333

OCCURRENCE AND SIGNIFICANCE OF TRIMETHYLAMINE OXIDE IN MARINE ANIMALS

SPECIAL SCIENTIFIC REPORT–FISHERIES No. 333

UNITED STATES DEPARTMENT OF THE INTERIOR
FISH AND WILDLIFE SERVICE

Explanatory Note

The Special Scientific Report series comprises results of investigations, usually of restricted scope, intended to aid or direct management or utilization practices and as guides for administrative or legislative action. Reports in this series are issued in limited quantities for the official use of Federal, State, or other cooperating agencies, and in processed form for economy and to avoid delay in publication.

United States Department of the Interior, Fred A. Seaton, Secretary
Fish and Wildlife Service, Arnie J. Suomela, Commissioner

THE OCCURRENCE AND SIGNIFICANCE OF TRIMETHYLAMINE OXIDE IN MARINE ANIMALS

By

Herman S. Groninger
Biochemist
Bureau of Commercial Fisheries

This is contribution No. 569 from the technological laboratories of the Bureau of Commercial Fisheries, Fish and Wildlife Service, U. S. Department of the Interior.

United States Fish and Wildlife Service
Special Scientific Report--Fisheries No. 333

Washington, D. C.
December 1959

CONTENTS

TABLES

ABSTRACT

Some pertinent information on the occurrence of trimethylamine oxide in marine animals is reported, and the current ideas on the origin and function of the oxide are examined.

THE OCCURRENCE AND SIGNIFICANCE OF TRIMETHYLAMINE OXIDE IN MARINE ANIMALS

by

Herman S. Groninger
Biochemist
U. S. Fish and Wildlife Service
Seattle, Washington

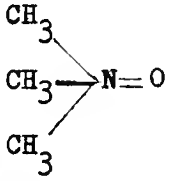
INTRODUCTION

The occurrence of trimethylamine oxide in marine animals is interesting because of its indirect effect on the quality of seafood.

The purposes of this review are (1) to compile pertinent information on the occurrence of trimethylamine oxide in marine animals and (2) to examine current ideas on the origin and function of trimethylamine oxide in these animals.

CHEMICAL NATURE OF TRIMETHYLAMINE OXIDE

Trimethylamine oxide has the following structure:



The oxide is a solid with a melting point of 257° C. It is soluble in water, acting as a weak base with a dissociation constant for the cationic acid of 4.65 (Ronald and Jakobsen 1947). The salts of the oxide have a marked buffering action in the region of pH 4.5 (Castell 1949a, Suyama 1958). In aqueous solution, the oxide is stable between pH 3 and 9 when heated at 107° for 7 hours (Ronald and Jakobsen 1947). The oxide can act as a hydrogen acceptor; it is reduced to trimethylamine in the presence of iron or hemoglobin catalyst and cysteine (Vaisey 1956) and also by an enzyme triamineoxidease (Tarr 1940), which is present in a number of different species of bacteria (Castell 1949b, Tarr 1939). Trimethylamine oxide is not toxic to animals.

OCCURRENCE

Trimethylamine oxide is found with other kinds of nonprotein nitrogen compounds in the fluids and tissues of marine and fresh-water animals. Although this report is concerned primarily with trimethylamine oxide in marine animals, a short discussion of its occurrence in fresh-water animals is included for comparison.

Occurrence in Marine Animals

Table 1 gives the content of trimethylamine oxide in a number of marine animals. The oxide has been found in a coelenterate, an echinoderm, some molluscs, all crustacea, all elasmobranchs, most teleosts, a reptile, and two mannals. Trimethylamine oxide is not found in marine plants; however, trimethylamine and dimethylamine are found in marine algae (Kapeller-Adler and Vering 1931).

Table 3 gives the mean, range, and standard deviation of trimethylamine oxide in the major groups of marine animals. The mean content of trimethylamine oxide is 134.5 mg.N/100 g. in the elasmobranchs. The mean content of the oxide in the marine teleosts, crustacea, molluscs, and fresh-water teleosts is, respectively, about 0.41, 0.32, 0.26, and <0.1 that of the elasmobranchs.

There is a wide variation in oxide content among the species that compose each group of animals. Values of 38 to 64 mg.N/100 g. have been reported for sprat, *Clupea sprattus* (Ronald and Jakobsen 1947), whereas values of 3 and 4 mg. N/100 g. have been reported for albacore, *Germo alalunga*, and bluefin tuna, *Thunnus thynnus*, respectively (Kawabata 1953). Shewan (1951) has reported that the mean content of oxide in Arctic specimens is higher than that found in North Sea specimens of the same species.

Seasonal variations in oxide content of herring have been observed by Ronald and Jakobsen (1947). They have reported that the oxide content in the tissue in winter is up to 100 percent greater than that found in summer.

The mean content of oxide in cod, *Gadus morrhua*, haddock, *Gadus aeglefinus*, and whiting, *Gadus merlangus*, is greater in large than in small fish.

Uneven distribution of the oxide in different parts of the fish has been observed (Shewan 1951, Suyama and Tokuhiro 1954). The dark lateral line of the flesh of herring and tunny contains only about half as much trimethylamine oxide as is found in the rest of the muscle.

Aside from the differences in diet among the various species, no explanation can be given for the differences in trimethylamine oxide content correlated with geographic area, season, or size of the fish. The finding of a tissue trimethylamine oxide reductase in the dark meat of albacore, *Germo alalunga*, and frigate-mackerel tissue, *Auxis tapeinosoma*, (Kawabata 1953) suggests that the variation in content between different parts of the fish is due to the enzymatic breakdown of trimethylamine oxide to trimethylamine. Other investigators have been unable to show that trimethylamine oxide is reduced in swordfish, cod, and halibut muscle (Anderson and Fellers 1949, Shewan and Jones 1957, Tarr 1939).

Table 1.--Trimethylamine oxide content of marine animals

Species	Common name	Concentration	Reference
		Mg. N/100 g. wet tissue	
Coelenterata			
Scyphozoa			
Cyanea capillata	Jellyfish	Present	Mohr 1937
Echinodermata			
Asteroidea			
Asterias vulgaris	Starfish	20	Dyer 1952
	Atlantic starfish	0	Shewan 1951
Echinoidea			
Strongylocentrotus franciscanus	Sea urchin	0	Norris and Benoit 1945a
Holothuroidea			
Cucumaria frondosa	Sea cucumber (Atlantic)	76, 86	Dyer 1952
Cucumaria miniata	Sea cucumber (Pacific)	0	Norris and Benoit 1945a
Stichopus californicus	Sea cucumber	0	Norris and Benoit 1945a
Mollusca			
Amphineura			
Katharina tunicata	Black chiton	0	Norris and Benoit 1945a
Cryptochiton stelleri	Giant chiton	0	Norris and Benoit 1945a
Gastropoda			
Anisodoris nobilis	Sea slug	0	Norris and Benoit 1945a
Thais lamellosa	Snail	0	Norris and Benoit 1945a
Littorina silchana	Snail	0	Norris and Benoit 1945a
Polinices heros	Moonshell	0	Dyer 1952
Haliotis gigontea	Ear-shell	61	Simidu et al. 1953
Turbo cornutus	Wreath-shell	20	Simidu et al. 1953

Table 1.--Trimethylamine oxide content of marine animals (continued)

Species	Common name	Concentration	Reference
		Mg.N/100 g. wet tissue	
Mollusca (cont.)			
Pelecypoda			
Mya arenaria	Clam	0	Dyer 1952
Spissula solidissima	Bar clam	0	Dyer 1952
Paphia straminea	Clam	0	Norris and Benoit 1945a
Paphia undulata	Clam	38	Simidu and Hibiki 1957
Soxidomus giganteus	Clam	0	Norris and Benoit 1945a
Macoma inquinata	Clam	0	Norris and Benoit 1945a
Mytilus edulis	Mussel (Pacific)	0	Norris and Benoit 1945a
Mytilus edulis	Mussel (Atlantic)	0	Dyer 1952
Ostrea gigas	Oyster (Pacific)	2	Simidu and Hibiki 1957
Ostrea japonica	Oyster (Pacific)	0	Norris and Benoit 1945a
Ostrea virginica	Oyster (Atlantic)	0	Dyer 1952
Ostrea cucullata	Oyster (Indian)	33	Velankar and Govindan 1958
Cardium californiense	Cockle	15, 18	Norris and Benoit 1945a
Cardium corbis	Cockle	32	Norris and Benoit 1945a
Pecten hericius	Scallop (muscle)	45-73, 63 (9)*	Norris and Benoit 1945a
	(organs)	3-6, 4 (4)	Norris and Benoit 1945a
Pecten hindsii	Scallop (muscle)	42-110, 71 (7)	Norris and Benoit 1945a
Pecten jordanii	Scallop	45	Norris and Benoit 1945a
Pecten grandis	Scallop (Atlantic)	79-84, 82 (3)	Dyer 1952
Cephalopoda			
Polypus hongkongensis	Octopus	24	Norris and Benoit 1945a
Octopus vulgaris	Octopus (basal foot muscle)	11-15, 13 (3)	Asano and Sato 1954
	(terminal foot muscle)	10-11	Asano and Sato 1954
	(abdominal muscle)	10-11	Asano and Sato 1954
	(digestive tract)	2-4	Asano and Sato 1954

---

* The figures 45-73, 63 (9) means that the range of values was from 45 to 73, that the average value was 63, and that the number of samples was 9.

Table 1.--Trimethylamine oxide content of marine animals (continued)

Species	Common name	Concentration	Reference
		Mg. N/100 g. wet tissue	
Mollusca (cont.)			
Cephalopoda			
Octopus fangsiao	Octopus (basal foot muscle)	27	Asano and Sato 1954
	(terminal foot muscle)	24	Asano and Sato 1954
	(abdominal muscle)	28	Asano and Sato 1954
	(digestive tract)	6	Asano and Sato 1954
Octopus dofleini	Octopus (basal foot muscle)	22	Asano and Sato 1954
	(terminal foot muscle)	32	Asano and Sato 1954
	(abdominal muscle)	40	Asano and Sato 1954
Loligo opalescens	Squid	150-156, *153*	Norris and Benoit 1945a
Loligo pealeii	Squid	110-122, *116*	Dyer 1952
Loligo kensaki	Squid	87,66,261, *138*	Simidu et al. 1953
Sepioteuthis lessoniana	Squid	195	Simidu et al. 1953
Ommastrephes sloanipacificus	Squid	73	Simidu et al. 1953
Arthropoda (Crustacea)			
Copepoda			
(A mixture largely of) *Corycaeus affinis* *Calanus finmarchius* *Tortanus discaudatis* *Epidabiocera amphrites*	Zooplankton	63	Norris and Benoit 1945a

Table 1.--Trimethylamine oxide content of marine animals (continued)

Species	Common name	Concentration	Reference
		Mg. N/100 g. wet tissue	
Arthropoda (Crustacea) (cont.)			
Copepoda			
(A mixture of nearly completely) *Corycaeus affinis*	Zooplankton	22	Norris and Benoit 1945a
(Mainly) decapod larvae	Zooplankton	47	Shewan 1951
Cirripedia			
Balanus cariosus	Barnacles	24	Norris and Benoit 1945a
Balanus nubila	Barnacles	59-99, 80 (4)	Norris and Benoit 1945a
Amphipoda sp.	Sand-flea	3	Norris and Benoit 1945a
Decapoda			
Pandalus danae	Shrimp	39-88, 64 (3)	Norris and Benoit 1945a
Pagurus alaskensis	Hermit crab	36	Norris and Benoit 1945a
Pagurus ochotensis	Hermit crab	66	Norris and Benoit 1945a
Pagurus setosis	Hermit crab	64	Norris and Benoit 1945a
Pagurus tenuimanus	Hermit crab	38, 42, 40, 40	Norris and Benoit 1945a
Oregonia gracilis	Spider crab	18	Norris and Benoit 1945a
Pugettia gracilis	Spider crab	13	Norris and Benoit 1945a
Cancer gracilis	Crab	31	Norris and Benoit 1945a
Cancer productus	Crab	64	Norris and Benoit 1945a
Hemigrapsus nudus	Shore crab	15	Norris and Benoit 1945a
Homarus vulgaris	Lobster (muscle)	100-110, 105 (4)	Kermack, Lees and Wood 1955
	(hepatopancreas)	17, 21	Kermack, Lees and Wood 1955
	Lobster (Europe) (tail muscle)	19	Hoppe-Seyler 1934
Fishes			
Holocephali			
Hydrolagus colliei	Ratfish (muscle)	169, 188, 178	Norris and Benoit 1945a
	(blood serum)	10, 11	Norris and Benoit 1945a

Table 1.--Trimethylamine oxide content of marine animals (continued)

Species	Common name	Concentration	Reference
		Mg. N/100 g. wet tissue	
Fishes (continued)			
Marsipobranchii			
Myxine glutinosa	Hagfish	63	Dyer 1952
Elasmobranchii			
Lamna nasus	Porbeagle	200	Dyer 1952
Squalus acanthias	Spiny dogfish (muscle)	190	Dyer 1952
	(Atlantic) (blood plasma)	104	Cohan, Krupp and Chidsay 1958
	Spiny dogfish (muscle)	189, 123, 156	Norris and Benoit 1945a
	(blood serum)	90	Norris and Benoit 1945a
	(kidney)	59	Norris and Benoit 1945a
	(liver)	25, 11, 18	Norris and Benoit 1945a
	(pancreas)	10	Norris and Benoit 1945a
	(spleen)	87	Norris and Benoit 1945a
	(stomach)	12	Norris and Benoit 1945a
Raja erinacea	Little skate	86	Dyer 1952
Raja seabrata	Prickly skate (Atlantic)	37-160, 73 (5) 51	Dyer 1952 Beatty and Gibbons 1937
Raja senta	Smooth skate	99	Beatty and Gibbons 1937
Raja batis	Skate	206	Love et al. 1959
Raja hollandi	Skate	254	Suyama and Tokuhiro 1954
Raja laevis	Barndoor skate	234	Beatty and Gibbons 1937
Mustelus manazo	--	275	Suyama and Tokuhiro 1954
Scoliodon sp.	--	72	Velankar and Govindan 1958
Chiloscyllium griscum	--	83	Velankar and Govindan 1958
Sphyrna mallius	--	107	Velankar and Govindan 1958
Trygon microps	--	49	Velankar and Govindan 1958
Trygon urnak	--	104	Velankar and Govindan 1958
Myliobatis maculata	--	29	Velankar and Govindan 1958
Rhynchobatus djeddensis	--	87	Velankar and Govindan 1958
Tygron imbricata	--	46	Velankar and Govindan 1958

Table 1.--Trimethylamine oxide content of marine animals (continued)

Species	Common name	Concentration	Reference
		Mg. N/100 g. wet tissue	
Fishes (continued)			
Teleostei			
Tarpon atlanticus	Tarpon	43	Dyer 1952
Clupea harengus	Herring (Atlantic)	67-81, *74* (7)	Dyer 1952
Clupea sprattus	Herring (whole)	38-64, *50* (20)	Ronald and Jakobsen 1947
	(entrails)	0	Ronald and Jakobsen 1947
	(milts)	31-96, *75*	Ronald and Jakobsen 1947
	(roe)	4-26, *18*	Ronald and Jakobsen 1947
Clupea pallasii	Herring (Pacific)	69	Dyer 1952
Oncorhynchus tschawytscha	King salmon	9, 10	Norris and Benoit 1945a
		25	Dyer 1952
Oncorhynchus kisutch	Silver salmon	9-12, *10* (3)	Norris and Benoit 1945a
Salmo salar	Atlantic salmon	24	Dyer 1952
Menidia notata	Atlantic silverside	28	Dyer 1952
Scomber scombrus	Atlantic mackerel	31-38, *34* (3)	Dyer 1952
		41-54, *48* (10)	Beatty and Gibbons 1937
Thunnus thynnus	Bluefin tuna	4	Dyer 1952
Germo alalunga (G)	Albacore (dark meat)	3.2	Kawabata 1953
	(white meat)	2.7	Kawabata 1953
Auxis tapeinosoma	Frigate-mackerel		
	(dark meat)	15.9	Kawabata 1953
	(white meat)	1.6	Kawabata 1953
Gadus aeglefinus	Cod (Europe)	19-51, *44* (135)	Love et al. 1959
	(Arctic)	60-140, *81* (136)	Love et al. 1959
Gadus callarias	Cod (Europe)	41-73, *62* (33)	Love et al. 1959
	(Arctic)	19-200, *103* (260)	Love et al. 1959
Gadus merlangus	Cod (Europe)	44-57, *51* (70)	Love et al. 1959
Gadus virens	Cod (Europe)	40	Love et al. 1959
	(Arctic)	56-98, *77* (13)	Love et al. 1959

Table 1.--Trimethylamine oxide content of marine animals (continued)

Species	Common name	Concentration	Reference
		Mg. N/100 g. wet tissue	
Fishes (continued)			
Teleostei			
Gadus morhua	Atlantic cod	67-115, 95 (25)	Dyer 1952
		110-146, 125 (50)	Beatty 1939
		44-86, 61 (7)	Ronald and Jakobsen 1947
Melanogrammus aeglefinus	Haddock	43-83, 70 (9)	Dyer 1952
		63-80, 71 (8)	Beatty 1939
		30-53, 43 (5)	Ronald and Jakobsen 1947
Urophycis chuss	Squirrel hake	100-140, 120 (5)	Dyer 1952
		147-176, 166 (5)	Beatty 1939
Brosme brosme	Cusk	65	Dyer 1952
Macrourus berglax	Smooth spined rat-tail	85	Dyer 1952
Hippoglossus hippoglossus	Atlantic halibut	65-75, 70 (5)	Dyer 1952
Hippoglossus stenolepis	Pacific halibut	65	Dyer 1952
Hippoglossus platessoides	American plaice	93	Dyer 1952
Limanda ferruginea	Rusty dab	21-88, 62 (10)	Dyer 1952
		78	Beatty 1939
Limanda limanda	--	30-37, 34 (3)	Love et al. 1959
Pseudopleuronectes americanus	Winter flounder	64-82, 70 (30)	Dyer 1952
Glyptocephalus microcephalus	Lemon sole	36	Ronald and Jakobsen 1947
Glyptocephalus cynoglossus	Witch	43-105, 58 (6)	Beatty 1939
Lophius piscatorius	Goosefish	42-75, 59 (2)	Dyer 1952
Panpus argenteus	--	55	Velankar and Govindan 1958
Mugil sp.	--	24	Velankar and Govindan 1958
Chirocentrus dorab	--	36	Velankar and Govindan 1958
Cypsilurus sp.	--	25	Velankar and Govindan 1958
Sardinella albella	--	54	Velankar and Govindan 1958
Athlennes hians	--	43	Velankar and Govindan 1958

Table 1.--Trimethylamine oxide content of marine animals (continued)

Species	Common name	Concentration	Reference
		Mg. N/100 g. wet tissue	
Fishes (continued)			
Teleostei			
Scoberomorus commersonii	--	28	Velankar and Govindan 1958
Platophrys pantherina	--	36	Velankar and Govindan 1958
Cynoglossus bengalensis	--	36	Velankar and Govindan 1958
Saurida tumbil	--	55	Velankar and Govindan 1958
Sillago sihamer	--	21	Velankar and Govindan 1958
Synagris japonicus	--	40	Velankar and Govindan 1958
Lethrinus cinereus	--	55	Velankar and Govindan 1958
Caranx leptolepsis	--	54	Velankar and Govindan 1958
Gerres sp.	--	40	Velankar and Govindan 1958
Mulloides flavolineatus	--	51	Velankar and Govindan 1958
Arius sona	--	36	Velankar and Govindan 1958
Scatophagus argus	--	33	Velankar and Govindan 1958
Reptilia			
Chelone imbricata	Sea turtle	5	Velankar and Govindan 1958
Mammalia			
--	Whale	Present	Shewan 1951
--	Porpoise	Present	Shewan 1951
Phocaena phocaena	Porpoise	0	Dyer 1952
Balaenoptera musculus	Blue whale	0	Dyer 1952

Occurrence in Fresh-Water Animals

Fresh-water animals, in comparison to marine animals, contain little trimethylamine oxide (tables 2 and 3). The mean content of trimethylamine oxide in marine teleosts was 54.9, whereas in fresh-water teleosts it was only 7.5 mg. trimethylamine oxide nitrogen per 100 g. Trimethylamine oxide is not found in fresh-water plants or in fresh-water zooplankton (Kapeller-Adler and Vering 1931). Dimethyl- and trimethylamine are also absent from fresh-water algae, but methylamine is present. Tertiary amines have been reported in a number of land plants (Challinor 1914, Cromwell 1950, Gessner 1950, Guggenheim 1951, Henry and Grindly 1949, Smith and Young 1953, and Steiner and Stein 1954).

PHYSIOLOGICAL AND BIOCHEMICAL SIGNIFICANCE OF TRIMETHYLAMINE OXIDE

It is not unreasonable to expect that trimethylamine oxide, which is widely distributed in marine organisms, is associated with some function or functions in these animals. It has, however, been difficult to show these expected relationships. Consideration is given in the following sections to the probably functions of trimethylamine oxide in different animals and to the theories on the origin of the oxide.

Osmoregulation and Excretion

Since there are differences in osmoregulation and excretion among the groups of marine and fresh-water animals, the possible relationship of trimethylamine oxide to these functions will be discussed for each group of animals.

Elasmobranchs.--The osmotic pressure of the tissue fluids of the marine elasmobranch is greater than that of sea water or of the urine of these animals (Baldwin 1948, Smith 1931 and 1936). A urea content of 2.0 to 2.5 percent plus trimethylamine oxide may constitute 42 to 55 percent of the total osmotic pressure of elasmobranch plasma (Cohen, Krupp, and Chidsey 1958). Trimethylamine oxide contributes 7 to 12 percent of this total (Hoppe-Seyler 1930). Although this contribution is small, it appears to be important, since renal conservation of the oxide has been shown in elasmobranchs. The filtered oxide is almost completely reabsorbed in the kidney. Hoppe-Seyler (1930) observed that the concentration of the oxide in the urine was 10 percent,or less, of that in the plasma. The mean concentration of oxide in the plasma for 39 specimens of dogfish was 99 ± 14 mg. N/100 ml. Over a 24-hour period, this concentration only varied 6 to 9 mg. N/100 ml. per specimen. This shows that the concentration is controlled over a narrow range (Cohen, Krupp, and Chidsey 1958).

Table 2.--Trimethylamine oxide content of fresh-water animals

Species	Common name	Concentration	Reference
		Mg. N/100 g. wet tissue	
Arthropoda (Crustacea) (Mainly)			
Daphnia and *Coelsphaerium*	Zooplankton	0	Shewan 1951
---	River crab	9	Hoppe-Seyler 1934
Fishes			
Teleostei			
Micropterus dolomieu	Small mouth bass	5	Anderson and Fellers 1952
Esox niger	Pickerel	11	Anderson and Fellers 1952
Morone americana	White perch	12	Anderson and Fellers 1952
Perca flurescens	Yellow perch	0	Anderson and Fellers 1952
Pomoxis nigro-maculatus	Black crappie	15	Anderson and Fellers 1952
Lepomis gibbosus	Sun fish	6	Anderson and Fellers 1952
Lepomis m. machrochirus	Blue gill	17	Anderson and Fellers 1952
Notemigonus c. crysoleucas	Eastern golden shiner	0	Anderson and Fellers 1952
Ameiurus sp.	Bullhead	0	Anderson and Fellers 1952
Esox lucius	Pike	4	Lintzel et al. 1939
Tinca vulgaris	--	5	Lintzel et al. 1939
Trutta fario	Trout	14	Lintzel et al. 1939
Abramas vimba	--	17	Lintzel et al. 1939
Barbus flur	--	6	Lintzel et al. 1939
Anguilla vulgaris	Eel	6	Lintzel et al. 1939
Carassius vulgaris	Crucian carp	5	Lintzel et al. 1939
Perca fluviatilis	Perch	10	Lintzel et al. 1939
Leuciscus erythrophthalmus	Redeye	11	Lintzel et al. 1939
Aspius rapax	--	7	Lintzel et al. 1939
Leuciscus cephalus	Chub	4	Lintzel et al. 1939
Abramas blicca	--	2	Lintzel et al. 1939
Amphibia			
Necturus	--	Present	Wilson and Wolff 1938

Table 3.--Summary of trimethylamine oxide content of marine and fresh-water animals

Kind	No. of species	Concentration		
		Range [1]	Mean	Std. dev. [1] [2]
		Mg. N/100 g.	Mg. N/100 g.	Mg. N/100 g.
Molluscs	35	0-195	35.2	51.3
Crustacea	18	3-105	43.0	27.6
Elasmobranchs	19	29-275	134.5	65.0
Marine teleosts	72	0-143	54.9	26.1
Fresh-water teleosts	21	0-17	7.5	5.4

[1] Where two or more values are reported for a species the mean of these was used to represent the range and to calculate the standard deviation.

[2] The standard deviation was calculated using the relationship

$$\sqrt{\frac{\Sigma x^2 - \frac{(\Sigma x)^2}{N}}{N-1}}$$

where (x) is the content of oxide in milligrams per 100 g. and (N) is the number of species considered.

Teleosts.--The mean freezing-point depression for the fluids of the marine teleost is about -0.7° C., and that for sea water is about -2.0° C. or more. Since the depression of the freezing point varies proportionately with osmotic pressure, the marine teleost must eliminate metabolic products from its tissues against an osmotic-pressure gradient (Baldwin 1951).

The content of trimethylamine oxide in blood plasma of the teleost is low compared to that in the blood plasma of the elasmobranch. The following values have been reported: *Lophius piscatorius* 12-17 mg. N/100 ml. (Brull and Nizet 1954), *Sebastodes* sp. negligible, *Scorpaenichthys marmoratus* negligible, and *Pleuronectidae* sp. negligible (Norris and Benoit 1945a). The small amount of oxide in the teleost blood and the lower osmotic pressure of this blood compared with sea water make it appear unlikely that trimethylamine oxide functions as an osmoregulator. There are, however, several observations that suggest this function. Smith (1958), in the course of Arctic studies, found that the freezing point depression for fluid in the tissue of Arctic teleosts that live near the surface fluctuates between approximately -1.5° in winter and -0.8° C. in summer. The content of tissue oxide in herring in winter is about double that found in summer (Ronald and Jakobsen 1947). The tissue of Arctic fish has a greater content of oxide than does that of fish of the same species caught in the North Sea (Shewan 1951). These data suggest the possibility that trimethylamine oxide could function to increase osmotic pressure to allow the teleost to live in cold water without the danger of becoming frozen.

Since the tissue fluids of the teleost are at a lower osmotic pressure than is its sea-water environment, the animal is in constant threat of being dehydrated. Teleost urine has been shown to be hypotonic to plasma. The teleost conserves water that would go into the production of urine by excreting up to 90 percent of its excretory nitrogen through the gills in the form principally of ammonia (Baldwin 1948). About one-half of the urinary nitrogen excreted by Lophius piscatorius was trimethylamine oxide (Grollman 1929). Using the same animal, Brull and Nizet (1954), however, reported that the main nitrogenous constituent in the urine was creatinine. Evidence is also lacking on the permeability of gill tissue to trimethylamine oxide. It has been presumed that the oxide is not excreted via the gills.

Other marine animals.--The physiological function of trimethylamine oxide in marine invertebrates is not known. Marine invertebrates are generally in osmotic equilibrium with their environment (Florkin 1949). These animals apparently maintain this equilibrium by transfer of inorganic salts and water across membranes; therefore, the oxide probably does not have an important osmotic function in these animals.

Fresh-water teleosts.--The depression of the freezing point of fresh water is seldom greater than -0.02°,whereas that for the tissues of the fresh-water teleost is approximately -0.6° (Baldwin 1951). These data show that the tissues of the fresh-water teleost are at a greater osmotic pressure than is their environment. Considering the low concentration of trimethylamine oxide in fresh-water fish (tables 2 and 3), it does not appear that the small degree of osmotic pressure contributed by the oxide could be considered an important function in these animals.

Nitrogen Metabolism

Although the origin of trimethylamine oxide is unknown, a number of suggestions have been made that relate the occurrence of trimethylamine oxide in an animal to metabolism of nitrogen-containing compounds.

Trimethylamine oxide as a product of protein metabolism.--Hoppe-Seyler (1930) has suggested that trimethylamine oxide is the nontoxic end product of protein metabolism. Thus far no metabolic pathway for synthesis of trimethylamine oxide has been worked out for the fish or for any other living organism. Baldwin (1951) suggested that trimethylamine oxide may be endogenous in origin, since some marine teleosts can excrete up to 30 percent of their total excretory nitrogen in the form of trimethylamine oxide. There are data to show that not all teleosts excrete large amounts of trimethylamine oxide. Wood (1958) showed that trimethylamine oxide made up only a small fraction of the excreta of sculpin, *Leptocottus armatus*, starry flounder, *Platichthys stellatus*, and blue sea-perch, *Taeniotoca lateralis*.

Ogilvie and Warren (1957) reported that trimethylamine oxide may originate in the killifish, *Fundulus heteroclitus*, by an endogenous process. They offered this as an explanation for the apparent accumulation of the oxide in the tissues of fasting animals and of animals on an oxide-free diet. This interpretation is probably incorrect, since the oxide content was reported in units of mg. N/100 g. flesh and no consideration was given the possibility that the animals may have lost weight and thus have shown an apparent increase in tissue oxide.

It has been presumed that if trimethylamine oxide is synthesized by marine teleosts, the last step will involve the conversion of trimethylamine to trimethylamine oxide. Kapeller-Adler and Vering (1931) reported that an enzyme system to catalyze this reaction is lacking in teleosts and amphibia. Such an enzyme system has been demonstrated in man and in other mammals (Lintzel 1935, Norris and Benoit 1945b, Tarr 1941).

Trimethylamine oxide from an exogenous source.--Benoit and Norris (1945) showed that young salmon raised in a marine environment on an oxide-free diet do not accumulate trimethylamine oxide in muscle tissue. When the salmon were fed a diet containing oxide, some retention resulted. Hashimoto and Okaichi (1958a, b) reported that dietary trimethylamine oxide is accumulated in the muscles of the goldfish, *Carassius auratus*, and the eel, *Anguilla japonica*; however, when these fish were fed an oxide-free diet, the oxide was not found in the muscle. Okaichi, Manabe, and Hashimoto (1959) reported that the globefish, *Fugu niphobles*, and filefish, *Monacanthus cerrhifer*, accumulate ingested trimethylamine oxide in their tissues, whereas the jack mackerel, *Trachurus japonicus*, does not.

If we accept this idea that trimethylamine oxide in the food is accumulated in the tissue of fishes, then we should look for the synthetic or metabolic source of the oxide at some point in the food chain. Considering the food chain in reverse, we find that the larger teleosts utilize smaller fishes and other larger marine animals as food; these animals utilize the zooplankton, the zooplankton utilize the phytoplankton, and the phytoplankton synthesize their food by photosynthesis. The first point in the food chain where trimethylamine oxide is found is in the zooplankton. The oxide found in zooplankton could get there by two routes. The simpler would be the conversion of the trimethylamine found in the food--marine plants (Channing and Young 1953, Kapeller-Adler and Vering 1931)--to the oxide by the zooplankton. The more complex route would involve the synthesis of the oxide in the zooplankton from smaller fragments. It appears that a study of zooplankton in which the trimethylamine oxidase system and transmethylation systems are investigated might give valuable information on this aspect of the problem.

Trimethylamine oxide as a methyl donor and its relationship to other methylated compounds.--Barrenscheen and Pantlitschko (1950) have shown that trimethylamine oxide acts as a methyl donor for the synthesis of choline in the muscle brei of guinea pig. This system, which is called cholinepherase, has not been reported in marine animals.

Since trimethylamine and trimethylamine oxide do not prevent lipotropic changes in the livers of experimental animals as does choline (Moyer and du Vigneaud 1942), the transmethylation functions that have been given to choline have not been ascribed to trimethylamine oxide (Arnstein 1955).

Bach (1945) suggests that some marine animals possess a large methylating capacity, as is shown by the occurrence of tetramine, $\left[(CH_3)_4\ N\right]^+$, in the *Actinia* (sea anemone). A number of other methylated compounds have been found in marine animals. Some of these are choline, glycine betaine, gamma-butyrobetaine, homarine, trigonelline, stachydrine (Shewan 1951), dimethylthetin (Patton 1958), methionine, and dimethylbetaine (Welsh and Prock 1958).

Other tertiary amine oxides.--Tertiary amine oxides, the family of compounds to which trimethylamine oxide belongs, are known to occur as components of plants and animals (Fish, Sweeley, and Horning 1956). It is not known, however, what the function of these oxides might be in the plants and animals. The frequency with which they occur in living organisms suggests that they are something more than terminal oxidation products of amines (Fish, Johnson, and Horning 1956). It has been found possible to carry out a ferric-ion induced rearrangement of tertiary-amine oxides under mild conditions to yield as products a secondary amine plus formaldehyde or formic acid. This rearrangement has been shown for N,N-dimethyl-tryptamine oxide (Fish, Johnson, and Horning 1956) and trimethylamine oxide (Vaisey 1956).

Formation of trimethylamine oxide in mammals as a result of degradation of choline or choline-containing derivatives.--Choline fed to rats is excreted in the urine as trimethylamine oxide (De la Huerga and Popper 1952, Norris and Benoit 1945b). This is caused by bacterial breakdown of choline to trimethylamine in the intestinal tract (Dyer and Wood 1947); and following absorption, the trimethylamine is converted to the oxide by trimethylamine oxidase. Very small amounts of trimethylamine oxide and trimethylamine are present in mammal tissues; however, no function has been reported for these compounds.

SUMMARY

Trimethylamine oxide has been reported in a coelenterate, an echinoderm, some molluscs, all crustaceans, all elasmobranchs, most

teleosts, a reptile, and mammals from a marine environment and in a crustacean, most teleosts, and an amphibian from a fresh-water environment. In general, animals from a marine environment contain much greater amounts of the oxide than do animals from a fresh-water environment. The oxide is unevenly distributed among marine animals. In marine teleosts, the factors of geographic environment, species, season, size, and location in the animal affect the content of oxide.

Trimethylamine oxide has not been reported in plants; however, other tertiary amine oxides and tertiary amines have been reported.

Trimethylamine oxide appears to contribute to the osmotic pressure of the elasmobranch. It is not known if the oxide functions as an osmotic pressure agent in the teleost.

Certain marine animals possess a strong methylating capacity. It has been reported that trimethylamine oxide will methylate choline in a system isolated from a mammal; however, no such system has been isolated from a marine animal.

It has been suggested that trimethylamine oxide is endogenous in origin and that it might be a product of protein metabolism. There is, however, no direct evidence to prove this point. Mechanisms for the synthesis of trimethylamine oxides have not been demonstrated in animals or plants.

In higher marine animals, the occurrence of trimethylamine oxide can be caused by an exogenous source, since it has been shown that the oxide can accumulate in the muscles of fish from trimethylamine oxide in the food. The zooplankton are the first animals in the food chain that contain the oxide. Zooplankton could obtain the oxide by converting exogenous trimethylamine to the oxide or by synthesizing the oxide from smaller fragments. The primary origin, however, of trimethylamine oxide in marine animals is still to be explained.

LITERATURE CITED

ANDERSON, D. W., JR., AND C. R. FELLERS
1949. Some aspects of trimethylamine formation in swordfish. Food Technology, vol. 3, no. 8, August, pp. 271-273.

ANDERSON, D. W., AND C. R. FELLERS
1952. The occurrence of trimethylamine and trimethylamine oxide in fresh-water fishes. Food Research, vol. 17, no. 6, November-December, pp. 472-474.

ARNSTEIN, H. R. V.
1955. The function of vitamin B_{12} in animal metabolism. In "The Biochemistry of Vitamin B_{12}," Biochemical Society Symposia No. 13, pp. 92-108, University Press, Cambridge, England.

ASANO, M., AND H. SATO
1954. Biochemical studies on octopus. I - Trimethylamine and trimethylamine oxide contents of octopus. Tohoku Journal of Agricultural Research, vol. 5, no. 3, December, pp. 191-195.

BACH, S. J.
1945. Biological methylation. Biological Reviews, vol. 20, no. 4, October, pp. 158-76.

BALDWIN, E.
1948. Dynamic aspects of biochemistry. Cambridge University Press, New York, New York, 544 pp.

BALDWIN, E.
1951. Comparative aspects of biochemistry in fish. In "The Biochemistry of Fish," Biochemical Society Symposia No. 6, pp. 3-7, University Press, Cambridge, England.

BARRENSCHEEN, H. K., AND M. PANTLITSCHKO
1950. Methylation by means of plant and animal tissues. VII - The mechanism of methyl transfer for choline. Chemical Abstracts, vol. 44, no. 15, August 10, p. 6943^g.

BEATTY, S. A., AND N. E. GIBBONS
1937. The measurement of spoilage in fish. Journal of the Biological Board of Canada, vol. 3, no. 1, November-December, pp. 77-91.

BEATTY, S. A.
1939. Studies of fish spoilage. III - The trimethylamine oxide content of the muscles of Nova Scotia fish. Journal of the Fisheries Research Board of Canada, vol. 4, no. 4, March, pp. 229-232.

BENOIT, G. J., JR., AND E. R. NORRIS
1945. Studies on trimethylamine oxide. II - The origin of trimethylamine oxide in young salmon. Journal of Biological Chemistry, vol. 158, no. 2, April, pp. 439-442.

BRULL, L., AND E. NIZET
1954. Blood and urine constituents of *Lophius piscatorius*. Chemical Abstracts, vol. 48, no. 5, March 10, p. 2930^e.

CASTELL, C. H.
1949a. The influence of trimethylamine oxide on the bacterial reduction of redox indicators. Journal of the Fisheries Research Board of Canada, vol. 7, no. 10, January, pp. 567-575.

CASTELL, C. H.
1949b. Bacteria and the reduction of trimethylamine oxide. Food in Canada, vol. 9, no. 10, September, pp. 11-14.

CHALLINOR, R. W.
1914. The occurrence of trimethylamine and its origin in the Australian salt bush, *Rhagodia hastata*, R. Br. Chemical Abstracts, vol. 8, no. 13, July 10, p. 2454.

CHANNING, D. M., AND G. T. YOUNG
1953. Amino acids and peptides. X - The nitrogenous constituents of some marine algae. Journal of the Chemical Society, August, pp. 2481-2491.

COHEN, J. J., M. A. KRUPP, AND C. A. CHIDSEY
1958. Renal conservation of trimethylamine oxide by the spiny dogfish, *Squalus acanthias*. American Journal of Physiology, vol. 194, no. 2, August, pp. 229-235.

COOK, A. S.
1931. A study of the occurrence of trimethylamine in marine animals. Canadian Chemistry and Metallurgy, vol. 15, no. 1, January, pp. 22-23.

CROMWELL, B. T.
1950. The micro-estimation and origin of trimethylamine in Chenopodium vulvaria L. The Biochemical Journal, vol. 46, no. 5, May, pp. 578-582.

DE LA HUERGA, J., AND H. POPPER
1952. Factors influencing choline absorption in the intestinal tract. Journal Clinical Investigation, vol. 31, no. 6, June, pp. 598-603.

DYER, F. E., AND A. J. WOOD
1947. Action of Enterobacteriaceae on choline and related compounds. Journal of the Fisheries Research Board of Canada, vol. 7, no. 1, February, pp. 17-21.

DYER, W. T.
1952. Amines in fish muscle. VI - Trimethylamine oxide content of fish and marine invertebrates. Journal of the Fisheries Research Board of Canada, vol. 8, no. 5, January, pp. 314-324.

FISH, M. S., N. M. JOHNSON, AND E. C. HORNING
1956. t-Amine oxide rearrangements. N,N-dimethyltryptamine oxide. Journal of the American Chemical Society, vol. 78, no. 15, August 15, pp. 3668-3671.

FISH, M. S., C. C. SWEELEY, AND E. C. HORNING
1956. Tert-amine oxice rearrangements. Chemistry and Industry, April, p. R24.

FLORKIN, M.
1949. Biochemical evolution. Academic Press, Inc., New York, New York, chapter V, p. 72.

GESSNER, O.
1950. Pharmacology of Arnica montana. Chemical Abstracts, vol. 44, no. 5, March 10, p. 2121^{g}.

GROLLMAN, A.
1929. The urine of the goosefish (Lophius piscatorius); its nitrogenous constituents with special reference to the presence in it of trimethylamine oxide. Journal of Biological Chemistry, vol. 81, no. 2, February, pp. 267-278.

GUGGENHEIM, M.
1951. Die biogenen amine. S. Karger, Publisher, New York, New York, pp. 67-72.

HASHIMOTO, Y., AND T. OKAICHI
1958a. Trimethylamine oxide in fish muscle. I - The origin of trimethylamine oxide in goldfish muscle. Bulletin of the Japanese Society of Scientific Fisheries, vol. 24, no. 8, pp. 640-4.

HASHIMOTO, Y., AND T. OKAICHI
1958b. Trimethylamine oxide in fish muscle. II - Trimethylamine oxide in the muscle of eels kept in fresh and brackish water. Bulletin of the Japanese Society of Scientific Fisheries, vol. 24, no. 8, pp. 645-647.

HENRY, A. J., AND D. N. GRINDLEY
1949. Courbonia virgata - its chemical compound and basic constituents. Journal of the Society of Chemistry and Industry, vol. 68, no. 1, January, pp. 9-12.

HOPPE-SEYLER, F. A.
1930. The limitations and significance of biological methylation processes. Zeitschrift fur Bioligie, vol. 90, no. 5, September, pp. 433-466.

HOPPE-SEYLER, F. A.
1934. Trimethylamine oxide and other nitrogen bases in crab muscles. Chemical Abstracts, vol. 28, no. 1, January 10, p. 2169.

KAPELLER-ADLER, R., AND F. VERING
1931. The occurrence of methylated ammonium compounds in sea tangle and some feeding experiments with trimethylamine performed in cold blooded animals. Biochemische Zeitschrift, vol. 243, no. 4-6, December, pp. 292-309.

KAWABATA, T.
1953. Studies on the trimethylamine oxide reductase. I - Reduction of trimethylamine oxide in the dark muscle of pelagic migrating fish under aseptic condition. Bulletin of the Japanese Society of Scientific Fisheries, vol. 19, no. 4, pp. 505-12.

KERMACK, W. O., H. LEES, AND J. D. WOOD

1955. Some non-protein constituents of the tissues of the lobster. The Biochemical Journal, vol. 60, no. 3, July, pp. 424-428.

LINTZEL, W., H. PFEIFFER, AND I. ZIPPEL
1939. Studies on trimethylammonium bases. IV - The occurrence of trimethylamine oxide in the muscles of fresh-water fish. Chemical Abstracts, vol. 33, no. 21, November 10, p. 8827.

LINTZEL, W.
1935. Studies on trimethylammonium bases. III - Trimethylammonium bases in human urine. Chemical Abstracts, vol. 29, no. 4, February 20, p. 1121.

LOVE, R. M., J. A. LOVERN, AND N. R. JONES

1959. The chemical composition of fish tissues. Food Investigation Special Report No. 69, pp. 37-38, Her Majesty's Stationery Office, London.

MOHR, M.
1937. Nitrogen-containing constituents of the jellyfish (*Cyanea capillata*). Zeitschrift fur Biologie, vol. 98, no. 2, May, pp. 120-4.

MOYER, A. W., AND V. du VIGNEAUD
1942. The structural specificity of choline and betaine in transmethylation. Journal of Biological Chemistry, vol. 143, no. 2, April, pp. 373-382.

NORRIS, E. R., AND G. J. BENOIT, JR.
1945a. Studies on trimethylamine oxide. I - Occurrence of trimethylamine oxide in marine organisms. Journal of Biological Chemistry, vol. 158, no. 2, April, pp. 433-438.

NORRIS, E. R., AND G. J. BENOIT, JR.
1945b. III - Trimethylamine oxide excretion by the rat. Journal of Biological Chemistry, vol. 158, no. 2, April, pp. 443-448.

OGILVIE, J.M.G., AND A. A. WARREN
1957. The occurrence of trimethylamine oxide in *Fundulus heteroclitus*. Canadian Journal of Zoology, vol. 35, no. 6, December, pp. 735-745.

OKAICHI, T., M. MANABE, AND Y. HASHIMOTO
1959. Trimethylamine oxide in fish muscle. III - The origin of trimethylamine oxide in marine fish muscle. Bulletin of the Japanese Society of Scientific Fisheries, vol. 25, no. 2, June, pp. 136-140.

PATTON, S.
1958. Chemical aspects of flavor research on milk and its products. In "Flavor Research and Food Acceptance," p. 315, Reinhold Publishing Corporation, New York, New York.

RONALD, O. A., AND F. JAKOBSEN
1947. Trimethylamine oxide in marine products. Journal of the Society of Chemistry and Industry, vol. 66, no. 5, May, pp. 160-166.

SHEWAN, J. M.
1951. The chemistry and metabolism of the nitrogenous extractives in fish. In "The Biochemistry of Fish," Biochemical Society Symposia No. 6, pp. 28-48, University Press, Cambridge, England.

SHEWAN, J. M., AND N. R. JONES
1957. Chemical changes occurring in cod muscle during chill storage and their possible use as objective indices of quality. Journal of the Science of Food Agriculture, vol. 8, no. 8, August, pp. 491-498.

SIMIDU, W., S. HIBIKI, S. SIBATA, AND K. TAKEDA
1953. Studies on muscle of aquatic animals. XVI - Distribution of extractive nitrogens in muscles of several kinds of gastropods. Bulletin of the Japanese Society of Scientific Fisheries, vol. 19, no. 8, pp. 871-876.

SIMIDU, W., AND S. HIBIKI
1957. Studies on putrefaction of aquatic products. XXIV - On spoilage of some shellfishes. Bulletin of the Japanese Society of Scientific Fisheries, vol. 23, no. 5, pp. 255-259.

SMITH, A. V.
1958. Life at low temperatures. Nature, vol. 182, no. 4640, October 4, pp. 911-913.

SMITH, D. G., AND E. G. YOUNG
1953. The nitrogenous constituents of *Fucus vesiculosus*. Journal of Biological Chemistry, vol. 205, no. 2, December, pp. 849-58.

SMITH, H. W.
1931. The absorption and excretion of water and salts by the elasmobranch fishes. II - Marine elasmobranchs. American Journal of Physiology, vol. 98, no. 2, September, pp. 296-310.

SMITH, H. W.
1936. The retention and physiological role of urea in the elasmobranchii. Biological Reviews, vol. 11, no. 1, January, pp. 49-82.

STEINER, M., AND E. STEIN
1954. Paper chromatographic detection of primary, secondary, and tertiary amines in plants. Chemical Abstracts, vol. 48, no. 14, July 25, p. 8333^{f}.

SUYAMA, M., AND T. TOKUHIRO
1954. Urea content and ammonium formation of the muscle of cartilagenous fishes. III - The distribution of urea and trimethylamine oxide in different parts of the body. Bulletin of the Japanese Society of Scientific Fisheries, vol. 19, no. 10, pp. 1003-1006.

SUYAMA, M.
1958. Studies on the buffering capacity of fish muscle. II - Effects of trimethylamine oxide and urea on buffering capacity. III - Effects of trimethylamine and ammonia on the buffering capacity of fish muscle and the change in the capacity during degradation. Bulletin of the Japanese Society of Scientific Fisheries, vol. 24, no. 4, pp. 271-276.

TARR, H.L.A.
1939. The bacterial reduction of trimethylamine oxide to trimethylamine. Journal of the Fisheries Research Board of Canada, vol. 4, no. 5, February, pp. 367-377.

TARR, H.L.A.
1940. Specificity of triamineoxidease. Journal of the Fisheries Research Board of Canada, vol. 5, no. 2, December, pp. 187-196.

TARR, H.L.A.
1941. The fate of trimethylamine oxide and trimethylamine in man. Journal of the Fisheries Research Board of Canada, vol. 5, no. 3, January, pp. 211-216.

VAISEY, E. B.
1956. The non-enzymic reduction of trimethylamine oxide to trimethylamine, dimethylamine and formaldehyde. Canadian Journal of Biochemistry and Physiology, vol. 34, no. 6, November, pp. 1085-1090.

VELANKAR, N. K., AND T. K. GOVINDAN
1958. Preliminary study of the distribution of nonprotein nitrogen in some marine fishes and invertebrates. Proceedings of the Indian Academy of Science, vol. 47B, no. 4, April, pp. 202-209.

WELSH, J. H., AND P. B. PROCK
1958. Quaternary ammonium bases in the coelenterates. Biological Bulletin, vol. 115, no. 3, December, pp. 551-561.

WILSON, D. W., AND W. A. WOLFF
1938. Basic nitrogenous extractives of Necturus muscle. Journal of Biological Chemistry, vol. 124, no. 1, June, pp. 103-106.

WOOD, J. D.
1958. Nitrogen excretion in some marine teleosts. Canadian Journal of Biochemistry and Physiology, vol. 36, no. 12, December, pp. 1237-1242.