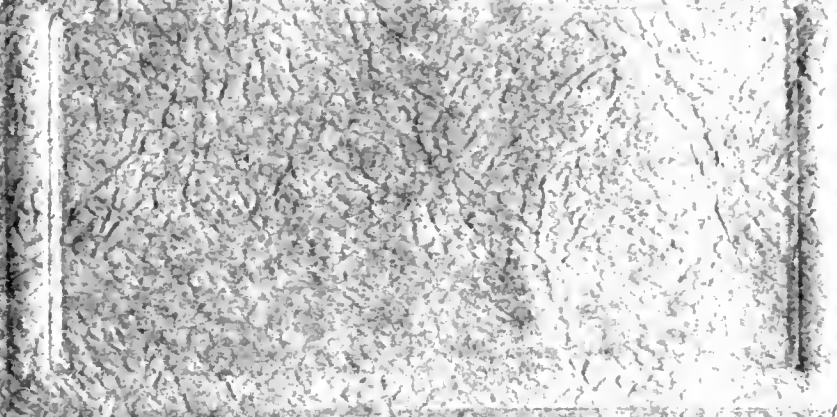


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II. PERTURBATIONS THROUGH BILLINGS

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PREPARED FOR PRESENTATION AT THE THIRTY-SIXTH ANNUAL MEETING  
OF THE MONTANA ACADEMY OF SCIENCES, APRIL 24, 1976 IN HARVE



# MICROFLORA OF THE YELLOWSTONE RIVER. II. PERTURBATIONS THROUGH BILLINGS

Loren L. Bahls

## INTRODUCTION

The last floristic survey of the Yellowstone River through Billings was accomplished 20 years ago when the river was receiving a pollution burden much greater than it receives today. In 1955, no bottom organisms occurred in the first 11 miles below waste outfalls at Billings; sewage "fungus" was commonplace; and taste and odor problems were chronic (10).

Today, due largely to the application of pollution control technology, the situation is much improved and the river is getting cleaner (2). Nevertheless, the Yellowstone River from Laurel to Billings remains water quality limited because of discharges from the Laurel and Billings sewage treatment plants, and wastewater discharges from three oil refineries, a sugar beet factory and a coal-fired steam-electric plant. There are also a number of non-point source sediment and oil problems in this reach of the river (5).

This paper describes the response of Yellowstone River algae to a variety of waste discharges originating in the Laurel-Billings municipal/industrial complex. Emphasis is placed on the relationship between the structure of benthic diatom associations and ambient concentrations of selected algal nutrients. This investigation was conducted as a contribution to the biological portion of a waste load allocation study being prepared for this section of the Yellowstone by the Montana Department of Health and Environmental Sciences.



## METHODS

### Sampling Stations and Schedule

Nine stations were sampled from Laurel downstream to Huntley, including the Clarks Fork River and Yegen Drain:

- I. Yellowstone River at Laurel (above Laurel wastewater discharge and confluence with the Clarks Fork River).
- II. Clarks Fork River at mouth.
- III. Yellowstone River at Duck Creek Bridge.
- IV. Yellowstone River at South Bridge (Billings).
- V. Yellowstone River below Corette plant.
- VI. Yellowstone River at East Bridge (Billings).
- VII. Yegen Drain at mouth.
- VIII. Yellowstone River below Yegen Drain (above Billings wastewater discharge).
- IX. Yellowstone River near Huntley.

Periphyton samples and water samples for algal nutrients were collected at these stations on the dates listed in Table I.

Table I. Sampling Schedule (All samples taken in 1975 except the nutrient sample at Station V, which was collected in 1974)

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	<u>Station and Date</u>								
	<u>I</u>	<u>II</u>	<u>III</u>	<u>IV</u>	<u>V</u>	<u>VI</u>	<u>VII</u>	<u>VIII</u>	<u>IX</u>
Nutrient Samples	9/9	9/9	9/9	9/9	7/22	9/9	10/23	10/7	9/9
Periphyton Samples	9/9	9/9	9/9	9/9	9/16	9/9	11/1	11/1	9/9

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## Field and Laboratory Procedures

At each station, periphyton samples were obtained by sampling natural substrates in proportion to the surface area of each type that was exposed for colonization. (Rocks predominated at most stations.) Substrates from both sluggish and rapidly flowing water were sampled in order to minimize possible bias caused by current effects. This procedure allows for collection of a composite sample that is representative of the range of physical conditions prevailing at each site at the time of collection.

From each sample, a subsample was taken and scanned microscopically to determine the presence and relative importance of non-diatom algae. Then, in a manner prescribed by the Environmental Protection Agency (4), each sample was acidified and oxidized, a permanent mount was prepared, and a diatom species proportional count was performed.

Nutrient analyses were performed at the Department of Health and Environmental Sciences' water laboratory in Helena following methods outlined by the American Health Association (1).

## Diversity Measures

Two diversity indexes were applied to the diatom species relative abundance figures obtained from the proportional counts: Margalef's index (9),

$$d = \frac{s - 1}{\ln N}$$

and Shannon's index (12),

$$D = - \sum_i^S (N_i/N) \log (N_i/N)$$



A series of corollary measures derived from the Shannon index were also applied:

$$D_{MAX} = \log (s)$$

$$D_{MIN} = \log (N) - \left( \frac{N - s + 1}{N} \right) \log (N - s + 1)$$

$$RD = \frac{D_{MAX} - D}{D_{MAX} - D_{MIN}}$$

$$EV = D/D_{MAX}$$

where  $s$  = number of taxa in the sample,  $N_i$  = number of individuals in taxon  $i$ , and  $N$  = total number of individuals counted.  $D_{MAX}$  and  $D_{MIN}$  are theoretical maximum and minimum diversities.  $RD$  or redundancy is an expression of dominance by one or more species and is inversely proportional to the wealth of species. (A value of zero is obtained if each individual belongs to a different species and a value of one is obtained if all individuals belong to the same species.)  $EV$  or evenness measures the equality of species abundances in a sample; the greater the disparities among species abundances, the smaller will be the evenness.

Another derived measure is Lloyd and Ghelardi's "equitability" (6),

$$e = \frac{s'}{s}$$

where  $s'$  is the number of taxa required to produce the observed Shannon index ( $D$ ) if the taxa are distributed according to MacArthur's "broken stick" model (8). Equitability is more sensitive to pollution than is  $D$ ; even slight levels of degradation have been found to reduce equitability below 0.5 and generally to a range of 0.0 to 0.3 (4).



## RESULTS

### Algal Nutrients

The results of algal nutrient analyses are presented in Table II. The Yegen Drain (VII) was a major contributor of all species of nitrogen and phosphorus. The Clarks Fork River (II) introduced appreciably higher levels of nitrate while phosphate was elevated below the Corette plant (V). Overall, comparing nutrient concentrations at Laurel (I) and Huntley (IX), nitrogen species were not appreciably concentrated by discharges through Billings, but phosphate and total phosphorous were.

### Non-Diatom Algae

Diatoms dominated the flora at all stations except in and below Yegen Drain where Oscillatoria and Stigeoclonium were the dominant algae, respectively. Euglena and a filamentous bacterium resembling Sphaerotilus were also evident at these two sites. Cladophora glomerata was abundant at Laurel and below the Corette discharge. The remaining 11 genera of non-diatom algae--all greens and blue-greens--were relatively uncommon.

### Diatoms

The structure of benthic diatom associations at the nine sites is given in Table III.

The seven major taxa are those that contributed 10 percent or more relative abundance in one or more collections. Pollution tolerances for these taxa were obtained from Cholnoky (3) and Lowe (7). Generally, Achnanthes minutissima and Cymbella affinis are intolerant of organic



Table II. Algal Nutrients in the Yellowstone River Through Billings (All values in mg/l)

Nutrient	Station								
	I	II	III	IV	V	VI	VII	VIII	IX
NO <sub>3</sub> + NO <sub>2</sub> (Total as N)	0.05	0.38	0.05	0.02	0.06	0.05	0.34	0.12	0.06
Ammonia (Total as N)	<0.01	<0.01	<0.01	<0.01	--	<0.01	3.2	0.46	<0.01
Nitrogen (Kjeldahl, Total as N)	0.18	0.45	0.24	0.26	--	0.15	4.0	1.0	0.24
Phosphate (PO <sub>4</sub> as P)	0.006	0.010	0.009	0.004	0.023	0.005	0.100	0.022	0.020
Phosphorus (Total as P)	0.013	0.016	0.013	0.016	--	0.014	0.124	0.032	0.027





Table III. Structure of Benthic Diatom Associations in the Yellowstone River Through Billings

Parameter	<u>Station</u>								
	I	II	III	IV	V	VI	VII	VIII	IX
Major Taxa (%)									
<u>Achnanthes minutissima</u>	2.0	35.8	25.4	25.0	18.6	21.0	7.4	1.7	9.9
<u>Cymbella affinis</u>	11.6	18.7	23.7	21.2	42.3	15.8	14.4	1.7	16.5
<u>Diatoma vulgare</u>	15.6	0.3	6.7	1.9	3.0	5.7	3.8	2.8	4.9
<u>Navicula cryptocephala v. veneta</u>	4.6	8.7	3.8	5.2	3.0	4.1	6.3	29.7	5.5
<u>Navicula mutica</u>								11.7	
<u>Nitzschia dissipata</u>	25.8	8.7	7.3	11.1	16.9	18.0	15.9	5.6	19.7
<u>Nitzschia palea</u>	3.7	2.0	4.7	3.0	1.2	2.7	9.0	27.8	4.3
Total <u>Nitzschia</u> species (%)	30.7	15.8	16.8	21.0	20.5	27.1	34.5	41.7	35.5
Taxa Observed	46	42	51	52	39	47	58	55	46
Taxa Counted (s)	32	28	33	37	27	36	49	37	31
Cells Counted (N)	352	358	342	368	338	366	367	360	345



Table III. (Continued)

Parameter	<u>Station</u>								
	I	II	III	IV	V	VI	VII	VIII	IX
Margalef Diversity (d)	5.29	4.59	5.48	6.09	4.47	5.93	8.13	6.12	5.13
Shannon Diversity (D)	3.74	3.27	3.66	3.81	2.80	3.83	4.47	3.31	3.87
Maximum Diversity (DMAX)	5.00	4.81	5.04	5.21	4.75	5.17	5.61	5.21	4.95
Minimum Diversity (DMIN)	0.87	0.74	0.92	0.97	0.75	0.95	1.29	0.99	0.85
Redundancy (RD)	0.30	0.38	0.33	0.33	0.49	0.32	0.27	0.45	0.26
Evenness (EV)	0.75	0.68	0.73	0.73	0.59	0.74	0.30	0.63	0.78
Equitability (e')	0.59	0.50	0.55	0.54	0.37	0.58	0.67	0.38	0.68



pollution; Diatoma vulgare and Nitzschia dissipata will tolerate only weak organic pollution but thrive where oxidation is complete; and Navicula cryptocephala var. veneta and Nitzschia palea are tolerant of organic pollution. The characteristically aerophilous Navicula mutica is an anomaly in the Yellowstone River. The total abundance of Nitzschia species is generally regarded as a suitable indicator of nitrogenous pollution. With one exception, to be discussed later, these indicator taxa behaved as expected considering their pollution tolerances and the nature and amount of enrichment. Relative abundance values for all major taxa were reasonably close at the stations bracketing the study section (stations I and IX).

The most striking feature about the diversity measures in Table III is the position held by the Yegen Drain collection (VII). Here, taxa observed, taxa counted, Margalef, Shannon, maximum and minimum diversities, evenness and equitability were all conspicuously and unexpectedly maximum. On the other hand, stations in the Clarks Fork River (II), below the Corette plant (V), and below Yegen Drain (VIII) all had depressed diversity levels indicating they were subject to some perturbation. As with relative abundances of the major species, values for diversity measures at the most upstream and downstream stations (I and IX) were fairly close (Table III).



## DISCUSSION AND CONCLUSIONS

On the basis of diversity measures, the most severely impacted station on the Yellowstone through Billings was below the Corette plant (V). The great abundance (42.3%) of Cymbella affinis here helped depress diatom diversity to the lowest levels recorded in the present study. Although nutrient data at this site are incomplete and dated, phosphate does appear to be significantly more concentrated here than upstream (Table II). However, the abundance of the saprophobic diatoms C. affinis and Achnanthes minutissima and the relatively minor importance of Nitzschiae indicate chemical water quality below the Corette plant to be rather good. Because C. affinis is a summer diatom, i.e., it prefers warmer waters (7), the stress causing depressed diversity at this location appears to be brought on by elevated temperature from the thermal discharge rather than by some chemical constituent introduced from the ash pond.

While burdened with a much heavier nutrient load (Table II), Yegen Drain (VII) had significantly higher diversity values than other study sites (Table III). Yet when this load was released into the Yellowstone River, diversity values were slightly depressed (Station VIII). One explanation might be that Yegen Drain offers a greater diversity of habitats and a physical environment, in terms of substrate, depth, temperature and flow regime, favorable to a larger variety of benthic diatoms. This situation deserves more attention and illustrates the fact that factors other than pollution load are responsible for biological diversity levels in streams, making them difficult to compare on this basis alone.





Nitzschia palea (Nitzsch)

organic nitrogen compounds. It is apparently in response to the high nitrogen load in the Snake River Drain (Tables II and III). The diatoms are able to utilize nitric acids and liberate free ammonia to some extent. This may be responsible for the rapid reduction in nitrate in the Snake River Drain and the eventual recovery to normal levels. With four times the nitrogen load, it is not surprising that the diatoms had only one-third the N. palea population in the Snake River downstream (VIII). This may be due to a combination of physical factors less favorable to N. palea in the Snake River.

Except for nitrate (Table II), water quality in the Snake River at Clarks Fork River (Station II) appears to be similar to that in the Yellowstone River. The relatively low diversity values (Table III) in the Snake River discharges through Laurel have no discernible effect on the Yellowstone River periphyton at Duck Creek Bridge.

The phytoplankton data from the Snake River are not strictly comparable to the data on the Yellowstone River. It is evident that water quality has improved in the Snake River in 20 years. On the whole, comparison of the periphyton of the Yellowstone River periphyton was not significantly different from that through Billings in 1975. Self-cleaning of the Snake River flora from pollution may be considered.



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