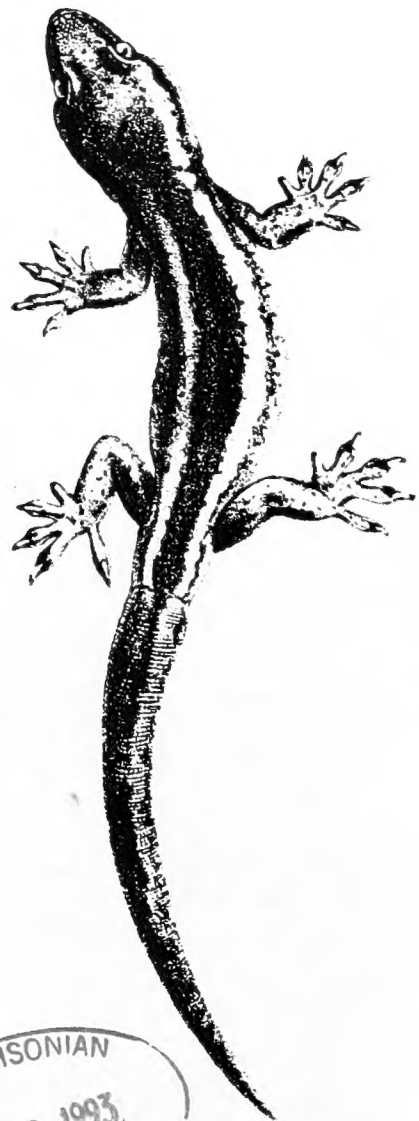


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~~K. P. SCHUMER~~

CHECKLIST
OF
GEKKONOID LIZARDS



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&
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INTRODUCTION

The following list includes all genus and species group names of extant gekkonoid lizards, their authors and years of publication. Most of the taxa I recognize are based on current common usage, and no attempt has been made to assess the monophyly of any genus. Junior objective and subjective synonyms are placed in parentheses. According to Article 80c of the ICZN, I follow existing usage in those cases under consideration by the Commission (e.g., Case 2527; BZN 46:38-40, 46-48, regarding *Nactus arnouxii* A. Duméril [1851] and *Nactus pelagicus* Girard [1858]). An asterisk is applied to all recognized subspecies, taxa followed by a question mark require further study, and a nomen nudum is followed by n.n. Not all unjustified emendations are noted in this abbreviated compilation. The list was derived from my manuscript of gekkonoid synonymies, wherein complete literature references, type localities and primary types are given. I would greatly appreciate notification of additions and corrections to the following list so that the complete version can be published with minimal errors. I wish to thank Aaron Bauer, Ross Sadlier and Glenn Shea for their assistance.

CHECKLIST

- Aclys* Kluge, 1974
 concinna Kluge, 1974 (*major** Storr, 1987)
- Aeluroscalabotes* Boulenger, 1887 (*Aelurosaurus* Boulenger, 1885)
 felinus Günther, 1864 (*borneensis* Günther, 1864;
 dorsalis W. Peters, 1871; *longicaudatus*
 Andersson, 1924; *multituberculatus** [?]
 Kopstein, 1927)
- Afroedura* Loveridge, 1944
 africana Boulenger, 1888 (*namaquensis** FitzSimons,
 1938; *tirasensis** Haacke, 1965)
 amatolica Hewitt, 1925
 bogerti Loveridge, 1944
 hawequensis Mouton and Mostert, 1985
 karroica Hewitt, 1925 (*wilmoti* Hewitt, 1926;
 *halli** Hewitt, 1935)
 nivaria Boulenger, 1894
 pondolia Hewitt, 1925 (*multiporis** Hewitt, 1925;
 *langi** FitzSimons, 1930; *marleyi** FitzSimons,
 1930; *haackei** Onderstall, 1984; *major**
 Onderstall, 1984)
 tembulica Hewitt, 1926
 transvaalica Hewitt, 1925 (*platyceps* Hewitt, 1925;
 *loveridgei** Broadly, 1963)
- Agamura* Blanford, 1874 (*Rhinogekko* de Witte, 1973)
 femoralis M. A. Smith, 1933

gastropholis Werner, 1917
misonnei de Witte, 1973
persica A. Duméril, 1856 (*cruralis* Blanford, 1874)

Ailuroonyx Fitzinger, 1843 (*Theconyx* Gray, 1845;
Aeluroonyx Agassiz, 1846)
seychellensis Duméril and Bibron, 1836
trachygaster A. Duméril, 1851

Alsophylax Fitzinger, 1843 (*Altiphylax* Jeriomtschenko
and Shcherbak, 1984)
laevis Nikolsky, 1907 (*kashkarovi* Andrushko, 1968)
loricatus Strauch, 1887 (*szczerbaki** Golubev and
Sattarov, 1979)
pipiens Pallas, 1811
przewalskii Strauch, 1887 (*microtis* Blanford,
1875)
tadjikiensis Golubev, 1979
tokobajevi Jeriomtschenko and Shcherbak, 1984

Aprasia Gray, 1839 (*Ophioseps* Barboza du Bocage, 1873;
Ophiopsiseps Boulenger, 1887)
aurita Kluge, 1974
fusca Storr, 1979
haroldi Storr, 1978
inaurita Kluge, 1974
parapulchella Kluge, 1974
pseudopulchella Kluge, 1974
pulchella Gray, 1839 (*nasutus* Barboza du Bocage,
1873; *brevirostris* Werner, 1909)
repens Fry, 1914
rostrata Parker, 1956
smithi Storr, 1970
striolata Lütken, 1863 (*lineolata* Lütken, 1863;
octolineata W. Peters, 1864; *glauerti* Parker,
1956)

Aristelliger Cope, 1862 (*Idiodactylus* Bocourt, 1870;
Aristelligella Noble and Klingel, 1932)
barbouri Noble and Klingel, 1932
cochranae Grant, 1931 (*expectatus** Cochran, 1933)
georgeensis Bocourt, 1873 (*irregularis* Cope, 1885)
hechti Schwartz and Crombie, 1975
lar Cope, 1862
praesignis Hallowell, 1857 (*nelsoni** Barbour,
1914)

Asaccus Dixon and S. Anderson, 1973
elisae Werner, 1895 (*eugeniae* Nikolskij, 1907;
ingae Eiselt, 1973)
gallagheri Arnold, 1972

- griseonotus* Dixon and S. Anderson, 1973
- Asiocolotes* Golubev, 1984
depressus Minton and J. A. Anderson, 1965
levitoni Golubev and Shcherbak, 1979
- Bavayia* Roux, 1913
crassicollis Roux, 1913)
cyclura Günther, 1872 (*neocaledonicus* Barboza du Bocage, 1873; *bavayi* Sauvage, 1878)
montana Roux, 1913
ornata Roux, 1913
sauvagii Boulenger, 1883
septuiclavis Sadlier, 1988
validiclavis Sadlier, 1988
- Bogertia* Loveridge, 1941
lutzae Loveridge, 1941
- Briba* Amaral, 1935
brasiliiana Amaral, 1935
- Bunopus* Blanford, 1874 (*Trachydactylus* Haas and Battersby, 1959)
crassicauda Nikolsky, 1907
spatalurus J. Anderson, 1901 (*jolensis* Haas and Battersby, 1959; *hajarensis** Arnold, 1980)
tuberculatus Blanford, 1874 (*blanfordii* Strauch, 1887; *gabrielis* Werner, 1936; *biporus* Werner, 1938; *abudhabi* Leviton and S. Anderson, 1967)
- Calodactylodes* Strand, 1870 (*Calodactylus* Beddome, 1870)
aureus Beddome, 1870
illingworthi Deraniyagala, 1953
- Carinatogekko* Golubev and Shcherbak, 1981
aspratilis S. Anderson, 1973
heteropholis Minton, S. Anderson, and J. A. Anderson, 1970
- Carphodactylus* Günther, 1897
laevis Günther, 1897
- Chondrodactylus* W. Peters, 1870
angulifer W. Peters, 1870 (*weirei* Boulenger, 1887; *namibensis** Haacke, 1975)
- Christinus* Wells and Wellington, 1984 (*Ridegekko* Wells and Wellington, 1985)
guentheri Boulenger, 1885
marmoratus Gray, 1845 (*peronii* Fitzinger, 1843; *australis* Gray, 1845; *macrodactylus* Boulenger, 1885; *affinis* Boulenger, 1885;

biggsae Wells and Wellington, 1985;
*alexanderi** Storr, 1987)

- Cnemaspis* Strauch, 1887 (*Ancylodactylus* L. Müller, 1907; *Paragonatodes* Noble, 1921)
affinis Stoliczka, 1870 (*penangensis* Flower, 1896)
africana Werner, 1895
argus Dring, 1978
barbouri Perret, 1986
beddomei Theobald, 1870 (*marmoratus* Beddome, 1870)
boiei Gray, 1842
boulengerii Strauch, 1887 (*glaucus* M. A. Smith, 1921)
dickersoni K. P. Schmidt, 1919
dilepis Perret, 1963
elgonensis Loveridge, 1935
lavolineata Nicholls, 1949
gigas Perret, 1986
goaensis Sharma, 1976
indica Gray, 1846
jerdonii Theobald, 1868 (*scalpensis** Ferguson, 1877)
kandiana Kelaart, 1852 (*malabaricus* Jerdon, 1853; *gracilis* Beddome, 1870; *wicksii* Stoliczka, 1873; *humei* Theobald, 1876; *andersonii* Annandale, 1904)
kendalli Gray, 1845
koehlerii Mertens, 1937
kumpoli Taylor, 1963
littoralis Jerdon, 1853 (*planiceps* Beddome, 1871)
mysoriensis Jerdon, 1853
nairi Inger, Marx and Koshy, 1984
nigridius M. A. Smith, 1925
occidentalis Angel, 1943
ornata Beddome, 1870
petrodroma Perret, 1986
podihuna Deraniyagala, 1944
quattuorseriata Sternfeld, 1912 (*bohmanni* L. Müller and Uthmoller, 1950)
siamensis M. A. Smith, 1925
sisparensis Theobald, 1876 (*maculatus* Beddome, 1870; *bireticulatus* Annandale, 1915)
spinicollis L. Müller, 1907
timoriensis Duméril and Bibron, 1836
tropidogaster Boulenger, 1885
uzungwae Perret, 1986
wynadensis Beddome, 1870
- Coleodactylus* Parker, 1926
amazonicus Andersson, 1918 (*zernyi* Wettstein, 1928)
brachystoma Amaral, 1935 (*pfrimeri*

Miranda-Ribeiro, 1939)
guimaraesi Vanzolini, 1957
meridionalis Boulenger, 1888
septentrionalis Vanzolini, 1980

Coleonyx Gray, 1845 (*Brachydactylus* W. Peters, 1863;
Anarbylus Murphy, 1974)
brevis Stejneger, 1893
elegans Gray, 1845 (*scapularis* A. Duméril, 1851;
coleonyx A. Duméril, 1856; *nemoralis**
 Klauber, 1945)
fasciatus Boulenger, 1885
mitratus W. Peters, 1863 (*dovii* Boulenger, 1885)
reticulatus Davis and Dixon, 1958
switaki Murphy, 1974 (*gypticolus** Grismer and
 Ottley, 1988)
variegatus Baird, 1859 (*abbotti** Klauber, 1945;
*bogerti** Klauber, 1945; *peninsularis**
 Klauber, 1945; *slevini** Klauber, 1945;
*sonoriensis** Klauber, 1945; *utahensis**
 Klauber, 1945)

Colopus W. Peters, 1869
wahlbergii W. Peters, 1869 (*kalaharicus*
 FitzSimons, 1932; *furcifer** Haacke, 1976)

Cosymbotus Fitzinger, 1843 (*Platyurus* Oken, 1836;
Nycteridium Günther, 1864; *Mimetozone*
 Boulenger, 1896)
craspedotus Mocquard, 1890 (*floweri* Boulenger,
 1896)
platyurus Schneider, 1792 (*schneideriana* Shaw,
 1802; *marginatus* Cuvier, 1829; *schneideri*
 Günther, 1864; *himalayanum* J. Anderson, 1871;
nepalensis Annandale, 1907)

Crenadactylus Dixon and Kluge, 1964
ocellatus Gray, 1845 (*bilineatus* Gray, 1845;
*horni** Lucas and Frost, 1895; *naso** Storr,
 1978; *rostralis** Storr, 1978)

Crossobamon Boettger, 1888 (*Ptenodactylus* Strauch, 1887)
eversmanni Wiegmann, 1834 (*atropunctatus*
 Lichtenstein and von Martens, 1856; *lumsdenii*
 Boulenger, 1887; *maynardi** M. A. Smith, 1933)
orientalis Blanford, 1876 (*dunstervillei* Murray,
 1884)

Cyrtopodion Fitzinger, 1843 (*Dinosauria* Gistel, 1868;
Mesodactylus Shcherbak and Golubev, 1984)
agamuroides Nikolsky, 1900
elongatus Blanford, 1875

kachhensis Stoliczka, 1872 (*petrensis* Murray,
1884)
montiumsalsorum Annandale, 1913
scaber Heyden, 1827 (*basoglui* Baran and Gruber,
1982)
watsoni Murray, 1892 (*ingoldbyi* Procter, 1923)

Delma Gray, 1831 (*Nisara* Gray, 1867; *Pseudodelma* Fischer, 1882)
australis Kluge, 1974
borea Kluge, 1974
butleri Storr, 1987 (*haroldi* Storr, 1987)
elegans Kluge, 1974
fraseri Gray, 1831
grayii A. Smith, 1849
impar Fischer, 1882 (*lineata* Rosen, 1905)
inornata Kluge, 1974
labialis Shea, 1987
mitella Shea, 1987
molleri Lütken, 1863
nasuta Kluge, 1974
pax Kluge, 1974
plebeia De Vis, 1888 (*wollemi* [n.n.] Wells and
Wellington, 1985)
tincta De Vis, 1888 (*reticulata* Garman, 1901)
torquata Kluge, 1974

Diplodactylus Gray, 1832 (*Stenodactylopsis*
Steindachner, 1870; *Lucasius* Kinghorn, 1929;
Lucasium Wermuth, 1965; *Turnerdactylus* Wells
and Wellington, 1989; *Ozziedactylus* Wells and
Wellington, 1989; *Manwellisaurus* Wells and
Wellington, 1989)
alboguttatus Werner, 1910
byrnei Lucas and Frost, 1896 (*dorotheae* Wells and
Wellington, 1985)
conspicillatus Lucas and Frost, 1897 (*hillii*
Longman, 1915; *laevis* Sternfeld, 1924;
platyurus Parker, 1926)
damaeus Lucas and Frost, 1896
fulleri Storr, 1978
galeatus Kluge, 1963
granariensis Storr, 1979 (*rex** Storr, 1988)
immaculatus Storr, 1988
kenneallyi Storr, 1988
maini Kluge, 1962
mitchelli Kluge, 1963
occultus King, Braithwaite and Wombey, 1982
ornatus Gray, 1845
polyophthalmus Günther, 1867
pulcher Steindachner, 1870 (*bilineatus* Lucas and
Frost, 1903; *dorsotaeniata* Pellegrin, 1909;
dorsalis Werner, 1910; *lucasi* Fry, 1914)

savagei Kluge, 1963
squarrosus Kluge, 1962
steindachneri Boulenger, 1885 (*stenurus* Werner,
 1909; *jonathoni* Wells and Wellington, 1985)
stenodactylus Boulenger, 1896 (*woodwardi* Fry,
 1914)
taenicauda De Vis, 1886 (*taeniocauda* Boulenger,
 1887)
tessellatus Günther, 1875 (*pachyurus* Werner, 1909)
vittatus Gray, 1832 (*furcosus* W. Peters, 1864;
barbouri Angel, 1936)
wombeyi Storr, 1978

Dravidogecko M. A. Smith, 1933
anamallensis Günther, 1875

Ebenavia Boettger, 1878
inunguis Boettger, 1878 (*boettgeri* Boulenger,
 1885)

Eublepharis Gray, 1827
angramainyu S. Anderson and Leviton, 1966 (*ensafi*
 Baloutch and Thireau, 1986)
hardwickii Gray, 1827 (*lunatus* Blyth, 1847)
macularius Blyth, 1854 (*fasciolatus** Günther,
 1864; *afghanicus** Borner, 1974; *gracilis*
 Borner, 1974; *montanus** Borner, 1976; *fuscus**
 Borner, 1981; *smithi** Borner, 1981)
turcmenicus Darevsky, 1978

Eurydactylodes Wermuth, 1965 (*Eurydactylus* Sauvage, 1878)
symmetricus Andersson, 1908
vieillardii Bavay, 1869

Geckoella Gray, 1867
collegalensis Beddome, 1870 (*speciosus* Beddome,
 1870)
deccanensis Günther, 1864 (*albofasciatus*
 Boulenger, 1885; *dekkansensis* M. A. Smith,
 1935)
jeyporensis Beddome, 1877
madarensis Sharma, 1980
nebulosus Beddome, 1870
triedrus Günther, 1864 (*punctata* Gray, 1867)
yakhuna Deraniyagala, 1945 (*zonatus* Deraniyagala,
 1945)

Geckolepis Grandidier, 1867
anomala Mocquard, 1902
maculata W. Peters, 1880 (*humbloti* Vaillant, 1887)
modesta Methuen and Hewitt, 1913
petiti Angel, 1942

polylepis Boettger, 1893
typica Grandidier, 1867

Geckonia Mocquard, 1895

chazaliae Mocquard, 1895 (*malazodes* Ahl, 1930)

Gehyra Gray, 1834 (*Peropus* Wiegmann, 1835; *Phyria* Gray, 1842; *Dactyloperus* Fitzinger, 1843; *Perodactylus* Fitzinger, 1843; *Peripia* Gray, 1845; *Chalinocnemis* Duges, 1883; *Spasmocnemis* Duges, 1883; *Phyriadora* Wells and Wellington, 1985)

angusticaudata Taylor, 1963

australis Gray, 1845 (*punctulata* Gray, 1842)

baliola A. Duméril, 1851 (*marmorata* Macleay, 1877; *brevicaudis* Macleay, 1877)

barea Kopstein, 1926

borroloola King, 1983

brevipalmata W. Peters, 1874

butleri Boulenger, 1900

catenata Low, 1979

dubia Macleay, 1877 (*longicaudis* Macleay, 1877; *torresiana* Günther, 1877)

fehlmanni Taylor, 1962

intermedia Brown, 1902

interstitialis Oudemans, 1894

kimberleyi Borner and Schuttler, 1983

lacerata Taylor, 1962

lampei Andersson, 1913

leopoldi Brongersma, 1930

marginata Boulenger, 1887 (*fischeri* Strauch, 1887; *marginata* Boettger, 1895)

membranacruralis King and Horner, 1989

minuta King, 1982

montium Storr, 1982

mutilata Wiegmann, 1835 (*peronii* Duméril and Bibron, 1836; *platurus* Bleeker, 1858;

insulensis Girard, 1858; *pardus* Tytler, 1865;

perdicolor Cope, 1868; *packardii* Cope, 1869;

navarri Duges, 1883; *beebei* Annandale, 1913)

nana Storr, 1978

occidentalis King, 1984

oceanica Lesson, 1830 (*pacifica* Gray, 1834;

oualensis Duméril and Bibron, 1836; *gularis* Gray, 1842)

pamela King, 1982

papuana Meyer, 1874

pilbara Mitchell, 1965 (*cognata* Borner and Schuttler, 1982)

punctata Fry, 1914 (*fenestra* Mitchell, 1965)

purpurascens Storr, 1982

robusta King, 1983

- variegata* Duméril and Bibron, 1836 (*grayi* Steindachner, 1867; *pusillus* [?] Cope, 1868; *mestoni* de Vis, 1890; *annetteae* Wells and Wellington, 1985; *kingi* Wells and Wellington, 1985; *lazelli* Wells and Wellington, 1985)
vorax Girard, 1858
xenopus Storr, 1978
- Gekko Laurenti*, 1768 (*Gecko* Brongniart, 1800; *Gecus* Rafinesque, 1810; *Platydactylus* Goldfuss, 1820; *Gecco* Ritgen, 1828; *Lomatodactylus* van der Hoeven, 1833; *Scelotretus* Fitzinger, 1843)
athymus Brown and Alcalá, 1962
auriverrucosus Zhou and Liu, 1982
chinensis Gray, 1842 (*similignum* M. A. Smith, 1923)
gecko Linnaeus, 1758 (*verticillatus* Laurenti, 1768; *teres* Laurenti, 1768; *guttatus* Daudin, 1802; *verus* Merrem, 1820; *annulatus* Kuhl, 1820; *reevesii* Gray, 1831; *tenuis* Hallowell, 1857; *indicus* Girard, 1858; *azhari** Mertens, 1955)
gigante Brown and Alcalá, 1978
hokouensis Pope, 1928 (*amissus* Taylor, 1962)
japonicus Schlegel (*in* Duméril and Bibron), 1836 (*jamori* Temminck and Schlegel, 1838; *nanus* Cantor, 1842; *yamori* Fritze, 1891)
kikuchii Oshima, 1912
liboensis Zhou and Li, 1982
mindorensis Taylor, 1919
palawanensis Taylor, 1925
palmatus Boulenger, 1907
petricolus Taylor, 1962
porosus Taylor, 1922
romblon Brown and Alcalá, 1978
scabridus Liu and Zhou, 1982
smithii Gray, 1842 (*stentor* Cantor, 1847; *albomaculatus* Giebel, 1861; *albofasciolatus* Günther, 1867)
subpalmatus Günther, 1864 (*melli* Vogt, 1922)
swinhonis Günther, 1864
taibaiensis Mingtao, 1985
tawaensis Okada, 1956
tuberculatus Daudin, 1802 (*monarchus* Schlegel, *in* Duméril and Bibron, 1836; *burmeisteri* Giebel, 1861; *deissneri* Giebel, 1861)
verreauxii Tytler, 1865
vittatus Houttuyn, 1782 (*unistriata* Shaw, 1792; *bivittatus* Duméril and Bibron, 1836; *trachyloemus* W. Peters, 1872)
yakuensis Matsui and Okada, 1968

Gonatodes Fitzinger, 1843

- albogularis* Duméril and Bibron, 1836 (*fuscus** Hallowell, 1855; *varius* A. Duméril, 1856; *notatus** Reinhardt and Lütken, 1862; *maculatus* Steindachner, 1867; *braconnieri* O'Shaughnessy, 1875; *bodinii** Rivero Blanco, 1964)
- annularis* Boulenger, 1887 (*boonii* Lidth de Jeude, 1904; *beebei* Noble, 1923)
- antillensis* Lidth de Jeude, 1887
- atricucullaris* Noble, 1921
- caudiscutatus* Günther, 1859 (*collaris* Garman, 1892)
- ceciliae* Donoso-Barros, 1965
- concinatus* O'Shaughnessy, 1881 (*buckleyi* O'Shaughnessy, 1881; *ligiae* Donoso-Barros, 1967)
- eladioi* Nascimento, Avila-Pires and Cunha, 1987
- falconensis* Shreve, 1947
- hasemani* Griffen, 1917 (*spinulosus* Amaral, 1932)
- humeralis* Guichenot, 1855 (*ferriugineus* 24Cope, 1864; *incertus* W. Peters, 1871; *sulcatus* O'Shaughnessy, 1875)
- ocellatus* Gray, 1831 (*confidentalis* Marcuzzi, 1950)
- petersi* Donoso-Barros, 1967
- seigliei* Donoso-Barros, 1965
- taniae* Roze, 1963
- tapajonicus* Rodrigues, 1980
- vittatus* Wiegmann, 1856 (*gilli* Cope, 1864; *roquensis** Roze, 1956)

Goniurosaurus Barbour, 1908 (*Amamisaurus* Borner, 1981)

- kuroiwae* Namiye, 1912 (*orientalis** Maki, 1931; *yamashinae* Okada, 1936; *splendens** Nakamura and Ueno, 1959)
- lichtenfelderi* Mocquard, 1897 (*hainanensis** Barbour, 1908)

Gonydactylus Kuhl and van Hasselt, 1822 (*Goniodactylus*

- Schlegel, 1826; *Cyrtodactylus* Gray, 1827; *Gonyodactylus* Wagler, 1830; *Goniodactylus* Gray, 1842; *Gonyodactylus* Fitzinger, 1843; *Puellula* Blyth, 1860; *Quantasia* Wells and Wellington, 1985)
- agusanensis* Taylor, 1915
- angularis* M. A. Smith, 1921
- annulatus* Taylor, 1915
- baluensis* Mocquard, 1890
- biordinis* Brown and McCoy, 1980
- brevipalmatus* M. A. Smith, 1923

cavernicolus Inger and King, 1961 (*kinabalensis*
Malkmus, 1989)
chitralensis M. A. Smith, 1935
condorensis M. A. Smith, 1921
consobrinoides Annandale, 1905
consobrinus W. Peters, 1871
darmandvillei Weber, 1890 (*defossei* Dunn, 1927)
dattanensis Khan, 1980
derongo Brown and Parker, 1973
deveti Brongersma, 1948
elok Dring, 1978
fasciolatus Blyth, 1860
feae Boulenger, 1893
fraenatus Günther, 1864
fumosus F. Müller, 1895 (*halmahericus** Mertens,
1929)
gubernatoris Annandale, 1913
himalayansis Duda and Sahi, 1978
indusoani Khan, 1988
ingeri Hikida, 1990
intermedius M. A. Smith, 1917
irregularis M. A. Smith, 1921
jellesmae Boulenger, 1897
khasiensis Jerdon, 1870 (*himalayicus* Annandale,
1906; *tamaiensis** M. A. Smith, 1940)
kirmanensis Nikolsky, 1900
laevigatus Darevsky, 1964 (*uniformis** Auffenberg,
1980)
lateralis Werner, 1896
lawderanus Stoliczka, 1871 (*himalayensis*
Annandale, 1913)
loriae Boulenger, 1897
lousiadensis de Vis, 1892 (*tuberculatus* Lucas and
Frost, 1900; *olivii* Garman, 1901)
malcolmsmithi Constable, 1949
malayanus de Rooij, 1915
mansarulus Duda and Sahi, 1978
marmoratus Gray, 1831 (*agamensis* Bleeker, 1860)
matsuii Hikida, 1990
mimikanus Boulenger, 1914
mintoni Golubev and Shcherbak, 1981
novaeguineae Schlegel, 1844
oldhami Theobald, 1876
papuensis Brongersma, 1928 (*novaeguineae*
Brongersma, 1928)
peguensis Boulenger, 1893 (*zebraicus** Taylor,
1962)
philippinicus Steindachner, 1867
pubisulcus Inger, 1957
pulchella Gray, 1827
quadrivirgatus Taylor, 1962
rohtasfortai Khan and Tasnim, 1990

redimiculus King, 1962
rubidus Blyth, 1860 (*tigris* Tytler, 1864)
sadleiri Wells and Wellington, 1985
sermowaiensis de Rooij, 1915
stoliczkai Steindachner, 1867 (*yarkandensis* J. Anderson, 1872; *walli* Ingoldby, 1922)
sworderi M. A. Smith, 1925
tibetinus Boulenger, 1905
variegatus Blyth, 1859
wetariensis Dunn, 1927
yoshii Hikida, 1990

Gymnodactylus Spix, 1825 (*Dasyderma* Fitzinger, 1843)
geckoides Spix, 1825 (*spixii* Gray, 1831;
gymnodactylus Schinz, 1833; *spinulosus*
 Fitzinger, 1843; *darwinii** Gray, 1845;
girardi Steindachner, 1867; *amarali** Barbour,
 1925; *conspicuus* Amaral, 1932; *helgae* Amaral,
 1950)
guttulatus Vanzolini, 1982

Hemidactylus Gray, 1825 (*Gecus* Rafinesque, 1810;
Hemidact Oken, 1817; *Boltalia* Gray, 1842;
Hoplopodion Fitzinger, 1843; *Microdactylus*
 Fitzinger, 1843; *Onychopus* Fitzinger, 1843;
Tachybates Fitzinger, 1843; *Pnoepus*
 Fitzinger, 1843; *Doryura* Gray, 1845;
Velernesia Gray, 1845; *Leiurus* Gray, 1845;
Nubilia Gray, 1845; *Eurhous* Fitzinger, 1861;
Liurus Cope, 1862; *Emydactylus* Bocourt, 1870;
Bunocnemis Günther, 1894; *Lophopholis* M. A.
 Smith and Deraniyagala, 1934; *Aliurus* Dunn
 and Dunn, 1940)
agrius Vanzolini, 1978
albopunctatus Loveridge, 1942
aporus Boulenger, 1906
arnoldi Lanza, 1978
barodanus Boulenger, 1901
bavazzanoi Lanza, 1978
bayonii Barboza du Bocage, 1893
bouvieri Bocourt, 1870 (*cessacii* Barboza du
 Bocage, 1873; *boavistensis** Boulenger, 1906;
chevalieri Angel, 1935; *razoensis** Gruber and
 Schleich, 1982)
bowringii Gray, 1845 (*berdmorei* Blyth, 1853)
brookii Gray, 1845 (*angulatus** Hallowell, 1854;
tytleri Tytler, 1865; *guineensis* W. Peters,
 1868; *affinis* Steindachner, 1870;
kushmorensis Murray, 1884; *gleadowi* Murray,
 1884; *stellatus* Boulenger, 1885; *murrayi*
 Gleadow, 1887; *tenkatei* Lidth de Jeude, 1895;
togoensis Werner, 1897; *haitianus** Meerwarth,

- 1901; *subtriedroides** Annandale, 1905;
*leightoni** Boulenger, 1911; *luzonensis*
 Taylor, 1915; *neotropicalis* Shreve, 1936;
*parvimaculatus** Deraniyagala, 1953)
citernii Boulenger, 1912
curlei Parker, 1942
depressus Gray, 1842 (*argentii* Gray, 1845;
pieresii Kelaart, 1853)
echinus O'Shaughnessy, 1875
fasciatus Gray, 1842 (*ornatus* Gray, 1845; *formosus*
 Hallowell, 1857; *ituriensis** K. P. Schmidt,
 1919)
flaviviridis Rüppell, 1835 (*coctaei* Duméril and
 Bibron, 1836; *sublevis* Gray, 1842; *cocteau*
 Fitzinger, 1843; *ruppelli* Fitzinger, 1843;
bengaliensis J. Anderson, 1871; *sublaevis*
 Boulenger, 1885; *bengalensis* Boulenger, 1885;
zollii Scortecci, 1929)
forbesii Boulenger, 1899
frenatus Duméril and Bibron, 1836 (*bojeri*
 Fitzinger, 1843; *javanicus* Fitzinger, 1843;
vittatus Gray, 1845; *punctatus* Jerdon, 1853;
inornatus Hallowell, 1861; *pumilus* Hallowell,
 1861; *caracal* Tytler, 1865; *chaus* Tytler,
 1865; *hexaspis* Cope, 1869; *longiceps* Cope,
 1869; *papuensis* Macleay, 1877; *tristis*
 Sauvage, 1878; *fragilis* Calabresi, 1915;
vandermeermohri Brongersma, 1928; *okinawensis*
 Okada, 1936)
funaiolii Lanza, 1978
garnotii Duméril and Bibron, 1836 (*peruvianus*
 Wiegmann, 1835; *vulpecula* Girard, 1858;
ludekingii Bleeker, 1858; *guadama* Theobald,
 1868; *mortoni* Theobald, 1868; *mandelianus*
 Stoliczka, 1871; *blanfordii* Boulenger, 1885)
giganteus Stoliczka, 1871
gracilis Blanford, 1870 (*platyceps* Annandale,
 1912)
granchii Lanza, 1978
granti Boulenger, 1899
greefi Barboza du Bocage, 1886
homoeolepis Blanford, 1881
intestinalis Werner, 1897 (*ansorgii* Boulenger,
 1901)
isolepis Boulenger, 1895
jubensis Boulenger, 1895
karenorum Theobald, 1868
klauberi Scortecci, 1948
laevis Boulenger, 1901
laticaudatus Andersson, 1910 (*fossatii** Scortecci,
 1928)
lemurinus Arnold, 1980

- leschenaultii* Duméril and Bibron, 1836 (*bellii* Gray, 1845; *pustulosus* Lichtenstein and von Martens, 1856; *kelaarti* Theobald, 1868; *marmoratus* Blanford, 1870)
longicephalus Barboza du Bocage, 1873 (*longiceps* O'Shaughnessy, 1873; *bocagii* Boulenger, 1885; *molleri* Bedriaga, 1892; *hecqui* Boulenger, 1901; *steindachneri* Tornier, 1902)
mabouia Moreau de Jonnès, 1818 (*tuberculosus* Raddi, 1823; *incanescens* Wied-Neuwied, 1824; *armatus* Wied-Neuwied, 1824; *aculeatus* Spix, 1825; *cruciger* Spix, 1825; *mabuia* Cuvier, 1829; *mabuya* Fitzinger, 1843; *calabaricus* Boettger, 1878; *benguellensis* Barboza du Bocage, 1893)
macropholis Boulenger, 1896
maculatus Duméril and Bibron, 1836 (*sykesii* Günther, 1864; *hunae** Deraniyagala, 1937)
marmoratus Hallowell, 1861
matschiei Toriner, 1901
megalops Parker, 1932
mercatorius Gray, 1842 (*gardineri* Boulenger, 1909; *persimilis* Barbour, 1928; *mandanus* Loveridge, 1936)
modestus Günther, 1894
muriceus W. Peters, 1870
newtoni Ferreira, 1897
nigriventris Lidth de Jeude, 1905
ophiolepis Boulenger, 1903
ophiolepidoides Lanza, 1978
oxyrhinus Boulenger, 1899
palaichthus Kluge, 1969
persicus J. Anderson, 1872 (*bornmuelleri* Werner, 1895)
platycephalus W. Peters, 1854
prashadi M. A. Smith, 1935
puccionii Calabresi, 1927
pumilio Boulenger, 1899 (*pumilus* Boulenger, 1899)
reticulatus Beddome, 1870
richardsonii Gray, 1845
ruspolii Boulenger, 1896 (*erlangeri* Steindachner, 1907)
scabriceps Annandale, 1906
sinaitus Boulenger, 1885 (*shugraensis* Haas and Battersby, 1959)
smithi Boulenger, 1895
somalicus Parker, 1932
squamulatus Tornier, 1896 (*bocagei* Toriner, 1896; *wernerii* Tornier, 1897; *tornieri* Mocquard, 1902; *floweri** Werner, 1908; *alluandi* Angel, 1923; *barbouri** Loveridge, 1942)
stejnegeri Ota and Hikida, 1989

- subtriedrus* Jerdon, 1853
tanganicus Loveridge, 1929
tasmani Hewitt, 1932
taylori Parker, 1932
triedrus Daudin, 1802 (*lankae** Deraniyagala, 1953)
tropidolepis Mocquard, 1888
turcicus Linnaeus, 1758 (*cyanodactylus* Rafinesque, 1810; *meridionalis* Risso, 1826; *granosus* Heyden, 1827; *robustus* Heyden, 1827; *veruculatus* Cuvier, 1829; *verrucosus* Gray, 1831; *karachiensis* Murray, 1884; *exsul* Barbour and Cole, 1906; *parkeri** Loveridge, 1936; *spinalis** Buchholz, 1954)
vietnamensis Darevsky and Kupriyanova, 1984
yerburii J. Anderson, 1895 (*pauciporosus** Lanza, 1978)
- Hemiphyllodactylus* Bleeker, 1860 (*Spathodactylus* Günther, 1872; *Spathoscalabotes* Boulenger, 1885; *Cainodactylus* Barbour, 1924)
harterti Werner, 1900 (*larutensis* Boulenger, 1900)
insularis Taylor, 1918
typus Bleeker, 1860 (*gracilis* Bleeker, 1860; *crepuscularis* Bavay, 1869; *aurantiacus** Beddome, 1870; *mutilatus* Günther, 1872; *ceylonensis* Boulenger, 1885; *leucostictus* Stejneger, 1899; *margarethae* Brongersma, 1931; *chapaensis** Bourret, 1937; *pallidus** Auffenberg, 1980)
yunnanensis Boulenger, 1903 (*dushanensis** Zhou and Liu, 1981; *jinpingsensis** Zhou and Liu, 1981; *longlingsensis** Zhou and Liu, 1981)
- Hemitheconyx* Stejneger, 1893 (*Psilodactylus* Gray, 1864)
caudicinctus A. Duméril, 1851
taylori Parker, 1930
- Heteronotia* Wermuth, 1965 (*Heteronota* Gray, 1845)
binoei Gray, 1845 (*derbianus* Gray, 1845; *australis* Steindachner, 1867; *anomalus* W. Peters, 1868; *annulatus* Macleay, 1877; *horneri* Wells and Wellington, 1985; *wadei* Wells and Wellington, 1985)
planiceps Storr, 1989
spelea Kluge, 1963
- Holodactylus* Boettger, 1893
africanus Boettger, 1893 (*aculeatus* Calabresi, 1915)
cornii Scortecci, 1931

- Homonota* Gray, 1845 (*Cubina* Gray, 1845; *Cubinia* Gray, 1845; *Wallsaurus* Underwood, 1954; *Garthia* Donoso-Barros and Vanzolini, 1965; *Garthia* Vanzolini and Donoso-Barros, 1966)
andicola Cei, 1978
borellii Peracca, 1897
darwinii Boulenger, 1885 (*macrocephala** Cei, 1978)
fasciatus Duméril and Bibron, 1836 (*pasteuri* Wermuth, 1965)
gaudichaudii Duméril and Bibron, 1836 (*dorbignii** Duméril and Bibron, 1836; *klugei** Donoso-Barros, 1970)
horrida Burmeister, 1861 (*mattogrossensis* Berg, 1895)
penai Donoso-Barros, 1965
underwoodi Kluge, 1964
uruguayensis Vaz-Ferreira and Soriano, 1961
whitii Boulenger, 1885
- Homopholis* Boulenger, 1885 (*Platypholis* Boulenger, 1890; *Blaesodactylus* Boettger, 1893)
antongilensis Bohme and Meier, 1980
boivini A. Duméril, 1856
fasciata Boulenger, 1890 (*erlangeri** Steindachner, 1906)
mulleri Visser, 1987
sakalava Grandidier, 1867 (*heterolepis* Boulenger, 1896)
walbergii A. Smith, 1849 (*macrolepis* Boulenger, 1885; *arnoldi* Loveridge, 1944)
- Hoplodactylus* Fitzinger, 1843 (*Pentadactylus* Gray, 1845; *Dactylocnemis* Steindachner, 1867; *Woodworthia* Garman, 1901)
chrysosireticus Robb, 1980
delcourti Bauer and Russell, 1986
duvaucelii Duméril and Bibron, 1836
granulatus Gray, 1845 (*brevidactylus* Gray, 1845; *greyii* Knox, 1870; *sylvestris* Buller, 1881; *versicolor* Colenso, 1885; *nebulosus* McCann, 1955)
kahutarae Whitaker, 1985
maculatus Boulenger, 1885
pacificus Gray, 1842 (*pomarii* Girard, 1858; *wullerstorfi* Fitzinger, 1861; *brunneus* Cope, 1868; *digitatus* Garman, 1901)
rakiurae Thomas, 1981
stephensi Robb, 1980
- Kaokogecko* Steyn and Haacke, 1966
vanzylis Steyn and Haacke, 1966

- Lepidoblepharis* Peracca, 1897 (*Lathrogecko* Ruthven, 1916)
buchwaldi Werner, 1910
colombianus Mechler, 1968
duolepis Ayala and Castro, 1983
festae Peracca, 1897
grandis Miyata, 1985
heyerorum Vanzolini, 1978
intermedius Boulenger, 1914
miyata Lamar, 1985
microlepis Noble, 1923
oxycephalus Werner, 1894
peraccae Boulenger, 1908
ruthveni Parker, 1926
sanctaemartae Ruthven, 1916 (*fugax* Ruthven, 1928)
williamsi Ayala and Serna, 1986
xanthostigma Noble, 1916
- Lepidodactylus* Fitzinger, 1843 (*Amydosaurus* Gray, 1845)
aureolineatus Taylor, 1915
balioburius Ota and Crombie, 1989
browni Pernetta and Black, 1983
christiani Taylor, 1917
euaensis Gibbons and Brown, 1988
gardineri Boulenger, 1897
guppyi Boulenger, 1884
herrei Taylor, 1923 (*medianus** Brown and Alcalá, 1978)
listeri Boulenger, 1889
lugubris Duméril and Bibron, 1836 (*neglectus* Girard, 1858; *meijeri* Bleeker, 1859; *pomareae* Fitzinger, 1861; *cantoris* Günther, 1864; *harrieti* Tytler, 1865; *moestus* W. Peters, 1867; *roseus* Cope, 1869; *caudeloti* Bavay, 1869; *mysorensis* Meyer, 1874; *ornata* Macleay, 1877; *divergens* Taylor, 1918; *lombocensis* Mertens, 1929; *ogasawarasimae* Okada, 1930; *intermedius** Darevsky, 1964)
magnus Brown and Parker, 1977
manni K. P. Schmidt, 1923
mutahi Brown and Parker, 1977
novaeguineae Brown and Parker, 1977
oorti Kopstein, 1926
orientalis Brown and Parker, 1977
planicaudus Stejneger, 1905 (*naujanensis* Taylor, 1919)
pulcher Boulenger, 1885
pumilus Boulenger, 1885
pusillus [?] Cope, 1868
ranauensis Ota and Hikida, 1988
shebae Brown and Tanner, 1949
woodfordi Boulenger, 1887

yami Ota, 1987

Lialis Gray, 1835 (*Ophiophthalmus* Fitzinger, 1843;
Alopecosaurus Lindholm, 1905)
burtonis Gray, 1835 (*bicatenata* Gray, 1842;
punctulata Gray, 1842; *leptorhyncha* W.
 Peters, 1874; *concolor* W. Peters, 1874)
jicari Boulenger, 1903 (*cuneirostris* Lindholm,
 1905; *inornata* Lindholm, 1905)

Luperosaurus Gray, 1845
brooksii Boulenger, 1920
browni Russell, 1979
cumingii Gray, 1845
joloensis Taylor, 1918
macgregori Stejneger, 1907
palawanensis Brown and Alcalá, 1974

Lygodactylus Gray, 1864 (*Scalabotes* W. Peters, 1880;
Vanzoia H. M. Smith, Martin and Swain, 1977)
angolensis Barboza du Bocage, 1896 [1897] (*laurae*
 K. P. Schmidt, 1933)
angularis Günther, 1893 (*heeneeni** DeWitte, 1933;
*grzimeki** Bannikov and Darevsky, 1969)
arnoulti Pasteur, 1964
bernardi FitzSimons, 1958 (*bonsi** Pasteur, 1962)
blanci Pasteur, 1967
bradfieldi Hewitt, 1932
capensis A. Smith, 1849 (*strigatus* Gray, 1864;
*grotei** Sternfeld, 1911; *ngamiensis*
 FitzSimons, 1932; *pakenhami** Loveridge, 1941)
chobiensis FitzSimons, 1932
conradti Matschie, 1892
conraui Tornier, 1902 (*strongi* Barbour and
 Loveridge, 1927)
decaryi Angel, 1930
depressus K. P. Schmidt, 1919
expectatus Pasteur and Blanc, 1967
fischeri Boulenger, 1890
grandisonae Pasteur, 1962
gravis Pasteur, 1964
guibei Pasteur, 1964
gutturalis Barboza du Bocage, 1873 (*paurospilus**
 Laurent, 1952; *dysmicus** Perret, 1963)
heterurus Boettger, 1913
inexpectatus Pasteur, 1964
insularis Boettger, 1913
keniensis Parker, 1936
klemmeri Pasteur, 1964
klugei H. M. Smith, Martin and Swain, 1977
lawrencei Hewitt, 1926
luteopicturatus Pasteur, 1964 (*zanzibaritis**)

Pasteur, 1964)
madagascariensis Boettger, 1881 (*pictus* W. Peters, 1883; *petteri** Pasteur and Blanc, 1967)
manni Loveridge, 1928
methueni FitzSimons, 1937
miops Günther, 1891 (*spinulifer* Boettger, 1913)
montanus Pasteur, 1964
ocellatus Roux, 1907
ornatus Pasteur, 1964
picturatus W. Peters, 1868 (*variegatus* W. Peters, 1868; *griseus* Toriner, 1896; *septemlineatus* Toriner, 1896; *quinquelineatus* Tornier, 1896; *mombasicus** Loveridge, 1935; *sudanensis** Loveridge, 1935; *ukerewensis* Loveridge, 1935)
pictus W. Peters, 1883 (*robustus* Boettger, 1913)
rarus Pasteur and Blanc, 1973
rex Broadly, 1963
scheffleri Sternfeld, 1912 (*laterimaculatus** Pasteur, 1964; *compositus** Pasteur, 1964; *ulugurensis** Pasteur, 1964)
scorteccii Pasteur, 1959
septemtuberculatus Angel, 1942
somalicus Loveridge, 1931 (*fisheri* Scorteccii, 1931; *annectans* Loveridge, 1935; *battersbyi** Pasteur, 1962; *howelli** Pasteur and Broadly, 1988)
stevensoni Hewitt, 1926
thomensis W. Peters, 1880 (*rolasi* Greef, 1885; *delicatus** Pasteur, 1962; *wermuthi** Pasteur, 1962)
tolampyae Grandidier, 1872 (*tuberifer* Boettger, 1913)
tuberosus Mertens, 1965
verticillatus Mocquard, 1895
wetzeli H. M. Smith, Martin and Swain, 1977
williamsi Loveridge, 1952

Mediodactylus Shcherbak and Golubev, 1977

amictopholis Hoofien, 1967
brevipes Blanford, 1874
heterocercus Blanford, 1874 (*mardinensis** Mertens, 1924)
kotschyi Steindachner, 1870 (*cyprius* Fitzinger, 1843; *concolor** Bedriaga, 1882; *maculatus** Bedriaga, 1882; *danilewskii** Strauch, 1887; *oertzeni** Boettger, 1888; *colchicus** Nikolsky, 1902; *bartoni** Stepanek, 1934; *fitzingeri** Stepanek, 1937; *orientalis** Stepanek, 1937; *solerii** Wettstein, 1937; *steindachneri** Stepanek, 1937; *stepaneki** Wettstein, 1937; *syriacus** Stepanek, 1937; *unicolor* Wettstein, 1937; *wettsteini**

Stepanek, 1937; *buresschi* Stepanek, 1937;
*rumelicus** L. Müller, 1939; *saronicus**
 Werner, 1939; *kalypsae** Stepanek, 1939;
*lycaonicus** Mertens, 1952; *rarus* Wettstein,
 1952; *stubbei* Wettstein, 1952; *christianae*
 Buchholz, 1955; *skopjensis** Karaman, 1965;
*buchholzi** Beutler and Gruber, 1977;
*schultzewestrumi** Beutler and Gruber, 1977;
gruberi Beutler and Gruber, 1977; *fuchsi**
 Beutler and Gruber, 1977; *bibroni** Beutler
 and Gruber, 1977; *adelpheisensis** Beutler and
 Gruber, 1978; *bileki* Tiedemann and Haupl,
 1980; *tinensis** Beutler and Fror, 1980;
*beutleri** Baran and Gruber, 1981; *karabagi**
 Baran and Gruber, 1982; *ciliciensis** Baran
 and Gruber, 1982; *ponticus** Baran and Gruber,
 1982)

russowii Strauch, 1887 (*zarudnyi** Nikolsky, 1899;
copalensis Schnitnikow, 1928)
sagittifer Nikolsky, 1899
spinicauda Nikolsky, 1887

Microgecko Nikolsky, 1907

helenae Nikolsky, 1907 (*fasciatus** Schmidtler and
 Schmidtler, 1972)
latifi Leviton and S. Anderson, 1972
persicus Nikolsky, 1903 (*bakhtiari** Minton, S.
 Anderson, J. A. Anderson, 1970;
*euphorbiacola** Minton, S. Anderson, and J. A.
 Anderson, 1970)

Microscalabotes Boulenger, 1883

bivittis W. Peters, 1883 (*hildebrandti* W. Peters,
 1883; *cowanii* Boulenger, 1883)

Millotisaurus Pasteur, 1962

mirabilis Pasteur, 1962

Nactus Kluge, 1983 (*Mascarenogecko* Ulber and Gericke, 1988)

pelagicus Girard, 1858 (*arnouxii* A. Duméril, 1851;
multicarinatus Günther, 1872; *arfakianus*
 Meyer, 1874; *fasciata* Macleay, 1877;
marmorata Macleay, 1877; *eboracensis* Macleay,
 1877; *cheverti* Boulenger, 1885; *heteronotus*
 Boulenger, 1885; *undulatus** Kopstein, 1926)
coindermirensis Bullock, Arnold and Bloxan, 1985
galgajuga Ingram, 1978
serpensinsula Loveridge, 1951
vankampeni Brongersma, 1933

Narudasia Methuen and Hewitt, 1914

festiva Methuen and Hewitt, 1914

- Naultinus* Gray, 1842 (*Heteropholis* Fischer, 1882;
Naultinulus Chrapliwy, H. M. Smith, and
 Grant, 1961)
elegans Gray, 1842 (*punctatus** Gray, 1843;
lineatus Gray, 1869; *sulphurus* Buller, 1871;
pentagonalis Colenso, 1880; *ocellatus* McCann,
 1955)
gemmeus McCann, 1955
grayii Bell, 1843 (*simpsoni* Robb, 1980)
manukanus McCann, 1955
poecilochlorus Robb, 1980
rudis Fischer, 1881 [1882]
stellatus Hutton, 1872 (*pulcherrimus* Buller, 1877)
tuberculatus McCann, 1955
- Nephrurus* Günther 1876 (*Underwoodisaurus* Wermuth, 1965)
asper Günther, 1876
deleani Harvey, 1983
laevissimus Mertens, 1958
laevis De Vis, 1886 (*platyurus* Boulenger, 1886;
*occidentalis** Storr, 1963; *pilbarensis**
 Storr, 1963)
milii Bory de Saint-Vincent, 1825 (*miliusii*
 Duméril and Bibron, 1836; *blavieri* de
 Rochebrune, 1884; *asper* Boulenger, 1913;
husbandi Wells and Wellington, 1984)
sphyrurus Ogilby, 1892 (*walshi* Kinghorn, 1931)
stellatus Storr, 1968
vertebralis Storr, 1963
wheeleri Loveridge, 1932 (*cinctus** Storr, 1963)
- Oedura* Gray, 1842 (*Pachyurus* Fitzinger, 1843; *Amalasia*
 Wells and Wellington, 1984)
castelnaui Thomillot, 1889 (*mayeri* Garman, 1901)
coggeri Bustard, 1966
filicipoda King, 1984
gemmata King and Gow, 1983
gracilis King, 1984
lesueurii Duméril and Bibron, 1836 (*phillipsi*
 Wells and Wellington, 1985)
marmorata Gray, 1842 (*fracticolor* De Vis, 1884;
cincta De Vis, 1888; *attenboroughi* Wells and
 Wellington, 1985; *derelicta* Wells and
 Wellington, 1985; *greeri* [n.n.] Wells and
 Wellington, 1985)
monilis De Vis, 1888
obscura King, 1984
reticulata Bustard, 1969
rhubifer Gray, 1845
robusta Boulenger, 1885
tryoni De Vis, 1884 (*ocellata* Boulenger, 1885)

verrilli [?] Cope, 1868

Ophidiocephalus Lucas and Frost, 1897
taeniatus Lucas and Frost, 1897

Pachydactylus Wiegmann, 1834 (*Colobopus* Fitzinger, 1843; *Cantina* Gray, 1845; *Homodactylus* Gray, 1864; *Elasmodactylus* Boulenger, 1894)

amoenus Werner, 1910

austeni Hewitt, 1923

bibronii A. Smith, 1845 [1846] (*turneri* Gray, 1864; *stellatus* Werner, 1910; *pulitzerae** K. P. Schmidt, 1933)

bicolor Hewitt, 1926

capensis A. Smith, 1846 (*elegans* Gray, 1845; *mentomarginatus* A. Smith, 1849; *oshaughnessyi** Boulenger, 1885; *obscurus* Thomiot, 1889; *affinis** Boulenger, 1896; *tessellatus* Werner, 1910; *leopardinus* Sternfeld, 1911; *levyi* FitzSimons, 1933; *vansoni** FitzSimons, 1933; *katanganus** DeWitte, 1953)

caraculicus FitzSimons, 1959

fasciatus Boulenger, 1888

formosus [?] A. Smith, 1849

gaiasensis Steyn and Mitchell, 1967

geitje Sparrman, 1778 (*ocellatus* Cuvier, 1817; *inunguis* Cuvier, 1817; *bergii* Wiegmann, 1834)

kobosensis FitzSimons, 1938

kochii FitzSimons, 1959

labialis FitzSimons, 1938

laevigatus Fischer, 1888 (*tessellatus* FitzSimons, 1938; *fitzsimonsi** Loveridge, 1947)

maculatus Gray, 1845 (*albomarginatus* Hewitt, 1932; *microlepis* Hewitt, 1935)

mariquensis A. Smith, 1849 (*latirostris** Hewitt, 1923; *macrolepis* FitzSimons, 1939)

mentalis Hewitt, 1926

monticolus [?] FitzSimons, 1943

namaquensis Sclater, 1898

oculatus Hewitt, 1927

oreophilus McLachlan and Spence, 1967

punctatus W. Peters, 1854 (*brunnthaleri* Werner, 1913; *langi* FitzSimons, 1932; *amoenoides** Hewitt, 1935; *scherzi** Mertens, 1954)

rugosus A. Smith, 1849 (*frater* Hewitt, 1935; *barnardi** FitzSimons, 1941)

sansteyni Steyn and Mitchell, 1967

scutatus Hewitt, 1927 (*robertsi** FitzSimons, 1938; *angolensis** Loveridge, 1944)

serval Werner, 1910 (*purcelli** Boulenger, 1910; *pardus* Sternfeld, 1911; *montanus* Methuen and

- Hewitt, 1914; *onscepensis** Hewitt, 1935)
tetensis Loveridge, 1953
tigrinus Van Dam, 1921 (*rhodesianus* Loveridge,
 1947)
tsodiloensis Haacke, 1966
tuberculosus Boulenger, 1894 (*triedrus* Boulenger,
 1913; *boulengeri* Toriner, 1896)
weberi Roux, 1907 (*gariesensis* Hewitt, 1932;
*weneri** Hewitt, 1935; *acuminatus**
 FitzSimons, 1941)
- Palmatogeko* Andersson, 1908 (*Syndactylosaura* Werner, 1910)
rangei Andersson, 1908 (*schultzei* Werner, 1910)
- Paradelma* Kinghorn, 1926
orientalis Günther, 1876
- Paragehyra* Angel, 1929
petiti Angel, 1929
- Paroedura* Günther, 1879
androyensis Grandidier, 1867 (*porogaster*
 Boulenger, 1896)
bastardi Mocquard, 1900
gracilis Boulenger, 1896
guibae Dixon and Kroll, 1974
homalorhinus Angel, 1936
oviceps Boettger, 1881
pictus W. Peters, 1854 (*madagascariensis* Mocquard,
 1895; *robustus* Boulenger, 1896)
sanctijohannis Günther, 1879
stumpffi Boettger, 1879
- Perochirus* Boulenger, 1885
ateles A. Duméril, 1856 (*articulatus* Fischer,
 1882; *depressus* Fischer, 1882; *klugei*
 Wermuth, 1965)
guentheri Boulenger, 1885
scutellatus Fischer, 1882
- Phelsuma* Gray, 1825 (*Anoplopus* Wagler, 1830; *Chiroperus*
 Wiegmann, 1834; *Rhoptropella* Hewitt, 1937;
Macrophelsuma Hoffstetter, 1946;
Archaeophelsuma Borner, 1972; *Neophelsuma*
 Borner, 1972)
abbotti Stejneger, 1893 (*sumptio** Cheke, 1982)
andamanensis Blyth, 1860 (*chamaelon* Tytler, 1864)
astriata Tornier, 1901 (*carinatum* [?] Rendahl,
 1939; *maheense* Rendahl, 1939; *astovei**
 FitzSimons, 1948; *semicarinata** Cheke, 1982)
barbouri Loveridge, 1942
befotakensis Borner and Minuth, 1982

- borbonica* Mertens, 1966 (*agalegae** Cheke, 1975)
cepediana Merrem, 1820
chekei Borner and Minuth, 1982
comorensis Boettger, 1913
dubia Boettger, 1881
flavigularis Mertens, 1962
gigas Lienard, 1842 (*newtonii* Günther, 1877;
newtonii Boulenger, 1884; *edwardnewtonii*
Vinson and Vinson, 1969)
guentheri Boulenger, 1885
guimbeaui Mertens, 1963 (*rosangularis** Vinson and
Vinson, 1969)
guttata Kaudern, 1922
laticauda Boettger, 1880 (*angularus** Mertens,
1964)
lineata Gray, 1842 (*bifasciata** Boettger, 1913;
*chloroscelis** Mertens, 1962; *bombetokensis**
Mertens, 1964; *dorsivittata** Mertens, 1964;
*pusilla** Mertens, 1964; *punctulata** Mertens,
1970; *leiogaster* Mertens, 1973)
madagascariensis Gray, 1831 (*tetradactylus* [?]
Schneider, 1797; *sarroubea* [?] Daudin, 1802;
*grandis** Gray, 1870; *kochi** Mertens, 1954;
martensi Mertens, 1962; *venusta** Mertens,
1964; *notissima* Mertens, 1970; *boehmei**
Meier, 1982)
minuthi Borner, 1980
modesta Mertens, 1970
mutabilis Grandidier, 1869 (*breviceps** Boettger,
1894; *androyense* Mocquard, 1901; *micropholis*
Boettger, 1913)
nigristriata Meier, 1984
ocellata Boulenger, 1885
ornata Gray, 1825 (*vinsoni* Mertens, 1963;
*inexpectata** Mertens, 1966)
parkeri Loveridge, 1941
quadriocellata W. Peters, 1883 (*bimaculata**
Kaudern, 1922; *leiura** Meier, 1983; *parva**
Meier, 1983)
robertmertensi Meier, 1980
serraticauda Mertens, 1963
standingi Methuen and Hewitt, 1913
sundbergi Rendahl, 1939 (*longinsulae** Rendahl,
1939; *pulchra* Rendahl, 1939; *cousinense*
Rendahl 1939; *menaiensis* Mertens, 1966;
*ladiguensis** Bohme and Meier, 1981; *rubra*
Borner and Minuth, 1982; *umbrae* Borner and
Minuth, 1982)
trilineata Gray, 1842
v-nigra Boettger, 1913 (*pasteuri** Meier, 1984)

- Phyllodactylus* Gray, 1828 (*Euleptes* Fitzinger, 1843;
Discodactylus Fitzinger, 1843; *Gerrhopygus*
 Gray, 1845)
angelensis Dixon, 1966
angustidigitus Dixon and Huey, 1970
ansorgii Boulenger, 1907
apricus Dixon, 1966
barringtonensis Van Denburgh, 1912
baurii Garman, 1892 (*gorii** Lanza, 1973)
bordai Taylor, 1942
bugastrolepis Dixon, 1966
clinatus Dixon and Huey, 1970
darwini Taylor, 1942
davisi Dixon, 1964
delcampi Mosauer, 1936
dixoni Rivero-Blanco and Lancini, 1968
duellmani Dixon, 1960
europaeus Gene, 1839 (*wagleri* Fitzinger, 1843;
doriae Lataste, 1877)
galapagensis W. Peters, 1869 (*daphnensis** Van
 Denburgh, 1912; *duncanensis* Van Denburgh,
 1912; *maresi** Lanza, 1973; *olschkii** Lanza,
 1973)
gerrhopygus Wiegmann, 1835 (*gymnopygus* Duméril and
 Bibron, 1836)
gilberti Heller, 1903
heterurus Werner, 1907
homolepidurus H. M. Smith, 1935 (*nolascoensis**
 Dixon, 1964)
inaequalis Cope, 1875
insularis Dixon, 1960
interandinus Dixon and Huey, 1970
johnwrighti Dixon and Huey, 1970
julieni Cope, 1885
kofordi Dixon and Huey, 1970
lanei H. M. Smith, 1935 (*magnatuberculatus* Taylor,
 1940; *rupinus** Dixon, 1964)
leei Cope, 1889
lepidopygus Tschudi, 1845 (*phacophorus* Tschudi,
 1845; *nigrofasciatus* Cope, 1878; *variegatus*
 Werner, 1900)
lineatus Gray, 1838 (*essexi** Hewitt, 1925;
*rupicolus** FitzSimons, 1938)
martini Lidth de Jeude, 1887
melanostictus Taylor, 1962
microlepidotus FitzSimons, 1939
microphyllus Cope, 1876
muralis Taylor, 1939 (*isthmus** Dixon, 1964)
palmeus Dixon, 1968
partidus Dixon, 1966
paucituberculatus Dixon, 1960

peringueyi [?] Boulenger, 1910
porphyreus Daudin, 1802 (*brevipes* Mocquard, 1900;
namaquensis Hewitt, 1935; *cronwrighti* Hewitt,
 1937)
pulcher Gray, 1828 (*spatulatus* Cope, 1863)
pumilis Dixon and Huey, 1970
reissii W. Peters, 1862 (*baessleri* Werner, 1900;
guayaquilensis Werner, 1910; *abrupteseriatus*
 Werner, 1913; *magister* Noble, 1924)
riebeckii W. Peters, 1882
rutteni Hummelinck, 1947
santacruzensis Dixon, 1966
sentosus Dixon and Huey, 1970
siamensis Boulenger, 1898 (*paviei* Mocquard, 1904;
burmanicus Annandale, 1905)
tinklei Dixon, 1966
trachyrhinus Boulenger, 1899
transversalis Huey, 1975
tuberculosis Wiegmann, 1835 (*magnus** Taylor, 1940;
edwardofischeri Mertens, 1952; *ingeri** Dixon,
 1964; *saxatilis** Dixon, 1964)
unctus Cope, 1864
ventralis O'Shaughnessy, 1875 (*mulleri* Parker,
 1935; *underwoodi* Dixon, 1962)
wirshingi Kerster and H. M. Smith, 1955
 (*hispaniolae** Schwartz, 1979; *sommeri**
 Schwartz, 1979)
xanti Cope, 1864 (*mentalis* Werner, 1910;
*nocticolus** Dixon, 1964; *zweifeli** Dixon,
 1964; *acorius** Dixon, 1966; *angulus** Dixon,
 1966; *circus** Dixon, 1966; *estabanensis**
 Dixon, 1966; *coronatus** Dixon, 1966; *sloani*
 Bostic, 1971)

Phyllopezus W. Peters, 1877
periosus Rodrigues, 1986
pollicaris Spix, 1825 (*goyazensis* W. Peters, 1877;
*przewalskii** Koslowsky, 1894; *spixii*
 Schlegel, 1933)

Phyllurus Goldfuss, 1820 (*Geckoides* Peron, 1807;
Phyllurus [n.n.] Oken, 1817; *Phyllura* Kaup,
 1827; *Anomalurus* Fitzinger, 1843)
caudiannulatus Covacevich, 1975
cornutus Ogilby, 1892 (*lichenosus* Günther, 1897;
swaini Wells and Wellington, 1985)
platurus Shaw, 1790 (*phyllurus* Schneider, 1797;
discosura Merrem, 1820; *platyura* Merrem,
 1820; *novae-hollandiae* Cuvier, 1822; *whitii*
 Gray, 1825; *cuvieri* Bory de Saint-Vincent,
 1828; *platycaudus* Schinz, 1835; *australis*
 Swainson, 1839; *inermis* Gray, 1845)

salebrosus Covacevich, 1975

Pletholax Cope, 1864

gracilis Cope, 1864 (*edelensis** Storr, 1978)

Pristurus Rüppell, 1835 (*Spatalura* Gray, 1863;

Geisopristurus Lanza and Sassi, 1968)

abdelkuri Arnold, 1986

carteri Gray, 1863 (*tuberculatus** Parker, 1931)

celerrimus Arnold, 1977

collaris Steindachner, 1867

crucifer Valenciennes, 1861 (*longipes* W. Peters,

1871; *stefaninii* Calabresi, 1927;

laticephalus Scortecci, 1933)

flavipunctatus Rüppell, 1835 (*percristatus*

Boulenger, 1896; *pseudoflavipunctatus*

Scortecci, 1933)

gasperetti Arnold, 1986 (*gallagheri** Arnold, 1986)

guichardi Arnold, 1986

insignis Blanford, 1881

insignoides Arnold, 1986

minimus Arnold, 1977

ornithocephalus Arnold, 1986

phillipsii Boulenger, 1895 (*somalicus* Parker,

1932)

popovi Arnold, 1982

rupestris Blanford, 1874 (*migiurtinicus* Scortecci,

1933; *guweirensis** Haas, 1951; *iranicus** K.

P. Schmidt, 1952)

saada Arnold, 1986

simonettai Lanza and Sassi, 1968

sokotranus Parker, 1938

Pseudogekko Taylor, 1922

brevipes Boettger, 1897

compressicarpus Taylor, 1915

labialis W. Peters, 1867

smaragdinus Taylor, 1922

Pseudogonatodes Ruthven, 1915

barbouri Noble, 1921

furvus Ruthven, 1915

guianensis Parker, 1935 (*amazonicus** Vanzolini,

1967)

lunulatus Roux, 1927

peruvianus Huey and Dixon, 1970

Ptenopus Gray, 1865

carpi Brain, 1962

garrulus A. Smith, 1849 (*maculatus** Gray, 1865)

kochi Haacke, 1964

- Ptychozoon* Kuhl, 1822 (*Ptyxozoon* Kuhl and Hasselt, 1822; *Pteropleura* Gray, 1827)
horsfieldii Gray, 1827
intermedium Taylor, 1915
kuhli Stejneger, 1902 (*homalocephala* Creveldt, 1809)
lionotum Annandale, 1905
rhacophorus Boulenger, 1899
- Ptyodactylus* Goldfuss, 1820
guttatus Heyden, 1827 (*maculatus* Schinz, 1833; *lacazzi* Boutan, 1892; *siphonorhina* J. Anderson, 1896)
hasselquistii Donndorff, 1798 (*ascalabotes* Merrem, 1820; *lobatus* Geoffroy, 1823; *ragazzi** J. Anderson, 1898; *togoensis* Tornier, 1901)
homolepis Blanford, 1876 (*socotranus** Steindachner, 1902)
oudrii Lataste, 1880
puiseuxi Bouton, 1893 (*bischoffsheimi* Boutan, 1893; *montmahoui* Bouton, 1893; *barroisi* Bouton, 1893; *syriacus* Peracca, 1894; *sancti-montis* Barbour, 1914)
- Pygopus* Merrem, 1820 (*Hysteropus* A. Duméril, in Duméril and Bibron, 1839; *Cryptodelma* Fischer, 1882)
lepidopodus Lacepede, 1804 (*novaehollandiae* Oppel, 1811; *lepidopus* Merrem, 1820; *squamiceps* Gray, 1845; *longicaudatus* Tepper, 1882)
nigriceps Fischer, 1882 (*baileyi* Günther, 1897; *schraderi** Boulenger, 1913; *klugei* Wells and Wellington, 1985; *territorianus* [n.n.] Wells and Wellington, 1985)
- Quedenfeldtia* Boettger, 1883
trachyblepharus Boettger, 1874 (*moerens* Chabanaud, 1916)
- Rhacodactylus* Fitzinger, 1843 (*Correlophus* Guichenot, 1866; *Ceratolophus* Barboza du Bocage, 1873; *Chamaeleonurus* Boulenger, 1878; *Chamaeleonurus* Carus, 1889; *Pseudothecadactylus* Brongersma, 1936; *Torresia* Brongersma, 1934)
auriculatus Bavay, 1869 (*hexaceros* Barboza du Bocage, 1873)
australis Günther, 1877
cavaticus Cogger, 1975
chahoua Bavay, 1869
ciliatus Guichenot, 1866
leachianus Cuvier, 1829 (*aubryanus* Barboza du Bocage, 1873)
lindneri Cogger, 1975

- sarasinorum* Roux, 1913
trachyrhynchus Barboza du Bocage, 1873
 (*trachycephalus* Boulenger, 1878)
- Rhoptropus* W. Peters, 1869 (*Dactychylikion* Thominot, 1878)
afer W. Peters, 1869
barnardi Hewitt, 1926
biporosus FitzSimons, 1957
boultoni K. P. Schmidt, 1933 (*benguellensis**
 Mertens, 1938; *montanus** Laurent, 1964)
braconnieri Thominot, 1878
bradfieldi Hewitt, 1935 (*diporus** Haacke, 1965)
taeniostictus Laurent, 1964
- Rhynchoedura* Günther, 1867
ornata Günther, 1867 (*paraornatus* Wells and
 Wellington, 1984; *ormsbyi* Wells and
 Wellington, 1985)
- Saurodactylus* Fitzinger, 1843
fasciatus Werner, 1931
mauritanicus Duméril and Bibron, 1836 (*desertorum*
 Fitzinger, 1843; *brosseti** Bons and Pasteur,
 1957)
- Sphaerodactylus* Wagler, 1830 (*Sphaeriodactylus* Gray, 1831)
alayi Grant, 1959
altavelensis Noble and Hassler, 1933
 (*brevirostratus** Shreve, 1968;
*enriguilloensis** Shreve, 1968; *lucioi* Thomas
 and Schwartz, 1983)
argivus Garman, 1888 (*bartschi** Cochran, 1934;
*lewisi** Grant, 1941)
argus Gosse, 1850 (*henriquesi* Grant, 1940;
*andresensis** Dunn and Saxe, 1950)
armasi Schwartz and Garrido, 1974
armstrongi Noble and Hassler, 1933 (*hypsinephes**
 Thomas and Schwartz, 1983)
asterulus Schwartz and Graham, 1980
beattyi Grant, 1937 (*seamani** Thomas and Schwartz,
 1966)
becki K. P. Schmidt, 1919
bromeliarum G. Peters and Schwartz, 1972
caicosensis Cochran, 1934
callocricus Schwartz, 1976
cinereus Wagler, 1830
clenchi Shreve, 1968 (*apocoptus** Schwartz, 1983)
cochranae Ruibal, 1946
copei Steindachner, 1867 (*anthracinus* Cope, 1862;
*picturatus** Garman, 1887; *asper* Garman, 1888;
*cataplexis** Schwartz and Thomas, 1965;
*enochrus** Schwartz and Thomas, 1965;

- polyommatus** Thomas, 1968; *astreptus**
 Schwartz, 1975; *pelates** Schwartz, 1975;
*websteri** Schwartz, 1975; *deuterus** Schwartz,
 1975)
corticola Garman, 1888 (*aporrox** Schwartz, 1968;
*campter** Schwartz, 1968; *soter** Schwartz,
 1968)
cryphius Thomas and Schwartz, 1977
darlingtoni Shreve, 1968 (*noblei** Shreve, 1968;
*bobilini** Thomas and Schwartz, 1983;
*mekistus** Thomas and Schwartz, 1983)
difficilis Barbour, 1914 (*lycauges** Schwartz,
 1983; *euopter** Schwartz, 1983; *typhlops**
 Schwartz, 1983; *peratus** Schwartz, 1983;
*diolenius** Schwartz, 1983; *anthracomus**
 Schwartz, 1983)
dunni K. P. Schmidt, 1936
elasmorhynchus Thomas, 1966
elegans Macleay, 1834 (*punctatissimus* Duméril and
 Bibron, 1836; *alopex* Cope, 1862)
elegantulus Barbour, 1917
fantasticus Duméril and Bibron, 1836
 (*ligniservulus** King, 1962; *anidrotus**
 Thomas, 1965; *fuga** Thomas, 1965; *hippomanes**
 Thomas, 1965; *karukera** Thomas, 1965;
*orescius** Thomas, 1965; *phyzacinus** Thomas,
 1965; *tartaropylorus** Thomas, 1965)
gaigeae Grant, 1932
gilvitorques Cope, 1862
glaucus Cope, 1865 (*inornatus* W. Peters, 1873;
torquatus Strauch, 1887)
goniorhynchus Cope, 1895
graptolaemus Harris and Kluge, 1983
heliconiae Harris, 1982
homolepis Cope, 1886 (*carinatus* Andersson, 1916;
imbricatus Andersson, 1916; *mertensi* Wermuth,
 1965)
inaguae Noble and Klingel, 1932
intermedius Barbour and Ramsden, 1919 (*drapetiscus*
 Schwartz, 1958)
klauberi Grant, 1931
ladae Thomas and Hedges, 1988
lazelli Shreve, 1968
leucaster Schwartz, 1973
levinsi Heatwole, 1968
lineolatus Lichtenstein and von Martens, 1856
 (*casicolus* Cope, 1862)
macrolepis Günther, 1859 (*grandisquamis* Stejneger,
 1904; *danforthi* Grant, 1931; *parvus** King,
 1962; *ateles** Thomas and Schwartz, 1966;
*guarionex** Thomas and Schwartz, 1966; *inigoii**
 Thomas and Schwartz, 1966; *mimetes** Thomas

and Schwartz, 1966; *phoberus** Thomas and Schwartz, 1966; *spanius** Thomas and Schwartz, 1966; *stibarus** Thomas and Schwartz, 1966)
mariguanae Cochran, 1934
microlepis Reinhardt and Lütken, 1863
 (*melanospilos* Bocourt, 1873; *thomasi** Schwartz, 1965)
micropithecus Schwartz, 1977
millepunctatus Hallowell, 1861 (*continentalis* Werner, 1896)
molei Boettger, 1894 (*buergeri* Werner, 1900; *venezuelanus* Roux, 1927; *boettgeri* [n.n.] Donoso Barros, 1968)
monensis Meerwarth, 1901
nicholsi Grant, 1931
nigropunctatus Gray, 1845 (*decoratus** Garman, 1888; *flavicauda** Barbour, 1904; *gibbus** Barbour, 1921; *atessares* Thomas and Schwartz, 1966; *granti** Thomas and Schwartz, 1966; *lissodesmus** Thomas and Schwartz, 1966; *strategus** Thomas and Schwartz, 1966; *porrasi** Schwartz, 1972)
notatus Baird, 1858 (*exsul** Barbour, 1914; *amaurus** Schwartz, 1966; *atactus** Schwartz, 1966; *peltastes** Schwartz, 1966)
nycteropus Thomas and Schwartz, 1977
ocoae Schwartz and Thomas, 1977
oliveri Grant, 1944 (*storeyae** Grant, 1944)
omoglaux Thomas, 1982
oxyrhinus Gosse, 1850 (*dacnicolor** Barbour, 1910)
pacificus Stejneger, 1903
parkeri Grant, 1939
parthenopion Thomas, 1965
perissodactylus Thomas and Hedges, 1988
ramsdeni Ruibal, 1959
randi Shreve, 1968 (*methorius** Schwartz, 1977; *strahmi** Schwartz, 1977)
rhabdotus Schwartz, 1970
richardsoni Gray, 1845 (*gossei** Grant, 1939)
roosevelti Grant, 1931
rosaurae Parker, 1940
ruibali Grant, 1959
sabanus Cochran, 1938
samanensis Cochran, 1932
savagei Shreve, 1968 (*juanilloensis** Shreve, 1968)
scaber Barbour and Ramsden, 1919
scapularis Boulenger, 1902
semasiops Thomas, 1975
shrevei Lazell, 1961
sommeri Graham, 1981
sputator Sparrman, 1784 (*pictus* Garman, 1888)
streptophorus Thomas and Schwartz, 1977

(*sphenophanes** Thomas and Schwartz, 1983)
stejnegeri Cochran, 1931
thompsoni Schwartz and Franz, 1976
torrei Barbour, 1914 (*spielmani** Grant, 1958;
*ocujal** Thomas and Schwartz, 1966)
townsendi Grant, 1931
underwoodi Schwartz, 1968
vincenti Boulenger, 1891 (*festus** Barbour, 1915;
*monilifer** Barbour, 1921; *adamus** Schwartz,
 1965; *diamesus** Schwartz, 1965; *josephinae**
 Schwartz, 1965; *pheristus** Schwartz, 1965;
*psammius** Schwartz, 1965; *ronaldi** Schwartz,
 1965)
williamsi Thomas and Schwartz, 1983
zygaena Schwartz and Thomas, 1977

Stenodactylus Fitzinger, 1826 (*Goniodactylus* Schlegel,
 1826; *Tolarenta* Gray, 1842; *Ceramodactylus*
 Blanford, 1872; *Pseudoceramodactylus* Haas,
 1957; *Trigonodactylus* Haas, 1957; *Garzoniella*
 Perret, 1976)
affinis Murray, 1884
arabicus Haas, 1957
doriae Blanford, 1874 (*major* Parker, 1930)
grandiceps Haas, 1952
khobarensis Haas, 1957
leptocosymbotes Leviton and S. Anderson, 1967
petrii J. Anderson, 1896 (*stenurus* Werner, 1899;
elimensis Barbour, 1914)
pulcher J. Anderson, 1896
slevini Haas, 1957 (*arabicus* Haas, 1957; *hassi*
 Kluge, 1967)
sthenodactylus Lichtenstein, 1823 (*elegans*
 Fitzinger, 1826; *savignyi* Audouin, 1827;
guttatus Cuvier, 1829; *wilkinsonii* Gray,
 1842; *mauritanicus* Guichenot, 1850; *hirouxii*
 Doumergue, 1899; *longipes* Perret, 1976)
yemenensis Arnold, 1980

Strophurus Fitzinger, 1843 (*Strophura* Gray, 1845;
Oedurella Lönnberg and Andersson, 1913;
Eremiastrophurus Wells and Wellington, 1985)
assimilis Storr, 1988
ciliaris Boulenger, 1885 (*aberrans** Glauert, 1952)
elderi Stirling and Zietz, 1893 (*mahoodi* Wells and
 Wellington, 1985)
intermedius Ogilby, 1892
jeanae Storr, 1988
mcmillani Storr, 1978
michaelseni Werner, 1910
rankini Storr, 1979
spinigerus Gray, 1842 (*inornatus** Storr, 1988)

strophurus Duméril and Bibron, 1836 (*dumerilii* Fitzinger, 1843)
taeniatus Lönnberg and Andersson, 1913
wellingtonae Storr, 1988
williamsi Kluge, 1963
wilsoni Storr, 1983

Tarentola Gray, 1825 (*Makariogecko* Jogger, 1984; *Sahelogecko* Jogger, 1984; *Saharogecko* Jogger, 1984; *Neotarentola* Jogger 1984)
americana Gray, 1831 (*milberti* Duméril and Bibron, 1836; *cubana* Gundlach and W. Peters, 1865; *warreni** Schwartz, 1968)
angustimentalis Steindachner, 1891
annularis I. Geoffroy, 1827 (*savignyi* Audouin, 1827; *aegyptiacus* Cuvier, 1829; *senegalensis* Boulenger, 1885; *quadraticauda* Toriner, 1905; *relicta** Jogger, 1984)
boehmei Jogger, 1984
boettgeri Steindachner, 1891 (*hierrensis** Jogger and Bischoff, 1983; *bischoffi** Jogger, 1984)
borneensis Gray, 1845 (*gigas** Barboza du Bocage, 1875; *rudis** Boulenger, 1906; *protogigas** Jogger, 1984; *maioensis** Schleich, 1984; *brancoensis* Schleich, 1984)
caboverdianus Schleich, 1984 (*substituta** Schleich, 1984; *antoensis* Jogger, 1984; *razianus** Schleich, 1984; *nicolauensis** Schleich, 1984)
clypeata [?] Gray, 1842
darwinii Jogger, 1984
delalandii Duméril and Bibron, 1836
deserti Boulenger, 1891
ephippiata O'Shaughnessy, 1875 (*hoggarensis** Werner, 1937; *panousei* Pasteur, 1959; *senegambiae** Jogger, 1984; *nikolausi** Jogger, 1984)
gomerensis Jogger and Bischoff, 1983
mauritanica Linnaeus, 1758 (*muricatus* Laurenti, 1768; *fascicularis** Daudin, 1802; *stellio* Merrem, 1820; *muralis* Duméril and Bibron, 1836; *facetanus* Strauch, 1862; *atlantica* Doumergue, 1899; *gracilis* Doumergue, 1899; *lissoide* Doumergue, 1899; *saharae* Doumergue, 1899; *tuberculata* Rosen, 1905; *juliae** Jogger, 1984)
neglecta Strauch, 1887 (*angusticeps* Strauch, 1887; *greyi** Jogger, 1984)
parvicarinata Jogger, 1980

Tenuidactylus Shcherbak and Golubev, 1984
caspius Eichwald, 1831 (*fasciatus* Menetries, 1832;

insularis Akhmedow and Shcherbak, 1978)
fedtschenkoi Strauch, 1887
longipes Nikolsky, 1896 (*microlepis** Lantz, 1918)
turcmenicus Shcherbak, 1978
voraginosus Leviton and S. Anderson, 1984

Teratolepis Günther, 1870
albofasciatus Grandison and Soman, 1963
fasciata Blyth, 1853

Teratoscincus Strauch, 1863
bedriagai Nikolsky, 1899
microlepis Nikolsky, 1899
przewalskii Strauch, 1887
scincus Schlegel, 1858 (*keyserlingii** Strauch, 1863; *zarudnyi* Nikolsky, 1896; *roborowskii* Bedriaga, 1905; *rustamowi** Shcherbak, 1979)

Thecadactylus Oken, 1817 (*Thecodactylus* Wagler, 1830)
rapicauda Houttuyn, 1782 (*perfoliatus* Schneider, 1793; *laevis* Daudin, 1802; *surinamensis* Daudin, 1802; *theconyx* Duméril and Bibron, 1836; *tristis* Hallowell, 1854)

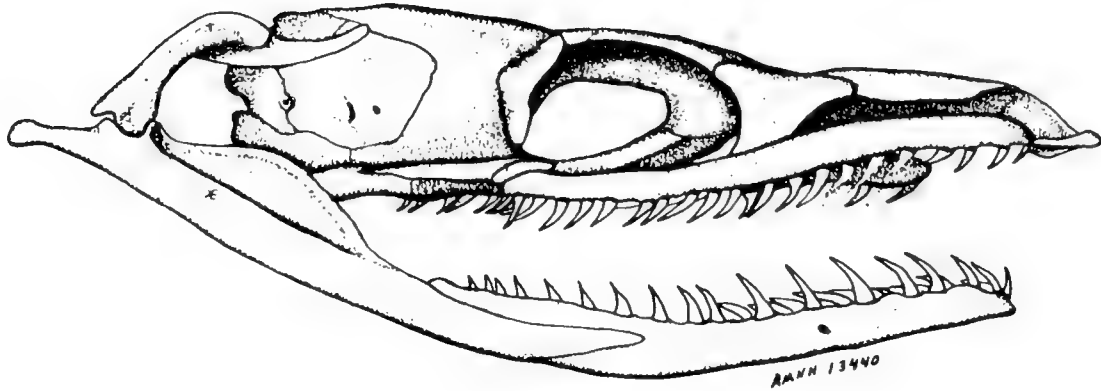
Tropiocolotes W. Peters, 1880
persicus Nikolsky, 1903
scorteccii Cherchi and Spano, 1963
steudneri W. Peters, 1869 (*petersii* Boulenger, 1891; *natterei* Steindachner, 1900)
tripolitanus W. Peters, 1880 (*occidentalis** Parker, 1942; *somalicus** Parker, 1942; *algericus** Loveridge, 1947; *apoklomax* Papenfuss, 1970)

Urocotyledon Kluge, 1983
inexpectata Stejneger, 1893
palmata Mocquard, 1902
weileri L. Müller, 1909
wolterstorffi Tornier, 1900

Uroplatus A. Duméril, 1805 [1806] (*Uroplates* Gray, 1825; *Rhacoessa* Wagler, 1830; *Oiacurus* Fitzinger, 1843; *Lonchurus* Fitzinger, 1843)
alluaudi Mocquard, 1894
ebenau Boettger, 1879 (*boettgeri* Fischer, 1883 [1884]; *phantasticus* Boulenger, 1888; *schneideri* Lamberton, 1913)
fimbriatus Schneider, 1797 (*hypoxantha* Wagler, 1833)
guentheri Mocquard, 1908
lineatus Duméril and Bibron, 1836
sikorae Boettger, 1913

40
666
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(SERPENTES: COLUBRIDAE)



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INTRODUCTION

Compiling a bibliography which spans over two and a half centuries and includes scholarly works from a score of countries is both time-consuming and rewarding. I began accumulating references on neotropical vine snakes of the genus Oxybelis in 1965 when I was a student at Louisiana State University. My initial reference was Bogert and Oliver's 1945: "A Preliminary Analysis of the Herpetofauna of Sonora."

Five species make up the genus Oxybelis Wagler. These are: O. aeneus, O. argenteus, O. boulengeri, O. brevirostris, and O. fulgidus. The species of the genus are distributed from the extreme southwestern United States (Arizona), south through tropical Mexico and Central America, to Brazil, Bolivia, and Peru in South America. The southern limits of the range are poorly known.

This bibliography lists known references to the genus, to recognized species of the genus, and to all junior synonyms. Publications mentioning species erroneously placed within Oxybelis and now assigned elsewhere are likewise cited. A few titles which mention no specific Oxybelis taxa but which are considered particularly relevant to their taxonomy are also included. In this latter category are several papers which have been used in the literature to establish priority dates for determining sequences of synonyms of Oxybelis taxa.

Although some subjective interpretations are necessary, Seba (1735) and Schuechzer (1735a and/or 1735b) may be the earliest references to these snakes of which I am aware. By the mid-1800's, a considerable literature existed. Brief commentaries on distribution, locality records, or anatomy made up the bulk of this. Papers on snakes of the genus Oxybelis have increased in recent decades and considerably more detailed studies have occasionally appeared.

Although more recent titles have been added, intensive literature searching ceased in November of 1989. Omissions from this list may be due to my having overlooked citations or not having had the opportunity to verify sources in time for inclusion herein. The bibliography is reasonably comprehensive and it should prove a useful tool for anyone working with these snakes.

Over the years, many persons have assisted me in searching the literature, in translating non-English papers, and in interpreting older literature. Others have helped by calling my attention to papers in less prominent journals. To list their names at this time would undoubtedly result in unintended omissions of oversight. Nevertheless, I extend my thanks to all who have been of assistance. I am especially grateful to two persons who were particularly helpful during the final stages of

preparing this manuscript. Marinus S. Hoogmoed provided invaluable comments on plates in the Schuechzer papers (1735a, 1735b, 1738), which had proven particularly difficult to interpret. Kenneth L. Williams supplied a number of references I had not previously encountered and also spent considerable time and effort copying and/or verifying reference sources for me.

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TYPES AND HISTORICALLY

IMPORTANT SPECIMENS OF RATTLESNAKES
IN THE
MUSÉUM NATIONAL D'HISTOIRE NATURELLE (PARIS)

MICHEL THIREAU

Laboratoire des Reptiles et Amphibiens
Muséum National d'Histoire Naturelle

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The cover illustration is from Lacepede 1788:pl. 18.

INTRODUCTION

According to the classification of Hoge and Romano-Hoge (1981), rattlesnakes, "serpents à sonnettes," are members of the Viperidae Laurenti 1768, which consists of four subfamilies: Viperinae Laurenti 1768, Azemiopinae Liem, Marx and Rabb 1971, Causinae Cope 1860, and Crotalinae Oppel 1871. The Crotalinae is further subdivided into the tribes Agkistrodontini Gray 1825 and Crotalini Gray 1825. One group of Crotalini contains two genera: Sistrurus Garman 1883 and Crotalus Linné 1758; the former represents the primitive rattlesnakes, the latter the derived ones. The evolution of the rattlesnakes has been discussed in several important monographs (e.g., Gloyd, 1940, Klauber, 1956 & 1972, Brattstrom, 1964).

The purpose of this report is to summarize the information uncovered during my search for lost types and taxonomically-nomenclaturally important specimens. The information is presented in two parts. The first part examines the type specimens of nominal species, whether or not the specimen's presence has been confirmed in our collections. The second part examines historical specimens, i.e., those specimens used by A. M. C. Duméril, A. H. A. Duméril, and G. Bibron in their redescription of species named by their predecessors. The types represent eight binominals and one trinomial; the historical specimens were used in eight binominals. In each case, the species are arranged alphabetically. All the specimens discussed herein are stored in alcohol and are housed in the Museum national d'Histoire naturelle, Paris (MNHN).

The information for each species is arranged in the following manner: 1) the author, date of publication, reference (abbreviated) with page of original description and identification of any associated illustrations, occasionally outside text or as a painting on vellum; 2) the current status of the taxon; 3) the catalogue number in the registry of the Laboratoire de Zoologie (Reptiles et Amphibiens); 4) the status of the type; 5) the original citation of the type locality and subsequent restrictions; 6) the name of the collector and/or donor, sometimes with date(s) of collection/donation; 7) the length of the body (TL, from tip of snout to base of the rattle); 8) supplementary comments.

Finally, as much as possible, my purpose is to trace lost types as well as the historically important specimens.

TYPES

Crotalus atricaudatus Latreille In Sonnini et Latreille, 1801, Hist. nat. des Rept., 3:209, 4:323, 402.
=Crotalus horridus Linnaeus, 1758; fide Collins & Knight, 1980.

No types specimens were located. Type locality, "Caroline"; restricted to "Charleston, South Carolina" by Schmidt (1953:228). Bosc was the collector of the type series. He proposed the name atricaudatus in a manuscript used by Latreille. Most likely, no specimens were sent to Paris (cf. also Daudin, 1803, 5:316 n.i.p.).

Crotalus boiquira Lacepède, 1789, Hist. Nat. Quadr. ovipares et des Serpens. In 4^o, 2:130, 390, pl.18, fig. 1. In 12^o, 4:6, 142, 209, 356, pl.7, fig. 1.
=Crotalus horridus Linnaeus, 1758, fide Collins & Knight, 1980.

No types specimens were located. One specimen was deposited in the Royal Cabinet. Type locality, "Nouveau Monde," "depuis la terre de Magellan jusqu'au lac Champlain, vers le quarante-cinquième degré de latitude septentrionale," near "de la grande chute d'eau du Niagara," "Caroline." TL 132 cm (this is 4 feet, 10 lines and 4 inches, according to Lacepède (12^o, 4:142)).

Crotalus durissus var. melanurus Garman, 1883, Mem. Mus. Comp. Zool. 3(3):175.
=Crotalus horridus Linnaeus, 1758; fide Collins & Knight, 1980.

Holotype, MNHN 821. Type locality, "Caroline du Sud." Collector and donor unknown. TL 91.5 cm. The synonymy derives from Pisani et al. (1973). Crotalus durissus var. melanurus Jan, 1859:153 is a nomen nudum, which has no official status according to the International Code of Zoological Nomenclature (1985:285-286). This holotype was received by the Paris museum in exchange from Milan (cf. e.g., Duméril, A.H.A., In Jan, 1858:438-439; Jan, 1858:515; Duméril, A.H.A., 1865:47-50).

Crotalus immaculatus Latreille In Sonnini et Latreille, 1801, Hist. nat. des Rept., 3:201, 4:323, 402.
=Crotalus durissus terrificus (Laurenti, 1768); fide Klauber, 1972.

No type specimens were located. A specimen was maintained in the galleries of the Muséum d'Histoire naturelle. Daudin

(1803, 5:319) recorded its length as approximately two French ft, thus TL about 65 cm. Latreille incorrectly indicated that the type series derived from "Indes orientales," in his reference to Séba (1735, 2: pl. 95, no. 3 "Vipera Caudisona Ceilanica. Autre vipère à queue sonnante, de Ceylon, 100" and pl. 96, no. 1. "Vipera Orientalis, maxima, Caudisona, Foemina. Vipère à queue sonnante, Orientale, femelle, très grande, 101").

Crotalus jimenezii Dugès, 1877, *Naturelleza*, 4:23, pl. h.t. 1, fig. 18-20.
 =Crotalus polystictus (Cope, 1865); fide Hoge and Romano-Hoge, 1981.

Syntype?, MNHN 1883.284. Catalogue locality, "Guanajuato, 200 km à l'Est de Guadalajara [Mexico]." Donor, Dugès (14 December 1888). TL 89.5 cm.

Syntype?, MNHN 1883.288. Catalogue locality, "Guanajuato, 200 km à l'Est de Guadalajara [Mexico]." Donor, Dugès (12 May 1883). TL 32 cm.

The date of publication for this new taxon was determined by Smith (1942:96). The type localities are "Silao [Guanajuato], Colima et Guadalajara," all three in Mexico. Smith and Taylor (1950:334) restricted the type locality to Guadalajara, Jalisco. Smith and Necker (1943:179) stated that Dugès sent ". . . specimens to many specialists in Europe and the United States to aid in the description and elucidation of the Mexican fauna." Smith and Necker (1943:180) reported "Cotypes of . . . Crotalus jimenezii are present in United States Collection, although lacking in the Dugès Muséum" and on a following page (1943:187) "A specimen in the U.S. National Museum (No 46508, from Guanajuato), is said to be a 'duplicate type' of jimenezii. It is a small specimen, presented to Nelson and Goldman by Dugès in 1896. Two other specimens in the Nat. Mus. presented by Dugès (Nos 24448, 26152), may be cotypes. The latter also are from 'Guanajuato.'" Cochran (1961:170) did not mention these latter two specimens, nor did McCranie (1976:180.1).

Crotalus lugubris Jan, 1859, *Plan d'une Iconographie descriptive des Ophidiens*, *Rev. Mag. Zool.*, 22ème sér., (11):153, 156, pl. E. (4ème fig.); off-print:28, 31, pl. E (4ème fig.).
 =Crotalus triseriatus triseriatus (Wagler, 1830); fide Hoge and Romano-Hoge, 1981.

Paralectotype, MNHN 7754. Type locality, "Mexique." Collector and donor unknown. TL 50 cm. The type specimen in the Paris museum was described specifically by Jan in his

table of quantitative data. The heterogeneity of the type series caused Klauber (1940:17) to designate a lectotype of C. lugubris, and he selected No. 1414 deposited in the Milan museum and described by Jan in his table (second column). Klauber later (1972:48) indicated that this specimen (originally from Mexico) had been destroyed.

Crotalus rhombifer Latreille, In Sonnini et Latreille, 1801, Hist. nat. des Rept. 3:197, 4:322, 402.
=Crotalus adamanteus Palisot de Beauvois, 1799; fide McCranie, 1980.

No type specimens were located. Type locality, "Amérique"; restricted to Gainesville, Florida [USA] by Schmidt (1953:230). Specimen(s) were supposedly present in the "Collection des serpens du Muséum d'Histoire naturelle de Paris" (fide Latreille).

Crotalus simus Latreille, In Sonnini et Latreille, 1801, Hist. nat. des Rept. 3:202, 4:323, 402.
=Crotalus durissus durissus (Linnaeus, 1758); fide Hoge and Romano-Hoge, 1981.

No type specimens were located. Type locality, "Ceylan". According to Latreille, "Un individu de cette espèce [est] conservé au Muséum national." According to Daudin (1803, 5:321), the type specimen of Paris was part of the Séba collection.

Crotalus strepitans Daudin, 1803, Hist. Nat. Gén. et Part. des Reptiles 5:318.
=Crotalus durissus terrificus (Laurenti, 1768); fide Klauber, 1972.

No type specimens were found. Type locality, "continent de l'Amérique". TL approx. 65 cm. According to Daudin " . . . l'individu, placé dans la galerie du muséum d'histoire naturelle de Paris, n'a que deux pieds environs de longueur."

HISTORICALLY IMPORTANT SPECIMEN

Crotalus atrox: DB&D, 1854, Erp. Gén. 7(2):1464, 1482.
=Crotalus atrox Baird and Girard, 1853.

MNHN 4034. Locality, Texas, USA. Donated by Trécul (August 1850?). Described also by A.H.A. Duméril (1857:45).

Crotalus confluentes: DB&D, 1854, Erp. Gén. 7(2):1464, 1475. Atlas Erp. Gén. 1854: pl. 84^{bis}, fig. 4. For Crotalus confluentes Say, 1823.

=Crotalus viridis (Rafinesque, 1818).

MNHN 4035. Locality, Texas, USA. Donated by Trécul (August 1850?). TL 89.5 cm.

MNHN 4036. Locality, Texas, USA. Donated by Trécul (August 1850?). TL 113 cm. Illustrated in Atlas.

Crotalus durissus: DB&D, 1854, Erp. Gén. 7(2):1462, 1465. For Crotalus durissus Linnaeus, 1758.

=Crotalus adamanteus Palisot de Beauvois, 1799.

MNHN 7750. Locality, "Amérique septentrionale" = southern USA. Collected and donated by Bosc. TL 93.5 cm.

=Crotalus durissus Linnaeus, 1758.

MNHN 7752. Locality, Mexico. Collected and donated by Bosc. TL 121.5 cm.

Crotalus durissus: DB&D, 1854, Erp. Gén. 7(2):1465. Atlas, Erp. Gén. 1854: pl. 84^{bis}, fig. 1. Duméril, A.H.A., 1854, Arch. Mus. natn. Hist. nat., Paris 7:283. For Crotalus durissus Linnaeus, 1758.

=Crotalus horridus Linnaeus, 1758.

MNHN 883. Locality, "Amérique septentrionale" = southern USA. Donated by Harpert (February 1842). TL 104 cm. Atlas illustration not reported by Collins and Knight, 1980.

Crotalus durissus: DB&D, 1854, Erp. Gén. 7(2):1465, 1466. Duméril, A.H.A., 1854, Arch. Mus. natn. Hist. nat., Paris 7:282. Duméril, A.H.A., 1861, Arch. Mus. natn. Hist. nat., Paris 10:439. For Crotalus durissus Linnaeus, 1758.

=Crotalus horridus Linnaeus, 1758.

MNHN 3660. Locality, New York (center of Milbert's scientific operations, fide Boitard and Janin, 1842:37). Collected and donated by Milbert. TL 143.5 cm. Vél. 88(4)* drawn by Chazal from the living specimen in 1833. At this time, the tail has seven rattle segments, compared to 13 when deposited in the collection. [* A painting on vellum (tome 88, no. 4) held in the central library/MNHN, Paris.]

Crotalus horridus: DB&D, 1854, Erp. Gén. 7(2):1472. For Crotalus horridus Linnaeus, 1758.
= Crotalus durissus Linnaeus, 1758.

MNHN 3211. Locality, Brazil. Donated by Clossen (March 1844). TL 109 cm.

MNHN 4033. Locality, Ste. Marthe, District de Cundimarcha, about 700 km N Bogota, 11°19'N 76°24'W [Colombia]. Donated by son of Fontanier (July 1853). TL 37.5 cm.

Crotalus Kirtlandii: DB&D, 1854, Erp. Gén. 7(2):1482. Duméril, A.H.A., 1857, Cat. Rept. Coll. M.H.N.P. :45. For Crotalus kirtland Holbrook, 1842.
= Sistrurus catenatus (Rafinesque, 1818).

MNHN 890. Locality, Ohio [USA]. Donated by Hallowell (June 1855). TL 48 cm.

Crotalus miliarius: Duméril, A.H.A., 1857, Cat. Rept. Coll. M.H.N.P. :45.
= Sistrurus miliarius (Linnaeus, 1758).

MNHN 886. Locality, "Amérique septentrionale" = southern USA. Donated by Harlan. TL 35.5 cm. Palmer (1978:220.1) summarized only the AngloSaxon literature, except for two of 51 references cited.

Crotalus rhombifer: DB&D, 1854, Erp. Gén. 7(2):1470. Atlas, Erp. Gén. 1854: pl. 84^{bis}, fig. 3. For Crotalus rhombifer Latreille In Sonnini and Latreille, 1801.
= Crotalus adamanteus Palisot de Beauvois, 1799.

MNHN 882. Locality, United States. TL 138 cm. Atlas figure 3 depicts the dorsal pattern of MNHN 882 but with several modifications to the symmetry of the dorsorostral head scalation. McCranie (1980: 252.1) did not report the existence of this illustration.

Crotalus tergeminus: DB&D, 1854, Erp. Gén. 7(2):1479. Atlas, Erp. Gén., 1854: pl. 84^{bis}, fig. 5. For Crotalus tergeminus Say, 1823.
= Sistrurus catenatus (Rafinesque, 1818).

MNHN 887. Locality, Charlestown, north of Louisville, Indiana [USA]. Donated by Holbrook (January 1846). TL 55.5 cm. This is the specimen illustrated in the Atlas. Minton

(1983:332.1) did not report the existence of this illustration.

MNHN 2074. Locality, Charlestown, north of Louisville, Indiana [USA]. Donated by Holbrook (January 1846). TL 25 cm.

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I wish to thank my colleagues, the researchers and technicians in the laboratory, and also Y. Laissus and C. Hustache (Bibliothèque centrale) for their assistance in examining the Museum's rare books. Mr. Polls worked with me on the analysis of Sistrurus miliarius. A. Dubois critically reviewed an earlier draft of this manuscript.



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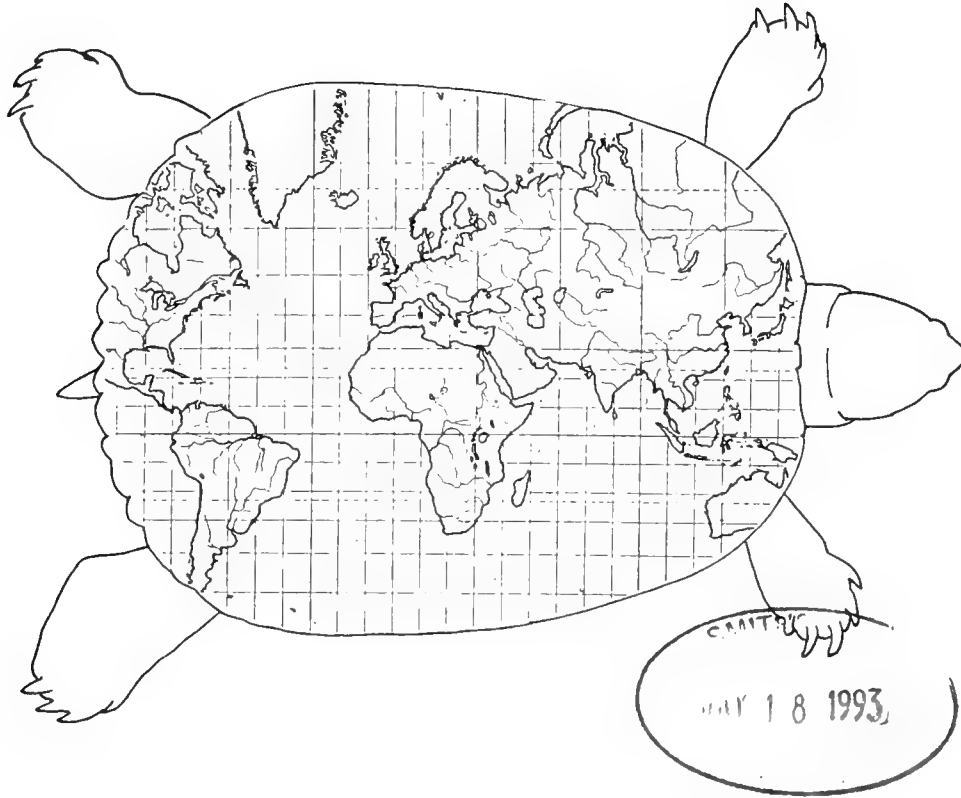
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SPECIES RICHNESS MAPS
OF THE
FRESHWATER AND TERRESTRIAL TURTLES
OF THE WORLD



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INTRODUCTION

With the publication of my turtle checklist (Iverson, 1986) it was finally possible to begin preliminary analyses of the patterns and correlates of species richness in turtles on a global scale. As a first step I prepared species density maps for each turtle family by outlining the range of each turtle species as plotted in Iverson (1986), and manually overlaying those range maps to produce species richness isopleth (= species density) maps. This was undertaken with the clear realization that the taxonomy and distribution of turtles in some areas of the world was still not well studied (especially southeast Asia), but that general patterns of species density would not change radically even with much further study.

The species density maps that were generated revealed a number of interesting zoogeographic patterns, and stimulated a pilot study of the correlates of maximum freshwater and terrestrial turtle species density in 42 river basins across five continents (Iverson, 1991). However, because of journal space limitations, it was not possible to publish the species density maps along with that analysis. Those maps are reproduced here in hope of stimulating further analyses of the distribution and zoogeography of turtles. The maps appear in the same order as the taxa appeared in Iverson (1986).

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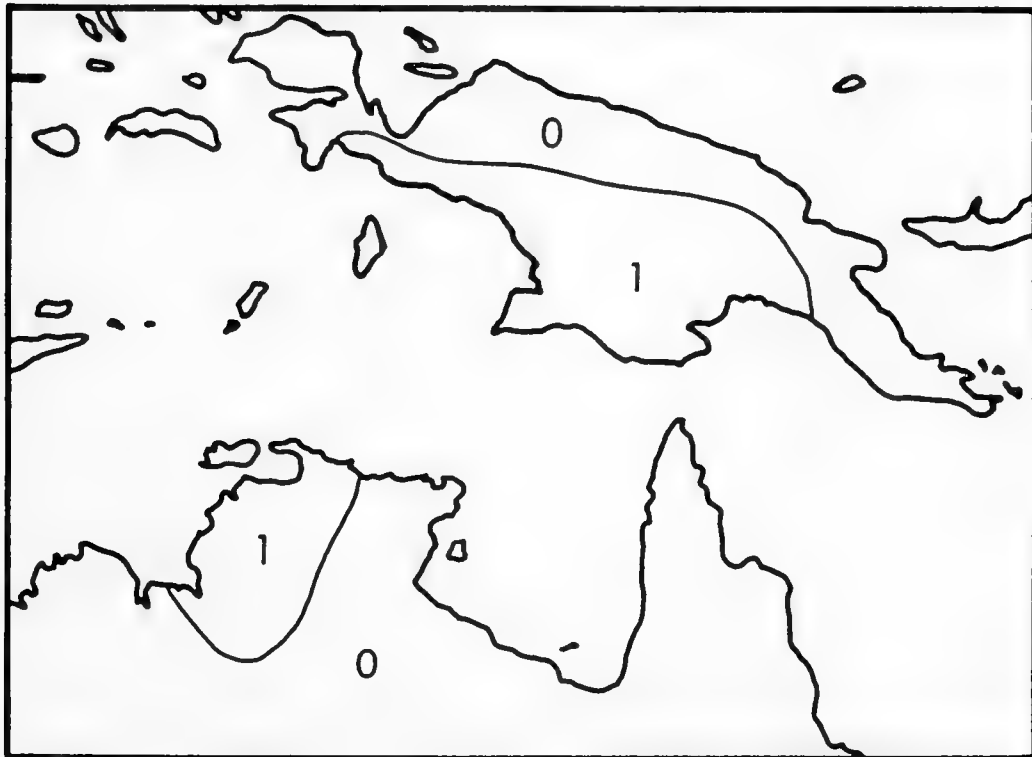


Fig. 1. Species richness of the family Carettochelyidae on New Guinea and Northern Australia.



Fig. 2. Species richness of the family Chelydridae on North and Central America.

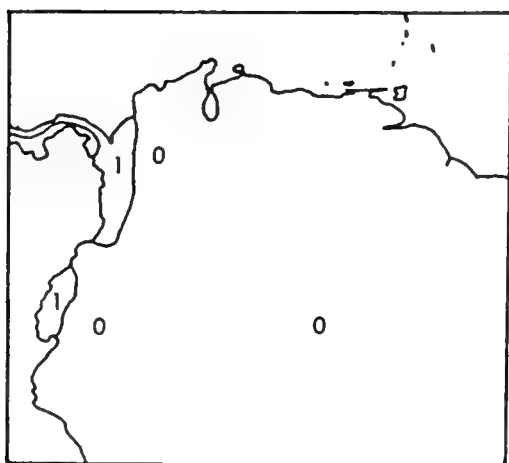


Fig. 3. Species richness of the family Chelydridae on northwestern South America.



Fig. 4. Species richness of the family Dermatemydidae on Mexico and Central America.



Fig. 5. Species richness of the subfamily Batagurinae (family Emydidae) on Asia.

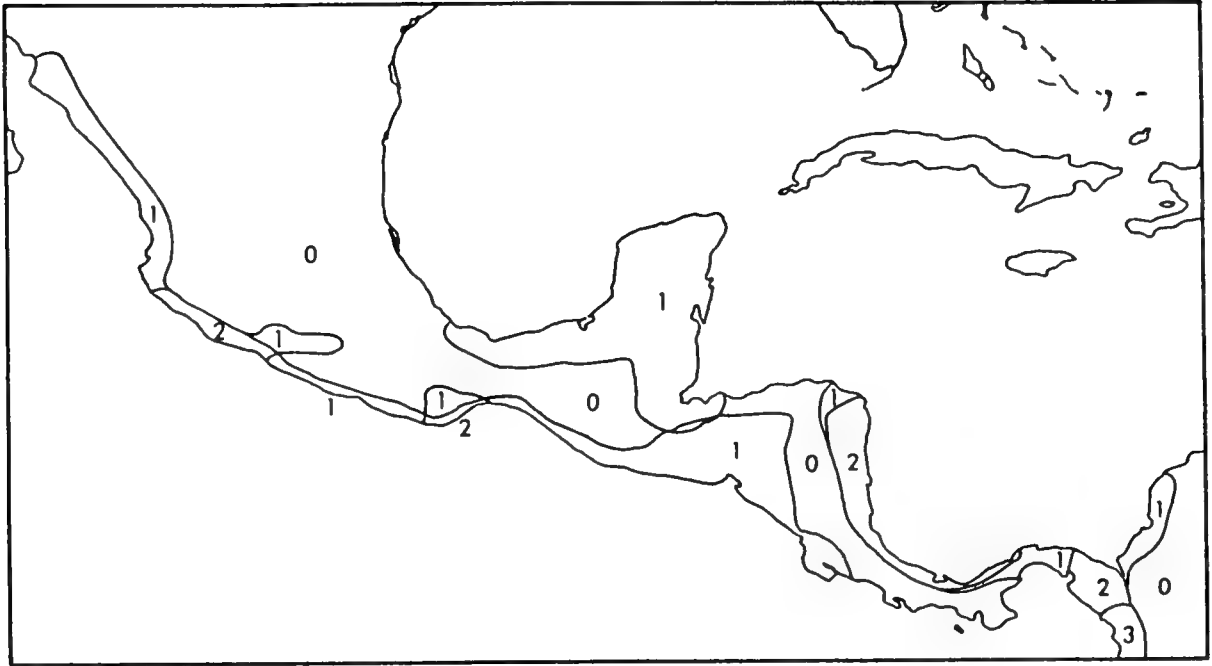


Fig. 6. Species richness of the subfamily Batagurinae (family Emydidae) on Central America.

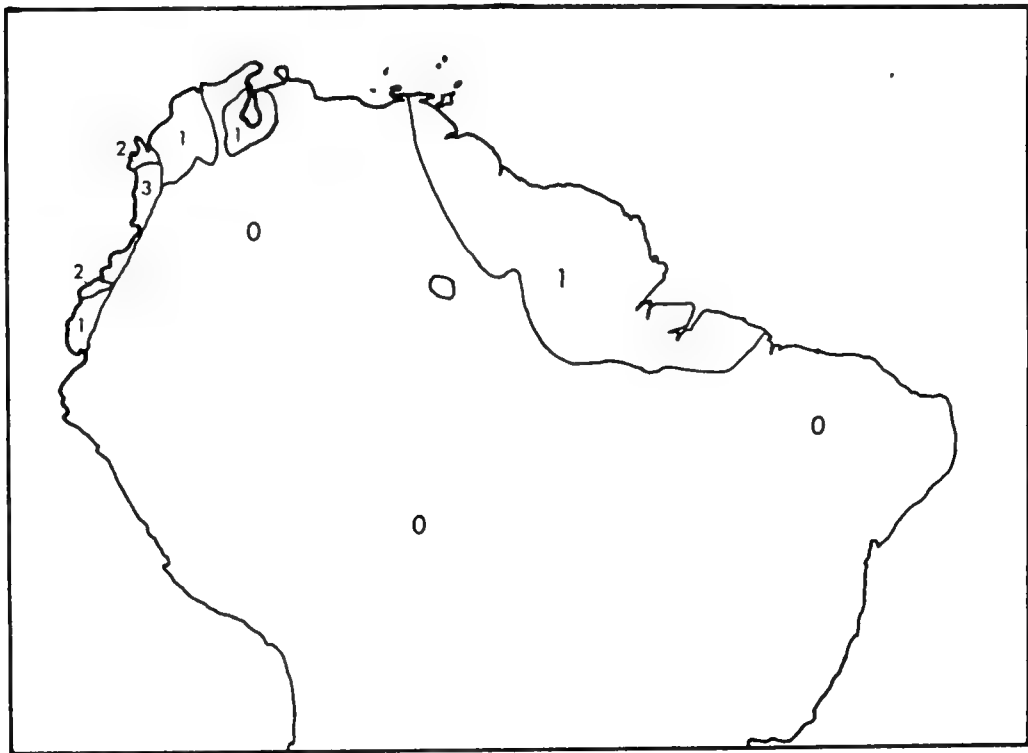


Fig. 7. Species richness of the subfamily Batagurinae (family Emydidae) on South America.

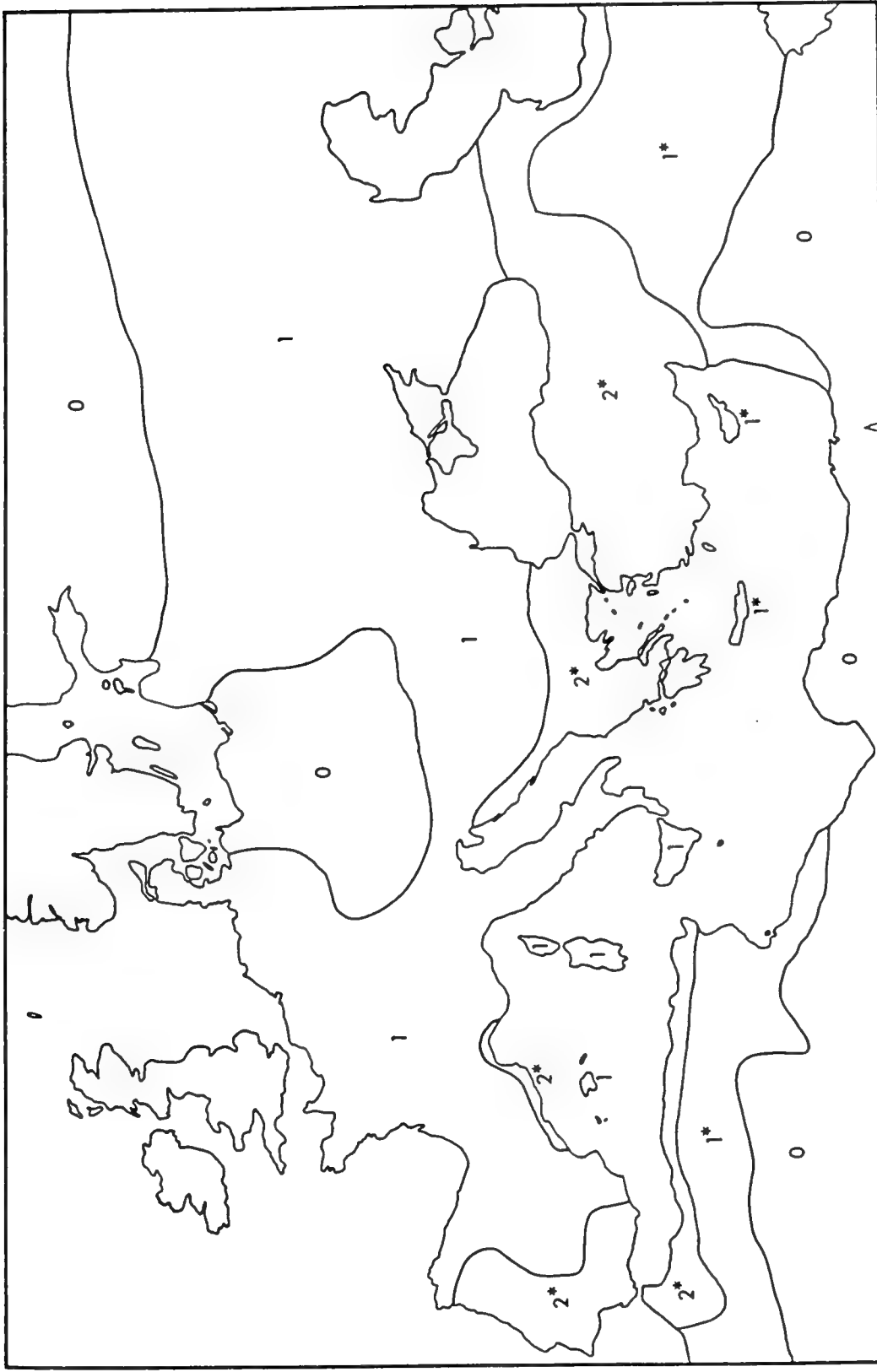


Fig. 8. Species richness of the subfamilies Batagurinae and Emydinae (family Emydidae) on Europe. Richness values marked with an asterisk indicate the presence of a single batagurine species.



Fig. 9. Species richness of the subfamily Emydinae (family Emydidae) on North and Central America, excluding the box turtle genus *Terrapene* and the salt marsh terrapin genus *Malaclemys* (see Fig. 10).

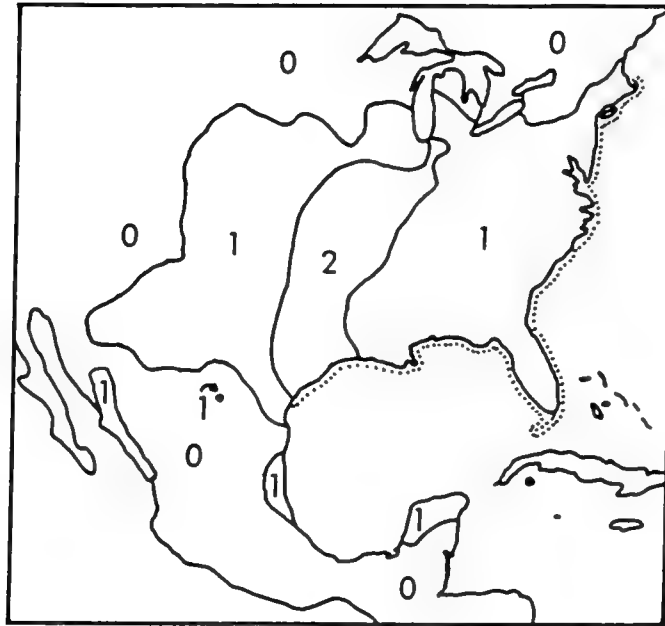


Fig. 10. Species richness of the box turtles of the genus *Terrapene* and the salt marsh terrapin of the genus *Malaclemys* (subfamily Emydinae; family Emydidae) on North America (see Fig. 9). Dotted line parallels the coastline along which a single species of *Malaclemys* occurs.



Fig. 11. Species richness of the subfamily Emydinae (family Emydidae) on northern South America.

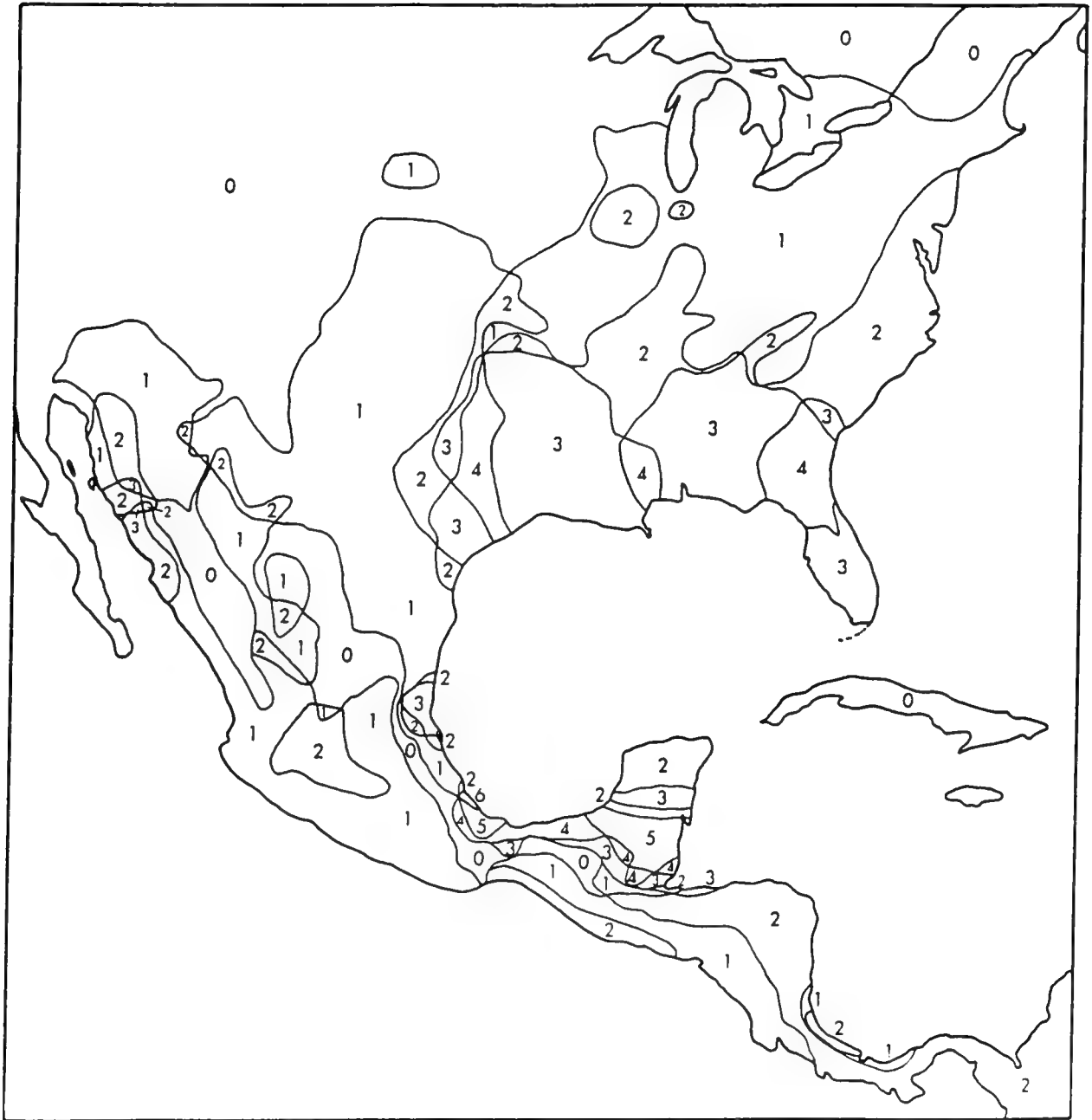


Fig. 12. Species richness of the family Kinostemidae on North and Central America.

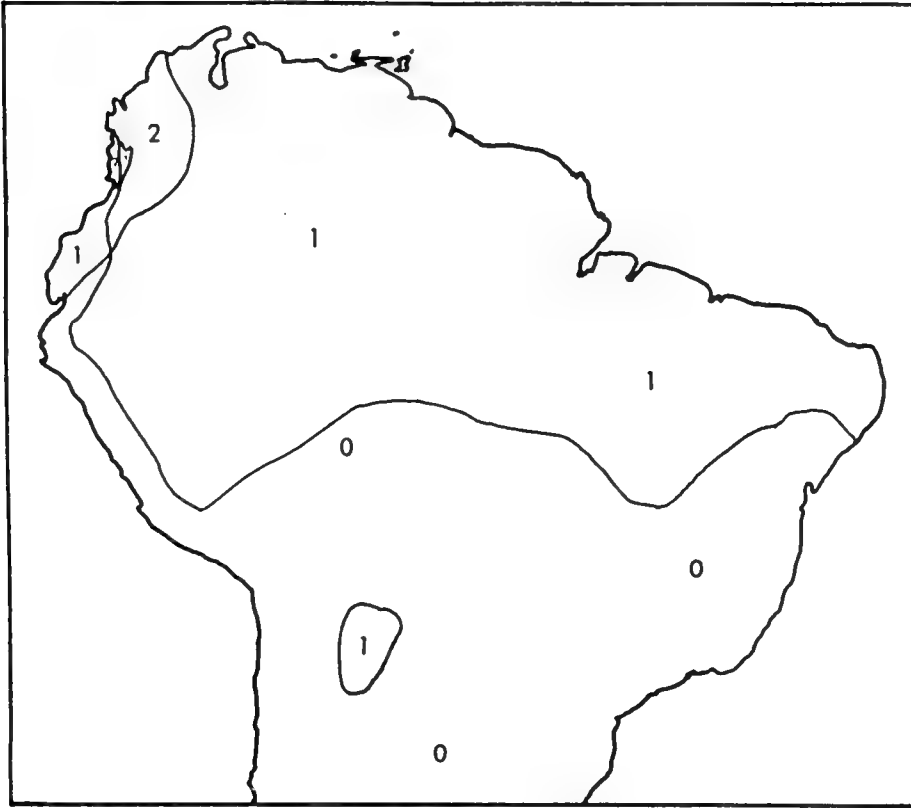


Fig. 13. Species richness of the family Kinosternidae on South America.

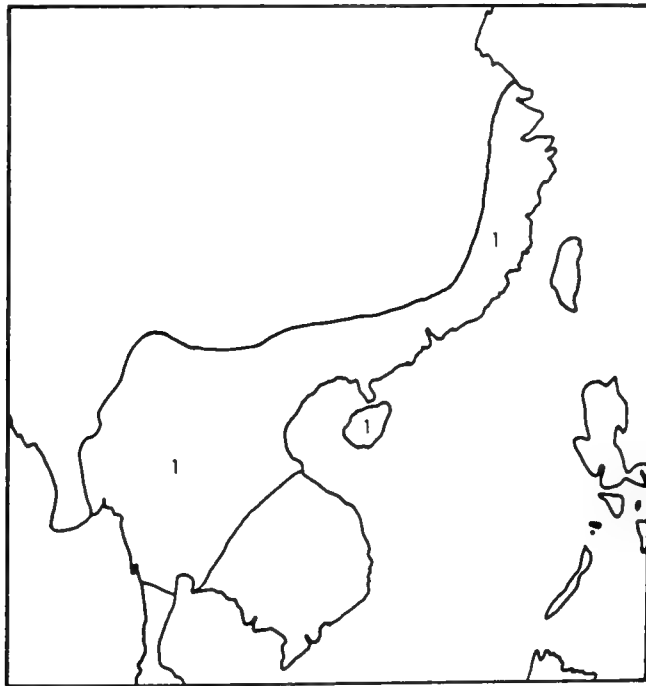


Fig. 14. Species richness of the family Platysternidae on Asia.

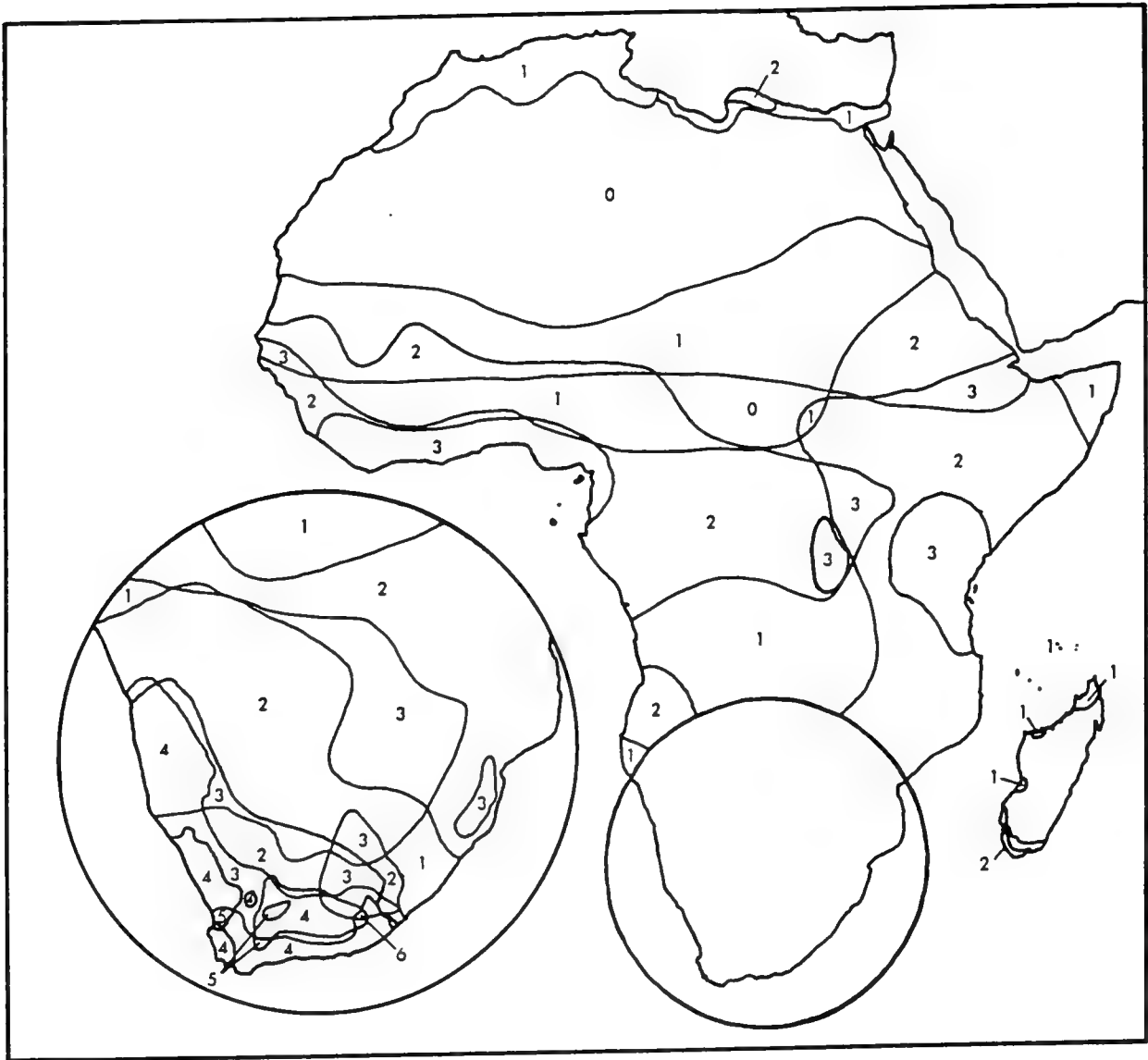


Fig. 15. Species richness of the family Testudinidae on Africa.

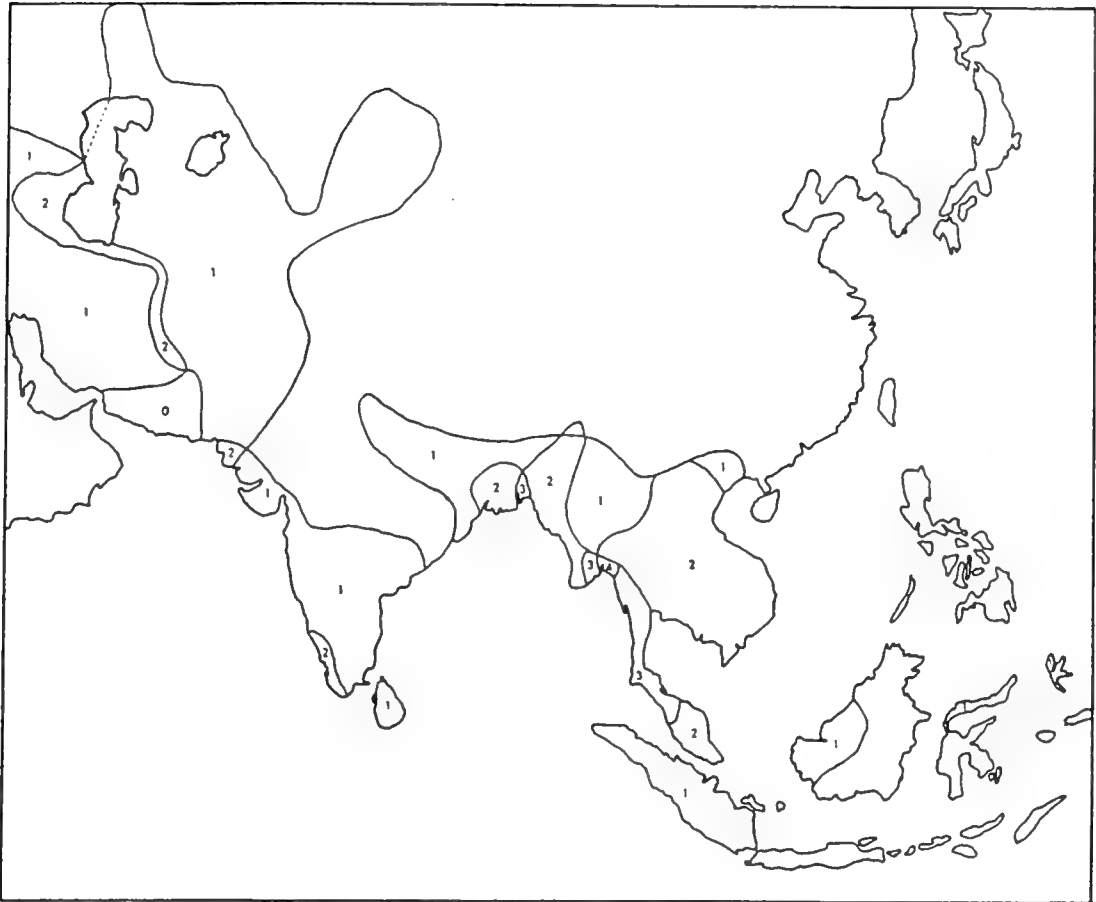


Fig. 16. Species richness of the family Testudinidae on Asia.

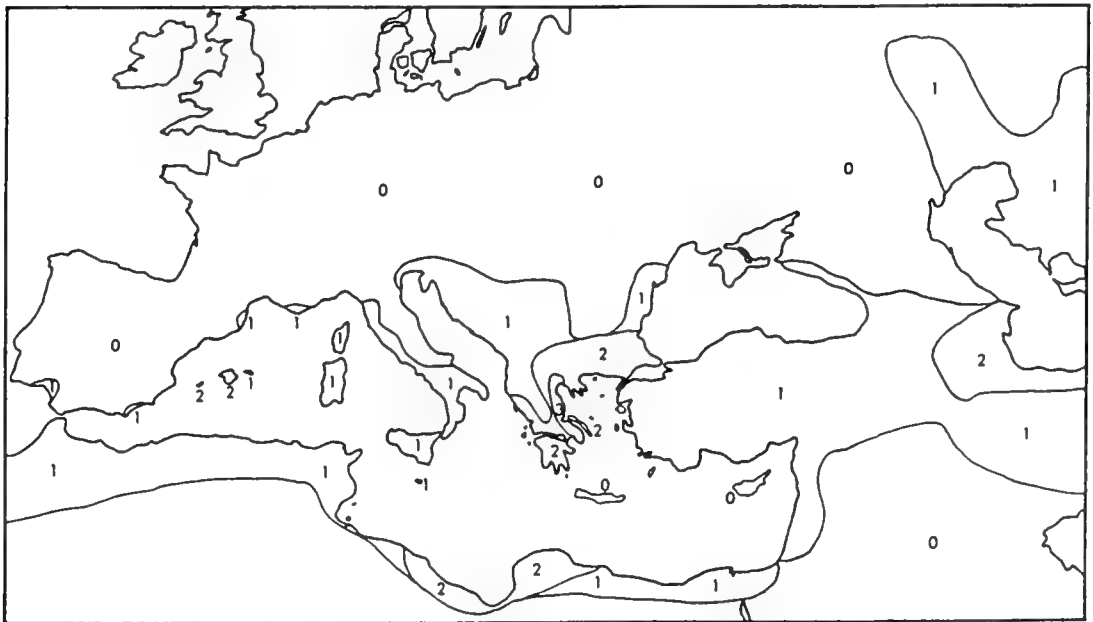


Fig. 17. Species richness of the family Testudinidae on Europe.

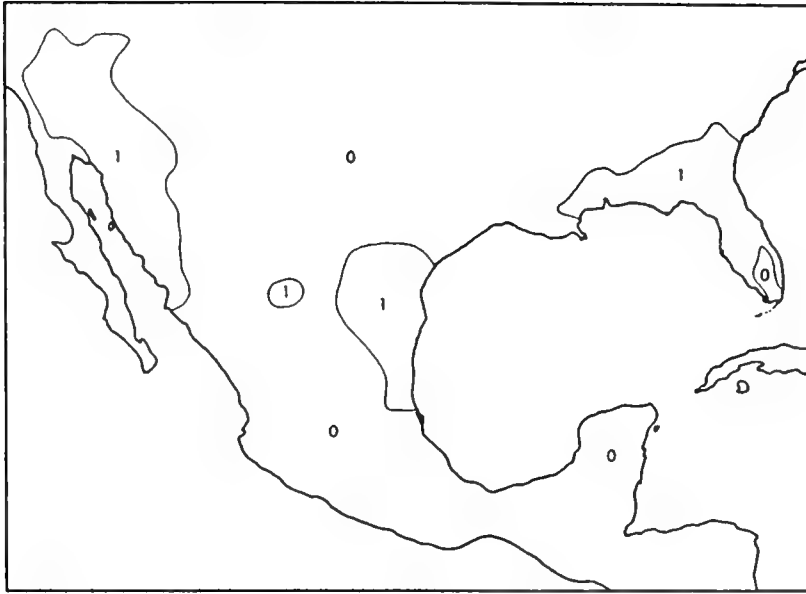


Fig. 18. Species richness of the family Testudinidae on North America.



Fig. 19. Species richness of the family Testudinidae on South America, excluding the Galapagos Islands where species richness never exceeds one.

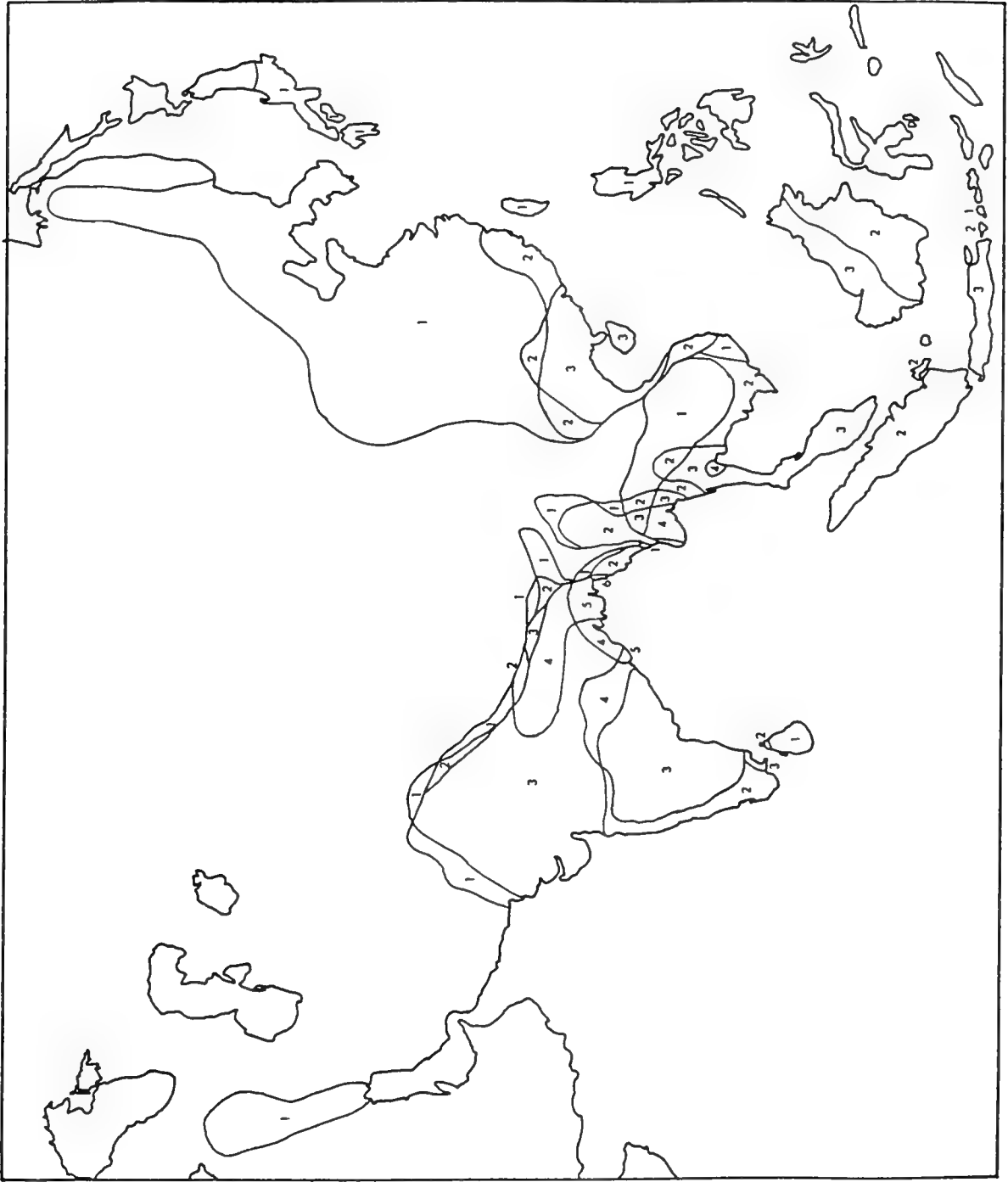


Fig. 20. Species richness of the family Trionychidae on Asia (see also Fig. 21).

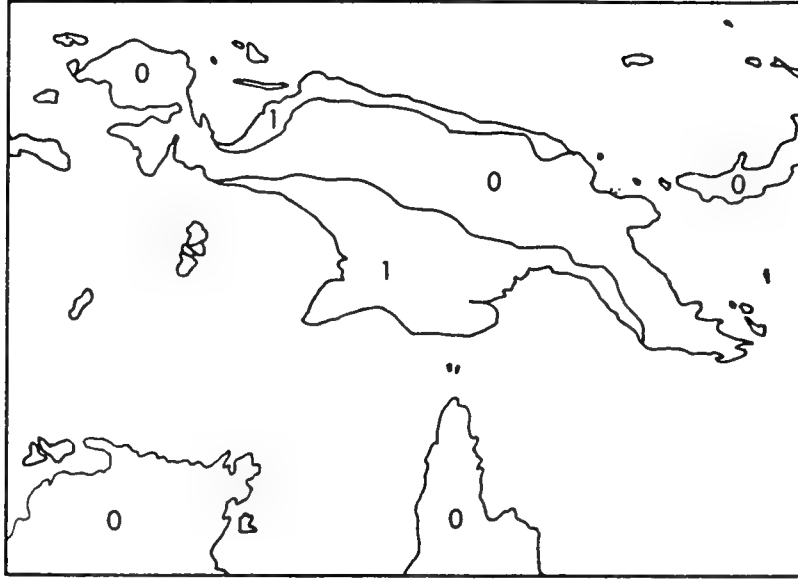


Fig. 21. Species richness of the family Trionychidae on New Guinea.

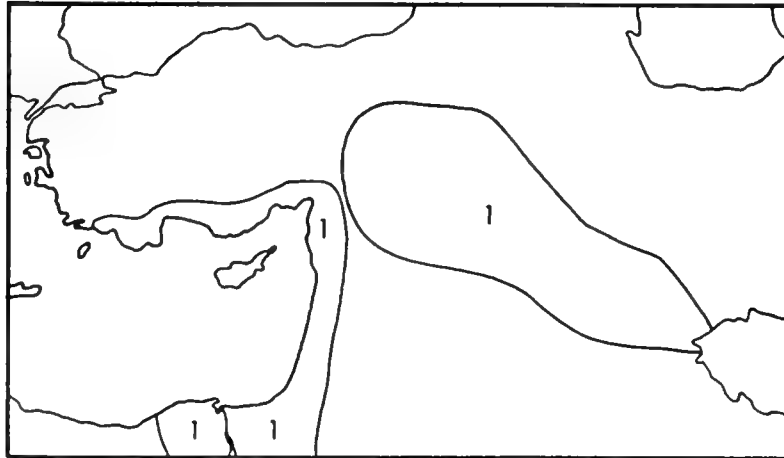


Fig. 22. Species richness of the family Trionychidae in the Middle East.

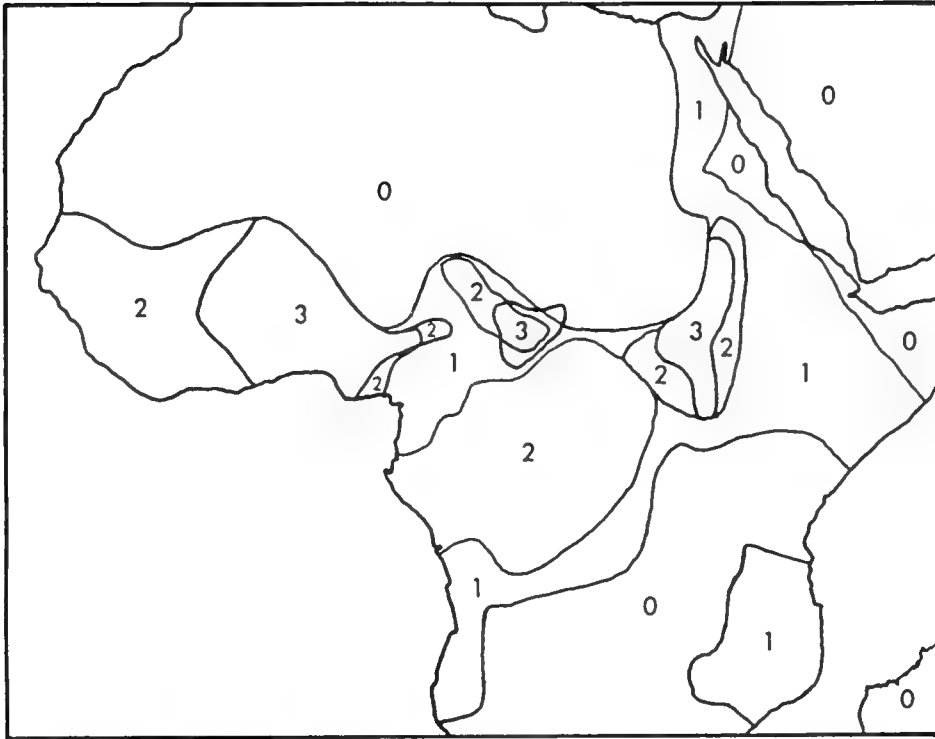


Fig. 23. Species richness of the family Trionychidae on Africa.

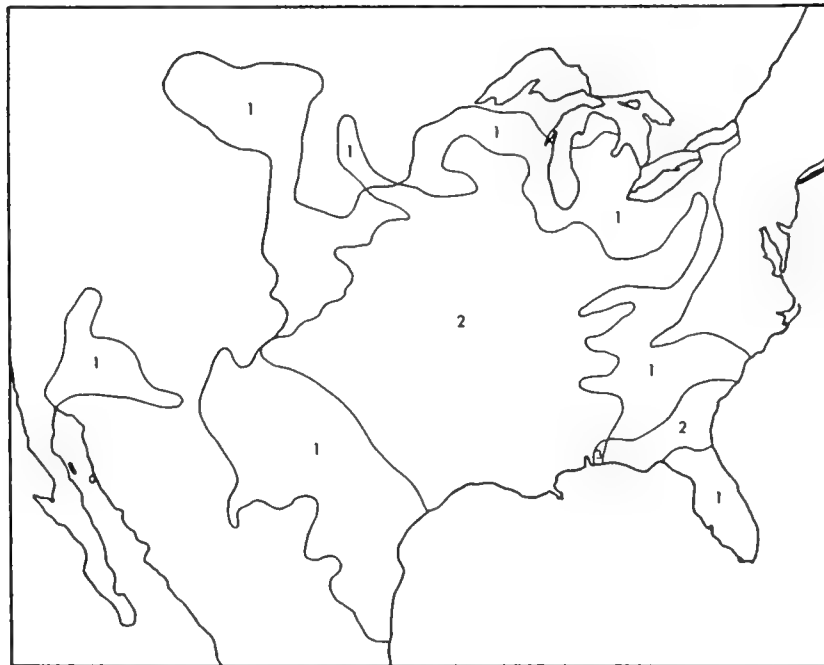


Fig. 24. Species richness of the family Trionychidae on North America.

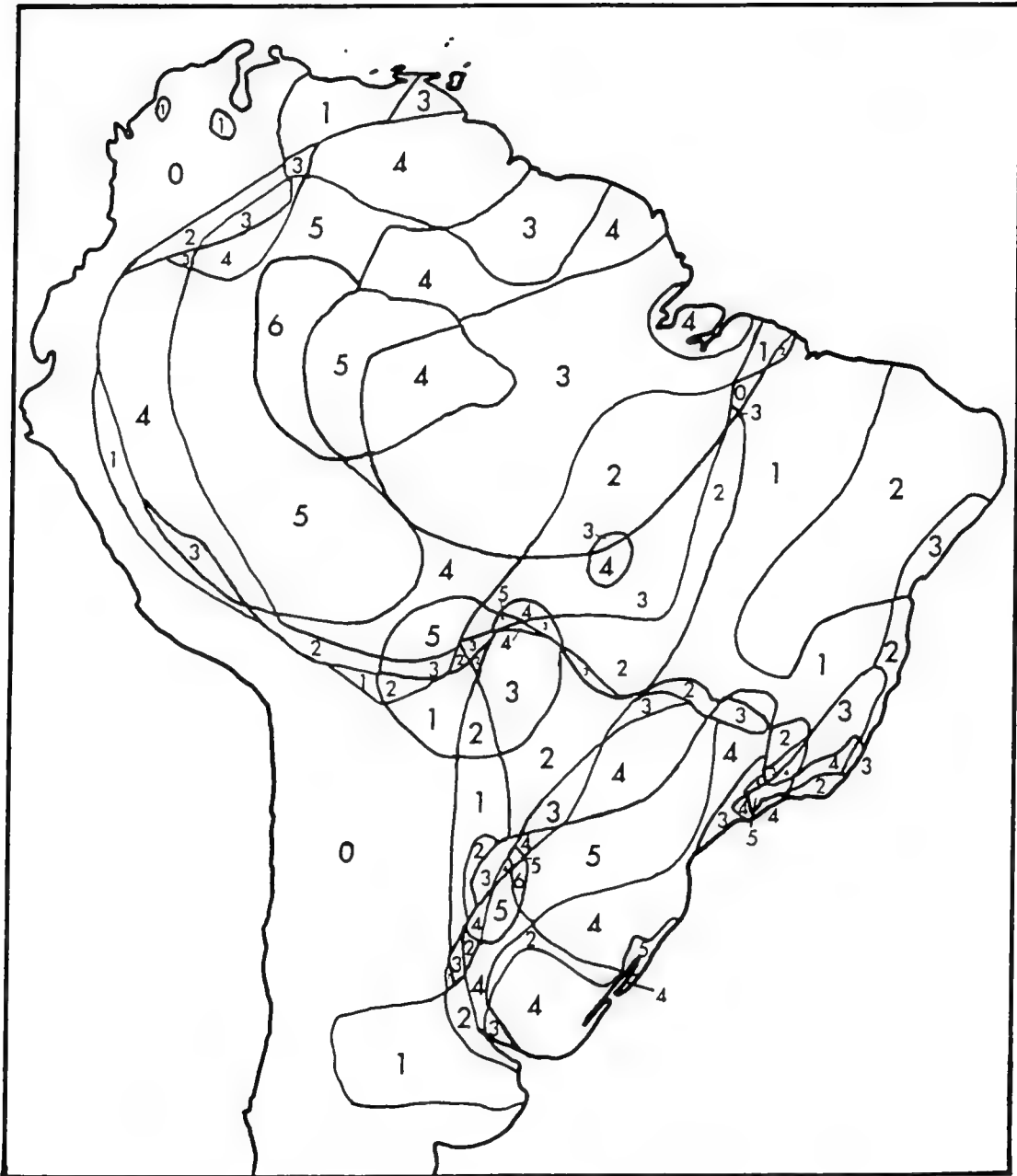


Fig. 25. Species richness of the family Chelidae on South America.

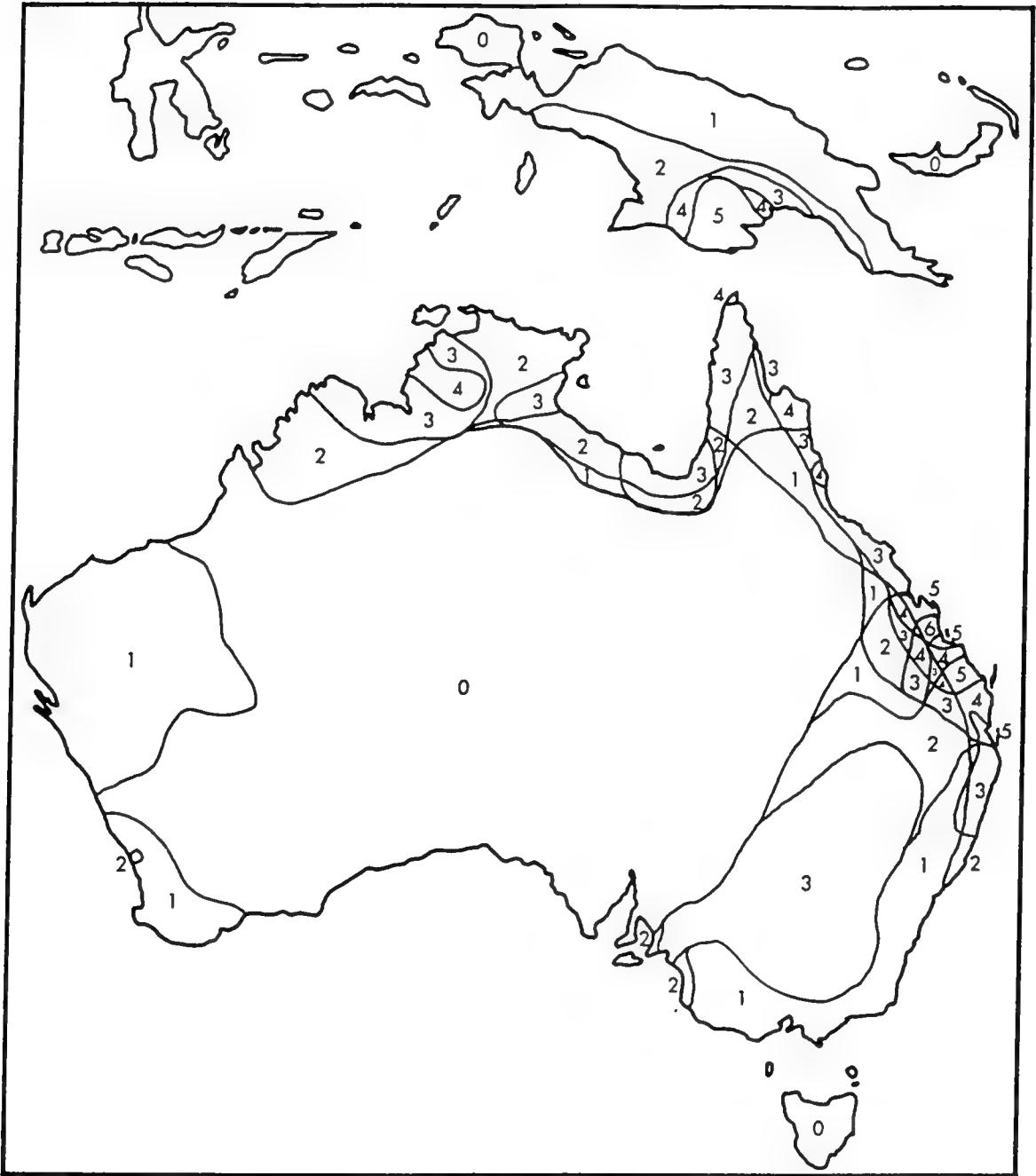


Fig. 26. Species richness of the family Chelidae on Australia.

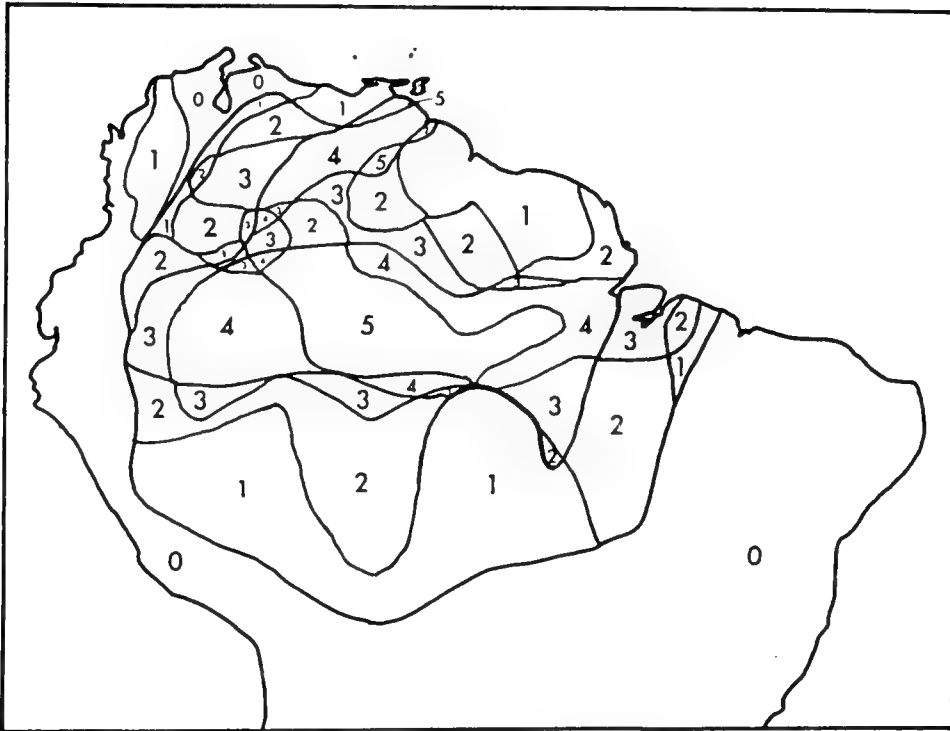


Fig. 27. Species richness of the family Pelomedusidae on South America.

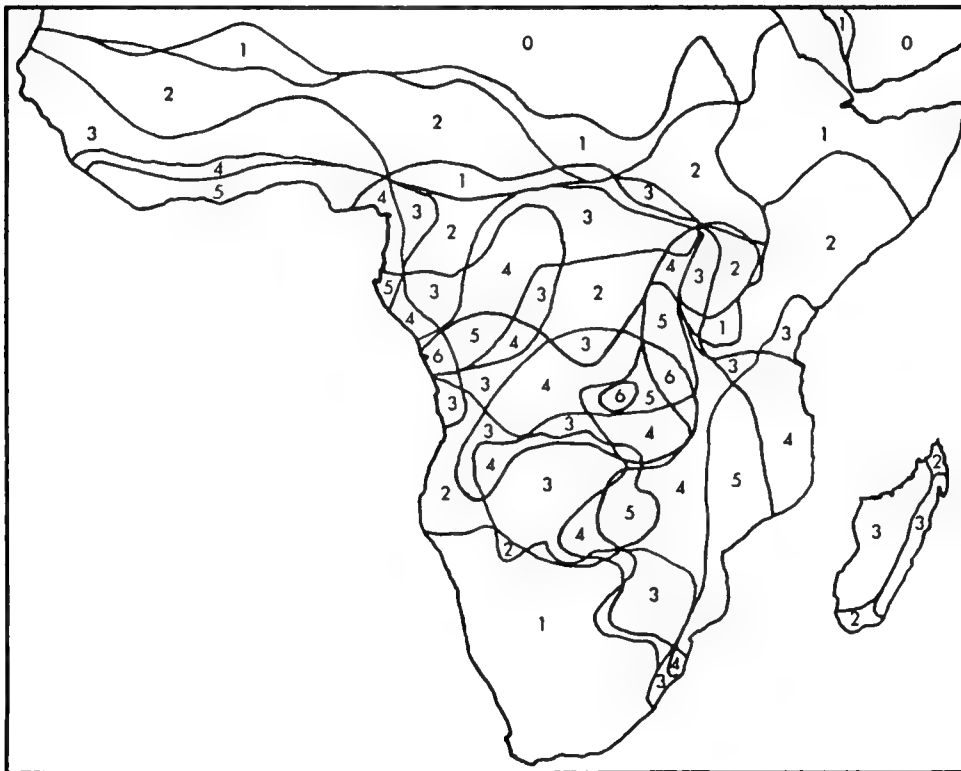
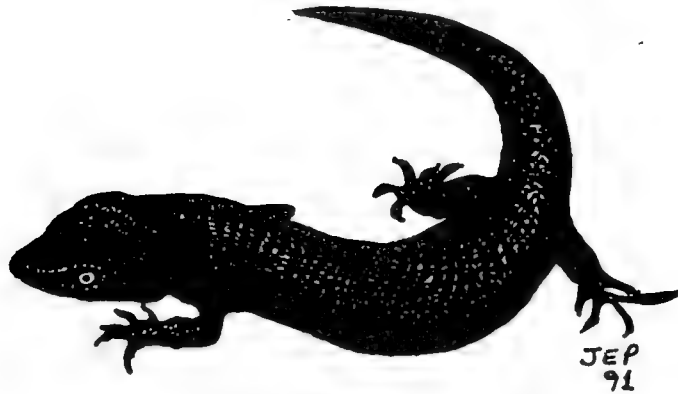


Fig. 28. Species richness of the family Pelomedusidae on Africa.

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CHECKLIST AND BIBLIOGRAPHY (1960-85)
OF THE
VENEZUELAN HERPETOFAUNA



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I N T R O D U C T I O N

The Venezuelan herpetofauna is fairly large compared to any other belonging to a tropical country of similar area. The diversity is due to both a complex physiography and an active speciation process.

The present checklist includes, to the best of my knowledge, all species recorded for Venezuela and described through December 1990. Of the 490 recorded taxa, 15% have been described in the last two decades. The process of description could be stronger if a checklist were available; however, there is no such list. Because many additional species are known but not described and many additional ones awaiting discovery, I offer this checklist as a base line reference tool, realizing that it will require continuing modifications to keep it current with new research discoveries and systematic rearrangements.

In spite of their usefulness, only two other similar checklists are available for South American (Ecuador - Miyata, 1982, SHIS 54: 1-70; Chile - Veloso & Navarro, 1988, Bol. Mus. Reg. Sci. Nat. Torino 6 (2):481-539).

The accompanying bibliography covers the period 1960-1985, attempting to include the fully diversity of topics dealing with the Venezuelan herpetofauna. Undoubtedly, some references have been involuntarily omitted, for which I apologize. References prior to 1960 are covered by Duellman (1979), Duellman & Trueb (1985), Hoogmoed (1973), Medem (1981, 1983), Pritchard & Trebbau (1984), Rivero (1961), Roze (1966), and Vanzolini (1977, 1978) books. We recommend readers to these latter references for the older literature. The 1985 limit was selected because after that year the economical situation in Venezuela forced our libraries to discontinue many subscription and cease or delay the publication of in-country periodicals. Thus since then, bibliographic searches are a haphazard affair.

I hope that this list and bibliography will help the young generations of Venezuelan (and others) herpetologists to more expeditiously begin their studies of this interesting vertebrate fauna.

ACKNOWLEDGMENTS

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Many people and institutions provided help or furnished information, but I want in particular to express my thanks to the Grupo de Ecología Animal and the Library of the Faculty of Sciences, Universidad de Los Andes. Nancy M. Sierra help me through the whole process of collecting bibliographic references. Also, Pedro Hidalgo, Robinson Pérez, John E. Simmons, Pedro Durant, María José Praderio, John D. Lynch and George R. Zug were helpful in all or part of the elaboration of this work.

Several manuscript versions were typed by Ana Isabel Osorio. The final draft was typed into a computer by Belkis Rivas. My thanks to them as well.

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A M P H I B I A

A N U R A

BUFONIDAE

- Atelopus carbonerensis Rivero 1972
 A. cruciger cruciger (Lichtenstein & Martens 1856)
 A. mucubajiensis Rivero 1974
 A. oxyrhynchus Boulenger 1903
 A. pinangoi Rivero 1980
 A. sorianoi La Marca 1983
 Bufo ceratophrys Boulenger 1882
 B. granulatus barbouri Gallardo 1965
 B. g. beebei Gallardo 1965
 B. g. humboldti Gallardo 1965
 B. g. merianae Gallardo 1965
 B. guttatus Schneider 1799
 B. marinus (Linnaeus 1758)
 B. nasicus Werner 1903
 B. sternosignatus Gunther 1858
 B. typhonus alatus Thomiot 1884
 B. t. typhonus (Linnaeus 1758)
 Oreophrynella huberi Diego-Aransay & Gorzula 1987
 O. macconnelli Boulenger 1900
 O. quelchii quelchii (Boulenger 1895)

CENTROLENIDAE

- Centrolenella altitudinalis Rivero 1968
 C. andina Rivero 1968
 C. anthisthenesi Goin 1963
 C. buckleyi venezuelensis Rivero 1968
 C. durantii Rivero 1985
 C. estevesi Rivero 1968
 C. fleischmanni (Boettger 1893)
 C. fragilis Rivero 1985
 C. lentiginosa Rivero 1985
 C. loreocarinata Rivero 1985
 C. orientalis Rivero 1968
 C. ostracodermoides Rivero 1985
 C. pallida Rivero 1985
 C. pleurolineata Rivero 1985
 C. pulidoi Rivero 1968
 C. revocata Rivero 1985

DENDROBATIDAE

- Colostethus alboguttatus (Boulenger 1903)
 C. bromelicola (Test 1956)
 C. brunneus (Cope 1887)
 C. collaris (Boulenger 1912)
 C. dunni (Rivero 1961)
 C. durantei Péfaur 1985
 C. haydeae Rivero 1976
 C. herminae (Boettger 1893)
 C. humilis Rivero 1978
 C. leopardalis Rivero 1976
 C. mandelorum (Schmidt 1932)
 C. mayorgai Rivero 1978
 C. meridensis Dole & Durant 1972
 C. molinarii La Marca 1985
 C. neblina (Test 1956)
 C. oblitteratus Rivero 1984
 C. orostoma Rivero 1976
 C. riveroi (Donoso - Barros 1964)
 C. sanmartini Rivero, Langone & Prigioni 1986
 C. saltuensis Rivero 1978
 C. serranus Péfaur 1985
 C. shrevei (Rivero 1961)
 C. trinitatis (Garman 1887)
 Dendrobates leucomelas Steindachner 1864
 Epipedobates pictus (Tschudi 1838)
 Minyobates steyermarki (Rivero 1971)

HYLIDAE

- Aparasphenodon venezolanus (Mertens 1950)
 Flectonotus fitzgeraldi Parker 1934
 F. pygmaeus (Boettger 1893)
 Gastrotheca helenae Dunn 1944
 G. nicefori Gaige 1933
 G. ovifera (Weinland 1854)
 G. walkeri Duellman 1980
 Hyla albomarginata Spix 1824
 H. albopunctata Spix 1824
 H. alemani Rivero 1964
 H. battersbyi Rivero 1961
 H. benitezi Rivero 1961
 H. boans (Linnaeus 1758)
 H. crepitans Wied 1824
 H. geographica Spix 1824
 H. granosa Boulenger 1882
 H. jahni Rivero 1961
 H. labialis meridensis Rivero 1961

- H. lanciformis guerreroi Rivero 1971
 H. lascinia Rivero 1969
 H. lemai Rivero 1972
 H. loveridgei Rivero 1961
 H. luteocellata Roux 1927
 H. marmorata (Laurenti 1768)
 H. microcephala misera Werner 1903
 H. minuscula Rivero 1971
 H. minuta Peters 1872
 H. multifasciata Gunther 1858
 H. pelidna Duellman 1989
 H. platydactyla Boulenger 1905
 H. punctata punctata (Schneider 1799)
 H. sibleszi Rivero 1972
 H. vigilans Solano 1971
 Ololygon baumgardneri (Rivero 1961)
 O. boesemani (Goin 1966)
 O. boulengeri (Cope 1887)
 O. danae Duellman 1986
 O. exigua Duellman 1986
 O. garbei (Miranda-Ribeiro 1926)
 O. nebulosa Spix 1824
 O. rostrata (Peters 1863)
 O. rubra rubra (Laurenti 1768)
 O. trilineata Hoogmoed & Gorzula 1979
 O. x-signata (Spix 1824)
 Osteocephalus rodriguezi (Rivero 1968)
 O. taurinus Steindachner 1862
 Phrynohyas venulosa (Laurenti 1768)
 Phyllomedusa bicolor (Boddaert 1772)
 P. hypocondrialis hypocondrialis (Daudin 1803)
 P. medinae Funkhouser 1962
 P. trinitatis Mertens 1926
 Sphaenorhynchus lacteus (Daudin 1801)
 Stefania evansi (Boulenger 1904)
 S. ginesi Rivero 1966
 S. goini Rivero 1966
 S. marahuaquensis (Rivero 1961)
 S. riae Duellman & Hoogmoed 1984

ALLOPHRYNIDAE

Allophryne ruthveni Gaige 1926

LEPTODACTYLIDAE

- Adelophryne gutturosa Hoogmoed & Lescure 1984
 Adenomera andreae (Muller 1923)
 A. hylaedactyla (Cope 1868)

- Ceratophrys calcarata* Boulenger 1890
 C. *cornuta* (Linnaeus 1758)
Dischidodactylus duidensis (Rivero 1968)
 D. *colonnelloi* Ayarzagüena 1983
Eleutherodactylus anotis Walker & Test 1955
 E. *bicumulus* (Peters 1863)
 E. *boconoensis* Rivero & Mayorga 1973
 E. *briceni* (Boulenger 1903)
 E. *chlorosoma* Rivero 1982
 E. *colostichos* La Marca & Smith 1982
 E. *cornutus* (Jiménez de la Espada 1871)
 E. *johnstonei* Barbour 1914
 E. *ginesi* (Rivero 1964)
 E. *lancinii* Donoso-Barros 1968
 E. *lentiginosus* Rivero 1982
 E. *marmoratus* (Boulenger 1900)
 E. *maussi* (Boettger 1893)
 E. *melanoproctus* Rivero 1982
 E. *mondolfii* Rivero 1982
 E. *nicefori* Cochran & Goin 1970
 E. *orocostalis* Rivero 1961
 E. *paramerus* Rivero 1982
 E. *pleurostriatus* Rivero 1982
 E. *prolixodiscus* Lynch 1978
 E. *pulidoi* Rivero 1982
 E. *pulvinatus* Rivero 1968
 E. *racenisi* Rivero 1961
 E. *reticulatus* Walker & Test 1955
 E. *rozei* Rivero 1961
 E. *stenodiscus* Walker & Test 1955
 E. *terraebolivaris* Rivero 1961
 E. *tubernasus* Rivero 1982
 E. *turumiquirensis* Rivero 1961
 E. *urichi urichi* (Boettger 1894)
 E. *vanadise* La Marca 1984
 E. *vilarsi* (Melin 1941)
 E. *williamsi* Rivero 1961
Leptodactylus bolivianus Boulenger 1898
 L. *fragilis* (Brocchi 1877)
 L. *fuscus* (Schneider 1799)
 L. *knudseni* Heyer 1972
 L. *labyrinthicus* (Spix 1824)
 L. *longirostris* Boulenger 1882
 L. *macrosternum* Miranda-Ribeiro 1926
 L. *mystaceus* (Spix 1824)
 L. *pentadactylus* (Laurenti 1768)
 L. *poecilochilus* (Cope 1862)
 L. *rhodomystax* Boulenger 1883
 L. *riveroi* Heyer & Pyburn 1983

- L. rugosus Noble 1923
 L. wagneri (Peters 1862)
 Lithodytes lineatus (Schneider 1799)
 Physalaemus enesevae Heatwole, Solano & Heatwole 1965
 P. fischeri (Boulenger 1896)
 P. pustulosus ruthveni (Netting 1930)
 Pleurodema brachyops (Cope 1868)
 Pseudopaludicola boliviana Parker 1927
 P. llanera Lynch 1989
 P. pusilla (Ruthven 1916)

MICROHYLIDAE

- Adelastes hylonomus Zweifel 1986
 Elachistocleis ovalis (Schneider 1979)
 E. surinamensis (Daudin 1802)
 Otophryne robusta robusta Boulenger 1900
 O. r. steyermarki Rivero 1967

PIPIDAE

- Pipa arrabali Izecksohn 1976
 P. parva Ruthven & Gaige 1923
 P. pipa (Linnaeus 1758)

RANIDAE

- Rana palmipes Spix 1824

PSEUDIDAE

- Pseudis paradoxa caribensis Gallardo 1961

C A U D A T A

PLETHODONTIDAE

- Bolitoglossa borburata Trapido 1942
 B. orestes Brame & Wake 1962

G Y M N O P H I O N A

CAECILIAIDAE

- Caecilia flavopunctata Roze & Solano 1963
 C. gracilis Shaw 1802
 C. subnigricans Dunn 1942
 C. tentaculata Linnaeus 1758

POTOMOTYPHLIDAE

Potomotyphlus kaupii (Berthold 1859)

RHINATREMATIDAE

Epicrionops nigrus (Dunn 1942)

SIPHONOPIDAE

Microcaecilia rabei (Roze & Solano 1963)
Siphonops annulatus (Mikan 1820)

TYPHLONECTIDAE

Nectocaecilia haydee (Roze 1963)
Typhlonectes venezuelense Fuhrmann 1914

R E P T I L I A

T E S T U D I N E S

PELOMEDUSIDAE

Podocnemis erythrocephala (Spix 1824)
P. expansa (Schweigger 1812)
P. unifilis Troschel 1848
P. vogli Muller 1935
Peltocephalus dumerilianus (Schweigger 1812)

CHELIDAE

Chelus fimbriatus (Schneider 1783)
Phrynops (Phrynops) geoffroanus (Schweigger 1812)
P. (Mesoclemmys) gibbus (Schweigger 1812)
P. (Batrachemys) nasutus (Schweigger 1812)
P. zuliae Pritchard & Trebbau 1984
Platemys platycephala (Schneider 1792)

EMYDIDAE

Rhinoclemmys diademata (Mertens 1954)
R. punctularia flammigera Paolillo 1985
R. p. punctularia (Daudin 1801)
Trachemys scripta callirostris (Gray 1855)
T. s. chichiriviche Pritchard & Trebbau 1984

TESTUDINIDAE

Geochelone (Chelonoidis) carbonaria (Spix 1824)
 G. denticulata (Linnaeus 1766)

KINOSTERNIDAE

Kinosternon scorpioides scorpioides (Linnaeus 1766)

DERMOCHELYIDAE

Dermochelys coriacea (Linnaeus 1766)

CHELONIIDAE

Chelonia mydas (Linnaeus 1758)
 Caretta caretta (Linnaeus 1758)
 Eretmochelys imbricata imbricata (Linnaeus 1766)
 Lepidochelys olivacea (Eschscholz 1829)

C R O C O D Y L I A

CROCODYLIDAE

Crocodylus acutus (Cuvier 1807)
 C. intermedius Graves 1819

ALLIGATORIDAE

Caiman crocodilus crocodilus Linnaeus 1758
 C. c. fuscus (Cope 1868)
 Paleosuchus palpebrosus (Cuvier 1807)
 P. trigonatus (Schneider 1801)

A M P H I S B A E N I A

AMPHISBAENIDAE

Amphisbaena alba Linnaeus 1758
 A. fuliginosa fuliginosa Linnaeus 1758
 A. f. varia Laurenti 1768
 A. rozei Lancini 1963
 A. spurrelli Boulenger 1915
 Mesobaena huebneri Mertens 1925

L A C E R T I L I A

SCINCIDAE

- Mabuya bistriata (Spix 1825)
 M. croizati Horton 1973
 M. ficta Rebouças-Spieker 1981

GEKKONIDAE

- Coleodactylus amazonicus (Anderson 1918)
 C. septentrionalis Vanzolini 1980
 Gonatodes albogularis albogularis (Dumeril & Bibron 1836)
 G. annularis Boulenger 1887
 G. antillensis (Van Lidth de Jeude 1887)
 G. bodinii Rivero-Blanco 1964
 G. ceciliae Donoso-Barros 1966
 G. concinnatus (O'Shaughnessy 1881)
 G. falconensis Shreve 1947
 G. humeralis (Guichenot 1855)
 G. ocellatus (Gray 1831)
 G. petersi Donoso-Barros 1967
 G. seigliei Donoso-Barros 1966
 G. taniae Roze 1963
 G. vittatus (Lichtenstein 1856)
 Hemidactylus mabouia (Moreau de Jonnes 1818)
 H. palaichthus Kluge 1969
 Phyllodactylus dixonii Rivero-Blanco & Lancini 1967
 P. ventralis O'Shaughnessy 1875
 Pseudogonatodes guianensis Parker 1935
 P. lunulatus (Roux 1927)
 Sphaerodactylus molei Boettger 1894
 Thecadactylus rapicaudus (Houttuyn 1782)

IGUANIDAE

- Anolis auratus Daudin 1802
 A. chrysolepis Dumeril & Bibron 1837
 A. frenatus Cope 1899
 A. fuscoauratus klugeri Roux 1929
 A. gibbiceps Cope 1864
 A. jacare Boulenger 1903
 A. onca (O'Shaughnessy 1875)
 A. punctatus Daudin 1802
 A. roquet (Bonnaterre 1789)
 A. sagrei Dumeril & Bibron 1837
 A. squamulatus Peters 1863
 A. transversalis Dumeril 1851
 A. tropidogaster Hallowell 1857

Basiliscus basiliscus barbouri Ruthven 1914
 B. b. *basiliscus* (Linnaeus 1758)
Iguana iguana iguana (Linnaeus 1758)
Phenacosaurus nicefori Dunn 1944
Plica plica (Linnaeus 1758)
 P. *umbra umbra* (Linnaeus 1758)
Polychrus marmoratus (Linnaeus 1758)
Tropidodactylus onca (O'Shaughnessy 1875)
Tropidurus bogerti Roze 1958
 T. *hispidus* (Spix 1825)
Uracentron weneri Mertens 1925
Uranoscodon superciliosum (Linnaeus 1758)

TEIIDAE

Ameiva ameiva (Linnaeus 1758)
 A. *bifrontata* Cope 1862
Anadia bitaeniata Boulenger 1903
 A. *blakei* Schmidt 1932
 A. *bogotensis* (Peters 1862)
 A. *brevifrontalis* (Boulenger 1903)
 A. *hobarti* La Marca & García-Pérez 1990
 A. *marmorata* (Gray 1846)
 A. *pamplonensis* Dunn 1944
 A. *steyeri* Nieden 1914
Arthrosaura reticulata (O'Shaughnessy 1881)
Bachia bicolor (Cope 1896)
 B. *flavescens flavescens* (Bonnaterre 1789)
 B. f. *schlegeli* (Dumeril & Bibron 1839)
 B. *guianensis* Hoogmoed & Dixon 1977
 B. *heteropa lineata* Boulenger 1903
 B. h. *marcelae* Donoso-Barros & Garrido 1964
 B. h. *trinitatis* (Barbour 1914)
 B. *monodactyla* (Daudin 1802)
Cercosaura ocellata ocellata Wagler 1830
Cnemidophorus lemniscatus lemniscatus (Linnaeus 1758)
 C. l. *nigricolor* Peters 1873
 C. *gramivagus* McCrystal & Dixon 1987
Euspondylus acutirostris (Peters 1862)
 E. *phelpsi* Lancini 1968
Gymnophthalmus speciosus (Hallowell 1861)
 G. *underwoodi* Grant 1958
Kentropyx borckianus Peters 1862
 K. *calcarata* Spix 1825
 K. *striata striata* Daudin 1802
Leposoma percarinatum (Muller 1923)
Neusticurus bicarinatus (Linnaeus 1758)
 N. *racenisi* Roze 1958
 N. *rudis* Boulenger 1900

N. tatei (Burt & Burt 1931)
 Prionodactylus ampuedai (Lancini 1968)
 Proctoporus achlyens Uzzell 1958
 P. luctuosus (Peters 1862)
 Ptychoglossus kugleri Roux 1927
 P. brevifrontalis (Boulenger 1903)
 Riolama leucostictus (Boulenger 1900)
 Tretioscincus bifasciatus (Dumeril 1851)
 Tupinambis teguixin (Linnaeus 1758)

S E R P E N T E S

BOIDAE

Boa constrictor constrictor Linnaeus 1758
 Corallus caninus (Linnaeus 1758)
 C. enydris enydris (Linnaeus 1758)
 C. e. cooki Gray 1842
 Epicrates cenchria cenchria (Linnaeus 1758)
 E. c. maurus Gray 1849
 Eunectes murinus gigas (Latreille 1802)

ANILIIDAE

Anilius scytale phelpsorum Roze 1958

LEPTOTYPHLOPIDAE

Leptotyphlops affinis (Boulenger 1884)
 L. amazonicus Orejas-Miranda 1969
 L. dimidiatus (Jan 1861)
 L. goudotti goudotti (Dumeril & Bibron 1854)
 L. macrolepis (Peters 1857)
 L. septemstriatus (Schneider 1801)
 L. tenellus Klauber 1939

ANOMALEPIDAE

Helminthophis flavotermiatus (Peters 1857)
 Liotyphlops albirostris (Peters 1881)

TYPHLOPIDAE

Typhlops brongersmianus Vanzolini 1972
 T. lehneri Roux 1926
 T. minuisquamus Dixon & Hendricks 1979
 T. reticulatus (Linnaeus 1758)

COLUBRIDAE

- Atractus badius* (Boie 1827)
 A. *duidensis* Roze 1961
 A. *elaps* (Gunther 1858)
 A. *emigdioi* González-Sponga 1971
 A. *erythromelas* Boulenger 1903
 A. *fuliginosus* (Hallowell 1845)
 A. *insipidus* Roze 1961
 A. *lancinii* Roze 1967
 A. *major* Boulenger 1894
 A. *mariselae* Lancini 1969
 A. *riveroi* Roze 1961
 A. *steyermarki* Roze 1958
 A. *trilineatus* Wagler 1828
 A. *univittatum* (Jan 1862)
 A. *ventrimaculatus* Boulenger 1905
 A. *vittatus* Boulenger 1894
Chironius carinatus (Linnaeus 1758)
 C. *exoletus* Linnaeus 1758
 C. *fuscus fuscus* (Linnaeus 1758)
 C. *monticola* Roze 1952
 C. *multiventris multiventris* Schmidt & Walker 1943
 C. *m. septentrionalis* Wiest 1978
 C. *scurrulus* (Wagler 1824)
Clelia clelia clelia (Daudin 1803)
Dendrophidion dendrophis (Schlegel 1837)
 D. *nuchalis* (Peters 1864)
Dipsas catesbyi (Sentzen 1796)
 D. *copei* (Gunther 1872)
 D. *latifrontalis* (Boulenger 1905)
 D. *pavonina* Schlegel 1837
 D. *perijanensis* (Aleman 1953)
 D. *variegata variegata* (Dumeril, Bibron & Dumeril 1854)
Drymarchon corais corais (Boie 1827)
 D. *c. margaritae* Roze 1959
 D. *c. melanurus* (Dumeril, Bibron & Dumeril 1854)
Drymobius rhombifer (Gunther 1860)
Drymoluber dichrous (Peters 1863)
Erythrolamprus aesculapii aesculapii (Linnaeus 1758)
 E. *bizona* Jan 1863
 E. *pseudocorallus* Roze 1959
Helicops angulatus (Linnaeus 1758)
 H. *hagmanni* Roux 1910
 H. *hogeii* Lancini 1964
 H. *pastazae* Shreve 1934
 H. *scalaris* Jan 1865
Hydrodynastes bicinctus bicinctus (Hermann 1804)
Hydrops triangularis venezuelensis Roze 1957

Imantodes cenchoa (Linnaeus 1758)
 Lampropeltis triangulum (Cope 1860)
 Leptodeira annulata annulata (Linnaeus 1758)
 L. a. ashmeadii (Hallowell 1845)
 L. septentrionalis ornata (Bocourt 1884)
 Leptophis ahaetulla coeruleodorsus Oliver 1942
 L. a. copei Oliver 1942
 L. a. occidentalis (Gunther 1859)
 Liophis breviceps breviceps Cope 1860
 L. b. canaimus Roze 1957
 L. cobellus cobellus (Linnaeus 1758)
 L. c. trebbauai Roze 1958
 L. epinephalus opisthotaenius (Boulenger 1908)
 L. lineatus (Linnaeus 1758)
 L. melanotus melanotus (Shaw 1802)
 L. poecilogyrus (Wied 1825)
 L. reginae semilineatus (Wagler 1824)
 L. r. zweifeli Roze 1959
 L. typhlus typhlus (Linnaeus 1758)
 L. williamsi (Roze 1958)
 Masticophis mentovarius centralis (Roze 1953)
 M. m. suborbitalis (Peters 1868)
 Mastigodryas amarali (Stuart 1938)
 M. bifossatus striatus (Amaral 1931)
 M. boddaerti boddaerti (Sentzen 1796)
 M. pleei (Dumeril, Bibron & Dumeril 1854)
 Ninia atrata (Hallowell 1845)
 Oxybelis aeneus Wagler in Spix 1824
 O. argenteus (Daudin 1803)
 O. fulgidus (Daudin 1803)
 Oxyrhopus petola petola (Linnaeus 1758)
 O. p. digitalis (Reuss 1834)
 O. trigeminus Dumeril, Bibron & Dumeril 1854
 O. venezuelanus Shreve 1947
 Philodryas olfersii herbens (Wied 1825)
 P. viridissimus (Linnaeus 1758)
 Phimophis guianensis (Troschel in Schomburgh 1848)
 Pseudoboa coronata Schneider 1801
 P. neuwieddi (Dumeril, Bibron & Dumeril 1854)
 Pseudoeryx plicatilis plicatilis (Linnaeus 1758)
 Pseustes poecilonotus polylepis (Peters 1867)
 P. sulphureus sulphureus (Wagler in Spix 1824)
 Rhadinaea brevirostris (Peters 1863)
 R. multilineata (Peters 1859)
 Rhinobothryum bovalli Andersson 1926
 Sibon nebulata nebulata (Linnaeus 1758)
 Spilotes pullatus pullatus (Linnaeus 1758)
 Stenorrhina degenhardtii ocellata Jan 1876
 Tantilla melanocephala (Linnaeus 1758)

- T. semicincta (Dumeril, Bibron & Dumeril 1854)
 Thamnodynastes chimanta Roze 1958
 T. pallidus (Linnaeus 1758)
 T. strigilis (Thunberg 1787)
 Tripanurgos compressus (Daudin 1803)
 Umbrivaga mertensi Roze 1964
 Xenodon rabdocephalus (Wied 1824)
 X. severus (Linnaeus 1758)

ELAPIDAE

- Leptomicrurus collaris breviventris Roze & Bernarl-Carlo 1987
 Micrurus dissoleucus dissoleucus (Cope 1860)
 M. d. meridensis Roze 1989
 M. dumerilii carinicaudus Schmidt 1936
 M. d. venezuelensis Roze 1989
 M. hemprichii hemprichii (Jan 1858)
 M. h. ortonii Schmidt 1953
 M. isozonus (Cope 1860)
 M. lemniscatus diutius Burger 1955
 M. l. helleri Schmidt & Schmidt 1925
 M. mipartitus semipartitus (Jan 1858)
 M. psyches psyches (Daudin 1803)
 M. p. remotus Roze 1987
 M. spixii obscurus (Jan in Jan & Sordelli 1872)
 M. surinamensis nattereri Schmidt 1952

VIPERIDAE

- Bothrops atrox atrox (Linnaeus 1758)
 B. a. colombiensis (Hallowell 1845)
 B. bilineatus bilineatus (Wied 1825)
 B. b. smaragdinus Hoge 1966
 B. brazili Hoge 1953
 B. lansbergii rozei Peters 1959
 B. medusa Sternfeld 1920
 B. schlegelii (Berthold 1846)
 B. taeniatus lichenosus Roze 1958
 B. t. taeniatus Wagler 1824
 B. venezuelensis Sandner-Montilla 1952
 Crotalus durissus cumananensis Humboldt 1811
 C. d. pifanorum Sandner-Montilla 1980
 C. d. ruruima Hoge 1965
 C. vegrandis Klauber 1941
 Lachesis muta muta (Linnaeus 1766)

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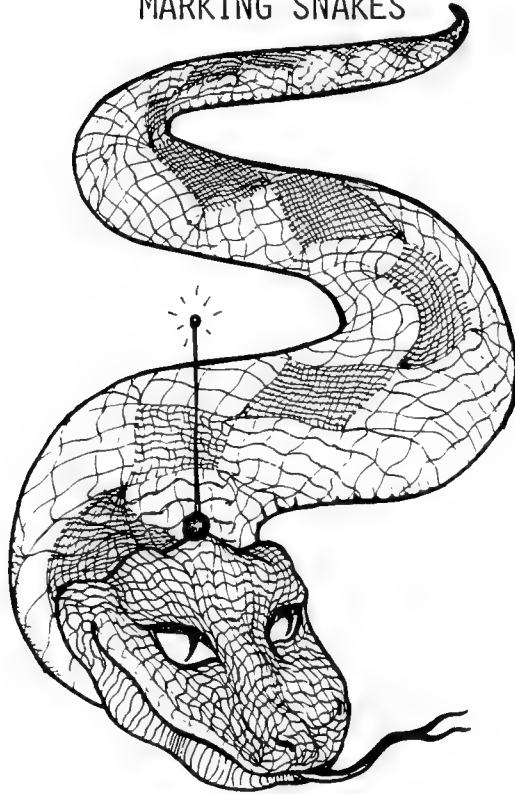
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A REVIEW OF TECHNIQUES
FOR
MARKING SNAKES



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COVER ILLUSTRATION. Radiotrackers' ideal snake with a built-in antenna; drawn by Stephanie Ersek.

INTRODUCTION

Snakes present a serious difficulty to ecologists wishing to study them in the field. Due to their secretive behaviour and cryptic coloration, they are generally difficult to find, and because of their solitary lifestyle, they are usually only found as isolated individuals. Whereas turtles, lizards, and crocodilians lend themselves easily to marking methods, snakes are the most difficult reptiles to mark effectively for field identification. Nevertheless, researchers have ingenuously developed a variety of techniques for marking snakes. No less than twenty different methods have been used with more than fifty snake species (Appendix). These techniques fall into several categories: 1) scale-clipping; 2) tagging; 3) painting/coloring; 4) tattooing, 5) branding; 6) recording of integumentary and scale patterns; 7) incising; and radiotelemetry. These are summarized in a subsequent section.

An ideal marking method for snakes should satisfy most of the following criteria, as enumerated by Lewke and Stroud (1974) and Ricker (1956). It should: 1) be as free of stress and pain as possible; 2) not affect the mortality rate of the animals; 3) afford minimal opportunity for infection; 4) not affect behaviour; 5) not inhibit normal movement; 6) not inhibit the shedding process; 7) be permanent or at least long-lasting; 8) be easily read to identify accurately individuals; 9) be adaptable to all sizes of animals; 10) be easily employed in the field or laboratory situation; and 11) involve equipment and materials that are easily made or obtainable at a minimal cost.

Discussions of preferred methods of marking can be found in Brown and Parker (1976b), Clark (1971), Ferner (1979), Fitch (1949A; 1987), Kroll et al. (1973), Lewke and Stroud (1974), McGinnis (1967), Pendlebury (1972), Pough (1970), Reinert and Cundall (1982), Schmidt and Davies (1941), Spellerberg and Prestt (1978), Swingland (1978), Turner (1977), Weary (1969) and Woodbury (1948; 1956).

Considering the aforementioned references, enough information is available for ecologists to choose an effective tagging method for their investigations according to their budget and the hypotheses to be investigated. To permit ready access and evaluation of the different methodologies, I offer the following summaries of the various marking techniques along with the pertinent literature.

MARKING PROTOCOLS

Scale-Clipping

Subcaudal clipping (1).

The first method ever used to mark snakes for an ecological study was devised by Blanchard and Finster (1933). It consisted of completely removing three subcaudal scales with a pair of sharp scissors, using a code to designate those scales clipped. By counting caudad from the cloacal plate, and placing the left and right subcaudals on the respective sides of a dash, a typical code of "2,5-4" represents the second and fifth subcaudal on the left and the fourth subcaudal on the right as being marked.

This method or variations thereof have been used, at least until the recent advances in radiotelemetry, for the past fifty years by the overwhelming majority of ecologists. Published reports indicate that close to 50,000 snakes have been marked by subcaudal clippings. It has been employed in studies by Blaesing (1979), Blanchard et al. (1979), Carpenter (1952), Fitch (1949b, 1958), Fitch and Fleet (1970), Fitch and Glading (1947), Fukada (1959; 1960; 1978), Parker (1974), Reynolds and Scott (1982), Siebert (1950), Siebert and Hagen (1947), Test et al. (1966), and Tinkle (1957).

Although Blanchard and Finster (1933) warned that scales only partially removed would regenerate through time, subsequent studies demonstrated that certain species have the ability to completely regenerate even entirely removed subcaudals. Calström and Edelstam (1946) were the first to demonstrate this, and further noted how difficult and harmful subcaudal clipping was with juveniles. Conant (1948) described a zoo specimen of Elaphe obsoleta in which regeneration obliterated the original marks after five years. Regeneration was further verified by Fitch (1982), who studied twelve species of snakes over a 30 year period (22,000 individuals marked). He showed that regeneration of clipped scales was more rapid and complete in Elaphe obsoleta than in any other species that he studied. After a few years, many clipped scales were indistinguishable from natural injuries (Fitch, 1963a). Other species also possess good regeneration. For example, Diadophis punctatus are scarred by bites from predators, and these scars are indistinguishable from those of clipping after two or three years (Fitch, 1975). Similarly, Fitch (1963b) observed regeneration in some Coluber constrictor after two or three years.

Ventral clipping (2).

Woodbury (1951) was the first to use ventral scales. He clipped a portion of the ventral scale on either side, designating the right side with a capital letter and the left side with a small letter. Scales were lettered cranial from the anal plate, and sex was indicated by M or F. Thus a code combination of "DaM" referred to a male snake with a clipped fourth ventral on the right and the first ventral on the left.

Brown and Parker (1976b) modified Woodbury's methodology to fulfill the eight criteria of Lewke and Stroud (1974) for a good marking method. The modified method is claimed to be satisfactory for at least four years and was used by Brown (1973), Brown and Parker (1974; 1982), Henderson et al. (1980), Jacob and Painter (1980) and Parker and Brown (1973; 1974a-b). The Brown and Parker protocol remove half of a ventral in combination of one to three clips with a total of 989 unique combinations. Ventrals are counted anteriorly from the anal plate on the left side in units from 1-9; on the right side they are counted in series of 10's (10-90), 100's (100-900), and 1000's (if necessary). A snake whose belly was clipped on the left side of ventral nine and the right side of ventral three and eleven would be number 239. Regeneration does not appear to occur as readily in the ventrals as in the subcaudals. No ill effects were noticed in markings of more than 1000 Coluber constrictor (Brown, 1973). This methodology appears superior for scale-clipping, although some ambiguity may occur such as misidentification of individual marks (Fitch, 1987).

Ventral and subcaudal clipping (3).

Fitch (1958, 1960) proposed a combination of subcaudals and ventral scale clippings. The first twenty subcaudals allow 362 unique combinations. A typical code of "U 5L 2R" represents markings on the fifth left and the second right subcaudals. Ventrals are used to indicate the next series of 362 marks. Counting cranial from the anal plate, "G 1L" designates the first ventral on the left or "G 1R" the first right ventral, etc. Fitch employed this technique in his later studies (1963a,b; 1965; 1975; 1982), and it was also used by Clark (1970) and Platt (1969).

Another variant of scale clipping was devised by Clark (1971). Only two marks are necessary with the designation of a basal starting point. For example, "BL-2L 3R" indicated a base mark on the left and the subsequent ventrals second left and third right are marked. To improve reliability, a scale is removed from the first dorsal scale row immediately adjacent to the basal marked ventral.

Henderson (1974), Lang (1969), and Saint-Girons (1964) used combined clipping methodologies, but did not provide full details on the numbering protocol.

The disadvantages of all clipping methods are the time required to mark each snake, the frequently drawing blood and possibility of infection, and the difficulty in marking small snakes (Weary, 1969).

Most recent researchers favor the combined ventral-subcaudal methodology of Prestt (1971) and Brown and Parker (1984). This method has been used successfully in studies on the population dynamics of colubrids and viperids (e.g. Brown and Parker, 1984; Feaver, 1977; Parker and Brown, 1974 a,b, 1980; Prestt, 1971; Spellerberg and Phelps, 1977).

Tags

Metal tags (4).

Hirth (1966) was the first to attach tags to snakes. He used uniquely numbered stainless steel tags clamped to the corner of the mouth. Fortunately, this location has not been used subsequently. Voris et al. (1983) attached numbered metal tags to the tail tips of sea snakes. He discontinued the tagging when it became apparent that the tags collected debris, wore holes in the tails, and were often lost.

Plastic plugs (5).

Pough (1970) described the use of colored plastic plugs or serially numbered metal tags attached to the tail via a buttoner device. The technique is presumably a quick and permanent method of marking snakes. The tags were partially inserted into the caudal musculature through the lateral subcaudal scales. Those inserted into ventral scales fell off. Plugs cannot be attached to snakes smaller than 250 mm SV. During Pough's studies there was no evidence of infection nor any indication that the plug interferes with shedding or locomotion.

Plastic disk (6).

Pendlebury (1972) initiated the attachment of colored disks to rattlesnake's rattles. A pair of disks are sewn on either side of the basal rattle segment. With the use of a pair of disks in ten color combinations 100 specimens can be uniquely marked. Two pairs of disks on each snake allows 10,000 unique combinations. This method, although limited to studies of rattlesnakes, has been used successfully by JACOB and Painter (1980), Reinert and Kodrich (1982), Stark (1984) and Brown et al. (1984).

Plastic tag/colored beads (7).

Thin, yellow plastic tags were successfully used by Voris et al. (1983) to mark sea snakes; these tags were embedded subcutaneously in front of the tail on the right side of the body. Similar red tags hooked through the tail were found to be less valuable. Hudnall (1982) suggested the use of colored beads to mark snakes.

Radioactive wire (8).

Barbour et al. (1969) first used a radioactive wire to mark snakes. A thin wire is inserted subcutaneously in the tail via a modified hypodermic syringe. The wires were radioactive Cobalt (Co^{60}) with a 50-80 microcurie dose. Hirth et al. (1969) and later Fitch (fide Ferner, 1979) used radioactive Tantalum (Ta^{182}) wire with 400 microcurie amounts. Snakes so marked can be located nine meters away when on the surface and three meters away when 30 cm below the surface. Tantalum and Cobalt have been the preferred source of radiation. Cobalt has a more powerful gamma radiation with a half-life of 5 years, whereas Tantalum has a half-life of only 6 months. The disadvantages of radioactive tags are that they may injure and eventually kill the snakes carrying them. Furthermore they are readily shed and then become hazardous to other animals or humans (Fitch, 1987). Radioactive tags are preferable to radiotransmitters in smaller snakes. The use of radioactive tags (and telemetry) are reviewed in Spellerberg and Prestt (1978), Swingland (1978) and Ferner (1979).

The major problem for the effective use of external tags is the choice of an adequate attachment site. The elongated, limbless body of snakes offers no satisfactory attachment sites. The possible sites are the free edges of the anterior ventrals, the cloacal plate, the lower jaw, or the base of the rattle in rattlesnakes. The tags must be attached in a manner to avoid interference with normal behavior and to reduce abrasion or loss of the tags.

The aforementioned tags can be stitched to the skin with heavy thread, thin wire or nylon. Utmost care must be exercised when attaching tags to avoid injury. For example, pinched skin undergoes necrosis thereby modifying the snake's normal behavior and increasing the likelihood of tag loss (Fitch, 1987).

Painting/coloring

Brush paint (9).

Fitch (1960) painted red, yellow, orange, and blue enamel marks on snakes to check on their shedding cycle. Pough (1966) used

quick drying waterproof paint to number the basal rattle segment of rattlesnakes. This method has been further altered for rattlesnakes by Brown et al. (1984). The head and neck of snakes were painted by Parker (1976) for field identification.

Spray painting (10).

Henderson et al. (1981) used rapid-drying fluorescent spray paint (non-toxic) to mark arboreal snakes. Three quarters of the animals spray-painted were later observed. Plummer (1985) also used spray paint to mark Opheodrys.

Water color pencil (11).

Stebbins (1966) suggested the use of Mongol water color pencils to temporarily mark specimens for field observations.

The preceding three methods produce temporary marking, nonetheless they facilitate the recognition of individuals without recapture in a variety of field situations. The marks are lost with the shed exuvia or through wear. As Fitch (1987) points out, these methods have additional useful applications, such as for group markings of snakes to obtain capture-recapture ratios that can serve for a Petersen index census. In population studies based on permanent marking systems, these temporary marks provide the best method for investigating ecdysis cycles.

Most species of snakes however are too secretive for regular or long-term observations, so the short duration of paint marking is of little use. Another major disadvantage is that the colored marks render marked individuals more conspicuous to visually oriented predators, hereby distorting the capture-recapture ratio (Fitch, 1987).

Tattooing

Battery powered tattooing (12).

Imler (1945) was the first researcher to tattoo snakes with a battery-operated unit. He tattooed the venter of bullsnakes with a special code number. This method was subsequently redescribed by Woodbury (1948, 1951) for marking rattlesnakes at a den. Using a portable tattooing outfit and India ink, he tattooed large numbers of snakes on their throats, subcaudals and/or ventrals, and areas lacking pigments. Disadvantages include the tendencies of the tattoos to fade and the necessity of tattooing unpigmented or lightly pigmented areas.

Electric powered tattooing (13).

Weary (1969) improved the tattooing method by employing an

inexpensive pyrographic needle (50 W unit operated from a 110 volt source) or a soldering pencil (powered by a 12 volt battery) to burn/scar scales without drawing blood.

Tattooing is more labor-intensive than scale clipping and usually involves the transport of bulky equipment. Also a major drawback is that during marking, there is a high risk of injury and even death if the needle penetrates too deeply and pierces the body cavity. Fitch (1987) points out two additional problems limiting the widespread use of these techniques. First, in darkly pigmented individuals and species of snakes, the numbers are difficult to read accurately and second, especially in snakes marked as juveniles the ink spreads and sometimes blurs the numbers.

Branding

Hot (flame) branding (14).

Clark (1971) described the use of a heat source, a Bunsen burner in the laboratory or a small propane torch in the field, to brand numbers on snakes by heating a branding wire of 20% chromium/80% nickel (Chromel A) and applying it to the scales.

Cold (freeze) branding (15).

Lewke and Stroud (1974) employed a superchilled branding instrument of copper wire to mark snakes. The technique was devised by Farrel (1966) for use with livestock. Of the three coolants tested (dry ice and 95% ethyl alcohol, Freon 12, and Freon 22), dry ice and alcohol was the most successful. Chromatophores are destroyed by quick-freezing the skin surface. The brand appears as a white area (Fitch, 1987).

Although this method is a good field-marking technique, a disadvantage is that the mark is not evident until after ecdysis and there is a minimal size limit of the branding iron used. Branding shares the same problems and disadvantages with tattooing. It is labor-intensive, sometimes causes injuries and may be difficult to read with heavily pigmented snakes.

Recording integumentary and scale patterns

Natural marks (16).

Calström and Edelstam (1946) employed the innovation of recording a snake's color pattern because each has a unique pattern. They photographed or sketched the fourth through eighth

ventrals (from anterior end) of Natrix natrix. These patterns were sufficient, in conjunction with other scutellation data to reidentify individuals. Snakes with distinctive skin patterns like Coronella and Vipera were treated in the same manner by recording their dorsal patterns. The use of this technique was considered feasible for Pelamis by Kropach (1973). Fitch (1987) suggests a methodology for recognition of individual Agkistrodon contortrix based on the arrangement of complete and incomplete "hourglass" marks on the body.

Scale formulae (17).

Blanchard and Finster (1933) mentioned an unsuccessful attempt to recognize individuals with scutellation data. However, Fukada (1978) described recording the arrangement of temporal scales in Elaphe climacophora as an adjunct to scale-clipping.

Both of these pattern-recognition techniques are labor intensive in recording and recognizing specific patterns and furthermore prone to reading errors. As Fitch (1987) suggests, natural marks should be noted and used to confirm the identification of individuals based on other marks or tags.

Incisions

Tail-notching (18).

In Kropach's unpublished dissertation (1973), he described a tail notching protocol for seasnakes. He (Kropach, 1975) later elaborated on this technique.

Radiotelemetry

Fitch (1987) recommends two fundamental criteria that should be met for the use of radiotransmitters. The transmitters should be small enough not to burden the snake during its normal activities such as feeding and locomotion, yet large enough to have a sufficient transmission range for easy localization.

Oral insertion (force fed) (19).

McGinnis and Moore (1969) pioneered the use of radiotelemetry in snake studies, adapting the technique used for lizards by McGinnis (1967). They force-fed a Boa constrictor with a miniature temperature sensitive transmitter with a range of 100 meters. Various types of radio transmitters, antennae and battery sources have been used subsequently in snake studies. Transmitters are coated with paraffin/beeswax mixture or silicone, and are either palpated into the animal's stomach or sewn into a food item and

then force-fed. Battery life ranges from a few days to several months. Orally inserted radiotransmitters have been used by Brown and Parker (1976a), Brown et al. (1982), Fitch and Shirer (1971), Galligan and Dunson (1979), Hammerson (1979), Henderson et al. (1976), Jacob and McDonald (1975), Jacob and Painter (1980), Johnson (1972), Landreth (1973), Montgomery and Rand (1978), Moore (1978), Nickerson et al. (1978), Osgood (1970), Parker and Brown (1972), Reinert (1981) and Reinert and Kodrich (1982).

Some disadvantages of this method are that the effective range of the transmitter is small, the transmitter may be regurgitated, and the transmitter in the stomach may cause behavioral changes in the snake. Fitch and Shirer (1971) tied a string around the body and sewed it to a ventral scale to prevent their snakes from losing the transmitters. Early studies demonstrated that the beeswax covering are digested by the snake, causing the transmitter to malfunction.

Surgical insertion (20). In order to alleviate the above mentioned problems, Fitch and Shirer (1971) surgically implanted transmitters into the abdomens of snakes. Brown and Parker (1976a) encapsulated their transmitters in polyethylene and inserted them abdominally with minimal bleeding. The sutures healed completely within two weeks. Other workers using this method include Brown and Parker (1982), Henderson et al. (1980), Jacob and Painter (1980). Reinert and Cundall (1982) improved the transmission of radiotelemeter by the implantation of both the transmitter and the antenna, the latter subcutaneously. Their transmitters had a maximum range of 1.5 km, a vast improvement over previous techniques. Further examples of studies using this technique are those by Reinert et al. (1984) and Madsen (1984).

Surgical implantation method involves anesthetizing the snake and inserting a foreign object into the body cavity. Although the healing process and fibrotic encapsulation may be rapid, behavioral alterations may occur (Fitch, 1987).

During the past two decades, the quality, longevity and distance of the emitted signals have been ameliorated. In conjunction with better radiotelemetry units, more efficient computerized radio-telemetric systems are now available to monitor free-ranging snakes (Stanmer, 1988). Furthermore, the implantation techniques are more refined (Weatherhead et al., 1984).

Fitch (1987) suggests that radiotelemetry may be the choice technique for most ecological research involving the marking of snakes. It allows for quick and effective monitoring of individuals in open terrain. In closed biotopes such as swamps, subterranean

habitats or rainforests, the situation is less favorable.

SUMMARY

Ecologists have a variety of different techniques at their disposal for the marking of snakes for field studies. With the sophistication now present in radiotelemetry, snakes can be tracked over long distances and recordings can be made for lengthy periods of time, such as overwintering in hibernacula.

The following appendix and bibliography identify the various published marking techniques and the primary literature sources.

The most widely used method for marking snakes may remain the ventral and subcaudal scale clipping. It is permanent, cost-efficient, and it allows the unique marking of each individual in a population. This method also has no permanent ill effect on the marked animals.

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Appendix - This list is a partial overview at best of the species of snakes that have been tagged for field studies. Many more species have been marked in Masters theses and Doctoral dissertations as well as other publications that are not listed in the zoological records. See numbers in text for the identification of methods.

TAXON

METHODS USED

FAMILY BOIIDAE

<u>Boa constrictor</u>	19
<u>Morelia spilotes</u>	19

FAMILY COLUBRIDAE

<u>Carphophis amoenus</u>	8
<u>Carphophis vermis</u>	1,3
<u>Coluber constrictor</u>	1,2,3,4,8,12,19,20
<u>Coronella sp.</u>	16
<u>Coronella austriaca</u>	3
<u>Diadophis punctatus</u>	1,2,3,8,17
<u>Elaphe climacophora</u>	1
<u>Elaphe obsoleta</u>	1,3,19
<u>Elaphe quadrivirgata</u>	1
<u>Elaphe subocularis</u>	1
<u>Helicops angulatus</u>	19
<u>Heterodon nasicus</u>	3
<u>Heterodon platyrhinus</u>	3
<u>Lampropeltis calligaster</u>	1,3,19
<u>Lampropeltis triangulum</u>	1,2,20
<u>Leptophis depressirostris</u>	19
<u>Liophis reginae</u>	1
<u>Masticophis lateralis</u>	19
<u>Masticophis taeniatus</u>	2,3,4,8,9,12,19
<u>Natrix natrix</u>	16,17,20
<u>Nerodia fasciata</u>	19
<u>Nerodia sipedon</u>	1,3,19
<u>Nerodia taxispilota</u>	19
<u>Opheodrys aestivus</u>	10
<u>Opheodrys vernalis</u>	1
<u>Oxybelis aeneus</u>	3
<u>Pituophis melanoleucus</u>	1,2,3,12,19
<u>Rhabdophis tigrinus</u>	1
<u>Spilotes pullatus</u>	19
<u>Storeria dekayi</u>	1,3,8

<u>Storeria occipitomaculata</u>	1
<u>Thamnophis butleri</u>	1
<u>Thamnophis elegans</u>	1
<u>Thamnophis ordinoides</u>	1
<u>Thamnophis radix</u>	1
<u>Thamnophis sirtalis</u>	1,3,5,19
<u>Uromacer catesbyi</u>	10
<u>Uromacer oxyrhynchus</u>	10

FAMILY HYDROPHIIDAE

<u>Enhydrina schistosa</u>	4,7
<u>Laticauda colubrina</u>	3
<u>Pelamis platurus</u>	18

FAMILY VIPERIDAE

<u>Agkistrodon contortrix</u>	1, 3, 9, 16,19
<u>Bothrops atrox</u>	19
<u>Crotalus atrox</u>	1, 9, 19
<u>Crotalus cerastes</u>	19
<u>Crotalus horridus</u>	1, 2, 3, 19, 20
<u>Crotalus mitchelli</u>	19
<u>Crotalus molossus</u>	1, 9
<u>Crotalus scutulatus</u>	1, 9
<u>Crotalus viridis</u>	1, 2, 6, 8, 12, 19, 20
<u>Sistrurus catenatus</u>	6, 19
<u>Trimeresurus flavoviridis</u>	19,20
<u>Vipera sp.</u>	16
<u>Vipera berus</u>	20

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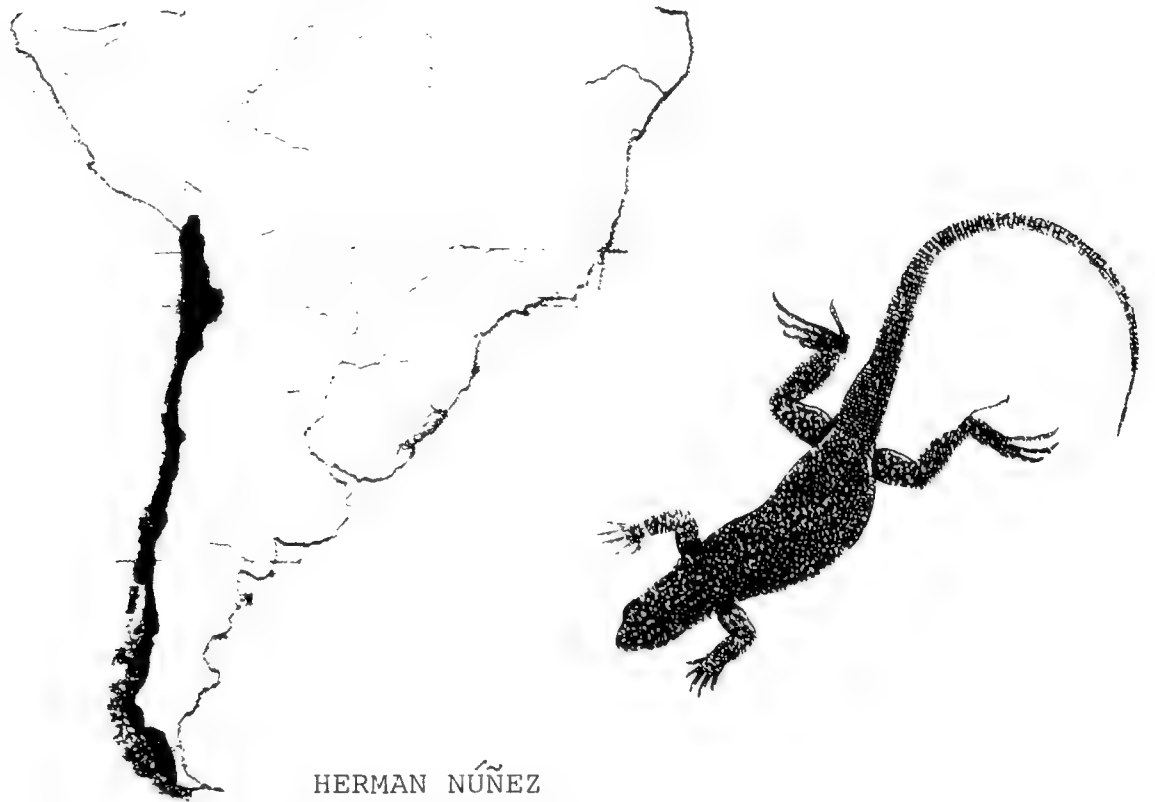
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GEOGRAPHICAL DATA OF CHILEAN
LIZARDS AND SNAKES IN THE
MUSEO NACIONAL DE HISTORIA NATURAL
SANTIAGO, CHILE



HERMAN NÚÑEZ

Sección Zoología
Museo Nacional de Historia Natural



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INTRODUCTION

The herpetological collections of the Museo Nacional de Historia Natural (MNHNC) contains about 3500 amphibians and reptiles. Nearly 90% of the specimens are lizards, and of these, most are Liolaemus, the most diversified member of the Chilean herpetofauna. The specimens of Liolaemus derive mainly from "central Chile", i.e., the area between the city of La Serena and the Biobio River. Both northern and southern Chile are relatively unexplored; thus, the taxonomy and composition of these herpetofaunas is less well known.

The distribution of Chilean lizards and snakes has not received much attention (however, see Valencia & Velosa 1981, Velosa & Navarro 1988) beyond the general information provided by Peters & Donoso-Barros (1970) and Donoso-Barros (1966, 1970). The latter do not provide sufficient information to define the exact ranges, especially lacking altitudinal data, and such data are critical for Chile, which is a narrow country bordered on the east by mountains and a complex system of ridges and valleys with numerous isolated populations of lizards and snakes.

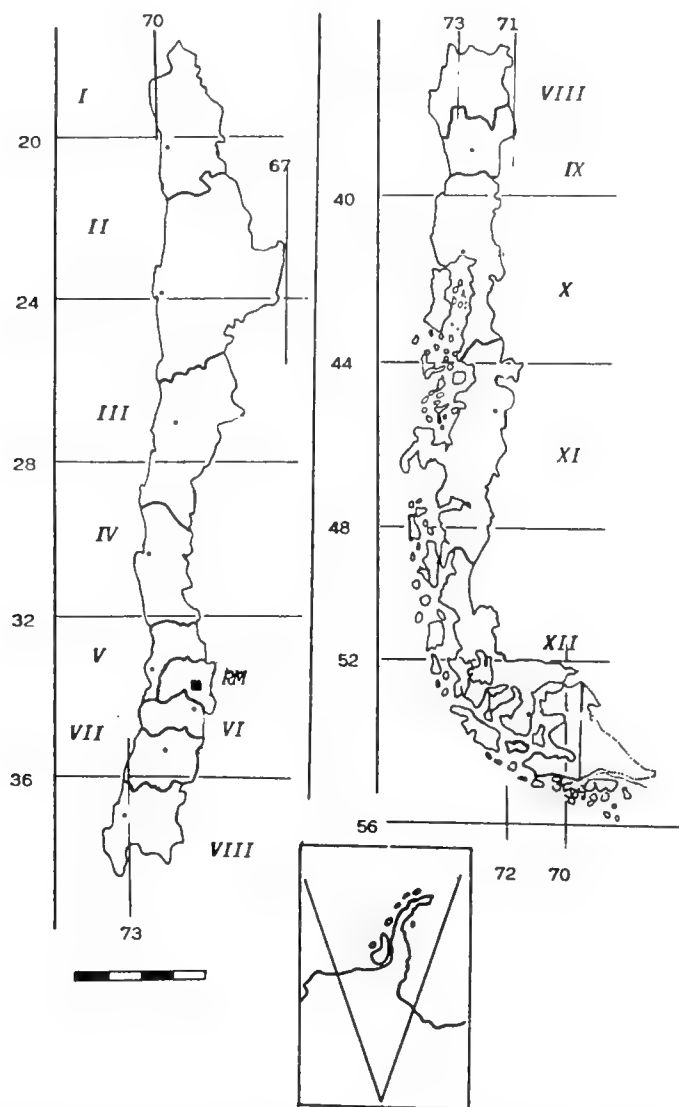
Accurate distributional data are essential in clarifying biogeographical problems, ecological consequences of human intervention, and physiological aspects of latitudinal and altitudinal adaptations. Since the MNHNC collection has 76 species of lizards and snakes, representing about 75% of Chilean species, the distributional data of these specimens provide a good baseline for such investigations. My goal has been to provide accurate locality data for the Chilean species of lizards and snakes vouchered by one or more MNHNC specimens. Uncertainties of taxonomy or locality are indicated by numbers in parentheses; these items are discussed at end of the taxonomic section.

Taxa are listed alphabetically by family, genus, species, and finally by locality. Some localities are too small to appear in the **Indice de Nombres Geográficos de Chile** (Instituto Geográfico Militar, no date) or in the **Ruiso Patrón Index** (1942). Similar problems arise when the locality data are too broad (e.g., Magallanes); in these cases, I mark the data with *sensu lato* and report only the administrative region. Latitude and longitude are south and west, respectively, and the first two digits are degrees, the second two are minutes. Altitude is in meters above sea level. The final column provides a major city, town, or other geographic entity near to the specimen's origin.

Chile contains a Metropolitan Region (RM) and twelve

administrative regions: Primera Región, Tarapacá; Segunda Reg., Antofagasta; Tercera Reg., Atacama; Cuarta Reg., Coquimbo; Quinta Reg., Aconcagua; Sexta Reg., Libertador General Bernardo O'Higgins; Séptima Reg., Maule; Octava Reg., Biobio; Novena Reg., Araucanía; Décima Reg., Los Lagos; Décimo Primera Reg., General Carlos Ibáñez del Campo; Décimo Segunda Reg., Magallanes y Antártica Chilena. These regions are identified by number in the Reg. column.

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Map of Chile with the administration regions designated by Roman numerals (listed in Introduction). Scale, each segment = 100 km.

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COLUBRIDAE

Phylodryas chamissonis

	Lat.	Long.	Alt.	Reg.	
Aucó	3131	7106	500	4	14 km NE Illapel
Camarico	3604	7225	150	7	Chipa tributary+ Cauquenes river
Cartagena	3333	7136	60	5	3 km NE San Antonio
Cavilolén, cuesta	3146	7119	700	4	24 km NE Los Vilos
Cerrillos	3330	7046	470	RM	15 km SW Santiago
Chena	3335	7044	950	RM	4.5 km W San Bernardo
Cobquecura	3608	7249	0	8	1 km W Cobquecura
Colina	3320	7048	470	RM	6 km W Lampa
Concón	3256	7132	80	5	15 km S Quintero
Concepción	3649	7303	100	8	Capital 8th Region
Coquimbo	2957	7120	50	4	11 km S La Serena
Culimó	3204	7114	390	4	5 km W Tilama
del Flaco, Baños	3458	7028	1520	6	70 km SE San Fernando
Fray Jorge, Parque Nacional	3039	7140	0	4	14 km W Salala
Km 300 Panam. Norte	ca3100	ca7140	100	4	South of Maitencillo
La Reina, Cerros de	3354	7031	1600	RM	Ravines E Santiago
Lo Arcava	3341	7035	650	RM	8 km Puente Alto
Lo Barnechea	3321	7030	816	RM	14 km NE Santiago
Los Vilos	3154	7131	5	4	0.5 km S Los Vilos
Macul, Quebrada de	3329	7028	500- 1800	RM	18 km SE Santiago
Melipilla	3341	7113	170	RM	Melipilla town
Peñalolén	3328	7032	800	RM	11 km SE Santiago
Peñuelas, Lago	3309	7132	343	5	20 km SE Valparaíso
Pichidangui	3208	7132	5	4	4 km SW Quilimarí
Pudahuel	3321	7052	490	RM	13 km NW Santiago
Quintero	3247	7132	70	5	30 km N Quillota
Rapel	3356	7144	40	6	42 km SW San Antonio
San Antonio	3335	7137	25	5	Capital San Antonio Province
San Luis de Macul	3332	7033	600	RM	13 km SE Santiago
San Pedro, Laguna	3650	7305	13	8	4 km SW Concepción
Santa Lucía, Cerro	3327	7038	600	RM	Capital of Chile
Santa Rita	3340	7031	650	RM	8 km SW Puente Alto
Santiago	3327	7038	600	RM	Capital of Chile
Santo Domingo, Rocas de	3328	7137	15	5	6 km S San Antonio
Taco de Lampa	3314	7055	500	RM	Polpaico tributary + Chacabuco
Vilches Alto	3536	7105	1150	7	9 km SE Vilches
<i>Tachymenis chilensis chilensis</i>					
del Flaco, Baños	3458	7028	1520	6	70 km SE San Fernando
El Ingenio	3346	7015	1200	RM	50 km SE Santiago

<i>T. ch. chilensis</i> (Cont.)	Lat.	Long.	Alt.	Reg.	
Frav Jorge, Parque Nacional	3039	7140	0	4	14 km W Salala
Los Cipreses, Valle del río	3425	7026	900	6	Tributary of Cachapoal river
Macul, Quebrada de	3329	7028	500– 1800	RM	18 km SE Santiago
Morado, Valle del	3347	7002	2800	RM	North tributary of El Volcán river
Putaendo	3237	7043	813	5	13 km N San Felipe
Ramón, Cerro de	3330	7026	3050	RM	25 km E Santiago
Talinay	3050	7137	762	4	9 km NW Peñablanca
Vilches, Alto	3536	7105	1150	7	9 km SE Vilches

GEKKONIDAE

<i>Phyllodactylus gerrhopygus</i>					
Antofagasta (<i>sensu lato</i>)	2338	7042	30	2	Capital 2nd Region
Tocopilla	2250	7012	5	2	Capital Tocopilla province

POLICHRIDAE

<i>Diplolaemus bibroni</i>					
Chile Chico	4633	7140	100	11	80 km S Balmaceda
<i>Diplolaemus darwini</i>					
Ultima Esperanza	5130	7300	100	12	Coastal of Seno Ultima Esperanza
<i>Pristidactylus alvaroi</i>					
El Roble, Cerro	3538	7102	2200	5	25 km SE La Calera
<i>Pristidactylus torquatus</i>					
Bellavista, Sierras de "Bosque Valdiviano" (<i>sensu lato</i>)	3436	7032	3500	6	41 km E San Fernando
Valdivia (<i>sensu lato</i>)	10
Vilches Alto	3949	7314	10	10	Capital of 10th Region
	3536	7105	1150	7	9 km SE Vilches
<i>Pristidactylus valeriae</i>					
Cantillana, Pies del Cerro	3358	7058	600	6	24 km W San Francisco de Mostazal
<i>Pristidactylus volcanensis</i>					
El Volcán	3349	7010	1416	RM	60 km SE Santiago

TEIIDAE

<i>Callopiastes palluma palluma</i>					
Almendrillo	3202	7035	1650	4	39 km NE Petorca

<i>C. p. palluma</i> (Cont.)	Lat.	Long.	Alt.	Reg.	
Aucó	3131	7106	500	4	14 km NE Illapel
Blanco, Cerro	3309	7049	690	RM	12 km SE Tiltil
Colina	3320	7048	470	RM	6 km W Lampa
Cuesta La Dormida	3303	7102	800	RM	37 km E Quilpué
El Roble, Cerro	3258	7102	2200	5	25 km SE La Calera
Graneros	3404	7044	475	6	12 km N Rancagua
La Serena (sensu lato)	2954	7115	50	4	470 km N Santiago
Lipangue (Pudahuel)	3321	7052	490	RM	13 km NW Santiago
Macul, Quebrada de	3329	7028	500- 1800	RM	18 km SE Santiago
Palqui	3044	7057	500	4	5 km S Monte Patria
Pichidanguí (sensu lato)	3208	7132	5	4	4 km SW Quilimarí
Polpaico	3309	7047	580	RM	15 km SE Tiltil
San Luis de Macul	3332	7033	600	RM	13 km SE Santiago
Taco de Lampa	3314	7055	500	RM	Polpaico tributary + Chacabuco

Callopiestes palluma manni

Copiapó (sector Loreto)	2722	7020	450	3	Capital 3d Region
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TROPIDURIDAE

Ceiolaemus fabiani

San Pedro de Atacama	2255	6812	2450	2	3 km SE Vilama
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Liolaemus alticolor

Chungará	1814	6909	4570	1	20 km S Caquena
Isluga (Suricayo)	1915	6843	4000	1	3 km N Alsore

Liolaemus altissimus altissimus

Cantillana, Cerro (1)	3358	7058	600	6	24 km W San Francisco de Mostazal
El Teniente, Mina (1)	3404	7022	2110	6	37 km NE Rancagua
Farellones (sensu lato)	3320	7019	2308	RM	35 km E Santiago
Lagunillas	3337	7018	2000	RM	35 km SE Santiago
Maule, Laguna del (1)	3604	7030	2100	7	11 km W Paso Pehuenche
Ramón, Cerro de	3330	7026	3050	RM	25 km E Santiago
San Francisco, Valle del río	3320	7022	2500	RM	North tributary of Mapocho river
Sewell (1)	3405	7023	2600	6	31 km NE Rancagua

Liolaemus altissimus moradoensis

Colina, Baños de	3356	7000	2500	RM	SE tributary of Volcán river
El Yeso, Embalse	3341	7007	3370	RM	NE tributary of Maipo river
Morado, Valle del	3347	7002	2800	RM	North tributary of El Volcán river
Negra, Laguna	3339	7008	2200	RM	60 km SE Santiago

Liolaemus bibroni

Chile Chico	4633	7140	100	11	80 km S Balmaceda
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Liolaemus bisignatus

	Lat.	Long.	Alt.	Reg.	
Caldera, Playa Negra	2704	7049	19	3	Caldera town

Liolaemus buergeri

El Planchón (<i>sensu lato</i>)	3514	7034	3000	7	67 km SE Curicó
Maule, Laguna del	3604	7030	2100	7	11 km SW Paso Pehuenche
San Pedro (Vergara)	3510	7030	2200	7	Teno Lake + SW Nacimiento river

Liolaemus cf ceii

Las Damas, Río de	3455	7021	2000		
			3000	6	73 km E San Fernando

Liolaemus chiliensis

Algarrobo	3323	7141	30	5	2 km SW Algarrobo
Cartagena	3333	7136	60	5	3 km N San Antonio
Cobquecura	3608	7249	0	8	1 km W Cobquecura
Cuesta la Dormida	3303	7102	800	RM	37 km E Quilpué
Curamávida	3717	7244	700-		
			1200	8	2 km W Laja
El Belloto	3312	7114	630	5	26 km SE Quilpué
El Manzano	3335	7024	1068	RM	28 km S Santiago
El Pangue	3315	7112	600	RM	18 km NW Curacaví
El Teniente, Mina	3404	7022	1870	6	37 km NE Rancagua
El Volcán	3349	7010	1416	RM	60 km SE Santiago
La Pintana	3337	7039	620	RM	7 km SE San Bernardo
Leyda	3328	7127	190	5	12 km W Puangue
Macul, Quebrada de	3329	7028	500-		
			1800	RM	18 km SE Santiago
Pucón	3916	7159	280	9	73 km SE Temuco
Putendo	3237	7043	813	5	13 km N San Felipe
Santiago (Quinta Normal)	3327	7038	600	RM	Capital of Chile
Santo Domingo, Rocas de	3338	7137	15	5	6 km S San Antonio
Talca ("4 km E de La Mina")	3550	7047	959	7	15 km E Curillínque
Tulahuén	3101	7044	1096	4	33 km NE Combarbalá
Valparaíso (<i>sensu lato</i>)	3303	7107	194	5	Capital of 5th Province
Vilches Alto	3536	7105	1150	7	9 km SE Vilches
Zapallar	3133	7106	450	4	10 km NE Illapel

Liolaemus constanzae

San Pedro de Atacama	2255	6812	2450	2	3 km SE Vilama
Peine	2341	6802	2400	2	20 km E Laguna Brava

Liolaemus copiapoensis

Huasco (Playa grande)	2228	7131	25	3	Huasco town
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Liolaemus cristiani

El Peine, Cerro	3537	7102	2448	7	14 km E Vilches
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Liolaemus curis

	Lat.	Long.	Alt.	Reg.	
del Flaco, Baños	3458	7028	1520	6	70 km SE San Fernando
Las Damas, Río de	3455	7021	2000		
			3000	6	73 km SE San Fernando

Liolaemus cyanogaster cyanogaster

Cobquecura	3608	7249	0	8	1 km W Cobquecura
Neltume	3915	7213	225	9	3 km NE Villarrica
Villarrica	3916	7207	220	9	68 km E Temuco

Liolaemus darwini

Chile Chico	4633	7140	100	11	80 km S Balmaceda
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Liolaemus donosoi

Taltal, Quebrada de	2528	7025	750	2	Coastal range of Antofagasta
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Liolaemus dorbigni (=L. puritamensis)

Puritama	2243	6802	3550	2	40 km NE San Pedro de Atacama
Toconao	2311	6800	2475	2	8 km SE Zaparquilla

Liolaemus eleodori

Negro Francisco, Laguna del	2727	6913	4125	3	12 km W Portezuelo Astaburuaga
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Liolaemus fitzgeraldi

del Inca, Laguna	3250	7009	3500	5	42 km E Los Andes
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Liolaemus fitzingeri

Chile Chico	4633	7140	100	11	90 km S Balmaceda
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Liolaemus fuscus

Aucó	3131	7106	500	4	14 km NE Illapel
Blanco, Cerro	3309	7049	690	RM	12 km SE Tilttil
Caleu	3300	7100	1200	RM	11 km NE Tilttil
Culimó	3204	7114	390	4	5 km W Tilama
Fray Jorge, Parque Nacional	3039	7140	0	4	14 km W Salala
Illapel	3138	7110	350	4	50 km S Combarbalá
La Campana, Cerro	3257	7107	300	5	20 km SE La Calera
La Plata, Quebrada de	3329	7054	900	RM	Ravine at W of Santiago
Macul, Quebrada de	3329	7028	500–		
			1800	RM	18 km SE Santiago
Pichidangui	3208	7132	5	4	4 km SW Quilimarí
Ruinas	3105	7138	100	4	South of Maitencillo
San Luis de Macul	3332	7033	600	RM	13 km SE Santiago
Talca	3525	7139	90	7	Capital &th Region

Liolaemus gravenhorsti

	Lat.	Long.	Alt.	Reg.	
Santiago (<i>sensu lato</i>)	3327	7038	600	RM	Capital of Chile

Liolaemus hellmichi

Moreno, Cerro	2400	6953	1785	2	20 km NW Antofagasta
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Liolaemus hernani

del Flaco, Baños	3458	7028	1520	6	70 km SE San Fernando
Los Cipreses, Valle del Río	3425	7026	900	6	Tributary of Cachapoal river
Sewell	3405	7023	2600	6	31 km NE Rancagua

Liolaemus islugensis

Colchane	1917	6838	3850	1	2 km W Pisiga Chile
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Liolaemus kingi

Buitreras, Estación	ca5140	ca6940	100	12	"on the road to Río Gallegos" (<i>sic</i>)
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Liolaemus kuhlmani

Concón	3256	7132	80	5	15 km S Quintero
Coquimbo	2957	7120	50	4	11 km S La Serena
La Serena (<i>sensu lato</i>)	2954	7115	50	4	470 km N Santiago
Limarí	3040	7132	62	4	1 km N Salala
Los Vilos	3154	7131	5	4	0.5 km S Los Vilos
Quintay	3312	7142	40	5	20 km S Valparaíso
Quintero (relictual forest)	3247	7132	70	5	30 km NW Quillota
Totalillo	3004	7122	5	4	14 km W Guanaqueros

Liolaemus lemniscatus

Cartagena	3333	7136	60	5	3 km NE San Antonio
Clarillo, Valle del río	3341	7034	650	RM	South tributary of Maipo river
Cobquecura	3608	7249	0	8	1 km W Cobquecura
Concón	3256	7132	80	5	15 km S Quintero
Culimó	3204	7114	390	4	5 km W Tilama
El Belloto	3312	7114	630	5	26 km SE Quilpué
El Principal	3342	7034	670	RM	9 km S Puente Alto
El Volcán	3349	7010	1416	RM	60 km SE Santiago
Huentelauquén	3135	7132	25	4	35 km N Los Vilos
La Plata, Quebrada de	3329	7054	900	RM	Ravine at W Santiago
Las Palmas, Rancho	3301	7104	500	5	34 km E Quilpué
Llolleo, Laguna de	3336	7137	10	5	2 km N Llolleo
Los Cipreses, Valle del río	3425	7026	900	6	Tributary of Cachapoal river
Los Dominicos	3323	7031	784	RM	11 km NE Santiago
Macul, Quebrada de	3329	7028	500–1800	RM	18 km SE Santiago

<i>L. lemniscatus</i> (Cont.)	Lat.	Long.	Alt.	Reg.	
Pichidangui	3208	7132	5	4	4 km SW Quilimarí
Quintero	3247	7132	70	5	30 km N Quillota
Renca, Cerros de	3324	7042	903	RM	8.5 km NW Santiago
San Antonio	3335	7137	25	5	Capital San Antonio Province
Taco de Lampa	3314	7055	500	RM	Polpaico tributary + Chacabuco
Tagua-Tagua, San Vicente de	3426	7105	207	6	18 km NW San Fernando
Talca	3525	7139	90	7	Capital 7th province
Talca (4 km E de La Mina")	3550	7047	959	7	17 km E Curillinque
Vilches Alto	3536	7105	1150	7	9 km SE Vilches
<i>Liolaemus leopardinus leopardinus</i>					
El Teniente, Mina (1)	3404	7022	2110	6	37 km NE Rancagua
Farellones	3320	7019	2380	RM	35 km E Santiago
San Francisco, Valle del Río	3320	7022	2500	RM	North tributary of Mapocho River
<i>Liolaemus leopardinus ramonensis</i>					
Ramón, Cerro de	3330	7026	3050	RM	25 km E Santiago
<i>Liolaemus leopardinus spp.</i>					
El Yeso, Embalse (2)	3341	7007	3370	RM	NE tributary of Maipo river
Sewell (2)	3405	7023	2600	6	31 km NE Rancagua
<i>Liolaemus leopardinus valdesianus</i>					
Lo Valdés	3351	7003	1800	RM	South Tributary of El Volcán river
<i>Liolaemus lineomaculatus</i>					
Amorosa, Laguna	12	...
<i>Liolaemus lorenzmuelleri</i>					
La Laguna, Embalse	3013	7004	3275	4	53 km SE Guanta
<i>Liolaemus magellanicus</i>					
Balmaceda	4550	7138	20	11	120 km SE Puerto Aysén
Catalina, Bahía	5305	7050	0	12	Between Ensenada Laredo and Punta Arenas
Josefina, Caleta	5321	6934	0	12	100 km E Bahía Inútil
Dos Lagunas	12	...
Manantiales	12	...
Magallanes (<i>sensu lato</i>)	5300	7040	...	12	12th Región
Punta Arenas (<i>sensu lato</i>)	5310	7054	12	12	Capital of 12th Región
San Gregorio	5239	7013	64	12	North of Segunda Angostura
Ultima Esperanza (Laguna Azul)	5130	7300	100	12	Coastal of Seno Ultima Esperanza

Liolaemus maldonadae

	Lat.	Long.	Alt.	RM	
Los Molles	3043	7039	2600		
			2800	4	53 km E Ovalle

Liolaemus monticola monticola

Almendrillo	3202	7035	1650	4	39 km NE Petorca
Caleu	3300	7100	1200	RM	11 km NE Tiltil
El Roble, Cerro	3258	7102	2200	5	25 km SE La Calera
El Volcán	3349	7010	1416	RM	60 km SE Santiago
Farellones (Curva 20)	ca3320	ca7019	1800	RM	30 km E Santiago
Los Dominicos	3323	7031	784	RM	11 km NE Santiago
La Plata, Quebrada de	3329	7054	900	RM	Ravine at W Santiago
Macul. Quebrada de	3329	7028	500-		
			1800	RM	18 km SE Santiago
San Luis de Macul	3332	7033	600	RM	13 km SE Santiago

Liolaemus nigriceps (3)

Río Frío	2500	6904	1000	2	High plateau of Antofagasta
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Liolaemus nigromaculatus nigromaculatus

Caldera (playa Negra)	2704	7049	19	3	Caldera town
Copiapó, río Huasco	2722	7020	450	3	Capital 3d Region
Pan de Azúcar, Cerro	2807	7056	756	3	4 km N Cantera de Agua

Liolaemus nigromaculatus atacamensis

Antofagasta (11 km al norte)	2338	7043	30	2	Capital 2nd Region
Pan de Azúcar, Cerro	2807	7056	756	3	4 km N Cantera de Agua
Taltal	2524	7029	115	2	190 km S Antofagasta

Liolaemus nigroviridis campanae

El Roble, Cerro	3258	7102	2200	5	25 km SE La Calera
La Campana, Cerro	3257	7107	1900	5	20 km SE La Calera

Liolaemus nigroviridis minor

Colina, Baños de	3356	7000	2500	RM	SE tributary of El Volcán river
El Volcán	3349	7010	1416	RM	60 km SE Santiago
El Yeso, Embalse	3341	7007	3370	RM	NE tributary of Maipo river
Lo Valdés	3351	7003	1800	RM	South tributary of El Volcán river
Morado, Valle del	3347	7002	2800	RM	North tributary of El Volcán river
Morales, Baños de	3347	7005	2000	RM	North tributary of El Volcán River
Negra, Laguna	3339	7008	2200	RM	60 km SE Santiago

Liolaemus nigroviridis nigroroseus

	Lat.	Long.	Alt.	Reg.	
Chiuchiu	2220	6839	2520	2	3 km N Salado river
Licancabur, Volcán (on the road to Sulphur mine)	2250	6753	4000	2	11 km N Casablanca
Peine	2341	6802	2400	2	20 km E Laguna Brava
San Pedro de Atacama	2255	6812	2450	2	3 km SE Vilama
Talabre (4)	ca2311	ca6800	2500	2	8 km SE Zaparpilla

Liolaemus nigroviridis nigroviridis

Arrayán, Quebrada del	ca3321	ca7028	1800	RM	15 km E Santiago
Cantillana, Cerro (1)	3358	7058	2281	6	24 km W San Francisco de Mostazal
Farellones	3320	7019	2308	RM	35 km E Santiago
Lagunillas	3337	7018	2000	RM	35 km SE Santiago
Macul. Quebrada de	3329	7028	500–		
			1800	RM	18 km SE Santiago
Ramón, Cerro de	3330	7026	3050	RM	25 km E Santiago
Sewell (1)	3405	7023	2600	6	31 km NE Rancagua

Liolaemus nitidus

Aucó	3131	7106	500	4	14 km NE Illapel
Cartagena	3333	7136	60	5	3 km NE San Antonio
Colina	3320	7048	470	RM	6 km W Lampa
Culimó	3204	7114	390	4	5 km W Tilama
El Roble, Cerro	3258	7102	2200	5	25 km SE La Calera
El Volcán	3349	7010	1416	RM	60 km SE Santiago
Farellones	3320	7019	2380	RM	35 km E Santiago
Farellones (curva 20)	ca3320	ca7019	2380	RM	35 km NE Santiago
Fray Jorge, Parque Nacional	3039	7140	0	4	14 km W Salala
Illapel	3138	7110	350	4	50 km S Combarbalá
Lo Valdés	3351	7003	1800	RM	South tributary of El Volcán river
Macul, Quebrada de	3329	7028	500–		
			1800	RM	18 km SE Santiago
Pichidangui	3208	7132	5	4	4 km SW Quilimarí
Ramón, Cerro de (5)	3330	7026	3050	RM	25 km E Santiago
San Antonio	3335	7137	25	5	Capital San Antonio Province
Santo Domingo, Rocas de	3338	7137	15	5	6 km S San Antonio
Sewell	3405	7023	2600	6	31 km NE Rancagua
Talinay	3050	7137	762	4	9 km NW Peñablanca
Tulahuén	3101	7044	1096	4	33 km NE Combarbalá

Liolaemus ornatus

Colchane	1917	6838	3850	1	2 km W Pisiga Chile
Lampaipata	ca1917	ca6838	3850	1	Near Colchane

Liolaemus pantherinus

	Lat.	Long.	Alt.	Reg.	
Ollagüe	2113	6815	3700	2	350 km NE Antofagasta

Liolaemus paulinae

Calama, (las vegas)	2227	6856	2250	2	Capital El Loa Province
San Pedro de Atacama (6)	2255	6812	2450	2	3 km E Vilama

Liolaemus pictus chiloensis

Chiloé island (sensu lato)	10	3 km S Puerto Montt
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Liolaemus pictus pictus

Valdivia	3949	7314	10	10	Capital Valdivia province
Neltume	3915	7213	225	9	3 km NE Villarrica
Vilches Alto (1)	3536	7105	1150	7	9 km SE Vilches

Liolaemus platei platei

Aucó	3131	7106	500	4	14 km NE Illapel
Copiapó (sector Loreto)	2722	7020	450	3	Capital 3d Region
El Teniente	3100	7138	200	4	South of Maitencillo
Fray Jorge, Parque Nacional	3039	7140	0	4	14 km W Salala
Morrillos	3011	7133	5	4	5 km NE Guanaqueros
Tulahuén	3101	7044	1096	4	33 km NE Combarbalá

Liolaemus pseudolemniscatus

Aucó	3131	7106	500	4	14 km NE Illapel
Camión	3103	7139	100	4	Near Maitencillo town
Culimó	3204	7114	390	4	5 km W Tilama
El Teniente	3100	7138	100	4	South of Maitencillo
Illapel	3138	7110	350	4	50 km S Combarbalá
Ruinas	3105	7138	100	4	South of Maitencillo

Liolaemus schmidti

Lejía, Laguna	2329	6741	4350	2	18 km Angostura Pili
Antofagasta, Altiplano de	2	2nd region
Tara, Laguna	2259	6717	4350	2	18 km SE Laguna Blanca
Punta Negra, Salar de	2428	6854	2960	2	E of Pique San Carlos

Liolaemus schroederi

Cuesta la Dormida	3303	7102	800	RM	37 km E Quilpué
El Roble, Cerro	3258	7102	2200	5	25 km SE La Calera
El Teniente, Mina	3404	7022	2110	6	37 km NE Rancagua
Farellones	3320	7019	2380	RM	35 km E Santiago
La Campana, Cerro	3257	7107	300	5	20 km SE La Calera
La Disputada	3320	7002	2500	RM	North tributary of Mapocho river
Macul, Quebrada de	3329	7028	500- 1800	RM	18 km SE Santiago

<i>L. schroederi</i> (Cont.)	Lat.	Long.	Alt.	Reg.	
San Francisco, Valle del río	3320	7022	2500	RM	North tributary of Mapocho river
Santiago	3327	7038	600	RM	Capital of Chile
Tunca	3423	7106	200	6	7 km Peumo
Vilches Alto	3536	7105	1150	7	9 km SE Vilches
Villarrica (6)	3916	7207	220	9	68 km SE Temuco
<i>Liolaemus silvai</i>					
Copiapó	2722	7020	450	3	Capital 3d region
<i>Liolaemus tenuis punctatissimus</i>					
Parque Lota	3705	7309	20	8	30 km S Concepción
<i>Liolaemus tenuis tenuis</i>					
Algarrobo	3323	7141	30	5	2 km SW Algarrobo
Cartagena	3333	7136	60	5	3 km NE San Antonio
Clarillo, Valle del río	3341	7034	650	RM	South tributary of Maipo river
Culimó	3204	7114	390	4	5 km W Tilama
La Campana, Cerro	3257	7107	300	5	20 km SE La Calera
Las Palmas, Rancho	3301	7104	500	5	34 km E Quilpué
Macul, Quebrada de	3329	7028	500-		
			1800	RM	18 km SE Santiago
Pichidangui (cerro El Carmen)	3208	7132	800	4	4 km SW Quilimarí
San Antonio	3335	7137	25	5	Capital San Antonio Province
Santa Lucía Cerro	3327	7038	600	RM	Capital of Chile
Santiago	3327	7038	600	RM	Capital of Chile
Tagua-Tagua, San Vicente de	3426	7105	207	6	18 km NW San Fernando
Talca ("4 km E de La Mina")	3550	7047	959	7	15 km E Curillinque
Trapa-Trapa	3744	7113	1400	8	70 km E Santa Bárbara
Vilches Alto (1)	3536	7105	1150	7	9 km SE Vilches
<i>Liolaemus walkeri</i>					
Licancabur, Volcán (on the road to Sulphur mine)	2250	6753	4000	2	11 km N Casablanca
San Pedro de Atacama	2255	6812	2450	2	3 km SE Vilama
<i>Liolaemus zapallarensis zapallarensis</i>					
Fray Jorge, Parque Nacional	3039	7140	0	4	14 km W Salala
Pichidangui	3208	7132	5	4	4 km SW Quilimarí
Talinay Alto	3050	7137	762	4	9 km NW Peñablanca
<i>Phrynosaura audituvelata</i> (7)					
San Pedro de Atacama	2255	6812	2450	2	3 km SE Vilama
<i>Phrynosaura reichei</i>					
Atacama, Subcordillera de	3	3d region
Punta Negra, salar de	2428	6854	2960	2	E Pique San Carlos

Phymaturus flagellifer

	Lat.	Long.	Alt.	Reg.	
del Flaco, Baños	3458	7026	1520	6	70 km SE San Fernando
El Planchón	3514	7034	3000	7	67 km SE Curicó
Las Damas, Río de	3455	7021	1500		
			3000	6	73 km E San Fernando
Los Andes (8)	3250	7036	820	5	65 km N Santiago
Los Cipreses, Valle del río	3425	7026	900	6	Tributary of Cachapoal river

Tropidurus atacamensis

Antofagasta	2338	7024	30	2	Capital 2nd region
Copiapó (sector Loreto)	2722	7020	450	3	Capital 3d region
Moreno, cerro (at foot)	2400	6953	1785	2	20 km NW Antofagasta
Santa María, islote	2400	6953	10	2	Small islet near Cerro Moreno
Tocopilla	2250	7012	5	2	Capital Tocopilla province

Tropidurus theresioides (?)

Yalquincha, Quebrada de	2227	6856	2250	2	ca Calama town
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Velosaura jamesi

Chungará	1814	6909	4570	1	20 km S Caquena
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Notes

- (1) These lizards are atypical, although they belong to the nominal group.
- (2) Both differ from each other, as well as from the nominal form.
- (3) **Insertae sedis.**
- (4) These lizards (only two specimens) are rather atypical, they are included in this species only for convenience.
- (5) Exceptionally only one specimen was captured at the summit of Cerro de Ramón.
- (6) Uncertain locality.
- (7) This species was formerly described in the genus **Ctenoblepharys**, its inclusion into the genus **Phrynosaura**, demands the shift of the species name desinence in accordance with the new one (Art. 31, letter b, of the International Code of Zoological Nomenclature, 3d ed., 1985).
- (8) Very Uncertain locality.

Locality(Lat./Long./Alt./Region)

Species

Algarrobo (3323/7141/30/5/2 km SW Algarrobo)

Liolaemus chiliensis

Liolaemus tenuis tenuis

Almendrillo (3202/7035/1650/4/39 km NE Petorca)

Callopistes palluma palluma

Liolaemus monticola monticola

Amorosa, Laguna (.../.../.../12/...)

Liolaemus lineomaculatus

Antofagasta (2338/7024/30/2/Capital of 2nd region)

Liolaemus nigromaculatus atacamensis

Phyllodactylus gerrhopygus

Tropidurus atacamensis

Antofagasta, Altiplano de (.../.../1000/2/range of Antofagasta)

Liolaemus nigriceps (insertae sedis)

Liolaemus schmidti

Arrayán, Quebrada del (ca3321/ca7028/1800/RM/15 km E Santiago)

Liolaemus nigroviridis nigroviridis

Atacama, subcordillera de (.../.../500-1000)/3/Range of Atacama)

Phrynosaura reichei

Aucó (3131/7106/500/4/14 km NE Illapel)

Callopistes palluma palluma

Liolaemus fuscus

Liolaemus nitidus

Liolaemus platei platei

Liolaemus pseudolemniscatus

Philodryas chamissonis

Balmaceda (ca 4550/ca 7138/200/11/120 km SE Puerto Aysén)

Liolaemus magellanicus

Bellavista, Sierras de (3436/7032/3500/6/41 km E San Fernando)

Pristidactylus torquatus

Blanco, Cerro (3309/7049/690/RM/12 km SE Tilttil)

Callopistes palluma palluma

Liolaemus fuscus

Buitreras, Estación (ca 5140/ca 6940/100/12/"on the road to Río Gallegos" -sic-)

Liolaemus kingi

Calama (2227/6856/2250/2/Capital of El Loa Province)
Liolaemus paulinae

Caldera (2704/7049/19/3/Caldera town)
Liolaemus bisignatus
Liolaemus nigromaculatus nigromaculatus

Caleu (3300/7100/1200/RM/11 km NE Tiltitl)
Liolaemus fuscus
Liolaemus lemniscatus
Liolaemus monticola monticola

Camarico (3604/7225/150/7/Estero Chipa + Cauquenes river)
Philodryas chamissonis

Camión (3103/7139/100/4/Near Maitencillo town)
Liolaemus pseudolemniscatus

Cantillana, Cerro (3358/7058/2281/6/24 km W San Francisco de Mostazal)
Liolaemus altissimus (atypical form)
Liolaemus nigroviridis (atypical form)
Pristidactylus valeriae -at foot of Cerro Cantillana-

Cartagena (3333/7136/60/5/3 km N San Antonio)
Liolaemus chiliensis
Liolaemus lemniscatus
Liolaemus nitidus
Liolaemus tenuis tenuis
Philodryas chamissonis

Catalina, Bahía (5305/7050/0/12/Between Ensenada Laredo and Punta Arenas)
Liolaemus magellanicus

Cavilolén, Cuesta (3146/7119/700/4/24 km NE Los Vilos)
Philodryas chamissonis

Chena (3335/7044/950/RM/4.5 km W San Bernardo)
Philodryas chamissonis

Chile Chico (4633/7140/100/11/80 km S Balmaceda)
Diplolaemus bibroni
Liolaemus bibroni
Liolaemus darwini
Liolaemus fitzingeri

Chiloé, Island (.../.../.../10/The biggest island S Puerto Montt)
Liolaemus pictus chiloensis

Chiuchiu (2220/6839/2520/2/3 km N Salado river)
Liolaemus nigroviridis nigroroseus

Chungará (1814/6909/4570/1/20 km S Caquena)

Liolaemus alticolor

Velosaura jamesi

Cerrillos (3330/7046/470/RM/15 km SW Santiago)

Philodryas chamissonis

Clarillo, Valle del río (3341/7043/650/RM/South tributary of Maipo river)

Liolaemus lemniscatus

Liolaemus tenuis tenuis -a local form with reddish throat-

Cobquecura (3608/7249/0/8/1 km W Cobquecura town)

Liolaemus chiliensis

Liolaemus cyanogaster cyanogaster

Liolaemus lemniscatus

Philodryas chamissonis

Colchane (1917/6838/3850/1/2 km W Pisiga Chile)

Liolaemus islugensis

Liolaemus ornatus

Colina (3320/7048/470/RM/6 km W Lampa)

Callopistes palluma palluma

Liolaemus nitidus

Philodryas chamissonis

Colina, Baños de (3356/7000/2500/RM/SE tributary of El Volcán river)

Liolaemus altissimus moradoensis

Liolaemus nigroviridis minor

Concepción (3649/7303/100/8/Capital of 8th region)

Philodryas chamissonis

Concón (3256/7132/80/5/15 km S Quintero)

Liolaemus kuhlmani

Liolaemus lemniscatus

Philodryas chamissonis

Copiapó (2722/7020/450/3/Capital 3d region)

Callopistes palluma manni

Liolaemus nigromaculatus nigromaculatus

Liolaemus platei platei

Liolaemus silvai

Tropidurus atacamensis

Coquimbo (2957/7120/50/4/11 km S La Serena)

Liolaemus kuhlmani

Philodryas chamissonis

Cuesta La Dormida (3303/7102/800/RM/37 km E Quilpué)

Callopistes palluma palluma

Liolaemus chiliensis

Liolaemus schroederi

Culimó (3204/7114/390/4/5 km W Tilama)

Liolaemus fuscus

Liolaemus lemniscatus

Liolaemus nitidus

Liolaemus pseudolemniscatus

Liolaemus tenuis tenuis

Philodryas chamissonis

Curamávida (3717/7244/700-1200/8/2 km W Laja)

Liolaemus chiliensis

del Flaco, Baños (3458/7028/1520/6/70 km SE San Fernando)

Liolaemus curis

Liolaemus hernani

Phymaturus flagellifer

Philodryas chamissonis

Tachymenis chilensis chilensis

del Inca, Laguna -Portillo- (3250/7009/3500/5/42/ E Los Andes)

Liolaemus fitzgeraldi

Dos Lagunas (.../.../.../12?)

Liolaemus magellanicus

El Belloto (3312/7114/630/5/26 km SE Quilpué)

Liolaemus chiliensis

Liolaemus lemniscatus

El Ingenio (3346/7015/1200/RM/20 km SE Santiago)

Tachymenis chilensis chilensis

El Manzano (3335/7024/1068/RM/28 km S Santiago)

Liolaemus chiliensis

El Pangue (3315/7112/600/RM/18 km NW Curacaví)

Liolaemus chiliensis

El Peine, cerro (3537/7102/2448/7/4 km E Vilches)

Liolaemus cristiani

El Planchón (3514/7034/3000/7/67 km SE Curicó)

Liolaemus buergeri

Phymaturus flagellifer

El Principal (3342/7034/670/RM/9 km S Puente Alto)
Liolaemus lemniscatus

El Roble, cerro (3258/7102/2200/5/25 km SE La Calera)
Callopistes palluma palluma
Liolaemus monticola monticola
Liolaemus nigroviridis campanae
Liolaemus nitidus
Liolaemus schroederi
Pristidactylus alvaroi

El Teniente (3100/7138/100/4/South of Maitencillo)
Liolaemus platei platei
Liolaemus pseudolemniscatus

El Teniente, Mina (3404/7022/1870/6/37 km NE Rancagua)
Liolaemus altissimus altissimus (atypical form)
Liolaemus chiliensis - 1870 masl -
Liolaemus leopardinus leopardinus (atypical form)
Liolaemus schroederi

El Volcán (3349/7010/1416/RM/60 km SE Santiago)
Liolaemus chiliensis
Liolaemus lemniscatus
Liolaemus monticola monticola
Liolaemus nigroviridis minor
Liolaemus nitidus
Pristidactylus volcanensis

El Yeso, Embalse (3341/7007/3370/RM/NE tributary of Maipo river)
Liolaemus altissimus moradoensis
Liolaemus leopardinus ssp.
Liolaemus nigroviridis minor

Farellones (3320/7019/2200-3000/RM/35 km E Santiago)
Liolaemus altissimus altissimus
Liolaemus leopardinus leopardinus
Liolaemus nigroviridis nigroviridis
Liolaemus nitidus
Liolaemus schroederi

Farellones, (Curva 20) (ca 3320/ca7019/1800/RM/30 km E Santiago)
Liolaemus monticola monticola
Liolaemus nitidus

Fray Jorge, Parque Nacional (3039/7140/0-400/4/14 km W Salala)
Liolaemus fuscus
Liolaemus nitidus
Liolaemus platei platei

Liolaemus zapallarensis zapallarensis
Philodryas chamissonis
Tachymenis chilensis chilensis

Graneros (3404/7044/475/6/12 km N Rancagua)
Callopistes palluma palluma

Huasco, Puerto de (2228/7131/25/3/NE Vallenar)
Liolaemus copiapoensis
Liolaemus nigromaculatus nigromaculatus

Huentelauquén (3135/7132/25/4/35 km N Los Vilos)
Liolaemus lemniscatus

Illapel (3138/7110/350/4/50 km S Combarbalá)
Liolaemus fuscus
Liolaemus nitidus
Liolaemus pseudolemniscatus

Isluga (1915/6843/4000/1/3 km N Alsore)
Liolaemus alticolor

Josefina, Caleta (5321/6934/0/12/100 km of Bahía Inútil)
Liolaemus magellanicus

Kilómetro 300 Panam. Norte (ca3100/ca7140/100/4/South of Maitencillo)
Philodryas chamissonis

La Campana, cerro (3257/7107/300-1900/5/20 km SE La Calera)
Callopistes palluma palluma
Liolaemus fuscus
Liolaemus nigroviridis campanae
Liolaemus schroederi
Liolaemus tenuis tenuis

La Disputada, Mina (3320/7002/2500/RM/N tributary of Mapocho river)
Liolaemus schroederi

La Laguna, Embalse (3013/7004/3275/4/53 km SE Guanta)
Liolaemus lorenzmuelleri

La Pintana (3337/7039/620/RM/7 km SE San Bernardo)
Liolaemus chiliensis

La Plata, Quebrada de (3329/7054/900/RM/Ravine at W Santiago)
Liolaemus fuscus
Liolaemus lemniscatus
Liolaemus monticola monticola

La Reina, Cerros de (3354/7031/1600/RM/Ravines at E Santiago)
Philodryas chamissonis

La Serena (2954/7115/50/4/470 km N Santiago)
Callopistes palluma palluma
Liolaemus kuhlmani

Las Damas, Río de (3455/7021/2000-3000/6/73 km E San Fernando)
Liolaemus cf ceii
Liolaemus curis
Phymaturus flagellifer

Las Palmas, Rancho (3301/7104/500/5/34 km E Quilpué)
Liolaemus lemniscatus
Liolaemus tenuis tenuis

Lagunillas (3337/7018/2000/RM/35 km SE Santiago)
Liolaemus altissimus altissimus
Liolaemus nigroviridis nigroviridis

Lampaipata, Quebrada (ca1917/ca6838/3850/1/2 km W Pisiga Chile)
Liolaemus ornatus

Lejía, Laguna (2329/6741/4350/2/18 km Angostura de Pili)
Liolaemus schmidtii

Leyda (3328/7127/190/5/12 km W Puangue)
Liolaemus chiliensis

Licancabur, Volcán (2250/6753/4000/2/11 km N Casablanca)
Liolaemus nigroviridis nigroroseus
Liolaemus walkeri

Limarí (3040/7132/62/4/1 km N Salala)
Liolaemus kuhlmani

Lipangue (3321/7052/490/RM/13 km NW Santiago)
Callopistes palluma palluma

Llolleo (=Llo-lleo), Laguna de (3336/7137/10/5/2 km N Llolleo)
Liolaemus lemniscatus

Lo Arcaya (3341/7035/650/RM/8 km S Puente Alto)
Philodryas chamissonis

Lo Barnechea (3321/7030/816/RM/14 km NE Santiago)
Philodryas chamissonis

Lo Valdés (3351/7003/1800/RM/South tributary of El Volcán river)

Liolaemus leopardinus valdesianus

Liolaemus nigroviridis minor

Liolaemus nitidus

Los Andes (3250/7036/820/5/65 km N Santiago)

Phymaturus flagellifer (so uncertain locality)

Los Cipreses, Río (3425/7026/900/6/Tributary of Cachapoal river)

Liolaemus hernani

Liolaemus lemniscatus

Phymaturus flagellifer

Tachymenis chilensis chilensis

Los Dominicos (3323/7031/784/RM/11 km NE Santiago)

Liolaemus lemniscatus

Liolaemus monticola monticola

Los Molles (3043/7039/2600–2800/4/83 km E Vallenar)

Liolaemus maldonadae

Los Vilos (3154/7131/5/4/0.5 km S Los Vilos town)

Liolaemus kuhlmani

Philodryas chamissonis

Lota, Parque (3705/7309/20/8/30 km S Concepción)

Liolaemus tenuis punctatissimus

Macul, Quebrada de (3329/7028/500–1800/RM/18 km SE Santiago)

Callopistes palluma palluma

Liolaemus chiliensis

Liolaemus fuscus

Liolaemus lemniscatus

Liolaemus monticola monticola

Liolaemus nigroviridis nigroviridis

Liolaemus nitidus

Liolaemus schroederi

Liolaemus tenuis tenuis

Philodryas chamissonis

Tachymenis chilensis chilensis

Magallanes –sensu lato– (5300/7040/.../12/12 th Region)

Liolaemus magellanicus

Maipú (3330/7046/47/RM/15 km SW Santiago)

Philodryas chamissonis

Manantiales (.../.../.../12/...)

Liolaemus magellanicus

Maule, Laguna del (3604/7030/2100/7/11 km Paso Pehuenche)
Liolaemus altissimus altissimus (atypical form)
Liolaemus buergeri

Melipilla (3341/7113/170/RM/Melipilla city Westward of Santiago)
Philodryas chamissonis

Morado, Valle del -(sector Panimávidas)-
 (3347/7002/2800/RM/North tributary of Volcán River)
Liolaemus altissimus moradoensis
Liolaemus leopardinus valdesianus
Liolaemus nigroviridis minor
Tachymenis chilensis chilensis

Morales, Baños de (3347/7005/2000/RM/North tributary of Volcán River)
Liolaemus nigroviridis minor
Tachymenis chilensis chilensis

Moreno, Cerro (2400/6953/1785/2/20 km NW Antofagasta)
Liolaemus hellmichi
Tropidurus atacamensis at foot of C^o Moreno

Morrillos (3011/7133/5/4/5 km NE Guanaqueros)
Liolaemus platei platei

Munaipata, Cerro de (ca1917/ca6838/3850/1/...)
Liolaemus ornatus

Negra, Laguna (3339/7008/2220/RM/60 km SE Santiago)
Liolaemus altissimus moradoensis
Liolaemus nigroviridis minor

Negro Francisco, Laguna del (2727/6913/4125/3/12 km portezuelo Astaburuaga)
Liolaemus eleodori

Neltume (3915/7213/225/9/3 km NE Villarrica)
Liolaemus cyanogaster cyanogaster
Liolaemus pictus pictus

Ollagüe (2113/6815/3700/2/350 km NE Antofagasta)
Liolaemus pantherinus

Pajonales (2423/6837/3858/2/3 km N Salar Pajonales)
Liolaemus nigriceps -insertae sedis-

Palqui (3044/7057/500/4/5 km S Monte Patria)
Callopiastes palluma palluma

Pan de Azúcar, Cerro (2807/7056/754/3/4 km Cantera del Agua)

Liolaemus nigromaculatus atacamensis
Liolaemus nigromaculatus nigromaculatus
 (Both subspecies are found together)

Peine (2341/6802/2400/2/20 km E Laguna Brava)

Liolaemus constanzae
Liolaemus nigroviridis nigroroseus

Peñalolén (3328/7032/800/RM/11 km SE Santiago)

Philodryas chamissonis

Peñuelas, Lago (3309/7132/343/5/20 km SE Valparaíso)

Philodryas chamissonis

Pichidangui (3208/7132/5/4/4 km SW Quilimarí)

Callopistes palluma palluma
Liolaemus fuscus
Liolaemus lemniscatus
Liolaemus nitidus
Liolaemus tenuis tenuis
Liolaemus zapallarensis zapallarensis
Philodryas chamissonis

Polpaico (3309/7047/580/RM/15 km SE Tiltil)

Callopistes palluma palluma

Pucón (3916/7159/280/9/70 km SE Temuco)

Liolaemus chiliensis

Pudahuel (3321/7052/490/RM/13 km NW Santiago)

Philodryas chamissonis

Punta Arenas (5310/7054/12/12/Capital 12th region)

Liolaemus magellanicus

Punta Negra, Salar de (2428/6854/2960/2/E of Pique San Carlos)

Liolaemus schmidtii
Phrynosaura reichei

Puritama (2243/6802/3550/2/40 km NE San Pedro de Atacama)

Liolaemus dorbignii (= *L. puritamensis*)

Putendo (3237/7043/813/5/13 km N San Felipe)

Liolaemus chiliensis
Tachymenis chilensis chilensis

Quintay (3312/7142/40/5/20 km S Valparaíso)

Liolaemus kuhlmani

Quintero (3247/7132/70/5/30 km NW Quillota)

Liolaemus kuhlmani

Liolaemus lemniscatus (Relictual forest)

Philodryas chamissonis (Relictual forest)

Ramón, Cerro de (3330/7026/3050/RM/25 km E Santiago)

Liolaemus altissimus altissimus

Liolaemus leopardinus ramonensis

Liolaemus nigroviridis nigroviridis

Liolaemus nitidus (exceptionally only one specimen)

Tachymenis chilensis chilensis

Rapel (3356/7144/40/6/42 km SW San Antonio)

Philodryas chamissonis

Renca, Cerros de (3324/7042/903/RM/85 km NW Santiago)

Liolaemus lemniscatus

Río Frío (2500/6904/1000/2/High plateau of Antofagasta)

Liolaemus nigriceps

Ruínas (3105/7138/100/4/South of Maitencillo)

Liolaemus fuscus

Liolaemus pseudolemniscatus

San Antonio (3335/7137/25/5/ Capital San Antonio province)

Liolaemus lemniscatus

Liolaemus nitidus

Liolaemus tenuis tenuis

Philodryas chamissonis

San Francisco, Valle del río (3320/7022/2500/RM/North tributary of Mapocho river)

Liolaemus altissimus altissimus

Liolaemus leopardinus leopardinus

Liolaemus schroederi

San Gregorio (5239/7013/64/12/N Segunda Angostura)

Liolaemus magellanicus

San Luis de Macul (3332/7033/600/RM/13 km SE Santiago)

Callopistes palluma palluma

Liolaemus fuscus

Liolaemus monticola monticola

Philodryas chamissonis

San Pedro, Laguna (3650/7305/13/8/4 km SW Concepción)

Philodryas chamissonis

San Pedro, Pichuante, Vergara (3510/7030/2200/7/Teno lake + SW Nacimiento river)

Liolaemus buergeri

San Pedro de Atacama (2255/6812/2450/2/3 km SE Vilama)

Ceiolaemus fabiani

Liolaemus constanzae

Liolaemus nigroviridis nigroroseus

Liolaemus paulinae (uncertain occurrence)

Liolaemus walkeri

Phrynosaura audituvelata

Santa Lucía, Cerro (3327/7038/600/RM/Small hill in Santiago city)

Liolaemus tenuis tenuis

Philodryas chamissonis

Santa María, Islote (ca2400/ca6953/0/2/20 km NW Antofagasta)

Tropidurus atacamensis

Santa Rita (3340/7031/650/RM/8 km SW Puente Alto)

Philodryas chamissonis

Santiago, Ciudad de (3327/7038/600/RM/Capital of Chile)

Liolaemus chiliensis

Liolaemus gravenhorsti

Liolaemus schroederi (unusual finding)

Liolaemus tenuis tenuis

Philodryas chamissonis

Santo Domingo, Rocas de (3338/7137/15/5/6 km S San Antonio)

Liolaemus chiliensis

Liolaemus nitidus

Philodryas chamissonis

Sewell (3405/7023/2600/6/31 km NE Rancagua)

Liolaemus altissimus altissimus (atypical form)

Liolaemus hernani

Liolaemus leopardinus ssp. (atypical form)

Liolaemus nigroviridis nigroviridis (atypical form)

Liolaemus nitidus

Taco de Lampa (3314/7055/500/RM/E of Polpaico and Chacabuco)

Callopistes palluma palluma

Liolaemus lemniscatus

Philodryas chamissonis

Tagua-Tagua, San Vicente de (3426/7105/207/6/18 km NW San Fernando)

Liolaemus lemniscatus

Liolaemus tenuis tenuis

Philodryas chamissonis

Talabre (ca2311/ca6800/2500/2/8 km SE Zaparilla)

Liolaemus nigroviridis nigroroseus (very atypical form)

Talca (3525/7139/90/ Capital 7th region)

Liolaemus fuscus

Liolaemus lemniscatus

Talca ("4 km E de la Mina") -SIC- (3550/7047/959/7/15 km E Curillínque)

Liolaemus chiliensis
Liolaemus tenuis tenuis

Talinay (3050/7137/762/4/9/ km NW Peñablanca)

Liolaemus nitidus
Liolaemus zapallarensis zapallarensis
Tachymenis chilensis chilensis

Taltal (2524/7029/115/2/190 km S Antofagasta)

Liolaemus nigromaculatus atacamensis

Taltal, Quebrada de (2528/7025/750/2/Oasis in costal range)

Liolaemus donoso

Tara, Laguna (2259/6717/4350/2/18 km SE Laguna Blanca)

Liolaemus schmidt

Toconao (2311/6800/2475/2/8 km SE Zaparpilla)

Liolaemus dorbigni (=L. puritamensis)

Tocopilla (2250/7012/5/2/Capital Tocopilla province)

Phyllodactylus gerrhopygus
Tropidurus atacamensis

Totalillo (3004/7122/5/4/14 km W Guanaqueros)

Liolaemus kuhlman

Trapa-Trapa (3744/7113/1400/8/70 km E Santa Bárbara)

Liolaemus tenuis tenuis

Tulahuén (3101/7044/1096/4/33 km NE Combarbalá)

Liolaemus chiliensis
Liolaemus nitidus
Liolaemus platei platei

Tunca (3423/7106/200?/6/7 km of Peumo)

Liolaemus schroeder

Ultima Esperanza (Laguna Azul) 5130/7300/100/12/Coastal of Seno Ultima Esperanza)

Diplolaemus darwini
Liolaemus magellanicus

Valdivia (3949/7314/10/10/Capital of Valdivia province)

Liolaemus pictus pictus
Pristidactylus torquatus

"Valdiviano, Bosque" (.../.../.../9th and 10th/...)
Pristidactylus torquatus

Valparaíso, Ciudad de (3303/7107/194/5/Principal port of Chile)
Liolaemus chiliensis

Vilches Alto (3536/7105/1150/7/9 km SE Vilches)
Liolaemus chiliensis
Liolaemus lemniscatus
Liolaemus pictus pictus (atypical form)
Liolaemus schroederi
Liolaemus tenuis tenuis
Pristidactylus torquatus
Philodryas chamissonis
Tachymenis chilensis chilensis

Villarrica (3916/7207/220/9/68 km E Temuco)
Liolaemus cyanogaster cyanogaster
Liolaemus schroederi (uncertain locality)

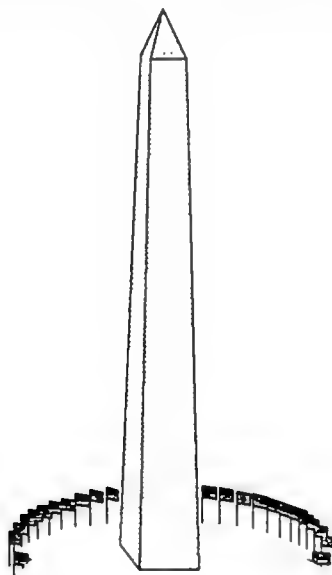
Yalquincha, Quebrada de (2227/6856/2250/2/ca Calama)
Tropidurus theresioides (?)

Zapallar (3133/7106/450/4/10 km NE Illapel)
Liolaemus chiliensis



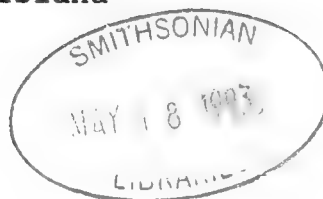
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BIBLIOGRAPHY AND SCIENTIFIC NAME INDEX
TO AMPHIBIANS AND REPTILES
IN THE
PUBLICATIONS OF THE
BIOLOGICAL SOCIETY OF WASHINGTON
BULLETIN 1-8, 1918-1988
AND
PROCEEDINGS 1-100, 1882-1987



ERNEST A. LINER

Houma, Louisiana



SMITHSONIAN
HERPETOLOGICAL INFORMATION
SERVICE
NO. 92

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INTRODUCTION

The present alphabetical listing by author(s) covers all papers bearing on herpetology that have appeared in Volume 1-100, 1882-1987, of the Proceedings of the Biological Society of Washington and the four numbers of the Bulletin series concerning reference to amphibians and reptiles.

From Volume 1 through 82 (in part), the articles were issued as separates with only the volume number, page numbers and year printed on each. Articles in Volume 82 (in part) through 89 were issued with volume number, article number, page numbers and year. Beginning with Volume 90, the Proceedings became a quarterly journal citing the volume number, issue number, page numbers and the year. Thus, the seeming inconsistencies in citations derive from the historically variable citation within the Proceedings. The Bulletin has been issued in numbers only.

All junior authors are listed alphabetically and cross referenced to the senior author for ease in locating their publications. As a result any article in this bibliography can be located by knowing the title or the author(s). All articles with original names are preceded by an * (asterisk).

All scientific names of amphibians and reptiles are listed alphabetically and referenced to the article(s) where they are mentioned. No names in bibliographic references are included. All original spellings in the articles have been maintained except those that ended with i or ii. Double ii has been substituted when both have appeared. Proceedings articles are cited with only the volume and first page number; Bulletin articles are preceded by a "B". All original names in the Proceedings are indicated in **bold**. No original names were proposed in the Bulletin. This arrangement makes it easy to distinguish articles with original names. The page number identifies the article, and not the actual page of occurrence of the name.

The author wishes to thank C. Gans for suggesting this project and for suggesting the addition of a scientific name index G. R. Zug and W. R. Heyer.

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- *----. 1913. Reptiles and amphibians from eastern Sudan. 26: 145-150.
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- *----. 1916a. A new Tantilla from Mexico. 29: 93-94.
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- . 1922d. Note on Lampropeltis mexicana (Garman). 35: 226.
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SCIENTIFIC NAME INDEX

- Abastor erythrogramus 22:
 129; 58: 131.
 Ablabes 23:169.
 Ablephararus boutoni keiensis
 24:15.
 Abronia deppi 74:37.
 Acris crepitans 74: 157.
 Acris gryllus 15: 121; 22:
 129; 23: 9; 33: 129; 44: 11;
 50: 137.
 Acris gryllus **blanchardi** 60:
 39.
 Acris gryllus crepitans 20:
 1: 21: 47; 21: 53; 21: 69;
 60: 39.
 Acris gryllus gryllus 20: 1;
 60: 39.
 Adelphicos quadrivirgatus
 sargii 62:165.
 Adenomera 87: 81.
 Adenomera andreae 87: 327;
 88: 315; 89: 581.
 Adenomera bokermanni 88: 315;
 89: 581.
 Adenomera hylaedactyla 87:
 327; 88: 315; 89: 581.
 Adenomera **lutzi** 88: 315; 89:
 581.
 Adenomera marmorata 87: 327;
 88: 315; 89: 581.
 Adenomera martinezi 88: 318;
 89: 581.
 Agalychnis 67: 159.
 Agalychnis calcarifer 93:
 1177.
 Agalychnis callidryas 93:
 153.
 Agalychnis dacnicolor 93:
 153.
 Agalychnis litodryas 93:
 1177.
 Agalychnis moreletii 37: 65;
 93: 153.
 Agalychnis spurrelli 37: 65.
 Agama colonorum 26: 145.
 Agama doriae 26: 145.
 Agama hartmanni 26: 145.
 Agama kirkii **fitzsimonsi** 63:
 127.
 Agama nupta. 83: 477.
 Agama occipitalis 63: 127.
 Agama spinosa 26: 145.
 Agkistrodon acutus 91: 963.
 Agkistrodon bilineatus 91:
 963.
 Agkistrodon bilineatus
 bilineatus 84: 327; 92:
 135.
 Agkistrodon bilineatus
howardgloydi 97: 135.
 Agkistrodon bilineatus
russeolus 84: 327; 97: 135.
 Agkistrodon bilineatus taylori
 84: 327; 97: 135.
 Agkistrodon blomhoffii 85:
 557; 91: 963.
 Agkistrodon blomhoffii
 brevicaudatus 90: 1002.
 Agkistrodon blomhoffii
 brevicaudus 85: 557; 90:
 1002.
 Agkistrodon blomhoffii
dubitatus 90: 1002.
 Agkistrodon blomhoffii
 intermedius 85: 557.
 Agkistrodon blomhoffii
siniticus 90: 1002.
 Agkistrodon brevicaudus 85:
 557.
 Agkistrodon **caliginosus** 85:
 557; 91: 963.
 Agkistrodon contortrix 15:
 121; 91: 960.
 Agkistrodon contortrix
 contortrix 82: 219.
 Agkistrodon contortrix
 laticinctus 82: 219.
 Agkistrodon contortrix mokeson
 82: 219.
 Agkistrodon contortrix
phaeogaster 82: 219.
 Agkistrodon contortrix
 pictigaster 82: 219.
 Agkistrodon halys 91: 963.
 Agkistrodon halys brevicaudus
 85: 557; 90: 1002.
 Agkistrodon halys caroganus
 90: 1002.
 Agkistrodon halys **cognatus** 90:
 1002.
 Agkistrodon halys halys 90:
 1002.
 Agkistrodon halys intermedius
 90: 1002.
 Agkistrodon himalayanus 91:
 963.

- Agkistrodon intermedius 85: 557; 91: 963.
 Agkistrodon mokasen 33: 129; 50: 137.
 Agkistrodon mokeson mokeson 59: 165; 82: 219.
 Agkistrodon monticola 91: 963.
 Agkistrodon nepa 90: 1002.
 Agkistrodon piscivorus 91: 963.
 Agkistrodon piscivorus **conanti** 82: 219.
 Agkistrodon piscivorus leucostoma 82: 219.
 Agkistrodon piscivorus piscivorus 82: 219.
 Agkistrodon rhodostoma 91: 963.
 Agkistrodon saxatilis 85: 557.
 Agkistrodon stejnegeri 85: 557.
 Agkistrodon strauchi 91: 963.
 Agkistrodon ussuriensis 85: 557.
 Alligator lacordairei 87: 231.
 Alligator mississippiensis 21: 47; 22: 129; 23: 9; 23: 115; 76: 65.
 Allophryne ruthveni 99: 42.
 Alopoglossus buckleyi 99: 214.
 Alsodes 95: 594; 98: 774.
 Alsodes gargola 93: 920.
 Alsodes illotus 93: 920.
 Alsodes laevis 93: 920.
 Alsodes montanus 93: 920.
 Alsodes monticola 93: 920.
 Alsodes nodosus 93: 920.
 Alsodes vanzolinii 93: 920.
 Alsophis 35: 219.
 Alsophis anegadae 30: 97.
 Alsophis angulifer 70: 209.
 Alsophis anomalus 45: 189.
 Alsophis antillensis 30: 97.
 Alsophis bruesi 28: 71.
 Alsophis leucomelas 28: 71.
 Alsophis leucomelas **danforthi** 51: 147.
 Alsophis leucomelas manselli 51: 147.
 Alsophis leucomelas sanctorum 51: 147.
 Alsophis melanichnus 41: 127.
 Alsophis portoricensis variegatus 90: 985.
 Alsophis pulcher 28: 71.
 Alsophis rufiventris 51: 147.
 Alsophis rudii 29: 215.
 Alsophis **sanctonum** 28: 71.
 Alsiophylax persicus 83: 477.
 Altirana 55: 49.
 Amblycephalus vertebralis 51: 43.
 Amblymetopon variegatum 49: 51.
 Amblystoma talpoideum 23: 9.
 Ambystoma annulatum 21: 85.
 Ambystoma argus 15: 239.
 Ambystoma carolinae 15: 239.
 Ambystoma copeiannum 20: 1.
 Ambystoma gracile 43: 55; 47: 167.
 Ambystoma jeffersonianum 20: 1; 25: 135; 31: 51; 50: 137; 69: 93; 76: 159.
 Ambystoma jeffersonianum fuscum 15: 121; 31: 1; B1: 1.
 Ambystoma macrodactylum 43: 55.
 Ambystoma maculatum 15: 239; 33: 129; 50: 137; 59: 165; B1: 1.
 Ambystoma microstomum 20: 1; 21: 47; 21: 53; 21: 69; 22: 115.
 Ambystoma opacum 15: 121; 15: 239; 20: 1; 21: 53; 21: 69; 21: 85; 33: 129; 50: 137.
 Ambystoma prosperina 21: 53.
 Ambystoma punctatum 15: 239; 20: 1; 21: 53; 25: 135; B1: 1.
 Ambystoma talpoideum 25: 135.
 Ambystoma texanum 58: 39.
 Ambystoma texanus 21: 53; 21: 69.
 Ambystoma tigrinum 15: 121; 20: 1.
 Ambystoma tigrinum californiense 21: 53; 43: 55; 67: 243.
 Ambystoma tigrinum diaboli 76: 159.
 Ambystoma tigrinum mavortium 67: 243.

- Ambystoma tigrinum nebulosum* 67: 243.
Ambystoma tigrinum stebbinsi 67: 243.
Ambystoma tigrinum tigrinum 67: 243.
Ambystoma tigrinum velasci 67: 243.
Ambystoma vehiculum 47: 169.
Ameiva 28: 71; 42: 153; 92: 272.
Ameiva abbotti 44: 89.
Ameiva aquilina 51: 147; 86: 231.
Ameiva ameiva aquilina 86: 231.
Ameiva ameiva atrigularis 86: 231.
Ameiva ameiva tobagana 86: 231.
Ameiva amivoides 55: 143.
Ameiva atrigularis 86: 231.
Ameiva auberi 81: 23.
Ameiva barbouri 41: 53.
Ameiva beatensis 44: 89.
Ameiva chaitzami 55: 143.
Ameiva chrysolaeama 41: 53.
Ameiva dorsalis 29: 215.
Ameiva exsul 30: 97; 51: 147.
Ameiva exsul alboguttata 90: 985.
Ameiva festiva miadis 55: 143.
Ameiva griswoldi 29: 215; 30: 163.
Ameiva lebereri 88: 367.
Ameiva leptophrys 55: 143.
Ameiva lincolota 26: 69; 41: 53.
Ameiva pluvianotata 51: 147.
Ameiva polops 26: 69; 30: 97.
Ameiva pulchra 55: 143.
Ameiva sackii 44: 73.
Ameiva surinamensis tobaganus 86: 231.
Ameiva taeniura 41: 53; 82: 777.
Ameiva thoracica 29: 215.
Ameiva tobagana 29: 221; 86: 231.
Ameiva tobaganus 29: 221.
Ameiva undulata 53: 119.
Ameiva undulata dextra 74: 37.
Ameiva undulata hartwegi 53: 55; 55: 143.
Ameiva undulata parva 53: 55; 55: 143.
Ameiva undulata pulchra 55: 143.
Ameiva undulata stuarti 53: 55; 55: 143.
Ameiva undulata undulata 53: 55; 55: 143.
Ameiva wetmorei 26: 69; 30: 97.
Amiva suranamensis tobaganus 86: 231.
Amolops 39: 53.
Amphibolurus muricatus 27: 201.
Amphisbaena 59: 73.
Amphisbaena caudalis 41: 53.
Amphisbaena innocens 41: 53.
Amphisbaena punctata 29: 85.
Amphiuma means 22: 129; 23: 9.
Amphiuma tridactyla 22: 129; 23: 9.
Amyda 28: 71.
Anadia 90: 60.
Anadia angusticeps 42: 99.
Anadia bitaeniata 42: 99; 50: 11.
Anadia bogotensis 42: 99.
Anadia metallica 42: 99.
Anadia nicefori 42: 99.
Anadia ocellata 42: 99.
Anadia pulchella 42: 99.
Anadia pulchra 50: 11.
Anadia rhombifera 42: 99.
Anadia steyeri 42: 99.
Anadia vittata 42: 99.
Ancistrodon bilineatus 84: 327.
Ancistrodon blomhoffi brevicaudus 85: 557.
Ancistrodon blomhoffii 90: 1002.
Ancistrodon contortrix 20: 1; 21: 48; 21: 69; 21: 85; 22: 115; 22: 129; 23: 9.
Ancistrodon halys brevicaudus 85: 557.
Ancistrodon halys intermedius 85: 557.
Ancistrodon halys stejnegeri 85: 557.
Ancistrodon hypnale 90: 1002.

- Ancistrodon intermedius* 84: 557.
Ancistrodon millardi 90: 1002.
Ancistrodon nepa 90: 1002.
Ancistrodon piscivorus 21: 47; 21: 69; 21: 85; 22: 129; 23: 9.
Ancistrodon saxatilis 85: 557.
Ancylocranium barkeri 59: 73.
Ancylocranium somalica 59: 73.
Ancylocranium somalicum 59: 73.
Andrias 86: 105.
Aneides aeneus 61: 127.
Aneides ferreus 43: 55; 61: 127.
Aneides flavipunctatus 43: 55; 62: 57.
Aneides flavipunctatus flavipunctatus 61: 127.
Aneides flavipunctatus niger 61: 127.
Aneides lugubris 61: 127.
Aneides lugubris lugubris 43: 55.
Anilius leachii 79: 255.
Anolis albipalpebralis 29: 215.
Anolis alligator 29: 121; 86: 231.
Anolis alutaceus 85: 509.
Anolis antiquae 28: 71; 29: 215.
Anolis argillaceus 85: 509.
Anolis asper 51: 147.
Anolis bartschi 81: 123.
Anolis bimaculatus 28: 71.
Anolis bimaculatus leachi 79: 255.
Anolis brunneus 23: 99.
Anolis carolinensis 21: 47; 21: 69; 21: 85; 22: 115; 22: 129; 23: 99; 33: 129; 61: 159.
Anolis cepedii 28: 71.
Anolis chlorocyaneus 82: 777.
Anolis chrysolaeama 82: 777.
Anolis cobanensis 55: 159.
Anolis coelestinus 41: 53.
Anolis cooperi 61: 159.
Anolis copei 49: 43.
Anolis cristatellus 28: 71; 29: 215; 30: 97; 90: 985.
Anolis cristatellus cristatellus 51: 147.
Anolis cumingii 56: 109.
Anolis cyanopleurus 85: 509.
Anolis cybotes 38: 101; 41: 53; 82: 777.
Anolis distichus 41: 53; 45: 183.
Anolis distichus altavelensis 61: 159.
Anolis distichus caudalis 61: 159.
Anolis distichus distichoides 61: 159.
Anolis distichus distichus 61: 159.
Anolis distichus dominicensis 61: 159.
Anolis distichus floridanus 61: 159.
Anolis distichus juliae 61: 159.
Anolis distichus wetmorei 61: 159.
Anolis dominicensis caudalis 45: 183.
Anolis dominicensis wetmorei 44: 89.
Anolis doris 38: 101; 41: 53.
Anolis dunni 74: 37.
Anolis ferreus 28: 71; 30: 97.
Anolis gadovi 74: 37.
Anolis hendersoni bahorucoensis 78: 39.
Anolis hendersoni hendersoni 78: 39.
Anolis krugi 30: 97.
Anolis krugi gingivinus 51: 147.
Anolis krugi wattsi 51: 147.
Anolis leachii 28: 71.
Anolis leachii lividus 51: 147.
Anolis lindeni 25: 163.
Anolis liogaster 74: 37.
Anolis longitibialis 44: 89; 88: 367.
Anolis loveridgei 49: 43.
Anolis lucius 81: 123.
Anolis marmoratus 28: 71.
Anolis megapholidotus 74: 37.
Anolis mestrei 29: 19.

- Anolis microlepidotus* 74: 37.
Anolis monensis 29: 215; 90: 985.
Anolis nebuloides 53: 119; 74: 37.
Anolis nebulosus 53: 119; 74: 37.
Anolis olsoni 41: 53.
Anolis omiltemanus 74: 37.
Anolis ordinatus 29: 215.
Anolis petersii 49: 43.
Anolis porcatus 23: 99.
Anolis principalis 23: 9.
Anolis principalis brunneus 23: 99.
Anolis pulchellus 30: 97.
Anolis richardii 30: 97; 51: 147.
Anolis ricordi 82: 777.
Anolis roquet 30: 97.
Anolis roquet gentilis 51: 147.
Anolis roquet roquet 86: 231.
Anolis roquet vincentii 51: 147.
Anolis sabanus 51: 147.
Anolis sagrei 61: 159.
Anolis semilineatus 41: 53.
Anolis spectrum spectrum 85: 509.
Anolis spectrum sumiderensis 85: 509.
Anolis stejnegeri 61: 159.
Anolis stratulus 30: 97.
Anolis subocularis 74: 37.
Anolis terrae-altae 28: 71.
Anolis townsendi 16: 3.
Anolis trossulus 29: 221.
Anolis vanidicus rejectus 85: 509.
Anolis vandicus vandicus 85: 509.
Anomalepis 45: 173.
Anomalepis mexicana 36: 185.
Anops kingii 29: 85.
Anopsibaena 29: 85.
Anotheca 52: 187.
Anotheca coronata 52: 187.
Aparasphenodon 55: 151.
Aporophis melanocephalus 35: 219.
Aristelligella barbouri 46: 33.
Aristelliger barbouri 88: 305.
Aristelliger cochranæ 46: 33; 88: 305.
Aristelliger expectatus 46: 33.
Aristelliger georgeensis 88: 305.
Aristelliger hechti 88: 305.
Aristelliger irregularis 46: 33.
Aristelliger lar 41: 53; 46: 33; 86: 35; 88: 305.
Aristelliger praesignis 46: 33; 88: 305.
Aristelliger titan 88: 305.
Arizona elegans phillipi 67: 69.
Aromochelys carinatus 23: 9.
Aromochelys odorata 20: 1.
Aromochelys odoratum 15: 235; 23: 9.
Aromochelys odoratus 15: 121; 22: 129.
Aromochelys tristycha 21: 47; 21: 69; 23: 9.
Arrhyton bivittatum 78: 99.
Arrhyton dolichurum 78: 99.
Arrhyton fulvum 78: 99.
Arrhyton taeniatum 78: 99.
Arrhyton vittatum landoi 78: 99.
Arrhyton vittatum vittatum 78: 99.
Arthroleptides 55: 49.
Arthroleptis adolfi-friderici 45: 61.
Arthroleptis lonnbergi 45: 61.
Arthroleptis methneri 45: 61.
Arthroleptis stenodactylus uluguruensis 45: 61.
Arthroleptis stenodactylus variabilis 45: 61.
Aryncus venustus 95: 688.
Ascaphus 40: 123.
Ascaphus truei 30: 123.
Ascaphus truei californicus 62: 57.
Ascaphus truei montanus 62: 57.
Ascaphus truei truei 62: 57.
Aspidonectes emoryi 21: 47; 21: 69; 23: 115.
Aspidonectes ferox 23: 115.
Aspidonectes spinifer 23: 115.

- Aspidoscelis* 42: 153.
Astylosternus 55: 49.
Atelopus 93: 1177.
Atelopus ebenoides 90: 60.6
Atelopus varius 37: 65.
Atomarchus multimaculatus 53: 125.
Atopophrynus 95: 557; 99: 214.
Atopophrynus syntomopus 95: 557.
Atractaspis alterima 26: 145.
Atractaspis irregularis 26: 145.
Atractaspis leucomelas 25: 145.
Atractaspis phillipsi 26: 145.
Atractus badius 59: 17; 68: 11.
Atractus bocourti 68: 11.
Atractus collaris 68: 11.
Atractus duboisi 68: 11.
Atractus dunni 68: 11.
Atractus ecuadorensis 68: 11.
Atractus gaigeae 68: 11.
Atractus maculata 68: 11.
Atractus occidentalis 68: 11.
Atractus occipitoalbus 68: 11.
Atractus orcesi 68: 11.
Atractus trilineatus 29: 221; 68: 11.
Austrralocrinia riparia 95: 423.
Australocrinia tasmaniensis 95: 423.
- Babina* 37: 65.
Bachia alleni alleni 78: 141.
Bachia alleni parviceps 78: 141.
Bachia alleni trinitatis 78: 141.
Bachia cuvieri 78: 141.
Bachia pallidicepa 78: 141.
Bachia trinitatus 78: 141.
Baikia africana 29: 85; 59: 73.
Barisia gadovi gadovi 74: 37.
Barycholos 84: 163.
Barycholos pulcher 87: 381.
Basanitia 92: 377.
Basanitia gehrti 98: 657.
Basanitia lactea 98: 657.
- Basanitia nigriventris* 98: 657.
Bascanion constrictor 15: 121; 23: 9.
Bascanion flagellum 23: 9.
Bascanium constrictor 22: 129.
Basiliscus americanus 27: 9.
Basiliscus barbouri 27: 9.
Basiliscus goodridgii 27: 9.
Basiliscus vittatus 53: 119; 74: 37.
Batrachemys 22: 125.
Batrachemys nasuta 22: 125.
Batrachoseps attenuatus 43: 55.
Batrachoseps attenuatus leucopus 43; 55.
Batrachyla 98: 774.
Batrachyla leptopus 95: 594.
Batrachyla taeniata 95: 594.
Bipes canaliculatus 74: 37.
Boa hortulana cookii 51: 147.
Bogertia 54: 195.
Bogertia lutzae 54: 195.
Boiga kraepelini 15: 15.
Bolieria 64: 91.
Bolitoglossa 75: 71: 90: 60.
Bolitoglossa alvaradoi 76: 289.
Bolitoglossa arborescandens 76: 289.
Bolitoghlossa bellii 54: 77; 54: 81.
Bolitoglossa borburata 76: 289.
Bolitoglossa cephalica 54: 81.
Bolitoglossa chica 89: 289.
Bolitoglossa colonea 76: 289; 89: 289.
Bolitoglossa cuchumatana 89: 289.
Bolitoglossa diminuta 89: 289.
Bolitoglossa epimela 89: 289.
Bolitoglossa flaviventris 76: 289.
Bolitoglossa gadovii 54: 81.
Bolitoglossa galaenae 54: 81.
Bolitoglossa lignicolor 76: 289.
Bolitoglossa manni 54: 81.
Bolitoglossa melanomolga 54: 81.

- Bolitoglossa mexicana* 76: 289.
Bolitoglossa palustris 76: 289.
Bolitoglossa peruviana 89: 289.
Bolitoglossa platydactyla 76: 289.
Bolitoglossa riletta 89: 289.
Bolitoglossa rostrata 89: 289.
Bolitoglossa rufescens 76: 289; 89: 289.
Bolitoglossa salvinii 76: 289.
Bolitoglossa smithi 54: 81.
Bolitoglossa striatula 76: 289.
Bolitoglossa unguidatis 54: 81.
Bolitoglossa yucatanica 76: 289.
Borborocoetes calcaratus 95: 594.
Borborocoetes insularis 95: 594.
Borborocoetes mexicanus 54: 87.
Borborocoetes miliaris 37: 65.
Borborocoetes quixensis 37: 65.
Borborocoetes stejnegeri 37: 65.
Bothriechis nigroviridis 32: 213.
Bothriopsis castelnaudi 32: 213.
Bothriopsis quadriscutatus 32: 213.
Bothrops atrox 36: 185.
Bothrops atrox atrox 81: 315.
Bothrops barbouri 72: 79.
Bothrops chloromelas 59: 17.
Bothrops lanceolatus 86: 231.
Bothrops lansbergii rozei 81: 315.
Bothrops lansbergii venezuelensis 81: 319.
Bothrops neglecta 50: 11.
Bothrops Neuvoiedii venezuelensis 81: 319.
Bothrops oligolepis 59: 17.
Bothrops pifanoi 81: 319.
Bothrops undulatus 72: 79.
Bothrops venezuelae 81: 319.
Bothrops venezuelensis 81: 319.
Briba 54: 195.
Breviceps 23: 165.
Buergeria ijimae 14: 189.
Buergeria ishikawae 14: 189.
Bufo 30: 97; 37: 141; 93: 1177.
Bufo aduncus 21: 53.
Bufo americanus 21: 69; 21: 199; 22: 129; 33: 129; 44: 11; 69: 93; 76: 159.
Bufo americanus americanus 50: 137; 76: 159.
Bufo bankorensis 51: 159.
Bufo biporcatus 24: 15.
Bufo boreas 62: 57.
Bufo boreas boreas 43: 55.
Bufo bufo miyakonis 51: 159.
Bufo californicus 43: 73; 44: 11.
Bufo canagicus canagicus 43: 55.
Bufo canagicus halophilus 43: 55; 43: 73.
Bufo canorus 43: 55.
Bufo cataulaciceps 72: 109; 73: 45.
Bufo cavator 24: 15.
Bufo coccifer 67: 159; 74: 195.
Bufo cognatus 21: 53; 44: 11; 76: 159.
Bufo cognatus californicus 43: 73.
Bufo compactilis 21: 47; 21: 53; 21: 69; 21: 199; 74: 157.
Bufo copei 69: 93.
Bufo debilis 21: 47; 21: 69; 21: 199; 22: 115.
Bufo empusus 72: 109; 73: 45.
Bufo fowleri 33: 129; 50: 137.
Bufo funereus 74: 195.
Bufo granulatus 37: 65.
Bufo gundlachi 72: 109; 73: 45.
Bufo guntheri 72: 109.
Bufo hemiophrys 76: 159.
Bufo ibarraii 67: 159; 74: 195.
Bufo lemur 30: 97; 72: 109.
Bufo lentiginosus 22: 129; 23: 9.

- Bufo lentiginosus americanus* 15: 121; 20: 1; 21: 47; 21: 53; 21: 69; 21: 85; 76: 159.
Bufo lentiginosus woodhousei 21: 53; 21: 69; 76: 159.
Bufo leptopus 51: 191.
Bufo longinasus 73: 45.
Bufo longinasus dunni 72: 109.
Bufo longinasus longinasus 72: 109.
Bufo longinasus ramsdeni 72: 109.
Bufo marinus 28: 71; 29: 221; 30: 97; 44: 11; 80: 105.
Bufo melanostictus 51: 159.
Bufo microtus 67: 159.
Bufo monksiae 53: 125.
Bufo peltacephalus 72: 109.
Bufo peltacephalus fustiger 73: 45.
Bufo peltacephalus peltacephalus 73: 45.
Bufo punctatus 21: 53; 21: 69; 21: 199; 23: 115.
Bufo quercicus 22: 129; 23: 9; 72: 109.
Bufo regularis 26: 145. 74: 195.
Bufo rubropunctatus 93: 920.
Bufo simus 53: 125.
Bufo spinulosus 98: 774.
Bufo taladai 73: 45.
Bufo terrestris americanus 69: 93.
Bufo terrestris copei 69: 93.
Bufo turpis 30: 97.
Bufo valliceps 21: 47; 21: 53; 21: 199; 22: 115; 37: 65; 67: 159; 74: 195.
Bufo valliceps macrocristatus 74: 195.
Bufo valliceps valliceps 74: 195.
Bufo valliceps wilsoni 74: 195.
Bufo venustus 95: 688.
Bufo viridis arabicus 83: 477.
Bufo woodhousei 44: 11; 76: 159.
Bufo woodhousei fowleri 59: 165.
Bufo woodhousei hemiophrys 76: 159.
- Bungarus flaviceps baluensis* 51: 43.
Bungarus flaviceps flaviceps 51: 43.
Bungarus flaviceps formosus 51: 43.
- Cacopoides borealis* 22: 89.
Cacotus calceratus 95: 594.
Cadea blanoides 29: 85; 70: 209; 71: 37.
Caecilia pachynema 59: 117.
Calamaria 49: 51.
Calamaria brachyura 40: 127.
Calamaria egregia 40: 127.
Calamaria electa 40: 127.
Calamaria gracillima 36: 91.
Calamaria leucocephala 51: 43.
Calamaria lovii 36: 91.
Calamaria lumbricoidea griswoldi 51: 43.
Calamaria lumholtzii 36: 91.
Calamaria pfefferi 14: 189.
Calamaria raveni 36: 91.
Calamaria sumatrana 40: 127.
Calamaria ventralis 36: 91.
Calamita cinereus evittata 50: 9.
Callisaurus ventralis ventralis 36: 79.
Callopeltis guttatus guttatus 15: 121.
Callopeltis obsoletus B1: 1.
Callopeltis obsoletus obsoletus 15: 121.
Callopettis guttatus B1: 1.
Callophis hughi 40: 179.
Callophis maculiceps maculiceps 40: 179.
Callophis maculiceps univirgatus 40: 179.
Callopistes 42: 153.
Calloselasma acutis 91: 963.
Calloselasma rhodostoma 91: 963.
Callula verrucosa 22: 89.
Calyptahyla crucialis 90: 194.
Camariolus 95: 423.
Candoia bibroni 98: 221.
Cardioglossa 51: 191.
Carphophiops amoenus 22: 129.
Carphophis amoena 33: 129.

- Carphophis amoena amoena 50:
 137.
 Carphophis amoenus 15: 121.
 Carphopiops amoenus 20: 1.
 Carpodacus vittatus 78: 99.
 Casarea 64: 91.
 Celestus 40: 91.
 Celestus costatus 41: 53; 92:1.
 Celestus curtissi 92: 1.
 Celestus stenurus 92: 1.
 Cemophora coccinea 18: 73;
 23: 9; 31: 99; 50: 89; B1:1.
 Centrolene geckoideum 37: 65;
 64: 34.
 Centrolene prosoblepon 37:
 65; 64: 33.
 Centrolenella 93: 1177.
 Centrolenella albolunica 93:
 714.
 Centrolenella antioquiensis
 64: 33; 93: 714.
 Centrolenella **bejaranoi** 93:
 714.
 Centrolenella **bergeri** 93:
 714.
 Centrolenella buckleyi 93:
 714.
 Centrolenella eurygnatha 93:
 714.
 Centrolenella fleischmanni
 37: 65; 93: 714.
 Centrolenella granulosa 64:
 33.
 Centrolenella johnelsi 93:
 714.
 Centrolenella munozorum 93:
 714.
 Centrolenella ocellata 93:
 714.
 Centrolenella pellucida 93:
 714.
 Centrolenella prosoblepon 93:
 714.
 Centrolenella pulverata 93:
 714.
 Centrolenella resplendens 93:
 714.
 Centrolenella siren 93: 714.
 Centrolenella spiculata 93:
 714.
 Centrolenella spinosa 64: 33.
 Centrolenella truebae 93:
 714.
 Centrolenella vanzolinii 93:
 714.
 Cerathyla bubalus 30: 31.
 Cerathyla **panamensis** 30: 31.
 Ceratophrys stolzmanni
scaphiopeza 80: 105; 84:163.
 Ceratophrys stolzmanni
 stolzmanni 80: 105.
 Cercosaurus rhombifer 42: 99.
 Chaibassie tricarineta 15:
 235.
 Chalcides ocellatus 26: 145.
 Chaleidolepis metallicus 42: 99.
 Chamaeleo **angusticoronatus**
 16: 61.
 Chamaeleo basiliscus 26: 145.
 Chamaeleo **macrorhinus** 16: 61.
 Chamaeleo tenuis **excubitor** 24:
 219.
 Chamaeleo tenuis tenuis 24: 219
 Chamaelinorops 45: 191.
 Chamaelinorops barbouri 41: 45.
 Chamaelinorops **wetmorei** 41: 45.
 Chamaello dilepis 16: 61.
 Champsia acuta 100: 713.
 Chaperina fusca 51: 191.
 Chaporina visaya 80: 65.
 Chelodina expansa 22: 125.
 Chelodina longicollis 22:
 125; 99: 472.
 Chelodina novae-guineae 22:
 125.
 Chelodina oblonga 22: 125.
 Chelodina siebenrocki 22:
 125; 99: 477.
 Chelonia depressa 27: 201.
 Chelonia japonica 27: 201.
 Chelonia mydas 27: 201; B8:
 116.
 Chelopus guttatus 22: 129;
 23: 9.
 Chelopus insculptus 20: 65.
 Chelys fimbriatus 99: 477.
 Chelydra **osceola** 31: 89.
 Chelydra serpentina 15: 121;
 20: 1; 21: 47; 21: 69; 21:
 85; 22: 129; 23: 9; 31: 89;
 33: 129; 44: 11; 50: 137.
 Chelydra serpentina serpentina
 76: 159.
 Chersydrus granulatus
 granulatus 51: 209.
 Chersydrus granulatus
luzonensis 51: 209.
 Chilomeniscus 56: 109; 67:
 69.

- Chionactis michoacensis* 56: 109.
Chionactis occipitalis annulatus 56: 109.
Chionactis occipitalis klauberi 56: 109.
Chionactis occipitalis occipitalis 56: 109.
Chionactis occipitalis palastrois 56: 109.
Chirixalus doriae 40: 179.
Chironius fuscus 59: 17.
Chiropterotriton 89: 289.
Chiropterotriton abscondens 61: 177.
Chorophilus feriarum 48: 107.
Chorophilus nigritus 20: 1; 23: 9.
Chorophilus nigritus feriarum 15: 121.
Chorophilus nigritus septentrionalis 69: 169.
Chorophilus nigritus verrucosus 48: 107.
Chorophilus occidentalis 21: 53; 21: 85; 23: 9.
Chorophilus ornatus 21: 53; 23: 9.
Chorophilus septentrionalis 69: 169.
Chorophilus triseriatus 21: 53; 22: 115; 48: 107; 69: 169.
Chorophilus triseriatus clarkii 21: 47; 21: 69.
Chorophilus verrucosus 48: 107.
Chrysemys alabamensis 99: 472.
Chrysemys concinna 51: 173; 93: 339.
Chrysemys concinna suwanniensis 99: 472.
Chrysemys elegans 20: 1.
Chrysemys floridana 93: 339.
Chrysemys floridanus 23: 9.
Chrysemys hieroglyphica 20: 1.
Chrysemys marginata 20: 1.
Chrysemys mobiliensis 23: 9.
Chrysemys picta 15: 121; 33: 129; 93: 339; B8: 108.
Chrysemys picta bellii 76: 159; 90: 685.
Chrysemys picta marginata 90: 685.
Chrysemys picta picta 50: 137; 56: 168.
Chrysemys rubriventris 23: 9; 93: 339.
Chrysemys rubriventris rubriventris 99: 472.
Chrysemys scripta 23: 9.
Chrysemys troosti 20: 1; 23: 9.
Cinosternum bauri 23: 9.
Cinosternum flavescens 23: 9.
Cinosternum louisianae 23: 9.
Cinosternum pennsylvanicum 23: 9.
Cinosternum steindachneri 23: 9.
Cistuda caroline 20: 1.
Cistuda clausa 15: 235.
Cistuda odorata 15: 235.
Cistuda pennsylvanica 15: 235.
Clelia bitorquata 36: 185.
Clelia clelia 59: 17.
Clelia clelia clelia 72: 79.
Clelia cloelia 29: 221; 36: 185.
Clelia fitzeringi 36: 185.
Clemmys guttata 31: 51; 50: 137; 76: 305.
Clemmys guttatus 15: 121.
Clemmys insculpta 31: 51; 43: 13; 50: 137; B1: 1.
Clemmys marmorata 31: 51.
Clemmys muhlenbergii 31: 51; 50: 137.
Clemmys nuchalis 31: 51.
Cnemidophorus burti 53: 119.
Cnemidophorus deppei 53: 119.
Cnemidophorus deppei deppei 42: 153; 44: 73; 74: 37.
Cnemidophorus deppei infernalis 74: 37.
Cnemidophorus gigas 74: 37.
Cnemidophorus grahami 53: 119.
Cnemidophorus gularis 21: 47; 21: 69; 21: 85; 21: 165; 53: 119.
Cnemidophorus gularis sericeus 42: 153.
Cnemidophorus guttatus 44: 73; 53: 119.
Cnemidophorus guttatus immutabilis 74: 37.

- Cnemidophorus hyperythrus danheimae* 42: 153.
Cnemidophorus lemniscatus 29: 221.
Cnemidophorus metanostethus 53: 119.
Cnemidophorus murinus 42: 153.
Cnemidophorus perplexus 42: 153; 53: 119; 53: 125.
Cnemidophorus pictus 67: 69.
Cnemidophorus sacki sacki 74: 37.
Cnemidophorus sackii 44: 73.
Cnemidophorus sexlineatus 15: 121; 20: 1; 21: 69; 33: 129; 36: 85; 42: 153; 53: 119; 57: 124; 58: 89; B1: 1.
Cnemidophorus sexlineatus gularis 44: 73.
Cnemidophorus sexlineatus perplexus 44: 73.
Cnemidophorus sexlineatus sackii 44: 73.
Cnemidophorus sexlineatus tessellatus 44: 73.
Cnemidophorus tessellatus 21: 165; 35: 1; 44: 73.
Cnemidophorus tessellatus tessellatus 36: 85; 42: 153; 53: 119.
Cnemidophorus tigris 35: 1.
Cnemidophorus undulatus 55: 143.
Cochranella 64: 33.
Cochranella albomaculata 64: 33.
Cochranella buckleyi 64: 33.
Cochranella chrysops 64: 33.
Cochranella colymbiphyllum 64: 33.
Cochranella eurygnatha 64: 33.
Cochranella fleischmanni 64: 33.
Cochranella parambae 64: 33.
Cochranella parvula 64: 33.
Cochranella pulverata 64: 33.
Cochranella uranoscopa 64: 33.
Cochranella valeroi 64: 33.
Cochranella viridissima 64: 33.
Coleonyx brevis 71: 14.
Coleonyx elegans 71: 149.
- Coleonyx elegans nemoralis* 74: 37.
Coleonyx fasciatus 71: 149.
Coleonyx mitratus 71: 149.
Coleonyx reticulatus 71: 149.
Coleonyx variegatus 71: 149.
Colorhogia redimita 78: 99.
Colostethus 93: 1177.
Colostethus abditaurantiacus 95: 557.
Colostethus alagoanus 84: 147.
Colostethus alboguttatus 84: 147.
Colostethus anthonyi 84: 147.
Colostethus anthracinus 84: 147.
Colostethus beebei 84: 147.
Colostethus bocagei 84: 147.
Colostethus bromelicola 84: 147.
Colostethus brunneus 84: 147.
Colostethus capixaba 84: 147.
Colostethus carioca 84: 147.
Colostethus chocoensis 84: 147.
Colostethus collaris 84: 147.
Colostethus dunni 84: 147.
Colostethus elachyhistus 84: 147.
Colostethus festae 84: 147.
Colostethus fuliginosus 84: 147.
Colostethus granuliventris 84: 147.
Colostethus herminae 84: 147.
Colostethus infraguttatus 84: 147.
Colostethus inguinalis 84: 147.
Colostethus intermedius 84: 147.
Colostethus kingsburyi 84: 147.
Colostethus latinasus 84: 147.
Colostethus mandelorum 84: 147.
Colostethus maechesianus 84: 147.
Colostethus mertensi 84: 147.
Colostethus neblina 84: 147.
Colostethus nexipus 99: 214.
Colostethus nubicola 84: 147; 93: 1177.
Colostethus olfersioides 84: 147.
Colostethus palmatus 84: 147; 95: 557.

- Colostethus peruvianus* 84: 147.
Colostethus pratti 84: 147.
Colostethus pulchellum 84: 147.
Colostethus ranoides 84: 147.
Colostethus riocosangae 84: 147.
Colostethus riveroi 84: 147.
Colostethus shrevei 84: 147.
Colostethus subpunctatus 84: 147; 90: 60.
Colostethus sylvatica 84: 147.
Colostethus taeniatus 84: 147.
Colostethus talamancae 84: 147.
Colostethus taeniatus 84: 147.
Colostethus tricolor 84: 147.
Colostethus trilineatus 84: 147.
Colostethus trinitatus 84: 147.
Colostethus variabilis 84: 147.
Colostethus vergeli 84: 147; 95: 557.
Colostethus vertebralis 84: 147.
Colostethus whimperi 84: 147.
Coluber bicinctus 82: 847.
Coluber chlorosoma 53: 125.
Coluber coccineus 31: 99.
Coluber constrictor 33: 129; B1: 1.
Coluber constrictor constrictor 50: 137.
Coluber constrictor flaviventris 44: 11.
Coluber doliatus 31: 99.
Coluber elapsoides 31: 99.
Coluber emoryi 23: 115.
Coluber erythrogaster B1: 1.
Coluber flagellum 33: 129.
Coluber guttatus 20: 1; 22: 129; 23: 9; B1: 1.
Coluber irregularis 15: 15.
Coluber obsoletus 20: 1; B1: 1.
Coluber obsoletus confinis 23: 9.
Coluber obsoletus confinus 23: 115.
Coluber obsoletus lindheimeri 21: 69.
Coluber quadrivittatus 15: 36; 22: 129; 23: 9.
Coluber rosaceus 46: 153.
Coluber spiloides 21: 47; 21: 69; 23: 115.
Condylosaurus subtessellatus 35: 229.
Coniophanes 35: 219; 56: 109.
Coniophanes bipunctatus bipunctatus 53: 55.
Coniophanes bipunctatus biseriatus 53: 55.
Coniophanes fissidens dispersus 72: 79.
Conophis vittatus viduus 72: 79; 74: 37.
Conophis vittatus vittatus 72: 79.
Conopsis 56: 109.
Conopsis nasus 53: 125.
Constrictor constrictor 29: 221; 36: 185.
Constrictor constrictor imperator 72: 79.
Contia episcopa isozona 56: 109.
Contia isozona 56: 109.
Contia michoacanensis 56: 109.
Contia nasus 53: 125.
Contia taylori 56: 109.
Contia tenuis 56: 109.
Cophoscincus subvittatus 76: 69.
Cornufer boulengeri 81: 69.
Cornufer corrugatus 80: 65.
Cornufer corrugatus corrugatus 21: 165.
Cornufer corrugatus rubristriatus 21: 189.
Cornufer nova-britannica 81: 69.
Cornufer punctata 91: 965.
Cornufer tenasserimensis 55: 71.
Coronella coccinea 31: 99.
Coronella cyclura 82: 763.
Coronella jaegeri 98: 295.
Corythomantis 52: 187; 55: 151.
Cosmiosophis 82: 847.
Crepidophryne 93: 1177.
Crepidophryne epioticus 99: 42.
Crepidus epioticus 99: 42.
Crinia bilingua 95: 423.
Crinia deserticola 95: 423.
Crinia georgiana 95: 423.
Crinia glauerti 95: 423.

- Crinia haswelli* 95: 423.
Crinia insignifera 95: 423.
Crinia parinsignifera 95: 423.
Crinia pseudinsignifera 95: 423.
Crinia remota 95: 423.
Crinia riparia 95: 423.
Crinia signifera 95: 423.
Crinia sloanei 95: 423.
Crinia subinsignifera 95: 423.
Crinia tinnula 95: 423.
Crinia victoriana 95: 423.
Crocodylus 42: 153.
Crocodylus americanus mexicanus 100: 713.
Crocodylus americanus moreletii 100: 713.
Crocodylus mexicanus 100: 713.
Crocodylus acutus 76: 65; 87: 231.
Crocodylus acutus acutus 100: 713.
Crocodylus moreletii 87: 231; 100: 713.
Crossodactylodes 51: 41.
Crossodactylodes pintoii 51: 41.
Crossodactylus 98: 774; 99: 42; 99: 100.
Crotalus adamanteus 21: 47; 22: 129; 23: 9.
Crotalus atrox 21: 47; 21: 69.
Crotalus durissus culminatus 72: 79.
Crotalus horridus 15: 121; 20: 1; 21: 47; 21: 69; 21: 85; 22: 129; 33: 129; 57: 33; B1: 1.
Crotalus lepidus 40: 57; 53: 125.
Crotalus molossus mollosus 53: 125.
Crotalus pricei 32: 213.
Crotalus stejnegeri 32: 213.
Crotalus tigris 32: 213.
Crotalus triseriatus 32: 213.
Crotalus triseriatus pricei 53: 125.
Crotalus willardi 53: 125.
Crotaphopeltis 35: 229.
Crotaphopeltis duchesnii 45: 83.
Crotaphopeltis hotamboeia hotamboeia 45: 83.
Crotaphopeltis shrevei 45: 83.
Crotaphytus 21: 53.
Crotaphytus collaris 21: 69; 44: 11.
Crotaphytus collaris baileyi 36: 83; 53: 119.
Crotaphytus collaris collaris 53: 119.
Crotaphytus reticulatus 53: 119.
Crotaphytus wislizenii 35: 1; 53: 119.
Cryptoblepharus 93: 350; 98: 221.
Cryptoblepharus boutonii balinensis 24: 15; 27: 201.
Cryptoblepharus boutonii cursor 24: 15; 27: 201.
Cryptoblepharus boutonii furcata 24: 15.
Cryptoblepharus boutonii keiensis 24: 15; 27: 201.
Cryptoblepharus boutonii nigropunctatus 24: 15.
Cryptoblepharus boutonii paschalis 24: 15.
Cryptoblepharus boutonii peronii 24: 15; 27: 201.
Cryptoblepharus nigropunctatus 24: 15.
Cryptoblepharus poecilopleurus 24: 15.
Cryptobranchus alleghaniensis 20: 1; 25: 135.
Ctenosaura pectinata 74: 37.
Ctenosaurus acanthura 53: 119.
Cuora chriskaronnarum 100: 624.
Cuora pani 100: 624.
Cuora trifasciatus 100: 624.
Cuora tungia 96: 567.
Cuora yunnanensis 100: 624.
Cyclagras bicinctus 82: 847.
Cyclagras gigas 82: 847.
Cyclemys orbiculata 15: 235.
Cyclemys trifasciata 15: 235.
Cyclodina whitakeri 98: 221.

- Cyclophis aestivus* 20: 1; 21: 47; 21: 69; 21: 85; 23: 9; B1: 1.
Cyclorhamphus 62: 57.
Cycloramphus asper 96: 548.
Cycloramphus boraceiensis 96: 548.
Cycloramphus cedrensis 96: 548.
Cycloramphus duseni 96: 548.
Cycloramphus fuliginosus 96: 548.
Cycloramphus granulatus 96: 548.
Cycloramphus izecksohni 96: 548.
Cycloramphus lutzorum 96: 548.
Cycloramphus mirandaribeiroi 96: 548.
Cycloramphus ohausi 96: 548.
Cycloramphus rhyakonastes 96: 548.
Cycloramphus semipalmatus 96: 548.
Cycloramphus valae 96: 548.
Cyclorana alboguttatus 63: 131.
Cyclorana australis 63: 131.
Cyclorana slevini 63: 131.
Cyclura 88: 367.
Cyclura bocolopha 16: 129; 32: 145.
Cyclura carinata 16: 129; 29: 215; 30: 97; 32: 145.
Cyclura caymanensis 32: 145.
Cyclura collei 30: 97; 32: 145.
Cyclura cornuta 16: 129; 32: 145; 41: 53.
Cyclura cornuta stejnegeri 90: 985.
Cyclura cyclura 16: 129.
Cyclura inornata 29: 215; 32: 145.
Cyclura macleayi 32: 145.
Cyclura mattea 32: 145.
Cyclura nigerrima 30: 97; 32: 145.
Cyclura nuchalis 32: 145.
Cyclura pinguis 30: 97; 32: 145.
Cyclura portoricensis 32: 145.
Cyclura rileyi 16: 129; 32: 145.
Cyclura stejnegeri 30: 97; 32: 145.
Cyrtodactylus pelagicus 98: 221.
Cystignathus fernandezi 95: 594.
Cystignathus insularis 95: 594.
Cystignathus oxyglossus 95: 594.
Dolophia 59: 73.
Dasia smaragdina 27: 201.
Dasia smaragdinum moluccarum 24: 15.
Dasia smaragdinum smaragdinum 24: 15.
Dasia smaragdinum viridipunctum 24: 15.
Dasia vittata 58: 47.
Deinagkistrodon 91: 963.
Deinagkistrodon acutus 91: 963.
Deirochelys reticulata 23: 9.
Deiroptyx bartschi 41: 169; 71: 37.
Deiroptyx vermiculata 41: 169.
Deiroptyx vermiculatus 71: 37.
Dendraspis jamesoni jamesoni 49: 63.
Dendraspis jamesoni kaimosae 49: 63.
Dendrelaphis caudolineatus 51: 43.
Dendrobates 55: 49; 84: 147; 93: 1177; 99: 214.
Dendrobates anthonyi 94: 67.
Dendrobates boulengeri 94: 67.
Dendrobates espinosal 94: 67.
Dendrobates femoralis 94: 67.
Dendrobates histrionicus 95: 557.
Dendrobates lugubris 34: 157.
Dendrobates myersi 94: 67.
Dendrobates parvulus 55: 151.
Dendrobates pumilio 93: 1177.
Dendrobates ranoides 84: 147.
Dendrobates talamancae 34: 157.
Dendrobates tricolor 94: 67.

- Dendrobates truncatus* 34: 157.
Dendrobates zaparo 94: 67.
Dendrophidian clarki 54: 73.
Dendrophidian dendrophis 54: 73.
Dendrophidian paucicarinatus 54: 73.
Dendrophidian percarinatus 54: 73.
Dendrophidian vinitor 54: 73.
Dendrophis calligaster 27: 201.
Dendrophis pictus 51: 43.
Desmognathus brimleyorum 21: 85.
Desmognathus fusca 20: 1; 22: 129; 25: 135; 29: 73; B1: 1.
Desmognathus fusca auriculata 23: 9.
Desmognathus fuscus 61: 127.
Desmognathus fuscus fuscus 15: 121; 33: 129; 50: 137; 58: 39.
Desmognathus monticola 29: 73; 33: 129; 58: 39; 71: 153.
Desmognathus nigra 25: 135.
Desmognathus ochrophaea 25: 135.
Desmognathus ochrophaea carolinensis 29: 73; 33: 129.
Desmognathus ochrophaea ochrophaea 29: 73.
Desmognathus phoca 58: 39.
Desmognathus quadramaculata 25: 135; 29: 73; 76: 153.
Desmognathus quadra-maculatus 33: 129.
Diadophis punctatus 15: 121; 20: 1; 22: 129; 23: 9.
Diadophis punctatus edwardsii 33: 129; 50: 137.
Diadophis punctatus punctatus 58: 131.
Diadophis regalis 21: 69; 21: 85.
Diadophis regalis arizonae 53: 125.
Diadophis regalis arnyi 21: 69.
Diaglena 52: 187.
Diaglena reticulata 55: 151.
Diaglena spatulata 55: 151.
Dicamptodon ensatus 43: 55; 61: 127.
Dicrodon 42: 153.
Dicroglossus blythii 55: 49.
Dicroglossus brevipalmatus 55: 49.
Dicroglossus cancrivora 55: 49.
Dicroglossus corrugata 55: 49.
Dicroglossus cyanophlyctis 55: 49.
Dicroglossus doriae 55: 49.
Dicroglossus galamensis 55: 49.
Dicroglossus grayi 55: 49.
Dicroglossus grunniens 55: 49.
Dicroglossus kuhlii 55: 49.
Dicroglossus leytensis 55: 49.
Dicroglossus limnocharis 55: 49.
Dicroglossus macrodon 55: 49.
Dicroglossus microdisca 55: 49.
Dicroglossus microtypanum 55: 49.
Dicroglossus modesta 55: 49.
Dicroglossus occipitalis 55: 49.
Dicroglossus tigerina 55: 49.
Diemyctylus viridescens 15: 121.
Diemyctylus ensicauda 51: 159.
Diemyctylus meridionalis 21: 47.
Diemyctylus viridescens 20: 1; 21: 47; 23: 9; 25: 135.
Diemyctylus viridescens meridionalis 21: 53; 21: 69.
Diemyctylus viridescens miniatus 21: 53.
Diploglossus analpistus 92: 1.
Diploglossus barbouri 82: 777.
Diploglossus costatus 82: 777.
Diploglossus cruscus 82: 777.

- Diploglossus curtissi* 82: 777.
Diploglossus darlingtoni 82: 777.
Diploglossus delasagra 82: 777.
Diploglossus duquesneyi 82: 777.
Diploglossus hewardi 82: 777.
Diploglossus microblepharis 82: 777.
Diploglossus montisserrati 82: 777.
Diploglossus occiduus 82: 777.
Diploglossus pleii 82: 777.
Diploglossus sepsoides 82: 777.
Diploglossus stenurus alloeides 82: 777.
Diploglossus stenurus rugosus 82: 777.
Diploglossus warreni 82: 777; 92: 1.
Diplolaemus bibronii 34: 139.
Diplolaemus darwinii 34: 139.
Diporophora bilineata 27: 201.
Dipsadomorphus 15: 15.
Dipsas latifasciatus 59: 17.
Dipsas schunkii 59: 17.
Dipsosaurus dorsalis carmenensis 67: 69.
Dipso-saurus dorsalis sonoriensis 53: 119.
Discodeles bufoniformis 91: 965.
Discodeles guppyi 91: 965.
Discodeles tenasserimensis 55: 71.
Disteira godeffroyi 60; 1.
Disteira orientalis 14: 189.
Doliophis bilineata 57: 105.
Dracaena 42: 153.
Draco cornutus 16: 59.
Draco gracilis 16: 59.
Dromica 35: 219.
Dromicus amazonicus 35: 219.
Dromicus w-nigrum 41: 127.
Dryadophis melanolomus stuarti 72: 79.
Dryadophis pleii B2: 199.
Drymarchon 76: 153.
Drymarchon corais 29: 221.
Drymarchon corais rubidus 72: 79.
Drymobius boddaertii 28: 71; 29: 221; 36: 185; 86: 231.
Drymobius boddaertii bruesi 51: 147.
Drymobius chloroticus 54: 73.
Drymobius margaritiferus fistulosus 72: 79.
Dugandia 82: 847.
Echis carinatus pyramidum 83: 477.
Ecpleopus affinis 57: 97.
Ecpleopus bogotensis 42: 99.
Ecpleopus gaudichaudii 57: 97.
Ecpleopus lutzae 57: 97.
Edalorhina perezii 55: 151.
Eirenis persica 83: 477.
Eirenis punctatolineata 83: 477.
Elachistocleis ovalis 55: 151.
Elaphe bairdi 43: 73.
Elaphe chlorosoma 53: 125.
Elaphe conspicillata 42: 129.
Elaphe guttata 45: 5; 46: 153; 58: 89.
Elaphe laeta 44: 11.
Elaphe laeta intermontanus 55: 133.
Elaphe laeta laeta 55: 133.
Elaphe melanura 51: 43.
Elaphe obsoleta confinis 33: 129.
Elaphe obsoleta obsoleta 50: 137.
Elaphe periacea 42: 129.
Elaphe quadrivittata deckerti 45: 5.
Elaphe quadrivittata quadrivittata 45: 5.
Elaphe rosacea 45: 5; 46: 153.
Elaphe triaspis intermedia 72: 779.
Elapochrous deppei 54: 119.
Elapomorphus blumii 27: 199.
Elapomorphus michoacanensis 56: 109.
Elapomorphus nuchalis 27: 199.
Elaps fitzingeri 55: 63.
Elaps fulvius 21: 47; 21: 69; 23: 9.

- Elaps schrankii* 82: 847.
Eleutherodactylus 28: 131;
 53: 125; 93: 1177; 94: 413;
 95: 557; 98: 774.
Eleutherodactylus abbotti 36:
 93; 78: 165.
Eleutherodactylus achatinus
 87: 381; 90: 424; 92: 498.
Eleutherodactylus acuminatus
 87: 381; 88: 351.
Eleutherodactylus affinis 90:
 60.
Eleutherodactylus alcoae 88:
 367.
Eleutherodactylus alfredi 52:
 187; 54: 197; 80: 211; 93:
 928.
Eleutherodactylus altae 93:
 928; 94: 413.
Eleutherodactylus
altamazonicus 34: 157; 91:
 418
Eleutherodactylus alticola
 90: 194.
Eleutherodactylus andicola
 100: 337.
Eleutherodactylus andrewsi
 90: 194.
Eleutherodactylus anotis 93:
 327.
Eleutherodactylus antillensis
 30: 97.
Eleutherodactylus anzuetoii
 54: 197.
Eleutherodactylus
appendiculatus 83: 221.
Eleutherodactylus areolatus
 87: 381.
Eleutherodactylus armstrongi
 78: 165.
Eleutherodactylus
auriculatoides 45: 191.
Eleutherodactylus auriculatus
 70: 209; 71: 37; 78: 165;
 90: 194; 90: 424.
Eleutherodactylus baryecus
 93: 327.
Eleutherodactylus
berkenbuschii 93: 928.
Eleutherodactylus binotatus
 98: 657.
Eleutherodactylus biporcatus
 83: 221.
Eleutherodactylus bogotensis
 90: 60.
Eleutherodactylus bransfordi
 34: 157.
Eleutherodactylus brocchi 37:
 65; 54: 197; 93: 928.
Eleutherodactylus bufonoides
 80: 211.
Eleutherodactylus cabrerai
 93: 327.
Eleutherodactylus cactorum
 53: 55.
Eleutherodactylus calcarulatus
 92: 498.
Eleutherodactylus carvalhoi
 87: 381.
Eleutherodactylus
caryophyllaceus 93: 928;
 94: 413.
Eleutherodactylus cavernicola
 90: 194.
Eleutherodactylus cerasinus
 93: 928; 94: 413.
Eleutherodactylus ceresinus
 34: 157.
Eleutherodactylus chalceus
 92: 498.
Eleutherodactylus chiastonotus
 90: 424.
Eleutherodactylus colodactylus
 93: 327.
Eleutherodactylus
conspicillatus 35: 111; 83:
 221; 90: 424; 91: 418; 100:
 337.
Eleutherodactylus coqui 91:
 123.
Eleutherodactylus cornutus
 83: 221.
Eleutherodactylus cosnipatae
 91: 418; 100: 337.
Eleutherodactylus cremmobates
 92: 498.
Eleutherodactylus crenunguis
 92: 498.
Eleutherodactylus
croceoinguinis 87: 381.
Eleutherodactylus cruentus
 92: 498; 93: 928; 94: 413.
Eleutherodactylus cruralis
 100: 337.
Eleutherodactylus cundalli
 90: 194.
Eleutherodactylus danae 91:
 418; 100: 337.
Eleutherodactylus decoratus
 80: 211; 93: 928.

- Eleutherodactylus diastema* 87: 381.
Eleutherodactylus diastemma 34: 157.
Eleutherodactylus dimidiatus 70: 209; 71: 37; 78: 165.
Eleutherodactylus discoidalis 100: 337.
Eleutherodactylus dubitus 94: 413.
Eleutherodactylus duellmani 93: 327.
Eleutherodactylus dunnii 35: 111.
Eleutherodactylus eileenae 71: 37.
Eleutherodactylus elegans 90: 60.
Eleutherodactylus fenestratus 90: 424; 94: 404; 100: 337.
Eleutherodactylus fitzingeri 83: 221; 90: 424; 91: 418; 92: 498; 93: 928; 94: 404.
Eleutherodactylus flavescens 51: 93.
Eleutherodactylus fleishmanni 34: 157.
Eleutherodactylus frater 93: 928; 94: 404.
Eleutherodactylus fraudator 100: 337.
Eleutherodactylus fuscus 90: 194.
Eleutherodactylus galdi 83: 221.
Eleutherodactylus gehrmanni 71: 37.
Eleutherodactylus glandulosus 88: 351.
Eleutherodactylus goldmani 37: 65.
Eleutherodactylus gollmeri 54: 197; 83: 221; 90: 424.
Eleutherodactylus gossei 90: 194.
Eleutherodactylus grabhami 90: 194.
Eleutherodactylus grandoculis 90: 424.
Eleutherodactylus granulatus 91: 418; 100: 337.
Eleutherodactylus greyi 70: 209; 71: 37.
Eleutherodactylus guentheri 83: 221; 98: 657.
Eleutherodactylus guerreroensis 80: 211.
Eleutherodactylus gularis 87: 381.
Eleutherodactylus gundlachi 78: 165.
Eleutherodactylus gutturalis 90: 424; 94: 404.
Eleutherodactylus heminota 78: 165.
Eleutherodactylus heterodactylum 90: 424.
Eleutherodactylus hidalgoensis 80: 211.
Eleutherodactylus hoehnei 98: 657.
Eleutherodactylus holti 98: 657.
Eleutherodactylus humeralis 34: 157.
Eleutherodactylus incomptus 94: 404.
Eleutherodactylus inguinalis 90: 424.
Eleutherodactylus inoptatus 30: 97; 41: 53; 78: 165.
Eleutherodactylus insignatus 30: 97.
Eleutherodactylus jamaicensis 90: 194.
Eleutherodactylus johnstonei 51: 147; 90: 194.
Eleutherodactylus jugans 78: 165.
Eleutherodactylus junori 90: 194.
Eleutherodactylus klinikowskii 78: 165.
Eleutherodactylus lacrimosus 90: 424.
Eleutherodactylus lactea 98: 657.
Eleutherodactylus lanciformis 34: 157.
Eleutherodactylus lanthanites 90: 424; 94: 404.
Eleutherodactylus latidiscus 87: 381; 94: 413.
Eleutherodactylus latidiscus tamsitti 92: 498.
Eleutherodactylus lentus 28: 71; 30: 97.
Eleutherodactylus leptodactyloides 84: 163.

- Eleutherodactylus leucopus* 88: 351.
Eleutherodactylus lindae 91: 418; 100: 337.
Eleutherodactylus longirostris 37: 65.
Eleutherodactylus loustes 92: 498.
Eleutherodactylus luteolus 90: 194.
Eleutherodactylus lutosus lutosus 94: 413.
Eleutherodactylus lutosus molinoi 94: 413.
Eleutherodactylus lymani 83:221.
Eleutherodactylus lynchi 90:60.
Eleutherodactylus marmoratus 90: 424; 94: 404.
Eleutherodactylus marshae 94: 413.
Eleutherodactylus martiae 87: 381; 94: 404.
Eleutherodactylus martinicensis 29: 215; 29: 221; 51: 147; 86: 231.
Eleutherodactylus melanostictus 54: 197.; 93: 928; 94: 413.
Eleutherodactylus mendax 91: 418; 100: 337.
Eleutherodactylus mercedesae 100: 337.
Eleutherodactylus mexicanus 54: 87.
Eleutherodactylus milesi 49:43.
Eleutherodactylus monensis 90: 985.
Eleutherodactylus monnichorum 93: 928.
Eleutherodactylus moro 87: 381; 93: 928.
Eleutherodactylus necerus 92: 498.
Eleutherodactylus neodreptus 78: 165.
Eleutherodactylus nicefori 90: 60.
Eleutherodactylus nigriventris 95: 377; 98: 657.
Eleutherodactylus nigrovittatus 83: 221.
Eleutherodactylus noblei 34:157.
Eleutherodactylus nubicola 90: 194.
Eleutherodactylus nyctophylax 100: 337.
Eleutherodactylus occidentalis 54: 87.
Eleutherodactylus orcutti 90: 194.
Eleutherodactylus ornatissimus 87: 381.
Eleutherodactylus orphnolaimus 83: 221.
Eleutherodactylus palmeri 100: 337.
Eleutherodactylus pantoni 90: 194.
Eleutherodactylus parabotes 78: 165.
Eleutherodactylus pardalis 93: 928; 94: 413.
Eleutherodactylus parvus 98: 657.
Eleutherodactylus paululus 87: 381; 88: 351.
Eleutherodactylus peraltae 94: 413.
Eleutherodactylus peruvianus 91: 418.
Eleutherodactylus pharangobates 91: 418; 100: 337.
Eleutherodactylus planirostris 90: 194.
Eleutherodactylus platydactylus 91: 418; 100: 337.
Eleutherodactylus platyrhynchus 93: 928.
Eleutherodactylus polyptychus 34: 157.
Eleutherodactylus poolei 51: 93.
Eleutherodactylus pseudoacuuminatus 87: 381.
Eleutherodactylus pugnax 93: 327.
Eleutherodactylus pulvinatus 83: 221; 90: 424.
Eleutherodactylus pusillus 98: 657.
Eleutherodactylus quixensis 84: 163.
Eleutherodactylus randorum 98: 657.
Eleutherodactylus rayo 93:928.
Eleutherodactylus rhabdolaemus 100: 337.

- Eleutherodactylus rhodopus* 34: 157; 37: 65.
Eleutherodactylus ricordi planirostris 70: 209.
Eleutherodactylus ricordi rogersi 79: 209.
Eleutherodactylus ricordii 23: 100; 71: 37; 78: 165; 90: 194.
Eleutherodactylus ridens 93: 928; 94: 413.
Eleutherodactylus rostralis 54: 197.
Eleutherodactylus rubicundus 92: 498.
Eleutherodactylus rufifemoralis 78: 165.
Eleutherodactylus rugosus 34: 157.
Eleutherodactylus rugulosus 83: 221; 93: 928.
Eleutherodactylus ruidus 93: 327.
Eleutherodactylus ruthae 78: 165.
Eleutherodactylus salaputium 91: 418; 100: 337.
Eleutherodactylus sallaei 35: 111; 37: 65.
Eleutherodactylus saltator 54: 87.
Eleutherodactylus savagei 94: 404.
Eleutherodactylus scitulus 100: 337.
Eleutherodactylus silvicola 80: 211.
Eleutherodactylus sisypodemus 90: 194.
Eleutherodactylus sobetes 93: 327.
Eleutherodactylus sonans 71: 37.
Eleutherodactylus spanios 98: 657.
Eleutherodactylus spatulatus 52: 187; 54: 197; 80: 211.
Eleutherodactylus stadelmani 49: 43.
Eleutherodactylus surdus cabrerai 93: 327.
Eleutherodactylus surdus surdus 93: 327.
Eleutherodactylus symingtoni 70: 209; 71: 37.
Eleutherodactylus taeniatus 94: 404; 94: 413.
Eleutherodactylus talamancae 93: 928.
Eleutherodactylus terraebolivaris 90: 424; 94: 404.
Eleutherodactylus tiptoni 94: 413.
Eleutherodactylus trachyblepharis 87: 381.
Eleutherodactylus unistrigatus 90: 60; 90: 424; 91: 418; 92: 498; 93: 327; 93: 928; 94: 404; 100: 337.
Eleutherodactylus unistrigatus hosti 98: 657.
Eleutherodactylus urichii 51: 147; 90: 424.
Eleutherodactylus varians 71: 37.
Eleutherodactylus varleyi 70: 209; 71: 37.
Eleutherodactylus venancioi 98: 657.
Eleutherodactylus ventrilineatus 78: 165.
Eleutherodactylus ventrimarmoratus 91: 418; 94: 413.
Eleutherodactylus vertebralis 90: 60.
Eleutherodactylus vilarsi 90: 424; 94: 404.
Eleutherodactylus vocalis 93: 928.
Eleutherodactylus walkeri 87: 381.
Eleutherodactylus wetmorei 45: 191.
Eleutherodactylus w-nigrum 92: 498.
Eleutherodactylus xucanebi 54: 197.
Eleutherodactylus zeuctotylus 90: 424.
Eleutherodactylus zeus 71: 37.
Elgaria formosa 15: 37.
Elosia aspera 95: 377.
Elosia bufonium 95: 377.
Elosia glabra 95: 377; 99: 100.
Elosia lateristrigata 99: 100.
Elosia lateristrigatus 95: 377.
Elosia magalhaesi 95: 377.
Elosia nasus meridionalis 95: 377.
Elosia nasuta 95: 377.
Elosia ornata 95: 377.
Elosia perplicata 95: 377.
Elosia pulchra 99: 100.

- Emoia caeruleocauda* 93: 350.
Emoia concolor 93: 350.
Emoia cyanura 93: 350; 98:221.
Emoia murphyi 93: 350.
Emoia nigra 93: 350.
Emoia parkeri 93: 350.
Emoia samoensis 93:350; 98:221.
Emoia speiseri 93: 350.
Emys castanea 15: 235.
Emys geoffroana 22: 125.
Emys gibba 22; 125.
Emys mobilensis 51: 173.
Emys nasuta 22: 125.
Emys picta B1: 1.
Emys radiolata 22: 125.
Emys ruficeps 22: 125.
Emys subnigra 15: 235.
Emys virgulata B1: 1.
Endryas quinquilineatus 50: 11.
Engystoma areolata 22: 115;
23: 115.
Engystoma carolinense 21: 47;
21: 53; 21: 69; 21: 85; 21:
199; 22: 115; 23: 9; 23:
165.
Engystoma ornata 51: 159.
Engystoma ovale 27; 77.
Engystoma rugosum 23: 165.
Engystoma texense 21: 53; 22:
115; 23: 115.
Ensatina croceater 43: 55.
Ensatina eschscholtzii 43: 55;
47: 169.
Ensatina klauberi 43: 55.
Ensatina sierrae 43: 55.
Enulius unicolor 72: 79.
Epicrates cenchria 29: 221.
Epicrates chrysogaster 29: 215.
Epicrates fordii 29: 215.
Epicrates monensis monensis
90: 985.
Eremias guttulata watsonae
83: 477.
Ergilemys 96: 567.
Erythrolamprus aesculapii
aesculapii 49: 43.
Erythrolamprus aesculapii
impar 49: 43.
Endryas amarali 86: 231.
Eugonglylus 98: 221.
Eumeces altamirani 49: 55.
Eumeces anthracinus 23: 115;
46: 129; 57: 123.
Eumeces brevilineatus 21:
165; 23: 115.
Eumeces brevirostris 46: 129;
46: 175.
Eumeces callicephalus 46:
175; 49: 55; 53: 125.
Eumeces copei 46: 129; 49:
55; 53: 119.
Eumeces dicei 46: 175.
Eumeces fasciatus 15: 121;
21:165; 22:129; 23:9; 50:137.
Eumeces guttulatus 23: 115.
Eumeces humilis 46: 175.
Eumeces inexpectatus 58: 131.
Eumeces kishinouyei 14: 189.
Eumeces laticeps 59:165; 64:59.
Eumeces lynxe lynxe 56: 109.
Eumeces managuae 46:67; 49:55.
Eumeces multivirgatus 23: 115.
Eumeces obsoletus 21:69; 23:115.
Eumeces ochoteranae 46: 129;
46: 175; 74: 37.
Eumeces pachyurus 23: 115.
Eumeces parviauriculatus 46:
175; 53: 125.
Eumeces parvulus 46: 175.
Eumeces quinquelineatus 20:
1; 21: 47; 21: 69; 21: 85;
21: 165; 22: 115; 23: 115.
Eumeces schmidti 46: 67.
Eumeces schneideri 83: 477.
Eumeces schwartzei 46:67; 49:55.
Eumeces scutatus 46: 67.
Eumeces septentrionalis 46:
129; 76: 159.
Eumeces taeniolatus 46: 67.
Eumeces tetragrammus 21: 47;
23: 115.
Eupemphix 84: 147.
Eupemphix bolbodactyla 98: 657.
Eupemphix olfersioidews 84:147.
Eupemphix pustulosus 37: 65;
55: 151.
Eupemphix schereri 55: 151.
Eupemphix trinitatus 29: 221.
Eupsophus calcaratus 95: 594;
98: 411.
Eupsophus coppingeri 93: 920.
Eupsophus grayi insularis 95:
594.
Eupsophus insularis 95: 594;
98: 411.
Eupsophus juminensis 98: 774.
Eupsophus migueli 95: 594;
98: 411.
Eupsophus peruanua 98: 774.

- Eupsophus roseus* 95: 594; 98: 411.
Eupsophus vanzolinii 93: 920.
Eupsophus vittatus 95: 594; 98: 411.
Eupsophus wettsteini 98: 774.
Eurycea 40: 123.
Eurycea bislineata bislineata 33: 129; 50: 137; 50: 141; 62: 89; 69: 93.
Eurycea bislineata cirrigera 33: 129; 62: 89.
Eurycea bislineata major 62: 89.
Eurycea bislineata rivicola 62: 89.
Eurycea bislineata wilderae 33: 129; 62: 89.
Eurycea gutto-lineata 33: 129.
Eurycea latitans 63: 51.
Eurycea longicauda 33: 129; 50: 137.
Eurycea longicauda guttolineata 56: 167.
Eurycea lucifuga 57: 31.
Eurycea melanopleura 50: 141.
Eurycea multiplicata 54: 77.
Eurycea nana 63: 51.
Eurycea neotenes 50: 141; 63: 51.
Eurycea pterophila 63: 51.
Eurycea quadridigitata quadridigitata 50: 141.
Euspondylus guentheri 57: 97; 99: 214.
Eutaenia brachystoma 58: 147.
Eutaenia butleri 58: 147.
Eutaenia dorsalis 21: 69.
Eutaenia elegans marcia 21: 47.
Eutaenia eques 21: 69; 21: 85.
Eutaenia marcia 21: 69.
Eutaenia proxima 21: 47; 21: 69; 21: 85.
Eutaenia sackeni 23: 9.
Eutaenia sirtalis 20: 1; 21: 47; 23: 9.
Eutaenia sirtalis parietalis 21: 69; 21: 85.
Eutaenia sirtalis sirtalis 21: 69.
Eutaenia saurita 20: 1.
Eutania brachystoma 16: 83.
Eutania sirtalis 16: 83.
Exostinus lancensis 86: 105.
Exostinus rugosus 86: 105.
Exostinus serratus 86: 105.
Farancia abacura 21: 47; 22: 129; 23: 9.
Ficimia 56: 109.
Ficimia desertorum 49: 51.
Ficimia elalocroma 49: 51.
Ficimia olivacea 49: 51.
Ficimia ornata 49: 51.
Ficimia publia 49: 51.
Ficimia quadrangularis 49: 51.
Ficimia variegata 49: 51.
Garbeana 55: 151.
Gastrophryne aequatorialis 23: 165.
Gastrophryne albopunctata 23: 165.
Gastrophryne areolata 23: 165.
Gastrophryne borneensis 23: 165.
Gastrophryne carolinensis 23: 165; 58: 89; 74: 157.
Gastrophryne elegans 23: 165.
Gastrophryne leucosticta 23: 165.
Gastrophryne microps 23: 165.
Gastrophryne muelleri 23: 165.
Gastrophryne olivacea 74: 157.
Gastrophryne ovale 27: 77.
Gastrophryne ovalis 23: 165.
Gastrophryne pictiventris 23: 165.
Gastrophryne texana 23: 165.
Gastrophryne usta 23: 165.
Gastrotheca 90: 60.
Gastrotheca coronata 52: 187.
Gastrotheca griswoldi 98: 774.
Gastrotheca marsupiata 100: 337.
Geckolepis 57: 1.
Gehyra 54: 195.
Gekko athymus 75: 67.
Gekko pumilus 75: 67.
Geochelone demissa 96: 567.
Geochelone elephantopus abingdoni B8: 108.
Geochelone emys 96: 567.
Geochelone gigantea B8: 108; B8: 129.
Geochelone impressa 96: 567.
Geochelone insolitus 96: 567.
Geochelone kalsanensis 96: 567.
Geochelone lunanensis 96: 567.
Geochelone oskarkuhni 96: 567.
Geochelone radiata B8: 108.
Geochelone sulcata 96: 567.
Geochelone ulanensis 96: 567.
Geochelone yniphora B8: 108.
Geochelone yunanensis 96: 567.
Geocrinia laevis 95: 423.

- Geoemyda spengleri* 15: 235.
Geoemyda spinosa 15: 235.
Geoemys spengleri 15: 235.
Geophis anocularis 33: 127.
Geophis dubius 33: 127.
Geophis godmani 33: 127.
Geophis nasalis 62: 165.
Geophis omiltemana 72: 79.
Geophis rostralis 33: 127.
Gerrhonotus burnettii 15: 37.
Gerrhonotus caeruleus 15: 37.
Gerrhonotus caeruleus
caeruleus 58: 5.
Gerrhonotus caeruleus palmeri
58: 5.
Gerrhonotus caeruleus
principis 58: 5.
Gerrhonotus caeruleus
shastensis 58: 5.
Gerrhonotus caeruleus
utahensis 58: 5.
Gerrhonotus kingii 53: 125.
Gerrhonotus liocephalus
liocephalus 74: 37.
Gerrhonotus nobilis 53: 125.
Glauconia dulcis 21:47; 21:69.
Glauconia emini 54: 175.
Glyphodon tristis 27: 201.
Gonatodes notatus 41: 53.
Gonyocephalus spinipes 27: 201.
Gopherus agassizi B8: 108.
Gopherus berlandieri 96: 567.
Gopherus flavomarginatus B8:108.
Gopherus polyphemus 48:79;B8:108.
Graptemys 93: 339.
Graptemys barbouri 76: 153.
Graptemys flavimaculata 76: 153.
Graptemys geographica 21: 69.
Graptemys nigrinoda 76: 153.
Graptemys oculifera 76: 153.
Graptemys pseudogeographica B1:1.
Graptemys pseudogeographica
pseudogeographica 56: 168.
Graptemys pulchra 23: 9; 76:
153.
Gymnodactylus amarali 38: 101.
Gymnodactylus pelagicus 27: 201.
Gymnodactylus rubidus 64: 91.
Gymnodactylus serpensinsula
64: 91.
Gymnophis nicefori 37: 125.
Gymnophthalmus 53: 19.
Gymnophthalmus birdi 78: 141.
Gymnophthalmus lineatus 78: 141.
Gymnophthalmus luetkeni 78: 141.
Gymnophthalmus pleei 78: 141.
Gymnophthalmus pleei luetkeni
78: 141.
Gymnophthalmus pleei nesydrion
78: 141.
Gymnophthalmus pleei pleei
78: 141.
Gymnophthalmus speciosus 78:141.
Gymnophthalmus sumichrasti
78: 141.
Gymnophthalmus underwoodi 78:141.
Gyrinophilus danielsi 33: 129.
Gyrinophilus porphyriticus
25: 135; 33: 129.
Hadrianus 96: 567.
Haideotriton wallacei 76: 153.
Haldea striatula 21: 47; 21:
69; 21: 85; 22: 129; 23: 9;
23: 115.
Helminthophis albirostris 45:173.
Helminthophis bondensis 45: 173.
Helminthophis canellei 45: 173.
Helminthophis emunctus 45: 173.
Helminthophis flavoterminalis
39: 123.
Helminthophis frontalis 39: 123;
45: 173.
Helminthophis petersii 45: 173.
Helminthopis 36: 185.
Helocaetes feriarum 68: 155.
Heloderma horridum 44: 73;
53: 119; 74: 37.
Heloderma suspectum 53: 119.
Hemidactylum 43: 55.
Hemidactylum scutatum 15:
121; 31: 51; 50: 137.
Hemidactylum scutatum 20: 1.
Hemidactylus 57: 1.
Hemidactylus brookii 49: 59.
Hemidactylus citernil 49: 59.
Hemidactylus frenatus 27:
201; 74: 37.
Hemidactylus greeffi 54: 195.
Hemidactylus maboia 28: 71.
Hemidactylus mabouia 49: 59;
51: 147; 90: 985.
Hemidactylus mandanus 49: 59.
Hemidactylus parkeri 49: 59.
Hemidactylus puccuonii 49: 59.
Hemidactylus turcicus 61: 159.
Hemiphractus 52: 187.
Heosemys 15: 235.
Heosemys depressa 15: 235.
Heosemys grandis 15: 235.

- Heosemys spinosa* 15: 235.
Herpetodryas carinatus 28: 71.
Hesperotestudo 96: 567.
Heterodon 18: 73.
Heterodon browni 16: 123.
Heterodon contortrix 33: 129;
 50: 137.
Heterodon nasicus nasicus
 76:159.
Heterodon platirrhinos 15: 121.
Heterodon platyrhinos 20: 1;
 21: 47; 21: 69; 21: 85; 22:
 129; 23: 9.
Heterodon simus 22:129; 23:9.
Hinulia elegans 58: 47.
Hispaniolus 41: 49.
Hispaniolus pratensis 41: 49.
Holbrookia 53: 119.
Holbrookia maculata
 approximans 53: 125.
Holbrookia maculata lacerata
 21: 69.
Holbrookia maculata maculata
 44: 11.
Holbrookia propinqua 21:47;
 40: 57.
Holbrookia propinqua propinqua
 45: 15.
Holbrookia propinqua stonei
 45: 15.
Holbrookia texana 21:69; 40:57.
Holoectes clarkii 21: 53.
Homalocranium atrocinctum
 56: 109.
Homalocranium michoacanense
 56: 109.
Homopholis wahlbergii arnoldi
 57: 1.
Homopholis wahlbergii
 wahlbergii 57: 1.
Homolepida crassicauda 27: 201
Homophis bergeri B8: 108.
Hydraspis galeata 22: 125.
Hydrodynastes bicinctus 82:
 847.
Hydromedusa tectifera 99: 472.
Hydrophis ornatus inornatus 60:1.
Hydrophis ornatus maresinensis
 60: 1.
Hydrophis ornatus ocellatus
 60: 1.
Hydrophis ornatus ornatus 60: 1.
Hydrophis semperi 51: 209.
Hydrosaurus amboinensis 24: 15.
Hydrosaurus microlophus 24: 15.
Hydrosaurus pustulosus 24: 15.
Hydrosaurus weberi 24: 15.
Hyla 23:115; 29:85; 44:11; 52:187
Hyla alleei 67: 159.
Hyla angularis 58: 53.
Hyla angustilineata 93: 1177.
Hyla arborea 74: 157.
Hyla arboricola 67: 159.
Hyla arenicolor 21: 53; 43:
 55; 43: 73; 53: 125; 74: 157.
Hyla arfakiana 58: 53.
Hyla astartea 98: 657.
Hyla aurea aurea 63: 131.
Hyla aurea cyclorhynchus 63:131.
Hyla aurea raniformis 63: 131.
Hyla aurea ulongae 63: 131.
Hyla avivoca 41: 89; 74: 157.
Hyla baudinii 53: 19; 74: 157.
Hyla becki 58: 53.
Hyla bistrincta 50: 43.
Hyla blairi 93: 153.
Hyla bocourti 67: 159.
Hyla brachypus 91: 965.
Hyla brongersmani 58: 53.
Hyla canadensis 69: 169.
Hyla cardenasi 67: 159.
Hyla carolinensis 8: 75.
Hyla carvalhoi 98: 657.
Hyla ceratophrys 55: 151.
Hyla chica 37: 65.
Hyla chloronota 58: 53.
Hyla cinerea 13: 75; 21: 47;
 21: 53; 21: 69; 22: 115; 22:
 129; 23: 9; 37: 141; 74: 157.
Hyla cinerea cinerea 50: 9.
Hyla cinerea evittata 33: 129.
Hyla circumdata 98: 657.
Hyla coerulea 27: 201.
Hyla copii 21: 53.
Hyla chrysoscelis 23: 115.
Hyla crepitans 29:221; 93:153.
Hyla crucifer 33: 129; 41:
 89; 50: 137; 74: 157; 91:123.
Hyla crucifer crucifer 59:165.
Hyla crucifera crucifera 69: 93;
 76: 159.
Hyla darlingtoni 58: 53.
Hyla debilis 93: 1177.
Hyla dolichophis 91: 965.
Hyla dominicensis 45: 191.
Hyla echinata 93: 1177.
Hyla elacochroa 93: 1177.
Hyla erythromma 50: 43.
Hyla euphorbeacea 67: 159.

- Hyla euthysanota* 41: 123; 49: 43; 67: 159.
Hyla everetti 58: 53.
Hyla evittata 13: 75; 15: 199; 15: 121; 50: 9; B1: 1.
Hyla eximia 67: 159.
Hyla femoralis 21: 53; 22: 129; 23: 9; 41: 89; 74: 157.
Hyla femoralis chrysoscelis 23: 115; 60: 47.
Hyla fimbrimembra 93: 1177.
Hyla gabbii 37: 65; 49: 43.
Hyla giesleri 93: 655.
Hyla gratiosa 23: 9; 74: 157.
Hyla heilprini 37: 65.
Hyla hobbsi 91: 123.
Hyla hylax 98: 657.
Hyla infrafronata militaria 91: 965.
Hyla izecksohni 98: 657.
Hyla kinghorni 63: 131.
Hyla krefftii 27: 201.
Hyla labialis 90: 60.
Hyla lafrantzi 67: 159.
Hyla larinopygion 95: 557.
Hyla latopalmata 63: 131.
Hyla legleri 93: 1177.
Hyla leonhard-schultzi 59: 43.
Hyla leucophyllata 93: 153.
Hyla loquax 93: 1177.
Hyla lythrodes 93: 1177; 99: 51.
Hyla marianae 90: 194.
Hyla mathiassoni 91: 123.
Hyla microps 93: 655.
Hyla miliaria 93: 1177.
Hyla militaria 91: 965.
Hyla montana 58: 53.
Hyla nanuzae 98: 657.
Hyla nasus 95: 377; 99: 42.
Hyla ocellifera 64: 33.
Hyla papuensis 58: 53.
Hyla phaeocrypta 41: 89.
Hyla phlebodes 50: 43.
Hyla pickeringii 13: 75; 15: 121; 20: 1.
Hyla pictipes 93: 1177.
Hyla pinorum 50: 43.
Hyla prosoblepon 37: 65; 64: 33.
Hyla pseudopuma 93: 1177.
Hyla pugnax 93: 1177.
Hyla punctata 91: 123; 93: 153.
Hyla raniformis 74: 157.
Hyla ranoides 95: 377; 99: 42.
Hyla regilla 21: 53; 43: 55; 43: 73; 67: 159; 74: 157.
Hyla reticulata 55: 151.
Hyla richardtaylori 93: 1177.
Hyla rivularis 93: 1177.
Hyla robertmertensi 50: 43.
Hyla rosenbergi 37: 65.
Hyla rostrata 93: 153.
Hyla rozellae 67: 159.
Hyla ruficollis 67: 159.
Hyla rufiocularis 93: 1177; 99: 51.
Hyla salvadorensis 67: 159.
Hyla salvavida 99: 51.
Hyla schmidtorum 99: 51.
Hyla semifasciata 21: 53.
Hyla septentrionalis 29: 215; 69: 169.
Hyla smaragdina 56: 49.
Hyla soralia 99: 51.
Hyla spinipollex 49: 43; 67: 159.
Hyla squirella 21: 47; 21: 53; 22: 115; 22: 129; 23: 9; 74: 154.
Hyla stadelmani 49: 43.
Hyla staufferi 50: 43; 53: 19; 93: 1177.
Hyla taeniopus 50: 43.
Hyla thysanota 93: 1177.
Hyla tica 93: 1177.
Hyla triseriata 44: 11; 68: 155.
Hyla underwoodi 50: 43.
Hyla uranochroa 67: 159; 93: 1177; 99: 51.
Hyla vagabunda 58: 53.
Hyla valancifer 93: 1177.
Hyla vanvlietti 21: 53.
Hyla versicolor 15: 121; 20: 1; 23: 9; 33: 129; 41: 89; 74: 157.
Hyla versicolor chrysoscelis 21: 53; 21: 69; 21: 85; 23: 115; 60: 47.
Hyla versicolor phaeocrypta 41: 89.
Hyla versicolor sandersi 60: 47.
Hyla versicolor versicolor 50: 137; 60: 47; 76: 159.
Hyla walkeri 67: 159.
Hyla wandae 91: 123; 93: 153.
Hyla wilderi 90: 194.
Hyla wirzi 58: 53.
Hyla wrightorum 67: 159.
Hyla xanthosticta 93: 1177.
Hylactophryne augusti 93: 928.
Hylaplesia nigriventris 98: 657.
Hylarana 55: 49.
Hylella azteca 56: 49.
Hylella fleischmanni 37: 65.
Hylella parabambae 64: 33.

- Hylella parambae* 64: 33.
Hylella picta 50: 43; 56: 49.
Hylella prosoblepon 37: 65.
Hylella sumichrasti 56: 49.
Hylella wolterstorffi 58: 53.
Hylixelalus beebei 84: 147.
Hylixelalus chochoensis 84: 147.
Hylixelalus collaris 84: 147.
Hylixelalus granuliventris 84:147.
Hylodes 87:81; 98:657; 98:774.
Hylodes babax 95: 377; 99:100.
Hylodes brocchi 93: 928.
Hylodes calcitrans 54: 87.
Hylodes charadranaetes 99:100.
Hylodes cruentus 94: 413.
Hylodes frater 94: 404.
Hylodes glabrus 99: 100.
Hylodes gravenhorstii 99: 42.
Hylodes lateristrigatus 95: 377; 99: 100.
Hylodes maculatus 69: 169; 76: 159.
Hylodes magalhaesi 99: 100.
Hylodes martinicensis 86: 231; 99: 42.
Hylodes melanostictus 93:928.
Hylodes mertensi 95: 377.
Hylodes nasus 95: 377.
Hylodes ornatus 95:377; 99:100.
Hylodes otavioi 99: 100.
Hylodes phyllodes 99: 100.
Hylodes platyrhynchus 93: 928.
Hylodes pulcher 95:377; 99:100.
Hylodes reguis 95:377; 99:100.
Hylodes surdus 93: 327.
Hylodes truncatus 95: 377.
Hylodes vanzolinii 95:377;99:100.
Hylopsis buckleyi 95: 557.
Hyloxalus bocagei 84: 147.
Hyloxalus fuliginosus 55: 151; 84: 147.
Hyloxalus vergeli 84: 147.
Hynobius 30: 123.
Hyperolius coccotis 51: 213.
Hyperolius microps 45: 61; 51: 213.
Hyperolius parkeri 51: 213.
Hyperolius petersii 45: 61.
Hyperolius poweri 51: 213.
Hyperolius pusillus 51: 213.
Hyperolius pygmaeus 45: 61.
Hyperolius translucens 51:213.
Hyperolius tuberilinguis 51:213.
Hyperolius usaramoae 45: 61; 51: 213.
Hypnale hypnale 91: 963.
Hypnale nepa 90:1002; 91:963.
Hypnale walli 90:1002; 91:963.
Hypodictyon ridens 94: 413.
Hypopachus 50: 43.
Hypopachus aquae 65: 1.
Hypopachus barberi 53: 19; 54: 125; 65: 1.
Hypopachus championi 53: 19; 54: 125; 67: 159; 74: 195.
Hypopachus cuneus 21:53; 53:19.
Hypopachus cuneus cuneus 54:125.
Hypopachus cuneus nigroreticulatus 54: 125.
Hypopachus globosus 53: 19.
Hypopachus globulosus 54: 125; 65: 1.
Hypopachus inguinalis 53: 19; 54: 125; 65: 1.
Hypopachus maculata 67: 159.
Hypopachus oxyrhinus 53: 19.
Hypopachus pearsei 27: 77.
Hypopachus simus 54:125; 65:1.
Hypopachus variolosus 53: 19.
Hypsigena 67: 69.
Ialtris dorsalis 41: 53; 41: 127; 45: 189.
Ialtris parishii 45: 189.
Idiotyphlops emunctus 45: 173.
Idiotyphlops flavoterminalis 39: 123.
Iguana delicatissima 30: 97.
Iguana iguana 29: 221.
Iguana iguana iguana 51: 147.
Iguana rhinolopha 30:97; 53:119.
Iguana tuberculata 29: 221.
Indotestudo kaiseni 96: 567.
Indotestudo nanus 96: 567.
Isocelis maculata 68: 11.
Istiurus 24: 15.
Ixalus fergusonii 53: 105.
Ixalus fuscus 55: 71.
Ixalus pallidipes 21: 189.
Japalura brevipes 49: 117.
Japalura polygonata 49: 117.
Japalura splendida 49: 117.
Japalura swinhonis 49: 117.
Japalura yunnanensis 49: 117.
Kalophrynus pleurostigma pleurostigma 80: 67.
Kaloula pulchra hainana 51:127.
Kaloula pulchra pulchra 51:127.

- Kansuchelys chiayukuanensis* 96: 567.
Kansuchelys ovalis 96: 567.
Kansuchelys tsiyuanensis 96:567.
Kentropyx pelviceps 42: 153.
Kinosternon brevicaudatum 15: 235.
Kinosternon flavescens 21: 47; 21: 69.
Kinosternon flavescens flavescens 70: 2011.
Kinosternon flavescens spooneri 70: 201.
Kinosternon flavescens stejneri 70: 201.
Kinosternon integrum 74: 37.
Kinosternon longicaudatum 15: 235.
Kinosternon louisianae 21: 47; 21: 69.
Kinosternon odoratum 33: 129.
Kinosternon pennsylvanicum 15: 121; 20: 1.
Kinosternon sonoriense 70: 201.
Kinosternon subrubrum 33: 129.
Kinosternon subrubrum subrubrum 50: 137.
Kinosternon tricarinata 15:235.
- Lacerta amboinensis* 24: 15.
Lacerta homalocephala 15: 37.
Lacerta maculata 15: 239.
Lacerta punctata 15: 239.
Lacerta subviolacea 15: 239.
Lachesis atrox 32: 213.
Lachesis aurifer 32: 213.
Lachesis barbouri 32: 213.
Lachesis brachystoma 32: 213.
Lachesis godmani 32: 213.
Lachesis lanceolatus 29: 221; 32: 213.
Lachesis lansbergii 32: 213.
Lachesis mutus 32: 213.
Lachesis nummifer 32: 213.
Lachesis undulatus 32: 213.
Lampropeltis 18: 73; 53: 125.
Lampropeltis annulata 31: 99.
Lampropeltis calligaster 21:69; 44: 11.
Lampropeltis coccinea 32: 99.
Lampropeltis doliata 31: 99.
Lampropeltis doliata blanchardi 72: 79.
Lampropeltis doliatus 31: 99.
- Lampropeltis doliatus collaris* 15: 121.
Lampropeltis doliatus doliatus 15:121; 21: 47; 21: 69; 21:85.
Lampropeltis doliatus gentilis 21: 47.
Lampropeltis doliatus triangulus 15: 121.
Lampropeltis elapsoides 31: 99.
Lampropeltis getula sayi 21: 69; 21: 85.
Lampropeltis getulus getulus 15: 121; 33: 129; 58: 89.
Lampropeltis getulus holbrooki 21: 47.
Lampropeltis getulus niger 33: 129.
Lampropeltis mexicana 35: 226.
Lampropeltis rhombomaculata 33: 129; 57: 124.
Lampropeltis rhombomaculatus 15: 121.
Lampropeltis triangulum amaaura 31: 99.
Lampropeltis triangulum gentilis 31: 99.
Lampropeltis triangulum triangulum 31: 99; 33: 129; 50: 137.
Lampropholis delicata 98: 221.
Lampropholis quichenoti 98: 221.
Lamprosoma annulatum 56: 109.
Lamprosoma episcopum 56: 109.
Latastie longicaudata 26: 145.
Lathrogecko sanctae-martae 29: 87.
Lathrogecko xanthostigma 29: 87.
Leimadophis 35: 219.
Leimadophis alleni 41: 53.
Leimadophis andicolus 28: 149.
Leimadophis parvifrons lincolni 44: 89.
Leimadophis parvifrons parvifrons 44: 89.
Leimadophis parvifrons proteus 41: 53.
Leimadophis parvifrons tortiganus 44: 89.
Leimadophis taeniurus 28: 149.
Leimadophis taeniurus bipraeocularis 50: 11.
Leiocephalus anonymus 97: 827.
Leiocephalus apertosulcus 97: 827.
Leiocephalus astictus 73: 103.

- Leiocephalus barahonensis* 41: 53; 92: 272.
Leiocephalus beatanus 41: 53; 44: 89.
Leiocephalus carinatus 29: 215; 73: 103; 97: 827.
Leiocephalus cubensis 72: 139; 73: 67; 97: 827.
Leiocephalus cubensis cubensis 73: 103.
Leiocephalus cuneus 97: 827.
Leiocephalus etheridgei 97: 827.
Leiocephalus jamaicensis 97: 827.
Leiocephalus koopmani 81: 23.
Leiocephalus lunatus 92: 272.
Leiocephalus macropus 73: 103; 81: 23.
Leiocephalus macrops asbolomus 81: 23.
Leiocephalus macropus hoplites 72: 139.
Leiocephalus macropus hyacinthurus 72: 139.
Leiocephalus macropus immaculatus 72: 139.
Leiocephalus macropus koopmani 72: 139.
Leiocephalus macropus macropus 72: 139; 73: 67.
Leiocephalus melanochlorus 29:215; 45:177; 92:272; 97:827.
Leiocephalus partius 97: 827.
Leiocephalus personatus 41: 49; 41: 53; 82: 777; 92: 272.
Leiocephalus personatus barahonensis 45: 177.
Leiocephalus personatus beatanus 45: 177.
Leiocephalus personatus mentalis 45: 177.
Leiocephalus personatus personatus 45: 177.
Leiocephalus personatus scalaris 45: 177.
Leiocephalus personatus semilineatus 45: 177.
Leiocephalus personatus vinculum 45: 177.
Leiocephalus pratensis 92: 272.
Leiocephalus raviceps 97: 827.
Leiocephalus raviceps jaumei 81: 23.
Leiocephalus raviceps klinikowskii 73: 67; 81: 23.
Leiocephalus raviceps raviceps 73: 67; 81: 23.
Leiocephalus raviceps uzzelli 73: 67; 81: 23.
Leiocephalus rhtidira 92: 272.
Leiocephalus schreibersii 41:53; 45: 177; 92: 272.
Leiocephalus semilineatus 41:53; 94: 827; 92: 272.
Leiocephalus stictigaster 72: 139; 73: 67; 81: 23.
Leiocephalus stictigaster exotheotus 73: 103.
Leiocephalus stictigaster lucianus 73: 103.
Leiocephalus strictigaster sierrae 73: 103.
Leiocephalus stictigaster stictigaster 73: 103.
Leiocephalus vinculum 41: 53.
Leiocephalus vinculum altavelensis 92: 272.
Leiocephalus vinculum endomycnus 92: 272.
Leiocephalus vinculum vinculum 92: 272.
Leiocephalus vittatus 73: 67.
Leiopisma albertisii 27:201.
Leiopisma beccarii 24: 15.
Leiopisma cyanogaster 27:201.
Leiopisma fuscum 24: 15; 27: 201.
Leiopisma hawaiiensis 52: 1.
Leiopisma laterale 21: 165; 22: 115; 23: 115.
Leiopisma noctua 52: 1.
Leiopisma peronii 27: 201.
Leiopisma pullum 24: 15.
Leiopisma 76: 69.
Leiopisma acrinasum 98:221.
Leiopisma alazon 98: 221.
Leiopisma assatum assatum 52: 187.
Leiopisma austrocaledonicum 98: 221.
Leiopisma baudini 98: 221.
Leiopisma cherriei 52: 187.
Leiopisma chloronoton 98:221.
Leiopisma coventryi 98:221.
Leiopisma deplanchei 98:221.
Leiopisma duperreyi 98:221.
Leiopisma elegantoides lobulus 58: 47.
Leiopisma entrecasteauxii 98: 221.

- Leiolopisma eunica* 40: 179.
Leiolopisma euryotis 98: 221.
Leiolopisma fallai 98: 221.
Leiolopisma fasciolare 98:221.
Leiolopisma gracile 54: 193.
Leiolopisma gracilicorpus 98:221.
Leiolopisma grande 98: 221.
Leiolopisma greeri 98: 221.
Leiolopisma homalonotum 98:221.
Leiolopisma infrapunctatum
 98; 221.
Leiolopisma kohtaoensis 40: 179.
Leiolopisma laterale 33: 129;
 40: 179; 44: 11.
Leiolopisma lichenigerum 98:221.
Leiolopisma lineocellatum 98:
 221.
Leiolopisma mariae 54: 193.
Leiolopisma melanostictum 40:
 179.
Leiolopisma metallicum 98:221.
Leiolopisma nigriplantare 98:
 221.
Leiolopisma nigrofasciolatum
 54: 193; 98: 221.
Leiolopisma noctua 52: 1.
Leiolopisma novacaledonicum
 98: 221.
Leiolopisma ocellatum 98: 221.
Leiolopisma otagense 98: 221.
Leiolopisma palfreymani 98:221.
Leiolopisma platynotum 98:221.
Leiolopisma prehensicauda 58:47.
Leiolopisma pretiosum 98: 221.
Leiolopisma pulchellum
levitoni 76: 69.
Leiolopisma rupicola 40: 179.
Leiolopisma semperi 76: 69.
Leiolopisma spenceri 98: 221.
Leiolopisma steindachneri 98:221.
Leiolopisma subvittatum 76: 69.
Leiolopisma suteri 98: 221.
Leiolopisma telfairi 98: 221.
Leiolopisma tricolor 98: 221.
Leiolopisma trilineatum 98:221.
Leiolopisma unicolor 53: 119;
 58: 89.
Leiolopisma variabile 98: 221.
Leiolopisma zamboangensis 76:69.
Leiolopisma zia 98: 221.
Leiopelma archeyi 62: 57.
Leiopelma hamiltoni 62: 57.
Leiopelma hochstetteri 62: 57.
Leiosaurus 34: 139.
Leiosophis gigas 82: 847.
- Lejosophis bicinctus* 82: 847.
Lejosophis gigas 82: 847.
Lemiadophis melanotus 29: 221.
Lemiadophis stahli 30: 97.
Leptodactylus 44: 11; 93:
 1177; 99: 42.
Leptodactylus albilabris 28: 71;
 30: 97; 87: 327; 96: 270.
Leptodactylus amazonicus 96:270.
Leptodactylus andicola 100: 337.
Leptodactylus bolivianus 37: 65;
 87: 327; 96: 560.
Leptodactylus bufonius 87: 327.
Leptodactylus caliginosus 84:
 163.
Leptodactylus chaquensis 87:
 327; 96: 560.
Leptodactylus curtus 84: 163.
Leptodactylus dantasi 96: 560.
Leptodactylus discodactylus
 84: 163; 87: 81.
Leptodactylus elenae 96: 270.
Leptodactylus fragilis 96: 270.
Leptodactylus furnarius 96:270.
Leptodactylus fuscus 83: 221;
 84:163; 87:327; 96:270; B2:199.
Leptodactylus gaigeae 51: 41.
Leptodactylus geminus 87: 327;
 96: 270.
Leptodactylus goliath 84: 163.
Leptodactylus gracilis 87: 327;
 96: 270.
Leptodactylus hemidactyloides
 84: 163.
Leptodactylus hylaedactylus
 84: 163.
Leptodactylus inoptatus 30: 97.
Leptodactylus knudseni 87: 327.
Leptodactylus labialis 30: 97;
 87: 327.
Leptodactylus labrosus 84: 163.
Leptodactylus latinasus 87: 327.
Leptodactylus laurae 96: 270.
Leptodactylus lineatus 84: 163.
Leptodactylus longirostris
 96: 270.
Leptodactylus macrosternum
 96: 560.
Leptodactylus marmoratus 51:
 41; 84: 163; 87: 327.
Leptodactylus melanonotus 84:163;
 87:81: 87: 327; 96: 560.
Leptodactylus mystaceus 84:
 163; 87: 327; 96: 270.
Leptodactylus mystacinus 87:327.

- Leptodactylus "natalensis"* 87: 327.
Leptodactylus nigrescens 84:163.
Leptodactylus notoakites 96:270.
Leptodactylus ocellatus 87: 327; 96: 560.
Leptodactylus pentadactylus 87: 327.
Leptodactylus pentadactylus rubidoides 84: 163.
Leptodactylus podicipinus 87: 327; 93: 928; 96: 560.
Leptodactylus poecilochilus 96: 270.
Leptodactylus pulcher 84: 163.
Leptodactylus pustulatus 96:560.
Leptodactylus rhodomystax 84: 163; 96: 560.
Leptodactylus rhodonotus 84: 163; 87: 327.
Leptodactylus riveroi 96: 506.
Leptodactylus rubido 84: 163.
Leptodactylus sibilatrix 83: 221; B2: 199.
Leptodactylus spixi 96: 270.
Leptodactylus stenodema 84:163.
Leptodactylus tuberculosus 84: 163.
Leptodactylus validus 51: 147.
Leptodactylus ventrimaculatus 84: 163.
Leptodactylus vividis 96: 560.
Leptodactylus wagneri 84: 163; 87: 327; 93: 928; 96: 560.
Leptodeira 45: 83.
Leptodeira annulata 29: 221; 36: 185.
Leptodeira annulata cussuliris 72: 79.
Leptodeira annulata polysticta 54: 115.
Leptodeira annulata septentrionalis 54: 115.
Leptodeira annulata taylori 54: 115.
Leptodeira frenata 52: 187.
Leptodeira latifasciata 72: 79.
Leptodeira maculata 54: 115.
Leptodeira rhombifera 50: 11.
Leptodeira septentrionalis polysticta 72: 79.
Leptognathus atypicus 36: 185.
Leptognathus boettgeri 36: 185.
Leptognathus peruana 36: 185.
Leptognathus schunkii 36: 185.
Leptognathus latifasciatus 36: 185.
Leptognathus vagus 36: 185.
Leptomicrurus narduccii 59: 17.
Leptophis liocercus 29: 221.
Leptotyphlops albifrons 36: 185; 58: 29; 87: 167.
Leptotyphlops asbolepis 98: 204.
Leptotyphlops bilineata 28: 71; 79: 255.
Leptotyphlops bilineatus 98: 204.
Leptotyphlops calypso 98: 204.
Leptotyphlops columbi 58: 29; 98: 204.
Leptotyphlops dulcis 45: 151; 90: 209.
Leptotyphlops emini emini 54:175.
Leptotyphlops emini pembae 54: 175.
Leptotyphlops goudotii 98: 204.
Leptotyphlops humilis 45: 151; 98: 204.
Leptotyphlops leptepileptus 98: 204
Leptotyphlops maximus 45: 151.
Leptotyphlops munoai 90: 209.
Leptotyphlops myopica 45: 151.
Leptotyphlops nigricans 98: 204.
Leptotyphlops phenops bakewelli 72: 79.
Leptotyphlops pyrites 79: 255; 88: 367; 98: 204.
Leptotyphlops rubrolineatus 87: 167.
Leptotyphlops rufidorsum 58: 29.
Leptotyphlops rufidorsus 87: 167.
Leptotyphlops striatula 58: 29.
Leptotyphlops teaguei 87: 167.
Leptotyphlops tenella 58: 29.
Leptotyphlops tenellus 87: 167.
Leptotyphlops tessellatus 87: 167.
Leptotyphlops tricolor 87: 167.
Leuiperus mexicanus 54: 87.
Leurognathus marmoratus 25: 135; 33: 129; 76: 153.
Levirana vibicaria 35: 221.
Lialis burtonii 27: 201.
Liasis childreni 27: 201.
Liasis clarki 27: 201.
Liasis olivaceus 27: 201.
Limnodynastes 99: 42.
Linguelaps annulatus 21: 85.
Lioccephalus arenarius 29: 215.
Liodytes alleni 23: 9.
Liolaemus boulengeri 34: 139.

- Liolaemus elongatus* 34: 139.
Liolaemus kingii 34: 139.
Liolaemus lineomaculatus 34:139.
Liolaemus magellanicus 34: 139.
Liolaemus pictus 95: 594.
Liolaemus rothi 34: 139.
Liolepisma laterale 21: 47;
21:69; 21:85; 22:129; 23:9.
Liopelma 40; 123; 62: 57.
Liopeltis libertatis 23: 169.
Liopeltis major 23: 169.
Liopeltis vernalis 15:121; 20:1.
Liophis 82: 847.
Liophis almedensis 98: 295.
Liophis atahuallpae 35: 219.
Liophis dilepis 98: 295.
Liophis elapoides diastema
54: 119.
Liophis jaegeri 98: 295.
Liophis maryellenae 98: 295.
Liophis miliaris 98: 295.
Liophis poecilogyrus 98: 295.
Liophis taeniurus 36: 185.
Liophis tricinctus 54: 119.
Liophis viridis 98: 295.
Liotyphlops albirostris 45: 173.
Liotyphlops anops 50: 11.
Lipinia noctua 93: 350; 98: 221.
Lipinia pulchella 76: 69.
Lipinia semperi 76: 69.
Lithodytes 28:131; 98:774; 99:42.
Lithodytes discodactylus 87: 81.
Lithodytes latrans 21: 53;
21: 69; 23: 115.
Lithodytes lineatus 84:163; 87:81.
Lithodytes melanostictus 93: 928.
Lithodytes ricordii 23: 9.
Litoria bicolor 91: 965.
Litoria freycineti 99: 42.
Litoria infrafrenata militaria
91: 965.
Litoria militaria 91: 965.
Litoria splendida 99: 42.
Litoria thesaurensis 91: 965.
Lophosaura goodridgii 27: 9.
Lophura 24: 15.
Lophura concinna 25: 191.
Loxocemus sumichrasti 72: 79.
Lygodactylus capensis 26: 145.
Lygodactylus capensis
scheffleri 48: 195.
Lygodactylus fischeri
scheffleri 48: 195.
Lygodactylus grotei pakenhami
54: 175.
Lygodactylus ocellatus 48:195.
Lygodactylus picturatus
gutturalis 26: 145; 28: 195.
Lygodactylus picturatus
mombasicus 48: 195.
Lygodactylus picturatus
picturatus 26: 145; 48: 195.
Lygodactylus picturatus
sudanensis 48: 195.
Lygodactylus picturatus
ukerewensis 48: 195.
Lygodactylus somalicus
annectens 48: 195.
Lygodactylus somalicus
somalicus 48: 195.
Lygodactylus thomensis 48: 195.
Lygophus lineatus B2: 199.
Lygosoma annamiticum 40: 179.
Lygosoma anolis 58: 47.
Lygosoma arborens 80: 69.
Lygosoma assata taylori 74: 37.
Lygosoma assatum 54: 181.
Lygosoma auriculatum 76: 69.
Lygosoma cherriei cherriei
54: 181.
Lygosoma cherriei stuarti 54:181.
Lygosoma decipiens 80: 69.
Lygosoma diwata 80: 69.
Lygosoma elegans 58: 47.
Lygosoma elegantoides lobulus
58: 47.
Lygosoma flavipes 58: 47.
Lygosoma gracile 54: 193.
Lygosoma herberti 40: 179.
Lygosoma helenae 40: 179.
Lygosoma infralineolatum 76: 69.
Lygosoma jagori 80: 69.
Lygosoma laterale 15:121; B1: 1.
Lygosoma lednickyi 80: 69.
Lygosoma mariae 54: 193.
Lygosoma mindanensis 80: 69.
Lygosoma nigrofasciolatum 54:193.
Lygosoma nitens 76: 69.
Lygosoma parkeri 58: 47.
Lygosoma prehensicauda 58: 47.
Lygosoma pulchellum 58: 47.
Lygosoma pulchellum levitoni
76: 69.
Lygosoma pulchellum pulchellum
76: 69.
Lygosoma pulchellum taylori
76: 69.
Lygosoma quadrivittatum 76: 76.
Lygosoma quadrivittatum
infralineolatum 76: 69.

- Lygosoma quadrivittatum*
quadrivittatum 76: 69.
Lygosoma relictum 76: 69.
Lygosoma semperi 76: 69.
Lygosoma slevini 54: 193.
Lygosoma steerei 80: 69.
Lygosoma subvittatum 76: 69.
Lygosoma surdum 76: 69.
Lygosoma variegatum 80: 69.
Lygosoma zamboangensis 76: 69.
Lysoptychus lateralis 17: 17.
- Mabuya aenea* 29: 221.
Mabuya agilis 29: 215; 53:119.
Mabuya aurata 29: 221.
Mabuya aurata affinis 83: 477.
Mabuya brachypoda 74: 37.
Mabuya cepedei 29: 215.
Mabuya mabouya mabouya 51: 147.
Mabuya mabouya sloanei 90: 985.
Mabuya maculilabris albotaenia
albotaeniata 54: 175.
Mabuya quinquetaeniata 26: 145.
Mabuya sloanii 29: 215.
Macrolemys temminckii 99: 472.
Magnadigita adspersa 65: 1.
Magnadigita cerroensis 75: 71.
Magnadigita cuchumantana 65: 1.
Magnadigita dunni 65: 1.
Magnadigita engelhardti 65: 1.
Magnadigita flavimembris 65: 1.
Magnadigita franklini 65: 1.
Magnadigita helmrichi 65: 1.
Magnadigita hypacra 75: 71.
Magnadigita lincolni 65: 1.
Magnadigita macrinii 65: 1.
Magnadigita marmorea 75: 71.
Magnadigita morio 65: 1.
Magnadigita nigrescens 75: 71.
Magnadigita nigroflavescens 65:1.
Magnadigita omniumsanctorum 65:1.
Magnadigita pesrubra 75: 71.
Magnadigita robusta 65:1; 75:71.
Magnadigita rostrata 65: 1.
Magnadigita subpalmata 65: 1;
 75: 71.
Magnadigita torresi 75: 71.
Malaclemmys centrata 15: 121;
 B1: 1.
Malaclemmys littoralis
rhizophoarum 68: 157.
Malaclemmys macrospilota 23: 9.
Malaclemys geographica 20: 1.
Malaclemys pseudo-geographica
 20: 1.
- Malaclemys terrapin* 99: 472.
Malaclemys terrapin centrata
 68: 157.
Malaclemys terrapin
macrospilota 68: 157.
Malaclemys terrapin
rhizophorarum 68: 157.
Malaclemys terrapin tequesta
 68: 157.
Malacochersus tornieri 96: 567.
Manculus 50: 141.
Manolepis putnami 72: 79.
Manouria chiayukuanensis 96: 567.
Manouria ovalis 96: 567.
Manouria sharanensis 96: 567.
Manouria sichuanensis 96: 567.
Manouria tsiuyunensis 96: 567.
Manouria yushensis 96: 567.
Masticophis flagellum cingulum
 67: 247.
Masticophis flagellum
flavigularis 67: 247.
Masticophis flagellum lineatus
 72: 79.
Masticophis flagellum piceus
 67: 247.
Mastigodryas boddaerti dunni
 86: 231.
Mastigodryas bruesi 86: 231.
Mastigodryas pleei 86: 231.
Maticora bivirgata bivirgata
 57: 105.
Maticora bivirgata tetrataenia
 57: 105.
Maticora intestinalis
bilineata 57: IV; 57: 105.
Maticora intestinalis everetti
 57: 105.
Maticora intestinalis
immaculata 57: 105.
Maticora intestinalis
intestinalis 57: IV; 57: 105.
Maticora intestinalis
nigrotaeniata 57: 105.
Maticora philippina 57: 105.
Maticora philippinus 57: 105.
Megaelosia 98: 774.
Megaelosia goeldi 99: 100.
Megophrys boettgeri 39: 53.
Megophrys minor 39: 53.
Melanochelys trijuga 15: 235.
Melanosaurus maximus 86: 105.
Micrixalus borealis 55: 71.
Micrixalus dimunitiva 55: 71.
Micrixalus fuscus 55: 71.

- Micrixalus herrei* 55: 71.
Micrixalus opistborhodus 55: 71.
Micrixalus sarasinorum 55: 71.
Micrixalus saxicola 55: 71.
Micrixalus silvaticus 55: 71.
Micrixalus tenasserimensis
 55: 71.
Micrixalus torrentis 55: 71.
Microbatrachylus lineatissimus
 54: 87.
Microbatrachylus oaxacae 54: 87.
Microgecko helenae 83: 477.
Microhyla berdmorei 40: 179.
Microhyla fissipes 14: 189;
 51: 159.
Microhyla leucostigma 51: 191.
Microhyla malcolmi 40: 179.
Microhyla okinavensis 14: 189;
 51: 159.
Microhyla ornata 51: 159.
Microhyla undulata 51: 159.
Micropholis stowii 98: 1028.
Microps unicolor 23: 165.
Micrurus 49: 51.
Micrurus affinis affinis 55: 63.
Micrurus annellatus 59: 17.
Micrurus corallinus 36: 185.
Micrurus fitzingeri fitzingeri
 55: 63.
Micrurus fitzingeri
microgalbineus 55: 63.
Micrurus fulvius tenere 55: 63.
Micrurus langsdorffii 59: 17.
Micrurus nigrocinctus browni
 72: 79.
Molge pyrrhogaster ensicauda
 51: 159.
Molge pyrrhogastra
ensicaudatus 51: 159.
Monopeltis 59: 73.
Monoplocus 42: 153.

Nannobatrachus anamollaiensis
 55: 49.
Nannobatrachus beddomii 55: 49.
Nannobatrachus kempholeyiensis
 55: 49.
Nannophrys 55: 49.
Nanorana 55: 49.
Natrix 53: 125.
Natrix bisecta 15: 121.; B1: 1.
Natrix boulengeri 50: 125.
Natrix compressicauda 23: 9.
Natrix craspedogaster 50: 125.
Natrix erythrogaster 64: 60.

Natrix erythrogaster alta 76:
 169.
Natrix erythrogaster bogerti
 76: 169.
Natrix erythrogaster
transversa 76: 169.
Natrix fasciata 23: 9.
Natrix fasciata erythrogaster
 22: 129.
Natrix fasciata sipedon 20: 1.
Natrix inas 50: 125.
Natrix khasiensis gilhodesi
 50: 125.
Natrix kirklandi 20: 1.
Natrix leberis 20: 1.
Natrix mairii 23: 89.
Natrix rhombifera 20: 1.
Natrix septemvittata 33: 129;
 59: 137.
Natrix sipedon 15: 121; 22:
 129; 28: 61; 33: 129.
Natrix sipedon sipedon 50: 137.
Natrix taxispilota 22: 129.
Necrosaurus 86: 105.
Nectes obscurus 16: 51.
Nectes subasper 17: 51.
Nectophryne afra 51: 191.
Nectophryne batesi 51: 191.
Nectophryne gardineri 51: 191.
Nectophryne misera 51: 191.
Nectophryne parvipalmata 51: 191.
Nectophryne picturata 51: 191.
Nectophryne sundana 51: 191.
Nectophryne werthi 51: 191.
Nectophrynoides tornieri 51: 191.
Necturus alabamensis 51: 143.
Necturus beyeri 51: 143.
Necturus louisianensis 51: 143.
Necturus maculatus 20: 1.
Necturus maculosus 51: 143.
Necturus maculosus maculosus
 76: 159.
Neoseps 76: 153.
Niceforonia montium 98: 774.
Niceforonia wettsteini 98: 774.
Nicoria 15: 235.
Ninia sebae sebae 62: 165.
Nordenosaurus 86: 105.
Nordenosaurus magnus 86: 105.
Nyctibatrachus major 55: 49.
Nyctimantis 52: 187.
Nyctimantis papua 29: 85.
Nyctimantis rugiceps 29: 85.
Nyctimystes 29: 85.
Nyctimystes milneana 58: 53.

- Nyctimystes montana* 58: 53.
Nyctimystes papua 29:85; 58:53.
Nyctimystes semipalmata 58: 53.
- Odontophrynus* 80: 105.
Oedipina 75: 71.
Oedipina alforoi 34: 143; 65: 1.
Oedipina ignea 65: 1.
Oedipina poelzi 89: 289.
Oedipina syndactyla 61: 177.
Oedipina taylori 65: 1.
Oedipina uniformis 34: 143.
Oedipina vermicularis 34: 143.
Oedipus ahli 76: 289.
Oedipus barbouri 49: 43.
Oedipus chiropterus 49: 43.
Oedipus flavimembris 67: 159.
Oedipus franklini 67: 159.
Oedipus helmrichi 55: 159.
Oedipus lignicolor 76: 289.
Oedipus picadoi 61: 177.
Oedipus rex 34: 143; 61: 17.
Oedipus rostralis 61: 17.
Oedipus rufescens 52: 187.
Oedipus sulcatus 34: 143.
Oedipus townsendi 35: 5.
Oeidozuga laevis laevis 80: 65.
Oligodon analepticos 82: 763.
Oligodon brevicauda 82: 768.
Oligodon cyclurus 82: 763.
Oligosoma 98: 221.
Ololygon catharinae
 bocainensis 93: 655.
Ololygon catharinae brieni
 93:655.
Ololygon catharinae
 obtriangula 93: 655.
Ololygon catharinae simplex
 93: 655.
Ololygon catharinae
 trapicheiroi 93: 655.
Ololygon humilis 93: 655.
Ololygon opalina 93: 655.
Ololygon rubra 96: 59
Onychodactylus 30: 123.
Opheodrys aestivus 15: 121.
Opheodrys vernalis blanchardi
 76: 159.
Opheodrys vernalis vernalis
 76: 305.
Opheosaurus ventralis 21: 47;
 21: 69.
Ophibolus coccineus 23: 9.
Ophibolus gentilis 31: 99.
Ophibolus getulus 22: 129.
Ophibolus getulus sayi 23: 9.
Ophibolus rhombomaculatus 15: 36;
 15: 90.
Ophibolus triangulum mexicanus
 35: 226.
Ophisaurus harti 32: 142.
Ophisaurus ventralis 20: 1;
 21: 69; 22: 129; 23: 9; 53:
 119.
Opophisops elegans elegans
 80: 477.
Ophryoessoides tridescens 80:
 105.
Opisthotropis typica 51: 43.
Oreocalamus hanitschi 51: 43.
Oreophis boulengeri 35: 226.
Oreophryne annulata 80: 65.
Oreophryne brachypus 91: 965.
Oreophryne jeffersoniana 80: 65.
Oreophryne nana 80: 65.
Oreophryne rookmaakeri 80: 65.
Oreophryne visaya 80: 65.
Oreophryne variabilis 80: 65.
Oreophryne zimmeri 80: 65.
Osceola 31: 99.
Osceola doliata doliata 21: 69.
Osceola doliata triangula 20: 1.
Osteocephalus taurinus 91: 123.
Osteopilus brunneus 90: 194.
Oxybelis acuminatus 29: 221;
 36: 185.
Oxybelis aeneus auratus 72: 79.
Oxyrhina varians 53: 125.
Oxyrhopus petola 59: 17.
- Paludicola bufonina* 34: 139.
Paludicola illothus 35: 111.
Paludicola imitator 34: 157.
Paracrinia haswelli 95: 423.
Pedostibes altitudinus 51: 191.
Pedostibes everetti 51: 191.
Pedostibes hosii 51: 191.
Pedostibes kempfi 51: 191.
Pedostibes tuberculosus 51:191.
Parvimolge richardi 61: 177.
Pelamis platurus B2:199; B2:267.
Pelobius 99: 42.
Pelodryas caeruleus 99: 42.
Pelodryas militarius 91: 965.
Pelomedusa subrufa 99: 472.
Pelophryne 51: 191.
Pelophryne albotaeniata 51:191.
Pelophryne brevipes 51: 191.
Pelophryne guentheri 51: 191.
Pelophryne lighti 51: 191.

- Pelophryne macrotis* 51: 191.
Pelophryne maculata 51: 191.
Pelophryne misera 51: 191.
Pelophryne signata 51: 191.
Pelusios castaneus castanoides 99: 472.
Pelusios nigricans 15: 235.
Pelusios subniger 99: 472.
Perochirus 54: 195.
Petalognathus 36: 185.
Petalognathus nebulatus 29: 221.
Phaeognathus hubrichti 76: 153.
Phelsuma abboti abotti B8: 101.
Phelsuma abboti sumptio B8: 101.
Phelsuma astriata astriata B8: 101.
Phelsuma astriata semicarinata B8: 101.
Phelsuma laticauda B8: 101.
Phelsuma madagascariensis parkeri 54: 175.
Phelsuma sundbergi longinsulae B8: 101.
Phelsuma sundbergi sundbergi B8: 101.
Philautus doriae 40: 179.
Philautus eximius 53: 105.
Philautus femoralis 53: 105.
Philautus hansenae 40: 179.
Philautus nongkhorensis 40: 179.
Philautus palpebralis 40: 179.
Philautus variabilis 53: 105.
Phrynella pollicaris 51: 191.
Phryniscus 23: 165.
Phrynobatrachus acridoides 54: 175.
Phrynobatrachus natalensis 26: 145.
Phrynobatrachus pakenhami 54: 175.
Phrynohyas venulosa 93: 153.
Phrynops geoffroana 22: 125.
Phrynops hilarii 22: 125.
Phrynops rufipes 22: 125.
Phrynops tuberosa 22: 125.
Phrynops wagleri 22: 125.
Phrynopus juninensis 98: 774.
Phrynopus montium 98: 774.
Phrynopus nanus 90: 60.
Phrynopus peruanus 98: 774.
Phrynopus wettsteini 98: 774.
Phrynosoma 53: 119.
Phrynosoma asio 74: 37.
Phrynosoma brevicornis 45: 73.
Phrynosoma cornutum 21: 47; 21: 69; 21: 165; 45: 73.
Phrynosoma douglassii douglassii 53: 125.
Phrynosoma douglassii hernandesi 21: 165; 53: 125.
Phrynosoma orbiculare alticola 66: 27.
Phrynosoma orbiculare cortezii 66: 27.
Phrynosoma orbiculare orbiculare 53: 125; 66: 27.
Phrynosoma platyrhinos 35: 1.
Phrynosoma taurus 74: 37.
Phyllobates 84: 147; 93: 1177; 95: 557; 99: 42; 99: 214.
Phyllobates alagoanus 84: 147.
Phyllobates alboguttatus 84: 147.
Phyllobates anthonyi 84: 147.
Phyllobates aurotaenia 94: 67.
Phyllobates beatriciae 34: 157.
Phyllobates bromelicola 84: 147.
Phyllobates capixaba 84: 147.
Phyllobates carioca 84: 147.
Phyllobates infraguttatus 84: 147.
Phyllobates intermedius 84: 147.
Phyllobates kingsburyi 84: 147.
Phyllobates latinasus 84: 147.
Phyllobates lugubris 34: 157.
Phyllobates mandelorum 84: 147.
Phyllobates marchesianus 84: 147.
Phyllobates mertensi 84: 147.
Phyllobates nubicola 55: 71.
Phyllobates palmatus 84: 147.
Phyllobates peruvianus 84: 147.
Phyllobates pratti 84: 147.
Phyllobates ridens 94: 413.
Phyllobates riocosangae 84: 147.
Phyllobates sylvatica 84: 147.
Phyllobates taeniatus 84: 147.
Phyllobates talamancae 34: 157.
Phyllobates tatinasus 37: 65.
Phyllobates terribilis 94: 67.
Phyllobates trilineatus 84: 147.
Phyllobates trinitatus 84: 147.
Phyllobates truncatus 34: 157.
Phyllobates vertebralis 84: 147.
Phyllobates vittatus 94: 67.
Phyllodactylus bordai 74: 37.
Phyllodactylus delcampi 74: 37.
Phyllodactylus insularis 81: 419.
Phyllodactylus lanei 74: 37.
Phyllodactylus magnus 74: 37.
Phyllodactylus palmeus 81: 419.
Phyllodactylus tuberculatus 81: 419.
Phyllodromus pulchellum 84: 147.

- Phyllodromus vertebralis 84:147.
 Phyllomedusa boliviana 96: 59.
 Phyllomedusa duellmani 96: 59.
 Phyllomedusa hypochondrialis
 93: 153; 96: 59.
 Phyllomedusa pailona 96: 59.
 Phyllomedusa sauvagii 93: 153.
 Phyllomedusa tarsius 96: 59.
 Phyllomedusa trinitatis 96: 59.
 Phyllomedusa venusta 93: 1177;
 96: 59.
 Phyllorhynchus arenicola 67: 69.
 Phyllorhynchus browni browni
 67: 69.
 Phyllorhynchus browni fortitus
 67: 69.
 Phyllorhynchus browni lucidus
 67: 69.
 Phyllorhynchus decurtatus
 decurtatus 67: 69.
 Phyllorhynchus decurtatus
 norrisi 67: 69.
 Phyllorhynchus decurtatus
 nubilis 67: 69.
 Phyllorhynchus decurtatus
 perkinsi 67: 69.
 Phymaturus flagellifer 34: 139.
 Phymaturus palluma 34: 139.
 Phymaturus patagonicus 34: 139.
 Phymaturus spurcus 34: 139.
 Physalaemus 93: 1177.
 Physalaemus cuvieri 98: 657.
 Physalaemus franciscaae 98: 657.
 Physalaemus maculiventris 98:657.
 Physalaemus nanus 98: 657.
 Physalaemus obtectus 98: 657.
 Physalaemus olfersi 98: 657.
 Physalaemus signiferus 98: 657.
 Physignathus cochinchinensis
 25: 191.
 Physignathus cocincinus
 caudicinctus 25: 191.
 Physignathus cocincinus
 cocincinus 25: 191.
 Physignathus cocincinus
 mentager 25: 191.
 Physignathus mentager 25: 191.
 Pipa pipa 44: 11.
 Pithecopus boliviana 96: 59.
 Pithecopus pailonus 96: 59.
 Pituophis deppei gibsoni 67: 159.
 Pituophis deppei lineaticollis
 67: 159; 72: 79.
 Pituophis melanoleucus
 melanoleucus 58: 89.
 Pituophis sayi sayi 44: 11.
 Pityophis catenifer sayi 21:
 47; 21: 69.
 Pityophis melanoleucus 23: 9.
 Platydictylus americana 81: 123.
 Platydictylus americanus
 cubanus 81: 123.
 Platydictylus milbertii 81: 123.
 Platymantis 99: 42.
 Platymantis acrochordus 81: 69.
 Platymantis aculeodactylus 81:69.
 Platymantis akarithymus 81: 69;
 91: 965.
 Platymantis boulengeri 81: 69;
 91: 965.
 Platymantis cornutus 95: 386.
 Platymantis corrugatus 95: 386.
 Platymantis corrugatus
 papuensis 81: 69.
 Platymantis dorsalis 81: 69;
 95: 386.
 Platymantis gilliardi 81: 69;
 91: 965.
 Platymantis guentheri 95: 386.
 Platymantis hazelae 95: 386.
 Platymantis ingeri 95: 386.
 Platymantis insulatus 95: 386.
 Platymantis lawtoni 95: 386.
 Platymantis levigatus 95: 386.
 Platymantis macroceles 91: 965.
 Platymantis macrops 81: 69.
 Platymantis magnus 91: 965.
 Platymantis meyersi 81: 69.
 Platymantis mimicus 81: 69;
 91: 965.
 Platymantis nexipus 91: 965.
 Platymantis nova-britanica
 81: 69.
 Platymantis papuensis
 papuensis 81: 69.
 Platymantis papuensis schmidti
 81: 69; 91: 965.
 Platymantis papuensis weberi
 81: 69.
 Platymantis parkeri 81: 69.
 Platymantis pelewensis 81: 69.
 Platymantis polillensis 95: 386.
 Platymantis rhipiphalcus 81: 69;
 91: 965.
 Platymantis solomonis 81: 69;
 91: 965.
 Platymantis spelaeus 95: 386.
 Platymantis subterrestris 95:386.
 Platymantis vitianus 91: 965.
 Platymantis weberi 81: 69.

- Platypholis altamirani* 49: 55.
Platypholis heterolepis 57: 1.
Platyplacopus kuehnei
carinatus 51: 127.
Platyplacopus kuehnei kuehnei
51: 127.
Platyplectrurus madurensis 55:
49.
Platysternon megacephalum
megacephalum 100: 624.
Platysternon megacephalum
peguense 100: 624.
Platysternon megacephalum
rogeli 100: 624.
Platysternon megacephalum
shiui 100: 624.
Platysternon megacephalum
trisernalis 100: 624.
Platythyra flavescens 70: 201.
Plectrohyla 64: 33.
Plectrohyla avia 65: 1.
Plectrohyla cotzicensis 61: 17;
65: 1.
Plectrohyla guatemalensis 61: 17;
65: 1.
Plectrohyla sagorum 65: 1.
Plectrohyla cotzicensis 61: 17.
Plectromantis andicola 100: 337.
Plectromantis wagneri 84: 163.
Plestiodon fasciatus 33: 129.
Plethodon 40: 123.
Plethodon aeneus 25: 135.
Plethodon cinereus 15: 121;
33: 129; 50: 137.
Plethodon cinereus cineurus
20:1; 59:157; 62:135; 69:93.
Plethodon cinereus dorsalis 20:1.
Plethodon cinereus
erythronotus 20: 1.
Plethodon dunnii 47: 169.
Plethodon erythronatus 29: 73.
Plethodon erythronotus 25: 135.
Plethodon glutinosus 15: 121;
20: 1; 21: 53; 21: 85; 22:
129; 23: 9; 25: 135; 43: 55;
50: 137; 61: 127.
Plethodon glutinosus
glutinosus 33: 129; 59: 157.
Plethodon hardii 54: 77.
Plethodon huldae 62: 135.
Plethodon idahoensis 54: 77.
Plethodon iecanus 61: 127.
Plethodon intermedius 47: 169.
Plethodon jordani 25: 135;
33: 129.
- Plethodon metcalfi* 25: 135;
33: 129.
Plethodon nettingi 59: 157;
62: 135.
Plethodon richmondi 62: 135.
Plethodon shermani 25: 135;
33: 129.
Plethodon vehiculus 47: 169.
Plethodon wehrlei 59: 157;
62: 135.
Plethodon welleri 62: 135.
Plethodon yonahlossee 33: 129.
Pleurodema 93: 1177.
Pleurodema brachyops B2: 199.
Pliocercus aequalis 55: 159.
Pliocercus andrewsi 55: 159.
Pliocercus bicolor 54: 119;
55: 159.
Pliocercus dimidiatus 54: 119.
Pliocercus elapoides aequalis
54: 119.
Pliocercus elapoides diastemus
54: 119; 55: 159.
Pliocercus elapoides elapoides
54: 119; 55: 159.
Pliocercus elapoides
laticollaris 54: 119.
Pliocercus elapoides schmidti
55: 159.
Pliocercus elapoides
semicinctus 55: 159.
Pliocercus euryzonus 54: 119.
Polychrus marmoratus 29: 221.
Polypedates anodon 21: 189.
Polypedates edentulus 21: 189.
Polypedates moltrechtii 51: 159.
Polypedates robustus 51: 159.
Polypedates schlegelii owstoni
51: 159.
Pomatops 23: 89.
Pomatops valvifera 23: 89.
Procinura aemula 56: 109.
Prostherapis 84: 147.
Prostherapis boulengeri 22: 89.
Prostherapis brunneus 84: 147.
Prostherapis dunnii 84: 147.
Prostherapis femoralis 22: 87.
Prostherapis festae 84: 147.
Prostherapis herminae 84: 147.
Prostherapis inguinalis 84: 147.
Prostherapis neblina 84: 147.
Prostherapis riveroi 84: 147.
Prostherapis shrevei 84: 147.
Prostherapis subpunctatus 84:147.
Prostherapis variabilis 84: 147.

- Prostherapis vertebralis* 84:147.
Prostherapis whimperi 84: 147.
Prostheraspis femoralis 22: 89.
Protestudo 96: 567.
Psammobates geometricus B8: 108.
Psammophis brevirostris 45: 83.
Psammophis sibilans 26: 145.
Pseudacris clarki 74: 157.
Pseudacris feriarum 31: 51.
Pseudacris nigrita 74: 157.
Pseudacris nigrita canadensis
 69: 169.
Pseudacris nigrita feriarum
 50: 137; 68: 155; 69: 169.
Pseudacris nigrita maculata
 69: 169.
Pseudacris nigrita nigrita
 48: 107.
Pseudacris nigrita
septentrionalis 69: 169;
 76: 159.
Pseudacris nigrita triseriata
 68: 155; 69: 169; 76: 159.
Pseudacris nigritus verrucosa
 48: 107.
Pseudacris ocularis 48: 107.
Pseudacris septentrionalis
 69: 169.
Pseudacris streckeri 74: 157.
Pseudemoia 98: 221.
Pseudemys alabamensis 99: 472.
Pseudemys concinna 21: 69; 21:
 85; 33: 129; 93: 339; B1: 1.
Pseudemys concinna hoyi 51: 173.
Pseudemys elegans 21: 47; 21: 69;
 33: 129.
Pseudemys floridana 22: 129;
 93: 339.
Pseudemys floridana concinna
 50: 137.
Pseudemys mobilensis 21: 69.
Pseudemys nelsoni 93: 339.
Pseudemys ornata 51: 173.
Pseudemys rubriventris 15:
 121; 50: 137; 57: 124.
Pseudemys rubriventris
alabamensis 51: 173.
Pseudemys scripta 22: 129;
 93: 339.
Pseudemys texana 21: 47; 21:
 69; 51: 173.
Pseudis paradoxus 93: 153.
Pseudoboa neuwiedii 50: 11.
Pseudobranchius striatus 23: 9.
Pseudoeurycea barbouri 67: 159.
Pseudoeurycea expectata 67: 159.
Pseudoeurycea gadovii 67: 159.
Pseudoeurycea goebeli 67: 159.
Pseudoeurycea leprosa 67: 159.
Pseudoeurycea rex 67: 159.
Pseudoeurycea smithi 67: 159.
Pseudoficimia 56: 109.
Pseudopareas atypicus 36: 185.
Pseudopareas spilogaster 36: 185.
Pseudopareas vagrans 36: 185.
Pseudopareas vagus 36: 185.
Pseudotriton montanus 33: 129.
Pseudotriton ruber nitidus
 33: 129.
Pseudotriton ruber ruber 33:
 129; 50: 137.
Pseudotriton ruber schencki
 33: 129.
Pseudotriton schencki 33: 129.
Pseudoxenodon bambusicola 49:117.
Pseudoxenodon karlschmidti
 49: 117.
Pseudoxenodon popei 49: 117.
Pterohyla 52: 187.
Pterohyla fodiens 55: 151.
Pterophryne 95: 423.
Pterophrynus 95: 423.
Ptychemys concinna 51: 173.
Ptychemys hoyi 51: 173.
Ptychemys mobiliensis 51: 173.
Ptychohyla adipoventris 67: 159.
Ptychohyla alleei
Ptychohyla bogerti 67: 159.
Ptychohyla schmidtorum 67: 159.
Ptychohyla uranachroa 67: 159.
Ptychozoon homalocephalum 15: 37.
Ptychozoon horsfieldii 15: 37.
Ptychozoon kuhii 15: 37.
Pyxicephalus 55: 49.
Pyxis 96: 567.

Ramphiophis multimaculatus
 35: 229.
Rana 48: 107; 93: 153.
Rana adenopleura 51: 159.
Rana aesopus 23: 9; 37: 141;
 48: 79; 69: 135.
Rana afghana 39: 53.
Rana agilis 37: 73.
Rana amurensis 37: 73.
Rana areolata 69: 135.
Rana areolata aesopus 48: 79.
Rana areolata areolata 21: 53.
Rana aurora 62: 57.
Rana aurora draytonii 43: 55.

- Rana beddomii* 55: 49; 55: 71.
Rana boulengeri 21: 189.
Rana boyllii boyllii 43: 55.
Rana boyllii muscosa 43: 55.
Rana boyllii sierrae 43: 55.
Rana brevipalmatus 54: 197.
Rana bufoniformis 91: 965.
Rana burnsi 35: 107.
Rana caerulea 99: 42.
Rana caerulea punctata 35: 221.
Rana canagica 43: 55.
Rana cancrivora 54: 197; 90: 669.
Rana cancrivora cancrivora 80: 65.
Rana cantabrigensis 43: 55; 76: 159.
Rana capito 48: 79.
Rana capito capito 69: 135.
Rana capito sevosa 69: 135.
Rana capito stertens 69: 135.
Rana catesbeiana 15: 121; 21: 85; 33: 129; 37: 141; 43: 55.
Rana catesbiana 20: 1; 21: 47; 21: 53; 21: 69; 22: 129; 23: 9.
Rana clamata 15: 121; 20: 1; 22: 129.
Rana clamitans 21: 85; 33: 129; 37: 141; 50: 137; 69: 93.
Rana clamitans melanota 76: 159.
Rana corrugata 54: 197.
Rana cyanophlyctis 54: 197.
Rana dalmatina 37: 73.
Rana diuata 90: 669.
Rana doriae 54: 197.
Rana erythraea 90: 669.
Rana feae 21: 189.
Rana galamensis 54: 197.
Rana gibbosa 23: 165.
Rana godmani 35: 221.
Rana gracilipes 51: 159.
Rana gracilis 51: 159.
Rana grayi 54: 197.
Rana grunniens 54: 197.
Rana grylio 37: 141.
Rana guentheri 51: 159.
Rana halecina 44: 11.
Rana heckscheri 37: 141.
Rana himalayana 39: 53.
Rana holsti 21: 189.
Rana japonica ornativentris 37: 73.
Rana jugans 39: 53.
Rana kandiyohi 35: 107.
Rana kuhlii 54: 197; 90: 669.
Rana leytensis 80: 65; 90: 669.
Rana liebigii 21: 189.
Rana limnocharis 51: 159; 54: 197; 90: 669.
Rana longicrus 37: 73.
Rana macrodon 54: 197; 90: 669.
Rana magna 90: 664.
Rana magna magna 80: 65.
Rana martensi 37: 73.
Rana mascareniensis 26: 145.
Rana microdisca 54: 197; 90: 669.
Rana microtympanum 54: 197.
Rana modesta 54: 197.
Rana mystacea 96: 270.
Rana namiyei 14: 189.
Rana narina 14: 189.
Rana nicobarensis 90: 669.
Rana occipitalis 54: 197.
Rana ocellata 44: 11.
Rana ornativentris 37: 73.
Rana ovalis 23: 165.
Rana palmipes 35: 221; 80: 105; 84: 163.
Rana palustris 15: 121; 20: 1; 33: 129; 50: 137; 76: 153; 76: 159.
Rana papua 27: 201.
Rana papua novaebritanniae 91: 965.
Rana pipiens 15: 121; 21: 47; 21: 69; 21: 85; 22: 115; 22: 129; 23: 9; 26: 53; 35: 107; 44: 11; 50: 43; 53: 125; 93: 1177.
Rana pipiens austriicola 21: 53.
Rana pipiens brachycephala 20: 1.
Rana pipiens pipiens 20: 1; 69: 93; 76: 159.
Rana pipiens sphenoccephala 20: 1.
Rana pretiosa 62: 57.
Rana pretiosa luteiventris 26: 53; 43: 55.
Rana pustulosa 35: 221.
Rana pyxicephalus 54: 197.
Rana rugata 81: 69.
Rana septentrionalis 35: 107; 37: 141; 69: 93; 76: 159.
Rana sevosa 69: 135.
Rana signata 90: 669.
Rana signata grandicula 80: 65.
Rana sphenoccephala 21: 53; 21: 69; 33: 129.
Rana subaspera 21: 189.

- Rana swinhoana* 51: 159.
Rana sylvatica 15: 121; 20: 137; 76: 159.
Rana sylvatica cantabrigensis 69: 93; 76: 159.
Rana sylvatica sylvatica 69: 93.
Rana temporaria 37: 73.
Rana temporaria japonica 37: 73.
Rana temporaria ornativentris 37: 73.
Rana temporalis 55: 49.
Rana tenasserimensis 55: 71.
Rana tigerina 54: 197.
Rana tsushimensis 37: 73.
Rana typhonia 96: 270.
Rana verrucosa 55: 49.
Rana vibicaria 35: 221.
Rana virescens austriaca 21: 69.
Rana virescens brachycephala 21: 69.
Rana virgatipes 22: 129; 37: 141; 64: 59.
Rana virginia 44: 11.
Rana vittigera 51: 159.
Rana woodworthi 90: 669.
Ranidella riparia 95: 423.
Ranidella signifera 95: 423.
Ranidella tasmaniensis 95: 423.
Ranidens 30: 123.
Ranodon kessleri 30: 123.
Ranodon olympicus 30: 123.
Ranodon sibiricus 30: 123.
Ranula chrysoprasina 35: 221.
Regina leberis 15: 121.
Rhabdosoma maculatum 68: 11.
Rhacophorus dimbullae 53: 105.
Rhacophorus fergusonianus 53: 105.
Rhacophorus fergusonii 53: 105.
Rhacophorus leucomystax quadrilineatus 80: 65.
Rhacophorus moltrechti 51: 159.
Rhacophorus owstoni 51: 159.
Rhacophorus robustus 51: 159.
Rhacophorus stictomerus 53: 105.
Rhadinaea 56: 109; 67: 159.
Rhadinaea aemula 72: 79.
Rhadinaea crassa 55: 185.
Rhadinaea decorata 55: 185.
Rhadinaea flavilata 23: 9.
Rhadinaea forbesi 55: 185.
Rhadinaea gaigeae 55: 185.
Rhadinaea hempsteadae 55: 159.
Rhadinaea hesperia baileyi 55: 185.
Rhadinaea hesperia hesperia 55: 185; 72: 79.
Rhadinaea hesperia hesperioides 55: 185.
Rhadinaea lachrymans 62: 165.
Rhadinaea lineata 98: 295.
Rhadinaea quinquelineata 55: 159.
Rhadinaea veraepacis 62: 165.
Rhadinea binotata 35: 219.
Rhadinea steinbachi 35: 219.
Rhadinea undulata 35: 219.
Rhamnophis jacksonii 49: 63.
Rhamphophryne 95: 557.
Rhinemys nasuta 22: 125.
Rhinemys rufipes 22: 125.
Rhineura 76: 153.
Rhineura floridana 23: 9.
Rhinocheilus lecontei 21: 69.
Rhinoderma darwinii 93: 920.
Rhinoderma signifera 98: 657.
Rhinoleptus 98: 204.
Rhinotyphlops albirostris 39: 123; 45: 173.
Rhinotyphlops lalandii 45: 173.
Riopa punctata 15: 239.
Salamandra bislineata 62: 89.
Salamandra margaritifera 15: 239.
Salamandra palustris 15: 239.
Salamandra phoca 58: 39.
Salamandra venenosa 15: 239.
Salamandrella 30: 123.
Salamandrella wasnessenskyi 61: 127.
Salvadora 56: 109.
Salvadora grahamiae 21: 47; 21: 69.
Salvadora intermedia intermedia 72: 79.
Salvadora mexicana 72: 79.
Sauresia 78: 39; 92: 1.
Sauresia sepsoides 40: 91; 41: 53.
Sauromaulus hispidus 53: 119.
Sauromaulus obesus 53: 119.
Sauromaulus townsendi 53: 119.
Scaphiopus 62: 57; 80: 105.
Scaphiopus couchii 21: 47; 21: 53; 21: 69; 21: 199; 22: 115; 23: 115; 43: 73.
Scaphiopus hammondii 21: 53; 21: 199; 23: 115; 43: 55.

- Scaphiopus hammondii*
bombifrons 21: 53.
Scaphiopus holbrookii 15:
 121; 21: 47; 21: 53; 21:
 199; 23: 9; 23: 115.
Scaphiopus hurterii 23: 115.
Sceloporus 53: 119; 56: 109;
 87: 327.
Sceloporus aeneus 47: 121.
Sceloporus asper 74: 37.
Sceloporus bulleri 49: 87.
Sceloporus carinatus 49: 87;
 67: 159.
Sceloporus cautus 84: 307.
Sceloporus chrysostictus 49: 87.
Sceloporus cochranæ 49: 87.
Sceloporus consobrinus 21: 47;
 21: 69; 21: 85.
Sceloporus couchii 17: 17;
 40: 57; 47: 121.
Sceloporus dispar 29: 227.
Sceloporus disparilis 29: 227.
Sceloporus exsul 84: 307.
Sceloporus ferrariperezi 49: 87.
Sceloporus formosus 49: 87.
Sceloporus formosus scitulus
 74: 37.
Sceloporus gadoviae 74: 37.
Sceloporus graciosus 47: 121.
Sceloporus graciosus graciosus
 34: 63; 35: 1.
Sceloporus grammicus grammicus
 74: 37.
Sceloporus graciosus 47: 121.
Sceloporus guentheri 31: 89;
 49: 87.
Sceloporus horridus horridus
 74: 37.
Sceloporus horridus oligoporus
 74: 37.
Sceloporus irazuensis 49: 87.
Sceloporus jalapæ 47:121; 49:87.
Sceloporus jarrovii 49: 87.
Sceloporus jarrovii jarrovii
 53: 125.
Sceloporus lineolateralis 49: 87.
Sceloporus lunæi 49: 87.
Sceloporus maculosus 49: 87.
Sceloporus magister
monserratensis 67: 69.
Sceloporus malachiticus 49: 87.
Sceloporus marmoratus 47: 121.
Sceloporus melanorhinus 31: 89.
Sceloporus melanorhinus
calligaster 74: 37.
Sceloporus merriami 17: 17;
 40: 57; 47: 121.
Sceloporus merriami annulatus
 50: 83.
Sceloporus merriami merriami
 50: 83.
Sceloporus microlepidotus 29:227.
Sceloporus microlepidotis
disparilis 53: 125.
Sceloporus microlepidotus
disparillis 53: 125.
Sceloporus mucronatus
omiltemanus 74: 37.
Sceloporus ochoterenæ 49: 87.
Sceloporus ochoterenai 74: 37.
Sceloporus ornatus 29: 227.
Sceloporus poinsettii 53: 125.
Sceloporus pyrocephalus 74: 37.
Sceloporus salvini 49: 87.
Sceloporus scalaris 17: 17;
 47: 121.
Sceloporus schmidti 49: 87.
Sceloporus serrifer 31:89; 49:87.
Sceloporus siniferus 49: 87.
Sceloporus siniferus siniferus
 74: 37.
Sceloporus smaragdinus 49: 87.
Sceloporus spinosus 21: 47;
 21: 69; 21: 165.
Sceloporus spinosus apicalis
 64: 101.
Sceloporus spinosus
caeruleopunctatus 64: 101;
 74: 37.
Sceloporus spinosus floridanus
 31: 89.
Sceloporus spinosus spinosus
 44: 73; 64: 101; 74: 37.
Sceloporus squamosus 49: 87;
 67: 159.
Sceloporus stejneri 74: 37.
Sceloporus taeniocnemis 49: 87.
Sceloporus torquatus 29: 227;
 31: 89; 49: 87.
Sceloporus torquatus
cyanogenys 44: 129.
Sceloporus torquatus
omiltemanus 44: 129.
Sceloporus torquatus poinsetii
 44: 129.
Sceloporus torquatus poinsetti
 44: 129.
Sceloporus torquatus torquatus
 44: 129.

- Sceloporus undulatus* 15: 121;
 20: 1; 31: 89; 33: 129; 50:
 137; 84: 307.
Sceloporus undulatus virgatus
 53: 125.
Sceloporus variabilis 17: 17;
 40: 57; 49: 87.
Sceloporus variabilis
marmoratus 47: 121.
Sceloporus variabilis
variabilis 47: 121.
Sceloporus virgatus 84: 397.
Sceloporus viviparus 21: 165;
 49: 87.
Sceloporus woodi 31: 89.
Scincus viridipunctus 24: 15.
Scolecophis atrocinctus 56: 109.
Scolecophis michoacanensis
 56: 109.
Scolecosaurus 78: 141.
Scotobleps 55: 49.
Seminatrix pygaea 23: 9.
Sepsina weberi 35: 229.
Shinisaurus crocodilurus 86: 105.
Siaphos aequalis 54: 193.
Siaphos meleagris helleri 45:113.
Sibon frenatum 52: 187.
Sibon nebulatus 72: 79.
Sibynomorphus 36: 185.
Sibynomorphus annulata 34: 157.
Sibynomorphus argus 34: 157.
Sibynomorphus articulata 34: 157.
Sibynomorphus bicolor 34: 157.
Sibynomorphus pictiventris
 34: 157.
Sibynomorphus ruthveni 34: 157.
Sibynophis annulatus annulatus
 49: 43.
Sibynophis annulatus
hondurensis 49: 43.
Siluboura nigra 55: 49.
Silybura 55: 49.
Simotes brevicauda 82: 763.
Simotes cyclurus 82: 763.
Sinohadrianus sichuanensis
 96: 567.
Siren lacertina 15: 121; 21: 47;
 21: 53; 22: 129; 23: 9; B1: 1.
Siren lacertura 20: 1.
Sistrurus catenatus 20: 1.
Sistrurus catenatus consors
 21: 47.
Sistrurus miliarius 21: 47;
 21: 69; 23: 9; 33: 129.
Smilisca baudinii 21: 47; 21:
 53; 74: 195; 93: 1177.
Smilisca phaeota 93: 1177.
Smilisca puma 93: 1177.
Smilisca sila 93: 1177.
Sminthillus 98: 774.
Sonora episcopa episcopa 56: 109.
Sonora episcopa taylori 56: 109.
Sonora erythura 56: 109.
Sonora michoacanensis
michoacanensis 56: 109.
Sonora michoacanensis
mutabilis 56: 109.
Sonora miniata linearis 56: 109.
Sonora miniata miniata 56: 109.
Sonora mosaueri 56: 109.
Sonora occipitalis 56: 109.
Sonora semiannulata blanchardi
 56: 109.
Sonora semiannulata gloydi
 56: 109.
Sonora semiannulata isozona
 56: 109.
Sonora semiannulata linearis
 56: 109.
Sonora semiannulata
semiannulata 56: 109.
Sooglossus 51: 191.
Spalerosophis diadema
cliffordi 83: 477.
Spelerpes 22: 129.
Spelerpes bilineatus 25: 135.
Spelerpes bislineatus 15: 121;
 20: 1.
Spelerpes danielsi 25: 135.
Spelerpes guttolineatus 15:
 121; 23: 9; 25: 135; B1: 1.
Spelerpes lignicolor 76: 289.
Spelerpes longicaudus 15: 121;
 20: 1; B1: 1.
Spelerpes maculicaudus 20: 1.
Spelerpes montanus 30: 87.
Spelerpes multiplicatus 21: 85.
Spelerpes porphyriticus 17: 102.
Spelerpes ruber 30: 87.
Spelerpes ruber ruber 15:
 121; 25: 135.
Spelerpes ruber schencki 25:135.
Spelerpes vermicularis 65: 1.
Sphaerodactylus alayoi 87: 337.
Sphaerodactylus alopex 29: 215.
Sphaerodactylus altavelensis
 90: 243; 95: 81.
Sphaerodactylus argus 71: 27.

- Sphaerodactylus argus*
continentalis 49: 43.
Sphaerodactylus armasi 87: 337.
Sphaerodactylus armstrongi
 88: 367; 90: 243; 95: 81.
Sphaerodactylus asper 28: 71.
Sphaerodactylus brevirostratus
 86: 35.
Sphaerodactylus bromeliarum
 95: 392.
Sphaerodactylus calicara 95:
 392.
Sphaerodactylus cinereus 41:
 53; 71: 27; 87: 337; 95: 81.
Sphaerodactylus copei 41: 53;
 90: 243; 95: 81.
Sphaerodactylus copei enochrus
 88: 367.
Sphaerodactylus copei
picturatus 88: 367.
Sphaerodactylus copei websteri
 88: 367.
Sphaerodactylus corticolus
 29: 215.
Sphaerodactylus cryphius 90:
 243; 95: 81.
Sphaerodactylus darlingtoni
 95: 81.
Sphaerodactylus decoratus 29:
 215; 87: 337.
Sphaerodactylus decoratus
decoratus 71: 27.
Sphaerodactylus decoratus
drapetiscus 71: 27.
Sphaerodactylus decoratus
gibbus 71: 27.
Sphaerodactylus decoratus
torrei 71: 27.
Sphaerodactylus difficilis 49:
 43; 86: 35; 90: 985.
Sphaerodactylus difficilis
randi 88: 367; 90: 243.
Sphaerodactylus dunni 49: 43.
Sphaerodactylus elegans 28:
 71; 30: 163; 95: 81.
Sphaerodactylus elegantulus
 30: 163; 51: 147.
Sphaerodactylus fantasticus
 28: 71; 51: 147.
Sphaerodactylus festus 28: 71.
Sphaerodactylus flavicaudis
 71: 27.
Sphaerodactylus gibbus 71: 27.
Sphaerodactylus glaucus 49: 43.
Sphaerodactylus grandisquamis
 30: 97.
Sphaerodactylus intermedius
 87: 337.
Sphaerodactylus leucaster 86:
 35; 90: 243.
Sphaerodactylus levinsi 90: 985.
Sphaerodactylus lineolatus
 16: 3; 49: 43.
Sphaerodactylus macrolepis 28:
 71; 30: 97; 45: 183; 90: 985.
Sphaerodactylus mariguanae
 90: 985.
Sphaerodactylus melanospilus
 28: 71.
Sphaerodactylus micropithecus
 90: 985.
Sphaerodactylus monensis 90: 985.
Sphaerodactylus nigropunctatus
 71: 27.
Sphaerodactylus nigropunctatus
strategus 87: 337.
Sphaerodactylus notatus 29:
 215; 71: 27.
Sphaerodactylus notatus
atactus 87: 337; 95: 392.
Sphaerodactylus notatus randi
 90: 243.
Sphaerodactylus nycteropus 95: 81.
Sphaerodactylus oliveri 71: 27.
Sphaerodactylus omoglaux 95: 81.
Sphaerodactylus pacificus 16: 3.
Sphaerodactylus picturatus 28: 71.
Sphaerodactylus randi
methorius 90: 243.
Sphaerodactylus randi randi
 90: 243.
Sphaerodactylus randi strahmi
 90: 243.
Sphaerodactylus rhabdotus 86:
 35; 90: 243.
Sphaerodactylus richardsonii
 28: 71; 45: 183.
Sphaerodactylus ruibali 87: 337.
Sphaerodactylus sabanus 51: 147.
Sphaerodactylus samanensis
 45: 183.
Sphaerodactylus savagei 90: 985.
Sphaerodactylus scaber 71: 27;
 79: 255.
Sphaerodactylus shrevei 86: 35.
Sphaerodactylus sputator 28: 71;
 30: 163; 51: 147.
Sphaerodactylus stejneri
 71: 27; 87: 337.

- Sphaerodactylus streptophorus* 90: 243; 95: 81.
Sphaerodactylus thompsoni 88: 367; 90: 243.
Sphaerodactylus torrei 28: 71; 30: 163; 71: 27; 87: 337.
Sphaerodactylus vincenti 51: 147.
Sphenomorphus arborens 80: 69.
Sphenomorphus decipiens 80: 69.
Sphenomorphus diwata 80: 69.
Sphenomorphus helenae 40: 179.
Sphenomorphus jagori 80: 69.
Sphenomorphus kinabaluensis 80: 69.
Sphenomorphus lednickyi 80: 69.
Sphenomorphus mindanensis 80: 69.
Sphenomorphus multisquamatus 80: 69.
Sphenomorphus quoyi 27: 201.
Sphenomorphus sabanus 80: 69.
Sphenomorphus steerei 80: 69.
Sphenomorphus variegatum 80: 69.
Sphenophis 31: 99.
Sphenophryne mehelyi 91: 965.
Sphenosoma bowringi 40: 179.
Sphenosoma hughi 40: 179.
Spilotes corais couperi 23: 9.
Spilotes pullatus 29: 221.
Spilotes pullatus dichrous 36: 185.
Staurios 55: 49.
Staurios afghana 55: 71.
Staurios larutensis 55: 71.
Staurios natator 55: 71.
Staurois parvus 95: 621.
Stenocephalus 23: 165.
Stenodactylus 57: 1.
Stenorhina 56: 109.
Stenorrhina freminivilli freminivilli 72: 79.
Stenorrhina freminivilli lactea 72: 79.
Stereochilus marginatus 22: 129.
Sternoteirus 15: 235.
Sternothaerus boscii 15: 235.
Sternothaerus leachianus 15: 235.
Sternothaerus odorata 15: 235.
Sternothaerus odoratus 15: 235.
Sternothaerus trifasciatus 15: 235.
Sternotherus odorata 15: 235.
Sternotherus odoratus 15: 235; 50: 137; 56: 168.
Sternotherus pennsylvanica 15: 235.
Stilosoma 76: 153.
Stilosoma extenuatum 23: 9.
Storeria dekayi 15: 121; 20: 1; 21: 47; 21: 69; 21: 85; 23: 9; 50: 137.
Storeria occipitomaculata 15: 121; 76: 297.
Storeria occipitomaculata occipitomaculata 76: 159.
Stupendemys geographicus 99: 472.
Stylemys 96: 567.
Syrrhophus 64: 33.
Syrrhophus campi 28: 131.
Syrrhophus juninensis 98: 774.
Syrrhophus latodactylus 54: 87.
Syrrhophus leprus 28: 131; 53: 95.
Syrrhophus lutosus 34: 157; 94: 413.
Syrrhophus montium 98: 774.
Syrrhophus omiltemanus 54: 87.
Syrrhophus pipilans 53: 95.
Syrrhophus ridens 94: 413.
Syrrhophus marnochii 21: 53; 28: 131.
Syrrhophus molinoi 94: 413.
Syrrhophus mystaceus 35: 111.
Systema 23: 165.
Tachymenis elongata 35: 219.
Tachymenis surinamensis 35: 219.
Tantilla 49: 51.
Tantilla atriceps 53: 55.
Tantilla bocourti 29: 93; 53: 55; 72: 78.
Tantilla boulengeri 56: 109.
Tantilla breve 53: 55.
Tantilla brevissima 53: 55.
Tantilla calamarina 53: 55.
Tantilla canula 53: 55.
Tantilla coronadoi 72: 79.
Tantilla coronata 22: 129; 23: 9; 29: 93; 53: 55; 56: 109.
Tantilla coronata coronata 76: 305.
Tantilla deviatix 29: 93.
Tantilla eiseni 53: 55.
Tantilla flavilineata 64: 97.
Tantilla gracilis 21: 69; 21: 85; 53: 55; 56: 109.
Tantilla jani 64: 97.
Tantilla lintoni 53: 55.
Tantilla martindelcampoi 53: 55.
Tantilla melanocephala 36: 185.

- Tantilla nigriceps nigriceps* 53: 55.
Tantilla rubra 56: 109.
Tantilla rubrum 53: 55.
Tantilla schistosa 53: 55.
Tantilla semicineta 50: 11; 56: 109.
Tantilla striata 64: 97.
Tantilla taeniata 64: 97.
Tantilla triseriata 64: 97.
Tantilla wilcoxi 29: 93; 53: 55; 53: 125.
Tarbophis 45: 83.
Tarbophis beetzii 35: 229.
Tarbophis semiannulatus 35: 229.
Tarentola americana americana 81: 123.
Tarentola americana warreni 81: 123.
Tarentola annularis 26: 145.
Tarentola cubana 29:215; 81:123.
Teius 42: 153.
Teius teyou teyou 44: 73.
Telmatobius 37: 65; 62: 57.
Telmatobius arequipensis 98: 774.
Telmatobius barrioi 98: 774.
Telmatobius cirrhacelis 98: 774.
Telmatobius duseni 96: 548.
Telmatobius jelskii 98: 774.
Telmatobius juninensis 98: 774.
Telmatobius lintoni 98: 774.
Telmatobius marmoratus 98: 774.
Telmatobius niger 98: 774.
Telmatobius peruvianus 98: 774.
Telmatobius schreiteri 98: 774.
Telmatobius simonsi 98: 774.
Telmatobufo australis 95: 688.
Telmatobufo bullocki 95: 688.
Telmatobufo venustus 93: 920; 95: 688.
Teratohyla 64: 33.
Teratohyla spinosa 64: 33.
Terrapene amboinensis 15: 235.
Terrapene bauri 23: 9.
Terrapene bosicii 15: 235.
Terrapene carolina 15: 121; 22: 129; 23: 9; 23: 124; 33: 129; 50: 137.
Terrapene carolina carolina 39: 19.
Terrapene carolina triunguis 21: 85; 23: 115.
Terrapene clausa 15: 235.
Terrapene goldmani 46: 119.
Terrapene major 23: 9.
Terrapene nigricans 15: 235.
Terrapene odorata 15: 235.
Terrapene ornata 15: 235; 21: 47; 21: 69; 44: 11.
Terrapene pennsylvanica 15: 235.
Terrapene sinica 96: 567.
Terrapene tricarinata 15: 235.
Terrapene triunguis 23: 9.
Terraphene clausa 15: 235.
Testudo antiqua 96: 567.
Testudo chienfutungensis 96: 567.
Testudo concentrica B1: 1.
Testudo demissa 96: 567.
Testudo ferox B1: 1.
Testudo galeata 22: 125.
Testudo graeca 96: 567; B8: 108.
Testudo hermanni 96: 567; B8:108.
Testudo hipporionum 96: 567.
Testudo hanenensis 96: 567.
Testudo horsfieldii 96: 567.
Testudo hypercostata 96: 567.
Testudo insolita 96: 567.
Testudo insolitus 96: 567.
Testudo kaiseni 96: 567.
Testudo kalganensis 96: 567.
Testudo kegenica 96: 567.
Testudo longicollis 22: 125.
Testudo lunanensis 96: 567.
Testudo nanus 96: 567.
Testudo picta B1: 1.
Testudo polyphemus 23: 9.
Testudo scorpioides 15: 235.
Testudo shansiensis 96: 567.
Testudo spengleri 15: 235.
Testudo sphaerica 96: 567.
Testudo tabulata 51: 147.
Testudo tungia 96: 567.
Testudo turgida 96: 567.
Testudo tunhuanensis 96: 567.
Testudo ulanensis 96: 567.
Testudo yunnanensis 96: 567.
Testudo yushensis 96: 567.
Tetraprion jordani 55: 151.
Thalassochelys caretta 23: 9.
Thalerophis diplotropis 72: 79.
Thamnophis angustirostris 53:125.
Thamnophis arabdotus 53: 55.
Thamnophis brachystoma 58: 147; 69: 71.
Thamnophis butleri 45: 195; 58: 147; 69: 71.
Thamnophis chrysocephalus 72: 79.
Thamnophis cyrtopsis cyclides 72: 79.
Thamnophis elegans 45: 195.

- Thamnophis eques 53: 55; 53: 125.
 Thamnophis lineatus 45: 195.
 Thamnophis marcianus 45: 195.
 Thamnophis multimaculata 53: 125.
 Thamnophis ordinoides
 ordinoides 45: 195.
 Thamnophis ordinoides vagrans
 45: 195.
 Thamnophis proximus 45: 195.
 Thamnophis radix 69: 71.
 Thamnophis radix butleri 45:
 195; 58: 147.
 Thamnophis radix haydeni 76: 159.
 Thamnophis radix radix 45: 195.
 Thamnophis **rozellae** 53: 55.
 Thamnophis sauritus 15: 121;
 33: 129.
 Thamnophis scalaris godmani
 72: 79.
 Thamnophis sirtalis 15: 121;
 33: 129; 45: 195.
 Thamnophis sirtalis dorsalis
 15: 121.
 Thamnophis sirtalis parietalis
 44: 11; 76: 159.
 Thamnophis sirtalis sirtalis
 50: 137; 69: 93.
 Thamnophis sumichrasti 53: 55.
 Thecodactylus rapicaudus 28:
 71; 51: 147.
 Therapene 15: 235.
 Thoropa 96: 548; 98: 774.
 Thoropa **lutzi** 51: 41.
 Thoropa miliaris 51: 41.
 Thrasops jacksonii jacksonii
 49: 63.
 Thrasops jacksonii **schmidti**
 49: 63.
 Thrasops rothschildi 49: 63.
 Toluca 56: 109.
 Toluca conica 72: 79.
 Tomodactylus 98: 774.
 Trachemys scripta 93: 339.
 Trichobatrachus 55: 49.
 Trionocephalus intermedius
 85: 557.
 Trimeresurus chaseni 51: 43.
 Trimeresurus **garciai** 53: 55.
 Trimeresurus gramineus 51: 43.
 Trimeresurus lansbergii
 annectens 49: 43.
 Trimeresurus lansbergii
 lansbergii 49: 43.
 Trimeresurus lansbergii
 ophryomegas 49: 43.
 Trimeresurus nasutus 49: 43.
 Trimeresurus nummifer nummifer
 52: 165.
 Trimeresurus nummifer **picadoi**
 52: 165.
 Trimeresurus ophyromegas 49: 43.
 Trimeresurus sumatranus
 malcolmi 51: 43.
 Trimeresurus sumatranus
 sumatranus 51: 43.
 Trimeresurus undulatus 53: 55.
 Trimerorhinus tritaeniatus
 multisquamis 45: 83.
 Trimerorhinus tritaeniatus
 tritaeniatus 45: 83.
 Trimerorhinus variabilis 45: 83.
 Trimetopon barbouri 62: 165;
 67: 159.
 Trimetopon gracile 62:165; 67:159
 Trimetopon **hannsteini** 62:
 165; 67: 159.
 Trimetopon **pilonaorum** 67: 159.
 Trimetopon pliolepis 62: 165.
 Trimetopon posadasi 62: 165;
 67: 159.
 Trimetopon simile 62:165; 67:159.
 Trimetopon sleveni 62:165; 67:159
 Trimetopon veraepacis 62: 165;
 67: 159.
 Trimetopon viquezi 62:165; 67:159
 Trimorphodon 56: 109.
 Trimorphodon biscutatus
 semirutus 72: 79.
 Trimorphodon latifascia 72: 79.
 Trionyx ferox 23: 9; 23: 115.
 Trionyx muticus 20: 1.
 Trionyx spiniferus 20: 1.
 Triprion 52: 187.
 Triprion petasatus 55: 151.
 Triton ensicauda 51: 159.
 Triton sibiricus 30: 123.
 Triton subcristatus 51: 159.
 Triturus ensicaudus 51: 159.
 Triturus granulatus 62: 57.
 Triturus rivularis 62: 57.
 Triturus torosus 43: 55.
 Triturus viridescens 33: 129.
 Triturus viridescens
 viridescens 50: 137; 59:
 165; 59: 166.
 Tropidoclonion lineatum
 annectens 76: 297.
 Tropidoclonion lineatum
 lineatum 76: 297.

- Tropidoclonion lineatum texanum* 76: 297.
Tropidoclonium lineatum 21: 69; 45: 195; 76: 297.
Tropidoclonium lineatum iowae 76: 297.
Tropidodipsas 36: 185.
Tropidodipsas fasciata 55: 159.
Tropidodipsas fischeri 55: 159.
Tropidodipsas guerreroensis 72: 79.
Tropidodipsas kidderi 55: 159.
Tropidodipsas sartorii 55: 159.
Tropidogaster spilogaster 36: 185.
Tropidonotus clarkii 21: 47.
Tropidonotus erythrogaster 21: 69.
Tropidonotus fasciatus 21: 69.
Tropidonotus grahami 21: 69.
Tropidonotus johannis 50: 125.
Tropidonotus leberis 21: 85.
Tropidonotus modestus 50: 125.
Tropidonotus rhombifer 21: 47; 21: 69.
Tropidonotus sipedon fasciatus 21: 47; 21: 69; 21: 85.
Tropidonotus sipedon transversus 21: 47; 21: 69; 21: 85.
Tropidonotus transversus 21: 69.
Tropidophis canus 88: 77.
Tropidophis feicki 71: 37; 88: 77.
Tropidophis haetiana 41: 53.
Tropidophis haetianus 88: 77.
Tropidophis maculatus pilsbryi 88: 77.
Tropidophis melanurus 88: 77.
Tropidophis nigriventris hardyi 88: 77.
Tropidophis nigriventris nigriventris 88: 77.
Tropidophis pardalis 88: 77.
Tropidophis paucisquamis 59: 17.
Tropidophis pilsbryi galacelidus 88: 77.
Tropidophis pilsbryi pilsbryi 88: 77.
Tropidophis semicinctus 88: 77.
Tropidophis taczanowskyi 59: 17.
Tropidophis wrighti 88: 77.
Tropidophorus beccarii 58: 47.
Tropidophorus berdmorei 58: 47.
Tropidophorus darlingtoni 58: 47.
Tropidophorus laotus 58: 47.
Tropidophorus mocquardi 58: 47.
Tropiocolotes helenae 83: 477.
Tropiocolotes steudneri 83: 477.
Tropiocolotes tripolitanus 83: 477.
Tupinambis 42: 153.
Tupinambis nigropunctatus 29: 221.
Typhlomolge rathbuni 21: 53.
Typhlonectes natans 37: 125.
Typhlops 36: 185.
Typhlops biminiensis 78: 121; 79: 255.
Typhlops braminus 72: 79.
Typhlops broomi 58: 111.
Typhlops capitulata 79: 255.
Typhlops caymanensis 78: 121; 79: 255.
Typhlops costaricensis 78: 121.
Typhlops diversus 58: 111.
Typhlops dominicana 79: 255.
Typhlops emunctus 45: 173.
Typhlops flavoterminalis 39: 123.
Typhlops frontalis 45: 173.
Typhlops granti 79: 255.
Typhlops haitiensis 79: 255.
Typhlops hectus 87: 11.
Typhlops jamaicensis 79: 255.
Typhlops lehneri 78: 121.
Typhlops lumbricalis 28: 71; 41: 53; 78: 121; 79: 255; 87: 11.
Typhlops monastus 79: 255.
Typhlops monastus geotomus 79: 255.
Typhlops monastus monastus 79: 255.
Typhlops monensis 79: 255; 90: 985.
Typhlops pusilla 79: 255.
Typhlops pusillus 87: 11.
Typhlops richardi platycephalus 79: 255.
Typhlops richardi richardi 79: 255.
Typhlops reticulatus 78: 121.
Typhlops rostellata 79: 255.
Typhlops stadelmani 49: 43.
Typhlops sulcata 79: 255.
Typhlops syntherus 79: 255; 87: 11; 88: 367.
Typhlops tenuis 49: 43.
Typhlops tovelli 58: 111.
Typhlops trinitatus 78: 121.
Typhlops unilineatus 78: 121.
Typhlotriton 50: 141.

- Uma 73: 67.
 Uperolia marmorata 27: 201.
 Ungalia brasiliensis 59: 17.
 Ungalia paucisquamis 59: 17.
 Ungalia taczanowskyi 59: 17.
 Ungaliophis danieli 59: 17.
 Uromacer catesbyi 41: 53; 44: 89.
 Uromacer dorsalis 41: 53; 44: 89.
 Uromacer frenatus 41: 53; 44: 89.
 Uromacer oxyrhynchus 41: 53;
 44: 89.
 Uromacer scandax 44: 89.
 Uromacer **wetmorei** 44: 89.
 Urosaurus bicarinatus
 anonymorphus 74: 37.
 Urosaurus bicarinatus
 bicarinatus 74: 37.
 Urotheca bicinctus 82: 847.
 Urotheca elapoides 54: 119.
 Urotheca euryzonus 54: 119.
 Uta 47: 121; 53: 119.
 Uta ornata **chiricahuae** 54: 165.
 Uta ornata linearis 54: 165.
 Uta stansburiana 35: 1.
 Uta stansburiana **nevadensis**
 26: 27.

Vanzolinus 87: 81.
 Vanzolinus discodactylus 87:
 81; 87: 327.
 Varanus griseus 83: 477.
 Verticaria caerulea 42: 153.
 Verticaria ceralbensis 42: 153.
 Verticaria hyperythra
 hyperythra 42: 153.
 Verticaria picta 42: 153.
 Verticaria sericea 42: 153.
 Virginia elegans 20: 1; 23: 9.
 Virginia valeriae 15: 121; 23: 9.
 Virginia valeriae valeriae
 50: 137.

Wetmorena 40: 91; 92: 1.
 Wetmorena **haetiana** 40: 91.
 Wetmorena haetiana haetiana
 78: 39.
 Wetmorena haetiana **mulica** 78: 39.
 Wetmorena haetiana **surda** 78: 39.
 Woodworthia 25: 187.

 Xenodon colubrinus 53: 55.
 Xenodon gigas 82: 947.
 Xenodon **mexicanus** 53: 55.
 Xenodon rabdocephalus 59: 17.
 Xenopus laevis **bunyoniensis**
 45: 113.
 Xenopus laevis laevis 45: 113.
 Xenopus laevis victorianus
 45: 113.
 Xenosaurus grandis 54: 47; 54: 217.
 Xenosaurus grandis arboreus
 86: 105.
 Xenosaurus newmanorum 86: 105.
 Xenosaurus platyceps 86: 105.
 Xenosaurus **rackhami** 54: 47;
 54: 217.
 Xerobates berlandieri 21: 47.
 Xestosaurus bogotensis 42: 99.

 Zachaenys pervulus 37: 65.
 Zamenis constrictor 20: 1; 21: 85.
 Zamenis constrictor
 constrictor 21: 69.
 Zamenis constrictor
 flaviventris 21: 65; 23: 9.
 Zamenis flagellum 21: 69, 115.
 Zamenis flagellum flagellum 21:
 47.
 Zamenis florulentus 26: 145.

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A BIBLIOGRAPHY OF CHILDREN'S LITERATURE AND
EDUCATIONAL RESOURCES ON AMPHIBIANS AND REPTILES



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Division of Natural Sciences
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INTRODUCTION

This bibliography of children's literature and educational resources is for both educators and scientists. Often in an attempt to stimulate and facilitate learning, we encourage students to read. This bibliography offers young readers, their parents, and teachers a selection of excellent books from which to choose. For the scientists called into schools or nature centers to share her/his knowledge, this compilation provides a list of resources to help bridge the gap between the presentation and the classroom program.

Section I is a list of 135 children's books (primarily nonfiction). Entries are alphabetical by author and numbered sequentially. Those entries followed by an asterisk* are judged particularly useful and accurate. The indices following the bibliography provide cross-reference by grade level and taxonomic group and direct the user to specific books by their numeric code. Section II provides an additional list of resources to assist teachers in their classroom activities.

This bibliography is not all inclusive. I offer it as a starting place to assist educators and scientists in their interactions with young readers and novice naturalists. I hope that it will stimulate further interest in the study and conservation of amphibians and reptiles.

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Grade Level Index

All books listed in the bibliography were reviewed by the author. Books were assigned to grade levels based on content and reading level. Some books assigned to each grade level will likely be beyond the capabilities of some young readers. However, they are appropriate selections for teachers to read to/with their classes.

Grade 1: 2, 3, 4, 5, 6, 7, 8, 9, 10, 19, 21, 24, 30, 36, 46, 47, 48, 49, 50, 53, 66, 67, 69, 75, 76, 79, 82, 84, 98, 110, 116, 117, 121, 122, 123, 125, 126, 127.

Grade 2: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 16, 18, 19, 21, 24, 27, 30, 36, 46, 47, 48, 49, 50, 53, 66, 67, 69, 75, 76, 79, 82, 84, 96, 97, 98, 110, 112, 114, 115, 116, 117, 121, 122, 123, 125, 126, 127, 130.

Grade 3: 1, 3, 4, 5, 6, 7, 8, 9, 16, 18, 20, 21, 24, 27, 36, 42, 45, 46, 47, 48, 52, 55, 56, 58, 61, 68, 75, 79, 81, 84, 85, 86, 87, 88, 90, 91, 92, 93, 94, 96, 97, 98, 99, 110, 111, 112, 113, 114, 115, 117, 119, 121, 122, 124, 126, 127, 130, 133, 134.

Grade 4: 1, 3, 4, 5, 6, 7, 8, 9, 11, 12, 18, 20, 27, 29, 31, 32, 34, 42, 45, 52, 54, 55, 56, 58, 61, 68, 73, 74, 80, 81, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 96, 97, 99, 104, 105, 111, 112, 113, 114, 119, 121, 122, 124, 126, 127, 128, 130, 132, 133, 134, 135.

Grade 5: 1, 11, 12, 14, 16, 20, 22, 23, 25, 26, 29, 31, 32, 34, 35, 37, 38, 39, 40, 41, 42, 43, 44, 45, 52, 54, 56, 57, 58, 59, 61, 62, 68, 71, 73, 74, 77, 80, 81, 85, 86, 87, 88, 89, 92, 93, 94, 99, 100, 102, 104, 105, 111, 112, 113, 119, 121, 122, 124, 126, 128, 132, 133, 134, 135.

Grade 6: 11, 12, 14, 15, 17, 22, 23, 25, 26, 28, 29, 31, 32, 33, 34, 35, 37, 38, 39, 40, 41, 42, 43, 44, 45, 52, 54, 57, 59, 60, 62, 63, 64, 68, 70, 71, 73, 74, 77, 80, 81, 83, 87, 88, 89, 100, 102, 104, 105, 108, 109, 113, 118, 119, 120, 121, 122, 124, 126, 128, 129, 132, 133, 134, 135.

Grade 7: 7, 11, 14, 15, 17, 22, 23, 25, 26, 28, 29, 31, 33, 34, 35, 38, 39, 40, 41, 43, 44, 51, 54, 57, 59, 60, 62, 63, 64, 65, 68, 70, 71, 74, 77, 78, 80, 83, 88, 100, 102, 104, 105, 106, 108, 109, 118, 119, 120, 121, 122, 126, 129, 131, 132, 133, 134, 135.

Grade 8: 14, 15, 17, 22, 23, 26, 28, 33, 34, 35, 37, 38, 39, 44, 51, 54, 57, 59, 60, 62, 63, 64, 65, 70, 71, 77, 78, 83, 88, 100, 102, 103, 106, 108, 109, 118, 119, 120, 121, 122, 126, 129, 131, 132, 133, 134, 135.

Taxonomic Index

Salamanders: 11, 13, 34, 39, 104, 112, 131, 135

Frogs and Toads: 2, 11, 13, 14, 16, 21, 22, 28, 30, 34, 36,
44, 46, 47, 48, 50, 56, 57, 63, 64, 66, 67,
70, 73, 76, 77, 79, 82, 84, 85, 90, 92, 93,
98, 104, 107, 112, 118, 123, 125, 128, 130,
131, 133, 135.

Crocodilians: 11, 13, 15, 24, 27, 32, 34, 43, 49, 55, 61, 65,
69, 71, 74, 81, 87, 97, 99, 100, 101, 104, 109,
111, 117, 120, 121, 134, 135.

Turtles: 1, 3, 11, 13, 24, 27, 29, 32, 34, 59, 61, 65, 69, 71,
87, 88, 94, 99, 100, 101, 102, 104, 108, 110, 111, 115,
120, 121, 127, 135.

Lizards: 3, 11, 12, 13, 18, 19, 24, 25, 26, 27, 32, 34, 44, 45,
51, 53, 60, 61, 65, 68, 69, 71, 78, 80, 87, 88, 91, 99,
100, 101, 104, 106, 111, 114, 116, 120, 121, 129, 135.

Snakes: 3, 4, 5, 6, 7, 8, 9, 10, 11, 13, 17, 20, 21, 22, 23,
26, 27, 31, 32, 33, 34, 35, 37, 38, 40, 41, 42, 52, 54,
58, 61, 62, 69, 71, 72, 75, 83, 86, 87, 88, 89, 91, 96,
99, 100, 101, 103, 104, 105, 111, 113, 119, 120, 121,
122, 124, 126, 132, 135.

Section II: Educational Resources

This section includes a listing of curriculum resources, journal articles and other items the author has found valuable when working with educators interested in amphibians and reptiles. All items have been annotated.

These materials can be obtained at many academic libraries or through interlibrary loan services.

Anonymous. 1983. Snakes! Snakes! Snakes! **Nature Naturally** 7(1):1-6.

Notes: Instructional unit for grades 4-6. Teacher and student pages provide illustrations and information about snakes in a variety of formats. References and answer key provided.

_____. 1987. Hands on Herps. **Science Activities** 24(4): 27-30.

Notes: Hands-on activity for grades 1-8. Compares general characteristics of reptiles and amphibians. Outlines procedures for using teaching stations in classroom.

Chattin, S.S. 1983. Snaking through science. **Science Teacher** 50(4):20-23.

Notes: Article encourages activities with live animals, especially snakes. Offers some suggestions for care and feeding.

Kramer, D.C. 1984. The classroom animal: garter snakes. **Science & Children** 22(2):34-35.

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_____. 1987. The classroom animal: snapping turtles. **Science & Children** 24(5):22-24.

_____. 1987. The classroom animal: newts. **Science & Children** 25(3):42-43.

Notes: These articles will be useful to classroom teachers. Life history information. Tips on feeding, housing and care needs. Safety concerns addressed where appropriate. Some suggest classroom activities.

National Wildlife Federation. 1987. Hurray for Herps!
Naturescope 3(4):1-69.

Notes: Grades K-8. Many teaching activities. Includes reproducible handouts and background information for students and teachers. Relatively inexpensive.

Nethery, L.B. 1984. Children and humane behavior: are snakes included? **Children's Environments Quarterly** 1(3):27-29.

Notes: "Food for thought" for all educators teaching about animals.

Ohanian, S. 1988. Leaps in learning. **Learning** 16(5):48-52.

Notes: Uses frog lore as the basis for enrichment in language arts, science, social studies and the arts. Useful list of reference books included.

Shaw, J.M. 1985. Ideas. **Arithmetic Teacher** 32(7):27-32.

Notes: Four work sheets for practicing math skills.

Smith, R.W. 1987. The class menagerie: using small animals to enrich your science curriculum. **Learning** 15(5):64-66.

Notes: Art, writing and science activities with live snails, lizards, frogs and toads. Nice approach to integrating subjects.

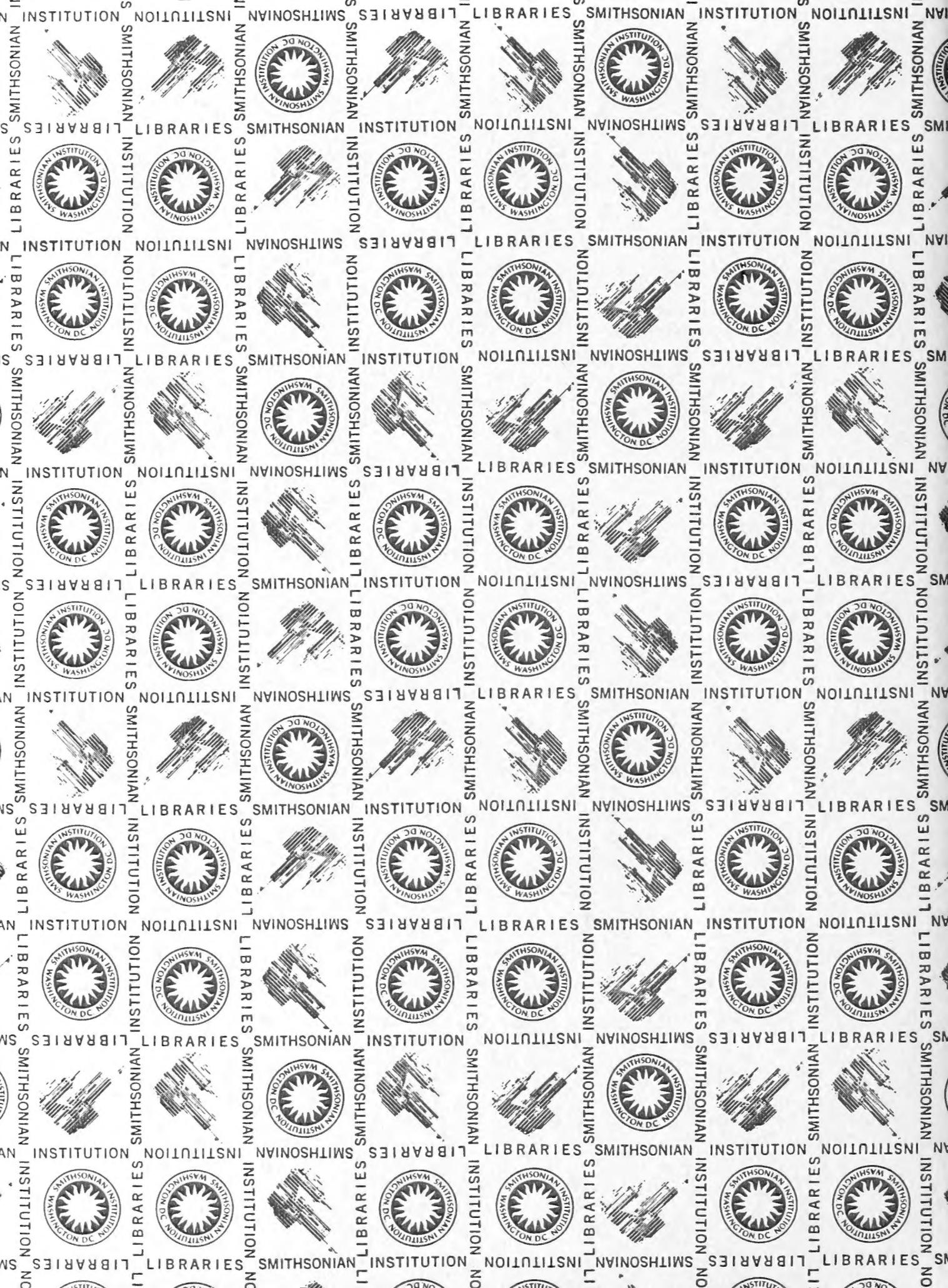
Yingling, P.S. 1983. Where the wildlife is. **Perspectives for Teachers of the Hearing Impaired** 1(3):2-3.

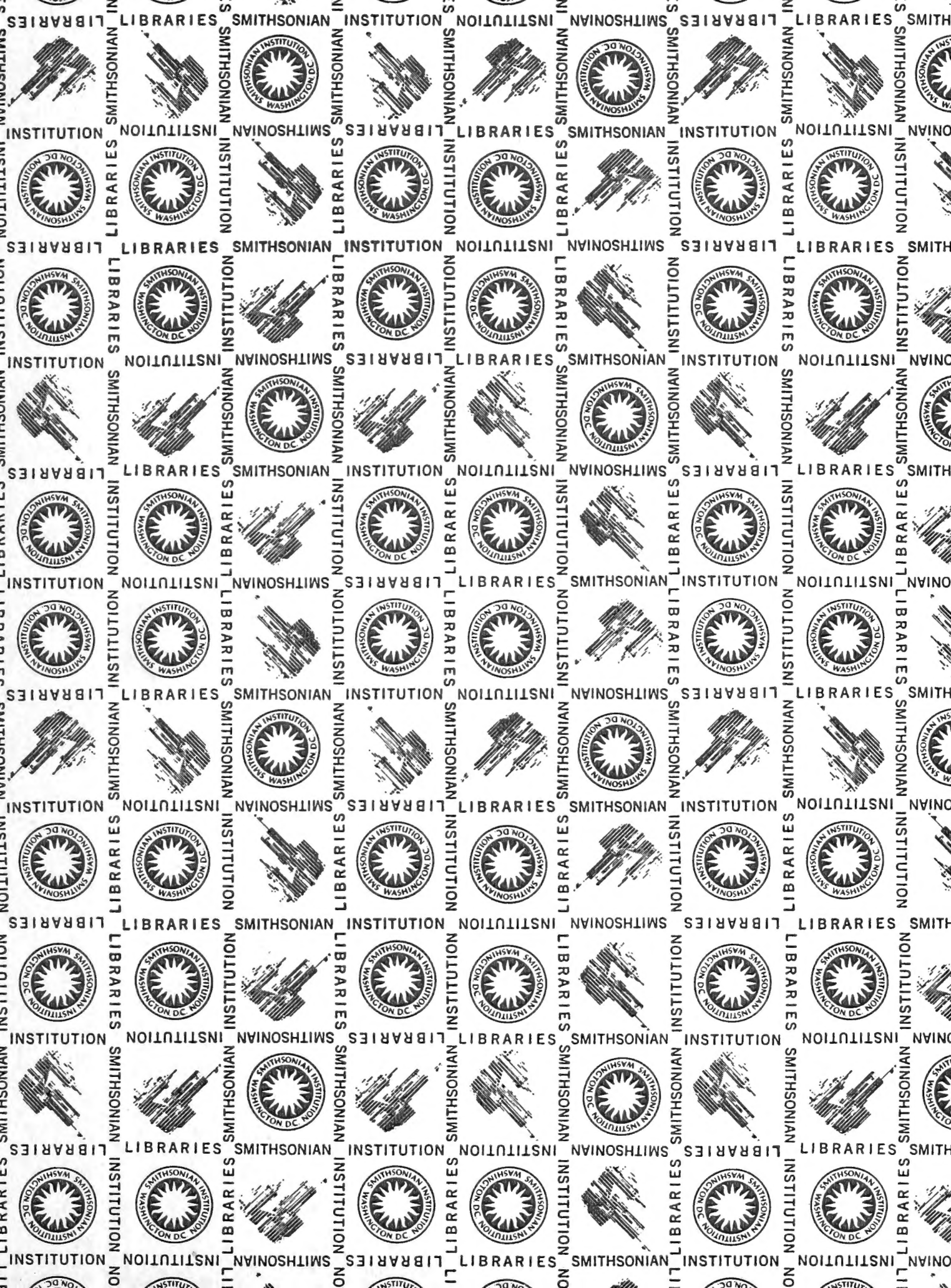
Notes: Describes techniques using live animals to stimulate understanding of life cycle concepts for hearing impaired individuals. The suggestions here are valuable to all teachers, not just specialists!

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