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Revisionary Notes on Australian Microhylid Frogs of the Genus Sphenophryne

By RICHARD G. ZWEIFEL¹

In a review of the microhylid frogs of Australia (Zweifel, 1962), I recognized three species of Sphenophryne and concluded that these were the forms named Austrochaperina robusta, A. gracilipes, and A. brevipes by Fry (1912, 1915). I proposed a substitute name, Sphenophryne fryi, for A. brevipes, because the name brevipes was preoccupied in the genus Sphenophryne, but I presumed that the other two species were valid.

Recently I was privileged to examine the specimens in the Australian Museum and the Macleay Museum in Sydney and the Zoölogisch Museum in Amsterdam on which Fry based his descriptions. I find that, contrary to my assumption, Fry did not name the three species that I recognized but named one species twice. Therefore, it is necessary to provide a new name for the species diagnosed in my previous paper as Sphenophryne robusta (Fry) and to allocate the name robusta to the proper species.

Methods of making measurements were detailed in my previous paper and need not be repeated here. I am grateful to Mr. S. Daan for assisting me in the examination of specimens at the Zoölogisch Museum, Amsterdam. Mr. Harold Cogger not only made the type specimens in the Australian Museum available to me, but also facilitated my study of specimens in the Macleay Museum, Sydney. My visits to the museums in Amsterdam

 $^{^{\}rm 1}\,{\rm Associate}$ Curator, Department of Herpetology, the American Museum of Natural History.

and Sydney were made possible by National Science Foundation Grant GB-2217. The following abbreviations are used:

A.M., Australian Museum, Sydney
A.M.N.H., the American Museum of Natural History
C.N.H.M., Chicago Natural History Museum
M.C.Z., Museum of Comparative Zoölogy at Harvard College
M.M., Macleay Museum, Sydney
Z.M.A., Zoölogisch Museum, Amsterdam

Sphenophryne robusta (Fry)

Austrochaperina robusta Fry, 1912, p. 89, fig. 37, pl. 9.
Austrochaperina gracilipes Fry, 1912, p. 93, pl. 8, figs. 1, 1a, 1b.
Sphenophryne gracilipes: Parker, 1934, p. 155. Loveridge, 1935, p. 56. Zweifel, 1956, p. 13; 1962, p. 30.

Fry based his original description of *robusta* on 10 specimens; nine he assigned to variety A and the tenth to variety B. Subsequently (1915), he gave variety B the specific name *brevipes*, and it now stands as *Sphenophryne fryi* Zweifel, 1962. The description of *gracilipes* was based on a single specimen.

I have examined the type specimen of gracilipes (A.M. No. 4536) and the type series of robusta. The type of robusta is A.M. No. 5295. Seven additional specimens of the type series are grouped together as M.M. No. 53, and one specimen (Z.M.A., No. 5877) is in Amsterdam. In addition there is A.M. No. 5296, an alcoholic specimen largely skeletonized. The specimens in the Macleay and Amsterdam museums are listed as cotypes, but evidently Fry used this term in the way "paratype" is used today, for in the catalogue of the Australian Museum he specifically designated one specimen as the type and others as cotypes.

Upon plotting the eye-naris and internarial measurements of the specimens in the type series of *robusta* on scatter diagrams previously published (Zweifel, 1962, figs. 9, 11), I found that these measurements did not coincide with those of the individuals that I had referred to this species. Rather, the type series of *robusta* appears to be composed of larger individuals of the form that I had called *gracilipes*. The distance from eye to naris and the internarial distance are both shorter in the types of *robusta* than that in specimens of comparable size that I referred to *robusta*. It appears, then, that I applied the name *robusta* incorrectly. The specimens to which I applied this name evidently represent an undescribed species which was unrepresented in Fry's collections. This species is named below in the present paper.

The question remains whether robusta and gracilipes are distinct species.

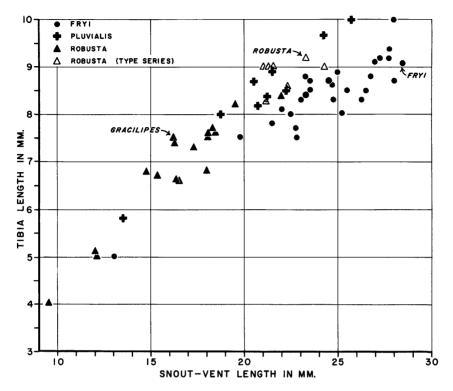


Fig. 1. Relationship of tibia length to body length in Australian Sphenophryne. Type specimens are indicated by names and arrows.

Although Fry gave detailed descriptions of each of them, he compared robusta and gracilipes only in a brief key (1912, p. 89) in which robusta falls in the couplet "Snout slightly prominent" and gracilipes in "Snout very prominent, longer than the orbital diameter." I find no differences between the type of gracilipes and the type series of robusta in the internarial or eye-naris measurements that best express snout shape. The ratio of internarial distance to snout-to-vent length (IN/S-V) in gracilipes is 0.099, whereas in eight specimens of robusta the average is 0.101, range 0.093-0.108. The ratio of eye-to-naris distance to snout-to-vent length (E-N/S-V) is 0.068 in gracilipes and averages 0.069 in eight specimens of robusta, range 0.064-0.077. The ratio of distance from eye to naris to internarial distance (E-N/IN) is 0.687 in gracilipes and averages 0.683, range 0.62-0.75, in eight individuals of robusta.

The legs of the type of gracilipes are proportionately slightly longer than those of any frog in the type series of robusta. The ratio of tibia length to snout-to-vent length (TL/S-V) is 0.463 in gracilipes, 0.399 (0.37-0.43) in

eight specimens of *robusta*. The two samples are similar in the ratio of head width to snout-to-vent length: 0.357 (0.32-0.40) in seven specimens of *robusta* and 0.333 in *gracilipes*.

The type specimens of both forms are now rather bleached, but there is nothing in the original descriptions indicative of striking differences in color or pattern.

I find no differences, therefore, between gracilipes and robusta that would confirm their specific distinctness. When the additional specimens that

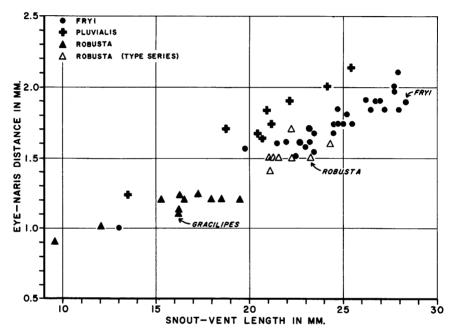


Fig. 2. Relationship of distance from eye to naris to body length in Australian Sphenophryne. Type specimens are indicated by names and arrows.

I had earlier referred to gracilipes are considered, this view is strengthened. The scatter diagrams of pertinent measurements of this group of specimens (figs. 1-3) suggest that only one species is represented. If robusta and gracilipes are indeed distinct species, as Fry supposed, I can find no way of diagnosing them.

The names gracilipes and robusta were proposed in the same publication. I follow page priority in choosing the name robusta for the species.

DIAGNOSIS: Sphenophryne robusta differs from S. fryi chiefly in leg length. The scatter diagram (fig. 1) shows little overlap in distribution, a fact that is confirmed by the ratio of tibia length to snout-to-vent length:

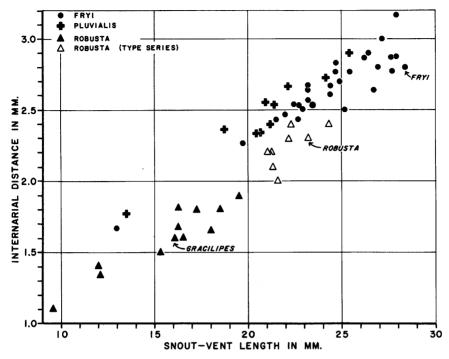


Fig. 3. Relationship of internarial distance to body length in Australian Sphenophryne. Type specimens are indicated by names and arrows.

robusta 0.413 ± 0.005 (0.37-0.46) $\mathcal{N}=25$; fryi 0.346 ± 0.004 (0.31-0.38) $\mathcal{N}=29$. Only eight specimens, four of each species, have ratios in the common zone of 0.37-0.38.

Sphenophryne robusta is compared with S. pluvialis in the description of the latter species that follows. Comparisons in my previous paper (Zweifel, 1962, pp. 35-36) of gracilipes with other species of Sphenophryne living in New Guinea will stand for robusta and those species.

DESCRIPTION: The description of gracilipes provided earlier (Zweifel, 1962, pp. 31-33) will serve for robusta as defined here.

DISTRIBUTION: Sphenophryne robusta is the most widely distributed Australian microhylid. It occurs from southern New Guinea to the type locality, the Russell River (between Innisfail and Cairns), Queensland. Specific locality records and a list of specimens examined were given under S. gracilipes in my previous paper (Zweifel, 1962, p. 36). To this list are to be added the type specimen (A.M. No. 5295) and eight "cotypes" (paratypes), all from the type locality: M.M. No. 53 (six specimens), A.M. No. 5296, and Z.M.A. No. 5877.

Sphenophryne pluvialis, new species

Austrochaperina robusta: Andersson, 1916, p. 7 (?part). Uperolia marmorata: Barbour, 1914, p. 202. Sphenophryne robustsa [sic]: Parker, 1934, p. 157 (?). Sphenophryne polysticta: Loveridge, 1935, p. 57 (part). Sphenophryne robusta: Zweifel, 1962, p. 19, fig. 7.

Type: A.M.N.H. No. 54195, collected by the Archbold Cape York Expedition at Speewah, elevation 1500 feet, Queensland, on April 9, 1948.

DIAGNOSIS: Sphenophryne pluvialis is distinguished from S. fryi by its relatively longer hind legs, longer snout, and slightly smaller maximum size. The mean ratio of tibia length to snout-to-vent length is 0.407 in pluvialis and 0.346 in fryi, and the minimum observed in pluvialis, 0.38,

TABLE 1

RATIOS OF TIBIA LENGTH AND HEAD WIDTH TO SNOUT-TO-VENT LENGTH
IN AUSTRALIAN SPECIES OF Sphenophryne

	TL/S-V			HW/S-V				
	Mean	σ m	Range	$\mathcal N$	Mean	σm	Range	\mathcal{N}
S. fryi	0.346 ±	0.004	(0.31-0.38)	29	0.397 ±	0.003	(0.33-0.42)	29
S. pluvialis	0.407 ±	0.007	(0.38-0.45)	10	0.377 ±	0.009	(0.36-0.45)	10
S. robusta	0.413 ±	0.005	(0.37-0.46)	25	0.343 ±	0.002	(0.30-0.41)	19

is equal to the maximum in fryi. Difference in snout shape is best expressed by the ratio of eye-to-naris distance to snout-to-vent length. The mean in pluvialis is 0.084 and in fryi is 0.071; as in leg length, the minimum ratio observed in pluvialis is equal to the maximum in fryi, 0.079. The largest specimen of fryi is the type, 28.4 mm. snout to vent, whereas the maximum length for any specimen in my series of pluvialis is 25.5 mm.

Sphenophryne pluvialis differs from its other Australian congener, S. robusta, in having a relatively shorter internarial distance and relatively less distance between eye and naris. Although ranges of the ratios E-N/S-V and IN/S-V (see table 2) overlap broadly if all specimens are considered, the distribution of these measurements on the scatter diagrams (figs. 2, 3) shows no overlap. The overlap in ratios results from changes in ratios with growth that produce curvilinear regression when plotted. When specimens less than 16 mm. in snout-to-vent length are omitted from the calculations, the ratios show no overlap: E-N/S-V, pluvialis 0.083 (0.079-0.090) $\mathcal{N}=9$, robusta 0.068 (0.064-0.077) $\mathcal{N}=16$; IN/S-V, pluvialis 0.117 (0.112-0.126) $\mathcal{N}=9$, robusta 0.100 (0.092-0.110) $\mathcal{N}=16$. Com-

TABLE 2	Ratios of Eye-to-Naris Distance and of Internarial Distance to Snout-to-Vent Length,	and of Eye-to-Naris Distance to Internarial Distance in Australian Species of Sphenophryne

29 10 20

 $\begin{array}{c} 0.109 \pm 0.001 \; (0.098 \hbox{--}0.127) \\ 0.118 \pm 0.002 \; (0.112 \hbox{--}0.131) \\ 0.103 \pm 0.001 \; (0.092 \hbox{--}0.117) \end{array}$

29 10 19

 $0.084 \pm 0.001 \ (0.079-0.091) \ 0.071 \pm 0.002 \ (0.064-0.094)$ $0.071 \pm 0.003 \ (0.065 - 0.079)$

29 10 19

 $\begin{array}{c} 0.649 \pm 0.006 \; (0.59 \hbox{--}0.72) \\ 0.714 \pm 0.005 \; (0.68 \hbox{--}0.74) \\ 0.698 \pm 0.012 \; (0.62 \hbox{--}0.82) \end{array}$

S. fryi S. pluvi ilis S. robusta

V-S/NI

E

Mean

E-N /S-V

Mean

≿

Range

Mean

E-N/IN дш parisons with species of Sphenophryne found in New Guinea were made in my previous paper (Zweifel, 1962, pp. 23-25) under the name S. robusta.

Description: The description of *Sphenophyrne robusta* (Zweifel, 1962, pp. 20-22) was based on the same specimens referred here to the new species, so it is unnecessary to repeat that description.

The name pluvialis alludes to the rain-forest habitat of this species.

DISTRIBUTION: Sphenophryne pluvialis is known from only three localities in the general vicinity of Cairns, Queensland: Kuranda (C.N.H.M. Nos. 29026, 29028-29032, M.C.Z. Nos. 7603, 19256, paratypes); Lake Eacham (C.N.H.M. No. 29034, paratype); Speewah, 1500 feet (A.M.N.H. No. 54165, holotype).

Sphenophryne fryi Zweifel

The status of this species remains unchanged. It may be added that my examination of the type specimen (A.M. No. 2285) confirms the conspecificity of the type and the specimens that I referred to this species. The similarity of the type to the other specimens is seen in figures 1-3.

KEY TO THE SPECIES OF Sphenophryne IN AUSTRALIA

The key that appeared in my previous paper (Zweifel, 1962, p. 38) requires modification, as is indicated below. The reader should consult the earlier paper for comments on the generic identification of Australian microhylid frogs. If measurements are made according to the methods I used (Zweifel, 1962, p. 4), the key that follows provides a means of distinguishing the forms included. The differences between species are sufficiently slight that even minor variation in the technique of measuring may give significantly different ratios.

1.	TL/S-V in mo	st specimens less	s than 0.38 (1	mean, 0.346)) S. fryi
2.	TL/S-V in mo	st specimens 0.3	8 or greater	(mean, 0.41))

	12/5 / m most specimens of or greater (mean, or m)
3.	E-N/S-V of adult specimens 0.079 or greater (mean, 0.083), IN/S-V 0.112
	or greater (mean, 0.117)
	E-N/S-V of adult specimens less than 0.079 (mean, 0.068), IN/S-V less than
	0.112 (mean, 0.100)

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