Northeastern United States  
Higher Fungi: Ascomycetes,  
Deuteromycetes, and  
Basidiomycetes

A. R. Cavaliere

March 1977

U.S. DEPARTMENT OF COMMERCE

Juanita M. Kreps, Secretary

National Oceanic and Atmospheric Administration

Robert M. White, Administrator

National Marine Fisheries Service

Robert W. Schoning, Director

## FOREWORD

This issue of the "Circulars" is part of a subseries entitled "Marine Flora and Fauna of the Northeastern United States." This subseries will consist of original, illustrated, modern manuals on the identification, classification, and general biology of the estuarine and coastal marine plants and animals of the Northeastern United States. Manuals will be published at irregular intervals on as many taxa of the region as there are specialists available to collaborate in their preparation.

The manuals are an outgrowth of the widely used "Keys to Marine Invertebrates of the Woods Hole Region," edited by R. I. Smith, published in 1964, and produced under the auspices of the Systematics-Ecology Program, Marine Biological Laboratory, Woods Hole, Mass. Instead of revising the "Woods Hole Keys," the staff of the Systematics-Ecology Program decided to expand the geographic coverage and bathymetric range and produce the keys in an entirely new set of expanded publications.

The "Marine Flora and Fauna of the Northeastern United States" is being prepared in collaboration with systematic specialists in the United States and abroad. Each manual will be based primarily on recent and ongoing revisionary systematic research and a fresh examination of the plants and animals. Each major taxon, treated in a separate manual, will include an introduction, illustrated glossary, uniform originally illustrated keys, annotated check list with information when available on distribution, habitat, life history, and related biology, references to the major literature of the group, and a systematic index.

These manuals are intended for use by biology students, biologists, biological oceanographers, informed laymen, and others wishing to identify coastal organisms for this region. In many instances the manuals will serve as a guide to additional information about the species or the group.

Geographic coverage of the "Marine Flora and Fauna of the Northeastern United States" is planned to include organisms from the headwaters of estuaries seaward to approximately the 200 m depth on the continental shelf from Maine to Virginia, but may vary somewhat with each major taxon and the interests of collaborators. Whenever possible representative specimens dealt with in the manuals will be deposited in the reference collections of major museums.

After a sufficient number of manuals of related taxonomic groups have been published, the manuals will be revised, grouped, and issued as special volumes. These volumes will thus consist of compilations of individual manuals within phyla such as the Coelenterata, Arthropoda, and Mollusca, or of groups of phyla.

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# Marine Flora and Fauna of the Northeastern United States.

## Higher Fungi: Ascomycetes, Deuteromycetes, and Basidiomycetes

A. R. CAVALIERE<sup>1</sup>

### ABSTRACT

This manual provides an illustrated key and alphabetical listing, with brief descriptions, of common genera of higher marine fungi in the classes Ascomycetes, Deuteromycetes (Fungi Imperfecti), and a single member of the Basidiomycetes. A glossary and selected bibliography complement the key. Information on methods of harvesting, incubation, and studying these fungi is also included.

### INTRODUCTION

This manual is a guide to the genera of higher marine fungi that inhabit the intertidal zone of the Atlantic waters extending from North Carolina to Nova Scotia. Keys, descriptions, and illustrations are included for genera of Ascomycetes, Deuteromycetes (Fungi Imperfecti), and a single member of the Basidiomycetes. Less conspicuous saline forms of zoosporic fungi, the so-called marine Phycomycetes, as well as parasitic fungi, amoeboid forms, and those inhabiting the intestinal tract of arthropods, are not included in this account. In addition, species of higher fungi are excluded which have been reported only once or are rare or inconspicuous in the mycological flora.

Fungi inhabiting the saline environment appear to be cosmopolitan in distribution with only a few species having been shown to be endemic to one particular region. Some members of the marine mycoflora which are tropical, however, tend to be associated only with mangrove communities or other phanerogams which are restricted to warmer waters.

### METHODS OF HARVESTING AND STUDYING

Marine Ascomycetes, Basidiomycetes, and Fungi Imperfecti occur as saprophytes on driftwood, cordage, and other cellulosic material, or as weak parasites infesting dying species of marine phanerogams or algae. Various plant parts, grass culms, driftwood, and algae are best collected along the shore at low tide and kept submerged in a container of seawater until studied. Fungi may also be induced to grow on substrates introduced into the seawater. A  $\frac{1}{4}$ -inch hole is drilled through the center of small, 4 × 6 inch panels of various

kinds of wood. These are then attached, in linear fashion, to a knotted, polyethylene or nylon line and submerged at or below the low tide limit for a 2-4 mo period. When panels are harvested, they are scraped of all macroscopic fouling organisms and rinsed in seawater. They are then examined for fungal growth and/or incubated individually for an additional period of one to several months in sterile, dry, air-tight aquaria or other glass containers. Several methods of harvesting and incubating lignicolous fungi are described in the literature.

Ascocarps, dense mycelial growth as well as conidial clusters, are located with a dissecting scope utilizing a high intensity light source. Conidial heads may also be located by placing thin strips of substrate in a drop of seawater on a slide and observing the preparation directly under a compound microscope. Semipermanent mounts are made by placing fruiting structures into a drop of lactophenol (20 g phenol crystals; 20 g lactic acid; 40 g glycerol; 20 g distilled water; 0.01 g cotton blue or acid fuchsin) or Hoyer's medium (made by soaking 30 g of flake gum arabic in 50 ml of distilled water for 24 h, dissolving 200 g of chloral hydrate into the mixture, and then stirring in 20 ml of glycerol. Allow the mixture to settle before using.) When examining ascocarps or pycnidial structures, it is best to crush the fruiting bodies to expose the centrum. This is best accomplished by lightly tapping the cover slip with the handle end of a dissecting needle or the eraser end of a pencil. Spores of several of the marine species have gelatinous appendages which are best observed in a seawater mount under reduced light intensity or by phase contrast microscopy. Gelatinous appendages are deliquescent in most cases, short-lived, and, unfortunately, not retained satisfactorily in any known mounting medium.

Pure cultures of marine Ascomycetes and Fungi Imperfecti are initiated by introducing spores or centrum cells from several ascocarps onto low nutrient level, seawater agar media and incubating at room temperature. Kirk (1969) offers the most useful account

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of the isolation and culture of lignicolous marine fungi (0.1% glucose; 0.01% yeast extract; 1.8% agar in aged seawater adjusted to approximately 20‰; 0.03% U.S.P. streptomycin sulfate). White birch applicator sticks, balsa strips, filter paper, or toweling paper added to the culture tubes serve as an additional cellulosic substrate. Methods of preparing specimens for embedding, serial sectioning, and differential staining are outlined elsewhere (Cavaliere 1966, 1973).

Several additional works covering various aspects of the biology and taxonomy of marine Ascomycetes may be useful to the student (Barghoorn and Linder 1944; Johnson and Sparrow 1961; Cavaliere and Johnson 1966; Kohlmeyer and Kohlmeyer 1971).

## GLOSSARY

The use of mycological terminology has been reduced to a minimum. Several terms are defined as well as illustrated. More complete definitions are found in Snell and Dick (1957) and Ainsworth and Bisby (1971).

*Acuminate* Gradually narrowing to a point.  
*Amerospore* One-celled spore; spore without partitions.  
*Amorphous* Without definite shape or structure.  
*Anastomose* To form a network of interconnecting hyphae.  
*Antepenultimate* Refers to the third to the last cell in any row of cells.  
*Apiculate* Having one to many sharp points or denticles.  
*Appendage* A process of any kind; a structure which adheres.  
*Ascocarp* A fructification in Ascomycetes bearing asci and ascospores (see Figs. 1-3).  
*Ascospore* A spore, typical of the sexual stage or cycle in Ascomycetes, borne in an ascus.  
*Ascus* A reproductive cell in Ascomycetes; a structure, within which are produced, by meiosis, normally 4-8 haploid spores (see Figs. 6, 7).  
*Attenuate* Gradually narrowing or thinning.  
*Awl-shaped* Gradually tapering from the base to a sharp, flexible or semirigid point.  
*Bacilliform* Refers to spores which are rod-shaped (see Fig. 24).  
*Basidiocarp* A fructification in Basidiomycetes producing basidia and basidiospores.  
*Basidiospore* A sexual spore; produced by meiosis and borne on a basidium.  
*Basidium* A cell within which nuclei first undergo reduction division then pass onto extensions externally forming basidiospores.  
*Bitunicate* Refers to asci having two walls; double-layered (see Fig. 7).  
*Biturbinate* Refers to 2-celled spores having both ends conical and slightly curved.  
*Catenulate* Attached in chains.  
*Clavate* Club-shaped or thickened at the apex (see Fig. 13).  
*Cleistothecium* A more or less spherical covering en-

closing asci; a fruiting structure in the Ascomycetes (Plectomycetes) produced as a result of sexual reproduction and opening at maturity by a rupture (see Fig. 1).

*Concolorous* Refers to the pigmentation being the same color throughout.

*Conidiophore* A specialized hypha or cell bearing conidia.

*Conidium* Spores produced by the Deuteromycetes; a spore borne on or in a specialized hypha termed a conidiophore; asexually produced spores.

*Deciduous* Short-lived; falling away; not persistent.

*Deliquescent* Dissolving or liquifying.

*Denticulate* Having small teeth.

*Dictyospore* A spore with many transverse and longitudinal septations; a muriform spore (see Fig. 23).

*Didymospore* Two-celled spores; spores with a single partition.

*Echinulate* With minute spines.

*Ellipsoidal* Refers to spores having the shape of an ellipse; generally rounded at both ends and having curved sides (see Fig. 19).

*Elongate* Longer than broad, having parallel sides (see Fig. 18).

*Endogenous* Borne or developing within.

*Epispore* The outer layer of the spore wall.

*Erumpent* Breaking through the surface of the substrate; refers to the position of the fruiting body with relationship to the substrate.

*Eucarpic* Refers to a condition in which only part of the somatic thallus is converted in the formation of fructifications.

*Filiform* Refers to spores which are slender and thread-like (see Fig. 22).

*Fuscous* Drab, gray or smokey in color.

*Fusiform* Spindle-shaped or tapering at both ends (see Fig. 12).

*Fusoid* Somewhat fusiform.

*Globose* Refers to spores which are spherical (see Fig. 8).

*Guttule* Oily, spherical globule.

*Helicospore* A spiral or helicoid spore (see Fig. 20).

*Holocarpic* Refers to a condition in which the entire somatic thallus is converted into a fructification.

*Hyaline* Transparent or translucent, colorless.

*Inequilateral* Having unequal sides.

*Innate* Occurring below the surface of the substrate. Refers to the position of the fruiting body with relationship to the substrate.

*Intercalary* Borne or developed between the base and apex of hyphae.

*Involuted* Rolled or enrolled.

*Lenticular* Refers to spores shaped like a double convex lens (see Fig. 11).

*Muriform* Refers to spores having both transverse and longitudinal septations (see Fig. 23).

*Oblong* Longer than broad with approximately parallel sides (see Fig. 15).

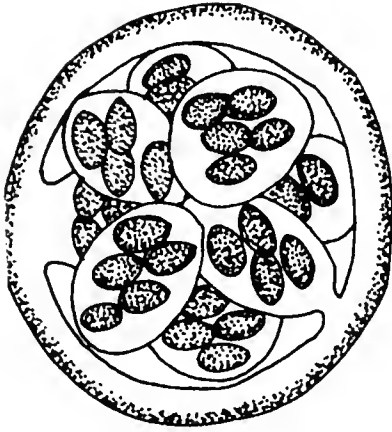
*Obpyriform* Reverse pear-shaped (see Fig. 17).

*Obtuse* Blunt or rounded, not pointed.

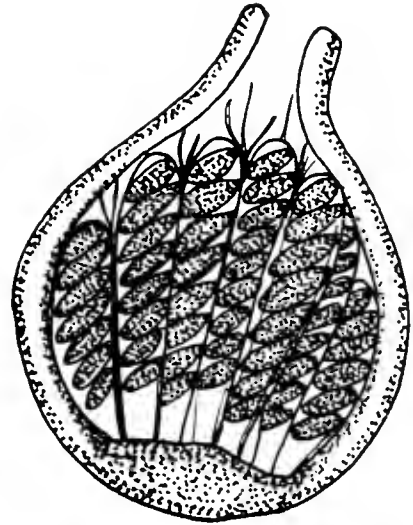


- Ovoid* Egg-shaped (see Fig. 10).
- Pedicellate* Borne on a pedicel or stalk.
- Penultimate* Refers to the next to the last cell in any row of cells.
- Perithecium* A more or less flask-shaped, papillate or beaked covering enclosing asci; a fruiting structure in the Ascomycetes (Pyrenomycetes) produced as a result of sexual reproduction (see Fig. 2).
- Peritrichous* Having flagellumlike hairs surrounding the structure.
- Phialide* Specialized conidiophore or cell within which conidia are produced and released (see Fig. 5).
- Phragmospore* A spore having two or more transverse septations.
- Pleomorphic* Having more than one form; polymorphic.
- Polymorphic* Occurring in several forms.
- Pseudothecium* A unilocular stroma. A cavity or locule formed by the dissolution of stromatic tissue (see Fig. 3).
- Pycnidium* In Deuteromycetes, a more or less flask-shaped structure bearing conidia on conidiophores internally (see Fig. 4).
- Pyriiform* Pear-shaped (see Fig. 16).
- Rhomboidal* Refers to spores having oblique angles and equal or unequal adjacent sides; more or less diamond-shaped.
- Scolecospore* A long, thread-shaped, filiform or vermicular spore (see Fig. 22).
- Septate* Having crosswalls or partitions.
- Sessile* Without a pedicel or stalk.
- Seta* Slender, bristle-shaped structure.
- Sheath* A covering or envelope.
- Staurospore* A spore which is more or less star-shaped (see Fig. 21).
- Stroma* A more or less tightly interwoven mass of hyphae within or on which reproductive structures are formed.
- Subglobose* Refers to spores or ascocarps which are nearly spherical; sides slightly flattened or compressed from the top (see Fig. 9).
- Truncate* Ending abruptly, squared off at the apex.
- Undulant* Wavy sheath or covering.
- Unitunicate* Refers to asci having a single wall (see Fig. 6).
- Verrucose* Covered with warts or marks.
- Verruculose* Covered with minute warts.

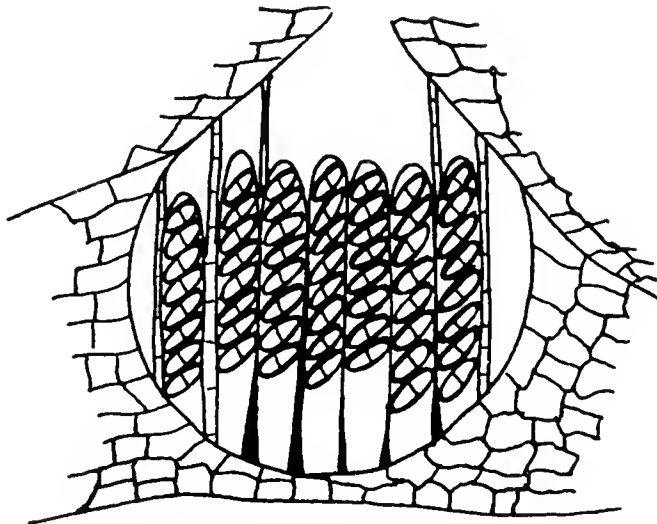
FIGURES OF REPRODUCTIVE STRUCTURES AND SPORE SHAPES



1. cleistothecium

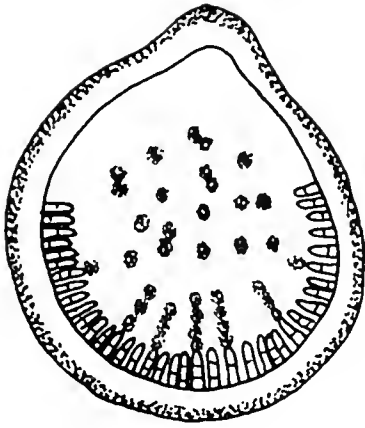


2. perithecium

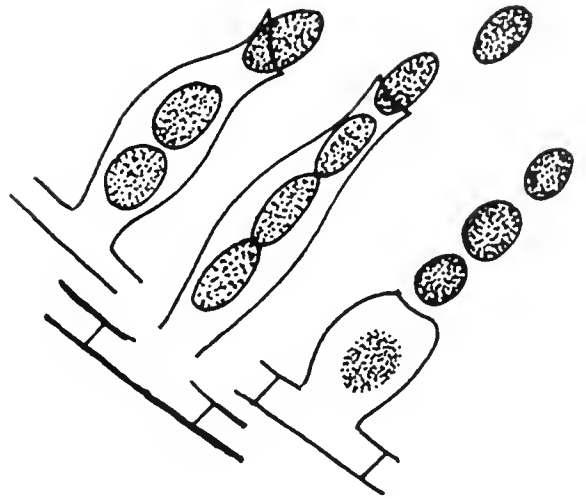


3. pseudothecium

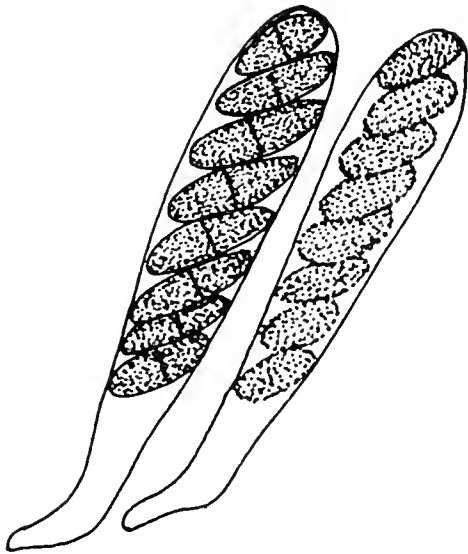
Figures 1-3.—Reproductive structures in Ascomycetes.



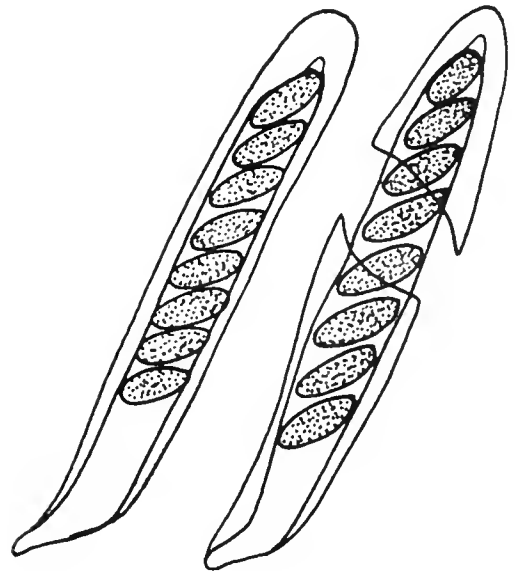
4. pycnidium



5. phialide

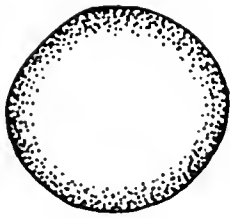


6. unitunicate asci

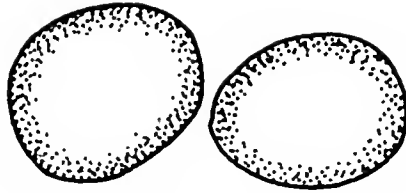


7. bitunicate asci

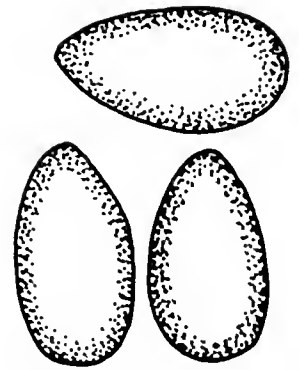
Figures 4-7.—Reproductive structures in Ascomycetes and Deuteromycetes.



8. globose



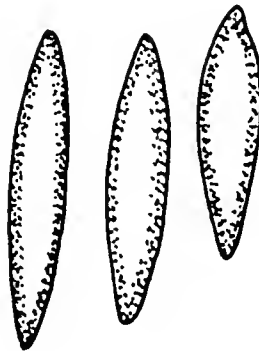
9. subglobose



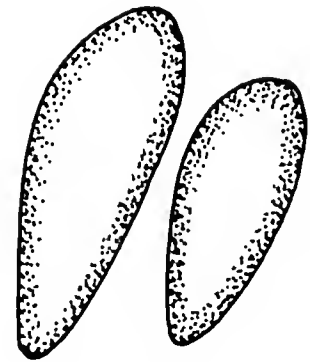
10. ovoid



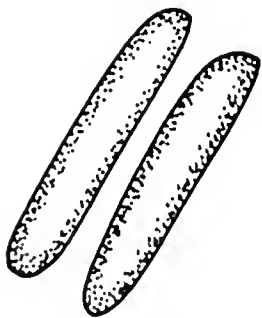
11. lenticular



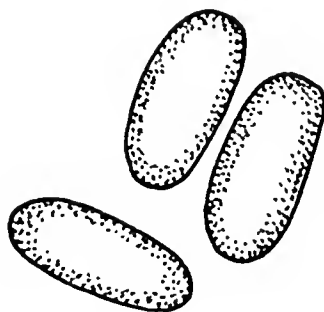
12. fusiform



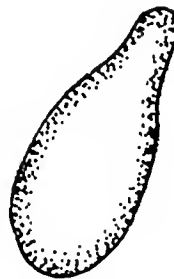
13. clavate



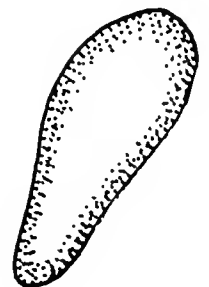
14. cylindrical



15. oblong

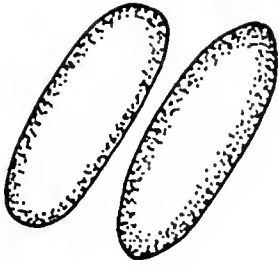


16. pyriform

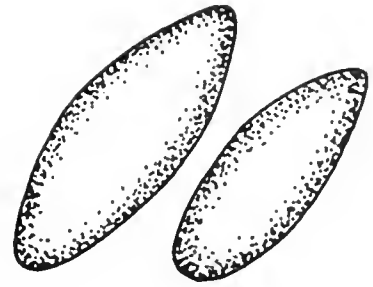


17. obpyriform

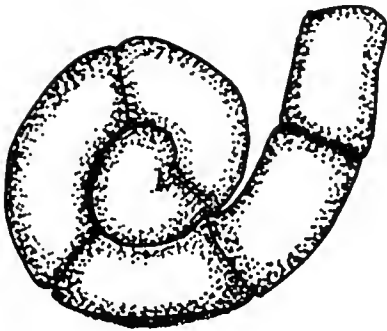
Figures 8-17.—Spore shapes in fungi.



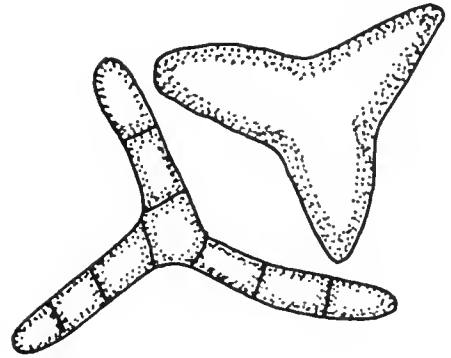
18. elongate



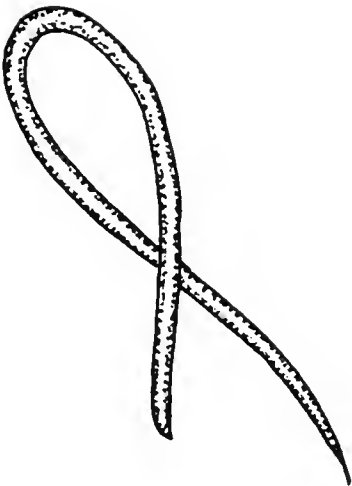
19. ellipsoidal



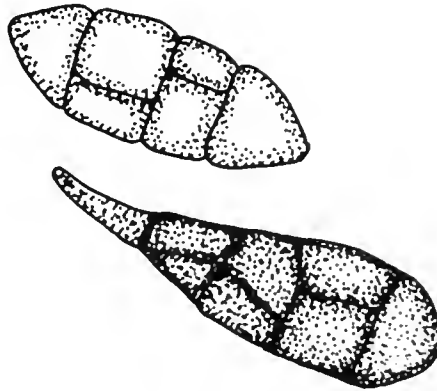
20. helicoid



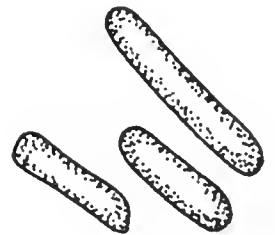
21. staurospore



22. filiform



23. muriform



24. bacilliform

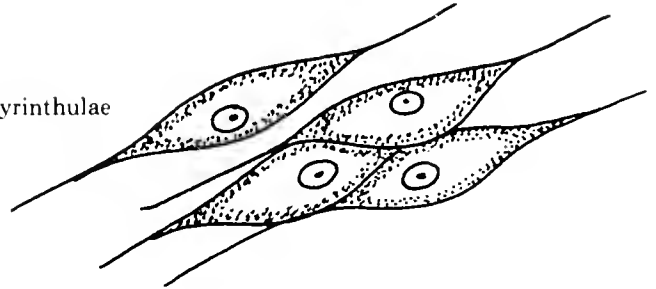
Figures 18-24.—Spore shapes in fungi.

# KEY TO MAJOR GROUPS OF FUNGI OCCURRING IN THE MARINE ENVIRONMENT

The preliminary key separates the nine major groups of fungi occurring in the marine environment: Labyrinthulaceae, Chytridiomycetes, Hyphochytridiomycetes, Plasmodiophoromycetes, Oomycetes, Trichomycetes, Deuteromycetes, Ascomycetes, and Basidiomycetes (*Nia vibrissa*, the only representative of the Basidiomycetes included in this treatment, is keyed out directly). Both the classes Ascomycetes and Deuteromycetes are separated into genera utilizing the Saccardo sporological system. Morphology and color of the spores

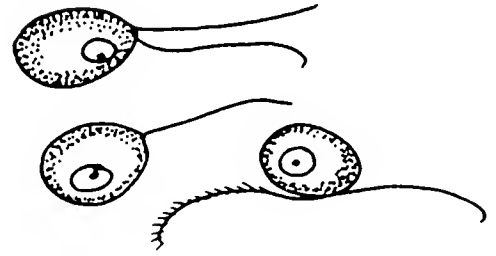
and conidia are the essential features employed in the Saccardo system. While the Deuteromycetes are keyed directly to genera or species, the Ascomycetes are divided into spore groups (Amerosporae, Dictyosporae, Didymosporae, Phragmosporae, Scolecosporae). Each spore group is then separated into the various genera. Genera of Ascomycetes are finally delimited into species based primarily on spore size and on the nature of spore appendages in those species possessing them.

1 Vegetative phase or stage entirely amoeboid . . . . . Labyrinthulaceae



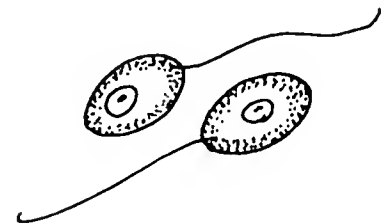
1 Vegetative phase or stage one- to many-celled, holocarpic or eucarpic . . . . . 2

2 (1) Flagellated cells present . . . . . 3

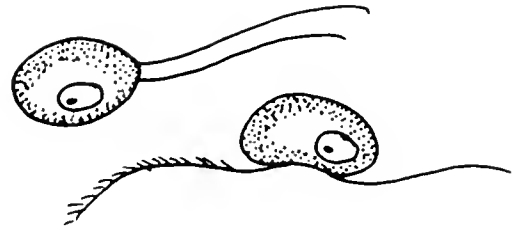


2 (1) Flagellated cells lacking . . . . . 6

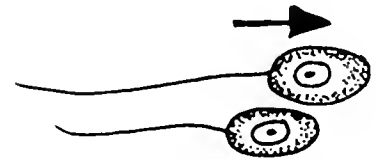
3 (2) Motile cells uniflagellate . . . . . 4



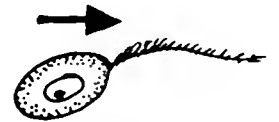
3 (2) Motile cells biflagellate . . . . . 5



4 (3) Flagellum of the whiplash type, posteriorly inserted . . .  
. . . . . Chytridiomycetes



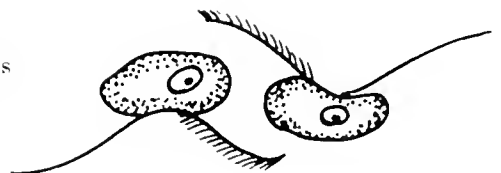
4 (3) Flagellum of the tinsel type, anteriorly inserted . . . . . Hyphochytridiomycetes



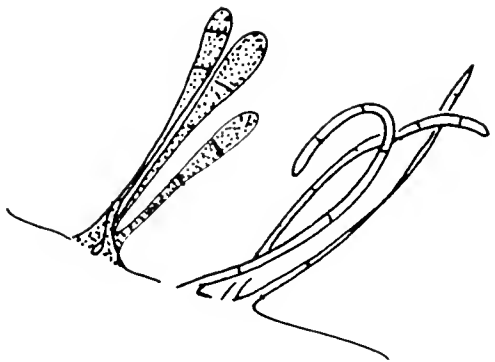
5 (3) Flagella heterokont, both of the whiplash type . . . . .Plasmodiophoromycetes



5 (3) Flagella nearly isokont, one of the whiplash type,  
the other tinsel . . . . . Oomycetes

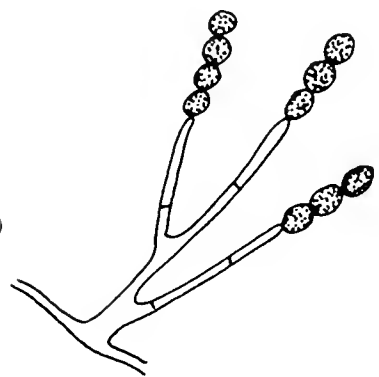


6 (2) Hyphae attached to the digestive tract or cuticle of arthropods . . . . . Trichomycetes



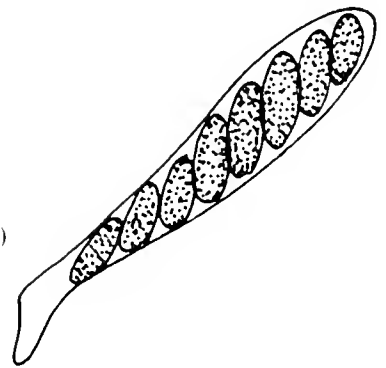
6 (2) Hyphae saprobic or parasitic on algae, phanerogams or other cellulosic materials . . . . . 7

7 (6) Sexually formed spores lacking; conidia usually formed in pycnidia (Fig. 4) or directly on vegetative hyphae . . . . Deuteromycetes (p. 32)



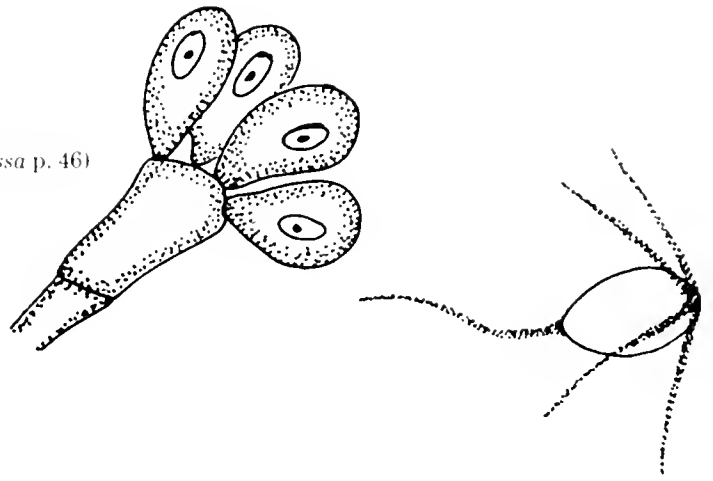
7 (6) Sexually formed spores present . . . . . 8

8 (7) Sexual spores, usually 8 in number, produced endogenously within asci . . . . . Ascomycetes (p. 11)



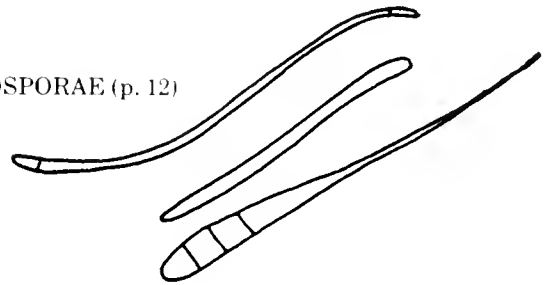


- 8 (7) Sexual spores, usually 2-4 in number,  
 exogenously produced on basidia . . .  
 . . . . . Basidiomycetes (*Nia vibrissa* p. 46)



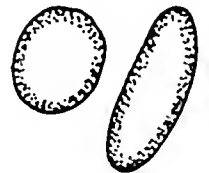
**KEY TO SPORE GROUPS OF ASCOMYCETES**

- 1 Ascospores filiform or slender; more than 10 times as  
 long as broad . . . . . SCOLECOSPORAE (p. 12)



- 1 Ascospores not filiform or slender; never more than 10 times as long as broad . . . . . 2

- 2 (1) Ascospores single-celled (nonseptate) . . . . . AMEROSPORAE (p. 14)



- 2 (1) Ascospores two- or more-celled (septate) . . . . . 3

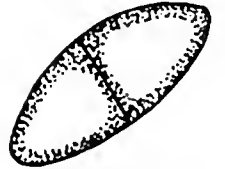


3 (2) Ascospores with transverse septa only . . . . . 4

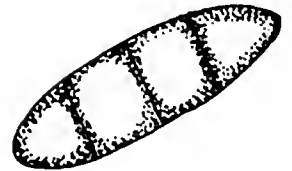
3 (2) Ascospores with transverse and longitudinal septations; muriform . . . . .  
DICTYOSPORAE (p. 15)



4 (3) Ascospores 2-celled (one septation only) . . . . . DIDYMOSPORAE (p. 16)



4 (3) Ascospores three- or more-celled (multiseptations) . . . . .  
PHRAGMOSPORAE (p. 27)

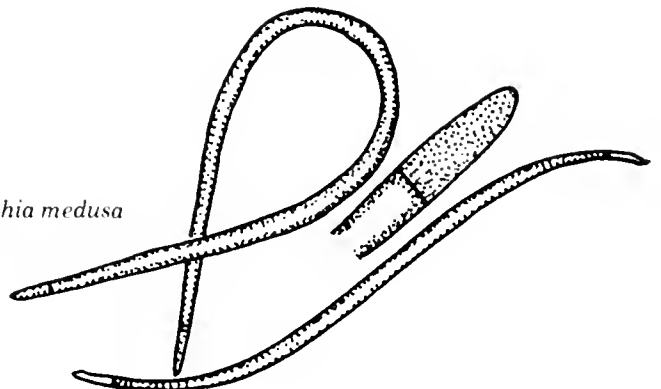


### KEY TO GENERA AND SPECIES OF SCOLECOSPORAE

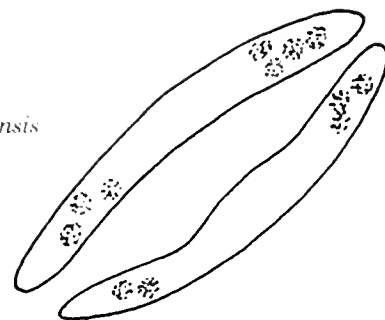
1 Ascospores single-celled (nonseptate) . . . . . 2

1 Ascospores three- to many-celled (multiseptate) . . . . . 3

2 (1) Ascospores 50-750  $\mu$ ; provided with a conoid cell or appendage at each end . . . . . *Lulworthia medusa*



- 2 (1) Ascospores usually shorter than  $30\mu$ ; without appendages . . . (see also Amerosporae) . . . . . *Halonectria milfordensis*



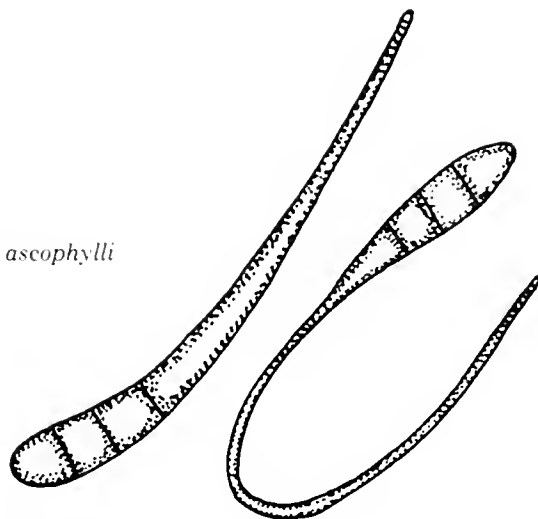
- 3 (1) Ascospores with a caplike, bulbous or threadlike appendage at each end . . . . . 4

- 3 (1) Ascospores without appendages . . . . . 5

- 4 (3) Appendages bulbous, subspherical . . . . . *Lindra* (p. 14)

- 4 (3) Appendages caplike or threadlike . . . (see also Phragmosporae) . . . . . *Haligena* (p. 29)

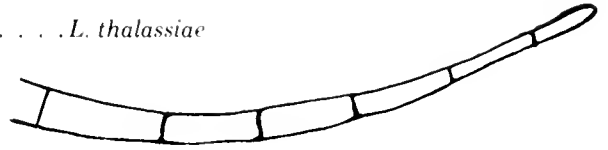
- 5 (3) Ascospores to  $100\mu$  long, 3- to 4-celled, tapering at one end . . . . . *Trailia ascophylli*



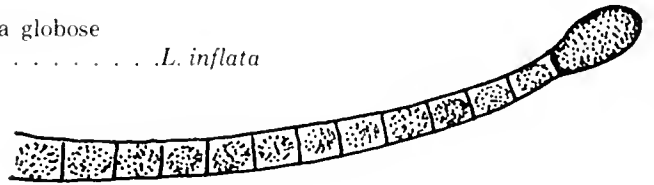
- 5 (3) Ascospores to  $400\mu$  long, filiform, cylindrical or tapering at both ends . . . . . *Lindra* (p. 14)

## Key to Species of *Lindra*

- 1 Ascospores 15- to 19-celled, tips slightly inflated . . . . . *L. thalassiae*

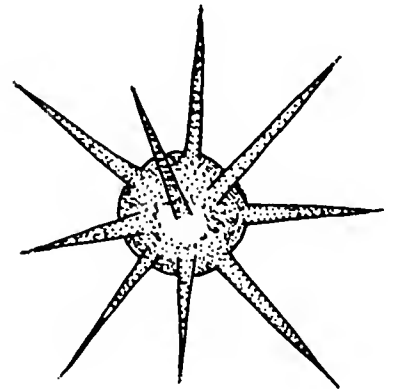


- 1 Ascospores 28- to 52-celled, tips provided with a globose appendage . . . . . *L. inflata*



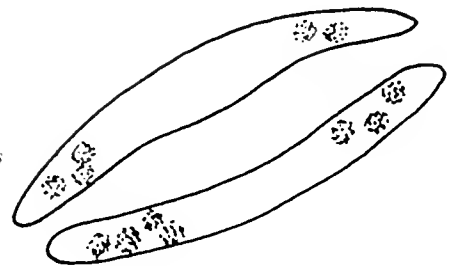
## KEY TO GENERA AND SPECIES OF AMEROSPORAE

- 1 Ascospores with several rigid, awl-shaped appendages arising from several positions along the spore wall . . . . . *Amylocarpus encephaloides*



- 1 Ascospores not provided with appendages . . . . . 2

- 2 (1) Ascospores many times longer than broad, fusiform or cylindrical, straight or curved . . . (see also Scolecosporae . . . . . *Halonectria milfordensis*



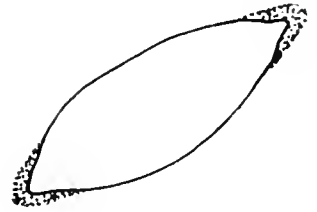
2 (1) Ascospores more or less ellipsoidal, not many times longer than broad . . . . . *Haloguignardia* (p. 15)

**Key to Species of *Haloguignardia***

1 Ascospores 30  $\mu$  or shorter, ends rounded . . . . . *H. oceanica*

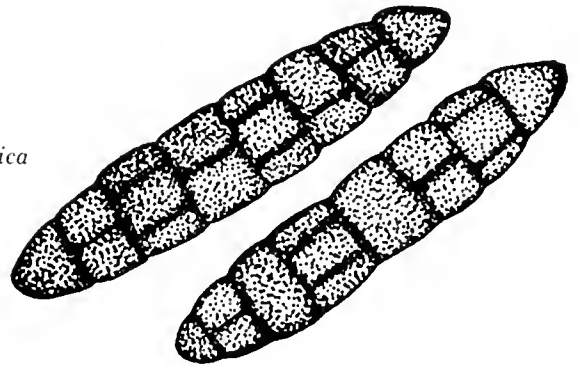


1 Ascospores 30  $\mu$  or longer, ends tapered . . . . . *H. tumefaciens*



**KEY TO GENERA AND SPECIES OF DICTYOSPORAE (*PLEOSPORA*)**

1 Ascospores more than 35  $\mu$  long, 7-9 transverse septa . . . . . *Pleospora pelagica*



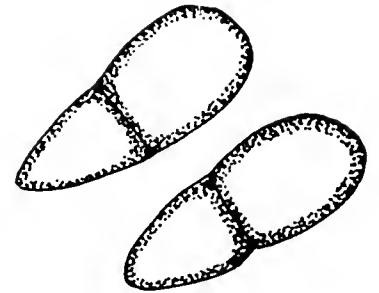
1 Ascospores less than 35  $\mu$  long, 3-7 transverse septa . . . . . *Pleospora pelvetiae*



## KEY TO GENERA AND SPECIES OF DIDYMOSPORAE

- 1      Ascospores surrounded by a gelatinous sheath and/or provided with appendages . . . . . 11
- 1      Ascospores with neither a gelatinous sheath nor appendages of any kind . . . . . 2
- 2 (1) Ascospores hyaline . . . . . 3
- 2 (1) Ascospores yellow, yellow-brown, brown, or dark brown . . . . . 6
- 3 (2) Ascocarp a pseudothecium containing bitunicate asci . . . . . 4
- 3 (2) Ascocarp a perithecium containing unitunicate asci . . . . . 5

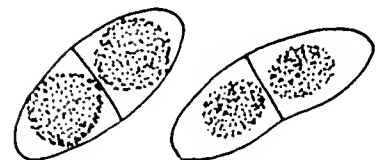
4 (3) Ascospores ellipsoidal, inequilateral . . . . . *Didymella fucicola*



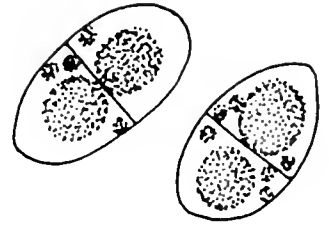
4 (3) Ascospores fusiform to oblong, equilateral . . . . .  
 . . . . . *Mycosphaerella ascophylli*



5 (3) Ascospores fusiform to ellipsoidal, never more than 10  $\mu$   
 in diameter . . . . . *Lignicola laevis*



5 (3) Ascospores broadly ellipsoidal or ovoid; 10-15  $\mu$  in diameter . . . . . *Nais inornata*



6 (2) Ascospores exclusively 2-celled . . . . . 7

6 (2) Ascospores 2- to 4-celled . . . (see also Phragmosporae) . . . . . *Leptosphaeria* (p. 29)

7 (6) Ascospore wall verruculose . . . . . *Didymosphaeria* (p. 23)

7 (6) Ascospore wall smooth . . . . . 8

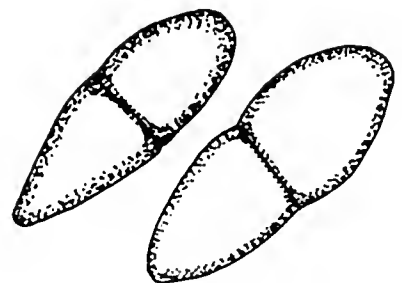
8 (7) Ascospores predominantly larger than 33  $\mu$  . . . . . 9

8 (7) Ascospores usually smaller than 33  $\mu$  . . . . . 10

9 (8) Ascospore cells unequal in size and shape; lower half cylindrical; upper half pointed . . . . . *Didymosphaeria* (p. 23)

9 (8) Ascospore cells equal in size and shape, straight or slightly curved . . . . . *Leptosphaeria* (p. 29)

10 (8) Ascospore 2-celled, usually unequal in size, occasionally biturbinate . . . . . *Microthelia linderi*

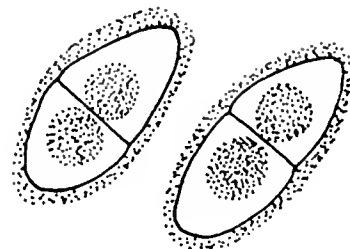


10 (8) Ascospore more than 2-celled, equal in size . . . . . *Leptosphaeria* (p. 29)

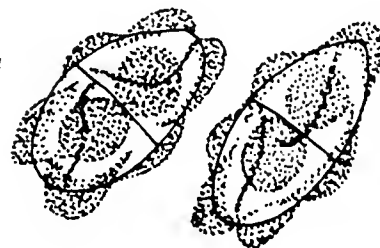
11 (I) Ascospores surrounded by a thin or thick gelatinous sheath but without protruding appendages of any kind . . . . . 12

11 (I) Ascospores may lack a gelatinous sheath but possessing apical and/or equatorial appendages . . . . . 13

12 (II) Ascospores to 15  $\mu$  long, pigmented, surrounded by a thin gelatinous sheath . . . . . *Massariella maritima*



12 (II) Ascospores to 35  $\mu$  long, hyaline, surrounded by an undulant, gelatinous sheath . . . . . *Didymosamarospora euryhalina*



13 (II) Ascospores provided with appendages or processes in both apical and equatorial positions . . . . . 23

13 (II) Ascospores provided with appendages or processes at either one or both apices but without such structures around the equator or midseptum . . . . . 14

14 (I3) Spore appendages usually at one end only . . . . . 18

14 (I3) Spore appendages always occurring at both ends . . . . . 15

15 (I4) Spores with a single appendage at each end . . . . . 20

15 (I4) Spores with 2-7 appendages at each end . . . . . 16

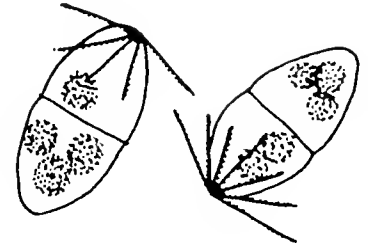
16 (I5) Ascospores with 2 (or a pair of) appendages at each end . . . . . *Halosphaeria* (p. 23)

16 (I5) Ascospores with 3-7 appendages at each end . . . . . 17



- 17 (16) Appendages usually 3 at each end (sometimes 4-7), terminal or subterminal, stiff, thin, tapering, sometimes curved, to 40  $\mu$  long . . . . . *Corollospora* (p. 22)
- 17 (16) Appendages 4-6 at each end, bulbous at base, radiating, attenuate, semirigid, mostly to 30  $\mu$  long . . . . . *Halosphaeria* (p. 23)

- 18 (14) Appendages radiating, stiff or bristlelike . . . . .  
 . . . . . *Keissleriella blepharosporea*

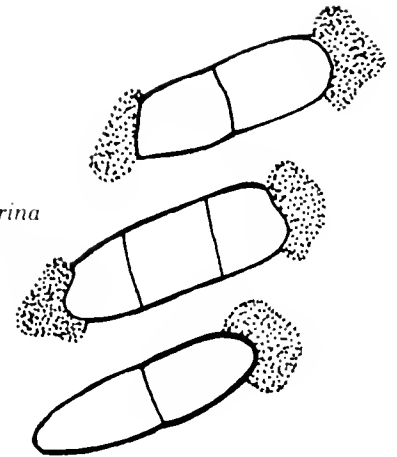


- 18 (14) Appendages gelatinous, short, caplike, subspherical or long . . . . . 19

- 19 (18) Spores cylindrical, to 70  $\mu$  long . . . . . *Halosphaeria* (p. 23)

- 19 (18) Spores ellipsoidal, thick-walled, to 35  $\mu$  long . . . (see also Phragmosporae) . . . . . *Lentescospora submarina*

- 20 (15) Ascospores with a short, gelatinous appendage at each end (see also Phragmosporae) . . . . . *Lentescospora submarina*

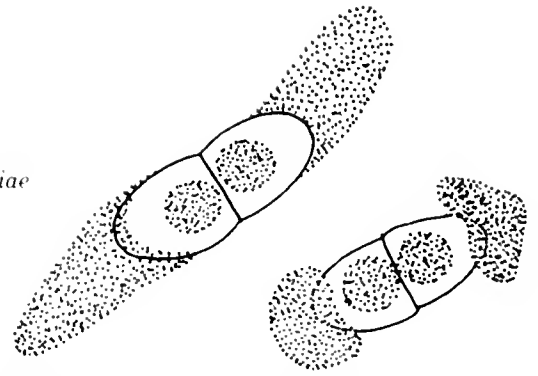


- 20 (15) Ascospore appendages variously shaped . . . . . 21

- 21 (20) Ascospores with a gelatinous sheath; appendages tapering or filiform . . . . . *Ceriosporopsis* (p. 21)

- 21 (20) Ascospores without a gelatinous sheath; appendages subspherical or caplike, blunt or attenuate, stiff or hamate . . . . . 22

- 22 (21) Ascospores ellipsoidal, never elongate, under 30  $\mu$  in length; with a caplike or globose, thick, polymorphic appendage at each end; ascocarp stromatic . . . . . *Phycomelaina laminariae*

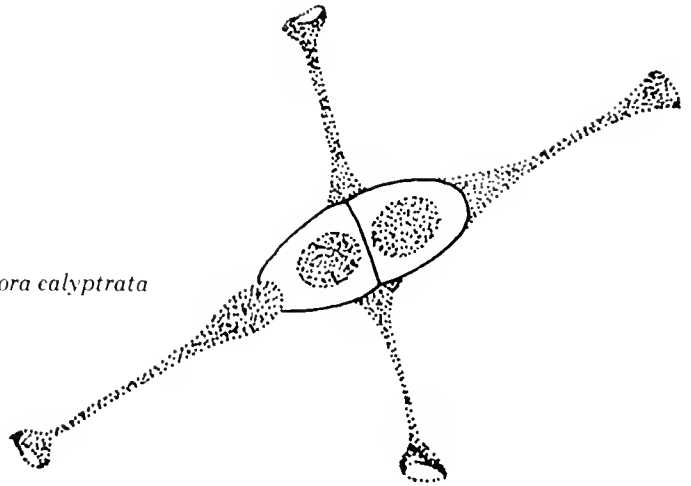


- 22 (21) Ascospores ellipsoidal to elongate, to 70  $\mu$  in length; with a subspherical, tapering or blunt, straight or curved appendage at each end; ascocarp nonstromatic . . . . . *Halosphaeria* (p. 23)

- 23 (13) Equatorial appendages setae or flagellalike, arising peritrichously . . . . . *Corollospora* (p. 22)

- 23 (13) Equatorial appendages not flagellalike, not arising peritrichously . . . . . 24

- 24 (23) Equatorial and apical appendages provided with a small involuted cap or tip . . . . . *Marinospora calyptrata*



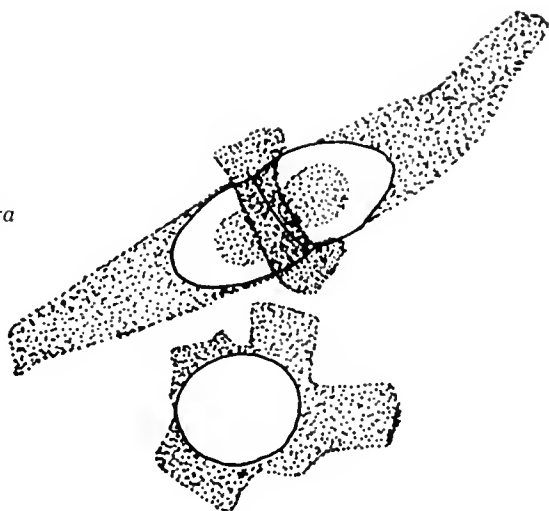
- 24 (23) Equatorial and apical appendages not provided with small involuted caps or tips . . . . . 25

- 25 (24) Ascospores totally surrounded by a gelatinous sheath; apical appendages broad, cylindrical or hollow . . . . . *Ceriosporopsis* (p. 21)

- 25 (24) Ascospores not surrounded by a gelatinous sheath; apical appendages thin, tapering, attenuate or blunt and subspherical . . . . . *Halosphaeria* (p. 23)

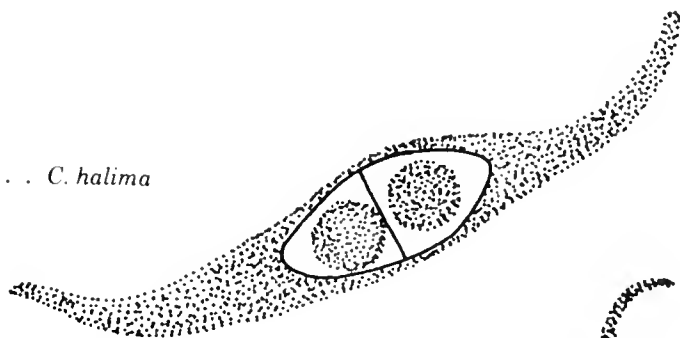
## Key to Species of *Ceriosporopsis*

- 1 Ascospores provided with both a tapering, gelatinous appendage at each end, and a thick, collarlike or several wedge-shaped processes around the septum . . . . . *C. tubulifera*

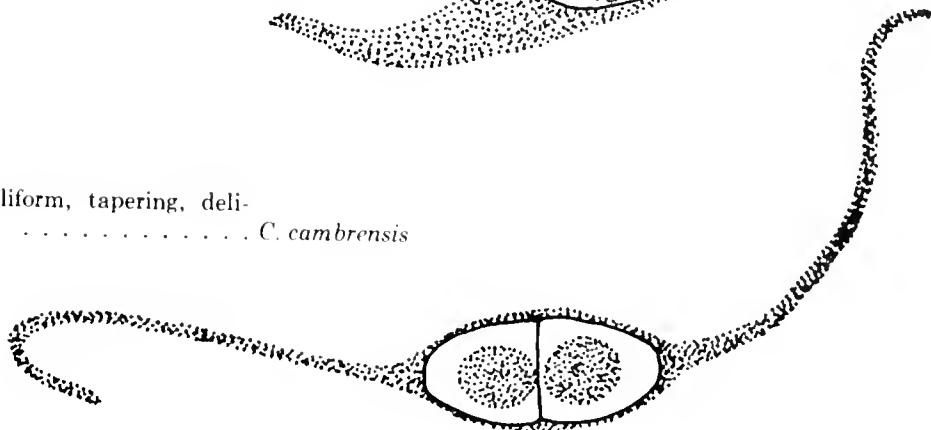


- 1 Ascospores provided with only a tapering, gelatinous appendage at each end . . . . . 2

- 2 (1) Appendages broadly filiform, tapering, to 8  $\mu$  thick at base . . . . . *C. halima*



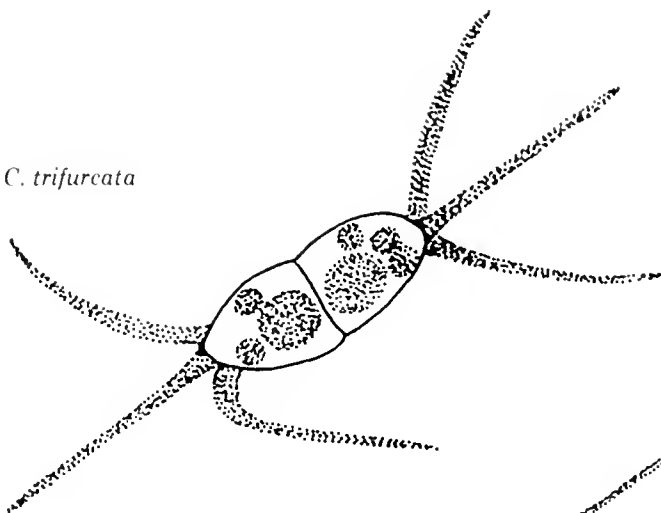
- 2 (1) Appendages thin, filiform, tapering, deliquescent, to 45  $\mu$  long . . . . . *C. cambrensis*



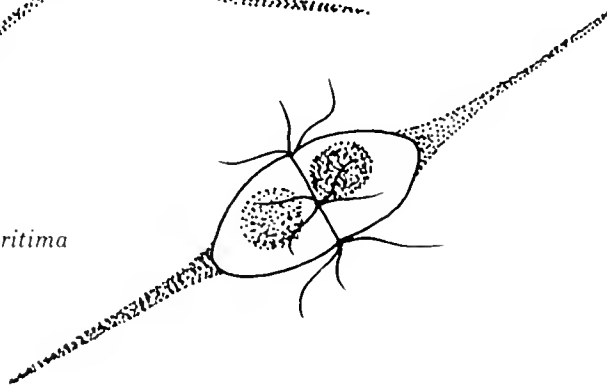
## Key to Species of *Corollospora*

- 1    Ascospores 2-celled . . . . . 2
- 1    Ascospores 4- to 6-celled . . . . . 3

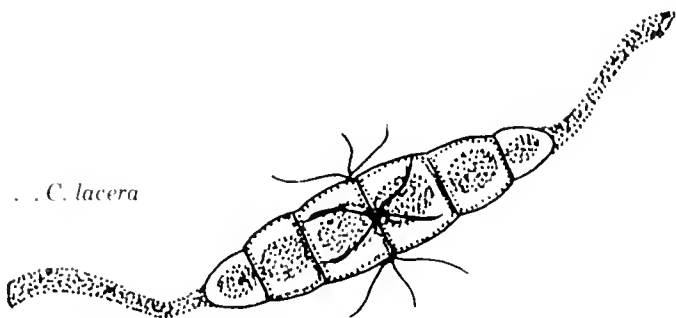
- 2 (1) Spores with 3 (sometimes more) terminal or subterminal appendages at each end . . . . . *C. trifurcata*



- 2 (1) Spores with one slender appendage at each end and several hairlike ones equatorially . . . . . *C. maritima*

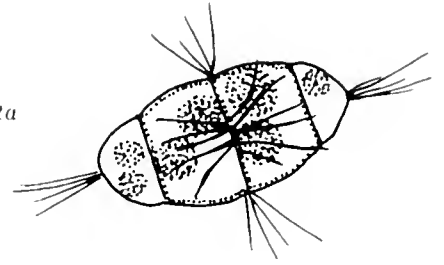


- 3 (1) Apical appendages single, cylindrical, horn-like, thick; equatorial ones hairlike . . . . . *C. lacera*

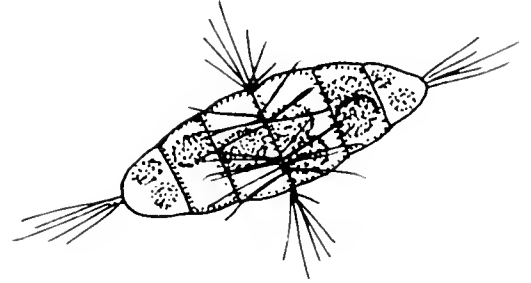


- 3 (1) Apical and equatorial appendages hairlike . . . . . 4

4 (3) Spores predominantly 4-celled . . . . . *C. cristata*

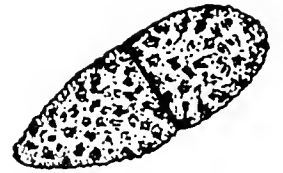


4 (3) Spores predominantly 6-celled . . . . . *C. comata*

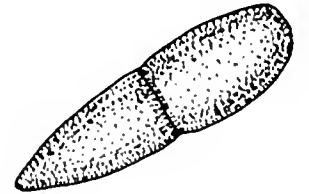


**Key to Species of *Didymosphaeria***

1 Ascospores dark brown, verruculose . . . . . *D. enalia*



1 Ascospores hyaline to light yellow, smooth . . . . . *D. danica*

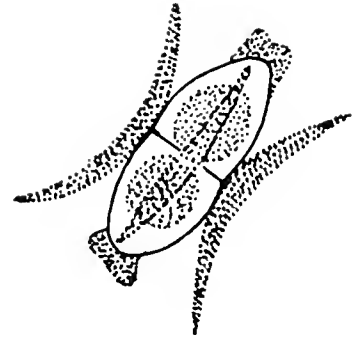


**Key to Species of *Halosphaeria***

1 Ascospores with appendages or processes in both apical and equatorial positions . . . . . 2

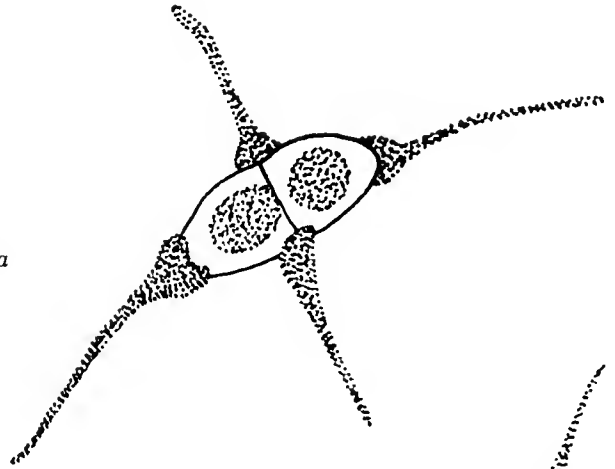
1 Ascospores with appendages or processes in apical positions only . . . . . 4

2 (1) Apical appendages short, caplike or cup-shaped; equatorial ones 3 or 4 in number, crescent-shaped, and more or less rigid . . . . . *H. mediosetigera*

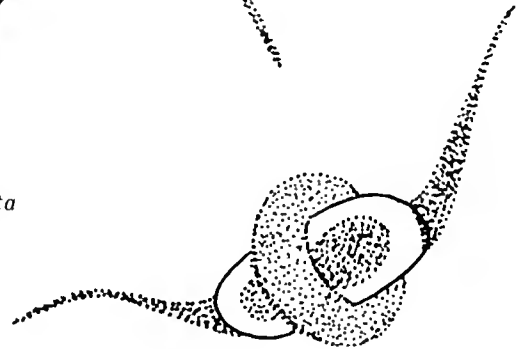


2 (1) Apical appendages long, narrow, flexible; equatorial ones similar, or a collarlike annulus . . . . . 3

3 (2) Equatorial appendages 3 or 4, narrow and flexible . . . . . *H. appendiculata*



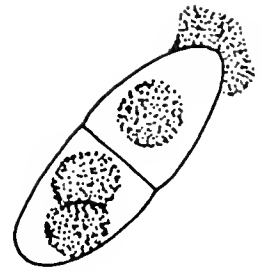
3 (2) Equatorial appendage a collarlike annulus . . . . . *H. torquata*



4 (1) Ascospores with a single appendage at one or both apices; these caplike or subspherical, terminal or subterminal, blunt, tapering or scooplike . . . . . 5

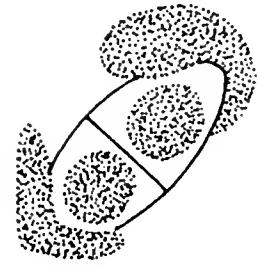
4 (1) Ascospores with two to several appendages at each apex; these fingerlike, stellate, mustache-shaped, hooked, or pleomorphic . . . . . 7

- 5 (4) Appendage short, subspherical, usually only at one end; spores to 69  $\mu$  long . . . . . *H. cucullata*

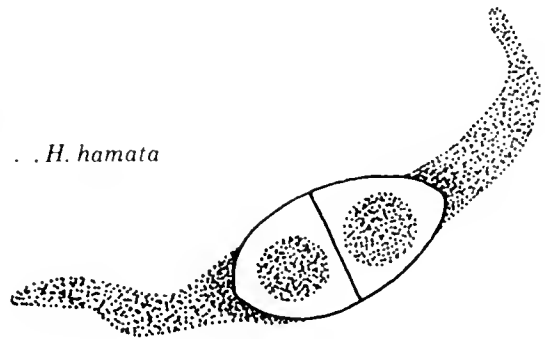


- 5 (4) Appendages blunt, attenuate, hooked or tapering; spores to 35  $\mu$  long . . . . . 6

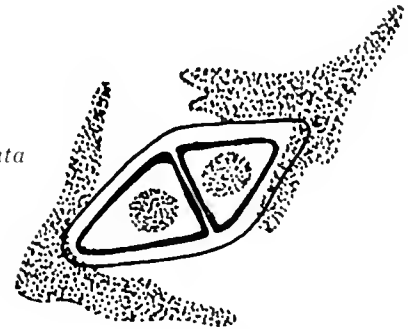
- 6 (5) Appendages blunt, scooplike, occasionally subspherical, to 10  $\mu$  wide . . . . . *H. trullifera*



- 6 (5) Appendages terminal or subterminal, hooked, curved or straight, may be subspherical at first, then becoming tapered . . . . . *H. hamata*



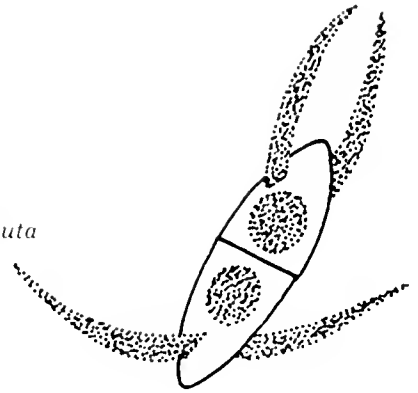
- 7 (4) Ascospores rhomboidal or diamond-shaped; thick-walled . . . . . *H. pilleata*



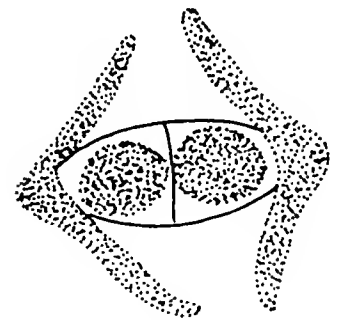
- 7 (4) Ascospores more or less ellipsoidal; not thick-walled . . . . . 8

- 8 (7) Appendages paired; these being either gelatinous or rigid . . . . . 9
- 8 (7) Appendages 3-6 at each end; these being gelatinous or semirigid . . . . . 10

- 9 (8) Appendages rigid, acuminate, directed parallel or at right angles to each other . . . . . *H. quadricornuta*



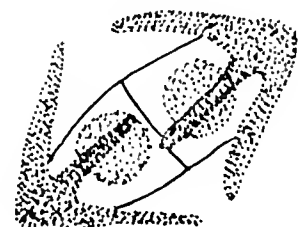
- 9 (8) Appendages gelatinous, stout or tapered, mustache-shaped . . . . . *H. maritima*



- 10 (8) Spores provided with 3-5 terminal or subterminal, straight or curved, tapering, bulbous appendages to 20  $\mu$  long; these being individually attached . . . . . *H. salina*



- 10 (8) Spores provided with 4-6 terminal, attenuate, tapering, semirigid, radiating appendages to 35  $\mu$  long; these forming a common palmlike base at the spore apices . . . . . *H. quadriremis*

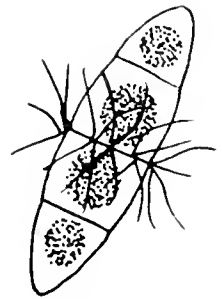




## KEY TO GENERA AND SPECIES OF PHRAGMOSPORAE

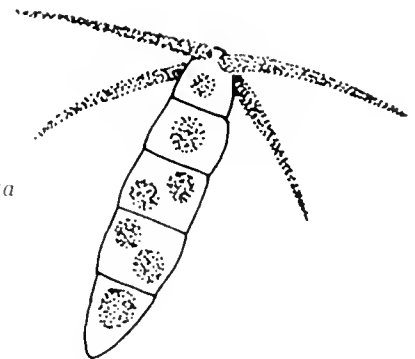
- 1     Ascospores provided with appendages . . . . . 2
- 1     Ascospores without appendages; with or without a gelatinous sheath . . . . . 6
- 2 (1) Ascospores provided with apical or equatorial appendages, but not both . . . . . 3
- 2 (1) Ascospores provided with apical as well as equatorial appendages . . . . . *Corollospora* (p. 22)

- 3 (2) Appendages positioned around the equatorial septum . . . . .  
       . . . . . *Chaetosphaeria chaetosa*



- 3 (2) Appendages positioned at one or both spore apices . . . . . 4

- 4 (3) Ascospores provided with 3 or 4 slender, radiating appendages at one end . . . . . *Torpedospora radiata*



- 4 (3) Ascospores provided with a single appendage at one or both apices . . . . . 5

5 (4) Appendages long, slender, attenuate, may be inflated at tips . . . . . *Haligena* (p. 29)

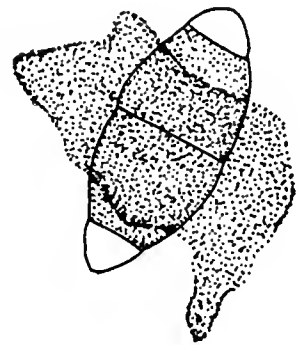
5 (4) Appendages short, blunt, at one or both ends . . . (see also *Didymosporae*) . . . . . *Lentescospora submarina*



6 (1) Spores 2- to 4-celled . . . . . 7

6 (1) Spores 4- to 8-celled . . . . . 9

7 (6) Ascospores provided with an amorphous, gelatinous sheath generally surrounding the midportion of, and usually as thick as, the spore . . . . . *Carbosphaerella leptosphaerioides*



7 (6) Ascospores either without a sheath, or if one is present, not amorphous, but thin and surrounding the entire spore . . . . . 8

8 (7) All cells hyaline and more or less equal in size . . . . . *Sphaerulina* (p. 31)

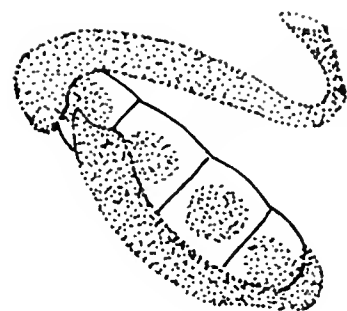
8 (7) All or some cells yellow to yellow-brown, or brown to dark brown; if all hyaline, penultimate cell largest . . . . . *Leptosphaeria* (p. 29)

9 (6) Ascospores hyaline, end cells attenuate and curved . . . . . *Sphaerulina* (p. 31)

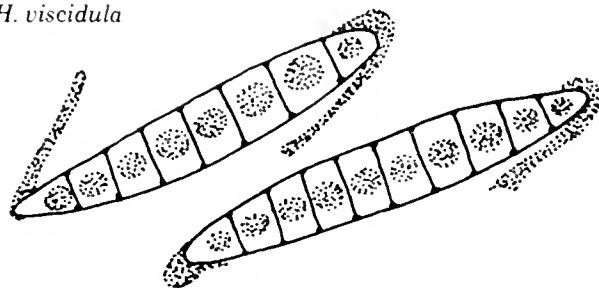
9 (6) Ascospores hyaline or yellow-brown, end cells not attenuate and curved . . . . . *Leptosphaeria* (p. 29)

### Key to Species of *Haligena*

1      Ascospores 4- or 5-celled . . . . . *H. elaterophora*



1      Ascospores 6- to 12-celled . . . . . *H. viscidula*

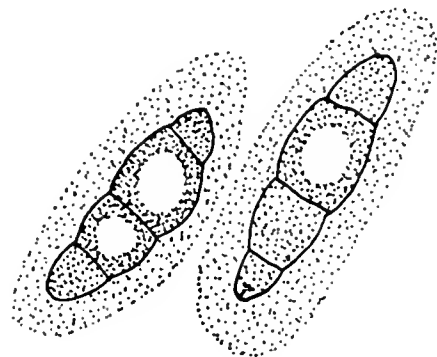


### Key to Species of *Leptosphaeria*

1      Ascospores 2- to 4-celled . . . . . 2

1      Ascospores more than 4-celled . . . . . 6

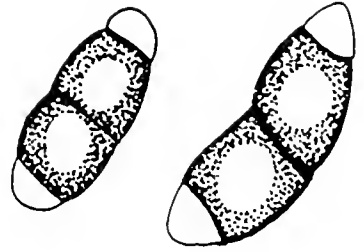
2 (1) Ascospores surrounded by a gelatinous sheath . . . . . *L. halima*



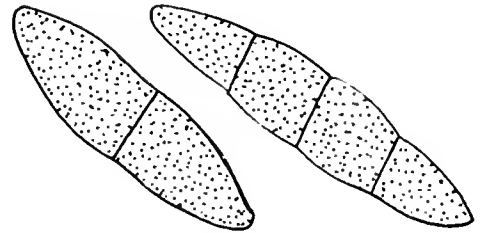
2 (1) Ascospores without a gelatinous sheath . . . . . 3

3 (2) Spores concolorous throughout . . . . . 4

3 (2) Spores with hyaline end-cells and yellow-brown to brown midcells . . . . . *L. discors*

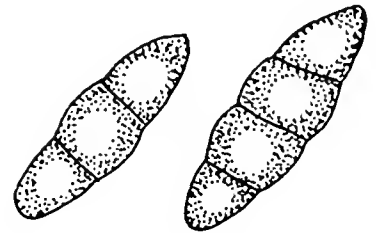


4 (3) Mature spores longer than 35  $\mu$  . . . . . *L. marina*

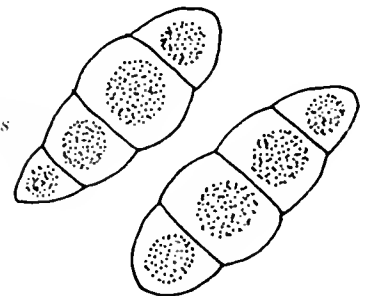


4 (3) Mature spores shorter than 35  $\mu$  . . . . . 5

5 (4) Ascospores 2- to 4-celled, brown or dark brown . . . . . *L. oraemaris*

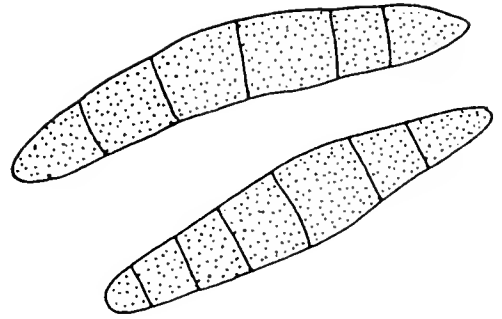


5 (4) Ascospores 4-celled, hyaline, penultimate cell usually largest . . . . . *L. australiensis*

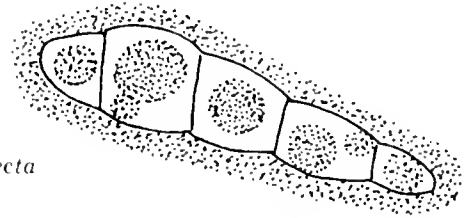


6 (1) Spores surrounded by a gelatinous sheath . . . . . 7

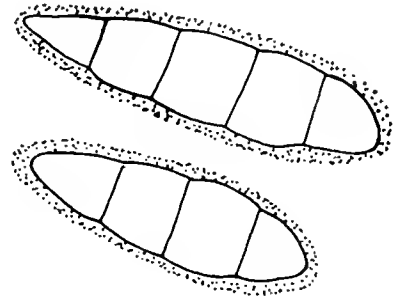
6 (1) Spores without a sheath . . . . . *L. albopunctata*



7 (6) Ascospores 4- or 5-celled, hyaline, penultimate cell usually largest; sheath thick, to 11  $\mu$  . . . . . *L. contecta*

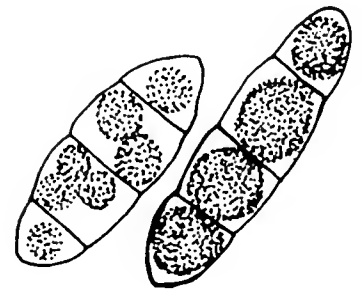


7 (6) Ascospores 4- to 6-celled, yellow to yellow-brown, midcells largest, sheath thin . . . . . *L. maritima*

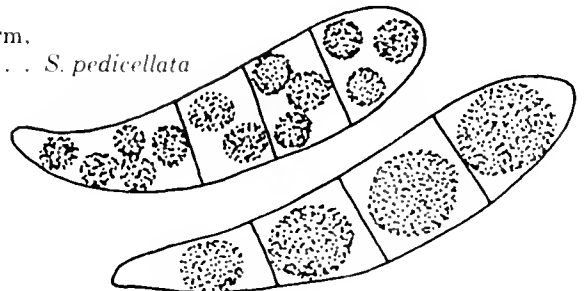


### Key to Species of *Sphaerulina*

1 Ascospores 4-celled, to 36  $\mu$  long, fusoid, both ends rounded . . . . . *S. oraemaris*

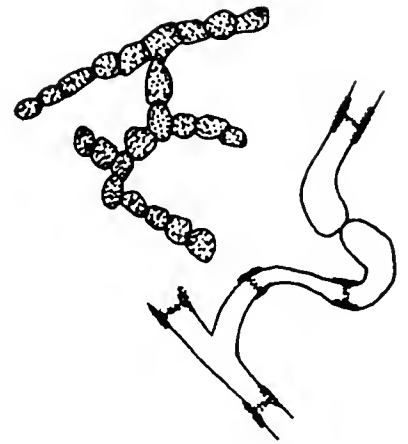


1 Ascospores 4- to 6-celled, to 54  $\mu$  long, elongate-pyriform, one end-cell rounded, the other attenuate and curved . . . . *S. pedicellata*



## KEY TO GENERA AND SPECIES OF DEUTEROMYCETES

1 Conidia produced in pycnidia or directly from vegetative hyphae . . . . . 2

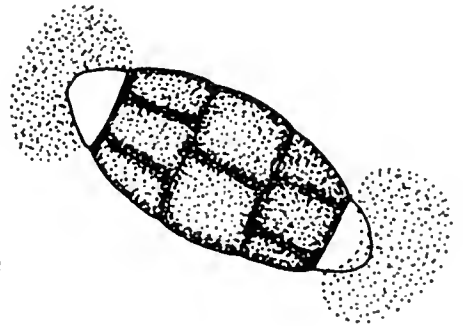


1 Conidia not produced; vegetative hyphae only . . .  
 . . . . . *Papulaspora halima*

2 (1) Conidia produced in pycnidia . . . . . 3

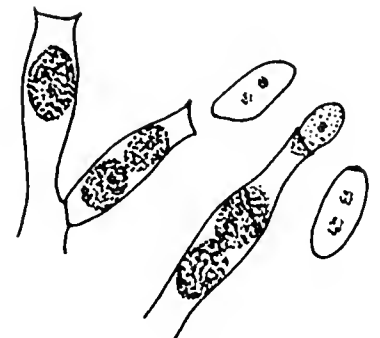
2 (1) Conidia produced directly from vegetative hyphae . . . . . 7

3 (2) Conidia muriform, fuscous or yellow-brown . . . . .  
 . . . . . *Camarosporium metableticum*



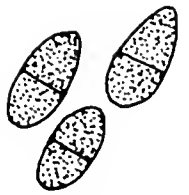
3 (2) Conidia 1- or 2-celled, hyaline to yellow-brown . . . . . 4

4 (3) Conidia produced endogenously within bottle-shaped  
 phialides . . . . . *Phialophorophoma litoralis*



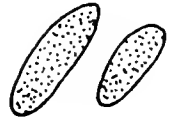
4 (3) Conidia produced exogenously on conidiophores . . . . . 5

5 (4) Conidia 2-celled, rarely 3-celled . . . . . *Diplodia oraeamaris*

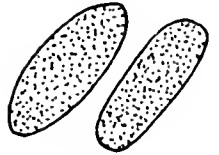


5 (4) Conidia single-celled . . . . . 6

6 (5) Conidia ellipsoidal to bacilliform under 7.5 μ . . . . . *Phoma* (p. 46)



6 (5) Conidia ovate to elongate, over 15 μ long . . . . . *Macrophoma* (p. 45)



7 (2) Conidia 1- or 2-celled . . . . . 8

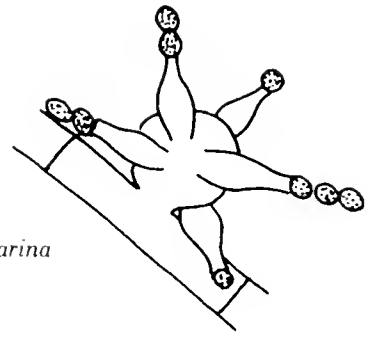
7 (2) Conidia 2- to 12-celled, or muriform . . . . . 15

8 (7) Conidia catenulate or clustered . . . . . 9

8 (7) Conidia produced individually on conidiophores or in phialides . . . . . 14

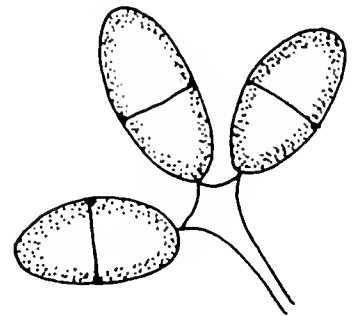
9 (8) Conidia clustered on common basal cells . . . . . 10

9 (8) Conidia catenulate or clustered on individual conidiophores . . . . . 12

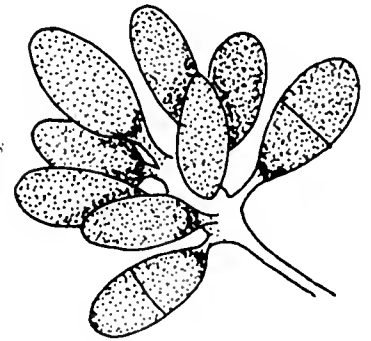


10 (9) Conidia small, under  $5\ \mu$  . . . . . *Botryophialophora marina*

10 (9) Conidia large,  $9-23\ \mu$  long . . . . . 11



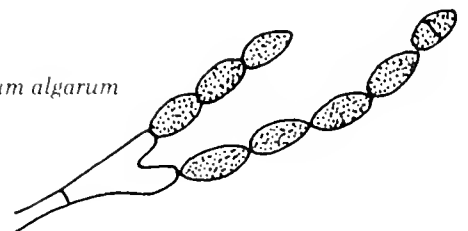
11 (10) Conidia hyaline, 2-celled, rarely single-celled, borne on peg-like teeth of conidiophores . . . . . *Arthrobotrys superba*



11 (10) Conidia fuscous, single-celled, rarely 2-celled, borne on basidiumlike conidiophores . . . . . *Asteromyces cruciatus*

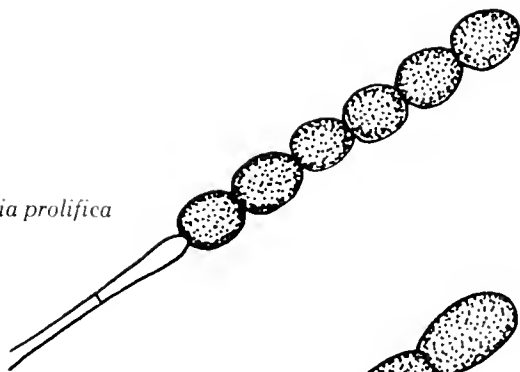
12 (9) Conidia clustered or in long chains, spherical . . . . . 13

12 (9) Conidia in short chains, subglobose or ellipsoidal . . . . .  
 . . . . . *Cladosporium algarum*

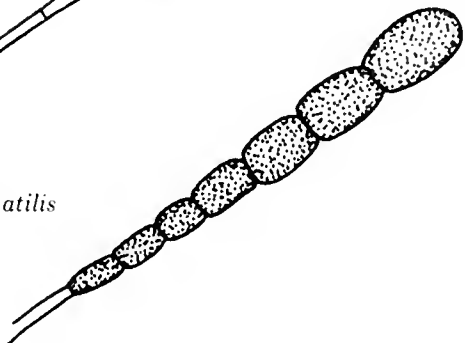




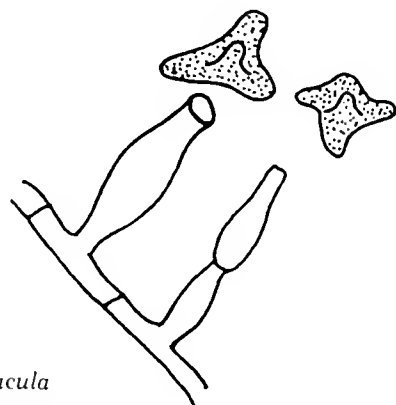
13 (12) Conidial cells all uniform in diameter; conidiophores long, distinct . . . . . *Periconia prolifica*



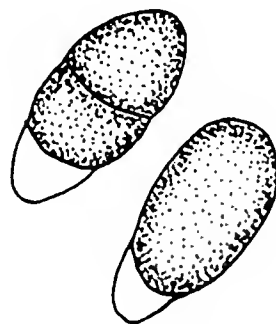
13 (12) Conidial cells increasing in diameter from base to apex; conidiophores short, indistinct . . . . . *Cremasteria cymatilis*



14 (8) Conidia staurosporous, single-celled, borne in phialides with collarlike rim . . . . . *Clavatospora stellatacula*



14 (18) Conidia obpyriform, 1- or 2-celled, produced on conidiophores . . . . . *Humicola alopallonella*

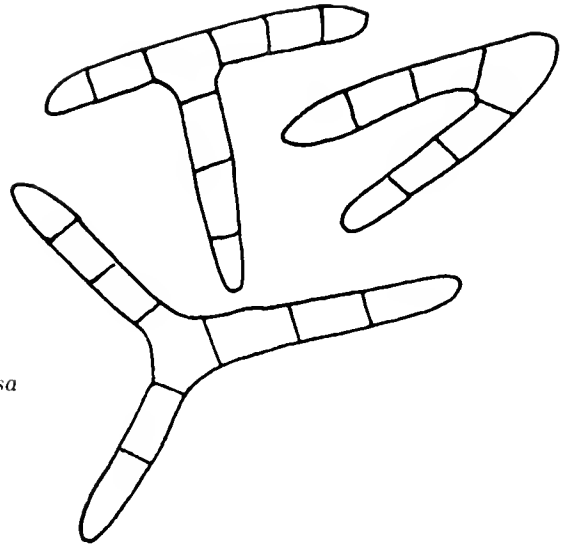


15 (7) Conidia staurosporous or muriform . . . . . 16

15 (7) Conidia ellipsoidal, elongate, curved or helicoid . . . . . 22

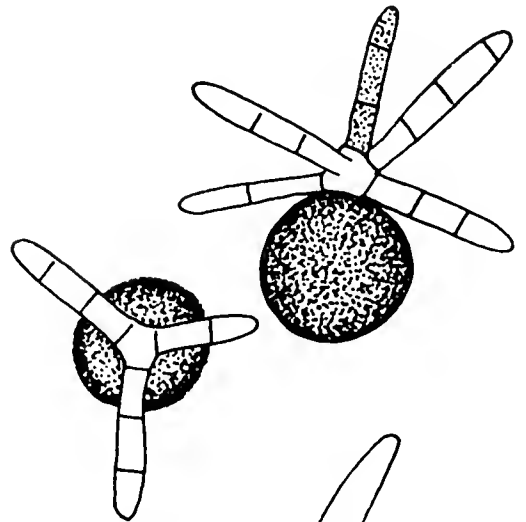
16 (15) Conidia staurosporous . . . . . 17

16 (15) Conidia muriform . . . . . 19



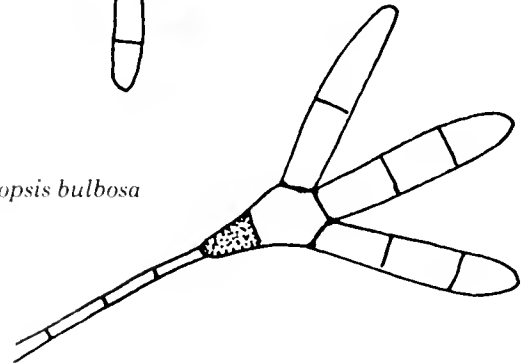
17 (16) Conidial cells hyaline throughout . . . . .  
. . . . . *Varicosporina ramulosa*

17 (16) Conidial cells not hyaline throughout . . . . . 18

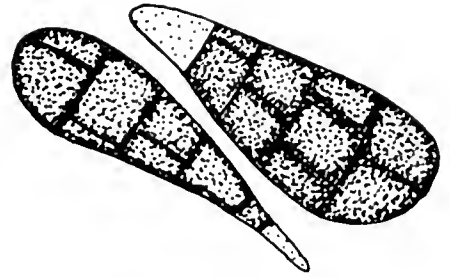


18 (17) Basal conidial cell spherical, dark brown  
to black, 20  $\mu$  in diameter or larger . . . . .  
. . . . . *Orbimyces spectabilis*

18 (17) Basal conidial cell ellipsoidal, light brown, 20  $\mu$  in dia-  
meter or smaller . . . . . *Clavariopsis bulbosa*

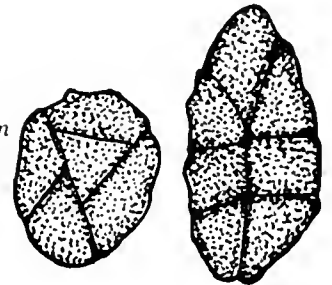


19 (16) Conidia pyriform, apical cell often attenuate, to 80  $\mu$  long . . . . . *Alternaria maritima*



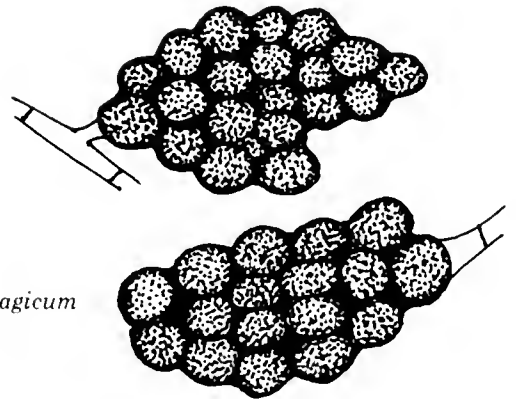
19 (16) Conidia variously shaped, apical cell not attenuate, under 50  $\mu$  long . . . . . 20

20 (19) Conidia globose to ellipsoidal, 19-32  $\mu$  long; conidiophores absent or reduced to simple pedicels . . . . . *Stemphylium maritimum*

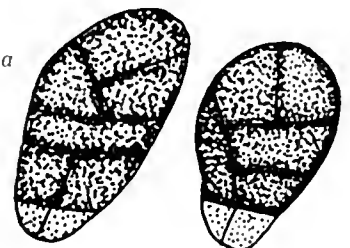


20 (19) Conidia clavate, pyriform; conidiophores distinct . . . . . 21

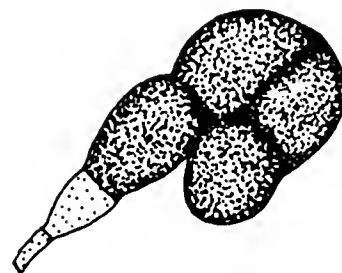
21 (20) Conidia fuscous or dark brown, cells more or less parallel arising from a single basal cell . . . . .  
 . . . . . *Dictyosporium pelagicum*



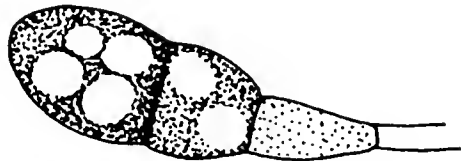
21 (20) Conidia dark brown to black, cells irregular; 1 to 3 light basal cells may occur . . . . . *Monodictys pelagica*



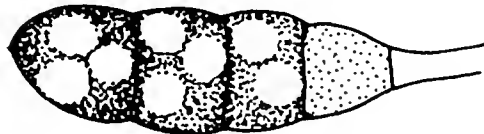
- 22 (15) Conidia curved, bent or straight . . . . . 23
- 22 (15) Conidia coiled one to several times . . . . . *Zalerion* (p. 39)
- 23 (22) Conidia cylindrical to ellipsoidal, straight or slightly curved, smooth or slightly echinulate:  
apical tip may be slightly thickened . . . . . *Dendryphiella* (p. 38)
- 23 (22) Conidia clavate, curved or straight; cells increasing in size from base to apex . . . . . 24



- 24 (23) Conidia 3- to 9-celled, curved . . . . . *Cirrenalia macrocephala*

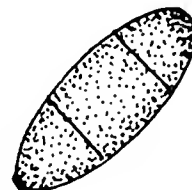


- 24 (23) Conidia 3- to 6-celled, straight . . . . .  
. . . . . *Trichocladium achrasporum*

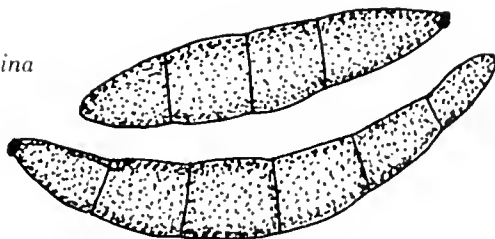


**Key to Species of *Dendryphiella***

- 1 Conidia 2- to 4-celled, to 20  $\mu$  long . . . . . *D. arenaria*

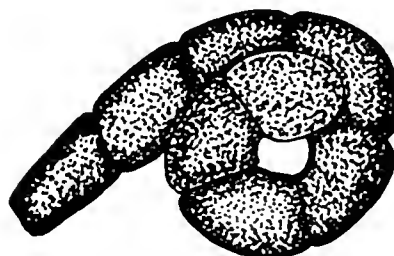


- 1 Conidia 2- to 12-celled, to 75  $\mu$  long . . . . . *D. salina*

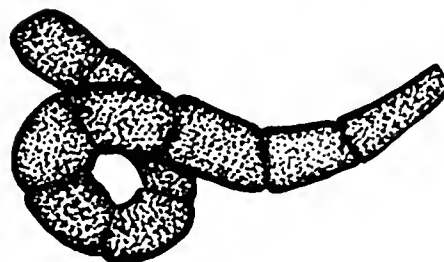


**Key to Species of Zalerion**

- 1 Conidial filament forming a more or less perfect 1- to 3-coiled spiral . . . . . *Z. maritimum*



- 1 Conidial filament forming a variable, irregular spiral in 3 planes . . . . . *Z. varium*



**ANNOTATED LIST OF SPECIES**

The following list of descriptions is arranged alphabetically. The name of each species is followed by the authority and the journal in which the original description or current valid name of the organism was first published. Lists of synonyms have been restricted to those which have come into existence since 1970. For a recent, thorough treatment of the systematics and taxonomic positions of higher marine fungi see Kohlmeyer (1972) and Hughes (1975).

**PHYLUM EUMYCOPHYTA**

**CLASS ASCOMYCETES**

*Amylocarpus encephaloides* Currey  
 Proc. R. Soc. Lond. 9:119. 1857-1859.  
 Ascospores hyaline, globose to subglobose, single-celled, guttulate, 8-16  $\mu$  in diameter, with 10-15 rigid, awl-

shaped appendages arising from various positions along the wall, 5-10  $\mu$  long.

Ascocarp a cleistothecium 160-500  $\times$  200-670  $\mu$ .  
 In driftwood and other cellulosic material.

*Carbosphaerella leptosphaerioides* Schmidt  
 Nat. Nat. Mecklenburg 7:9-10. 1969.

Ascospores 4-celled, 25-40  $\times$  16-20  $\mu$ , broadly ellipsoidal; the two central cells large, dark brown; apical cells small, hyaline; constricted at septa; provided with a gelatinous, hyaline amorphous sheath.  
 In driftwood and other cellulosic material.

*Ceriosporopsis cambrensis* Wilson

Trans. Br. Mycol. Soc. 37:276. 1954.  
 Ascospores hyaline, ellipsoidal to subcylindrical, ends obtuse, 2-celled, one large guttule per cell, may or may not be constricted at the septum, 22-33  $\times$  5-14.5  $\mu$ ; provided at each end with a single, filiform, deliques-

cent appendage to 45  $\mu$  in length; wholly or partially surrounded by a gelatinous sheath.  
In driftwood, reeds, and other cellulosic material.

*Ceriosporopsis halima* Linder  
Farlowia 1:409. 1944.

Ascospores hyaline, ellipsoidal, oblong-ellipsoidal or fusiform, 2-celled, 1-2 guttules per cell, may be constricted at septum, 17-40  $\times$  6-14  $\mu$ ; provided at each end with a single, broadly filiform, tapering appendage 5-8  $\mu$ ; in diameter, wholly or partially surrounded by a gelatinous sheath.  
In driftwood, reeds, and other cellulosic material.

*Ceriosporopsis tubulifera* (Kohlm.) Kirk in Kohlm.  
Can. J. Bot. 50:1953. 1972.  
Syn. *Halosphaeria tubulifera* Kohlm.  
Nova Hedwigia 2:312. 1960.

Ascospores hyaline, ellipsoidal, 2-celled, slightly constricted at septum, 1 or 2 guttules per cell, 14.5-23  $\times$  8.5-11  $\mu$ ; provided at each end with a broad, flared, tapering, tubular appendage, 9-15  $\times$  5-8.5  $\mu$  long, in addition, with a 4.5-8.5  $\mu$  thick equatorial collar which eventually splits into several wedge-shaped fragments.  
In driftwood, reeds, and other cellulosic material.

*Chaetosphaeria chaetosa* Kohlm.  
Nova Hedwigia 6:307-308. 1963.

Ascospores hyaline, fusiform, or elongate to ellipsoidal, 4-celled, each cell guttulate, constricted at septa, 25.5-36.5  $\times$  7.5-11.5  $\mu$ ; provided around the midseptum with numerous, stiff setae, 6-7.5  $\mu$  long.  
Ascus provided with a disc-shaped, perforated apparatus.  
In driftwood and other cellulosic material.

*Corollospora comata* (Kohlm.) Kohlm.  
Ber. Dtsch. Bot. Ges. 75:126. 1962.

Ascospores predominately 6-celled, fuscous to gray around midseptum, end cells hyaline, fusoid, multiguttulate, constricted at septa, 31-54  $\times$  12-17  $\mu$ ; provided with a tuft of up to 15 hairs at each end and additionally with hairlike tufts at the midseptum; apical hairs to 15  $\mu$  long, equatorial ones to 20  $\mu$  long.  
In driftwood and other cellulosic material.

*Corollospora cristata* (Kohlm.) Kohlm.  
Ber. Dtsch. Bot. Ges. 75:126. 1962.

Ascospores predominately 4-celled, ellipsoidal, fuscous to gray around midseptum, end cells hyaline, multiguttulate, constricted at septa, 24-41  $\times$  8.5-16.5  $\mu$ ; provided with a tuft of hairs at each end and additionally with hairlike tufts at the midseptum; apical hairs to 8  $\mu$  long, equatorial ones to 9  $\mu$  long.  
In driftwood and other cellulosic material.

*Corollospora lacera* (Linder) Kohlm.  
Ber. Dtsch. Bot. Ges. 75:126. 1962.

Ascospores 4- to 6-celled (predominately 5-celled) fusoid

or oblong-ellipsoidal, fuscous to brownish around midseptum, end cells hyaline, constricted at septa, 1 large guttule per cell, 39-63  $\times$  10-15  $\mu$ ; provided at each end with a slender, subcylindrical, slightly curved appendage, approximately hornlike in general shape, to 20  $\mu$  long; in addition, the midseptum is provided with several flexible setae arising peritrichously.

In driftwood, reeds, calcareous substrates associated with sandy beaches. Rare.

*Corollospora maritima* Werdermann  
Notizbl. Bot. Gart. Mus. Berlin-Dahlem 8:248. 1922.

Ascospores 2-celled, hyaline to pale yellow around septum, fusoid to ellipsoidal, 1 or 2 guttules per cell, conspicuously constricted at septum, 20-53  $\times$  6.5-14  $\mu$ ; each end provided with a slender, tapering, rigid or semirigid appendage to 20  $\mu$  long; in addition, the septum is provided with several flexible setae arising peritrichously.

In driftwood, cellulosic materials, and calcareous fragments.

*Corollospora trifurcata* (Höhnk) Kohlm.  
Ber. Dtsch. Bot. Ges. 75:126. 1962.

Ascospores 2-celled, 26-38.5  $\times$  9-16.5  $\mu$ , hyaline, fusoid to ellipsoidal, constricted at septum, 1-4 guttules in each cell; provided at each end with 3 (sometimes 4-7) terminal or subterminal, tapering, curved, deciduous appendages to 39  $\mu$  long.

In driftwood, cellulosic materials, and calcareous fragments.

*Didymella fucicola* (Suth.) Kohlm.  
Phytopathol. Z. 63:342. 1968.

Ascospores 2-celled, 16-23  $\times$  6-8  $\mu$ , hyaline, ovoid to ellipsoidal, constricted at septum, one cell slightly larger than the other.

Ascocarp a pseudothecium; asci bitunicate, 55-115  $\times$  10-15  $\mu$ .

Reported only in *Fucus*.

*Didymosamarospora euryhalina* Johnson & Gold  
J. Elisha Mitchell Sci. Soc. 73:104. 1957.

Ascospores 2-celled, 25-34  $\times$  12-17  $\mu$ , hyaline, broadly ovoid or broadly fusoid, ends rounded or slightly tapered, 1 guttule per cell, may be slightly constricted at septum, surrounded by a gelatinous, undulant sheath.

In *Juncus* and other culms.

*Didymosphaeria danica* (Berlese) Wilson & Knoble  
Trans. Br. Mycol. Soc. 44:57. 1961.

Ascospores 2-celled, 33-44  $\times$  5-8  $\mu$ , hyaline to slightly yellow, broadly ellipsoidal or fusiform-ellipsoidal, lower half of spore shorter and cylindrical, the upper half slightly larger and pointed.

Ascocarp a pseudothecium; asci bitunicate, 83-94  $\times$  12-14  $\mu$ .

Reported only in *Chondrus*.

*Didymosphaeria enalia* Kohlm.

Ber. Dtsch. Bot. Ges. 79:28. 1966.

Ascospores 2-celled,  $15.5\text{-}23 \times 6.5\text{-}11 \mu$ , dark brown, ellipsoidal to broadly ellipsoidal, constricted at septum, verruculose.

Ascocarp a pseudothecium; asci bitunicate  $117\text{-}135 \times 12.5\text{-}15.5 \mu$ .

In driftwood. Uncommon in colder waters.

*Haligena elaterophora* Kohlm.

Nova Hedwigia 3:87-88. 1961.

Ascospores 4- or 5-celled, ellipsoidal to oblong,  $24\text{-}54.5 \times 10\text{-}19.5 \mu$ , hyaline, ends rounded; provided at each end with a single, long, curved, grooved, terminally inflated appendage.

In wood and other cellulosic material.

*Haligena viscidula* J. & E. Kohlm.

Nova Hedwigia 9:92. 1965.

Ascospores 6- to 12-celled,  $37.5\text{-}89 \times 4\text{-}6.5 \mu$ , hyaline, fusiform to fusiform-cylindrical, each cell with a single guttule, may be slightly constricted at septa; provided at each end with a short, caplike or threadlike appendage  $6\text{-}10 \mu$  long.

In driftwood and other cellulosic material.

*Haloguignardia oceanica* (Ferd. & Winge) Kohlm.

Mar. Bio. 8:344. 1971.

Syn. *Phyllachorella oceanica* Ferd. & Winge  
Mycologia 12:102-103. 1920.

Ascospores single-celled,  $20\text{-}31.5 \times 9\text{-}14 \mu$ , hyaline, ellipsoidal to fusiform, ends rounded; provided at each end with a small, gelatinous, deciduous appendage.

Ascocarp a stromatic structure with unitunicate asci.

In *Sargassum*.

*Haloguignardia tumefaciens* (Cribb & Herbert) Cribb & Cribb

Pap. Univ. Queensland Dep. Bot. 3:98. 1956.

Ascospores single-celled,  $30\text{-}59 \times 10\text{-}21.5 \mu$ , hyaline, broadly ellipsoidal or fusiform-ellipsoidal, ends tapered, may be densely granular around the equator; provided at each end with an acute, nipplelike appendage surrounding the peglike apical tip of the spore.

In *Sargassum*.

*Halonectria milfordensis* Jones

Trans. Br. Mycol. Soc. 48:287. 1965.

Ascospores single-celled,  $16.5\text{-}29 \times 2\text{-}3.5 \mu$ , hyaline, fusiform to fusiform-cylindrical, straight or curved, equatorial area of spore sometimes broader than ends; multiguttulate.

In driftwood and other cellulosic material. Rare.

*Halosphaeria appendiculata* Linder

Farlowia 1:412. 1944.

Ascospores 2-celled,  $16\text{-}30 \times 6\text{-}14 \mu$ , hyaline, ellipsoidal to ellipsoidal-cylindrical, each cell with one or more guttules, constricted at septum; provided at each end

with a single, narrow, flexible, gelatinous, deliquescent appendage, and 3 or 4 similar ones equatorially, to  $18 \mu$  in length.

In reeds, driftwood, and other cellulosic material.

*Halosphaeria cucullata* (Kohlm.) Kohlm.

Can. J. Bot. 50:1956. 1972.

Syn. *Remispora cucullata* Kohlm.

Mycologia 56:770. 1964.

Ascospores 2-celled,  $21\text{-}69 \times 6\text{-}12 \mu$ , hyaline, ellipsoidal or cylindrical, one to several guttules per cell, may be slightly constricted at septum; provided with a gelatinous, caplike or subspherical, deciduous appendage  $5\text{-}10 \mu$  in diameter at one end of the spore.

In driftwood.

*Halosphaeria hamata* (Höhnk) Kohlm.

Can. J. Bot. 50:1956. 1972.

Syn. *Remispora hamata* (Höhnk) Kohlm.

Ber. Dtsch. Bot. Ges. 74:305. 1961.

Ascospores 2-celled,  $16\text{-}31 \times 6.5\text{-}11 \mu$ , hyaline, ellipsoidal to fusiform-ellipsoidal, 1 or 2 guttules per cell, may be constricted at septum; provided at each end with a terminal or subterminal, gelatinous, hooked, curved, or straight appendage; appendage may be caplike or rounded at first, later becoming tapered.

In *Typha*, reeds, driftwood, and other cellulosic material.

*Halosphaeria maritima* (Linder) Kohlm.

Can. J. Bot. 50:1956. 1972.

Syn. *Remispora maritima* Linder.

Farlowia 1:410. 1944.

Ascospores 2-celled,  $16\text{-}35 \times 8.5\text{-}15 \mu$ , hyaline, ovoid, ellipsoidal or elongate, 1 large guttule per cell, may be constricted at septum; provided at each end with a mustache-shaped, or pair of stout, tapering, gelatinous appendages.

In driftwood and other cellulosic material.

*Halosphaeria mediosetigera* Cribb & Cribb

Pap. Univ. Queensland Dep. Bot. 3:100. 1956.

Ascospores 2-celled,  $24\text{-}44.5 \times 8\text{-}20 \mu$ , hyaline, fusiform-ellipsoidal or ellipsoidal, ends rounded, may be slightly constricted at septum, one or more guttules per cell; provided at each end with a small cap-shaped or cup-shaped gelatinous appendage which may become free; in addition, with 3 or 4 crescent-shaped, rigid, equatorial appendages attached laterally along the septum.

In driftwood and other cellulosic material.

*Halosphaeria pilleata* (Kohlm.) Kohlm.

Can. J. Bot. 50:1957. 1972.

Syn. *Remispora pilleata* Kohlm.

Nova Hedwigia 6:319. 1963.

Ascospores 2-celled,  $24\text{-}36.5 \times 12.5\text{-}19 \mu$ , hyaline, rhomboidal or diamond-shaped, thick-walled, usually 1 guttule per cell; provided at each end with a gelatinous, pleomorphic, veil-like or sheath-like appendage.

In driftwood.

*Halosphaeria quadricornuta* Cribb & Cribb

Pap. Univ. Queensland Dep. Bot. 3:99. 1956.

Ascospores 2-celled,  $20\text{-}36 \times 6\text{-}11.5 \mu$ , hyaline, ellipsoidal, fusiform-ellipsoidal or elongate, one to several guttules per cell, may be constricted at septum, occasionally covered by a gelatinous sheath; provided at each end with a pair of acuminate, rigid, occasionally subterminal appendages directed either parallel or at right angles to each other, to  $37 \mu$  long.

In driftwood and other cellulosic material. Uncommon in colder waters.

*Halosphaeria quadriremis* (Höhnk) Kohlm.

Can. J. Bot. 50:1957. 1972.

Syn. *Remispora quadri-remis* (Höhnk) Kohlm.

Nova Hedwigia 2:332. 1960.

Ascospores 2-celled,  $18\text{-}34 \times 8\text{-}15.5 \mu$ , hyaline, ellipsoidal to ovoid, ends rounded, 1 or 2 large guttules per cell, may be constricted at septum; provided at each end with 4-6 gelatinous, attenuate, semirigid, tapering appendages, to  $35 \mu$  long.

In driftwood and other cellulosic material.

*Halosphaeria salina* (Meyers) Kohlm.

Can. J. Bot. 50:1957. 1972.

Syn. *Remispora salina* (Meyers) Kohlm.

Mycologia 60:262. 1968.

Ascospores 2-celled,  $19\text{-}28 \times 8\text{-}13.5 \mu$ , hyaline, ellipsoidal to oblong-ellipsoidal, ends rounded, may be slightly constricted at septum, usually 1 large guttule per cell; provided at each end with 3 or 4 (occasionally 5), terminal or subterminal, radiating, curved or straight, tapering, more or less bulbous appendages to  $20 \mu$  long.

In driftwood and other cellulosic material. Less common in colder waters.

*Halosphaeria torquata* Kohlm.

Nova Hedwigia 2:311. 1960.

Ascospores 2-celled,  $20\text{-}30.5 \times 10\text{-}16 \mu$ , hyaline, broadly ellipsoidal, 1 large guttule per cell, may be constricted at septum; provided at each end with a gelatinous, tapering, semirigid appendage to  $15 \mu$  long; in addition, with an equatorial cylindrical annulus around the septum.

In driftwood and other cellulosic material.

*Halosphaeria trullifera* (Kohlm.) Kohlm.

Can. J. Bot. 50:1957. 1972.

Syn. *Remispora trullifera* Kohlm.

Nova Hedwigia 6:321. 1963.

Ascospores 2-celled,  $23\text{-}35 \times 14\text{-}20 \mu$ , hyaline, ellipsoidal to oblong-ellipsoidal, ends rounded, 1 large guttule per cell, may be constricted at septum; provided at each end with a gelatinous slightly tapering or blunt, scoop-like or caplike appendage, to  $10 \mu$  wide.

In driftwood.

*Keissleriella blepharospora* J. & E. Kohlm.

Nova Hedwigia 9:97. 1965.

Ascospores 2-celled,  $12\text{-}21 \times 6\text{-}8 \mu$ , hyaline, ellipsoidal, ends rounded, one to several guttules per cell, may be constricted at septum; provided at one end with 4-7 terminal, radiating, stiff or bristlelike appendages to  $13 \mu$  long.

Known only from bark of *Rhizophora*. Collected on drift floating northward.

*Lentescospora submarina* Linder

Farlowia 1:411. 1944.

Ascospores 2-celled (may become 3-celled prior to germination), thick-walled,  $15.5\text{-}35 \times 6\text{-}15 \mu$ , hyaline, ellipsoidal or elongate-ellipsoidal, occasionally each cell with a large guttule, may be slightly constricted at septum; provided at each end, only rarely at a single end, with a short, gelatinous appendage, to  $7 \mu$  in diameter.

In driftwood and other cellulosic material.

*Leptosphaeria albopunctata* (West.) Sacc.

Sylloge Fungorum 2:72. 1883.

Ascospores 6- to 8-celled, antepenultimate cell largest,  $26\text{-}55 \times 10\text{-}15 \mu$ , light yellow to yellow-brown, fusiform, to broadly fusiform, ends rounded, slightly constricted at septa, curved or straight.

Ascocarp a pseudothecium; asci bitunicate.

In *Juncus*, *Spartina*, reeds, driftwood, and other cellulosic material.

*Leptosphaeria australiensis* (Cribb & Cribb) Hughes

Sysis 2:132. 1968.

Ascospores 4-celled, penultimate cell usually largest,  $19\text{-}27 \times 5.5\text{-}9 \mu$ , hyaline, fusiform to ellipsoidal, 1 guttule per cell, constricted at septa.

Ascocarp a pseudothecium; asci bitunicate.

In floating drift of *Rhizophora*, *Avicennia*, driftwood, and other cellulosic material. Less common in colder waters.

*Leptosphaeria contecta* Kohlm.

Nova Hedwigia 6:314. 1963.

Ascospores 4- to 5-celled, penultimate cell usually largest,  $32.5\text{-}44 \times 8.5\text{-}11.5 \mu$ , hyaline, fusiform to fusiform-ellipsoidal, straight or curved, cells guttulate, constricted at septa, covered by a gelatinous, hyaline sheath  $9\text{-}11 \mu$  thick.

Ascocarp a pseudothecium; asci bitunicate.

In driftwood, reeds, and other cellulosic material.

*Leptosphaeria discors* Saccardo & Ellis in Saccardo

Michelia 2:567. 1882.

Ascospores 4-celled,  $28\text{-}36 \times 8\text{-}14 \mu$ , broadly fusoid or broadly ellipsoidal, straight or curved, end cells rounded and hyaline, midcells yellow-brown or brown and each containing a large guttule, constricted at septa.

Ascocarp a pseudothecium; asci bitunicate.

In *Juncus*, *Spartina*, floating culms, reeds, driftwood, and other cellulosic material.

*Leptosphaeria halima* Johnson

Mycologia 48:502. 1956.



- Ascospores 4-celled, penultimate cell largest,  $12-18 \times 5-8 \mu$ , yellow-brown, straight or curved, fusiform or cylindrical, constricted at septa, enclosed in a hyaline, gelatinous sheath  $11-17 \mu$  thick.  
 Ascocarp a pseudothecium; asci bitunicate.  
 In *Spartina*, reeds, driftwood, and other cellulosic material.
- Leptosphaeria marina* Ellis & Ever.  
 J. Mycol. 1:43. 1885.  
 Ascospores 2- to 4-celled,  $48-72 \times 10-14 \mu$ , yellow to yellow-brown, fusiform or elongate-ellipsoidal, ends rounded, straight or curved, constricted at septa.  
 Ascocarp a pseudothecium; asci bitunicate.  
 In *Juncus*, *Spartina*, reeds, driftwood, and other cellulosic material.
- Leptosphaeria maritima* (Cooke & Plow.) Sacc.  
 Sylloge Fungorum 2:73. 1883.  
 Ascospores 4- to 6-celled,  $30-45 \times 6-14 \mu$ , yellow to yellow-brown, fusiform or ellipsoidal, straight or slightly curved, constricted at septa, conspicuously at mid-septum; midcells of unequal size; covered by a thin gelatinous sheath.  
 Ascocarp a pseudothecium; asci bitunicate.  
 In *Juncus*, *Spartina*, reeds, driftwood, and other cellulosic material.
- Leptosphaeria oraemaris* Linder  
 Farlowia 1:413. 1944.  
 Ascospores 2- to 4-celled,  $17-32 \times 4-8 \mu$ , brown or dark brown, fusoid or ellipsoidal, straight or curved, constricted at septa.  
 Ascocarp a pseudothecium; asci bitunicate.  
 In *Spartina*, reeds, floating culms, driftwood, and other cellulosic material.
- Lignicola laevis* Höhnk  
 Veröff. Inst. Meeresforsch. Bremerhaven 3:216. 1955.  
 Ascospores 2-celled,  $14-24 \times 5.5-9 \mu$ , hyaline, fusiform or ellipsoidal-oblong, straight or slightly curved, 1 large guttule per cell, constricted at septum.  
 In driftwood and other cellulosic material.
- Lindra inflata* Wilson  
 Trans. Br. Mycol. Soc. 39:411. 1956.  
 Ascospores multiseptate, 28- to 52-celled, septa narrowly spaced,  $210-415 \times 4-6 \mu$ , filiform, hyaline, diameter uniform throughout, multiguttulate; provided at each end with a short, clavate or subglobose, hyaline, gelatinous appendage.  
 In reeds, driftwood, and other cellulosic material.
- Lindra thalassiae* Orpurt, Meyers, Boral, & Sims  
 Bull. Mar. Sci. Gulf Caribb. 14:406. 1964.  
 Ascospores multiseptate, 15- to 19-celled,  $230-390 \times 3-6 \mu$ , filiform, curved, s-shaped or u-shaped, hyaline, attenuate, tips slightly inflated.  
 In *Thalassia* and *Sargassum*.
- Lulworthia medusa* (Ell. & Ever.) Cribb & Cribb emend. Cavaliere & Johnson  
 Nova Hedwigia 10:434-435. 1966.  
 Ascospores single-celled,  $60-760 \mu$ , hyaline, filiform, tapering, curved, s-shaped, u-shaped or  $\sigma$ -shaped; provided at each end with a hyaline, tapering, conoid end chamber or appendage of variable length.  
*Lulworthia medusa*—small spore variety "fucicola-type"  $60-150 \mu$ .  
*Lulworthia medusa*—medium spore variety "medusa-type"  $230-500 \mu$ .  
*Lulworthia medusa*—large spore variety "grandispora-type"  $500-760 \mu$ .  
 In *Spartina*, *Typha*, *Zostera*, *Fucus*, stems of various phanerogams, driftwood, reeds, and other cellulosic material.
- Marinospora calyprata* (Kohlm.) Cavaliere  
 Nova Hedwigia 11:548. 1966.  
 Ascospores 2-celled,  $22-35 \times 9-11 \mu$ , hyaline, ellipsoidal, ends rounded, 1 large guttule per cell, constricted at septum; provided with an appendage at each end, and 3 or 4 of them around the septum, each up to  $30 \mu$  in length, hyaline, gelatinous, straight or curved, and terminated with a small involuted cap or tip.  
 Ascocarp a stromatic structure with unitunicate asci.  
 In driftwood, reeds, and other cellulosic material.
- Massariella maritima* Johnson  
 Mycologia 48:846. 1956.  
 Ascospores 2-celled,  $10-15 \times 5-10 \mu$ , yellow-brown, ellipsoidal or oblong-ellipsoidal, 1 large guttule per cell, constricted at septum, surrounded by a relatively thin gelatinous sheath.  
 In driftwood. Rare.
- Microthelia linderi* Kohlm.  
 Trans. Br. Mycol. Soc. 57:483-484. 1971.  
 Syn. *Microthelia maritima* (Linder) Kohlm.  
 Nova Hedwigia 2:322. 1960.  
 Ascospores 2-celled,  $13.5-21 \times 5-7.5 \mu$ , brown or light brown, ellipsoidal, clavate or fusiform-ellipsoidal, occasionally curved, one cell usually larger than the other, cells may be minutely guttulate, constricted at septum.  
 Ascocarp a pseudothecium with bitunicate asci.  
 In driftwood, culms, and other cellulosic material.
- Mycosphaerella ascophylli* Cotton  
 Trans. Br. Mycol. Soc. 3:96. 1908.  
 Ascospores 2-celled,  $15-25 \times 4-6.5 \mu$ , hyaline, fusiform to oblong, ends tapering, guttulate, may be slightly constricted at septum.  
 Ascocarp a pseudothecium with bitunicate asci.  
 In *Ascophyllum*.
- Nais inornata* Kohlm.  
 Nova Hedwigia 4:409. 1962.  
 Ascospores 2-celled,  $22-30 \times 11.5-15.5 \mu$ , hyaline, broadly ellipsoidal or broadly oval, ends rounded, one large

and several small guttules per cell, may be slightly constricted at septum.

In driftwood.

*Phycomelaina laminariae* (Rost.) Kohlm.

Phytopathol. Z. 63:350-356. 1968.

Ascospores 2-celled,  $17.5-27.5 \times 5.5-8 \mu$ , hyaline, fusiform-ellipsoidal or ellipsoidal-cylindrical, straight or slightly curved, usually 1 large guttule per cell, constricted at septum; provided at each end with a cap-like or globose, elongate or attenuate, gelatinous, more or less polymorphic appendage to  $30 \mu$  long.

Ascocarp a stromatic structure with unitunicate asci.

In *Laminaria*.

*Pleospora pelagica* Johnson

Mycologia 48:504. 1956.

Ascospores muriform, 7-9 transverse septa, 0-8 longitudinal septa,  $35-52 \times 9-15 \mu$ , yellow-brown, fusiform, ellipsoidal or clavate-ellipsoidal, usually inequilateral, ends broadly rounded, constricted at midseptum as well as at most others.

Ascocarp a pseudothecium with bitunicate asci.

In *Spartina*, driftwood, and other cellulosic material.

*Pleospora pelvetiae* Sutherland

New Phytol. 14:41. 1915.

Ascospores muriform, 3-7 (predominantly 7) transverse septations, several longitudinal ones,  $20-35 \times 7-12 \mu$ , yellow-brown, ellipsoidal or elongate, ends rounded, constricted at septa.

In various floating culms, reeds, and algae. Uncommon.

*Sphaerulina oraemaris* Linder

Farlowia 1:413. 1944.

Ascospores 4-celled,  $26-36 \times 5-7 \mu$ , hyaline, fusoid, ends broadly rounded, may be slightly constricted at septa.

In *Spartina*, various culms, driftwood, and other cellulosic material.

*Sphaerulina pedicellata* Johnson

Mycologia 48:846. 1956.

Ascospores 4- to 6-celled,  $36-54 \times 6-15 \mu$ , hyaline, elongate-ovoid to elongate-pyriform, one end cell attenuate and curved; may be slightly constricted at septa, usually densely guttulate.

In *Spartina*, various culms, driftwood, and other cellulosic material.

*Torpedospora radiata* Meyers

Mycologia 49:496. 1957.

Ascospores 4- or 5-celled,  $30-52 \times 4-9 \mu$ , hyaline, oblong or elongate-clavate or cylindrical, usually 2 guttules per cell, slightly constricted at septa; provided at one end with 3 or 4 slender, acuminate, semirigid, radiating appendages to  $40 \mu$  long.

In driftwood.

*Trailia ascophylli* Sutherland

Trans. Br. Mycol. Soc. 5:149-151. 1915.

Ascospores long, filamentous and tapering,  $90-100 \times 4$  at the widest diameter, to  $1 \mu$  at the narrowest portion, 2- to 4-celled; septations restricted to broad area of spore, narrow area nonseptate, long and attenuate, occasionally bent or coiled.

In *Ascophyllum* and *Fucus*.

## PHYLUM EUMYCOPHYTA

### CLASS DEUTEROMYCETES

*Alternaria maritima* Sutherland

New Phytol. 15:46. 1916.

Conidia muriform, pyriform; apical cell often attenuate,  $14-80 \times 8-18 \mu$ , dark brown except for hyaline or pale, attenuate apical cell, constricted at septa, catenulate or simple, smooth or verrucose

Conidiophores simple, erect, short.

In *Laminaria*, *Juncus*, *Spartina*, driftwood, and other cellulosic material.

*Arthrobotrys superba* Corda

Pracht-Flora Euro. Schimmel. p. 43. 1839.

Conidia 2-celled, broadly ellipsoidal, ovoid, or slightly pyriform, upper cell usually larger than lower one,  $16-23 \times 7-9.5 \mu$ ; hyaline, slightly constricted at septum. Conidiophores scattered, bearing 4-10 conidia on slightly expanded tips or short, peglike teeth or denticles.

In driftwood.

*Asteromyces cruciatus* F. and Mme. Moreau ex Hennebert

Can. J. Bot. 40:1213. 1962.

Conidia single-celled, rarely 2-celled, ellipsoidal, ovoid or broadly pyriform,  $9-20 \times 4-9 \mu$  fuscous.

Conidiophores basidiallike, hyphae radiate from a bulbous base, bearing 1-13 conidia.

Spores collected in foam.

*Botryophialophora marina* Linder

Farlowia 1:404. 1944.

Conidia single-celled, globose or subglobose,  $2-3 \mu$  in diameter, hyaline or light, in dense, cottonlike clusters. Conidiophores (phialides) arising from a subglobose, basal cell, 3-20 in number,  $3.5-8.5 \times 1.5-2.5 \mu$ , simple or vase-shaped.

In driftwood and other cellulosic material.

*Camarosporium metableticum* Trail

Scott. Nat. 8:267. 1886.

Conidia muriform, broadly ellipsoidal, ends tapering or round,  $27-40 \times 11-16 \mu$ , fuscous or yellow-brown, 3-9 transverse septa, constricted at septa, apical cells lighter; provided at each end with a gelatinous, cap-like or subglobose appendage.

Conidiophores cylindrical, simple,  $5-10 \mu$  long, bearing single conidia.

Pycnidia  $150-216 \times 210-330 \mu$ , subglobose, solitary, black, innate.

In *Ammophila*.

- Cirrenalia macrocephala* (Kohlm.) Meyers & Moore  
Am. J. Bot. 47:346. 1960.  
Conidia 3- to 9-celled, recurved, bent, rarely straight, 10-41 × 18-28 μ; basal cells smaller and hyaline or light, terminal cells larger and fuscous to brown; conspicuously constricted at septa; terminal cell largest, 5-15 μ in diameter.  
Conidiophores simple, short, to 3 septate, hyaline or pale yellow, bearing single conidia.  
In driftwood and other cellulosic material.
- Cladosporium algarum* Sutherland  
New Phytol. 15:37. 1916.  
Conidia 1- or 2-celled, subglobose, oblong or slightly ellipsoidal, smooth, 5-15 × 5-10 μ, pale olive to brown, constricted at septum.  
Conidiophores erect, branched, ends truncate or rounded, septate, olive to brown.  
In *Laminaria*, driftwood, and other cellulosic material.
- Clavariopsis bulbosa* Anastasiou  
Mycologia 53:11. 1961.  
Conidia staurosporous, irregular, 3- or 4-armed, septate, 20-70 × 6-8.5 μ; basal cell brownish; ellipsoidal or ovoid; central cell slightly branched, arms divergent, cylindrical.  
Conidiophores long, cylindrical, hyaline, septate, simple or branched.  
In driftwood and other cellulosic material.
- Clavatospora stellatacula* Kirk  
Mycologia 61:178-181. 1969.  
Conidia single-celled, staurosporous, triangular, 5-7 × 6-7 μ; 4 or 5 subapical projections to 2.5 μ long; hyaline.  
Conidiophores simple, may be slightly branched; phialides with a collarlike rim.  
In driftwood and other cellulosic material.
- Cremasteria cymatilis* Meyers & Moore  
Am. J. Bot. 47:348. 1960.  
Conidia single-celled, subglobose to ellipsoidal, 7.5-27 × 7.5-16 μ, rusty brown to light brown, forming chains (catenulate) 2-20 conidia in length; chain strongly constricted at each cell; cells increasing in diameter from base to apex.  
Conidiophores short, lateral, single or branched.  
In driftwood and other cellulosic materials.
- Dendryphiella arenaria* Nicot  
Rev. Mycol. 23:93. 1958.  
Conidia 2- to 4-celled, ovoid to ellipsoidal, oblong, or slightly cylindrical, 9-20 × 3.5-6 μ, brown, smooth or slightly echinulate, may be slightly constricted at septa; conidial epispore thickened and forms a conspicuous annulus or knot at its point of attachment to the conidiophore.  
Conidiophores simple or branched, septate, apical cell slightly inflated, light brown to subhyaline, 15-90 μ long.  
In *Sargassum*, floating *Thalassia*, *Ammophila*, driftwood, and other cellulosic material.
- Dendryphiella salina* (Suth.) Pugh & Nicot  
Trans. Br. Mycol. Soc. 47:226. 1944.  
Conidia 2- to 12-celled (predominantly 4- to 6-celled), elongate, cylindrical or fusiform-ellipsoidal, straight or curved, ends rounded or tapered, 14.5-75 × 5.5-10.5 μ, light brown or olive, may be slightly constricted at septa; solitary or catenulate; epispore of conidia thickened at point of attachment to the conidiophore.  
Conidiophores simple or branched, septate, apical cell slightly inflated, light brown to yellowish.  
In *Chondrus*, *Laminaria*, *Spartina*.
- Dictyosporium pelagicum* (Linder) G. C. Hughes ex Johnson & Sparrow  
Fungi in Oceans and Estuaries, p. 391-392. 1961.  
Conidia muriform; form variable, ovoid, clavate, branched (boxing-glovelike), irregular; several branches may arise from a single basal cell; 12-66 × 10-35 μ, dark brown, occasionally black; conspicuously constricted at septa.  
Conidiophores simple, pale brown or dark brown, septate, 15-35 × 1-2.5 μ.  
In driftwood and other cellulosic material.
- Diplodia oraemaris* Linder  
Farlowia 1:403. 1944.  
Conidia 2-celled (rarely single-celled), ovoid or ellipsoidal 6-11 × 3.5-7.5 μ, yellow-brown, may be slightly constricted at septum.  
Conidiophores simple, single-celled, hyaline.  
Pycnidia 135-365 × 140-470 μ, flattened, black above, cream to hyaline below.  
In driftwood and other cellulosic material.
- Humicola alopallonella* Meyers & Moore  
Am. J. Bot. 47:346. 1960.  
Conidia 1- or 2-celled (occasionally 3-celled), obpyriform to clavate, 8-17.5 × 15-37.5 μ apical cell largest and brownish, basal cell small and yellow to hyaline, constricted at septum.  
Conidiophores one to several celled, simple, hyaline to light fuscous.  
In driftwood and other cellulosic material.
- Macrophoma* (Saccardo) Berlese & Voglio  
Sylloge Fungorum 1-4:306. 1886.  
Conidia single-celled, ovate to elongate, over 15 μ long, hyaline or subhyaline.  
Conidiophores simple, short, filiform.  
Pycnidia innate, erumpent, lenticular or subglobose.  
In driftwood and other cellulosic material.
- Monodictys pelagica* (Johnson) Jones  
Trans. Br. Mycol. Soc. 46:138. 1963.  
Conidia muriform, ovoid or spherical, more commonly obpyriform, symmetrical or asymmetrical, 15-44 × 12.5-37 μ; 1-3 brown or yellowish basal cells,

brownish or blackish terminal cells, or concolorous black throughout; may be conspicuously constricted at septa. Conidiophores dark fuscous, septate, 1- to 3-celled, simple.

In driftwood and other cellulosic material.

*Orbimyces spectabilis* Linder

Farlowia 1:404. 1944.

Conidia staurosporous, multicellular; basal cell globose to subglobose, shining black, 22-42  $\mu$  in diameter; provided with one or two sets of radiating, fingerlike, septate filaments to 6  $\mu$  wide and to 50  $\mu$  long; each set of radiating filaments join a common dark basal cell; radiating cells may lighten in color from base to apex.

In driftwood and other cellulosic material.

*Papulospora halima* Anastasiou

Nova Hedwigia 6:266. 1963.

Hyphae septate, hyaline or brown, anastomosing, to 4.8  $\mu$  in diameter, with hyphal thickenings, chainlike; large masses of hyphae or bulbils formed, subglobose, compact, to 870  $\mu$  in diameter, dark brown or black.

No spores formed.

In driftwood and other cellulosic material.

*Periconia prolifica* Anastasiou

Nova Hedwigia 6:260. 1963.

Conidia single-celled, globose to subglobose, 6-20  $\mu$  in diameter, light brown to brown, light red to rust, thick-walled, smooth, catenulate.

Conidiophores hyaline, septate, simple or branched, to 200  $\mu$  long.

In driftwood and other cellulosic material.

*Phialophorophoma litoralis* Linder

Farlowia 1:403. 1944.

Conidia single-celled, hyaline, ellipsoidal or pyriform, 2.5-4.5  $\times$  1.5-2  $\mu$ , endogenous.

Conidiophores (phialides) simple or branched, nonseptate, hyaline, bottle-shaped, with a collarlike, flaring neck.

Pycnidia innate, ovoid or ellipsoidal, 110-175  $\mu$ .

In driftwood and other cellulosic material.

*Phoma* Saccardo

Michelia 2:4. 1880.

Conidia single-celled, ellipsoidal or bacilliform, hyaline, 1.5-7.5  $\mu$ .

Pycnidia innate, ellipsoidal or subglobose.

In *Laminaria*, *Spartina*, *Juncus*, various reeds, driftwood, and other cellulosic material.

*Stemphylium maritimum* Johnson

Mycologia 48:844. 1956.

Conidia muriform, ellipsoidal, globose, pyramidal, or irregular, 19-32  $\times$  17-29  $\mu$ , dark brown, smooth or verrucose, constricted at septa.

Conidiophores simple, apiculate.

In driftwood and other cellulosic material.

*Trichocladium achrasporum* (Meyers & Moore) Dixon  
Mycologia 63:237. 1971.

Syn. *Culcitalna achraspora* Meyers & Moore

Am. J. Bot. 47:349. 1960.

Conidia 3- to 6-celled, clavate, ovate, or broadly obpyriform, 15-45  $\times$  8-24  $\mu$ ; cells increase in size and darken from base to apex, terminal cells dark brown; dark cells guttulate; constricted at septa.

Conidiophores simple, short or long, septate, hyaline, pale yellow or brownish, straight or curved.

In driftwood and other cellulosic material.

*Varicosporina ramulosa* Meyers & Kohlm.

Can. J. Bot. 43:916. 1965.

Conidia staurosporous, cylindrical, multiseptate, several radiating arms usually at right angles to each other, hyaline; most commonly seen as 3 radiating, multicellular, cylindrical hyphal strands to 45  $\mu$  long.

In *Sargassum*, *Thalassia*, and *Typha*. Absent in colder waters north of New Jersey.

*Zalerion maritimum* (Linder) Anastasiou

Can. J. Bot. 41:1136. 1963.

Conidia helicosporous, 1-3 times coiled, multiseptate, all cells approximately similar in size, to 14 cells long, occasionally longer, deeply constricted at septa, to 12  $\mu$  in diameter, fuscous to black.

Conidiophores simple or once-branched, hyaline, yellow, or pale fuscous, septate.

In driftwood and other cellulosic material.

*Zalerion varium* Anastasiou

Can. J. Bot. 41:1136. 1963.

Conidia helicosporous, coiled in 3 planes, single or branched; cells 5.5-13  $\times$  4.5-10.5  $\mu$ , dark brown to black; clusters to 65  $\mu$  in diameter, occasionally grape-like clusters.

Conidia simple, fuscous to black.

In driftwood and other cellulosic material.

## PHYLUM EUMYCOPHYTA

### CLASS BASIDIOMYCETES

*Nia vibrissa* Moore & Meyers

Mycologia 51:874. 1959.

Basidiospores single-celled, ovoid to ellipsoidal, or broadly ellipsoidal, 7.5-15  $\times$  6.5-15  $\mu$ , hyaline; provided at one end with a single, gelatinous, filiform appendage to 50  $\mu$  long, and 3-5 additional ones similar in shape but slightly shorter at the opposite end.

Basidiocarp orange to orange-brown.

In *Spartina*, driftwood, and other cellulosic material.

## SELECTED BIBLIOGRAPHY

AINSWORTH, G. C.

1971. Ainsworth & Bisby's dictionary of the fungi. 6th ed. Commonwealth Mycol. Inst., Kew, Surrey, Engl., 663 p.

- BARGHOORN, E. S., and D. H. LINDER.  
1944. Marine fungi: Their taxonomy and biology. *Farlowia* 1:395-467.
- CAVALIERE, A. R.  
1966. Marine Ascomycetes: Ascocarp morphology and its application to taxonomy. I. *Nova Hedwigia* 10:387-398.  
1973. Ascomycetes. In Peter Gray (editor), *Encyclopedia of microscopy and microtechnique*, 638 p. Van Nostrand Reinhold Co.
- CAVALIERE, A. R., and T. W. JOHNSON, JR.  
1966. Marine Ascomycetes: Ascocarp morphology and its application to taxonomy. II-V. *Nova Hedwigia* 10:399-461.
- HUGHES, G. C.  
1975. Studies of fungi in oceans and estuaries since 1961. I. Lignicolous, caulicolous and foliicolous species. *Oceanogr. Mar. Biol. Ann. Rev.* 13:69-180.
- JOHNSON, T. W., JR., and F. K. SPARROW, JR.  
1961. Fungi in oceans and estuaries. J. Cramer Weinheim, 668 p.
- KIRK, P. W., Jr.  
1969. Isolation and culture of lignicolous marine fungi. *Mycologia* 61:174-177.
- KOHLMEYER, J.  
1972. A revision of Halosphaeriaceae. *Can. J. Bot.* 50:1951-1963.
- KOHLMEYER, J., and E. KOHLMEYER.  
1971. Synoptic plates of higher marine fungi. 3rd ed. Lehre Cramer, 87 p.
- SNELL, W. H., and E. A. DICK.  
1957. A glossary of mycology. Harvard Univ. Press, Cambridge, Mass., 171 p.

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to collaborate in the preparation of manuals, reviews manuscripts, and advises the Scientific Editor of the National Marine Fisheries Service.

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## COORDINATING EDITOR'S COMMENTS

Publication of the "Marine Flora and Fauna of the Northeastern United States" is most timely in view of the growing universal emphasis on environmental work and the urgent need for more precise and complete identification of coastal organisms than has been available. It is mandatory, wherever possible, that organisms be identified accurately to species. Accurate scientific names unlock the great quantities of biological information stored in libraries, obviate duplication of research already done, and make possible prediction of attributes of organisms that have been inadequately studied.

A. R. Cavaliere began his studies on the taxonomy and morphology of marine Ascomycetes in 1961 as a graduate student at Duke University. Since that time his work has expanded to include the occurrence and systematics of marine fungi along selected areas of the Pacific and Atlantic coasts of the United States, as well as Nova Scotia and Iceland.

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