(


# A LABORATORY MANUAL <br> OF <br> INVERTEBRATE ZOÖLOGY 

## GILMAN A. DREW, Ph.D.

PROFESSOR OF BIOLOGY AT THE UNIVERSITY OF MAINE; IN CHARGE OF ZOÖLOGICAL INSTRUCTION AT THE MARINE BIOLOGICAL LABORATORY, WOODS HOLL, MASSACHUSETTS

## WITH THE AID OF

MEMBERS OF THE ZOÖLOGICAL STAFF OF Instructors of the marine biological LABORATORY, WOODS HOLL, MASS.

## PHILADELPHIA AND LONDON

## W. B. SAUNDERS COMPANY 1907



$$
\frac{679397}{2.7 .58}
$$

## PREFACE.

The present manual has for its basis a set of laboratory directions prepared by members of the staff of instructors to meet the needs of the class in general zoölogy at the Marine Biological Laboratory of Woods Holl, Massachusetts. Those who were associated with me in the preparation of the first notes were Dr. Robert W. Hall, Dr. James H. McGregor, Mr. Robert A. Budington and Dr. Caswell Grave. Other members of the staff who have either aided me in modifying the original notes or who have added others are Dr. Winterton C. Curtis, Dr. D. H. Tennant, Dr. Otto C. Glaser, Dr. Grant Smith, Dr. John H. McClellan and Dr. Lorande L. Woodruff. Each year for the past six years the directions have been changed where experiences indicated changes should be made.

Probably few instructors will find it desirable for their students to follow closely all that is given in this manual, but it has seemed better to arrange the matter in a logical order, and in some of the forms to call attention to only the important points of anatomy or adaptation, than to try to make the directions for each form complete in themselves. To make the directions for each form complete would necessarily add much labor for the student and would, by the repetition of well-known facts, tend to blunt some of the new and important points to be gained.

The type method of laboratory study has for many years been the prevailing method, but care needs to be exercised to keep students from making everything conform to type, and in leading them to see the wonderful adaptations that fit the different animals for their particular lives. The manual is not intended to lead students to a knowledge of comparative anatomy alone, but to an appreciation of adaptation as well.

It has fallen on me year by year to see that desirable changes were made in the directions, and it has finally been my lot to put them into their present form, but much of the credit belongs to the men who have been associated with me in the instruction work at the Marine Biological Laboratory.

> The Author.


## ont. de UNIV. TURUViV

## CONTENTS.

Page.
PROTOZOA ..... 1
Sarcodina ..... 3
Amœba proteus ..... 3
Foraminifera ..... 4
Actinosphærium or Actinophrys ..... 5
Mastigophora ..... 6
Euglena ..... 6
Volvox ..... 7
Ceratium ..... 8
Noctiluca ..... 8
Sporozoa ..... 9
Gregarina ..... 9
Infusoria ..... 10
Paramecium ..... 10
Spirostomum ..... 11
Vorticella ..... 11
Oxytricha ..... 13
PORIFERA ..... 14
Grantia ..... 15
CGELENTERATA ..... 18
Hydrozoa ..... 20
Hydra (Fresh-water Polyp) ..... 20
Obelia ..... 22
Parypha ..... 24
Gonionemus ..... 25
Hydrocorallina ..... 27
Siphonophora ..... 27
Scyphozoa ..... 27
Aurelia ..... 27
Actinozoa ..... 30
Metridium (Sea-Anemone) ..... 30
Ctenophora ..... 32
Mnemiopsis ..... 32
PLATYHELMINTHES ..... 35
Turbellaria ..... 36
Planaria maculata ..... 36
Bdelloura or Syncœlidium ..... 37
Trematoda ..... 39
Hæmatolæchus (Distomum) ..... 39
Cestoda ..... 41
Crossobothrium laciniatum ..... 41
Platyhelminthes (Continued). Page.
Nemertinea ..... 44
Tetrastemma ..... 44
NEMATHELMINTHES ..... 46
Ascaris ..... 46
Trichina ..... 47
TROCHELMINTHES ..... 48
Rotifera ..... 48
Brachionus (A Rotifer) ..... 48
MOLLUSCOIDA ..... 50
Polyzoa ..... 50
Bugula ..... 50
Plumatella ..... 52
Brachiopoda. ..... 52
Terebratulina ..... 52
ECHINODERMATA ..... 54
Asteroidea ..... 55
Asterias (Starfish) ..... 55
Ophiuroidea ..... 60
Ophiura (Serpent-Star) ..... 60
Echinoidea ..... 61
Arbacia (Sea-Urchin) ..... 61
Holothuroidea ..... 67
Thyone (Sea-Cucumber) ..... 67
ANNELIDA ..... 70
Chetopoda ..... 71
Nereis virens (Clam-Worm) ..... 71
Lumbricus (Earthworm) ..... 74
Autolytus cornutus ..... 80
Lepidonotus squamatus ..... 81
Diopatra cuprea ..... 81
Chrtopterus ..... 82
Amphitrite ornata ..... 83
Cistenides gouldii ..... 84
Clymenella torquata ..... 84
Arenicola cristata ..... 85
Sabella microthalma ..... 85
Hydroides ..... 86
Spirorbis borealis ..... 86
Gephyrea ..... 86
Phascolosoma ..... 86
MOLLUSCA ..... 89
Lamellibranchiata ..... 91
Venus mercenaria (Quohog) ..... 91
Yoldia limatula ..... 99
Mytilus or Modiola (Mussels) ..... 100
Pecten irradians (Scallop) ..... 101
Ostrea virginiana (Oyster) ..... 103
Solenomya ..... 103
Mya arenaria (Long Clam) ..... 104
Ensis directus (Razor-shell Clam) ..... 105
Mollusca (Continued). Page.
Amphineura ..... 106
Chætopleura ..... 106
Gastropoda ..... 107
Sycotypus ..... 107
Loligo pealii (Squid) ..... 115
ARTHROPODA ..... 124
Crustacea ..... 128
Homarus americanus (Lobster) ..... 128
Callinectes hastatus (Blue Crab) ..... 134
Eupagurus (Hermit Crab) ..... 138
Hippa (Sand Mole) ..... 139
Squilla ..... 139
Mysis ..... 141
Talorchestia (Beach-Flea) ..... 141
Porcellio or Oniscus (Sow-Bug) ..... 142
Caprella ..... 143
Branchipus (Fairy Shrimp) ..... 143
Daphnia ..... 144
Cyclops (Water-Flea) ..... 145
Argulus (Fish-Louse) ..... 146
Lepas (Goose-Barnacle) ..... 146
Arachnoidea ..... 147
Limulus (Horseshoe Crab) ..... 147
Buthus (Scorpion) ..... 149
Epeira (Round-Web Spider) ..... 150
Phoxichilidium ..... 152
Myriapoda ..... 152
Lithobius (Centipede, Earwig) ..... 152
Julus (Thousand-legs) ..... 153
Insecta ..... 154
Acridium (Grasshopper) ..... 154
Apis mellifica (Honey-Bee) ..... 159
CHORDATA ..... 164
UROchorda ..... 165
Molgula manhattensis ..... 165
Perophora ..... 168
Botryllus ..... 169
Amarœcium (Sea-Pork) ..... 170
Salpa cordiformis ..... 172
Acrania ..... 173
Amphioxus lanceolatus ..... 173
NOTES FOR GUIDANCE IN MAKING PERMANENT PREPARA- TIONS ..... 175
GLOSSARY ..... 181
INDEX ..... 195
(1)

## DIUL. BE UAMY TORUR

## INVERTEBRATE ZOÖLOGY.

## PROTOZOA.

Unicellular Animals.
Class 1. Sarcodina.
With changeable pseudopodia, during adult life. Reproduction by simple division and by sporeformation.
Subclass 1. Rhizopoda.
With lobose or reticulate pseudopodia.
Order 1. Amœbida.
With lobose pseudopodia. (Amœeba, Arcella, Difflugia.)
Order 2. Reticulariida.
With fine branching and anastomosing pseudopodia. Shells, when present, usually calcareous. (Microgromia, the Foraminifera.)
Subclass 2. Heliozoa.
Typically spherical in form. The pseudopodia, which radiate from the entire surface of the body, are ray-like, seldom changeable, and usually possess an axial filament. (Actinophrys, Actinosphærium, Clathrulina.)
Subclass 3. Radiolaria.
With ray-like pseudopodia, and with a chitinous capsule inclosing the nuclei. The skeleton, when present, is formed of silica or acanthin. All are marine. (Thallassicolla.)
Class 2. Mastigophora.
Motile organs in the form of flagella. Reproduction by longitudinal division. Colony formation is frequent.
Subclass 1. Flagellidia.
With a definite anterior end on which there are
one or more flagella. The members of one order (Choanoflagellidia) have one or more collar-like processes about the base of the single flagellum. (Mastigamœba, Proterospongia, Euglena, Peranema.)
Subclass 2. Dinoflagellidia.
Usually with two flagella, one encircling and the other directed away from the body. (Peridinium, Ceratium.)
Subclass 3. Cystoflagellidia.
With two flagella, one of which is modified into a "tentacle," while the other is short and contained within the gullet. (Noctiluca.)
Class 3. Sporozoa.
Without flagella or cilia in the adult period of the life-cycle. Reproduction is by spore-formation. All are endoparasites.
Subclass 1. Telosporidia.
Reproductive phase of the life-cycle is distinct from, and follows the trophic phase.
Order 1. Gregarinida.
The young stages are intracellular parasites, while the adults are free and motile in the digestive tract or body-cavity of the host. Sporulation occurs within a cyst during the free period of the life-cycle. (Gregarina.)
Order 2. Coccidiida.
Without a free and motile adult stage. Sporulation occurs within a cyst, during the intracellular period of the life-cycle. (Coccidium.)
Order 3. Hæmosporidiida.
Living chiefly in the blood-corpuscles of vertebrates. In many forms the entire sexual period of the life-cycle takes place in an intermediate host, as the mosquito. (Laverania malariæ.)
Subclass 2. Neosporidia.
Reproduction takes place during the trophic phase of the life-cycle.
Order 1. Myxosporidiida.
The initial free stage occurs in the tissues or the cavities of the organs of the host. The adult form is amœeboid. (Myxidium.)

Order 2. Sarcosporidiida.
The initial stage of the life-cycle occurs in the muscle-cells of vertebrates. (Sarcocystis.)
Class 4. Infusoria.
With motile organs in the form of cilia during all or part of the life-cycle. Nucleus dimorphic (macronucleus and micronucleus). Reproduction is by simple transverse division or by budding.
Subclass. 1 Ciliata.
With cilia throughout the life-cycle.
Order 1. Holotrichida.
The cilia are of approximately equal length and thickness and equally distributed over the body. Trichocysts are present. (Prorodon, Paramecium.)
Order 2. Heterotrichida.
With a uniform covering of cilia, together with an "adoral zone" formed of cilia fused into membranelles. (Spirostomum, Stentor, Halteria.)
Order 3. Hypotrichida.
The cilia are limited to the ventral surface of a dorso-ventrally flattened body. Cilia often fused into cirri, membranelles, etc. (Oxytricha, Pleurotricha, Euplotes, Peritromus.)
Order 4. Peritrichida.
More of less bell-shaped in form. Cilia usually reduced to those constituting the adoral zone. (Vorticella, Zoöthamnium, Lichnophora.)
Subclass 2. Suctoria.
Usually possessing cilia only during the embryonic stages of the life-cycle. Tentacles adapted for piercing and sucking are present. (Podophrya, Ephelota, Acineta.)

## SARCODINA.

## AMOEBA PROTEUS.

Amœbæ are usually easily discernible under the low power of the microscope as irregular, semi-transparent, granular bodies. Find a specimen in the material provided, which is known to contain amœbæ, and determine the following points:

1. With the high power observe the peculiar method of loco-
motion, the constant but slow change in the shape of the body by means of projections, pseudopodia, or "false feet."

Make sketches at intervals of one or two minutes to show the changes in the form of the body.
2. Observe the peripheral zone of hyaline protoplasm, the ectoplasm, and compare this with the inner protoplasm, the endoplasm. Observe in detail the formation of a pseudopodium. Does the endoplasm extend into the pseudopodium? Can you explain how the movement is caused?
3. Find a clear space which appears and disappears at intervals; this is the contractile vacuole. Determine the length of time between successive contractions. Are the intervals regular? When the vacuole contracts what becomes of the contents? Do you know its supposed function?
4. Note the oval or rounded nucleus moving with the flowing endoplasm. What is its structure?
5. Food materials in process of digestion are readily seen. Of what do they consist? They are contained in gastric vacuoles. By careful watching, it is often possible to observe the manner in which food is ingested and the manner in which the undigested matter is egested.

Make a careful drawing of an Amœba.
Amœbæ of various kinds represent in many respects the simplest type of protozoan, and are therefore placed in the first class of these animals, the Sarcodina. The individuals of this class all possess pseudopodia, but many are quite unlike those of Amœba. Look over the figures of various Rhizopoda.

If time and material permit, study Amœba verrucosa, Arcella, and Diffugia, and compare them with Amœba proteus. Do you understand how the shells of the last two genera are made, and of what service they are? Why are not shells good for all forms?

Drawings of these forms are desirable.

## FORAMINIFERA.

With very few exceptions Foraminifera are marine and provided with shells. Empty shells from deep-sea dredgings or from
the sand beaches of such islands as the Bermudas may be had for study. Examine them with a low power by reflected light.

1. Carefully examine various shells, compare them with each other and with figures. Notice the great variety in size and shape and determine how the chambers must have been added during growth.
2. Observe a single opening in a shell, and determine whether the general surface has any finer perforations. Be sure to understand the relation of the live animal to the shell. (Refer to Calkins, pp. 71-78, for a general discussion of the shells of the Sarcodina.)

Make drawings of several types of shells.

## ACTINOSPHAERIUM OR ACTINOPHRYS.

Find, as usual, with the low power, and increase the magnification as occasion demands.

1. Note the many fine radiating pseudopodia. These are quite stiff compared with those of $A m c e b a$ and for a considerable time show little change, not being pushed out and retracted constantly as in Amaba. Is the animal flat or spherical?
2. Both ectoplasm and endoplasm are so filled with vacuoles that they present a frothy appearance characteristic of most Heliozoa. The endoplasm of all Protozoa is alveolar in structure, but in Actinosphærium the vacuoles are exceptionally large, though not as large as those in the ectoplasm.
3. The nucleus is present in the center of the organism, but it is somewhat difficult to demonstrate in the live animal.
4. At some point near the periphery, the contractile vacuole can usually be seen. When it is found notice its action, and immediately after it has contracted look among the pseudopodia of that region for indications of its extruded contents.

Draw a specimen, indicating all of the points observed.
5. When the contractile vacuole discharges, or when any foreign body touches the ends of the pseudopodia, notice the way in which this type of pseudopodium is moved. What does this indicate in regard to its structure? How far do the pseudo-
podia extend? They may be seen to contain minute granules when studied with the high power and best light.
6. If possible, observe the process of catching food with the tips of the pseudopodia and the manner in which it is drawn toward the body. Note any motion on the surface of the body as the food is drawn closer, and also the manner in which the food is finally ingested. Are there any indications that the pseudopodia extend as still finer filaments beyond the point to which it is possible to trace them with the highest magnification at hand? If the capturing of food is observed, make a series of diagrams to illustrate the process.

If possible, observe a specimen undergoing division. Draw.
It is desirable to examine Clathrulina, noting the stalk and skeleton. Look over figures.

## MASTIGOPHORA.

## EUGLENA.

Understand its habitat and with what forms it is usually associated.

1. Observe the free-swimming movements of the organism, and the euglenoid changes in the form of the body.

Make drawings showing the changes in the shape of a single individual.
2. Distinguish anterior and posterior ends. Is there any dorso-ventral differentiation? Note the motile organ, the flagellum. Where is it attached? What relation does it bear to the gullet? How is it directed during locomotion of the organism. Does it serve any other purpose besides locomotion?
3. The green color of Euglena is due to chlorophyl, and this enables the animal to live in the clearest water, being nourished like a typical green plant, but minute particles of food are also taken into the endoplasm through the gullet, and thus Euglena combines holozoic and holophytic methods of nutrition. Consider the bearing of this on the position of Euglena and its allies in the protozoan scale.
4. Note the absence of color near the anterior and posterior ends of the organism. Near the anterior end also notice the red pigment spot, or stigma. What is its probable function?
5. Stain a specimen with iodin and look for the nucleus. It is obscured by the chlorophyl.

Make a drawing showing all of the points observed.
Look through the stock cultures for other forms of Mastigophora, such as Trachelomonas, Peranema, Phacus, etc.

It is desirable to make drawings of the different forms.

## volvox.

Volvox globator is better for study than V. aurens. It may be distinguished from the latter by the larger size of the colony, the greater number of cells that compose it (about 15,000 ), the angular shape of the individual cells, and the stout connecting processes of protoplasm, into which chromatophores may enter.

Observe the movements of colonies in a watch-glass of water, with the naked eye and with a low power of the microscope.

1. Do the colonies tend to collect toward a particular side of the dish? What reason is there for the reaction?
2. Place a number of colonies on a slide with enough water to allow them to be covered without crushing them. Study first with the low and then with the high power and determine the species. Understand the relation of the individual cells to the colony. (See Parker and Haswell, Fig. 50.)

Draw a figure showing several cells and their protoplasmic connections.
3. Compare in detail an individual cell with Euglena.
4. Observe, if possible, certain cells, called parthenogonidia, which are specialized for asexual reproduction. These divide and form the daughter colonies, which become detached and swim in the interior of the parent colony. They are finally liberated by the rupture of the wall of the parent colony.

Make a figure of a parent colony that incloses several daughter colonies of different sizes.
5. V. globator is monœcious. Look for eggs and bundles of spermatozoa.

## Figure them.

6. Be sure to recognize the significance of the fact that the cells of Volvox are differentiated into somatic and germ cells, and to understand the resulting physiological division of labor. (See Calkins, p. 232.)
7. Consider the reasons for and against regarding Volvox and allied organisms as animals rather than plants.

## CERATIUM.

1. Examine this form with a high power, and in a favorable specimen notice the sculptured outer surface of the cellulose test. The living animals are green or brown owing to the presence of chromatophores in the protoplasm.
2. Note the furrow encircling the body. Does it extend completely around it? Is there a short furrow on one side at right angles to the first, or a depression of considerable size? Understand the position of the flagella.

- Draw the animal, showing the points observed.

Look for examples of the earlier stages of division, and of later stages, which appear as chains of fully formed individuals attached together.

## NOC'TILUCA.

If living specimens are not to be had for study, material preserved in alcohol, after suitable fixation, can be used. Specimens are best examined in a cell-slide under a cover-glass.

1. Observe the nearly globular shape, and on one side a groove from which arises a large flagellum or "tentacle." Is there a deep groove near it? At the bottom of this groove it is possible to see the mouth in a living specimen. Another smaller flagellum is visible in living specimens inserted at the bottom of the mouth, but in preserving the organism it is usually destroyed.
2. Note the appearance of the preserved protoplasm. The endoplasm appears parenchymatous. At one point a more com-
pact mass is seen, from which strands appear to radiate. This has been found to contain the nucleus.

Noctiluca is phosphorescent, and frequently causes very brilliant displays.

Make a drawing.

## SPOROZOA.

GREGARINA.
Remove the head and posterior end of a larval or adult meal beetle and pull out the digestive tract with a pair of forceps. Place the digestive tract on a slide, split it open lengthwise with a sharp scalpel, and then spread it out, with the inner wall exposed, and cover. The operation should be performed rapidly to prevent the material from drying. If the beetle is infected, numerous gregarines will be visible under the microscope. Study with low and high powers.

1. Does the animal move? A great number of refractive granules are present in the protoplasm. They are regarded as reserve nourishment. They can be removed with acid.
2. Note that the body is covered with a membrane, and is divided into a dense superficial layer, the ectoplasm, and a central, more fluid mass, the endoplasm.
3. The endoplasm is separated into two parts by a portion of the ectoplasm. The anterior part is termed the protomerite, and the posterior part the deutomerite. In which is the nucleus situated?
4. Is it possible to distinguish a layer of myonemes just external to the endoplasm?
5. Is there another section of the body just anterior to the protomerite? If so, this is the epimerite.

Before reproduction Gregarina throws off the epimerite, leaves it in the cell-host, and falls into the lumen of the digestive tract. It then encysts, and the protomerite and the deutomerite form one spore-producing individual. The attached stage in the life-history of Gregarina is termed the cephalont, and the detached stage, the sporont. (See Calkins, Fig. 77.)

Make a drawing.

## INFUSORIA.

## PARAMECIUM.

Place a drop of the culture on a slide, cover, and examine with the low power.

1. In an animal not closely confined note the shape and movements. Is it possible to distinguish an anterior and a posterior end? A forward and backward movement? Is one side of the animal kept constantly uppermost? Is there a dorsal and ventral surface? Do the animals change their shape either permanently or temporarily? Individuals tend to collect about air-bubbles and at the edge of the cover-glass. Why?

Indicate by a sketch all the points which can be determined with the low power.
2. Draw off all superfluous water by means of filter-paper, add a trace of powdered carmine, and then find a specimen which is narrowly confined and examine it with the high power.

The particles of carmine are taken into the body. Determine how and where. Note that the carmine collects in gastric vacuoles. What do you think is probably the nature of the fluid in the vacuoles? In watching them do you notice any definite movement of the protoplasm? Try to see the undigested material ejected.
3. Determine the arrangement of the cilia, and the nature of their motion. Is there a reversal of the direction of the stroke, etc. ? ${ }^{1}$
4. Observe the contractile vacuoles. How many are there? Is their position constant? What is their action? In compressed specimens the contractile vacuoles and their reservoirs are usually conspicuous. Note the order of appearance and disappearance of the vacuoles and reservoirs.
5. Focus carefully on the margin of the body and note a very thin outer cuticle. A thick layer, the ectoplasm, devoid of gran-

[^0]ules but containing radially arranged, minute, oval bodies, the trichocysts, is just internal to the cuticle. The inner mass of protoplasm, containing the contractile and gastric vacuoles, and small granules, is the endoplasm.
6. If possible distinguish the clear, centrally located nucleus (macronucleus).

Make a sketch showing all of the above points.
7. Kill the animal by running a drop of methyl-green under the cover-glass. What happens to the cilia? To the trichocysts?

Sketch the trichocysts with the threads protruded, and also note and sketch the macronucleus and the micronucleus.
8. Observe, if possible, animals dividing and conjugating.
9. Study demonstrations of permanently stained specimens for finer structure.

## SPIROSTOMUM.

1. Compare Spirostomum with Paramecium, noting the method of locomotion, the shape of the body, the ciliation, the buccal groove and mouth, and the large excretory reservoir, filling the posterior end of the body and in communication with the anterior end of the body by a canal.
2. Note the highly refractive, long, band-like (moniliform) macronucleus. In another species of Spirostomum the macronucleus is similar to that of Paramecium. It is desirable to examine stained specimens of the two species of Spirostomum.
3. Note the sudden contractions of the body. When these occur spiral lines appear on the surface. Can you distinguish these lines when the animal is extended? These are primitive structures (myonemes) functioning as muscles.

Make a drawing of the extended animal and a diagram showing the form when contracted.

## VORTICELLA.

Place a number of individuals on a slide and cover loosely to avoid crushing. As usual, study first with the low power and then with the high.

1. Notice that the body of Vorticella has the general shape
of an inverted bell. The covering of the body is a very thin transparent layer, the cuticle, underneath which is the peripheral layer of ectoplasm enveloping the more fluid and granular endoplasm.
2. The peristome is the rounded rim about the base of the bell.
3. The elevated and inclined area included within the peristome, and ciliated around the edge, is the disk. It is somewhat convex.
4. The marked depression between the disk and the peristome is the vestibule. It is also lined with cilia. The vestibule defines the ventral surface of the animal.
5. The gullet, a slender canal, leads from the vestibule toward the center of the body.
6. The anus occurs at the side of the vestibule. It is a temporary opening from which the undigested products are passed into the vestibule and so to the exterior.
7. Within the endoplasm are situated the clear contractile vacuole, several gastric vacuoles, the long U-shaped macronucleus, and the small round micronucleus. The macronucleus may be made more distinct by treating with methyl-green.
8. The stalk is composed of a sheath, which is continuous with the cuticle of the body, and, within the sheath, the contractile axis or myoneme, which is continuous with the body ectoplasm. Notice that this myoneme is situated within the sheath in a very loose spiral, and that the stalk quickly contracts into a close spiral when the animal is stimulated. Observe also the manner in which the peristome folds over simultaneously with the contraction of the stalk. What purpose does the contraction of the stalk serve?

Vorticella is distinguished from its allied genera by its simple unbranched stalk and also by the spiral form assumed by the contracted stalk. In which order of the Ciliata does the ciliation of Vorticella place it?

Make a drawing of an expanded individual and a sketch to show the condition when contracted.
9. Study, by means of finely powdered carmine, the vortex
currents set up by the cilia. Note how the particles are collected in the gullet, and at intervals are forced in rounded masses into the endoplasm to form gastric vacuoles. Is there a definite circulation in the endoplasm?
10. Endeavor to find several stages of reproduction by division.

Large fresh-water species of Vorticella are preferable for study, but marine species may be substituted when necessary. If time and material permit, study Lichnophora, a marine peritrichous form parasitic on Crepidula. (See Calkins, p. 208.)

## OXYTRICHA.

Infusoria belonging to the genus Oxytricha, or the genera Stylonychia, Pleurotricha, Euplotes, etc., may be used for the following study. These forms belong to the order Hypotrichida. Hypotrichous forms are the most highly organized of the class Infusoria, as well as of the entire phylum of Protozoa, and present a complexity of structure and function which is not found probably within the limits of a single cell elsewhere in the animal series.

1. In an animal which is becoming quiet, note the mode of locomotion, the shape of the body, the buccal groove, the contractile vacuole, etc., as in other forms studied. Compare the ciliation with that of other forms. Refer to Calkins, Fig. 98, and understand the relation of cirri, membranelles, etc., to cilia.

Draw, showing the structure in detail.
2. Run some methyl-green under the cover-glass. What is the shape of the macronucleus? The shape varies considerably in the different genera. Is it possible to distinguish the micronucleus?
3. Prepare a fresh slide and observe in detail the characteristic movements and manner of creeping over various objects. As the animal turns sidewise, note the marked dorso-ventral compression of the body.

Represent this diagrammatically beside the previous drawing.
It is desirable to examine permanently stained preparations for division stages, finer details of the nuclei, etc.

## PORIFERA.

Cells not differentiated to form definite organs. Water admitted through surface pores and ejected through an osculum or through oscula.

Class 1. Calcarea.
With a skeleton composed of calcareous spicules.
Subclass 1. Homocoela.
With the gastreal layer continuous so the col-。 lar cells line the whole gastreal cavity. (Leucosolenia.)
Subclass 2. Heterocela.
Gastreal layer discontinuous. Collar cells restricted to the flagellated chambers. (Grantia.)
Class 2. Hexactinellida.
With a skeleton composed of siliceous six-rayed spicules.
Order 1. Lyssacina.
Spicules separate or becoming united. (Euplectella.)
Order 2. Dictyonina.
Spicules united from the first into a firm framework. (Eurete.)
Class 3. Demospongix.
Great diversity of structure. Dominant forms of today.
Subclass 1. Tetraxonida.
Typically with four-rayed spicules. (Corticella.)
Subclass 2. Monaxonida.
Simple, usually unbranched spicules. Spongin frequently present. (Cliona, Suberites, Chalina, Spongilla.)
Subclass 3. Keratosa.
Skeleton of spongin fibers. No true spicules. (Euspongia, Aplysina.)
Subclass 4. Myxospongida.
Without skeleton. (Oscarella.)

## GRANTIA.

This form is quite common along the New England coast, where it occurs attached to rocks, seaweeds, and submerged woodwork from just below the lowest tide-mark to a number of fathoms in depth. You should visit an old wharf where specimens may be found, and study their relation to the forms with which they are associated. Specimens will be found to vary considerably in size. The largest sometimes reach an inch in length.

1. Examine a dry specimen and notice its general shape, manner of attachment, and osculum. The osculum is surrounded by a funnel of rather long spicules. Distributed over the general surface, more or less hidden by the numerous spicules, are many small pores. Their presence may be demonstrated more satisfactorily later.
2. Look for indications of budding. If your specimen does not show this, examine others.

Make an enlarged drawing of a sponge.
With a razor or sharp scalpel cut a dry specimen into halves, with a stroke from base to osculum, and notice:
3. The central cavity or cloaca.
4. Many apopyles, the inner openings of tubes that are embedded in the walls of the sponge, will be seen opening into the cloaca. Are the apopyles arranged in any order?

5 . With the low power of your microscope (with the light turned off) examine the cut wall and find that it is traversed by parallel tubes. Determine that these tubes are of two kinds.
(a) Regular, nearly cylindrical tubes that open into the cloaca through the apopyles and that bear tufts of spicules on their closed ends, at the surface of the body. These are the radial canals. It is frequently hard to see their openings into the cloaca, as the apopyles are narrow, so the section only occasionally passes through them.
(b) Smaller and less regular tubes that open on the outer surface between the clusters of spicules, and do not open into
the cloaca. These are the incurrent canals. In life there are small pores, prosopyles, that open from the incurrent canals into the radial canals. These openings are very minute and are apparently capable of being closed. They are never visible in dried material.
6. Examine thin, transverse sections of a dry sponge and determine the positions of radial and incurrent canals.

Make a drawing that will show the arrangement of the canals.
7. Examine the spicules and determine their positions as regards canals. Boil a portion of a sponge in caustic potash until only the spicules remain and examine the spicules. See if more than one kind occurs.

Draw specimens of the spicules.

## Living and sectioned material.

1. Place a living sponge in a watch-glass of sea-water, add a little powdered carmine, and examine it with the low power of your microscope for currents of water. See if particles are moving in a definite direction near the general surface and near the osculum.
2. With a moderately sharp razor cut tangential sections of the wall, as thin as possible, mount in sea-water under a cover, and examine with a low power. This will show both incurrent and radial canals in cross-section. How can you distinguish one from the other? In a favorable place look for moving flagella. Are flagella in all of the canals? In favorable situations it can be easily seen that the cells that have flagella possess collars also. (Collars may be withdrawn by cells so they protrude but slightly.) You see now what causes the current of water. Do you understand how a sponge feeds? Compare the choanocytes of the sponge with choanoflagellate protozoans.

Make a drawing showing the arrangement of choanocytes.
Examine transverse sections of a specimen that has been decalcified and stained.

1. The cloacal chamber is lined by a pavement of epithelium.
2. The radial canals are lined by more conspicuous cells, the gastral epithelium, or choanocytes.
3. The incurrent canals and the outer surface of the sponge are covered with flattened cells, the dermal epithelium.
4. In a part of the section where a considerable area of choanocytes appear in surface view, look for the prosopyles, through which the water passes from the incurrent to the radial canals. (They may not be found.)
5. Make out any structures you can in the area lying between the dermal and gastreal layers. What cells are found here?

Make a drawing of several adjacent canals to show the above points and indicate the course of the water by arrows.
6. In the stained sections, look for single ova and for spheres containing many spermatozoa, the sperm-spheres. Look also for segmenting eggs, which are frequently to be found. The ova are fertilized while still lying where they have developed, just within the choanocyte layer. Remaining in place, they undergo cleavage and develop so far as the amphiblastula stage (see figures in the text-books). They then break through the choanocyte layer into the radial canals and pass out with the current of water. Living specimens are frequently found with such embryos issuing from the osculum in the outgoing current of water. The sperm-spheres, when fully developed, also break through the choanocyte layer and, separating into their component spermatozoa, pass out with the outgoing water.

Ova and sperm are present in the same individual, and the animal is therefore hermaphroditic. Whether self-fertilization is prevented, as in many other hermaphroditic forms, by the ripening of one element before the other, has not been fully established.

If the time allows, draw ova, sperm-spheres, segmenting eggs, and embryos.

It is desirable to examine specimens of Lencosolenia, a still simpler sponge, and of some of the more complicated forms, like commercial sponges, Spongilla, Cliona, and Chalina. Why is more than a single osculum desirable in such forms? Understand the relation of the internal structure of the complicated forms to the more simple forms. What reason is there for the complication?

## COELENTERATA.

With a single continuous coelenteron or gastro-vascular cavity. With the exception of the Ctenophora all have nettle cells. There are two cellular layers and a mesoglea.

## Class 1. Hydrozoa.

Coelenteron simple, without septa. Gonads usually ectodermal. Fully formed medusæ have a velum.
Order 1. Leptolinæ.
With a fixed zoöphyte stage.
Suborder 1. Anthomedusx.
Without hydrothecæ or gonothecæ. The medusa bears gonads on the manubrium. (Hydra, Parypha.)
Suborder 2. Leptomedusæ.
With hydrothecx and gonothecx. The medusa bears gonads on the radial canal. (Obelia, Gonionemus.)
Order 2. Trachylinæ.
Without fixed zoöphyte stage.
Suborder 1. Trachymeduse.
Tentacles from the margin of the umbrella. Gonads on the radial canals. (Petasus.)
Suborder 2. Narcomedusæ.
Tentacles from the exumbrella. Gonads on the manubrium. (Æginopsis.)
Order 3. Hydrocorallina.
Massive calcareous exoskeleton. (Millepora.)
Order 4. Siphonophora.
Pelagic. Colonial. Colony usually shows extreme polymorphism of its zoöids. (Physalia.)
Class 2. Scyphozoa.
Body-wall of polyp thrown into four ridges (tænioles) which project into the coelenteron. Medusæ without velum and with gastric tentacles. Medusoid form predominating.

Order 1. Stauromedusæ.
Conical or vase-shaped umbrella. No tentaculocysts. (Tessera.)
Order 2. Peromedusæ.
Conical umbrella with transverse constriction. Four inter-radial tentaculocysts. (Pericolpa.)
Order 3. Cubomedusæ.
Four-sided umbrella. With per-radial tentaculocysts. (Charybdea.)
Order 4. Discomedusæ.
Saucer-shaped umbrella. Per-radial and interradial tentaculocysts. (Aurelia.)
Class 3. Actinozoa.
With a stomodæum, and with mesenteries extending into the cœelenteron. Fixed forms.
Subclass 1. Zoantharia.
Mesenteries and tentacles usually very numerous.
Order 1. Actiniaria.
Usually single. No skeleton. (Metridium. Sagartia.)
Order 2. Madreporaria.
Usually form colonies and always have calcareous exoskeleton. (Astrangia, Orbicella, Meandrina.)
Order 3. Antipatharia.
Tree-like. Mesenteries and tentacles comparatively few. Chitinoid skeleton. (Cirripathes.)
Subclass 2. Alcyonaria.
Mesenteries and tentacles eight in number. Tentacles branched.
Order 1. Alcyonacea.
Skeleton in the form of small, irregular bodies, frequently calcareous spicules. (Alcyonium, Tubipora.)
Order 2. Gorgonacea.
Tree-like, with calcareous or horny exoskeleton. No syphonoglyphes. (Gorgonia.)
Order 3. Pennatulacea.
Colony with one end usually embedded in the sea-bottom. (Pennatula, Renilla.)
Class 4. Ctenophora.
Single. Pelagic. Eight rows of meridional swimming plates. No nettle cells.


#### Abstract

Order 1. Cydippida. Nearly circular. Two tentacles, each of which may be retracted into a sheath. (Pleurobrachia, Mnemiopsis.) Order 2. Lobata. Compressed in the vertical plane. Two large oral lobes. No tentacle-sheaths. (Deiopea.) Order 3. Cestida. Ribbon-shaped. Two tentacles with sheaths, and numerous other tentacles. (Cestus.) Order 4. Beroida. Laterally compressed. Without tentacles. (Berœ.)


## HYDROZOA.

## HYDRA. (Fresh-water Polyp.)

Hydra, the only common fresh-water cœlenterate, is frequently found in jars of water taken from quiet pools or sluggish streams that contain lily-pads, decaying leaves, and other vegetable matter. The animals may frequently be found by examining the surfaces of submerged leaves, but it is usually better to allow such material to stand in glass jars for a day or two, as the animals then tend to collect on the lighter sides of the vessels. They are easily kept in balanced aquaria.

Examine specimens in an aquarium and find what you can about their mode of life. Do they form colonies?

Place a specimen in a watch-glass of water and examine it with a lens.

1. What is its shape and color? Is it attached? If so, by what part of the body? Notice the circlet of tentacles. How many are there? Compare notes with others and see if all have the same number. How are they placed?
2. Does the Hydra move its body or tentacles? Is it sensitive? How do you know?
3. Examine with à low power of the microscope and review the above points. You may also be able to see the mouth around which the tentacles are arranged.

Make two drawings, one showing the animal expanded and the other contracted.

Place your specimen on a slide under a cover-glass that is supported by the edge of another cover-glass, so it can be examined with a high power. Be careful not to crush it. Notice:
4. The outer layer, ectoderm. What is its color? Is it continuous over the whole outer surface? Does it vary in thickness? Are the cells of which it is composed apparently all alike?
5. The inner layer, endoderm. What is its color? If color is present, is it evenly diffused or is it collected in special bodies? Are the cells of which the endoderm is composed apparently all alike? Do they differ in appearances from those of the ectoderm other than in color? If the specimen is not deeply colored, look for flagella moving in the internal cavity.
6. Examine the ectoderm of the tentacles carefully and notice that each of the large, rounded, clear cells, the nematocysts, shows a rather indefinite streak running from its outer end, back into the interior. See if you can find the trigger (cnidocil) on any of these cells.

Draw a portion of a tentacle showing the distribution of the nematocysts.
7. Place your specimen under the low power of the microscope, carefully run in a drop of saffranin, and see if any of the nematocysts are discharged when the saffranin touches them. Examine with a high power and notice the appearance of the thread. Notice the change in the shape of the nematocysts that have discharged. See if you can find two kinds.

Make an enlarged drawing of an exploded nematocyst.
8. Examine prepared transverse sections of Hydra. Notice that the body is composed of two layers of cells, between which is an almost structureless thin layer. Do the cells of the two layers differ in size, shape, and structure? Do you find more than one kind of cell in each or either of these layers? Where are they? What are they?

Make a careful drawing of the section showing the arrangement as you see it.

Examine longitudinal sections, for differences in the character of the ectoderm and endoderm in different parts of the body.
9. Reproduction. Examine living specimens in a watchglass of water for bud formation and for sexual organs. Spermaries are just beneath the tentacles; ovaries, lower down; buds may be found at different levels. What layers of cells is involved in the formation of each of these?

Eggs are not formed at all seasons of the year and vary greatly in appearance according to their stage of development.

Make drawings of the stages of reproduction that you find.

## OBELIA.

These small, colonial animals are common on submerged or floating wood, stones, and seaweeds, where the water is rather free from sediments. With the aid of a glass-bottomed pail they, in company with many other forms, may usually be seen about old wharfs.

Note the appearance of large colonies of this form that are growing on stones or on pieces of board.

1. Notice the tree-like form of any single stem. Do the branches have a definite size and arrangement?
2. At the extremities of the branches are the single individuals, hydranths or zoöids. Each is similar to a single Hydra in certain ways, but is inclosed in a vase-like formation, the hydrotheca.
3. The latter is a continuation of a tough, membranous sheath, the perisarc, which covers each part of the whole colony.

Do you notice any modifications of the perisare below the hydrotheca? Do the modifications serve any purpose?
4. Trace the stem down to the creeping, stolon-like portion of the colony, the hydrorhiza.

Make a drawing of a colony.
5. The fleshy continuation of the zoöid down into the stalk is termed the conosarc. Is it in close contact with the perisarc?
6. In an expanded hydranth, note the mouth, the arrangement of the tentacles, and the number of tentacles. How is the
individual supported in its cup? Can you trace the colenteric cavities down through the stalks, and prove them to be continuous with each other? Motion in the fluid contents of living specimens makes this easy to observe.
7. Examine a hydranth with a high power and look for the cell-layers characteristic of cœlenterates. Determine how its tentacles differ from the tentacles of Hydra, and explode nematocysts as with Hydra.

Make a drawing of a hydranth.
8. Look for certain extremities which show neither tentacles nor any opening in the perisarcal covering. Such a condition signifies either an undeveloped hydranth or a reproductive individual. If the latter, it is considerably swollen and is termed a gonangium. The central core or cœnosare of a gonangium, the blastostyle, should be examined for medusce buds. This may require a high power.

Make a drawing of a gonangium.
9. You may find free medusæ swimming in the dish where material is kept. If you do, you should examine one, but it will not prove as satisfactory for study as a larger form, like Gonionemus, directions for the study of which are given further over.

In comparing it with Gonionemus notice the small size of the velum, and the ease with which the bell turns wrong side out, so the manubrium appears like a handle. At Woods Holl, Obelia apparently does not always liberate its medusæ, and it is not uncommon to find planulæ escaping from gonothecæ. Frequently those medusæ that are liberated have previously shed their eggs.
10. Study the cellular structure of a hydranth and of a gonangium, as seen in cross and longitudinal sections.

Make a drawing of each.
For comparison use any thecate forms, which may be offered as loose material or as demonstrations, such as Campanularia, Sertularia, and Plumularia.

## PARYPHA.

This form is frequently abundant on the piles of old wharfs, where the colored colonies form conspicuous masses just below low-water mark.

Examine the general form of a colony and note, either with a hand lens or with the naked eye, the stem, or hydrocaulus, as it arises from the branching, matted hydrorhizal portion of the colony. The parts of the colony will be seen to differ from the Leptomedusan (Campanularian) form studied, especially in branching, rigidity, hydrothecæ, and gonangia.

Make a drawing to show the formation of the colony.

1. How does a hydranth differ from the hydranth of Obelia in the matter of tentacles? Is a hydrotheca present?
2. The mouth is terminal and is situated at the end of a manubrium.
3. The short but rather large body of the hydranth passes back to the perisarc as the fleshy axis, conosarc.
4. Notice the medusa buds on the manubrium between the rows of tentacles. What is their arrangement? This is a form in which the medusæ are not set free, but remain vestigial. The gonads ripen on the partially developed manubrium of the medusa. The sexes are separate.

Make a drawing of a hydranth.
5. The arrangement of the attached medusæ is best seen in sections. In the male medusæ numerous spermatozoa will be found, while the female individuals have a much smaller number of large ova, which are likely to be in advanced stages of segmentation.

The sections show the same body layers as Hydra, and the derivation of the medusa as an outpocketing of the wall of the hydranth is evident.

Make drawings of sections of male and female reproductive organs (medusa buds).

For comparison, study Pennaria, Margelis, Hydractinia, Clava, and Eudendrium.

## GONIONEMUS.

As has been seen, the medusæ buds are usually produced from the walls of a specialized hydranth (Leptomedusæ) or from the manubrium wall of an ordinary hydranth (Anthomedusæ). A series of these buds in various stages would show the formation of the umbrella-shaped individual with the growth of the marginal tentacles around the edge. The life-history of this form is not known, but from its structure we are led to believe that it belongs to the suborder Leptomedusæ. ${ }^{1}$ It is found in considerable numbers throughout the summer in the border of eelgrass in the Eel Pond at Woods Holl, where it may be obtained with a dip-net. It is more satisfactory to study than the medusa of Obelia, as it is much larger and its movements and organs are more easily observed. In plan of structure the two are quite similar.

Put a living specimen in a flat-sided jar containing sea-water, or in a finger-bowl, with a black tile beneath, and notice:

1. Its method of locomotion. To the contraction of what part of the bell is movement due? How large is the jet of water that is delivered from the bell? Why is the jet made narrow? Does the jet necessarily leave at the center or may it be thrown from one side? Should it be thrown from one side, what would be the result?
2. Its position in the water when quiet. Why is this position more desirable than the opposite? With a needle-point prove that various parts of the body are sensitive.

With either fresh or preserved material notice:

1. Its flattened dome-shape. The convex face is called the ex-umbrella (aboral), while the concave portion is termed the sub-umbrella (oral).
2. The velum is the perforated diaphragm that partly closes in the sub-umbrella. All medusæ possessing this structure are classed as Craspedota. Do you understand its use?

[^1]3. In the center of the sub-umbrella is seen the large pendent manubrium, at the extremity of which is a wide-lipped mouth. If the medusa is alive, feed it with small bits of clam meat.
4. From the capacious sac at the base of the cavity of the manubrium, the stomach, the four radiating chymiferous tubes, or canals, lead to the periphery of the disk, where they open into the very delicate circular circumferential canal. The four radii marked out by these canals are called the per-radii. Do you understand the use of these canals?
5. The gonads hang from beneath the chymiferous tubes into the sub-umbrellar space. They are lobulated in structure, and more or less prominent according to maturity and the breeding season. The eggs or spermatozoa, as the case may be, are dehisced from these into the water directly.
6. The tentacles. Is their arrangement a radially symmetrical one? How are the nematocysts arranged on them? Look for adhesive organs on them. Of what use are such organs?

Turn your specimen with the velum side toward you and study the edge of the medusa with a low-power objective for the sense organs. These are of two kinds:
(a) The larger, round bodies at the bases of the tentacles communicate with the circumferential canal (which may possibly be seen along the edge of the bell). They are filled with a layer of strongly pigmented endoderm cells and are probably lightpercipient organs.
(b) Other small sessile and transparent outgrowths, situated between the bases of the tentacles, are the so-called otocysts, which are probably static organs.

All of the tentacles are abundantly supplied with tactile, sensory cells. There is a well-established circumvelar nerve ring (not easily determined in living material) derived from the ectoderm, also scattering nerve cells beneath the ectoderm in connection with the muscular tissue. Ex-umbrellar and subumbrellar layers of muscle fibers are also present.

Make a drawing from the side, slightly tipped, to show the velum, and another as seen from the oral surface.

## HYDROCORALLINA.

To this group belong forms that have heavy calcareous exoskeletons. While material is generally not at hand to study the polyps, it is desirable to study and sketch the characteristic forms of colonies such as Millepora and Stylaster, and to note the difference in the distribution of pores. Later you will see how decidedly these differ from the ordinary stony corals.

## SIPHONOPHORA.

Examine living or preserved specimens of Physalia, and sketch the type with reference to showing, if possible, the following structures: (a) pneumatophore, (b) dactylozoöids, (c) gastrozoöids, (d) gonodendrons, (e) tentacles. It will be well to refer to a text-book to find the positions and functions of each of these.

## SCYPHOZOA.

## AURELIA.

This form is one of the common jelly-fishes, and is found floating freely in the water. It is frequently washed up on shore. To be appreciated these medusæ should be seen as they occur at the surface of the sea, before they have been handled or injured. Frequently vast numbers may be seen together, all gently pulsating and thus keeping near the surface. The movement is very different from that of most hydrozoan medusæ, being very deliberate and graceful.

If living material is offered, study the method of locomotion and compare it with the locomotion of Gonionemus. Like the latter, the discoid animal presents ex-umbrellar (aboral) and subumbrellar (oral) surfaces, but the edges of the disk are indented, fringed with very numerous short tentacles, and a velum is wanting. What difference does the velum make in locomotion?

The ex-umbrellar surface presents little of interest. In the
live specimens, however, prove that the animal is sensitive over this area as elsewhere.

Preserved and hardened material is better than living for the study of the rest of the anatomy of this form. With a specimen in water in a finger-bowl, with a black tile for the background, find the following from the sub-umbrellar surface:

1. The shape of the animal. Is the margin perfectly circular or regularly indented? Are all of the marginal portions similar?
2. Four large, fringed oral arms or lips hang from the corners of the nearly square mouth, which is located in the center. Notice how each arm is similar to a long, narrow leaf, with the sides folded especially along their margins. Examine the arms for nematocysts. Do you understand how the animal gets its food? If the arm edges appear to be covered with dark specks and granules, examine to see if embryos may not be entangled.
3. The mouth is found to lead by a short gullet into a rather spacious stomach, which is produced in the region between each two corners of the mouth to form a gastric pouch. Determine the shape of the stomach.
4. The remaining parts of the digestive (and also in this case circulatory) system include the numerous radial canals and the single circumferential canal.
(a) Directly beneath each oral arm a per-radial canal is given off, which, at a short distance from the stomach, gives off a branch on either side. The per-radial canal proper usually continues straight to the marginal circumferential canal, without further subdivision, but the two side branches above mentioned in turn subdivide several times.
(b) From the peripheral wall of each gastric pouch three canals pass toward the margin; the middle one (inter-radial canal) branches somewhat after the manner of the per-radial canals, but the other two (ad-radial canals) continue to the circular canal without further branching. ${ }^{1}$

[^2]5. The position of the gastric pouches is made clearly manifest by the gonads, which lie on the floor of the pouches, as frilllike structures, horseshoe-shaped, with their open sides toward the mouth. The ova or spermatozoa are shed into the stomach and pass out of the mouth. Embryos in various stages of development may frequently be found adhering to the oral arms. The sexes are separate. On the sub-umbrellar surface, opposite each gonad, is a little pocket, the sub-genital pit, which opens freely to the outside. Whatever purpose this may serve, it does not function to conduct the genital products to the outside.
6. Parallel with the inner or concave border of each gonad is a row of delicate gastric filaments. These are supplied with stinging cells, and they may aid in killing live food taken into the stomach. These structures are not present in the Hydrozoan medusa.
7. At the marginal extremity of each per-radial and interradial canal there is an incision on the edge of the animal, in which there are sensory organs. In each incision find:
(a) A tentaculocyst in the form of a short, club-like structure containing a prolongation of the circular canal. At its outer extremity are calcareous concretions or lithites, and a pig-ment-spot or ocellus. Each tentaculocyst is protected aborally by a hood-like projection, and on the sides by marginal lappets.
(b) Two depressions, one above and the other below the tentaculocyst. These have been assigned olfactory functions, and are called the olfactory pits.

Make a drawing showing the profile of the entire animal, and show the structure of at least one quadrant, as seen from the oral surface.

If time permits study a developmental stage, "ephyra," and compare it with the adult.

By way of comparison, examine demonstrations of Cyanea, Dactylometra, Lucernaria, or other forms belonging to this group.

## ACTINOZOA.

METRIDIUM. (Sea-Anemone.)
Specimens are quite common on piles, as well as on rocky bottoms, and may be easily observed by means of a glassbottomed pail. Most of the observations can be made much better on specimens in aquaria, but it is desirable to see their natural surroundings.

1. Notice the shape and attachment of expanded, living specimens in an aquarium, or in a deep finger-bowl. The free end, called the disk or peristome, is fringed with tentacles, and the elongated mouth is located in the middle of this area. At one or both angles of the mouth the lips are thickened into what is called a siphonoglyph.

Make a drawing of the animal.
2. Feed a specimen with bits of mashed clam to ascertain its manner of taking in food. Drop bits on the tentacles at one time, and disk at another.

Endeavor also to determine whether there are currents constantly passing in or out of the mouth that are due to ciliary action.
3. Irritate the animal and observe its manner of contraction. When fully contracted, if the irritation is continued, threadlike structures, acontia, are thrust out through minute pores, cinclides, in the body-wall.

Make a drawing of the contracted animal.
Internal Anatomy.-Using preserved material, place the edge of a razor across the peristomial area, at right angles to the mouth-slit, and divide the animal from disk to base into halves.

1. Note the extent of the esophagus and siphonoglyphes; they lead into the coelenteric chamber. Find the extent of this chamber, and the method of its subdivision by delicate partitions, the mesenteries, or septa. Are all of the mesenteries alike?
2. Forming the free edges of the mesenteries, below the esophagus, are the convoluted mesenteric filaments, which are
secretory organs that are probably equivalent to the gastric filaments of the Scyphozoa.
3. Quite near the bases of the mesenteries are the attachments of the acontia. What relation have they to the mesenteric filaments?
4. Also located on the mesenteries, and arranged parallel to the filaments, but back from the edge a bit, are the reproductive organs or gonads. Are they found on all of the mesenteries? The ova or spermatozoa are shed into the coelenteric chamber and pass out through the mouth.

Cut one of the halves of your specimen transversely in the region of the esophagus, and study the arrangements of the mesenteries, their attachments, etc.
5. How many pairs of primary mesenteries, i. e., those attached both to the outer body-wall and to the esophagus, are there? The directive septa are those at the angles of the esophageal tube. The portion of the colenteric cavity between any two pairs of mesenteries is termed an inter-radial chamber. The space between the two mesenteries of each pair is called an intraradial chamber.
6. Carefully determine the disposition of the longitudinal retractor muscles on the mesenteries. Do they occupy similar positions on all of the mesenteries?
7. Examine the upper parts of the mesenteries for openings, septal stomata, that put the chambers in communication
8. Are the tentacles solid or hollow?

Make a drawing of a longitudinal section and another of a cross-section. Put into these all of the points of the anatomy you have seen.

If time and opportunity permit, it is very desirable that this form should be compared with specimens of the order Madreporaria, and later with the Alcyonaria. Such a form as Astrangia may easily be obtained either alive or properly preserved, and will serve to show the relation of the hard parts of the coral to the polyp. You should understand the relation of the septa
and the mesenteries, and of the polyps to each other. If specimens are at hand, compare such forms as Orbicella, Favia, and Meandrina, or any forms that show gradations from separate calices to fused groups, and understand the positions of mouths, the arrangement of the colenteric chambers, and the way in which the colony has come to its present form. You should also examine large branching colonies and determine why branches are formed and how they arise.

Examine the structure of an Alcyonarian colony and see how the polyps are placed. The structure of the expanded polyps is nicely shown by Renilla. The spicules of such forms as Gorgonia may be obtained by boiling a portion of a colony in caustic potash. What purpose do such spicules serve?

## CTENOPHORA.

## MNEMIOPSIS.

This form belongs to the group of 'animals popularly called "comb-jellies," and occurs along the coast in irregular abundance during the summer months. Specimens are very phosphorescent when disturbed, so, when they are abundant, the display caused by them while rowing at night is sometimes brilliant. They may frequently be seen during the daytime and can often be satisfactorily observed in the shade of a wharf when the water is calm.

Unmutilated, living material can be studied to best advantage, but preserved material may be had that is quite satisfactory for anatomic study.

1. In general appearance a specimen resembles a hydrozoan medusa, with its aboral surface elongated until, as a whole, it approaches the shape of a fowl's egg.
2. The broader or oral end bears two heavy terminal lobes, between the bases of which is the slit-like mouth. We may consider the elongation of the mouth to be in the antero-posterior line. A bilateral symmetry is thus evident.
3. On each lateral surface of the animal, midway between
the terminal lobes, at a short distance from the mouth, notice a small opaque spot, an undeveloped tentacle in its sheath.
4. At the aboral pole is a depression, in the bottom of which is the " sensory body."
5. Leading away from this and extending as meridional lines toward the oral pole, are eight ctenophoral rows, or swimming plates. Examine the plates with a hand-lens and determine their structure and function. A pair of rows (arising from the pit of the sensory body as a single row) extends down over each terminal lobe. Another pair passes down each lateral surface of the animal. Each of these lateral rows, after passing half-way to the mouth, changes its appearance somewhat, and leaves the surface of the body proper, being continued down one edge and up the other of a finger-like process which hangs orally, called an auricle. Each ctenophore then possesses four auricles, which are somewhat covered over by the loose edges of the terminal lobes. The fringe of cilia which borders each auricle is continuous with a fringe which extends up and down the inner, lateral edge of each terminal lobe. Do you understand how the animal gets its food?

Digestive System.-With a pipet inject a solution of carmine into the mouth opening.

1. You can then more plainly see the long ribbon-like esophagus, which leads to a very small stomach just beneath the sensory body.
2. From the stomach are given off the canals, which in a successful injection will be seen to be as follows:
(a) Two short "excretory canals," opening into the pit where the sensory body is located.
(b) Two esophageal canals, one on each side, passing down along the esophagus.
(c) Two tentacular canals, one on each side, passing to the tentacular structure of that side.
(d) Four meridional canals, each of which bifurcates. The eight thus formed pass down the animal superficially, just beneath the ctenophoral rows.

Reproductive System.-The ctenophore is hermaphroditic and ova and spermatozoa are proliferated from the walls of the meridional vessels.

A portion of a ctenophoral row should be cut off, and examined under a microscope, to ascertain the arrangement and relation of plates and cilia.

Make a drawing of a side view.
Make a diagram that will show the appearance of a meridional cross-section.

## PLATYHELMINTHES.

Body elongated, flattened and unsegmented. Anus generally absent.

Class 1. Turbellaria.
Outer surface ciliated. Free living.
Order 1. Polycladida.
Intestine complexly branched. No separate vitellaria. (Planocera, Leptoplana, Stylochus.)
Order 2. Tricladida.
Intestine with anterior median, and two posterior lateral limbs. Vitellaria numerous. (Planaria, Bdelloura, Syncoelidium.)
Order 3. Rhabdoceelida.
Simple, sac-like intestine. Body usually elongated. (Polychœorus, Microstomum.)
Class 2. Trematoda.
Parasitic. Generally with sucking disks. Welldeveloped digestive system.
Order 1. Monogenetica.
Ectoparasitic. Direct development. Three or mere suckers. (Polystomum.)
Order 2. Digenetica.
Endoparasitic. Complicated development. Never more than two suckers. (Distomum.)
Class 3. Cestoda.
Endoparasitic. Without digestive cavity. Usually having a scolex, bearing clinging organs (suckers or hooks).
Order 1. Monozoa.
Body not divided into proglottids. (Caryophyllæus.)
Order 2. Polyzoa.
Body consisting of scolex and proglottids. (Tænia, Crossobothrium.)

Class 4. Nemertinea.
Elongated, ciliated, with eversible proboscis not directly connected with the alimentary canal. Intestine usually with lateral diverticula. Anus present. (Tetrastemma, Cerebratulus.)

## TURBELLARIA.

## PLANARIA MACULATA.

This form is very common in fresh-water ponds throughout the United States. It is found during the day on the lower or shaded surfaces of stones and other submerged objects, a fact which suggests that it is nocturnal in its habits. Most freshwater planarians have very opaque bodies and their internal organization cannot be studied in the fresh specimens.

1. Notice the general shape of the body.
2. The methods of locomotion. Look for cilia.
3. The pharynx and mouth near the middle of the ventral surface.
4. The eye-spots on the anterior dorsal surface.
5. Try feeding specimens by crushing a live pond-snail and putting the fragments in the dish with them. If any of the worms are at rest, set them in motion by lifting one end of each with a bit of wood or some blunt instrument. Observe the animals at intervals of a few minutes and see if any of them begin to feed. If so, by turning them over quickly with a blunt instrument, try to see how the pharynx is used. If not successful, try turning a specimen ventral side up, and placing a small bit of snail meat on its body in the region of the pharynx.
6. Look among the specimens in the dishes on the preparation table for animals that show marks of normal fission.
7. Clean a heavy watch-glass thoroughly and pour it about two-thirds full of clean pond-water from the jar on the preparation table. Transfer all of the specimens to this dish, lifting each carefully with a bit of wood. With a scalpel mutilate them in various ways; cut one in two transversely, another longitudinally, another into several pieces of various shapes. Make
memoranda, if necessary, of the shapes of the various pieces. Carefully cover the dish and set it away. Examine the pieces with a hand-lens every twenty-four hours for the next week or ten days. If the water in the dish begins to show signs of becoming foul, transfer the pieces to a clean dish of fresh pond-water. Do not use water from the tap.

## BDELLOURA OR SYNCOELIDIUM.

Most triclads are free-living, but a few are ectoparasites. The above-mentioned forms are found upon the proximal joints of the walking legs and in the gill-books of Limulus. Owing to the absence of pigment, they are very favorable for the study of internal structure, and may be used to demonstrate the structures not observed in Planaria maculata.

1. Observe the movements of the living worms in a watchglass of sea-water; then place a specimen on a slide, dorsal side uppermost, and cover with a slip.

If any of the points of structure mentioned for Planaria have not been observed, try to find them on this form.
2. Notice that the gut with its three main branches (triclad type) and many secondary diverticula is easily recognizable. The mouth can sometimes be made out as a small circular opening leading ventrally from the posterior end of the pharyngeal sheath.

Compress the specimen as much as possible by drawing off the water with filter-paper and look for:
3. The cerebral ganglia, a bilobed structure beneath the eyespots, that appears as a slightly lighter area.
4. From the cerebral ganglia two longitudinal nerve cords pass backward, and several smaller nerves pass off in front. Examine the specimen by reflected light, looking particularly at the nervous system and pharynx. What relation have the nerve cords behind?
5. With the high power and good light, look for the watervascular tubules. The region anterior to the cerebral ganglia is a favorable place. They form a clear, branching tracery, a
little lighter than the surrounding tissue. The flicker of the flame cells can usually be seen, but they may be more easily seen in Crossobothrium. Examine chart and text-book figures of the water-vascular system.

Make a good-sized drawing of a worm, showing the above points.

Reproductive Organs.-Turbellarian worms are hermaphroditic. In this form the various organs are so crowded together that it will be best to follow each system separately. Compress a specimen under the slip and find the male organs as follows:
(a) The testes are the numerous rounded masses between the lateral branches of the gut. They are connected by means of fine tubes which cannot be seen in fresh specimens.
(b) The vasa deferentia, two large tubes, one on either side of the pharynx, that unite posteriorly near the base of the penis.
(c) The genital atrium, within which the penis lies withdrawn, is situated behind the pharynx. The penis and atrium may be considered as a replica, in miniature, of the pharynx and its sheath.

If the above structures cannot be satisfactorily seen, try preserved, stained, and mounted specimens. ${ }^{1}$

Draw the male reproductive system. Refer to charts and text-books for anything that is obscure.

The female organs are as follows:
(a) Opening into the genital atrium are the two large uteri. Each has a separate opening into the atrium but has no direct connection with any other part of the reproductive system.
(b) Place a worm ventral side up and look carefully be-
${ }^{1}$ Specimens may be readily killed by compressing under a slip, being careful to draw the excess of fluid out on one side so that the animal cannot contract, and running in killing fluid. (Sublimate acetic is good.) As soon as they become opaque white, put on enough killing fluid to float the slip off and transfer the specimens to a dish of the fixative for five minutes, then 50 percent alcohol a few minutes, 70 percent several hours, stain with borax carmine or Delafield's hematoxylin; dehydrate, clear and mount in balsam. (See directions in the appendix for making permanent preparations.)
tween the second and third anterior gut diverticula on either side of the main anterior ramus for the two ovaries.
(c) The oviducts pass backward from the ovaries, parallel to the vasa deferentia, and unite posterior to the penis. The common duct thus formed enters the posterior part of the genital atrium. The oviduct is difficult to demonstrate and it may be necessary to try both fresh and stained material.
(d) Along the margins of the animal, between the diverticula of the gut, are rounded bodies, the vitellaria. These discharge their products into the oviducts. Do you know what they are for?

Draw the female reproductive system.
Study stained and mounted specimens for any points which have not been found, and particularly examine the nervous system. Look for the marginal nerve running along the edge of the body, and for numerous transverse commissural nerves. How many of these are there? How regular is their arrangement? ${ }^{1}$

## TREMATODA.

Trematodes are flat worms which lead a wholly parasitic life, but which have retained, to a greater or less degree, those organs that characterize free-living animals. Some Trematodes are parasitic upon the outside (or ectoderm) of other animals, and are hence called "ectoparasites."

## HAEMA'T'OLOECHUS (DISTOMUM).

This form is found as a parasite in the lungs of frogs. In some localities a large proportion of the frogs are infested and several specimens are frequently found in one frog. The host of the asexual generation of this species is not known, but in a closely allied species the asexual generation lives in the pond-

[^3]snail. The living worm is cylindrical and pointed at both anterior and posterior ends. With a low-power objective note:

1. The anterior sucker, surrounding the mouth.
2. The ventral sucker, near the middle.
3. Do you find eyes?
4. The alimentary canal.
(a) Mouth.
(b) The muscular pharynx.
(c) Soon after leaving the pharynx the intestine divides into two equal branches, which pass, one on the left and one on the right side, to near the end of the body. These intestinal branches do not send out lateral branches as they do in Bdelloura.

The Water-vascular System.-A small opening will be found at the posterior end of the body from which a duct passes forward in a median position to a point a little posterior to the median sucker. Here it divides and sends a branch on either side of the worm to near the anterior end.

The Nervous System.-This is difficult to see, but on either side of the pharynx a small, deeply stained mass, the cerebral ganglia, may be visible. Three pairs of longitudinal nerves pass back to near the posterior end of the body.

Make a drawing showing the above structures as far as you have seen them.

The Reproductive Organs.-Male: Two large bodies, the testes, very definite in outline, occupy the posterior end of the worm. A duct from each, the vas deferens, passes forward, and the two unite just posterior to the point where the intestine branches. By means of a median, common duct, they open to the exterior through the male genital opening. This is situated on the ventral surface, just below the point where the intestine branches.

Female: Some of the ducts are difficult to see, and in many cases they cannot be followed, but some of the organs can be found in most of the specimens.

The ovary is a lobed organ lying a little to one side of the
middle of the animal, and just anterior to the testes. Lying against it is the sac-like oötype, into which the ovary opens. From the posterior end of the oötype the long, coiled, ductlike uterus passes backward to near the posterior end of the worm, turns and passes forward, and finally opens at a point on the ventral surface near the male opening. The uterus of an adult worm usually contains embryos and fills the body, so as to obscure the other parts.

The vitellaria consist of numerous small, rounded masses that lie near the margins of the animal. The products of these organs are emptied into the oötype through a short common duct, just ventral to the oötype. Do you know what they are for? Laurer's canal is a short duct which leads from the oötype to the exterior. Its function is doubtful.

## CESTODA.

The Cestoda are endoparasites which possess very few of those organs that are characteristic of free-living animals. They have no alimentary canal, no organs of special sense, and, except in the head, the nervous system is feebly developed. On the other hand, the organs needed for the reproduction of the species are enormously developed, so that in the more mature portions of the animal, the ovaries, testes, and accessory organs occupy nearly the whole space. Can you explain why this is true?

## CROSSOBOTHRIUM LACINIATUM.

This form passes its adult life in the intestine (spiral valve) of the sand-shark. Cestode larvæ which may be the young of this species are abundant in the cystic duct of the squeteague. How the developing eggs and embryos are conveyed from the shark to the squeteague is not known. The transfer of the larvæ from the squeteague to the alimentary canal of the shark can be easily understood.

Adult Stage.-1. Notice specimens that are attached to the wall of the intestine of the shark.
2. Observe movements of specimens in a dish of sea-water.

Do the suckers have independent movements?
3. With a low power of the compound microscope, or with a hand-lens, note that the worm is made up of a head portion, the scolex, and of numerous segments, the proglottids. What is the relative size of the proglottids in the different regions of any specimen? Where are new proglottids necessarily produced if the largest are always the oldest? Are the proglottids attached to one another with equal firmness in all parts of the body? Note their peculiar shape, and how they are connected together. In the above examination, if living material is used it will often be desirable to stretch portions of the animal very gently with your forceps.
4. Note the number and arrangement of the disk-like suckers. How are they borne on the scolex? Do you find each sucker to be entirely simple?

Draw the adult worm.
5. Cut from the head-end of a living specimen a piece consisting of a scolex and not more than one or two proglottids. Place this on a slide, cover, being careful not to compress too much at first, and examine the scolex carefully again to make sure you understand its structure.
6. Look for transparent tubes coiling about in the scolex and its suckers. Compress the specimen by drawing off as much water as possible with filter-paper, and look again for the transparent tubes. These are portions of the water-vascular system. Recall the description of this system given in the lecture or in text-books. The finer branches which lead from the main trunks are difficult to identify with certainty, but by using the high power of your microscope, and focusing just below the surface in the more transparent portions of the scolex, the flame cells may easily be seen. The "flame" appears like a short, thick whip lost in continual vibration. Find such flames and watch them carefully. If not found at once, let the preparation stand and examine in about half an hour. In the older preparation they are frequently easier to find.
7. In both scolex and proglottids of fresh specimens many clear, transparent, thread-like muscle fibers may be seen. There will also be found an abundance of clear, rounded granules of lime.
8. Watch the movements of the large, detached proglottids. Pull proglottids from the posterior end of the specimen to see how easily they may be detached. Very many tape-worms have these " motile proglottids," which in some cases remain alive for so long after being detached as to seem almost like independent animals.

Mount stained specimens of proglottids in balsam and study the reproductive organs. ${ }^{1}$

1. On one side of the proglottid the lateral genital aperture will be seen. The penis is a long, slender organ, found protruding, or lying in its sheath near the lateral aperture. The vas deferens, a long, convoluted tube, extends from the penis to the testes, which form many rounded, deeply stained structures that lie about the oval outline of the uterus. On leaving the penis the vas deferens extends toward the pointed end of the proglottid, along the side of the uterus, until it reaches a point anterior to it, where it may sometimes be seen sending branches to the testes, but is frequently lost. Throughout its length it is greatly convoluted and is generally filled with spermatozoa.
2. At the base of the penis, in the lateral genital aperture, is the external opening of the female organs. From this point a small tube, the vagina, leads to a point below the sac-like uterus, which is sometimes very large and sometimes collapsed and small. The vagina ends in a small pouch, the oötype, from which a short canal (sometimes visible, but more often obscured by the vagina, which lies above or below it) leads to the uterus.
3. The ovary consists of a large many-fingered mass in a median position, near the posterior end of the proglottid. It surrounds, more or less completely, the end of the vagina and oötype.

[^4]4. The vitelline glands occupy the posterior corners of the proglottid and may extend anteriorly along its margins, by the sides of the testes, nearly to its anterior extremity. The ducts from the vitelline glands unite and join the oötype.
5. The shell gland is a small median mass that is situated between the lobes of the ovary around the oötype.

From the lecture and figures, understand the relation of the ducts of these glands and the vagina to the oötype and uterus, how and where the eggs are fertilized, and how they are finally lodged in the uterus. Why should hermaphroditism occur in this form?

Draw a figure of the proglottid showing all of the parts you have seen.

Larval Stage.-Examine and draw a specimen of the larva found in the cystic duct of the squeteague. The scolex with its suckers at the anterior end, and the opening of the watervascular system at the posterior end, are readily seen. Compress slightly if the trunks of the water-vascular system are not easily seen. They can always be seen in preserved and stained specimens that have been killed under pressure. If you have trouble in seeing them, examine such a specimen. Do you find proglottids? Understand the relation of this larva to a true cysticercoid.

## NEMERTINEA.

Several representatives of this group are rather easily obtained. Some of these, as some species of Cerebratulus and Meckelia, are large, but they are generally unsatisfactory for anatomic study, as they are opaque and filled with a connectivetissue parenchyma that binds the organs together. Furthermore, they are especially likely to cut themselves into small pieces by contraction of muscles in the body-wall.

## TETRASTEMMA.

This small animal lives among the forms that are generally found attached to piles. Specimens can usually be found by plac-
ing scrapings from piles in a glass jar with a little sea-water and allowing them to stand from a half hour to three hours. The animals may then be found, with the aid of a lens, on the sides of the dish, usually near the surface.

With a pipet transfer a specimen to a slide, cover it, and examine with low and high powers of the microscope. Notice:

1. The shape of the body, the four eye-spots, and the ciliated grooves.
2. The straight alimentary canal. The diverticula of the intestine and the terminal anus.
3. The enormous proboscis, consisting of a large anterior eversible portion, and a smaller posterior portion that is not eversible. When the proboscis is retracted it is bent upon itself. Stylets are present in the eversible portion, near its inner end. Can you determine how the proboscis is protruded and retracted? Does the proboscis have anything to do with the digestive system?
4. Beneath the posterior eye-spots are the cerebral ganglia, from which lateral nerve cords extend posteriorly.
5. If the specimen happens to contain eggs, they will lie between the diverticula of the intestine. They are comparatively very large.

## NEMATHELMINTHES.

Body elongated, cylindrical, and not segmented. Many are parasitic forms. Anus usually present. Coelom not filled with parenchyma. The classes may not be genetically related.

Class 1. Nematoda.
Many are internal parasites, but others are found in fresh and salt water and in damp earth. Body pointed at both ends. Mouth terminal, anus ventral. (Ascaris, Trichina, Gordius.)
Class 2. Acanthocephala.
Formidable intestinal parasites. Proboscis bearing hooks. No alimentary canal. (Echinorhynchus.)
Class 3. Chætognatha.
Marine, and all but one species pelagic. With caudal and lateral fins and bristle-like jaws. (Sagitta.)

## ASCARIS.

Animals belonging to this genus are common in the intestine of the horse and pig, and are not uncommon in man. Examine specimens and see if they have any organs that would aid them in clinging to the intestinal wall. How can they retain their positions?

1. Can you determine which is anterior and which is posterior? Is there any indication of segmentation? Can the ventral side be told from the dorsal?
2. Find the mouth and see that it is bounded by three lips. Notice how these are placed and find the papillæ on the ventral ones. Find the anus and note its position. This serves also as a reproductive aperture for the male. In the female the reproductive aperture is situated about one-third back from the anterior end. It can be seen only in favorable specimens.
3. Open a well preserved or fresh specimen along the dorsal line and notice the definite colom, and the straight alimentary canal. If the specimen is a female, find the Y-shaped genital organs, the free, ovarian ends of which are slender and some-
what tangled. The position of the external genital opening has already been noted. In the male there is a single, tangled, threadlike testis, which joins the enlarged seminal vesicle that extends to the cloaca. The nervous system consists of a circum-esophageal ring, six longitudinal nerves, the dorsal and ventral of which are larger than the others, and anterior nerves. It is not easily seen.

A drawing is desirable.

## TRICHINA.

Encysted specimens may frequently be found by examining thin pieces of pig muscle obtained from the meat market. Pigs fattened in small pens and fed on table waste, or in slaughteryards and fed on the offal of butchered animals, are much more likely to be infected than others. Scavenger rats and cats are frequently infected.

1. Flatten a piece of muscle containing trichinæ between two slides in a little glycerin and notice the relation of the animal to the muscle fibers. Notice the cyst that surrounds it and see if you can determine whether this was formed by the host or the parasite. There are frequently fat cells at the ends of the cyst. Just after the parasites are encysted, the cysts are surrounded by capillaries that may be injected by injecting the vessels of the host. These may be found only at a definite stage after encystment. Why are they formed? Do they indicate how the cysts were formed? If the trichinæ are abundant see if you can find more than one in a cyst.
2. Notice the shape that is assumed by the parasite. Is the coiling always the same? If your material is fresh, mount some of the muscle between slides without glycerin, warm the slide, and see if the encysted animals will move.
3. Are the anterior and posterior ends alike? Is there any indication of a mouth? The large cells that form the intestine can frequently be seen. It should be borne in mind that the encysted specimen is not fully adult and that the animal grows after reaching the alimentary canal of the next host.

Make a drawing of an encysted animal.

## TROCHELMINTHES.

Minute animals whose adult structure seems to be related to that of the trochophore larva. Mouth usually surrounded by a circlet of cilia. Three classes (Rotifera, Dinophilea, and Gastrotricha) are referred to this phylum, but they may not be genetically related.

## ROTIFERA.

Mostly fresh-water forms, but a few are marine. All are of microscopic size. The pharynx is provided with a masticatory apparatus, and the anterior end bears a trochal disk. Most rotifers are free, but a few are permanently attached, and some, as Melocerta, live in tubes of their own formation.

## BRACHIONUS (A Rotifer).

These animals are frequently quite abundant in ponds and aquaria. They are not very active, and spend most of their time near the bottom among the plants and debris. Owing to their minute size, they must be studied with a high power of the microscope.

1. The body is divided into a trunk, which is inclosed in a transparent cuticular lorica, and a movable tail or foot. The tail is tipped with two processes which form forceps, by means of which it attaches itself to plants. Can you see how these are used? Why does the animal need to attach itself?
2. Projecting anteriorly from the lorica is the retractile trochal disk. Notice the cilia on the margin of this disk. Is the disk used in locomotion? Does the animal always move when the cilia are active? What other use has the disk? Is the animal entirely dependent upon the cilia of the disk for locomotion?
3. The mouth is at the ventral border of the trochal disk and leads by a short buccal cavity to the mastax, which is a muscular apparatus provided with three chitinous trophi (a median incus and two mallei). It is used in grinding the food. The grinding movements are easily seen. A very short gullet leads from the mastax to the large stomach. The intestine is short and thick and opens into a cloaca. The anus is near the base of the tail, on the dorsal surface.
4. The reproductive and excretory systems are not easily seen. An ovary and a large vitellarium are present. The oviduct opens into the cloaca. Two long nephridial tubes open into a contractile vesicle that in turn opens into the cloaca.
5. There is a single ganglion in the anterior dorsal region, immediately beneath two red eye-spots. Anterior to the eyespots is a dorsal feeler, which is a tactile organ.

There are many common rotifers that have no lorica and some of them have the trochal disk two-lobed.

## MOLLUSCOIDA.

Lophophore present. Mouth and anus closely approximated. Colom usually present.

## Class 1. Polyzoa.

Usually colonial. Zoöids of small size and protected by a firm cuticle.
Subclass 1. Ectoprocta.
Anus outside of the lophophore.
Order 1. Gymnolæmata.
Marine. Circular lophophore. No epistome present. (Bugula, Membranipora.)
Order 2. Phylactolæmata.
Fresh-water. Horseshoe-shaped lophophore. Epistome present. (Plumatella, Pectinatella.)
Subclass 2. Endoprocta.
Colonial or solitary. Anus and mouth both inside of the lophophore. (Loxosoma, Pedicellina.)
Class 2. Phoronida.
Marine. Solitary. Lophophore horseshoe-shaped with each end coiled. (Phoronis.)
Class 3. Brachiopoda.
Marine. Solitary. Bivalve shell. Usually attached by a peduncle.
Order 1. Inarticulata.
Valves not united by a hinge. (Lingula.)
Order 2. Articulata.
Valves hinged. Usually with a shelly loop to support the lophophore. (Terebratulina.)

## POLYZOA. <br> BUGULA.

The colonies are very common in shallow water along shore, attached to rocks and piles. They may be examined with the aid of a glass-bottomed pail in the positions they occupy on the
sides of the piles of almost any old wharf. What must be the source of their food? What part of the colony is likely to be best nourished? Collect specimens by scraping the piles and see what forms are associated with them.

1. Examine a colony in a dish of water and see how it branches. Does it present any regularity?

Make a drawing of a colony.
2. Remove one of the flat branches, place it in a watch-glass of water, and examine it with a low power. What more can be observed regarding the branches? How are the cups arranged? Are the cups on the two sides of a twig placed in definite relations to each other? Where are the empty cups found? Why? Can you find connections between the cups of the two sides?

Make a drawing showing the arrangement of the cups.
3. Allow a living branch to remain undisturbed for a few moments and with a microscope see how the thin outer margins of the cups are unfolded as the zoöids protrude.
4. Mount a specimen on a slide, cover, and compare the tentacles of an expanded zoöid with those of the hydroids that you studied. How do they differ? How must the animal feed?
5. How are the tentacles arranged around the distal end of the body? How many tentacles are there? Look for the mouth.
6. Can you see the parts of the alimentary canal? Is there food in the stomach? How does the zoöid pull itself back into its cup?
7. Look for avincularia and observe their movements and structure. Where is the jaw hinged? Where are the muscles that open it? Where are the muscles that close it? Of these muscles, which are largest? Why? See if "sense hairs" can be found between the jaws. What is their probable use?

Draw an avicularium.
8. Occia with embryos will be found in some specimens. Where are they placed?
9. Put powdered carmine in the water with a living branch and see if the zoöids will eat it.
10. Put a small living branch in a drop of sea-water under a supported cover-glass and see if any of the zoöids will expand. If any do expand they may be examined, with a high power, to good advantage.

Study specimens that have been killed while expanded. Stain with iodin, wash in water, mount in glycerin, study with a high power. Find the retractor muscles, the funiculus, germ cells, and, if possible, the shape of the alimentary canal. As the alimentary canal bears a definite relation to the position of the zoöid on the branch, its shape can be readily determined only when the branch happens to be twisted so the zoöid is to be seen in side view.

Make a drawing showing the structure.
If time permits it will be desirable to examine an incrusting form to determine its method of branching and the way in which the cups are closed.

## PLUMATELLA. ${ }^{1}$

If the zoöids of this fresh-water form will expand in a watchglass of fresh water, notice the shape of the lophophore and the position of the epistome. In such a specimen the ganglion may be seen as a rounded mass just beneath the lophophore, between the mouth and the anus. Study the statoblasts with a microscope.

## BRACHIOPODA.

## TEREBRATULINA.

Examine specimens on the demonstration table and notice:

1. Shell. The difference in the size and shape of the two valves and their position in relation to the body. How are the valves articulated? How are they opened?
2. Peduncle. Its position. What is its use?
${ }^{1}$ Slices of the large gelatinous form, Pectinatella, placed in watchglasses of fresh water, make very satisfactory objects for study, as the zoöids will soon expand, and they are then in the best possible position for study.
3. Muscles. Those used in opening and closing the shell.
4. Lophophore. Consisting of two elongated arms with a double row of tentacles on each.
5. Mouth. Notice its relation to the grooves running between the rows of tentacles on each of the arms of the lophophore.

## ECHINODERMATA.

Radially symmetrical animals, with calcareous plates in the integument. Water-vascular system always present.

Class 1. Asteroidea.
With radiating arms not sharply defined from the central disk. Ambulacral feet in grooves on the oral side.
Order 1. Phanerozonia.
With large marginal ossicles. (Astropecten.)
Order 2. Cryptozonia.
Marginal ossicles inconspicuous. (Asterias.)
Class 2. Ophiuroidea.
With slender radiating arms sharply defined from the central disk. No ambulacral grooves.
Order 1. Ophiurida.
Arms not branched. (Ophiura.)
Order 2. Euryalida.
Arms branched. (Astrophyton.)
Class 3. Echinoidea.
Globular, or somewhat disk-shaped, spiny bodies.
Shell or test composed of close-fitting plates.
Order 1. Regularia.
Nearly globular test. Spines rather large. Mouth and anus polar. Jaws present. (Arbacia, Strongylocentrotus.)
Order 2. Clypeastroidea.
More or less flattened test. Spines very small.
Anus not polar. Jaws present. (Echinarachnius.)
Order 3. Spatangoidea.
Somewhat flattened and elongated. Spines very small. Neither mouth nor anus polar.
Class 4. Holothuroidea.
Bodies soft, elongated and cylindrical. Mouth and anus polar, the former surrounded by a circlet of large oral tentacles.

# Order 1. Elasipoda. <br> Well-marked bilateral symmetry. Tube feet on ventral and papillæ on dorsal surface. Deep sea only. 

Order 2. Pedata.
Ambulacral feet in rows or scattered. (Thyone, Cucumaria.)
Order 3. Apoda.
Without tube feet. Worm-like. (Synapta.)
Class 5. Crinoidea.
Temporarily or permanently attached by a stalk. With five branching arms radiating from a small disk.
Order 1. Neo-Crinoidea.
Characters as above. (Antedon, Pentacrinus.)

## ASTEROIDEA.

## ASTERIAS. (Starfish.)

Starfishes are rather common along most coasts and are among the worst enemies of oysters. They can generally be most satisfactorily examined on shallow-water mussel-beds or on rocks covered with barnacles. Places where starfish occur should be visited, and the conditions under which they live examined. Determine:

1. Upon what and how they feed.
2. What their enemies must be.
3. How their arms are repaired when injured. Do you find specimens that are growing new tips to injured arms or are such arms apparently replaced? When an arm is injured how must the animal proceed to repair it?
4. Do specimens ever conceal themselves? See if specimens can be found with pieces of grass and weeds covering them. Try picking these pieces off to see if they adhere.
5. Do the animals have other means of protection?

Examine a specimen and notice:

1. That the surface by which the animal clings, the oral surface, is different from the other, aboral surface, and that
both surfaces are covered with short spines. What is the use of the spines?
2. It consists of radiating arms and a central disk.
3. On the aboral surface of the disk, near the junction of the two arms, a small, frequently conspicuously colored, circular body, the madreporic plate. The two arms adjacent to this plate are sometimes referred to as the bivium, and the remaining three as the trivium. The radial symmetry of the animal is disturbed externally only by the madreporic plate. Examine this plate with a lens and determine its structure.
4. On the oral surface, the mouth. Note its size and see if it is provided with jaws of any kind. Would you expect jaws? Why?
5. Radiating from the mouth are the ambulacral grooves, one on each arm. In these grooves are the ambulacral or tube feet. Do they have a definite arrangement? Along the sides of the grooves are slender spines that differ from the general body-spines in being movable.
6. Scrape the tube feet from a portion of an ambulacral groove of a dried specimen and notice the pores through which the feet are attached to organs inside the arm. Notice also the exposed ambulacral plates and determine their relation to the pores.

Draw figures of the aboral and oral surfaces of a starfish, and a diagram to show the relation of the ambulacral plates and pores.

Place a living starfish in a dish of sea-water.

1. Study its method of locomotion. How are the ambulacral feet used? How far can they be protruded?
2. Tear the starfish quickly from the bottom. Do any of the feet remain behind? Understand how they are attached.
3. Place the starfish on its aboral surface and watch it turn over.
4. Find the thread-like dermal branchice projecting through the body integument. Are they withdrawn when touched? What is their function?
5. Stroke the starfish with a camel's-hair brush and notice how the hairs are caught. Can you determine by what and
how they are held? With a hand-lens examine around the bases of the spines, and see the arrangement of the pedicellaric. Their function is obscure, but they enable the starfish to hold small objects firmly and they may be of service in dealing with possible surface parasites.
6. Remove some of the pedicellarice with forceps and examine them under the microscope. Is there more than one kind?

## Draw a pedicellaria.

Internal Structure.-Make the dissection under water, and in cutting through the integument be careful not to injure the underlying soft parts.

With strong scissors cut through the aboral body-wall near the tips of the rays of the trivium. Carry the cuts forward along the sides of the rays to the disk.

Lift up the integument at the tip of each arm and carefully snip away the mesenteries which attach the organs to it. Cut the membranes that extend into the disk opposite the junctions of the arms, and remove the three-rayed flap of integument thus freed, cutting as close as possible to the madreporite, but leaving this in place.

Digestive System.-In studying this system you should constantly bear in mind the peculiar method by which the animal feeds, as the digestive system is highly modified to suit this method.

1. The short, cone-shaped intestine and the intestinal cжеса were probably removed with the integument. The intestine probably does not function, and may be regarded as a vestige. It opens near the center of the disk, on the aboral side, by a very minute anus that is very hard to see.
2. The stomach, which occupies the greater part of the space in the disk, is composed of a small aboral portion, the pyloric division, that receives the ducts from the hepatic cæca, and a larger, lobed, cardiac division, into which the mouth opens. The cardiac portion may be everted through the mouth, thus being turned wrong side out. Five pairs of muscles, which draw this portion of the stomach back into place, may be seen at-
tached to the ridges formed by the ambulacral plates in each arm. How is it possible for the stomach to be everted? What reason is there for two divisions?
3. In each arm is a pair of long, glandular organs, the hepatic cocca. The ducts of each pair unite and join the pyloric division of the stomach by a common duct. These are digestive glands. What reason is there for having ten enormous digestive glands? Does this have anything to do with the method of feeding?

Make a drawing of the digestive system of the disk and one arm.

Reproductive System.-Turn the hepatic cæca to one side and notice the ovaries or testes. The sexes are separate, but the organs have the same general appearance in both sexes. They vary in size according to the season of the year, sometimes being so small that they are not easily found, and again being nearly or quite as large as the hepatic cæca. With a pair of forceps lift up one of these organs and see where it is attached. It is at this point that the reproductive cells reach the exterior. How many gonads are there?

Draw the gonads into anothcr arm of your figure.
Water-vascular System. ${ }^{1}$ - 1 . Carefully remove the side of the stomach next to the bivium, being very careful not to disturb the stone canal, which runs from the madreporic plate to the margin of the membrane around the mouth. By the side of the stone canal is a thin band of tissue formerly supposed to be a heart. It is now generally believed to be connected with the reproductive system, and is frequently referred to as the axial organ. It has nothing to do with the system now under consideration.
2. The circular canal, which is joined by the stone canal at the outer margin of the peristomial membrane, follows the margin of the membrane and so encircles the mouth. Originating

[^5]from it at points very near the ampullæ of the first tube feet are nine small vesicles, Tiedemann bodies. They are smaller than the ampullæ and project in toward the mouth. The position where the tenth Tiedemann body might be expected, is taken by the stone canal.
3. Leaving the circular canal are five radial water tubes, one for each arm. These tubes lie along the oral surfaces of the ambulacral plates, and are accordingly not visible on the inside of the animal. The position of the tube can best be understood by making a transverse section of an arm. It will then be seen either in injected or uninjected specimens, lying immediately below the ambulacral plates. In injected specimens it may be followed by dissecting from the oral side, from the circular canal to the extremity of the arm, where it ends in a small tentacle.
4. Along the sides of the ambulacral ridges, within the bodycavity, are rows of little bag-like ampullce. Determine their relation to the ambulacral pores. If the specimen is fresh, press a few ampullæ and see if the corresponding tube feet are affected. Can you determine their function? In a dissection it is hard to find the connecting tubes that join the radial tubes to the tube feet, but they can sometimes be seen in sections of arms of injected specimens. They can readily be seen in microscopic preparations.

The water-vascular system is very distinctive for the Echinodermata, and you should understand perfectly:
(a) How the tube feet are extended.
(b) What causes them to adhere.
(c) The connection between tube feet, ampullæ, connecting canals, radial water tubes, circular canal, stone canal, and madreporic plate.
(d) How it is possible to extend one foot without extending others.

Make a drawing showing the arrangement of the water-vascular system.

Nervous System.-This is not easily studied by dissection.

It consists of a nerve ring which encircles the mouth and lies just ventral to the circular water canal, and five radial nerves that extend down the arms just beneath the radial water tubes, to end at the tips of the arms in pigment spots, the eyespots. The whole central nervous system is superficial and forms a portion of the outer covering of the body. The radial nerves can be seen by separating the rows of ambulacral feet, but it is much more satisfactory to study them in prepared sections.

Muscular System.-Examine the walls of the starfish for its muscular system. If time permits, it will be desirable to macerate a portion of an arm to see the skeleton to which these muscles are attached.

Study prepared sections of the arm of a small starfish and determine the relation of organs.

1. The hepatic cæca. How are they supported? What is their structure?
2. The radial canal, connecting tubes, tube feet, and ampullæ.
3. The thickened, deeply stained, radial nerve between the tube feet and below the radial water tube.
4. The perihæmal canal, divided by a thin partition, that lies between the radial water tube and the radial nerve.

Make a drawing of a section of an arm that will show these points.

Understand how a starfish can open an oyster or a mussel and how it digests it when open. How can it digest a barnacle or a small snail? How does it respire?

## OPHIUROIDEA.

## OPHIURA. (Serpent-Star.)

These animals live more or less concealed in crevices, shells, eel-grass, etc., and may be obtained either by dredging or by pulling a dip-net through eel-grass. They are not conspicuous objects along the shore, as are starfish, and they differ essen-
tially from starfish in their method of locomotion and their method of feeding.

Examine a specimen and notice:

1. The appearance of the disk and arms. Are the spines similar to those of Asterias? The arms are more flexible. In what direction do they bend easiest?
2. Do you find a madreporic plate?
3. The size and shape of the mouth.
4. The ambulacral groove. Is it distinct?
5. The ambulacral feet. Do they have suckers? How are they arranged?

Draw an oral view of a specimen.
Place a living specimen in a dish of sea-water and watch its movements.

1. Compare the rate and method of movement with Asterias.
2. Are all of the arms used in progressing in the same way?
3. See if the arms can be used interchangeably or if a certain one is always directed forward.
4. Are the ambulacral feet of any service? Do they adhere? The internal structure shows that the stomach is not eversible and that the hepatic cæca do not extend into the arms. Is there any correlation between these two facts?

The nervous and water-vascular systems are very similar to those of Asterias, but here the former lies within instead of on the surface of the arm, the entire arm being encased with four or more rows of shields. They can be studied best in sections.

## ECHINOIDEA.

## ARBACIA. ${ }^{1}$ (Sea-Urchin.)

In some localities sea-urchins can be found in tide pools or near low-tide mark, where they may be-very abundant. In other localities they can be obtained only by dredging. When possible they should be observed in their native places and the conditions noted.

[^6]1. What apparently serves as food for the animal? Can you determine how this is obtained?
2. Do you find attempts at concealment?
3. Are the animals able to climb?

Put a living sea-urchin in a dish of sea-water and study its movements.

1. When placed on its back, how does it turn over?
2. What is the normal method of progression?
3. How are the spines arranged when the animal is creeping on the bottom?
4. What difference do you note between the spines on the lower and upper surfaces?
5. How long are the tube feet? Are they used with the spines in moving or do both sets of organs act independently?
6. Grasp a spine with your forceps and see if neighboring spines respond. Do they form a good defensive armor?
7. In what directions may a spine be moved? Remove a spine from a preserved specimen and determine how it was attached and how the muscles that moved it were attached to the spine and to the test.

Make a diagram showing the arrangement.
8. Do the spines have any definite arrangement?
9. By means of the tube feet, notice that there are five ambulacral areas, between which are five inter-ambulacral areas.
10. Notice an area on the aboral surface which is free from spines. This is the periproct.
11. Notice the membrane around the mouth, the peristome.
12. Look for pedicellarice on the peristome. In what other places are pedicellarix found? Do they differ from those of the starfish?

Draw one.
13. Notice the tentacles (modified tube feet) on the peristome.
14. The dermal branchice are shrub-like appendages at the outer edge of the peristome. They are situated opposite each inter-ambulacral area.

Skeleton. ${ }^{1}$-Examine the aboral surface of a cleaned "test" and note:

1. The periproct has four triangular plates which cover the anal opening. (Scattered plates in Strongylocentrotus.)
2. Around these anal plates are five large ones, that form the apices of the inter-ambulacral series of plates. These are the genital plates, and each is perforated by a small opening, the genital pore.
3. That one of the genital plates is larger than the others and is full of very minute pores. This is the madreporite, which is homologous with the madreporite of the starfish. Determine its structure with a lens.
4. Between the genital plates are five smaller ocular plates, also perforated, which form the apices of the ambulacral series of plates. These plates and the genital plates, together form what is known as the apical system.
5. In the ambulacral series of plates, the arrangement of the openings (ambulacral pores) through which the tube feet protrude.
6. Do all of the plates bear balls to which spines were articulated? Are the balls of equal size? Do they have a definite arrangement?

Can you homologize the positions of the ambulacral, interambulacral, ocular, and genital plates in the sea-urchin and starfish? What portion of the starfish is represented by the periproct of the sea-urchin?

Make a drawing of the test, showing the ambulacral, interambulacral, and apical systems of plates.
7. Around the peristome, on the inside of the test, note the five auricles forming arches or bridges over the bases of the ambulacral areas. Their purpose will be seen later.

[^7]With a scalpel or strong scissors, cut around the equatorial region of an alcoholic specimen, taking care to cut through the test only. Break or cut the aboral portion away bit by bit until near the genital plates, freeing the fragments from the internal organs without disturbing their positions.

Reproductive System.-How were the gonads (their appearance is the same in both sexes) attached to the test? How many are there? Opposite what areas of the test are they placed? Where do they open to the exterior?

Digestive System.-Remove the gonads from the three areas farthest from the madreporic plate, lift the remaining aboral portion of the test slightly, and examine the alimentary canal.

1. The large and conspicuous jaws, frequently called the lantern. They will be studied later.
2. The esophagus, passing between the jaws, and bending over to one side to join the intestine.
3. The intestine. Notice its size and its shape. Do its loops have any relation to the positions of the gonads?
4. The intestinal siphon, lying along the intestine and attached to it at both ends.
5. The rectum, running from the end of the intestine to the anus.
6. The mesenteries which hold the various organs in place.

Make a drawing to show the reproductive and digestive organs.
Water-vascular System.-1. The stone canal leads from the madreporite to the circular canal, which encircles the esophagus at a point just above the lantern.
2. From the circular canal radial tubes pass over the top and down the sides of the lantern, to pass through the auricles and up the ambulacral tract, to the ocular plates. They can be easily seen along the sides of the test, but are difficult to see before they leave the lantern.
3. Along the course of each radial canal, the ampullar, which supply the tube feet, are to be seen. The relations of the tube feet and radial canals are practically the same as in the starfish except that the removal of the radial tubes to the inner sides
of the ambulacral plates causes two perforations for each foot here, while the starfish has only one. One of these perforations is for the connection between the ampulla and the foot, the other is for the connecting tube between the radial canal and the foot. The connecting tube joins the foot outside of the plate (as in the starfish), while it joins the radial canal inside of the plates (different from the starfish).

Remove the intestine and study the lantern and its attachments.

1. In shape the lantern is a five-sided pyramid, having the "teeth" at its apex projecting through the peristome. The base of the pyramid may be compared with a wheel, in which the five epiphyses are the tire and the five radially directed rotu$l x e$ are the spokes, each one of which has a more slender bar, forked at the free extremity, the radius lying over it. Each of the five segments represents a jaw that is articulated to its neighbors at its base, near the esophagus. The points of the teeth can thus be separated and closed, and the jaws protruded and retracted by means of muscles.
2. The whole lantern is inclosed in a delicate membrane, the peripharyngeal or lantern membrane.
3. Connecting adjacent alveoli from top to bottom are the inter-alveolar muscles, that by their combined action close the jaws.
4. To each of the arms of the radius fork a muscle is attached. Where is it attached at the other end?
5. A pair of protractor muscles pass down from each epiphysis. To what are they attached? They are used in protruding the jaws?
6. A pair of retractors is attached to the tip of each alveolus. They can be used in opening the jaws or in retracting the jaws. Do you see how?

Understand how the jaws may be protruded, opened, closed, and retracted by means of these muscles.

Make a drawing to illustrate the arrangement of the muscles.
7. Remove the lantern by cutting the peristome, clear away the external tissues, and examine its construction. With a scalpel cut the inter-alveolar muscles so the jaws may be separated. Find:
(a) The large V -shaped alveoli (a straight suture indicates that each is formed by the fusion of two parts). Notice the roughenings on their esophageal sides. What purpose can they serve? Why should the alveoli be so large and the inter-alveolar muscles so strong?
(b) The epiphysis, which is fused with the upper corners of each alveolus and extend in to form a bar over its base, thus being functionally a part of the alveolus itself. The sutures between it and the alveolus proper can usually be seen.
(c) The rotula, one of which joins the ends of each epiphysis and extends to the position of the esophagus. The five rotulæ of the lantern articulate with each other around the esophagus, and each rotula articulates with the epiphyses of two adjacent jaws. Do you understand how the jaws move on the rotulæ?
(d) The radii,lying over the rotulæ, are slender and bifurcated at their outer ends.
(e) Inclosed in each alveolus is a tooth. Examine both extremities of it and determine why the inner end is soft.

Understand thoroughly how the jaws are used and why the animal needs them. Why does the sea-urchin not need large hepatic cæca?

The Nervous System.-The nervous system is difficult to demonstrate in dissections, but is easy to trace in sections. It consists of:

1. A nerve ring that encircles the esophagus at a point just above the mouth.
2. Five radial nerves that pass from the ring, along the inside of the ambulacral areas of the test, to the ocular plates.

The radial water tubes will be found in sections adjacent to the radial nerves. The two are separated only by a narrow space, the pseudohæmal canal. Between the radial nerves
and the tissue of the test there is another narrow cavity, the epineural sinus.

If time permits, students will find a dissection of the sanddollar, Echinarachnius, valuable for purposes of comparison. Special notes will not be necessary. Its shape and restricted ambulacral areas should be studied in the light of its habits and food-supply. How does the animal move?

## HOLOTHUROIDEA.

## THYONE. (Sea-Cucumber.)

These animals may be found in protected and usually muddy places, concealed in eel-grass. They are generally so effectually concealed that they cannot be satisfactorily studied in their native places. It is desirable to visit places where they occur and find specimens by feeling for them near the bottom. It is then possible to realize the life for which they are adapted.

Examine living expanded specimens in an aquarium (taking care not to disturb them) and note:

1. How the tentacles are used. What kind of food would it get by this means? Compare the method of food-getting with the starfish and sea-urchin.
2. The respiratory movements of the body. Notice the strength of the current of water ejected.
3. The general shape of the body when expanded. Does it seem to rest on a particular side?
4. The number and arrangement of the tentacles. To what do they probably correspond in the sea-urchin?

Kill the specimen by catching it with strong forceps behind the mouth, when the tentacles are expanded, and holding it in hot water. Note that:

1. The body is covered with papilliform ambulacral suckers. It is possible in some cases to see that they are arranged in five broad, longitudinal bands.
2. The suckers are less abundant on the dorsal (upper) surface than on the ventral.
3. A small papilla is to be found on the dorsal surface, between the tentacles. On it is the genital opening. This will be referred to again.

Make a drawing of the animal as seen from the side, indicating all of the points of structure that have been seen.

With a pair of scissors, open the animal longitudinally along the middle of the ventral (lower) surface.

Digestive System.-1. Note the delicate perforated mesentery, which attaches it to the walls of the body.
2. The esophagus, leading from the mouth through a cartilaginous structure, which recalls the lantern of the sea-urchin. Examine and see if the arrangement is similar to that of the sea-urchin lantern.
3. The thin-walled and enlarged stomach.
4. The coiled intestine, which leads to the cloaca.

Draw the alimentary canal in position.
Cut the alimentary canal just in front of the stomach, and close to the cloaca, and as you remove it notice the blood-vessel that runs along the intestine.

Respiratory and Excretory System.-Arising laterally from either side of the cloaca are the two respiratory trees. They are branched and project far forward into the body-cavity. With a pipet inject them with starch-mass. The strong jets of water ejected by the living specimen were thrown from these tubes. Can you understand how they serve for respiration? The walls of the tubes composing the trees are glandular and may thus serve to excrete wastes.

Make a drawing of the cloaca and respiratory trees.
Reproductive System.-The single gonad (ovary or testis) occupies a median dorsal position in the anterior part of the body-cavity. It is composed of a multitude of filaments, which join to make a brush. This brush projects backward into the body-cavity. The duct of the organ lies along the dorsal midline, between the two muscle bands, and leads to the opening
upon the small papilla near the mouth that has already been noticed.

Water-vascular System.-1. The circular canal can be found in favorable specimens, surrounding the deeper portions of the esophagus. It gives rise to one or two Polian vesicles, which are very large and hang down into the body-cavity.
2. Ten forwardly directed canals leave the water-ring and pass into the tentacles, which may be homologized with tube feet.
3. The five radial canals (homologous with the radial canals of the starfish and sea-urchin) originate from the water-ring and pass backward, ending near the cloaca. The course of each radial canal is easily followed by means of the numerous small, elongated ampullæ which supply the tube feet.
4. The stone canal and madreporite are much reduced in holothurians. The madreporite, except in larvæ and very young specimens, is not found on the outer surface. The stone canal leads obliquely backward from the water-ring, toward the bodywall, to join a small calcareous body, the madreporite, which lies in the body-cavity. Does this give you a reason for the presence of large Polian vesicles? The liquid in the water-vascular system is not sea-water. Notice its color.

Make a diagram of the water-vascular system.
The classes of the Echinodermata show exceptionally well how a general type of structure may be retained and still modified in certain regards for special habits. Compare, for instance, the feeding habits of the starfish, sea-urchin, and sea-cucumber.

## ANNELIDA.

Body elongated, generally divided into somites. Colom usually extensive. Appendages when present form parapodia.

Class 1. Archi-annelida.
Without setæ or parapodia. Nervous system not separate from the epidermis. (Polygordius.)
Class 2. Chætopoda.
With numerous, distinct somites that are provided with setæ.
Subclass 1. Archi-chætopoda.
Setæ retractile. Nervous system not separate from the epidermis. (Saccocirrus.)
Subclass 2. Polychæta.
With numerous sete. With a great variety of structure. (Nereis, Autolytus, Lepidonotus, Diapatra, Chætopterus, Cistenides, Spirorbis, Clymenella, Sabella, Hydroides, Arenicola, Amphitrite.)
Subclass 3. Myzostomida.
Disk-shaped. Without external segmentation.
Parasites on Echinodermata. (Myzostoma.)
Subclass 4. Oligochæta.
Without parapodia. Setæ few and simple.
(Tubifex, Lumbricus.)
Class 3. Gephyrea.
No segmentation. With or without setæ. With introvert or proboscis.
Order 1. Inermia.
With introvert. Anus dorsal. No setæ. (Phascolosoma.)
Order 2. Armata.
With proboscis. Anus posterior. Setæ few. (Echiurus.)
Class 4. Hirudinea.
Somites constant in number, with more external annuli than there are somites. With sucking mouth and posterior sucker.

Order 1. Rhynchobdellida.
Anterior end of body forming a proboscis or introvert. No jaws. (Glossiphonia, Pontobdella, Clepsine.)
Order 2. Gnathobdellida.
No proboscis or introvert. Mouth usually with three teeth. (Hirudo.)

## CHETOPODA.

## NEREIS VIRENS. (Clam-Worm.)

These animals may be found inhabiting mud-flats from which the water flows at low tide. Occasionally they may be seen with their head ends protruding from their burrows, but generally specimens will have to be dug. Notice the conditions under which the animals live and the forms with which they are associated. It should also be understood that many of their worst enemies are present only when the water covers their burrows.

External Structure.-1. Examine a living worm in a dish of sea-water, noting the motions of the body and of the parapodia or swimming feet.

Make a drawing of the animal.
2. Hold it down against the bottom of the dish to induce it to protrude the proboscis, the protrusible anterior portion of the alimentary canal. This is lined with chitin and armed with numerous denticles and a pair of lateral jaws.
3. Is the general surface clean or slimy? Compare with the earthworm in this respect and explain the basis for the difference.
4. Determine the direction of the peristaltic waves in the dorsal blood-vessel.
5. Is the median ventral nerve cord visible through the bodywall?
6. In a freshly killed or preserved worm, count the segments or metameres and compare it with your neighbor's to ascertain whether the number is constant. What segments, if any, are devoid of parapodia? Why?
7. In the head distinguish the prostomium, which bears the
four eyes and a pair of short terminal tentacles. At each side of the prostomium is a thick palp. Determine whether the worm has the sense of vision, and whether its sense of touch is delicate.
8. Also in the head find the peristomium, the segment which surrounds the mouth and bears four pairs of lateral tentacles or cirri. Stretch the mouth with forceps.

Make an enlarged drawing of the head.
9. Find the small terminal anus and a pair of caudal cirri on the last segment.
10. With scissors cut off a parapodium close to the body and observe that it has a dorsal blade and a ventral blade (notopodium and neuropodium). Each of these contains a bundle of bristles or setce. What use can you ascribe to the setæ? In each bundle is one very thick seta, the aciculum, which extends into the body and is attached to muscles. Of what use is the aciculum? Examine a few of the small setæ microscopically. What is their structure? Why is it desirable to have so many of the small setæ? Why does this animal need more than an earthworm needs?

Observe that each parapodium has a small dorsal and a small ventral cirrus, and that the main portion of both notopodium and neuroporium has the form of a flattened blade, somewhat divided into lobes. The largest lobe of the notopodium is very thin and vascular. What function can you ascribe to it?

Draw a parapodium.
11. Look for the nephridiopores, minute apertures which are segmentally placed on the ventral surface near the neuropodial cirri.

Internal Structure.-For dissection use a specimen that has been killed and fasten it down by a pin through the head and one through the posterior part. With scissors cut through the bodywall, longitudinally, near the mid-dorsal line.

Find the dissepiments which divide the coelom, or body-cavity, into metameric chambers. Cut through the dissepiments with scissors and pin the edges of the body-wall apart, progressing toward the head.

Circulatory System.-The dorsal blood-vessel lies along the dorsal surface of the alimentary canal and gives off branches in each segment, which ramify through the body-wall and viscera and connect with the longitudinal, ventral blood-vessel. The blood-plasma contains red coloring-matter in solution.

Digestive System.-The mouth-cavity leads into a muscular pharynx, a portion of which is protrusible as the proboscis. Examine carefully the muscles of the pharynx, protractors and retractors, and ascertain their attachments. Posterior to the pharynx find a narrow esophagus and a small dilation, the crop, with a digestive gland at each side. Where does the duct of the gland open? In the very long stomach-intestine, which follows the crop, note the constrictions and their relations to the dissepiments. Can you demonstrate dorsal or ventral mesenteries? Cut open the pharynx and the anterior end of the stomachintestine and note the character of their walls.

Make a drawing of the digestive system.
Muscular System.-How many distinct bands of longitudinal muscles can be distinguished? Examine with a hand-lens the parapodial muscles attached to the base of the acicula. Can you make out a layer of circular muscles? Of what layers does the body-wall consist?

Excretory System.-The nephridia are not nearly as easily found or studied as they are in the earthworm. Near or just beneath the lateral edges of the ventral muscle-bands find the minute pear-shaped nephridia. Determine their distribution in the body. Each nephridium consists of a tortuous canal in a multi-nucleate mass of protoplasm. The external opening is the nephridiopore above mentioned. The inner end perforates the dissepiment anterior to the body of the nephridium and opens into the cœelomic cavity of the segment next in front, by a ciliated funnel, the nephrostome. With a hand-lens try to find the nephrostome. Remove a nephridium by means of fine forceps and examine it with a microscope.

Reproductive System.-The sexes are separate, but no permanent gonads are present. At the breeding season the ova
or spermatozoa are proliferated from the cœlomic epithelium of a large number of segments and escape by rupture of the body-wall.

Nervous System. ${ }^{1}$-On lifting the alimentary canal you will see the ventral ganglionated nerve cord. Note the nerves passing off laterally from the ganglia. How many pairs of nerves per segment are there, and how are they placed? Are the ganglia metameric? Is there any indication that the nerve cord is double? At the anterior extremity of the cord note the infraesophageal ganglia and, extending from them and encircling the anterior end of the alimentary canal, the circum-esophageal connectives which unite above in the bilobed brain or supra-esophageal ganglia. Sensory nerves connect the brain with the eyes, tentacles, and palps.

Make a drawing of the nervous system.

## LUMBRICUS. (Earthworm.)

Earthworms feed mostly at night. What reason is there for this habit? You should look for earthworms with a lantern some mild, calm summer evening when the ground is quite moist. See if they leave their burrows entirely. How much of the body is generally protruded? Can you determine what the worms are doing? Are they disturbed by walking near them? Are they ever disturbed by flashing the light suddenly upon them? Of what service to them is the ability to distinguish light? Look for castings near the burrows. During daylight look for castings and thus determine the relative abundance of worms in lawns, gardens, etc. (As the worms come to the surface only when it is moist, castings will be abundant only at such times.) Do the castings indicate anything about the feeding habits.

[^8]Place a living specimen upon moist filter-paper and observe the direction and method of movement. How can it reverse its direction? Gently touch different parts of the body to see which are the most sensitive.

Observe the movement of the blood in the dorsal vessel. In what direction does it move? Does the vessel change in shape?

Place a preserved specimen in a dish with a little water and notice:

1. The difference in shape of the two ends of the body. The mouth is at the anterior end, below the protruding lobe of the prostomium. The anus is a vertical slit at the end of the last somite.
2. The dorsal and ventral sides. How do they differ?
3. The right and left sides are symmetrical. Count the somites of the body, compare with others, and record the result.
4. On the anterior third of the body certain somites are swollen and form the clitellum. What somites are swollen? The clitellum is not present in young individuals. It is used in making egg-cases and providing food for developing embryos. Understand how this is accomplished.
5. Small swollen areas on the ventral side of the fifteenth somite, where the vasa deferentia open.
6. Setoe project slightly from the surface of each somite. These light colored spines are easily felt with the fingers. See if you can determine the number and position of the rows by stroking gently. How are they used?

Draw a ventral view of the anterior end, including the clitellum, and another view of the posterior end.

Taking care not to cut deep, with fine scissors cut through the dorsal wall of the body, and extend the cut the whole length of the body. Carefully spread and pin the animal open. In doing this you must tear or cut the septa, but be careful not to tear or break the organs that perforate them.

Alimentary Canal.-This consists of a straight tube that runs the length of the body.

1. Immediately behind the mouth is a muscular, white organ,
the pharynx. Through how many somites does this extend? It is connected with the body-wall by numerous, radiating muscle fibers. What function do these fibers perform?
2. Behind the pharynx is the narrow and long esophagus. This runs posteriorly between lobed, light colored organs, the seminal vesicles, that will be studied in connection with the reproductive organs. Press these aside and notice the small calciferous glands.
3. The esophagus leads to the crop, which lies just anterior to and in contact with the gizzard. In what somites are these organs placed? What is their shape? Do you understand the function of each?
4. Leaving the gizzard is the stomach-intestine, which runs through the remainder of the body, giving off lateral diverticula in each somite. Do you know its function?

Notice the relation of the septa to the alimentary canal.
Circulatory System. - 1 . Lying dorsal to the alimentary canal is the blood-vessel that could be seen pulsating in the living specimen. In most cases this vessel is full of blood and appears brown.
2. Near the anterior end of the body large side branches, the aortic arches, are given off on either side and pass down around the esophagus. How many aortic arches do you find? In what somites are they placed?
3. Examine with a lens and see whether you find other vessels connected with the dorsal aorta. If you do determine, how are they placed? Do they appear like the aortic arches?

Make a drawing of the anterior end of the body, showing the points you have seen.
4. Gently press the stomach-intestine to one side and see if you find a blood-vessel beneath it. Do the aortic arches join this? Other connections between blood-vessels are too small to be studied in dissections, but you should understand from textbooks or lectures what they are, and the probable course of circulation.

Excretory System.-1. A pair of nephridia occurs in each
somite, one nephridium on either side of the alimentary canal. (The first three or four somites are not provided with nephridia.) Each nephridium is a coiled tube, appearing to the unaided eye as a fluffy mass, that opens externally between the groups of setæ, in the position already observed, and internally by a small opening, the funnel. The inner opening is not in the somite in which the most of the tube lies, but in the somite anterior to it. That is, the nephridium that occupies the space in somite twenty, opens externally on somite twenty, but internally perforates the septum directly anterior and opens into somite nineteen.
2. Remove a nephridium with your forceps and examine it with your microscope. Notice that it consists of a coiled tube of varying diameter. The funnel is not easy to find and is hard to remove. It may be found by removing the portion of the septum through which the nephridium passes and examining it with a microscope.

Draw the nephridia into your previous figure.
Cut the stomach-intestine behind the gizzard and pull it forward, carefully separating the tissue from it as it is drawn forward, so underlying organs will not be disturbed. In this way free the alimentary canal to the position of the pharynx.

You can now see the extent of the nephridia, and possibly see where they perforate the septa.

Reproductive System.-1. The seminal vesicles are large white bodies, united in the median line. They send three lobes on either side, that normally overlap the posterior part of the esophagus. In what somites do the lobes occur?
2. Carefully open the seminal vesicles near the median dorsal line and examine their contents microscopically.
3. With a pipet wash out the contents and notice the two pairs of convoluted funnels, the inner openings of the vasa deferentia. The testes are hard to find, as they are the same color as the coagulated mass that filled the seminal vesicles. They are attached to the septa just anterior to the funnels. The
narrow tubes of the vasa deferentia may sometimes be seen leaving the seminal vesicles. They open externally on somite fifteen.
4. The ovaries are a pair of very small organs attached to the posterior surface of the septum that separates the twelfth from the thirteenth somite, near the mid-ventral line. They may sometimes be found with a lens, but are not usually visible otherwise. If possible, remove an ovary and examine it with a microscope to see its shape, and to find which portion has the most mature eggs. The oviducts open into the cavity of the thirteenth somite and externally through the ventral wall of the fourteenth somite, in line with the nephridia. They can seldom be seen in dissections.
5. Between the ninth and tenth and the tenth and eleventh somites, on the ventral side, are two pairs of white, rounded pouches, the seminal receptacles, that open externally but not internally. Understand their function.

Nervous System.-1. On the dorsal surface of the pharynx, near its anterior end, are the two cerebral ganglia. They lie on either side of the median line and are connected by a stout commissure. In what somite do they lie?
2. The remainder of the ganglia lie ventral to the alimentary canal. The first ventral ganglia are connected with the cerebral ganglia by connectives that pass around the sides of the pharynx. Adjacent ganglia of the ventral chain are united by connectives. The ganglia of each somite, and the cords that connect those of adjacent somites, are fused so that the original paired condition is not very apparent. How far does the ventral chain of ganglia extend? Where do nerves leave it?

Draw the nervous system into the figure that shows the reproductive system.

Notice the sacs that inclose the setæ and indicate them in the above figure.

## Examine prepared serial microscopic sections. ${ }^{1}$

[^9]1. The cuticle will probably be absent in most sections, in which case the outer covering will be the cellular hypodermis or skin. How many cells thick is this layer? Look for the gland cells that keep the living worm moist. Do you know how the cuticle is formed?
2. Beneath the hypodermis is the circular muscle layer, which is followed by the longitudinal muscle layer. The fibers of the latter are arranged in conspicuous bundles. Lining the bodywall is the thin peritoneal layer. Do you understand the function of each of these layers? How is the body elongated?
3. Find the setce and determine where they are placed, how many are in each group, how many groups there are, how they pierce the body-wall, and what muscles are attached to them. Why are setæ not in every section?
4. The alimentary canal consists of a lining epithelium, followed by connective tissue and muscle, and, on its outer wall, peritoneal cells, which in the region of the stomach-intestine are large, very numerous, and are known as the chloragog cells.
5. Lying in the mid-ventral line, beneath the alimentary canal and close to the body-wall, is the ventral nerve cord. Examine its structure. See if any of the sections show nerves leaving it.
6. Dorsal to the alimentary canal is the dorsal blood-vessel, on its ventral side is the ventral blood-vessel, and ventral to the nerve cord the sub-neural vessel.
7. Find sections of the nephridia. Where are they placed? How do the sections appear? Why?

Other organs will appear in most of the sections. See if you can identify them.

Draw an enlarged cross-section.
from grit, before they are preserved for sectioning. It is well to narcotize them by placing them in a small quantity of water and adding a little alcohol from time to time (never enough to make the worms squirm violently) until they cease to move. They may then be killed with sublimate acetic or other killing agent and treated in the usual manner.

## AUTOLYTUS CORNUTUS.

This polychrete lives in cylindrical tubes of its own construction that it attaches to seaweeds and hydroids, and is especially interesting because of its method of reproduction, by budding.

Study live and preserved specimens with the naked eye and with the hand-lens, in order to form a correct idea of its natural color, size, and movements, and then study stained specimens with the low power.

1. Observe two individuals attached end to end. The anterior one is a non-sexual zoöid (or original "stock") and is giving rise to a new sexual zoöid by budding. Counting the peristomium as one somite, on what somite does the bud begin and what does it represent?
2. Study the head of the anterior, non-sexual zoöid. Find three prostomial tentacles. How are they arranged? Find the eyes. How many pairs are there? Do you find palps? On the peristomium find the two tentacles and a tentacular cirrus.
3. On the succeeding somites study the parapodia. Observe the large dorsal cirri and the knob-like notopodium with the short unjointed setæ. There is no neuropodium.
4. Identify the pharynx, gizzard, and intestine.
5. Compare the sexual bud with the non-sexual individual. The adult male and female differ. The outer prostomial tentacles of the male are forked. Is this bud to be a male or a female? In an older sexual individual make out a so-called thoracic region in which the setæ are short, and an abdominal region in which the setæ are long. Look for evidences of germ cells in the bodycavity, between the intestine and body-wall. There is a ventral brood-pouch on the adult female and the young partly develop in it. Find the anal cirri.

A drawing illustrating the method of reproduction should be made.

## LEPIDONOTUS SQUAMATUS.

The family Polynoidæ, to which this belongs, can be distinguished from all others by the presence of peculiar plates (elytra) on the dorsal surface. They lead sluggish lives under stones and are carnivorous. Note the size, color, and shape of the worm.

1. The elytra. How are they arranged? What purpose do they serve? How many are there? With a hand-lens observe the fringed condition of the outer edge and the small tubercles covering the surface. Note the color of the elytra and the notches in the inner edges of the posterior pair. L. sublevis differs in that its elytra are merely punctate and the posterior pair are not so deeply notched.
2. The head is hidden by the first pair of elytra. If the elytra are removed you will find a small reddish prostomium with a pair of eyes, three tentacles (the middle one large and clubshaped), and a pair of palps.
3. Find the mouth, which is placed ventrally. Note the median, red streak along the ventral surface that is due to pigmented cells which surround the nerve cord. If the pharynx is everted, observe the fringe of papillæ surrounding a dorsal and a ventral pair of teeth. There are no septa in the anterior region of the body where the withdrawn pharynx lies.
4. The anus is dorsally placed, and can be found beneath the notches in the last pair of elytra.

A drawing is desirable.

## DIOPATRA CUPREA.

This worm belongs to the family Eunicidæ. Specimens live on mud- and sand-flats, sometimes above low-tide mark, but usually where the burrows are covered by water. This form is especially interesting because of its feeding and tubebuilding habits, parapodial gills, and complex jaw-apparatus. Study the preserved specimens for the structure and specimens in an aquarium for the habits. Notice the construction of the tube and determine how it is formed.

1. Notice the size of the body, also its gradual attenuation posteriorly. Account for this condition. Observe how degenerate the parapodia are posteriorly from the same cause.
2. The prostomium. Identify the tentacles. What is their number and arrangement? Find a pair of eyes dorsally placed behind the tentacles, also a pair of palps in front of them. Note a second, larger pair of palps which serve as an upper lip.
3. The peristomium. What appendages does it carry? Note the lower lip formed from the ventral edge of the peristomium.
4. The position of the jaw-apparatus can be identified as being in a pouch ventral to the buccal region. Find both by means of a probe. What kind of food are such jaws fitted for?
5. The parapodia vary greatly, depending upon their position on the body. Notice that the notopodia are vestigial, being represented only by the dorsal cirri and, toward the anterior end, branchial cirri or gills, acicula, can be seen projecting into the base of the dorsal cirrus. The neuropodium shows two kinds of setæ: (a) stiff and unjointed, (b) crochets. It also bears an accessory cirrus and the ventral cirri, which are curiously modified in most cases as glands for use in tube-building. Make out all these modifications and where they occur.

## CHAETOPTERUS.

This is one of the most aberrant of our Polychætas. It lives on mud-flats below low tide in a U -shaped, parchment-like tube, both ends of which protrude above the sand. In the body three regions can be distinguished. Examine a tube and see the size of its outer openings. Specimens may be made to live in tubes of glass, bent to correspond to their tubes, and their normal movements may thus be studied in aquaria. What must be the source of the animal's food.

1. The anterior region. Identify ten modified parapodia, the fourth of which is supplied with a group of much stouter setæ. Observe that the funnel-like mouth is placed dorsally and surrounded ventrally and laterally with flaring peristomial
lips. Find the pair of peristomial cirri. The region between these cirri represents the prostomium.
2. The middle region consists of five somites. The first, the eleventh segment, is marked by the great pair of wings which are used to bring food to the mouth. Their dorsal surfaces are grooved and supplied with cilia, as is the median dorsal line. Hence a current of water passes continually toward the mouth. The twelfth somite is marked by a dorsal and a ventral sucker, which are modified parapodia. Somites thirteen, fourteen, and fifteen carry notopodial folds or fans, for keeping up a stream of water through the tube. Their neuropodia are mere knobs.
3. The posterior region is less highly modified. Of how many segments does it consist? Notice their gradual diminution in size. Homologize the parts of their appendages.
4. The living Chætopterus contains a green coloring-matter and is very phosphorescent. A commensal polynoid often lives in its tube.

A drawing is desirable.

## AMPHITRITE ORNATA.

This belongs to the family Terebellidæ and lives under stones, or in mud or sand, along shore in stout muddy tubes.

1. Find the prostomium, which forms an upper lip and bears a transverse group of long, retractile tentacles.
2. The peristomium forms the under lip, but bears no appendages.
3. Find three pairs of ramose gills. These are modifications of the dorsal cirri. (Terebella has three pairs, but they are of unequal size.)
4. Notice again the feeble development of the parapodia and the absence of ventral cirri and neuropodial setæ. Setæ are not found posteriorly. On what somite do they begin?
5. Find the ventral shield glands which are concerned in building the tube. How many are there?
6. The live worm is of a bright pinkish color, due to its red
blood. There is only one internal septum and it forms a socalled diaphragm. Anterior to the diaphragm the nephridia are large and excretory in function. Posterior to the diaphragm the nephridia serve as genital tubes.

A drawing is desirable.

## CISTENIDES GOULDII.

This very aberrant worm belongs to the family Amphictenidæ.

1. Study the beautiful tube of sand and the manner in which the grains are fitted together. It is said that the worms can carry the tubes about.
2. See how the peristomium and the large golden seto close the shell. The setæ are said to belong to the second somite. Notice the ends of the tentacles protruding from the tube.
3. Find the tentacles, two pairs of gills, and the parapodia. Notice how the latter diminish in size posteriorly and how each typically consists of a ridge-like notopodium without setæ and a reduced neuropodium with long golden setæ. If the specimen is complete you can see a much degenerated portion (the scapha) at the posterior end, which serves to close the small end of the tube.

A drawing is desirable.

## CLYMENELLA TORQUATA.

This worm belongs to the family Maldanidæ. It makes tubes of sand and generally lives in sheltered places on sandy or muddy shores.

1. Study the structure of the tube; observe how the animal protrudes at either end of the tube.
2. Observe the diameter and length of the worm, the small number of somites, their great length as compared to somites of Nereis, and the reduced parapodia provided with simple setæ. Notice the characteristic collar on the fifth somite, and the funnel at the posterior end, with the anus within it. The mouth is more or less ventral and is overhung by a narrow prostomium surrounded by a peristomial rim.

A drawing is desirable.

## ARENICOLA CRISTATA.

This remarkable worm, called the "lug-worm" by fishermen, belongs to the family Arenicolidæ.

1. Notice the color, and the gradual diminution in size posteriorly. Also notice the false annulations between the appendages, the arborescent gills representing modifications of certain notopodia, the reduced parapodia, and the character of the setæ.
2. If the buccal region is everted, observe the papillæ which cover it. The prostomium is an inconspicuous dorsal knob and it is fused with the peristomium. At the sides of the prostomium is the ciliated nuchal groove.
3. On what somites can you find indications of neuropodia? of gills? of setæ? Notice the cirriform papillæ of the "tail." Find large nephridiopores on certain somites about an eighth of an inch below each notopodium. What is the distribution of the pores?

A drawing is desirable.

## SABELLA MICROTHALMA.

This worm belongs to the family Sabellidæ. It builds leathery, muddy tubes on piles, among tunicates, algæ, etc.

1. In addition to the general size, form, and color of the worm, observe the reduced condition of the parapodia, and the arrangement and general structure of the branchice or gills. These structures are modifications of the palps and not of the parapodia, as in the other species which have been studied. Observe the two irregular rows of small ocelli or eye-spots. Account for the presence of eyes in their position. A pair of short tentacles can be seen by spreading the branchiæ aside.
2. Find a collar which is used in smoothing the orifice of the tube. This is a peristomial structure and is so extensively developed in some species as to hide the prostomium entirely.
3. Identify eight setigerous somites anteriorly, in which the capillary setce are in the notopodium and the uncini are in the neuropodium. With the peristomium they form a "thorax" of nine somites. In the somites which follow, the "abdo-
men," observe that the uncini and the capillary setæ stand in the reverse order.
4. Find the ventral shield-glands. A furrow (sulcus or fæcal groove) divides them into pairs toward the posterior end of the worm.

A drawing is desirable.

## HYDROIDES.

This is a member of the family Serpulidæ. Study living specimens and their heavy calcareous tubes. Notice the banded branchice (modified palps) and the dorsally placed operculum, a modified gill filament. Look for "eyes" on the gill filaments. A drawing is desirable.

## SPIRORBIS BOREALIS.

This animal is also a member of the family Serpulidæ. Specimens are very abundant along the shore, attached to Fucus.

1. Study the tube and notice the way in which it "parallels" the form of a small snail-shell.
2. Remove a live specimen from the Fucus on which it grows and crack the tube away with a needle. Study the animal in a watch-glass with a low power. Identify the gills, the operculum (which serves as a "brood-pouch"), the setcx, and the collar. Are there any "eyes" on the gills?
3. Study the egg-strings which are lodged in the tube, and the young embryos which are to be found in the brood-pouch.

A drawing is desirable.

## GEPHYREA.

## PHASCOLOSOMA.

This form is commonly found buried in sand between tidemarks. Specimens sometimes occur on the same flats with Nereis, but they are generally more abundant where the mud is of a slightly different, more sticky character.

1. Handle a living specimen and see how turgid it is. If you touch a specimen that has been allowed to expand in a dish
of sea-water you will find it is rather soft, but becomes turgid immediately upon being touched. How is this accomplished?
2. Examine a living animal in a dish of sea-water. The anterior portion of the body, the introvert, is drawn in, but may occasionally be extended, when it will be seen to bear at the anterior extremity a crescentic tentacular fold, which partly surrounds the mouth.
3. Compare with a preserved specimen which has been killed with the introvert extended.

Make drawings showing the animal with the introvert protruded and with the introvert concealed.
4. The anus is located on a dorsal papilla, anterior to the middle of the body. Near the anus a pair of lateral papillæ mark the position of the nephridiopores. The coiled intestine and brown nephridial tubes can probably be seen through the body-wall. Note carefully the character of the skin. Is there any indication of spines, appendages, or eye-spots?

For dissection use both fresh and preserved specimens.
With scissors open the worm from end to end near the middorsal line, and pin the body-wall out flat.
5. In opening the fresh worm, note the pinkish colomic fluid which fills the coelom. Examine a drop under the microscope. What functions has this fluid to perform?

Alimentary Canal.-Trace the alimentary canal (stomachintestine) from mouth to anus. Do any digestive glands open into it at any point? Note the mesenterial thread which runs through the axis of the intestine spiral. Where is it attached? Does it seem to be contractile in the fresh worm?

Muscular System.-Note the silvery-white longitudinal muscles composing the inner layer of the body-wall. Are they arranged in distinct bands or in a continuous sheet? Remove some of these muscles carefully to expose the layer of circular muscles. How many retractor muscles of the introvert are there? How is the mechanism of protrusion of the introvert to be explained?

Circulatory System.-This system is very difficult to observe. Dorsal and ventral blood-sinuses are present, and communicate anteriorly by a circular sinus.

Excretory System.-Find a pair of brown nephridia, an inch or more in length. Cut off a nephridium (from the fresh worm) as close as possible to the body-wall, and examine it under a microscope. Near the cut (the attached) end find the coelomic opening or nephrostome. Is it ciliated?

Reproductive System.-The sexes are separate. There are no permanent gonads, but at the breeding season sex-cells are proliferated from the cœlomic epithelium, at the points where the ventral retractor muscles are attached to the body-wall. These cells become detached and mature while floating in the cœlomic fluid. They pass out through the nephridia, which function as gonoducts.

Nervous System.-Does the ventral nerve-cord seem to be double? Is it ganglionated? Does it give off lateral nerve branches? Trace the circum-esophageal connectives to the supraesophageal ganglion. The ganglion is small and situated behind the tentacular fold, to which sensory nerves extend. Does any system of organs show segmentation?

Make a drawing to show the internal anatomy.

## MOLLUSCA.

Unsegmented. Usually provided with a calcareous protecting shell and a ventral foot.

Class 1. Lamellibranchiata.
Bivalve shell. Gills adapted for gathering food as well as for respiration. Foot usually adapted for burrowing. No hard mouth parts.
Order 1. Protobranchia.
Gills composed of a series of transverse plates. Foot apparently split at the end. Two adductor muscles, posterior frequently the smaller. (Nucula, Yoldia.)
Order 2. Filibranchia.
Gills lamelliform. Filaments united by modified cilia. Anterior adductor muscle frequently greatly reduced. (Mytilus, Modiola.)
Order 3. Pseudo-lamellibranchia.
Gills lamelliform. Inter-filamentar junctions usually not very extensive, may be either ciliary or vascular. Only one adductor muscle. (Pecten, Ostrea.)
Order 4. Eulamellibranchia.
Gills lamelliform. Inter-filamentar junctions extensive and vascular. Adductor muscles of nearly equal size. (Venus, Unio, Mya.)
Order 5. Septibranchia.
Gills reduced to a horizontal partition. Two adductor muscles. Deep sea forms. (Silenia, Cuspidaria.)
Class 2. Amphineura.
Bilaterally symmetrical, elongated. Nervous system not concentrated. Radula sometimes present. Shell, when present, composed of eight transverse pieces.
Order 1. Placophora.
Dorsal shell, composed of eight transverse pieces.

Foot broad. Gills simple, lateral. (Chiton, Chætopleura, Trachydermon.)
Order 2. Aplacophora.
Body elongated, covered by a mantle. Adult without shell but with spicules. No true foot. Gills posterior. (Neomenia, Dondersia.)
Class 3. Gastropoda.
Body unsymmetrical, usually covered by a spiral shell. Foot usually flattened and adapted for creeping. Radula usually present.
Subclass 1. Streptoneura.
Nervous system twisted into the form of a figure 8. Sexes distinct.

Order 1. Aspidobranchia.
Nervous system not concentrated. Gills usually present and paired. Auricles paired. (Acmæa, Patella, Haliotus.)
Order 2. Pectinibranchia.
Nervous system somewhat concentrated. Single gill. Single auricle. (Sycotypus, Buccinum, Crepidula.)
Subclass 2. Euthyneura.
Nervous system not twisted into the form of a figure 8. Sexes united.
Order 1. Opisthobranchia.
Aquatic respiration. Shell when present rather delicate. (Bulla, Æolis.)
Order 2. Pulmonata.
Air-breathers. Live on land or in fresh water. Aperture to mantle cavity narrow and contractile. (Limax, Limnæa, Helix.)
Class 4. Scaphopoda.
Bilaterally symmetrical. Shell tubular, elongated dorso-ventrally and open at both ends. Foot conical. (Dentalium.)
Class 5. Cephalopoda.
Bilaterally symmetrical. Shell chambered or reduced and internal. Distinct head with arms bearing suckers.
Subclass 1. Dibranchiata.
Arms forming a circlet around the mouth. Funnel a complete tube. Shell usually internal. Two gills.

Order 1. Decapoda.
Ten arms, two of which are elongated, suckers on stalks. (Loligo, Sepia, Spirula.)
Order 2. Octopoda.
Eight arms, suckers sessile. (Octopus, Argonauta.)
Subclass 2. Tetrabranchiata.
Tentacles numerous. External chambered shell. Funnel open along one side. Only one living genus. (Nautilus.)

## LAMELLIBRANCHIATA.

## VENUS MERCENARIA. (Quohog. ${ }^{1}$ )

Animals of this species wander around over muddy bottoms in rather shallow water, keeping the siphon end, at least, above the surface of the mud. If possible, you should find specimens in their native places and watch their movements. Specimens placed in water and left undisturbed for some hours are likely to protrude the siphons, and the foot may be protruded in some cases. ${ }^{2}$ Allow powdered carmine to slowly settle past the openings of the siphons and determine the direction of the current of water for each. Touch portions of the animal and find what parts are most sensitive.

Shell.-Note its general shape, and that it is composed of two symmetrical parts, the valves. For each valve notice:

1. The outline.
2. A swelling, the umbo, ending in a point, the beak, from which growth has proceeded.
3. The lines of growth. Were the valve cut off along one of these lines, the shape would not be changed. Why are the lines arranged in this manner? How were they formed?

The two valves are joined by the ligament. The margin

[^10]bearing the ligament is dorsal, and that toward which the beaks point is anterior. Which valve is right and which is left?

Draw a valve, showing the points observed.
Pry the two valves apart and insert a knife-blade between the mantle and one valve of the shell. Notice that the lobes of the mantle are loosely attached to the shell along their margins, and more firmly attached a half inch or more from the margins.

Separate the mantle from one valve, and cut the adductors where they are attached to this valve. Why do the valves gape now? Press them together, and notice that they stay closed only while held. Remove a valve and study its interior.

1. Find the large scars where the anterior and posterior adductor muscles were attached.
2. Find smaller scars where the anterior and posterior foot muscles were attached. The anterior scar is dorsal and a little posterior to the corresponding adductor muscle scar. (Not the position for Unio.) The posterior scar connects with the dorsal portion of the corresponding adductor muscle scar.
3. The ventral borders of the adductor muscle scars are connected by a distinct line, the pallial line. What forms it? The posterior end of this line is indented to form the pallial sinus. (Not true for Unio.) What is the meaning of this sinus?
4. Along the dorsal margin of the valve notice prominences, the teeth. There are two kinds of teeth. The anterior, cardinal, consist of short elevations. The posterior, lateral, are not very prominent, but are comparatively long and extend along the dorsal margin. Notice that the teeth on the two valves interlock. What is their function?

Draw a valve as seen from the inside.
5. By examining the inside of a shell of Unio or Mytilus near its margin, the typical three layers of which it is composed can be seen. How is it possible for all three layers to be secreted by the mantle, which lines the inside of the shell? Can you find any reason for more than one layer?

Mantle.-This consists of two lobes (one of which is normally
applied to the inner surface of each valve of the shell) that are united dorsally.

1. The free border of each lobe is thickened, and contains muscles that were attached to the shell along the pallial line. What function do these muscles perform?
2. The posterior portions of the lobes of the mantle are thickened and united to each other so as to form two tubes (in Unio the ventral tube is formed by contact only), the siphons, through which water passes into and out of the shell.
3. See how the muscles of the siphons are arranged and attached. Does the attachment bear any relation to the pallial sinus?

Visceral Mass and Foot.-These portions form the large median mass. The viscera are contained in the dorsal portion.

1. The ventral portion is hard and muscular, and forms the foot.
2. Besides the crossing muscle fibers of which the foot is largely composed, it is supplied with two pairs of muscles that are attached to the shell. The cut ends of these muscles, the anterior and the posterior foot muscles, may be seen protruding through the lobe of the mantle. They correspond in position to the scars on the shell.

Do you understand by what means the foot is protruded?
Gills.-These consist of two pairs of thin, striated, somewhat brownish organs, a pair lying on each side of the visceral mass, between it and the lobes of the mantle.

1. Each gill extends from the wall that separates the two siphons, anteriorly and dorsally to a point nearly opposite the beaks of the shell, and is attached by its dorsal margin only.
2. Each outer gill is attached along its dorsal border to the corresponding mantle lobe on the outer side. The inner gills, besides being attached to the dorsal margins of the outer gills, are on their inner sides attached to each other and to the visceral mass. (For some distance the inner side of the inner gill lies against the visceral mass, but is not attached to it.)

By this arrangement the space between the lobes of the
mantle, which is known as the mantle chamber, is divided into a ventral and a dorsal portion. The ventral portion is much the larger, communicates with the ventral siphon, and because the gills hang into it, it is known as the branchial chamber. The dorsal chamber is known as the cloacal chamber. The siphons are frequently referred to by names corresponding to the chambers with which they communicate. The minute structure of the gills will be studied later.
3. Place a little powdered carmine on the gill of a specimen that is submerged in sea-water and see what becomes of it.

Labial Palps.-These consist of a pair of rather small triangular flaps on each side of the visceral mass.

1. The two outer palps are united above the mouth, which is situated just posterior to the ventral border of the anterior adductor muscle, to form a small fold that corresponds in position to an upper lip.
2. The two inner palps likewise unite to form a fold corresponding in position to an under lip.

Make a drawing showing the arrangement of the soft parts.
Structure of a Gill.-Cut off a piece of the edge of a gill, put it on a slide with a drop of sea-water, and examine with a low power of the microscope. Notice:

1. The cilia on the edge and surface of the gill.
2. The surface is marked by a series of parallel ridges, the filaments, with grooves between them. ${ }^{1}$

The filaments are joined together laterally by series of bridges (you will see them later), the inter-filamentar junctions, with the pores, inhalant ostia, between them. Each side of the gill is thus composed of a single layer of united parallel filaments, which together form what is known as a lamella. Each gill is composed of two such lamellæ, one on each side. These lamellæ are united at intervals by bridges that run the whole width of the gill (dorsal to ventral), parallel to the filaments, and at

[^11]right angles to the inter-filamentar junctions. These are called the inter-lamellar junctions. By means of the inter-lamellar junctions, the space between the two lamellæ is divided into a series of water tubes. The openings of these tubes into the cloacal chamber may easily be seen after the cloacal chamber has been cut open.
3. Separate a small piece of one lamella from the other. This can most readily be done by catching the free dorsal border of the inner lamella of an inner gill with the forceps, and either tearing off a piece or freeing it by cutting with scissors while it is being removed with the forceps. Mount this piece, with the outer surface up, without pressing it, under a coverglass in a drop of sea-water and observe with a low power:
(a) Filaments, that run the width of the gill.
(b) Inter-filamentar junctions, which form bridges connecting the filaments.
(c) Inhalant ostia. The opening bounded by filaments and inter-filamentar junctions.
(d) The position of the torn inter-lamellar junctions, appearing as indefinite dark stripes running in the same direction as the filaments.

With a high power observe:
(a) The chitinous rods that lie inside of, and stiffen the filaments.
(b) The cilia on the sides of the filaments. These are of two kinds: (1) Surface cilia that form currents of water along the filaments. These will be seen waving back and forth, or if still moving rapidly, apparently moving along the sides of the filaments. (2) Deeper cilia that are down between the filaments and can be seen by changing the focus. These move at right angles to the others, and apparently become longer and shorter. Why?

Draw a surface view of a piece of a lamella.
Examine a piece of the gill of Mytilus for the above structures. In this form the inter-filamentar junctions are small and composed of modified cilia only, and the inhalant ostia
are correspondingly large. By pressing the gill the inter-filamentar junctions can be pulled apart.

Study prepared sections of the gill of Venus and notice:

1. Lamellæ.
2. Inter-lamellar junctions.
3. Water tubes.
4. Filaments.
5. Inter-filamentar junctions.
6. Cilia.
7. Inhalant ostia.
8. Blood spaces.
9. Chitinous rods.

Draw.
Understand the direction taken by water in passing from the branchial to the cloacal siphon. What makes the water move?

Labial Palps.-The positions of these organs have already been noted.

1. Examine a piece of the palp with a microscope, and notice that the side turned toward the adjacent palp is thrown into ridges and grooves, and is densely ciliated.
2. The space between each outer and inner palp is continuous with the "corners" of the mouth. The free margins come close to the borders of the gills and normally inclose them.

Understand how food is gathered and carried to the mouth.
Circulatory System.-The pericardium, in which the heart lies, is a somewhat triangular space that appears clear, through the mantle. It lies just anterior to the posterior adductor muscle. Open the pericardium, and notice the beating of the heart. The heart consists of three parts:

1. A central portion, the ventricle, that surrounds the intestine and gives rise to a blood-vessel at each end.
2. Two triangular portions, the auricles, that receive blood from the gills and open into the sides of the ventricle.

Notice the sequence and power of the contractions.
Just posterior to the pericardium is an enlarged portion of the alimentary canal. This has no relation to the heart, for which it is sometimes mistaken.

Excretory and Genital Systems.-The excretory system consists of a pair of dark colored glandular organs that lie beneath the pericardium. Each communicates with the pericardium by a small opening that is not easy to demonstrate in dissections, and with the cloacal chamber by another small opening.

By turning the two gills (of Venus) dorsally a very small papilla may be seen, just beneath the free border of the inner gill, lying in the cloacal chamber. On the tip of this papilla are two openings. The inner one is the opening of the excretory organ. The outer one is the opening of the genital duct.

The genital glands are light colored organs that, during the breeding season, extend through the principal part of the visceral mass. Neither the genital nor the excretory systems can be profitably studied in a general dissection of this form. In Unio the excretory organs are more satisfactory for study. Do you understand the supposed significance of their connection with the pericardium?

Nervous System.-1. Carefully remove the body-wall by the side of the esophagus and notice the cerebral ganglion of the corresponding side. This is a rounded, slightly yellow organ, about the size of a pin-head, lying just posterior to the dorsal border of the anterior adductor muscle. (In Unio it is more ventral in position.) The cerebral ganglia of the two sides are united by a commissure that passes anterior to the esophagus. Two connectives leave each cerebral ganglion. One passes posteriorly to join the visceral ganglion of the corresponding side. The other passes into the foot to join the pedal ganglion of the corresponding side.
2. Cut the united lamellæ of the inner gills ventral to the posterior adductor muscle. This will expose the visceral ganglia. They are pear-shaped bodies lying just beneath the posterior adductor muscle, connected with each other by a short commissure, and connected with the cerebral ganglia by connectives that may be traced a short distance forward without dissection. A large nerve leaves the posterior end of each ganglion and
supplies the posterior end of the corresponding lobe of the mantle. Smaller nerves go to the posterior adductor muscle and gills.
3. With a razor or sharp scalpel make a median sagittal section of the foot, extending it some distance into the visceral mass. This will expose the pedal ganglia, that lie just anterior to a loop of the intestine, and dorsal to the muscular portion of the foot. The pedal ganglia are connected with each other by a broad commissure and with the cerebral ganglia by connectives.

By careful dissection it is possible to trace the connectives and many of the nerves. The razor clam, Ensis, is especially favorable for dissections of the nervous system, as the ganglia, connectives, and many important nerves lie very near the surface and can be seen without cutting the tissues above them.

Make a drawing, indicating the position of the ganglia.
Digestive System.-This may be traced by following a guarded bristle that has been inserted into the mouth of a specimen that has been killed in hot water (not boiling), or by very carefully picking off the tissue from one side. The intestine where it penetrates the heart has already been seen, and may easily be followed to the anus.

The general arrangement of the alimentary canal is well shown by a median sagittal section of a preserved specimen.

The brownish digestive gland, commonly called the "liver," will be seen surrounding a portion of the stomach.

The enlargement on the intestine just posterior to the pericardium is of unknown function. In some forms a special diverticulum from the stomach bears a transparent cylindrical rod, the crystalline style. This can easily be found in Mya. Probably all lamellibranchs have similar structures more or less well developed, but many do not have special pouches for their formation.

Draw the alimentary canal. (This may be included with your sketch of the nervous system.)

Cut a preserved specimen into transverse sections about a quarter of an inch thick, and place the sections in their proper
order and position. (They should be placed in a dissecting pan in a very little water.)

Study these sections for the arrangement of organs. The relation of the gills to the branchial and the cloacal chambers should be understood.

Make drawings of sections that pass through the heart and through the posterior adductor muscle.

If time permits, it will be desirable to become acquainted with some of the structures of theoretic importance and with some of the adaptations of lamellibranchs for the lives they live. For this purpose a few forms have been selected, and directions for the study of the particular parts in question are given.

## YOLDIA LIMATULA.

This form belongs to the order Protobranchia, and is supposed to be one of the most primitive of living lamellibranchs. It lives in soft mud, such as is found in quiet coves and bays. (It is abundant in the Eel Pond at Woods Holl.) Although it burrows in the mud, it lives near the surface, and frequently has the posterior end above the mud.

1. Place a specimen in a dish of sea-water, and notice the movements and shape of the foot. See if the movements are always alike. What would happen if such movements were made by a specimen lying on soft mud? Place a specimen on mud and watch the results.
2. Leave a specimen in an aquarium in which two inches of bottom mud has been placed, and see if it is feeding in the morning.
3. Place a young, transparent specimen in a watch-glass of sea-water and study the parts. The foot has already been observed. Its motions will probably be seen again here. It has been considered a creeping organ. Do you find evidence that confirms or opposes the view? With a low power of the microscope notice:
4. The palps. These are very large. The outer palp on
each side is provided with a long appendage that may be protruded from between the valves of the shell. This is the feeding appendage.
5. The gills. These are quite small and are composed of parallel plates that are capable of being moved. They are situated behind the palps, are attached dorsally by muscular membranes to the body-wall, and posteriorly to the wall that separates the siphons. They illustrate what is supposed to be the most primitive type of lamellibranch gill. Watch their movements and see if you can determine how they cause the jets of water to leave the cloacal siphon. What reason is there for forming such strong jets of water?
6. The heart and ganglia are nicely shown in such a specimen.
7. Remove one of the shell valves of an adult specimen and examine the organs. An elongated sense tentacle occurs on one or the other side of the base of the branchial siphon, between the wall of the siphon and the corresponding mantle lobe.

A drawing of the organs will prove profitable.

## MYTILUS OR MODIOLA. (Mussels.)

These animals belong to the order Filibranchia, and show comparatively simple gills, as well as interesting modifications for their manner of living. They live attached to stones, shells, piles, or even to sand grains, sometimes in moderately deep water, but frequently between low- and high-tide marks. The two forms may easily be distinguished by the positions of their beaks. The beaks of Mytilus form the anterior end of the shell. Those of Modiola are placed a short distance posteriorly. You should visit "mussel beds," and see where and how they are attached and on what they must depend for food.

1. Place young specimens in dishes of sea-water and see if they will attach themselves by their byssal threads. (They will generally require some hours.) If you can get them to attach to slides, the attachment may be microscopically examined.
2. Test the strength of the byssal threads of a rather old
specimen. Are they elastic? How would elasticity aid the animal in remaining attached?
3. Leave specimens in sea-water for some hours, and see if they change their positions.
4. Notice the margins of the mantle. Are they fused? Why are siphons not necessary? See if you can find where water passes in and out.
5. Wedge the valves of a specimen apart, cut the adductor muscles (take note of their relative size), and examine the arrangement of organs.
6. Find where the byssal threads are attached.
7. Notice the relatively small foot, and compare it with the powerful foot muscles. Why are such powerful foot muscles necessary?
8. See how the gills are attached to the body. The filaments of the gills of this form are very loosely attached to each other by modified clumps of cilia, that represent the inter-filamentar junctions. Cut off a piece of a gill, mount it in sea-water under a cover, and examine with low and high powers. Find places where filaments are attached by the bunches of cilia. Find places where the cilia have pulled apart. Notice the size and shape of the ostia and find the two kinds of movable cilia.

9 . This form usually shows the way food is gathered especially well. Place powdered carmine on the surface of a gill and see what becomes of it.
10. Notice the thickened condition of the mantle. The gonads extend into them, and the thickening is due to sexual products.

Drawings of the arrangement of the organs, and especially of the microscopic structure of the gill, will prove profitable.

## PECTEN IRRADIANS. (Scallop.)

This species belongs in the order Pseudo-lamellibranchia and lives on muddy or sandy bottoms, generally where the water is from a few inches to several fathoms deep. It has the power of swimming pretty well developed. At rest on the bottom it always lies on the right valve of the shell.

1. Do the valves of the shell differ in color or shape?
2. On each side of the beak of each valve is a flattened projection frequently called an "ear" or "wing," the posterior of which slopes backward, while the anterior, especially the one on the right valve, is somewhat separated from the body of the shell by a notch.

Place specimens in dishes of sea-water, and when they have opened their shells notice:
3. The bright specks, the pallial eyes, along the margins of the mantle. Are they placed in any order?
4. The arrangement of the tentacles on the margins of the mantle. Why should sense organs be placed in this position?
5. The mantle and see if it is sensitive. How far can it be drawn back into the shell? What muscles are used in withdrawing it? Why is it necessary to withdraw it?
6. Specimens in aquaria will often swim. If possible, notice how this is accomplished.

Wedge the valves of a specimen apart and notice the single large adductor muscle. What need has Pecten for such a large adductor? Notice the foot and compare it with the foot of Venus.

How are the gills attached to the body? What would be the effect on the gills if they were attached to the mantle and to each other, as in most forms, when water is ejected in swimming.

Examine the structure of the gill and notice how much larger the inter-filamentar junctions are near the inter-lamellar junctions than elsewhere. Near the margins of the gills the junctions are frequently simple bunches of cilia, as in Mytilus. Observe the muscular movements of the gills. The gills of this form need to be muscular so they can be drawn together when the animal swims.

Drawings to show the arrangement of the organs and the structure of the gill are desirable.

## OSTREA VIRGINIANA, (Oyster.)

This also belongs to the order Pseudo-lamellibranchia. It forms a good example of adaptations for a sedentary life. It occurs, fastened to rocks and other shells, in positions where it is much exposed to attacks of the enemies of lamellibranchs.

1. Notice the difference in the size and shape of the valves. Why is this desirable?
2. Notice the thickness of the valves and the completeness with which they come in contact when the shell is closed. Would such a heavy or tight-closing shell be satisfactory for the scallop or the razor-shell clam?
3. Open the shell by breaking the edge, inserting a knifeblade through the opening, and cutting the adductor muscle away from the flattened left valve of the shell and notice the single adductor, extensive gills, and the absence of a foot.

## SOLENOMYA.

This form, a member of the order Protobranchia, with much the same structure as Yoldia, shows an interesting method of swimming that should be compared with Pecten, and with the jets of water formed by Mya. Specimens may be dug at low tide from mud or sandy mud, placed in a dish of sea-water, and observed. Does the posterior opening in the mantle chamber correspond to typical siphons? See if you can find how the animal swims. Is the movement continuous or jerky? Does the animal move forward or backward? Is the foot active? Are jets of water thrown from the shell? Is the animal adapted to forming jets of water?

Examine a specimen that has the valves closely drawn together and see how rounded the margins appear. Examine a shell from which the animal has been removed by maceration and see the relation of the shell cuticle and the calcareous portion of the shell. What becomes of the marginal cuticle when the shell is closed? Can this have anything to do with throwing jets of water from the shell?

## MYA ARENARIA. (Long Clam.)

This animal belongs to the order Eulamellibranchia, as does Venus, and is introduced because of adaptations for its manner of living. It lives buried in the mud, in which as an adult it remains stationary. You should find a "clam bed" along the shore, and after noticing the pits in the surface of the mud, and the jets of water that are sometimes thrown from the pits, dig down and see how the animals are placed. If the water is calm, see if you cannot find the openings of the siphons at the surface of the mud, of specimens that are still covered by water. You will need to walk very carefully so as to disturb mud and water as little as possible, as the siphons are otherwise closed and withdrawn.

1. Why does this animal not need a shell that is as heavy and closes as tightly as that of Venus? Does it show the same points regarding the valves (umbos, beaks, lines of growth, and ligament)? Later, when the shell is removed, the large cartilage pit on the left valve will be seen.
2. The ventral borders of the mantle lobes are united except near the anterior end, where there is a space through which the foot may be seen.
3. The siphons are large and muscular and may be retracted, as in the specimen that you are handling, or may be greatly extended, as may sometimes be seen in aquarium specimens. Why does Mya need larger siphons than Venus does?
4. Pick up a specimen that has the siphons extended and notice the powerful ejection of water. Is it ejected from one or both openings? How is this accomplished? Of what service can such jets be to the animal? Why are powerful jets of this nature of more service to Mya than to Venus?

Notice the cartilage in the cartilage pit on the left valve. What function does it perform? Why is there no need for a large and powerful foot? It is much easier to trace the alimentary canal and the ganglion connectives in this form than in $V e$ nus?

## ENSIS DIRECTUS. (Razor-shell Clam.)

This species is another representative of the order Eulamellibranchia and is introduced because of its adaptation for a burrowing habit, and because of the great ease with which its nervous system can be studied. Individuals are not uncommon on mud- or sand-flats from which the water flows at low tide. They may sometimes be seen protruding above the surface of the mud, but are hard to approach because of their great sensitiveness. Upon being disturbed they quickly disappear beneath the surface of the mud.

1. Notice the shape of the shell, the way it gapes at both ends, and the way the lobes of the mantle are fused.
2. With a pencil-point or seeker stroke the tentacles around the ventral mantle opening, and around the siphon openings, while the animal is being held anterior end downward. This will cause it to perform the burrowing movements. Study the movements carefully and see what the effects would be were they performed in mud. Thrust the anterior end of the shell in mud and watch the result of the movements.
3. Water is ejected by the sides of the foot to aid in burrowing or to enable the animal to swim, but observations on its method of ejecting it are not easily made, and are sure to take much time. Notice the way the anterior margins of the lobes of the mantle scrape mud from the foot when the foot is being withdrawn.
4. With a scalpel separate the united margins of the mantle throughout their length. Slowly pry the valves apart, lift up the free end of the foot and pull it posteriorly.

The cerebral ganglia are plainly visible without further cutting. They lie just posterior to the anterior adductor muscle and in front of the mouth, and are widely separated. They are connected by a narrow commissure, and each gives rise to a cerebro-visceral and a cerebro-pedal connective and to a number of nerves. The nerves that supply the anterior part of the mantle and the anterior adductor muscle are especially easily seen.
5. If the specimen is one that is nearly or quite dead, it is, by cutting, easy to follow the cerebro-pedal connectives to the pedal ganglia, which are not far from the base of the foot and not deeply embedded.
6. Allow the foot to return to its normal position and cut along the line of union of the inner gills. Without further cutting the visceral ganglia may be studied. Their connectives, which may be followed easily as far forward as the palps, and the posterior pallial and the branchial nerves may be seen.

A drawing of the nervous system should be made.

## AMPHINEURA.

## CHAETOPLEURA.

It will be profitable to study only external features, unless time is to be had for cutting and studying sections, as the species is small and difficult to dissect. Its apparently generalized structure, and its adaptations, make it desirable for students to understand from descriptions and figures the main features of its anatomy.

1. Examine specimens that are attached to stones and shells and see how nicely they adapt their shapes to the shapes of the objects to which they are attached. How is this possible?
2. Remove a specimen and quickly transfer it to a clean glass slide, applying its ventral side to the glass. Put your finger in its back and prevent it from curling for a minute. It will then generally remain attached to the slide and may be studied from both sides.
3. How many plates are there? What is the shape of each? Do they apparently join edge to edge or do they overlap? Do the plates extend clear to the margin of the animal? What reason is there for having plates instead of a solid dorsal shell?
4. Notice the thickened margin of the animal, and see that dorsally it bears spicules, while ventrally it is smooth and is applied closely to the slide.
5. Notice the flattened elliptical foot. Do you understand how the animal creeps and adheres?
6. In front of the foot is the head fold in which the mouth can be seen.
7. In the furrow bordering the foot are the gills.
8. Remove the animal from the slide and see how it curls up. Try to unroll it.
9. If you care to see the radula, the organ that especially indicates affinity to the Gastropoda, it can be pulled out by grasping just behind the mouth with pointed forceps and pulling forward. When removed it may be mounted on a slide with water and studied with the microscope.

## GASTROPODA.

A majority of the Gastropoda have the body protected by a spirally wound shell, and crawl around by means of a flattened muscular foot that forms the ventral portion of the body.

Examine specimens of Tritia or any other active form and notice:

1. The relation of the animal to its shell when retracted and when extended.
2. Movements. Can you determine how are they performed?
3. The movements of the tentacles and proboscis. What do the movements accomplish?
4. Touch a specimen and see what positions the parts take when it retracts into the shell. If the animal has an operculum see where it is borne and how it fits into the aperture of the shell.

## SYCOTYPUS.

This large gastropod lives in comparatively shallow water and depends largely on other Mollusca for its food. Examine a retracted specimen and see how the shell is closed by a horny lid, the operculum. Examine expanded specimens in the aquaria, and see where the operculum is placed. What position
must the animal assume in the shell to bring the operculum in position?

Shell.-A somewhat conical tube, spirally wound, somewhat like a spiral stairway. Observe the following parts:

1. The apex, forming the closed end of the tube.
2. The spire. How many whorls are there? Do they differ in number in different specimens? In what direction are the whorls wound? (Hold the apex toward you in determining this point.) Examine old and young specimens and see if there is evidence that the apex is worn off.
3. The body-whorl. The one that opens to the outside.
4. The columella. The axis around which the whorls are wound. This is best studied in a broken shell.
5. The aperture, which is bounded by the inner lip on the columellar side and by the outer lip along the free edge.
6. The siphonal canal, which forms the spout-like prolongation of the shell.
7. The lines of growth. What do they represent? Do they show evidence of injuries that have befallen the shell during the life of the individual?
8. In structure the shell presents three layers. In a broken shell notice: (a) the cuticle, worn away from the greater portion of the shell; (b) the nacre, smooth and lining the inner surface of shell; (c) the middle layer. How can three layers be secreted by the mantle that lines one of them?

Draw two figures, one of a perfect and one of a broken shell.
9. Compare the shell with the shells of other forms, such as Lunatia, Bulla, Haliotus, Crepidula, and Acmæa.

Soft Parts.-Examine an animal that has been removed from its shell and killed while more or less expanded ${ }^{1}$ and see in what

[^12]position it was placed in the shell. Compare the number of whorls made by it to the number in the shell. Understand which is right and which is left for the coiled part of the body. Which side was applied to the columella? In determining the position of organs, constantly keep the sides in mind.

Before beginning the dissection, note the following parts:

1. The visceral dome. The portion that extended into the spire of the shell.
2. The mantle, which is thin and closely applied to the visceral dome, and raised to form a thickened collar that extends entirely around the body along a line that corresponds to the aperture of the shell.
3. The siphon, which is a spout-like prolongation of the collar. Into what portion of the shell does it fit?
4. The mantle chamber. This can be seen by raising the edge of the collar of the mantle.
5. The head, which forms an anterior prolongation.
6. The tentacles, forming two triangular projections on the head.
7. The eyes, pigmented spots on the outer edges of the tentacles.
8. The proboscis, which, when extended, protrudes from beneath the portion that bears the tentacles. What is its size, shape, and general appearance? It may be retracted entirely into the body.
9. The mouth, at the end of the proboscis. The end of the odontophore may frequently be seen protruding from the mouth.
10. The foot. What is its position, consistency, color, and shape?
11. The opening of the pedal gland, on the sole of the foot. Is the pedal gland well developed in both sexes? Do you know its function?
12. The operculum. Notice its position and attachment.
13. If the specimen is a male, the large, somewhat flattened and bent penis, a little to the right and posterior to the right tentacle.

A number of organs may be seen through the somewhat transparent mantle. These are:
14. The liver, which forms the first two whorls of the spire. Notice its color.
15. The gonad, which is borne on the dorsal surface of the liver, and differs in individuals from red and brown to yellow.
16. The stomach, which lies on the left (external) surface of the liver. It is curved and light colored and is frequently rather indistinct.
17. The kidney, which lies on the dorsal surface, and a little to the left side, on the anterior end of the liver. It is somewhat rectangular in shape and differs in color from a yellowish-brown to a chocolate color. The kidney is composed of two parts, the large acinous portion, and the smaller tubuliferous portion. The latter lies along the left side of the former, by the side of the pericardium.
18. The pericardium lies to the left of the anterior end of the kidney. Through its dorsal wall the yellowish heart can generally be seen.
19. The columellar muscle, which attaches the animal to its shell, and enables it to withdraw, can be traced to the foot.
20. If the specimen being examined is a female, the large yellowish nidamental gland will be seen near the right side.
21. The large, brownish gill lies to the left of the nidamental gland in the female and anterior to the heart.
22. The osphradium is a small, brownish organ to the left of the anterior end of the gill and at the base of the siphon.
23. The hypobranchial gland is a glandular portion of the mantle, to the right of the gill (between the gill and the nidamental gland, in the female).

Make a drawing of the animal as a whole, showing as many of the observed points as possible.

Open the mantle chamber by cutting the mantle along the right side of the gill to the limit of the cavity, reflect the flaps, and notice the position and structure of the gill, osphradium, hypobranchial gland (cut in opening the mantle cavity), and, if
the specimen is a female, the nidamental gland. The opening of the rectum will be seen at the end of a short papilla in the right side of the mantle cavity. The opening of the nidamental gland will be seen on an elevation a little to the right and anterior to the anus. If possible, insert a guarded bristle into this opening and see what becomes of it. Trace the oviduct from the ovary along the columellar side of the liver. See what becomes of it. Examine the inside of the nidamental gland and see its relation to the oviduct.

If the specimen is a male, follow the vas deferens from the testis to the base of the penis.

Circulatory System.-Remove the thin membrane that forms the roof of the pericardial chamber.

1. The heart consists of : (a) the large, rounded ventricle; (b) the smaller, conical, thin-walled auricle.
2. The auricle receives blood by two vessels. One, returning blood from the gill, runs along the left side of the gill to its posterior end, where it bends abruptly to the right along the margin of the pericardial cavity, and enters the auricle. The other returns blood from the tubuliferous portion of the kidney and follows the right side of the pericardium to the auricle.
3. The gill receives its blood through a vessel that borders its right side. This vessel receives the blood from a portion of the mantle, and from the large, acinous portion of the kidney.
4. The blood leaves the ventricle by a single vessel, the aorta, that almost immediately gives rise to the visceral artery which supplies the visceral hump. Trace its distribution.

The aorta makes an abrupt turn downward and forward and enlarges to form the secondary heart which lies alongside the esophagus. Follow the course of the aorta and its branches.

The course of general circulation is, beginning with the heart, (a) system, (b) kidney, (c) gill, and (d) back again to the heart. Why is such a course of circulation better than the reverse?

Draw a figure showing the vascular system.
Excretory System.-The two portions of the kidney have
already been noticed. Cut along their common line of union and examine the inner surface of each part.

1. Notice the parallel lines of tubules that form the substance of the tubuliferous portion, and the lobules that form the comparatively thick walls of the acinous portion.
2. Find the slit-like opening that leads from the kidney to the mantle cavity. It is at a point between the two portions of the kidney and is easily found from the mantle chamber. A small opening leads into the pericardium, but it is hard to find it in dissections.

Digestive System.-1. Remove part of the integument at the base of the proboscis and find the muscles that retract it. How many are there and how are they attached? Do you understand how the proboscis is extended?
2. With a pair of scissors open the extended proboscis along the ventral line, pin it open, and notice that the exposed muscular mass, the buccal mass, is attached to the wall of the proboscis in the region of the mouth, at its base, and by means of fibers, along its sides.
3. Push the muscular mass slightly to one side and notice the esophagus, which is closely applied to the dorsal wall of the proboscis. Notice the muscle fibers that extend from it to the proboscis. What is their function?
4. Examine the buccal mass.
(a) The strands of muscles that run from its sides forward to be inserted on the walls of the proboscis are attached to a cartilage, the odontophoral cartilage. These are the cartilage protractors.
(b) Running lengthwise of the buccal mass, on its ventral side, are three pairs of slender muscles, one pair median and the others covering the horns of the odontophoral cartilage that has just been observed. Find to what the muscles are attached anteriorly and posteriorly. If the animal is fresh, pull on them with the forceps and see what moves. These are the radula protractors.
(c) Beneath the radula protractors observe the sheet of crossfibers that bind the horns of the odontophoral cartilage together.

Make a drawing showing the ventral side of the buccal mass.
(d) A portion of the radula is visible near the anterior end of the proboscis. Introduce a bristle into the esophagus and determine its relation to the exposed radula.
(e) Loosen the anterior end of the buccal mass from the wall of the proboscis, turn it back and see how the radula passes around the odontophoral cartilage. With a hand-lens notice the teeth on the open radula, ventral to the cartilage, and see how the radula is folded as it passes over the dorsal side of the cartilage so the teeth are turned in. What reason is there for folding the radula in this manner?
(f) Cut the cartilage protractors and reflect the buccal mass. It is attached to the wall of the proboscis at its posterior end by strong muscles, the radula retractors. These may be studied after cutting the sheath of cross-fibers that hold the mass together. Determine how they are attached to the sides of the radula. Why do they need to be so powerful?
(g) The retractors of the cartilage are attached to the ends of the horns and extend posteriorly to the walls of the proboscis.

Make a drawing of the buccal mass as seen from the dorsal side.
(h) Pull away the muscles and examine the shape of the odontophoral cartilage and its relation to the radula.
(i) Remove the radula, unfold it, and examine it microscopically. Do the teeth differ in any way at the two ends? Why is the radula so long?

Draw a portion.
The radula is the organ upon which most gastropods depend for getting food. You should understand how:

1. The proboscis is protruded and retracted.
2. The odontophoral cartilage is protruded and retracted.
3. The radula is protracted and retracted.
4. The radula is folded by the cartilage and spread for action.
5. The food is torn off and taken into the mouth.

Near the base of the proboscis is a pair of large, yellow
salivary glands, the ducts from which extend on either side of the esophagus to the mouth. Further back, on the right side of the esophagus, is the delicate pancreas.

Trace the esophagus to the stomach and the intestine to the anus.

Nervous System.-Most of the ganglia are grouped around the esophagus, about three-quarters of an inch posterior to the base of the proboscis. They are all brown and accordingly conspicuous. Cut around its base so the proboscis may be turned back, and the ganglia on the ventral surface of the esophagus may be seen. Carefully pick away the tissue that covers the ganglia and notice on the ventral side of the esophagus:

1. The small but conspicuous buccal ganglia. These are united with each other and with the cerebral ganglia and send nerves to the mouth apparatus.
2. The large pedal ganglia, fused together but distinctly paired, lying posterior to the buccal ganglia and sending nerves to the two sides of the foot. Each is united by connectives with the corresponding cerebral and pleural ganglia.

From the dorsal side a number of ganglia may be seen, more of which lie to the right than to the left of the median line.

1. On the left side there are two ganglia that are in rather close union with each other. The most anterior, the left cerebral, is the larger of the two. The left pleural joins it posteriorly and ventrally and extends nearly to the ventral side of the esophagus.
2. On the right side four ganglia may be distinguished. The right cerebral and right pleural are fused to form one mass, but there is a marked constriction between them. Posteriorly and dorsally the right pleural is connected by a connective with the right visceral, which lies very close to it. The remaining ganglion, the left visceral, which is almost in contact with the right pleural and right pedal ganglia, lies ventrally and to the right of the right visceral ganglion. It is connected with the left pleural ganglion by a connective that runs behind the pedal ganglia. There seems also to be a connection with the right pleural ganglion, but this must be considered a secondary connection. Do you understand why this ganglion occupies this position?
3. Another ganglionic mass, the abdominal ganglion, possibly formed by the fusion of two ganglia, lies just below the external opening of the kidney, where it can be seen as a brown mass, through the body-wall. It lies on the elongated commissure that connects the two visceral ganglia. The commissure may be followed by dissection.

The cerebral ganglia are the most centralized. Besides being connected with each other by a commissure dorsal to the esophagus, and being intimately connected with the pleural ganglia, each cerebral ganglion is connected with the corresponding buccal and pedal ganglion, and, through the pleural, with the visceral ganglion. The visceral ganglia are connected with each other by a long commissure on which the abdominal ganglion is placed. Each pedal ganglion receives connectives from the cerebral and from the pleural ganglion of the corresponding side.

Draw figures of the nervous system and compare them with the clay model. ${ }^{1}$

## LOLIGO PEALII. (The Squid.)

Specimens of this or closely related species are rather common along the Atlantic coast of the United States. They are active swimmers, but may occasionally be seen in shallow, quiet water near the shore. The movements and positions of adult specimens in aquaria should be studied. Do you know what they eat and how they capture their food?

Study a small living specimen in a jar of sea-water and notice:

1. Its general shape and distinct head.
2. Its position in the water. For convenience, the lower surface may be referred to as ventral, but this is not to be considered as morphologically the same as the ventral surface of other Mollusca. What parts are kept moving? Why is water pumped when the animal is not swimming?

[^13]3. In what direction it can swim best. Can it swim in the other direction? How does it swim? Is the funnel movable? How does it guide its movements?
4. Its color. Irritate it and see what happens. What purpose does the change in color serve? What is the ink for?
5. What happens when the end of a finger is placed within the circlet of tentacles of an animal about two inches long that is being held firmly?

Using an adult specimen, observe:
6. The arrangement of the arms on the head. Are they arranged in any definite order? Are they all alike?
7. The suckers of the arms. Do they follow the same order on all of the arms?
8. The structure of a sucker. Notice the peduncle, outer thin margin, horny ring, and piston. Is the horny ring smooth? What is its function? How does the sucker work? Split one and draw the cut surface.
9. The mouth. Where is it placed? Notice the tips of the horny beak. Which jaw is longest?
10. The eyes.
11. The fold of tissue behind each eye. These have been called the olfactory organs.
12. The attachment of the head and the extent of the mantle opening around the neck.
13. The funnel protruding from beneath the mantle on the ventral surface. Notice the position and character of its opening.
14. The median dorsal projection of the mantle.
15. The tail-fin, its position, shape, and color. What is its function?

Draw the animal as seen from the ventral side.
Carefully open your specimen by cutting through the mantle a little to one side of the mid-ventral line, and expose the mantle chamber.

Notice:

1. The thickness and character of the mantle and its relation to the rest of the body. Why does it need to be so muscular?
2. The arrangement of the funnel. Why does it have a thin posterior edge? How is it held in position against the mantle? Does it have a valve? Is the siphon movable in the living animal?
3. The free edge of the mantle and its relation to the folds beneath the eyes. Do you understand how the water gets into and out of the mantle cavity?
4. The large retractor muscles of the funnel. How many are there? How can the funnel be pointed in different directions? What need is there for such a provision?
5. The retractor muscles of the head. How many are there? Are they used in swimming in any way?
6. The rectum, opening near the base of the funnel between two small lateral flaps of tissue.
7. The ink-bag, dorsal to the rectum and opening into it near the anus.
8. The gills, extending from a point about midway of the body toward the free edge of the mantle. How many are there? How are they attached? Why does an animal that is not swimming continually pump water through the mantle chamber?
9. The branchial hearts, at the bases of the gills, rounded, light colored organs that can be seen through the membrane covering them.
10. The median ventral mesentery.

If the specimen is a male, notice:

1. The slender, tapering penis, to the left of the rectum.
2. The kidneys, white organs to be seen through the membranous covering, between the bases of the gills. From this position they taper anteriorly for half an inch or more and send small lobes posteriorly.
3. The openings of the kidneys near their anterior ends, on small papillæ.
4. The conical posterior portion of the viscera. This is composed of a large visceral sac and portions of the sexual organs.

Draw the animal, showing the points observed.
If the animal is a female, notice:

1. The pair of large, white nidamental glands that cover a portion of the rectum and the greater part of the ink-bag.
2. The openings of these glands at their anterior ends. Do you know the function of these glands?
3. The opening of the oviduct dorsal to, and a little to the left of, the left nidamental gland.
4. The mass of eggs that fills the posterior portion of the body. These are in the ovary and oviduct.

Draw the animal, showing the points observed.
Excretory System.-If the animal is a female, remove the nidamental glands, and the kidneys will be seen in the position described for the male. The kidneys consist of:

1. The white, somewhat triangular, glandular portions already noticed, extending from the region of each branchial heart anteriorly, and forming a portion of the walls of the pre-cavæ.
2. The cavities of the organs lying ventrally, and at the sides of the glandular portions.
3. The external openings, at the ends of small papillæ, on either side of the rectum near the anterior ends of the kidneys.

Digestive System.-Remove the funnel and its retractor muscles and carefully lay the head open, along the ventral side, to expose the buccal mass. This is a rounded, muscular organ, with a ring of tissue at its anterior end that surrounds the horny jaws. Examine the jaws and see which is the larger. Trace the narrow esophagus from the posterior end of the buccal mass backward. At the base of the head it enters the liver, a large, white organ that lies between the retractor muscles of the head, and extends from the base of the head to a point dorsal to the external openings of the kidneys. Lying close to the esophagus and covered by the anterior end of the liver is an elongated median salivary gland, the duct from which follows the esophagus into the head. The esophagus leaves the liver about midway of its length, and follows along the ventral surface nearly to the stomach. Before entering the stomach the esophagus passes the pancreas, a white, lobed organ that lies just beneath the glandular
portion of the kidneys, and the systemic heart, a roughly diamondshaped organ that lies between the branchial hearts.

The stomach proper is a rather small, thick-walled sac that lies on the right side of the body, dorsal and posterior to the right branchial heart. From the left side of the stomach a rather large opening leads into a thin-walled blind sac, the visceral sac, that when filled with partly digested food, as it frequently is, extends posteriorly to the end of the body and occupies a considerable part of the conical portion of the body. When empty, it is quite small and inconspicuous.

The intestine leaves the stomach very near the point the esophagus enters, and just anterior to the opening that leads into the visceral sac. It passes ventrally, and becomes visible from the surface, where its position has already been noted.

Draw a figure showing the digestive system.
Cut a median sagittal section of the buccal mass and notice the mouth cavity, the jaws, the muscles that move the jaws, the tongue, and the position of the radula. Is the radula arranged in the strap-over-pulley manner that it is in Sycotypus?

Draw a figure of the section.
Male Reproductive System.-The position of the penis has already been noticed. Cut the tissue away from the base of the penis and notice the swollen spermatophoric sac in which the spermatophores are formed. Through the walls of the base of the penis and the spermatophoric sac, groups of slender, lightcolored spermatophores can be seen. They will be studied later. The vas deferens consists of three distinct parts:
(a) A narrow, straight portion that extends anteriorly from near the pointed, posterior end of the spermatophoric sac and lies along its left side. This is sometimes called the vas efferens.
(b) A swollen portion that lies on the left of the anterior end of the spermatophoric sac, the seminal vesicle.
(c) A narrow and convoluted tube that leads posteriorly from the seminal vesicle alongside the portion described in (a), to the capsule of the testis. This is sometimes referred to separately as the vas deferens.

The testis is a large, white organ that extends, from the region of the stomach, posteriorly to the end of the body, dorsal to the visceral sac.

Draw a figure of the male reproductive system.
Open the base of the penis and remove a number of spermatophores. Place them in a watch-glass in water and examine with a microscope. Notice:

1. The sheath.
2. The packet of spermatozoa.
3. The spiral discharging portion.

Draw.
Why are the spermatozoa contained in spermatophores?
Female Reproductive System.-The opening of the oviduct has already been noticed. Observe:
(a) The large, swollen portion, the oviducal gland, that lies dorsal to the left branchial heart.
(b) The oviduct leaving the dorsal portion of the gland. The oviduct is long and convoluted, and is frequently filled wrth eggs. This depends on the season of the year.
(c) The lighter colored, greatly lobulated ovary, also frequently filled with eggs, lying dorsal to the oviduct and visceral sac and extending from the region of the stomach to the end of the body. The ovary is inclosed in a capsule from which the oviduct leads.

The nidamental glands have been studied and removed.
Draw a figure of the female reproductive system.
Circulatory System.-An injected specimen is desirable. The blood that has been supplied to the body in general is collected by veins and carried to the branchial hearts. The vessels that collect the blood are:

1. The pre-cava. A single vessel carries the blood from the head to the anterior ends of the kidneys. Here the vessel divides into right and left pre-cavæ that are intimately connected with the kidneys. The pre-cavæ diverge near the posterior ends of the kidneys and enter the corresponding branchial hearts.
2. The post-cava. A pair of very large vessels that return
blood from the posterior end of the body. They join the corresponding pre-cavæ near the anterior borders of the branchial hearts.
3. The mantle-veins. These return blood to the branchial hearts from the anterior portion of the mantle.

The blood that is received by each branchial heart is sent into the corresponding gill through a vessel that leaves the heart near the opening of the mantle vein, and runs along the side of the gill that is attached to the mantle.

The blood is collected from each gill by a large branchial vein that runs along the ventral side of the gill, and enters the systemic heart.

Draw a figure showing the vessels connected with the branchial hearts.

Expose the systemic heart by carefully removing the superficial tissue between the branchial hearts, and notice that it is not symmetrical. Its lateral angles receive the branchial veins and it gives rise to an artery from each of the other two angles.

The posterior aorta divides almost immediately into three large vessels. These are:
(a) The median mantle artery which follows the edge of the ventral mesentery to the mantle.
(b) A pair of lateral mantle arteries which diverge posteriorly and supply the two sides of the mantle. Besides these large vessels there is a small vessel that runs anteriorly over the ventral surface of the heart and supplies the ink gland and rectum, and another one that runs dorsally and posteriorly to supply part of the reproductive system.
(c) From the dorsal surface of the heart, near its anterior end, a small vessel passes over the anterior and dorsal surfaces of the stomach and finally passes into the gonad.

The anterior aorta is larger than the posterior aorta. From the anterior angle of the heart, which is to the right of the median line, it follows a straight course alongside the esophagus to the head. A number of small vessels are given off along its course, and it is finally distributed to the head and arms.

Draw the vessels connected with the systemic heart, into the figure you have just made.

Nervous System.-The stellate ganglia may be seen through the transparent lining of the mantle, on either side of the neck, where the body joins the mantle. They send nerves to the mantle and are joined to ganglia in the head (the infra-esophageal) by connectives. Why does the mantle need such large, special ganglia? Other small ganglia are situated in the body, but the large and important ones are grouped in the head, where they are supported and protected by cartilages.

With a razor make a median sagittal section of the head of a squid and notice:

1. Dorsal to the esophagus a rounded mass, the supra-esophageal'ganglion, which is supposed to represent the fused cerebral ganglia.
2. Ventral to the esophagus the elongated infra-esophageal ganglion, which is supposed to represent the fused pedal and visceral ganglia and (together with the masses that connect the supra- and infra-esophageal ganglia around the esophagus) the pleural ganglia.
3. The anterior prolongation of the infra-esophageal ganglion to form the pro-pedal portion, which supplies nerves to the arms.
4. The small supra-buccal ganglia, lying dorsal to the esophagus, and a little further anterior than the ends of the pro-pedal portion. These are connected by connectives with the supraesophageal ganglia.
5. The infra-buccal ganglia, about the same size as, and lying ventral to, the supra-buccal ganglia, and connected with them by connectives that run around the esophagus.

Draw a figure of a sagittal section of the head.
Two large ganglia, the optic ganglia, lie against the eyes and will be seen in cross-sections of the head that will be studied later. A dissection of one side of the head will show one.

Open the animal along the mid-dorsal line and find the pen which is embedded in the mantle. After exposing it for its
full length, turn the flaps aside and see that it lies in a pocket. It probably represents a modified shell that has become entirely inclosed by the mantle. What is its function?

Pull the pen out of the mantle and draw it.
With a razor make cross-sections of a squid, a quarter of an inch thick, and arrange them in order, in a little water, as they are made. Identify the parts you have found in dissection.

Make drawings of the sections that pass through the infra-esophageal ganglion, through the eyes, through the liver, and through the heart.

If time permits, study prepared sections that have previously been made. The structure of the eye and the positions of the parts of the nervous system should receive special attention.

Specimens of other cephalopods, such as Octopus and Nautilus, should be compared with the squid and the adaptations that fit them for their particular lives noted.

## ARTHROPODA.

With segmented bodies that are provided with segmented appendages.

Class 1. Crustacea.
Usually aquatic. With a more or less hardened outer covering and many thoracic appendages.
Subclass 1. Entomostraca.
Usually small. Appendages little differentiated. The number of post-cephalic segments variable.
Order 1. Phyllopoda.
Appendages with leaf-like expansions. (Branchipus, Daphnia.)
Order 2. Ostracoda.
Free-swimming, with the body inclosed in a bivalve shell. Seven pairs of appendages. (Cypris.)
Order 3. Copepoda.
Body elongated and distinctly segmented (except in parasitic forms). Four or five pairs of biramous appendages. (Cyclops, Argulus.)
Order 4. Cirripedia.
Comparatively large and usually attached. Usually with six pairs of biramous appendages. Forms that are not parasitic are covered by calcareous plates. (Lepas, Balanus.)
Subclass 2. Malacostraca.
Usually of considerable size and generally highly organized. Except in one order, thorax of eight and abdomen of seven segments.
Order 1. Phyllocarida.
Body inclosed in a large, bivalve cephalic carapace. Abdomen of eight segments. Thoracic segments free from the head. (Nebalia.)
Order 2. Schizopoda.
Thoracic appendages all biramous. Shrimp-like in shape. (Mysis.)

Order 3. Decapoda.
Thoracic segments united with the head. Three pairs of maxillipeds and five pairs of legs, of which the first bear heavy pincers. (Homarus, Cambarus, Crangon, Eupagurus, Hippa, Callinectes, Cancer.)
Order 4. Stomatopoda.
Five anterior thoracic legs are maxillipeds. Eyes stalked. Gills borne on abdominal segments. (Squilla.)
Order 5. Cumacea.
Two anterior thoracic legs are maxillipeds. Eyes sessile. Small shrimp-like forms. (Diastylis.)
Order 6. Arthostraca.
First and sometimes second thoracic segments fused with the head. Eyes usually sessile. (Talorchestia, Gammarus, Caprella, Porcellio.)
Class 2. Arachnoidea.
Head and thorax fused and bearing six pairs of appendages. Respiratory organs lamellate and abdominal or replaced by tracheæ.
Subclass 1. Gigantostraca.
With lamellate abdominal gills. Coxal joints of legs used as jaws. Marine. (Limulus.)
Subclass 2. Arachnida.
Respiration by lamellate lungs or tracheæ. Four pairs of walking legs. Mostly terrestrial.
Order 1. Scorpionida.
Abdomen segmented, posterior portion slender and very flexible, ending in a sting. Four pairs of lung-books. Pedipalpi chelate. (Buthus.)
Order 2. Pseudoscorpionida.
Abdomen segmented, without slender posterior portion or sting. Pedipalpi chelate. Respiration by tracheæ. (Chelifer.)
Order 3. Pedipalpida.
Abdomen flattened and segmented. Pedipalpi simple or chelate. Two pairs of lung-books. (Phrynus.)
Order 4. Solpugida.
Body composed of three segments. Cheliceræ chelate. Pedipalpi elongate, simple. Respiration by tracheæ. (Galeodes.)

Order 5. Phalangida.
Body short and oval. Abdomen composed of six segments. Cheliceræ chelate. Pedipalpi and legs very long and slender. Respiration by tracheæ. (Phalangium.)
Order 6. Araneida.
Abdomen unsegmented and usually distinct. Cheliceræ end in claws that are provided with poison glands. Lung-books and sometimes tracheæ also are present. Spinnerets present on the abdomen. (Epeira, Agalena.)
Order 7. Acarida.
Body not divided into regions. Biting or piercing and sucking mouth-parts. Respiration by tracheæ or through integument. (Sarcoptes, Dermacentor.)
Supplementary to the Arachnoidea.
Pycnogonida.
(Doubtfully referred to the group.) Body composed of segmented cephalothorax and vestigial abdomen. Legs very long, angular, and containing portions of the viscera. No special respiratory organs. (Phoxichilidium.)
Class 3. Onychophora.
Elongated bodies with some annelid-like characters. Appendages short, numerous, and creased rather than jointed. Respiration by means of tracheæ. (Peripatus.)
Class 4. Myriapoda.
Generally elongated bodies with numerous jointed appendages. A distinct head bearing ocelli, antennæ, and jaws is present. Respiration by means of tracheæ.
Order 1. Symphyla.
With not more than twelve leg-bearing trunk segments. A single pair of branching tracheæ. (Scolopendrella.)
Order 2. Chilopoda.
With numerous trunk segments, each with a single pair of legs. First pair of trunk appendages forming poison jaws. Body dorso-ventrally compressed. (Lithobius.)

Order 3. Diplopoda.
With numerous trunk segments, each with two pairs of legs. No poison jaws. Body not compressed. (Julus.)
Order 4. Pauropoda.
With ten trunk segments and nine pairs of legs. (Pauropus.)

## Class 5. Insecta.

Body divided into head, thorax, and abdomen. Three pairs of thoracic legs and generally one or two pairs of wings.
Order 1. Thysanura.
No metamorphosis. No wings. Mouth-parts usually mandibulate. Some forms show vestigial abdominal appendages. (Lepisma, Sminthurus.)
Order 2. Orthoptera.
Metamorphosis direct. Two pairs of wings usually present, of which the anterior are usually tough and protect the more delicate posterior ones. Mouth-parts mandibulate. (Acridium, Gryllus, Periplaneta.)
Order 3. Neuroptera.
Metamorphosis direct. Two pairs of netted veined wings usually present. Mouth-parts mandibulate. Prothorax free. (Libellula, Termes, Hexagenia.)
Order 4. Hemiptera.
Metamorphosis direct. Two pairs of wings usually present. Mouth-parts piercing and suctorial. (Benacus, Cicada, Pediculus, Aphis.)
Order 5. Diptera.
Metamorphosis indirect. Wings, when present, one pair and membranous. Mouth-parts suctorial. (Culex, Tabanus, Musca.)
Order 6. Lepidoptera.
Metamorphosis indirect. Two pairs of scaly wings. Mouth-parts suctorial. (Platysamia, Anosia, Philampelus.)
Order 7. Coleoptera.
Metamorphosis indirect. Membranous hind wings folded and covered by modified fore wings, the elytra. Mouth-parts mandibulate. (Lachnosterna, Doryphora.)

## Order 8. Hymenoptera. <br> Metamorphosis indirect. Two pairs of membranous wings. Mouth-parts suctorial and mandibulate. (Apis, Vespa.)

## CRUSTACEA.

## HOMARUS AMERICANUS. (Lobster. ${ }^{1}$ )

These animals are not generally found where they can be readily observed in nature, but many valuable observations can be made on specimens confined in aquaria. If other animals are present in the aquarium notice the position of defense that is taken. In nature the animal probably spends much of its time under rocks with the anterior end of the body turned toward the opening. In this position both sense organs and weapons are in the proper position for attack or defense. Notice how the appendages are used. Are the sense organs moved frequently? Why are the eyes on stalks? What appendages are used in walking? Are all of these appendages used in just the same way? Does the animal move equally well in all directions? Perhaps you can make the animal swim; if so, observe the method. Feed a specimen with portions of a clam or fish, and see how food is torn to pieces and transferred to the mouth, and determine, if possible, how the mouth appendages are used.

External Anatomy.-As in Nereis, the body is segmented. The metameres of the head and thorax, however, are immovably fused to form a cephalo-thorax. This is covered dorsally by a single piece, the carapace.

1. Note, on the carapace, the cervical groove between the head and thorax, and the beak or rostrum forming an anterior spine. The ventro-lateral edge of the carapace is not attached. A flat object thrust between it and the body passes into the gill chamber. This free plate of the carapace is called the gill-cover. Do you know why the edge of the carapace is free? Notice the

[^14]hair-like spines along its free border. What purpose do these serve?
2. The abdomen is composed of seven movable segments, each bearing a pair of jointed appendages except the last, which is sometimes not considered a true segment and is called the telson. Each abdominal segment consists of a dorsal piece, the tergum, which is continued as a free plate laterally (the pleuron), and of a ventral piece, the sternum. Move the abdominal segments and see where they are hinged. How are the terga and sterna arranged to allow free movement? In the thorax the sterna, though fused, can be distinguished. There are eight segments in the thorax.
3. Appendages.-Aside from the stalked eyes, whose homology with true appendages is doubtful, there are nineteen pairs. These are, counting from before backward: antennules, antennю, six pairs of mouth appendages, five pairs of walking legs (pereiopods), of which the first are the claws or chelce, and six pairs of swimmerets (pleopods). In the male, the first two pairs of pleopods are modified to form copulatory organs.
(a) Turn one of the fifth pair of pleopods forward and examine its posterior aspect. It consists of a basal piece, the protopod, a lateral branch, the exopod, and a median branch, the endopod. This branched type of appendage is designated as biramous. What is its use? Compare with this the modified sixth pair of pleopods, called the uropods.

Make a drawing of one of the fifth pleopods.
(b) In front of the chelæ will be seen the sixth pair of mouth appendages, the third maxillipeds. Remove that of the right side and compare it with the fifth pleopod. In addition to the protopod, exopod, and endopod, it bears a long blade, the epipod, which extended into the gill chamber. The protopod is composed of two segments, coxopod and basipod; the endopod of five segments, ischipod, meropod, carpopod, propod, and dactylopod. The exopod is composed of one long and many short segments. How is the appendage modified to serve in feeding?

Make a drawing of the third maxilliped.
(c) Remove the remaining five mouth appendages and compare each with the third maxilliped. These are, beginning posteriorly, the second maxilliped, first maxilliped, second maxilla (with a broad paddle, the scaphognathite, the use of which should be understood), first maxilla, and the mandible. Just back of the mandibles are two small flaps, the paragnatha, which are not true appendages. Do you understand the use of each of these appendages? Most of the appendages have parts that may be compared with the typical biramous appendage, but they are much modified to serve special functions, and the exact homologies are not important. Between the mandibles note the mouth, bounded in front by the labrum.

Drawings of these appendages may be made if time permits.
(d) The antennex are biramous. Notice on the ventral side of the basal joint of an antenna the opening of the green gland or nephridium.
(e) The antennules, though branched, are not considered to be of the biramous type. Do you know why? Remove one and note on the dorsal surface of the basal joint a groove at whose median extremity is a small hole, the opening into the otocyst. Do you know the probable function of the antennules and of the otocysts? What reason is there for having both antennules and antennæ?
(f) Compare the pereiopods with the third maxilliped. Which is lacking, endopod or exopod? Examine each of the joints of one of these appendages and see in what directions the appendage may be moved. Are there any ball-and-socket joints? Compare the chelæ with the other pereiopods and see how they differ. To what part of a chela does the last segment of the last pereiopod correspond? What reason is there for having these appendages different? Do you think the arrangement of the appendages would aid the lobster in climbing over rough bottom?

Open one of the large chelæ and determine how the muscles are arranged to control its opening and closing. Which mus-
cles are strongest? Find how the muscles are attached to the "thumb."

Find the openings of the sexual ducts on the basal joints of the pereiopods; the last pair in the male, the next to the last pair in the female. In the female there is an opening into a seminal receptacle through a triangular elevation on the ventral side of the thorax.
4. Gills.-Remove the gill-cover of the left side, being careful not to injure the gills. Extending up into the gill cavity are seven epipods belonging to the three maxillipeds and the four anterior pereiopods. They separate the gills into groups. Each group will be seen to correspond to a segment. The gills show three sorts of attachments: (a) to the appendages themselves (podobranchs), (b) to the articular membrane between appendages and body-wall (arthrobranchs), and (c) to the bodywall itself (pleurobranchs). There are two arthrobranchs in some segments, one behind and above the other. How is the current of water forced through the gill-chamber? What is the function of the epipods? What direction must the water take through the gill chamber? Examine the structure of a gill. Move one of the appendages to which a gill is attached and see the effect on the gill.

Internal Anatomy.-Remove the carapace (beginning at the middle of the posterior margin and cutting forward, holding the cartilage knife parallel with the surface) as far laterally as the upper limits of the gill chambers and anteriorly to the base of the rostrum. What is the pigmented membrane for? Dissect it off so underlying organs may be seen.

1. The chitinous stomach lies near the anterior end with the ophthalmic artery running along its mid-dorsal line. Beside and behind the stomach are two masses of muscle which you have severed from the carapace. These are the mandibular muscles, and each is divided into an anterior and a posterior bundle. Lateral to these muscle masses are the yellow-green digestive glands, commonly called liver. Between and in front of the posterior mandibular bundle note the gonads, and follow one
forward by pressing aside the muscle mass. In the male the testis is a slender, white, convoluted cord, which ends blindly against the side of the stomach. The extent and position of the far thicker yellow ovary is much the same (unless the animal be mature, in which case it will be found greatly enlarged and orange).
2. The heart extends through the posterior third of the thorax. Remove the upper part of the delicate pericardium surrounding it, cut its arterial and other connections, and place it in water. Note the shape, the origin of the arteries, and the three pairs of valvular ostia. Do you understand how it receives blood?
3. Trace the gonads as far as the abdomen, noting the single anastomosis between those of opposite sides just in front of the heart. Beneath the heart the sexual ducts are given off-vasa deferentia in the male, oviducts in the female. Trace one outward and downward to its opening by removing a portion of the body-wall and of the basal joint of the proper leg.
4. Remove the posterior lateral body-wall forward to a position opposite the anterior third of the stomach. Pull the anterior lobe of the liver, which extends beneath the stomach, outward and backward. The liver will be seen to be attached to the pyloric end of the stomach (i.e., the smaller part, where the stomach passes into the intestine). Cut this attachment and note that it is really where the liver opens into the stomach. Just back of this point the right and left lobes of the liver are connected by a cross-branch passing beneath the intestine. Remove one liver lobe back to the abdomen. After having the circum-esophageal connectives pointed out, remove the stomach by cutting the esophagus, the intestine, and the bands of muscles attached to the stomach. Examine it in water, noting the cardiac and pyloric parts, the chitinous grinding and straining apparatus in the interior, and the muscles and plates that cause the movements of the grinding apparatus. Why does a lobster with chelæ and six pairs of mouth appendages need a gastric mill?
5. Between the circum-esophageal connectives medially and
the large antennary muscles laterally, note the oval excretory organs, called the green glands. They are covered by a very delicate membrane. Poke a small hole in one of the membranes and with a blowpipe show that it is really a thin bladder. Its opening on the antenna has already been seen.
6. Remove the dorsal wall of the abdomen and trace the posterior portions of the gonads, liver lobes, and intestine. In the sixth abdominal segment the intestine swells to form the chitin-lined rectum and gives off the blind intestinal caccum.

Circulatory and Nervous Systems. ${ }^{1}$-Remove the carapace of an injected specimen as before, also the gill-cover and gills on one side.

1. There can generally be seen, through the transparent body-wall, efferent branchial veins, which return the blood from the gills. These unite into six large ones which open into the pericardium at the side. Find these openings if possible. Do you understand how blood gets into the heart?
2. Note, at the anterior end of the heart, the ophthalmic artery and the two antennary arteries. Trace the former forward to the rostrum, cut it on the stomach and turn it forward for future study. Trace the antennary arteries to the mandibular muscles and cut them near the heart. Press the front end of the heart back and note the two small hepatic arteries. Each branches immediately, one division passing between the gonads, and the other laterally.
3. Remove the muscles on one side of the heart and examine it from the side, noting the great sternal artery extending downward, and the smaller dorsal abdominal artery running back above the intestine. Follow the latter through the abdomen.
4. Cut all arteries and remove the heart. Trace the antennaries through the mandibular muscles, noting the branch to the stomach.

[^15]5. Remove the thoracic viscera as before, follow the circumesophageal connectives forward and identify the cerebral ganglia in order not to destroy them.
6. Follow one antennary artery to the green gland, antennary muscle, eye muscle, etc.
7. Follow the distribution of the ophthalmic artery.
8. Remove the intestine and muscles of the abdomen, and find and trace forward the ventral nerve chain. Notice the position of the ganglia and the nerves that leave them and the connectives. In the thorax the ventral nerve chain passes beneath a system of chitinous plates (the endo-phragmal skeleton) and lies in a cavity, the ventral blood sinus. Note the enlarged subesophageal ganglion, the cross commissure just back of the esophagus, the nerves to the mouth appendages, nerves from the cerebral ganglia, and nerves from the other ganglia. What indication is there that the sub-esophageal ganglia represent more than a single pair?

Sketch the nervous system.
9. The sternal artery passes through the ventral nerve chain and then extends backward and forward as the ventral longitudinal artery. Remove the nervous system and follow this artery.

Draw a diagrammatic cross-section through the thorax, putting in one drawing the circulation from the heart through the sternal artery to the limbs and back through the gills to the heart.

## CALLINECTES HASTATUS. (Blue Crab*)

Crabs may be found in shallow water along shore, where they may be easily observed on quiet days. In what direction does the animal normally move? How are the legs used? What is the attitude of defense? Determine how the blue crab swims. What do crabs apparently use for food? Do they conceal themselves, are they protectively colored, or do they depend entirely upon their weapons for defense?

In studying the anatomy of the crab, constant comparisons should be made with the lobster.

External Anatomy.-1. The body is composed of cephalothorax and abdomen. Dorsally note the shape of the carapace and the position of the abdomen. The size of the abdomen differs in male and female. Why should it be larger in the female?
2. Note the antennce, antennules, and eyes, and see how they are packed away in recesses in the carapace. In the living animal see if any of these are frequently moved.
3. The third maxillipeds are flattened and cover the other mouth appendages.
4. Straighten the abdomen and note the anus. Compare the abdomen of a male with that of a female and both with that of the lobster. The dorsal side of each segment is covered by a tergum. The covering between each pair of pleopods is the sternum, the immovable flap lateral to them is the pleuron. Compare the abdominal appendages or pleopods of a male and a female.
5. The ventral side of the cephalo-thorax is covered by the sternal plastron. Note the eight sterna and six pairs of lateral episterna.
6. In the female find the openings of the oviducts in the fifth sternum.

Make a drawing of the ventral side.
7. Expose the gill chamber and compare the gill distribution with that of the lobster.
8. Remove the left third maxilliped entire, and compare it with the same appendage of a lobster. The protopod is composed of two segments (coxopod and basipod). The endopod has five pieces (ischipod, meropod, carpopod, propod, and dactylopod). The exopod has two large and many small segments. Attached to the coxopod laterally is an epipod which extended into the gill chamber.
9. Remove the remaining mouth appendages on the left side and compare them with the third maxilliped. They are: second maxilliped bearing epipod and two small gills; first maxilliped with an epipod; second maxilla with a flattened exopod, called the scaphoghathite, which has a special function that should be
understood; first maxilla, thin and leaf-like; mandible with two hard rods for the attachment of muscles.
10. Detach and examine one each of the eyes, antennules, and antennce. On the flattest side of the basal joint of the antennule note a dark suture-the scar of the former opening into the otocyst. Do you understand what function is performed by the otocysts? Near the base of the antenna find the opening of the renal organ (green gland).
11. Compare each of the five walking legs (pereiopods) with the third maxilliped. Which part, endopod or exopod, is lacking? Which bear forceps or chelæ? Why is this so? Note in the male the openings of the sperm ducts on the coxopods of the fifth pair.

Internal Anatomy.-Remove the entire dorsal part of the carapace.

1. Postero-laterally are two firm prominences, the flanks, containing muscles. What are these muscles for? Anterior to these are the gill chambers covered by a thin cuticle. Remove this and note the gills with their tips converging medially.
2. Between the gill chambers and flanks is the delicate pericardium. Remove this and find the heart with its ostia. Anteriorly it sends out an ophthalmic artery and two antennary arteries. Just anterior to the heart are muscles which were attached to the shell. What organs do they supply? The antennary arteries pass through the heads of a pair of the muscles.
3. In front of the gill chambers are the gonads. In the female the orange ovary will be seen lying on the yellow liver. In the male the slender, wavy, white cord, the testis, lies in approximately the same position.
4. The heart is attached to the pericardium by muscular strands. Cut these, and the three anterior arteries, and remove the heart, noting the two hepatic arteries beneath the antennary arteries, the great sternal artery passing downward from the under side, and the small abdominal artery just behind the last.

Draw dorsal and ventral views of the heart to show the ostia and the origins of arteries.
5. Cut across a gill and notice its afferent and efferent vessels. The latter is continuous with one of the sinuses which empty into the pericardial cavity. Can you determine how many sinuses there are? Do you understand how the heart receives blood?

Reproductive System.-Beginning antero-laterally, on one side, dissect out the reproductive organs, noting at the same time the distribution of arteries.
(a) Female reproductive organs.-Each ovary passes inward and backward, anastomoses with the one of the other side behind the stomach, and extends back to the abdomen. On a level with the posterior part of the stomach a branch passes downward and outward and is continuous with a dense, white organ, the seminal receptacle. Leave this receptacle in place but remove the entire ovary.
(b) Male reproductive organs.-The slender testis passes inward and backward, anastomoses with its fellow of the other side behind the stomach, and is continued as a thick, muchcoiled tube, the vas deferens, to the median side of the flank. It then runs forward nearly to the stomach, turns back again, and enters the substance of the flank. By removing the top of the flank and the upper side of the coxopod of the swimming leg, it can be followed to its external opening.

Digestive System.-(a) The liver fills all of the body-cavity not occupied by other organs. Remove the portion of it that is in the region of, and anterior to, the stomach, noting its connection with the alimentary tract.
(b) The stomach is a chitinous box divided into a larger cardiac and a smaller pyloric portion. Find the duct from the liver, and a slender, white, coiled tube, the pyloric cсгсит.
(c) Follow the delicate intestine back beneath the heart. Between the posterior edges of the flank is a white mass composed of a coiled tube, the intestinal ccecum. Remove the terga of the abdominal segments, follow this cæcum to its connection with the intestine, and follow the latter to the anus, noting its chitinous lining.
(d) Cut out the alimentary tract, open the stomach, and examine the grinding and straining apparatus.

Make a drawing of the alimentary canal.
Excretory Organs.-Examine the antennary gland (green gland) on the inside of the carapace opposite the base of the antenna. It consists of a thin bladder, and, anterior to this, a mass composed of a coiled tube which opens at the base of the antenna.

Nervous System.-Find the ring of ganglia around the ventral end of the sternal artery. ${ }^{1}$ Trace the nerves from this to the appendages and to the small abdomen. Trace the circumesophageal connectives around the gullet (they anastomose just behind it) to the cerebral ganglia. Along with the distribution of the ophthalmic and antennary arteries, trace the nerves from the cerebral ganglia to the eyes, antennæ, antennules, etc. Why should the nervous system be more concentrated than it is in the lobster?

Make a drawing of the nervous system.

## EUPAGURUS. (Hermit Crab.)

Examine a living specimen and see how it moves, and how the aperture of the shell is closed by the two large claws when the animal withdraws.

With a hammer crack the shell away from the animal and examine the twisted abdomen.

1. Has it lost its symmetry in appendages as well as in shape?
2. How many of the appendages have been retained? What is the function of these appendages?
3. Remove several other specimens from their shells and place them in a dish of sea-water together? Do they seem to understand that they are not protected?
4. Place an empty shell in the dish and see what happens.
5. Put more empty shells in the dish, but be sure they are not quite large enough for the crabs. Then add some larger

[^16]shells and watch the crabs test them to see which will serve best.

A drawing is desirable.

## HIPPA. (Sand Mole.)

On sand beaches, between low- and high-water mark, there may frequently be seen the shallow depressions that mark the places where these animals have burrowed. They may be dug out with a shovel, but they quickly disappear again.

1. Notice their shape and the ease and rapidity with which they burrow.
2. Place specimens in a dish containing sand and a little seawater and try to determine just how the burrowing is done. This may frequently be done by holding a specimen so it just touches the sand. Which end goes into the sand first? Notice the positions in which the appendages are held. Does this have anything to do with the direction in which it burrows? Does the animal jump or crawl? In what direction and how can it swim?
3. Examine the body and see if it is divided into head, thorax, and abdomen. Why has the telson such a peculiar shape?
4. Examine the appendages.
(a) The stalked eyes.
(b) The biramous antennules and the exceedingly long, feathery antennce. What is the usual position of the antennæ?
(c) The mouth appendages. Are strong, hard mandibles present? What must the character of the food be?
(d) The thoracic appendages. How many are there? Are they similar? Are there any chelæ?
(e) The abdominal appendages. Are they all alike? What functions are performed by them?

Make a drawing.

## SQUILLA.

Compare the animal carefully with the lobster, noting all of the important differences. The posterior three thoracic segments are free. The male possesses a copulatory organ on the basal joint of the last thoracic leg. In the female the opening of
the oviducts is in the mid-ventral line, on the antepenultimate thoracic segment. Examine the chelæ and compare them with the chelæ of a lobster. Are they homologous appendages in the two animals? If you have living specimens, study their movements while walking and swimming.

## A drawing of a side or ventral view will be profitable.

Internal Anatomy.-1. Remove the top of the carapace and abdomen. Beneath the muscles note the elongated, white tube, the heart, which extends from the stomach to the fifth abdominal segment. The anterior end is slightly enlarged and gives rise to the anterior aorta. The posterior end gives rise to a posterior aorta. Note lateral arteries and ostia. Remove the heart.
2. Beneath the heart, in the male, is a whitish, pigmented, flattened mass which consists of two convoluted tubes, the testes. Cut this mass across between the second and third abdominal segments and force it posteriorly. The two testes are continuous posteriorly. Follow them anteriorly and find the slender, dense, coiled vasa deferentia passing outward and downward at the posterior end of the third thoracic segment. Cut them and lay them back where they can be dissected later. The testes extend forward to the region of the stomach. Remove the testes.
3. Beneath the heart, in the female, are the two ovaries. Trace them forward and backward, and find the very slender oviduct that extends from each outward and downward in the region of the antepenultimate thoracic segment. Remove the ovaries, deferring the tracing of the oviduct.
4. Beneath the reproductive organs, in both sexes, is the granular liver. This consists of two lobes which extend from the stomach to the end of the telson. They form saccular diverticula between segments and in the telson. Where do they open into the alimentary tract?
5. Free the intestine, which is between the lobes of the liver. The rectum is in the sixth abdominal segment.
6. Pull back the anterior end of the stomach, identify the circum-esophageal connectives, in order not to destroy them,
and free the stomach by cutting the esophagus and intestine. Examine the stomach under water.
7. Trace the nerve chain. What ventral ganglia are fused? The cerebral ganglia are most easily exposed by slicing away, very superficially, the dorsal surface of the rostrum and pressing the eye muscles apart.

A drawing of the nervous system will be profitable.
8. Trace the genital ducts to their external openings.

## MYSIS.

If living specimens are to be had, watch them swim, and determine what parts are used in swimming. Does the animal swim in one direction or in both?

1. Compare the body with that of a lobster.
2. Are appendages present on each of the divisions of the body? Compare them with the appendages of the lobster? How do the thoracic appendages differ?
3. Notice the otocysts in the tail fin. What are their positions?
4. The living animal is transparent, and many internal organs, such as heart, gills, and portions of the alimentary canal, can be seen.

If time permits, make a drawing.

## TALORCHESTIA. (Beach-Flea.)

These active little animals inhabit sand beaches, where they burrow in the sand and hide in the decaying vegetable matter that accumulates along such beaches near high-water mark. Turn over some of this material and notice the activity of the animals that are disturbed. Most of them probably belong to another closely related genus, but their movements are much the same. How far can a specimen leap? Are the leaps of an individual continuously in one direction, so it may get away from the point of danger? Is each leap straight forward or does the animal whirl in the air? What purpose is served by the leaping? Try to catch a specimen. Determine how the leaping is accomplished. Determine how the specimens burrow.

If you will walk along a beach some quiet night with a lantern you will probably see something of the night activities of these animals.

1. Select a large specimen and count the number of segments. Is the body divisible into head, thorax, and abdomen?
2. The eyes are not stalked. Are they compound?
3. The second antennex of the male are very large. Compare them with the first antennæ and with the antennæ of a female.
4. Around the mouth are the labrum, forming an upper lip, the first maxillipeds (fused), forming a lower lip, and between them the mandibles, first maxillue, and second maxille.
5. Examine the appendages behind the mouth. How many are there? How many bear claws? Compare these claws with those of a lobster, and see how they differ. Which appendages are used in crawling? Why are some of the appendages arranged so they can be twisted around by the sides of the animal? What are the remaining appendages used for? Watch an animal jump.
6. Spread the appendages apart and find the gills, which are attached to the bases of the appendages.

Make a drawing of the animal.

## PORCELLIO OR ONISCUS. (Sow-Bug.)

These animals occur in damp places, such as under stones, logs, etc., and in cellars. They live for the most part on decaying vegetable matter. To what class of the Arthropoda do they belong?

1. Notice the shape. Is this an adaptation?
2. Is the body divisible into head, thorax, and abdomen? Count the number of segments. Is there any evidence of fusion at the posterior end of the body?
3. Examine the appendages.
(a) Are the eyes stalked or sessile?
(b) Only one pair of antennæ is present, the first pair being rudimentary.
(c) The mouth appendages are small. They consist of mandibles, two pairs of maxille, and one pair of maxillipeds.
(d) How many walking legs are there? Are these all alike?
(e) Notice the character and number of the abdominal appendages. On the posterior surface of all but the last pair, which are modified to form anal feelers, are gills. These are the only respiratory organs. Why must these animals live in damp places?

Make a drawing of the animal from the ventral side.

## CAPRELLA.

These animals are very common on hydroids, but because of their peculiar shape and slow motions are rather inconspicuous. Watch the animals and see how they move. Is the body kept at rest and moved by the action of the appendages, or how is movement from place to place effected? Are the appendages adapted for grasping? Why are they arranged at the two ends of the body? Watch specimens and see if you can determine on what they feed

The form is of interest because of its extreme modification to suit it to the needs of its life. There is some difference in the structure of the male and the female. The adult structure does not enable us to determine the homology of appendages.

1. Count the segments of the body. Do they differ in number and shape in male and female? The first represents the head with two fused thoracic segments. The abdomen forms a minute protuberance at the posterior end of the body.
2. Examine the appendages.
(a) At the anterior end of the body are the eyes, two pairs of antenno, a pair of maxillipeds, and a pair of legs.
(b) At the hinder part of the body are three pairs of legs.
(c) Near the middle of the body of the female, and near the anterior end in the male, is another pair of legs.
(d) On two of the segments which do not bear legs are gills. If time permits make a drawing.

## BRANCHIPUS. (Fairy Shrimp.)

These animals may be found in pools of fresh water in the early spring, just as the ice is leaving. Their method of swim-
ming by means of the large, expanded appendages should be observed.

1. Into what parts does the body seem to be divided? Do all of these parts show segmentation?
2. Find the following organs.
(a) The stalked, prominent eyes.
(b) The antennce. In the female the first are slender and the second vestigial. In the male the first are slender and the second are enormously enlarged to form a clasping organ.
(c) The labrum (not an appendage) forms an upper lip.
(d) The mandibles, beneath the labrum and by the sides of the mouth. Do they have cutting-edges?
(e) Vestigial maxillce behind the mouth.
(f) Swimming appendages. How many are there? Notice the fringe of hairs on each. What are these for? Remove one and examine it with a microscope. The lobes have been described as exopodite and endopodite, but their exact relationship is not certain.

A drawing is desirable.

## DAPHNIA.

This small fresh-water form frequently occurs in large numbers in small pools and brooks. Determine how it swims. Being small and transparent, it may be satisfactorily studied with a compound microscope.

1. Notice the shape and extent of the protective covering. To what part of other crustaceans does this correspond? Are the appendages and the abdomen capable of being thrust out? Are there any signs of segmentation of the body?
2. Determine what parts are used in keeping a current of water passing through the shell. Why is such a current needed?
3. If the animal carries young, notice how they are kept in the brood chamber by a spine that extends up from the dorsal portion of the base of the abdomen.
4. Notice the beating of the heart.
5. Examine the appendages.
(a) Are the eyes stalked or sessile? They frequently show a peculiar reaction to light. If the light is cut off from the microscope, the eye will be seen to rotate on its axis. . If the light is admitted again, the eye rotates back to its original position.
(b) The first antennce are very small and project ventrally. What is the chief function of the second antennæ?
(c) Several appendages will be seen inside of the shell, but it is hard to determine their exact relation. The functions of some of them may be apparent.

A drawing is desirable.

## CYCLOPS. (Water-Flea.)

Most any free-swimming copepod, either fresh-water or marine, will answer quite as well as the fresh-water Cyclops.

Cyclops may be found in almost any pool of fresh water and the marine forms are among the most abundant of the animals of the sea. Surface skimming of the sea, made with a net composed of cheese-cloth or silk bolting-cloth, will yield an abundance of material.

1. Watch the animals and see how they swim. With a pipet try to catch a certain individual and see whether the jerky movements probably aid these animals in escaping enemies. Determine what organs are used in swimming.
2. Examine specimens that have been confined under a coverglass with a microscope, and notice the shape of the body. Into what parts is it divided? Count the number of segments. Look for evidence of fused segments. Notice how the spines on the abdomen are arranged.
3. Examine the appendages.
(a) Do you find eyes that are equivalent to the usual arthropod eyes? Do you find an eye-spot? If such a spot is found, determine its position and shape.
(b) Which pair of antennce is largest? Why are the large antennee fringed with spines?
(c) Are there thoracic or abdominal appendages? Are any appendages other than the first antennæ used in swimming?
(d) The mouth parts consist of mandibles and two maxillx.
4. If the specimen is a female it may have two large egg sacs attached to the sides of the base of the abdomen. The female has two of the abdominal segments fused. In the male the segments are free.

A drawing of the specimen is desirable.

## ARGULUS. (Fish-Louse.)

These animals may be found on many species of fresh-water and marine fish. Notice their shape and determine how they cling to their host. Are they able to crawl? Can they swim?

1. Into what regions can the body be divided?
2. Examine the appendages and find:
(a) The eyes, the eye-spot, and the two pairs of small antennce.
(b) The sucking proboscis, composed of mandibles and maxillce, which lies between the suckers.
(c) The suckers, which are the modified second maxilla.
(d) The posterior (third) maxillipeds just behind the suckers.
(e) Four pairs of biramous thoracic appendages. What is their function?

Make a drawing of the animal.

## LEPAS. (Goose-Barnacle.)

If possible, examine a cluster of specimens as they naturally occur attached to floating timber.

1. Why are the peduncles much larger in some specimens than in others? Are they contractile so the body may be moved into different positions? Would such movements be of value?
2. Notice the abdominal appendages. Can they be thrust from the shell? What is their character? What are their characteristic movements? Drop a small piece of clam meat on these appendages of a living specimen and see what happens. What kind of food would they naturally collect?
3. Examine the portions of the shell. The portion on the closed, dorsal margin is the carina, laterally and near the base
of the peduncle are the scuta, and near the extremity the terga. Why are there so many pieces? Notice the lines of growth and determine the direction of growth of each piece.

Draw the animal as seen from one side.
With a scalpel or razor cut a preserved specimen into right and left halves, extending the cut through the peduncle.
4. The mouth will be seen at the end of a rather thick prolongation which is extended to near the bases of the abdominal appendages. On the margin of this prolongation are the small scale-like mandibles, first maxillce, and second maxillx. The stomach is rather large and the small intestine leads to the posterior end of the abdomen, where it opens between the abdominal appendages.
5. The nervous system, consisting of a large pair of cerebral ganglia and a short ventral chain of ganglia, should be seen in such a section.
6. The animal is hermaphroditic. The testes lie dorsal to the stomach and communicate with a conspicuous coiled vas deferens that is continued to the elongated penis at the end of the abdomen. What reason is there for such a long penis? The ovary occupies the interior of the peduncle. The oviducts are inconspicuous and hard to follow. They open near the bases of the anterior thoracic appendages.
7. Examine the appendages carefully and be sure that you understand the relation of parts. What part must the peduncle represent? Understand the beautiful adaptation of the animal for its life.

A drawing showing the organs is desirable.

## ARACHNOIDEA.

LIMULUS. (Horseshoe Crab.)
Notice the way in which the animal crawls upon the bottom. Is it well protected from enemies? Examine it carefully for parasites and for animals that are attached to it. Disturb it and see if it will swim. The animals are usually quite active
in the evening, and if you visit a car in which they are kept, at this time of the day, you are likely to find them crawling up the sides, falling over and swimming on their backs. In this position it is easy to determine how they swim. The animals are very hardy and will stand even complete removal from the water for days at a time. During the spring and early summer, eggs are deposited in the sand; the male holding to the edge of the abdomen of the female with claws modified for the purpose, is dragged after her. If possible, the method of egg deposition and fertilization should be observed.

1. The animal consists of a hoof-shaped cephalothorax, an abdomen, and a caudal spine. How are these joined? Is there any indication of segmentation of any of them?
2. Examine the eyes with a lens and see that they are compound.
3. On the lower side of the cephalothorax notice the appendages. Are they all built on the same plan? Compare them in male and female. Do you know what the modifications are for? Compare the pincers with those of a lobster. The first pair of appendages is called the chelicerce. Between the bases of the last pair of walking legs are the chilaria. Behind the chilaria is the operculum. Does this show evidence of being modified appendages? What is its function?
4. Between the bases of the cephalothoracic appendages is the mouth. Do the bases of the appendages show any modifications that may serve as teeth? Can the pincer-bearing appendages be so bent as to be used in feeding?
5. Along the sides of the abdomen notice the movable spines. How many are there?
6. Under the operculum are the gills. How many groups are there? Are they arranged in pairs? How are they attached to the body? Are they movable? What reason is there for moving them? Examine a bunch of gills, frequently called a gill-book, and see how it is formed.
7. At the base of the caudal spine notice the anus.

Make a drawing of the ventral surface.

Internal Anatomy.-This shows no very special adaptation and can be pretty well understood by studying a longitudinal section of a small preserved specimen.

In such a section the following organs may be found:

1. The dorsal extensor, the ventral flexor, and the leg muscles.
2. The elongated tubular heart just beneath the dorsal covering, in the posterior end of the cephalothorax and the anterior end of the abdomen.
3. The alimentary canal, consisting of the esophagus and the anterior and posterior portions of the stomach, which extends posteriorly without much change to the anus. The liver, which surrounds the stomach and fills the greater portion of the cephalothorax, sends its secretions to the stomach.
4. The cerebral ganglia, near the bases of the cheliceræ, and the ventral chain of ganglia should also be seen in satisfactory sections.

A drawing is desirable.

## BUTHUS. (Scorpion.)

Living specimens of these animals are not usually available for laboratory study. They live for the most part concealed during the day in crevices and holes and are active at night. Their food is largely spiders and insects which are seized by the claws and killed with the abdominal sting.

1. Into what parts is the body divided? How many segments are recognizable? Which are the most freely movable?
2. Look for eyes. Do you find any besides the large pair?
3. Find four pairs of slit-like openings on the ventral side of the pre-abdomen. These are the stigmata, the openings of the lung-books.
4. Find the following appendages:
(a) The chelicerce. What is their structure and where are they placed?
(b) The pedipalpi. Compare them with the cheliceræ and count their segments.
(c) Four pairs of walking legs. Count their segments and see if they are armed with claws.
(d) The comb-shaped pectines. Are they on the thorax or the abdomen? Their function is doubtful.
5. Examine the mouth. Are there any jaws? Is a labrum present?
6. Find the position of the anus. The terminal spine is provided with a poison gland and serves as a sting. In the living animal, the post-abdomen is habitually carried over the back.

Make a drawing of the under side of a specimen.

## EPEIRA. (Round-Web Spider.)

Examine the webs of different species of spiders and see how they are constructed. Do all of the webs have places for the concealment of the owners? Do all spiders seem to construct definite webs for the capture of insects? How do spiders entangle insects in their webs? Do different kinds use different methods? What parts of insects are eaten?

By destroying webs that are occupied by spiders that are in convenient places for observation, the construction of new webs may be observed. Notice how the framework is laid and then how the threads are attached to the framework. Are any of the legs used in handling the thread? Are spiders equally active at all times of the day?

Spiders' webs may frequently be seen floating in the air, especially in the late summer or autumn. By watching spiders that are on fences and bushes the formation of these threads may be observed. Watch such a spider and see if you can determine the use to which the thread is put.

Capture a spider and watch it descend by a thread. Where is the thread formed? Does the spider hold to it with its legs? Keep taking the thread up so that the spider cannot reach the ground, and see if there is a limit to the amount that can be formed. When the spider starts to climb the thread see how this is done, and whether the thread is taken up as the animal climbs or is allowed to float free.

Find where spiders lay their eggs. Some carry them. If you can find a specimen with an egg-sac, see how it is carried and whether it will drop its eggs when frightened. Remove the egg-sac and see if the spider will accept it again. Open several egg-sacs and see if the eggs all appear to be in the same stage of development.

Study the movements of the animal and see how many of the appendages are used in locomotion. Are any of the appendages used sometimes for locomotion and sometimes for feeling?

Examine the external structure of Epeira.

1. Into what parts is the body divided? Do both parts bear appendages?
2. Look for eyes on the anterior end of the body. How many are there? Do they seem to be simple or compound? Determine whether a specimen can see.
3. The following appendages should be found:
(a) The chelicerce or mandibles. Notice their structure and see that each ends in a sharp claw. The poison-gland discharges at the tip of this claw.
(b) The pedipalpi or palpi. How many segments have they? Examine their tips for claws. What are they apparently used for?
(c) Four pairs of legs. Are they all alike? Count the segments and examine their tips for claws.
(d) On the abdomen, three pairs of spinnerets. Notice their positions and see if they are segmented. Understand their function and whether they are all used at the same time. They are probably true abdominal appendages.
4. On the lower surface of the abdomen, near its anterior end, are two slits, the openings into the lung-sacs or lung-books. They are respiratory in function.
5. Just in front of the spinnerets is a minute median pore, the spiracle, that is often very hard to find. It is the external opening of a series of abdominal tracheæ.

Make a drawing of a ventral view.

## PHOXICHILIDIUM.

The exact affinities of the pygnogonids to other forms is not known, but they have certain characters that have suggested a possible relationship to the Arachnoidea. They are frequently found in considerable abundance on the material that is attached to piles. Notice their movements and see how they cling to the material on which they are moving.

1. The body is very slender and is composed of a number of free segments that form the head and thorax and a small, vestigial abdomen. How many free segments are there? At the anterior end is a rather prominent proboscis, with the mouth at its end.
2. The following appendages will be found:
(a) The chelicerce. What is their structure? Are they armed with pincers?
(b) Four pairs of long walking legs. How many segments have they? The viscera extends into the bases of these appendages.
(c) The male is provided with a pair of ventral appendages called the ovigerous legs, by means of which the eggs are collected as they are laid by the female. These appendages are not present in the female.

Make a drawing of the under side of a specimen.

## MYRIAPODA.

## LITHOBIUS. (Centipede, Earwig.)

These animals may frequently be found under stones, logs or boards, or about rubbish or manure heaps. They live largely on insects, larvæ, and small worms, and are very active.

1. Notice the shape of the body and count the number of segments. Is there a distinct head? Are the segments very movable?
2. How many appendages does each segment possess? Are all of the segments provided with appendages? Allow the ani-
mal to run and see how the legs are used. Do those of a side all move in the same direction at the same time? Are all of the legs alike? Notice the pair of appendages just behind the head and see how they differ from the others. These appendages are organs of prehension that are used in grasping the prey. They are provided with poison glands that open on their inner sides near their free ends.
3. Examine the head and find the eyes, antennor, and mouth parts. The latter consist of a labrum, a pair of mandibles, and two pairs of maxilloc, the last pair of which are united to form a labium.
4. Understand how the animal breathes. The stigmata are situated near the bases of the legs, but are hard to see except in favorable specimens.

Make a drawing of the animal.

## JULUS. (Thousand-legs.)

These animals are frequently very abundant under the dead bark of logs or stumps, in decaying wood, and in decaying heaps of grass. In the autumn they frequently congregate under boards and in corners. They feed on decaying vegetable matter.

1. Disturb a specimen and see how it rolls up. Can this be protective? See if there is any odor when it is disturbed. What purpose can such an odor serve?
2. What is the shape of the body? Is it hard or soft? How many segments are there?
3. How many appendages are borne on a segment? Do all of the segments bear appendages? Does the animal move rapidly? Why does it not need to move as rapidly as the preceding form? Do the first pair of appendages behind the head show modifications for prehension? Does this animal need such an organ?
4. Compare the organs of the head with those of the preceding form.

Make a drawing of the under side of one segment.

## INSECTA:

## ACRIDIUM. (Grasshopper.)

Study grasshoppers as they occur in nature and determine as far as possible the following points:

1. Do they see or hear? Are they equally sensitive to touch on all parts of the body? Why should the animal be well provided with sense organs?
2. What is their food? Are all plants eaten or are some avoided? See how the mouth parts are used in feeding.
3. What are the important enemies of grasshoppers? How do they escape their enemies? Do they hide? Are they protectively colored? How does jumping serve them better than crawling? How many times its length can a grasshopper jump? Why are wings needed?
4. During late summer and autumn you may find individuals depositing eggs. See if you can determine how the end of the body is worked into the ground.

For study it is desirable to use a rather large, freshly killed or alcoholic specimen.

The body is divided into three well-marked regions.

1. The Head.-Is it movable? Does it need to be as movable as your own head? It bears several organs.
(a) The compound eyes. Examine one with a lens or remove its outer covering and examine it with a compound microscope. You should understand the structure of the whole eye and how it gives a single visual image.
(b) The ocelli, three in number, one near the middle of the front part of the head and the others placed near the bases of the antennæ.
(c) The antennce. Why are they so flexible? Examine one with a microscope and notice the spines. What are these for?
(d) Mouth parts. These should be studied later.
2. The Thorax.-Why should it be large and comparatively firm? This portion is more or less distinctly divided into three parts, each of which carries a pair of legs.
(a) Compare the three legs of one side. Do they have the same number of segments? Do all of the joints of the leg move in the same plane? The five divisions of a leg are, beginning with the basal end: coxa, trochanter (immovably joined to the coxa in the leaping legs), femur, tibia, and tarsus, which is composed of four movable pieces. Why do the femurs of the leaping legs differ from the femurs of the other legs? Determine how the foot is arranged to hold to objects. Have you noticed a grasshopper settle its feet preparatory to jumping? Examine the joint between the femur and tibia.
(b) Examine the wings and notice their size, shape, places of attachment, and general character. Do they apparently have different functions to perform? Notice how the posterior wings are folded so they may be covered by the anterior. Does this seem to greatly reduce their strength ${ }^{1}$
3. The Abdomen.-Count the number of segments. Each one is covered dorsally by a tergum and ventrally by a sternum. Why should the abdomen be more movable than the other portions? The posterior ends of the abdomens of male and female differ. This portion of the female is modified to form the ovipositor, which consists of two large pairs of plates that inclose a smaller pair of plates. It is between these plates that the oviduct opens. Why do the larger plates possess hard tips? Along the sides of the abdomen notice the stomata, the external openings of the respiratory system. Do you find stomata on other parts of the body?

Draw an enlarged side view of a grasshopper, placing the appendages in their proper positions.

Mouth Parts.-It has already been noticed that the mouth parts serve to cut off pieces of leaves, which are then passed directly into the alimentary canal. For such a purpose there should be holding as well as cutting parts.

1. Pass a needle under the labrum, which forms the upper lip, and notice that it is hinged and that the end is lobed. It is

[^17]not supposed to be homologous with usual arthropod appendages. With fine scissors remove it and place it in a watch-glass containing water.
2. Immediately behind the labrum is a pair of hard, darkcolored organs, the mandibles, that are used in cutting the food. Their position should be carefully noted, but it will be better to leave them in position until the other mouth appendages have been removed.
3. Situated by the side of the mouth and just behind the mandibles are the maxillce. With a needle push one to one side and notice that it consists of a somewhat flattened portion with a jointed maxillary palp at one side. Carefully determine the positions of the maxillæ with relation to other parts. What possible uses are served by the two parts? Remove them with scissors and place them in the watch-glass with the labrum, in approximately their relative positions and study carefully.
4. Pass a needle behind the remaining appendage, the labium, and see that it is hinged and forms the lower lip. Remove it with scissors and place it in position in the watch-glass. You will find that it bears a pair of labial palpi, and that there is a deep cleft along the middle line. These are indications that the appendage is the result of the fusion of a pair of appendages.
5. Remove the mandibles and examine their cutting margins. Place them in position in the watch-glass.

Make a drawing showing the structure of each of these appendages. Arrange your figures as nearly as possible in the relative positions of the parts. ${ }^{1}$

Internal Structure.-Remove the wings, and before opening the body notice the rather large, somewhat transparent tympanum on each side of the first abdominal segment, very near the base of the leaping leg. The structure of the auditory organ may be easily studied by staining, clearing, and mounting in balsam. (See Packard's "Text-Book of Entomology" or

[^18]Brooks's "Hand-book of Invertebrate Zoölogy.") Remove the dorsal portion of the wall of the abdomen and thorax, and notice:

1. The heart, which will be found attached to the portion of the wall of the abdomen that has been removed, by means of numerous radiating muscle fibers. You probably will not be able to determine the structure of the heart in the dissection. Read this up, and determine what the radiating muscle fibers are for.
2. The space between the muscles and the viscera is filled more or less completely by the fat-body and the trachere. With a lens notice how the tracheæ connect with the spiracles and how they branch. Remove a portion of the tissue in which you can see tracheæ, mount it in water under a cover, and examine it microscopically. Each tracheal tube is marked by striations wound around it. Do you know what causes this appearance and what the arrangement is for? Do you understand how the tracheal system is arranged? Why is it extended all over the body and how is the air made to go in and out?
3. Near the dorsal surface of the posterior part of the abdomen, surrounded by the tissues already mentioned, are the gonads. These differ in size and shape according to the sex. In the male the vasa deferentia may be seen leaving the lobulated testes. In the female the oviducts pass around the sides of the intestine. They may be followed later.
4. Loosen the anterior ends of the gonads and turn them posteriorly to expose the hinder part of the alimentary canal. ${ }^{1}$
(a) The esophagus, which bends backward from the mouth, gradually enlarges as it enters the thorax.
(b) The crop, which is not sharply separated from the esophagus, gradually narrows posteriorly.
(c) Following the constriction posterior to the crop is the elongated stomach, frequently called the ventriculus. Surrounding the anterior end of this portion are a series of rather large

[^19]diverticula, the gastric cacca, that extend both anteriorly and posteriorly from the points where they open into the stomach.
(d) Some distance behind the posterior ends of the hepatic cæca, quite concealed by the mass of small uriniferous tubes, is a slight constriction and hardening of the alimentary canal that marks the division between the stomach and intestine. It is at this point that the uriniferous tubes join the alimentary canal.
(e) Behind the intestine the alimentary canal becomes much smaller and is known as the hind intestine or colon.
(f) Behind the colon, forming the hinder portion of the alimentary canal, is the slightly enlarged rectum. The rectum cannot be seen until the ovary is removed, which should be deferred until the ducts have been seen.

Make a drawing showing the position of the parts of the alimentary canal in side view.

Cut the intestine and turn the alimentary canal posteriorly and anteriorly.
5. Notice the muscles:
(a) That move the abdominal segments.
(b) That move the legs (those that supply the wings have been destroyed).
(c) That move the jaws.

Do you understand now why the thorax needs to be comparatively large and firm?
6. The nervous system is directly comparable to that of the lobster, but the connectives between the ganglia will be found to be distinctly double and the ganglia to be somewhat differently arranged. ${ }^{1}$

The ventral chain will be found to consist of a pair of subesophageal, three pairs of thoracic, and five pairs of abdominal ganglia with the connectives between them. Which of these are largest? Why is this the case? Trace the nerves from them and see what organs they supply.

[^20]Trace the connectives forward from the sub-esophageal ganglia and see that they pass around the esophagus, thus forming the circum-esophageal connectives. Cut away the dorsal portion of the head and expose the cerebral ganglia.

Add the nervous system to the figure that shows the alimentary canal.
7. Trace the oviducts down around the sides of the body and notice that they unite with each other ventral to the nervous system, to form the vagina. This may be traced to its opening between the plates of the ovipositor. Dorsal to the vagina, opening to the exterior very near it, is a small sac, the spermatheca, which serves to store the spermatozoa received from the male until the eggs are laid.

The reproductive organs may also be added to your figure showing internal anatomy.

## APIS MELLIFICA. (Honey-Bee.)

The life of this form is so different from that of the grasshopper that, should time permit, a study of its complete anatomy would be profitable, but attention will here be confined to a few of the more general adaptations that fit it for its life.

Bees at work on flowers should be examined and the methods of getting honey and pollen noticed.

1. Catch by the wings a bee that has been gorging itself and bend the abdomen forward with your thumb-nail until the bee disgorges. Notice where the fluid comes from and how much there is of it. When the abdomen is released watch the bee as it swallows the drop it has disgorged.
2. Notice where the pollen is carried, and see if you can determine how it is attached. Examine bees working on different flowers, or watch them as they enter their hives, and see if the pollen is always of the same color. Do you understand what the pollen is and what the bees use it for?
3. You may find bees gathering pitch from buds, knots, boards, or freshly varnished furniture, and fastening it on their legs. Do you know what this is used for?
4. Watch the entrance of a bee-hive and see if those going in are ever challenged. Perhaps you may see the method of defense. If so, you will notice that the stranger simply tries to get away. You may also see how dead bees and foreign materials are removed.
5. It is desirable to see something of the activities in the hive. This can be most satisfactorily done with an observatory hive, by means of which comb-building, honey-storing, egg-laying, brood-rearing, etc., can be very satisfactorily-studied.

Directions for the study of the mouth parts and the sting are all that seem necessary, but the wings should be examined microscopically to see how those of a side are joined together, and a hind-leg should be examined to see how the hairs on the tibia form a pollen basket.

Mouth Parts. -1 . With a lens notice that there is a pair of hard jaws, the mandibles, situated on the sides of the head at the base of the tongue. These mandibles are directly homologous with the mandibles of the grasshopper. Between the bases of the mandibles is a labrum, and extending from beneath the end of the labrum is a small epipharynx.
2. With scissors remove the tongue, which is normally carried against the lower surface of the thorax, and transfer it to a watch-glass. It may now be dehydrated, passed into oil of cloves, placed in position on a slide, and mounted in balsam, when it can be studied best, or it may be immediately spread under a cover or between slides in glycerin.
3. The central portion is the hairy, segmented labium (the hypopharynx of some authors), bearing at its end a little pad called the spoon. The labium is folded lengthwise so as to form a pair of fine ducts which run from tip to base. The arrangement is such that the bee may, through blood-pressure, unfold the labium. This probably is an adaptation for cleaning it. Attached to a median rod, the mentum, which forms the base of the labium, is a pair of flattened appendages, the labial palps, that are hinged so that they may be drawn together to inclose the labium and thus form a rather large tube, which is made
more complete by means of the remaining pair of flattened appendages, the maxillce. On the outer margin of each maxilla is a small protuberance, the maxillary palp. When sipping from an abundance of liquid the extemporized tube formed by the labial palps and maxillæ around the labium is used, the liquid being drawn in by means of the sucking stomach. When the liquid is in very small quantities it is apparently lapped up by the spoon and transferred through the labium. ${ }^{1}$

A figure of the mouth parts is desirable.
Sting.-The sting is to be regarded as a modified ovipositor that is no longer concerned in depositing eggs, but has become a weapon of offense and defense. It is accordingly present only in the female. The queen never uses her sting except in combat with other queens.

Remove the dorsal integument of the abdomen of either a fresh or preserved specimen, and find the dark brown shaft of the sting, near the posterior end. Grasp the shaft with a pair of fine forceps and forcibly remove it. A considerable mass of tissue will be removed adhering to the base of the shaft, but this consists for the most part of accessory organs that must be understood. Spread the sting upon a slide, and either dehydrate and mount in balsam, or mount in glycerin. The balsam mount will prove more satisfactory, but the cover must be clamped down until the balsam hardens.

1. The shaft consists of three parts:
(a) A heavy support, called the awl or sheath, pointed at its extremity and sending a pair of arms or arches from its base, which normally bend ventrally, but are here forced to the sides. At its extremity each of these arches enlarges to form a rather large flattened plate, the sheath plate, to which strong muscles are attached.
(b) A pair of lancets which are fastened to the dorsal surface of the sheath and the sheath arches by tongue and groove joints (each tongue is enlarged along its inner margin so that it is held

[^21]firmly in the groove). Each lancet is pointed at its free extremity, and its sides near the point are set with barbs that point toward the base of the sting. The arch of each lancet is continued past the end of the corresponding sheath arch, and is there articulated to one corner of a somewhat triangular plate. The remaining corners of each are articulated respectively to the large sheath plate and to another plate, the oval plate. Determine the attachment of the muscles to the plates and find what movements of the lancet the contraction of the different sets of muscles would cause. You must understand that the lancets are elastic and bend quite easily.

The large muscles attached to the sheath plates were attached to the wall of the abdomen and function to give the thrust that sets the sting. After the sting is drawn from the body of the bee, the muscles attached to the plates continue active, and the sting works deeper and deeper in. Understand why it works in instead of out.
2. Lying near the base of the shaft is a large poison sac or reservoir, which is very muscular. It receives its poison from the poison gland, a long and narrow coiled tube that is bifurcated near its free end. It discharges the poison by means of the contraction of the muscles of its walls through a rather large, short duct into the space inclosed by the sheath and the two barbs. Each barb bears a prominence that serves as an injector, which moves backward and forward with the barb to which it is attached, in an enlargement of the basal portion of the sheath. It may be seen in the preparation. In this way poison is forced into the wound. Poison may also be admitted to the cavities of the lancets, which are hollow, and escape through minute pores near the barbs.
3. Lying near the base of the shaft of the sting, sometimes covered by the poison sac, may nearly always be found the last pair of abdominal ganglia, from which nerves may be traced to the muscles that are attached to the plates.

Understand the whole mechanism, how it is operated and its use.
4. Catch a living bee by the wings and press the end of the abdomen against a piece of soft leather, such as a leather-covered book. Pull the bee away and with a lens watch the movements of the sting, which will remain in the leather. Observe the spasmodic contractions of the poison sac. See how long and how energetically the movements are continued and how deep the sting is worked in. This should remind you that a sting should be removed immediately, and that it should not be pulled out, as grasping the poison sac will aid in injecting the poison, but scraped off with a finger-nail or some other instrument.

A drawing showing the mechanism of the sting is desirable.

## CHORDATA.

With notochord, dorsal nerve cord, and branchial clefts during some period of existence.

Sub-phylum 1. Adelochorda.
Body somewhat worm-like and divided into proboscis, collar, and trunk. Notochordal development slight.
Class Adelochorda.
Characters as above. (Balanoglossus.)
Sub-phylum 2. Urochorda.
Adult body inclosed in a tunic. Larvæ usually motile. Notochord confined to the tail region.
Class Urochorda.
Characters as above.
Order 1. Larvacea.
Swim throughout life by means of a tail. The pharynx has a single pair of branchial slits. (Appendicularia.)
Order 2. Thaliacea.
Swim by forming currents of water. The pharynx has two or many branchial slits. (Salpa, Dololium.)
Order 3. Ascidiacea.
Mostly fixed. Pharynx large, provided with many branchial slits. (Molgula, Perophora, Botryllus, Amarœcium.)
Sub-phylum 3. Vertebrata.
Notochord or vertebral column present in the adult practically throughout the length of the body. Central nervous system forms a dorsal nerve tube.
Division 1. Acrania.
Without true skull or highly complex brain.
Class 1. Acrania.
Characters as above. (Amphioxus.)
Division 2. Craniata.
With true skull and highly developed brain.
(Six Classes and as many as fifty-six Orders may be recognized under this Division.)

## UROCHORDA.

## MOLGULA MANHATTENSIS.

Specimens of this simple ascidian may be found attached to old piles, associated with many other forms. In some localities they may be so abundant as to practically incrust the piles, and crowd each other out of shape. Examine such a mass and see how different sized individuals are associated. Pull them apart and see if there is any tissue connection between them that would indicate a definite relation between neighbors. Do you understand how the individuals get started in the places where they are attached? With a glass-bottomed pail you can see the expanded individuals on the piles; but they can be more satisfactorily studied in small dishes of sea-water.

1. Observe the contraction and closure of the two siphons when the animal is irritated.
2. Add a little powdered carmine to the water to determine which is the incurrent or oral and which the excurrent or atrial siphon.
3. Ascertain the number of lobes at the extremity of each siphon. Are pigment-spots present on the siphonal lobes?

Certain organs are distinguishable through the tough tunic which incloses the body. The endostyle in the mid-ventral line of the pharynx or branchial basket will serve as a guide in orienting the animal. Determine dorsal, ventral, anterior, right and left aspects.

Make a drawing of an expanded animal.
4. The tunic or test can be removed by cutting through it with scissors, taking care not to injure the mantle or body-wall. Enlarge the opening made in the tunic and strip it from the body. Where is the tunic most firmly attached? Examine a small piece of the tunic microscopically. Are blood-vessels visible in it? Does it contain any cells?
5. For further study use both fresh and preserved material from which the tunic has been removed. Identify as many organs as possible through the mantle. In a living specimen
note the beating of the heart (the heart is on the right side) and the frequent reversal of the direction of the pulsations. The endostyle, longitudinal pharyngeal folds, intestine, gonads, gonoducts, renal organ, and subneural gland are also visible through the mantle.
6. Note the muscle bands of the mantle which serve to contract the body and especially the siphons. Where are the muscles best developed? Is there any definite arrangement of the muscle-bands?
7. Fix a large specimen by pins through the siphons, and with scissors and fine forceps remove a large section of the mantle from the left side, injuring the underlying pharynx as little as possible. The large space between the pharynx and the mantle, laterally and dorsally, is the atrium, or peribranchial chamber, which is formed as an ectodermal involution. Into this atrial cavity open the intestine and the gonoducts, and also the numerous stigmata of the pharynx.

Alimentary Canal.-Cut out a piece of the wall of the pharynx from a very fresh specimen and examine in sea-water with a microscope.

1. Note the meshwork of blood-vessels, and the openings or stigmata, lined with actively moving cilia. Of what use are these cilia?
2. On each side of the pharynx six longitudinal pharyngeal folds will be seen.
3. The endostyle is a ciliated groove along the mid-ventral wall. Anteriorly it is continuous with the peri-pharyngeal ciliated bands, which encircle the oral end of the pharynx. From the point where they unite dorsally the dorsal lamina extends backward along the mid-dorsal line of the pharynx. At its posterior end will be seen the small opening into the esophagus.

Do you understand how the animal captures its food and how the endostyle, peri-pharyngeal bands, and dorsal lamina are used?
4. In front of the anterior end of the dorsal lamina note the small, volute-shaped dorsal tubercle. This is the extremity of
the hypophysis, a tube connecting the subneural gland with the oral cavity.
5. A ring of oral tentacles will be seen in the mouth, anterior to the peri-pharyngeal bands. Of what use are sensory tentacles in the mouth? How many tentacles are there?
6. The very short esophagus opens into the stomach, which will be recognized by the brown digestive glands that cover it. From the stomach the intestine forms a loop on the left side, and is easily traced to the anus, which opens dorsal to the pharynx in the atrial chamber. A longitudinal fold, the typhlosole, extends throughout the intestine. What is the use of such a fold?

Reproductive System.-On each side of the body, adherent to the inside of the mantle, is an elongate hermaphrodite gland. Each gland consists of a lighter part, the testis, and a darker part, the ovary. The gonoducts open on the outer wall of the atrial cavity near the base of the atrial siphon. Each consists of two ducts, oviduct and vas deferens. Microscopic examination of the oviduct may show the presence of eggs.

Excretory System.-The renal organ is a conspicuous, elongated sac on the right side. It contains a brownish fluid and usually some solid matter. It does not possess a duct.

Nervous System.-The cerebral ganglion, which in Molgula is almost completely surrounded by the subneural gland, lies close to the mantle, between the two siphons, and is thus dorsal to the mouth. Nerves can be seen passing from the ganglion to the two siphons. The hypophysis, a tube leading from the subneural gland, opens as the dorsal tubercle above mentioned.

Circulatory System.-1. The heart, which lies on the right side between the hermaphrodite gland and the renal organ, is inclosed within a pericardium which is a portion of the colom. It should be studied in a living specimen, with the aid of a handlens.
2. If a very small Molgula (one-eighth of an inch in length) is studied alive in a watch-glass with the microscope, the course
of the circulation, and the frequent reversal of its direction, can be observed.
3. From the dorsal end of the heart a cardio-visceral vessel runs to the visceral mass, where it divides into smaller vessels. These, reuniting, form the viscero-branchial vessel which extends along the dorsal surface of the pharynx above the dorsal lamina. Numerous small branchial vessels in the pharyngeal wall connect this vessel with the branchio-cardiac, which lies ventral to the endostyle and unites with the ventral end of the heart. The frequent reversal of the current can be readily seen both in the heart and in the vessels.

The relation of the parts will be more clearly understood if a second large specimen is dissected as follows: With scissors cut off the atrial siphon, thus exposing the atrium; then similarly remove by a single cut the oral siphon, together with the anterior end of the pharynx (the piece thus cut off should contain the ganglion, dorsal tubercle, peri-pharyngeal bands, oral tentacles, anterior portion of the endostyle, dorsal lamina, etc.).

Make drawings that will show the structure.

## PEROPHORA.

This colonial simple ascidian occurs on piles and other submerged materials, and is commonly attached by branching stolons to seaweeds, simple tunicates, or other sessile animals. Material should be quite fresh for satisfactory study, and should be carefully handled to avoid crushing. Study in a watch-glass of sea-water (or support the cover-glass), with a low power of the microscope.

1. Notice that the individuals are essentially very much like miniature Molgulas. Identify as many of the organs that were seen in Molgula as possible, noting the differences.
2. The form illustrates the type (Clavelinidæ) in which a colony is formed by budding from a stolon, but in which the individuals retain their identity to a great degree and have separate tunics.
3. Study the stolon with its flattened epicardiac tube. This tube is derived from the branchial sac and is accordingly endodermic.
4. Study buds of various sizes and see how the inner vesicles arise from the epicardiac tube.
5. Try to make out the entire course of the circulation of the blood. Notice especially the heart, branchial vessels, vessels of the mantle, and the circulation of the stolon. Watch the pulsations of the heart and see the reversal of the blood-current. Is the heart-beat synchronous in different individuals? What part of the blood is colored?
6. Study the action of the cilia in the gill clefts.

Drawings of a colony and of an individual are desirable.

## BOTRYLLUS.

The small, radially arranged colonies of this composite ascidian are common on eel-grass, from which they may be separated by means of a knife, and studied alive in a watch-glass with a low power of the microscope. The cleaner and more transparent colonies should be selected.

1. Note the character which makes the form a "composite" ascidian-the common tunic or test. Find the mouths and the common cloacal cavity. Would it be correct to say that a common atrium is present?
2. Find the annular blood-vessel and its numerous ampullec. Do you observe any striking facts regarding the circulation? What function have the ampullæ?
3. With your knowledge of Molgula as a guide, identify as many of the organs as possible. (This is sometimes difficult because of pigment.)
4. Very young colonies, with only the first one or two generations of buds, may also be found on eel-grass, appearing as transparent hemispherical lumps about a millimeter in diameter. These should be fixed and stained on the eel-grass, and later mounted (either still attached or removed) in balsam. These will show very clearly the formation of buds of the "parietal"
or "peribranchial" type. (In this type the outer vesicle arises from the integument, and the inner vesicle from the parietal wall of the atrial cavity.) The inner vesicle may be seen partly constricted into three divisions-the pharynx and the two atrial sacs. From which "germ layer" then are these parts in the bud derived?
5. Look for the tailed larvæ or "tadpoles" near the surface and on the side turned toward the light, in a dish in which Botryllus has been kept for an hour or two. Is this positive phototrophism advantageous? Examine a larva under a microscope.

Drawings of the adult, the young colony, and the larva are desirable.

## AMAROECIUM. (Sea-Pork.)

Different species of this composite ascidian live at different depths and show minor structural differences, especially in the tests. Colonies may be found abundantly on piles and they are frequently brought up with a dredge.

1. Compare the grouping of the individuals in the colony with Botryllus. Is there any regularity in the number of a group connected with a common cloacal cavity?
2. With a sharp knife, cut a section vertical to the surface of the mass, and two or three millimeters thick, and study it with a low power of the microscope. Other pieces should be squeezed in a finger-bowl half full of sea-water, the expressed material (adult animals, fragments, embryos, etc.) allowed to settle, and then rinsed with clean sea-water. A few entire adults may be picked out with a pipet.

In the adult animal you may find:
(a) Oral and atrial openings.
(b) Pharynx, with the peri-pharyngeal bands and endostyle, esophagus, the orange-brown corrugated stomach, and intestine.
(c) The cerebral ganglia.
(d) The long post-abdomen, with its hollow epicardium connected with the pharynx. (The post-abdomen is really a stolon. Recall Perophora.) If complete, the red-pigmented tip will be seen.
(e) The slowly pulsating U-shaped heart, situated very near the tip of the post-abdomen.
3. In the atrium, which serves as a brood-pouch, embryos in all stages may be found. How do the eggs compare in size with those of Molgula?
4. Look for buds formed by segmentation of the post-abdomen (stolon). The "inner vesicle" of these buds, which gives rise to the alimentary canal and atrial sacs, comes from the endodermic epicardium, as in Perophora. Compare this with Botryllus.
5. If the material squeezed in the finger-bowl was quite fresh, living embryos in all stages of development can be found, and the tailed larvæ will probably be found hatching during the first hour or two. These swim rapidly, and usually swim away from the light. Does this correspond with Botryllus? Is this negative phototrophism adaptive?

The tailed larvæ may be picked up with a pipet while swimming (the dead ones on the bottom of the dish should not be used), dropped into fixing fluid, and finally stained and mounted in balsam. Some larvæ will be found attached to the dish by their adhesive organs. Notice where these organs are situated.

In larvæ that have been previously stained and mounted observe:
(a) The shape of the animal and its division into body and tail.
(b) The thick test, and the oral and atrial openings.
(c) The adhesive organs. How many are there?
(d) The notochord. How far does it extend?
(e) The tail muscles.
(f) The pharynx, with as yet few gill clefts, the endostyle, esophagus, stomach, intestine, and yolk-mass.
(g) The cerebral vesicle with the eye-spot and otolith.

If young individuals that have been attached but a short time, but have lost their tails, are stained and mounted, they will be found very instructive when compared with the larva.

The complete degeneration of the tail and the final rotation into the position of the adult can be traced in a series of individuals.

Drawings of an adult individual, of a larva, and of a young individual are desirable.

## SALPA CORDIFORMIS.

Examine a specimen in a bowl of water without dissecting. Use a hand-lens.

Sexual form (occurring in chains):

1. Note the transverse muscle bands. How many bands are there? Are they complete or interrupted? Do you know what they are for?
2. The oral aperture is dorsal and far forward. Are there any muscles for opening and closing it?
3. What is the form and position of the cloacal aperture? Is it provided with muscles?
4. Observe the processes of the tunic, one anterior, one midventral, and two posterior. These processes (except the dorsal posterior) serve to unite the individuals of the chain.
5. Does the animal show perfect bilateral symmetry?
6. Posterior to the mouth, the ganglion and the pigmented eye-spot may be found. Immediately anterior to these is the elongate hypophysis.
7. Note the endostyle in the floor of the pharynx, and the dorsal lamina between the pharynx and atrial cavity. From the anterior end of the dorsal lamina the peri-pharyngeal bands extend to the anterior end of the endostyle.
8. The pharynx communicates laterally with the atrium by means of two very large stigmata. These are probably homologous with the numerous stigmata of Molgula.
9. The "nucleus," the large mass in the posterior end of the body, contains the stomach and intestine.

The ova are fertilized by spermatozoa from individuals of another chain, since in the same chain the spermatozoa mature much later than the ova. The fertilized ova migrate to
a spot in the right wall of the atrium, where they develop into the solitary non-sexual Salpa.

In this species as many as three or four embryos may be seen attached by "placente" to the cloacal wall on the right side. The placental connection finally separates, and the embryo passes out through the cloacal aperture.

Make an enlarged drawing (a latero-dorsal view is best).

## ACRANIA.

## AMPHIOXUS LANCEOLATUS.

While living material is not easily provided for laboratory work, it should be understood that this form spends most of its time in the sand of the bottom, in which it burrows with great ease by movements of the body.

1. In an alcoholic specimen note the dorsal, ventral, and caudal regions of the median fin, metapleural folds, muscle plates or myotomes, buccal cavity fringed with cirri, atriopore, and anus.
2. Using a specimen that has been macerated in 20 percent nitric acid, remove the skin and myotomes from the right side very carefully, by means of needles, exposing the notochord, nerve cord, gonads, and the entire alimentary canal (pharynx, intestine, and digestive diverticulum or "liver," which lies along the right side of the pharynx).
3. Examine microscopically and notice:
(a) The nerve cord, cerebral vesicle, cerebral nerves, eye-spot, and pigment cells. Note also the alternate metamerism of the spinal nerves.
(b) The buccal skeleton.
(c) A large piece of the pharyngeal wall.
4. Examine an Amphionus one centimeter in length, stained and mounted.

Identify as many as possible of the structures mentioned above, and in addition note: the olfactory pit, oral velum with velar tentacles, and "taste organ" in the buccal cavity.

A drawing showing the general structure is desirable.
5. Make thick free-hand sections of various regions and study with a low power in a watch-glass, to supplement the study of stained sections.
6. Prepared sections should be studied that show the following five regions: (a) buccal cavity; (b) anterior part of pharynx; (c) posterior part of pharynx with gonads and liver; (d) atriopore; (e) anus.

The five sections should be studied with a low power and drawn. In (b) (anterior part of pharynx), note especially the limits of the colom and atrium, the lymph-spaces in the metapleural folds, the two dorsal aorte, the ventral aorta, the epibranchial groove, the endostyle, the sub-endostylar colom, and the two kinds of gill-bars, primary and tongue-bars.

With a high power study the nerve cord (best shown in region $a$ ) and the gill-bars and endostyle (best shown in region b).

Drawings of these regions are desirable.

## NOTES FOR GUIDANCE IN MAKING PERMANENT PREPARATIONS.

Only very simple directions are here given, such as will serve to aid students who have had no experience in preparing objects for microscopic examination to make preparations when this is desirable for proper laboratory study. Those who desire to prepare material for serial sections, or who wish to make whole mounts of delicate material, are referred to Lee's "Microtomist's Vade Mecum."

The steps taken in preparing total mounts include:

1. Fixing, or killing.
2. Washing.
3. Dehydrating and staining.
4. Clearing.
5. Mounting.

Fixing.-This is necessary to keep the cells and tissues as nearly as possible in their natural position, shape, and structure, and in order that the protoplasm composing them may be kept in condition to stain satisfactorily.

In selecting a fixing agent remember:

1. If the material is highly irritable and contractile, it will have to be killed practically instantly with hot solutions, or be previously narcotized.
2. If there is much lime, an agent that contains much acid should not be used, as the lime will be dissolved and the bubbles of gas are likely to tear or distort tissues.
3. Where rapid fixation is desirable, as in expanded hydroids and the like, sublimate-acetic (hot) is preferable. Where the tissue, or the animal, is not specially muscular, or liable to contraction, any of the fluids can be used. The time objects should be left in the killing solution varies, approximately, directly as
their size. Three minutes will suffice for killing hydroids in sublimate-acetic.

Washing.-All objects must be thoroughly washed, after using most killing agents. This may be done with repeated changes of fresh water or with alcohol, beginning with a low grade and gradually working up to 70 percent. With most small objects alcohol is preferable, but if the object is large this is too expensive.

In case a fixing agent is used which is an alcoholic solution, wash out in the same grade of alcohol used in making the fixing agent.

Dehydrating and Staining.-From water, all objects should be placed successively in 35 percent, 50 percent, and 70 percent alcohol, five to fifteen minutes in each for small objects. In subsequent changes from one grade to another allow about the same time. All tissues killed in a corrosive sublimate mixture should now be treated with a weak solution of iodin, to dissolve the corrosive sublimate that still remains, and thus prevent the later formation of crystals of that substance. Such crystals would not appear immediately, but ever increasingly, as the preparation is kept. Put a few drops of iodin into the 70 percent alcohol containing the object, leave a few minutes, and, if the yellow color caused by the iodin has disappeared, turn off the alcohol and use more 70 percent alcohol with iodin, as before. 'The bleaching indicates that some corrosive sublimate remains. Repeat until the yellow color does not fade. Then wash in clear 70 percent alcohol. At this point either staining, or preparation for so doing, begins.

In case the stain you wish to use is a 70 percent alcoholic solution, it may be used immediately. Otherwise, the object must be run through the grades of alcohol, up or down as the case may be, to that medium in which the stain to be used is dissolved. If an aqueous stain such as alum-carmine is to be used, pass through 50 percent and 35 percent alcohol to water. If a 95 percent alcoholic stain is to be used, pass through 80 percent and 95 percent alcohol.

The time an object should be treated with stain varies with the stain and the size of the object. Alum-carmine should be used from six to twenty hours, according to circumstances. Borax-carmine should be used from five minutes to half an hour. Aceto-carmine, used for killing and staining, acts very rapidly. Delafield's hematoxylin (a dark wine-colored solution in water) requires ten minutes to half an hour. In all these cases, examination of the objects themselves is the only means of deciding when staining is sufficient. It is usually best to slightly overstain and then to bleach out, as certain parts of the protoplasmic structure will retain the stain better than others and thus better differentiation will be secured. After staining, bring the tissues gradually into 70 percent alcohol, and then treat with acidulated alcohol to remove excess of stain. After this, every trace of the acid must be removed by washing in clean alcohol, or the tissues will continue to bleach after they are mounted. The specimen is now ready for final dehydration. In damp climates, as at the seashore, your stronger alcohols must be kept closely covered all of the time or they will take water from the atmosphere and be unfit for the purpose. Run through 80 percent, 95 percent, and 100 percent alcohol, thus completing dehydration. Every trace of water must be removed and then kept out.

Clearing and Mounting.-From absolute alcohol, place objects in some clearing fluid (clove oil, cedar oil, or xylol) and leave till they have a clear, translucent appearance, after which place on a clean slide, with come canada balsam or dammar, and cover with a cover-glass.

If the object turns cloudy or milky when placed in the cleaning fluid, it is evidence that all of the water has not been removed, and it should be returned to absolute alcohol for complete dehydration. Tissues left in the clove oil or xylol for any great length of time will become hard and brittle. In case tissues in the process of preparation must necessarily be left untreated for several days, they should be left in a 70 percent or 80 percent alcoholic medium.

Sectioned Material.-In a few cases sectioned material may be distributed to the class. Be sure that the slide on which you intend mounting the sections is thoroughly clean. Remove any greasy substance with 95 percent alcohol. On a cleaned slide, smear a very little albumen with your finger-tip and remove all except the thinnest film. Now place the sections on the albumen over an area the size of the cover-glass to be used, and press them down flat with the tip of a clean, dry finger. ${ }^{1}$ Warm the slide over an alcohol lamp very carefully until the paraffin in which the sections are embedded is just melted. While the paraffin is still melted treat it with xylol (a jar containing xylol for this purpose is desirable). This will dissolve the paraffin and leave the sections alone adhering to the slide. When the paraffin is completely dissolved (this will take but a few seconds), drain off the xylol, apply a drop of balsam, and cover as in total mounts. The preparation is now ready for use, and is permanent, but must be handled carefully while fresh.

## Application of above directions in the case of a hydroid :

Hot corrosive, fifteen seconds.
Cold corrosive, five minutes.
Water or alcohol, four changes, three or four minutes each.
Thirty-five percent, 50 percent, and 70 percent alcohol, five minutes each.
Seventy percent alcohol plus iodin, as in directions above.
One-half of your material may now be placed in borax-carmine. Leave the material in this till objects have taken on a good color. (Ask an instructor about this.) When sufficiently stained, put into acidulated alcohol till the color assumes a brilliant appearance, but do not allow it to fade too far. Wash in 70 percent and

[^22]then run through 80 percent, 95 percent, and 100 percent alcohol, five minutes in each, thence into clove oil, or cedar oil, keeping all reagents carefully covered, and leave till the object is thoroughly penetrated. This latter process may take five to ten minutes.

If, on putting your objects into the clearing medium, the latter exhibits a milky-white appearance, the material is not sufficiently dehydrated, and must be returned to 100 percent alcohol.

After clearing is completed, put the object on a clean slide with a little balsam and cover.

The material not treated with borax-carmine may be run back through 50 percent and 35 percent alcohols to water, to which a few drops of hematoxylin have been added, or put from water into alum-carmine. The former stain, if dense, should not require over twenty to thirty minutes, but objects must be left in alum-carmine ten to twenty hours. When a good color is obtained, run the material through the grades of alcohol, from the lowest to the highest (five minutes in each), and mount as in the case of the borax-carmine objects.

Objects stained in alum-carmine will probably not overstain; but excess of hematoxylin should be extracted with acidulated alcohol when the 70 percent grade is reached, after which it is very essential that all of the acid be removed by repeated changes of 70 percent alcohol. Otherwise the objects will fade.

## GLOSSARY.

Abdomen. The posterior division of the body of an arthropod.
Aboral. The surface away from the mouth.
Aciculum. A supporting rod in a parapodium of an annelid.
Acinous. Saccular or granular.
Acontium. One of the filaments that are attached to the mesenteries of such forms as Metridium.
Adductor muscle. A closing or withdrawing muscle.
Adhesive organ. A sucker or sticky pad that will adhere.
Ad-radial canal. A canal in a medusa that lies between adjacent perand inter-radial canals.
Afferent. Coming toward, as a vessel that leads to an organ.
Alga. A simple plant.
Alimentary canal. Digestive tube.
Alternation of generation. Alternation of sexual and asexual generations in the life cycle of an organism.
Alveolus. A little sac or cavity; also one of the plates that bears the teeth in an echinoid.
Ambulacral area. The region bearing the tube feet of an echinoderm.
Ambulacral foot. A tube foot of an echinoderm.
Ambulacral groove. One of the depressions in which the tube feet of a starfish are placed.
Ambulacral plate. One of the plates of an ambulacral area.
Ambulacral pore. The opening through which a tube foot projects.
Ambulacral ridge. The elevation in the colom of a starfish arm, caused by the ambulacral plates.
Ambulacral sucker. The sucker at the end of a tube foot.
Amphiblastula. An embryonic stage of a sponge.
Ampulla. A reservoir connected with the tube foot of an echinoderm.
Anal plate. One of the plates in the periproct of an echinoid.
Analogous. Similar in function.
Anastomosis. The joining together, as of vessels and nerves.
Antenna. A sensory head appendage of an arthropod.
Antennule. A sensory head appendage of an arthropod, placed just anterior to the antenna when present.

Anterior. Front or head end.
Antero-posterior. Lengthwise of the body.
Anus. The posterior opening of the alimentary canal.
Aorta. In invertebrates used to designate the chief blood-vessel.
Apical system. A group of plates surrounding the periproct of an echinoid.
Apopyle. The opening of a radial canal of a sponge into the gastreal cavity or cloaca.
Arthrobranch. A gill of a crustacean borne by the articular membrane at the base of an appendage.
Asexual. Reproduction by other than sexual methods, as by budding or division.
Atriopore. External opening of the atrium of Amphioxus.
Auricle. A division of the heart.
Avicularium. A structure shaped like a bird's head, to be found on some of the Polyzoa.
Axial organ. A structure near the stone canal of echinoderms that is apparently connected with the genital organs.

Basipod. Second segment from the body of a crustacean limb.
Beak. Horny mouth parts; the point from which growth has proceeded in a clam shell.
Bilateral symmetry. Right and left sides alike.
Biramous. Composed of two branches, as a typical crustacean appendage.
Bivalve. Having two valves or pieces, as a clam shell.
Bivium. The two rays of a starfish that are nearest the madreporic plate. ${ }^{\circ}$
Blastostyle. The axial tissue of a reproductive polyp of certain Hydrozoa.
Body-cavity. Cœlom; the cavity between the alimentary canal and bodywall.
Body-wall. The outer wall of the body.
Brain. In invertebrates frequently applied to the cerebral ganglia.
Branchiæ. Gills; organs adapted for aquatic respiration.
Branchial heart. An accessory heart placed at the base of the gill, as in the squid.
Brood sac. A cavity or pouch in which developing embryos are carried.
Bud. An outgrowth of an animal that will become a new individual.
Byssal thread. One of the threads by which certain lamellibranchs attach themselves.

Cæcum. A sac-like appendage of the alimentary canal.
Calciferous glands. Esophageal glands of some annelids.
Carapace. Head and thoracic covering of some crustaceans.

Cardiac division of stomach. Anterior or first division.
Carpopod. Fifth segment from the body of the leg of a crustacean.
Cellulose. The material that forms the walls of plant cells.
Cephalont. Attached stage in the life-history of Gregarina.
Cephalothorax. Fused head and thorax of many crustaceans.
Cervical groove. A groove that marks the boundary between the head and thorax of some crustaceans.
Chela. Large claw of many crustaceans; also applied to pincer-like claws on other appendages.
Chelate. Bearing pincer-like claws.
Chelicera. One of the anterior pair of mouth appendages of Arachnoidea.
Chitin. The material that forms the outer covering of insects and other animals.
Chlorogog. Excretory cells on the intestine of certain annelids.
Chlorophyl. The green coloring-matter of plants.
Choanocyte. A cell provided with a "collar."
Chromatophore. A body in which chlorophyl is lodged.
Cilia. Small vibrating appendages of cells.
Cinclides. Minute openings in the body-wall of coelenterates.
Circular canal. Marginal canal of a medusa; also applied to the water canal that surrounds the mouth of an echinoderm.
Circumferential canal. Marginal canal of a medusa.
Cirrus. A soft tactile appendage.
Cleavage. The act of cell division.
Clitellum. The thickened glandular region on an earthworm that secretes the egg case.
Cloaca. A space that receives the discharge from the alimentary canal and kidneys, and frequently from other organs.
Cnidocil. The trigger of a nematocyst.
Cœlenteron. The internal space of a colenterate.
Cœlom. Body-cavity; the cavity between the alimentary canal and body-wall.
Cœnosarc. The fleshy continuation of a hydroid into the stalk.
Collar-cell. A cell provided with a collar; choanocyte.
Colon. Hinder part of the alimentary canal.
Columella. Axis around which the spire of a gastropod shell is wound.
Commensal. Organisms living together and usually partaking of the same food.
Commissure. A nerve connecting two ganglia of a pair.
Compound eye. Eye of an arthropod that is composed of many similar pieces, called omatidia.
Connecting canal. The canal that joins the tube foot to the radial canal of an echinoderm.

Connective. A nerve connecting two ganglia not of a pair.
Contractile vesicle. Contractile excretory organ of Protozoa.
Copulation. Sexual union.
Coxa. Basal segment of the leg of an insect.
Coxopod. Basal segment of a leg of a crustacean.
Crop. An enlargement of the alimentary canal used to store food.
Crystalline style. A transparent rod frequently found in the alimentary canal of lamellibranchs.
Ctenophoral row. A row of swimming plates on a ctenophore.
Cuticle. Outside protective covering.
Cyst. A sac or pouch.
Cystic duct. The duct that leads away from the gall-bladder.
Cysticercus. A stage in the development of many tapeworms.
Dactylopod. Last segment, seventh, of a crustacean leg.
Dactylozoöid. Elongated tentacle-like zoöid of a siphonophore.
Denticle. Small, tooth-like protuberance, as in the pharynx of some annelids.
Dermal branchiæ. Projections on the surface of the body that are used for respiration. See starfish.
Development. The series of changes that lead from the fertilized egg to the mature animal.
Digestive gland. Any gland that secretes a digestive fluid.
Dimorphism. Two distinct forms of individuals in the colony or species.
Diœcious. Sexes in two separate individuals.
Directive septa. Those placed opposite the syphonoglyphes of an actinozoan.
Disk. The central portion of a starfish.
Dissepiment. A transverse partition in an annelid.
Distal. Remote from the point of origin or attachment.
Diverticulum. An out-pocketing from a tube.
Dorsal. Back.
Dorsal lamina. A ciliated ridge on the dorsal side of the pharynx of an ascidian.
Dorso-ventrally. From the dorsal to the ventral position.
Ectoderm. The outer embryonic layer.
Ectoparasite. A parasite on the outside of the body.
Ectoplasm. Outer layer of Amœba and of other Protozoa.
Efferent. Going away, as a vessel that leads away from an organ.
Elytra. The modified fore-wings of a beetle.
Embryo. An immature animal.
Encyst. To inclose in a cyst.

Endoderm. The inner embryonic layer.
Endoparasite. A parasite inside of the body.
Endo-phragmal skeleton. Chitinous plates that cover the nerve-chain and ventral blood-sinus in the thorax of certain crustaceans.
Endoplasm. Inner portion of an Amoba and other Protozoa.
Endopod. The branch of a biramous appendage of an arthropod that is nearest the mid-line of the body.
Endoskeleton. An internal skeleton.
Endostyle. A ciliated groove in the ventral wall of the pharynx of an ascidian.
Ephyra. An embryonic stage of Discomedusx.
Epicardium. A hollow process from the pharynx of some ascidians. See Amarœcium.
Epipharynx. A projection from the roof of the mouth of some insects.
Epiphysis. A plate joined to the base of the alveolus in the mouth-parts of an echinoid.
Epipod. A membranous projection found on certain crustacean limbs, that extends into the gill chamber.
Episternum. A lateral piece next to the sternum of arthropods.
Epistome. A projection above the mouth. See Pectinatella.
Esophagus. The portion of the alimentary canal that leads back from the mouth or pharynx.
Euglenoid. Similar to Euglena, especially in movements.
Exopod. The branch of the biramous appendage of an arthropod that is away from the mid-line of the body.
Exoskeleton. An outer covering, as a shell.
Exumbrella. The convex side of a medusa.
Eye-spot. A pigment spot generally supposed to be associated with perception of light.

Femur. The third segment from the body of the leg of an insect.
Fission. A method of asexual reproduction by division.
Flagellum. An elongated vibratory projection of a cell.
Flame-cell. The terminal cell of one of the excretory tubes of the Plathelminthes.
Foot. A locomotor organ. See Venus.
Funiculus. A strand of connective tissue that connects the stomach with the body-wall in Polyzoa.
Funnel. The tube through which water is expelled from the mantle chamber by cephalopods.

Ganglion. A group of nerve cells.
Gastric filament. One of the filaments in the stomach of Scyphozoa.

Gastro-vascular. Digestive and circulatory in function, as the gastrovascular cavities of coelenterates.
Gastrozoöid. Feeding individuals of hydroids.
Genital atrium. A space receiving the genital ducts. . See Bdelloura.
Genital gland. A gonad.
Genital plate. One of the plates through which the gonads open in echinoderms.
Genital pore. The opening in the genital plate or other external opening of a gonoduct.
Gill. Aquatic respiratory organ.
Gizzard. A muscular triturating division of the alimentary canal.
Gonad. A generative tissue, a germ gland.
Gonangium. A reproductive individual of the Hydrozoa.
Gonotheca. The transparent covering of a gonangium.
Green gland. One of the excretory glands of certain crustaceans.
Gullet. Esophagus; the tube leading back from the mouth or pharynx.
Gut. Digestive tube.

Head. The anterior division of the body of higher animals
Hepatic cæcum. Digestive gland of a starfish.
Hermaphrodite. With male and female sexual organs.
Holophytic. The nutrition characteristic of plants.
Holozoic. The nutrition characteristic of animals.
Homologous. Of similar structure.
Host. The animal that harbors a parasite.
Hyaline. Transparent, glassy.
Hydranth. An individual of a hydroid colony.
Hydrocaulus. The stem of a hydroid colony.
Hydrorhiza. The projections by which hydroid colonies are attached.
Hydrotheca. The outer secreted covering or cup of a hydranth.
Hypobranchial gland. A gland near the gill of some gastropods. Some lamellibranchs have glands that bear the same name.
Hypodermis. A cellular layer that lies just beneath the cuticle of annelids, arthropods and some other animals.
Hypopharynx. A projection borne on the lower side of the pharynx of some insects.
Hypophysis. A ventral projection from the brain of Chordata.

Incurrent canal. A canal that admits water to a sponge.
Integument. Skin; outer covering.
Inter-ambulacral area. One of the areas of an echinoderm that lies between adjacent ambulacral areas.

Inter-filamentar junction. A connection between adjacent filaments in a lamellibranch gill.
Inter-lamellar junction. A connection between adjacent lamellæ in a lamellibranch gill.
Inter-radial canals. The canals of a medusa that lie midway between the per-radial canals.
Intestine. One of the divisions of the alimentary canal.
Introvert. A portion that will turn inward, as the anterior end of Phascolosoma.
Ischipod. The third segment of a typical crustacean leg.

Kidney. Frequently applied to the excretory organ of an invertebrate.
Labrum. The appendages that form the lower lip of insects and some other arthropods.
Lamella. One of the two sides that form a lamellibranch gill; a flat structure.
Lamelliform. Like a lamella; thin and flat.
Lamina. A thin plate or a scale.
Lancet. A sharp structure; a portion of the sting of a bee.
Larva. An embryo; a stage in the development of an animal.
Lateral. At or toward the side.
Ligament. The portion that unites the valves of a clam shell; an elastic connection.
Lithite. One of the concretions in a tentaculocyst of a medusa.
Liver. Frequently applied to the largest digestive gland of an invertebrate.
Lophophore. The disk that surrounds the mouth and bears the tentacles in the Molluscoida.
Lorica. The transparent covering of a rotifer.
Macronucleus. The larger of the two nuclei of certain Protozoa.
Madreporic plate. The perforated plate through which the water-vascular system of an echinoderm is put in communication with the sea-water.
Mandible. One of a pair of mouth appendages of an arthropod.
Mandibulate. Possessing mandibles.
Mantle. The outer fold of tissue of many animals; in many mollusks and tunicates it secretes a protective covering.
Manubrium. The projection at the end of which the mouth is situated in celenterates.
Marginal lappets. Small flaps of tissue near the sense organs of Discomedusw.
Mastax. A division of the alimentary canal of a rotifer.

Maxilla. One of the mouth appendages of arthropods.
Medusa. Jelly-fish; the sexual stage of certain hydroids.
Membranells. Structures formed of fused cilia found in some Ciliata.
Meropod. The fourth segment from the body of a crustacean leg.
Mesenteric filament. The modified free edge of a mesentery of Actinozoa.
Mesentery. A membrane that supports the intestine; one of the partitions of Actinozoa.
Mesoglea. The jelly-like substance that separates the ectoderm and endoderm of a coelenterate.
Metamere. One of the serial body-segments of an animal, as in annelids.
Metamorphosis. A change, especially in form, of an animal.
Metapleural fold. One of a pair of folds on the sides of Amphioxus.
Micronucleus. The smaller of the two nuclei of some Protozoa.
Moniliform. Resembling a string of beads.
Monœcious. Sexes united in one individual.
Mouth. The opening through which food is taken.
Myoneme. A contractile fiber. See Vorticella.
Nacre. The inner layer of a mollusk shell.
Nematocyst. A weapon of a coelenterate; nettle cell.
Nephridiopore. The external opening of a nephridium.
Nephrostome. The inner opening of a nephridium.
Nerve commissure. A nerve connecting two ganglia of a pair.
Nerve connective. A nerve connecting two ganglia not of a pair.
Nettle cell. Nematocyst; a weapon of a coelenterate.
Neuropodium. The ventral division of a parapodium of an annelid.
Nidamental gland. An accessory reproductive gland possessed by some females, especially gastropods and cephalopods.
Notochord. A supporting structure characteristic of Chordata.
Notopodium. The dorsal division of a parapodium of an annelid.
Nuchal groove. A groove in the neck.
Nucleus. An organ of a cell.
Odontophore. A special structure in the mouth of most mollusca except lamellibranchs. The name is applied to the whole structure, cartilage, radula and muscles. (It is used by some authors as the equivalent of radula.)
Ocellus. A simple eye of an arthropod.
Ocular plate. A plate at the end of an ambulacral area of an echinoderm.
Olfactory organ. An organ to distinguish odors.
Oœcium. A structure in Polyzoa in which the embryo develops.

Oötype. The space in flat-worms where the eggs are supplied with shells.
Operculum. The horny lid that fits into the aperture of the shell of some gastropods.
Oral. The mouth side.
Osculum. The opening of a sponge through which water escapes.
Osphradium. A supposed sense organ of Mollusca.
Ossicle. A small hard plate.
Ostium. A small pore; in lamellibranchs one of the pores in the gills through which water is passed.
Otocyst. A sense organ, probably static in function.
Otolith. A hard body in an otocyst.
Ovary. A female sexual gland.
Oviducal gland. A glandular division of an oviduct. See squid.
Oviduct. A female sexual duct that leads from the ovary.
Ovipositor. A modified portion of some insects that is used in depositing eggs.
Ovum. Female germ cell.
Pallial line. The depression in the shell of a lamellibranch that is caused by the attachment of pallial muscles.
Pallial sinus. The indentation in the pallial line of some lamellibranchs, caused by the insertion of the retractor muscles of the siphons.
Pancreas. A digestive gland.
Papilla. A small projection.
Paragnatha. Lamellæ behind the mandibles of some Crustacea.
Parapodium. An appendage on a somite of an annelid.
Parenchyma. A soft tissue; that which fills the body-cavities of flatworms.
Pectine. One of a pair of appendages of scorpions.
Pedicellaria. A minute pincer-like organ that is present on some echinoderms.
Pedipalpi. Appendages of Arachnoidea.
Peduncle. A short stalk.
Pelagic. Organisms that live at or near the surface of the water.
Pen. Vestigial shell of a cephalopod. See squid.
Penis. Male intromittent organ.
Pericardium. A membrane surrounding the heart.
Periopod. A walking-leg of a crustacean.
Peri-pharyngeal bands. The ciliated bands that connect the endostyle and dorsal lamina of an ascidian around the mouth.
Periproct. The region around the anus (especially applied to the echinoderms).
Perisarc. The secreted outer covering of a hydroid.

Peristaltic. The motion caused by the successive contraction of the muscle fibers in the walls of a tube.
Peristome. The region around the mouth (especially applied to echinoderms).
Peristomium. The somite of an annelid that bears the mouth.
Peritoneum. The membrane that lines the celom.
Per-radial canals. The canals of a medusa that lie opposite the corners of the mouth.
Pharynx. An anterior division of the alimentary canal.
Planula. A young coelenterate embryo.
Pleopod. An abdominal appendage of a crustacean.
Pleurobranch. A gill of a crustacean that is borne on the body-wall.
Pleuron. One of the lateral pieces or processes of a somite of an arthropod.
Podobranch. A gill of a crustacean that is borne on the basal joint of an appendage.
Polymorphism. Many distinct forms of individuals.
Polyp. An individual of a hydroid stage of a ceelenterate.
Pore. A small opening.
Post-cava. A blood-vessel that leads to the heart from the posterior portion of the body. See squid.
Posterior. Hinder; anal end.
Pre-cava. A blood-vessel that leads to the heart from the anterior end of the body. See squid.
Primary mesentery. One of the vertical muscular partitions that extends from the body-wall to the esophagus of an actinozoan.
Proboscis. Applied to various tube-like organs around the head that are usually capable of being everted or protruded.
Proglottid. One of the pieces that compose the body of a cestode.
Propod. The next to the last segment, sixth, of a typical crustacean limb.
Prosopyle. One of the pores through which water passes from an incurrent to a radial canal of a sponge.
Prostomium. The anterior process that overhangs the mouth of an annelid.
Prothorax. Anterior division of the thorax of an insect.
Protomerite. The anterior part of Gregarina.
Protoplasm. Cell substance; living matter.
Protopod. The first two segments of a crustacean limb (the protopod bears the exopod and endopod).
Proximal. Toward the origin or attachment.
Pseudopodium. One of the changeable protoplasmic projections of the Sarcodina.
Pyloric division of stomach. Posterior or second division.

Radial symmetry. With the parts symmetrically radiating from a common center.
Radius. One of the parts of the jaw apparatus of an echinoid; from center to periphery.
Radula. The flexible membrane of an odontophore that bears the teeth.
Ray. One of the arms of a starfish.
Rectum. The posterior division of the alimentary canal.
Renal organ. An organ that excretes nitrogenous wastes and other materials.
Reservoir. A place where anything is stored; the poison sac of a bee.
Respiratory tree. The respiratory mechanism of some holothurians.
Retractor muscle. A muscle that withdraws an organ or portion.
Rostrum. The anterior spine of a lobster and of other crustaceans.
Rotula. One of the calcareous pieces of the jaw of an echinoid.
Rudimentary. When applied to adult animals means permanently undeveloped; vestigial.

Sagittal. In or parallel to the mesial plane.
Salivary gland. In invertebrates applied to any gland that opens into the mouth cavity.
Scaphognathite. The epipod of the second maxilla of certain Crustacea, that is used in baling water.
Scolex. Anterior portion of the tapeworm.
Segment. One of a series of divisions of an animal's body or appendage.
Segmentation. Frequently applied to the cleavage of an embryo.
Seminal receptacle. A sac in which spermatozoa are stored.
Seminal vesicles. The sacs that inclose the testes of an earthworm.
Septum. A plate that divides two spaces. See Nereis.
Sessile. Fixed; without the power of locomotion.
Seta. A small spine of an annelid that is usually of service in locomotion.
Setigerous gland. A gland that forms setæ.
Sexual. Of or pertaining to sex or sexes.
Shell gland. A gland that secretes the shell; sometimes applied to the kidneys of Entomostraca.
Siphon. Tubes for the transmission of water in Mollusea.
Somite. Metamere; one of the serial body-segments of an animal as an annelid.
Sperm. Spermatozoön; male reproductive cell.
Spermary. Male reproductive body.
Spermatheca. A seminal receptacle, used for storing spermatozoa in the female.
Spermatozoön. Male reproductive cell.
Sperm-sphere. A mass of spermatozoa in the earthworm.

Spicules. Minute skeletal bodies. See Grantia.
Spinneret. One of the organs by means of which a spider spins its thread.
Spiracle. Breathing pore; external opening of the tracheal system.
Spiral valve. The complicated posterior portion of the alimentary canal of a shark.
Spongin. The material of which the fibers of the commercial sponges are composed.
Spore. A small reproductive body formed asexually.
Sporont. The detached stage of Gregarina.
Sporulation. The act of forming spores.
Stalk. A stem or a peduncle.
Statoblast. Asexual reproductive body of certain Polyzoa.
Sternum. The ventral covering of a segment of an arthropod.
Stigma. One of the external openings of the trachea; one of the apertures in the pharynx of an ascidian.
Stolon. An extension of the body-wall from which buds are developed.
Stomach. A division of the alimentary canal.
Stomach-intestine. A division of the alimentary canal that functions as both stomach and intestine. See earthworm.
Stomodæum. The anterior portion of the alimentary canal that is ectodermal in origin.
Stone canal. The tube that leads from the madreporic plate to the circular water canal in echinoderms.
Stylet. A small sharp instrument.
Sub-genital pits. The pouches adjacent to the gonads of the Discomedusæ, on the subumbrellar side.
Sub-neural gland. A glandular body in ascidians.
Sub-umbrella. The concave (oral) surface of a medusa.
Sulcus. A furrow or groove.
Suture. An immovable union between plates or ossicles.
Swimmeret. Pleopod; an abdominal appendage of a crustacean
Swimming plate. One of the swimming organs of a ctenophore.
Synchronous. Happening at the same time.
Syphonoglyphe. A ciliated groove in some of the actinozoa.
Systemic heart. A heart that sends blood to the system. See squid.
Tactile Capable of feeling.
Tarsus. The segmented foot of an insect.
Telson. Hinder division or segment of a crustacean.
Tentacle. An elongated, unsegmented tactile organ
Tentaculocyst. A sense organ of certain medusæ.
Tergum. The dorsal covering of a segment of an arthropod.
Test. Shell of an echinoid; tunic of an ascidian.

Testis. Male genital gland.
Thorax. The body division of arthropods posterior to the head.
Tibia. The segment of the leg of an insect that is between the femur and tarsus.
Trachea. One of the respiratory tubes of certain arthropods.
Trichocyst. An infusorian defensive organ.
Trivium. The three rays of a starfish that are farthest from the madreporic plate.
Trochal disk. The ciliated disk of a rotifer.
Trochanter. The second segment from the body, of the leg of an insect.
Trochophore. An embryo of certain forms, such as the Annelida and Mollusca.
Tubercle. A small knob-like prominence.
Tunic. The outer covering of an ascidian.
Tympanum. A membrane of an auditory organ.
Typhlosole. A longitudinal ridge in the intestine.

Umbo. The raised portion of the valve of a clam shell that ends in the beak.
Umbrella. Applied to the arched portion of a medusa.
Uriniferous tube. One of the tubes of a kidney.
Uropod. One of the pair of abdominal appendages that, with the telson, form the tail-fin of a crustacean.
Uterus. A female organ in which young develop.

Vacuole, contractile. An excretory organ of Protozoa.
Vacuole, food. A temporary space in Protozoa in which food is digested.
Vagina. The terminal division of the female reproductive duct.
Vas deferens. The duct that leads away from the testicle.
Vas efferens. Sometimes applied to one of the divisions of the male reproductive duct of the squid.
Velum. The circular muscular membrane of a medusa.
Ventral. Under surface; belly.
Ventricle. A division of the heart which forces blood to the body.
Ventriculus. A division of the alimentary canal of an insect.
Vestibule. A depression near the mouth in certain Protozoa. See Vorticella.
Vestigial. An organ that remains undeveloped and has no function; rudimentary as applied in anatomy.
Viscera. Internal organs taken collectively.
Visceral mass. Applied to the portion of a mollusk that contains stomach, intestine, liver, gonads, etc.

Vitellarium. A female reproductive gland that supplies cells to be used as food for developing embryos. See Bdelloura.
Vitelline glands. Same as vitellarium.
Water tube. One of the tubes between the lamellæ of a lamellibranch gill.
Whorl. A turn of the shell of a gastropod.
Yolk-mass. A mass of food material for the nourishment of an embryo.
Zoöphyte. An animal that is somewhat plant-like in appearance.
Zoöid. One of the individuals in a united colony of animals. See Obelia and Bugula.

## INDEX.

References to directions for the study of forms are indicated by the use of boldface type for the page numbers.

Acanthocephala, 46
Acarida, 126
Acineta, 3
Acmæa, 90
Acrania, 164, 173
Acridium, 127, 154
Actiniaria, 19
Actinophrys, 1, 4
Actinosphærium, 1, 5
Actinozoa, 19, 30
Adelochorda, 164
Æginopsis, 18
Æolis, 90
Agalena, 126
Alcyonacea, 19
Alcyonaria, 19, 32
Alcyonium, 19
Amarœecium, 164, 170
Amoeba, 1, 3
Amœebida, 1
Amphineura, 89, 106
Amphioxus, 164, 173
Amphitrite, 70, 83
Annelida, 70
Anosia, 127
Antedon, 55
Anthomedusæ, 18
Antipatharia, 19
Aphis, 127
Apis, 128, 159
Aplacophora, 90
Aplysina, 14

Apoda, 55
Appendicularia, 164
Appendix, 175
Arachnida, 125
Arachnoidea, 125, 147
Araneida, 126
Arbacia, 54, 61
Arcella, 1, 4
Archi-annelida, 70
Archi-chætopoda, 70
Arenicola, 70, 85
Argonauta, 91
Argulus, 124, 146
Armata, 70
Arthostraca, 125
Articulata, 50
Ascaris, 46
Ascidiacea, 164
Ascidian, 165
Aspidobranchia, 90
Asterias, 54, 55
Asteroidea, 54, 55
Astrangia, 19, 31
Astropecten, 54
Astrophyton, 54
Aurelia, 19, 27
Autolytus, 70, 80
Balanoglossus, 164
Balanus, 124
Barnacle, 146
Bdelloura, 35, 37

Beach-flea, 141
Bee, 159
Beetle, 155
Benacus, 127
Berœ, 20
Beroida, 20
Blue crab, 134
Botryllus, 164, 169
Brachionus, 48
Brachiopoda, 50, 52
Branchipus, 124, 143
Buccinum, 90
Bug, 156
Bugula, 50
Bulla, 90
Buthus, 125, 149
Butterfly, 156, 161
Calcarea, 14
Callinectes, 125, 134
Cambarus, 125, 128
Campanularia, 23
Cancer, 125
Caprella, 125, 143
Caryophyllæus, 35
Centipede, 152
Cephalopoda, 90
Ceratium, 2, 8
Cerebratulus, 36, 44
Cestida, 20
Cestoda, 35, 41
Cestus, 20
Chætognatha, 46
Chætopleura, 90, 106
Chætopoda, 70, 71
Chætopterus, 70, 82
Chalina, 14, 17
Charybdea, 19
Chelifer, 125
Chilopoda, 126
Chiton, 90, 106
Chordata, 164
Cicada, 127

Ciliata, 3
Cirripathes, 19
Cirripedia, 124
Cistenides, 70, 84
Clam, 91, 104, 105
Clam-worm, 71
Clathrulina, 1, 6
Clava, 24
Clearing, 177
Clepsine, 71
Cliona, 14, 17
Clymenella, 70, 84
Clypeastroidea, 54
Coccidiida, 2
Coccidium, 2
Colenterata, 18
Coleoptera, 127
Copepoda, 124
Corticella, 14
Crab, 134, 138
Crab, horseshoe, 147
Crangon, 125
Craniata, 164
Crayfish, 128 $\qquad$
Crepidula, 90, 108
Crinoidea, 55
Crossobothrium, 35, 41
Crustacea, 124, 128
Cryptozonia, 54
Ctenophora, 19, 32
Cubomedusæ, 19
Culex, 127
Cumacea, 125
Cumingia, 91
Cuspidaria, 89
Cyclops, 124, 145
Cydippida, 20
Cypris, 124
Cystoflagellidia, 2
Daphnia, 124, 144
Decapoda (Arthropoda), 125
Decapoda (Mollusca), 91

Dehydrating, 176
Deiopea, 20
Demispongix, 14
Dentalium, 90
Dermacentor, 126
Diastylis, 125
Dibranchiata, 90
Dictyonina, 14
Difflugia, 1, 4
Digenetica, 35
Dinoflagellidia, 2
Dinophilea, 48
Diopatra, 70, 81
Diplopoda, 127
Diptera, 127
Discomedusx, 19
Distomum, 39
Dololium, 164
Dondersia, 90
Doryphora, 127
Earthworm, 74
Earwig, 152
Echinarachnius, 54
Echinodermata, 54
Echinoidea, 54, 61
Echinorhynchus, 46
Echiurus, 70
Ectoprocta, 50
Elasipoda, 55
Endoprocta, 50
Ensis, 91, 105
Entomostrica, 124
Epeira, 126, 150
Ephelota, 3
Eudendrium, 24
Euglena, 2, 6
Eulamellibranchia, 89
Eupagurus, 125, 138
Euplectella, 14
Euplotes, 3, 13
Eurete, 14
Euryalida 54

Euspongia, 14
Euthyneura, 90
Fairy shrimp, 143
Favia, 32
Filibranchia, 89
Fish-louse, 146
Fixing, 175
Flagellidia, 1
Fly, 156, 161
Foraminifera, 1, 4
Fresh-water mussel, 91
Fresh-water polyp, 20
Galeodes, 125
Gammarus, 125
Gastropoda, 90, 107
Gastrotricha, 48
Gephyrea, 70, 86
Gigantostraca, 125
Glossary, 180
Glossiphonia, 71
Gnathobdellida, 71
Gonionemus, 18, 25
Goose-barnacle, 146
Gordius, 46
Gorgonacea, 19
Gorgonia, 19, 32
Grantia, 14, 15
Grasshopper, 154
Gregarina, 2, 9
Gregarinida, 2
Gryllus, 127
Gymnolæmata, 50
Hematolechus, 39
Hæmosporidiida, 2
Haliotus, 90
Halteria, 3
Heliozoa, 1
Helix, 90
Hemiptera, 127
Hermit crab, 138

## INDEX.

Heterocœela, 14
Heterotrichida, 3
Hexactinellida, 14
Hexagenia, 127
Hippa, 125, 139
Hirudinea, 70
Hirudo, 71
Holothuroidea, 54, 67
Holotrichida, 3
Homarus, 125, 128
Homocœela, 14
Honey bee, 159
Horseshoe crab, 147
Hydra, 18, 20
Hydractinia, 24
Hydrocorallina, 18, 27
Hydroides, 70, 86
Hydrozoa, 18, 20
Hymenoptera, 128
Hypotrichida, 3
Inarticulata, 50
Inermia, 70
Infusoria, 3, 10
Insecta, 127, 154
Julus, 127, 153
Keratosa, 14
Lachnosterna, 127
Lamellibranchiata, 89, 91
Larvacea, 164
Laverania, 2
Lepas, 124, 146
Lepidonotus, 70, 81
Lepidoptera, 127
Lepisma, 127
Leptolinæ, 18
Leptomedusæ, 18
Leptoplana, 35
Leucosolenia, 14, 17
Libellula, 127

Lichnophora, 3, 13
Limax, 90
Limnæа, 90
Limulus, 125, 147
Lingula, 50
Lithobius, 126, 152
Lobata, 20
Lobster, 128
Loligo, 91, 115
Long clam, 104
Loxosoma, 50
Lumbricus, 70, 74
Lyssacina, 14

Madreporaria, 19, 31
Malacostraca, 124
Margelis, 24
Mastigamœba, 2
Mastigophora, 1, 6
Meandrina, 19, 32
Meckelia, 44
Melocerta, 48
Membranipora, 50
Metridium, 19, 30
Microstomum, 35
Millepora, 18, 27
Mnemiopsis, 20, 32
Modiola, 89, 100
Molgula, 164, 165
Mollusca, 89
Molluscoida, 50
Monaxonida, 14
Monogenetica, 35
Monozoa, 35
Mosquito, 161
Mounting, 177
Musca, 127
Mussel, 100
Mya, 89, 104
Myriapoda, 126, 152
Mysis, 124, 141
Mytilus, 89, 91, 100
Myxidium, 2

Myxospongida, 14
Myxosporidiida, 2
Myzostoma, 70
Myzostomida, 70
Narcomeduste, 18
Nautilus, 91, 123
Nebalia, 124
Nemathelminthes, 46
Nematoda, 46
Nemertinea, 36, 44
Neo-crinoidea, 55
Neomenia, 90
Neosporidia, 2
Nereis, 70, 71
Neuroptera, 127
Noctiluca, 2, 8
Nucula, 89
Obelia, 18, 22
Octopoda, 91
Octopus, 91, 123
Oligochæta, 70
Oniscus, 142
Onychophora, 126
Ophiura, 54, 60
Ophiurida, 54
Ophiuroidea, 54, 60
Opisthobranchia, 90
Orbicella, 19, 32
Orthoptera, 127
Oscarella, 14
Ostracoda, 124
Ostrea, 89, 103
Oxytricha, 3, 13
Oyster, 103
Paramectum, 3, 10
Parypha, 18, 24
Patella, 90
Pauropoda, 127
Pauropus, 127
Pecten, 89, 94101

Pectinatella, 50, 52
Pectinibranchia, 90
Pedata, 55
Pedicellina, 50
Pediculus, 127
Pedipalpida, 125
Pennaria, 24
Pennatula, 19
Pennatulacea, 19
Pentacrinus, 55
Peranema, 2, 7
Pericolpa, 19
Peridium, 2
Peripatus, 126
Periplaneta, 127
Peritromus, 3
Peromedusæ, 19
Perophora, 164, 168
Petasus, 18
Phacus, 7
Phalangida, 126
Phalangium, 126
Phanerozonia, 54
Phascolosoma, 70, 86
Philampelus, 127
Phoronida, 50
Phoronis, 50
Phoxichilidium, 126, 152
Phrynus, 125
Phylactolæmata, 50
Phyllapoda, 124
Phyllocarida, 124
Physalia, 18, 27
Placophora, 89
Planaria, 35, 36
Planocera, 35, 39
Platyhelminthes, 35
Platysamia, 127
Pleurobrachia, 20
Pleurotricha, 3, 13
Plumatella, 50, 52
Plumularia, 23
Podophyra, 3

Polychæta, 70
Polychœerus, 35
Polycladida, 35
Polygordius, 70
Polystomum, 35
Polyzoa (Cestoda), 35
Polyzoa (Molluscoida), 50
Pontobdella, 71
Porcellio, 125, 142
Porifera, 14
Preparations, 175
Prorodon, 3
Proterospongia, 2
Protobranchia, 89
Protozoa, 1
Pseudo-lamellibranchia, 89
Pseudo-scorpionida, 125
Pulmonata, 90
Pycnogonidia, 126, 152
Quоноя, 91
Radiolaria, 1
Razor-shell clam, 105
Regularia, 54
Renilla, 19, 32
Reticulariida, 1
Rhabdocœlida, 35
Rhizopoda, 1
Rhynchobdellida, 71
Rotifer, 48
Rotifera, 48
Round-web spider, 150
Sabella, 70, 85
Saccocirrus, 70
Sagartia, 19
Sagitta, 46
Salpa, 164, 172
Sand mole, 139
Sarcocystis, 3
Sarcodina, 1, 3
Sarcoptes, 126

Sarcosporidiida, 3
Scallop, 101
Scaphopoda, 90
Schizopoda, 124
Scolopendrella, 126
Scorpion, 149
Scorpionida, 125
Scyphozoa, 18, 27
Sea-anemone, 30
Sea-cucumber, 67
Sea-pork, 170
Sea-urchin, 61
Sections, 178
Sepia, 91
Septibranchia, 89
Serpent-star, 60
Sertularia, 23
Shrimp (fairy), $\mathbf{1 4 3}$
Silenia, 89
Siphonophora, 18, 27
Sminthurus, 127
Solenomya, 103
Solpugida, 125
Sow-bug, 142
Spatangoidea, 54
Spider, 150
Spirorbis, 70, 86
Spirostomum, 3, 11
Spirula, 91
Spongilla, 14, 17
Sporozoa, 2, 9
Squid, 115
Squilla, 125, 139
Staining, 176
Starfish, 55
Stauromedusæ, 19
Stentor, 3
Stomatopoda, 125
Streptoneura, 90
Strongylocentrotus, 54, 63
Stylaster, 27
Stylochus, 35
Stylonychia, 13

Suberites, 14
Sycotypus, 90, 107
Symphyla, 126
Synapta, 55
Syncœlidium, 35, 37
Tabanus, 127
Tænia, 35
Talorchestia, 125, 141
Telosporidia, 2
Terebratulina, 50, 52
Termes, 127
Tessera, 19
Tetrabranchiata, 91
Tetrastemma, 36, 44
Tetraxonida, 14
Thaliacea, 164
Thallassicolla, 1
Thousand-legs, 153
Thyone, 55, 67
Thysanura, 127
Trachelomonas, 7
Trachydermon, 90
Trachylinæ, 18
Trematoda, 35, 39

Trichina, 46, 47
Tricladida, 35
Tritia, 107
Trochelminthes, 48
Tubifex, 70
Tubipora, 19
Tunicate, 165
Turbellaria, 35, 36
Unio, 89, 91
Urochorda, 164, 165
Venus, 89, 91
Vertebrata, 164
Vespa, 128
Volvox, 7
Vorticella, 3, 11
Washing, 176
Water-flea, 145
Yoldia, 89, 91, 99
Zoantharia, 19
Zoöthamnium, 3

## SAUNDERS' BOOKS

## $\longrightarrow$ on

## Pathology, Physiology Histology, Embryology and Bacteriology

W. B. SAUNDERS COMPANY

925 Walnut Street
9. Henrietta Street

Philadelphia
Covent Garden, London

## LITERARY SUPERIORITY

THE excellent judgment displayed in the publications of the house at the very beginning of its career, and the success of the modern business methods employed by it, at once attracted the attention of leading men in the profession, and many of the most prominent writers of America offered their books for publication. Thus, there were produced in rapid succession a number of works that immediately placed the house in the front rank. One need only cite such instances as Keen's five volume work on Surgery, Kelly and Noble's Gynecology and Alsdominal Surgery, Fowler's Surgery, Ashton's Gynecology, Moynihan's works on Abdominal Operations and on Gallstones, Sahli, Kinnicutt, and Potter's Diagnostic Methods, Stengel's Pathology, Hir:t's Obstetrics, Anders' Practice, DaCosta's Surgery, Church and Peterson's Nervous and Mental Diseases, and the International Text-Book of Surgery, edited by Warren and Gould. These books have made for themselves a place among the best works on their several subjects.

[^23]
## American Text-Book of Pathology

American Text-Book of Pathology. Edited by Ludvig Hektoen, M. D., Professor of Pathology, Rush Medical College, in affiliation with the University of Chicago ; and David Riesman, M. D., Professor of Clinical Medicine, Philadelphia Polyclinic. Handsome imperial octavo, 1245 pages, 443 illustrations, 66 in colors. Cloth, $\$ 7.50$ net ; Sheep or Half Morocco, $\$ 9.00$ net.

## SUMPTUOUSLY ILLUSTRATED

The present work is the most representative treatise on the subject that has appeared in English. It furnishes practitioners and students with a comprehensive text-book on the essential principles and facts in General Pathology and Pathologic Anatomy, with especial emphasis on the relations of the latter to practical medicine. The pictorial feature of the work forms a complete atlas of pathologic anatomy and histology.

## Quarterly Medical Journal, Sheffield, England

" As to the illustrations, we can only say that whilst all of them are good, most of them are really beautiful, and for them alone the book is worth having. Both colored and plain, they are distributed so profusely as to add very largely to the interest of the reader and to help the student."

## McConnell's Pathology

A Manual of Pathology. By Guthrie McConnell, M.D., Pathologist to the Skin and Cancer Hospital, St. Louis. 12 mo of 523 pages, with 170 illustrations. Flexible leather, $\$ 2.50$ net.

## RECENTLY ISSUED

Dr. McConnell has discussed his subject wi h a clearness and precision of style that render the work of great assistance to both student and practitioner. The illustrations, many of them original, have been introduced for their practical value.

## Well.' Chemical Pathology

Chemical Pathology. By H. Gideon Wells, Ph.D., M.D., Assistant Professor of Pathology in the University of Chicago. Octavo of 549 pages. Cloth, $\$ 3.25$ net ; Half Morocco, \$4.75 net.

## JUST ISSUED

Dr. Wells discusses in the introductory chapter the chemistry and physics of the animal cell, giving the essential facts of the composition of proteids, and of ionization, diffusion, osmotic pressure, etc., and the relation of these facts to cellular activities. Special chapter are devoted to Diabetes and to Uric-acid Metabolism and Gout.

Wm. H. Welch, M. D.,
Professor of Pathology, Johns Hopkins Universily.
"The work fills a real need in the English literature of a very important subject. I shall be glad to recommend it."

## McFarland's Pathology

A Text-Book of Pathology. By Joseph McFarland, M.D., Professor of Pathology and Bacteriology in the MedicoChirurgical College of Philadelphia. Octavo of 8r8 pages, with 350 illustrations, many in colors. Cloth, $\$ 5.00$ net ; Half Morocco, $\$ 6.50$ net.

## RECENTLY ISSUED-BEAUTIFULLY ILLUSTRATED

Unlike most works on pathology, this work treats the subject, not from the professor's point of view, but from that of the student.

## American Medicine

"We feel confident in saying no other recent treatise, not encyclopedic in character on any subject, contains so much direct and correlated information on the branch with which it deals."

# Dürck and Hektoen's General Pathologic Histology 

Atlas and Epitome of General Pathologic Histology. By Pr. Dr. H. Dürck, of Munich. Edited, with additions, by Ludvig Hektoen, M. D., Professor of Pathology, Rush Medical College, Chicago. With 172 colored figures on 77 lithographic plates, 36 text-figures, many in colors, and 353 pages of text. In Saunders' Atlas Series. Cloth, $\$ 5.00$ net.

## RECENTLY ISSUED

This new atlas gives the accepted views in regard to the significance of pathologic processes, conflicting theories having been omitted. All the illustrations have been made from original specimens, in many cases as high as twenty-six colors having been required.

## W. T. Councilman, M.D.,

Professor of Pathologic Anatomy, Harvard University.
"I have seen no plates which impress me as so truly representing histologic appearances as do these. The book is a valuable one."

## Howell's Physiolog'y

A Text-Book of Physiology. By William H. Howell, Ph. D., M. D., Professor of Physiology in the Johns Hopkins University, Baltimore, Md. Octavo of 915 pages, with 275 illustrations. Cloth, $\$ 4.00$ net.

## JUST READY-THE NEW (2d) EDITION

Dr. Howell has had many years of experience as a teacher of physiology in several of the leading medical schools, and is therefore exceedingly well fitted to write a text-book on this subject. Main emphasis has been laid upon those facts and views which will be directly helpful in the practical branches of medicine. At the same time, however, sufficient consideration has been given to the experimental side of the science.

## The London Lancet

"This is one of the best recent books on physiology, and we warmly commend it to the attention of those who desire to obtain by reading a general all-round, yet concise survey of the scope, facts, theories, and speculations that make up its subject matter."

## Böhm, Davidoff, and Huber's Histology

A Text-Book of Human Histology. Including Microscopic Technic. By Dr. A. A. Böнm and Dr. M. von Davidoff, of Munich, and G. Carl Huber, M. D., Professor of Histology and Embryology in the University of Michigan, Ann Arbor. Handsome octavo of 528 pages, with 377 beautiful original illustrations. Flexible cloth, $\$ 3.50$ net.

## RECENTLY ISSUED-SECOND REVISED EDITION

The work of Drs. Böhm and Davidoff is well known in the German edition, and has been considered one of the most practically useful books on the subject of Human Histology. This American edition has been in great part rewritten and very much enlarged by Dr. Huber, who has also added over one hundred original illustrations. Dr. Huber's extensive additions have rendered the work the most complete students' text-book on Histology in existence.

## Drew' 5 Invertebrate Zoölogy

A Laboratory Manual of Invertebrate Zoölogy. By Gilman A. Drew, Ph. D., Professor of Biology at the University of Maine. With the aid of Members of the Zoölogical Staff of Instructors at the Marine Biological Laboratory, Woods Holl, Mass. 12 mo of 200 pages.

Cloth, \$1.25 net.

## JUST READY

The author has had extensive experience in the classroom and the laboratory, being in charge of zoological instruction at the Marine Biological Laboratory at Woods Holl, Mass. This training has fitted him most admirably to write such a book as this. The subject is presented in a logical way, and the type study has been following, as this method has been the prevailing one for many years. The typography is such that the student will find no difficulty whatever in following the classification. The work is very practical.

## GET THE BEST <br> American <br> THE NEW STANDARD Illustrated Dictionary <br> Just Issued-The New (4th) Edition

The American Illustrated Medical Dictionary. A new and complete dictionary of the terms used in Medicine, Surgery, Dentistry, Pharmacy, Chemistry, and kindred branches; with over 100 new and elaborate tables and many handsome illustrations. By W. A. Newman Dorland, M. D., Editor of "The American Pocket Medical Dictionary." Large octavo, nearly 850 pages, bound in full flexible leather. Price, $\$ 4.50$ net ; with thumb index, $\$ 5.00$ net.

## Gives a Maximum Amount of Matter in a Minimum Space, and at the Lowest Possible Cost <br> WITH 2000 NEW TERMS

The immediate success of this work is due to the special features that distinguish it from other books of its kind. It gives a maximum of matter in a minimum space and at the lowest possible cost. Though it is practically unabridged, yet by the use of thin bible paper and flexible morocco binding it is only $13 / 4$ inches thick. In this new edition the book has been thoroughly revised, and upward of two thousand new terms have been added, thus bringing the book absolutely up to date. The book contains hundreds of terms not to be found in any other dictionary, over 100 original tables, and many handsome illustrations.

## PERSONAL OPINIONS

[^24]
# Stengel's Text-Book of Pathology 

 New (5th) Edition-Just IssuedA Text-Book of Pathology. By Alfred Stengel, M. D., Professor of Clinical Medicine in the University of Pennsylvania. Octavo volume of 979 pages, with 400 text-illustrations, many in colors, and 7 full-page colored plates. Cloth, $\$ 5.00$ net ; Half Morocco, \$6.50 net.

## WITH 400 TEXT-CUTS, MANY IN COLORS, AND 7 COLORED PLATES

In this work the practical application of pathologic facts to clinical medicine is considered more fully than is customary in works on pathology. In this edition the section dealing with General Pathology has been most extensively revised, several of the important chapters having been practically rewritten. A very useful addition is an Appendix treating of the technic of pathologic methods, giving briefly the most important methods at present in use for the study of pathology, including, however, only those methods capable of giving satisfactory results.

## PERSONAL AND PRESS OPINIONS

## William H. Welch, M. D.,

Professor of Pathology, Johns Hopkins University, Baltimore, Md.
" I consider the work abreast of modern pathology, and useful to both students and practitioners. It presents in a concise and well-considered form the essential facts of general and special pathologic anatomy, with more than usual emphasis upon pathologic physiology."

## Ludvig Hektoen, M. D.,

Professor of Pathology, Rush Medical College, Chicago.
"I regard it as the most serviceable text-book for students on this subject yet written by an American author."

## The Lancet, London

" This volume is intended to present the subject of pathology in as practical a form as possible, and more especially from the point of view of the 'clinical pathologist.' These subjects have been faithfully carried out, and a valuable text-book is the result. We can most favorably recommend it to our readers as a thoroughly practical work on clinical pathology."

# Mallory and Wright's Pathologic Technique 

Recently Issued-Third Edition, Enlarged

Pathologic Technique. A Practical Manual for Workers in Pathologic Histology, including Directions for the Performance of Autopsies and for Clinical Diagnosis by Laboratory Methods By Frank B. Mallory, M. D., Associate Professor of Pathology, Harvard University ; and James H. Wright, M. D., Director of the Clinico-Pathologic Laboratories, Massachusetts General Hospital. Octavo of 469 pages, with 138 illustrations. Cloth, $\$ 3.00$ net.

## WITH CHAPTERS ON POST-MORTEM TECHNIQUE AND AUTOPSIES

In revising the book for the new edition the authors have kept in view the needs of the laboratory worker, whether student, practitioner, or pathologist, for a practical manual of histologic and bacteriologic methods in the study of pathologic material. Many parts have been rewritten, many new methods have been added, and the number of illustrations has been considerably increased. Among the many changes and additions may be mentioned the amplification of the description of the Parasite of Actinomycosis and the insertion of descriptions of the Bacillus of Bubonic Plague, of the Parasite of Mycetoma, and Wright's methods for the cultivation of Anaërobic Bacteria. There have also been added new staining methods for elastic tissue by -Weigert, for bone by Schmorl, and for connective tissue by Mallory.

## PERSONAL AND PRESS OPINIONS

William H. Welch, M.D.,<br>Professor of Pathology, Johns Hopkins University, Baltimore, Md.<br>"I have been looking forward to the publication of this book, and I am glad to say that 1 find it a most useful laboratory and post-mortem guide, full of practical information and well up to date."

## Boston Medical and Surgical Journal

"This manual, since its first appearance, has been recognized as the standard guide in pathological technique, and has become well-nigh indispensable to the laboratory worker."

# Heisler's <br> Text-Book of Embryology 

Just Ready-The New (3d) Edition

A Text-Book of Embryology. By John C. Heisler, M. D., Professor of Anatomy in the Medico-Chirurgical College, Philadelphia. Octavo volume of 435 pages, with 212 illustrations, 32 of them in colors. Cloth, $\$ 3.00$ net.

## WITH 212 ILLUSTRATIONS, 32 IN COLORS

The fact of embryology having acquired in recent years such great interest in connection with the teaching and with the proper comprehension of human anatomy, it is of first importance to the student of medicine that a concise and yet sufficiently full text-book upon the subject be available. The new edition of this work represents all the latest advances recently made in the science of embryology. Many portions have been entirely rewritten, and a great deal of new and important matter added. A number of new illustrations have also heen prepared which will prove valuable. Heisler's Embryology has become a standard work.

## PERSONAL AND PRESS OPINIONS

## G. Carl Huber, M. D.,

Professor of Histology and Embryology, University of Michigan, Ann Arbor.
"I find the second edition of 'A Text-Book of Embryology' by Dr. Heisler an improvement on the first. The figures added increase greatly the value of the work. I am agaiz recommending it to our students."

## William Wathen, M.D.,

Professor of Obstetrics, Abdominal Surgery, and Gynecology, and Dean, Kentucky School of Medicine, Louisville, Ky.
"It is systematic, scientific, full of simplicity, and just such a work as a medical student will be able to comprehend."

## Birmingham Medical Review, England

" We can most confidently recommend Dr. Heisler's book to the student of biology or medicine for his careful study, if his aim be to acquire a sound and practical acquaintance with the subject of embryology."

## McFarland's Pathogenic Bacteria

The New(5th)Edition, Revised

A Text-Book upon the Pathogenic Bacteria. By Joseph McFarland, M. D., Professor of Pathology and Bacteriology in the Medico-Chirurgical College of Philadelphia; Pathologist to the Medico-Chirurgical Hospital, Philadelphia, etc. Octavo volume of 647 pages, finely illustrated. Cloth, $\$ 3.50$ net.

## JUST ISSUED

This book gives a concise account of the technical procedures necessary in the study of bacteriology, a brief description of the life-history of the important pathogenic bacteria, and sufficient description of the pathologic lesions accompanying the micro-organismal invasions to give an idea of the origin of symptoms and the causes of death. The illustrations are mainly reproductions of the best the world affords, and are beautifully and accurately executed.

## H. B. Anderson, M. D.,

Associate I'rofessor of Clinical Medicine, University of Toronto.
"The book is a satisfactory one, and I shall take pleasure in recommending it to the students of the University."

## Hill's

## Histology and Organography

A Manual of Histology and Organography. By Charles Hill, M. D., Professor of Histology and Embryology, Northwestern University, Chicago. 12 mo of $46_{3}$ pages, with 313 illustrations. Flexible leather, $\$ 2.00$ net.

## RECENTLY ISSUED

Dr. Hill's fifteen years' experience as a teacher of histology has enabled him to present a work characterized by clearness and brevity of style and a completeness of discussion rarely met in a book of its pretensions. The mouth and teeth are given particular consideration.

## Eyre's Bacteriologic Technique

The Elements of Bacteriologic Technique. A Laboratory Guide for the Medical, Dental, and Technical Student. By J. W. H. Eyre, M. D., F. R. S. Edin., Bacteriologist to Guy's Hospital, London, and Lecturer on Bacteriology, Medical and Dental Schools, etc. Octavo, 375 pages, with 170 illustrations. Cloth, $\$ 2.50$ net.

This book presents, concisely yet clearly, the various methods at present in use for the study of bacteria, and elucidates such points in their life-histories as are debatable or still undetermined. The illustrations are numerous and practical.

## Medical News, New York

"Of the many laboratory guides constantly being issued, this book is undoubtedly the best that has reached us."

## Warren's

## Pathology and Therapeutics

Surgical Pathology and Therapeutics. By John Collins Warren, M. D., LL.D., F. R. C. S. (Hon.), Professor of Surgery, Harvard Medical School. Octavo, 873 pages, i36 relief and lithographic illustrations, 33 in colors. With an Appendix on Scientific Aids to Surgical Diagnosis, and a series of articles on Regional Bacteriology. Cloth, $\$ 5.00$ net ; Sheep or Half Morocco, $\$ 6.50$ net.

## SECOND EDITION, WITH AN APPENDIX

In the second edition of this book all the important changes have been embodied in a new Appendix. In addition to an enumeration of the scientific aids to surgical diagnosis there is presented a series of sections on regional bacteriology, in which are given a description of the flora of the affected part, and the general principles of treating the affections they produce.

## Roswell Park, M. D.,

In the Harvard Graduate Magazine.
"I think it is the most creditable book on surgical pathology, and the most beautiful medical illustration of the bookmakers' art that has ever been issued from the American press."

## Dürck anə Hektoen's Special Pathologic Histology

Atlas and Epitome of Special Pathologic Histology. By Dr. H. Dürck, of Munich. Edited, with additions, by Ludvig Hertoen, M. D., Professor of Pathology, Rush Medical College, Chicago. In two parts. Part I.-Circulatory, Respiratory, and Gastro-intestinal Tracts. 120 colored figures on 62 plates, and ${ }^{1} 58$ pages of text. Part II.-Liver, Urinary and Sexual Organs, Nervous System, Skin, Muscles, and Bones. 123 colored figures on 60 plates, and 192 pages of text. Per part : Cloth, $\$ 3.00$ net. In Saunders' Hand-Atlas Series.

The great value of these plates is that they represent in the exact colors the effect of the stains, which is of such great importance for the differentiation of tissue. The text portion of the book is admirable.

## William H. Welch, M. D.,

Professor of Pathology, Johns Hopkins University, Baltimore.
" I consider Diurck's 'Atlas of Special Pathologic Histology,' edited by Hektoen, a very useful book for students and others. The plates are admirable."

## Sobotta and Huber's Human Histology

Atlas and Epitome of Human Histology and Microscopic Anatomy. By Privatdocent Dr. J. Sobotta, of Würzburg. Edited, with additions, by G. Carl Huber, M. D., Professor of Histology and Embryology, and Director of the Histological Laboratory, University of Michigan, Ann Arbor. With 214 colored figures on 80 plates, 68 text-illustrations, and 248 pages of text. Cloth, $\$ 4.50$ net. In Saunders' Hand-Atlas Series.

[^25]
## American Text-Book of Physiology

American Text-Book of Physiology. In two volumes. Edited by William H. Howell, Ph. D., M. D., Professor of Physiology in the Johns Hopkins University, Baltimore, Md. Two royal octavo volumes of about 600 pages each, fully illustrated. Per volume: Cloth, $\$ 3.00$ net ; Sheep or Half Morocco, $\$ 4.25$ net.

## SECOND EDITION, REVISED AND ENLARGED

Even in the short time that has elapsed since the first edition of this work there has been much progress in Physiology, and in this edition the book has been thoroughly revised to keep pace with this progress. The chapter upon the Central Nervous System has been entirely rewritten. A section on Physical Chemistry forms a valuable addition, since these views are taking a large part in current discussion in physiologic and medical literature.

## The Medical News

"The work will stand as a work of reference on physiology. To him who desires to know the status of modern physiology, who expects to obtain suggestions as to further physiologic inquiry, we know of none in English which so eminently meets such a demand."

## Stewart's Physiology

## A Manual of Physiology, with Practical Exercises.

 For Students and Practitioners. By G. N. Stewart, M. A., M. D., D. Sc., Professor of Physiology in the University of Chicago. Octavo volume of 911 pages, with 395 text-illustrations and colored plates. Cloth, $\$ 4.00$ net.
## RECENTLY ISSUED-NEW (5th) EDITION

This work is written in a plain and attractive style that renders it particularly suited to the needs of students. The systematic portion is so treated that it can be used independently of the practical exercises, which constitute an important feature of the book. In the present edition a considerable amount of new matter has been added, especially to the chapters on Blood, Digestion, and the Central Nervous System.

## Philadelphia Medical Journal

"Those familiar with the attainments of Prof. Stewart as an original investigator, as a teacher and a writer, need no assurance that in this volume he has presented in a terse, concise, accurate manner the essential and best established facts of physiology in a most attractive manner."

# Levy and Klemperer's Clinical Bacteriology 

The Elements of Clinical Bacteriology. By Drs. Ernst Levy and Felix Klemperer, of the University of Strasburg. Translated and edited by Augustus A. Eshner, M. D., Professor of Clinical Medicine, Philadelphia Polyclinic. Octavo volume of 440 pages, fully illustrated. Cloth, $\$ 2.50$ net.

## Lehmann, Neumann, and Weaver's Bacteriolog'y

Atlas and Epitome of Bacteriology : including a TextBook of Special Bacteriologic Diagnosis. By Prof. Dr. K. B. Lehmann and Dr. R. O. Neumann, of Würzburg. From the Second Revised and Enlarged German Edition. Edited, with additions, by G. H. Weaver, M. D., Assistant Professor of Pathology and Bacteriology, Rush Medical College, Chicago. In two parts. Part I.-632 colored figures on 69 lithographic plates. Part II.-5II pages of text, illustrated. Per part: Cloth, $\$ 2.50$ net. In Saunders' Hand-Atlas Series.

## Lewis' Anatomy and Physiology

Anatomy and Physiology for Nurses. By LeRoy Lewis, M. D., Surgeon to and Lecturer on Anatomy and Physiology for Nurses at the Lewis Hospital, Bay City, Michigan. 12 mo of 317 pages, with 146 illustrations. Cloth, \$1.75 net.

## JUST ISSUED

## Nurses Journal of the Pacific Coast

" It is not in any sense rudimentary, but comprehensive in its treatment of the subjects in hand."

## Senn's Tumors

Second Revised Edition
Pathology and Surgical Treatment of Tumors. By Nicholas Senn, M. D., Ph. D., LL.D., Professor of Surgery, Rush Medical College, Chicago. Handsome octavo, 718 pages, with 478 engravings, including 12 full-page colored plates. Cloth, $\$ 5.00$ net; Sheep or Half Morocco, \$6.50 net.
"The most exhaustive of any recent book in English on this subject. It is well illustrated, and will doubtless remain as the principal monograph on the subject in our language for some years."-Journal of the American Medical Association.

## Stoney's Bacteriolog'y and Technic $\begin{gathered}\text { Recently Issued } \\ \text { New } \\ (2 d) \\ \text { Ed. }\end{gathered}$

Bacteriology and Surgical Technic for Nurses. By Emily M. A. Stoney, Superintendent of the Training School for Nurses at the Carney Hospital, South Boston, Mass. Revised by Frederic R. GrifFith, M.D., Surgeon, New York. 12 mo of 278 pages, profusely illustrated. Cloth, \$1.50 net.
"These subjects are treated most accurately and up to date, without the superfiuous reading which is so often employed. . . . Nurses will find this book of the greatest value."-The Trained Nurse and Hospital Review.

## Clarkson's Histolog'y

A Text-Book of Histology. Descriptive and Practical. For the Use of Students. By Arthur Clarkson, M. B., C. M. Edin., formerly Demonstrator of Physiology in the Owen's College, Manchester, England. Octavo, 554 pages, with 174 colored original illustrations. Cloth, $\$ 4.00$ net.
"The volume in the hands of students will greatly aid in the comprehension of a subject which in most instances is found rather difficult. . . . The work must be considered a valuable addition to the list of available text-books, and is to be highly recommended." -New York Medical Journal.

## Gorham's Bacteriology

A Laboratory Course in Bacteriology. For the Use of Medical, Agricultural, and Industrial Students. By Frederic P. Gorham, A. M., Associate Professor of Biology in Brown University, Providence, R. I., etc. 12 mo of 192 pages, with 97 illustrations. Cloth, $\$ \mathbf{I} .25$ net.
"One of the best students' laboratory guides to the study of bacteriology on the market. ... The technic is thoroughly modern and amply sufficient for all practical purposes."-American Journal of the Medical Sciences.

## Raymond's Physiology

Recently Issued New (3d) Edition

Human Physiology. By Joseph H. Raymond, A. M., M. D., Professor of Physiology and Hygiene, Long Island College Hospital, New York. Octavo of 685 pages, with 444 illustrations. Cloth, $\$ 3.50$ net.

[^26]
## Ball's Bacteriology Recently Issued-Fifth Edition, Revised

Essentials of Bacteriology : being a concise and systematic introduction to the Study of Micro-organisms. By M. V. Ball, M. D., late Bacteriologist to St. Agnes' Hospital, Philadelphia. $12 m o$ of 236 pages, with 96 illustrations, some in colors, and 5 plates. Cloth, \$1.co net. In Saunders' Question-Compend Series.
"The technic with regard to media, staining, mounting, and the like is culled from the latest authoritative works." - The Medical Times, New York.

## Budgett's Physiology <br> Recently Issued <br> New (2d) Edition

Essentials of Physioiogy. Prepared especially for Students of Medicine, and arranged with questions following each chapter. By Sidney P. Bulgett, M. D., Professor of Physiology, Medical Department of Washington University, St. Louis. 16 mo volume of 245 pages, finely illustrated with many full-page half-tones. Cloth, \$1.00 net. In Saunders' Question-Compend Series.
"He has an excellent conception of his subject . . It is one of the most satisfactory books of this class.-University of Pennsylvania Medical Bulletin.

## Leroy's Histology <br> Recently Issued <br> New (3d) Edition

Essentials of Histology. By Louis Leroy, M. D., Professor of Histology and Pathology, Vanderbilt University, Nashville, Tennessee. $12 \mathrm{mo}, 275$ pages, with 92 original illustrations. Cloths $\$ 1 . c o$ net. In Saunders' Question-Compend Series.
" The work in its present form stands as a model of what a student's aid should be; and we unhesitatingly say that the practitioner as well would find a glance through the book of lasting benefit."-The Medical World, Philadelphia.

## Bastin's Botany

Laboratory Exercises in Botany. By the late Edson S. Bastin, M. A. Octavo, 536 pages, with 87 plates. Cloth, $\$ 2.00$ net.

## Barton and Wells' Medical Thesaurus

A Thesaurus of Medical Words and Phrases. By Wilfred M. Barton, M. D., Assistant Professor of Materia Medica and Therapeutics; and Walter A. Wells, M. D., Demonstrator of Laryngology, Georgetown University, Washington, D. C. 12mo, 534 pages. Flexible leather, $\$ 2.50$ net.

## American Pocket Dictionary

Fifth Edition, Revised Just Issued

American Pocket Medical Dictionary. Edited by W. A. Newman Dorland, M. D., Assistant Obstetrician to the Hospital of the University of Pennsylvania. Containing the pronunciation and deffnition of the principal words used in medicine and kindred sciences, with 64 extensive tables. Handsomely bound in flexible leather, with gold edges, $\$ 1.00$ net; with patent thumb index, $\$ 1.25$ net.
"I can recommend it to our students without reserve."-J. H. Hollakd, M. D of the Yefferson Medical College, Philadelphia.

$$
0
$$



25


[^0]:    ${ }^{1}$ It is possible to decrease the rate of movement of both animal and cilia by placing it in a solution of gum arabic. Specimens so treated remain alive for some time.

[^1]:    ${ }^{1}$ There is some reason for thinking that the polyp stage of this form develops directly into the medusa. (See Perkins, Proc. Acad. Nat. Sci., Philadelphia, 1902.)

[^2]:    ${ }^{1}$ In most cases the foregoing canals are very evident, but if they are not, they may be injected with water in which carmine is mixed, by inserting a large-mouthed pipet into the stomach.

[^3]:    ${ }^{1}$ A Polyclad, Planocera, can be obtained without difficulty from the mantle chamber of Sycotypus. If Sycotypus is allowed to remain out of water for some hours the Planocera usually crawl out. The form is fairly satisfactory for study.

[^4]:    ${ }^{1}$ Specimens may be killed in the manner described for Bdelloura. Enough pressure should be used to flatten the proglottids decidedly.

[^5]:    ${ }^{1}$ This may be injected in fresh specimens, either with gelatin or fine starch-mass, by picking up one of the radial canals with a hypodermic syringe and injecting toward the disk.

[^6]:    ${ }^{1}$ These directions will serve for any of our common sea-urchins.

[^7]:    ${ }^{1}$ If a preserved specimen of Strongylocentrotus be placed in a solution of nitric acid (about 15 percent) from five to ten minutes, the plates of the test can be more easily seen, especially after drying. This is apparently due to the coloring-matter in the animal itself. Arbacia is not helped by the treatment.

[^8]:    ${ }^{1}$ The nervous system can be most readily studied by tearing it out with needles in a specimen which has been macerated in 20 percent nitric acid for twenty-four hours. Sensory cells and their neurites can be identified in the parapodia by placing them in a 1 percent solution of ammonium picrate after having let vigorous worms crawl around for three or four hours in a small amount of 1 percent solution of methylen-blue. Mounts of the parapodia should be made in a mixture of glycerin and ammonium picrate solution.

[^9]:    ${ }^{1}$ Small worms should be kept in a dish and fed on clean moistened filter-paper, which they will eat readily, until the alimentary canal is free

[^10]:    ${ }^{1}$ Points in which the fresh-water mussel differ have been noted, so the directions may be used for that form.
    ${ }^{2}$ Other species of lamellibranchs are more satisfactory than Venus for studying movements, as they expand quickly after being disturbed. Among the common ones that may be mentioned are Ensis, Cumingia, Yoldia, and Mytilus.

[^11]:    ${ }^{1}$ The general surface features are especially easily seen in Pecten, where the inter-filamentar junctions are small and well marked, and the inhalant ostia are correspondingly large and distinct.

[^12]:    ${ }^{1}$ This can be accomplished by breaking the shell away with the blade of a hatchet, and when enough of the shell has been removed, loosening the muscle from the columella with the thumb, and then pulling and twisting the animal out. When free from the shell place the animal in sea-water to which has been added about one-tenth its volume of alcohol and a little turpentine (about 10 c.c. of turpentine to each 100 c.c. of alcohol) and leave for several hours. An animal treated in this way will usually die with its proboscis extended. For the method we are indebted to Mr. Geo. M. Gray, Curator at the Marine Biological Laboratory, Woods Holl, Mass.

[^13]:    ${ }^{1}$ Instructors will find that a model prepared by sticking lumps and strands of modeling clay on a cylindrical graduate, to illustrate the positions of the ganglia and connectives on the esophagus, will greatly aid the students.

[^14]:    ${ }^{1}$ These directions may be used for the crayfish without much modification. The smaller size of these animals will make it more difficult to trace some of the nerves and blood-vessels

[^15]:    ${ }^{1}$ The circulatory system of a fresh specimen may be satisfactorily injected with starch-mass by inserting the needle of a hypodermic syringe into the pericardium from the posterior margin of the carapace. The operation is easily performed when the distance to the pericardium is understood.

[^16]:    ${ }^{1}$ In a fresh specimen the ganglia can be more easily studied after treating them with strong alcohol for a moment.

[^17]:    ${ }^{1}$ You should examine the posterior wing of a beetle and see how it is folded.

[^18]:    ${ }^{1}$ The mouth parts of all insects that depend on biting off portions of plants for food are quite similar. Directions for the study of the mouth parts of the honey-bee are given further on, but the mouth parts of other forms, such as the fly, butterfly, and bug, should be studied.

[^19]:    ${ }^{1}$ There is great diversity in the parts of the alimentary canals of different insects. The great differences in feeding habits render this necessary.

[^20]:    ${ }^{1}$ The arrangement of the ganglia in insects is very variable, showing many gradations in concentration.

[^21]:    ${ }^{1}$ The comparative study of the mouth parts of a butterfly, horse-fly, house-fly, and mosquito will prove valuable.

[^22]:    ${ }^{1}$ If the sections are not needed for study for a day or more, they may be floated out on water placed over the film of albumen. Heat the water until the sections stretch out flat, but do not melt the paraffin. In not less than twelve hours after the slide is thoroughly dry, it may be treated as directed in the other case. The value of this method is that it gives perfectly flat sections.

[^23]:    A Complete Catalogue of Our Publications will be Sent upon Request

[^24]:    Howard A. Kelly, M. D.,
    Professor of Gynecology, Johns Hopkins University, Baltimore.
    "Dr. Dorland's dictionary is admirable. It is so well gotten up and of such convenient size. No errors have been found in my use of it."
    J. Collins Warren, M.D.,

    Professor of Surgery, Harvard Medical School.
    "I regard it as" a valuable aid to my medical literary work. It is very complete and of convenient size to handle comfortably. I use it in preference to any other."

[^25]:    Lewellys F. Barker, M. D.,
    Professor of the Principles and Practice of Medicine, Johns Hopkins University.
    "I congratulate you upon the appearance of this volume. The illustrations are certainly very fine, and Dr. Huber has made important contributions to the text. The book should have a large sale."

[^26]:    "The book is well gotten up and well printed, and may be regarded as a trustworthy guide for the student and a useful work of reference, for the general practitioner. The illustrations are numerous and are well executed."-The Lancet, London.

